



Complete Genome Sequence of a Clinical *Campylobacter* Isolate Identical to a Novel *Campylobacter* Species

Birgitta Duim,^{a,b} Linda van der Graaf-van Bloois,^{a,b} Arjen Timmerman,^{a,b} Jaap A. Wagenaar,^{a,b,c} Jacky Flipse,^d Janny Wallinga,^d Peter Bloembergen,^d William G. Miller,^e Aldert L. Zomer^{a,b}

^aFaculty of Veterinary Medicine, Department of Infectious Diseases and Immunology, Utrecht University, Utrecht, the Netherlands

^bWHO Collaborating Center for Campylobacter/OIE Reference Laboratory for Campylobacteriosis, Utrecht, the Netherlands

^cWageningen Bioveterinary Research, Lelystad, the Netherlands

^dLaboratory of Medical Microbiology and Infectious Diseases, Isala Clinics, Zwolle, the Netherlands

^eProduce Safety and Microbiology Research Unit, Agricultural Research Service, U.S. Department of Agriculture, Albany, California, USA

ABSTRACT Here, we present the complete genome sequence of a *Campylobacter* strain isolated in the Netherlands from a patient with gastroenteritis. The strain showed >98% sequence identity to the novel *Campylobacter* species sequence recently recovered from metagenomic data, isolated from breastfed infants with diarrheal disease, and named “*Candidatus Campylobacter infans*.”

Here, we present the genome sequence of *Campylobacter* strain 19S00001, which was isolated from a 42-year-old male in the Netherlands with relapsing diarrhea whose stool samples were repeatedly positive by a *Campylobacter*-specific quantitative PCR (qPCR) (1, 2). To isolate this *Campylobacter* strain, 1 g of fecal material was suspended in 1.5 ml phosphate-buffered saline (PBS), and subsequently, 300 μ l of this suspension was added onto a cellulose acetate membrane filter (0.65 μ m; Millipore) placed on Columbia agar with 5% sheep blood (Oxoid, Thermo Scientific, Inc.) (3). After 30 min, the filter was removed and the plate was incubated under microaerobic conditions (83.3% N₂, 7.1% CO₂, 3.6% H₂, and 6% O₂) at 37°C for 72 h. One *Campylobacter*-suspected colony was detected and subcultured on Columbia agar with 5% sheep blood under microaerobic conditions at 37°C for 5 days. DNA was isolated using the DNeasy UltraClean microbial kit (Qiagen, Venlo, the Netherlands), was not sheared, and was size selected for sequencing. The Illumina library was prepared using the Nextera kit (Illumina, San Diego, CA, USA). Pooled libraries were sequenced using a NextSeq system providing 1,068,916 reads (*N*₅₀ of 150 bp) that were trimmed using TrimGalore v0.4.4 (<https://github.com/FelixKrueger/TrimGalore>). Nanopore sequencing was performed according to protocol SQK-LSK109 with flow cell type R9.4.1 on a MinION device (FLO-MIN106D; Oxford Nanopore, Oxford, United Kingdom), using the live (fast) basecalling method in MinKNOW v19.12.1 on a MinIT device, providing 2,484,000 reads (*N*₅₀ of 6.4 kb) that were trimmed and downsampled to 200 \times coverage using filtlong (<https://github.com/rwwick/Filtlong>), and resulting in 17,582 reads (*N*₅₀ of 23 kb) that were assembled using metaFlye v2.7 (4) into a single scaffold. The genome was circularized and rotated using a DnaA database in Unicycler v0.4.7 (5) with overlapping Illumina and Nanopore reads. This resulted in two circular contigs, representing a 1,754.5-kb chromosome with a GC content of 35.9% and a 5,856-kb plasmid. Prokka v1.13 (6) was used for annotation with “*Candidatus Campylobacter infans*” as an additional custom database (GenBank accession number [SPMW000000001](https://www.ncbi.nlm.nih.gov/nuccore/SPMW000000001)) (7, 8). A core genome phylogeny without correction for recombination was reconstructed using FastTree (9), based on a 365,157-bp core gene superalignment of 351 core genes of which the protein sequences had at least 35% sequence identity, as determined by

Citation Duim B, van der Graaf-van Bloois L, Timmerman A, Wagenaar JA, Flipse J, Wallinga J, Bloembergen P, Miller WG, Zomer AL. 2021. Complete genome sequence of a clinical *Campylobacter* isolate identical to a novel *Campylobacter* species. *Microbiol Resour Announc* 10:e00721-20. <https://doi.org/10.1128/MRA.00721-20>.

Editor Vincent Bruno, University of Maryland School of Medicine

Copyright © 2021 Duim et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Birgitta Duim, b.duim@uu.nl.

