

Transmission dynamics of lumpy skin disease in Ethiopia

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24	Transmission dynamics of lumpy skin disease in Ethiopia
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48	Running title: Lumpy skin disease transmission dynamics

49 Summary

Lumpy skin disease (LSD) is a severe disease of cattle caused by a Capripoxvirus and often 50 caused epidemics in Ethiopia and many other countries. This study was undertaken to 51 quantify the transmission between animals and to estimate the infection reproduction ratio in a 52 predominantly mixed crop-livestock system and in intensive commercial herd types. The 53 transmission parameters were based on a SIR epidemic model with environmental 54 transmission and estimated using generalized linear models. The transmission parameters 55 were estimated using a survival rate of infectious virus in the environment equal to 0.325 per 56 day, a value based on the best fitting statistical model. The transmission rate parameter 57 between animals was 0.072 (95% CI: 0.068-0.076) per day in the crop-livestock production 58 system, whereas this transmission rate in intensive production system was 0.076 (95% CI: 59 0.068-0.085) per day. The reproduction ratio (R) of LSD between animals in the crop-60 livestock production system was 1.07, whereas it was 1.09 between animals in the intensive 61 production system. The calculated R provides a baseline against which various control options 62 can be assessed for efficacy. 63 Key words: Cattle, Ethiopia, LSD, Transmission, Reproduction ratio 64 65 66 67 68

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73 **1. Introduction**

Lumpy skin disease (LSD) is a severe viral disease of cattle, which often occurs as regional epidemics within a larger area in which it is endemic. It is caused by *Lumpy skin disease virus* (LSDV) which is of the genus *Capripoxvirus* of family *Poxvirida*e. LSDV is one of the most important animal poxviruses because of the serious economic consequences in cattle [1, 2]. The disease is characterized by lachrymation, fever, nodular lesions on the skin and mucosal surfaces, lymph node enlargement, inflammatory and oedematous swelling of the legs and lameness [1, 3].

81

The disease was reported for the first time in Zambia in 1929 and was confined to Africa until an outbreak occurred in Israel in 1989 [1]. However, currently, the disease is found in most African and Middle East countries and recently it has spread to eastern and south eastern European countries. LSDV is clearly on the move in expanding its territory and increasingly becoming a risk for other Asian and European countries [4].

87

Though the mechanism of LSDV transmission has not yet been clearly established, it is 88 hypothesized that the main mode of transmission of LSDV is via blood feeding arthropods 89 [5]. Experimentally, female Aedes aegypti mosquitoes have been shown to transmit LSDV 90 mechanically from infected to susceptible cattle [6]. The potential role of ixodid ticks in 91 transmission of LSDV has also been demonstrated in transmission studies including 92 mechanical transmission between cattle for Amblyomma hebraeum and Rhipicephalus 93 appendiculatus, trans-stadial transmission for A. hebraeum, and transovarial transmission for 94 Rhipicephalus (Boophilus) decoloratus [7-11]. Transmission of LSDV between infected and 95 susceptible animals by direct contact is considered to be inefficient [5, 12]. 96

97

98 Data from infectious disease outbreaks are usually incomplete and highly dependent.

Incomplete because the infection process is only partially observable, i.e. not all cases may be 99 included due to under-reporting or because of asymptomatic cases, the number of susceptible 100 animals may not be known exactly, individuals who enter or leave the study population may 101 not be recorded accurately, there may be misdiagnosis of cases and flaws in data collection. 102 Data such as daily or weekly case numbers are obviously dependent [13, 14]. However, 103 transmission under field conditions can be estimated from the number of infections that 104 occurred during the study period or at certain intervals by mathematical modelling using 105 exactly that dependence [15, 16]. 106

