

## Indigenous fermented foods

Food microbiology: fundamentals and frontiers, 3rd edition

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## Indigenous Fermented Foods

A food is considered fermented when one or more of its constituents have been acted upon by selected microorganisms or their enzymes to produce a significantly altered final product desirable for human consumption. Most fermentations are caused by molds, yeasts, or bacteria, either singularly or in combination. Indigenous or traditional fermented foods have been prepared and consumed for hundreds of years and are strongly linked to cultures and traditions of millions of people around the world, especially in rural communities. The origins of most fermentation technologies have been lost in the mists of history. Some products and practices no doubt fell by the wayside; those that remain today have survived the test of time. Fermented food products are important components of the diet as staples, adjuncts to staples, condiments, and beverages.

This oldest form of food biotechnology originated as a necessity for enhancing the keeping quality of diverse plant and animal food materials through organic acid, alcoholic, and alkaline fermentations. Fermentation, a relatively efficient low-energy preservation process, also improves digestibility, flavor, appearance, nutrient

contents, and other quality attributes and reduces antinutritional components of the substrates and cooking time. Many fermented foods are now receiving global attention for their health-promoting or disease-preventing or -curing effects. Whereas a considerable number of food fermentation processes have been scaled up for commercial purposes, most types of fermented foods are still produced on a home scale. Such products often contain mixed genera and populations of microorganisms because of the lack of controlled processing facilities. In view of larger-scale industrialized food fermentation, microbial ecology and metabolic activities of functional microorganisms must be investigated. Present-day developments in molecular methods enable accurate characterization of strains and development of tailor-made fermented food products.

A variety of fermented foods can be found widespread over the world. Following the sequence in Table 38.1, some of them will be described in this chapter, mainly to illustrate the complexity of biochemical, nutritional, and sensorial changes that result from an array of microbial activities in a range of raw materials.

Table 38.1 Some important indigenous fermented foods

Product(s)	Country(ies) and/or area(s)	Substrate(s)	Functional microflora <sup>a</sup>	Type(s) of fermentation <sup>b</sup>	Description and usage
Cereal and starch crop products					
Ang-kak	China, Southeast Asia	Rice	Molds	SSF, TS	Dry purple-red powder; colorant
Banku	Egypt	Maize or cassava	Yeasts, LAB	SSF, N	Dumpling; staple
Ben-saalga	Burkina Faso	Pearl millet	Bacteria, LAB	SmF, N	Sour, thin gruel; breakfast staple and infant food
Bhatura	India, Pakistan	Wheat	LAB, yeasts	SSF, TS	Deep-fried, flat, leavened bread; snack
Bouza	Egypt	Wheat	Yeasts, LAB	SmF, TS	Pale yellow, thick, sour alcoholic drink
Breads (leavened yeast and sour-dough breads)	Worldwide	Wheat and/or rye	Bakers' yeast or yeast-LAB mixed cultures	SSF	Baked leavened dough; staple
Busaa	Kenya, Uganda	Maize, finger millet	Yeasts, LAB	SmF, N	Sour alcoholic drink
Chicha	South America	Maize	Molds, yeasts, LAB, AAB	SmF, N	Clear, yellowish, effervescent, sour alcoholic drink
Deguè	Burkina Faso	Pearl millet	Bacteria, molds	SSF, N	Balls diluted with milk or water to make porridge
Gari	Nigeria, West Africa	Cassava	LAB, yeasts	SSF, N	Granulate; precooked instant porridge; breakfast staple
Injera	Ethiopia, Sudan	Teff (or corn or sorghum)	LAB, yeasts	SSF, N	Sour, soft, steam-baked, flat pancake; staple
Jalebi	India	Wheat	LAB, yeasts	SSF, N	Pretzel-like syrupy confection
Jnard	India, Nepal, Bhutan	Finger millet	Molds, yeasts, LAB	SSF, TS	Sweet-sour alcoholic drink
Kenkey	Ghana	Maize	LAB, yeasts	SSF, N	Sour dumpling; cooked; staple
Lafun	Nigeria, West Africa	Cassava	LAB	SmF, N	White flour made into a stiff porridge; staple
Lao-chao	China, Indonesia	Rice	Molds, yeasts	SmF, TS	Sweet-sour, juicy, alcoholic snack
Mahewu	South Africa	Maize	LAB	SmF, N	Sour, nonalcoholic drink
Mawè	Benin, Togo	Maize	LAB, yeasts	SSF, N	Sourdough made into porridge or gruel; staple
Merissa	Sudan	Sorghum	LAB, yeasts	SSF-SmF, N	Thick, sour alcoholic drink
Minchin	China, Thailand	Wheat (or rice)	Molds	SSF, N	Thin strips or noodles; staple
Munkoyo	Zambia, Zaire	Maize	LAB, yeasts	SmF, N	Sweet-sour alcoholic drink
Naan	Afghanistan, Iran, Pakistan, India	Wheat	Yeasts, LAB	SSF, N	Flat, baked bread; staple
Ogi	Nigeria, West Africa	Maize or sorghum or millet	Molds, yeasts, AAB, LAB	SmF, N	Sour gruel; staple
Pito	Nigeria, Ghana	Sorghum or maize	Molds, LAB, yeasts	SmF, TS	Sweet-sour alcoholic drink
Poi	Hawaii	Taro corm	LAB, yeasts	SSF, N	Sour porridge; staple or condiment
Poto-poto	Congo	Maize	LAB, yeasts	SSF, N	Sourdough balls made into porridge or gruel; staple
Pozol	Mexico	Nixtamal <sup>c</sup>	Molds, yeasts, LAB, other bacteria	SSF, N	Balls diluted with water to make sour, nonalcoholic porridge
Puto	Philippines	Rice	LAB, yeasts	SSF, N	Spongy, steamed cake; snack
Ruou nep than	Vietnam	Rice	Molds, yeasts	SSF-SmF, TS	Alcoholic drink

(Continued)

Table 38.1 (Continued)

Product(s)	Country(ies) and/or area(s)	Substrate(s)	Functional microflora <sup>a</sup>	Type(s) of fermentation <sup>b</sup>	Description and usage
Saké	Japan	Rice	Molds, yeasts, bacteria, LAB	SmF, TS	Alcoholic drink
Tapé	Indonesia	Cassava or rice	Molds, yeasts	SSF, TS	Sweet-sour alcoholic snack
Tapuy	Philippines	Rice	Yeasts, LAB	SSF, TS	Sour-sweet alcoholic drink
Yakju and Takju	Korea	Rice	Molds, yeasts, LAB, other bacteria	SSF-SmF, TS	Alcoholic drink
Legume products					
Daddawa	West Africa, Nigeria	African locust bean	Bacteria	SSF, N	Flavoring agent; soup and stew ingredient
Inyu	Taiwan, China, Hong Kong	Black soybeans	Molds, LAB, yeasts	SmF, N	Syrup; flavor enhancer
Kecap asin	Indonesia	Soybeans	Molds, LAB, yeasts	SSF-SmF, TS	Thin, transparent, light brown salty liquid; condiment
Kecap manis	Indonesia	Soybeans, palm sugar, herbs	Molds, LAB, yeasts	SSF-SmF, TS	Thick, dark brown syrup; sweet condiment
Kinema	India, Nepal	Soybeans	Bacteria	SSF, N	Paste made into thick curry; side dish
Meitauza	China, Taiwan	Soybean press cake	Molds	SSF, N	Cake, fried or cooked; side dish
Meju	Korea	Black soybeans	Molds, LAB, yeasts	SSF-SmF, N	Syrup; seasoning agent
Natto	Japan	Soybeans	Bacteria	SSF, PS	Mucilaginous snack
Oncom	Indonesia	Peanut press cake	Molds	SSF, TS	Cake, deep fried or roasted; side dish or soup ingredient
Papad	India, Pakistan, Bangladesh	Black gram	Yeasts	SSF, N	Deep-fried or roasted snack or condiment
Sufu	China, Taiwan	Soybean curd	Molds	SSF, N	Paste; condiment
Tempeh	Indonesia	Soybeans	Molds, bacteria	SSF, TS	White, mold-penetrated and covered cake, stewed or deep fried; side dish, snack, or soup ingredient
Wadi	India, Pakistan, Bangladesh	Black gram	LAB, yeasts	SSF, N	Balls or cones; condiment
Cereal-legume mixture products					
Dhokla	India	Rice, Bengal gram	LAB, yeasts	SSF, N	Steamed, soft cake; snack
Idli	India, Sri Lanka	Rice, black gram	LAB, yeasts	SSF, N	Steamed, spongy cake; snack
Miso	Japan	Soybeans, rice	Molds, yeasts, LAB	SSF, TS	Paste; soup base or seasoning agent
Soy sauces	East and South-east Asia	Soybeans, wheat	Molds, LAB, yeasts	SSF-SmF, TS	Brown, salty liquid; seasoning agent
Taoco	Indonesia	Soybeans, cereals	Molds, LAB, yeasts	SSF-SmF, TS	Yellow paste; seasoning agent
Vegetable products					
Gundruk	India, Nepal	Mustard leaves	LAB	SSF, N	Shreds; soup ingredient or pickle
Soibum, Mesu, Naw-Mai-Dong	India, Nepal, Thailand	Young bamboo shoot	LAB, yeasts	SSF, N	Cubes; consumed as a pickle or made into curry

(Continued)

Table 38.1 Some important indigenous fermented foods (Continued)

