



Transcriptional Feedback in Plant Growth and Defense by PIFs, BZR1, HY5, and MYC Transcription Factors

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Abstract

Growth of *Arabidopsis* is controlled by the activity of a set of bHLH and bZIP transcription factors of which phytochrome interacting factor4 (*PIF4*), BRASSINAZOLE-RESISTANT 1 (*BZR1*), and elongated hypocotyl 5 (*HY5*) have been most extensively studied. Defense responses are controlled by a set of MYC transcription factors of which *MYC2* is best characterized. Moreover, hundreds of additional proteins (here named co-factors) have been identified which (in)directly may affect the expression or activity of these TFs. Thus, regulation of expression of genes encoding these co-factors becomes an integral part of understanding the molecular control of growth and defense. Here, we review RNA-seq data related to PIF, BZR1, HY5, or MYC activity, which indicate that 125 co-factor genes affecting PIFs, HY5, BZR1, or MYCs are themselves under transcriptional control by these TFs, thus revealing potential feedback regulation in growth and defense. The transcriptional feedback on co-factor genes related to *PIF4*, *BZR1*, and *MYC2* by PIFs, BZR1, or MYCs, mostly results in negative feedback on *PIF4*, *BZR1*, or *MYC2* activity. In contrast, transcription feedback on co-factor genes for *HY5* by *HY5* mostly results in positive feedback on *HY5* activity. PIF4 and BZR1 exert a balanced regulating of photoreceptor-gene expression, whose products directly or indirectly affect PIF4, HY5, and MYC2 protein stability as a function of light. Growth itself is balanced by both multiple positive and multiple negative feedback on *PIF4* and *BZR1* activity. The balance between growth and defense is mostly through direct cross-regulation between *HY5* and *MYC2* as previously described, but also through potential transcriptional feedback on co-factor genes for *MYC2* by *PIF4*, *BZR1*, and *HY5* and through transcriptional feedback of co-factors for PIF4 and BZR1 by *MYC2*. The interlocking feed-forward and feed-backward transcriptional regulation of *PIF4*, *BZR1*, *HY5*, and *MYC2* co-factors is a signature of robust and temporal control of signaling related to growth and defense.

Keywords PIF4 · BZR1 · HY5 · MYC2 · Transcriptional feedback · Growth · Defense

Introduction

Key Message

- DNA-ChIP and RNA-seq meta data analysis shows that co-factors that affect transcriptional activity of PIFs, BZR1, and HY5 and MYCs are under (potential) transcriptional feedback control by these same transcription factors, revealing novel and redundant feedback in growth and defense at the level of transcription.

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Key Transcription Factors Acting in Growth and Defense in *Arabidopsis*: PIFs, BZR1, HY5, and MYCs

The molecular components involved in cell elongation and defense responses in plants have most extensively been studied in the model plant *Arabidopsis*. Numerous transcription factors (TFs) are involved in growth control, of which the basic helix-loop-helix transcription factors phytochrome interacting factors (PIFs) and the brassinolide activated transcription factor protein BRASSINAZOLE-RESISTANT 1 (BZR1) and its homolog BRI1-EMS suppressor 1 (BES1) have been shown to be key factors in stimulating growth under different environmental conditions (darkness, heat, shade) (Leivar et al. 2008; Quint et al. 2016; Li et al. 2020a).

In contrast, suppression of growth in the light is mainly by the basic leucine zipper transcription factors elongated hypocotyl 5 (HY5) and its homolog (HYH) (Gangappa and Botto 2016). Table S1 shows the number of publications for TFs listed in combination with terms related to the environmental conditions of darkness, light, heat, shade, or defense, represented by the terms skotomorphogenesis, photomorphogenesis, thermomorphogenesis, and defense, respectively. The inventory shows that from the PIFs, PIF4 is the most representative/studied TF; from the brassinolide (BR) regulated TFs, BZR1 has been most extensively studied; and from the growth-suppressing TFs, HY5 has most extensively been studied (Table S1). Therefore, the focus here is on the transcriptional regulation of PIF4, BZR1, and HY5 during growth. Biotic stress responses triggered by the Jasmonic acid signaling pathway result in activation of MYC transcription factors, which target defense-related genes. Of the different defense-related *MYC* genes (*MYC2*, *MYC3*, and *MYC4*), specifically the action of *MYC2* is key and has been best studied in *Arabidopsis* (Table S1) (Fernandez-Calvo et al. 2011). Therefore, in this study on transcriptional regulation for defense responses, the focus is on *MYC2*. Our aim is to summarize the literature on transcriptional regulation of the main transcription factors in growth and defense and use available omics-data sets to uncover previously unnoticed potential for transcriptional positive and negative feedback regulation.

Different Levels of Potential Feedback on PIF4, BZR1, HY5, or MYC2 Activity

When PIFs, BZR1, HY5, and MYCs are key TFs in growth and defense, the TFs that act on the promoter of the genes encoding these key TFs are expected to be higher order TFs that regulate growth and defense, unless these key TFs play a dominant role in regulating their own expression. Indeed, the genes for each of these key TFs have sequences in their promoter for potential self-binding, indicating the potential for auto-feedback (as shown as an example for PIF4, Fig. 1A, direct feedback a). In addition, each of the key TFs may affect the expression of other TF genes that bind to the promoter of the key TFs (as shown for PIF4, Fig. 1A, in-direct feedback b). While transcription and translation may determine the level of key TF proteins, and thus the potential capacity for key TF activity, the actual key TF activity is determined by additional factors acting at the post-transcriptional level (reviewed in (Koini et al. 2009, Fernandez-Calvo et al. 2011, Gangappa and Botto 2016, Ibanez et al. 2018, Martinez et al. 2018; Qiu 2020)). The genes encoding these co-factors may themselves be (partially) under the control of the key TF itself, thus forming potential additional indirect feedback on each

key TF (as shown for PIF4 in Fig. 1A, feedback c. In the same way, three levels of potential feedback on BZR1, HY5, and MYC2 may be drawn).

The literature describes different types of co-factors that affect either PIF4, BZR1, HY5 or MYC2 activity (Fig. 1B). Our inventory yielded 102 co-factor genes for PIF4 (Table S2), 93 co-factor genes for BZR1 (Table S3), 36 co-factor genes for HY5 (Table S4) and 57 co-factor genes for MYC2 (Table S5). Tables S2-S5 also give a brief description of each co-factor acting on PIF4, BZR1, HY5, or MYC2 and refers to the individual papers describing the experimental conditions under which these factors affect the TF activity. Among these co-factors are other TFs (e.g., certain clock genes) that may either aid or block the transcriptional activity of PIF4, BZR1, HY5, or MYC2. Some co-factors are part of a gene family such as the family of JAZ repressors (12 members), or the family of DELLA proteins (5 members) with sequestering activity. While the interaction with our key TFs may only have been shown for some family members, here all gene family members are considered potential co-factor (see Tables S2-5). The activity of BZR1 and MYC2 is coupled to brassinosteroid (BR) biosynthesis and signaling and Jasmonic acid (JA) biosynthesis and signaling, respectively. Therefore, genes involved in BR biosynthesis and signaling were included as potential positive acting co-factor genes for BZR1 and JA biosynthesis genes and signaling were included as potential additional positive acting co-factor genes for MYC2 (see Tables S2-5). The activity of phosphorylated BZR1 and PIFs is effectuated by the binding of 14-3-3 proteins (Gampala et al. 2007, Wang et al. 2011, Wang et al. 2013a, b, Srivastava, Srivastava et al. 2020). Binding of 14-3-3 to phosphorylated BZR1 results in cytosolic retention of BZR1 and thus 14-3-3 proteins are negative co-factors for BZR1 activity in the nucleus. Also, for PIF proteins, 14-3-3 binding results in reduced PIF activity (Adams et al. 2014, Camoni et al. 2018, Huang et al. 2018). Therefore, the set of 14-3-3 genes is included as potential negative acting co-factor genes for both PIF4 and BZR1. In addition, co-factors may be part of protein complexes involved in the specific degradation of one of the four TF proteins. The summary in Fig. 1B shows the potential positive (green arrows) or negative (red arrows) feedback of the different types of co-factors on either PIF4, BZR1, HY5, or MYC2. The regulation of the genes encoding the numerous co-factors that modulate PIF4, BZR1, HY5, or MYC2 protein activity, by these same key TFs, form the potential of indirect feedback as described in Fig. 1A (feedback c). This potential for feedback regulation through co-factor genes has not been studied systematically and is here determined from available transcriptome data related to the transcriptional activity of PIFs, BZR1, HY5, or MYCs (see below).

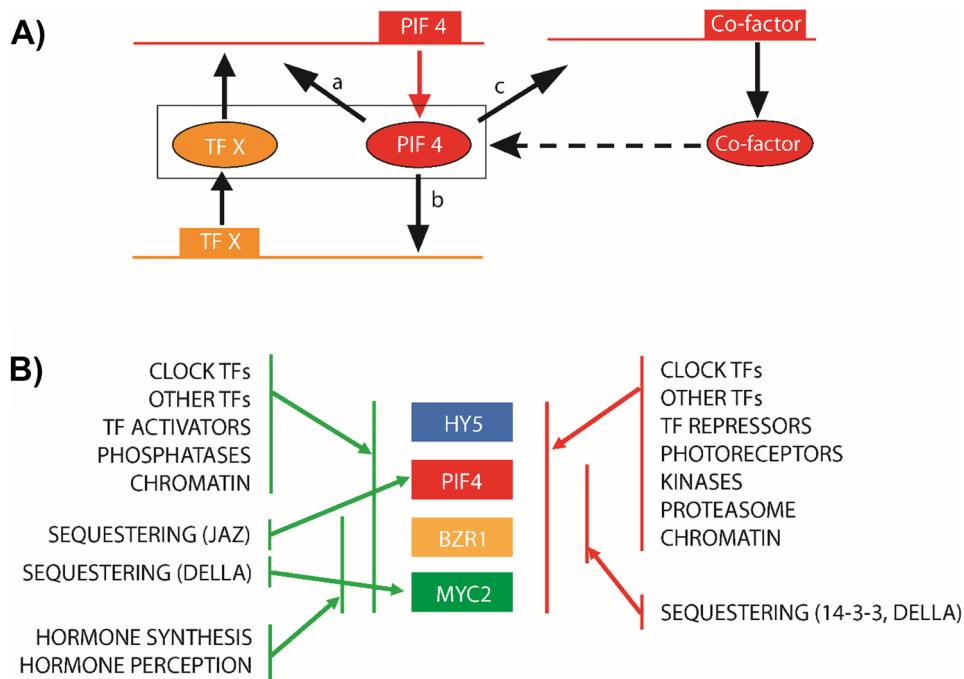


Fig. 1 Multiple potential (in)direct feedback on key TF activity. **A** Potential (in)direct feedback on key TF activity is illustrated for PIF4, but a similar scheme can be drawn for the other key TFs. PIF4 gene (coding sequence and promoter), PIF4 related co-factor gene (coding sequence and promoter), and genes for TF X acting on PIF4 (coding sequence and promoter) are indicated by line (promoter) and rectangle (coding sequence). PIF4 protein, PIF4-related co-factor protein, and TF X protein are indicated by an oval. PIF4 has binding sites for PIF4 in its own promoter, indicating a potential role for PIF4 in its own transcription (potential direct auto-feedback regulation loop (**a**)). Multiple TFs are involved in the transcription of PIF4, of which the corresponding genes may be targeted by PIF4 (potential indirect transcriptional feedback loops (**b**)). In addition, multiple co-factors may act on the activity/stability of PIF4 or TF X acting on PIF4 (dashed arrow). The genes encoding for these co-factors may be tar-

get of PIF4 (potential indirect autofeedback regulation on PIF4: (**c**)). Potential transcriptional feedback indicated by the solid black arrows, potential post-transcriptional feedback on PIF4 or TF X protein indicated by a dashed black arrow. Similar feedback interaction schemes may be drawn for BZR1, HY5, and MYC2. **B** Different types of co-factors acting on PIF4, BZR1, HY5, or MYC2 protein. Left: factors with positive feedback on the key TFs (Green arrows), Right: factors with negative feedback on the key TFs (Red arrows). TF activators and repressors: Proteins that interact with the key TFs in transcription complexes to either stimulate or block transcriptional activity. Chromatin: chromatin modifying factors. Proteasome: components of multi-protein complexes that target proteins for degradation. Sequestering: proteins that interact with the key TFs to prevent their transcriptional activity

