

Potential of Chitin Nanocrystals

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1 **ABSTRACT**

2

3 Chitin is a naturally occurring polysaccharide that can be found in shrimp shells, a waste product
4 from food industry. Due to its abundance and interesting properties, chitin is a hot topic within
5 both academia and industry, with research in many fields. The aim of this thesis was to explore
6 the use of chitin as a catalyst support material. We compared the shrimp shells (SHSH) to its
7 counterpart hydrolysed chitin nanocrystals (CHNC) as a support material for a copper catalyst. In
8 this work, the importance of the catalyst support preparation has been investigated and the effect
9 of drying on the properties of the copper catalyst. Drying at 60 °C resulted in homogeneous dried
10 impregnated samples. Moreover, various reduction conditions were also studied to obtain evenly
11 dispersed metallic copper for catalysis.

12

13 Different properties of the catalyst and its support were measured, including support material
14 pore size and copper particle size, thermal stability and crystallinity. Thermal gravimetric
15 analysis (TGA) showed that the degradation of SHSH began at a higher temperature than the
16 degradation of CHNC. Transmission electron microscopy (TEM) was applied to analyse the
17 morphology of CHNC and also the copper particle size distribution, which when embedded on a
18 CHNC support, ranged from 15 to 25 nm. X-ray diffraction (XRD) was used to show the
19 formation of copper oxides and metallic copper by various calcination/reduction conditions.
20 Moreover, physisorption revealed that the surface area of the CHNC support was greater than
21 SHSH, a beneficial property for a catalyst carrier. Considering all analysis performed, CHNC
22 had preferable catalyst carrier properties in comparison to SHSH, however, further work is
23 necessary to compare and evaluate copper catalysis on a CHNC support in contrast to another
24 support system.

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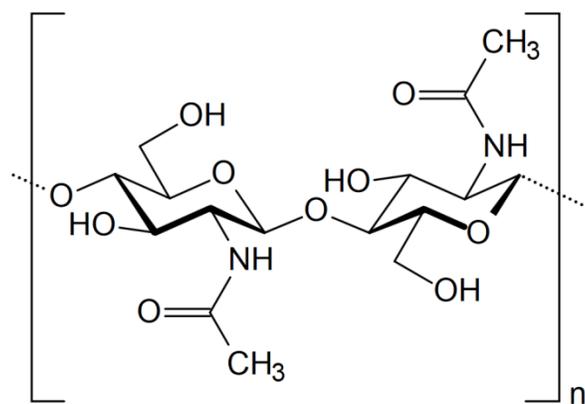
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62 **Abbreviations**

63	AFM	Atomic force microscopy
64	CHNC	Chitin nanocrystals
65	FTIR	Fourier transform infrared
66	HPLC	High performance liquid chromatography
67	IWI	Incipient wetness impregnation ²
68	SEM	Scanning electron microscopy
69	SHSH	Shrimp shells
70	TEM	Transmission electron microscopy
71	TGA	Thermogravimetric analysis
72	XRD	X-ray diffraction

73 1 INTRODUCTION

74 Chitin is the second most abundant polysaccharide found in nature, next to cellulose, acting as a
75 structural building block. Chitin can be found in a number of natural sources: algae, cell walls of
76 fungi, exoskeletons of crustaceans, insects, invertebrates, marine diatoms, molluscs and yeasts
77 (Synowiecki & Al-Khateeb, 2003). Chitin is a polymer consisting of repeating units of N-
78 acetylglucosamine and the chemical structure is shown in Figure 1. It can be found in three
79 different polymorphic forms: α , β , and γ . The form found is dependent upon the source and
80 differs in the arrangement of the individual chitin chains and the resulting side chain interactions.
81 In the α -chitin form, the chains are organised in an anti-parallel fashion, whereas in the β -form,
82 the chains align in parallel (Jang *et al.*, 2004). Intra- and inter-chain hydrogen bonding between
83 the N-H \cdots O=C groups leads to a rigid, highly ordered crystalline structure (Jang *et al.*, 2004;
84 Sikorski *et al.*, 2009). γ -chitin is a mixture of the two arrangements. Chitosan is a derivative of
85 chitin, which differs in the amount of acetyl groups (CH₃CO) in the side chain; above 50 %
86 acetylation is termed chitin, and values below are termed chitosan (Khor *et al.*, 2003).



87 *Figure 1. Chemical structure of the chitin molecule, showing two of the N-acetylglucosamine subunits that repeat to form the*
88 *polysaccharide.*

89 The largest source of chitin currently comes in the α -form as a waste stream from the food
90 industry during preparation of shellfish. The typical composition of shellfish waste is 20-30 wt%
91 chitin, 30-40 wt% proteins, 30-50 wt% calcium carbonate, and lipids and astaxanthin (<1 wt%)
92 (Zeng *et al.*, 2011). The annual quantity of shellfish harvested is estimated to be 29.900.000
93 tonnes per year, ranging in chitin content from 2-12 wt% (wet basis) of the crustaceans total
94 body mass (Synowiecki & Al-Khateeb, 2003). This corresponds to a minimum annual chitin
95 production of approximately 6.000.000 tonnes, based on the lowest chitin content. Instead of
96 sending the shrimp shells to landfill or discarding in the sea, it is potentially economically and
97 environmentally desirable to valorise the shrimp shell waste stream (Yan & Chen, 2015).

98 This polysaccharide has a number of interesting properties that has led to increased interest
99 within academia and industry, especially in the fields of biomedicine and food. According to Yu
100 *et al.* (2014), chitin has excellent mechanical properties due to its light weight and high strength,

101 as well as good biocompatibility. Some examples of the applications of chitin within the
 102 literature include: emulsion stabilisation (Perrin *et al.*, 2014), wound healing and drug delivery
 103 (Khor & Lim, 2003), fruit juice clarification and gel formation (Rinaudo, 2006). Several studies
 104 showed the use of chitin as a biosorption material for the removal of heavy metals during waste
 105 water treatment. Pinto *et al.* (2011) investigated two commercially available chitin products for
 106 heavy metal removal during waste water treatment from mining. Under alkali and neutral
 107 conditions, it was possible to completely remove iron, lead and zinc, and partially remove
 108 cadmium, cobalt, copper and manganese at a chitin load of 2 g/L (Chitorem SC-20) (Pinto *et al.*,
 109 2011). Currently, there are many uses for chitin being explored within the literature, yet none are
 110 carried out, to the author's knowledge, on a commercial scale.

111 Another new promising area under investigation involves the use of materials on a nanoscale
 112 derived from natural sources, including chitin. Chitin particles on the nanoscale size range are
 113 referred to as whiskers, nanocrystals, nanofibers or nanorods. Within this thesis, chitin
 114 derivatives of a nano-size will be referred to as chitin nanocrystals (CHNC). Nanocrystals are
 115 defined by having a length or width of less than 100 nm in at least one dimension. Table 1 shows
 116 the sizes of CHNC found within the literature and also the methods utilised to measure it.

117
 118 *Table 1. Nanocrystal size comparison from various literature sources, including the microscopy technique used for measurement.*

Length (nm)	Width (nm)	Method	Source
100-600	4-40	TEM	Gopalan Nair & Dufresne, 2003
100-700	5-30	TEM	Lin <i>et al.</i> , 2012
240	18	TEM	Tzoumaki <i>et al.</i> , 2010
200-500	10-15	TEM	Goodrich & Winter, 2007

119
 120 The use of CHNC is becoming popular because of their low cost, rigidity, high aspect ratio and
 121 highly crystalline structure (Lin *et al.*, 2012). Isolated Chitin consists of a high crystalline
 122 structure compared its unhydrolysed counterpart (Zhang *et al.*, 2015). CHNC also have the
 123 additional features of a larger surface area and Young's modulus compared to crude chitin
 124 (Perrin *et al.*, 2014), thermal and chemical stability (Wang *et al.*, 2015).

125 To obtain CHNC, different temperatures, types of acid, acid concentration and duration have
 126 been employed in the literature, varying these conditions adjusts the degree of hydrolysis (Lin *et al.*,
 127 2012). The principle steps to prepare CHNC include: an alkali treatment to remove
 128 contaminating proteins, bleaching to breakdown contaminating light emitting chromophore
 129 molecules, hydrolysis with an acid at an elevated temperature to reduce the chitin chain length,
 130 reducing to a nanoscale size, as well as demineralising using washing and centrifugation, and
 131 finally sonication to break up the chitin aggregates (Tzoumaki *et al.*, 2010).

132 CHNC have been studied for a number of different applications. Gels from natural sources have
133 been studied due to their many applications in the food, pharmaceutical, cosmetic and paint
134 industries (Tzoumaki *et al.*, 2010). One area of interest which has not been widely studied is the
135 use of CHNC as a catalyst support material. For an optimum catalyst support the following
136 criteria are necessary: large surface area, cheap, unreactive in reaction medium, thermal stability
137 (reaction and preparation dependent), widely available, recoverable (catalyst remains on the
138 support) and non-toxic. Little research is available on the use of chitin or CHNC as catalyst
139 carriers in contrast to cellulose, the most abundant polysaccharide in nature. Using chitin
140 supported ruthenium, Matsuoka *et al.* (2015) found high yields during the hydration of various
141 nitriles to amides under aqueous conditions. As well as achieving high yields, the catalyst
142 preparation was relatively easy and tested on a variety of nitriles with aromatic, heteroaromatic
143 and aliphatic substituents. Wang *et al.* (2015) produced chitin microspheres by employing a sol
144 gel method. The microspheres were loaded with the catalyst α -amylase. After using the catalyst
145 for 10 repeated cycles, an activity of 95 % of the enzyme was still achieved.

146 In this thesis it is hypothesised that CHNC have superior properties as a catalyst support in
147 contrast to crude chitin in terms of porosity and available surface area. No previous research has
148 been found on the use of CHNC in the field of catalyst supports. The aim of this study was to
149 examine the effects of support preparation and calcination/reduction. Analysis to gauge the
150 success of the catalyst support includes thermal degradation analysis, physisorption,
151 microstructural investigations and XRD to understand the most viable route to prepare chitin as a
152 catalyst support material. Catalysis using metallic copper (Cu^0) can be employed to produce
153 sorbitol from glucose and fructose (Liu *et al.*, 2014b). The goal was to obtain metallic copper on
154 the chitin support materials with the least copper oxides.

155 2 MATERIALS & METHODS

156 2.1 MATERIALS

157 Coarse flake shrimp shells were obtained from Sigma-Aldrich (St. Louis, MO). Distilled water
158 was used in all experiments. Sigma-Aldrich was the supplier of hydrochloric acid (HCl,
159 concentrated 37 % v/v), sodium acetate (C₂H₃NaO₂), glacial acetic acid, potassium hydroxide
160 (KOH), sodium chlorite (NaClO₂), dialysis tubing cellulose membrane (cut-off: 14,000 Da),
161 copper nitrate trihydrate (Cu(NO₃)₂) and chitin from shrimp shells (Sigma-Aldrich: C9752).

