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Plant ontogeny determines strength and associated plant fitness consequences of plant-mediated interactions between herbivores and flower visitors

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Abstract

- Plants show ontogenetic variation in growth-defence strategies to maximize reproductive output within a community context. Most work on plant ontogenetic variation in growth-defence trade-offs has focussed on interactions with antagonistic insect herbivores. Plants respond to herbivore attack with phenotypic changes. Despite the knowledge that plant responses to herbivory affect plant mutualistic interactions with pollinators required for reproduction, indirect interactions between herbivores and pollinators have not been included in the evaluation of how ontogenetic growth-defence trajectories affect plant fitness.
- 2. In a common garden experiment with the annual *Brassica nigra*, we investigated whether exposure to various herbivore species on different plant ontogenetic stages (vegetative, bud or flowering stage) affects plant flowering traits, interactions with flower visitors and results in fitness consequences for the plant.
- 3. Effects of herbivory on flowering plant traits and interactions with flower visitors depended on plant ontogeny. Plant exposure in the vegetative stage to the caterpillar *Pieris brassicae* and aphid *Brevicoryne brassicae* led to reduced flowering time and flower production, and resulted in reduced pollinator attraction, pollen beetle colonization, total seed production and seed weight. When plants had buds, infestation by most herbivore species tested reduced flower production and pollen beetle colonization. Pollinator attraction was either increased or reduced. Plants infested in the flowering stage with *P. brassicae* or *Lipaphis erysimi* flowered longer, while infestation by any of the herbivore species tested increased the number of flower visits by pollinators.
- 4. Our results show that the outcome of herbivore-flower visitor interactions in *B. nigra* is specific for the combination of herbivore species and plant ontogenetic stage. Consequences of herbivory for flowering traits and reproductive output were strongest when plants were attacked early in life. Such differences in selection pressures imposed by herbivores to specific plant ontogenetic stages may drive the evolution of distinct ontogenetic trajectories in growth-defence-reproduction strategies and include indirect interactions between herbivores and flower visitors.

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5. Synthesis. Plant ontogeny can define the direct and indirect consequences of herbivory. Our study shows that the ontogenetic stage of plant individuals determined the effects of herbivory on plant flowering traits, interactions with flower visitors and plant fitness.

KEYWORDS

florivores, herbivore-induced plant responses, indirect interactions, ontogenetic trajectories, ontogenetic variation, plant defence, plant reproduction, pollinators

1 | INTRODUCTION

Interactions between species are the foundations of ecological communities. Ontogenetic stages affect the role that individuals of a species play within a community (Miller & Rudolf, 2011; Nakazawa, 2015). For example, plants gradually develop from seedling through pre-reproductive and reproductive stages, to eventually senesce and die (Boege & Marquis, 2005). During each of these stages, plants interact with different community members such as antagonistic herbivores, competing plants or beneficial pollinators. To maximize their fitness, plants may display ontogenetic variation in: resistance to herbivores (Barton & Koricheva, 2010; Boege & Marquis, 2005), investment in growth to outcompete neighbours for light (Tonnabel, David, & Pannell, 2017; Zhang, Zhou, Huang, Japhet, & Sun, 2008) or investment in the recruitment of natural enemies of herbivores to reduce the impact of herbivore attack (Quintero, Barton, & Boege, 2013; Quintero, Lampert, & Bowers, 2014).

Plants are part of dynamic communities that constantly shift the cost-benefit balance between plant growth and defence. Distinct ontogenetic trajectories in growth-defence strategies may allow plants to optimize responses to such shifts (Barton & Boege, 2017). However, ontogenetic growth-defence trajectories come with important implications. First, investment in one component may energetically tradeoff against other components of the trajectory. Second, investment in one component may alter plant interactions with other community members (de Vries, Evers, & Poelman, 2017; Dutton, Luo, Cembrowski, Shore, & Frederickson, 2016; Lucas-Barbosa, 2016; Villamil, 2017). For example, large plants may be more apparent to herbivores as a result of increased investment in growth induced by competition for light (de Vries, Evers, Dicke, & Poelman, 2019). Plant defence responses to herbivory can affect plant reproduction by changes in flowering time, flower abundance and plant interactions with flower visitors (Rusman, Lucas-Barbosa, Poelman, & Dicke, 2019). Surprisingly, despite the fact that pollinators are essential for the successful reproduction of most plant species, these have seldom been included in theoretical frameworks of growth-defence trade-offs through plant ontogeny (but see Villamil, 2017).

Herbivores may affect flower visitors in various ways. For example, herbivores may directly affect flower visitation by physically repelling pollinators (Canela & Sazima, 2003), or indirectly by removing flower biomass that makes plants less attractive to flower visitors (Söber, Moora, & Teder, 2010). The effects of herbivores on pollinators may be more extensive through indirect interactions mediated by herbivore-induced plant responses. Defence and reproductive traits are physiologically linked via multiple mechanisms such as resource trade-offs, shared phytohormonal signalling pathways, shared genetic and biochemical pathways (Jacobsen & Raguso, 2018; Lucas-Barbosa, 2016; Rusman, Lucas-Barbosa, et al., 2019). In addition, individual traits can have both defensive and reproductive functions. As a consequence, plant defensive responses induced by herbivores affect flower traits. For example, flowering plants under attack by caterpillars change floral volatile emission to attract natural enemies of the caterpillars. These changes at the same time reduce the attraction of bumblebees that use floral volatiles during foraging (Schiestl, Kirk, Bigler, Cozzolino, & Desurmont, 2014). Indeed, herbivore-induced changes in expression of flower traits affect the visitation of mutualistic and antagonistic flower visitors (McArt, Halitschke, Salminen, & Thaler, 2013; Rusman, Poelman, Nowrin, Polder, & Lucas-Barbosa, 2019; Stam, Dicke, & Poelman, 2018). Because flower visitors directly interact with the reproductive organs of the plant, herbivore-flower visitor interactions may come with important plant fitness consequences (Chautá, Whitehead, Amaya-Márquez, & Poveda, 2017; Moreira, Castagneyrol, Abdala-Roberts, & Traveset, 2019; Rusman, Lucas-Barbosa, & Poelman, 2018).

Plant ontogeny is a crucial factor that may determines the outcome and fitness consequences of herbivore-flower visitor interactions. The costs of herbivory vary over plant ontogeny, as well as plant responses to herbivory (Boege, Dirzo, Siemens, & Brown, 2007; Brütting et al., 2017; Diezel, Allmann, & Baldwin, 2011; Rostás & Eggert, 2008). For example, juvenile/vegetative plants may experience higher costs of herbivory compared to flowering plants due to the consumption of important photosynthetic tissues. Juvenile/vegetative plants may respond to herbivore attack with expensive resistance traits to protect those valuable tissues, whereas flowering plants may rely on constitutive defences and/or tolerance mechanisms (Boege et al., 2007; Boege & Marquis, 2005; Lucas-Barbosa et al., 2017; Ochoa-López, Villamil, Zedillo-Avelleyra, & Boege, 2015). Hence, attack early in plant life might reduce resource availability for reproduction. Resource trade-offs between herbivore resistance and reproduction become apparent in expensive flower traits, such as flowering time, flower abundance, nectar and pollen production (Barber et al., 2015; Poveda, Steffan-Dewenter, Scheu, & Tscharntke, 2005; Quesada, Bollman, & Stephenson, 1995; Strauss, Conner, & Rush, 1996). Indeed, plants attacked early in development produced smaller inflorescences as compared with non-damaged plants and plants attacked late in development (Hoffmeister, Wittköpper, & Junker, 2016). In addition to resource-based mechanisms, flowering plants change flower traits as part of their defensive response induced by herbivory (Rusman, Lucas-Barbosa, et al., 2019). Such changes are apparent in traits that function in both defence and reproduction, such as flower volatiles and colour (Desurmont, Laplanche, Schiestl, & Turlings, 2015; Rusman, Poelman, et al., 2019). For example, flowering turnip plants change floral volatile emission upon herbivore attack to increase the attraction of natural enemies of the herbivores, but these changes reduce pollinator attraction (Schiestl et al., 2014). Herbivore attack early in development might alter flower traits primarily via resource-based mechanisms, while attack late in development may change flower traits to optimize defensive functions while reducing reproductive functions. Therefore, we expect ontogenetic variation in plant-mediated herbivore-flower visitor interactions. So far, ontogenetic variation in indirect herbivore-flower visitor interactions and associated fitness consequences have not been investigated.

In this study on the annual plant Brassica nigra, we investigated whether exposure of plants at different ontogenetic stages to various herbivore species affects plant flowering traits, interactions with flower visitors and results in fitness consequences for the plant. In a manipulative experiment, we exposed plants in the vegetative, bud or flowering stage to one of six herbivore species. More specifically, we studied whether herbivore attack to plants at these three ontogenetic stages affects (a) plant phenological traits and flower abundance, (b) visitation rates of mutualists (pollinators), (c) abundance of antagonists (florivorous pollen beetles, Meligethes spp.) and (d) seed production. By studying ontogenetic variation of effects of herbivory on plant reproduction we aimed at elucidating whether the selection pressures imposed by herbivores vary depending on the plant ontogenetic stage in which the plant was attacked. Such ontogenetic variation in selection pressures potentially drive the evolution of plant defence through their ontogeny (Barton & Boege, 2017; Ochoa-López, Rebollo, Barton, Fornoni, & Boege, 2018; Poelman & Kessler, 2016).

