

Effect of 2020–21 and 2021–22 Highly Pathogenic Avian Influenza H5 Epidemics on Wild Birds, the Netherlands

Valentina Caliendo, Erik Kleyheeg, Nancy Beerens, Kees C.J. Camphuysen, Rommert Cazemier, Armin R.W. Elbers, Ron A.M. Fouchier, Leon Kelder, Thijs Kuiken, Mardik Leopold, Roy Slaterus, Marcel A.H. Spierenburg, Henk van der Jeugd, Hans Verdaat, Jolianne M. Rijks

The number of highly pathogenic avian influenza (HPAI) H5-related infections and deaths of wild birds in Europe was high during October 1, 2020–September 30, 2022. To quantify deaths among wild species groups with known susceptibility for HPAI H5 during those epidemics, we collected and recorded mortality data of wild birds in the Netherlands. HPAI virus infection was reported in 51 bird species. The species with the highest numbers of reported dead and infected birds varied per epidemic year; in 2020–21, they were within the Anatidae family, in particular barnacle geese (*Branta leucopsis*) and in 2021–22, they were within the sea bird group, particularly Sandwich terns (*Thalasseus sandvicensis*) and northern gannet (*Morus bassanus*). Because of the difficulty of anticipating and modeling the future trends of HPAI among wild birds, we recommend monitoring live and dead wild birds as a tool for surveillance of the changing dynamics of HPAI.

The dynamics of highly pathogenic avian influenza (HPAI) virus infection of the H5 GS/GD lineage (clade 2.3.4.4b) have dramatically changed for wild birds. For 2 recent epidemic seasons (2020–21 and 2021–22), HPAI H5 viruses have adapted to survive long term in wild bird populations; they are now considered enzootic in wild birds (1–3). This change in status was supported by the shift in HPAI epidemiol-

ogy during summer 2021, as the virus circulated continuously in northwestern Europe and Scandinavia (1,4). High rates of virus detection in wild and captive birds continued in 2022 for the largest epidemic observed to date in Europe (4). The circulation of HPAI virus during the 2022 breeding season exposed several colony-breeding seabird species along the northwest coast of Europe to infection (4–7), culminating in a high number of HPAI virus detections in dead wild birds during June–August 2022. At that time several seabird species exhibited widespread and massive deaths from HPAI H5N1 virus at their breeding colonies in Germany, the Netherlands, France, and the United Kingdom (4–8). Authorities have recommended reporting the number of wild birds found dead or ill during HPAI-associated dieoffs, both to contribute to the understanding of the ecologic effect of HPAI outbreaks and for targeted, evidence-based policy making (4,9).

The extent to which bird species are associated with HPAI largely depends on how often each species has tested positive (10). Several factors play a role in this assessment: species-specific susceptibility to clinical disease, local population size, geographic and climate circumstances, reporter effort, and number of birds screened during surveillance.

Author affiliations: Utrecht University, Utrecht, the Netherlands (V. Caliendo, J.M. Rijks); Sovon, Dutch Centre for Field Ornithology, Nijmegen, the Netherlands (E. Kleyheeg, R. Slaterus); Wageningen Bioveterinary Research, Lelystad, the Netherlands (N. Beerens); Royal Netherlands Institute for Sea Research, Den Burg, the Netherlands (K.C.J. Camphuysen); Wetterskip Fryslan, Leeuwarden, the Netherlands (R. Cazemier); Wageningen Bioveterinary Research, Lelystad (A.R.W. Elbers); Erasmus University Medical Center, Rotterdam, the Netherlands

(R.A.M. Fouchier, T. Kuiken); Staatsbosbeheer, Amersfoort, the Netherlands (L. Kelder); Wageningen Marine Research, Den Helder, the Netherlands (M. Leopold, H. Verdaat); Netherlands Food and Consumer Product Safety Authority, Utrecht (M.A.H. Spierenburg); Vogeltrekstation—Dutch Centre for Avian Migration and Demography NIOO-KNAW, Wageningen, the Netherlands. (H. van der Jeugd); Waarneming.nl, Stichting Observation International, Den Helder (H. Verdaat)
DOI: <http://doi.org/10.3201/eid3001.230970>

In the Netherlands, the AI-Impact working group, a consortium of ornithologists, virologists, epidemiologists, nature managers, and animal health organizations, has been active since 2020 to provide up-to-date information on wild bird mortalities during HPAI outbreaks. The aims of the consortium are to identify the range of wild bird species exposed and affected by HPAI, and to estimate the HPAI-associated level of mortality of wild birds, also relative to their population size.

For this study, we collected dead-bird reports and virologic diagnoses from a variety of sources to estimate species-specific mortality prevalence caused by the 2020–21 and 2021–22 HPAI H5 epidemics in the Netherlands. During the epidemics, mortality data were collected monthly and provided near-real-time information on the trend of the epidemic for interested organizations.

Methods

The methods for this study were similar to those used by Kleyheeg et al. (11). In brief, we collected wild bird mortality data from 2 complete bird influenza seasons, October 1, 2020–September 30, 2021, and October 1, 2021–September 30, 2022. The national competent authorities reported dead wild birds to the AI-Impact consortium as part of the national animal infectious disease surveillance program and by citizen scientists who were invited to report sightings of dead birds on the web platforms of the AI-Impact members (<https://dwhc.nl/dode-vogels-melden/>, <https://www.sovon.nl/nl/content/vogelgriep>, <https://www.nvwa.nl/onderwerpen/vogelgriep-preventie-en-bestrijding>, <https://waarneming.nl>). In addition, all seabird strandings data along the Dutch North Sea and Wadden Sea coasts were analyzed as part of a long-term monitoring project and checked for unusually high numbers of stranded birds with effort correction (number per km of coastline searched, $n \text{ km}^{-1}$) (12,13). Unusually high densities (i.e., stranded birds per area) were ≥ 5 times background densities, as measured using identical surveys from the previous 40 years in any given month. Double counts did not occur in this dataset because carcasses were marked.

We categorized reports by bird species, date, and location of finding. Double counts (e.g., multiple entries for the same species on the same date, at the same location from the same observer) were excluded as much as possible. Consistent with similar studies, we found it highly likely that the number of reported carcasses substantially underestimates actual deaths; for example, collection rates of water bird carcasses during typical avian botulism outbreaks are 10%–25% (14).