Potential for Auto-Feedback and Indirect Feedback on PIFs, BZR1, HY5, and MYCs

The inventory of validated TFs binding to promoters of *PIF4*, *BZR1*, *HY5*, and *MYC2* from ChIP-seq experiments as available at PlantPan3.0 show 34 TFs with binding to the *PIF4* promoter, 36 TFs with binding to the *BZR1* promoter, 26 TFs with binding to the *HY5* promoter and 32 TFs with binding to the *MYC2* promoter (Fig. 2A). Actually, no less than 16 TFs share binding to each of the four promoters, one of which is PIF4 itself (Fig. 2A). Although this indicates a potential central role for PIF4 in the expression of all four selected TFs, no evidence is found in RNA-seq data that PIF4 or PIFs are limiting in the transcription of BZR1, HY5, or MYC2, suggesting a redundant function of PIFs acting on these promoters. As described above, there is an overlap for co-factors that act on PIF4, BZR1, MYC2, or

HY5 protein (Fig. 2B). This suggests a potential for coordinated activity between these four key TFs as effectuated by co-factor action. (Venn diagram of Tables S2-S5: Fig. 2B). PIF4 and BZR1 share 36 co-factor genes. PIF4 and MYC2 share 27 co-factor genes, BZR1 and MYC2 share 12 co-factor genes, and HY5 and MYC2 only share 8 co-factor genes (Fig. 2B). The inventory indicates PIF4 as an indirect co-factor for BZR1 and HY5 activity, BZR1 as co-factor for its own activity and PIF4 activity and MYC2 is a direct co-factor for its own activity and indirect co-factor for PIF4 and HY5 activity (Fig. 2B). HY5, COP1, and photoreceptors PHYB, CRY1, and CRY2 have a central position as shared co-factors for all four selected TFs. HY5 can affect PIF4 by competitive binding of HY5 at PIF4 target sites (Delker et al. 2014; Gangappa and Kumar 2017). HY5 potentially affects BZR1 by sequestering de-phosphorylated (active) BZR1/BES1 (Li and He 2016), HY5 potentially affects its

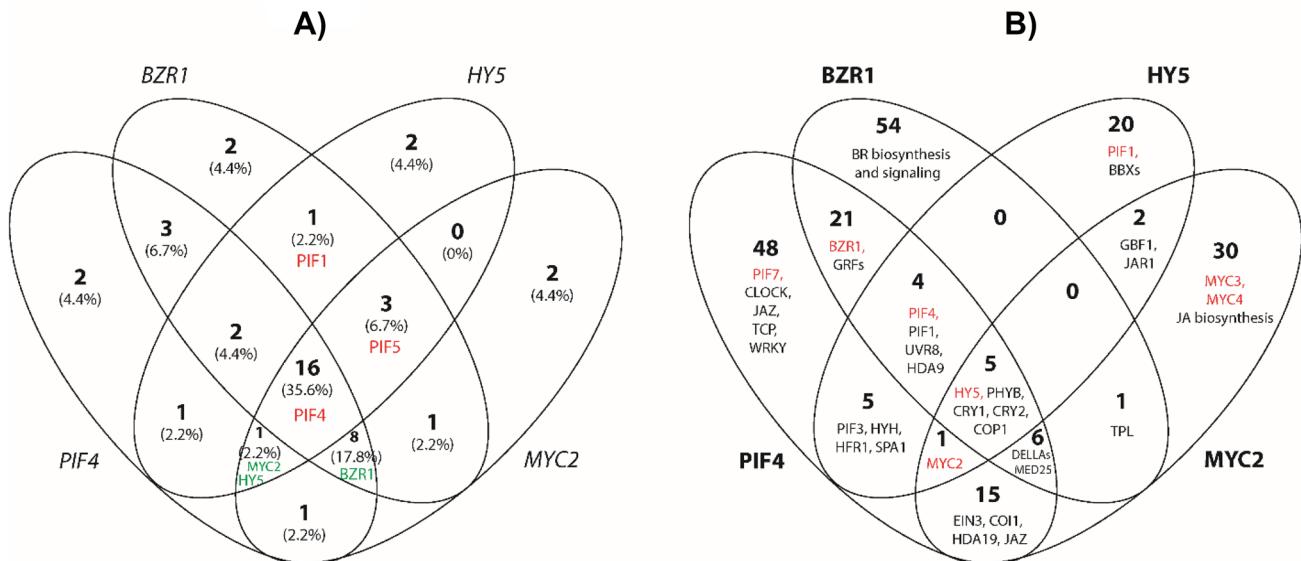


Fig. 2 Overlap in transcription factors and co-factors acting on *PIF4*, *BZR1*, *HY5*, or *MYC2*. **A** Overlap in validated TFs binding to the promoter of *PIF4*, *BZR1*, *HY5*, or *MYC2* (data from PlantPan3.0). Positions of PIF1, PIF4 and PIF5 are indicated in red. Positions of MYC2, HY5, and BZR1 are indicated in green: data from (Lee et al. 2007, Yu et al. 2011, Binkert et al. 2014, Oh et al. 2014, You et al. 2019). PIF4 interacts with the promoter of both *PIF4*, *BZR1*, *HY5*, and *MYC2*. However, PIF4 alone does not affect the expression of these genes (see Table S2). The 15 additional TFs binding to the promoter

of both *PIF4*, *BZR1*, *HY5*, and *MYC2* are ABF1, ABF3, ANAC032, AT5G04760, BBM, GBF2, HB5, HB6, LEC1, MYB44, NFYC2, RD26, ZAT6. For a description of the action of these co-factors see Table S2-5. Although DNA ChIP experiments do not show the binding of PIF3 to any of the four key TF genes, PIF3 is involved in the expression of *PIF4* and *HY5*. **B** Overlap in co-factors acting on *PIF4*, *BZR1*, *HY5*, or *MYC2* protein (see Table S2-S5) The positions of PIFs, MYCs, BZR1, and HY5 are indicated in red

own protein stability by activating COP1 transcription under UV light (Huang et al. 2012), and HY5 represses transcription of MYC2 (Chakraborty et al. 2019). COP1 is required for the stabilization of PIF proteins in the dark (Gangappa and Kumar 2017) and COP1 interacts with and degrades the phosphorylated (inactive) BZR1 protein. While this decreases the total pool of BZR1 protein, this may increase the chance of active BZR1 homodimer formation (Kim et al. 2014). COP1 has a negative effect on HY5 protein levels, as it targets HY5 and the positive acting TFs for HY5 transcription (BBX, LAF1, HFR1) for degradation (Saijo et al. 2003; Gangappa and Botto 2016). Under UV light, COP1 stabilizes GBF1, a negative acting TF for HY5 transcription, but also HYH protein, a positive acting TF for HY5 transcription (Gangappa and Botto 2016). Finally, COP1 is also required for the degradation of MYC2 in the dark (Chico et al. 2014). The red-light activated PHYB^{Pfr} interacts with PIF proteins and targets them for degradation (Huq and Quail 2002; Lorrain et al. 2008). Because BZR1 and PIF4 share common co-factor target genes, the effect of PHYB on PIF4 protein levels indirectly affects the downstream activity of BZR1 (Kim et al. 2014). PHYB^{Pfr} prevents the interaction of HY5 protein with the COP1/SPA complex and thus results in the stabilization of HY5 protein (Lu et al. 2015). Finally, PHYB^{Pfr} increases MYC2 protein stability, but at the same time enhances the stability of JAZ repressor proteins,

resulting in suppression of MYC2 target gene expression (Chico et al. 2014). Figure 1B summarizes the different types of co-factors that may have either positive or negative effect on PIF4, BZR1, HY5, or MYC2 protein activity. Co-factors that influence PIF4, BZR1, HY5, or MYC2 may vary from TFs, kinases, phosphatases, components of protein complexes involved in protein degradation and proteins that can sequester these TFs or enzymes involved in hormone biosynthesis that leads to activation of BZR1 or MYC2.

Omics Meta-Analysis to Uncover Potential Feedback Regulation on PIF4, BZR1, HY5, and MYC2 Activity

The inventory of TFs and co-factors acting on PIF4, BZR1, HY5, and MYC2 was used to uncover the potential for feedback regulation. PIF4, BZR1, HY5, and MYC2 may feedback on their own activity through transcriptional feedback on the genes encoding co-factors that act on PIF4, BZR1, HY5, or MYC2 protein. The potential for (in)direct auto-feedback on the four TFs through their respective co-factor genes is determined from available ChIP-seq sequence data using tagged PIFs (Oh et al. 2012; Pfeiffer et al. 2014), BZR1 (Oh et al. 2012, Oh et al. 2014) or BES1 (Yu et al. 2011), HY5 (Lee et al. 2007), or MYC2 protein (You et al. 2019). Whether binding of the TF to the promoter of its co-factor gene actually results in transcriptional regulation was

determined from a comparison of the transcriptome in WT and the different *pif* mutants (Leivar et al. 2012; Oh et al. 2012; Huai et al. 2018) *bzr1-1D* mutant (Oh et al. 2014), *hy5* mutant (Zhao et al. 2019), or *myc2/3/4* triple mutant (Van Moerkerke et al. 2019). When PIF4 binds to the promoter of a TF-gene or co-factor gene involved in regulating *PIF4* transcription or activity, this is counted as a potential for transcriptional auto-feedback regulation by PIF4 (idem for BZR1, HY5, and MYC2). In addition to *PIF4* ChIP-seq data, also ChIP-seq binding data by PIF1, PIF3, or PIF5 protein is included in the inventory because of potential redundancy by other PIFs in co-regulation of PIF4 target genes. The contribution of just PIF4 in regulating transcription of a target gene may not be noticeable in a *pif4* single-mutant background. Therefore, transcriptome analysis of the *pif1/3/4/5* quadruple (*pifq*) mutant was included in the transcriptome analysis (Pfeiffer et al. 2014). Altered expression of PIF4-related co-factor genes in *pifq* mutant background was treated here as an indication of potential feedback regulation by PIF4. Similarly, the contribution of only MYC2 in regulating transcription of a target gene may not be noticeable in a *myc2* single-mutant background. Therefore, regulation of MYC2 co-factor genes by MYC2 was derived from the comparison of the transcriptome in *myc2/3/4* triple mutant versus WT (Schweizer et al. 2013; Van Moerkerke et al. 2019) or 35S-MYC2 lines versus WT (Chini et al. 2007). Since the full knock-out mutant for *BZR1* is lethal, transcriptional regulation by BZR1 was from comparing WT with the *bzr1-1D* mutants, in which BZR1 protein is constitutively active due to point mutation in the phosphorylation site (Wang et al. 2002) and from WT plants treated with an inhibitor of endogenous BR biosynthesis which prevents activation of endogenous BZR1/BES1 and from the comparison of *pifq/bzr1-1D* versus *pifq* (Oh et al. 2012; Oh et al. 2014).

Using a comprehensive inventory of relevant ChIP-seq and RNA-seq data sets the potential for direct and indirect feedback on the four selected TFs was determined (see Supplement Methods). Although some feedback on the four selected TFs has been described in the literature, the inventory identifies numerous new potential feedbacks on PIF4, BZR1, HY5, and MYC2 at (post)-transcriptional level and indicates for each TF both positive and negative feedback that may help to maintain homeostasis. Tables 1, 2, 3, and 4 list all the co-factor genes for which either binding to the promoter or transcriptional regulation by PIF4, BZR1, HY5, or MYC2 was identified. In each table, the transcriptional regulation of co-factor genes that results in negative feedback on *PIF4*, *BZR1*, *HY5*, or *MYC2* transcription/activity respectively is indicated in red, while transcriptional regulation of co-factor genes that result in positive feedback on *PIF4*, *BZR1*, *HY5*, or *MYC2* transcription/activity respectively is indicated in green. Overall, there is little overlap between the binding of PIF4, BZR1, HY5, and MYC2 to

their respective co-factor genes (ChIP-seq data) and regulation of their own co-factor genes (RNA-seq data), confirming a general lack of overlap between the ChIP-seq and RNA-seq data sets that have been observed before (Pfeiffer et al. 2014). However, since here data was collected from different publications, the lack of overlap in ChIP-seq and RNA-seq data may also be due to different experimental conditions used in ChIP-seq and RNA-seq experiments.

For discussion of results, a change in transcription of a co-factor gene is assumed to be limiting for feedback on the TF that regulates its expression. Because of the redundancy in the regulation of genes by PIFs, we will treat the transcriptional results for *pifq* mutants as being relevant for regulation by PIF4. Similarly, because of the redundancy in the regulation of genes by MYCs results for the triple mutant *myc2/3/4* (*myct*) are treated as being relevant for regulation by MYC2.

