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Unraveling the prevalence of soil-borne fungal pathogens in the North China Plain: a global analysis approach

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Abstract Soil-borne fungal plant pathogens lead to worldwide economic yield losses. However, despite the agricultural importance of the North China Plain (NCP), little is known about the occurrence and severity of soil-borne fungal pathogens that could potentially affect the yields of three main crops in this area: wheat, maize and soybean. By combining searches in public databases, we identified a set of main soil-borne fungal pathogen species potentially affecting crop production in the NCP. We investigated their distribution patterns at three scales: globally, in China and in the NCP, and evaluated how these patterns were determined by a range of variables related to climate, spatial drivers and crop area density of the three crop species. We found 25 main soil-borne fungal pathogens that could potentially affect the yields

of wheat, maize and soybean in the NCP. Twenty of these fungal pathogens were prevalent across the globe, and 13 were very frequent in the NCP. Significant positive relationships between pathogen prevalence and crop area density were only found for two specialist pathogens. Our work provides an overview of the main soil-borne fungal pathogens in one of China's most intensively cultivated regions, shedding light on potential, previously overlooked, fungal threats for agricultural production in the area. This baseline information may contribute to the development of sustainable management strategies aimed at mitigating disease outbreaks caused by soil-borne pathogens in the future.

Keywords Wheat · Maize · Soybean · Plant pathogen distribution · Host availability · Environmental drivers

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Introduction

Many plant pathogens cause serious impacts on crops globally, leading to major yield losses (Savary et al., 2019; Ristaino et al., 2021). This includes soil-borne fungal pathogens, many of which have global distributions (Scherin et al., 2013; Termorshuizen, 2014; De Coninck et al., 2015; Delgado-Baquerizo et al., 2020; Al-Sadi, 2021; Marquez et al., 2021) and can lead to economically important losses in agriculture worldwide (De Coninck et al., 2015). Examples of globally important soil-borne pathogens are the fungus *Fusarium graminearum*, the causal agent of Fusarium head blight of cereals (Summerell et al., 2010; Chen et al., 2019a; Karlsson et al., 2021), reducing yields of crops such as wheat and maize (Savary et al., 2019); and *Rhizoctonia solani*, which causes root rot in many crops including wheat, maize, soybean and rice (Ghosh et al., 2017; Ajayi-Oyetunde & Bradley, 2018; Lin et al., 2021).

The North China Plain (NCP) is an important and intensively managed agricultural production area in China (National Bureau of Statistics of China, 2021). The main crops cultivated in the NCP are wheat, maize and soybean, altogether accounting for 63% of the total planted area (National Bureau of Statistics of China, 2021). The highly intense agricultural production in the region is achieved through the profuse use of irrigation, fertilizers and pesticides (Miao et al., 2011; Sun et al., 2011; Brauns et al., 2018; Wang et al., 2019; Liu et al., 2020; Mu et al., 2022), all of which are likely to favour the incidence of soil-borne diseases. For instance, the enrichment of nitrogen in soils caused by overuse of fertilizers has been shown to favour the accumulation of soil-borne pathogens in agro-systems (Walters & Bingham, 2007; Paungfoo-Lonhienne et al., 2015; Wei et al., 2018), and the widespread use of fungicides may induce the plant pathogens' tolerance to them (Mikaberidze et al., 2017; Corkley et al., 2022). *Fusarium* species have been isolated from diseased wheat stems in randomly selected wheat fields of the NCP (Xu et al., 2018; Zhang et al., 2015, 2020), and also isolated in maize (Meng, 2019) and soybean (Yang, 2012) fields in this region. Moreover, these soil-borne diseases have been reported to reduce yields of these main crops in the NCP (Fu, 2017; Zhang et al., 2020). However, apart from these well-known pathogens, other soil-borne fungal pathogens in the region remain understudied.

Numerous studies conducted in the NCP have primarily focused on documenting the symptoms associated with soil-borne pathogens (Xue, 2006; Zhu, 2012; Zhang et al., 2012; Zhao et al., 2015; Ji & Han, 2016; Li, 2018; Xin & Yu, 2021; Zhou et al., 2021), and many are published in Chinese journals with limited scope (CNKI, <https://www.cnki.net>). However, there is a dearth of information regarding the specific causal agents responsible for these symptoms. Furthermore, although a few studies have employed cultivation techniques to identify certain pathogens in the NCP (Wang, 2019; Liu et al., 2021), these investigations have been limited to specific locations. To manage the soil-borne diseases effectively and promote sustainable agricultural development in the NCP, it is crucial to unravel the prevalence of pathogen species that pose a potential threat to crop production.

Characteristic agricultural areas such as the NCP may have specific assemblages of fungal pathogens owing to the peculiarities of the dominant production systems—including the main crop species and management practices—in addition to the general environmental factors that govern the distribution of all fungi. In particular, the density of host crop species can be a crucial factor in recruiting and multiplying crop-specific pathogens (Zhu et al., 2000; Hiddink et al., 2010), with monocultures tending to accumulate specific pathogens in soils, as compared to more diverse plant communities where the presence of such pathogens becomes diluted (Zhu et al., 2000; Hiddink et al., 2010; Boudreau, 2013; Ampt et al., 2018; Mommer et al., 2018). For instance, monocultures of wheat, maize and soybean have been shown to accumulate high abundances of soil-borne fungal pathogens specialized in attacking each crop, such as *Gaeumannomyces graminis* in wheat (Ramanauskienė et al., 2018; Gargouri et al., 2020), *Stenocarpella maydis* in maize (Flett & Wehner, 1991; Pinto et al., 2022), and *Macrophomina phaseolina* in soybean (Pérez-Brandán et al., 2014). On the other hand, the intensive use of fungicides (Price et al., 2015; Ons et al., 2020; Mu et al., 2022), as well as other management practices associated with the main crops in the area (Li et al., 2018; Wang et al., 2019; Chen et al., 2019b; Liu et al., 2019; Sun et al., 2020), could have effects on the regional soil-borne

fungal communities that are difficult to predict and remain understudied.

Irrespective of the effects regional agricultural production systems may have on pathogen distribution, fungal occurrence will also be affected by major environmental factors that may confound the effects of cropping systems. Different regions of the globe have characteristic assemblages of fungi, including soil-borne pathogens (Vujanovic et al., 2006; Teder-soo et al., 2014, 2022), due to variation in major environmental drivers that operate at different spatial scales, including factors such as climatic conditions (Teder-soo et al., 2014; Větrovský et al., 2019; Delgado-Baquerizo et al., 2020) or large-scale changes in soil properties (Teder-soo et al., 2014, 2022). In addition to these well-defined variables, other spatial drivers of fungal distribution comprise variables that are difficult to quantify, such as the differential dispersal abilities of fungal species (e.g. whether they produce propagules enabling long-range or very restricted dispersal, or are likely affected by geographic barriers; Peay et al., 2010; Wardle & Lindahl, 2014; Bever et al., 2015; Chaudhary et al., 2022), and historical factors such as human-driven transportation of crop seeds across landscapes or changes in land use (Desprez-Loustau et al., 2010). These spatial drivers that lead to differences in the pathogens' distribution due to different dispersal ability should be accounted for. Examples of environmental patterns in soil-borne pathogen distribution include those of *Fusarium* spp. in asparagus fields in eastern Canada, determined by temperature and soil properties (Vujanovic et al., 2006); the restricted distribution of *Colletotrichum kahawae* attributed to geographic barriers (Silva et al., 2012); or the relatively well-documented spread of *Verticillium dahliae* due to anthropogenic movement (Short et al., 2015).

Here, to understand the potential threats of pathogens occurrence in the NCP, we assessed the distribution of soil-borne fungal pathogens on crops that are dominant in the NCP—maize, wheat, soybean—at three levels: global, China and the NCP. We studied the contribution of climatic, other spatial factors (e.g. related to the dispersal ability of fungi or geographic barriers), and crop area density to explain prevalence at these fungi at different scales. In particular, our specific aims were to 1) identify the major soil-borne fungal pathogens that could potentially affect maize, wheat and soybean in the NCP; 2) evaluate

the prevalence of these pathogens in the NCP as compared to their distribution at increasingly larger scales (i.e. within China and globally); and 3) identify the main predictors influencing the distribution of these pathogens, with a particular emphasis on the availability of the specific host crop species (evaluated through regional crop area density).

Methods

Selection of focal pathogens

We first compiled a list of potential pathogenic fungal species with relevance for the production of wheat, maize or soybean based on the following criteria: (1) they have been reported in the literature as causal agents of disease in at least one of these crop species, (2) they are soil-borne (i.e. transmitted via soil), and (3) the diseases that they cause have been reported to cause agricultural yield losses in the NCP.

We used two approaches to select potential pathogens fulfilling those criteria. First, we identified important pathogens worldwide for the three focal crops by searching in the 'Fungal-Host Distributions' database of the US Department of Agriculture (USDA; Farr & Rossman 2021) in December 2021, using "*Triticum aestivum*" (wheat), "*Zea mays*" (maize) and "*Glycine max*" (soybean) as search terms in the 'Host Name' field. We retained for further analysis only those pathogens with relatively widespread global distributions, i.e. detected in at least 20 countries.

In the second approach, we looked for specific literature reporting pathogens of the focal crops in the NCP in December 2021, by searching in the China National Knowledge Infrastructure database (CNKI, <https://www.cnki.net>) and in Web of Science (WoS, <https://www.webofscience.com/wos/woscc/basic-search>). We used the query string "(('wheat' OR 'maize' OR 'soybean') AND 'disease' AND 'China')". We reviewed these publications and retained those reporting pathogens in the NCP. In a second round of literature searches, we queried the CNKI database using the string "pathogen species" AND "yield losses", and retained the publications related to the yield losses in the NCP caused by those pathogens.

We combined the information obtained from the two approaches described above to compile a final list of soil-borne fungal pathogen species of potential

relevance for the main crops produced in the NCP (Table 1). To determine the phylogenetic relationships among the selected pathogens, we built a Maximum Likelihood phylogenetic tree based on representative sequences retrieved from the NCBI GenBank database (<http://www.ncbi.nlm.nih.gov>; Table S1) using the software MEGA7 v7.0.26 (Kumar et al., 2016).

