

Viewpoint

Exploiting differences in the energy budget among C₄ subtypes to improve crop productivity

Summary

C₄ crops of agricultural importance all belong to the NADP-malic enzyme (ME) subtype, and this subtype has been the template for C_4 introductions into C_3 crops, like rice, to improve their productivity. However, the ATP cost for the C₄ cycle in both NADP-ME and NAD-ME subtypes accounts for > 40% of the total ATP requirement for CO₂ assimilation. These high ATP costs, and the associated need for intense cyclic electron transport and low intrinsic quantum yield Φ_{CO_2} , are major constraints in realizing strong improvements of canopy photosynthesis and crop productivity. Based on mathematical modelling, we propose a C_4 ideotype that utilizes low chloroplastic ATP requirements present in the nondomesticated phosphoeno/pyruvate carboxykinase (PEP-CK) subtype. The ideotype is a mixed form of NAD(P)-ME and PEP-CK types, requires no cyclic electron transport under low irradiances, and its theoretical Φ_{CO_2} is c. 25% higher than that of a C₄ crop type. Its cell-typespecific ATP and NADPH requirements can be fulfilled by local energy production. The ideotype is projected to have c. 10% yield advantage over NADP-ME-type crops and > 50% advantage over C₃ counterparts. The ideotype provides a unique (theoretical) case where Φ_{CO_2} could be improved, thereby paving a new avenue for improving photosynthesis in both C_3 and C_4 crops.

Introduction

Increasing yields of major food crops, like wheat and rice, to cope with increasing populations is a major challenge in agriculture, especially in the face of an accelerating climate change (Fischer *et al.*, 2014). Crop yield improvement by the Green Revolution in the 1960s has resulted almost exclusively from the increased harvest index by introducing (semi)dwarfing genes, which allowed more fertilizer input without the risk of having more lodging (Fischer *et al.*, 2014). Afterwards, crop yield potential has been improved progressively; for example, by optimizing morphological characteristics and exploiting heterosis (Yuan, 2017). Since the benefit from approaches such as improving harvest index may have reached a maximum value, further improvement of crop productivity has to



come from significantly improved crop photosynthesis competence, either by increasing leaf photosynthesis efficiency or by improving light interception, or by optimizing the profile of photosynthetic resources in the crop canopy (Zhu *et al.*, 2010).

Whereas conventional breeding may have selected photosynthesis-related traits that likely contributed partly to recent yield progress (Fischer et al., 2014), the natural variation in leaf photosynthesis above the check variety within a species is generally small (Gu et al., 2014). Therefore, several research consortia have aimed to introduce C₄ photosynthesis (as in maize) into C₃ crops (like rice) via genetic engineering (e.g. von Caemmerer et al., 2012; Schuler et al., 2016), and significant progress has already been made - see the recent reviews of Weber & Bar-Even (2019) and Ermakova et al. (2020). The rationale for these endeavours is the CO₂-concentrating mechanism (CCM) in C₄ leaves that largely suppresses photorespiration, thereby resulting in higher lightsaturated maximum leaf photosynthetic capacity A_{max} (see Supporting Information Table S1 for symbol definitions) in C₄ than C₃ leaves. It was hoped that rice productivity could be increased by 50% when initially conceiving the C_4 rice project (Mitchell & Sheehy, 2006). However, more recent analysis, using a crop model based on more detailed physiological parameters, showed that potential productivity of rice when grown under the current climatic condition could be increased by only 33-42% by introducing the complete maize-like C4 mechanism, and the advantage was even lower under future high-CO2 atmospheric conditions (Yin & Struik, 2017). This less-than-expected advantage, as also predicted by Bellasio & Farquhar (2019), has been explained by little gain of quantum yield of photosynthetic CO₂ assimilation Φ_{CO_2} in maize-C₄ relative to C₃ plants, because the photosynthetic rate of lower leaves in a full canopy is largely determined by Φ_{CO_2} rather than by A_{max} . Many studies (e.g. Day & Chalabi, 1988; Gu et al., 2014) showed that full-canopy photosynthesis generally depends more on Φ_{CO_2} than on A_{max} .

The small advantage of Φ_{CO_2} in maize relative to rice leaves is because the operation of CCM in maize leaves requires ATP and the production of ATP is limited under low light conditions prevalent in lower leaves of a canopy. The Calvin cycle for assimilating 1 mol CO₂ requires 2 mol NADPH and 3 mol ATP, whereas the C₄ cycle in maize additionally requires 2 mol ATP because of the regeneration of phosphoenolpyruvate (PEP) by pyruvate, phosphate dikinase (PPDK) in the C₄ cycle (Hatch, 1987). This means that the C_4 cycle accounts for *c*. 40% of the total ATP consumption (von Caemmerer & Furbank, 1999). As the generally believed stoichiometry of linear electron transport (LET) in chloroplasts gives an ATP : NADPH ratio of 1.5 (Allen, 2003), the ratio exactly matching the required ratio by the Calvin cycle, the ATP required for the C₄ cycle has to come from the cyclic electron transport (CET; Yin & Struik, 2012). In the C₄ cycle, bicarbonate ions, converted from CO₂, are fixed in mesophyll (M) cells by PEP

© 2020 The Authors

New Phytologist © 2020 New Phytologist Foundation

This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

carboxylase (PEPc) to produce oxaloacetate (OAA); OAA is quickly reduced to malate or converted to aspartate. These C₄ acids move to bundle-sheath (BS) cells, where they are decarboxylated to deliver CO_2 to Rubisco to start the Calvin cycle. There are different enzymes for decarboxylating C₄ acids in BS compartments, and C₄ species have been classified traditionally into three subtypes according to the decarboxylating enzyme: NADP-malic enzyme (ME) in chloroplasts, NAD-ME in mitochondria, and PEPcarboxykinase (CK) in the cytosol (Hatch, 1987; Fig. S1).

In NADP-ME and NAD-ME subtypes, decarboxylating malate into CO₂ produces pyruvate. Pyruvate moves directly (in the NADP-ME subtype) or via aminotransferase (in the NAD-ME subtype) to M cells, where pyruvate reacts to regenerate PEP by PPDK (Fig. S1a,b). Based on the NADPH and ATP requirement and generally estimated leakiness ϕ , the fraction for the CO₂ leakage from BS to M cells, Yin & Struik (2018) theoretically calculated that Φ_{CO_2} of the two ME subtypes under the atmospheric condition was c. 0.064 mol mol⁻¹ (Table 1), comparable to the measured Φ_{CO_2} (Ehleringer & Pearcy, 1983). These values are not much higher than Φ_{CO_2} of C₃ species (Ehleringer & Pearcy, 1983) if adjusted for reference temperature 25°C under the current atmospheric CO₂ conditions. This indicates that, at low irradiances, the gain from CCM in minimizing photorespiration is largely cancelled out by the extra ATP requirement for CCM. C₄ crops of agricultural importance for food and energy (maize, sorghum, millet, sugarcane, and *Miscanthus* × giganteus) all belong to the NADP-ME subtype (Hatch, 1987; Sage, 2016), and the subtype has been used as the template for developing C₄ rice (Karki et al., 2013).

