

Crop Cycle Length Determines Optimal Transplanting Date for Seedlings from Hybrid True Potato Seeds



Luuk C. M. van Dijk^{1,2} · Olivia C. Kacheyo² · Michiel E. de Vries² · Willemien J. M. Lommen¹ · Paul C. Struik¹

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Abstract

The technology of hybrid breeding in diploid potatoes creates opportunities to design novel and improved cultivation systems based on hybrid true potato seeds. A promising cultivation pathway to produce seed or ware tubers is by transplanting greenhouse-raised seedlings into the field. This study explored the effects of transplanting date and seedling age on tuber yield, using greenhouse-raised seedlings. Field trials with experimental hybrid genotypes were conducted in three consecutive years. In 2017 and 2018, 4- and 6-week-old seedlings were transplanted at four dates: March, April, May and June. In 2019, transplanting dates included April, May and June and seedling age was 5 weeks. In 2018, the March planting experienced severe frost during the initial field period resulting in crop failure. In 2017 and 2019, plants could withstand shorter and less severe frost events. Seedling age did not significantly affect tuber parameters. Transplanting in June resulted in lower marketable yield (>28 mm) compared with earlier transplanting dates when crops were harvested in September. At full crop senescence, no differences in marketable yield were observed. The optimal transplanting window, taking into account weather-related risks, is approximately between early April and end May. For some genotypes, crop cycle length was observed to be a more important yield-determining factor than transplanting date.

Keywords Greenhouse nursery \cdot Hybrid cultivation systems \cdot Hybrid TPS \cdot Potato cropping cycle \cdot Seedling age \cdot *Solanum tuberosum* L. \cdot Transplanting date \cdot True Potato Seed

Luuk C. M. van Dijk luuk.vandijk@wur.nl

¹ Centre for Crop Systems Analysis, Wageningen University and Research, Bornsesteeg 48, 6708 PE Wageningen, The Netherlands

² Solynta, Dreijenlaan 2, 6703 HA Wageningen, The Netherlands

Introduction

The novel technology of diploid hybrid breeding reported by Lindhout et al. (2011) has led to the introduction of hybrid true potato seeds (TPS) for potato production. To derive seed and ware tubers from hybrid TPS, various cultivation systems can be used. These include direct field sowing, field transplanting of greenhouse-raised seedlings and planting of seedling tubers derived from either transplanted or directly sown plants. All these systems can be used for further seed tuber multiplication or the production of ware tubers (Almekinders et al. 1996, 2009; van Dijk et al. 2021).

Greenhouse-raised seedlings for field production in potato have been widely used since the introduction of TPS. Transplanting is favoured over direct sowing as it shortens the duration of the field growing period compared with direct sowing in the field and advances growth at the start of the crop cycle (Wiersema 1984; Almekinders et al. 2009). Until now, potato crops derived from TPS-grown plant materials were not able to penetrate the existing potato value chains that are based on seed-tuber grown crops; hybrid TPS is expected to accelerate the transition from a conventional seed tuber-based system to a TPS-based system (Lindhout et al. 2011, 2018; Jansky et al 2016; Stokstad 2019). Presently, the first yield data of diploid hybrid TPS grown from greenhouse-raised seedlings have been reported as ranging between 25 and 30 Mg/ha (van Dijk et al. 2021). Crops grown from seedling tubers derived from greenhouse-raised seedlings of diploid hybrid TPS yield between 16 and 52 Mg/ha (Stockem et al. 2020).

Transplanting of seedlings and other transplants, e.g. cuttings, is associated with the risk of poor seedling establishment in the field mostly influenced by transplant shock (cf. Snoek et al. 1985; Reekie et al. 2005, 2007). Potato seedlings are highly sensitive to transplant shocks and the field conditions under which transplanting is carried out (Almekinders et al. 1996; Muthoni et al. 2014; van Dijk et al. 2021). Moreover, transplants show a slower initial growth in the field than plants grown from seed tubers leading to a reduced interception of radiation (cf. Lommen 1999). A longer growth duration in the field might give transplants the opportunity to catch up with seed-tuber grown plants. Environmental factors such as temperature, daylength and global radiation are known to affect the length of the growth cycle by affecting the speed and duration of vegetative growth, tuber initiation and tuber bulking (Ewing and Struik 1992; Kooman et al. 1996a, b). One would expect a strong interaction between these environmental factors and genotype, resulting from specific responses of cultivars to accumulated environmental factors (combined environmental factors received in time) during the crop cycle, especially when planting seedlings on different dates.

Early transplanting and the use of larger-sized seedlings (cf. Everaarts et al. 1993; Kerbiriou et al. 2013) may provide possible solutions to advancing the crop growth cycle. Seedlings of larger size, due to advanced vegetative growth during the greenhouse nursery phase, lead to faster crop establishment in the field. Early transplanting, however, presents a risk of transplanting coinciding

with unfavourable conditions such as frost (cf. Snoek et al 1985; Timmer et al. 1989; Neuvel et al. 1991), for which potato is sensitive (Hijmans et al. 2003; Bethke et al. 2019). Large, over-developed transplants (BBCH stage 109 and further, Kacheyo et al. 2021) are also associated with high risk of mechanical damage during transplanting, and with increased transpiration rates (Reekie et al. 2005, 2007) which can cause wilting of the seedling (Snoek et al. 1985). On the other hand, transplanting of small, under-developed seedlings (BBCH stage < 102, Kacheyo et al. 2021) in open field conditions leads to delayed establishment and growth of seedlings, which negatively impacts the length of the crop growth cycle (Everaarts et al. 1993; Kerbiriou et al. 2013).

The typical Dutch planting season for arable crops, from mid-March to mid-May, is known for its fluctuating daily minimum and maximum temperatures; both frost during the day and temperatures above 25 °C are possible. This necessitates the need to investigate how well a potato crop grown from transplants will perform when transplanted before mid-May.

Understanding the accumulated effects of environmental factors during a cropping cycle is necessary for defining an optimal period for field transplanting. Similarly, exploring the effects of seedling age on crop development and ultimately tuber yield is essential for defining the ideal transplantable seedling. For hybrid TPS-derived seedlings, the accumulated environmental effects during the cropping cycle, the timing of transplanting during the field season, the environmental conditions at the time of transplanting and the age of the hybrid seedlings at the time of transplanting have not yet been studied, but this knowledge is needed for successful adoption of hybrid TPS by farmers. It is assumed that an early transplanting date might be beneficial to crop yield, but comes with the risks of frost. A late transplanting date might reduce crop yield due to a shorter field period compared with earlier dates.

The aim of this study was to establish — under Dutch agronomic conditions — the effects of (1) the *timing* of field transplanting and (2) the seedling age at the date of transplanting on tuber yield in different size classes and to understand (3) how accumulated environmental factors impact the effects of the transplanting date. Several field experiments were carried out across three seasons with multiple experimental hybrid genotypes. In the final experimental year, an additional *harvest* was added at *complete crop senescence* to observe the yield differences between a fully matured crop compared with a harvest at a *conventional date in September*.