Retrieval of pathogen distribution, climatic, spatial and host availability data

We retrieved data on the global distribution of the selected pathogen species from the GlobalFungi database (<https://globalfungi.com>; Větrovský et al., 2020), which compiles high-throughput ITS (internal transcribed spacer) amplicon sequencing data from studies conducted across the globe and published between

2009 and 2022. These include quantitative datasets on the worldwide distribution of fungal taxa based on molecular sequencing of DNA barcode regions from environmental samples, enabling the characterization of whole fungal communities with a certain degree of taxonomic precision (Nilsson et al., 2019). By compiling these amplicon sequencing data, the database enables tracking of fungal taxa worldwide in environmental samples, such as soil or plant tissues (Bálint et al., 2016; Nilsson et al., 2019). We searched for each species in the GlobalFungi database and downloaded their global occurrence data and associated metadata, including geographical co-ordinates, biome (e.g. originating from cropland or other natural habitats), climatic variables (mean annual temperature, MAT; mean annual precipitation, MAP), and relative abundance per study in December 2021.

Table 1 List of target soil-borne fungal pathogens and their host specificity (i.e. wheat, maize, and soybean), based on database or literature searches

Pathogen	Class	Wheat	Maize	Soybean	Sources ¹
<i>Fusarium asiaticum</i>	Sordariomycetes	✓			Zhang et al., 2012; Qi 2015; Gao 2016
<i>Fusarium culmorum</i>	Sordariomycetes	✓			USDA; Ren 2011; Zhou et al., 2019
<i>Fusarium graminearum</i>	Sordariomycetes	✓	✓	✓	USDA; Sun et al., 2014; Zhou 2014
<i>Fusarium oxysporum</i>	Sordariomycetes	✓	✓	✓	USDA; Zhang 2012; He 2016
<i>Fusarium proliferatum</i>	Sordariomycetes	✓	✓	✓	Yang, 2012; Sun et al., 2014; Wang, 2019
<i>Fusarium fujikuroi</i>	Sordariomycetes		✓		Shan 2017
<i>Fusarium solani</i>	Sordariomycetes			✓	USDA; Yang, 2012; Wang, 2019
<i>Fusarium avenaceum</i>	Sordariomycetes	✓		✓	USDA; Zhang 2012
<i>Colletotrichum truncatum</i>	Sordariomycetes			✓	USDA
<i>Phomopsis longicolla</i>	Sordariomycetes			✓	Wang, 2019; Ye et al., 2020
<i>Macrophomina phaseolina</i>	Dothideomycetes			✓	USDA; Zhang et al., 2009; Wang, 2019
<i>Zyloseptoria tritici</i>	Dothideomycetes	✓			USDA
<i>Cercospora kikuchii</i>	Dothideomycetes			✓	USDA
<i>Cercospora sojina</i>	Dothideomycetes			✓	USDA
<i>Exserohilum turcicum</i>	Dothideomycetes		✓		USDA
<i>Pyrenophora tritici-repentis</i>	Dothideomycetes	✓			USDA
<i>Curvularia lunata</i>	Dothideomycetes		✓		USDA
<i>Bipolaris zeicola</i>	Dothideomycetes		✓		USDA
<i>Bipolaris sorokiniana</i>	Dothideomycetes	✓			USDA; Zhou 2014; Xu et al., 2018; Kang et al., 2020
<i>Bipolaris maydis</i>	Dothideomycetes		✓		USDA
<i>Physoderma maydis</i>	Blastocladiomycetes		✓		USDA
<i>Rhizoctonia solani</i>	Basidiomycetes		✓	✓	USDA; Ye et al., 2020
<i>Urocystis tritici</i> *	Ustilaginomycetes	✓	✓		USDA
<i>Sporisorium reilianum</i> *	Microbotryomycetes		✓		USDA; Ma 2006
<i>Ustilago maydis</i> *	Ustilaginomycetes		✓		USDA

¹Only a selection of references is shown. The full list is provided in Table S2. *All species are necrotrophs, except the last three, which are biotrophs

We obtained data about agricultural production of the three focal crop species per country, including crop area density (cultivated area of crops) for each target crop, land area and cropland area from FAOSTAT (<https://www.fao.org/faostat/en/#data>; FAOSTAT, 2022). In January 2022, we downloaded the data of host variables from the ‘Crops and livestock products’ database of the FAOSTAT, using “all countries” in the countries field; “Area harvested” in the elements field; “Wheat”, “Maize” and “Soybean” in the items field; and “from 2000 to 2020” in the Years field. We also downloaded the data of land area and cropland area in the ‘Land Use’ database of the FAOSTAT, using “all countries” in the countries field; “Area” in the elements field; “Land area” and “Cropland” in the items field; and “from 2000 to 2020” in the years field.

We mainly targeted the effects of climatic, spatial and host availability variables on soil-borne fungal pathogen distributions, but we also accounted for additional variables that may affect pathogen distribution. We controlled for 1) differences in sampling effort across biomes (i.e. leading to a higher probability of detecting the focal fungi in countries with more studies at cropland biome) by calculating the proportion of cropland studies as the number of studies at cropland biome divided by the total number of studies in a given country; 2) for the country area (i.e. higher chances to detect the fungi in larger countries) by calculating the proportion of cropland area per country with respect to the total land area of each country; and 3) for other spatially-structured, unmeasured sources of variation that could explain patterns in the dataset. We accounted for the latter spatial factors using Principal Co-ordinates of Neighbor Matrices (PCNM; Dray et al., 2006) using package *vegan* v2.6–2 (Oksanen et al., 2020), which summarize spatial patterns of variation at different scales, that could represent factors either intrinsic to the organisms (e.g. dispersal), geographic (e.g. geographic barriers), or environmental (e.g. climatic variables other than those explicitly considered; Dray et al., 2006; Griffith & Peres-Neto, 2006; Peres-Neto & Legendre, 2010).

Analysis of pathogen distributions

All analyses of pathogen distributions were conducted in R v4.1.2 (R Core Team, 2021), using the package *rnatlearn* v0.1.0 (South, 2017) to collect additional geographical information on the samples’

origin, such as the country (for globe and China) and province (for the NCP), and the centroid co-ordinates per country or province. In each case, the relative pathogen abundance per study was calculated as the number of reads for the target fungus divided by the total number of reads per study. However, since this metric is prone to bias due to methodological differences across studies, we mainly used prevalence data, defined as the proportion of studies per country/province where the target pathogen was detected. For this, the number of studies in each country where the pathogen was detected was divided by the total number of studies in that area. Thus, the higher the prevalence of the pathogen at a given scale (e.g. global, China, the NCP), the higher the probability of detecting that pathogen at that scale.

We identified pathogens with a particularly high representation in the NCP compared to the rest of China or the globe by comparing their normalized prevalence across the three areas. We defined normalized prevalence as the ratio of prevalence per pathogen with respect to the maximum prevalence per scale (i.e. ranging from zero to one, respectively indicating no or maximum prevalence in the area). We considered pathogens to be overrepresented in the NCP with respect to the rest of China or the world when the normalized prevalence observed at NCP was at least three times larger than in the other areas. We arbitrarily selected this cut-off value based on the distribution of the prevalence data, because it yielded a conservative selection of pathogens with the highest representation at NCP.

Statistical analysis

All statistical analyses were performed with R. To test if the prevalence of each pathogen followed a proportional distribution across the three scales considered (global, China and NCP), between-group significant differences in pathogen prevalence were tested using pairwise Pearson’s Chi-squared (R package *fifer* v1.2; Fife & Fife, 2017). We used linear models to estimate the effects of climatic, spatial and host availability variables on pathogen distributions, and the proportion of cropland area and the proportion of cropland studies were considered as covariates. The climatic predictors used were retrieved from the GlobalFungi dataset, including mean annual temperature (MAT) and mean annual precipitation (MAP) averaged per

country/province. The spatial factors captured were those derived from PCNM variables based on the samples in the dataset (see above). For host availability, the mean annual harvest area (crop area density) of each target crop per country retrieved from FAOSTAT was considered. Firstly, for each selected pathogen, stepwise model selections using package MASS v7.3–54 (Venables & Ripley, 2013) were first performed to select only the PCNM variables that are significantly associated with pathogen prevalence, to avoid inflation of models caused by inclusion of too many PCNM variables (Gilbert & Bennett, 2010). Next, linear models were fitted to estimate the effects of climatic, the selected spatial, host availability and covariates variable on pathogen prevalence data. The analysis pipeline in this study can be found in Fig. S1

Results and discussion

Description of the datasets

The USDA database searches resulted in 819 records of pathogen species for wheat, 989 for maize and 597 for soybean, globally. From these, we selected 12 widespread pathogen species for wheat, 18 for maize and 10 for soybean for subsequent analyses (Table S2).

The two literature searches (one in WoS and one in CNKI) initially resulted in 100 publications describing pathogens of wheat in the NCP, 174 of maize, and 136 of soybean. After further analysis of the literature records, we identified 20 publications on wheat, 5 on maize and 6 on soybean from which we recorded the pathogen species and related information concerning the detection method (either isolation or direct sequencing), the host, the type of disease and the relative abundance. Finally, 23 publications related to yield loss caused by those pathogens in the NCP were retrieved from the second search in WoS and CNKI (Table S2). The references in this dataset were also listed in Table S3.

A final list of 25 target soil-borne fungal pathogens representing six taxonomic classes, and their host specificity towards each of the focal crop species, was compiled by collating the results from the above approaches (Table 1). Seven soil-borne fungal pathogens affecting yield were found for wheat, nine for maize and six for soybean (Table 1). Three out

of 25 target pathogens were biotrophs, i.e. *Urocystis tritici*, *Sporisorium reilianum* and *Ustilago maydis* (Table 1).

Distribution of selected soil-borne fungal pathogens at global and regional scales

To investigate the distribution patterns of the 25 selected soil-borne pathogen species, we retrieved data from the GlobalFungi database representing 36,684 records of sampling sites across 367 studies (Table S4). The distribution of 20 pathogen species showed widespread occurrences, in most cases spanning the globe (more than three continents; Fig. 1a, b, S2), while five species (*Phomopsis longicolla*, *Bipolaris maydis*, *Physoderma maydis*, *Sporisorium reilianum* and *Ustilago maydis*) were concentrated in China and the NCP (Fig. 1d, e, S2). There were clear biases in the representation of geographical areas globally, e.g. with only a few records of sampling sites originating from Africa compared to Europe or Asia (Fig. 1c, f).