Product(s)	Country(ies) and/or area(s)	Substrate(s)	Functional microflora <sup>a</sup>	Type(s) of fermentation <sup>b</sup>	Description and usage
Kanji	India, Pakistan, Israel	Carrot or beet	Yeasts	SmF, N	Deep purple, sour, alcoholic drink
Kimchi	Korea	Cabbage (or radish taproot), garlic	LAB	SSF, N or TS	Sour, carbonated vegetable; staple
Sauerkraut	Europe, Russia, United States	White cabbage	LAB	SSF-SmF, N or PS	Sour shreds; consumed raw or cooked with meat or sausages
Sinki	India, Nepal	Radish taproot	LAB	SSF, N	Sour shreds; pickle or soup ingredient
Dairy products					
Gorgonzola, Blue Stilton, Roquefort cheese	Italy, United Kingdom, France	Cow's or sheep's milk	LAB, molds	SSF, TS or PS	Blue-veined cheese; strong-flavored side dish or cooking ingredient
Camembert cheese	France	Cow's milk	LAB, molds	SSF, TS or PS	Mold surface-ripened cheese; soft-texture side dish with gradually developing strong flavor
Dahi	India, Pakistan, Bangladesh, Sri Lanka	Cow's or buffalo's milk	LAB	SmF, N or PS	Thick gel; dessert
Gouda cheese	The Netherlands	Cow's milk	LAB	SSF, TS or PS	Small-eyed or blind cheese; multipurpose protein food
Kefir	Scandinavia, Russia	Goat's, sheep's, or cow's milk	Yeasts, LAB	SmF, TS	Effervescent, sour, mild alcoholic drink
Koumiss	Russia	Mare's milk	LAB, yeasts	SmF, N	Effervescent, cloudy, sour, alcoholic drink
Lassi	India	Cow's or buffalo's milk	LAB	SmF, N	Sour drink
Yogurt	Europe, worldwide	Cow's milk	LAB	SmF, TS or PS	Viscous or thick gel; dessert or side dish
Fish products					
Bagoong	Philippines	Fish or shrimp or oyster	Bacteria	SSF, N	Brown paste; condiment
Izushi	Japan	Fish, rice, vegetable	LAB	SSF, N	Pickle
Katsuobushi	Japan	Bonito or skipjack tuna	Molds	SSF, N	Strips, dried; seasoning agent
Som-fak	Thailand	Fish fillet, rice, garlic	LAB	SSF, N	Served raw or cooked; main course or snack
Meat products					
Country-cured ham	Europe, United States	Pork	Bacteria, LAB, molds	SSF, N	Cured meat; ham slices consumed raw or cooked
Nem	Vietnam	Pork, garlic	Bacteria, LAB	SSF, N or TS	Meat cubes, fried; side dish
Nham	Thailand	Pork, cooked rice	LAB	SSF, N	Sour slices, deep fried, crispy; snack
Salami	Europe, United States	Pork and beef	LAB	SSF, N or PS	Sausage

(Continued)

Table 38.1 (Continued)

Product(s)	Country(ies) and/or area(s)	Substrate(s)	Functional microflora <sup>a</sup>	Type(s) of fermentation <sup>b</sup>	Description and usage
Miscellaneous products					
Balao balao	Philippines	Rice, shrimp	LAB	SSF, N	Main dish or sauce
Basi	Philippines	Sugarcane juice	Yeasts, LAB	SmF, TS	Sweet-sour, effervescent, cloudy, alcoholic drink
Bongkrek	Indonesia	Coconut press cake	Molds	SSF, TS	Bars, roasted or fried; snack or soup ingredient
Kishk	Egypt, Syria, Lebanon, Jordan, Iraq, North Africa	Wheat, milk	LAB, yeasts, other bacteria	SSF, N	Brownish, sour, dried balls; snack or soup ingredient
Kombucha	Japan, Indonesia, China, Russia	Tea liquor, sugar	AAB, yeasts	SmF, TS	Sour, mildly alcoholic drink
Miang, or Leppet-So	Myanmar, Thailand	Tea leaves	LAB	SSF, N	Sour-bitter tasting soft snack
Tarhana, or Trahana	Turkey, Greece	Tomatoes, wheat flour	Yeasts	SSF, N or PS	Tomato dough, dehydrated granulate; soup ingredient
Palm wines	All tropical palm-growing countries	Sap of coconut, date, palmyra, oil, nipa, raphia, or kithul palm	Yeasts, LAB, bacteria, AAB	SmF, N	Sweet-sour alcoholic drink
Ugba, or Ogiri	Nigeria, West and Central Africa	African oil bean or castor oil beans or melon or sesame seeds	Bacteria	SSF, N	Dark brown balls; salad ingredient or flavoring agent in soups, stews, and sauces

<sup>a</sup> AAB, acetic acid bacteria; LAB, lactic acid bacteria.

<sup>b</sup> SSF, solid-state fermentation; SmF, submerged fermentation; SSF-SmF, solid-state fermentation followed by submerged fermentation; N, natural and/or backslotted fermentation; TS, traditional undefined starter; PS, pure culture starter.

<sup>c</sup> Corn grains cooked in alkaline water.

## CEREAL AND STARCH CROP PRODUCTS

### Bakery Products

Bread, in various forms, has been a staple in the diets of many population groups for many centuries. The history of bread traces back to about 3,000 B.C. The development of cereal foods has proceeded through several stages, from roasted grain to gruels to flat breads and finally to leavened bread loaves. Early Egyptians developed the use of fermentation for breads and constructed baking ovens. We will focus here on leavened breads owing their sensorial properties, at least in part, to fermentative activities of microorganisms.

In principle, bread is made from dough that is fermented and baked. The essential ingredients are wheat or rye flour, salt, water, and a leavening agent (26). Usually some fat, sugar, milk solids, and bread-improving emulsifiers are added, but these are not essential. The function of water (50 to 60% of flour weight) is to hydrate

the starch and gluten (extensible and elastic proteins in wheat), enabling the mixing and kneading of a viscoelastic dough that retains the carbon dioxide gas formed during fermentation. The most commonly used leavening agent is bakers' yeast, *Saccharomyces cerevisiae* (27), which is commercially available as dehydrated granules (instant dry yeast), fresh yeast cake, or yeast cream (a suspension). Dry yeast must first be reactivated in a flour-water suspension for about 20 min. Yeast cream and cake have the advantage that no activation is required, but they are prone to spoilage by lactic acid bacteria (LAB) and thus have limited shelf lives. Based on flour weight, about 1 to 6% yeast dry matter is required. The function of salt (1 to 2%) is to moderate the fermentation rate in order to obtain a steady production of gas that can be adequately retained in the dough. After fermentation for several hours at 25 to 30°C, doughs are remixed to obtain a homogenous distribution of gas cells. The dough is portioned to the required weight or size and is

molded and put into baking pans. After another period of fermentation, the dough has at least doubled its volume and is baked in a hot-air or steam oven for 20 to 40 min at temperatures ranging from 180 to 230°C.

Sourdough bread is slightly acidic because of the leavening agent, sourdough. In contrast to the pure-culture bakers' yeast, sourdough comprises a stable, mixed microflora containing  $10^7$  to  $10^9$  LAB CFU/g, predominantly *Lactobacillus sanfransiscensis* and occasionally *Lactobacillus pontis*, *Lactobacillus panis*, *Lactobacillus frumenti*, or *Lactobacillus reuteri*, and  $10^5$  to  $10^7$  yeast CFU (predominantly *Candida milleri*)/g, obtained by repeated propagation of sourdough fermentations by reinoculation. Long-term propagation of a sourdough during the last seven decades has been documented (16), and anecdotal reports exist of sourdoughs maintained over several centuries. Commercial sourdough starters have been developed and are available as dehydrated granules and (semi)dried preferments. Sourdoughs are required for rye breads to achieve bakeability, and they are widely used in rye and wheat breads because of the high sensory quality they impart to these breads. Sourdough contributes to the characteristic flavor (61), improves texture, and delays staling and microbial spoilage of bread (60).

Naan (naan) is made by mixing white wheat flour with sugar, salt, backslop (called khamira), and water. The hand-kneaded dough is left in an earthen jar to ferment for 12 to 24 h. After maturity, the leavened dough is made into balls, placed on a smooth surface sprinkled with flour, and flattened by a wooden rolling pin. Smoothly flattened round dough is transferred onto a circular pad of cotton cloth and is slapped onto the inner wall of the clay-clad brick oven, called the tandoor, where it sticks for baking at 120 to 150°C until the dough is puffed off and light brown. The bread is speared with a skewer and removed from the oven wall to be served hot, usually along with meat or chicken preparations. From a new dough (pH 5.9) for making naan,  $10^5$  CFU of yeasts/g and  $10^2$  CFU of LAB/g can be obtained compared with respective counts of  $10^8$  and  $10^9$  CFU/g from ripe, fermented dough (pH 4.8) (5). *S. cerevisiae* is the predominant yeast. Presently, bakers' yeast and dahi are added to shorten the fermentation period.

### Kenkey

Kenkey is a dense, sour-tasting, cooked mass, served as thick slices at breakfast combined with tea, sardines, or other foods. Cleaned whole corn (maize, *Zea mays*) kernels are soaked in water for 2 days; during this period, the kernels soften (43), which is essential during the next operation, i.e., coarse wet milling. The resulting wet grits

are kneaded into a stiff dough, which is covered and left to ferment at ambient temperature (25 to 30°C) for 2 to 4 days. Dominant microorganisms are obligate heterofermentative lactobacilli, e.g., *Lactobacillus fermentum*, and yeasts, mainly *Candida krusei* and *S. cerevisiae*. When fermented according to local preference for odor and acidity (28), the dough is divided into two equal portions, and one portion is cooked, with the addition of some water, while being kneaded continuously with a cooking stick into an elastic gelatinized paste called aflata. The aflata is then mixed through the remaining uncooked dough. The resulting mass is molded by hand into units of 200 to 400 g and wrapped in banana leaves (Fanti kenkey) or corn sheaths (Ga kenkey). The packages are cooked by immersion in boiling water for a few hours. The function of the aflata is twofold: it acts as an adhesive, keeping the mixture in shape, and it carries the water needed for the swelling (gelatinization) of the uncooked, gritty dough. During the fermentation, the level of available lysine (and thus protein quality) and nutrient bioavailability increase, and flavor compounds (2,3-butanediol, butanoic acid, lactic acid, 3-methylbutanoic acid, octanoic acid, 2-phenylethanol, and propionic acid) are formed (28).

### Mawè

Mawè is an intermediate product used for the preparation of, e.g., ablo, a steam-cooked corn bread, and porridge, e.g., aklui (23). To prepare mawè, cleaned, dry, whole corn kernels are milled into grits, partly reground to obtain a fine grind, mixed with water, kneaded into a dough, covered, and allowed to ferment naturally during 2 to 4 days at ambient temperature (30°C). The pH decreases to 3.7 to 3.8, and an attractive freshly sour flavor is formed due mainly to heterofermentative lactobacilli (*Lb. fermentum* and *Lactobacillus cellobiosus*) and *C. krusei*.

### Ogi

To prepare ogi, kernels of corn are soaked in warm water for 1 to 3 days, after which they are wet milled and sieved with water through a screen to remove fiber, hulls, and much of the germ. The filtrate is fermented to yield a sour, white, starchy sediment. Fermentation is by lactobacilli (*Lb. fermentum*, *Lb. cellobiosus*, *Lactobacillus brevis*, and *Lactobacillus plantarum*) originating from the environment, although other bacteria (*Enterobacter sakazakii* and *Corynebacterium* spp.) and yeasts (*C. krusei*, *Candida kefir*, and *Rhodotorula* spp.) are also involved (42). Ogi may be diluted in water to 8 to 10% solids and boiled into a pap or cooked and turned into a stiff gel (eko) before eating.

