

512-A / 1989 04

# Plant Resources of South-East Asia

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No 12(1)

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Medicinal and poisonous plants 1

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L.S. de Padua, N. Bunyaphatsara and  
R.H.M.J. Lemmens (Editors)

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Backhuys Publishers, Leiden 1999

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ISBN 90-5782-042-0  
NUGI 835  
Design: Frits Stoepman bNO.

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Printed in the Netherlands by Veenman drukkers, Ede.  
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# Contents

Editors and contributors 11

Prosea Board of Trustees and Personnel 17

Foreword 19

1 Introduction 21

1.1 Definitions 21

1.1.1 *Systems of medicine* 21

1.1.2 *The importance of medicinal plants* 21

1.1.3 *Aromatic plants* 22

1.1.4 *Poisonous plants used as medicine and pesticide* 22

1.2 How the medicinal and poisonous plants have been grouped 23

1.3 Role of medicinal and poisonous plants 24

1.3.1 *Traditional and modern medicine in South-East Asia* 24

1.3.2 *Production and trade* 32

1.4 Phytochemistry 32

1.4.1 *Carbohydrates* 33

1.4.2 *Lipids* 34

1.4.2.1 *Vegetable oils* 34

1.4.2.2 *Acetogenins* 34

1.4.3 *Amino acids and their derivatives* 34

1.4.3.1 *Amino acids* 34

1.4.3.2 *Cyanogenic glycosides* 35

1.4.3.3 *Sulphur-containing compounds* 35

1.4.3.4 *Lectins* 35

1.4.3.5 *Enzymes* 36

1.4.4 *Alkaloids* 36

1.4.5 *Phenols and phenolic glycosides* 36

1.4.5.1 *Simple phenolic compounds* 38

1.4.5.2 *Tannins* 38

1.4.5.3 *Coumarins and their glycosides* 40

1.4.5.4 *Quinones* 40

1.4.5.5 *Flavonoids* 41

1.4.5.6 *Anthocyanins* 42

1.4.5.7 *Phloroglucinols* 43

1.4.5.8 *Lignans and related compounds* 43

- 1.4.6 *Terpenoids and steroids* 43
  - 1.4.6.1 *Monoterpenes* 43
  - 1.4.6.2 *Sesquiterpenes* 45
  - 1.4.6.3 *Diterpenes* 45
  - 1.4.6.4 *Triterpenes and steroids* 46
  - 1.4.6.5 *Carotenoids* 48
- 1.5 *Biological and pharmacological activity and therapeutical applications* 48
  - 1.5.1 *Factors affecting biological activity* 48
    - 1.5.1.1 *Physicochemical properties* 49
    - 1.5.1.2 *Chemical parameters* 49
    - 1.5.1.3 *Spatial considerations* 50
  - 1.5.2 *Bio-assaying* 51
    - 1.5.2.1 *Requirements for screening medicinal plant material* 51
    - 1.5.2.2 *Common pharmacological screening methods* 53
  - 1.5.3 *Surveys of bioactivity, pharmacological and therapeutic categories* 54
  - 1.5.4 *Future developments in research on bioactivity* 54
- 1.6 *Botany* 55
  - 1.6.1 *Plants used in medicine* 55
  - 1.6.2 *Weedy and forest species* 56
  - 1.6.3 *Chemotaxonomy* 56
- 1.7 *Eology* 57
- 1.8 *Agronomy* 58
  - 1.8.1 *Production systems* 58
    - 1.8.1.1 *Collection of medicinal plants from the wild* 58
    - 1.8.1.2 *Cultivation of medicinal and poisonous plants* 58
  - 1.8.2 *Propagation* 59
  - 1.8.3 *Husbandry* 59
- 1.9 *Harvesting and handling after harvest* 60
  - 1.9.1 *Harvesting* 60
  - 1.9.2 *Drying and cleaning* 60
  - 1.9.3 *Storage conditions* 61
- 1.10 *Processing, utilization and quality control* 62
  - 1.10.1 *Extraction methods* 62
  - 1.10.2 *New industrial standards* 63
  - 1.10.3 *Household preparation* 64
    - 1.11 *Genetic resources and breeding* 64
      - 1.11.1 *Plant diversity and conservation* 65
      - 1.11.2 *Breeding* 65
    - 1.12 *Research and development* 66
      - 1.12.1 *Main research topics* 66
      - 1.12.2 *Main institutions* 66
  - 1.13 *From plant to drug* 68
  - 1.14 *Prospects* 69

## 2 Alphabetical treatment of genera and species 71

<i>Abrus</i>	: Indian liquorice 73
<i>Achillea millefolium</i>	: yarrow 77
<i>Acorus calamus</i>	: sweet flag 81
<i>Aerva</i>	: aerva 86
<i>Ageratum</i>	: goatweed 88
<i>Allium</i>	: onion 93
<i>Aloe</i>	: aloe 100
<i>Alternanthera</i>	: alternanthera 105
<i>Amaranthus spinosus</i>	: spiny amaranth 110
<i>Amomum</i>	: amomum 113
<i>Andrographis paniculata</i>	: creat 119
<i>Angelica acutiloba</i>	: tang kui 123
<i>Antiaris toxicaria</i>	: upas tree 126
<i>Arcangelisia flava</i>	: yellow-fruited moonseed 129
<i>Aristolochia</i>	: birthwort 133
<i>Artemisia</i>	: wormwood, mugwort 139
<i>Belamcanda chinensis</i>	: blackberry lily 148
<i>Bidens</i>	: beggar-tick, bur-marigold 150
<i>Blumea</i>	: ngai camphor plant 155
<i>Brucea javanica</i>	: kuwalot 160
<i>Bryophyllum</i>	: life plant 163
<i>Cannabis sativa</i>	: hemp 167
<i>Cardiospermum</i>	
<i>halicacabum</i>	: balloon vine 176
<i>Carmona retusa</i>	: kinangan 178
<i>Cassia</i>	: golden shower, horse cassia 181
<i>Catharanthus roseus</i>	: Madagascar periwinkle 185
<i>Centella asiatica</i>	: Asiatic pennywort 190
<i>Chenopodium ambrosioides</i>	: wormseed 194
<i>Cinchona</i>	: cinchona, quinine 198
<i>Cissampelos pareira</i>	: sansau 205
<i>Curculigo orchioides</i>	: taloangi 207
<i>Curcuma</i>	: curcuma, turmeric 210
<i>Cyclea</i>	: cyclea 219
<i>Cyperus</i>	: nut sedge 222
<i>Datura</i>	: thorn apple 229
<i>Derris</i>	: derris 234
<i>Desmodium</i>	: tick clovers 242
<i>Elephantopus</i>	: elephant's foot 250
<i>Embelia</i>	: embelia 254
<i>Erythroxylum</i>	: coca 258
<i>Euphorbia</i>	: spurge 263
<i>Eurycoma</i>	: eurycoma 272
<i>Fatoua villosa</i>	: sikir 275
<i>Ficus</i>	: ficus, fig 277
<i>Gloriosa superba</i>	: flame lily 289
<i>Heliotropium</i>	: heliotrope 292

<i>Holarrhena</i>	: holarrhena 296
<i>Hydnocarpus</i>	: sausage tree 299
<i>Hypericum</i>	: St John's wort 303
<i>Imperata</i>	: cogon grass 307
<i>Ixora</i>	: ixora 311
<i>Jasminum</i>	: jasmine 315
<i>Jatropha</i>	: jatropha, physic nut 320
<i>Justicia</i>	: justicia 327
<i>Kaempferia</i>	: kaempferia, galangal 331
<i>Kalanchoe</i>	: kalanchoe 335
<i>Lantana</i>	: sage 338
<i>Melochia corchorifolia</i>	: chocolate weed 342
<i>Mentha arvensis</i>	: mint 344
<i>Mimosa pudica</i>	: sensitive plant 349
<i>Momordica</i>	: momordica 353
<i>Morus</i>	: mulberry 359
<i>Oldenlandia</i>	: oldenlandia 364
<i>Orthosiphon aristatus</i>	: Java tea 368
<i>Oxalis corniculata</i>	: Indian sorrel 371
<i>Papaver</i>	: poppy 373
<i>Peperomia pellucida</i>	: ketumpangan air 379
<i>Phyllanthus</i>	: phyllanthus 381
<i>Phytolacca</i>	: pokeweed 392
<i>Plantago</i>	: plantain, psyllium 397
<i>Plectranthus</i>	: country borage, painted nettle 403
<i>Plumbago</i>	: leadwort 409
<i>Premna</i>	: premna 413
<i>Pueraria montana</i>	: kudzu 417
<i>Quisqualis</i>	: Burma creeper 421
<i>Rauwolfia</i>	: snakewood 424
<i>Rhinacanthus nasutus</i>	: tarebak 431
<i>Schefflera</i>	: schefflera 433
<i>Scutellaria</i>	: skullcap 438
<i>Senna</i>	: ringworm bush, foetid cassia 442
<i>Smilax</i>	: sarsaparilla, China root 447
<i>Solanum</i>	: solanum, nightshade 453
<i>Sophora tomentosa</i>	: seacoast laburnum 460
<i>Stephania</i>	: stephania 463
<i>Strychnos</i>	: Saint Ignatius bean, nux-vomica tree 467
<i>Styphnolobium japonicum</i>	: Japanese pagoda tree 472
<i>Taraxacum officinale</i>	: dandelion 475
<i>Tinospora</i>	: tinospora 479
<i>Trichosanthes</i>	: snake gourd 484
<i>Verbena officinalis</i>	: vervain 491
<i>Vernonia</i>	: vernonia 493
<i>Vitex</i>	: vitex 497

Literature 503

Acknowledgments 608

Acronyms of organizations 610

Glossary 611

Sources of illustrations 647

Index of compounds 658

Index of pharmaceutical terms 663

Index of scientific plant names 673

Index of vernacular plant names 691

The Prosea Foundation 707

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  - Anas Subarnas, Department of Pharmacy, Faculty of Mathematics and Natural Sciences, Padjadjaran University, Sumedang 45363, Indonesia (*Cannabis sativa*)
  - Diah Sulistiarini, Puslitbang Biologi LIPI, Jl. Ir. H. Juanda 22, Bogor 16122, Indonesia (*Allium*)
  - S. Susiarti, Puslitbang Biologi LIPI, Jl. Ir. H. Juanda 22, Bogor 16122, Indonesia (*Jatropha*)
  - H. Sutarno, PROSEA Indonesia, Herbarium Bogoriense, Jl. Ir. H. Juanda 22, Bogor 16122, Indonesia (*Catharanthus roseus*)
  - Stephen P. Teo, Sarawak Herbarium, Sarawak Forestry Department, Badrudin Road, 93660 Kuching, Sarawak, Malaysia (*Smilax*)
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## Foreword

Although many people in rural areas in the tropics – including South-East Asia – rely on locally available plants for their daily health care, the importance of these plants has, until recently, been underestimated. This has now changed, partly because of the demand for plant-derived medicines, and partly because it is now known that medicinal and poisonous plants are a rich source of promising chemical compounds. The resulting resurgence of interest in medicinal plants in South-East Asia, particularly in Thailand and the Philippines, is also fuelled by interest in environmentally friendly plant-derived pesticides. This renaissance of medicinal and poisonous plants has created a demand for detailed knowledge from new research and for an overview of existing published information. As interesting compounds are more likely to be found in plants with known medicinal uses, the best way to find new applications of plant-derived drugs would seem to be to combine local knowledge with the results of modern research on the properties of plant-derived medicines. It is here that this Prosea handbook volume on medicinal and poisonous plants is so valuable. It provides the latest information on the botanical, agricultural, chemical and medicinal aspects of these plants and, as such, will be an invaluable resource for those involved with medicinal and poisonous plants, whether working in research and industry or in education and extension.

Bangkok, June 1999

Professor Dr. Her Royal Highness Princess Chulabhorn Mahidol  
President  
The Chulabhorn Research Institute, Bangkok, Thailand

# 1 Introduction

## 1.1 Definitions

True to its title *Prosea 12* is devoted to medicinal and poisonous plants used in South-East Asia. Following common practice in literature, the two categories of plants have been combined into one commodity group. This is because many poisonous plants are used medicinally; at lower doses their toxic constituents are often beneficial.

### *1.1.1 Systems of medicine*

It is important to distinguish between three different types of medicine: traditional, herbal and pharmaceutical. A plant may be consumed as a medicinal tea by members of a community living in the area where the plant is indigenous, the same plant may be cultivated and processed in the country of origin into a formulation of a herbal medicine sold in western countries, and it may provide a lead compound for a pharmaceutical product. These systems of medicine are complementary in health care and can in no way substitute one another (Balick et al., 1996). In this volume, the role of the species in each of the three systems is distinguished whenever possible.

### *1.1.2 The importance of medicinal plants*

Medicinal plants are of great economic importance. They are used as raw materials for the extraction of active constituents in pure form (e.g. quinine and quinidine from *Cinchona* bark), as precursors for synthetic vitamins (e.g. fixed oil for vitamin E) and steroids (e.g. *Dioscorea* and *Smilax* roots), and as preparations for herbal and indigenous medicines. Plants are not only the major source of energy-rich foods in most societies, but are also an indispensable source of vitamins and other substances promoting healthy growth. But note that though the consumption of certain plant parts may be prophylactic, e.g. scurvy is prevented by eating citrus fruits (in which vitamin C is the active factor), plant species with these properties are generally treated in other *Prosea* volumes, particularly *Prosea 2*: 'Edible fruits and nuts' and *Prosea 8*: 'Vegetables'.

The present volume covers only those species of plants whose medicinal uses and properties are described in the literature. The discovery of important medicinal properties (e.g. oncolytic properties of the alkaloids in *Catharanthus roseus* (L.) G. Don) in the last 40 years has resulted in thousands of scientific reports on South-East Asian medicinal plants. The few existing handbooks on medicinal plants of South-East Asia (e.g. Perry, 1980) only give information on



the plants' uses and properties, but the team of authors and editors responsible for this volume has combined the information in the reports and manuals with botanical and agronomic information.

Many plants used in medicine or as poison have other uses. In this volume, however, generally only those species with primarily medicinal or poisonous functions are dealt with.

This compilation of the published data will be especially useful to researchers developing new drugs. It is more efficient to screen plants used in traditional medicine, as these yield a much higher output of interesting substances than plants sampled at random. This is why many pharmaceutical companies incorporate ethnobotanical information into their research and development programmes. Much knowledge still only exists as oral tradition and many species still need to be investigated to find out their constituents and biological effects. Numerous plant species are used in traditional veterinary medicine. Most small farmers in South-East Asia rely on herbal medicines to treat their sick animals, since these are easily available and affordable. Written information on veterinary uses is scarce; most information is still transmitted orally.

### *1.1.3 Aromatic plants*

Many kinds of aromatic plants are traditionally used medicinally, usually prepared as teas. The volatile components of essential oils are often present in the preparations and these are often thought to be responsible for the biological activity.

It was sometimes difficult to decide which species should be covered in Prosea 12: 'Medicinal and poisonous plants', Prosea 19: 'Essential-oil plants', and Prosea 13: 'Spices' (the latter includes numerous aromatic plants). Wherever possible it is tried to avoid an overlap in the species treated in these 3 volumes, with the result that the final choice is somewhat subjective. Furthermore, though lower plants (fungi, algae, mosses, lichens and ferns) are sometimes also used medicinally, they have not been included in this volume, even when they have no other use; instead, they appear in Prosea 15: 'Cryptogams'. Several plant species used medicinally also produce dyes or tannins, and are treated in Prosea 3: 'Dye and tannin-producing plants'.

### *1.1.4 Poisonous plants used as medicine and pesticide*

Only those poisonous plants used medicinally or as pesticides are dealt with in this volume. The pests that plant extracts can be used against are rodents, birds, insects, molluscs, nematodes, fungi, bacteria, algae, viruses and weeds. The preparations used to protect against these pests are known respectively as rodenticides, avicides, insecticides, molluscicides, nematocides, fungicides, bactericides, algacides, virucides and herbicides. Though traditional applications of pesticides of plant origin were gradually ousted by chemical pesticides, there has recently been a resurgence of interest in plant-based pesticides. This is partly because chemical pesticides have been found to have disadvantages, including being health hazards to farmers and consumers. Furthermore, there is concern about the accumulation of residues in soil, groundwater and animals, and the build-up of resistance in pests. In contrast, plant-based pesticides are

usually not accumulative, so are environmentally benign. Moreover, they are often highly selective, their toxicity to non-target mammals is usually low, and pests do not appear to develop resistance to them because these pesticides contain many active ingredients, even when derived from a single plant source (Chomchalow in Chomchalow & Henle, 1993). Examples of plant species used as a source of plant-based pesticides are neem (*Azadirachta indica* A.H.L. Juss.), derris (*Derris elliptica* (Wallich) Benth.), turmeric (*Curcuma longa* L.) and citronella (*Cymbopogon nardus* Rendle). However, because plant-based pesticides are so biodegradable they are rather unstable, which means that they have a short shelf-life, must be used soon after preparation, and have low persistence after application. Many plant-based pesticides are made domestically and more cheaply than chemical pesticides. Since only crude extracts are used to make them, such pesticides are definitely much less effective than chemical pesticides that have been formulated and purified to a high concentration.

## 1.2 How the medicinal and poisonous plants have been grouped

In 1996 a Prosea Task Force on Medicinal and Poisonous Plants was appointed to delineate the large commodity group and to propose how this group could best be dealt with in the Prosea handbook. The Task Force, which consisted of L.S. de Padua (University of the Philippines, chairwoman), R.H.M.J. Lemmens (Prosea Publication Office, the Netherlands, secretary), N. Wulijarni-Soetjipto (Prosea Network Office, Indonesia), N. Bunyapraphatsara (Mahidol University, Thailand), A.M. Latiff (Universiti Kebangsaan, Malaysia), Nguyen Tien Ban (Institute of Ecology and Biological Resources, Vietnam), Sjamsul Arifin Achmad (Institut Teknologi Bandung, Indonesia), R.P. Labadie (Utrecht University, the Netherlands) and D.K. Holdsworth (Norwich, United Kingdom) presented its report in March 1996.

It proposed that Prosea 12: 'Medicinal and poisonous plants' should consist of three parts, published separately. The three parts would essentially reflect the importance accorded to the species: the most important species would be treated in part 1, the least important ones in part 3. In spite of its shortcomings, this approach has important advantages over alphabetical treatment: it enables important and well-known medicinal and poisonous plants to be dealt with in greater detail than unimportant and lesser-known ones, and allows for any omissions from the first two parts to be made good in the third part. Preliminary lists for the 3 parts were drawn up more or less subjectively, based on existing handbooks on medicinal or useful plants for South-East Asia like Burkill (1966), Dharma (1981), Heyne (1927), Holdsworth (1977), Nguyen Van Duong (1993) and Quisumbing (1978). The list was finalized after being critically reviewed by the members of the Task Force.

The plants in Prosea 12: 'Medicinal and poisonous plants' are dealt with primarily by genus rather than by species. This is because the properties of different species within one genus and hence their uses are often similar. Furthermore, the genus approach reduces the large commodity group to manageable proportions. After each genus description, selected species are described briefly. If only one species of a genus is important in South-East Asia, however, it is dealt with as a species.

### 1.3 Role of medicinal and poisonous plants

Since time immemorial people have used plants and other materials that were not part of their usual diet to treat illness. They arrived at these treatments by trial and error and accumulated tradition and experience. All cultures have long histories of the use of plants in folk medicine, recorded in ancient herbals from which most of the present-day pharmacopoeias have been derived. Archaeological evidence for the use of herbal remedies goes back 60 000 years: in a Neanderthal cave burial site, excavated in Iraq in 1960, pollen from 8 plant species was found around human bones. These plants were evidently intentionally collected and placed there; 7 of them are medicinal plants still used today.

In the last 30 years there has been a resurgence of interest in the use of plants as medicines. However, although the world population is increasing and plant-rich habitats such as the tropical forests are dwindling steadily (Latiff, 1991) there appears to be no concerted effort in research and conservation. The renewed interest in medicinal plants is also clearly noticeable in South-East Asia and has resulted in research and development programmes in several countries, and also in joint efforts such as the Asian Network on Medicinal and Aromatic Plants (ANMAP).

Forests have long been regarded primarily as a source of timber, but now the value of non-wood forest products is becoming increasingly appreciated. Medicinal plants are important non-wood forest products and should therefore be a priority in forest protection measures. It is therefore gratifying that biodiversity prospecting and its policy implications for medicinal plants are now recognized as an important issue in conservation.

#### 1.3.1 Traditional and modern medicine in South-East Asia

Below, the history, status and role of medicinal plants is described briefly for each country in South-East Asia; likely future developments are also noted. For practical reasons (the cross-border distribution of ethnic groups) the islands of Borneo and New Guinea are treated separately instead of under Malaysia, Brunei, Indonesia and Papua New Guinea.

##### *Indonesia*

The traditional use of plants for healing in Indonesia dates back to prehistoric times. The art and knowledge of the uses of plants as medicine have been handed down orally from generation to generation. Some plants still used in traditional medicine can be found depicted in reliefs on the walls of ancient temples in Java, such as those of Borobudur, Prambanan, Penataran and Sukuh. They include *Aegle marmelos* (L.) Correa, *Antidesma bunius* (L.) Sprengel, *Borassus flabellifer* L., *Calophyllum inophyllum* L., *Datura metel* L. and *Syzygium cumini* (L.) Skeels.

The earliest written references to the local uses of plants in Indonesia are in the early 16th Century reports of Portuguese explorers. The first endeavour to gather data on Java's medicinal plants was by Bontius (1658). His work includes some 60 plates of plants with descriptions of their healing powers and uses. Rumphius's work ('Herbarium Amboinense', 1741-1755), a special study

of the flora of Ambon (Moluccas) was more important. It describes hundreds of plants, giving extensive details on their medicinal use and properties. Horsfield (1816) published one of the first monographs on the medicinal plants of Java.

Many publications on medicinal plants appeared in the 19th and early 20th Century. Of these, Greshoff's publications (in the period 1890–1914) focused mainly on poisons but also included plants with medicinal properties. Kloppenburg-Versteeg (1907, 1911) wrote books in Dutch, giving hints and advice on using Indonesian plants. The second edition of Heyne's book on the useful plants of Indonesia (1927) gave extensive information on the medicinal uses. Since then, numerous papers and books have been published on the medicinal plants of parts and islands of Indonesia, but these usually either relate to one or a few species, or summarize recorded traditional uses in a confined region (e.g. Avé & Sunito (1990) for Siberut and Bell & van Houten (1993) for Central Seram). There have also been some books dealing summarily with larger numbers of medicinal plants from Indonesia, e.g. Kasahara & Hemmi (1995) and Syamsuhidayat & Hutapea (1991).

In rural areas, 'dukuns', i.e. persons with putative expertise in medical matters and who use medicinal plants in their preparations, still play an important role in primary health care. So-called 'jamus' – complex mixtures of herbs – are still widely and commonly used in Java. The ingredients are well pounded and mixed, and steeped in hot water. Alternatively, they may be dried, and then boiled when required, to yield a decoction for use. Jamus may be preserved in powder form, after drying over heat in an iron pan. Most jamus have a long history of traditional use and some have been tested empirically and shown to be effective. However, they are often not used as medicine for a given disease but to keep the body healthy, in a holistic approach. Sometimes they are used for cosmetic purposes. It is often recommended to take jamus regularly. The composition of a jamu used to treat a certain disease and having a certain standard name may vary, depending on the custom or view of the person preparing the mixture. Jamus are in general prepared and traded by women from Central Java, whose good healthy complexions advertise the efficacy of their products. Some jamus are given to livestock. It is estimated that 1000–1300 plant species are used in the preparation of jamus. Most of these are collected from the wild. Manufactured products are also widely available over the counter throughout Indonesia. Jamus produced industrially have also been exported from Indonesia for a number of years in the form of powders and tablets. The knowledge on medicinal plants and jamus has been kept in families in the form of hand-written records. The original manuscript on Javanese traditional medicine, called 'serat kawruh bab jampi-jampi Jawi' and written around 1831 was kept in the library of the Surakarta Palace. It contains 1166 prescriptions, 922 of which are jamu preparations.

Research on medicinal plants has been conducted in Indonesia for more than 50 years. The studies have included the collection of samples, the inventory of genetic resources, ethnobotany, biotechnology, agronomy, chemical properties, pharmacological and toxicological screenings, product standardization, formulation and plant conservation. Several institutes working on medicinal plants have been established since 1950, as well as working groups and committees. Numerous scientific meetings have been organized on the subject.

Recent developments point to an increased interest in medicinal plants. A national working group on medicinal plants was established in 1990 as a follow-up to a national seminar. To promote the development and socialization of the use of medicinal plants, the Ministry of Health has issued lists of recommended medicinal species to be planted in family gardens. Gardens with medicinal plants have been established throughout the country. Various institutions have made a germplasm inventory of medicinal and aromatic plants (Wahid in Chomchalow & Henle, 1993).

### *Peninsular Malaysia*

Traditional medicine has been important to Malaysians of all ethnic groups for centuries. The influence of the cultures of China, India and Java is strong. For instance, almost all the medicinal products sold in the Chinese community are imported from China, and the influence of Javanese medicine is still important among the local Javanese communities in Selangor. The classic work by Burkill (1935) is still the standard reference for traditional medicine.

Much of the knowledge on traditional medicine still dominant in the culture of various ethnic groups is unrecorded, and handed down from one generation to the next. The practitioners of traditional Malay medicine have vast knowledge about the identification and classification of plants, folk nomenclature and, above all, the medicinal properties. This knowledge has not yet been tapped systematically to develop medicines based on traditional remedies (Latiff, 1991). However, since 1981 researchers at the University Kebangsaan Malaysia have conducted many multidisciplinary projects on medicinal plants. Although the cultivation of medicinal plants was advocated to ensure a continuous and reliable supply of products for local consumption or export, no large companies have shown interest; small-scale cultivation for local consumption or sale has started, however (Latiff, 1991).

The approaches summarized by Latiff (1991) as being vital for future research in traditional medicine in Malaysia remain valid today. They are:

- An inventory and therapeutic classification of the medicinal plants used.
- The development of scientific criteria and methods for assessing the safety of medicinal plant products and their efficacy in the treatment of diseases.
- The introduction of national standards and specifications for identity, purity, strength and manufacturing practices.
- The designation of research and training centres for the study of medicinal plants.

### *Borneo*

The original inhabitants of Borneo were probably the forest dwelling Punan people. Other groups, such as the Iban, immigrated over the centuries. In more recent history, Malay people have immigrated to coastal areas, followed by Chinese settlers later. The traditional medicine of these peoples of Borneo reflects their origins.

Many Chinese inhabitants of Borneo use imported dried herbs packaged in China, which have often been recorded as Chinese herbal medicine for several thousand years. These are supplemented to some extent by local plants. There

are Chinese medicine shops in most coastal towns in Borneo. Malays recognize the similarities of the flora of Borneo and Peninsular Malaysia. Malay traditional medicine has only been recorded in this century.

Many of the non-Chinese people of Borneo have a tradition of a 'bomor' or 'shaman': persons who have accumulated the medicinal lore of their people, who are regarded as healers. These persons may still use incantations, invoke animistic spirits in trances and be well versed in local superstitions and native psychology. Above all, they use plants either from the rain forest or cultivated in their garden. In contrast to Chinese herbal medicine the plants used are invariably collected fresh and used externally or internally as decoctions. Though Malay medicine is similar in origin, it has been modified by the influence and teachings of Islam.

Until very recently, there have been few studies of the traditional medicine of the indigenous tribal peoples of Borneo. This still represents a challenge to botanists, pharmacologists, pharmacognosists, anthropologists and phytochemists. There were some studies of traditional medicine done by Dutch explorers when Indonesia was still a Dutch colony, but these were almost exclusively confined to the economically more important islands of Java and Sumatra. The difficulties of learning the languages of the people of Borneo contributed to the general lack of scientific studies. In the early 1990s, some inventories and bio-assay screening of medicinal plants used by the Kenyah Dayak people, under the auspices of World Wide Fund for Nature, resulted in a publication (Leaman et al., 1991).

The same might be said of British explorers in the former British colonies of North Borneo (present-day Sabah) and Sarawak. Traditional medicine was of little interest when western drugs and quinine could be imported. The establishment of the Sarawak Museum in Kuching led to an interest in ethnic cultures and in the medicinal properties of indigenous plants associated with these cultures. Recently some research has been done by the Sarawak Forest Department (unpublished), and a publication by Ahmad & Holdsworth (1994) is available. For Sabah, there are two recent publications by Ahmad & Holdsworth (1994, 1995). In Brunei some interest in publishing data on medicinal plants began in the early 1990s. There are several recent publications on the medicinal plants of Brunei (Holdsworth, 1991; Mohiddin, Wong Chin & Holdsworth, 1991, 1992). Recent studies by the staff of the Herbarium Bogoriense resulted in seminar reports on medicinal plants of Kalimantan in 1995. The increased cooperation of nations in the area should result in future joint studies of the medicinal plants and plant lore in different areas of Borneo.

### *New Guinea*

There is a rich heritage of traditional knowledge on the use of plants as medicines in New Guinea. The first New Guineans, who possibly arrived from South-East Asia and settled some 60 000 years ago, may have brought medicinal plants with them, or the knowledge of how to use the familiar plants they found on arrival. This would have been followed over thousands of years by systematic trial and error experimentation in the coastal and highland areas. Plants that had effective medicinal properties would be used again, and in this way a local tribal pharmacopoeia would be built up.

A few scattered records of medicinal plants were made by explorers and botanists in Dutch, German and Australian New Guinea at the beginning of the 20th Century. The first collection of some importance was that of Father Futscher, a Catholic priest on New Britain in the 1950s. He collected about 80 plant species used as medicine, noted the local Kuanua language names and tried to identify the plants. The results were published in German in 1959. In the early 1970s many plants from Papua New Guinea were tested in the Chemistry Department of the University of Papua New Guinea (Port Moresby) for the presence of alkaloids. In 1973 it was decided to concentrate on testing traditional medicinal plants. It was shown that these plants – particularly those used internally to treat malaria and fevers – were more likely to contain alkaloids than plants chosen randomly. The medicinal plants collected were identified at the Papua New Guinea National Herbarium at Lae. In over 20 years of fieldwork, Holdsworth collected several thousand specimens of medicinal plants in many different areas of Papua New Guinea; their uses were noted, and the plants were identified, mainly at the PNG National Herbarium in Lae. To date, the survey has yielded over 600 species of medicinal plants. It is interesting to crosscheck the medicinal uses of the plants in different areas of New Guinea (with its numerous languages and consequently little exchange of information between tribal communities) and other tropical countries, by studying the available literature. A first survey was published in 1977 (Holdsworth, 1977), and a more complete overview is in preparation.

In New Guinea, fresh plant material is invariably used for medicinal applications. One plant is often used alone, and sap from its leaves or bark is drunk, or rubbed on the body. Decoctions of bark, twigs or fresh leaves are also drunk. Many traditional medicines have a strong or bitter taste, which suggests that they contain alkaloids, although other substances might account for this taste too. A healer may also choose plants for a treatment based on the Doctrine of Signatures, a practice favoured by many mediaeval physicians.

Traditional beliefs are strong in rural areas throughout New Guinea and healing methods that use medicinal plants are still widely practised. Most tribes have a traditional healer who has been trained by an elder close relative in the uses of certain effective and secret medicinal plants. Often the healer was regarded as a sorcerer and practitioner of traditional medical psychology. Papua New Guinea is currently experiencing rapid changes. Young men and women leave their villages to attend school and to seek employment in towns. They are not available to learn the traditions and medicinal knowledge of their relatives and ancestors, so many traditional customs and practices are disappearing in village communities or are being replaced. The demonstrative effectiveness of a shot of antibiotics and other modern medicines at the first aid post, health clinic or hospital has reduced reliance on plant medicines and is accelerating the disappearance of traditional medical practice in many areas. It is essential to record the traditional uses of medicinal plants before they are lost forever.

### *The Philippines*

Plants have played an important role in traditional medicine in the Philippines since ancient times. When the Spaniards colonized the Philippines in 1521, they were surprised to find many medicinal plants. It is possible that Chinese

traders who came to the Philippines before the Spaniards introduced their herbal medicines and traded them with the Filipinos for other goods. Several manuscripts written during the Spanish regime (1521–1898) survive; well known among these are Father Blanco's 'Flora de Filipinas' (1737), and 'Plantas Medicinales de Filipinas' by Trinidad Pardo de Tavera (1892). Much research on medicinal plants was conducted in the years of American occupation (1898–1935) at the University of the Philippines and at Government Laboratories, now the Department of Science and Technology. Guerrero led these scientific activities, and working with him were scientists such as Merrill, Brown, Elmer, Sulit, Valenzuela, Maranon, Santos, Concepcion and Quisumbing, all well known in the field of medicinal plants. It was during this period that Brown's 'Useful plants of the Philippines' and Quisumbing's 'Medicinal plants of the Philippines' were written, surveys were extended to unexplored regions and chemical analyses and clinical investigations were done. During the Second World War, Filipinos depended entirely on plants as sources of medicines, and came to realize not only that the Philippines abounds with a wide variety of medicinal plants, but also that research on these plants was still much needed.

Medicinal plants are part of the cultural heritage and the 'herbolario' (herb doctor) is a respected member in his community. The 'herbolario' and the 'hilot' (midwife) are traditional practitioners who learn and pass down their craft from generation to generation. They use plants for the treatment of diseases and to relieve pain and physical suffering. Traditional practice was coupled with native beliefs and superstition. The common belief was that disease is due to the presence of evil spirits in the human body which could only be removed by using some bitter-tasting substances, usually derived from plants. Although the practice of the traditional practitioners was empirical in nature, most of the original information on drug-producing plants was derived from them and forms the basis for recent scientific studies and for the present research and development programmes on medicinal plants.

When modern western drugs became available, many Filipinos, especially those in the urban centres, lost touch with their herbal heritage. It became very difficult for herbalists and researchers on medicinal plants to attract interest and support for their work, particularly for the wide acceptance of the use of these plants. The persistence and dedication of Filipino scientists to the efforts of providing adequate health care to poorer sectors of the population through intensive research and dissemination of information have brought about major changes in attitudes and in the health care system. Many western-trained doctors now prescribe herbal medicines for their patients, and medicinal plants continue to provide basic and alternative health care to the Filipino people, particularly in remote areas and islands where the lack of medicines is critically felt. As in most developing countries, herbal medicines still play a major role, with more than 80% of the population using herbal remedies. Dosage forms such as tablets, capsules, syrups, ointments, liniments, tinctures, lozenges, lotions and herbal teas are available, and herbal soaps, shampoos and other body care products are popular. Some of these products are even exported. Remarkable progress and new developments have taken place in recent years, strengthened by the active participation of government and private agencies. Some pharmaceutical companies have expanded into herbal medi-



cines and body care products, and the government has established 4 factories in different regions to manufacture herbal medicines. As a result of intensive research, several medicinal plants are being promoted for use and cultivation in the countryside. Dissemination of information about medicinal plants and herbal products is actively pursued through publications, presentations, seminars, training programmes, exhibitions and educational displays. Many publications are available on Philippine medicinal plants, e.g. 'the National Formulary' by Concha (1978), 'the Guidebook on the proper use of medicinal plants' by Maramba, and the four volumes of 'the Handbook on Philippine medicinal plants' by de Padua, Lugod & Pancho. The population now has access to safe, effective and affordable herbal medicines, and meanwhile there is renewed and accelerating interest in the industrialization and exploitation of Philippine medicinal plants.

### *Thailand*

Medicinal plants have constituted an important part of Thai traditional medicine since that system of medicine, which was adapted from the Ayurvedic system incorporating Thai culture and tradition, was introduced 700–1000 years ago. The earliest knowledge is not well documented, as only a few prescriptions survive. The system of traditional medicine was revised and compiled during the reigns of Kings Rama I and II of Ratanakosin in the 18th and early 19th Century. The prescriptions were recorded on the stone plaques and walls of the Wat Poh temple. Under King Rama V (1868–1910) the royal prescriptions were again compiled, revised and published, and this served as the basis for the Thai traditional medicine of today.

The use of medicinal plants fell sharply when western drugs became available. It is difficult to reverse the decline, because the texts on traditional medicine are too vague. Most of the texts contain the name of the plants, their indications and recipes, but lack detailed information on the preparation. This is probably because traditional doctors jealously guarded their knowledge. Most of the recipes are composed of many ingredients, and each ingredient is added for a specific purpose. At present, most doctors are trained in western medicine and the health care system relies almost totally on imported western drugs. This leads to the following problems:

- A large amount of foreign exchange is lost.
- The drugs are too expensive for people with low incomes.
- The ease of using modern medicines allows abuse.
- A shortage of drug supplies may arise in times of civil unrest.
- The government cannot provide equal health care between rural and urban areas, especially in terms of drug supply.

These problems prompted the government to consider revitalizing the use of local medicinal plants. The first development plan was set up in 1982. Studies by the Faculty of Pharmacy of the Mahidol University suggested that medicinal plants should be developed for use in primary health care, for the pharmaceutical industry and for export. Of these, the development for primary health care has been most successful, with about 55 plant species being promoted for use. The use of single species of plants was investigated, to make it easier to determine or trace the cause of any adverse effect that might occur.

The plants were selected carefully, using the following criteria:

- Only species for which there is scientific evidence of beneficial activity are selected, and only those with pharmacologically confirmed efficacy are recommended for use in primary health care.
- The plants must have passed toxicity tests, including mutagenic and teratogenic tests, or must also be used as food.
- The plants are used to treat symptoms for which self-diagnosis is possible.
- The method of preparation is simple.
- The plants are locally available, so as to ensure year-round supply.

Health workers were trained in the proper use of medicinal plants and their cultivation, and are responsible for encouraging their use at village level. The programme of developing medicinal plants for primary health care was successful in rural areas where there is no easy access to drugstores and hospitals. At present the government is preparing to add some medicinal plants to the national list of essential drugs. It is also drawing up good manufacturing practices for the factories preparing traditional medicine, to improve the quality of herbal drugs.

Development for export is progressing slowly, because there is no large-scale cultivation technology and also because of the rapid change in world demand. Medicinal plants are recommended as catch crops. Many of them can be grown in areas where the more common commercial crops cannot be cultivated, and they can play a role in forest conservation.

### *Vietnam*

Up until the 18th Century the history of the Vietnamese traditional medicine, which goes back more than 2000 years, is interlarded with the names of famous physicians such as Tue Tinh (14th Century). Only two books from this period have survived: 'Nam Duoc Than Hieu' ('The miraculous effect of traditional medicine'), which includes 580 ingredients of traditional medicine, and 'Hong Nghia Giac Tu Y Thu' ('Hong Nghia's summary of using traditional medicine'), which covers 600 ingredients. In the 18th Century, Le Huu Trac added another 330 ingredients to the former book, and published it as 'Linh Nam Ban Thao' ('Linh Nam's traditional medicine book'). This formed the basis for traditional medicine in Vietnam. Since the beginning of the 19th Century there have been many publications on Vietnamese medicinal plants.

Since the mid-1950s, the Vietnamese government has stimulated research on medicinal plant resources for use in primary health care. From 1960 to date, over 200 species of medicinal plants have been commercialized. The plants are collected from natural resources or from cultivation in quantities in excess of 100 000 t/year. Most of them are used in traditional medicine forms such as pastes, powders, pills and liquids, but some plant compounds are extracted industrially, the total quantity being over 2000 t/year. The number of research and development institutes dealing with traditional medicine is increasing, as is their input, and the availability of experienced physicians is further helping increase the popularity of traditional medicine. The Vietnamese system of traditional medicine has contributed greatly to community health care.

### 1.3.2 Production and trade

Statistics on international trade usually refer to commodity groups. Exact figures for individual plant drugs can be obtained for only a few cases. Often it is not even possible to distinguish between medicinal plants used in the pharmaceutical industry and plant species used in other industries. Medicinal and aromatic applications are lumped together (e.g. *Piper nigrum* L., *Zingiber officinale* Roscoe). According to trade figures of the United Nations Conference on Trade and Development, the international trade in plant-based drugs has a value of US\$ 800 million annually, on average. China is by far the leading country: between 1992 and 1995, its average annual exports were more than 120 000 t, with a value of over US\$ 250 million. India followed (33 000 t, US\$ 46 million), and then Germany (14 000 t, US\$ 68 million). Singapore exports the largest amount of plant-based drugs of the South-East Asian countries: 13 200 t annually with a value of US\$ 54 million in the period between 1992 and 1995. Thailand is also amongst the top 12 countries in the world: its annual exports are 3300 t, with a value of US\$ 7 million (Lange & Schippmann, 1997). The export value of Indonesian medicinal plants amounted to just US\$ 2 million in 1991, whereas the internal market was worth US\$ 45 million. The major countries importing plant-based drugs are Hong Kong (China) with 77 000 t worth US\$ 134 million annually between 1992 and 1995, Japan (43 000 t, US\$ 114 million) and Germany (43 000 t, US\$ 96 million). Singapore is market leader in South-East Asia with an average annual import of 7300 t and a value of US\$ 36 million.

However, the trade in medicinal plants is vast and largely unmonitored. Moreover, most people in developing countries depend on the direct use of plants for their health care, and thus the total trade in plant-based medicines may be a hundred times more than the volume of the international trade.

## 1.4 Phytochemistry

Phytochemistry deals with the chemistry of plant metabolites and their derivatives. The metabolic system of a plant may be regarded as being constituted of regulated processes within which biochemical conversions and mass transfer take place. Our understanding in this field has advanced to a stage in which definite metabolic processes, biosynthetic pathways and their interconnection are distinguished and studied in the context of their function and genetic control.

The metabolic performance of living organisms can be distinguished into primary metabolism and secondary metabolism. Primary metabolism is associated with fundamental life processes common to all plants. It comprises processes such as photosynthesis, pentose cycle, glycolysis, the citric acid cycle, electron transport, phosphorylation and energy regulation and management. Primary metabolites are produced and converted molecular entities, that are needed in anabolic pathways to build, maintain and reproduce the living cell. In catabolic pathways, primary metabolites (and food products) provide the chemical energy and precursors for biosynthesis.

Primary and secondary metabolism are interconnected in the sense that the biosynthesis of accumulating secondary metabolites can be traced back to ubiq-

uitous primary metabolites. However, in contrast to primary metabolites, secondary metabolites represent features that can be expressed in terms of ecological, taxonomic and biochemical differentiation and diversity. The biosynthesis and accumulation of secondary metabolites provide a basis for biochemical systematics and chemosystematics. In addition, the wide molecular diversity of secondary metabolites throughout the plant kingdom represents an extremely rich biogenic resource for the discovery of novel drugs and for developing innovative drugs. Not only do plant species yield raw material for useful compounds; the molecular biology and biochemistry provide pointers for rational drug development.

Primary and secondary metabolites can be classified on the basis of their chemical structure into much the same categories of chemical compounds: carbohydrates, lipids, amino acids, peptides, proteins, enzymes, purine and pyrimidine derivatives. Within such compound classes, secondary metabolites generally show greater individuality and diversity in their molecular structure than primary metabolites. On the other hand, certain compound classes appear to be extraordinarily rich in secondary metabolites. Examples are the structurally diverse groups of alkaloids, phenolics, acetogenins and terpenoids. Ubiquitary primary metabolites belonging to these compound classes seem to be restricted to only a limited number of key compounds functioning as biosynthetic precursors.

Most of the plant compounds that have been found to be medicinally useful and interesting tend to be secondary metabolites. Nonetheless, the discussion of compound classes that follows has been arranged according to chemical structure classes usually clustered as such.

#### *1.4.1 Carbohydrates*

The first products plants produce by photosynthesis are carbohydrates. They are formed from water and carbon dioxide and can be grouped into sugars and polysaccharides. The sugars are either monosaccharides such as glucose and fructose, or oligosaccharides containing up to 5 or 6 monosaccharide units. Monosaccharides are classified according to the number of carbon atoms they contain; thus, trioses, tetroses, pentoses, hexoses and heptoses are  $C_3$  to  $C_7$  compounds. The polysaccharides are macromolecules, containing a large number of monosaccharide residues.

Carbohydrates constitute a large portion of plant biomass, e.g. cellulose as part of the cellular framework, and starch as a food reserve.

Sugars can unite with a wide variety of compounds to form glycosides, increasing the water solubility of the compounds. Glycosides vary in chemical structure and pharmacological activity due to their aglycone component.

In addition to their use as bulking agents in pharmaceuticals, carbohydrates have recently been recognized to have useful pharmacological properties. Several polysaccharides exhibit immunomodulatory, antitumour, anticoagulant (e.g. heparin), hypoglycaemic or antiviral activities. The various carbohydrate products traded include fibre, cellulose and its derivatives, starch (glucose polymers) and its derivatives, dextrans, fructans (fructose polymers; e.g. inulin), algenic acids, agar and gums.

## 1.4.2 Lipids

### 1.4.2.1 Vegetable oils

Vegetable oils are major sources of  $\beta$ -sitosterol, which is a steroid drug precursor. One vegetable oil, obtained from groundnut, yields lecithins, which are used to enhance food digestibility. Lecithins are also used in pharmaceutical formulations. Recently, some vegetable oils have been found to be rich in  $\gamma$ -linolenic acid (see Figure 1), which is the precursor of prostaglandins, leukotrienes and thromboxanes. All these compounds are involved in platelet aggregation and inflammatory processes. Only members of *Onagraceae*, *Saxifragaceae* and *Boraginaceae* contain  $\gamma$ -linolenic acid.

Vegetable oils are significant in both the food and pharmaceutical industries. Some are used as solvents for lipid-soluble drugs such as vitamins and antibiotics. Others, e.g. almond oil and olive oil, are used in cosmetics. Castor oil is well known for its purgative activity, but has fallen out of favour because of its unpleasant taste.

### 1.4.2.2 Acetogenins

Acetogenins are long-chain aliphatic compounds with 35–39 carbon atoms, ending with a  $\gamma$ -lactone, most often unsaturated and cyclized into one or two tetrahydrofuran rings that may or may not be adjacent. They are characteristic of *Annonaceae* (e.g. *Annona*, *Goniothalamus*, *Rollinia* and *Uvaria*). The potential application of acetogenins is linked to their antitumour (e.g. asimicin, bullatacine), antibacterial (e.g. cherimolin) and insecticidal (e.g. asimicin, annonin, annonacin) properties. See Figure 2 for the structure of annonacin, as an example of an acetogenin.

## 1.4.3 Amino acids and their derivatives

Amino acids are constituents of peptides, proteins and enzymes, but also the precursors of a large variety of secondary metabolites including alkaloids and phenolic compounds, which are both discussed separately.

### 1.4.3.1 Amino acids

The function of amino acids is not only for protein synthesis; they are also considered to be a form of nitrogen storage (e.g. cannavanine, hemoarginine) and a germination inhibitor. The few studies of the pharmacological activities of amino acids include reports of curcubatine being used as taeniocide. Many toxic amino acids have been identified; examples include  $\beta$ -( $\gamma$ -L-glutamylamino)propionitrile and  $\gamma$ -N-oxalyl-L- $\alpha$ , $\beta$ -diaminopropionic acid which are responsible



Figure 1. Structure of  $\gamma$ -linolenic acid.

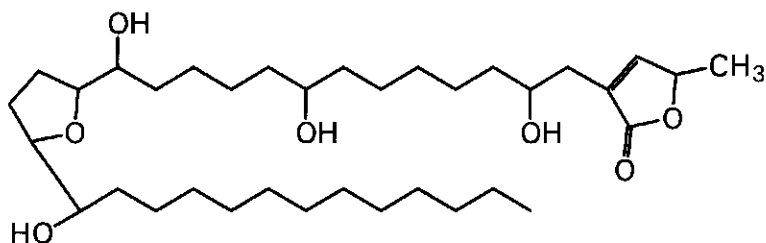


Figure 2. Structure of annonacin.

for the toxicity of grass pea (*Lathyrus sativus* L.) that brings about osteo-lathyrism and neuro-lathyrism in livestock, and mimosine from *Leucaena* inhibiting protein and nucleic acid synthesis which results in livestock losing appetite and weight, and their growth being inhibited.

#### 1.4.3.2 Cyanogenic glycosides

Cyanogenic glycosides are compounds derived from amino acids. Hydrolysis of these compounds by enzymes or acids yields hydrocyanic acid, a toxic principle. Biosynthetically, the aglycones of cyanogenic glycosides are derived from L-amino acids. Cyanogenic glycosides are prevalent in the families *Rosaceae*, *Leguminosae*, *Gramineae*, *Araceae*, *Euphorbiaceae* and *Passifloraceae*. Examples are linamarin, amygdalin and prunasin.

#### 1.4.3.3 Sulphur-containing compounds

The sulphur-containing compounds of pharmaceutical significance are allein, allicin, ajoene and other related compounds isolated from garlic. Allicin and ajoene (the latter is a condensation product of allicin) exhibit many biological activities, including antihypercholesterolaemic, antiplatelet aggregation, anti-hypertensive, fibrinolytic and antifungal activities. Recently, diallyl cysteine, an odourless active ingredient of garlic, was found to be biosynthesizable.

#### 1.4.3.4 Lectins

Lectins are proteins or glycoproteins that are able to bind with the carbohydrate moiety on cell membranes in a specific and reversible fashion, without displaying enzymatic activity. Most lectins in higher plants are located in seeds. They are commonly found in legumes such as groundnut, soya bean and common bean.

Some lectins have the ability to agglutinate red blood cells of a specific blood group. These lectins are referred as phytohaemagglutinin. The haemagglutination activity is important in immunological studies. Some lectins are toxic, e.g. ricin from castor (*Ricinus communis* L.) seeds and abrin from jequirity bean (*Abrus precatorius* L.) seeds.

#### 1.4.3.5 Enzymes

Plant-derived enzymes used as drugs include papain and bromelain. Both are proteolytic enzymes useful as an anti-inflammatory drug. Ficin has similar properties.

#### 1.4.4 Alkaloids

It is not easy to define the term 'alkaloid' precisely, since there is no sharp border between alkaloids and naturally occurring complex amines. At present, the term is used for plant-derived compounds containing one or more nitrogen atoms (usually in a heterocyclic ring), and usually having a marked physiological action on humans or animals. The term 'proto-alkaloids' or 'pseudo-alkaloids' is sometimes applied to compounds that lack one or more of the properties of the typical alkaloids, e.g. the nitrogen in a heterocyclic ring system; examples include mescaline and ephedrine. To avoid problems with this common definition of alkaloids, some authors propose a more narrow definition: an alkaloid is a cyclic organic compound containing nitrogen in a negative oxidation state, which has limited distribution in living organisms.

Based on their chemical structures, alkaloids are divided into several subgroups: non-heterocyclic alkaloids, and heterocyclic alkaloids which are again divided into 12 major groups according to their basic ring structure. Figure 3 shows some examples; mescaline is an example of a non-heterocyclic or pseudo-alkaloid, tetrandrine of a bisbenzylisoquinoline alkaloid and solasodine of a triterpene alkaloid. Free alkaloids are soluble in organic solvents such as ether or chloroform. Alkaloids will furthermore react with acids to form water-soluble salts. There are a few exceptions to this general rule. In certain alkaloids, e.g. in ricinine, the lone pair of electrons on the nitrogen atom can be protonated. Another example is berberine, a quaternary ammonium alkaloid; the free base is already water-soluble. Physically, most alkaloids exist in solid form, but some are liquid, e.g. nicotine.

Alkaloids in plants are believed to be waste products and a nitrogen source. They are thought to play a role in plant protection and germination, and to be plant growth stimulants. Alkaloids are more common in dicotyledons than in monocotyledons; families rich in them are *Amaryllidaceae*, *Liliaceae* s.l., *Apo-cynaceae*, *Berberidaceae*, *Leguminosae*, *Papaveraceae*, *Ranunculaceae*, *Rubiaceae* and *Solanaceae*.

Many alkaloids are pharmaceutically significant, e.g. morphine as a narcotic analgesic, codeine in the treatment of coughs, colchicine in the treatment of gout, quinine as an antimalarial, quinidine as an anti-arrhythmic and L-hyoscyamine (in the form of its racemic mixture known as atropine) as anti-spasmodic and for pupil dilation.

#### 1.4.5 Phenols and phenolic glycosides

Phenols probably constitute the largest group of secondary plant metabolites. They range from simple structures with one aromatic ring to complex polymers such as tannins and lignins. Examples of phenolic classes of pharmaceutical interest are (1) simple phenolic compounds, (2) tannins, (3) coumarins and their

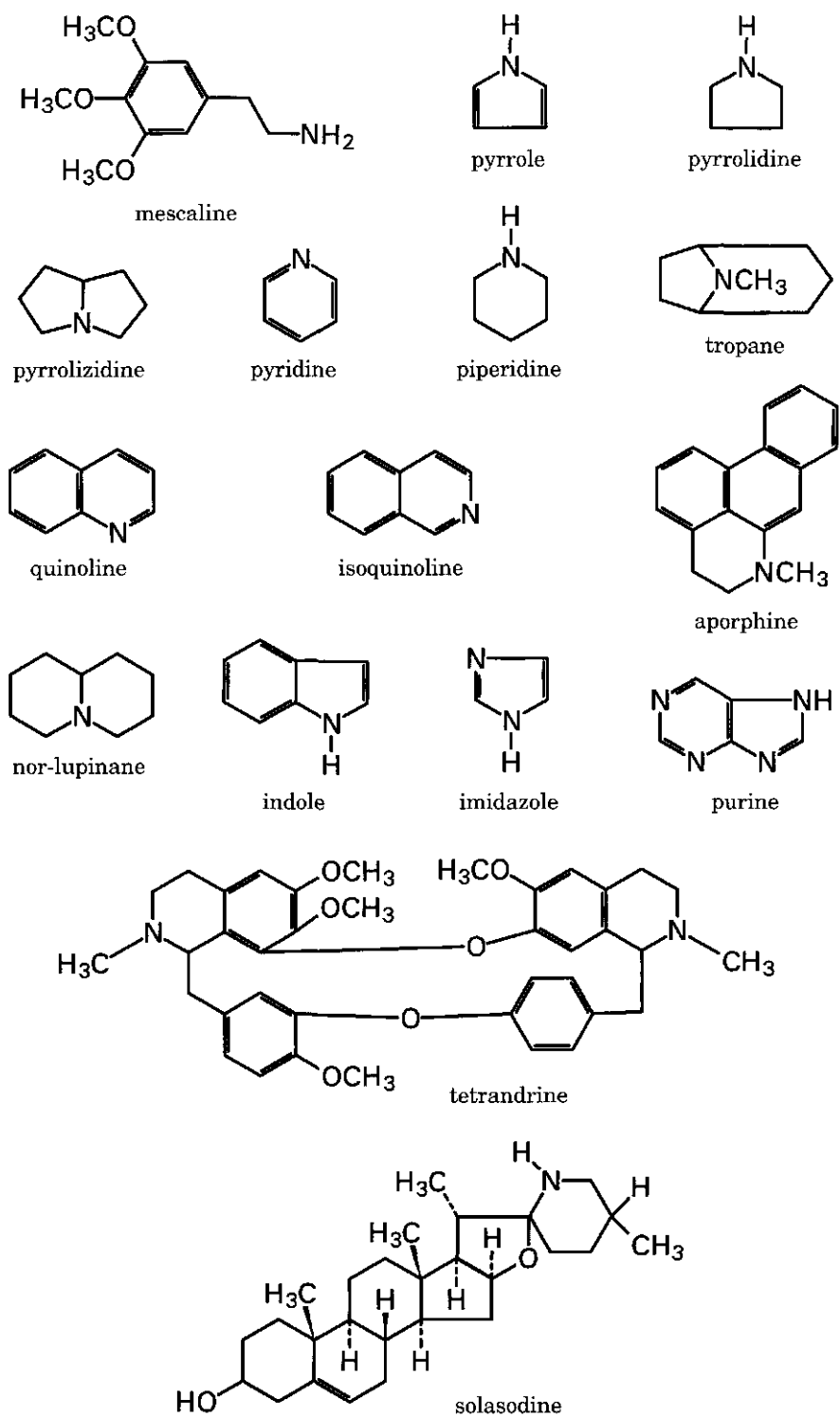


Figure 3. Alkaloids: basic structures and some examples.



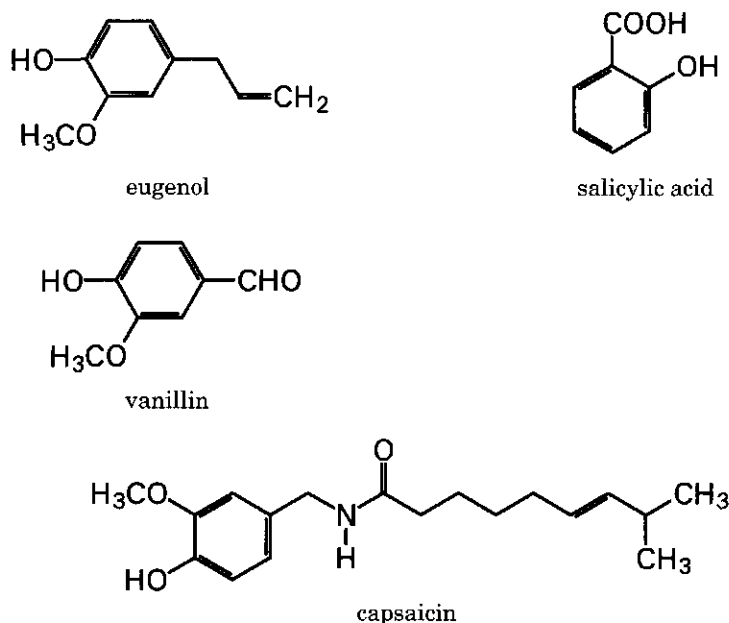


Figure 4. Examples of simple phenolic compounds.

glycosides, (4) quinones, (5) flavonoids, (6) anthocyanins, (7) phloroglucinols, and (8) lignans and related compounds. These phenolic compounds are biosynthesized via the shikimic acid or acetate pathways.

#### 1.4.5.1 Simple phenolic compounds

Compounds in this group have a monocyclic aromatic ring with an alcoholic, aldehydic or carboxylic group. They may have a short hydrocarbon chain. Figure 4 shows some examples; capsaicin is a vanillyl amide of isodecenoic acid. Eugenol is widely used in dentistry due to its antibacterial, anti-inflammatory and local anaesthetic activities. Vanillin is commonly used as a food flavouring. For salicylic acid anti-inflammatory properties have been reported. Capsaicin, a compound isolated from *Capsicum*, is now marketed as an analgesic.

#### 1.4.5.2 Tannins

The chemistry of tannins is complex. The distinction made in the literature between hydrolysable tannins and condensed tannins is based on whether acids or enzymes can hydrolyse the components or whether they condense the components to polymers. Although not watertight, this distinction largely corresponds to groups based on gallic acid and those based on flavane-related components. Numerous vegetable tannins have been discovered, but only the major tanning constituents of the most important groups of tannins are listed here, i.e. the group of gallotannins and ellagitannins, and the group of proanthocyanidins. Gallotannins and ellagitannins are esters of gallic acid or its dimers digallic acid and ellagic acid with glucose and other polyols. Proanthocyanidins

are oligomers of 3-flavanols (catechins) and 3,4-flavandiols (leucoanthocyanidins); see Figure 5 in which R = H or OH.

Tannins are able to react with proteins. On being treated with a tannin, a hide absorbs the stain and is protected against putrefaction, thereby being converted into leather. For more information, see Prosea 3: 'Dye and tannin-producing plants'.

Though tannins are widespread in plants, their role in plants is still unclear. They may be an effective defence against herbivores, but it is likely that their major role in evolution has been to protect plants against fungal and bacterial attack. The high concentrations of tannins in the non-living cells of many trees (heartwood, bark), which would otherwise readily succumb to saprophytes, have been cited in support of this hypothesis. Some authorities consider tannins to be waste products, and it has also been suggested that leaf tannins are active metabolites used in the growing tissues. However, tannins in different plant species probably have different functions.

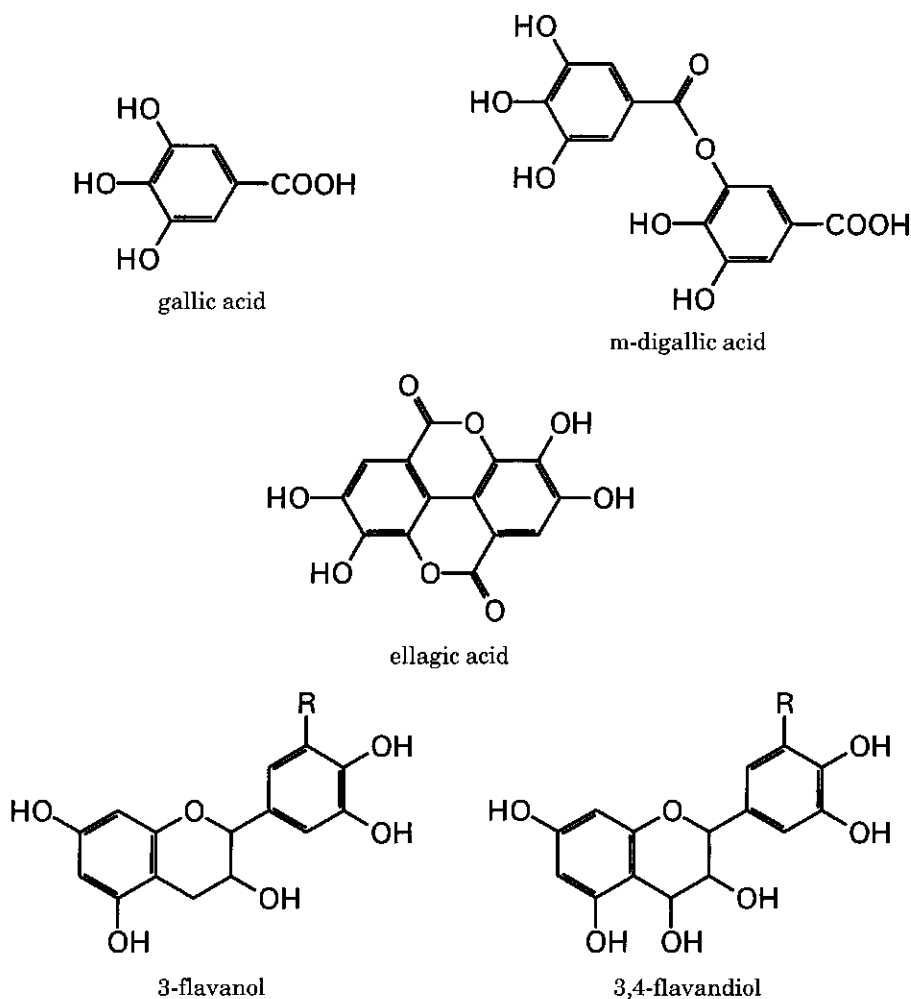


Figure 5. Structures of some tannins.

Tannins are used against diarrhoea and as antidotes in poisoning by heavy metals. Their use declined after the discovery of the hepatotoxic effect of absorbed tannic acid. Recent studies have reported that tannins have anti-cancer and anti-HIV activities.

#### 1.4.5.3 Coumarins and their glycosides

Coumarins are benzo- $\alpha$ -pyrone derivatives that are common in plants both in a free state and as glycosides. They give a characteristic odour of new-mown hay and occur, for instance, in many *Leguminosae*. They are biosynthetically derived via the shikimic acid pathway. Figure 6 shows the structure of coumarin. Common derivatives are umbelliferone, herniarin, aesculetin, scopoletin, fraxin and chicorin.

The biological activities reported are spasmolytic, cytostatic, molluscicidal, antihistaminic and antifertility.

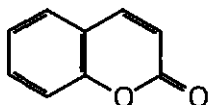


Figure 6. Structure of coumarin.

#### 1.4.5.4 Quinones

Quinones are oxygen-containing compounds that are oxidized homologues of aromatic derivatives and are characterized by a 1,4-diketo-cyclohexa-2,5-diene pattern (paraquinones) or by a 1,2-diketo-cyclohexa-3,5-diene pattern (orthoquinones). In naturally occurring quinones, the dione is conjugated to an aromatic nucleus (benzoquinones) or to a condensed polycyclic aromatic system: naphthalene (naphthoquinones), anthracene (anthraquinones), 1,2-benzanthracene (anthracyclinones), naphthodianthrene (naphthodianthrone), pyrene, phenanthrene and abietane-quinone. See Figure 7.

Naphthoquinones and anthraquinones have some importance medicinally; see below.

#### Naphthoquinones

Naphthoquinones are yellow or orange pigments from plants. Most are 1,4-

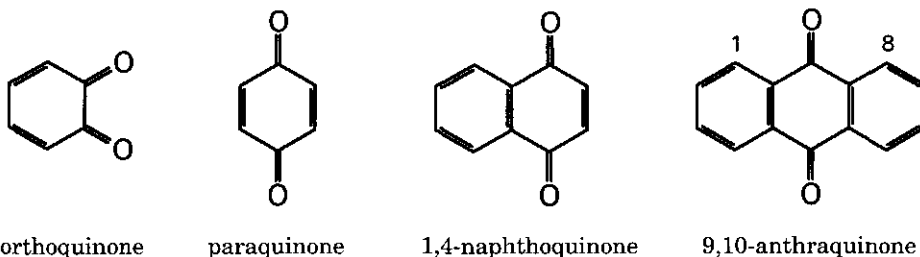


Figure 7. Basic structures of some quinones.

naphthoquinones; 1,2-naphthoquinones are rarely found. Hydroxyl and methyl substitutions at C-2 are common. Biosynthetically, the naphthoquinones are almost exclusively derived via the shikimic acid pathway.

The occurrence of naphthoquinones is limited in fungi and sporadic in Angiosperms. They are found in species of the families *Bignoniaceae*, *Ebenaceae*, *Droseraceae*, *Juglandaceae*, *Plumbaginaceae*, *Boraginaceae*, *Lythraceae*, *Proteaceae* and *Verbenaceae*.

The pharmaceutical significance of this group of quinones is limited. Plumbagin exhibits antibacterial and cytotoxic activities. Lawsone from henna (*Lawsonia inermis* L.) is a powerful fungicide and hair colourant.

### *Anthraquinones*

Anthraquinones are characterized by the presence of phenolic and glycoside moieties, derived from anthracene, and have a variable degree of oxidation. They have a common double hydroxylation in the positions 1 and 8 (see Figure 7). The glycosidic linkage may be C- or O-bonding. The anthraquinones are mostly biosynthesized via the acetate pathway, although some examples may be derived via the shikimic acid pathway.

Anthraquinones are found in species of the families *Rubiaceae*, *Leguminosae*, *Polygonaceae*, *Rhamnaceae*, *Ericaceae*, *Euphorbiaceae*, *Lythraceae*, *Saxifragaceae*, *Scrophulariaceae* and *Verbenaceae*. In monocotyledons, they are found only in *Liliaceae* s.l.

Anthraquinones isolated from plants with laxative activity include sennosides, aloins and emodin. The therapeutic use of anthraquinones as laxatives is very well recognized. The products are sold commercially. Common medicinal plants which contain anthraquinones are *Senna* and *Aloe* species.

### 1.4.5.5 *Flavonoids*

Flavonoids are the compounds responsible for the colour of flowers, fruits and sometimes leaves. Some, such as chalcones and flavonols, are yellow. The name refers to the Latin word 'flavus', which means yellow. Some may contribute to the colour by acting as co-pigment. Flavonoids protect the plant from UV-damaging effects and play a role in pollination by attracting animals by their colours.

The basic structure of flavonoids is 2-phenyl chromane or an Ar-C<sub>3</sub>-Ar skeleton. Biosynthetically they are derived from a combination of the shikimic acid and acetate pathways. Small differences in basic substitution patterns give rise to several sub-groups; in the plant flavonoids can either occur as aglycones or as O- or C-glycosides. See Figure 8 for basic structures.

Recently, flavonoids have attracted interest due to the discovery of their pharmacological activities as anti-inflammatory, analgesic, antitumour, anti-HIV, anti-diarrhoeal, anti-hepatotoxic, antifungal, antilipolytic, anti-oxidant, vasodilator, immunostimulant and anti-ulcerogenic. Examples of biologically active flavonoids are hesperidin and rutin for decreasing capillary fragility, and quercetin as anti-diarrhoeal.

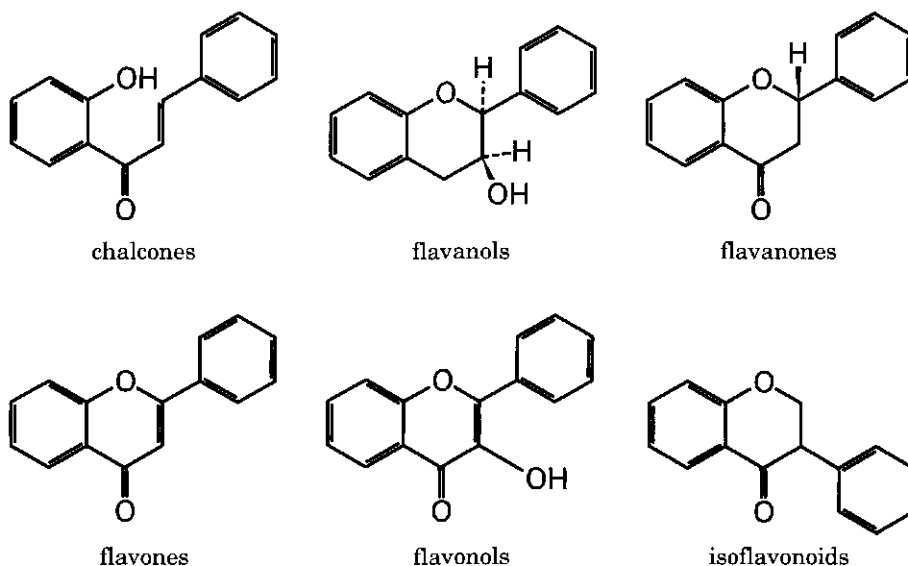


Figure 8. Basic structures of some flavonoids.

#### 1.4.5.6 Anthocyanins

Anthocyanins are the compounds responsible for the red, pink, mauve, purple, blue or violet colours of most flowers and fruits. These water-soluble pigments occur as glycosides (anthocyanins *sensu stricto*) and their aglycone (anthocyanidins). They are derived from the 2-phenyl benzopyrylium cation, more commonly referred to as the flavylium cation. Cyanin (see Figure 9) is an example of an anthocyanin.

Anthocyanins are found in all Angiosperms, except for most species of the order *Caryophyllales*: only species of the families *Caryophyllaceae* and *Molluginaceae* contain them; in other families (e.g. *Chenopodiaceae*, *Cactaceae*), the pigmentation is due to betalains.

The application of anthocyanins is as food additive, e.g. in beverages, jams and confectionary products. The pharmacological activities are similar to flavonoids; for instance for decreasing capillary permeability and fragility, and as anti-oedema.

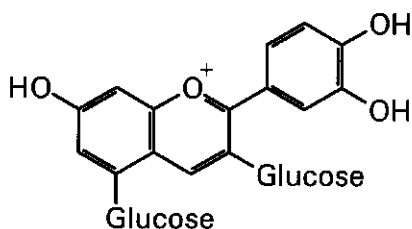


Figure 9. Structure of cyanin (cyanidin-3,5-diglucoside), as an example of an anthocyanin.

#### 1.4.5.7 Phloroglucinols

Phloroglucinols are derivatives of 1,3,5-trihydroxybenzene, which e.g. are found in *Cannabis sativa* L., a well-known stimulant of the central nervous system.

Tetrahydrocannabinol and its derivatives influence behaviour, inducing euphoria and relaxation at low doses, but at higher doses, they may induce anxiety, sometimes to panic proportions. Sometimes hallucination and tinnitus are observed. Other effects are bronchodilation and a lowering of intra-ocular pressure.

#### 1.4.5.8 Lignans and related compounds

Lignans and related compounds are derived from condensation of phenylpropane units. Formerly, the term referred to compounds whose skeleton results from bonding between  $\beta$ -carbons of the side chain of two units derived from 1-phenylpropane (8-8' bond). Neolignans are also condensation products of phenylpropanoid units, but the actual bond varies and involves no more than one  $\beta$ -carbon (8-3', 8-1', 3-3', 8-0-4' for example). The term 'oligomers' is incorrect; designated lignans or neolignans result from the condensation of 2-5 phenylpropanoid units (e.g. sesquilignans and dilignans, lithospermic acid). Norlignans are probably specific to gymnosperms and have a C<sub>17</sub> skeleton.

Lignins are substances deposited at the end of the formation of the primary and secondary cell walls. Chemically, they are polymers arising from copolymerization of alcohol with a p-hydroxycinnamic structure (p-hydroxycinnamyl, coniferyl or sinapyl alcohol). Lignins are always combined with polysaccharides.

The pharmacological activity of lignans is antitumour. Kadsurenone, a neolignan, exhibits anti-allergic and antirheumatic activity. The major application of lignins is as a precursor of vanillin, which is widely used in the food industry.

#### 1.4.6 Terpenoids and steroids

Terpenoids and steroids are derived from isoprene (a 5-carbon unit), which is biosynthesized from acetate via mevalonic acid.

##### 1.4.6.1 Monoterpenes

Monoterpenes are the most simple constituents in the terpene series and are C<sub>10</sub> compounds. They arise from the head to tail coupling of two isoprene units. They are commonly found in essential oils. Iridoids and pyrethrins are included in this group. Examples of monoterpenes found in essential oils are shown in Figure 10.

Iridoids are monoterpenes characterized by a cyclopenta [C] pyranoid skeleton, also known as the iridane skeleton (cis-2-oxo-bicyclo-[4,3,0]-nonane). Secoiridoids, which arise from iridoids by cleavage of the 7,8-bond of the cyclopentane ring, are also included in the iridoids. Examples of secoiridoids are the bitter constituents of gentian, e.g. gentiopicroside, amarogentin and esters of sweroside and swertiamarin.

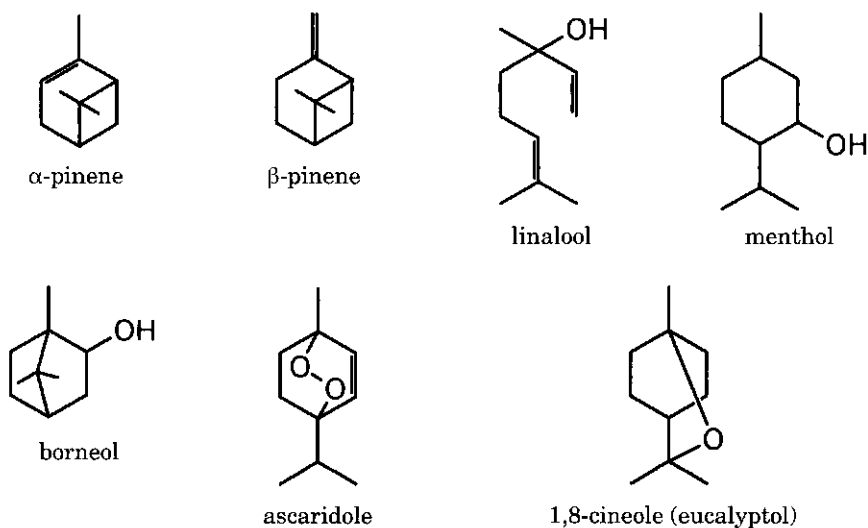


Figure 10. Examples of monoterpenes.

Pyrethrins are irregular monoterpenes arising from the non-classic coupling of isopentenyl pyrophosphate and dimethylallyl pyrophosphate. Some are found in essential oils. Figure 11 gives the basic structures of iridoids and secoiridoids and an example of pyrethrins.

The pharmacological properties of iridoids are quite limited: the iridoid-containing drugs currently used do not yield any major active principle. However, there are reports on analgesic and anti-inflammatory activities of some iridoids, e.g. harpagoside. Pyrethrins are toxic for coldblooded animals such as fish, amphibians and insects. They are widely used as insecticides.

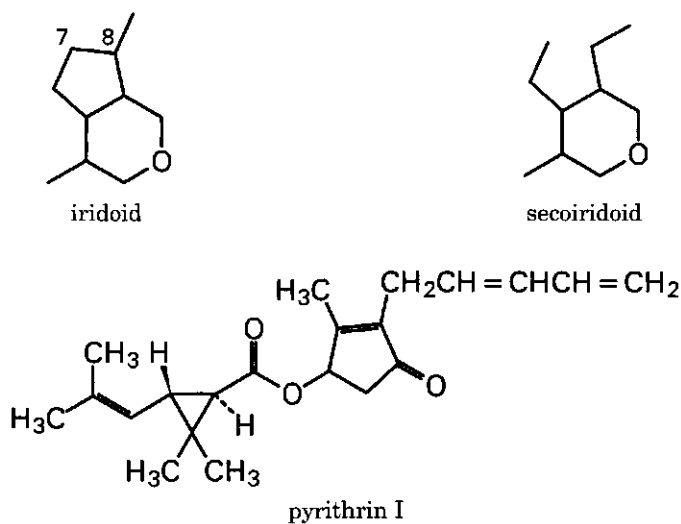


Figure 11. Basic structures of iridoids and secoiridoids, and the structure of pyrethrin I as an example of pyrethrins.

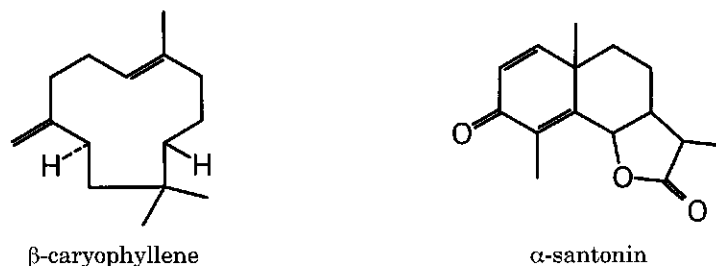


Figure 12. Examples of sesquiterpenes.

#### 1.4.6.2 Sesquiterpenes

Sesquiterpenes are also constituents of essential oils of many plants, e.g. bisabolol, humulene and caryophyllene. Figure 12 shows two examples of sesquiterpenes. Sesquiterpenoid lactones are well known as bitter principles. They occur in fungi, bryophytes and angiosperms (especially common in *Compositae*).

Sesquiterpenes possess a broad range of biological activities due to the  $\alpha$ -methylene- $\gamma$ -lactone moiety and epoxides. Their pharmacological activities are antibacterial, antifungal, anthelmintic, antimalarial and molluscicidal. Examples are santonin used as an anthelmintic and artemisinin as an antimalarial.

#### 1.4.6.3 Diterpenes

Diterpenes constitute a vast group of  $C_{20}$  compounds arising from the metabolism of 2E,6E,10E-geranylgeranyl pyrophosphate. They are present in some animals and plants; they are particularly abundant in the orders *Lamiales* and *Asterales*.

Diterpenes have some therapeutic applications. For instance, taxol (see Figure 13) and its derivatives from *Taxus* are anti-cancer drugs. Other examples are forskolin, with antihypertensive activity, zoapatanol, as an abortifacient, and stevioside, as a sweetening agent.

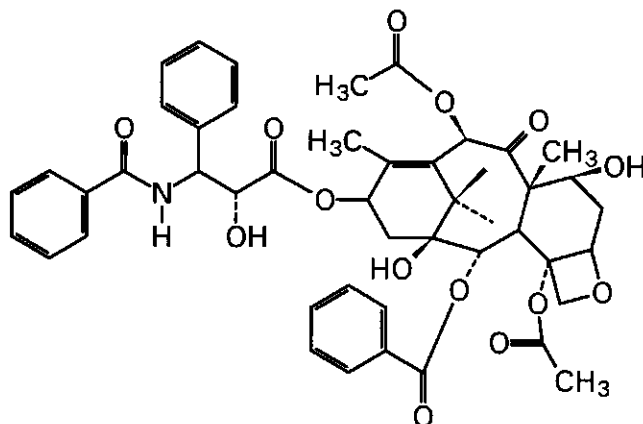


Figure 13. Structure of taxol as an example of a diterpene.



#### 1.4.6.4 Triterpenes and steroids

Triterpenes are  $C_{30}$  compounds arising from the cyclization of 3S-2,3-epoxy,2,3-squalene. The basic skeletons are shown in Figure 14: oleanane is an example of a pentacyclic triterpene, quassin of a tetracyclic triterpene and testosterone of a steroid. Tetracyclic triterpenes and steroids have similar structures, but their biosynthetic pathway is different. Steroids contain a ring system of three 6-membered and one 5-membered ring; because of the profound biological activities encountered, many natural steroids together with a considerable number of synthetic and semi-synthetic steroidal compounds are employed in medicine (e.g. steroidal saponins, cardioactive glycosides, corticosteroid hormones, mammalian sex hormones).

The pharmaceutical applications of triterpenes and steroids are considerable. Cardiac glycosides have been used in medicine without replacement by synthetic drugs. Saponins from ginseng and liquorice exhibit many therapeutic effects.

#### Saponins

Saponins constitute a vast group of glycosides which occur in many plants. They are characterized by their surfactant properties; they dissolve in water and, when shaken, form a foamy solution. Saponins are classified by their aglycone structure into triterpenoid and steroid saponins; most triterpenoid saponins are derivatives of one of the triterpenes oleanane, ursane and lupane, while steroid saponins generally possess the typical steroid skeleton enlarged with 2 extra rings E, a furan structure and F, a pyran structure, respectively. Examples of 2 aglycones are shown in Figure 15. In saponins, sugar and/or uronic acid residues are attached to the aglycones via the C-3 hydroxyl group.

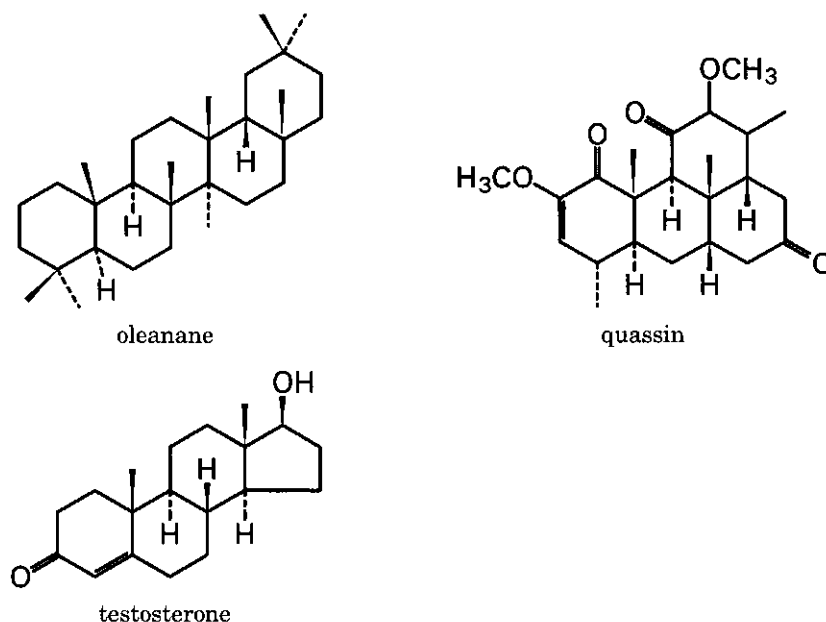


Figure 14. Basic structures of triterpenes and steroids.

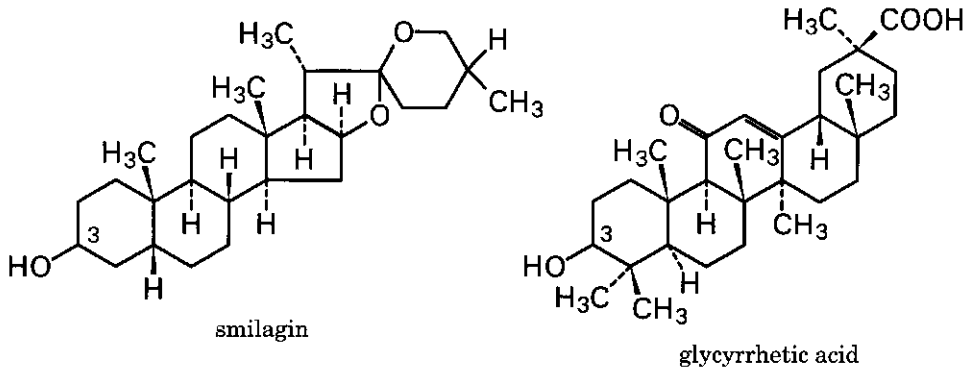


Figure 15. Structures of two saponins (aglycone structure).

Most saponins have haemolytic properties and are toxic to coldblooded animals, especially fish. The steroidal saponins are important precursors for steroid drugs, including anti-inflammatory agents, androgens, oestrogens and progestins. Well-known steroid sapogenins are diosgenin from *Dioscorea*, hecogenin from *Agave* and smilagenin from *Smilax*.

Triterpene saponins exhibit various pharmacological activities: anti-inflammatory, molluscicidal, antitussive, expectorant, analgesic and cytotoxic. Examples include the ginsenosides, which are responsible for some of the pharmacological activity of ginseng, and the active triterpenoid saponins from liquorice.

### Cardiac glycosides

The aglycone part of cardiac glycosides is a tetracyclic steroid with an attached unsaturated lactone ring that may have 5 or 6 members. Cardiac glycosides are classified into two groups according to the lactone ring: the  $C_{23}$  cardenolides with an  $\alpha,\beta$ -unsaturated d- $\gamma$ -lactone (= butenolide), and the  $C_{24}$  bufadienolides with a di-unsaturated  $\gamma$ -lactone (= pentadienolide). The sugar moiety is normally attached via the C-3 hydroxyl group of the aglycone. The majority of the saccharides found in cardiac glycosides are highly specific. They are 2,6-dideoxyhexoses, such as D-digitoxose, L-oleandrose or D-diginose. These sugars give a positive reaction with the Keller-killiani reagent.

Cardiac glycosides have been used as drugs for the treatment of cardiac insufficiency. An example is digitoxin from *Digitalis*, where the sugar moiety is attached to the aglycone digitoxigenin (see Figure 16) via the C-3 hydroxyl group.

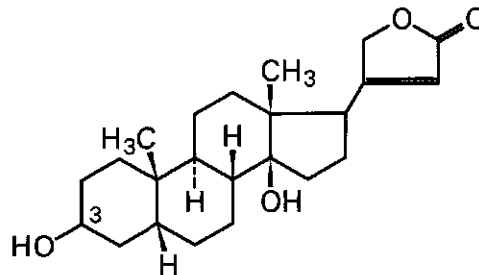


Figure 16. Structure of digitoxigenin.

#### 1.4.6.5 Carotenoids

Carotenoids contain 8 isoprene ( $C_{40}$ ) units that are responsible for the yellow or orange colour of some vegetables and fruits. Among these compounds, the hydrocarbons are collectively referred to as carotenes and the hydroxylated derivatives as xanthophylls. Carotenoids are either acyclic (e.g. lycopene) or comprise one or two pentacyclic or hexacyclic rings at one end or the other (e.g.  $\beta,\psi$ -carotene), or at both ends (e.g.  $\beta,\beta$ -carotene).

Carotenoids became interesting agents after the discovery of a negative correlation between the plasma concentration of  $\beta$ -carotene and the prevalence of certain forms of cancer. Some doctors prescribe  $\beta$ -carotene for cancer patients. Furthermore, in the intestine  $\beta$ -carotenes are converted to retinol (vitamin A). They can be used for the treatment of photosensitization, retinal disease and glaucoma. Carotenoids are also safe colouring agents for food and cosmetics.

### 1.5 Biological and pharmacological activity and therapeutical applications

Among the many classical examples of biological action of plant material in man are the different tastes (sweet, bitter, sour, astringent), sensations (irritating, itchy, pungent, acrid) and the types of euphoria and hallucinations. However, only recently biological activity is understood in terms of molecular interactions. Plants and plant constituents have a key position in the advancement of modern studies and knowledge on biological activity of substances. There are several reasons for this. Firstly, plant species, whether traditionally used or not, continue to be important sources of food, medicines and supplementary health products. Secondly, the bioactive plant compounds are themselves products (or derived products) of metabolism, and hence function in life processes in a similar way to compounds that operate in humans and animals. Researchers hoping to develop drugs from plants need to understand the basics of such functions and mechanisms in relation to the bioactive molecular entities. Thirdly, plants also yield products which are auxiliaries in medicine and pharmacy and sustain or condition pharmacological activity and therapeutic efficacy. In addition, a series of these auxiliary substances are used in biomedical research and in clinical tests.

Testing the biological activity of medicinal or potentially medicinal plant material demands a special approach. Investigations may be focused on understanding the bioactivity of a compounded plant extract or simply directed at isolating a single bioactive chemical compound. In the latter case, results often lead to oversimplification or wrong explanations of the bioactivity of extract preparations. On the other hand, thorough studies on single bioactive constituents provide important information for plant drug research. However, the much more complex array of molecular interactions and bioactivity mechanisms that arises from plant extracts represents a much greater and more fascinating challenge to science.

#### 1.5.1 Factors affecting biological activity

Various aspects of bioactivity apply to any chemical, whether of natural or syn-

thetic origin. These aspects will now be described briefly. They fall into three categories (Gringauz, 1997; Gubernator & Böhm, 1998; Krogsgaard-Larsen & Bundgaard, 1991):

- Physicochemical properties such as solubility, partition coefficients and ionization.
- Chemical parameters such as resonance, inductive effects, oxidation–reduction potentials, types of bonding and isosterism.
- Spatial considerations such as molecular dimensions, interatomic distances and stereochemistry.

#### *1.5.1.1 Physicochemical properties*

These relate to the transport of the bioactive compound to its site of action, usually a receptor or other biomacromolecule at cellular or subcellular level. Under experimental (in vivo or clinical) or real life conditions the extent to which a drug passes through semipermeable membranes before reaching its site of action depends on its solubility. Under in vitro conditions many of these barriers are absent. In vitro bioactivity therefore represents only a stage in the basic assessment of pharmacological effects. In plant drug research, the solubility of active constituents may be revealed from extraction procedures. Extraction programmes separate lipophilic constituents from water-soluble compounds. Further fractionation of an extract may lead to further refinement of physicochemical properties. After the bioactive molecular entity has been identified, detailed data on solubility, partition coefficients and the electrolytic behaviour can be determined. Solubility characters are closely related to drug absorption, and the degree of absorption is an important determinant of drug action. Many bioactive plant constituents are weak acids and bases, and their degree of ionization, when dissolved, is of great importance to their bioactivity. As a rule, the ionic form is more water-soluble.

These factors are important when bioactivity is regarded in the context of drug distribution between intestine and plasma, between kidney tubules and urine, and between plasma and other body compartments. Generally, but simplified, one may say that only the lipid-soluble and undissociated forms of a bioactive molecule will pass through membranes. However, at the site of action, bioactive compounds may generate their action by binding to a receptor on the cell membrane.

#### *1.5.1.2 Chemical parameters*

The structural features of a compound can be related to its pharmacological properties, either qualitatively or quantitatively. The principles, concepts and numerical rules governing qualitative and quantitative relationships between structure and activity help explain the pharmacological activity of a new compound, which is why it is important to elucidate the structure of a newly isolated plant compound. The basic aspects of molecular structures involved in bioactivity include:

- *Resonance*. This is the phenomenon that a molecule can be represented by two or more structures that differ only in their electron, but not atomic, arrangement. So, electron density and electron distribution patterns help ex-

plain the molecule's reactivity and hence its molecular interaction and bioactivity.

- *Inductive effects.* These are measurable electrostatic phenomena caused by actual electron shifts or displacements along chemical bonds. Either negative or positive inductive effects may lead to changes in bioactivity.
- *Oxidation-reduction potential.* This phenomenon represents the tendency of a compound to lose electrons (oxidation) or gain electrons (reduction). Without electron transfers, various systems in the living cell would not function. Through the nature of their chemical structure, bioactive compounds may affect these systems. Note that bioactive compounds derived from plant sources function in enzyme systems in the plant which are similar to those in the humans or animals treated.
- *Types of bonding.* Basically the phenomenon of biological activity is concerned with covalent and noncovalent molecular interactions. Firstly, covalent bonds (single, double and triple bonds) are common to all biomolecules. Under the physiological conditions of living organisms, covalent bonds form enzymatically. As a rule, however, further biochemical functionality of biomolecules proceeds through noncovalent interactions. Hydrogen bonds, ionic forces, hydrophobic (or lipophilic) bonding, and charge-transfer interactions, all representing noncovalent interactions, are also common to functional life processes. Thus bioactivity as encountered when a given compound, whether biogenic or xenobiotic, comes in contact with a living system (in vitro, in vivo, clinically, or unintentionally) interferes with ongoing life processes. However, the molecular interactions will still be in terms of covalent or noncovalent principles. Agents that affect physiological functions by forming irreversible covalent bonds with target biomacromolecules are usually very toxic at cellular level, and would be difficult to control clinically and medically. So in plant drug research, constituents that exert their activity through much weaker and reversible bonding processes are much more desirable. Noncovalent and reversible covalent binding of target molecules are preferable and, moreover, are characterized by equilibrium thermodynamics (Gubernator & Böhm, 1998). Association constants can be determined and are reproducible. In the case of high-molecular weight ligands (e.g. a large bioactive plant molecule) the association rate of these ligands with the target biomolecule slows down and it becomes unpractical to determine an equilibrium. In cases of reversible covalent interactions, measurable activity constants (e.g.  $IC_{50}$  values = concentration giving 50% inhibition) are very dependent on experimental conditions such as the concentration of constituents, incubation time, temperature, acidity, etc. The systematic search for desirable plant ligands not only strives for bioactivity through reversible covalent or noncovalent interactions, but also for selectivity. The latter conditions the specificity of action, and reduces toxicity and side-effects. From the vast number of studies of biological and pharmacological activity and its molecular basis, it is clear that it is still a long way to total understanding and sufficient explanations.

#### 1.5.1.3 Spatial considerations

It appears important to have good steric and electronic complementarity be-

tween ligand and target biomolecule (Gubernator & Böhm, 1998). A bioactive compound interacting with DNA, a receptor molecule or an enzyme fits sterically into a binding pocket, the space sterically provided by these targets. The molecular dimensions, interatomic distances, arrangements of electrons and the stereochemical properties of both ligand and target are decisive. In other words, a molecular 'docking' mechanism is at the basis of biological and pharmacological activity. This is illustrated by an example.

(-)-Huperzine A is a potent and reversible inhibitor of acetylcholinesterase. However, this plant compound does not show muscarinic effects. The lack of these undesirable side-effects suggests that (-)-huperzine A has potential for treating 'cholinergic insufficiency' disorders, such as Alzheimer's disease. This compound has been isolated from the club moss *Huperzia serrata* (Thunb. ex Murray) Trevisan (synonym: *Lycopodium serratum* Thunb. ex Murray), which is used medicinally (Raves et al., 1997). See Figure 17 for the structure diagram. The (-) isomer of this chiral molecule has a more potent bioactivity than either the (+) isomer or the racemic mixture.

Such detailed molecular orientation and interaction data are not available for most known bioactive plant ligands. However, the target biomolecule of many other plant compounds is known, and, if not, the pharmacological effects have been studied (Bierhaus et al., 1997; Colegate & Molyneux, 1993; Hassig et al., 1997; Hung et al., 1996; Raves et al., 1997). Table 1 gives examples of plant ligands whose target biomolecules are known.

### 1.5.2 Bio-assaying

#### 1.5.2.1 Requirements for screening medicinal plant material

The biological and pharmacological effects caused by ligand-target interactions can be studied and assessed in specifically designed bio-assays. The huge range of bio-assay literature cannot be covered within the scope of this brief treatise, but a few essential remarks are in place here. When searching for plant ligands that are not only effective but also selective, specific and reversible in their interactions, bio-assays should meet certain requirements.

Firstly, one or more appropriate assays should be selected or developed for initial screening of extracts. 'Appropriate' means rapid and simple to perform, and functional and specific in their goal. The functions of the biochemical factors involved under physiological and pathophysiological conditions must be clear. In other words, bio-assaying is a way to relate bioactivity to factors and conditions relevant to disorders and homeostasis. As a result, bio-assaying

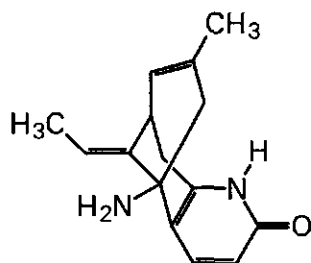


Figure 17. Structure of huperzine A.

Table 1. Examples of plant ligands with known target molecules.

Target category	Target biomolecule	Plant ligand
Enzymes	Acetylcholinesterase	Physostigmine
	Adenylate cyclase	Forskolin
	Cyclooxygenase	Salicylic acid
	Glycosidase I	Castanospermine
	Histone deacetylase	Butyric acid
	Mannosidase I	Deoxymannojirimycin
	Na <sup>+</sup> K <sup>+</sup> ATPase	Cardiac glycosides
	Trypanothione reductase	Kukoamine A
Receptors	Acetylcholine receptor	Hyoscyamine, atropin
	Adrenergic receptors	Ephedrine, cathinone
	– Agonists	Reserpine
	– Blocking	Morphine
	Opiate receptors PAF-receptor	Kadsurenone
Nucleic acids	DNA	Xanthotoxin
Other proteins and glycoproteins	Transcription factor AP-1 and NF-kappa B	Curcumin

Various sources.

should result in a closer understanding of a biological or pharmacological effect of a single compound. By giving an integral picture of interactions and effects, bio-assaying elucidates the pharmacological actions of plant extracts and their ethnomedical uses.

In the initial stage, *in vitro* testing should have priority over *in vivo* studies using laboratory animal models. Such a decision can be based on purely scientific as well as economic and ethical reasons. *In vivo* studies may be preferable at later stages of research, but this depends on the amount and nature of evidence of bioactivity already collected by means of *in vitro* studies, and the quest for additional information under life conditions. A bioactive plant compound and a bioactive plant preparation that are candidates for therapeutic application will still have to undergo extensive clinical and toxicological screening programmes before they can be registered as medicines.

Medicinal plant material is screened for bioactive compounds for many different considerations and using a variety of approaches ranging from selecting plant sources randomly to more systematic approaches. Some background features of screening approaches are listed below (Bierer et al., 1996; Colegate & Molyneux, 1993; Sills, 1996):

- *Phytochemistry directed screening approaches.* The typical feature of such approaches is that isolation and structure elucidation studies always precede work on biological activity as two unconnected experimental stages. The focus may be on a specific class of compounds (e.g. alkaloids), or a specific subgroup of compounds, or even a subgroup within a certain plant family or genus. Such approaches may have been inspired by pharmacological data on

related substances or by ethnomedical data, but usually there is no direct correlation with the research approach.

- *Ethnomedicine and ethnobotany directed screening approaches.* These follow up clues on bioactivity that have been derived from evaluative studies on traditional medicine and folkloric practices. In the set-up, the choices made for bio-assaying are strongly influenced by pharmaceutical, medical, health and cultural considerations.
- *Randomized screening approaches.* The most characteristic feature of these approaches is the absence of any clues. The plant material is simply a carrier of potential bioactive substances. Randomly selected and collected plant samples are extracted according to general protocols, and subjected to specialized bio-assays.
- *Integral screening approach.* Basically the bioactivity of a plant ligand should be seen and explored within the full context of its phenomenal existence, i.e. its biology, biochemistry, molecular biology, chemistry and biophysics. The approach is based on all life-science aspects and on the biocultural empirical experience with a plant source. A disease, a medical indication or an illness is usually the starting-point for the bio-assay. From the pathophysiology of a disease the relevant *in vitro* bio-assays and *in vivo* models are derived and refined. Furthermore, the experimental part of the bio-assay involves bioactivity-guided fractionation and isolation. This method represents a very appropriate and rational link between the detection of bioactivity and phytochemical methods.

#### 1.5.2.2 Common pharmacological screening methods

There are many types of pharmacological screens, most of which have to be carried out in a well-equipped laboratory. There are screens for specific bacteria, fungi, protozoa, intestinal worms, viruses and spirochaetes. The efficacy of compounds against specific health problems such as cancer and inflammation is also often probed, and the effect on various physiological and anatomical systems such as reproduction, digestion and circulation can be judged. The brine shrimp screen, the antibacterial screen, the brewers' yeast screen and the Hippocratic screen are commonly applied simple techniques.

Brine shrimps are small aquatic animals that can be grown in solutions resembling seawater. In order to test the potential toxicity of a plant - and thus its probability of containing an anti-cancer agent - measured amounts of plant extract are added to containers holding known numbers of brine shrimps. The surviving brine shrimps are counted after 6 hours and 24 hours, and the acute and chronic LD<sub>50</sub> values are calculated, respectively; this corresponds to the concentration of the compound in solution that kills 50% of the brine shrimps.

Bacteria can be grown on agar medium in Petri dishes. When measured amounts of a plant extract are placed on paper disks set on the surface of the bacteria-inoculated agar under sterile conditions, after 18–24 hours bacteria-free circles can be observed around some of the paper disks, indicating that the extract has inhibited the microbes. Plant extracts can be tested for phototoxic and fungicidal activity against brewers' yeast (*Saccharomyces cerevisiae*).

These tests yield much less information than the sophisticated assays that are



done in fully equipped laboratories, and can only detect a limited range of biological activities (Martin, 1995).

Hippocratic screening is a simple observational technique. Only crude plant material is used. Dried plant material is chopped and run through a mill. The resulting powder is suspended by trituration in a sterile 0.25% agar solution in double-distilled water, and injected intraperitoneally into rats. Only rats can be used for Hippocratic screening of crude natural products because they effectively resist both infection and peritonitis. A log-dose series of injections is made, ranging from 1 g/kg downwards, and the presence or absence of a large number of symptoms is recorded within certain intervals. The result is a unique pharmacological 'fingerprint' for each class of drug. Virtually all known drug types can be detected as active by the Hippocratic screen conducted in rats, with the exception of the various chemotherapeutic drugs such as antibiotics (Malone, 1981).

### *1.5.3 Surveys of bioactivity, pharmacological and therapeutic categories*

There is a need for comprehensive surveys to cope with the enormous expansion of information on diverse types of bioactivity and novel bioactive structures. There are published surveys on ligand-target interactions, in pharmacological categories and at the level of pharmacotherapeutical grouping. Examples of ligand-target categories are: inhibitors of HIV-1 reverse transcriptase, inhibitors of acetylcholinesterase, inhibitors of protein kinases and inhibitors of glycosidases.

Examples of pharmacological categories are: agents acting at synaptic and neuroeffector junctional sites, agents acting on the central nervous system, agents affecting renal and cardiovascular function and agents interfering with inflammatory processes.

Examples of pharmacotherapeutic groups are: emetics, analgesics, anti-inflammatories, anaesthetics, anti-cancer drugs, psychoactive drugs and anti-AIDS drugs.

Table 2 lists some plant compounds with their bioactivity categories.

### *1.5.4 Future developments in research on bioactivity*

Future advances in plant drug research will provide information on bioactivity in terms of molecular interactions with target biopolymers on a broad scale, all this within the context of homeostasis and pathophysiological conditions. Developments in the fields of genetics, molecular biology, bioinformatics and techniques used in determining the steric structure of plant metabolites and target macromolecules appear important today. In addition, fundamental understanding of molecular biodiversity seems important in the process of using plants as resources for drug development. Some animal models will be replaced by testing on cell cultures, because new techniques of culturing cells and tissues have become available. These new testing methods require smaller amounts of test compounds and will provide information at cellular level. This will lead to extensive studies on medicinal plants.

Table 2. Examples of plant compounds with their bioactivity.

Plant source (origin)	Compound name	Bioactivity pharmacological therapeutic category
<i>Areca catechu</i> L. (Asia)	Arecoline	Cholinergic agonist, veterinary anthelmintic
<i>Andrographis paniculata</i> Nees (Asia)	Andrographolide	Antibacterial
<i>Curcuma longa</i> L. (Asia)	Curcumin	Choleretic, anti-oxidant, anti-inflammatory, apoptosis inducer, inhibitor of angiogenic differentiation
<i>Hypericum perforatum</i> L. (Europe)	Hypericin	Protein kinase C-inhibitor, photodynamic antiviral agent
<i>Phyllanthus myrtifolius</i> Moon (Sri Lanka)	Phyllamycin B	HIV-1 RT inhibitor
<i>Phyllanthus myrtifolius</i> Moon (Sri Lanka)	Retrojusticidin B	HIV-1 RT inhibitor
<i>Physostigma venenosum</i> Balf. (Africa)	Physostigmine	AChE inhibitor, parasympathomimetic, miotic in primary glaucoma
<i>Picrorhiza kurroa</i> Benth. (Asia)	Apocynin	Multiple-immunomodulator, inhibition of: <ul style="list-style-type: none"> <li>- STZ-induced ROS</li> <li>- platelet aggregation</li> <li>- TNF<math>\alpha</math> release</li> </ul>
<i>Pilocarpus jaborandi</i> Holmes (South America)	Pilocarpine	Cholinergic agonist, miotic in glaucoma
<i>Zanthoxylum zanthoxyloides</i> (Lamk) B. Zepernick & F.K. Timler (Africa)	Fagaronine	HIV-1 RT inhibitor

Various sources.

## 1.6 Botany

### 1.6.1 Plants used in medicine

The World Health Organization has compiled a list of more than 21 000 plant species purportedly used globally in medicine. It is estimated that 2000–3000 species are used for medicinal purposes in South-East Asia. The number of medicinal plants in Indonesia is estimated at 1000, out of a total flora of 28 000 species. In Malaysia, approximately 1200 trees, shrubs and herbs of the about 12 000 species have been reported to have traditional medicinal properties (Soepadmo, 1991). The number of medicinal plant species in Philippine hand-

books (e.g. Quisumbing, 1978) is about 850, out of a total number of higher plant species in the Philippines roughly estimated as 8000. It is estimated that there are more than 10 000 plant species in Thailand, of which about 1800 are listed as medicinal in the Thai Traditional Materia Medica. However, only 1100 of these are botanically identified. Over 1800 plant species have been identified in Vietnam as useful for medicinal purposes. Medicinal plants are also numerous in adjacent regions. In India, the number of plant species used in traditional medical systems is estimated at 1100–1500, and about 700 species of medicinal plants grow wild in Nepal. An estimated 300 plant species are used in traditional medicine in Pakistan, and about 550 flowering plants in Sri Lanka.

### 1.6.2 Weedy and forest species

Approximately half (125000) of the world's flowering plant species live in tropical forests. The tropical rain forests continue to support a vast reservoir of potential drug species. They can provide natural product chemists with invaluable compounds or starting points for developing new drugs. Less than 1% (Balick et al., 1996) of tropical species have been studied for their pharmaceutical potential; this proportion is even lower for species confined to the tropical rain forest. To date, about 50 major drugs have come from tropical plants. The existence of undiscovered pharmaceuticals for modern medicine has often been cited as one of the most important reasons for protecting tropical forests, so the high annual extinction rate of an estimated 3000 plant species is a matter of great concern.

It is notable that the more important medicinal and poisonous plants include many weedy species. It seems most likely that this is because these species are so widely distributed and common that they are the most obvious plants to be tried for medicinal purposes. Also, their toxic effects manifest comparatively easily because of the presence of livestock. The weeds include species that are pantropical or even cosmopolitan (e.g. *Achillea millefolium* L.), and that therefore occur in regions where research on medicinal plants is more common than in South-East Asia (e.g. in India, China and Europe). There is thus much literature on these species and they are more highly valued.

More advanced defence mechanisms by secondary metabolites to prevent browsing by livestock might also account for the high proportion of weedy species among the more important medicinal plants.

### 1.6.3 Chemotaxonomy

The medicinal and poisonous plants in South-East Asia form an extremely diverse group taxonomically. Some families are, however, comparatively rich in species used medicinally, usually because of the common occurrence of certain types or classes of chemical compounds. Examples are *Apocynaceae* and *Menispermaceae* with their alkaloids, and *Compositae* and *Umbelliferae* that contain essential oils.

The great diversity in the taxonomy of medicinal and poisonous plants is also reflected in the growth forms, which range from small herbs to large trees, and in life cycles, ranging from annuals (e.g. *Artemisia annua* L.) to slow-growing trees (e.g. *Cinchona* spp.).

No study of medicinal plants can be started without a proper botanical identifi-

cation of the species. As there is a relationship between taxonomy and the chemical profile, taxonomic botany is important when attempts are made to find a species that yields a desired substance. A certain compound (or a compound close to it) is more likely to be present in a species related to a species known to contain this compound. Taxonomic studies can therefore help predict the presence of active substances in certain groups of plants (Hedberg in Leeuwenberg, 1987). Constraints are the lack of herbaria and qualified botanists in many tropical countries.

It is essential to use the correct scientific name of medicinal plants. Vernacular names are often very confusing. Erroneous namings are quite common. It should be compulsory for a voucher specimen to be deposited in a public herbarium accompanying a scientific publication on medicinal plants or their compounds.

Plants to be evaluated for medicinal properties (e.g. for anti-cancer activity, as has been done for a long time by the National Cancer Institute, United States) should cover a wide taxonomic range as this will provide a great diversity of types of chemical structures, thereby increasing the likelihood of finding active compounds.

### 1.7 Ecology

Temperature, rainfall, photoperiod and altitude are factors of great importance for the development of plants. The day length may considerably influence the growth and yield of constituents. *Mentha x piperita* L., for instance, does not grow well under short-day conditions. Most medicinal plants in cultivation need plenty of sun to be of high quality. Most aromatic herbs do not tolerate more than half-day shade, and for these plants the sunniest sites should be chosen in somewhat cool and cloudy climates, though a shady site is acceptable in sunnier climates. Altitude may have a definite effect on growth and on yield of constituents. For example, *Cinchona*, grows slowly at altitudes above 2000 m, and the quinine yield is low when cultivated at altitudes below 800 m. A number of medicinal plants thrive best at high altitudes, and some species only flower at higher elevations and never in the lowland.

Many medicinal plants are easy to grow because they are tolerant of and adaptable to poor conditions. However, unfavourable climatic conditions can cause the synthesis and accumulation of excessive levels of undesirable compounds and/or low levels of desirable compounds. In oil-yielding plants, for instance, poor oil quality may result. Alkaloid yields are affected by environmental factors such as altitude, temperature, moisture, light, soil type, together with handling after harvest and genetic factors. In many alkaloid-yielding plants (e.g. *Datura metel* L.), the alkaloid content peaks in the dry season and is lowest in the rainy season.

The soil requirements vary with the species. Certain species, like most *Zingiberaceae*, prefer loose, moist and humus-rich soils with a pH of 6–7.5, while others e.g. most *Labiatae*, thrive best in dry, well-drained and sandy soils enriched with organic matter. Most medicinal plants are intolerant of water-logging. In general, alluvial and clayey soils that are slightly acid and have a good water-retaining capacity are suitable for growing medicinal plants in regions with not too much rain.

## 1.8 Agronomy

### 1.8.1 Production systems

Medicinal plants are often collected from the wild. Many medicinal plants, especially the aromatic herbs, are grown in home gardens but some are cultivated as field crops, either in sole cropping or in intercropping systems, and rarely as plantation crops.

#### 1.8.1.1 Collection of medicinal plants from the wild

In most countries, the collection of medicinal plants from wild sources is still the rule. Although the natural flora has been used as the major source of medicinal plants throughout history, it is neither possible nor desirable to base a medium-scale industry on it. Attempts to do this have caused depletion of species and even eradication. Furthermore, the quality of raw material obtained from spontaneous growth may vary considerably with respect to their constituents. It is possible to use the wild flora in a way that enhances and preserves the plant resources, but this requires strict regulations and control. Such use must be preceded by an evaluation of the abundance of selected species within a large area, resulting in recommendations about how much raw material may be harvested in a certain area. Local people often transplant and cultivate wild medicinal plants in and around their homes and villages.

#### 1.8.1.2 Cultivation of medicinal and poisonous plants

Several countries in South-East Asia including Indonesia, the Philippines, Vietnam and Thailand grow medicinal crops for domestic use and for export. The number of medicinal crops grown varies from country to country. Although cropping of medicinal and poisonous plants is common practice, it is usually on a small scale. Large-scale cultivation is dictated by the requirements of the pharmaceutical industry, the main user of the raw material.

The demand for medicinal plant material is fickle; large-scale cultivation often brings down the price, and the market situation often changes drastically when the industry's search for cheaper alternative source materials for drugs is successful. The great variation in the demand and supply have acted as a damper to developing efficient crop production systems. Moreover, many medicinal plants are labour-intensive in propagation, husbandry, harvesting, post-harvest processing and packing. On the other hand, pharmaceuticals are often commodities of high value and low bulk, which makes them attractive crops for small-holder farmers in communities where transport constraints restrict bulky cash crops.

South-East Asia has a long practice of traditional farming systems, but these have not been developed to include medicinal and poisonous crops. Commercial cultivation of medicinal crops has to be based on a sound scientific footing. However, research carried out on cropping systems including medicinal crops is rather limited in South-East Asia, whereas very little scientific information is available on the medicinal crops themselves. Farmers in India have long grown complementary crops to derive maximum benefits of existing soil mois-

ture and nutrients. In cropping systems based on opium poppy in India, groundnut and black gram proved a profitable combination which is recommended to the growers. The results of trials with *Rauvolfia serpentina* (L.) Benth. ex Kurz in India showed that the yield was highest when grown as a sole crop and that intercropping often depressed yields considerably. However, it is reported that vegetable crops, soya bean, garlic and onion have little effect on root yields of *Rauvolfia* and add to the overall economy of the system.

There is extensive experience with the cultivation of *Cinchona* in Indonesia, although this crop has declined in importance since the Second World War. The cropping system practised is a long-term one, with a cutting cycle of 7–8 years, leaving a coppice to produce new shoots. Leguminous cover crops are sometimes planted in between the *Cinchona* rows or on the contour to prevent erosion, but *Cinchona* may also be cultivated under the shade of spared rain forest trees.

The success of a new crop in a cropping system depends upon its growing period, soil nutrient and irrigation requirements, disease and pest tolerance or resistance, and its yield. For example, mint (*Mentha* spp.) and opium poppy (*Papaver somniferum* L.) make heavy demands on irrigation and fertilizers, whereas psyllium (*Plantago* spp.) and periwinkle (*Catharanthus roseus* (L.) G. Don) need light irrigation and less fertilizer. *Cinchona* produces an economic yield from 7 years after planting onward, whereas jasmine (*Jasminum*), also a perennial crop, starts yielding already from the second year onward, and *Rauvolfia* after 18 months. Some species are grown as annual crops, e.g. opium poppy, mint and psyllium. These annuals may rather easily fit into existing crop rotations and modes of cultivation.

### 1.8.2 Propagation

Several species are mostly propagated by seed. Examples are found amongst commercially important crops like hemp (*Cannabis sativa* L.), coca (*Erythroxylum novogranatense* (Morris) Hieron.) and opium poppy (*Papaver somniferum* L.). Disadvantages of this method of propagation may be the great genetic diversity of the progeny and the rapid decrease in seed viability that sometimes occurs. Some species, e.g. *Mentha arvensis* L. and *Erythroxylum coca* Lamk, are commonly propagated by cuttings. Rooting can be stimulated by application of growth regulators. *Bryophyllum pinnatum* (Lamk) Oken is propagated from foliar embryos. Although in vitro propagation techniques are commonly used for ornamentals, they are still rarely used for medicinal crops, though they may be very advantageous in providing homogeneous plant material.

### 1.8.3 Husbandry

Most medicinal crops are grown in gardens, but sometimes they are grown in pots. Certain cropping techniques can considerably influence the yield of both dry matter and pharmacologically active constituents. The quantity and quality of chemical fertilizers, for instance, may influence the content of secondary metabolites in plants. For example in *Datura stramonium* L., where the alkaloid biosynthesis is increased by replacing part of the  $\text{NO}_3^-$  in fertilizer by  $\text{NH}_4^+$  (Demeyer & Dejaegre, 1993).

Fertilizers and pesticides may contaminate crops. Therefore, manure should preferably be applied before planting short-duration crops, because harvested plant material may be contaminated with bacteria if it is applied later. Chemical fertilizers are usually not used when growing medicinal crops, because plants usually take up too much sodium and potassium, resulting in excessive concentrations of these elements in decoctions. Very often, compost is used. Biological pest control by companion planting (e.g. with *Tagetes*) is often practised.

## **1.9 Harvesting and handling after harvest**

### *1.9.1 Harvesting*

Medicinal plants, especially those growing in the wild, are often harvested by hand. Even when cultivated, manual harvesting is often more practical, e.g. for harvesting bark and fruits that do not mature simultaneously (e.g. *Senna* spp.). Leafy plant material is often labour-intensive to harvest; it is easier to harvest it from cultivated plants, as the individual plants are in approximately the same stage of development and grow close together in smaller areas.

The amount of a constituent is usually not constant throughout the life cycle of a plant. Therefore, the stage at which a plant is harvested is very important for the yield of the desired constituent. There may be seasonal variations, but in perennial plants the age may also be important. It is generally assumed that the best time for harvesting is when the organ in question has reached its optimal state of development. Roots and rhizomes are usually collected at the end of the growing period, bark often at the beginning of the growing season, when it is easier to strip because of the abundance of soft cells near the cambium, leaves before flowering, flowers at anthesis, and fruits and seeds when fully ripe.

Uncontrolled stripping of bark will easily destroy trees, and in several cases has seriously threatened the diversity and abundance of species. It is therefore important, but not easy, to harvest bark in a sustainable non-destructive fashion.

### *1.9.2 Drying and cleaning*

After harvest and cleaning, usually by washing, some plant materials have to be dried in the sun, others in the shade. Flowers are usually dried immediately after picking, in the shade; they are regularly turned over to prevent browning. Drying is the most common method of preserving plant material. Rapid removal of moisture largely prevents degradation of the constituents, since enzymatic processes require the presence of water. Drying also lessens the risk of external attack, e.g. by moulds (Samuelsson, 1992). In some extreme cases, soaking in ethanol is required to deactivate the enzyme. The desired constituents are often damaged by heat, so it is often advantageous to dry at moderate temperatures (45–50°C). The best method of drying depends on the plant material, and uncontrolled drying may cause severe loss of quality. Irradiation is sometimes practised in manufacturing to avoid contamination by bacteria and fungi. The most efficient drying is achieved in large driers of the tunnel

type. The material is spread out on trays placed on mobile racks and transported into a tunnel where they meet a stream of air. However, most often plant material is simply dried in the open. Mechanical driers are used for bulk operation. Sizeable losses still occur in drying and subsequent post-harvest handling.

Most materials have to be crushed or ground, coarsely or finely, before they can be loaded into an extractor or distillation vessel. Most extracts will be dried by distillation under vacuum or spray drying. Freeze-drying (lyophilization) is an adequate method for drying water extracts containing heat-sensitive substances such as antibiotics and proteins, but it requires relatively complicated and expensive apparatus.

### *1.9.3 Storage conditions*

Raw material in the form of dried plant material can be stored only for a limited period of time and provided certain requirements are met. Storage time can be minimized when the processing is planned in such a way that the harvested raw material is used as soon as possible. Preparation of material for distillation varies with the properties of the material. Some materials like flowers should be distilled immediately after harvesting, whereas others such as foliage parts are best stored for some days before distilling. Some materials can be stored indefinitely before distillation.

There are great differences in the stability of crude drugs. Drugs containing glycosides, esters and essential oils are usually less stable than those containing alkaloids and tannins (Samuelsson, 1992). Optimal storage conditions must be employed to prevent deterioration. The enzymes able to break down constituents are rendered inactive in properly dried plant material, but they may become active again if not well protected from moisture during storage. Humidity should therefore be controlled, and wilted leafy plant material should be kept dry and cool to prevent fermentation or mould growth. A concrete floor under shade is often used. The moisture content of the material should stay under 10%. To avoid insect and fungal attacks, the material is often redried in the sun. In order to reduce undesirable microbial contamination and to prevent the development of other organisms, some plant materials are sterilized before storage. Ethylene oxide or methyl chloride may be used, and drugs so treated should comply with an acceptable limit for toxic residues.

The storage of end products such as extracts, also requires care. Storage vessels should be well cleaned prior to being filled. Sometimes it is necessary to fill the vessel to capacity to prevent oxidation, and to run an inert gas over the top to eliminate traces of oxygen. Post-processing oxidation of essential oils is a common problem to be avoided by exclusion of air, trace metals and sunlight. Light-sensitive products such as essential oils are stored in vessels in the dark. Under suitable conditions, however, most essential oils can be stored for long periods. Larger quantities are stored in metal drums lined with polyethylene. It is important that the essential oil is not in contact with rubber or plastic because chemical contamination may occur.



## 1.10 Processing, utilization and quality control

### 1.10.1 Extraction methods

Active constituents can be extracted from the plant material by maceration, percolation or continuous extraction. Extraction is the first step in isolating the desired constituents from plant material, using a solvent. Sometimes it is sufficient to achieve an equilibrium of concentration between drug components and the solution within set limits, as in the case of tinctures, tisanes, decoctions and teas. In other cases the drug is extracted to exhaustion, i.e. until all solvent extractables are removed by the solvent. The latter method is mostly used in industry.

In all industrial procedures the raw material is pretreated with solvent outside the extractor, preventing sudden changes in bulk and accelerating the penetration of the solvent through the cell walls to release the extractables. To facilitate extraction, the solvent should diffuse inside the cell and the desired substance must be sufficiently soluble in the solvent. The ideal solvent for extraction is one in which the extractive is most soluble and selective, so that the desired constituent will be extracted with minimum impurities. Alcohol is often used, but because of its great extractive power it is often the least selective in that it extracts all soluble constituents. The ratio of alcohol and water used varies, depending on the polarity of the active compounds.

Most alkaloids can easily be extracted with organic solvent after the powdered drug has been mixed with water and alkali. The alkali will liberate the alkaloid from its salts. However, some volatile alkaloids, protoalkaloids and quaternary ammonium alkaloids should not be extracted by this method. Even though the alkaloids are soluble in acids, the use of acids is not appropriate for industry because of the large volumes required for exhaustive extraction. Some herbs are extracted with volatile organic solvents to produce oleoresins.

The equipment used for extraction with solvents comprises the following components (Wijesekera in Chomchalow & Henle, 1993):

- An extraction vessel with a heating jacket for steam heating or fitted with electrical devices.
- A condenser in a reflux position.
- A solvent reservoir.
- A facility to convert to reboiler position or a separate reboiler.
- A short column for solvent recovery.

There are 3 basic types of essential oil distillation:

- Water distillation (hydrodistillation).
- Wet steam (water and steam) distillation.
- Dry steam (steam) distillation.

Stills of the first type are the simplest and are used by small producers. The plant material is immersed in boiling water. Steam distillation is an improved method to avoid prolonged contact of the material with heat. The still contains a grid which keeps the plant material above the water level (wet steam distillation) or steam is provided from a separate boiler (dry steam distillation). Stills should be insulated to reduce heat losses.