162 2.2 ISOLATION OF CHITIN NANOCRYSTALS (CHNC)

163 The procedure described by Tzoumaki *et al.* (2009) was altered in terms of quantity of shrimp
164 shells used, centrifugation time and rotation speed. The nanocrystal dispersion did not settle after
165 numerous of centrifugation attempts, therefore higher G forces were used and longer
166 centrifugation times. In the procedure it was unclear in which step glacial acetic acid was used.
167 Thereby the assumption has been made that the acetic acid is used to alter the pH of the sodium
168 acetate buffer solution.

169 Raw shrimp shells were purified based on a protocol described by Tzoumaki *et al.* (2009) and
170 Nair & Dufresne (2003) to obtain chitin nanocrystals (CHNC). Approximately 60 g of shrimp
171 shells were suspended in 1400 mL of 5 % w/w KOH solution and boiled for 6 h under stirring to
172 remove contaminating proteins. This dispersion was kept at room temperature overnight under
173 continuous stirring. The dispersion was filtered and washed several times with distilled water
174 with the use of a Büchner funnel. The material remaining after the above procedure, was
175 bleached in 700 mL of a NaClO₂ solution (17 g of NaClO₂ in 1 L of 0.3 M sodium acetate buffer,
176 the pH was adjusted to 4.0 with the use of glacial acetic acid), at 80 °C for 6 h. The bleaching
177 solution was changed every 2 h, followed by rinsing the sample with distilled water.
178 Subsequently, the resulting material was kept in approximately 700 mL of 5 % w/w KOH
179 solution for 48 h to remove any residual proteins. The resulting dispersion was centrifuged at
180 2500 × g for 15 min. The chitin nanocrystal dispersion was prepared by hydrolysing the purified
181 chitin sample with 3 N HCl at a boiling temperature for 90 min under continues stirring. The
182 ratio of 3 N HCl to chitin solids was 30 mL /g chitin. After acid hydrolysis, the dispersion was
183 diluted with distilled water and followed by centrifugation at 2500 × g for 15 min. This
184 centrifugation process was repeated three times. Afterward, the dispersion was transferred to a
185 dialysis bag and dialyzed under running tap water for 2 h and left overnight into distilled water.
186 Appropriate volumes of 1 N HCl solution were used to adjust the final pH of the dispersion to 3.
187 To break the possible CHNC aggregates, the dispersion was finally subjected to an ultrasonic
188 treatment for 45 min. To avoid overheating of the sample, intervals of 5 min were given. The
189 nanocrystals were subsequently stored at 4 °C.

190 2.3 FREEZE DRYING

191 The procedure by Tzoumaki *et al.* (2009) resulted in the production of an aqueous dispersion.
192 For further analysis and impregnation, the nanocrystals had to be dried. Freeze drying was
193 chosen as the most sensible route to produce dried CHNC. Prior to freeze drying the samples
194 were frozen at -20 °C. Wu & Meredith (2014) investigated different freezing temperatures of
195 chitin nanofibers prior to freeze drying (-20,-80 and -196 °C); the highest temperature for
196 freezing resulted in the smallest pore size in a range of 0.33 – 3.2 µm.

197 The chitin dispersions were stored at -20 °C for 24 h before freeze drying. The frozen chitin was
198 further cooled down to -80 °C and then dried in a freeze-dryer (Christ alpha 2-4 LDplus) at
199 0.011 mbar for 48 h and for the final drying step at 0.001 mbar for 24 h.

200 2.4 FOURIER TRANSFORM INFRARED SPECTROSCOPY (FTIR)

201 To confirm the correct functional groups are present within the samples, Fourier Transform
202 Infrared Spectroscopy (FTIR) was used to compare to literature, FTIR spectra of the samples
203 were obtained on a Varian Scimitar 1000 FT-IR spectrometer equipped with a DTSG-detector
204 and a PIKE MIRacle ATR. The sample was placed on a diamond w/ZnSe lens single reflection
205 plate. The measurement resolution was set at 4 cm⁻¹, and the spectra were collected in the ATR
206 (Attenuated Total Reflection) mode in the range 4000-650 cm⁻¹ with 64 co-added scans. The
207 obtained data were transferred into excel for further analysis.

208 2.5 CATALYST IMPREGNATION, INCIPIENT WETNESS IMPREGNATION (IWI)

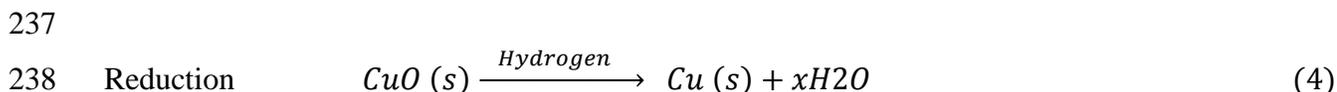
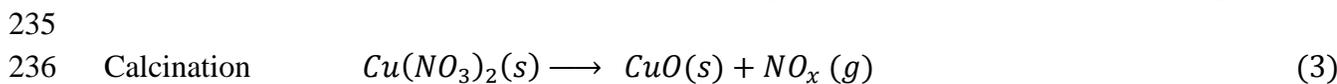
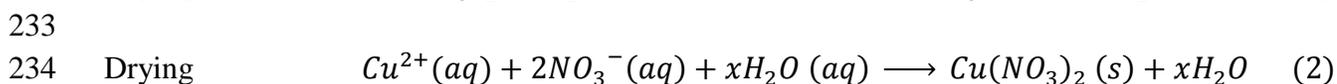
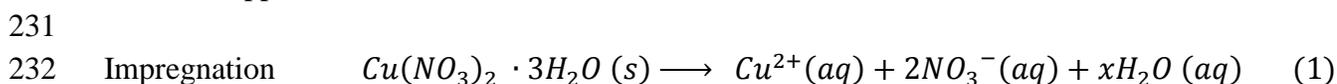
209 Dry CHNC and SHSH were impregnated with an aqueous solution of copper nitrate trihydrate.
210 To achieve a sufficient wetness, liquid was added to the catalyst support dropwise until complete
211 saturation on a visual basis. A liquid/support ratio of 3.6 mL/g was determined for both CHNC
212 and SHSH. To obtain a 5 wt% copper solution to add to the support, it was calculated that 19 g
213 of copper nitrate trihydrate was required, based on molar masses. After impregnation, the
214 samples were dried for 3 h at 60 °C, and will be referred to as 5Cu-CHNC and 5Cu-SHSH for
215 impregnated CHNC and impregnated chitin from shrimp shells, respectively.

216 2.6 CALCINATION/REDUCTION

217 Calcination/reduction of the catalyst impregnated support was done in a tubular furnace. A
218 quartz weighing boat was used to carry the catalyst/support which weighed about 1 g ±0.3. The
219 sample was heated with a heating rate of 5 °C/min to 250 °C and left for 2 h. A gas hourly space
220 velocity (GHSV) of 26 % N₂ and 30 % H₂ was used. Afterwards the H₂ flow was turned off and
221 flushed for 30 min with N₂ to clear out the volatile and harmful H₂ gas.

222 A 5 wt% Cu sample was prepared by impregnation of the hydrolysed chitin support to incipient
223 wetness (see section 2.5). Two different routes were taken, (i) direct reduction and a (ii) two-step

224 reduction. The first route (i) involved transferring the impregnated sample to a tubular oven,
 225 where the sample was heated to 250 °C (5 °C min⁻¹) in a the gas hourly space velocity (GHSV)
 226 during thermal treatment, N₂/ H₂, air, or N₂ was used and subsequently left in the same gas flow
 227 for 2 h. Alternatively (ii), the impregnate was dried at 200 °C in a static air oven for 2 h and the
 228 sample is subsequently reduced in a tubular oven with a flow of N₂/H₂, air, or N₂. Equations 1-4
 229 show the intermediate steps (impregnation, drying, calcination and reduction) necessary to obtain
 230 metallic copper.



239
 240 The calcination/reduction temperatures was held at the given temperature °C for 2 h.

241 Various conditions of drying and calcination/reduction were tested to study the preparation
 242 effects on the resulting impregnated samples. These conditions can be found in Table 2. In this
 243 thesis, chitin nanocrystal samples (CHNC) are denoted as CHNC-X, where X stands for the gas
 244 or gas combination used during reduction and/or calcination. This principle accounts the same
 245 for shrimp shells (SHSH) these are denoted as SHSH-x. CHNC reduced in H₂ (g) are denoted as
 246 CHNC-H2 or SHSH-H2, respectively. Impregnated samples denoted with “5Cu”, corresponds to
 247 5 % copper impregnation to the support material. Thus samples denoted as 5Cu-CHNC-H2 refers
 248 to 5 % copper impregnated onto a CHNC support and reduced with hydrogen gas.

249

250 Table 2. Sample preparation, drying temperature/time and metal particle size in the impregnated Cu/CHNC samples.

Sample	Drying		Calcination/reduction		
	Temp (°C)	Duration (h)	Temp (°C)	Gas	GHSV (mL/min)
5Cu-CHNC	120	3	250	N ₂ /H ₂	63/30
5Cu-CHNC	60	4	200	N ₂ /H ₂	63/30
5Cu-CHNC	60	4	150	N ₂ /H ₂	63/30
5Cu-SHSH	120	3	250	N ₂	63
5Cu-SHSH	60	3	200	H ₂	30
5Cu-SHSH	120	3	200	N ₂ /H ₂	63/30
5Cu-SHSH	60	3	200	Air	-

251 2.7 THERMAL DEGRADATION (TGA)

252 To determine the degradation temperature profiles a TGA/DSC1 STARe system Mettler Toledo
253 was used. The initial mass of each sample was 10 ± 0.5 mg and placed into a 70 μ L aluminium
254 crucible without lid, then loaded into the TGA and heated from 30 °C to 900 °C. The degradation
255 temperature of the samples were studied at a heating rate of 20 °C/min in a nitrogen atmosphere
256 with a purge rate of 50 mL/min. Data was acquired by means of *STARe* software and analysed
257 using Microsoft Excel.

258 2.8 X-RAY DIFFRACTION (XRD)

259 To determine the crystallinity of the nanomaterials and catalyst, powder X-ray diffraction
260 patterns were obtained by means of X-ray diffraction (XRD). With the use of the following
261 operating conditions: 40 kV, 40mA, the relative intensity was recorded at a scattering range (2θ)
262 of 10° to 90°. The measuring time was 1s/step. Crystal lattices and their specific peaks can be
263 compared with the measured samples.

264 2.9 TRANSMISSION ELECTRON MICROSCOPY (TEM)

265 Transmission electron microscopy imaging was performed on a JEOL JEM-2000FX
266 transmission electron microscope operated at an acceleration voltage of 80 kV. One drop of the
267 CHNC in methanol suspension (0.025 w/w %) was deposited on a carbon-coated copper grid and
268 allowed to air dry. Optical micrographs of the CHNC dispersions were captured by an Olympus
269 BX 51 polarizing optical microscope fitted with a digital camera (Olympus, DP 50). The
270 specimens were prepared by placing a thin layer of sample between a microscope slide and a
271 coverslip.