Ogi is a major breakfast cereal for adults and a traditional food for weaning babies. As a result of the preparation method, significant (40%) losses of protein occur but the digestibility of the remaining protein is improved by 20% (42). In Nigeria, industrialization of ogi manufacture has taken place (54), enabling better control of quality and hygiene. Based on upgraded village technology, the final product is packaged and distributed as a long-shelf-life dehydrated powder, obtained by rotary drying or spray drying of the fermented wet cake. The nutritional value of ogi can be improved by enrichment with soybeans to obtain a 15% protein content (54).

### Pozol

Pozol, which dates back to the Aztec period, is made from nixtamal, which consists of corn kernels that have been boiled in lime water containing about 10% calcium hydroxide. This treatment, which probably evolved from the use of naturally occurring alkaline water of volcanic origin, facilitates the swelling of the corn and removal of pericarps (decortication). The resulting cooked endosperms are washed, drained, and milled to obtain masa, a coarse paste which is molded into balls (51), wrapped in banana leaves, and left to ferment for 1 to 2 weeks at ambient temperature (22 to 27°C). During the process, the pH increases to about 7.5 after nixtamalization and then gradually decreases to 3.8 to 4.0 after 1 week due to fermentative acidification, dominated by LAB. *Streptococcus* spp. account for 25 to 50% of the microflora, and *Lb. plantarum* and *Lb. fermentum*, together with *Leuconostoc* and *Weissella* species, are the other dominant microorganisms (4). Yeasts, including *Candida* spp. and *Trichosporon cutaneum*, are encountered in combination with the LAB (51). When left to ferment for longer periods, yeasts and molds (*Geotrichum candidum* and *Rhizopus* spp.) develop on the surface (51), imparting a musty flavor. In addition to the development of desired flavor, fermentation also contributes to the digestibility and increased riboflavin, niacin, and tryptophan contents of the product.

### Ang-Kak

Red kojic rice (ankak, or anka) is made by solid-state fermentation of cooked rice with the ascomycetous molds *Monascus* spp. Rice is washed, steamed for about 1 h, cooled to 36°C, inoculated with starter, heaped to ferment until the temperature rises to 42°C, and then spread and shelved. It is used in the fermentation industry for coloring red rice wine and foods such as sufu, fish sauce, and red soybean curd. The azaphilone pigments produced by *Monascus ruber*, *Monascus pilosus*, and *Monascus purpureus* include the orange rubropunctatin and monascorubrin, purple rubropunctamin and monascorubramin,

and yellow ankaflavin and monascin (59), which are heat stable over a wide range of pHs and thus of interest as bio-colorants in foods. Several other secondary metabolites are produced, including xanthomonasins, monascumic acid, monascusones, monacolins, and  $\gamma$ -aminobutyric acid. The pleasant flavor of ang-kak is derived from alcohols, aldehydes, ketones, esters, and terpenoid compounds.

Of recent interest are the potential health-promoting effects of ang-kak, such as cholesterol-lowering ability due to mevonolin (monacolin K), hypotensive effects due to  $\gamma$ -aminobutyric acid, and anti-inflammatory effects (3). The optimum cultural conditions for the production of pigments by a *Monascus* sp. isolated from the solid koji of Kaoliang liquor are reported to be pH 6.0 for a 3-day incubation at 32°C. Among the carbon sources tested, starch, maltose, and galactose are suitable for pigment production; a starch content of 3.5% (5% rice powder) and a sodium or potassium nitrate content of 0.5% gave maximum yield of pigment in laboratory media. Zinc may act as a growth inhibitor of *Monascus purpureus* and concomitantly as a stimulant for glucose uptake and the synthesis of secondary metabolites such as pigments.

### Ragi

The Indonesian word ragi refers to a starter or inoculum, and the name following ragi indicates the intended use of the starter, e.g., ragi-tempe, ragi-tapé, ragi-peuyeum, and ragi-tapai. Similar starters are Indian bakhar, Nepalese murcha, Thai loog-pang, Vietnamese men, Philippine bubod, Malaysian jui-paing, Chinese chu, Japanese tane koji, and Korean nuruk. To prepare ragi, rice flour is mixed with a variety of herbs, spices, and water to make dough which is inoculated by dusting with powdered ragi from a previous batch, flattened into cakes (about 3 cm in diameter and 1 cm thick), placed on a bamboo tray, covered with leaves or a cloth, incubated for 2 to 5 days at ambient temperature (20 to 30°C), air or sun dried, and preserved until needed.

A widely used type of ragi and ragi-like starters combines three groups of microorganisms, mucoraceous fungi, yeasts, and LAB (45). Ragi contains molds, namely *Amylomyces rouxii* and *Aspergillus*, *Mucor*, and *Rhizopus* spp. *Amylomyces* reproduces through thick-walled chlamydospores which ensure survival when the starter cakes are dried and stored prior to being used. Among the diverse yeast species in ragi, *Saccharomycopsis fibuligera* and *Pichia anomala* are the principal amyolytic and ethanol-producing yeasts, respectively. *Pediococcus pentosaceus*, *Weissella* spp., *Lactobacillus curvatus*, and *Enterococcus faecium* form the LAB component of ragi microflora (70). The microflora of ragi varies with the location and additives used. The molds and several

yeasts convert starchy materials into fermentable sugars, which are subsequently converted into ethanol by the yeasts and organic acids by the LAB and molds.

### Puto

To make puto, rice grains are soaked, ground to a semi-paste consistency (called galapong), mixed with starter (called lebadura), and fermented at two different stages, during which time the volume and lactic acid content increase 3- and 20-fold, respectively. The fermented batter is poured into molds and steam cooked for 15 to 30 min to make puto. The predominant microorganism in the fermenting batter is always *Leuconostoc mesenteroides* (30), followed by *Enterococcus faecalis* and then *S. cerevisiae* and *Pediococcus dextrinicus*. The yeasts produce low levels of ethanol and, along with *L. mesenteroides*, leaven the batter, rendering a spongy texture to the product.

### Rice Beers

Although the term “rice wine” is also in use for a rice-fermented alcoholic drink, the term “rice beer” is technically correct because, like beer, rice beer is produced from grain rather than fruit and it undergoes a two-stage fermentation process wherein starches in the rice are broken down into sugars which are then converted into alcohol. While most of these processes still follow indigenous technology, significant development in the manufacturing process has been made in Japan, China, and Korea; modern Japanese saké manufacture is highly

sophisticated. The manufacture of rice beers can be characterized as a biotechnological process which includes steaming, inoculation with starter, mashing, and fermentation. The microorganisms involved in the fermentations of some rice beers are listed in Table 38.2. Depending on the fermentation performance, the ethanol content varies and can reach up to 15% (vol/vol) (14, 15).

### Injera

To prepare injera, teff flour and water are combined with irsho, a fermented yellow fluid saved from a previous batch. The resultant thin, watery paste is generally incubated for 1 to 3 days. A portion of the fermented paste is then mixed with 3 parts water and boiled to give a product called absit, which is in turn mixed with a portion of the original fermented flour to yield a thin injera. Thick injera (aflegna) is a teff paste that has undergone only minimal fermentation (12 to 24 h) and is characterized by a sweet flavor and a reddish color. A third type of injera (komtata) is made from overfermented paste and, consequently, has a sour taste, probably due to extensive growth of LAB (17). Although the microflora compositions responsible for fermentation of the sweeter types of injera have not been fully determined, *Candida guilliermondii* is apparently a primary yeast in this process. The carbon sources for fermentation originate from the grain. Initially, a rise in free sugars, mainly sucrose, takes place, followed by a decline due to microbial assimilation (72). Regardless of the method used to prepare injera,

Table 38.2 Microorganisms involved in the production of Asian rice beers

Beer(s)	Starter	Functional microorganisms
Brem	Ragi-tapé	<i>Amylomyces</i> spp., <i>Mucor</i> spp., <i>Rhizopus</i> spp., <i>Saccharomyces cerevisiae</i> , <i>Candida glabrata</i> , <i>Pichia anomala</i> , <i>Issatchenkia orientalis</i>
Ruou nep than	Men	<i>Amylomyces rouxii</i> , <i>Rhizopus</i> spp., <i>Saccharomyces cerevisiae</i> , <i>Pichia anomala</i>
Saké, Mirin	Tane-koji	<i>Aspergillus oryzae</i> , <i>Saccharomyces sake</i> , <i>Pichia anomala</i> , <i>Lactobacillus sakei</i>
Sato, Ou, Nam-Khao	Loog-pang	<i>Mucor</i> spp., <i>Rhizopus</i> spp., <i>Candida</i> spp., <i>Saccharomyces</i> spp.
Shaoshing	Chu	<i>Aspergillus oryzae</i> , <i>Rhizopus</i> spp., <i>Saccharomyces cerevisiae</i>
Takju, Yakju	Nuruk	<i>Aspergillus</i> spp., <i>Mucor</i> spp., <i>Rhizopus</i> spp., <i>Saccharomyces cerevisiae</i> , <i>Pichia anomala</i> , <i>Hansenula subpelliculosa</i> , <i>Candida</i> spp., <i>Debaryomyces polymorphus</i> , <i>Lactobacillus plantarum</i> , <i>Leuconostoc mesenteroides</i>
Tapai	Jui-piang	<i>Amylomyces rouxii</i> , <i>Rhizopus oryzae</i> , <i>Mucor</i> spp., <i>Saccharomycopsis fibuligera</i> , <i>Pichia anomala</i>
Tapuy	Bubod	<i>Rhizopus oryzae</i> , <i>Amylomyces rouxii</i> , <i>Saccharomycopsis fibuligera</i> , <i>Rhodotorula glutinis</i> , <i>Debaryomyces hansenii</i> , <i>Candida</i> spp., <i>Lactobacillus plantarum</i> , <i>Leuconostoc</i> spp.

the fermented dough with batter consistency is baked on a hot, oiled clay griddle for a few minutes, resulting in a large, pancake-like bread injera.

### Gari

Fermented root of the cassava plant (*Manihot esculenta*) is known as gari in the rain forest belt of West Africa. To prepare gari, the corky outer peel and the thick cortex are removed and the inner portion of the root is grated. The pulp is then packed into jute bags, and weights are applied to express some of the juice. After 3 to 4 days of fermentation, cassava is sieved and heated while constantly turning over a hot steel pan. This process is known as garification. The final product contains 10 to 15% moisture, 80 to 85% starch, 0.1% fat, 1 to 1.5% crude protein, and 1.5 to 2.5% crude fiber. Palm oil may be added as a colorant just before or after drying. For the production of 1 ton of gari, 4 tons of cassava roots are required. Cassava is a highly perishable crop once harvested; garification is a clever approach to achieve a safe, shelf-stable product. In Nigeria, gari production has been industrialized (55).