To account for the variable representation of data points across geographic areas, we calculated prevalence as the proportion of studies per country or province (in China) in which each pathogen was detected. We found a low prevalence for all pathogens at the global scale as compared to scales encompassing all of China and, particularly, the NCP (Fig. 2). At the global scale, members of the Sordariomycetes showed a higher prevalence than fungi from other classes, a trend that was consistent at the scale of China but was somewhat lost in the NCP, which showed an elevated frequency of species in the Dothideomycetes, such as *Bipolaris sorokiniana* and *Cercospora sojina* (Fig. 2). A comparison of the prevalence patterns of pathogens across the three scales using Chi-square tests showed that no significant difference ($X^2=0.07-0.9$, $P=1$). Based on a threshold of a ratio ≥ 3 between the normalized prevalence in the NCP respect to the global and China scales, we identified 13 pathogen species to be disproportionately overrepresented in the NCP: *Fusarium graminearum*, *Phomopsis longicolla*, *Cercospora kikuchii*, *C. sojina*, *Exserohilum turcicum*, *Pyrenophora tritici-repentis*, *Bipolaris zeicola*, *B. sorokiniana*, *B. maydis*, *Physoderma maydis*, *Urocystis tritici*, *Sporisorium reilianum* and *Ustilago maydis* (Fig. 2). In contrast, two pathogen species were found to have extremely low representation in the NCP,

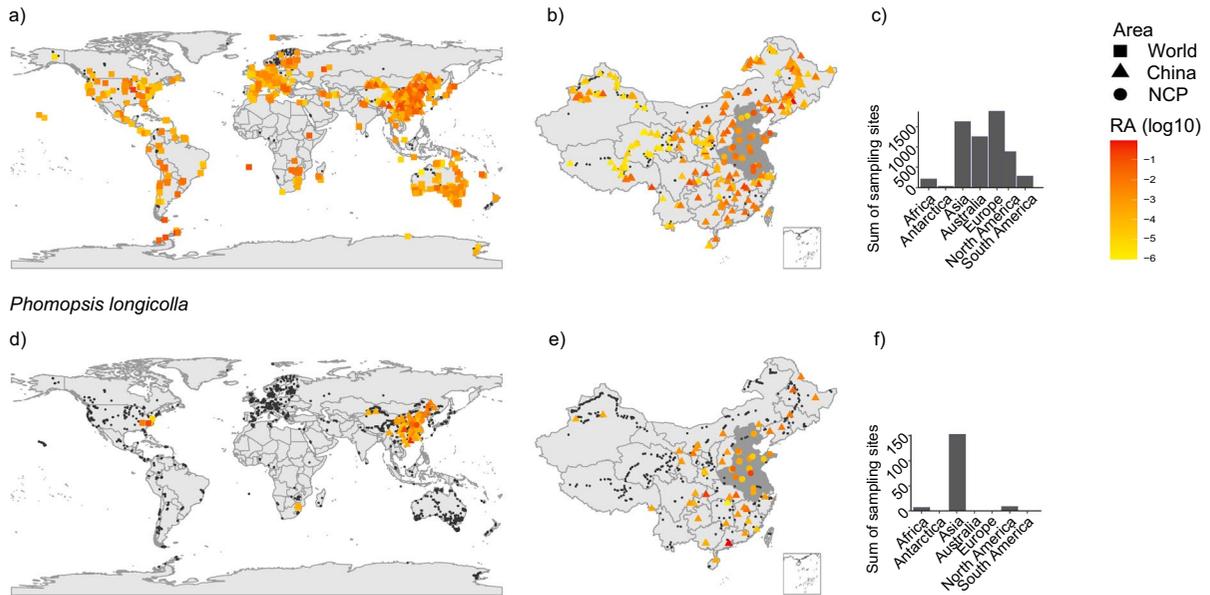
Fusarium oxysporum

Fig. 1 Examples of the distribution of soil-borne fungal pathogens at the three different scales (Global, China and the North China Plain). The maps show the detection across the GlobalFungi dataset of two species selected as examples based on their contrasting distribution ranges: *Fusarium oxysporum* (a-c) and *Phomopsis longicolla* (d-f) (For the distributions of the remaining 23 pathogens selected in this study, see Figure S2). In each map, points indicate sampling sites, with black points and heat-coloured points indicating absence or presence

of the target fungal species, respectively. In coloured points, the colour correlates with the relative abundance (RA) of the pathogen species (\log_{10} -scale, see colour key). Note that the North China Plain is highlighted in dark grey in the maps of China (b, e). The bar plots in c and f show the sum of records of sampling sites detecting *Fusarium oxysporum* (c) and *Phomopsis longicolla* (f) across continents (note the different scales in the y axes)

Fusarium culmorum and *Zymoseptoria tritici*, for which the ratios were zero.

Ecological determinants of pathogen prevalence

We estimated the contribution of variables indicative of climatic, spatial and host availability conditions to the global levels of prevalence of the target soil-borne fungal pathogens using linear regression models (Fig. 3). We did not find a general pattern in the association of fungal groups (i.e. phylogenetically related species or with similar host preferences) with particular sets of climatic and spatial factors, even though more species were affected by spatial factors (e.g. geographical barriers, dispersal limitation, or historical factors) than by climatic conditions (Fig. 3). Likewise, we did not find a relationship between pathogen prevalence and the availability of the respective host crop species, estimated as the crop area density (Fig. 3). Of all pathogen species, only *Urocystis tritici*

and *Sporisorium reilianum* (both biotrophic Basidiomycota) were positively related to regional crop area density of their respective host crops. In the case of *Fusarium proliferatum*, its prevalence was related to crop area density of its host wheat but not of its other hosts maize and soybean, whereas *Cercospora kikuchii* was related to crop area density of its host soybean, but also of the non-host wheat (Fig. 3).

Discussion

We identified 25 soil-borne fungal pathogen species that could potentially affect the productivity of the three main crops produced in the NCP, wheat, maize and soybean. Twenty of these pathogens had widespread distributions across the globe, but 13 species showed prevalence patterns associated with the NCP. An investigation of the underlying ecological factors that could drive the global distribution of these

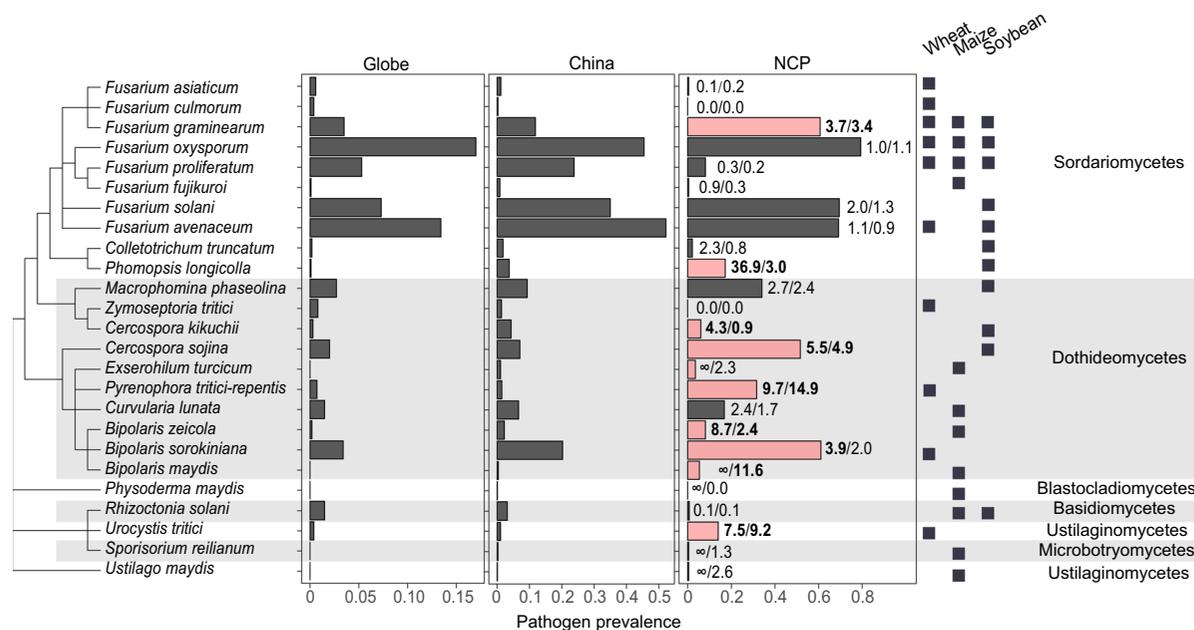


Fig. 2 Prevalence profile of the 25 selected soil-borne fungal pathogens at the three scales considered (Global, China, and North China Plain). Species and class names of the selected pathogens, and their phylogenetic relationships inferred by a Maximum Likelihood phylogeny are shown. The bar plots show the prevalence of soil-borne fungal pathogens at the global, China, and the NCP scale, defined as the proportion of studies per region where the target pathogen was detected (note the different scales in the x axes of each panel). The red bars highlight the pathogens identified as being overpre-

sented in the NCP as compared to the globe and China scales, based on ratios of normalized prevalence (prevalence of each pathogen divided by the maximum prevalence of a pathogen at each scale) above three (values next to bars, globe/China). The ∞ symbol indicates infinite ratio values, which occur when there are zero detections outside of China. The black squares rightwards to the bar plots show the pathogen's host specificity towards the three focal crops (wheat, maize and soybean), based on literature records

pathogens did not reveal consistent relationships between pathogen prevalence and climatic or spatial variables. In addition, the crop area density of the three main crops did not account for the prevalence of the pathogens, except for two specialist pathogens, *Urocystis tritici* and *Sporisorium reilianum*. Based on these data we conclude that other factors not accounted for in our study may be better predictors for the prevalence of potential fungal pathogens in wheat, maize, and soybean in the NCP.

Soil-borne pathogens: taxonomy and fungal lifestyles

Twenty of the 25 studied soil-borne pathogen species belong to the classes of Sordariomycetes and Dothideomycetes, which are known to contain a high proportion of plant pathogenic species (Hyde et al., 2013; Maharachchikumbura et al., 2016). Examples of these pathogenic species from Dothideomycetes are *Zymoseptoria*

tritici (see Table S2; Torriani et al., 2015) and *Bipolaris maydis* (Table S2; Aregbesola et al., 2020); from the class Sordariomycetes—*Phomopsis longicolla* (Table S2; Mengistu et al., 2009). All these pathogens are frequently reported as ubiquitous pathogens in agriculture (Zhang et al., 2006; Haridas et al., 2020).