272 2.10 SCANNING ELECTRON MICROSCOPY (SEM)

273 The surface morphology of dry samples suspended in HPLC grade methanol was determined
274 using a scanning electron microscope (SEM). The samples were mounted on an aluminium
275 mount and sputtered with tungsten. The samples were scanned at an accelerating voltage of 10
276 kV, with a working distance of ± 40 mm, as appropriate. EDX measurements were taken
277 throughout various scans to determine the localisation of various elements within the samples.

278 2.11 PHYSISORPTION

279 Both adsorption and desorption isotherms were measured with the Micromeritics TriStar II plus.
280 The samples were dried overnight at high vacuum at 120 °C prior to measurement with nitrogen.
281 The specific surface areas were calculated according to the Brunauer-Emmett-Teller (BET)
282 method.

283 3 RESULTS & DISCUSSION

284 3.1 CHITIN NANOCRYSTALS AND SHRIMP SHELL PREPARATION

285 The procedure to obtain CHNC is an adaptation from the procedure presented by Tzoumaki *et al.*
286 (2009). The resultant CHNC that were produced after acid hydrolysis, purification and freeze
287 drying the hydrolysed were fluffy and white, the structure can be seen in Figure 2.



288
289
290 *Figure 2. Photograph of chitin nanocrystals (CHNC) after freeze drying.*

291 3.1.1 FTIR

292 To ensure that a pure CHNC sample had been prepared, FTIR was used to identify the functional
293 groups that were present. Figure 3 shows the spectra recorded for SHSH, CHNC and a reference
294 SHSH of the samples show a similar FTIR spectra, all of the peaks present in the reference
295 sample are present in the SHSH and CHNC sample. Previously in the literature (Goodrich &
296 Winter, 2007), the FTIR spectra of crude chitin and CHNC have been presented. The assignment
297 of the peaks at 1656, 1621 & 1556 cm^{-1} correspond to two amide regions within the chitin chain

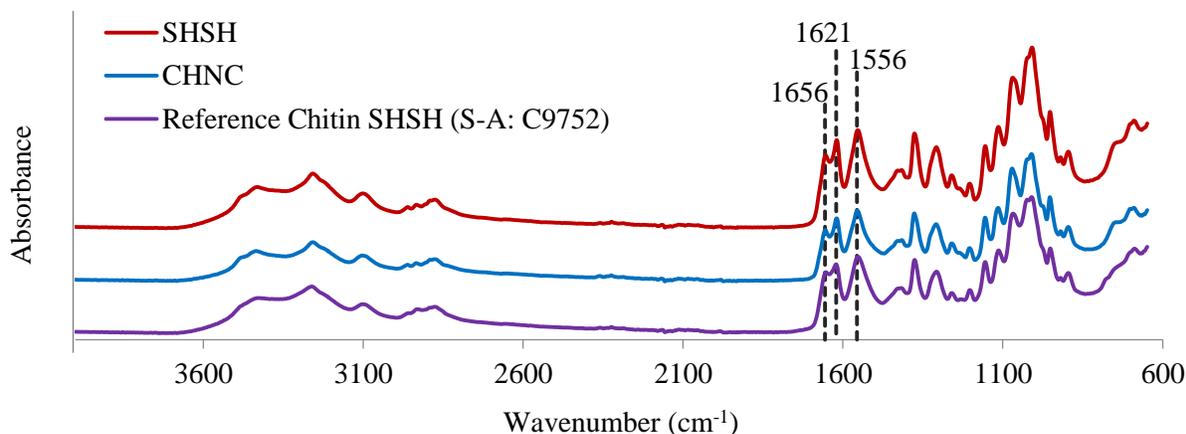


Figure 3. FTIR spectra of various chitin based samples. Shrimp shells (SHSH), chitin nanocrystals (CHNC). Amide peaks are assigned using a literature source (Goodrich & Winter, 2007).

298 Structure (Goodrich & Winter, 2007). These spectra indicate that the crude chitin was already
299 pure without contamination from protein or any other substance. An assumption is made that the
300 deprotonation step with KOH in the procedure of Tzoumaki *et al.* (2010) can be discarded in
301 future work.

302

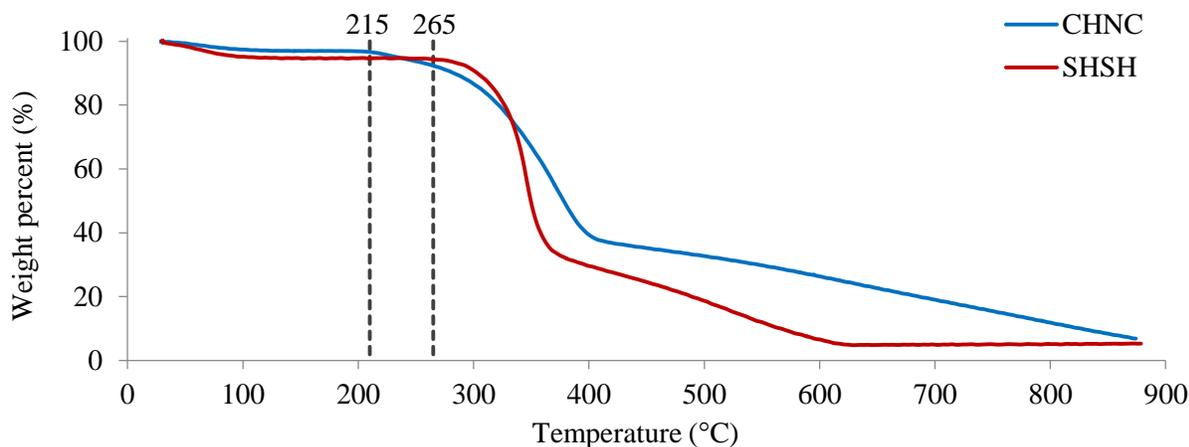
303 3.1.2 TGA

304 Thermal gravimetric analysis (TGA) gives us information about the degradation of samples
305 during thermal treatment. This is useful information because if the calcination/reduction after
306 impregnation is carried out at temperatures above the degradation temperature of the catalyst
307 carrier, then the support is not suitable. This is also useful information for if the reaction to be
308 catalysed occurs at elevated temperatures.

309 Figure 4 shows the TGA profiles of pyrolysis for CHNC and crude chitin SHSH at a heating rate
310 of 20 °C/min. The onset of water evaporation was coupled with weight-loss centred at about
311 100 °C (Wanjan *et al.*, 2005). From Figure 4, the water content of SHSH and CHNC can be
312 measured as 5 % and 3 %, respectively.

313 For SHSH, the onset of chitin degradation occurred at 265 °C, compared to CHNC which
314 occurred at 215 °C. It was found that 61 % of the CHNC was degraded during the second weight
315 loss episode in the TGA curve. The weight loss for SHSH during heating up to 370 °C was
316 measured at 66 %. The thermal stability of chitin is lower than that of crude chitin sample
317 (SHSH), as degradation occurred at a lower temperature. At these temperatures,
318 depolymerisation of the chitin occurs, which resulted in the production of volatile components
319 and char (Wanjan *et al.*, 2005). Köll *et al.* (1991) confirmed that the deacetylation of chitin,
320 which increased with increasing temperature, was not the largest contributing factor to weight
321 loss. The degree of acetylation was measured using HPLC. This means that inter-chain hydrogen
322 bonding is still present between the chitin chains after thermal treatment.

323 The maximum decomposition temperature of α -chitin was found in the literature to have a value
324 of 419.4 °C at a heating rate of 20 °C/min (Jang *et al.*, 2004). Changes to the heating rate during
325 TGA leads to variation in the maximum decomposition temperature (T_m). The maximum
326 decomposition temperature of CHNC found within this thesis is comparable to the literature
327 source measured at the same heating rate (Jang *et al.*, 2004).



328 Figure 4. TGA curves to show the effect of increasing temperature at a rate of 20 °C/min on the degradation of chitin
 329 nanocrystals (CHNC) and shrimp shells (SHSH).

330

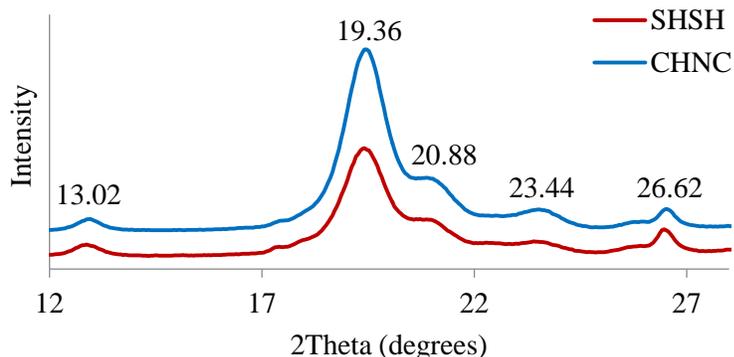
331 3.1.3 XRD

332 The intensity and position of the peaks in an XRD pattern provides information on the crystal
 333 structure of the examined sample. XRD patterns were recorded to compare the effects of CHNC
 334 preparation versus the crude SHSH prior to hydrolysis. The comparison can be seen in Figure 5,
 335 including the annotation of the observed 2Theta values. The XRD patterns for CHNC and the
 336 crude SHSH contained peaks at similar 2Theta values. Crystallinity of CHNC has also been
 337 studied previously in the literature (Goodrich & Winter, 2007), including the assignment of
 338 2Theta values to crude shrimp shells and CHNC to peaks measured in the range of 7-30°. Similar
 339 2Theta values were also found for CHNC prepared by Liu *et al.* (2014a) for use in waste water
 340 treatment. These 2Theta values are similar to those found within this study (Table 3). The peaks
 341 within the XRD pattern correspond to the Miller indices (hkl), which correspond to different
 342 planes within the molecule under investigation. Different planes intersect atoms differently,
 343 hence why a number of peaks are seen per crystal structure.

344 The intensity of the peak at the 2Theta value of 19.36° was greater for CHNC than for SHSH,
 345 indicating a difference in the crystallinity between the two samples. The preparation of CHNC
 346 resulted in the formation of a more crystalline material than SHSH, confirmed by XRD. This was
 347 also found by Goodrich & Winter (2007), where acid hydrolysis resulted in an increase of
 348 crystallinity of chitin by 8 %. This value was measured as the ratio of the sum of areas under the
 349 crystalline diffraction peaks of chitin to the total area under the curve.