This study was part of the Potarei project; this project explored scenarios for novel and improved cultivation pathways for potato based on diploid hybrid true potato seeds, using experimental hybrid genotypes to gain first insights. The project aimed to lay foundation for further research in potato cultivation pathways for hybrid potato. Therefore, this study will also elaborate on additional studies on specific aspects which are important to operationalise a practical cultivation system for transplanted hybrid potato seedlings.

Materials and Methods

Hybrid Potato Genotypes

In the course of 3 years, seven experimental hybrids originating from the hybrid potato breeding company Solynta were used for experiments. Lindhout et al. (2018) gave a detailed description of the genetic background of the breeding program from which the used experimental hybrids originated. Due to the experimental nature of the genetic material, large commercial size seed quantities were not available. Therefore, it was decided to use the most advanced and most homogeneous material available in each year. In 2017, *genotypes H01* and *H02* were used, in 2018 *H03* and *H04* and in 2019 *H03*, *H07*, *H08* and *H09*. Known characteristics related to the ontology of the experimental *genotypes* used are described in Supplementary Table S1. The numeric code of the *genotypes* signifies advanced' hybrids compared with the other hybrids in subsequent years.

Production and Transplanting of Seedlings

Seedlings were raised in a greenhouse compartment following the protocol described by van Dijk et al. (2021). Thereafter, they were transferred to a screenhouse for a 1-week hardening-off phase to reduce the transplant shock (cf. Snoek et al. 1985; Poll et al. 1994). On the dates of field planting in 2017 and 2018, seedlings of different *age* treatments had been raised for 4 or 6 weeks after sowing. In 2019, all seedlings past a 5-week-long nursery period.

One day prior to transplanting, uniform, transplantable seedlings were selected. Selection criteria were as follows: *4-week*-old seedlings with a stem of 2–3 cm and 2–4 true leaves, *5-week*-old seedlings with a stem of 7–12 cm and 5–8 leaves and *6-week*-old seedlings with a stem > 14 cm and > 8 true leaves (BBCH stages 102–104, 105–108 and 109, respectively; Kacheyo et al. 2021).

Field Locations and Experimental Design

Field experiments were conducted at Nergena (51°59′40″N, 5°39′24″E), the Netherlands, on a sandy soil, in three seasons (2017, 2018, 2019).

In both 2017 and 2018, a field experiment was conducted to assess the effects of the *transplanting date* and the *age* of the seedlings at transplanting. The experiment was a split-plot experiment containing four replicated blocks. Four *transplanting dates (March, April, May, June)* were randomly assigned to the main plots (Table 1). Two seedling *ages (4 weeks* and *6 weeks* at transplanting in both years) and two *genotypes (H01, H02* in 2017 and *H03, H04* in 2018) were assigned to the sub-plots (Table 1). Gross plots $(3.00 \times 1.80 \text{ m})$ consisted of four rows, spaced at 0.75 m, and nine seedlings per row, spaced at 0.20 m, creating a planting density of 6.67 plants/m². Net plots included the middle two rows with five seedlings each $(1.50 \times 1.00 \text{ m})$.

	1					
Year ^a	Coding of planting	Genotype	Seedling age	Dates of:		
	dates			Planting	Harvest	
					September	Senescence
2017	March, April, May, June	H01, H02	4 weeks, 6 weeks	21 March 13 April 11 May 7 June	21 September	
2018	March, April, May, June	H03, H04	4 weeks, 6 weeks	14 March ^b 10 April 9 May 6 June	20 September	
2019	April, May, June	H03, H07, H08, H09	5 weeks	9 April 7 May 4 June	4 September 10 September 10 September	10 October 10 October 9 November

Table 1 Experimental details

^aIn 2017 and 2018, experiments were laid out in 4 blocks, in 2019 in 2 blocks

^bThe March planting in 2018 got killed by frost during the first 2 weeks in the field

Seedlings were hand-transplanted in small ridges with the seedlings' peat moss plug covered with about 1 cm of soil (see van Dijk et al. 2021 for more details); see Table 1 and Figs. 1a–d, 2a–b. Harvest of all *transplanting dates* was conducted on 21 September in 2017 and on 20 September in 2018 (Table 1 and Figs. 1a–d, 2a–b), in line with the *conventional harvest date* in Dutch potato production. At the date of harvest, the crop was in principal growth stages 907, 905, 905 and 901 according to the BBCH scale for *transplanting dates March* to *June*, respectively (Kacheyo et al. 2021). In both years, all tubers (> 20 mm) obtained from the net plots were used for the necessary measurements.

In 2019, a field experiment was conducted to assess the effects of *transplanting* date and harvest on tuber yield. The experiment had a split-plot with two replicated blocks. Three transplanting dates (April, May and June) were assigned to the main plots and four genotypes (H03, H07, H08 and H09) plus the two harvest dates, September harvest and — full — senescence harvest (Table 1), were assigned to the sub-plots. Plots $(3.00 \times 1.96 \text{ m})$ consisted of four rows, spaced at 75 cm, and seven seedlings per row, spaced at 28 cm, creating a plant density of 4.76 plants/m². The September harvest took place when crop stages were comparable with those of the harvest dates in 2017 and 2018 (Table 1 and Figs. 1e-f, 2c). Hence, the tubers of the April and May transplanting dates were harvested on 4 and 10 September respectively when the haulm was > 50%, but not yet completely senesced (BBCH stage 905-906, Kacheyo et al. 2021). The June transplanting date was harvested on 10 September, when the complete haulm had sagged and senescing had just started (BBCH stage 901, Kacheyo et al. 2021). For all transplanting dates in 2019, the senescence harvest took place when the whole haulm was fully senesced (BBCH stage 907, Kacheyo et al. 2021; Table 1 and Figs. 1e-f, 2c), which occurred on 10 October for transplanting dates April and May and on 9 November for transplanting date June. During all harvest dates in 2019, tubers of whole plots (5.88 m²) consisting of 28 plants were lifted. Moreover, at the September harvest, all tubers > 10 mm

were included in the measurements whereas at full *senescence*, all tubers > 20 mm were taken into account.

Cultural Practices

Immediately after transplanting, seedlings were irrigated and received at least 0.5 L water per m row (at least 0.1 L/seedling), as described in van Dijk et al. (2021). This promoted fast recovery from transplanting as water-limited conditions were avoided. Furthermore, irrigation to supplement precipitation when necessary was applied by a boom irrigation system, see Fig. 2. When daily minimum temperatures were forecasted to decrease to <1–2 °C, plants were covered with a transparent perforated plastic film, as used in conventional Dutch vegetable crop production (Snoek et al. 1985; Timmer et al. 1989; Neuvel et al. 1991; de Kraker et al. 1993); see Fig. 1b, d, f. The film was removed when temperatures were forecasted to rise > 15 °C for a period of > 3 h.