The passage of steam through plant material causes volatile oil to distil over with the steam. The compounds can be distilled out of the plant material at

around 100°C. When the condensate cools through a condenser, the oil, dispersed in water, separates from the aqueous phase, forming two layers which can then be separated easily. It is important that the separator has a large volume to minimize turbulence, because significant amounts of oil can be lost with the distillate water if the oil is not allowed to separate completely. The best material for stills, condensers and separators is stainless steel. The method of distillation chosen must be suited to the particular essential oil, and has to be determined experimentally.

Hydrodiffusion is another, more recent process in which low-temperature, low-pressure steam is used to extract the essential oils. Essential oils from the more fragile flower material can be obtained by enfleurage, a process in which successive batches of freshly picked flowers are exposed to layers of grease coated on stacked glass plates, and finally the resulting pomade is extracted, usually with alcohol, to obtain absolute. This is an almost superseded method practised in the perfume industry, for flowers that continue to produce aroma compounds for several days after they have been picked. At present it is only used for the most expensive perfumes. See Prosea 19: 'Essential-oil plants' for more detailed information on essential oil distillation.

### *1.10.2 New industrial standards*

The practice in industry is to judge plants according to their content of important constituents, but the extent to which the desired secondary metabolites are technically and biologically exploitable is also a governing consideration. Tissue culture techniques have much to offer. They can produce more homogeneous plant material and are also of interest for the industrial production of plant-derived natural substances, including drugs. The main reason why plant tissue and cell cultures have not yet become important for the production of pharmacologically active natural products is the high cost of producing the desired substances by this method. The breakthrough will depend on basic research in molecular biology to clarify how the plants regulate the formation of secondary metabolites and how this is connected with the development of organs (Samuelsson, 1992). Often, high-yielding cell lines are selected, taking advantage of the somaclonal variation. This requires rapid and sensitive methods for analysing the desired metabolites.

Standards are available for several herbal products and guidelines specifying the requirements have been formulated by individual countries, the World Health Organization (WHO) and United Nations Industrial Development Organization (UNIDO). One of the key points on which the medicinal plant-based industry differs from any other agro-industry is the requirement of sophisticated facilities for chemical analyses at all stages. Quality control and assessment are needed, and analytical research has to go hand in hand with plant breeding. A range of quick, reliable and acceptable methods is available, and the institutions dealing with medicinal plants need to have a well-equipped laboratory.

For the quality control of crude drugs the identity of the crude drug must be known, and also the content of active constituents and impurities. Descriptions of the macro- and micromorphology of crude drugs are given in pharmacopoeias and handbooks. A laboratory carrying out the quality assurance of crude drugs should have a well-documented collection of reference materials.

### 1.10.3 Household preparation

In some countries of South-East Asia, standard pharmaceutical methods have been modified to enable herbal medicines to be made without sophisticated and expensive laboratory equipment. The materials employed in the procedures are those found in rural kitchens. In the Philippines, for instance, these methods of preparation, referred to as 'kitchen technologies', are being taught nationwide and are part of the health care system. The most common preparations made are infusions or teas, decoctions, syrups, liniments, ointments, pills, herbal soaps and lozenges.

The following general guidelines are given for household preparations of medicinal plants in the Philippines:

- Be sure of the identity of the plant.
- Use only one plant drug at a time.
- Use only the recommended plant parts.
- Collect only those plant parts that look healthy: no insect damage, discoloration or other signs of abnormality.
- Follow the recommended methods of preparation.
- When using dried drugs, use only half the amount prescribed for fresh plant drugs.
- Infusions and decoctions should be freshly prepared. A dose for one day may be prepared and kept in a thermos flask.
- Use containers made of inert materials; for cooking: earthenware pots, enamel-lined, pyrex, not metallic utensils.
- Sterilization of medicine bottles is very important. This may be done by heating the bottles, caps or bottled products in a double boiler for at least 20 minutes.
- Observe care and cleanliness at all times.

An example of 'kitchen technology' as described in the Philippines is a sweetened preparation from the leaves of *Vitex negundo* L. called lagundi syrup, and is indicated below:

- Materials: cooking pot, ladle, cup, stove, strainer, medicine bottles, labels, lagundi leaves, sugar/honey, water.
- Proportion: 1 cup chopped lagundi leaves to 2 cups water.
- Procedure: (1) Prepare a decoction by boiling the leaves in water in an uncovered pot for 20 minutes or until the water has decreased to half of the original volume. (2) Cool and strain. (3) Measure the amount of decoction produced. One-third of this volume is the amount of sugar/honey to be added. (4) Add the sweetener, stirring gently. The mixture can be put back on the stove, on low heat, until all the sweetener is dissolved/blended with the mixture. This is the syrup. (5) Transfer the syrup to the sterilized medicine bottles. Seal and label properly. (6) Store the bottled syrup in a clean, cool, dry place away from light.

## 1.11 Genetic resources and breeding

The need for a comprehensive inventory of medicinal plants is felt in countries where such plants are starting to play a role in the primary health care system. In most countries in South-East Asia the extent of genetic erosion is being in-

ventoried, but this is not being done as well as it should be, because of the huge cost involved.

#### *1.11.1 Plant diversity and conservation*

Several species occurring in primary forest are rare and endangered because of large-scale forest destruction and/or over-collecting. Forest destruction may easily endanger species with a narrow area of distribution and may result in genetic erosion. The discovery and use of plants by the herbal and pharmaceutical industries often lead to degradation of the resources. Often both habitat loss and over-harvesting reinforce each other as synergetic factors contributing to the species's overall endangered status. However, in only a few cases is it known whether certain medicinal plants are already or potentially endangered (Lange & Schippmann, 1997). It has been necessary to protect some species, e.g. *Rauvolfia serpentina* (L.) Benth. ex Kurz and *Aloe* spp., by including them in the Appendices of the Convention on International Trade in Endangered Species of Wild Fauna and Flora (CITES). Several species with a large area of distribution (e.g. *Acorus calamus* L.) need not be considered as at risk from genetic erosion; others have a remarkable capacity to regenerate.

Although medicinal plants have been cultivated for centuries, their germplasm collection has been very limited. There are no germplasm collections for most species and very little effort has been made towards conserving the genetic variation. Gene banks are a way of conserving genetic diversity, but they generally cover major food crops and include hardly any medicinal plants. Gene banks of medicinal and poisonous plants with limited collections (up to 500 accessions) are listed for most of the countries in the region, but the coverage of the geographical and botanical diversity is far from complete. In situ conservation of valuable species in natural parks and reserves and in botanical gardens is too little focused on medicinal plants. In the few important medicinal plant gardens in South-East Asia, the diversity of species has mostly decreased in the last 30 years.

#### *1.11.2 Breeding*

Unlike other commercial crops, medicinal crops continue to be cultivated in the same way as they were hundreds or sometimes thousands of years ago, with few exceptions. Very little has been done to genetically improve these crops, despite their long history of domestication. There seems to be great potential for improving the yield and quality of medicinal crops. Breeding work on medicinal crops whose wild forms have great genetic variability often leads to spectacular successes, even in the first few cycles of selection. Therapeutic value and yield are important criteria for selection and breeding. For industrial-scale production of raw material it is vital to select cultivars with the required characteristics.

Research on breeding of medicinal crops thus lags far behind that of food crops and other commodities. The objectives of breeding in medicinal crops are to obtain a high yield of active constituents, to improve adaptability and quality, and also to obtain resistance to diseases and pests and stress tolerance. The trend in several countries towards using plants in primary health care in-

creases the need for improvement of crops through breeding. However, for many species there is still no information available.

Examples of existing breeding programmes include:

- The development of genotypes of *Rauwolfia* with a short maturation period and a high root yield for the production of reserpine and ajmalicine.
- The development of genotypes of *Catharanthus roseus* (L.) G. Don with a high root yield and total alkaloid content.

Biotechnology, plant cell culture and molecular techniques are important for the improvement of medicinal plants. For many medicinal species, complete plants have been regenerated from callus cultures, excised plant organs and isolated protoplasts, whereas selections have been made for cell lines with high alkaloid content. Natural or artificially induced mutations have been used to develop plants that produce desired types of alkaloids (e.g. in *Datura* and *Papaver*). Sophisticated hybridization techniques have been applied to several plants of pharmaceutical interest, for the purpose of combining certain desirable characteristics or for producing entirely new characteristics not found in either parent. Gene transfer is possible from related wild species to cultivated plants.

The most profitable system for the synthesis of secondary metabolites is plant cell suspension culture, but the yields of medicinally important alkaloids are lower than in whole plants grown in the field. Hairy root cultures are promising, but quite expensive. Genetic transformation may lead to better in vitro production of secondary metabolites.

## **1.12 Research and development**

### *1.12.1 Main research topics*

Research priorities in South-East Asia for medicinal and poisonous plants are:

- Ethnobotanical research on traditionally used plants.
- Agronomy and commercialization of traditional medicinal plants.
- Medical, biological, microbiological and biochemical screening and standardization.
- Issues related to the use, conservation and socio-economic aspects of traditionally used medicinal plants.
- Legislation and management locally, regionally, nationally and internationally.

### *1.12.2 Main institutions*

The main institutes and universities conducting research on medicinal and poisonous plants in the respective countries in South-East Asia are:

#### *Indonesia*

- Agency for Development and Application of Technology (Badan Pengkajian dan Penerapan Teknologi), Serpong
- Central Institute for Research and Development of Agrobased Industry (Balai Besar Penelitian dan Pengembangan Industri Hasil Pertanian), Bogor

- Research and Development Centre for Biology (Pusat Penelitian dan Pengembangan Biologi), Bogor
- Research and Development Centre for Industrial Crops (Pusat Penelitian dan Pengembangan Tanaman Industri), Bogor
- Research and Development Centre for Pharmacy (Pusat Penelitian dan Pengembangan Farmasi), Jakarta
- Research Institute for Spices and Medicinal Crops (RISMRC), Bogor (including 14 experimental gardens)
- Research Institute for Veterinary Medicine, Bogor
- Tawangmangu Research Institute for Medicinal Crops (Balai Penelitian Tanaman Obat Tawangmangu), Surakarta
- various academic institutions throughout Indonesia including Institut Teknologi Bandung, Universitas Airlangga (Surabaya), Universitas Gadjah Mada (Yogyakarta), Universitas Jenderal Sudirman (Purwokerto), Universitas Udayana (Denpasar).

### *Malaysia*

- Forest Research Institute Malaysia (FRIM), Kepong
- Malaysian Agricultural Research and Development Institute (MARDI), Serdang
- Universiti Putra Malaysia (UPM), Serdang
- University of Malaya (UM), Kuala Lumpur
- Universiti Kebangsaan Malaysia (UKM), Bangi
- Universiti Sains Malaysia (USM), Penang

### *Papua New Guinea*

- Wau Ecology Institute, Wau
- University of Papua New Guinea, Port Moresby

### *The Philippines*

- University of the Philippines System: various institutes, colleges and departments at UP Los Baños, UP Manila and UP Diliman, Quezon City
- Department of Science and Technology: Philippine Council for Health Research and Development (PCHRD), Philippine Council on Agriculture and Forestry Resources Research and Development (PCARRD), Integrated Technology Development Institute (ITDI), Forest Products Research and Development Institute (FPRDI), Food and Nutrition Research Institute (FNRI), National Research Council of the Philippines (NRCP)
- Department of Health: Traditional Medicine Unit (TRADMED), Philippine Institute for Traditional and Alternative Health Care (PITAHAC), Bureau of Food and Drugs (BFAD), Institute of Tropical Health (ITM)
- various academic institutions including University of Santo Tomas and De la Salle University in Manila, Philippine Institute for Pure and Applied Chemistry and Ateneo de Manila University in Quezon City

*Thailand*

- Department of Agriculture: research stations at Chiang Rai, Chiang Mai, Chanthaburi, Chumphon, Ubon Ratchathani, Chai Nat, Suphan Buri
- Department of Medical Sciences: botanical garden at Chanthaburi
- Royal Forestry Department: Phu Khae Botanic Gardens and various National Parks
- Thailand Institute of Scientific and Technological Research (TISTR), Bangkok
- various academic institutions, in particular Mahidol University, Faculty of Pharmacy, Bangkok

*Vietnam*

- Institute Materia Medica, Hanoi
- Institute of Natural Products Chemistry, Hanoi
- Institute of Chemistry, Ho Chi Minh City
- Ministry of Health, Hanoi
- Science Production Centre of Vietnamese Ginseng, Ho Chi Minh City
- Institute of Tropical Biology, Ho Chi Minh City

**1.13 From plant to drug**

Since the 1950s, the pharmaceutical industry has relied primarily on new synthesized compounds, with the exception of most antibiotics which are derived from micro-organisms. There was almost no interest in using plants for drug development, but in the last few years that situation has changed slowly.

Plants are still an overwhelming source of novel chemical structures, and substances within plants widely used by humans are less likely to be seriously toxic than synthetic chemical compounds. There is renewed recognition that traditional systems of medicine are appropriate starting points for the development of modern medicines. Another reason for a shift from synthetics to plant-derived products is the increased interest of the public in using medicines from plant sources.

When one or more active constituents have been isolated, studies are performed in animal species (rodents, other mammals) to investigate the mechanism of action. Acute and chronic toxicity studies are required, to ensure the safety of the drug. The most suitable preparation must be determined, i.e. one that provides the proper dose of the drug and is stable enough to be launched on the market. Then clinical studies are carried out. The first clinical phase deals with a small group of healthy subjects, to observe the efficacy and possible side-effects. In the second clinical phase the drug is tested on a small group of patients, and finally a complete clinical study is carried out. All this involves much time and money.

Drug discovery programmes have become less efficient, and the costs involved in developing a drug have escalated rapidly with the increasing requirements associated with the demonstration of the safety and efficacy of a compound. The minimum costs are US\$ 100 million (Horrobin & Lapinskas in Prendergast et al., 1998), but may amount to US\$ 2000 million. The high costs discour-

age companies from entering a drug development programme unless there is a fair chance that the returns will eventually be much higher and there is protection of intellectual property.

Although the patenting of new pharmaceutical uses of known compounds is now possible in many countries, new plant-source drugs often cannot be patent-protected. They may arise from traditional sources or from scientific publications, and in such cases marketing protection may be necessary to make the necessary investment attractive for companies to take the drug through the approval process. This is often in the form of a market monopoly for a period up to 10 years.

Important issues to be addressed to develop a plant source successfully as a pharmaceutical are:

- A chemical substance must be shown to be safe and to have a reasonable prospect of being effective. A problem arises because in many cases, the activity of isolated compounds is equated with the efficacy of the preparation without considering the possibly important modifying action of other drug constituents.
- A financial assessment should be made to compare the costs for developing the drug with the revenue expected from sale of the product.
- Where possible, it is usually preferable to prepare the compound concerned at reasonable cost synthetically. Where this is not possible, plants should be comparatively easy to cultivate and the plant should contain a reasonable amount of the compound, preferably in easily harvested parts such as leaves or seeds.

### 1.14 Prospects

The South-East Asian region abounds with medicinal and poisonous plants, many providing drugs for various therapeutic categories and having revolutionized medical science over the years. Accounts of positive effects of herbal preparations are no longer just folklore; they are backed by extensive scientific research, and many modern-day medicines have been derived from medicinal plants. More significant cures for major health problems remain to be discovered.

The sharing of benefits is at present a sensitive issue for the cooperation between drug-resource countries and drug-producing countries. Standard regulations should be set up by resource countries to gain benefit from commercial production of drugs from plant resources.

To harness the full potentials of medicinal plants research should focus not only on the validation of safety and efficacy, combined with development and conservation efforts, but also on chemistry, biological activity, formulation of drugs, clinical trials and cultivation technology. The need for state-of-the-art facilities, adequate funding for research and training of medical researchers are issues to be addressed. Concerted efforts along these lines are critical if long-term objectives are to improve the health of man and provide good health care to all.

There is worldwide concern about the side-effects produced by purified compounds and synthetic drugs. Dangerous side-effects of medicinal plants tend to be limited, but are often concentration-dependent. The long-continued use of



numerous medicinal plants with apparent positive effects and no evidence of detrimental side-effects validates their safety and efficacy and supports their position in the medical practice today. Furthermore, some optically active asymmetrical compounds cannot be synthesized chemically. Many plants are used as pesticides in farms, gardens and homes. People concerned with maintaining ecological balances prefer pesticides from plant products over synthetic formulations.

This implies that the prospects for medicinal and poisonous plants are promising. In fact, there is already a trend in South-East Asia towards a reevaluation of the use of medicinal plants in primary health care. In some countries, there are already initiatives to promote their use and to disseminate information on proper applications. In other countries, the recent economic crisis is forcing governments to seriously consider the use of medicinal plants. Sometimes regulations have to be modified to make it easier to register locally produced herbal drugs. At the same time, appropriate manufacturing practices should be introduced to assure the quality of such drugs.

It is envisaged that certain medicinal plants may become commercial crops. However, the demand for medicinal plant products is often not as large as expected and can change rapidly. Oversupply must be avoided by attuning supply to demand in and between countries.

It has been estimated that the higher plants in the world's tropical forests contain about 375 potential pharmaceuticals of which about 50 have already been discovered. It has been suggested that the complete collection and screening of all tropical forest species may cost about US\$ 3–4 billion to a private pharmaceutical company, and as much as US\$ 147 billion to society as a whole (Mendelsohn & Balick, 1995). The potential value of undiscovered drugs is an additional incentive to conserve species-rich tropical forests.

L.S. de Padua, N. Bunyaphratsara & R.H.M.J. Lemmens

with contributions from D.K. Holdsworth (traditional and modern medicine in South-East Asia: Borneo, New Guinea), R.P. Labadie (phytochemistry (introductory part), biological and pharmacological activity and therapeutic applications), Nguyen Tien Ban (traditional and modern medicine in South-East Asia: Vietnam), N. Wulijarni-Soetjipto (traditional and modern medicine in South-East Asia: Indonesia) & J.L.C.H. van Valkenburg (definitions, botany, research and development)

## 2 Alphabetical treatment of genera and species

## Abrus Adanson

Fam. pl. 2: 327, 511 (1763).

LEGUMINOSAE

$x = 10, 11, 12$ ; *A. fruticulosus*:  $n = 12, 24, 2n = 22$ ,

*A. precatorius*:  $n = 11, 2n = 22$

**Major species** *Abrus fruticulosus* Wight & Arn., *A. precatorius* L.

**Vernacular names** Indonesia, Malaysia, the Philippines: saga.

**Origin and geographic distribution** There are 4–20 species in *Abrus*, the number depending on how a species is defined. The genus is distributed pantropically, with 2 species in South-East Asia including the whole of Malesia.

**Uses** An extract of roots and leaves is used in traditional medicine to treat cough, hoarseness, digestive disorders such as gastralgia, and aphtha; it is also used as a diuretic. The seeds of *A. precatorius* have also played an important role in the treatment of conjunctivitis in various parts of the world. Macerating 3–5 seeds in 1 l water and applying drops of the liquid to the eye produces an inflammation of the conjunctiva, which used to be considered a cure for granular conjunctivitis. This practice has been abandoned because it appeared to be dangerous and uncontrollable. Although extremely toxic, the seeds have been used against malaria and dysentery, in conjunction with other drugs. In India, *A. precatorius* has a considerable reputation in the Ayurvedic, homeopathic, Unani and allopathic systems of medicine. The seed is one of the components of an oil applied to expel worms and against itching and skin diseases. It is also used to prepare an aphrodisiac and for a paste to remove piles, as an antidote when poisoned, and to treat glandular swellings and ulcers. It is used to induce abortion. Extracts from the seeds are used in Africa for the treatment of urinary schistosomiasis.

Leaves and roots have been used in various countries to sweeten foods. The leaves of *A. fruticulosus* are employed to sweeten traditional medicines used in central Thailand. The attractively coloured seeds of *A. precatorius* are often used as objects of art, ornaments or mascots; they are used as beads in rosaries and necklaces. They have also been used in soldering jewellery; when macerated in water they become mucous, and this sticky substance was mixed with solder to distribute it evenly and as a temporary cement prior to heating. In the Philippines, powdered seeds made into a paste have been used to poison darts and arrows. The stems have been used to tie together

materials in harbour works. *A. precatorius* is often cultivated in Java as an ornamental.

**Production and international trade** Although *A. precatorius* is sometimes cultivated for its medicinal uses or for the sweetening properties of the leaves, there are no statistics on production.

**Properties** *A. precatorius* seeds are extremely poisonous. They contain a toxic lectin fraction, usually called abrin, which is a complex mixture of toxic abrins and relatively non-toxic *Abrus* agglutinins. There are several methods for isolating the different toxins and agglutinins. However, there are so many isomorphs of the toxins (glycoproteins) that a number of subfractions (e.g. abrins A, B, C, D or abrins a, b or abrins I, II, III – all with slightly different characteristics) can be isolated, depending on the procedure used. Very little information is available on relations between the toxins isolated by the different methods; a gross relationship has been established on the basis of subunit compositions and sepharose 4B binding. From this, it can be concluded that abrin III probably corresponds to abrin C, and abrin I might resemble abrin A.

Typically, 100 g of seed kernels yield approximately 120 mg of abrin I, 150 mg of abrin II and 240 mg of abrin III. All abrins are glycoproteins ( $M_r$  63 000–67 000), composed of 2 polypeptide chains (A and B chains) linked through a single disulphide bond. The smaller A chain inhibits protein synthesis at nanomolar concentrations, and causes cell death. It is an N-glycosidase which inactivates eucaryotic ribosomes by cleaving the N-glycoside link of the residue at A<sup>4324</sup> of 28S-rRNA. It does not directly affect protein synthesis under the same conditions. Because of its ribosome-inactivating properties, studies of its biological activity have mainly focused on its potential as an immunotoxin in cancer therapy. It has been shown to be more toxic to tumour cells than to normal cells, and to provide therapeutic protection against Ehrlich ascites tumour and fibrosarcoma in mice and Yoshida sarcoma in rats, and to have an inhibitory effect in mice with solid human tumours. The larger B chain of the abrins is a galactose-specific lectin that binds to galactose-containing receptors on the cell plasma membrane. *Abrus* agglutinin is a tetramer with  $M_r$  134 900. It can be separated on DEAE-Sephacel into 2 fractions (APA-I, -II) that have several isomorphs. It is non-toxic to animal cells, but it is a potent haemagglutinator. When tested on mouse spleen cells, abrin was demonstrated to be a potent lymphocyte mitogen. Fresh preparations from the seeds of *A. pre-*

*catorius* are extraordinarily toxic, but are not mitogenic. However, after being stored for several months at 4°C it seems that such preparations are relatively non-toxic and are effective mitogens. The lectins from the seeds have blastogenic properties on human blood lymphocytes in vitro.

Abrin is one of the two most toxic substances of plant origin known (the other being ricin from the seeds of *Ricinus communis* L.). The LD<sub>50</sub> value of abrin in mice is as low as 20 µg/kg (intra peritoneal) for the purified substance, and goats fed with a daily amount of 1 g/kg body weight of the seeds die within a few days. The symptoms are loss of appetite, bloody diarrhoea, dyspnoea, dehydration, loss of condition and recumbency caused by fatty change and necrosis of hepatocytes and renal convoluted tubules, pulmonary haemorrhage, oedema and emphysema, and erosions of the abomasal and intestinal epithelium. Intoxication of dogs resulted in death after 15–40 hours. When non-lethal doses of abrin were given to mice and dogs, the symptoms were reversible. The animals recovered, apparently completely, in 1–3 weeks. In addition to its toxic effect, the aqueous seed extract also has antigenic, abortive and teratogenic properties. Human poisoning is characterized by a latent period of several hours to days, followed by severe gastro-enteritis with erosion, necrosis of the liver, kidneys, spleen and lymphatic tract. After ingestion of seeds, for instance by children, immediate emesis is essential; less than one seed can be fatal when it is thoroughly masticated and the tough seed-coat has been damaged. Seeds swallowed whole with intact testa remain innocuous. The toxins are heat-stable to incubation at 60°C for 30 minutes; at 80°C, however, most of the toxicity is lost within 30 minutes.

*Abrus* seeds are also a rich source of alkaloids, among which abrin (N-methyl-L-tryptophan), hypaphorine, N,N-dimethyl tryptophan methylester, precatorine, choline and trigonelline. The insecticidal properties of seed extracts are attributed to the alkaloids (rotenoids), with hypaphorine being the most effective. The alkaloids reduce the fecundity of female mites and also deter feeding. The seeds contain an indole fraction that inhibits the growth of several plant species, e.g. of germinating lettuce. The active component of this fraction differs from the indole alkaloids abrine and hypaphorine, and has been determined as N,N-dimethyl tryptophan.

A methanol extract of the seeds furthermore showed a concentration-related inhibitory effect on the motility of human spermatozoa, and re-

duced sperm viability; this offers prospects for application as a human contraceptive. Seed extracts also fully inhibit germination of spores of the fungi *Botrytis cinerea* and *Colletotrichum gloeosporioides*. An almost 100% inhibition of tobacco mosaic virus was found on *Capsicum* pepper plants in vitro.

The roots and leaves of *A. precatorius* are known to contain constituents that have anti-inflammatory, anti-tumour, antitoxic, antitussive, antithrombotic and antibiotic properties. Phytochemical investigations of the herb and roots have revealed the presence of a series of isoflavanquinones: abruquinones A, B and C from the herb, and abruquinones A, B, D, E and F from the roots. Abruquinones A, B and D were found to exhibit remarkable inhibitory effects on platelet aggregation. The IC<sub>50</sub> of abruquinones A and B for the inhibition of the platelet aggregation induced by arachidonic acid and collagen were less than 5 µg/ml, whereas that of abruquinone D was less than 10 µg/ml for the aggregation induced by arachidonic acid. Abruquinones A, B, D and F also showed strong anti-inflammatory and anti-allergic effects: superoxide formation was inhibited at a dose of less than 0.3 µg/ml, and the release of both β-glucuronidase and lysozyme from rat neutrophils and of both β-glucuronidase and histamine from mast cells were inhibited at a dose of less than 1 µg/ml. All these effects were measured in vitro.

The roots are also known to have antioestrogenic activity. Tests with laboratory animals (hamsters) infected with *Schistosoma haematobium* seemed to confirm the activity against urinary schistosomiasis. After oral application of a root extract of *A. precatorius* both the egg count and worm load were significantly reduced when compared to the controls.

The powdered drug from the leaves of *A. fruticosus* used in Thailand is pale green and has a sweet taste. The sweet constituents of the leaves (of *A. precatorius* and *A. fruticosus*) have been characterized as the cycloartane glycosides abrusosides A, B, C and D. Their aglucone, obtained by acid hydrolysis has been identified as abrusogenin. Abrusosides exhibit sweetness potencies in the range of 3–100 times greater than sugar (2% sucrose solution). In preliminary safety tests they were found to be non-toxic for rodents and were non-mutagenic. The yield of abrusosides A–D from *A. fruticosus* leaves (0.33%) is slightly lower than from *A. precatorius* leaves (0.39%), but the concentration of abrusoside B, the sweetest com-

pound, is higher in *A. fruticosus* (0.08% versus 0.03% in *A. precatorius*).

The ethanol extract of the leaves has been found to inhibit acetylcholine-induced contractions of preparations of toad rectus abdominis and rat phrenic nerve-diaphragm muscle. The effects were concentration-dependent and reversible. Furthermore there were no effects on direct electrical stimulation of the rat diaphragm. Thus the ethanol extract is similar to d-tubocurarine with respect to its pattern of neuromuscular blockade. Leaves and roots have been reported to contain some abrin.

**Adulterations and substitutes** As already mentioned, the toxicity of abrin is paralleled by ricin from *Ricinus communis*, which has similar properties. Other vegetable proteins with pharmacological potential that are broadly comparable with abrin are found in the *Cucurbitaceae* genera *Luffa*, *Momordica* and *Trichosanthes*. They also have abortifacient and antitumour properties.

The roots of true liquorice (*Glycyrrhiza glabra* L.) contain the sweetener glycyrrhizine, which is used worldwide on a much larger scale than the sweetening substances in *Abrus* leaves or roots. It has similar medicinal properties to the abrusosides (e.g. anti-inflammatory and antitussive properties) and is used to cure coughs, bronchitis and gastralgia.

**Description** Woody subshrubs or lianas up to 6(-9) m long, stems often reaching 1.5 cm in diameter, often slender-branched. Leaves alternate, paripinnate with opposite leaflets, the rachis projecting beyond the last pair of leaflets; stipules small, usually persistent. Inflorescence axillary or terminal, pseudoracemose with the flowers in clusters on short reduced wart-like branchlets often arranged unilaterally on the rachis. Flowers sessile or subsessile, bisexual, 5-merous; calyx tube almost toothless or with 5 short teeth, the upper pair partly joined; corolla papilionaceous, much longer than the calyx, white, yellow, pink to dark purple, standard ovate-orbicular with a short claw and notched at the apex, wings oblong-falcate with long claws, keel longer than wings; stamens 9, filaments joined into a tube but free in upper part, staminal tube at the base adnate to the standard; ovary superior, subsessile, pubescent, 1-loculate, with numerous ovules, style curved, usually persistent, stigma capitate. Fruit an oblong to linear pod, flattened or inflated, beaked, pubescent, dehiscent, (1-)3-12-seeded, more or less septate between the seeds. Seeds subglobose, ovoid to ellipsoid, sometimes compressed, usually shiny.

Seedling with epigeal germination; hypocotyl elongated.

**Growth and development** The swollen wart-like branchlets of the inflorescence of *A. precatorius* are visited by ants.

**Other botanical information** *Abrus* is usually considered to have an isolated position within the subfamily *Papilionoideae* and placed in a separate tribe *Abreae*. It is characterized by the combination of woody stems with a tendency to twine, paripinnate leaves, pseudoracemes and 9 stamens. Some taxonomists adhere to a wide species concept, resulting in the acceptance of only 4 species worldwide. Others prefer a more narrow species concept and accept up to 20 species. *A. fruticosus* is an extremely polymorphic and widely distributed species, which is sometimes considered as a complex of several separate species.

**Ecology** *Abrus* occurs rather frequently in grasslands, cropped land (also as a weed), thickets, edges of rain forest and gallery forest, up to 1500 m altitude.

**Propagation and planting** The seed weight of *A. precatorius* averages 150-410 mg. When untreated, the germination rate is about 40% in 18 days. After seeds have been soaked in concentrated  $H_2SO_4$  for 3 hours at 30°C the germination is about 85% in 10 days. Mechanical scarification results in a germination rate of 97%. Seed can be sown directly in the field or in a nursery. When sown directly in the field, 40 kg/ha is needed to obtain 40 000-50 000 plants. 2-3 seeds are planted per hole. Plants from seed sown in a nursery are planted into the field at 25 cm x 60 cm when 3-4 months old. Young plants should preferably be shaded by trees. In Indonesia, *Gliricidia sepium* (Jacq.) Kunth ex Walp. and *Leucaena leucocephala* (Lamk) de Wit are commonly used as shade trees. Propagation by cuttings is easy. *A. precatorius* can be raised in sole cropping or as a cover crop e.g. in rubber plantations in Peninsular Malaysia.

**In vitro production of active compounds** Ribosome-inactivating proteins and agglutinins have been isolated from callus and cell suspension cultures established from seed explants of *A. precatorius*. Biosynthesis of these lectins is positively correlated with the growth of the cultures. The lectins can be purified from the culture, and their electrophoretic mobility and biological activity are comparable with those of the lectins purified from the seeds. The cultures can be maintained on revised Murashige and Skoog medium. The rotenoid content of leaves, stems and seeds is, however,

greater than in tissue cultures. Small amounts of tephrosin and deguelin can be found in the tissue culture.

**Husbandry** Farmyard manure can be given early at 10–15 t/ha. Plants 2–3 months old can be fertilized with 150 kg urea + 150 kg triplesuperphosphate + 50–100 kg KCl per ha. Experiments with planting *A. precatorius* without support gave a higher yield of leaves (14.5%) and facilitated harvesting.

**Diseases and pests** The most serious disease of *A. precatorius* in Indonesia is *Rhizoctonia solani*, a fungus that causes stem rot. Witches broom disease caused by a mycoplasma-like organism has been reported on *A. precatorius* in Taiwan.

**Harvesting** The first harvest can be obtained when *A. precatorius* plants are 6–8 months old. Plants are cut 25–30 cm above the ground; 4–6 harvests per year can be expected.

**Yield** When harvested 4–6 times per year, a total of 4.7 t/ha of fresh leaves and twigs per year is obtained, which is equivalent to 0.6–1 t of dry leaves. Roots are harvested when plants are 30–36 months old, yielding 2.5–3.5 t of fresh roots per ha.

**Genetic resources and breeding** Both Malaysian *Abrus* species have large areas of distribution and often inhabit anthropogenic localities. They do not seem to be at risk of genetic erosion. In India, the populations of *A. precatorius* have been gradually depleted because of the extensive use in local medicine. No germplasm collections are known to exist, except in botanical gardens. In Indonesia, living plant material is available at the Research Institute for Spices and Medicinal Plants, Bogor and the Research Institute for Medicinal Plants at Tawangmangu.

**Prospects** Extensive research has been carried out on *A. precatorius* to elucidate the chemical composition, structure and properties of the seed constituents and to a lesser extent of those in the leaves and roots. It appears that certain constituents exhibit anti-cancer and anti-leukaemia effects. Moreover, they may influence fertility as well, whereas the toxic properties may be applicable as effective insecticide. In short, *Abrus* shows promise.

**Literature** |1| Breteler, F.J., 1960. Revision of *Abrus* Adanson (Pap.) with special reference to Africa. *Blumea* 10(2): 607–624. |2| D'Silva, I., Vaidyanathan, C.S. & Podder, S.K., 1993. Ribosome-inactivating proteins and agglutinins from callus and suspension cultures of *Ricinus communis* L. and *Abrus precatorius* L. *Plant Science* (Limerick) 94(1–2): 161–172. |3| Fullas, F., Choi,

Y.-H., Kinghorn, A.D. & Bunyapraphatsara, N., 1990. Sweet-tasting triterpene glycoside constituents of *Abrus fruticosus*. *Planta Medica* 56(3): 332–333. |4| Kaushik, P. & Khanna, P., 1992. Insecticidal substances from in vivo and in vitro tissue culture of *Abrus precatorius* L. *Advances in Plant Sciences* 5(2): 464–469. |5| Kinamore, P.A., Jaeger, R.W. & de Castro, F.J., 1980. *Abrus* and *Ricinus* ingestion: management of three cases. *Clinical Toxicology* 17(3): 401–405. |6| Ratnasooriya, W.D., Amarasekera, A.S., Perera, N.S.D. & Premakumara, G.A.S., 1991. Sperm antimotility properties of a seed extract of *Abrus precatorius*. *Journal of Ethnopharmacology* 33(1–2): 85–90. |7| Sheng, C.K., Sheng, C.C., Lin, H.C., Jin, B.W., Jih, P.W. & Che, M.T., 1995. Potent antiplatelet, anti-inflammatory and antiallergic isoflavanquinones from the roots of *Abrus precatorius*. *Planta Medica* 61: 307–312. |8| Singh, K. & Kumar, S., 1984. Ecophysiological observations on Indian medicinal plants. I. Seed germination responses to certain physical and chemical treatments. *Acta Botanica Indica* 12(2): 216–219. |9| Thuân, N.V., Dy Phon, P. & Niyomdham, C., 1987. Leguminosae (Fabaceae) Papilionoideae. In: Lescot, M. (Editor): *Flore du Cambodge, du Laos et du Vietnam*. Vol. 23. Muséum National d'Histoire Naturelle, Paris, France. pp. 50–55. |10| Verdcourt, B., 1979. A manual of New Guinea legumes. *Botany Bulletin* No 11. Office of Forests, Division of Botany, Lae, Papua New Guinea. pp. 305–308.

#### *Selection of species*

#### ***Abrus fruticosus* Wight & Arn.**

Prod. Fl. Penins. Ind. Or. 1: 236 (1834).

**Synonyms** *Abrus melanospermus* Hassk. (1844), *Abrus pulchellus* Wallich ex Thwaites (1859).

**Vernacular names** Indonesia: saga areuy, areuy si hayam (Sundanese), daun sambang (Javanese). Malaysia: saga negri, akar kacang inai (Peninsular). Thailand: ma klam phueak (Chiang Mai), kho kiu (Chanthaburi), ma khaam yaan (Trang). Vietnam: k[ee] c[oo]s[t th[ar]].

**Distribution** Africa, India, Sri Lanka, Burma (Myanmar), Indo-China, China, Thailand and Malesia (with certainty in Peninsular Malaysia, Java, Borneo, the Philippines, Sulawesi and New Guinea).

**Uses** The roots are used to treat digestive disorders. The leaves are employed to sweeten traditional medicines used in the central regions of

Thailand. The stems have been used to tie together materials in harbour works.

**Observations** A prostrate subshrub or woody climber up to 6 m long; leaves with (10–)12–34 oblong, obovate-oblong or ovate leaflets, truncate to broadly rounded and mucronulate at apex; inflorescence comparatively slender, usually straight, flowers in clusters on cushion-like reduced branchlets; fruit oblong to linear-oblong, compressed, 4–12-seeded; seeds usually strongly compressed, greyish-brown to reddish-brown, sometimes speckled. *A. fruticulosus* is extremely polymorphic and often subdivided into several separate species and subspecies. It occurs in roadsides, along streams, in thickets and edges of lowland rain forest up to 1000 m altitude.

**Selected sources** 202, 262, 451, 580, 1519, 1520.

### **Abrus precatorius L.**

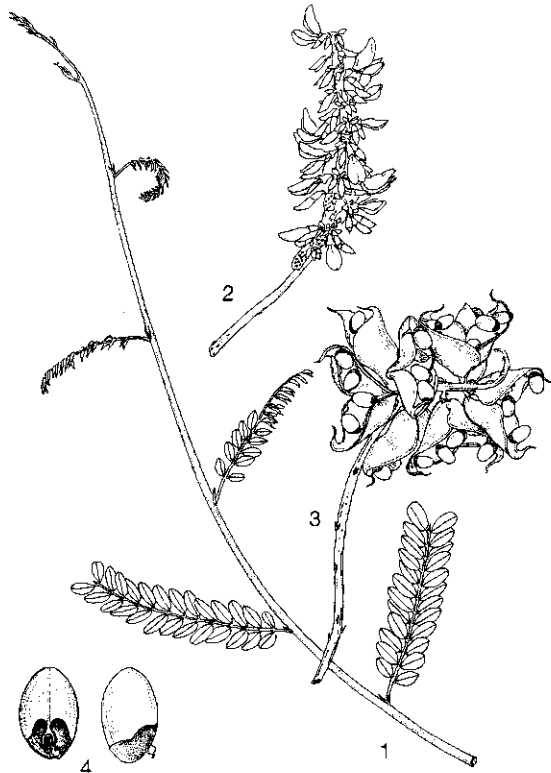
Syst. nat. ed. 12: 472 (1767).

**Vernacular names** Indian liquorice, jequirity bean, crab's eye (En). Jéquirity, liane réglisse (Fr). Indonesia: saga, saga manis (general), saga telik (Javanese). Malaysia: akar saga. Philippines: saga, kansasaga, bangati (general). Burma (Myanmar): ywe-nge. Cambodia: ângkreem, kre:m krâ:m (Kompong Thom). Laos: khua sa em, makam. Thailand: ma klam taanuu (Bangkok), klam khrua (Chiang Mai), ma khaam thao (Trang). Vietnam: d[aa]y c[uw][ow]f m th[ar]o, cam th[ar]o d[aa]y, d[aa]y t[uw][ow]ng t[uw].

**Distribution** Africa, tropical and subtropical America (introduced), tropical Asia, Australia and the Pacific Islands; throughout South-East Asia.

**Uses** The seeds have played an important role in the treatment of conjunctivitis in various parts of the world. An extract of roots and leaves is a traditional cure for aphtha. In coastal areas of Papua New Guinea, leaves are chewed for a week as a traditional treatment for asthma. The leaves are also used in the same way as liquorice. The seeds are used in ornaments.

**Observations** A woody climber up to 6(–9) m long, stems often attaining 1.5 cm in diameter; leaves with 16–34 oblong, obovate-oblong or ovate leaflets, obtuse to acuminate at apex; inflorescence thick and robust, usually curved, flowers in dense clusters on cushion-like nodes; fruit oblong, inflated, 1–7-seeded; seeds ovoid, scarlet with area around the hilum black, rarely entirely black, whitish or yellowish. The African material has been separated as subsp. *africanus* Verdc., based on minor differences in pod characteristics from



*Abrus precatorius* L. – 1, young shoot; 2, inflorescence; 3, infructescence; 4, seeds.

the Asian subsp. *precatorius*. *A. precatorius* occurs in grasslands, cropped land (also as a weed), thickets, edges of monsoon rain forest and gallery forest up to 1500 m altitude.

**Selected sources** 65, 130, 190, 193, 196, 202, 255, 268, 350, 353, 301, 363, 398, 417, 557, 568, 580, 597, 633, 696, 706, 711, 712, 714, 736, 738, 811, 896, 948, 1015, 1035, 1059, 1060, 1170, 1178, 1206, 1328, 1519, 1520, 1541, 1563, 1612.

R.H.M.J. Lemmens & F.J. Breteler

### **Achillea millefolium L.**

Sp. pl. 2: 899 (1753).

COMPOSITAE

2n = 18, 36, 54, 72

**Vernacular names** Yarrow, milfoil, thousand weed (En). Achillée millefeuille (Fr). Vietnam: d[uw] [ow]ng klyf th[ar]o.

**Origin and geographic distribution** *A. millefolium* is considered as a complex of difficult-to-separate taxa found primarily throughout the

temperate and boreal zones of the Northern Hemisphere and, to a lesser extent, the Southern Hemisphere. It is cultivated, usually as an ornamental, in mountainous areas of some parts of Malesia (e.g. locally in the Philippines and in Java), and is locally naturalized there. It grows wild or is naturalized in Indo-China.

**Uses** The flowering tops of yarrow (and the herb) are listed in many Western pharmacopoeias. In traditional medicine they are considered to possess antiphlogistic, spasmolytic, stomachic, carminative and choleric properties. Yarrow is used internally for the treatment of gastro-intestinal complaints (inflammation, diarrhoea, flatulence, cramps, poor digestion), as a bitter aromatic (to counter loss of appetite), to stimulate the secretion of bile (choleric activity) and to enhance the renal elimination function. In the Unani system of medicine, the flowers are used as abortifacient and emmenagogue. Externally the herb is used in poultices or preparations with alcohol (percolates, fluid extracts) in the treatment of inflammation of the skin and mucous membranes, as well as for healing wounds and relief of itches. In folk medicine, the drug is often employed as a haemostyptic, e.g. in bleeding from haemorrhoids and in menstrual disorders.

The ethereal luteolin oil has found widespread use in cosmetic and dermatological preparations. It is also used as a rinse to strengthen the hair, and as a shampoo to prevent baldness. In the form of a bath, yarrow or its oil are applied to remove perspiration.

Yarrow has been reported to be used medicinally in Java, usually as a decoction of fresh flower heads and leaves in water, internally and externally, for the same purposes as in traditional medicine in Europe. Yarrow is often one of the ingredients in herbal mixtures. In India, it is used in a powdered mixture of 18 medicinal plants, which has shown anti-viral activity in mice experimentally infected with encephalitis virus.

Besides its numerous medicinal uses, yarrow has been used as a ceremonial smoke, snuff and beverage by North American Indians, as an occasional substitute for cinnamon or nutmeg, and as a substitute for hops in the brewing of beer. It is a common ingredient of herbal candies. Yarrow is commonly planted as an ornamental, particularly the forms with reddish flowering heads.

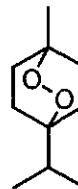
**Production and international trade** Yarrow is grown commercially in Albania (700 kg/year) and Hungary (100 kg/year) and in small quantities in the northern United States and Canada.

The quantity of oil produced annually is less than 1000 kg worldwide.

**Properties** Approximately 150 different compounds have been isolated and identified from yarrow. The essential oil (content 0.2% to more than 1%) has been much investigated chemically because of its pharmacological properties. The essential oil is found in different concentrations in all aboveground parts of the plant: the leaves, however, contain less than the flower-heads. It contains sesquiterpene lactones which are precursors of azulenes (proazulenes, e.g. achillicin); upon steam distillation, they yield azulene and chamazulene. Oils rich in azulene and chamazulene (up to 51% of the oil) have a characteristic deep blue colour. Other sesquiterpenes include achillin, achillifolin, matricarin, millefin, dihydroparthenolide, germacrene D (up to 54%),  $\beta$ -caryophyllene (up to 8%), balchanolide, and farnesol. The monoterpenes present in the oil include, depending on the origin of the oil: terpinen-4-ol, terpineol,  $\alpha$ -pinene (up to 2% of the oil),  $\beta$ -pinene (up to 10%), 1,8-cineole (up to 6%), sabinene (up to 17%), myrcene (up to 15%), camphor (up to 7%), linalool (up to 26%),  $\beta$ -thujone (up to 14%),  $\alpha$ -thujone (up to 20%),  $\alpha$ -phellandrene (up to 12%) and limonene (up to 3%).

Several monoterpenes and sesquiterpenes have been identified in the essential oil from plants from Greece, with ascaridole as the main component (47%) and lesser amounts of 1,8-cineole (10%), p-cymene (7%),  $\alpha$ -terpinene (7%) and camphor (8%); other populations from Greece had camphor as the main constituent (22%), with smaller amounts of 1,8-cineole (12%), lavandulol (7%) and borneol (8%), and small amounts of another 80 compounds.

The flavonoids have also received considerable attention. Yarrow contains flavonoids (apigenin and luteolin-7-O-glucoside), glycosylflavones (especially swertisin, vicenin-2 and -3, schaftoside and isoschaftoside) and 6-methoxylated or di- and tri-methylated flavonols (such as pectolarigin, 3-methylbetuletol and 3,6,4'-methylquercetagine).



ascaridole



The roots of *A. millefolium* are reported to contain polyynes (e.g. pontica epoxide) and alkylamides (e.g. undeca-2E,4E-diene-8,10-dienoic acid isobutylamide). Other nitrogen-containing compounds isolated from the herb include betaine (0.05%), betonicine (= achilleine = L-(-)-hydroxystachydrine, a compound with reported haemostatic activity) and L-(-)-stachydrine. Triterpenes, saponins, coumarins (0.35%) and tannins (3–4%) have also been reported.

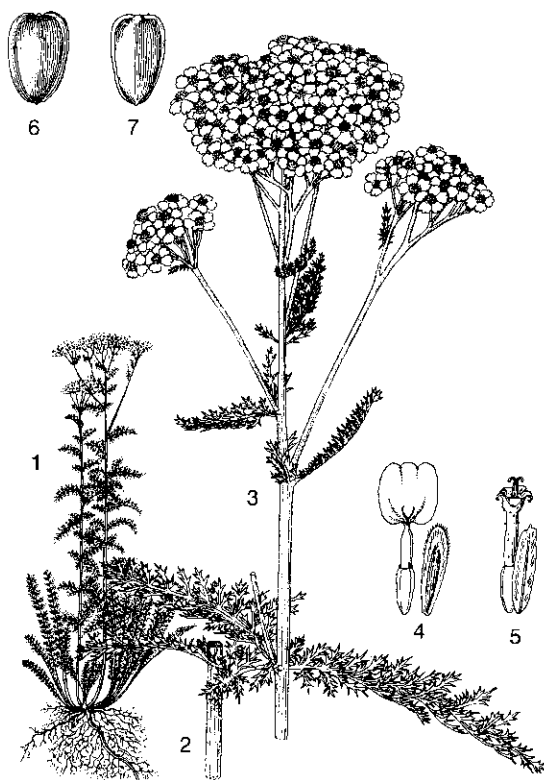
The anti-inflammatory and antispasmodic effects have been confirmed in respectively a mouse-footpad oedema model and an isolated rabbit-ileum model. These properties are tentatively attributed to the flavonoids, which are known to have such activity, but azulene and related compounds have also been claimed to possess anti-inflammatory together with antipyretic activity. The mechanism of anti-inflammatory activity has been suggested by the synthesis of anti-prostaglandin. Sesquiterpene lactones exhibit interesting biological effects, including antimicrobial and cytotoxic and anti-cancer activity. The methyl esters of the sesquiterpenes achimillic acid A, B and C (isolated from *A. millefolium* flowers) were found to be active against mouse P-388 leukaemia cells *in vivo*.

Ascaridole (a monoterpene-peroxide) and  $\alpha$ -peroxyachifolide (a sesquiterpene-peroxide) both showed *in vitro* activity against malaria parasites (*Plasmodium falciparum*). The activity of ascaridole is about the same as for artemisinin from *Artemisia annua* L. The activity of  $\alpha$ -peroxyachifolide is much less than that of the latter compound ( $EC_{50}$  1  $\mu\text{g/ml}$  and 0.01  $\mu\text{g/ml}$ , respectively). Besides the anti-protozoal effects, ascaridole also has anthelmintic activity against *Ankylostoma*, *Ascaris*, *Necator* and *Trichuris* worms. Thujone is known to possess abortifacient activity.

Yarrow has been reported to contain substances that inhibit seed germination, are antibacterial and act as a mosquito larvicide, and a volatile oil that evokes sex pheromone-like responses in male cockroaches. An ethanol extract of plants from Sweden showed repelling properties against the mosquito *Aedes aegypti*. The most active compounds were identified as stachydrine, caffeic, chlorogenic and salicylic acids, and the phenolic compound pyrocatechol. These substances seem to be fairly safe for cutaneous application, but more toxicity studies are desirable. Yarrow may induce allergic dermatitis, which is usually attributed to the sesquiterpene lactones such as guaianolide peroxides. Furanocoumarins might be the cause of phytophotodermatitis.

**Adulterations and substitutes** Matricaria or German chamomile, *Matricaria recutita* L. (synonyms: *Chamomilla recutita* (L.) Rauschert, *Matricaria chamomilla* auct. non L.) is another plant species with an essential oil containing a sesquiterpenoid lactone, i.e. matricin, which may decompose to chamazulene. Wormseed (*Chenopodium ambrosioides* L.) also contains ascaridole in larger amounts.

**Description** A perennial herb, (8–)30–90 cm tall, with aromatic odour and greyish-green colour from the numerous small hairs; stem angular. Leaves alternate, clustered at the base of the stem and with smaller leaves upwards, highly dissected, up to 3-pinnatifid, lanceolate to linear in outline, up to 20 cm  $\times$  6 cm. Flowering heads (capitula) in a flat-topped corymb, small, pedunculate, varying in colour from white to pink, magenta and red; involucre bracts in few rows, the outer somewhat shorter than the inner, with a scarious mar-



*Achillea millefolium* L. - 1, plant habit; 2, middle part of stem with leaf; 3, upper part of flowering stem; 4, ray flower and involucre bract; 5, disk flower and receptacle scale; 6, achene from disk flower; 7, achene from ray flower.

gin. Outer florets in each capitulum usually 5, female, ligulate with more or less 3-dentate, patent ligules; inner florets hermaphrodite, 5-lobed, with compressed corolla tube and a receptacle scale at the base. Fruit a compressed achene, oblong or obovate, without pappus.

**Growth and development** Plants of yarrow reproduce by suckers and consequently usually grow in groups. The suckers develop into rhizomes. Yarrow is an obligate cross-pollinator, and is pollinated by insects. The fruits are distributed by wind.

**Other botanical information** In the literature the name *A. millefolium* may refer to several species of the complex. In West European literature it usually means *A. millefolium* sensu stricto; in Eastern Europe it is mainly *A. millefolium* s.s. and *A. collina* J. Becker ex Reichenb.; in North America usually *A. lanulosa* Nutt and, to a lesser extent, *A. millefolium* s.s. is meant.

*A. collina* (also described as *A. millefolium* subsp. *collina* (J. Becker ex Reichenb.) Weiss) is tetraploid and *A. millefolium* s.s. is hexaploid, but they cannot be clearly distinguished by morphological characteristics. Diploid, tetraploid, hexaploid and octoploid taxa have been reported within the complex *A. millefolium*. The most common form in Western Europe is the hexaploid with white flowers.

**Ecology** *A. millefolium* generally grows in open, unshaded areas ranging from cliffs and alpine pastures to lowland meadows, lawns, roadsides and waste ground. The stems and rhizomes are tough and withstand trampling well; yarrow often grows close to paths and roads. In closed grassland it is usually suppressed by the grasses. Tetraploid populations containing proazulenes have only been found in meadow habitats and not in roadside vegetations and pioneer habitats, where the hexaploid type occurs with little or no proazulenes. Shoot height and aboveground fresh matter decrease sharply on acid soils.

In South-East Asia, yarrow is locally naturalized in grassy habitats, such as roadsides and lawns, and in mountainous areas.

**Propagation and planting** Yarrow can easily be grown from seed. Vegetative propagation from suckers is also possible.

**In vitro production of active compounds** Friable calli were obtained from hypocotyls of yarrow in Gamborg B5 medium. They were used for the production of cell suspension cultures in the same liquid in the dark supplemented with 1.5 mg/l dichlorophenoxyacetic acid, 0.1 mg/l kinetin

and 2% sucrose. Cultures grown as such had a doubling time of 35–40 hours. Analysis of the volatile component produced by yarrow cell suspension cultures showed the presence of monoterpenes and sesquiterpenes, some of them not present in plant extracts.

**Husbandry** Presumably, yarrow plants with reddish flowering heads were favoured for cultivation as medicinal plant. Where yarrow is cultivated, seeds are planted in early spring, and the crop is grown as an annual.

**Harvesting** Planted yarrow is usually harvested late summer when in full bloom.

**Yield** There is a considerable variation in the contents of oil and azulene in yarrow, due to provenance of the material, the plant part used, its age, season of collection, climatic and soil conditions, and the inverse relationship between the amounts of oil and azulene produced. In the Ukraine, *A. collina* cultivated experimentally showed very promising results: 6.3 t/ha of fresh matter, 0.2% essential oil of which 42% azulene. In experiments with yarrow in Brazil, manuring resulted in an average increase of biomass production from 47 to 134 g/plant, and of essential oil yield from 0.04 to 0.15 ml/100 g of fresh flower heads.

**Handling after harvest** After harvesting, yarrow should be wilted for 24–48 hours but kept dry to prevent fermentation. Whole plants are distilled by steam distillation, which takes 6–10 hours. Cohabitation is recommended. The oil should be stored in dark containers under cool conditions.

**Genetic resources and breeding** Tetraploid yarrow in the *A. millefolium* complex contains fair amounts of proazulenes, yielding azulenes upon steam distillation. Except for a few hexaploid populations the essential oil of hexaploids contains no more than traces of proazulenes. Diploid yarrow is reported to lack flavones and flavonols. The widely varying medicinal properties and uses of yarrow in different regions is the result of the occurrence of several genotypes in the *A. millefolium* complex with different chemical composition.

**Prospects** Western pharmacopoeias prescribe determining the essential oil content (>0.3%) for yarrow and characterizing azulenes in the drug. Only certain taxa and populations within the *A. millefolium* complex can meet these requirements. The relation between chemical composition, ploidy level and ecological conditions is very complicated and makes standardization extremely difficult. Moreover, unambiguous identification is hampered by the frequent occurrence of hybridization

and aneuploids. Yarrow and its properties are poorly known in South-East Asia. Information on the chemical composition is urgently needed. Yarrow is easy to propagate and grow and might be well cultivated in home gardens at higher altitudes in Malesia.

**Literature** |1| Axtell, B.L. & Fairman, R.M., 1992. Minor oil crops. Part III - Essential oils. FAO Agricultural Services Bulletin No 94: 215-216. |2| Banh-Nhu, C., Gacs-Baitz, E., Radics, L., Tamas, J., Ujszaszy, K. & Verzar-Petri, G., 1979. Achillicin, the first proazulene from *Achillea millefolium*. *Phytochemistry* 18(2): 331-332. |3| Bruneton, J., 1995. Pharmacognosy, phytochemistry, medicinal plants. Technique & Documentation Lavoisier, Paris, France. pp. 289-291. |4| Chandler, R.F., Hooper, S.N. & Harvey, M.J., 1982. Ethnobotany and phytochemistry of yarrow, *Achillea millefolium*, Compositae. *Economic Botany* 36(2): 203-223. |5| Figueiredo, A.C.S. & Pais, M.S.S., 1991. *Achillea millefolium* (yarrow) cell suspension cultures: establishment and growth conditions. *Biotechnology Letters* 13(1): 63-68. |6| Hofmann, L. & Fritz, D., 1993. Genetical, ontogenetical and environmental caused variability of the essential oil of different types of the *Achillea millefolium* 'complex'. In: Bernáth, J., Craker, L.E. & Levy, A. (Editors): *Botanical Aspects, Genetic Resources, Genetics, Breeding, Biotechnology. Proceedings of the First World Congress on Medicinal and Aromatic Plants for Human Welfare*. Maastricht, Netherlands, July 19-25, 1992. *Acta Horticulturae* 330: 147-157. |7| Hofmann, L., Fritz, D., Nitz, S., Kollmannsberger, H. & Drawert, F., 1992. Essential oil composition of three polyploids in the *Achillea millefolium* 'complex'. *Phytochemistry* 31(2): 537-542. |8| Kokkalou, E., Kokkini, S. & Handilou, E., 1992. Volatile constituents of *Achillea millefolium* in relation to their infraspecific variation. *Biochemical Systematics and Ecology* 20(7): 665-670. |9| Schaffer, M.C., Ronzelli, P. & Koehler, H.S., 1993. Influence of organic fertilization on the biomass, yield and composition of the essential oil of *Achillea millefolium* L. In: Palevitch, D., Simon, J.E. & Mathé, A. (Editors): *Raw Material Production, Product Introduction. Proceedings of the First World Congress on Medicinal and Aromatic Plants for Human Welfare*. Maastricht, Netherlands, July 19-25, 1992. *Acta Horticulturae* 331: 109-114. |10| Tunón, H., Thorsell, W. & Bohlin, L., 1994. Mosquito repelling activity of compounds occurring in *Achillea millefolium* L. (Asteraceae). *Economic Botany* 48(2): 111-120.

**Other selected sources** 139, 197, 240, 549, 580, 589, 750, 1035, 1149, 1178, 1262, 1474, 1566.

R.H.M.J. Lemmens & N. Bunyaphatsara

### **Acorus calamus L.**

Sp. pl. 1: 324 (1753).

ACORACEAE

2n = 24, 36, 48

**Synonyms** *Acorus terrestris* Spreng. (1825), *Acorus asiaticus* Nakai (1936).

**Vernacular names** Sweet flag, sweet root, calamus (En). Calamus, acore odorant, acore vrai (Fr). Indonesia: daringo (Sundanese), dringo (Javanese), jerango (Sumatra). Malaysia: jerangau, deringu, jerangoh (Peninsular). Papua New Guinea: lepe (Angi, Enga), esue (Mendi, Southern Highlands), wamala (Aroma, Central Province). Philippines: lubigan (Tagalog, Bisaya), acoro (Spanish), daraw (Iloko). Laos: hang khao nam. Thailand: kha Chiang chee (northern), wan nam (central), haang khao phaa (Chiang Mai). Vietnam: th[ur]y x[uw]low]ng bloof, x[uw]low]ng b[loof], b[loof] bloof] n[lees]p.

**Origin and geographic distribution** Sweet flag is probably a native of China and India. Its use as a medicinal plant dates back to Egypt, Greek and Roman times. Sweet flag was distributed from its native range by rhizomes through trade and commerce, and arrived in Europe in the 16th Century. In the Malesian region, it is considered as naturalized and not truly wild. It is found in many parts of Indonesia, Malaysia, and Papua New Guinea and locally in the Philippines (Bontoc and Benguet Provinces), and outside Malesia in Indo-China and Thailand. It is also cultivated here and there.

**Uses** The rhizomes of sweet flag have been used extensively in traditional medicine by Chinese, Indians, American Indians and others, and are still used in many regions. In Roman and Arabic civilizations aphrodisiac properties were attributed to the rhizome and it was used in North America and Europe as a panacea; in India, sweet flag has, for centuries, been an important medicinal aid for stomach complaints and colic in children. Since ancient times it has been reputed for its stimulant digestive virtues. In India, the rhizomes are traditionally used in an infusion to treat diarrhoea, dysentery, atonic dyspepsia and asthma, and for their carminative, expectorant, nauseant, anti-spasmodic, stomachic, vermifuge, sedative and emetic properties. In Vietnam, sweet flag is used

to treat respiratory disorders (asthma, inflammation), rheumatism, remittent fevers, snake bites and as sedative. In Indonesia and Malaysia, the rhizomes are usually used externally to treat inflammation, rheumatism, lumbago and skin diseases, and internally after childbirth. In Java, sweet flag is an ingredient of certain 'jamus', and in the Philippines it is used as a masticatory against toothache and as a stimulant, carminative and antirheumatic. In Papua New Guinea, the leaves of sweet flag are taken as a tonic, and chewed to relieve toothache. The crushed rhizome is rubbed into the hair to kill lice. In Brunei, sweet flag is used to treat gastritis and diarrhoea and also as a poison antidote. In Thailand, the rhizomes are used as carminative, analgesic, anthelmintic, and to treat diarrhoea and dysentery. In Japan, *Acorus* oil is used as bathing agent, considered to be effective against skin diseases and to improve blood circulation. In the Unani (Greco-Arab) system of medicine, sweet flag is used to treat cardiovascular diseases. In Vietnam, a dose of 2–5 g/day is administered in decoction. In modern phytotherapy, sweet flag rhizomes ('calami rhizoma') can, on the basis of their constituents, be called a bitter aromatic. This is principally used as a stomachic and carminative (internally) and externally as a rubefacient and in the treatment of seborrhoea (as a bath).

The fragrant oil obtained from the rhizome is not only used medicinally, but also for flavouring alcoholic beverages (e.g. vermouths), fish, sweets and cakes, in perfumes and sacred oils and as an insecticide. As an insecticide, it is often used as emulsified foliage spray. The use of rhizome powder in warehouses and on the farm to protect stored grain, rice and pulses from insect pests has proved fairly effective and economical; powdered rhizomes may also reduce the extent of fungal and bacterial contamination. The hydroalcoholic extract is important in food technology, whereas the essential oil is important in perfumery. The extract is also useful as an antibacterial and antifungal agent. In ancient times the fragrant leaves were used as a strewing herb to remove disagreeable odours and to deter pests. The candied rhizome was a confection in Europe and America. Sweet flag is used in magic rituals in New Guinea and it was also used in snuff rituals by American Indians.

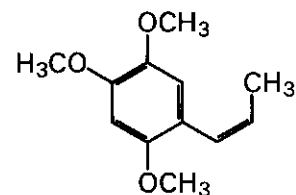
**Production and international trade** The dried rhizomes of sweet flag are traded locally on markets. Nowadays this trade is not very important, but the extremely large area of distribution

resulting from former cultivation indicates that it must have been considerable in the past. The oil is traded in somewhat larger amounts in Europe, mainly for flavouring alcoholic drinks. It is reported that annually about 200 t of rhizomes are used for manufacturing medicines in the Ukraine, and about 20 t in Germany. Almost 30 medicinal preparations which contain sweet flag are available in Europe.

**Properties** The rhizome of sweet flag is aromatic, smelling of citrus, with a bitter spicy taste. The rhizome contains 2–6(–9)% of a pale yellow to pale brown essential oil with a woody spicy odour with increasingly sweet afternotes and great tenacity. It is normally obtained by steam distillation of fresh or dried unpeeled rhizomes.

Thanks to the great amount of research done on the chemical compounds in the rhizome it is known that sweet flag oil is a source of oxygenated sesquiterpenes of great structural variety. The major chemical constituents of the essential oil are phenylpropanes, monoterpenes and thermolabile sesquiterpenoids. As many as 250 or so volatile components have been detected in the oil of the triploid European var. *calamus*, and about 100 in the tetraploid var. *angustatus*. The major constituents include  $\beta$ -asarone (cis-isoasarone), methyleugenol, cis-methylisoeugenol, geranylacetate,  $\beta$ -farnesene, shyobunone, epishyobunone, isoshyobunone, calamusenone and acorenone. The proportion of each chemical component in the oil varies among the varieties, depending on the degree of polyploidy. The concentration of  $\beta$ -asarone varies markedly; it may form as much as 4–8% of the rhizome and up to 96% of the essential oil in tetraploid Asiatic plants, but only about 0.3% of the rhizome and up to 5% of the oil in triploid European plants, but is absent (or undetectable) in diploid North American plants. The asarone is odourless, so the minor components are decisive in the fragrance of the oil.

The 2 stereoisomers  $\alpha$ -asarone (trans-isoasarone) and  $\beta$ -asarone (cis-isoasarone) are reported to have psychoactive effects. This has been attrib-



$\beta$ -asarone (cis-isoasarone)

uted to the structure, which is similar to that of amphetamines and of the hallucinogenic compound mescaline. Asarone has a relaxing effect on smooth muscle tissue, and the oil has been found to induce spasmolytic activity in rabbit intestines, aortae and uteri. Experiments with guinea-pig ilia have demonstrated that the cortex of the rhizome acts as an antispasmodic agent. In tests on laboratory animals it also showed antihistamine, anti-convulsant and antipyretic activity. It has also been found to act as neuroleptic enhancer, central nervous system depressant, carcinogen, hypothermic, hypotensive, analgesic, anti-inflammatory, bronchodilator, respiration inhibitor, hepatotoxin and antifibrillatory. However, other tests have shown negative results for many of these activities. Although  $\beta$ -asarone is reported to relax smooth muscle tissue, the American drug that does not contain this component has also been shown to be spasmolytically active. These results suggest that  $\beta$ -asarone cannot be solely responsible for the effect and that other antispasmodic compounds must also be present.

The oil is reported effective for hypotensive relief in cats and as an anticonvulsant in pregnant mice. An oral dose of 500 mg/kg of the ethanolic extract showed significant anti-secretory and anti-ulcerogenic activity in rats subjected to pyloric ligation, reserpine and cysteamine administration, and had a highly significant protective effect against cytotoxic agents; these results support the use of sweet flag for the treatment of gastropathy in traditional medicine. Extracts have shown effective antifungal and antibacterial activity, and are reported to be effective against leeches. The rhizome has shown insecticidal activity against a wide range of insect species. Both antifeedant activity and contact toxicity have been reported, and the oil can cause sterility in some insects.

The oil has shown anti-amoebic activity against *Paramecium caudatum* and nematocidal activity against *Ascaris lumbricoides*, *Toxocara canis* and *Meloidogyne incognita*, as well as acaricidal properties against the tick *Boophilus microplus*. It has also been found to inhibit the germination of weeds in cotton. Tests with ground rhizomes mixed with cotton seeds showed promising results for the use as seed protectant against the fungus *Sclerotium rolfsii* that causes damping-off disease. The active compound seems to be  $\beta$ -asarone which has toxic and sterilizing effects. Since the amount of  $\beta$ -asarone is highly dependent on the source of the plant material, care should be taken to carefully record this source. Dried rhizomes have been

found to exhibit no antiviral and antitumour activity.

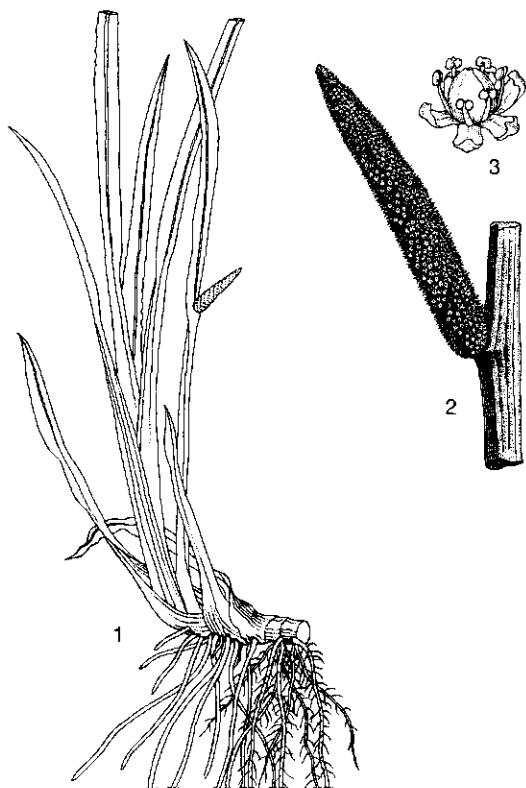
Tannins, starches, mucin, soft gums and resins are also present. Rhizomes contain approximately 10% moisture, 8% sugar, 16% protein, 2% nitrogen, 6.5% ash and 28% ethanol-soluble extractive. Under certain conditions sweet flag is poisonous, causing disturbed digestion, gastro-enteritis and persistent constipation, followed by diarrhoea and passage of blood into the faeces. The use of sweet flag is prohibited in the United States and Canada, because cancerous tumours were found in laboratory animals treated with sweet flag for long periods. The carcinogenic agent seems to be  $\beta$ -asarone, from which mutagenic (demonstrated on *Salmonella typhimurium*) and chromosome damaging properties have also been reported. In general the diploid (North American) variety void of  $\beta$ -asarone should be used for pharmaceutical applications. However, triploid European forms poor in  $\beta$ -asarone (< 0.5%) are acceptable, providing they are not used for prolonged periods. The recommended limit in Europe for flavouring is 0.1 mg/kg in foods and 1 mg/kg in alcoholic beverages and spice mixtures. A rapid and reliable thin-layer chromatographic method, allowing determination down to 0.01 mg/l is available. In some cases, the oil has been known to cause dermatitis when in contact with the skin.

The properties of the mainland Asiatic *A. gramineus* have been studied much less than those of *A. calamus*. It seems to contain less essential oil, but this oil has a high concentration of  $\beta$ -asarone (63–81%), whereas  $\alpha$ -asarone has also been reported as one of the important principles of the dry rhizome. The hexane fraction from methanolic extracts revealed potent inhibitory activity against the resistance of multi-drug resistant *Staphylococcus aureus*; benzoic acid phenylmethyl ester (benzyl benzoate) has been identified as active principle. A water extract of the dry rhizome decreased the locomotor activity of mice and increased the pentobarbital-induced sleeping time, in a dose dependent way.

**Adulterations and substitutes** In India, the rhizomes of *Alpinia galanga* (L.) Willd. and *A. officinarum* Hance are commonly used as adulterant for medicinal purposes. Neem seed oil (from *Azadirachta indica* A.H.L. Juss.) has similar insecticidal properties, as do extracts from leaves and fruits of *Melia azedarach* L., from the rhizome of turmeric (*Curcuma longa* L.) and from garlic (*Allium sativum* L.), and the oils from basil (*Ocimum* spp.), star anise (*Illicium verum* Hook.f.)

and nutmeg (*Myristica fragrans* Houtt.).

**Description** A perennial glabrous herb up to 150 cm tall; rhizome creeping, extensively branched, up to 3 cm in diameter, pale yellowish to pinkish-brown outside, whitish, sometimes slightly pinkish inside, upper surface marked with large V-shaped leaf-scars, longitudinally furrowed, under surface with circular pitted scars of rootlets arranged in irregular lines. Leaves erect, linear-ensiform, with obliquely acuminate apex, often characteristically corrugated at one side in the upper part, with distinct midrib and numerous thin parallel veins, glossy green but often reddish towards base, aromatic. Inflorescence arising from the rhizome, erect, with a cylindrical, straight or slightly curved spadix up to 10 cm long and produced from about the middle of an apparent leaf consisting of the compressed trigonous leaf-like peduncle and the leaf-like spathe forming a continuation of the peduncle. Flowers densely arranged on the spadix, bisexual, 3-merous; tepals 6, in 2 whorls, free, narrowly oblong, 2-3 mm long;



*Acorus calamus* L. - 1, plant habit; 2, inflorescence; 3, flower.

stamens 6, free, about 3 mm long, with strap-shaped filaments and orbicular-elliptical anthers dehiscent by a longitudinal slit; ovary superior, subquadrangular, 2-3-celled, stigma sessile, subconical. Fruit a 2-3-celled berry, turbinate and prismatic with pyramidal top, few-seeded, reddish. Seeds ellipsoid.

**Growth and development** Rhizomes of sweet flag can rapidly develop leaves and inflorescences under favourable conditions, e.g. in spring in temperate climates. Rhizomes show a remarkable tolerance of anaerobic conditions. Plants are usually exposed to periods of flooding and consequently to anaerobic conditions, and can survive for about 2 months in the complete absence of oxygen. The physiology of the plant is adapted to these conditions: the expression of genes enclosing glycolytic enzymes is induced during periods of submergence.

In certain populations plants often do not flower for years. In Malesia, sweet flag is even reported to flower rarely. In many areas the plant does not develop fruits. In Java, the tetraploid plants do fruit sometimes.

**Other botanical information** *Acorus* has traditionally been placed in the family *Araceae*, where it is included in the subfamily *Pothoideae* together with e.g. *Pothos* and *Anthurium* and forms the tribe *Acoreae*. However, recent taxonomic studies suggest that on the basis of morphological, anatomical, developmental and molecular evidence the genus should be placed in the monotypic family *Acoraceae*. The recent suggestion that *Acorus* is a member of the oldest extant lineage of monocotyledons is based on phylogenetic analysis from DNA sequences; there is supporting morphological, anatomical and embryological evidence. *Acorus* is generally considered to consist of 2 species, but a third species has been distinguished in China.

*A. calamus* is highly variable in many respects. The size and shape of rhizomes, leaves and spadices are greatly affected by growth conditions. The species has been subdivided primarily on the basis of genome differences. *Var. americanus* (Raf.) Wulff is diploid and fertile and occurs from North America to Siberia, *var. calamus* is triploid and sterile and occurs in Europe, the Himalayas and temperate India and certain parts of the United States of America, and *var. angustatus* Bess. is tetraploid and partly fertile and occurs in eastern and southern Asia from Japan and China to the Malesian region. The tropical ecotype of this last variety is sometimes called *var. verus* L.

Another polytypic species, *A. gramineus* Soland. ex Aiton, is diploid. It differs from *A. calamus* in its usually very narrow worm-like spadix, leaves without distinct midrib but with ribs on the margins, and tougher rhizomes and leaves. It is native to mainland south-eastern Asia (from India, Thailand and Indo-China to China) and Japan, and is used there for similar purposes as *A. calamus*.

**Ecology** Sweet flag is a component of semi-aquatic habitats, usually in eutrophic locations. It can be a vigorous invader of new sites. In Malesia, it is found along ditches, pools, fish-ponds and marshes, and is sometimes cultivated. In Java, it is found up to 2100 m altitude. Sweet flag can be planted on clayey loams and light alluvial soils.

**Propagation and planting** Sweet flag can be propagated easily from pieces of rhizome. The field is ploughed and watered prior to planting, and sometimes green manure is incorporated. The rhizome pieces to be planted are generally 6 cm long and have growing tops. They are planted at 20 cm × 20 cm. Roots start to develop 10–15 days after planting, and are soon followed by leaves. In India, sweet flag has been successfully intercropped with poplar (*Populus* sp.).

**Harvesting** Sweet flag can be harvested within one year after planting. The timing strongly affects the yield of essential oil. In temperate climates, the best period for harvesting is autumn, and the least suitable period is spring.

**Yield** In India, the average weight of individual green rhizomes harvested 10 months after planting was 175 g (95 g after drying). They contained 1.4% essential oil on average, and the highest yield was 10.4 kg/ha.

**Handling after harvest** After harvesting, the rhizomes are freed from leaves and roots and washed; they are dried unpeeled, cut in pieces and sold on the market. Storing powdered rhizomes for 2 months at 29°C and 65–75% relative humidity did not reduce their effectiveness as an insecticide.

**Genetic resources and breeding** Sweet flag has a very large area of distribution and is common in many parts of the world in habitats which are not at risk of destruction. However, locally (e.g. in certain parts of India) it is endangered or even on the verge of extinction. The great genetic variability which is correlated with differences in chemical composition should be taken into account when making germplasm collections and when breeding for special purposes.

**Prospects** Research findings suggest that sweet flag may have applications for several ailments

for which it has a historical record of use. It may still have beneficial applications in modern medicine. Its use in perfumes, foods and beverages is limited because of the carcinogenic phenylpropane derivative  $\beta$ -asarone present in the extract, but the presence of diploid populations which seem to be free from this component offers new opportunities for more extensive use in the future after selection. Sweet flag might have good prospects for commercial exploitation as a pesticide of plant origin.

**Literature** |1| Bruneton, J., 1995. Pharmacognosy, phytochemistry, medicinal plants. Lavoisier Publishing, Paris, France. pp. 463–464. |2| Dey, D. & Das, M.N., 1982. Pharmacognostic studies of *Acorus calamus* and its adulterants. *Acta Botanica Indica* 10(1): 28–35. |3| Grayum, M.H., 1987. A summary of evidence and arguments supporting the removal of *Acorus* from the Araceae. *Taxon* 36(4): 723–729. |4| Mazza, G., 1985. Gas chromatographic and mass spectrometric studies of the constituents of the rhizome of *calamus*. *Journal of Chromatography* 328: 179–206. |5| Motley, T.J., 1994. The ethnobotany of sweet flag, *Acorus calamus* (Araceae). *Economic Botany* 48(4): 397–412. |6| Phillip, J., Nair, G.S., Premalatha & Sudhadevi, P.K., 1992. Effect of planting materials and time of harvest on yield and essential oil content of rhizomes in *Acorus calamus*. *Indian Cocoa, Arecanut and Spices Journal* 16(2): 63–65. |7| Rafatulah, S., Tariq, M., Mossa, J.S., Al-Yahya, M.A., Al-Said, M.S. & Ageel, A.M., 1994. Anti-secretagogue, anti-ulcer and cytoprotective properties of *Acorus calamus* in rats. *Fitoterapia* 65(1): 19–23. |8| Röst, L.C.M., 1979. Biosystematic investigations with *Acorus*. 4. Communication. A synthetic approach to the classification of the genus. *Planta Medica* 37(4): 289–307. |9| Röst, L.C.M. & Bos, R., 1979. Biosystematic investigations with *Acorus* L. 3. Communication. Constituents of essential oils. *Planta Medica* 36(4): 350–361. |10| Stahl, E. & Keller, K., 1981. Zur Klassifizierung handelsüblicher Kalmusdrogen [About the classification of commercial *Acorus calamus* drugs]. *Planta Medica* 43(2): 128–140.

**Other selected sources** 97, 190, 194, 202, 297, 332, 348, 350, 386, 580, 597, 964, 1035, 1178, 1312, 1566.

Nguyen Van Dzu

## **Aerva Forssk.**

Fl. aegypt.-arab.: 170, cxxii (1775).

AMARANTHACEAE

*x* = unknown; *A. lanata*:  $2n = 16$ , *A. sanguinolenta*:  $2n = 36, 42, 44, 52$

**Major species** *Aerva lanata* (L.) A.L. Juss. ex Schultes, *A. sanguinolenta* (L.) Blume.

**Origin and geographic distribution** *Aerva* consists of approximately 10 species and occurs in tropical and subtropical regions of Africa and Asia. Africa is considered the centre of diversity. Two species occur within the Malesian region, especially in regions with a monsoon climate.

**Uses** Common medicinal uses of *Aerva* include applications as diuretic (valued in cases of lithiasis and catarrh of the bladder, and for prostatic ailments) and as vermifuge. In India the dried plants are used against diabetes and malaria. The seeds are used against bronchitis. In Sri Lanka small clumps of *A. lanata* are frequently grown in gardens to make a medicinal tea. The use of cultivated reddish forms of *A. sanguinolenta* to treat haematuria and menstruation problems has been considered as doctrine of the signature. In India the leaves are made into a paste and applied for the treatment of wounds. *A. sanguinolenta* is also cultivated as an ornamental.

**Production and international trade** *Aerva* is usually cultivated in home gardens for use in local medicine and as an ornamental, and the plants do not enter the international trade.

**Properties** Several alkaloids have been isolated, including aervine, methylaervine, aervoside, aervolanine, canthine-6-one and  $\beta$ -carboline-1-propionic acid in *A. lanata*. Several flavonoid glycosides have also been isolated and identified from this species, e.g. narcissin, aervitrin and 4 flavonoid  $\beta$ -coumaroylglycosides, together with 2 feruloylamides and other phenolic compounds. The compounds  $\beta$ -sitosterol, campesterol and chrysin have been isolated from *A. lanata* plants cultivated in Egypt and  $\beta$ -sitosterol, daucosterol, syringic acid, vanillic acid, feruloyltyramine and feruloylhomovanillylamine have been isolated from plants cultivated in Russia. Glucose, galactose and xylose were the predominant carbohydrates in hydrolysates from leaves and flowers.

Preliminary tests in India on rats to study the effects of fresh juice and aqueous extracts of *A. lanata* on the chemically induced nephrotic syndrome, and as anti-inflammatory, diuretic and steroidal agent showed positive results. An ethylacetate extract of the dried whole plant exhibited antimalar-

ial activity. The diuretic effect has also been studied in humans. An extract (200 ml of a 50 g/l infusion) was reported to induce diuresis; the urine output was significantly elevated with extract from fresh plants, when compared with the control group which received the same quantities of water or isotonic saline. Furthermore, of the different parts of the plants tested, the flowers were found to be most effective in inducing diuresis. However, the nature of the diuresis (aquaretic or saluretic) was not defined, since only the urine flow was estimated. In a more comprehensive double-blind setting with healthy volunteers these results could not be repeated: the *A. lanata* extract did not significantly increase urine flow, sodium excretion, potassium excretion, or urine and plasma osmolality as compared to water or a diluted infusion of tea prepared to resemble the *A. lanata* extract as closely as possible in colour and taste. The influence on the formation of urinary (bladder) stones was investigated in the rat. It was shown that *A. lanata* did not affect the formation or dissolution of urinary stones of the phosphate type.

Leaves of *A. sanguinolenta* contain a potent inhibitor of plant virus infection. Leaf extracts mixed with the virus inoculum of tobacco mosaic virus and sunhamp rosette virus caused almost total inhibition of virus infectivity. The resistance to virus infection induced by the extract is systemic and long lasting. Leaf extracts of *A. sanguinolenta*, applied as 4% foliar spray, delayed the appearance of disease symptoms caused by yellow mosaic virus on mung bean, but they could not be used to control the disease.

**Adulterations and substitutes** Java tea (*Orthosiphon aristatus* (Blume) Miq.) is another herb which is reputed for its diuretic activity, and which is prescribed to treat similar complaints and in similar preparations.

**Description** Perennial, erect or somewhat clambering, dioecious or polygamous herbs up to 200 cm tall, often almost woody at base, often divided from near the base into ascending or erect branches; branches often unbranched for a considerable length, terete, densely clothed with appressed or patent whitish hairs. Leaves alternate or opposite (often on a single specimen), simple and entire, densely clothed with appressed white hairs on both surfaces (but especially beneath); petiole short, stipules absent. Inflorescence an axillary and terminal spike, solitary or fascicled and sometimes forming a paniculate inflorescence. Flowers small, usually bisexual but sometimes unisexual, solitary in the axil of persistent bracts,



subtended by 2 bracteoles, white or tinged with purple; tepals 5, free, hairy; stamens 5, filaments connate at base, free parts subulate, about half as long as tepals, alternating with shorter stamens, anthers 2-celled; ovary superior, 1-celled, compressed, glabrous, style very short with 1-2 stigmas. Fruit a much compressed utricle, about 1 mm in diameter, bursting irregularly, 1-seeded. Seed reniform, shiny black.

**Growth and development** *A. lanata* is often reported not to flower before the second year, but flowering sometimes does occur in the first year. In India two forms have been distinguished, one reddish with a deep penetrating taproot having numerous lateral roots, short branches and leaves and comparatively long and numerous spikes, and the other green in colour with a short, thin taproot having few lateral roots, longer branches, larger leaves and comparatively short and few spikes. The first form is apparently an adaptation to drier conditions. Intermediate forms have been found.

**Other botanical information** Three *Aerva* species have been reported to occur in the Malesian area, but one of these (*A. curtisii* Oliv.) has been transferred to a new genus *Psilotrichopsis* together with the continental South-East Asian species *A. cochinchinensis* Gagn.

**Ecology** Both *Aerva* species occur especially in periodically dry areas and in dry locations. They are found in sunny or slightly shaded sites, e.g. along roadsides, in waste places, in brushwood and hedges, and are common in many regions. They can be a weed on cropped land.

**Propagation and planting** *A. sanguinolenta* may be propagated by stem cuttings. Plant growth regulators (e.g. ethyl hydrogen-1-propyl phosphonate) promote root formation and subsequent shoot growth.

**Genetic resources and breeding** *Aerva* species occur widespread in anthropogenic habitats and are not at risk from genetic erosion.

**Prospects** *Aerva* may have some prospects as a medicinal plant that is easy to cultivate on a small scale e.g. in home gardens. The diuretic properties that have been claimed since antiquity, however, should be clarified first to define their exact nature, before recommendations can be given for a possible application of *A. lanata* as a drug.

**Literature** [1] Amin, K.M.Y., Ahmed, S. & Khan, N.A., 1994. Anti-nephrotic syndrome ethnic drug Bishiri Booti (*Aerva lanata*) – experimental study of relevant pharmacological actions. Ethnobiology in human welfare: abstracts of the fourth international congress of ethnobiology, Lucknow,

Uttar Pradesh, India, 17–21 November, 1994. p. 94. [2] Backer, C.A., 1949. Amaranthaceae. In: van Steenis, C.G.G.J. (Editor): Flora Malesiana. Series 1, Vol. 4. Noordhoff-Kolff N.V., Djakarta, Indonesia. pp. 84–86. [3] de Padua, L.S., Lugod, G.C. & Pancho, J.V., 1981. Handbook on Philippine medicinal plants. Vol. 3. Technical Bulletin Vol. V, No 2. Documentation and Information Section, Office of the Director of Research, University of the Philippines at Los Baños, the Philippines. p. 5. [4] Nirmala Mary, T. & Aruna Prabha, S., 1980. Ecology of *Aerva lanata* Juss. Geobios 7(6): 285–286. [5] Pervykh, L.N., Karasartov, B.S. & Zapesochnaya, G.G., 1992. A study of the herb *Aerva lanata* IV. Flavonoid glycosides. Chemistry of Natural Compounds 28(5): 509–510. [6] Singh, S.S., 1992. Rooting and regeneration potential of stem cuttings of *Aerva sanguinolenta* L. as influenced by ethyl hydrogen-1-propyl phosphonate (Niagara). Acta Botanica Indica 20(2): 294–296. [7] Udupihille, M. & Jiffry, M.T.M., 1986. Diuretic effect of *Aerva lanata* with water normal saline and coriander as controls. Indian Journal of Physiology and Pharmacology 30(1): 91–97. [8] Verma, H.N. & Srivastava, A., 1985. A potent systematic inhibitor of plant virus infection from *Aerva sanguinolenta* Blume. Current Science (Bangalore) 54(11): 526–528. [9] Zapesochnaya, G.G., Kurkin, V.A., Okhanov, V.V. & Miroshnikov, A.I., 1992. Canthin-6-one and  $\beta$ -carboline alkaloids from *Aerva lanata*. Planta Medica 58(2): 192–196. [10] Zapesochnaya, G.G., Pervykh, L.N. & Kurkin, V.A., 1991. A study of the herb *Aerva lanata*. Chemistry of Natural Compounds 27(3): 336–340.

#### *Selection of species*

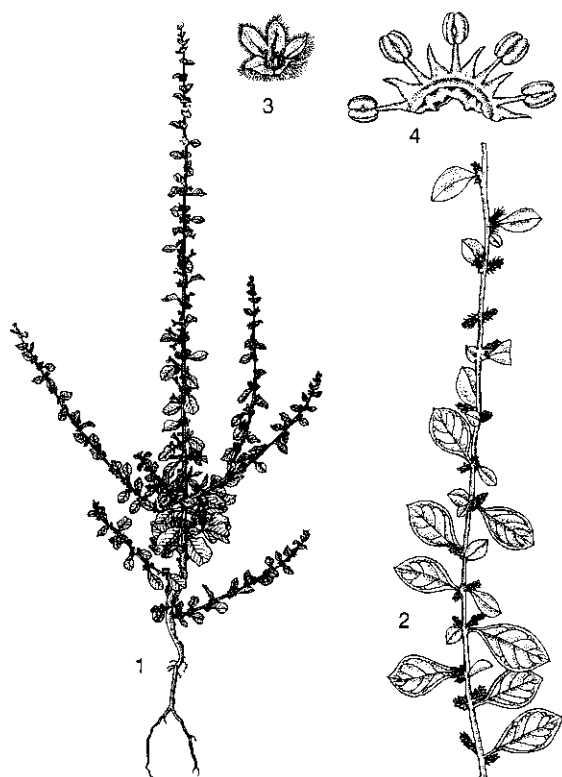
***Aerva lanata* (L.) A.L. Juss. ex Schultes**  
Roemer & Schultes, Syst. veg. 5: 564 (1819).

**Synonyms** *Achyranthes lanata* L. (1753), *Illecebrum lanatum* (L.) L. (1771), *Achyranthes villosa* Forssk. (1775).

**Vernacular names** Indonesia: katumpangan uler, rumput upas-upasan. Philippines: tabangahas, apugapugan, pamaynap (Tagalog), karlatan (Iloko). Vietnam: mao v[ix] l[oo]ng.

**Distribution** Africa, Madagascar, the Seychelles and other islands in the Indian Ocean and southern Asia from Arabia to India, Sri Lanka, Indo-China and Malesia (Sumatra, Bangka, Java, the Philippines, Timor, the Aru Islands and southern and south-eastern New Guinea).

**Uses** In the Philippines, a decoction is consid-



*Aerva lanata* (L.) A.L. Juss. ex Schultes - 1, plant habit; 2, branchlet with young inflorescences; 3, flower; 4, opened staminal tube with stamens and staminodes.

ered to be an efficacious diuretic and is used against catarrh of the bladder and gonorrhoea. Leaves steeped in hot water are applied to swellings in Indonesia. In India, the roots are additionally used to treat headache, as demulcent, to cure coughs and as a vermifuge.

**Observations** A perennial erect herb up to 110 cm tall, main branches and upper part of the stem often unbranched for a considerable length, leafy and flowering almost throughout, internodes usually shorter than 2 cm; leaves alternate, ovate-elliptical to obovate, 0.5–5 cm × 0.3–3 cm; spikes up to 2.5 cm long, in the axil of normal leaves, never forming a loosely branched paniculate inflorescence; tepals 1–1.5 mm long, stigmas 2, spreading; fruiting spike easily breaking up. *A. lanata* is locally common along roadsides, in abandoned fields and waste places, in Java up to 100 m altitude.

**Selected sources** 57, 93, 96, 190, 332, 497, 580, 891, 1041, 1127, 1178, 1202, 1469, 1470, 1483, 1641, 1642, 1643, 1644.

### *Aerva sanguinolenta* (L.) Blume

Bijdr. fl. Ned. Ind.: 547 (1826).

**Synonyms** *Achyranthes sanguinolenta* L. (1762), *Aerva scandens* (Roxb.) Wallich ex Moq. (1849), *Aerva timorensis* Moq. (1849).

**Vernacular names** Indonesia: ki sambang (Sundanese), sambang colok, gondang kasih (Javanese). Laos: do:k khaix ped (Luang Prabang). Thailand: khrua khao tok (northern), yaa dok khao (central), phan nguu yai (Saraburi). Vietnam: m[oo]ng g[af] (Phu Khanh), rau chua, mao v[ix] d[or].

**Distribution** From India, Indo-China, southern China and Taiwan to Thailand and the Malesian region (Java, the Lesser Sunda Islands, Sulawesi, the Moluccas and the Philippines).

**Uses** Decoctions of young branches are used internally against haematuria and irregular or painful menstruation. *A. sanguinolenta* is cultivated as an ornamental, particularly the purplish tinged form, e.g. in Java.

**Observations** A perennial, erect or clambering herb up to 200 cm tall, sometimes branched, internodes often longer than 3 cm; leaves alternate or opposite, ovate-elliptical, oblong or lanceolate, 1.5–7.5 cm × 0.5–4.5 cm; spikes up to 5 cm long, partly in the axil of normal leaves, partly in the axil of bracts and often forming a loosely branched paniculate inflorescence; tepals 2–2.5 mm long, stigma 1, entire or obscurely 2-lobed; fruiting spike dense and thick, not easily breaking up. *A. sanguinolenta* is locally common in abandoned fields, brushwood and hedges, in Java up to 200 m altitude, but in Indo-China up to 2000 m.

**Selected sources** 93, 96, 580, 750, 816, 868, 1367, 1368, 1469, 1470, 1522.

N. Bunyapraphatsara & R.H.M.J. Lemmens

### *Ageratum* L.

Sp. pl. 2: 839 (1753); Gen. pl. ed. 5: 363 (1754).

COMPOSITAE

$x = 10$ ; *A. conyzoides*:  $2n = 20, 40$ , *A. houstonianum*:  $2n = 20$

**Major species** *Ageratum conyzoides* L.

**Vernacular names** Goatweed, billy goat weed (En).

**Origin and geographic distribution** *Ageratum* comprises approximately 30 species, all of which – except for *A. conyzoides* and *A. houstonianum* – are restricted to the Americas and adjacent West Indies. Goatweed is a pantropical weed which originates from South and Central Ameri-

ca. It extends about 30° north and south of the equator.

*A. conyzoides* was already being cultivated as an ornamental at the end of the 17th Century, in Europe. *A. houstonianum* was not cultivated before about 1820, but is now a very popular garden plant, whereas *A. conyzoides* has fallen out of favour. Both species occur as a weed in South-East Asia, but *A. conyzoides* is more widespread and common.

**Uses** *A. conyzoides* plays a role in traditional medicine in many parts of the world. It is widely used externally to treat skin diseases, wounds (both disinfection and haemostasis), ulcers and boils; internally as febrifuge and to treat diarrhoea and haemorrhages. Local uses reported include applications as an emetic (internally), and to treat eye diseases (externally), pneumonia (externally), sore throat (infusion as syrup), stomach-ache (internally), gonorrhoea and catarrh (internally). In Indonesia, an infusion of the roots is used against fever, and an infusion of the leaves to wash sore eyes, and to treat stomach-ache and wounds. In Malaysia, leaves are used externally to heal wounds, cuts, scratches and itches and to alleviate tooth-ache; a decoction of the root is taken for treating coughs and a decoction of the whole plant is taken against asthma. The juice of fresh leaves is widely used as a vulnerary in the Philippines, and leaves cooked in coconut oil are also applied to wounds. A decoction of the herb is used to treat stomach troubles. In Papua New Guinea, a solution of crushed leaves is taken to treat diarrhoea and juice from squeezed leaves is used to treat sore eyes; the leaves are used in New Britain internally against fever and dysentery and externally to heal wounds. In Thailand, the leaves are used for treating wounds, itching and eye inflammations. The whole plant is used as antipyretic, diuretic, carminative, anti-amoebic and emmenagogue. In Vietnam, *A. conyzoides* is reported to be effective in the treatment of allergic sinusitis and rhinitis; it is also used in hair care. Extracts may be used as insecticide.

Both *A. conyzoides* and *A. houstonianum* are sometimes grown as ground cover, e.g. in rubber plantations in Java. In China, *A. conyzoides* is used as a cover crop in citrus plantations, and is reputed to be effective in the biocontrol of mites as it hosts predacious mites. *A. houstonianum* is often cultivated as annual ornamental, in tropical as well as temperate regions.

**Production and international trade** Goatweed is only used locally medicinally and is not

traded on the international market. It is only planted on a fairly large scale in China. Goatweed is rarely cultivated on a larger scale to obtain insecticides, since the active compounds can be synthesized.

**Properties** Goatweed has a disagreeable odour which has been described as the smell of a billy-goat or of salty shrimps. When dried it smells of coumarin.

Both goatweed species contain an essential oil (in *A. conyzoides* 1.6% w/v), which can be isolated from the fresh leaves and stems. This oil is characterized by the presence of volatile chromene derivatives (up to 85%) of the precocene (e.g. 7-methoxy-2,2-dimethylchromene (precocene I), and ageratochromene (precocene II)) and enecalin type. The proportions of precocene I and II differ between the species: the oil of *A. conyzoides* is rich in precocene I (about 80%) and poor in precocene II (less than 1%), and that of *A. houstonianum* contains approximately equal amounts (23–32% of precocene I and 24–44% of precocene II). The presence of terpenes as well as of chromenes has been established;  $\beta$ -caryophyllene is the main constituent (10% in *A. houstonianum*) after the precocenes of both oils.

Analysis of the aerial parts of *A. conyzoides* yielded 11 chromenes in total, and also the lignan (+)-sesamin and the sesquiterpene caryophyllene-epoxide (caryophyllene oxide). Another study found the largest amounts of precocenes in the leaves, followed by the flowering heads, whereas stems and roots had only minor amounts.

The genus *Ageratum* is known to be rich in flavonoids, especially of the polymethoxygenated flavone type. The following have been identified in *A. conyzoides*: 5,6,7,5'-tetramethoxy-3'4'-methylenedioxyflavone, 5,6,7,8-tetramethoxy-3'4'-methylenedioxyflavone (= linderoflavone B), 5,6,7,8,5'-pentamethoxy-3'4'-methylenedioxyflavone (= eupalestin), 5,6,7,8,3',4',5'-heptamethoxyflavone (= 5'-methoxynobiletin), 5,6,7,8,3',4'-hexamethoxyflavone (= nobiletin), 5,6,7,3',4',5'-hexamethoxyflavone, 5,6,7,3',4'-pentamethoxyflavone (= sinen-setin) and 5,6,7,3',4',5'-hexamethoxy-8-hydroxyflavone. Flavones from *A. houstonianum* include 5,6,7,8-tetramethoxy-3'4'-methylenedioxyflavone (= lucidin dimethylether), eupalestin, 5,6,7,8,2',4',5'-heptamethoxyflavone (= agecorynin C), 5,6,7,8,2',3',4',5'-octamethoxyflavone (= agehoustonin A) and 5,6,7,2',3',4',5'-heptamethoxyflavone (= agehoustonin B).

The pyrrolizidine alkaloids 9-angeloylretronecine, lycopsamine and echimidine have been isolated

from *A. conyzoides*. Furthermore, both species contain friedelin,  $\beta$ -sitosterol and stigmasterol; *A. houstonianum* additionally contains friedelan-3 $\beta$ -ol and also accumulates benzofuran derivatives in the roots.

Precocenes I and II have been synthesized in a single step in 2 hours, starting from a substituted monophenol and 3-methyl-2-butenal in pyridine at 140°C.

Tests with oral administration of a leaf extract to rats in Africa showed that *A. conyzoides* has analgesic properties, but less than morphine. In tests with mice, an extract of whole plants induced hemagglutination against sheep red blood cells at a concentration of 24  $\mu$ g/ml. A fairly good antiphlogistic effect by comparison with hydrocortison and inhibiting activity on experimentally induced tumour development in mice have been reported from Vietnam. The flavones have been suggested as the substances responsible for the promotion of wound healing. Crude material isolated from *A. conyzoides* leaves exhibited antibacterial activity in vitro against *Staphylococcus aureus*. *Ageratum* oil showed antibacterial activity against 20 bacteria in a test using 22 bacteria.

The essential oil has antifungal properties; in tests it inhibited the growth of *Alternaria alternata*, *Aspergillus* spp., *Colletotrichum truncatum*, *Fusarium oxysporum*, *Helminthosporium tericum*, *Penicillium italicum*, *Rhizoctonia solani* and *Trichoderma viride*. An aqueous extract of *A. conyzoides* also showed nematocidal activity against *Meloidogyne incognita*. Moreover, it exhibited insecticidal activity against diamondback moth (*Plutella xylostella*) in cruciferous crops, pulse beetles (*Callosobruchus chinensis* and *C. maculatus*) in stored leguminous seed, and corn weevils and red flour beetles (*Tribolium castaneum*) in stored maize. At concentrations of less than 10 mg/l it significantly inhibits emergence of adult mosquitoes; this inhibition has been attributed to the antijuvenile hormone activity of the chromenes. Precocene I and precocene II are able to induce precocious metamorphosis, cause sterilization and/or force diapause in certain insects, especially Heteropterans. The methoxyflavones possibly also have toxic effects on insects. A chloroform extract of leaves showed insecticidal activity against *Drosophila melanogaster* and *Dysdercus cingulatus* which was comparable to the standard insecticide malathion and better than the activity of the natural insecticide rotenone. Extracts derived from flowers of *A. houstonianum* have also shown ovicidal, antifeedant and repellent proper-

ties against several insects; acetone extracts were more effective than ethyl acetate extracts, and leaf and shoot extracts were less effective than flower extracts. Precocene II showed toxicity for American dog ticks (*Dermacentor variabilis*). A 2 cm thick layer of powdered leaves can control potato tuber moth (*Phthorimaea operculella*) in stored potatoes for up to 120 days. Locusts (*Locusta migratoria*) fed on goatweed during the nymphal stage were sterile; this implies that goatweed might be used for biological control if planted near breeding sites or on possible migration routes. Whitefly was disrupted in its development by *A. houstonianum*, though female insects were attracted by the plant to lay their eggs; this too indicates good prospects for biological control. Antiviral activity against some legume viruses has also been reported.

In tests with rats in the Philippines, crude aqueous extracts of *A. conyzoides* applied through stomach intubation caused severe poisoning at concentrations of 15–20% after 5 days. The extract retarded follicular development in the ovary, and caused inflammation and degeneration of liver cells. Goatweed is also suspected of hepatotoxicity in livestock. Cattle deaths from liver damage in northern Sumatra have been attributed to pyrrolizidine alkaloid poisoning.

Goatweed shows allelopathic effects on crops such as rice and wheat by inhibiting seed germination and root elongation of seedlings.

Goatweed pollen is reported to cause commonly respiratory tract allergy, even in very low concentration.

**Adulterations and substitutes** Other plant sources of insecticides include *Azadirachta indica* A.H.L. Juss., *Derris elliptica* (Wallich) Benth. and other composites such as *Tagetes* spp. and *Tanacetum cinerariifolium* (Trev.) Schultz-Bip. Flavones with supposedly anti-inflammatory activity are also found in other *Compositae* such as *Achillea millefolium* L.

**Description** Annual erect herbs, at the base sometimes decumbent and rooting, up to 120 (–150) cm tall; roots fibrous. Leaves opposite in lower part of plant, higher ones alternate, simple, serrate-crenate above the entire base, with sparse long hairs above and below, glandular on lower side, pinnately veined or faintly 3-veined, distinctly petiolate; stipules absent. Inflorescence consisting of (1–)4–18 peduncled heads arranged in cymose clusters; involucre campanulate, involucral bracts 2–3-seriate, 2-ribbed, green with a pale or reddish-violet top; head 60–100-flowered. Flowers

usually bisexual, 5-merous; corolla tubular in varying shades of blue and lavender or white, corolla lobes short, triangular; stamens inserted on inner side of the corolla, with fused anthers forming a ring around the style, with an apical appendage; ovary inferior, 1-celled, with 1 erect ovule on the bottom of the cell, style with 2 long filiform arms with thickened hairy tops, far exerted from the corolla. Fruit an oblong achene, dry and indehiscent, 1–2 mm long, 5-angular, scabrous on the angles, blackish with a pale base, on the outer margin of the top with a pappus consisting of 5(–6) free, membranous, awn-tipped, off-white scales 1.5–3 mm long. Seedling with epigeal germination; cotyledons leafy, orbicular, glabrous; hypocotyl up to 5 mm long, epicotyl absent or extremely short.

**Growth and development** A single plant of goatweed can produce up to 40 000 seeds. There is no marked dormancy and germination is promoted by light and inhibited by burial. Seed of *A. conyzoides* can germinate at comparatively low temperatures (10–20°C), which explains its occurrence at higher altitudes, whereas maximum temperature for germination is around 30°C. For seed of *A. houstonianum*, the minimum temperature for germination is reported as 20°C and the maximum temperature 35°C.

The life cycle can be completed in less than 2 months. Flowers and fruits may be present throughout the year. The occurrence of male sterility has been documented for both *A. conyzoides* and *A. houstonianum*. The usually male fertile *A. conyzoides* is self-pollinated and cross-pollinated by insects. *A. houstonianum* has been reported as self-incompatible. Fruits are dispersed by wind but may also cling to the fur of animals.

**Other botanical information** *A. conyzoides* and *A. houstonianum* are closely related, and the latter has even been considered as merely a variety of the former (*A. conyzoides* var. *houstonianum* (Miller) Sahu). Although both resemble each other closely and are often confused, there are some reliable morphological distinguishing characteristics, particularly in the involucre bracts and leaf-bases. There are also minor differences in number of flowers per head (on average more in *A. houstonianum*), and length of corolla, anthers and style (on average longer in *A. houstonianum*). Both grow in similar habitats and both may become weeds. *A. conyzoides* has a short-day ecotype and a day-neutral ecotype.

Several cultivars of *A. houstonianum* are popular

garden plants, e.g. cultivar 'Blaue Donau' in Europe.

**Ecology** Goatweed can be found from sea-level up to 2500 m altitude. It is a very common weed in numerous annual and perennial crops. It commonly grows in roadsides, waste places and grassy fields. In shifting cultivation in Thailand, *A. conyzoides* is one of the dominant species in the second year after clearing of the forest, together with *Chromolaena odorata* (L.) R. King & H. Robinson. *A. houstonianum* has a more restricted altitudinal range than *A. conyzoides*; in Java it occurs up to 1700 m altitude.

**Propagation and planting** Propagation by stem cuttings is more efficient in *A. houstonianum* than by seed or root, resulting in rapid growth and reproduction. Protoplasts have been isolated from leaves, stems and callus of *A. houstonianum*, and callus has regenerated.

**Diseases and pests** *A. conyzoides* is the host to several pathogens causing serious diseases in commercial crops: for *Pseudomonas solanacearum* attacking Irish potato in India and for the nematodes *Meloidogyne incognita* (despite the reported nematocidal activity against the same species) and *M. javanica* causing yellowing and wilting in black pepper and vegetables in the Philippines and *Pratylenchus* sp. attacking upland rice in the Philippines. A geminivirus called ageratum yellow vein virus, which is transmitted by the whitefly *Bemisia tabaci*, causes a vein-yellowing disease in *A. conyzoides*; it has been reported from Malaysia and Singapore. A similar virus, the tobacco krupuk virus, is reported for Indonesia. *A. conyzoides* is a host of cotton bollworm in Thailand and for *Nysius inconspicuus* in India (a pest on sesame). *A. houstonianum* has been reported in India as a host for green scale (*Coccus viridis*) which is a major pest of coffee.

Both species are important weeds in arable and plantation crops. The lepidopterous insect *Parauchaetes pseudoinsulata* can complete its life cycle on *A. conyzoides* and might be a promising agent to control this species biologically in areas where it is a serious weed.

**Genetic resources and breeding** Both goatweed species are widespread in anthropogenic habitats, suggesting a broad genetic variability. The existence of different photoperiodic ecotypes is of interest for breeding, as it provides an opportunity to obtain planting material adapted to specific conditions. Breeding programmes could focus on optimum medicinal and insecticidal activity which is reflected in high concentrations of active

compounds such as precocenes. The breeding value of male sterility is rated as low.

**Prospects** Goatweed has several widespread and comparatively well-documented medicinal applications. It has outstanding prospects as an insecticide. The insecticidal properties offer possibilities for the control of mosquitoes that are vectors of malaria and filariasis, and for the control of insects like locusts, bugs and mites. The effect on phytophagous mites in citrus deserves further research. Goatweed is easy to cultivate, which makes it suitable for large-scale production for industrial use as well as for home gardening for local use.

Since goatweed establishes spontaneously, it is a cheap and beneficial soil cover crop. Moreover, certain forms of goatweed are attractive ornamental plants.

**Literature** |1| Adesogan, E.K. & Okunade, A.L., 1979. A new flavone from *Ageratum conyzoides*. *Phytochemistry* 18(11): 1863–1864. |2| Bioka, D. & Mabika, A., 1993. Effet analgésique d'un extrait brut d'*Ageratum conyzoides* chez le rat [Analgesic effect of a crude extract from *Ageratum conyzoides* on the rat]. In: Schilcher, H., Phillipson, J.D. & Loew, D. (Editors): *First World Congress on Medicinal and Aromatic Plants for Human Welfare*, Maastricht, the Netherlands, July 19–25, 1992. Pharmacology, phytotherapy, human welfare, regional aspects. *Acta Horticulturae* No 332: 171–176. |3| Durodola, J.I., 1977. Antibacterial property of crude extracts from a herbal wound healing remedy – *Ageratum conyzoides*. *Planta Medica* 32(4): 388–390. |4| Gonzalez, A.G., Aguiar, Z.E., Grillo, T.A., Luis, J.G., Rivera, A. & Calle, J., 1991. Chromenes from *Ageratum conyzoides*. *Phytochemistry* 30(4): 1137–1139. |5| Gonzalez, A.G., Aguiar, Z.E., Grillo, T.A., Luis, J.G., Rivera, A. & Calle, J., 1991. Methoxyflavones from *Ageratum conyzoides*. *Phytochemistry* 30(4): 1269–1271. |6| Johnson, M.F., 1971. A monograph of the genus *Ageratum* L. (Compositae-Eupatorieae). *Annals of the Missouri Botanical Garden* 58(1): 6–88. |7| Kaul, M.L.H. & Neelangini, 1989. Male sterility in diploid *Ageratum conyzoides* L. *Cytologia* 54(3): 445–448. |8| Ohta, T., Kuhr, R.J. & Bowers, W.S., 1977. Radiosynthesis and metabolism of the insect antijuvenile hormone precocene II. *Journal of Agricultural and Food Chemistry* 25(3): 478–481. |9| Sharma, V.S., 1987. Comments on the identity of *Ageratum conyzoides* L., and *A. houstonianum* Mill. – two naturalized weeds in India. *Feddes Repertorium* 98(11–12): 557–560. |10| Siebertz, R., Proksch, P. & Witte, L., 1990. Accumulation and

biosynthesis of the chromenes precocene I and II in *Ageratum houstonianum*. *Phytochemistry* 29(7): 2135–2138.

#### *Selection of species*

### ***Ageratum conyzoides* L.**

Sp. pl. 2: 839 (1753).

**Vernacular names** Goatweed (En). Eupatoire bleue (Fr). Indonesia: babadotan (Sundanese), wedusan (Javanese), dus-bedusan (Madurese). Malaysia: tahi anjing, rumput pereh jarang, rumput sekedok (Peninsular). Philippines: bulakmanok (Tagalog), singilan (Iloko), bahug-bahug (Panay Bisaya). Thailand: thiam mae haang (Loei), saapraeng saapkaa (Chiang Mai), ya saap raeng (Ratchaburi). Vietnam: c[aa]y b[oo]ng c[uws]t heo, c[aa]y hoa c[uws]t l[ow]j[n], c[aa]y b[oo]ng th[us]i.

**Distribution** Originating from Central and South America, but now a pantropical weed that is very common throughout India, Burma (Myanmar), Indo-China, southern China, Thailand and Malesia.

**Uses** The most widespread medicinal uses are externally to heal wounds and to treat skin diseases, and internally to treat diarrhoea, as a febrifuge and as an anti-allergenic agent. The plant yields an insecticide. It is sometimes planted as a ground cover in plantations, e.g. of rubber and citrus.

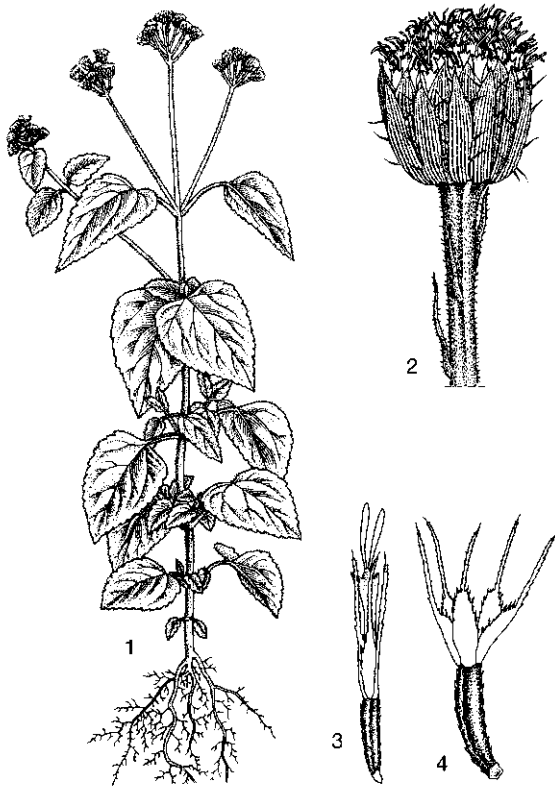
**Observations** An annual erect herb, at the base sometimes decumbent and rooting, up to 120(–150) cm tall, stems with rather long hairs on the nodes and younger parts; leaves ovate, triangular-ovate or rhomboid-ovate, (0.5–)1–10 cm × 0.5–7 cm, with obtuse or rounded base; head 4–6 mm long, 60–75-flowered, outermost involucre bracts beset with only simple eglandular hairs, inner involucre bracts with abruptly contracted apex; corolla 1–2.5 mm long; fruit glabrous or very sparingly hairy. *A. conyzoides* is very common in fields, roadsides and waste places up to 2500 m altitude.

**Selected sources** 18, 96, 156, 202, 332, 350, 383, 495, 496, 549, 580, 597, 614, 616, 685, 713, 876, 928, 929, 996, 1034, 1035, 1090, 1126, 1131, 1178, 1246, 1268, 1294, 1324, 1386, 1408, 1517, 1570, 1593, 1659.

### ***Ageratum houstonianum* Miller**

Gard. dict. ed. 8: *Ageratum* No 2 (1768).

**Synonyms** *Ageratum mexicanum* Sims (1825).



*Ageratum conyzoides* L. - 1, plant habit; 2, flowering head; 3, flower; 4, fruit with pappus.

**Vernacular names** Goatweed (En). Probably many of the vernacular names listed under *A. conyzoides* also refer to *A. houstonianum*.

**Distribution** Originating from Central America, but cultivated in tropical, subtropical and temperate regions. It is found naturalized in many warmer regions including China, India and locally in South-East Asia (e.g. Peninsular Malaysia, Java, the Philippines, Vietnam).

**Uses** Probably *A. houstonianum* has similar medicinal applications as *A. conyzoides*, but there is little information in literature about its actual uses. The plant yields an insecticide, and it is commonly planted as ornamental.

**Observations** An annual erect herb, at the base sometimes creeping and rooting, up to 70(-90) cm tall, stems clothed with patent white hairs; leaves subtriangular, ovate to deltoid, 2-8.5(-9.5) cm × (1.5-)3-6.5(-8) cm, with cordate to truncate base; head 5.5-7 mm long, 75-100-flowered, outermost involucre bracts with both simple hairs and shorter glandular hairs, inner involucre bracts gradually tapering into an acute

apex; corolla 2.5-3 mm long; fruit hairy. *A. houstonianum* is locally common in fields, roadsides and waste places up to 1700 m altitude.

**Selected sources** 96, 232, 549, 614, 685, 929, 966, 1073, 1175, 1268, 1324, 1354, 1438.

Slamet Sutanti Budi Rahayu, Rina Ratnasih  
Irwanto & L.J.G. van der Maesen

## Allium L.

Sp. pl. 1: 294 (1753); Gen. pl. ed. 5: 143 (1754).

ALLIACEAE

$x = 8$ ; *A. cepa*:  $2n = 16$ , *A. chinense*:  $2n = 32$ , *A. fistulosum*:  $2n = 16$ , *A. sativum*:  $2n = 16$ , *A. tuberosum*:  $2n = 32$

**Major species** *Allium cepa* L., *A. sativum* L.

**Vernacular names** Onion (En). Indonesia: bawang.

**Origin and geographic distribution** *Allium* comprises 500-700 species and is mainly distributed in the Northern Hemisphere. Centres of diversity are in regions that are seasonally dry, particularly around the Mediterranean in Europe, in central Asia and North America. The Malesian region has no indigenous species, but several introduced species are cultivated.

**Uses** *Allium* species have been used since antiquity for their antiseptic properties. Besides its antibacterial properties, onion (*A. cepa*) juice is known for its diuretic, lipid- and blood-pressure lowering and anti-asthmatic/anti-allergic properties. Onion is used in traditional medicine to treat boils, wounds, stings and felons externally, and internally to relieve coughs, bronchitis, asthma, gastro-intestinal disorders (e.g. flatulence, diarrhoea) and headache.

Garlic (*A. sativum*) is valued worldwide as a 'panacea' to cure an array of diseases and to strengthen the body. Phytotherapeutic products based on garlic are traditionally used to treat minor vascular disorders. Garlic is recommended against high blood pressure. Oral administration of garlic (and, to a lesser extent, onion) juice or oil has been reported to prevent hyperlipemia, arteriosclerosis and myocardial infarction. Other traditional uses include the treatment of coughs, bronchitis and gastro-intestinal disorders (flatulence). Garlic oil is used as rubefacient (to treat muscle pains, lumbago, arthritis and ischias) and as vermifuge (especially against enterobiasis). Garlic juice is used externally in the treatment of *Taenia versicolor*, ringworms and chronic wounds. It is also believed that garlic juice promotes

longevity and has aphrodisiac properties.

*A. chinense* ('rakkyo') is used against fever, stomach-ache and eye-infections. In China, it is reputed to be effective in the treatment of stenocardia, angina pectoris and so-called stagnant blood, and it is included in some traditional preparations. *A. tuberosum* (Chinese chives) is reportedly effective against tumours, toothache and intestinal disorders. Welsh onion (*A. fistulosum*) is considered to have strong stimulant properties.

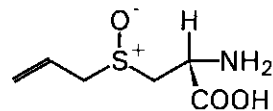
*Allium* species are well known for their worldwide use as a vegetable and condiment, which often goes hand in hand with their attributed medicinal properties.

**Production and international trade** Onion (*A. cepa*) is economically the most important *Allium* crop in South-East Asia. In 1990 Indonesia produced about 500 000 t onion (mainly shallot) and 100 000 t garlic. In that year the Philippines exported 12 000 t onion and Thailand 18 000 t, but other countries of South-East Asia imported large amounts: Indonesia 16 000 t, Malaysia 125 000 t, Singapore 65 000 t (of which 34 000 t was re-exported), Brunei 2000 t and Papua New Guinea 2000 t. Shallot bulbs are traded fresh, fried or pickled, whereas garlic is sold fresh or dry and in the form of pills, drinks and powders based on extract.

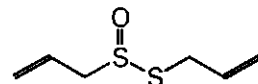
**Properties** *Allium* species release characteristic odours when the tissue is damaged (the 'onion odour' or 'garlic odour'). They are known for their sulphur-containing compounds, such as S-alkyl-L-cysteine sulfoxides (alkyl is e.g. methyl, propyl, vinyl, allyl) and  $\gamma$ -glutamyl-S-alkyl-cysteines.

The main constituent of fresh undamaged garlic is alliin (S-allyl-L-(+)-cysteine sulfoxide, > 0.3%), which is degraded by the enzyme alliinase (C-S-lyase) to pyruvic acid and 2-propenesulphenic acid upon cutting or bruising the tissue (alliin and alliinase are localized in separated compartments in the undamaged plant material). 2-Propenesulphenic acid is immediately transformed into allicin (diallyldisulphide-mono-S-oxide). Air oxidation of allicin leads to diallyldisulphide (1,7-dithioocta-4,5-diene), which is the chief constituent of garlic volatile oil, and together with related tri- and oligosulphides is responsible for the characteristic garlic smell. Allicin condensation products such as ajoenes and vinylthiines are also present in alcoholic garlic extracts. The drug also contains carbohydrates (fructans) and steroidal saponins.

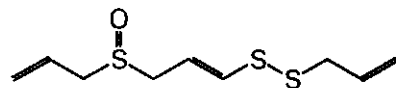
Fresh onion bulbs also contain fructans, and also flavonoids and sulphur compounds (trans-(+)-S-(1-



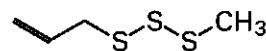
alliin



allicin



(E)-ajoene



methyl-allyl-trisulphide

propenyl)-L-cysteine sulfoxide (about 0.2%) and other cysteine derivatives). Upon bruising the bulb, the sulfoxides are degraded by alliinase to release pyruvic acid and alkyl-thiosulphinates, which rapidly turn into disulphides, the predominant compounds in *A. cepa* extracts. The volatile alliinase split product (Z)-thiopropenal-S-oxide (from trans-(+)-S-(1-propenyl)-L-cysteine sulfoxide) is the well-known lacrimatory factor in onions.

The growing conditions affect the proportion of different thiosulphinates; for instance, garlic grown in cooler climates shows a higher allyl to methyl ratio than garlic grown in warmer climates.

Several effects of *A. cepa*, *A. sativum* and other *Allium* species have been well investigated. In in vitro experiments, garlic, onion and Chinese chives showed antibacterial and antifungal activity against both gram-positive and gram-negative bacteria (including enteropathogens), pathogenic yeasts (*Candida* spp.) and some skin-pathogenic fungi. In an in vivo study, rabbits and guinea-pigs with experimentally induced dermatophyte infections (*Microsporum canis*, *Trichophyton rubrum*) were treated locally with a garlic extract. After 7 days of treatment, it took another 7–10 days for the skin lesions to completely recover.

Tests with rabbits and rats demonstrated that garlic extract lowers blood cholesterol and triglyceride and also has antihypertensive and anti-hyperglycaemic effects. Onion juice has been shown



to have anti-hyperglycaemic activity and anti-asthmatic activity in guinea-pigs. Furthermore, the saponin fraction prepared from the methanolic extract of rakkyo bulbs, is reported to exhibit inhibitory activities on cyclic AMP phosphodiesterase and Na<sup>+</sup>/K<sup>+</sup>-ATPase. The action was almost as potent as that of papaverine; this makes the saponins interesting for their cardiotonic effect.

The best investigated activity, however, is the effect on thrombocyte aggregation. Garlic and onion extracts have been found to show *in vitro* activity against platelet aggregation. Excessive platelet aggregation is recognized to be a dangerous contributory factor to thrombosis and arteriosclerosis, possibly leading to myocardial and cerebral infarctions. Several compounds have been reported as active principles against thrombocyte aggregation; for instance, the activity has been attributed to the ajoenes (inhibiting lipoxygenases), but also to methyl-allyl-trisulphide (the inhibitory effect depends on the content of this compound in the oil). Dimethyl- and diphenylthiosulphinate (from *A. cepa*) inhibited thromboxane synthesis, whereas the acid amides N-p-coumaroyltyramine and N-trans-feruloyltyramine, lunularic acid and p-coumaric acid, all sulphurless compounds isolated from the ethylacetate-soluble fraction of *A. chinense* bulbs, were shown to inhibit prostaglandin and thromboxane synthetases. Compared with aspirin, the compounds from *A. chinense* were more potent. Adenoside was isolated from the n-butanol soluble fraction of *A. sativum* and *A. chinense* bulbs; it showed very significant inhibitory activity against the aggregation of human platelets *in vitro*. Finally, chinenosides (furostanol saponins from *A. chinense* bulbs) inhibited ADP-induced aggregation of human blood platelets, the effect being comparable to that of aspirin.

