



Draft Genome Sequences of the *Vibrio parahaemolyticus* Strains VHT1 and VHT2, Pasteurization-Resistant Isolates from Environmental Seafood

Guadalupe Meza,^{a,b} Hussain Majrshi,^b Kumar Saurabh Singh,^c  Ratnakar Deole,^d  Hung King Tiong^b

^aDepartment of Biological and Environmental Sciences, University of West Alabama, Livingston, Alabama, USA

^bClinical Laboratory, Huntsville Hospital, Huntsville, Alabama, USA

^cBioinformatics Group, Wageningen University, Wageningen, The Netherlands

^dOklahoma State University, Center for Health Sciences, Department of Biochemistry and Microbiology, Tulsa, Oklahoma, USA

ABSTRACT Two pasteurization-resistant strains, VHT1 and VHT2, of environmental, viable but nonculturable, pathogenic *Vibrio parahaemolyticus* were isolated from environmental oysters. Their whole-genome sequences were constructed. The genome sizes for VHT1 and VHT2 are 5.11 Mbp and 5.26 Mbp, respectively.

Vibrio parahaemolyticus is an opportunistic, viable but nonculturable (VBNC), seafood-borne pathogenic bacterium (1) that has been closely monitored by the CDC's Cholera and Other *Vibrio* Illness Surveillance (COVIS) system since 1989 and that is frequently difficult to detect in the food system (2). In the present study, two VBNC, pasteurization-resistant strains of environmental *V. parahaemolyticus*, VHT1 and VHT2 (3), were analyzed and their whole-genome sequences determined. Here, we present the draft genome sequences of *V. parahaemolyticus* VHT1 and VHT2.

Previous work by Meza et al. (3) used a modified two-step enrichment technique to recover VHT1 and VHT2 from environmental oysters. Briefly, *Vibrio* contaminants in a 25-g sample were homogenized in 225 g saline peptone water (SPW) in a stomacher bag with a Stomacher 400 Circulator lab blender and incubated for 48 h for enrichment at 35°C. Subsequently, the enrichment solution was heated at 80°C for 20 min and refrigerated overnight, and culture dilutions made in sterile buffered peptone water (BPW) were plated onto thiosulfate-citrate-bile salts-sucrose agar medium, followed by incubation for 72 h at 35°C. Visible green colonies of presumptive *V. parahaemolyticus* were then streaked onto Brain Heart Infusion (BHI) agar supplemented with 3% NaCl for purification, and the pure cultures were resuspended in BHI storage solution before being stored at -70°C. Bacterial 16S sequences (400 to 600 nucleotides) were identified using the NCBI nucleotide BLAST (blastn) platform. The sequences matched those of *V. parahaemolyticus* strain DHO76 (GenBank accession no. [CP066246.1](https://doi.org/10.1093/genbank/CP066246.1)) perfectly.

Using the Qiagen DNeasy blood and tissue kit, the VHT1 and VHT2 genomes were extracted from the fresh cultures prepared in BHI broth supplemented with 3% NaCl, followed by overnight incubation at 35°C and a repeated subculture (3). The DNA extracts were analyzed using the Quant-iT PicoGreen double-stranded DNA (dsDNA) assay kit coupled with the Qubit fluorometric quantifier and the NanoDrop spectrophotometer (DNA quantity and purity measurement). Paired-end libraries with 2,930,163 (VHT1) or 2,627,568 (VHT2) reads were generated using the Nextera XT DNA sample preparation kit, followed by whole-genome sequencing (WGS) using the Illumina MiSeq platform (2 × 250 bp, with the MiSeq reagent Nano kit v2) at Molecular Cloning Laboratories. FastQC genome reads with quality (Phred score) values of >20 (4) were used for subsequent processing with Trimmomatic v. 0.32 (5). Genome assemblies were constructed using SPAdes v. 3.13.0 (6) and evaluated using the basic metrics listed in Table 1. Annotations of genome assemblies and putative pathways were

Editor Frank J. Stewart, Montana State University

Copyright © 2022 Meza et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Hung King Tiong, htiong@uwa.edu.

The authors declare no conflict of interest.

