

On farm observations to increase genetic gain in breeding schemes for village poultry production–A simulation study

Chu, T. T., Bastiaansen, J. W. M., Norberg, E., & Berg, P.

This is a "Post-Print" accepted manuscript, which has been published in "Acta Agriculturae Scandinavica A: Animal Sciences"

This version is distributed under a non-commercial no derivatives Creative Commons (CC-BY-NC-ND) user license, which permits use, distribution, and reproduction in any medium, provided the original work is properly cited and not used for commercial purposes. Further, the restriction applies that if you remix, transform, or build upon the material, you may not distribute the modified material.

Please cite this publication as follows:

Chu, T. T., Bastiaansen, J. W. M., Norberg, E., & Berg, P. (2018). On farm observations to increase genetic gain in breeding schemes for village poultry production–A simulation study. Acta Agriculturae Scandinavica A: Animal Sciences. DOI: 10.1080/09064702.2018.1543444

You can download the published version at:

https://doi.org/10.1080/09064702.2018.1543444

1	On farm observations to increase genetic gain in breeding schemes for village
2	poultry production – A simulation study
3	Thinh Tuan Chu ^{1,2,4} , John W.M. Bastiaansen ² , Elise Norberg ^{1,3} & Peer Berg ^{1,3}
4	
5	¹ Center for Quantitative Genetics and Genomics, Department of Molecular Biology and
6	Genetics, Aarhus University, 8830 Tjele, Denmark
7	² Wageningen University & Research Animal Breeding and Genomics, 6709 PG Wageningen,
8	The Netherlands
9	³ Department of Animal and Aquacultural Sciences, Norwegian University of Life Sciences, 1432
10	Ås, Norway
11	⁴ Corresponding author's email: <u>chu.thinh@mbg.au.dk</u>
12	

14 Abstract

To improve genetic gain of breeding programs for village poultry production, breeding schemes 15 with observations obtained in village production systems using individual (VIO) and group 16 17 recording (VGO) were examined under different levels of genotype-by-environment-interactions (GxE). GxE was modelled by varying the correlation between traits measured in the breeding 18 station and village environments for bodyweight (r_{g_BW}) and egg production (r_{g_EP}) . Relative and 19 20 absolute genetic gains obtained from VIO and VGO were used for comparison between the schemes. Results showed that village observations significantly improved genetic gains compared 21 to the scheme without birds tested in the village. The improvement was only slightly larger with 22 individual observations than with group observations. Higher rg BW and rg EP led to lower relative 23 genetic gain, but higher absolute gain of VIO and VGO. It is recommended to apply a breeding 24 scheme using group recording of village performance when strong GxE in breeding for village 25 poultry is expected. 26

27 *Key words: breeding scheme; GxE; group recording; stochastic simulation; village poultry*

28 Introduction

Introduction of exotic breeds or high yielding hybrids has failed to upgrade the genetic level of the current chicken populations in Ethiopia due to various reasons, such as farmer preference, lack of required input and chicken adaptability (Dana et al., 2010; Wondmeneh et al., 2015). In addition, the application of exotic breeds in an intensive or semi-intensive production system for smallholder villagers brings in lower economic returns than the use of indigenous chicken under a scavenging production system (FAO, 2010; Okeno et al., 2013). Moreover, one of the biggest advantages of indigenous chicken is their disease resistance and adaptability to harsh conditions (Dessie et al., 2000). Therefore, a key approach for delivering a productive and adapted chicken suitable for the
production system and acceptable to the farmers, is to improve the indigenous chicken through
breeding programs.

A selective breeding program was initiated in 2008 at the Debre Zeit Agricultural Research Centre 39 40 in Ethiopia (Dana et al., 2011). The ultimate objective of the breeding program is an improved dual-purpose chicken (Horro) for growth and egg production, which also is well-adapted to the 41 semi-scavenging environment of village poultry production. However, the breeding scheme of the 42 program has revealed to be suboptimal as it has shown slow genetic progress and signs of losing 43 44 adaptability of indigenous chicken after 7 generations of selection (Wondmeneh, 2015). The differences between the conditions at the research station and villages might cause genotype by 45 environment interaction (GxE). At the research station, birds are selected under hygienic 46 conditions, nutritionally adequate diets and well-protected cages, whereas at the villages, birds are 47 48 subjected to a combination of low food availability, sub-optimal diet, prevalence of diseases and other social interaction factors. 49

Significant GxE in poultry has been reported in a number of studies (Bekele et al., 2009; Chen et 50 al., 2009; Horst, 1985; Kapell et al., 2012; Mathur & Horst, 1994; N'Dri et al., 2007) and reviews 51 (FAO, 2010; Mathur, 2003). GxE could reduce potential genetic gains of a breeding program. 52 There are, however, only a few studies (Bijma & Arendonk, 1998; Mulder & Bijma, 2005) on 53 design or evaluation of breeding schemes in the presence of GxE, and they are mainly designed 54 for other species than poultry and for commercial production instead of village production. A big 55 challenge for implementing breeding schemes for village poultry is the need for routine collection 56 of observations on individual animals. Group mean of full-sibs and half-sibs can be a possible 57 alternative for village phenotype recording. Studies on pooled data has illustrated that selection 58

based on estimated breeding values (EBV) from pooled observations can be effective, particularly
when group members have close relationships (Biscarini et al., 2008; Nurgiartiningsih et al., 2004;
Olson et al., 2006; Peeters et al., 2013). However, the use of pooled observations in breeding
programs where GxE is present, and where animals with the pooled observations are not candidates
of selection, has not been demonstrated.

This paper proposes breeding schemes for village dual-purpose poultry production in the presence of GxE. Stochastic simulation is applied to compare breeding schemes on genetic gain considering group and individual recording and to optimize the data recording effort in villages versus stations. GxE was modelled by varying the correlation between traits measured in station and village environments.

