

RESEARCH ARTICLE

Potential tradeoffs between effects of arbuscular mycorrhizal fungi inoculation, soil organic matter content and fertilizer application in raspberry production

Ke Chen ^{*}, Jeroen Scheper, Thijs P. M. Fijen, David Kleijn

Plant Ecology and Nature Conservation Group, Wageningen University, Wageningen, The Netherlands

^{*} chenkeji@gmail.com

Abstract

Ecological intensification has been proposed as an alternative paradigm for intensive agriculture to boost yield sustainably through utilizing ecosystem services. A prerequisite to achieving this is to understand the relations between multiple ecosystem services and production, while taking growth conditions such as nutrient availability into consideration. Here, we conducted a pot-field experiment to study the interactive effects of soil organic matter (SOM) content and arbuscular mycorrhizal fungi (AMF) inoculation on the production of raspberry (*Rubus idaeus* L.) under four levels of fertilizer application. Raspberry flower number, fruit number and yield only significantly increased with fertilizer inputs but were not impacted by SOM content or AMF inoculation. Fruit set and single berry weight were influenced by both SOM content and AMF inoculation, in complex three-way interactions with fertilizer application. Fruit set of AMF inoculated plants increased with fertilizer inputs in low SOM soils, but decreased with fertilizer inputs under high SOM soils, with the highest fruit set occurring at no fertilizer inputs. In low SOM soils, the relation between single berry weight and fertilizer application was more pronounced in inoculated plants than in non-inoculated plants, while in high SOM soils the relative benefits of AMF inoculation on single berry weight decreased with increasing fertilizer inputs. We attribute the lack of effects of AMF inoculation and SOM content on flower number, fruit number and yield mainly to potential tradeoffs between the experimental variables that all influence resource uptake by plant root systems. Our results suggest that potentially beneficial effects of AMF and SOM can be offset by each other, probably driven by the dynamic relations between AMF and the host plants. The findings reveal fundamental implications for managing AMF inoculation and SOM management simultaneously in real-world agricultural systems.

OPEN ACCESS

Citation: Chen K, Scheper J, Fijen TPM, Kleijn D (2022) Potential tradeoffs between effects of arbuscular mycorrhizal fungi inoculation, soil organic matter content and fertilizer application in raspberry production. PLoS ONE 17(7): e0269751. <https://doi.org/10.1371/journal.pone.0269751>

Editor: Sergio Saia, University of Pisa: Università degli Studi di Pisa, ITALY

Received: January 28, 2022

Accepted: May 27, 2022

Published: July 18, 2022

Peer Review History: PLOS recognizes the benefits of transparency in the peer review process; therefore, we enable the publication of all of the content of peer review and author responses alongside final, published articles. The editorial history of this article is available here: <https://doi.org/10.1371/journal.pone.0269751>

Copyright: © 2022 Chen et al. This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement: All relevant data are within the paper and its [Supporting Information](#) files.

Introduction

Conventional agricultural intensification cannot meet the twofold challenge facing agriculture: increasing yield to feed the growing world population while minimizing negative externalities

Funding: Ke Chen was financially supported through the China Scholarship Council (File No. 201706990023). The funder had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing interests: The authors have declared that no competing interests exist.

on the environment [1,2]. It is increasingly difficult to further promote productivity through mainstream intensive farming practices [3,4], because the production is increasingly limited by critical natural ecosystem services, such as insect pollination [5,6] and soil formation [7]. Additionally, these intensive farming practices have caused severe environmental problems, such as soil and water pollution [8,9] and biodiversity loss [10,11], which are threatening human-wellbeing [3]. Ecological intensification has been proposed as a promising alternative for conventional intensive agriculture. It is based on managing multiple ecosystem services to complement and/or replace artificial inputs to maintain or enhance productivity while reducing negative environmental impacts [12,13]. Ecological intensification has been advocated as an environmentally friendly way towards food security [14,15] and an increasing number of studies provide proof of concept for this paradigm [16–18]. There are still knowledge gaps between theory and practice, however, which limit the adoption of ecological intensification by the agricultural sector [13]. For example, when multiple ecosystem services are managed in conjunction, their effects on production could interact synergistically, negatively or not at all [19,20]. Understanding whether and how different ecosystem services interact in shaping crop production is of importance to maximize the benefits of ecological intensification and promote its adoption [13].

Soil organic matter (SOM) and arbuscular mycorrhizal fungi (AMF) are two natural factors that provide or influence vital ecosystem services in cropping systems [12,17,21]. SOM is often used as a proxy for soil services, as it is able to mediate the flow of soil ecosystem services [12,22], and it strongly affects almost all soil properties [23]. Examples include soil structural stability and water-holding capacity (physical properties), cation exchange capacity and pH regulation (chemical properties), and nutrient supply for microbial communities (biological properties) [23]. SOM content, therefore, often relates positively to crop production [17,24].

AMF are widespread soil microorganisms from the phylum *Glomeromycota*, and they can form symbiotic associations with the majority of the cultivated crops [25,26]. AMF develop an extensive hyphal network through proliferating their hyphae inside plant roots (intracellular hyphae) as well as within the soil (extraradical hyphae), thus acting as a bridge between plant and soil [27–29]. AMF mainly help their host plants exploit poorly mobile ions (notably inorganic phosphate) that are beyond the root zone, in exchange for photosynthetic products from the host for metabolic needs [29]. Besides assisting with resource uptake, AMF colonization can also benefit the hosts by enhancing their tolerance to abiotic and biotic stresses, such as drought, salinity, diseases and pathogens [30,31]. Indirectly, AMF can benefit the hosts via improving soil structure and soil aggregation [32]. Inoculation of AMF has been found to promote crop yield [21,33,34], especially where the indigenous AMF communities have been degraded by agricultural practices [35,36].