2 | MATERIALS AND METHODS

2.1 | Plant and insects

Black mustard *Brassica nigra* L. is an annual plant belonging to the cabbage and mustards family (Brassicaceae). Plants grow often in high-density stands on open river banks and floodplains, and as early successional species in disturbed areas. This species is considered to be an obligate outcrossing species (Conner & Neumeier, 1995), with a generalized pollination system (Lucas-Barbosa, van Loon, Gols, Beek, & Dicke, 2013; Rusman et al., 2018). Plants flower for several weeks in which hundreds of small yellow flowers with four petals are

produced. New flowers open daily, with a relatively short longevity of 3–5 days. Flowers are hermaphroditic, that is contain both male and female structures.

We used seeds of a Black mustard accession (CGN06619) that originates from the Centre for Genetic Resources (CGN). Seeds were propagated by open field pollination and germinated in trays. One-week-old plants were transplanted to and cultivated in pots (Ø 17 cm; 2 L) under greenhouse conditions ($23 \pm 2^{\circ}$ C, 50%–70% r.h., L16:D8). Pots were filled with potting soil (Lentse potgrond) and sand in a 1:1 volume ratio. When plants were 2-week-old they were transferred to an outside area protected by an insect screen. Three-week-old plants were transplanted into the field.

Black mustard plants are colonized by a diverse herbivore community, comprising more than 30 species, of which most are specialist herbivores. We exposed plants to six herbivore species from three herbivore functional groups (HFGs), namely two chewing herbivores (the caterpillar Pieris brassicae and sawfly Athalia rosae), two sap-feeding herbivores (the aphids Brevicoryne brassicae and Lipaphis erysimi) and two root-feeding herbivores (the cabbage root fly Delia radicum and nematode Heterodera schachtii). In the field, the above-ground herbivores colonize plants throughout the season when plants are still vegetative seedlings till large flowering plants (E. Poelman and Q. Rusman, pers. obs.). This information is largely lacking for the root herbivores. Natural colonization densities for above-ground herbivores range between 1 and 30 chewing herbivore larvae or adult aphids per plant (E. Poelman and Q. Rusman, pers. obs.), for the cabbage root fly five to nine larvae per plant (Soler, 2009) and for nematodes about 1,000 eggs and juveniles per 100 g soil (H. Helder, pers. comm.).

The caterpillar P. brassicae, the sawfly A. rosae and the aphids B. brassicae and L. erysimi are routinely reared in the Laboratory of Entomology (Wageningen University) under greenhouse conditions (22 ± 1°C, 50%-70% r.h., L16:D8). Pieris brassicae and B. brassicae were reared on Brussels sprouts Brassica oleracea variety gemmifera cultivar Cyrus; A. rosae and L. erysimi were reared on Raphanus sativus. Larvae of the cabbage root fly D. radicum were reared on turnips Brassica rapa or rutabaga (Brassica napus) in a climate cabinet (22 ± 1°C, 50%-70% r.h., L16:D8). Nematodes were reared on rapeseed B. napus cultivar Jennifer. Cysts were hatched in the laboratory using a 3 mM ZnCl₂ solution (Rusman et al., 2018). After hatching, nematodes were flushed out of the hatching sieve using tap water, and solutions containing about 1,000 nematodes (J2 stage) in 4 ml of water were used to infest plants. The caterpillar P. brassicae, and the aphids B. brassicae and L. erysimi were originated from the surroundings of Wageningen (The Netherlands), while the sawfly A. rosae originated from surroundings of Würzburg (Bavaria, Germany), the cabbage root fly D. radicum from Zeewolde (The Netherlands) and cysts of the nematode H. schachtii from the rearing of the Institute for Rational Sugar production (IRS; Bergen op Zoom, The Netherlands). The population used was IRS 07-01-04.02 and originated from Woensdrecht, The Netherlands.

2.2 | Common garden experiment–Field design

A common garden experiment was designed to investigate whether herbivore infestation of plants at different developmental stages affected flowering traits (number of inflorescences and phenological traits), flower visitors (mutualistic pollinators and antagonistic pollen beetles, Meligethes spp.) and plant seed production. We planted 160 plots of B. nigra in a field of the experimental farm of Wageningen University, The Netherlands (51°59'N, 5°39'E). Plots were organized in 10 rows and 16 columns, and each plot was composed of five plants-one central plant and four plants surrounding the central plant-at a distance of 20 cm. Distance between central plants of neighbouring plots was 1.5 m. Each day 24 plots were planted, except for day 7, when we planted 16 plots. Plots of columns 1-8 were planted between days 1 and 4, column 5 was kept empty, and plants of columns 9-16 were planted between days 4 and 7. Treatments were randomly assigned over plots using a Latin square design, that is each combination of herbivore species and plant developmental stage never occurred twice in the same row or column. Treatments were equally divided over the planting dates and replicated eight times. The experiment was performed from the beginning of May to the end of August (2016).

2.3 | Plant treatments

Plants were infested with herbivores at different developmental stages, either in the vegetative, bud or flowering stage (Figure 1). Plots in the vegetative stage, growth stage 14-17 (based on *B. napus*; Meier, 2001) were infested one day after planting. Plots in the

bud or flowering stage were infested one day after three of the five plants of a plot had reached the bud or flowering stage, including the central plant. We considered that a plant had reached the bud stage when buds of the first flowering stalk rose above the leaves (growth stage 53; Meier, 2001). Plants were considered flowering when the first flower opened (growth stage 60; Meier, 2001). We placed a mesh tent (95 I \times 95 w \times 190 hr/cm) for 24 hr over each plot for infestation, to provide the necessary time for the herbivores to settle on the plants. Uninfested control plots were also covered with a mesh tent for 24 hr right after planting, when all five plants were still in the vegetative stage.

We infested *B. nigra* plants by placing 10 first instar-chewing herbivores or 10 adult sap-feeding herbivores on two true leaves (five per leaf), or by placing 10 first instar *D. radicum* larvae at the base of the plant stem. To infest a plant with nematodes, 4 ml of solution containing about 1,000 nematodes was added in four holes (1 ml per hole) around the main stem of the plant (Rusman et al., 2018). Such holes were approximately 10 cm deep with a diameter of 0.5 cm, and were made at 2 cm from the stem of each of the five plants. For all insect herbivores we used densities that were representative for intermediate natural infestation densities.

Six days after infestation we monitored the survival of all aboveground herbivore species for each plant. Plants were re-infested with five second instar-chewing larvae or five adult-sucking herbivores when two or fewer herbivores were recorded. At the end of the experiment, root samples were taken from approximately 80 plants and checked for damage by *D. radicum* or nematode presence.



FIGURE 1 Schematic representation of the timeline of the experiment. All *Brassica nigra* plants were planted at the same time in the field. Plants exposed to herbivores in the vegetative stage (light green bar) were infested 1 day after planting. Plants exposed to herbivores in the bud stage (dark green bar) were infested 1 day after plots had reached the bud stage. Plants exposed to herbivores in the flowering stage (yellow bar) were infested 1 day after plots had reached the flowering stage [Colour figure can be viewed at wileyonlinelibrary.com]

2.4 | Effect of herbivore infestation and plant ontogeny on flower abundance and phenological traits

To investigate if flower abundance and several phenological traits of B. nigra were influenced by herbivore exposure at different plant developmental stages, we recorded flower abundance and followed plant development of infested and uninfested plants. Flower abundance was assessed by counting all inflorescences of each plant at three time points (Figure 1): (a) 7-9 days after plants had entered the bud stage, and at (b) 7-9 and (c) 14-16 days after plants had started flowering. Plant phenological traits were assessed by monitoring plant development daily. We recorded the start of the bud stage, the start of the flowering stage and the end of flowering. Plants were considered finished with flowering when all buds and flowers were gone, and only developing and ripe siliques remained on the flower stalks. We calculated the duration of the bud stage by subtracting the number of days needed to reach the bud stage from the number of days needed to reach the flowering stage. We calculated the duration of the flowering stage by subtracting the number of days needed to reach the flowering stage from the number of days to the termination of flowering.

2.5 | Effects of herbivore infestation and plant ontogeny on floral mutualists and antagonists

To investigate if pollinator visitation to B. nigra flowers was influenced by herbivore exposure of plants at different developmental stages, we recorded pollinator behaviour in plots of infested and uninfested plants at two time points during the following flowering stage: (a) between 7 and 9 days and (b) between 14 and 16 days (Figure 1). Pollinator visitation to a plot was monitored for 10 min. When a pollinator entered the plot and had contact with a flower, identity of the pollinator, number of flowers visited and time spent per flower were recorded (Rusman et al., 2018). The identity of other pollinators that visited the plot during the observation of a pollinator was recorded as well. If the same pollinator individual returned to the plot under observation after having visited a different plot, we scored that visit as a new visit (Rusman et al., 2018). Pollinators were placed in one of the following six pollinator groups: honeybees (HB) Apis mellifera, bumblebees Bombus terrestris, Bo. lapidarius, Bo. pascuorum and other Bombus spp., syrphid flies (SF; several Eristalis spp. and several other syrphid species), solitary bees (several Andrena and Lasioglossum species but also other Apidae excluding Bombus spp.), other flies (non-syrphid Diptera) and butterflies (Pieris spp. and other Lepidoptera). Recordings were done using a handheld computer (Psion Workabout Protm 3) programmed with The Observer XT software (version 10, Noldus Information Technology, Wageningen, The Netherlands). Recordings were done during the day (between 9 a.m. and 1 p.m., or 2 p.m. and 5 p.m.) and only when weather conditions were favourable for pollinator activity (15-30°C and wind speed ≤6 m/s; Rusman et al., 2018).