69 Materials and methods

70 Breeding schemes

The stochastic simulation program ADAM (Pedersen et al., 2009) was used to simulate 100 71 replicates for each scenario. The simulation mimicked the situation of the Horro chicken breeding 72 73 population at the Debre Zeit Agricultural Research Centre, Ethiopia (Dana et al., 2011). The schemes were designed for dual-purpose village poultry production, by including body weight 74 (BW) and egg production (EP) in the breeding goal (Figure 1). The breeding structure consisted 75 76 of 30 roosters and 300 hens. In each generation, a hen had 4 offspring that were candidates for selection and an additional number of offspring for testing. Sex was randomly assigned to offspring 77 at a 50:50 ratio. The candidates for selection were reared in a research station. Under the station 78 79 conditions, the birds had phenotypes defined as "station" traits. Birds for testing were transferred to village small holders for recording of phenotypes, which were defined as "village" traits. The 80

village tested birds were not considered as selection candidates, but only gave information for
evaluating station selection candidates.

In each generation, selection candidates went through two selection rounds. In the first selection 83 round, 150 of all male candidates were selected after phenotypes for BW were measured both in 84 station and in village. No selection was applied in the females. This round was to ensure a high 85 selection response and to reduce costs of keeping all male candidates until EP was recorded. In the 86 second round, 30 males were selected out of the remaining 150 candidates and 300 females were 87 selected out of all female candidates. Selection round 2 was performed after phenotypes for EP 88 were realized. BLUP selection was applied for both the selection rounds, so information about 89 90 relatives both in station and in village was used. Selection was simulated for 20 discrete generations. 91

92 Trait simulation

Phenotypes of BW and EP were simulated and BW was observed for both male and female birds while EP was observed in females only. In the station environment, BW and EP were denoted as BW_s and EP_s, respectively, whereas in the village environment, the phenotypes were denoted as BW_v and EP_v, respectively. Observations on BW_s and EP_s were realized individually, while BW_v and EP_v were recorded as either group mean or individually. Group records were the average of the simulated phenotypes of 10 paternal-sibs, which were randomly selected from the 40 offspring of a sire. Therefore, members of a group could have both full-sib and half-sib relationship.

100 The genetic parameters assumed for all traits are shown in Table 1. The parameters of 101 (co)variances, correlation and heritabilities of BW_s and EP_s were based on literature for indigenous 102 chicken in Africa (Dana et al., 2011; Lwelamira et al., 2009; Niknafs et al., 2012; Oleforuh-Okoleh, 103 2011). We assumed that the village scavenging system would result in a larger environmental variance and a lower heritability compared to the conditions on station. The heritabilities for 104 village traits was set to half the values for the station traits. Additive genetic variances of BW_s and 105 EPs were assumed equal to those of BWv and EPv, respectively. Genetic correlation between BWs 106 and EP_s was also equal to that of BW_y and EP_y. Genetic correlations between the village and station 107 environments for BW (rg BW) and EP (rg EP) were varied to reflect different extent of GxE. To 108 ensure a positive-definite matrix of genetic covariance, the genetic correlation between BWs and 109 EP_v was approximated by multiplying the average of $r_{g BW}$ and $r_{g EP}$ by correlation between BWs 110 111 and EP_s (Table 1). This approximation came from assuming that the link between BW_s and EP_v might be through either one of two paths. One path was through correlation between BWs and EPs 112 and correlation between EP_s and EP_y, and another path was through correlation between BW_s and 113 BW_v and correlation between BW_v and EP_v . The genetic correlation between BW_v and EP_s was 114 approximated in the same way. Environmental correlation between BW_v and EP_v was assumed to 115 be equal to that between BWs and EPs. Other environmental correlations between traits were set 116 to 0 because birds only had records either on the station or in the village environment. 117

True breeding values of BW_s, EP_s, BW_v and EP_v traits of a bird *i* at generation 0 were scaled to 118 achieve an initial genetic covariance matrix by following equation: $\mathbf{tbv}_i = \mathbf{L}' \times \mathbf{r}$, where \mathbf{tbv}_i is a 119 vector of true breeding values of bird *i*; \mathbf{L}' is the Cholesky decomposition of the initial genetic 120 covariance matrix; and \mathbf{r} is a vector of random numbers from a standardized normal distribution. 121 Means of the traits were 0. Simulation of environmental values of the traits was similar to 122 simulation of true breeding values, with a Cholesky decomposition of the environmental 123 covariance matrix. Phenotypic observation of a trait for an individual was the sum of true breeding 124 125 value and environmental value. Environmental (co)variances were kept constant through the simulations whereas genetic (co)variance and heritability decreased due to Bulmer effect of selection and inbreeding. True breeding value of the descendants was half of true breeding values of their parents plus Mendelian sampling terms. Mendelian sampling variance of the offspring was determined based on the inbreeding of the parents.

130 Simulation of group mean observations was done in two steps. The first step was simulation of individual phenotypic observations as described above. The second step was to compute group 131 mean observations. All offspring birds of a sire in a village were randomly assigned into groups 132 of 10 birds. Individual phenotypic observations of those offspring birds were used to calculate 133 group means. Subsequently, the individual phenotypic observations were replaced by group 134 means. For BW_v , 10 paternal-sib of a group had the same group mean observation. For EP_v , 135 phenotypic observations of females of the 10 paternal-sib group were used to calculate the group 136 mean, and phenotypic observations of these females were replaced by the mean. 137