A wealth of studies have shown that AMF and SOM can influence each other [37,38]. AMF are able to positively influence SOM content directly, through producing glomalin-related soil proteins [39,40], which are significant components of SOM [41,42]. Additionally, AMF has been found to affect the decomposition of SOM negatively [38] or positively [43]. On the other hand, various organic compounds released from the decomposition of SOM have been shown to influence AMF growth and activity, either positively [44] or negatively [45]. However, as far as we know, so far no studies ever clearly tested whether and how their effects on crop production interact. Furthermore, agricultural practices, in particular artificial fertilizer application, can influence the effects of both AMF [46] and SOM [24] as it also influences nutrient availability of crop plants. It is therefore essential to take fertilizer inputs into consideration when test the interacting effects of AMF and SOM on crop production. Here, we examined (1) the combined effects of AMF inoculation and SOM content on the production of raspberry (*Rubus idaeus* L.) and (2) how they are affected by fertilizer application.

Materials and methods

(a) Study system

Raspberry was used as the study crop, which is an important perennial fruit crop, with growing consumer interest due to its health benefits and flavours [47,48]. We selected the commercial cultivar 'Tulameen', as it is among the most popular raspberry cultivars in a range of climatic conditions [49] and is locally available. The study was conducted in an experimental field of Wageningen University & Research in the Netherlands, from August 2019 to September 2020.

(b) Experimental setup

We adopted a randomized complete block design to account for potential confounding gradients in the experimental field, and we combined all of the following three crossed factors: (i) low SOM content vs high SOM content, (ii) AMF inoculated vs non-inoculated and (iii) four levels of fertilizer application (i.e. 16 plants per block). The SOM treatments were obtained by mixing different proportions of two types of sandy soils which had different SOM content (0.3% vs 4.6%) resulting in either 1.95% SOM content soils ('low SOM' treatment; available N: 14.0 mg/kg, available P: 0.6 mg/kg, available K: 19.4 mg/kg, pH: 6.6) or 3.96% SOM content soils ('high SOM' treatment; available N: 43.1 mg/kg, available P: 0.6 mg/kg, available K: 26.6 mg/kg, pH: 5.9). As for AMF treatments, we used *Rhizophagus intraradices* inoculum (MYKOS® Xtreme Gardening, Canada: 300 propagules/gram). Half of the original inoculum was autoclaved at 121°C for two hours as sterilized inoculum for non-inoculated treatments [50]. The four levels of fertilizer treatments represented the equivalent 0, 33, 66 and 99 kg ha⁻¹ of N per year, ranging from no to optimum N inputs [51]. The fertilizer used was a compound fertilizer (Fertilizers® Cropsolutions, The Netherlands), containing 10.80% N, 13.44% K and 5.89% P.

We purchased 160 raspberry cuttings from a local supplier, with an average height of ca. 60 cm. To avoid the influence from the original peaty substrate, we carefully washed away the soil adhering to the roots in early August 2019. We added the recommended dose of AMF inoculum (25 grams) or an equal volume of sterilized inoculum evenly to the washed roots of the plants. The plants were then transplanted to a 10-litre plastic pot (upper diameter 28 cm, holes in the bottom for drainage but covered with cloth to minimize root growth out of the pot) and filled with low or high SOM soils according to the experimental design. However, higher than expected mortality occurred, possibly due to the cuttings being damaged during the roots washing process combined with the late summer heat. Only 56 plants survived out of the 160 plants, and 48 of them were of good health and thus were selected for further experimentation in three blocks. To carry out the experiment with sufficient replication, we additionally purchased another 160 raspberry cuttings in early October 2019. Because 60 cm cuttings were no longer available, we used plants with an average height of ca. 25 cm. Strictly following the earlier described protocol and using the same materials, the new batch of cuttings were washed, inoculated and transplanted into the low or high SOM soils. In this round, 110 out of the 160 new cuttings survived. These 110 plants were arranged into seven blocks. In total, the experiment therefore started out with 158 potted raspberry plants in 10 blocks. Plants with different treatments were placed randomly within each block. Plants were spaced with one meter within and between rows. Pots were dug into the soil to protect the roots from extremely high or low temperatures. The fertilizer treatments were applied by splitting the annual dose (0, 33, 66 and 99 kg ha⁻¹ of N) into three applications: the first one in the autumn, the second one at bud break in early spring of the following year and the last one at early flowering. All plants received equal and ample irrigation (depending on the weather conditions), and weeds in the

pots were regularly removed. Prior to berry ripening, all plants were bagged with mesh bags to avoid predation by animals. We harvested and weighed the ripe berries when they had just turned bright red. We summed up the berry weight from the same plant to get the total yield and fruit number. Additionally, we carefully counted the wilted or aborted flowers that failed to develop into fruits, which in combination with the fruit number allowed us to estimate the flower number. Because we could only have taken root samples at the end of the experiment, and earlier analyses showed AMF colonization rate of raspberry plants from different treatments did not differ after almost a year's growth [52], we did not measure AMF root colonization rates.

(c) Data analysis

Until harvest, 41 plants from the first batch survived and developed fruits; all 110 plants from the second batch survived, but only 25 of them developed fruits. Since we mainly focus on the effects of treatments on production, only the plants that produced fruits were involved in the data analysis (sample size $n = 66$, S1 Table). We ran separate linear mixed-effects models using the function `lme()` of the `nlme` package in R [53] to study the interacting effects of SOM, AMF and fertilizer on flower number, fruit number, single berry weight (g/fruit) and total yield (g/plant), and included "block" as a random factor. We included the origins of the plants as a covariable in all models, to account for differences between plants from the first and the second batch. Because the fruit set followed a binomial distribution, we used the function `glmmTMB()` to run the same models assuming a binomial distribution [54]. Single berry weight was averaged per plant to avoid pseudoreplication, and response variables were transformed if necessary to meet the normality and homoscedasticity assumptions of the models.

Full models were simplified by removing non-significant predictors (backward elimination) using likelihood ratio tests with removal thresholds of $p > 0.05$, until the resulting minimum adequate model consisted only of variables that contributed significantly to the outcome [55,56].