Transcriptional Feedback on and by PIF4

Of the 34 different TFs with validated binding to the promoter of *PIF4* (Fig. 2), the literature search identified 13 positive acting TFs and eight negative acting TFs on *PIF4* expression (Fig. 3). Although PIF4 and other PIFs bind to the *PIF4* promoter, the role of PIF4 (and other PIFs) in the transcription of the *PIF4* gene has not been studied extensively. Ectopic overexpression of PIF4 results in suppression of the endogenous *PIF4* gene (Shapulatov 2019). Moreover, *PIF4* expression is also upregulated in a *pif3* mutant compared to WT, but not in a *pif1* or *pif5* single mutant (Zhang et al. 2013). Overall, this suggests that PIF4 and PIF3 may be negative regulators of *PIF4* transcription. Some of the negative TFs acting on *PIF4* transcription are part of the clock: components of the evening complex (*LUX*, *ELF3*, *ELF4*: (Nusinow et al. 2011, Mizuno et al. 2014, Nieto et al. 2015, Raschke et al. 2015, Ezer et al. 2017)), clock genes *PRR9* (Zhu et al. 2016; Li, Zhang et al. 2020a, b, c), *PRR7* (Mizuno et al. 2014), *PRR5* (Zhu et al. 2016), and the clock gene *GI* (Zhu et al. 2016). In contrast, clock genes *LHY* and *CCA1* have a positive effect on *PIF4* transcription (Sun et al. 2019). The transcription feedback by PIF4 on some of the clock components (Fig. 3, feedback 3 and 4) suggests an intricated balance between growth and circadian timekeeping that may result in finetuned gating of PIF4 activity over the day. Gating of PIF4 activity by the clock has also been attributed to the interaction between TOC1 and PIF4 (Soy et al. 2016; Zhu et al. 2016), but transcription of *TOC1* is not under direct feedback regulation by PIF4.

The transcription factors MYB30 (Bertoni 2020; Yan et al. 2020), BZR1 (Ibanez et al. 2018) (Oh et al. 2012), a subset of TCPs (Han et al. 2019), a subset of WRKYs (Foreman et al. 2011; Sun et al. 2020), and COG1 (Wei et al. 2017) all have been identified as positive regulators of PIF4

Table 1 (Potential) Transcriptional feedback by PIF4 on positive/negative regulators of PIF4. ChIP PIF bound: genes bound by PIF proteins in DNA ChIP-seq experiments (positive hits indicated in grey; numbers indicate bound PIF family member(s)). Transcriptome data red cell: transcriptional regulation of co-factor gene by PIF4/PIFs, resulting in negative feedback on PIF4. Green cell: transcrip-

tional regulation of co-factor gene by PIF4/PIFs, resulting in positive feedback on PIF4. *: Feedback by PIF4 itself: data from (Zhang et al. 2013; Shapulatov 2019). References: 1: (Leivar et al. 2009; Oh et al. 2012); 2: (Oh et al. 2012); 3: (Pfeiffer et al. 2014); 4: (Leivar et al. 2009); 5: (Leivar et al. 2009)

			1	2 5d D on BRZ	3 2d D	4 2d L/D	5 2d D	6 5d cR
Pos. regulators	GENE ID	CHIP- SEQ PIFs	PIF reg.	WT/pif q	pifq/W T	pifq/W T	pif4/W T	WT/pif 4
LHY	AT1G01060		D					
CCA1	AT2G46830		C	U	U			
BZR1	AT1G75080		D					
SHB1	AT4G25350		D					
PIF7	AT5G61270			D	U			
ARF6	AT1G30330	1/4/5	U					
TCP5	AT5G60970		D					
TCP13	AT3G02150	1/3/4/5	D	D	U			
CBF1	AT4G25490	5	D		D			
WRKY33	AT2G38470	5	D		U			
WRKY26	AT5G07100							D
HLS1	AT4G37580		C	U				
SPA1	AT2G46340	1/4/5	D		U			
COP1	AT2G32950		D					
JAZ8	AT1G30135				D		D	D
JAZ9	AT1G70700		D	D				
JAZ6	AT1G72450	1/4/5						
JAZ7	AT2G34600		D		D			
JAZ3	AT3G17860	1/3/5	D					
JAZ5	AT1G17380	5						
JAZ10	AT5G13220		D					
JAZ1	AT1G19180	1/3/4/5	U				D	D
HMR	AT2G34640	5		D	D			
PRE1	AT5G39860	1/5	D					U
PHYA	AT1G09570	4 (Oh, Zhu et al. 2012)						
GID1B	AT3G63010	1/4/5	D					
Neg. regulators	GENE ID	CHIP PIF	PIF reg.	WT/pif q	pifq/W T	pifq/W T	pif4/W T	WT/pif 4
BOP2	AT2G41370	1						
PRR9	AT2G46790	3						
PRR7	AT5G02810			U				D
PRR5	AT5G24470	4			U			
GI	AT1G22770	3						
HY5	AT5G11260	1/4	U					
HYH	AT3G17609			D				
PIF4	AT2G43010	5						*
PIL1	AT2G46970		U	U	D			U
HFR1	AT1G02340	1/3/4/5	U	U	D			U
MYC2	AT1G32640		U		D			
SPCH	AT5G53210			D				D
PAR1	AT2G42870	1/3/4/5	U	U				D
PAR2	AT3G58850	1/4/5	U	U				D
HEC1	AT5G67060							D
HEC2	AT3G50330			U				D
SOB3/AHL29	AT1G76500	1/3/4/5	U	U				
ESC/AHL27	AT1G20900	1/3/4/5	U	U	D			
RGA1	AT2G01570	1/3/4/5	U	U				
RGA2	AT1G14920	1/3/4/5	U	U				
RGL1	AT1G66350	1/5	U					U
RGL2	AT3G03450				D			
RGL3	AT5G17490	1						U
PHYB	AT2G18790	1/3/4/5	U	U				
CRY2	AT1G04400		U	U				
UVR8	At5G63860		U	U				
HDA15	AT3G18520		U					
BIN2	AT4G18710	1/5						
GRF3	AT5G38480	1/4						
GRF8	AT5G65430		U	U				
GRF13	AT1G78220	3						
GRF2	AT1G78300	1/5						

Table 2 (Potential)

Transcriptional feedback by BZR1 on positive/negative regulators of BZR1. ChIP BZ1/BE1: genes bound by BZR1 (BZ1) or BES1 (BE1) proteins in DNA ChIP-seq experiments (positive hits indicated in grey). Transcriptome data: Red cell: transcriptional regulation of co-factor gene by BZR1/BES1, resulting in negative feedback on BZR1. Green: transcriptional regulation of co-factor gene by BZR1/BES1, resulting in positive feedback on BZR1. References: 1: (Yu et al. 2011; Oh et al. 2014); 2: (Sun et al. 2010); 3: (Oh et al. 2012); 4: (Oh et al. 2012)

		1	2 5d dark/ 5d dark on BR	3 5d dark on 2 μ M BRZ	4 5d dark on 2 μ M BRZ
Pos. regulators	Gene ID	ChIP BZ1/BE1	WT on BR /WT	<i>bzr1-ID / Col-0</i>	<i>pifq;bzr1-ID/pifq</i>
BIM1	AT5G08130	BZ1		D	
COP1	AT2G32950	BZ1		D	
PIF4	AT2G43010	BZ1		D	
MYB30	AT3G28910	BE1	U	D	
BES1	AT1G19350		U		
BZR1	AT1G75080	BZ1	U		
ARF6	AT1G30330	BZ1/BE1	D		
ARF8	AT5G37020	BZ1	D		
PIF1/PIL5	AT2G20180	BZ1			
WRKY46	AT2G46400	BZ1		U	
WRKY54	AT2G40750	BZ1			
WRKY70	AT2G40750	BZ1			
ELF6	AT5G04240		D		
MED25	AT1G25540	BZ1/BS1			
BSU1	AT1G03445			U	
OCTOPUS	AT3G09070	BZ1			
Polar	AT4G31805	BZ1		D	
BASL	AT5G60880	BZ1		D	
BRI1	AT4G39400	BZ1/BE1	D	D	
BRL1	AT1G55610	BZ1			
SERK3/BA K1	AT4G33430	BZ1/BE1		D	
BIR3	AT1G27190		D		
BSK3	AT4G00710	BZ1		D	
DWF4	AT3G50660	BZ1/BE1	D	D	D
CPD/DWF3	AT5G05690	BZ1/BE1	D		
CYP90D1	AT3G13730	BZ1/BE1	D	D	D
ROT3	AT4G36380	BZ1/BE1	D	D	
BR60x2	AT3G30180	BZ1	D	D	D
DWF7	AT3G02580	BZ1		D	
Neg. regulators	Gene ID	ChIP BZ1/BE1	WT on BR /WT	<i>bzr1-ID / Col-0</i>	<i>pifq;bzr1-ID/pifq</i>
SINAT2	AT3G58040		D		
ATG8F	AT4G16520	BZ1			
AtD14	AT3G03990	BZ1		D	
MYBL2	AT1G71030	BZ1/BE1	D	D	
HY5	AT5G11260	BZ1			
HAT1	AT4G17460	BZ1/BE1		D	
ATAF2/AN AC081	AT5G08790	BZ1/BS1		D	
RD26	AT4G27410	BZ1/BE1	D	D	
TINY/ERF0 40	AT5G25810	BZ1		D	
TINY2	AT5G11590	BZ1			
ERF72	AT3G16770	BZ1	U		
BES1	AT1G19350		U		
BZR1	AT1G75080	BZ1	U		
BOP1	AT3G57130	BZ1		D	
BOP2	AT2G41370	BZ1		D	
RGA1	AT2G01570	BZ1			
RGA2	AT1G14920	BZ1/BE1	D		
RGL1	AT1G66350	BZ1	U		
RGL2	AT3G03450	BZ1	U		
RGL3	AT5G17490	BZ1			
BKI1	AT5G42750	BZ1	D		
UVR8	AT5G63860	BZ1		D	
CRY1	AT4G08920	BZ1		D	
PHYB	AT2G18790	BZ1	D		
TPL	AT1G15750	BZ1/BE1	D		
HDA15	AT3G18520		D	D	
BIN2	AT4G18710	BZ1	D		
MKK5	AT3G21220	BZ1			
GRF3	AT5G38480	BZ1			
GRF8	AT5G65430	BS1			

Table 3 (Potential)

Transcriptional feedback by HY5 on positive/ negative regulators of HY5. ChIP-seq HY5 bound: genes bound by HY5 under white light (W), blue light (b), or red light (r). Transcriptome data is from a comparison of 2-day-old seedling WT versus *hy5* grown under continuous R light (cR). UV: upregulation of COP1 by HY5 under UV light. Red: transcriptional regulation by HY5, resulting in negative feedback on HY5. Green: transcriptional regulation by HY5, resulting in positive feedback on HY5. References: 1: (Lee et al. 2007; Binkert et al. 2014); 2: (Zhao et al. 2019); 3: (Binkert et al. 2014); 4: (Huang et al. 2012)

Pos. regulators	Gene ID	1	2
		ChIP HY5	2d cR WT/ <i>hy5</i>
HY5	AT5G11260	b/r	U (3)
HYH	AT3G17609	W/b/r	U (3)
MYC2	AT1G32640	W	D
TCP2	AT4G18390	b/r	
HFR1	AT1G02340	W	
BBX11	AT2G47890	W/b/r	U
BBX21	AT1G75540	W	
BBX22	AT1G78600	W	
PHYA	AT1G09570	W	
PHYB	AT2G18790	b	
FIN219/JAR1	AT2G46370	W/b	
COP1	AT2G32950	b/r	
Neg. regulators	Gene ID	ChIP HY5	Wt/ <i>hy5</i>
BBX24	AT1G06040	W/b/r	
BBX32	AT3G21150	W/b	D
BBX28	AT4G27310	W/b/r	
HY5	AT5G11260	b/r	U
PIF4	AT2G43010	W	
MYC2	AT1G32640	W	D
GBF1	AT4G36730	W/b/r	
WRKY36	AT1G69810	b/r	
SPA1	AT2G46340	b/r	
COP1	AT2G32950	b/r	U (UV) (4)
RUP1	AT5G52250	W/b	
RUP2	AT5G23730	W/b	U
HDA9	AT3G44680	b	

expression. The action of BZR1, CCA1, TCPs, and WRKYs on PIF4 transcription becomes limiting during thermomorphogenesis. The extensive redundancy in TFs regulating PIF4 transcription may hide the contribution of these TFs when the expression of PIF4 is low at normal temperature.

