



# Host plant resistance to thrips (Thysanoptera: Thripidae) – current state of art and future research avenues

Sanae Mouden and Kirsten A Leiss

Integrated Pest Management (IPM) is endorsed as the future standard for crop protection worldwide. This holistic concept integrates preventative and curative measures amongst which host plant resistance (HPR) plays an essential role. Up to now HPR has been a somewhat under-utilized tool in pest management due to widespread use of pesticides and technological hindrance. Thrips are key pests in agriculture and horticulture worldwide. Here we provide an overview on the current status of research on constitutive and induced HPR including thrips–host relationships and thrips as virus vectors. We stress modulation of plant defense responses by abiotic and biotic elicitors to increase HPR and provide an outlook on the increasing potential of HPR inspired by the fast advancement of –omics techniques.

## Address

Wageningen University & Research, Business Unit Horticulture, Violierenweg 1, 2665 MV Bleiswijk, The Netherlands

Corresponding author: Leiss, Kirsten A ([kirsten.leiss@wur.nl](mailto:kirsten.leiss@wur.nl))

Current Opinion in Insect Science 2021,45:xx–yy

This review comes from a themed issue on **Pests and resistance**

Edited by **Andrew Michel** and **Marion Olney Marris**

<https://doi.org/10.1016/j.cois.2020.11.011>

2214-5745/© 2020 The Authors. Published by Elsevier Inc. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

## Introduction

Global agriculture is at an important juncture and in a period of great change, whereby Integrated Pest Management (IPM) is endorsed as the future paradigm for crop protection [1]. The holistic concept of IPM emphasizes a systems approach integrating preventative and curative intervention measures amongst which host plant resistance (HPR) plays an essential role. Up to now HPR has been a somewhat under-utilized tool in pest management. Selection for yield, taste or appearance has greatly reduced phenotypic and genetic diversity of cultivated plants often accompanied by a loss of HPR [2]. This was reasonable using pesticides on a large scale. Widely spreading pesticide resistance and, especially for the EU, more stringent pesticide regulations related to apparent risks for human health, beneficial organisms and the environment, reduce pesticide availability which makes

established cropping systems highly vulnerable. HPR incorporates varieties which hamper preference, reproduction, feeding and/or transmission of insect-borne pathogens. Thrips species of the family Thripidae are particularly polyphagous and contain key pests of a large variety of ornamentals and vegetables worldwide [3], including western flower thrips (WFT: *Frankliniella occidentalis*), onion thrips (*Thrips tabaci*) and melon thrips (*Thrips palmi*). In addition to direct feeding damage these species are virus vectors of which tomato spotted wilt virus (TSWV) is the economically most important [4\*\*]. The present review provides an interdisciplinary overview of the current body of knowledge on constitutive and inducible components of HPR to thrips, and discusses the potential of elicitors to modulate plant defenses. Finally, we provide an outlook on the future exploitation of HPR inspired by the fast advancement of –omics techniques.