Results

The number of flowers per plant was only influenced by fertilizer inputs (Table 1). Plants receiving 99 kg N·ha⁻¹ produced 32% more flowers than plants without any fertilizer inputs (Fig 1A). Similarly, fruit number and total yield per plant were only affected by fertilizer inputs (Table 1). The fruit number of the plants grown with the highest fertilizer inputs was 69% higher than that of plants receiving no fertilizer (Fig 1B). Increasing fertilizer inputs from 0 to

Table 1. Effects of arbuscular mycorrhizal fungi (AMF; inoculated vs non-inoculated), soil organic matter (high vs low SOM content) and fertilizer application rates (0, 33, 66, 99 kg N·ha⁻¹·year⁻¹) on raspberry fruit production variables ($n = 66$). Bold values represent significant effects ($P < 0.05$).

	Flower number (sqrt transformed)		Fruit set		Fruit number (ln transformed)		Single berry weight		Yield (ln transformed)	
	$\chi^2_{(1)}$	P	$\chi^2_{(1)}$	P	$\chi^2_{(1)}$	P	$\chi^2_{(1)}$	P	$\chi^2_{(1)}$	P
AMF	0.185	0.667	1.614	0.204	0.232	0.630	2.070	0.150	0.005	0.943
SOM	0.601	0.438	22.136	0.000	0.936	0.333	1.304	0.254	0.137	0.711
Fertilizer	5.107	0.024	23.883	0.000	6.433	0.011	10.593	0.001	14.914	0.000
Origin	29.620	0.000	17.136	0.000	27.739	0.000	13.807	0.000	28.936	0.000
AMF:fertilizer	0.014	0.907	13.303	0.000	0.671	0.413	0.670	0.413	0.241	0.624
AMF:SOM	0.033	0.855	1.054	0.305	0.292	0.589	3.356	0.067	0.246	0.620
SOM:fertilizer	1.132	0.287	0.715	0.398	0.008	0.928	0.073	0.787	0.019	0.891
AMF:SOM:fertilizer	0.768	0.381	16.053	0.000	0.047	0.829	4.722	0.030	1.438	0.230

<https://doi.org/10.1371/journal.pone.0269751.t001>

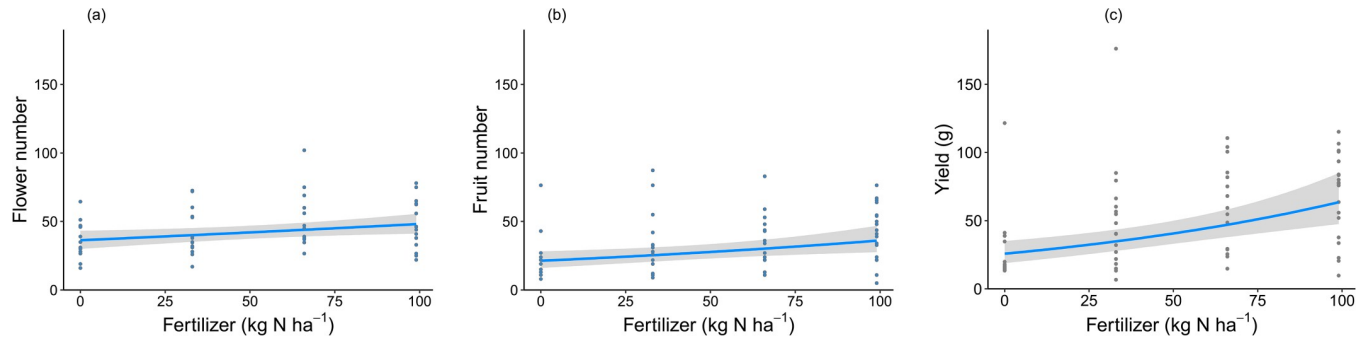


Fig 1. Effects of fertilizer application rates on flower number (a), fruit number (b) and yield (c) per plant. Graphs show conditional partial regression plots based on the minimum adequate models. Shadings show the 95% confidence interval, and points represent partial residuals.

<https://doi.org/10.1371/journal.pone.0269751.g001>

99 kg N·ha⁻¹ increased yield from 25.7 g to 63.5 g (Fig 1C). SOM content or AMF inoculation did not affect these yield parameters, nor did they influence the effect of fertilizer (no significant interactions; Table 1).

A three-way interaction was found between the effects of AMF inoculation, SOM content and fertilizer inputs on fruit set (Table 1). In low SOM soils, the fruit set increased with increasing fertilizer inputs, for both AMF inoculated and non-inoculated plants (Fig 2A). In high SOM soils, the fruit set of non-inoculated plants showed a positive relationship with fertilizer inputs, while the fruit set of inoculated plants was highest in unfertilized soils and decreased with increasing fertilizer inputs (Fig 2B).

There was also a three-way interaction between the three experimental variables on the single berry weight per plant (Table 1). In low SOM soils, the relationship with fertilizer application rate was much more pronounced for AMF inoculated plants than for non-inoculated plants (Fig 3A). In high SOM soils, single berry weight was consistently higher in AMF inoculated plants than in non-inoculated plants, although the difference seemed to decrease with increasing fertilizer inputs (Fig 3B).

