



OPEN ACCESS

EDITED BY

Gloria A. Otunola,
University of Fort Hare, South Africa

REVIEWED BY

Birsen Yilmaz,
Çukurova University, Turkey
Gordana Kendel Jovanovic,
Teaching Institute of Public Health
of Primorsko-Goranska County,
Croatia
Sanja Majanovic,
University of Rijeka, Croatia

*CORRESPONDENCE

Katherine J. Li
katherine.li@wur.nl

SPECIALTY SECTION

This article was submitted to
Food Chemistry,
a section of the journal
Frontiers in Nutrition

RECEIVED 22 June 2022

ACCEPTED 30 August 2022

PUBLISHED 20 September 2022

CITATION

Li KJ, Burton-Pimentel KJ, Vergères G,
Feskens EJM and Brouwer-Brolsma EM
(2022) Fermented foods
and cardiometabolic health:
Definitions, current evidence,
and future perspectives.
Front. Nutr. 9:976020.
doi: 10.3389/fnut.2022.976020

COPYRIGHT

© 2022 Li, Burton-Pimentel, Vergères,
Feskens and Brouwer-Brolsma. This is
an open-access article distributed
under the terms of the [Creative
Commons Attribution License \(CC BY\)](https://creativecommons.org/licenses/by/4.0/).
The use, distribution or reproduction in
other forums is permitted, provided
the original author(s) and the copyright
owner(s) are credited and that the
original publication in this journal is
cited, in accordance with accepted
academic practice. No use, distribution
or reproduction is permitted which
does not comply with these terms.

Fermented foods and cardiometabolic health: Definitions, current evidence, and future perspectives

Katherine J. Li^{1,2*}, Kathryn J. Burton-Pimentel²,
Guy Vergères², Edith J. M. Feskens¹ and
Elske M. Brouwer-Brolsma¹

¹Division of Human Nutrition and Health, Wageningen University & Research, Wageningen, Netherlands, ²Agroscope, Bern, Switzerland

Unhealthy diets contribute to the increasing burden of non-communicable diseases. Annually, over 11 million deaths worldwide are attributed to dietary risk factors, with the vast majority of deaths resulting from cardiometabolic diseases (CMDs) including cardiovascular disease (~10 million) and type II diabetes (~339,000). As such, defining diets and dietary patterns that mitigate CMD risk is of great public health importance. Recently, the consumption of fermented foods has emerged as an important dietary strategy for improving cardiometabolic health. Fermented foods have been present in the human diet for over 10,000 years, but knowledge on whether their consumption benefits human health, and the molecular and microbiological mechanisms underpinning their purported health benefits, is relatively nascent. This review provides an overview of the definitions of fermented foods, types and qualities of fermented foods consumed in Europe and globally, possible mechanisms between the consumption of fermented foods and cardiometabolic health, as well as the current state of the epidemiological evidence on fermented food intake and cardiometabolic health. Finally, we outline future perspectives and opportunities for improving the role of fermented foods in human diets.

KEYWORDS

fermented foods, microorganisms, cardiometabolic disease, biomarkers, dietary assessment

Introduction

What is a fermented food?

For millennia, fermentation has been used as an effective method of food preservation and alcohol production (1). Fermentation of raw agricultural products can improve their nutritional qualities as well as impart new aromas and tastes. However, the increase in the popularity of fermented foods over the past decades has led to widespread

misconceptions about what is required for a food to be considered “fermented.” Several definitions of fermented foods have been proposed over the years. One of the earlier documented descriptions, by Steinkraus (2), captures the biological complexity and transformative nature of food fermentation: “Fermented foods are food substrates that are invaded or overgrown by edible microorganisms whose enzymes, particularly amylases, proteases and lipases hydrolyze the polysaccharides, proteins and lipids to non-toxic products with flavors, aromas and textures pleasant and attractive to the human consumer.” However, this definition does not capture the intentional nature of food fermentation processes, and seemingly limits the transformative components in food to macronutrients. In 2021, the International Scientific Association for Probiotics and Prebiotics (ISAPP) provided a consensus statement on fermented foods, broadly defining fermented foods as: “Foods made through desired microbial growth and enzymatic conversions of food components” (3). Under this definition, fermented foods are those formed through a controlled process involving “desired” microorganisms. Importantly, the broad reference to conversions of “food components” suggests that the fermentation process could generate novel dietary compounds with distinct functional properties.

All fermented foods are procured *via* the actions of fermentative microorganisms. Microorganisms naturally present in the raw food matrix or the surrounding environment can initiate “spontaneous” fermentations, such as during the fermentation of cocoa beans using indigenous yeasts, lactic acid bacteria (LAB), and acetic acid bacteria (AAB) (4). However, large-scale industrial fermentations, which are commonplace, typically use starter cultures to ensure consistency in the end food product (5). Advances in food technology has also led to the application of alternative processing of foods that were traditionally fermented, resulting in non-fermented products, notably: pickled vegetables preserved in vinegar or brine, meat or fish preserved using salt, bread leavened using baking powder, and fresh cheeses curdled with vinegar or citric acid (Table 1).

While the consumption of fermented foods tends to be synonymous with consuming a “dose” of live microorganisms, this is not always the case. The presence of live microorganisms in the ready-to-consume product is dependent on several factors: the phase of the fermentation (e.g., fermentation of cocoa beans in the food preparation phase versus fermentation of yogurt in the final product phase), whether the fermented food is heat-treated or if the microorganisms are intentionally removed (e.g., filtration of wine), as well as personal food preferences (e.g., cooked sauerkraut consumed in the Netherlands) (6). These factors create a key delineation (i.e., presence/absence of microorganisms) for the classification of different types of fermented foods with relevance to their health impacts.

TABLE 1 Classification of fermented foods adapted from Marco et al. (3).

Fermented

Live microorganisms present

- Yogurt
- Sour cream
- Kefir
- Most cheeses
- Miso
- Natto
- Tempeh
- Non-heated fermented vegetables
- Non-heated salami, pepperoni and other fermented sausages
- Boza, bushera and other fermented cereals
- Most kombuchas
- Some beers

Live microorganisms absent

- Bread
- Heat-treated or pasteurized fermented vegetables, sausage, soy sauce, vinegar, and some kombuchas
- Wine, most beers, and distilled spirits
- Coffee and chocolate beans (after roasting)
- Chocolate (after heat processing)

Not fermented

- Chemically leavened bread
- Fresh sausage
- Vegetables pickled in brine and/or vinegar
- Chemically produced soy sauce
- Salted or cured processed meats and fish

Fermented foods as a source of live microorganisms

In Western societies, the resurgent interest in the consumption of fermented foods can be credited to the explosion of research into the human microbiome (7, 8). Several studies have demonstrated that diet influences the structure and function of the gut microbiota (9, 10). It is believed that the consumption of fermented foods containing probiotics – “live microorganisms which, when administered in adequate amounts, confer a health benefit on the host” (11) – is an effective way to introduce potentially beneficial microorganisms to the intestinal tract and help manage a wide range of disorders associated with gut microbial dysbiosis. These include both intestinal disorders, such as behavior and (gut-)brain disorders, inflammatory bowel disease, irritable bowel syndrome, and coeliac disease, as well as extra-intestinal disorders, including allergy, asthma, obesity, metabolic syndrome, and cardiovascular disease (CVD) (12, 13).

The diversity of microorganisms found in fermented foods produced globally, as well as their functional properties, have been the subject of several comprehensive reviews (14, 15). Primarily, these include gram-positive (particularly LAB) and gram-negative bacteria, filamentous molds, and enzyme-

and alcohol-producing yeasts (14). These microorganisms and their enzymes have varied functional roles, such as acting as antimicrobial agents (16), antioxidants (17), and fibrinolytic agents (15, 18). Recent advances in (meta)genomic sequencing are further expanding our understanding of the microbial diversity and functional potential of fermented foods, in particular bacterial and fungal species that have been less well-characterized due to difficulties in culturing these species (8, 19).

Another important aspect affecting the health impact of fermented foods is the amount of live microorganisms provided by the consumption of fermented foods. In a review by Rezac et al. (6), many fermented foods (cheese, yogurt, sausages, vegetables, cereals, sour beer, kombucha, fermented fish, and tempeh) were found to contain 10^5 – 10^7 colony forming units (CFU) of LAB/(mL or g), with cultured dairy products containing up to 10^9 CFU/(mL or g). However, considerable variation was observed based on geographical region and sampling time, in addition to the manufacturing, processing, and storage conditions (6). Recently, Marco et al. (20) classified foods from the U.S. National Health and Nutrition Examination Survey (NHANES) 2001–2018 based on expert opinions on their levels of live microorganisms: low ($< 10^4$ CFU/g), medium (10^4 – 10^7 CFU/g), and high ($> 10^7$ CFU/g). They found that over 50% of children and adults were consumers of foods containing high levels of live microorganisms, and that the proportions of people consuming live microorganisms and per capita intake increased significantly over time (20). Although guidelines are lacking for the minimum dose of live microorganisms that should be consumed, the European Union (EU) health claim for yogurt and “improved lactose tolerance” stipulates that at least 10^8 CFU of live starter microorganisms per gram of yogurt (21). Further, Derrien et al. (22) predicted that ingesting a dose of 10^{10} ingested bacterial cells would be sufficient to drastically shift the composition of the gut microbiota and impact the immune and neuroendocrine responses of the host.