In the PEP-CK subtype, a portion of OAA is reduced to malate, using NADPH from M chloroplasts, and its remaining portion moves, via aspartate, to the BS cytosol. The OAA in the BS cytosol is directly decarboxylated by PEP-CK while NAD-ME

simultaneously decarboxylates malate in the BS mitochondria (Hatch, 1987; Fig. S1c). Although this PEP-CK subtype requires additional NADPH to operate, it is the most efficient C₄ type in terms of ATP requirement from the chloroplast electron transport chain. First, only 1 mol ATP is required per mole of OAA decarboxylated by PEP-CK into CO₂ and PEP (Hatch, 1987; Kanai & Edwards, 1999), and the PEP returns directly to the M cytosol, thereby minimizing the use of PPDK that would need 2 mol ATP to regenerate 1 mol PEP. Second, the ATP required by PEP-CK can be exclusively produced by NADH oxidation in the respiratory chain of BS mitochondria associated with malate decarboxylation (Hatch, 1987; Burnell & Hatch, 1988). As the stoichiometry of ATP production per oxidation of NADH in mitochondria (the ATP : NADH ratio n) is believed to be either 3 (von Caemmerer & Furbank, 1999) or 2.5 (Kanai & Edwards, 1999) - much higher than the generally believed ATP : NADPH ratio of 1.5 for the LET in chloroplasts - the cost for additional NADPH in this subtype is actually overcompensated for by its involvement of the mitochondrial electron transport chain. The only extra chloroplastic ATP required in this PEP-CK subtype arises from the regeneration of PEP by PPDK from malate decarboxylation-derived pyruvate, which is 0.5 mol (if n = 3) or 0.572 mol (if n = 2.5) ATP per mole CO₂ assimilated (Yin & Struik, 2018). Therefore, the proportion x of the chloroplastic ATP consumed by the C_4 cycle is theoretically 0.15–0.17 in the PEP-CK subtype, considerably lower than the 0.4 for the NADP-ME and NAD-ME subtypes (Table 1). Based on the NADPH and ATP budget, and assuming the same proportions of leakiness and photorespiration, Yin & Struik (2018) theoretically calculated that the Φ_{CO_2} of the PEP-CK subtype is c. 0.080–0.081 mol mol⁻¹ (Table 1), c. 26% higher than the Φ_{CO_2} of the NADP-ME and NAD-ME subtypes. Such an advantage, if engineered into C₄ crops, would significantly increase canopy photosynthesis and,

Table 1 NADPH and ATP requirement from chloroplastic electron transport (mol mol⁻¹ CO₂) for the Calvin cycle (before '+' sign) and for the C₄ cycle (after '+' sign), and theoretical quantum yield of CO₂ assimilation (Φ_{CO_2} , mol mol⁻¹) for various (proposed) types of C₄ photosynthesis, using Φ_{CO_2} of C₃ photosynthesis under the nonphotorespiratory (NPR) condition as the reference.

			Minimum energy required from chloroplast ^a				
			NADPH	ATP	f _{CET}	Φ_{CO_2}	x
C ₃ (NPR)			2	3	0	0.100	na
Classical C ₄ subtypes	NADP-ME		2	3 + 2	0.5	0.064	0.41
	NAD-ME		2	3 + 2	0.5	0.064	0.41
	PEP-CK	<i>n</i> = 3	2 + 0.25	3 + 0.50	0.05	0.081	0.15
		<i>n</i> = 2.5	2 + 0.286	3 + 0.572	0.06	0.080	0.17
NH ₂ -balanced PEP-CK type ^b $n = 3$ n = 2.5		<i>n</i> = 3	2 + 0.5	3 + 0.0	0	0.075	0.00
		<i>n</i> = 2.5	2 + 0.5	3 + 0.25	0	0.075	0.09
C ₄ ideotype ^b		<i>n</i> = 3	2 + 0.36	3 + 0.55	0	0.079	0.16
		n = 2.5	2 + 0.4	3 + 0.6	0	0.078	0.17

x, the proportion of the chloroplastic ATP that is used to support the C_4 or CO_2 -concentrating mechanism cycle; na, not applicable.

n, stoichiometric coefficient for the number of ATP generated per NADH oxidation; f_{CET} , Φ_{CO_2} , and x in the last three columns were estimated assuming that all C₄ types had the same leakiness (0.16; Yin & Struik, 2012) and the same proportions for alternative energy sinks (see Supporting Information Table S2), to allow objective comparison of their intrinsic quantum yield.

^aThe word 'Minimum' is used because data in these columns do not include energy loss due to leakiness and energy requirements by alternative energy sinks. ^bNADPH and ATP requirements for these types are derived from discussion in the text; f_{CET}, fraction for cyclic electron transport. thus, crop yield. Indeed, the simulation of Yin & Struik (2017) showed that crop productivity can be increased almost linearly with decreasing ATP demands by the CCM.

However, literature-reported values of Φ_{CO_2} measured under ambient conditions are not significantly higher in the PEP-CK species than in C_4 species of the other subtypes, especially not when compared with the NADP-ME species (Ehleringer & Pearcy, 1983). The gaps between theoretically expected and experimentally measured Φ_{CO_2} in the PEP-CK species have not been explained yet. Here, we first propose hypotheses that could explain the small advantage of the PEP-CK subtype in Φ_{CO_2} over the other subtypes. On this basis, we describe a scheme for the energy budget in the PEP-CK subtype and extend the model of Yin & Struik (2012) to calculate the Φ_{CO_2} of this PEP-CK subtype. By exploring theoretical margins using this extended model, we propose a C₄ ideotype and assess whether this would have advantages over the current crop C₄ type (NADP-ME) at the canopy level. With this approach, we hope to open a new horizon by redesigning strategies to introduce the C₄-ness into C₃ crops, given that most attention so far has been paid to how to make C₄ enzymes and structures function and little attention has been paid to the C4 energetics (Ermakova et al., 2020).

Hypotheses

Hypothesis 1: flux of amino groups between mesophyll and bundle-sheath cells needs to be balanced

The classical PEP-CK subtype (Hatch, 1987; Burnell & Hatch, 1988) was defined assuming that the ATP required by PEP-CK could be exclusively produced by NADH oxidation in the respiratory chain of BS mitochondria associated with NAD-MEdependent malate decarboxylation. If n = 3, only 25% of the CO₂ from C₄-acid decarboxylation in BS cells is from the NAD-ME activity; and if n = 2.5, the NAD-ME-associated decarboxylation will account for 28.6% of the CO2 generated. This definition of the PEP-CK subtype did not consider the phosphate balance and the required amino (NH₂)-flux balance. Phosphate is generally believed to freely diffuse, and its availability is carefully regulated (Heldt et al., 1987). However, the NH2-flux balance between M and BS cells is highly relevant for both NAD-ME and PEP-CK subtypes. In the NAD-ME subtype, OAA reacts with glutamate (with NH₂) to form aspartate, which is transported to BS mitochondria where it is deaminated to release OAA. The OAA is then converted to malate, and the malate decarboxylation product, pyruvate, is converted to alanine, which is shuttled to the M cytosol; thereby, the NH₂ flux is balanced between M and BS cells (Fig. S1b). The direct OAA decarboxylation by PEP-CK cannot achieve such a balance because its product, PEP, returns directly to the M cells (Fig. S1c). This inability to achieve the balance by direct OAA decarboxylation is one of the reasons why the 'PEP-CK-only' C4 type has hardly been identified in nature (Furbank, 2011). The parallel running of malate decarboxylation via NAD-ME and direct OAA decarboxylation via PEP-CK only alleviates the imbalance in the PEP-CK subtype. The balance can be achieved only if the two decarboxylation pathways are equal in using the initial OAA (i.e. 50% each).