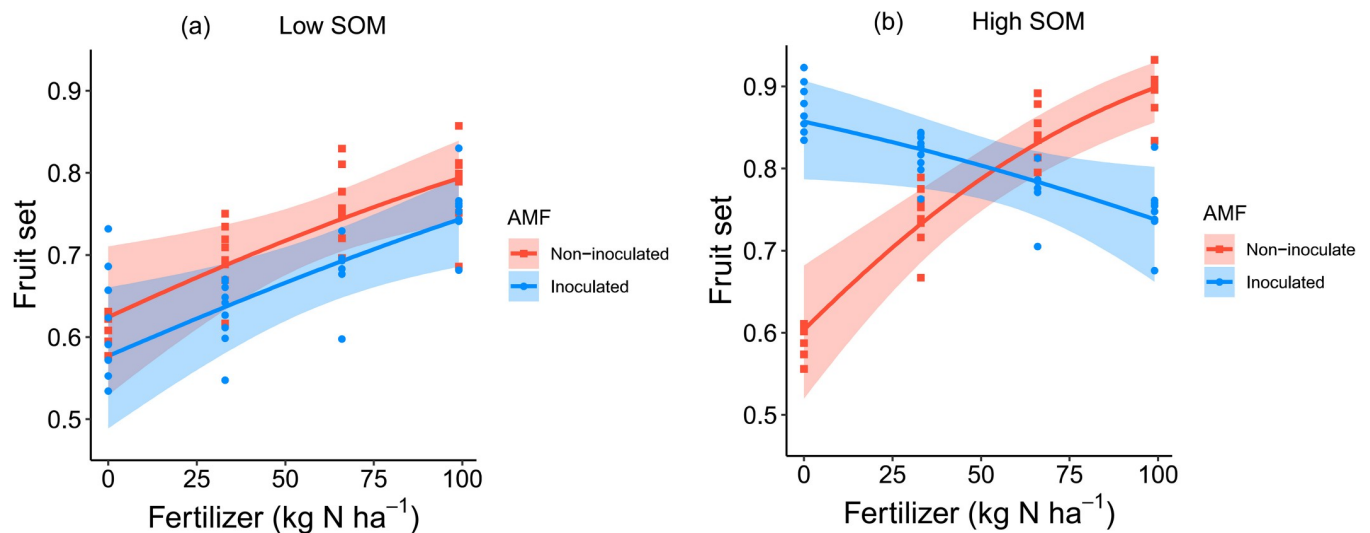


Fig 2. Interactive effects of AMF inoculation, SOM and fertilizer application rates on fruit set per plant. Graphs show conditional partial regression plots based on the minimum adequate model; shadings show the 95% confidence interval, and points represent partial residuals.

<https://doi.org/10.1371/journal.pone.0269751.g002>

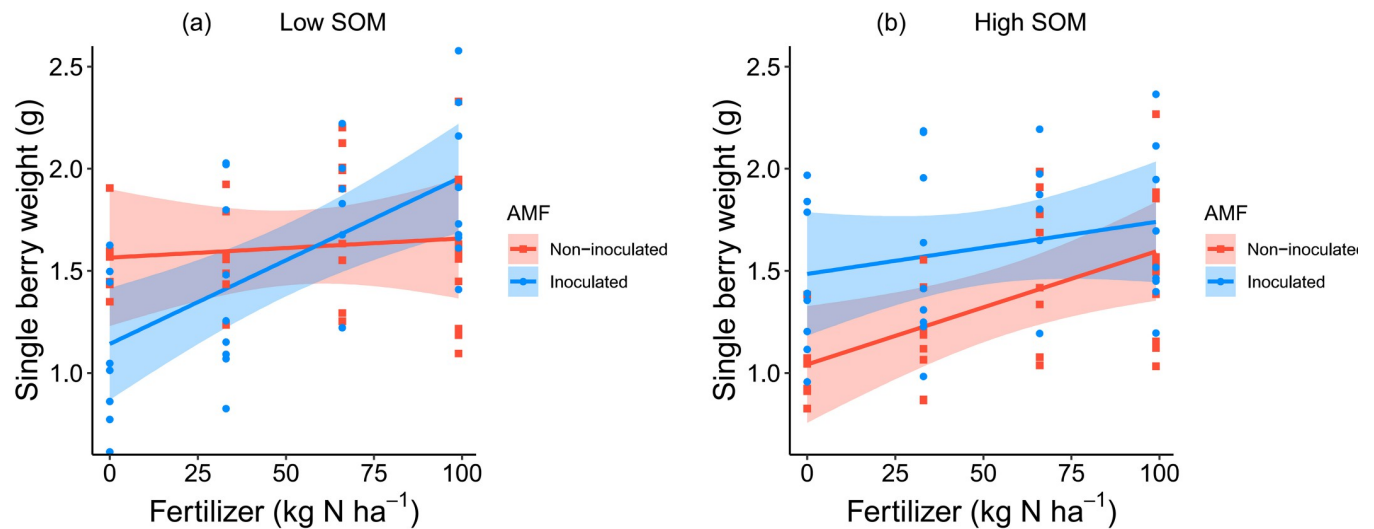


Fig 3. Interactive effects of AMF inoculation, SOM and fertilizer application rates on average single berry weight (g) per plant. Graphs show conditional partial regression plots based on the minimum adequate model; shadings show the 95% confidence interval, and points represent partial residuals.

<https://doi.org/10.1371/journal.pone.0269751.g003>

Discussion

In this study, we found that the numbers of flowers and fruit, as well as the most important parameter from the perspective of farmers, yield per plant, were only driven by fertilizer inputs and were not significantly impacted by AMF inoculation or SOM content. The positive relation between fertilizer application and fruit set in AMF inoculated plants in low SOM soils, changed into a negative relation in high SOM soils. Similarly, at low SOM the relation between fertilizer application and single berry weight was more pronounced in inoculated plants than in non-inoculated plants, but in high SOM soils it was the other way around. This suggests that the effects of AMF and SOM on these yield parameters cancel each other out and as a result did not contribute to the final yield.

At first glance, the lack of effects of AMF inoculation and SOM content on yield, flower number and fruit number, may seem at odds with results of earlier studies done using this same study system. For example, Chen, Kleijn (52) found significant positive effects of AMF inoculation on raspberry flower number, fruit number and yield. Furthermore, in wild raspberry populations, we found that yield was positively related to the SOM content (Chen K. et al., unpublished results, January 2022). However, the first study was only done at low SOM content levels (1.95%; the same as the current low SOM content treatment), while the second study was exclusively done in high SOM content soils (mean 7.4%, range 3.2–13.1%), and neither of these studies simultaneously manipulated both SOM content and AMF inoculation. Potential tradeoffs between the effects of the two factors on raspberry yield would therefore not become apparent in these studies. This is further supported by the fact that Fig 3A is almost an exact copy of Fig 3 in [52]. Both these graphs show the effects of fertilizer and AMF on single berry weight under the same low SOM content levels. Furthermore, in our previous experiments we showed that part of the effects of AMF and SOM could be explained by their positive influence on flower visitation rate by pollinators [52,57]. Because pollinators were not considered in this study, this may have left unexplained any potential indirect effects of AMF inoculation and SOM content on flower visitation rate and consequently the final yield.