Clinical tests have been conducted (inhibition of thrombocyte aggregation, lipid-lowering activity), but the results are usually conflicting or inconsistent, possibly because of the non-standardized preparations or questionable protocols used. In a double-blind, placebo-controlled experiment on the inhibition of blood-platelet aggregation in a group of patients in which this parameter was constantly and/or spontaneously increased the results were significant. After taking a standardized garlic preparation (800 mg daily, containing 1.3% allicin) for a period of 4 weeks, the spontaneous thrombocyte aggregation disappeared, and several other parameters (e.g. microcirculation of the skin and plasma viscosity) improved.

Furthermore, another placebo-controlled, double-blind experiment seemed to demonstrate the effectiveness of a standardized garlic powder (containing 1.3% alliin, 800 mg daily, administered for 4 months) as a blood-cholesterol lowering agent. Other trials seem to indicate a fibrinolytic activity for both garlic and onion.

An inverse correlation has been reported between regular consumption of garlic and onion and the risk of stomach cancer. S-allylcysteine has been found a chemopreventive agent for hepatocarcinogenesis in rats. From tests with mice it was suggested that garlic may provide an effective form of therapy for transitional cell carcinoma of the bladder. Other studies gave evidence of a direct effect of S-allylmercaptocysteine, one of the stable components present in aged garlic extract, on established cancer cells of breast and prostate.

In a test with alloxan diabetic rats, treatment with S-allyl-cysteine sulphoxide isolated from garlic ameliorated the diabetic condition almost to the same extent as did insulin. A garlic extract cured experimentally infected mice of trypanosomiasis in 4 days, with diallyldisulphide as the probable active compound. Garlic may improve arterial oxygenation and symptoms in patients with hepatopulmonary syndrome; this warrants further investigation. Extracts of *A. tuberosum* have been reported to have anti-tumour activity *in vitro* and *in vivo*.

Extracts of welsh onion showed allelopathic effects; in tests, they inhibited the growth of *Compositae* crops such as lettuce, marigold, *Aster* and *Chrysanthemum*. However, a 1% extract was highly effective in stimulating mycelial growth of the edible mushroom *Lentinus edodes*.

**Description** Perennial or biennial herbs, often grown as annual from bulbs or seed, up to 100(-150) cm tall; roots adventitious, up to 30 cm long; bulb usually present and distinct, with papery or chartaceous protective coats (tunics); real stem very short, forming a disk at the base of the bulb; pseudostem on top of bulb formed by the sheathing bases of successive leaves. Leaves basally concentrated but sometimes sheathing the scape for a considerable distance and then appearing cauline, alternate, often distichous, blades flat, terete, fistulose or angular, usually filiform-linear with acute apex, glabrous. Inflorescence umbellate, spherical or subspherical, on top of a terete or fistulose scape usually exceeding the leaves, initially surrounded by 2 hyaline bracts which are normally almost fused, composed of flowers, bulbils or both. Flowers bisexual, often

with slender pedicel, actinomorphic, campanulate to urceolate; tepals 6, in 2 whorls, free or almost free; stamens 6, filaments inserted at the base of the tepals, anthers dehiscing with longitudinal slits; ovary superior, 3-locular, style simple, often subgynobasic. Fruit a globular capsule, splitting loculicidally, several-seeded. Seeds broad and triangular, often blackish and wrinkled after drying; embryo more or less curved.

**Growth and development** Bulbs of *Allium* species are formed from the lower parts of the leaf-sheaths, as a result of photosynthate mobilization from the leaf-blade to the base of the leaves. A bulb is only formed when the plant has reached a certain stage of growth and when the daylength is long enough and temperature sufficiently high. When the bulb has reached full maturity, the leaf-blades start to wither. The bulbils that are often present in the inflorescence can be useful for vegetative propagation, especially in taxa with poor seed-setting.

Pollination is by insects such as bees, bumble-bees and hover flies. Onion is a facultative cross-pollinator, the percentage of selfing amounting to 10–20%. Flowers are often protandrous. In Chinese chives, more than 90% of the seed develops apomictically. Flower induction is controlled by temperature and daylength, and for several species (particularly rakkyo, welsh onion, Chinese chives) flowering is rare in the tropics.

**Other botanical information** There are numerous cultivars, particularly of garlic and onion. Only short-day cultivars are of interest to the tropics. There is no appropriate cultivar classification for South-East Asia. The presumed wild ancestors and some related species of cultivated *Allium* have been determined, e.g. *A. longicuspis* Regel which is the presumed wild ancestor of garlic, and *A. altaicum* Pallas which is a closely related species of welsh onion.

**Ecology** Tropical shallot requires an average day temperature of 20–26°C and a daylength of at least 11 hours, whereas common onion prefers slightly lower temperatures and a daylength of at least 13 hours. In Indonesia, shallot is mainly grown in the lowlands below 450 m altitude, preferably on well-drained alluvial clay soil, whereas Chinese chives, welsh onion and garlic are grown in the highlands up to 2200 m altitude. Most tropical onions are grown during the dry season, as too much rain will result in a high incidence of fungal diseases. They require well-drained soils.

**Propagation and planting** Several *Allium*

species (shallot, rakkyo) are commonly propagated by bulbs. To avoid problems of dormancy, the bulbs should first be stored for some months. Garlic is normally propagated by lateral bulbs (cloves) and seldom by bulbils from the inflorescence. In South-East Asia, welsh onion is propagated mainly from basal tillers, and Chinese chives by division of clumps. Propagation by seed may enhance the size and shape of bulbs and minimize diseases. The seed for propagating common onion is produced in the subtropics, where the climatic conditions are more favourable. Some cultivars are direct-seeded, whereas others are transplanted from nurseries where the seed is usually sown under a mulch. Planting distance varies according to species and conditions, but is usually 10–15 cm × 15–20 cm. Intercropping with other vegetables, e.g. hot pepper, carrot, Irish potato and mustard, is common.

**Husbandry** Weeds are often a serious problem and weeding should be done regularly; it is mostly done by hand, although chemical weed control is increasing. The crop must be irrigated regularly during dry weather. Organic and/or chemical fertilizers are generally applied. Crop rotation is important to avoid the build-up of diseases and pests such as *Fusarium*, *Sclerotium* and nematodes.

**Diseases and pests** Fungal diseases are common, particularly during the rainy season. *Alternaria*, *Fusarium*, *Stemphylium*, *Aspergillus* and *Colletotrichum* species may cause severe losses in onion. Viruses may also cause problems, which may be overcome by visually inspecting the planting material in the field and destroying any affected plants, or by the use of true-seed cultivars. Nematodes can be very harmful, especially in upland soils at higher altitudes, and without adequate crop rotation. Army worms (*Spodoptera exigua*) and thrips (*Thrips tabaci*) are reported as serious pests in shallot, common onion, welsh onion and garlic.

**Harvesting** Bulbs are often harvested after the leaves have wilted, usually 2–3 months after planting, but after 3–5 months in common onion and garlic. The bulbs are usually pulled out by hand, tied into bunches, and dried in the sun, usually with the bulbs covered by the leaves to protect them. Often, some bulbs are kept as planting material for the next growing season. Chinese chives is a ratoon crop; leaves are harvested repeatedly from the same plants the year round.

**Yield** Yields of common onion in South-East Asia range from 7–20 t/ha; average yields of shallot (6 t/ha), rakkyo, welsh onion (7 t/ha) and garlic

(2.5–4.5 t/ha) are generally lower.

**Handling after harvest** After drying, the bulbs (or plants of welsh onion) are tied into bunches which are sold directly or stored, often by hanging them on racks in well-ventilated places. For long-distance transport, the dry leaves are cut off and the bulbs are packed in bags or crates. The bulk of rakkyo is steeped in brine and subsequently processed into sweet or sour pickles. Chinese chives leaves are marketed as fresh as possible.

**Genetic resources and breeding** Germplasm collections of common onion, garlic and welsh onion are maintained at several institutes in Europe, the United States and Japan. Smaller collections of shallot and garlic are available at research institutes in South-East Asia, e.g. Lembang Horticultural Research Institute (LEHRI), Indonesia. Evaluation of the South-East Asian germplasm collections led to the recommendation of some local cultivars, e.g. of shallot. Breeding objectives are resistance to diseases and improvement of bulb quality and yield. It would be a breakthrough to find seed-producing selections, e.g. in rakkyo and garlic.

**Prospects** *Allium* species have an outstanding reputation as phytotherapeutic. The ability to inhibit thrombosis seems to be of remarkable medicinal value. The medicinal reputation might further stimulate the interest in *Allium* as vegetable as well. Some medicinal properties have been confirmed by in vitro tests, animal tests and/or clinical tests (e.g. antimicrobial activity, activity against platelet aggregation, blood cholesterol lowering activity), but other attributed properties (e.g. diuretic activity, anti-cancer activity) have not been demonstrated conclusively. Well executed clinical tests with standardized preparations are needed to confirm these properties. Garlic has potential for the preservation of processed foods because of the inhibitory activity against pathogenic bacteria such as *Salmonella typhi* and *Escherichia coli*.

**Literature** [1] Bruneton, J., 1995. Pharmacognosy, phytochemistry, medicinal plants. Technique & Documentation Lavoisier, Paris, France. pp. 180–183. [2] Buijsen, J.R.M., 1993. Alliaceae. In: Kalkman, C. et al. (Editors): Flora Malesiana. Series 1, Vol. 11(2). Foundation Flora Malesiana. Rijksherbarium/Hortus Botanicus, Leiden University, the Netherlands. pp. 375–384. [3] de Padua, L.S. & Pancho, J.V., 1983. Handbook on Philippine medicinal plants. Vol. 4. Technical Bulletin Vol. VI No 1. Documentation and Information Section, Office of the Director of Research, University

of the Philippines at Los Baños, the Philippines. pp. 6–8. [4] Goda, Y., Shibuya, M. & Sankawa, U., 1987. Inhibitors of the arachidonate cascade from *Allium chinense* and their effect on in vitro platelet aggregation. Chemical and Pharmaceutical Bulletin 35(7): 2668–2674. [5] Kosuge, T., Yokota, M., Sugiyama, K., Yamamoto, T., Ni, M.Y. & Yan, S.C., 1985. Studies on antitumour activities and antitumour principles of Chinese herbs (in Japanese). Yakugaku Zasshi 105(8): 791–795. [6] Kuroda, M., Mimaki, Y., Kameyama, A., Sashida, Y. & Nikaido, T., 1995. Steroidal saponins from *Allium chinense* and their inhibitory activities on cyclic AMP phosphodiesterase and Na<sup>+</sup>/K<sup>+</sup> ATPase. Phytochemistry 40(4): 1071–1076. [7] Okuyama, T., Fujita, K., Shibata, S., Hoson, M., Kawada, T., Masaki, M. & Yamate, N., 1989. Effects of Chinese drugs 'xiebai' and 'dasuan' on human platelet aggregation (*Allium bakeri*, *A. sativum*). Planta Medica 55(3): 242–244. [8] Okuyama, T., Shibata, S., Hoson, M., Kawada, T., Osada, H. & Noguchi, T., 1986. Effect of oriental plant drugs on platelet aggregation. III. Effect of Chinese drug 'xiebai' on human platelet aggregation. Planta Medica 52(3): 171–175. [9] Peng, J.-P., Yao, X.-S., Tezuka, Y. & Kikuchi, T., 1996. Furostanol glycosides from bulbs of *Allium chinense*. Phytochemistry 41(1): 283–285. [10] Siemonsma, J.S. & Kasem Piluek (Editors), 1993. Plant Resources of South-East Asia No 8. Vegetables. Pudoc Scientific Publishers, Wageningen, the Netherlands. pp. 62–82.

#### *Selection of species*

#### **Allium cepa L.**

Sp. pl. 1: 300 (1753).

**Vernacular names** Onion, common onion, shallot onion (En). Echalote, oignon (Fr). Brunei: bawang besar. Indonesia: bawang merah, bawang beureum, bawang bombay. Malaysia: bawang merah, bawang kecil, bawang besar. Papua New Guinea: lip anian, anian (Pidgin). Philippines: sibuyas tagalog (Tagalog), bauang pula (Tagalog), lasona (Iloko). Cambodia: khtüm krähââm, khtüm barang. Laos: hoom bwax, bwax fälangx. Thailand: hom farang, hom hua yai (central). Vietnam: h[af]nh t[aa]y, h[af]nh c[ur], h[af]nh t[aw]m.

**Distribution** Probably originating from central Asia, but nowhere truly wild. Cultivated all over the world; cv. group *Aggregatum* (shallot) predominates in the tropical lowland of South-East Asia, but cv. group *Common Onion* is grown in the

Philippines, Papua New Guinea and Thailand.

**Uses** Onion plays an important role in traditional medicine; it is used as diuretic and suppresses the blood sugar level and platelet aggregation. Shallot is used traditionally as febrifuge and as a poultice to cure wounds. In the Philippines the bulbs are considered anthelmintic and stomachic, and used to treat diarrhoea, headache, earache and amenorrhoea. Shallot stimulates the appetite.

**Observations** A biennial herb, usually grown as an annual, up to 100 cm tall; mature bulb up to 15 cm in diameter; leaves 3-8, blades semiterete, at first solid, later becoming hollow, glaucous; scape 1-several, terete, often inflated in the middle or in the lower part, hollow, inflorescence with up to 2000 flowers; flowers with greenish-white to purplish tepals, stamens sometimes slightly exceeding tepals, style shorter than stamens at anthesis; fruit 4-6 mm in diameter, containing up to 6 seeds; seeds about 3 mm × 2 mm. In cv. group Common Onion the bulbs are large and normally single, and plants reproduce from seed or from seed-grown bulbils; in cv. group *Aggregatum*

(shallot) the bulbs are smaller, several to many forming an aggregated cluster, and plants reproduce vegetatively via lateral bulbils.

**Selected sources** 76, 193, 205, 332, 549, 977, 1178, 1356, 1446.

### *Allium chinense* G. Don

Mem. Wernerian Nat. Hist. Soc. Edinb. 6: 83 (1827).

**Synonyms** *Allium bakeri* Regel (1875), *Allium schoenoprasum* auct. non L.

**Vernacular names** Rakkyo (En). Indonesia: bawang ganda. Malaysia: lokyo. Thailand: krathiam-chin (Bangkok), hom-prang (central), hom-paenyuak (northern). Vietnam: ki[ee]ju.

**Distribution** Native to central and eastern China; widely grown in China and Japan and to a limited extent in South-East Asia.

**Uses** Medicinally, the bulbs are of interest in the prevention of thrombosis. They are used for the treatment of heart failures in Chinese medicine. Rakkyo is also used against fever, stomachache and eye infections.

**Observations** A biennial herb, up to 60 cm tall; bulb ellipsoid, up to 4 cm × 1.5 cm; leaves 3-4(-5), D-shaped or nearly triangular in transverse section, 3-5-ridged; scape terete, solid, inflorescence with up to 30 flowers, without bulbils; flowers with purplish tepals, stamens and style much longer than tepals.

**Selected sources** 76, 205, 490, 549, 799, 1081, 1082, 1123, 1356.

### *Allium fistulosum* L.

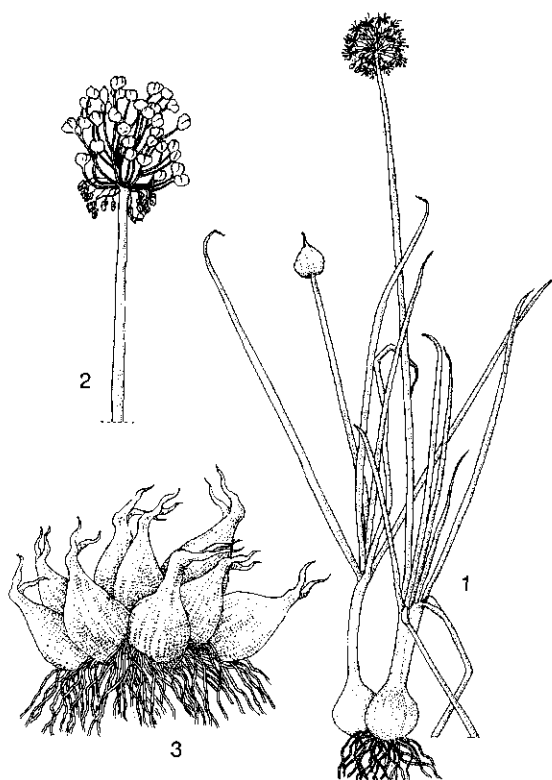
Sp. pl. 1: 301 (1753).

**Synonyms** *Allium bouddhae* O. Debeaux (1877), *Allium bakeri* Hoop. (1929) non Regel.

**Vernacular names** Welsh onion, (Japanese) bunching onion, spring onion (En). Ciboule (Fr). Indonesia: bawang daun, bawang bakung (Sundanese), bawang ongang (Javanese). Malaysia: daun bawang. Philippines: buyah (Ifugao). Cambodia: khtüm sänlök. Thailand: hom-ton (central), hom-cheen (peninsular). Vietnam: h[af]nh hoa, h[af]nh h[uw][ow]ng.

**Distribution** Probably originating from north-western China, but nowhere truly wild; cultivated all over the world.

**Uses** The therapeutic qualities attributed to the Welsh onion are many, especially in Chinese medicine. It is used to improve the functioning of internal organs and the metabolism, and to prolong life. It is further reported to improve eyesight, to aid digestion and perspiration, and to enhance re-



*Allium cepa* L. cv. group *Aggregatum* - 1, flowering plants; 2, inflorescence; 3, mature bulbs.

covery from common colds, headaches, wounds and festering sores. The plants are said to reduce or prevent white ant infestation in gardens, and in China the diluted juice is used against aphids.

**Observations** A perennial herb, usually grown as an annual or biennial, up to 150 cm tall; bulb indistinct, gradually passing into the scape; leaves 4-12 (actively growing green leaves 3-6), blades terete, hollow, bluish-green; scape 1, terete, hollow, inflorescence composed solely of flowers or of bulbils; flowers with greenish-white to white tepals, stamens and style much longer than tepals; fruit about 5 mm in diameter; seeds 3-4 mm × 2-2.5 mm.

**Selected sources** 76, 205, 549, 1356.

**Allium sativum L.**

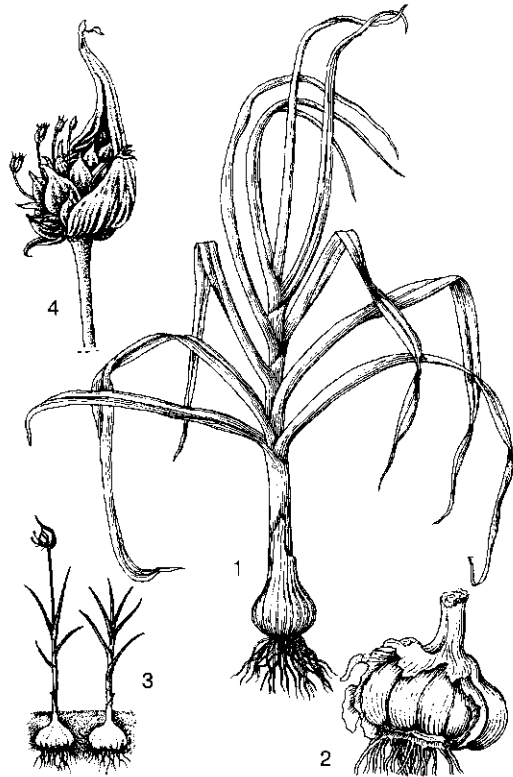
Sp. pl. 1: 297 (1753).

**Vernacular names** Garlic (En). Ail (Fr). Indonesia: bawang putih, bawang bodas (Sundanese). Malaysia: bawang putih. Papua New Guinea: galik (Pidgin). Philippines: bawang (Tagalog, Ilocano), ajos (Bisaya), ahus (Ibanag). Cambodia: khtüm sââ. Laos: kath'iem. Thailand: krathiam (general), hom-tiam (northern). Vietnam: t[or]i.

**Distribution** Probably originating from central Asia (Tien Shan), but nowhere truly wild; cultivated all over the world at latitudes between 5-50° in both hemispheres.

**Uses** Garlic is much valued as a medicinal crop. It has a strong reputation for lowering the blood sugar and cholesterol levels and inhibiting thrombus formation. It is used externally to cure headache, insect bites, rheumatism and toothache, and a decoction internally as febrifuge. Leaves and bulbs are considered to have hypotensive, carminative, antiseptic, anthelmintic, diaphoretic and expectorant properties. Many attributed prophylactic activities are questionable, but have resulted in a rich supply of and demand for medicinal pills, drinks and powders based on garlic extracts.

**Observations** An erect herb, usually grown as an annual, up to 150 cm tall; bulb composed of (1-4)-15 lateral bulbs (cloves), up to 7 cm in diameter; leaves 4-10, blades flat or V-shaped in transverse section; scape 1, solid, inflorescence composed solely of bulbils or of bulbils and flowers; flowers often rudimentary or absent, with greenish-pink to purplish tepals, stamens and style shorter than tepals; fruit abortive, seedless. Two groups of cultivars are often distinguished: cv. group Common Garlic with a straight scape, and



*Allium sativum L.* - 1, plant habit; 2, bulb; 3, plantlets; 4, inflorescence.

cv. group *Ophioscorodon* (rocambolè or serpent garlic) with a scape having a distinct curve or coil towards the top.

**Selected sources** 5, 76, 82, 193, 205, 332, 549, 729, 794, 825, 883, 1046, 1081, 1178, 1231, 1356, 1357, 1440, 1446.

**Allium tuberosum Rottler ex Sprengel**  
Syst. veg. 2: 38 (1825).

**Synonyms** *Allium uliginosum* G. Don (1827), *Allium senescens* Miq. (1867), *Allium odorum* auct. non L.

**Vernacular names** Chinese chives (En). Indonesia: kucai. Malaysia: kuchai. Philippines: kutsay (Tagalog), ganda (Bisaya), amput di imayyaw (Ifugao). Cambodia: kachaay. Thailand: kuichai (Bangkok), hom-paen (northern). Vietnam: h[ej], n[es]n t[af]u.

**Distribution** Probably native in eastern Asia; cultivated in the United States, Nepal, India, China, Taiwan, Korea, Japan, Indo-China, Thailand, Indonesia and the Philippines.

**Uses** Chinese chives is used medicinally against

tumours and intestinal disorders, as a stomachic and as bactericide in pulmonary infections, and is considered to promote recovery from fatigue. Leaves and bulbs are used in the Philippines as antiseptic and vulnerary. The seed is used in Thailand and Indo-China against toothache, as an antiseptic mouthwash.

**Observations** A perennial herb forming dense clumps, up to 50 cm tall; bulb indistinct, narrowly ovoid, about 2 cm × 1.5 cm; leaves 4–9, blades flat above, slightly keeled below; scape 1, compressed, solid, inflorescence many-flowered, without bulbils; flowers with white tepals, stamens and style about as long as tepals; fruit 5–6 mm in diameter; seeds 3–4 mm long.

**Selected sources** 76, 205, 332, 549, 778, 1035, 1178, 1356.

Diah Sulistiarini, Juliasri Djamal  
& Iman Raharjo

## Aloe L.

Sp. pl. 1: 319 (1753); Gen. pl. ed. 5: 150 (1754).

ASPHODELACEAE

$x = 7$ ; *A. ferox*, *A. vera*:  $2n = 14$

**Major species** *Aloe vera* (L.) Burm.f.

**Vernacular names** Aloe (En). Indonesia: lidah buaya. Vietnam: [oo] h[oo]ji.

**Origin and geographic distribution** The approximately 330 species of *Aloe* originate from Africa, Madagascar and Arabia. The centres of diversity are South Africa (Transvaal) and the region of Eritrea, Ethiopia and northern Somalia. Over 100 species are cultivated and there is an overwhelming number of hybrids and cultivars. All Malesian *Aloe* have been introduced and are popular garden and pot plants. *A. vera* was formerly produced on Barbados, where it had been introduced early in the 16th Century. Nowadays, it is cultivated commercially in the United States, Mexico, the Caribbean, Israel, Australia, Thailand and South Kalimantan. Commercial plantations of *A. ferox* have been established in Albertinia in South Africa.

**Uses** *Aloe* juice, contained in the pericyclic cells of the vascular bundles in the leaf, is used to make the laxative drug known as 'aloe', 'aloes' or 'bitter aloes'. *Aloe* gel, the mucilage from polyhedral cells in the central part of the leaf, is claimed to have healing properties as well.

The three main types of commercial aloe drug are 'Curaçao aloe', from *A. vera*, 'Cape aloe', mainly from *A. ferox* and 'Socotrine aloe', from *A. perryi*

Baker. *Aloe* was already known to the Greeks as early as the 4th Century B.C. The drug was used by Alexander the Great, Dioscorides, Celsus and Pliny and by later Greek and Arabian physicians. It is used as a laxative, purgative and vermifuge. An ordinary dose takes 15–18 hours to produce an effect. It acts mainly on the large intestine. The effect of aloe can be strong, and nowadays it is used in combination with weaker purgatives or antispasmodic drugs. 'Cape aloe' has more powerful purgative properties than the other two; 'Socotrine aloe' is the mildest. In Thai traditional medicine, aloe is always included in the drug recipe because traditional doctors believe that patients will recover faster if unwanted material is expelled from the body. Both 'Curaçao aloe' and 'Cape aloe' are used for veterinary purposes in Europe.

Externally, aloe gel has proven effective in the treatment of skin burns by X-ray radiation. Furthermore, *A. vera* gel has gained popularity as a folk remedy worldwide, and numerous claims have been made for its medicinal properties. However, experimental results only support its applications for wound healing, treatment of burns, anti-inflammatory and diabetic activities. *A. vera* gel has been approved by the US Federal Drug Administration only for first aid treatment of burns and cuts. *Aloe* gel is widely used as a hydrating and skin-protecting agent in creams and liquids such as sun lotion, shaving cream, lip balm and healing ointments. It is also gaining importance as a refreshing and nutritive ingredient in food and drinks in Indonesia and Thailand.

Many other *Aloe* species are popular pot plants or garden ornamentals in the Malesian region (e.g. *A. arborescens* Miller, *A. saponaria* (Aiton) Haw.). Apart from their ornamental value they are often used in folk medicine as hair lotion and to promote wound healing. The sticky leaf sap of *A. camperii* Schweinf. is reported to be used in Papua New Guinea to treat severe burns. In Vietnam, an *Aloe* species is applied as a laxative and emmenagogue, whereas the leaves are eaten cooked with sugar or made into a potage.

**Production and international trade** In 1992 the local *Aloe* industry based on *A. ferox* in South Africa was worth about US\$ 0.5 million. Though considerable quantities are marketed and used locally, most of the 'Cape aloe' produced in South Africa is exported to Europe, especially to Italy, France and Germany.

The international market for *Aloe* gel is not very open. Most gel is bought by the cosmetic industry

which demands high quality. An export permit is compulsory because all *Aloe* species are listed in the Convention on International Trade in Endangered Species of Wild Fauna and Flora (CITES) Appendix II.

**Properties** The aloe drug contains 15–40% aloe-emodin-anthrone 10-C-glucosides (hydroxy-anthraquinone derivatives) such as aloin, hydroxyaloin and (in *A. ferox*) aloinoside. Aloin, sometimes referred to as barbaloin, is a mixture of aloin A and aloin B, which interconvert through the anthranol form. Furthermore, the juice contains a pyrone derivative (aloenin) and resins: free and 8-C-glucosylated-2-acetyl-7-hydroxy-5-methylchromones (e.g. aloesone, furoaloesone, aloeresin A, aloeresin B (aloesin) and aloeresin C). *A. ferox* also contains free or glycosylated feroxidin (a tetralin) and feradolide (a dihydroisocoumarin). 'Cape aloe' drugs must contain at least 18% hydroxy-anthraquinone derivatives, 'Curaçao aloe' at least 28%. 'Curaçao aloe' is almost entirely soluble in 60% alcohol and does not contain more than 30% of substances insoluble in water. It should not contain more than 12% moisture and 3% ash. 'Cape aloe' should not contain more than 12% moisture and 2% ash, and should have a water-soluble fraction of at least 45%.

The compound responsible for the laxative properties is aloin. Experiments with rats have shown that aloin itself is inactive as a laxative, but that it is activated to aloe-emodin anthrone, a purgative component, by *Eubacterium* sp. In the diarrhoea induced by aloin, the increase in water content might be a more important factor than the stimulation of peristalsis. Daily and prolonged use of laxatives of this type may lead to serious problems, such as dependence and 'cathartic colon'. Anthraquinone laxatives should not be used longer than 8–10 days, or by children younger than age 12. Contra-indications for aloe drugs include pregnancy, breast-feeding, intestinal inflammations and haemorrhoids. Preferably, aloin should be administered together with an antispasmodic, to moderate its griping action. Possible side-effects of aloin include congestion and irritation of the pelvic organs. Anthranoid-containing laxatives such as aloe may play a role in colorectal cancer. There are some published data on the genotoxic potential of anthranoids, and there is evidence of a tumorigenic potential in rodents.

*A. vera* gel has earned a reputation as a miracle drug. It is very rich in water, but does not appear to contain very specific compounds. The major

constituents are polysaccharides (pectins, hemicelluloses), plus amino acids, lipids, sterols and enzymes. It has been proven to be effective for burn treatment, because of its anti-inflammatory and wound-healing properties. The active constituents are aloctin A, aloctin B, bradykininase and magnesium lactate. Its healing properties may also be due to hydrating, insulating and protective activities resulting from the high water content. Furthermore, it has been claimed that acemannan, the major carbohydrate fraction of *A. vera* gel, has various therapeutic properties, including acceleration of wound healing, immune stimulation, anti-cancer and antiviral effects, and that acemannan may partly function through macrophage activation. In experiments with rats both topical and oral treatments with *A. vera* had a positive influence on the synthesis of glycosamino glycans, thereby beneficially modulating wound healing. The healing of dermal wounds in diabetic rats is also positively influenced by oral and topical application of *A. vera*. The antidiabetic effects of aloe gel probably have an influence on phases such as inflammation, fibroplasia, collagen-synthesis and maturation and wound contraction. These antidiabetic effects have been confirmed in animal experiments. Subsequently, clinical trials in Thailand gave satisfactory results. Active constituents identified in this research were polysaccharides and glycoprotein. Besides the activities mentioned above, aloe gel also exhibited antiviral effects. A placebo-controlled, double-blind study showed that a topically applied *A. vera* extract of 0.5% in a hydrophylic cream is beneficial for patients suffering from psoriasis. It did not show toxic or any other objective side-effect. A clinical test with patients with advanced solid tumours, for whom no standard effective therapy is available, suggested that *A. vera* extract in combination with the immunomodulating neurohormone melatonin may produce some therapeutic benefits, at least in terms of stabilization of disease and survival.

**Adulterations and substitutes** Psyllium (*Plantago* spp.), which is a natural bulk laxative, is one of the substitutes for anthraquinone-containing laxative drugs such as aloe, that may cause dependency and/or cathartic colon. However, in Thailand, anthraquinone-containing preparations from *Senna alata* (L.) Roxb. and other *Senna* and *Cassia* species are also sometimes recommended as substitutes for aloe. *Centella asiatica* (L.) Urb. is recommended as a substitute for aloe gels in wound treatments. Its triterpenes

have exhibited both wound healing and antibacterial activities.

**Description** More or less succulent shrubby perennials, often with very short stem and fleshy fibrous roots. Leaves arranged spirally in a rosette, sometimes distichous, linear to lanceolate or triangular, very thick and fleshy, sheathing at the base, margins usually sinuate-dentate, often spiny apically, sometimes entire, surface sometimes spiny, containing colourless, yellow, brown or purple sap. Inflorescence a pseudo-lateral, simple or branched, long-cylindrical raceme. Flowers bisexual, protandrous, pedicellate; tepals 6, usually connate into a tube, sometimes outer 3 free, fleshy, apices sub-acute to obtuse; stamens 6, in 2 rows of 3; ovary superior, 3-locular, style filiform, longer than stamens, stigma small. Fruit a loculicidal capsule, many-seeded. Seeds elongate and ovoid, grey or black, arillate.

**Growth and development** *Aloe* species follow the Crassulacean acid metabolism (CAM) pathway. CAM plants are able to fix CO<sub>2</sub> at night and to photosynthesize with closed stomata during the day, thus minimizing water loss under arid conditions. This, plus their succulent leaves, stems and the presence of a thick cuticle, makes them well adapted to dry conditions.

The flower morphology of *A. ferox* suggests that the flowers are pollinated by birds. However, in Africa honey bees also play a role in the pollination. *A. ferox* is self-incompatible and only a few flowers per raceme flower simultaneously, apparently to promote cross-pollination. The stamens are exerted in the morning, then wither and withdraw in the afternoon, whereas the style is exerted on the second day of anthesis. In Africa, *A. vera* flowers and fruits normally. In India and other areas outside its natural range, however, fruit formation is rare, and any fruits that do form have seeds which do not germinate. This failure to set fruit is presumed to be caused by pollen sterility and self-incompatibility.

**Other botanical information** *Aloe* was formerly (and sometimes still is) included in the family *Liliaceae* s.l. Nowadays, it is usually placed in *Asphodelaceae* and sometimes in a separate family *Aloaceae*. *Aloe* and related genera like *Haworthia*, *Gasteria* and *Astroloba* can be distinguished by their succulent leaves, vascular bundles in a ring around ground parenchyma, a cap of aloin cells at the phloem pole, chemical properties of the often coloured and/or pungent sap, and the homogeneity of the chromosome composition. The numbers of species described in *Aloe* differs consider-

ably and synonymy and names for subspecific rank are overwhelming. The names *A. vera* and *A. barbadensis* are still contentious among specialists.

**Ecology** *Aloe* grows in a wide range of climatic conditions. Though the root system is shallow, it can be grown under dry conditions. Waterlogging should be avoided and *Aloe* thrives best on well-drained, rich soils.

*A. ferox* is one of the dominant species in the 'succulent bushland' vegetation in South Africa. It is especially abundant on arid rocky hillsides up to about 1000 m elevation, where mean temperatures range from 27–31°C and annual rainfall from 50–300 mm. Severe drought is said to stop juice production.

**Propagation and planting** *Aloe* can be propagated vegetatively, through suckers, offsets or cuttings, or by seed.

In *A. vera* cultivation, vegetative propagation is usually preferred above propagation by seed, because of the poor seedling emergence and the faster initial growth of suckers. Water deficiency may lead to decreased sucker formation. Suckers can be cut from the mother plant when they are 15–20 cm long. They may be grown in a nursery during the first year. After transplanting, the distances between and within rows in large-scale *Aloe* plantations are usually at least 0.5 m.

In vitro reproduction of *Aloe* is also possible, but has received little attention. With *A. ferox*, only plant regeneration from root and embryo tissue has succeeded. In the case of *A. vera*, micropropagation through in vitro culture of vegetative meristems, as well as in vitro regeneration of leaf base explants appears to be possible.

**In vitro production of active compounds** Aloin, aloesin and aloeresin have been found in *A. ferox* plantlets regenerated in vitro from root and embryo tissue, but could not be detected in the callus.

**Husbandry** *Aloe* species need to be left 2–3 years after transplanting before they can be used to obtain juice. However, for the production of gel in Thailand the first cut is made 6–12 months after transplanting. Weeding may be carried out mechanically or chemically; glyphosate is reported not to cause damage to *A. vera*. As the formation of too many suckers may retard growth and reduce yields of the mother plant, early removal of suckers is recommended, to obtain larger leaves. Water requirement for gel production is much higher than that for juice production, as gel yields are directly related to the amount of water avail-



able for the crop. Therefore, gel production may be increased through irrigation. The application of nitrogen may result in higher gel yields, because of faster leaf formation and higher yields per leaf. However, applying large amounts of water and nitrogen may reduce gel quality. Other cultural practices that may favour leaf production are mulching, shading and furrow cultivation.

**Diseases and pests** No serious diseases are known for *A. vera*. In India, *Alternaria alternata* and *Fusarium solani* are causes of leaf spot disease. In Aruba, leaf rot caused by *Erwinia chrysanthemi* occurs occasionally.

**Harvesting** *Aloe* juice is often collected by cutting off the leaves transversely close to the stem and positioning them in such a way that the juice drains into pots, tubs, vessels or even a simple canvas placed over a depression. The juice may also be obtained by squeezing the leaves or by warm or cold water retting. In South Africa, *A. ferox* is preferably tapped during the rainy season, because then the juice is more abundant, but tapping is also carried out in other periods of the year, except for the driest months. The leaves are usually cut in the morning and it takes 4–5 hours for the juice to drain from a pile of leaves. Only older leaves are cut; younger ones and growing tips are spared. In *A. ferox* in South Africa, the aloin content of the leaf juice was found to differ markedly between provenances. Large variations in aloin content have been found in *A. vera* too, with the highest concentration in exudates from younger mature leaves.

*Aloe* gel is obtained after eliminating the outer tissues of the leaf. In Aruba, gel is obtained by cutting open the leaves lengthwise and scraping the gel from the leaf blade. The youngest leaves (< 25 cm) are not suitable because of the small amount of gel, but the leaves should not be too old, because gel quantity and quality may decline. Individual *A. vera* leaves in Aruba reach their maximum fresh weight after about 40 weeks of growth. In a system where only selected leaves are cut, the possibilities for mechanical harvesting are limited.

**Yield** In Aruba, *A. vera* plants can produce 16–20 leaves per year under optimal moisture supply and sufficient nitrogen fertilization. With a plant density of 50 000 plants/ha and an average fresh leaf weight of 0.2 kg, this implies a gel yield of 160–200 t (560–840 kg dry matter) per ha.

**Handling after harvest** Collected *Aloe* juice is usually concentrated by boiling and then cooling. 'Curaçao aloe' may also be vacuum evaporated and then concentrated. On cooling, a solid, amor-

phous extract forms, which constitutes the drug. Its appearance varies with the concentration process used and the species. If the juice has been concentrated slowly, for instance in the sun or over a low fire, the cooled extract is opaque and waxy ('hepatic aloe') and aloin crystals are visible under the microscope. If, on the other hand, the juice has been concentrated rapidly, for instance over a fierce fire, the cooled extract is semi-transparent ('glassy aloe') and no aloin crystals are visible under the microscope. Aloe from *A. vera* is usually 'hepatic', aloe from *A. ferox* 'glassy'. Collected gel in Aruba is purified by centrifugal removal of cell wall material, and the liquid pure gel is stabilized by adding chemicals.

**Genetic resources and breeding** As a result of the continuous vegetative propagation, the genetic variability within *A. vera* in cultivation seems to be rather small. Nevertheless, high-yielding plants of *A. ferox* and *A. vera* may be selected and propagated for commercial cultivation. Many *Aloe* species hybridize in the wild if their area of distribution and period of flowering overlap, and it is easy to produce hybrids in cultivation. Because of their popularity as a greenhouse plant various *Aloe* species are potentially at risk of extinction. No germplasm collections and breeding programmes are known to exist.

**Prospects** *A. vera* and *A. ferox* might be potential crops in arid regions. *A. vera* is of particular interest, since its gel can be used in burn and wound care. It still remains beneficial as a household remedy: fresh gel can easily be prepared and applied to the wound. In its use as a laxative, however, the drug is tending to be replaced by other laxatives such as purified anthraquinones from *Cassia* and *Senna* species and bulk laxatives as from *Plantago* spp.

**Literature** |1| Bruneton, J., 1995. Pharmacognosy, phytochemistry, medicinal plants. Technique & Documentation Lavoisier, Paris, France. pp. 349–364. |2| Cavallini, A., Natali, L. & Castorena Sanchez, I., 1991. *Aloe barbadensis* Mill. (= *A. vera* L.). In: Bajaj, Y.P.S. (Editor): Biotechnology in agriculture and forestry. Vol. 15. Medicinal and aromatic plants III. Springer-Verlag, Berlin, Germany. pp. 95–106. |3| Grieve, M., 1994. A modern herbal. Tiger Books International, London, United Kingdom. pp. 26–29. |4| Grindlay, D. & Reynolds, T., 1986. The *Aloe vera* phenomenon: a review of the properties and modern uses of the leaf parenchyma gel. *Journal of Ethnopharmacology* 16(2–3): 117–151. |5| Hodge, W.H., 1953. The drug aloes of commerce, with special reference to

the Cape species. *Economic Botany* 7: 99–129. |6| Ishii, Y., Tanizawa, H. & Takino, Y., 1994. Studies of aloe. IV. Mechanism of cathartic effect (3). *Biological and Pharmaceutical Bulletin* 17(4): 495–497. |7| Racchi, M.L., 1988. Using in vitro culture to study the biosynthesis of secondary products in *Aloe ferox*. *Rivista di Agricoltura Subtropicale e Tropicale* 82(4): 707–714. |8| Reynolds, G.W., 1966. The aloes of tropical Africa and Madagascar. The Trustees the Aloe Book Fund, Swaziland, South Africa. 537 pp. |9| Reynolds, G.W., 1982. The aloes of South Africa. 4th Edition. A.A. Balkema, Rotterdam, the Netherlands. 538 pp. |10| van Schaik, A.H., 1994. Growing *Aloe vera* for gel production: report on four years of agricultural research on Aruba. Department of Agronomy, Wageningen Agricultural University, the Netherlands, & Department of Agriculture, Husbandry and Fisheries, Aruba, Dutch Caribbean. 58 pp.

#### *Selection of species*

#### ***Aloe ferox* Miller**

Gard. Dict. ed. 8: n. 22 (1768).

**Vernacular names** Cape aloe (En).

**Distribution** Indigenous over a vast area in South Africa. Plants introduced in the Malesian region probably originate from the Dutch East India Company's garden in the Cape. *A. ferox* is cultivated in Java.

**Uses** *A. ferox* is the source of 'Cape aloe'. Medicinal uses of fresh *A. ferox* in South-East Asia are probably similar to those reported for *A. vera*. In Africa, the inspissated leaf juice is used as a purgative in human and veterinary medicine and fresh juice is applied in cases of ophthalmia and syphilis. The flower nectar is said to be narcotic. The leaves have also been used in South Africa to make a jam tasting like watermelon jam.

**Observations** A perennial succulent shrub, up to 3(–5) m tall, usually with a single stem, densely covered with the persistent remains of the old leaves; leaves 50–60 in a dense capitate rosette, lanceolate, up to 100 cm × 15 cm, fleshy, upper surface flat, lower surface convex, smooth to spiny, margins sinuate-dentate, dull green, sometimes tinged red; inflorescence branched with 5–8 erect racemes, racemes very densely flowered, 50–80 cm × 9–12 cm; flowers with a dark red to orange perianth, filaments and style exerted. In its native area *A. ferox* is a rather variable species.

**Selected sources** 202, 350, 580, 596, 900, 1084, 1180, 1218, 1219, 1222, 1510, 1554, 1574, 1584.

#### ***Aloe vera* (L.) Burm.f.**

Fl. ind.: 83 (1768).

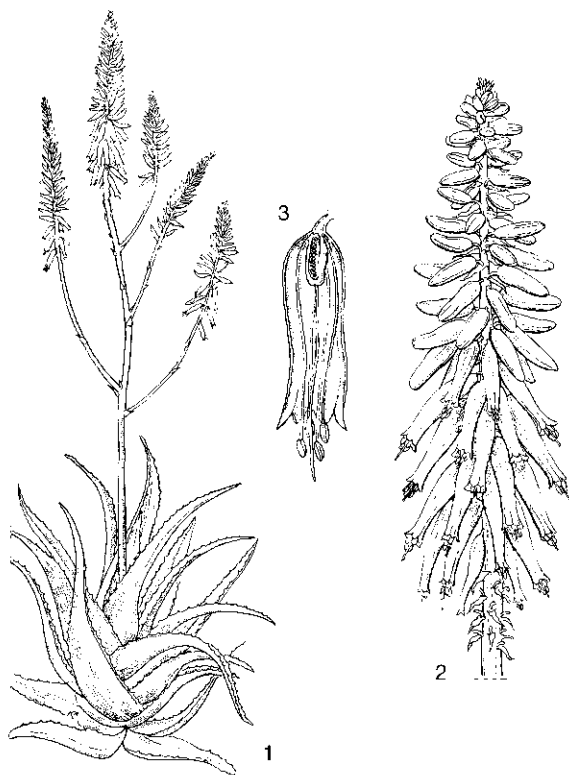
**Synonyms** *Aloe perfoliata* L. var. *vera* L. (1753), *Aloe barbadensis* Miller (1768).

**Vernacular names** Barbados aloe, Curaçao aloe (En). Indonesia: lidah buaya. Malaysia: lidah buaya (Peninsular), bunga raja raja (Sabah). Philippines: sabila (Tagalog), dilang-buwaya (Bikol), dilang-halo (Bisaya). Thailand: waan faimai (northern), waan hang chorakhe, haang takhe (central). Vietnam: [oo] h[oo]ji, [uw] h[oo]ji, nha d[ar]m.

**Distribution** Origin unknown; some authors presume the Macaronesian region, others prefer Arabia. At present *A. vera* is widely distributed throughout the tropics and subtropics. It was already common in India in the first Century AD. *A. vera* is grown as a pot plant and ornamental throughout Malaysia.

**Uses** The fresh yellow leaf juice is often used as a laxative or purgative and refrigerant. Externally, it is often used to treat burns, wounds, abrasions, skin diseases, irritations and alopecia. Fresh *A. vera* has a multitude of medicinal applications in South-East Asia. The leaf sap or juice is applied externally to treat pimples, blackheads or cuts. The sap mixed with other ingredients to mask its bitter taste is taken in Indonesia against asthma and to treat coughs. In the Philippines, similar mixtures are taken to cure dysentery and kidney problems or against dyspepsia. In Indo-China, the fresh leaf juice is considered purgative, anthelmintic, depurative and an emmenagogue. In Papua New Guinea, the juice is used internally to treat stomach ulcers. The leaf gel or peeled leaves are generally externally applied to treat skin afflictions and as a poultice on contusions or as a general refrigerant. The gel may also be applied externally on haemorrhoids. It is furthermore used as a hairwash to promote hair growth and as a general cosmetic to improve the complexion and to smoothen the skin. Sometimes the peeled leaves are eaten to relieve sore throat and coughs and as a mild laxative. *A. vera* is the source of the 'Curaçao aloe' drug, which is used as a purgative, vermifuge, emmenagogue and stomachic. The aloe drugs ('jadam') used in Malaysia and South-East Asia are mostly imported. 'Jadam' is used as an aperient, but it is also put on wounds and swellings, and daubed on the abdomen in the case of fever and after confinement. Furthermore a dye can be obtained from the dried leaf sap.

**Observations** A perennial shrub, with very short stem, taproot 5–10 cm long with many sec-



*Aloe vera* (L.) Burm. f. – 1, plant habit; 2, part of inflorescence; 3, flower in longitudinal section.

ondary roots in the upper soil, freely suckering and forming dense groups; leaves about 16, erect to slightly spreading, narrowly triangular, 40–50 cm × 6–7 cm, upper surface grey-green to pale green with few to many spots, lower surface generally lighter, margin with firm deltoid pale teeth of 2 mm; inflorescence simple or sparsely branched, 60–100 cm tall, racemes 30–40 cm × 5–6 cm, densely flowered; flowers with yellow, orange or red perianth, stiffly pendulous, anthers and stigma exserted. The long history of cultivation has led to various selections that are sometimes given formal ranking.

**Selected sources** 27, 36, 97, 104, 143, 190, 193, 198, 199, 201, 202, 204, 216, 222, 223, 250, 251, 275, 287, 322, 323, 332, 350, 378, 408, 426, 439, 508, 509, 510, 531, 570, 580, 585, 587, 597, 602, 603, 638, 659, 669, 679, 847, 869, 900, 978, 979, 1035, 1084, 1126, 1128, 1178, 1192, 1218, 1219, 1222, 1248, 1256, 1269, 1355, 1426, 1429, 1430, 1451, 1476, 1506, 1574, 1575, 1635, 1651.

N.O. Aguilar & M. Brink

## *Alternanthera* Forssk.

Fl. aegypt.-arab.: 28, lix (1775).

AMARANTHACEAE

$x$  = unknown; *A. ficoidea*:  $n = 34$ , *A. philoxeroides*:  $2n = 28, 100$ , *A. sessilis*:  $n = 17, 18, 20$ ,  $2n = 34, 96$

**Major species** *Alternanthera sessilis* (L.) DC.

**Origin and geographic distribution** *Alternanthera* comprises approximately 150 species and is distributed in all tropical and subtropical regions, but the New World tropics are by far richest in species. Seven species have been found in Malesia, only one of which (*A. sessilis*) is indigenous; the other species have been introduced and are often locally naturalized.

**Uses** An infusion of the entire plant of *A. sessilis* is used in Indonesia as a remedy against intestinal cramps, diarrhoea and dysentery, and externally as a cooling agent to treat fever. In Malaysia it is used internally against intestinal inflammation and fever, and externally to treat wounds. *A. sessilis* is used in local medicine in Taiwan, often in mixtures with other medicinal plants such as *Eclipta prostrata* (L.) L., *Hypericum ascyron* L. and *Wollastonia chinensis* (Osbeck) Merr., to treat hepatitis, tight chest, bronchitis, asthma and lung troubles, to stop bleeding and as a hair tonic. It is used locally in Thailand and India against dysentery, as cholagogue, abortifacient and febrifuge, and to treat snake bites, inflamed wounds and boils, and in Thailand and Sri Lanka as a galactagogue. An extract from *A. philoxeroides* is used medicinally in India to treat 'female diseases'. *A. pungens* is reported in India to be useful as diuretic and to treat gonorrhoea.

The densely matted growth of some species (e.g. *A. ficoidea*) makes them useful for protecting soil against erosion. *A. philoxeroides* can be used as a tertiary filtration system for domestic sewage. Several species are planted in gardens as ornamentals. Some *Alternanthera* species are valued for aquarium decoration. Cooked leaves of *A. sessilis* are sometimes eaten as vegetable.

**Production and international trade** Dried plants of *Alternanthera* are only occasionally traded on local markets and by herbalists. They are not traded on the international market.

**Properties** Little is known about the chemical constituents of the various *Alternanthera* species. A C-glycosylated flavonoid, alternanthin, has been isolated from *A. philoxeroides*. The triterpenes  $\alpha$ -spinasterol and  $\beta$ -spinasterol have been demonstrated to occur in several *Alternan-*

*thera* species, among which *A. sessilis*. This species is also reported to contain stigmaterol,  $\beta$ -sitosterol, oleanotic acid and its derivatives, and saturated (aliphatic) esters. A high iron content (about 2%) has been found in *A. philoxeroides*.

*A. sessilis* leaves contain 12 g/100 g dietary fibre. Incorporation of about 75 g of this vegetable fibre in the daily diet of diabetics, significantly reduced the postprandial blood glucose level. The leaves of *A. sessilis* are rich in  $\beta$ -carotene.

In tests in India, leaf pastes of *A. sessilis* exhibited inhibition of mutagenicity in *Salmonella typhimurium* strains. They inhibited the formation of the potent environmental carcinogen nitrosodietanolamine from its precursors such as triethanolamine. The aqueous alcohol extract of the entire plant exhibits hypothermic and histaminergic activities and relaxes smooth muscles. An ether extract of *A. sessilis* yielded an active principle having anti-ulcerative properties.

An aqueous extract of *A. philoxeroides* inhibited the growth of the human immunodeficiency virus (HIV-1) in vitro at concentrations non-toxic to the host cells (H9 cells, human T-helper lymphocytes). It was also inhibitory to the growth of the herpes simplex (HSV) and respiratory syncytial viruses (RSV). The infectivity of extracellular virions of HIV-1, HSV and RSV was partially destroyed by the extract, but no activity on extracellular virions, or on virus growth was found for vesicular stomatitis virus (VSV), adenovirus (AV) or polio virus (PV). Chemical studies have indicated that the active anti-HIV-1 component was heat-stable, water-soluble and non-dialysable. Preliminary chemical characterization revealed that it might be a partially sulphonated polysaccharide.

*A. philoxeroides* extracts can markedly protect suckling mice from being infected by epidemic haemorrhagic fever virus (EHFV). After the infected mice were treated, their survival rate increased and pathological lesions and virus antigen in the tissues mitigated as compared with the controls. However, therapeutical doses caused slight deformations of the hepatic cells. Preliminary chemical investigations revealed that the active component might be a coumarin analogue.

Extracts prepared (leaves, cold water) from *A. brasiliensis* and *A. ficoidea* showed antiviral activity against herpes simplex (HSV-1) virus in HEP-2 cells in vitro at non-toxic concentrations. The activity is however, influenced by the process of extracting. Using hot water or a lyophilization procedure might drastically decrease the activity.

Crude aqueous extracts of *A. brasiliensis* and *A. ficoidea* also showed antiviral activity against Aujeszky disease virus (ADV) in IB-RS-2 pig cell cultures, and bovine diarrhoea virus (BVD) in GBK bovine cell lines. The antiviral activity might vary depending on the temperature used during extraction.

Antiviral activity against tobacco mosaic virus in beans and tobacco has also been demonstrated for several *Alternanthera* species.

**Description** Annual or perennial herbs, erect, ascending, creeping, clambering or floating, often much branched, often hairy with dentate or smooth hairs. Leaves opposite, simple and entire, with short petiole; stipules absent. Inflorescence an axillary or rarely terminal head, sessile or stalked, sometimes a short spike. Flowers bisexual, solitary in axils of bracts, subtended by 2 scarious bracteoles; perianth often dorsally compressed, with 5 free, equal or unequal tepals; stamens 2-5, sometimes some without anthers, filaments united at the base into a short cup or longer tube, usually alternating with staminodes, anthers small, 1-celled; ovary superior, 1-celled, often compressed, style short with capitate stigma. Fruit an indehiscent utricle, sometimes corky, 1-seeded, falling off with the perianth and with or without bracteoles. Seed variably lenticular. Seedling with epigeal germination; cotyledons leafy, glabrous, apex rounded; hypocotyl and epicotyl elongated, purplish.

**Growth and development** *A. philoxeroides* is a  $C_3$  plant. Each stem node is capable of producing a new plant under favourable conditions.

The dark brown, corky fruits of *A. sessilis* often float in great quantities on the water. Some of the introduced species (*A. ficoidea*, *A. philoxeroides*) do not set fruit in Malasia.

**Other botanical information** *Alternanthera* is closely related to *Gomphrena*, which differs particularly in the shape of the androecium.

There is much confusion about the correct name for the taxon which is dealt with here under the name *A. ficoidea*. In 1989 the Committee for Spermatophyta of the International Association for Plant Taxonomy recommended maintaining the much used name *A. ficoidea* for this species, even though it is nomenclaturally incorrect (the correct name is *A. tenella*), in order to stabilize the nomenclature. *A. ficoidea* cultivated in South-East Asia should be considered as a cultigen of the South American species.

**Ecology** *Alternanthera* species are usually found in moist localities. *A. sessilis* prefers open

places such as roadsides, gardens and along rice fields, but *A. brasiliensis* shows a preference for shaded localities (e.g. on slopes). *A. philoxeroides* can tolerate a large range of habitats, growing out over water as dense floating mats or on moist soil as individual plants. Plants of this species can grow at NaCl concentrations of 400 mol/m<sup>3</sup>. *A. pungens* is adapted to more dry locations.

Several species are noxious weeds, e.g. *A. sessilis* in upland rice, carrot and tomato, *A. philoxeroides* in irrigated rice and *A. brasiliensis* in coffee. *A. ficoidea* is a dreaded weed in beans, soybeans, groundnuts, plums and cotton in America.

The leaves of some *Alternanthera* (e.g. *A. sessilis*) can be purplish pigmented by the occurrence of anthocyanins and betalains. An inverse correlation has been observed between pigmentation intensity and soil moisture.

**Propagation and planting** *A. ficoidea* and *A. philoxeroides* do not produce ripe seeds in Malaysia. However, *A. ficoidea* can easily be propagated from cuttings or divisions, whereas *A. philoxeroides* can rapidly develop new plants from stem parts. *A. sessilis* can probably also be propagated very easily from seed and vegetatively.

#### **In vitro production of active compounds**

Cell suspension cultures of *A. philoxeroides* have been derived from leaf callus grown at 25°C in the dark in Murashige and Skoog medium containing 1 mg/ml 2,4-D. After an abrupt and substantial increase in salinity, the suspension cells exhibited a rapid attainment of water balance and cell growth. Concomitant with the ability of these cultures to withstand an increase in NaCl are the abilities to produce betaine and to take up exogenous betaine.

**Husbandry** In general, *Alternanthera* can stand pruning well.

**Diseases and pests** A leaf spot disease of *A. sessilis* caused by *Fusarium pallidoroseum* has been described in Nigeria. It may spread to crops in which *A. sessilis* occurs as a weed, e.g. okra, yams, potatoes, onions and carrots.

Introduction of the lepidopterous *Vogtia malloi* from Argentina and the alligator weed flea beetle (*Agasicles hygrophila*) from the United States into several countries has successfully reduced populations of the noxious aquatic weed *A. philoxeroides*.

**Genetic resources and breeding** *Alternanthera* species are not endangered. On the contrary, they are often noxious weeds expanding their area of distribution (e.g. *A. philoxeroides* in Australia). However, the genetic variability of the

species originally introduced in South-East Asia is rather small because of the limited number of introductions and the vegetative mode of reproduction. It is unclear whether the medicinal properties attributed to these introduced species apply to the present South-East Asian populations as well.

**Prospects** Some of the medicinal properties of *Alternanthera* warrant further research. The antiviral effects of *A. philoxeroides*, and the anti-ulcerative properties of *A. sessilis* seem to be the most promising.

**Literature** [1] Backer, C.A., 1949. Amaranthaceae. In: van Steenis, C.G.G.J. (Editor): Flora Malesiana. Series 1, Vol. 4. Noordhoff-Kolff N.V., Djakarta, Indonesia. pp. 91-94. [2] Council of Scientific and Industrial Research (various editors), 1985. The wealth of India. Revised Edition. Vol. 1. Publications and Information Directorate, New Delhi, India. pp. 206-207. [3] Dharma, A.P., 1981. Indonesische geneeskrachtige planten [Indonesian medicinal plants]. De Driehoek, Amsterdam, the Netherlands. p. 21. [4] Larsen, K., 1989. Caryophyllales. In: Lescot, M. (Editor): Flore du Cambodge, du Laos et du Vietnam. Vol. 24. Muséum National d'Histoire Naturelle, Paris, France. pp. 46-52. [5] Koseki, I., Simoni, I.C., Nakamura, I.T., Noronha, A.B. & Costa, S.S., 1990. Antiviral activity of plant extracts against aphthovirus, pseudorabies virus and pestivirus in cell cultures. Microbios Letters 44(173): 19-30. [6] Krishnakumar, A., Sivaramakrishnan, V.M. & Sivaswamy, S.N., 1991. Inhibition of nitrosation reaction by some spices/leafy vegetables. Advances in Plant Sciences 4(1): 189-193. [7] Lagrota, M.H.C., Wigg, M.D., Miranda, M.M.F.S., Santos, M.G.M. & Costa, S.S., 1995. Inhibition of herpes simplex virus replication by different extracts of Caryophyllales. Biomedical Letters 51(202): 127-135. [8] Noronha, A.B., Amelia, M., Alexandre, V., De Gaetano, R. & Vicente, M., 1993. Protection against tobacco mosaic virus induced by some Caryophyllales plant extracts. Microbios 73(294): 75-80. [9] Si-man, Z., Yong-sheng, H., Tappa, H.D. & Smith, K.M., 1988. Inhibitor against the human immunodeficiency virus in aqueous extracts of *Alternanthera philoxeroides*. Chinese Medical Journal 101: 861-886. [10] Sreedevi & Chaturvedi, A., 1993. Effect of vegetable fibre on post prandial glycemia. Plant Foods for Human Nutrition 44(1): 71-78.

*Selection of species****Alternanthera brasiliana* (Torner) O. Kuntze**

Revis. gen. pl. 2: 537 (1891).

**Synonyms** *Alternanthera strigosa* Hassk. (1839).

**Distribution** Native of tropical America; introduced and naturalized in western and central Java.

**Uses** *A. brasiliana* might be useful as an antiviral agent.

**Observations** A perennial herb up to 3 m tall, decumbent at base, ascending-erect or clambering higher up, often widely branched, stems solid; hairs minutely dentate; leaves ovate-lanceolate, 3.5–10 cm × 1–4 cm, densely appressed pilose on both surfaces when young but later slowly glabrescent, petiole 0.5–1.5 cm long; flowering heads stalked; tepals strongly 3-veined, yellowish-white, filaments united at base into a distinct tube; fruit ellipsoidal, about 2 mm long. *A. brasiliana* occurs locally gregariously in Java, on moist, shaded localities on ravine slopes and stream banks, at 200–600 m altitude.

**Selected sources** 47, 93, 97, 773, 805, 1056.

***Alternanthera ficoidea* (L.) P. Beauv.**

Fl. Oware 2: 66, pl. 99, fig. 1 (1818; 'ficoidea').

**Synonyms** *Gomphrena ficoidea* L. (1753), *Alternanthera tenella* Colla (1828).

**Vernacular names** Indonesia: bayam merah (general), jukut jatinangor (Sundanese), kecicak abang (Javanese). Thailand: phakpet daeng, phrommi daeng, phakpet farang. Vietnam: rau d[ee]u d[or].

**Distribution** Native of tropical South America; introduced in Malesia (e.g. Sumatra, Java, Papua New Guinea) and elsewhere in South-East Asia as an ornamental.

**Uses** *A. ficoidea* might be useful as an antiviral agent. The densely matted growth makes it suitable to be used to protect soil against erosion. It is commonly planted in gardens as ornamental. The leaves are sometimes eaten as a vegetable, e.g. in Sri Lanka.

**Observations** A perennial herb up to 50 cm tall, erect or decumbent and rooting in lower part, often much branched and forming dense tufts, stems solid; hairs dentate; leaves oblong, oblong-obovate to spatulate, 1–6 cm × 0.5–2 cm, finely pilose when young but later glabrescent, often variegated with brownish-red, red, pink or yellow, petiole 1–4 mm long; flowering heads sessile; 3 outer

tepals distinctly 3-veined, shiny white or yellowish, filaments united at base into a very short cup; fruit not produced in Malesia. In South-East Asia only var. *versicolor* (Lem.) Backer (synonyms: *Alternanthera amoena* Backer & v. Slooten, *A. bettzickiana* (Regel) Nicholson, *A. ficoidea* (L.) P. Beauv. var. *bettzickiana* (Nicholson) Backer, *A. manillensis* (Walp.) Kanis (1972), *A. tenella* Colla var. *versicolor* (Lem.) Veldk.) is cultivated. In fact, this taxon should be considered as a cultivar. It hardly ever sets fruit and besides being cultivated as an ornamental, is known as a non-persisting escape.

**Selected sources** 47, 93, 97, 580, 773, 805, 816, 1055, 1470.

***Alternanthera philoxeroides* (Mart.) Griseb.**

Goett. Abh. 24: 36 (1879).

**Synonyms** *Telanthera philoxeroides* (Mart.) Moq. (1849).

**Vernacular names** Alligator weed (En). Indonesia: tolod, krokot (Java). Burma (Myanmar): kanabaw. Thailand: phakpet-nam, phakpet.

**Distribution** Native of tropical South America; introduced and naturalized in western Java, and very locally elsewhere in Malesia (e.g. found in south-eastern Kalimantan).

**Uses** An extract is used medicinally in India to treat 'female diseases'. In Indonesia young tops are eaten raw or cooked. It can be used as a tertiary filtration system for domestic sewage. In China *A. philoxeroides* is cultivated for compost-making, whereas in the United States it is grown as food for lobsters. In watercourses it is a noxious weed.

**Observations** A perennial herb up to 1 m tall, ascending from a creeping or floating, rooting base, often much branched and forming a dense mass, stems fistulose in lower part; hairs smooth; leaves oblong or oblong-obovate, 2–8 cm × 0.5–2.5 cm, glabrous or ciliate, petiole 3–6 mm long; flowering heads stalked or occasionally sessile; tepals 1-veined, shining white, filaments united at base into a distinct tube; fruit not produced in Malesia. *A. philoxeroides* occurs locally gregariously in Java, in stagnant or slow-moving water in pools and ditches.

**Selected sources** 93, 97, 106, 288, 868, 888, 1056, 1174, 1344, 1356, 1616.

***Alternanthera pungens* Kunth**

Humb., Bonpland & Kunth, Nov. gen. sp. 2: 206 (1818).

**Synonyms** *Alternanthera repens* (L.) Link

(1821) non Gmelin, *Telanthera pungens* (Kunth) Moq. (1849).

**Vernacular names** Thailand: khok krasun, khok krasun lek.

**Distribution** Native of tropical America; introduced and naturalized locally in Java, Thailand and India.

**Uses** In India, it is said to possess diuretic properties, and a decoction is used internally to treat gonorrhoea.

**Observations** A perennial prostrate herb up to 50 cm tall, with robust taproot, often woody at base of stem, much branched, stems solid; hairs minutely dentate; leaves elliptical-obovate, 1–4.5 cm × 0.5–2 cm, glabrous or slightly appressed pilose, petiole 2–10 mm long; flowering heads sessile; tepals 3-veined in the lower half, white, filaments united at base into a very short cup; fruit broadly ovoid, much compressed, about 1.5 mm long. *A. pungens* occurs locally in Java and Papua New Guinea, on waste land, dockyards and roadsides in the lowland near the coast. In India it is a weed of pebbly soils and dry locations.

**Selected sources** 93, 97, 288, 704, 816, 1470.

### *Alternanthera sessilis* (L.) DC.

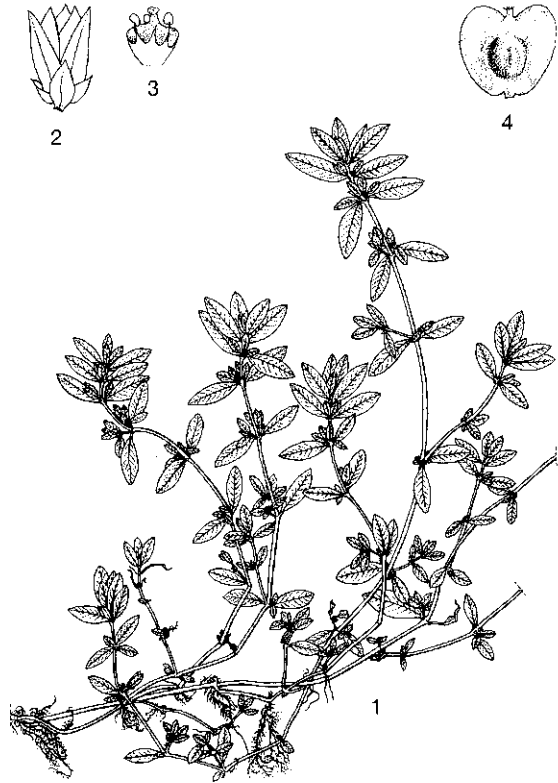
Cat. pl. horti monsp.: 77 (1813).

**Synonyms** *Alternanthera triandra* Lamk (1783), *Alternanthera denticulata* R.Br. (1810), *Alternanthera nodiflora* R.Br. (1810).

**Vernacular names** Indonesia: kremek (Sundanese), bayem kremah (Javanese), daun tolod (Moluccas). Malaysia: keremak, pudoh, kermak bukit. Philippines: bunga-bunga (Tagalog), bilanamanut (Magindanao), gogoat (Bontok), Cambodia: ch̄ng b̄ngko:ng (Kompong Thom). Laos: khaix ped, phak ph'è:w (Louang Prabang), n̄: ti:d kho:x (Vientiane). Thailand: phakpet khaao, phakpet thai. Vietnam: rau d[ee]ju (general).

**Distribution** Throughout the tropics and subtropics of America, Africa and Asia; throughout Malasia.

**Uses** An infusion of the entire plant is used in Indonesia as a remedy against intestinal cramps, diarrhoea and dysentery, and externally as a cooling agent to treat fever. In Malaysia it is used internally against intestinal inflammation and fever, and externally to treat wounds. *A. sessilis* is used in local medicine in Taiwan, often in mixtures with other medicinal plants, to treat hepatitis, tight chest, bronchitis, asthma, and lung troubles, to stop bleeding and as a hair tonic. It is used locally in India against dysentery, as a cholagogue, abortifacient and febrifuge and to treat snake bites, in-



*Alternanthera sessilis* (L.) DC. – 1, plant habit; 2, flower with bract and bracteoles; 3, flower with tepals removed; 4, fruit.

flamed wounds and boils, and in Thailand and Sri Lanka as galactagogue. It is eaten as a vegetable, e.g. in Vietnam and Sri Lanka.

**Observations** A perennial, sometimes annual herb up to 1 m tall, erect, ascending or creeping, often widely branched, taproot robust, stems solid, sometimes floating in water and then stems fistulose in lower part; hairs smooth; leaves linear-lanceolate, oblong to ovate or obovate, 1–15 cm × 0.5–3 cm, glabrous or sparsely pilose, petiole 1–5 mm long; flowering heads sessile; tepals 1-veined or only 3-veined at the very base, shiny white or purplish, filaments united at base into a very short cup; fruit obreniform, corky, about 2 mm long. *A. sessilis* is a very common plant of constant or periodically humid, open localities in roadsides, gardens, ditches, swamps, rice fields and tea plantations, up to 1250 m altitude.

**Selected sources** 93, 97, 202, 288, 350, 580, 704, 783, 816, 860, 868, 1035, 1083, 1178, 1370, 1394, 1470.

R.H.M.J. Lemmens & S.F.A.J. Horsten

**Amaranthus spinosus L.**

Sp. pl. 2: 991 (1753).

AMARANTHACEAE

2n = 34

**Vernacular names** Spiny amaranth, prickly amaranth, spiny pigweed (En). Epinard malabre (Fr). Brunei: bayam berduri (Malay). Indonesia: bayam duri (general), bayem eri (Javanese), senggang cucuk (Sundanese). Malaysia: bayam duri, bayam hutan (general). Philippines: urai (Tagalog), harum (Bisaya), kalunai (Iloko). Cambodia: phti: bânla: (Pursat). Laos: hôm hna:m (general). Thailand: mang-lang-du (Karen-Mae Hong Son), pa-tue (Khmer), phak hom nam (peninsular). Vietnam: rau d[ee]f[n] gai (general), gileef[n] gai (Ha Nam Ninh).

**Origin and geographic distribution** *A. spinosus* occurs in all tropical and subtropical regions, including the whole of South-East Asia, often gregariously and as a weed. It is sometimes found in temperate zones as well. It has been suggested that spiny amaranth originates from lowland tropical South and Central America, and that it was introduced in other warmer parts of the world from about 1700 AD onwards. Nowadays it is rarely cultivated.

**Uses** The root of spiny amaranth is known as an effective diuretic. In the Philippines, India, Thailand and Indo-China, a decoction of the root is used to treat gonorrhoea. It is also applied as an emmenagogue and antipyretic. In many countries, including Indonesia, the bruised leaves are applied externally in cases of eczema, burns, wounds and boils. The leaves are considered a good emollient. In Malaysia, spiny amaranth is used as an expectorant and to relieve breathing in acute bronchitis. In mainland South-East Asia, it is also used as a sudorific, febrifuge, antidote to snake poison, galactagogue, and to treat menorrhagia. Spiny amaranth is reported to be used to treat haemorrhoids in Africa. Some tribes in India apply spiny amaranth to induce abortion.

In Indo-China and India, spiny amaranth is used as forage, and it is said to increase the yield of milk in cattle. However, cases of poisoning in cattle have also been reported. Spiny amaranth is browsed by sheep and goats and is a highly nutritious feed at any time during the year. The young leaves are sometimes eaten as a vegetable.

**Production and international trade** Spiny amaranth is not traded commercially, and is rarely found in local markets.

**Properties** Little is known about the specific

constituents of *A. spinosus*. The roots contain  $\alpha$ -spinasterol and some saponins. Sterols ( $\beta$ -sitosterol, stigmasterol, campesterol and cholesterol), n-alkanes, fatty acids (e.g. stearic-, oleic- and linoleic acid) and free alcohols have also been found in the petroleum-ether extract of the herb. The flavonoid rutin has been found in the above-ground matter of spiny amaranth in a concentration up to 1.9%, and traces of hydrocyanic acid in the leaves. Spiny amaranth is furthermore reported to contain a considerable amount of potassium, up to 4.5% in the dried leaves, which might explain the known diuretic properties.

A lectin has been purified from the seeds by means of chromatographic procedures. Its reaction was non-specific in general: it reacted with human and various animal erythrocytes. Its unique carbohydrate specificity will prove useful in biochemistry. Lecithins are also known to occur in the seeds of several other *Amaranthus* species, e.g. *A. caudatus* L. and *A. leucocarpus* S. Watson.

Spiny amaranth possesses a strong phagocytic effect. No antibacterial activity has been demonstrated, but crude aqueous extracts showed fungicidal activity against *Cercospora cruenta*, which causes a leafspot disease in mung bean (*Vigna radiata* (L.) Wilczek). However, these extracts were inferior to benomyl in controlling the disease. Furthermore they showed antiviral activity against Aujeszky virus (ADV) in IB-RS-2 pig cell cultures and bovine diarrhoea virus (BVD) in GBK bovine cell lines. The antiviral activity against BVD, however, was lost upon heating the extract for 30 minutes at 50–60°C. Thus it is possible that at least a part of the antiviral activity resides in proteins or glycoproteins that are largely inactivated by heating for 10 minutes above 60°C.

Spiny amaranth has considerable nutritional value. The high foliar content of the amino acid lysine could make it a valuable protein supplement in cereal-based diets. However, cases of spontaneous poisoning by spiny amaranth in cattle have been reported, particularly after severe droughts when few other forages were available. It was suggested that the spiny amaranth caused renal failure.

*A. spinosus* pollen may cause hay fever, but the reaction is usually milder than that caused by some grass pollen. Hypo-sensitization injection treatment with a mixture of 3 allergenic grasses and spiny amaranth for at least one year gave significant improvement in patients with allergic rhinitis and/or seasonal asthma in the Philippines. Allelochemicals have been isolated and identified



from aerial plant parts. These are volatile aliphatic compounds which inhibit germination of seeds of crops like carrot, tomato and onion. The most active alcohols present in spiny amaranth are 3-methyl-1-butanol and 3-hexen-1-ol; the most active aldehyde is 3-methylbutanal, and the most active ketone is 2-heptanone.

**Adulterations and substitutes** The *Amaranthaceae* genus *Aerva* has similar properties to spiny amaranth and is used for similar complaints. It seems probable that *Amaranthus* and *Aerva* are related not only botanically but also chemically.

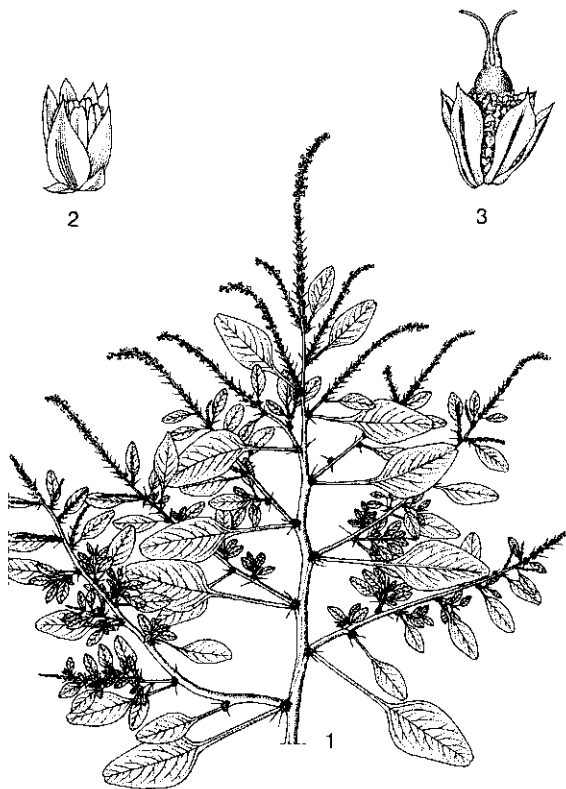
**Description** An annual, erect monoecious herb up to 100(–130) cm tall, much branched; stem terete or obtusely angular, glabrous or slightly pubescent, green or variably suffused with purple. Leaves alternate, simple and entire, ovate-lanceolate to rhomboid, 3.5–11 cm × 1–4.5 cm, acute and often slightly decurrent at base, obtuse, rounded or slightly retuse and often short mucronate at apex, glabrous or slightly pubescent on veins

when young; petiole rather long, approximately as long as leaf-blade; stipules absent. Inflorescence consisting of dense clusters, lower ones axillary, higher ones often collected in an axillary and terminal spike which is often branched in its lower part; axillary clusters usually armed with (1–)2(–3) very sharp spines up to 2 cm long. Flowers solitary in the axil of a bract, subtended by 2 bracteoles, bracts and bracteoles scarious, mucronate from a broad base, shorter or as long as the perianth, unisexual; male flowers usually arranged in a terminal spike above the base of the inflorescence, green; tepals 5 or in male flowers often 3, free, subequal, ovate-oblong to oblong-spatulate, up to 2.5 mm long, very convex, membranous, with transparent margins and green or purple median band; stamens 5, about as long as tepals; ovary superior, oblong, 1-celled, styles 2–3, ultimately recurved. Fruit an oblong utricle with persisting styles, circumscissile a little below the middle or indehiscent, 1-seeded. Seed about 1 mm in diameter, shiny black or brownish-black with thin margin. Seedling with epigeal germination; cotyledons leafy, glabrous, apex rounded to slightly acute; hypocotyl up to 12 mm long, epicotyl absent.

**Growth and development** In India spiny amaranth flowers twice a year. Seeds mature in about one month after flowering. They are scattered around the mother plants or distributed by animals feeding on the plants. It has been observed that large numbers of seedlings emerge from decaying cattle faecal deposits. Seeds are eaten by birds.

In India, seeds germinate throughout the year but seedlings exhibit a high degree of mortality. Less than 1% of them reach the first leaf stage; less than 5% of these reach the 4-leaf stage and continue growing.

**Other botanical information** Some other species of the genus *Amaranthus* (about 40 species worldwide) are also used medicinally, but have other primary uses. The leaves of the well-known vegetable *A. tricolor* L. are considered as a good emollient in Vietnam, and are used in Malaysia to treat haemorrhagia. The leaves of the lesser-known vegetable *A. viridis* L. are used in Africa as a febrifuge and as a poultice to treat inflammations, boils and abscesses. In South and Central America, they are used as a diuretic and galactagogue (applied as an infusion) and as emollient. In India, they are used to treat snake bites and scorpion stings, and in New Britain to treat mosquito bites and insect stings. The seeds of the grain



*Amaranthus spinosus* L. – 1, part of flowering plant; 2, male flower with bracteoles; 3, fruit.

amaranth *A. cruentus* L. are used in India as diuretic and to treat scrofulous sores.

**Ecology** Spiny amaranth is adapted to a wide range of climatic and edaphic factors. It grows best in the sun or in light shade; a light intensity of less than 30% completely suppresses flowering. Flowering is earliest and most abundant in areas with daylengths of 11–12 hours. It is nitrophilous and prefers soils with a high organic matter content, but is also able to grow on sandy soils. Optimal growth is obtained on soils with moderate moisture content, but spiny amaranth is capable of growing on wet soils as well. It is drought-resistant and can even grow under arid conditions.

Spiny amaranth is a very noxious weed in many parts of the world. It is, for instance, troublesome in upland rice, sugar cane and carrot in Indonesia, in maize in the Philippines, in groundnut and soya bean in Taiwan, and in tomato and field pea in India. In South-East Asia, it is very common in roadsides, waste places, railway yards, cropped land and gardens, up to 1400 m altitude.

**Propagation and planting** Spiny amaranth is propagated by seed. Some types are known to produce 235 000 seeds per plant. The weight of 100 seeds is 14–25 mg. Freshly collected seeds may germinate at temperatures as high as 40°C, with a germination rate of up to 95%. After storage, however, temperature requirements are lower. Seeds stored for one month at room temperature have almost 100% germination, and after 5 months they have approximately 90% germination. When they are stored for one year at 20°C the germination rate will drop to about 50%, but storage at lower temperatures gives a higher rate.

**Husbandry** As a weed in tomato in India spiny amaranth has been successfully controlled by the application of geraniol, which completely blocked the germination of the weed without affecting the the tomato crop. An ethanolic extract of seeds of *Coffea arabica* L. (with 1,3,7-trimethylxanthine as active ingredient) at a concentration of 1.2 g/l, completely inhibited germination of spiny amaranth in a crop of black gram (*Vigna mungo* (L.) Hepper) without negative effects for this pulse crop.

**Diseases and pests** Spiny amaranth is a host plant for, among others, tobacco mosaic virus, groundnut rosette virus, cucumber mosaic virus and root-knot nematodes (*Meloidogyne* spp.), which attack some commercial crops. When the world's worst weeds are ranked on the basis of the number of pests hosted, spiny amaranth is placed number 6, hosting 15 pests that may affect crops.

Some natural insect enemies of spiny amaranth have been recorded from Mexico: the pyralid *Herpetogramma bipunctalis* and the curculionid *Conotrachelus seniculus*. These might be useful for biological control. In India, the bud weevil *Ceuthorrhynchus asperulus*, a pest of pigeon pea (*Cajanus cajan* (L.) Millsp.), has been found feeding on *Amaranthus* species including spiny amaranth.

**Genetic resources and breeding** The genetic variability of spiny amaranth is great because of its enormous area of distribution and its wide ecological adaptation.

**Prospects** The medicinal properties of spiny amaranth have received very little attention. The diuretic and anti-inflammatory properties in particular deserve more research, as these properties are valued in many different regions of the world. Moreover, the reputed high nutritional value of the leaves offers good prospects for more common use as a vegetable or a forage.

**Literature** |1| Backer, C.A., 1949. Amaranthaceae. In: van Steenis, C.G.G.J. (Editor): Flora Malesiana. Series 1, Vol. 4. Noordhoff-Kolff N.V., Djakarta, Indonesia. pp. 78–79. |2| Connick, W.J., Bradow, J.M. & Legendre, M.G., 1989. Identification and bioactivity of volatile allelochemicals from Amaranth residues. *Journal of Agricultural and Food Chemistry* 37(3): 792–796. |3| Council of Scientific and Industrial Research (various editors), 1985. The wealth of India. Revised Edition. Vol. 1. Publications and Information Directorate, New Delhi, India. p. 219. |4| Dharma, A.P., 1981. Indonesische geneeskrachtige planten [Indonesian medicinal plants]. De Driehoek, Amsterdam, the Netherlands. p. 25. |5| Gopal, B., 1974. Autecological study of *Amaranthus spinosus* L. *Annals of Arid Zone* 13(3): 187–195. |6| Koseki, I., Simoni, I.C., Nakamura, I.T., Noronha, A.B. & Costa, S.S., 1990. Antiviral activity of plant extracts against aphthovirus, pseudorabies virus and pestivirus in cell cultures. *Microbios Letters* 44(173): 19–30. |7| Larsen, K., 1989. Caryophyllales. In: Lescot, M. (Editor): Flore du Cambodge, du Laos et du Vietnam. Vol. 24. Muséum National d'Histoire Naturelle, Paris, France. pp. 20–21. |8| Maroon, C.J.M., Poina, O.S. & Molina, A.B., 1984. Evaluation of crude plant extracts as fungicides for the control of *Cercospora* leafspot of mungbean (*Vigna radiata* (L.) Wilczek). *Philippine Phytopathology* 20(1–2): 27–38. |9| Oliveira, J.S. & de Carvalho, M.F., 1975. Nutritional value of some edible leaves used in Mozambique. *Economic Botany* 29(3): 255–263. |10| Singh, J., Kamboj, K.K., Kam-

boj, S.S., Sandhu, R.S. & Shangary, S., 1993. Affinity, purification and characterization of lectins from two *Amaranthus* species. *Plant Science* 94(1-2): 47-53.

**Other selected sources** 190, 202, 303, 332, 580, 868, 1035, 1169, 1178, 1234, 1424, 1470.

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## Amomum Roxb.

Fl. Ind. 1: 317 (1820); Pl. Coromandel 3: 75 (1820).

### ZINGIBERACEAE

$x = 24$ ; *A. aculeatum*:  $n = 24$ , *A. krervanh*:  $2n = 48$ , *A. longiligulare*:  $2n = 48$ , *A. squarrosum*:  $n = 24$ , *A. uliginosum*:  $n = 24$ , *A. villosum*:  $2n = 48$ , *A. xanthioides*:  $2n = 48$

**Major species** *Amomum krervanh* Pierre ex Gagnep., *A. villosum* Lour., *A. xanthioides* Wallich ex Baker.

**Vernacular names** Thailand: krawaan. Vietnam: sa nh[aa]n.

**Origin and geographic distribution** *Amomum* consists of about 100 species and is distributed in eastern Asia, from India and China, throughout the Malesian region, to tropical Australia. In Vietnam 22 species have been found. The total number of species in Malesia is difficult to estimate, but Peninsular Malaysia has approximately 18 species, Java about 13, and Borneo about 30; the distribution of the genus in other areas is very incompletely known.

**Uses** Several *Amomum* species (for instance *A. krervanh*, *A. villosum* and *A. xanthioides*) are well known medicinal herbs in China, Indo-China and Thailand, particularly their fruits. They are mainly used to treat gastric and digestive disorders (e.g. vomiting, poor appetite, poor digestion, colic, diarrhoea). Besides their tonic, carminative and stomachic properties, the fruits are also considered to have emmenagogue and febrifugal properties. They are also sometimes prescribed to treat tuberculosis with hemoptysis, liver and uterine affections, and rheumatism. Seeds of *A. villosum* are known as 'saren' in China, and the fruit shells as 'sake'. Some medicinal uses have been reported from the Malesian region; *Amomum* seeds are often included in stomachics and in preparations for coughs and colds.

The fruits of several species are commonly used as cooking ingredient, and are sometimes also used in perfumery. They may serve as antioxidant in foods.

**Production and international trade** The fruits have importance in southern China (Yunnan). The total cultivated area is estimated to be over 13 000 ha. Around 1993 the dried fruits of *A. villosum* had a value of US\$ 5.5/kg. Vietnam exports seed to China yearly, e.g. of *A. villosum* and *A. xanthioides*. In Cambodia, the fruits are mainly collected from the natural forest, whereas in Thailand the plants are mainly cultivated.

**Properties** Fruits contain an essential oil, which is colourless and transparent and has the characteristic odour of the fruit and the seed. The fruits of *A. krervanh* contain approximately 3-4% essential oil with 1,8-cineol (eucalyptol) as main compound (60-80%). Further constituents are camphene, p-cymene,  $\alpha$ -humulene, limonene,  $\alpha$ -pinene, terpinene and  $\alpha$ -terpineol. The pharmacological activities reported for *A. krervanh* include antifungal, antipyretic, smooth muscle relaxant and hypotensive activity. Fruits of *A. villosum* are reported to contain 3% essential oil consisting of bornylacetate (34%), camphor (27%), borneol (13%), camphene (10%), limonene (7%) and the minor compounds  $\alpha$ - and  $\beta$ -pinene and myrcene.

Fruits of *A. xanthioides* contain 1.7-3% oil. The essential oils from *A. xanthioides* can be divided into 3 groups, based on the nature of their main compounds; the first group mainly containing camphor and bornylacetate, the second one mainly composed of linalool and nerolidol, and the third one with a high content of  $\beta$ -caryophyllene. The oil of the first group in general consists of camphor (29%), bornylacetate (22%), camphene (13.5%), limonene (10%), myrcene (4%),  $\beta$ -pinene (4%) and  $\alpha$ -pinene (3%).

Investigations of *A. villosum* cultivated in Yunnan (China) revealed the presence of ethyl-octacosate, docosyl hexylate, stigmast-4-ene-1,3-dione,  $\beta$ -sitosterol and daucosterol in the roots and rhizomes.

A diterpene peroxide has been isolated from *A. krervanh* fruits in Thailand. This compound exhibited potent activity against *Plasmodium falciparum* and is thus of interest in combating malaria. In vitro experiments showed that the compound has roughly one-tenth the activity of artemisinin (from *Artemisia annua* L.) and the same level of activity as arteflene, which is an effective synthetic antimalarial agent structurally related to artemisinin. It is interesting to note that *A. fenzlii* Kurz is used to treat malarial fever in India, as is *A. tsao-ko* Crevost & Lem. in China. Crude drugs prepared from *A. xanthioides* showed antifungal activity using organic solvents. At 3000

ppm the Indian *A. subulatum* Roxb. showed 100% inhibition for the storage fungus *Aspergillus flavus*; it showed a broad range of fungitoxicity in tests with plant pathogens, and had no adverse effect on the germination of rice; the seed oil was also highly active against keratinophilic fungi. *A. subulatum* also showed some insecticidal activity against the storage pest *Sitophilus oryzae*.

It has been reported from the Andaman Islands that sap of an *Amomum* species (doubtfully identified as *A. aculeatum*) acts as a tranquillizer for bees (rock bees, *Apis dorsata*), and is used in harvesting honey. When a mangled stem is held near the bee hives, the sap tranquillizes the vindictive worker bees to such an extent, that they do not sting. Reportedly, the sap's tranquillizing effect is specific to rock bees.

**Adulterations and substitutes** Fruits and seeds of several other *Zingiberaceae* (e.g. *Alpinia*, *Elettaria* and *Globba* species) are used for similar purposes, and are sometimes mixed with *Amomum* seeds, but often they are less valuable.

**Description** Medium-sized to large aromatic herbs up to 400 cm tall, with creeping rhizome near the soil surface or above ground, occasionally elevated on short stilt roots, sometimes emitting long stolons, several-stemmed; spurious stems erect, swollen near base. Leaves distichous, numerous, usually lanceolate, finely veined, distinctly or indistinctly petiolate, lower ones sheathing with the sheath open on the side opposite the lamina, sheath apically produced into an erect, short or long ligule. Inflorescence either lateral immediately from the rhizome near the base of a leafy stem, or lateral on stolons arising from the rhizome, capitulate, usually globose to ovoid, sometimes elongate; peduncle often short and shallowly hidden under earth or litter, with biseriate, persistent scales; bracts often numerous, arranged spirally, persistent or becoming slimy and disappearing, each bract embracing one usually tubular-sheathing bracteole and one flower. Flowers bisexual, zygomorphic, 3-merous; calyx tubular, indented on one side, unequally 3-dentate on the other side; corolla with tube shorter to longer than calyx and lobes about as long as tube, superior lobe broadest and hollowed near apex, lateral lobes appressed against the labellum; labellum longer than corolla-lobes, very variable, lower part erect, tubular and connate with base of filament, upper part spreading, essentially 3-lobed and folded over the stamen, usually yellow or orange in the centre and with purplish markings, the sides often white; functional stamen 1, usually with dis-

tinct filament, anther elongate with connective developed into a large 3-lobed appendage, stamino-odes 2, small or absent; ovary inferior, 3-locular with axile placentation, style filiform with nectaries at base, stigma widened at apex, fimbriate. Fruit berry-like or capsular, indehiscent or dehiscent with 3 valves, aculeate, ribbed or smooth, many-seeded. Seeds angular, surrounded by an aril.

**Growth and development** When cultivated, *Amomum* usually forms a dense ground cover. Plants are shallow-rooted, with about 80% of the root mass in the upper 10 cm of the soil. Flowering and fruiting start 4-5 years after planting.

The flowers, which usually last less than one day, are pollinated by insects, whose numbers and frequency of visits seriously affect seed production. Shedding of young fruits can be serious but can be reduced by proper use of 2,4-D and colchicine at the time of flowering. The fruits ripen in 3-4 months.

**Other botanical information** *Amomum* belongs to the tribe *Alpinieae*, which also includes the small genus *Elettaria*, the large genus *Alpinia*, and the medium-sized genus *Riedelia*, which is centred in New Guinea.

*A. compactum* Soland. ex Maton is mainly cultivated for its seeds, which are used as a condiment, but it also has medicinal properties. *A. subulatum* Roxb. and *A. fenzlii* Kurz are used medicinally in India and Nepal. *A. tsao-ko* Crevost & Lem., whose fruits are used as a condiment, is also applied in traditional medicine in northern Vietnam and southern China. Its fruits are mixed with other medicinal plants to produce a reputed remedy for malaria. *A. cardamomum* L. is a synonym of *Elettaria cardamomum* (L.) Maton; several parts of this spice are used medicinally.

**Ecology** Like other members of the *Zingiberaceae*, *Amomum* is almost a characteristic element of the ground flora of primary rain forest, often in moist locations and in lower montane forest (in Vietnam even up to 2200 m altitude). *Amomum* is humid-thermophilous and moderately shade tolerant, but intolerant of drought. The climatic conditions most suitable for *A. villosum* are 1000-2400 mm annual rainfall, about 80% relative humidity and a mean annual temperature of 19-22°C. It prefers loose, moist and humus-rich soils with a pH 6-7.5. These requirements seem to be applicable to many other *Amomum* spp.

Most *Amomum*-underplanted forests are located in valleys or on hillsides with a moderate slope at 500-1400 m altitude. The fruit production of *A.*

*krervanh* in the lowland is inferior to that in the highlands.

**Propagation and planting** In southern China, *Amomum* is preferably propagated by cuttings from stolons of 1–2-year-old plants with 2 horizontal branches. These cuttings may produce 3–4 shoots in the first year and 20–30 stems/m<sup>2</sup> within 3 years, which are ready for flowering and fruiting. Plants propagated from cuttings flower approximately 1 year earlier than seedlings. Sometimes rhizomes for planting are collected from the natural forest.

In southern China, shade-tolerant *Amomum* spp. like *A. villosum* are underplanted either in natural forest or as part of an agroforestry system. In natural forest, planting of seedlings or cuttings is preceded by thinning to give 30–40% light intensity, with retention of shade trees. In agroforestry, *Amomum* spp. are planted in plantations of rubber (*Hevea brasiliensis* (Willd. ex A.L. Juss.) Muell.-Arg.), *Cinnamomum porrectum* (Roxb.) Kosterm., mango (*Mangifera indica* L.), *Albizia chinensis* (Osbeck) Merr., *Paraserianthes falcataria* (L.) Nielsen and *Cassia siamea* Lamk. Satisfactory results were obtained by planting *A. villosum* seedlings or cuttings at 1 m × 1 m in 4 years old rubber plantings, spaced at least 6 m × 6 m.

**Husbandry** Newly planted *Amomum* must be weeded frequently. In China, the application of fertilizer containing traces of manganese sulphate often effectively prevents leaf yellowing in *A. villosum* plantations.

**Diseases and pests** Leaf blight of *A. villosum* caused by *Glomerella cingulata* occurs in China. Other diseases reported to affect *A. villosum* include seedling anthracnose (caused by *Colletotrichum zingiberis*) and fruit rot (caused by *Rhizoctonia solani* and *Fusarium* spp.). The insect pest *Prodiocetes* sp. seriously affects cultivated *A. compactum* in West Java.

**Harvesting** Fruits are usually hand-picked when they start darkening.

**Yield** The mean annual yield of *A. villosum* planted in natural forest in southern China is about 375 kg/ha, with a maximum of 650 kg/ha. A 1-ha plot of *A. villosum* underplanted in rubber yielded about 190 kg/year of dried fruits.

**Handling after harvest** After harvest fruits need to be dried immediately. To avoid the quality reduction of the seed and the oil evaporating rapidly they should not be peeled until after drying. Dried seeds are kept in jute or nylon bags stored in cool, dry and well-ventilated conditions.

In Thailand, the volatile seed oil is obtained by water distillation.

**Genetic resources and breeding** Many *Amomum* species seem to have a limited geographical distribution, but lack of botanical knowledge might be at least partly responsible for this. Most *Amomum* species occur in primary rain forest, and large-scale destruction of this forest type undoubtedly puts them at risk of genetic erosion or even extinction.

There is great market demand for *Amomum* in Indo-China and China, and natural populations are dwindling rapidly. *A. villosum* had become so scarce in China by the mid 1980s that considerable areas have since been planted with this species. In Vietnam and elsewhere, the scope for cultivation is increasing, which helps to protect *Amomum* species from genetic erosion.

**Prospects** Although *Amomum* has a considerable reputation in Chinese medicine, it is not much used medicinally in South-East Asia. Little research has been done on the pharmacological characteristics, and more is needed to confirm the claimed medicinal properties. The antimalarial activity of fruit extracts of some *Amomum* species deserves further attention.

The successful cultivation of *Amomum* spp. in agroforestry systems in tropical China might also be applicable to South-East Asia, if there is a potential market for their products. The annual revenue from an agroforestry system of rubber and *Amomum* in tropical China is about 4 times that of a pure rubber plantation.

A thorough taxonomical revision of *Amomum* is badly needed. The species most interesting medicinally, now often cultivated in Indo-China and Thailand, could probably also be cultivated at higher elevations in Malesia.

**Literature** [1] Backer, C.A. & Bakhuizen van den Brink, R.C., 1968. Flora of Java. Vol. 3. Noordhoff, Groningen, the Netherlands. pp. 51–58. [2] Dao Lan Phuong, 1990. Chemical composition of Vietnamese *Amomum xanthioides* essential oil. Tap chi Duoc hoc [Journal of Pharmacy] 1: 17–19 (in Vietnamese). [3] Do Tat Loi, 1995. Medicinal plants and ingredients of Vietnam. Science and Technology Publishing House, Hanoi, Vietnam. pp. 516–523 (in Vietnamese). [4] Dutta, T.R., Ahmed, R., Abbas, S.R. & Rao, M.K.V., 1985. Plants used by Andaman aborigines in gathering rock-bee honey. Economic Botany 39(2): 130–138. [5] Ho Phamh, 1993. Zingiberaceae. Cay co Vietnam [Flora of Vietnam]. Vol. 3(1). Mekong Printing, United States. pp. 535–571. [6] Holttum, R.E.,

1950. The Zingiberaceae of the Malay Peninsula. Gardens' Bulletin Singapore 13(1): 1-249. |7| Kamchonwongpaisan, S. et al., 1995. An anti-malarial peroxide from *Amomum krervanh* Pierre. Tetrahedron Letters 36(11): 1821-1824. |8| Perry, L.M., 1980. Medicinal plants of East and Southeast Asia. Attributed properties and uses. The MIT Press, Cambridge, Massachusetts, Unites States & London, United Kingdom. pp. 437-438. |9| Smith, R.M., 1985. A review of Bornean Zingiberaceae: 1 (Alpineae p.p.). Notes from the Royal Botanic Garden Edinburgh 42(2): 261-314. |10| Zhou, S., 1993. Cultivation of *Amomum villosum* in tropical forests. Forest Ecology and Management 60(1-2): 157-162.

#### *Selection of species*

#### **Amomum aculeatum Roxb.**

Asiat. Res. 11: 344, t. 6 (1810).

**Synonyms** *Amomum ciliatum* Blume (1827), *Amomum flavum* Ridley (1909).

**Vernacular names** Indonesia: parahulu, prahulu (Sundanese), wola waliyan (Javanese). Papua New Guinea: apiyanga (Gulf), qulengapaie (Morobe).

**Distribution** Vietnam, Peninsular Malaysia (Penang), Sumatra, Java, Papua New Guinea; sometimes cultivated in Java.

**Uses** In Java drops of juice from the leaf-stalks are applied to the eyes of women after childbirth. In Papua New Guinea, leaves are chewed in combination with other plants and traditional salt to soothe headaches and backache. The sap is drunk to treat fever and influenza. The sourish and sweet fruits are edible.

**Observations** A large herb up to 400 cm tall, with stout and long underground rhizome and rather slender, up to 1.5 cm thick, leafy stems; leaves lanceolate, 10-60 cm × 2-9 cm; inflorescence with base in the ground, up to 10 cm long, dense and rounded, bracts about 3.5 cm × 1.5 cm, thin, brownish and soon disintegrating, bracteoles about 1 cm long, tubular at the base; flowers pedicelled, far exserted from bracts, corolla tube about as long as calyx, lobes pale flesh-coloured to orange, labellum orange-yellow with many small crimson spots and lines, forming a closed cup with dorsal petal, anther with considerably spreading crest; fruit 2-3.5 cm × 1.5-2 cm, dark purplish, covered with fleshy greenish spines. *A. aculeatum* occurs in Java in primary forest and teak forest up to 800 m altitude.

**Selected sources** 97, 115, 580, 609, 615, 1227, 1494.

#### **Amomum gracile Blume**

Enum. pl. Javae 1: 49 (1827).

**Vernacular names** Indonesia: ela-ela (Java).

**Distribution** Western and central Java.

**Uses** Fruits are chewed to treat nausea and indigestion.

**Observations** A medium-sized to fairly large herb up to 110(-200) cm tall, with poorly developed rhizome and slender, up to 0.6 cm thick, branched stolons and slender, up to 1 cm thick, leafy stems which are bulbous at base; leaves lanceolate, larger ones 25-35 cm × 3-4 cm; inflorescence short, up to 5.5 cm long including peduncle, few-flowered, bracts few, up to 1.5 cm long, outer 1-2 sterile, bracteoles about 1 cm long, tubular at the base; flowers more or less sessile, exserted from bracts, corolla tube about as long as calyx, labellum white with red tubercles at base, a red band in the centre and margins edged with red, stamen much shorter than labellum, anther with large 3-lobed appendage; fruit 1-1.5 cm long, 3-grooved, red, densely covered with minute straight prickles. *A. gracile* occurs below 100 m altitude, often in teak forest.

**Selected sources** 97, 580, 1494.

#### **Amomum hochreutineri Valetou**

Icones Bogoriensis 2(4): t. 195 (1906).

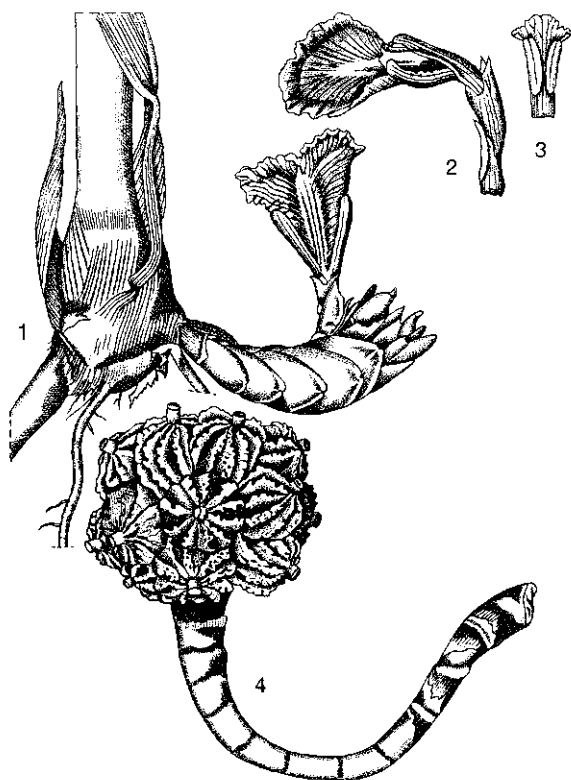
**Vernacular names** Indonesia: kihitir, cacabutan (Sundanese).

**Distribution** Very locally in western Java.

**Uses** Rhizomes and fruits are used as poultice to treat lumbago.

**Observations** A medium-sized herb up to 100 cm tall, with stout and long rhizome, whitish and up to 1 cm thick, and slender, red leafy stems; leaves lanceolate, larger ones 55-70 cm × 7.5-9 cm; inflorescence ascending, hardly raised above the ground, up to 30 cm long, few-flowered, bracts few, up to 3.5 cm long, outer 1-2 sterile, bracteoles about 2 cm long, hardly sheathing; flowers sessile, exserted from bracts, corolla tube about as long as calyx, labellum erect, much exserted, white, greenish-yellow in the centre and red-streaked, stamen hardly shorter than corolla, anther with 3-lobed appendage; fruit about 1.5 cm long, 9-12-ribbed, ribs with curled-crenate margins, white with red stripes. *A. hochreutineri* occurs in primary lower montane forest at 1000-1400 m altitude.

**Selected sources** 97, 580.



*Amomum hochreutineri* Valetton - 1, stem base with inflorescence; 2, flower; 3, stamen; 4, infructescence.

***Amomum krervanh* Pierre ex Gagnep.**

Bull. Soc. Bot. Fr. 53: 138 (1906).

**Vernacular names** Round Siam cardamom, camphor seed (En). Cambodia: kreko krervanh, karvanh, krewanh. Thailand: krawaan khao (central). Vietnam: b[aj]ch d[aa]j u kh[aa]s u, sa nh[aa]n nam vang.

**Distribution** Laos, Cambodia, Vietnam, southern China and Thailand; also cultivated there.

**Uses** Fruits are used to treat indigestion, liver and uterus diseases, rheumatism, diarrhoea and asthenic after dysentery, and as febrifuge, antiemetic and antitoxic of alcohol. In Indo-China, they are traded as a condiment and spice.

**Observations** A large herb up to 300 cm tall, with superficial rhizome; leaves lanceolate, up to 60 cm × 12 cm; inflorescence cylindrical to conical, up to 11 cm long, bracts about 4 cm long, bractlets tubular at the base; flowers with corolla tube about as long as calyx, labellum elliptical, rounded at apex, white with a yellow patch in the centre and yellow at margins, anther with a 3-lobed ap-

pendage; fruit about 1.5 cm in diameter, slightly ribbed. *A. krervanh* occurs in Cambodia and Vietnam in mountainous regions, and could well be cultivated in mountainous areas in Malasia. It is possibly conspecific with *A. testaceum* Ridley from Thailand, Peninsular Malaysia and Borneo.

**Selected sources** 455, 699, 1035, 1126, 1227.

***Amomum ligulatum* R.M. Smith**

Notes Roy. Bot. Gard. Edinb. 42(2): 298 (1985).

**Vernacular names** Indonesia: ubut bele sa'ai (Kenyah Dayak, East Kalimantan).

**Distribution** Borneo (Sabah, East Kalimantan).

**Uses** Locally in East Kalimantan, the tender inner pith of leafy stems is roasted or boiled and eaten to treat stomach-ache and diarrhoea.

**Observations** A large herb up to 250 cm tall; leaves narrowly lanceolate, 25–80 cm × 3–5 cm; inflorescence narrowly ellipsoid, about 12 cm long, on peduncle up to 10 cm long, bracts about 2 cm long; flowers yellowish-orange, with pedicels up to 1 cm long, corolla tube about as long as calyx, labellum broad, orange with darker spot, anther with an undulate crest; fruit ovoid, about 2 cm long, pale yellowish-brown. *A. ligulatum* occurs in forest up to 1200 m altitude.

**Selected sources** 829, 1379.

***Amomum longiligulare* T.L. Wu**

Fl. Hainan. 4: 533 (1977).

**Vernacular names** Vietnam: sa nh[aa]n, m[ef] tr[as] l[af].

**Distribution** Hainan, northern and central Vietnam; also cultivated in Vietnam.

**Uses** Seeds are used to treat indigestion, diarrhoea, vomiting and toothache, whereas the roots are applied against rheumatism.

**Observations** A large herb up to 250 cm tall, with rhizome creeping on the ground; leaves narrowly elliptical, 20–30 cm × 5–6 cm; inflorescence small, on a 2–4 cm long peduncle, few-flowered, bracts few; flowers subsessile, corolla tube about as long as calyx, labellum rounded and concave, with yellow margins, anther with clearly 3-lobed appendage; fruit 1.5–2 cm in diameter, brownish-purple, short-thorned. In Vietnam, *A. longiligulare* occurs in mountainous areas.

**Selected sources** 1037.

***Amomum squarrosus* Ridley**

Journ. Roy. As. Soc. Straits Br. 57: 104 (1910).

**Vernacular names** Malaysia: puar tadah embun (Peninsular).

**Distribution** Peninsular Malaysia.

**Uses** Flowers are made into a poultice which is applied to the head to treat giddiness.

**Observations** A large herb up to 400 cm tall, with rhizome sometimes supported by stilt roots, leafy shoots close together; leaves narrowly lanceolate, up to 55 cm × 6 cm; inflorescence oblong, up to 10 cm long, on peduncle up to 15 cm long, bracts about 2 cm long, almost persistent, bracteoles up to 2 cm long, funnel-shaped; flowers with corolla tube shorter than calyx, labellum distinctly 3-lobed, white with a yellow median band flanked by red lines, anther with a 3-lobed appendage; fruit about 1.3 cm in diameter, smooth or slightly ribbed towards the apex. *A. squarrosus* occurs locally in the lowland, up to 300 m altitude.

**Selected sources** 202, 615, 1227.

### ***Amomum stenocarpum* Veleton**

Bull. Jard. Bot., ser. 3, vol. 2: 354 (1920).

**Vernacular names** Indonesia: kaol haol (Simeuluë).

**Distribution** Simeuluë (Indonesia).

**Uses** The stem juice is used to treat cough. The sourish fruits are edible.

**Observations** A large herb up to 300 cm tall; leaves lanceolate, larger ones up to 90 cm × 13 cm; inflorescence elongate, with distinct peduncle, bracts up to 4.5 cm long, persistent; flowers unknown; fruit narrowly fusiform, about 2.5 cm long, greenish. *A. stenocarpum* is found in secondary forest.

**Selected sources** 580, 1497.

### ***Amomum uliginosum* J.G. König ex Retz.**

Observ. bot. 3: 56 (1783).

**Vernacular names** Malaysia: puar hijau, puar gajah, tepus merah (Peninsular). Thailand: krawaan paa (Pattani).

**Distribution** Thailand and Peninsular Malaysia.

**Uses** Seeds are used medicinally in Thailand, whereas rhizomes are possibly used as a stomachic. Fruits are edible, and leaves are sometimes used for making temporary shelters.

**Observations** A large herb up to 300 cm tall, with subterranean, long and much branched rhizome, leafy shoots widely apart; leaves narrowly lanceolate, up to 50 cm × 7 cm, with caudate apex; inflorescence small and globose, up to 5 cm long, on peduncle up to 10 cm long, bracts 2.5–3 cm long, bracteoles about 2 cm long, tubular at the base; flowers with corolla tube as long as or slight-

ly longer than calyx, labellum ovate and strongly concave, white, sometimes with 2 dark red spots at base and with a dark crimson stripe on each side, anther with a 3-lobed appendage having spreading side lobes; fruit up to 2 cm long, covered by slender and soft red spines. *A. uliginosum* is locally abundant in lowland forest and on river banks, up to 1000 m altitude.

**Selected sources** 202, 615, 1227.

### ***Amomum villosum* Lour.**

Fl. cochinch.: 4 (1790).

**Synonyms** *Amomum echinosphaera* K. Schumann ex Gagnep. (1902).

**Vernacular names** Thailand: reo dong (Trat). Vietnam: sa nh[aa]n, m[ef] tr[es] b[af], d[uw] [ow]ng xu[aa]n sa.

**Distribution** Southern China, Vietnam and Thailand; also cultivated there.

**Uses** In Chinese and Vietnamese traditional medicine the fruits are used to treat indigestion, diarrhoea, flatulence, toothache, and as febrifuge and antiseptic. They are also used as spice and condiment.

**Observations** A large herb up to 300 cm tall, with thick rhizome; leaves narrowly ovate-lanceolate, up to 40 cm × 9 cm; inflorescence ascending, on peduncle up to 8 cm long, with few flowers scattered from base to top, bracts membranous, bracteoles tubular at the base; flowers with corolla tube slightly longer than calyx, labellum spoon-shaped to almost circular and concave with emarginate apex, white with prominent middle vein, anther with a 3-lobed appendage having ear-shaped side lobes; fruit up to 2 cm long, reddish-brown and covered by small flexuous spines, difficult to break into 3 fragments. *A. villosum* occurs in forest, often in mountainous areas, and usually on wet soils.

**Selected sources** 455, 1035, 1126, 1658.

### ***Amomum xanthioides* Wallich ex Baker**

Hook.f., Fl. Brit. India 6: 239 (1892).

**Synonyms** *Amomum villosum* Lour. var. *xanthioides* (Wallich ex Baker) T.L. Wu & S.J. Chen (1978).

**Vernacular names** Bastard cardamom, tavoy cardamom (En). Thailand: phaa laa (Shan, Chiang Mai), maak ee (Chiang Mai), neak naeng (north-eastern). Vietnam: sa nh[aa]n, s[us]c sa m[aa]jt.

**Distribution** India, Laos, Cambodia, Vietnam, southern China and Thailand.

**Uses** In Chinese and Vietnamese traditional



**Andrographis paniculata (Burm.f.)  
Wallich ex Nees**

Wallich, Pl. asiat. rar. 3: 116 (1832).

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**Synonym** *Andrographis subspathulata* C.B. Clarke (1884).

**Vernacular names** Creat, green chireta (En). Roi des amers (Fr). Indonesia: ki oray (Sundanese), sambilata (Javanese), ampadu (Padang). Philippines: aluy, lekha (Tagalog), sinta (Bikol). Thailand: fa thalaai (Bangkok), khee-pang-hee (Chinese), yaa kannguu (Songkhla). Vietnam: c[oo]ng c[oo]ng, kh[oor] di[ee]p, xuy[ee]n t[laa]m li[ee]n.

**Origin and geographic distribution** *A. paniculata* is probably native to India, but has been introduced and cultivated as a medicinal plant in many parts of Asia including Indo-China, China, Thailand, Peninsular Malaysia, Indonesia, the Philippines and Australia. It is now widely naturalized in most of these regions. It has also been introduced, possibly for its ornamental value, in the West Indies and Central America.

**Uses** The roots and leaves of *A. paniculata* have a well-known application in traditional medicine in India, various parts of South-East Asia, Central America and the Caribbean. It used to be considered an effective remedy against snake bites; in India, it is locally still used for this. It has also been reported as useful to treat insect bites and, in combination with *Orthosiphon aristatus* (Blume) Miq., as a remedy for diabetes. An infusion or sap from the crushed leaves has been recommended for the treatment of fever, as a tonic, and for itching skin eruptions. A decoction of the leaves or roots is used against stomach-ache, dysentery, typhus, cholera, influenza and bronchitis, as a vermifuge, and is considered a diuretic. Another use is as a poultice on swollen legs or feet, vitiligo and piles. Pills or infusions are also recommended to treat female disorders, dyspepsia, hypertension, rheumatism, gonorrhoea, amenorrhoea, torpid liver and jaundice. Furthermore, *A. paniculata* is considered to be anti-inflammatory and immunosuppressive, but reports on antibacterial activity are contradictory.

**Properties** Phytochemical studies on *A. paniculata* have resulted in the isolation of a number of diterpenes from the aerial parts, of which the most important are:

- Diterpenoids of the ent-labdane type, e.g. andrographolide, 14-epi-andrographolide, isoandro-



*Amomum villosum* Lour. - 1, plant base with infructescence; 2, part of leafy stem; 3, flower.

medicine, fruits are used to treat indigestion, diarrhoea, flatulence, gastralgia, toothache, and as febrifuge and antiseptic. They are also used as a spice and condiment. Thai traditional doctors also apply the fruit to treat cough, against asthma and as an anti-emetic.

**Observations** A large herb up to 300 cm tall, with thick rhizome; leaves narrowly ovate-lanceolate, up to 40 cm × 9 cm; inflorescence ascending, on peduncle up to 8 cm long, with few flowers, bracts membraneous, bracteoles tubular at the base; flowers with corolla tube slightly longer than calyx, labellum spoon-shaped to almost circular and concave with emarginate apex, white with prominent middle vein, anther with a 3-lobed appendage having ear-shaped side lobes; fruit 1.5-2 cm long, yellowish-green and covered by small spines, difficult to break into 3 fragments. *A. xanthioides* occurs in forest, often in mountainous areas, and usually on wet soils.

**Selected sources** 313, 1035, 1126.

Nguyen Quoc Binh

grapholide, 14-deoxyandrographolide, 14-deoxy-12-methoxyandrographolide, 12-epi-14-deoxy-12-methoxyandrographolide, 14-deoxy-12-hydroxyandrographolide, 14-deoxy-11-hydroxyandrographolide, 14-deoxy-11,12-dihydroandrographolide, ent-14 $\beta$ -hydroxy-8(17),12-labdadien-15,16-olide-3 $\beta$ ,19-oxide.

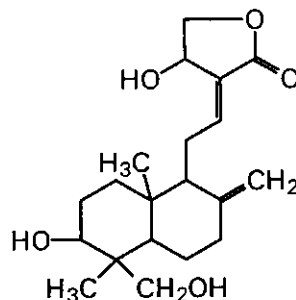
- Diterpene glucosides, e.g. andrographiside, deoxyandrographiside, 14-deoxy-11,12-dihydroandrographiside, neoandrographolide, 6'-acetyl-neoandrographolide.

- Bis-andrographolides A, B, C, and D.

The presence of flavones in the root has also been reported. Extracts and purified diterpenes and flavonoids have been investigated for a multitude of pharmacological effects.

In a placebo-controlled double-blind study, performed as a pilot trial in 50 patients, the effect of a standardized *A. paniculata* extract was evaluated in the initial treatment of the common cold and sinusitis. Patients were advised to take 4 tablets (containing 85 mg of the extract, or placebo) 3 times daily. Furthermore, they were given a self-monitoring form, and were assessed at the clinic after 5 days. In the *A. paniculata* group the subjective symptoms and symptom duration were both significantly reduced. In another double-blind study, patients with common cold were treated with *A. paniculata* extract (dose 1200 mg powdered leaves/day) or placebo. A significant reduction in clinical symptoms was observed in the treated group on day 4 of administration. It was concluded that powdered *A. paniculata* leaves have the capacity to significantly shorten the duration of common colds. When the efficacy of a high dose of powdered *A. paniculata* leaves (6 g/day) was compared with either a low dose of powdered leaves (3 g/day) or paracetamol (acetaminophen, 3 g/day) in a randomized double blind design in patients with pharyngotonsillitis, the paracetamol and the high dose of the powdered leaves produced significantly better effects than the low dose of *A. paniculata* on day 3, in terms of relief of fever and sore throat. On day 7 the clinical effects were no longer different. Furthermore, minimal, self-limiting side effects were found in about 20% of each of the groups.

The anti-inflammatory effect (carrageenin-induced oedema model) of an orally administered infusion of leaves at 51.4 mg/100 g bodyweight has been tested in mice; it was similar to 10 mg phenylbutazon/100 g. Andrographolide at 100 or 300 mg/kg also shows anti-inflammatory activity and significantly inhibits hind paw oedema in-



andrographolide

duced by carrageenin, kaolin and nystatin. Furthermore, an ethanol extract stimulates both antigen-specific and nonspecific immune responses in mice more than the purified andrographolides.

An ethanolic extract of *A. paniculata* administered orally to rats with yeast-induced fever showed significant antipyretic activity. Andrographolide at 100 or 300 mg/kg also exhibits significant antipyretic properties in rats. Evaluated for its analgesic effects in mice, a dose of 8 mg/kg of a 10% infusion of the herb, applied intraperitoneally, is comparable with 48 mg/kg phenylbutazon. At a per oral dose of 300 mg/kg, purified andrographolide shows significant analgesic activity in the acetic acid-induced writhing test in mice and in Randall Selitt's test in rats.

The crude water extract of *A. paniculata* as well as the semi-purified n-butanol and aqueous fractions produced a significant fall in mean arterial blood pressure in anaesthetized Sprague-Dawley rats; the ED<sub>50</sub> values were respectively 11.4 mg/kg, 5.0 mg/kg and 8.6 mg/kg. The aqueous extract exhibits a dose-dependent hypotensive activity on the systolic blood pressure in spontaneously hypertensive rats, when chronic intraperitoneal infusions are administered by osmotic pumps. Mechanistic studies indicate that the effect might be due to a reduction of circulating angiotensin-converting enzyme levels, as well as to a reduction of numbers of free radicals in the kidneys. Furthermore, a 10% infusion of the herb applied intravenously to rabbits at 1 ml/kg bodyweight reduced the blood pressure by 6–10 mm Hg in 10–20 seconds.

An *A. paniculata* extract was found to significantly alleviate atherosclerotic iliac artery stenosis induced by both de-endothelialization and high cholesterol diet in rabbits. It may therefore play an important role in preventing restenosis after coronary angioplasty, which normally can be 30–40%. Observations in dogs with experimentally induced

myocardium infarction indicate that an aqueous extract may also limit the expansion of localized anaemia of the myocardium and exert a marked protective effect on reversibly ischemic myocardium. In these experiments, antithrombotic effects were also observed in the animals, e.g. inhibition of thromboxane (TBX<sub>2</sub>) synthesis and of platelet aggregation. It is believed that these effects might be at least partially due to flavones present in the extract.

When administered intraperitoneally (100 mg/kg) in mice, andrographiside and neoandrographolide have a significant protective effect on hepatotoxicity induced by carbontetrachloride or tert-butylhydroperoxide. Andrographolide is also hepatoprotective against galactosamine and paracetamol induced liver damage in rats. An extract of *A. paniculata* showed anti-diarrhoeal activity in rabbit and guinea-pig ileal loop models. The diterpenes andrographolide and neoandrographolide isolated from the alcoholic extract showed potent antisecretory activity against *Escherichia coli* enterotoxin induced secretions. Andrographolide at 100 or 300 mg/kg is also known to exhibit significant anti-ulcer properties in rats. Furthermore, apigenin 7,4'-di-o-methyl-ether (a flavonoid) shows anti-ulcer effects in experimentally induced ulcers in guinea-pigs and rats. It is suggested that this effect is due to the antisecretory activity and protective effect on the gastric mucosa.

Other pharmacological effects reported in literature include:

- The water extract administered at 10 mg/kg can significantly prevent glucose-induced hyperglycaemia in rabbits, but has no effect on adrenaline-induced hyperglycaemia. Furthermore, a hypoglycaemic effect in rabbits was observed when a 20% infusion was administered orally at doses of 12.5 and 37.5 ml/kg.
- The chloroform extract administered orally in rats at a dose of 8 mg/kg has a diuretic effect similar to 25 mg furosemide/kg.
- Platelet aggregation in humans was significantly reduced by *A. paniculata* extracts.
- Oral administration of 20 mg of the dry leaf powder for 60 days has an antifertility effect (antispermatogenic and/or anti-androgenic) in male albino rats.
- The alcoholic extract of the rhizomes of *A. paniculata* exhibits good in vitro anthelmintic activity against *Ascaris lumbricoides*.
- Andrographolide exhibits a strong choleric effect when administered intraperitoneally to albino rats.

- Neoandrographolide isolated from *A. paniculata* exhibits significant antimalarial activity against *Plasmodium berghei* NK 65 in the mouse *Mastomys natalensis*.

- Dehydroandrographolide succinic acid monoester, derived from andrographolide, has been found to inhibit the human immunodeficiency virus (HIV) in vitro.