350
351

Table 3. Observed and literature (Goodrich & Winter, 2007) reported 2Theta values

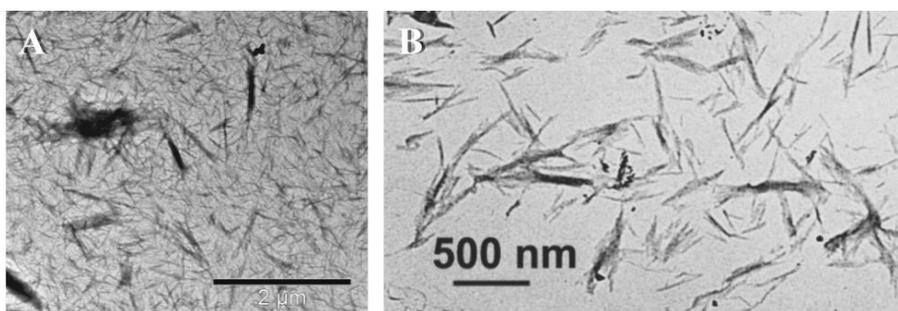


2Theta _{literature}	2Theta _{observed}
12.82	13.02
19.40	19.36
20.88	20.88
22.47	23.44
26.65	26.62

352 Figure 5. A comparison of XRD patterns of crude shrimp shells (SHSH) and
353 chitin nanocrystals (CHNC).

354 3.1.4 Transmission electron microscopy (TEM)

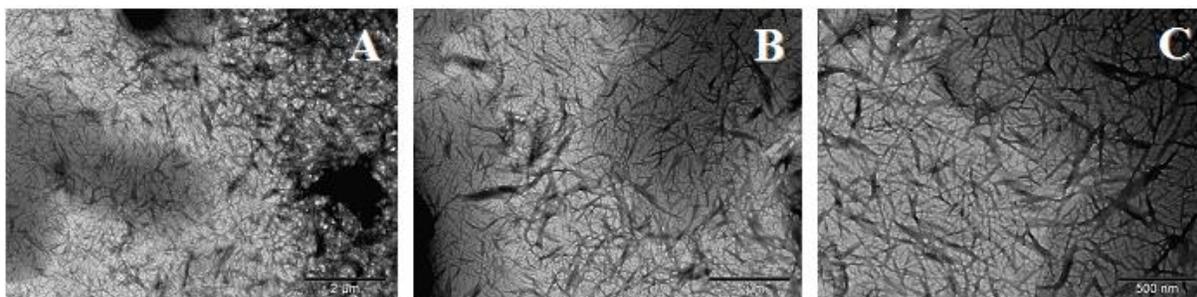
355 To characterise the CHNC, TEM was carried out to determine the size and organisation of the
356 nanocrystals. Table 1 shows the size ranges of CHNC found within the literature. Figure 6A
357 shows a transmission electron micrograph (TEM) of a dilute suspension 0.025 wt% of CHNC.
358 TEM confirms the presence of evenly dispersed CHNC in the sample. Within this study, it was
359 not possible to measure the length or width of the crystals due to oversaturation. Tzoumaki *et al.*
360 (2010) made a dispersion of 0.02 wt% CHNC, and obtained a more sparse dispersion. The
361 differences between the literature dispersion by Tzoumaki *et al.* (2010) and that found within this
362 study can be seen in Figure 6B. Thereby, a more suitable dilution of the CHNC sample should be
363 used for further chitin microscopic investigation e.g. 0.01 wt%. No microscopic images were
364 possible of the crude SHSH due to their large size on the millimetre scale.



365
366 Figure 6. A: TEM micrograph (2 μm) of obtained chitin nanocrystals with a 0.025 wt%, B: A TEM micrograph of 0.02 wt%
367 CHNC taken from Tzoumaki *et al.* (2010).

368

369 In Figure 7, TEM micrographs of CHNC are presented, at various magnifications. Figure 7A
 370 shows a magnification with a scale bar of 2 μm , with a broad nanocrystal network, dark patches
 371 can be seen to the right of the image where the assumption is made that the layers of nanocrystals
 372 were overcrowded. Looking into another part of the grid, with a scale bar annotated of 1 μm
 373 (Figure 7B). At the greatest magnification of 500 nm (Figure 7C), it was possible to see
 374 individual CHNC organised in a network, varying in width and length.
 375



376
 377 Figure 7. TEM micrographs of chitin nanocrystals (CHNC), prior to impregnation. Size of scale bar: A 2 μm ; B 1 μm ; C 500 nm.

378 3.2 IMPREGNATION OF SAMPLES

379 After impregnation of the support materials, drying is necessary for further analysis. The
 380 relevance of the drying step is usually neglected in the literature. However, a study was found in
 381 the literature highlighting the importance of the drying step (Toupance *et al.*, 2000). Figure 8
 382 shows the process for impregnating the support with the copper catalyst.

383 Figure 8A corresponds copper nitrate trihydrate was weighed out. A dilution was made with
 384 miliQ to obtain a 5 % wt copper solution, shown in Figure 8B, details of the copper solution
 385 preparation were detailed in Equation 5. Chitin was impregnated by the means of incipient wet
 386 impregnation. It was observed that the chitin saturation point was about 3.6 mL of solution per
 387 gram chitin (see Figure 8C for completely wetted CHNC). After the addition of this amount, all
 388 copper solution was absorbed by the chitin support. Other catalyst supports were studied by
 389 Delannoy *et al.* (2006), 0.8 mL/g of different catalyst supports were used. CHNC seems to excel
 390 in solution uptake compared to conventional catalyst support.
 391

$$392 \quad M = \frac{C_{Cu}}{100} * \frac{V}{mw_{Cu}} * mw_{Cu(NO_3)_2} \quad (5)$$

393 Where:

394	M	mass of copper nitrate trihydrate (g)
395	C_{Cu}	copper in the solution (%)
396	V	volume of MiliQ water (mL)
397	mw_{Cu}	molecular weight of copper (g/mol)
398	$mw_{Cu(NO_3)_2}$	molecular weight of copper nitrate trihydrate (g/mol)

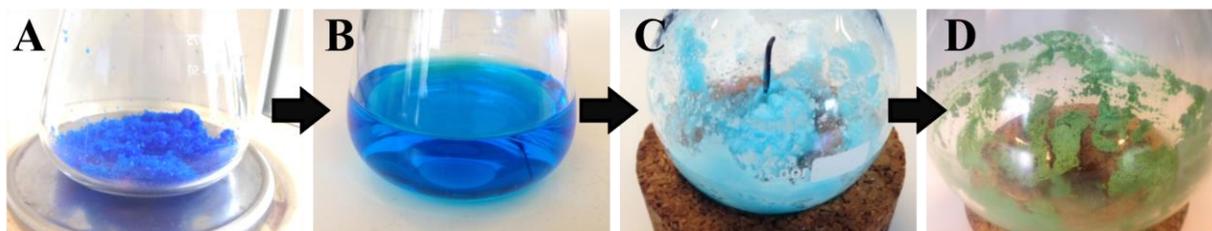


Figure 8. From left to right, A: Weighing copper; B: Addition of miliQ water to copper; C: Mixed impregnated chitin with copper; D: Drying of chitin.

399

400

401 A concentration of 5 % copper was used as this value has been used previously in the literature
 402 (Chang *et al.*, 2003). Figure 8C shows the impregnated CHNC after homogenisation by vigorous
 403 shaking on a rotational vortex machine. Subsequently the impregnated sample was dried in an
 404 oven, the resultant catalyst and its carrier was a dried green substance consisting of impregnated
 405 CHNC, Figure 8D.

406 Initially impregnated SHSH and CHNC samples were dried at 120 °C in order to perform further
 407 analysis. After heating at 120 °C for 3 h, an inhomogeneous colouration was observed for the
 408 CHNC only (Figure 9). A comparison of the XRD patterns was performed to compare the
 409 crystallinity of both regions in the sample (Figure 10). The brown coloured 5Cu-CHNC(Brown)
 410 had reduced chitin peaks (10-30°), in comparison to the green coloured sample. An increase in
 411 different formations of copper oxides (Cu^{2+}) was also found for the brown coloured sample,
 412 which were undesirable because metallic copper (Cu^0) was the required catalyst. Metallic copper
 413 (Cu^0) has been previously studies in the literature using XRD, the 2Theta values correspond well
 414 to those found in this thesis (Park *et al.*, 2007).



Figure 9. Impregnated nanocrystals (5Cu-CHNC) dried at 120 °C.

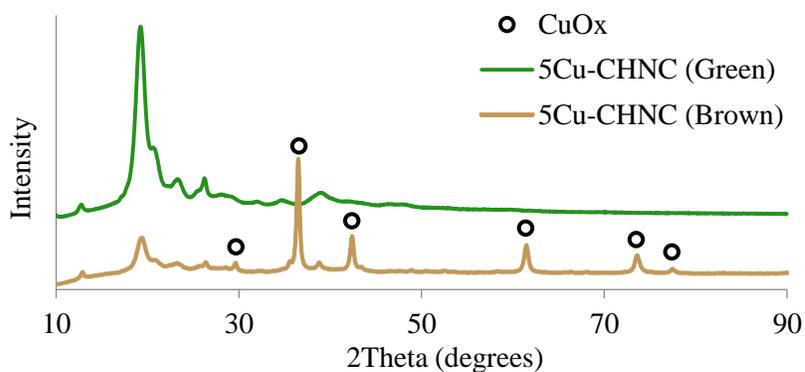


Figure 10. XRD patterns of impregnated chitin nanocrystals (5Cu-CHNC) dried at 120 °C. ○ indicates 2Theta values corresponding to undesirable copper oxides.

415

416 Crude SHSH and CHNC were then dried at a lower temperature of 60 °C. There was no effect of
417 inhomogeneity in colour of the impregnated chitin at 60 °C observed, see Figure 11. When
418 considering the process of drying chitin, it is also vital to consider the residence time in the oven.
419 It was found that leaving chitin in the oven for 4 h at 60 °C gives an evenly dried sample,
420 completely green in colour throughout (see Figure 11). A sample was dried overnight (>15 h) at
421 60 °C, XRD analysis (Figure 12) showed there was no indication that copper oxides were formed.
422 Therefore if the sample is dried at 60 °C, we can assume that the drying time does not influence
423 the production of copper oxides.



Figure 11. Impregnated chitin (5Cu-CHNC) nanocrystals dried at 60 °C.

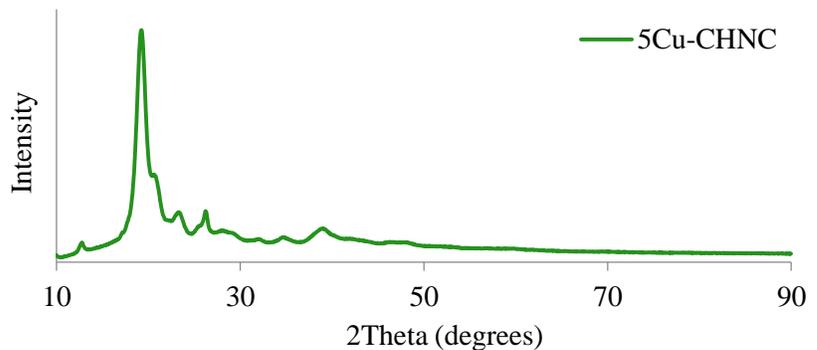


Figure 12. XRD analysis of impregnated chitin nanocrystals, dried at 60 °C.

424

425 A drying temperature of 60 °C was employed for the following experiments due to the reduced
426 production of copper oxides in the dried, impregnated CHNC.