## Host plant resistance to thrips

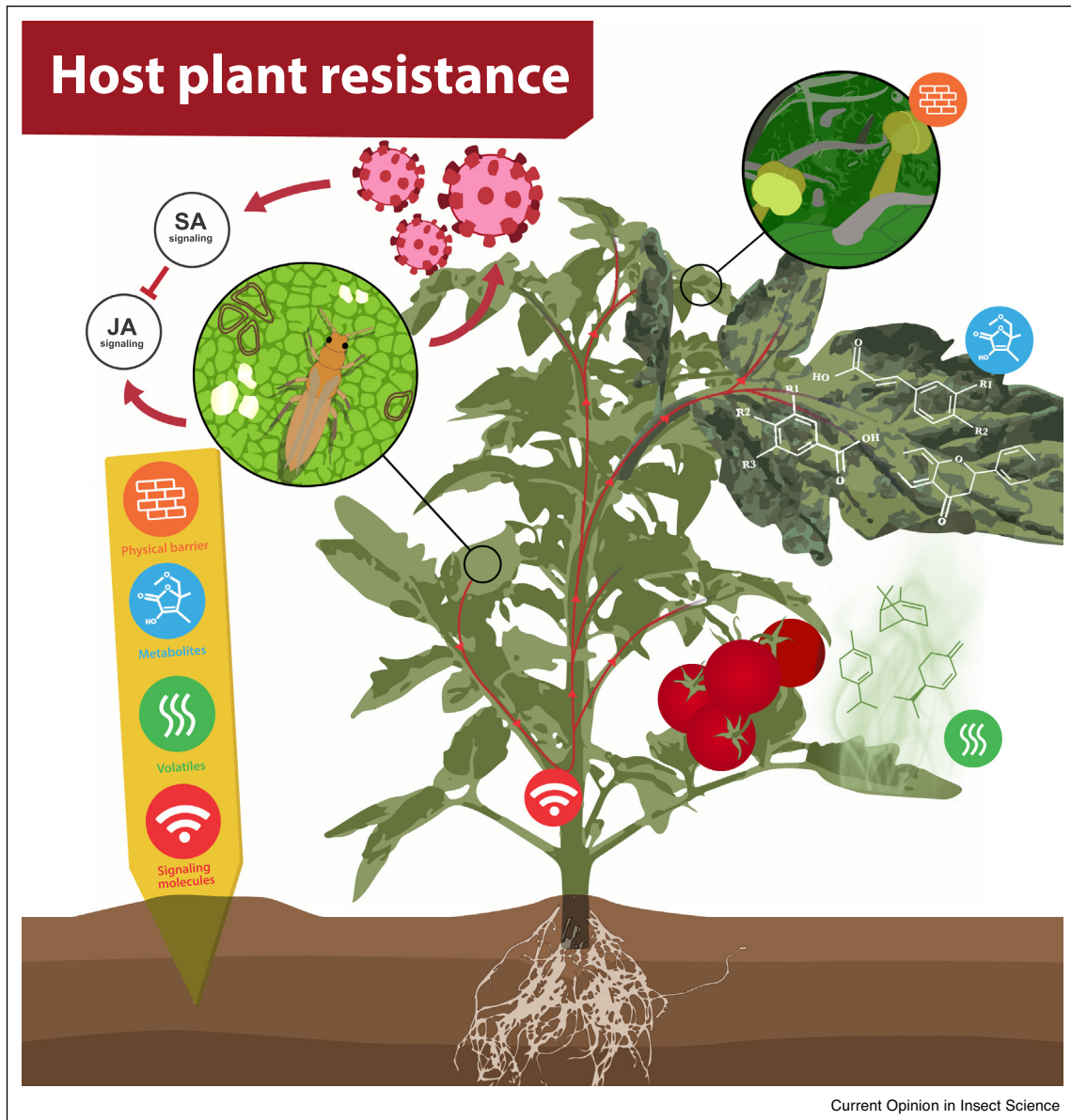
Over the past decades, great effort has been devoted developing plant varieties with resistance to viruses although, resistance to thrips feeding is recently gaining attention [4\*,5]. Empirical approaches to HPR often involves phenotypic screening, that is, evaluation of germplasm for resistant genotypes [6–14]. Advances in phenotyping technologies support high-throughput characterization of crop resistance to thrips [8,15]. Moreover, the affordability of current omics technologies fosters a major advancement in connecting genotype–phenotype relationships to enhance our understanding of plant–thrips defense mechanisms [16–21].

HPR results from a complex set of interdependent mechanisms (Figure 1) ranging from morphological barriers to the complex array of signaling molecules of the plant's immune system. These plant-defensive responses are associated with constitutive, thus always present traits, or induced plant traits shaped upon attack or elicitation in the plant's morphology, physiology and/or chemistry producing plant secondary metabolites [13,22]. Both ways are not mutually exclusive and lead to direct or indirect effects through attraction of natural enemies.

## Constitutive defense

Constitutive barriers, including morphological leaf traits, form a first line of defense against thrips [7,11,14,23,24]. Many studies are correlative with little consensus concerning the role of leaf traits. For instance, feeding damage by the viruliferous soybean thrips (*Neohydatothrips variabilis*)

Figure 1



A schematic model for host plant resistance (HPR) to thrips. Thrips face direct constitutive plant defenses such as physical barriers (i.e. leaf pubescence or trichomes), secondary defensive plant metabolites or indirect defenses such as the emission of volatile organic compounds. Upon thrips feeding, damage typically appears as silver scars whereas tospoviruses (depicted by red virus particles), vectored by thrips, lead to chlorotic ringspots (upper left part). Thrips attack activates local plant immune responses by jasmonic acid (JA) production which is systemically transported to undamaged plant tissues (red arrow). Tospoviruses induce the salicylic acid (SA) pathway and are capable of manipulating host defenses through SA-JA antagonism to benefit their vector.

was reduced in genotypes with low pubescence [14] whereas, in another study Abdelmaksoud *et al.* [24] reported lower WFT infestations in strawberry cultivars with high densities of non-glandular trichomes. However, in both studies only a small number of genotypes was analyzed, therefore, probably additional not measured traits were interfering. In search for markers associated with thrips resistance, Bac-Molenaar *et al.* [25<sup>••</sup>] reported that foliar resistance to WFT was independent from glandular

trichome density and derived volatile compounds in a mapping population derived from a resistant and non-resistant tomato line. Metabolomic analysis identified leaf  $\alpha$ -tomatine and a non-elucidated phenolic compound as putative defense metabolites related to WFT resistance. In addition, glandular trichomes are known for their production of sticky resinous substances, such as acylsugars. Their importance to WFT resistance has been shown by Vosman *et al.* [26]. Ben-Mahmoud *et al.* [27] demonstrated that

particular combinations of acylsugars with specific fatty acid profiles, not only reduced WFT oviposition but also lowered incidence of TSVW. Regarding defense compounds Maharija *et al.* [17] reported two capsiariosides and a flavonoid to correlate with WFT resistance using metabolite quantitative trait loci (QTL) mapping. Concurrently, capsiariosides were negatively correlated with WFT preference and damage in an untargeted metabolomic profiling approach in *Capsicum* whereas, sucrose and malonylated flavone glycosides were related to susceptibility [18]. In addition, plant olfactory cues play a role in direct defenses as composition and magnitude of volatile emission shapes host localization or preferences [28–32]. Moreover, thrips herbivory may also affect plant biochemistry through changes in nutritional host quality [33,34] or by inducing defenses.