138 *Simulated scenarios*

A reference breeding scheme and 2 alternative breeding schemes were simulated (Table 2). The 139 140 reference breeding scheme had 1200 candidates for selection and 600, 1200 or 1800 tested birds. Both the selection candidates and tested birds provided information of station phenotypes. For the 141 two alternative schemes, the tested birds did not provide information of station phenotypes but 142 were transferred to village environment to get village phenotypes. In one of the alternatives 143 (breeding scheme VIO), the village birds had individual observations. In the other (breeding 144 scheme VGO), the birds had group mean observation of 10 paternal-sibs. As suggested in Cahaner 145 et al. (1993), Kapell et al. (2012), Mathur and Horst (1994), Mathur (2003) and Chen et al. (2009), 146 a stronger GxE interaction was simulated for EP than for BW, and therefore a lower genetic 147

correlation between station and village measures. The lower correlation for EP than BW came from assumption that traits of reproduction have stronger GxE interaction than traits of production, and traits with lower heritability generally display higher GxE (Mathur, 2003). The values of r_{g_BW} were set at 0.5, 0.7 and 0.9 and r_{g_BP} were 0.1, 0.3 and 0.5.

- 152 As a consequence, there were 4 factors investigated: type of breeding schemes, number of tested
- birds, r_{g BW} and r_{g EP}. All three breeding schemes were simulated with all three numbers of tested
- birds and all 9 combinations of r_{g_BW} and r_{g_EP} resulting in a total of 81 simulated scenarios.

155 Selection criteria

Breeding was done to optimize production in the village environment and therefore the breedinggoal was as follows:

158
$$H = 0^* BW_s + 0^* EP_s + 0.078^* BW_v + 9.080^* EP_v$$
 (1)

An economic value of 0 was assigned to the station traits of BW_s and EP_s with the assumption that only village performance mattered. Economic values given to BW_v and EP_v were from Okeno et al. (2012). Unit of BW was measured in grammes, and EP was cumulative number of eggs produced until 40 weeks of age.

Breeding values were estimated based on data from VIO and VGO using multivariate best linear
unbiased prediction (BLUP) models. For individual phenotypic observation, the model was:

$$165 \quad \mathbf{y} = \mathbf{X}\mathbf{b} + \mathbf{Z}\mathbf{a} + \mathbf{e} \tag{2}$$

where **y** is a vector of individual phenotypic records of traits of BW_s, EP_s, BW_v and EP_v; **b** is a vector of fixed year effects; **a** is a vector of animal breeding values of the traits to be estimated assumed **a** ~ $MVN[0, \mathbf{A} \otimes \mathbf{G}]$, where MVN is the multivariate normal distribution, **A** is the additive genetic relationship matrix among individuals and **G** is the additive genetic (co)variance matrix among the traits as a 4x4 matrix; **X** and **Z** are incidence matrices relating fixed effects and breeding values to phenotypic observations of birds; and **e** is a vector of residuals of the traits assumed $\mathbf{e} \sim MVN \begin{bmatrix} 0 & \mathbf{I}_s \otimes \mathbf{E}_s & 0 \\ 0' & 0 & \mathbf{I}_v \otimes \mathbf{E}_v \end{bmatrix}$, where \mathbf{I}_s and \mathbf{I}_v are identity matrices of station and village observations, respectively, \mathbf{E}_s and \mathbf{E}_v are the environmental covariance matrices (2x2) of the station traits (BW_s and EP_s) and the village traits (BW_v and EP_v), respectively.

For group observations, the same model as (2) was used, except that group averages of the 10
paternal-sibs were treated as if they were individual phenotypic records of each of the ten birds.
This is an approximate approach described in Olson et al. (2006).

Selection for the reference breeding scheme was also based on breeding values estimated using
model (2), except that the model was a bivariate model applied for 2 traits of BW_s and EP_s only.
Selection for the reference scheme was indirect selection, in which selection index was:

181
$$I = 0.078 * BW_s + 9.080 * EP_s$$
 (3)

However, (1) was still used as true breeding goal to assess genetic gain of all scenarios.

A combined measure of GxE, which represents the correlation between performances in the two environments, were based on the values of r_{g_BW} and r_{g_EP} with their economic indexes. Genetic correlation between (1) and (3) (r_{g_HI}) was calculated as:

186
$$r_{g_HI} = \frac{Cov(H;I)}{\sqrt{Var(H) \times Var(I)}}$$
(4)

187 Where *Cov(H; I)* is genetic covariance between *H* and *I*; *Var(H)* is genetic variance of *H*; *Var(I)*188 is genetic variance of *I*.

189 Data analysis

For all scenarios, simulated output of total index genetic merit and the inbreeding coefficient from generation 5 to 20 were used for analyses. The index genetic merit of a scenario was the sum of true breeding values indexed with their economic values as in (1). The genetic merit of generation t, G_t , was the average of index true breeding values of all new-born individuals at generation t. Similarly, the inbreeding coefficient at generation t, F_t , was the average of inbreeding coefficients of individuals calculated by pedigree information.

For each replicate, genetic gain per generation (ΔG) was computed as the difference between G_{20} and G_5 . The relative genetic gain per generation (RG) of VIO and VGO scenarios was calculated as the differences between their genetic gains and the mean of genetic gain of the corresponding reference scenarios divided by the mean of genetic gain of the reference scenarios.

200
$$RG = \frac{\Delta G_{alternative \, scenario} - Average \, \Delta G_{reference \, scenario}}{Average \, \Delta G_{reference \, scenario}} \times 100\%$$

Where *RG* is relative genetic gain per generation of VIO or VGO scenario over the reference scenario; $\Delta G_{alternative scenario}$ is genetic gain per generation of a replicate of VIO or VGO scenario; *Average* $\Delta G_{reference scenario}$ is the mean of genetic gain of 100 replicates of the reference scenario corresponding to the VIO or VGO scheme that had the same number of tested animals, and same r_{g_BW} and r_{g_EP} .

Rate of inbreeding per generation were computed as the negative of the slope of the regression of $ln(1-F_t)$ on *t* for F_{5} - F_{20} (Nirea et al., 2012).