107

One parameter often used to characterize transmission is the basic reproduction ratio (R_0) with 108 the effective reproduction ratio (R_e) being the parameter for the transmission after 109 intervention. R₀ is defined as the average number of secondary infections caused by one 110 typical infectious individual in a fully susceptible population during its entire infectious 111 period [17], whereas Re reflects the transmission parameter in a partially susceptible 112 (previously exposed or vaccinated) population [18]. The reproduction ratio (R) is frequently 113 used to describe the behaviour of transmission just after introduction of a disease. Whether an 114 115 outbreak spreads or dies out depends on whether the reproduction ratio is greater than, or less than, 1 respectively. If R exceeds 1, a typical (i.e. average) infected animal infects on average 116 more than one susceptible animal, and thus it may cause a major outbreak, while if R is 117 smaller than 1 the disease will die out or it will at most produce a minor outbreak [16, 19]. 118 119

Despite a large number of LSD outbreaks in many African and Middle East countries, its dynamics are not well studied. Only one study, undertaken by Magori-Cohen et al. [12] in a dairy herd of Israel, reports an estimate for the reproduction ratio of LSDV ($R_0 = 15.7$).

Therefore, the current study was undertaken with the objectives to better understand the
 LSDV outbreak dynamics and to quantify the transmission rate parameter and the
 reproduction ratio between animals.

126

127 **2. Materials and Methods**

128

129 **2.1. Study area, farms and animals contact patterns**

130

The study was carried out from 28 April 2014 to 1 February 2015 in the central and north-131 western parts of Ethiopia. In the north-western part, it involves the cattle population in Mota 132 town and parts of the surrounding five Kebeles (Kebele is the smallest administrative unit in 133 Ethiopia covering an approximate area of 53 km²) in Hulet Ejju Enessie district, and 134 Debremarkos University dairy farm in Gozamn district. In the central part, the following 135 herds were enrolled: Selale Dairy Development Private Limited Company (Selale Dairy Dev't 136 PLC) in Wuchale district, Aser Dev't PLC in Sululta district, Ambo University dairy farm in 137 Ambo district, Holeta agricultural research centre farm (Holeta A.R.C) and Holeta special 138 cattle breeding centre (Holeta S.C.B.C) in Welmera district, Selam children village dairy farm 139 in Addis Ababa and Jenesis dairy farm in Ada'a district (Figure 1). Mota area (Mota town and 140 parts of the surrounding five Kebeles) covers an area of about five km radius. The production 141 system in the Mota area is mainly mixed crop-livestock while the other herds were 142 commercial dairy herds. Most of the animals in the mixed crop-livestock type of herds were 143 of local Zebu breed whilst the intensive herds consisted of Holstein-Zebu cross. Farms were 144 categorized into small (<10 cattle), medium (10-50 cattle), large (51-300 cattle), very large 145 (301-700 cattle) and extra-large (>700 cattle) based on the number of cattle they comprised. 146

The cattle contact network depends on a number of factors including housing system, size and 147 nature of grazing lands, water points, cattle density, and frequency and duration of contacts. 148 This study was undertaken at the family herd (group of animals owned by a family for 149 subsistence) and commercial farm (group of animals owned by a private or public 150 organization for commercial purpose) levels. All smallholder herds enrolled in the study were 151 in the Mota area, but the intensive commercial farms were located in different areas. Since the 152 smallholder herds in the subsistence crop-livestock system (Mota area) are managed 153 extensively, they regularly mixed at shared pastures and watering points so that they had to be 154 considered as one epidemiological unit. Animals in the intensive commercial farms, however, 155 did not have direct contact with animals in other farms in their surroundings and most of them 156 were located in districts far apart from each other. 157

158

159 **2.2. Period of the epidemic**

160

To assess the association between LSD epidemics and the season of the outbreak (which has a 161 strong relation with arthropod dynamics), the outbreak duration was categorized into three 162 periods, Belg (period 1), Kiremt (period 2) and Bega (period 3) following the meteorological 163 seasons of Ethiopia. Belg is a short rainy period from February to May over much of the 164 Belg-growing areas. However, over the north-western parts of the country (where Mota area 165 is located) this season is predominantly dry except for the month of May. Kiremt is the period 166 from June to September; it is the main rainy season in which the major food crops of the 167 country are produced. The magnitude of rainfall during Kiremt is higher as compared to the 168 other seasons for many parts of the country. Bega is the period from October to January. It is 169 normally a dry season characterized by cool nights and hot days over various parts of the 170 country [20]. 171