### Inducible defense – signaling and manipulation

While constitutive defenses have gathered substantial attention in the majority of screening studies, induced resistance is rapidly gaining interest [35]. In contrast to constitutive defenses induced defenses are associated with lower metabolic costs. All plants, whether resistant or susceptible, respond to insect attack by induction of a coordinated defense strategy primarily mediated by endogenous phytohormonal signaling, followed by biochemical cascades activating local and/or systemic defense mechanisms (Figure 1). Among the early plant responses, jasmonic acid (JA) acts as a main regulator of induced thrips defenses [22,36,37]. Pepper plants colonized by WFT enhanced the expression of the JA-related marker genes *CaLOX2* and *CAP1N II* [37]. Moreover, WFT are susceptible to externally induced JA-mediated defenses. Foliar JA application impaired feeding of WFT [38,39] and increased duration of developmental stages [40]. Unlike adults, feeding activities of WFT larvae were observed to tolerate naturally induced levels of JA-defenses suggesting that artificial induction of JA-regulated defenses overamplifies rather than mimics natural response [41]. Late plant responses include the production of plant secondary metabolites [38,39].

In contrast, thrips-vector viruses such as TSWV induce the salicylic acid (SA) pathway which indirectly enhances fitness and performance of the vector by exploiting SA-JA antagonism [21]. In addition to immunological plant modulation, viruses modify vector behavioral responses to facilitate virus transmission [42,43]. Such behavioral responses can be mediated by plant-derived cues such as volatile alterations aimed at attracting [44] or repelling vectors [45].

Whether defenses actually enhance plant resistance depends on whether thrips have evolved counter adaptations such as the ability to detoxify and degrade plant toxins [34,46,47]. The underlying molecular components of how thrips feeding modulates induced responses still

remains unclear [35]. There are indications that plant responses to thrips are influenced by potential elicitors such as saliva components [48<sup>\*</sup>] and endosymbiotic-borne cues [49]. The latter points at the potential of different bacterial groups in manipulating plant defense responses to thrips, highlighting the importance of a multitrophic context when considering thrips–plant interactions.

In complex food webs omnivorous thrips can affect plant-mediated interactions of co-existing herbivorous arthropods such as whiteflies [50] or aphids [51<sup>\*</sup>]. Such dual interactions may even cascade up to the third trophic level by influencing preference and performance of natural enemies [51<sup>\*</sup>]. A study by Ataide *et al.* [41], aimed to disentangle direct and indirect effects of plant defenses, demonstrated that feeding by predatory omnivorous WFT larvae was independent from JA-mediated indirect defense effects on spider mite prey quality.

### Modulating defenses by plant resistance inducers

Plant signals inducing resistance are highly conserved among plant species. Accordingly, modulation of plant defenses to increase HPR, through biotic and abiotic elicitors (Figure 2), has a great potential [35].