208 Summary statistics for *RG* of VIO and VGO scenarios were based on 100 replicates. ANOVA 209 were used to test direct and interaction effects of various factors on *RG*. The differences between

scenarios were tested for significance using Tukey's HSD (honest significant difference, P < 0.05). Summary statistics for rate of inbreeding of scenarios of VIO, VGO and reference schemes were also computed.

213 **Results**

The 4-way interaction of breeding scheme, r_{g_BW} , r_{g_EP} and number of tested animals were significant on *RG* with *p* <0.0001. As can be seen in Figure 2, all scenarios of VIO and VGO breeding schemes had genetic gain greater than the scenarios of the corresponding reference scheme. Relative genetic gains ranged from 21 to 268%.

The addition of birds tested in the village condition increased *RG*. When number of village tested birds was 600, 1200 and 1800, on average, *RG* was 84, 98 and 112%, respectively.

The VIO breeding scheme had higher *RG* than VGO breeding scheme. On average, *RG* of VIO was 102% while it was 94% for VGO. With 600, 1200 and 1800 tested birds, *RG* of VGO were 81, 94 and 107%, respectively, and *RG* of VIO were 87, 102 and 117%, respectively. In all cases with the same number of village tested birds and the same correlations of r_{g_BW} and r_{g_EP} , *RG* of VGO was lower than that of VIO.

Lower genetic correlations between traits measured on station and village environments, lead to higher *RG*. With the maximum values for $r_{g_{EP}}$ of 0.5 and $r_{g_{BW}}$ of 0.9, on average, *RG* was 28% whereas *RG* was 225% for scenarios with the minimum values for $r_{g_{EP}}$ of 0.1 and $r_{g_{BW}}$ of 0.5. It seems that the magnitude of increase in *RG* was higher with a reduction in $r_{g_{EP}}$ than with the reduction in $r_{g_{BW}}$. To have a better explanation of the trend of *RG*, $r_{g_{HI}}$ can be used as an assessment of indirect selection of selection index I to the true breeding goal. *RG* decreased with increasing $r_{g_{HI}}$ (Table 3). Genetic gains per generation of VIO and VGO breeding scenarios can be seen in Table 4. Similar to *RG*, ΔG of VIO and VGO scenarios increased with increasing number of village tested animals. Genetic gains of VIO scenarios were also higher than that of VGO scenarios. However, unlike *RG*, lower genetic correlations between traits measured in station and village environments ($r_{g_{EP}}$ and $r_{g_{BW}}$) resulted in lower ΔG . Meanwhile, ΔG increases with a higher genetic correlation between environments ($r_{g_{HI}}$), but at a lower relative increase as can be seen in Table 4.

The rates of inbreeding decreased as number of tested animals increased. They, on average, were 2.00, 1.99 and 1.97% for scenarios with 600, 900 and 1800 tested animals, respectively. Higher rates of inbreeding were found in the VGO scenarios than in the VIO scenarios. The rates of inbreeding, on average, were 1.70% for the reference scenarios, 2.10% for VIO scenarios and 2.15% for VGO scenarios. The rates of inbreeding had a reducing tendency as $r_{g_{e}}P$ and $r_{g_{e}}P$ increased.

244 **Discussion**

In this study, breeding schemes for village dual-purpose poultry using group and individual recordings of village and station performances at different levels of GxE interaction were compared. Results showed that village observations significantly improved genetic gains of VIO and VGO compared to the reference breeding scheme. The improvement was larger in VIO than in VGO. Increasing number of village tested birds also increased genetic gain. Higher genetic correlations between traits measured in station and village environments lead to lower relative genetic gain, but higher absolute genetic gain.

252 *Effects of village observation on genetic gain*

Increasing the number of animals tested in village improved accuracy of selection, and thus genetic gains of VIO and VGO schemes compared to the reference schemes. The main difference between the reference breeding scheme and its alternatives was the type of tested information. Village observations were direct phenotypes while station observations were correlated phenotypes. The reference scheme had only station performance while its alternatives had both station and village performances. Selection in the reference scheme is an indirect selection approach, and therefore, it results in the lowest accuracy of selection compared to its alternatives.

The value of village observations increased when the genetic correlation between station and village traits was lower. The scenarios with r_{g_BW} of 0.9 and r_{g_EP} of 0.5 gave lower relative genetic gain than those with r_{g_BW} of 0.5 and r_{g_EP} of 0.1. Nonetheless, the absolute genetic gain was larger in scenarios with higher genetic correlations as contribution of both station and village observations to accuracy of selection increased.

It has been suggested that if the genetic correlation between performance in the selection and 265 266 production environments is less than 0.8, breeding program with information from the production environment would be worthwhile to improve genetic gain (Robertson, 1959). Other studies have 267 also shown that a significantly higher genetic gain can be achieved with performance information 268 269 from the production environment (Bijma & Arendonk, 1998; Mulder & Bijma, 2005). However, when genetic correlation between the performance in selection and production environments is 270 high, for example 0.9, a large number of animals need to be tested in the production environment 271 for a significant improvement in genetic gain. 272

In our study, relative genetic gains were positive in all scenarios of VIO and VGO with any number of birds tested in village, r_{g_BW} or r_{g_EP} . In other studies, to model GxE, a single trait in two

275 environments is often used (Bijma & Arendonk, 1998; Mulder & Bijma, 2005). To be comparable to other studies, instead of rg BW and rg EP, rg HI should be used as a representative of genetic 276 correlation between station and village environments. It takes into account the variances and 277 covariances of BW and EP traits measured in the two environments with their economic indexes. 278 The value of rg_HI reflects the magnitude of indirect selection on the selection index I to the true 279 breeding goal. It describes the extent of GxE when more than one trait is measured in two 280 environments. In the simulation, rg HI was 0.16-0.56, which might explain the high relative genetic 281 gains of all VIO and VGO scenarios. 282

It was expected that both the increases of relative genetic gain and decreases in absolute genetic gain would correspond to increases of r_{g_HI} . However, an increasing tendency of absolute genetic gain did not correspond to the increase of r_{g_HI} (Table 4). Possible explanations may include the two-stage selection for BW in males and that EP is a sex limited trait (50% fewer records for EP than for BW), thus a change of r_{g_EP} has a different impact on absolute genetic gain than a change of r_{g_BW} .