Fermented foods as a source of fermentation-derived metabolites

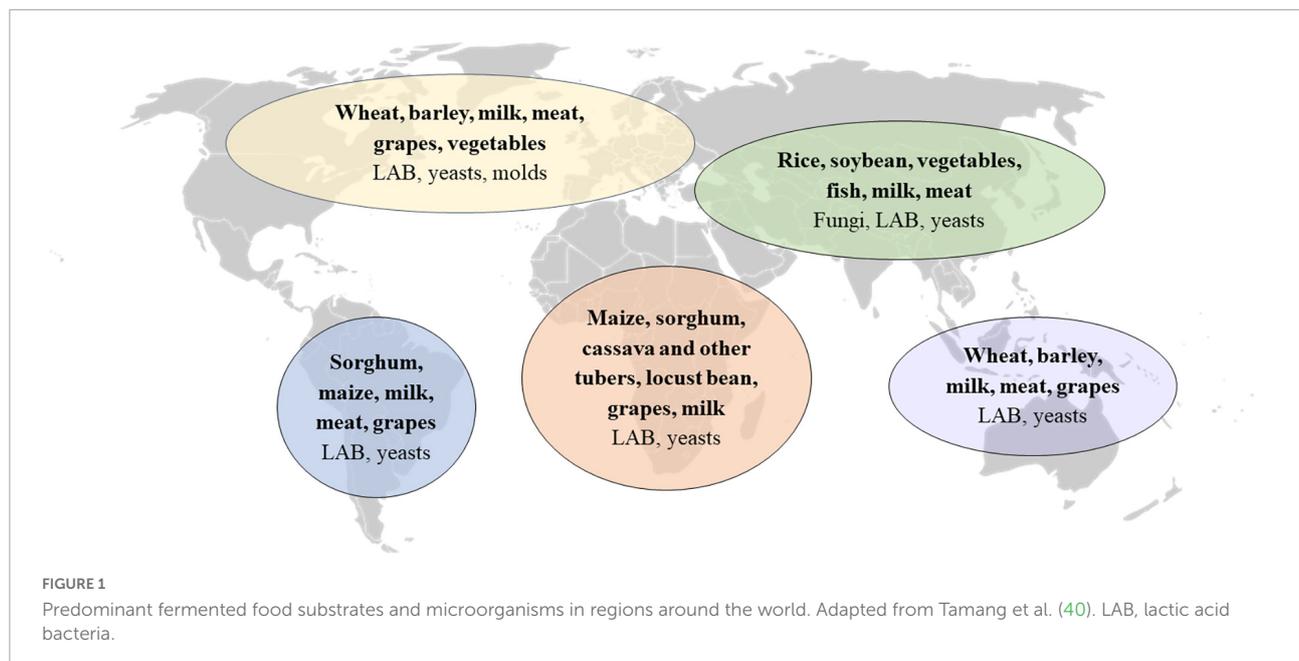
While not all fermented foods contain live microorganisms at the time of consumption, microbial activity during fermentation can still produce bioactive metabolites that could be beneficial to human health (23). The main fermentation processes can be grouped by the primary metabolites of fermenting microorganisms: alcohol and carbon dioxide produced by yeasts, acetic acid produced by AAB, lactic acid produced by LAB, ammonia and fatty acids produced by *Bacilli* and molds, and propionic acid produced by propionic acid bacteria (24). These metabolites parallel the end-products of the fermentation of undigested carbohydrate and protein by the gut microbiota, some of which (e.g., organic acids) have been

positively associated with host gastrointestinal and immune health, lipid and protein metabolism, and appetite control (25).

Additionally, various secondary “bioactive” metabolites produced during fermentation are receiving increasing scientific interest for their functional properties. The types of metabolites produced depends on both the substrate and type of fermentation. For example, fermentation of milk results in the production of α 1- and β -casein peptide fragments from milk casein, which have been detected in several varieties of cheese (26). These bioactive peptides have angiotensin-converting enzyme (ACE) inhibitory activity and have also been reported to modulate opioid receptors in the gut epithelium (26). On the other hand, fermentation of multiple foods (e.g., cheese, sauerkraut) *via* a common fermentation pathway using LAB generates phenyllactic acids which help with food preservation as well as serve a physiological role of immune modulation (27, 28). Gamma-aminobutyric acid (GABA), which is enriched in sourdough bread through fermentation with LAB, has been shown to help control blood pressure and protect against CVD (29). Additionally, short-chain fatty acids (e.g., acetate, propionate, and butyrate) produced during microbial fermentation have shown hypocholesterolemic properties (30).

Fermentation to enhance the nutritional quality of foods

In addition to introducing novel fermentation-derived metabolites and directly influencing nutrients, fermentation can also enhance the nutritional composition of the end food product or improve nutrient bioavailability. Various fermented foods have been shown to have enhanced nutritional attributes compared to their non-fermented counterparts (31–34). For example, it has been observed that levels of flavonoids, anthocyanins, and triterpenoids progressively increased during the fermentation of raw radishes, beets, and peppers (32). Fermentation of milk into cheese and yogurt has been shown to increase the levels of free amino acids detected in plasma, including α -amino butyric acid, alanine, asparagine, cysteine, glycine, glutamine, histidine, isoleucine, leucine, lysine, methionine, ornithine, phenylalanine, proline, serine, threonine, tryptophan, tyrosine, and valine (33, 34). Many micronutrients (calcium, phosphorus, A and B vitamins, potassium, zinc, and choline) also have a higher bioavailability in yogurt than in raw milk due to the acidity and fermentation process (35). Further, fermentation can also inactivate toxic dietary components and degrade anti-nutritional factors (36). Phytates present in cereals, legumes, and tubers are decreased during fermentation as a result of the activity of microbial phytases (36). The combination of fermentation and cooking also inactivates lectins from legumes that hinder nutrient absorption from the gastrointestinal tract (36).



Fermented foods consumed in global and European diets

Over 5,000 types of fermented foods and beverages have been estimated to exist worldwide, contributing to 5–40% of the human diet (3, 37, 38). Every region and culture produces distinct fermented foods based largely on accessibility to different raw materials (Figure 1). The specific types and qualities of fermented foods consumed in different cultures around the world have been the subject of several comprehensive reviews (14, 39, 40). In the present review, we focus on fermented foods consumed in Europe.

In Europe, fermented foods were traditionally derived from fermentation of milk, wheat, barley, meat, grapes, and vegetables. Multiple varieties of cheeses, wine, beer, and dried/salted meats (including fermented meats) are designated as goods with “specific geographical origin” in the EU and have distinct historical and cultural significance (41). Several other fermented foods, such as coffee and chocolate, have origins in other cultures but have been normalized in European diets from ancient trade routes or colonization. It is expected that the varieties of fermented foods in European diets will continue to expand with globalization and consumer tastes for healthy and exotic flavors. A non-exhaustive overview of the types of fermented foods commonly consumed in modern European diets, as well as their raw materials and fermenting microorganisms, is provided in Table 2. It should be noted that although spirits (e.g., brandy, gin, whisky) are also technically considered to be fermented beverages, they were not further evaluated in this review, as their high alcohol content dominates the presence of other metabolites

of fermentation that are interesting from a nutritional point of view.

In a recent quantitative evaluation of food lists from food frequency questionnaires and 24-h recalls, it was determined that approximately 20% of the Dutch adult diet comprises fermented food items (42). These estimates are likely to reflect the prevalence of fermented food consumption in other European countries with similar diets. The fermented foods consumed primarily consisted of coffee, beer, wine, yeast-leavened bread products, chocolate, cheese, yogurt, quark, and buttermilk. Several less commonly consumed fermented foods were also captured by 24-h recalls, including certain fermented dairy products (sour cream, crème fraîche, yakult), sausages (salami and chorizo), fermented fish (salted herring, shrimp paste), vegetables (sauerkraut, fermented pickled vegetables), soy (miso, tempeh, soy sauce), and yeast-fermented desserts (doughnuts, pastries).

The evidence between fermented foods and cardiometabolic health

Proposed mechanisms

Several mechanisms have been proposed that support a role of fermented foods in promoting cardiometabolic health (Figure 2). As mentioned previously, fermented foods containing live microorganisms at the time of consumption may provide a source of probiotics, which can modulate both the composition and function of the host’s gut microbiota (22). Changes in gut microbial composition could enhance

TABLE 2 Substrates and microorganisms of predominant fermented foods consumed in European diets.