172

173 **2.3. Infection status of animals**

174

Herds were visited every week to check whether or not animals showing symptoms of LSD 175 were present. If so, the infection chain within the herd was monitored by visiting the affected 176 herd twice a week throughout the study period and the LSD status (susceptible, infected or 177 recovered) of all animals was determined. At the start of the study all cattle were assumed to be 178 susceptible. The start of the infectious period was considered to be the day following that on 179 which an animal was first reported with clinical signs of LSD. Infected animals were assumed 180 to stay infectious on average for 10 days taking the duration of viraemia as a proxy for period 181 of infectivity [5, 21, 22]. An infected animal becomes most infective during the viraemic phase 182 of the disease because the amount of virus in various body tissues and secretions and excretions 183 of the animal become the highest in this phase [22]. Animals that died before the infectious 184 period was completed were considered infectious for the days they lived after being considered 185 infectious. 186

187

The contribution of environment (E) to the transmission of LSDV was established by
determining a per day survival rate of LSD virus shed into the environment by infected
animals. This was done by fitting a GLM model to the collected data by varying the survival
rate from 0.1 to 0.9 and selecting the best fitting model with the lowest AIC value.

192

Nodular samples were collected from few affected cattle in each herd to confirm the outbreak
by using conventional and snapback real-time PCR (polymerase chain reaction) techniques

following the procedure described by Gelaye et al. [23].

196

197 **2.4. Estimation of the transmission parameters**

The transmission parameters were estimated based on a SIR epidemic model in which 198 individuals are either susceptible (S), infected and infectious (I) or recovered and immune or 199 dead (R). During the study, the numbers of I and S observed in each herd were recorded at the 200 start of each observation interval. Animals were registered as a new case (C) on the date they 201 were reported with LSD and as infectious (I) on the next day. Transmission of LSDV between 202 animals has been estimated from the relationship between the number of infectious animals at 203 204 the start of the time interval and the number of newly infected animals at the end of the time interval. Every new infection is related to the number of animals that were infectious at the 205 time of infection. 206

207

The transmission parameters were estimated by a generalized linear model (GLM) [24-27]. 208 The transmission dynamics of LSD between individuals are described by the change in the 209 number of susceptible (S), infectious (I), and recovered (R) animals. Susceptible cattle 210 become infected with a rate of $\beta \cdot S_t \cdot (I_t + E_t) / N_t$. Here, β is the transmission rate which can be 211 interpreted as the average number of new infections caused by a typical infectious animal in a 212 fully susceptible population per unit of time, St is the number of susceptible animals, It the 213 number of infectious animals, Et contribution of the environment to the transmission, and Nt 214 is the total number of animals at time t, and they are assessed at the start of each observation 215 period. The number of infectious contacts encountered by one individual in a period of length 216 Δt follows a Poisson distribution with parameter ($\beta \cdot (I_t + E_t) / N_t \cdot \Delta t$). Hence, the probability 217 of a susceptible animal escaping infection, during a period Δt is $e^{-\beta \cdot \Delta t \cdot (I_t + E_t)/N_t}$, and thus the 218 probability to become infected is $1-e^{-\beta \cdot \Delta t \cdot (I_t + E_t)/N_t}$. This implies that the number of new cases 219 (C) in a period Δt follows a binomial distribution. Consequently, the relation between the 220 expected number of cases per unit of time E(C), and I_t , E_t , N_t , β , and S_t can be formulated as 221