In low SOM soils, AMF-inoculated plants produced smaller raspberries than the non-inoculated plants under low fertilizer inputs, while the beneficial effects of AMF inoculation on

berry weight only became apparent at adequate fertilizer inputs. One possible explanation for this is that under nutrient deficiency AMF have to compete for the limiting nitrogen for their hyphae development against the host plants, reducing the resources that host plants can allocate to fruit development [58,59]. In high SOM soils and under low fertilizer inputs, AMF inoculation increased both single berry weight and fruit set compared to those of the non-inoculated treatments (Figs 2B and 3B), likely because AMF could help acquire nutrients from soil organic matter to compensate for the effects of artificial inputs [60]. However, the benefits of AMF inoculation tended to decrease (Fig 3B) or even change into parasitic effects (Fig 2B) with increasing fertilizer application rate, a pattern found in previous studies as well [61–63]. The demonstrated tradeoff between effects of AMF inoculation and SOM content at different fertilizer application rates on berry weight and fruit set might explain why we didn't observe any effect of these factors on flower number, fruit number and yield. The negative interaction could be explained by the cost-benefit relation between AMF and the host plants [61–63]. Host plants share up to 20% of total photosynthetic carbon with AMF, as the cost to maintain the symbiotic associations [64], while receiving mineral nutrients and other resources absorbed by AMF as the benefit [29]. The cost-benefit relations vary from positive to negative, depending on the environmental context and the identity of AMF and the host plants [65]. Under high fertilizer inputs and high SOM soils, the host plants might obtain adequate nutrients via their own root systems [66,67], which decreases the dependence on the assistance of AMF over nutrients acquisition. However, if the associated cost does not decrease, or less strongly, this may result in a net negative benefit which may explain the decreasing benefits of AMF for single berry weight and fruit set with increasing fertilizer levels in the present study. In addition, the decreasing benefits of AMF inoculation might also be explained by the direct suppressing effects of the host plants on AMF growth. When plants obtain sufficient nutrients and water via their own root system in high nutrients soils (high SOM and fertilizer in this study), they may suppress AMF development [68]. Consequently, the suppressed AMF contributed less to production and this might indirectly constrain the benefits delivery of SOM since AMF can enhance the decomposition of SOM [69,70].

Although our study is based on only one study in one crop species, it is the first one to explore the interactive effects of AMF and SOM under a range of fertilizer application rates. Our results provide an indication that the benefits of AMF and SOM on crop yield offset each other. This finding contributes to the understanding of the dynamic effects of AMF inoculation on crop production. For example, Yamawaki, Matsumura [71] found significant positive effects of AMF inoculation on turmeric (*Curcuma longa* L.) production under greenhouse conditions but no effects were found under field conditions, and they attributed the differing outcomes to the influence of indigenous AMF. However, the lack of beneficial effects of AMF inoculation under field conditions could also be caused by tradeoffs due to the interactive effects between AMF, SOM and fertilizer, according to our findings. Therefore, our findings may have important implications for applying AMF as biofertilizers in practical cropping systems, which has been increasingly proposed as a key solution for accomplishing sustainable agriculture [72,73]. For example, when SOM content is high, inoculating AMF might not be such a good idea as when SOM content is low, unless with reduced fertilizer inputs. This study starts the exploration of the combined effects of AMF and SOM on raspberry production under several fertilizer inputs, and further research over a wider range of contexts (e.g. crop, soil type, climate, irrigation and fungicides) is needed to identify their interactive effects under real-world conditions.

Supporting information

S1 Table. The number of replicated raspberry plants per treatment combination.

(PDF)

S1 Data.

(XLSX)

S1 File. R code for the data analysis.

(R)

Acknowledgments

We thank Emiel van Riet for his assistance with the fieldwork.

Author Contributions

Conceptualization: Jeroen Scheper, Thijs P. M. Fijen, David Kleijn.

Formal analysis: Ke Chen, Jeroen Scheper, Thijs P. M. Fijen, David Kleijn.

Funding acquisition: David Kleijn.

Investigation: Ke Chen, David Kleijn.

Methodology: Ke Chen, Jeroen Scheper, Thijs P. M. Fijen, David Kleijn.

Project administration: Ke Chen, David Kleijn.

Supervision: Jeroen Scheper, Thijs P. M. Fijen, David Kleijn.

Validation: Jeroen Scheper, Thijs P. M. Fijen, David Kleijn.

Visualization: Ke Chen, Jeroen Scheper, Thijs P. M. Fijen, David Kleijn.

Writing – original draft: Ke Chen.

Writing – review & editing: Ke Chen, Jeroen Scheper, Thijs P. M. Fijen, David Kleijn.

References

1. Tilman D, Cassman KG, Matson PA, Naylor R, Polasky S. Agricultural sustainability and intensive production practices. *Nature*. 2002; 418(6898):671–7. <https://doi.org/10.1038/nature01014> PMID: 12167873
2. Godfray HCJ, Beddington JR, Crute IR, Haddad L, Lawrence D, Muir JF, et al. Food security: the challenge of feeding 9 billion people. *science*. 2010; 327(5967):812–8. <https://doi.org/10.1126/science.1185383> PMID: 20110467
3. Cassman KG, Grassini P, van Wart J. Crop yield potential, yield trends, and global food security in a changing climate. *Handbook of climate change and agroecosystems* Imperial College Press, London. 2010:37–51.
4. Grassini P, Eskridge KM, Cassman KG. Distinguishing between yield advances and yield plateaus in historical crop production trends. *Nature communications*. 2013; 4(1):1–11. <https://doi.org/10.1038/ncomms3918> PMID: 24346131
5. Lebuhn G, Droege S, Connor EF, Gemmill-Herren B, Potts SG, Minckley RL, et al. Detecting insect pollinator declines on regional and global scales. *Conservation Biology*. 2013; 27(1):113–20. <https://doi.org/10.1111/j.1523-1739.2012.01962.x> PMID: 23240651
6. Deguines N, Jono C, Baude M, Henry M, Julliard R, Fontaine C. Large-scale trade-off between agricultural intensification and crop pollination services. *Frontiers in Ecology and the Environment*. 2014; 12(4):212–7.
7. Pimentel D, Burgess M. Soil erosion threatens food production. *Agriculture*. 2013; 3(3):443–63.