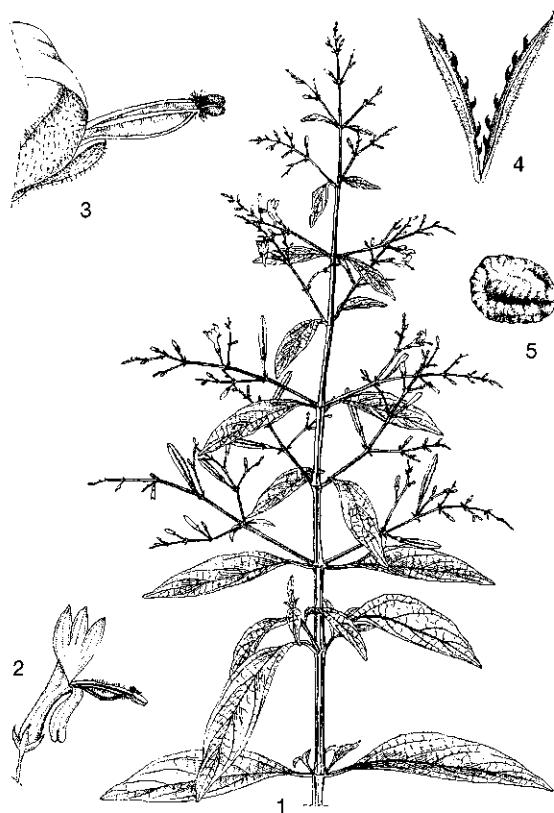
- Several of the ent-labdane type diterpenoids (e.g. andrographolide, 14-epi-andrographolide, isoandrographolide, 12-epi-14-deoxy-12-methoxy-andrographolide) show potent induction of cell differentiation towards M1 cells. In general, the activity of the dimers (bis-andrographolides A, B, C) is even more potent in this model system; the glucosides are only weakly active.

It has also been found that the ether extract of *A. paniculata* leaves has a fairly high anti-alkylating effect against ethyl-methane sulphonate. In general, anti-alkylating substances are associated with anti-carcinogenicity.

A leaf infusion administered intraperitoneally in mice, has an LD<sub>50</sub> at 71.1 mg/10 g body weight (acute toxicity). At a concentration of 1 mg/kg it lowers the body temperature at least 2°C. In guinea-pigs, a leaf infusion of 5%, 10% and 15% at a dose of 8 ml/kg lowers the body temperature by 0.9–1.1°C. The possible testicular toxicity of a dried extract of *A. paniculata* was investigated in male Sprague Dawley rats. No testicular toxicity was found with the treatments of 20, 200 and 1000 mg/kg during 60 days as evaluated by reproductive organ weight, testicular histology, ultrastructural analysis of leydig cells and testosterone levels after 60 days of treatment. It was concluded that *A. paniculata* extract did not produce sub-chronic testicular toxicity in male rats.

Finally, *A. paniculata* extract is reported to have antifeedant and anti-oviposition activity against a number of crop pests like *Callosobrunchus chinensis*, *Darcus dorsalis*, *Nephotettix cincticeps*, *Plutella xylostella*, *Sitophilus oryzae* and *Spodoptera litura*.

**Description** A perennial herb 30–100 cm tall; stems distinctly 4-angular, glabrous apart from a few hairs at the nodes. Leaves opposite, simple, narrowly ovate to lanceolate, 5–10 cm × 1.2–2.5 cm, base long attenuate, apex long acuminate, margin entire, glabrous but often gland-dotted; petiole short, up to 6 mm long, connected with the opposite one by transverse ridges. Flowers in lax, axillary and terminal racemes or panicles combined into a pyramidal inflorescence, with 2 small bracteoles at base of the 1–7 mm long pedicel, bi-



*Andrographis paniculata* (Burm. f.) Wallich ex Nees - 1, flowering stem; 2, flower; 3, flower with detail of style and stamen arrangement; 4, dehiscent fruit; 5, seed.

sexual, zygomorphic; calyx segments 5, jointed at base, with glandular and aglandular hairs; corolla bilabiate, white or rose with purple markings on the upper lip, tube 5-6 mm long, slightly enlarged below the limb, lower lip 4-6 mm long, oblong, 2-toothed at the apex, upper lip deeply 3-lobed, as long as the lower; stamens 2, inserted at the corolla tube apex and exserted, filaments hairy, anthers inserted at equal level, basally connate, bearded at base, deep purple to black; ovary superior, 2-locular with 3-7 ovules in each cell, style curved upwards, stigma entire. Fruit an erect, loculicidal, narrowly ellipsoidal, glandular hairy capsule, 14-20 mm × 3-3.5 mm, many-seeded. Seeds held up on well-developed hooks (retinaculæ), almost rectangular, rugosely furrowed, with 2 deep furrows.

**Growth and development** In India, leaves of *A. paniculata* start to redden in October and are shed in December, after which the plants dry from

the top down, leaving only a small portion of the stem green. New flushes emerge towards the end of the dry season. There are two distinct flowering periods: in India October and March-May, although plants growing under shaded conditions flower from October through to May. In Java, flowers and fruits of *A. paniculata* have been observed throughout the year. In northern Australia, flowering and fruiting specimens have been collected from November to June.

**Other botanical information** *Andrographis* comprises about 18 species occurring originally from India to China and western Malesia. It belongs to the small tribe *Andrographideae* of the subfamily *Acanthoideae*. This tribe is characterized by its articulated shoots and epidermis with cystoliths in combination with a 2-lipped corolla with ascending lobes and many ovules per locule. The pollen shows a unique structure as well. Within various populations of *A. paniculata* in India and Bangladesh 9 different cytotypes have been identified. These proved to be related to the environment, particularly to soil conditions.

**Ecology** *A. paniculata* is locally common and often gregarious. It exhibits weedy characteristics and occurs from sea-level up to 1600 m altitude in village groves, roadsides, waste places, open sandy locations and fields, but also in monsoon and teak forest receiving only 10-20% of full light.

**Propagation and planting** Propagation of *A. paniculata* by seed is possible. Seed should be soaked during 24 hours and dried before being sown. Germination starts after 1 week and the mean germination rate is about 80%. Cuttings consisting of 3 nodes taken from the upper third of 1-year-old plants have given the best results in vegetative propagation, with 80-90% rooting. In India, seed is sown in May-June, and seedlings are transplanted at 60 cm × 30 cm.

**In vitro production of active compounds** A distinct cell line from callus culture of *A. paniculata* proved capable of synthesizing andrographolide in greater quantity than in the intact plant. Plantlets differentiated from this line were also high-yielding.

**Husbandry** In shading experiments with *A. paniculata* the optimal proved to be 20% shade with average dry-matter production of 13.2 g per 5-month-old plant. In India, irrigation may be necessary during dry periods.

**Diseases and pests** *Sclerotium* sp. occasionally causes wilt disease in *A. paniculata* during the rainy season. Eugenol at 200 ppm or clove powder containing 0.2 % eugenol could inhibit the growth

and development of *Sclerotium* myceliae; its establishment is inhibited by eugenol at a concentration of 300 ppm or by clove powder containing 0.3% eugenol.

**Harvesting** Leaves of *A. paniculata* should be harvested when the inflorescence axis starts to grow, because the maximum accumulation of andrographolide is at that stage. In India, harvesting is in February-March, i.e. 9 months after planting. In Vietnam, where the crop is grown as an annual, the leaves are hand picked before flowering and roots are harvested when leaves start discolouring or wilting.

**Yield** Yields of 1–1.5 kg fresh weight/plant are obtained from 7-month-old *A. paniculata*.

**Handling after harvest** In general, harvested plant parts of *A. paniculata* are used fresh and consumed within a few days after collection. However, leaves and roots should be washed and dried in the sun or artificially before storage.

**Genetic resources and breeding** *A. paniculata* is relatively widespread and has a tendency to naturalize in areas where it has been introduced. Locally it is cultivated both for its ornamental and medicinal value. Therefore, the risk of genetic erosion seems rather limited. Selection should be directed to plant material with a higher content of medicinally important constituents. Preliminary results from callus culture techniques show considerable potential for improvement.

**Prospects** Extracts of *A. paniculata* as well as isolated compounds show a broad range of interesting pharmacological effects. In particular, the treatments for the common cold, the anti-inflammatory effects and the prevention of restenosis after coronary angioplasty deserve further attention. Furthermore, *A. paniculata* preparations may remain of local importance as a general tonic.

**Literature** |1| Barker, R.M., 1986. A taxonomic revision of Australian Acanthaceae. *Journal of the Adelaide Botanic Garden* 9: 130–131. |2| Hancke, J., Burgos, R., Caceres, D. & Wikham, G., 1995. A double-blind study with a new monodrug Kan Jang: decrease of symptoms in the recovery from common colds. *Phytotherapy Research* 9(8): 559–562. |3| Kapil, A., Koul, I.B., Banerjee, S.K. & Gupta, B.D., 1993. Antihepatotoxic effects of major diterpenoid constituents of *Andrographis paniculata*. *Biochemical Pharmacology* 46(1): 182–185. |4| Madav, S., Tandan, S.K., Lal, J. & Tripathi, H.C., 1996. Anti-inflammatory activity of andrographolide. *Fitoterapia* 67(5): 452–458. |5| Matsuda, T., Kuroyanagi, M., Sugiyama, S., Umehara,

K., Ueno, A. & Nishi, K., 1994. Cell differentiation-inducing diterpenes from *Andrographis paniculata* Nees. *Chemical and Pharmaceutical Bulletin (Tokyo)* 42(6): 1216–1225. |6| Nuratmi, B., Adjirni & Paramita, D.I., 1996. Beberapa penelitian farmakologi sambiloto (*Andrographis paniculata* Nees) (kumpulan abstrak) [Some pharmacological studies on sambiloto (*Andrographis paniculata* Nees) (collection of abstracts)]. *Warta Tumbuhan Obat Indonesia* 3(1): 23–24. |7| Roy Shamal, K., 1991. Andrographolide content in leaves and stems of *Andrographis paniculata* at different phases of growth. *Bangladesh Journal of Scientific Research* 9(1): 13–17. |8| Wang, D.W. & Zhao, H.Y., 1994. Prevention of atherosclerotic arterial stenosis and restenosis after angioplasty with *Andrographis paniculata* Nees and fish oil. *Experimental studies of effects and mechanisms. Chinese Medical Journal* 107(6): 464–470. |9| Zhang, C.Y. & Tan, B.K., 1997. Mechanisms of cardiovascular activity of *Andrographis paniculata* in the anaesthetized rat. *Journal of Ethnopharmacology* 56(2): 97–101. |10| Zhao, H.Y. & Fang, W.Y., 1990. Protective effects of *Andrographis paniculata* Nees on post-infarction myocardium in experimental dogs. *Journal of the Tongji Medical University* 10(4): 212–217.

**Other selected sources** 37, 45, 97, 114, 142, 173, 183, 200, 202, 236, 287, 350, 363, 527, 529, 531, 541, 543, 544, 550, 580, 695, 741, 882, 921, 926, 952, 979, 985, 1035, 1126, 1128, 1141, 1166, 1227, 1258, 1259, 1282, 1287, 1360, 1410, 1453, 1477, 1508, 1514, 1529, 1531, 1537, 1544, 1649, 1650, 1653.

Sri Sugati Sjamsuhidajat, Sudjaswadi Wiryowidagdo, Rini Sasanti & Wien Winarno

### **Angelica acutiloba (Siebold & Zucc.) Kitagawa**

*Bot. Mag., Tokyo* 51: 658 (1937).

UMBELLIFERAE

2n = 22

**Synonym** *Ligusticum acutilobum* Siebold & Zucc. (1845).

**Vernacular names** Thailand: tang kui.

**Origin and geographic distribution** *Angelica* comprises about 110 species, widely distributed in the temperate regions of the northern hemisphere. *A. acutiloba* is indigenous in Japan and was introduced for cultivation in mountainous regions in West Java in the 1970s.

**Uses** The root of *A. acutiloba* ('*Angelica Radix*')

is traditionally used in Japan and China as a general tonic and prescribed in the treatment of dysmenorrhoea, phthisis and haemorrhage. Furthermore, it is used as an emmenagogue, a remedy for anaemia, to alleviate pain during parturition and, in Indo-China, as a carminative and galactagogue. Two other *Angelica* species with a long history of medicinal use have been successfully introduced in Vietnam: *A. dahurica* (Fisch. ex Hoffm.) Benth. & Hook.f. ex Franchet & Savat. and *A. polymorpha* Maxim. (synonym: *A. sinensis* (Oliv.) Diels). In Western Europe various parts of *A. archangelica* L. have been used since ancient times as a vegetable and medicinal plant (to treat dyspeptic syndromes and as appetite stimulant).

**Production and international trade** Although there is a long tradition of trade of *A. acutiloba* from Japan to China and Indo-China, no trade figures are available.

**Properties** Extensive chemical studies on *A. acutiloba* have revealed a wide array of compounds including polysaccharides, lactones and alkynes, some of which are medicinally active. The roots of *A. acutiloba* contain two anticholinergic compounds: ligustilide and butylidenephthalide. They also contain seven analgesic compounds: falcarinol, falcarindiol and falcarinolone (polyacetylenes), and choline, scopoletin, umbelliferone and vanillic acid. The latter compounds inhibit writhing in mice induced by acetic acid. The three polyacetylenes were the most active in the writhing test. Falcarindiol and choline also showed anti-nociceptive activities in the retrograde injection test of bradykinin into a carotid artery on rats. The polysaccharide fraction of a hot water extract of the roots of *A. acutiloba* showed a mitogenic activity on B-lymphocytes, polyclonal B cell activator activity, antitumour activity against Ehrlich ascites cells, interferon-inducing activity and anticomplementary activity. These biological activities are caused by different water-soluble fractions. The action of *Angelica* immunostimulating polysaccharide (AIP) fraction on murine lymphocytes participating in antibody responses has been investigated. When AIP fraction was injected concomitantly into mice immunized with antigens, immunoglobulins G and M (IgG, IgM), antibody responses against sheep erythrocytes increased significantly, but IgM response against specific T-independent antigens did not augment. Furthermore, murine B lymphocytes were polyclonally activated in vitro and in vivo by AIP fraction to differentiate into antibody-forming cells as functionally matured cells. The differenti-

ation of B lymphocytes to an intermediate stage capable of responding to helper T-lymphocytes was also stimulated by administering AIP fraction to CDF1 and C3H/HeJ mice. Stimulation of T-lymphocytes was also found.

An anticomplementary active arabinogalactan IIb-1 (AGIIb-1) and an inactive arabinogalactan IIb-2 (AGIIb-2) were isolated from an extract of the root of *A. acutiloba*. AGIIb-1 mainly consisted of arabinose, galactose, rhamnose and galacturonic acid in a molar ratio of 2.2:1.0:0.3:0.4. AGIIb-1 was found to form molecular self aggregation, caused by hydrogen bonding and ionic interaction. However, it was independent of  $Ca^{2+}$  ions. The anticomplementary activity of AGIIb-1 seemed to be dependent upon the degree of molecular aggregation; the aggregate in water showed the greatest activity. Further separations of AGIIb-1 revealed that it consisted of one neutral unit (N-I), one neutral arabinan unit (N-II) and two acidic arabinogalactan (A-I and A-II) units. N-I showed the most potent anticomplementary activity. AGIIb-1, A-I, and A-II had similar moderate activities, but N-II had weak activity. Digestion products prepared from the latter fractions by treatment with purified enzymes yielded several polysaccharides that also showed (modified) anticomplementary activity. In general, the anticomplementary activity of AGIIb-1 was expressed mainly through the classical pathway, whereas some modified polysaccharides had markedly increased activity through the alternative pathway.

The crude polysaccharide fraction (AR-1), prepared by ethanol precipitation and dialysis of the hot water extract from the root of *A. acutiloba* showed a potent antitumour activity against ascitic forms of sarcoma-180, IMC carcinoma and Meth A fibrosarcoma, as well as the solid form of MM-46 tumour. AR-1 was further fractionated into a pectic (AR-2) and a arabinogalactan (AR-4) fraction. An antitumour active component from AR-4 was identified as polysaccharide AR-4E-2, which consisted of arabinose, galactose and rhamnose in the molar ratio of 3.3:1.0:0.7. It also contains a rhamno-galacturonan moiety in which 2,4-di-substituted rhamnose residues are attached to 4-substituted galacturonic acid through position 2 of rhamnose. Furthermore, four pectic polysaccharides with anticomplementary activity were isolated from fraction AR-2. These compounds have been named AR-2 Ila-IIId.

A root extract of *A. acutiloba* showed potent inhibitory effects on 12-O-tetradecanoylphorbol-13-acetate stimulated  $^{32}P$  incorporation into phospho-

lipids of cultured cells. The hot water extract of roots also exhibited antipyretic, central nervous system depressant, cardiac depressant, and hypotensive, antispasmodic and radioprotective activities.

**Description** A medium-sized perennial herb up to 70(–100) cm tall, tuberous root short with a few thickened secondary roots; stems glabrous, striate, solid. Leaves alternate, long-petiolate in lower part of stem, 1–2 ternately pinnate, deltoid in outline, 10–25 cm long, glabrous, leaf sheaths present, stipules absent; leaflets trifid, 5–10 cm × 1–2.5 cm, cuneate, truncate or rounded at base, ultimate segments lanceolate, long-acuminate, margin dentate, reticulate venation prominent. Inflorescence a compound many-flowered umbel, papillate, rays 30–40, 3–8 cm long, secondary rays (pedicels) 0.7–1.8 cm long; involucre consisting of a few filiform-linear bracts. Flowers with obsolete calyx, petals unobtrusive white, ovary inferior, styles 2. Fruit consisting of two 1-seeds mericarps, oblong, 4–5 mm long, slightly compressed, nar-

rowed towards the base, mericarps with slender ribs, lateral ribs slightly winged, vittae 3–4 in the intervals, 4 on the commissure.

**Other botanical information** Several subspecies and varieties have been distinguished within *A. acutiloba*. This variation can be partly attributed to geographical origin, but also the long tradition of cultivation has resulted in selections that can be morphologically distinguished. The differences are to some extent supported by karyological research.

**Ecology** *A. acutiloba* occurs naturally at higher elevations in mountainous regions. It is cultivated under similar conditions in Java.

**Propagation and planting** *A. acutiloba* can be propagated by seed as well as by somatic embryogenesis from hypocotyl and cotyledon material in Murashige and Skoog medium with 3% sucrose supplemented with 2,4-D (0.5–2 ppm) and kinetin (0.5–1 ppm). Regenerated plants can be obtained from the embryoids in Murashige and Skoog medium with 2% sucrose. Seedlings are usually transplanted. The spacing in experimental plantings in Java is 40 cm × 40 cm.

**Harvesting** Whole plants are uprooted, the tuberous roots separated, small rootlets removed, and finally the tuberous roots are washed and dried in the sun or shade.

**Yield** Intercropping *A. acutiloba* planted at 40 cm × 40 cm with maize and cabbage did not affect the yield of *Angelica* tuberous roots. In a field experiment in West Java, plantings of cultivar 'Siguyama Hikino' were harvested 6, 8, 10, 12, 14 and 16 months after transplanting. Root yields were 65–70% greater after 10 months than after 6–8 months.

**Handling after harvest** Roots of *A. acutiloba* were experimentally dried to a moisture content of about 11% and extracted with ethanol. The quality of the roots harvested after 10 months was of export grade with an ethanol extract percentage of 42%.

**Genetic resources and breeding** Breeding efforts in *A. acutiloba* are aimed at increasing the yield of tuberous roots and the concentration of active compounds. Experimental crosses between the cultivars 'Yamato Toki' (var. *acutiloba*) and 'Ibuki Toki' (var. *iwatenis* (Kitagawa) Hikino) showed a heterosis effect in the F-1 with respect to yield of tuberous roots and their sucrose content. However, the concentration of pharmacologically important compounds was intermediate between those of the parents. For industrial processing, breeding efforts should focus on obtaining plant



*Angelica acutiloba* (Siebold & Zucc.) Kitagawa - 1, root; 2, stem with flowers and fruits.

material with a uniform concentration of pharmacologically important compounds in the roots. Somatic embryogenesis appears to be a successful method for this purpose. Breeding programmes in Vietnam to promote the cropping of *Angelica* at lower elevations through adapted cultivars have been successful with *A. dahurica* and *A. polymorpha*.

**Prospects** The polysaccharides from *A. acutiloba* show promise in antitumour activity and immunology (anticomplementary activity). Research on their actions thus deserves further attention.

**Literature** |1| Kiyohara, H., Cyong, J.C. & Yamada, H., 1988. Structure and anticomplementary activity of pectic polysaccharides isolated from the root of *Angelica acutiloba* Kitagawa. *Carbohydrate Research* 182(2): 259-275. |2| Kumazawa, Y., Nakatsuru, Y., Fujisawa, H., Nishimura, C., Mizunoe, K., Otsuka, Y. & Nomoto, K., 1985. Lymphocyte activation by a polysaccharide fraction separated from hot water extracts of *Angelica acutiloba* Kitagawa. *Journal of Pharmacobiodynamics* 8(6): 417-424. |3| Nguyen Van Duong, 1993. *Medicinal plants of Vietnam, Cambodia and Laos*. Mekong Printing, Santa Ana, California, United States. pp. 412-413, 418. |4| Ohwi, J., 1965. *Flora of Japan*. Smithsonian Institution, Washington D.C., United States. pp. 680-682. |5| Okuyama, T., Takata, M., Nishino, H., Nishino, A., Takayasu, J. & Iwashima, A., 1990. Studies on the antitumor-promoting activity of naturally occurring substances. II. Inhibition of tumor-promoter-enhanced phospholipid metabolism by umbelliferous materials. *Chemical and Pharmaceutical Bulletin* 38(4): 1084-1086. |6| Perry, L.M., 1980. *Medicinal plants of East and Southeast Asia*. Attributed properties and uses. The MIT Press, Cambridge, Massachusetts, United States & London, United Kingdom. pp. 410-411. |7| Sada, Y., Tanaka, S., Tabata, M., Ozaki, K. & Komiya, T., 1993. Evaluation of intervarietal F-1 hybrids of the medicinal plant, *Angelica acutiloba*, clonally propagated by somatic embryogenesis. *Shoyakugaku Zasshi* 47(3): 235-242 (in Japanese). |8| Sudiarso, 1990. Effect of harvesting time on root yield of touki (*Angelica acutiloba*) in West Java. *Industrial Crops Research Journal* 2(2): 9-12. |9| Tanaka, S., Ikeshiro, Y., Tabata, M. & Konoshima, M., 1977. Anti-nociceptive substances from the roots of *Angelica acutiloba*. *Arzneimittelforschung* 27(11): 2039-2045. |10| Yamada, H., Komiya, K., Kiyohara, H., Cyong, J.C., Hirakawa, Y. & Otsuka, Y., 1990. Structural characterization and antitumor activity of a pectic poly-

saccharide from the roots of *Angelica acutiloba*. *Planta Medica* 56: 182-186, 420.

**Other selected sources** 74, 363, 397, 746, 747, 777, 881, 958, 1071, 1128, 1270, 1330, 1466, 1636.

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### ***Antiaris toxicaria* Lesch.**

Ann. Mus. Natl. Hist. Nat. 16: 478 (1810).

MORACEAE

2n = 24, 28

**Synonyms** *Antiaris macrophylla* R.Br. (1814), *Antiaris africana* Engl. (1902), *Antiaris welwitschii* Engl. (1902).

**Vernacular names** Upas tree, sacking tree (En). Indonesia: upas (general), ancar (Javanese), tatau (Sumatra). Malaysia: ipoh (Peninsular), tasem (Sarawak). Papua New Guinea: antiaris. Philippines: dalit (Tagalog), ipo (Tagalog, Bisaya). Burma (Myanmar): aseik, hymaseik. Laos: 'nong<sup>2</sup>, nong. Thailand: yang nong (central, northern), yuan (peninsular). Vietnam: [a]y sui, thu[oo]s[b]aws]n.

**Origin and geographic distribution** *Antiaris* is a monotypic genus. The only species, *A. toxicaria* is found throughout the Old World tropics, from West Africa to Madagascar, and in Sri Lanka, India, Indo-China, southern China, Thailand, throughout the Malesian region, the Pacific (east to Fiji and Tonga), and northern Australia.

**Uses** The latex of *A. toxicaria* obtained from the bark is one of the principal components of dart and arrow poisons in South-East Asia, used by many peoples for hunting and warfare. It is usually mixed with poison obtained from *Strychnos* species and components from other plants or poisonous animals. Its effectiveness is thought to be enhanced by the synergy of its poisonous and irritating components. The latex is also reported to be used as fish poison and birdlime. Although a single species, old reports claim that trees from regions outside South-East Asia are less poisonous or even innocuous. It is possible that these reports refer to the latex being used differently, not as a dart or arrow poison, and thus not entering the bloodstream.

Seeds, leaves and bark are used as a febrifuge and the seeds also as an antidysenteric. The latex is reported to be a mild circulatory and cardiac stimulant when used in very small amounts, but in large amounts it is a myocardial poison. In the Philippines, the soft wood is macerated and the



fluid is used as a poultice for swellings. In India, upas tree is used as a febrifuge and to treat dysentery and epilepsy. In Vietnam, Cambodia and Laos, it is not used medicinally.

*A. toxicaria* is a sacred tree among some South-East Asian peoples. The fruit contains latex, but is reported to be edible. The bark yields fibre to make clothing, cordage, sacks, mats and paper. The bark has also been used for dyeing. The wood is used in light construction, furniture, interior finish, pallets, crates and plywood.

**Production and international trade** The wood of *A. toxicaria* enters international trade, but the other products are for local use only.

**Properties** The active principles in the latex of *A. toxicaria* are the cardiac glycosides (cardenolides) e.g.  $\alpha$ ,  $\beta$  and  $\gamma$ -antiarin, which have digitalis-like effects on the heart. In larger amounts they lead to cardiac arrest and secondary effects such as vomiting and convulsions. Reports on lethal dosage, administered intravenously, specify 0.3 mg as lethal within 12 minutes for a rabbit, and 1 mg to cause death within 3–9 minutes in dogs.

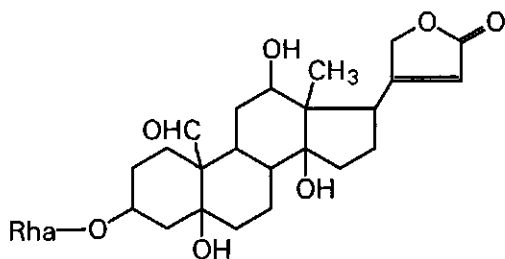
When administered to anaesthetized rats, the crude latex of *A. toxicaria* results in changes in the electrocardiogram (ECG) and systemic blood pressure. The extract inhibited the  $\text{Na}^+\text{K}^+\text{ATPase}$  that was partially purified from guinea-pig heart muscle. When the extract, and ouabain as a reference compound were applied to isolated frog heart muscles a decrease of twitch frequency together with an increased twitch tension were observed. All these facts together suggest that the main components of the latex are cardiac glycosides, which affect  $\text{Na}^+\text{K}^+\text{ATPase}$  activity of the heart muscle membrane. The poison must enter the bloodstream to be effective; the latex can be ingested without any effects. However, a fatal case of rhabdomyolysis and acute oliguric renal failure following oral ingestion of blowpipe dart poison prepared from *Antiaris* and *Strychnos* has been

reported. The influence of intravenous administration or otherwise promoting the poison to enter the bloodstream was illustrated in an experiment with dogs. When administered subcutaneously in pure form, there was no permanent toxic effect; when diluted with a decoction of *Strychnos ignatii* Bergius and administered in the same way the latex provoked a higher frequency of respiration, vomiting, convulsions and a rapid death. The sequence of the effects on toads were mild convulsions, violent peristalsis, an acceleration of the heartbeat followed by a deceleration, a contraction of the blood vessels, and death.

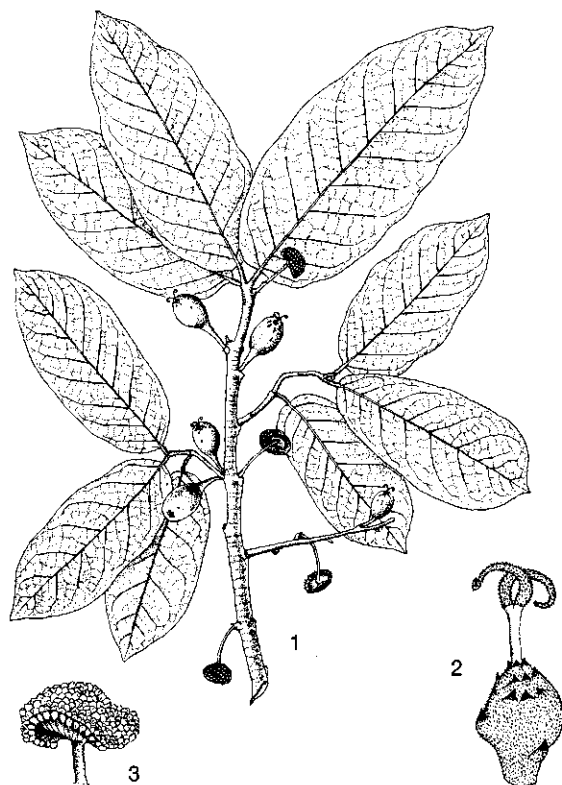
Prenylaurones (antiarone A and B), prenylchalcones (antiarone C, D and E) and prenylflavanones (antiarone F, G, H and I) have been isolated from the root bark. An aqueous ethanol extract of the bark exhibited cytotoxic activity against CA-9KB cells.

**Adulterations and substitutes** Cardiac glycosides are found in several dozen genera. In Vietnam, for example, other plants containing cardiac glycosides include *Asclepias curassavica* L., *Calotropis gigantea* (L.) Dryander, *Cerbera odollam* Gaertner, *Corchorus capsularis* L., *C. olitorius* L., *Digitalis* spp., *Nerium oleander* L., *Strophanthus* spp. and *Thevetia peruviana* (Pers.) K. Schumann. In western medical practice, pure glycosides produced by the extraction industry are used instead of crude plant products.

**Description** A monoecious, small to large tree up to 45(–60) m tall; bole straight, up to 180 cm in diameter, sometimes with steep buttresses up to 3 m high; bark surface smooth becoming slightly fissured, greyish-white, inner bark soft and fibrous, exuding a creamy copious latex which soon darkens to dirty brown and becomes granular upon exposure; twigs hairy. Leaves alternate, distichous, rounded to slightly heart-shaped, ovate or oblong, 7.5–20 cm  $\times$  3.5–8.5 cm, simple, slightly unequal at base, entire to denticulate; petiole 0.2–1 cm long, hairy; stipules free, caducous. Inflorescence on a short shoot, in leaf axils or below the leaves, subtended by involucre bracts, solitary or in groups of 2–4, the male ones below the female ones on the same twig. Male inflorescence a stalked discoid head with many flowers; each flower with 2–7 tepals and 2–4 stamens. Female inflorescence with 1–2 flowers, sessile or stalked; flower pear-shaped; perianth 4-lobed; ovary adnate to the perianth, 1-locular with a single ovule, styles 2. Fruit forming a drupaceous whole together with the enlarged, fleshy receptacle, ellipsoidal to pear-shaped, velvety. Seed one, cotyledon



$\beta$ -antiarin (Rha = rhamnose)



*Antiaris toxicaria* Lesch. – 1, fertile twig; 2, female inflorescence; 3, male inflorescence.

fleshy. Seedling with hypogeal germination, the epicotyl with a few scale leaves, followed by spirally arranged, conduplicate, dentate leaves.

**Growth and development** Trees of *A. toxicaria* develop according to Roux's architectural tree model, characterized by a continuously growing monopodial orthotropic trunk and plagiotropic branches. In a 27-year-old trial in Indonesia trees measured on average 17 m in height and 27 cm in diameter. In Java the trees flower in June on the new shoots.

**Other botanical information** Formerly, *Antiaris* comprised several species, but is now regarded monotypic. The variable species *A. toxicaria* has been divided into 5 subspecies. Subsp. *toxicaria* and subsp. *macrophylla* (R.Br.) C.C. Berg occur within the Malesian region; the first is found from Sri Lanka to Sulawesi, the second from the Philippines to Tonga. The size of the fruit increases from Africa to Polynesia.

The vernacular names 'upas' and 'ipoh' refer to plant poisons acting on the blood in general. Similarly, these names are used for *Strychnos* ('ipoh

akar') and *Sophora tomentosa* L. ('upas biji' or 'upas kamarunggi').

**Ecology** *A. toxicaria* is a rare, scattered tree in primary forest up to 1500 m altitude. It is occasionally found in grassy savanna and on coastal plateaus. The morphological variation as observed in habit and various parts of the plant may well be linked to environmental factors. In Africa, it occurs under semi-arid conditions as well as in rain forest areas, or even in swamp forest.

**Propagation and planting** *A. toxicaria* can be propagated by seed. About 70–90% of sown stones germinate in 18–89 days.

**Husbandry** Trees of *A. toxicaria* have a good self-pruning ability; they are not resistant to fire.

**Harvesting** The latex of *A. toxicaria* is tapped by making scores in the bark with a knife. It is only collected when required, as it cannot be stored and must be used fresh. The bark is harvested by stripping from the tree.

**Yield** The latex yield of a scarred tree may be 100–500 g in 2 days.

**Handling after harvest** An extensive list of traditional preparations and mixtures of 'upas poison' can be made. In general, the latex from the root-bark or bark is mixed with other ingredients such as bark or roots of *Strychnos*, *Derris* and other presumably irritating substances. The mixture is boiled over a fire to obtain a thick paste in which the dart and arrow points are dipped. The time over which the poison retains its potency is rather variable, apparently depending on mixture and method of preparation.

In Malaysia, bark cloth is obtained by shaving off the outer part from bark stripped from the tree, and beating and washing the inner fibrous part. Careful preparation is required, because traces of latex may irritate the skin.

**Genetic resources and breeding** Genetic erosion of *A. toxicaria* is difficult to assess, as trees are not widely harvested throughout their natural area of distribution, but generally occur at low densities.

**Prospects** The latex of *A. toxicaria* has been proposed as a medicine for heart diseases. As the crude drug extract varies in concentration and composition of the constituents, and given the extreme toxicity of the latex, it is rather difficult to standardize clinical applications.

**Literature** |1| Berg, C.C., 1977. Revisions of African Moraceae (excluding *Dorstenia*, *Ficus*, *Musanga* and *Myrianthus*). Bulletin du Jardin Botanique National de Belgique 47: 267–407. |2| Bisset, N.G., 1962. Cardiac glycosides: Part VI.

Moraceae: The genus *Antiaris* Lesch. *Planta Medica* 10: 143–151. [3] Boer, E. & Sosef, M.S.M., 1998. *Antiaris* Lesch. In: Sosef, M.S.M., Hong, L.T. & Prawirohatmodjo, S. (Editors): *Plant Resources of South-East Asia No 5(3)*. Timber trees: Lesser-known timbers. Backhuys Publishers, Leiden, the Netherlands. pp. 73–75. [4] Browne, F.G., 1955. *Forest trees of Sarawak and Brunei and their products*. Government Printing Office, Kuching, Malaysia. pp. 348–349. [5] Burkill, I.H., 1966. *A dictionary of the economic products of the Malay Peninsula*. Revised reprint volume 1 (A-H). Ministry of Agriculture and Cooperatives, Kuala Lumpur, Malaysia. pp. 175–185. [6] Council of Scientific and Industrial Research, 1948. *The wealth of India: a dictionary of Indian raw materials & industrial products*. Volume 1. Publications and Information Directorate, New Delhi, India. pp. 83–84. [7] Dolder, F., Tamm, C. & Reichstein, T., 1955. Die Glykoside von *Antiaris toxicaria* Lesch. *Glykoside und Aglycone*, 150 [Glycosides of *Antiaris toxicaria* Lesch. Glycoside and aglycones, 150]. *Helvetica Chimica Acta* 38(6): 1364–1396. [8] Hano, Y., Mitsui, P. & Nomura, T., 1990. Seven prenylphenols, antiarones C, D, E, F, G, H and I from the root bark of *Antiaris toxicaria* Lesch. *Heterocycles* 31(7): 1315–1324. [9] Pételot, A., 1954. *Les plantes médicinales du Cambodge, du Laos et du Vietnam*. [The medicinal plants of Cambodia, Laos and Vietnam]. Vol. 3. Centre National de Recherches Scientifiques et Techniques, Saigon, Vietnam. pp. 126–127. [10] Quisumbing, E., 1978. *Medicinal plants of the Philippines*. Katha Publishing Co., Quezon City, the Philippines. pp. 224–226.

**Other selected sources** 97, 162, 193, 248, 280, 283, 284, 438, 459, 546, 580, 594, 769, 1035, 1126, 1167, 1237, 1342, 1476, 1564.

E. Boer, M. Brink & M.S.M. Sosef

### *Arcangelisia flava* (L.) Merr.

Interpr. Herb. amboin.: 222 (1917).

MENISPERMACEAE

2n = unknown

**Synonyms** *Arcangelisia lemniscata* (Miers) Becc. (1877), *Arcangelisia loureiri* (Pierre) Diels (1910).

**Vernacular names** Yellow-fruited moonseed (En). Indonesia: areuy ki koneng (Sundanese), sirawan (Javanese), daun bulan (Moluccas). Malaysia: mengkunyit. Philippines: abutra (Ilokano, Bisaya), suma (Tagalog, Pampango). Thailand:

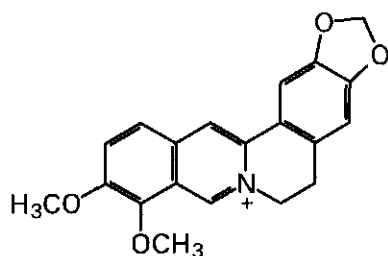
khamin khrua (Chanthaburi), kamphaeng jedchunum. Vietnam: v[ar]ly d[aw]s[ng].

**Origin and geographic distribution** *Arcangelisia* consists of only 2 species. Yellow-fruited moonseed is widely distributed from Hainan (China), Indo-China, southern peninsular Thailand, Peninsular Malaysia, Sumatra, Java, Borneo, the Philippines, Sulawesi, the northern Moluccas to New Guinea.

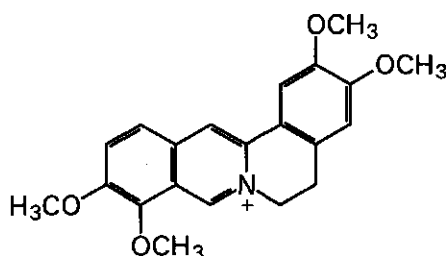
**Uses** Yellow-fruited moonseed is mainly used medicinally. In Peninsular Malaysia, a decoction of the stem is taken internally for jaundice, worms, indigestion and other intestinal complaints. The smoke from the burning wood is inhaled for troubles of the mucous membrane of the nose and mouth. In the Philippines, yellow-fruited moonseed is a popular antiseptic: a decoction of the wood is used to clean wounds, ulcers and other skin irritations. Traditional applications include the use of a decoction or infusion of the stem as a stomachic, febrifuge, expectorant, tonic, and emmenagogue or abortivum (depending on the quantity administered). In Indonesia, the stems are sold as 'kayu seriawan', meaning 'wood against sprue'. The sap which flows abundantly from cut stems is drunk against fever and sprue. In Thailand, the stems are used against indigestion, as a tonic and emmenagogue; the flowers are used to treat dysentery. In the Philippines, the Moluccas and New Guinea a yellow dye is extracted from the woody stem. The use of the fruits as a fish poison is questionable.

**Properties** *Menispermaceae* species are well-known to contain mixtures of (bis-)benzylisoquinoline-type alkaloids, which are biosynthetically derived from the amino acids phenylalanine or tyrosine. Alkaloids found in *A. flava* are: berberine, 8-hydroxyberberine, columbamine, jatrorrhizine, palmatine, thalifendine, dehydrocorydalmine, shobakunine (all of the quaternary protoberberine type), and (–)R,R-limacine, (+)R,S-homoaromaline and pycnarrhine (of the bisbenzylisoquinoline type). The pharmacological effects and the yellow colour of the dye extracted from the plant, are largely attributable to berberine, which is present in concentrations of up to 5% in the stem (dry weight).

The pharmacological effects of berberine have been fairly well investigated. Berberine (as the chloride) has been found to be active against a number of gram-positive as well as gram-negative bacteria, such as *Diplococcus pneumoniae*, *Escherichia coli*, *Neisseria gonorrhoeae*, *Salmonella typhosa*, *Shigella dysenteriae*, *Staphylococcus au-*



berberine



palmatine

*reus*, *S. hemolyticus* and *S. paradysenteria* in different media. It had about the same antibacterial activity as some sulphonamides; berberine also had an effect in broth supplemented with serum, whereas the sulphonamides were antagonized. However, it was found possible for the microorganisms to acquire resistance when left in contact with berberine for a long time.

Berberine (as the sulphate) has been shown to be bactericidal to *Vibrio cholerae* at a concentration of 35 µg/ml and bacteriostatic to *Staphylococcus aureus* at a concentration of 50 µg/ml. In both these organisms berberine at the concentrations mentioned inhibited RNA and protein synthesis almost immediately after addition. Cell-free preparations made from vibrios pretreated with berberine did not produce choleraic symptoms in infant rabbits, suggesting that the toxin was either inactivated or neutralized. Oral administration of berberine to infant rabbits 18–24 h before a single fatal intra-intestinal dose of choleraic toxin prevented toxin-induced diarrhoea and consequently prolonged survival when compared with untreated choleraic animals. The quaternary ammonium group in berberine seems necessary for its antibacterial activity. Derivatives without the quaternary ammonium group, such as tetrahydroberberine, showed only little antibacterial effect. Berberine (sulphate) in concentrations of 10–25 mg/ml inhibited the growth of the fungi *Alternaria* spp., *Aspergillus flavus*, *A. fumigatus*,

*Candida albicans*, *Curvularia* spp., *Drechslera* spp., *Fusarium* spp., *Mucor* spp., *Penicillium* spp., *Rhizopus oryzae* and *Scopulariopsis* spp. Oral administration of berberine sulphate at doses of 350–700 mg/kg was effective in treating *Candida albicans* infections of the intestine in mice.

Berberine (sulphate) administered to rats at doses of 100 mg/kg body weight, 10 days after experimentally induced intestinal amoebiasis was effective in 80% of the animals. It completely inhibited the growth to trophozoites of *Entamoeba histolytica* at concentrations of 0.5–1 mg/ml in vitro, and was active in vivo against infections with *E. histolytica* in hamsters and rats. Berberine has also been found to be trypanocidal against *Trypanosoma brucei rhodesiense*. In vitro activities with IC<sub>50</sub> values of 0.4 µg/ml were determined.

Both berberine sulphate (50 µg/ml) and berberine chloride (25 µg/ml) showed growth inhibition of Ehrlich and lymphoma ascites tumour cells. The presence of berberine in granules inside the cells was detected by its fluorescence. The cytotoxic ED<sub>50</sub> values in HeLa cell cultures were 3.5–30 µg/ml, and in KB cells a 70% inhibition of protein synthesis was found at a concentration of 1 µg/ml. Berberine chloride inhibited the formation of DNA, RNA, proteins and lipids, as well as the oxidation of [<sup>14</sup>C]glucose to <sup>14</sup>CO<sub>2</sub> when incubated with S180 (Swiss mouse ascites sarcoma) cells in vitro. Protein and RNA syntheses were most sensitive to berberine. However, berberine failed to inhibit the growth of S180 ascites tumours in mice, which may be explained by the effect of different glucose levels in biological fluids. The binding of the alkaloid to DNA was investigated by means of spectroscopy. Calf thymus DNA produced systematic changes in the absorption spectrum of berberine, which suggests that berberine forms a complex with DNA and binds to the extent of one alkaloid molecule per two base pairs. These binding properties seem to be influenced by the presence of charge and the position and type of substituents in the molecule. From other experiments it was also concluded that berberine is a potent activator for macrophages, to induce inhibition of tumour cells in vitro.

Intravenous infusion of berberine sulphate to rats was found to lower the blood pressure in a dose-dependent manner. A significant hypotensive effect was followed by bradycardia. These effects were also observed in bilaterally vagotomized rats. Berberine chloride at doses of 0.5–5.0 mg/kg administered to rabbits anaesthetized with urethane produced a long lasting, dose-related de-

crease in blood pressure. The berberine induced hypotension seems attributable to  $\alpha$ -adrenoceptor blockade, and not to a direct relaxant effect on vascular smooth muscle. Berberine had no direct vasodilatory effects on isolated rabbit pulmonary and cat coronary arteries either, however, the alkaloid reversed vasoconstriction mediated by  $\alpha$ -adrenergic agents in both preparations.

Both berberine and palmatine inhibited specific cholinesterase in rabbit spleen and pseudo-cholinesterase in normal horse serum. Both compounds were less effective inhibitory agents than neostigmine, but palmatine exhibited lower toxicity than berberine. Tetrahydropalmatine and tetrahydroberberine had no anticholinesterase effect, suggesting that the quaternary ammonium group is crucial for the effect of isoquinoline alkaloids on this enzyme.

Some metabolic and toxicological data on berberine are available from experiments on rats. The blood level of orally administered [ $^3\text{H}$ ]berberine chloride plateaued after 4–24 h, and maximal levels in the liver and muscles were achieved at 12 h. Urinary berberine excretion reached a maximum at 12–24 h. Excretion in the urine and faeces at 48 h amounted to respectively 2.7% and 86% of the administered dose. Faecal elimination as the main excretion route indicates that berberine is not readily absorbed by the gastro-intestinal tract. The biological half-life of berberine chloride was 5.2 h after intraperitoneal administration and 5.4 h after oral administration. Perfusion experiments (in dogs and rabbits) indicated oxidation of berberine chloride in the liver. The  $\text{LD}_{50}$  value of berberine sulphate was more than 1 g/kg after oral administration in the rat and about 90 mg/kg after intraperitoneal administration. Histopathological examinations revealed no changes in tissues and organs, even in cases when berberine sulphate had been given for 6 weeks at daily doses of 500 mg/kg.

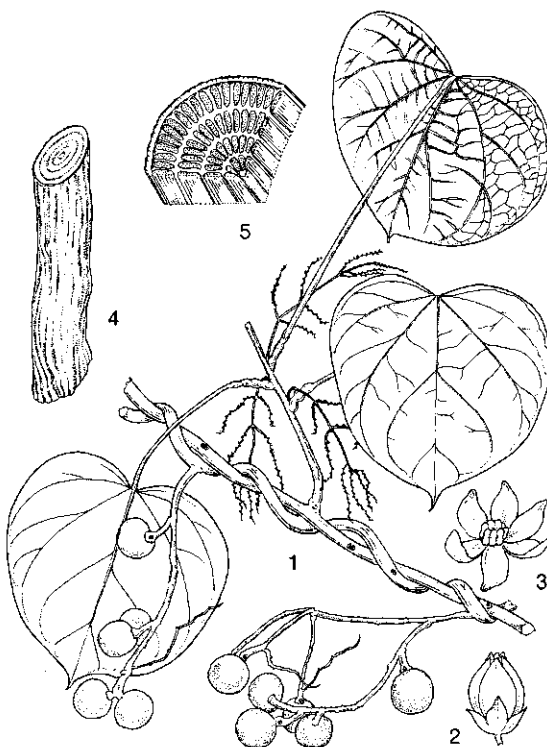
Of the bisbenzylisoquinoline alkaloids investigated, (+)-homoaromaline showed inhibition of the histamine production by RBL-2H3 cells *in vitro*, and both (+)-homoaromaline and (–)-limacine were capable of inhibiting the growth of cultured *Plasmodium falciparum* strains and tumour cell lines. However, their 'selectivity index' (activity against mammalian cells / activity against cultured *P. falciparum* strains) typically ranges from 2–100; a selectivity index of >1000 appears to indicate that a component merits further investigation as an anti-malarial.

Crude aqueous extracts of *A. flava* showed slight

insecticidal activity against cotton bollworm (*Helicoverpa armigera*) in the Philippines. Bollworms fed with treated cotton bolls were significantly smaller and shorter than their controls.

**Adulterations and substitutes** Several *Menispermaceae*, e.g. *Coscinium fenestratum* (Gaertner) Colebr., *Fibraurea tinctoria* Lour., *Limacia* spp. and *Tinospora* spp. contain berberine or related compounds and are used for similar purposes. *Coptis teeta* Wallich (*Ranunculaceae*) is another species containing berberine and with similar uses. Berberine has been named after the genus *Berberis* (*Berberidaceae*) in which the compound was found first. In India and Vietnam, for example, *Berberis* spp. are used against similar diseases as *A. flava*.

**Description** A large, woody, glabrous, dioecious liana, up to 20 m long; stem up to 5 cm in diameter, wood yellow, exuding yellow sap when cut. Leaves usually ovate, (10–)12–25 cm  $\times$  (5.5–)8–19 cm, coriaceous, palmately 5-veined at the base;



*Arcangelisia flava* (L.) Merr. – 1, intertwining flowering and fruiting stems; 2, closed male flower; 3, opened male flower; 4, part of woody stem; 5, detail of stem in cross and longitudinal section.

petiole (4–7–15–20) cm long, swollen at both ends; stipules absent. Inflorescence axillary or cauliflorous, paniculate, slender, 10–50 cm long, lateral branches spicate to subspicate. Flowers unisexual, with 3–4 minute outer sepals and 6 larger inner sepals, petals absent; male flower subsessile, with a sessile, globose cluster of 9–12 anthers; female flower with 3 carpels and a number of staminodes. Fruit a slightly laterally compressed drupe, transversely subovoid, 2–3 cm in diameter, yellow, with a club-shaped stalk; endocarp woody, covered with a dense mat of radial fibres. Seed broadly ellipsoidal, with ruminant endosperm, cotyledons much folded.

**Growth and development** The fruits are eaten and dispersed by primates such as macaques, gibbons and orang-utans, and probably other arboreal mammals.

**Other botanical information** Some confusion exists in the literature between *Arcangelisia flava* and *Anamirta cocculus* (L.) Wight & Arnott. *A. flava* has yellow wood and is used predominantly as a medicine, *Anamirta cocculus* has white wood and the fruits are used as a fish poison and an insecticide, while its bark is used as rope. The second *Arcangelisia* species (*A. tympanoda* (Lauterb. & K. Schumann) Diels) is apparently endemic to New Guinea and is poorly known.

**Ecology** *A. flava* occurs in forests at altitudes up to 1000 m, sometimes near river banks. In Sulawesi, it is reported on limestone.

**In vitro production of active compounds** In vitro production of berberine is possible. Callus cultures of *A. flava* have been established in Thailand, using revised tobacco medium supplemented with phytohormones (naphthalene acetic acid at 2 mg/l, indole-butyric acid at 2 mg/l and kinetin at 1 mg/l). The production can be increased on media containing coconut milk, casein hydrolysate, tyrosine (a berberine precursor), manganese sulphate and aluminium sulphate. The intensity of the yellow colouration of the callus is an indication of the amount of alkaloid produced.

**Harvesting** In Indonesia, the stems are cut in the early morning to obtain the sap which is drunk to cure fever and sprue.

**Yield** In the Philippines, it has been reported that as much as 1 kg of berberine can be obtained from a single plant.

**Genetic resources and breeding** Yellow-fruited moonseed is widespread and seems not to be easily liable to genetic erosion. There are no known germplasm collections or breeding programmes. Future breeding work may focus on the

selection of types with high alkaloid content.

**Prospects** Yellow-fruited moonseed is considered an interesting medicinal plant and may become important in cancer control. The development of proper biotechnological methods to produce alkaloids in tissue culture might provide the tools for large-scale production of alkaloids from *A. flava*.

**Literature** |1| Brown, W.H., 1951. Useful plants of the Philippines. Vol. 1. Technical Bulletin 10. Department of Agriculture and Natural Resources. Bureau of Printing, Manila, the Philippines. pp. 530–531. |2| Burkill, I.H., 1966. A dictionary of the economic products of the Malay Peninsula. 2nd edition, Vol. 1. Ministry of Agriculture and Co-operatives, Kuala Lumpur, Malaysia. pp. 215–217. |3| Chi, C.W., Chang, Y.F., Chao, T.W., Chiang, S.H., P'Eng, F.K., Lui, W.Y. & Liu, T.Y., 1994. Flowcytometric analysis of the effect of berberine on the expression of glucocorticoid receptors in human hepatoma HepG2 cells. *Life Sciences* 54(26): 2099–2107. |4| Creasey, W.A., 1979. Biochemical effects of berberine. *Biochemical Pharmacology* 28(7): 1081–1084. |5| de Padua, L.S., Lugod, G.C. & Pancho, J.V., 1977. Handbook on Philippine medicinal plants. Vol. 1. Documentation and Information Section, Office of the Director of Research, University of the Philippines, Los Baños, the Philippines. p. 43. |6| Forman, L.L., 1986. Menispermaceae. In: van Steenis, C.G.G.J. & de Wilde, W.J.J.O. (Editors): *Flora Malesiana*. Series 1, Vol. 10. Kluwer Academic Publishers, Dordrecht, Boston, London. pp. 209–211. |7| Heyne, K., 1927. *De nuttige planten van Nederlandsch Indië* [The useful plants of the Dutch East Indies]. 2nd edition. Vol. 1. Departement van Landbouw, Nijverheid & Handel in Nederlandsch-Indië. p. 621. |8| Quisumbing, E., 1978. *Medicinal plants of the Philippines*. Katha Publishing Co., Quezon City, the Philippines. pp. 293–294. |9| Tang, W. & Eisenbrand, G., 1992. *Chinese drugs of plant origin*. Springer Verlag, Berlin, Heidelberg, New York. pp. 361–371. |10| Yanpaisan, W., 1989. The effect of phytohormones and some additives on tissue culture establishment and in vitro production of alkaloids from *Arcangelisia flava* Merr. *Journal of the National Research Council of Thailand* 21(1): 1–27.

**Other selected sources** 433, 531, 750, 857, 859, 863, 1005, 1287, 1389.

E.H. Mandia, C.E. Ridsdale,  
S.F.A.J. Horsten & A.M. Aguinaldo

## Aristolochia L.

Sp. pl. 2: 960 (1753); Gen. pl. ed. 5: 410 (1754).

ARISTOLOCHIACEAE

$x = 7$ ; *A. tagala*:  $2n = 14$

**Major species** *Aristolochia tagala* Cham.

**Vernacular names** Birthwort, Dutchman's pipe (En). Snakeroot (Am). Aristoloche (Fr).

**Origin and geographic distribution** *Aristolochia* consists of approximately 300 species and is mainly distributed throughout the tropics, but some species occur in warmer temperate regions. The greatest diversity in species is found in Central and South America. In the Malesian region 28 species have been found; *A. tagala* has the largest area of distribution, occurring from India and China, throughout South-East Asia, to Australia.

**Uses** *Aristolochia* is not much used in local medicine in South-East Asia. The use of a decoction of the roots as a stomachic, emmenagogue and febrifuge is most common. A poultice of the leaves is sometimes used to treat skin diseases. Some extra-Malesian species are, however, renowned in Chinese, Indian and South American health care systems, and are included in pharmacopoeias.

The Chinese Pharmacopoeia lists e.g. the dry ripe fruits of *A. contorta* Bunge and of *A. debilis* Sieb. & Zucc. which are used in the treatment of respiratory diseases as an antitussive and antiasthmatic. Their dry aerial parts are also used as a diuretic against oedema and as an antirheumatic. The root of *A. fangchi* Y.C. Wu ex L.D. Chou & S.M. Hwang is valued as an antirheumatic and diuretic and the dried vine of *A. mandshuriensis* Kom. is a well-known diuretic and antiphlogistic for treatment of oedema and rheumatic pain. In China and Japan, extracts from the roots of *A. debilis* are furthermore used to treat high blood pressure.

*A. bracteolata* Lamk (synonym: *A. bracteata* Retz.) and *A. indica* L. have a considerable reputation in India. *A. bracteolata* is reputed for its purgative and anthelmintic properties; a root decoction is employed to expel roundworms, and is known as an emmenagogue. In Africa, *A. bracteolata* has a local reputation as a powerful anthelmintic mostly used in veterinary practice. The dried rhizomes and roots of *A. indica* constitute an important drug in India, much esteemed as a gastric stimulant and bitter tonic and used to treat intermittent fevers. The drug is prescribed as a tincture, and sometimes administered as a powder. The juice of fresh leaves is used to treat coughs, and the seeds to treat inflammations, biliousness and cough. Juice from the leaves is applied to ulcers

and, mixed with castor oil, to eczema. *A. indica* is also used in India as an antidote for snakebites and scorpion stings. In traditional Thai medicine, the roots are used as antipyretic, emmenagogue, expectorant and tonic. The leaves are used in the treatment of snakebites. Its roots possess antifertility activity. *A. tagala* roots are considered in India to be tonic, carminative and emmenagogue. They are frequently used there to adulterate *A. indica* for use in medicine. *A. elegans* Masters from tropical America has medicinal and insecticidal properties and is locally cultivated in South-East Asia, e.g. in the Philippines and Thailand. In Vietnam, some species are imported from China to be used in local medicine, e.g. *A. heterophylla* Hemsley and *A. westlandii* Hemsley, which are used as diuretic and prescribed to treat oedema and dysuria. *A. serpentaria* L. is used in local medicine in North America.

Several American species are cultivated as ornamentals for their beautiful flowers. Some species are cultivated as food plants for the larvae of commercially traded swallowtail and birdwing butterflies.

**Production and international trade** None of the Malesian *Aristolochia* species are traded, but fruits of *A. debilis* are imported from China and sold in Chinese medicine shops in Peninsular Malaysia.

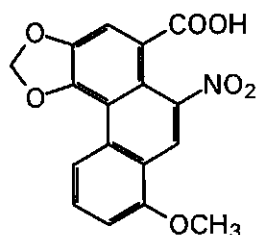
**Properties** The chemical constituents of *Aristolochia* species can generally be divided into three chemical groups: aristolochic acids (derivatives), alkaloids and sesquiterpenes.

The aristolochic acids are a family of at least 14 closely related structures, derived from the phenanthrene system and bearing a carboxyl function and a nitro substituent. These compounds are intensely bitter, and it is hypothesized that in their biosynthesis oxidative coupling of orientaline gives prestephanine, which is converted into stephanine; oxidative cleavage of stephanine then furnishes aristolochic acids. When the nitro group is replaced biosynthetically by an amino group, the carboxyl group forms a lactam ring, giving a number of aristolactams. Aristolochic acid can furthermore be converted to aristolic acid by a one-step removal of the nitro group.

One of the alkaloids isolated from some *Aristolochia* species is magnoflorine (an aporphine type alkaloid derived from phenylalanine/tyrosine), which is structurally and phylogenetically strongly related to aristolochic acid derivatives. The sesquiterpenes are mainly constituents of the volatile oil.

Not much is known about the properties and chemical constituents of the South-East Asian species, except for some information on *A. tagala* from the root of which aristolochic acid I (= aristolochic acid A), aristolochic acid IIIa (aristolochic acid C), 9-hydroxyaristolochic acid I and allantoin have been isolated. Much more research has been done on some Indian and Chinese species.

Aristolochic acid I (0.1–0.6%) and debilic acid have been isolated from the roots of *A. debilis*, together with 9-hydroxy and 9-methoxyaristolochic acid I, aristolochic acid II, aristolochic acid IIIa, aristolochic acid IV, aristolochic acid IVa (= aristolochic acid D) and some aristolactams. Furthermore isolation of the alkaloidal constituents magnoflorine, cyclanoline, tetrandrine, N-acetyl-nornuciferine and allantoin has been reported. The sesquiterpenes isolated are mainly of the aristolane type: e.g. aristolone (about 0.42%), 9-aristolene, 1(10)-aristolene and debilone. Aristolochic acid I methyl ester, aristolochic acid IV methyl ester and aristolochic acid IVa were isolated from the stems of *A. mandshuriensis*, and magnoflorine detected. In addition, a new glycoside of aristolochic acid D was also isolated and named aristolose or aristolochin. Phytochemical analysis of the constituents in the roots of *A. fangchi* revealed amongst others the presence of aristolochic acid I, aristolochic acid IVa, aristolochic acid IV methyl ester, magnoflorine, p-coumaric acid and N-(p-hydroxyphenyl)-p-coumaramide in the ethanol extract. Together with aristolochic acid I, magnoflorine and allantoin, a new compound aristolochic acid E was isolated from the root of *A. contorta*. Several aristolochic acids and derivatives (e.g. aristolic acid), sesquiterpenes, alkaloids (e.g. l-curine), steroids, p-coumaric acid and a naphthaquinone (aristolindiquinone) have been identified from the roots of *A. indica*. A phytochemical investigation of the leaves of *A. elegans* indicated the presence of sterols and 5 alkaloids. The steroidal material was isolated and identified as  $\beta$ -sitosterol. The seeds of *A. bracteolata* contain



aristolochic acid I

aristolochic acid and magnoflorine. Aristolochic acid I and IVa, magnoflorine, allantoin and  $\beta$ -sitosterol have been isolated and identified from the roots of *A. heterophylla*.

The biological activities of aristolochic acid I have been studied extensively. A number of gram-positive bacteria (including *Bacillus*, *Diplococcus*, *Mycobacterium*, *Sarcina*, *Staphylococcus* and *Streptococcus*) are inhibited by this acid at a concentration of 50–200  $\mu\text{g/ml}$ . A concentration of aristolochic acid I higher than 200  $\mu\text{g/ml}$  was needed to inhibit gram-negative bacteria and fungi. Mice infected with *Diplococcus pneumoniae*, *Staphylococcus aureus* or *Streptococcus pyogenes* were found to be protected from disease by intraperitoneal administration of aristolochic acid I at a dose of 50  $\mu\text{g/kg}$ . The phagocytic activity of peritoneal macrophages of treated mice was markedly stimulated. Some results of studies on the immunostimulating and antitumour activities of aristolochic acid I contradicted each other. It has been reported that the survival time of mice bearing ascitic sarcoma-37 tumours treated with aristolochic acid by intraperitoneal administration at a daily dose of 1–5  $\text{mg/kg}$  for 5 days was appreciably prolonged. Growth of mouse sarcoma-37 cells was found to be completely inhibited by incubation with aristolochic acid I. Treating mice with aristolochic acid I at a daily dose of 2.5–5  $\text{mg/kg}$  for 3 days after subcutaneous implantation of sarcoma-37 cells resulted in 40–50% inhibition of tumour growth. Aristolochic acid I, administered orally, reduced the number of tumours induced by methylcholanthrene in mice. Aristolochic acid I increased oxygen consumption in a dose-dependent manner in liver cells and splenocytes of mice. The metabolic activity of guinea-pig peritoneal macrophages and human leucocytes was also enhanced by aristolochic acid I, as shown by measuring oxygen consumption. Both aristolochic acid I and II exhibited a stimulation of lucigenine-enhanced, opsonized Zymosan-induced neutrophil chemiluminescence as a sensitive assay for immunomodulating activity. In a leucocyte adherence inhibition test, an activity of aristolochic acid I could also be demonstrated; however, it was weaker than that of prednisolone. Following the administration of aristolochic acid I to guinea-pigs immunized with Q-fever antigen, the antigen-induced decrease in bone marrow lymphocyte count was restored to normal levels much faster than was observed in untreated immunized controls. In contrast to these results, there is also a report that aristolochic acid I did not prolong the sur-



vival time of tumour-bearing mice or enhance the immune function of the mouse reticuloendothelial system, or the phagocytic activity of mouse peritoneal macrophages. Aristolochic acid has been shown to be a non-competitive inhibitor of phospholipase A2 from snake venom. It inhibited the oedema-inducing and haemolytic activity of this compound in the venom, but failed in tests to inhibit other pathological activities of the enzyme.

Aristolochic acids are known for their nephrotoxicity in humans and several animal species, and their mutagenic and carcinogenic activities have also been extensively studied. Aristolochic acid I has been proved to be a direct mutagen in *Salmonella typhimurium* strains TA 1537 and TA 100, but had no mutagenic effect on TA 1535, TA 1538 or TA 98. Aristolochic acid II had almost equal mutagenic potency, and the aristolactams were mutagenic in both strains too, when a metabolizing system was present. The mutagenic activity of aristolochic acid I was also tested in the granuloma pouch assay, which detects gene mutations induced in subcutaneous granuloma tissue of rats. After direct exposure of the target tissue, aristolochic acid I induced high frequencies of mutations at a relatively low cytotoxic level. After oral administration of aristolochic acid I to rats, a dose dependent mutagenic activity was registered. The carcinogenic activity of aristolochic acid I has been demonstrated in experimental animals. Male and female rats treated orally with aristolochic acid I at daily doses of 0.1, 1.0 or 10.0 mg/kg developed a high incidence of tumours, dependent on dose and time. Rats treated for 3 months with either 1.0 or 10.0 mg/kg aristolochic acid I developed severe papillomatosis of the forestomach, with occasional signs of malignancy. Without further treatment, the rats developed squamous cell carcinomas in the forestomach 3–6 months later and formation of metastases. For these reasons, many European countries (e.g. Germany and France) have restrictive regulations for preparations containing *Aristolochia*, even including homeopathic preparations with their great dilution.

The alkaloid magnoflorine is a hypotensive principle. In rabbits it decreased arterial blood pressure and induced hypothermia. In anaesthetized cats, intravenous injection of 2 mg/kg magnoflorine produced a prompt and significant fall in blood pressure. Oral administration at a dose of 20–40 mg/kg also resulted in hypotension. The acute LD<sub>50</sub> of magnoflorine by intravenous injection in mice was 20 mg/kg. Oral administration with a

tenfold higher dose daily for 4 weeks did not elicit any toxic symptoms or retard growth.

Aristolic acid (biosynthetically derived from aristolochic acid through removal of the nitro group) obtained from *A. indica* disrupted nidation in mice when administered on the first day of pregnancy. It showed implantation-inhibiting effects. Possibly this compound interferes with steroidal conditioning of the uterus. Furthermore, both aristolochic acid I and magnoflorine induced contractions in isolated pregnant rat uterus and stimulated guinea-pig ileum. p-Coumaric acid isolated from *A. indica* roots is known to be an inhibitor of prolactin secretion.

Ethanol extracts of *A. indica* roots decreased fertility in rats and hamsters. The petroleum ether extract of the roots showed 100% interceptive activity in mice at a single dose of 100 mg/kg. The sesquiterpene (12S)-7,12-secoishwaran-12-ol is reported as another active principle. Other laboratory experiments have failed to demonstrate activity on uterine contraction, and experiments on abortifacient activity have been inconclusive.

Tests on rats with ethanolic extracts of *A. indica* showed no antipyretic activity. The ethanol extract of *A. bracteolata* exhibited uterine stimulant and anthelmintic properties. Aqueous and alcohol extracts of *A. debilis* were found to be highly effective against herpes simplex virus in vitro, and also showed some effect on respiratory syncytial virus and coxsackie virus. Extracts of *A. mandshuriensis* showed significant inhibitory activity on the mutagenicity of 3-amino-1,4-dimethyl-5H-pyrido-(4,3-b)-indole, and angiotensin converting enzyme.

The aristolochic acids extracted from *A. bracteolata* showed chemosterilizing effects on several insect species including mosquitoes. In the Philippines, extracts from *A. tagala* showed insecticidal properties against the yponomeutid crucifer pest *Plutella xylostella*, the maize pest *Ostrinia furnacalis* and the common cutworm (*Spodoptera litura*). A methanolic extract of *A. bracteolata* showed significant inhibitory effect on the aflatoxin production of *Aspergillus flavus*, and the plant may have potential as an antifungal agent.

**Adulterations and substitutes** There are indications that *Aristolochia* extracts are sometimes mistaken for *Menispermaceae* extracts. Rapidly progressive renal fibrosis has been described in young women who had taken Chinese herbs as part of a slimming regime. Aristolochic acid was suspected as its causal factor, but this compound is not present in the *Stephania* extract, which is

one of the ingredients of the drug. Possibly an *Aristolochia* extract and not a *Stephania* extract was used to prepare the drug in question.

**Description** Woody or herbaceous perennial climbers or erect, scandent to scrambling shrubs often woody at base; tuberous or prostrate rhizomes or rootstocks often present; older woody stems usually with a thick corky and fissured bark. Leaves arranged spirally or alternate, simple, usually entire but sometimes 3-lobed, venation palmate or pinnate, secondary veins often extending obliquely towards the leaf margin; petiole grooved above; stipules absent. Flowers solitary or in fascicles or in cymose, racemose, spicate or paniculate inflorescences, in the axils of leaves or borne on the stems, bisexual, zygomorphic (rarely actinomorphic), bracts usually present and persistent, pedicel usually hardly distinct from the ovary; perianth consisting of 3 parts with a basal inflated part (utricle), a straight or curved cylindrical tube, and the expanded 3(-6)-lobed limb with valvate or induplicate lobes or (usually) 1-lipped; stamens 6(-10), in a single whorl, adnate to the style column to form a gynostemium, anthers extrorse and dehiscing longitudinally; ovary inferior, oblong or elongate, 6-celled, style column 6-lobed. Fruit a 6-celled capsule, dehiscing septically, usually basally towards the apex, many-seeded. Seeds ovate, deltoid or triangular, flat, often winged, testa crustaceous or hard, finely verrucose or smooth, funicle often thickened and covering the whole seed, with fleshy and copious albumen and minute embryo. Seedling with epigeal germination; cotyledons rather fleshy; first 2 leaves opposite, subsequent ones alternate.

**Growth and development** In *Aristolochia* the flowers of an inflorescence open only singly or very few at a time. Their form is related to the fly-trap pollination mechanism. The flowers are insect-pollinated, usually by flies and sometimes by ants attracted by the putrid odour of the flowers and trapped in a kettle-like part (utricle), after having passed a 'slide zone' on the limb. The flower tube in between the limb and utricle is usually provided with retrorse hairs preventing insects from leaving the utricle. The flowers are protogynous and the ripe stigmas may be pollinated by insects when entering the utricle, where glands provide feed to keep them alive until the stamens have ripened. Mostly flowers open around daybreak and wither after about 24 hours, but sometimes they last longer. After the stamens have shed their pollen, the flower withers, the hairs in the tube lose turgescence, and the insects

can leave the utricle and possibly visit another flower, leading to cross-pollination. However, some *Aristolochia* species are not furnished with such specialized systems to promote cross-pollination and are then self-fertilized.

The fleshy funicle of the seed forms an elaiosome which is probably attractive for ants dispersing the seeds. The seed wing present in some *Aristolochia* (e.g. *A. tagala*) may serve wind dispersal.

**Other botanical information** Flowering material of *Aristolochia* is generally needed for correct identification, but flowers are often scantily represented in herbaria, and are commonly deformed by drying.

**Ecology** *Aristolochia* usually occurs scattered, often in primary forest, but some species (e.g. those discussed here) are also found in secondary forest and scrub vegetation. Most species are confined to lowland forest, but some occur above 1500 m altitude.

Caterpillars of several butterfly species (particularly swallowtail butterflies of the family *Papilionidae*) are known to feed exclusively on leaves and young shoots of *Aristolochia*. They use chemical compounds in the plants to become poisonous for predators. For instance, it has been shown that aristolochic acid I present in larvae feeding on *A. debilis* deterred feeding of tree sparrows, but also triggered cannibalistic activity of the larvae against eggs and pupae, which also contain the compound. It has also been demonstrated that aristolochic acid induces a significant oviposition response in female swallowtail butterflies.

**Propagation and planting** In India, *Aristolochia* is usually propagated by seed, which germinates in about 2 weeks. Sometimes it is grown from rhizomes collected from the wild.

Experimental in vitro propagation has been successful for *A. bracteolata* in India. The formation of callus was observed from young leaves and nodes placed on Murashige and Skoog medium supplemented with kinetin, naphthalene acetic acid and indole acetic acid. Roots were initiated from the callus when the concentrations of kinetin and indole acetic acid were increased. From a single nodal segment 1-4 shoots were raised, and shoots grew 5-6 cm tall within 30 days.

**Diseases and pests** Although caterpillars of various butterfly species feed on the leaves of *Aristolochia*, they are rarely reported to cause extensive defoliation.

**Harvesting** In India, *A. indica* is allowed to grow for 2 years to yield rootstocks of marketable size.

**Yield** The yield of *A. indica* rootstocks in India is estimated at 4.5–5.6 t/ha in 2-year-old plantations.

**Handling after harvest** No information available.

**Genetic resources and breeding** Most *Aristolochia* species have a limited area of distribution and occur very scattered in lowland forest. This makes them very vulnerable to genetic erosion due to rapid changes in land use. Some species (e.g. *A. indica* in India) have already become rare in the wild because of their popularity for medicinal purposes.

**Prospects** The medicinal uses of *Aristolochia* are extremely local in South-East Asia and in addition many of them were reported more than 100 years ago. However, *Aristolochia* is widely used medicinally, particularly in China and India, and the activity for several applications has been demonstrated by research. The South-East Asian *Aristolochia* probably have similar properties, but these still have to be confirmed by experiments. A major drawback for use in medicine is the carcinogenic activity of some of the major active compounds, limiting the application in modern medicine. The viricidal, fungicidal and insecticidal properties might offer prospects for wider use.

**Literature** |1| Bruneton, J., 1995. Pharmacognosy, phytochemistry, medicinal plants. Lavoisier Publishing, Paris, France. pp. 748–749. |2| Council of Scientific and Industrial Research (various editors), 1985. The wealth of India. Revised Edition. Vol. 1. Publications and Information Directorate, New Delhi, India. pp. 422–427. |3| Ding Hou, 1984. Aristolochiaceae. In: van Steenis, C.G.G.J. & de Wilde, W.J.J.O. (Editors): Flora Malesiana. Series 1, Vol. 10. Kluwer Academic Publishers, Dordrecht, Boston, London. pp. 53–108. |4| El Tahir, K.E.H., 1991. Pharmacological actions of magnoflorine and aristolochic acid-1 isolated from the seeds of *Aristolochia bracteata*. International Journal of Pharmacognosy 29(2): 101–111. |5| Ganguly, T., Pakrashi, A. & Pal, A.K., 1986. Disruption of pregnancy in mouse by aristolochic acid I. Plausible explanation: relation to early pregnancy events. Contraception 34(6): 625–638. |6| Phuphanaphong, L., 1987. Aristolochiaceae. In: Smitinand, T. & Larsen, K. (Editors): Flora of Thailand. Vol. 5, Part 1. The Forest Herbarium, Royal Forest Department, Bangkok. pp. 1–31. |7| Quisumbing, E., 1978. Medicinal plants of the Philippines. Katha Publishing Co., Quezon City, the Philippines. pp. 254–256. |8| Remeshree, A.B., Hariharan, M. & Unnikrishnan, K., 1994. Micro-

propagation and callus induction of *Aristolochia bracteolata* Lam. – a medicinal plant. Phytomorphology 44(3–4): 247–252. |9| Schmeiser, H.H., Bieler, C.A., Wiessler, M., van Ypersele de Strihou, C. & Cosyns, J.P., 1996. Detection of DNA adducts formed by aristolochic acid in renal tissue from patients with Chinese herbs nephropathy. Cancer Research 56(9): 2025–2028. |10| Tang, W. & Eisenbrand, G., 1992. Chinese drugs of plant origin. Springer Verlag, Berlin, Heidelberg, New York. pp. 145–157.

#### *Selection of species*

#### ***Aristolochia philippinensis* Warb.**

Perkins, Fragm. fl. Philipp.: 170 (1905).

**Vernacular names** Philippines: barubo (Negrito), puso-pusoan (Tagalog), tambal-balanding (Zambales).

**Distribution** The Philippines (Luzon, Mindoro, Bancalan Island, Mindanao).

**Uses** A decoction of the roots is used in traditional medicine in the Philippines as a stomachic and emmenagogue.

**Observations** An erect shrubby plant up to about 1 m tall, with terete old stems up to 4 cm in diameter, slightly irregularly ridged, and slightly striate branches; leaves elliptical, lanceolate to oblanceolate, 8.5–24 cm × 3.5–8.5 cm, obtuse, sometimes slightly cuneate at base, usually glabrous on both surfaces, with 1 pair of faint basal veins, 5–8 pairs of secondary veins and loosely reticulate tertiary veins; flowers in a spicate or racemose inflorescence, perianth 1-lipped, distinctly veined; fruit subglobose, shortly cylindrical to oblong-ellipsoidal, up to 2.5 cm long; seeds not winged. *A. philippinensis* occurs in thickets and forest up to 900 m altitude.

**Selected sources** 356, 1126, 1178.

#### ***Aristolochia rumphii* Kostel.**

Allg. med.-pharm. Fl. 2: 465 (1833).

**Vernacular names** Indonesia: akar pulurun, tuhe tutunu, warosbot (Moluccas).

**Distribution** The Lesser Sunda Islands, southwestern Sulawesi and the Moluccas.

**Uses** A decoction of the roots (and sometimes of the twigs but this is less powerful) is used to treat stomach-ache, spasm, constipation and intermittent fever.

**Observations** A climber, woody at base and with long slender stem; leaves elliptical-oblong or ovate-oblong to narrowly lanceolate, 7–12.5 cm ×

(1)–3–5.5 cm, obtuse, sometimes slightly cuneate at base, minutely hairy beneath, with 1 pair of basal veins, 4–5 pairs of secondary veins and loosely transverse or reticulate tertiary veins; flowers in a short racemose inflorescence, perianth 1-lipped, green with brown limb; fruit short cylindrical, about 2.2 cm long; seeds not winged. *A. rumphii* occurs in open forest, thickets and grassland up to 100 m altitude.

**Selected sources** 356, 580.

### *Aristolochia sericea* Blanco

Fl. Filip.: 283 (1837).

**Synonyms** *Aristolochia imbricata* Masters (1875), *Aristolochia membranacea* Merr. (1919).

**Vernacular names** Philippines: bangisi, pangisi (Iloko).

**Distribution** The Philippines (Luzon).

**Uses** Roots are chewed to treat gastralgia, and, macerated in spirituous liquor, the drug is administered as a uterine tonic after childbirth; it is a violent abortive. The entire fresh plant is used as carminative, emmenagogue and febrifuge.

**Observations** An erect shrubby plant up to about 0.5 m tall, with terete stems about 3 mm in diameter, initially densely pubescent; leaves lanceolate or oblong-lanceolate, 7–15 cm × 2–5 cm, usually shallowly cordate at base, sparsely hairy beneath, with 2 pairs of basal veins and about 5 pairs of faint secondary veins; flowers in a few-flowered and short inflorescence, perianth 1-lipped, with rather distinct reticulation; fruit subglobose, about 1 cm in diameter, initially densely hairy; seeds not winged. *A. sericea* occurs in dry thickets up to 350 m altitude.

**Selected sources** 190, 356, 1126, 1178.

### *Aristolochia tagala* Cham.

Linnaea 7: 207, t. 5, f. 3 (1832).

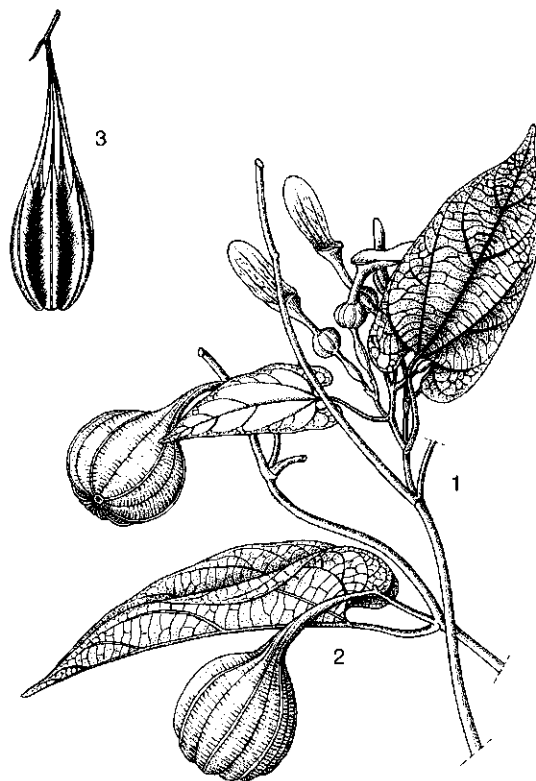
**Synonyms** *Aristolochia roxburghiana* Klotzsch (1859), *Aristolochia megalophylla* K. Schumann (1889), *Aristolochia mindanaensis* Warb. (1905).

**Vernacular names** Indonesia: kalayar (Sundanese), puyan (Javanese), kunit (Sulawesi). Malaysia: akar ketola hutan, akar petola hutan (Peninsular). Philippines: timbangan (Tagalog), goan-goan (Bisaya), nagerus (Iloko). Thailand: krachao pheemot, krachao mot (central). Vietnam: ph[of]ng k[yr], d[aa]y kh[oo]s r[as]ch.

**Distribution** From India, Sri Lanka and Bangladesh, through Burma (Myanmar), Indo-China (Cambodia, Vietnam), China and Thailand, to the whole of Malesia, the Solomon Islands and Australia (Queensland).

**Uses** Powdered roots are used as a tonic, carminative and emmenagogue, and to treat infantile tympanites in the Philippines. In Malaysia, pounded leaves are applied to the head to treat fever. In Papua New Guinea, leaves are rubbed over a patient's head and subsequently mixed with water and given to the patient to drink. In the Moluccas, leaves ground with *Curcuma* are warmed and applied externally to treat swollen limbs, colics and skin diseases.

**Observations** A climber up to 20 m long, with terete, slightly furrowed branches up to 5 mm in diameter; leaves usually ovate to ovate-oblong, 6–20(–27) cm × 4–10(–16) cm, cordate at base with rounded auricles, sparsely short-haired to subglabrous beneath, with 2 pairs of basal veins, 3–5 pairs of secondary veins and loosely reticulate or crossbar-like tertiary veins; flowers in a racemose or paniculate inflorescence, perianth 1-lipped, with faint venation, pale yellowish or greenish to purplish or dark reddish-brown; fruit subglobose, slightly pyriform or oblong, up to 4 cm long; seeds winged. *A. tagala* occurs in forest and thickets,



*Aristolochia tagala* Cham. – 1, flowering stem; 2, fruiting stem; 3, dehiscent fruit.

usually up to 800 m altitude, but in Thailand up to 1050 m and in New Guinea up to 1350 m.

**Selected sources** 202, 288, 359, 364, 580, 597, 1126, 1137, 1178.

R. Kiew

## Artemisia L.

Sp. pl. 2: 845 (1753); Gen. pl. ed. 5: 367 (1754).

COMPOSITAE

$x = 8, 9$ ; *A. annua*:  $2n = 18$ , *A. apiacea*:  $2n = 18$ , *A. capillaris*:  $2n = 18, 36$ , *A. vulgaris*:  $2n = 16, 18, 24, 54$

**Major species** *Artemisia annua* L., *A. capillaris* Thunb., *A. vulgaris* L.

**Vernacular names** Mugwort, wormwood (En).

**Origin and geographic distribution** *Artemisia* consists of approximately 200 species (some estimates are up to 400 species), most of which are native to dry grassland regions of Eurasia and North America. The regions of central and southwestern Asia are particularly rich in species; the genus is thought to have originated here, and to have migrated to North America. Several species have been introduced in the Malesian area, usually as ornamentals; some have naturalized.

**Uses** *Artemisia* is well known in phytotherapy all over the world. Numerous species are used in local medicine. The rediscovery in the 1970s of *A. annua* as remedy against malaria was spectacular. It had already been used for over 1000 years in China to treat malarial fever, but modern researchers became interested in its properties when the search for new antimalarial medicines started in response to the growing resistance of malaria-causing agents (*Plasmodium* spp.) to the industrial antimalarial drugs currently in use. The active compound, artemisinin (or 'qinghao-su' in China), a sesquiterpene lactone endoperoxide, may be administered in tablets or suppositories. A series of artemisinin derivatives has been semi-synthesized, often with improved pharmacological, pharmaceutical, technological or pharmacokinetic properties; some of them are also used clinically now. The methyl-ether derivative, artemether, and the ethyl-ether derivative artemotil (proposed INN name, previously  $\beta$ -arteether), which are better lipid soluble, are available for intra-muscular injection. The sodium salt of the hemisuccinate ester, also known as (sodium) artesunate is water soluble; it is administered orally or by intravenous injection.

Leaves and flowering tops of *A. vulgaris* are tradi-

tionally used to stimulate the appetite, as a sedative and as a vermifuge. However, the use of excessive doses over long periods may lead to digestive and urinary disorders. A gel containing *A. vulgaris* extract is considered a useful skin care product for dry and pruritic skin conditions. *A. vulgaris* (or a related species) is used in local medicine in India to treat rheumatism. Leaves are used in Chinese medicine as a remedy against haemorrhage and diarrhoea. In Vietnam, a decoction is prescribed to treat menorrhagia.

The buds of *A. capillaris* ('*Artemisiae Capillaris Flos*') have been used since antiquity in Chinese and Japanese medicine, mainly in the treatment of liver and related diseases, e.g. inflammation of the liver, jaundice and cholecystitis. The drug is also applied as cholagogue, anti-inflammatory drug, analgesic, antipyretic and diuretic.

*A. absinthium* L. is used in traditional medicine in several countries, e.g. in India to treat chronic fever, swellings and inflammation of the liver and as a tonic and stimulant. In Cuba it is applied to treat various diseases caused by parasites, whereas formerly in Europe it was used as a digestive, in the treatment of gastritis, against stomach cramps, stomach and intestinal atony, and as anthelmintic. Formerly it was an ingredient of a popular alcoholic drink in France, but its use in absinthe has been banned because of the suspected neurotoxicity of one of the chemical constituents, i.e. thujone. *A. dracuncululus* L. and *A. maritima* L. are used as an aperient, stomachic, stimulant and febrifuge in India. The flowering heads of the latter and *A. cina* Berg ex Poljakov produce santonin, valued as an anthelmintic. In India, *A. nilagirica* (C.B. Clarke) Pampan. is considered an emmenagogue, anthelmintic, stomachic and febrifuge, and is also used to treat skin diseases and ulcers. *A. scoparia* Waldst. & Kit. is a source of scoparone, which exhibits significant hypotensive and tranquilizing activity.

*A. vulgaris* is sold as a vegetable on markets in Sarawak and Thailand.

**Production and international trade** Artemisinin and its derivative artemether are produced commercially in several countries. Artemisinin is extracted industrially from cultivated *A. annua* in Vietnam, and pharmaceutical companies in China and France produce artemether industrially. Artemether is, for instance, marketed in ampules of 80 mg/ml in vegetable oil, to be administered by intra-muscular injection. Artemotil (proposed INN name, previously  $\beta$ -arteether) will be commercially available in 1 ml ampules containing 50

mg or 150 mg per ml sesame oil for intra-muscular injection in cases of severe *Plasmodium falciparum* malaria, as soon as marketing authorisation has been obtained in the Netherlands.

**Properties** Extracts of *A. annua* show anti-malarial activity, which can be attributed to the fraction containing sesquiterpene lactones (based on cadinane and closely related carbon frameworks). The most important active compound of *A. annua* against malaria from this fraction is the endoperoxide artemisinin, but some related compounds from *A. annua* and other *Artemisia* species also show some activity, e.g. arteannuin B and other peroxides such as artemisitene and arteinculton. However, artemisinin has significantly greater activity than the other peroxide compounds, having an  $EC_{50}$  of 0.01  $\mu\text{g}/\text{ml}$ , compared with 1–10  $\mu\text{g}/\text{ml}$  for the other compounds.

Artemisinic acid plays a pivotal role in the biosynthetic pathway of artemisinin. Artemisinic acid originates from mevalonic acid and farnesyl-diphosphate, yielding a cadinane skeleton as a close precursor of this compound. The arteannuin B formed as a result of various processes, one being lactonization, is considered an intermediate in the bioconversion of artemisinic acid to artemisinin. Artemisinin is too complex to be synthesized on a large scale, and it is generally obtained by isolation from plant material. Leaves originating from the Washington (Virginia, United States) area and Europe contain 0.05–0.1% artemisinin (on dry-weight basis), while leaves originating from southern China and the northern provinces of Vietnam contain up to 1.3% artemisinin (on dry-weight basis). A liquid-liquid extraction technique, that allows the use of recovered solvents, has been developed for large-scale production of artemisinin.

Several analytical methods are in use to assay the biosynthetic precursors and metabolites of artemisinin. A simple TLC method is available for screening. HPLC with electrochemical detection, GC-MS and thermospray LC-MS allow very effi-

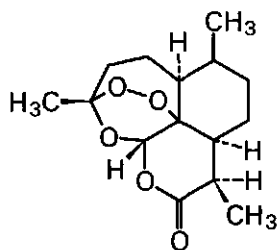
cient detection of artemisinin and structurally related compounds.

Clinical tests in China, Cameroon, the Gambia, the Netherlands, Thailand, Vietnam, Zambia and other countries on volunteers and thousands of patients with severe and non-severe *Plasmodium falciparum* malaria indicated that artemisinin and its derivatives are safe and effective.

For instance  $\beta$ -dihydroartemisinin shows a rapid absorption and distribution, and depending on the dosis, blood levels peak in 1–2 hours, while the biological half-life lies between 1–2 hours too. Multiple dose treatment via the intra-muscular route with artemotil ( $\beta$ -arteether) attained a steady state level after 8–14 hours. The half-life varied between 35–45 hours. The artemisinin group of drugs is only valuable to treat cerebral or *Plasmodium falciparum* malaria.

Malaria is caused by a parasitic *Plasmodium* protozoan, which uses mosquitoes of the genus *Anopheles* as an intermediary host. When an infected mosquito bites a person, sporozoites enter the blood, but they disappear rapidly from the circulation to localize in the parenchymal cells of the liver in which they grow and segment to merozoites. This stage of the infection lasts for 5–16 days, depending on the *Plasmodium* species. On reaching maturity these merozoites are released from the liver cells and penetrate erythrocytes where further division and development takes place. When this process is complete, the erythrocytes burst open and the merozoites enter the blood stream. It is this periodic breaking of erythrocytes that causes the chill so characteristic of malaria. The fever following the chill is due to the liberated foreign protein and cell products. Some of the merozoites infect new blood corpuscles, while others develop into the sexual form, called gametes. The gametes can pass to a healthy mosquito when it bites a person suffering from malaria. The gametes conjugate in the mosquito, forming sporozoites, and the circle is complete.

Artemisinin acts as a so-called blood schizonticide on the asexual erythrocytic stage of the parasites. A critical step in the mechanism of action of artemisinin (and related drugs) seems a hemin-catalysed reduction of the peroxide moiety, resulting in more cytotoxic compounds, such as free radicals and reactive aldehydes that subsequently kill malarial parasites. Membrane damage, alkylation and oxidation of proteins, oxidation of fats, inhibition of the protein and nucleic acid synthesis have been found in these parasites, as well as interaction with cytochrome oxidase and with the



artemisinin

glutamine transport system. The hemin-rich internal environment of the parasites in the erythrocyte is assumed to be responsible for the apparent selective toxicity of artemisinin towards these organisms. Artemisinin rapidly clears the blood from parasites (elimination and improvement of symptoms occur sooner than with chloroquine, and good results have also been obtained with patients who were no longer responsive to chloroquine), but it is inactive against liver stages of the parasite. Due to this spectrum, the drug should not be used as prophylactic; this also greatly reduces the risk that resistance will develop to this new class of antimalarials. Artemisinin and several derivatives have furthermore been found to kill early stages of gametocytes of *Plasmodium falciparum* too. This gametocidal effect may play a role in the interruption of malaria transmission. There are no reports of serious toxicity in humans. Toxicity to the myocardium in macaques after extreme high doses has been reported, while in a test on dogs the highest dose caused deaths. However, when the latter study was repeated under conditions of good laboratory practice, no mortality occurred, and the no toxic effect level was 3 mg/kg in dogs treated daily during 4 weeks with artemotil.

In addition to the antimalarial activity, some other biological activities of artemisinin (and related structures) and of other *Artemisia*-constituents have also been investigated, e.g. cytotoxicity to Ehrlich ascites tumour cells in vitro. All compounds (including artemisinin, artemether and sodium artesunate) showed cytotoxicity, with  $IC_{50}$  values ranging from 12–30  $\mu$ M. The variations in effect between the structurally strongly related compounds mostly correlated well with the theoretical capacity of radical formation and stabilization. Artemisinin, artemisinic acid, arteannuin B, a series of friedelane-type triterpenoids, and the flavonoid quercetagetin-6,7,3',4'-tetramethylether showed positive test results for in vitro cytotoxicity in a series of tumour cell lines (P-388 murine lymphocytic leukaemia, A-549 human lung carcinoma, MCF-7 human breast adenocarcinoma, HT-29 human colon adenocarcinoma and KB human nasopharynx carcinoma). Artemisinin and artesunate have been found effective against experimental schistosomiasis in mice and dogs.

Extracts of *A. annua* showed a strong inhibitory effect on tobacco mosaic virus; the inhibitory agents were identified as the sterols sitosterol and stigmasterol. The extracts also showed in vitro anticoccidial effect against *Eimeria tenella*, which

causes a serious disease in poultry. Artemisinin has allelopathic activity, and inhibits seed germination, seedling growth and root induction of crops such as lettuce and beans.

An extract of 5 g dry powder of *A. absinthium* in 50 ml water, diluted 1:35, showed 90% growth inhibition of *Plasmodium falciparum* in a test in Cuba. An  $LD_{50}$  of 31  $\mu$ g/ml was detected for the sesquiterpene lactone fraction. The test method available for the evaluation of crude extracts assesses the ability of the extract to inhibit [ $G$ - $^3H$ ]-hypoxanthine uptake into *Plasmodium falciparum*.

*A. vulgaris* extracts show insecticidal, insect-repellent, antimutagenic and anthelmintic activities; reports on antimalarial activity are contradictory. The efficacy of a gel containing *A. vulgaris* (or a closely related species) extract has been studied in Japan on 56 patients having pruritic skin lesions. Excellent clinical improvement was obtained in 67% of the cases of pruritic dermatitis, in 56% of atopic dermatitis and 73% of senile xerosis; poor response was observed in 2 cases of contact dermatitis. No side effects were observed.

An aqueous extract of *A. vulgaris* markedly inhibited the growth of both gram-positive and gram-negative bacteria in vitro. It inhibited the growth of the cariogenic bacterium *Streptococcus mutans* considerably. The essential oil from fresh leaves tested at 5000 ppm against the storage fungus *Aspergillus flavus* showed 67% growth inhibition. The dehydromatricaria esters present in the plant showed some antifungal activity, but in general their biological activity is slight. Roots of *A. vulgaris* showed mild toxic activity against the oriental fruit fly. An extract of *A. vulgaris* (particularly of young leaves) inhibits germination and seedling growth of other plants, e.g. of lucerne; it showed some retarding effect on the growth of tea, but it increased the growth of the fungus *Pythium myriotylum*.

The essential oil from several species (e.g. *A. annua*, *A. vulgaris*) is suitable for use in the perfume and cosmetics industry. The oil content of dried flowering parts of *A. annua* is approximately 0.6%. However, *A. vulgaris* normally has a low volatile oil content (0.03%), which accounts for its palatability and digestibility for animals as compared with other *Artemisia* species; the protein content is about 32% and the average in vitro digestibility 67%. *A. vulgaris* is, however, suspected of causing bladder cancer in cattle. The oil content can be much higher in certain types, e.g. in the Philippines where a yield of 0.3% from air-dried

leaves has been reported. The volatile oil is yellowish-greenish with an intense and persistent fragrance. More than 70 compounds, mainly monoterpenes and sesquiterpenes have been identified structurally from the essential oil from flowering parts of *A. vulgaris*. The oil production is seasonally dependent and chemical composition is highly variable; 1,8-cineole, camphor, terpinen-4-ol,  $\beta$ -pinene, (+)- and (-)-borneol, myrcene and vulgarin are invariably present, but thujones ( $\alpha$ - and  $\gamma$ -) are only present in traces or absent. A rather high concentration of thujones is present in oil from *A. absinthium*; habitual use or large doses of absinthe beverages causes absinthism, characterized by neurotoxic symptoms such as restlessness, tremors and convulsions. Whether thujones are the sole cause of these symptoms remains an open question since absinthe formerly also contained cupric sulphate and indigo-based colorants. Almost 50 components were identified in leaves of *A. vulgaris* from Vietnam, the major ones being  $\beta$ -caryophyllene (24%) and  $\beta$ -cubebene (12%).

In *A. capillaris* it is the seed that contains most oil. In the essential oil of *A. capillaris* 25 terpenoids (e.g.  $\beta$ -pinene, limonene and  $\gamma$ -terpinene), 6 phenylacetylenes, 7 phenols and 15 fatty acids have been identified; capillen (a phenylacetylene) is the main component. The main components of drugs prepared from *A. capillaris* are scoparone (6,7-dimethoxycoumarin) and capillarisin. Several other flavonoids and the coumarin 6,7-dimethylesculetin have been identified in the active fraction of the methanol extract. An extract of *A. capillaris* inhibited bovine lens aldose aldehyde reductase and rabbit platelet aggregation; this may be of interest in the prevention of diabetes complications. Scoparone and scopoletin exhibited a potent inhibitory effect on rabbit platelet aggregation, and capillarisin did likewise on bovine lens aldose reductase. Scoparone and capillarisin have choleric action. The flowers, as one of the ingredients in a herbal medicine, helped change the bile flow to almost normal level in  $\alpha$ -naphthyl isothiocyanate-induced cholestasis in rats. In tests with mice, the buds and leaves of *A. capillaris* showed significant protective effect against liver lesions induced by carbon tetrachloride. The active principles were shown to be the flavones eupatolitin and arcapillin. An extract inhibited the adherence of *Streptococcus mutans*, a bacterium which causes dental caries, to teeth surfaces. Tests on isolated rat heart indicate that scoparone possesses anti-anginal action as a vasodilator. Furthermore, kinetic experiments using rabbit thoracic aorta

showed that scoparone has a marked inhibitory effect on the contractions induced by norepinephrine (noradrenaline), 5-hydroxytryptamine, histamine and angiotensin II. Like nitroglycerin, scoparone appeared to be a competitive antagonist of norepinephrine.

Polymers of caffeoylquinic acids are the main polyphenolic components in *A. capillaris*; caffeic acid can be produced by partial hydrolysis of these compounds. Extracts containing these 'caffeetannins' and related compounds have protective action against liver damage. Species used as haemostatic generally also contain caffeetannins.

Extracts of *A. capillaris* have been found to be positive in the chromosomal aberration and micronucleus assays in mice. Leaf and stem extracts of *A. capillaris* showed pronounced nematocidal activity against *Bursaphelenchus lignicolus*. The phenylacetylenes capillen (1-phenyl-2,4-hexadiyne) and 2,4-pentadiynylbenzene (1-phenyl-2,4-pentadiyne) have been isolated from *A. capillaris* roots and buds; capillen inhibits the germination of seeds of, for instance, millet, cabbage and carrot, and both compounds have an antifeeding activity on cabbage butterfly (*Pieris rapae*) larvae, as do certain other minor constituents in growing buds such as capillarin, methyleugenol, ar-curcumene and bornyl acetate. A factor promoting root growth, capillarol, has also been isolated from the leaves; it increased root growth of rice by 80%. *A. capillaris* extracts showed antimicrobial action and were effective in suppressing the growth of food-poisoning bacteria, *Lactobacillaceae* and mycotoxigenic moulds. The flavonoid fraction has been patented for anti-acne treatment and the coumaric fraction for use as a hair stimulant.

The aerial parts of *A. cina*, which contain flavonoids (e.g. hispidulin, quercetin, rutin and caffeic acid), phenol acids and coumarins, and certain other *Artemisia* species indigenous in Russia showed anti-tumour activity in animal tests; these species may be useful in the treatment of Ehrlich carcinoma, breast adenocarcinoma, sarcoma and Walker carcinosarcoma. An aqueous extract of flowering parts of *A. cina* was found to be lethal to larvae of the mosquito *Culex pipiens*; it had an  $EC_{50}$  value of 4 g/l 24 hours after treatment. At a dose of 40 ppm the extract is able to give a significant control of the root-knot nematode *Meloidogyne incognita*. Twenty components have been identified in *A. cina* oil including  $\alpha$ -pinene,  $\beta$ -pinene, myrcene, camphene,  $\beta$ -ocimene, sabinene and limonene.

The large amounts of pollen produced by the



wind-pollinated plants can cause allergic reactions in susceptible persons. Contact dermatitis caused by *A. vulgaris* has been reported.

**Adulterations and substitutes** Synthetic antimalarials derived from quinine are widely used, as are related alkaloids from *Cinchona* spp. The search for new antimalarials in response to the growing resistance of malaria-causing agents to industrial drugs has not only resulted in interest in *A. annua*, but also in interest in certain other promising plant resources used in traditional medicine to treat malaria, e.g. *Azadirachta indica* A.H.L. Juss., *Brucea javanica* (L.) Merr., *Cyclea barbata* Miers and *Dichroa febrifuga* Lour.

**Description** Erect or ascending aromatic annual or perennial herbs or subshrubs, usually densely hairy. Leaves alternate, usually divided or 1-3-pinnate; stipules absent. Flowering heads numerous and small, in spicate, racemose or paniculate inflorescences or sometimes solitary, usually nodding at anthesis, discoid, greenish or yellowish, rather few-flowered; involucre campanulate or subglobose to ovoid, with bracts imbricate in 1-3 series and scarious at margins, the outer bracts usually smaller; receptacle flat or conical to hemispherical, glabrous or pubescent. Flowers of two types, with 1 series of marginal pistillate ray flowers, and bisexual or functionally male tubular disk flowers in the centre of the head; pappus absent; corolla of ray flowers tubular and 2-3-fid, that of disk flowers tubular and 5-fid; stamens 5, inserted on the corolla, with distinct filaments and connate anthers forming a tube around the style, anthers often tipped with acute appendages; ovary inferior, 1-celled, style bifid and exerted in pistillate flowers and often dilated or penicillate in disk flowers. Fruit an obovoid or oblong, terete achene, rounded and with a disk at the apex, striate or 2-ribbed, glabrous or pubescent.

**Growth and development** The life cycle of the Chinese-Vietnamese material of *A. annua*, under natural conditions, is completed within 10 months. Seeds germinate in January to March and fruits can be harvested in October-November. The harvest of plants for extraction of artemisinin takes place in July. Initially, growth is slow and seedlings reach a height of about 5 cm after one month and 25-30 cm after 3 months. Growth is much more rapid from the fourth month. The vegetative period lasts 6-8 months. By August, *A. annua* has become strongly branched, and flower buds become visible. The flowers do not secrete nectar and are wind pollinated; they produce pollen abundantly. The life-cycle of European-

American material of *A. annua* is completed within 6 months. Germination takes place in May, while seeds can be harvested in November.

Chinese-Vietnamese material grown in Europe in the open cannot be planted earlier than May-June because of night frost. In October one can observe elongation of stems and small branches as a prelude to flowering. Flowering is in general frustrated by early night frost and bad weather. Under green-house conditions seed can be produced.

*A. annua* is basically self-fertilizing, but considerable cross-pollination may occur. In some *Artemisia* species, the inner flowers in a flowering head are functionally male and do not set fruit (e.g. in *A. capillaris*). This is also reported for *A. annua*, but in fact the inner flowers do produce seeds, but these are less viable and seedlings often die shortly after germinating.

**Other botanical information** *Artemisia* is in desperate need of a thorough and complete taxonomical revision. Several closely related (or perhaps conspecific) species are often confused in eastern Asia, particularly in the group *A. campestris* L., *A. capillaris* Thunb. and *A. scoparia* Waldst. & Kit. of the section *Dracuncululus*, and in the group of *A. indica* Willd. (synonym: *A. princeps* Pampan.), *A. nilagirica* (C.B. Clarke) Pampan. and *A. vulgaris* L. of the section *Abrotanum*. Consequently, the literature is often difficult to interpret, and information on *A. vulgaris* from eastern Asia probably often refers to other related species. In fact, some authors consider *A. vulgaris* to be a single very variable and widespread species, whereas many others consider it as a complex of up to about 100 closely related species.

*A. annua*, *A. apiacea* (both of the section *Abrotanum*) and *A. capillaris* are sometimes confused. *A. annua* can be identified by its strongly branched panicle and small, subglobose heads. *A. capillaris* is a subshrub, the other two species are annuals.

*A. annua* material from European-American and Chinese-Vietnamese origin shows some clear differences. The European-American type has the ability to produce inflorescences 3-4 months after sowing, while the Chinese-Vietnamese type will not do so when planted in Europe in the open. In order to produce seed of the Chinese-Vietnamese type in Europe, one has to grow it under green-house conditions. Furthermore, the artemisinin content of the Chinese-Vietnamese type is ten-fold higher compared to the European-American type, also when raised under European conditions.

**Ecology** *Artemisia* prefers full sunlight. It is often found in roadsides, waste places and fields. *A. vulgaris* is locally a noxious weed, e.g. in tea plantations.

In cultivation, *A. annua* demands fertile and moisture-retentive soils for optimal growth. It does not tolerate dry conditions or waterlogging, and it usually dies within 2–3 days of flooding. It tolerates neutral to slightly acid soils (pH no lower than 5), and is usually cultivated on rich sandy loams or alluvial soils.

Variation in acetylene content has been found in different ecotypes of *A. capillaris*; the phenylacetynes capillen and capillin were found to be the main constituents in the roots and leaves of plants growing along freshwater rivers, but were only found in the roots of plants growing in a saline environment.

**Propagation and planting** *A. annua* is propagated by seed. In northern Vietnam, seeds are collected in November and sown in February–April or in July–August (southern Vietnam). One gram contains 20 000–22 000 seeds. Before sowing, seeds are soaked in warm water (45–50°C) for 2–3 hours or in a 0.1% gibberellin solution for 15–20 minutes. Under optimal soil moisture and temperature (20–25°C) conditions pretreated seeds start germinating 4–8 days after planting, untreated seeds 10–20 days after planting. The germination rate is usually 50–60% when fresh seed is sown, but drops to 2–3% after 6 months of storage. The usual rate for broadcasting seed in the field is 300–500 g per ha. About 40–50 days after sowing plants are thinned to 20–40 cm × 20–40 cm. Nowadays, seed is preferably sown in nurseries and when seedlings are 15–20 cm tall they are planted into the field at 20–40 cm × 20–40 cm. The latter method of propagation is preferred to direct seeding because it shortens the crop cycle by about 2 months and secures a better and more uniform stand.

Seedbeds are 1–1.2 m wide and 15–20 cm high and provided with a layer of fine-textured topsoil. The application of 5–10 t/ha of green manure or organic manure before planting is beneficial.

**In vitro production of active compounds** Callus from *A. annua* seed has been initiated by transfer onto agar containing Murashige and Skoog basal salts with 5% sucrose, 0.1 mg/l kinetin and 1 mg/l 2,4-dichlorophenoxyacetic acid. The callus cultures were maintained at 25°C under constant illumination and subcultured every 4 weeks. After 3 subcultures, the calli were inoculated into liquid medium of the same ingredients

(without agar) and maintained under the same conditions on a rotary shaker. The cell suspension culture was used for the isolation of constituents. The cell suspension cultures exhibited antimalarial activity in vitro, both in the n-hexane extract of the plant cell culture medium and in the chloroform extract of the cells. Trace amounts of artemisinin may account for the activity of the n-hexane fraction, but only the methoxylated flavonoids artemetin, chrysopenetin, chrysopenol-D and cirsilineol can account for the activity of the chloroform extract. However, the activity of these flavonoids is much lower than for artemisinin.

**Husbandry** *A. annua* responds well to fertilization. N fertilizers are usually applied twice, each time 90–110 kg/ha, the first time about 2 months after direct sowing in the field or 2 weeks after transplanting from the nursery, the second time one month before harvesting. P and K fertilizer is sometimes also applied. However, the artemisinin content of the plants does not increase under these favourable growth conditions. In Vietnam, it has been reported that the artemisinin content of cultivated *A. annua* plants can be comparable to that in plants growing in the wild.

*A. vulgaris* has been intercropped successfully with poplar (*Populus* sp.) in India. It responds well to the application of complete fertilizer (10 g/plant).

**Diseases and pests** Several years of experience with trial plantations of *A. annua* in Vietnam did not reveal serious diseases or pests. Minor pests are ants carrying away seeds after sowing, crickets damaging seedlings, and caterpillars and aphids feeding on the crop.

Weedy *Artemisia* can serve as a host to pathogens and pests that can seriously affect crops. *A. vulgaris*, for instance is a host for cucumber mosaic virus, the worm *Ostrinia nubilalis* and the European corn borer. *A. annua* has been reported as a host of nematodes (*Meloidogyne* spp.).

**Harvesting** The highest leaf yield and the highest foliar artemisinin content in *A. annua* (up to 0.9%) are obtained when the crop is 5 months old. In Vietnam, the harvest is usually in August in the north and in November in the south. Harvesting should preferably be on dry, sunny days.

In Japan, tests with material of *A. capillaris* harvested on different dates showed considerable variation in activity of the drug and in the content of capillarisin and dimethylesculetin. In that country, the capillarisin and 6,7-dimethylesculetin contents reach maximum levels in leaves just before the appearance of flower buds (end of

July), and one month later in the heads (end of August). The best time for harvesting is between the flower bud stage and early flowering, which is from late August to early September in Japan. The flavonoid content of *A. cina* is low during the vegetative period, increases during bud formation and flowering, and decreases again during fruiting.

Small quantities of *A. vulgaris* are usually collected all year round.

**Yield** Yields of cultivated *A. annua* in Vietnam range between 25 and 45 t/ha of fresh material. Yields are generally lower in northern Vietnam than in southern Vietnam: 1.5–2.5 t/ha of dried cleaned material (0.5–0.9% artemisinin) in the north, 2–4 t/ha (0.3–0.6% artemisinin) in the south.

**Handling after harvest** After harvesting, plants are usually sun-dried on brick or cement yards or on asphalt roads for 1–2 days. Broken parts of roots and stems are subsequently separated mechanically, and the remaining material is further dried to below 12% moisture content. Dried leaves of *A. annua* should be stored and packed in jute bags under air conditioning (low relative humidity). In experiments, where the relative moisture of the material varied between 4–16%, the artemisinin content was slightly affected over a period of one year. Material with a moisture content of 16% and stored in jute bags under air conditioning showed a decrease from 5.9–5.2% artemisinin only. Thus, if described storage conditions are maintained, there will be ample time to extract artemisinin before the next crop arrives, while extra electricity costs for air conditioning will be compensated by higher artemisinin extraction yields.

Moreover, there is advantage because of the fact that there is no pressure to extract the crop immediately. As highly volatile extraction solvents are used, chemical extraction can be carried out during the winter season in northern Vietnam and not during summer and autumn when high temperatures complicate cooling of these solvents.

Artemisinin has a low solubility in aqueous and oily solvents, thus causing problems for clinical application. The derivative sodium artesunate is readily water-soluble, and can be processed more easily to medicaments.

**Genetic resources and breeding** Although *A. capillaris* has for centuries been considered useful as a medicinal plant in China and Japan, it has hardly been cultivated, and the crude drug has been largely derived from wild plants. *A. apiacea*

occurs rather scattered and is poorly known. It has been reported that protection of *A. cina* is necessary in parts of Russia because of overcollecting for medicinal purposes. *A. annua* is increasingly being planted for artemisinin production (e.g. in Vietnam and China); in other regions (e.g. Java) it was already being planted for ornamental purposes. *A. vulgaris* is widespread and is not at risk of genetic erosion. There are no records of *Artemisia* species in germplasm banks, and there are no known breeding programmes.

**Prospects** Interest in *A. annua* as an anti-malaria crop is increasing enormously. Tests indicate that artemisinin and its derivatives have a rapid and effective action and low toxicity. To date, no resistance of *Plasmodium* against artemisinin and its active derivatives has been found. It is advisable to control the prescription strictly and not to use these compounds for preventive treatment (which is in any case not self-evident, given that the biological half-life of artemisinin is approximately 4 hours). The therapeutic indications should be provisionally the treatment of severe malaria or when resistance to other antimalarials is suspected. *A. annua* has great prospects in malaria control, and research should continue to optimize its utilization. Artemisinin yields are about 2 kg/ha and should be raised to at least 50 kg/ha to make a cheap anti-malarial.

Several other *Artemisia* species have interesting medicinal properties and deserve more research. For instance, *A. capillaris* and *A. vulgaris* are of interest as an anti-inflammatory for treating skin complaints and for the prevention of dental caries, whereas the former species may be useful in the prevention of complications caused by diabetes and for its protective action against liver damage.

**Literature** [1] Bruneton, J., 1995. Pharmacognosy, phytochemistry, medicinal plants. Lavoisier Publishing, Paris, France. pp. 458–459, 506–508. [2] Ikenaga, T., Hizako, M., Tajima, M. & Nakashima, K., 1994. Production of choleric substances in the capitulum, leaf and stem of *Artemisia capillaris* during the plant growth cycle. *Biological and Pharmaceutical Bulletin* 17(1): 150–151. [3] Kimura, Y., Okuda, H., Okuda, T., Hatano, T., Agata, I. & Arichi, S., 1985. Studies on the activities of tannins and related compounds from medicinal plants and drugs. VII. Effects of extracts of leaves of *Artemisia* species, and caffeic acid and chlorogenic acid on lipid metabolic injury in rats fed peroxidized oil. *Chemical and Pharma-*

ceutical Bulletin 33(5): 2028–2034. |4| Liu, K.C.-S.C., Yang, S.-L., Roberts, M.F., Elford, B.C. & Phillipson, J.D., 1992. Antimalarial activity of *Artemisia annua* flavonoids from whole plants and cell cultures. *Plant Cell Reports* 11(12): 637–640. |5| Luo, X.-D. & Shen, C.-C., 1987. The chemistry, pharmacology and clinical applications of quinghaosu (artemisinin) and its derivatives. *Medical Research Reviews* 7: 29–52. |6| Michaelis, K., Vostrowsky, O., Paulini, H., Zintl, R. & Knobloch, K., 1982. Das ätherische Öl aus Blüten von *Artemisia vulgaris* L. [Essential oil from flowers of *Artemisia vulgaris*]. *Zeitschrift für Naturforschung, Section C*, 37(3/4): 152–158. |7| Nguyen Tien Ban, Le Kim Bien & Vu Xuan Phuong, 1990. The discovery and experimental cultivation of *Artemisia annua* L. for medicaments against malaria. In: *Selected collection of scientific reports on ecology and biological resources. Science and Technics Publishing House, Hanoi, Vietnam*. pp. 213–218 (in Vietnamese). |8| Tan, R.X., Zheng, W.F. & Tang, H.Q., 1998. Biologically active substances from the genus *Artemisia*. *Planta Medica* 64(4): 295–302. |9| Woerdenbag, H.J., Pras, N., van Uden, W., Wallaart, T.E., Beekman, A.C. & Lugt, C.B., 1994. Progress in the research of artemisinin-related antimalarials: an update. *Pharmacy World & Science* 16: 169–180. |10| Yamahara, J., Kobayashi, G., Matsuda, H., Katayama, T. & Fujimura, H., 1989. The effect of scoparone, a coumarin derivative isolated from the Chinese crude drug *Artemisiae Capillaris Flos*, on the heart. *Chemical and Pharmaceutical Bulletin* 37(5): 1297–1299.

#### *Selection of species*

#### ***Artemisia annua* L.**

Sp. pl. 2: 847 (1753).

**Vernacular names** Sweet wormwood (Am). Vietnam: thanh hao, thanh hao hoa v[af]ng, ng[air] si.

**Distribution** Eastern Europe to India, Indo-China, China and Taiwan, naturalized in Japan and North America, sometimes as an adventive plant in western Europe; locally cultivated as an ornamental in Java, but on a larger scale for medicinal purposes in Vietnam and China.

**Uses** An extract is highly valued as a cure for malaria; the isolated active compound (artemisinin) is the basis for commercially traded medicaments. The plant is also used in folk medicine to treat jaundice and anorexia. The seeds are used in

China to treat flatulence, dyspepsia and tuberculosis, and plants in the bud stage in China and Indo-China as febrifuge and to treat boils and skin diseases.

**Observations** An annual branched herb up to 150 cm tall (in cultivation sometimes up to 300 cm tall) with ribbed stem; leaves bipinnatifid or tripinnatifid, up to 12 cm long, with linear, 0.3–1 mm wide pectinately dentate segments, glabrous; heads in rather large panicles, 1.5–2.5 mm long, central flowers bisexual, corolla yellowish; fruit obovoid, 0.6–1 mm long and glabrous. *A. annua* is locally a common weed on waste grounds and in fields. It is reported that only plants from southern China (Sichuan Province) and northern Vietnam contain abundant artemisinin. This intraspecific variation ranges up to a ten-fold higher artemisinin content compared to plants from other regions.

**Selected sources** 10, 11, 12, 38, 39, 97, 193, 246, 361, 394, 724, 748, 872, 878, 903, 1032, 1033, 1035, 1061, 1126, 1130, 1262, 1287, 1585, 1654.



*Artemisia annua* L. – 1, flowering stem; 2, stem base and roots; 3, flower head.

**Artemisia apiacea Hance**

Walp., Ann. bot. syst. 2: 895 (1852).

**Vernacular names** Vietnam: rau bao, thanh cao, th[ar]o cao.

**Distribution** India, Indo-China, China, Korea, Mongolia and Japan.

**Uses** In Vietnam and China, the aerial plant parts are considered febrifuge, haemostatic, tonic and stomachic and prescribed to treat tuberculosis, malaria, epistaxis, anorexia and neurasthenia, and externally to treat furuncles, haemorrhoids and dermatosis.

**Observations** A branched annual or biennial herb up to 150 cm tall; leaves bipinnatifid, up to 15 cm long, with pectinately dentate lanceolate to linear, 1.5–2 mm wide segments, glabrous; heads in panicles, 4–6 mm long, central flowers bisexual, corolla pale yellowish; fruit obovoid, nearly 1 mm long, glabrous. *A. apiacea* is often found along rivers.

**Selected sources** 1035, 1126, 1130.

**Artemisia capillaris Thunb.**

Fl. jap.: 309 (1784).

**Vernacular names** Malaysia: rumput roman (Peninsular). Vietnam: ng[ar]i l[as] kim, nh[aa]n tr[aa]n b[aw]s[c].

**Distribution** India, China, Korea, Mongolia, Japan, the Ryukyu Islands and the Philippines; locally cultivated in gardens in Peninsular Malaysia.

**Uses** The buds have been used since antiquity in Chinese and Japanese medicine as a cholagogue, anti-inflammatory, antipyretic and diuretic in jaundice, inflammation of the liver and cholecystitis; dried plants are imported by Vietnam from China. It has been reported in Peninsular Malaysia that the leaves used to be applied in poultices to cure headache.

**Observations** A much-branched subshrub up to 100 cm tall, stem somewhat woody at base, sterile as well as flowering branches present; leaves bipinnate, up to 9 cm long, with linear-filiform, 0.5–2 mm wide segments, initially densely villous but glabrescent; heads in large panicles, 1.5–2 mm long, central flowers male; fruit oblong, about 0.8 mm long and glabrous. *A. capillaris* closely resembles the widely distributed *A. campestris* L. and *A. scoparia* Waldst. & Kit., but these species differ in respectively their larger and smaller heads. It is especially found along seashores and rivers.

**Selected sources** 202, 645, 737, 743, 744, 745, 959, 1035, 1047, 1075, 1130, 1484, 1606, 1607, 1617, 1618, 1619, 1620.

**Artemisia vulgaris L.**

Sp. pl. 2: 848 (1753).

**Vernacular names** Mugwort (En). Indonesia: baru cina (Sumatra), beunghar kucing (Sundanese), suket ganjahan (Javanese). Malaysia: hiya, bunga ayam hutan bateh, baru cina (Peninsular). Philippines: damong-maria (Tagalog), erbaka (Iloko), gilbas (Cebu-Bisaya). Thailand: kot chulaalamphuaua. Vietnam: ng[ar]i c[uws]u, thu[oo]s[c] c[uws]u.

**Distribution** Native in Europe, continental Asia and North America, introduced and naturalized locally in South-East Asia, e.g. in Java; locally cultivated, e.g. in Peninsular Malaysia and the Philippines.

**Uses** Leaves and flowering tops are used traditionally to stimulate the appetite, as a sedative and as a vermifuge. A gel containing the extract is considered a useful skin care product for dry and pruritic skin conditions. In the Philippines, a decoction or infusion of the leaves is used as a vulnerary, expectorant, stomachic and emmenagogue. In Indonesia, the herb is used as a diuretic, to treat haemorrhoids, diarrhoea and (externally) to treat skin diseases and sores. In Malaysia, it is also used to treat sores. In Thailand, the roots are used as anthelmintic, the leaves as an anti-asthma, antipyretic, expectorant, emmenagogue and to cure diarrhoea; the flowers are used as an anti-asthma and expectorant. In Vietnam, it is considered haemostatic, emmenagogue and stomachic, and is used externally for poulticing ulcers and sores. *A. vulgaris* is used in local medicine in India to treat rheumatism. In Chinese medicine the leaves are used as a remedy against haemorrhage and diarrhoea. Leaves are used to flavour food.

**Observations** A perennial, often ascending and branched herb up to 200 cm tall, with subterranean stolons and grooved stem; leaves pinnatifid to bipinnate, up to 10.5(–14) cm long, with lanceolate, 1–7 mm wide segments, with dense white lanate-arachnoid hairs beneath; heads in panicles with spiciform branches, 3.5–5 mm long, central flowers bisexual, corolla pale green; fruit glabrous, but not developing in Java. *A. vulgaris* is locally a common weed in open localities, in fields and roadsides, in Java at 250–3000 m altitude. *A. vulgaris* is a complex species that many authors have divided into numerous species, but that others consider to represent a single very variable and widespread species. Information on *A. vulgaris* from eastern Asia probably often refers to other related species (mainly *A. indica*?).

**Selected sources** 97, 193, 202, 243, 244, 332,

350, 361, 580, 944, 1008, 1035, 1126, 1130, 1178, 1448, 1572.

Nguyen Tien Ban, Vu Xuan Phuong  
& Charles B. Lugt

### **Belamcanda chinensis (L.) DC.**

Redouté, Liliac. 3: t. 121 (1805).

IRIDACEAE

$n = 16, 64, 2n = 32$

**Synonyms** *Ixia chinensis* L. (1753), *Belamcanda punctata* Moench (1794), *Pardanthus chinensis* (L.) Ker Gawler (1804), *Gemmingia chinensis* (L.) O. Kuntze (1891).

**Vernacular names** Blackberry lily, leopard lily, leopard flower (En). Indonesia: brojo lintang, jamaka (Sundanese), semprit (Javanese). Philippines: abaniko (Tagalog), abanico (Spanish), palma (Spanish, Tagalog). Thailand: waan meetyap (northern), waan haangchaang (Bangkok). Vietnam: x[aj] can, r[er] qu[aj]t, la cho.

**Origin and geographic distribution** *B. chinensis* occurs in northern India, northern Vietnam, eastern China, Korea and southern Japan. It is often planted as an ornamental, and has locally semi-naturalized on a small scale, e.g. in Sumatra, Java, Sulawesi and the Moluccas. It is also planted for ornamental purposes in the Philippines, but has not naturalized there. In China and Japan it is planted as a medicinal plant. It is also locally naturalized in North America.

*B. chinensis* was introduced as an ornamental from China into Great Britain and continental Europe at the end of the 17th Century or early 18th Century, and to North America in the late 18th Century. Now it is a common garden plant in North America and Japan.