Between transplanting and 100% ground cover, ridges were earthened-up in two operations to the size of a conventional potato ridge. Weeding was done by hand to avoid undesirable and unpredictable effects of herbicides on the experimental potato hybrids. Fertiliser and fungicide applications were carried out following Dutch potato cultivation recommendations.

Observations, Measurements and Calculations

Daily weather data were recorded at weather station the Veenkampen, located at 2.9 km West from the trial fields. Data included minimum and maximum temperatures (T-min & T-max, Fig. 1b, d, f) and precipitation (Fig. 2). Daily incident photosynthetically active radiation (PAR) was calculated from the mean incoming solar radiation per day ($Q_{Global(IN)}$)

$$PAR = \frac{0.45 \times Q_{Global(IN)} \times D}{10^6}$$

where *PAR* is in MJ/m²/day, $Q_{Global(IN)}$ was measured in W/m² and *D* corresponds with the total number of seconds per day. The accumulation of received PAR per *transplanting date* per season is shown in Fig. 1a, c, e as PAR- Σ . Applied irrigation was measured in the field using two rain gauges. The total received water (irrigation + precipitation) per *transplanting date* per season is shown as received water- Σ in Fig. 2. The total received temperature (with a base temperature (T-base) of 0 °C) per *transplanting date* per season is shown as T- Σ in Fig. 1b, d, f.

In 2017's and at 2019's *September harvest*, tuber measurements were carried out by hand. First, samples were cleaned per plot to remove sand, other plant parts and small tubers, using thresholds of 20 mm in 2017 and 10 mm in



Fig. 1 Daily incident photosynthetic active radiation (PAR) from March to mid-November (field season) and PAR sum (PAR- Σ) per *transplanting date* are depicted in panels **a**, **c** and **e** for the respective years. Panels **b**, **d** and **f** show the daily minimum (T-min) and maximum (T-max) temperature and the temperature sum (T- Σ , T-base=0 °C) per *transplanting date*. Periods when plastic film was applied are highlighted with a grey blocked pattern. Across the panels, vertical arrows indicate the events of reaching 100% ground cover and harvest per *transplanting date*, which are additionally flagged with their estimated percentage of crop senescence



Fig. 2 Daily precipitation, applied irrigation and total received water sum (water- Σ) during the field period from March to mid-November for each of the *transplanting dates*. The date of 100% ground cover per *transplanting date* is indicated with vertical arrows corresponding with *transplanting date* curve pattern. For all years, the *harvest dates* in *September* are indicated with black diamonds. The *harvests* for the fully *senesced* crops in 2019 are indicated with black circles

September 2019. Secondly, total tuber fresh weight (FW) and number of tubers were assessed per plot. Individual tubers were graded into tuber size categories, using a square measure. Per tuber size class, tuber FW and number of tubers were assessed. In 2017, tubers were graded into the size classes (sc) 20 < sc < 28, $28 < sc \le 40$, $40 < sc \le 50$ and sc > 50 mm. During 2019's September harvest, tubers were graded into $10 < sc \le 28$, $28 < sc \le 40$ and sc > 40 mm. In 2018 and at 2019's senescence harvest, tubers were processed by an automated phenotyping line, as explained by Stockem et al. (2020) and detailed by van Dijk et al. (2021). The tuber phenotyping line included a 3D camera to assess tuber size and volume per individual tuber, which were used to calculate tuber FW and number of tubers per size class. The set size categories of the phenotyping line were equal to 2017's measurements, which were done by hand. Measured tuber FW and numbers of tubers per plot were converted into data per ha. For 2017, 2018 and 2019's senescence harvest, the tuber size thresholds of > 20 (= total tubers), > 28 (= marketable tubers), > 40, > 50 mm and $28 < sc \le 50$ mm (= seed tubers) were used to obtain insights into tuber yield in different size classes. As the tuber grades at 2019's September harvest were slightly different, tuber size thresholds of > 10 (= total tubers), > 28 (= marketable tubers) and > 40 mm were used.

Statistical Analysis

The programme GenStat 19th edition (VSN International Ltd. 2019) was used for statistical analyses. All experiments were analysed as spit-plot designs with a general analysis of variance. In 2017 and 2018, the significance of main effects and interactions of *date*, *age* and *genotype* were assessed. At the lowest level (sub-plot level), the *age*×*genotype* combinations were clustered and randomised within the *date* (main plot level), which was randomised in the block factor.

In 2019, the significance of main effects of *date* and *genotype* and their interactions were assessed for the *September* and *senescence harvests* separately. All significant factor or interaction means were compared using Fisher's protected LSD test ($\alpha = 0.05$).

Results

Field Management and Weather Conditions During the Three Experimental Years

In 2017, the seedlings were transplanted in ridges with a different ridge shape and height resulting in a different seedling position relative to the ridge top and ground level compared with later seasons, where initial ridges were smaller (van Dijk et al. 2021). In 2018 and 2019, a distinct V-shape in the ridge centre improved the ridges and allowed transplanting of seedlings deep in the ridge and close to the ridge base, thereby protecting seedlings against strong winds in a sheltered micro-environment (van Dijk et al. 2021). The V-shape was also able to support the plastic film, in contrast to the flatter ridge in 2017 where the seedlings supported the film resulting in damage to the seedlings.

Temperature In 2017, for *transplanting dates March* and *April*, minimum temperatures of <2 °C were observed for at least 10 days during the first 4 weeks after transplanting (Fig. 1b). Seedlings were covered with a plastic film, which, however, pressed down the plant tops and the leaves. The contact of film and seedlings caused visible frost damage: brownish discolouration followed by senescence of the damaged leaves (cf. Snoek et al. 1985). Although damaged, seedlings managed to recover when daily maximum temperatures increased above c. 15 °C.

In 2018, the first field-week after the *March transplanting*, on 14 March, included low day and night temperatures, dropping to a minimum of -4.9 °C (Fig. 1d). Despite the improved ridges and covering with a plastic film, the seedlings were heavily affected by the frost period. Increased temperatures in the second week after transplanting and onwards (see Fig. 1c, d) did not allow seedling recovery and all plants had died by 25 March. Analysis of the results of 2018 therefore excluded the *March planting date* as no data were collected.

In 2019, no *March* planting was carried out anymore. In this year, the *April date* coincided with minimum temperatures < 2 °C on a daily base in the first week after transplanting (Fig. 1f). Even with improved ridges, covered by a plastic film, top leaves of seedlings were damaged by frost. From the second week onwards,

minimum and maximum temperatures gradually increased (Fig. 1f) resulting in recovery of the seedlings of 2019's *April date*.

During the 2017, 2018 and 2019 field periods, recorded maximum temperatures were 31.3, 35.2 and 38.0 °C (Fig. 1b, d, f). The number of days with temperatures > 30 °C was 3, 10 and 11 days, respectively.