The stimulation by PIF4 of BR biosynthesis gene expression during thermomorphogenesis and the resulting activation of BZR1 forms a positive feedback loop on PIF4 transcription by BZR1, which has previously been recognized (Ibanez et al. 2018) (Fig. 3: positive feedback 1). Our inventory identifies additional potential positive feedback on PIF4 through the positive regulator of PIF4, COG1 (Fig. 3, positive feedback 2) and through downregulation by PIF4 of the repressors of PIF4: *SPCH*, *PRR7*, and *PRR5* (Lau et al. 2018) (Fig. 3, positive feedback 3). It has previously been recognized that positive feedback on PIF4 transcription poses a potential danger for out-of-control upregulation of PIF4 transcription (Ibanez et al. 2018). However, our inventory identifies multiple negative feedback interactions on PIF4 transcription or PIF4 transcriptional activity that may balance the positive feedback loops: PIF4 represses transcription of the positive TFs *TCP13*, *LHY*, *WRKY26*, *WRKY33*, and *PIF7* (Fig. 3: negative feedback 4). In addition to these multiple potential negative feedback on transcription

of the *PIF4* gene, there is also multiple negative feedback on the activity of PIF4 protein, as PIF4 upregulates six TFs that interfere with PIF4 transcriptional activity (Fig. 3: negative feedback 5). Finally, the PIF4 protein itself may play an important role in limiting the upregulation of PIF4 transcription as PIF4 is a negative regulator of its own expression (Fig. 3: negative feedback 6).

Because feedback on the photoreceptors which affect the stability of PIF4 protein is both through PIF4 and BZR1 this is discussed separately below (Fig. 8).

Transcriptional Feedback on and by BZR1

For the 36 different TFs with validated binding to the promoter of *BZR1* (Fig. 2), evidence for actual transcription regulation of *BZR1* is not available. The binding of PIF1, PIF4, PIF5, and HY5 to the promoter of *BZR1* does not result in altered transcription of *BZR1* in *PIF* or *HY5* mutants (Table 2). *BZR1* can bind to its own promoter (Fig. 2) and *BZR1* gene transcription is upregulated by the application of BR, which activates BZR1, suggesting that transcription of *BZR1* is under positive auto-feedback regulation (Fig. 4; positive feedback 1). However, transcription of *BZR1* is not upregulated in the *bzr1-1D* mutant

Table 4 (Potential) Transcriptional feedback by MYC2 on positive/ negative regulators of MYC2. ChIP MYC bound: genes bound by MYC2. Transcriptome data red cell: transcriptional regulation of co-factor gene by MYC2/3/4 resulting in negative feedback on MYC2. Green: tran-

scriptional regulation of co-factor gene by MYC2/3/4, resulting in positive feedback on MYC2. References: 1: (You et al. 2019); 2: (Chini et al. 2007); 3: (Schweizer et al. 2013); 4: (Van Moerkerke et al. 2019)

		28d 16L/8D	1 14d 16L/8D	2 28d 16L/8D	3 15d 12L/12D
Pos. regulator	Gene ID	CHIP MYC2	35S- MYC2	WT/myc2/3/4	WT/myc2/3/4
MYC2	AT1G32640			U	
MYC4	AT4G17880			U	
LOX3	AT1G17420				
AOC1	AT3G25760			U	
AOC2	AT3G25770			U	
AOC3	AT3G25780			U	
OPR3	AT2G06050			U	
CRY2	AT1G04400				
RGA2	AT1G14920				
GA2OX2	AT1G30040				
GSTU20	AT1G78370			U	D
Neg. regulator	Gene ID	CHIP MYC2	35S- MYC2	WT/myc2/3/4	WT/myc2/3/4
JAM2/bhlh13	AT1G01260				
MYC2	AT1G32640			U	
JAZ8	AT1G30135		-		
JAZ9	AT1G70700		U	U	
JAZ7	AT2G34600		U		
JAZ3	AT3G17860		U		
JAZ1	AT1G19180		U		
JAZ5	AT1G17380		U		
JAZ10	AT5G13220		U	U	
JAZ12	AT5G20900		U		
TPL	AT1G15750				
GA2OX2	AT1G30040				
RGA2	AT1G14920				

background, suggesting that BZR1 alone is not sufficient for auto-feedback on *BZR1* transcription. In total six positive feedback loops on BZR1 are identified, involving 13 co-factor genes (Fig. 4). BZR1 upregulates transcription of TF *WRKY36*, which enhances BZR1 target gene expression (Chen et al. 2017). Since BZR1 may target its own expression the upregulation of *WRKY36* forms potential positive feedback on BZR1 (Fig. 4, positive feedback 2). The negative regulators of BZR1 protein HAT1, RD26, MYBL2, TINY, and HDA15 all are downregulated by BZR1, thus forming additional potential positive feedback on BZR1 (Fig. 4; positive feedback 3). The transcriptional feedback on BZR1-related co-factor genes was investigated in dark grown seedlings treated with BR, which activates BZR1 and BES1 (Sun et al. 2010) and in dark grown *bzr1-1D*

mutant seedlings treated with Brassinazole (BRZ) to inhibit endogenous BR biosynthesis. The former experiment reveals targets of both BZR1 and BES1. The latter may reveal the direct targets of only BZR1 (in the form of constitutive active *bzr1-1D*), since activation of BES1 is limited through inhibition of endogenous BR biosynthesis by BRZ (Oh et al. 2012). The brassinosteroid-insensitive 2 (BIN2) is a kinase and a negative regulator of BZR1 protein stability and nuclear transport. The activity of BIN2 is inhibited by the activity of phosphatase BSU1 (Kim et al. 2009, 2011) and transcription of BSU1 is stimulated by BZR1, which thus forms potential positive feedback on BZR1 activity (Fig. 4, positive feedback 6). The potential reduction in BIN2 activity through upregulation of BSU1 may also lead to reduced stability of the BES1/

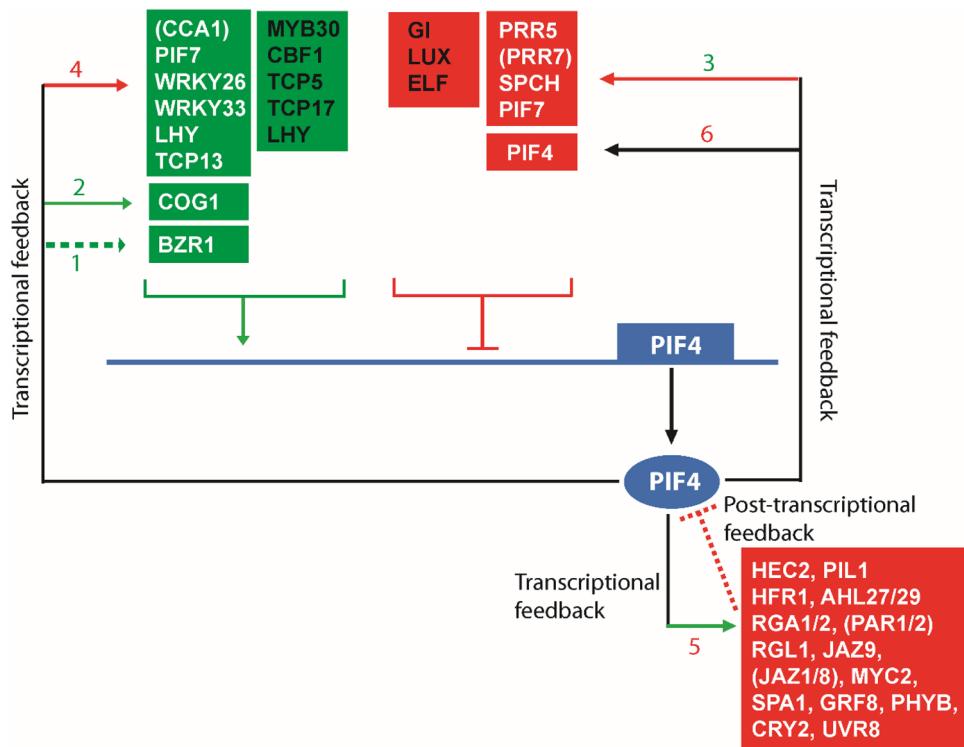


Fig. 3 Potential feedback interactions between TFs and co-factors acting on PIF4. PIF4 gene is indicated by blue line (promoter) and blue rectangle (coding sequence). PIF4 protein is indicated by blue oval. TFs with positive/negative action on transcription of the PIF4 gene are indicated in green/red blocks. TFs in black are not under transcriptional feedback by PIF4. Solid red/green arrows: negative/positive transcriptional feedback by PIF4. Dashed red lines: negative/positive feedback on transcriptional activity of PIF4 protein. Dashed green arrow: indirect positive feedback on PIF4 transcription by PIF4 through upregulation BR synthesis and signaling which activates BZR1 (posi-

tive feedback 1). COG1 is upregulated by PIFs in a *bzr1-1D* mutant background (Oh et al. 2012) and interaction is assumed to also take place in a WT background (potential positive feedback 2). Negative feedback 5 indicates upregulation of co-factors repressing PIF4 protein activity (for factors between parenthesis transcriptional feedback by PIF4 is condition dependent; see Table 1). Black arrow: negative feedback of PIF4 on its own expression (negative feedback 6). Red/green numbers: negative/positive feedback on PIF4. Sources for RNA-seq data related to PIFs are in legend Table 1

BZR1 corepressors HAT1, RD26, TINY, and MYBL2 as all these co-repressors are substrates of BIN2 (Ye et al. 2012; Zhang et al. 2014; Jiang et al. 2019; Xie et al. 2019). While phosphorylation of BES1/BZR1 by BIN2 leads to protein degradation, BIN2 phosphorylation of HAT1, RD26, TINY, and MYBL2 stabilizes these corepressors of BZR1 activity. Therefore, reduced BIN2 activity could reduce the suppressing effect of HAT1, RD26, TINY, and MYBL2 on BZR1, resulting in an indirect positive feedback on BZR1 (Fig. 4; positive feedback through a). BZR1 is also involved in direct downregulation of *HAT1*, *RD26*, *TINY*, and *MYBL2* (Table 2), resulting in another potential positive feedback on BZR1 (Fig. 4, positive feedback 3). BOP1 (and by analogy BOP2) inhibits the transport of BZR1 from the cytosol to the nucleus and thus negatively regulates BZR1 activity. The downregulation of BOP1 and BOP2 by BZR1 thus forms additional positive feedback on BZR1 (Fig. 4, positive feedback 4). Active BRs are conjugated by ATAF2, and upregulation of ATAF2 by BZR1

thus forms another potential positive feedback on BZR1 (Fig. 4, positive feedback 5). Thus, the negative feedback on BZR1 through BR biosynthesis and direct action of BIN2 on BZR1 (Fig. 4, negative feedback 7) is balanced by multiple potential positive feedback loops on BZR1 (Fig. 4, positive feedback 1–6, 8 and a). It has already been described that transcription of some key genes in BR biosynthesis is under negative control by BES1 (Kim et al. 2009) and transcriptome analysis of the *bzr1-1D* mutant indicates a similar role for BZR1 in the suppression of BR biosynthesis genes (Table 2). The transcriptome of the *bzr1-1D* mutant reveals that also several genes encoding components of the BR receptor complex are also under negative transcriptional feedback by BZR1 (Table 2). Because feedback on the photoreceptors which affect the stability of BZR1 protein (Fig. 4 positive feedback 8) is both through BZR1 and PIF4 this is discussed separately below (see Fig. 8).