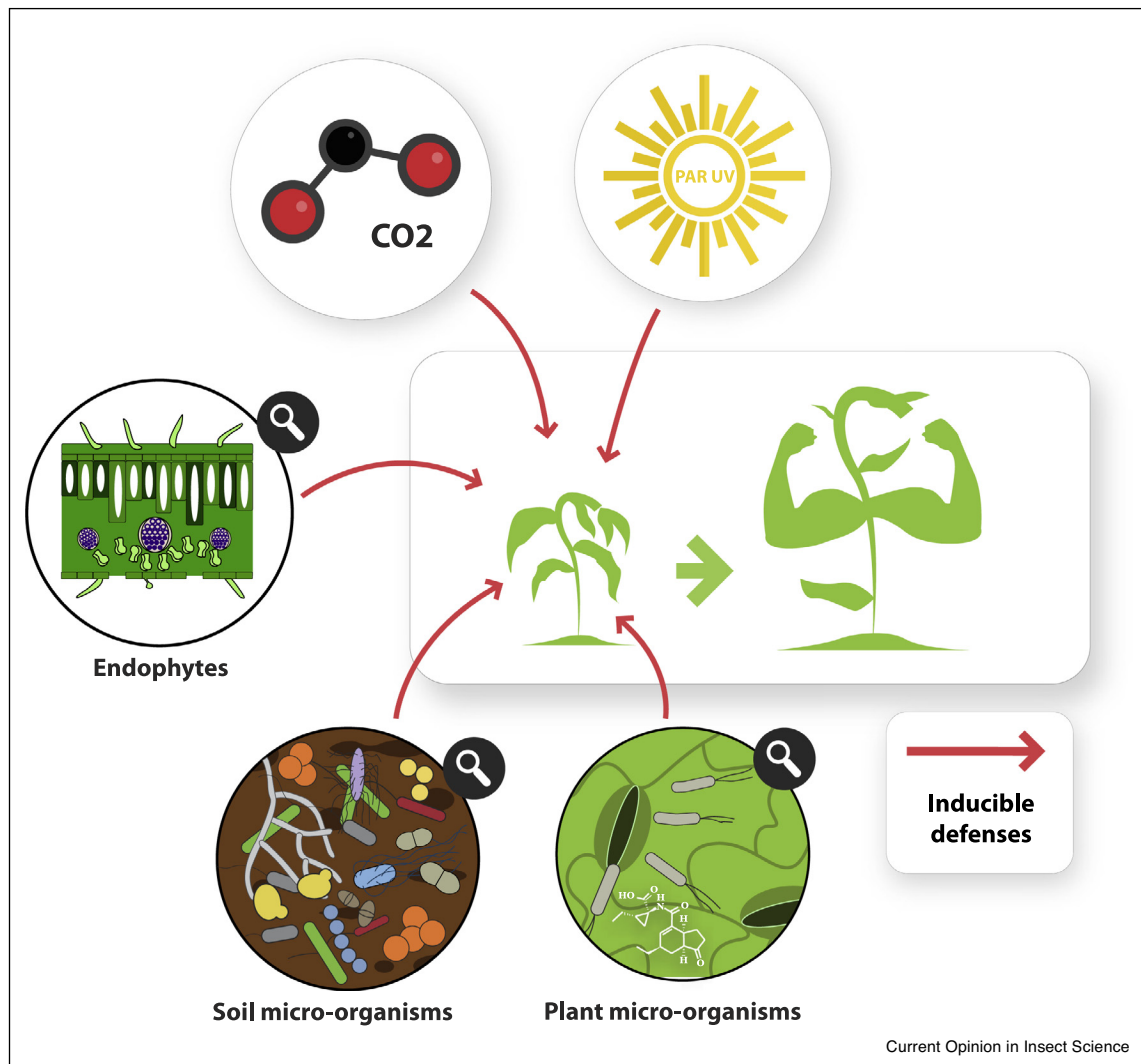
#### Abiotic elicitors

Environmental factors such as photosynthetically active radiation [52] and ultraviolet light [22,36,53] can mediate HPR to thrips. Exogenously applied calcium chloride improved resistance of kidney bean plants to WFT by extending the larval developmental time [54]. Calcium application only slightly induced the activities of defense-related enzymes whereas more potent defense responses were induced upon subsequent attack, a phenomenon generally referred to as priming. Priming has also been observed in UV-treated chrysanthemum, which experienced a stronger induction of JA signaling after thrips infestation [22]. While JA or its volatile derivative methyl jasmonate are commonly tested synthetic elicitors to mimic thrips induced plant responses [39], not much progress has been made in the development of products based on this mechanism.

#### Biotic elicitors

Regarding biotic elicitors, a growing body of research is demonstrating how microbial communities mediate plant defense through direct and indirect changes to plant immunity and/or nutrition. Infiltration of tomato leaves with *Pseudomonas syringae* (Pst), producing the JA analogue coronatine, reduced WFT feeding damage by reinforcing plant defense metabolites, such as phenols [55]. Pst-derived and filtered culture medium, as well as a coronatine defective Pst mutant increased tomato resistance to thrips suggesting that induction of resistance did not require a living Pst population but likely depends on other additional defense elicitors. Earthworm-driven modifications of soil properties, particularly

Figure 2



Modulation of plant inducible defenses. Biotic elicitors (soil microbes, plant microbes and endophytes) as well as abiotic elicitors such as ultraviolet light (UV), photosynthetically active radiation (PAR) and CO<sub>2</sub> can be exploited as strategies to induce resistance to thrips.

through increased nitrogen availability and microbial activity, decreased thrips abundance by inducing phenols as defense compounds [56]. Muvea *et al.* [57] reported that colonization of onions by the fungal endophyte *Hypocrea lixii* not only enhanced resistance against *T. tabaci* but also reduced incidence of thrips-transmitted *iris yellow spot virus* (IYSV). There is a growing interest in manipulating soil microbiomes to improve above-ground pest management. Soil microbiomes, conditioned by non-crop plants, induced aboveground resistance in chrysanthemums to WFT [58<sup>\*</sup>], which was positively correlated with the amount of leaf chlorogenic acid as a secondary defense compound.