Water The received water- Σ in Fig. 2 shows that among all seasons, from planting to September harvest, 2019 received the most water. In 2017, *transplanting dates March, April, May* and *June* received on average 2.8, 3.0, 3.2 and 3.4 mm/day, respectively. For 2018, *March* excluded, these values were 2.6, 2.7 and 2.9 mm/day. In 2019, the planting dates *April, May* and *June* received on average 3.9, 4.0 and 3.9 mm/day, respectively, which was for every *transplanting date* 0.5–1.3 mm/day more than in previous years.

Besides the *September harvest*, a *full-senescence harvest* was carried out in 2019. From October, in the final phase of the season, it was observed that the difference in crop development between *planting dates April* and *May* became smaller. Therefore, the date of full crop senescence (BBCH stage 907, Kacheyo et al. 2021) was around 10 October for both planting dates (Fig. 1e, f), which was 184 days after transplanting (DAT) for the *April* planting date and 156 DAT for the *May* date. The *June transplanting date* took one more month before complete *senescence*. It was observed that night frost late October (Fig. 1f) accelerated the process of crop senescence.

Effects of Seedling Age on Yield and Number of Tubers

No significant effects of *age* were found on total yield and number of tubers in 2017 and 2018 (Table 2), nor were there any significant interactions between *seedling age* and the *transplanting date* or *genotype*. In these years, 4- and 6-week-old seedlings were transplanted on each *date*. In 2019, only 5-week-old seedlings were used on each *transplanting date* because of the lack of *seedling age* effect observed in the previous years.

Effects of Transplanting Date on Yield and Number of Tubers on a Conventional Harvest Date

Following the conventional date of potato harvest in the Netherlands, all three experimental years included a *September harvest*, see Table 1. Significantly lower yields were generally observed in the *June transplanting date* as compared with the earlier *transplanting dates* in 2017 and 2019 (see also Fig. 3a–h). On the other hand, no significant differences in yield were observed between the *March* and *May transplanting dates* in 2017 and *April* to *May dates* in 2018 and 2019 (Tables 2 and 3).

Marketable Tubers (>28 mm) In 2017, significant effects of *transplanting date* were observed on both yield and number of *marketable tubers* per ha (Table 2). The

thresholds and classes	at the harvests	s in 2017 and 20	118. Relevant n	nain effects or j	interactions ($P <$	0.05) are listed	in bold			
Year and treatment	Tuber fresh	weight (Mg/ha)) per tuber size	threshold or cl	ass	Tuber numb	ers (× 1000/ha)	per tuber size t	threshold or cla	SS
	> 20 mm	>28 mm	> 40 mm	> 50 mm	28–50 mm	>20 mm	>28 mm	>40 mm	> 50 mm	28–50 mm
2017										
Date (D)	< 0.001	< 0.001	0.083	0.717	< 0.001	< 0.001	< 0.001	0.084	0.627	< 0.001
Genotype (G)	< 0.001	< 0.001	< 0.001	< 0.001	< 0.001	< 0.001	< 0.001	< 0.001	< 0.001	< 0.001
Age (A)	0.160	0.086	0.087	0.713	0.079	0.752	0.152	0.055	0.753	0.153
D×G	0.249	0.245	0.474	0.588	0.147	0.072	0.101	0.393	0.447	0.088
$D \times A$	0.627	0.572	0.589	0.363	0.748	0.785	0.779	0.490	0.357	0.853
G×A	0.405	0.426	0.853	0.997	0.382	0.613	0.535	0.630	0.964	0.520
$D \times G \times A$	0.958	0.927	0.561	0.331	0.978	0.756	0.893	0.446	0.494	0.850
2018										
Date (D)	0.159	0.127	0.092	0.744	0.123	0.487	0.246	0.134	0.831	0.234
Genotype (G)	< 0.001	< 0.001	< 0.001	< 0.001	< 0.001	0.338	0.033	< 0.001	< 0.001	0.108
Age (A)	0.906	0.922	0.778	0.858	0.831	0.820	0.728	0.650	0.951	0.715
D×G	0.111	0.203	0.785	0.619	0.129	0.077	0.130	0.658	0.581	0.124
D×A	0.778	0.629	0.518	0.419	0.749	0.827	0.893	0.478	0.442	0.908
G×A	0.664	0.671	0.869	0.793	0.703	0.541	0.505	0.850	0.951	0.501
D×G×A	0.727	0.576	0.533	0.773	0.462	0.848	0.563	0.397	0.870	0.534



Fig. 3 Tuber yield (panels **a**–**l**) and number of tubers (panels **m**–**x**) per ha split by genotype in 2017, 2018 and 2019 *September* and *senescence harvest*. 2017 H01 (**a** and **m**), 2017 H02 (**b** and **n**), 2018 H03 (**c** and **o**), 2018 H04 (**d** and **p**), 2019 September harvest H03 (**e** and **q**), 2019 September harvest H07 (**f** and **r**), 2019 September harvest H08 (**g** and **s**), 2019 September harvest H09 (**h** and **t**), 2019 Senescence harvest H03 (**i** and **u**), 2019 senescence harvest H07 (**j** and **v**), 2019 senescence harvest H08 (**k** and **w**), 2019 senescence harvest H09 (**l** and **x**)

Table 3Significanand classes per har	ces of main el vest date in 20	ffects of transp 119. Relevant n	<i>lanting date</i> an nain effects or in	d <i>genotype</i> and nteractions (P <	their interactic (0.05) are listed	on for tuber fre: l in bold	sh weight and n	number of tubers	s of main tube	r size thresholds
Harvest date and	Tuber fresh	weight (Mg/hi	a) per tuber size	threshold or cla	ass	Tuber numbe	srs (×1000/ha) p	ber tuber size thr	eshold or class	
treatment	>10 mm	>28 mm	>40 mm ^a			>10 mm	> 28 mm	> 40 mm ^a		
2019 Harvest Septe	amber									
Date (D)	0.016	0.036	0.143			0.060	0.010	0.356		
Genotype (G)	< 0.001	< 0.001	< 0.001			< 0.001	< 0.001	< 0.001		
D×G	0.096	0.035	0.047			0.159	0.166	0.140		
	> 20 mm	> 28 mm	>40 mm	>50 mm	28-50 mm	> 20 mm	> 28 mm	> 40 mm	> 50 mm	28-50 mm
2019 Harvest senes	scence									
Date (D)	0.819	0.461	0.008	0.023	0.290	0.293	0.312	0.035	0.036	0.187
Genotype (G)	< 0.001	0.002	0.007	0.007	< 0.001	0.293	< 0.001	0.006	0.008	< 0.001
D×G	0.847	0.840	0.692	0.429	0.902	0.100	0.768	0.826	0.578	0.676
^a During 2019's Sel	otember harve	st, only three d	ifferent tuber siz	zes were measu	red					