Fermented food	Substrate ^a	Fermenting microorganisms	Live microorganisms at time of consumption	References
Cheese (Afuega'l Pitu, Armada, Asiago, blue cheese, Brie, Burgos, Cabrales, Camembert, Cheddar, Comte, Danbo, Edam, Feta, Fontina, Galotyri, Gorgonzola, Gouda, Gubeen, Grana Padano, Havarti, Livarot, Limburger, Manchego, Monterey Jack, Mozzarella, Muenster, Parmesan, Puzzone di Moena, Pecorino Romano, Provolone, Stilton, Swiss, Swiss Gruyere, Tilsit)	Milk (bovine)	<ul style="list-style-type: none"> • LAB: <i>Lb. paracasei</i>, <i>Lb. rhamnosus</i>, <i>Lb. delbrueckii</i> subsp. <i>Bulgarius</i>, <i>Lb. delbrueckii</i> subsp. <i>delbrueckii</i>, <i>Lb. delbrueckii</i> subsp. <i>lactis</i>, <i>Lb. helveticus</i>, <i>Lb. casei</i>, <i>Lb. plantarum</i>, <i>Lb. salivarius</i>, <i>Lc. lactis</i> subsp. <i>cremoris</i>, <i>Lc. lactis</i> subsp. <i>lactis</i>, <i>Leuc. spp.</i>, <i>Ent. spp.</i> (<i>Ent. durans</i>, <i>Ent. faecium</i>), <i>Strep. thermophilus</i> • Other gram-positive bacteria: <i>Staph. spp.</i>, <i>Brevibacterium linens</i>, <i>Propionibacterium freudenreichii</i> • Fungi: <i>Debaryomyces hansenii</i>, <i>Geotrichum candidum</i>, <i>P. camemberti</i>, <i>P. roqueforti</i> 	Yes (most)	(6, 14)
Yogurt	Milk (bovine)	<ul style="list-style-type: none"> • LAB: <i>S. thermophilus</i>, <i>Lb. bulgaricus</i>, <i>Lb. acidophilus</i>, <i>Lb. casei</i>, <i>Lb. rhamnosus</i>, <i>Lb. delbrueckii</i> subsp. <i>bulgaricus</i>, <i>Lb. gasseri</i>, <i>Lb. johnsonii</i> • Other gram-positive bacteria: <i>B. lactis</i>, <i>B. bifidum</i> 	Yes	(6, 14)
Buttermilk	Milk (bovine)	<ul style="list-style-type: none"> • LAB: <i>Lactococci</i>, <i>Lactobacilli</i> 	Yes	(1, 6)
Kefir	Milk (bovine, ewe, goat, buffalo), kefir grains	<ul style="list-style-type: none"> • LAB: <i>Lb. paracasei</i> ssp. <i>paracasei</i>, <i>Lb. acidophilus</i>, <i>Lb. delbrueckii</i> ssp. <i>bulgaricus</i>, <i>Lb. plantarum</i>, and <i>Lb. kefirifaciens</i>, <i>Lb. kefirii</i>, <i>Lb. brevis</i>, <i>Lb. casei</i> ssp. <i>pseudoplantarum</i>, <i>Lb. helveticus</i>, <i>Lb. lactis</i>, <i>Lb. lactis</i> ssp. <i>lactis</i>, <i>Enterococcus sp.</i>, <i>Lc. Lacti</i>, <i>Lc. lactis</i> ssp. <i>cremoris</i>, <i>Leuc. mesenteroides</i> • Other gram-positive bacteria: <i>Bacillus sp.</i>, <i>S. thermophilus</i> • AAB: <i>A. Aceti</i>, <i>A. rasens</i>, <i>A. fabarum</i>, <i>A. lovaniensis</i>, <i>A. orientalis</i>, <i>Gluconobacter frateurii</i> • Yeasts: <i>Sacc. cerevisiae</i>, <i>C. inconspicua</i>, <i>C. maris</i>, <i>Kluyveromyces marxianus</i>, <i>Kluyveromyces marxianus</i> var. <i>lactis</i>, <i>Dekkera anomala</i>, <i>Kazachstania kefir</i>, <i>Naumovozyma sp.</i> 	Yes	(116)
Bread (white, wholegrain, sourdough)	Barley, rye, wheat	<ul style="list-style-type: none"> • Yeasts: <i>Sacc. cerevisiae</i>, <i>C. humili</i>, <i>Issatchenkia orientalis</i> • LAB (sourdough only): <i>Lb. sanfranciscensis</i>, <i>Lb. alimentarius</i>, <i>Lb. buchneri</i>, <i>Lb. casei</i>, <i>Lb. delbrueckii</i>, <i>Lb. fructivorans</i>, <i>Lb. plantarum</i>, <i>Lb. reuteri</i>, <i>Lb. johnsonii</i> 	No	(14)
Dried fermented sausage (Salami, Salsiccia, Soppressata, Alheiras, Botillo, Chorizo, Salchicón, Pepperoni)	Pork or beef	<ul style="list-style-type: none"> • LAB: <i>Lb. plantarum</i>, <i>Lb. paraplantarum</i>, <i>Lb. brevis</i>, <i>Lb. rhamnosus</i>, <i>Lb. sakei</i>, <i>Lb. zeae</i>, <i>Lb. paracasei</i>, <i>Ent. faecalis</i>, <i>Ent. faecium</i>, <i>Leuc. mesenteroides</i>, <i>Ped. pentosaceus</i>, <i>Ped. acidilactici</i>, <i>W. cibaria</i>, <i>W. viridescens</i>, <i>Lb. sake</i>, <i>Lb. curvatus</i>, <i>Lb. plantarum</i> • Other gram-positive bacteria: <i>Micrococcus spp.</i>, <i>Staph. spp.</i> • Gram-negative bacteria: <i>Enterobacteriaceae</i> • Yeasts and molds 	Yes	(6)
Sauerkraut	Cabbage	<ul style="list-style-type: none"> • LAB: <i>Leuc. mesenteroides</i>, <i>Ped. pentosaceus</i>; <i>Lb. brevis</i>, <i>Lb. plantarum</i>, <i>Lb. sakei</i> 	Yes	(6, 14)
Fermented cucumbers	Cucumbers	<ul style="list-style-type: none"> • LAB: <i>Leuc. mesenteroides</i>, <i>Ped. cerevisiae</i>, <i>Ped. acidilactici</i>, <i>Lb. plantarum</i>, <i>Lb. brevis</i> 	Yes	(14)
Fermented olives	Olives	<ul style="list-style-type: none"> • LAB: <i>Leuc. mesenteroides</i>, <i>Ped. pentosaceus</i>; <i>Lb. plantarum</i> <i>Lb. pentosus</i>/<i>Lb. plantarum</i>, <i>Lb. paracollinoides</i>, <i>Lb. vaccinostercus</i>, <i>Lb. suebicus</i>, <i>Ped. sp.</i> • Other gram-positive bacteria: <i>Gordonia sp.</i> • Gram-negative bacteria: <i>Pseudomonas sp.</i>, <i>Sphingomonas sp.</i>, <i>Sphingobium sp.</i>, <i>Sphingopyxis sp.</i>, <i>Thalassomonas agarivorans</i>, • Yeasts: <i>C. cf. apicola</i>, <i>Pic. sp.</i>, <i>Pic. Manshurica</i>, <i>Pic. galeiformis</i>, <i>Sacc. cerevisiae</i> • Archaea: <i>Halorubrum orientalis</i>, <i>Halosarcina pallid</i> 	Yes	(14)
Coffee	Coffee cherries	<ul style="list-style-type: none"> • LAB: <i>Lc. Lactis</i>, <i>Leuc. Mesenteroides</i>, <i>Lb. plantarum</i>, <i>Lb. brevis</i> • Other gram-positive bacteria: <i>Bc. cereus</i>, <i>Bc. megaterium</i>, <i>Bc. subtilis</i>, <i>Bc. Macerans</i> 	No	(117)

(Continued)

TABLE 2 (Continued)

Fermented food	Substrate ^a	Fermenting microorganisms	Live microorganisms at time of consumption	References
		<ul style="list-style-type: none"> • Gram-negative bacteria: <i>Serratia</i> sp., <i>Enterobacter agglomerans</i>, <i>Klebsiella pneumonia</i>, <i>Erwinia herbicola</i>, <i>Acinetobacter</i> sp., <i>Escherichia coli</i> • Yeasts: <i>Pic. anomala</i>, <i>Torulasporea delbrueckii</i>, <i>Rhodotorula mucilaginosa</i>, <i>C. ernobii</i>, <i>C. carpophila</i>, <i>Saccharomyces</i> sp., <i>Pic. caribbica</i>, <i>C. membranifaciens</i>, <i>Arxula</i> sp., <i>Hanseniaspora uvarum</i>, <i>Kluyveromyces</i> sp., <i>Kloeckera</i> sp., <i>C. xestobii</i> 		
Chocolate	Cocoa pods	<ul style="list-style-type: none"> • AAB: <i>A. pasteurianus</i>, <i>A. senegalensis</i> • LAB: <i>Lb. fermentum</i>, <i>Lb. ghanensis</i>, <i>Lb. brevis</i>, <i>Leuc. mesenteroides</i>, <i>Leuc. pseudomesenteroides</i>, <i>W. ghanensis</i>, <i>Lb. cacaonum</i>, <i>Lb. fabifermentans</i>, <i>W. fabaria</i>, <i>Fructobacillus pseudoficulneus</i>, <i>Lb. plantarum</i> • Gram-negative bacteria: <i>Enterobacteria</i>, <i>Tatumella ptyseos</i>, <i>Tatumella citrea</i> • Yeasts: <i>Sacc. cerevisiae</i>, <i>Kluyveromyces</i>, <i>Hanseniaspora uvarum</i>, <i>Hanseniaspora guilliermondii</i>, <i>Issatchenkia orientalis</i> (<i>C. krusei</i>), <i>Pic. membranifaciens</i> 	No	(14, 118)
Wine	Grapes	<ul style="list-style-type: none"> • Yeasts: <i>Sacc. cerevisiae</i>, <i>C. colliculosa</i>, <i>C. stellata</i>, <i>Hanseniaspora uvarum</i>, <i>Kloeckera apiculata</i>, <i>Kl. thermotolerans</i>, <i>Torulasporea delbrueckii</i>, <i>Metschnikowia pulcherrima</i>, <i>Candida</i> sp. and <i>Cladosporium</i> sp. 	No	(14)
Beer	Barley, hops	<ul style="list-style-type: none"> • Yeasts: <i>Sacc. cerevisiae</i>, <i>Sacc. carlsbergensis</i>, <i>Sacc. pastorianus</i> 	No	(14)

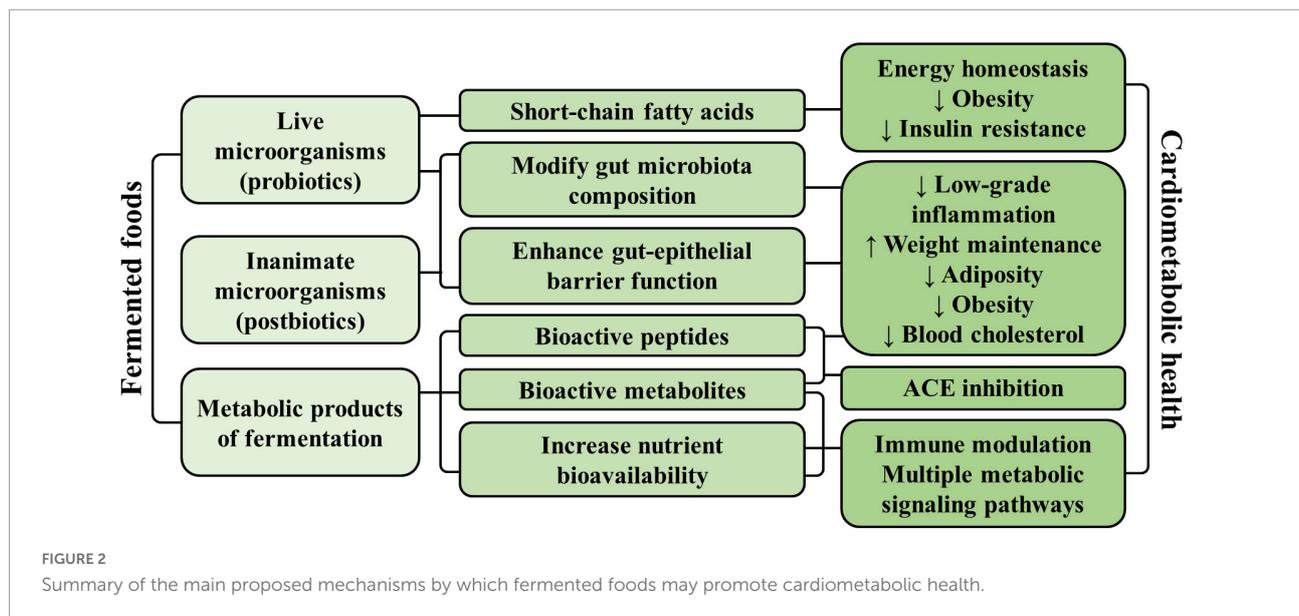
A., *Acetobacter*; AAB, acetic acid bacteria; B., *Bifidobacterium*; Bc., *Bacillus*; C., *Candida*; LAB, lactic acid bacteria; Ent., *Enterococcus*; Lb, *Lactobacillus*; Lc, *Lactococcus*; Leuc. *Leuconostoc*; P., *Penicillium*; Ped., *Pediococcus*; Pic., *Pichia*; S, *Streptococcus*; Sacc., *Saccharomyces*; Staph., *Staphylococcus*; W., *Weissella*. ^aMost common substrates listed, other substrates can also be used.

the integrity of the intestinal barrier and reduce low-grade inflammation associated with endotoxemia, which is speculated to be a mediator of obesity-related diseases (43). In animal models, consumption of dairy products with probiotics demonstrated greater cardiometabolic health benefits compared to consumption of dairy products without probiotics. In one such study, C57BL/6 mice on high-fat diets given kefir (a fermented dairy product) had reduced weight gain, hepatic steatosis, and low-density lipoprotein (LDL)-cholesterol levels compared to mice given milk (44). Mice given kefir also had higher levels of *Lactobacillus*, *Lactococcus*, total yeast, and *Candida* in the gut, which was strongly correlated with upregulated expression of fatty acid oxidation genes (*AOX*, *PPAR- α*) in both hepatic and adipose tissues. Reduced plasma levels of the pro-inflammatory cytokine IL-6 and down-regulation of the inflammation gene *MCPI* in adipose tissue was also observed. Evidence from several human trials also support a promising role for certain probiotic strains (primarily *Lactobacillus*) on weight maintenance, adiposity, obesity, and cholesterol levels, although further evidence is needed for their clinical relevance (45–49). Moreover, consumption of probiotics seems to modulate the function of the gut microbiota by increasing the production of short-chain fatty acids that impact energy homeostasis, obesity, and insulin resistance (50, 51). Given that there is a clear overlap between fermentation

products and microbial activities in fermented foods and the gut microbiota, the literature on the health impact of the gut microbiota feeds back into the potential health benefits of fermented foods.