We categorized wild bird mortality reports into 4 groups: Anatidae (i.e., geese, swans, ducks), other water birds (including gulls, grebes, herons, cormorants, waders, rallids), raptors, and other land birds. We classified birds of the families Podicipedidae, Laridae, Stercorariidae, Alcidae, Gaviidae, Procellariidae, Sulidae, and Phalacrocoracidae, in 1 subgroup, sea birds. We analyzed mortality data of selected species individually, because they experienced particularly high mortality rates during either the 2020–21 epidemic (i.e., barnacle goose [*Branta leucopsis*], common buzzard [*Buteo buteo*], peregrine falcon [*Falco peregrinus*], great black-backed gull [*Larus marinus*]), or during the 2021–22 epidemic (i.e., Sandwich tern [*Thalasseus sandwicensis*], northern gannet [*Morus bassanus*]). We used data from the public database of Sovon (Dutch Centre for Field Ornithology, Nijmegen, the Netherlands; <https://www.sovon.nl>) to compare the number of reported dead wild birds per avian group during October–March (classified as winter mortality) and April–September (classified as summer mortality) between the 2020–21 and 2021–22 epidemics; we then compared data for the same months of 2010–11 with 2015–16 as described by Kleyheeg et al. (11). In the later period (2010–2016), there had been no outbreaks of HPAI in wild birds in the Netherlands. We tested a limited number of wild bird carcasses (Appendix 1 Table, <https://wwwnc.cdc.gov/EID/article/30/1/23-0970-App1.xlsx>) for HPAI virus by real-time reverse transcription PCR on oropharyngeal and cloacal swabs as previously described (15,16). We submitted groups of ≥ 3 dead wild birds of certain categories (i.e., Anatidae, water birds) found dead at the same location, and single birds of other susceptible species (i.e., raptors) that were suspected of being HPAI virus-infected, for virologic analysis.

We used species data on live population estimates from the public database of Sovon to evaluate mortality rates by bird species (Table; Appendix 1 Table). Population size represents the estimated lowest and highest number of birds wintering in the Netherlands, based on census data for 2013–2020 from Sovon. For summer migratory species, population size represents the estimated lowest and highest number of birds migrating to the Netherlands, based on census data for 2016–2021 from Sovon.

Results

A total of 16,631 wild birds of 160 species were reported dead in the Netherlands in October 1, 2020–September 30, 2021. Water birds including Anatidae accounted for 70% of the total deaths reported and land birds, including raptors, the remaining 30% (Table 1).

RESEARCH

Table. Reported bird species, nonbreeding population size estimates, number of carcasses, and RT-PCR test results for wild birds sampled during 2020–21 and 2021–22 HPAI epidemics, the Netherlands*

Avian group and species	Maximum estimated nonbreeding population size, ×1,000	No. carcasses (mortality rate, %) [†]		No. carcasses HPAI positive/no. tested	
		2020–21	2021–22	2020–21	2021–22
Anatidae		7,901	14,309	361/628	173/416
Geese		4,802	8,867	234/332	154/260
Barnacle goose	710–870	3,435 (1.5–4.8)	5,310 (2.4–7.4)	147/171	77/104
Graylag goose	550–670	390 (0.2–0.7)	1,054 (0.7–1.9)	30/59	53/60
Unidentified species	NA	607	1,653	35/59	36/60
Swans		996	1,453	60/136	2/17
Mute swan	41–48	305 (2.5–7.4)	479 (3.9–11)	38/93	0
Unidentified species	NA	629	969	19/54	2/17
Ducks		2,103	3,985	67/160	17/139
Eurasian wigeon	820–950	125 (<0–0.01)	300 (0.1–0.3)	12/13	1/9
Tufted duck	220–280	45 (0.6–2.5)	34 (0.01–0.1)	1/19	0
Other waterbirds		4,068	21,895	19/162	95/245
Grebes	NA	62	164	0/2	4/10
Hérons	NA	250	232	0/33	3/23
Cormorants	NA	234	371	2/14	2/35
Waders	NA	1,045	1,713	9/49	10/14
Rallids	NA	327	472	0/2	1/23
Sea bird		2,371	19,340	16/102	75/140
Gulls		1,074	5,538	7/61	37/100
Great black-backed gull	25–100	137 (0.01–5.4)	372 (1.4–14.8)	1/1	1/3
Sandwich tern	27–120 [‡]	0	5,166 (17.2–>90) [§]	0	29/33
Northern gannet	4–27	203	2,215 (32.8–>90)	0	6/11
Raptors		1,011	763	42/155	83/149
Common buzzard	30–50	365 (2.9–12.1)	363 (2.9–12.1)	27/91	55/81
Peregrine falcon	0.5–0.6	27 (18–54)	28 (18–56)	4/5	9/11
Other land birds		3,651	3,850	2/40	6/59
Corvids	NA	271	363	1/24	4/26
Total		16,631	41,519	427/985	357/869

*Data from Sovon (Dutch Centre for Field Ornithology, Nijmegen, the Netherlands). HPAI, highly pathogenic avian influenza; NA, not available; RT-PCR, reverse transcription PCR.

[†]Expressed as fraction of the nonbreeding population. Lower and higher values are calculated considering the 10%–25% collection rates, as described by Kleyheeg et al. (11).

[‡]Estimated migration maximum.

[§]Expressed as fraction of the migrant population.

Anatidae by themselves represented 50% of the total deaths reported. Of the bird carcasses identified to species, by far the highest number of deaths were reported for barnacle geese ($n = 3,435$). The next highest numbers of dead animals were reported for graylag geese ($n = 390$), common buzzards ($n = 365$), and mute swans (*Cygnus olor*) ($n = 305$). HPAI virus infection was reported in 45 species (Appendix 1 Table). The species with the highest numbers of reported dead and infected birds were within the Anatidae group: barnacle geese, graylag geese, and mute swans. Common buzzard was the species with the highest numbers of reported dead and infected birds within the raptor group. Expressed as fraction of the nonbreeding population, and after accounting for detection probability, the reported dead birds represented up to 4.8% of barnacle geese, 0.7% of graylag geese, and 7.4% of mute swans (Table 1). We found the highest mortality rates occurred in raptors and scavenging species: relative to their wintering populations, up to 54% of peregrine falcons, 12.1% of common buzzards, and 5.4% of great black-backed gulls may have died.

A total of 41,519 wild birds of 158 species were reported dead in the Netherlands during October 1, 2021–September 30, 2022. Water birds including Anatidae accounted for 80% and land birds including raptors for the remaining 20% of the total deaths reported (Table 1). Sea birds represented >40% and Anatidae 30% of the total deaths. Of the bird carcasses identified to species, the highest number was reported for the barnacle goose ($n = 5,310$). The next highest numbers of dead individuals were reported for Sandwich terns ($n = 5,166$), and northern gannets ($n = 2,215$). HPAI virus infection was confirmed in 51 species (Appendix 1 Table). The species with the highest numbers of reported dead and infected birds were within the sea bird and Anatidae groups, and the species most represented were the Sandwich tern and the barnacle goose. Expressed as a fraction of the nonbreeding population, and after accounting for detection probability, the reported dead birds represented 32.8%–90% of northern gannets and up to 7.4% of barnacle geese (Table 1). The Sandwich tern appears as a summer breeder in the Netherlands;

after accounting for detection probability, the reported dead birds represented 17.2%–90% of the estimated migrant population of Sandwich terns. We found that high mortality rates also occurred in raptors: up to 56% of wintering populations of peregrine falcons and 12.1% of common buzzards may have died. Mortality rates in winter or summer months were higher than the average estimates in previous years (i.e., compared to the same timeframe in 2011–2016, years in which major wild bird mortalities from outbreaks of HPAI virus did not occur). In particular, the number of reported carcasses was >50 times higher for geese in winter 2022 and >1,000 times higher for Sandwich terns in summer 2022 (Figure 1).