To investigate if pollen beetle (*Meligethes* spp.) colonization was influenced by plant exposure to herbivores in different plant developmental stages, we monitored pollen beetle abundance on plots of infested and uninfested plants. We counted the number of adult pollen beetles on each plant of a given plot at the same three time points we assessed flower abundance (Figure 1). Recordings were done during the day (2–6 p.m.) and only when weather conditions were favourable for pollen beetle activity (15–30°C and wind speed ≤6 m/s; Rusman et al., 2018).

2.6 | Effects of herbivore infestation at different plant ontogenetic stages on plant seed production

To investigate if life-time seed production was influenced by herbivory during different plant developmental stages, we assessed seed number and biomass of the plants. We harvested seeds of three plants for each plot; the central plant and two side plants (randomly selected and not adjacent to each other). First harvesting date for each plant was selected before the first siliques would lose their seeds (Rusman et al., 2018). At first harvest, we collected all ripe siliques and left immature siliques and flowers on the plant. Then, plants were checked weekly and siliques harvested when ripe. Siliques were stored in paper bags in a dry storage room until seeds were manually extracted from the siliques. We calculated total number of seeds per plant by weighing 100 seeds, and the total weight of seeds harvested per plant (Rusman et al., 2018). We estimated the total number of seeds by dividing total seed weight by the weight of 100 seeds and multiplied the result by 100. The weight of one seed was estimated by dividing the weight of 100 seeds by 100.

2.7 | Statistical analysis

For count data such as the number of insects, flowers, days and seeds, we used GL(M)M with a Poisson distribution and a log link function, or negative binomial distribution with a log link function to correct for overdispersion. We ran two models: The first model included herbivore treatment and plant developmental stage and their interactions as fixed factors. In the case of numbers of insects or flowers, this model was run for each time point separately because (a) not all time points included all plant developmental stages and (b) we were interested in species-specific effects of herbivory on plant flowering traits and interactions with flower visitors, and exploring patterns of higher functional levels (i.e. plant ontogeny) rather than changes over time. Uninfested control plants were excluded from these analyses, because they could not be assigned to any plant developmental stage. The second model included herbivore treatment nested in HFG, time point (except for phenological traits or seeds) and the interaction between herbivore treatment and time point as fixed factors. This model was run for each plant developmental stage separately. Interactions were removed from the models if they were statistically non-significant (p > .05). For post hoc analyses we used Tukey's post hoc tests. Random factors were selected using a backward approach; all random factors

such as day (not for flowering traits or seeds), time (morning vs. afternoon; only for pollinators), plot (not for pollinators), plant position (not for pollinators), day*treatment (not for flowering traits or seeds) were added to the model and removed if they explained <3% of the variation or were statistically non-significant (p > .05). We used the LME4 (Bates, Maechler, Bolker, & Walker, 2015), MULTCOMP (Hothorn, Bretz, & Westfall, 2008) and LSMEANS (Lenth, 2016) packages for these analyses. For continuous data such as time spent per visit and per flower by pollinators, we used linear (mixed) models with a Gaussian distribution and identity link function or a Gamma distribution with a log link function if the data did not follow a normal distribution. The same fixed factors, random factor selection approach and software packages as for count data were used. We analysed pollinator community composition by comparing the pollinator community composition of infested and uninfested plots with a chi-squared test (Rusman et al., 2018). Expected pollinator community composition was calculated by summing pollinators within each pollinator group for all plots and dividing this number by the total number of pollinators. This results in an expected percentage for each pollinator group. This percentage was then multiplied by the total number of pollinators recorded for infested or uninfested plots (Rusman et al., 2018). We calculated expected community composition based on pollinators visiting all treatment groups because the pollinator community distributes over the different treatments including the uninfested plots based on pollinator preference in the choice situation, for example the community composition of the uninfested plots is affected by the presence of the infested plots (Rusman et al., 2018). If pollinator community composition was explained by plant exposure to herbivores, pairwise comparisons among all herbivores within one plant developmental stage were performed, and pairwise comparisons among the three plant developmental stages for each herbivore species were performed using chi-squared tests. To correct for multiple tests of pairwise comparisons, we adjusted the *p*-values using the false discovery rate correction. We used the fifer package for these analyses (Fife, 2014). In addition, to assess which pollinator groups contributed to differences between herbivore species and plant developmental stages, we calculated the standardized residuals for each pollinator group in each treatment (Rusman et al., 2018). We used a threshold value of ±2, for example residual values higher than +2 or lower than -2 indicate a significant contribution of that pollinator group to the differences in pollinator community composition (Sharpe, 2015). Pollinator groups which composed <1% of the community were excluded from the analysis, for example other flies (0.3%), butterflies (0.04%). For correlations between the number of inflorescences and insects, we computed the correlation coefficient r using the Pearson or Kendall method, depending on the distribution of the data. For correlations between the number of inflorescences and pollinators, we averaged the number of inflorescences per plot. Correlation graphs were made using the GGPUBR package (Kassambara, 2018). All analyses were carried out in R (version 3.4.3 × 64, 2017, The R Foundation for Statistical Computing Platform).

3 | RESULTS

Plant ontogeny determined the effects of herbivory on flowering plant traits, interactions with flower visitors and plant seed production. Below we provide details for the effects of herbivory at different plant ontogenetic stages on flower abundance, pollinator attraction, pollen beetle colonization and plant reproduction. Effects on plant phenological traits, pollinator community composition, pollinator visitation and correlations between numbers of pollinators/pollen beetles and flowers are described in the Supporting Information.

3.1 | Effects of herbivore infestation and plant ontogeny on flower abundance

Plant ontogeny determined the effects of plant exposure to herbivores on the number of flowers produced by B. nigra, and these effects were herbivore species-specific and not associated with the HFG (Figure 2; Tables S3 and S4). One week after plants had started to produce buds, plants exposed to herbivores in the vegetative stage had fewer inflorescence on display compared with plants exposed to herbivores in the bud stage (Tukey's post hoc test, p < .001), and this effect was particularly strong for plants exposed to P. brassicae caterpillars (Tukey's post hoc test, p < .001). Effects of herbivore species on the number of inflorescences were always negative, and observed for plants exposed to P. brassicae caterpillars in the vegetative stage and H. schachtii nematodes in the bud stage 1 week after plants had started to produce buds (Figure 2). These negative effects were still apparent 2 weeks after plants had started flowering for plants exposed to P. brassicae caterpillars in the vegetative stage, and by this time, plants exposed to B. brassicae aphids in the vegetative stage also had fewer inflorescences compared to uninfested plants. Effects of P. brassicae, B. brassicae and L. erysimi on the number of inflorescences 2 weeks after plants had started flowering varied depending on the ontogenetic stage in which the plant was attacked (Table S2).

3.2 | Effects of herbivore infestation and plant ontogeny on floral mutualists—Pollinator attraction

One week after plants had started flowering, plots were on average visited by 32 pollinators which visited 89 flowers during the 10-min observation time, and 2 weeks after plants had started flowering this increased to 44 pollinators and 101 flower visits. Plant ontogeny determined the effects of plant exposure to herbivores on the number of pollinators visiting plots and the number of flowers visited by pollinators, and these effects mostly depended on herbivore identity rather than HFG (Figure 3; Figures S3–S5 and S8–S10; Tables S3 and S4). One week after plants had started flowering, plants exposed to herbivores in the bud stage were visited by a larger number of pollinators, especially honeybees, compared with plants exposed to herbivores in the flowering stage (Tukey's post hoc tests, total pollinators [TP]: p = .025, HB: p = .034), and effects were particularly strong when plants



FIGURE 2 Number of inflorescences of uninfested *Brassica nigra* plants and plants infested with herbivores at different plant ontogenetic stages. Boxplots show median (line), first and third quartiles, minimum and maximum. Outliers (1.5 times the interquartile range below the first or above the third quartile) are represented by circles. Plants were monitored for the number of inflorescences at three time points: between 7 and 9 days after plants had reached the bud stage, and between 7 and 9 days, and 14 and 16 days after plants had started flowering. Number of replicates per herbivore treatment varied between 28 and 45 plants, and between 76 and 78 for uninfested plants. Letter groups (a,b) and (k,l) above bars indicate significant differences ($p \le .05$) between herbivore species within a plant ontogenetic stage based on Tukey's post hoc tests, and small or capital letters were used for different time points. Greek letters above lines indicate significant differences at ($p \le .05$) between no indicates no differences [Colour figure can be viewed at wileyonlinelibrary.com]

were exposed to larvae of the sawfly A. *rosae* (Tukey's post hoc tests, TP: p = .025, HB: p = .034), *L. erysimi* aphids (Tukey's post hoc test, TP: p = .007) or *H. schachtii* nematodes (Tukey's post hoc tests, TP: p < .001, HB: p < .001). However, plants exposed to herbivores in the flowering stage received more syrphid fly visits compared to plants exposed in the vegetative stage (Tukey's post hoc test, p = .036). In contrast to the number of pollinator visits, more flowers were visited by all pollinators for plants exposed to herbivores in the flowering (Tukey's post hoc tests, TP: p = .006, HB: p = .002) and compared to plants exposed in the vegetative stage on both time points (Tukey's post hoc tests, TP: p = .006, HB: p = .002) and compared to plants exposed in the vegetative stage on both time points (Tukey's post hoc tests, 1 week: TP: p < .001, HB: p < .001, 2 weeks: TP: p < .001, HB: p = .005). This was true for four of the six herbivore species (Table S2). The effects of individual herbivore species on the number of TP, honeybees and SF varied with plant ontogeny for both time points (Table S2).

