# Marker imputation and linkage map construction using a skim sequenced diploid potato population

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# **Background**

With the low cost of next-generation sequencing and the development of algorithm adapted to outcrossing crops [1], genotyping-by-sequencing strategies based on skim sequencing and imputation have become increasingly popular. Unfortunately low depth skim sequencing results predominantly in missing values. To overcome this problem we implemented in R an imputation pipeline based on clustering of parent specific SNPs in bins of 100 kb. We applied it to a population of 1536 full-sibs descending from heterozygous diploid potato sequenced at an average read depth 1.7x.

### Results

- We imputed 4893 female and 4735 male segregating markers and used these to construct parental linkage maps.
- Linkage maps displayed high collinearity with the physical map, except for recently discovered 5.8-Mb inversion on chr03 [2].
- Local failure of imputation coincided with low parent specific SNP density (e.g. male chr07) or extreme skewed segregation (e.g. male chr01).
- Skewed segregation was identified on eight chromosomes. Some distortion colocalized with known loci (e.g. S-locus, StSIEL) [3].
- Using the imputed markers in a single marker regression model we identified a major QTL for yield colocalizing with *StCDF1*.

### **Methods**

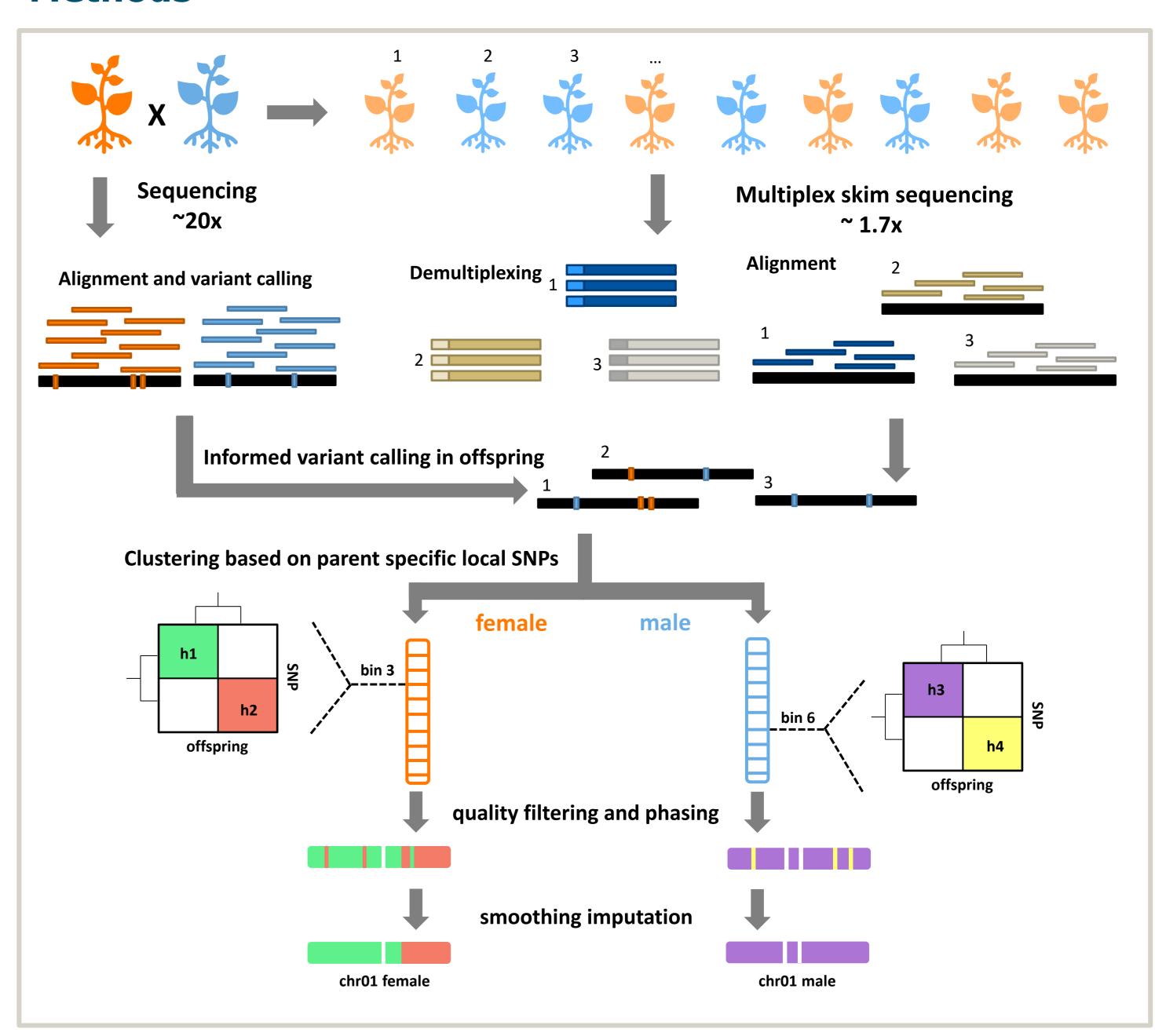


Figure 1. Imputation pipeline (modified from OutcrossSeq [1])