### Host plant resistance – quo vadis?

Development of HPR has been hampered for technological reasons. *A-priori* knowledge on plant metabolites of

the crop in question was required for adequate chemical analysis. Concurrently, large-scale screening of thrips HPR on breeding populations was almost impossible since only *in-vivo* bioassays could be performed. The fast advancement of omics-techniques, however, enables a much quicker and comprehensive understanding of HPR to thrips. Genetic engineering has resulted in the first-ever transgenic plant trait targeting thrips [59]. Transgenic cotton expressing a *Bacillus thuringiensis* protein showed substantially less damage to WFT and tobacco thrips (*Frankliniella fusca*) [60<sup>\*</sup>,61]. Gene-editing tools such as CRISPR can further accelerate the introduction of thrips HPR into crops. However, in Europe, varieties based on this technology are subject to the same stringent regulations as conventional genetically modified organisms. Therefore, the combination of metabolomics

and QTL mapping identifying markers as selection tools for thrips HPR in conventional breeding programs is a very promising way forward. Especially since quantitative traits depend on the cumulative action of several different genes making it more difficult for thrips to break through this defense. Because of the rapid progress in next generation sequencing more genomic crop sequences are becoming available. However, ornamentals are often polyploid and, although current techniques are fast, data analysis and interpretation are still challenging. The use of metabolomic markers shows potential for thrips HPR screening in these commodities. Next to breeding for conventional HPR, breeding for induced HPR offers a completely new avenue, which needs to be explored. Besides breeding, crop management techniques inducing HPR look promising. The transition to more efficient but energy saving LED light in greenhouses enables the use of ultraviolet light as an abiotic elicitor. Also beneficial microbes, used as biotic elicitors, registered as plant protection products or biostimulants, offer great potential. However, there is still relatively little known about their molecular mode of action and interplay with the plant. While HPR becomes increasingly important in thrips management we should not look at this one-sided. Viruses, vectored by thrips, can impact plant–thrips interactions. Also, HPR may impact biological control organisms. We advocate that HPR to thrips needs to be placed into an IPM context. To fully exploit the potential of HPR, also horizontal integration, thus concurrent protection to a broader spectrum of pests should be taken into consideration.

### Conflict of interest statement

Nothing declared.

### Acknowledgements

This research did not receive any specific grant from funding agencies in the public, commercial, or not-for-profit sectors. Johanna Bac-Molenaar is thanked for comments on the manuscript and Lucas Frantzen for designing the graphical illustrations.

### References and recommended reading

Papers of particular interest, published within the period of review, have been highlighted as