8. Carpenter SR, Caraco NF, Correll DL, Howarth RW, Sharpley AN, Smith VH. Nonpoint pollution of surface waters with phosphorus and nitrogen. *Ecological applications*. 1998; 8(3):559–68.
9. Rodríguez-Eugenio N, McLaughlin M, Pennock D. Soil pollution: a hidden reality: FAO; 2018.
10. Potts SG, Biesmeijer JC, Kremen C, Neumann P, Schweiger O, Kunin WE. Global pollinator declines: trends, impacts and drivers. *Trends in ecology & evolution*. 2010; 25(6):345–53.
11. Ponge J-F, Pérès G, Guernion M, Ruiz-Camacho N, Cortet J, Pernin C, et al. The impact of agricultural practices on soil biota: a regional study. *Soil Biology and Biochemistry*. 2013; 67:271–84.
12. Bommarco R, Kleijn D, Potts SG. Ecological intensification: harnessing ecosystem services for food security. *Trends Ecol Evol*. 2013; 28(4):230–8. Epub 2012/11/17. <https://doi.org/10.1016/j.tree.2012.10.012> PMID: 23153724.
13. Kleijn D, Bommarco R, Fijen TP, Garibaldi LA, Potts SG, van der Putten WH. Ecological intensification: bridging the gap between science and practice. *Trends in ecology & evolution*. 2019; 34(2):154–66. <https://doi.org/10.1016/j.tree.2018.11.002> PMID: 30509848
14. Pywell RF, Heard MS, Woodcock BA, Hinsley S, Ridding L, Nowakowski M, et al. Wildlife-friendly farming increases crop yield: evidence for ecological intensification. *Proceedings of the Royal Society B: Biological Sciences*. 2015; 282(1816):20151740. <https://doi.org/10.1098/rspb.2015.1740> PMID: 26423846
15. IPBES. The assessment report of the Intergovernmental Science-Policy Platform on Biodiversity and Ecosystem Services on pollinators, pollination and food production. Bonn, Germany: Secretariat of the Intergovernmental Science-Policy Platform on Biodiversity and Ecosystem Services, 2016 9280735675.
16. Tamburini G, De Simone S, Sigura M, Boscutti F, Marini L. Conservation tillage mitigates the negative effect of landscape simplification on biological control. *Journal of Applied Ecology*. 2016; 53(1):233–41.
17. Garratt MP, Bommarco R, Kleijn D, Martin E, Mortimer SR, Redlich S, et al. Enhancing soil organic matter as a route to the ecological intensification of European arable systems. *Ecosystems*. 2018; 21(7):1404–15.
18. Tittone P. Ecological intensification of agriculture—sustainable by nature. *Current Opinion in Environmental Sustainability*. 2014; 8:53–61.
19. Garibaldi LA, Andersson GK, Requier F, Fijen TP, Hipólito J, Kleijn D, et al. Complementarity and synergisms among ecosystem services supporting crop yield. *Global food security*. 2018; 17:38–47.
20. Tamburini G, Bommarco R, Kleijn D, van der Putten WH, Marini L. Pollination contribution to crop yield is often context-dependent: A review of experimental evidence. *Agriculture, ecosystems & environment*. 2019; 280:16–23.
21. Zhang S, Lehmann A, Zheng W, You Z, Rillig MC. Arbuscular mycorrhizal fungi increase grain yields: A meta-analysis. *New Phytologist*. 2019; 222(1):543–55. <https://doi.org/10.1111/nph.15570> PMID: 30372522
22. Williams A, Hedlund K. Indicators and trade-offs of ecosystem services in agricultural soils along a landscape heterogeneity gradient. *Applied soil ecology*. 2014; 77:1–8.
23. Krull ES, Skjemstad JO, Baldock JA. Functions of soil organic matter and the effect on soil properties: Cooperative Research Centre for Greenhouse Accounting Canberra; 2004.
24. Oldfield EE, Wood SA, Bradford MA. Direct evidence using a controlled greenhouse study for threshold effects of soil organic matter on crop growth. *Ecological Applications*. 2020:e02073. <https://doi.org/10.1002/eap.2073> PMID: 31965653
25. Tawarayama K. Arbuscular mycorrhizal dependency of different plant species and cultivars. *Soil Science and Plant Nutrition*. 2003; 49(5):655–68.
26. Rillig MC, Aguilar-Trigueros CA, Camenzind T, Cavagnaro TR, Degrune F, Hohmann P, et al. Why farmers should manage the arbuscular mycorrhizal symbiosis. *New Phytologist*. 2019; 222(3):1171–5. <https://doi.org/10.1111/nph.15602> PMID: 30657593
27. Jeffries P, Gianinazzi S, Perotto S, Turnau K, Barea J-M. The contribution of arbuscular mycorrhizal fungi in sustainable maintenance of plant health and soil fertility. *Biology and fertility of soils*. 2003; 37(1):1–16.
28. Rajtor M, Piotrowska-Seget Z. Prospects for arbuscular mycorrhizal fungi (AMF) to assist in phytoremediation of soil hydrocarbon contaminants. *Chemosphere*. 2016; 162:105–16. <https://doi.org/10.1016/j.chemosphere.2016.07.071> PMID: 27487095
29. Smith SE, Read DJ. *Mycorrhizal symbiosis*: Academic press; 2010.
30. Begum N, Qin C, Ahanger MA, Raza S, Khan MI, Ashraf M, et al. Role of arbuscular mycorrhizal fungi in plant growth regulation: implications in abiotic stress tolerance. *Frontiers in plant science*. 2019; 10:1068. <https://doi.org/10.3389/fpls.2019.01068> PMID: 31608075