Table 4 Means of ma ferent means $(P < 0.0)$ For <i>P</i> -values of the an	in yield param 5) are listed in alysis of varia	leters for tuber fre bold . Different la nce, see Table 2	ssh weight and etters indicate	number of tul significant dif	bers in 2017 and Terences betweer	2018 as influenc 1 <i>transplanting d</i>	ed by <i>transplan</i> l <i>ates</i> according	<i>ting date</i> and <i>g</i> to Fisher's prot	<i>tenotype</i> . Sign tected LSD te	ificantly dif- it ($\alpha = 0.05$).
Year and treatment	Tuber fresh v	veight (Mg/ha) p	er tuber size th	nreshold or class	SS	Tuber number	s (× 1000/ha) pe	er tuber size th	reshold or class	S
	>20 mm	>28 mm	> 40 mm	> 50 mm	28–50 mm	> 20 mm	>28 mm	> 40 mm	> 50 mm	28–50 mm
2017										
Mean	31.6	25.6	9.6	2.5	23.1	<i>LL</i> 6	559	120	19	540
Date (D)										
March	36.4 bc	29.0 b	11.0	2.5	26.5 b	1202 с	663 b	136	19	643 b
April	37.9 с	30.8 b	12.1	3.2	27.7 b	1144 c	654 b	143	25	629 b
May	32.2 b	26.3 b	9.6	2.3	24.0 b	970 b	580 b	116	18	563 b
June	20.1 a	16.2 a	6.8	2.1	14.1 a	593 a	340 a	83	16	324 a
Genotype (G)										
H01	23.9	17.6	4.6	0.5	17.1	719	349	47	3	346
H02	39.4	33.5	15.2	4.6	29.0	1235	769	192	36	733
2018										
Mean	26.9	23.0	8.7	2.1	20.9	730	468	91	13	455
Date (D)										
March										
April	28.6	24.6	10.3	2.2	22.3	767	487	108	14	473
May	28.3	24.4	9.2	2.2	22.2	737	494	96	13	480
June	23.8	20.0	6.7	1.8	18.1	685	424	70	12	412
Genotype (G)										
H03	32.1	28.9	13.5	3.7	25.1	706	503	131	23	480
H04	21.7	17.1	4.0	0.5	16.7	753	434	51	3	430

transplanting date June produced significantly lower *marketable yield*, 16 Mg/ha, compared with the earlier *transplanting dates*, producing between 26 and 31 Mg/ ha, which did not differ significantly from each other (Fig. 3a–b, Table 4). The number of *marketable tubers* followed a similar pattern as yield; *transplanting date June* produced significantly fewer tubers compared with earlier dates (Fig. 3m–n, Table 4).

In 2018, no significant effects of *transplanting date* on *marketable yield* and number of tubers were observed, although a similar trend was visible as in 2017 (Table 2). A mean *marketable yield* of 23 Mg/ha was obtained across the *transplanting dates April*, *May* and *June* (Table 4, Fig. 3c–d).

In 2019, during the September harvest, interaction between the factors transplanting date and genotype was significant for marketable yield (Table 3). Highest marketable yields were obtained in genotype H07 transplanted in May or June, 40 and 37 Mg/ha, respectively (Fig. 3f, Table 5). The April planting of H07 produced a yield of 33 Mg/ha, which was similar to the range of yields for genotype H03, the second highest yielding genotype, in all three planting dates (32–35 Mg/ha, Fig. 3e, Table 5). Marketable yields of genotypes H08 and H09 were more affected by late transplanting than other genotypes (Table 5, Fig. 3g–h). A significant decrease in marketable yield of H09 was observed with every subsequent transplanting date, from 28 Mg/ha when transplanted in April to only 16 Mg/ha when transplanted in June (Fig. 3h). The number of tubers harvested in September 2019 significantly differed among the transplanting dates: the earlier the transplanting was carried out, the more tubers were harvested (Table 5, Fig. 3q–t).

Seed Tubers (28–50 mm) The *seed tuber* size class was analysed for the years 2017 and 2018. Significant *transplanting date* effects were observed for *seed tuber* yield and number of seed tubers in 2017 only (Table 2). In 2017, the highest *seed tuber* yield and highest number of tubers were produced when the crop was *transplanted* between *March* and *May*, within which *seed tuber* yields ranged between 24 and 28 Mg/ha. The *June transplanting date* produced, significantly, the lowest seed tuber yield (14 Mg/ha) and the lowest number of seed tubers compared with the other *transplanting dates* (Table 4). In 2018, the mean *seed tuber* yield across *transplanting dates* was 21 Mg/ha (Table 4).

Total Tubers Highest *total tuber* yield in 2017 was attained by the *transplanting dates* of *March* and *April*, which produced 36 and 38 Mg/ha, respectively (Fig. 3a-b). At 32 Mg/ha, the *May transplanting date* produced, compared with *April*, significantly lower yield. However, *May*'s *total tuber yield* did not significantly differ from that obtained for *March*, but was significantly higher than that of *June* (20 Mg/ha, Table 4, Fig. 3a-b). Total number of tubers produced by the *planting dates* in 2017 showed a clear trend: significantly more tubers were produced in *March* and *April*, *May* produced significantly fewer tubers compared with the earlier *transplanting dates* and *June* produced the lowest number of tubers (Fig. 3m-n, Table 4).

In 2018, no significant effects of *transplanting date* were found on *total yield* or number of tubers (Table 2).

	Tuber fres	sh weight ((Mg/ha) J	per tuber	size ^a	Tuber numbers (×1000/ha) per tuber size ^a				
	Total ^b	>28	>40	>50	28–50	Total ^b	>28	>40	> 50	28–50
2019 Harvest Se	eptember									
Mean	32.9	26.8	10.2			1149	523	110		
Date (D)										
April	35.9 b	28.1 b	8.2			1327	645 c	150		
May	33.5 b	28.1 b	11.3			849	513 b	135		
June	29.4 a	24.1 a	10.9			895	481 a	155		
Genotype (C	3)									
H03	37.4 B	32.6 C	14.9 C			1024 B	546 B	147 B		
H07	45.2 C	36.4 D	14.2 C			1717 C	768 C	158 B		
H08	23.1 A	16.2 A	3.5 A			1110 B	395 A	55 A		
H09	26.0 A	21.9 B	8.0 B			747 A	383 A	83 A		
D×G										
H03										
April	40.7	34.5 e	14.0 def	2		1327	645	150		
May	35.8	31.8 de	14.0 ef			849	513	135		
June	35.8	31.5 de	16.6 f			895	481	155		
H07										
April	44.2	33.0 de	7.6 bc			2117	867	100		
May	49.1	39.8 f	15.8 f			1729	797	163		
June	42.3	36.5 ef	19.3 f			1305	641	209		
H08										
April	23.8	16.9 abc	3.0 ab			1118	432	39		
May	25.5	18.9 bc	5.9 abo	:		1055	412	68		
June	19.8	12.7 a	1.5 a			1156	342	57		
H09										
April	34.8	28.1 d	8.3 bcd	l		988	508	80		
May	23.7	22.0 с	9.7 cde	e		615	358	105		
June	19.5	15.6 ab	6.1 abo	:		638	283	64		
2019 Harvest se	nescence									
Mean	37.4	33.3	17.1	5.6	27.7	813	532	153	31	501
Date (D)										
April	36.7	31.5	13.5 a	3.1 a	28.5	913	563	132 a	19 a	544
May	38.5	34.4	17.3 b	5.3 a	29.1	832	549	156 b	29 a	520
June	37.0	34.0	20.4 c	8.3 b	25.6	695	485	172 b	45 b	440
Genotype (C	i)									
H03	37.9 B	35.2 B	22.3 B	9.8 C	25.4 A	685 AB	489 A	185 BC	52 C	437 A
H07	53.1 C	46.7 C	22.8 B	7.3 BC	39.4 B	1249 C	804 B	217 C	43 BC	761 B
H08	28.2 A	23.5 A	8.1 A	1.2 A	22.3 A	758 B	441 A	81 A	8 A	433 A
H09	30.3 AB	27.8 AB	15.1 AB	3.9 AB	23.8 A	560 A	395 A	131 AB	22 AB	373 A