During the 2020–21 epidemic in the Netherlands, wild bird deaths clustered in 2 peaks, the first in November 2020 and the second, smaller peak in April–May 2021 (Figure 2). During both peaks, barnacle geese were among the species most severely affected. During the 2021–22 epidemic in the Netherlands, wild bird deaths also showed 2 peaks, the first in January 2022 and the second in June 2022 (Figure 2). During the first peak, barnacle geese were again among the species most severely affected, and during the second peak, sea birds were the most severely

affected. The virus was still detected in October 2022, but that date was considered the start of the new HPAI 2022–23 outbreak.

Discussion

HPAI dynamics in wild birds are constantly evolving. The 2020–21 HPAI H5 epidemic was more devastating than earlier HPAI H5 epidemics in Europe, causing high numbers of HPAI infections and deaths in many species of wild birds (4,9,17–19). Goose species, such as the barnacle goose, accounted for the highest number of casualties. During that epidemic, high prevalence of infection in geese was also reported in Germany and United Kingdom (2). In our study, several duck species consistently tested positive for HPAI H5 virus during the epidemic; however, reported deaths for those species were lower than for goose species. This finding represents a different scenario than that of the 2016–17 HPAI H5 epidemic, in which duck species, such as Eurasian wigeons (*Mareca penelope*) and tufted ducks (*Aythya fuligula*), experienced the highest number of deaths (11). The high mortality rate of barnacle geese during the 2020–21 HPAI H5 epidemic is unprecedented. Barnacle geese are one of the most abundant geese species in the Netherlands

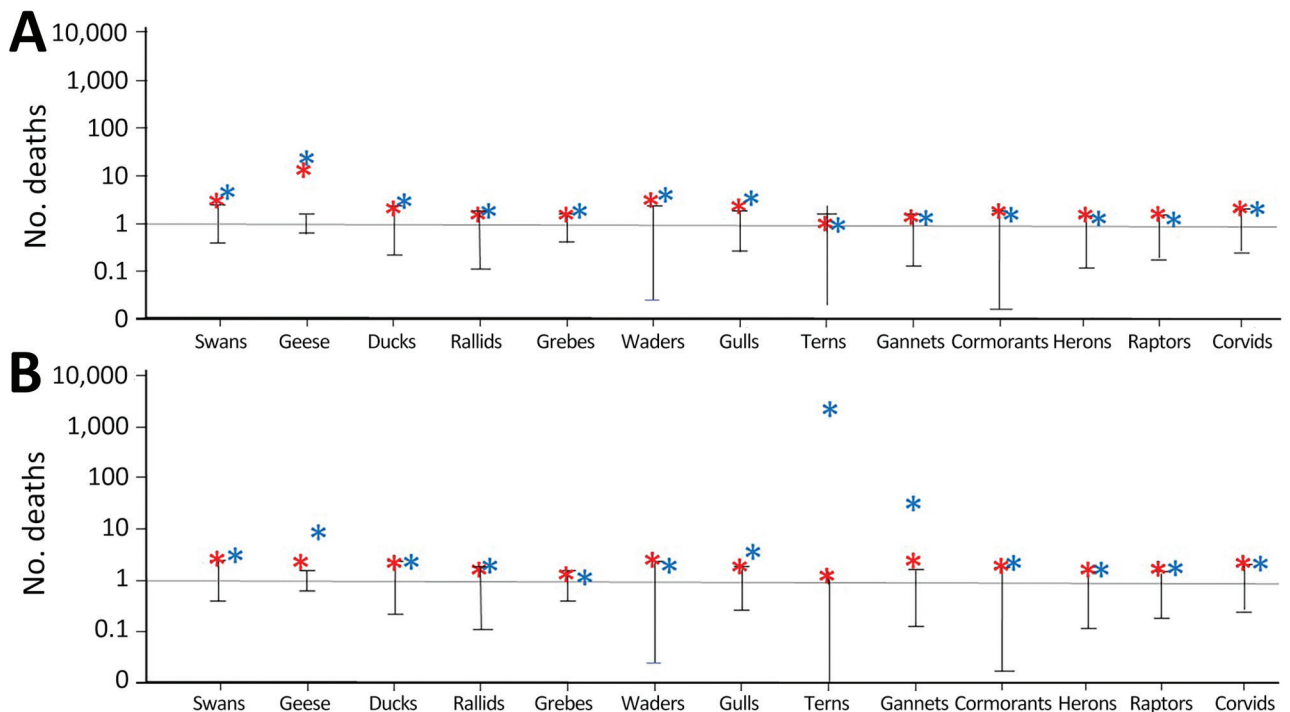


Figure 1. Relative number of reported deaths during the highly pathogenic avian influenza epidemics of 2020–21 (red asterisks), and 2021–22 (blue asterisks), the Netherlands. Deaths are relative to the normalized number of deaths reported over the same period from 2011–2012 to 2015–2016 (average is 1; data from Sovon, Dutch Centre for Field Ornithology, Nijmegen, the Netherlands). A) Deaths reported in the winter months, October–March. B) Deaths reported in the summer months, April–September. The y-axis is on a log scale; reported relative number of deaths among geese during winter 2021–22 was >50 larger than in the previous years. Error bars indicate maximum and minimum deaths.