Fresh cassava roots of bitter varieties contain cyanogenic glycosides, viz., linamarin and lotaustralin, that decompose during the fermentation of gari with the liberation of gaseous hydrocyanic acid. The hydrolysis of cyanogenic glycosides is due mainly to endogenous linamarinase, reducing cyanide levels from 300 mg (initial) to 10 to 20 mg of HCN/kg of product (55). *Lb. plantarum* and other LAB contribute significantly to decreasing the pH (74). The acid condition favors fungal growth, mainly that of *Galactomyces candidum*, which contributes to the characteristic aroma and flavor by its aldehydes and esters.

### Tapé

Tapé ketan (rather similar to lao-chao) and tapé ketella are prepared by fermenting rice and cassava, respectively. Glutinous rice or peeled and chopped cassava root is soaked, steam cooked until soft, spread in thin layers onto bamboo trays, inoculated with powdered ragi-tapé (starter), and left to ferment under cover for 1 to 3 days at 27 to 30°C to produce a soft, white mass (65). The essential biochemical changes, caused by *Amylomyces rouxii* and *Pichia burtonii*, are the hydrolysis of starch into maltose and glucose and the conversion of a part of the sugars into alcohol and organic acids, thereby imparting a sweet-sour taste to the product.

### Poi

To prepare poi, the corm of the taro plant (*Colocasia esculenta*) is cooked for several hours by baking or steaming, peeled, ground, combined with water to make a smooth,

sticky paste, and stored airtight for 2 to 3 days at ambient temperature (8). *Lactobacillus delbrueckii* and *Lactococcus lactis* predominate in the early stages of fermentation, lowering the pH to 3.8 to 4.0. These bacteria, along with *Saccharomyces kefir*, produce a large amount of lactic acid and moderate amounts of acetic, propionic, succinic, and formic acids. *Candida vini* and *Geotrichum candidum*, which are prevalent in the later stages of fermentation, impart a pleasant fruity aroma to mature poi.

## LEGUME PRODUCTS

### Daddawa

Daddawa (dawadawa) preparation is still largely a family art practiced at home. Seeds of the African locust bean tree (*Parkia biglobosa*) are washed to remove yellow powdery pulp, leaving black beans which are then boiled in water in a covered container for 18 to 24 h, with occasional replenishing of water to swell the seeds and soften the very tough seed coats, which are then removed by pounding and rinsing. The cotyledons are reboiled for 30 min to 2 h when a native softening agent called kuru (containing mostly potash) is added. The cotyledons are drained, heaped (10 to 15 cm deep) in calabash trays or dumped in a hole in the ground, and covered with locally available leaves and sackcloth. Beans are left to ferment for 2 to 4 days at 25 to 35°C, during which time they become dark brown and covered in a sticky mucilaginous layer and develop a pungent odor. The bean mass is air dried in the sun or hot shade, where the beans darken further, and the beans are then used loose or shaped into balls or pyramids (9, 63).

Although daddawa is dominated and produced mainly by *Bacillus subtilis*, it contains several other species of *Bacillus* and *Leuconostoc* (52, 63). During fermentation, the temperature of 25°C and the pH of 7.0 of the beans increase to 45°C and pH 8.1 at 36 h. The content of free amino acids increases fivefold due to extensive proteolysis. Certain antinutritional factors such as oligosaccharides, phytic acid, and oxalate decrease during fermentation (52).

### Kinema

Kinema resembles natto except that, although in natto intact whole soybeans are used, in kinema the beans are crushed to form grits about half the size of cotyledons. Kinema is a naturally fermented product, containing *Bacillus* spp., enterococci, and yeasts. However, *B. subtilis* is the principal bacterium in the microflora and is largely responsible for the production of kinema. Spores of *B. subtilis*, which are normally present on soybeans, survive the cooking treatment to initiate and carry out

the fermentation. Strong proteolytic activity causes an increase in pH from an initial pH of 6.9 to a pH of 8.6 at the end of fermentation (62, 63).

### Meitauza

Meitauza is prepared from okara (insoluble carbohydrate residue that is left over after the production of soy milk or tofu) which is ground, steeped, strained, and formed into round cakes 10 to 14 cm in diameter and 2 to 3 cm thick at the middle and 1 to 1.3 cm thick at the edges. The cakes are placed in a vessel and left to ferment with moderate aeration until, after 10 to 15 days, they are covered with white mycelium of *Actinomucor elegans*. The molded cakes are then sun dried. Meitauza is served either fried in vegetable oil or cooked with vegetables as a flavoring agent (56).

### Natto

Natto, a popular breakfast and dinner item in Japan, is usually eaten with rice along with soy sauce and spicy mustard. It is the only food in the category of alkaline fermentations that has been industrialized (62). With the use of whole soybeans, three types of natto are prepared. Yukiwari-natto and hama-natto are koji (*Aspergillus oryzae*)-based products, while the more common itohiki-natto is a *Bacillus*-fermented product. Itohiki-natto, generally referred to as natto, is popular in the eastern Kanto region (Tokyo). Natto soybeans are small (up to 5.5 mm in diameter) with a clear hilum, thin seed coat, and high carbohydrate content. Smaller beans are preferred, as the fermentation process reaches the center of the beans easier. To prepare natto traditionally, soybeans are washed, soaked overnight, boiled until tender (approximately 15 min), drained, partially air dried for 20 min over bamboo trays, and put into shallow paper containers covered with wax paper. The containers are stacked in large wooden boxes, covered with straw mats, and left near an oven at approximately 36°C to ferment for 1 day. Intentional inoculation is not necessary because straw contains the fermenting microorganism, *B. subtilis*. However, not all strains of *B. subtilis* are suitable for making good natto (53).

Yukiwari-natto is made by mixing itohiki-natto with rice koji and salt and then aging at 25 to 30°C for about 2 weeks. To prepare hama-natto, washed soybeans are soaked in water for 4 h and steamed for 1 h. After cooling, the beans are inoculated with koji, fermented for about 20 h, dried to a moisture content of 12%, submerged in brine, and aged for 6 to 12 months.

Natto is prized for its high nutritional value and improved digestibility, both resulting from fermentation. The nature of the free amino acid profile of natto is similar

to that of kinema (44). Natto has a characteristic pungent but pleasant aroma. Sulfur-containing compounds deriving from the cooked soybeans and pyrazines formed during fermentation are the main contributors to the characteristic natto odor. The sulfur compounds include 4-ethyl-2-methylthiazole, 3,5-dimethyl-1,2,4-trithiolane, and thialdine. The pyrazines present at the highest concentrations include tetramethyl, trimethyl, and 2,5-dimethyl derivatives (57). Natto is also characterized by the presence of a sticky paste on its surface. When stirred, the paste increases in volume and becomes stickier and is held together like a spider web by gossamer-like threads. Natto mucin contains 22% fructan and 78% poly-DL-glutamic acid with a  $\gamma$ -peptide linkage ( $\gamma$ -PGA) which has a high viscoelasticity and is spinnable due to the formation of network structures of randomly coiled  $\gamma$ -PGA through intermolecular H bondings in the presence of fructan (25). Production of  $\gamma$ -PGA in the natto strains of *B. subtilis* is regulated by the *comQXPA* quorum-sensing system and is genetically unstable because of the translocation of *IS4Bsu1* into the *comP* gene at a high frequency (41). The *IS4Bsu1* is widely distributed among *B. subtilis* strains in other similar soybean-fermented foods, such as kinema, Thai thua-nao, Chinese douchi, Korean chungkuk-jang, and Burmese chine pepoke (24). Natto mucin can absorb 5,000 times its weight in water, and this remarkable property has been put to use in cosmetics and wrappings of food products.

### Oncom

Oncom is a by-product of peanut oil pressing, produced by soaking peanut (*Arachis hypogaea*) press cake for 1 day, mixing with starchy ingredients such as cassava residues, steaming for about 1 h, cooling, and inoculating with pregrown fungal mycelium, usually *Neurospora sitophila* or *Neurospora intermedia*. The inoculated dough is molded to form brick-shaped pieces that are incubated for a few days in banana leaves at ambient temperature (25 to 30°C) (31). Oncom hitam (black oncom) and oncom merah (yellow-red oncom) contain different mycofloras. The merah type contains mainly *Neurospora*, whereas hitam is dominated by *Rhizopus* spp. *Rhizopus* sporangiospores are black due to melanoids, and carotenoids form the basis of the orange-yellow color of *Neurospora*.

Oncom flavor has been described as fruity and somewhat alcoholic; after frying, mincemeat or almond flavors can be detected. The enzymatic activities (lipases, proteases) provoke an increase in free fatty acids and degradation of proteins, resulting in improved protein digestibility which is relevant for consumers with digestive disorders. Recently, experiments with oncom-miso made

from soybeans and oncom demonstrated increased anti-oxidative and antimutagenic activity associated with the enzymatic release of isoflavone-aglycones (37). Instead of dry, spore-based starters, starters used for oncom are propagated and maintained by mycelial growth in a kind of fed-batch solid-state fermentation kept active by the processors. Although very little controlled experimentation has been done on this fermentation, it is presumed that a method of vegetative propagation is needed because the *Neurospora* spores have limited viability when stored dry and have poor germination ability.

### Sufu

*Actinomucor elegans* (7, 21) and *Actinomucor taiwanensis* (11) are used as pure-culture starters in the manufacture of Chinese fu-ru, or sufu. The process of preparing sufu starts with the production of soy milk by soaking dehulled soybeans, grinding, sieving, and cooking the watery extract, the latter to inactivate trypsin inhibitors and reduce some of the undesirable beany flavor. Next, a coagulation step is carried out, by adding salts (calcium or magnesium sulfate) or acid, in order to obtain a precipitate of mainly soy protein and entrapped lipids. This precipitate is collected and pressed to obtain sheets of tofu (soybean curd) of the required moisture content and firmness. After cutting of the tofu into cubes (dices), the tofu is inoculated with a suspension of mold spores. Incubation for a few days usually results in luxuriant mycelial development, giving the dices a fluffy appearance. These are now called pehtze, containing about 74% water, 12% protein, and 4.3% lipid. After flattening of the mycelium to form a protective skin on the cubes, they are submerged in a maturation mix and stored for several months to develop into a flavorsome, soft, cheese-like product. The main functions of the maturation mix are preservation, flavoring, and coloring. Preservation of sufu is achieved by a combination of salt and alcohol (rice beer may be used), whereas ang-kak and other ingredients impart specific flavor and color to the product (21). Depending upon the desired flavor and color, pehtzes may be submerged in salted, fermented rice or soybean mash, fermented soybean paste, or a solution containing 5 to 12% sodium chloride, red rice, and 10% ethanol. Red rice and soybean mash impart a red color to sufu. Use of brine containing high levels of ethanol results in sufu with a marked alcoholic bouquet. The major functions of the molds in this process are the formation of a protective layer of mycelial biomass surrounding the pehtze cubes and, most importantly, the release of several enzymes that are responsible for the partial degradation of the protein, fiber, and lipid fractions in pehtze during the maturation. This degradation results in softening of

the texture, solubilization of constituents, and accumulation of flavor-enhancing compounds, such as glycine and glutamic acid (20, 36). In view of the optimization of industrial sufu-making processes, the response of *Actinomucor elegans* to temperature, salt, and alcohol has been studied. The higher the salt and alcohol levels during the maturation, the slower the enzymatic reactions take place, thus requiring longer maturation times. With the objective of accelerating the maturation, the salt and alcohol levels could be lowered. This is feasible to a level of about 10% alcohol in combination with 6% salt; at lower levels the product is susceptible to spoilage by LAB, however, as well as survival of pathogens and enterotoxin formation by *Staphylococcus aureus*.