Table 5 Means of main yield parameters in 2019 at both harvests influenced by *transplanting date* and *genotype*. Bold means significantly differ (P < 0.05). Significant differences between *transplanting dates*, *genotypes* and $D \times G$ interaction are shown with different letters, using Fisher's protected LSD test ($\alpha = 0.05$)

^asize threshold or class in mm;

^bTotal >10 mm for *September*, >20 mm for *senescence*

During the *September harvest* in 2019, the *April* and *May transplanting dates* produced significantly higher *total yields* of 36 and 34 Mg/ha, respectively. *June* followed with 29 Mg/ha, see Table 5.

Large Tubers No significant effects of *transplanting date* were observed in the *larger tuber size* classes with thresholds of >40 mm and >50 mm in 2017 and 2018 (Table 2). However, in 2019, a *date* × *genotype* interaction was significant for yield at the tuber size threshold >40 mm (Tables 3 and 5). Significantly higher yields of tubers >40 mm were harvested for the *May* and *June transplanting date* for *H07* than for its *April* date (Fig. 3f). The tuber yields >40 mm of *H03* ranged between 14 and 19 Mg/ha (Fig. 3e) but were not significantly different between the *transplanting dates*. Compared with *H03* and *H07*, *genotypes H08* and *H09* produced significantly lower tuber yield >40 mm, without any significant effects of *transplanting date* within the *genotypes H08* or *H09* (Fig. 3g–h, Table 5).

Genotypic Differences in Yield and Number of Tubers on a Conventional Harvest Date

In 2017, the *genotypes* showed significant differences in yield and number of tubers, as shown in Table 2. For all parameters, *genotype H02* produced a higher yield and more tubers than *H01* (Table 4, Fig. 3a–b, m–n). *Marketable yield* of *H02* was 34 Mg/ha, almost twice as much as the 18 Mg/ha produced by *H01* (Fig. 3a–b). *H02*'s *seed tuber* yield was 29 Mg/ha compared with 17 Mg/ha produced by *H01* (Table 4).

In 2018, *genotypes* significantly differed for all parameters except for number of tubers > 20 mm and number of *seed tubers* (Table 2). For all significantly different variables, *genotype H03* produced more yield and more tubers than *H04* (Table 4, Fig. 3c–d, o–p). *H03* produced 29 and 25 Mg/ha for *marketable* and *seed tuber* yield, respectively, compared with 17 Mg/ha for *H04 marketable* and *seed tuber* yield (Table 4, Fig. 3c–d).

During the *September harvest* in 2019, significant differences between *genotypes* were observed for number of *marketable tubers*, *total tuber yield* and total number of tubers as well as for number of tubers > 40 mm (Tables 3 and 5). The highest *total tuber yield*, 45 Mg/ha, was produced by *genotype H07*, followed by *H03* which produced 37 Mg/ha. *Genotypes H08* and *H09* produced the lowest *total tuber yield*, 23 and 26 Mg/ha, respectively (Fig. 3e–h, Table 5).

Transplanting Date and Genotypic Effects on Yield and Number of Tubers During a Prolonged Cropping Cycle

In contrast to the 2019 September harvest, which was before crop senescence, the fully senesced crop showed no significant effects of transplanting date or $date \times genotype$ interactions for the marketable and total tuber yield and numbers of tubers. The seed tuber size class yield and number of tubers were also not affected by transplanting date (Table 3). Measured mean yields were 33, 28 and 37 Mg/ha

for *marketable*, *seed tuber* and *total yields*, respectively (Table 5). The yield and number of *larger-sized* tubers, >40 and >50 mm, significantly differed among *transplanting dates* (Table 3). The yield of tubers >40 mm increased significantly with every subsequent *transplanting date* (Table 5). *Transplanting* in *April*, *May* and *June* produced 14, 17 and 20 Mg/ha tuber yield >40 mm, respectively. Yield of tubers > 50 mm was significantly higher when *transplanted* in *June* (8 Mg/ha, Table 5, Fig. 3i–l) than when *transplanted* in *April* or *May* (3–5 Mg/ha).

Significant *genotype* effects were observed for all measured variables at the *senescence harvest* (Table 3). Similar to 2019's *September harvest*, H07 produced, significantly, the highest *marketable* and *total tuber yields* (47 and 53 Mg/ha, respectively; Table 5, Fig. 3j). The second highest *marketable* and *total tuber yields* were produced by H03, which produced 35 and 38 Mg/ha, respectively (Fig. 3i). H08 and H09 produced significantly lower *marketable yield* and *total tuber yield* compared with the other two *genotypes* (Table 5). The highest *seed tuber yield* was produced by H07, 39 Mg/ha (Table 5). Significantly lower *seed tuber yields* were produced by H03, which produced 5). Significantly lower seed tuber yields were produced by H03, H08 and H09, with yields between 22 and 25 Mg/ha, which did not significantly differ from each other (Table 5).

Discussion

The aim of this study was to assess and understand — under Dutch field conditions — the effects of the *date of transplanting* of greenhouse-raised hybrid potato seedlings and the *age of seedlings* at *transplanting* on potato yield. The possibilities of growing a full potato crop from transplanted hybrid seedlings and possible options for producing optimum yields while reducing the risk of unfavourable conditions will be discussed.

Over 3 years of experiments, all *transplanting dates* were able to produce reasonable yields, whereas highest measured *marketable yields* over *transplanting dates* ranged between 29 and 34 Mg/ha (Tables 4, 5). Highest *seed tuber yields* attained were between 26 and 29 Mg/ha. The current study shows that earlier *transplanting*, in *March* and *April*, does not obviously result in higher *marketable* or *seed tuber yields*. The earliest studied *date*, *March*, was highly prone to risks of crop failure due to low temperatures and (night) frost. Under Dutch circumstances, night frost risks and incidental colder temperatures are present until mid-May. However, the effect of *transplanting date* was much smaller and less pronounced than anticipated, rather, the use of different experimental hybrid *genotypes* determined the differences in yield levels, which partly might have been caused by their differences in crop cycle length.