427 3.3 CALCINATION/REDUCTION OF IMPREGNATED SAMPLES

428 In order to produce the copper catalyst in the correct form (Cu^0), calcination/reduction was
429 performed using various gas combinations and temperatures. Details of these conditions can be
430 found in Table 2. Various methods were used (XRD, physisorption and microscopic
431 investigation) to compare and contrast crude SHSH and CHNC.

432 3.3.1 XRD patterns of SHSH

433 Figure 13 shows crude SHSH, dried impregnated SHSH with 5 wt% copper solution (5Cu-
434 SHSH) and an impregnated sample (5Cu-SHSH-N₂H₂(250)) reduced at 250 °C with combined
435 nitrogen (N₂) and hydrogen (H₂) gases. Drying at 120 °C (5Cu-SHSH) resulted in a decrease in
436 the 20° peak compared to the crude SHSH sample. Thereby indicating a less crystalline form of
437 chitin upon impregnation and drying. Copper was reduced simultaneously with N₂ and H₂ gases
438 to prepare sample 5Cu-SHSH-N₂H₂(250). After the reduction process, copper peaks can clearly
439 be seen Figure 13 5Cu-SHSH-N₂H₂(250) at a 2Theta value of 43° and 50°. For copper (Cu)
440 another peak should be visible at a 2Theta value of 74° according to Park *et al.* (2007), but up to
441 this value was not measured due to boundary measurements.

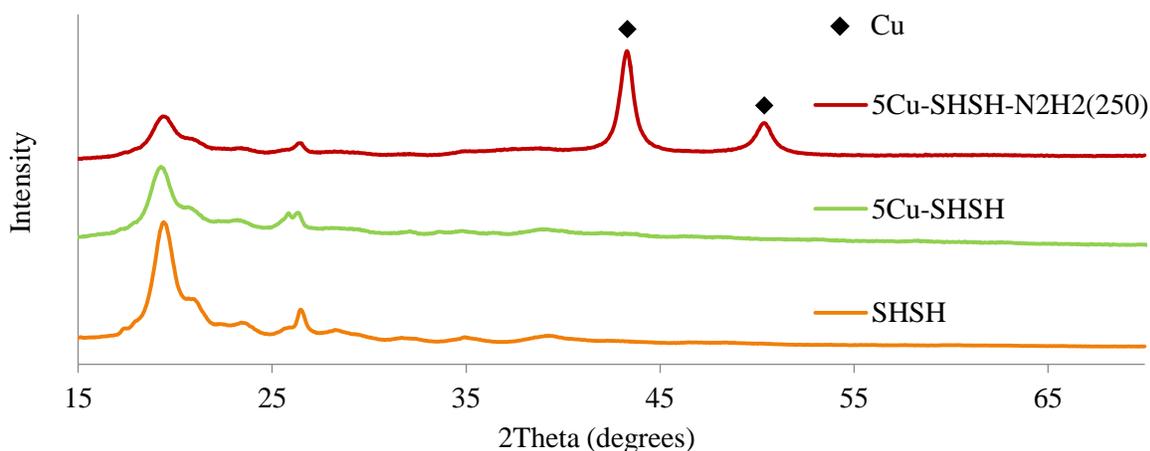


Figure 13. XRD patterns of SHSH, 5-CuSHSH and 5Cu-SHSH-N₂H₂(250), ◆ indicates 2Theta values corresponding to metallic copper.

442
443 Indirect reduction (2.6ii) at lower temperatures was performed to study the effects on the
444 crystallinity of chitin and copper. A lower temperature of 200 °C was chosen for investigation as
445 this falls below the thermal degradation temperature (T_m) of SHSH. The chitin peaks which were
446 found between 2Theta values of 10° and 30° in Figure 14 were not visible for any of the SHSH
447 samples prepared at lower temperature with indirect reduction (Figure 14). Although chitin is
448 still present within the sample, this result suggests that the crystalline form of chitin is lost and
449 must become amorphous. Formation of copper oxides occurred during N₂ treatment (5Cu-SHSH-

451 N₂(200)) or during air followed by nitrogen treatment (5Cu-SHSH-Air(200)-N₂(200)). Copper
452 oxides are highlighted on the XRD patterns by open circles (○). When H₂ gas was subsequently
453 passed over the same sample (5Cu-SHSH-Air(200)-N₂(200)-H₂(200)), metallic copper (Cu⁰)
454 was present within the sample, however that was the only crystalline material that was found
455 using XRD.

456

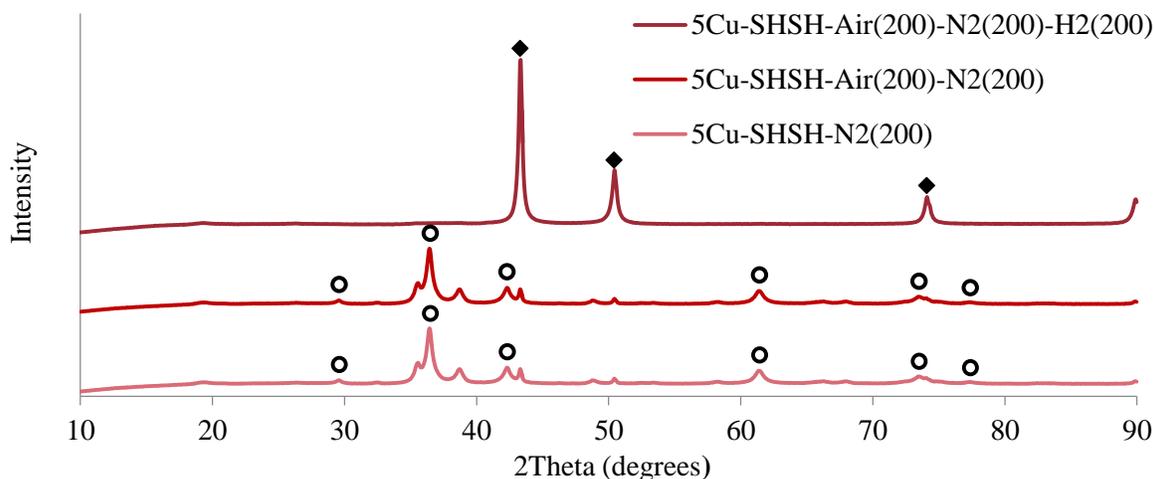


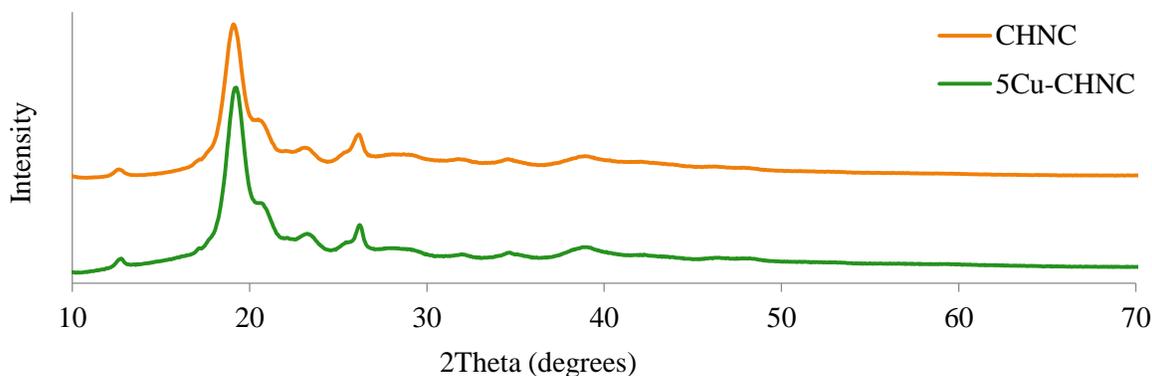
Figure 14. XRD patterns to show the effects of reduction/calcination on the crystallinity of the samples versus a dried sample without reduction/calcination. ◆ Indicates 2Theta values corresponding to copper, Copper oxide formations were indicated by ○.

457

458

459 3.3.2 XRD patterns of CHNC

460 In this section CHNC, impregnated CHNC and reduced CHNC samples were analysed using
461 XRD to determine their crystallinity. Initially CHNC was compared to the impregnated by a 5 %
462 copper solution (5Cu-CHNC). The 5Cu-CHNC went through a heat treatment of 60 °C to
463 remove the water prior to analysis, whereas CHNC was dried via freeze drying. No decrease in
464 the crystallinity of CHNC was observed. In addition to impregnation and drying no copper peaks
465 were observed indicating the absence of copper in a crystalline form.



466

Figure 15. XRD patterns of CHNC, and impregnated 5Cu-CHNC dried at 60 °C.

467 After direct reduction by N₂ and H₂ gas (2.6i), metallic copper peaks (Cu) can be seen in figure
 468 16 at 2Theta values of 43°, 50° and 74° with reduction carried out at 200 or 250 °C (5Cu-CHNC-
 469 N₂H₂(200) & 5Cu-CHNC-N₂H₂(250)). When reduction was performed at 150 °C (5Cu-CHNC-
 470 N₂H₂(150)), copper oxides were produced. This indicates that the reduction of copper oxides to
 471 copper occurs between in the region of 150-200 °C. Another peak should be visible at a 2Theta
 472 value of 94°, but that region was not analysed during this measurement.

473

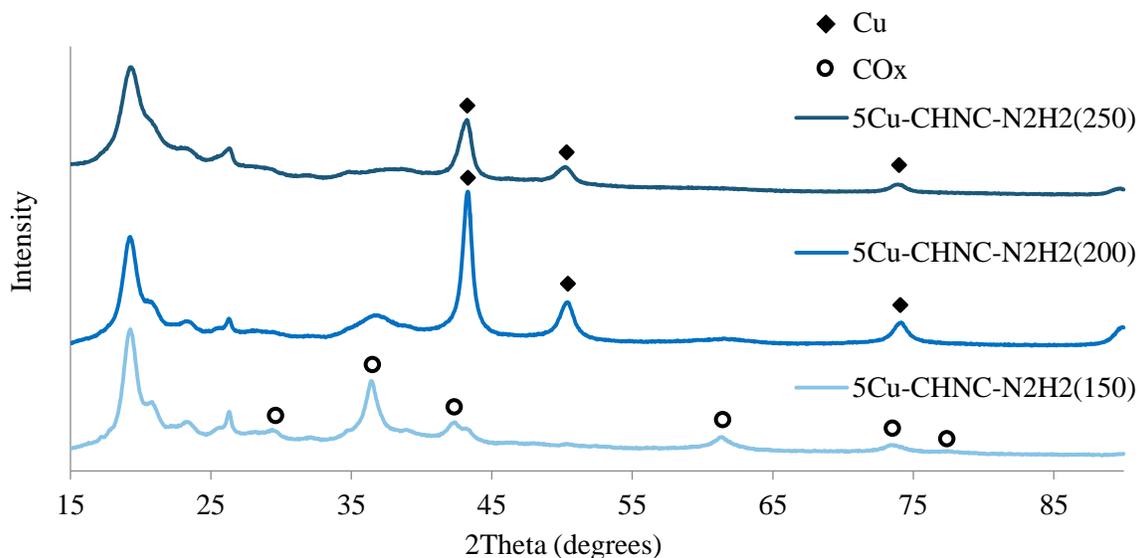


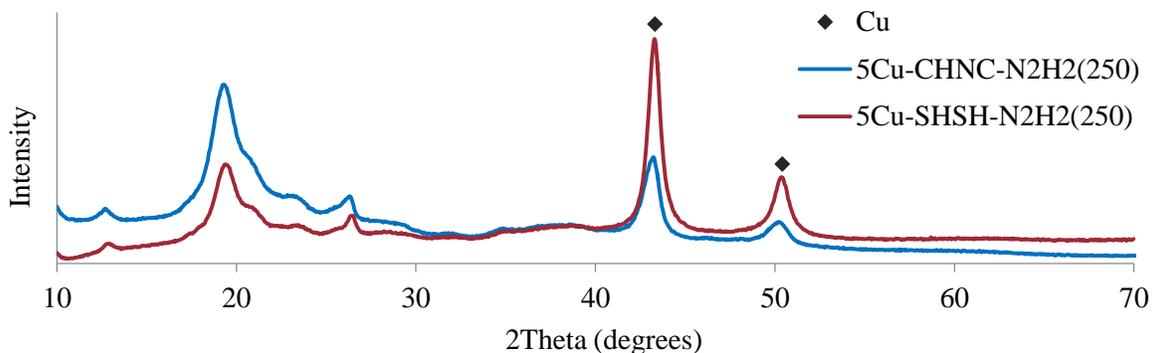
Figure 16. XRD patterns to show the effect of direct reduction temperature on the impregnated chitin nanocrystal (CHNC). Copper is indicated by ♦, Copper oxide formations were indicated by ○.