**Uses** The rhizome of *B. chinensis* is much used against inflammations of the throat and upper respiratory tract such as laryngitis, pharyngitis, tonsillitis, cough and asthma. It is given for purifying the blood and in Vietnam also against swollen liver and spleen, and to treat snake bites. The rhizome is recommended as an expectorant, antitussive, deobstruent and carminative, and is used in tonics and as a purgative. In Vietnam it is also considered diuretic. It is reported as a remedy for gonorrhoea in Malaysia. In Indonesia, it has been reported to be chewed with *Piper betle* L. leaves after childbirth in Sumatra, and to be used as a poultice to treat lumbago; it is also applied as a medicinal plant in northern Sulawesi. Locally in Peninsular Malaysia the rhizome has been used

in a medicinal bath after childbirth.

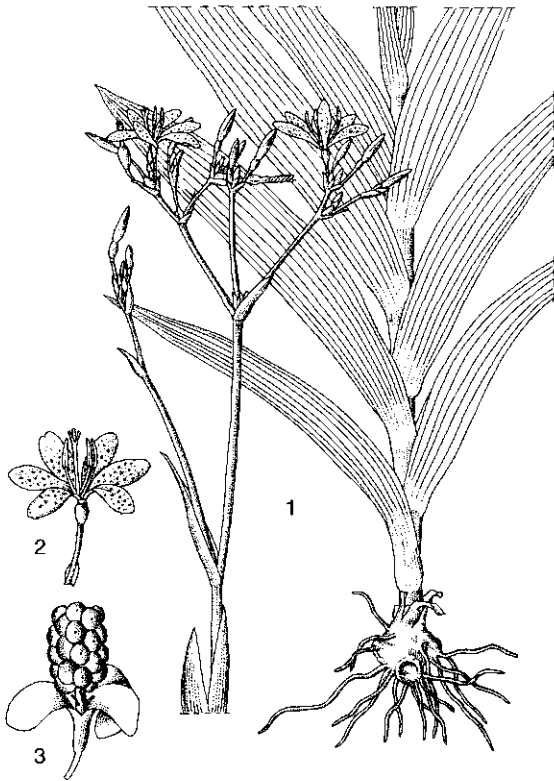
With its attractively blotched orange flowers and its fruits showing the glossy black seeds, blackberry lily is commonly planted as an ornamental in the tropics and in more temperate regions.

**Production and international trade** The trade in rhizomes of *B. chinensis*, often dried and sliced, is mainly from China, but no statistics are available.

**Properties** The rhizomes of *B. chinensis* taste bitter and acrid. Several highly oxygenated isoflavonoids have been isolated from the rhizomes, including tectorigenin, irigenin, iristectorigenin, belamcanidin, methyl-irisolidone, irisfloreantin and noririsfloreantin, which are thought to be responsible for the allergy-inhibiting activity. Furthermore, nine iridals have been isolated, the most important of which is belamcandal (28-acetoxy-14,15-dihydro-26-hydroxy-19-methylidene-spiroirida-15,17-dienal). This compound stimulates the throat membrane, but it is unstable and decomposes during drying processes; no stimulation is thus reported when the dried rhizomes are applied. An aqueous extract of the rhizomes has been screened for possible inhibitory activity against HIV-1 proteases, using a fluorogenic assay. In this test, the effective concentration was found to be 25 µg/ml. The dimeric 1,4-benzoquinones belamcandaquinones A and B have been isolated from the seeds; the first of these compounds showed specific cyclo-oxygenase inhibitory activity. Belamcandal A and B, two alkenyl- (pentadecyl-) phenols and ardisianone A, an alkenyl-1,4-benzoquinone, have also been isolated from the seeds. Using the cytosol of isolated guinea-pig polymorphonuclear leucocytes, belamcandal A and ardisianone A were found to be specific 5-lipoxygenase inhibitors. The belamcandaquinones are probably derived from ardisianone A and belamcandal B.

**Adulterations and substitutes** The rhizome of *Iris* species, such as *I. japonica* Thunb., also contains iridals, has the same activity on the throat membrane and is used for disorders of the throat.

**Description** A perennial, erect, glabrous herb, 50–150 cm tall, with short, creeping, stoloniferous rhizome with round scars of old stems, usually without leaf remnants; stem corymbosely branched in the upper part. Most leaves crowded in the lower half of the stem, distichous and folding over, ensiform and obliquely linear-lanceolate, 20–60 cm × 2–4.5 cm, nearly erect, with long spatheaceous base and somewhat distinct veins,



*Belamcanda chinensis* (L.) DC. - 1, flowering plant habit; 2, flower; 3, dehiscent fruit showing the seeds.

vivid green to glaucous, pellucid-margined; one short leaf present at the base of each branch of stem. Inflorescence a cyme, only top branches with flowers, each cyme with 2 membranous floral sheaths (spathes), (3-)6-12 flowered. Flowers rather small, shrivelling spirally after flowering; pedicel terete, 2-4 cm long, persistent, with indistinct articulation below the ovary; perianth actinomorphic, tepals short connate at base, oblong, 2-3.5 cm long, outer 3 slightly longer than inner 3, clawed, spreading, yellow outside with orange margins, bright orange with dark red blotches inside, outer tepals with longitudinal dark red nectaries at base of upper side; stamens 3, placed before the outer tepals, 1.5-2 cm long, with free filiform filaments and linear, basifixed anthers; ovary inferior, ovoid and slightly trigonous, 8-10 mm long, 3-celled, short-beaked, style about 15 mm long, orange-yellow, 3-fid with style-arms gradually thickened upwards. Fruit an oblong or obovoid, trigonous capsule with 3 deep longitudinal furrows, 1.5-3 cm long, opening with 3 loculi-

cidal valves, many-seeded. Seeds subglobose, 4-5 mm in diameter, glossy black, remaining attached to the central placenta by the raphe for some time after the fruit opens.

**Growth and development** After germination, *B. chinensis* grows slowly but steadily, and may flower within one year of sowing the seed. The flowers are open for a few hours only, from dawn to noon, and 1-2 flowers open at a time within one cyme. It has been reported that apomixis may occur in *B. chinensis*. The plant is short-lived.

**Other botanical information** The monotypic genus *Belamcanda* belongs to the tribe *Irideae*. Morphological, anatomical, cytological and palynological investigations suggest it is particularly closely related to *Iris dichotoma* Pall. An intergeneric hybrid between *B. chinensis* and *I. dichotoma* is available from commercial plant growers; the hybrid is vigorous and partly fertile.

The root tip karyotypes may exhibit inconstancy in their chromosome complement. Cells with  $2n = 28$ , 30 and 32 can be present;  $2n = 30$  has been reported to be most frequent. This type of polymorphism along with aneusomy within different cells of the same tissue has been attributed to the often vegetative means of propagation; it is thought that cells with altered karyotype enter the growing tips of daughter shoots during propagation.

**Ecology** In Java, *B. chinensis* is semi-naturalized in forest edges, brushwood and waste places at 750-2100 m altitude. In Vietnam, it is usually found in savannas. It grows well in the full sun, but also in partial shade. It tolerates moderate frost, but should be protected from severe cold (below  $-15^{\circ}\text{C}$ ). In more temperate regions, *B. chinensis* easily adapts to a wide range of soils.

**Propagation and planting** *B. chinensis* is usually propagated by dividing the rootstock in spring or early autumn in temperate regions, or from seed. In the United States it was found that after stratifying the seed for 60 days at a temperature of  $5^{\circ}\text{C}$  germination occurred one month later. Controls of seed without stratification failed to germinate. Seed can be sown in an equal mixture of *Sphagnum* moss and quartz sand. Seedlings require little care.

**Diseases and pests** Tomato spotted wilt virus can attack *B. chinensis* plants. A mosaic virus found on *Iris fulva* Ker Gawler can also infect *B. chinensis*, causing yellowish streaks and sometimes faint general yellowing of leaves and occasionally downward curling or twisting of young leaves and deforming of fruits. *Pestalotiopsis disseminata* has been reported to cause leaf spots in

India, and the leaf miner *Amauromyza belamcandae* has been reported in Japan. Aphid colonies may cluster on the leaves.

**Handling after harvest** In markets in China rhizomes of *B. chinensis* are sold in hard longitudinal slices which are dark brown outside with transverse markings and a few rootlets, and pale yellowish-brown inside. However, it has been reported that some of the active compounds (e.g. belamcandol) decompose easily on drying and so it is recommended to use fresh rhizomes for medicinal purposes.

**Genetic resources and breeding** *B. chinensis* is planted widespread as an ornamental and locally also as a medicinal plant. It is assumed that the genetic diversity is not endangered, and germplasm collections are not known to exist.

**Prospects** The presence of some active compounds in rhizomes and seed of *B. chinensis* makes it an interesting medicinal plant for home-garden use and perhaps also for large-scale planting. Its ornamental value adds to the promising prospects for planting in South-East Asia, where it seems most suited for mountainous regions. The reported inhibitory activity against HIV-1 protease deserves more research.

**Literature** |1| Abe, F., Chen, R.-F. & Yamauchi, T., 1991. Iridals from *Belamcanda chinensis* and *Iris japonica*. *Phytochemistry* 30(10): 3379-3382. |2| Chimpamba, B.B., 1973. Intergeneric hybridization between *Iris dichotoma* Pall. and *Belamcanda chinensis* Leman. *Cytologia* 38(3): 539-547. |3| de Padua, L.S., Lugod, G.C. & Pancho, J.V., 1981. Handbook on Philippine medicinal plants. Vol. 3. Documentation and Information Section, Office of the Director of Research, University of the Philippines at Los Baños, the Philippines. p. 29. |4| Fukuyama, Y., Kiriya, Y., Okino, J. & Kodama, M., 1993. Belamcandaquinones A and B, novel dimeric 1,4-benzoquinone derivatives possessing cyclooxygenase inhibitory activity. *Tetrahedron Letters* 34(47): 7633-7636. |5| Fukuyama, Y., Okino, J. & Kodama, M., 1991. Structures of belamcandols A and B isolated from the seed of *Belamcanda chinensis*. *Chemical and Pharmaceutical Bulletin* 39(7): 1877-1879. |6| Geerinck, D.J.L., 1977. Iridaceae. In: van Steenis, C.G.G.J. (Editor): *Flora Malesiana*. Series 1, Vol. 8. Sijthoff & Noordhoff International Publishers, Alphen aan den Rijn, the Netherlands. p. 82. |7| Schulze, W., 1971. Die systematische Stellung der Iridaceen-Gattung *Belamcanda* Adans. [The taxonomic position of the Iridaceae genus *Belamcanda* Adans.]. *Feddes Repertorium* 81(8-9): 519-526. |8|

Scott, B., 1987. Plants for the landscape: blackberry lilies. *American Horticulturist* 66(8): 10-11, 13. |9| Xu, H.X., Wan, M., Loh, B.N., Kon, O.L., Chow, P.W. & Sim, K.Y., 1996. Screening of traditional medicines for their inhibitory activity against HIV-1 protease. *Phytotherapy Research* 10(3): 207-210. |10| Yamaki, M., Kato, T., Kashihara, M. & Takagi, S., 1990. Isoflavones of *Belamcanda chinensis*. *Planta Medica* 56(3): 335.

**Other selected sources** 97, 128, 202, 287, 580, 947, 1035, 1178, 1396.

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## Bidens L.

Sp. pl. 2: 831 (1753); Gen. pl. ed. 5: 362 (1754).

COMPOSITAE

$x = 12$ ; *B. bipinnata*:  $2n = 24, 36, 48, 72$ , *B. biter-nata*:  $2n = 24, 48, 72$ , *B. pilosa*:  $2n = 24, 36, 48, 72$ , *B. tripartita*:  $2n = 48, 72$

**Major species** *Bidens pilosa* L.

**Vernacular names** Beggar-tick, bur-marigold (En). Bident (Fr). Indonesia: hareuga (Sundanese), ketul (Javanese). Thailand: noksai.

**Origin and geographic distribution** *Bidens* is a large genus of about 200 species and has a worldwide distribution. Its centres of diversity are located in tropical and subtropical regions of North America and Africa. Only 4-5 species are found within the Malesian area.

**Uses** *Bidens* is widely used in traditional medicine, against numerous complaints, often to soothe pain. Applications may be ascribed to its antiseptic and astringent properties. An infusion or a decoction of roots, leaves or flowers, or the juice of the leaves is used against coughs, headache, fever, constipation, diarrhoea, intestinal worms, stomach-ache, toothache, poisoning, muscular pains and as a bath to treat itching and rheumatic pains. Crushed leaves or flower-heads, sometimes heated over a fire, are applied on the skin to treat inflammations, burns, ulcers, boils or skin affections in general and as a haemostatic on wounds. A decoction of the leaves or the roots is applied on eyelids to treat eye infections. Roots are chewed against toothache, and tinctures of the flowers and leaves are applied as a mouthwash against toothache. Furthermore, the roots and seeds are used as an expectorant, emmenagogue, diuretic, and also against kidney-stones and gall-stones. Seeds are sometimes used as an anaesthetic. An infusion of the leaves of *B. pilosa* is used in diabetes.



In Europe, *B. tripartita* was formerly valued for its diuretic and astringent properties, and used against fevers, gravel stone, bladder and kidney troubles and as a good styptic and remedy for ruptured blood vessels. In North America, the roots and seeds of *B. tripartita* are used as emmenagogue and in laryngeal and bronchial diseases. In Brazil, *B. pilosa* is used in the treatment of malaria.

The flowers of *B. pilosa* are used in the Philippines in the production of a kind of wine called 'sinitsit'. In Mexico, the leaves are used as a substitute for tea as a tonic and stimulant. In Indonesia and in Africa young, 2–5 cm long shoots and young leaves are eaten raw or cooked as a vegetable. They have a bitter astringent taste and are much relished in some regions. *B. pilosa* is readily browsed by domestic livestock, including poultry, and is sometimes used as a fodder. It is said to have a high nutritional value. The flowers are rich in nectar which yields a high-quality, reddish honey. In Thailand, *B. bipinnata* is considered an ornamental. Young shoots of *B. biternata* are eaten raw or steamed. Plants of *B. tripartita* yield a black dye which is used as a hair dye in China.

**Production and international trade** In most regions *B. pilosa* is available as a weed to meet daily needs. In Indonesia, young shoots are for sale on local markets but no statistics are available.

**Properties** Extracts of *B. pilosa* show anti-malarial activity both in vitro and in vivo. The crude ethanol extract from *B. pilosa* (50 µg/ml) causes up to 90% inhibition of *Plasmodium falciparum* growth in vitro, compared with respectively 86–94% and 68–79% inhibition for the chloroform and butanol fraction (both at 50 µg/ml). In vivo, the crude ethanol extract and the chloroform fraction cause about 40% reduction of *P. berghei* parasitaemia in mice. Phenylacetylenes and flavonoids have been found in the ethanol extract from the leaves and the roots. The major component of the chloroform fractions from the roots was the phenylacetylene 1-phenylhepta-1,3-diyne-5-en-7-ol-acetate. Other *Bidens* species with aliphatic and phenylacetylenes and related compounds (thiophenes) were also found to be very active in vitro: 50 µg/ml of *B. tripartita*, which contains 13 acetylenes, reduced *P. falciparum* growth by 87%, and a similar dose of *B. bipinnata*, which has 9 acetylenes, gave a 70% reduction. However, extracts of *B. biternata* containing only 3 acetylenes showed only 38% inhibition at the same dose. The results indicate that the anti-

malarial activity of *Bidens* may be attributed to the presence of acetylene compounds. The therapeutic usefulness of these compounds seems limited, since they are easily oxidized by air and light. Furthermore, acetylenes also have antimicrobial activity. A number of polyacetylenes, e.g. phenylhepta-1,3,5-triyne from petroleum ether and methanol/water extracts of *B. pilosa* are toxic to yeasts and some bacteria. This compound, which can also be isolated from the aqueous methanolic extract of leaves, flowers and achenes of *B. bipinnata* is an active anti-parasitic and exhibited marked insecticidal activity with LC<sub>50</sub> of 204 ng/cm<sup>2</sup> for the first instar larvae of the fall armyworm (*Spodoptera frugiperda*).

In literature, the polyacetylene 7-phenylhepta-2,4,6-triyne is reported phytotoxic to fibroblast cells. The polyacetylene β-D-glucopyranosyloxy-3-hydroxy-6(E)-tetradecen-8,10,12-triyne from *B. pilosa* shows overgrowing action against normal and transformed human cell lines in culture. Dried leaves of *B. pilosa* have a co-carcinogenic action for oesophageal tumours induced in rats. Consumption of the leaves, as in South Africa, has been found to promote the development of oesophageal cancer.

In addition to the acetylenes, other compounds such as phytosterols (β-sitosterol), triterpenes (friedelin and friedelan-3β-ol) and caffeic acid(s) are also reported from *B. pilosa*. The main flavonoids from leaf extracts of *B. pilosa* are aurones and chalcones. Since friedelin and friedelan-3β-ol, as well as several flavonoids have known anti-inflammatory properties, their detection in extracts from *B. pilosa*, together with the presence of the described acetylenes, may rationalize the use of *B. pilosa* in traditional medicine, especially for treating wounds, against inflammations and against bacterial infections of the gastro-intestinal tract.

The ethanolic extract of *B. pilosa* showed a very high inhibition of prostaglandin synthesis in an in vitro assay for cyclo-oxygenase inhibitors. The methanol extract of *B. pilosa* showed radio-protective activity for bone marrow. Besides the above-mentioned pharmacological activity, antihyperglycaemic, immunomodulator, anti-ulcer and hypotensive activity are reported.

**Adulterations and substitutes** 1-Phenylhepta-1,3-diyne-5-en-7-ol-acetate has also been isolated from *Coreopsis* species.

**Description** Annual or perennial, usually erect herbs; stem branched, terete to 4-angled. Leaves opposite or rarely whorled, upper leaves some-

times alternate, simple to deeply 2–3-pinnatisect, margins entire to dentate or serrate or variously incised, sessile or petiolate; stipules absent. Inflorescence a terminal or axillary, capitulum, solitary or arranged in corymbose or panicle-like cymes; receptacle flat to conical, set with scales (paleae); involucre campanulate to hemispherical, 2-seriate, outer involucre bracts often much larger than the inner ones. Ray flowers absent or present, sterile or female, ligulate, yellow, purple to lilac or white; disk flowers bisexual, with yellow to brownish-orange or purple, tubular, 4–5-toothed corolla; stamens 4–5, anthers fused brown to black, caudate to sagittate at base; style bifurcate with short to long arms. Fruit a dorsiventrally compressed or 3–4(–6)-angled achene, linear-oblong to ellipsoidal or broadly obovate, not beaked, margins setulose, thickened or sometimes winged; pappus absent or composed of up to 4(–5) usually barbed bristles. Seedling with epigeal germination; cotyledons free, strap-shaped to spatulate; hypocotyl elongated; leaves opposite.

**Growth and development** *B. pilosa* produces seed (achenes) abundantly; one infructescence can produce 50–70 seeds, one plant up to 6000. The seed has no dormancy and germinates within 3–4 days. Seed viability is high and even 3–5-year-old seeds still have a viability of 80%. Phytochrome controls germination in *B. pilosa* and seeds germinate in darkness when the level of the pre-existing active form of phytochrome is above a certain threshold. The optimum range in day/night temperatures for germination is 25/20°C–35/30°C at a photoperiod of 12 h. Temperatures below 15/10°C and above 45/40°C influence germination negatively. Flooding following seeding, even for a day, will reduce emergence to about 25%. Seedling emergence decreases further sharply with extended periods of flooding. In some areas 3–4 generations per year are possible, making *B. pilosa* often a noxious weed in cropped land. Flowering starts about 1.5–2 months after sowing; plants are self-fertile and seed is mature 1 month after flowering. Flowering and fruiting is throughout the year. The seed is easily distributed by animals and people because of the barbed bristles of the pappus which adhere to fur or to clothes. The effective way of dispersal has contributed to *B. pilosa* developing into a worldwide weed. Plants have the highest biomass of leaves at about flowering time.

**Other botanical information** *Bidens* belongs to the tribe *Heliantheae* and is closely related to *Coreopsis*, with which some authors advocate

uniting it. The taxonomy of *Bidens* is still unsatisfactory.

Due to its worldwide distribution, *B. pilosa* is a highly variable species: plants are erect or decumbent, leaves are simple to highly dissected with entire to dentate-serrate margins, heads may be discoid or radiate, ray flowers may be yellow, white or pinkish and short to long, the achenes may be awnless or have 2–5 bristles. In the past *B. pilosa* has been subdivided into 7 varieties, some with a number of formae. However, in America and South Africa it has been discovered that most of the taxa distinguished may occur in one population. Subdivision of the species is no longer considered useful, particularly because different ploidy levels seem to play a role in addition to the morphological variation. It is possible that what is now considered to represent *B. pilosa* in South-East Asia in fact consists of several species, as has appeared to be the case in North and Central America, but more biosystematic research is needed.

*B. bipinnata* closely resembles *B. biternata* and has been confused occasionally in literature because of erroneous identification of the material.

**Ecology** All South-East Asian *Bidens* species are known as weeds of cropped land and appear also in roadsides, along watercourses, in brushwood and thickets, up to 2500 m altitude. They prefer sunny to slightly shaded places and moist soils. *B. pilosa* is a cosmopolitan weed of more than 30 crops and often becomes dominant after the eradication of perennial grasses. It displays allelopathic effects on a number of crops.

**Propagation and planting** Optimal emergence occurred when planting seeds less than 1 cm deep, with no emergence when planted as deep as 10 cm.

**Harvesting** Leaves and flower-heads of *Bidens* are simply collected from the wild whenever the need arises. Plants are readily available in the surroundings of human habitation.

**Genetic resources and breeding** In view of its widespread distribution and weedy nature, *Bidens* occurring in South-East Asia is unlikely to be at risk of genetic erosion.

**Prospects** The traditional application of *Bidens* for its antiseptic and anti-inflammatory properties will remain of local importance, especially because plants are always readily available. The antimalarial properties deserve further attention.

**Literature** |1| Alvarez, L., Marquina, S., Villareal, M.L., Alonso, D., Arranda, E. & Delgado, G., 1996. Bioactive polyacetylenes from *Bidens* pi-

losa. *Planta Medica* 62(4): 355–357. |2| Ballard, R., 1986. *Bidens pilosa* complex (Asteraceae) in North and Central America. *American Journal of Botany* 73: 1452–1465. |3| Brandao, M.G.L., Krettli, A.U., Soares, L.S.R., Nery, C.G.C. & Marinuzzi, H.C., 1997. Antimalarial activity of extracts and fractions from *Bidens pilosa* and other *Bidens* species (Asteraceae) correlated with the presence of acetylene and flavonoid compounds. *Journal of Ethnopharmacology* 57: 131–138. |4| Geissberger, P. & Sequin, U., 1991. Constituents of *Bidens pilosa* L.: do the components found so far explain the use of this plant in traditional medicine? *Acta Tropica* 48(4): 251–261. |5| Holm, L.G., Plucknett, D.L., Pancho, J.V. & Herberger, J.P., 1977. The world's worst weeds. Distribution and biology. East-West Center, University Press of Hawaii, Honolulu, United States. pp. 185–189. |6| Jager, A.K., Hutchings, A. & Van Staden, J., 1996. Screening of Zulu medicinal plants for prostaglandin-synthesis inhibitors. *Journal of Ethnopharmacology* 52(2): 95–100. |7| Mesfin, T., 1993. An account of *Bidens* (Compositae: Heliantheae) for Africa. *Kew Bulletin* 48: 437–516. |8| Mirvish, S.S., Salmasi, S., Lawson, T.A., Pour, P. & Sutherland, D., 1985. Test of catechol, tannic acid, *Bidens pilosa*, croton oil, and phorbol for cocarcinogenesis of esophageal tumors induced in rats by methyl-*n*-amyl nitrosamine. *Journal of the National Cancer Institute* 74(6): 1283–1290. |9| N'Dounga, M., Balansard, G., Babadjamian, A., David, P.T. & Gasquet, M., 1983. Study on *Bidens pilosa* L. Identification and antiparasitic activity of 1-phenyl-1,3,5-heptatriyne. *Plantes Médicinales et Phytothérapie* 17: 64–75. |10| Wat, C.K., Biswas, R., Graham, E., Bohm, L. & Towers, G.H.N., 1978. UV-mediated antibiotic activity of phenylheptatriyne in *Bidens pilosa*. *Planta Medica* 33(3): 309–310.

*Selection of species*

***Bidens bipinnata* L.**

Sp. pl. 2: 832 (1753).

**Synonyms** *Bidens pilosa* L. var. *bipinnata* (L.) Hook.f. (1881).

**Vernacular names** Spanish needles (En). Bident bipenné (Fr). Vietnam: v[a]n th[oj] t[aa]y, song nha k[es]p.

**Distribution** Native to North America and eastern Asia; introduced and occurring as a weed in southern Europe, Central and South America, Africa, Australia and elsewhere in Asia. In Male-

sia, only known from the Philippines.

**Uses** The warmed juice of the leaves is used to treat conjunctivitis, against earache and as a styp-tic on wounds. The roots and seeds are used as an emmenagogue, expectorant, stimulant and anti-spasmodic, and also to treat asthma. In China, *B. bipinnata* is used to treat stings of insects, snake bites, and unhealthy granulations of wounds. In Taiwan, a decoction of the entire plant is used as an antidiarrhetic.

**Observations** An annual, erect herb up to 1.5(–2.5) m tall, stem 4-angular, glabrous or minutely hispid in the upper part; leaves opposite, occasionally alternate towards the apex, bipinnatisect with the lower segment often 2–3-cleft or pinnatifid, 11–20 cm long, the segments ovate to deltoid or the terminal one lanceolate, margin crenate-serrate, petiolate; capitula in lax paniculate cymes, radiate, 4–6 mm broad, outer involucre bracts 7–10, 3–5 mm × 0.5 mm, shorter than the inner ones; ray flowers 3–5, corolla 5–6 mm long, yellow, disk flowers with yellow, 4–5 mm long corolla; achene linear-fusiform, 4-angular, 7–18 mm long, with (2–)4 retrorsely barbed bristles of 2–4 mm. *B. bipinnata* is found on roadsides, wasteland, and field margins, up to 1400 m altitude; in the Philippines it has been found on dry slopes at 1300 m.

**Selected sources** 289, 458, 852, 935, 938, 1126, 1329.

***Bidens biternata* (Lour.) Merr. & Sherff**

Bot. Gaz. 88: 293 (1929).

**Synonyms** *Coreopsis biternata* Lour. (1790), *Bidens chinensis* Willd. (1804), *Bidens abyssinica* Sch. Bip. (1846).

**Vernacular names** Indonesia: hareuga (Sundanese), ketul (Javanese), daun jarong (Malay, Moluccas). Thailand: koncham (Nakhon Ratchasima).

**Distribution** Widely distributed in tropical and subtropical regions of Africa, Asia including the Malesian region, and Australia; introduced and locally naturalized in temperate Europe.

**Uses** The leaf juice is used to treat eye and ear affections. The rubbed leaves are applied to skin affections in general, as a haemostatic on wounds, and wrapped around the umbilical cord of babies. The seeds are applied as an anthelmintic in animals.

**Observations** An annual, erect herb up to 1.5(–2) m tall, stem 4-angular, glabrous to pubescent; leaves opposite or rarely alternate towards the apex, pinnately (3–)5–9-lobed, 9–19 cm long,

glabrous to densely tomentose, the segments ovate to ovate-lanceolate, the lower ones often pinnatifid, crenate-serrate or rarely lobulate-dentate, petiolate; capitula in lax paniculate cymes, usually radiate, 5–7 mm broad, outer involucre bracts (4–)5–10(–15), (3–)4–7(–12) mm × 0.4–0.7 mm, much narrower than the inner ones; ray flowers (0–)2–5, sterile, corolla 3–6 mm long, yellow, disk flowers with yellow, 3–5.5 mm long corolla; achene linear, 4–8-ribbed, 6–16(–20) mm long, with (2–)3–4(–5) retrorsely barbed bristles of (1–)2–4 mm long. *B. biternata* is common and occurs in fields, gardens, roadsides, thickets, teak forest and along watercourses, up to 2300 m altitude.

**Selected sources** 97, 202, 289, 580, 852, 937 938, 1126, 1329, 1380.

### *Bidens pilosa* L.

Sp. pl. 2: 832 (1753).

**Synonyms** *Bidens sundaica* Blume (1826), *Bidens leucorrhiza* (Lour.) DC. (1836), *Bidens pilosa* L. var. *minor* (Blume) Sherff (1925).

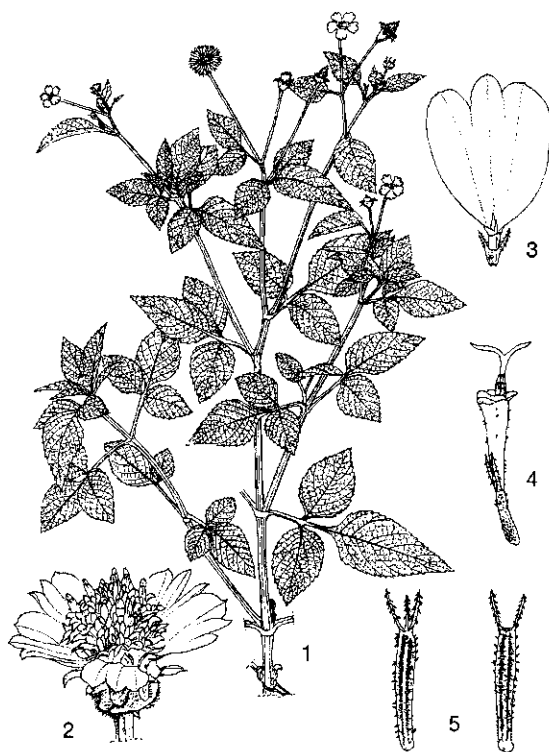
**Vernacular names** Black jack (En). Sornet (Fr). Indonesia: ajeran, hareuga (Sundanese), jaringan, ketul (Javanese). Malaysia: kancing baju, pau-pau pasir, keroten. Papua New Guinea: ivu na mag (Gunantuna, New Britain), rakot (Kurtatchi, Bougainville). Philippines: dadayem (Ibanag), burburtak (Ilocano), pisau-pisau (Bisaya). Thailand: puen noksai (northern), kee nok sai, yaa koncham khaao (central). Vietnam: d[ow]n bu[oo]st, t[uwr] t[oo] hoang, q[ur]y tr[aa]m th[ar]o.

**Distribution** *B. pilosa* originates from tropical America but is now distributed and naturalized as a weed in most tropical and subtropical regions of the world, even sometimes extending into some temperate areas. In South-East Asia it is common in many places, except in Kalimantan and the Moluccas.

**Uses** *B. pilosa* is widely used in traditional medicine against numerous complaints, often to soothe pain. An infusion or decoction, or the juice of the leaves is used against coughs, angina, headache, fever, diabetes, constipation, diarrhoea, intestinal worms, stomach-ache, toothache, poisoning, muscular pains and as a bath to treat itching and rheumatic pains. Crushed leaves, sometimes heated over a fire, are applied on the skin to treat inflammations, burns, on wounds to stop bleeding and on ulcers. In Papua New Guinea, crushed flower-heads are often used externally to extract pus from boils. A decoction of the leaves or

of the roots is applied on eyelids to treat eye infections. A tincture of the flowers and leaves is used as a mouthwash against toothache. Roots are chewed against toothache. In Indonesia, the Philippines and Africa, young shoots and young leaves are eaten raw or cooked as a vegetable. Leaves as vegetable in the daily diet have been observed to prevent goitre in the Philippines. The plant is sometimes used as fodder.

**Observations** An annual, usually erect herb up to 1(–2) m tall, stem 4-angled, glabrous or sparsely pubescent; leaves opposite, pinnately 3–5-lobed, occasionally the lower and/or upper leaves simple, up to 15(–20) cm long, glabrous or sparsely pubescent on both surfaces, margin usually serrate or crenate-serrate, the segments ovate to ovate-lanceolate, the terminal one largest, petiolate; capitula solitary or in lax paniculate cymes, usually radiate, 5–12 mm broad, outer involucre bracts 7–10, spatulate, reflexed at anthesis, 3–4 mm × 0.5–0.8 mm, inner ones ovate-lanceolate; ray flowers absent or 4–8, sterile, corolla 7–15 mm long, white to yellow or pinkish, disk flowers with 3.5–5



*Bidens pilosa* L. – 1, flowering and fruiting plant; 2, flowering head; 3, ray floret; 4, disk floret; 5, achenes.

mm long, yellow corolla; achenes linear, 4–6-ribbed, 4–13 mm long, with 2–3(–5) retrorsely barbed bristles of 2–4 mm long. *B. pilosa* is a very common weed of sunny, often disturbed places like roadsides, fields, thickets and along water-courses, up to 2500 m altitude.

**Selected sources** 52, 53, 85, 97, 111, 168, 180, 184, 202, 224, 332, 350, 458, 475, 580, 597, 598, 599, 610, 614, 663, 818, 852, 920, 937, 1035, 1124, 1126, 1128, 1178, 1329, 1380, 1386, 1543, 1551, 1572.

### ***Bidens tripartita* L.**

Sp. pl. 2: 831 (1753).

**Vernacular names** Trifid bur-marigold, three-lobed butterbur (En). *Bident tripartiti* (Fr). Vietnam: th[ur]ly song nha.

**Distribution** From Europe and northern Africa to the Himalayas, China, Japan, Taiwan, Vietnam, the Philippines, West Java and New Guinea (Irian Jaya).

**Uses** In China, a decoction of *B. tripartita* is recommended in treating chronic dysentery, heart ailments, as a refrigerant and as a wash to treat chronic eczema. The roots and seeds are used as emmenagogue, expectorant, diuretic and also against kidney-stones and gallstones. *B. tripartita* also yields a black dye which is used as a hair dye in China.

**Observations** An annual, erect herb up to 1(–1.5) m tall, stem 4-angular, glabrous or sparsely pubescent; leaves opposite, the upper ones sometimes alternate, pinnately 3–5-lobed or rarely entire, up to 13 cm long, sparsely pubescent below, margins coarsely serrate, terminal segment larger, sessile or on a short and winged petiole; capitula solitary, not radiate, 10–20 mm broad, outer involucre bracts 5–10, foliaceous, oblanceolate, 10–35(–45) mm long, inner ones ovate-lanceolate, 6–12 mm long; ray flowers absent, disk flowers with 4–4.5 mm long, yellow corolla; achene narrowly obovate, compressed, 6–11 mm long, with 2(–3) retrorsely barbed bristles. *B. tripartita* occurs in moist, open habitats like watersides, swampy pastures and paddy fields, up to 1700 m altitude.

**Selected sources** 97, 184, 259, 289, 508, 852, 1126, 1329.

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### ***Blumea* DC.**

Guill., Arch. Bot. (Paris) 2: 514 (1833).

COMPOSITAE

$x = 9, 10, 11$ ; *B. balsamifera*:  $2n = 18, 20$ , *B. lacera*:  $2n = 18, 20, 22, 36$ , *B. lanceolaria*:  $2n = 18, 20, 54$

**Major species** *Blumea balsamifera* (L.) DC., *B. lacera* (Burm.f.) DC., *B. lanceolaria* (Roxb.) Druce.

**Vernacular names** Indonesia: sembung (general), capo (Sulawesi), capa (Sumatra). Malaysia: sembung (general), chapa. Philippines: sambong (Filipino). Laos: 'nat. Thailand: naat.

**Origin and geographic distribution** *Blumea* comprises about 50 species, most of which are confined to tropical Asia from Sri Lanka to China and the Malesian region, some extending to Africa in the west, Australia in the south, and the Pacific as far as Hawaii in the east. Some of the species reported from Africa may, however, well belong to other related genera. The majority of the species occur in South-East Asia, which seems the most likely centre of origin. *Blumea* is found throughout the Malesian region, with the Philippines being the richest with 19 species (4 endemics), followed by Indonesia with 18 (2 endemics), New Guinea with 13 (2 endemics) and Peninsular Malaysia with 6 species.

**Uses** A decoction of leaves or flowers of several *Blumea* species is used throughout the Malesian region to treat asthma, bronchitis and catarrhal affections in general. The leaf juice of several *Blumea* species is applied in various countries of the Malesian region to treat sores, boils, aphthae and sore eyes, probably due to its astringent properties. The leaves of *B. balsamifera* have been used in Chinese medicine since ancient times as a carminative, mild stimulant, vermifuge and as topical application for septic ulcers. In South-East Asia it is one of the most common and widely used medicinal plants for a number of ailments, mainly as a stomachic, antispasmodic, vermifuge and sudorific. In the Philippines, a diuretic and kidney-stone medicine is prepared commercially from *B. balsamifera* leaves. In Thailand, cigarettes containing the chopped, dried leaves are smoked to relieve the pain of sinusitis, whereas a decoction of fresh leaves, alone or in combination with other plant preparations, is used as a bath for women after childbirth and also for young children. In northern Vietnam leaves are also used as a medicinal condiment. A decoction of the roots of *B. riparia* is drunk to treat colic and the leaves and roots of *B. arfakiana* are similarly used in the

treatment of stomach-ache. Furthermore, *B. lacera* is used in the treatment of haemorrhages and as an anthelmintic, febrifuge, deobstuent, diuretic and stimulant. *B. lanceolaria* is also used as a sudorific and applied externally as a poultice against rheumatism. The young leaves of *B. arfakiana* are eaten to treat anaemia.

*B. balsamifera* yields a high quality camphor oil, known as 'ngai camphor'. Camphor is used in liniments against rheumatic pains; a 3% ethanol solution is used to sooth itching. *B. lacera* also contains a camphor-like oil which is reputedly stronger than that of *B. balsamifera*. Its leaves can be eaten as a vegetable. *B. lanceolaria* is sometimes cultivated in gardens for culinary purposes as a seasoning.

**Properties** *B. balsamifera* is well known for its medicinally important essential oil. Plants from the Philippines yield about 0.1–0.4% essential oil, whereas those from Burma (Myanmar) are reported to contain 1.9% oil. The high quality *B. balsamifera* oil ('Ngai camphor') from Chinese sources consists almost entirely of (–)-borneol (1-borneol), that of Burmese origin consists of 75% of (–)-camphor (1-camphor) and 25% of (–)-borneol. These or related compounds can be found in varying concentrations in the essential oils of other *Blumea* species too. *B. lacera* from India yields 0.085% of an essential oil containing 'Blumea-camphor', which is probably a mixture of camphor, borneol and/or related components. The main constituent of *B. lacera* essential oil from Nigeria was thymoquinol-dimethyl-ether. The essential oil of *B. lanceolaria* consists for 95% of methyl-thymol; in another sample, however, p-cymene was identified as the major component (99%).

Phytochemical investigations have furthermore revealed the presence of flavonoids (5-hydroxy-3,6,7,3',4'-pentamethoxyflavone, 5,3',4'-trihydroxy-3,6,7-trimethoxyflavone, and a very small quantity of another flavone) in the leaves of *B. lacera*. Campesterol (a sterol) has also been isolated from the aboveground parts. Two glycosides, the triterpenoid glycoside 19 $\alpha$ -hydroxy-urs-12-ene-24,28-dioate-3-O- $\beta$ -D-xylopyranoside and the phenol glycoside 2-isoprenyl-5-isopropylphenol-4-O- $\beta$ -D-xylopyranoside have been isolated from the whole plant of *B. lacera*.

The flavonoid blumeatin (5,3',5'-trihydroxy-7-methoxy-dihydro-flavone) has been isolated from *B. balsamifera*. Intraperitoneal injections of this component in CCl<sub>4</sub>-intoxicated rats inhibited the increase of serum alanine aminotransferase (ALAT, SGPT), liver triglyceride level and in-

creased serum triglyceride,  $\beta$ -lipoprotein, and liver glycogen content. The histological lesions of the liver of treated rats were less severe than those of their hepatic injury control. After intraperitoneal injection blumeatin also shortened the pentobarbital sleeping time in CCl<sub>4</sub>-intoxicated mice. In thioacetamide-intoxicated mice, intraperitoneal injection of the compound inhibited the increase of serum alanine aminotransferase and liver triglyceride. These findings suggest that blumeatin could protect the liver against injury induced by CCl<sub>4</sub> and thioacetamide. Other flavonoids, flavone and quercetin derivatives have been reported in addition to blumeatin. Three sesquiterpene lactones isolated from *B. balsamifera* showed antitumour activity against Yoshida sarcoma cells in tissue culture. An extract from *B. balsamifera* reduced the mutagenicity potential of mitomycin C, dimethylnitrosamine and tetracycline in mice, and exhibited antimutagenic effects.

Other biological activities include antihistamine release properties by *B. balsamifera*, and antifungal activity of the ethanolic extract of *B. balsamifera* against *Epidermophyton floccosum* with a minimum inhibitory concentration of less than 10 mg plant material/ml. Water extracts of the entire plant showed a diuretic activity similar to coffee and tea.

The essential oil of *B. lacera* has a synergistic effect on the insecticidal activity of pyrethrum; the essential oil alone does not have insecticidal activity. *B. balsamifera* has insecticidal properties against the yponomeutid crucifer pest *Plutella xylostella*, but the compound responsible for the insecticidal activity is not known. It also effectively protected stored garlic against a number of insect pests. Moreover, when topically applied its essential oil is toxic to a number of insects and to golden snails (*Pomacea* spp.).

**Adulterations and substitutes** Essential oils of several other plant groups have similar applications and contain related compounds. Examples for camphor and borneol, without specification of stereo specificity, can be found in *Cinnamomum camphora* (L.) J.S. Presl and in *Compositae* like *Achillea* and *Artemisia*.

**Description** Herbs or shrubs, up to 4 m tall; stem simple or branched, erect to ascending, sometimes sprawling. Leaves alternate, simple, often pinnately lobed, linear-lanceolate to oblanceolate, margin serrate, dentate or denticulate, base usually rounded to tapering, sessile or shortly petiolate, exstipulate. Inflorescence a discoid capitulum arranged in lax to compact corymbs or

panicles or occasionally solitary; receptacle honey-combed, epaleate, glabrous or hairy; involucre campanulate to hemispherical, involucre bracts multiseriate, narrow, pubescent on the back, outer ones much shorter than inner ones. Flowers with a tubular corolla, often yellow, rarely white or pale purple; marginal flowers female, corolla filiform, 2–4-lobed, in several rows; disk flowers bisexual, corolla (4–)5-lobed, stamens (4–)5, alternating with the corolla lobes, tailed at base, ovary inferior, style exserted, bifid. Fruit an oblong achene, terete or obscurely 4-angled, with 5 or 10 ribs; pappus uniseriate, composed of numerous, slender, toothed bristles, white or yellowish-white to red. Seedling with epigeal germination; paracotyledons free, opposite; hypocotyl not elongated; first pair of leaves opposite, subsequent leaves alternate.

**Growth and development** *B. balsamifera* is evergreen. Pollination is probably by insects. In New Guinea, *B. arfakiana* flowers from March to October, and *B. arnakidophora* from September to January. *B. balsamifera*, *B. lacera* and *B. riparia* flower throughout the year.

**Other botanical information** *Blumea* belongs to the tribe *Inuleae* and seemed to be closely related to the genera *Laggera* and *Blumeopsis*. It has even been proposed to merge these three genera, but recent studies place the latter two in the tribe *Plucheeae*. There has been considerable confusion about the identity of *B. chinensis* and *B. pubigera*. Most literature on these species pertains to *B. riparia*.

**Ecology** Most *Blumea* species have a weedy habit and are found in various ruderal and strongly secondarized habitats such as roadsides and fields, in the lowland and mountainous regions up to 3000 m altitude. Most *Blumea* species can tolerate drought and are found in regions with a slight to pronounced dry season. *B. balsamifera* tolerates fire, after which it readily sprouts from underground parts.

**Propagation and planting** *B. balsamifera* can be propagated by seed and by root or stem cuttings. Cuttings are placed in containers under shade. Water should be given with care, as too much watering is harmful. After about 2 weeks the plants can be transplanted in a place receiving full sunlight. After transplanting into the field, young plantings should be weeded regularly. Other *Blumea* species are usually propagated by seed only. 100 achenes of *B. lacera* weigh 4.8 mg; their germination rate is about 95%, falling to about 10% when stored for 12 months. The optimum

temperature for germination is 30°C, but achenes can germinate at 20–50°C.

**Husbandry** In general, *Blumea* species are considered weeds, so more effort is geared towards eradication rather than towards cropping. Full sunlight is optimal for *B. balsamifera* for growth and for the production of essential oil. In the Philippines it responds well to a fertilizer application of 40 g ammonium sulphate or 100 g solophos (0-18-0) per plant.

**Diseases and pests** In the Philippines *B. balsamifera* suffers from leaf rust caused by *Endophyllum blumeae*, resulting in premature defoliation when the attack is severe. During the rainy season circular leaf spot caused by *Cercospora* sp. may lead to serious losses. Orange leaf spot also caused by a *Cercospora* sp. occurs occasionally. In Indonesia, *Gloeosporium* sp. has been found to cause anthracnose in *B. balsamifera*. Both *B. balsamifera* and *B. lacera* are occasionally suffering from *Endophyllum blumeae*. *B. balsamifera* is reported to be a host of the mites *Amblyseius* sp., *Brevipalpus obovatus* and *Typhlodromus jackmicleyi*.

**Harvesting** In gardens, leaves of *B. balsamifera* are collected when required. On a larger scale, either whole plants are harvested or leaves are picked up to four times a year.

**Yield** In Vietnam yields of 50 t/ha of fresh leaves of *B. balsamifera* have been reported, yielding 50–200 kg borneol.

**Handling after harvest** For home consumption fresh leaves are washed, finely chopped and given to patients. Commercially, leaves are dried in the shade, in a well ventilated place before use. On distillation the leaves can yield 0.1–0.5% of a yellow oil.

**Genetic resources and breeding** In view of the weedy nature of most *Blumea*, the risk of genetic erosion appears rather limited. No germplasm collections of *B. balsamifera* are known to be maintained and no breeding work has been done.

**Prospects** The essential oil contains interesting components such as (–)-borneol and (–)-camphor, which may find use e.g. in perfumery and/or chemical synthesis (due to their more uncommon stereochemical conformation). Furthermore, the presence of flavonoids like blumeatin merits further research for their protective effects on the liver.

**Literature** |1| Aguilar, N.O., 1999. *Blumea balsamifera* (L.) DC. In: Oyen, L.P.A. & Nguyen Xuan Dung (Editors): Plant Resources of South-East Asia No 19. Essential-oil plants. Backhuys Pub-

lishers, Leiden, the Netherlands. pp. 68–70. |2| Dutta, S.C., Saha, B.N., Pathak, M.G. & Mathur, R.K., 1989. Essential oil of *Blumea lanceolaria* (Roxb.) Druce. *Indian Perfumer* 33(1): 38–39. |3| Gupta, S.C., Khanolkar, U.M., Koul, O. & Saxena, B.N., 1977. Pyrethrin synergistic activity by the essential oils of a few *Blumea* species. *Current Science* 46(9): 304–305. |4| Koster, J.T., 1972. The Compositae of New Guinea III. *Blumea* 20: 211–225. |5| Lim-Sylianco, C.Y., Concha, J.A., Jorcano, A.P. & Lim, C.M., 1986. Antimutagenic effects of eighteen Philippine plants. *Philippine Journal of Science* 115(4): 293–298. |6| Nguyen Xuan Dung, Do Tat Loi, Do Tat Hung & Leclercq, P.A., 1991. Chemical composition of the oil of *Blumea lanceolaria* (Roxb.) Druce from Vietnam. *Journal of Essential Oil Research* 3(4): 285–286. |7| Perry, L.M., 1980. *Medicinal plants of East and Southeast Asia*. Attributed properties and uses. The MIT Press, Cambridge, Massachusetts, United States & London, United Kingdom. pp. 87–88. |8| Quisumbing, E., 1978. *Medicinal plants of the Philippines*. Katha Publishing Co., Quezon City, the Philippines. pp. 964–967. |9| Randeria, A.J., 1960. The composite genus *Blumea*, a taxonomic revision. *Blumea* 10: 176–317. |10| Xu, S.B., Chen, W.F., Liang, H.Q., Lin, Y.C., Deng, Y.J. & Long, K.H., 1993. Protective action of blumeatin against experimental liver injuries. *Acta Pharmacologica Sinica* 14(4): 376–378.

#### *Selection of species*

#### ***Blumea arfakiana* Martelli**

Nuov. Giorn. Bot. Ital. 15: 292 (1883).

**Synonyms** *Blumea balfourii* Hemsl. (1894).

**Distribution** The Moluccas, New Guinea, the Solomon Islands, Palau and Samoa.

**Uses** In Papua New Guinea, the young leaves are eaten to treat anaemia, and leaves and roots are used against stomach-ache.

**Observations** A herb or low shrub up to 2 m tall, stems erect, sparsely pubescent; leaves broadly elliptical to oblanceolate, 6–35 cm × 3–11 cm, long-tapering at base, margin serrate, scabrid above, sparsely pubescent below, sessile or nearly so; capitula in large terminal panicles, 10–14 mm in diameter, peduncle 4–18 mm long, involucre 8-seriate, 9–10 mm long, involucre bracts elliptical-lanceolate, glandular; marginal flowers 8 mm long, disk flowers 2–8, 5–8 mm long; achene 1.5 mm long, ribbed, pilose, pappus 5–6 mm long, whitish to pale red. *B. arfakiana* occurs in open

places in forests, along rivers and creeks and in fallow fields, up to 900(–1600) m altitude.

**Selected sources** 611, 775, 1198.

#### ***Blumea arnakidophora* Mattf.**

Bot. Jahrb. Syst. 69: 286 (1938).

**Distribution** Borneo (Mt Kinabalu) and New Guinea.

**Uses** In Papua New Guinea the leaf juice is used to treat boils, sores and sore eyes, whereas in New Britain, the leaves and roots are used against stomach-ache.

**Observations** A small shrub up to 2 m tall, stems erect, woolly tomentose; leaves lanceolate to oblanceolate, 4–29 cm × 1.5–8 cm, tapering at base, margin mucronulate-serrate, velutinous above, woolly tomentose below, sessile or nearly so; capitula in large terminal panicles, 12–15 mm in diameter, peduncle 2–15 mm long, involucre 9–10 mm long, involucre bracts linear to linear-lanceolate, densely lanuginose; marginal flowers 4.5–6 mm long, disk flowers 7–18, 4.5–6 mm long; achene 1 mm long, ribbed, sparsely pubescent, pappus 4–6 mm long, pale reddish-yellow. *B. arnakidophora* occurs in grassy, secondary growth, forest edges and roadsides, at 1300–2700 m altitude.

**Selected sources** 597, 610, 755, 1198.

#### ***Blumea balsamifera* (L.) DC.**

Prodr. 5: 447 (1836).

**Synonyms** *Blumea appendiculata* (Blume) DC. (1836), *Blumea grandis* (Wallich) DC. (1836), *Blumea zollingeriana* C.B. Clarke (1876).

**Vernacular names** Ngai camphor plant (En). Camphrier (Fr). Indonesia: sembung (general), sembung utan (Sundanese), sembung gantung (Javanese). Malaysia: chapa, chapor, sembung. Philippines: sambong (Tagalog), lakadbulan (Bikol), subsub (Ilocano). Burma (Myanmar): poung-ma-theing. Cambodia: bai mat. Laos: 'nat, phi ma 'sen. Thailand: kam phung (northern), naat yai (central). Vietnam: d[aj]i bi, t[uw]f bi, b[aw]ng phi[ees]n.

**Distribution** From India, Burma (Myanmar), Indo-China, southern China and Taiwan to Thailand, Malaysia, Indonesia and the Philippines.

**Uses** In South-East Asia *B. balsamifera* is widely used for a number of ailments, mainly as a stomachic, antispasmodic, vermifuge and sudorific. In the Philippines a diuretic and kidney-stone medicine is prepared commercially from *B. balsamifera*. In Thailand cigarettes containing the chopped, dried leaves are smoked to relieve the



pain of sinusitis, whereas a decoction of fresh leaves, alone or in combination with other plant preparations, is used as a bath for women after childbirth and also for young children. In northern Vietnam leaves are also used as a medicinal condiment.

**Observations** A shrub or sometimes a herb, up to 4 m tall, stems erect, densely woolly-villous; leaves usually narrowly oblong-lanceolate or sometimes oblong-ovate or oblong-obovate, 6–30 cm × 1.5–12 cm, tapering at base, margin entire, serrate, serrulate to pinnately lobed, rugose and pilose above, densely silky-woolly below, sessile to petiolate; capitula in large axillary or terminal panicles, 6–10 mm in diameter, peduncle 3–10 mm long, involucre 7–9 mm long, involucral bracts linear, densely woolly; marginal flowers up to 6 mm long, disk flowers 8–28, 5–7 mm long; achene ribbed, about 1 mm long, pubescent, pappus 4–6 mm long, whitish or reddish-yellow. *B. balsamifera* grows, sometimes gregariously, along roadsides, in fields, grasslands, *Imperata* fields, brushwood and forest, including bamboo and teak forest and sometimes in wet places on river banks, from sea-level up to 2200 m altitude. *B. balsamifera* is very variable in its foliage and degree of pubescence; the flower heads and florets, however, are remarkably uniform in size and morphology throughout its range. Plants growing in montane habitats appear more woolly than those from the lowland.

**Selected sources** 97, 184, 202, 287, 332, 344, 364, 437, 531, 580, 651, 775, 852, 858, 885, 974, 1035, 1126, 1128, 1178, 1198, 1232, 1261, 1380, 1411, 1503, 1525, 1601.

### *Blumea lacera* (Burm.f.) DC.

Wight, Contr. bot. India: 14 (1834).

**Synonyms** *Blumea runcinata* DC. (1836), *Blumea thyrsoidea* Sch. Bip. (1866), *Blumea bodinieri* Vaniot (1903).

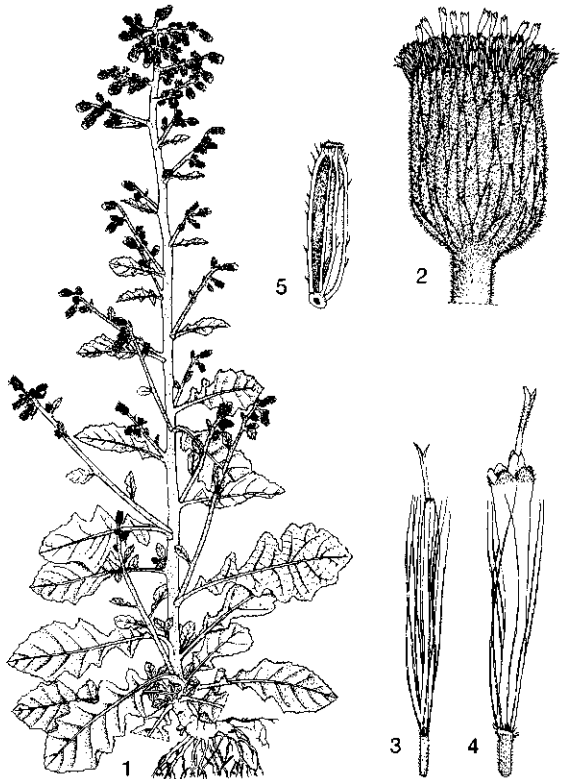
**Vernacular names** Indonesia: batu linca, sembung lalaki (Sundanese), kremahan (Javanese). Malaysia: lumai hutan (Peninsular). Philippines: damong-mabaho, tubang-kabayo (Tagalog), lamlampaka (Bontoc). Thailand: naat wua. Vietnam: c[ar]l[ma], d[a]j[bi] r[as]h.

**Distribution** The Cape Verde Islands, tropical Africa, Pakistan, India, Sri Lanka, Burma (Myanmar), Indo-China, China, the Ryukyu Islands, Taiwan, Thailand, throughout the Malesian region towards the Marianas (Guam), the Solomon Islands, New Caledonia and Australia.

**Uses** In the Philippines, a decoction of fresh

flowers of *B. lacera* is given before meals to treat bronchitis. Its leaf juice is a useful anthelmintic and is given, mixed with black pepper, for haemorrhages. It is also used as a febrifuge, astringent, deobstruent and stimulant. An astringent eye-lotion has also been prepared from the leaves. Furthermore, it is applied as a diuretic and is useful in several catarrhal afflictions. In Vietnam it is used for its wound-healing properties and in the treatment of sores. The leaves can be eaten as a vegetable. *B. lacera* contains a strong, camphor-like oil.

**Observations** A herb up to 2.5 m tall, stems erect, tomentose to densely velutinous; leaves elliptical-oblong to oblanceolate or obovate, 3–21 cm × 1–13.5 cm, acute to tapering at base, margin entire to lyrate lobed, coarsely to finely dentate, tomentose to velutinous above, tomentose to woolly below, sessile or shortly petiolate; capitula in axillary and terminal, dense to lax panicles, 5–6.5 mm in diameter, peduncle 2–10 mm long, involucre 4–10 mm long, 5–6-seriate, outer involucral bracts



*Blumea lacera* (Burm. f.) DC. – 1, plant habit; 2, capitulum; 3, marginal flower; 4, disk flower; 5, achene with pappus removed.

oblanceolate, inner ones linear-lanceolate, all densely velutinous; marginal flowers 3–4 mm long, disk flowers 8–30, 3–4.5 mm long; achene 1–1.3 mm long, ribbed, sparsely pilose, pappus 4–5 mm long, whitish. *B. lacera* is highly variable and occurs in sunny to slightly shaded places in grasslands, fields, roadsides and forest edges, up to 2900 m altitude.

**Selected sources** 24, 26, 97, 202, 287, 530, 580, 637, 775, 804, 852, 1126, 1128, 1178, 1198, 1199, 1365, 1386, 1398, 1532, 1576.

### **Blumea lanceolaria (Roxb.) Druce**

Rep. Bot. Exch. Club Brit. Isles 4: 609 (1917).

**Synonyms** *Blumea myriocephala* DC. (1836), *Blumea laxiflora* Elmer (1906), *Blumea conspicua* Hayata (1911).

**Vernacular names** Malaysia: chapa, kepijit (Peninsular). Vietnam: rau [ax]n g[ox]i, x[uw] [ow]ng s[oo]ng.

**Distribution** From Sri Lanka, India and Bangladesh to Indo-China, China, the Ryukyu Islands, Taiwan, Peninsular Malaysia, Sumatra, Java and the Philippines.

**Uses** The leaves are sudorific and used against bronchitis, aphthae and asthma. They have also been applied externally as a poultice to treat rheumatism. *B. lanceolaria* is sometimes cultivated for culinary purposes as a seasoning.

**Observations** A herb or small shrub up to 2 m tall, stems erect, generally unbranched, glabrous though puberulous above; leaves simple, generally elliptical-oblanceolate, 6–35 cm × 1–9 cm, long-tapering at base, margin minutely to coarsely serrate-dentate, rugose and lustrous above, glabrous or puberulous below, sessile; capitula in terminal panicles, 6–11 mm in diameter, almost sessile and clustered on up to 2 cm long peduncles, involucre 8–9 mm long, 4–5-seriate, outer involucral bracts ovate-lanceolate, inner ones narrowly lanceolate, all pubescent on the back; flowers 5–6 mm long; achene ribbed, pubescent, pappus up to 5 mm long, yellowish-white to pale red. *B. lanceolaria* is highly variable and occurs in humid shaded places, along stream banks, up to 1500 m altitude.

**Selected sources** 97, 202, 384, 852, 1035, 1038, 1126, 1128, 1198, 1397, 1398.

### **Blumea riparia (Blume) DC.**

Prodr. 5: 444 (1836).

**Synonyms** *Blumea chinensis* auct. non (L.) DC., *Blumea pubigera* auct. non (L.) Merr.

**Vernacular names** Indonesia: tombak-tombak (Malay), jonge areuy, lalangkapan (Sundanese).

Papua New Guinea: mulmul (Wapenamanda, Enga), mungla (Mt Hagen, Western Highlands). Philippines: katarai (Sulu), lankat (Manobo), pagang-pagang (Cebu Bisaya). Laos: phang nhot pang. Thailand: mu masang, kamu maeng.

**Distribution** From India and Burma (Myanmar) to Indo-China, southern China, Taiwan, Thailand, throughout the Malesian region to the Solomon Islands.

**Uses** In Peninsular Malaysia, a decoction of the roots is taken to cure colic. In Papua New Guinea, leaf juice is used to treat sores, boils and sore eyes.

**Observations** A scandent shrub, stems sprawling, glabrous though sparsely puberulous above; leaves simple, narrowly elliptical to narrowly obovate, entire, 2.5–13 cm × 1.3–5 cm, rounded at base, margin mucronulate-denticulate, both surfaces glabrous or with a few hairs, shortly petiolate, petiole up to 8 mm long; capitula in terminal and axillary, few-headed racemes, 8–18 mm in diameter, on up to 2.5 cm long peduncles, involucre 8–10 mm long, 5-seriate, outer involucral bracts narrowly ovate, inner ones linear-lanceolate, all pilose; marginal flowers 4–5.5 mm long, disk flowers 5–8, 5–6 mm long; achene about 1 mm long, prominently ribbed, pubescent, pappus 4–6 mm long, white. *B. riparia* is found in thickets, open grassy places, forest clearings, along streams and rivers and in light rain forest, up to 2000 m altitude.

**Selected sources** 97, 580, 597, 610, 775, 852, 1126, 1178, 1198, 1525.

D.S. Alonzo

### **Brucea javanica (L.) Merr.**

Journ. Arn. Arb. 9: 3 (1928).

SIMAROUBACEAE

2n = unknown

**Synonyms** *Brucea sumatrana* Roxb. (1814), *Brucea amarissima* (Lour.) Desv. ex Gomes (1872).

**Vernacular names** Indonesia: kuwalot (Sundanese, Javanese), malur (Batak), tambara marica (South Sulawesi). Malaysia: embalau padang, kusum, lada pahit (Peninsular). Philippines: balaniog (general), magkapayos (Samar-Leyte Bisaya), manongao-bobi (Cebu Bisaya). Cambodia: damli thnang, pramat monus. Laos: ich kone, kom roi, phia<sup>2</sup> fan. Thailand: ratchadat (peninsular), ka chaplak (northern), dee khon (central). Vietnam: c[aa]y su[oo]st, c[uws]t chu[oo]t, s[aa]f[u] d[aa]ju.

**Origin and geographic distribution** *B. javanica* is widespread and occurs from Sri Lanka and India towards Indo-China, southern China, Taiwan, Thailand, and although rare in the Moluccas and New Guinea throughout the Malesian region to northern Australia. Its patchy distribution in eastern Malesia suggests that it was introduced here by man long ago. It has certainly been introduced in Micronesia (Ponape) and Fiji.

**Uses** All parts of *B. javanica*, but most often the pyrenes and roots, are used medicinally, mainly against amoebic dysentery, diarrhoea, malaria and as a febrifuge. It is known in Chinese traditional medicine, where it is additionally applied for the treatment of haemorrhoids, corns, warts, ulcers and cancer. The pyrenes are well-known under the name 'Macassar kernels', and are also applied as an insecticide. The leaves are applied as a poultice against enlarged spleen, scurf, ring-worm, boils and centipede bites. A decoction of the roots is also used to treat abdominal pains, coughs and as an important remedy for internal poisoning. In Australia, the bark and roots have been used by Aborigines to treat toothache.

**Production and international trade** The pyrenes, roots and occasionally other parts of *B. javanica* are traded on local markets, but are not of great commercial importance.

**Properties** Several quassinoids have been isolated from the fruits of different *Brucea* species. In general, these compounds have been shown to have strong anti-amoebic, antimalarial and/or cytotoxic (anti-cancer) properties.

One of the major quassinoids found in *B. javanica* and *B. antidysenterica* J.F. Miller from Africa is bruceantin. Anti-amoebic, antimalarial and anti-cancer properties of this compound are reported in literature; the antimalarial activity is not simply due to cytotoxic effects. Furthermore, quassinoids (e.g. brusatol) from the fruits, as well as the triterpenoids bruceajavanin A, dihydrobruceajavanin A

and bruceajavanin B (from the stems of *B. javanica*) have been shown to inhibit the growth of the chloroquine-resistant strain *Plasmodium falciparum* K1 in vitro. Some of the quassinoids (bruceine A, B and D, brusatol) also showed in vivo activity against *P. bergeri* infections in mice after oral dosing. Finally, the quassinoids bruceine A, B and C, present in a chloroform extract of *B. javanica* fruits, had a very potent activity in vitro against a multi-drug resistant *P. falciparum* strain with an ID<sub>50</sub> of 8.66, 8.15 and 1.95 ng/ml, respectively, in comparison with 6.26 ng/ml for the mefloquine reference.

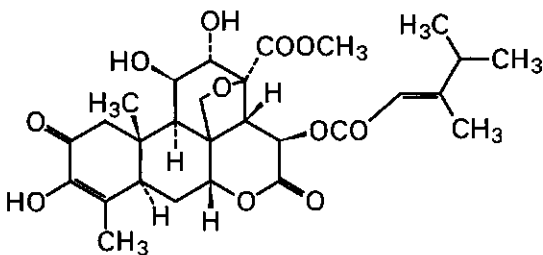
Quassinoids (e.g. bruceolides, bruceantin and bruceantinol) are reported to show inhibitory action against lymphocytic leukaemia and lung carcinoma. Bruceoside A and B (quassinoids) were found to possess lethal toxicity when the methanol extract of *B. javanica* was administered to mice. Bruceoside C showed potent cytotoxicities against KB, A-549, RPMI and TE-671 tumour cell lines, and bruceosides D, E, and F show selective cytotoxicity in leukaemia and non-small cell lung, colon, central nervous system, melanoma and ovarian cancer cell lines. Other quassinoids found in *B. javanica* with cytotoxic effects and with potential for cancer therapy include e.g. brusatol and the yadanziosides A-H, O and P.

*B. antidysenterica* also contains quassinoids with similar cytotoxic effects (e.g. bruceanols A, B, D, E, F, G, H, bruceantinosides A-C, yadanziosides G, N, M, P) as well as bruceanic acids (A, the methyl ester of A, B-D). Cytotoxic canthin-6-one alkaloids, biosynthetically derived from tryptophan, are also reported in the literature.

Clinical observations of improvement in clinical manifestations after administering *B. javanica* oil emulsion intravenously to patients with brain metastasis from lung cancer have been experimentally confirmed in rabbits by positive effects on intracranial hypertension.

The crude extract of *B. javanica* has been found to be very effective against the internal parasite *Blastocystis hominis* at an active concentration of 500 µg/ml as compared with 10 µg/ml for metronidazole, the active standard drug for *B. hominis*. Bruceoside D showed in vitro anti-tuberculosis activity. However, the activity on the test organism, *Mycobacterium tuberculosis*, was low, with 7% inhibition at 12.5 µg/ml.

**Adulterations and substitutes** The *Cinchona* alkaloid quinine, and the synthetic antimalarials derived from it (e.g. chloroquine) are widely in use for the classical treatment of malaria. Artemisinin



bruceantin

(a sesquiterpene lactone from *Artemisia annua* L.) and its synthetic analogues (e.g.  $\beta$ -artemeter and sodium artesunate) comprise a class of new anti-malarials, which are of interest, since resistance to the malaria-causing parasites is developing fast. Other *Simaroubaceae* that might be of interest in this respect include *Eurycoma*, *Picrasma* and *Quassia*, whereas *Dichroa* species (*Saxifragaceae*) warrant some attention as well.

**Description** A monoecious or dioecious shrub or small tree up to 10 m tall with soft-haired twigs and leaves. Leaves arranged spirally, imparipinnate, 20–50 cm long, exstipulate; leaflets 3–15, opposite, short petiolulate, ovate-oblong to ovate-lanceolate, margin bluntly serrate or crenate, secondary veins unbranched and terminating in a marginal gland. Inflorescence axillary, pubescent, composed of small cymes united into bracteate, raceme-like thyrses. Flowers unisexual, 4-merous, small, greenish-white to greenish-red or purple; sepals connate at base; petals free; disk intrastaminal, thick, 4-lobed; stamens short, vestigial or absent in female flowers; ovaries superior, free,

each with a single, pendent ovule, styles free or coherent at base, subulate, bent outwards over the top of the ovary. Fruit consisting of 1–4 hardly fleshy drupelets; drupelet 2-ribbed, dry, purplish-black and 4–5(–7) mm long when mature, pyrene with wrinkled endocarp. Seed ovoid, with thin testa and very thin endosperm. Seedling with epigeal germination; cotyledons emergent, leafy; hypocotyl elongated; first two leaves opposite, subsequent ones arranged spirally; first few leaves 3-foliolate, later ones with increasing number of leaflets.

**Growth and development** In a germination test in Peninsular Malaysia fruits of *B. javanica* had a germination rate of about 35% within 11–273 days.

All shoots are orthotropic. Growth is rapid and flowering starts early. Flowering and fruiting can be throughout the year. Pollination is probably by insects. Small fruit bats eat the fruits and thus disperse the seed. However, the life span is only a few years and plants have to be regenerated from seed.

**Other botanical information** A second Malaysian species, *B. mollis* Wallich ex Kurz (synonyms: *B. luzoniensis* S.Vidal, *B. macrobotrys* Merr., *B. acuminata* Li), is found in Indo-China, Thailand and the Philippines and may contain similar medicinally active compounds. The African *B. anti-dysenterica* is a well-known medicinal plant with similar applications.

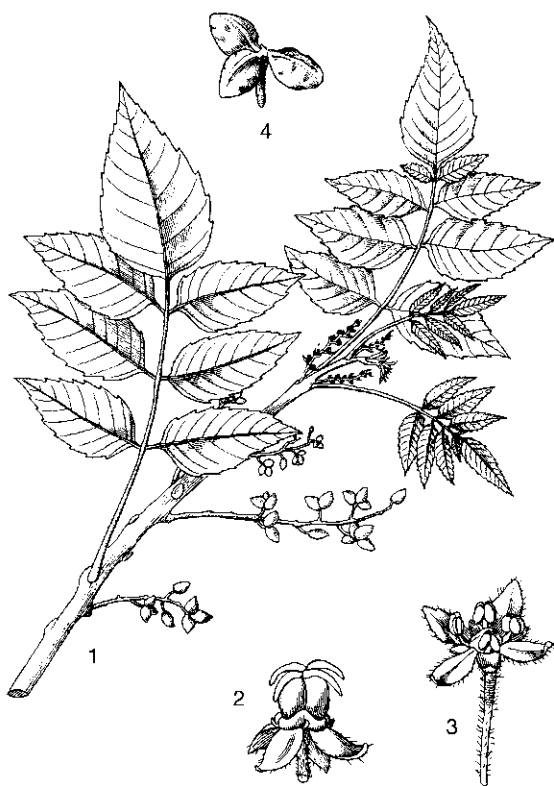
**Ecology** *B. javanica* is very common preferring open localities such as light secondary forest and thickets, forest edges, ridges, and even occurring in sunny places on sandy dunes and on limestone. It grows under both per-humid and seasonal conditions, from sea-level up to 900 m altitude.

**In vitro production of active compounds** Canthin-6-one alkaloids can be produced by cell suspension cultures of *B. javanica*. The total yield of alkaloids produced in cells and medium is in excess of 2.0 mg/g on a dry weight basis. The major alkaloids produced are canthin-6-one, 11-hydroxycanthin-6-one, 5-methoxycanthin-6-one and 11-methoxycanthin-6-one.

**Handling after harvest** After the mature fruits have been collected, the fruit pulp is removed. The pyrenes that remain are washed and dried in the sun.

**Genetic resources and breeding** Since *B. javanica* is common in anthropogenic habitats and has a large area of distribution, the risk of genetic erosion seems limited.

**Prospects** The various quassinoids found in the



*Brucea javanica* (L.) Merr. – 1, twig with flowers and fruits; 2, female flower; 3, male flower; 4, fruit.

seed of *B. javanica* that possess both antimalarial and anti-cancer activity merit further research. Because of the growing resistance of malaria parasites to the well-known, and even newer, antimalarials currently in use, there is a continuous need to develop new compounds to control this almost global infectious disease. The quassinoids and the canthin-6-one alkaloids might also have good potential for the development of a cytostatic drug to treat various cancers.

**Literature** |1| Anderson, M.M., O'Neill, M.J., Phillipson, J.D. & Warhurst, D.C., 1991. In vitro cytotoxicity of a series of quassinoids from *Brucea javanica* fruits against KB cells. *Planta Medica* 57(1): 62-64. |2| Fukamiya, N., Okano, M., Miyamoto, M., Tagahara, K. & Lee, K.H., 1992. Antitumor agents, 127. Bruceoside C, a new cytotoxic quassinoid glucoside, and related compounds from *Brucea javanica*. *Journal of Natural Products* 55(4): 468-475. |3| Kitagawa, I., Mahmud, T., Simanjuntak, P., Hori, K., Uji, T. & Shibuya, H., 1994. Indonesian medicinal plants. VIII. Chemical structures of three new triterpenoids, bruceajavanin A, dihydrobruceajavanin A, and bruceajavanin B, and a new alkaloidal glycoside, bruceacanthinoside, from the stems of *Brucea javanica* (Simaroubaceae). *Chemical and Pharmaceutical Bulletin* 42(7): 1416-1421. |4| Kupchan, S.M., Britton, R.W., Lacadie, J.A., Ziegler, M.F. & Sigel, C.W., 1975. The isolation and structural elucidation of bruceantin and bruceantinol, new potent antileukemic quassinoids from *Brucea antidysenterica*. *Journal of Organic Chemistry* 40(5): 648-654. |5| Lee, K.H., Tani, S. & Imakura, Y., 1987. Antimalarial agents, 4. Synthesis of a brusatol analog and biological activity of brusatol-related compounds. *Journal of Natural Products* 50(5): 847-851. |6| Lu, J.B., Shu, S.Y. & Cai, J.Q., 1994. Experimental study on the effect of *Brucea javanica* oil emulsion on rabbit intracranial pressure. *Chung Kuo Chung Hsi I Chieh Ho Tsa Chih* 14(10): 610-611 (in Chinese). |7| Nooteboom, H.P., 1962. Simaroubaceae. In: van Steenis, C.G.G.J. (Editor): *Flora Malesiana*. Series I, Vol. 6. Wolters-Noordhoff Publishing, Groningen, the Netherlands. pp. 209-212. |8| Ohnishi, S., Fukamiya, N., Okano, M., Tagahara, K. & Lee, K.H., 1995. Bruceosides D, E, and F, three new cytotoxic quassinoid glucosides from *Brucea javanica*. *Journal of Natural Products* 58(7): 1032-1038. |9| O'Neill, M.J., Bray, D.H., Boardman, P., Chan, K.L., Phillipson, J.D., Warhurst, D.C. & Peters, W., 1987. Plants as sources of antimalarial drugs, Part 4: Activity of *Brucea javanica* fruits against

chloroquine-resistant *Plasmodium falciparum* in vitro and against *Plasmodium berghei* in vivo. *Journal of Natural Products* 50(1): 41-48. |10| Pavanand, K., Nutakul, W., Dechatiwongse, T., Yoshihira, K., Yongvanitchit, K., Scovill, J.P., Flippen-Anderson, J.L., Gilardi, R., George, C., Kanchanapee, P. & Webster, H.K., 1986. In vitro antimalarial activity of *Brucea javanica* against multi-drug resistant *Plasmodium falciparum*. *Planta Medica* 52(2): 108-111.

**Other selected sources** 202, 284, 363, 441, 442, 443, 488, 579, 580, 648, 649, 834, 1020, 1035, 1053, 1076, 1077, 1078, 1080, 1096, 1126, 1128, 1165, 1178, 1271, 1272, 1273, 1287, 1383, 1473, 1513, 1548, 1571, 1614.

Arbayah H. Siregar

### **Bryophyllum Salisb.**

Parad. Lond.: t. 3 (1805).

CRASSULACEAE

$x = 17, 18$ ; *B. pinnatum*:  $2n = 36, 40$ , *B. proliferum*:  $2n = 34$

**Major species** *Bryophyllum pinnatum* (Lamk) Oken.

**Vernacular names** Indonesia: buntiris (Sundanese). Malaysia: sedingin, seringin (Peninsular).

**Origin and geographic distribution** *Bryophyllum* comprises about 30 species. Almost all species are restricted to or originate from Madagascar. However, the exact origin of *B. pinnatum* is unknown. In Malesia 2 species are found naturalized.

**Uses** The main medicinal use of *B. pinnatum* in South-East Asia is in the treatment of boils, wounds, burns and scalds. In Indonesia, pounded leaves are used as a diuretic, and leaves are used externally to treat sores and pain in back and feet; a poultice is sometimes applied on sore eyes or to relieve headache; a decoction is used internally to treat fever and oedema; an extraction of dried pounded leaves in water is used against haemorrhoids. In Malaysia, the crushed leaves are applied to the forehead to treat headache and to the chest to treat coughs and pains. In Brunei, a leaf infusion is drunk as a febrifuge. In the Philippines, the leaves are used as an astringent, antiseptic, and against insect bites. Fresh, pounded leaves are applied to burns and as poultices on boils. Leaf juice is used (mixed with lard) to treat diarrhoea, dysentery, cholera and phthisis. The leaves are also used as topicals to treat disloca-

tions, equimosis and callosities. In Papua New Guinea, young leaves are heated over a fire and placed on sores, or applied as a poultice to boils, sores and swellings. In Vietnam, Cambodia, Laos and Thailand, fresh leaves are applied to burns, scalds, wounds, boils, skin diseases and corns and to treat ophthalmia, phlegm, rheumatism, neuralgia and pain.

The use of *B. pinnatum* for medicinal purposes is also widespread outside South-East Asia. In India, the leaves are applied to wounds, bruises, boils and, in the form of poultice or powder, to ulcers, whereas leaf juice is given to treat bilious diarrhoea and lithiasis. In West Africa, the juice is used as a diuretic and for the treatment of earache and ophthalmia. The leaves are rubbed or tied on the head against headache, and the roots are used to make a cough medicine. In Brazil, the leaves are used as an emollient and refrigerant over a face swollen from neuralgia or tooth trouble, and in Puerto Rico leaf juice is used to treat acute nephritis. *B. pinnatum* is also used as an ornamental plant and in ceremonies.

*B. proliferum* is sometimes cultivated in South-East Asia as a hedge plant, but there are no reports of medicinal uses.

**Properties** The leaves of *B. pinnatum* are reported to contain 'bryophyllin', a mixture of bufadienolides, with antibacterial effects on gram-positive and gram-negative bacteria. They are used to treat intestinal problems caused by such bacteria, and also externally. Further phytochemical investigations have revealed the presence of the bufadienolides bryophyllin A (= bryotoxin C), bryophyllin B and bersaldegenin-3-acetate in fresh whole plant material of *B. pinnatum*. Bryophyllin A can be converted into bryophyllin B by the addition of a catalytic amount of acid ( $\pm$ )-10-camphor sulphonic acid). All 3 compounds have shown strong in vitro cytotoxicity against KB-tumour cells; bryophyllin A and bersaldegenin-3-acetate were also cytotoxic to A-549 and HCT-8 cells.

Both *B. pinnatum* and *B. proliferum* can be poisonous to cattle and sheep when eaten in large amounts. The toxic syndrome is known as cotyledonosis (or 'krimpsieke'), a neurotoxic syndrome, which, together with cardiac glycoside poisoning can be caused by *Crassulaceae*-bufadienolides. The bufadienolides bryotoxin A, B and C (= bryophyllin A) have been isolated from different parts of both species. High pressure liquid chromatography (HPLC) analysis has revealed concentrations of 61 mg/kg bryotoxin (total of A, B

and C) in the flowerheads of *B. proliferum*, and 24, 52 and 141 mg/kg bryotoxin (total of A, B and C), respectively, in the flowerheads, leaves/stems and roots of *B. pinnatum*. Experiments with the structurally comparable bufadienolides daigremontin and bersaldegenin-1,3,5-orthoacetate (from *B. daigremontianum* (Hamet & Perr.) Berger and *B. tubiflorum* Harv.) showed a pronounced sedative effect in mice at low doses (0.1–0.5 mg/kg; motility test), but a toxic effect at higher concentrations, inducing paralysis and spasmodic muscle contractions. A pronounced positive inotropic activity is also seen on the heart.

Furthermore, methanolic leaf extracts of *B. pinnatum* showed anti-inflammatory activity in rats and mice (carrageenin-induced oedema, cotton pellet granuloma, formaldehyde-induced arthritis, adjuvant-induced arthritis and turpentine-induced joint oedema). It is thought that the active constituents,  $\beta$ -sitosterol and some aliphatic alcohols might contribute to the effects. Methanolic leaf extracts also showed anti-ulcer activity in rats and guinea-pigs by providing protection against gastric lesions induced by e.g. aspirin, indomethacin, serotonin, reserpine, ethanol, histamine, acetic acid or stress. In mice infected with *Leishmania amazoniensis*, oral treatment with an aqueous leaf extract from *B. pinnatum* significantly decreased lesion growth and the number of viable parasites.

Ethyl acetate and petroleum ether extracts of leaves of *B. pinnatum* exhibited potent antimutagenic activities at non-toxic concentrations against reversion mutations induced by ethyl methane-sulphonate in *Salmonella typhimurium* strains TA 100 and TA 102. Further fractionation into non-polar lipid and polar lipid fractions and investigations of these suggest that there may be different types of antimutagenic compounds in *B. pinnatum*.

Besides the bufadienolides mentioned, a variety of other compounds have been found in *B. pinnatum*: sterols (e.g. 24-ethyl-desmosterol, sitosterol, clerosterol, isofucoesterol), triterpenes (e.g.  $\alpha$ -amyrin,  $\beta$ -amyrin, 18 $\alpha$ -oleanane, bryophollone, bryophynol,  $\psi$ -taraxasterol), phenanthrenes (e.g. 2(9-decenyl)phenanthrene, 2(9-undecenyl)phenanthrene), alkanes (C<sub>25</sub>–C<sub>35</sub>, with C<sub>31</sub> and C<sub>33</sub> predominating), alkanols (C<sub>26</sub>–C<sub>34</sub>, with C<sub>32</sub> as major component), phenolic compounds (e.g. p-coumaric acid, ferulic acid, syringic acid, caffeic acid, p-hydroxybenzoic acid) and flavonoid glycosides (quercetin-3-di-arabinoside, kaempferol-3-glucoside).

**Adulterations and substitutes** Other *Crassu-*

laceae, notably *Kalanchoe ceratophylla* Haw. and *K. crenata* (Andrews) Haw., have comparable medicinal uses.

**Description** Shrubs or shrublets with branches erect, cartilaginous, but usually somewhat woody, rarely regenerating from the base. Leaves opposite, simple or imparipinnate, fleshy, persistent, usually producing pseudobulbils on the margins. Inflorescence terminal or terminal with axillary paniculate cymes; bracts on peduncle distinct, abruptly shorter than leaves. Flowers bisexual, pendulous, 4-merous; calyx with sepals usually fused for more than half the length; corolla fused into a tube longer than the spreading lobes; stamens 8 in 2 whorls, filaments glabrous and fused to corolla tube in lower third, anthers usually protruding, with terminal appendage; ovary superior, consisting of 4 free carpels, abruptly constricted into longer styles. Fruit a many-seeded follicle. Seeds ellipsoid, with a constriction and abruptly widening at the blunt proximal end.

**Growth and development** The formation of foliar embryos in the notches of the leaf is typical for *Bryophyllum*. Contrary to some other *Bryophyllum* spp., which develop plantlets under long days, foliar embryos in *B. pinnatum* normally develop into plantlets only when the leaf is detached or injured, or when cytokinin is applied to the attached leaf. Inhibition of plantlet formation in intact plants is probably related to high auxin levels. Release from dormancy is also reported to be initiated by external conditions, such as high humidity, absence of light or water stress, but the production of plantlets of *B. pinnatum* is not influenced by photoperiodicity.

The young plants on the leaves fall off after having formed roots and a thin stem with a few leaves. They may be transported by rain-wash. In Malesia, *B. pinnatum* never sets fruit. *B. proliferum* never flowers in lowland Java, and rarely in mountainous regions during the period March-August. It seems that no fruits are produced.

*B. pinnatum* was the first plant in which Crassulacean Acid Metabolism (CAM) was described. CAM plants are able to fix CO<sub>2</sub> at night and have photosynthesis with closed stomata during the day, to minimize water loss.

**Other botanical information** *Bryophyllum* is closely related to *Kitchingia* and *Kalanchoe*; sometimes the first two genera are united in *Kalanchoe*. However, several vegetative and floral characters can be used to distinguish *Bryophyllum* from *Kalanchoe*. *Bryophyllum* usually has pendulous flowers, larger, campanulate-globular

calyces with fused sepals, corolla tubes constricted above the ovary, stamens inserted at the base of the corolla tube, carpels shorter than the styles, large epigynous scales, foliar embryos on the leaf margins in 50% of the species, a basic chromosome number of 17 for most species and almost all species are from Madagascar.

**Ecology** *Bryophyllum* is very hardy, and will survive under a low water supply. In South-East Asia, *B. pinnatum* is found up to 1000 m altitude, in sunny or slightly shaded locations. The habitat can be stony, is always dry and never far from human habitation. Naturalized *B. proliferum* is found in Java between 1000 m and 1600 m altitude. It is cultivated at lower elevations as well.

**Propagation and planting** The easiest way of propagation of *Bryophyllum* is by foliar embryos. When leaves are cut and kept under warm and moist conditions, plantlets soon form. When these have formed roots and a thin stem with a few leaves, they can be separated from the parent leaf and planted. *Bryophyllum* can also be propagated through stem cuttings and seed.

**Husbandry** *Bryophyllum* species are collected from the wild or grown in small quantities for home use, and information on specific care is absent.

**Diseases and pests** In India, *B. pinnatum* is reported to be susceptible to the fungi *Alternaria alternata*, *Glomerella cingulata* and *Colletotrichum dematium*.

**Harvesting** The leaves are simply cut and used fresh.

**Genetic resources and breeding** *Bryophyllum* species occurring in the Malesian region are widely distributed in other parts of the world. This widespread distribution, the relatively easy propagation and common use as a pot plant limits the risk of extinction. However, as both species are almost exclusively propagated vegetatively, the genetic basis in South-East Asia may be very narrow.

**Prospects** *Bryophyllum* species will remain of some importance as a readily available traditional antiseptic and counterirritant, and will be collected and/or grown in small quantities for home use. The antimutagenic activity merits further research.

**Literature** [1] Akihisa, T., Kokke, W.C.M.C., Tamura, T. & Matsumoto, T., 1991. Sterols of *Kalanchoe pinnata*: first report of the isolation of both C-24 epimers of 24-alkyl- $\delta^{25}$ -sterols from a higher plant. *Lipids* 26(8): 660-665. [2] Backer, C.A., 1951. Crassulaceae. In: van Steenis,

C.G.G.J. (General editor): Flora Malesiana. Series 1, Vol. 4. Noordhoff-Kolff N.V., Djakarta, Indonesia. pp. 197-202. |3| Burkill, I.H., 1966. A dictionary of the economic products of the Malay Peninsula. Revised reprint. Vol. 1. Ministry of Agriculture and Cooperatives, Kuala Lumpur, Malaysia. pp. 380-381. |4| Gaiind, K.N. & Gupta, R.L., 1973. Phenolic components from the leaves of *Kalanchoe pinnata*. *Planta Medica* 23(2): 149-153. |5| Houck, D.F. & Rieseberg, L.H., 1983. Hormonal regulation of epiphyllous bud release and development in *Bryophyllum calycinum*. *American Journal of Botany* 70(6): 912-915. |6| Lauzac-Marchal, M., 1974. Taxonomie végétale.- Réhabilitation du genre *Bryophyllum* Salisb. (Crassulacées Kalanchoïdées). [Plant taxonomy.-Réhabilitation of the genus *Bryophyllum* Salisb. (Crassulaceae Kalanchoideae)]. *Comptes rendus hebdomadaires des séances de l'Académie des Sciences Paris. Série D. Sciences naturelles*. Vol. 278: 2505-2508. |7| Perry, L.M., 1980. Medicinal plants of East and Southeast Asia. Attributed properties and uses. The MIT Press, Cambridge, Massachusetts, United States & London, United Kingdom. pp. 109-110. |8| Quisumbing, E., 1978. Medicinal plants of the Philippines. Katha Publishing Co., Quezon City, the Philippines. pp. 349-351. |9| Sidharta, P. & Chaudhuri, A.K.N., 1991. Studies on the anti-ulcer activity of a *Bryophyllum pinnatum* leaf extract in experimental animals. *Journal of Ethnopharmacology* 33: 97-102. |10| Yamagishi, T., Haruna, M., Yan, X.Z., Chang, J.J. & Lee, K.H., 1989. Antitumor agents, 110. *Bryophyllin B*, a novel potent cytotoxic bufadienolide from *Bryophyllum pinnatum*. *Journal of Natural Products* 52(5): 1071-1079.

#### *Selection of species*

#### ***Bryophyllum pinnatum* (Lamk) Oken**

Allg. Naturgeschichte Vol. III(3): 1966 (1841).

**Synonyms** *Cotyledon pinnata* Lamk (1786), *Bryophyllum calycinum* Salisb. (1805), *Kalanchoe pinnata* (Lamk) Pers. (1805).

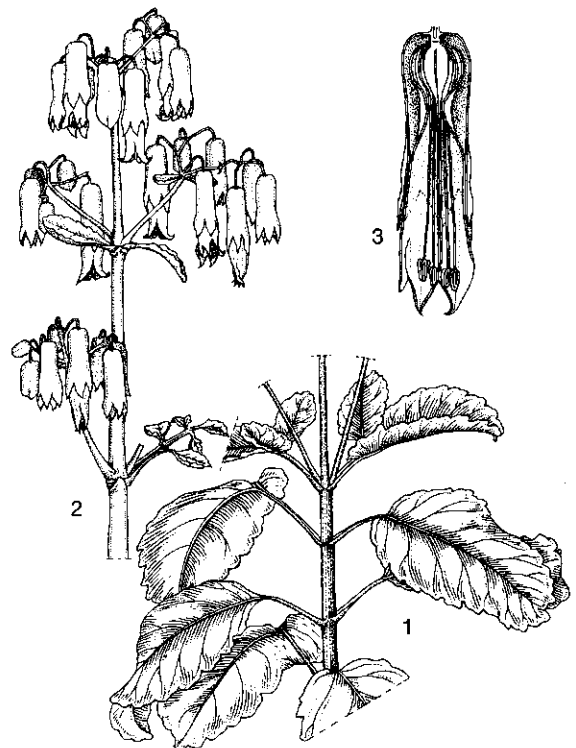
**Vernacular names** Life plant, floppers (En). Brunei: bendingin, serigen. Indonesia: daun sejuk (Malay, Palembang), buntiris (Sundanese), sosor bebek (Javanese). Malaysia: sedingin, seringin, setawar padang. Philippines: karitana (Bisaya), abisrana (Iloko), katakataka (Tagalog). Burma (Myanmar): yoekiyapinba. Laos: poun tay, poun po. Thailand: benchachat (central), ton tai bai pen, khwum taai ngaai pen. Vietnam: c[aa]y

thu[oo]c b[or]ng, c[aa]y tr[uw][ow]ng sinh, l[aj]c d[i]j]a sinh c[aw]n.

**Distribution** *B. pinnatum* has a pantropical distribution. In Malesia it is naturalized throughout the region. In many places it is a weed; on the other hand it is reported to be cultivated in Indonesia, Malaysia, the Philippines and Indo-China.

**Uses** The fresh leaves are commonly used as a poultice in the treatment of boils, wounds, burns and scalds.

**Observations** A robust, unbranched herb, 30-200 cm tall, glabrous; leaves fleshy, leathery when older, earlier ones simple, ovate, with cordate or rounded base, upper ones pinnate, 3-5-foliolate, 5-9(-20) cm x 2.5-5 cm, base cuneate, apex obtuse, margin crenate, petiole semi-amplexicaulous; inflorescence a lax terminal cyme, 10-80 cm long; flowers pendulous, with a cylindrical calyx, up to 25 mm x 8 mm, slightly indented at the base, lobes ovate-triangular, 7-12 mm long, very acutely acuminate, corolla tube cylindrical, about 30 mm long, strongly constricted at about 8 mm



*Bryophyllum pinnatum* (Lamk) Oken - 1, upper part of stem; 2, inflorescence; 3, flower in longitudinal section.



from the base, corolla lobes oblong-ovate, about 6 mm long, abruptly long-acuminate, in lower half green, in upper half red. *B. pinnatum* is found in dry rather sunny locations near human habitation up to 1000 m altitude.

**Selected sources** 95, 97, 202, 302, 332, 350, 363, 461, 462, 574, 597, 619, 741, 824, 900, 923, 924, 979, 1011, 1035, 1063, 1126, 1128, 1178, 1203, 1346, 1347, 1351, 1569, 1584, 1604, 1605.

### **Bryophyllum proliferum** Bowie

Curtis's Bot. Mag., ser. III, 15: t. 5147 (1859).

**Synonyms** *Kalanchoe prolifera* (Bowie) Hamet (1908).

**Vernacular names** Indonesia: buntiris (Sundanese).

**Distribution** *B. proliferum* is native to Madagascar and was introduced into Java long ago. It is naturalized in mountainous parts of West Java.

**Uses** The fresh leaves of *B. proliferum* may well be used in a similar way as with *B. pinnatum*. It is sometimes cultivated as a hedge plant.

**Observations** An erect shrub 50–200 cm tall, quite glabrous; lower and topmost leaves of flowering plants not deeply divided, middle ones deeply pinnatisect with oblong segments, 5–23 cm × 2–8 cm, slightly or rather deeply crenate, very fleshy, petiole robust, semi-amplexicaulous; inflorescence a terminal cyme, 50–80 cm long; flowers pendulous, with broad and cuspidate calyx lobes, corolla tube cylindrical, 2–2.5 cm long, distinctly constricted above the base, shortly 4-lobed, lobes ovate, 3–5 mm long, shortly acuminate, red. *B. proliferum* is found naturalized in hedges, thickets and roadsides between 1000 m and 1600 m in Java.

**Selected sources** 65, 97, 824, 900, 923, 1536.

Wardah & M. Brink

### **Cannabis sativa** L.

Sp. pl. 2: 1027 (1753).

CANNABACEAE

2n = 20, 40, 80

**Vernacular names** Hemp, Indian hemp, marihuana (En). Chanvre (Fr). Indonesia: ganja (general), ginje jawa (Javanese). Malaysia: ganja. Thailand: kancha, kancha cheen (general), paang (Shan-Mae Hong Son). Vietnam: gai m[ef]o, lanh m[as]n, c[aa]f[n] xa.

**Origin and geographic distribution** *C. sativa* is the only species in *Cannabis*. It is a native of the temperate parts of Asia: near the Caspian

Sea, in Iran, the Kirghiz steppe, southern Siberia and probably also the Himalayas and northern India. It is one of the oldest of cultivated plants. Hemp was valued by the Chinese 8500 years ago, and it may be one of the oldest non-food crops. It was introduced into western Asia and Egypt, and subsequently Europe during the period 2000–1000 B.C., and cultivation in Europe became widespread from about 500 AD onwards. From Central Asia it spread eastward to China, Indo-China, Thailand and the Malesian region. It might have occurred in Java already 1000 years ago, in Malaysia more than 300 years ago, and in the Philippines more than 200 years ago. It was introduced into South America in 1545 and into North America in 1606. Nowadays, it is cultivated in many parts of the temperate, subtropical and tropical regions. In many countries including those in South-East Asia, however, its cultivation is prohibited by law.

**Uses** Hemp provides different products: therapeutics and narcotics (flowers and leaves), fibre (stems), oil (seeds) and food for humans and animals (seeds). It was probably first used as a source of fibre: the oldest remains of cloth made from hemp date back 6000 years. The use for seed oil is more recent, but began at least 3000 years ago, and the earliest reference to narcotic use appears to date from 5000 years ago in China. The earliest recorded medicinal use of hemp is found in a Chinese pharmacopoeia of 4700 years old.

All parts of the plant are used in Chinese medicine. The seeds are considered useful as a tonic, alterative, emmenagogue, laxative, demulcent, diuretic, anthelmintic, narcotic and anodyne. They are prescribed in fluxes, post-partum difficulties, obstinate vomiting, and used externally on eruptions, ulcers, wounds and favus. The specially prepared seeds are prescribed for uterine prolapse and to aid parturition, and as a febrifuge. Hemp is a sedative of the stomach, used to treat dyspepsia with painful symptoms, cancers and ulcers. It is also used to treat migraine, neuralgia, tetanus and rheumatism.

In western medicine, hemp preparations were extensively used between the middle of the 19th Century and the Second World War as an anticonvulsant, analgesic, sedative, and soporific, and to treat tetanus, neuralgia, uterine haemorrhage, rheumatism, epilepsy, migraine, convulsions, spasms and miscellaneous pains. It was considered a milder and less dangerous analgesic than opium.

The inconsistency of its therapeutic activity, the

poor keepability of its preparations, the difficulty in deciding optimal doses and the emergence of synthetic analgesics and hypnotics led to the use of hemp being gradually abandoned in the first half of the 20th Century, and there is very limited authorized medicinal use today. In recent years, hemp drugs have been advocated as very useful to treat spasm in patients suffering from multiple sclerosis, to treat increased pressure within the eyeball and to treat loss of appetite in AIDS patients. The best known application, however, is its use as a sedative in cancer patients, and to treat side-effects of cancer chemotherapy e.g. nausea, vomiting and convulsions. One of the cannabinoids,  $\Delta^9$ -tetrahydrocannabinol, the best known active compound of hemp, is marketed as an anti-emetic (sometimes also called dronabinol) e.g. in the United States. Other potential applications of isolated cannabinoids include the use as antiglaucoma, anti-asthmatic, anticonvulsant, spasmolytic and analgesic.

The narcotic use varies between cultures, and many descriptive terms exist. In India, where the use of *Cannabis* as a drug became more important in the last millennium than anywhere else in the world, three types of preparations are distinguished: 'bhang' (dried, powdered plant, made into a drink with milk or water), 'ganja' (dried flowering tips of female plants, usually smoked, sometimes eaten or drunk) and 'charas' (crude resin scraped from the plant, which is smoked, sometimes eaten). In the western world, 'marihuana' usually refers to a preparation comprising crumbled leaves, small twigs and flowering parts of female plants, whereas 'hashish' is a stronger preparation, with more resin and little recognizable plant material. All these drug types contain a resin from the glandular hairs on leaves, stems and inflorescences.

The bast fibres of *C. sativa* are traditionally used to make yarns, twines, ropes, nets and paper, while the wooden core of the stems is normally used as animal bedding or fuel. Hemp yarns are mainly made by wet spinning the long fibres. Improved yarn quality can be obtained by 'cottonization' of hemp, involving the chemical or mechanical rearrangement of bast fibres. This makes it possible to process the fibres on cotton machines. However, the hemp fibres and yarns currently on the market do not meet the requirements of the textile industry with respect to fibre fineness, homogeneity, flexibility and distribution of fibre length. In Malesia, hemp is not important as a fibre crop, but in Thailand its fibres are used to

make ropes and textile, especially in the northern part of the country.

Carton can be made from pulp of the different fibre types of hemp. High quality paper (monetary bills) and specialty applications (diapers, bandage) may be envisaged using pulp from bast or core.

Current fibre applications in building and construction materials are fibre and particle boards, panels and inorganic matrix composites (IMC). Boards and panels are mostly used indoors as a non-structural material for insulation. The applications of IMCs include plaster boards, tiles, concrete, mortars and plasters. Important arguments for using plant fibres in IMCs are asbestos substitution, saving of weight, waste management and the good compatibility with the matrix due to the hydrophylic properties of plant fibres.

One technologically innovative application of bast fibres of hemp is in fibre reinforced structural materials called composites, where the fibres replace the glass fibres that are normally used. Combined with the application of biodegradable resins such as cellulose, starch or casein, the use of natural fibres in composites has a clear ecological advantage over traditional materials. Markets for these often specialized and costly products are manufacturers of automotive and aircraft interior parts, of machines, of sports and leisure goods and of biomedical aids, and the construction industry.

The oil in the seeds can be used as a substitute for linseed oil in paints and soap. Other applications of the oil are in cosmetics and as surfactants in detergents.

Hemp seed is edible, and used as human food on a limited scale. It is also used as bird and poultry feed. The press cake remaining after oil extraction is used to feed livestock.

**Production and international trade** The production areas of hemp differ, depending on the product required. Hemp for narcotic properties is mainly grown in warmer climates. It is cultivated for 'ganja' in India (Bengal, Madras and Mysore) by a few licensed growers, and the drug is a monopoly of the Indian Government. In many countries it is grown for narcotics even though this is illegal.

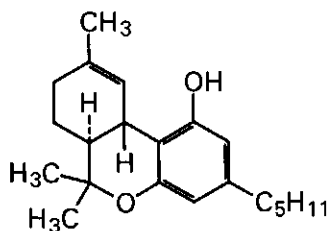
In 1991–1993, the annual world production of hemp fibre and tow was 120 000 t, of which 80 000–90 000 t was produced in Asia, mainly in India (45 000 t) and China (20 000–25 000 t). The annual world production of hemp seed in 1991–1993 was around 40 000 t, of which 22 000–25 000 t was produced in China.

There are no statistics on production in South-East Asia as the cultivation, possession, preservation, distribution, transportation and trade of hemp are forbidden by law e.g. in Singapore since 1870, in Burma (Myanmar) since 1873 and in Indonesia since 1927.

**Properties** Hundreds of different components have been isolated from hemp, and the amount of literature on their chemistry and biological activity is overwhelming. The most interesting compounds for medicinal purposes are the cannabinoids, which are mainly present in the leaves and flowering tops of female plants, and accumulate in the bracts and resin, but are absent in the seeds and stems.

Cannabinoids are terpenophenolics, classified into several groups on basis of their structures. At present, some 60 of these compounds are known, and the main representatives of each of these groups are: cannabigerol (CBG), cannabidiol (CBD), cannabichromene (CBC), cannabicydol (CBL), cannabielsoin (CBE), cannabinol (CBN), cannabiodiol (CBDL), cannabitrilol (CBTL), (-)- $\Delta^8$ -trans-tetrahydrocannabinol ( $\Delta^8$ -THC) and (-)- $\Delta^9$ -trans-tetrahydrocannabinol ( $\Delta^9$ -THC). In the latter compound,  $\Delta^9$ -THC refers to the more common dibenzopyran system of numbering; when the less common monoterpenoid system of numbering is used, this compound is called  $\Delta^1$ -THC. In each group, the cannabinoids can be present either as neutral phenolics/phenolmethylethers or as one or more isomeric acidic analogues, differing only in the presence of a carboxyl group (e.g. cannabigerolic acid, cannabidiolic acid, cannabinolic acid).  $\Delta^9$ -THC has two acidic analogues:  $\Delta^9$ -tetrahydrocannabinolic acids A (carboxyl group at position 2) and B (carboxyl group at position 4). Acidic cannabinoids are regarded as the genuine compounds; the carboxylic group, however, is very unstable: decarboxylation readily occurs, e.g. during growth of the plant, storage of plant products or upon analysis.

The biosynthesis of the cannabinoids starts with the condensation of geranylpyrophosphate and olivetolic acid (a polyketo-acid) into cannabigerolic acid. Cannabigerolic acid is an intermediate of major importance in the formation of several types of cannabinoids e.g. CBC and CBD. The latter undergoes a second cyclization to yield  $\Delta^9$ -THC-acid; decarboxylation finally gives  $\Delta^9$ -THC.  $\Delta^9$ -THC itself is not quite stable either: e.g. on prolonged storage the compound is converted into CBN via formation of an additional aromatic structure.



$\Delta^9$ -tetrahydrocannabinol

Besides the cannabinoids, the presence of various other components in *C. sativa* is documented: flavonoids (e.g. canniflavon-1, canniflavon-2), phenolic spiroindanes, dihydrostilbenes, dihydrophenanthrenes and spermidine alkaloids (cannabisativin, anhydrocannabisativin in the leaves, stems and roots). *C. sativa* contains an essential oil whose main components are  $\beta$ -caryophyllene, humulene,  $\alpha$ -pinene,  $\beta$ -pinene, limonene, myrcene and cis- $\beta$ -ocimene.

The pharmacological activity of *C. sativa* is mainly based on  $\Delta^9$ -tetrahydrocannabinol; other cannabinoids seem to have less, if any biological activity, although many of them have never been studied well. Of the two major *Cannabis* products, good quality marihuana contains 0.1–2.7%  $\Delta^9$ -THC and hashish 4–10%  $\Delta^9$ -THC. CBD and cannabidiolic acid are the main components of the glandular hairs (up to 15%); the remaining cannabinoids occur in smaller amounts.

Various preparations of *Cannabis* or  $\Delta^9$ -THC have traditionally been used for their psychological manifestations. The predominant central-nervous-system (CNS) response to  $\Delta^9$ -THC in humans include analgesia and anti-emesis, as well as a 'psychological high' state with alterations in cognition and memory, and a decrement in psychomotor performance. The acute toxicity of  $\Delta^9$ -THC is reported to be very low (e.g. 128 mg/kg intravenous in the monkey); there are no documented cases of human death caused by this component or hemp. The initial effects caused by a common 'dose' of inhaling one cigarette with 2%  $\Delta^9$ -THC, or by an oral application of 20 mg of the purified compound are described as a feeling of well-being, euphoria and relaxation, and effects on the sensorium, sense of time, short-term memory and motivation. Higher doses may induce anxiety which may become panic, dysphoria and hallucinations. Tolerance to many of these effects has been found in test animals: use in humans also led to rapid development of tolerance. Chronic use of hemp leads to a weak physical dependence, but psycho-

logical dependence is substantial and dependent on the user's history. Chronic use may also lead to paranoid psychosis. An overdose is mainly marked by a psychotic state (anxiety, suicidal tendencies, deep mental confusion) which may last for a week. Interruption of drug intake in chronic users may cause withdrawal syndrome which subsides rapidly in 3–4 days. Since the 1970s, many studies have been carried out to determine the impact of the use of hemp on health, but the long-term effects are still not well known.

Studies on the relations between the structure and activity of cannabinoids have shown that prerequisites for the psychotropic activity are the pyran structure, a stereochemical (–)-configuration, a trans configuration of the  $\Delta^9$ -bond and a free phenolic group. This might explain why both  $\Delta^9$ -THC and its metabolites formed by hydroxylation (e.g. 11-hydroxy- $\Delta^9$ -THC) are active, whereas CBG, CBD and CBC are inactive. At present, the exact action of cannabinoid drugs in the brain is still poorly understood. Very little is known yet about the neuroanatomical location of the cells responsible, or the cellular mechanisms involved. Cannabinoid drugs have been found to inhibit adenylate cyclase activity in a model neuronal system; this ability was furthermore related to the ability of these compounds to produce effects on the central nervous system. These results led to the identification of the presence of a specialized cannabinoid receptor in brain homogenates of the rat.

Besides the psychological effects, a vast array of other effects is known from the cannabinoids affecting e.g. the immune system, the hormonal system, cell growth and cell structures.

The effects on the immune system have been studied in mice, using sheep red blood cells (SRBC) as the antigen. Animals treated with  $\Delta^9$ -THC (10 and 15 mg/kg) during the primary immunization period exhibited a suppression of the primary humoral immune response. Mice treated with  $\Delta^9$ -THC during the secondary immunization period showed no measurable suppression of the secondary humoral immune response to SRBC. However, when mice were given  $\Delta^9$ -THC (10 and 15 mg/kg) during primary immunization, the secondary humoral was suppressed (existence of a memory aspect). In all experiments CBD or CBN were inactive (10 and 25 mg/kg).

When effects of cannabinoids on the hypothalamic-pituitary axis were studied in vivo in the rat,  $\Delta^9$ -THC and CBN both produced an acute suppression of plasma luteinizing hormone (LH), plasma

testosterone and hypothalamic noradrenaline (norepinefrine) metabolism. There were no effects on plasma follicle-stimulating hormone or hypothalamic LH-releasing hormone (LHRH). These results therefore suggest that decrease of LH secretion is due to reductions in noradrenaline stimulated LHRH release (hypothalamic level), rather than to changes in LHRH synthesis or pituitary LHRH response.

Cannabinoids have been found to affect the growth, proliferation and division of a variety of cell types. Reduction of cell growth and division has been observed in protozoans. Other cell systems sensitive to cannabinoid-induced effects on cell growth include HeLa cervical carcinoma cells ( $\Delta^9$ -THC,  $\Delta^8$ -THC, 11-hydroxy- $\Delta^9$ -THC and CBN), Lewis lung carcinoma cells ( $\Delta^9$ -THC,  $\Delta^8$ -THC and CBN; but CBD appeared to stimulate growth) and B103 neuroblastoma cells ( $\Delta^9$ -THC).

Cannabinoids furthermore are highly lipophilic molecules. This property can lead e.g. to cannabinoids partitioning into the lipid phase of biological membranes. Interaction has been shown with subcellular structures such as mitochondria, lysosomes and the mitotic apparatus. A reorganization of microtubules, microfilaments and neurofilaments was reported in B103 neuroblastoma cells, following treatment with  $\Delta^9$ -THC (1–100  $\mu$ M). On B103 cells, the change in cytoskeleton corresponded with changes to the overall morphology of the cells.

Numerous investigations are also available on the application of *Cannabis* or cannabinoids as drugs. The licensed medicinal use of  $\Delta^9$ -THC (dronabinol) in e.g. the United States and the United Kingdom is for prevention of nausea and vomiting in patients undergoing cancer chemotherapy. The effects of *Cannabis* on the gastro-intestinal propulsion and motility have been studied in detail in rodents. In mice and rats, intravenous injection of  $\Delta^9$ -THC slowed the rate of gastric emptying and small intestine transit. In the rat, the substance inhibited gastric emptying and small intestinal transit more than large bowel transit, indicating a selectivity for the more proximal sections of the gut. A decrease in frequency of both gastric and intestinal contractions without altering the intraluminal pressure was also found. Such changes probably reflect a decrease in propulsive activity, without changes in basal tone. CBD had no effect on gastric emptying or intestinal transit. In patients, cannabinoids as anti-emetics are as effective as the well known phenothiazines. Side-effects are relatively common. In some studies a

third of patients experienced dysphoria, and up to 80% had somnolence. It was also found that  $\Delta^9$ -THC is more rapidly and reliably absorbed from the lungs than from the gut, and patients taking the drug by smoking can thus titrate their own dose.

*Cannabis* has been reported to reduce muscle spasm and tremors in patients suffering from cerebral palsy or multiple sclerosis (MS). On the other hand, it has also been found to impair posture and balance in patients with spastic MS. There have been 3 trials with oral  $\Delta^9$ -THC in patients with multiple sclerosis. In a placebo controlled study of 9 patients, doses of 5 or 10 mg  $\Delta^9$ -THC improved spasticity compared with the placebo. Four patients exhibited an objectively measured benefit, described as substantial, 2 patients also claimed subjective improvement of symptoms. One further patient claimed subjective improvement, but this was not confirmed objectively. In a second study 2 out of 8 patients receiving doses of 5–15 mg  $\Delta^9$ -THC experienced both subjective and objective improvement in tremor. A further 5 claimed mild subjective improvement in tremor and general well-being, but this was not confirmed objectively. The third study included 13 patients with multiple sclerosis spasticity that proved untreatable with standard muscle relaxants. Using doses of 2.5–15 mg  $\Delta^9$ -THC in a double blind, placebo controlled trial, patients considered that spasticity had improved; however, neurologists blinded to the treatments could not differentiate between  $\Delta^9$ -THC and placebo. Furthermore, doses over 7.5 mg were relatively poorly tolerated, with symptoms of weakness or psychoactive effects.

In one experiment CBD was given orally to 5 patients with dystonic movement disorders. A dose-related improvement in dystonia, ranging from 20–50% was observed in all patients. In 2 patients, however, the higher doses worsened co-existing parkinsonism. CBD also appeared promising as an anticonvulsant in epilepsy. In a controlled study, adding this cannabinoid to the prescribed anticonvulsants produced improvement in 7 patients with grand mal; 3 of them showing substantial improvement.

*Cannabis* has been reported to cause bronchodilation, so *Cannabis* derivatives have therefore been tested as anti-asthma drugs. There has been preliminary research on  $\Delta^9$ -THC in the form of an aerosol spray, but other cannabinoids may also be of interest. One interesting finding for future research is that cannabinoids may affect the bronchi

by a mechanism differing from that of the more familiar anti-asthmatic drugs.

Cannabinoids might be used to treat wide-angle glaucoma, which is a major cause of blindness. In a number of patients, *Cannabis* caused a dose-related, clinically significant decrease in intra-ocular pressure, lasting several hours. Though it does not cure the disease, *Cannabis* can slow down the progressive loss of sight when conventional medicines fail and surgery is too risky. CBN and CBG have been administered to cats, topically or chronically. Whereas CBN had a modest effect on intra-ocular pressure after a single dose, it caused a more significant reduction in ocular tension during chronic administration. CBG had similar effects, but its chronic administration induced a larger response. CBN caused ocular toxicity (conjunctival erythema and hyperaemia). CBG, however, lacked these toxicities; its ocular hypotensive effect, therefore, is most interesting.

Finally, *Cannabis* and/or  $\Delta^9$ -THC may help to increase food intake and slow weight loss in e.g. cancer and AIDS patients. Preliminary clinical trials on this are showing promising results.

Aqueous extracts of hemp seeds have furthermore showed strong nematocidal activity on the larvae, eggs and cysts of the nematode *Heterodera schachtii*. An aqueous extract from the flowers had less activity, and extracts from leaves, stems and roots showed no activity. The nematocidal activity is probably due to the compounds 7-methyl-1,2,3,4-tetrahydroxy-phenazine and 3-acetyl-4-hydroxy-6-methyl-2-pyridone.

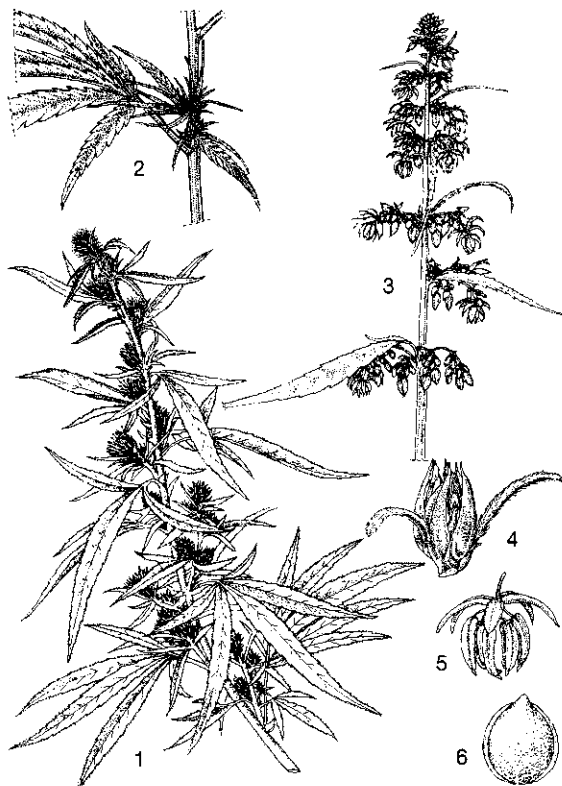
The bast of the plant cultivated for fibre contains primary and secondary bast fibres rich in cellulose but low in hemicellulose and lignin. Today, lines are available containing 67% of cellulose, 13% of hemicellulose and 4% of lignin. Primary bast fibres are 5–40 mm long and heterogenous, secondary bast fibres are smaller and more uniform with an average length of 2 mm. The woody core contains parenchyma, vessels and libriform short fibres with an average length of 0.55 mm. The chemical composition of the core fibres resembles that of hardboard with typical values of 40% cellulose, 20% hemicellulose and 20% lignin.

The seeds of hemp contain 29–35% oil, 20–24% protein and 20–30% carbohydrates. The dry seed contains up to 15% fibre, 4–6% minerals and about 6% water. No narcotically active compounds accumulate in the seed, but contamination may occur due to contact with the glands of flower bracts and leaves containing cannabinoids. Washing is sufficient to remove contamination. The

most abundant fatty acid in the seed is omega 6 linoleic acid; it comprises 54–70% of total fatty acids present.

**Adulterations and substitutes** Because the most important compounds, the cannabinoids, have not been found in plant genera other than *Cannabis*, there are no natural adulterations and substitutes. Synthetic analogues of  $\Delta^9$ -tetrahydrocannabinol have been developed, notably nabilone and levonantradol. They have undergone successful clinical trials, but still have considerable side-effects.

**Description** An annual, tall (generally 1–1.5 m tall, sometimes much taller) erect herb, usually branched, dioecious or sometimes monoecious, rather densely appressed-pubescent when young. Leaves opposite near base of stem, arranged spirally higher up, palmately compound, long-petioled; stipules free, filiform or narrowly subulate, about 0.5 cm long; leaflets (3–)5–7(–11), upper leaves often with only 1 leaflet, lanceolate from a



*Cannabis sativa* L. – 1, branch of female plant; 2, part of branch with female inflorescence; 3, part of branch of male plant; 4, female flower; 5, male flower; 6, fruit.

narrowed base, 6–14 cm × 0.3–1.5 cm, sessile, long-acuminate, coarsely serrate, on the upper surface very scabrid with short stiff hairs, on the lower surface appressed-pubescent, rather densely beset with sessile glands. Flowers unisexual; male flowers in short, dense cymes, united into foliate, terminal panicles, very shortly pedicelled, with 5 free tepals, oblong, about 5 mm long, membranous, imbricate, finely appressed-pubescent, greenish-white with pellucid white margins, stamens 5, epitepalous, with erect linear filaments and comparatively large 3–4 mm long, basifixed, 2-celled, yellow anthers; female flowers solitary in the axil of a small, primary, membranous, entire bract enveloping the ovary, each enveloped by a spathaceous, conspicuous, acuminate secondary bract, perianth absent, ovary sessile, 1-celled with a solitary pendulous ovule, style central, stigmas 2, up to 7.5 mm long, filiform, caducous. Fruit a broadly oval, much compressed achene, 4–5 mm long, with a concave rimmed base, faintly keeled on the lateral margins, smooth, shiny, yellowish or brown, closely enveloped by the secondary bract; pericarp hard, crustaceous, easily splitting into 2 halves. Seed with unilateral, scanty and fleshy albumen; embryo large, horseshoe-shaped, cotyledons large, radicle long. Seedling with epigeal germination.

**Growth and development** Hemp is normally dioecious, but monoecious cultivars have been bred; the two sexes are normally indistinguishable before flowering. In a dioecious crop, male and female plants are generally present in similar numbers, but, depending on cultivar and growing conditions, there may be up to 50% more female than male plants. Male plants die soon after anthesis, whereas female plants live 3 to 5 weeks longer than male plants, until the seed is ripe. The flowers are wind-pollinated. The total growth duration of hemp strongly depends on photoperiod and temperature. The sensitivity to photoperiod starts after formation of a few pairs of leaves and after a certain amount of heat has been received. In practice this means that the optimal production of fibre hemp is limited to regions with relatively long days, in which hemp is able to extend its vegetative growth phase sufficiently to produce long stems. The length of the flowering phase of hemp also depends on photoperiod and temperature. Between plants and in individual plants, flowering is more synchronized at shorter daylength and higher temperatures. The relatively expensive metabolites like cannabinoids and oil formed during and after flowering, may lead to a reduction of the

amount of dry mass formed per unit of intercepted radiation. The degree of branching depends on propagation methods and conditions of cultivation.

**Other botanical information** Great variation exists in hemp as a result of selection for fibre, oilseed and/or resin. This variation is further enhanced by the ease of crossing between these plant types. The widespread intergrading between different types makes all classifications inexact. A geographical classification is in use for cultivated hemp, in which North European, Central Russian, Mediterranean and Asiatic types are distinguished. The North European hemp is characterized by a short stem (< 1.5 m) and a premature flowering. Fibre and seed yields are generally low. Central-Russian hemp is cultivated in Europe and Asia between 50–60° latitude. Total growth duration is 90–110 days, with stems reaching 1.3–3 m height. Fibre yields of these types are average, but high seed yields may be obtained. Mediterranean hemp is normally cultivated south of 50° latitude in Europe, although very high fibre production may be obtained by growing these types further north. The total growth duration is 130–150 days, with stems reaching 2.5–4.5 m. Hence, fibre production can be high and of good quality. Seed yields are average because of their relatively long vegetative growth phase. Asiatic type hemp plants form branched stems of 2.5–3 m with short internodes. Growth duration is 150–170 days, but may vary considerably between populations. For practical purposes, three types can be distinguished, based on the concentrations of  $\Delta^9$ -tetrahydrocannabinol and cannabidiol: the drug (resin) type, with high  $\Delta^9$ -tetrahydrocannabinol concentration (> 1%) and no cannabidiol; the hemp (fibre) type with very low  $\Delta^9$ -tetrahydrocannabinol content (< 0.3%) and high cannabidiol concentration; and the intermediate type, with high concentrations of both compounds. However, concentrations may change during the growing season, and other components may also play a role.

**Ecology** Hemp can be grown over a great range of altitudes, climates and soils. It requires humid tropical climates to produce narcotic resin. For the production of fibre, climates of the temperate regions with temperatures of 15–27°C during the growing season are optimal. Hemp thrives on moderately to very fertile soils provided there is enough water. It is suitable for alluvial soils along streams or loamy soils with rather high rainfall. The only reliable ecological data available is for European hemp cultivated for fibre. For these cul-

tivars, the temperature requirement to reach the onset of photoperiod-sensitive phase has been quantified at 482 C°d from sowing on, and critical daylengths of 14–14.5 h have been established. Daylengths longer than the critical one extend the length of the photoperiod-sensitive phase, prolonging the vegetative growth phase. For the lower latitudes in the northern hemisphere, this sensitivity to short days limits the potential yield of hemp, while at higher latitudes low temperatures in spring are the main constraint to yield. Hemp has relatively horizontally oriented leaves, resulting in a high degree of light interception by the top of the canopy. This leads to intense shading. Although this has the advantage that weeds are suppressed, it reduces overall photosynthesis of the crop. When oil and resins are also being formed during the reproductive phase, radiation use efficiency may drop significantly. The optimal soils for hemp are sandy loams high in organic matter, with a pH around 6.

**Propagation and planting** Hemp is usually raised from seed. The seed germinates at low temperatures, but not below 1°C. Soil temperatures of 10–12°C are required for optimal crop establishment. Emergence is seriously hampered by unfavourable conditions such as soil compaction and waterlogging. Healthy seed should give 90% germination, and if properly stored it will remain viable for up to 2 years. Vegetative propagation using cuttings has been successful. However, there are morphological and biochemical differences between plants derived from seed and vegetative propagules. Vegetative propagules have higher concentrations of  $\Delta^9$ -tetrahydrocannabinol than plants raised from seed, and better developed lateral branches.