- of special interest
- of outstanding interest

1. Dara SK: **The new integrated pest management paradigm for the modern age.** *J Integr Pest Manag* 2019, **10**:12.
2. Whitehead SR, Turcotte MM, Poveda K: **Domestication impacts on plant–herbivore interactions: a meta-analysis.** *Philos Trans R Soc B* 2017, **372**:20160034.
3. Wu SY, Xing ZL, Ma TT, Xu DW, Li YY, Lei ZR, Gao YL: **Competitive interaction between *Frankliniella occidentalis* and locally present thrips species: a global review.** *J Pest Sci* 2020:1-12.
4. Reitz SR, Gao Y, Kirk WD, Hoddle MS, Leiss KA, Funderburk JE: **Invasion biology, ecology, and management of western flower thrips.** *Annu Rev Entomol* 2020, **65**:17-37.
5. Srinivasan R, Abney MR, Lai PC, Culbreath AK, Tallury S, Leal-Bertioli S: **Resistance to thrips in peanut and implications for management of thrips and thrips-transmitted orthotospoviruses in peanut.** *Front Plant Sci* 2018, **9**:1604.
6. Badenes-Pérez FR, López-Pérez JA: **Resistance and susceptibility to powdery mildew, root-knot nematode, and western flower thrips in two types of winter cress (*Brassicaceae*).** *Crop Prot* 2018, **10**:41-47.
7. Kaur B, Kuraparthi V, Bacheler J, Fang H, Bowman DT: **Screening germplasm and quantification of components contributing to thrips resistance in cotton.** *J Econ Entomol* 2018, **111**:2426-2434.
8. Visschers IG, van Dam NM, Peters JL: **An objective high-throughput screening method for thrips damage quantitation using Ilastik and ImageJ.** *Entomol Exp Appl* 2018, **166**:508-515.
9. Beuzelin JM, VanWeelden MT, Soto-Adames FN, Sandhu HS, Davidson RW, Baucum L, Swanson S: **Effect of sugarcane cultivar and foliar insecticide treatment on infestations of the invasive sugarcane thrips, *Fulmekiola serrata* (Thysanoptera: Thripidae), in Florida.** *J Econ Entomol* 2019, **112**:2703-2712.
10. Togola A, Boukar O, Chamarthi S, Belko N, Tamò M, Oigiangbe N, Ojo J, Ibikunle M, Fatokun C: **Evaluation of cowpea mini core accessions for resistance to flower bud thrips *Megalurothrips sjostedti* trybom (Thysanoptera: Thripidae).** *J Appl Entomol* 2019, **143**:683-692.
11. Westmore GW, Allen GR, Wilson CR: **Colour preference and potato cultivar oviposition choice by onion thrips, *Thrips tabaci* (Thysanoptera: Thripidae).** *J Asia Pac Entomol* 2019, **22**:25-32.
12. Lagos-Kutz D, Pawlowski ML, Haudenschild J, Han J, Domier LL, Hartman GL: **Evaluation of soybean for resistance to *Neohydatothrips variabilis* (Thysanoptera: Thripidae) noninfested and infected with soybean vein necrosis virus.** *J Econ Entomol* 2020, **113**:949-955.
13. Romero B, Dillon FM, Zavala JA: **Different soybean cultivars respond differentially to damage in a herbivore-specific manner and decrease herbivore performance.** *Arthropod Plant Interact* 2020, **14**:89-99.
14. Zhou J, Johnson DT, Tzanetakis IE: **Assessing soybean genotypes for feeding damage by *Neohydatothrips variabilis* (Thysanoptera: Thripidae).** *Crop Prot* 2020, **128**:104983.
15. Jongsma MA, Thoen MP, Poleij LM, Wieggers GL, Goedhart PW, Dicke M, Noldus LP, Kruisselbrink JW: **An integrated system for the automated recording and analysis of insect behavior in T-maze arrays.** *Front Plant Sci* 2019, **10**:20.
16. Smeda JR, Schillmiller AL, Anderson T, Ben-Mahmoud S, Ullman DE, Chappell TM, Kessler A, Mutschler MA: **Combination of acylglucose QTL reveals additive and epistatic genetic interactions and impacts insect oviposition and virus infection.** *Mol Breed* 2018, **38**:3.
17. Maharajaya A, Vosman B, Pelgrom K, Wahyuni Y, de Vos RC, Voorrips RE: **Genetic variation in phytochemicals in leaves of pepper (*Capsicum*) in relation to thrips resistance.** *Arthropod Plant Interact* 2019, **13**:1-9.
18. Macel M, Visschers IG, Peters JL, Kappers IF, de Vos RC, van Dam NM: **Metabolomics of thrips resistance in pepper (*Capsicum* spp.) reveals monomer and dimer acyclic diterpene glycosides as potential chemical defenses.** *J Chem Ecol* 2019, **45**:490-501.
19. Shrestha A, Champagne DE, Culbreath AK, Abney MR, Srinivasan R: **Comparison of transcriptomes of an orthotospovirus vector and non-vector thrips species.** *PLoS One* 2019, **14**:e0223438.
20. Gupta R, Min CW, Kim SW, Yoo JS, Moon AR, Shin AY, Kwon SY, Kim ST: **A TMT-based quantitative proteome analysis to elucidate the TSWV induced signaling cascade in susceptible and resistant cultivars of *Solanum lycopersicum*.** *Plants* 2020, **9**:290.
21. Nachappa P, Challacombe JF, Margolies DC, Nichols JR, Whitfield AE, Rotenberg D: **Tomato spotted wilt virus benefits**