Specific effects of HFG were observed when plants were exposed to herbivory in the flowering stage (total number of flowers visited by HB) 1 week after plants had started flowering (Table S4). Honeybees visited more flowers of plants exposed to sap-feeding or root herbivores in the flowering stage compared to uninfested plants (Tukey's post hoc tests, p = .006 and p = .003 respectively). Specific herbivores led to either increased or decreased pollinator visitation when plants were exposed in the vegetative stage (the number of flowers visited by SF), bud stage (the number of TP, HB and SF, and number of flowers visited by TP, HB and SF) or flowering stage (the number of TP and HB, and number of flowers visited by SF) if compared with uninfested plants (Figure 3; Figures S3–S5 and Figures S8–S10; Tables S3 and S4).

3.3 | Effects of herbivore infestation and plant ontogeny on a floral antagonist

We observed on average one pollen beetle adult per plant 1 week after plants had started to produce buds; this number increased to an average of seven beetles per plant 1 week after plants had started flowering, and declined to about three beetles per plant 2 weeks



FIGURE 3 Number of pollinator visits observed on flowers of uninfested plots (control) of *Brassica nigra* plants and on flowers of plots infested by herbivores at different plant ontogenetic stages. Boxplots show median (line), first and third quartiles, minimum and maximum. Outliers (1.5 times the interquartile range below the first or above the third quartile) are represented by circles. Observations lasted for 10 min and were made at two time points: between 7 and 9 days, and 14 and 16 days after plots had started flowerings. For 7–9 days after plots had started flowering, the number of replicates per herbivore treatment varied between 7 and 9, and was 16 for uninfested plants. For 14–16 days after plots had started flowering, the number of replicates per herbivore treatment varied between 2 and 6, and was 10 for uninfested plants. Letters groups (a–d, k–n, x–z) above bars indicate significant differences at ($p \le .05$) between herbivore species within a plant ontogenetic stage based on Tukey's post hoc tests, and small or capital letters were used for different time points. Greek letters above lines indicate significant differences ($p \le .05$) between a minimum and startes no differences [Colour figure can be viewed at wileyonlinelibrary.com]

after plants had started flowering. Plant ontogeny influenced the effects of plant exposure to herbivores on the number of pollen beetle adults observed per plant, and these effects depended on HFG and herbivore identity (Figure 4; Tables S3 and S4). Plants exposed in the vegetative stage contained fewer adult beetles compared to plants exposed in the bud stage 1 week after plants had started to produce buds (Tukey's post hoc test, p = .035) and 1 week after plants had started flowering (Tukey's post hoc test, p = .019). Interestingly, this was true for four of the six individual herbivore species 1 week after plants had started to produce buds, and for the other two herbivore species 1 week after plants had started flowering (Table S2).

Compared with uninfested plants, effects of the herbivores *A. rosae*, *P. brassicae* and *B. brassicae* were always negative, and observed for plants exposed in the vegetative stage (1 week after plants had started to produce buds and 1 week after plants had started flowering), and the bud stage (1 week after plants had started to produce buds and 2 weeks after plants had started flowering), but not for plants exposed in the flowering stage (Figure 4; Tables S3 and S4). The effects on plants exposed in the bud stage to herbivores depended on the HFG. One week after plants started to produce buds, plant exposed to chewing herbivores were more heavily colonized by pollen beetles than plants exposed to



FIGURE 4 Number of pollen beetle adults (*Meligethes* spp.) observed on flowers of uninfested (control) *Brassica nigra* plants and plants infested by herbivores at different plant ontogenetic stages. Boxplots show median (line), first and third quartiles, minimum and maximum. Outliers (1.5 times the interquartile range below the first or above the third quartile) are represented by circles. Plants were monitored for the number of pollen beetle adults at three time points: between 7 and 9 days after plants had reached the bud stage, and between 7 and 9 days, and 14 and 16 days after plants had started flowering. Number of replicates per herbivore treatment varied between 28 and 45 plants, and between 76 and 78 for uninfested plants. Letters groups (a,b) and (k,l) above bars indicate significant differences at ($p \le .05$) between herbivore species within a plant ontogenetic stage based on Tukey's post hoc tests, and small or capital letters were used for different time points. Greek letters above lines indicate significant differences at ($p \le .05$) between plant ontogenetic stages based on Tukey's post hoc tests, whereas ns indicates no differences [Colour figure can be viewed at wileyonlinelibrary.com]

sap-feeding herbivores or uninfested plants (Tukey's post hoc tests, p = .002 and p = .003 respectively). However, 1 week after plants started flowering, plant exposed to chewing and root herbivores had fewer pollen beetles than plants exposed to sap-feeding herbivores (Tukey's post hoc tests, p < .001 and p < .001 respectively) or uninfested plants (Tukey's post hoc tests, p < .001 and p < .001 and p < .001 respectively). Two weeks after plants started flowering, plants exposed to root herbivores had higher numbers of pollen beetles compared to plants (Tukey's post hoc tests, p < .001, p < .001 and p = .001 and p = .001 respectively).

3.4 | Effects of herbivore infestation and plant ontogeny on plant seed production

Brassica nigra plants produced on average 10,000 seeds, with an average individual seed weight of 1 mg. Overall, plant ontogeny

influenced the effects of herbivory on seed numbers and weight (Table S6). Plant ontogeny determined the effects of plant exposure to all herbivores when it comes to side plants, while for central plants and at the plot level these effects were herbivore specific (Figure 5; Figure S20; Table S6). Side plants exposed to herbivores in the vegetative stage produced fewer seeds than plants exposed to herbivores in the flowering stage (Tukey's post hoc test, p < .010). This was especially true for plants exposed to B. brassicae aphids in the vegetative stage if compared with plants that were exposed to these aphids in the flowering stage (Tukey's post hoc test, p < .001) but also in the bud stage (Tukey's post hoc test, p < .001). For the average number of seeds produced per plant per plot and seed weight, the effect of individual herbivore species varied with plant ontogeny (Table S2). This was also the case for the number of seeds produced by central plants. Herbivore species-specific effects on the number of seeds produced were restricted to plants exposed in the vegetative stage (Figure 5a; Figure S20). Plant exposure to L. erysimi aphids



FIGURE 5 Number of seeds (a) and seed weight (b) of seeds produced by uninfested (control) *Brassica nigra* plants and plants infested by herbivores at different plant ontogenetic stages. Boxplots show median (line), first and third quartiles, minimum and maximum. Outliers (1.5 times the interquartile range below the first or above the third quartile) are represented by circles. Number of seeds and seed weight were averaged for three plants per plot (one central plant and two side plants). The number of replicates per herbivore treatment varied between 43 and 50, and between 88 and 91 for uninfested plants. Letters above bars indicate significant differences at ($p \le .05$) between herbivore species within a plant ontogenetic stage based on Tukey's post hoc tests, whereas ns above lines indicates no differences between plant developmental stages [Colour figure can be viewed at wileyonlinelibrary.com]

increased the number of seeds produced, while seed numbers were reduced by exposure to *P. brassicae* caterpillars or *B. brassicae* aphids. Effects of herbivory on seed weight were restricted to plants exposed in the vegetative stage and depended on HFG and herbivore identity (Table S6). Plants exposed to sap-feeding herbivores produced lighter seeds compared with plants exposed to chewing herbivores, root herbivores or uninfested plants (Tukey's post hoc tests, p = .004, p < .001 and p = .031). Especially the aphid *B. brassicae* reduced the seed weight (Figure 5b).

