



IMPART: an update of the project

K. T. Veldman¹ • J.S. Slettemeås² • S.A. Granier³ • A. Perrin-Guyomard³ • T. Hechard³ • C. Dierikx⁴ • S. Maurischat⁵ • and the IMPART consortium

1. Wageningen Bioveterinary Research (WBVR), Lelystad, the Netherlands • 2. Norwegian Veterinary Institute (NVI), Oslo, Norway • 3. French Agency for Food, Environmental and Occupational Health & Safety (ANSES), Fougères, France • 4. Centre for Infectious Disease Control (RIVM), Bilthoven, the Netherlands • 5. German Federal Institute for Risk Assessment (BfR), Berlin, Germany

IMPART (IMproving Phenotypic Antimicrobial Resistance Testing) includes four topics related to the harmonization of phenotypic methods for detection of antimicrobial resistance: selective isolation and detection of **colistin-resistant *Enterobacteriaceae* (WP1)**, selective isolation and detection of **carbapenemase-producing *Enterobacteriaceae* (WP2)**, setting **ECOFFs** for specific pathogen/antibiotic combinations (**WP3**) and development of a standardized **disk diffusion method** for susceptibility testing of ***Clostridium difficile* (WP4)**.

METHODS

To determine the optimal selective culturing method for isolation of colistin-resistant (WP1) and carbapenemase-producing *Enterobacteriaceae* (WP2), two pre-ring trials were organised in 2018. A small number of participants were involved (Anses, NVI, RIVM and WBVR) to test several conditions and most selective media available on the European market as of September 2018 (Fig 1 and Fig 2).

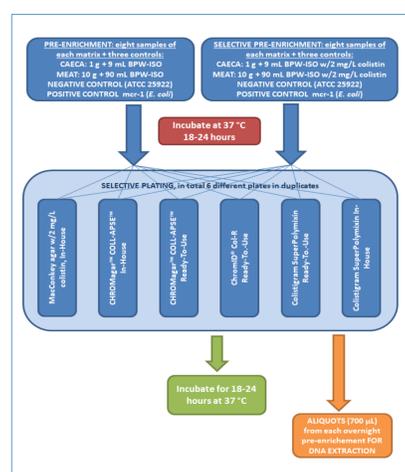


Fig 1 FLOW CHART WP1: colistin-resistant *Enterobacteriaceae*

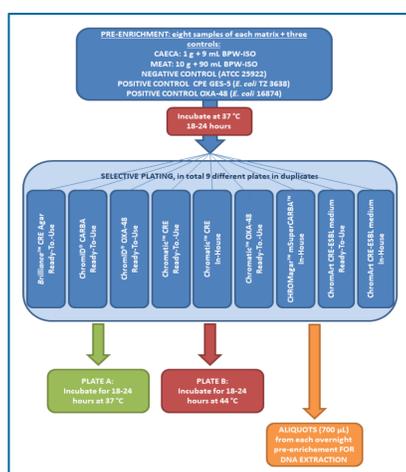


Fig 2 FLOW CHART WP2: carbapenemase-producing *Enterobacteriaceae*

Within WP3, susceptibility testing of veterinary bacteria will be performed with broth microdilution using panels as described in Fig 3. MIC data will be shared and epidemiological cut-off values (ECOFFs) will be assigned according to EUCAST methods.

In WP4, *C. difficile* strains were first collected and characterized. A well-characterised, multi-variant *C. difficile* strain collection was then used to test an optimised disk diffusion protocol, analysing different media and conditions.

Fig 3 Antimicrobial panels for WP3: NLDGNS for testing Gram-negative bacteria, NLDGPS for testing Gram-positive bacteria and NLD1MAC for testing both Gram-positive and Gram-negative bacteria.

RESULTS

The robustness of the sample preparation protocol was confirmed for ring trials in WP1 and WP2: artificially contaminated samples were stable and homogenous. The three participants were able to recover the expected bacteria, but the different selective media performed unevenly. Nonetheless, the WP1 protocol still requires amendments to eliminate false positive colonies.

For WP3, MIC testing just started to a delay in the delivery of the microtiter plates (Sensititre). We expect to produce > 15.000 MIC-values and establish a significant number of new ECOFFs.

Within WP4, the collection and characterisation of *C. difficile* isolates was completed and an optimized disk diffusion test protocol was developed.



CONCLUSION

The pre-ring trials of WP1 and WP2 delivered essential information to finalize the protocols for the final ring trial to be held in 2019 for all participants.

In WP3, new ECOFFs will be established of antimicrobials for veterinary use.

In WP4, a ring trial will elucidate the interlaboratory reproducibility of the optimized method.