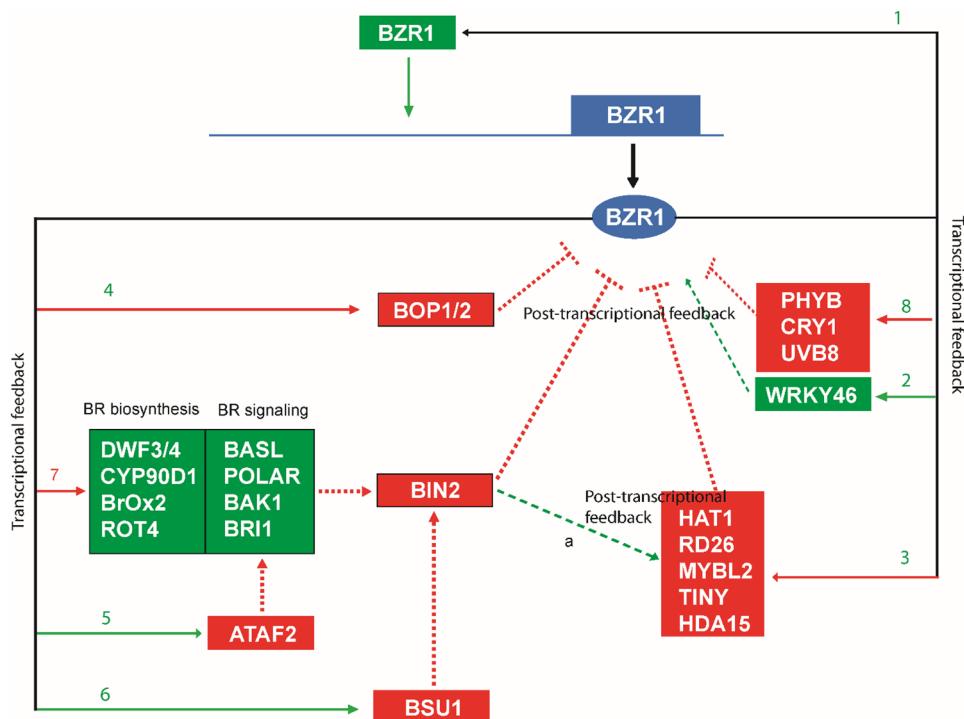


Fig. 4 Potential feedback interactions between TFs and co-factors acting on BZR1. BZR1 gene is indicated by a blue line (promoter) and blue rectangle (coding sequence). BZR1 protein is indicated by blue oval. In the presence of BR signaling, BZR1 may show positive feedback on its own transcription through feedback loops 1, 2, 3, 4, 5, 6, and 8. These potential feedforward loops are balanced by negative feedback through 7, which results in suppression of BR biosynthesis

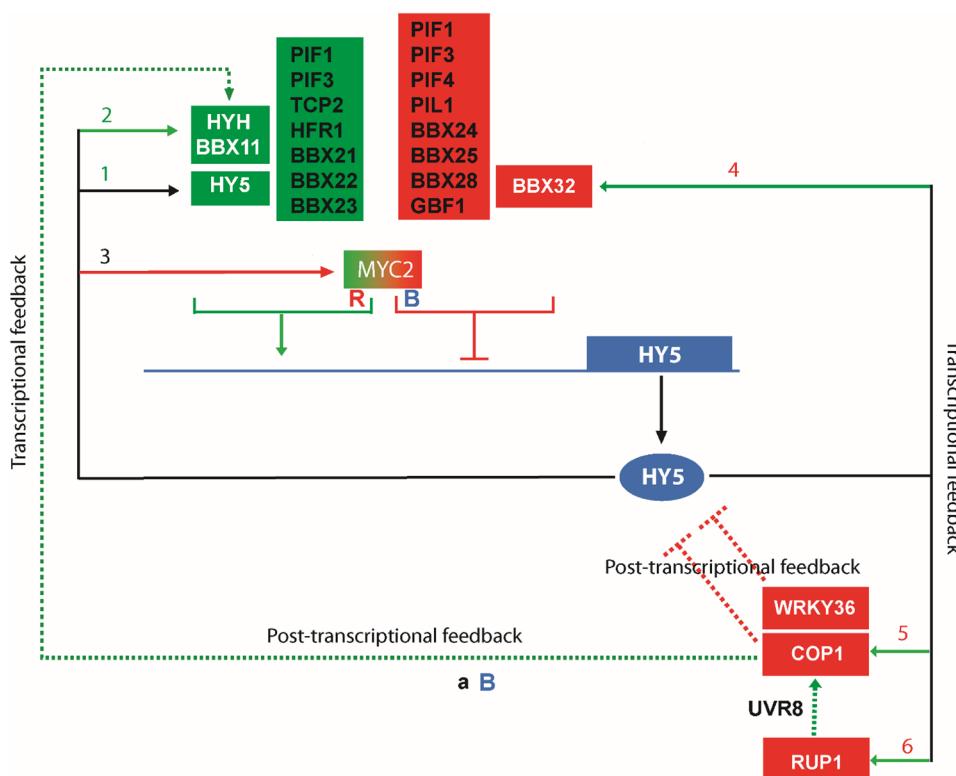
and signaling and downregulation of BOP1/2 which targets BZR1. Solid red/green arrows: negative/positive transcriptional feedback by BZR1. Dashed red lines: indirect negative feedback on BZR1 protein. Dashed green arrow: indirect positive feedback on BZR1 protein. Red/green numbers: negative/positive feedback loops on BZR1. Sources for RNA-seq data related to BZR1 are in legend Table 2

Transcriptional Feedback on and by HY5

From the 26 different TFs with validated binding to the promoter of *HY5* (Fig. 2), our literature search identifies 20 transcription factors for which either positive or negative transcription regulation of the *HY5* gene has been demonstrated. However, most of these validated TFs acting on *HY5* are not under transcriptional feedback by *HY5* (Fig. 5, indicated in black; see Table S4). The binding of PIF4 to the promoter of *HY5* does not correlate to the transcriptional regulation of *HY5* by PIF4 or PIFs (Table 3). A literature search identifies nine positive-acting TFs and nine negative-acting TFs for *HY5* transcription, while the action of TF MYC2 on *HY5* transcription depends on light color (Ortigosa et al. 2020). The PlantPan3.0 software does not predict a *HY5* binding site in the promoter of *HY5* and ChIP-seq data show *HY5* binding to the promoter of *HYH* (a close homolog of *HY5*), but no binding to the promoter of *HY5* itself (Lee et al. 2007). However, binding of *HY5* (and *HYH*) to T/G box in the promoter of *HY5* was detected by Binkert, using dedicated ChIP qPCR analysis rather than ChIP-seq, and a potential positive auto-feedback regulation of *HY5* transcription by *HY5* was demonstrated (Binkert

et al. 2014) (Fig. 5: positive feedback 1). In addition, there is potential positive feedback through upregulation of positive acting TFs *HYH* and *BBX11* by *HY5*, especially under UVB light (Binkert et al. 2014; Yang et al. 2018; Chakraborty et al. 2019) (Fig. 5; positive feedback 2). *BBX32* protein suppresses the activity of positive action of TF *BBX21* on *HY5* (Holtan et al. 2011). Therefore, suppression of *BBX32* transcription by *HY5* constitutes another potential indirect positive feedback regulation on *HY5* activity (Fig. 5; positive feedback 4). Transcriptional regulation of *HY5* by *MYC2* is complicated as *MYC2* stimulates *HY5* expression under Red (R) light (Ortigosa et al. 2020), but represses *HY5* transcription under Blue (B) light (Chakraborty et al. 2019) (Fig. 5; feedback 3). Under B light, the negative feedback on *MYC2* thus results in potential positive feedback on *HY5*, while under R light the negative feedback on *MYC2* results in potential negative feedback on *HY5*. In addition, under B light *COP1* enhances the stabilization of *HYH* (Gangappa and Botto 2016), resulting in potential positive feedback on *HY5* transcription by *HYH* (Fig. 5, positive feedback a). Under white light, the actual feedback may depend on the relative contribution of R and B components in the white light. The different positive feedback on *HY5* is compensated

Fig. 5 Potential feedback interactions between TFs and co-factors acting on HY5. HY5 gene is indicated by blue line (promoter) and blue rectangle (coding sequence). HY5 protein is indicated by blue oval. TFs with positive action on HY5 are indicated in green blocks, and TFs with negative action on HY5 are indicated in red blocks. TFs in black letters are not under transcriptional feedback by HY5. Solid red/green arrows: negative/positive transcriptional feedback by HY5. Dashed red lines: indirect negative feedback on HY5 protein. Dashed green arrow a: indirect positive feedback on HY5 under B light. Red/green numbers: negative/positive feedback loops on HY5. R: red light, B: blue light. The feedback through RUP1 and UVR8 is relevant under UV light. Sources for RNA-seq data related to HY5 are in legend Table 3



by negative feedback through upregulation of TF *WRKY36*, which blocks HY5 transcriptional activity, and upregulation of *COP1*, which targets HY5 protein for degradation (Fig. 5; negative feedback 5). The upregulation of *RUP2* by HY5 was only uncovered by dedicated qPCR experiments under UV light conditions (Binkert et al. 2014). *RUP2*, together with *RUP1* simulates UVR8 dimer formation, ending signaling of the UVR8 monomer in response to UV-B light. UVR8 monomer interacts with and represses *WRKY36*, a negative transcriptional regulator of HY5 and the UVR8 monomer disrupts *COP1* function for degradation of HY5. Thus, UV light initially may result in enhanced transcription and protein stability of HY5. *RUP1* and *RUP2* are two proteins that promote UVR8 dimer formation and are involved in the termination of UVR8 monomer signaling. The transcriptional upregulation of both *COP1* and *RUP2* by HY5 (Huang et al. 2012), potentially terminates and limits UVR8 monomer activity on *WRKY49* and *COP1* (Binkert et al. 2014) (Fig. 5; negative feedback 6).

Transcriptional Feedback on and by MYC2

For most of the different TFs with validated binding to the promoter of *MYC2* (Fig. 2) evidence for actual transcription regulation of *MYC2* is not available. The binding of *PIF4* to the promoter of *PIF4* does not correlate to transcriptional regulation of *MYC2* by *PIF4* or *PIFs* (Table 4). It has been shown that binding *MYC2* to its own promoter does indeed

result in a direct positive auto-feedback regulation by *MYC2* during short-term JA signaling (Van Moerkerke et al. 2019; Wang et al. 2019). However, under prolonged JA signaling *MYC2* exerts negative feedback on its own transcription (Wang et al. 2019). In addition, HY5 has been identified as a negative regulator of *MYC2* transcription in combination with an unknown factor that facilitates (indirect) binding of HY5 to the *MYC2* promoter (Chakraborty et al. 2019) (Fig. 6. Feedback 1). *MYC2* upregulates the expression of *MYC4*, which assists *MYC2* in targeting downstream genes (Table 4; Fig. 6, positive feedback 2). In addition, there is positive feedback through JA/JAile signaling by transcriptional upregulation of JA biosynthesis genes by *MYCs*, and upregulation of *GSTU20* which stimulates the conversion of JA to the active signaling component JAile. The potential higher JA and JAile levels may increase JA signaling which targets *JAZ* proteins (transcriptional inhibitors of *MYC2* activity) for destruction (Fig. 6, positive feedback 3). The feedback on JA signaling enhances *MYC2* activity, as activation of *MYC2* by JA-ile signaling leads to the destruction of repressor *JAZ* proteins that block *MYC2* activity. These positive feedback loops on *MYC2* are compensated by two negative feedbacks on *MYC2*: *MYC2* activity results in the upregulation of several *JAZ* genes, potentially enhancing the suppression of *MYC2* activity (Fig. 6, negative feedback 4) (Chini et al. 2007). Also, HY5 is upregulated by *MYC2*, while HY5 together with an unidentified factor downregulates *MYC2* transcription (Chakraborty et al. 2019) (Fig. 6;

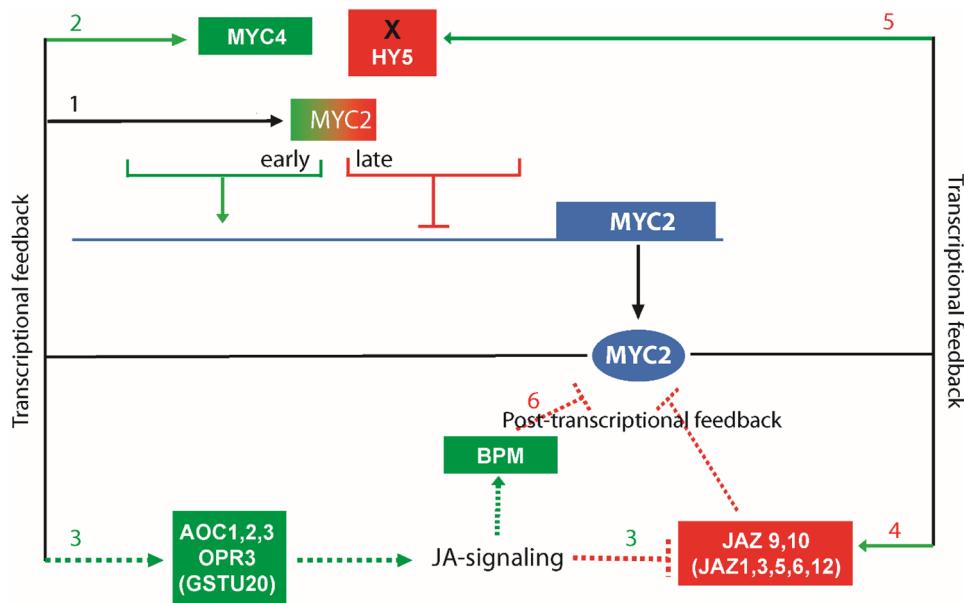


Fig. 6 Potential feedback interactions between TFs and co-factors acting on MYC2. MYC2 gene is indicated by blue line (promoter) and blue rectangle (coding sequence). MYC2 protein is indicated by blue oval. TFs with positive/negative action on MYC2 are indicated in green/red. TF X works together with HY5 on the transcription of MYC2. Solid red/green arrows: negative/positive transcriptional feedback by MYC2. Dashed red lines: negative interaction on MYC2 protein. Dashed green arrow a: positive effect on JA by co-factor protein

negative feedback 5). BPM1 is part of a protein complex that targets MYC2 for degradation and JA signaling stabilizes BPM1 (Chico et al. 2020). Therefore, the positive feedback on JA synthesis and subsequent signaling can result in both a positive effect on MYC2 (through the removal of JAZ proteins) and negative effect on MYC2 (through stabilization of BPM1) (Fig. 6, positive feedback 3, negative feedback 6). The inventory of RNA-seq data shows that transcriptional feedback on MYC2 co-factor genes seems to be more extensive in rosette plants than in seedlings (Table 4). Moreover, feedback on GSTU20 by MYC2/MYCs was the opposite in seedlings and rosette plants (Table 4).