4 | DISCUSSION

The findings of our study illustrate that plant ontogeny determines the effects of herbivory on flowering traits, interactions with pollinator mutualists and flower-feeding antagonists, and plant reproductive output (Figure 6). Effects of herbivory were mostly species-specific. In few cases-visitation times and flower visits by SF, seed weight-effects depended on the HFG of the herbivore. Plants exposed in the vegetative stage to P. brassicae caterpillars or B. brassicae aphids resulted in reduced flowering time and flower abundance. Plants infested with these herbivores had reduced pollinator attraction and plant colonization by pollen beetles. Overall, this negatively affected the number and weight of seeds produced. Interestingly, plants exposed in the vegetative stage to L. erysimi aphids increased seed production. When plants were exposed in the bud stage to A. rosae, B. brassicae, L. erysimi or H. schachtii, herbivory led to reduced flower abundance and pollen beetle colonization. Plants infested with these herbivores received either more or less pollinator visits. Plants exposed to herbivores in the flowering stage received more flower visits by pollinators than plants exposed at the vegetative and bud stages, irrespective of which herbivore we used as inducer. Plants infested in the flowering stage with P. brassicae caterpillars or L. erysimi aphids flowered longer than uninfested plants or plants infested with the other herbivores. Plant ontogenetic stage defined the effects of herbivory on changes in pollinator flower visitation behaviour, that is increases or decreases in the number of flowers visited, time spent per visit and flower. Taken together, both plant ontogeny and herbivore identity shaped the effects of herbivory on flowering traits, the outcome of indirect interactions with flower visitors, and the consequences for plant fitness.

Our data show that plant ontogeny is a major determinant of indirect interactions between herbivores and flower visitors. Plantmediated interactions were specific for the identity of the interaction partners (herbivores and flower visitors) and the direction of the interactions—positive, negative or neutral—varied with plant ontogeny. The resulting indirect interaction web between herbivores and flower visitors appears dynamic and variable over plant ontogeny. Ontogenetic variation in indirect interaction webs is evident from aquatic systems (predatory fish and pelagic and benthic food webs), and systems which include both aquatic and terrestrial components (amphibians or aquatic insects; Miller & Rudolf, 2011; Nakazawa, 2015). However, variation in indirect interaction



FIGURE 6 Illustration summarizing the effects of herbivore attack at different plant ontogenetic stages (vegetative. bud, flowering) on flowering traits, seed production and plant interactions with flower visitors. The sign in the circle shows the direction of effect as compared with uninfested plants, where (+) is a positive and (-) a negative effect. No arrow indicates no significant effects as compared with uninfested plants. Effects of herbivory in the vegetative stage were mainly identified for the caterpillar Pieris brassicae and aphid Brevicoryne brassicae, while effects of herbivory in the bud and flowering stage were spread among the six herbivores used: the sawfly Athalia rosae, the caterpillar P. brassicae, the aphid B. brassicae, the aphid Lipaphis erysimi, the cabbage root fly Delia radicum and the nematode Heterodera schachtii [Colour figure can be viewed at wileyonlinelibrary. coml

webs with ontogeny has received limited attention in plant-insect systems (Waltz & Whitham, 1997). Ontogenetic variation in indirect herbivore-flower visitor interactions can be explained by two nonmutually exclusive mechanisms: different plant responses over plant ontogeny or varying effects of herbivory on plants determined by the timing of herbivore encounter. Indeed, plant responses to herbivory vary with plant ontogeny (Diezel et al., 2011; Ochoa-López et al., 2015; Rostás & Eggert, 2008), which includes changes in flower traits (Desurmont et al., 2015; Hoffmeister et al., 2016). The timing of events such as herbivory determines the strength of interactions between herbivores and plants, and subsequent effects on flower visitors (Chase, 2003; Stam et al., 2018; Vannette & Fukami, 2014). Herbivory during the vegetative stage of a plant may be costlier as compared to herbivory during the flowering stage, due to increased investments in resistance and loss of important photosynthetic tissues early in life (Boege et al., 2007; de Vries et al., 2019; de Vries, Poelman, Anten, & Evers, 2018). This may lead to reduced investments in flowers because of resource limitations (Barber et al., 2015; Poveda et al., 2005; Quesada et al., 1995; Strauss et al., 1996). Herbivory affects organisms that engage in interactions with the plant under attack soon after the event. Moreover, new interactions established over the rest of the life of the plant are affected until the plant dies (Stam et al., 2018; Stam, Kos, Dicke, & Poelman, 2019; Van Zandt & Agrawal, 2004). The ontogeny of the herbivores themselves likely influences the outcomes of the effects of plant ontogeny on flower-visitor interactions and plant fitness (Barber, Adler, Theis, Hazzard, & Kiers, 2012; Pineda, Soler, Pastor, Li, & Dicke, 2017). Herbivore species differ in whether their population and damage decrease or increase over time. Caterpillar damage will increase overtime, but their numbers will decrease because of predation

and dispersion (Lucas-Barbosa et al., 2013). Aphids will grow exponentially to become strong nutrient sinks, while nematodes may need time to settle and reproduce before heavy damage is inflicted. Hence, herbivore attack will echo through the indirect interaction web differently depending on the arrival time of a specific herbivore with plant ontogeny.

While herbivore infestation in various plant ontogenetic stages affected plant-mediated interactions with flower visitors, effects of herbivory on plant flowering traits were most pronounced when herbivores colonized plants early in life, that is during their vegetative stage. Different effects of herbivore infestation on plant traits and on flower visitors suggest the importance of other traits than flower abundance for interactions with flower visitors. Indeed, variation in flower traits such as flower scent and morphology, which we did not assess in this study (but see Rusman, Poelman, et al., 2019), may explain part of the variation in flower-visitor communities (Kuppler, Höfers, Wiesmann, & Junker, 2016; Soper Gorden & Adler, 2016). Interestingly, plant exposure to herbivores affected the correlation between the number of inflorescences and pollinator mutualists, but not the correlation between the number of inflorescences and antagonistic pollen beetles. This may indicate variable importance of resource quantity and quality for mutualists and antagonists (Cariveau, Irwin, Brody, Garcia-Mayeya, & Ohe, 2004; Wenninger, Kim, Spiesman, & Gratton, 2016). Floral antagonists may prefer/better assess resource quantity than quality (Althoff, Xiao, Sumoski, & Segraves, 2013; Ekbom & Borg, 1996; Rusman et al., 2018; Wenninger et al., 2016), whereas both may be important for floral mutualists during foraging (Kuppler et al., 2016). Still, we found cases where herbivore exposure did not affect the number of inflorescences but did affect pollen beetle colonization. This could

be due to changes in traits that determine flower apparency, such as floral volatiles, which floral antagonists use to locate resources (Jönsson, Rosdahl, & Anderson, 2007; Theis & Adler, 2012). This suggests that antagonist-antagonist interactions are not limited by the dependence of antagonists on resource quantity (Rusman et al., 2018). Herbivory affects flower quantity and quality differently (Hoffmeister et al., 2016; Rusman, Poelman, et al., 2019), and mutualists and antagonists have contrasting effects on plant reproduction (Grass, Bohle, Tscharntke, & Westphal, 2018; Soper Gorden & Adler, 2018). Therefore, variable importance of resource quantity and quality for mutualists and antagonists is likely important for indirect plant fitness consequences of herbivory.

Our data show that fitness consequences of herbivory can be specific to the plant ontogenetic stage that is attacked. Feeding by specific herbivores only affected plant reproduction when plants were colonized by herbivores early in life, while still in the vegetative stage. This indicates that the potential trade-off between plant growth/ reproduction and defence is limited to herbivore attack in specific plant developmental stages (Lucas-Barbosa, 2016; Lucas-Barbosa, Loon, & Dicke, 2011). Differences in fitness consequences of herbivory through plant ontogeny can result from direct effects such as allocation costs or developmental constraints (Barton & Boege, 2017), and from indirect effects via plant-mediated interactions (Lucas-Barbosa, 2016; Poelman & Kessler, 2016; Strauss, 1997). For annual plants, the main defence strategy early in plant development is resistance, while later in plant development this switches to tolerance (Boege et al., 2007). Increased investment of black mustard in resistance and loss of important photosynthetic tissues due to herbivore damage early in life will be especially expensive (de Vries et al., 2019, 2018). This can explain our observed reduction in flowers, the number of seeds produced and seed weight for plants exposed in the vegetative stage to P. brassicae caterpillars or B. brassicae aphids. Alternatively, expression of the most effective defence strategy against P. brassicae and B. brassicae may be limited early in plant development due to developmental constraints (Barton & Boege, 2017; Quintero et al., 2013) or the absence of natural enemies of herbivores early in the season (Gómez-Marco, Tena, Jaques, & García, 2016; Mira & Bernays, 2002). Herbivory also affects seed production indirectly via interactions with plant-associated antagonists and mutualists (McArt et al., 2013; Pashalidou et al., 2015; Strauss, Rudgers, Lau, & Irwin, 2002). The reduction in pollinator visitation for plants exposed in the vegetative stage to P. brassicae caterpillars or B. brassicae aphids likely contributes to the reduced number of seeds produced. Alternatively, plant responses to herbivory may render plants more attractive for subsequent arriving herbivores with associated plant fitness costs (Erwin, Züst, Ali, & Agrawal, 2014), although we did not observe this for specialist pollen beetles (Meligethes spp.). Plant interactions with higher trophic levels are also affected by herbivory (Soler, Bezemer, Van Der Putten, Vet, & Harvey, 2005). Plant responses to herbivory may render plants less attractive to natural enemies of other herbivores (Pierre, Dugravot, et al., 2011; de Rijk, Yang, Engel, Dicke, & Poelman, 2016), potentially reducing plant fitness (Hoballah & Turlings, 2001; Pashalidou et al., 2015).