289 Group versus individual observation

VGO breeding scheme was similar to VIO, except that recordings of village performance were in groups of 10 paternal-sibs. Our findings showed that VGO had lower *RG* than VIO, which is due to a lower accuracy of prediction of breeding values using group recording. Pooling birds in groups reduced the amount of information that was provided for each individual. Nonetheless, VGO had substantially increased genetic gains compared to the reference breeding scheme and reduction of the absolute genetic gain in comparison to the corresponding VIO scenario was at most 6% (Table 4.). Other studies have analysed pooled data, in which pooled observations were groups of random animals, full-sibs, half-sibs and descendants of maternal grand sire (Biscarini et al., 2008; Nurgiartiningsih et al., 2004; Olson et al., 2006; Peeters et al., 2013). From these studies, it can be concluded that estimation of breeding values from pooled data is theoretically and practically feasible for selection, particularly when the pooled observations are groups of closely related animals.

Biscarini et al. (2008) illustrated that correlations between EBV based on individual observation 303 and the pooled observation of 4 half-sib animals were 0.703-0.748 for EBV of the own animals, 304 305 0.814-0.891 for EBV of their sires with more than 10 offspring and 0.847-0.880 for EBV of their 306 dams with more than 4 offspring. Nurgiartiningsih et al. (2004) also demonstrated high correlations between EBV based on individual and group observations which were, on average, 0.844 for EBV 307 of the animals and 0.943 for EBV of their sires. Olson et al. (2006) studied accuracies of predicting 308 309 breeding values from individual and group observation using simulation. They found that in the 310 absence of pen effects, accuracies of EBV of animals themselves or their sires would be improved when animals allocated in a group were more related and when size of each group was smaller 311 given the same total number of animals. 312

In our study, to estimate EBV, selection candidates of VGO scenarios could have indirect information from individual observations of the correlated traits (BW_s and EP_s) of their own and parents' performance and direct information from pooled observations of the desired village traits (BW_v and EP_v) of their sibs. The pooled observations were groups of birds that had half-sib and full-sib relationship to the selection candidates. By averaging observations of the sib mixture, effects of dams mated to a sire on their offspring cannot be distinguished. The pooled observations can be only approximated as average of half-sibs. Meanwhile, effects of dams, full-sib and halfsib relationships can be taken into account in predicting EBV of selection candidates in VIO
scenarios, which resulted in a higher genetic gain in VIO than in VGO scenarios.

Nonetheless, the differences between accuracy of selection of VIO and VGO were not substantial. With r_{g_BW} of 0.5 and r_{g_EP} of 0.1, accuracy of EBVs of selection candidates was 0.863, 0.892 and 0.917 for VGO with 600, 1200 and 1800 village tested birds, respectively, while the accuracy of EBVs was 0.868, 0.908 and 0.925 for VIO with 600, 1200 and 1800 village tested birds, respectively. With r_{g_BW} of 0.9 and r_{g_EP} of 0.5, the accuracy of EBVs was 0.917, 0.927 and 0.935 for VGO with 600, 1200 and 1800 village tested birds, respectively; and 0.920, 0.934 and 0.940 for VGO with 600, 1200 and 1800 village tested birds, respectively.

329 *Methodology*

In our study, high relative genetic gains were achieved for VIO and VGO scenarios, and none of their replicates had negative relative genetic gains. This is due to 3 important assumptions including strong GxE, unchanged number of selection candidates and no common maternal effects.

GxE was modelled for BW at rg BW of 0.5-0.9 and EP at rg EP of 0.1-0.5, which represents quite 333 334 strong interactions. Conventional breeding programs are usually carried out under conditions most favourable for the expression of genotypes. One of the important reasons for this is that GxE is 335 often small, especially for commercial breeds where production animals are reared in enclosed, 336 highly controlled conditions, similar to the station situation. However, the differences between 337 338 village and breeding station are likely to be more substantial. Therefore, if birds are selected under station conditions of sufficient and balanced diets, absence of infectious diseases and minimum of 339 340 stress, strong GxE will be expected.

Number of selection candidates was assumed to be unchanged, even for the reference breeding 341 scheme in which tested birds were assumed to have station observations. This assumption is not 342 reasonable in practice, but it was included to quantify benefit of village observations. In theory, as 343 long as genetic correlation between traits measured in station and village environments is less than 344 1, village observations would provide additional genetic gains for VIO and VGO. Alternatively, if 345 346 the combined number of birds for selection and village testing was constant, the use of birds for village testing in VIO schemes would not be beneficial for genetic gain with rg HI above 0.8 due 347 to reduced selection intensity (Chu et al., 2018; Mulder & Bijma, 2005; Robertson, 1959). The use 348 of birds for village testing in VGO schemes would only become beneficial when rg HI was even 349 lower than the $r_{g HI}$ of VIO schemes. 350