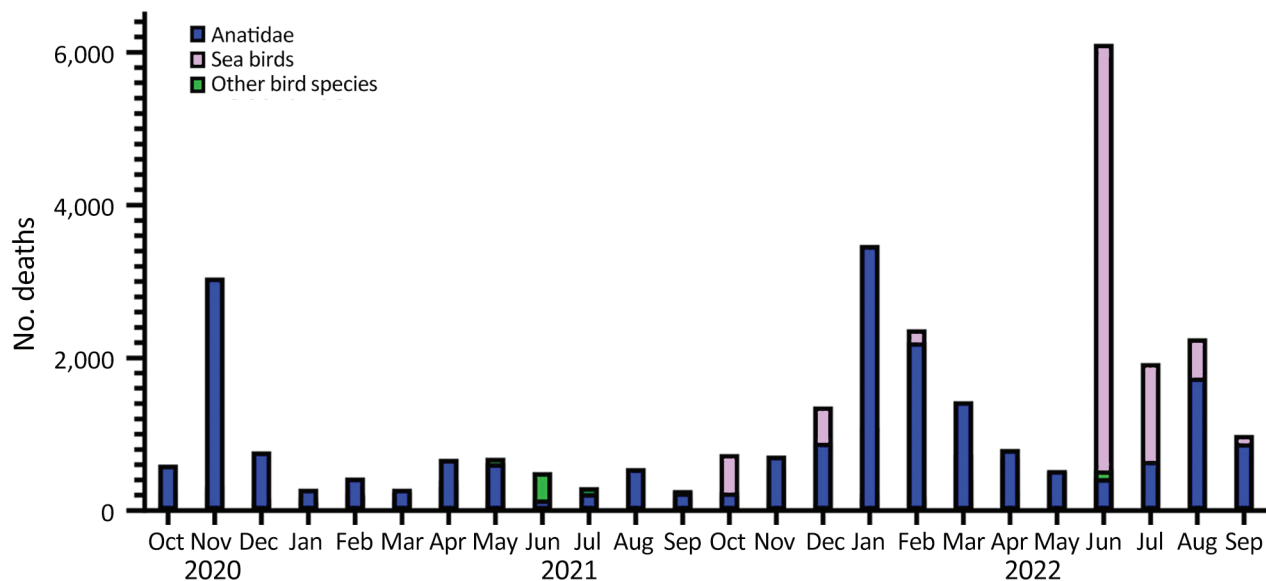


Figure 2. Temporal pattern of wild bird deaths during the 2020–21 and 2021–22 highly pathogenic avian influenza epidemics, the Netherlands. Sea birds include the bird families of Podicipedidae, Laridae, Stercorariidae, Alcidae, Gaviidae, Procellariidae, Sulidae, and Phalacrocoracidae.

(19,20); they are gregarious, herbivorous birds with a preference for coastal grassland and water-rich agricultural fields (21,22). Barnacle geese share their wintering habitat with other herbivorous birds, such as Eurasian wigeons and mute swans (*Cygnus olor*). The large number of geese and their gregarious behavior likely enabled the intraspecies transmission of the virus by direct or indirect contact (e.g., contaminated grass, contaminated water). The abundant circulation of HPAI H5 virus in new host species indicates that the virus has become well adapted to transmission in those species. During the 2020–21 epidemic, HPAI H5 virus was recovered from wild bird populations in the Netherlands for >1 year, indicating that it can be spread and maintained long-term in those populations (4,9), a new observation since the 2016–17 HPAI H5 epidemic, in which viral circulation was mainly limited to autumn and winter (11). A consequence of the unusual persistence of the virus into summer was that naive, newly hatched birds, especially juvenile Anseriformes such as mute swans and graylag geese and raptors such as white-tailed eagle (*Haliaeetus albicilla*), were exposed to the virus and died from infection during spring and summer 2021 (9). The large number of infected wild birds, either ill or dead, was a likely determinant for interspecies viral transmission to hunting or scavenger bird species. Raptors, for example, are exposed to infection by ingesting infected prey (23). During the epidemic, 11 different raptor species were found to be infected, and the highest number of infections and deaths occurred in the

common buzzard (Table; Appendix 1 Table). Among the nonraptor scavenger species, we found 6 different species of gulls (Laridae) and 4 species of corvid (Corvidae) to be infected (Table; Appendix). Because the populations of certain raptor species, such as the peregrine falcon, are relatively small, HPAI may represent a new threat for their conservation. Clinical signs of the infection in wild raptors were mainly neurologic, such as incoordination, body tremors, and torticollis, and were associated with brain lesions and a high level of neurotropism (23).

The 2021–22 HPAI H5 virus epidemic has caused the highest number of casualties among wild birds ever recorded in Europe (4). A significant change in the dynamic of the infections was that, since summer 2021, the virus has been enzootic in wild bird populations in Europe (1,2). This unprecedented, continuous circulation of virus during spring and summer also exposed colonial sea birds to infection (5). During the spring, colonial sea birds congregate in high number at their coastal breeding grounds. In this setting the virus could spread widely within and between breeding colonies, causing outbreaks that resulted in high adult and chick deaths (6,7). Sandwich terns were among the sea bird species that were more severely affected by the HPAI epidemic in the Netherlands. The sandwich tern is a vulnerable, migratory species that only breeds in a limited number of colonies across Europe seasonally, during April–September. Infection-associated mass mortality, with a mortality peak in June, was seen in 9 of the 10 Sandwich tern

breeding colonies in the Netherlands (6). The HPAI-associated mass mortality event is a severe threat for the conservation of this species. Mass dieoffs in the breeding colonies will most likely have long-term repercussions for the Sandwich tern population (6). Constant monitoring of the surviving birds will be necessary to assess the long-term effect of HPAI on this species in the coming years.

The northern gannet is another colonial sea bird species that seasonally occurs off the coast of the Netherlands, although it does not breed in the Netherlands, and that was severely affected by the HPAI epidemic. The HPAI-associated infections started in April and reached a peak in July 2022. We recorded high mortality of breeding gannets on nests; large numbers of carcasses of gannets were sighted afloat near several of the largest or most important breeding colonies and widespread in the North Sea basin (7). The ecology and pattern of mortalities of northern gannets have been studied in the Netherlands since 1980. Data from this long-term study enabled accurate evaluation of the 2022 mortality event in relation to background mortality and corrected per observer effort (24). For the northern gannet the number of reported corpses in July 2022 was 66 times larger than average in previous years, the highest spike in deaths over the past 40 years (24).

During the 2021–22 epidemic, high mortality rates in sea bird species were also reported in other countries in and outside of Europe. For example, Dalmatian pelicans (*Pelecanus crispus*) in Greece and great skuas (*Stercorarius skua*) on Foula, United Kingdom, both had 60%–70% declines of their populations because of HPAI virus infection during colonial breeding (7,8,25). The high density of birds and their close contact during colonial breeding probably enabled the rapid spread of the infection within the colonies. Field data suggest that HPAI-positive birds could shed virus for some period before death, providing opportunities for direct bird-to-bird or environmental transmission (26). Bird species such as great skuas have been reported to bathe and socialize at freshwater lochs and pools, providing possible opportunities for conspecific infection (7). Scavenging activities are another possible source of infection. Unattended chicks from dead parents most likely died because of lack of parental care. Maternal antibodies have been described in chicks of previously infected parents, but clinical protection is short-lived and requires high maternal antibody titers (27,28). Furthermore, maternal antibodies are only relevant if the infection has occurred before egg laying. Infected birds of certain Anatidae species can survive HPAI virus

infection with limited clinical consequences (29,30). Experimentally serially infected ducks can develop a long-term immunity that confers protection from subsequent HPAI virus infection (29). It is possible that sea bird species will also develop flock immunity protective at future reinfection. The surviving birds should be tested for the presence of serum antibodies to gather data about flock immunity over the next several years.

The massive number of dead birds at colonies posed a biosecurity issue through the risk for viral spillover; cleaning up was an overwhelming task for the involved authorities. The AI-Impact consortium, together with the competent health authorities, provided a decision tree for the cleanup of dead birds to reduce the environmental contamination with minimal disturbance for the remaining birds (Appendix 2, <https://wwwnc.cdc.gov/EID/article/30/1/23-0970-App2.pdf>; Appendix 3, <https://wwwnc.cdc.gov/EID/article/30/1/23-0970-App3.pdf>). Carcass removal is necessary to reduce the amount of infected material that could spread the infection in the environment (6). Thus, we recommend controlled studies to optimize carcass removal.