Based on our results of reviewing literature records, five out of the studied 25 pathogens were generalists (see Table 1). These species can infect a wide range of crops from different families (*F. graminearum*: Chongo et al., 2001; *F. oxysporum*, *F. proliferatum*, *F. avenaceum*: Aoki et al., 2014; O'Donnell et al., 2015; *R. solani*: Anderson, 1982). The other 20 pathogen species were reported to affect only single crops (Table 1; Wheat: Strelkov & Lamari, 2003; Jahani et al., 2014; Maize: Kumar et al., 2022; Xia et al., 2022; Soybean: Goodwin et al., 2001; Li et al., 2010). Of these 20 pathogens associated with single crops, 17 have necrotrophic

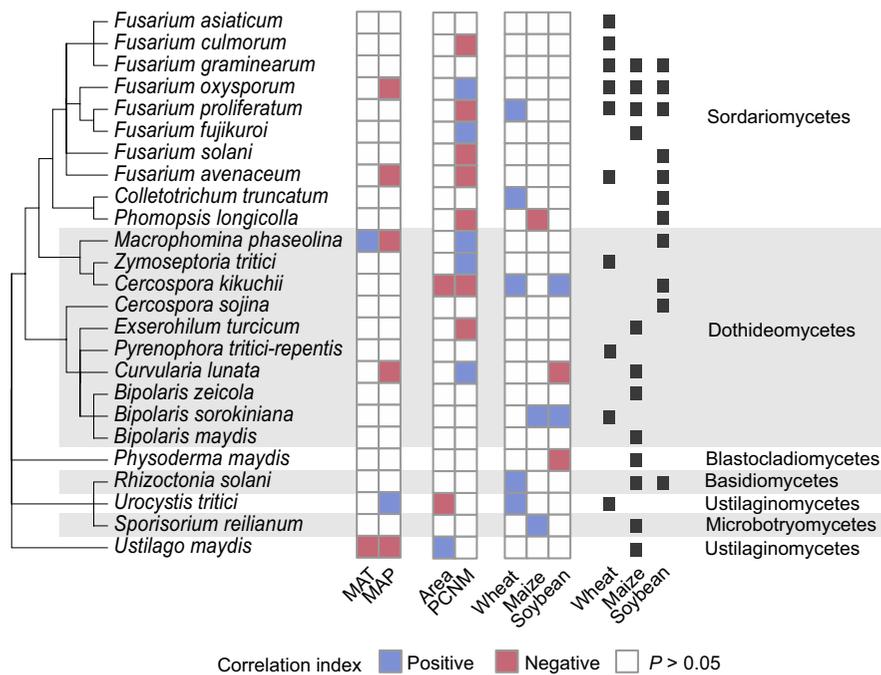


Fig. 3 Relationship between ecological drivers and the prevalence of selected soil-borne fungal pathogen species. The heatmaps summarize results from linear regressions between ecological drivers (i.e. climate, space, and host availability variables) and the prevalence per country of each fungal species. The colours within the cells represent significant positive or negative regression coefficients in the relationship between pathogen prevalence and each variable (see key), thus indicating positive or negative relationships, respectively. The contributions of different PCNMs were combined in a single column, for simplicity, indicative of a correlation between

pathogens prevalence and spatial factors (see Table S5 for the effects of individual PCNMs). Species and class names of the selected pathogens, and their phylogenetic relationships are shown. The black squares on the right of the heatmaps show the pathogens’ host specificity towards the three focal crops (wheat, maize, and soybean), based on literature records. Abbreviations: MAT, mean annual temperature; MAP, mean annual precipitation; PCNM, principal co-ordinates of neighbor matrices, calculated from the countries’ centroid co-ordinates

lifestyles (Table 1), meaning that they can also survive and grow on crop debris or other organic matter in the soils (Glazebrook, 2005; Horbach et al., 2011). For example, Zhen et al. (2009) observed an increased soil-borne disease of wheat when higher amounts of straw amendments were used as inputs to improve soil structure.

Two important soil-borne pathogens for wheat in many regions across the world—*Fusarium culmorum* and *Zymoseptoria tritici* (Wagacha & Muthomi, 2007; Torriani et al., 2015; Li et al., 2016; Zhong et al., 2017) were found to be particularly underrepresented in the NCP (see Fig. 2). In the case of *F. culmorum*, this could be due to its apparent preference for high moisture, as it is observed in wheat fields under high moisture climates in Australia (Backhouse & Burgess, 2002). However,

wheat in the NCP is grown with flood irrigation in the dry part of the season as most rain falls during summer monsoons after wheat harvest and before wheat sowing (Sun et al., 2010). *Z. tritici* is more problematic in temperate climates in Europe than in the NCP, probably for the same reason: the relatively dry growing conditions with little rainfall (Fones & Gurr, 2015; Boixel et al., 2022). It is important to acknowledge that certain regions of the world have been underrepresented in studies on global fungal distribution, which is a common limitation of studies focusing on microbial biogeography (Větrovský et al., 2019; Tedersoo et al., 2022; Makhalyane et al., 2023). Future research should prioritize addressing this gap by expanding the geographic scope and increasing the inclusion of underexplored regions.

Relationships between soil-borne fungal pathogens and host cultivated area

For two out of the 25 fungal species, the biotrophic species *Urocystis tritici* and *Sporisorium reilianum*, we found positively significant relationships between their prevalence and crop area density of their respective host crops, wheat and maize, respectively. These two fungal species are known as true specialists because they depend on their host species to complete their life cycles (*U. tritici*: Zhao et al., 2019; *S. reilianum*: Poloni & Schirawski, 2016), thus highlighting the importance of host density in driving pathogen pressure, which is consistent with predictions in plant epidemiology (Gilbert, 2002; Halliday et al., 2017; Parker et al., 2015).

For the remaining pathogen species (i.e. 23 out of 25) we did not find relationships between their prevalence and crop area density. One possible explanation could be that these soil-borne fungal pathogens have much broader host ranges than biotrophic species, thus their occurrence may depend less on the presence of specific plant species as they can colonize multiple hosts, often without inducing disease symptoms (Jibril et al., 2016; Lofgren et al., 2018; Maciá-Vicente et al., 2020; Newman & Derbyshire, 2020; Maciá-Vicente et al., 2020). It is well known that presumed non-host crop species may serve as pathogen reservoirs (Malcolm et al., 2013; Glynou et al., 2018; Lofgren et al., 2018). For example, *Macrophomina phaseolina* has been found to colonize more than 500 host species (Khan, 2007).

Rather than being the cause of the lack of correlations, it can also be that crop area density is not a good descriptor of pathogen occurrence. Xu et al. (in preparation) observed that in addition to wheat, maize and soybean, 28 other crops are grown in the NCP, such as stevia, chili pepper, cabbage and mung bean (see Table S6 for full list). In addition, the diversity of agricultural practices is immense, ranging from labour-driven systems managed by smallholders to very intensive, mechanized companies that optimize yields (Pan et al., 2014; Zhang et al., 2016; Liang et al., 2022). This diversity of crops and management practices may provide another reason for the small effects of specific host availability on the occurrence of soil-borne pathogens. Therefore, further research is needed to test the actual relationships between fungal pathogens, crops and management practices, in order to provide a baseline to start designing more sustainable cropping systems.

Environmental drivers of pathogen occurrence

Our work reveals inconsistent patterns in the effects of climatic conditions and spatial factors on the distribution of fungal pathogens. Previous studies have emphasized the roles of climate and spatial distance in shaping soil fungal communities (Peay et al., 2010; Tedersoo et al., 2014; Coleman-Derr et al., 2016; Glynou et al., 2016; Maciá-Vicente & Popa, 2022), and this also holds true for soil-borne fungal pathogens (van Agtmaal et al., 2017; Delgado-Baquerizo et al. 2020). However, our finding shows that the soil-borne fungal pathogen species at the three different scales were not consistently affected by environmental conditions. Instead, the distribution of each species responded differently to different sets of conditions, in line with multiple studies investigating the environmental drivers of individual plant-associated fungi (Glynou et al., 2016, 2018; Maciá-Vicente & Popa, 2022). This result is further supported by previous studies in the NCP showing that the occurrence of different pathogen species (*Fusarium* spp.) was affected differently by distinct environmental factors (Dong et al., 2016; Xu et al., 2018). The variation in response to environmental factors among these different pathogen species may be attributed to their genetic diversity, leading to different genotypes with varying sensitivities to environmental factors (Jump et al., 2009; Mann et al., 2014).

Conclusions

Our study identified 13 out of 25 soil-borne fungal pathogens that exhibited a disproportionately high prevalence in the NCP, potentially leading to yield losses in wheat, maize, and soybean crops. We found inconsistent correlations between these 13 pathogen species and environmental factors, indicating individual ecological preferences across fungal species. Notably, two of these 13 species showed a significantly positive correlation with their respective host crop's area density. These findings highlight the importance of proactive management strategies to mitigate yield losses caused by these fungal pathogens in the NCP. Understanding the specific interactions between these pathogens and their host crops, as well as the environmental factors that determine their occurrence and function, will enable the development

of targeted strategies for disease management. Overall, our study provides valuable insights for implementing approaches to enhance crop sustainability in the NCP. By establishing an inventory of pathogen prevalence in the region, our research serves as a foundation for developing sustainable agricultural systems that are more resilient against soil-borne fungal diseases in the future.

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Data Availability The datasets generated and/or analyzed during the current study are available from the corresponding author on reasonable request.

Declarations

Conflict of interest All authors declare that they have no conflict of interest.