Common maternal effects were not included in our simulation. The inclusion of the common 351 maternal effects would have relatively slight effects on genetic gain of VIO if birds from different 352 353 families are randomly distributed to smallholders. In contrast, it would reduce considerable genetic gains of VGO as members of the group with pooled observations were paternal-sibs. However, the 354 common maternal effects are negligible for the traits of selection in breeding program for village 355 poultry. The traits for selection are often at relatively old age, for example, BW at 16 or 20 weeks 356 of age and EP at 40 or 44 weeks of age. At these ages, common maternal effects for BW and EP 357 would be insignificant. Common maternal effects for BW reduce as birds age (Begli et al., 2016; 358 Dana et al., 2011; Prado-Gonzalez et al., 2003). The dam effects of BW disappeared at 8 weeks of 359 age (Prado-Gonzalez et al., 2003) and 12 weeks of age (Dana et al., 2011). Common dam effects 360 are usually not included in the model for EP traits as they are expressed late in bird life. 361

362 *Application of breeding schemes for village poultry production*

363 Poultry breeding for village production by poor and nutritionally insecure people in the rural and peri-urban regions of the Sub-Saharan Africa must accept the reality that people prefer dual-364 purpose chicken in a scavenging or semi-scavenging system (Dana et al., 2010). High investment 365 for commercial housing shed, supplementation of feed and expanded flock size can lead to 366 unsteady net returns. Such a risky investment was one of the main reasons that village farmers 367 were reluctant to spend on the inputs (Wondmeneh, 2015). It is shown that the use of the 368 scavenging production system for smallholders brings in higher economic returns than the use of 369 the semi-intensive or intensive system (FAO, 2010). Therefore, to improve the livelihood of the 370 targeted people, a proper breeding program for village poultry production is required. 371

Using village observations, breeding schemes VIO and VGO would be appropriate for improving 372 373 genetic gain of a breeding program and possibly maintaining adaptability traits which are major advantages of indigenous chicken in village production. However, implementation of VIO requires 374 375 individual records of pedigree and measurement of phenotypes under village conditions. Routine 376 recording phenotypes for individual birds is most likely not possible in village production systems. Measurement of individual phenotypes by smallholder farmers often has low accuracy 377 378 (Lwelamira, 2012). Implementation of VGO is simpler in practice compared to VIO. Although lower genetic gain is predicted for VGO, the increased accuracy of data recording in VGO may 379 make up for this. Group recording in the VGO breeding scheme reduces the complexity of tracing 380 and recording process. Therefore, the recommended breeding scheme for village poultry 381 production is VGO. Testing 600 birds in the village environment results in significant genetic gain 382 383 for the program, compared to testing them on station.

384 Conclusions

385 Village observations significantly increased genetic gain compared to station observations. The improvement was only slightly larger with individual observations (VIO) than with group 386 observations (VGO). Higher genetic correlations between traits measured in station and village 387 environments led to higher genetic gain, but lower relative genetic gain in VIO and VGO scenarios. 388 In assessing relative genetic gains from village observations for a breeding program in presence of 389 GxE, rg HI, the genetic correlation between station and village breeding objective, should be used 390 to model GxE as it explained better the magnitude of GxE than rg BW or rg EP alone. Breeding 391 schemes that use village group recording are applicable for breeding indigenous dual-purpose 392 393 poultry where a strong GxE is expected.

394 Acknowledgements

Thinh T. Chu benefits from a joint grant from the European Commission within the framework of the Erasmus-Mundus joint doctorate "EGS-ABG" and Graduate School of Science and Technology, Aarhus University.

398 Conflict of interests

399 The authors declare that they have no conflict of interests.

400 Authors' contributions

- 401 TTC, EN and PB designed and coordinated the study. TTC, PB and JB designed breeding schemes.
- 402 TTC and PB conducted all simulations. TTC drafted the manuscript, and all authors read and 403 approved the final manuscript.

404 **References**:

405	Begli, H. E., Torshizi, R. V., Masoudi, A. A., Ehsani, A., & Jensen, J. (2016). Longitudinal analysis
406	of body weight, feed intake and residual feed intake in F2 chickens. Livestock Science, 184,
407	28-34.

- Bekele, F., Gjoen, H. M., Kathle, J., Adnoy, T., & Abebe, G. (2009). Genotype x environment
 interaction in two breeds of chickens kept under two management systems in Southern
 Ethiopia. *Tropical Animal Health and Production*, 41, 1101-1114.
- Bijma, P., & Arendonk, J. A. M. (1998). Maximizing genetic gain for the sire line of a
 crossbreeding scheme utilizing both purebred and crossbred information. *Animal Science*,
 66, 529-542.
- Biscarini, F., Bovenhuis, H., & van Arendonk, J. A. (2008). Estimation of variance components
 and prediction of breeding values using pooled data. *Journal of animal science*, 86, 28452852.
- Cahaner, A., Deeb, N., & Gutman, M. (1993). Effect of the plumage-reducing naked-neck (Na)
 gene on the performance of fast-growing broilers at normal and high ambient temperatures. *Poultry science*, 72.
- Chen, C.F., Gourichon, D., Huang, N.Z., Lee, Y.P., Bordas, A., & Tixier-Boichard, M. (2009).
 Performance comparison of dwarf laying hens segregating for the naked neck gene in
 temperate and subtropical environments. *Genetics Selection Evolution*, 41, 1-11.
- Chu, T.T., Alemu, S.W., Norberg, E., Sørensen, A.C., Henshall, J., Hawken, R., & Jensen, J.
 (2018) Benefits of testing birds in both bio-secure and production environments in genomic
 selection breeding programs for commercial broiler chicken, *Genetics Selection Evolution*,
 vol. 50, no. 1, pp. 52.

427	Dana, N., van der Waaij, L. H., Dessie, T., & van Arendonk, J. A. M. (2010). Production objectives
428	and trait preferences of village poultry producers of Ethiopia: implications for designing
429	breeding schemes utilizing indigenous chicken genetic resources. Tropical Animal Health
430	and Production, 42, 1519-1529.