31. Diagne N, Ngom M, Djighaly PI, Fall D, Hocher V, Svistoonoff S. Roles of arbuscular mycorrhizal fungi on plant growth and performance: Importance in biotic and abiotic stressed regulation. *Diversity*. 2020; 12(10):370.
32. Rillig MC, Mummey DL. Mycorrhizas and soil structure. *New Phytologist*. 2006; 171(1):41–53. <https://doi.org/10.1111/j.1469-8137.2006.01750.x> PMID: 16771981
33. Baum C, El-Tohamy W, Gruda N. Increasing the productivity and product quality of vegetable crops using arbuscular mycorrhizal fungi: a review. *Scientia horticulturae*. 2015; 187:131–41.
34. Srivastava A, Malhotra S, Krishna Kumar N. Exploiting nutrient-microbe synergy in unlocking productivity potential of perennial fruits: A review. *Indian J Agric Sci*. 2015; 85(4):459–81.
35. Jin H, Germida JJ, Walley FL. Suppressive effects of seed-applied fungicides on arbuscular mycorrhizal fungi (AMF) differ with fungicide mode of action and AMF species. *Applied Soil Ecology*. 2013; 72:22–30.
36. Manoharan L, Rosenstock NP, Williams A, Hedlund K. Agricultural management practices influence AMF diversity and community composition with cascading effects on plant productivity. *Applied Soil Ecology*. 2017; 115:53–9.
37. Joner E, Jakobsen I. Growth and extracellular phosphatase activity of arbuscular mycorrhizal hyphae as influenced by soil organic matter. *Soil Biology and Biochemistry*. 1995; 27(9):1153–9.
38. Zhou J, Zang H, Loeppmann S, Gube M, Kuzyakov Y, Pausch J. Arbuscular mycorrhiza enhances rhizodeposition and reduces the rhizosphere priming effect on the decomposition of soil organic matter. *Soil Biology and Biochemistry*. 2020; 140:107641.
39. Wright SF, Upadhyaya A. Extraction of an abundant and unusual protein from soil and comparison with hyphal protein of arbuscular mycorrhizal fungi. *Soil science*. 1996; 161(9):575–86.
40. Rillig MC. Arbuscular mycorrhizae, glomalin, and soil aggregation. *Canadian Journal of Soil Science*. 2004; 84(4):355–63.
41. Jones DL, Hodge A, Kuzyakov Y. Plant and mycorrhizal regulation of rhizodeposition. *New phytologist*. 2004; 163(3):459–80. <https://doi.org/10.1111/j.1469-8137.2004.01130.x> PMID: 33873745
42. Zhang J, Tang X, Zhong S, Yin G, Gao Y, He X. Recalcitrant carbon components in glomalin-related soil protein facilitate soil organic carbon preservation in tropical forests. *Scientific reports*. 2017; 7(1):1–9.
43. Paterson E, Sim A, Davidson J, Daniell TJ. Arbuscular mycorrhizal hyphae promote priming of native soil organic matter mineralisation. *Plant and Soil*. 2016; 408(1):243–54.
44. Gryndler M, Hršelová H, Cajthaml T, Havránková M, Řezáčová V, Gryndlerová H, et al. Influence of soil organic matter decomposition on arbuscular mycorrhizal fungi in terms of asymbiotic hyphal growth and root colonization. *Mycorrhiza*. 2009; 19(4):255–66. <https://doi.org/10.1007/s00572-008-0217-y> PMID: 19104847
45. Ravnkov S, Larsen J, OLSSON PA, Jakobsen I. Effects of various organic compounds on growth and phosphorus uptake of an arbuscular mycorrhizal fungus. *New Phytologist*. 1999; 141(3):517–24.
46. Bakhshandeh S, Corneo PE, Mariotte P, Kertesz MA, Dijkstra FA. Effect of crop rotation on mycorrhizal colonization and wheat yield under different fertilizer treatments. *Agriculture, Ecosystems & Environment*. 2017; 247:130–6. <https://doi.org/10.1016/j.agee.2017.06.027>
47. Burton-Freeman BM, Sandhu AK, Edirisinghe I. Red raspberries and their bioactive polyphenols: cardiometabolic and neuronal health links. *Advances in Nutrition*. 2016; 7(1):44–65. <https://doi.org/10.3945/an.115.009639> PMID: 26773014
48. Giuffrè AM, Louadj L, Rizzo P, De Salvo E, Sicari V. The Influence of Film and Storage on the Phenolic and Antioxidant Properties of Red Raspberries (*Rubus idaeus* L.) cv. Erika. *Antioxidants*. 2019; 8(8):254. <https://doi.org/10.3390/antiox8080254> PMID: 31366095
49. Aprea E, Biasioli F, Carlin S, Endrizzi I, Gasperi F. Investigation of Volatile Compounds in Two Raspberry Cultivars by Two Headspace Techniques: Solid-Phase Microextraction/Gas Chromatography – Mass Spectrometry (SPME/GC–MS) and Proton-Transfer Reaction– Mass Spectrometry (PTR–MS). *Journal of agricultural and food chemistry*. 2009; 57(10):4011–8. <https://doi.org/10.1021/jf803998c> PMID: 19348421
50. Changey F, Megloulou H, Fontaine J, Magnin-Robert M, Tisserant B, Lerch TZ, et al. Initial microbial status modulates mycorrhizal inoculation effect on rhizosphere microbial communities. *Mycorrhiza*. 2019; 29(5):475–87. <https://doi.org/10.1007/s00572-019-00914-1> PMID: 31506745
51. Strik BC. A review of nitrogen nutrition of *Rubus*. *Acta Horticulturae*; 2005; 403–410.
52. Chen K, Kleijn D, Scheper J, Fijen TP. Additive and synergistic effects of arbuscular mycorrhizal fungi, insect pollination and nutrient availability in a perennial fruit crop. *Agriculture, Ecosystems & Environment*. 2022; 325:107742.