Potential for Cross-Regulation of Co-factor Genes by PIF4, BZR1, HY5, and MYC2

Besides feedback of each TF on its own co-factor genes (Fig. 1A pathway b), each of the four TFs may also interact through regulation of each other's co-factor genes (Fig. 1A, pathway c). Figure 7A shows which BZR1-, HY5- and MYC-related co-factor genes are upregulated or downregulated by PIFs/PIF4. Results indicate the potential for interaction between the PIF4 activity through indirect feedback on the other TFs. However, there is no preferred positive or negative feedback by PIF4 on either BZR1, HY5, or

action. Early: positive feedback by MYC2 on its transcription in early response, late: negative feedback by MYC2 on its own expression in late response to JA. Red/green numbers: negative/positive feedback loops on HY5. (GSTU20): transcriptional feedback by MYC2 is different in seedlings and rosette plants (Table 4). (JAZ1,3,5,6,12): these JAZ genes are under negative transcriptional control in 35S-MYC2 overexpression lines (Table 4). Sources for RNA-seq data related to MYCs are in legend Table 4

MYC2 transcription/activity through transcriptional regulation of their respective co-factor genes by PIF4 (Fig. 7A). Similarly, BZR1 regulates a substantial number of PIF4-related and MYC2-related co-factor genes. However, regulation of these genes by BZR1 also does not show a preference for positive or negative feedback on either PIF4, HY5, or MYC2 (Fig. 7B). Interestingly, PIF4 upregulates DELLA genes RGL1 and RGL3, while BZR1 upregulates RGA2, but downregulates DELLA genes RGL1 and RGL2 (Fig. 7A). DELLA proteins can sequester PIF4 and JAZ proteins. The feedback by DELLAAs through transcriptional by PIF4 and BZR1 on growth and defense is therefore complex. HY5 is involved in the transcription of a set of negative co-factors for PIF4 and BZR1, but with no preference for positive or negative feedback on PIF4 or BZR1, respectively (Fig. 7C). As described before, MYC2 induces the transcription of HY5, while HY5 downregulates transcription of MYC2 (Chakraborty et al. 2019, Li et al. 2020). Combined this would balance the transcription of both HY5 and MYC2, but this requires activation of MYC2 through JA-ile signaling. Indeed, this is realized through the mutual feedback on JA biosynthesis by both HY5 and MYC2 (Fig. 7C). Previously, HY5 was recognized to be involved in the expression of LOX3 (Wasternack and Hause 2013), but RNA-seq data shows that also LOX2 is

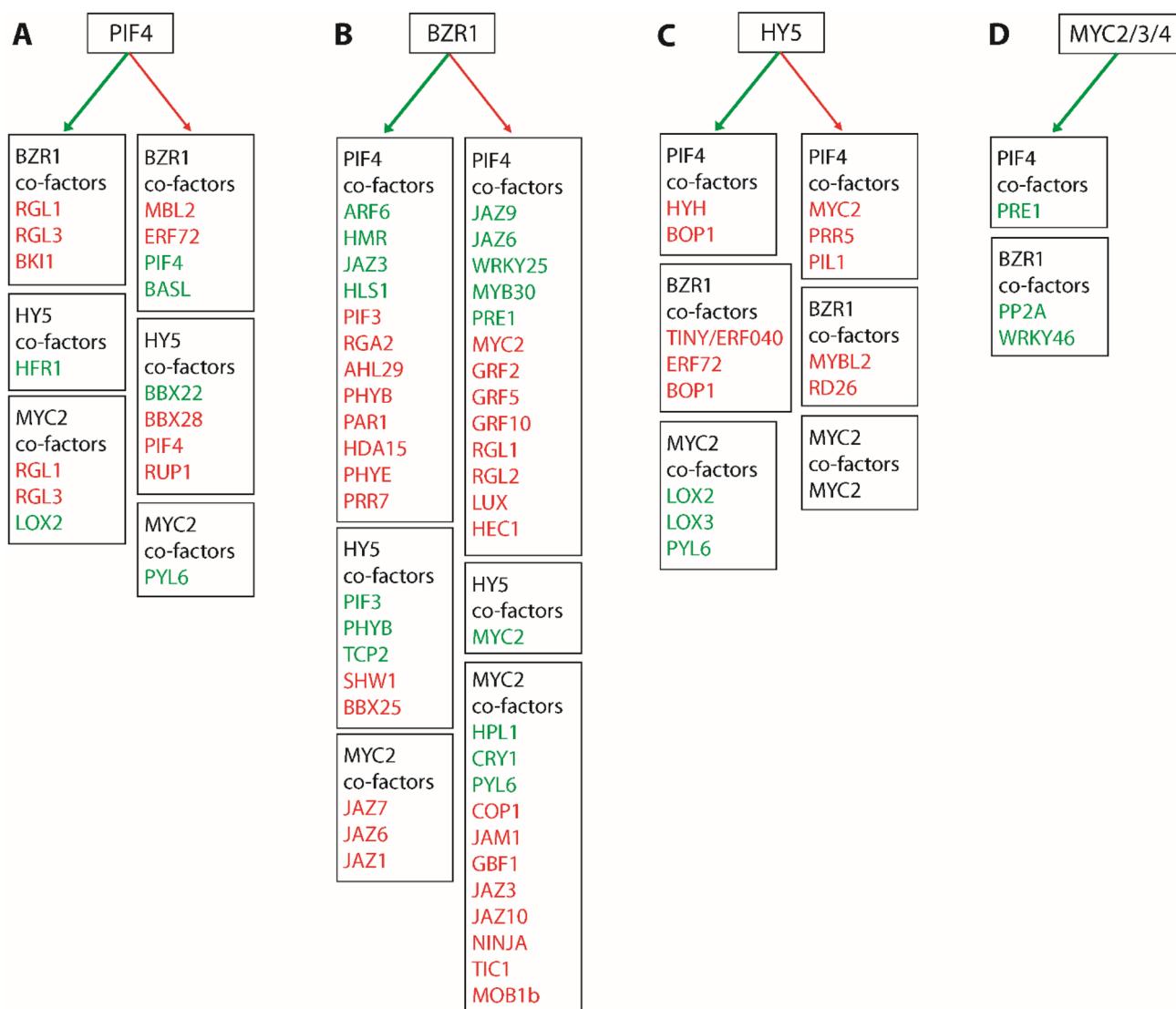


Fig. 7 Cross regulation of PIF4, BZR1, HY5, and MYC2 co-factor genes. **A** Cross-regulation of BZR1, HY5, and MYC2 co-factor genes by PIF4. **B** Cross-regulation of PIF4, HY5, and MYC2 co-factor genes by BZR1. **C** Cross-regulation of PIF4, BZR1, and MYC2 co-factor genes by HY5. **D** Cross regulation of PIF4, BZR1, and HY5

co-factor genes by MYC2. Positive acting co-factors are shown in green, negative acting co-factors are in red. Green arrow: transcriptional upregulation of co-factor gene by key TF. Red arrow: transcriptional downregulation by key TF

regulated by HY5 (Fig. 7C). Moreover, HY5 is a positive and PIF4 is a negative regulator of PYL6, a modulator of MYC2 transcriptional activity on JAZ6 and JAZ8 (Aleman et al. 2016; Li, Shi et al. 2020a, b, c)). Combined these results could indicate reduced sensitivity to ABA-induced PYL6/MYC2 heterodimer activity in the dark (when PIF4 is active) and increased ABA sensitivity to ABA-induced PYL6/MYC2 heterodimer activity in the light (when HY5 is active) (Fig. 7C). The biological meaning of modulating MYC2 activity through differential regulation of PYL6 by PIF4 and HY5 needs further study. MYCs (MYC2) enhance

the expression of one positive acting co-factor gene for PIF4 (PRE1) and two positive acting co-factor genes for BZR1 (PP2A and WRKY46) (Fig. 7D). This reveals new levels of growth and defense interaction previously not recognized: while activation of MYC2 suppresses elongation through HY5, it also increases the potential for PIF4 and BZR1 activity through upregulation of PRE1, PP2A, and WRKY46 (Fig. 7D). It could be that such activity functions as priming the plant for regaining growth when defense signaling is over (Fig. 7D).

Integration of Feedback on Light and Hormone Signaling by PIF4, BZR1, HY5, and MYC2

Of the five co-factor genes shared by all four TFs (HY5, PHYB, CRY1, CRY2, COP1; Fig. 2B), PHYB is co-regulated by PIF4 and BZR1, COP1 is co-regulated by BZR1 and HY5, HY5 is coregulated by PIFs and HY5, while CRY1 is only regulated by BZR1 and CRY2 only by PIF4 (Tables 1, 2, 3, and 4). Combined, this demonstrates how transcriptional regulation of growth (by PIF4, BZR1, HY5) and defense (by MYC2) potentially is linked through feedback on shared co-factor genes. This reveals multiple levels that effectuate the trade-off between growth and defense, many of which have not been recognized before.

PIF4 protein is stabilized by nuclear COP1 activity, while the nuclear complex SCF^{COP1/SPA1} targets HY5 for protein degradation. Nuclear COP1 is destabilized by activated PHY, CRY or UVR8 photoreceptors, respectively (Huq and Quail 2002; Lorrain et al. 2008; Hayes et al. 2017; Tavridou et al. 2020), so in general, growth is stimulated in the dark (stable PIF4) and suppressed in the light (stable HY5). BZR1 protein is by default targeted for degradation after phosphorylation by the kinase BIN2 (He et al. 2002) and BZR1 protein is stabilized when the activity of BIN2 is inhibited under BR signaling (Sellaro et al. 2009). BZR1 protein may also be targeted for degradation in response to specific phytohormones, such as through interaction with the SCF^{MAX2/D14} strigolactone receptor complex (Wang et al. 2013a, b, Li et al. 2017). MYC

proteins are targeted by the CUL3^{BPM} E3 ubiquitin ligases, while the stability of BPM protein is enhanced by Jasmonic Acid signaling. Thus, JA-signaling alleviates repression of MYC2 activity but limits MYC2 activity in a negative feedback loop that regulates MYC protein stability (Jung et al. 2015; Chico et al. 2020). A summary of our inventory shows that PIF4 and BZR1 act antagonistically at multiple levels to balance positive and negative feedback on growth (Fig. 8). BR biosynthesis gene expression is upregulated by PIF4 but downregulated by BZR1. The upregulation of BR synthesis by PIF4 and potential subsequent BR signaling results in positive feedback on growth through BZR1 activated PIF4 expression during thermomorphogenesis (Ibanez et al. 2018) (Fig. 8). Such positive feedback is potentially dangerous, as it may result in uncontrolled upregulation of PIF4 by BZR1 (Ibanez et al. 2018). However, PIF4 transcription is restrained through transcriptional feedback through numerous negative acting co-factor genes. While the contribution of each single co-factor to negative feedback on PIF4 may be small, the combined feedback may help control PIF4 expression and activity, especially during heat stress.