### Tempeh

Tempeh (the Indonesian spelling is “tempe”) is made from cooked seeds (those of soybeans, cereals, or others) or seed-processing by-products by solid-state fungal fermentation (47, 48). Tempeh is an attractive nonmeat protein food that can be used as an ingredient in a large variety of traditional Indonesian dishes as well as in Western-style spreads, snacks, and burgers.

Soybeans are soaked in water at ambient temperature overnight or until hulls (testae) can be easily removed by hand. LAB and yeasts predominant in water in which soybeans have been soaked are *Lactobacillus casei*, *Lactococcus* spp., *Pichia burtonii*, *Candida diddensiae*, and *Rhodotorula mucilaginosa* (40). Fermentative acidification during the soaking stage has been shown to suppress the growth of spoilage and pathogenic bacteria (48).

After removal of the hulls from the soaked soybeans, cotyledons are cooked for 30 to 60 min, drained, and cooled. In the traditional tempeh process, simple methods are employed for the inoculation of the cooked beans. In principle, it is possible to use some previously made tempeh as inoculum (32); however, as tempeh contains a considerable population of bacteria other than those desired for fermentation, the reuse of tempeh as an inoculum incurs the risk of fermentation failure due to bacterial overgrowth. Therefore, professional tempeh manufacturers use traditional mold spore concentrates. These are, e.g., harvested from cooked rice on which selected strains of *Rhizopus oligosporus* have been cultured or cooked soybeans that have been held between leaves of *Hibiscus tiliaceus* (the waru tree). The latter type of widely used starter, made by specialized households, is available in the public markets in Indonesia.

The inoculated beans are then spread onto bamboo frames, wrapped in a punctured plastic sheet or between banana leaves, and allowed to ferment at ambient temperature (25 to 30°C) for 1 to 2 days. At this point, the

soybeans are covered with white *Rhizopus oligosporus* mycelium and bound together as a cake (32). Aerobic mold growth requires oxygen and produces heat and carbon dioxide. Care should be taken that the beans do not dehydrate and that no overheating ( $>40^{\circ}\text{C}$ ) occurs. This is achieved by allowing only restricted access of air to the beans and by limiting the thickness of the bean layers or packages (47).

Several factors may limit the acceptability and shelf life. These include the production of black sporangia and spores, indicating inadequate fermentation conditions, which results in an undesirable gray color, and enzymatic browning, comparable to the browning of cut apples, which is initiated by prolonged storage or mechanical abuse. Whereas freshly fermented tempeh has an attractive mushroom-like flavor, prolonged storage may lead to yeasty off-odors or ammoniacal odors resulting from excessive protein and amino acid degradation.

During fermentation, carbohydrases, lipases, proteases, phytases, and other enzymes degrade macromolecular substrates, resulting in very significant increases in water-soluble nutrients for enhanced digestion, biosynthesis of B vitamins such as folate (18), and transformation of soy isoflavones into health-promoting antioxidant compounds (47).

### Wadi and Papad

Wadi (wari) is prepared by soaking dal (dehusked split beans), generally of black gram (*Phaseolus mungo*), in water for 6 to 12 h, draining, grinding into a smooth soft dough, and fermenting for 1 to 3 days at 20 to 27°C, with or without spices but with salt and backslop added. In an alternative method, the dough is combined with shredded waxgourd (*Benincasa hispida*) and whisked vigorously until it becomes light and fluffy due to the incorporation of air. The fermented or whisked dough is hand-molded into cones or balls (3 to 8 cm in diameter), deposited onto bamboo or palm mats smeared with oil, and sun dried for 4 to 8 days (6). The surface of the cones or balls becomes covered with a mucilaginous coating which retains the gas formed during fermentation within them. The wadis look hollow with many air pockets and yeast spherules in the interior and have a characteristic surface crust.

Initially the microflora includes LAB, *Bacillus* spp., flavobacteria, and yeasts. Gradually, a domination by gas-producing *L. mesenteroides*, *Lb. fermentum*, *S. cerevisiae*, and *Trichosporon cutaneum* is achieved. *Candida vartiovaarae* and *Kluyveromyces marxianus* are also often found. Summer is more favorable for the prevalence of bacteria, and winter is more favorable for the

yeasts (67). The production of acid and gas results in a decrease of pH from 5.6 to 3.2, an increase in total acid (as lactic acid) from 0.5 to 1.5%, and a twofold increase in the volume of the dough. The LAB are mainly responsible for acidification of dough, a condition which favors the growth of yeasts and leavening. Fermentation brings about a significant increase in soluble solids, non-protein nitrogen, soluble nitrogen, free amino acids, and B vitamins. Most of these changes cause improvement in digestibility and nutritional value. Increase in total acidity during fermentation helps to enhance the shelf life of the product (66).

Papad (papadam, or appalam) is a thin, usually circular, wafer-like product used to prepare curry or eaten as a crackly snack or appetizer with meals after roasting or deep frying. In the indigenous method of preparation, black gram flour alone or blended with Bengal gram (*Cicer arietinum*), lentil (*Lens culinaris*), red gram, or green gram (*Phaseolus aureus*) flour is hand-kneaded with a small quantity of peanut oil, common salt (about 8%), papad khar (a natural additive), and water and then beaten or pounded into a stiff paste. The paste may be seasoned with spices. The dough (sometimes with backslop added) is left to ferment for 1 to 6 h and then formed into long cylinders and cut and shaped into small balls which are rolled into thin, circular flat sheets (10 to 24 cm in diameter, 0.2 to 1.2 mm thick) by using a wooden rolling pin and generally dried under shade to 12 to 17% moisture content (6, 64). *C. krusei* and *S. cerevisiae* are involved in fermenting the dough, presumably resulting in modest leavening.

## CEREAL-LEGUME MIXTURE PRODUCTS

### Idli

Idli is a classical example of cereal-legume mixture food that provides an improved balance of carbohydrates and proteins. Because of its appealing sour flavor, spongy texture, nutritional quality, and easy digestibility, idli is also fed to infants as a complementary food and is used as a main dish in diets provided to patients in hospitals (49).

The substrates used in preparing idli are white polished rice and black gram dal (1:4 to 4:1), which are washed and soaked in water separately at ambient temperature for 5 to 10 h. While rice is coarsely ground, the dal is ground into a smooth, mucilaginous paste. The two slurries are combined generally in the ratio of 2:1 and stirred well with added salt (0.8%) to form a thick batter which is put in a closed container and left in a warm place (25 to 35°C) to ferment overnight or longer (14 to 24 h). The fermentation period must allow a definite leavening (two- to threefold increase in volume) of

the batter and development of a pleasant acid flavor. The fermented batter is poured into small cups (8 to 10 cm in diameter) and steamed in a covered pan to yield a soft, spongy product (39). The open texture is attributed to the protein (globulin) and polysaccharide (arabinogalactan) in black gram (71).

Although bacteria and yeasts that participate in the fermentation are generally introduced by the substrates, it is often the practice to add backslop to the newly ground substrates. With the progress of fermentation, both bacterial and yeast cell numbers increase significantly, with a concomitant decrease in pH and an increase in the volume of the batter and its amylase and protease activities. *L. mesenteroides* is the most commonly encountered bacterium, followed by *Lb. fermentum*, *Enterococcus faecalis*, and *Pediococcus dextrinicus* (39). During fermentation, along with *L. mesenteroides*, yeasts such as *S. cerevisiae*, *Debaryomyces hansenii*, *Pichia anomala*, and *Trichosporon pullulans* are predominant, and *Trichosporon cutaneum* develops subsequently. *S. cerevisiae* is the only yeast that eventually persists (68).

The major functions of the fermentation of idli include the leavening of batter and improvement of flavor and nutritional value. The role of LAB is to reduce the pH of the batter from an initial 6.0 to an optimum level (4.1 to 4.5) for yeast activity. The LAB may also play a role in the breakdown of phytate present in black gram. *L. mesenteroides* isolated from soy idli secretes  $\beta$ -N-acetylglucosaminidase and  $\alpha$ -D-mannosidase, which are involved in the hydrolysis of hemagglutinin. Yeasts help in the degradation of starch, a process that cannot be carried out by *L. mesenteroides*, into maltose and glucose by producing extracellular amylolytic enzymes. Yeasts also produce carbon dioxide and play a significant role in leavening. Fermentation of batter by inoculating the ingredients with yeasts individually and in combination with *L. mesenteroides* has revealed that yeasts contribute not only to gas production, resulting in good texture, but also to sensory qualities. The higher activity of amylases and levels of B vitamins and free amino acids attained in yeast-enriched fermentations represent positive contributions of yeasts (73).

### Miso

Fermented soybean pastes are known as miso in Japan, chiang in China, jang or doenjang in Korea, taoco in Indonesia, and tao chieo in Thailand. In addition to soybeans and salt, most of these products contain cereals such as rice or barley (38). Miso is fermented using *Aspergillus oryzae* and a yeast, *Zygosaccharomyces rouxii*. Sometimes, *Tetragenococcus halophila* and *Enterococcus faecalis* are also involved in the fermentation. Heat-treated rice and/

or soybeans are used to prepare shinshu or rice-soybean miso. After the initial solid-state fermentation dominated by *Aspergillus oryzae*, salt (38% of the original weight of dry soybeans) is added to the koji and mixed thoroughly. The mixture is inoculated with *Z. rouxii*; traditionally, sound miso from a previous batch is used to inoculate the koji-soybean-salt mixture prior to fermentation. Although halophilic yeasts such as *Torulopsis versatilis* may be present, only *Z. rouxii* produces the desired metabolites for an acceptable product. The mixture, known as green miso, is packed into vats or tanks to undergo anaerobic fermentation and aging at 25 to 30°C. White miso takes about 1 week, salty miso takes 1 to 3 months, and soybean miso takes over 1 year. White miso contains 4 to 8% salt, which permits rapid fermentation, and yellow or brown misos contain 11 to 13% salt. Moisture content ranges from 44 to 52%, protein content ranges from 8 to 19%, carbohydrate content ranges from 6 to 30%, and fat content ranges from 2 to 10%, depending on the ratio of soybeans, rice, and barley used as ingredients.