On the other hand, fermented foods that do not contain live microorganisms at the time of consumption can still be a source of postbiotics, defined as a “preparation of inanimate microorganisms and/or their components that confers a health benefit on the host” (52). The mechanisms of the non-viable microorganisms have been postulated to be similar to probiotics, such as helping to modulate the gut microbiota and enhance epithelial barrier function, as well as modulating host immune, metabolic, and signaling responses (52). In mouse models, consumption of heat-inactivated postbiotic preparation consisting of *Limosilactobacillus fermentum* and *Lactobacillus delbrueckii* resulted in altered gut microbiota composition and intestinal structure, indicating that the inactivated microorganisms maintain biological activity (53).

Importantly, both fermented foods containing live probiotics or inactive postbiotics contain end-products of fermentation (i.e., metabolites derived from or enhanced by the fermentation process), which could modulate multiple metabolic signaling pathways to improve overall cardiometabolic health. Many fermented foods are produced using LAB, which generates lactic acid, short-chain fatty acids,



bioactive peptides, and polyamines with potential effects on cardiovascular, immune and metabolic health (52, 54). Fermenting bacteria can also influence cardiometabolic health by improving nutrient bioavailability, such as the bacterial production of vitamin K2 from vitamin K1, leading to a more potent activation of vitamin K-dependent proteins that affect multiple metabolic pathways (55). Other compounds may be present in fermented foods due to the presence of specific bacteria. For example, conjugated linoleic acid (CLA) that is associated with improved energy homeostasis (56) and which is already present in non-fermented dairy, may be elevated in fermented dairy foods due to the action of LAB or *Bifidobacteria* strains on linoleic acid (57).

Epidemiological evidence for fermented foods and cardiometabolic health

Total (bacterial) fermented food intake

In the EPIC-NL cohort ($n = 34,409$) (58), total bacterial fermented food intake [which includes 78% dairy (yogurt, buttermilk and quark), 16% cheese, 4% meat (dried sausage), 2% vegetables (sauerkraut, pickles, olives), 0.1% soya (tempeh), and 0.2% vinegar] was not significantly associated with both all-cause mortality [hazard ratio (HR) 1.00, 95% CI 0.88–1.13], and CVD-related mortality (HR 1.04, 95% CI 0.83–1.30). Subgroup analyses did show a moderate inverse association between cheese and CVD mortality, particularly from stroke (HR 0.59, 95% CI 0.38–0.92). In this study, fermented foods made using yeast as the main starter culture (bread, wine, beer, alcoholic drinks) or by endogenous enzymes/microorganisms (cocoa, coffee, tea) were intentionally excluded. Aside from this report,

no studies have investigated the impact of total fermented food consumption on CMDs and associated risk factors.

Fermented foods containing live microorganisms

In Europe, fermented dairy products comprise the largest group of fermented foods consumed that contain live microorganisms. Thus, numerous studies have examined the cardio-protective effects of fermented dairy foods, both in the context of and separately to total dairy intake. For instance, the Prospective Urban Rural Epidemiology (PURE) study ($n = 136,384$, 21 countries, five continents) (59) showed that higher intake of total dairy foods (> 2 servings/day versus no consumption) was associated with significantly lower risks of total CVD (HR 0.84, 95% CI 0.75–0.94), cardiovascular mortality (HR 95% CI 0.77, 0.58–1.01), major CVD events (HR 0.78, 95% CI 0.67–0.90), and stroke (HR 0.66, 95% CI 0.53–0.82), which seemed to be particularly driven by intakes of yogurt (HR total CVD 0.86, 95% CI 0.75–0.99) and milk (HR total CVD 0.90, 95% CI 0.82–0.99). In contrast, the Northern Sweden Health and Disease Study ($n = 108,065$) showed that higher intakes of milk were associated with an increased risk of myocardial infarction (HR 1.17, 95% CI 1.03–1.34) and type II diabetes (HR 1.23, 95% CI 1.10–1.37) in men, but not women (60). Intakes of fermented milk, butter and cheese were not significantly associated with these CVD-related outcomes. Stratified analysis by fat content suggested that lower fat dairy products were associated with increased risk of the CVD-related outcomes assessed. The authors hypothesized that this may be attributed to other factors (i.e., compensation of calories from dairy with that of other food groups, consumption of other foods in the diet in addition to low-fat dairy, or to possible cardio-protective effects of dairy fats). Effects of fermented dairy

TABLE 3 Summary of the systematic reviews and meta-analyses between dairy foods and various cardiometabolic disease risk parameters^a [adapted from Gille et al. (79)].

	Total dairy	Milk	Cheese	Yogurt
Prospective studies				
CVD	●	●	●	●
CAD/CHD	●	●	●	●
Stroke	●	●	●	●
Hypertension	●	●	●	●
Metabolic syndrome	●	●	●	●
Type II diabetes	●	●	●	●
Intervention studies				
LDL-cholesterol	●	●	●	●
HDL-cholesterol	●	●	●	●
Fasting triglycerides	●	●	●	●
Postprandial triglycerides	●	●	●	●
LDL size	●	●	●	●
Apolipoprotein B	●	●	●	●
Non-HDL cholesterol	●	●	●	●
Cholesterol ratios	●	●	●	●
Inflammation	●	●	●	●
Insulin resistance	●	●	●	●
Blood pressure	●	●	●	●
Vascular function	●	●	●	●

CAD, coronary arterial disease; CHD, coronary heart disease; CVD, cardiovascular disease; HDL, high-density lipoprotein; LDL, low-density lipoprotein.

^a Colors represent the overall findings of the studies: favorable (green), neutral or no effect (yellow), uncertain or undetermined (gray). No adverse associations were observed.

intake on individual CMD risk factors have been examined as well, including blood lipids (61–65), hypertension (66–70), body mass index and obesity (71–76), type II diabetes, glycemia and insulin homeostasis (60, 74, 77, 78), also with conflicting findings. A review of 16 meta-analyses by Gille et al. (79) showed weakly beneficial albeit inconsistent links between fermented dairy products and several CMD risk factors (Table 3). The strongest evidence in the review was observed between yogurt on risk factors of type II diabetes (79).

Fermented foods containing inactivated microorganisms

Several other fermented foods whereby live microorganisms have been inactivated or removed (coffee, wine, beer, and chocolate) have also been investigated for their effects on CMD risk. Due to the abundance of literature on these foods, we focused on summarizing the results of systematic reviews and meta-analyses. In terms of coffee, previous prospective studies and meta-analyses did not show associations between coffee consumption and coronary heart disease (CHD) risk (80, 81). However, a recently updated systematic review and meta-analysis of prospective cohort studies observed an inverse non-linear association between moderate habitual coffee consumption and CVD risk (82) with the lowest risk reported

at 3 to 5 cups of coffee per day [relative risk (RR) 0.85, 95% CI 0.8–0.9]. Here, genetic polymorphisms may be an important consideration to further interpret the heterogeneous impacts of coffee consumption on CVD-related outcomes (83).

Several meta-analyses have shown that moderate intake of red wine (e.g., 270 mL/day) may be protective effect against CHD, while extremely high intakes have deleterious effects (84–86). Using data from the EPIC Spanish cohort ($n = 15,630$ men and $n = 25,808$ women), Arriola et al. (84) found that “moderate” (5–30 g of alcohol/day) and “high” (30–90 g alcohol/day) intakes of wine by men reduced CHD risk compared to low or no intakes (HR 0.40; 95% CI 0.25–0.64 for moderate intakes; HR 0.44; 95% CI 0.28–0.69 for high intakes). Similarly, a systematic review of prospective studies indicated that moderate beer consumption of up to 16 g alcohol/day (equivalent to 1 drink/day) for women and 28 g/day (1–2 drinks/day) for men may also have a protective effect against CVD and mortality compared to non-alcohol or occasional drinkers (87). However, a recent meta-analysis of controlled clinical trials of beer intake found that beer drinkers had elevated total cholesterol, high-density lipoprotein (HDL)-cholesterol and apolipoprotein A1, and flow mediated dilation compared to non-beer drinkers, but no differences in LDL-cholesterol, triglycerides, blood pressure, or other biochemical markers of inflammation (88). Overall, the current state of the evidence on the impact of wine and beer intake on different CMD risk parameters remain equivocal.

For chocolate, multiple meta-analyses have found that consumption of chocolate was inversely associated with CHD, stroke, and type II diabetes; however, effective categories or levels of intake differed among the studies and need to be further clarified, and several aspects of study quality needs to be improved (89–92).

Aside from these abovementioned fermented foods, to our knowledge, no well-designed prospective studies have examined the cardiometabolic health impacts of other fermented food products consumed in European (or global) diets.