222	$E(C_t)=S_t \cdot (1-e^{-\beta \cdot \Delta t \cdot (I_t+E_t)/N_t})$. The transmission parameter β ($\beta = e^b$, where b is the regression
223	coefficient of the intercept of the model) was estimated using a GLM with a complementary-
224	log-log link function and $log\left(\Delta t \cdot \frac{I_t + E_t}{N_t}\right)$ as offset. Finally, we obtained R by multiplying β
225	with the average length of the infectious period [19, 24, 27] times a factor of $(1-E)^{-1}$ which
226	incorporates the environmental contribution.
227	
228	The Chi-square test was used to test the association of morbidity and mortality with
229	production systems and GLM to compare transmission rates between the three meteorological
230	periods, production systems and herd sizes.
231	
232	All analyses were carried out in Stata 14.
233	
234	3. Results
235	
236	3.1. Descriptive statistics
237	
238	During the study period, a total of 14,319 individual animals from 2,446 herds were followed
239	for LSD occurrence. 12,509 animals (in 2,438 herds) were kept in the crop-livestock system
240	and 1,810 animals (in 8 herds) in the intensive production system (Table 1).
241	
242	The number of animals and herds affected, morbidity and mortality due to LSD per
243	production system are indicated in Table 1. The morbidity was significantly higher in the
244	intensive (17.5%) compared to the crop-livestock (10.1%) system. The mortality was also
245	significantly higher in the intensive (4.0%) than in the crop-livestock (0.7%) system (Table
246	1).

247

In the Mota area, the LSD outbreak started at the end of April 2014 but in the other study farms the outbreak started later and continued until the first week of February 2015. The epidemic curve of the LSD outbreak in the Mota area is presented in Figure 2.

251

252 3.2. Transmission of LSD between animals

253

The contribution of the environment to the transmission (E) and the number of C, I and S 254 animals in the Mota area are listed for each day of the epidemic (Supplementary Table S1). 255 The transmission rate parameter between animals in the dominantly subsistent crop-livestock 256 production system was 0.072 (95% CI: 0.068-0.076) per day (Table 2) whereas in the 257 intensive production system it was 0.076 (0.068-0.085) per day (Table 3). The survival rate of 258 infectious LSD virus in the environment was estimated as 0.325 per day based on the best 259 fitting statistical model and this value was used to account for the indirect transmission 260 (excluding the immediate or direct transmission) of the virus. The average LSD infectious 261 periods for animals are indicated in Table 2 and 3 for both production systems. 262

263

Based on the survival rate of LSDV in the environment, the multiplication factor of R was 1.5. Then a reproduction ratio of 1.07 between animals was calculated in the crop-livestock production system in the Mota area (Table 2). R values between animals vary from 0.90 (Aser dairy farm) to 1.15 (Ambo university) in the eight intensive farms while the overall R value for intensive dairy farms was 1.09 (Table 3). Major outbreaks have been observed in Ambo University, Holeta S.C.B.C, Holeta A.R.C, Selale Dairy Dev't PLC, Selam children village dairy herds and Mota area (Table 3, Supplementary Table S2). Transmission parameter rates (β) between animals for subsistence crop-livestock production system in the Mota area showed significant differences between period two and three (P <0.05) (Table 2). However, the transmission rates did not significantly differ between production systems and herd sizes.

275

276 **4. Discussion**

277

The 10.1% and 17.5% animal level morbidity of LSD reported in the current study in the 278 subsistence crop-livestock production system and intensive system, respectively, are within 279 the range of what has been reported in previous works [1, 28]. Similarly, the mortality was 280 higher in the intensive production system than in the crop-livestock system. These significant 281 differences in morbidity and mortality between animals in the two systems might be explained 282 by the breed of cattle raised in the two systems. In the intensive system, Holstein-Friesian 283 local cross was the dominant breed which is more susceptible and more severely affected by 284 LSD than the local Zebu breed [1, 29], which is the breed commonly found in the crop-285 livestock production system. The other reason might be related to the way we calculated the 286 morbidity and mortality in both systems. In the crop-livestock system, all animals in the Mota 287 area whether or not they were within an infected herd or not, were included in the 288 denominator, whereas in the intensive system only the number of animals in infected herds 289 were in the denominator to calculate the morbidity and mortality. 290