Effects of Seedling Age at Transplanting

In 2017 and 2018, field *transplanting* of 4- and 6-week-old seedlings showed no significant effects of *seedling age* on yield and number of tubers (Table 2). Due to the lack of significant influence of the *seedling ages* on yield parameters over the first

two trial seasons, an optimal *age* for field *transplanting* could not be determined in this study. Therefore, and due to adequate performance of 5-week-old seedlings in recent studies (Kacheyo et al. 2021; van Dijk et al. 2021), only 5-week-old seedlings were used in 2019.

Seedling age at transplanting is entangled with the seedling development and phenology at transplanting. The use of transplant age to characterise transplantable seedlings was largely based on the time taken from sowing, through hardening off, up until transplanting. To assure a uniform selection of transplantable seedlings and eliminate off-types, an additional selection based on seedling growth stage, vigour and size was carried out. This resulted in seedlings with a stem of 2-3 cm and 2-4 true leaves, seedlings with a stem of 7-12 cm and 5-8 leaves and seedlings with a stem>14 cm and>8 true leaves (see 'Materials and Methods') being selected for the different age classes (4, 5 and 6 weeks) under the experimental conditions. However, seed quality, precision of sowing and conditions during germination (BBCH stage 01-05, Kacheyo et al. 2021) and emergence (BBCH stage 07-09, Kacheyo et al. 2021) (Almekinders 1995; Lamichhane et al. 2018) among other factors do influence the growth of seedlings in a nursery (Buishand et al. 1985; Everaarts et al. 1993; Kerbiriou et al. 2013). At the moment, the conditions for optimal growth of hybrid seedlings in the greenhouse nursery as well as the optimal period and conditions of the seedling hardening-off phase have not been studied in detail, nor defined.

Night Frost Impacted the Effects of Transplanting Date

The choice of the time to transplant (Table 1) was guided by possible adverse effects of the environmental conditions, especially in the initial stages of crop growth. Fig. 4 shows details on the minimum temperatures and frost sums around the different planting dates in the three experimental years. Field establishment of seedlings from the earliest date, March, was only achieved in 2017, when only sparse occurrences of frost were observed in March (Fig. 4a) and the mild frost conditions allowed the transplants to establish and withstand the frosts. In 2018, the March planted seedlings were heavily affected by frost (Fig. 4b) despite the covering with a plastic film during the period when temperatures were <2 °C. Plants did not manage to survive the stresses induced by the frost, leading to the loss of all plants of this transplanting date. The different effects of frost in the 2 years might be attributed to a combination of incidence of frost at the *date of transplanting* and a frost sum (frost- Σ) of – 20 °Cd within the first 1.5 week in the field in 2018 (Fig. 4b). Additionally, in 2017, the April date also coincided with frost occurrences, although not on the day of transplanting and also less violent than in 2018's March date (Fig. 4a). In April 2019, frost occurred the second night after transplanting, and a frost- Σ of -6.6 °Cd was reached on the fifth day in the field (Fig. 4c). Plants were visibly affected by frost, but recovered in the following weeks. Therefore, transplanting in March, but also early in April, is precarious as any prolonged occurrence of frost may result in accumulated effects which may eventually cause irreversible damage



Fig. 4 Detailed zoom-in on the colder (between – 5 and 10 °C) daily minimum temperatures (T-min), the temperature sum $(T-\Sigma)$ per *transplanting* (TP) *date* and the sum of negative temperatures (frost- Σ) per *transplanting date* for the periods from 10 March until the latest night frost event (10 May) in the three experimental years (for all Σ T-base = 0 °C). *Dates of transplanting* are indicated with stars

to the plants. In addition, under Dutch conditions, the local field situation might be too wet to prepare a decent planting bed or ridges in early spring (Poll et al. 1994).

Contribution of Transplanting Date to Tuber Yield on a Conventional Harvest Date

The duration of a crop's growing period strongly influences crop yield. For a potato crop, temperature and radiation are important yield-determining factors when grown under non-limiting conditions. Furthermore, the crop cycle length of a cultivar may also influence the yield attained at the time of harvest, with long cycle cultivars requiring a longer growing period (Kooman 1995).

Transplanting in *March* as well as *April* advanced the crop growth cycle from *transplanting* up to the *conventional September harvest*, but coincided with the risk of frost. When comparing the abovementioned advanced crop growth cycles — starting in *March* or *April* — with starting in May, the additional accumulated PAR and temperature (Fig. 1) did not result in more yield (Tables 4 and 5). Thus, on average, the crop formed less tuber FW per day in the field when transplanted early than when transplanted in *May*. When transplanted in *June*, on the other hand,

accumulation of tuber FW per day was high, but when harvested in *September*, the crop cycle duration was too short to make similar yields as the other *transplanting dates*. Therefore, prospects for field transplanting exercises for a successful growing period in the conventional cropping cycle — *September harvest* — gravitate towards transplanting on the *April* or *May* dates. Hence, this study defines the optimal window for field transplanting of hybrid potato seedlings between the beginning of *April* and the end of *May*. A defined optimal *transplanting date* is an important ingredient to add to the novel method for production of seed(ling) tubers for the current Dutch potato sector.

Contribution of Crop Cycle Length to Experimental Hybrid Genotypes Tuber Yield

Fig. 5 shows a general trend (not significantly tested) where a shorter crop cycle results in lower *marketable yields*. However, the figure also shows that the individual experimental genotypes used in this study differ in the number of days needed to reach their maximum yield.

For instance, interpolation between *planting dates* of *H01* in 2017 shows that maximum marketable yield was attained at around 130 DAT, compared with 160 DAT for *H02*, which reflects *transplanting* in *May* and *April*, respectively, when harvested in September (Fig. 5a). Also in 2018 (Fig. 5b), the effects of additional days in the crop cycle on the marketable yield in September seemed to be largely *genotype* dependent; *genotype H04* reached its maximum yield already at about 100 DAT (*transplanting* in *June*) — a longer growing period did not result in additional yield — and *genotype H03* needed at least 130 days in the field (*May* date) to reach maximum yield.

In 2019, when four *genotypes* were used and two *harvests* were conducted — in *September* and at crop *senescence* — differences in yield and number of marketable tubers among *genotypes* were clearly observed (Fig. 3e–l, Table 5). Additionally, the effect of a prolonged crop cycle was observed per *genotype* for each *transplanting date* (Fig. 5c–f). The amount of additional marketable yield from the early (*September*) to the late (*senescence*) *harvest* differed much among *genotypes*, increasing the length of the crop cycle showed most effect for the late planting in *June*.