Limitations of the current evidence

While a plethora of studies have examined associations between the intake of certain fermented foods (fermented dairy, coffee, wine, beer, and cocoa) and cardiometabolic health outcomes, closer examination of the methods of these studies reveals limitations and inaccuracies which could obscure the interpretations of the results. In many studies, non-fermented foods have been misclassified as fermented food products. For example, in an association study between fermented dairy intake, diet quality, and cardiometabolic profile, Mena-Sánchez et al. (93) defined fermented dairy products as “low-fat yogurt, whole-fat yogurt, and all types of cheese (petit Swiss; ricotta; fresh cheese; cottage; and semi-cured and cured cheeses such as Cheddar, Manchego, and Emmental).” Similarly, Kostinen

et al. (94) included cottage cheese under total fermented dairy products when examining associations between fermented and non-fermented dairy products and risk of coronary heart disease. Ricotta, cottage cheese, and fresh cheeses are produced by curdling of milk with acid, and are thus generally not considered to be fermented (95). In light of the recently clarified definitions of fermented foods, a more critical classification scheme is thus required to correctly classify fermented foods prior to examining associations between their intake and health outcomes. Furthermore, almost all studies to date have relied on self-report measures to assess the intake of fermented foods (58, 93, 94). The subjective nature of these tools can result in inaccurate estimates of the levels of fermented food intake. The characteristics and limitations of self-report dietary assessment tools are described in detail below, as well as an opportunity to improve the accuracy of assessing fermented food intake through more objective measures.

Opportunities and future perspectives in research for fermented foods

Improving the dietary assessment of fermented foods

One of the criticisms of population-based studies in nutritional epidemiology is the inability to accurately capture the foods consumed in the diet and their levels of intake, contributing to inconsistent evidence between fermented foods and health, and weakening their potential translation to clinical and public health applications (96, 97). Common dietary assessment methods such as 24-h recalls (i.e., a detailed account of all foods consumed in the previous 24 h self-reported by participants or inventoried by trained interviewers) and food frequency questionnaires (FFQ) (self-reported intakes of a pre-determined list of foods, typically in the past month to year), have advantages and drawbacks (98). They rely on (subjective) self-reporting, and their precision depends upon the devotion, diligence, and memory of the participants. Such errors can lead to reduced power from non-systematic reporting errors, systematic reporting errors, as well as underestimated or overestimated findings in association studies.

To circumvent the limitations of self-report dietary measurement tools, researchers have looked to food intake biomarkers (FIBs) as objective measures of dietary intake (99). The identification of new FIBs has been driven by the application of metabolomics in nutritional research, which allows for a comprehensive measurement of all low molecular weight molecules in biological samples (100). Identifying a single FIB for a specific food can be a challenging task, since most dietary compounds commonly occur in many different

foods. Different food sources may also contain common FIBs from a shared food processing method or metabolic pathway. This is particularly relevant to fermented foods, where common microbial fermentation pathways (e.g., lactic fermentations) can result in the production of similar sets of metabolites. At the same time, the diverse raw material substrates used for fermentation can be a source of unique parent compounds. In these cases, multi-marker approaches, consisting of a combination of non-specific yet complementary biomarkers, could better inform the intake of fermented foods (101). Multi-metabolite panels have been suggested for wine, cocoa, and bread (101), but remain to be exploited for other fermented foods. Thus, there is value in identifying FIBs for fermented foods that could improve the dietary assessment of these foods for future studies, inform on their nutritional quality, and elucidate the mechanisms of action that underpin the health benefits of fermented foods. In addition, there is a need to validate candidate FIBs for fermented foods that have emerged from non-targeted and targeted nutritional metabolomics studies in real-life, non-controlled situations.

Expanding the discovery of food intake biomarkers for globally consumed fermented foods

There is a bias in the literature for investigating a narrow list of fermented foods. In a recent systematic review of FIBs of fermented foods, it was shown that the vast majority of studies were conducted for coffee, wine, cocoa, beer, and bread, while only a smattering of other types of fermented foods were represented (102). For some foods, such as fermented soy, there appears to be ample research but the studies primarily target a list of food group-level biomarkers (e.g., isoflavones found in all soy products), whereas FIBs specifically associated with fermented soy intake were not investigated. While these are common foods consumed in Western/European diets, due to globalization, many of nutritious and delightful fermented food products that are indigenous to other parts of the world are now available locally. Thus, the current literature on FIBs of fermented foods presents an “incomplete” picture of the diversity of fermented foods consumed in modern globalized diets. To address this, further discovery-driven studies on the identification of FIBs for less common fermented foods and condiments in Europe (sourdough, sauerkraut, salami, Worcestershire sauce) and globally (kombucha, kefir, tempeh, kimchi, soy sauce) are warranted. Further, quantification of the most promising FIBs could help in order to use these biomarkers to calibrate self-reported fermented food intakes to clarify diet-disease associations (103). Expanding such research could eventually have an impact on public health, such as through refining dietary guidelines for fermented foods.

Development of a fermented food-specific food frequency questionnaires

While self-report dietary assessment tools are often perceived to be inaccurate due to response bias and measurement error, their qualities of being non-intrusive and non-resource intensive still supports their use in large population-based studies. In addition, well-designed FFQs can produce valid and reproducible estimates to rank participants according to their levels of intake (42). Since most FFQs are not designed to capture fermented foods but rather common foods in the diet of a specific population, there is conceivable value in developing a FFQ specific for fermented foods, which could better capture the habitual intake of fermented foods.

In order for a fermented food-specific FFQ to be effective in accurately capturing the types and levels of fermented foods, researchers must first agree on what foods are considered to be “fermented.” Just recently in 2021, the International Scientific Association for Probiotics and Prebiotics (ISAPP) convened an expert panel to create a common definition for fermented foods (3), which is a step toward unifying future research on the compositional and health qualities of fermented foods. Additionally, consumer misconceptions about fermented foods could be simultaneously addressed. A survey conducted among ~200 university students revealed that nearly two-thirds were unfamiliar with the term “fermented dairy products,” and a similar percentage were unsure whether several cultured dairy products were fermented (104). Such unfamiliarity with fermented foods would yield inaccurate estimates of the types and levels of fermented foods consumed in the diet. To circumvent this, improvements in food legislation could make it easier for consumers to identify fermented foods in the marketplace based on clear food labeling (3). Such food labeling should clearly indicate a food or certain food brands as being “fermented,” and provide additional nutritional information, such as the presence or absence of live microorganisms, as well as compositional aspects of the fermented food. This information could also be linked to a comprehensive, food composition database to help facilitate future research.

Better understanding the composition of different fermented foods and their documentation in food databases

Fermented food products are complex and multi-faceted. The composition of fermented foods reflects untransformed compounds from the raw food substrate, transformed raw material compounds by the fermentation process, and novel metabolites produced by fermentative microorganisms. Additionally, the presence of live microorganisms (or even

inactivated microorganisms which can still be bioactive) presents a unique additional compositional layer (3, 52). Metabolomics and metagenomics analyses coupled with bioinformatics and machine learning could better capture the molecular composition of fermented foods. Concurrently, documenting the composition of different fermented foods and their meta-data in food composition databases (new or existing) is a critical step. Food composition databases are widely used in nutrition and health research to provide information on the nutritional content of foods. However, current food composition databases only report a limited number of nutrients, which only represent a minute fraction of the thousands of distinct chemicals in our food (105). Additionally, in the modern food supply there exists a diverse range of manufactured fermented food products. The same generic fermented food can have many unique formulations depending on the manufacturing source. Here, the use of branded food databases (where information on food composition is provided by manufacturers, food monitoring, and crowdsourcing) are gaining momentum and could be a valuable resource in comprehensively capturing the ingredients within branded fermented foods (106–108). Documenting the composition of fermented foods could open new research avenues for understanding how their consumption affects health and disease. Additionally, such information could also benefit food industries to formulate products with improved nutritional qualities.

Understanding the contribution of fermented foods to healthy and sustainable diets: fermentation as “Agriculture 2.0”

Finally, food fermentation could be thought of as a versatile tool to help extend the world’s food supply. Global food and agriculture production accounts for a quarter of the world’s greenhouse gas emissions, and the global population is poised to reach 10 billion by 2050 (109, 110). With so many mouths to feed, there is a pressing need to reflect on how to improve the sustainability of human diets to best protect the natural environment. A recent report from the EAT-Lancet Commission advised consumers to eat more plant-based foods and less animal-based foods in order to relieve the human footprint on food systems (111). Fermented foods could play an important role in achieving healthy and sustainable plant-based diets. For millennia, food fermentation has been used as a strategy to avoid food waste by increasing the shelf life of fresh foods. In the modern food supply chain, fermentation of plant-based foods can be an effective strategy to improve their shelf life, storage, and transport (112). The consumption of many protein-rich vegetable or legume alternatives to meat are constrained by the high presence of anti-nutrients, which can

be reduced by food fermentation. In addition, fermentation can have an effect on increasing the palatability of plant-based foods by introducing new flavors and reducing unpleasant flavors. On the other hand, fermentation may be used as a strategy to reduce meat consumption (rather than replacing it completely), through the development of novel fermented food products with mixed protein and animal sources (113).

Additionally, fermented foods have an important role in promoting a circular economy, by utilizing waste products from the food industry (e.g., peels, seeds, pulp) and generating new, value-added foods (114). While it is known that fermentation of foods can also increase their nutrient density (thus reducing the need to consume large amounts of foods), industrial fermentations also require a lot of energy (115). Thus, more work is required to elucidate the nutritional and sustainable trade-offs of fermented food production and consumption. Overall, if fermented foods can be consumed as part of a sustainable dietary pattern, they can help improve the environmental impact of food production, promote human health (reducing the risk of CMDs), and in turn, promote economic prosperity.

Conclusion

Fermented foods have been consumed by humans for over 10,000 years, and their renewed popularity in modern diets emphasizes the need to fully elucidate their effects on cardiometabolic health. Although some studies have suggested that the consumption of fermented foods could benefit cardiometabolic health, the evidence is still unclear. This could be partly due to the misclassification of fermented foods, and the use of subjective dietary assessment tools to estimate their intake, in large epidemiological studies. Several future research directions could help disentangle the relationships between fermented foods and cardiometabolic health: improving the dietary assessment of fermented foods (through identification

of FIBs and development of a fermented food FFQ), better understanding the composition of different fermented food products, and exploring the role of fermented foods as part of healthy, sustainable diets.

Author contributions

KL wrote the original draft of the review and prepared the figures and tables. EB-B, KB-P, GV, and EF critically reviewed the manuscript. All authors contributed to the study concept, and read and approved the final manuscript.