Received 23 July 2020

Accepted 20 January 2021

Published 18 February 2021

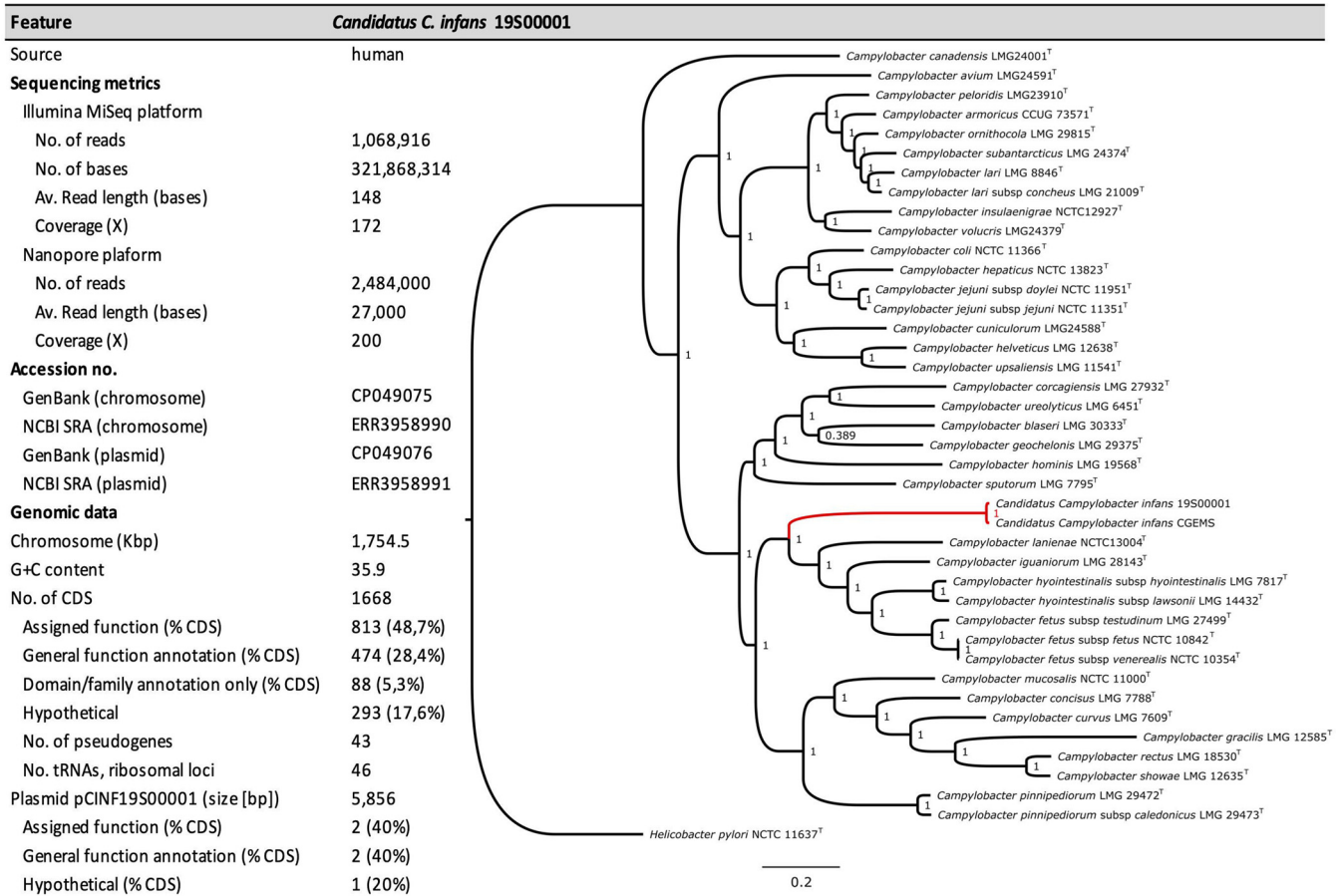


FIG 1 Sequencing metrics, genomic data, and phylogenetic dendrogram based on the core genome single nucleotide polymorphisms (SNPs). *Helicobacter pylori* J99 is used as an outgroup and root. Local support values, calculated with the Shimodaira-Hasegawa test (14), for all branches are given. Bar, 0.2 substitutions per nucleotide position in the core gene superalignment.

Roary v3.12.0 (10). The average nucleotide identity (ANI) was identified using JSpecies v1.2.1 (11). Default parameters were used for all software unless specified otherwise.

The sequence clustered with the genome of the recently identified novel putative species belonging to the *Campylobacter fetus* group “*Candidatus Campylobacter infans*” (Fig. 1) (7) with a 98.27% ANI.

The 19S00001 genome contains a type II-B CRISPR-Cas system (12). A putative integrated phage containing genes for a type IV (T4SS) conjugative transfer system (13) and a second phage integration containing a zonula occludens toxin (Zot) island were identified. The S-layer secretion system gene *sapDEF*, as well as D-arabinose 5-phosphate isomerase (*kpsF*), and flagellin-associated genes were present as identified in the genome of “*Candidatus Campylobacter infans*” (7). This study shows the first complete genome and plasmid sequences of a clinical isolate with high sequence identity to a novel *Campylobacter* species.