In nature, plants can be colonized by multiple herbivores at the same time or in close sequence. This can lead to complex interactive effects of herbivory on plant-associated organisms with consequences for plant seed production. For example, plants infested with H. schachtii or Pratylenchus penetrans nematodes can reduce aphid population growth, while Meloidogyne hapla nematodes enhance aphid population growth (Hol, Raaijmakers, Mons, Meyer, & Dam, 2016; van Dam, Wondafrash, Mathur, & Tytgat, 2018). Simultaneous colonization by nematodes and aphids on vegetative plants can thereby negatively or positively influence the effects that aphids have on seed production. Simultaneous herbivory can induce different plant responses as compared to induction by a single herbivore (Pierre, Jansen, et al., 2011; Ponzio, Papazian, Albrectsen, Dicke, & Gols, 2017). This can have consequences for plant-mediated interactions (Chrétien et al., 2018; Soler et al., 2012; Stam, Chrétien, Dicke, & Poelman, 2017; Stam et al., 2018) and plant seed production (Stam et al., 2019). Ontogenetic variation in networks of indirect plant-mediated interactions includes these complex interactive effects on plant fitness (Poelman & Kessler, 2016; Rusman et al., 2018; Soper Gorden & Adler, 2018; Stam et al., 2019). Taken together, variation in direct and indirect consequences of herbivory during plant ontogeny likely imposes selection pressures that drive the evolution of plant defence ontogenetic trajectories (Barton & Boege, 2017; Ochoa-López et al., 2018).

Plant ontogeny is important for direct and indirect consequences of herbivory. Therefore, studies on the evolution of plant defences need to consider ecologically relevant timing of herbivory. Plants can be particularly vulnerable to specific herbivores during certain stages in life, and herbivores that arrive on plants in specific ontogenetic stages can generate particularly strong selection pressures. Plant traits can be effective anti-herbivore defences during some plant developmental stages, but mediate ecological costs of herbivory in other plant development stages (Barton & Boege, 2017). The adaptive value of traits can therefore only be assessed when considering the complete life cycle of the organisms, and their interactions based on ecologically relevant timing. By determining direct and indirect interactions, ontogeny creates developmental stagespecific communities which may have profound effects on overall community structure and dynamics (Miller & Rudolf, 2011; Nakazawa, 2015). Moreover, community structure and dynamics may affect trait evolution (Agrawal, Hastings, Johnson, Maron, & Salminen, 2012; Guimarães Jr., Pires, Jordano, Bascompte, & Thompson, 2017; Siepielski & Benkman, 2004; Utsumi, Ando, Roininen, Takahashi, & Ohgushi, 2013), resulting in eco-evolutionary dynamics driven by ontogenetic variation (Ohgushi, 2016).

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AUTHORS' CONTRIBUTIONS

Q.R., D.L.-B. and E.H.P. planned and designed the study, interpreted the data and wrote the manuscript; Q.R. and K.H. collected the data; Q.R. analysed the data.

DATA AVAILABILITY STATEMENT

Data have been deposited in Dryad Digital Repository: https://doi. org/10.5061/dryad.j3tx95x94 (Rusman, Lucas-Barbosa, Hassan, & Poelman, 2020).

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REFERENCES

- Agrawal, A. A., Hastings, A. P., Johnson, M. T. J., Maron, J. L., & Salminen, J.-P. (2012). Insect herbivores drive real-time ecological and evolutionary change in plant populations. *Science*, 338, 113–116. https:// doi.org/10.1126/science.1225977
- Althoff, D. M., Xiao, W., Sumoski, S., & Segraves, K. A. (2013). Florivore impacts on plant reproductive success and pollinator mortality in an obligate pollination mutualism. *Oecologia*, 173, 1345–1354. https:// doi.org/10.1007/s00442-013-2694-8
- Barber, N. A., Adler, L. S., Theis, N., Hazzard, R. V., & Kiers, E. T. (2012). Herbivory reduces plant interactions with above-and belowground antagonists and mutualists. *Ecology*, 93, 1560–1570. https://doi. org/10.1890/11-1691.1
- Barber, N. A., Milano, N. J., Kiers, E. T., Theis, N., Bartolo, V., Hazzard, R. V., & Adler, L. S. (2015). Root herbivory indirectly affects aboveand below-ground community members and directly reduces plant performance. *Journal of Ecology*, 103, 1509–1518. https://doi. org/10.1111/1365-2745.12464
- Barton, K. E., & Boege, K. (2017). Future directions in the ontogeny of plant defence: Understanding the evolutionary causes and consequences. *Ecology Letters*, 20, 403–411. https://doi.org/10.1111/ ele.12744
- Barton, K. E., & Koricheva, J. (2010). The ontogeny of plant defense and herbivory: Characterizing general patterns using meta-analysis. *The American Naturalist*, 175, 481–493. https://doi.org/10.1086/650722
- Bates, D., Maechler, M., Bolker, B., & Walker, S. (2015). Fitting linear mixed-effects models using lme4. *Journal of Statistical Software*, 67, 1–48.
- Boege, K., Dirzo, R., Siemens, D., & Brown, P. (2007). Ontogenetic switches from plant resistance to tolerance: Minimizing costs with age? *Ecology Letters*, 10, 177–187. https://doi.org/10.1111/j.1461-0248.2006.01012.x
- Boege, K., & Marquis, R. J. (2005). Facing herbivory as you grow up: The ontogeny of resistance in plants. *Trends in Ecology & Evolution*, 20, 441–448. https://doi.org/10.1016/j.tree.2005.05.001
- Brütting, C., Schäfer, M., Vanková, R., Gase, K., Baldwin, I. T., & Meldau, S. (2017). Changes in cytokinins are sufficient to alter developmental patterns of defense metabolites in *Nicotiana*

attenuata. The Plant Journal, 89, 15–30. https://doi.org/10.1111/tpj. 13316

- Canela, M. B. F., & Sazima, M. (2003). Florivory by the crab Armases angustipes (Grapsidae) influences hummingbird visits to Aechmea pectinata (Bromeliaceae)1. Biotropica, 35, 289–294. https://doi. org/10.1111/j.1744-7429.2003.tb00287.x
- Cariveau, D., Irwin, R. E., Brody, A. K., Garcia-Mayeya, L. S., & Von Der Ohe, A. (2004). Direct and indirect effects of pollinators and seed predators to selection on plant and floral traits. *Oikos*, 104, 15–26. https://doi.org/10.1111/j.0030-1299.2004.12641.x|
- Chase, J. M. (2003). Community assembly: When should history matter? *Oecologia*, 136, 489–498. https://doi.org/10.1007/s00442-003-1311-7
- Chautá, A., Whitehead, S., Amaya-Márquez, M., & Poveda, K. (2017). Leaf herbivory imposes fitness costs mediated by hummingbird and insect pollinators. *PLoS ONE*, *12*, e0188408. https://doi.org/10.1371/journal. pone.0188408
- Chrétien, L. T. S., David, A., Daikou, E., Boland, W., Gershenzon, J., Giron, D., ... Lucas-Barbosa, D. (2018). Caterpillars induce jasmonates in flowers and alter plant responses to a second attacker. New Phytologist, 217, 1279–1291. https://doi.org/10.1111/nph.14904
- Conner, J. K., & Neumeier, R. (1995). Effects of black mustard population size on the taxonomic composition of pollinators. *Oecologia*, 104, 218–224. https://doi.org/10.1007/BF00328586
- de Rijk, M., Yang, D., Engel, B., Dicke, M., & Poelman, E. H. (2016). Feeding guild of non-host community members affects host-foraging efficiency of a parasitic wasp. *Ecology*, 97, 1388–1399. https://doi.org/ 10.1890/15-1300.1
- de Vries, J., Evers, J. B., Dicke, M., & Poelman, E. H. (2019). Ecological interactions shape the adaptive value of plant defence: Herbivore attack versus competition for light. *Functional Ecology*, 33, 129–138. https://doi.org/10.1111/1365-2435.13234
- de Vries, J., Evers, J. B., & Poelman, E. H. (2017). Dynamic plant-plantherbivore interactions govern plant growth-defence integration. *Trends in Plant Science*, 22, 329–337. https://doi.org/10.1016/j.tplants. 2016.12.006
- de Vries, J., Poelman, E. H., Anten, N., & Evers, J. B. (2018). Elucidating the interaction between light competition and herbivore feeding patterns using functional-structural plant modelling. *Annals of Botany*, 121, 1019-1031. https://doi.org/10.1093/aob/mcx212
- Desurmont, G. A., Laplanche, D., Schiestl, F. P., & Turlings, T. C. J. (2015). Floral volatiles interfere with plant attraction of parasitoids: Ontogeny-dependent infochemical dynamics in *Brassica rapa*. *BMC Ecology*, 15, 17. https://doi.org/10.1186/s12898-015-0047-7
- Diezel, C., Allmann, S., & Baldwin, I. T. (2011). Mechanisms of optimal defense patterns in *Nicotiana attenuata*: Flowering attenuates herbivory-elicited ethylene and jasmonate signaling. *Journal of Integrative Plant Biology*, 53, 971–983. https://doi.org/10.1111/j.1744-7909.2011.01086.x
- Dutton, E. M., Luo, E. Y., Cembrowski, A. R., Shore, J. S., & Frederickson, M. E. (2016). Three's a crowd: trade-offs between attracting pollinators and ant bodyguards with nectar rewards in *Turnera*. *The American Naturalist*, 188, 38–51. https://doi.org/10.1086/686766
- Ekbom, B., & Borg, A. (1996). Pollen beetle (Meligethes aeneus) oviposition and feeding preference on different host plant species. *Entomologia Experimentalis et Applicata*, 78, 291–299. https://doi.org/ 10.1111/j.1570-7458.1996.tb00793.x|
- Erwin, A. C., Züst, T., Ali, J. G., & Agrawal, A. A. (2014). Above-ground herbivory by red milkweed beetles facilitates above- and below-ground conspecific insects and reduces fruit production in common milkweed. *Journal of Ecology*, 102, 1038–1047. https://doi.org/10.1111/ 1365-2745.12248
- Fife, D. (2014). fifer: A collection of miscellaneous functions. R package version 1.0.
- Gómez-Marco, F., Tena, A., Jaques, J. A., & García, A. U. (2016). Early arrival of predators controls *Aphis spiraecola* colonies in citrus clementines.