474

475

476 A comparison of the XRD patterns for reduced SHSH and CHNC can be seen in Figure 17. It
 477 was possible to observe crystalline chitin peaks in the region of 10-30° in both of the samples

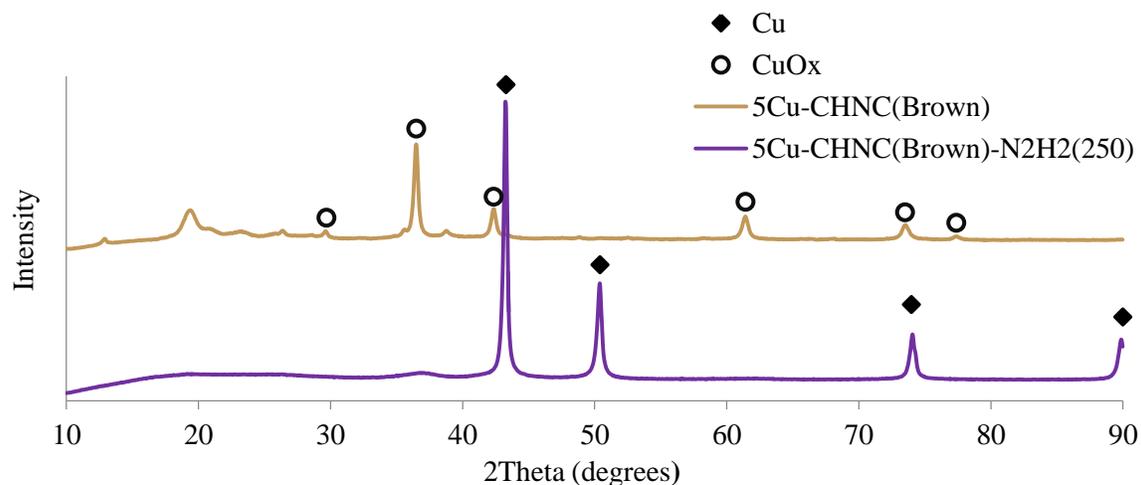
478 analysed. Metallic copper peaks were also observed at the same 2Theta values found previously
 479 within the literature (Park *et al.*, 2007). In terms of metallic copper, the crystallinity in the SHSH
 480 sample was greater than that found in the CHNC sample.



481
 482 *Figure 17. XRD patterns of reduced impregnated chitin from shrimp shells (5Cu-SHSH-N2H2(250)) and reduced impregnated*
 483 *chitin nanocrystals (5Cu-CHNC-N2H2(250)). Copper is indicated by ◆.*

484 3.3.3 XRD patterns of 5Cu-CHNC-N2H2(brown) sample

485 The resultant 5Cu-CHNC(Brown) chitin sample after drying at 120 °C was reduced with N₂ and
 486 H₂ gas to determine the catalyst carrier properties compared to the 5Cu-CHNC(green) sample
 487 after reduction (Figure 18). It can be seen that the crystallinity of chitin observed in the sample
 488 was reduced upon reduction in relation to the copper peaks (5Cu-CHNC(Brown)-N2H2(250)).
 489 Copper oxides were found within the impregnated sample prior to reduction (5Cu-
 490 CHNC(Brown)). Upon reduction, strong metallic copper peaks can be seen, which directly
 491 resulted from reduction of the copper oxides (Equation 4).



492
 493 *Figure 18. XRD analysis of dried impregnated chitin nanocrystals which turned brown at 120 °C (5Cu-CHNC (Brown)) and*
 494 *reduced chitin nanocrystal 5Cu-CHNC(Brown)-N2H2(250), Copper is indicated by ◆, Copper oxide formations were indicated*
 495 *by ○.*

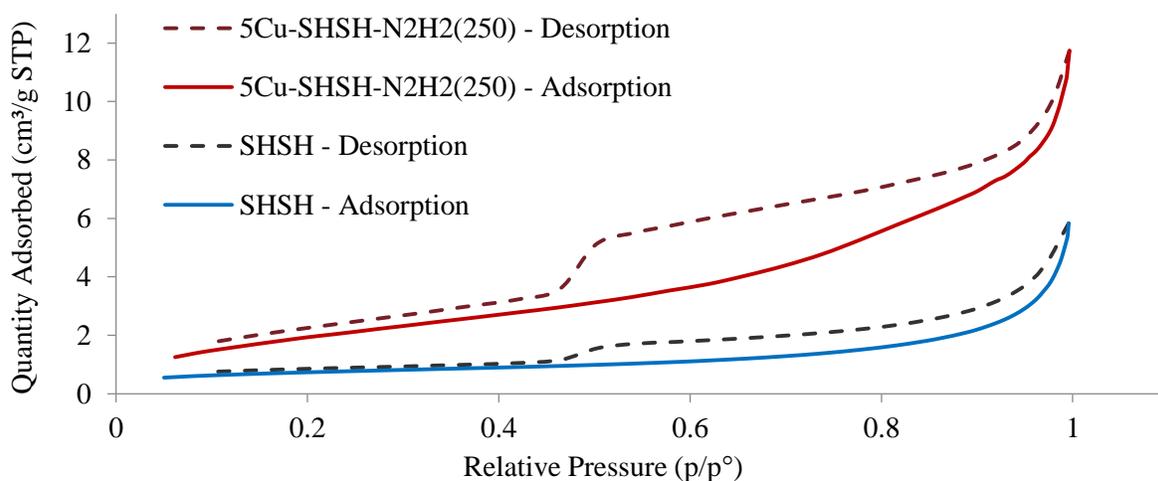
493 **3.3.4 Physisorption**

494 Physisorption was carried out with inert nitrogen gas to study the effects of drying and
495 calcination/reduction conditions on the sample characteristics. Table 2 shows the different
496 conditions for copper supported on both chitin source (SHSH & CHNC) which were tested.

497 **3.3.4.1 SHSH**

498 Figure 19 gives a comparison of crude SHSH versus a reduced impregnated SHSH sample (5Cu-
499 SHSH-N₂H₂(250)). The physisorption graph gives us information on absorption, the amount of
500 gas absorbed by the sample, and desorption describes the release of the gas. The reduced
501 impregnated SHSH sample (5Cu-SHSH-N₂H₂(250)) was able to absorb more N₂ gas than the
502 crude SHSH sample. This could be explained by more space for the gas to enter the catalyst
503 support in the impregnated reduced sample in comparison to the crude SHSH. This could
504 correspond to a structural change upon reduction. Both samples experienced an episode during
505 desorption, a barrier was present which occurred at a relative pressure of 0.5. More information
506 on the pore size and surface area (BET) of the materials can be found below in Table 4.

507



508

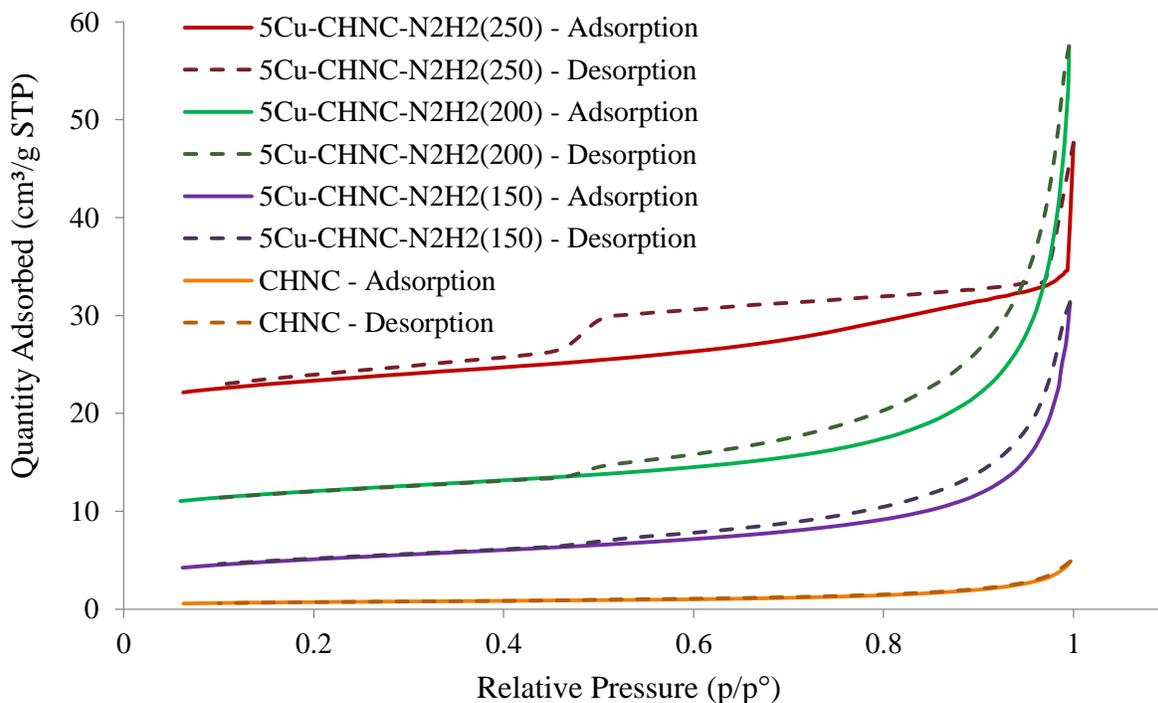
509 *Figure 19. Physisorption graph comparison of crude chitin from shrimp shells (SHSH) and reduced, impregnated SHSH.*

510

511 3.3.4.2 CHNC

512 Physisorption data was collected for CHNC after preparation and the effect of temperature (150,
513 200, & 250 °C) during direct reduction was also studied. The results can be found in Figure 20.
514 When comparing CHNC to SHSH, without impregnation or reduction, it can be seen that both
515 materials absorb the same volume of gas. However, upon desorption, SHSH showed a barrier
516 had to be overcome, whereas CHNC did not. When studying the effect of thermal treatment
517 during reduction (150, 200 and 250 °C) of CHNC, a stepwise increase in the amount of N₂ gas
518 absorbed was found with increasing temperature. The increase was found to be stepwise with the
519 greatest volume of gas absorbed by the temperature reduced at 250 °C (5Cu-CHNC-N₂H₂(250)).
520 At a relative pressure value of 1, this stepwise increase in the volume absorbed with increasing
521 reduction temperature was not observed. This could be due to a discrepancy in the measurement
522 of the sample prepared with reduction at 250 °C. This should be investigated further. The
523 desorption was also affected by the reduction temperature (Figure 20). A desorption barrier was
524 found upon reduction at a relative pressure of 0.5 and the intensity of the barrier increased with
525 increased temperature.