During spring 2021, for the first time since the 2005–06 HPAI H5 epidemic, the virus was detected in Europe in several carnivore species, European foxes (*Vulpes vulpes*), gray seals (*Halichoerus grypus*), and harbor seals (*Phoca vitulina*); they were most likely infected through contact with or ingestion of infected wild birds (9,31). HPAI H5 viruses were once again detected in wild mammal species in Europe during the 2021–22 season and showed genetic markers of adaptation to replication in mammals (16). Therefore, we recommend planned year-round active and passive surveillance of wild mammals. The zoonotic risk for infection for humans of this particular H5 virus strain is considered low for the general population and low to medium for occupationally exposed workers, such as culling operators, wild animal rehabilitators, and workers involved in carcass removal (16). Persons at occupational risk should wear adequate personal protective equipment and be immunized with preventive annual vaccination against human influenza to avoid reassortment with HPAI H5 virus. In case of potential infection, those persons should be monitored for respiratory symptoms, neurologic symptoms, or conjunctivitis for 10 days after exposure (16), and diagnostic testing, if necessary, should be conducted at the competent national health authority.

Since the end of 2016, mass mortality events among wild birds caused by HPAI H5 infection in the

Netherlands have occurred in various species in various years, including Eurasian wigeon (2016), tufted duck (2016), barnacle goose (2020–2022), Sandwich tern (2022), and black-headed gull (*Chroicocephalus ridibundus*) (2023) (16). One characteristic those species have in common is that they live in dense groups at certain times of the year (10) and live close to open water. We suspected that this combination is an important risk factor for infection, because such groups have more opportunities for virus exposure and transmission and for possible species-specific adaptation of the virus (10). However, susceptibility to disease from HPAI virus infection seems to vary enormously between species. For example, disease and death can peak in one species while other species similarly present in the same area show hardly any signs of disease (10).

Because it remains difficult to anticipate and to model the future trends of HPAI among wild birds, we recommend constant monitoring of live and dead wild birds as an essential tool for surveillance of the evolving dynamics of HPAI. This method has several limitations; one is that it is difficult to exclude double-counted reports. Another is that not all the reported dead birds can be tested for HPAI virus infection, and not all will have died from HPAI infection. Two main improvements that we propose for HPAI surveillance in wild birds are long-term monitoring of HPAI-associated wild bird deaths, corrected for observer effort, and testing apparently healthy wild birds, particularly candidate reservoir species, for both HPAI virus and antibodies to HPAI virus. For the constant monitoring of wild bird deaths in the Netherlands during the 2020–21 and 2021–22 HPAI H5 epidemics, citizen scientists were a fundamental resource and made it possible to obtain a wider impression of the actual scale of mortality in wild birds, which otherwise would have been limited to the data from official surveillance. In addition to surveillance for HPAI, we recommend recording of wild bird deaths and encouraging and systematically endorsing the work of citizen scientists and international citizen-science platforms.

Acknowledgments

We thank the members of the AI-Impact group for their contribution.

All data are available in the article and appendices.

This research was financed by Horizon 2020 Framework Programme Deltaflu (grant no. 727922), and in part by the Dutch Ministry of Agriculture, Nature and Food Quality (project no. WOT-01-003-012).

Author contributions: Conceptualization: V.C., E.K., J.R., T.K. Methodology: V.C., E.K., J.R. Investigation: all authors. Visualization: V.C., E.K. Supervision: J.R., T.K. Writing, original draft: V.C. Writing, review and editing: all authors.

About the Author

Dr. Caliendo is a postdoctoral researcher at the Dutch Wildlife Health Centre in Utrecht, the Netherlands. Her primary research interests are wildlife medicine and diseases.

References

1. Pohlmann A, King J, Fusaro A, Zecchin B, Banyard AC, Brown IH, et al. Has epizootic become enzootic? Evidence for a fundamental change in the infection dynamics of highly pathogenic avian influenza in Europe, 2021. *MBio*. 2022;13:e0060922. <https://doi.org/10.1128/mbio.00609-22>
2. King J, Harder T, Globig A, Stacker L, Günther A, Grund C, et al. Highly pathogenic avian influenza virus incursions of subtype H5N8, H5N5, H5N1, H5N4, and H5N3 in Germany during 2020–21. *Virus Evol*. 2022;8:veac035. <https://doi.org/10.1093/ve/veac035>
3. Caliendo V, Lewis NS, Pohlmann A, Baillie SR, Banyard AC, Beer M, et al. Transatlantic spread of highly pathogenic avian influenza H5N1 by wild birds from Europe to North America in 2021. *Sci Rep*. 2022;12:11729. <https://doi.org/10.1038/s41598-022-13447-z>
4. European Food Safety Authority; European Centre for Disease Prevention and Control; European Reference Laboratory for Avian Influenza; Adlhoch C, Fusaro A, Gonzales JL, Kuiken T, Marangon S, Niqueux É, et al. Avian influenza overview June–September 2022. *EFSA Journal* 2022;20:7597. <https://doi.org/10.2903/j.efsa.2022.7597>
5. Wille M, Barr IG. Resurgence of avian influenza virus. *Science*. 2022;376:459–60. <https://doi.org/10.1126/science.abo1232>
6. Rijks JM, Leopold MF, Kühn S, In 't Veld R, Schenk F, Brenninkmeijer A, et al. Mass mortality caused by highly pathogenic influenza A(H5N1) virus in Sandwich terns, the Netherlands, 2022. *Emerg Infect Dis*. 2022;28:2538–42. <https://doi.org/10.3201/eid2812.221292>
7. Camphuysen CJ, Gear SC, Furness RW. Avian influenza leads to mass mortality of adult great skuas in Foula in summer 2022. *Scott. Birds*. 2022;42:312–23.
8. Banyard AC, Lean FZX, Robinson C, Howie F, Tyler G, Nisbet C, et al. Detection of highly pathogenic avian influenza virus H5N1 Clade 2.3.4.4b in great skuas: a species of conservation concern in Great Britain. *Viruses*. 2022;14:212. <https://doi.org/10.3390/v14020212>
9. European Food Safety Authority, European Centre for Disease Prevention and Control, European Reference Laboratory for Avian Influenza; Adlhoch C, Fusaro A, Gonzales JL, Kuiken T, Marangon S, Niqueux E, et al. Avian influenza overview September–December 2021. 2021 Dec 22. <https://doi.org/10.2903/j.efsa.2021.7108>
10. Slaterus R, Schekkerman H, Kleyheeg E, Sierdsema H, Foppen R. Impact of highly pathogenic avian influenza in bird populations in the Netherlands [in Dutch]. Nijmegen (the Netherlands): Sovon Vogelonderzoek Nederland; 2022.