The positive feedback through BR synthesis and signaling on PIF4 is balanced by the negative regulation of BR biosynthesis and BR signaling genes by BZR1. In addition, the expression of PHYB and UVR8 is upregulated by PIF4 (Tables 1–2), which potentially increases negative feedback on growth: Light-activated PHYB directly interacts with PIF4 protein, resulting in increased mutual turnover (Huq

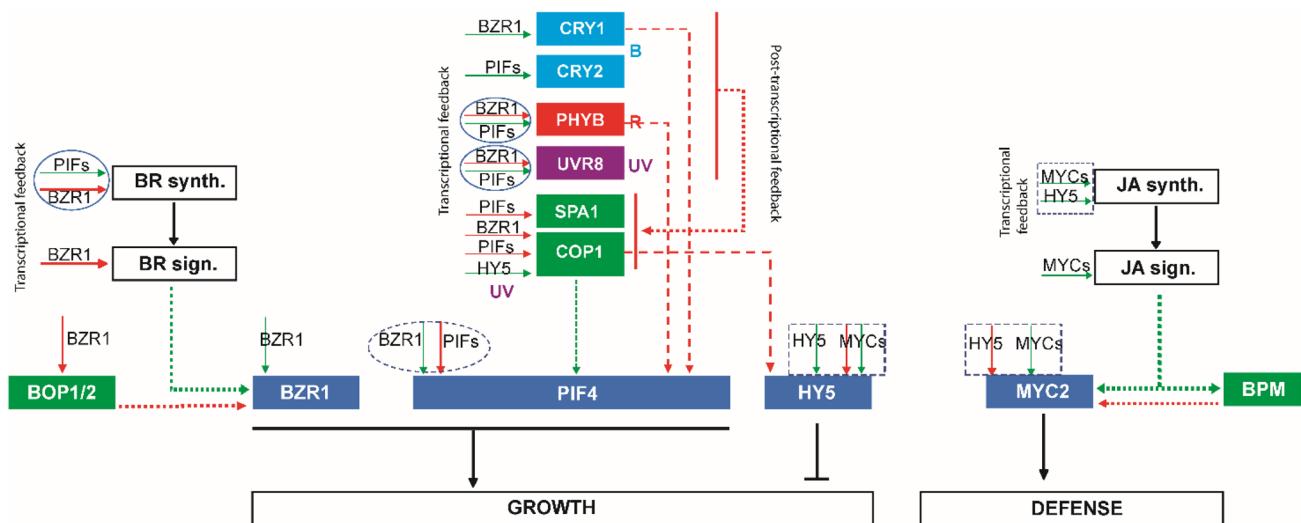


Fig. 8 Integrated feedback in growth by PIF4, BZR1, and HY5 and balancing growth with defense by HY5 and MYC2. Green/red arrows: positive/negative transcriptional feedback. Solid ovals: dual regulation by PIFs and BZR1. Dashed rectangles: dual regulation by HY5 and MYC2. The figure illustrates potential feedback in growth through dual feedback on BR synthesis (PIFs and BZR1), JA (HY5 and MYCs),

photoreceptors PHYB (PIFs and BZR1), UVR8 (PIFs and BZR1) and triple feedback on COP1 (PIFs, BZR1, HY5) and the balance between growth and defense through potential dual feedback on HY5 (HY5 and MYCs), MYC2 (HY5 and MYCs) and JA biosynthesis (HY5 and MYCs)

and Quail 2002; Ni et al. 2014; Ma et al. 2016; Pedmale et al. 2016). PHYB and UVR8, when light activated, also reduce SPA1/COP1 activity. SPA1 phosphorylates PIF4 in vitro and SPA1 activity is required for the stabilization of PIF4, especially during thermomorphogenesis (Lee et al. 2020). Reduced SPA1/COP1 activity therefore results in negative feedback on PIF4, but positive feedback on HY5, both resulting in suppression of growth (Huq and Quail 2002; Lorrain et al. 2008; Hayes et al. 2017; Tavridou et al. 2020). The negative feedback on growth through PHYB and UVR8 by PIF4 is balanced by the activity of BZR1, which upregulates the expression of PHYB and UVR8 (Fig. 8). Regulation of expression of COP1 provides negative feedback growth as PIF4 and BZR1 both downregulate the expression of COP1. Under UV light HY5 can also induce transcription of COP1. Potentially this could result in negative feedback on HY5 activity and positive feedback on PIF4 activity. However, under UV the HY5 protein COP1/SPA interaction is interrupted by UVR8. No contribution of HY5 to COP1 expression is detected under other light conditions, but if a single contribution of HY5 is low, COP1 will not be detected as a differentially expressed gene.

The dual regulation by PIF4 and BZR1 has been described before BR biosynthesis genes and relates to different functions of BZR1 homodimers and PIF4/BZR1 heterodimer, respectively (Oh et al. 2012). Supposedly, BES1 and BZR1 monomers act as repressors when they bind to the promoter of BR biosynthesis genes, especially during the day when PIF4 protein levels are low (Sun et al. 2010; Wang et al. 2012). However, at night, when PIF4 protein levels are high, PIF4 binds to BES1/BZR1 to form a heterodimer which activates transcription of BR biosynthesis genes (Sun et al. 2010; Oh et al. 2012). Further studies are needed to determine whether a similar mechanism operates for diurnal PIF4/BZR1 mediated transcriptional regulation of the photoreceptor genes PHYB and UVR8. The synergistic interaction between BZR1 and PIF4 also operates downstream of PIF4 and BZR1, as thousands of common downstream target genes are synergistically co-regulated by PIF4 and BZR1 (Oh et al. 2012). SPA1 is also under dual regulation by PIF4 and BZR1, but here PIF4 represses, while BZR1 stimulates transcription SPA1.

Most central to balance activity of PIF4 may be the regulation by PIF4 itself. BZR1 upregulates the expression of PIF4. BZR1 and PIF4 were shown to synergistically stimulate the expression of thousands of common target genes, but the set of common target genes does not include PIF4 (Oh et al. 2012). However, because of the extensive redundant regulation of PIF4 transcription and activity, such dual regulation by PIF4/BZR1 may go unnoticed in single mutant studies. When expressed at high levels, ectopic PIF4 downregulates endogenous PIF4 expression, so PIF4 itself

may be the most crucial factor to restrain uncontrolled PIF4 expression (Shapulatov 2019). Further studies are needed to determine the role of specific PIF4/BZR1 homodimers and heterodimers in the regulation of expression of SPA1 and PIF4. Of the blue light receptors, only CRY2 is under positive transcriptional feedback by PIFs, and under this condition, CRY2 may limit PIF4 activity. CRY2 protein is light-labile rapidly down-regulated by blue light as a function of light intensity (Lin et al. 1998). The positive transcriptional regulation of CRY2 by the light labile PIF proteins, and negative feedback of activated CRY proteins on PIF4, may therefore be most relevant under low light intensities. At higher B light intensities, the negative feedback on PIF4 through CRY1 becomes more important, and this is stimulated by BZR1 (Fig. 8).

Growth and defense are mostly balanced through mutual transcriptional regulation of HY5 and MYC2, by HY5 and MYC2 as previously described (Ortigosa et al. 2020). Positive feedback in defense is through upregulation of JA biosynthesis genes and signaling genes by MYC2, but also upregulation of some JA biosynthesis genes by HY5 (Figs. 7 and 8). In addition, defense and growth are linked through transcriptional regulation of PIF4 and BZR1 related positive acting co-factor genes by MYC2, resulting in potential positive feedback on growth by MYCs (Fig. 7). This may partly compensate for the negative effect on growth by MYC2 through HY5.

Recently the temporal transcriptional response to W, B, R, or FR light was determined for *Arabidopsis* seedlings (Kurihara et al. 2020). Because light plays an important role in the regulation of PIF4 and HY5 protein stability, the role of light quality on potential feedback through co-factor genes on PIF4 or HY5, respectively, was determined. Of the 102 co-factors identified for PIF4 (Table S2), 58 genes are under transcriptional feedback by PIF4 (Table 1). PIF4 protein is unstable in light, and therefore, it may be expected that transcription these co-factor genes for PIF4 is affected by light (Table S10). Indeed, 39 co-factor genes show light regulation (Table S10) and 27 of these co-factor genes show both regulations by PIF4 and by light (Table S10, grey cells). As expected, many co-factor genes seem to be under reciprocal control by PIFs and light. For instance, HSL1 is upregulated by PIFs (Table 1) but downregulated by light conditions that result in the destabilization of PIFs (Table S10A). In contrast, negative co-factor genes PRR9, HYH, PIL1, HFR1, PAR1, PAR2, HEC2, AHL27/29, and RGA1/2 are downregulated by PIFs (Table 1), but upregulated by light (Table S10B), indicating that PIFs may act as a suppressor for these genes in the dark and that other TFs beside PIF4 are responsible for the upregulation in the light. Overall, in the light regulation of PIF4-related co-factor genes, there is no consistent positive or negative feedback on PIF4 (positive

feedback indicated by green cells, negative feedback indicated by red cells in Table S7-S8).

Of the 57 co-factor genes identified for HY5 (Table S3), 13 co-factor genes are under some form of light-dependent transcriptional control (Table S11). Twelve of these 13 co-factor genes are also under direct transcriptional control of HY5 (Table S11, grey cells). Transcriptional light regulation of HY5 co-factor genes results mostly in potential negative feedback on HY5, both by suppression of all positive acting, light-regulated, co-factor genes and by upregulation of most negative acting, light-regulated, co-factor genes (Table S11). Because HY5 protein is stabilized in the light, this suggests that HY5 acts as a suppressor for the positive acting co-factor genes and as an activator for the negative acting co-factor genes. Transcription of *HY5* and *HYH* is upregulated by HY5 and light, and this positive feedback on HY5 is balanced by the transcriptional upregulation of negative acting co-factors COP1 and RUP2. In addition, ten HY5 co-factor genes under that are under transcriptional feedback by HY5 (Table 3) are not light-regulated.

Comparing PIF-, BZR1-, HY5-, and MYC-Related ChIP-seq and RNA-seq Data

Although DNA ChIP-seq data indicate a central role for PIF4 by binding to the promoters of *PIF4*, *BZR1*, *HY5*, and *MYC2* (Fig. 2), only the PIF4 gene itself is under redundant transcriptional control by PIFs (Table 1). Of the combined set of 213 co-factor genes that affect the activity of PIF4, BZR1, HY5, and/or MYC2 protein activity, 72 co-factor genes are positive targets in the ChIP-seq experiments. Of these 72 co-factor genes, only 22 genes are under transcriptional control by one or more of these TFs. In contrast, 43 co-factor genes are transcriptionally affected by either PIF4, BZR1, HY5, and/or MYC2, without being the target of these TFs in the DNA ChIP-seq experiments. This may be in part due to detection limitations in DNA ChIP-seq experiments. For instance, the binding of HY5 to the HY5 and MYC2 promoter was only detected using the more dedicated ChIP-PCR experiments (Binkert et al. 2014; Chakraborty et al. 2019). Alternatively, transcription of these co-factor genes may be indirectly regulated by either PIF4, BZR1, HY5, and/or MYC2. Some genes bound by one or more PIF proteins do not show altered expression in the *pif4* or *pif-q* mutant, indicating that the PIF binding does not influence the expression of these genes. Indeed, most PIF4-related co-factor genes that show altered expression in single or multiple PIF mutant backgrounds are not on the list of targets of PIF protein binding (Table 1). This either indicates that these transcriptional feedback

interactions are indirect, or it indicates that DNA ChIP data are incomplete, or that conditions of DNA ChIP data differed from conditions used for transcriptome profiling (Tables 1, 2, 3, and 4).