- its thrips vector by modulating metabolic and plant defense pathways in tomato. *Front Plant Sci* 2020, **11**:575564.
22. Escobar-Bravo R, Nederpel C, Naranjo S, Kim HK, Rodríguez-López MJ, Chen G, Glauser G, Leiss KA, Klinkhamer PG: **Ultraviolet radiation modulates both constitutive and inducible plant defenses against thrips but is dose and plant genotype dependent.** *J Pest Sci* 2019:1-13.
  23. Jacob TK, Senthil Kumar CM, Devasahayam S, D'Silva S, Kumar RS, Biju CN, Praveena R, Ankegowda SK: **Plant morphological traits associated with field resistance to cardamom thrips (*Sciothrips cardamomi*) in cardamom (*Elettaria cardamomum*).** *Ann Appl Biol* 2020, **177**:143-1451.
  24. Abdelmaksoud EM, El-Refai SA, Mahmoud KW, Ragab ME: **Susceptibility of some new strawberry genotypes to infestation by western flower thrips, *Frankliniella occidentalis* (Pergande) (Thysanoptera: Thripidae) in the nursery.** *Ann Agric Sci* 2020, **65**:144-148.
  25. Bac-Molenaar JA, Mol S, Verlaan MG, Van Elven J, Kim HK, Klinkhamer PG, Leiss KA, Vrieling K: **Trichome independent resistance against western flower thrips in tomato.** *Plant Cell Physiol* 2019, **60**:1011-1024.
  26. Vosman B, van 't Westende WP, Henken B, van Eekelen HD, de Vos RC, Voorrips RE: **Broad spectrum insect resistance and metabolites in close relatives of the cultivated tomato.** *Euphytica* 2018, **214**:46.
  27. Ben-Mahmoud S, Anderson T, Chappell TM, Smeda JR, Mutschler MA, Kennedy GG, Ullman DE: **A thrips vector of tomato spotted wilt virus responds to tomato acylsugar chemical diversity with reduced oviposition and virus inoculation.** *Sci Rep* 2019, **9**.
  28. Avellaneda J, Díaz M, Coy-Barrera E, Rodríguez D, Osorio C: **Rose volatile compounds allow the design of new control strategies for the western flower thrips (*Frankliniella occidentalis*).** *J Pest Sci* 2019:1-14.
  29. Cao Y, Li C, Yang H, Li J, Li S, Wang Y, Gao Y: **Laboratory and field investigation on the orientation of *Frankliniella occidentalis* (Thysanoptera: Thripidae) to more suitable host plants driven by volatiles and component analysis of volatiles.** *Pest Manag Sci* 2019, **75**:598-606.
  30. Diabate S, Deletre E, Murungi LK, Fiaboe KK, Subramanian S, Wesonga J, Martin T: **Behavioural responses of bean flower thrips (*Megalurothrips sjostedti*) to vegetative and floral volatiles from different cowpea cultivars.** *Chemoeology* 2019, **29**:73-88.
  31. Ren X, Wu S, Xing Z, Xu R, Cai W, Lei Z: **Behavioral responses of western flower thrips (*Frankliniella occidentalis*) to visual and olfactory cues at short distances.** *Insects* 2020, **11**:177.
  32. Ren X, Wu S, Xing Z, Gao Y, Cai W, Lei Z: **Abundances of thrips on plants in vegetative and flowering stages are related to plant volatiles.** *J Appl Entomol* 2020, **114**:732-742.
  33. Yu D, Huang P, Chen Y, Lin Y, Akutse KS, Lan Y, Wei H: **Effects of flower thrips (Thysanoptera: Thripidae) on nutritional quality of banana (Zingiberales: Musaceae) buds.** *PLoS One* 2018, **13**:e0202199.
  34. Qian L, He S, Liu X, Huang Z, Chen F, Gui F: **Effect of elevated CO<sub>2</sub> on the interaction between invasive thrips, *Frankliniella occidentalis*, and its host kidney bean, *Phaseolus vulgaris*.** *Pest Manag Sci* 2018, **74**:2773-2782.
  35. Steenbergen M, Abd-el-Hallem A, Bleeker P, Dicke M, Escobar-Bravo R, Cheng G, Haring MA, Kant MR, Kappers IF, Klinkhamer PG *et al.*: **Thrips advisor: exploiting thrips-induced defences to combat pests on crops.** *J Exp Bot* 2018, **69**:1837-1848.
  36. Dillon FM, Chludil HD, Reichelt M, Mithöfer A, Zavala JA: **Field-grown soybean induces jasmonates and defensive compounds in response to thrips feeding and solar UV-B radiation.** *Environ Exp Bot* 2018, **156**:1-7.
  37. Sarde SJ, Bouwmeester K, Venegas-Molina J, David A, Boland W, Dicke M: **Involvement of sweet pepper CaLOX2 in jasmonate-dependent induced defence against western flower thrips.** *J Integr Plant Biol* 2019, **61**:1085-1098.
  38. Chen G, Klinkhamer PG, Escobar-Bravo R, Leiss KA: **Type VI glandular trichome density and their derived volatiles are differently induced by jasmonic acid in developing and fully developed tomato leaves: implications for thrips resistance.** *Plant Sci* 2018, **276**:87-98.
  39. Chen G, Kim HK, Klinkhamer PG, Escobar-Bravo R: **Site-dependent induction of jasmonic acid-associated chemical defenses against western flower thrips in *Chrysanthemum*.** *Planta* 2020, **251**:8.
  40. Zheng X, Chen Y, Zhao L, Chen Y, Zheng L, Zheng K, Mu Y, Zhao X, Gao Y, Zhang J: **Tripartite interactions between jasmonicsalicylic acid pathways, western flower thrips, and thrips-transmitted tomato zonate spot virus infection in *Capsicum annuum*.** *Arthropod Plant Interact* 2019, **13**:289-297.
  41. Ataide LM, Dias CR, Schimmel BC, van Erp T, Pallini A, Kant MR: **Food decisions of an omnivorous thrips are independent from the indirect effects of jasmonate-inducible plant defences on prey quality.** *Sci Rep* 2019, **9**:1-12.
  42. Wan Y, Hussain S, Merchant A, Xu B, Xie W, Wen X, Wang S, Zhang Y, Zhou X, Wu Q: **Tomato spotted wilt orthotospovirus influences the reproduction of its insect vector, western flower thrips, *Frankliniella occidentalis*, to facilitate transmission.** *Pest Manag Sci* 2020, **76**:2406-2414.
  43. Leach A, Fuchs M, Harding R, Nault BA: **Iris yellow spot virus prolongs the adult lifespan of its primary vector, onion thrips (*Thrips tabaci*) (Thysanoptera: Thripidae).** *J Insect Sci* 2019, **19**:8.
  44. Mwando NL, Tamiru A, Nyasani JO, Obonyo MA, Caulfield JC, Bruce TJ, Subramanian S: **Maize chlorotic mottle virus induces changes in host plant volatiles that attract vector thrips species.** *J Chem Ecol* 2018, **44**:681-689.
  45. Wu X, Xu S, Zhao P, Zhang X, Yao X, Sun Y, Fang R, Ye J: **The orthotospovirus nonstructural protein NSs suppresses plant MYC-regulated jasmonate signaling leading to enhanced vector attraction and performance.** *PLoS Pathog* 2019, **15**:e1007897.
  46. Liu L, Hou XL, Yue WB, Xie W, Zhang T, Zhi JR: **Response of protective enzymes in western flower thrips (Thysanoptera: Thripidae) to two leguminous plants.** *Environ Entomol* 2020, **2020** <http://dx.doi.org/10.1093/ee/nvaa090>.
  47. Cao Y, Yang H, Gao Y, Wang L, Li J, Wang C, Li C: **Effect of elevated CO<sub>2</sub> on the population development of the invasive species *Frankliniella occidentalis* and native species *Thrips hawaiiensis* and activities of their detoxifying enzymes.** *J Pest Sci* 2020:1-14.
  48. Abd-El-Hallem AM, Hoogstrate SW, Schuurink RC: **A robust functional genomics approach to identify effector genes required for thrips (*Frankliniella occidentalis*) reproductive performance on tomato leaf discs.** *Front Plant Sci* 2018, **9**:1852.
  49. Schausberger P: **Herbivore-associated bacteria as potential mediators and modifiers of induced plant defense against spider mites and thrips.** *Front Plant Sci* 2018, **9**:1107.
  50. Pappas ML, Tavlaki G, Triantafyllou A, Broufas G: **Omnivore-herbivore interactions: thrips and whiteflies compete via the shared host plant.** *Sci Rep* 2018, **8**:3996.
  51. Vaello T, Pineda A, Marcos-García M: **Role of thrips omnivory and their aggregation pheromone on multitrophic interactions between sweet pepper plants, aphids, and hoverflies.** *Front Ecol Evol* 2019, **6**:240.
  52. Escobar-Bravo R, Ruijgrok J, Kim HK, Grosser K, Van Dam NM, Klinkhamer PG, Leiss KA: **Light intensity-mediated induction of trichome-associated allelochemicals increases resistance against thrips in tomato.** *Plant Cell Physiol* 2018, **59**:2462-2475.
  53. Escobar-Bravo R, Chen G, Kim HK, Grosser K, van Dam NM, Leiss KA, Klinkhamer PG: **Ultraviolet radiation exposure time and intensity modulate tomato resistance to herbivory through activation of jasmonic acid signaling.** *J Exp Bot* 2019, **70**:315-327.