11. Kleyheeg E, Slaterus R, Bodewes R, Rijks JM, Spiereburg MAH, Beerens N, et al. Deaths among wild birds during highly pathogenic avian influenza A(H5N8) virus outbreak, the Netherlands. *Emerg Infect Dis*. 2017;23:2050–4. <https://doi.org/10.3201/eid2312.171086>
12. Camphuysen CJ, Heubeck M. Marine oil pollution and beached bird surveys: the development of a sensitive monitoring instrument. *Environ Pollut*. 2001;112:443–61. [https://doi.org/10.1016/S0269-7491\(00\)00138-X](https://doi.org/10.1016/S0269-7491(00)00138-X)
13. Camphuysen CJ. Beached bird surveys in the Netherlands, autumn 2021 and winter 2021/22. NIOZ Report, RWS Centrale Informatievoorziening BM 22.18. Texel (the Netherlands); Royal Netherlands Institute for Sea Research; 2022.
14. Bollinger TK, Evelsizer DD, Dufour KW, Soos C, Clark RG, Wobeser G, et al. Ecology and management of avian botulism on the Canadian prairies. 2011 [cited 2017 Jun 20]. http://www.phjv.ca/pdf/BotulismReport_FINAL_FullReport_Aug2011.pdf
15. Poen MJ, Venkatesh D, Bestebroer TM, Vuong O, Scheuer RD, Oude Munnink BB, et al. Co-circulation of genetically distinct highly pathogenic avian influenza A clade 2.3.4.4 (H5N6) viruses in wild waterfowl and poultry in Europe and East Asia, 2017–18. *Virus Evol*. 2019;5:vez004. <https://doi.org/10.1093/ve/vez004>
16. Beerens N, Heutink R, Bergervoet SA, Harders F, Bossers A, Koch G. Multiple reassorted viruses as cause of highly pathogenic avian influenza A(H5N8) virus epidemic, the Netherlands, 2016. *Emerg Infect Dis*. 2017;23:1974–81. <https://doi.org/10.3201/eid2312.171062>
17. Adlhoch C, Baldinelli F, Fusaro A, Terregino C. Avian influenza, a new threat to public health in Europe? *Clin Microbiol Infect*. 2022;28:149–51. <https://doi.org/10.1016/j.cmi.2021.11.005>
18. European Food Safety Authority, European Centre for Disease Prevention and Control, European Reference Laboratory for Avian Influenza; Adlhoch, C, Fusaro, A, Gonzales, JL, Kuiken, T, Marangon, S, Stahl, K, et al., 2023. Scientific report: avian influenza overview December 2022–March 2023. *EFSA Journal*. 2023;21:7917. <https://doi.org/10.2903/j.efsa.2023.7917>
19. van der Jeugd HP, Kwak A. Management of a Dutch resident barnacle goose *Branta leucopsis* population: How can results from counts, ringing and hunting bag statistics be reconciled? *Ambio*. 2017;46(Suppl 2):251–61. <https://doi.org/10.1007/s13280-017-0900-3>
20. Voslamber B, van der Jeugd H, Koffijberg K. Numbers, trends, and distribution of breeding goose populations in the Netherlands. *Limosa*. 2010;78:1–17.
21. Koffijberg K, Schekkerman H, van der Jeugd H, Hornman M, van Winden E. Responses of wintering geese to the designation of goose foraging areas in The Netherlands. *Ambio*. 2017;46(Suppl 2):241–50. <https://doi.org/10.1007/s13280-016-0885-3>
22. Si Y, Skidmore A, Wang T, de Boer WF, Toxopeus AG, Schlerf M, et al. Distribution of barnacle geese *Branta leucopsis* in relation to food resources, distance to roosts, and the location of refuges. *Ardea*. 2011;99:217–26. <https://doi.org/10.5253/078.099.0212>
23. Caliendo V, Leijten L, van de Bildt MWG, Fouchier RAM, Rijks JM, Kuiken T. Pathology and virology of natural highly pathogenic avian influenza H5N8 infection in wild common buzzards (*Buteo buteo*). *Sci Rep*. 2022;12:920. <https://doi.org/10.1038/s41598-022-04896-7>
24. Camphuysen KCJ, Kelder L, Zuhorn C, Fouchier R. Avian influenza panzootic leads to mass strandings of northern gannets in the Netherlands, April–October 2022. *Limosa*. 2022;95:4.
25. Alexandrou O, Malakou M, Catsadorakis G. The impact of avian influenza 2022 on Dalmatian pelicans was the worst ever wildlife disaster in Greece. *Oryx*. 2022;56:813. <https://doi.org/10.1017/S0030605322001041>
26. Prosser DJ, Schley HL, Simmons N, Sullivan JD, Homyack J, Weegman M, et al. A lesser scaup (*Aythya affinis*) naturally infected with Eurasian 2.3.4.4 highly pathogenic H5N1 avian influenza virus: movement ecology and host factors. *Transbound Emerg Dis*. 2022;69:e2653–60. <https://doi.org/10.1111/tbed.14614>
27. Christie KF, Poulson RL, Seixas JS, Hernandez SM. Avian influenza virus status and maternal antibodies in nestling white ibis (*Eudocimus albus*). *Microorganisms*. 2021;9:2468. <https://doi.org/10.3390/microorganisms9122468>
28. Maas R, Rosema S, van Zoelen D, Venema S. Maternal immunity against avian influenza H5N1 in chickens: limited protection and interference with vaccine efficacy. *Avian Pathol*. 2011;40:87–92. <https://doi.org/10.1080/03079457.2010.541226>
29. Caliendo V, Leijten L, van de Bildt MWG, Poen MJ, Kok A, Bestebroer T, et al. Long-term protective effect of serial infections with H5N8 highly pathogenic avian influenza virus in wild ducks. *J Virol*. 2022;96:e0123322. <https://doi.org/10.1128/jvi.01233-22>
30. Gobbo F, Fornasiero D, De Marco MA, Zecchin B, Mulatti P, Delogu M, et al. Active surveillance for highly pathogenic avian influenza viruses in wintering waterbirds in northeast Italy, 2020–2021. *Microorganisms*. 2021;9:2188. <https://doi.org/10.3390/microorganisms9112188>
31. Rijks JM, Hesselink H, Lollinga P, Wesselman R, Prins P, Weesendorp E, et al. Highly pathogenic avian influenza A(H5N1) virus in wild red foxes, the Netherlands, 2021. *Emerg Infect Dis*. 2021;27:2960–2. <https://doi.org/10.3201/eid2711.211281>

Address for correspondence: Valentina Caliendo, Dutch Wildlife Health Centre, Faculteit Diergeneeskunde, Universiteit Utrecht, Yalelaan 1, 3584 CL, Utrecht, the Netherlands; email: v.caliendo@uu.nl

Effect of 2020–21 and 2021–22 Highly Pathogenic Avian Influenza H5 Epidemics on Wild Birds, the Netherlands

Appendix 2



ADVICE FOR CLEARING WILD BIRD CARCASSES IN OUTDOOR AREAS DURING THE HIGH PATHOGENIC BIRD FLU OUTBREAK H5N1 2020–2021

Working group Almpact2021, 13 November 2020

Reason for advice

The first step in finding dead wild birds is reporting through the appropriate official channels: NVWA (045–546 31 88) on >3 ducks, geese, swans in (more or less) the same location, or >20 birds from other bird species in (more or less) the same location; in other cases Sovon (www.sovon.nl/nl/content/vogel-en-zoogdierdoelen) and DWHC (www.dwhc.nl/meldsvorm/). The reports together give these organizations a picture of the course of the outbreak.