Received 9 August 2022

Accepted 29 September 2022

Published 17 October 2022

TABLE 1 Assembly and annotation metrics of *V. parahaemolyticus* VHT1 and VHT2

Statistic	Data for strain:	
	VHT1	VHT2
No. of quality-filtered paired-end reads	2,930,163	2,627,568
No. of scaffolds	57	271
N_{50} (bp)	807,591	531,181
Largest scaffold length (bp)	1,194,203	1,737,051
Assembly length (bp)	5,115,539	5,261,907
Avg genome coverage (×)	181.3	112.5
G+C content (%)	45.24	45.12
No. of genes	4,769	5,141
No. of protein-coding genes	4,586	4,909
No. of rRNA genes	18	17
No. of tRNA genes	114	119
No. of pseudogenes	47	92
No. of genes associated with KEGG pathways	1,009	985
No. of genes associated with KEGG orthology	2,457	2,415
GenBank accession no.	JACERE000000000	JACERD000000000
SRA accession no.	SRR12395870	SRR12395869
BioProject accession no.	PRJNA608845	PRJNA608845

carried out using the NCBI Prokaryotic Genome Annotation Pipeline (PGAP) v. 4.12 (7) and KEGG tools (8), respectively, and are listed in Table 1. Default parameters were used for all software unless otherwise specified.

The sequencing data statistics are tabulated in Table 1. The genome sizes of both VHT1 and VHT2 are within the range of those for *V. parahaemolyticus* variant strains, namely, between 4.29 and 6.47 Mbp (NCBI; accessed 10 July 2021), and they have G+C contents of 45.24% (VHT1) and 45.12% (VHT2).

Data availability. This paired-end whole-genome MiSeq project has been deposited at GenBank under the BioProject accession no. [PRJNA608845](#) (VHT1 and VHT2). The reads have been deposited in the SRA under the accession no. [SRR12395870](#) (VHT1) and [SRR12395869](#) (VHT2). The 16S rRNA gene sequences were deposited at the NCBI under the GenBank accession no. [OP090156](#) (VHT1) and [OP106961](#) (VHT2). Bacterial culture material, genomic, and identification details are available at <https://doi.org/10.6084/m9.figshare.20457063>.

ACKNOWLEDGMENTS

The University of West Alabama (UWA) Office of Sponsor Program and Research (OSPR) Faculty seed grants supported the sample collection and characterization work. Oklahoma State University faculty startup funds supported the WGS work.

We thank Kevin Morse (UWA Department of Biological and Environmental Sciences faculty) for manuscript review.

REFERENCES

- Mizunoe Y, Wai SN, Ishikawa T, Takade A, Yoshida S. 2000. Resuscitation of viable but nonculturable cells of *Vibrio parahaemolyticus* induced at low temperature under starvation. *FEMS Microbiol Lett* 186:115–120. <https://doi.org/10.1111/j.1574-6968.2000.tb09091.x>.
- Centers for Disease Control and Prevention (CDC). 2019. *Vibrio* species causing vibriosis. <https://www.cdc.gov/vibrio/faq.html>. Accessed 29 May 2021.
- Meza G, Majrshi H, Tiong HK. 2022. Recovery of pasteurization-resistant *Vibrio parahaemolyticus* from seafoods using a modified, two-step enrichment. *Foods* 11:764. <https://doi.org/10.3390/foods11050764>.
- Wingett SW, Andrews S. 2018. FastQ Screen: a tool for multi-genome mapping and quality control. *F1000Res* 7:1338. <https://doi.org/10.12688/f1000research.15931.2>.
- Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. *Bioinformatics* 30:2114–2120. <https://doi.org/10.1093/bioinformatics/btu170>.
- Nurk S, Bankevich A, Antipov D, Gurevich A, Korobeynikov A, Lapidus A, Prjibelsky A, Pyskhin A, Sirotkin A, Sirotkin Y, Stepanauskas R. 2013. Assembling genomes and mini-metagenomes from highly chimeric reads, p 158–170. *In* Deng M, Jiang R, Sun F, Zhang X (ed), *Research in computational molecular biology*. RECOMB 2013. Lecture notes in computer science, vol 7821. Springer, Berlin, Germany. https://doi.org/10.1007/978-3-642-37195-0_13.
- Tatusova T, DiCuccio M, Badretdin A, Chetvermin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI Prokaryotic Genome Annotation Pipeline. *Nucleic Acids Res* 44:6614–6624. <https://doi.org/10.1093/nar/gkw569>.
- Kanehisa M, Sato Y, Morishima K. 2016. BlastKOALA and GhostKOALA: KEGG tools for functional characterization of genome and metagenome sequences. *J Mol Biol* 428:726–731. <https://doi.org/10.1016/j.jmb.2015.11.006>.