References

- Ajayi-Oyetunde, O. O., & Bradley, C. A. (2018). *Rhizoctonia solani*: Taxonomy, population biology and management of rhizoctonia seedling disease of soybean. *Plant Pathology*, *67*(1), 3–17. <https://doi.org/10.1111/ppa.12733>
- Al-Sadi, A. M. (2021). *Bipolaris sorokiniana*-induced black point, common root rot, and spot blotch diseases of wheat: A review. *Frontiers in Cellular and Infection Microbiology*, *11*, 584899. <https://doi.org/10.3389/fcimb.2021.584899>
- Ampt, E. A., van Ruijven, J., Raaijmakers, J. M., Termorshuizen, A. J., & Mommer, L. (2018). Linking ecology and plant pathology to unravel the importance of soil-borne fungal pathogens in species-rich grasslands. *European Journal of Plant Pathology*, *154*(1), 141–156. <https://doi.org/10.1007/s10658-018-1573-x>
- Anderson, N. A. (1982). The genetics and pathology of *Rhizoctonia solani*. *Annual Review of Phytopathology*, *20*(1), 329–347.
- Aoki, T., O'Donnell, K., & Geiser, D. M. (2014). Systematics of key phytopathogenic *Fusarium* species: Current status and future challenges. *Journal of General Plant Pathology*, *80*(3), 189–201. <https://doi.org/10.1007/s10327-014-0509-3>
- Aregbesola, E., Ortega-Beltran, A., Falade, T., Jonathan, G., Hearne, S., & Bandyopadhyay, R. (2020). A detached leaf assay to rapidly screen for resistance of maize to *Bipolaris maydis*, the causal agent of southern corn leaf blight. *European Journal of Plant Pathology*, *156*, 133–145. <https://doi.org/10.1007/s10658-019-01870-4>
- Backhouse, D., & Burgess, L. W. (2002). Climatic analysis of the distribution of *Fusarium graminearum*, *F. pseudograminearum* and *F. culmorum* on cereals in Australia. *Australasian Plant Pathology*, *31*, 321–327. <https://doi.org/10.1071/AP02026>
- Bálint, M., Bahram, M., Eren, A. M., Faust, K., Fuhrman, J. A., Lindahl, B., ... & Tedersoo, L. (2016). Millions of reads, thousands of taxa: microbial community structure and associations analyzed via marker genes. *FEMS Microbiology Reviews*, *40*(5), 686–700. <https://doi.org/10.1093/femsre/fuw017>
- Bever, J. D., Mangan, S., & Alexander, H. M. (2015). Maintenance of plant species diversity by pathogens. *Annual Review of Ecology, Evolution, and Systematics*, *46*(2015), 305–325. <https://doi.org/10.1146/annurev-ecolsys-112414-054306>
- Boixel, A. L., Gélisse, S., Marcel, T. C., & Suffert, F. (2022). Differential tolerance of *Zymoseptoria tritici* to altered optimal moisture conditions during the early stages of wheat infection. *Journal of Plant Pathology*, *104*(2), 495–507. <https://doi.org/10.1007/s42161-021-01025-7>
- Boudreau, M. A. (2013). Diseases in intercropping systems. *Annual Review of Phytopathology*, *51*, 499–519.
- Brauns, B., Jakobsen, R., Song, X., & Bjerg, P. L. (2018). Pesticide use in the wheat-maize double cropping systems of the North China Plain: Assessment, field study, and implications. *Science of the Total Environment*, *616–617*, 1307–1316. <https://doi.org/10.1016/j.scitotenv.2017.10.187>
- Chaudhary, V. B., Aguilar-Trigueros, C. A., Mansour, I., & Rillig, M. C. (2022). Fungal dispersal across spatial scales. *Annual Review of Ecology, Evolution, and Systematics*, *53*(1), 69–85. <https://doi.org/10.1146/annurev-ecolsys-012622-021604>
- Chen, Y., Kistler, H. C., & Ma, Z. (2019a). *Fusarium graminearum* trichothecene mycotoxins: Biosynthesis, regulation, and management. *Annual Review of Phytopathology*, *57*, 15–39. <https://doi.org/10.1146/annurev-phyto-082718-100318>
- Chen, S., Waghmode, T. R., Sun, R., Kuramae, E. E., Hu, C., & Liu, B. (2019b). Root-associated microbiomes of wheat under the combined effect of plant development and nitrogen fertilization. *Microbiome*, *7*(1), 136. <https://doi.org/10.1186/s40168-019-0750-2>
- Chongo, G., Gossen, B. D., Kutcher, H. R., Gilbert, J., Turkington, T. K., Fernandez, M. R., & McLaren, D. (2001). Reaction of seedling roots of 14 crop species to *Fusarium graminearum* from wheat heads. *Canadian Journal of Plant Pathology*, *23*(2), 132–137. <https://doi.org/10.1080/07060660109506920>
- Coleman-Derr, D., Desgarennes, D., Fonseca-Garcia, C., Gross, S., Clingenpeel, S., ... & Tringe, S. G. (2016). Plant compartment and biogeography affect microbiome composition in cultivated and native *Agave* species. *New Phytologist*, *209*(2), 798–811. <https://doi.org/10.1111/nph.13697>
- Corkley, I., Fraaije, B., & Hawkins, N. (2022). Fungicide resistance management: Maximizing the effective life of plant protection products. *Plant Pathology*, *71*(1), 150–169. <https://doi.org/10.1111/ppa.13467>

- De Coninck, B., Timmermans, P., Vos, C., Cammue, B. P., & Kazan, K. (2015). What lies beneath: Belowground defense strategies in plants. *Trends in Plant Science*, 20(2), 91–101. <https://doi.org/10.1016/j.tplants.2014.09.007>
- Delgado-Baquerizo, M., Guerra, C. A., Cano-Díaz, C., Egidi, E., Wang, J.-T., Eisenhauer, N., Singh, B. K., & Maestre, F. T. (2020). The proportion of soil-borne pathogens increases with warming at the global scale. *Nature Climate Change*, 10(6), 550–554. <https://doi.org/10.1038/s41558-020-0759-3>
- Desprez-Loustau, M. L., Courtecuisse, R., Robin, C., Husson, C., Moreau, P. A., Blancard, D., ... & Sache, I. (2010). Species diversity and drivers of spread of alien fungi (sensu lato) in Europe with a particular focus on France. *Biological Invasions*, 12, 157–172. <https://doi.org/10.1007/s10530-009-9439-y>
- Dong, F., Qiu, J., Xu, J., Yu, M., Wang, S., Sun, Y., ... & Shi, J. (2016). Effect of environmental factors on *Fusarium* population and associated trichothecenes in wheat grain grown in Jiangsu province, China. *International Journal of Food Microbiology*, 230, 58–63. <https://doi.org/10.1016/j.ijfoodmicro.2016.04.020>
- Dray, S., Legendre, P., & Peres-Neto, P. R. (2006). Spatial modelling: A comprehensive framework for principal coordinate analysis of neighbour matrices (PCNM). *Ecological Modelling*, 196(3–4), 483–493. <https://doi.org/10.1016/j.ecolmodel.2006.02.015>
- FAOSTAT. (2022). Food and agriculture organization of the United Nations (FAO), *FAOSTAT Database*. Retrieved January 13, 2022, from <https://www.fao.org/faostat/en/#data/QCL>
- Farr, D., & Rossman, A. (2021) “Fungal Databases.” Systematic Mycology and Microbiology Laboratory, ARS, USDA. Retrieved December 8, 2021, from <http://nt.ars-grin.gov/fungaldatabases/>
- Fife, D., & Fife, M. D. (2017). Package ‘fifer’: a biostatisticians toolbox for various activities, including plotting, data cleanup, and data analysis. Retrieved January 26, 2022, from <http://cran.nexr.com/web/packages/fifer/index.html>
- Flett, B., & Wehner, F. (1991). Incidence of stenocarpella and *Fusarium* cob rots in monoculture maize under different tillage systems. *Journal of Phytopathology*, 133(4), 327–333. <https://doi.org/10.1111/j.1439-0434.1991.tb00168.x>
- Fones, H., & Gurr, S. (2015). The impact of *Septoria tritici* Blotch disease on wheat: An EU perspective. *Fungal Genetics and Biology*, 79, 3–7. <https://doi.org/10.1016/j.fgb.2015.04.004>
- Fu, J. (2017). Occurrence and control measures of corn sheath blight in Cheng’an city of Hebei (in Chinese). *China Agricultural Information*, 15, 42–44. Retrieved December 1, 2021, from <http://www.cqvip.com/QK/98116B/201715/673148145.html>
- Gao, X. (2016). *Identification on the population structure and mycotoxin chemotypes of Fusarium head blight as well as its sensivity of fungicide in Shandong (in Chinese)*. Master thesis, Shandong Agricultural University.
- Gargouri, S., Khemir, E., Souissi, A., Paulitz, T., Murray, T., Fakhfakh, M., Achour, I., Chekali, S., Mliki, Y., & Burgess, L. (2020). Survey of take-all (*Gaeumannomyces tritici*) on cereals in Tunisia and impact of crop sequences. *Crop Protection*, 135, 105189. <https://doi.org/10.1016/j.cropro.2020.105189>
- Ghosh, S., Kanwar, P., & Jha, G. (2017). Alterations in rice chloroplast integrity, photosynthesis and metabolome associated with pathogenesis of *Rhizoctonia solani*. *Scientific Reports*, 7(1), 1–12. <https://doi.org/10.1016/10.1038/srep41610>
- Gilbert, G. S. (2002). Evolutionary ecology of plant diseases in natural ecosystems. *Annual Review of Phytopathology*, 40, 13–43. <https://doi.org/10.1146/annurev.phyto.40.021202.110417>
- Gilbert, B., & Bennett, J. R. (2010). Partitioning variation in ecological communities: Do the numbers add up? *Journal of Applied Ecology*, 47(5), 1071–1082. <https://doi.org/10.1111/j.1365-2664.2010.01861.x>
- Glazebrook, J. (2005). Contrasting mechanisms of defense against biotrophic and necrotrophic pathogens. *Annual Review of Phytopathology*, 43, 205–227. <https://doi.org/10.1146/annurev.phyto.43.040204.135923>
- Glynou, K., Nam, B., Thines, M., & Maciá-Vicente, J. G. (2018). Facultative root-colonizing fungi dominate endophytic assemblages in roots of nonmycorrhizal *Microthlaspi* species. *New Phytologist*, 217(3), 1190–1202. <https://doi.org/10.1111/nph.14873>
- Glynou, K., Ali, T., Buch, A. K., Haghi Kia, S., Ploch, S., Xia, X., ... & Maciá-Vicente, J. G. (2016). The local environment determines the assembly of root endophytic fungi at a continental scale. *Environmental Microbiology*, 18(8), 2418–2434. <https://doi.org/10.1111/1462-2920.13112>
- Goodwin, S. B., Dunkle, L. D., & Zismann, V. L. (2001). Phylogenetic analysis of *Cercospora* and *Mycosphaerella* based on the internal transcribed spacer region of ribosomal DNA. *Phytopathology*, 91(7), 648–658. <https://doi.org/10.1094/PHYTO.2001.91.7.648>
- Griffith, D. A., & Peres-Neto, P. R. (2006). Spatial modeling in ecology: The flexibility of eigenfunction spatial analyses. *Ecology*, 87(10), 2603–2613. [https://doi.org/10.1890/0012-9658\(2006\)87\[2603:SMIETF\]2.0.CO;2](https://doi.org/10.1890/0012-9658(2006)87[2603:SMIETF]2.0.CO;2)
- Halliday, F. W., Heckman, R. W., Wilfahrt, P. A., & Mitchell, C. E. (2017). A multivariate test of disease risk reveals conditions leading to disease amplification. *Proceedings of the Royal Society B: Biological Sciences*, 284(1865), 20171340. <https://doi.org/10.1098/rspb.2017.1340>
- Haridas, S., Albert, R., Binder, M., Bloem, J., LaButti, K., Salamov, A., ... & Grigoriev, I. V. (2020). 101 Dothideomycetes genomes: a test case for predicting lifestyles and emergence of pathogens. *Studies in Mycology*, 96(1), 141–153. <https://doi.org/10.1016/j.simyco.2020.01.003>
- He, X. (2016). *Genetic diversity and pathogenicity analysis of Fusarium pseudograminearum that is dominant pathogen causing crown rot of wheat (in Chinese)*. Master thesis, Henan agricultural University.
- Hiddink, G. A., Termorshuizen, A. J., & Van Bruggen, A. H. C. (2010). Mixed cropping and suppression of Soilborne diseases. In E. Lichtfouse (Eds.), *Genetic Engineering, Biofertilisation, Soil Quality and Organic Farming. Sustainable Agriculture Reviews (Vol. 4, 119–146)*. Dordrecht: Springer.
- Horbach, R., Navarro-Quesada, A. R., Knogge, W., & Deising, H. B. (2011). When and how to kill a plant cell: Infection