The agronomic methods depend on the product desired. For 'ganja' production in India, seed is sown in rows 1.2 m apart at a seed rate of 3–5 kg/ha, followed by a thinning when the plants are 20 cm tall. For fibre production, seed is sown densely at a rate of 30–40 kg/ha, either broadcast or in drills. Optimal plant densities of 90–120 plants/m<sup>2</sup> with row distances of 12–20 cm seem optimal, but row distances of more than 1 m and inter-plant distances between 15–50 cm are used for seed production. In China seed plants are sometimes sown in clusters.

**Husbandry** Weeding is rarely necessary because the dense canopy shades out weeds. However, weeds may be a problem in the establishing crop and in gaps. Practical experience with fibre hemp on optimal soils has shown that for the pro-

duction of 1000 kg stem material an annual fertilization of 15–20 kg N, 4–5 kg P<sub>2</sub>O<sub>5</sub> and 15–20 kg K<sub>2</sub>O is required. The need for nitrogen is highest during the vegetative growth phase in which green leaf material is produced. Requirements for phosphorus and potassium increase gradually during vegetative growth, peaking around flowering before slowly declining again. In an increasing order and within certain limits, fibre quality is improved by the nitrogen, phosphorus and potassium contents of the hemp plant. Hemp is a suitable crop for rotation with almost any crop, though some problems may occur in rotations with beets, because of nematode infestations. Hemp suppresses weeds and loosens the soil for the following crops. Moreover, root and leaf material may be left in the field to serve as organic material for the next crop. The male plants produce the best fibre and are sometimes harvested first; the female plants are sometimes allowed to stand to set seed for oil production. The Asian practice of removing male plants is not because of their lower content of  $\Delta^9$ -tetrahydrocannabinol, but to prevent seed production in female plants, which would reduce resin production.

**Diseases and pests** Diseases and pests in hemp may be plant specific or general. Reported damage by hemp-specific organisms are from *Grapholita delineaana*, *Melasporea cannabina*, *Phorodon cannabis*, *Psylliodes attenuata* and *Sep-toria cannabis*. Seeds of hemp may be infected by the parasitic *Orobanche racemosa* L. or hemp killer. The most important non-specific diseases and pests in hemp are: *Botrytis cinerea*, *Ostrinia nubilalis* and *Sclerotinia sclerotiorum*. Yield may also be depressed by *Cuscuta europaea* L., *Fusarium* spp., *Ditylenchus dipsaci*, *Tetranychus urticae*, some insects of the *Noctuidae* and larvae of *Agriotes lineata*, *Melolontha melolontha* and *Tipula paludosa*. Hemp may also suffer from nematodes such as *Meloidogyne hapla* in northern Europe and, in northern India *Neottolenchus clarus* and *Quinsulcius similis*. The role played by essential terpenoid substances in repelling insects and exudates of cannabinoids (as antibiotics) has been neglected and deserves investigation. Another of hemp's natural defence mechanisms - its covering of non-glandular trichomes - might serve as a mechanical defence against predators.

**Harvesting** How hemp is harvested depends on the product. For 'ganja' production, male plants are pulled out as soon as they are recognized and before the pollen is shed. Unfertilized female plants are left, and harvested when flower stalks

begin to turn yellow, at about 5 months after sowing. For the production of 'charas', the resin is collected by men who run through the plantings clad in leather garments. The resin sticks to the garments and is then scraped off. Another method is to collect the resin by squeezing plant tops between the palms of the hand. Plants are harvested for fibre manually or by machinery. In China, manual harvest is by cutting the stems; the branched plants are left on the edges of the field and their seed is harvested for sowing.

**Yield** The average yield of 'ganja' in India is about 280 kg/ha. Stem yields (yielding approximately 25–35% fibre) are usually between 3–8 t/ha, with a potential of 20 t/ha. When hemp is grown solely for seed production, yields of 1300–1700 kg seed per ha may be obtained.

**Handling after harvest** For the production of 'ganja', harvested inflorescences are trodden and pressed into flat cakes. For fibre production the cut stems are graded by pulling out the longest and medium stems, respectively. The short and twisted stems are for local use. Leaves are stripped off with a knife. The stems are then dried on the field for 2–4 days. About 200 stems are then bundled and immersed in water for a 3-day retting. After retting, another 2–3 days are used for field drying. Depending on the degree of retting, a second retting is carried out or stems are allowed to dry further in the field, this time in bundles. Fibre bundles are subsequently stripped off by hand from the partly wet stems and dried on lines before marketing. Another method to obtain the fibres is to dry the stems completely before breaking and to comb them. The yield of ribbon with this dry method is slightly higher than with the semi-wet method; 10% versus 6% relative to dry stem mass. In Europe, hemp is harvested mechanically by cutting, drying in swathes on the field for a few weeks, and subsequently pressing and baling in one operation. The yield of ribbon is on average 15%. The centres for commercial production of sowing seed for fibre hemp are in France, Hungary and Poland. The fineness and coarseness of the fibre and cleanness or degree of retting are decisive for traditional processing of ropes and textiles. Sometimes, male and female plants are therefore separated and processed differently. The stems are also graded during harvest. On modern spinning machines, in which production speeds are the determining factor, fibre length may be a limitation to the use of bast fibre. Parallel processing of the fibrous raw material is required, to prevent entanglement; this adds sub-



stantially to the costs of the fabrics. Hemp fibres may easily be used for paper production in which the chemical content of the fibres mainly determines the quality of the pulp. The drainability and bleachability of the pulp and sheet forming are among the many factors that determine the applicability of the pulp. For the application of fibre in composite material the following properties are of interest: high tensile strength, rigidity, impact resistance, small volume shrinkage during curing, resistance to corrosion, low density, non-toxic, recyclability, ease of disposal and economic price. These quality characteristics are partly inherent to the natural fibre, but those like tensile strength and rigidity are determined by cultivar choice, growing conditions, environment, mechanization and processing.

**Genetic resources and breeding** There has been limited preservation of germplasm of *C. sativa* in gene banks because of its bad image as a narcotic. Together with the declining interest in breeding and maintaining of cultivars, this has led to an impoverishment of germplasm resources. However, a large reservoir of natural variation is maintained by wild forms.

The difference in timing of anthesis of male and female plants promotes outbreeding. Production of large amounts of pollen and wind pollination tend to lead to extensive genetic exchange between different domesticated forms and between domesticated and wild plants. Breeding has mostly focused on the creation of monoecious varieties. In Europe, breeding and selection work is directed at obtaining hemp types with a bast fibre content higher than 30% and  $\Delta^9$ -tetrahydrocannabinol levels below 0.3%.

**Prospects** Although the resin present in hemp has been recognized to have therapeutic value, the use of hemp as a medicinal plant is limited. This is mainly because the cultivation, possession, preservation, transportation and trade of hemp is prohibited in most countries. However, research is increasingly being initiated on hemp drugs for relieving patients suffering from diseases such as multiple sclerosis, cancer, AIDS and glaucoma. Since many young people in big cities smoke 'ganja' or 'marihuana', the drug is probably being smuggled into South-East Asia, and illegal cultivation of hemp in remote areas may still be found. Hemp grown for fibre has a long history and still has very good prospects. The plant fibre products have major advantages in various branches of industry. They are biocompatible and biodegradable, thereby reducing the environmental burden

caused by consumer goods and disposables, building and construction material and civil engineering. The research priorities include: determining optimal primary production techniques, developing field fibre extraction techniques, quantifying the relationships between primary production conditions, processing and fibre quality, and optimizing the management of production chains to best use all components of hemp.

**Literature** |1| Coffman, C.B. & Gentner, W.A., 1979. Greenhouse propagation of *Cannabis sativa* L. by vegetative cuttings. *Economic Botany* 33(2): 124-127. |2| Devane, W.A., Dysarz, F.A., Johnson, M.R., Melvin, L.S. & Howlett, A.C., 1988. Determination and characterization of a cannabinoid receptor in rat brain. *Molecular Pharmacology* 34: 605-613. |3| Gray, C., 1995. Cannabis - The therapeutic potential. *Pharmaceutical Journal* 254: 771-773. |4| Grinspoon, L., 1995. The hemp plant as a source of medicine. In: *Biorohstoff Hanf. Proceedings of the symposium, 2-5 March 1995*. 2nd edition. Nova-Institut, Köln, Germany. pp. 568-575. |5| Small, E., 1979. The species problem in *Cannabis*; science and semantics. Vol. 2: *Semantics*. Corpus, Toronto, Canada. 156 pp. |6| Small, E., 1995. Hemp. In: Smartt, J. & Simmonds, N.W. (Editors): *Evolution of crop plants*. 2nd edition. Longman, London, United Kingdom. pp. 222-223. |7| Small, E. & Cronquist, A., 1976. A practical and natural taxonomy for *Cannabis*. *Taxon* 25(4): 405-435. |8| Turner, C.E., Elsohly, M.A. & Boeren, E.G., 1980. Constituents of *Cannabis sativa* L. 17. A review of the natural constituents. *Journal of Natural Products* 43(2): 169-234. |9| van der Werf, H.M.G., 1994. *Crop physiology of fiber hemp (Cannabis sativa L.)*. PhD thesis, Wageningen Agricultural University, the Netherlands. 153 pp. |10| van Soest, L.J.M., Mastebroek, H.D. & de Meijer, E.P.M., 1993. Genetic resources and breeding: a necessity for the success of industrial crops. *Industrial Crops and Products* 1: 283-288.

**Other selected sources** 94, 97, 98, 174, 193, 202, 273, 402, 406, 491, 492, 549, 621, 667, 693, 709, 1126, 1133, 1167, 1172, 1277, 1338, 1339, 1401, 1437, 1580.

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**Cardiospermum halicacabum L.**

Sp. pl. 1: 366 (1753).

SAPINDACEAE

2n = 22

**Synonyms** *Cardiospermum corindum* L. (1762), *Cardiospermum microcarpum* Humb., Bonpl. & Kunth (1841), *Cardiospermum luridum* Blume (1847).

**Vernacular names** Balloon vine, heart pea (En). *Coeur des Indes*, *pois de coeur* (Fr). Indonesia: ketipes (Javanese), paria gunung (Sundanese), cenet (Malay, Western Sumatra). Malaysia: peria bulan, uban kayu, bintang berahi. Philippines: parol-parolan (Tagalog), kana (Cebu Bisaya), paria-aso (Iloko). Thailand: kok kra om (central), pho om (Pattani), luupleep khrua (northern). Vietnam: t[aa]flm phong, ch[uf]lm phong.

**Origin and geographic distribution** *C. halicacabum* probably originates from the New World tropics. At present, it is a common weed in tropical and subtropical regions throughout the world, and is common throughout Malesia. It is reported to be cultivated in the Philippines and Burma (Myanmar).

**Uses** The root is the most important plant part used for medicinal purposes. In South-East Asia it is considered to be diaphoretic, diuretic, emetic, antipyretic and purgative. The roots are reported to be used in Indonesia and the Philippines against catarrh of the bladder and urinary tract. The leaves are considered antirheumatic in the Philippines, where they are either taken internally or applied externally. In Indonesia, the bruised leaves are used as a cooling compress for nervous headaches. The leaves are reported to be used to treat eye problems in Malaysia, Thailand and Vietnam.

In India, the root is considered diuretic, diaphoretic, emetic, laxative, emmenagogic and rubefacient and is sometimes used to treat rheumatism, lumbago and nervous diseases. The leaves are reported to be rubefacient and useful as a poultice to treat rheumatism, and the leaf juice to be a cure for earache. In Bangladesh, pills made from a paste of the whole plant are used to treat asthma. In China, a decoction of the plant is used as a post-surgery wash, and a tea of the leaves is rubbed on itching skin. In Taiwan, leaves are applied to swellings, together with salt. In southern Africa an infusion of the leaves and stalk is applied as an enema to cure diarrhoea and dysentery; a similar method has been reported for In-

donesia. The vapour of the crushed leaves is inhaled to relieve headache.

Apart from its medicinal uses, *C. halicacabum* is eaten as a vegetable, the stems serve to make baskets and the seeds are used as beads. The leaves are reported to be used for washing clothes and the head. An edible oil can be obtained from the seed.

**Properties** The seeds contain about 30% oil, consisting of the glycerides of arachidic, lignoceric, stearic, oleic and linoleic acids and glycerol. Furthermore, the seed oil has been found to contain large amounts (55%) of cyanolipids (e.g. cardiospermin) and to consist for a large part (45%) of C<sub>20</sub> acids. Cyanolipids have not been found in families other than the *Sapindaceae*. Together with cyanogenous glycosides, these compounds provide plants with the ability to produce hydrocyanic acid, often through the use of enzymes that are activated when plant tissue is damaged. The extremely poisonous nature of hydrocyanic acid can thus protect plants, for instance against damage by insects. Hydrocyanic acid is also dangerous to man; cyanide acts by inhibiting the cytochrome oxidase system for oxygen utilization in cells. Other respiratory enzymes are also inhibited, but to a lesser degree. However, large amounts of plant material often have to be consumed to achieve dangerous concentrations (0.5–3.5 g/kg).

The unsaponifiable fraction of the pericarp is reported to contain pigments and flavones, whereas that of the inner parts of the seeds is characterized by triterpenoids and steroids. The latter fraction showed anti-inflammatory activity in rats.

An ethanolic extract of the aerial parts of *C. halicacabum* has shown anti-inflammatory activity in rats using the carrageenin-induced rat paw oedema test. The mechanism of action is most probably either the inhibition of phospholipase-II activity, resulting in reduced availability of arachidonic acid, or the stabilization of the lysosomal membrane system. An ethanolic leaf extract produced depression of the central nervous system, fall in blood pressure and bradycardia in isolated organ preparations and in vivo. The fall in blood pressure was partly antagonized by atropine and antihistaminics. A possible mechanism may be the inhibition of angiotensin-converting enzyme. On the guinea-pig ileum preparation, the extracts produced a strong contraction, which could also be partly antagonized by atropine and antihistaminics.

Another pharmacological experiment was an in vivo test in rats, in which an extract produced mild

analgesia and showed proconvulsant and anti-inflammatory activity as tested by the granuloma pouch and cotton pellet implantation test. The essential oil and the water-soluble fraction of a dried alcoholic leaf extract gave an immediate fall of blood pressure in anaesthetized dogs; this hypotensive action was not affected by atropine. Furthermore, the water-soluble fraction of a dried alcoholic extract of the seeds produced an initial depression, followed by a marked stimulation of isolated frog heart. Finally, methanolic extracts of dried plants have shown antisickling and anticrenation activity of erythrocytes, with observed reversals being more potent than the effect of testosterone propionate.

In addition to the above-mentioned cyanolipids and cyanogenic glycosides, phytochemical investigations of *C. halicacabum* have revealed a variety of compounds: quebrachitol, steroids (stigmasterol,  $\beta$ -sitosterol), flavonoids (apigenin and acacetin), phenolic acids (p-hydroxybenzoic acid, vanillic acid, melilotic acid, p-coumaric acid and ferulic acid), alkaloids, tannins, proanthocyanidins, aromatic nitrosulfones and saponins. The saponins may account for the diuretic properties and make external preparations slimy for poulticing. Furthermore, saponins characteristically form foamy solutions in water, and saponin-containing seeds are often used as detergents. Most saponins have haemolytic properties, and thus are toxic to cold-blooded animals, hence employed as a fish poison. They are usually only weakly toxic for warm-blooded species when taken orally, but more dangerous when given parenterally, because of their direct haemolytic action.

**Description** An annual or perennial climbing herb or subshrub, up to 3 m tall, often much branched, especially near the base; stems deeply 5-grooved, slender, glabrous to sparsely hairy. Leaves alternate, compound, biternate, 5–8 cm  $\times$  5–8 cm, petiole 1.5–3 cm long, grooved, slender, with minute stipules at the base; leaflets mostly 3-partite and pinnately lobed, lobes and apex aristulate, subglabrous to sparsely covered with short appressed hairs, petiolules narrowly winged, petiolule of terminal leaflet about 1 cm long, those of lateral ones about 0.5 cm. Inflorescence thyrsoid, axillary, 5–14 cm long, patent, sparsely short-hairy, with a pair of tendrils, peduncle 7–10 cm long, slender, slightly above the tendrils terminated by a pseudo-whorl of 3 uniparous, helicoid cymes, which are bracteate, spreading, long-stalked and few-flowered; bracts lanceolate to elliptical, 1–2 mm long. Flowers unisexual, 2–3.5 cm



*Cardiospermum halicacabum* L. - 1, plant habit; 2, male flower with sepals and petals removed; 3, pistillode of male flower; 4, female flower with sepals and petals removed; 5, pistil of female flower; 6, fruit; 7, seed.

long, obliquely zygomorphic, with slender pedicel; sepals 4, broadly ovate to broadly elliptical, 1–2.5 mm  $\times$  1.2–2 mm, free, imbricate, green, tinged red with white margins, subglabrous; petals 4, obovate-cuneate to orbicular, 1.5–2.5 mm  $\times$  1–2 mm, with a scale inside above the base of each petal, white to creamish with yellowish margin, almost glabrous; stamens 8, slightly curved upwards, unequal, filaments 0.8–2.5 mm long, only slightly reduced in female flowers, slightly hairy, anthers 0.5 mm long; ovary superior, obovoid, 2–3 mm long, 3-angled, 3-celled, with 1 ovule per cell, variably pubescent, with a short columnar style and 3-lobed stigma, in male flowers pistil strongly reduced. Fruit a globular capsule, inflated, 1.5–4 cm in diameter, 3-lobed, 3-celled, green, reddish at base or with reddish veins, papyraceous. Seeds subglobular, about 4 mm in diameter, dull-black, smooth, glabrous, hilum prominent, white, cordate, rather large.

**Growth and development** *C. halicacabum* can be found flowering and fruiting throughout the year, except for prolonged periods of drought. The first flower in every cyme is usually female, all others male. Fruits are dispersed by water currents, over short distances by wind but mainly as a result of human activities.

**Other botanical information** *C. halicacabum* belongs to a genus of about 12 species, most of them restricted to tropical and subtropical America. *C. grandiflorum* Swartz, found in Africa and the Americas, is cultivated as an ornamental in Malesia and sometimes naturalized. It also contains cyanogenic compounds in its leaves.

**Ecology** *C. halicacabum* is found under a wide range of ecological conditions: in everwet or seasonal climates, on acid and basic soils, and in dry, marshy or periodically flooded places. It prefers sunny places, such as wasteland, roadsides, grassland, scrub, hedges and forest edges, at altitudes up to 1500 m.

**Propagation and planting** *C. halicacabum* can be propagated by seed and softwood cuttings. Seeds germinate at temperatures from 15–40°C with an optimum of 35°C, and taking about 3 weeks. Scarification with concentrated sulphuric acid may facilitate germination.

**Diseases and pests** Balloon vine is a major problem in soya bean cropping in the United States. The size and shape of the seeds is very similar to those of soya bean seeds, which makes mechanical separation of the two very difficult.

**Harvesting** *C. halicacabum* is mostly collected from the wild.

**Genetic resources and breeding** As *C. halicacabum* is a common weed throughout the tropics and subtropics, the risk of genetic erosion appears limited.

**Prospects** Because *C. halicacabum* contains potentially toxic compounds such as saponins, cyanolipids and cyanogenic glycosides, it seems advisable to carry out research on its toxicity for humans before promoting its use for medicinal purposes.

**Literature** [1] Gopalokrishnan, C., Dhananjayan, R. & Kameswaran, L., 1976. Studies on the pharmacological actions of *Cardiospermum halicacabum*. *Indian Journal of Physiology and Pharmacology* 20(4): 203–208. [2] Gurib-Fakim, A. & Sewraj, M.D., 1992. Studies on the antisickling properties of extracts of *Sideroxylon puberulum*, *Faujasiaopsis flexuosa*, *Cardiospermum halicacabum*, and *Pelargonium graveolens*. *Planta Medica* 58(7 suppl. 1): A648–A649. [3] Hegnauer, R.,

1973. *Chemotaxonomie der Pflanzen* [Chemotaxonomy of plants]. Vol. 6. Birkhäuser Verlag, Basel, Boston, Stuttgart. pp. 271–287. [4] Johnston, S.K., Murray, D.S. & Williams, J.C., 1979. Germination and emergence of balloonvine (*Cardiospermum halicacabum*). *Weed Science* 27(1): 73–76. [5] Leenhouts, P.W., 1994. *Cardiospermum*. In: de Wilde, W.J.J.O., Nooteboom, H.P. & Kalkman, C. (Editors): *Flora Malesiana*. Series 1, Vol. 11(3). Rijksherbarium/Hortus Botanicus, Leiden, the Netherlands. pp. 483–486. [6] Mikolajczak, K.L., Smith, C.R. & Tjarks, L.W., 1970. Cyanolipids of *Cardiospermum halicacabum* L. and other sapindaceous seed oils. *Lipids* 5(10): 812–817. [7] Plouvier, V., 1949. Nouvelles recherches sur le quérachitol des Sapindacées et Hippocastanacées, le dulcitol des Celastracées et le saccharose de quelques autres familles [New researches on quérachitol of Sapindaceae and Hippocastanaceae, dulcitol of Celastraceae and saccharose of some other families]. *Comptes Rendues hebdomadaires des Séances de l'Académie des Sciences* 228: 1886–1888. [8] Sadique, J., Chandra, T., Thenmozhi, V. & Elango, V., 1987. Biochemical modes of action of *Cassia occidentalis* and *Cardiospermum halicacabum* in inflammation. *Journal of Ethnopharmacology* 19(2): 201–212. [9] Umadevi, I. & Daniel, M., 1991. *Chemosystematics of the Sapindaceae*. *Feddes Repertorium* 102(7–8): 607–612. [10] Wijayakusuma, H.M.H., Dalimartha, S. & Wirian, S.W., 1994. *Tanaman berkhasiat obat di Indonesia* [Plants yielding medicine in Indonesia]. Vol. 3. Pustaka Kartini, Indonesia. pp. 99–101.

**Other selected sources** 1, 19, 46, 97, 104, 190, 193, 202, 242, 287, 332, 412, 531, 580, 551, 900, 921, 955, 1035, 1085, 1126, 1128, 1178, 1221, 1305, 1380, 1554.

J.P. Rojo & F.C. Pitargue

### **Carmona retusa (Vahl) Masam.**

*Trans. Nat. Hist. Soc. Formosa* 30: 61 (1940).

BORAGINACEAE

2n = 32

**Synonyms** *Ehretia microphylla* Lamk (1792), *Ehretia buxifolia* Roxb. (1796), *Carmona microphylla* (Lamk) G. Don (1837).

**Vernacular names** Indonesia: kinangan, serut lanang (Javanese), pinaan (Madurese). Philippines: putputai (Bikol), alangit (Bisaya), tsaang gubat (Tagalog). Thailand: khoi cheen (Bangkok), chaa yeepun (central), chaa (Chiang Mai). Viet-

nam: kim li[ee]n, c[uf]m r[uj]n, b[uf]m r[uj]n.

**Origin and geographic distribution** *Carmona* is a monotypic genus. The only species *C. retusa* is found from India eastward to southern China, Taiwan and Japan, and further south throughout Malesia to New Guinea and the Solomon Islands. It is often grown as an ornamental.

**Uses** In the Philippines an infusion of the leaves of *C. retusa* is taken as a substitute for tea. It is considered stomachic, antidiarrhoeal and as a remedy for dysentery and cough. In Ternate an infusion of the leaves is taken as a febrifuge. A decoction of the leaves is taken against stomach troubles and cough in the Philippines. In Madura the roots are reported to be ingested to clean the body after childbirth. In India, the plant is considered an antidote against plant-based poisoning and an alterative in cachexia and syphilis. Furthermore, it is traditionally used to stop the haemorrhaging resulting from the bite of the viper *Echis carinatus*.

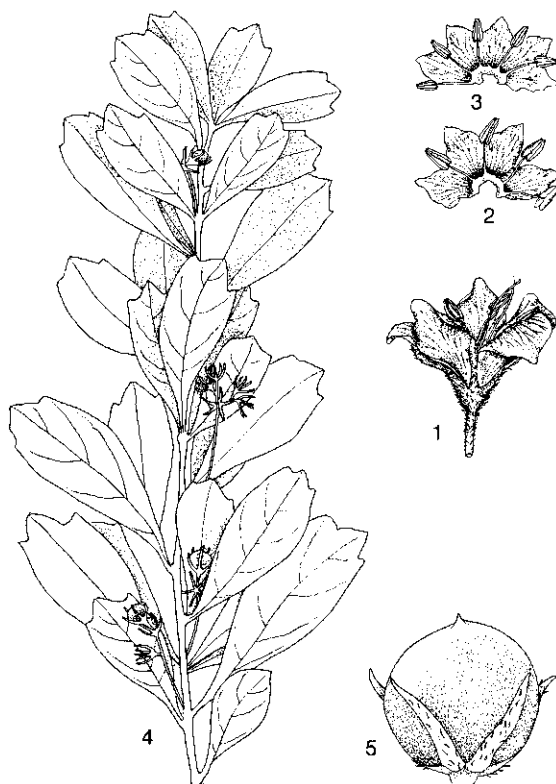
**Properties** *Boraginaceae* commonly contain pyrrolizidine alkaloids, biosynthesized from the amino acid ornithine, and quinoid or phenolic compounds, derived from C-prenylated, C-geranylated or C-farnesylated 4-hydroxybenzoic acid. The isomeric red pigments alkannin and shikonin are the best known representatives of such hydroxybenzoic acid derivatives.

The portion of the methanol extract of the aerial parts of *C. retusa* that is soluble in ethyl acetate has shown inhibitory activity on exocytosis of hexosaminidase in antigen-stimulated rat basophils. Activity guided studies have shown 5 dimeric prenylbenzoquinones as active compounds: microphyllone (4a,5,8,8a-tetrahydro-11,14-dihydroxy-7-methyl-4a-(3-methyl-2-butenyl)-5,8a-o-benzo-1,4-naphthoquinone) and 4 closely related components. Furthermore, the methanol extract of the leaves showed strong antihistamine release properties. Rosmarinic acid, a phenylacrylic acid derivative, which is a known inhibitor of histamine release, has been isolated as an active constituent. Astragalol, nicotiflorin (both flavonoid glycosides), and  $\alpha$ -amyrin,  $\beta$ -amyrin and baurenol (triterpenoids) were also isolated.

Ehretianone, a quinonoid xanthene, (7-hydroxy-9a- $\alpha$ -(3-methylbut-2-enyl)-4a, $\alpha$ 9- $\alpha$ -(2-methylprop-2-enyl)-4a,9a-dihydro-1,4-dioxoxanthene) was isolated from a methanol extract of the root bark of *C. retusa*. The compound, administered before and after administration of *Echis carinatus* venom, was shown to protect mice against the action of the snake venom.

Finally, both microphyllone and ehretianone isolated from root-bark material showed antibacterial activity against a panel of bacteria. In an experiment in the Philippines, tablets from the dried leaves reduced the formation of micronucleated polychromatic erythrocytes induced by mitomycin C, tetracycline, and dimethylnitrosamine. This suggests that these tablets possess antimutagenic activity.

**Description** A shrub or much-branched small tree, 1-4(-10) m tall; young branches hispid, with buds or short shoots producing clusters of leaves and inflorescences. Leaves simple, alternate, obovate to spatulate, 1-6(-10) cm  $\times$  0.5-2.5(-4) cm, thick, gradually narrowing towards base, toothed or crenate towards apex, with short rigid hairs, lateral veins about 5, arching; petiole 0.1-0.5(-1) cm long, stipules absent. Inflorescence in axil of leaves or on apex of short shoots, flowers in fascicles of 2-6 or in a cyme. Flowers actinomorphic, bisexual, (4-)5-merous, pedicelled; calyx 3-6 mm long, with (4-)5 linear lobes, densely hairy inside;



*Carmona retusa* (Vahl) Masam. - 1, flower; 2, 4-merous corolla; 3, 5-merous corolla; 4, fruiting twig; 5, fruit.

corolla sub-rotate, white, 6–9 mm in diameter, tube about 2 mm long, widening, lobes spreading, 2.5–4.5 mm long; stamens (4–)5 with filaments 2.5–3.5 mm long and anthers oblong; ovary superior, globose, about 1 mm in diameter, style deeply bifid, 4.5–6 mm long. Fruit drupaceous, globose, 5–6 mm in diameter, red or yellow, with 1–4 seeds, not breaking up into pyrenes. Seeds with a straight or slightly curved embryo, embedded in thin albumen. Seedling with epigeal germination; cotyledons leafy, green, hypocotyl elongated.

**Growth and development** Branch shoots are developed at every node, but are of two types: short shoots that do not elongate and long shoots that resemble the leader shoot. The fruits are reported to be dispersed by birds.

**Other botanical information** In the view of most authors, the *Boraginaceae* family can be divided into five subfamilies. *Carmona* belongs to the subfamily *Ehretioideae*. Together with the other Malesian representatives of this subfamily (*Coldenia*, *Ehretia* and *Rotula*) and with the subfamily *Cordioideae* (represented by *Cordia* in Malesia) *Carmona* is sometimes placed in the family *Cordiaceae*. Most authors have included *Carmona* in *Ehretia*. However, *Carmona* can be distinguished by the undivided, short-beaked endocarp, and its distinctive growth habit and general appearance.

**Ecology** *C. retusa* is a rare or locally common species found in open, dry, sunny habitats, such as thickets, shrub vegetation and teak forest at low and moderate elevations.

**Propagation and planting** *C. retusa* can be propagated by cuttings, preferably top shoots or young leafy shoots. Roots develop slowly (1–2 months). Planting is at 1 m intervals with 2 m between rows.

**In vitro production of active compounds** Using leaf tissue of *C. retusa* as explant, callus growth can be observed after 2 days. The callus contains alkaloids.

**Harvesting** Mature leaves of *C. retusa* are harvested and senescent leaves discarded.

**Handling after harvest** In the Philippines, leaves of *C. retusa* are air-dried for 4–5 days in shallow containers with screened bases. Containers are kept in well-ventilated rooms. The dried leaves are powdered and processed into pills.

**Genetic resources and breeding** *C. retusa* is a widespread species, and there are no reports of overexploitation. The risk of genetic erosion seems to be limited, and in addition it is cultivated as a garden ornamental.

**Prospects** *C. retusa* ranks among the top 10 of medicinal plants with potential in the Philippines, and thus a small-scale industry has developed. Its constituents, e.g. microphyllone and ehretianone show interesting biological activities, which merit further research. Furthermore, alkannin esters of various organic acids have recently been introduced in therapy for their wound-healing properties.

**Literature** |1| Agarwal, S.K., Rastogi, R.P., Van Koningsveld, H., Goubitz, K. & Olthof, G.J., 1980. The molecular structure of 4a,5,8,8a-tetrahydro-11,14-dimethoxy-7-methyl-4a-(3-methyl-2-butenyl)-5,8a-o-benzo-1,4-naphthoquinone. *Tetrahedron* 36(10): 1435–1438. |2| Bruneton, J., 1995. Pharmacognosy, phytochemistry, medicinal plants. Technique & Documentation Lavoisier, Paris, France. pp. 675–679. |3| Gopal, R.H., Balakrishna, K., Ramkumar, V., Rao, R.R., Selvanayagam, Z.E. & Gnanavendhan, S., 1997. Antibacterial activity of *Ehretia buxifolia*. *Fitoterapia* 68(1): 85–86. |4| Gutierrez, H.G., 1982. An illustrated manual of Philippine materia medica. Vol. 2. National Research Council of the Philippines, Tagig, Metro Manila, the Philippines. pp. 445–446. |5| Lim-Sylianco, C.Y., Blanco, F.R.B. & Lim, C.M., 1987. Mutagenicity, clastogenicity and antimutagenicity of medicinal plant tablets produced by the NSTA Pilot Plant IV. Tsaang gubat tablets. *The Philippine Journal of Science* 116(3): 273–280. |6| Quintana, E.G., Saludez, J.D., Batoon, M.P. & Generalao, M.L., 1982. Agricultural production of selected medicinal plants: propagation to postharvest handling. *PCARRD Monitor* 10(4): 8–10. |7| Riedl, H., 1997. *Boraginaceae*. In: Kalkman, C. et al. (Editors): *Flora Malesiana*. Series 1. Vol. 13. *Rijksherbarium/Hortus Botanicus*, Leiden, the Netherlands. pp. 43–144. |8| Rimando, A.M., Inoshiri, S., Otsuka, H., Kohda, H., Yamasaki, K., Padolina, W.G., Torres, L., Quintana, E.G. & Cantoria, M.C., 1987. Screening for mast cell histamine release inhibitory activity of Philippine medicinal plants active constituent of *Ehretia microphylla*. *Shoyakugaku Zasshi* 41(3): 242–247. |9| Selvanayagam, Z.E., Gnanavendhan, S.G., Balakrishna, K., Rao, R.B., Sivaraman, J., Subramanian, K., Puri, R. & Puri, R.K., 1996. Ehretianone, a novel quinonoid xanthene from *Ehretia buxifolia* with antisnake venom activity. *Journal of Natural Products* 59(7): 664–667. |10| Yamamura, S., Simpol, L.R., Ozawa, K., Ohtani, K., Otsuka, H., Kasai, R., Yamasaki, K. & Padolina, W.G., 1995. Antiallergic dimeric prenylbenzoquinones from *Ehretia microphylla*. *Phytochemistry* 39(1): 105–110.

**Other selected sources** 97, 190, 202, 213, 332, 686, 921, 1020, 1126, 1178, 1295, 1564.

A.P. Guevara

## Cassia L.

Sp. pl. 1: 376 (1753); Gen. pl. ed. 5: 178 (1754).

LEGUMINOSAE

$x = 12, 14$ ; *C. fistula*:  $2n = 24, 28$ , *C. grandis*:  $2n = 28$

**Major species** *Cassia fistula* L., *C. grandis* L.f.

**Origin and geographic distribution** *Cassia* comprises about 30 species and is pantropical. There is probably only one indigenous species in Malesia (*C. javanica*) and three other species have been introduced from mainland Asia or South America, originally for ornamental purposes: *C. bakeriana* Craib, *C. fistula* and *C. grandis*. The medicinal use of *C. fistula* dates from ancient times and has been the main factor in its spread.

**Uses** The ripe pods and seeds of *C. fistula*, *C. grandis* and *C. javanica* are used as a laxative. Other plant parts (root-bark, leaves, flowers) of *C. fistula* also have purgative properties, but to a lesser extent. A decoction of the roots may be used to purify wounds and ulcers. The bark of the trunk is reported to be used in Java and India to treat skin problems, whereas in the Philippines the leaves are applied on fungal skin infections. In India, the roots are used to treat fever. Fresh juice of the leaves of *C. grandis* is used externally in the treatment of ringworm. Leaf decoctions are also used as laxative and to treat lumbago. In Panama, *C. fistula* is used in folk medicine for the treatment of diabetes.

In modern medicine, the pulp of *C. fistula* is sometimes used as a mild laxative in pediatrics. However, laxative drugs of this type should be used with caution, because daily and prolonged use may lead to dependence and 'cathartic colon'.

Apart from its medicinal properties, *Cassia* has many other uses. *C. bakeriana*, *C. fistula*, *C. grandis* and *C. javanica* are all planted as ornamentals. The latter three species and especially *C. javanica* provide hard multipurpose timber. In Thailand, the heartwood of *C. fistula* is used as a masticatory. The bark of *C. fistula* and *C. javanica* is also used for tanning, but the latter species is less valued. The seeds of *C. fistula*, *C. grandis* and *C. javanica* are a potential commercial source of gums. Seed gum is a potential binder for the pharmaceutical industry.

**Production and international trade** Al-

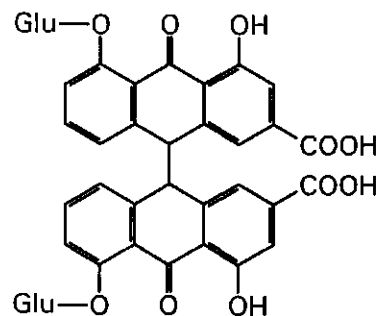
though *C. fistula* pods have been traded to Europe for centuries, no recent trade information is available.

**Properties** The fruit pulp of *C. fistula* is rich in pectins and mucilages. Furthermore, the laxative properties of fruit pulp and leaves are mainly due to the content of anthraquinone derivatives (about 2%), e.g. rhein (an anthraquinone), the sennidins (dianthrone) and the sennosides (the corresponding dianthrone-glycosides). In a study in Mexico, sennoside contents were found up to 1.5% in the leaves, and up to 1.9% in the fruits. Anthraquinone drugs are used as laxatives. The sugar moiety in the glycosides increases water solubility of the molecule, and thus facilitates transport to the site of action: the colon. In the colon, bacteria hydrolyse the glycosides and dianthrone to anthraquinones, a reaction which is immediately followed by the local reduction of the anthraquinones to their corresponding anthrones. The latter compounds act directly on the large intestine, to stimulate peristalsis. Anthraquinones are also found in *C. javanica* (anthraquinone glycosides) and *C. grandis* (aloe-emodin).

In an Indian study, fatty acids in *C. fistula* were found to consist mainly of linoleic acid (52.5%), oleic acid (18.1%) and palmitic acid (16%). Other fatty acids were: vernolic acid (6.1%), stearic acid (3.4%), sterculic acid (2%) malvalic acid (1.5%) and myristic acid (0.4%).

Compounds isolated from *C. grandis* include centaureidine, catechin, myristicin, 2,4-dihydroxybenzaldehyde, 3,4,5-trimethoxybenzaldehyde, 2,4,6-trimethoxybenzaldehyde,  $\beta$ -sitosterol, koku-saginine (6,7-dimethoxyfuroquinoline) and fabio-line (1,1'-bipiperidine).

In vitro and in vivo tests showed that seed powder of ripe *C. fistula* fruits has amoebicidal and cysticidal properties against *Entamoeba histolytica* and that it could cure intestinal and hepatic



sennoside A (Glu = glucose)

amoebiasis of laboratory animals and intestinal amoebiasis of humans. *C. fistula* also exhibits further anti-amoebic, insecticide and anthelmintic activities.

The hypocholesterolaemic effect of *C. fistula* has been investigated using hypercholesterolaemic male albino rats. Hypercholesterolaemia was induced by feeding on a mixture of cholesterol plus cholic acid for a 12-week period. Hypercholesterolaemia was characterized by significant increase in the average levels of total lipids, total cholesterol and triglycerides, and significant decrease in phospholipid content. Administration of *C. fistula* significantly reduced blood and liver total lipids. Brain, spleen, kidneys and heart followed nearly the same trend but with moderate effect. Blood, liver, kidneys, spleen and heart total cholesterol were significantly reduced, while that of the brain was not affected. The level of triglycerides was markedly improved. There was a moderate rise, however, in phospholipid content in all organs studied; that is, there was marked progress in the correction of lipid metabolism. Administration of *C. fistula* also induced a significant decrease in the high activities of serum GOT, GPT, alkaline and acid phosphatase; the values nearly returned to the initial values. Total serum protein, albumin (A), globulin (G), A/G, free amino acids, uric acid and creatinine were also determined. Their values were improved and nearly attained the normal values of the control group.

The aqueous fraction of *C. fistula* produced a significant decrease in glycaemia in mice ( $p < 0.001$ ) at 4 and 24 hours with doses of 300 and 500 mg/kg, and at 1 and 4 hours after the dose of 1000 mg/kg ( $p < 0.001$ ). In the glucose tolerance test, the aqueous fraction of *C. fistula* produced a significant decrease ( $p < 0.05$ ) in glucose tolerance at a dose of 500 mg/kg, but a significant increase ( $p < 0.001$ ) at a dose of 1000 mg/kg.

The ethanol extract of the leaves and bark of *C. grandis* showed in vitro antifungal activity against *Epidermophyton floccosum*, *Microsporum gypseum* and *Trichophyton rubrum* in pure culture at a minimal inhibitory concentration of 50 µg/ml.

**Adulterations and substitutes** Anthraquinone glycosides and sennosides are also found in *Senna* species, which are also used for their laxative and purgative properties.

**Description** Large shrubs or small to medium-sized trees up to 30(–40) m tall; bole up to 60 cm in diameter. Leaves arranged spirally, often distichous, paripinnate, without extrafloral nectaries;

stipules present. Inflorescence terminal on main shoots or short side shoots or axillary, racemose. Flowers having pedicels with 2 bracteoles at or just above the base; hypanthium present but variable; calyx 5-merous, sepals reflexed at anthesis; corolla 5-merous, zygomorphic; androecium zygomorphic, 10-merous, filaments of 3 abaxial stamens sigmoidally curved, generally longer than their anthers, the other 7 filaments straight and short; ovary superior, style variable. Fruit an elongated pod, cylindrical or compressed, indehiscent, many-seeded. Seeds 1- or 2-seriate, funicle filiform. Seedling with epigeal germination; cotyledons emergent, semi-fleshy.

**Growth and development** *C. fistula* is a slow growing, deciduous tree. Generally, it takes 8–10 years from sowing to flowering. This can be reduced by vegetative propagation. In Singapore, *C. fistula* sheds its leaves at 9–10 months' intervals and the inflorescences develop with the new leaves. At the beginning of flowering, the whole crown is covered with flowers; sporadic flowering continues up to 3 months. Experiments have shown that pollen is still viable after 4 weeks of storage.

Seedling development in *C. grandis* is initially rapid, but slows down after 2 months. It is reported evergreen in Java and deciduous in northern Malaysia and Indo-China, where the leaves fall at the beginning of the dry season. The tree flowers before new leaves appear. In Costa Rica, fruit takes 10–12 months to mature.

The roots of *C. fistula* and *C. javanica* lack nodulating ability, but for *C. grandis* this is not clear. The concentration of sennoside in the leaves of *C. fistula* was highest soon after the onset of the rainy season, when new leaves had appeared and flowering started. The sennoside content of the pods was highest at the midstage of fruit maturation, when the pods were pale brown.

**Other botanical information** Until the beginning of the 1980s *Cassia* was considered to be a very large genus of over 500 species, but then it was split into 3 genera: *Cassia* sensu stricto, *Senna* and *Chamaecrista*. *Cassia* now has only about 30 species, whereas *Senna* and *Chamaecrista* comprise about equal numbers of species (about 260 and 270 respectively). *C. fistula* is able to hybridize with *C. javanica*.

**Ecology** *Cassia* is found in forests at low altitudes. *C. fistula* occurs in Java in light forest below 400 m altitude, in the Philippines in open grasslands at low and medium altitudes. It seems to favour calcareous and red, volcanic soils, but in



Thailand it is also found on sandy and loamy soils. *C. javanica* occurs in moist evergreen forest, deciduous monsoon forest and in more open or even savanna-like habitats. The various subspecies of *C. javanica* show preferences for either dry or moist habitats on a wide variety of soils.

**Propagation and planting** *C. fistula*, *C. grandis* and *C. javanica* can be propagated by seed and vegetatively through cuttings and layering. They have a hard seed-coat and germination is improved considerably by mechanical scarification or treatment with concentrated sulphuric acid for at least 45 minutes. Seed can be stored for prolonged periods without loss of viability. *C. fistula* seed should be sown in full light, and adequate water supply is required for optimal germination. The seed can increase three times in weight by absorbing water. Direct sowing is practised in Asia, but preliminary weeding is important. In Costa Rica, *C. grandis* is propagated by means of large cuttings ('apicormic shoots'). Vertical shoots of 15 cm in diameter are cut, trimmed to a length of 2.5 m. These are laid out in the shade for a week and then stacked vertically for three weeks. Then they are planted, with the lower ends buried 50 cm deep. *C. javanica* is usually propagated by seed, with 50% of the seed producing healthy plants.

**Diseases and pests** *Colletotrichum gloeosporioides* causes brown pinhead spot disease in *C. fistula* in Malaysia. In the Philippines, *C. fistula* is attacked by the psyllid *Heteropsylla cubana*. Symptoms include leaf curling, defoliation, stunted shoot growth and death.

**Harvesting** Pods of *Cassia* are harvested when mature, and in general simply collected from the ground.

**Handling after harvest** For domestic use of *C. fistula*, the pulp is scraped from the fresh pods. Pods intended for trade are dried. Prolonged boiling of the pulp leads to loss of the purgative properties.

**Genetic resources and breeding** In view of their wide distribution *C. fistula*, *C. grandis* and *C. javanica* are not endangered or liable to genetic erosion.

**Prospects** *C. fistula*, *C. grandis* and *C. javanica* may be interesting multipurpose trees for South-East Asian farmers, and have a high ornamental value. With regard to the frequent use of *Cassia* as laxative, caution seems to be justified. The hypocholesterolaemic, antifungal and anti-amoebic properties warrant further research.

**Literature** [1] Asseleih, L.M.C., Hernandez, O.H. & Sanchez, J.R., 1990. Seasonal variations

in the content of sennosides in leaves and pods of two *Cassia fistula* populations. *Phytochemistry* 29: 3095-3099. [2] Bruneton, J., 1995. Pharmacognosy, phytochemistry, medicinal plants. Lavoisier Publishing, Paris, France. pp. 349-366. [3] Caceres, A., Lopez, B., Juarez, X., del Aguila, J. & Garcia, S., 1993. Plants used in Guatemala for the treatment of dermatophytic infections. 2. Evaluation of antifungal activity of seven American plants. *Journal of Ethnopharmacology* 40(3): 207-213. [4] El-Saadany, S.S., el-Massry, R.A., Labib, S.M. & Sitohy, M.Z.A.D., 1991. The biochemical role and hypocholesterolaemic potential of the legume *Cassia fistula* in hypercholesterolaemic rats. *Nahrung* 35(8): 807-815. [5] Esposito-Avella, M., Diaz, A., de Gracia, I., de Tello, R. & Gupta, M.P., 1991. Evaluacion de la medicina tradicional: efectos de *Cajanus cajan* L. (Guandu) y de *Cassia fistula* L. (canafistula) en el metabolismo de los carbohidratos en el raton [Evaluation of traditional medicine: effects of *Cajanus cajan* L. and of *Cassia fistula* L. on carbohydrate metabolism in mice]. *Revista Medica de Panama* 16(1): 39-45. [6] Gonzalez, A.G., Bermejo, J. & Valencia, E., 1996. A new C6-C3 compound from *Cassia grandis*. *Planta Medica* 62(2): 176-177. [7] Irwin, H.S. & Barneby, R.C., 1982. The American Cassiinae. A synoptical revision of Leguminosea tribe Cassieae subtribe Cassiinae in the New World. *Memoirs of the New York Botanical Garden* 35(2): 64-635. [8] Larsen, K. & Ding Hou, 1996. *Cassia*. In: Kalkman, C., Kurkop, D.W., Nooteboom, H.P., Stevens, P.F. & de Wilde, W.J.J.O. (Editors): *Flora Malesiana*. Series 1, Vol. 12(2). Rijksherbarium/Hortus Botanicus, Leiden University, the Netherlands. pp. 556-565. [9] Monif, T., Malhotra, A.K. & Kapoor, V.P., 1992. *Cassia fistula* seed galactomannan: potential binding agent for pharmaceutical formulation. *Indian Journal of Pharmaceutical Sciences* 54(6): 234-240. [10] Shukla, S.C. & Das, S.R., 1988. Cure of amoebiasis by seed powder of *Cassia fistula*. *International Journal of Crude Drug Research* 26(3): 141-144.

#### *Selection of species*

#### **Cassia fistula L.**

Sp. pl. 1: 377 (1753).

**Vernacular names** Golden shower, Indian laburnum (En). Caneficier (Fr). Indonesia: trengguli (Javanese), bobondelan (Sundanese), klobop (Madurese). Malaysia: bereksa, tengguli, rajah kayu. Philippines: fistula (Tagalog, Cebu Bisaya),

kana-pistula (Tagalog), bitsula (Cebu Bisaya). Cambodia: reach, reach speu, reach chhpoeus. Laos: khoun (general). Thailand: khuun (central, northern), lom laeng (northern), ratchaphruek (central). Vietnam: c[aa]y b[of] c[aj]p n[luw]lows[c, mu[oof]ng ho[af]ng y[ees]n.

**Distribution** Widespread in the tropics; in Java often cultivated as an ornamental, in the Philippines planted as a medicinal or ornamental plant, cultivated throughout New Guinea.

**Uses** Since ancient times *C. fistula* has been used as a laxative throughout the tropics. In Papua New Guinea, Central Province, broken bones and tropical ulcers are bandaged with bark scrapings and leaf sap. In Thailand, the heartwood is traditionally applied as an anthelmintic. The wood is occasionally used e.g. for posts, carts and agricultural implements. The bark is used for tanning and as an ingredient in betel paste.

**Observations** A small to medium-sized tree, 10–15 m tall or sometimes more, deciduous or se-

mi-deciduous, branches spreading, young twigs glabrous; leaves with 3–7 pairs of leaflets, petiole 5–8 cm long, terete, leaflets ovate-oblong, 7–12 cm × 4–8 cm, subcoriaceous, base broadly cuneate, apex acute, with shiny upper surface, glabrous when mature; inflorescence an axillary, pendulous, lax raceme, 20–40(–60) cm long, many-flowered; flowers fragrant, sepals 7–10 mm long, petals broadly ovate, golden-yellow, stamens 10, 3 long with filaments 3–4 cm long, 4 shorter with filaments 6–10 mm long, 3 reduced with filaments 3–4 mm long and minute anthers; fruit pendent, terete, 20–60 cm long, 1.5–2 cm in diameter, black, glabrous, indehiscent; seeds numerous, separated by papery septa and embedded in black, glutinous pulp. *C. fistula* occurs in open forest and grassland at lower altitudes.

**Selected sources** 91, 97, 133, 181, 190, 284, 319, 332, 336, 357, 402, 409, 580, 653, 692, 726, 817, 846, 973, 1023, 1128, 1178, 1277, 1287, 1343, 1520.

### *Cassia grandis* L.f.

Suppl.: 230 (1781).

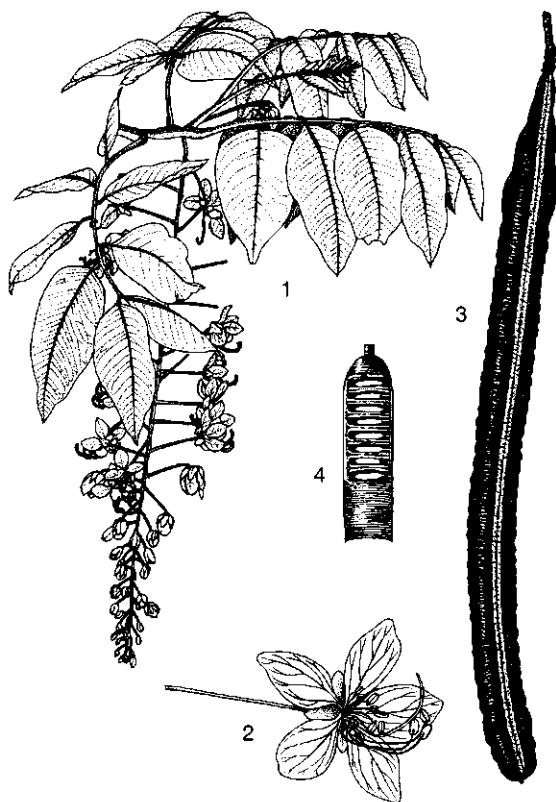
**Synonyms** *Cassia pachycarpa* de Wit (1956).

**Vernacular names** Horse cassia, pink shower (En). Malaysia: kotek, kotek mamak. Cambodia: sac phle, kreete. Laos: brai xiem, may khoum. Thailand: kanpaphruek (Bangkok). Vietnam: b[oof] c[aj]p d[or], [oo] m[oo]i.

**Distribution** Originating from tropical America, but introduced throughout the tropics; abundant in Cambodia and southern Vietnam, common as an ornamental and escape in Malaysia, Java and New Guinea.

**Uses** The fruit pulp is used as a laxative similar to *C. fistula* and reported to be more powerful. A decoction of the leaves is used as a laxative and in the treatment of lumbago. *C. grandis* is also reported to give strong multipurpose wood.

**Observations** A medium-sized tree, up to 20(–30) m tall, semi-deciduous, young branches and inflorescence covered with rusty lanate indumentum; leaves with 10–20 pairs of leaflets, petiole 2–3 cm long, lanate, leaflets sessile, elliptical-oblong, 3–5 cm × 1–2 cm, subcoriaceous, rounded at both ends; inflorescence a lateral raceme, 10–20 cm long, 20–40-flowered; flowers with sepals 5–8 mm long, petals initially red, fading to pink and later orange, the median one red with a yellow patch, stamens 10 with hirsute anthers, 3 long ones with filaments up to 30 mm and anthers 2–3 mm long, 5 short ones with filaments 7–9 mm and anthers 1–1.5 mm long, 2 reduced ones with



*Cassia fistula* L. – 1, branch with inflorescence; 2, flower; 3, mature pod; 4, section of pod showing seeds.

filaments about 2 mm long; fruit pendent, compressed, 20–40(–60) cm long, 3–5 cm in diameter, blackish, glabrous, woody, rugose; seeds 20–40 per pod, surrounded by sweetish pulp. *C. grandis* is a common ornamental in villages at lower altitude.

**Selected sources** 97, 284, 336, 357, 409, 416, 653, 688, 817, 1035, 1493.

### **Cassia javanica L.**

Sp. pl. 1: 379 (1753).

**Synonyms** *Cassia nodosa* Roxb. (1832), *Cassia bartonii* F.M. Bailey (1901), *Cassia agnes* (de Wit) Brenan (1958).

**Vernacular names** Indonesia: bobondelan (Sundanese), boking-boking (Sumatra), trengguli (Javanese). Malaysia: bebusok, busok-busok (Peninsular). Philippines: pink shower (En), antsoan (Bikol). Cambodia: bô prùk'. Laos: khoun loy<sup>2</sup>. Thailand: chaiaphruk (central), kalalphruk (central, northern), lak khoei lak kluea (Trang). Vietnam: b[uf] c[aj]p, mu[oof]ng b[off] c[aj]p.

**Distribution** Widespread in the Malesian area, wild and cultivated. Widely cultivated all over tropical Asia. The various subspecies are geographically confined to parts of the Malesian area, though sometimes cultivated outside their original area of distribution.

**Uses** The ripe pods and seeds are used as a traditional laxative throughout the Malesian area. In Thailand, bark and seeds are also used as antipyretics. However, it was noted that emesis may be observed. The bark is used for tanning leather. *C. javanica*, in particular subsp. *agnes* (de Wit) K. Larsen and subsp. *nodosa* (Roxb.) K. & S.S. Larsen, is also widely planted as an ornamental. The wood is used for general construction, furniture and cabinet making.

**Description** A small to medium-sized tree up to 25(–40) m tall, deciduous or semi-deciduous, trunk of young trees either smooth or armed with stump-remnants of branches; leaves with 5–15(–20) pairs of leaflets, petiole 1.5–4 cm long, leaflets elliptical-ovate to oblong, 2.5–5 cm × 1.5–2.5 cm, base broadly rounded, apex acute or rounded to obtuse; inflorescence a raceme or panicle, terminal on leafy shoots or lateral on short side branches, up to 16 cm long, many-flowered; flowers with sepals 4–10 mm long, green to dark red, petals 15–35 mm long, whitish to reddish or buff, stamens 10, 3 longer ones with filaments 2 cm long, 4 shorter with filaments about 1 cm long and 3 reduced with filaments about 1 cm long and minute anthers; fruit pendent, terete, 20–60 cm long, 1–1.5(–2.5) cm in diameter, indehiscent;

seeds numerous, embedded in a flat disk. *C. javanica* is a very polymorphic species and several subspecies are distinguished. It has a wide ecological amplitude and is generally found at lower elevations.

**Selected sources** 97, 202, 284, 336, 357, 580, 640, 653, 817, 1520, 1564.

Anny Victor Toruan-Purba

### **Catharanthus roseus (L.) G. Don**

Gen. hist. 4(1): 95 (1837).

APOCYNACEAE

2n = 16

**Synonyms** *Vinca rosea* L. (1759), *Lochnera rosea* (L.) Reichenb. ex Endl. (1838).

**Vernacular names** Madagascar periwinkle (En). Indonesia: bunga serdadu, kembang tembaga, tapak dara (general). Malaysia: kemunting china, rumput jalang, tahi ayam (Peninsular). Philippines: chichirica (Sp), kantotai, amnias (Tagalog). Thailand: nom in (Surat Thani), phakpot bok (northern), phaengphuai bok (Bangkok). Vietnam: c[aa]y b[oo]ng d[uwf]a, d[uwf]a c[aj]n, hoa h[ar]i d[awf]ng.

**Origin and geographic distribution** *C. roseus* belongs to a small genus of 8 species, all originating from Madagascar except for *C. pusillus* (Murr.) G. Don, which is restricted to India and Sri Lanka. For centuries, Madagascar periwinkle has been cultivated as an ornamental throughout the tropics and occasionally in the subtropics; it has become naturalized in many regions. It was brought into cultivation in the first half of the 18th Century in Paris, from seeds collected in Madagascar, and was later distributed from European botanical gardens to the tropics as an ornamental.

**Uses** In traditional medicine, a decoction of all parts of the plant is used to treat malaria, diarrhoea, diabetes, cancer and skin diseases. Madagascar periwinkle is well known as an oral hypoglycaemic agent. Extracts prepared from the leaves have been used as an antiseptic agent for the healing of wounds, against haemorrhage and as a mouthwash to treat toothache. Madagascar periwinkle is also considered to be a diaphoretic and diuretic and is used to relieve indigestion, dyspepsia, dysentery, toothache and wasp stings, and as a vomitive, purgative, vermifuge, depurative and haemostatic.

The aerial parts of the plant are used for alkaloid extraction (vincristine, vinblastine, vindesine, vi-

norelbine). The alkaloids are prescribed in anti-cancer chemotherapy, usually as part of complex chemotherapy protocols. They are administered intravenously, via injection or infusion. Vincristine (sulphate) is indicated in the treatment of acute leukaemia, Hodgkin's disease, non-Hodgkin's lymphoma, small-cell bronchial cancer, neuro and nephroblastomas, metastasized breast cancer and various sarcomas (especially rhabdomyosarcoma). A normal dose for adults is 1–1.4 mg/m<sup>2</sup> of body surface area, usually at a frequency of once a week, or once monthly in combination chemotherapy. The indications for vinblastine (sulphate) are Hodgkin's disease, non-Hodgkin's lymphoma, advanced testicular cancer, Kaposi's sarcoma, and sometimes choriocarcinomas and some cases of histiocytosis (especially the Lettere-Siwe syndrome). A normal dose for adults is 4–6 mg/m<sup>2</sup> of body surface area weekly. Vindesine, a semisynthetic derivative of vinblastine, is indicated in the treatment of acute lymphatic leukaemia (especially in children) and refractory lymphomas and melanomas. When used alone, a normal dose of vindesine is 3 mg/m<sup>2</sup> of body surface area every 7–10 days for one month, and then at intervals of 15 days. Combination chemotherapy protocols often allow lower doses. The semisynthesized vinorelbine has breast cancer and bronchial cancer as current indications, and is often administered to adults in a dose of 25–30 mg/m<sup>2</sup> of body surface area in monotherapy, and in lower doses in combination chemotherapy protocols. The major toxic side effect of vinblastine is myelosuppression, especially leukopenia. Neurotoxicity is the dose-limiting toxicity of vincristine. Both neurotoxicity and myelosuppression are observed after administration of vindesine, but they are less severe than with vincristine and vinblastine. All four agents may induce thrombocytopaenia and alopecia.

The dried root is an industrial source of ajmalicine (raubasine), which increases the blood flow in the brain and peripheral parts of the body. Preparations of it are used to treat the psychological and behavioural problems of senility, sensory problems (dizziness, tinnitus), cerebrovascular accidents, cranial traumas and their neurological sequelae.

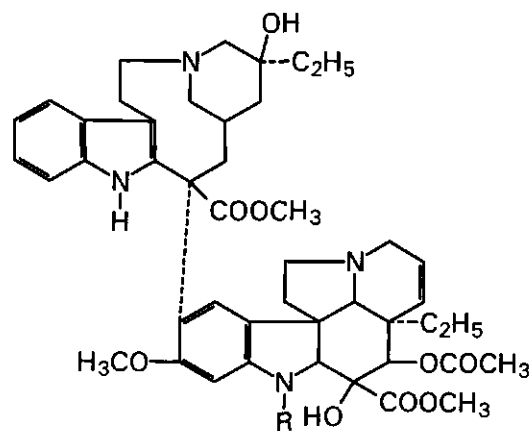
Madagascar periwinkle is a popular garden ornamental, grown as a perennial in tropical regions and as an annual in temperate regions. It is valued for its bushy habit and large flowers carried above dark green foliage.

**Production and international trade** The dimeric alkaloids extracted from the aerial parts

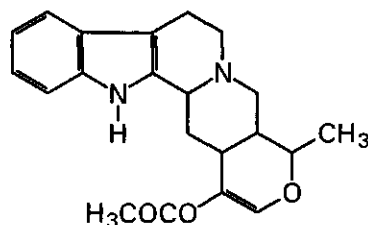
of Madagascar periwinkle are marketed as a lyophilisate or a solution of a salt designed for the sole intravenous route (direct intravenous or through infusion tubing). Vindesine and vinorelbine, which are semisynthesized derivatives of vinblastine, are marketed as a sulphate and a bitartrate, respectively, for injectable solutions. All these drugs are prescription drugs that pharmacists in western countries cannot issue without the direct authorization of a physician.

The price of vincristine was reported to be over US\$ 200 000/kg in 1993; the world market consumes 250–300 kg annually with a value of US\$ 50–80 million.

**Properties** Madagascar periwinkle has been found to contain as many as about 100 constituents with an indole or dihydroindole structure. The principal constituent is vindoline (up to 0.5%); other major compounds are serpentine, catharanthine, ajmalicine (raubasine), akuammine, lochnerine and tetrahydroalstonine. Ajmalicine and serpentine are essentially present in the roots, whereas catharanthine and vindoline accumulate in aerial parts. The aerial parts contain 0.2–1% alkaloids.



vinblastine (R = CH<sub>3</sub>) and vincristine (R = CHO)



ajmalicine

The substances of pharmacological interest are the dimeric alkaloids which show a coupling of an indole and a dihydroindole. Of the separate parts, the indole/dihydroindole moiety is derived from the amino acid tryptophan, which is coupled to a monoterpene residue. Several of these dimeric alkaloids have cytostatic properties, but they occur in very small amounts: vincristine (= leurocristine) in up to 3 g/t of dried drug, and vinblastine (= vincalcucoblastine) in a slightly larger amount. Other active compounds are leurosidine (= 20'-epivinblastine) and leurosine (= 15',20'-epoxyvinblastine).

Vincristine and vinblastine are antimetotics. They bind to tubulin and prevent the formation of microtubules that assist in the formation of the mitotic spindle; in this way, they block mitosis in the metaphase. These compounds have a non-trivial toxicity; they both have neurotoxic activity (especially vincristine) because the microtubule assembly also plays a role in neurotransmission. Their peripheral neurotoxic effects are neuralgia, myalgia, paresthesia, loss of the tendon reflexes, depression and headache, and their central neurotoxic effects are convulsive episodes and respiratory difficulties. Other side-effects are multiple and include alopecia, gastro-intestinal distress including constipation, buccal ulcerations, amenorrhoea and azoospermia. As vinblastine is highly leucopenic, its dosage must be carefully controlled. The alkaloids are very irritating; if extravasation accidentally occurs there is a risk of tissue necrosis. It is possible to limit the side-effects by carefully guiding the dose and administration, and intensively monitoring the treatment. In common with all teratogenic chemotherapeutics, pregnancy and breast-feeding are strictly contra-indicated. Semisynthetic derivatives whose structure is closely related to that of the naturally occurring dimeric alkaloids are also used as anti-cancer drugs. Vindesine can be prepared from vinblastine, and it is also a potent antimetotic. Its side-effects include a transient granulocytopenia and effects comparable to those caused by vincristine and vinblastine, although the neurological symptoms are less obvious. Vinorelbine (= noranhydrovinblastine) is obtained from anhydrovinblastine. It acts preferentially on mitotic microtubules and not so much on neuronal microtubules, and consequently its neurological toxicity is limited. However, its haematotoxic activity is substantial, so its dosage must be carefully controlled.

Some of the alkaloids (e.g. leurosine and vindoline) exhibit a moderate hypoglycaemic action.

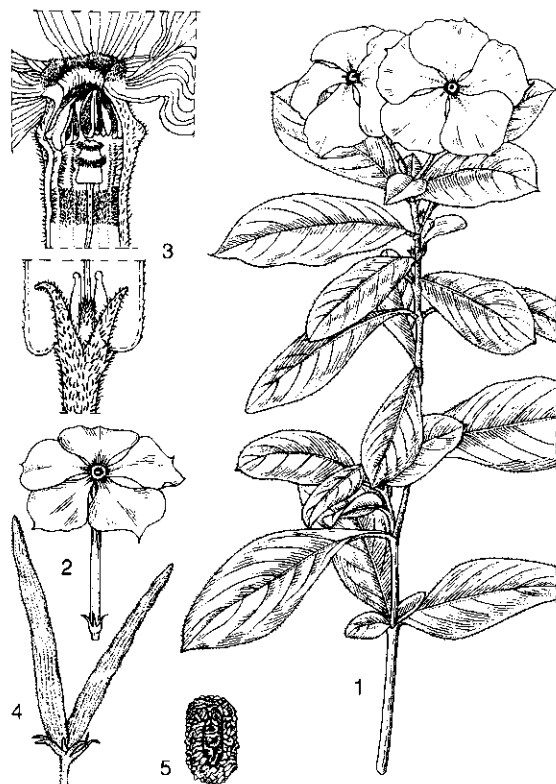
However, most experiments to confirm the reputed positive effect on diabetes have had disappointing results. Vinblastine markedly inhibited in vitro the reproduction of *Trypanosoma cruzi*, the organism responsible for Chagas' disease, which is a major health problem in Central and South America.

Roots to be used in pharmacy must contain at least 0.4% ajmalicine and serpentine (the quaternary base corresponding to ajmalicine; ajmalicine can be derived from it). These compounds can easily be characterized and quantified by two thin-layer chromatography analyses. Ajmalicine is an  $\alpha$ -adrenergic blocking spasmolytic, which at high doses reverses the effects of adrenaline and moderates the activity of the vasomotor centres, especially in the brain stem. It temporarily increases the blood flow to the brain.

Antiviral activity has been reported in vitro for some *Catharanthus* alkaloids. Extracts showed fungicidal activity (e.g. against *Fusarium solani* that causes wilt in aubergine and *Sclerotium rolfsii* that causes damping-off disease in tomato) and nematocidal activity (e.g. against *Meloidogyne incognita* and *M. javanica*).

**Adulterations and substitutes** Ajmalicine and derivatives are also found in other *Apocynaceae*, such as *Rauwolfia* spp.

**Description** An erect or decumbent, deciduous undershrub up to 100(-200) cm tall, usually with white latex; roots up to 70 cm long; stems often woody at base. Leaves decussate, simple, elliptical to obovate or narrowly obovate, (3-)4-9 cm  $\times$  (1-)1.5-3.5 cm, herbaceous to thinly leathery, cuneate and sometimes oblique at base, obtuse or acute at apex with a mucronate tip, entire, glossy green above and pale green below, laxly pubescent to glabrous on both sides, secondary veins 7-11 on both sides of midrib and more or less conspicuous, tertiary venation inconspicuous; petiole (0.1-)0.3-1 cm long, with a fringe of colleters in the axil; true stipules absent. Inflorescence terminal, but apparently lateral because of alternating development of one of the axillary buds of the apical leaf-pair, 1-2-flowered. Flowers actinomorphic, bisexual, 5-merous, subsessile; sepals slightly connate at base, (2-)3-5 mm  $\times$  1-1.5 mm, green; corolla salver-shaped, pink, rose-purple or white with a purple, red, pink, pale yellow or white centre, tube 2-3 cm long and widening near the top, laxly puberulous to glabrescent outside, with a densely strigose ring of hairs in the throat and with a sericeous ring of hairs lower down the tube, lobes broadly obovate, 1-2(-3) cm long, mucronate



*Catharanthus roseus* (L.) G. Don - 1, flowering twig; 2, flower; 3, base and top of corolla tube in longitudinal section; 4, fruit; 5, seed.

at apex, glabrous, spreading, in bud overlapping to the left; stamens included in the corolla tube, inserted just below the corolla throat, filaments very short, anthers free, introrse; ovary superior, consisting of 2 very narrowly oblong carpels coherent at base, style filiform, with a cylindrical pistil head provided at base with a reflexed hyaline frill ('petticoat') and with rings of woolly hairs at base and apex, stigma glabrous; disk composed of 2 glands, often longer than ovary. Fruit composed of 2 cylindrical and acute follicles 1-4 cm long, striate, laxly puberulous to glabrous, green, dehiscent at adaxial side, many-seeded. Seeds oblong, 1-2 mm long, with rugose testa and lateral hilum, black; cotyledons flat, slightly shorter than radicle; endosperm scanty.

**Growth and development** Madagascar periwinkle is self-incompatible. The seeds usually fall close to the mother plant, but are sometimes transported by ants.

**Other botanical information** *Catharanthus* is very closely related to *Vinca* but differs in gen-

eral appearance (*Vinca* produces trailing or floppy, long-lived stems with persistent, leathery leaves) and in flower characteristics (flowers in *Vinca* solitary, corolla infundibular, stamens with long filaments, glands at base of ovary small, pistil head without 'petticoat').

The flower types present in Madagascar periwinkle differ in whether the corolla is pink, rose-purple, white, or white but red-eyed. Usually, no qualitative differences have been found in alkaloid composition between these flower types, but tests in the Philippines showed that plants with rose-purple flowers had a higher alkaloid production than white-flowered plants. Likewise, tests in Thailand revealed the white red-eyed plants to produce the highest yields of alkaloids and vinblastine. Nowadays, these types are usually classified as cultivars, e.g. cv. *Albus*.

*C. lanceus* (Bojer ex A.DC.) Pichon is used medicinally in Madagascar and South Africa; the leaves are used as astringent and emetic, the aerial parts as galactagogue and vomitive, and the roots as purgative. The alkaloid fractions of *C. lanceus* have shown hypotensive activity (mainly caused by yohimbine, a potent  $\alpha$ -adrenergic blocker). A lyophilized aqueous extract was found to show anti-tumour activity (leurosine is most potent), whereas some alkaloids have a hypoglycaemic effect.

**Ecology** Madagascar periwinkle often occurs in sandy locations along the coast, but also inland on river banks, in savanna vegetation and in dry waste places and roadsides, sometimes in open forest or scrub, usually on sandy soils, but sometimes also on rocky soils. It is highly salt-tolerant, and is mostly found near sea-level, but occasionally up to 1500 m altitude. It can stand drought well, but not severe heat. Under severe water stress the alkaloid content of mature leaves was found to double, but it did not change in stems and immature leaves and it decreased in roots.

**Propagation and planting** Madagascar periwinkle is usually propagated by seed. Seed may remain dormant for several weeks after maturity. The optimum temperature for germination is about 25°C. The germination rate of fresh seed is reported as 40%, but pretreatment with chemical stimulants like a potassium nitrate solution may enhance it to 90%. Madagascar periwinkle can also be propagated vegetatively by greenwood or semi-ripe cuttings rooted in a closed container with bottom heat.

**In vitro production of active compounds** Callus tissue of Madagascar periwinkle can be

cultured on supplemented White's medium or supplemented Heller's medium, usually solidified with agar. Tests in Thailand showed that the best callus initiation and greatest weight of callus from stem explants was obtained in a Murashige and Skoog basal medium supplemented with 0.5 mg/l 2,4-D and 1 mg/l kinetin. In the Philippines, successful trials have been carried out using floral explants.

Cell suspension cultures are grown in Gamborg's B5 medium or in LS medium containing 2 mg/l naphthalene acetic acid, 0.2 mg/l kinetin and 30 g/l sucrose, under constant light at 25°C and constant shaking. Cell cultures of Madagascar periwinkle produce a variety of monoterpenoid alkaloids. The alkaloid spectra of root and shoot cultures are similar to those of roots and aerial parts, respectively, of whole plants. Ajmalicine, serpentine and catharanthine are usually the major constituents. Much higher yields of serpentine and ajmalicine, the hypotensive agents, can be produced in cell cultures than in whole plants: up to 2% on dry weight basis versus 0.3% in whole plants. The dimeric anti-cancer alkaloids vinblastine and vincristine are almost undetectable in cultured cells, so attention has turned to the production of catharanthine and vindoline, which can be used as precursors for their synthesis. However, in experiments the production of vindoline appeared not to be stable in cell culture. The lack of vindoline accumulation in cell suspension cultures has been correlated with the lack of expression of the enzymes which catalyse the last steps of vindoline biosynthesis. These enzymes are considered to express only at later development stages and occur in above-ground plant parts; the last steps in biosynthesis only occur in seedlings grown in light.

Multiple shoot cultures induced from seedlings produce vindoline and catharanthine in rather higher levels. Murashige and Skoog medium supplemented with 7 mg/l benzyladenine and 1 mg/l  $\alpha$ -naphthalene acetic acid strongly stimulates the formation of shoots, whereas medium supplemented with 2,4-dichlorophenoxyacetic acid suppresses the formation of shoots.

Another possible method of vindoline production is by cultures of selected hairy roots. These can be established by infecting seedlings with *Agrobacterium rhizogenes*. Some clones not only showed levels of ajmalicine, serpentine and catharanthine comparable to those of cell suspension cultures, but also about 3 times more vindoline than usually found in cell cultures.

**Husbandry** In South-East Asia, Madagascar

periwinkle is usually cultivated as an ornamental; there are no records of large-scale cultivation for medicinal purposes. It responds well to N fertilizers, but can also grow and persist on poor soils.

In India, Madagascar periwinkle is largely cultivated as a 200-day crop for its leaves (for the extraction of vinblastine and vincristine) and its roots (for the extraction of ajmalicine). The crop needs little irrigation and fertilizer.

**Diseases and pests** In Malaysia, Madagascar periwinkle has been reported to be infected with so-called Malaysian periwinkle yellow. Symptoms include excessive yellowing of foliage, virescence, phyllody, bunchy top and stunted flowers and leaves, suggesting infection by a mycoplasma-like organism. Similar diseases have been reported from China, Taiwan, North America and Europe. Mycoplasma-like organisms can be transferred to Madagascar periwinkle by parasitic plants of the genus *Cuscuta*, and perhaps also by leafhoppers. Container-grown plants in the United States cultivated as ornamentals have been reported susceptible to *Phytophthora parasitica* that causes root and stem rot.

**Yield** The alkaloids in Madagascar periwinkle used in cancer chemotherapy occur in very small amounts: vincristine in an amount of up to 3 g/t of dried drug, and vinblastine in a slightly larger amount. These dimeric alkaloids are almost undetectable in cultured cells. The serpentine and ajmalicine production can be much higher in cell cultures than in whole plants: 1.3% on a dry weight basis versus 0.3% in whole plants.

**Handling after harvest** The aerial parts of Madagascar periwinkle are separated from the roots; both are dried at low temperatures, then packed for shipment. Potted plants for use as ornamentals are usually traded in sealed packages. They are marketable in this condition for 18 days, and do not require watering during this period.

**Genetic resources and breeding** Although Madagascar periwinkle probably originated from a limited area in south-eastern Madagascar, it is now widely planted and naturalized in all tropical areas, and is certainly not endangered. However, protection of the wild populations in Madagascar is desirable to ensure the conservation of the genetic diversity, which might be of interest for breeding purposes in the future.

Tetraploid plants, induced with colchicine, have been found to have a much higher alkaloid content than diploid plants, but the doubling of chromosomes was found to result in reduced pollen fertility and poor seed set.

*C. roseus* has been successfully crossed with *C. trichophyllus* (Baker) Pichon, with the F<sub>1</sub> having a high seed set and good viability when *C. trichophyllus* was the female parent. The alkaloid profiles of the two species are different, and alkaloid production seems to be higher in hybrids than in the parent species. A possible strategy for improving alkaloid production in *Catharanthus* could be to breed for hybrids with a high alkaloid content.

**Prospects** The possibility of accessing active dimeric alkaloids by biomimetic synthesis has recently attracted much attention. It is now conceivable that vinblastine could be obtained from starting materials such as catharanthine and vindoline that are neither rare nor too expensive. These latter two compounds can be produced in sufficient amounts in *in vitro* cultures of Madagascar periwinkle. Studies on analogues of the well-known alkaloids suggest good prospects for new developments vis-à-vis *Catharanthus* alkaloids.

Madagascar periwinkle may have some prospects as a protectant of stored grain, since tests in the Philippines have demonstrated that its use results in improved germination of treated maize kernels and in vigorous seedlings. Corn weevil and flour beetle infestation were also reduced. However, possible toxicity and teratogenicity of residues could be a serious drawback.

**Literature** |1| Bhadra, R., Vani, S. & Shanks, J.V., 1993. Production of indole alkaloids by selected hairy root lines of *Catharanthus roseus*. *Biotechnology and Bioengineering* 41(5): 581-592. |2| Bruneton, J., 1995. *Pharmacognosy, phytochemistry, medicinal plants*. Lavoisier Publishing, Paris, France. pp. 832-838. |3| Cavin, J.C., Krassner, S.M. & Rodriguez, E., 1987. Plant-derived alkaloids active against *Trypanosoma cruzi*. *Journal of Ethnopharmacology* 19(1): 89-94. |4| Marfori, E.C. & Alejar, A.A., 1993. Alkaloid yield variation in callus cultures derived from different plant parts of the white and rosy-purple periwinkle, *Catharanthus roseus* (L.) G. Don. *Philippine Journal of Biotechnology* 4(1): 1-8. |5| Plaizier, A.C., 1981. A revision of *Catharanthus roseus* (L.) G. Don (Apocynaceae). *Mededelingen Landbouwhogeschool Wageningen, Nederland* 81-9. 12 pp. |6| Schütte, H.R., 1991. III. Secondary plant substances: monoterpenoid indole alkaloids. *Progress in Botany* 52: 84-96. |7| Sevestre-Rigouzzo, M., Nef-Campa, C., Ghesquière, A. & Chrestin, H., 1993. Genetic diversity and alkaloid production in *Catharanthus roseus*, *C. trichophyllus* and their hybrids. *Euphytica* 66: 151-159. |8| Taylor, W.I. &

Farnsworth, N.R. (Editors), 1975. *The Catharanthus alkaloids: botany, chemistry, pharmacology and clinical use*. Marcel Dekker Inc., New York, United States. |9| Wibowo, A.R., 1991. Efek hipoglikemik akar tapak doro (*Catharanthus roseus* (L.) Don) [Hypoglycaemic effect of roots of white flowered tapak doro (*Catharanthus roseus* (L.) Don)]. S1 Thesis. Faculty of Pharmacy, Gadjah Mada University, Yogyakarta, Indonesia. 83 pp. |10| Yuan, Y.-J., Hu, T.-T. & Yang, Y.-M., 1994. Effects of auxins and cytokinins on formation of *Catharanthus roseus* G. Don multiple shoots. *Plant Cell, Tissue and Organ Culture* 37(2): 193-196.

**Other selected sources** 202, 299, 332, 347, 350, 1035, 1178, 1381, 1498, 1595.

H. Sutarno & Rudjiman

### *Centella asiatica* (L.) Urb.

Mart., *Fl. bras.* 11, 1: 287, pl. 78, fig. 1 (1879).

UMBELLIFERAE

2n = 18

**Synonyms** *Hydrocotyle asiatica* L. (1753).

**Vernacular names** Asiatic pennywort, Indian pennywort, gotu-cola (En). *Hydrocotyle asiatica* (Fr). Brunei: pegaga. Indonesia: daun kaki kuda, pegagan (general), antanan gede (Sundanese). Malaysia: pegaga (general). Philippines: takip-kohol, tappingan-daga (Tagalog), hahang-halo (Bisaya). Singapore: pegaga. Burma (Myanmar): min-kuabin. Cambodia: trachiek kranh. Laos: phak nok. Thailand: bua bok (central), pa-na-e khaa-doh (Karen, Mae Hong Son), phak waen (peninsular). Vietnam: rau m[as], t[is]ch tuy[ees]t[th[ar]o].

**Origin and geographic distribution** *Centella* comprises approximately 40 species with an amazing diversity in South Africa, where all species are confined except *C. asiatica* which has a pantropical distribution including South-East Asia and extending into some subtropical regions.

**Uses** Asiatic pennywort has been used in southern Asia, India and China since prehistoric times for a wide range of complaints. In China, it has been known for many centuries as a medicinal plant with tonic and cooling properties, and (together with *Hydrocotyle* spp.) is known in pharmacology as 'Folia Hydrocotyles'.

The most important use of the whole plant is in skin-related diseases. Fresh leaves, fresh juice, a decoction or an extract are applied, depending on the complaints. Several over-the-counter prepara-



tions recommended for skin care are available, containing various extracts of the plant or one of its constituents asiaticoside. Extracts are applied topically in the adjunct treatment of surgical wounds and minor burns. The extract is used effectively in the treatment of keloids, leg ulcers, phlebitis, slow-healing wounds, scleroderma, lupus, leprosy, surgical lesions, striae distensae, cellulitis and aphthae. Purified extracts are known to accelerate cicatrizing and skin grafting.

Orally, the extract is indicated to relieve the symptoms of venous and lymphatic vessel insufficiency, and used to treat atonic wounds and for hypertrophic healing. The direct application of Asiatic pennywort as a drug is only rarely prescribed in Western medicine. More frequently, standardized extracts are utilized; these are applied orally, intramuscularly or subcutaneously. Asiatic pennywort has a considerable reputation for treating epilepsy. In Ayurvedic medicine in India, it is one of the ingredients of a non-alcoholic anti-epileptic syrup which showed significant anti-epileptic activity in tests with rats. It is also used in India and Thailand as a tonic and to treat dysentery. In Sri Lanka, an extract is used in traditional medicine as a galactagogue. Dry leaves are one of the ingredients of pills taken in Vietnam against senility, and are also used successfully in a complex preparation against acute infective hepatitis.

Asiatic pennywort is a relished vegetable in many South-East Asian countries (with the exception of the Philippines) and also in Sri Lanka. The slightly bitter leaves are eaten raw or cooked. In Thailand, Vietnam, Cambodia and Laos a softdrink is made from the juice of the leaves; the juice is diluted with water and some sugar is added. The popularity of Asiatic pennywort as a vegetable and soft drink is certainly related to its medicinal properties.

**Production and international trade** The reported quantities of dried plants used by traditional drug manufacturers in Indonesia in the period 1991 to 1994 vary between 19–125 t/year. Most of Asiatic pennywort for commercial medicinal preparations comes from Madagascar. From 1979 to 1988, 26–96 t/year was exported from this country to all parts of the world for pharmaceutical use.

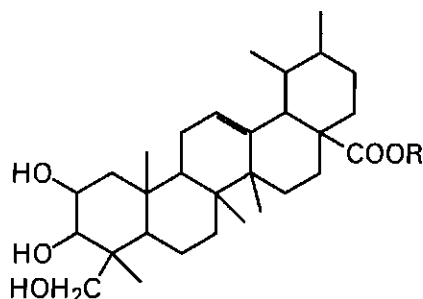
Fresh Asiatic pennywort is a common product on vegetable markets in South-East Asia. Usually plants are gathered from the wild, but it is cultivated commercially in Sri Lanka. No statistics are available. In Thailand a locally produced soft

drink made from Asiatic pennywort is sold canned in supermarkets.

**Properties** Several triterpenoid compounds have been isolated from Asiatic pennywort, the most important ones being asiaticoside, madecassoside, asiatic acid and madecassic acid. These are considered the pharmacologically active principles. In ethanolic extracts compounds such as  $\beta$ -sitosterol and stigmasterol have been demonstrated.

Several animal tests have confirmed that extracts have a wound-healing effect, and this activity has also been reported from clinical studies with humans. Patients with burns, cellulitis, leprotic infections and skin ulcers have been treated successfully in controlled studies. Asiatic acid, madecassic acid and asiaticoside have been tested separately and in combination on skin human fibroblast collagen I synthesis *in vitro*. The mixture as well as each individual component stimulated collagen I synthesis to a similar extent. Collagen I is involved in wound healing. A mixture of brahmoside and brahminoside exhibits antispasmodic, antipyretic, central nervous system-depressant and hypotensive activity.

Asiatic pennywort has shown promising narcotic analgesic activity mediated through opioidergic receptors. The ethanolic extract exhibited anti-stress activity and activity against stress-induced gastric ulcer formation in rats, comparable to that of diazepam. In a clinical test in Italy, patients with chronic venous insufficiency of various etiology were treated with Asiatic pennywort at a dose of 60 mg/day during 4 months. For most patients the drug was effective against the subjective symptoms, but did not show significant changes in conjunctival capillaroscopy. In a multicentre, double-blind placebo-controlled study in France with patients suffering from venous insufficiency of the lower limbs, a significant difference was found in favour of a titrated extract of Asiatic pennywort



asiaticoside (R = glucose-glucose-rhamnose)

for the symptoms of heaviness in the lower limbs and oedema. The venous distensibility was improved by the extract. The triterpene fraction of Asiatic pennywort has an effect on the metabolism in the connective tissue of the vascular wall and on the microcirculation. Treatment with the triterpene fraction for 3 weeks caused a significant reduction in the number of circulating endothelial cells in patients with post-phlebotic syndrome.

An aqueous extract of Asiatic pennywort showed activity against the herpes simplex II virus. The recovery of guinea-pigs with experimentally induced tuberculosis was hastened by treatment with asiaticoside. This compound promotes healing through bacteriostatic activity and stimulation of the reticuloendothelium.

The anti-tumour effect of the crude extract of Asiatic pennywort and of partially purified fractions has been studied by *in vitro* short and long term chemosensitivity and by *in vivo* tumour model test systems. The proliferation of transformed cell lines was inhibited more by the partially purified fractions than by the crude extract. The 50% effective doses on 3-hour exposure to the fractions were 17 µg/ml for Ehrlich ascites tumour cells and 22 µg/ml for Dalton's lymphoma ascites tumour cells. Hardly any toxic effects were detected in normal human lymphocytes. At a concentration of 8 µg/ml the partially purified fractions also significantly suppressed the multiplication of mouse lung fibroblast cells. Oral administration of the extracts retarded the development of solid and ascites tumours and increased the lifespan of these tumour-bearing mice.

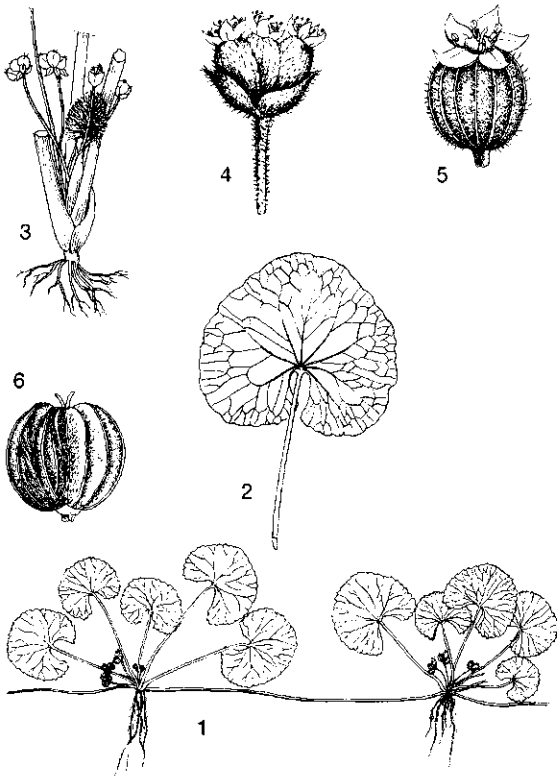
The sensitizing capacity of the raw extract and its triterpenic constituents asiaticoside, asiatic acid, madecassoside and madecassic acid has been studied in guinea-pigs. All were found to be very weak sensitizers; although applied frequently to damaged skin, the risk of acquiring contact sensitivity is low. The toxic dose of asiaticoside by intramuscular application to mice and rabbits is reported to be 40–50 mg/kg body weight. In oral applications, 1 g of asiaticoside per kg body weight has not proved toxic in rats. Asiatic pennywort is toxic in large overdose or as a result of long-term application. It can produce narcotic effects, headache, vertigo, and, occasionally in sensitive individuals, it may even lead to coma. Contra-indications include patients with cardiovascular disorders and internal bleeding. Besides the above activities, anti-amoebic and immunostimulant activity are also reported. Asiaticoside is related to

chemical compounds with known oncogenic activity and it has been found to exhibit slight tumour-producing properties in experimental animals, but possible carcinogenic effects have not yet been investigated thoroughly. Reduction of fertility in female mice has also been reported for asiaticoside. In the essential oil isolated by steam distillation from the aerial parts of Malaysian plants 41 compounds have been identified, with the sesquiterpenoids (80%) as the major category and β-caryophyllene (27%), α-humulene (34%) and germacrene-D (10%) as the most abundant. The major components in the essential oil from plants collected in Sri Lanka were α-copaene (14%), β-caryophyllene (12%), trans-β-farnesene (5%) and α-humulene (9%).

Per 100 g edible portion fresh leaves contain: water 88 g, protein 2 g, fat 0.2 g, carbohydrates 7 g, fibre 1.6 g, Ca 170 mg, P 32 mg, Fe 6 mg, provitamin A 4.5 mg and vitamin C 49 mg.

**Adulterations and substitutes** Asiatic acid and asiaticoside have also been demonstrated in the bark of *Schefflera heptaphylla* (L.) Frodin (syn. *S. octophylla* (Lour.) Harms), which is used in Vietnamese traditional medicine as a tonic and to treat rheumatism. Asiatic acid has also been found in ether extracts of the wood of *Terminalia brassii* Exell and *T. complanata* K. Schumann.

**Description** A small perennial herb, creeping with long stolons (up to 2.5 m long), rooting at the nodes; young parts more or less puberulous. Leaves in rosettes, simple, lamina orbicular-reniform, 1–7 cm in diameter, regularly crenate or crenate-dentate, palmately veined, subglabrous; petiole 1–40(–50) cm long, glabrous to puberulous, broadened at the base into a leaf-sheath; stipules absent. Inflorescence an axillary simple umbel, (1–)3(–7)-flowered with middle flower sessile and lateral flowers with a short pedicel and with involucre of 2 ovate bracts, 0.5–5 cm long peduncled, 1–5 together; scale-like leaves at base of peduncles about 3 mm long. Flowers bisexual, 5-merous; calyx obsolete; petals roundish to broadly obovate, 1–1.5 mm long, entire, greenish, pinkish or reddish; stamens alternate with the petals; disk 2-lobed, plane with elevated margin; ovary inferior, 2-celled, styles 2. Fruit consisting of 2 one-seeded mericarps connected by a narrow junction, separating when mature, oblate-rounded, strongly laterally compressed, 3 mm × 3–4 mm, mericarps distinctly 7–9-ribbed, ribs connected by veins, pubescent when young but often glabrescent. Seed laterally compressed. Seedling with epigeal germination; hypocotyl 2–4 mm long, glabrous; cotyledons



*Centella asiatica* (L.) Urb. - 1, plant habit; 2, leaf; 3, stem base with young leaf, flowers and fruits; 4, inflorescence; 5, flower; 6, fruit.

broadly ovate to elliptical, shallowly emarginate at apex, glabrous; epicotyl absent.

**Growth and development** Asiatic pennywort grows and flowers year round. In the Philippines, it is reported to be one of the main pollen sources for honeybees.

**Other botanical information** *C. asiatica* seems related to *Hydrocotyle* species, but there is morphological, anatomical, palynological and phytochemical evidence for retaining it in *Centella*. Morphologically, *Hydrocotyle* differs particularly in its 3-ribbed mericarps, its free stipules at the base of the petiole, and its peltate or reniform leaves, in the latter case with lobed margins.

In Sri Lanka, where Asiatic pennywort is cultivated as a vegetable, two cultivars are distinguished: a small creeping form, and an erect bushy form with large leaves and petioles. The latter cultivar is most popular.

**Ecology** Asiatic pennywort occurs in sunny or slightly shaded, damp localities on fertile soils (preferring sandy loams with much organic mat-

ter), e.g. along stream banks, on or near paths, alongside walls and in damp, open grassland, from sea-level up to 2500 m altitude. It is an early colonizer of fallowed land in shifting cultivation systems, but may occur also on recently disturbed habitats and even on undisturbed sites. It may carpet the ground completely, but in regions with a monsoon climate usually only during the rainy season.

**Propagation and planting** Asiatic pennywort can be easily propagated vegetatively by runners which root on the nodes, although reproduction by seed is possible. It often regenerates from fragments of stems buried in the soil during hoeing. Stem pieces with one node are planted directly in the field or first in a nursery. Shading is not necessary, but sufficient soil moisture is essential. In Sri Lanka, planting distance is 30 cm × 25 cm for the bush type cultivar and 15 cm × 15 cm for the creeping type cultivar. A planting distance of 50 cm × 60 cm has been reported from Indonesia and Malaysia. At planting, organic fertilizer is added at a rate of 1.5 kg/m<sup>2</sup>. Runners can be planted directly in the field ploughed to 20 cm deep.

**In vitro production of active compounds** Cell suspension cultures of Asiatic pennywort can be grown on a modified Murashige and Skoog medium supplemented with growth regulators, 30 g/l sucrose and 500 g/l casein hydrolysate. They can be subcultured at intervals of 14 days in flasks on a shaker at 24°C and a 16-hour photoperiod.

Cell suspension cultures are used to convert the cytotoxic compound thiocolchicine, a hemisynthetic substrate obtained from natural colchicine (from seeds of *Colchicum autumnale* L.), to thiocolchicoside (3-O-glucosylthiocolchicine), a drug used as a myorelaxant and analgesic. The thiocolchicine can be administered to 7-day-old suspension cultures, and the glucosides are localized intracellularly. Thiocolchicoside accounts for 85% of the glucosides. The cultures have also shown the ability to oxidize papaverine to papaveraldine. There is no known in vitro production of the active compounds of Asiatic pennywort.

**Husbandry** Cultivated Asiatic pennywort does not need much care. A planting can be maintained for 2-3 years if only leaves are harvested. Every 6 months it needs to be fertilized with 1.5 kg/m<sup>2</sup> of organic material. Sometimes small amounts of urea are given, to stimulate leaf growth.

**Diseases and pests** Asiatic pennywort is relatively little affected by diseases and pests. Bacterial wilt caused by *Pseudomonas solanacearum* has

been reported from Sri Lanka, and a leaf spot disease caused by *Cochliobolus geniculatus* has been reported from India.

**Harvesting** If circumstances are favourable, the first harvest can be obtained 2–3 months after planting. Subsequent harvests are possible every 2 months if only leaves are to be harvested. However, creeping types are usually harvested as whole plants. For medicinal use, whole plants are generally harvested at any suitable time of the year.

**Yield** Bushy types may reach 8 t/ha of fresh leaves for the first harvest, and 14 t/ha for each subsequent harvest. Yields of the creeping types are lower.

**Handling after harvest** After harvesting, plants to be used for medicinal purposes are stripped of their roots, cleaned with water and air-dried. The dried material should be kept in tightly closed containers and stored under dry conditions. Fresh leaves harvested as a vegetable are tied together in small bundles and marketed soon, as they wilt rapidly.

**Genetic resources and breeding** Considerable genetic variation may occur between natural populations, due to the wide distribution of Asiatic pennywort. However, there are no known germplasm collections.

In a study of the presence of asiaticoside in various Indian ecotypes it appeared that the asiaticoside content differed significantly among them, with the highest amount (about 0.11%) in ecotypes from subtemperate Himalaya. Selection of genotypes with high asiaticoside content should be considered for commercial exploitation for medicinal purposes. No breeding work is being done.

**Prospects** The medicinal value of Asiatic pennywort has been acknowledged in both traditional and modern medicine. Recent studies have confirmed the efficiency of the plant and its extracts in the treatment of skin injuries and diseases. Moreover, asiaticoside is reported to have considerable medicinal value against leprosy. Therefore, it is remarkable that hardly any commercial plantings exist. As a vegetable, however, cultivation on a larger scale is successful in Sri Lanka. Research on cultivation practices and breeding for specific purposes is needed prior to large-scale production in South-East Asia for medicinal purposes. A suitable method has already been developed for fingerprint analysis and standardization of all Asiatic pennywort preparations.

**Literature** [1] Babu, T.D., Kuttan, G. & Padikala, J., 1995. Cytotoxic and anti-tumour proper-

ties of certain taxa of Umbelliferae with special reference to *Centella asiatica* (L.) Urban. *Journal of Ethnopharmacology* 48(1): 53–57. [2] Bonte, F., Dunas, M., Chaudagne, C. & Meybeck, A., 1994. Influence of asiatic acid, madecassic acid and asiaticoside on human collagen I synthesis. *Planta Medica* 60(2): 133–135. [3] Buwalda, P., 1949. Umbelliferae. In: van Steenis, C.G.G.J. (Editor): *Flora Malesiana*. Series 1, Vol. 4. Noordhoff-Kolff N.V., Djakarta, Indonesia. pp. 116–117. [4] Kartnig, T., 1988. Clinical applications of *Centella asiatica* (L.) Urb. In: Craker, L.E. & Simon, J.E. (Editors): *Herbs, spices and medicinal plants: recent advances in botany, horticulture and pharmacology*. Vol. 3. Oryx Press, Phoenix, Arizona, United States. pp. 145–173. [5] Morelli, I., Bonari, E., Pagni, A.M., Tomei, P.E. & Menichini, F., 1983. Selected medicinal plants. *FAO Plant production and protection paper No 53/1*. Food and Agriculture Organization of the United Nations, Rome, Italy. pp. 33–35. [6] Peiris, K.H.S. & Kays, S.J., 1996. Asiatic pennywort (*Centella asiatica* (L.) Urb.): A little-known vegetable crop. *Hortotechnology* 6(1): 13–18. [7] Pointel, J.P., Boccalon, H., Cloarec, M., Ledevhat, C. & Joubert, M., 1987. Titrated extract of *Centella asiatica* (TECA) in the treatment of venous insufficiency of the lower limbs. *Angiology* 38(1): 46–50. [8] Sarma, D.N.K., Khosa, R.L., Chansauria, J.P.N. & Sahai, M., 1995. Antiulcer activity of *Tinospora cordifolia* Miers and *Centella asiatica* Linn. extracts. *Phytotherapy Research* 9(8): 589–590. [9] Sarma, D.N.K., Khosa, R.L., Chansauria, J.P.N. & Sahai, M., 1996. Antistress activity of *Tinospora cordifolia* Miers and *Centella asiatica* Linn. extracts. *Phytotherapy Research* 10(2): 181–183. [10] Solet, J.-M., Bister-Miel, F., Galons, H., Spagnoli, R., Guignard, J.-L. & Cosson, L., 1993. Glucosylation of thiocolchicine by a cell suspension culture of *Centella asiatica*. *Phytochemistry* 33(4): 817–820.

**Other selected sources** 97, 193, 202, 311, 315, 332, 350, 395, 560, 580, 597, 602, 705, 901, 1035, 1066, 1126, 1178, 1287, 1386, 1572, 1590.

Djoko Hargono, Pudji Lastari,  
Yun Astuti & M.H. van den Bergh

### **Chenopodium ambrosioides L.**

Sp. pl. 1: 219 (1753).

CHENOPODIACEAE

2n = 16, 32, 48, 64

**Vernacular names** Wormseed, Mexican tea (En). Philippines: alpasotis (general), adlabon (Ig-

erot), bubula (Bontok). Vietnam: c[aa]y d[aa]f[u] h[oo]l, c[aa]y d[aa]f[u] giun, th[oo]r kinh gi[ow]s[i].

**Origin and geographic distribution** *C. ambrosioides* is native to Central and South America, and has been introduced in Europe, Africa, Asia and Australia. In the Malesian region, it is naturalized in mountainous regions in Java, Sulawesi and the Philippines, and has occasionally been found in Papua New Guinea. Its cultivation in Java for medicinal purposes was abandoned because it was unprofitable. It is still cultivated in the Philippines.

**Uses** Wormseed is used all over the world as a vermifuge. It is an effective anthelmintic with a long history of use, and has been used in America since about 1800. Bruised fruits are administered in small doses or the juice from the plant is given undiluted or as a decoction in milk or water. It is effective against hookworms (*Ankylostoma duodenale*, *Necator americanus*), roundworm (*Ascaris lumbricoides*) and whipworm (*Trichuris trichiura*). The essential oil, which is generally considered as nervine and anti-rheumatic, is very effective against amoebae causing dysentery. Wormseed is also commonly used externally to treat ulcers, eczema and erysipelas. Crushed leaves are widely applied as poultices on bruises, insect bites and ulcers. In the Philippines, the leaves are also used as a carminative in poultices applied to the abdomen of children suffering from dyspepsia, and are also considered an emmenagogue. In Central America, it has been used as an antispasmodic and stomachic, and a decoction used to be administered as an internal haemostatic, as a remedy for indigestion and to treat ulcers. In Brazil, wormseed is used in the treatment of cutaneous leishmaniasis. Additional uses reported from southern Africa and Mexico include the application of an infusion against colds and stomach-ache, as an enema against intestinal ulceration, diuretic, emmenagogue and as sudorific. In India, the essential oil is employed in pectoral complaints and nervous affections. 'Di-fu-zi' is a Chinese drug widely found on markets in China. It is derived from *Kochia scoparia* (L.) Schrader (synonym: *Chenopodium scoparia* L.), but sometimes also from *C. ambrosioides* or *C. album* L.; the preparation is known for its diuretic and antifungal properties.

In Mexico, wormseed is used in animal health care, particularly to treat gastro-intestinal nematodes in sheep. Fish diseases caused by helminths (e.g. *Capillaria* spp., *Spirocamellanus* spp.) have been treated successfully in Mexico by using

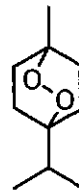
wormseed. Wormseed has been found to be unaffected by *Meloidogyne* spp., and can be used in crop rotations for sugar cane fields infested with these nematodes. In Africa Congo-Brazzaville, wormseed is traditionally used to protect groundnut from insect pests; it showed effective control of the beetle *Caryedon serratus*.

Wormseed oil or extract is used commercially as a fragrance component in lotions and perfumes. The herb is also used in Mexico as a condiment in soups, used sparingly to impart an acceptable flavour.

**Production and international trade** Wormseed was cultivated in the United States to control hookworms and roundworms until effective synthetic compounds became available in the 1950s. The annual world production of wormseed oil was estimated at 35 t by the end of the 1950s, but has since decreased. The price of the oil was US\$ 22–31 per kg in 1983.

**Properties** The glandular hairs present on leaves and fruits exude an essential oil. Oil yield is highest for fruits, up to 2.5%. The oil is colourless or pale yellow, has a peculiar disagreeable odour and a bitter, burning taste. About 50 compounds have been identified in the oil, accounting for 97%. The major compounds in the essential oil are p-cymene, limonene (up to 32.5%),  $\alpha$ -terpinene, trans-pinocarveol (up to 27%) and ascaridole (1,4-peroxido-p-menthene-2, up to 86%). The chemical composition of the oil seems to differ considerably depending on the origin of the plants and botanical variety. Limonene and trans-pinocarveol have been reported as the main constituents of the oil from Mexican plants, but high amounts of ascaridole have been reported in oil elsewhere. The flavonol glycosides kaempferol 3-rhamnoside-4'-xyloside and kaempferol 3-rhamnoside-7-xyloside, along with kaempferol, isorhamnetin and quercetin have been identified from the fruits. Per 100 g the leaves contain: 85 g water, 4 g protein, 0.7 g fat, 7.5 g total carbohydrates, 1.3 g fibre and 2.4 g ash.

The crushed plant, the expressed juice, the flower spikes, the seeds and the seed oil of *C. ambro-*



ascaridole

*sioides* are well known for their use as anthelmintics. The essential oil and its main component ascaridole, which is considered the main active principle, paralyze, but do not kill the intestinal worms; they must then be expelled by a laxative. Activity is reported against a variety of intestinal parasites, e.g. *Ankylostoma*, *Ascaris*, *Necator* and *Trichuris*. Leaf extracts also showed an in vitro activity against *Ascaris lumbricoides* eggs. The oil should be used with caution: mild reactions are headache, dizziness and nausea, but in overdose it can cause cardiac and respiratory disturbances, convulsions, drowsiness, vomiting and weakness. It should not be prescribed to persons with nervous, heart or kidney troubles, or to pregnant women.

The essential oil has fungitoxic activity. It has been found to show strong in vitro activity against the dermatophytes *Microsporium gypseum* and *Trichophyton rubrum*, and also against *Aspergillus fumigatus* and *Cladosporium trichoides*. Experimental ringworm infection in guinea-pigs was cured within 7–12 days by an ointment containing the oil. The essential oil inhibited growth of *Aspergillus flavus* effectively at 2000 ppm, and mycelial growth of *Rhizoctonia solani* (causing damping-off of seedlings of e.g. mungbean (*Vigna radiata* (L.) Wilczek) was totally inhibited by the oil at 1000 ppm on malt extract agar medium, without showing phytotoxic effects on germination and seedling growth. The lack of effect on germination and early growth of crops recorded in these experiments is contradictory to the allelopathic activity of wormseed oil and extract reported from other experiments. Storage for one year did not affect the fungitoxicity of the oil, neither did heating to 100°C. Dry residues of wormseed plants mixed with soil (10 g of residue with 90 g of soil) effectively controlled *Phytium aphanidermatum* and *Rhizoctonia solani* infection of common bean (*Phaseolus vulgaris* L.). At 1000 ppm wormseed oil was found to provide complete protection to stored wheat from all fungi investigated, without showing any phytotoxic effect. Ascaridole itself also has antifungal activity. In laboratory tests this compound gave over 50% inhibition of *Sclerotium rolfsii* (*Corticium rolfsii*) at a concentration of 1 mg/ml.

Experiments with mice showed that a methanol extract of the dried aerial parts of *C. ambrosioides* has a hypothermic effect at 2 g/kg, as well as inhibitive effect on acetic acid-induced writhing at 3 g/kg, suggesting that the plant has some sedative and/or analgesic effects. Separation and isolation

using these bioassays revealed that these effects can be attributed to ascaridole. Oral administration of ascaridole in mice at a dose of 100 mg/kg showed a significant hypothermic and analgesic effect, prolonged the anesthesia induced by sodium pentobarbital and reduced the locomotor activity enhanced by methamphetamine. Doses of 300 mg/kg, however, had lethal toxicity.

Ascaridole is also reported to be a potent inhibitor of plasmodial growth in lower concentrations, and to kill malarial parasites in higher concentrations. It is effective at about the same dose as chloroquine and artemisinin. Like artemisinin the compound is one of the few naturally occurring terpenes bearing a peroxide group. This peroxide group must be essential for the antimalarial activity of ascaridole, because 1,8-cineol (which has an epoxide group instead of a peroxide group) at identical concentrations is inactive.

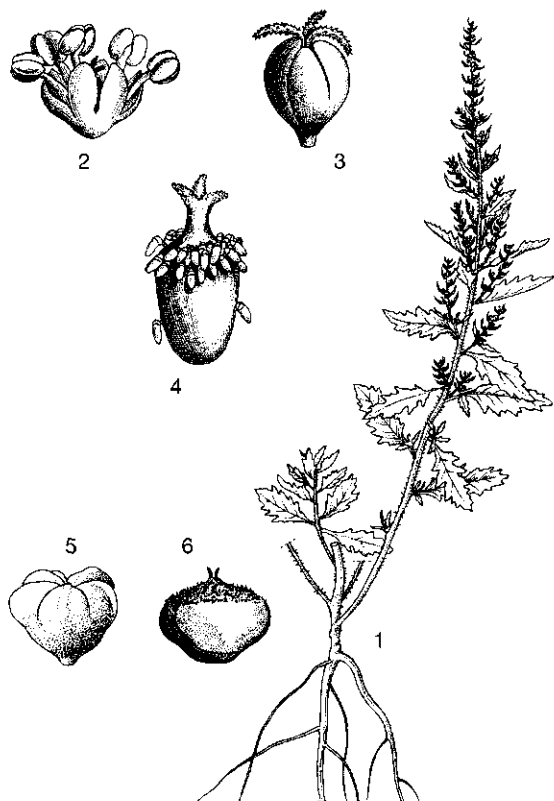
Wormseed contains saponin, which is located mostly in the roots; the aglycone fraction of the saponin is echinocystic acid. The saponin is reported to have antifungal and molluscicidal activity.

Wormseed kills and repels insects, and also acts as an antifeedant. Dried plants give stored products some protection against weevils and beetles. Foliage has been used successfully to control the gelechiid moth *Phthorimaea operculella*, a pest of stored Irish potato. Limonene is insecticidal against a variety of flies, mosquitoes, ants, beetles, weevils, fleas, wasps, crickets, ticks and mites.

The terpenes p-cymene, ascaridole and aritazone have an allelopathic effect, and may inhibit seed germination and seedling growth of other plants. Wormseed has a role in traditional agro-ecosystems in Mexico in controlling weeds and reducing nematodes. Farmers allow it to grow only when crops are ready to be harvested because of its allelopathic activity. Wormseed extracts showed a moderate antioxidant effect on feed fats. Leaf extracts are effective in inhibiting infection of crops such as common bean by tobacco mosaic virus.

**Adulterations and substitutes** In order to reduce the high ascaridole content to the minimum permissible requirement, synthetic chemicals corresponding to the constituents of the oil are frequently substituted for wormseed oil. Some *Artemisia* species were also popular as vermifuge in Europe, and commonly replaced wormseed there.

**Description** An erect or ascending annual herb up to 100(–150) cm tall, often very branched,



*Chenopodium ambrosioides* L. - 1, flowering plant; 2, bisexual flower; 3, female flower; 4, ovary with glands; 5, fruit enclosed by perianth; 6, fruit after removal of perianth.

strong-smelling; stem angularly ribbed, glabrous or finely pubescent. Leaves alternate, oblong-lanceolate, 1.5–15 cm × 0.5–5 cm, acute and often almost decurrent at base, acute to obtuse at apex, usually coarsely or shallowly serrate-dentate but highest leaves entire, herbaceous, bright green, lower surface variably densely studded with yellow glands, otherwise subglabrous, secondary veins thin; petiole short; stipules absent. Flowers in 3–25-flowered clusters in the axil of bractlike leaves, united in lax spikes together forming a leafy panicle, small, bisexual or some female, sometimes some male; perianth 4–5-cleft to near the base, 1–1.5 mm long, pale green with a paler base, with ovate-triangular, very concave segments; stamens (1–)4–5, filaments free, slightly exceeding the perianth; ovary superior, depressed globose, with many small, yellow glands on top, 1-celled, stigmas (2–)3–5; fruit a nut entirely concealed by the connivent tepals, 1-seeded. Seed

usually horizontally in fruit, broadly obovoid or ellipsoid, 0.6–0.8 mm in diameter, shiny brownish-black.

**Growth and development** Wormseed flowers and fruits throughout the year. It produces massive amounts of seed.

**Other botanical information** *Chenopodium* comprises perhaps up to 250 species. *C. ambrosioides* is very variable and shows an extremely large area of distribution. Several infraspecific taxa have been distinguished giving rise to at least 12 different varieties. Most important are var. *ambrosioides* and var. *anthelminticum* (L.) A. Gray (synonym: *C. anthelminticum* L.) which are commonly cultivated in many warmer parts of the world.

Several other *Chenopodium* species are used in folk medicine in Central and South America for similar purposes as wormseed, e.g. *C. chilense* Schrader, *C. graveolens* Willd. and *C. multifidum* L.

**Ecology** *C. ambrosioides* occurs locally abundantly along roadsides and in waste places, sometimes also in upland rice fields; in Java it occurs at 1600–2000 m altitude.

**Propagation and planting** Wormseed is propagated by seed. It is reported that 6–10 kg/ha of seed is sufficient. Seeds germinate 7–21 days after sowing. Germination is promoted by light and optimum temperatures are 15–35°C. Imbibed seed should be pretreated with low temperatures. The optimal planting distance in Java is 1 m between rows and 0.5 m within the row.

**Husbandry** Wormseed needs plenty of water during early growth, but later it is quite drought tolerant. Large doses of N fertilizer reduce the ascaridole content of the plants; it is common practice to apply N fertilizer at 40 kg/ha. A leguminous cover crop, e.g. *Vigna hosei* (Craib) Backer ex K. Heyne, is sometimes used in Indonesia, making N fertilization redundant.

**Diseases and pests** In South America, downy mildew caused by *Peronospora* spp. is an important disease in cultivated *Chenopodium* spp., including wormseed. Weedy wormseed can be a host of powdery mildew; the disease can spread to tomato.

**Harvesting** Wormseed is harvested when the fruits are ripe. Usually they are stripped off by hand in the early morning and in dry weather.

**Yield** An experimental plantation in Java yielded about 8 t/ha of dried fruits during a period of 1.5 years with 3 successive harvests; after each of the first two harvests the plants were cut and fer-

tilizer was applied. Fruits may yield 1–2% of oil on distillation, thus giving a yield of 80–150 kg of oil per ha. Experimental plantations in Germany yielded 70 t of fresh and 14 t of dried whole plant material per ha.

**Handling after harvest** Harvested fruits are sun-dried and cleaned from broken leaflets and flower remains by sieving. The oil is obtained by steam distillation at 130–140°C and at a pressure of 3.5–4 atmosphere. Pure oil can be obtained when cooled to 50°C or more.

**Genetic resources and breeding** Wormseed is spread worldwide and needs no protection measures. It exhibits considerable variation in morphology and chemical composition and offers potential for breeding for specific purposes (e.g. var. *anthelminticum* has a high proportion of ascari-dole).

**Prospects** Although the importance of wormseed as anthelmintic has waned, it may have promising prospects for various uses. The leaves seem safe to be used as a medicinal tea, as long as it is not overused. Stored products such as beans could possibly be treated with wormseed oil for pest protection, provided there are no toxic effects for mammals. The effects of a wormseed crop in the control of weeds and nematodes in a rotation with field crops should be further investigated.

**Literature** [1] Backer, C.A., 1949. Chenopodiaceae. In: van Steenis, C.G.G.J. (Editor): Flora Malesiana. Series 1, Vol. 4. Noordhoff-Kolff, Djakarta, Indonesia. pp. 99–106. [2] Badawy, E.-S.M., 1979. Ökologische und ontogenetische Einflüsse auf Drogenenertrag, Nährstoffentzug und arzneiliche Wirkstoffe von *Chenopodium ambrosioides* L., Traubenkraut [Ecological and ontogenetical influences on drug yield, nutrient consumption and medical agents in *Chenopodium ambrosioides* L.]. Thesis. Institut für Pflanzenbau und Pflanzenzüchtung, Giessen, Germany. 159 pp. [3] Jimenez-Osornio, F.M.V.Z.J., Kumamoto, J. & Wasser, C., 1996. Allelopathic activity of *Chenopodium ambrosioides* L. *Biochemical Systematics and Ecology* 24(3): 195–205. [4] Kishore, N., Dixit, S.N. & Dubey, N.K., 1989. Fungitoxic studies with *Chenopodium ambrosioides* for control of damping-off in *Phaseolus aureus* (Moong) caused by *Rhizoctonia solani*. *Tropical Science* 29(3): 171–176. [5] Kishore, N., Mishra, A.K. & Chansouria, J.P.N., 1993. Fungitoxicity of essential oils against dermatophytes. *Mycoses* 36(5–6): 211–215. [6] Okuyama, E., Umeyama, K., Saito, Y., Yamazaki, M. & Satake, M., 1993. Ascari-dole as a pharmacologically active principle of 'Paico', a medicinal Peru-

vian plant. *Chemical and Pharmaceutical Bulletin* 41(7): 1309–1311. [7] Paré, P.W., Zajicek, J., Ferracini, V.L. & Melo, I.S., 1993. Antifungal terpenoids from *Chenopodium ambrosioides*. *Biochemical Systematics and Ecology* 21(6–7): 649–653. [8] Quarles, W., 1992. Botanical pesticides from *Chenopodium*. *IPM Practitioner* 14(2): 1–11. [9] Sagrero-Nieves, L. & Bartley, J.P., 1995. Volatile constituents from the leaves of *Chenopodium ambrosioides* L. *Journal of Essential Oil Research* 7(2): 221–223. [10] Vázquez-Yanes, C. & Orozco-Segovia, A., 1990. Ecological significance of light controlled seed germination in two contrasting tropical habitats. *Oecologia* 83(2): 171–175.

**Other selected sources** 97, 332, 373, 374, 380, 733, 742, 979, 1035, 1126, 1149, 1178.

Undang A. Dasuki

## Cinchona L.

Sp. pl. 1: 172 (1753); Gen. pl. ed. 5: 79 (1754).

RUBIACEAE

$x = 17$ ; *C. officinalis*, *C. pubescens*:  $2n = 34$

**Major species** *Cinchona officinalis* L., *C. pubescens* Vahl.

**Vernacular names** *Cinchona*, quinine (En). Quinquina (Fr). Indonesia: kina. Malaysia: kuinin. Cambodia: kini:n. Thailand: quinin. Vietnam: canh ki na.

**Origin and geographic distribution** *Cinchona* comprises about 50 species. The centre of diversity lies along the Andes mountains of Bolivia, Peru, Ecuador, Colombia and Venezuela. There the species and natural hybrids flourish on the misty and humid eastern slopes at 800–3000(–3700) m altitude.

The collection of *Cinchona* germplasm for dispersal outside its centre of origin started in 1848. Hasskarl's expeditions to South America (1852–1854) provided the Dutch in Java (Indonesia) with plant material. A British expedition under Markham did likewise for India and Sri Lanka (1859). In both cases, the plant material adapted to the local conditions with no problems. However, the bark of the vigorously growing trees appeared to contain such small amounts of quinine that it was not profitable to start cultivation. Seeds Ledger (1864) collected in Bolivia produced weakly growing seedlings in Java, some of which had an unusually high percentage of quinine in the bark. Meanwhile, seed samples from South America and from importations from surrounding coun-



tries had arrived in nearly every country of South-East Asia.

*Cinchona* started to be distributed worldwide in the second part of the 19th Century. Around 1880, Sri Lanka had become a major producer of cinchona bark, albeit of low quality. By 1895 it had been superseded by the Dutch East Indies (Indonesia) as the main producer, mainly because of the better quality of the bark (*C. officinalis*). The crop was introduced into West and East Africa (Guinea, Cameroon, Kenya, Tanzania) and Central Africa (Congo Kinshasa, Rwanda) in the 1930s. In all of these countries, except for Congo Kinshasa, production has dwindled. After the rapid decline of bark production in South America around 1880, interest revived in Latin America around 1940, especially in Guatemala, but declined sharply after 1945.

In Asia, *Cinchona* cultivation is still important in Indonesia and India. The plantations in Burma (Myanmar), the Philippines, Sri Lanka and Vietnam have been abandoned, as have those in Australia (Queensland) and Papua New Guinea.

**Uses** The Spanish conquerors of Central and South America reported the use of cinchona bark by Indian miners in the Andes to suppress shivering from the cold in the mines. Later, Jesuits found that shivering was caused by fever. This led to the discovery of cinchona bark as a remedy against malaria. Anti-malarial drinks containing small quantities of quinine, one of the major alkaloids found in cinchona bark, were developed for use in the tropics especially in India; they are still very popular (e.g. tonic water).

Malaria is a disease mostly found in tropical areas, where it constitutes a major medical problem. It is characterized by attacks of severe fever, which recur at regular intervals. There is also a form with irregular attacks of severe fever. The disease is caused by a parasitic protozoan of the genus *Plasmodium*, which uses mosquitos of the genus *Anopheles* as an intermediary host. When an infected mosquito bites a person, sporozoites enter the blood, but they disappear rapidly from the circulation to localize in the parenchymal cells of the liver in which they grow and segment. On reaching maturity these merozoites are released from the liver cells and penetrate erythrocytes where further division and development takes place. When this process is complete, the erythrocytes burst open and the merozoites enter the blood stream. It is this periodic breaking of erythrocytes that induces the chill so characteristic of malaria. The fever following the chill is the body's

response to the liberated foreign protein and cell products.

Some of the merozoites infect new blood corpuscles, while others develop into the sexual form, called gametes. The gametes can pass to a healthy mosquito when it bites a person suffering from malaria. The gametes conjugate in the mosquito, forming sporozoites, and the circle is complete. Quinine acts by killing the merozoites in the blood, except for those in the reproductive stages. Quinine has largely been replaced by synthetic anti-malarials (e.g. chloroquine), which have fewer side-effects. In recent years, however, renewed interest has arisen in quinine and related alkaloids because of the growing resistance of malaria-causing agents (*Plasmodium* spp.) to industrial anti-malarial drugs currently in use. Additionally, large amounts of quinine are used as a bitter flavouring in soft drinks and in innumerable other products, such as hair oils and shampoos, sun-tan oil, insecticides, as a vulcanizing agent in the rubber industry, and in the preparation of certain metals.

Another cinchona alkaloid, quinidine (a stereoisomer of quinine) is also active against malaria, but is nowadays mainly employed as an anti-arrhythmic. Minor uses of cinchona alkaloids include the treatment of ophthalmia, internal haemorrhoids and hiccups. A tincture has been used as a bitter to stimulate appetite and digestion. The alkaloids are also used in insecticides and moth repellants. Other applications of cinchona alkaloids are in the asymmetrical catalysis of chemical reactions. After extraction of alkaloids, the bark is still useful for tanning leather.

**Production and international trade** Indonesia maintained an almost total monopoly on the production of cinchona bark for nearly 50 years, up to the Second World War. In terms of ready product, this amounted to roughly 800 t/year of quinine sulphate. Since the Second World War, Indonesia has been gradually caught up and surpassed by Congo Kinshasa, although substantial quantities of quinine sulphate are still being produced in Indonesia, Guatemala, Tanzania and other countries.

International trade figures are often difficult to interpret because of the varying ways the quinine content of cinchona bark is indicated. In the past SQ2 and SQ7 were used most frequently, indicating 2 and 7 water molecules respectively. Nowadays, percentages are generally indicated as QAA, the anhydrous form of quinine salts: 1% QAA = 1.206% SQ2 = 1.345% SQ7.

At present, world production of cinchona alkaloids is estimated at about 600 t/year of QAA, for which about 10 000 t of bark are extracted, with Congo Kinshasa producing about 55%, Indonesia 30%, India 8% and the other countries 7%. Stripe canker (*Phytophthora cinnamomi*) is a threat to production in East and Central Africa.

Although there are factories in bark-producing countries (Indonesia, India, Guinea, Congo Kinshasa and Rwanda) that extract alkaloids, most of the end-products are still manufactured in Europe (West Germany, the Netherlands, France). About 60% of the production is used for pharmaceuticals, most of the remaining 40% in the food and beverage industry.