291

The infectious period and survival of the virus in the environment are important parameters in estimating the reproduction ratio but these parameters were not reported in any of the previous studies. However, information about these parameters is essential for formulating

appropriate prevention and control strategies for LSD. In this study too we did not estimate 295 the infectious period of an infected animal and the survival rate of the virus in the 296 environment because the study set up did not allow us to do that; instead we parametrized the 297 infectious period from information obtained in the literature and the survival rate by searching 298 for the best fitting model. We set the infectious period to 10 days for an infected animal by 299 taking into account the duration of virus isolation in blood for 10-12 days [5, 21]. 300 Furthermore, there is no clear information when infected animals become infectious, which is 301 important to know for the quantification of transmission. Infectiousness may start before or 302 after the onset of clinical disease, but for this study we set the start of the infectious period as 303 24 h after the onset of the disease considering that LSDV isolation from blood and skin 304 samples were achieved in most of the cases after the affected animals showed fever [21]. 305 Regarding the survival rate of the virus in the environment, literature indicates that the virus 306 survives in air-dried hides for at least 18 days, in necrotic skin nodules for up to 33 days or 307 longer, and for up to 35 days in desiccated crust [30], but it is not clear whether the viruses 308 surviving in these foci contribute to the transmission of LSD. Taking this information into 309 consideration we fitted a model (by selecting the best fitting model) to our data and found a 310 survival rate of 0.325 per day, which was used in the offset to incorporate the contribution of 311 312 environment to the transmission of LSDV. The implication of this survival rate is that the infectivity is increased by almost 50%. 313

314

To our knowledge, this is the first field study in Ethiopia in which transmission rate parameters have been quantified. This knowledge is helpful to design sets of measures that efficiently eliminate the virus. In the study, LSDV transmission was modelled by considering it as direct transmission. It is widely believed that LSDV is transmitted from infected to susceptible hosts indirectly through mechanical arthropod vectors, though the importance of

the different types of arthropod vectors in the transmission of LSD virus in field conditions is not fully understood [5, 12]. If a blood feeding arthropod feeds briefly on viraemic cattle and is interrupted, a subsequent immediate feeding on a second animal could result in virus transmission. The virus does not replicate within the vector [31] which thus serves as a passive carrier to transmit the disease. The vector in this case serves only as a bridge for the transmission of LSDV from infected to susceptible cattle so that we did not incorporate the vectors in the transmission model.

327

During the outbreak, LSDV was transmitted between animals with a rate of 0.072 per day in the crop-livestock production system. The transmission chain from which specific infected cattle to which susceptible cattle was not clearly identified due to the free movement and mixing up of animals in the area and mechanical transmission of the disease by arthropods vectors. Hence, the transmission rate between animals was calculated by considering the cattle population in the area as one population.

334

In the Mota area, the transmission rate of LSD was also estimated for different time periods 335 and the results indicate a significant difference in daily transmission rates between periods. 336 The per day transmission rate between animals was higher at the beginning of the outbreak (in 337 period 1 and 2 compared to period 3). This was expected, because during these periods the 338 susceptible population was not yet depleted and no specific measures were taken to reduce 339 transmission. This result indicates that starting implementation of control measures at early 340 stage of the outbreak is necessary to halt the spread of the disease. We did not assess the 341 periodic variation of transmission rate in farms of intensive production system due to the fact 342 that the outbreaks in those farms were relatively short and it was not convenient to divide the 343 time into different periods as in most occasions the outbreak fell in one period. 344

In this study, we estimated an R value of 1.07 between animals in the crop-livestock area. The

R values within the intensive farms were also in the range of 0.90 to 1.15 with an overall
value of 1.09. These R values are low compared with the R value of 15.7 reported for indirect
transmission within a commercial dairy farm in Israel [12]. The difference might be explained
by the method how R is calculated, different study population, the environmental difference
and the production set up.