In addition, the NVWA and DWHC can decide whether it is necessary to collect carcasses for bird flu diagnosis by WBVR in Lelystad. This keeps people informed of the locations and sources of infection of bird flu.

Wild bird carcasses are cleaned up in populated areas and in outlying areas where a lot of public comes, for reasons of hygiene and the risk of the virus spreading. However, it is not so clear to site managers what the best course of action is to deal with wild bird carcasses in remote

areas where there is little public access. This advice is about the latter: the choice for or against clearing wild bird carcasses in rural areas with little public.

Relevant information about highly pathogenic bird flu virus

- In general, wild birds that live in or around water (waterfowl in an ecologic sense) can be infected with avian influenza virus (AIV) through direct contact with other infected birds, or through indirect contact with environmental material (eg water, mud, grass) that is contaminated with contaminated excrement (eg faeces, snot) (1). Waterfowl that have often been found dead in recent years with highly pathogenic avian influenza virus (HPAIV) are ducks, geese, swans, grebes, storks, herons, cormorants, coots, waders, and gulls (2).

- Some bird species can become infected by eating the muscles and organs of other birds with HPAIV infection, either because they prey on infected birds or because they eat carcasses of dead, infected birds (3–5). Birds of prey and scavengers that have often been found dead with HPAIV infection in recent years include white-tailed eagle, rough-legged hawk, buzzard, peregrine falcon, hawk, short-eared owl, great black-backed gull, herring gull and magpie (2) (www.oie.int).

- The full list of 68 bird species in which HPAIV has been detected during the major HPAIV outbreak of 2016/2017 can be found in Table A.3 of the relevant EFSA report (6). Only virus positive dead birds identified by species are included.

- AIV in surface waters of the wetlands of Alaska and Minnesota remained infectious to birds for more than 7 months during the winter months (September to April) (7).

- In chicken carcasses experimentally infected with HPAIV, the maximum survival times for infectious virus under laboratory conditions depended on temperature. At +4°C maximum survival was 8 months in feathers, 5 months in muscle and 3 weeks in liver. At +20°C, maximum survival was 1 month in feathers, 3 weeks in muscle and 3 days in liver (8).

- RIVM estimates that there is a low risk of people becoming infected with the HPAIV strains that have been found in wild birds and poultry farming in recent

weeks (<https://www.rijksoverheid.nl/documents/kamerstukken/2020/11/10/room-letter-update-bird-flu-3rd-infection-and-low-zoonosis-risk>).

Considerations for Disposing of Wild Bird Carcasses in Outlying Areas

- Carcasses of dead wild birds infected with HPAIV are a source of virus contamination for their environment. As a result, birds that rest or forage in that environment run an extra risk of becoming infected.
- Birds of prey and carrion-eating birds that eat HPAIV-infected carcasses are at extra risk of becoming infected.

Choice for cleaning up

- Outlying area is regularly used by raptors and carrion-eating bird species.
- Carcasses are located in outlying areas where birds rest or forage in high density (eg high water refuge, wetland area, pasture, puddle, lake).

Choice against cleaning up

- Carcasses are located in outlying areas where birds of prey or carrion-eating bird species are not or sporadically present (unsuitable habitat) or cannot easily reach (e.g. undergrowth).
- Carcasses are located in rural areas where waterfowl occur sporadically.
- There is no clear indication of carcasses, while entering the area would disturb large numbers of wild birds.

How many carcasses to clean up?

There is no specific number to indicate here. It depends on the location and the circumstances. The best advice is to clean up when there are more carcasses than normal and when they are found in situations such as those mentioned under 'Clearance choice'.

How often do you clean up carcasses?

No specific frequency can be specified here either. It depends on the course of the outbreak, and specifically the number of birds that die per day. This can best be determined by regularly (preferably daily) checking the area from a distance, and letting the frequency of clearing depend on the degree of bird death.

How to clean up?

- Enter the area with the minimum number of people needed to clean up quickly and efficiently, with as little disturbance as possible to the birds present, as this could be counterproductive. Clear high water refuges during low water to limit disturbance.
- Combine carcass removal with registration of bird deaths in the area (number per species via Sovon or DWHC Web sites. Waarneming.nl can also be used by people who are familiar with it. Preferably keep the person in charge of the registration 'clean', i.e., he refrains from direct contact with dead birds. Please report any rings present via www.vogeltrekstation.nl
- Coordinate entry into the area with an ornithologist or area manager known locally with bird knowledge (also for the registration of numbers, species and rings).
- Wear adequate personal protection, and adequate disinfection afterwards (including footwear!) to prevent contamination of other areas.
- Transport carcasses in double plastic bags to the appropriate place, to be transported to a rendering company.
- For specific details about personal protection and other hygiene measures, see: [https://www.nvwa.nl/onderwerpen/vogelgriep-preventiveness-en-control/documents/dier/dierspreiden/vogelgriep/protocols/handbook-voor-het-opruiming -found-dead-wild-waterfowl](https://www.nvwa.nl/onderwerpen/vogelgriep-preventiveness-en-control/documents/dier/dierspreiden/vogelgriep/protocols/handbook-voor-het-opruiming-found-dead-wild-waterfowl).

Composition of the Almpact2021 Working Group (13 November 2020)

Valentina Caliendo (Erasmus MC), Kees Camphuysen (NIOZ), Armin Elbers (WBVR), Koos Hartnack (RWS), Leon Kelder (SBB), Erik Kleyheeg (Sovon), Thijs Kuiken (Erasmus MC, chairman), Mardik Leopold (Wageningen Marine Research), Jolianne Rijks (DWHC), Timo Roeke (waarneming.nl), Roy Slaterus (Sovon), Marcel Spierenburg (NVWA), Henk van der Jeugd (NIOO), Peter van Tulden (WBVR), Hans Verdaat (Wageningen Marine Research).