Data availability. Genome and plasmid sequences have been deposited in GenBank under the accession numbers CP049075.1 and CP049076.1. The Nanopore reads have been deposited in SRA under the accession numbers ERR3958990 and ERR3958991 (Fig. 1).

ACKNOWLEDGMENTS

We thank Alejandro Baars and Heleen Zweerus for performing the Nanopore run.

REFERENCES

1. Buijnesteijn van Coppenraet LES, Dullaert-de Boer M, Ruijs GJHM, van der Reijden WA, van der Zanden AGM, Weel JFL, Schuurs TA. 2015. Case-control comparison of bacterial and protozoan microorganisms associated with gastroenteritis: application of molecular detection.

- Clin Microbiol Infect 21:592.e9–592.e19. <https://doi.org/10.1016/j.cmi.2015.02.007>.
2. Flipse J, Duim B, Wallinga J, de Wijkerslooth LRH, van der Graaf-van Bloois L, Timmerman AJ, Zomer AL, Veldman KT, Wagenaar JA, Bloembergen P. 2020. A case of persistent diarrhea in a man with the molecular detection of various *Campylobacter* species and the first isolation of *Candidatus Campylobacter infans*. Pathogens 9:1003. <https://doi.org/10.3390/pathogens9121003>.
 3. Steele TW, McDermott SN. 1984. The use of membrane filters applied directly to the surface of agar plates for the isolation of *Campylobacter jejuni* from feces. Pathology 16:263–265. <https://doi.org/10.3109/00313028409068535>.
 4. Kolmogorov M, Rayko M, Yuan J, Polevikov E, Pevzner P. 2019. metaFlye: scalable long-read metagenome assembly using repeat graphs. bioRxiv <https://doi.org/10.1101/637637>.
 5. Wick RR, Judd LM, Gorrie CL, Holt KE. 2017. Unicycler: resolving bacterial genome assemblies from short and long sequencing reads. PLoS Comput Biol 13:e1005595. <https://doi.org/10.1371/journal.pcbi.1005595>.
 6. Seemann T. 2014. Prokka: rapid prokaryotic genome annotation. Bioinformatics 30:2068–2069. <https://doi.org/10.1093/bioinformatics/btu153>.
 7. Bian X, Garber JM, Cooper KK, Huynh S, Jones J, Mills MK, Rafala D, Nasrin D, Kotloff KL, Parker CT, Tennant SM, Miller WG, Szymanski CM. 2020. *Campylobacter* abundance in breastfed infants and identification of a new species in the Global Enterics Multicenter Study. mSphere 5:e00735-19. <https://doi.org/10.1128/mSphere.00735-19>.
 8. Miller WG, Yee E, Bono JL. 2018. Complete genome sequence of the *Arcobacter molluscorum* type strain LMG 25693. Microbiol Resour Announc 7:e01293-18. <https://doi.org/10.1128/MRA.01293-18>.
 9. Price MN, Dehal PS, Arkin AP. 2009. FastTree: computing large minimum evolution trees with profiles instead of a distance matrix. Mol Biol Evol 26:1641–1650. <https://doi.org/10.1093/molbev/msp077>.
 10. Page AJ, Cummins CA, Hunt M, Wong VK, Reuter S, Holden MTG, Fookes M, Falush D, Keane JA, Parkhill J. 2015. Roary: rapid large-scale prokaryote pan genome analysis. Bioinformatics 31:3691–3693. <https://doi.org/10.1093/bioinformatics/btv421>.
 11. Richter M, Rosselló-Móra R. 2009. Shifting the genomic gold standard for the prokaryotic species definition. Proc Natl Acad Sci U S A 106:19126–19131. <https://doi.org/10.1073/pnas.0906412106>.
 12. Makarova KS, Koonin EV. 2015. Annotation and classification of CRISPR-Cas systems. Methods Mol Biol 1311:47–75. https://doi.org/10.1007/978-1-4939-2687-9_4.
 13. van der Graaf-van Bloois L, Miller WG, Yee E, Gorkiewicz G, Forbes KJ, Zomer AL, Wagenaar JA, Duim B. 2016. *Campylobacter fetus* subspecies contain conserved type IV secretion systems on multiple genomic islands and plasmids. PLoS One 11:e0152832. <https://doi.org/10.1371/journal.pone.0152832>.
 14. Shimodaira H, Hasegawa M. 1999. Multiple comparisons of log-likelihoods with applications to phylogenetic inference. Mol Biol Evol 16:1114–1116. <https://doi.org/10.1093/oxfordjournals.molbev.a026201>.