Funding

This research was funded by Agroscope, Federal Department of Economic Affairs, Education and Research (EAER), Federal Office for Agriculture (FOAG) (Bern, Switzerland) in the context of the Cardioferment Project.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

References

- Kok CR, Hutkins R. Yogurt and other fermented foods as sources of health-promoting bacteria. *Nutr Rev.* (2018) 76(Suppl. 1):4–15. doi: 10.1093/nutrit/nuy056
- Steinkraus KH. Classification of fermented foods: worldwide review of household fermentation techniques. *Food Control.* (1997) 8:311–7. doi: 10.1016/s0956-7135(97)00050-9
- Marco ML, Sanders ME, Gänzle M, Arrieta MC, Cotter PD, De Vuyst L, et al. The international scientific association for probiotics and prebiotics (ISAPP) consensus statement on fermented foods. *Nat Rev Gastroenterol Hepatol.* (2021) 18:196–208. doi: 10.1038/s41575-020-00390-5
- De Vuyst L, Weckx S. The cocoa bean fermentation process: from ecosystem analysis to starter culture development. *J Appl Microbiol.* (2016) 121:5–17. doi: 10.1111/jam.13045
- Johansen E. Use of natural selection and evolution to develop new starter cultures for fermented foods. *Annu Rev Food Sci Technol.* (2018) 9:411–28. doi: 10.1146/annurev-food-030117-012450
- Rezac S, Kok CR, Heermann M, Hutkins R. Fermented foods as a dietary source of live organisms. *Front Microbiol.* (2018) 9:1785. doi: 10.3389/fmicb.2018.01785
- McDonald D, Hyde E, Debelius JW, Morton JT, Gonzalez A, Ackermann G, et al. American gut: an open platform for citizen science microbiome research. *mSystems.* (2018) 3:e31–18. doi: 10.1128/mSystems.00031-18
- Wolfe BE, Dutton RJ. Fermented foods as experimentally tractable microbial ecosystems. *Cell.* (2015) 161:49–55. doi: 10.1016/j.cell.2015.02.034

9. Leeming ER, Johnson AJ, Spector TD, Le Roy CI. Effect of diet on the gut microbiota: rethinking intervention duration. *Nutrients*. (2019) 11:2862. doi: 10.3390/nu11122862
10. Pasolli E, De Filippis F, Mauriello IE, Cumbo F, Walsh AM, Leech J, et al. Large-scale genome-wide analysis links lactic acid bacteria from food with the gut microbiome. *Nat Commun*. (2020) 11:2610. doi: 10.1038/s41467-020-16438-8
11. Hill C, Guarner F, Reid G, Gibson GR, Merenstein DJ, Pot B, et al. The international scientific association for probiotics and prebiotics consensus statement on the scope and appropriate use of the term probiotic. *Nat Rev Gastro Hepat*. (2014) 11:506–14. doi: 10.1038/nrgastro.2014.66
12. Carding S, Verbeke K, Vipond DT, Corfe BM, Owen LJ. Dysbiosis of the gut microbiota in disease. *Microb Ecol Health Dis*. (2015) 26:26191. doi: 10.3402/mehd.v26.26191
13. Morais LH, Schreiber HL, Mazmanian SK. The gut microbiota-brain axis in behaviour and brain disorders. *Nat Rev Microbiol*. (2021) 19:241–55. doi: 10.1038/s41579-020-00460-0
14. Tamang JP, Watanabe K, Holzapfel WH. Review: diversity of microorganisms in global fermented foods and beverages. *Front Microbiol*. (2016) 7:377. doi: 10.3389/fmicb.2016.00377
15. Tamang JP, Shin DH, Jung SJ, Chae SW. Functional properties of microorganisms in fermented foods. *Front Microbiol*. (2016) 7:578. doi: 10.3389/fmicb.2016.00578
16. Meira SMM, Daroit DJ, Helfer VE, Correa APF, Segalin J, Carro S, et al. Bioactive peptides in water-soluble extracts of ovine cheeses from Southern Brazil and Uruguay. *Food Res Int*. (2012) 48:322–9. doi: 10.1016/j.foodres.2012.05.009
17. Perna A, Intaglietta I, Simonetti A, Gambacorta E. Effect of genetic type and casein haplotype on antioxidant activity of yogurts during storage. *J Dairy Sci*. (2013) 96:3435–41. doi: 10.3168/jds.2012-5859
18. Peng Y, Yang X, Zhang Y. Microbial fibrinolytic enzymes: an overview of source, production, properties, and thrombolytic activity in vivo. *Appl Microbiol Biotechnol*. (2005) 69:126–32. doi: 10.1007/s00253-005-0159-7
19. Leech J, Cabrera-Rubio R, Walsh AM, Macori G, Walsh CJ, Barton W, et al. Fermented-food metagenomics reveals substrate-associated differences in taxonomy and health-associated and antibiotic resistance determinants. *mSystems*. (2020) 5:e522–520. doi: 10.1128/mSystems.00522-20
20. Marco ML, Hutkins R, Hill C, Fulgoni VL, Cifelli CJ, Gahche J, et al. A classification system for defining and estimating dietary intake of live microbes in US adults and children. *J Nutr*. (2022) 152:1729–36. doi: 10.1093/jn/nxac074
21. EFSA Panel on Dietetic Products Nutrition and Allergies. Scientific opinion on the substantiation of health claims related to live yoghurt cultures and improved lactose digestion. *EFSA J*. (2010) 8:1763. doi: 10.2903/j.efsa.2010.1763
22. Derrien M, van Hylckama Vlieg JE. Fate, activity, and impact of ingested bacteria within the human gut microbiota. *Trends Microbiol*. (2015) 23:354–66. doi: 10.1016/j.tim.2015.03.002
23. Giraffa G. Studying the dynamics of microbial populations during food fermentation. *FEMS Microbiol Rev*. (2004) 28:251–60. doi: 10.1016/j.femsre.2003.10.005
24. Marco ML, Heeney D, Binda S, Cifelli CJ, Cotter PD, Foligne B, et al. Health benefits of fermented foods: microbiota and beyond. *Curr Opin Biotechnol*. (2017) 44:94–102. doi: 10.1016/j.copbio.2016.11.010
25. Verbeke KA, Boobis AR, Chiodini A, Edwards CA, Franck A, Kleerebezem M, et al. Towards microbial fermentation metabolites as markers for health benefits of prebiotics. *Nutr Res Rev*. (2015) 28:42–66. doi: 10.1017/S0954422415000037
26. Hayes M, Stanton C, Fitzgerald GF, Ross RP. Putting microbes to work: dairy fermentation, cell factories and bioactive peptides. Part II: bioactive peptide functions. *Biotechnol J*. (2007) 2:435–49. doi: 10.1002/biot.200700045
27. Valerio F, Lavermicocca P, Pascale M, Visconti A. Production of phenyllactic acid by lactic acid bacteria: an approach to the selection of strains contributing to food quality and preservation. *FEMS Microbiol Lett*. (2004) 233:289–95. doi: 10.1016/j.femsle.2004.02.020
28. Peters A, Krumbholz P, Jäger E, Heintz-Buschart A, Akir MV, Rothmund S, et al. Metabolites of lactic acid bacteria present in fermented foods are highly potent agonists of human hydroxycarboxylic acid receptor 3. *PLoS Genet*. (2019) 15:e1008145. doi: 10.1371/journal.pgen.1008145
29. Bhanwar S, Bannia M, Ghosh M, Ganguli A. Use of *Lactococcus lactis* to enrich sourdough bread with gamma-aminobutyric acid. *Int J Food Sci Nutr*. (2013) 64:77–81. doi: 10.3109/09637486.2012.700919
30. Kim HJ, Noh JS, Song YO. Beneficial effects of kimchi, a Korean fermented vegetable food, on pathophysiological factors related to atherosclerosis. *J Med Food*. (2018) 21:127–35. doi: 10.1089/jmf.2017.3946
31. Hong KJ, Lee CH, Kim SW. *Aspergillus oryzae* GB-107 fermentation improves nutritional quality of food soybeans and feed soybean meals. *J Med Food*. (2004) 7:430–5. doi: 10.1089/jmf.2004.7.430
32. Raghuvanshi R, Grayson AG, Schena I, Amanze O, Suwintono K, Quinn RA. Microbial transformations of organically fermented foods. *Metabolites*. (2019) 9:165. doi: 10.3390/metabo9080165
33. Pimentel G, Burnand D, Mürger LH, Pralong FP, Vionnet N, Portmann R, et al. Identification of milk and cheese intake biomarkers in healthy adults reveals high interindividual variability of lewis system-related oligosaccharides. *J Nutr*. (2020) 150:1058–67. doi: 10.1093/jn/nxaa029
34. Dougkas A, Miniñane AM, Givens DI, Reynolds CK, Yaqoob P. Differential effects of dairy snacks on appetite, but not overall energy intake. *Brit J Nutr*. (2012) 108:2274–85. doi: 10.1017/S0007114512000323
35. Fernandez MA, Picard-Deland M, Le Barz M, Daniel N, Marette A. Yoghurt and health. In: Frias J, Martínez-Villaluenga C, Peñas E editors. *Fermented foods in health and disease prevention*. Québec, QC: Elsevier Science & Technology Books (2016).
36. Reddy NR, Pierson MD. Reduction in antinutritional and toxic components in plant foods (a) by fermentation. *Food Res Int*. (1994) 27:281–90. doi: 10.1016/0963-9969(94)90096-5
37. Borresen EC, Henderson AJ, Kumar A, Weir TL, Ryan EP. Fermented foods: patented approaches and formulations for nutritional supplementation and health promotion. *Recent Pat Food Nutr Agric*. (2012) 4:134–40. doi: 10.2174/2212798411204020134
38. Campbell-Platt G. Fermented foods - a world perspective. *Food Res Int*. (1994) 27:253–7. doi: 10.1016/0963-9969(94)90093-0
39. Anal AK. Quality ingredients and safety concerns for traditional fermented foods and beverages from Asia: a review. *Fermentation Basel*. (2019) 5:8. doi: 10.3390/fermentation5010008
40. Tamang JP, Cotter PD, Endo A, Han NS, Kort R, Liu SQ, et al. Fermented foods in a global age: east meets west. *Compr Rev Food Sci Food Saf*. (2020) 19:184–217. doi: 10.1111/1541-4337.12520
41. Capozzi V, Spano G. Food microbial biodiversity and "microbes of protected origin". *Front Microbiol*. (2011) 2:237. doi: 10.3389/fmicb.2011.00237
42. Li KJ, Brouwer-Brolsma EM, Burton KJ, Vergères G, Feskens EJM. Prevalence of fermented foods in the Dutch adult diet and validation of a food frequency questionnaire for estimating their intake in the NQplus cohort. *BMC Nutr*. (2020) 6:69. doi: 10.1186/s40795-020-00394-z
43. Bron PA, Kleerebezem M, Brummer RJ, Cani PD, Mercenier A, MacDonald TT, et al. Can probiotics modulate human disease by impacting intestinal barrier function? *Br J Nutr*. (2017) 117:93–107. doi: 10.1017/S0007114516004037
44. Kim DH, Kim H, Jeong D, Kang IB, Chon JW, Kim HS, et al. Kefir alleviates obesity and hepatic steatosis in high-fat diet-fed mice by modulation of gut microbiota and mycobiota: targeted and untargeted community analysis with correlation of biomarkers. *J Nutr Biochem*. (2017) 44:35–43. doi: 10.1016/j.jnutbio.2017.02.014
45. Wang L, Guo MJ, Gao Q, Yang JF, Yang L, Pang XL, et al. The effects of probiotics on total cholesterol: a meta-analysis of randomized controlled trials. *Medicine (Baltimore)*. (2018) 97:e9679. doi: 10.1097/MD.0000000000009679
46. Kadooka Y, Sato M, Imaizumi K, Ogawa A, Ikuyama K, Akai Y, et al. Regulation of abdominal adiposity by probiotics (*Lactobacillus gasseri* SBT2055) in adults with obese tendencies in a randomized controlled trial. *Eur J Clin Nutr*. (2010) 64:636–43. doi: 10.1038/ejcn.2010.19
47. Kadooka Y, Sato M, Ogawa A, Miyoshi M, Uenishi H, Ogawa H, et al. Effect of *Lactobacillus gasseri* SBT2055 in fermented milk on abdominal adiposity in adults in a randomised controlled trial. *Br J Nutr*. (2013) 110:1696–703. doi: 10.1017/S0007114513001037
48. Sanchez M, Darimont C, Drapeau V, Emady-Azar S, Lepage M, Rezzonico E, et al. Effect of *Lactobacillus rhamnosus* CGMCC1.3724 supplementation on weight loss and maintenance in obese men and women. *Br J Nutr*. (2014) 111:1507–19. doi: 10.1017/S0007114513003875
49. Sharafedinov KK, Plotnikova OA, Alexeeva RI, Sentsova TB, Songisepp E, Stepetova J, et al. Hypocaloric diet supplemented with probiotic cheese improves body mass index and blood pressure indices of obese hypertensive patients—a randomized double-blind placebo-controlled pilot study. *Nutr J*. (2013) 12:138. doi: 10.1186/1475-2891-12-138
50. Shirouchi B, Nagao K, Umegatani M, Shiraiishi A, Morita Y, Kai S, et al. Probiotic *Lactobacillus gasseri* SBT2055 improves glucose tolerance and reduces body weight gain in rats by stimulating energy expenditure. *Br J Nutr*. (2016) 116:451–8. doi: 10.1017/S0007114516002245