Journal of Pest Science, 89, 69-79. https://doi.org/10.1007/s1034 0-015-0668-9

- Grass, I., Bohle, V., Tscharntke, T., & Westphal, C. (2018). How plant reproductive success is determined by the interplay of antagonists and mutualists. *Ecosphere*, *9*, e02106. https://doi.org/10.1002/ ecs2.2106
- Guimarães Jr., P. R., Pires, M. M., Jordano, P., Bascompte, J., & Thompson, J. N. (2017). Indirect effects drive coevolution in mutualistic networks. *Nature*, 550, 511–514. https://doi.org/10.1038/nature24273
- Hoballah, M. E. F., & Turlings, T. C. (2001). Experimental evidence that plants under caterpillar attack may benefit from attracting parasitoids. *Evolutionary Ecology Research*, *3*, 583–593.
- Hoffmeister, M., Wittköpper, N., & Junker, R. R. (2016). Herbivoreinduced changes in flower scent and morphology affect the structure of flower-visitor networks but not plant reproduction. *Oikos*, 125, 1241–1249. https://doi.org/10.1111/oik.02988
- Hol, W. H. G., Raaijmakers, C. E., Mons, I., Meyer, K. M., & van Dam, N. M.
 (2016). Root-lesion nematodes suppress cabbage aphid population development by reducing aphid daily reproduction. *Frontiers in Plant Science*, 7. https://doi.org/10.3389/fpls.2016.00111
- Hothorn, T., Bretz, F., & Westfall, P. (2008). Simultaneous inference in general parametric models. *Biometrical Journal*, 50, 346–363. https:// doi.org/10.1002/bimj.200810425
- Jacobsen, D. J., & Raguso, R. A. (2018). Lingering effects of herbivory and plant defenses on pollinators. *Current Biology*, 28, R1164–R1169. https://doi.org/10.1016/j.cub.2018.08.010
- Jönsson, M., Rosdahl, K., & Anderson, P. (2007). Responses to olfactory and visual cues by over-wintered and summer generations of the pollen beetle, *Meligethes aeneus*. *Physiological Entomology*, *32*, 188–193. https://doi.org/10.1111/j.1365-3032.2007.00562.x
- Kassambara, A. (2018). ggpubr: 'ggplot2' based publication ready plots. R package version 0.1.
- Kuppler, J., Höfers, M. K., Wiesmann, L., & Junker, R. R. (2016). Timeinvariant differences between plant individuals in interactions with arthropods correlate with intraspecific variation in plant phenology, morphology and floral scent. *New Phytologist*, 210, 1357–1368. https://doi.org/10.1111/nph.13858
- Lenth, R. (2016). Least-squares means: The R package Ismeans. *Journal of Statistical Software*, *69*, 1–33.
- Lucas-Barbosa, D. (2016). Integrating studies on plant-pollinator and plant-herbivore interactions. *Trends in Plant Science*, *21*, 125–133. https://doi.org/10.1016/j.tplants.2015.10.013
- Lucas-Barbosa, D., Dicke, M., Kranenburg, T., Aartsma, Y., van Beek, T. A., Huigens, M. E., & van Loon, J. J. A. (2017). Endure and call for help: Strategies of black mustard plants to deal with a specialized caterpillar. *Functional Ecology*, 31, 325–333. https://doi.org/ 10.1111/1365-2435.12756
- Lucas-Barbosa, D., van Loon, J. J. A., & Dicke, M. (2011). The effects of herbivore-induced plant volatiles on interactions between plants and flower-visiting insects. *Phytochemistry*, 72, 1647–1654. https://doi. org/10.1016/j.phytochem.2011.03.013
- Lucas-Barbosa, D., van Loon, J. J. A., Gols, R., Beek, T. A., & Dicke, M. (2013). Reproductive escape: Annual plant responds to butterfly eggs by accelerating seed production. *Functional Ecology*, 27, 245–254. https://doi.org/10.1111/1365-2435.12004
- McArt, S. H., Halitschke, R., Salminen, J.-P., & Thaler, J. S. (2013). Leaf herbivory increases plant fitness via induced resistance to seed predators. *Ecology*, 94, 966–975. https://doi.org/10.1890/12-1664.1
- Meier, U. (2001). Growth stages of mono-and dicotyledonous plants. Berlin, Germany: Blackwell Wissenschafts-Verlag.
- Miller, T. E. X., & Rudolf, V. H. W. (2011). Thinking inside the box: Community-level consequences of stage-structured populations. *Trends in Ecology & Evolution*, 26, 457–466. https://doi.org/10.1016/j. tree.2011.05.005

- Mira, A., & Bernays, E. A. (2002). Trade-offs in host use by *Manduca sexta*: Plant characters vs natural enemies. *Oikos*, *97*, 387–397. https://doi. org/10.1034/j.1600-0706.2002.970309.x
- Moreira, X., Castagneyrol, B., Abdala-Roberts, L., & Traveset, A. (2019). A meta-analysis of herbivore effects on plant attractiveness to pollinators. *Ecology*, 100, e02707. https://doi.org/10.1002/ecy.2707
- Nakazawa, T. (2015). Ontogenetic niche shifts matter in community ecology: A review and future perspectives. *Population Ecology*, *57*, 347–354. https://doi.org/10.1007/s10144-014-0448-z
- Ochoa-López, S., Rebollo, R., Barton, K. E., Fornoni, J., & Boege, K. (2018). Risk of herbivore attack and heritability of ontogenetic trajectories in plant defense. *Oecologia*, 187, 413–426. https://doi.org/10.1007/ s00442-018-4077-7
- Ochoa-López, S., Villamil, N., Zedillo-Avelleyra, P., & Boege, K. (2015). Plant defence as a complex and changing phenotype throughout ontogeny. *Annals of Botany*, 116, 797–806. https://doi.org/10.1093/ aob/mcv113
- Ohgushi, T. (2016). Eco-evolutionary dynamics of plant-herbivore communities: Incorporating plant phenotypic plasticity. *Current Opinion in Insect Science*, 14, 40–45. https://doi.org/10.1016/j.cois. 2016.01.006
- Pashalidou, F. G., Frago, E., Griese, E., Poelman, E. H., Loon, J. J. A., Dicke, M., & Fatouros, N. E. (2015). Early herbivore alert matters: Plantmediated effects of egg deposition on higher trophic levels benefit plant fitness. *Ecology Letters*, 18, 927–936. https://doi.org/10.1111/ ele.12470
- Pierre, P. S., Dugravot, S., Ferry, A., Soler, R., Van Dam, N. M., & Cortesero, A. M. (2011). Aboveground herbivory affects indirect defences of brassicaceous plants against the root feeder *Delia radicum* Linnaeus: Laboratory and field evidence. *Ecological Entomology*, 36, 326–334. https://doi.org/10.1111/j.1365-2311. 2011.01276.x
- Pierre, P. S., Jansen, J. J., Hordijk, C. A., Van Dam, N. M., Cortesero, A.-M., & Dugravot, S. (2011). Differences in volatile profiles of turnip plants subjected to single and dual herbivory above-and belowground. *Journal of Chemical Ecology*, 37, 368–377. https://doi.org/10.1007/ s10886-011-9934-3
- Pineda, A., Soler, R., Pastor, V., Li, Y., & Dicke, M. (2017). Plant-mediated species networks: The modulating role of herbivore density. *Ecological Entomology*, 42, 449–457. https://doi.org/10.1111/een.12404
- Poelman, E. H., & Kessler, A. (2016). Keystone herbivores and the evolution of plant defenses. *Trends in Plant Science*, 21, 477–485. https:// doi.org/10.1016/j.tplants.2016.01.007
- Ponzio, C., Papazian, S., Albrectsen, B. R., Dicke, M., & Gols, R. (2017). Dual herbivore attack and herbivore density affect metabolic profiles of *Brassica nigra* leaves. *Plant, Cell & Environment*, 40, 1356–1367. https://doi.org/10.1111/pce.12926
- Poveda, K., Steffan-Dewenter, I., Scheu, S., & Tscharntke, T. (2005). Effects of decomposers and herbivores on plant performance and aboveground plant-insect interactions. *Oikos*, 108, 503–510. https:// doi.org/10.1111/j.0030-1299.2005.13664.x
- Quesada, M., Bollman, K., & Stephenson, A. G. (1995). Leaf damage decreases pollen production and hinders pollen performance in *Cucurbita texana*. *Ecology*, 76, 437–443. https://doi.org/10.2307/1941202
- Quintero, C., Barton, K. E., & Boege, K. (2013). The ontogeny of plant indirect defenses. *Perspectives in Plant Ecology, Evolution and Systematics*, 15, 245–254. https://doi.org/10.1016/j.ppees.2013.08.003
- Quintero, C., Lampert, E. C., & Bowers, M. D. (2014). Time is of the essence: Direct and indirect effects of plant ontogenetic trajectories on higher trophic levels. *Ecology*, 95, 2589–2602. https://doi. org/10.1890/13-2249.1
- Rostás, M., & Eggert, K. (2008). Ontogenetic and spatio-temporal patterns of induced volatiles in *Glycine max* in the light of the optimal defence hypothesis. *Chemoecology*, 18, 29–38. https://doi.org/10.1007/ s00049-007-0390-z