For such a scenario where NAD-ME and PEP-CK-dependent decarboxylation each deliver 50% of CO₂ to Rubisco, the required ATP per C₄ cycle would be 1.5, consisting of 0.5 ATP for PEP-CK activity in BS cells and 1.0 ATP for the regeneration of 0.5 mol PEP in M cells by PPDK. The oxidation of 0.5 NADH will provide 1.50 ATP (n = 3) or 1.25 ATP (n = 2.5), exactly or almost sufficient already to satisfy the required ATP for running the C₄ cycle. However, the LET for supplying the extra 0.5 NADPH for this scenario also produces a certain amount of ATP, which can be calculated as $(2 \times 0.5)(2 + f_{O})/h$, where $(2 + f_{O})/h$ is ATP produced per LET (f_{O} , fraction of electrons at plastoquinone that follow the Q cycle; *h*, proton : ATP ratio) (Yin & Struik, 2012). Assuming the most likely values $f_{\Omega} = 1$ and b = 4 (Yin & Struik, 2012), the LET would produce 0.75 ATP, which is more than enough to fulfil the shortfall of 0.25 ATP if n = 2.5. The '50% each' scenario was also proposed by Bräutigam et al. (2014) as the model for energy and NH₂ balance of the PEP-CK subtype. However, they overestimated the extra ATP requirement of this scenario because their analysis ignored that the NADH oxidation provides more ATP than required by the PEP-CK activity and that the LET associated with 0.5 NADPH also produces ATP. Here, we indicate that, for this scenario, the CO2 assimilation of the PEP-CK type would be NADPH limited (rather than ATP limited), and there is no need at all to engage the CET that is required for generating ATP in other C₄ subtypes. The theoretical Φ_{CO_2} calculated using the approach of Yin & Struik (2018) for this type is $0.075 \text{ mol mol}^{-1}$ (Table 1), which is somewhat lower than the theoretical Φ_{CO_2} (0.080–0.081 mol mol⁻¹) for the classical PEP-CK subtype but still higher than the measured Φ_{CO_2} of the PEP-CK species.

Hypothesis 2: PEP-CK species represent various mixtures of the classically defined PEP-CK component with NAD(P)-ME and/or with less efficient PEP-CK forms

Apparently, the aforementioned scenario of 50% each for two parallel decarboxylating pathways does not predict the measured $\Phi_{\rm CO_2},$ nor can it generate the required balance in the ATP : NADPH ratio. We hypothesize that there must be a third, higher ATP-requiring decarboxylating way (or even ways) involved in the PEP-CK subtype.

First, there might be additional NAD-ME or NADP-ME-based decarboxylation. Since there is hardly any NADP-ME activity in PEP-CK species (Hatch, 1987; Kanai & Edwards, 1999; Bräutigam, 2014; Koteyava *et al.*, 2015), this third pathway likely involves NAD-ME. The latter enzyme is already involved in the classical PEP-CK subtype (associated with providing ATP for PEP-CK activity); but we hypothesize that the additional NAD-ME activity functions as it occurs in the classical NAD-ME subtype, in which the NH₂-flux is balanced (see earlier). As such, on the one hand, NAD-ME provides ATP for PEP-CK activity and, on the other hand, functions independently with an increased ATP cost of CCM. However, the possibility that the third decarboxylation is NADP-ME dependent or a mixed NAD-ME and NADP-ME type cannot be ruled out, as NADP-ME has been found in some PEP-CK species (Sonawane *et al.*, 2018). Because the NADP-ME itself

does not involve any transamination, the $\rm NH_2$ -flux balance can also be guaranteed.

Second, the PEP produced by PEP-CK in the BS cells is so far believed to move directly to the M cells (Fig. S1c). However, the fate of PEP is uncertain. The PEP can first be converted in BS cells to pyruvate, which moves via alanine aminotransferase to the M cells (Smith & Woolhouse, 1983), where pyruvate is converted to PEP by PPDK. The conversion of PEP to pyruvate in the BS cells could occur via pyruvate kinase, releasing one ATP per PEP converted (Schuler *et al.*, 2016). Thus, the ATP balance for the C₄ cycle is neutral in the BS cells as the one ATP required for PEP-CK is equal to the one ATP released by pyruvate kinase. For this case, Φ_{CO_2} would be the same as for the first case where PEP-CK is mixed with NAD-ME or/and NADP-ME.

Third, the conversion of PEP to pyruvate in BS cells could also occur via PEP phosphatase (Smith & Woolhouse, 1983). This enzyme releases orthophosphate, resulting in a loss of chemical energy to entropy (Bräutigam *et al.*, 2018), in contrast to pyruvate kinase that conserves energy. Therefore, this case would predict an even lower Φ_{CO_2} in the PEP-CK subtype than in the NADP-ME and NAD-ME subtypes, and works probably for the PEP-CK species where Φ_{CO_2} is intrinsically low, or/and where only low activity of pyruvate kinase is detectable in BS cells (Kanai & Edwards, 1999).

The PEP in BS cells might also be converted via enolase and phosphoglyceromutase to 3-phosphoglycerate (3-PGA), which could move to the M cells and be converted back to PEP via the same enzymes (Huber & Edwards, 1975). This reaction may be engaged when the conversion of PEP to pyruvate cannot account for all of the carbon flux through the C₄ cycle (Smith & Woolhouse, 1983). However, as this reaction does not involve any energy gain or loss, it would predict the same Φ_{CO_2} as the classical PEP-CK type. Furthermore, the conversion to 3-PGA cannot contribute to maintain the NH2-flux balance. So, this mechanism will not be considered further. By contrast, the aforementioned second and third mechanisms involving pyruvate kinase and PEP phosphatase, respectively, not only can bring the theoretical Φ_{CO_2} of the PEP-CK subtype equal to that of the ME subtypes but can also lower the BS : total ATP requirement ratio (compared with the previously discussed 'pure PEP-CK' type) and maintain the NH2-flux balance between M and BS cells. The high BS: total ATP requirement ratio (3:4, Yin & Struik, 2018) and the NH₂-flux imbalance (Furbank, 2011) were argued to explain why the 'pure PEP-CK' type was hardly found in nature. Our hypothesis suggests that there are multiple scenarios for pure PEP-CK and hints that a pure PEP-CK type, if involving pyruvate kinase or PEP phosphatase, may actually exist.

Model for the mixed PEP-CK form

A scheme for quantitative proportions of possible decarboxylating pathways in the PEP-CK subtype, which incorporates both Hypothesis 1 and Hypothesis 2, is provided in Fig. 1. This scheme both fulfils energetic demands and guarantees the NH_2 -flux balance. The value of parameter *a* (the proportion of the initial OAA that is reduced to malate in M chloroplasts to ultimately drive

mitochondrial electron transport in BS cells and produce ATP) is a crucial parameter for this mixed type. Its value has a strong impact on the cellular NADPH : ATP production ratio, and, thereby, on the required fraction for cyclic electron transport f_{CET} . Both parameters, *a* and f_{CET} , are hard to measure experimentally. We developed equations for calculating these two parameters using Φ_{CO_2} that can be easily measured. These equations and their derivations are given in Notes S1.

Note that photorespiration may be higher in species of the NAD-ME or PEP-CK subtypes than in the NADP-ME subtype, because of larger amounts of photosystem II (PSII) in BS cells (thereby relatively more oxygen (O₂) evolution as a result of LET), and because of decarboxylation taking place either in mitochondria or in cytosol (thereby, a higher chance for the released CO₂ to escape from the refixation by Rubisco in BS chloroplasts, the organelle where decarboxylation occurs in the NADP-ME subtype). Quantifying to what extent photorespiration is actually higher in the NAD-ME, PEP-CK, or mixed types relative to the exclusive NADP-ME type is beyond the scope of this paper to quantify. To bypass this complication, we used our unpublished data from four PEP-CK species on their Φ_{CO_2} measured using a gas mixture of 2% O₂ and 1000 µmol mol⁻¹ CO₂; that is, under conditions in which photorespiration is conceivably negligible.