51. Yadav H, Lee JH, Lloyd J, Walter P, Rane SG. Beneficial metabolic effects of a probiotic via butyrate-induced GLP-1 hormone secretion. *J Biol Chem.* (2013) 288:25088–97. doi: 10.1074/jbc.M113.452516
52. Salminen S, Collado MC, Endo A, Hill C, Lebeer S, Quigley EMM, et al. The international scientific association of probiotics and prebiotics (ISAPP) consensus statement on the definition and scope of postbiotics. *Nat Rev Gastroenterol Hepatol.* (2021) 9:649–67. doi: 10.1038/s41575-021-00440-6
53. Selma-Royo M, Tarrazó M, Garcia-Mantrana I, Gomez-Gallego C, Salminen S, Collado MC. Shaping microbiota during the first 1000 days of life. *Adv Exp Med Biol.* (2019) 1125:3–24. doi: 10.1007/5584_2018_312
54. Pessione E, Cirrincione S. Bioactive molecules released in food by lactic acid bacteria: encrypted peptides and biogenic amines. *Front Microbiol.* (2016) 7:876. doi: 10.3389/fmicb.2016.00876
55. Mozaffarian D, Wu JHY. Flavonoids, dairy foods, and cardiovascular and metabolic health: a review of emerging biologic pathways. *Circ Res.* (2018) 122:369–84. doi: 10.1161/CIRCRESAHA.117.309008
56. Lordan R, Tsoupras A, Mitra B, Zabetakis I. Dairy fats and cardiovascular disease: do we really need to be concerned? *Foods.* (2018) 7:29. doi: 10.3390/foods7030029
57. Meraz-Torres LSH-SH. Conjugated linoleic acid in dairy products: a review. *Am J Food Technol.* (2012) 7:176–9. doi: 10.3923/ajft.2012.176.179
58. Johansson I, Esberg A, Nilsson LM, Jansson JH, Wennberg P, Winkvist A. Dairy product intake and cardiometabolic diseases in Northern Sweden: a 33-year prospective cohort study. *Nutrients.* (2019) 11:284. doi: 10.3390/nu11020284
59. Praagman J, Dalmeijer GW, van der Schouw YT, Soedamah-Muthu SS, Monique Verschuren WM, Bas Bueno-de-Mesquita H, et al. The relationship between fermented food intake and mortality risk in the European Prospective Investigation into Cancer and Nutrition-Netherlands cohort. *Br J Nutr.* (2015) 113:498–506. doi: 10.1017/S0007114514003766
60. Dehghan M, Mente A, Rangarajan S, Sheridan P, Mohan V, Iqbal R, et al. Association of dairy intake with cardiovascular disease and mortality in 21 countries from five continents (PURE): a prospective cohort study. *Lancet.* (2018) 392:2288–97. doi: 10.1016/S0140-6736(18)31812-9
61. Nestle PJ, Mellett N, Pally S, Wong G, Barlow CK, Croft K, et al. Effects of low-fat or full-fat fermented and non-fermented dairy foods on selected cardiovascular biomarkers in overweight adults. *Br J Nutr.* (2013) 110:2242–9. doi: 10.1017/S0007114513001621
62. Watanabe D, Kuranuki S, Sunto A, Matsumoto N, Nakamura T. Daily yogurt consumption improves glucose metabolism and insulin sensitivity in young nondiabetic Japanese subjects with type-2 diabetes risk alleles. *Nutrients.* (2018) 10:1834. doi: 10.3390/nu10121834
63. Tholstrup T, Høy CE, Andersen LN, Christensen RDK, Sandstrom B. Does fat in milk, butter and cholesterol differently? *J Am Coll Nutr.* (2004) 23:169–76. doi: 10.1080/07315724.2004.10719358
64. Sadeghi M, Khosravi-Boroujeni H, Sarrafzadegan N, Asgary S, Roohafza H, Ghariipour M, et al. Cheese consumption in relation to cardiovascular risk factors among Iranian adults- IHHP Study. *Nutr Res Pract.* (2014) 8:336–41. doi: 10.4162/nrp.2014.8.3.336
65. Steffen LM, Jacobs DR. Relation between dairy food intake and plasma lipid levels: the CARDIA Study. *Aust J Dairy Technol.* (2003) 58:92–7.
66. Soedamah-Muthu SS, Verberne LD, Ding EL, Engberink ME, Geleijnse JM. Dairy consumption and incidence of hypertension: a dose-response meta-analysis of prospective cohort studies. *Hypertension.* (2012) 60:1131–7. doi: 10.1161/HYPERTENSIONAHA.112.195206
67. Wang H, Fox CS, Troy LM, McKeown NM, Jacques PF. Longitudinal association of dairy consumption with the changes in blood pressure and the risk of incident hypertension: the Framingham Heart Study. *Br J Nutr.* (2015) 114:1887–99. doi: 10.1017/S0007114515003578
68. Usinger L, Reimer C, Ibsen H. Fermented milk for hypertension. *Cochrane Database Syst Rev.* (2012) 4:CD008118. doi: 10.1002/14651858.CD008118.pub2
69. Beltrán-Barrientos LM, González-Córdova AF, Hernández-Mendoza A, Torres-Inguanzo EH, Astiazaran-García H, Esparza-Romero J, et al. Randomized double-blind controlled clinical trial of the blood pressure-lowering effect of fermented milk with *Lactococcus lactis*: a pilot study. *J Dairy Sci.* (2018) 101:2819–25. doi: 10.3168/jds.2017-13189
70. Schlienger JL, Paillard F, Lecerf JM, Romon M, Bonhomme C, Schmitt B, et al. Effect on blood lipids of two daily servings of Camembert cheese. An intervention trial in mildly hypercholesterolemic subjects. *Int J Food Sci Nutr.* (2014) 65:1013–8. doi: 10.3109/09637486.2014.945156
71. Brouwer-Brolsma EM, Sluik D, Singh-Povel CM, Feskens EJM. Dairy shows different associations with abdominal and BMI-defined overweight: cross-sectional analyses exploring a variety of dairy products. *Nutr Metab Cardiovasc.* (2018) 28:451–60. doi: 10.1016/j.numecd.2018.01.008
72. Schwingshackl L, Hoffmann G, Schwedhelm C, Kalle-Uhlmann T, Missbach B, Knüppel S, et al. Consumption of dairy products in relation to changes in anthropometric variables in adult populations: a systematic review and meta-analysis of cohort studies. *PLoS One.* (2016) 11:e0157461. doi: 10.1371/journal.pone.0157461
73. Panahi S, Gallant A, Tremblay A, Pérusse L, Despres JP, Drapeau V. The relationship between yogurt consumption, body weight, and metabolic profiles in youth with a familial predisposition to obesity. *Eur J Clin Nutr.* (2019) 73:541–8. doi: 10.1038/s41430-018-0166-2
74. Naito E, Yoshida Y, Kunihiro S, Makino K, Kasahara K, Kounoshi Y, et al. Effect of *Lactobacillus casei* strain Shirota-fermented milk on metabolic abnormalities in obese prediabetic Japanese men: a randomised, double-blind, placebo-controlled trial. *Biosci Microbiota Food Health.* (2018) 37:9–18. doi: 10.12938/bmfh.17-012
75. Zemel MB, Richards J, Mathis S, Milstead A, Gebhardt L, Silva E. Dairy augmentation of total and central fat loss in obese subjects. *Int J Obes (Lond).* (2005) 29:391–7. doi: 10.1038/sj.ijo.0802880
76. Brassard D, Tessier-Grenier M, Allaire J, Rajendiran E, She Y, Ramprasad V, et al. Comparison of the impact of SFAs from cheese and butter on cardiometabolic risk factors: a randomized controlled trial. *Am J Clin Nutr.* (2017) 105:800–9. doi: 10.3945/ajcn.116.150300
77. Gijsbers L, Ding EL, Malik VS, de Goede J, Geleijnse JM, Soedamah-Muthu SS. Consumption of dairy foods and diabetes incidence: a dose-response meta-analysis of observational studies. *Am J Clin Nutr.* (2016) 103:1111–24. doi: 10.3945/ajcn.115.123216
78. Ibsen DB, Laursen ASD, Lauritzen L, Tjønneland A, Overvad K, Jakobsen MU. Substitutions between dairy product subgroups and risk of type 2 diabetes: the Danish Diet, Cancer and Health cohort. *Br J Nutr.* (2017) 118:989–97. doi: 10.1017/s0007114517002896
79. Gille D, Schmid A, Walther B, Vergères G. Fermented food and non-communicable chronic diseases: a review. *Nutrients.* (2018) 10:448. doi: 10.3390/nu10040448
80. Kawachi I, Colditz GA, Stone CB. Does coffee-drinking increase the risk of coronary heart-disease - results from a metaanalysis. *Brit Heart J.* (1994) 72:269–75. doi: 10.1136/hrt.72.3.269
81. Greenland S. A meta-analysis of coffee, myocardial infarction, and coronary death. *Epidemiology.* (1993) 4:366–74. doi: 10.1097/00001648-199307000-00013
82. Ding M, Bhupathiraju SN, Satija A, van Dam RM, Hu FB. Long-term coffee consumption and risk of cardiovascular disease: a systematic review and a dose-response meta-analysis of prospective cohort studies. *Circulation.* (2014) 129:643–59. doi: 10.1161/CIRCULATIONAHA.113.005925
83. De Giuseppe R, Di Napoli I, Granata F, Mottola A, Cena H. Caffeine and blood pressure: a critical review perspective. *Nutr Res Rev.* (2019) 32:169–75. doi: 10.1017/S0954422419000015
84. Arriola L, Martinez-Cambor P, Larrañaga N, Basterretxea M, Amiano P, Moreno-Iribas C, et al. Alcohol intake and the risk of coronary heart disease in the Spanish EPIC cohort study. *Heart.* (2010) 96:124–30. doi: 10.1136/hrt.2009.173419
85. Ronksley PE, Brien SE, Turner BJ, Mukamal KJ, Ghali WA. Association of alcohol consumption with selected cardiovascular disease outcomes: a systematic review and meta-analysis. *BMJ.* (2011) 342:d671. doi: 10.1136/bmj.d671
86. Fardet A, Boirie Y. Associations between food and beverage groups and major diet-related chronic diseases: an exhaustive review of pooled/meta-analyses and systematic reviews. *Nutr Rev.* (2014) 72:741–62. doi: 10.1111/nure.12153
87. Marcos A, Serra-Majem L, Pérez-Jiménez F, Pascual V, Tinahones FJ, Estruch R. Moderate consumption of beer and its effects on cardiovascular and metabolic health: an updated review of recent scientific evidence. *Nutrients.* (2021) 13:879. doi: 10.3390/nu13030879
88. Spaggiari G, Cignarelli A, Sansone A, Baldi M, Santi D. To beer or not to beer: a meta-analysis of the effects of beer consumption on cardiovascular health. *PLoS One.* (2020) 15:e0233619. doi: 10.1371/journal.pone.0233619
89. Kwok CS, Boekholdt SM, Lentjes MA, Loke YK, Luben RN, Yeong JK, et al. Habitual chocolate consumption and risk of cardiovascular disease among healthy men and women. *Heart.* (2015) 101:1279–87. doi: 10.1136/heartjnl-2014-307050
90. Larsson SC, Akesson A, Gigante B, Wolk A. Chocolate consumption and risk of myocardial infarction: a prospective study and meta-analysis. *Heart.* (2016) 102:1017–22. doi: 10.1136/heartjnl-2015-309203