**Properties** More than 36 different alkaloids have been reported as constituents of various *Cinchona* barks, the most important being quinine, quinidine, cinchonine and cinchonidine. Biosynthetically, these alkaloids are derived from the amino acid tryptophan and a monoterpenoid skeleton of the *Corynanthe* type. Quinine and quinidine are stereoisomers, their difference being the configurations at C-8 and C-9. Cinchonine and cinchonidine form another set of stereoisomers which lack the methoxy group at the C-6' position. Quinine and quinidine are of medicinal value; they are used to treat malaria and cardiac arrhythmias, respectively. Approximately 30–50% of the quinine produced is chemically converted to quinidine. The other alkaloids are not used medicinally, although they show effects similar to qui-

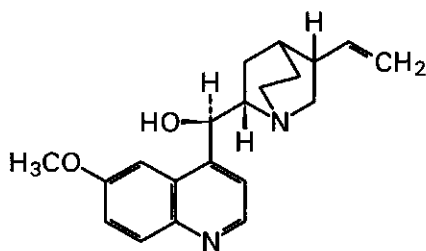
nine and quinidine. Cinchonine showed inhibition of human platelet aggregation.

The composition and content of alkaloids vary with species, genotype, environment and age of the bark. Generally, the alkaloid concentration is higher in bark from the bole than in branch bark. Selected clones of *C. officinalis* have been reported to yield as much as 14–16% quinine from dry bark. Normally, *Cinchona* bark has a total alkaloid content of 3–15%, and pharmaceutical bark must contain at least 6%. The quinidine content is generally low; about 1% of the total amount of alkaloids. Quinine can be converted into quinidine by means of a rather complicated chemical process. Alkaloids known as the cinchophyllines have been isolated from leaves of *C. officinalis*; they may be regarded as indole analogues of emetine and have shown in vitro amoebicidal activity. Research on the anti-microbial activity of a series of quasi-dimeric alkaloids found activity against gram-positive bacteria, but no activity against gram-negative bacteria, yeast and several fungi. A weak local anesthetic activity was found for cinchophyllamine, as well as some analgesic activity.

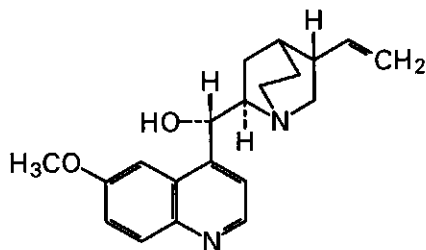
**Adulterations and substitutes** Numerous plant resources have been used and are still used in traditional medicine to treat malaria. One of the most important alternatives to cinchona is the herb *Artemisia annua* L. (or its isolated active compound artemisinin), which is widely used in Vietnam and China ('Quinghaosu'). Other species used in traditional medicine to treat malaria that have recently shown in vitro antiplasmodial activity are *Azadirachta indica* A.H.L. Juss., *Brucea javanica* (L.) Merr., *Cyclea barbata* Miers and *Dichroa febrifuga* Lour.

**Description** Evergreen woody shrubs or small to medium-sized trees, 8–16 m, occasionally up to 30 m tall; bark thick, greyish-brown to brown. Leaves opposite, oblong-elliptical, simple and entire; stipules interpetiolar, deciduous and leaving a characteristic scar. Inflorescence a terminal panicle, many-flowered. Flowers 1–2 cm long, fragrant, 5-merous, heterodistylous, pink or yellowish; calyx small, with pointed lobes; corolla tubular with spreading lobes with a fringe of hairs along the margins; stamens alternating with the corolla lobes and inserted in the corolla tube; ovary inferior, bilocular, style at the base with a circular disk, ending in a bifid stigma. Fruit a 1–3 cm long capsule containing 40–50 seeds. Seeds flat, winged, 4–5 mm × 1 mm.

**Growth and development** Freshly harvested *Cinchona* seeds include varying numbers of im-



quinine



quinidine

mature and deteriorated ones. After these have been removed the germination percentage is usually more than 90% after 2–3 weeks of incubation. The seeds remain viable over a year if stored dry, cool and dark. Light promotes the germination.

The tiny seedlings develop slowly at first but then speed up: after about 2 months 2–3 pairs of leaves have formed. Flowering starts after 4–7 years or even earlier under stress conditions. There is a periodicity in flowering which has not been fully investigated. Cross-pollination is by insects, mainly bees, butterflies and flies. Fruits mature about 7–8 months after flowering.

**Other botanical information** Most *Cinchona* cultivated in South-East Asia is known under the name *C. ledgeriana*, and most probably are high-yielding selections of *C. officinalis*. *Cinchona* known under the name *C. succirubra* belongs to *C. pubescens*.

The majority of the species and hybrids from the centre of diversity do not produce valuable chemical compounds, but might nevertheless be of interest for breeding.

**Ecology** In the natural habitat of *Cinchona*, high, evenly distributed annual rainfall (up to 4000 mm) and high relative humidity prevail. *Cinchona* grows optimally with a rainfall of 2500–3800 mm well distributed throughout the year. Nevertheless, *Cinchona* is known to grow under drier conditions as well (1500 mm, with distinct dry season) and it can stand an annual precipitation of 5000 mm, provided this is well distributed throughout the year. Low irradiation (misty slopes, forest canopy) is frequently encountered in regions where *Cinchona* occurs naturally. In Asia, *Cinchona* grows well in areas with an average minimum temperature of 14°C and an average maximum temperature of 21°C. Growth is hampered severely below 7°C and above 27°C. Altitudinal range is largely determined by the prevailing climatic conditions, but generally lies between 800–2000 m. It has been reported that the yield of quinine is low in plants cultivated under 800 m altitude, and that the plants are susceptible to diseases. Growth is slow at elevations above 2000 m. *Cinchona* cannot stand waterlogging.

Favourable soil types are slightly acid, well drained, with a good water-retaining capacity. *Cinchona* grows well on soils of volcanic origin. The most important species, *C. officinalis*, is very vulnerable to weed competition; *C. pubescens* is more competitive.

**Propagation and planting** *Cinchona* is propagated by seed as well as by vegetative means.

Seedbeds are carefully prepared to give a fine tilth. The small seeds are broadcast on the soil surface (3000–12 000/m<sup>2</sup> corresponding with 1–4 g/m<sup>2</sup>) and protected against wind, rain and direct sunlight. Germination starts within 2–3 weeks; after 4–6 months plantlets are 5–10 cm tall and are moved to nursery beds where they stay 6–7 months. The seedlings require temperature, light and ventilation to be carefully controlled, to avoid damping-off and other hazards. Young plants can be transplanted to the field when 1–1.5 years old. The rather delicate *C. officinalis* is often grafted on the more robust and vigorous *C. pubescens*. Seedlings of the latter reach the proper size for grafting after about 1 year. The scion is usually inserted by side-tongue grafting, but green-budding is also applied. Cuttings are difficult to root; cuttings taken from shoots formed after topping give better results.

The isolation and multiplication of high-yielding or disease-tolerant trees by in vitro culture techniques is a promising new method that may result in high-yielding and disease-resistant clones being available for planting in the near future. In vitro micro-grafting of *C. officinalis* on *C. pubescens* has proved to be successful and is comparatively simple.

*Cinchona* is almost exclusively grown as an estate crop, except in Congo Kinshasa where smallholders occasionally grow it. It is mostly grown as a sole crop, although in Congo Kinshasa it is occasionally intercropped with beans. *Cinchona* is planted in the field in holes of 50 cm × 50 cm × 50 cm, 80–150 cm apart, in rows or in a triangular arrangement depending mainly on the topography of the field. Before planting, the plants are pruned back about one-third, or defoliated by 50%. The planting out takes place at the beginning of the rainy season. Leguminous cover crops may be planted between the rows (e.g. *Desmodium* in Congo Kinshasa, *Crotalaria trichotoma* Bojer or *Shuteria vestita* Wight & Arnott in Indonesia) or on the contour to prevent erosion (e.g. *Leucaena leucocephala* (Lamk) de Wit).

**In vitro production of active compounds** In recent decades, much attention has been paid to the biosynthesis of cinchona alkaloids in in vitro cell, tissue and organ cultures. Fine cell suspensions do not produce alkaloids, and only cultures showing some form of differentiation produce alkaloids in reasonable amounts. Studies of possible biotechnological production of alkaloids with plant cell cultures are in progress, but have not yet led to large-scale processes.

Hypocotyl explants from seedlings can be induced to form callus on solid Gamborg B5 medium (0.7% agar), containing 2,4-dichlorophenoxyacetic acid (1 ppm) and kinetin (0.2 ppm). Alkaloid production is low and growth slow in cell and tissue cultures. It has been found that growth and indole alkaloid production (e.g. cinchonamine) was improved by increasing the auxin concentration in callus cultures, but anthraquinone production and quinoline alkaloid levels (e.g. quinidine) were highest when auxin concentrations were reduced. Low and medium cytokinin concentrations benefit the production of quinoline alkaloid. Adding the precursor tryptophan increases the amount of alkaloids produced, but reduces growth. The best growth was obtained in the light, although many media resulted in no growth at all in the light. From the results of the experiments with tissue culture it was concluded that the pathways leading to the various secondary products (anthraquinones, indole alkaloids and quinoline alkaloids) are, at least partly, regulated independently.

**Husbandry** Two systems of cultivation are applied:

- A short-term, intensive, high-production system with a relatively short production cycle of about 10 years from planting to harvesting. It is practised mainly in Congo Kinshasa. Planting is at densities of 10 000–12 000 plants/ha. Weeding is mostly by hand, although the use of herbicides is increasing. Around the third year after planting, weeds are shaded out because of the development of the canopy. At the same time pruning and thinning starts, producing the first harvest of low-quality bark. Thinning continues until, around 10 years after planting, a stand of 3000 well-shaped trees is left. These are then harvested completely, producing a minimum of 3.5 kg of high-quality bark per tree.
- A long-term, extensive, intermediate-production system with a longer occupation period. It is practised in Indonesia and Guatemala. Planting is at a density of 5000 plants/ha. Weeding is necessary over a longer period, while pruning is only carried out to shape the trees. After 7–8 years, when competition for light becomes a limiting factor, all trees are coppiced to a height of 15–20 cm. In maintaining a maximum of 2–3 shoots per stool, a new cycle is started which is treated in the same way as the first one. If proper care is taken and mortality after coppicing is not too high, this system of production can be maintained for several decades. It is also suitable for

*Cinchona* cultivation under the shade of rainforest trees, which are left to prevent serious erosion.

A combination of both systems is practised in West Bengal (India), where *C. officinalis* seedlings are first coppiced and then after completion of the second cycle, harvested completely. Modifications of these 2 systems have been developed to meet local conditions. One involves grafting clonal *C. officinalis* or hybrids on a rootstock of *C. pubescens*, giving uniform planting material, better growth and tolerance or resistance to *Phytophthora cinnamomi*. This method is practised in Indonesia and Guatemala.

Composite fertilizers such as NPK (20-10-10 or 15-15-15) are widely applied, although other compounds such as phosphates and oligo-elements are used as well, depending on the local conditions. In general, a final dressing of nitrogenous fertilizer (100–600 kg/ha of the above-mentioned NPK) about 6 months prior to harvest increases the alkaloid content of the bark. Where soils are low in organic matter, *Cinchona* responds well to the mulching.

Mechanization is not widespread in *Cinchona* cultivation, partly because of the fields are often undulating. For the time being, it is mainly limited to the application of herbicides and insecticides and, to a lesser extent, to the harvest and stripping of the trees. However, mechanization is becoming more important where labour is scarce.

**Diseases and pests** Seedlings are susceptible to *Pythium* spp., *Rhizoctonia solani* (causing damping-off), *Fusarium solani* (causing wilt), *Phytophthora cinnamomi*, *Sporotrichium* and *Verticillium* species (causing stem blight) and *Sclerotium rolfsii* (causing seedling blight). Attacks can easily be overcome by chemically sterilizing the seedbed, and by regularly shifting the nursery site. In later stages, *Cinchona* is vulnerable to *Phytophthora cinnamomi*, *P. parasitica* (causing top blight and girdle canker), *Corticium salmonicolor* (dieback of branches), and *Armillaria* sp. (root rot). Other fungi (*Alternaria*, *Cercospora* and *Sclerotium* spp.) are of little economic importance. In areas with *Phytophthora cinnamomi* and *P. parasitica*, a combination of cropping techniques (e.g. cover crops) should be practised to avoid infestation, because once these diseases have taken hold the application of fungicides is almost impossible and too expensive. *Phytophthora cinnamomi* can also be avoided by grafting on a *C. pubescens* rootstock. The outbreak of *Corticium salmonicolor* can be avoided by timely pruning of trees, and of *Armill-*

*laria* sp. and *Fomes noxius* by consistent removal of old stumps. There are indications that insufficient drainage and planting too deep may favour the incidence of a physiological canker.

The main pest in *Cinchona* is *Helopeltis* spp., which can cause considerable damage by sucking young shoots and leaves. *Helopeltis* outbreaks can be avoided by timely application of insecticides. Occasional outbreaks of other pests such as variou caterpillars (e.g. *Delephila nerii*) and borers occur, but are only of local importance.

**Harvesting** In general, two phases of harvest can be distinguished: pruning and thinning in the early years and the final harvest.

The bark is removed in various ways. In Indonesia and Congo Kinshasa, bark is removed by clubbing, but in Tanzania and Guatemala knives are used. Bark peeling machines are used occasionally.

**Yield** Pruning and thinning result in relatively low yields of bark and alkaloids. At the final harvest, yields of at least 10 t/ha of dry bark are obtained with the short-term production system. The bark from selected planting material may contain at least 7% QAA on average, resulting in more than 700 kg/ha of QAA. Both bark yield and alkaloid content vary considerably, as they are affected by various factors.

Yields from the long-term production system are generally lower in terms of production per ha per year. However, this system can be more advantageous in terms of return on investment.

As a guideline, industrial *Cinchona* plantations should produce an average of 50–100 kg/ha per year of QAA to give a safe return on investment. A plantation should be at least 300 ha in size to sustain the initial and overhead costs involved.

**Handling after harvest** The stripped bark is left to dry, preferably in the shade, or dried artificially. Drying in the open air has to be well supervised, because heating of wet bark may result in substantial losses of alkaloids. The bark should be spread thinly and turned over regularly. It is ready for further treatment when its moisture content is about 10%. Properly dried bark can be kept for several months without deterioration. It can be milled before packing, to facilitate shipment over long distances. Extraction and processing of the alkaloids to either totaquina, quinine bisulphate, quinine sulphate, quinine HCl or quinidine is mainly carried out in western Europe.

**Genetic resources and breeding** The dispersal of *Cinchona* seeds in the mid-19th Century is well documented. However, the limited survival rate of seeds and the destruction of earlier, low-

yielding introductions have resulted in a very limited genetic variation in the germplasm available outside the centre of diversity. Care should be taken to preserve the germplasm present in the centre of diversity for future use. Most work on breeding has been carried out in Indonesia. At an early stage it was concluded that besides a high quinine content, other parameters such as bark production, tree shape and vegetative growth were also important in determining yield. The 'ring method' was developed; this involves calculating the amount of quinine (in g) in a ring of bark 1 dm in width, at a height of 1 m, by multiplying the girth (in dm) at that height by the amount (in g) of water-free bark/dm<sup>2</sup> and the average quinine content of the bark. However, since the girth of a tree is a function of the plant density, this method proved insufficiently reliable for judging the amount of bark.

In 1931, some *C. officinalis* seeds of Indonesian origin had reached Congo Kinshasa to start a selection programme at the Mulungu experimental station near Bukavu. Elite trees were selected from the original population, vegetatively propagated, and planted in isolation. Seeds from these plots were harvested and distributed to local farmers and plantation enterprises. This policy resulted in rapid progress in bark production in Congo Kinshasa and neighbouring countries. Quinine percentages of up to 15% QAA were found in trees of about 10 years. However, there has been little further progress since the mid 1960s.

In India, breeding work has focused on selection of elite types, vegetative propagation of these types for industrial plantings, and controlled crosses between selected parents. Various methods of vegetative propagation have been tried: cuttings, air layering, budding, grafting, inarching. Budding and the production of cuttings by top-working have been most successful. This breeding programme has not been very successful, as the quality of the bark has not improved over the years. In the 1940s and 1950s a breeding programme was undertaken in Guatemala: hybrids of *C. pubescens* and *C. officinalis* were grafted on a *C. pubescens* rootstock or planted as cuttings. However, this programme was short-lived. Most breeding programmes have been abandoned (Congo Kinshasa, Guatemala) or give disappointing results (India, Indonesia). However, progress could be achieved for instance, by producing and distributing selected plant material, selecting suitable *C. pubescens* rootstocks to be used for grafting, and breeding for appropriate rooting architecture and disease re-

sistance in *C. officinalis*. Although not much progress is to be expected in obtaining higher quinine content in selected individuals, there is potential to increase QAA production per ha per year by at least 50%. In Indonesia, over 300 clones of *C. officinalis* and *C. pubescens* are maintained in germplasm collections.

**Prospects** Interest in *Cinchona* has recently been increasing. Vegetative propagation by means of tissue culture has provided a tool for more effective breeding programmes. It will play an increasingly important role in future plantings. The introduction of high-yielding, multi-line cultivars may improve productivity significantly. Improved cropping techniques (e.g. mechanization) will play an important role in the economics of the crop. Research has also been focused on the production of alkaloids by means of cell culture. Although stable cultures have been successfully established and small quantities of QAA have been produced, this method is still far removed from industrial application. *Cinchona* alkaloids have played a useful role in human life for more than 350 years. There are encouraging prospects of obtaining higher production levels at lower costs. This may be an important contribution in future malaria treatment, since the need for a cheap, effective therapy is becoming more important because of the increasing occurrence of this disease.

**Literature** |1| Chatterjee, S.K., 1977. Cultivation of quinine-yielding cinchona and emetine-yielding ipecac in India. Proceedings 4th Symposium Pharmacognosy and Chemistry of Natural Products, Leyden, the Netherlands. pp. 89-97. |2| Dürbeck, K., 1983. Anbau von *Cinchona* sp. zur Chinaringengewinnung [Cultivation of *Cinchona* sp. for production of quinine bark]. Thesis, Technische Universität, München, Germany. 101 pp. |3| Harkes, P.A.A., Krijbolder, L., Libbenga, K.R., Wijnsma, R., Nsengiyaremge, T. & Verpoorte, R., 1985. Influence of various media constituents on the growth of *Cinchona ledgeriana* tissue cultures and the production of alkaloids and anthraquinones therein. Plant Cell, Tissue and Organ Culture 4: 199-214. |4| Hunter, C.S., 1986. In vitro propagation and germplasm storage of *Cinchona* L. In: Withers, L.A. & Alderson, P.G. (Editors): Plant tissue culture and its agricultural applications. Butterworth, London, United Kingdom. pp. 291-301. |5| Kerbosch, M., 1948. De kina cultuur [The cultivation of quinine]. In: van Hall, C.J.J. & van de Koppel, C. (Editors): De landbouw in de Indische Archipel [The agriculture in the East Indian Archipel]. Vol. Ila. W. van Hoeve, the

Hague, the Netherlands. pp. 747-865. |6| McHale, D., 1986. The cinchona tree. Biologist 33: 45-53. |7| Phillipson, J.D., O'Neil, M.J., Wright, C.W., Bray, D.H. & Warhurst, D.C., 1987. Plants as sources of antimalarial and amoebicidal compounds. In: Leeuwenberg, A.J.M. (Editor): Medicinal and poisonous plants of the tropics. Proceedings of Symposium 5-35 of the 14th International Botanical Congress, Berlin, 24 July-1 August 1987. Pudoc Wageningen, the Netherlands. pp. 70-78. |8| Sharma, A., Tewari, R., Ganniyal, A.K. & Virmani, O.P., 1987. *Cinchona*: A review. Current Research on Medicinal and Aromatic Plants (CROMAP) 9: 34-56. |9| van Harten, A.M., 1969. *Cinchona* (*Cinchona* spp.). In: Ferwerda, F.R. & Wit, F. (Editors): Outlines of perennial crop breeding in the tropics. Miscellaneous Papers 4, Landbouwhogeschool, Wageningen, the Netherlands. pp. 111-128. |10| Verpoorte, R., Schripsema, J. & van der Leer, T., 1988. *Cinchona* alkaloids. The Alkaloids 34: 331-398.

#### *Selection of species*

### ***Cinchona officinalis* L.**

Sp. pl. 1: 172 (1753).

**Synonyms** *Cinchona calisaya* Wedd. (1848), *Cinchona ledgeriana* Moens ex Trimen (1881).

**Vernacular names** Crown cinchona, Ledger cinchona, yellow cinchona (En).

**Distribution** Naturally distributed in South America from Colombia to Bolivia; planted in many tropical countries, e.g. in India and Indonesia (Java).

**Uses** The bark is the traditional source of quinine, the classical industrial anti-malaria drug.

**Observations** A small tree, up to 16 m tall; leaves 7-28 cm × 2.5-13 cm, glabrous and with domatia beneath; flowers with glabrous calyx and yellowish-white to fleshy-coloured or red corolla, 8-17 mm long; fruit up to 25 mm long, glabrous or sparsely pubescent. *C. officinalis* occurs naturally in mountainous regions at 1200-3000 m altitude. In Java, it is planted at 800-2000 m.

**Selected sources** 97, 99, 202, 501, 580, 900, 1167, 1178, 1277.

### ***Cinchona pubescens* Vahl**

Skript. Naturh. Selsk. 1: 19 (1790).

**Synonyms** *Cinchona cordifolia* Mutis (1793), *Cinchona succirubra* Pav. ex Klotzsch (1858).

**Vernacular names** Red cinchona (En).

**Distribution** Naturally distributed in Central



*Cinchona officinalis* L. – 1, tree habit; 2, flowering twig; 3, flowers in longitudinal section showing heterodistily; 4, fruiting twig.

and South America from Costa Rica to Bolivia; planted in many tropical countries, e.g. in India and Indonesia (Java).

**Uses** The bitter constituents of the bark are a traditional anti-malarial of South-America, and have been used in the Malesian region since the 19th Century. The quinine is rather difficult to extract from the bark; also used as rootstock for *C. officinalis*.

**Observations** A small to medium-sized tree, up to 30 m tall; leaves 24–50 cm × 17–40 cm, pubescent and without domatia beneath; flowers with calyx pubescent outside and greenish-white corolla reddish towards lobes, 15–21 mm long; fruit (20–)30–40 mm long, initially pubescent but glabrescent. *C. pubescens* occurs naturally in mountainous regions at 1000–3700 m altitude. In Java, it is planted at (1000–)1250–1600(–2000) m.

**Selected sources** 97, 99, 202, 350, 501, 580, 900, 1167, 1178, 1277, 1640.

G. Staritsky, E. Huffnagel, A. Dharmadi & S.L. Dalimoenthe

**Cissampelos pareira** L.

Sp. pl. 2: 1031 (1753).

MENISPERMACEAE

*n* = 12

**Vernacular names** Indonesia: mangaloke (Moluccas). Philippines: sansau (Tagalog), sampare (Bisaya), kalaad (Iloko). Thailand: khong khamao (northern), khrueta ma noi (eastern), krung khamao (peninsular). Vietnam: d[aa]y m[oos]i, m[oos]i tr[of]n, ti[ees]t d[lee].

**Origin and geographic distribution** *Cissampelos* consists of 20–25 species and has a pantropical distribution.

Only the pantropical *C. pareira* var. *hirsuta* (Buch.-Ham. ex DC.) Forman occurs in Malesia. In Asia it is found from Nepal and India, through Burma (Myanmar), Indo-China, southern China, Thailand and Malesia (but is not known from Peninsular Malaysia, Sumatra and Java), to Australia (Queensland).

**Uses** Preparations of *C. pareira* have a large array of uses in tropical countries and are applied against a variety of complaints. For instance, the root decoction is used to treat complaints in the following organs:

- Urinary tract: diuretic (the Philippines, Thailand, India, Africa, Central and South America), acute and chronic cystitis (India), solvent of urinary calcifications (the Philippines, India, Africa, Madagascar, Central and South America).
- Gastro-intestinal tract: colic (antispasmodic action: Indo-China, South America), purgative (Africa, Madagascar), diarrhoea (Thailand, Africa), dysentery (India), emetic (Madagascar).
- Genital tract: emmenagogue (Thailand, Africa, Central and South America), painful menstruation and pre- and postnatal pains (antispasmodic action: South America), prevention of a threatened miscarriage (South America), to stop uterine haemorrhages (South America).
- In general: febrifuge (the Philippines, Thailand, India, Africa, Central and South America), pectoral (the Philippines), cough (Africa), blennorrhoea (Indo-China), heart trouble (India), rheumatism (Africa, Central and South America), jaundice (Thailand, Central and South America), treatment for snake bites (Africa, Central and South America), sores (externally, Thailand, India).

The (pounded) leaves are applied to snake bites (the Philippines), they are used to cure scabies (the Philippines), in the treatment of abscesses,

wounds and ulcers (Thailand, India, Africa, Central and South America) and as a stomachic (India). The plant is locally used by tribals in India in prevention of pregnancy.

The leaves, crushed in water, give a jelly which is used as a refreshment. In the Philippines, the fibres of the bark are made into ropes, which are used as a fish poison.

**Production and international trade** *C. pareira* is only used in local medicine and is not traded on the international market.

**Properties** *C. pareira* contains a number of alkaloids, especially bisbenzylisoquinoline alkaloids: hayatine (= d,l-bebeerine = d,l-curine), hayatidine (= d,l-4'-O-methylbebeerine = d,l-4'-O-methylcurine), hayatinine (= 1-4'-O-methylbebeerine = 1-4'-O-methylcurine), d-4'-O-methylbebeerine (= d-4'-O-methylcurine), l-bebeerine (= l-curine), d-isochondodendrine, d-dicentrine, d,l-dehydrodicentrine, d-insularine (all from roots), l-cycleanine (from roots and leaves) and cissampareine (from plants). Hayatinine (in the form of its methochloride) has been evaluated for its muscle-relaxant properties. The molecule is structurally very similar to that of d-tubocurarine from *Chondrodendron tomentosum* Ruiz & Pavón (*Menispermaceae*, South America) and also shows comparable neuromuscular blocking activities. Both have a non-depolarizing mode of action, and the blocking site is at the cholinergic (nicotinic) receptor in the post-synaptic membrane. Additionally, l-cycleanine has shown significant inhibition of nitric oxide production in vitro, and reduced the level of tumour necrosis factor in vivo, using a mouse model for fulminant hepatitis.

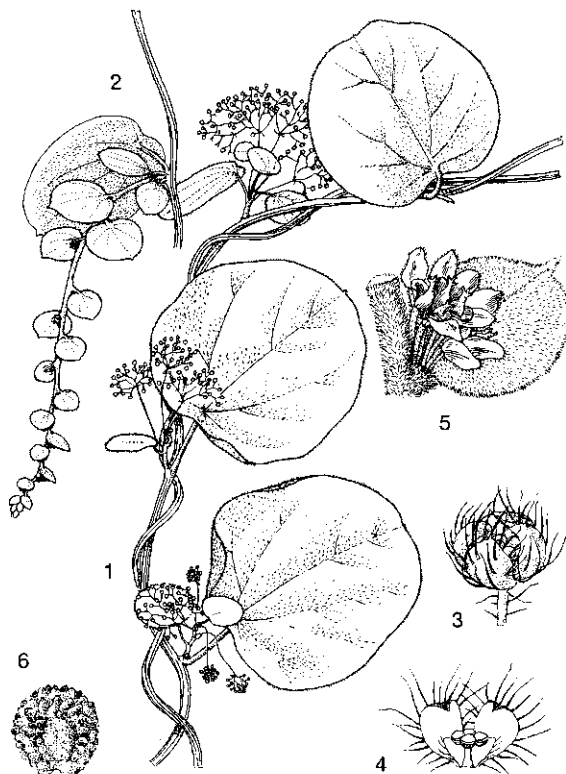
The roots have also been found to be a rich source of tropoloisoquinoline alkaloids. Using bioassay-directed purifications, guided by cytotoxicity against P388 cells, pareirubrine A, pareirubrine B, grandirubrine, isomerubrine and pareitropone have been isolated, all of which showed potent antileukemic activity. Furthermore, two cytotoxic azafluoranthene alkaloids, structurally strongly related to tropoloisoquinoline alkaloids, have been isolated from the same root extract. One of the tetrahydroprotoberberine group of alkaloids, cis-samine chloride (= cyclanoline chloride) has been isolated from the roots. *C. pareira* exhibits curare-like activity, depressing the central nervous systems, relaxing smooth muscles and with hypotensive and hypoglycaemic action.

In tests in Africa, extracts of *Cissampelos* roots and leaves controlled storage pest species, such as the small beetles *Acanthoscelides obtectus* on cow-

peas, *Prostephanus truncatus* on maize grains and *Sitophilus oryzae* on wheat grains.

**Adulterations and substitutes** Other *Menispermaceae* have similar or related alkaloids such as benzylisoquinolines, and similar applications. *Cyclea barbata* Miers is known to be used as a substitute.

**Description** A dioecious scandent shrub with woody older stems and slender leafy stems, glabrous to densely pubescent. Leaves arranged spirally, simple and entire, broadly ovate, 4.5–11 cm × 4.5–12 cm, with rounded, truncate or cordate base and acuminate to obtuse apex, mucronate at the tip, hairy below, sparsely pubescent above, palmately 5–7-veined; petiole 2–9 cm long, pubescent; stipules absent. Male inflorescence an axillary subcorymbose peduncled cyme, 2–4 cm long, solitary or a few together; female inflorescence axillary, thyrsoid, narrow, up to 18 cm long, composed of a pseudoraceme of fascicles with accrescent suborbicular bracts. Flowers unisexual, pedi-



*Cissampelos pareira* L. - 1, flowering stems of male plant; 2, flowering stem of female plant; 3, male flower; 4, sectioned male flower; 5, part of female inflorescence; 6, endocarp.



cel up to 2 mm long; male flower greenish or yellowish, with 4 sepals pilose outside, a cupuliform corolla and stamens completely fused, having 4 anther-cells; female flower with one sepal, one petal and one pilose carpel having a thick style with divaricately 3-lobed stigma. Fruit a pubescent, orange to red drupe, about 5 mm long, curved with style-scar near base; endocarp with 2 dorsal rows of very prominent transverse ridges. Seed horseshoe-shaped; embryo elongate, narrow, embedded in endosperm, cotyledons flattened.

**Growth and development** The flowers are probably pollinated by small insects such as flies and bees, and possibly small beetles and moths.

**Other botanical information** *C. pareira* has been subdivided into 2 varieties: var. *pareira* occurs in the West Indies, var. *hirsuta* (Buch.-Ham. ex DC.) Forman (synonyms: *C. pareira* L. var. *orbiculata* (DC.) Miq., *C. pareira* L. var. *peltata* Scheff., *C. pareira* L. var. *typica* Diels) is pantropical. *C. pareira* has been erroneously recorded from Peninsular Malaysia and Java. These records are based on misidentified specimens of *Pericampylus glaucus* (Lamk) Merr.

*C. owariensis* P. Beauv. ex DC. is cultivated in Africa as a medicinal plant.

**Ecology** *C. pareira* occurs in primary and secondary forest, in Thailand also in bamboo forest, and in thickets, up to 1300 m altitude. It climbs over trees and river banks.

**Genetic resources and breeding** *C. pareira* is very widespread and locally common (e.g. in the Philippines, Vietnam, Cambodia, Laos and Thailand). There is no reason to suppose any danger of genetic erosion. The quantity and composition of the alkaloids found in the roots seem to differ between accessions from different regions of the extremely large area of distribution. Although it cannot be excluded that this is partly due to misidentifications, it is possibly a result of great genetic diversity.

**Prospects** The alkaloids present in *C. pareira* have interesting properties, e.g. antileukaemic and neuromuscular blocking activity. The roots are used in traditional medicine in different parts of the world for similar purposes, which seems to confirm their effectiveness.

**Literature** [1] Brown, W.H., 1951. Useful plants of the Philippines. Reprint of the 1941–1943 edition. Vol. 1. Technical Bulletin 10. Department of Agriculture and Natural Resources. Bureau of Printing, Manila, the Philippines. p. 531. [2] Forman, L.L., 1986. Menispermaceae. In: van Stee-

nis, C.G.G.J. & de Wilde, W.J.J.O. (Editors): Flora Malesiana. Ser. 1, Vol. 10. Kluwer Academic Publishers, Dordrecht, Boston, London. pp. 234–236. [3] Forman, L.L., 1991. Menispermaceae. In: Smitinand, T. & Larsen, K. (Editors): Flora of Thailand. Vol. 5(3). The Forest Herbarium, Royal Forest Department, Bangkok. pp. 323–325. [4] Hegnauer, R., 1969. Chemotaxonomie der Pflanzen [Chemotaxonomy of plants]. Vol. 5. Birkhäuser Verlag, Basel, Boston, Stuttgart. p. 81. [5] Morita, H., Matsumoto, K., Takeya, K. & Itokawa, H., 1993. Azafluoranthene alkaloids from *Cissampelos pareira*. Chemical and Pharmaceutical Bulletin 41(7): 1307–1308. [6] Morita, H., Matsumoto, K., Takeya, K. & Itokawa, H., 1993. Conformation of tropolone ring in antileukemic tropoloisoquinoline alkaloids. Chemical and Pharmaceutical Bulletin 41(8): 1478–1480. [7] Morita, H., Matsumoto, K., Takeya, K., Itokawa, H. & Itaka, Y., 1993. Structures and solid state tautomeric forms of two novel antileukemic tropoloisoquinoline alkaloids, pareirubrines A and B, from *Cissampelos pareira*. Chemical and Pharmaceutical Bulletin 41(8): 1418–1422. [8] Nguyen Van Duong, 1993. Medicinal plants of Vietnam, Cambodia and Laos. Mekong Printing, Santa Ana, California, United States. pp. 257–258. [9] Niber, B.T., Helenius, J. & Varis, A.L., 1992. Toxicity of plant extracts to three storage beetles (Coleoptera). Journal of Applied Entomology 113(2): 202–208. [10] Quisumbing, E., 1978. Medicinal plants of the Philippines. Katha Publishing Co., Quezon City, the Philippines. pp. 294–296.

**Other selected sources** 979, 1554.

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### **Curculigo orchoides Gaertner**

Fruct. sem. pl. 1: 63 (1788).

HYPOXIDACEAE

$2n = 18, 36$

**Vernacular names** Papua New Guinea: tupaui (Kenemote, Eastern Highlands). Philippines: taloangi (Bagobo), tataluangi (Bukidnon), sulsulitik (Bontok). Thailand: waan phraao (northern). Vietnam: ng[ar]i cau, s[aa]m cau, ti[ee]n mao.

**Origin and geographic distribution** *C. orchoides* occurs from the subtropical Himalayas of Pakistan and India, to Cambodia, Vietnam and Laos, southern China, Taiwan, southern Japan, to Thailand and Malesia (at least known with certainty from Java and the Philippines), and possibly also to northern and eastern Australia. The

distribution in Malesia is very incompletely known.

**Uses** A decoction of the powdered rhizomes ('Curculiginis Rhizoma') is used in Chinese traditional medicine as a general tonic and analeptic in the treatment of decline (especially of physical strength). In the Philippines, Nepal and India, the rhizome is used as diuretic and aphrodisiac, and to cure skin diseases (externally), peptic ulcers, piles, gonorrhoea, leucorrhoea, asthma, jaundice, diarrhoea and headache. In Thailand, the rhizome is used as a diuretic and to treat diarrhoea. In Papua New Guinea, the rhizome and leaves are softened by being heated over a fire, before being rubbed on the body to serve as a contraceptive. In China, additional reported indications include the treatment of lumbago, arthritis, chronic nephritis, hypertension and the use as an emmenagogue, and in India, *C. orchoides* is used to induce abortion. Powdered rhizomes are normally used in decoction, but are also sometimes given with an equal quantity of sugar in a glass of milk. It is reported that the rhizomes are also used to produce flour in India.

**Production and international trade** Sliced and dried rhizomes of *C. orchoides* are traded in small quantities in local markets in China and Indo-China.

**Properties** The rhizome tastes slightly bitter and is mucilagenous. The alcoholic extract from the rhizome is reported to have adaptogenic, anti-inflammatory, anticonvulsive, sedative, androgenic and immuno-stimulating activity. The water extract of the rhizomes exhibits andrenergic receptor blocking ( $\alpha$  2), cholecystokin receptor binding, hypoxanthine-guanine phosphoribosyltransferase inhibition and uterine activating activity. Swelling of the tongue has been reported as a side-effect after drinking a decoction from the rhizome; in China the recommended antidote is a decoction of *Rheum tanguticum* Maxim. ex Balf. with sodium sulphate.

A series of 10 triterpenoidal saponins (curculigosaponins A-J) have been isolated from the rhizomes. All these compounds have curculigenine A ( $3\beta,11\alpha,16\beta$ -trihydroxycycloartane-24-one) as the aglycone. Pharmacological studies have shown that curculigosaponins C and F can promote the proliferation of spleen lymphocytes in mice very significantly, and that curculigosaponins F and G increase the weight of the thymus in vivo in mice. The triterpene alcohol, curculigol (24-methylcycloart-7-en- $3\beta,20$ -diol), whose structure is very similar to curculigenine A, has also been isolated.

Four phenolic glycosides have been isolated and identified: curculigoside, orcinol glycoside, curculigine A and corchioside A. Curculigoside from the rhizomes exerts immunological and protective effects. It has been found a characteristic constituent of 'Curculiginis Rhizoma', and a quantitative determination method using HPLC has been developed. The determination was performed indirectly by measuring the content of 2,6-dimethoxybenzoic acid, the hydrolysis product of curculigoside. Using this method, an average content of 0.2% curculigoside has been found in rhizomes from China.

Several aliphatic hydroxy-ketones (e.g. 27-hydroxytriacontan-6-one) have been reported from *C. orchoides*. The powdered rhizomes furthermore contain approximately 8% water, 4% alcohol-extractable matter, 1.5% ether-extractable matter, 15% crude fibre, 20% mucilage and 8.5% ash.

From the fruits of *Curculigo latifolia* Dryander, which grows wild in western Malaysia, 114 amino acids containing the peptide curculin were isolated. Curculin itself elicits a sweet taste (550 times sweeter than sucrose on a weight basis), which disappears rather rapidly after holding it in the mouth. Tasting a lemon (or ascorbic, citric or hydrochloric acid) afterwards then elicits a sweet, orange-like taste. This taste-modifying sensation lasts for about 10 minutes.

**Description** A perennial herb up to 50 cm tall, with vertical, more or less tuberous, blackish rhizome and rather stout roots. Leaves alternate, clustered and sessile on rhizome, narrowly lanceolate, 20-30 cm  $\times$  1-2 cm, long-tapering at base into a pseudo-petiole which is sheath-like at its base, and also long-tapering at apex, plicate, sparsely pilose with long hairs or glabrous, with few to several parallel veins. Inflorescence axillary, inconspicuous among the leaf-bases, spike-like, few-flowered or with a solitary flower, and with a very short scape or peduncle; bracts lanceolate, spatheaceous, 2-4 cm long, membranous, surpassing the peduncle and ovary. Flowers long-pilose, lower ones in the inflorescence bisexual, upper ones male; perianth with long slender tube 2-3 cm long (resembling a pedicel) and 6 equal, spreading lobes which are lanceolate to elliptical, 5-8 mm long, few-veined, pale outside and bright yellow inside; stamens 6, inserted on bases of perianth lobes, about half as long as perianth lobes, with short filaments attached to the bases of the linear anthers; ovary inferior, 3-locular, locules imperfect, style short and thick, with 3 stigmas. Fruit berry-like, rather fleshy, ellipsoid, about 1.5 cm in



*Curculigo orchioides* Gaertner – 1, flowering plant; 2, rhizome; 3, flower; 4, fruit; 5, seed.

diameter, surpassed by the bract, beaked by the persistent perianth tube, 1-4-seeded. Seeds subglobose to oblong, about 4 mm long, with beak (elaiosome) lateral to hilum; testa crustaceous, striate, black and shiny.

**Growth and development** Rhizomes of *Curculigo orchioides* may reach 30 cm × 11.5 cm. Only 3-5 leaves are found on the plant at a given time. The flowers and fruits are inconspicuous because they are close to the ground and partially covered by the bracts and leaves.

**Other botanical information** *C. orchioides* belongs to a genus of approximately 10 species with pantropical distribution. *Curculigo* has been variously included in *Amaryllidaceae* and *Liliaceae*, but is nowadays usually considered as belonging to the comparatively small family *Hypoxidaceae* with about 10 genera.

*C. ensifolia* R. Br., a species recorded for Australia, is possibly conspecific with *C. orchioides*.

**Ecology** *C. orchioides* occurs in open fields and grasslands. In Java it grows on periodically very dry, sunny or slightly shaded localities in grass-

lands and teak forest up to 400 m altitude. In the Philippines it is also found in grasslands, often dominated by *Imperata*.

**Propagation and planting** There is no information about tests on propagation of *C. orchioides*, but the method described for *C. latifolia* may be applicable. In trials with tissue culture of *C. latifolia* for propagation for ornamental purposes, cultures of rhizomes showed the best results. A half-strength Murashige and Skoog medium was used, supplemented with sucrose (30 g/l), thiamine (0.4 g/l), coconut water (150 ml/l), kinetin (5 mg/l) and indole-acetic acid (2.5 mg/l). About 90% of the plants potted out survived.

**Diseases and pests** In India, rust (*Puccinia hypoxidis*) is reported from *C. orchioides*.

**Harvesting** *C. orchioides* is not planted and rhizomes are collected from the wild.

**Handling after harvest** The rhizomes are washed, freed from roots, and sliced; the slices are dried in the shade. Usually the dried slices are powdered, and small amounts of powder are mixed in a glass of milk with sugar or used to prepare a decoction for drinking.

**Genetic resources and breeding** *C. orchioides* has been recorded amongst the rare and endangered ethno-medical plants in India. However, it has a large area of distribution, is locally rather common, is by no means restricted to endangered vegetation types, and therefore does not seem endangered or liable to genetic erosion.

**Prospects** *C. orchioides* may prove a valuable medicinal plant because its active compounds are known and comparatively well documented and a suitable method has been developed for quantifying one of them (curculigoside), which is important for quality control of the drug. However, more research is needed and appropriate cropping techniques should be developed so that *C. orchioides* can be cultivated.

**Literature** |1| Backer, C.A. & Bakhuizen van den Brink Jr., R.C., 1968. Flora of Java. Vol. 3. Wolters-Noordhoff, Groningen, the Netherlands. p. 209. |2| Bhaskaran, K. & Padmanabhan, D., 1983. Leaf development in *Curculigo orchioides*. *Phytomorphology* 31: 1-10. |3| Chee Len, L.-H., 1981. Tissue culture of *Curculigo latifolia* Dry. ex W.T. Ait. *Gardens' Bulletin Singapore* 34(2): 203-208. |4| Misra, T.N., Singh, R.S., Tripathi, D.M. & Sharma, S.C., 1990. Curculigol, a cycloartane triterpene alcohol from *Curculigo orchioides*. *Phytochemistry* 29(3): 929-931. |5| Nasir, E., 1980. *Amaryllidaceae*. In: Nasir, E. & Ali, S.I. (Editors): *Flora of Pakistan*. No 134. Department of

Botany, University of Karachi and National Herbarium, Pakistan Agricultural Research Council, Islamabad, Pakistan. pp. 4–5. |6| Nguyen Van Duong, 1993. Medicinal plants of Vietnam, Cambodia and Laos. Mekong Printing, Sata Ana, California, United States. pp. 44–45. |7| Quisumbing, E., 1978. Medicinal plants of the Philippines. Katha Publishing Co., Quezon City, the Philippines. pp. 172–174. |8| Xu, J.-P., Xu, R.-S. & Li, X.-Y., 1992. Four new cycloartane saponins from *Curculigo orchoides*. *Planta Medica* 58: 208–210. |9| Xu, J.-P., Xu, R.-S. & Li, X.-Y., 1992. Glycosides of a cycloartane saponin from *Curculigo orchoides*. *Phytochemistry* 31(1): 233–236. |10| Yamasaki, K., Hashimoto, A., Kokusenya, Y., Miyamoto, T., Matsuo, M. & Sato, T., 1994. Determination of curculigoside in *Curculiginis Rhizoma* by high performance liquid chromatography. *Chemical and Pharmaceutical Bulletin* 42(2): 395–397.

**Other selected sources** 190, 552, 597, 622, 895, 954, 1004, 1539, 1549.

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## Curcuma L.

Sp. pl. 1: 2 (1753); Gen. pl. ed. 5: 3 (1754).

ZINGIBERACEAE

$x = 16, 21$ ; *C. aurantiaca*:  $2n = 42$ , *C. longa*:  $2n = 32, 62-64$ , *C. petiolata*:  $2n = 64$ , *C. xanthorrhiza*:  $2n = 63$ , *C. zedoaria*:  $2n = 63, 64, 66$

**Major species** *Curcuma longa* L., *C. xanthorrhiza* Roxb., *C. zedoaria* (Christm.) Roscoe.

**Vernacular names** *Curcuma* (En, Fr), turmeric (En). Indonesia: temu. Malaysia: temu. Laos: kachièw, khminz. Thailand: khamin. Vietnam: ngh[eej].

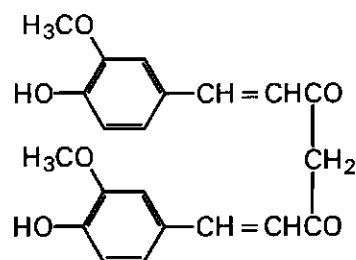
**Origin and geographic distribution** *Curcuma* comprises some 40–50 species and is native to the Indo-Malesian region, from India to Indo-China, Taiwan and Thailand, throughout Malesia, towards the Pacific and northern Australia. Some 20 species are present within Malesia. Several species have been introduced elsewhere in tropical and subtropical areas. The centre of diversity is located in India.

**Uses** Rhizomes of many *Curcuma* species are used medicinally, often to treat liver diseases (jaundice, gallstones), but also for various abdominal complaints. They are considered stomachic, carminative, haematic and styptic. Furthermore, they are sometimes applied to asthma, cough and bronchial catarrh, or to treat itch, scurf, skin in-

fections in general, or are applied to wounds and ulcers. The rhizomes of several *Curcuma* species are a well-known source of spice (turmeric), of starch and of a yellow-orange dye. The latter may be used in colouring clothing and food as such, or in the preparation of other dyes.

**Production and international trade** Annually about 20 000 t of cured dried whole rhizomes of turmeric (*C. longa*) enter into international trade. India is the largest producer with 400 000 t from 130 000 ha and dominates the international trade. Within Malesia, Indonesia is a major producer. All Asian producers are heavy consumers as well and some are even net importers. No information is available, except on *C. longa*.

**Properties** All *Curcuma* species are rich in essential oils. On distillation, rhizomes of *C. longa* yield 1.3–5.5% essential oil whose main constituents are sesquiterpenes called turmerones (about 60%, e.g. ar-turmerone,  $\alpha$ -,  $\beta$ -turmerone) and the sesquiterpene zingiberene (about 25%). Dried rhizomes of *C. xanthorrhiza* contain on average 3.8% essential oil with ar-curcumene, xanthorrhizol,  $\alpha$ -,  $\beta$ -curcumene and germacrene as major constituents. The compounds cyclo-isoprenemyrcene and p-tolylmethylcarbinol which are often mentioned as essential oil constituents in older literature are artifacts which originate from the distillation process and fractionation of oils at higher temperatures. The phenolic sesquiterpene xanthorrhizol is species specific: its presence can thus be used to distinguish *C. xanthorrhiza* from e.g. *C. longa*. In young rhizomes the essential oil content may be higher; 29.5% essential oil has been found in rhizomes that are just beginning to develop. Dried rhizomes of *C. zedoaria* contain about 1.5% essential oil with cineol, borneol, d-camphor, camphene and d- $\alpha$ -pinen as main constituents. A further group of constituents in the rhizomes of *Curcuma* species are the curcuminoids. Curcuminoids are referred to as curcumin (diferuloyl methane or curcumin I) and its deriva-



curcumin I

tives desmethoxy-curcumin (feruloyl-p-hydroxycinnamoyl methane or curcumin II) and bis-desmethoxy-curcumin (bis-(p-hydroxycinnamoyl)-methane or curcumin III). The name curcumin is also often used for the complex yellow-orange mixture of curcuminoids as a whole, isolated from the plant. Curcumin has some broad-spectrum antimicrobial activity, but therapeutic utility of *C. longa* for this indication has not been recognized. Curcumin I derived from *C. longa* inhibited the 5-lipoxygenase activity in rat peritoneal neutrophils as well as the 12-lipoxygenase and the cyclooxygenase activities in human platelets. In a cell-free peroxidation system, curcumin exerted strong antioxidative activity. Thus, its effects on the dioxygenases are probably due to its reducing capacity. Oral administration of the antioxidant curcumin, from *C. longa*, at a concentration of 200  $\mu\text{mol/kg}$  body weight significantly reduced the lung collagen hydroxyproline in whole-body  $\gamma$ -irradiated rats. Serum lipid and liver lipid peroxidation, which were increased by irradiation, were reduced significantly by the antioxidant treatment. The increased frequency of micronucleated polychromatic erythrocytes after whole-body irradiation of mice was significantly reduced by antioxidant treatment. The actual quantity of the three known curcuminoids, which in fact are all potent antioxidants, does not fully explain the antioxidant activity of the extracts of several *Curcuma* species. Three non-phenolic diarylheptanoids isolated from *C. xanthorrhiza* have been identified as trans,trans-1,7-diphenyl-1,3-heptadien-4-one (alnustone), trans-1,7-diphenyl-1,3-hepten-5-ol, and trans,trans-1,7-diphenyl-1,3-heptadien-5-ol. They all exerted significant anti-inflammatory activity in the carrageenin-induced hind paw oedema assay in rats. 1E,3E,1,7-Diphenylheptadien-5-one exerted potent anti-inflammatory activity ( $\text{ID}_{50}$  value of 67  $\mu\text{g/ear}$ , topically applied) in an ethylphenylpropiolate-induced ear oedema model in rats. The  $\text{ED}_{50}$  value of the water extract of *C. longa* after intraperitoneal administration was 4.7 mg/kg in carrageenin-induced rat paw oedema, that of the alcoholic extract 307 mg/kg, of the petroleum ether extract 40.7 mg/kg, of the sodium curcuminat 2.1 mg/kg and that of curcumin 8.7 mg/kg, but an  $\text{ED}_{50}$  of only 0.36 mg/kg has also been reported; the oral  $\text{ED}_{50}$  of curcumin was 100.2 mg/kg in mice and 48.0 mg/kg in rats. The mechanism by which curcumin inhibits inflammation is still poorly understood. Curcumin has been found to inhibit several types of phospholipases, notably phospholipase D, and it also inhibited

phospholipase D activation induced by 12-0-tetradecanoylphorbol-13-acetate. This suggests that the anti-inflammatory and anti-carcinogenic action of curcumin is partly due to the inhibition of phospholipase D and prostaglandin synthesis. The aqueous extracts of the crude drug of *C. aeruginosa* showed significant protective effects against  $\text{CCl}_4$ -induced liver injury in rats and D-galactosamine/lipopolysaccharide-induced liver injury in mice. The aqueous extract of the rhizomes of *C. xanthorrhiza* significantly reduced the acute elevation of serum glutamate oxaloacetate transaminase (alanine aminotransferase) and serum glutamate pyruvate transaminase (aspartate aminotransferase) induced by paracetamol or  $\text{CCl}_4$  in mice; it alleviated the degree of liver damage 24 hours after intraperitoneal administration of the hepatotoxic compounds. When the aqueous extract of rhizomes of *C. xanthorrhiza* (100 mg/kg by oral administration) was investigated in rats treated with  $\beta$ -D-galactosamine (288 mg/kg, intraperitoneally administered), the extract reduced the elevated concentrations of alanine aminotransferase and aspartate aminotransferase, and reduced histopathological changes induced 24 hours after administration. Antihepatotoxic activity of the rhizome of *C. longa* has also been reported.

Active principles other than curcuminoids from *C. xanthorrhiza* can modify the metabolism of lipids and lipoproteins.  $\alpha$ -Curcumene which can make up to about 65% of the essential oil of *C. xanthorrhiza* was shown to lower triglyceride levels in rats.

Streptozotocin-induced diabetic rats maintained for 8 weeks on a diet containing 0.5% curcumin showed a significant decrease in blood triglyceride and phospholipid levels, as do diabetic animals maintained on a high cholesterol diet, where hypercholesterolaemia and phospholipidemia is even more severe. In streptozotocin-induced diabetic rats which were fed on purified diets containing 5% *C. xanthorrhiza* the diabetic symptoms improved. This diet specifically modified the amount and composition of faecal bile acids. The essential oil of *C. longa* and *C. xanthorrhiza* and its component d-camphor caused a persistent increase of bile secretion in anaesthetized rats. p-Tolylmethylcarbinol activates the secretion of bile from the gall bladder and curcumin causes rhythmic contractions of the gall bladder. Furthermore, the liquid balm of *C. xanthorrhiza* was found to lower the total cholesterol content and the total lipid content in rabbits.

Furanogermenone and (4S,5S)-(+)-germacrone-4,5-epoxide had a potent preventive effect against stress ulceration. Moreover, oral administration of fractions of the methanol extract significantly inhibited the formation of both HCl-induced and indomethacin-induced gastric ulcers.

Water extracts of *C. zedoaria* inhibited the growth of mouse L5178Y leukaemia cells in a dose-dependent manner; inhibitory effects of alcohol extracts were not significant. The bisabolane sesquiterpenoids  $\alpha$ -curcumene, ar-turmerone,  $\beta$ -atlantone and xanthorrhizol were isolated as major antitumour constituents from the rhizomes of *C. xanthorrhiza*. Curcumin from *C. longa* inhibited the growth of hormone-dependent and -independent, and multi-drug resistant human breast tumour cell lines in a time- and dose-dependent way. The effect was correlated with the compound's inhibition of ornithine decarboxylase activity. A 5% turmeric diet significantly inhibited the tumour burden and tumour incidence in fore-stomach tumours induced by benzo-[a]-pyrene in Swiss mice and oral mucosal tumours induced by methyl-(acetoxymethyl)-nitrosamine in Syrian golden hamsters. Curcumin isolated from *C. longa* inhibited human colon cancer cell proliferation in vitro, mainly by causing cells to accumulate in the G2/M phase; this effect is independent of its ability to inhibit prostaglandin synthesis. In an assay studying the promotion of tumours induced by croton oil, 90% of the control animals had papillomas in the 10th week of tumour initiation, compared with only 10% of animals treated with curcumin III, 20% of the animals treated with curcumin II, and 40% of animals treated with curcumin I. Of the synthetic curcuminoids studied, salicylcurcuminoid, which had caused no papillomas by the 10th week, was the most potent anti-carcinogen. In vitro and in vivo tests with aqueous and ethanolic extracts of *C. longa* and *C. xanthorrhiza* showed marked antitumour activity. Topical application of curcumin together with 5 nmol of the tumour-promoting agent 12-O-tetradecanoylphorbol 13-acetate (TPA) twice weekly inhibited the number of TPA-induced tumours by 39% at a dose of 1  $\mu$ mol, 77% at a dose of 3  $\mu$ mol, and 98% at a dose of 10  $\mu$ mol.

When the antispasmodic activity of *C. longa* was tested using the isolated guinea-pig ileum, the ED<sub>50</sub> values of sodium curcuminates were 30.2  $\mu$ g/ml (nicotine), 77.2  $\mu$ g/ml (acetylcholine), 82.8  $\mu$ g/ml (5-hydroxytryptamine), 81.8  $\mu$ g/ml (histamine) and 171.4  $\mu$ g/ml (barium chloride), respectively. Concentrations of this magnitude will not

appear in the blood, because if curcumin is absorbed in the blood at all, it is rapidly metabolized in the liver and excreted through the bile.

ar-Turmerone isolated from *C. longa* is a potent antidote to snake bite. Furthermore, curcumin has a median inhibitory concentration (IC<sub>50</sub>) for strand transfer of 40  $\mu$ M on purified human immunodeficiency virus type 1 (HIV-1) integrase.

In a clinical study on turmeric, significant improvement was observed in patients with rheumatoid arthritis or with respiratory diseases. Limited clinical trials on the effects of orally administered turmeric on peptic ulcers showed promising results. Another clinical trial carried out in Thailand showed good results on dyspepsia.

The inhibitory activity of turmeric oil and curcumin isolated from *C. longa* was tested in *Trichophyton rubrum*-induced dermatophytosis in guinea-pigs. Turmeric oil at dilutions of 1:40–1:320 inhibited the dermatophytes. At dilutions of 1:40–1:80 it inhibited 4 isolates of pathogenic fungi. Curcumin had no inhibitory effect on either dermatophytes or pathogenic fungi. The essential oil from turmeric showed a moderate antibacterial activity against *Escherichia coli*.

Curcumin from *C. longa* has demonstrated phototoxicity to several species of bacteria and to mammalian cells, using a rat basophilic leukaemia cell model, under aerobic conditions. The rhizome powders of *C. longa* and *C. zedoaria* applied on *Cajanus cajan* (L.) Millsp. and *Vigna radiata* (L.) Wilczek seed before being stored were moderately effective to effective against the pulse beetle *Callosobruchus chinensis*; the LD<sub>50</sub> of the extract of *C. longa* was 0.05–0.1 ppm. Insecticidal as well as antifungal activity of *Curcuma* species has also been reported. The extract of the rhizome of *C. zedoaria* showed significant antifungal activity against *Cladosporium cladosporioides*.

In a contact residue bioassay the most active sesquiterpenoids xanthorrhizol and furanodienone showed pronounced toxicity against neonate larvae of *Spodoptera littoralis*. The LD<sub>50</sub> of xanthorrhizol following topical application was found to vary between 6.92 and 8.13  $\mu$ M/kg fresh weight irrespective of the larval stages studied. Xanthorrhizol, however, did not cause significant mortality of neonate larvae when incorporated into artificial diet, suggesting that the compounds are inactivated in the larval gut. The chloroform extract of *C. longa* proved economically useful in the treatment of *Trichophyton verrucosum* ringworm in cattle.

**Description** Perennial, rhizomatous, erect

herbs with spurious stems; subterranean parts fleshy, aromatic, roots fleshy, often bearing ellipsoidal tubers. Leafy shoots bearing bladeless sheaths forming a spurious stem on which less than 10, distichous, pinnately veined leaves develop; petiole well developed; ligule narrow. Inflorescence either terminal on a leafy shoot or on a separate shoot, spike-like, cylindrical; peduncle well developed; spike with large bracts which are joined for about half their length, forming pouches from which a cincinnus of 2-7(-10) flowers arises, uppermost bracts often larger and forming the 'coma'. Flowers bisexual, zygomorphic; bracteoles thin, not connate, enclosing the flower bud; calyx tubular, split unilaterally, about half as long as the corolla, unequally toothed; corolla tube united with the staminal tube, cylindrical below, cup-shaped above, lobes 3, translucent white or pink to purplish, the dorsal lobe hooded and ending in a hollow hairy point; staminodes 3, petaloid, the anterior one, called labellum, obovate, with a thickened median band, the 2 lateral ones elliptical-oblong, their inner edges folded under the hood of the dorsal corolla lobe; fertile stamen 1, filament short, broad, anther versatile, thecae parallel, often spurred at base, connective sometimes enlarged at the apex into a small crest; ovary inferior, 3-locular with axillary placentation and many ovules, style 1, filiform with a cup-shaped, 2-lobed stigma, held between the thecae. Fruit an ellipsoid capsule, crowned by the calyx remnants; pericarp thin, irregularly dehiscent. Seeds embedded in mucilage, ellipsoidal, with a lacerate aril of few segments free to the base.

**Growth and development** The primary rhizome of *Curcuma* is at first surrounded by small scales, that remain visible by their annular scars. Secondary and tertiary rhizomes develop from axillary buds of the primary and secondary rhizome, respectively. In Java, most *Curcuma* species flower in September-February(-March) and June-August; *C. aeruginosa*, *C. purpurascens* and *C. xanthorrhiza* flower almost throughout the year. Flowers generally open late in the afternoon and wither before the next morning. Although flowering is abundant and flowers have been observed to be frequently visited by insects searching for pollen, in Java only *C. aurantiaca* forms fruits. This is probably because it is diploid whereas virtually all other *Curcuma* are triploid. In Java, no dormancy period was observed after flowering, but this phenomenon has been reported from northern India, where it usually occurs during winter. Vesicular-arbuscular mycorrhizae have

been observed in *C. longa* and *C. zedoaria*; many of the sporulating mycorrhizae belong to the genus *Glomus*.

**Other botanical information** *Curcuma* belongs to the tribe *Hedychieae* and is characterized by the partly fused bracts. Its taxonomy is still unsatisfactory, the various species being very closely related and sometimes doubtfully distinct. Intensive cultivation, and possibly hybridization, makes it difficult to distinguish species and a thorough revision is badly needed. The fact that many 'species' appear to be triploids may be an indication of an origin from cultivation. Although the taxonomic treatment of the Flora of Java is not followed here, the rigorous lumping of species may eventually be justified. As *C. aurantiaca* is the only Javanese species producing fruit, it may indeed be the only indigenous species of Java. Two subgenera are distinguished: *Curcuma* with elongated rhizomes, a non-auriculate ligule, conspicuous coma bracts and longitudinally grooved and folded staminodes, and *Paracurcuma* with short rhizomes, an auriculate ligule, inconspicuous coma bracts and straight staminodes.

**Ecology** In the wild, most *Curcuma* are found in the undergrowth of tropical or subtropical forests or slightly shaded places such as forest margins and plantations, up to 1150 m altitude, but in the Himalayan foothills up to 2000 m. They grow best on well-drained, loamy or alluvial, fertile, friable soils and cannot stand waterlogging. They often occur in deciduous monsoon forest, in Java especially in teak forest, in areas with an annual rainfall of about 1000-2000 mm, exceptionally up to 4000 mm.

**Propagation and planting** Propagation of *Curcuma* is by mother (primary) rhizomes, cut mother rhizomes or by finger rhizomes (also referred to as daughter or lateral rhizomes). Seed rhizomes need to be stored for 2-3 months prior to planting. Finger rhizomes of *C. longa* store better, are more tolerant to wet soil conditions and can be planted at a lower rate. *C. longa* is planted in ridges at 30-40 cm distance or in flat beds usually at a spacing of 25 cm, although good results have been obtained using a spacing of 15 cm. *C. xanthorrhiza* is planted at a distance of 60 cm and *C. zedoaria* at 25-45 cm. *Curcuma* is best planted under partial shade and in soils that have been ploughed or turned over to a depth of 30 cm.

**Husbandry** Mulching increases resprouting of the *Curcuma* rhizomes and rhizome yield and should be done at planting and 2 months thereafter. *Curcuma* requires heavy manuring to ob-

tain a high yield; about 25 t/ha of manure is usually recommended. Recommendations for fertilization vary widely between locations.

**Diseases and pests** Leaf spot or leaf blotch caused by *Taphrina malucans* and rhizome rot caused by *Pythium aphanidermatum* are considered the most important diseases of turmeric. Bacterial wilt caused by *Pseudomonas solanacearum* has been found killing *C. mangga* in Java.

**Harvesting** *Curcuma* propagated by whole mother rhizomes can be harvested after 8–12 months; if propagated from cut mother rhizomes or finger rhizomes, plants can be lifted after 2 years. Rhizomes of *C. longa* begin to develop about 5 months after planting and can be lifted after 7–10 months when the lower leaves turn yellow. At harvesting, care should be taken not to damage the rhizomes; the finger rhizomes are separated from the mother rhizomes.

**Yield** Yields of *Curcuma* are 17–23 t/ha when irrigated. Under rainfed conditions 6.5–9 t/ha are obtained for *C. longa*, 20 t/ha for *C. xanthorrhiza* and 7.5–12 t/ha for *C. zedoaria*.

**Handling after harvest** Whole rhizomes of *Curcuma* are dried, or first cut in slices and then dried.

**Genetic resources and breeding** A germplasm collection of about 500–600 *C. longa* accessions is maintained in India, but crop improvement of turmeric is limited. Neither germplasm collections nor breeding activities are known to exist for the other *Curcuma* species.

**Prospects** Many *Curcuma* species are planted in home gardens by small farmers. Hardly any efforts have been undertaken in agronomy, plant breeding and pest management, not even in the well-known spice *C. longa*, to improve performance. *C. longa* and *C. xanthorrhiza* show promising prospects for medicinal applications but research is needed to establish their therapeutic value.

**Literature** [1] Ammon, H.P.T. & Wahl, M.A., 1991. Pharmacology of *Curcuma longa*. *Planta Medica* 57(1): 1–7. [2] Anto, R.J., George, J., Babu, K.V., Rajasekaran, K.N. & Kuttan, R., 1996. Antimutagenic and anti carcinogenic activity of natural and synthetic curcuminoids. *Mutation Research* 370(2): 127–131. [3] Dahal, K.R. & Salma Idris, in prep. *Curcuma longa* L. In: De Guzman, C.C. & Siemonsma, J.S. (Editors): *Plant Resources of South-East Asia 13. Spices*. [4] Halijah Ibrahim & Jansen, P.C.M., 1996. *Curcuma Roxburgh*, *Curcuma xanthorrhiza Roxburgh*, *Curcuma zedoaria* (Christmann) Roscoe. In: Flach, M.

& Rumawas, F. (Editors): *Plant Resources of South-East Asia No 9. Plants yielding non-seed carbohydrates*. Backhuys Publishers, Leiden, the Netherlands. pp. 72–78. [5] Hikino, H., 1985. Anti-hepatotoxic activity of crude drugs. *Yakugaku Zasshi* 105(2): 109–118. [6] Lin, S.C., Lin, C.C., Lin, Y.H., Supriyatna, S. & Teng, C.W., 1995. Protective and therapeutic effects of *Curcuma xanthorrhiza* on hepatotoxin-induced liver damage. *American Journal of Chinese Medicine* 23(3–4): 243–254. [7] Lin, S.C., Teng, C.W., Lin, C.C., Lin, Y.H. & Supriyatna, S., 1996. Protective and therapeutic effect of the Indonesian medicinal herb *Curcuma xanthorrhiza* on  $\beta$ -D-galactosamine-induced liver damage. *Phytotherapy Research* 10(2): 131–135. [8] Masuda, T., Isobe, J., Jitoe, A. & Nakatani, N., 1992. Antioxidative curcuminoids from rhizomes of *Curcuma xanthorrhiza*. *Phytochemistry* 31(10): 3645–3647. [9] Prucksumand, C., Indrasukesri, B., Leethochawalit, H., Nilvises, N., Prijavudhi, A. & Wimolwattanapun, S., 1986. Effect of the long turmeric (*Curcuma longa* Linn.) on healing of peptic ulcer: a preliminary report of 10 cases. *Thai Journal of Pharmacology* 8(3): 139–151. [10] Thamlikitkul, V. et al., 1989. Randomized double blind study of *Curcuma domestica* Val. for dyspepsia. *Journal of the Medical Association of Thailand* 72(11): 613–620.

#### *Selection of species*

#### ***Curcuma aeruginosa* Roxb.**

Asiat. Res. 11: 335 (1810).

**Vernacular names** Indonesia: temu hitam (general), temu ireng (Javanese, Balinese), ko-neng hideung (Sundanese). Malaysia: temu erang, temu hitam (Peninsular). Thailand: waan ma-haamek (central). Vietnam: ngh[eej] ten d[ooof]ng.

**Distribution** Burma (Myanmar), Vietnam, Cambodia, Thailand, Peninsular Malaysia, Sumatra and Java; cultivated at least in Vietnam, Cambodia, Java and the Moluccas.

**Uses** In Indo-China rhizomes are used as a medicine for colic. In Peninsular Malaysia they have been prescribed to treat asthma and cough, applied externally to scurvy, and suggested as an application for mental derangement. In Indonesia and Thailand, rhizomes are the chief ingredient of a decoction given to women after childbirth to accelerate the lochia. They are considered to be depurative and used both internally and externally for treating exanthema, and as a poultice to treat itch. Other medicinal applications are



against obesity, rheumatism, and as an anthelmintic. During periods of famine the starch extracted from the rhizomes is used as a substitute for cassava or maize. A dye can be obtained from the rhizome.

**Observations** A herb with rhizome up to 16 cm long and 3 cm thick, outside grey and shiny, tips pink, inside bluish or blue-green with white cortex; leaf sheaths to 50 cm long, blades elliptical to oblong-lanceolate, 30–80 cm × 9–20 cm, green with wide purplish-brown suffusion on each side of midrib on distal half; inflorescence on a separate shoot, bracts pale green, coma bracts purple; corolla about 4.5 cm long, deep crimson-pink; labellum about 17 mm × 17 mm, pale yellow with deep yellow median band, other staminodes longitudinally folded, pale yellow, anther spurred. *C. aeruginosa* is found in grassy places and teak forest, at 400–750 m altitude.

**Selected sources** 202, 314, 455, 558, 580 615, 681, 1126, 1128, 1380, 1496, 1507, 1552.

### **Curcuma aurantiaca v. Zijp**

Recueil Trav. Bot. Néerl. 12: 345 (1915).

**Vernacular names** Indonesia: koneng kalamasu (Sundanese), temu blobo, temu purot (Javanese).

**Distribution** Peninsular Malaysia and Java; possibly also in India.

**Uses** Rhizomes are used medicinally for their astringent properties and have a camphor scent. Locally, young inflorescences are eaten as a vegetable in 'sayur' (soup).

**Observations** A herb with rhizome not elongated, inside cream-coloured; leaf sheaths to 12 cm long, blades elliptical, 19–50 cm × 8–21 cm, dark green; inflorescence terminal on a leafy shoot, bracts yellow-green, coma bracts red to pink or purple; corolla about 4.5 cm long, orange to orange-yellow; labellum about 2 cm × 1.5 cm, orange with a darker centre, other staminodes straight, orange, anther not spurred. *C. aurantiaca* is fairly common in Java, where it is found in teak forest.

**Selected sources** 97, 202, 314, 380, 615, 1157, 1290, 1496.

### **Curcuma euchroma Valetton**

Bull. Jard. Bot. Buitenzorg, sér. 2, 27: 42 (1918).

**Vernacular names** Indonesia: kunir kebo, temu batok, temu ketek (Javanese).

**Distribution** Java.

**Uses** Rhizomes are sometimes used medicinally as a substitute for those of *C. longa*.

**Observations** A herb with branched rhizome,

outside and inside bright orange or orange-yellow, tips whitish; leaf sheaths up to 50 cm long, blades obovate-elliptical, 26–72 cm × 12–25 cm, green with a dark red-brown midrib; inflorescence terminal on a leafy shoot, bracts pale green to pale yellow-green, coma bracts slightly violet-dotted; corolla almost 2 cm long, pinkish; labellum about 17 mm × 16 mm, ochraceous with a darker median band, other staminodes longitudinally folded, ochraceous, anther with long spurs. *C. euchroma* is found in teak forest.

**Selected sources** 97, 202, 1496.

### **Curcuma heyneana Valetton & v. Zijp**

Bull. Jard. Bot. Buitenzorg, sér. 2, 27: 54 (1918).

**Vernacular names** Indonesia: temu giring (Javanese).

**Distribution** Java, mostly in Central and East Java, both wild and cultivated.

**Uses** Rhizomes are the principal ingredient of a body lotion or powder used in traditional Javanese skin treatment, often administered to the bride-to-be. The bitter rhizomes are given, together with other medicinal herbs, to treat fatty degeneration, also as a folk medicine for brides to combat fatigue. They are also often applied in modern beauty parlours. Rhizomes are also considered to be cooling and detergent, useful to treat skin diseases, abrasions and injuries, and are used as an anthelmintic, especially against pinworms, and against lipomatosis in combination with other plants. They yield starch that can be made into a porridge.

**Observations** A herb with much branched and elongated rhizome, outside pale yellow, inside whitish with yellowish centre to bright yellow throughout; leaf sheaths 22–35 cm long, blades elliptical, 17.5–42 cm × 7.5–13 cm, uniformly green; inflorescence on a separate shoot, bracts pale green, coma bracts pale pink with a dark tip; corolla about 4 cm long, whitish; labellum about 16 mm × 16 mm, white with a dark yellow median band to yellow, other staminodes longitudinally folded, whitish to yellow, anther with short spurs. *C. heyneana* grows wild in secondary forest, teak forest and abandoned places, up to 750 m altitude.

**Selected sources** 97, 202, 314, 380, 1496, 1507.

### **Curcuma longa L.**

Sp. pl. 1: 2 (1753).

**Synonyms** *Curcuma domestica* Valetton (1918).

**Vernacular names** Turmeric (En). Curcuma, safran des Indes, turmeric (Fr). Brunei: kunyit,

temu kuning, temu kunyit (Dusun, Malay). Indonesia: kunyit (general), kunir (Javanese), ko-neng (Sundanese). Malaysia: kunyit, temu kunyit, tius. Papua New Guinea: lavar, tamaravirua (Guntantuna, New Britain). Philippines: dilaw (Tagalog), kalabaga (Bisaya), kunik (Ibanag). Cambodia: ro miet. Laos: khi min, 'khmin<sup>2</sup> 'khun<sup>2</sup>. Thailand: khamin (general), khamin kaeng (northern), khamin chan (central). Vietnam: ngh[eej], ngh[eej] v[af]ng, u[aa]s[t] kim.

**Distribution** Turmeric probably originated from South or South-East Asia, most probably from India. It is not known in a true wild state although in some places it appears to have become naturalized (e.g. in teak forests of East Java). Turmeric has been grown in India since time immemorial. It reached China before the 7th Century, East Africa in the 8th Century and West Africa in the 13th Century. It was introduced into Jamaica in the 18th Century. Presently turmeric is widely cultivated throughout the tropics but cultivation on a considerable scale is largely confined to India and South-East Asia.

**Uses** The main rhizomes are stomachic, stimulant, carminative, haematic or styptic in all kinds of haemorrhages, and a remedy for certain types of jaundice and other liver trouble. Externally they are applied to relieve itch, small wounds, insect bites and certain skin eruptions and smallpox, also as a maturative. A decoction affords relief for a burning sensation in eye disease. The rhizomes are considered to be very good for irregular menstruation; they promote circulation, dissolve blood clots and are prescribed as a remedy for urinary infections and for abdominal, chest, and back pains. Turmeric is a remedy for diarrhoea, rheumatism, and to relieve cough and tuberculosis. It is further considered anti-spasmodic, and as a cure for inflammation of the gums. Turmeric also has insecticidal, fungicidal and nematocidal properties which make it a potential biocide. The leaves are used in preparing a special medicinal bread in Nepal and India, whereas in Papua New Guinea they are applied to skin pains, bruises, eye irritations, catarrh and colds. The major use of turmeric rhizomes is, however, as a spice. They are also applied as a yellow dye for clothing and food, and as a cosmetic and pH indicator. The rhizomes are an auspicious article in all religious observances in Hindu households, and have many other uses in daily life in connection with birth, marriage, and death, and in agriculture.

**Observations** A herb with branched rhizome, bright orange inside and outside, young tips



*Curcuma longa* L. - 1, rhizome; 2, flowering clump.

white; leaf sheaths up to 65 cm long, blades oblong-lanceolate to ovate-lanceolate, 7-70 cm × 3-20 cm, densely studded with pellucid dots; inflorescence terminal on a leafy shoot, bracts pale green with white streaks or white margins, coma bracts white, sometimes pink-tipped; corolla 4.5-5.5 cm long, white; labellum suborbicular to obovate, 12-22 mm in diameter, white with a yellow median band, other staminodes longitudinally folded, creamy white, anther with large spurs. *C. longa* is found naturalized mainly in teak forest, but also in sunny places, on clayey to sandy soils, up to 2000 m altitude.

**Selected sources** 31, 59, 73, 90, 97, 202, 287, 304, 314, 326, 350, 363, 413, 455, 479, 531, 580, 583, 597, 615, 681, 776, 897, 1035, 1066, 1097, 1126, 1128, 1158, 1163, 1178, 1337, 1380, 1421, 1450, 1452, 1458, 1467, 1481, 1496, 1507, 1525.

***Curcuma mangga* Valetton & v. Zijp**

Bull. Jard. Bot. Buitenzorg, sér. 2, 27: 50 (1918).

**Vernacular names** Indonesia: temu mangga

(general), koneng lalab (Sundanese), temo pao (Madurese). Malaysia: temu pauh (Peninsular). Thailand: khamin khao.

**Distribution** Only known from cultivation in Thailand, Peninsular Malaysia and Java.

**Uses** Rhizomes are part of a mixture given to treat continued fever. They are chewed to cause the womb to contract after childbirth. The starch is recommended by traditional healers to treat abdominal illness. The main use of *C. mangga* is as a vegetable.

**Observations** A herb with branched rhizome, yellowish outside, the top white, inside lemon-coloured to sulphur-yellow with a white outer layer; leaf sheaths 30–65 cm long, blades elliptical-oblong to oblong-oblancheolate, 15–95 cm × 5–23 cm, green; inflorescence on a separate shoot, bracts green, coma bracts white at base, purple towards the top; corolla 3–4 cm long, white; labellum 15–25 mm × 14–18 mm, white with a yellow median band, other staminodes longitudinally folded, white, anther with long, narrow spurs. *C. mangga* is cultivated in very fertile soils, up to 1000 m altitude.

**Selected sources** 97, 108, 202, 314, 380, 615, 681, 1066, 1126, 1356, 1420, 1496.

### **Curcuma petiolata** Roxb.

Fl. ind. (Carey ed.) 1: 36 (1820).

**Vernacular names** Indonesia: temu badur (Javanese), temu putri (Malay, Jakarta), temu tihing (Balinese).

**Distribution** Burma (Myanmar), Java and Bali; possibly also in the Moluccas.

**Uses** Rhizomes are used in the treatment of anorexia, puerperal fever and colic.

**Observations** A herb with rhizome not elongated, forming a compact mass, outside sallow-yellow, inside pale yellow; leaf sheaths 20–30 cm long, blades broadly elliptical-ovate to elliptical-lanceolate, 15–65 cm × 6–30 cm, green; inflorescence terminal on a leafy shoot, bracts greenish with violet streaks to entirely violet, coma bracts dark red to purplish; corolla 3–3.5 cm long, white with yellow or pink tips; labellum 13–17 mm × 15–17 mm, pale orange, other staminodes straight, pale orange, anther with short spurs. *C. petiolata* is naturalized in teak forest, upland fields, near bamboo stools and in other shaded places, up to 1600 m altitude. The status of this species needs further attention.

**Selected sources** 108, 314, 380, 1496, 1511.

### **Curcuma purpurascens** Blume

Enum. pl. Javae 1: 46 (1827).

**Vernacular names** Indonesia: koneng pinggang, koneng tinggang (Sundanese), temu tis (Javanese).

**Distribution** West and Central Java.

**Uses** Rhizomes are used against tussis and, when mixed with *Alyxia stellata* (Forst.) Roem. & Schultes, applied as a poultice after childbirth. The main tubers contain extractable starch. The tender central parts of the shoots and young rhizomes are eaten fresh or cooked as 'lalab'.

**Observations** A herb with branched rhizome, outside and inside orange-yellow with whitish tips; leaf blades elliptical, 55–70 cm × 19–23 cm, green but purple along the midrib above; inflorescence terminal on a leafy shoot, bracts pale green, coma bracts white at base and pale green towards the top or almost entirely white, outside pale brown spotted at the top; corolla about 5 cm long, white; labellum about 17 mm × 17 mm, pale creamy yellow with a dark yellow median band, other staminodes pale creamy yellow, anther with long spurs. *C. purpurascens* grows spontaneously in teak forest. It is cultivated up to 1000 m altitude.

**Selected sources** 97, 314, 580, 1066, 1496.

### **Curcuma soloensis** Valetton

Bull. Jard. Bot. Buitenzorg, sér. 2, 27: 46 (1918).

**Vernacular names** Indonesia: temu blenyeh, temu glenyeh, temu bayi (Javanese).

**Distribution** Java; fairly widely cultivated in Central Java.

**Uses** Rhizomes are used in the preparation of many traditional medicines against boils, scabies, cough and fever, and are especially used for children.

**Observations** A herb with branched rhizome, outside orange or orange-brown, inside orange-yellow; leaf blades elliptical, 29–55 cm × 12.5–19 cm, green; inflorescence terminal on a leafy shoot, bracts very pale green, coma bracts nearly white at base, dark violet towards the top; corolla about 5 cm long, pale pink; labellum 15–16 mm × 16–17 mm, orange, other staminodes longitudinally folded, orange, anther with long spurs. *C. soloensis* is found in teak forest and similar habitats, up to 900 m altitude.

**Selected sources** 97, 580, 1496.

### **Curcuma xanthorrhiza** Roxb.

Fl. ind. (Carey ed.) 1: 25 (1820).

**Vernacular names** Indonesia: koneng gede (Sundanese), temu lawak (Javanese), temo labak

(Madurese). Malaysia: temu lawas, temu raya (Peninsular). Thailand: wan chakmotluk (central). Vietnam: ngh[eej] v[af]ng, ngh[eej] r[leex] v[af]ng.

**Distribution** *C. xanthorrhiza* is native to Java, Bali and the Moluccas. It is commonly cultivated in Java, Peninsular Malaysia, the Philippines and Thailand, occasionally also in India.

**Uses** Rhizomes are used to treat various abdominal complaints and liver disorders (jaundice, gallstones, promoting the flow of bile). A decoction of the rhizome is also used as a remedy for fever and constipation, and taken by women as a galactagogue and to lessen uterine inflammation after giving birth. Other applications are against bloody diarrhoea, dysentery, inflammation of the rectum, haemorrhoids, stomach disorders caused by cold, infected wounds, skin eruptions, acne vulgaris, eczema, smallpox and anorexia. In Indonesia, rhizomes enter as an important ingredient into many 'jamus'. They yield a starch, and a yellow dye. Young stems and rhizome parts are eaten as a vegetable either raw or cooked. The inflorescences are eaten cooked. In Java, a soft drink called 'bir temu lawak' is prepared by cooking dried pieces of rhizomes.

**Observations** A herb with branched rhizome, outside dark yellow to reddish-brown, inside orange or orange-red; leaf sheaths up to 75 cm long, blades elliptical-oblong to oblong-lanceolate, 25–100 cm × 8–20 cm, green with a reddish-brown band along the midrib; inflorescence on a separate shoot, bracts pale green, coma bracts purple; corolla 4–6 cm long, pale red; labellum 2–2.5 cm × 1.5–2 cm, yellowish with a darker yellow median band, other staminodes longitudinally folded, yellowish-white, anther with long spurs. *C. xanthorrhiza* is found in thickets and teak forest, mainly on moist, fertile, humus-rich soils, up to 750 m altitude.

**Selected sources** 97, 202, 269, 270, 310, 314, 329, 350, 414, 580, 615, 660, 681, 776, 866, 867, 912, 1066, 1097, 1098, 1112, 1211, 1212, 1380, 1496, 1507, 1622, 1623, 1624.

### **Curcuma zedoaria (Christm.) Roscoe**

Trans. Linn. Soc. London 8: 354 (1807).

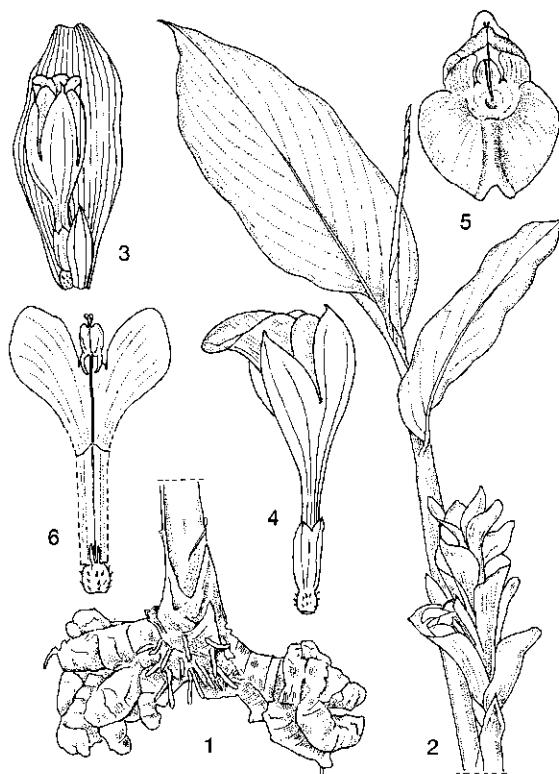
**Synonyms** *Curcuma pallida* Lour. (1790), *Curcuma zerumbet* Roxb. (1810).

**Vernacular names** Long zedoary, round zedoary, zedoary (En). Zédouaire (Fr). Indonesia: ko-neng tegal (Sundanese), temu putih (Malay, Jakarta). Malaysia: kunchur, temu kuning, temu lawak (Peninsular). Philippines: alimpuyas (Cebu Bisaya), barak (Tagalog), tamahilan (Bikol). Bur-

ma (Myanmar): thanuwen. Cambodia: prâtiël prèah 'angkaól. Laos: 'khmin<sup>2</sup> khai. Thailand: khamin khun (northern), khamin oi (central). Vietnam: ngh[eej] den, nga tru[aa]t, ng[ar]i t[is]m.

**Distribution** Probably native to north-eastern India. Distributed through cultivation to South and South-East Asia and probably throughout Malesia, China and Taiwan from where it easily escapes. Occasionally it is cultivated elsewhere (e.g. in Madagascar).

**Uses** Rhizomes are widely used as stimulant, stomachic, carminative, diuretic, anti-diarrhoeal, anti-emetic, anti-pyretic and depurative, the latter especially after childbirth, but also to clean and cure ulcers, wounds and other kinds of skin disorders. Rhizomes are also chewed against bad breath, and a decoction is drunk against stomach-ache, indigestion and colds. The major use of rhizomes is for starch. In Indonesia, the heart of young shoots is used as a vegetable, young rhi-



*Curcuma zedoaria* (Christm.) Roscoe – 1, rhizome; 2, leafy shoot and inflorescence; 3, flower with bract and bracteole; 4, flower in lateral view; 5, flower in front view; 6, stamen, lateral staminodes and pistil.

zome parts are eaten raw and inflorescences cooked. The leaves are used for flavouring foods. In India, rhizomes are also used in perfumery.

**Observations** A herb with branched rhizome, outside grey, inside pale yellowish to bright yellow; leaf sheaths 35–60 cm long, blades oblong to oblong-lanceolate, 25–75 cm × 7–20 cm, green with a purple band along the midrib; inflorescence on a separate shoot, bracts green or green with a purple margin, coma bracts purple or dark pink; corolla 3.5–4.5 cm long, yellowish-white; labellum 2–2.5 cm × 1.5–2 cm, yellowish-white with a darker yellow median band, other staminodes longitudinally folded, yellowish-white, anther with long spurs. *C. zedoaria* is found in various shady, damp localities on various soils, but prefers well-drained sandy soils, up to 1000 m altitude.

**Selected sources** 97, 117, 202, 203, 287, 314, 332, 363, 414, 455, 479, 531, 580, 615, 639, 897, 1035, 1066, 1112, 1126, 1128, 1178, 1211, 1212, 1287, 1380, 1496, 1507, 1525.

Trimurti H. Wardini & Budi Prakoso

## Cyclea Arn. ex Wight

III. Ind. Bot. 1: 22 (1840).

MENISPERMACEAE

*x* = unknown

**Major species** *Cyclea barbata* Miers and *C. laxiflora* Miers.

**Origin and geographic distribution** *Cyclea* includes about 30 species and occurs in India, Burma (Myanmar), Indo-China, southern China, Thailand and western and central Malesia (Peninsular Malaysia, Sumatra, Java, Borneo and the Philippines). In Malesia, 10 species have been found. *Cyclea* is occasionally planted, e.g. in Java.

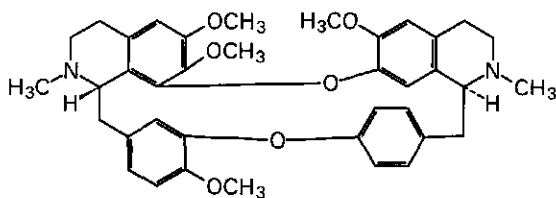
**Uses** A decoction of the roots is used in local medicine in Indonesia and Malaysia against fever and haemorrhoids, after childbirth and as a vermifuge for children. In Thailand, *C. barbata* ('krung khamao') is a rather well-known medicinal plant; the bitter decoction of its roots is used against (malarial) fever, in the treatment of lower abdominal pains, eye diseases, jaundice and as a tonic. *Cissampelos pareira* L., which has similar attributed uses and qualities, is also called 'krung khamao'. In Java, fresh crushed leaves of *C. barbata* are mixed with water, filtered and kept overnight to prepare a jelly called 'cincau' or 'cincau hijau' (green cincau), which is used as a refreshment and as a remedy against stomach complaints and fever as it has a cooling effect. In Thai-

land, a comparable preparation is used as appetizer and in the treatment of lower abdominal pains associated with malaria. The stem and roots of *Cyclea* are used in local medicine in southern China and Indo-China as diuretic, depurative, febrifuge and antidiysenteric, and to treat jaundice, eye diseases and framboesia. In Chinese medicine extracts from the roots are used as anaesthesia during surgery.

**Production and international trade** The roots are traded and used on a local scale, and have no importance on the international market. The jelly is traded in a small-scale syrup industry.

**Properties** Many *Menispermaceae* species are known to contain mixtures of alkaloids; in the case of *C. barbata* contents of 4–7% in the roots are reported. This mixture is of complex nature. The principal components are a series of related bisbenzylisoquinoline alkaloids, including (+)-S,S-tetrandrine (main alkaloid, up to 3% in the roots), (±)-tetrandrine, (+)-R,S-isotetrandrine, (-)-R,R-limacine, (±)-fangchinoline, (+)-R,S-isofangchinoline (= thalrugosine), (+)-R,S-berbamine, (+)-R,S-homoaromoline, (+)-S,S-N-methyltetrandrine, (+)-S,S-tetrandrine-2'-β-N-oxide, (-)-cycleapeltine, (-)-2'-norlimacine, (+)-cycleabarbatine, (-)-repandine, (+)-cycleanorine, (+)-daphnandrine, (+)-coclaurine, (-)-N-methylcoclaurine, (-)-curine (= (-)-berberine = R,R-chondodendrine), R,R-isochondodendrine and R,S-chondocurine. Besides the bisbenzylisoquinoline alkaloids, α-cyclanolin and its epimer β-cyclanolin (two tetrahydroprotoberberine type alkaloids) and magnoflorine (an aporphine type alkaloid) have also been isolated from *C. barbata* roots. Dicentrine, an alkaloid structurally related to magnoflorine has been isolated from the roots of *C. laxiflora*. The only non-alkaloid isolated from *C. barbata* roots is the sugar protoquercitol, which is often found in *Menispermaceae*.

An extract derived from the roots of *C. barbata* has demonstrated in vitro cytotoxic and anti-malarial activity. Using bioactivity guided isolation procedures, 5 bisbenzylisoquinoline alkaloids (i.e. (+)-tetrandrine, (-)-limacine, (+)-thalrugosine, (+)-homoaromoline and (-)-cycleapeltine) have been isolated as active principles. These compounds are capable of inhibiting the growth of both cultured *Plasmodium falciparum* strains and tumour cell lines. However, their 'selectivity index' (activity against mammalian cells/activity against cultured *P. falciparum* strains) typically ranges from 2 to 100, which is low compared to a selectivity index over 1000 for quinine or artemisinin.



(+)-S,S-tetrandrine

Methylation of (-)-curine has yielded dimethyl(-)-curine dimethochloride, whose chemical structure is closely related to (+)-tubocurarine from South American *Chondrodendron tomentosum* Ruiz & Pavón (*Menispermaceae*). The muscle-relaxant activity of dimethyl(-)-curine dimethochloride was found to be within the range of that of (+)-tubocurarine. Both have a non-depolarizing mode of action, and the blocking site is at the cholinergic (nicotinic) receptor in the postsynaptic membrane. The muscle-relaxant action and the inhibition of breathing could be antagonized by neostigmine. Side-effects are limited to mild hypotension and temporary facial cooling, whereas cardiovascular disturbances are insignificant. Dimethyl(-)-curine dimethochloride is therefore considered a safe muscle relaxant for surgery.

S,S-tetrandrine, one of the pure isolated bisbenzylisoquinolines, has undergone extensive pharmacological investigations. It has been shown that this compound is a new kind of blocker of the voltage-activated L-type  $\text{Ca}^{2+}$  channel in a variety of excitable cells, such as cardiac,  $\text{GH}_3$  anterior pituitary, neuroblastoma cells and rat neurohypophysial nerve terminals. As well as blocking L-type  $\text{Ca}^{2+}$ , it also blocks the voltage-dependent T-type  $\text{Ca}^{2+}$  channel. S,S-Tetrandrine's action in the treatment of cardiovascular diseases, including hypertension and supraventricular arrhythmia, is primarily due to its blocking of these voltage-activated  $\text{Ca}^{2+}$  channels. Furthermore, the alkaloid is a potent blocker of the  $\text{Ca}^{2+}$  activated  $\text{K}^+$  channels of neurohypophysial nerve terminals. There is no obvious clinical application of this action, but S,S-tetrandrine makes a promising ligand for the study of the  $\text{K}^+$  channel function.

Other pharmacological effects include anti-inflammation and immunosuppression. These actions often require higher doses than those needed to produce the cardiovascular effects. The mechanism(s) by which S,S-tetrandrine exerts these actions are unknown, although in vitro experiments have demonstrated inhibition of the production and release of inflammatory mediators and cytokines

such as histamine, prostaglandins, leukotrienes, platelet activating factor, interleukin-1, tumour necrosis factor and nitric oxide. The inhibition of tumour necrosis factor has also been shown in vivo, using a mouse model for fulminant hepatitis. A selective inhibition of T-cell dependent immune responses by S,S-tetrandrine has been observed in mice, and suppression of the chronic inflammation in an arthritis model in the rat.

In media containing calcium, the stereoisomer of S,S-tetrandrine, R,S-isotetrandrine, exerts similar, although less specific, calcium channel activities. In contrast to S,S-tetrandrine, the mechanism of action of R,S-isotetrandrine also involves intracellular mechanisms, since it has been shown that this component is also active in calcium-free media, whereas S,S-tetrandrine is not. Other activities of R,S-isotetrandrine are inhibition of histamine release (in vitro), inhibition of nitric oxide production (in vitro) and selective inhibition of T-cell dependent immune responses (in vivo, mouse). Isotetrandrine also markedly suppressed the tumour-promoting effect of 12-O-tetradecanoylphorbol-13-acetate in a mouse-skin carcinogenesis test.

Biological activities of R,S-chondocurine include suppression of nitric oxide production (in vitro), selective inhibition of T-cell dependent immune responses (in vivo, mouse), suppression of delayed type hypersensitivity (in vivo, mouse) and reduction of the level of tumour necrosis factor in a fulminant hepatitis model (in vivo, mouse).

Finally, (+)-homoaromoline and fangchinoline show inhibition of the histamine production by RBL-2H3 cells in vitro.

**Adulterations and substitutes** Other *Menispermaceae* have similar or related alkaloids such as bisbenzylisoquinolines and protoberberines, and have similar applications. There is a report of *C. barbata* being substituted by *Cissampelos pareira*. In Java, *C. barbata* leaves are sometimes substituted by the leaves of *Stephania capitata* (Blume) Sprengel or *Canthium horridum* Blume, for the production of 'cincau' jelly.

**Description** Slender dioecious lianas up to 15 m long; stem herbaceous or woody, hispid to glabrous; roots sometimes tuberous. Leaves arranged spirally, simple and entire, often peltate, palmately veined; stipules absent. Inflorescence axillary, terminal or cauliflorous, pseudoracemose or thyrsoid. Flowers unisexual, calyx with free or connate sepals, corolla with free or connate petals (rarely absent); male flower 4(-5)-merous, with stamens fused into a peltate synandrium with 4-5

anthers; female flower 2-3-merous, with 1 carpel having a 3-5-fid stigma, without staminodes. Fruit a curved, obovate to rotund drupe with style-scar near base; endocarp bony, dorsally ornamented with 3-6 rows of tubercles. Seed horse-shoe-shaped; embryo narrow, embedded in endosperm.

**Growth and development** The flowers are pollinated by insects such as small flies and bees and possibly also small beetles and moths, which are attracted by the scent of the flowers and possibly also by the smell of the leaves.

**Other botanical information** *Cyclea* is closely related to *Stephania*, which differs in its 2 whorls of sepals in the male flowers and its umbelliform or disciform inflorescences.

**Ecology** *Cyclea* occurs in forest, often in secondary forest, coastal forest, teak forest and bamboo forest, and in scrub vegetation, hedges, cultivated land and alang-alang (*Imperata cylindrica* (L.) Rauschel) fields, up to 1200 m altitude. Some *Cyclea* species have been found at altitudes as high as 2800 m.

**Propagation and planting** Cultivated *C. barbata* fruits erratically, so propagation is by stem or root cuttings. Cuttings from young woody stems, 25-30 cm long, are planted near hedges, trees or trellises, for support at a later stage. Cuttings may also be planted in the field at a spacing of 2 m x 1.5 m, supported by stakes, under shade trees. Before planting 5 kg manure and 10 g triple superphosphate is added per planting hole. Two weeks later an additional 10 g ammonium sulphate and 10 g potassium chlorite is given.

**Harvesting** A first harvest of *C. barbata* leaves from stem cuttings can be expected after 6-8 months. Consecutive harvests are at intervals of 2-3 months.

**Handling after harvest** To make good quality jelly, 'cincau hijau', young well-developed leaves should be used. The shelf-life of the jelly at ambient temperature is 1-2 days.

**Genetic resources and breeding** *C. barbata* and *C. laxiflora* are locally rather common, the former in Java and the latter in Peninsular Malaysia, and occur particularly in disturbed forest. They do not seem at risk of genetic erosion. However, some other *Cyclea* species are endemic to comparatively small areas (e.g. *C. kinabaluensis* Forman on Mount Kinabalu) and are more likely to become endangered.

**Prospects** *Cyclea* root extracts show some interesting properties that deserve more research. In particular the powerful neuromuscular block-

ing property comparable to d-tubocurarine chloride, and the antimalarial activity warrant more attention.

**Literature** |1| Forman, L.L., 1986. Menispermaceae. In: van Steenis, C.G.G.J. & de Wilde, W.J.J.O. (Editors): Flora Malesiana. Series 1, Vol. 10. Kluwer Academic Publishers, Dordrecht, Boston, London. pp. 237-243. |2| Forman, L.L., 1991. Menispermaceae. In: Smitinand, T. & Larsen, K. (Editors): Flora of Thailand. Vol. 5(3). The Forest Herbarium, Royal Forest Department, Bangkok, Thailand. pp. 325-331. |3| Guinaudeau, H., Lin, L.Z., Ruangrunsi, N. & Cordell, G.A., 1993. Bisbenzylisoquinoline alkaloids from *Cyclea barbata*. Journal of Natural Products 56(11): 1989-1992. |4| Klughardt, G. & Zymalkowski, F., 1982. Magnoflorin und Protoquercit als Inhaltsstoffen von *Cyclea barbata* Miers [Magnoflorine and protoquercitol as constituents of *Cyclea barbata* Miers]. Archiv der Pharmazie (Weinheim) 315(1): 7-11. |5| Lin, L.Z., et al., 1993. Cytotoxic and antimalarial bisbenzylisoquinoline alkaloids from *Cyclea barbata*. Journal of Natural Products 56(1): 22-29. |6| Martin, H.-J., Pachaly, P. & Zymalkowski, F., 1977. Alkaloide aus der thailändischen Menispermaceen-Droge Krung Kha Mao (*Cyclea barbata*), 6. Mitt. Isolierung und Strukturaufklärung quartärer Tetrahydroprotoberberin-Alkaloide [Alkaloids from the Thai Menispermaceae drug krung kha mao (*Cyclea barbata*), part 6. Isolation and structural elucidation of quaternary tetrahydroprotoberberine alkaloids]. Archiv der Pharmazie (Weinheim) 310(4): 314-319. |7| Nguyen Van Duong, 1993. Medicinal plants of Vietnam, Cambodia and Laos. Mekong Printing, Santa Ana, California, United States. p. 259. |8| Siwon, J., 1982. A pharmacognostical study of some Indonesian plants of the family Menispermaceae. PhD thesis, Leiden University, the Netherlands. pp. 10, 100-103. |9| Wu, J. & Xie, F.S., 1987. Dimethyl-1-curine dimethochloride an alternative for D-tubocurarine chloride. Chinese Medical Journal 100(3): 173-176. |10| Yang, Q.Z. & Lin, L.R., 1981. Mode of action of dimethyl-levo curine dimethochloride on neuro-muscular transmission. Acta Pharmacologica Sinica 2(1): 19-23.

#### *Selection of species*

#### ***Cyclea barbata* Miers**

Contr. bot. 3: 237 (1871).

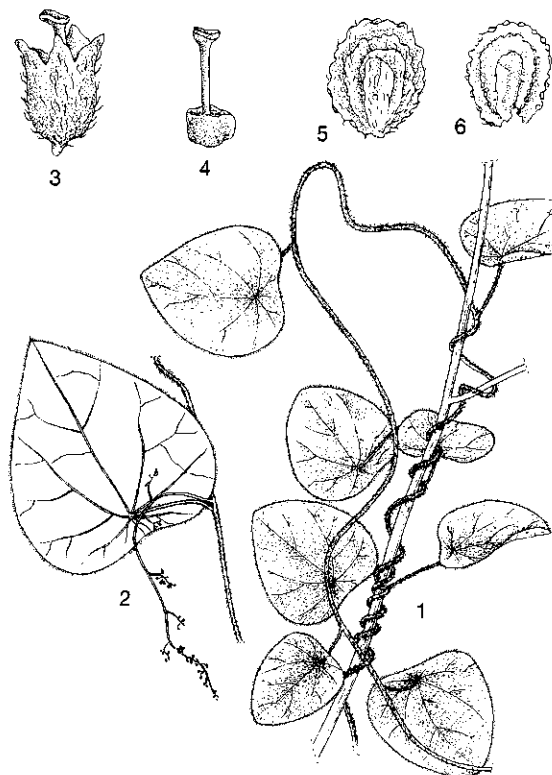
**Synonyms** *Cyclea peltata* auct. non (Lamk) Hook.f. & Thomson.

**Vernacular names** Indonesia: cincau (general), camcauh (Sundanese), camcao (Javanese). Thailand: krung khamao (peninsular), krung badan (south-eastern). Vietnam: d[aa]y s[aa]m, s[aa]m l[oo]ng.

**Distribution** India (Assam), Burma (Myanmar), Indo-China, Thailand, Simeuluë, islands in the Sunda Strait and Java.

**Uses** A decoction made of the roots is used in local medicine against fever. The leaves are used in Java to prepare a jelly called 'cincau' or 'cincau hijau' (green cincau), which is used as a refreshment and as a medicine against stomach complaints.

**Observations** A slender herbaceous or woody climber up to 8 m long, root fleshy, thick and long, pale brown outside, whitish or yellowish inside, stem hispid when young, later glabrescent; leaves ovate, deltoid-ovate or broadly ovate, up to 17.5 cm long, hairy below, petiole up to 6.5 cm long, hispid; male flowers with puberulous calyx and connate petals, female flowers tightly crowded in



*Cyclea barbata* Miers - 1, leafy stem; 2, stem with leaf and male inflorescence; 3, male flower; 4, male flower with calyx removed; 5, fruit; 6, endocarp.

subglobose heads, with tomentose carpels; fruit puberulous. *C. barbata* occurs in forest including teak forest and bamboo forest, and in grassland with scrub vegetation, sometimes on limestone, occasionally cultivated, up to 1100 m altitude.

**Selected sources** 305, 421, 422, 423, 493, 580, 751, 907, 1035, 1374, 1597, 1615, 1638.

### *Cyclea laxiflora* Miers

Contr. bot. 3: 241 (1871).

**Vernacular names** Malaysia: akar gasing bukit, akar rempenang, metimun tikus (Peninsular).

**Distribution** Thailand (peninsular), Peninsular Malaysia, Sumatra, Bangka and the Anambas Islands.

**Uses** A decoction made of the roots is used in local medicine in Peninsular Malaysia against fever and piles, after childbirth, as a vermifuge for children and applied externally to the nostrils in caries of the nasal bones.

**Observations** A slender herbaceous or slightly woody climber up to 15 m long, stem hispid to glabrous; leaves triangular-ovate, deltoid-ovate or broadly ovate, up to 16 cm long, hairy below, petiole up to 6 cm long, usually hispid; male flowers with glabrous or subglabrous calyx and free petals, female flowers in lax clusters, with glabrous or subglabrous carpels; fruit glabrous. *C. laxiflora* occurs in secondary and coastal forest, scrub vegetation, hedges and cultivated land, up to 1200 m altitude.

**Selected sources** 202, 421, 422, 423.

R.H.M.J. Lemmens & S.F.A.J. Horsten

### *Cyperus* L.

Sp. pl. 1: 44 (1753); Gen. pl. ed. 5: 26 (1754).

CYPERACEAE

$x = 8, 9$ ; *C. brevifolius*:  $n = 9, 2n = 32, 120$ , *C. iria*:  $n = 36, 56, 64, 2n = 18$ , *C. kyllingia*:  $n = 43, 62$ , *C. rotundus*:  $n = 40, 42, 48, 50, 54, 56, 64, 69, 76, 2n = 16, 26, 96, 108, 152$

**Major species** *Cyperus kyllingia* Endl., *C. rotundus* L.

**Vernacular names** Nut sedge, nut grass, cyperus (En). Indonesia: teki. Malaysia: teki. Thailand: ya kok, ya haeo. Vietnam: c[os]i, l[as]c.

**Origin and geographic distribution** *Cyperus* comprises about 600 species distributed worldwide, with the vast majority occurring in the tropics and subtropics. In Malesia 76 species occur, but most of them have a pantropical or paleotropical distribution.



**Uses** In general, *Cyperus* is economically unimportant and better known for its weeds than its useful plants; several species are pernicious weeds in agriculture.

Several of the weedy species, particularly *C. rotundus*, are used in local medicine. In South-East Asia, the most widespread medicinal applications of the tubers are against hepatic disorders, as an emmenagogue and for post-partum treatment, as a stomachic, as a febrifuge, against renal and urologic disorders, and to treat ulcers. It is interesting that *Cyperus* species are used in similar ways in other parts of the world, e.g. in South America. Other medicinal uses in South-East Asia are as a stimulant, diuretic, anthelmintic (in large doses), galactagogue, sudorific, as a mouthwash against diseases in the mouth and toothache, and as an astringent against diarrhoea and dysentery.

Usually the powdered tubers (often in decoctions) are used internally, but for ulcers, scorpion stings and snake bites a poultice is applied externally. *Cyperus* species are also well known in Chinese and Japanese traditional medicine and are reported to be used in India as stimulants, diuretics, anthelmintics and stomachics for example. In the Ayurvedic system of medicine the tubers are believed to be tonic, and to act as a nourisher and rejuvenator; they are part of several complex preparations.

Because of their aroma, the tubers are used to perfume clothing and to repel insects. The roasted tubers have also been used as a substitute for coffee and as an adulterant for cocoa.

The fatty oil extracted from the tubers was formerly used to make soap. The dried stems are used to make mats, ropes, baskets and other wickerwork. Tubers and aboveground parts can also serve as a forage. Some species are used for their edible tubers (especially in times of food scarcity), for their edible stems (as vegetable or as forage), and some are used as garden ornamentals.

**Production and international trade** All *Cyperus* species are considered as weeds that are difficult to eradicate. *C. rotundus* is considered to be one of the world's most pernicious weeds. The sole medicinal use of the tubers of this species is in local medicine.

**Properties** The underground parts of several weedy species contain essential oils, about 0.5–1% in the case of the fresh tubers of *C. rotundus*, mainly consisting of terpenoids or sesquiterpenoids (e.g. cyperone, cyperol, cyperolone, cyperene, copadiene, epoxy-guaiaene, rotundone, rotundol, patchoulone (= cyperotundon), kobuso-

ne, sugeonolacetate, sugetriol, oxido-eudesmenol, mustakone and  $\beta$ -selinene).

When Tanzanian medicinal plants were screened, *C. rotundus* showed activity in a test for in vitro antimalarial activity. Further phytochemical investigations revealed the presence of several active compounds:  $\alpha$ -cyperone, an auto-oxidation product of  $\beta$ -selinene, patchoulone, caryophyllene epoxide, 4,7-dimethyl-1-tetralone (all sesquiterpenes) and 10,12-peroxycalamenene (a sesquiterpene endoperoxide). There is evidence that the peroxide moiety of the latter molecule is important in the activity against malarial parasites, as are artemisinin (from *Artemisia annua* L.) and ascaridole (from *Chenopodium* oil).

The essential oils furthermore have nematocidal and insecticidal activity (e.g. against bugs, beetles and caterpillars), and the insect-repellent activity is comparable to that of citronella oil. The aggressive spread as a weed may also be partly attributed to the sesquiterpenes present in the oil, which inhibit the growth of seedlings of agricultural crops.

Several investigations have focused on the anti-inflammatory and/or anti-pyretic effects of *C. rotundus*. Preliminary observations were made on the ethanolic extracts of the roots. The mechanism of activity was subsequently found to be inhibition of prostaglandin synthesis. In addition to the effects mentioned, some antihistaminic, smooth muscle relaxant, anti-emetic, antihepatotoxic and antifungal activities were observed. A carrageen-induced oedema test in rat revealed that the triterpenoid in the light petroleum ether extract of the root is the main active compound with in vivo anti-inflammatory activity.

Small amounts of saponins have been reported from *C. rotundus*, and the main substance of the lipid fraction in tubers of *C. iria* has been shown to be hentriacontanol. In Malaysia, a compound isolated from a steam distillate of the whole plants of *C. iria* appeared to be identical to juvenile hormone produced by insects. Nymphs of the acridid *Melanoplus sanguinipes* feeding on *C. iria* showed pronounced morphogenetic effects when they moulted to adults, so the compound can be considered as the plant's defence mechanism against insects. The water extract of rhizomes exhibits bradycardia and cardia depressant, coronary vasodilator, hypotensive and diuretic activities. Clinical trials with root extracts in 64 obese patients found hypotensive activity in those patients with hypertension, but no effect in normal patients.

**Adulterations and substitutes** The following species combine several of the major applications of *Cyperus*: *Artemisia* spp. (e.g. emmenagogue, stomachic, febrifuge, diuretic and ulcers), *Desmodium* spp. (e.g. hepatic disorders, stomachic, febrifuge, diuretic and ulcers), *Hibiscus* spp. (e.g. emmenagogue, stomachic, febrifuge, diuretic and ulcers), *Elephantopus scaber* L. (e.g. emmenagogue, febrifuge, diuretic and ulcers), *Heliotropium indicum* L. (e.g. emmenagogue, stomachic, diuretic and ulcers), *Leonurus* spp. (e.g. emmenagogue and for post-partum treatment, febrifuge and diuretic) and *Tinospora* spp. (e.g. hepatic disorders, stomachic, febrifuge and ulcers). However, the active compounds are in general quite distinct and the similar medicinal use is not a result of chemical conformity. Chemically related substances are found in some grass genera such as *Cymbopogon*; their essential oils have very similar sesquiterpenes and some similar applications.

**Description** Perennial or annual herbs, tufted or with creeping rhizome or stolons, sometimes with tubers at intervals; stem usually erect, triangular in cross-section, solid, usually leafy only at the base. Leaves tristichous, narrowly linear, grass-like, the lower ones often scale-like. Inflorescence terminal, simple to decompound, umbel-like or capitate; rays subtended by a leaf-like bract forming an involucre; spikelets subcompressed, quadrangular to subterete, 1-many-flowered; glumes distichous, usually 2 basal ones empty. Flowers bisexual, the uppermost of the spikelet often male or sterile; stamens 1-3; style continuous with the ovary, with 2-3 stigmas. Fruit a sessile or short-stalked nut, trigonous or lenticular.

**Growth and development** Most *C. rotundus* plants originate from a tuber; seed production is possible but unimportant. The sprouting tuber produces a rhizome which terminates as a green aerial shoot. While emerging from the soil a swelling (usually called a 'basal bulb') appears on the rhizome, often near the surface, but up to a depth of 20 cm. Roots form on the basal bulb and rhizomes grow out from it horizontally for a distance of 1-30 cm before the tip turns up to produce a new aerial shoot with another basal bulb, or, alternatively, to form a subterranean tuber from which another rhizome appears at the apical end, thus forming chains of tubers. The basal bulb and aerial shoot population may increase fivefold in the first 4 weeks after a tuber has been planted. It is believed that there are no buds at the nodes of the rhizomes and that no new plants can grow from rhizome fragments. Rhizomes and tubers are

white and fleshy when young and some become firmly packed with starch. On aging they darken, harden, and most of the tissue outside the endodermis of the rhizomes sloughs off to leave a wiry structure resistant to desiccation and decay. Most tubers are found in the top 15 cm of the soil, and when planted at 90 cm depth are unable to grow to the surface. In cropped areas the starch reserves are greatest in tubers that are below the disturbed layer; in uncropped land they are greatest in tubers near the soil surface.

Short photoperiods stimulate flowering; the period from emergence to flowering varies between 3-8 weeks. Short photoperiods might also stimulate tuber formation and it is believed that tubers do not form until flowering begins. Flowers are cross-pollinated, mostly by wind. Although many seeds are formed, they are rarely viable: seed germination averages 1-5%.

*C. iria* spreads by seed that may germinate immediately as soon as it falls on the ground. Germination percentage is about 40% but is much lower if the soil is under water.

**Other botanical information** *Cyperus* is classified in the subfamily *Cyperoideae* and the tribe *Cypereae*. It seems to be most closely related to *Bulbostylis* and *Fimbristylis*. *Cyperus* is subdivided into 3 subgenera and numerous sections. Subgenus *Cyperus* includes most species in the Malesian Archipelago (about 60) and is characterized by a trigonous or, when lenticular, dorsoventrally compressed nut, and often 3 stigmas. Subgenus *Pycreus* includes 9 species in Malesia and has a laterally compressed nut, persistent rachilla and 2 stigmas. Subgenus *Kyllinga*, which has often been treated as a separate genus, includes 7 species in Malesia (e.g. *C. brevifolius* and *C. kyllingia*) and is characterized by its laterally compressed nut, disarticulating rachilla and 2 stigmas.

**Ecology** Most *Cyperus* species are hygrophilous and grow in moist or wet localities at low and medium altitudes, only a few occurring above 2000 m altitude in the tropics. Several species occur commonly in grasslands, on roadsides, river banks and waste places. Some species are noxious weeds in cropped land, and other species are colonizers of muddy and sandy flats near river mouths and along the coast. *C. rotundus* is found in fields, on roadsides, in neglected areas, at the edges of woods, along irrigation canals and streams, all over the world up to about 50° latitude in both hemispheres, beyond which low temperatures limit its further expansion. It grows readily at any elevation, humidity, in almost any soil type, soil

moisture and pH, and can survive very high temperatures. Only low temperatures, shade, and very saline soils can limit its growth, and the tubers can remain dormant for a long time to carry the plant through the most extreme conditions of heat, drought, flooding or lack of aeration.

**Propagation and planting** All *Cyperus* species can be propagated by seed, and tuber-forming species can be easily propagated by tubers.

**Husbandry** Some *Cyperus* species, especially *C. rotundus* are serious weeds of rice, sugar cane, maize and vegetables in South-East Asia. Digging up all rhizome parts and/or the planting of crops that produce continuous shade for several years can eradicate them. Biological control methods have not yet been successful.

**Diseases and pests** *C. rotundus* has been reported as a host plant for *Rhizoctonia* disease and for root-knot nematodes (*Meloidogyne* spp.).

**Yield** *C. rotundus* may produce up to 40 t subterranean plant material per ha per year.

**Handling after harvest** The tubers of *C. rotundus* are briefly scorched by fire to get rid of the fine roots. They are then washed and dried in the sun or by hot air.

**Genetic resources and breeding** No germplasm collections and breeding programmes of *Cyperus* are known to exist.

**Prospects** It is expected that in South-East Asia *Cyperus* will primarily remain a weed problem for which effective herbicides and biological control methods still have to be found. It is not recommended to plant weedy species for medicinal purposes. Comparatively little research has been done on the chemistry and pharmacology. More research seems worthwhile on those *Cyperus* species used for similar medicinal purposes in many areas of the world.

**Literature** |1| Bulman, J.C., 1989. *Cyperus rotundus* L. In: Westphal, E. & Jansen, P.C.M. (Editors): Plant Resources of South-East Asia. A selection. Pudoc, Wageningen, the Netherlands. pp. 108–109. |2| de Padua, L.S. & Pancho, J.V., 1983. Handbook on Philippine medicinal plants. Vol. 4. Technical Bulletin vol. VI No 1. Documentation and Information Section, Office of the Director of Research, University of the Philippines, Los Baños, the Philippines. p. 29. |3| Holm, L.G., Plucknett, D.L., Pancho, J.V. & Herberger, J.P., 1977. The world's worst weeds, distribution and biology. East-West Center, University Press of Hawaii, United States. pp. 8–24, 240–243. |4| Kern, J.H., 1974. Cyperaceae. In: van Steenis, C.G.G.J. (Editor): Flora Malesiana. Series 1, Vol.

7(3). Noordhoff International Publishing, Leyden, the Netherlands. pp. 592–661. |5| Mercado, B.L., 1979. A monograph on *Cyperus rotundus* L. Biotrop Bulletin No 15. Biotrop, SEAMEO Regional Center for Tropical Biology, Bogor, Indonesia. 63 pp. |6| Nguyen Van Duong, 1993. Medicinal plants of Vietnam, Cambodia and Laos. Mekong Printing, Santa Ana, California, United States. pp. 155–156. |7| Quisumbing, E., 1978. Medicinal plants of the Philippines. Katha Publishing Co., Quezon City, the Philippines. pp. 112–115. |8| Shanmugasundaram, E.R.B., Akbar, G.K.M. & Shanmugasundaram, K.R., 1991. Brahmighritam, an Ayurvedic herbal formula for the control of epilepsy. Journal of Ethnopharmacology 33(3): 269–276. |9| Toong, Y.C., Schooley, D.A. & Baker, F.C., 1988. Isolation of insect juvenile hormone III from a plant. Nature, United Kingdom 333(6169): 170–171. |10| Vedavathy, S. & Rao, K.N., 1991. Antipyretic activity of six indigenous medicinal plants of Tirumala Hills, Andhra Pradesh, India. Journal of Ethnopharmacology 33(1–2): 193–196.

#### *Selection of species*

#### ***Cyperus brevifolius* (Rottb.) Hassk.**

Cat. Hort. Bog.: 24 (1844).

**Synonyms** *Kyllinga brevifolia* Rottb. (1773).

**Vernacular names** Indonesia: jukut pendul (Sundanese), rumput sadanan (Javanese), rumput kapas (Sumatra). Malaysia: kanching bayu jantan, rumput sekepet burit, rumput teki (Peninsular). Philippines: boto-botonisan (Tagalog), kaddadot (Igorot), pugo-pugo (Central Bisaya). Thailand: yaa kok dok khaao (Bangkok), yaa hua mong (Surat Thani). Vietnam: b[a]j c d[aaf]u l[as] ng[aws]n.

**Distribution** Throughout the tropical and warm temperate regions of the world; very common throughout Malesia.

**Uses** The pounded rhizome is used as poultice for sores; it has anti-inflammatory properties. A decoction of the whole plant is used as diuretic and against malaria. Leaves are taken internally against diarrhoea. It is sometimes used as a fodder.

**Observations** A perennial herb with rhizome creeping horizontally under or close to the ground surface, stem 3–40(–50) cm long; leaves 1–3 mm wide, canaliculate, scabrid on the margins in the upper part, grass-green; inflorescence usually consisting of a single terminal head, involucre bracts (2–)3–4(–6), up to 6(–20) cm long, spikelets closely



*Cyperus brevifolius* (Rottb.) Hassk. - 1, plant habit; 2, inflorescence enclosed by involucre bracts; 3, spikelet; 4, nuts.

packed, 1(-2)-flowered; stamens 1-2(-3), stigmas 2; fruit biconvex, laterally compressed, obovoid or ellipsoid, yellowish-brown. *C. brevifolius* is extremely variable. It occurs up to 1500(-1900) m altitude in grasslands, along roads, in forest clearings and on river banks.

**Selected sources** 202, 332, 580, 722, 760, 1178.

### *Cyperus cyperoides* (L.) O. Kuntze

Rev. Gen. Pl. 3, 2: 333 (1898).

**Synonyms** *Mariscus sieberianus* Nees ex Clarke (1893).

**Vernacular names** Indonesia: jukut bebalean (Sundanese), suket lumbungan (Javanese), tetemung (southern Sumatra). Malaysia: menderong ekur tupai, rumput janggut baung, rumput mesiyang (Peninsular). Papua New Guinea: kaiga (Ialibu, Southern Highlands). Philippines: mangilang, kupiupi (Subanum), okokiang (Bontok). Thailand: yaa rang-kaa (Loei).

**Distribution** Throughout the tropics of Africa, Asia and Australia, also in the West Indies, but

there probably introduced; throughout Malesia.

**Uses** *C. cyperoides* may be used as a vermifuge; some occult uses are known in Papua New Guinea. Sometimes it is used as a fodder.

**Observations** A perennial herb with a very short rhizome, lacking stolons, stem 20-75 cm long; leaves 3-6 mm wide, flat, scabrid in the upper part; inflorescence usually simple, involucre bracts 5-10, up to 30 cm long, rays 5-17, up to 6(-10) cm long, spikelets linear; stamens 3, stigmas 3; fruit trigonous, linear, slightly curved, rufous to chestnut-coloured. *C. cyperoides* occurs up to 1800 m altitude in grassland, roadsides, in forest clearings and in secondary forest and thickets; it is a common weed in gardens.

**Selected sources** 202, 580, 597, 722, 1178.

### *Cyperus diffusus* Vahl

Enum. pl. 2: 321 (1805).

**Vernacular names** Indonesia: cekeng (Javanese), pukul sedapan (Riau). Malaysia: rumput bumbat, rumput chukor karbau, parah-parah (Peninsular). Philippines: tuhog-dalag (Tagalog), singao (Mindanao), barsanga-bakir (Iloko). Thailand: yaa teen kaa (Nakhon Ratchasima). Vietnam: c[or] glaaslu n[uw]lows[c].

**Distribution** From India, southern China and Indo-China to the Solomon Islands; throughout Malesia except the Lesser Sunda Islands.

**Uses** It has been reported that in the Philippines (Mindanao) the roots are used to treat diseased lips. In Thailand, the roots are used as an antipyretic, analgesic, cardiotoxic, diuretic and against coughing.

**Observations** A perennial herb with a very short rhizome, stem 15-80 cm long; leaves 5-10(-20) mm wide, channelled at the base, scabrid on the margins, grass-green above, pale green below; inflorescence decomposed, diffuse and lax, involucre bracts 4-10, up to 50 cm long, primary rays up to 20, unequal, up to 20 cm long, spikelets digitately arranged, (4)-6-12(-20)-flowered; stamens 3, stigmas 3; fruit triquetrous, ellipsoid or subpyramidal, dark brown to dusky black. *C. diffusus* occurs up to 700 m altitude in thickets and moist forests on river banks, and on shady roadsides.