- strategies of plant pathogenic fungi. *Journal of Plant Physiology*, 168(1), 51–62. <https://doi.org/10.1016/j.jplph.2010.06.014>
- Hyde, K. D., Jones, G., Liu, J. K., Ariyawansa, H., Boehm, E., Boonmee, S., ... & Zhang, M. (2013). Families of dothideomycetes. *Fungal Diversity*, 63(1), 1. <https://doi.org/10.1007/s13225-013-0263-4>
- Jahani, M., Aggarwal, R., Gupta, S., Sharma, S., & Dureja, P. (2014). Purification and characterization of a novel toxin from *Bipolaris sorokiniana*, causing spot blotch of wheat and analysis of variability in the pathogen. *Cereal Research Communications*, 42(2), 252–261. <https://doi.org/10.1556/crc.2013.0053>
- Ji, Y., & Han, Y. (2016). Causes and control strategies of wheat sheath blight in Wuqiao County in 2015 (in Chinese). *Hebei Agriculture*, 01, 32–34.
- Jibril, S. M., Jakada, B. H., Kutama, A. S., & Umar, H. Y. (2016). Plant and pathogens: pathogen recognition, invasion and plant defense mechanism. *International Journal of Current Microbiology and Applied Sciences*, 5(6), 247–257.
- Jump, A. S., Marchant, R., & Peñuelas, J. (2009). Environmental change and the option value of genetic diversity. *Trends in Plant Science*, 14(1), 51–58. <https://doi.org/10.1016/j.tplants.2008.10.002>
- Kang, R., Hu, Y., Wang, L., Xie, S., Li, Y., & Li, H. (2020). Pathogenicity variation and DNA polymorphism of *Bipolaris sorokiniana* infecting winter wheat in the Huanghuai floodplain of China. *Plant Pathology*, 70(1), 87–99.
- Karlsson, I., Persson, P., & Friberg, H. (2021). Fusarium head blight from a microbiome perspective. *Frontiers in Microbiology*, 12, 628373. <https://doi.org/10.3389/fmicb.2021.628373>
- Khan, S. N. (2007). *Macrophomina phaseolina* as causal agent for charcoal rot of sunflower. *Mycopath*, 5, 111–118.
- Kumar, S., Stecher, G., & Tamura, K. (2016). MEGA7: molecular evolutionary genetics analysis version 7.0 for bigger datasets. *Molecular Biology and Evolution*, 33(7), 1870–1874. <https://doi.org/10.1093/molbev/msw054>
- Kumar, B., Choudhary, M., Kumar, K., Kumar, ... & Rakshit, S. (2022). Maydis leaf blight of maize: Update on status, sustainable management and genetic architecture of its resistance. *Physiological and Molecular Plant Pathology*, 121, 101889. <https://doi.org/10.1016/j.pmpp.2022.101889>
- Li, S., Hartman, G. L., & Boykin, D. L. (2010). Aggressiveness of *Phomopsis longicolla* and other *Phomopsis* spp. on soybean. *Plant Disease*, 94(8), 1035–1040. <https://doi.org/10.1094/PDIS-94-8-1035>
- Li, H. L., He, X. L., Ding, S. L., Yuan, H. X., & Chen, L. L. (2016). First report of *Fusarium culmorum* causing crown rot of wheat in China. *Plant Disease*, 100(12), 2532–2532. <https://doi.org/10.1094/PDIS-05-16-0723-PDN>
- Li, H., Dai, M., Dai, S., & Dong, X. (2018). Current status and environment impact of direct straw return in China's cropland—a review. *Ecotoxicology and Environmental Safety*, 159, 293–300. <https://doi.org/10.1016/j.ecoenv.2018.05.014>
- Li, X. (2018). Occurrence status and comprehensive prevention and control measures of wheat soil-borne diseases in Tanghe County (in Chinese). *Modern Agriculture and Technology*, 11, 121.
- Liang, Z., van der Werf, W., Xu, Z., Cheng, J., Wang, C., Cong, W. F., ... & Groot, J. C. (2022). Identifying exemplary sustainable cropping systems using a positive deviance approach: Wheat-maize double cropping in the North China Plain. *Agricultural Systems*, 201, 103471. <https://doi.org/10.1016/j.agry.2022.103471>
- Lin, R., Xia, Y., Liu, Y., Zhang, D., Xiang, X., Niu, X., Jiang, L., Wang, X., & Zheng, A. (2021). Comparative mitogenomic analysis and the evolution of *Rhizoctonia solani* anastomosis groups. *Frontiers in Microbiology*, 12, 707281. <https://doi.org/10.3389/fmicb.2021.707281>
- Liu, W., Zhang, Y., Jiang, S., Murray, P. J., Liao, L., Li, X., & Zhang, J. (2019). Spatiotemporal differences in the arbuscular mycorrhizal fungi communities in soil and roots in response to long-term organic compost inputs in an intensive agricultural cropping system on the North China Plain. *Journal of Soils and Sediments*, 19(5), 2520–2533. <https://doi.org/10.1007/s11368-019-02244-3>
- Liu, S., Guo, N., Ma, H., Sun, H., Zheng, X., & Shi, J. (2021). First report of root rot caused by *Bipolaris zeicola* on maize in Hebei province. *Plant Disease*. <https://doi.org/10.1094/PDIS-01-21-0060-PDN>
- Liu, M., Zhang, W., Wang, X., Wang, F., Dong, W., Hu, C., ... & Sun, R. (2020). Nitrogen leaching greatly impacts bacterial community and denitrifiers abundance in subsoil under long-term fertilization. *Agriculture, Ecosystems & Environment*, 294, 106885. <https://doi.org/10.1016/j.agee.2020.106885>
- Lofgren, L. A., LeBlanc, N. R., Certano, A. K., Nachtigall, J., LaBine, K. M., Riddle, J., ... & Kistler, H. C. (2018). *Fusarium graminearum*: pathogen or endophyte of North American grasses?. *New Phytologist*, 217(3), 1203–1212. <https://doi.org/10.1111/nph.14894>
- Ma, J. (2006). *The region of physiological differentiation of Sporisorium reilianum in China (in Chinese)*. Master thesis. Jilin Agricultural University.
- Maciá-Vicente, J. G., & Popa, F. (2022). Local endemism and ecological generalism in the assembly of root-colonizing fungi. *Ecological Monographs*, 92(1), e01489. <https://doi.org/10.1002/ecm.1489>
- Maciá-Vicente, J. G., Nam, B., & Thines, M. (2020). Root filtering, rather than host identity or age, determines the composition of root-associated fungi and oomycetes in three naturally co-occurring Brassicaceae. *Soil Biology and Biochemistry*, 146, 107806. <https://doi.org/10.1016/j.soilbio.2020.107806>
- Maharachchikumbura, S. S., Hyde, K. D., Jones, E. G., McKenzie, E. H. C., Bhat, J. D., Dayarathne, M. C., ... & Wijayawardene, N. N. (2016). Families of sordariomycetes. *Fungal Diversity*, 79, 1–317. <https://doi.org/10.1007/s13225-016-0369-6>
- Makhalanyane, T. P., Bezuidt, O. K., Pierneef, R. E., Mizrahi, E., Zeze, A., Fossou, R. K., ... & Murrell, C. (2023). African microbiomes matter. *Nature Reviews Microbiology*, 1–3. <https://doi.org/10.1038/s41579-023-00925-y>
- Malcolm, G. M., Kuldau, G. A., Gugino, B. K., & del Mar Jiménez-Gasco, M. (2013). Hidden host plant associations of soilborne fungal pathogens: An ecological