54. Zeng G, Zhi J, Ye M, Yue W, Song J: **Inductive effects of exogenous calcium on the defense of kidney bean plants against *Frankliniella occidentalis* (Thysanoptera: Thripidae).** *Arthropod Plant Interact* 2020, **14**:473-480.
55. Chen G, Escobar-Bravo R, Kim HK, Leiss KA, Klinkhamer PG: **Induced resistance against western flower thrips by the *Pseudomonas syringae*-derived defense elicitors in tomato.** *Front Plant Sci* 2018, **2018**:1417.
56. Xiao Z, Jiang L, Chen X, Zhang Y, Defosse E, Hu F, Liu M, Rasmann S: **Earthworms suppress thrips attack on tomato plants by concomitantly modulating soil properties and plant chemistry.** *Soil Biol Biochem* 2019, **2019**:23-32.
57. Muvea AM, Subramanian S, Maniania NK, Poehling HM, Ekesi S, Meyhöfer R: **Endophytic colonization of onions induces resistance against viruliferous thrips and virus replication.** *Front Plant Sci* 2018, **9**:1785.
58. Pineda A, Kaplan I, Hannula SE, Ghanem W, Bezemer TM:
  - **Conditioning the soil microbiome through plant-soil feedbacks suppresses an aboveground insect pest.** *New Phytol* 2020, **226**:595-608.
59. Akbar W, Gowda A, Ahrens JE, Stelzer JW, Brown RS, Bollman SL, Greenplate JT, Gore J, Catchot A, Lorenz G: **First transgenic trait for control of plant bugs and thrips in cotton.** *Pest Manag Sci* 2019, **75**:867-877.
60. Huseth AS, D'Ambrosio DA, Yorke BT, Head GP, Kennedy GG:
  - **Novel mechanism of thrips suppression by Cry51Aa2. 834\_16 Bt toxin expressed in cotton.** *Pest Manag Sci* 2020, **76**:1492-1499.
61. D'Ambrosio DA, Kennedy GG, Huseth AS: **Feeding behavior of *Frankliniella fusca* on seedling cotton expressing Cry51Aa2. 834\_16 Bt toxin.** *Pest Manag Sci* 2020, **2020**:2781-2786.