Values of Φ_{CO_2} (Table 2) were all lower than the theoretical value 0.080-0.081 mol mol⁻¹ in Table 1 for the classically defined PEP-CK subtype. But they were slightly higher than the values $(0.060-0.067 \text{ mol mol}^{-1})$ reported in the literature for this subtype (Ehleringer & Pearcy, 1983), suggesting that the gas mixture indeed prevented the occurrence of photorespiration. Since they are not lower than 0.064 mol mol⁻¹, the theoretical $\Phi_{\rm CO_2}$ for the ME subtypes, it is unlikely that PEP-phosphatasebased conversion of PEP into pyruvate in BS cells played a significant role, although we cannot rule it out. For simplicity, we set parameter a'' = 0 (see Fig. 1). Based on this value and other input parameter values that are considered conserved (Table S2), the calculated a depended slightly on the value of n and varied from 0 to 0.16, all lower than the minimum value for this parameter of 0.25 defining the classical PEP-CK subtype. This means that, in order to meet the requirement of NH2-flux balance, the fraction of OAA following the direct OAA decarboxylation by PEP-CK was also from 0 to 0.16. The remaining dominant fraction (> 0.68) was to perform either NADP-ME and NAD-ME-based decarboxylation, or the PEP-CK decarboxylation involving pyruvate kinase. Though we were unable to quantitatively separate the three (but see later section), our calculations suggested that PEP-CK species might have substantial NAD(P)-ME activity. Bräutigam et al. (2014) did show that the NAD-ME activity was 35% higher than the PEP-CK activity in the PEP-CK species Panicum maximum. However, the current literature data on the activities of the three decarboxylating enzymes in the PEP-CK species (e.g. Hatch, 1987; Kanai & Edwards, 1999; Koteyava et al., 2015; Bräutigam et al., 2018; Sonawane et al., 2018) suggest that generally PEP-CK is still dominant over the other two decarboxylating enzymes. So, most likely, the PEP-CK decarboxylation involving pyruvate kinase accounts for the third source of CO₂ delivered to Rubisco.

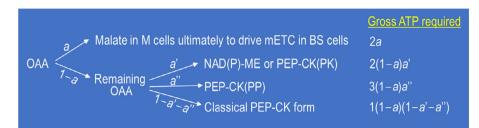


Fig. 1 A scheme illustrating multiple decarboxylating pathways of C₄photosynthesis. A fraction of OAA produced by phosphoeno/pyruvate (PEP) carboxylation in the mesophyll (M) cells, *a*, is reduced to malate, which will move to the bundle-sheath (BS) mitochondria where malate is decarboxylated, generating NADH that will drive the mitochondrial electron transport chain (mETC) and produce ATP. The remaining fraction of OAA in the M cells, (1 - a), follows various possible pathways: a fraction of this part of OAA (*a'*) follows the NAD-malic enzyme (ME) or NADP-ME decarboxylating pathway or the PEP-carboxykinase (CK) pathway where the PEP generated will be converted to pyruvate in the BS cells by PEP phosphatase (PP) – the pathway denoted as PEP-CK (PK); another fraction (*a''*) follows the PEP-core at the PEP generated from the aforementioned NADH-associated mETC. As discussed in the main text, the NH₂-flux balance associated with the PEP-CK type would require that *a* = (1 - *a*)(1 - *a'* - *a''*). The gross ATP requirements for each decarboxylating form can be formulated according to these fractions, as indicated in the figure, where NAD-ME, NADP-ME, and PEP-CK(PK) all require two ATP per decarboxylation (for PEP regeneration by pyruvate, phosphate dikinase), and therefore are lumped in the same category (but elsewhere in the text, and in Table 3 and Fig. 3, the fraction of this category for NADP-ME is denoted as *b*). Summing individual gross ATP requirements up, deducting the ATP production by the NADH oxidation, would give the net ATP requirement for the whole C₄ cycle. Combining this with the NH₂-balance equation *a* = (1 - *a*) (1 - *a'* - *a''*) and the since of the ATP requirement for the whole C₄ cycle as 2 + a'' - (n + 1 + a'')a in this mixed type.

Table 2 Values of parameters *a* and f_{CET} , calculated from unpublished estimates of Φ_{CO_2} (SEs of estimates in parentheses) from measurements for four phosphoeno/pyruvate carboxykinase species.

	Фар (1995)	r	<i>n</i> = 3		n = 2.5	
Species	$\Phi_{\text{CO}_2(\text{NPR})}$ (mol mol ⁻¹)	a	!	f _{CET}	а	$f_{\rm CET}$
Chloris gayana	0.068 (0.003)	C	0.01	0.509	0.01	0.508
<i>Melinis minutiflora</i>	0.065 (0.004)	C	00.0	0.517	0.00	0.517
Panicum maximum	0.072 (0.003)	C).13	0.386	0.16	0.373
Spartina gracilis	0.069 (0.004)	C	0.05	0.479	0.05	0.475

The calculation assumed that ϕ (leakiness) = 0.16; n = ATP : NADP ratio. *a*, proportion of the initial oxaloacetate for converting to malate in mesophyll cells that ultimately leads to mitochondrial electron transport in bundlesheath cells to produce ATP; f_{CET} , fraction for cyclic electron transport. Measurements for estimating $\Phi_{CO_2(NPR)}$ were conducted on leaves of four replicated plants for each species, using a gas mixture of 2% oxygen combined with 1000 µmol mol⁻¹ CO₂, with which a nearly nonphotorespiratory (NPR) condition is conceivably achieved.

Exploring theoretical margins to design an ideotype of $C_4\ photosynthesis$

The model described in Notes S1 for calculating parameters *a* and f_{CET} can be used to explore theoretical margins for the optimum combination of *a* and f_{CET} that gives the maximum Φ_{CO_2} within the physiologically relevant range of *a* and f_{CET} . This is illustrated in Fig. 2, where the physiologically relevant range is marked by the red arrow. The lowest Φ_{CO_2} of this range is defined by a = 0, which means that the mixed type would become the classically defined NADP-ME or NAD-ME subtype. The maximum Φ_{CO_2} of this range is defined by $f_{CET} = 0$, which gives the most efficient mixed type having the maximum Φ_{CO_2} , and the equivalent value of parameter $a \approx 0.36$ if n = 3 (Fig. 2) or $a \approx 0.40$ if n = 2.5. Beyond this point, f_{CET} would become negative. The NH₂-flux

balanced PEP-CK type (where a = 0.5) discussed earlier actually goes beyond this threshold that would need a negative $f_{CET} \approx -0.44$ (if n = 3) or $f_{CET} \approx -0.26$ (if n = 2.5). The logic for negative f_{CET} is that, with parameter *a* becoming higher, NADPH in the M cell is overused for reducing OAA to malate such that NADH oxidation and LET are both high, overproducing ATP

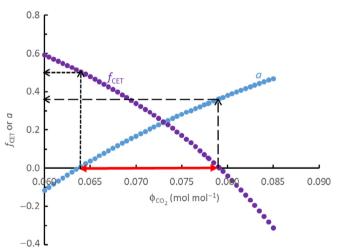


Fig. 2 Exploring theoretical margins of the proposed mixed phosphoenol pyruvate carboxykinase type to identify the optimum combination of parameters *a* (proportion of OAA reduced to malate in mesophyll cells for malate decarboxylation via NAD-malic enzyme (ME) to drive mitochondrial electron transport in bundle-sheath cells) and f_{CET} (fraction for cyclic electron transport) that maximizes Φ_{CO_2} within the physiologically relevant range of *a* and f_{CET} indicated by the thick red arrow; that is, the range when both are ≥ 0 . The maximum Φ_{CO_2} is achieved when $f_{CET} = 0$, and its equivalent value of parameter *a* in this illustration (where *n*, the ATP : NADH ratio, is set to 3.0) is *c*. 0.363, indicated by long-dashed black arrows. When a = 0, $f_{CET} \approx 0.5$ and $\Phi_{CO_2} \approx 0.064$ mol mol⁻¹, as indicated by short-dashed black arrows; this case corresponds to the classical NADP-ME subtype or NAD-ME subtype.

that would need a negative f_{CET} to balance the NADPH : ATP ratio. As a negative f_{CET} is obviously physiologically impossible, the NH₂-flux balanced PEP-CK type (where a = 0.5) discussed earlier should not exist in nature.