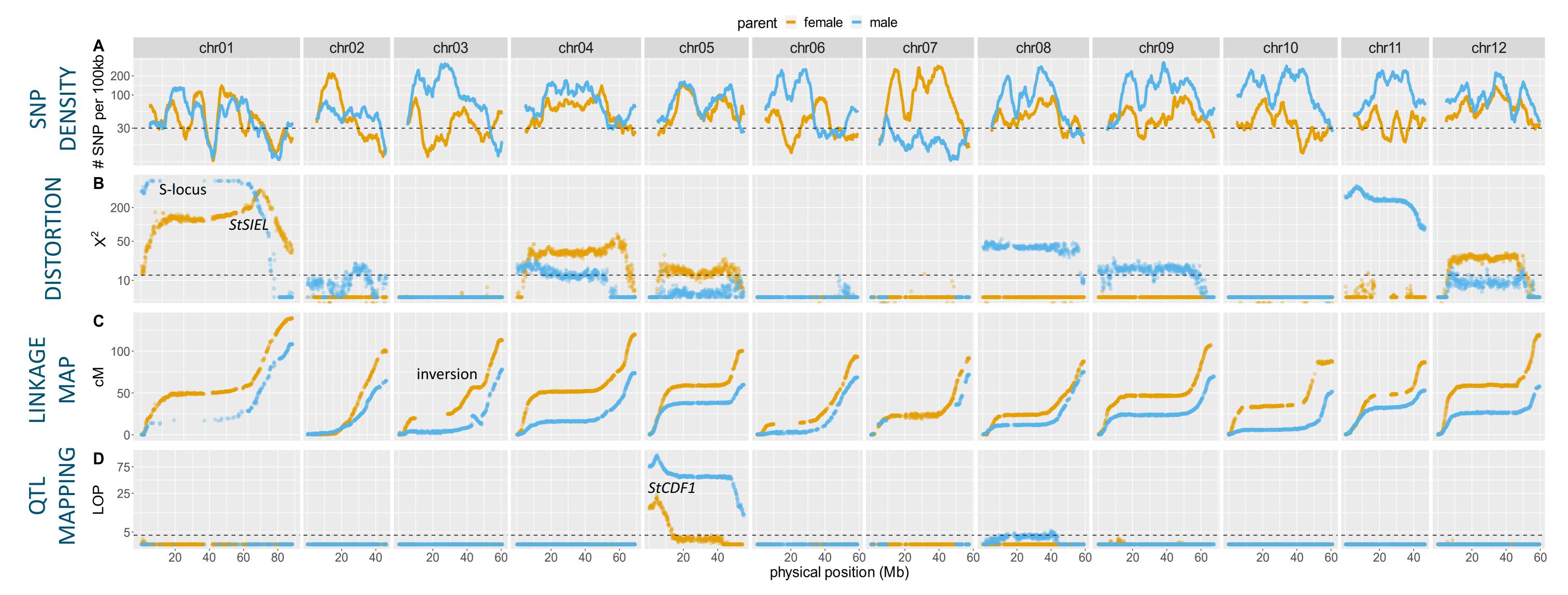


Figure 2. A. Number of parent specific SNPs per bins of 100kb used for imputation (moving average over 5Mbp). B. Deviation from Mendelian segregation ratio measured by chi-square goodness-of-fit test. C. Relationship between physical and genetic distance. D. -log(p-value) of single marker regression for yield in greenhouse (n=1534).

# Conclusion

The pipeline described in this poster facilitates the imputation of genotyping data from skim sequenced full-sib diploid populations. The resulting imputed markers can be used to construct high density linkage maps, explore transmission ratio distortion and identify loci regulating important traits.

# References

[1] Chen, M., Fan, W., Ji, F., Hua, H., Liu, J., Yan, M., Ma, Q., Fan, J., Wang, Q., Zhang, S. and Liu, G., 2021. Genome-wide identification of agronomically important genes in outcrossing crops using OutcrossSeq. Molecular plant, 14(4), pp.556-570.

[2] Tang, D., Jia, Y., Zhang, J., Li, H., Cheng, L., Wang, P., Bao, Z., Liu, Z., Feng, S., Zhu, X. and Li, D., 2022. Genome evolution and diversity of wild and cultivated potatoes. Nature, pp.1-7.

[3] Zhang, C., Wang, P., Tang, D., Yang, Z., Lu, F., Qi, J., Tawari, N.R., Shang, Y., Li, C. and Huang, S., 2019. The genetic basis of inbreeding depression in potato. Nature Genetics, 51(3), pp.374-378.