References

1. Olsen B, Munster VJ, Wallensten A, Waldenström J, Osterhaus AD, Fouchier RA. Global patterns of influenza A virus in wild birds. *Science*. 2006;312:384–8. [PubMed](#)
<https://doi.org/10.1126/science.1122438>
2. Brouwer A, Gonzales J, Huneau A, Mulatti P, Kuiken T, Staubach C, et al.; European Food Safety Authority (EFSA). Annual Report on surveillance for avian influenza in poultry and wild birds in Member States of the European Union in 2018. *EFSA J*. 2019;17:e05945. [PubMed](#)
3. Brown JD, Stallknecht DE, Swayne DE. Experimental infections of herring gulls (*Larus argentatus*) with H5N1 highly pathogenic avian influenza viruses by intranasal inoculation of virus and ingestion of virus-infected chicken meat. *Avian Pathol*. 2008;37:393–7. [PubMed](#)
<https://doi.org/10.1080/03079450802216595>
4. van den Brand JM, Krone O, Wolf PU, van de Bildt MW, van Amerongen G, Osterhaus AD, et al. Host-specific exposure and fatal neurologic disease in wild raptors from highly pathogenic avian influenza virus H5N1 during the 2006 outbreak in Germany. *Vet Res (Faisalabad)*. 2015;46:24. [PubMed](#) <https://doi.org/10.1186/s13567-015-0148-5>
5. Krone O, Globig A, Ulrich R, Harder T, Schinköthe J, Herrmann C, et al. White-Tailed Sea Eagle (*Haliaeetus albicilla*) Die-Off Due to Infection with Highly Pathogenic Avian Influenza Virus, Subtype H5N8, in Germany. *Viruses*. 2018;10:478. [PubMed](#) <https://doi.org/10.3390/v10090478>
6. Brown I, Mulatti P, Smietanka K, Staubach C, Willeberg P, Adlhoch C, et al.; European Food Safety Authority; European Centre for Disease Prevention and Control; European Union Reference Laboratory for Avian influenza. Avian influenza overview October 2016-August 2017. *EFSA J*. 2017;15:e05018. [PubMed](#)
7. Ramey AM, Reeves AB, Drexler JZ, Ackerman JT, De La Cruz S, Lang AS, et al. Influenza A viruses remain infectious for more than seven months in northern wetlands of North America. *Proc Biol Sci*. 2020;287:20201680. [PubMed](#) <https://doi.org/10.1098/rspb.2020.1680>
8. Yamamoto Y, Nakamura K, Mase M. Survival of Highly Pathogenic Avian Influenza H5N1 Virus in Tissues Derived from Experimentally Infected Chickens. *Appl Environ Microbiol*. 2017;83:e00604–17. [PubMed](#) <https://doi.org/10.1128/AEM.00604-17>

Effect of 2020–21 and 2021–22 Highly Pathogenic Avian Influenza H5 Epidemics on Wild Birds, the Netherlands

Appendix 3

ADVICE FOR REGISTRATION AND DISPOSAL OF WILD BIRD CARCASSES IN BREEDING COLONIES DURING HIGHLY PATHOGENIC AVIAN INFLUENZA OUTBREAK H5NX, 2022

Working group AI-Impact 21/22, 29 June 2022

In 2022, infection with highly pathogenic avian influenza virus H5 of the Goose/Guangdong lineage (further referred to as HPAI) more often has led to extensive mortality in breeding colonies of wild birds, including sandwich and common terns in the Netherlands, and northern gannets, great skuas and common guillemots in the UK. For this reason, we have written a protocol specifically aimed at HPAI in breeding colonies. It is important to realize that while all listed birds are ‘colonial’, breeding conditions differ from high densities on sand flats (e.g., terns), via relatively low densities (territories) on hill slopes (e.g., skuas), to high densities on steep cliffs or concentrations on flat tops of remote islands (e.g., gannets and auks).

The general principle is to leave wild birds with HPAI alone, so that the virus is not spread to other areas via disturbed birds. Two important reasons to visit a breeding colony where HPAI occurs are: 1, to limit virus spread in the breeding colony by removing carcasses from sites where infections may occur (colony itself, or bathing places for example); 2, to record scientific information about the outbreak, and 3. by removing carcasses from sites where they pose a risk for the general public or livestock. Below we give more details about the first two reasons.

(1) Limiting virus spread in the breeding colony by removing carcasses

Carcasses in a breeding colony can be a source of infection for other birds in the colony. A bird can be infected by touching the carcass (e.g., out of curiosity, seen in juvenile cormorants and pelicans; mating behavior, seen in Sandwich terns), by eating at the carcass (seen in great skuas), or by drinking water or bathing in water containing carcasses (seen in cormorants and great skuas). Avian influenza virus in carcasses and surface water can remain contagious for days to months, depending on environmental factors including the ambient temperature.

(2) Recording information about a HPAI outbreak in a breeding colony

In the current surveillance systems for HPAI in wild birds, the detection of HPAI virus in the event of mortality of wild birds is recorded, but not the number of wild birds found dead. Well-documented descriptions of HPAI outbreaks in wild birds are therefore important to better assess the impact of this disease on wild bird populations, and to help policymakers in making decisions to prevent future HPAI outbreaks, not only in poultry and humans, but also in wild birds.

Decision to visit a breeding colony

The final decision whether to visit a breeding colony should be taken depending on the local situation and in consultation with people with good knowledge of the breeding colony, and taking into consideration the following advantages and disadvantages:

Advantages

- removal of carcasses that may remain contagious for days to months
- more detailed recording of the outbreak than possible from a distance.

Disadvantages

- disturbance of birds, with a chance of virus being spread to other areas and/or chance that birds abandon the breeding colony
- (very small) risk of infection of people who visit the breeding colony.

Relevant information and samples to collect during a visit

To determine the extent of the outbreak, record:

- bird species
- age (external plumage characteristics)
- sex (external plumage characteristics)
- any ring numbers
- photos of sick and dead birds as supporting evidence.

To substantiate the cause of the outbreak:

- describe and/or record on film the clinical signs in affected birds.

Hygiene measures

The following hygiene measures are relevant if you choose to visit a breeding colony where HPAI occurs:

- Wear adequate personal protection, and perform adequate disinfection afterwards (including footwear!) to prevent contamination of other areas.
- Suitable disinfectants for HPAI virus include agents based on ethanol (e.g., Sterillium) or sodium hypochlorite (e.g., Clorox).
- Transport carcasses in double plastic bags to the place designated by the relevant authorities for further disposal
- For specific details on personal protection and other hygiene measures, see Web sites of the relevant authorities.

Composition of the AI Impact Working Group 21/22 (29 June 2022)

Nancy Beerens (WBVR), Valentina Caliendo (Erasmus MC), Kees Camphuysen (NIOZ), Rommert Cazemier (Wetterskip Fryslân), Armin Elbers (WBVR), Ron Fouchier (Erasmus MC), Jan Jelle Jongsma (Fryske Gea), Fedde Jonkman (Zuiderzeeland), Leon Kelder (SBB), Erik Kleyheeg (Sovon), Thijs Kuiken (Erasmus MC, chairman), Mardik Leopold (Wageningen Marine Research), Jolianne Rijks (DWHC), Ramon ter Schegget (RWS), Roy Slaterus (Sovon), Marcel Spiereburg (NVWA), Henk van der Jeugd (NIOO), Peter van Tulden (WBVR), Hans Verdaat (Wageningen Marine Research).