The maximum Φ_{CO_2} at $f_{CET} = 0$ was 0.079 mol mol⁻¹ (if n = 3) or 0.078 mol mol⁻¹ (n = 2.5), only slightly lower than Φ_{CO_2} of 0.080–0.081 mol mol⁻¹, the value calculated for the classically defined PEP-CK subtype (Table 1). However, as discussed earlier, the classically defined PEP-CK subtype does not meet the requirement for a balance of the NH₂-flux between M and BS cells. As a result, unlike the classically defined NADP-ME or NAD-ME subtypes (which are special cases when a = 0 in Fig. 2), the classically defined PEP-CK subtype cannot be a special case along the curves shown in Fig. 2. As the type with $f_{CET} = 0$ (Fig. 2) yields the highest Φ_{CO_2} , we shall call it the C₄ ideotype. This ideotype, potentially as a mixture of three C₄ subtypes, guarantees a balanced NH₂-flux and a balanced NADPH : ATP ratio as required by C₃ and C₄ cycles.

Can energy supply satisfy cell-type-specific energy demand in the designed ideotype?

It is necessary to examine if NADPH and ATP requirements can be met within M and BS cell types. To that end, we extended the celltype-specific models of Yin & Struik (2018) for light absorption, energy production, and requirement for the mixed C₄ type as defined in Fig. 1 (Notes S2). We run this extended NADPH and ATP production model for the C₄ ideotype with a = 0.36 (if n = 3) or a = 0.40 (if n = 2.5) and a'' = 0, using four sets of the three input parameters: leaf Chl content [CHL], fraction of [CHL] in BS cells, and fraction of photosystem I (PSI) in BS cells. These four sets of inputs were taken from measurements of Ghannoum *et al.* (2005) for two NADP-ME (sets I and II) and two NAD-ME (sets III and IV) species. The model can identify the physiologically relevant fraction of PSII in BS cells (Yin & Struik, 2018). Other output parameters relevant for our analysis here are the fraction of ATP production in BS cells and the fraction of NADPH production in BS cells. None of these three output parameters depended on the value of *n*, so they are given in a single value for each input set (Table 3). The model identified that the physiologically relevant fraction of PSII in BS cells had a single value and was equal to the input fraction of PSI in BS cells. This is because the proposed ideotype requires no CET; so, relative amounts of PSII in either cell type should agree with the relative amounts of PSI to perform LET only. For the same reason, the predicted fraction of ATP production in BS cells (Table 3), as LET produces a fixed ATP : NADPH ratio.

We then analysed whether ATP and NADPH supply satisfies cell-type-specific energy requirement in the C₄ ideotype, following the same method as used by Yin & Struik (2018); that is, by matching the fraction of ATP (NADPH) supply in BS cells with the fraction of ATP (NADPH) demand in BS cells. The formulae for the cell-type-specific demands are described in Table S3 (where *a*" should be set to zero for the ideotype). This matching gives rise to what fraction of the 3-PGA reduction in BS cells would be needed. Table S3 shows that the cell-type-specific ATP demand does not depend on parameter b (fraction of the second category of decarboxylation in Fig. 1 that is via NADP-ME) whereas the NADPH demand does. So, we first calculated the required fraction of the 3-PGA reduction in BS cells to have the ATP supply and demand in balance, indicated by the variable γ_{atp} . It varied from 0.35 to 0.73 when n = 3 or from 0.29 to 0.67 when n = 2.5(Table 3). The higher γ_{atp} when *n* is higher was because higher ATP supply from NADH oxidation in BS mitochondria can support a higher fraction of 3-PGA reduction in BS chloroplasts; and the transfer of ATP can be realized given available intracellular ATP shuttles (Shameer et al., 2019). The significantly higher fraction of

Table 3 Modelled output parameter values of the proposed C₄ ideotype assuming the ATP : NADH ratio n = 3 (before '/' sign) or = 2.5 (after '/' sign), using four sets (I–IV) of three input-parameter values.

Parameter		I	П	Ш	IV
Input	[CHL] (μ mol m ⁻²)	579	464	424	425
	Fraction of [CHL] in BS	0.33	0.38	0.60	0.59
	Fraction of PSI in BS	0.37	0.39	0.24	0.46
Output	Fraction of PSII in BS (α)	0.37	0.39	0.24	0.46
	Fraction of ATP produced in BS ^a	0.29	0.32	0.48	0.47
	Fraction of NADPH produced in BS ^a	0.29	0.32	0.48	0.47
	γatp	0.35/0.29	0.42/0.36	0.73/0.67	0.71/0.65
	b ^c	-0.04/-0.65	0.13/-0.43	0.79/0.48	0.75/0.44
	NADP-ME% in total decarboxylation ^d	-1.0/-13.2	3.4/-8.7	21.5/9.7	20.6/8.8
	γ_{nadph} (if $b = 0$) ^e	0.35/0.36	0.40/0.41	0.61/0.61	0.60/0.60

^aCell-type-specific production of ATP and NADPH quantified as the bundle-sheath (BS) : total ratio.

^b_{7 atp}, fraction of 3-phosphoglycerate (3-PGA) reduction in BS cells inferred from the balance that the BS : total ratio in ATP production is equal to the BS : total ratio in ATP demand.

^c*b*, the fraction of the second decarboxylation category (see Fig. 1) that is via NADP-malic enzyme (ME), inferred from assuming that γ_{nadph} equals γ_{atp} . ^dPercentage of the NADP-ME pathway accounting for the total decarboxylation, estimated as $[1 - 2a - (1 - a)a^n]b \times 100$ (see Supporting Information Notes S2), where a = 0.36 (if n = 3) or 0.40 (if n = 2.5), and $a^n = 0$ (see the text).

^eYnadph, fraction of 3-PGA reduction in BS cells inferred from the balance that the BS : total ratio in NADPH production is equal to the BS : total ratio in NADPH demand.

3-PGA reduction in BS cells in sets III and IV than in sets I and II was due to higher fractions of [CHL] in BS cells in sets III and IV, which enabled the capture of relatively more light energy and therefore to produce relatively more ATP in BS cells (Table 3).