During fermentation and aging, soybean protein is digested by proteases produced by *Aspergillus oryzae* in the koji. Amino acids and their salts, particularly sodium glutamate, contribute to flavor. The addition of commercial enzyme preparations to enhance fermentation has met with some success. The relative amount of carbohydrates in miso is a reflection of the amount of rice in the product. Starch is extensively saccharified by koji amylases to yield glucose and maltose, some of which is utilized as a source of energy by the microorganisms responsible for fermentation. Miso contains 0.6 to 1.5% acids, mainly lactic, succinic, and acetic. Esters of ethyl and higher alcohols with fatty acids in soybean lipid are important in giving miso its characteristic aroma. Up to 35% of the initial lipid content is degraded into fatty acids; the extent of maturation can be conveniently monitored by the levels of fatty acid ethyl esters (75). Furanones HEMF [4-hydroxy-2(or 5)-ethyl-5(or 2)-methyl-3(2H)-furanone] and HDMF [4-hydroxy-2, 5-dimethyl-3(2H)-furanone] produced by *Z. rouxii* have been identified as important flavor components in miso. Miso also contains B vitamins (riboflavin and cyanocobalamin) as a result of yeast fermentation. Miso is considered to be a health-promoting, functional food, offering protection against gastrointestinal disorders; cancers of the breast, stomach, and colon; and cholesterol-associated and degenerative diseases (38).

### Soy Sauces

Soy sauces are light to dark brown liquids with meat-like salty flavor used in cooking and as a table condiment. Traditionally made in the Orient, they are now also

produced in Europe and the Americas. In Japan, there are several distinguished types, including Koikuchi, Usukuchi, Tamari, Saishikomi, and Shiro soy sauces, all having characteristic colors and flavors. All contain relatively high levels (17 to 19%) of salt and are used to enhance the flavors of meats, seafoods, and vegetables. Typical ranges in other characteristics are as follows: pH 4.6 to 4.8 and 0.5 to 2.5 g of total nitrogen, 0.2 to 1.1 g of formol nitrogen, 3.8 to 2.0 g of reducing sugar, and traces to 2.2 ml of ethanol (per 100 ml).

There are two specific fermentation stages involved in soy sauce production, the first being an aerobic koji fermentation. Seed (tane) koji is produced by culturing single or mixed strains of *Aspergillus oryzae* or *Aspergillus sojae* on either steamed, polished rice or a mixture of wheat bran and soybean flour. Seed koji is added to a soybean-wheat mixture at a concentration of 0.1 to 0.2% and fermented into what is then simply called koji. The second stage is an anaerobic moromi or salt mash which undergoes LAB and yeast (*Z. rouxii*) fermentations.

The two main groups of enzymes produced by *Aspergillus oryzae* during koji fermentation are carbohydrases ( $\alpha$ -amylases, amyloglucosidase, maltase, sucrase, pectinase,  $\beta$ -galactosidase, cellulase, hemicellulase, and pentosan-degrading enzymes) and proteinases, although lipase activity has also been reported. These enzymes hydrolyze carbohydrates to sugars and proteins to produce amino acids and low-molecular-weight peptides. These soluble products are essential for yeast and bacterial activities during the moromi fermentation (2, 12). In the moromi

fermentation, *Tetragenococcus halophila* initially proliferates and produces lactic acid, which lowers the pH to 5.5 or less. This is followed by the growth of acid-tolerant yeasts, notably *Z. rouxii*, which produce about 3% alcohol and several compounds that contribute to the characteristic aroma of soy sauce.

Although *Z. rouxii* is the dominant moromi yeast, other yeasts such as *Candida versatilis* and *Candida etchellsii* also produce phenolic compounds which contribute to soy sauce aroma. Nearly 300 flavor compounds have been identified in Japanese soy sauce (50); major categories are summarized in Table 38.3. *Z. rouxii* produces flavor compounds, including alcohols, glycerol, esters, and furanones. Of the latter, HEMF produced by *Z. rouxii* and *Candida* spp. gives Japanese-type soy sauce its characteristic flavor (22). This compound is also reported to have antitumor and antioxidative properties (33). Notwithstanding their high salt contents, soy sauces require pasteurization and adequate bottling for preservation.

## VEGETABLE PRODUCTS

### Kimchi

Kimchi is a generic term used to denote a group of fermented vegetable foods in Korea. More specific names are used for these products, depending on the substrate, processing method, season, and locality. Each family has its own recipe handed down from generation to generation. T'ong baegu'u-kimchi, the most common kimchi, is

Table 38.3 Major flavor components in soy sauce

Type	Component(s)
Produced by <i>Zygosaccharomyces rouxii</i>	
Alcohols . . . . .	Benzyl alcohol, 1-butanol, ethanol, glycerol
Esters . . . . .	Bornyl acetate, ethyl acetate, ethyl lactate, ethyl myristate
Furanones . . . . .	4-Hydroxy-5-methyl-3(3 <i>H</i> )-furanone (HMMF), 4-hydroxy-2(or 5)-ethyl-5(or 2)-methyl-3(2 <i>H</i> )-furanone (HEMF)
Produced by <i>Candida versatilis</i> and <i>Candida etchellsii</i>	
Phenolic compounds . . . . .	2-Methoxy-4-ethylphenol, 2-methoxyphenol
Higher alcohols . . . . .	Isobutyl alcohol, isoamylalcohol, 2-phenylethanol
Miscellaneous	
Aldehydes . . . . .	Acetaldehyde, benzaldehyde, 2-methylpropanal
Volatile carboxylic acids . . . . .	Acetic acid, butanoic acid, 3-methylbutanoic acid
Ketones . . . . .	2-Hexanone, 3-hydroxyl-2-butanone
Furans . . . . .	2-Acetyl furan
Pyrroles . . . . .	2-Acetylpyrrole
Pyrazines . . . . .	2,3-Dimethylpyrazine, 3-ethyl-2,5-dimethylpyrazine, 2-methylpyrazine

made by cutting Chinese-type cabbage into two or four parts and adding salt (approximately 3%), followed by rinsing; draining; packing in layers with premixed garlic, onion, ginger, hot pepper powder, and salt; and fermenting for 2 to 3 days at around 20°C. The flavor of kimchi is dependent on the ingredients, fermentation conditions, and LAB involved in the fermentation (34). There is a succession of microfloras during kimchi production. *L. mesenteroides* is a major bacterium found in the initial to the middle stage of fermentation. *Weissella confusa* and *Leuconostoc citreum* present in the raw materials also initiate and remain present throughout the fermentation period, indicating their importance in kimchi production. *Lb. brevis* and *Lb. plantarum*, which appear at the middle stage of fermentation, are overtaken by *Lactobacillus sakei* and *Lb. curvatus*, which dominate the late stage of fermentation when the pH decreases to 3.9 (35). They produce various constituents such as lactic acid, acetic acid, ethanol, carbon dioxide, mannitol, and dextran that impart a good flavor to kimchi. Kimchi closely resembles sauerkraut. The optimal range in salt concentration in sauerkraut is 0.7 to 3.0%, and that in kimchi is 3.0 to 5.0%, and secondly, kimchi is less sour than sauerkraut but carbonated.

### FISH PRODUCTS

Basically salt-fermented fish products are popular in the Orient (34). Where large quantities (>20%) of salt are added, fermentation by microorganisms is largely suppressed and enzymes from fish flesh and gut cause proteolysis of the fish. To allow fermentation to gain more importance, a source of carbohydrate is added so that LAB prevail and ferment the carbohydrate into organic acids, which reduces the high buffering capacity of the fish to result in a rapid decrease in pH, aiding preservation and production of a tangy odor (58). To prepare low-salt fermented products like Korean sik-hae (side dish), whole fish is degutted; stripped; mixed with salt (6 to 7%); cured overnight; drained; mixed with cooked millet (7 to 8%), minced garlic (3 to 4%), and red pepper powder (8 to 9%); and packed in an earthen jar for fermentation at 20°C for 1 week. The jar is then kept in a cool place for 3 to 6 weeks to develop optimum flavor. The pH decreases rapidly during the first 3 to 5 days from 6.5 to <5.0, and softening begins after 3 days of fermentation. The amino-N concentration increases steadily up to 14 days, and this increase coincides with the attainment of optimum flavor. The important fermenting organisms are *L. mesenteroides* and *Lb. plantarum* (34). Garlic affects the microflora by inhibiting the growth of gram-negative bacteria, in particular, due to

its allicin content and by stimulating the growth of LAB due to the supply of fermentable fructose from its inulin reserve.

### MISCELLANEOUS PRODUCTS

#### Kombucha

For kombucha, black tea leaves are steeped in boiling water, combined with sugar (5 to 15%), cooled, inoculated, and left for 7 to 14 days, during which a film grows on the surface. When sufficiently thick (ca. 2.5 cm), the film falls to the bottom and further film develops on the surface. This biofilm is used as a starter for new batches; the liquid (pH of ca. 2.5) is claimed to benefit health (19). The cellulosic pellicle is a result of symbiotic associations of acetic acid bacteria, chiefly *Acetobacter aceti* subsp. *xylinus*, and various yeasts, e.g., *Brettanomyces*, *Zygosaccharomyces*, and *Saccharomyces* spp. Among them, *Zygosaccharomyces kombuchaensis* is the dominant species. Ethanol-producing yeasts are succeeded by acid-producing bacteria.

#### Palm Wines

The sap obtained from decapitated inflorescence of palm is fermented to produce wine. There is an art in binding the flower spathes, pounding to cause the sap to flow properly by cutting the spathe tip, and collecting the sap. The fermentation starts as soon as the sap flows into a container, and within a few hours it becomes reasonably high in alcohol content (up to 4%). The palm wine fermentation is always an alcoholic-lactic-acetic one, involving yeasts, LAB, and acetic acid bacteria. The microorganisms responsible are mainly *S. cerevisiae*, *Schizosaccharomyces pombe*, *Lb. plantarum*, and *L. mesenteroides*. The LAB are responsible for the consistency and soluble white coloration of palm wines through the production of glucans (29).