352

Knowledge of within herd transmission is necessary to assess the effectiveness of intervention 353 measures and to design effective monitoring programmes [32-34]. In this study, we estimated 354 that R was greater than 1 between animals in the dominantly crop-livestock system and in 355 some farms of the intensive production system. This sheds light on LSDV transmission and 356 further work should focus on the effect of control measures that add to bring R below the 357 threshold level. LSD control will be achieved if both reproduction ratios, among animals and 358 between herds are less than 1; and also if R among animals is greater than 1, but R, between 359 herds is below 1. Infections with low R values are less difficult to control than those with a 360 high R value [34]. Our estimates of R provides a baseline against which various control 361 options can be assessed for efficacy. In general, from this study it can be concluded that 362 transmission of LSDV between animals in Ethiopia is low. 363

364

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- 371 Conflict of interest
- 372 None
- 373
- 374 **5. References**
- Davies FG. Lumpy skin disease, an African capripox virus disease of cattle. *British Veterinary Journal* 1991; 147: 489-503.
- 2. **Carn.** Control of capripoxvirus infections. Vaccine 1993; **11**: 1275-1279.
- 378 3. **Tuppurainen ES, Oura CA.** Review: lumpy skin disease: an emerging threat to 379 Europe, the Middle East and Asia. *Transboundary and Emerging Diseases* 2012; **59**:
- 380 40-48.
- Tuppurainen ES, et al. Review: Capripoxvirus diseases: current status and
 opportunities for control. *Transboundary and Emerging Diseases*. *Published online:* 29 July 2015. doi:101111/tbed12444.
- 384 5. Carn VM, Kitching RP. An investigation of possible routes of transmission of lumpy
 385 skin disease virus (Neethling). *Epidemiology and Infection* 1995; 114: 219-226.
- Chihota CM, et al. Mechanical transmission of lumpy skin disease virus by *Aedes aegypti* (Diptera: *Culicidae*). *Epidemiology and Infection* 2001; 126: 317-321.
- Tuppurainen ES, et al. A potential role for ixodid (hard) tick vectors in the
 transmission of lumpy skin disease virus in cattle. *Transboundary and Emerging Diseases* 2011; 58: 93-104.
- Tuppurainen ES, et al. Mechanical transmission of lumpy skin disease virus by
 Rhipicephalus appendiculatus male ticks. *Epidemiology and Infection* 2013; 141: 425-430.
- 394 9. Tuppurainen ES, et al. Evidence of vertical transmission of lumpy skin disease virus
 395 in *Rhipicephalus decoloratus* ticks. *Ticks and Tick-borne Diseases* 2013; 4: 329-333.
- Lubinga JC, et al. Detection of lumpy skin disease virus in saliva of ticks fed on
 lumpy skin disease virus-infected cattle. *Experimental & Applied Acarology* 2013; 61:
 129-138.
- Lubinga JC, et al. Evidence of lumpy skin disease virus over-wintering by
 transstadial persistence in *Amblyomma hebraeum* and transovarial persistence in
 Rhipicephalus decoloratus ticks. *Experimental & Applied Acarology* 2014; 62: 77-90.
- Magori-Cohen R, et al. Mathematical modelling and evaluation of the different
 routes of transmission of lumpy skin disease virus. *Veterinary Research* 2012; 43. doi: 10.1186/1297-9716-43-1.
- Becker NG, Britton T. Statistical studies of infectious disease incidence. *Journal of the Royal Statistical Society: series B* 1999; 61: 287-307.
- 407 14. O'Neill PD. Introduction and snapshot review: Relating infectious disease
 408 transmission models to data. *Statistics in Medicine* 2010; 29: 2069-2077.
- Kroese AH, De Jong MCM. Design and analysis of transmission experiments. In:
 Menzies FD, Reid, S.W.J., ed. *Proceedings of the Society for Veterinary Epidemiology and Preventive Medicine*. Noordwijkerhout, The Netherlands, 2001, pp. xxi-xxxvii.
- Velthuis AGJ, et al. Quantification of transmission in one-to-one experiments. *Epidemiology and Infection* 2002; 128: 193-204.