- Dana, N., vander Waaij, E. H., & van Arendonk, J. A. M. (2011). Genetic and phenotypic
 parameter estimates for body weights and egg production in Horro chicken of Ethiopia. *Tropical Animal Health and Production*, 43, 21-28.
- 434 Dessie, T., Alemu, Y., & Peters, K. (2000). Indigenous chickens in Ethiopia: genetic potential and
 435 attempts at improvement. *World's Poultry Science Journal*, 56, 45-54.
- FAO. (2010). Chicken genetic resources used in smallholder production systems and opportunities
 for their development. In P. Sørensen (Ed.), *FAO Smallholder Poultry Production Paper No. 5.* Rome.
- Horst, P. (1985). *Effects of genotype × environment interactions on efficiency of improvement of egg production.* Paper presented at the Proceedings of the 18th Poultry Science Symposium
- and of the 25th British Poultry Breeders Round Table.
- 442 Kapell, D. N., Hill, W. G., Neeteson, A. M., McAdam, J., Koerhuis, A. N., & Avendano, S. (2012).
- Genetic parameters of foot-pad dermatitis and body weight in purebred broiler lines in 2
 contrasting environments. *Poultry science*, 91, 565-574.
- 445 Lwelamira, J. (2012). Genotype-Environment (GXE) Interaction for Body Weights for Kuchi
 446 Chicken Ecotype of Tanzania Reared Under Intensive and Extensive Management. *Global*
- 447 *Journal of Medical Research*, 12, 51-57.

448	Lwelamira, J., Kifaro, G. C., & Gwakisa, P. S. (2009). Genetic parameters for body weights, egg
449	traits and antibody response against Newcastle Disease Virus (NDV) vaccine among two
450	Tanzania chicken ecotypes. Tropical Animal Health and Production, 41, 51-59.

- 451 Mathur, P. K. (2003). Genotype-environment interactions: problems associated with selection for
- 452 increased production. In W. M. Muir & S. E. Aggrey (Eds.), *Poultry Genetics, Breeding*453 *and Biotechnology* (pp. 83-99): CAB International.
- Mathur, P. K., & Horst, P. (1994). Methods for evaluating genotype × environment interactions
 illustrated with laying hens. *Journal of Animal Breeding and Genetics*, 111.
- Mulder, H., & Bijma, P. (2005). Effects of genotype× environment interaction on genetic gain in
 breeding programs. *Journal of animal science*, 83, 49-61.
- N'Dri, A. L., Sellier, N., Tixier-Boichard, M., Beaumont, C., & Mignon-Grasteau, S. (2007).
 Genotype by environment interactions in relation to growth traits in slow growing
 chickens. *Genetics Selection Evolution*, 39, 513-528.
- Niknafs, S., Nejati-Javaremi, A., Mehrabani-Yeganeh, H., & Fatemi, S. A. (2012). Estimation of
 genetic parameters for body weight and egg production traits in Mazandaran native
 chicken. *Tropical Animal Health and Production*, 44, 1437-1443.
- 464 Nirea, K. G., Sonesson, A. K., Woolliams, J. A., & Meuwissen, T. H. (2012). Effect of non-random
 465 mating on genomic and BLUP selection schemes. *Genetics Selection Evolution*, 44, 11.
- 466 Nurgiartiningsih, V. M., Mielenz, N., Preisinger, R., Schmutz, M., & Schueler, L. (2004).
- 467 Estimation of genetic parameters based on individual and group mean records in laying
 468 hens. *British Poultry Science*, 45, 604-610.

469	Okeno, T. O., Magothe, T. M., Kahi, A. K., & Peters, K. J. (2012). Application of risk-rated profit
470	model functions in estimation of economic values for indigenous chicken breeding.
471	Tropical Animal Health and Production, 44, 1279-1287.

- Okeno, T. O., Magothe, T. M., Kahi, A. K., & Peters, K. J. (2013). Breeding objectives for
 indigenous chicken: model development and application to different production systems. *Tropical Animal Health and Production*, 45, 193-203.
- Oleforuh-Okoleh, V. U. (2011). Estimation of genetic parameters and selection for egg production
 traits in a Nigerian local chicken ecotype. *ARPN Journal of Agricultural and Biological Science*, 6, 12.
- Olson, K. M., Garrick, D. J., & Enns, R. M. (2006). Predicting breeding values and accuracies
 from group in comparison to individual observations. *Journal of animal science*, 84, 8892.
- Pedersen, L., Sørensen, A., Henryon, M., Ansari-Mahyari, S., & Berg, P. (2009). ADAM: A
 computer program to simulate selective breeding schemes for animals. *Livestock Science*,
 121, 343-344.
- Peeters, K., Ellen, E. D., & Bijma, P. (2013). Using pooled data to estimate variance components
 and breeding values for traits affected by social interactions. *Genetics Selection Evolution*,
 486 45, 27.
- 487 Prado-Gonzalez, E., Ramirez-Avila, L., & Segura-Correa, J. (2003). Genetic parameters for body
 488 weights of Creole chickens from Southeastern Mexico using an animal model. *Livestock* 489 *Research for Rural Development*, 15, 27-31.
- Robertson, A. (1959). The sampling variance of the genetic correlation coefficient. *Biometrics*, 15,
 469-485.

492	Wolc, A., Arango, J., Settar, P., Fulton, J., O'Sullivan, N., Preisinger, R., Dekkers, J. (2013).
493	Analysis of egg production in layer chickens using a random regression model with
494	genomic relationships. Poultry science, 92, 1486-1491.

- Wondmeneh, E. (2015). *Genetic improvement in indigenous chicken of Ethiopia*. (PhD Thesis),
 Wageningen University, Netherlands.
- Wondmeneh, E., van Der Waaij, E., Dessie, T., Udo, H., & Van Arendonk, J. (2015). Adoption of
 exotic chicken breeds by rural poultry keepers in Ethiopia. *Acta Agriculturae Scandinavica, Section A—Animal Science*, 64, 210-216.