Assuming that γ_{nadph} (fraction of the 3-PGA reduction in BS cells to have the NADPH supply and demand in balance) equalled $\gamma_{\rm atp}$, we were able to calculate *b*, which varied from -0.04 to 0.79 when n = 3 or from -0.65 to 0.48 when n = 2.5, and the resulting percentage of the NADP-ME pathway accounting for the total decarboxylation was also calculated (Table 3). The higher b when n = 3 was because this *n* resulted in higher γ_{atp} , which, when setting to be γ_{nadph} , would result in higher requirements for shuttling malate from M to BS cells by the NADP-ME-dependent pathway. Negative values of b and NADP-ME%, as occurred for sets I and II (Table 3), are physiologically impossible; their occurrence merely suggests that the input parameter value for the fraction of [CHL] in BS cells in set I and probably also in set II was not sufficiently high to realize the proposed C_4 ideotype. Assuming b = 0, we recalculated γ_{nadph} . Compared with γ_{atp} , the γ_{nadph} obtained was higher in cases where b was earlier found to be less than zero, but lower in cases where b was greater than zero (Table 3). 3-PGA as the product of RuBP carboxylation is produced in BS cells. Its reduction involves an intercellular shuttle (Hatch, 1987), during which 3-PGA is first transformed to 1,3-bisphosphoglycerate by phosphoglycerate kinase (the step that requires ATP) and then reduced to triose phosphate by glyceraldehyde 3-phosphate dehydrogenase (the step that requires NADPH). Higher γ_{atp} than γ_{nadph} , as calculated for sets III and IV, indicates the possibility that, relative to the second step, the first step occurs more in BS than in M cells. However, it is impossible to reconcile with a higher γ_{nadph} than $\gamma_{\rm atp}$, as calculated mostly for sets I and II, again suggesting that the proposed C₄ ideotype cannot be achieved with a low [CHL] fraction in BS cells.

Our model analysis herein suggests that cellular energy production can satisfy cell-type-specific ATP and NADPH requirement in the C_4 ideotype, provided that the fraction [CHL] in BS cells is greater than *c*. 0.4 and the proportion of 3-PGA reduction in BS cells and, optionally, the proportion of NADP-ME operation can adjust accordingly. The latter suggests an approach to estimate a maximum NADP-ME% in total decarboxylation (Table 3).

Advantages of the C $_4$ ideotype over the NADP-ME type at the canopy level of a crop

Finally, we evaluated whether the proposed C_4 ideotype can indeed have advantages over the current crop C_4 type (NADP-ME subtype) in a canopy and crop setting. To evaluate that, we run the crop model GECROS (Yin & Struik, 2017), for which we revised its C_4 submodel based on the algorithms developed here for the proposed C_4 ideotype (see Notes S3). We set the fraction of PSII in BS cells $\alpha = 0.10$ for the crop C_4 type as observed in maize and $\alpha = 0.40$ for the C_4 ideotype, the general value for the PEP-CK subtype (von Caemmerer & Furbank, 1999). We used the 31 yr (1980–2010) weather data of the International Rice Research Institute (in the Philippines), with GECROS input crop parameters as described by Yin & Struik (2017).

The simulated 31 yr average advantage of the crop type C_4 (NADP-ME) over the C3 rice in radiation use efficiency was 36.3%, and the advantage of the proposed C_4 ideotype was c. 50% when n = 3 and c. 48% when n = 2.5 (Table 4). Simulated advantages in total biomass all became somewhat higher, because an increased leaf area index was simulated as a consequence of the increased photosynthesis or radiation use efficiency, resulting in somewhat higher light interception during growing seasons. The advantage of the crop C₄ type was 42%, whereas that of the proposed C_4 ideotype was always >50% (Table 4). The required increase of α in the ideotype relative to the NADP-ME subtype is expected to increase photorespiration, but the impact of varying α on simulated advantages was small (results not shown). That was probably because, in the simulation, we assumed that the BS conductance (that determines the CO_2 leakage) in the proposed C_4 ideotype was as small as that found for the NADP-ME species, where decarboxylation occurs predominantly in chloroplasts. Whether or not this conductance becomes higher in the NAD-ME (decarboxylation in mitochondria) subtype and the PEP-CK subtype (decarboxylation in cytosol and mitochondria) is an important subject to investigate, although PEP-CK has been expressed to localize in chloroplasts of transgenic rice plants (Suzuki et al., 2000).

Concluding remarks

 C_4 crops of agricultural importance all belong to the NADP-ME subtype. Whether there is any causality behind this or whether it is just the coincidence of domestication is an interesting question to investigate. However, operating the CCM cycle in either NADP-ME or NAD-ME subtypes is energetically expensive, accounting for at least 40% of total ATP cost for CO₂ assimilation. Yin & Struik (2017) showed that the crop C₄ type, if being introduced into C₃ crops, would not be able to increase yield by 50%, the initially expected yield advantage of C₄ rice (Mitchell & Sheehy, 2006). We therefore proposed a C₄ ideotype by exploring theoretically lower energy requirements in the nondomesticated PEP-CK subtype.

Table 4 Simulated advantages (%) of the current crop C_4 type (the NADP-malic enzyme (ME) subtype) and the proposed C_4 ideotype over the C_3 rice in radiation use efficiency and total biomass production.

		Radiation use efficiency	Total biomass
$C_3 \text{ rice} C_4 \qquad \text{NADP-ME } C_4 type (\alpha = 0.1)$		2.95 g DM MJ ⁻¹ PAR 36.3%	22.6 t DM ha ⁻¹ 42.0%
	C_4 ideotype ($\alpha = 0.4$)	49.7%/47.5%	55.2%/52.7%

PAR, photosynthetically active radiation.

Radiation use efficiency is the ratio of total biomass to season-long accumulated PAR interception.

 α , fraction of photosystem II in bundle-sheath cells.

For the C₄ ideotype, the percentages before and after the '/' sign refer to the simulated advantages using n = 3 or n = 2.5, respectively (n, the number of ATP formed per NADH oxidation; see the text).

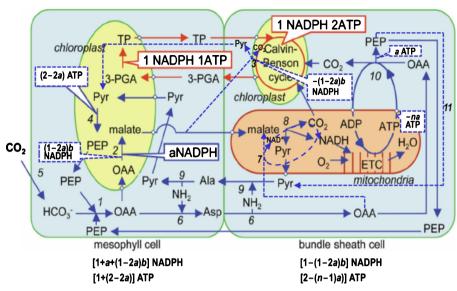


Fig. 3 The proposed mixed type of C₄ photosynthesis, and its NADPH and ATP requirements per CO₂ assimilation in mesophyll (M) and bundle-sheath (BS) cells. The red arrows show the Calvin cycle and its cell-type-specific NADPH and ATP requirements; the blue arrows show the C_4 cycle, where the full arrows show the cycle in the classically defined phosphoenol pyruvate (PEP)-carboxykinase (CK) subtype (see Supporting Information Fig. S1c; reprinted by permission from Springer Nature, Ishikawa et al., 2016) and the dashed ones show the new components added to form the mixed type. For illustrative simplicity, it is assumed that there is no leakiness, that the 3-phosphoglycerate (3-PGA) reduction of the Calvin cycle occurs in 50% in each of the two cell types, and that the PEP-CK pathway where the PEP generated is converted to pyruvate in the BS cells possibly by PEP phosphatase (PP) – the pathway denoted as PEP-CK(PP) in Fig. 1 – does not occur (for detailed algorithms for energy requirement without these assumptions, see Table S3). The coefficient a refers to the fraction of OAA produced by PEP carboxylation in the M cells that is reduced to malate moving to the BS mitochondria where malate decarboxylation generates NADH that will drive mitochondrial electron transport to produce ATP. This produces na ATP (where n is the ATP : NADH ratio). The coefficient b is the fraction of the second decarboxylation category defined in Fig. 1 that belongs to the NADP-malic enzyme (ME) type. If a = 0 and b = 1, then the mixed type becomes the NADP-ME subtype (see Fig. S1a); if a = 0 and b = 0, then the mixed type becomes the NAD-ME subtype (see Fig. S1b). Another special case if a = 0 and b = 0 is a hypothetical 'pure' PEP-CK type, where the PEP generated will be converted to pyruvate in the BS cells by pyruvate kinase – the pathway denoted as PEP-CK (PK) in Fig. 1. Numbers represent enzymes involved in the C₄ cycle: 1, PEP carboxylase (PEPc); 2, NADP-malate dehydrogenase (NADP-MDH); 3, NADP-ME; 4, pyruvate, phosphate dikinase (PPDK); 5, carbonic anhydrase (CA); 6, aspartate aminotransferase (AspAT); 7, NAD-malate dehydrogenase (NAD-MDH); 8, NAD-ME; 9, alanine aminotransferase (AlaAT); 10, PEP-CK; 11, pyruvate kinase (PK). Ala, alanine; Asp, aspartate; ETC, electron transport chain; Pyr, pyruvate; TP, triose phosphate.