The PIF4-, BZR1-, HY5-, and MYC2-related ChIP-seq and RNA-seq data sets used in this study were compared using cluster analysis (Fig. 9). The ChIP-seq data sets are all part of cluster B in Fig. 9, but do no cluster with most RNA-seq data sets, indicating that binding of TF to gene only shows low correlation to differential expression of that gene. The cluster analysis indicates three major clusters, each with some sub-cluster pairs. Table S6 shows the GO-enrichment analysis of the gene sets in the three major clusters. Only the gene set targeted by MYC2 clusters closely to the genes upregulated by MYC2 (Fig. 9, cluster B3). Moreover, the gene set targeted by HY5 closely clusters with the gene set targeted by MYC2, indicating the potential for interaction with the same genes between HY5 and MYC2 (Fig. 9, cluster B3). Most remarkably, the set of genes upregulated by MYC2 in 28-day-old plants clusters closely to genes downregulated by MYC2 in 15-day-old seedlings, indicating that the regulation of genes may change over time (Fig. 9, cluster A3). The GO-enrichment analysis of this cluster indicates that this mainly concerns genes involved in glucosinolate biosynthesis (Table S6). The gene sets in cluster A1 confirm the opposite regulation of PIFs/BZR1 and HY5 in growth (Fig. 9, cluster A1). Remarkably, both the PIF downregulated and BZR1 downregulated gene sets cluster closely to the BZR1 target gene set, suggesting that PIFs may play an additional role in the repressor function of BZR1 at target genes (Fig. 9, cluster B1). However, the GO-enrichment analysis does not indicate that this involves genes with a specified biological function as it mainly involves unclassified genes (Table S6). The gene set up-regulated by MYCs closely clusters with the set of genes down-regulated by PIF4 (Fig. 9, cluster C2). This mainly involves genes with function in ethylene signaling, hypoxia, and wounding (Table S6).

Gene sets downregulated by BZR1 in WT background and those down-regulated by *bzr1-1D* in *pifq* mutant background grown on BRZ do not cluster together (Fig. 9). This indicates that in a background where BR biosynthesis is blocked, BZR1 repressed target genes may require the additional action of PIFs (Sun et al. 2010). In contrast, the gene sets upregulated by BZR1 cluster closely with genes upregulated by *bzr1-1D* in the *pifq* mutant background, indicating that the positive action of BZR1 may not be dependent on PIFs (Fig. 9, cluster C1) where BZR1 by *bzr1-1D* require the co-action of PIFs (Sun et al. 2010).

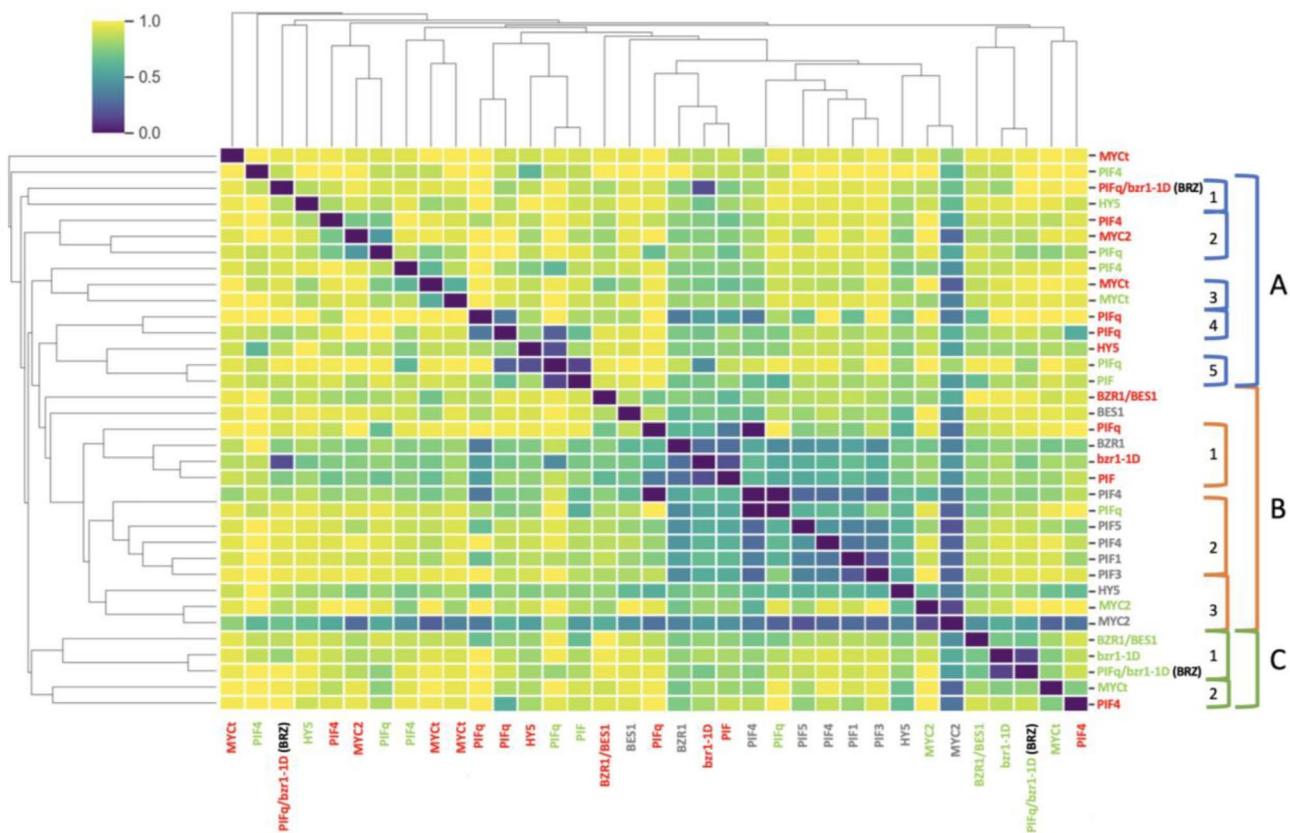


Fig. 9 Cluster analysis of ChIP-seq and RNA-seq data sets used in this study. Three main clusters (A, B, C) and sub-clusters are indicated. Gray: ChIP-seq data, Red: RNA-seq data: upregulated by TF or in genotype indicated, Green: RNA-seq data: upregulated by TF or in genotype indicated. For references see legends of Tables 1, 2, 3, and 4

Concluding Remarks

Each of the four selected TFs is under both feed forward and feed backward regulation by co-factors, and for PIF4 and BZR1 feedback through co-factors is most redundant. For PIF4, BZR1, and MYC2 the feedback through transcriptional regulation of their co-factor genes is mostly negative (downregulation of positive acting factors, upregulation of negative acting factors), while for HY5 the identified feedback is mostly positive (upregulation of positive acting factors and downregulation of negative acting factors) (Fig. 10). A feedforward regulation results in a prolonged activity of the TF(s), while the feed-back regulation may be needed to terminate the activity of the TFs (Lee et al. 2007). The extensive redundancy in negative feedback in the transcriptional regulation of growth strongly buffers against the effect of a single (a) biotic, mutant, or chemical perturbation. The implications of how growth and defense are linked by feedback transcription through co-factor genes should therefore be studied by probing or modeling combinatorial perturbations. The transcriptional regulation of PIF4, BZR1, HY5, and MYC2 is complicated by the fact that the action

of some TFs (e.g., PIF4 and BZR1/BES1) at common target genes depends on homodimer or heterodimer formation. For instance, growth BZR1 and PIF4 share a sub-set of downstream genes and presumably activate transcription of these genes as heterodimers (PIF4/BZR1) but may block transcription as homodimers (He et al. 2005; Oh et al. 2012, Planas-Riverola et al. 2019), while for another subset of common target genes they activate genes as homodimer (BZR1 or BES1), but block transcription as heterodimer with PIF4 (Martinez et al. 2018). The cluster analysis shows that most BZR1 repressed genes seem to require PIF co-action, while most positive targets of BZR1 seem to be independent of PIFs (Fig. 9 cluster C1). Therefore, BZR1-PIF4 heterodimer may mainly function as a transcription repressor for a subset of target genes, while BZR1 homodimer may function as an activator for another subset of target genes. The actual feedback by each of these TFs on target genes may be a function of their relative abundance, the potential for dimer formation, context, and sequence at the promoter binding site.

Major transcriptional feedback is through transcriptional control of photoreceptor co-factor genes and the SPA1/COP1 co-factor genes and integrates the action of

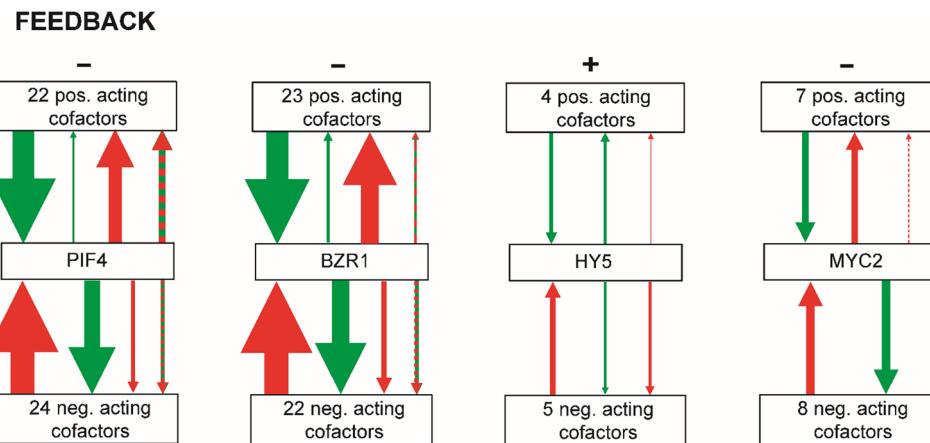


Fig. 10 Summary of transcriptional feedback on co-factor genes for PIF4, BZR1, HY5, and MYC2 (data from Tables 1, 2, 3, and 4). Solid arrows up/down from TF: co-factor genes under positive (green) or negative (red) transcriptional control by PIF4, BZR1, HY5, or MYC2. Striped arrows: positive or negative transcriptional control of co-factor gene depends on condition. The thickness of the arrow

relates to the total number of co-factors under transcriptional feedback by the TF. Arrows from co-factors to TF: positive (green) or negative (red) feedback by co-factors on TF. The + or - on top indicates an overall more positive (+) or negative (-) feedback on PIF4, BZR1, HY5, and MYC2 by their related co-factors, respectively

all four TFs (Fig. 8). The actual potential for coordinate transcription of PIF4, BZR1, HY5, and MYC2 may be larger than shown here because there are 15 other TFs besides PIF4 with shared validated binding to the promoter of these TF genes (Fig. 2) and it still needs to be determined if any of these have an active role in transcription of these four TFs and/or their related co-factor genes. The identified transcriptional feedback regulation by the co-factor genes may not be universal, but limited to certain tissue (Chaiwanon and Wang 2015) or cell type-specific (Lau et al. 2018); it may be limited to certain environmental conditions such as heat (Ibanez et al. 2018) or it may depend on developmental stage as shown for the *myct* mutant (Table S8; (Schweizer et al. 2013; Van Moerkercke et al. 2019)).

Genome-wide comparison of *Arabidopsis* and several crop species indicates that the key components of light signaling (PIFs, HY5) and BR signaling (BZR1 and BES1) are conserved in crops (Wang et al. 2017; Han et al. 2019a, b; Thalmann et al. 2019). Also, defense signaling pathway components are conserved between *Arabidopsis* and crops, but there are also significant differences (Anderson et al. 2005). For instance, PIFs, BZR1 and MYC genes from *Arabidopsis* equally relate to one or more rice genes and HY5 from *Arabidopsis* has multiple paralogues in rice (Wang et al. 2017). The feedback regulation on PIF4 through PRRs and HFR1 may be different in monocots, as rice has multiple paralogues for PRR5, no orthologue for PRR9, multiple paralogues for PRR7, and no orthologue for HFR1 (Wang et al. 2017). The myriad of potential positive and negative feedback, especially on PIF4, indicates the large pool of options that are available in nature to obtain

a balanced expression of PIFs to control growth. It is to be expected that the relative contribution of the potential positive and negative co-factors in the control of the activity of these TFs may differ in different plant species, while overall regulation of growth remains the same. Therefore, the better we understand the potential of interactions between growth and defense components, the better we may be able to interpret difference in transcriptional regulation in crop species.

Supplementary Information The online version contains supplementary material available at <https://doi.org/10.1007/s11105-022-01351-9>.

Author Contribution Omics ChIP-seq and RNA-seq retrieval and data analysis were done by SK and AvD. Data interpretation and figure preparation were done by US and the main text by SvdK and US.

Data Availability Public available data (ChIP-seq and RNA-seq) was used for the meta-analysis (see supplemental files of references in Tables 1–4). Additional tables in the Supplement are available online.

Declarations

Consent to Participate All authors consent to participate in the publication.

Consent for Publication All authors and WUR consent for the publication.

Conflict of Interest The authors declare no competing interests.

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