414	17.	Diekmann O, Heesterbeek JAP, Metz JAJ. On the definition and the computation of
415		the basic reproduction ratio R0 in models for infectious diseases in heterogeneous
416		populations. Journal of Mathematical Biology 1990; 28: 365-382.
417	18.	Chowell G, Nishiura H. Transmission dynamics and control of Ebola virus disease
418		(EVD): a review. BMC Medicine 2014; 12 . doi: 10.1186/s12916-014-0196-0.
419	19.	van Roermund HJW, et al. No between-pen transmission of foot-and-mouth disease
420		virus in vaccinated pigs. Vaccine 2010; 28: 4452-4461.
421	20.	NMA. Annual climate bulletin. National meteorological agency (NMA), Addis Ababa
422		Ethiopia.
423		http://www.ethiomet.gov.et/bulletins/view_pdf/348/2013annualbulletin.pdf.
424		Accessed 30 June, 2016. 2013.
425	21.	Tuppurainen ESM, Venter EH, Coetzer JAW. The detection of lumpy skin disease
426		virus in samples of experimentally infected cattle using different diagnostic
427		techniques. Onderstepoort Journal of Veterinary Research 2005; 72: 153-164.
428	22.	Woods JA. Lumpy skin disease virus. In: Dinter, Z., Morein, B. eds. Virus infections
429		of ruminants. Amesterdam: Elsevier Science publishers B. V., 1990, pp. 53-67.
430	23.	Gelaye E, et al. Development of a cost-effective method for capripoxvirus genotyping
431		using snapback primer and dsDNA intercalating dye. PLoS ONE 2013; 8: e75971.
432	24.	Velthuis AGJ, et al. Design and analysis of an Actinobacillus pleuropneumoniae
433		transmission experiment. Preventive Veterinary Medicine 2003; 60: 53-68.
434	25.	Heffernan JM, Smith RJ, Wahl LM. Perspectives on the basic reproductive ratio.
435		Journal of the Royal Society Interface 2005; 2: 281-293.
436	26.	Chowell G, Nishiura H, Bettencourt LM. Comparative estimation of the
437		reproduction number for pandemic influenza from daily case notification data. Journal
438		of the Royal Society Interface 2007; 4: 155-166.
439	27.	Bravo de Rueda C, et al. Quantification of transmission of foot-and-mouth disease
440		virus caused by an environment contaminated with secretions and excretions from
441		infected calves. Veterinary Research 2015; 46. doi: 10.1186/s13567-015-0156-5.
442	28.	Woods JA. Lumpy skin disease- A review. Tropical Animal Health and Production
443	• •	1988; 20 : 11-17.
444	29.	Manual of diagnostic tests and vaccines for terrestrial animals, chapter 2.4.14,
445		Lumpy skin disease. OIE, Paris.
446		(http://web.oie.int/eng/normes/MMANUAL/A_Index.htm) (accessed 26 February
447	20	2016).
448	30.	Lumpy skin disease.
449		(http://www.oie.int/fileadmin/Home/eng/Animal_Health_in_the_World/docs/pdf/Dise
450	21	ase_cards/LUMPY_SKIN_DISEASE_FINAL.pdf). Accessed 10 October 2016.
451	31.	Goddard J . Injectious Diseases and Arthropoas, 2 th ean. USA: Humana Press, 2008,
452	20	pp. 19-28. Stansman A at al Transmission of classical awing force within hands during the
453	32.	Stegeman A, et al. I ransmission of classical swine fever virus within herds during the
454		1997-1998 epidemic in The Netherlands. Preventive veterinary Medicine 1999; 42:
455	22	201-210. Creat FAM at al Modelling the effect of surveillance programmes on spread of
456	<i>33</i> .	by the herpequipue 1 between corrified cettle bards. Veteringry Microbiology 2001;
457		70 , 103 208
400 450	34	17. 175-200. Hage II at al Transmission of hoving herpesuirus 1 within and hetwaan hords on an
407 160	54.	island with a BHV1 control programme. Enidemiology and Infection 2003: 130: 541
400		557