The ideotype is the special form of the general mixed NAD(P)-ME and PEP-CK (Fig. 1) that results in the maximum Φ_{CO_2} (Fig. 2). The general mixed type considers the balance of the NH₂ groups between M and BS cells and the balance of NADPH and ATP as the metabolically required ratio. The classically defined PEP-CK subtype is not a special form of our general type, because it does not satisfy the first balance; neither is the NH₂-flux balanced type discussed earlier, as it does not satisfy the second balance. However, the other two classical subtypes are special cases of the general type. As species traditionally classified as NADP-ME or NAD-ME types often have some PEP-CK (Furbank, 2011), which can even be induced to increase by environmental changes (e.g. Sales *et al.*, 2018), our general type may be considered as a unified scheme of C₄ subtypes (Fig. 3).

The proposed C_4 ideotype does not need CET under low irradiances; so, probably few modifications of electron transport machinery are required because C_3 photosynthesis involves little CET. Furthermore, the PEP-CK is present in C_3 plants although it is responsible for nonphotosynthetic functions (Leegood & Walker, 2003), and genes encoding the evolution of C_4 PEP-CK have been reported (Christin *et al.*, 2009). However, the required engagement of multiple biochemical pathways (Fig. 3) might pose a challenge to engineering. Moreover, the required proportion of [CHL] in BS cells needs to be higher than that found in current C_4 crop species, and the vein spacing in C_3 leaves may not allow the housing of sufficient chloroplasts in BS cells (Karki *et al.*, 2013; Ermakova *et al.*, 2020). Wang *et al.* (2017) reported that transgenic rice lines expressing *Golden-like* transcription factor genes from maize displayed increased numbers and volumes of differentiated chloroplasts, and accumulated more [CHL], in BS and mestome sheath cells of the leaf. As long as the required [CHL] in BS cells can be reached, there are a range of options to achieve cellular energy balance of both ATP and NADPH; for example, via adjusted proportion of 3-PGA reduction in two cell types and adjusted engagement of the NADP-ME pathway (Table 3).

The required value of parameter *a* (proportion of OAA reduced to malate in M cells for malate decarboxylation via NAD-ME to drive mitochondrial electron transport) for the C₄ ideotype was *c*. 0.36 or 0.40. This is higher than the 0.25 or 0.286 required for the classically defined PEP-CK subtype (Hatch, 1987; Kanai & Edwards, 1999; Ishikawa *et al.*, 2016). As stated, the classical PEP-CK type does not meet the NH₂ balance and, thus, should not exist in nature. We do not know yet if there are C₄ species in nature that are physiologically the same as, or close to, the defined C₄ ideotype. Carnal *et al.* (1993) reported that maximum rates of malate decarboxylation via NAD-ME substantially exceeded the

minimum rates necessary for providing ATP for cytosolic OAA decarboxylation by PEP-CK. This suggests that PEP-CK species having high proportions (close to 0.36 or 0.40) of OAA reduced to malate in M cells for malate decarboxylation via NAD-ME may actually exist, although they may not necessarily have a high Φ_{CO_2} . Reasons for the low Φ_{CO_2} in PEP-CK species need to be examined in conjunction with our aforementioned hypotheses.

Photosynthetic Φ_{CO_2} is known to be recalcitrant to genetic improvement in both C_3 and C_4 types. However, canopy photosynthesis after canopy closure generally depends more on Φ_{CO_2} than on A_{max} (Day & Chalabi, 1988; Gu *et al.*, 2014), especially given recent molecular evidence that electron transport is a major limitation to photosynthesis in C_4 plants (Ermakova *et al.*, 2019). Our viewpoints provide a (theoretical) case where Φ_{CO_2} could be improved, thereby potentially paving a new way of redesigning and supercharging photosynthesis. If this is proven either from exploiting natural variations or by a synthetic biology approach, it would provide a new C_4 template not only for being introduced into C_3 crops but also for improving current C_4 crops.

Author contributions

XY conceived the viewpoints and designed the modelling. XY wrote the draft and finalized it with significant input from PCS.

ORCID

Paul C. Struik (D) https://orcid.org/0000-0003-2196-547X Xinyou Yin (D) https://orcid.org/0000-0001-8273-8022

Xinyou Yin* 🝺 and Paul C. Struik 🕩

Centre for Crop Systems Analysis, Department of Plant Sciences, Wageningen University & Research, PO Box 430, Wageningen, 6700 AK, the Netherlands (*Author for correspondence: email Xinyou.Yin@wur.nl)

References

- Allen JF. 2003. Cyclic, pseudocyclic and noncyclic photophosphorylation: new links in the chain. *Trends in Plant Science* 8: 15–19.
- Bellasio C, Farquhar GD. 2019. A leaf-level biochemical model simulating the introduction of C_2 and C_4 photosynthesis in C_3 rice: gains, losses and metabolite fluxes. *New Phytologist* 223: 150–166.
- Bräutigam A, Schliesky S, Külahoglu C, Osborne CP, Weber APM. 2014. Towards an integrative model of C₄ photosynthetic subtypes: insights from comparative transcriptome analysis of NAD-ME, NADP-ME, and PEP-CK C₄ species. *Journal of Experimental Botany* 65: 3579–3593.
- Bräutigam A, Schlüter U, Lundgren MR, Flachbart S, Ebenhöh O, Schönknecht G, Christin PA, Bleuler S, Droz JM, Osborne CP *et al.* 2018. Biochemical mechanisms driving rapid fluxes in C₄ photosynthesis. *BioRxiv*. doi: 10.1101/ 387431.
- **Burnell JN, Hatch MD. 1988.** Photosynthesis in phosphoenolpyruvate carboxykinase-type C₄ plants: pathways of C₄ acid decarboxylation in bundle sheath cells of *Urochloa panicoides. Archives of Biochemistry and Biophysics* **260**: 187–199.
- Carnal NW, Agostino A, Hatch MD. 1993. Photosynthesis in phospho*enol*pyruvate carboxykinase-type C_4 plants: mechanism and regulation of