- Rusman, Q., Lucas-Barbosa, D., Hassan, K., & Poelman, E. H. (2020). Data from: Plant ontogeny determines strength and associated plant fitness consequences of plant-mediated interactions between herbivores and flower visitors. *Dryad Digital Repository*, https://doi. org/10.5061/dryad.j3tx95x94
- Rusman, Q., Lucas-Barbosa, D., & Poelman, E. H. (2018). Dealing with mutualists and antagonists: Specificity of plant-mediated interactions between herbivores and flower visitors, and consequences for plant fitness. *Functional Ecology*, *32*, 1022–1035. https://doi. org/10.1111/1365-2435.13035
- Rusman, Q., Lucas-Barbosa, D., Poelman, E. H., & Dicke, M. (2019). Ecology of plastic flowers. *Trends in Plant Science*, 24, 725–740. https://doi.org/10.1016/j.tplants.2019.04.007
- Rusman, Q., Poelman, E. H., Nowrin, F., Polder, G., & Lucas-Barbosa, D. (2019). Floral plasticity: Herbivore-species-specific-induced changes in flower traits with contrasting effects on pollinator visitation. *Plant, Cell & Environment, 42, 1882–1896.* https://doi.org/10.1111/ pce.13520
- Schiestl, F. P., Kirk, H., Bigler, L., Cozzolino, S., & Desurmont, G. A. (2014). Herbivory and floral signaling: Phenotypic plasticity and tradeoffs between reproduction and indirect defense. *New Phytologist*, 203, 257–266. https://doi.org/10.1111/nph.12783
- Sharpe, D. (2015). Your chi-square test is statistically significant: Now what? Pratical Assessment, Research & Evaluation, 20, 2–10.
- Siepielski, A. M., & Benkman, C. W. (2004). Interactions among moths, crossbills, squirrels, and lodgepole pine in a geographic selection mosiac. *Evolution*, 58, 95–101. https://doi.org/10.1111/j.0014-3820. 2004.tb01576.x
- Söber, V., Moora, M., & Teder, T. (2010). Florivores decrease pollinator visitation in a self-incompatible plant. *Basic and Applied Ecology*, 11, 669–675. https://doi.org/10.1016/j.baae.2010.09.006
- Soler, R. G. (2009). Plant-mediated multitrophic interactions between aboveground and belowground insects. *Entomologische Berichten*, 69, 202–210.
- Soler, R., Badenes-Pérez, F. R., Broekgaarden, C., Zheng, S.-J., David, A., Boland, W., & Dicke, M. (2012). Plant-mediated facilitation between a leaf-feeding and a phloem-feeding insect in a brassicaceous plant: From insect performance to gene transcription. *Functional Ecology*, 26, 156–166. https://doi.org/10.1111/j.1365-2435.2011.01902.x
- Soler, R., Bezemer, T., Van Der Putten, W. H., Vet, L. E., & Harvey, J. A. (2005). Root herbivore effects on above-ground herbivore, parasitoid and hyperparasitoid performance via changes in plant quality. Journal of Animal Ecology, 74, 1121–1130. https://doi.org/ 10.1111/j.1365-2656.2005.01006.x
- Soper Gorden, N. L., & Adler, L. S. (2016). Florivory shapes both leaf and floral interactions. *Ecosphere*, 7, e01326. https://doi.org/10.1002/ ecs2.1326
- Soper Gorden, N. L., & Adler, L. S. (2018). Consequences of multiple flower-insect interactions for subsequent plant-insect interactions and plant reproduction. *American Journal of Botany*, 105, 1–12. https://doi.org/10.1002/ajb2.1182
- Stam, J. M., Chrétien, L., Dicke, M., & Poelman, E. H. (2017). Response of *Brassica oleracea* to temporal variation in attack by two herbivores affects preference and performance of a third herbivore. *Ecological Entomology*, 42, 803–815. https://doi.org/10.1111/een. 12455
- Stam, J. M., Dicke, M., & Poelman, E. H. (2018). Order of herbivore arrival on wild cabbage populations influences subsequent arthropod community development. *Oikos*, 127, 1482–1493. https://doi.org/ 10.1111/oik.05265
- Stam, J. M., Kos, M., Dicke, M., & Poelman, E. H. (2019). Crossseasonal legacy effects of arthropod community on plant fitness in perennial plants. *Journal of Ecology*, 107, 2451–2463. https://doi. org/10.1111/1365-2745.13231

- Strauss, S. Y. (1997). Floral characters link herbivores, pollinators, and plant fitness. *Ecology*, 78, 1640–1645. https://doi.org/10.1890/0012-9658(1997)078[1640:FCLHPA]2.0.CO;2
- Strauss, S. Y., Conner, J. K., & Rush, S. L. (1996). Foliar herbivory affects floral characters and plant attractiveness to pollinators: Implications for male and female plant fitness. *The American Naturalist*, 147, 1098–1107. https://doi.org/10.1086/285896
- Strauss, S. Y., Rudgers, J. A., Lau, J. A., & Irwin, R. E. (2002). Direct and ecological costs of resistance to herbivory. *Trends in Ecology* & *Evolution*, 17, 278–285. https://doi.org/10.1016/S0169-5347 (02)02483-7
- Theis, N., & Adler, L. S. (2012). Advertising to the enemy: Enhanced floral fragrance increases beetle attraction and reduces plant reproduction. *Ecology*, *93*, 430–435. https://doi.org/10.1890/11-0825.1
- Tonnabel, J., David, P., & Pannell, J. R. (2017). Sex-specific strategies of resource allocation in response to competition for light in a dioecious plant. *Oecologia*, 185, 675–686. https://doi.org/10.1007/s00442-017-3966-5
- Utsumi, S., Ando, Y., Roininen, H., Takahashi, J.-I., & Ohgushi, T. (2013). Herbivore community promotes trait evolution in a leaf beetle via induced plant response. *Ecology Letters*, 16, 362–370. https://doi. org/10.1111/ele.12051
- van Dam, N. M., Wondafrash, M., Mathur, V., & Tytgat, T. O. G. (2018). Differences in hormonal signaling triggered by two root-feeding nematode species result in contrasting effects on aphid population growth. Frontiers in Ecology and Evolution, 6. https://doi.org/10.3389/ fevo.2018.00088
- Van Zandt, P. A., & Agrawal, A. A. (2004). Community-wide impacts of herbivore-induced plant responses in milkweed (Asclepias syriaca). Ecology, 85, 2616–2629. https://doi.org/10.1890/03-0622
- Vannette, R. L., & Fukami, T. (2014). Historical contingency in species interactions: Towards niche-based predictions. *Ecology Letters*, 17, 115–124. https://doi.org/10.1111/ele.12204
- Villamil, N. (2017). Why are flowers sweeter than fruits or buds? Variation in extrafloral nectar secretion throughout the floral ontogeny of a myrmecophile. *Biotropica*, 49, 581–585. https://doi.org/10.1111/btp. 12463
- Waltz, A. M., & Whitham, T. G. (1997). Plant development affects arthropod communities: Opposing impacts of species removal. *Ecology*, 78, 2133–2144. https://doi.org/10.1890/0012-9658(1997)078[2133: PDAACO]2.0.CO;2
- Wenninger, A., Kim, T., Spiesman, B., & Gratton, C. (2016). Contrasting foraging patterns: Testing resource-concentration and dilution effects with pollinators and seed predators. *Insects*, 7, 23. https://doi. org/10.3390/insects7020023
- Zhang, H., Zhou, D., Huang, Y., Japhet, W., & Sun, D. (2008). Plasticity and allometry of meristem allocation in response to density in three annual plants with different architectures. *Botany-Botanique*, *86*, 1291–1298. https://doi.org/10.1139/b08-079

SUPPORTING INFORMATION

Additional supporting information may be found online in the Supporting Information section.

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