Area/Farm	Dominant	No. of	No. of	No. of	No. of	Morbidity	No.	Mortality
	Production	total	total	affected	infected	in %	died	in %
	system	herds	animals	herds	animals			
Mota area ^a	crop-livestock	2438	12509	841	1266	10.12	81	0.65
Ambo University	Intensive	1	86	1	24	27.91	6	6.98
Aser Dev't PLC	Intensive	1	50	1	5	10.00	0	0
Debremarkos University	Intensive	1	42	1	6	14.29	0	0
Holeta S.C.B.C	Intensive	1	429	1	88	20.51	19	4.43
Holeta A.R.C	Intensive	1	623	1	84	13.48	6	0.96
Jenesis dairy farm	Intensive	1	204	1	8	3.92	0	0
Selale Dairy Dev't PLC	Intensive	1	330	1	93	28.18	40	12.12
Selam Children village farm	Intensive	1	46	1	9	19.57	2	4.35
Intensive subtotal		8	1810	8	317	17.51	73	4.03

Table 1. LSD morbidity and mortality in subsistence crop-livestock and intensive commercial farms.

^a All herds and animals at risk considered

 χ^2 (1) = 87.89, P = 0.000 for differences in morbidity between animals in crop-livestock and intensive systems χ^2 (1) = 170.35, P = 0.000 for differences in mortality between animals in crop-livestock and intensive systems

Table 2. Transmission parameters of LSD virus between animals by meteorological period in dominantly crop-livestock system (Mota area),

Ethiopia, during the 2014 epidemic.

Transmission	Period	No. of	No. of	β (95% CI)	% CI) P-		R^a (95% CI)
		weeks	cases	per day	value	period in days	
Between animals	1 (18-22 ^b)	5	12	0.077 (0.043 - 0.139)	0.315	8.25	0.95 (0.53-1.72)
	2 (23-39)	17	887	0.080 (0.075 - 0.085)	0.000	9.03	1.08 (1.02-1.15)
	3 (40-47)	8	367	0.057 (0.051 - 0.063)	Ref	12.11	1.04 (0.93-1.14)
	Overall	30	1266	0.072 (0.068 - 0.076)		9.92	1.07 (1.01-1.13)

^aR is obtained after multiplying the product of β and infectious period by a factor of 1.5 which is a sum of the infectivity of the infected animal (1) and infectivity of the virus accumulated in the environment (0.5) at a particular date of the epidemic.

^bWeek number.

Table 3. Transmission parameters and reproduction ratios of LSD virus within eight intensive dairy herds and Mota area during the 2014/15

484 epidemic.

Area/Farm	Production	No. of	No. of	Outbreak	β (95% CI)	Average Inf.	R ^a
	system	animals	cases	dur. in weeks	per day	period in days	(95% CI)
Ambo University	Intensive	86	24	8	0.086 (0.057-0.130)	8.92	1.15 (0.76-1.74)
Aser Dev't PLC	Intensive	50	5	4	0.060 (0.022-0.159)	10	0.90 (0.33-2.39)
Debremarkos University	Intensive	42	6	4	0.064 (0.027-0.154)	10	0.96 (0.41-2.31)
Holeta S.C.B.C	Intensive	429	88	15	0.078 (0.063-0.096)	9.51	1.11 (0.90-1.37)
Holeta A.R.C	Intensive	623	84	17	0.071 (0.057-0.088)	9.96	1.06 (0.85-1.31)
Jenesis dairy farm	Intensive	204	8	8	0.061 (0.029-0.128)	10	0.92 (0.44-1.92)
Selale Dairy Dev't PLC	Intensive	330	93	21	0.082 (0.066-0.100)	9.24	1.14 (0.91-1.39)
Selam Children village farm	Intensive	46	9	7	0.068 (0.034-0.137)	10	1.02 (0.51-2.06)
Intensive total		1810	317	84	0.076 (0.068-0.085)	9.55	1.09 (0.97-1.22)
Mota area	Crop-livestock	12,509	1266	30	0.072 (0.068-0.076)	9.92	1.07 (1.01-1.13)
Overall		14,319	1583	114	0.073 (0.069-0.076)	9.84	1.08 (1.02-1.12)

^aR is obtained after multiplying the product of β and infectious period by a factor of 1.5, a sum of infectivity of the infected animal (1) and the infectivity of the virus accumulated in the environment (0.5) at a particular date of the epidemic.





504 Figure 2. Epidemic curve of lumpy skin disease in Mota area, Ethiopia, in 2014.