526



527

528

Figure 20. Physisorption graph of chitin nanocrystals (CHNC), with different reduction temperatures in the range of 150 to 250 °C

529 A comparison was made to determine which material, CHNC or SHSH, could absorb more gas
530 at the same reduction temperature. After impregnation and reduction at 250 °C, CHNC could
531 absorb a greater volume of gas due to structural changes upon heating in comparison to the
532 counterpart SHSH sample.

533 **3.3.4.3 Comparison of the resulting pore sizes in the catalyst carriers**

534 In terms of catalysis for any reaction, the greater the surface area of the catalyst, the more surface
535 is available for the reactants. It is also necessary to study the particle size of the catalyst support
536 material. In this study the surface area and average pore diameter were measured for SHSH and
537 CHNC, and also the influence of the reduction temperature was also studied (Table 4). The
538 calculated BET surface area indicates the area of catalyst carrier, not strictly the amount of
539 exposed copper available for catalysis. For SHSH, the BET surface area increased upon
540 reduction at a temperature of 250 °C. The average pore diameter of SHSH also decreased by
541 25 % upon reduction at 250 °C. Ideally, the copper particle size should be smaller than the
542 average pore diameter of the catalyst support. This will be discussed further in section 3.3.5.

543 Prior to reduction, CHNC had a comparable BET surface area value to the SHSH sample. The
544 average pore diameter was also similar. Upon reduction of CHNC at 150 °C, the BET surface
545 area increased and the average pore diameter reduced. With a reduction temperature of 200 °C,
546 the surface area increased, as well as the average pore diameter. Upon reduction at 250 °C, the
547 BET surface area and the pore diameter values both reduced. This could have been caused by a
548 discrepancy in the measurement which was discussed in the previous section (3.3.4.2). It could
549 also be explained through the thermal degradation temperature of CHNC; reduction was
550 performed at a temperature above the T_m of 215 °C.

551

552 *Table 4. Sample names and reduction temperatures, BET surface area, average pore width*

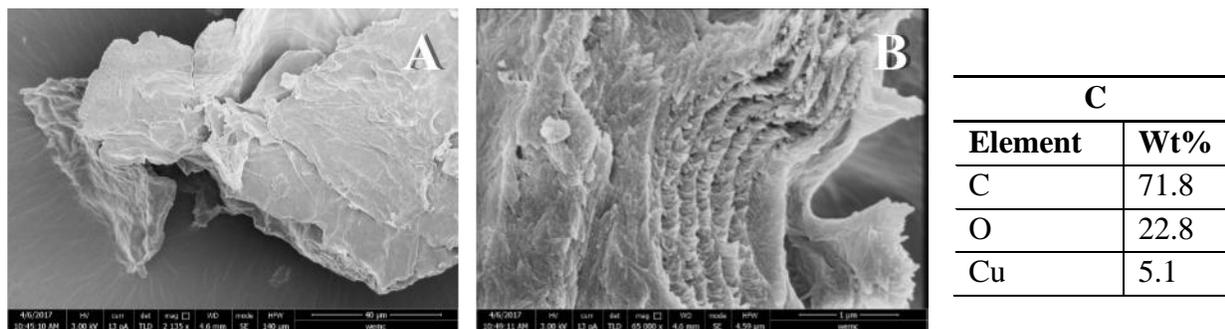
Sample	Reduction temperature (°C)	BET Surface Area (m²/g)	Average pore diameter (nm)
SHSH	-	2.65	12.10
5Cu-SHSH-N2H2	250	7.63	9.03
CHNC	-	2.46	11.57
5Cu-CHNC-N2H2	250	13.40	5.90
5Cu-CHNC-N2H2	200	14.54	8.49
5Cu-CHNC-N2H2	150	11.41	7.17

553

554 **3.3.5 Microscopic investigation**

555 Microscopy gave an insight into the size and how the nanocrystals were orientated, as well as the
556 size of the copper particles on the supporting material. SEM was used to give a visual
557 representation of CHNC after direction reduction (5Cu-CHNC-N₂H₂(200)). Using this
558 technique, it was not possible to see copper particles on the surface of the CHNC. In Figure 21A,
559 a particle was observed. It was not possible to distinguish individual nanocrystals, which could
560 be due to the magnification of the microscope. In Figure 21B, it is possible to see stacked layers
561 and order at a higher level of magnification (1µm) compared to Figure 21A (40µm). On Figure
562 21B, an EDX analysis was performed to give information on the elements which were present in
563 the visualised area of the sample (Figure 21C). From this analysis, it was possible to locate
564 copper at a concentration of 5.1 wt% in the visualised area of the sample. Also present was a
565 large concentration of carbon and oxygen, which corresponds to the chitin support material (see
566 Figure 1). Per chitin molecule found within the CHNC image analysed (containing 5 oxygen
567 molecules), there is roughly 1 copper particle present, based on weight (wt).

568



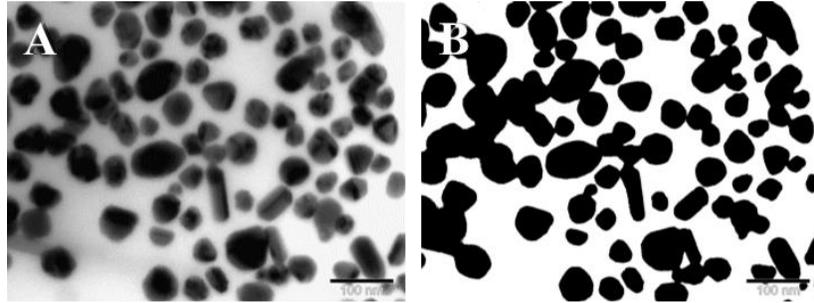
569

570 *Figure 21. A: SEM image of 5Cu-CHNC-N₂H₂(200), B: magnification of SEM image A SEM image. C:EDX spectrum of 5Cu-*
571 *CHNC-N₂H₂(200),*

572 One observation was that samples stored in methanol for a prolonged time began to discolour,
573 indicating that leaching of copper took place. Therefore fresh samples were used for all image
574 analysis.

575 **3.3.5.1 Sizing of copper particles**

576 The size of the copper particles could be measured using TEM micrographs. Particle size
577 measurement was done by a program called Fiji is Just, ImageJ. TEM images show dark
578 irregular and spherical shapes (Figure 22A). Particles were assumed to be overlapping each other,
579 therefore it was difficult to measure each individual particle. The contrast and brightness of the
580 surface area was altered for clear boundary definition for each particle, see Figure 22B.



581
582 *Figure 22. A: TEM micrograph of impregnated chitin nanocrystals (5Cu-CHNC-N2H2(200)), B simplified image of micrograph A*

583 The copper particle diameter (d_{Cu} in nanometres) was calculated by assuming spherical particle
584 shapes by means of the following equation.

585
$$d_{Cu} = 2\sqrt{\frac{a}{\pi}} \quad (6)$$

586 Where:

587 d_{Cu} diameter (nm)

588 a area (dependent of the size of TEM scale bar)

589 A sample of SHSH was also impregnated and reduced at a temperature of 250 °C (5Cu-SHSH-
590 N2H2(250)), the resulting sample was imaged using TEM, see Figure 23. An even dispersion of
591 copper was found throughout the impregnated and reduced sample. The carrier material (SHSH)
592 which supported the copper particles was randomly ordered. ImageJ software was used to
593 individually select every particle and a frequency count is made. These results can be seen in

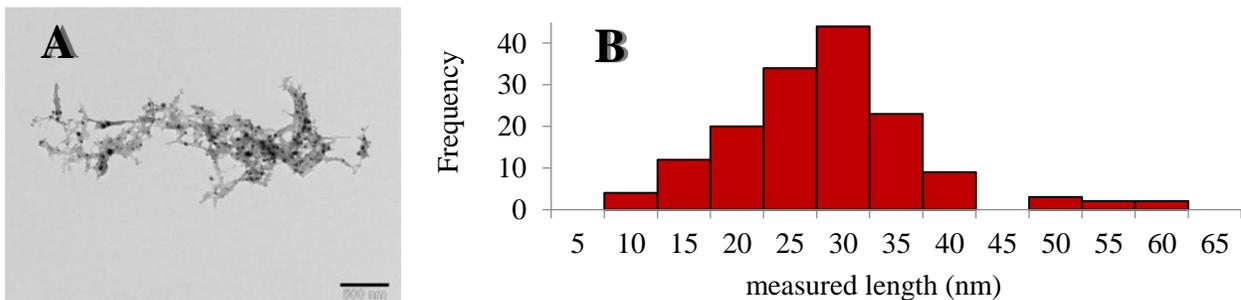


Figure 23. A: TEM image of 5Cu-SHSH-N2H2(250) at 500 nm. B: Histogram of sample count of copper particles for the sample 5Cu-SHSH-N2H2(250).

594 histogram (Figure 23B).

595 The copper particle size distribution on SHSH indicates copper particles ranging from 5 to 65 nm,
596 with an average of 30 nm.

597 The CHNC catalyst support was also studied with TEM to determine the size of the copper
598 particles. The sample was impregnated and reduced at a temperature of 200 °C (5Cu-CHNC-
599 N2H2(200)) (see Figure 24). Please note that the temperature of reduction for CHNC was lower
600 than that used for SHSH (200 vs. 250 °C). A broad dispersion of copper particles were visible
601 throughout the sample looking into different grids. In Figure 24, two distinct areas were observed,
602 each with a different particle size distribution of copper. A large cluster of smaller copper
603 particles are visible (Figure 24A) as well as a small cluster underneath it with larger particles
604 (Figure 24B).