Based on Fig. 5, *transplanting* in *June* and *harvesting* in *September* seems more feasible for a short cycle cultivar. *Transplanting dates April* and *May* would be more suitable for medium cycle cultivars when *harvested* in *September* and long crop cycle cultivars when *harvested* in October at crop *senescence*. The choices of the *date of transplanting* and the *date of harvest* are strongly influenced by the duration of the *genotype*'s crop cycle, the suitability of the *genotype* for a given set of environmental conditions and purpose.

Opportunities for Potato Cropping Systems Under Dutch Conditions

Fig. 5 indicates that the effects of crop cycle length on yield were *genotype* dependent. As Fig. 5 also suggests that both manipulation of the crop cycle duration at the



Fig. 5 Produced *marketable yield* (> 28 mm) plotted against the length of the growing period in the field. As in 2017 (**a**) and 2018 (**b**), the harvest of all *transplanting dates* was carried out on the same day in *September*, the shortest growing period was created by *transplanting date* in *June*, of which *marketable yield* is indicated with the most left marker. Panels **c**, **d**, **e** and **f** correspond with the 2019 transplanted genotypes *H03*, *H07*, *H08* and *H09*, respectively. Per *transplanting date*, the left-side marker corresponds with the *harvest* in *September* and the right-side marker with the *harvest* at crop *senescence*

start (planting) or at the end (harvest) could have considerable effects on the yield of some of the experimental hybrid genotypes used in this study. Besides, in more general, this study showed that early or late *transplantings (March* or *June*, respectively) were disadvantageous for the used experimental *genotypes*. Yet, combined with the innovation of hybrid potato breeding, early or late *transplanting* of hybrid diploid genotypes may give — hypothetically — a whole new set of opportunities to Dutch arable farmers.

The genotypes in this study could withstand short periods of colder and frosty conditions, as shown in Fig. 4. Because hybrid potato breeding at the diploid level gives breeders the possibilities for more targeted introduction or stacking of favourable genes (Su et al. 2020), possibilities arise for breeding both short and long crop cycle genotypes as well as cold or frost tolerant genotypes (Bethke et al. 2019). Short cycle hybrid genotypes will allow growers to cultivate and harvest a potato crop in a shorter crop cycle compared with intermediate or long crop cycle genotypes. It allows growers to cultivate besides (hybrid) potatoes also specific green manure and/or catch crops which are meant to increase soil organic matter, 'catch' the surplus of soil nitrogen and release it in the next season or reduce the number of plant parasitic nematodes by interrupting their life cycle. Growing such crops in the same growing season as a potato main crop fits perfectly in the new ambitions of the Dutch government and agricultural sector to strive for a more circular and agricultural system, depending less on external (chemical) inputs.

Moreover, short cycle hybrid potato will be an ideal new crop for vegetable growers in the Netherlands. Due to their experience with transplanted vegetable crops like cabbage, lettuce, spinach, leek, broccoli, endive and celery, integration of hybrid transplanted potato into their existing systems may be effortless. The aforementioned vegetables either have short cropping cycles and/or cold tolerance, which allows for early planting; or are hardy, thereby allowing for a late planting and later harvest during winter periods.

The machinery used in these vegetable cropping systems can also be used in transplanting and management of hybrid potato seedling crops. Furthermore, most agronomic practices such as cultivation practices, irrigation practices and management of frost by covering with plastic films can be adapted for new transplanted hybrid potato systems.

Additionally, short crop cycle varieties which tolerate late season transplanting including long haulm-stay-green traits will allow growers to include a potato cultivation — together with short-season vegetables — at the end of the cropping season. Ideally, these new hybrid potato varieties have the potential to serve the fresh potato market, focusing on consumer traits like taste, nutrition value and quality. Cultivation directly from hybrid TPS will allow growers to quickly anticipate for new hybrid varieties with improved resistances or changed preferences in the consumer markets compared with a traditionally bred crop grown from seed tubers.

Further Studies

The current study did not present a clear answer on the most suitable *trans-planting dates* — *March* to *June* — for yield and number of tubers. As significant effects of *genotype* and interactions of *genotype* and the *transplanting date* were present in every season, it is critical to determine the optimal *date of transplanting* hybrid potato seedlings. It is suggested to study this in a set of experiments solidly replicated in space and time. Especially, when considering the next step of innovation in hybrid potato breeding where the shift will be made from experimental hybrid genotypes to hybrid varieties for specific potato production markets. When new hybrid potato cultivars with distinct crop cycle types will be available, it is recommended to do more detailed studies on the feasible window of transplanting to produce optimum yields. As all *transplanting dates* in this study occurred in the first half of the month, it is also important to identify the optimal *date of transplanting* within the months of *April* and *May*.

In addition, more insight is needed into the ideal size of a seedling for transplanting. These studies should also take into account the nursery conditions needed to produce seedlings of a desired size and additional hardening-off and field management practices which also contribute to a minimal transplanting shock.

Conclusions

The choice of *transplanting date* greatly influences transplant establishment and yield because very early planting in *March* comes with increased risks of crop failure due to extended frost. However, hybrid potato breeding might anticipate on decreasing these risks in the future. No clear differences between *transplanting dates* have been observed for *April, May* and *June dates* when harvested at complete *senescence*. However, at the conventional harvest time in September, *transplanting dates April* and *May* were able to out-yield *June*. For *larger tuber sizes* (>40 mm), harvesting at earlier crop stages other than complete *senescence* showed interactions between *transplanting date* and the *genotype* whereas at complete *senescence*, a significant *transplanting date* effect was observed; late planting in *June* seemed to increase the share of *larger-sized* tubers and enhance yields.

The yields of 4- and 6-week-old seedlings were not significantly different. However, it is currently not known if 5-week-old seedlings might outperform the former due to a more ideal size or stage of development.

Genotypic effects were substantial in all seasons, and for some genotypes, the length of the cropping cycle might be more important than the date of transplanting. Harvesting in September, when transplanted in June, seemed to be too early to deliver a fully matured crop using its full yield potential. The choice of time to harvest especially for late planting dates is therefore influenced by the time of transplanting and the crop cycle length of the genotype.

Low minimum temperatures in early May did not seem to have much effect on the crops as they did not result in crop failure or lower yields compared with transplanting in June. Moreover, the experimental genotypes could withstand periods of low maximum day temperatures and several short events of night frost, which hints to a certain degree of cold tolerance in the diploid hybrid genotypes used. Introducing traits favouring short cropping cycles and cold/frost tolerance with hybrid breeding does create the opportunity to grow potato as a transplantable vegetable under Dutch agronomical conditions.

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Author Contribution LCMvD designed the experiments. LCMvD and OCK managed the field trials and collected the data. LCMvD analysed the data. Data interpretation and drafting the manuscript were done by LCMvD and OCK. The manuscript was revised based on input of all authors. All authors approved the final version.

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Declarations

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Conflict of Interest WJML is editor of Potato Research; PCS is editor-in-chief of Potato Research.

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