- perspective. *Phytopathology*, 103(6), 538–544. <https://doi.org/10.1094/PHYTO-08-12-0192-LE>
- Mann, M. B., Spadari, C. C., Feltrin, T., Frazzon, A. P. G., Germani, J. C., & Sand, S. T. (2014). Genetic variability of *Bipolaris sorokiniana* isolates using URP-PCR. *Tropical Plant Pathology*, 39, 163–171. <https://doi.org/10.1590/S1982-56762014000200007>
- Marquez, N., Giachero, M. L., Declerck, S., & Ducasse, D. A. (2021). *Macrophomina phaseolina*: General characteristics of pathogenicity and methods of control. *Frontiers in Plant Science*, 12, 634397. <https://doi.org/10.3389/fpls.2021.634397>
- Meng, C. (2019). Identification of pathogenic *Fusarium* spp. causing wheat crown rot and corn stalk rot under wheat-maize rotation system (in Chinese). Master thesis, *Shandong Agricultural University*.
- Mengistu, A., Castlebury, L., Smith, R., Ray, J., & Bellaloui, N. (2009). Seasonal progress of *Phomopsis longicolla* infection on soybean plant parts and its relationship to seed quality. *Plant Disease*, 93(10), 1009–1018. <https://doi.org/10.1094/PDIS-93-10-1009>
- Miao, Y., Stewart, B. A., & Zhang, F. (2011). Long-term experiments for sustainable nutrient management in China: A review. *Agronomy for Sustainable Development*, 31(2), 397–414. <https://doi.org/10.1051/agro/2010034>
- Mikaberidze, A., Paveley, N., Bonhoeffer, S., & van den Bosch, F. (2017). Emergence of resistance to fungicides: The role of fungicide dose. *Phytopathology*, 107(5), 545–560. <https://doi.org/10.1094/PHYTO-08-16-0297-R>
- Mommer, L., Cotton, T.E.A., Raaijmakers, J.M., Termorshuizen, A.J., van Ruijven, J., ... & Dumbrell, A. J. (2018). Lost in diversity: the interactions between soil-borne fungi, biodiversity and plant productivity. *New Phytologist*, 218(2), 542–553. <https://doi.org/10.1111/nph.15036>
- Mu, H., Zhang, J., Yang, X., Wang, K., Xu, W., Zhang, H., Liu, X., Ritsema, C. J., & Geissen, V. (2022). Pesticide screening and health risk assessment of residential dust in a rural region of the North China Plain. *Chemosphere*, 303(Pt 2), 135115. <https://doi.org/10.1016/j.chemosphere.2022.135115>
- National Bureau of Statistics of China, (2021). China Statistics Yearbook (in Chinese). Chinese Statistics Press, Beijing (2021).
- Newman, T. E., & Derbyshire, M. C. (2020). The evolutionary and molecular features of broad host-range necrotrophy in plant pathogenic fungi. *Frontiers in Plant Science*, 11, 591733. <https://doi.org/10.3389/fpls.2020.591733>
- Nilsson, R. H., Anslan, S., Bahram, M., Wurzbacher, C., Baldrian, P., & Tedersoo, L. (2019). Mycobiome diversity: High-throughput sequencing and identification of fungi. *Nature Reviews Microbiology*, 17(2), 95–109. <https://doi.org/10.1038/s41579-018-0116-y>
- O'Donnell, K., Ward, T. J., Robert, V. A., Crous, P. W., Geiser, D. M., & Kang, S. (2015). DNA sequence-based identification of *Fusarium*: Current status and future directions. *Phytoparasitica*, 43(5), 583–595. <https://doi.org/10.1094/PDIS-09-21-2035-SR>
- Oksanen, J., Blanchet, F. G., Kindt, R., Legendre, P., Minchin, P. R., O'hara, R., Simpson, G. L., Solymos, P., Stevens, M. H. H., & Wagner, H. (2020). Vegan: community ecology package. Retrieved January 26, 2022, from <https://CRAN.R-project.org/package=vegan>.
- Ons, L., Bylemans, D., Thevissen, K., & Cammue, B. P. (2020). Combining biocontrol agents with chemical fungicides for integrated plant fungal disease control. *Microorganisms*, 8(12), 1930. <https://doi.org/10.3390/microorganisms8121930>
- Pan, Y., Yu, Z., Holst, J., & Doluschitz, R. (2014). Integrated assessment of cropping patterns under different policy scenarios in Quzhou County, North China Plain. *Land Use Policy*, 40, 131–139. <https://doi.org/10.1016/j.landusepol.2013.11.015>
- Parker, I. M., Saunders, M., Bontrager, M., Weitz, A. P., Hendricks, R., Magarey, R., ... & Gilbert, G. S. (2015). Phylogenetic structure and host abundance drive disease pressure in communities. *Nature*, 520(7548), 542–544. <https://doi.org/10.1038/nature14372>
- Paungfoo-Lonhienne, C., Yeoh, Y. K., Kasinadhuni, N. R. P., Lonhienne, T. G., Robinson, N., Hugenholtz, P., ... & Schmidt, S. (2015). Nitrogen fertilizer dose alters fungal communities in sugarcane soil and rhizosphere. *Scientific Reports*, 5(1), 1–6. <https://doi.org/10.1038/srep08678>
- Peay, K. G., Garbelotto, M., & Bruns, T. D. (2010). Evidence of dispersal limitation in soil microorganisms: Isolation reduces species richness on mycorrhizal tree islands. *Ecology*, 91(12), 3631–3640. <https://doi.org/10.1890/09-2237.1>
- Peres-Neto, P. R., & Legendre, P. (2010). Estimating and controlling for spatial structure in the study of ecological communities. *Global Ecology and Biogeography*, 19(2), 174–184. <https://doi.org/10.1111/j.1466-8238.2009.00506.x>
- Pérez-Brandán, C., Huidobro, J., Grümberg, B., Scandiani, M.M., ... & Vargas-Gil, S. (2014). Soybean fungal soil-borne diseases: a parameter for measuring the effect of agricultural intensification on soil health. *Canadian Journal of Microbiology*, 60(2), 73–84. <https://doi.org/10.1139/cjm-2013-0792>
- Pinto, F. A. M. F., Porto, V. B. C., Guimarães, R. A., da Silva Siqueira C., ... & Medeiros, F.H.V.d., (2022). Detection and factors that induce *Stenocarpella* spp. survival in maize stubble and soil suppressiveness under tropical conditions. *Applied Sciences*, 12(10). <https://doi.org/10.3390/app12104974>.
- Poloni, A., & Schirawski, J. (2016). Host specificity in *Sporisorium reilianum* is determined by distinct mechanisms in maize and sorghum. *Molecular Plant Pathology*, 17(5), 741–754. <https://doi.org/10.1111/mp.12326>
- Price, C. L., Parker, J. E., Warrilow, A. G., Kelly, D. E., & Kelly, S. L. (2015). Azole fungicides—understanding resistance mechanisms in agricultural fungal pathogens. *Pest Management Science*, 71(8), 1054–1058. <https://doi.org/10.1002/ps.4029>
- Qi, X. (2015). *Study on the flora distribution and biological characteristics of Fusarium graminearum in Anhui (in Chinese)*. Master thesis. Anhui Agricultural University.
- R Core Team. (2021). R: a language and environment for statistical computing. *R Foundation for Statistical Computing*, Vienna, Austria. Retrieved January 26, 2022, from <http://www.r-project.org/>.
- Ramanauskienė, J., Semaškienė, R., Jonavičienė, A., & Ronis, A. (2018). The effect of crop rotation and fungicide seed

- treatment on take-all in winter cereals in Lithuania. *Crop Protection*, 110, 14–20. <https://doi.org/10.1016/j.cropro.2018.03.011>
- Ren, X. (2011). *Diversity analysis of Fusarium spp., the main causal agents of maize ear rot in China (in Chinese)*. Master thesis. Chinese Academy of Agricultural Sciences.
- Ristaino, J. B., Anderson, P. K., Bebbler, D. P., Brauman, K. A., Cunniffe, N. J., Fedoroff, N. V., ... & Wei, Q. (2021). The persistent threat of emerging plant disease pandemics to global food security. *Proceedings of the National Academy of Sciences*, 118(23), e2022239118. <https://doi.org/10.1073/pnas.2022239118>
- Savary, S., Willocquet, L., Pethybridge, S. J., Esker, P., McRoberts, N., & Nelson, A. (2019). The global burden of pathogens and pests on major food crops. *Nature Ecology & Evolution*, 3(3), 430–439. <https://doi.org/10.1038/s41559-018-0793-y>
- Scherm, B., Balmas, V., Spanu, F., Pani, G., Delogu, G., Pasquali, M., & Migheli, Q. (2013). *Fusarium culmorum*: Causal agent of foot and root rot and head blight on wheat. *Molecular Plant Pathology*, 14(4), 323–341. <https://doi.org/10.1111/mpp.12011>
- Shan, L. (2017). *Identification and diversity analysis of Fusarium spp. causing maize stem rot disease (in Chinese)*. Master thesis. Chinese Academy of Agricultural Sciences.
- Short, D. P., Gurung, S., Gladieux, P., Inderbitzin, P., Atallah, Z. K., ... & K. V. (2015). Globally invading populations of the fungal plant pathogen *Verticillium dahliae* are dominated by multiple divergent lineages. *Environmental Microbiology*, 17(8), 2824–2840. <https://doi.org/10.1111/1462-2920.12789>
- Silva, D.N., Talhinhas, P., Cai, L., Manuel, L., Gichuru, E.K., Loureiro, A., ... & Batista, D. (2012). Host-jump drives rapid and recent ecological speciation of the emergent fungal pathogen *Colletotrichum kahawae*. *Molecular Ecology*, 21(11), 2655–2670. <https://doi.org/10.1111/j.1365-294X.2012.05557.x>
- South, A. (2017). R.natural-earth: world map data from natural earth. *R package* version 0.1.0. 898, 79–88.
- Strelkov, S., & Lamari, L. (2003). Host-parasite interactions in tan spot *Pyrenophora tritici-repentis* of wheat. *Canadian Journal of Plant Pathology*, 25(4), 339–349. <https://doi.org/10.1080/07060660309507089>
- Summerell, B. A., Laurence, M. H., Liew, E. C., & Leslie, J. F. (2010). Biogeography and phylogeography of *Fusarium*: A review. *Fungal Diversity*, 44(1), 3–13. <https://doi.org/10.1007/s13225-010-0060-2>
- Sun, H., Shen, Y., Yu, Q., Flerchinger, G. N., Zhang, Y., Liu, C., & Zhang, X. (2010). Effect of precipitation change on water balance and WUE of the winter wheat–summer maize rotation in the North China Plain. *Agricultural Water Management*, 97(8), 1139–1145. <https://doi.org/10.1016/j.agwat.2009.06.004>
- Sun, Q., Kröbel, R., Müller, T., Römheld, V., Cui, Z., Zhang, F., & Chen, X. (2011). Optimization of yield and water-use of different cropping systems for sustainable groundwater use in North China Plain. *Agricultural Water Management*, 98(5), 808–814. <https://doi.org/10.1016/j.agwat.2010.12.007>
- Sun, J., Xie, S., Liu, J., Liu, J., Hao, J., & Deng, S. (2014). Morphological and molecular identification of *Fusarium* isolated from basal stalks of maize in Henan Province (in Chinese). *Acta Phytopathologica Sinica*, 44, 8–16.
- Sun, R., Chen, Y., Han, W., Dong, W., Zhang, Y., Hu, C., ... & Wang, F. (2020). Different contribution of species sorting and exogenous species immigration from manure to soil fungal diversity and community assemblage under long-term fertilization. *Soil Biology and Biochemistry*, 151, 108049. <https://doi.org/10.1016/j.soilbio.2020.108049>
- Tedersoo, L., Bahram, M., Pölmme, S., Kõljalg, U., Yorou, N. S., Wijesundera, R., Ruiz, L. V., Vasco-Palacios, A. M., Thu, P. Q., & Suija, A. (2014). Global diversity and geography of soil fungi. *Science*, 346(6213), 1256688. <https://doi.org/10.1126/science.1256688>
- Tedersoo, L., Mikryukov, V., Zizka, A., Bahram, M., Hagh-Doust, N., Anslan, S., Prylutskiy, & Abarenkov, K. (2022). Global patterns in endemism and vulnerability of soil fungi. *Global Change Biology*. <https://doi.org/10.1111/gcb.16398>
- Termorshuizen, A. J. (2014). Root pathogens. In: J Dighton, & JA Krumins (Eds.), *Interactions in soil: promoting plant growth. Biodiversity, community and ecosystems* (vol. 1, 119–137). Dordrecht, the Netherlands: Springer.
- Torriani, S. F., Melichar, J. P., Mills, C., Pain, N., Sierotzki, H., & Courbot, M. (2015). *Zymoseptoria tritici*: A major threat to wheat production, integrated approaches to control. *Fungal Genetics and Biology*, 79, 8–12. <https://doi.org/10.1016/j.fgb.2015.04.010>
- van Agtmaal, M., Straathof, A., Termorshuizen, A., Teurlincx, S., Hundscheid, & Boer, W. (2017). Exploring the reservoir of potential fungal plant pathogens in agricultural soil. *Applied Soil Ecology*, 121, 152–160. <https://doi.org/10.1016/j.apsoil.2017.09.032>
- Venables, W. N., & Ripley, B. D. (2013). *Modern applied statistics with S-PLUS*. Springer Science & Business Media.
- Větrovský, T., Kohout, P., Kopecký, M., Machac, A., Man, M., Bahnmann, B. D., ... & Baldrian, P. (2019). A meta-analysis of global fungal distribution reveals climate-driven patterns. *Nature Communications*, 10(1), 5142. <https://doi.org/10.1038/s41467-019-13164-8>
- Větrovský, T., Morais, D., Kohout, P., Lepinay, C., Algora, C., Awokunle Hollá, S., ... & Baldrian, P. (2020). GlobalFungi, a global database of fungal occurrences from high-throughput-sequencing metabarcoding studies. *Scientific Data*, 7(1), 228. <https://doi.org/10.1038/s41597-020-0567-7>
- Vujanovic, V., Hamel, C., Yergeau, E., & St-Arnaud, M. (2006). Biodiversity and biogeography of *Fusarium* species from northeastern North American asparagus fields based on microbiological and molecular approaches. *Microbial Ecology*, 51(2), 242–255. <https://doi.org/10.1007/s00248-005-0046-x>
- Wagacha, J. M., & Muthomi, J. W. (2007). *Fusarium culmorum*: Infection process, mechanisms of mycotoxin production and their role in pathogenesis in wheat. *Crop Protection*, 26(7), 877–885. <https://doi.org/10.1016/j.cropro.2006.09.003>
- Walters, D. R., & Bingham, I. J. (2007). Influence of nutrition on disease development caused by fungal pathogens: Implications for plant disease control. *Annals of Applied*