500 Tables

Table 1: Genetic parameters assumed for simulating body weight (BW) and egg number (EP) in

station (s) and village (v) environments: phenotypic variance, heritability (along the diagonal),

503 genetic correlations (above diagonal), and environmental correlations (below diagonal)

	σ_P^2	BW_s	EPs	BW_{v}	EPv
BW_{s}	291751	0.41	-0.12	r _{g_BW}	$-0.12 (r_{g_BW} + r_{g_EP})/2$
EPs	130.65	0.02	0.28	-0.12 $(r_{g_BW} + r_{g_EP})/2$	r_{g_EP}
BW_{v}	569610	0	0	0.21	-0.12
EP_{v}	261.29	0	0	0.02	0.14

504 Note: r_{g_BW} and r_{g_EP} , genetic correlation between traits of station and village environments, are variable factors.

Table 2: Breeding schemes and parameters of genetic correlations between station and village
traits

Alternative breeding scheme

Variables	Reference	Individual	Group observation	
	breeding	observation (VIO)	(VGO)	
	scheme			
Number of tested birds	600, 1200, 1800	600, 1200, 1800	600, 1200, 1800	
Type of observations on tested birds	Station	Village	Village	
Recording method	Individual	Individual	Group	
Genetic correlation between station	050709	050709	050709	
and village bodyweight (r_{g_BW})	,,	,,,	,,	
Genetic correlation between station	010305	010305	010305	
and village egg production (r_{g_EP})	0.1, 0.2, 0.2	0.1, 0.5, 0.5	0.1, 0.5, 0.5	

Table 3: Mean of relative genetic gain (*RG* %) of breeding scenarios with different genetic correlations between station and village bodyweight traits (r_{g_BW}) and egg production traits (r_{g_EP}) corresponding to genetic correlations between breeding goal H and index I (r_{g_HI}). S.E.M. was 1%.

r_{g_BW}	rg_ep	$r_{g_{HI}}$	Mean of RG
0.5	0.1	0.16	222
0.7	0.1	0.20	170
0.9	0.1	0.23	138
0.5	0.3	0.33	91
0.7	0.3	0.36	78
0.9	0.3	0.40	67
0.5	0.5	0.50	47
0.7	0.5	0.53	37

0.9	0.5	0.56	31	

Table 4: Mean of genetic gains per generation (ΔG) (± SD) of breeding scenarios with different genetic correlations between station and village bodyweight traits (r_{g_BW}) and egg production traits (r_{g_EP}) using either individual (VIO) or group recording (VGO) of village observations of 600, 1200 and 1800 birds.

r _a pw	DW Î a ED		ra ED ra UI		600		1200		1800	
rg_bw	ig_pr	ig_er	rg_III	VIO	VGO	VIO	VGO	VIO	VGO	
0.5	0.1	0.16	$20.4{\scriptstyle~\pm~2.4}$	$20.1{\scriptstyle\pm2.7}$	23.7 ± 2.1	22.2 ± 2.5	25.9 ± 1.8	$24.5{\scriptstyle~\pm~2.0}$		
0.7	0.1	0.20	21.9 ± 2.1	21.2 ± 2.2	25.1 ± 2.3	23.7 ± 2.1	26.6 ± 2.0	25.2 ± 2.2		
0.9	0.1	0.23	23.7 ± 2.3	$23.0{\scriptstyle~\pm~2.1}$	$26.5{\scriptstyle~\pm~2.1}$	25.3 ± 2.0	$28.0{\scriptstyle~\pm~2.0}$	26.9 ± 2.3		
0.5	0.3	0.33	21.1 ± 2.2	$20.5{\scriptstyle~\pm~2.2}$	$24.0{\scriptstyle~\pm~2.1}$	23.4 ± 2.3	25.9 ± 2.2	24.7 ± 2.3		
0.7	0.3	0.36	22.8 ± 2.3	21.5 ± 2.3	25.3 ± 2.2	24.6 ± 2.2	27.3 ± 2.1	25.9 ± 2.0		
0.9	0.3	0.40	24.7 ± 2.3	$24.0{\scriptstyle~\pm~2.3}$	26.6 ± 2.0	26.2 ± 1.6	28.2 ± 1.8	$27.1{\scriptstyle~\pm 1.9}$		
0.5	0.5	0.50	$23.0_{\pm2.1}$	$22.0{\scriptstyle~\pm~2.4}$	25.7 ± 2.2	24.7 ± 2.5	27.4 ± 2.1	$26.0{\scriptstyle~\pm~2.1}$		
0.7	0.5	0.53	$24.0{\scriptstyle~\pm~2.4}$	23.4 ± 2.3	26.7 ± 2.2	25.7 ± 2.1	28.1 ± 2.5	26.6 ± 2.1		
0.9	0.5	0.56	25.8 ± 2.1	$25.1{\scriptstyle~\pm 2.2}$	27.7 ± 2.0	$26.5{\scriptstyle~\pm~1.7}$	28.9 ± 1.9	$28.0{\scriptstyle~\pm~2.1}$		

514

515 Figure



516

Figure 1: Breeding cycle of a generation. ¹ Sex ratio of $1 \checkmark : 1 \heartsuit$; ² Bodyweight observed in both

518 \bigcirc and \bigcirc ; ³ Egg production observed in \bigcirc ; \rightarrow Birds reproduced/ selected, --- Observations

519 realized, ---- Information for selection



Figure 2: Means of relative genetic gains (%) (\pm SEM of 2%) of breeding scenarios with different genetic correlations between station and village bodyweight traits (r_{g_BW}) and egg production traits (r_{g_EP}) using either individual (VIO) or group recording (VGO) of 600, 1200 and 1800 village tested birds.