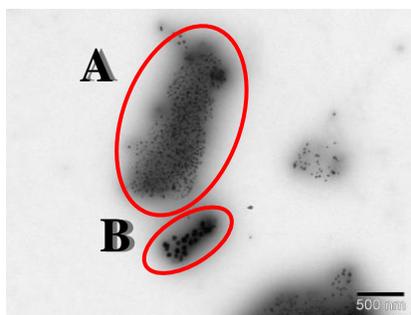


Figure 24. TEM micrograph of sample 5Cu-CHNC-N2H2(200)

605
606

607 Copper particles were measured in the large cluster of smaller particles (Figure 24A). A close-up
608 TEM micrograph of the copper particles was used for the size measurement (Figure 25A).
609 Within this region, an average copper particle size of 15 was observed (Figure 25B).

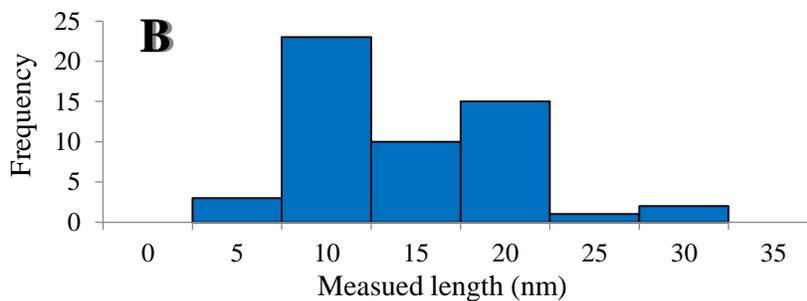
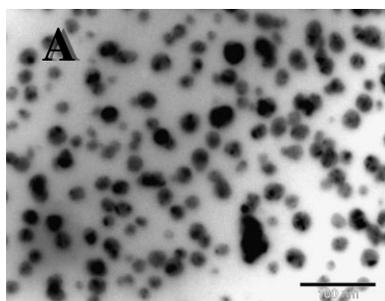


Figure 25. A: TEM image of 5Cu-CHNC-N2H2(200) at 100 nm. B: Histogram of sample count of copper particles for the sample 5Cu-CHNC-N2H2(200).

610

611 Within the smaller region of larger particle (Figure 24B), another copper particle size count was
612 performed using a close-up micrograph (Figure 26A). Within this area of the sample an average
613 copper particle size of 20 nm was measured (Figure 26B).

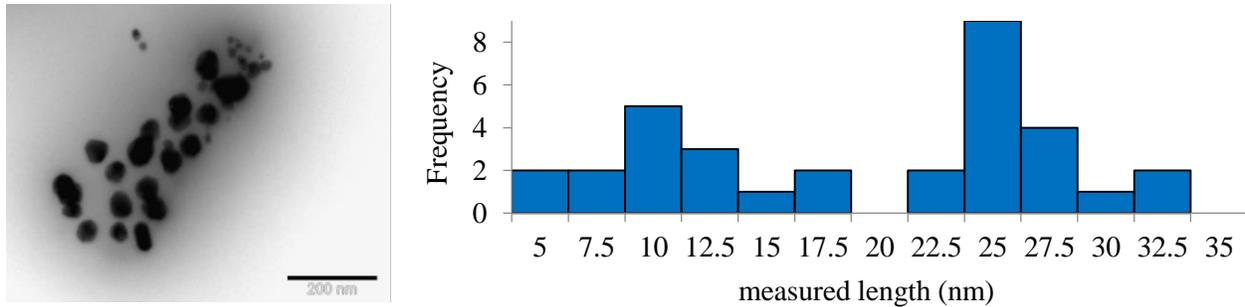


Figure 26. A: TEM image of 5Cu-CHNC-N₂H₂(200) at 200 nm. B: Histogram of sample count of copper particles in the sample 5Cu-CHNC-N₂H₂(200).

614

615 An average particle size of the large and small copper particle distributions for 5CU-CHNC-
616 N₂H₂(200) was calculated to be 20 nm. The results for sizing of the copper particles give an
617 indication of the average size, the micrographs presented here only take into consideration a very
618 small 2D image of the sample. However further analysis is necessary to give a true
619 representation of the size. According to the data presented, the copper particles are on average
620 larger than the average pore size diameters of both catalyst supports (SHSH and CHNC). It can
621 be assumed that some copper particles are small enough to enter the pores of the catalyst support
622 material. From the TEM micrographs, especially on the SHSH support (Figure 25A(SHSH
623 micrograph)), it could be assumed that copper particles are on the surface of the support material,
624 rather than within its pores. Further examination is needed to study how the catalyst carrier
625 performs and if copper is recoverable.

626 **3.4 CATALYSING THE HYDROGENATION OF GLUCOSE AND FRUCTOSE TO**
627 **SORBITOL – SUCCESS OF THE CATALYST CARRIERS**

628 In order to examine the success of the catalyst support material, a catalysed reaction needs to be
629 carried out with CHNC and SHSH materials. In this study, the hydrogenation of glucose to form
630 sorbitol was performed in the laboratory at 25 °C as an initial assessment of 5Cu-SHSH-
631 N2H2(250). High performance liquid chromatography (HPLC) was used to analyse the amount
632 of fructose and sorbitol formed during the reaction. A sample of the reaction liquid was taken
633 every 15 min for 4 h. HPLC data showed no decrease in glucose, nor production of fructose or
634 sorbitol peaks in any of the samples. Reasons why the reaction did not proceed could be linked to
635 the experimental set-up, including an incorrect temperature of the vessel, the concentration of
636 catalyst or reaction time. Also, the analytical method for measuring the levels of glucose,
637 fructose and sorbitol could also affect the data produced. Prior to analysis, there were issues with
638 the HPLC column, which may have influenced the results of the reaction. Please see section 5 for
639 more details of recommendations for future work.

640 4 CONCLUSIONS

641 The aim of this study was to investigate the potential of chitin nanocrystals (CHNC) and shrimp
642 shells (SHSH) as a catalyst support for metallic copper. A method was developed in this study to
643 impregnate, to dry and to calcinate/reduce the support material containing the copper catalyst.
644 Differences between CHNC and SHSH have been investigated and a comparison of their
645 physiochemical characteristics has been performed using techniques such as FTIR, XRD and
646 TGA.

647 Conditions for the impregnation, drying and calcination/reduction of the catalyst carrier materials
648 were studied and the most favourable were further investigated. It was possible to disperse small
649 copper particles ranging in length between 15-30 nm with chitin (both SHSH and CHNC) as a
650 catalyst support material by means of incipient wetness impregnation. Initially drying of the
651 impregnated sample was carried out at 120 °C, however this resulted in the production of
652 undesirable copper oxides. Drying the impregnated sample at 60 °C showed no copper oxide
653 formations. To obtain metallic copper, it was necessary to perform the calcination/reduction at a
654 temperature of 200 °C. Direct or indirect reduction and calcination were studied for SHSH, and it
655 was concluded that direct reduction (N_2/H_2) was more favourable due to chitin crystallinity and
656 the production of metallic copper.

657 Once the method for production of the catalyst carrier was developed, it was possible to
658 determine which carrier material was more suitable, SHSH or CHNC. Based on thermal analysis,
659 SHSH seems more favourable due to a higher decomposition temperature. By studying the
660 crystallinity of the catalyst support materials, CHNC was more favourable due to the presence of
661 crystalline chitin and metallic copper. The size of the copper particles on both of the catalyst
662 support materials was comparable, both materials contained small evenly dispersed copper
663 particles. Based on the physisorption analysis, the catalyst support materials have comparable
664 surface areas and average pore diameters. However after impregnation and direct reduction,
665 including thermal treatment, CHNC was more favourable due to a larger surface area and a
666 smaller average pore diameter.

667 Considering all of the analysis performed within this study, it is concluded that CHNC is a
668 suitable catalyst support material. The most promising conditions for catalyst support production
669 involved direct reduction using N_2 and H_2 gas at a temperature of 200 °C (5Cu-CHNC-
670 $N_2H_2(200)$). Further work is necessary to study the benefits of using a CHNC support material,
671 and to confirm the findings presented here.

672 **5 FUTURE WORK**

673 Future studies are recommended to further explore and to completely understand how the
674 catalyst support materials behave during the catalysed reaction. Many methods of catalyst carrier
675 production and analysis have been performed, however further clarification is necessary. During
676 this study, it was not possible to explore all interesting paths, due to time restrictions and
677 equipment failure. In this section, some ideas for continuation of the project are given.

678 **5.1 CATALYST SUPPORT PREPARATION**

679 The chitin from shrimp shells obtained from Sigma-Aldrich does not indicate the amount of
680 acetyl groups present or any residual proteins which could influence the pore size and
681 interactivity with the copper. With the knowledge obtained in this thesis, it is possible to
682 eliminate some steps from the procedure described in 3.1. The protonation step (addition of
683 KOH) could be discarded from the nanocrystal preparation as it is believed to have already been
684 carried out by the manufacturer.

685 Within this thesis, a solution of 5 wt% copper solution was used to impregnate the support
686 material, as this was considered to be suitable based on the literature. Different loading
687 percentages could be studied further as increasing the load of catalyst could improve the catalysis
688 yield and reaction time.

689 Another interesting aspect would be the effect of impregnation at different pH values. The
690 isoelectric point of a substance determines its orientation within aqueous conditions. In this work,
691 the chitin nanocrystals were freeze dried after preparation at a pH value of 3. In future work, the
692 effects of pH at the stage of freeze drying on the resulting CHNC sample should be studied
693 further. This could influence the average pore diameter of the support material, as well as the pH
694 of the catalyst solution chosen for impregnation of the carrier.

695 Comparable techniques as incipient wet impregnation, e.g. adsorption, wet impregnation,
696 homogeneous deposition precipitation can be done to investigate a faster/cheaper/higher yielding
697 way to impregnate a sample.

698 **5.2 ANALYTICAL TECHNIQUES TO BE CONSIDERED**

699 In order to measure the length and width of the CHNC, further dilution of the sample was
700 necessary to produce a clear TEM image. In this thesis, a CHNC concentration of 0.025 wt%
701 was initially used and resulted in a dense nanocrystal dispersion. As a starting point for future
702 work, a concentration of 0.01 wt% is suggested.

703 One technique that was not available during this project was chemisorption because the required
704 gas combination was not available. With this technique, it is possible to measure the strength of
705 the interaction between the copper catalyst and the support material. This technique can also be
706 used to measure the (copper) catalyst characteristics e.g. leaching capabilities of the catalyst.

707 X ray analysis , curve deconvolution of data, diffraction data smoothed by Savitsky-Golay filter
708 with the use of a second-order polynomial regression and fit with Gaussian-Lorentzian like shape
709 peak fit (Goodrich & Winter, 2007). With this data analysis, it is possible to determine the Miller
710 indices (hkl values), which provides data on the internal planes within a crystal lattice.

711 **5.3 PERFORMING CATALYSED REACTION**

712 In order to assess how the catalyst support material actually performs, it is necessary to carry out
713 the catalysed reaction. Due to the large volume of chitin which is readily available for
714 valorisation from shrimp shell waste, a number of different catalysts could be supported. Future
715 work with copper reactions should be carried out and compared to other catalysts and support
716 materials to determine the most beneficial conversion yields. Other catalysts could also be
717 explored using a chitin based support material.

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