- Biology*, 151(3), 307–324. <https://doi.org/10.1111/j.1744-7348.2007.00176.x>
- Wang, Y. (2019). Detection of soybean root pathogens and identification of soybean germplasm resistance to *Phytophthora* root rot from Huanghuaihai region. Nanjing Agricultural University.
- Wang, S., Hu, Y., Yuan, R., Feng, W., Pan, Y., & Yang, Y. (2019). Ensuring water security, food security, and clean water in the North China Plain - conflicting strategies. *Current Opinion in Environmental Sustainability*, 40, 63–71. <https://doi.org/10.1016/j.cosust.2019.09.008>
- Wardle, D. A., & Lindahl, B. D. (2014). Disentangling global soil fungal diversity. *Science*, 346(6213), 1052–1053. <https://doi.org/10.1126/science.aaa1185>
- Wei, W., Yang, M., Liu, Y., Huang, H., Ye, C., Zheng, J., ... & Zhu, S. (2018). Fertilizer N application rate impacts plant-soil feedback in a sanqi production system. *Science of the Total Environment*, 633, 796–807. <https://doi.org/10.1016/j.scitotenv.2018.03.219>
- Xia, X., Wang, Y., Zhou, S., Liu, W., & Wu, H. (2022). Genome sequence resource for *Bipolaris zeicola*, the cause of northern corn leaf spot disease. *Phytopathology*, 112(5), 1192–1195. <https://doi.org/10.1094/PHYTO-05-21-0196-A>
- Xin, B., & Yu, Y. (2021). Major wheat diseases and their comprehensive prevention and control techniques (in Chinese). *Nong Jia Can Mou*, 06, 52–52.
- Xu, F., Yang, G., Wang, J., Song, Y., Liu, L., Zhao, K., Li, Y., & Han, Z. (2018). Spatial distribution of root and crown rot fungi associated with winter wheat in the North China Plain and its relationship with climate variables. *Frontiers in Microbiology*, 9, 1054. <https://doi.org/10.3389/fmicb.2018.01054>
- Xue, L. (2006). Occurrence and control strategy of corn sheath blight (in Chinese). *Henan Agriculture*, 11.
- Yang, S. (2012). Soybean *Fusarium* root rot pathogen species and soybean germplasm screening (in Chinese). Master thesis, Nanjing Agricultural University.
- Ye, W., Zeng, D., Xu, M., Yang, J., Ma, J., Wang, Y., & Zheng, X. (2020). A LAMP-assay-based specific microbiota analysis reveals community dynamics and potential interactions of 13 major soybean root pathogens. *Journal of Integrative Agriculture*, 19(8), 2056–2063.
- Zhang, R. (2012). *Identification and biological characteristics of the causal organisms of soybean Fusarium root rot in Anhui (in Chinese)*. Master thesis. Anhui Agricultural University.
- Zhang, N., Castlebury, L. A., Miller, A. N., Huhndorf, S. M., Schoch, C. L., Seifert, K. A., ... & Sung, G. H. (2006). An overview of the systematics of the Sordariomycetes based on a four-gene phylogeny. *Mycologia*, 98(6), 1076–1087. <https://doi.org/10.1080/15572536.2006.11832635>
- Zhang, J., Cui, Y., Duan, C., Wu, X., Wang, X., & Zhu, Z. (2009). Identification of pathogen causing soybean charcoal rot (in Chinese). *Acta Agriculturae Boreali-Sinica*, 24, 192–196.
- Zhang, H., Wu, J., Zhou, W., Liu, Y., ... & Su, X. (2012). Occurrence characteristics and comprehensive control of major maize diseases, insect pests and weeds in Wafangdian City (in Chinese). *Modern Agriculture and Technology*, 2012(24), 147–147.
- Zhang, X. X., Sun, H. Y., Shen, C. M., Li, W., Yu, H. S., & Chen, H. G. (2015). Survey of *Fusarium* spp. causing wheat crown rot in major winter wheat growing regions of China. *Plant Disease*, 99(11), 1610–1615. <https://doi.org/10.1094/PDIS-04-14-0422-RE>
- Zhang, W., Cao, G., Li, X., Zhang, H., Wang, C., Liu, Q., Chen, X., Cui, Z., Shen, J., & Jiang, R. (2016). Closing yield gaps in China by empowering smallholder farmers. *Nature*, 537, 671–674. <https://doi.org/10.1038/nature19368>
- Zhang, L., Kou, K., Liu, Z., Wei, N., & Li, X. (2020). Occurrence characteristics and control strategies of wheat *Fusarium* crown rot in Cangzhou City (in Chinese). *Agriculture and Technology*, 40(12), 74–75.
- Zhao, X., Jin, S., Li, Z., Niu, N., & Wang, Y. (2015). Occurrence and control of soybean seedborne diseases in Hebei Province (in Chinese). *Journal of Hebei Agricultural Sciences*, 19(4), 46–48.
- Zhao, Y., Qin, F., Xu, F., Ma, J., Sun, Z., Song, Y., ... & Wang, H. (2019). Identification of *Tilletia foetida*, *Ustilago tritici*, and *Urocystis tritici* based on near-infrared spectroscopy. *Journal of Spectroscopy*, 2019. <https://doi.org/10.1155/2019/9753829>
- Zhen, W., Wang, S., Zhang, C., & Ma, Z. (2009). Influence of maize straw amendment on soil-borne diseases of winter wheat. *Frontiers of Agriculture in China*, 3, 7–12. <https://doi.org/10.1007/s11703-009-0003-4>
- Zhong, Z., Marcel, T. C., Hartmann, F. E., Ma, X., Plissonneau, C., Zala, M., ... & Palma-Guerrero, J. (2017). A small secreted protein in *Zymoseptoria tritici* is responsible for avirulence on wheat cultivars carrying the Stb6 resistance gene. *New Phytologist*, 214(2), 619–631. <https://doi.org/10.1111/nph.14434>
- Zhou, H. (2014). *The pathogen identification and pathogenicity of crown rot diseases in Huanghuai winter wheat growing region of China (in Chinese)*. Master thesis. Henan Agricultural University.
- Zhou, H., He, X., Wang, S., Ma, Q., & Li, H. (2019). Diversity of the *Fusarium* pathogens associated with crown rot in the Huanghuai wheat-growing region of China. *Environmental Microbiology*, 21(8), 2740–2754.
- Zhou, F., Hu, H., Fan, Y., Wang, J., Guo, Z., & Li, C. (2021). Identification of crown rot in wheat from Henan province and its sensitivity to 13 fungicides (in Chinese). *Journal of Henan Institute of Science and Technology (Natural Science Edition)*, 49(1), 1–13.
- Zhu, A. (2012). Main soybean diseases and comprehensive control techniques in Shangqiu City (in Chinese). *Bulletin of Agricultural Science and Technology*, 10, 158–160.
- Zhu, Y., Chen, H., Fan, J., Wang, Y., Li, Y., Chen, J., ... & Mundt, C. C. (2000). Genetic diversity and disease control in rice. *Nature*, 406(6797), 718–722. <https://doi.org/10.1038/35021046>

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