### NUTRITION AND HEALTH-PROMOTING ASPECTS

Many traditional fermented foods are staples in the diets of vast populations of people who would otherwise have less than minimum intakes of protein and/or calories. Although the quality or quantity of proteins in vegetable-based fermented foods generally is not dramatically increased over that in raw substrates, digestibility may be improved through fermentation. Antinutritional factors in plant materials, e.g., protease inhibitors and lectins in leguminous seeds and phytic acid in cereals and seeds, may actually be reduced by fermentation. Factors that inhibit digestive activity, or that form indigestible

complexes with proteins and minerals, are degraded during fermentation processes by endogenous (plant) and microbial enzymes. Naturally occurring toxic components, e.g., cyanogenic glycosides and glucosinolates, can be degraded substantially in fermented foods, rendering inedible materials into wholesome foods. Food components such as soybean isoflavones may be transformed into potent antioxidant aglycones, which have potential for reducing hypertension, cardiovascular ailments, and cancer, as a result of food fermentation. A significant synthesis of vitamins, especially B vitamins such as thiamine, niacin, folate, and riboflavin, contributes to the bioenrichment of foods (69).

The presence of large numbers of microorganisms (e.g., LAB) in fermented foods such as yogurt may have probiotic effects, i.e., supporting a healthy balance of the gut microbiota and offering enhanced colonization resistance and immunoresistance to the host (46). The positive contribution of traditional fermented foods to the nutritional well-being and health of those who consume them on a regular basis is widely recognized.

### FOOD SAFETY ASPECTS

Of interest to food microbiologists and sanitarians is the possibility of microorganisms' producing toxic substances or of pathogenic microorganisms' surviving during fermentation or storage of indigenous fermented foods. An investigation of the aflatoxin-forming ability of *Aspergilli* used in the Japanese food industry revealed that although fluorescent compounds were formed, none produced aflatoxin. Recently it was found that *Aspergillus oryzae* and *Aspergillus sojae*, typical "domesticated" industrial molds that have been used for centuries in the production of koji for soy sauce and miso, are incapable of forming aflatoxins. The *aflR* gene (aflatoxin pathway-specific regulatory gene) is impaired in its ability to activate transcription of aflatoxin biosynthetic genes and is unable to interact with *aflJ* (coactivator gene) (10). The inability of potentially toxigenic molds to form mycotoxins in food fermentations may be caused by altered gene expression, unfavorable food environment, or competition with other microorganisms.

A variety of pathogenic bacteria, viruses, and parasites may occur in raw ingredients or in fermented foods due to postprocessing contamination. Whereas many gram-negative bacteria cannot survive when exposed to acidic and/or elevated salt, alcohol, or sugar conditions, many other bacteria survive such stress conditions. Viruses and parasitic cysts can be eradicated only by adequate heat treatments. Safety of fermented foods must be evaluated and controlled on the basis of safety management

approaches, i.e., the hazard analysis and critical control point system (1) (see chapter 46 in this volume). Although traditional fermented foods have a history of safe use, adequate measures for hygienic processing and protection against contamination must be implemented.

### MODERN TOOLS IN RESEARCH AND DEVELOPMENT OF FERMENTED FOODS

The food industry is in continuous transition, adopting an innovative and market-oriented position (13). Within this trend, there are ample opportunities for genomics-based approaches to be applied in the fermented food industry. These include transcriptomics, proteomics, and other postgenomics approaches and are important new tools that may be applied to optimize processes and to gain a better insight into mechanisms. In view of the protection of origin (a.o.c., or certified origin of production), an unequivocal characterization of traditional fermented foods and their microflora will be required; this could be based on combinations of food compositional analysis and metabolomics (46).

In addition, these so-called "-omics" approaches offer opportunities to develop novel products and diversify processes. Innovative processes using nontraditional fermentation conditions, e.g., immobilized cells or agitated solid-state fermentors, or using pure-culture inoculation instead of multistrain natural fermentations, may invoke changes in secondary metabolite production. In view of maintaining the character of the food, as well as safeguarding safety for the consumer, the impact of novel processing should be investigated, understood, and possibly controlled. -Omics approaches will also offer opportunities to advance indigenous fermented food quality assurance and allow running cost to be reduced to more affordable levels. Finally, in the field of probiotics, -omics offer analytical approaches to substantiate health claims.

### References

1. Adams, M. R., and M. J. R. Nout (ed.). 2001. *Fermentation and Food Safety*. Aspen Publishers, Gaithersburg, Md.
2. Aidoo, K. E., J. E. Smith, and B. J. B. Wood. 1994. Industrial aspects of soy sauce fermentations using *Aspergillus*, p. 155–169. In K. A. Powell, A. Renwick, and J. F. Peberdy (ed.), *The Genus Aspergillus: from Taxonomy and Genetics to Industrial Application*. Plenum Press, New York, N.Y.
3. Akihisa, T., H. Tokuda, K. Yasukawa, M. Ukiya, A. Kiyota, N. Sakamoto, T. Suzuki, N. Tanabe, and H. Nishino. 2005. Azaphilones, furanoisophthalides, and amino acids from the extracts of *Monascus pilosus*-fermented rice (red-mold rice) and their chemopreventive effects. *J. Agric. Food Chem.* 53:562–565.

4. Ampe, F., N. Ben-Omar, C. Moizan, C. Wachter, and J. P. Guyot. 1999. Polyphasic study of the spatial distribution of microorganisms in Mexican pozol, a fermented maize dough, demonstrates the need for cultivation-independent methods to investigate traditional fermentations. *Appl. Environ. Microbiol.* 65:5464–5473.
5. Batra, L. R. 1981. Fermented cereals and grain legumes of India and vicinity, p. 547–553. In M. Moo-Young and C. W. Robinson (ed.), *Advances in Biotechnology*, vol. 2. *Fuels, Chemicals, Foods and Waste Treatment*. Pergamon Press, New York, N.Y.
6. Batra, L. R., and P. D. Millner. 1974. Some Asian fermented foods and beverages and associated fungi. *Mycologia* 66:942–950.
7. Benjamin, C. R., and C. W. Hesseltine. 1957. The genus *Actinomucor*. *Mycologia* 49:240–249.
8. Brown, A. C., and A. Valiere. 2004. The medicinal uses of poi. *Nutr. Clin. Care* 7:69–74.
9. Campbell-Platt, G. 1980. African locust bean (*Parkia* species) and its West African fermented food product, dawadawa. *Ecol. Food Nutr.* 9:123–132.
10. Chang, P. K. 2004. Lack of interaction between AFLR and AFLJ contributes to nonaflatoxigenicity of *Aspergillus sojae*. *J. Biotechnol.* 107:245–253.
11. Chou, C. C., F. M. Ho, and C. S. Tsai. 1988. Effects of temperature and relative humidity on the growth of and enzyme production by *Actinomucor taiwanensis* during sufu pehtze preparation. *Appl. Environ. Microbiol.* 54:688–692.
12. Chou, C.-C., and J.-H. Rwan. 1995. Mycelial propagation and enzyme production in koji prepared with *Aspergillus oryzae* on various rice extrudates and steamed rice. *J. Ferment. Bioeng.* 79:509–512.
13. De Vos, W. M. 2005. Frontiers in food biotechnology—fermentations and functionality. *Curr. Opin. Biotechnol.* 16:187–189.
14. Dung, N. T. P. 2004. Defined fungal starter granules for purple glutinous rice wine. Ph.D. thesis. Wageningen University, Wageningen, The Netherlands.
15. Fleet, G. H. 1998. The microbiology of alcoholic beverages, p. 217–262. In B. J. B. Wood (ed.), *Microbiology of Fermented Foods*, 2nd ed., vol. 1. Blackie Academic & Professional, London, United Kingdom.
16. Gänzle, M. G., and R. F. Vogel. 2002. Contribution of reutericyclin production to the stable persistence of *Lactobacillus reuteri* in an industrial sourdough fermentation. *Int. J. Food Microbiol.* 80:31–45.
17. Gashe, B. A. 1985. Involvement of lactic acid bacteria in the fermentation of tef (*Eragrostis tef*), an Ethiopian fermented food. *J. Food Sci.* 50:800–801.
18. Ginting, E., and J. Arcot. 2004. High-performance liquid chromatographic determination of naturally occurring folates during tempe preparation. *J. Agric. Food Chem.* 52:7752–7758.
19. Greenwalt, C. J., K. H. Steinkraus, and R. A. Ledford. 2000. Kombucha, the fermented tea: microbiology, composition, and claimed health effects. *J. Food Prot.* 63:976–981.
20. Han, B. Z., F. M. Rombouts, and M. J. R. Nout. 2004. Amino acid profiles of sufu, a Chinese fermented soybean food. *J. Food Comp. Anal.* 17:689–698.
21. Han, B. Z., F. M. Rombouts, and M. J. R. Nout. 2001. A Chinese fermented soybean food. *Int. J. Food Microbiol.* 65:1–10.
22. Hanya, Y., and T. Nakadai. 2003. Yeasts and soy products, p. 413–428. In T. Boekhout and V. Robert (ed.), *Yeasts in Food: Beneficial and Detrimental Aspects*. B. Behr's Verlag GmbH & Co. KG, Hamburg, Germany.
23. Hounhouigan, D. J., M. J. R. Nout, C. M. Nago, J. H. Houben, and F. M. Rombouts. 1993. Changes in the physico-chemical properties of maize during natural fermentation of mawe. *J. Cereal Sci.* 17:291–300.
24. Inatsu, Y., K. Kimura, and Y. Itoh. 2002. Characterization of *Bacillus subtilis* strains isolated from fermented soybean foods in Southeast Asia: comparison with *B. subtilis* (natto) starter strains. *Jpn. Agric. Res. Q.* 36:169–175.
25. Ishikawa, H., K. Okubo, and T. Oki. 1972. Characteristic spinnability of a natto mucin solution. *Nippon Kagaku Kaishi* 11:2171–2177.
26. Jakobsen, M., M. D. Cantor, and L. Jespersen. 2001. Production of bread, cheese and meat, p. 1–22. In H. D. Osiewacz (ed.), *The Mycota, Industrial Applications*, vol. 10. Springer-Verlag, New York, N.Y.
27. Jensen, I. 1998. Bread and bakers' yeast, p. 172–198. In B. J. B. Wood (ed.), *Microbiology of Fermented Foods*, 2nd ed., vol. 1. Blackie Academic & Professional, London, United Kingdom.
28. Jespersen, L., M. Halm, K. Kpodo, and M. Jakobsen. 1994. Significance of yeasts and moulds occurring in maize dough fermentation for “kenkey” production. *Int. J. Food Microbiol.* 24:239–248.
29. Joshi, V. K., D. K. Sandhu, and N. S. Thakur. 1999. Fruit based alcoholic beverages, p. 647–744. In V. K. Joshi and A. Pandey (ed.), *Biotechnology: Food Fermentation*, vol. 2. Educational Publishers, Ernakulam, India.
30. Kelly, W. J., R. V. Asmundson, G. L. Harrison, and C. M. Huang. 1995. Differentiation of dextran-producing *Leuconostoc* strains from fermented rice cake (puto) using pulsed-field gel electrophoresis. *Int. J. Food Microbiol.* 26:345–352.
31. Ko, S. D. 1986. Indonesian fermented foods not based on soybeans, p. 67–84. In C. W. Hesseltine and H. L. Wang (ed.), *Mycologia Memoir*, no. 11. *Indigenous Fermented Food of Non-Western Origin*. J. Cramer, Berlin, Germany.
32. Ko, S. D., and C. W. Hesseltine. 1979. Tempe and related foods, p. 115–140. In A. H. Rose (ed.), *Microbial Biomass*, vol. 4. Academic Press, London, United Kingdom.
33. Koga, T., K. Moro, and T. Matsudo. 1998. Antioxidative behaviors of 4-hydroxy-2,5-dimethyl-3(2H)-furanone and 4-hydroxy-2(or 5)-ethyl-5(or 2)-methyl-3(2H)-furanone against lipid peroxidation. *J. Agric. Food Chem.* 46:946–951.
34. Lee, C.-H. 1996. Lactic fermented foods and their benefits in Asia. *Food Biotechnol.* 5:187–197.
35. Lee, J.-S., G.-Y. Heo, J. W. Lee, Y.-J. Oh, J. A. Park, Y.-H. Park, Y.-R. Pyun, and J. S. Ahn. 2005. Analysis of kimchi microflora using denaturing gradient gel electrophoresis. *Int. J. Food Microbiol.* 102:143–150.