91. Yuan S, Li X, Jin Y, Lu J. Chocolate consumption and risk of coronary heart disease, stroke, and diabetes: a meta-analysis of prospective studies. *Nutrients*. (2017) 9:688. doi: 10.3390/nu9070688
92. Morze J, Schwedhelm C, Bencic A, Hoffmann G, Boeing H, Przybylowicz K, et al. Chocolate and risk of chronic disease: a systematic review and dose-response meta-analysis. *Eur J Nutr*. (2020) 59:389–97. doi: 10.1007/s00394-019-01914-9
93. Mena-Sánchez G, Babio N, Martínez-González MA, Corella D, Schroder H, Vioque J, et al. Fermented dairy products, diet quality, and cardio-metabolic profile of a Mediterranean cohort at high cardiovascular risk. *Nutr Metab Cardiovasc Dis*. (2018) 28:1002–11. doi: 10.1016/j.numecd.2018.05.006
94. Koskinen TT, Virtanen HEK, Voutilainen S, Tuomainen TP, Mursu J, Virtanen JK. Intake of fermented and non-fermented dairy products and risk of incident CHD: the Kuopio Ischaemic Heart Disease Risk Factor Study. *Br J Nutr*. (2018) 120:1288–97. doi: 10.1017/S0007114518002830
95. Kongo JM, Malcata FX. Cheese: types of cheeses – soft. In: Caballero B, Finglas PM, Toldra F editors. *Encyclopedia of Food and Health*. Oxford: Elsevier (2016). p. 768–73.
96. Taubes G. Epidemiology faces its limits. *Science*. (1995) 269:164–9. doi: 10.1126/science.7618077
97. Ioannidis JP. Implausible results in human nutrition research. *BMJ*. (2013) 347:f6698. doi: 10.1136/bmj.f6698
98. Willett WC. *Nutritional Epidemiology*. 3rd ed. New York, NY: Oxford University Press (2013).
99. Jenab M, Slimani N, Bictash M, Ferrari P, Bingham SA. Biomarkers in nutritional epidemiology: applications, needs and new horizons. *Hum Genet*. (2009) 125:507–25. doi: 10.1007/s00439-009-0662-5
100. Gibbons H, Brennan L. Metabolomics as a tool in the identification of dietary biomarkers. *Proc Nutr Soc*. (2017) 76:42–53. doi: 10.1017/S002966511600032X
101. Garcia-Aloy M, Rabassa M, Casas-Agustench P, Hidalgo-Liberona N, Llorach R, Andres-Lacueva C. Novel strategies for improving dietary exposure assessment: multiple data fusion is a more accurate measure than the traditional single-biomarker approach. *Trends Food Sci Tech*. (2017) 69:220–9. doi: 10.1016/j.tifs.2017.04.013
102. Li KJ, Brouwer-Brolsma EM, Burton-Pimentel KJ, Vergères G, Feskens EJM. A systematic review to identify biomarkers of intake for fermented food products. *Genes Nutr*. (2021) 16:5. doi: 10.1186/s12263-021-00686-4
103. D'Angelo S, Gormley IC, McNulty BA, Nugent AP, Walton J, Flynn A, et al. Combining biomarker and food intake data: calibration equations for citrus intake. *Am J Clin Nutr*. (2019) 110:977–83. doi: 10.1093/ajcn/nqz168
104. Hekmat S, Koba L. Fermented dairy products: knowledge and consumption. *Can J Diet Pract Res*. (2006) 67:199–201. doi: 10.3148/67.4.2006.199
105. Barabási A, Menichetti G, Loscalzo J. The unmapped chemical complexity of our diet. *Nat Food*. (2020) 1:33–7. doi: 10.1038/s43016-019-0005-1
106. Ter Borg S, Steenbergen E, Milder IEJ, Temme EHM. Evaluation of nutrient score in relation to dietary guidelines and food reformulation in The Netherlands. *Nutrients*. (2021) 13:4536. doi: 10.3390/nu13124536
107. Katsouri E, Zampelas A, Drosinos EH, Nychas GE. Labelling assessment of Greek "Quality Label" prepacked cheeses as the basis for a branded food composition database. *Nutrients*. (2022) 14:230. doi: 10.3390/nu14010230
108. Pravst I, Hribar M, Zmitek K, Blazica B, Korousic Seljak B, Kusar A. Branded foods databases as a tool to support nutrition research and monitoring of the food supply: insights from the slovenian composition and labeling information system. *Front Nutr*. (2021) 8:798576. doi: 10.3389/fnut.2021.798576
109. Springmann M, Clark M, Mason-D'Croz D, Wiebe K, Bodirsky BL, Lassaletta L, et al. Options for keeping the food system within environmental limits. *Nature*. (2018) 562:519–25. doi: 10.1038/s41586-018-0594-0
110. Tilman D, Clark M. Global diets link environmental sustainability and human health. *Nature*. (2014) 515:518–22. doi: 10.1038/nature13959
111. Willett W, Rockström J, Loken B, Springmann M, Lang T, Vermeulen S, et al. Food in the Anthropocene: the EAT–Lancet Commission on healthy diets from sustainable food systems. *Lancet*. (2019) 393:447–92. doi: 10.1016/S0140-6736(18)31788-4
112. Kinnunen P, Guillaume JHA, Taka M, D'Odorico P, Siebert S, Puma MJ, et al. Local food crop production can fulfill demand for less than one-third of the population. *Nat Food*. (2020) 1:229–37. doi: 10.1038/s43016-020-0060-7
113. Saint-Eve A, Irlinger F, Pénicaud C, Souchon I, Marette S. Consumer preferences for new fermented food products that mix animal and plant protein sources. *Food Qual Prefer*. (2021) 90:104117. doi: 10.1016/j.foodqual.2020.104117
114. Rastogi YR, Thakur R, Thakur P, Mittal A, Chakrabarti S, Siwal SS, et al. Food fermentation - Significance to public health and sustainability challenges of modern diet and food systems. *Int J Food Microbiol*. (2022) 371:109666. doi: 10.1016/j.jfoodmicro.2022.109666
115. Curran JS, Smith J, Holms W. Heat-and-power in industrial fermentation processes. *Appl Energy*. (1989) 34:9–20. doi: 10.1016/0306-2619(89)90051-2
116. Prado MR, Blandon LM, Vandenberghe LP, Rodrigues C, Castro GR, Thomaz-Soccol V, et al. Milk kefir: composition, microbial cultures, biological activities, and related products. *Front Microbiol*. (2015) 6:1177. doi: 10.3389/fmicb.2015.01177
117. Vilela DM, Pereira GVD, Silva CF, Batista LR, Schwan RF. Molecular ecology and polyphasic characterization of the microbiota associated with semi-dry processed coffee (*Coffea arabica* L.). *Food Microbiol*. (2010) 27:1128–35. doi: 10.1016/j.fm.2010.07.024
118. Meersman E, Steensels J, Mathawan M, Wittocx PJ, Saels V, Struyf N, et al. Detailed analysis of the microbial population in Malaysian spontaneous cocoa pulp fermentations reveals a core and variable microbiota. *PLoS One*. (2013) 8:e81559. doi: 10.1371/journal.pone.0081559