- Christin P-A, Petitpierre B, Salamin N, Büchi L, Besnard G. 2009. Evolution of C₄ phospho*enol*pyruvate carboxykinase in grasses, from genotype to phenotype. *Molecular Biology and Evolution* 26: 357–365.
- Day W, Chalabi ZS. 1988. Use of models to investigate the link between the modification of photosynthetic characteristics and improved crop yields. *Plant Physiology and Biochemistry* 26: 511–518.
- **Ehleringer J, Pearcy RW. 1983.** Variation in quantum yield for CO_2 uptake among C_3 and C_4 plants. *Plant Physiology* 73: 555–559.
- Ermakova M, Danila FR, Furbank RT, von Caemmerer S. 2020. On the road to C₄ rice: advances and perspectives. *The Plant Journal* 101: 940–950.
- Ermakova M, Lopez-Calcagno PE, Raines CA, Furbank RT, von Caemmerer S. 2019. Overexpression of the Rieske FeS protein of the Cytochrome *b*₆*f* complex increases C₄ photosynthesis in *Setaria viridis. Communications Biology* 2: e314.
- Fischer RA, Byerlee D, Edmeades GO. 2014. Crop yields and global food security: will yield increase continue to feed the world? ACIAR Monograph no. 158. Australian Centre for International Agricultural Research: Canberra, ACT, Australia.
- Furbank RT. 2011. Evolution of the C₄ photosynthetic mechanism: are there really three C₄ acid decarboxylation types? *Journal of Experimental Botany* 62: 3103–3208.
- Ghannoum O, Evans JR, Chow WS, Andrews J, Conroy J, von Caemmerer S. 2005. Faster Rubisco is the key to superior nitrogen-use efficiency in NADP-malic enzyme relative to NAD-malic enzyme C₄ grasses. *Plant Physiology* **137**: 638–650.
- Gu J, Yin X, Stomph TJ, Struik PC. 2014. Can exploiting natural genetic variation in leaf photosynthesis contribute to increasing rice productivity? A simulation analysis. *Plant, Cell & Environment* 37: 22–34.
- Hatch MD. 1987. C₄ photosynthesis: a unique blend of modified biochemistry, anatomy and ultrastructure. *Biochimica et Biophysica Acta* **895**: 81–106.
- Heldt HW, Flugge U, Daves D. 1987. Subcellular transport of metabolites in plant cells. *Biochemistry of Plants* 12: 49–85.
- Huber SC, Edwards GE. 1975. Regulation of oxaloacetate, aspartate, and malate formation in mesophyll protoplast extracts of three types of C₄ plants. *Plant Physiology* 50: 324–331.

Ishikawa N, Takabayashi A, Sato F, Endo T. 2016. Accumulation of the components of cyclic electron flow around photosystem I in C₄ plants, with respect to the requirements for ATP. *Photosynthesis Research* 129: 261–277.

- Kanai R, Edwards GE. 1999. The biochemistry of the C₄ photosynthesis. In: Sage RF, Monson RK, eds. C₄ plant biology. Toronto, ON, Canada: Academic Press, 49–87.
- Karki S, Rizal G, Quick WP. 2013. Improvement of photosynthesis in rice (*Oryza sativa* L.) by inserting the C₄ pathway. *Rice* 6: e28.
- Koteyeva NK, Voznesenskaya EV, Edwards GE. 2015. An assessment of the capacity for phospho*enol*pyruvate carboxykinase to contribute to C₄ photosynthesis. *Plant Science* 235: 70–80.
- Leegood RC, Walker RP. 2003. Regulation and roles of phosphoenolpyruvate carboxykinase in plants. Archives of Biochemistry and Biophysics 414: 204–210.
- Mitchell PL, Sheehy JE. 2006. Supercharging rice photosynthesis to increase yield. *New Phytologist* 171: 688–693.
- Sage RF. 2016. A portrait of the C₄ photosynthetic family on the 50th anniversary of its discovery: species number, evolutionary lineages, and Hall of Fame. *Journal of Experimental Botany* 67: 4039–4056.
- Sales CRG, Ribeiro RV, Hayashi AH, Marchiori PER, Silva KI, Martins MO, Silveira JAG, Silveira NM, Machado EC. 2018. Flexibility of C₄ decarboxylation and photosynthetic plasticity in sugarcane plants under shading. *Environmental* and Experimental Botany 149: 34–42.
- Schuler ML, Mantegazza O, Weber APM. 2016. Engineering C₄ photosynthesis into C₃ chassis in the synthetic biology age. *The Plant Journal* 87: 51–65.
- Shameer S, Ratcliffe RG, Sweetlove LJ. 2019. Leaf energy balance requires mitochondrial respiration and export of chloroplast NADPH in the light. *Plant Physiology* 180: 1947–1961.
- Smith AM, Woolhouse HW. 1983. Metabolism of phospho*enol*pyruvate in the C₄ cycle during photosynthesis in the phospho*enol*pyruvate-carboxykinase C₄ grass Spartina anglica Hubb. Planta 159: 570–578.
- Sonawane BV, Sharwood RE, von Caemmerer S, Whitney SM, Ghannoum O. 2017. Short-term thermal photosynthetic responses of C₄ grasses are independent of the biochemical subtype. *Journal of Experimental Botany* 20: 5583–5597.

Suzuki S, Murai N, Burnell JN, Arai M. 2000. Changes in photosynthetic carbon flow in transgenic rice plants that express C₄-type phospho*enol*pyruvate carboxykinase from *Urochloa panicoides. Plant Physiology* 124: 163–172.

von Caemmerer S, Furbank RT. 1999. Modeling C₄ photosynthesis. In: Sage RF, Monson RK, eds. C₄ plant biology. Toronto, ON, Canada: Academic Press, 173–211.

von Caemmerer S, Quick WP, Furbank RT. 2012. The development of C₄ rice: current progress and future challenges. *Science* 336: 1671–1672.

Wang P, Khoshravesh R, Karki S, Tapia R, Balahadia CP, Bandyopadhyay A, Quick WP, Furbank R, Sage TL, Langdale JA. 2017. Re-creation of a key step in the evolutionary switch from C_3 to C_4 leaf anatomy. *Current Biology* 27: 3278–3287.e6.

Weber APM, Bar-Even A. 2019. Improving the efficiency of photosynthetic carbon reactions. *Plant Physiology* 179: 803–812.

Yin X, Struik PC. 2012. Mathematical review of the energy transduction stoichiometries of C₄ leaf photosynthesis under limiting light. *Plant, Cell & Environment* 35: 1299–1312.

Yin X, Struik PC. 2017. Can increased leaf photosynthesis be converted into higher crop mass production? A simulation study for rice using the crop model GECROS. *Journal of Experimental Botany* 68: 2345–2360.

Yin X, Struik PC. 2018. The energy budget in C₄ photosynthesis: insights from a cell-type-specific electron transport model. *New Phytologist* 218: 986–998.

Yuan L. 2017. Progress in super-hybrid rice breeding. *Crop Journal* 5: 100–102. Zhu XG, Long SP, Ort DR. 2010. Improving photosynthetic efficiency for greater

yield. Annual Review of Plant Biology 61: 235–261.

Supporting Information

Additional Supporting Information may be found online in the Supporting Information section at the end of the article.

Fig. S1 The three classically-defined C_4 subtypes classified according to decarboxylation enzymes.

Notes S1 Deriving the equation for quantum yield for CO₂-assimilation (Φ_{CO2}), and equations for calculating parameters a and f_{CET} from the measured Φ_{CO2} .

Notes S2 Extending the model of Yin & Struik (2018) to accommodate the mixed type.

Notes S3 Extending the C_4 submodel in crop model GECROS to accommodate the C_4 ideotype.

Table S1 Definitions and units of model symbols.

Table S2 Indicative values of model input parameters used in the analysis.

Table S3 Formulae for calculating cell-type-specific NADPH and ATP demands per CO_2 assimilation in the mixed type.

Please note: Wiley Blackwell are not responsible for the content or functionality of any Supporting Information supplied by the authors. Any queries (other than missing material) should be directed to the *New Phytologist* Central Office.

Key words: C4 photosynthesis, crop improvement, energy budget, photosynthesis ideotype, quantum yield.

Received, 6 June 2020; accepted, 11 October 2020.