36. Liu, Y.-H., and C. C. Chou. 1994. Contents of various types of proteins and water soluble peptides in sufu during ageing and the amino acid composition of taste oligopeptides. *J. Chin. Agric. Chem. Soc.* 32:276–283.
37. Matsuo, M. 2004. Low-salt O-miso produced from koji fermentation of oncom improves redox state and cholesterolemia in rats more than low-salt soybean-miso. *J. Nutr. Sci. Vitaminol.* (Tokyo) 50:362–366.
38. Minamiyama, Y., and S. Okada. 2003. Miso: production, properties, and benefits to health, p. 277–286. In E. R. Farnworth (ed.), *Handbook of Fermented Functional Foods*. CRC Press, Boca Raton, Fla.
39. Mukherjee, S. K., M. N. Albury, C. S. Pederson, A. G. Van Veen, and K. H. Steinkraus. 1965. Role of *Leuconostoc mesenteroides* in leavening the batter of idli, a fermented food of India. *Appl. Microbiol.* 13:227–231.
40. Mulyowidarso, R. K., G. H. Fleet, and K. A. Buckle. 1989. The microbial ecology of soybean soaking for tempe production. *Int. J. Food Microbiol.* 8:35–46.
41. Nagai, T., L.-S. Phan Tran, Y. Inatsu, and Y. Itoh. 2000. A new IS4 family insertion sequence, IS4Bsu1, responsible for genetic instability of poly-gamma-glutamic acid production in *Bacillus subtilis*. *J. Bacteriol.* 182:2387–2392.
42. Nago, M. C., J. D. Hounhouigan, N. Akissoe, E. Zanou, and C. Mestres. 1998. Characterization of the Beninese traditional ogi, a fermented maize slurry: physicochemical and microbiological aspects. *Int. J. Food Sci. Technol.* 33:307–315.
43. Nche, P. F., G. T. Odamtten, M. J. R. Nout, and F. M. Rombouts. 1996. Soaking determines the quality of aflata for kenkey production. *J. Cereal Sci.* 24:291–297.
44. Nikkuni, S., T. B. Karki, K. S. Vilku, T. Suzuki, K. Shindoh, C. Suzuki, and N. Okada. 1995. Mineral and amino acid contents of kinema, a fermented soybean food prepared in Nepal. *Food Sci. Technol. Int.* 1:107–111.
45. Nout, M. J. R., and K. E. Aidoo. 2002. Asian fungal fermented food, p. 23–47. In H. D. Osiewacz (ed.), *The Mycota: Industrial Applications*, vol. 10. Springer-Verlag, New York, N.Y.
46. Nout, M. J. R., W. M. De Vos, and M. H. Zwietering (ed.). 2005. *Food Fermentation*. Wageningen Academic Publishers, Wageningen, The Netherlands.
47. Nout, M. J. R., and J. L. Kiers. 2005. Tempe fermentation, innovation and functionality: up-date into the 3rd millennium. *J. Appl. Microbiol.* 98:789–805.
48. Nout, M. J. R., and F. M. Rombouts. 1990. Recent developments in tempe research. *J. Appl. Bacteriol.* 69:609–633.
49. Nout, M. J. R., and P. K. Sarkar. 1999. Lactic acid food fermentation in tropical climates. *Antonie Leeuwenhoek* 76:395–401.
50. Nunomura, N., and M. Sasaki. 1992. Japanese soy sauce flavour with emphasis on off-flavours, p. 287–312. In G. Charalambous (ed.), *Off-flavours in Foods and Beverages*. Elsevier, Amsterdam, The Netherlands.
51. Nuraida, L., M. C. Wachter, and J. D. Owens. 1995. Microbiology of pozol, a Mexican fermented maize dough. *World J. Microbiol. Biotechnol.* 11:567–571.
52. Odunfa, S. A. 1988. Review: African fermented foods, from art to science. *MIRCEN J.* 4:259–273.
53. Ohta, T. 1986. Natto, p. 85–93. In N. R. Reddy, M. D. Pierson, and D. K. Salunkhe (ed.), *Legume-based Fermented Foods*. CRC Press, Boca Raton, Fla.
54. Onyekwere, O. O., I. A. Akinrele, and O. A. Koleoso. 1989. Industrialization of ogi fermentation, p. 329–362. In K. H. Steinkraus (ed.), *Industrialization of Indigenous Fermented Foods*. Marcel Dekker, New York, N.Y.
55. Onyekwere, O. O., I. A. Akinrele, O. A. Koleoso, and G. Heys. 1989. Industrialization of gari fermentation, p. 363–410. In K. H. Steinkraus (ed.), *Industrialization of Indigenous Fermented Foods*. Marcel Dekker, New York, N.Y.
56. O'Toole, D. 1999. Characteristics and use of okara, the soybean residue from soy milk production—a review. *J. Agric. Food Chem.* 47:363–371.
57. Owens, J. D., N. Allagheny, G. Kipping, and J. M. Ames. 1997. Formation of volatile compounds during *Bacillus subtilis* fermentation of soya beans. *J. Sci. Food Agric.* 74:132–140.
58. Owens, J. D., and L. S. Mendoza. 1985. Enzymatically hydrolysed and bacterially fermented fishery products. *J. Food Technol.* 20:237–293.
59. Pastrana, L., P. J. Blanc, A. L. Santerre, M. O. Loret, and G. Goma. 1995. Production of red pigments by *Monascus ruber* in synthetic media with a strictly controlled nitrogen source. *Process Biochem.* 30:333–341.
60. Pepe, O., G. Blaiotta, G. Moschetti, T. Greco, and F. Villani. 2003. Rope-producing strains of *Bacillus* spp. from wheat bread and strategy for their control by lactic acid bacteria. *Appl. Environ. Microbiol.* 69:2321–2329.
61. Ruiz, J. A., J. Quilez, M. Mestres, and J. Guasch. 2003. Solid-phase microextraction method for headspace analysis of volatile compounds in bread crumb. *Cereal Chem.* 80:255–259.
62. Sarkar, P. K. 2005. Microbiology of traditional alkaline fermented foods, p. 899–915. In T. Satyanarayana and B. N. Johri (ed.), *Microbial Diversity: Current Perspectives and Potential Applications*. I. K. International, New Delhi, India.
63. Sarkar, P. K., B. Hasenack, and M. J. R. Nout. 2002. Diversity and functionality of *Bacillus* and related genera isolated from spontaneously fermented soybeans (Indian kinema) and locust beans (African soumbala). *Int. J. Food Microbiol.* 77:175–186.
64. Shurpalekar, S. R. 1986. Papads, p. 191–217. In N. R. Reddy, M. D. Pierson, and D. K. Salunkhe (ed.), *Legume-Based Fermented Foods*. CRC Press, Boca Raton, Fla.
65. Siebenhandl, S., L. N. Lestario, D. Trimmel, and E. Berghofer. 2001. Studies on tape ketan—an Indonesian fermented rice food. *Int. J. Food Sci. Nutr.* 52:347–357.
66. Soni, S. K., and D. K. Sandhu. 1990. Biochemical and nutritional changes associated with Indian Punjabi warri fermentation. *J. Food Sci. Technol.* 27:82–85.
67. Soni, S. K., and D. K. Sandhu. 1999. Fermented cereal products, p. 895–949. In V. K. Joshi and A. Pandey (ed.), *Biotechnology: Food Fermentation*, vol. 2. Educational Publishers, Ernakulam, India.

68. Soni, S. K., and D. K. Sandhu. 1991. Role of yeast domination in Indian idli batter fermentation. *World J. Microbiol. Biotechnol.* 7:505–507.
69. Steinkraus, K. H. 1994. Nutritional significance of fermented foods. *Food Res. Int.* 27:259–267.
70. Sujaya, I. N., S. Amachi, K. Saito, A. Yokota, K. Asano, and F. Tomita. 2002. Specific enumeration of lactic acid bacteria in ragi tape by colony hybridization with specific oligonucleotide probes. *World J. Microbiol. Biotechnol.* 18:263–270.
71. Susheelamma, N. S., and M. V. L. Rao. 1979. Functional role of the arabinogalactan of black gram (*Phaseolus mungo*) in the texture of leavened foods (steamed puddings). *J. Food Sci.* 44:1309–1312, 1316.
72. Umeta, M., and R. M. Faulks. 1988. The effect of fermentation on the carbohydrates in tef (*Eragrostis tef*). *Food Chem.* 27:181–189.
73. Venkatasubbaiah, P., C. T. Dwarakanath, and V. S. Murthy. 1985. Involvement of yeast flora in idli batter fermentation. *J. Food Sci. Technol.* 22:88–90.
74. Westby, A., and D. R. Twiddy. 1992. Characterization of gari and fu-fu preparation procedures in Nigeria. *World J. Microbiol. Biotechnol.* 8:175–182.
75. Yamabe, S., K. Kaneko, H. Inoue, and T. Takita. 2004. Maturation of fermented rice-koji miso can be monitored by an increase in fatty acid ethyl ester. *Biosci. Biotechnol. Biochem.* 68:250–252.