53. R Core Team. R: A Language and Environment for Statistical Computing. R Foundation for Statistical Computing; 2020.
54. Brooks ME, Kristensen K, Van Benthem KJ, Magnusson A, Berg CW, Nielsen A, et al. glmmTMB balances speed and flexibility among packages for zero-inflated generalized linear mixed modeling. *The R journal*. 2017; 9(2):378–400.
55. Zuur A, Ieno EN, Walker N, Saveliev AA, Smith GM. *Mixed effects models and extensions in ecology with R*: Springer Science & Business Media; 2009.
56. Heinze G, Dunkler D. Five myths about variable selection. *Transplant International*. 2017; 30(1):6–10. <https://doi.org/10.1111/tri.12895> PMID: 27896874
57. Chen K, Fijen TP, Kleijn D, Scheper J. Insect pollination and soil organic matter improve raspberry production independently of the effects of fertilizers. *Agriculture, Ecosystems & Environment*. 2021; 309:107270.
58. Wang X-X, Wang X, Sun Y, Cheng Y, Liu S, Chen X, et al. Arbuscular mycorrhizal fungi negatively affect nitrogen acquisition and grain yield of maize in a N deficient soil. *Frontiers in microbiology*. 2018; 9:418. <https://doi.org/10.3389/fmicb.2018.00418> PMID: 29568292
59. Reynolds HL, Hartley AE, Vogelsang KM, Bever JD, Schultz P. Arbuscular mycorrhizal fungi do not enhance nitrogen acquisition and growth of old-field perennials under low nitrogen supply in glasshouse culture. *New Phytologist*. 2005; 167(3):869–80. <https://doi.org/10.1111/j.1469-8137.2005.01455.x> PMID: 16101923
60. Hodge A, Fitter AH. Substantial nitrogen acquisition by arbuscular mycorrhizal fungi from organic material has implications for N cycling. *Proc Natl Acad Sci U S A*. 2010; 107(31):13754–9. Epub 2010/07/16. <https://doi.org/10.1073/pnas.1005874107> PMID: 20631302; PubMed Central PMCID: PMC2922220.
61. Smith FA, Grace EJ, Smith SE. More than a carbon economy: nutrient trade and ecological sustainability in facultative arbuscular mycorrhizal symbioses. *New Phytologist*. 2009; 182(2):347–58. <https://doi.org/10.1111/j.1469-8137.2008.02753.x> PMID: 19207688
62. Jin L, Wang Q, Wang Q, Wang X, Gange AC. Mycorrhizal-induced growth depression in plants. *Symbiosis*. 2017; 72(2):81–8.
63. Hoeksema JD, Chaudhary VB, Gehring CA, Johnson NC, Karst J, Koide RT, et al. A meta-analysis of context-dependency in plant response to inoculation with mycorrhizal fungi. *Ecology letters*. 2010; 13(3):394–407. <https://doi.org/10.1111/j.1461-0248.2009.01430.x> PMID: 20100237
64. Willis A, Rodrigues B, Harris PJ. The ecology of arbuscular mycorrhizal fungi. *Critical Reviews in Plant Sciences*. 2013; 32(1):1–20.
65. Cavagnaro R, Oyarzabal M, Oesterheld M, Grimoldi A. Species-specific trade-offs between regrowth and mycorrhizas in the face of defoliation and phosphorus addition. *Fungal Ecology*. 2021; 51:101058.
66. El Kinany S, Achbani E, Faggroud M, Ouahmane L, El Hilali R, Haggoud A, et al. Effect of organic fertilizer and commercial arbuscular mycorrhizal fungi on the growth of micropropagated date palm cv. Fegouss. *Journal of the Saudi Society of Agricultural Sciences*. 2019; 18(4):411–7.
67. Wu S, Cao Z, Li Z, Cheung K, Wong MH. Effects of biofertilizer containing N-fixer, P and K solubilizers and AM fungi on maize growth: a greenhouse trial. *Geoderma*. 2005; 125(1–2):155–66.
68. Grman E. Plant species differ in their ability to reduce allocation to non-beneficial arbuscular mycorrhizal fungi. *Ecology*. 2012; 93(4):711–8. <https://doi.org/10.1890/11-1358.1> PMID: 22690621
69. Hodge A, Campbell CD, Fitter AH. An arbuscular mycorrhizal fungus accelerates decomposition and acquires nitrogen directly from organic material. *Nature*. 2001; 413(6853):297–9. Epub 2001/09/21. <https://doi.org/10.1038/35095041> PMID: 11565029.
70. Gui H, Hyde K, Xu J, Mortimer P. Arbuscular mycorrhiza enhance the rate of litter decomposition while inhibiting soil microbial community development. *Sci Rep*. 2017; 7:42184. Epub 2017/02/09. <https://doi.org/10.1038/srep42184> PMID: 28176855; PubMed Central PMCID: PMC5296878.
71. Yamawaki K, Matsumura A, Hattori R, Tarui A, Hossain MA, Ohashi Y, et al. Effect of inoculation with arbuscular mycorrhizal fungi on growth, nutrient uptake and curcumin production of turmeric (*Curcuma longa* L.). *Agricultural Sciences*. 2013; 4: 66–71.
72. Basu S, Rabara RC, Negi S. AMF: the future prospect for sustainable agriculture. *Physiological and Molecular Plant Pathology*. 2018; 102:36–45.
73. Igiehon NO, Babalola OO. Biofertilizers and sustainable agriculture: exploring arbuscular mycorrhizal fungi. *Applied microbiology and biotechnology*. 2017; 101(12):4871–81. <https://doi.org/10.1007/s00253-017-8344-z> PMID: 28547568