**Selected sources** 722.

### *Cyperus halpan* L.

Sp. pl. 1: 45 (1753; 'haspan').

**Vernacular names** Indonesia: rembang (Sumatra), para-para (Riau), rumput kudung (Kalimantan). Malaysia: rumput sumbu, rumput

bilis jantan (Peninsular). Philippines: bala-balangutan (Tagalog), barsanga (Iloko), bungot-bungot (Panay Bisaya). Thailand: kok naa (Trang), yaa kok chaai (Prachin Buri). Vietnam: c[us] c[ow]m.

**Distribution** Tropical and subtropical regions of the world; common throughout Indo-China, Thailand and Malesia.

**Uses** In Malaysia, smoke from the pith is used as febrifuge and a decoction of the plant is used internally against shingles. The pith has also been used for lamp wicks. *C. halpan* is sometimes used as fodder.

**Observations** A perennial herb, often flowering in the first year, with a short rhizome and reddish roots, stem usually slender, 10-40(-100) cm long; leaves 2-5 mm wide, flat, smooth or scaberulous at the top; inflorescence compound or decomposed, involucre bracts 2-3, up to 10(-15) cm long, primary rays up to 20, up to 15(-20) cm long, spikelets digitately arranged, 10-30(-40)-flowered; stamens 1-3, stigmas 3; fruit trigonous, broadly obovoid, whitish, later yellowish. Two subspecies are distinguished: subsp. *halpan* with slender and short stems, 1 stamen and small nut, and subsp. *juncooides* (Lamk) Kük. with more robust stems, (2-)3 stamens and slightly larger nut; the former is a common weed, the latter prefers more natural habits. *C. halpan* occurs up to 1900(-3150) m altitude in open wet locations such as muddy places in swamps; it is a characteristic weed in lowland rice fields.

**Selected sources** 202, 722.

### *Cyperus iria* L.

Sp. pl. 1: 45 (1753).

**Vernacular names** Indonesia: babawangan (Sundanese), rumput jekeng kungit (Javanese), rumput silupak (Sumatra). Philippines: sud-sud, alinang (Bikol), okokiang (Bontok). Thailand: kok huadaeng (Singburi), yaa kok saai (Nakhon Sawan), yaa kok lek (Ang Tong). Vietnam: c[os]i g[aj]o.

**Distribution** From Iran, Afghanistan, China and Japan to Australia, also in eastern Africa; common throughout Indo-China, Thailand and Malesia. Introduced and naturalized in the south-eastern United States and the West Indies.

**Uses** In India the plant is used as tonic, stimulant, stomachic, febrifuge, astringent, and for amenorrhoea. In Vietnam, the whole plant is used to treat rheumatism, to regulate menstruation; the rhizomes are used as a diuretic. It is sometimes used as fodder.

**Observations** An annual or sometimes perennial herb with fibrous, yellowish-red roots, stem (5-)15-50(-80) cm long; leaves 3-6(-8) mm wide, flat or channelled, scabrid on the margins in the upper part; inflorescence simple or compound, involucre bracts 3-5(-7), up to 40 cm long, primary rays 3-5(-8), very unequal, up to 10(-18) cm long, spikelets spicately arranged, 6-20(-24)-flowered; stamens 2-3, stigmas 3, minute; fruit triquetrous, obovoid or ellipsoid, shining dark brown to black. *C. iria* occurs up to 700(-1200) m altitude in open wet locations and is a characteristic weed in lowland rice fields, but it may also occur as a weed in vegetables, maize, sugar cane, groundnut and soya bean. It also occurs on roadsides and river banks.

**Selected sources** 202, 614, 722, 1178, 1465.

### *Cyperus kyllingia* Endl.

Cat. horti Vindob. 1: 94 (1842).

**Synonyms** *Kyllingia monocephala* Rottb. (1773).

**Vernacular names** Indonesia: jukut pendul bodas (Sundanese), melaran (Javanese), kembilikembili (Batak). Malaysia: rumput teki, rumput butang (Peninsular). Philippines: anuang (Tagalog), borobotones (Bisaya), borsa-nga-dadakkal (Iloko). Cambodia: smao kak kdam. Thailand: yaa tumhu (Bangkok), yaa nuai faai (Nakhon Si Thammarat). Vietnam: c[or] b[aj]c d[aa]u, c[or] n[us]t [as]o.

**Distribution** Common in tropical and warm-temperate Asia, less common in tropical Africa and Australia and rare in South America; common throughout Malesia.

**Uses** In India, the roots are considered an antidote to poisons. In Indonesia, they are used against diarrhoea and measles, and sometimes as a poultice for skin problems. In the Philippines, a decoction of the rhizomes is employed as a diuretic and, mixed with oil, it is applied externally to combat certain forms of dermatosis. In Vietnam, a poultice of the whole plant is used in the treatment of ulcers, sore throat, bacillary dysentery and fever. In Guam, the roots are used as abortifacient. *C. kyllingia* is sometimes used as a fodder.

**Observations** A perennial herb with horizontally creeping rhizome, stem 5-45 cm long; leaves 2-4(-5) mm wide, canaliculate, scabrid on the margins in the upper part, grass-green; inflorescence consisting of a terminal head with usually 1-3 much smaller, sessile heads at the base, involucre bracts 3-4, up to 30 cm long, spikelets very closely packed; stamens 3, stigmas 2; fruit biconvex, laterally compressed, oblong or oblong-

obovoid, yellowish-brown to chestnut-coloured. *C. kyllingia* occurs up to 1750 m altitude in open grassland, waste places, roadsides and regrowth.

**Selected sources** 190, 202, 580, 722, 760, 1035, 1178.

### *Cyperus malaccensis* Lamk

Tabl. encycl. 1(1): 146 (1791).

**Vernacular names** Indonesia: darengdeng (Sundanese), kumbu (Sumatra), rumput kuluwing (Sulawesi). Malaysia: menderong darat (Peninsular). Philippines: balangot (general), бага-as (Panay Bisaya), tikog (Manobo). Thailand: yaa saam liam (Bangkok). Vietnam: l[as]c, c[os]i.

**Distribution** From Irak through India to southern China, northern Australia and Polynesia; throughout Malesia.

**Uses** The rhizomes are used for post-partum treatment and against oedemas in Vietnam; they are considered to be diuretic. The stems are commonly used for making ropes, baskets, mats, hats and slippers.

**Observations** A perennial herb with stout stolons clothed with dark brown scales hardening into a woody rhizome, stem 60–175 cm long; leaves 5–10(–18) mm wide, scabrid at the top; inflorescence compound or subdecompound, involucre bracts 3–4, up to 30 cm long, primary rays 3–6(–10), very unequal, spikelets spicately arranged, 16–20(–40)-flowered; stamens 3, stigmas 3; fruit trigonous, slightly compressed dorsally, narrowly oblong, dark brown to black. *C. malaccensis* grows in moist localities, usually within the influence of salt or brackish water; it is an abundant colonizer on mud flats in estuaries and on sandy shores.

**Selected sources** 190, 202, 580, 722, 1035.

### *Cyperus rotundus* L.

Sp. pl. 1: 45 (1753).

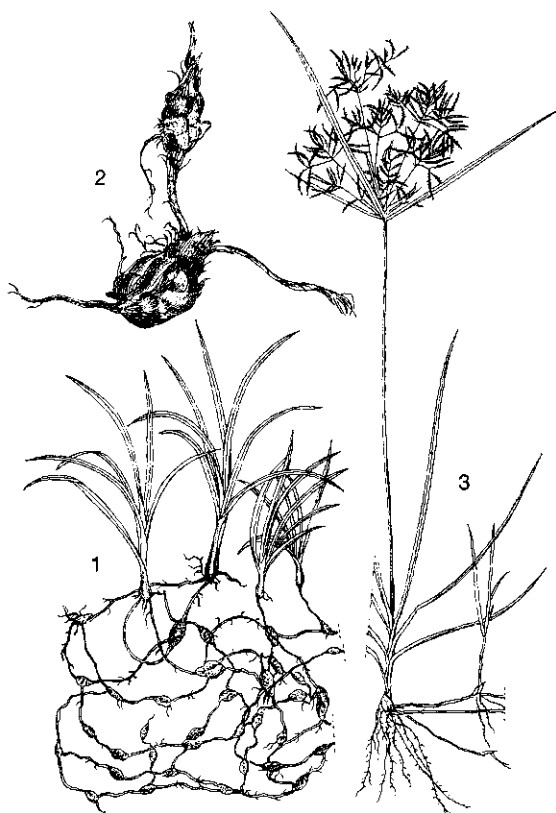
**Vernacular names** Purple nut grass, purple nut sedge (En). Souchet rond (Fr). Indonesia: teki (general), mota (Madura), karelawai (Sumba). Malaysia: teki, rumput haliya hitan. Philippines: mutha (Tagalog), ahos-ahos (Bisaya), boto-botones (Bikol). Burma (Myanmar): monhnyin-bin. Cambodia: kráva:nh chru:k. Laos: hèwz hmu:. Thailand: yaa haeo muu, yaa khon muu (general). Vietnam: c[ur] g[aa]s[u], h[uw] [ow]ng ph[uj].

**Distribution** *C. rotundus* is thought to originate from Africa. Now it is widely distributed throughout the warmer parts of the world and it is very common all over South-East Asia.

**Uses** *C. rotundus* is widely used medicinally for

various diseases and complaints (see the genus entry). It is also used as a fodder. The tubers are sometimes eaten. The oil extracted from the tubers was formerly used to make soap.

**Observations** A perennial herb with long, slender, stout, wiry, dark brown rhizomes giving rise at intervals of 5–25 cm to tubers, forming tuber chains, tubers subglobose or ellipsoid, 0.5–2.5 cm long, white and succulent when young, turning fibrous brown-blackish, stem slender, 15–30(–75) cm long; leaves 2–6 mm wide, flat, scabrid on the margins in the upper part; inflorescence simple or compound, involucre bracts 2–4(–6), up to 30 cm long, primary rays 3–9, very unequal, up to 10 cm long, spikelets spicately arranged, 10–40(–100)-flowered; stamens 3, stigmas 3; fruit trigonous, oblong-obovoid, brown to black, rarely maturing. Two subspecies are distinguished: subsp. *rotundus* and subsp. *retzii* (Nees) Kük. Subsp. *retzii* (synonym: *C. retzii* Nees) differs from subsp. *rotundus* by its stouter habit (stem 50–75 cm tall), the somewhat broader spikelets (about 2.5 mm



*Cyperus rotundus* L. - 1, young plant with rhizomes and tubers; 2, tubers; 3, flowering plant.

wide when ripe) and the paler elliptical-oblong glumes, 3.5–4 mm long; it appears in moist localities, sometimes as a weed, but never as a pest. Subsp. *rotundus* occurs up to 1000 m altitude in open grasslands, roadsides and waste places, and is often a serious pest in cultivated land, e.g. in lowland rice, maize, sugar cane and vegetables.

**Selected sources** 72, 202, 350, 414, 580, 597, 614, 664, 722, 757, 761, 930, 1035, 1178, 1315, 1454, 1514, 1533, 1558, 1559, 1563.

### **Cyperus stoloniferus Retz.**

Observ. bot. 4: 10 (1786).

**Vernacular names** Philippines: apulid-gapang (Tagalog). Vietnam: c[ur] g[aas]u bi[eer]n, h[uw] [ow]ng ph[uj] bi[eer]n.

**Distribution** Madagascar, Mauritius, whole South-East Asia, but only locally in the Philippines, to Melanesia and northern Australia.

**Uses** The tuber is used in India as a stomachic and heart stimulant. In Vietnam, *C. stoloniferus* is used as a substitute for *C. rotundus*, but has in general a stronger effect. It is considered useful as a sand binder.

**Observations** A perennial herb with long, creeping stolons hardening into a woody rhizome, forming stout tubers, stem 15–50 cm long; leaves 1.5–4 mm wide, scabrid in the upper part, glaucous; inflorescence simple or subcompound, involucre bracts 2–3, up to 30 cm long, primary rays 2–5, very unequal, 1–6 cm long, spikelets spicately arranged, 8–20-flowered; stamens 3, stigmas 3; fruit dorsoventrally compressed, broadly ovoid to ovoid, shining dark brown to blackish. *C. stoloniferus* is fairly common on coastal sands of dunes and beaches and occasionally occurs in saline, muddy locations.

**Selected sources** 202, 722, 1178, 1533.

Nguyen Khac Khoi

### **Datura L.**

Sp. pl. 1: 179 (1753); Gen. pl. ed. 5: 83 (1754).

SOLANACEAE

$x = 12$ ; *D. metel*:  $2n = 24, 48$ , *D. stramonium*:  $2n = 24$

**Major species** *Datura metel* L.

**Vernacular names** Thorn apple (En). Pomme épineuse, stramoine (Fr). Indonesia: kecubung. Malaysia: kechubong. Philippines: talong-punay (Tagalog). Thailand: ma khuea ba (northern), lamphong (central). Vietnam: c[af]d[ooj]c d[uw] [ow]c.

**Origin and geographic distribution** *Datura* consists of approximately 10 species. It is believed to have originated in the southern part of North America (Mexico) but has since been introduced throughout the world. Some species have a long history in the Old World; the first references to them are from the middle of the 16th Century, and it is probable that they were introduced soon after the discovery of America. Only *D. metel* and *D. stramonium* are found in South-East Asia.

**Uses** One of the main uses of *D. metel* in South-East Asia is for relieving asthma by smoking the dried leaves (and stems) or flowers. These 'asthma cigarettes' have been shown to be very efficient in some cases, but in other cases they had little or no effect. However, indiscriminate use can have serious, or even fatal consequences. Other traditional uses include the treatment of haemorrhoids, boils and sores, skin diseases, rheumatism, headache, toothache, cholera, parasites such as ringworm, and as an anaesthetic. In India, *D. metel* has similar medicinal applications, and it is used additionally to treat hydrophobia, syphilis, inflammations of the breasts, epilepsy, convulsion, smallpox, mumps and leprosy. The Chinese use the flowers as anaesthetic and to treat asthma, cough, eruptions and swellings, and as a pain reliever. In Africa, *D. metel* is used as an abortifacient and to treat asthma.

In South-East Asia, *D. stramonium*, which is only found in Java, where it occurs locally, is sometimes used in traditional medicine to treat fatigue, pain and for curing sprains. Elsewhere, e.g. in India, Europe and Africa, it is used similarly to *D. metel*.

*Datura* species contain (–)-hyoscyamine and (–)-scopolamine as the major alkaloids. The natural occurring, pharmacologically active (–)-hyoscyamine is easily racemized, yielding (±)-hyoscyamine, commonly known as atropine. Its pharmacological effect is the same as that of (–)-hyoscyamine, but it must be used in double doses, because the (+)-isomer is practically inactive. (–)-Hyoscyamine and atropine have an anti-cholinergic (parasympatholytic) action. Modern therapeutic uses of these alkaloids therefore include: as a remedy against spasms in skeletal muscles, the urinary tract (e.g. in the treatment of renal colic), and the respiratory tract (asthma, bronchitis), for the suppression of secretions (e.g. overproduction of hydrochloric acid in the stomach, and as pre-anaesthetic to reduce salivation and respiratory secretions), as an antidote (treatment of cholinesterase-inhibitor poisoning, e.g. by nerve gases or

certain insecticides), and in some special cases of Parkinsonism, acute myocardial infarction and bradycardia. (-)-Hyoscyamine and atropine are also available in eye drops for the treatment of uveitis and as a diagnostic aid (mydriatic) in certain eye examinations.

Like (-)-hyoscyamine, (-)-scopolamine has an anti-cholinergic effect, but unlike (-)-hyoscyamine, it is also a central nervous system depressant. Its uses include treatment of certain painful spasms, as a component of pre-anaesthetic medication and in ophthalmology, but the current main use is for the prevention of motion sickness (often applied as a skin patch behind the ear). The derivative scopolamine-butylbromide is frequently used in the treatment of gastro-intestinal spasms and renal or biliary colic.

The usefulness of *Datura* preparations in the treatment of asthma, and the route of application (cigarettes) is questionable and obsolete. A derivative, ipratropium bromide (isopropyl-atropine), is now commonly used instead. As an inhalation, it appears to be very effective in the treatment of chronic bronchitis, but somewhat less effective in asthma.

Hallucinogenic uses of *Datura* spp. are common among native tribes in Central and South America, but are apparently uncommon in South-East Asia. Although comparatively few cases of death have been recorded, its experimental use by, for example, adolescents, is very dangerous because the user often tries to harm himself or others. Criminals have used *Datura* to stupefy their victims. In Thailand, the seeds with a high concentration of scopolamine are used for this purpose.

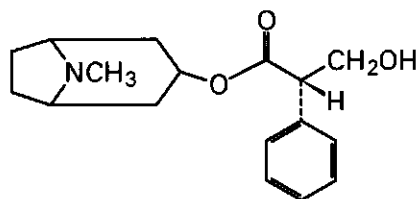
*Datura* is also used in veterinary medicine, e.g. to control ectoparasites. *D. metel* has been shown to be efficient as a pesticide. Extracts have been used successfully against the larvae of the lepidopterous *Plecoptera reflexa*, which is a serious pest in *Dalbergia sissoo* Roxb. ex DC. plantations in India, and against cotton pathogens. They also reduce insect damage in stored wheat and maize, and are useful to control the root-knot nematode *Meloidogyne javanica*. *D. metel* is commonly cultivated as an ornamental, particularly the forms with double and purplish flowers.

**Production and international trade** No trade statistics are available for *Datura*, although it is occasionally used for industrial extraction of tropane alkaloids. At the beginning of the 1980s the market prices of scopolamine hydrobromide and hyoscyamine sulphate were approximately

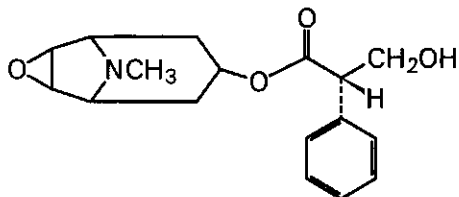
US\$ 850/kg and US\$ 470/kg, respectively. In South-East Asia, *Datura* is only traded on a small scale on local markets.

**Properties** The drugs used consist of the dried leaves, sometimes mixed with flowering tops and sometimes with fruits of *Datura* spp. The leaves are often rolled, wrinkled, agglomerated or broken in commercial samples. They have a characteristic, disagreeable odour and a slightly nauseous, bitter taste. The drug is rich in minerals (15–18%), and it must be emphasized that it is very toxic. The drug is generally used for the preparation of galenicals and not for the industrial extraction of alkaloids. The *D. stramonium* powder listed in the Dutch Pharmacopoeia (8<sup>th</sup> edition) is titrated to contain 0.23–0.27% total alkaloids. It is an ingredient of antitussive syrups, but is mostly used in the form of cigarettes to relieve respiratory difficulties, together with other drugs.

The concentration of total alkaloids in the leaves of *D. stramonium* is normally 0.2–0.5%, with (-)-hyoscyamine and (-)-scopolamine (= (-)-hyoscine) as the major compounds and apatropine, tropine, belladonnine and (-)-hyoscyamine-N-oxide as minor compounds. Biosynthetically these so-called tropane alkaloids are derived from the amino acid ornithine. An alkaloid content of about 0.6% has been reported for the flowers and fruits. The leaves of *D. metel* contain about 0.2–0.6% total alkaloids, the flowers 0.1–0.8% and the seeds 0.2–0.5%. (-)-Scopolamine is by far the chief constituent (up to 75% of the total alkaloid amount in mature leaves), together with (-)-hyoscyamine,



(-)-hyoscyamine



(-)-scopolamine



(-)-norhyoscyamine, (-)-norscopolamine, hydroxy-6(-)-hyoscyamine and meteloidine. Although also atropine ((±)-hyoscyamine) often has been reported to be present in *Datura*, it is very unclear whether it is really a natural product or an artefact obtained during isolation. Simply choosing different conditions throughout alkaloid extraction procedures (e.g. using sodium hydroxide instead of ammonia) can easily cause (-)-hyoscyamine to racemize into atropine.

Atropine, (-)-hyoscyamine and (-)-scopolamine are anti-cholinergics (parasympatholytics); they have been the starting point from which synthetic organic chemistry has created several other anti-cholinergics. Although (-)-hyoscyamine has a stronger activity than racemic atropine (the (+)-isomer is not active), the latter is more commonly prepared and used. These compounds act by competitively and reversibly inhibiting of the transmitter acetylcholine from binding to its (muscarinic) receptors, and this antagonism leads to sympathomimetic-like effects in the organs. They increase the heart rate by suppressing vagal inhibition, induce relaxation and motor inhibition in the smooth muscle fibres, decrease secretions, and induce passive mydriasis in the eyes which is very characteristic. Although at low doses their action tends to be depressant and sedative, at high doses they cause substantial excitation: agitation, disorientation, exaggerated reflexes, hallucinations, delirium, mental confusion and insomnia. (-)-Scopolamine has a similar activity, but less marked, especially on the myocardium. It potentiates neuroleptics, improves Parkinsonism, and at high doses causes speech and locomotor difficulties, affects intellectual faculties, and may lead to coma. *Datura* is also toxic to animals such as cattle, sheep and goats. The main clinical signs are disturbances in locomotion, fasciculation, hyperaesthesia, rapid respiration and reduced water intake.

*D. stramonium* has been tested for its ability to control pests, for which purpose *D. metel* is sometimes already used successfully.

**Adulterations and substitutes** Tropane alkaloids similar to those found in *Datura* are known from numerous *Solanaceae* (e.g. hyoscyamine and scopolamine in *Atropa belladonna* L. and *Hyoscyamus niger* L.). Scopolamine is found in high quantity in *Duboisia* spp., which are used for industrial production in Australia.

**Description** Annual herbs or short-lived perennials up to 2 m tall; stem dichotomously branched. Leaves alternate, simple, often with oblique base,

entire to repand, variously toothed or lobed, petiole, exstipulate. Flowers solitary in the branch forks, erect, large, bisexual, regular, usually 5-merous, shortly pedicelled; calyx tubular, circumscissile near the base after anthesis, lobed, base accrescent; corolla funnel-shaped or trumpet-shaped, lobes short, plicate and twisted in bud, with distinct acumens, sometimes appearing 10-lobed because of the presence of interacuminal lobules, white, yellow, purplish or violet; stamens adnate to the lower third of the corolla, anthers narrowly ellipsoid, basifixed, dehiscent longitudinally; ovary superior, 2-locular, often 4-locular at base because of a false septum, style filiform, stigma saddle-shaped and 2-lobed. Fruit an ovoid to globose capsule, 2-4-celled, with slender to stout spines, frilled at the base by the calyx base, dehiscent by 4 valves from the apex or breaking irregularly, green but becoming brown, many-seeded. Seeds almost D-shaped, compressed, testa finely pitted to coarsely sculptured, usually with well-developed funicular caruncle; endosperm present; embryo curved. Seedling with epigeal germination; hypocotyl long and slender; epicotyl growing straight upward between the narrowly ovate, petiole cotyledons; first 2-5 leaves smaller than subsequent ones.

**Growth and development** The vegetative axis is restricted to the basal portion of the plant, and branching is restricted to the flowering part which forms the major part of the plant and which does not revert to vegetative growth. The flowers are closed during the day and open in the evening, and are reported to be pollinated by hawkmoths and to be largely self-fertile.

In studies in India it was found that mature leaves of about the middle of the stems of *D. metel* had the maximum alkaloid content, and that very young fruits possessed a higher content of alkaloids than later developmental stages of the fruit. In *D. metel* grown experimentally in Iran, the highest scopolamine concentrations were found in the roots (0.2%) of 16-week-old plants, and in the stems (0.3%) and young leaves (0.25%) of 6-week-old plants.

It is known that the hyoscyamine/scopolamine ratio in *D. stramonium* is influenced by the developmental stage reached by the plants. In younger plants scopolamine is the main alkaloid, whereas hyoscyamine mostly becomes the dominant alkaloid when flower development has started. It is known that alkaloids are produced in the roots and transferred to the leaves, flowers and finally the fruits.

**Other botanical information** *Brugmansia* is considered here as a separate genus, although it is often treated as merely a section of *Datura* (sect. *Brugmansia* (Persoon) Bernh.). It mainly differs in its habit (a woody, comparatively long-lived arborescent shrub or small tree), its mode of growth (reproducing vegetatively by root suckers), its pendulous or inclined flowers open throughout anthesis for 4–6 days with spathe-like, not circumscissile calyx and long pedicel, and its fruit being a usually indehiscent berry with unarmed pericarp. Chemically, *Datura* and *Brugmansia* are similar, and consequently they have similar medicinal applications. However, the primary use of the *Brugmansia* species is as an ornamental.

*D. metel* is easily confused with other *Datura* species, especially *D. innoxia* Miller and sometimes also *D. stramonium*, but is characterized by the very short spines on the fruit and the glabrous stem and leaves. There is some evidence that *D. innoxia* can be successfully cultivated in the Philippines; it is a source of scopolamine and can be useful as antispasmodic, and is an important medicinal plant, for instance in India.

**Ecology** *Datura* species are usually found in more open locations: in grassland, roadsides, waste places, scrub vegetation and open forest. They tolerate various soil types but prefer clayey or loamy soils. *D. stramonium* can be a serious weed, e.g. in soya bean. The total alkaloid content of *D. metel* has been shown to peak in the hot and dry season in Nigeria, and it was lowest in the rainy season.

**Propagation and planting** *D. metel* is generally cultivated from seed sown either directly in the field or in a nursery bed. Soaking seed for one night in water may improve germination. Per hectare, 7–8 kg of seed is needed. Seed starts germinating after about 2 weeks, and germination is complete after one month. If the seed is sown in the nursery, seedlings are transplanted when 8–12 cm tall. Effective means of vegetative reproduction, such as root suckers in *Brugmansia*, are absent in *Datura*. Normal spacing is 70–100 cm.

**In vitro production of active compounds** Tropane alkaloids are biosynthesized in cell suspension cultures of *D. stramonium* in shake flasks and bioreactors. Calluses have been induced from leaves, stems and roots and cultured on Gamborg's B5 or Murashige and Skoog medium supplemented with growth regulators. The highest alkaloid content was produced in leaf calluses grown on a medium with low concentrations of growth regulators (0.1 mg/ml of benzyladenine

and 2,4-D), and in cultures grown in the dark. In tests, the cell structures responded very well to elicitors in the late exponential phase, whereas addition of cell-wall fragments of *Phytophthora megasperma* enhanced the final tropane alkaloid yield by 5-fold compared with the control culture. Supplying carbon and nitrogen to cell cultures in the early stationary phase suppressed tropane alkaloid production at comparatively low C:N ratio, but at a C:N ratio of over 100 the final product yield was increased. Total alkaloid production in the cell culture supplemented with phenylalanine and ornithine was 5 times higher than in the control culture, and higher ratios of tropine to tropic acid also stimulated alkaloid production (at a ratio of 20, the productivity was 7 times higher than that in the control culture). A hyoscyamine production of up to 7.5 mg/l daily was recorded in root cultures on full-strength Gamborg's B5 medium containing 5% sucrose at 20–25°C.

**Husbandry** The application of a nitrogenous fertilizer increases both growth and the hyoscyamine content of *Datura* spp. As the scopolamine content is inversely related to the increase in biomass, fertilization results in accelerated decrease in scopolamine, which is the major alkaloid in young plants. Methods leading to a retarded growth by e.g. indirect sowing or thinning will lead to a retarded shift of the hyoscyamine/scopolamine ratio to hyoscyamine. However, experiments in Burundi with *D. stramonium* showed that the application of chemical fertilizer but also of manure, resulted in increased production of total alkaloids. As a result of experiments in Burkina Faso, deflowering was recommended to increase the total alkaloid content in the leaves.

**Diseases and pests** *Datura* spp. can be infected by wilt and root rot caused by *Sclerotium rolfsii*, root and foot rot caused by *Corticium solani*, leaf spot caused by *Alternaria* spp., and by several mosaic and other virus diseases with symptoms like leaf curl, retarded growth and yellowing of leaves. Some of these diseases can be transmitted to solanaceous crops like tomato.

**Harvesting** The aerial parts are usually harvested at the beginning of flowering. Plants regrow after the first harvest and can be harvested once or twice again within the same season. In experiments in Burundi it was demonstrated that the best time for harvesting leaves of *D. stramonium* was 8 weeks after sowing, because alkaloid content was then maximal. From experiments in Burkina Faso, it was recommended to harvest the leaves in the early morning or late afternoon.

**Yield** In India, yields per ha of 10.5–14.5 t of fresh plant material and 750–1200 kg of seed of *D. metel* have been reported. In Pakistan, 400 kg/ha of seed of *D. stramonium* were harvested. Studies in Turkey showed leaf yields as high as 50 t/ha. The highest yields of scopolamine and hyoscyamine from *D. stramonium* reported for Algeria were 7.5 kg/ha and 21 kg/ha, respectively.

**Handling after harvest** The foliage of *Datura* spp. is dried in the shade immediately after harvesting. The fruits are first left in the sun to dry until they are open, then threshed to remove the seeds, which are subsequently sun-dried. Dried leaves and leaf powder should be stored in well-closed containers protected from light and moisture.

**Genetic resources and breeding** Both *D. metel* and *D. stramonium* have an extremely wide geographical distribution and prefer anthropogenic habitats. Therefore, they are not easily liable to genetic erosion. The genetic base of *D. stramonium* in South-East Asia is probably rather small because it only occurs very locally in Java. In order to develop industrial plantations for alkaloid extraction, breeding should particularly focus on high alkaloid content, combined with the development of proper means of vegetative propagation.

**Prospects** Although the tropane alkaloids scopolamine, hyoscyamine and atropine can be prepared synthetically, it is more economical to extract them from plants such as *Datura* spp. However, *Atropa* and *Duboisia* species are the major sources of raw materials. Moreover, local demand in South-East Asia for the pure chemicals is too small to justify commercial production of *Datura* spp. It is likely that local demand for *Datura* will be for tincture production only. On the world market for the pure tropane alkaloids, it will be difficult to compete with producers in China and India.

**Literature** |1| Afsharypuor, S., Mostajeran, A. & Mokhtary, R., 1995. Variation of scopolamine and atropine in different parts of *Datura metel* during development. *Planta Medica* 61(4): 383–384. |2| Avery, A.G., Satina, S. & Rietsema, J. (Editors), 1959. *Blakeslee: the genus Datura*. The Ronald Press Company, New York, United States. 289 pp. |3| Backer, C.A. & Bakhuizen van den Brink Jr., R.C., 1965. *Flora of Java*. Vol. 2. Noordhoff, Groningen, the Netherlands. pp. 477–478. |4| Bruneton, J., 1995. *Pharmacognosy, phytochemistry, medicinal plants*. Lavoisier Publishing, Paris, France. pp. 647–667. |5| Demeyer, K., Vanhaste, F., van de Velde, H. & Dejaegere, R., 1990.

Introductory study for the optimization of growth and alkaloid production by cell cultures of *Datura stramonium* L. *Acta Horticulturae* No 306: 210–218. |6| Dethier, M., Cordier, Y. & Demeyer, K., 1993. Cultivation of *Datura* species for scopolamine and hyoscyamine production in Burundi. *Acta Horticulturae* No 331: 39–48. |7| Gupta, S., Prabhakar, V.S. & Madan, C.L., 1973. The distribution of total alkaloids and major components in the different organs of *Datura metel* var. *fastuosa* at various stages of growth. *Planta Medica* 23(4): 370–376. |8| Hilton, M.G. & Rhodes, M.J.C., 1993. Factors affecting the growth and hyoscyamine production during batch culture of transformed roots of *Datura stramonium*. *Planta Medica* 59(4): 340–344. |9| International Trade Centre UNC-TAD/GATT, 1982. *Markets for selected medicinal plants and their derivatives*. Geneva, Switzerland. pp. 103–105. |10| Sobti, S.N. & Kaul, B.L., 1982. Cultivation of *Datura innoxia* and *D. metel* in India. In: Atal, C.K. & Kapur, B.M. (Editors): *Cultivation and utilization of medicinal plants*. Regional Research Laboratory, Council of Scientific and Industrial Research, Jammu-Tawi, India. pp. 259–261.

#### *Selection of species*

#### ***Datura metel* L.**

Sp. pl. 1: 179 (1753).

**Synonyms** *Datura fastuosa* L. (1759).

**Vernacular names** Downy thorn apple, metel thorn apple (En). Stramoine metel, concombre diable (Fr). Brunei: kecupong. Indonesia: kecubung (general), kucubung (Sundanese), kacubung (Javanese). Malaysia: kechubong, kechubong puteh, kechubong hitam. Philippines: talong-punay (Tagalog), kamkamaulau (Iloko), katchibong (Bisaya). Cambodia: slak. Laos: mak khuea ba. Thailand: ma khuea ba (northern), lamphong (central), liak (Khmer, Surin). Vietnam: c[af] d[ooj]c d[uw][owj]c, d[uw][ow]ng kim hoa, m[aj]n d[af] la.

**Distribution** Probably of American origin; widely cultivated and naturalized in all tropical and subtropical regions. In South-East Asia, *D. metel* is reported for Indo-China, Thailand, Peninsular Malaysia (cultivated and locally naturalized in the northern part), Brunei, Java (cultivated and naturalized), the Philippines (naturalized, sometimes cultivated) and Papua New Guinea (very locally cultivated and naturalized).

**Uses** Dried leaves of *D. metel* are traditionally used as 'asthma cigarettes' to relieve asthma in



*Datura metel* L. - 1, flowering stem and young fruit; 2, fruit; 3, seeds.

South-East Asia. In Brunei, an infusion of the flowers is drunk to relieve mental disorders.

**Observations** An annual or short-lived perennial herb up to 200 cm tall, stem short-haired to glabrous; leaves ovate to angular broad-ovate, 4–25 cm × 2–20 cm, entire to repand-dentate-lobed, petiole up to 16 cm long; flowers with corolla of (12–)14–18(–20) cm long, simple or double, white or purplish; fruit deflexed when mature, with numerous conical tubercles of approximately equal length. *D. fastuosa* is sometimes considered as a species distinct from *D. metel*, but more often as a variety or cultivar. *D. metel* is often cultivated in gardens and yards (often with double and purplish corolla), but also grows wild or naturalized (usually with simple and whitish corolla) and is locally abundant in waste places, roadsides, brushwood, open grassland, teak forest and on river banks, up to 1000 m altitude.

**Selected sources** 23, 78, 87, 97, 202, 332, 350, 528, 549, 561, 562, 580, 652, 847, 964, 1035, 1126, 1178, 1277, 1433.

### ***Datura stramonium* L.**

Sp. pl. 1: 179 (1753).

**Synonyms** *Datura tatula* L. (1762).

**Vernacular names** Thorn apple, jimsonweed, devil's apple (En). Pomme épineuse, stramoine (Fr). Indonesia: kucubung leutik (Sundanese), jarak pendek, kacubung wulung (Javanese). Thailand: lanphong khao.

**Distribution** Probably of American origin; now found all over the world, but particularly common in warm temperate regions. In South-East Asia, *D. stramonium* only occurs in some locations in Java, where it is naturalized, and also in mountainous regions in Thailand.

**Uses** In Java, *D. stramonium* is an ingredient of certain traditional medicines to treat fatigue, pain and for curing sprains.

**Observations** An annual herb up to 120 cm tall, stem glabrous or nearly so; leaves rhombic to angular-ovate or ovate-oblong, 6–20 cm × 3–15 cm, exculpted-dentate-pinnatilobed, petiole up to 10 cm long; flowers with corolla 5–9 cm long, simple, white or pale purplish; fruit always erect, densely covered with strong and long prickles. *D. stramonium* is locally naturalized in Java at higher altitudes (1000–2100 m) and grows in sunny and fertile locations such as on waste land and in tea plantations.

**Selected sources** 87, 97, 112, 193, 343, 346, 549, 561, 562, 580, 588, 652, 847, 1277.

Sri Hartati, Imastini Dinuriah  
& M.M. Blomqvist

### ***Derris* Lour.**

Fl. cochinch.: 432 (1790).

LEGUMINOSAE

$x = 10, 11, 12, 13$ ; *D. acuminata*:  $2n = 22$ , *D. elegans*:  $2n = 22$ , *D. elliptica*:  $2n = 20, 22, 24, 36$ , *D. malaccensis*:  $2n = 22, 24$ , *D. robusta*:  $n = 11$ , *D. scandens*:  $2n = 26$ , *D. trifoliata*:  $n = 10, 11, 2n = 22, 24$

**Major species** *Derris elliptica* (Wallich) Benth., *D. malaccensis* (Benth.) Prain, *D. trifoliata* Lour.

**Vernacular names** Derris, tuba root (En). Touba (Fr). Indonesia: tuba. Malaysia: tuba. Philippines: tubli. Vietnam: c[os]c k[ef]n.

**Origin and geographic distribution** *Derris* consists of about 55 species found throughout the Old World tropics. Most species (approximately 50) are found in South-East Asia, 3 in Australia and one species extends from Asia to eastern Africa. *D. elliptica* and *D. malaccensis* are culti-

vated, e.g. in India, southern China, Malaysia, Indonesia, the Philippines, New Guinea, tropical Africa and America.

**Uses** The rotenoids from *Derris* constitute an effective alternative to regular synthetic insecticide applications in horticulture and agriculture. They can be applied to control a large array of pests on a range of crops. The insecticide is applied as dust, spray, dip or bait. In cabbage it is used to control the lepidopterous *Plutella xylostella* (diamondback moth) and *Pieris rapae*. It is also used to control the aphid *Myzus persicae*, which is a major pest of vegetables and peaches in China. Furthermore, it is applied against borers, thrips and seedling maggots on maize, golden apple snails (*Pomacea* spp.) on rice, against the tenthredinid *Caliroa cerasi* on apple and oriental pear (*Pyrus pyrifolia* (N.L. Burman) Nakai), against cotton stainer, black bean aphids, common cutworm, earcutting caterpillar, and against *Helluta undalis* on broccoli. The extract is also effective against pests of stored garlic and rice. In poultry farming it can be used to kill ticks, fleas, lice, mites, flies and to control sticktight fleas. It is reported to be ineffective against bedbugs, cockroaches, scale insects and red spiders.

*D. elliptica* and *D. malaccensis* are used as a fish poison throughout southern Asia and the Pacific. The pounded root is considered the strongest fish poison in South-East Asia. Rotenone is used in fisheries in the Philippines, Bangladesh and India to remove predatory and other undesired fish from rearing ponds.

*D. elliptica* is traditionally used for antisepsis and applied to abscesses and against leprosy and itch, and sometimes as an abortifacient. The root of *D. scandens* is used in India to increase milk secretion after childbirth; it is crushed with or without water and the juice is given orally. The whole plant of *D. trifoliata* is used in India as a stimulant, anti-spasmodic and counter-irritant, and the bark against rheumatism, chronic paralysis and dysmenorrhoea, and in Papua New Guinea a decoction of the roots is applied externally against fever and internally against sores. A solution of crushed leaves of *D. elegans* is used to wash snake bites in Papua New Guinea. An extract from the roots of *D. elliptica* is reported to be employed as an ingredient of arrow poison in Borneo.

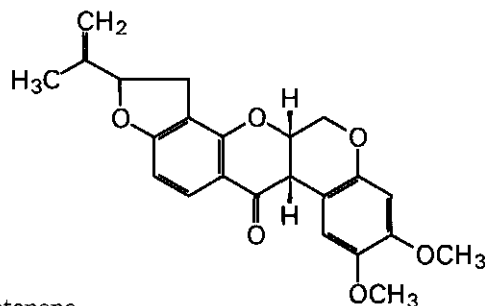
The stems are sometimes used for rough cordage. The wood of *D. robusta* trees is sometimes used for tea chests and implements. *Derris* can serve as soil improver. Lianas, including *D. elliptica*, may occur as weeds in forest plantations of

*Acacia*, *Eucalyptus* and *Swietenia*.

**Production and international trade** Until 1930 in Indonesia, *Derris* was grown as single plants near houses only. Cultivation of *Derris* was forbidden by law, to prevent fish being eradicated by too intensive use of the roots. Around 1930, several plantation companies planned to cultivate *Derris* on a large scale, to produce a pesticide for use on crops where the residues of synthetic insecticides could be harmful to the consumer. Within a few years *Derris* became an estate crop from what was practically a wild plant. In Java and Sumatra the area planted with *Derris* increased from 240 ha in 1935 to 10 000 ha in 1941. By 1947, however, no regular plantations remained, since everything had been harvested during the war and no new plantations had been established.

The main producing countries in South-East Asia are Malaysia, Indonesia and the Philippines. Imports in the United States of rotenoid-containing roots, mainly from *Derris*, exceeded 3000 t annually in the early 1950s, and in 1963 about 1500 t of crude *Derris* roots and 500 t of extract were imported. The introduction of synthetic insecticides, including pyrethrin analogues, has markedly reduced the demand for plant-based insecticides over the last 40–50 years.

**Properties** The roots are flexible and hard, with a slightly aromatic odour and somewhat bitter taste. Chemically, *Derris* species are characterized by the presence of toxic rotenoid compounds; rotenone as isoflavone derivative (0.3–12% in the root) is the most toxic compound, followed by deguelin (0.2–2.9%), elliptone (0.4–4.6%) and toxicarol (0–4.4%). These compounds are effective respiratory poisons; rotenone is a powerful mitochondrial inhibitor of electron transport. It exhibits a considerable degree of selective toxicity; it is very toxic to insects but only slightly toxic to mammals. *Derris* preparations are comparatively safe to natural enemies (parasites and predators) being used to control insect pests on crops, since



rotenone

they deteriorate when exposed to sunlight and air. To be effective they should contain not less than 3% rotenone on a dry-weight basis.

Compounds related to rotenone, but lacking the core B-C ring system have been synthesized. They are less active than rotenone, but often still have significant levels of inhibition.

The addition of soap solution (0.1%) lowers the toxic concentration of an aqueous extract of roots for snails from 2000 ppm to 100 ppm. Ethanolic and chloroform extracts cause 100% mortality at 100 ppm and 20 ppm, respectively. An ethanolic extract is more toxic than an aqueous extract because this solvent can extract more toxic constituents from the roots. However, this is not practical for use by farmers. Besides, the ethanolic extract is very toxic to fish. Rotenone at 20 ppm causes 83% and 100% mortality in snails after 24 hours and 48 hours, respectively. Rotenone-free extracts of *D. elliptica*, obtained by chloroform extraction and thermal heating were found to be still toxic to snails, but not to fish.

Studies have been performed with cultured cells in order to more fully characterize the bioactive potential of rotenone. Intense cytotoxic activity was observed in lymphocytic leukaemia, carcinoma of the nasopharynx, and a number of human cancer cells, e.g. fibrosarcoma, lung cancer, colon cancer, melanoma and breast cancer cell lines. Thus, rotenone has been evaluated as a potential antitumour agent. The growth-inhibiting effect has been demonstrated both with cultured cells and experimental tumours. Rotenone is broadly cytotoxic, but no cell-type specificity has been discerned.

Numerous other compounds have also been isolated and identified, e.g. flavonoids, deguelin and maackiain. For instance, roots of *D. elliptica* contain the rotenoid elliptinol and tubaic acid (0.01% of air-dried root). The latter compound showed anti-microbial activity, inhibiting the growth of *Bacillus subtilis*, *Staphylococcus aureus* and *Escherichia coli* at high concentrations. Other tests, however, showed no antibacterial activity of *D. elliptica* decoctions or macerations. Several amino acids (e.g. related to piperolic acid) have been isolated from its leaves, and also the amino-alcohol 2,5-dihydroxymethyl-3,4-dihydroxypyrrolidine (0.1% of fresh leaves). Compounds with a pyrrolidine ring structure might be of pharmacological interest, since some of them are known to have adrenolytic and vasodilating properties. Other related compounds have been found to possess antibacterial activity.

Nine pure compounds have been isolated from the roots of *D. robusta*: 4 isoflavones and 5 3-phenyl-4-hydroxycoumarins. The isoflavones include derubone, robustone and robustone methyl-ether; the 3-phenyl-4-hydroxycoumarins include robustic acid and robustin. Seven pure compounds have been isolated from the stems of *D. scandens*, e.g. the prenylated isoflavones warangalone, 8- $\gamma,\gamma$ -dimethylallylwighteone and 3'- $\gamma,\gamma$ -dimethylallylwighteone, and the 3-phenyl-4-hydroxycoumarin robustic acid. All the prenylated isoflavones and robustic acid were found to be potent inhibitors of the catalytic subunit of cyclic AMP-dependent protein kinase (cAK, from rat liver, in vitro). None of the compounds, however, was able to inhibit  $Ca^{2+}$ -dependent and phospholipid-dependent protein kinase C (PKC, from rat brain, in vitro). The flavonoid compound dehydrorotenone, lupeol and a straight-chain ketone have been isolated from the roots of *D. trifoliata*, but these compounds seem to have no appreciable insecticidal activity. Pentacyclic triterpenoids have also been isolated, whereas the leaves of *D. trifoliata* have yielded the flavonoid rhamnetin 3-O-neohesperidoside. Fresh leaves of *D. trifoliata* contain 25 mg/g of lipid, 3 mg/g sterol and 3 mg/g triterpene. The sterol fraction consists of 1.5% cholesterol, 7.5% campesterol, 9% stigmasterol, 21.5% sitosterol and 60.5% stigmast-7-en-3 $\beta$ -ol, the triterpene fraction of 10%  $\beta$ -amyrin, 12%  $\alpha$ -amyrin and 78% lupeol. The bark contains up to 9.5% tannin. Seeds of *D. robusta* contain the pyranoisoflavone derrone, the isoflavones derrugenin, robustigenin and 5-hydroxy-7-methoxyisoflavone, and rubone, a chalcone.

Aqueous extract of fruits of *D. trifoliata* with an  $LC_{50}$  value of 0.002–0.003 ppt showed toxicity to fish. The  $LC_{50}$  value for roots of *D. elliptica* for catfish in Bangladesh was 64–115 ppm. The toxicant in the root powder had completely detoxified in 6–7 days. Root powder is effective for eradicating predatory fishes in fish ponds at 5 ppm in fresh water and at 10–30 ppm in brackish water. *Derris* extract also has some fungicidal activity. The leaves of *D. elliptica* are said to be poisonous enough to kill cattle.

The wood of *D. robusta* is pale brown, with heartwood not distinctly demarcated from the sapwood. It is hard and heavy; the density is about 850 kg/m<sup>3</sup> at 15% moisture content.

**Adulterations and substitutes** Rotenoids are also obtained from the roots of other legumes such as *Lonchocarpus*, *Millettia*, *Piscidia* and *Tephrosia* spp. Other insecticides of plant origin used in

South-East Asia are present in seeds of *Croton tiglium* L., stem and roots of *Tinospora* spp., leaves of *Vitex negundo* L., *Nicotiana tabacum* L. and *Azadirachta indica* A.H.L. Juss. They are also found in leaves, fruits and bark of *Melia azedarach* L., whole plants of *Tanacetum cinerariifolium* (Trev.) Schultz-Bip., *Tagetes* spp. and *Lantana* spp., and in leaves, roots and seeds of *Annona squamosa* L. As a fungicide, star anise (*Illicium verum* Hook.f.) is more effective. Other piscicidal plants include *Croton tiglium*, *Myrica esculenta* Buch.-Ham. and *Sapindus saponaria* L.

**Description** Woody climbers or scandent shrubs, or sometimes trees (*D. robusta*) or erect shrubs; roots up to more than 2 m long and up to 2 cm in diameter, dark reddish-brown or greyish-brown; stem of lianas up to 20 m long and up to 10 cm in diameter, often ridged and densely lenticellate. Leaves alternate, imparipinnate with opposite leaflets, stipules small, stipels sometimes present. Inflorescence terminal or axillary, pseudoracemose or pseudopaniculate, sometimes contracted, with flowers crowded on the short ultimate branchlets or clustered at the nodes. Flowers bisexual, 5-merous; calyx tube usually cupular, almost toothless or with short teeth, the upper pair variably joined; corolla papilionaceous, much longer than the calyx, white, pink or purplish, standard often green at base, glabrous or hairy, wings adhering to the keel petals; stamens 10, united into a tube with openings at the base on either side of the upper filament; ovary superior, 1-loculate, with few ovules, style curved and tapering to a very small stigma. Fruit an oval or elliptical to linear-oblong, flattened, indehiscent pod with wings along upper edge or both edges, 1-few-seeded. Seeds usually reniform, smooth or wrinkled.

**Growth and development** *Derris* lianas may climb over trees and other vegetation forming a thick cover and thus act as a serious weed in forest plantations. *D. elliptica* may start flowering at 18 months of age. Wild plants flower and fruit normally. Pods ripen about 4 months after fertilization. The roots form nitrogen-fixing nodules. In cultivation fruiting is rare. Some cultivars like 'Ngawi' flower very rarely; others (e.g. 'Wulung' and 'Pantu') flower freely but seldom fruit.

**Other botanical information** *Derris* is placed in the tribe *Millettieae* within the subfamily *Papilionoideae*. It seems closely related to *Millettia* and *Lonchocarpus*, and to *Aganope* (or *Ostryocarpus* when this originally African genus is considered as congeneric). It is characterized by the winged

sutures of the pods. The genus is often subdivided into 3 sections: section *Derris* with about 50 species including *D. elegans* and *D. trifoliolata*, section *Brachypterum* (Wight & Arn.) Benth. with 3 species including *D. robusta* and *D. scandens*, and section *Paraderris* Miq. with 6 species including *D. elliptica* and *D. malaccensis*. The latter two sections have been raised to generic level, differing from the first one particularly in the the inflorescences and in the pods. However, new combinations for the *Derris* species involved have often not yet been made. Sometimes, a fourth section *Dipteroderris* Benth. is separated from section *Derris*. The 4 South American species closely related to *Derris* are usually classified in *Deguelia*.

Several other *Derris* species not discussed above have been mentioned as a fish poison and are occasionally used as an insecticide: *D. acuminata* Benth. (synonym: *D. pubipetala* Miq.), *D. multiflora* Benth. and *D. montana* Benth. for Indonesia, *D. philippinensis* Merr. for the Philippines, and *D. amoena* Benth. and *D. polyantha* Perk. for Peninsular Malaysia. *D. ferruginea* (Roxb.) Benth. is used in India as an insecticide.

Several cultivars of *D. elliptica* have been selected of which 'Sarawak Creeping', 'Changi No 3' and 'Ngawi' are reportedly commercially superior. Other cultivars include 'Pantu', 'Wulung', 'Putih' and 'Kotari'. They differ mainly in hairiness and leaflet shape, but also in rotenoid content, yield and susceptibility to diseases. 'Sarawak Erect' has been reported as the *D. malaccensis* cultivar with the highest content of rotenoids.

**Ecology** *Derris* grows best in regions with an annual precipitation of 2300–3300 mm and a mean annual temperature of 29°C. *D. elliptica* can survive dry periods of up to 4 months. This species is often confined to low altitudes, but locally (e.g. in Java) it can be found up to 1500 m altitude. *Derris* can be grown on a range of soils varying from coarse sand to heavy clay, but swampy and stony soils are unsuitable. *Derris* is sensitive to waterlogging. It prefers a rich friable loam and tolerates a pH 4.3–8. It is often found on river banks, in brushwood, forest borders and secondary forest. *D. trifoliolata* occurs near the coast in or near mangroves.

**Propagation and planting** Ripe dry seeds cannot be stored for long without losing their viability. They germinate immediately after sowing. For commercial production, however, woody stem cuttings 30–45 cm long and 0.5–1.5 cm in diameter and with 3 or more buds are used. In tests in Central America cuttings from mature stems with

a diameter of 2 cm and a length of 20–30 cm with at least 2 nodes gave the best results in rooting. Single-node cuttings can be employed if treated with root-inducing substances such as naphthalene acetic acid (0.2%). The cuttings are often first planted in nursery beds, to a depth of 15–20 cm. Shading and regular watering are needed during the dry season, and during the first weeks the beds should be kept free from weeds. When cuttings have 2 normal leaves, the shade is gradually removed. The cuttings are ready for transplanting into the field when about 6 weeks old, and are preferably transplanted in the rainy season. They are planted either in furrows 10 cm deep or in separate planting holes. The planting distance is 0.7–1 m × 0.7–1 m. Direct planting into the field is sometimes also practised, and should also be done in the rainy season. In Indonesia, *Tephrosia noctiflora* Bojer ex Baker is sometimes used as a shade during the first year, then pulled out and applied as a soil cover.

When planted specially for the production of cuttings, *Derris* is trained on a fence. Each year, these plantations produce enough material to plant an area at least ten times larger. The area to be planted can easily be enlarged at harvest by using cuttings from the branches of an established plantation.

In tests in the Philippines, tissue culture of *D. elliptica* was found to be not viable due to low explant decontamination, retarded shoot regeneration for alcohol propagation and unsuccessful shoot regeneration from calli.

#### **In vitro production of active compounds**

Tissue culture to obtain rotenoids appears to be rather difficult. When leaves of *D. elliptica* were used for callus induction, only trace amounts of rotenoids (3 µg/g on dry weight basis) were detected in the tissue subcultured for 4 months. Rotenoid biosynthesis decreased with frequent subcultures of callus tissue, and was finally lost. Callus with imperfectly differentiated rootlets induced from the leaves or stems by regulating plant hormones contained rotenoids which were identified as rotenone and deguelin. The rotenone content was 160 µg/g on dry weight basis. In other experiments in the Philippines root regeneration from calli induced from internodes, nodes and axillary buds of *D. elliptica* was obtained 3–5 weeks after inoculation on Shenck and Hildebrandt's medium supplemented with naphthalene acetic acid. Using Durham's test for rotenone, 60–100% of the calli and regenerated roots showed a positive reaction.

**Husbandry** *Derris* can be raised as a sole crop, or as an intercrop e.g. with rubber, kapok, coconut and cocoa. It needs full sunlight, however, so shading by the main crop should be avoided. Fertilizer application should be during the rainy season. The crop requires high K and P but low N. Trellising considerably increases the number of usable cuttings and the yield of roots.

The crop remains in the field for two years if the trailing cropping method is used. As the crop does not cover the soil completely during the first year, great care should be taken to prevent soil erosion. Hillsides are not suitable and the soil should be covered. To facilitate harvesting, soils must not be too heavy.

**Diseases and pests** Some fungal diseases are reported to damage planted *D. elliptica*: a rust (*Ustilago derrides*), a *Gloeosporium* sp. that causes the shoot tips to die, and an unidentified fungal disease that attacks cuttings in nursery beds. Pests are not serious and are easily controlled.

**Harvesting** In cultivation, the roots are often harvested before the plants flower, usually 2 years after planting. They generally grow no deeper than about 50 cm and are dug out carefully, taking care to minimize damage to the bark. On large plantations, the harvested area is cleaned and then replanted. In small plantings with trellised plants all roots are removed, except for those directly under the plants, leaving the top part intact. This root-pruning practice enables several harvests from the same plant. Regeneration, however, is slow.

**Yield** The yield of dried *D. elliptica* roots is 1100–1800 kg/ha, occasionally up to 3000 kg/ha, particularly when plants are trellised. The weight of a fresh *D. malaccensis* root is up to 900 g. Yields of rotenoids reported in literature vary widely, depending on species and cultivar, age of the plants and ecological conditions.

**Handling after harvest** After harvesting, the roots are cleaned, preferably in running water, and rapidly dried in the sun or in an oven at approximately 50°C to about 10% moisture content. Drying can be speeded up by cutting the roots into pieces up to 5 cm long. The roots can be stored in a cool and dry place. However, drying the roots seems to degrade the active constituent, particularly when stored too wet.

Rotenoid content is highest in roots 2–10 mm in diameter. So it is advisable to sort the roots into two groups before packing, those smaller and those larger than 1 cm in diameter. The roots are pressed into blocks of 100 kg or, if cut into chips,



packed in bags of 50 kg. The packing material should be waterproof because a high moisture content will cause rapid deterioration. *Dinoderus minutus* and *Sinoxylon anale* beetles feed on the dried roots and can cause considerable damage. Protection is possible by fumigation. If the product is to be used in dusting or spraying, the dried roots are ground into a fine powder. The powder remains effective for a long time if it is protected against air, sunlight and moisture.

Proper ventilation, the use of masks and strict attention to hygiene are a prerequisite when manufacturing *Derris* insecticides to prevent complaints such as dermatitis and irritation of mucous membranes.

Fresh roots are usually fermented with water for 24 hours to obtain an aqueous solution of the toxic principle. For application against snails in wet rice *Derris* roots are chopped and crushed, and then scattered over the field for at least 24 hours. This procedure is suitable for rice fields without fish. Often a solution is prepared for spraying. For this, chopped and crushed roots are first soaked in water (90 kg of roots per ha in 200–300 l water) with 0.1% soap solution and subsequently sieved and squeezed. The solution is boiled for one hour and immediately sprayed. It should not be kept for more than 24 hours. If it rains or the water level increases, the treatment should be repeated after 3–5 days.

For use in ponds against predatory fish, roots are chopped into pieces of 2–3 cm and ground (e.g. with a laboratory pulverizer) and then sieved to obtain the root powder, which can be used to selectively poison fish without killing prawns.

To use *Derris* as a fish poison in the Solomon Islands, leaves are put into a hole together with an equal amount of sand and pounded. The resulting granular green mixture is then spread under water, and poisoned fish exhibiting extreme disorientation are speared or collected from the surface of the water.

**Genetic resources and breeding** In South-East Asia, several *Derris* species are widely distributed as wild plants, but some species (particularly *D. elliptica* and, to a lesser extent, *D. malaccensis*) have been cultivated in gardens since antiquity. This has resulted in the present situation where, e.g. in Java, wild plants of *D. elliptica* vary widely but have a low rotenone content (about 0.5%), whereas the cultivated plants vary little but have a high rotenone content (12–13%). Collections of both provenances are available. Hybrids between *D. elliptica* and *D. malaccensis*

have shown promising results. Breeding trials have been hampered by the almost total self-incompatibility or cross-incompatibility of most cultivars of *D. elliptica*.

**Prospects** There is a renewed interest in the use of *Derris* as a pesticide because of the problems arising from repeated application of synthetic chemical insecticides, which are, however, often cheaper and more effective. Preparing a solution for use in the field is a simple procedure that can be done by the farmer. Moreover, the active compounds are completely detoxicated within one week and the effect on the environment is limited. *Derris* roots and their extracts are cheaper and more potent than pyrethrum, and are simple and convenient to apply. It has been reported that they have low toxicity for higher animals. However, determination of the toxicity of the insecticide to mammals and other non-target organisms needs further investigation. Because of the lack of convincing data, *Derris* insecticide has been banned in some countries (in the Netherlands, for instance) since about 1980.

**Literature** [1] Blasko, G., Shieh, H.-L., Pezzuto, J.M. & Cordell, G.A., 1989. <sup>13</sup>C-NMR spectral assignment and evaluation of the cytotoxic potential of rotenone. *Journal of Natural Products* 52(6): 1363–1366. [2] Council of Scientific and Industrial Research, 1952. *The wealth of India*. Volume 3. New Delhi, India. pp. 35–41. [3] Duke, J.A., 1981. *Handbook of legumes of world economic importance*. Plenum Press, New York and London. pp. 73–77. [4] Geesink, R., 1989. *Derris elliptica* (Sweet) Benth. In: Westphal, E. & Jansen, P.C.M. (Editors): *Plant Resources of South-East Asia, A selection*. Pudoc Wageningen, the Netherlands. pp. 112–114. [5] Guerrero, R.D., Guerrero, C.A. & Garcia, L.L., 1990. Use of indigenous plants as sources of fish toxicants for pond management in the Philippines. *Philippine Technology Journal* 15(2): 15–17. [6] Kodama, T., Yamakawa, T. & Minoda, Y., 1980. Rotenoid biosynthesis by tissue culture of *Derris elliptica*. *Agricultural and Biological Chemistry* 44(10): 2387–2390. [7] Maini, P.N. & Morallo-Rejesus, B., 1993. Molluscicidal activity of *Derris elliptica* (Fam. Leguminosae). *Philippine Journal of Science* 122(1): 61–75. [8] Rickard, P.P. & Cox, P.A., 1986. Use of *Derris* as a fish poison in Guadalcanal, Solomon Islands. *Economic Botany* 40(4): 479–484. [9] Spoon, W. & Toxopeus, H.J., 1950. *Derriswortel* [*Derris* root]. In: van Hall, C.J.J. & van de Koppel, C. (Editors): *De landbouw in de Indische Archipel* [Agriculture in the Indonesian Archipelago]. Vol.

3. Van Hove, 's-Gravenhage, the Netherlands. pp. 578-608. [10] Verdcourt, B., 1979. A manual of New Guinea legumes. Botany Bulletin No 11. Office of Forests, Division of Botany, Lae, Papua New Guinea. pp. 314-331.

#### *Selection of species*

#### ***Derris elegans* Graham ex Benth.**

Miq., Pl. jungh.: 252 (1852).

**Vernacular names** Malaysia: akar tuba (Peninsular). Papua New Guinea: imora (Kabali, Central Province).

**Distribution** India, Indo-China, Thailand, throughout Malesia, the Solomon Islands and the New Hebrides.

**Uses** *D. elegans* has been employed as a fish poison in New Guinea and possibly Peninsular Malaysia. A solution of crushed leaves in (green) coconut water is used in Papua New Guinea to wash snake bites.

**Observations** A liana or scandent shrub up to 10 m long, branches purplish-brown; leaflets 3-5(-7), glabrous or adpressed hairy below; inflorescence axillary or fascicled on older branches; flowers with brown, pink or red calyx and white corolla, standard without callosities at base, glabrous or pubescent at extreme tip; fruit elliptical or oblong-elliptical, with a narrow wing along one side, finally orange-brown. Three varieties are distinguished: var. *elegans* (synonym: *D. rufula* Lauterb. & K. Schumann) occurring throughout the area of the species, var. *gracillima* (Hemsley) Verdc. (synonyms: *D. cauliflora* Pulle, *D. momiensis* Kanehira & Hatusima, *D. papuana* Pulle) from the Moluccas, New Guinea, the Solomon Islands and the New Hebrides, and var. *vestita* (Baker) Prain (synonym: *D. vestita* Baker) from Malesia. *D. elegans* occurs in forest along the coast, on coastal plains, in sago swamps and in secondary and riverine forest, up to 600 m altitude; often on alluvial soils, but also on limestone soils.

**Selected sources** 202, 597, 1520.

#### ***Derris elliptica* (Wallich) Benth.**

Journ. Linn. Soc. 4, Suppl.: 111 (1860).

**Vernacular names** Derris, tuba root (En). Tuba (Fr). Brunei: tuba (Dusun, Malay). Indonesia: tuba, oyod tungkul (Javanese), tuwa leteng (Sundanese). Malaysia: tuba, akar tuba. Philippines: tubli, tugling-pula (Tagalog), upei (Bontok). Burma (Myanmar): hon. Cambodia: ca bia, k'biehs. Thailand: hang lai daeng, lai nam (northern),

kalamphoh (Phetchaburi). Vietnam: d[aa]y m[aa]jt, d[aa]y thu[oo]s]c c[as].

**Distribution** Bangladesh, Burma (Myanmar), Indo-China, Thailand, the Nicobar Islands and Malesia (apparently not wild in Borneo, Sulawesi and the Moluccas); cultivated in South-East Asia, India, tropical Africa and America.

**Uses** The powdered root of *D. elliptica* is widely used as an insecticide and fish poison. In Thailand, the roots are also used as an emmenagogue and the stems as a blood tonic. An extract of the root is reported to be used as an ingredient of dart poison in Borneo.

**Observations** A liana up to 16 m long, root reddish-brown, apical shoots often leafless for several m and rusty pubescent; leaflets 7-15, mostly densely rusty hairy on both surfaces when young; inflorescence axillary or fascicled on older branches; flowers with rusty pubescent calyx and pinkish corolla, standard with basal callosities, rusty silky hairy; fruit oblong or oblong-elliptical, with a narrow wing along both sides. *D. elliptica* is common-



*Derris elliptica* (Wallich) Benth. - 1, root system; 2, leaf; 3, inflorescences and young leaves; 4, pod.

ly found in forest edges, roadsides and along rivers, in Java up to 1500 m altitude.

**Selected sources** 56, 97, 190, 202, 287, 377, 494, 514, 580, 756, 782, 887, 1035, 1062, 1116, 1178, 1205, 1226, 1277, 1471, 1504, 1520, 1563.

**Derris malaccensis (Benth.) Prain**

Journ. Asiat. Soc. Bengal 66: 107 (1897).

**Synonyms** *Deguelia malaccensis* (Benth.) Blake (1929).

**Vernacular names** Malaysia: tuba merah (Peninsular). Vietnam: c[os]c k[ef]n.

**Distribution** Throughout Malesia; cultivated outside Malesia in India, southern China and tropical America.

**Uses** The roots of *D. malaccensis* are used as insecticide and fish poison.

**Observations** A liana up to at least 15 m long, root greyish-brown, young shoots adpressed pubescent; leaflets 5-9, glabrous above, adpressed pubescent beneath; inflorescence axillary; flowers with glabrous pink calyx and whitish or pinkish corolla, standard with basal callosities, glabrous; fruit oblong, with a narrow wing along both sides, rarely without wings. *D. malaccensis* often occurs in riverine rain forest, up to 250 m altitude.

**Selected sources** 202, 287, 377, 1226, 1277, 1520.

**Derris robusta (Roxb. ex DC.) Benth.**

Journ. Linn. Soc. 4, Suppl.: 104 (1860).

**Synonyms** *Derris polyphylla* (Miq.) Benth. (1860), *Brachypterum robustum* (Roxb. ex DC.) Dalz. & Gibs. (1861).

**Vernacular names** Indonesia: bekel, kedusan, wedusan (Javanese). Thailand: khang sai chang (Phitsanulok), kheemoot (Saraburi), haang khaao (northern). Vietnam: c[os]c k[ef]n m[aj]nh.

**Distribution** India, Burma (Myanmar), Thailand, Vietnam and Java.

**Uses** *D. robusta* is occasionally used as a shade tree and a green manure. The wood is used in India for e.g. tea chests, and locally in Java for handles of axes. The leaves may serve as fodder. Although it is chemically well investigated, no medicinal, insecticidal or piscicidal uses are known.

**Observations** A tree up to 20(-25) m tall, with bole up to 30(-65) cm in diameter, tops of branches finely pubescent; leaflets (7-)11-31, puberulous on both surfaces when young but often glabrescent above; inflorescence axillary; flowers with densely adpressed pubescent calyx and whitish or pale pink corolla, standard without basal callosities; fruit strap-shaped, with a wing

along one side. In Java, *D. robusta* occurs in deciduous forest, especially teak forest, up to 300 m altitude.

**Selected sources** 97, 287, 580.

**Derris scandens (Roxb.) Benth.**

Journ. Linn. Soc. 4, Suppl.: 103 (1860).

**Synonyms** *Brachypterum scandens* (Roxb.) Benth. (1838).

**Vernacular names** Hogcreeper (En). Indonesia: bendan, gobul (Javanese), sobi (Madura). Philippines: malasaga (Tagalog), lapak (Bikol). Thailand: khrua khao nang (Nakhon Ratchasima), thaowan priang (central), phan sanai (Chumphon). Vietnam: c[os]c k[ef]n leo.

**Distribution** From India to northern Australia; throughout Malesia, but apparently absent in New Guinea.

**Uses** *D. scandens* is used as a fish poison. In Thailand, the stems are used as a diuretic, laxative, expectorant, emmenagogue and in the treatment of common cold and backache. It is sometimes cultivated as an ornamental.

**Observations** A liana up to 20 m long or scandent shrub with drooping branches, young branches finely pubescent; leaflets (7-)9-13(-19), puberulous beneath; inflorescence axillary; flowers with pubescent purplish calyx and whitish or pinkish corolla, standard without basal callosities, glabrous or hairy on the back; fruit narrowly oblong or strap-shaped, with a broad wing along one side. *D. scandens* occurs in light forest and brushwood in the lowland (up to 200 m altitude in Java).

**Selected sources** 97, 202, 287, 580, 1009, 1542.

**Derris trifoliata Lour.**

Fl. cochinch.: 433 (1790).

**Synonyms** *Derris uliginosa* (Willd.) Benth. (1852), *Derris heterophylla* (Willd.) Backer ex K. Heyne (1927).

**Vernacular names** Indonesia: areuy ki tonggeret, tuwa areuy (Sundanese), gadel (Javanese). Malaysia: tuba bekut (Peninsular). Papua New Guinea: gamo (Zimanki, Western Province). Philippines: silasila, asiasimanan (Tagalog), butong (Bisaya). Thailand: khwaep thale, phak thaep (central), thopthaep thale (Phetchaburi). Vietnam: long k[es]n, c[os]c k[es]n n[uw][ows]c, d[aa]y c[os]c.

**Distribution** Eastern Africa, Madagascar, Mascarene Islands, South and South-East Asia including the whole of Malesia, the Pacific and Australia.

**Uses** *D. trifoliata* is used in local medicine in India as a stimulant, antispasmodic and counter-irritant, and against rheumatism, chronic paralysis and dysmenorrhoea, and in Papua New Guinea a decoction of the roots is used externally against fever and internally against sores. Thai traditional doctors use roots or stems as a laxative, carminative and anti-arthritis treatment. It serves as a fish poison. The leaves are sometimes used as fodder.

**Observations** A liana up to at least 15 m long, sometimes an erect shrub, branches soon becoming glabrous, dark red; leaflets 3-5(-7), variably glabrous; inflorescence terminal and axillary; flowers with glabrous, green calyx and white, pale pink or pale mauve corolla, standard without basal callosities, glabrous; fruit oblong or elliptical to broadly oval, with a narrow wing along one side. *D. trifoliata* grows near the coast in swampy scrub vegetation and forest behind beaches, on muddy foreshores and estuaries, and in edges of mangroves, sometimes in pure stands.

**Selected sources** 97, 202, 580, 597, 1035, 1178, 1520.

Auzay Hamid

## Desmodium Desv.

Journ. Bot. appl. 1: 122 (1813).

LEGUMINOSAE

$x = 10, 11$ ; *D. adscendens*, *D. gangeticum*, *D. microphyllum*, *D. triflorum*:  $2n = 22$

**Major species** *Desmodium adscendens* (Sw.) DC., *D. gangeticum* (L.) DC., *D. styracifolium* (Osbeck) Merr., *D. triflorum* (L.) DC.

**Vernacular names** Tick clovers (En). Vietnam: h[af]n the.

**Origin and geographic distribution** *Desmodium* consists of about 300 species found in the tropical and subtropical regions of Africa, Central and South America, East Asia and Oceania; some species occur in temperate regions. The centre of origin of the genus is South-East Asia. The highest species diversity is found in an area stretching from India eastward to western and south-western China and Malesia (25 species reported). Mexico is a second centre of diversity.

**Uses** *Desmodium* shows a wide range of medicinal uses. In South-East Asia, they are considered diuretic (*D. gangeticum*, *D. repandum*, *D. styracifolium*); other prominent uses are the treatment of diarrhoea, dysentery and stomach-ache (*D. difusum*, *D. gangeticum*, *D. heterophyllum* (Willd.)

DC., *D. triflorum*, *D. velutinum*), wounds, ulcers and other skin problems (*D. gangeticum*, *D. triflorum*, *D. sequax*), stones in the gall bladder, kidneys or bladder (*D. gangeticum*, *D. styracifolium*) and headache, toothache or other pains (*D. gangeticum*, *D. microphyllum*, *D. ormocarpoides*, *D. sequax*).

In the Philippines, a decoction of *D. triflorum* is used as a mouthwash and as an expectorant. In India, fresh leaves of *D. triflorum* are used internally as a galactagogue; and in Taiwan, the whole plant is used against fever, rheumatism, jaundice and gonorrhoea.

*D. auricomum* Grah. ex Benth. and *D. caudatum* (Thunb. ex Murray) DC. are reportedly used in local medicine in Indo-China, but no specific uses are mentioned. *D. adscendens*, *D. incanum* DC. (synonym: *D. canum* Schinz & Thell.) and *D. renifolium* Schindler do occur in South-East Asia, but reports of medicinal use are only available from outside the region. *D. incanum* is used as a diuretic, stomachic, febrifuge and hemostatic in Central America. *D. renifolium* is used as a febrifuge in Taiwan.

*D. heterocarpon* (L.) DC. and *D. heterophyllum* are primarily forages, but are also used medicinally in Malesia. The boiled roots of *D. heterocarpon* are used in Malaysia to poultice sore breasts, and a decoction of the plant is regarded as a tonic and a bechic. In Cambodia, the stems of *D. heterocarpon* subsp. *angustifolium* H. Ohashi are applied to fractures and snake bites. In Taiwan, a decoction of the roots is used against rickets in children. *D. heterophyllum* is applied in Malaysia to treat sores, earache, stomach-ache and abdominal complaints. In India, the roots are considered carminative, tonic and diuretic, the leaves are used as a galactagogue, and a decoction of the whole plant is used to treat stomach-ache and abdominal problems.

*Desmodium* contains a considerable number of species used as pasture and fodder crops, species used for ground cover and green manure; some of these have medicinal applications as well.

**Properties** *Desmodium* is very rich in alkaloids and related amino compounds, biosynthetically derived from different precursors.

From *D. gangeticum*, the following alkaloids and nitrogen-containing compounds have been isolated: hypaphorine, N,N-dimethyltryptamine, N,N-dimethyltryptamine-N<sub>6</sub>-oxide, 5-methoxy-N,N-dimethyltryptamine, 5-methoxy-N,N-dimethyltryptamine-N<sub>6</sub>-oxide (alkaloids, all derived from the amino acid tryptophan), N<sub>6</sub>-methyl-tetrahydroharman, 2-methyl-6-methoxy-β-carbolinium cation

(harman-type alkaloids, derived from the amino acid tryptophan), hordenine (= N,N-dimethyltyramine), N-methyltyramine, candicine (alkaloids derived from the amino acid tyrosine),  $\beta$ -phenylethylamine (amine, structurally related to tyramine) and choline (quaternary nitrogen compound). Of these alkaloids, hordenine is known to increase the urinary flow, and to be a remedy for diarrhoea and dysentery.

Other reported constituents are the pterocarpanoids (isoflavonoids) gangetin, gangetinin and desmodin. Gangetin isolated from the roots of *D. gangeticum* has been found to adversely affect the fertility and reproductive system of male rats. Gangetin caused a dose-dependent impairment of fertility, reduced the vaginal sperm count and enhanced pre-implantation losses. Aqueous root extracts have shown mild diuretic action, relaxant effects on intestine muscles of rats and dogs, and antibacterial, antifungal and anti-inflammatory activity. The alkaloid fraction of stems and leaves has shown relaxant (curariform) effects on frog rectal muscles. Methanolic extracts of Nigerian *D. gangeticum* plants have shown in vitro anti-leishmanial activity.

*D. styracifolium* contains triterpenoid saponins, of which soyasaponin I has been found to be effective against kidney stones. In a study with rats the extract inhibited the formation of calcium oxalate stones in kidneys by increasing the output of urine, decreasing the excretion of calcium and increasing the urinary excretion of citrate. Aqueous extracts have shown in vivo hypotensive action in rats through cholinergic receptor stimulation and autonomic ganglion and  $\alpha$ -adrenoreceptor blockade. Furthermore, the extract relaxed isolated methoxamine precontracted helical tail artery strips and was positive chronotropic without apparent effect on the contractile force. The flavonoid fraction exhibited hypotensive activity.

From the leaves of *D. adscendens*, alkaloids (tetrahydroisoquinolones, derived from tyrosine), amines ( $\beta$ -phenylethylamines, indole-3-alkylamines) and triterpenoid saponins (dehydrosoyasaponin I, soyasaponin I and soyasaponin III) have been isolated. The three saponins, especially dehydrosoyasaponin I, are potent potassium channel openers. This may be related to the reported use of *D. adscendens* against asthma and dysmenorrhoea, because opening of K-channels is expected to cause smooth muscle of the lung and uterus to relax. *D. adscendens* extracts have also been reported to inhibit the synthesis and release of histamines, prostaglandins and arachidonic acid. A

hot water extract causes a dose-dependent reduction in the amount of spasmogens released anaphylactically and in anaphylactic-induced contractions of ileal muscle in guinea-pigs. Ethanolic leaf extracts have shown analgesic and hypothermic activity in mice. In addition, they delayed the onset of pentylentetrazole forelimb clonus, and general seizures induced by kainic acid.

Alcoholic extracts of *D. triflorum* showed good in vitro anthelmintic activity against human *Ascaris lumbricoides*. Alkaloids and related nitrogen-containing compounds reported from *D. triflorum* (roots and/or leaves) include: hypaphorine, hypaphorine-methylester, N,N-dimethyltryptamine-N<sub>1</sub>-oxide, S-(+)-N,N-dimethyltryptophan-methylester (alkaloids derived from tryptophan), hordenine (= N,N-dimethyltyramine), 3,4-dihydroxyphenylethyl-trimethyl-ammoniumhydroxide (alkaloids derived from tyrosine), trigonelline, S(-)-stachydrine, tyramine,  $\beta$ -phenylethylamine, choline and betaine. The total alkaloid fraction has shown anti-spasmodic, sympathomimetic, central nervous system stimulant and curare-mimetic activity. Other reported constituents of *D. triflorum* include: indole-3-acetic acid, the flavonoid-C-glycosides vitexin, isovitexin and 2'-O-glucosylvitexin, the flavone apigenin, and the polyhydric alcohol (+)-pinitol.

From *D. caudatum* N,N-dimethyltryptamine, bufotenine, bufotenine-N-oxide (alkaloids derived from tryptophan), swertisin (flavonoid-C-glycoside) and the flavone desmodol have been isolated. From ethanolic root extracts of *D. incanum*, three anti-microbial isoflavones (desmodianones A, B and C) have been isolated, which have shown in vitro activity against *Bacillus subtilis*, *Mycobacterium smegmatis*, *Staphylococcus aureus* and *Streptococcus faecalis*.

**Description** Herbs, shrubs or subshrubs, rarely trees, mostly erect or ascendent, but frequently decumbent or subclimbing. Leaves alternate, 3-foliolate in most species, but frequently 1- and 3-foliolate, occasionally exclusively 1-foliolate and rarely 5-7-foliolate, petiole pulvinate, stipulate; leaflets variously shaped, lateral leaflets usually smaller than terminal one, chartaceous to coriaceous, reticulately veined, with 1 stipel at the base of lateral and 2 at the base of terminal leaflets. Inflorescence mostly racemose or paniculate, rarely fasciculate, terminal or terminal and axillary; bracts dimerous or rarely monomerous, early deciduous, primary bracts larger than secondary ones, usually narrowly ovate, secondary bracts mostly subulate or narrowly ovate; bracteoles pre-

sent or absent. Flowers pedicellate, calyx usually broadly campanulate, 4–5-lobed; corolla variously coloured, often pink to pale purple, papilionaceous, standard usually broadly obovate to almost orbicular, rounded or emarginate at the apex, not auriculate, sometimes short-clawed, wings short-clawed, with oblong lamina, rounded to obtuse at the apex, keel acute or obtuse at the apex; androecium monadelphous or diadelphous, stamens 10, anthers basifixed; ovary superior, narrowly oblong, sessile or stipitate, often many-ovuled, style inflexed or incurved, stigma terminal or lateral, capitate or minute. Fruit a jointed pod, often narrowly oblong or linear, flat or rarely turgid, mostly indehiscent, usually with straight or hooked hairs; articles usually elliptical to quadrangular. Seeds transversely broadly elliptical or broadly ovate to elliptical or depressed ovate, flat or turgid, hilum lateral. Seedling usually with epigeal germination; cotyledons thin leaf-like; first two leaves opposite and simple, subsequent ones alternate and similar to leaves of adult plants.

**Growth and development** *Desmodium* appears to be predominantly self-pollinating. However, when flowers are touched, they spring open and release pollen, which makes outcrossing possible. *D. diffusum*, *D. gangeticum*, *D. microphyllum*, *D. repandum*, *D. sequax*, *D. styracifolium*, *D. triflorum* and *D. velutinum* are reported to have nodulating ability.

**Other botanical information** *Desmodium* belongs to the subtribe *Desmodiinae* of the tribe *Coronilleae* within the subfamily *Papilionoideae*. The genus *Desmodium* in the broad sense has been divided into seven genera: *Codariocalyx*, *Dendrolobium*, *Desmodium* s.s., *Dicerma*, *Hegnere*, *Phyllodium* and *Tadehagi*. It is difficult to classify *Desmodium* s.s., because of the morphological variation and the continuity of morphological features across species borders; further taxonomic studies are needed. Here, the narrow sense of *Desmodium* is assumed.

**Ecology** *Desmodium* is mainly found in humid to sub-humid regions of the tropics and subtropics, on acid soils (pH < 6.5). The usual habitats are open woodland and forest clearings. In equatorial regions, *Desmodium* species are found from sea-level up to 3000 m altitude. The photoperiod sensitivity varies with the species.

**Propagation and planting** In general, propagation of *Desmodium* is by seed. The degree of scarification necessary for successful germination varies between species. The information available concerns species used as a forage or cover crop,

and no detailed information is available on species used for medicinal purposes.

**Diseases and pests** *Desmodium* is affected by a range of diseases. On a worldwide base, important pathogens are the fungi *Synchytrium desmodii* and *Phanerochaeta salmonicolor*, causing wart and pink disease respectively, the root-knot nematodes *Meloidogyne arenaria*, *M. hapla*, *M. incognita* and *M. javanica*, and the stem gall nematode *Pterotylenchus cecidogenus*. South-East Asian reports include the sooty moulds *Meliola bantamensis*, *M. bicornis* and *M. scabriseta* var. *integra* on *D. gangeticum*. A very wide range of diseases has been reported for *D. gangeticum* in India. *D. adscendens* has been reported to be resistant to attacks of *Meloidogyne* species.

**Genetic resources and breeding** The major germplasm collections of *Desmodium* in the world are found in Australia (CSIRO), Brazil (EMBRAPA and IRI), Colombia (CIAT) and the United States of America (University of Florida). In these collections, only a quarter of the known *Desmodium* species are present, and many of these with only a single entry. There have been only a few plant breeding efforts in *Desmodium*, and knowledge of the extent of heritable variation is limited. Interspecific hybridization is possible, but the degree of success depends on how closely related the parent species are.

**Prospects** *Desmodium* shows a broad range of traditional medicinal uses, some of which have already been related to the presence and activity of specific compounds. Further research on possible uses of the various *Desmodium* species seems worthwhile. Further taxonomic studies and germplasm collection are needed to exploit the potential of this genus.

**Literature** |1| Dy Phon, P., Ohashi, H. & Vidal, J.E., 1994. Légumineuses - Desmodiées [Leguminosae (Fabaceae) Papilionoideae - Desmodieae]. In: Flore du Cambodge, du Laos et du Viêt Nam [Flora of Cambodia, Laos and Vietnam]. Vol. 27. Muséum National d'Histoire Naturelle, Paris, France. pp. 62–142. |2| Ghosal, S. & Banerjee, P.K., 1969. Alkaloids of the roots of *Desmodium gangeticum*. Australian Journal of Chemistry 22: 2029–2031. |3| Ghosal, S., Srivastava, R.S., Bhattacharya, S.K. & Debnath, P.K., 1973. *Desmodium* alkaloids IV: chemical and pharmacological evaluation of *D. triflorum*. Planta Medica 23(4): 321–329. |4| Imrie, B.C., Jones, R.M. & Kerridge, P.C., 1983. *Desmodium*. In: Burt, R.L., Rotar, P.P., Walker, J.L. & Silvey, M.W. (Editors): The role of *Centrosema*, *Desmodium* and *Stylosanthes*

in improving tropical pastures. Westview Press, Boulder, Colorado, United States. pp. 97–140. |5| Kubo, T., Hamada, S., Nohara, T., Wang, Z., Hirayama, H., Ikegami, K., Yasukawa, K. & Takido, M., 1989. Study on the constituents of *Desmodium styracifolium*. *Chemical and Pharmaceutical Bulletin* 37(8): 2229–2231. |6| Lenné, J.M. & Stanton, J.M., 1990. Diseases of *Desmodium* species – a review. *Tropical Grasslands* 24: 1–14. |7| McManus, O.B., Harris, G.H., Giangiacomo, K.M., Feigenbaum, P., Reuben, J.P., Addy, M.E., Burka, J.F., Kaczorowski, G.J. & Garcia, M.L., 1993. An activator of calcium-dependent potassium channels isolated from a medicinal herb. *Biochemistry* 32(24): 6128–6133. |8| N'gouemo, P., Baldy-Moulinier, M. & Nguemby-Bina, C., 1996. Effects of an ethanolic extract of *Desmodium adscendens* on central nervous system in rodents. *Journal of Ethnopharmacology* 52(2): 77–83. |9| Ohashi, H., 1973. The Asiatic species of *Desmodium* and its allied genera (Leguminosae). *Ginkgoana* no. 1. 318 pp. |10| 't Mannetje, L. & Jones, R.M. (Editors), 1992. *Plant Resources of South-East Asia No 4. Forages*. Pudoc Scientific Publishers, Wageningen, the Netherlands. pp. 106–118.

#### *Selection of species*

#### ***Desmodium adscendens* (Sw.) DC.**

Prodr. 2: 332 (1825).

**Synonyms** *Hedysarum adscendens* Sw. (1788), *Desmodium oxalidifolium* G. Don (1832), *Desmodium trifoliolatum* Miq. (1855).

**Vernacular names** Papua New Guinea: roabe (Kurtachi, Bougainville). Philippines: pega pega. Vietnam: bai ngai.

**Distribution** Native in South America and Africa; introduced throughout South and South-East Asia and Melanesia.

**Uses** The leaves of *D. adscendens* are used in Papua New Guinea to treat stomach-ache. It is used against asthma and other diseases associated with smooth muscle contraction in Ghana, against fever, pain, epilepsy and stomach-ache in Congo. In Central America a decoction of the plant is used as a laxative and to treat convulsions, and to soothe urinary disorders in cases of venereal diseases. It was originally introduced in Asia as a cover crop in plantations.

**Observations** A creeping or ascending perennial herb or low shrub, up to 100 cm tall, stems terete, glabrescent; leaves 3-foliolate, stipules obliquely ovate-lanceolate, persistent; terminal

leaflet elliptical-obovate, (1.5–)2–4(–5.5) cm × 1–3 cm, larger than the lateral leaflets, upper surface glabrous, lower surface pilose, lateral veins 4–7, not reaching the margin; inflorescence a terminal or axillary raceme, 4–20 cm long, lax-flowered; flowers in pairs, calyx 5-lobed, covered in hairs, corolla white or purple to violet, androecium diadelphous; pod 10–25 mm × 3–4 mm, 3–6-jointed, dehiscent, densely covered in hairs; seeds flattened ellipsoid, 2.5–5.0 mm × 1.5 mm. *D. adscendens* is found in humid locations, provided that they are shady, e.g. on stream banks and bunds of rice fields, at 200–1000 m altitude.

**Selected sources** 14, 15, 16, 17, 97, 407, 979, 1069, 1120, 1520.

#### ***Desmodium diffusum* DC.**

Ann. Sci. nat. 4: 100 (1825).

**Synonyms** *Desmodium recurvatum* (Roxb.) Wight & Arn. (1834), *Desmodium laxiflorum* DC. subsp. *parvifolium* H. Ohashi & Chen (1983), *Desmodium laxiflorum* auct. non DC.

**Vernacular names** Laos: nha tük hma. Thailand: nieo maa. Vietnam: nha khau mau ri.

**Distribution** From India eastward to Burma (Myanmar), Thailand, Indo-China, China, Taiwan and throughout the Malesian region.

**Uses** In Laos, a decoction of the roots is used to treat stomach-ache in children. In India, it is used as a fodder and has been tried as a green manure.

**Observations** A prostrate or erect undershrub, 30–100 cm tall, branches hairy in the upper parts; leaves 3-foliolate, sometimes mixed with 1-foliolate leaves at the base of the branches, stipules narrowly triangular, 6–11 mm long, terminal leaflet ovate to broadly ovate, 2.5–11 cm × 1.5–6 cm, larger than the lateral ones, chartaceous, upper surface rather hairy, lower surface covered with silky hairs, lateral veins 7–9, extending to the margin; inflorescence a terminal and axillary raceme, 10–30 cm long; flowers in clusters of 3–5, calyx 4-lobed, hairy, corolla blue or violet, androecium diadelphous; pod linear, 3–3.8 cm × 0.2 cm, 5–9-jointed, articles asymmetrically oblong, covered with hooked hairs; seeds 2.5–3 mm × 1.1–1.3 mm. *D. diffusum* is found along roadsides, in grassland and forest margins, up to 2000 m altitude.

**Selected sources** 97, 287, 389, 629, 921, 1069.

#### ***Desmodium gangeticum* (L.) DC.**

Prodr. 2: 327 (1825).

**Synonyms** *Hedysarum gangeticum* L. (1753), *Meibomia gangetica* (L.) O. Kuntze (1891).

**Vernacular names** Indonesia: daun bulu ayam (Malay, Manado), daun picah (Sundanese), waliketupa (Javanese). Malaysia: akar katah, serengan, kemani bali. Philippines: dikit-dikit (Tagalog), pega-pega (Cebu Bisaya), andudukut (Sulu). Laos: tük hma:, do:yz tük hma:, ph'è:ng kh'am h'o:yz. Thailand: i-nio, yaa tuet maeo, nuut phra phuu (Trang). Vietnam: [c]aaly th[os]c l[es]p, c[or] ch[as]y, b[af]i ng[af]i.

**Distribution** Tropical Africa, tropical and subtropical Asia, throughout the Malesian region and Oceania; introduced in the West Indies.

**Uses** In Java, a decoction of the leaves is used against stones in the gall bladder, kidneys or bladder. In Malaysia, a decoction of the root is employed to treat diarrhoea or as a sedative for children. The roots are applied to the gums against toothache, and the leaves to the head against headache. In Vietnam, the roots are considered diuretic and prescribed in case of oedema. A decoction is used externally to clean wounds and ulcers. It is also used in Indo-China to treat coughs, swellings, dysentery and kidney problems. In Thailand, the roots are used as a diuretic and the whole plant as an anthelmintic. In India, the roots are reported to be applied as a febrifuge, bitter tonic, expectorant, alterative, diuretic and astringent, and in the treatment of diarrhoea, chronic fever, biliousness, snake bite and poisoning. The fibrous stems are reported to be useful for paper production, and in the Philippines they are used to make prawn traps. Conflicting information exists on its usefulness as a green manure.

**Observations** A much branched, erect shrub, or a prostrate to ascending subshrub 30–200 cm tall, rootstock thickened, young stems hairy; leaves 1-foliolate, stipules narrow and tapering to a point, 7–15 mm long, leaflet (1–)2.5–18 cm × (0.8–)2–8 cm, very variable in shape and size, chartaceous, upper surface variously hairy, lower surface densely hairy, lateral veins 6–12, usually extending to the margin, persistent; inflorescence a terminal or axillary raceme or panicle, 10–30 cm long; flowers in clusters of 2–4, calyx 4-lobed, densely covered with minute hooked hairs and longer straight hairs, corolla white to pale yellow or rose to violet, androecium diadelphous; pod linear, variably incurved, 1.2–2.5 cm × 2–2.5 mm, (4–)6–8-jointed, covered with minute hooked hairs, articles broadly oblong; seeds pale yellow, reniform, 1.5–1.8 mm × 2–2.5 mm. *D. gangeticum* is a very common weed in Malesia, mainly found in anthropogenic habitats in the lowlands, under everwet or seasonal conditions. In Indo-China it is



*Desmodium gangeticum* (L.) DC. – 1, plant habit; 2, branch with infructescence; 3, flower; 4, mature pod.

found in savannas and deforested terrains, in hedges and along forest paths, up to 1900 m altitude.

**Selected sources** 97, 119, 202, 287, 389, 481, 533, 580, 629, 661, 752, 819, 820, 921, 1035, 1069, 1120, 1168, 1178, 1393, 1476, 1520.

***Desmodium microphyllum* (Thunb. ex Murray) DC.**

Prodr. 2: 337 (1825).

**Synonyms** *Hedysarum microphyllum* Thunb. ex Murray (1784), *Desmodium parvifolium* DC. (1825).

**Vernacular names** Laos: hnha:z phak kè:b. Vietnam: tr[af]ng qu[ar] ba hoa.

**Distribution** From India and Sri Lanka eastward to China, Taiwan and Japan and southward to Malesia and Australia; throughout and common in the Malesian region.

**Uses** In Indo-China the whole plant is used in the treatment of eye problems and headache.

**Observations** A slender, much branched, erect



or creeping undershrub, up to 150 cm tall, stems glabrescent, terete; leaves 3-foliolate, usually partly 1-foliolate, stipules narrowly triangular to narrowly ovate, 2.4–4.5 mm long, terminal leaflet elliptical or obovate, 3–25 mm × 1–7 mm, larger than the lateral ones, thinly chartaceous, upper surface almost glabrous, lower surface sparsely hairy, lateral veins 4–5 on either side of the midrib, not extending to the margin; inflorescence terminal, racemose, 1–5 cm long, lax-flowered; flowers solitary, calyx very deeply 5-lobed, densely hairy, corolla variably coloured, keel longer than the wings, androecium diadelphous; pod 5–13(–25) mm × 2.5–3(–4) mm, (1–)3–4-jointed, flat, nearly glabrous or moderately hairy, articles obliquely broadly rhombic or elliptical, length 1–1.5 times the width; seeds 1.2–1.5 mm × 2–2.1 mm, arillate. *D. microphyllum* is a very common weed in the Malaysian region, and is indifferent to rainfall distribution. In Indo-China it is found at 0–2000 m altitude.

**Selected sources** 97, 389, 629, 752, 1120, 1126, 1520.

### **Desmodium ormocarpoides DC.**

Prodr. 2: 327 (1825).

**Synonyms** *Hedysarum adhaerens* Poir. (1817), *Hedysarum ormocarpum* Desv. ex Poir. (1817).

**Vernacular names** Papua New Guinea: agagil (Yunamami, East New Britain), digambi (Gaulim, East New Britain).

**Distribution** Sulawesi, the Moluccas, the Lesser Sunda Islands, New Guinea, Australia (Queensland), the New Hebrides and New Caledonia.

**Uses** In Papua New Guinea, the leaves are macerated in water and the solution is drunk to treat malaria and tuberculosis. The solution is also reported to be effective against pains and itches. Roots are chewed to prevent vomiting.

**Observations** A climbing shrub or undershrub, up to 150 cm tall, young stems covered with hooked hairs; leaves 1-foliolate, stipules narrowly triangular, 3.8–4.3 mm long, leaflet elliptical or ovate, 8–20 cm × 3.5–8 cm, chartaceous to coriaceous, both surfaces glabrous, lateral veins 6–8, prominent, not extending to the margin; inflorescence a terminal or axillary raceme, usually 15–70 cm long, lax-flowered; flowers in clusters of 2–4, calyx 4-lobed, minutely hairy, corolla white, androecium diadelphous, style spirally twisted; pod linear, stalked, 3–5-jointed, article length 4 times the width, separated by very narrow necks, covered with minute hooked hairs; seeds linear-ob-

long, 12 mm × 2 mm. In New Guinea, *D. ormocarpoides* is found at altitudes up to 1800 m, in savanna, scrubland, forest edges, plantations and roadsides.

**Selected sources** 533, 597, 752, 1069, 1126, 1520.

### **Desmodium repandum (Vahl) DC.**

Prodr. 2: 334 (1825).

**Synonyms** *Hedysarum repandum* Vahl (1791), *Desmodium scalpe* DC. (1825), *Meibomia repanda* (Vahl) O. Kuntze (1891).

**Vernacular names** Indonesia: waliketupa sapi (Javanese), potong kujang (Sundanese), leng-elen-gan (Madura). Laos: kh'ua s'a:thwa'.

**Distribution** Africa, Asia from Sri Lanka and India eastward to China and southward to Malasia, and some Pacific Islands; a common species throughout the Malasian region.

**Uses** In Indonesia, *D. repandum* is used in combination with other diuretic substances. It has also been used as a green manure and ground cover.

**Observations** An erect suffrutescent herb, usually 30–150 cm tall, often woody at the base, stems simple or branched, variously hairy, rootstock woody; leaves 3-foliolate, stipules narrowly ovate, (10–)15–20 mm long, terminal leaflet rhombic to obovate, (2–)5–8(–11) cm × (2–)3.5–6(–9) cm, lateral leaflets smaller than the terminal ones, chartaceous, upper surface variously hairy, lower surface densely hairy, lateral veins 4–5, extending to the margin, strongly oblique; inflorescence terminal or frequently terminal and axillary, racemose or sometimes paniculate, 15–30(–60) cm long, lax-flowered; flowers in clusters of 2–4, calyx 4-lobed, rather densely hairy, corolla orange to red, androecium diadelphous; pod (2–)3–4(–5)-jointed, article length 2 times the width, densely hairy on lateral surfaces, glabrescent or sparsely pubescent on sutures; seeds 2.5–3.5 mm × 5–5.6 mm. In Malasia and Indo-China, *D. repandum* is found in light forests and thickets at 700–2000 m altitude, almost always under everwet climatic conditions, also along roadsides.

**Selected sources** 97, 389, 580, 752, 921, 1069, 1120, 1520.

### **Desmodium sequax Wallich**

Pl. asiat. rar. 2: 46, t. 157 (1831).

**Synonyms** *Desmodium sinuatum* (Miq.) Blume ex Baker (1876), *Desmodium ancistrotrichum* K. Schumann & Lauterb. (1901).

**Vernacular names** Papua New Guinea: ufi pata (Kabiufa, Eastern Highlands).

**Distribution** From India eastward to China and Taiwan and southward to Malesia; in Malesia common throughout the region.

**Uses** In Papua New Guinea, the chewed leaf is spat onto wounds, apparently for its antiseptic properties. The root is included in mixtures used to alleviate toothache. In China, it is considered a diaphoretic.

**Observations** A shrub, up to 200 cm tall, young branches hairy; leaves 3-foliolate, stipules lanceolate, 4–7 mm long, terminal leaflet ovate to obovate or rhomboid, 3.5–14 cm × 2.3–8 cm, larger than the lateral ones, subcoriaceous, upper surface glabrescent, lower surface hairy; inflorescence a raceme, rarely branched; flowers in pairs, calyx 4-lobed, hairy, corolla pink, purple to violet-red, androecium monadelphous; pod 30–40 mm × 2.5–3.5 mm, (6–)8–13-jointed, articles 3–4 mm long, densely covered with hooked hairs; seeds 1.9–2.2 mm × 1.5–1.8 mm, rim-arillate. In Malesia, *D. sequax* is confined to higher altitudes, at 500–2400 m, mostly in open places, not in dry regions. In Vietnam it is found along water courses, in grassland, open places and forest margins at 200–1600 m altitude.

**Selected sources** 97, 389, 597, 629, 752, 1069, 1126, 1393, 1520.

### ***Desmodium strigillosum* Schindler**

Engl. Bot. Jahrb. 54: 57 (1916).

**Vernacular names** Cambodia: trôm sva;. Laos: taum pauv hma.

**Distribution** Burma (Myanmar), Cambodia, Laos and Vietnam.

**Uses** Macerated and crushed roots are applied as a poultice on swellings of the limbs.

**Observations** An erect shrub or subshrub, up to 60 cm tall, stems glabrescent; leaves 3-foliolate, stipules narrowly triangular, 7–8 mm long, terminal leaflet elliptical or obovate, 2–3.5(–4) cm × 1–1.7(–2) cm, larger than the lateral ones, thickly chartaceous to subcoriaceous, upper surface glabrous, lower surface densely hairy, lateral veins 5–7, not extending to the margin; inflorescence terminal, racemose, 3–6(–8) cm long, densely flowered; flowers in pairs, calyx 4-lobed, sparsely hairy, corolla blue, androecium diadelphous; pod narrowly oblong, compressed, 15–20 mm × 2–2.5 mm, 6–8-jointed, articles broadly oblong, densely covered with hooked and straight hairs; seeds 1.1–1.3 mm × 1.5–1.8 mm, rim-arillate. *D. strigillosum* is found on dry slopes and wasteland up to 800 m altitude. It may well be grown in the Malesian region.

**Selected sources** 389, 1069.

### ***Desmodium styracifolium* (Osbeck)**

**Merr.**

Amer. Journ. Bot. 3: 580 (1916).

**Synonyms** *Hedysarum styracifolium* Osbeck (1757), *Desmodium capitatum* Burm.f. (1825), *Desmodium retroflexum* (L.) DC. (1825).

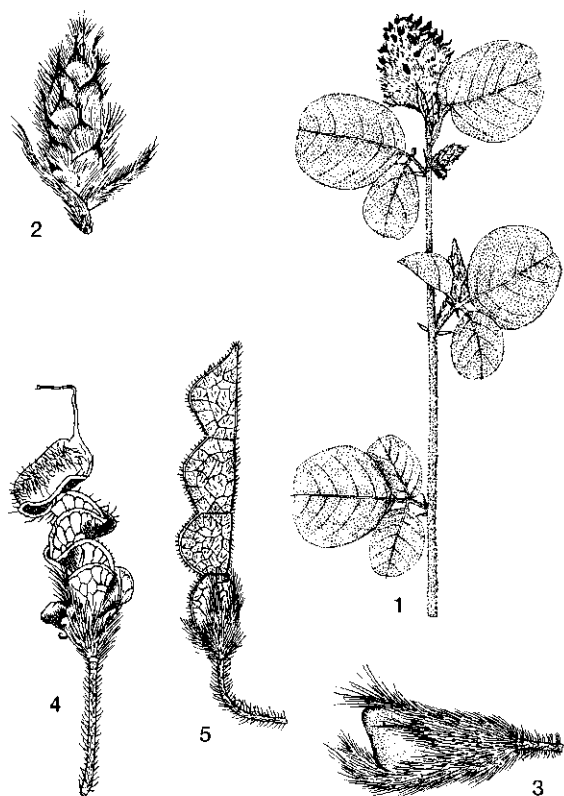
**Vernacular names** Indonesia: katepan, melikan (Javanese), susuukan (Sundanese). Malaysia: korat nasi. Cambodia: voë vè:t (general), smau srâlâb pôpôok, srâka: nièk (Battambang). Laos: kéng no:yz ngwâ liaz, pâdong fai. Thailand: ruk ku ning taa no, phee suea nam (Lampang). Vietnam: d[oo]fng ti[ee]n ([oo]ng), kim ti[ee]n th[ar]o, v[ar]ly r[oo]fng.

**Distribution** From India and Sri Lanka eastward to China, and southward to Malesia and Micronesia. Commonly found throughout the Malesian region, but no records from New Guinea.

**Uses** In Malaysia, *D. styracifolium* is used against colic, together with the central part of maize cobs. In Vietnam, the whole plant is considered diuretic, and applied against gall bladder and kidney stones and oedema. In southern China, it is also used to treat gall bladder and kidney stones. *D. styracifolium* is reported to be a good green manure, which covers the soil fast and produces numerous roots, nodules and seeds.

**Observations** A much branched, erect shrub or prostrate to ascending subshrub, 60–200 cm tall, young stems hairy; leaves 1- or 3-foliolate, stipules narrowly triangular, 8–11 mm long, terminal leaflet broadly elliptical or broadly obovate to orbicular, (1–)2–4.5(–5.5) cm × (1–)2–4.5(–5) cm, larger than the lateral ones, thick chartaceous to subcoriaceous, upper surface glabrous, lower surface densely hairy, lateral veins 8–10, not extending to the margin; inflorescence terminal and axillary, short racemose, 1–3(–5) cm long, very densely flowered; flowers in pairs, calyx 4-lobed, densely hairy, corolla purple or violet and fragrant, androecium diadelphous; pod narrowly oblong, compressed, (12–)15–20 mm × 2.5–3 mm, plicate-retroflexed when young, later straight, (4–)5–6-jointed, article quadrate or transversely broadly oblong, densely covered with hairs; seed reniform, depressed obovate or transversely elliptical, 1.3–1.8 mm × 2–2.2 mm. *D. styracifolium* is found in rather open habitats, in full sunlight as well as in shade. It grows on periodically wet and dry terrain, as well as in grasslands and abandoned rice fields from sea-level up to 600 m altitude.

**Selected sources** 50, 97, 202, 389, 580, 591, 593, 752, 1035, 1069, 1120, 1393, 1476, 1602.



*Desmodium styracifolium* (Osbeck) Merr. - 1, flowering stem; 2, young inflorescence; 3, flowerbud; 4, young pod; 5, mature pod.

### *Desmodium triflorum* (L.) DC.

Prodr. 2: 334 (1825).

**Synonyms** *Hedysarum triflorum* L. (1753), *Desmodium parvifolium* Blanco (1845).

**Vernacular names** Indonesia: daun mules (general), semanggan (Javanese), jukut jarem (Sundanese). Malaysia: rumput berek sisek putih, sisek tenggiling. Philippines: kaliskis-dalag (Tagalog), himbispuyo (Visaya), gumadep (Ifugao). Cambodia: smau kaè lolook. Thailand: ya-klethoi, ya-tanhoi (central), ya-tansai (northern). Vietnam: h[af]n the.

**Distribution** Pantropical; a common species throughout the Malesian region, Australia and the Pacific Islands.

**Uses** A decoction of *D. triflorum* is commonly used to treat diarrhoea and dysentery in Indonesia, Malaysia, the Philippines, Thailand, as well as China, India and Sri Lanka. In the Philippines, a decoction is also used as a mouthwash and as an expectorant. In Thailand, the whole plant is used

as an antipyretic and to quench thirst. In Indonesia, Malaysia, the Philippines, Laos and India the crushed plant or a poultice of the leaves is externally applied on wounds, ulcers, and for skin problems in general, apparently for its antiseptic properties. It is also used as a forage, as a green manure and ground cover.

**Observations** A much branched, mat-forming, prostrate, annual or perennial herb, 8–50 cm long, covered with hairs, rootstock woody; leaves 3-foliolate, stipules obliquely lanceolate, 3.5–6 mm long, persistent, terminal leaflet obovate-oblong, obovate or obcordate, 4–14 mm × 4–12 mm, larger than the lateral ones, variably hairy beneath; flowers 1–3 in leaf axils, calyx 5-lobed, hairy, corolla pink to purple, androecium diadelphous; pod flat, 6–18 mm × 2–3 mm, 3–5-jointed, constricted between the rectangular articles; seed quadrangular, 1.2 mm × 1.7 mm. *D. triflorum* is found on a wide range of soils, and most commonly in heavily grazed or closely cut areas.

**Selected sources** 20, 97, 119, 389, 482, 483, 629, 921, 1120, 1178, 1393, 1434, 1520.

### *Desmodium velutinum* (Willd.) DC.

Prodr. 2: 328 (1825).

**Synonyms** *Desmodium lasiocarpum* (P. Beauv.) DC. (1825), *Desmodium latifolium* (Roxb. ex Ker.) DC. (1825).

**Vernacular names** Thailand: yaa song plong (central, northern), nieo yai (southwestern).

**Distribution** Africa, Asia from India and Sri Lanka eastward to China and Taiwan and southward to Malesia; common throughout the Malesian region.

**Uses** In Papua New Guinea, squeezed leaves are chewed with salt to treat diarrhoea. Leaves and roots may be chewed (not swallowed) to relieve toothache.

**Observations** An erect shrub, usually 100–300 cm tall, branches often dark red, young parts densely hairy, rootstock thickened; leaves 1-foliolate, sometimes mixed with 3-foliolate leaves, stipules narrowly triangular, 2–15 mm long, leaflets very variable in size and shape, 4–20 cm × 2.5–13 cm, chartaceous to coriaceous, upper surface continuously appressed-pubescent, lower surface densely velutinous, lateral veins 8–10, extending to the margin; inflorescence terminal and axillary, racemose or paniculate, up to 20 cm long; flowers in clusters of 2–5, calyx 4-lobed, densely hairy, corolla pink, purple, blue or reddish-violet, androecium diadelphous; pod (1–)1.6–2.5 cm × 2.2–3.5 mm, (3–)5–7-jointed, articles broadly ob-

long, densely covered with hairs; seeds very broadly or depressed ovate, 1.3–1.6 mm × 1.8–2.5 mm. *D. velutinum* is subdivided in subspecies and varieties, of which *D. velutinum* subsp. *longibracteatum* (Schindler) H. Ohashi and *D. velutinum* subsp. *velutinum* are found in South-East Asia. *D. velutinum* is found in rather open habitats, savanna, secondary forest margins and as a garden weed from sea-level up to 1000 m altitude.

**Selected sources** 97, 389, 608, 612, 629, 752, 921, 1069, 1120, 1393, 1520.

N. Setyowati-Indarto & M. Brink

## Elephantopus L.

Sp. pl. 2: 814 (1753); Gen. pl. ed. 5: 355 (1754).

COMPOSITAE

$x = 11$ ; *E. mollis*:  $2n = 22, 44$ , *E. scaber*:  $2n = 22$ , *E. spicatus*:  $n = 10, 11, 13, 14$ ,  $2n = 22, 26$

**Major species** *Elephantopus mollis* Kunth, *E. scaber* L.

**Vernacular names** Elephant's foot (En). Pied d'éléphant (Fr). Indonesia: tapak liman. Malaysia: tutup bumi. Vietnam: c[us]c ch[ir] thi[ee]n.

**Origin and geographic distribution** *Elephantopus* comprises approximately 30 species and is mainly of tropical American origin. Some species have been introduced in South-East Asia. *E. scaber*, which is the most widespread in South-East Asia, is possibly indigenous there.

**Uses** In South-East Asian countries and India, *Elephantopus* species are well known in traditional medicine. Their primary use is as a diuretic and febrifuge, but other applications include as an anthelmintic (e.g. filariasis), antibiotic, anti-inflammatory, emollient, diaphoretic, emmenagogue and galactagogue. The plant is known as a remedy for coughs, headache, anaemia, dyspepsia, dysentery, colic pains, diarrhoea and leucorrhoea, and also reported to be beneficial during parturition: to speed birth and expulsion of the placenta, and afterwards to prevent inflammations. These uses are not confined to a certain part of the plant; preparations (such as decoctions or hot water extracts) of the (dried) roots, entire plants or leaves are mentioned in the literature.

*Elephantopus* preparations are also applied externally to heal wounds (astringent properties are attributed to the plant) and in cases of rheumatism (as a paste made from the entire plant).

In Africa and South America, elephants's foot has similar ethnomedical applications. In Chinese traditional medicine *Elephantopus* is used to treat in-

fluenza, colds, tonsillitis, pharyngitis, conjunctivitis, epidemic encephalitis, hepatitis, cirrhosis, ascites and eczema.

**Production and international trade** *Elephantopus* spp. are commonly collected from the wild, but there is no information on production and trade. A drug derived from *E. scaber* is available in China as tablets; it is marketed as a Chinese patent medicine.

**Properties** *Elephantopus* plants taste bitter. Most species (e.g. *E. mollis* and *E. scaber*) contain elephantopin and/or its related compounds (e.g. molephantin, molephantinin, phantomolin, deoxyelephantopin, isodeoxyelephantopin and 11,13-dihydrodeoxyelephantopin). These elephantopin derivatives (sesquiterpene lactones of the germacrane type, or germacranolides) have been found to be very characteristic of *Elephantopus*, and have attracted considerable interest because of their reported cytotoxic (anti-cancer) activities.

Molephantin, molephantinin and phantomolin (obtained from the ethanolic extract of *E. mollis*) have been shown to be active in various tumour assays in vitro (human epidermoid carcinoma of the larynx, human carcinoma of the nasopharynx) and in vivo (Ehrlich ascites carcinoma, Walker 256 carcinosarcoma, Lewis Lung carcinoma, P-388 lymphatic leukaemia). Deoxyelephantopin (from *E. scaber*) significantly inhibited the growth of Walker 256 carcinosarcoma in rats. Furthermore, dihydroelephantopin inhibited the growth of leukaemia tumour cells, and tomentophantin-A and -B (from *E. tomentosus* L.) displayed cytotoxic activity in the in vitro human nasopharynx carcinoma assay. Research has revealed the importance of an  $\alpha$ -methylene lactone function for the biological activity of sesquiterpenoid lactones.

Phytochemical investigations for the germacranolides have also revealed the presence of several other compounds. The guaianolides deacylcyanaropicrin, glucoazulanin-C and deacylcyanaropicrin-3 $\beta$ -glucopyranoside (= crepiside E) were isolated from the aqueous fraction of the total ethanolic extract of *E. scaber*. Stigmasteryl-3 $\beta$ -glucoside (a sterol) was isolated from the chloroform fraction. The triterpenes  $\beta$ -amyrin acetate, lupeol (acetate) and epifriedelanol, and the phytosterol stigmasterol were identified from ethanolic extracts of *E. mollis*.

Studies on the hepatoprotective effects of water extracts of *E. mollis*, *E. scaber* and *E. spicatus* against acute hepatic damage in rats induced by  $\beta$ -D-galactosamine and paracetamol (acetaminophen) demonstrated that these crude extracts

(500 mg/kg intraperitoneally) decrease the levels of serum glutamate-oxalate-transaminase (SGOT) and serum glutamate-pyruvate-transaminase (SGPT). This suggests that treatment with these extracts improves the induced hepatic lesions. In another analogous study, using  $\text{CCl}_4$ , it was furthermore found that hepatic fatty metamorphosis and necrosis of the central lobules were significantly improved after treatment with *E. mollis* and *E. scaber* extracts, but less after treatment with *E. spicatus* extract.

A preliminary pharmacological evaluation was conducted with aqueous and water/ethanol extracts of *E. scaber*. Doses of 0.3–6 g/kg induced writhing, loss of muscle tone, ataxia, prostration and death in mice. No analgesic effects of these extracts were detected using the mouse hot-plate ( $\leq 30$  mg/kg intraperitoneally (i.p.),  $\leq 600$  mg/kg peroral (p.o.)) or the acetic acid-induced writhing (mouse,  $\leq 300$  mg/kg p.o.) tests. Both extracts failed to modify diuresis in the rat ( $\leq 300$  mg/kg p.o.), to reduce carrageenan-induced rat paw oedema ( $\leq 600$  mg/kg p.o.) or to reduce yeast-induced hyperthermia in rats ( $\leq 600$  mg/kg p.o.). Aqueous extracts administered at doses of 25–100 mg/kg intravenously reduced blood pressure and heart rate in rats. Absence of diuretic effects was furthermore shown in a placebo controlled trial carried out on 10 healthy human volunteers.

Studies showed that *E. scaber* exhibits considerable antibacterial activity against the cariogenic bacterium *Streptococcus mutans* and against *Staphylococcus* sp. In general, *E. scaber* markedly inhibits the growth of both gram-positive and gram-negative bacteria. *Elephantopus* extract exhibits antiviral activity against e.g. polio virus and HIV. Protease and reverse transcriptase inhibition are also reported.

**Adulterations and substitutes** Sesquiterpene lactones are common in *Compositae*, but compounds of the elephantopin type seem characteristic for *Elephantopus*. In other genera of the tribe *Vernonieae*, such as *Vernonia*, the sesquiterpene lactones are mostly of the germacranolide type, and some of these compounds have also shown significant in vitro activity against cells derived from carcinomas (e.g. vernolepin from *Vernonia hymenolepis* A. Rich.).

**Description** Perennial herbs up to 120 cm tall, with a creeping rhizome or strong taproot; stems scabrous or pubescent. Leaves alternate, often rosulate or basal and cauline, becoming smaller upwards, simple, variously toothed, pinnately

veined, cuneately tapering or long-attenuate at base to a winged petiole, exstipulate. Inflorescence consisting of sessile, few-flowered, homogamous heads aggregated into a globose capitulate cluster or glomerule each of which is subtended by 2–4 foliaceous bracts; glomerules in corymbs or panicles or spicately arranged in the axils of small cauline leaves; involucre of head 2-seriate and consisting of about 8 narrowly oblong bracts. Flowers all tubular, bisexual, 5-merous; pappus of 4–5(–6) scabrid bristles; corolla with unequal, somewhat palmately spreading lobes, white, bluish or purplish; anthers connate and forming a tube around the style, obtuse at base and acute at apex; ovary inferior, 1-celled, style bifid with filiform, hairy arms. Fruit a truncate, 10-ribbed achene tipped by the 4–5 scabrid pappus bristles, which are dilated at base and equally long or with 2 longer ones sigmoidal at apex.

**Growth and development** *Elephantopus* can be found flowering throughout the year, but there is often a main flowering season (e.g. for *E. scaber* in Java from April to September). Flowers remain hidden in the involucre until shortly before anthesis. Within a head the usually 4 flowers expand almost simultaneously between 1 and 2 p.m., and close at about 5 p.m. The fruits are often distributed by mammals and people, when the scabrid bristles become caught in the fur or clothes.

**Other botanical information** *Elephantopus* is classified in the tribe *Vernonieae*, which includes about 70 genera and 1500 species. There is no modern comprehensive review of the tribe and consequently the affinity of *Elephantopus* is still obscure.

*Pseudelephantopus* is often kept separate from *Elephantopus* on the basis of the heads which are clustered in the axils of small cauline leaves and of 2 larger, sigmoid pappus hairs (in *Elephantopus* s.s. glomerules of heads terminal and usually long-pedunculate, all pappus hairs equal and straight). *E. spicatus* has been treated here and not separately because some recent publications have again treated *Pseudelephantopus* as if it is synonymous with *Elephantopus* (even though it is sometimes considered as a separate subgenus *Pseudelephantopus* (Rohr) C. Jeffrey). *Orthopappus* is nowadays also often included in *Elephantopus*.

**Ecology** Elephant's foot usually occurs in waste places, grasslands, roadsides and forest fringes. *E. scaber* is also found in the undergrowth of teak forest in Java. *Elephantopus* spp. are usually not noxious weeds, although they may occur on paths

in tea plantations, and in lawns, where they are sometimes considered as troublesome. In Hawaii, *E. mollis* and *E. spicatus* are classified as noxious weeds, which rapidly invade pastures but have no forage value. In South-East Asia, *Elephantopus* may occur up to 2000 m altitude.

**Diseases and pests** In the Philippines, a mosaic virus may cause mild chlorosis and prominent veins in *E. mollis*. There are indications that the disease can be transmitted by aphids to coconut and Manila hemp (*Musa textilis* Née), where symptoms can be more serious.

**Harvesting** Elephant's foot is collected from the wild.

**Handling after harvest** After collecting, the plants are washed, dried and stored until use. In general, they are collected fresh when needed.

**Genetic resources and breeding** *Elephantopus* spp. are widespread in anthropogenic habitats and seem not to be at risk of genetic erosion. No breeding work has been done to date.

**Prospects** Elephant's foot is considered to be an interesting medicinal plant resource with promising prospects for becoming a valuable adjunct in e.g. cancer research or treatment.

**Literature** |1| Chen, C.P., Lin, C.C. & Namba, T., 1989. Screening of Taiwanese crude drugs for antibacterial activity against *Streptococcus* mutants. *Journal of Ethnopharmacology* 27(3): 285–295. |2| de Silva, L.B., Herath, W.H.M.W., Jennings, R.C., Mahendran, M. & Wannigama, G.E., 1982. A new sesquiterpene lactone from *Elephantopus scaber*. *Phytochemistry* 21(5): 1173–1175. |3| Hayashi, T., Koyama, J., McPhail, A.T. & Lee, K.-H., 1987. Structure and absolute stereochemistry of tomenphantopin-A and -B, two cytotoxic sesquiterpene lactones from *Elephantopus tomentosus*. *Phytochemistry* 26(4): 1065–1068. |4| Hisam, A., Pieters, L., Claeys, M., Dommissie, R., Vanden Berghe, D. & Vlietinck, A., 1992. Guaianolide glucosides from *Elephantopus scaber*. *Planta Medica* 58(5): 474–475. |5| Jakupovic, J., Jia, Y., Zdero, C., Warning, U., Bohlmann, F. & Jones, S.B., 1987. Germacranolides from *Elephantopus* spp. *Phytochemistry* 26(5): 1467–1469. |6| Lee, K.-H., Ibuka, T., Furukawa, H., Kozuka, M., Wu, R.Y., Hall, I.H. & Huang, H.-C., 1980. Antitumor agents. XXXVIII: Isolation and structural elucidation of novel germacranolides and triterpenes from *Elephantopus mollis*. *Journal of Pharmaceutical Sciences* 69(9): 1050–1056. |7| Lin, C.C., Tsai, C.C. & Yen, M.H., 1995. The evaluation of hepatoprotective effects of Taiwan folk medicine 'Teng-Khia-U'. *Journal of Ethnopharma-*

*logy* 45(2): 113–123. |8| Lin, C.C., Yen, M.H. & Chiu, H.F., 1991. The pharmacological and pathological studies on Taiwan folk medicine VI. The effects of *Elephantopus scaber* ssp. *oblanceolata*, *Elephantopus mollis* and *Pseudoelephantopus spicatus*. *American Journal of Chinese Medicine* 19(1): 41–50. |9| McPhail, A.T., Onan, K.D., Lee, K.-H., Ibuka, T., Kozuka, M., Shingu, T. & Huang, H.-C., 1974. Structure and stereochemistry of the epoxide of phantomolin, a novel cytotoxic sesquiterpene lactone from *Elephantopus mollis*. *Tetrahedron Letters* 32: 2739–2741. |10| Poli, A., Nicolau, M., Simoes, C.M.O., Nicolau, R.M.R. & Zanin, M., 1992. Preliminary pharmacologic evaluation of crude whole plant extracts of *Elephantopus scaber*. Part I: in vivo studies. *Journal of Ethnopharmacology* 37(1): 71–76.

#### *Selection of species*

#### **Elephantopus mollis** Kunth

Humb., Bonpl. & Kunth, Nov. gen. sp. 4, ed. fol. 4: 20 (1818).

**Synonyms** *Elephantopus tomentosus* auct. non L.

**Vernacular names** Philippines: malatabako (Tagalog), tabtabako (Iloko), kaburon (Igorot).

**Distribution** Tropical America; introduced and widely naturalized in tropical Africa and Asia. Reported for Peninsular Malaysia, Borneo, Sulawesi and the Philippines.

**Uses** In the Philippines, the leaves (fresh and crushed or dried and powdered) are applied to wounds as a vulnerary. A decoction of the plant is prescribed as diuretic and febrifuge. It is also reported to act as an emetic. In Hong Kong, the entire plant is used for its diuretic, antihepatic and anti-bronchitis properties.

**Observations** A herb up to 100 cm tall, stems whitish pilose; leaves basal and cauline, elliptical-ovate or elliptical-obovate to elliptical-lanceolate, 8–22 cm × 3–7 cm; glomerules terminal, generally long-peduncled, glomerule bracts generally shorter than the involucral bracts; flowers with corolla about 5 mm long, whitish or sometimes pinkish or purplish; fruit 2.5–3 mm long with pappus bristles equal and 3.5–4 mm long. *E. mollis* occurs in open waste places and grasslands up to 2000 m altitude.

**Selected sources** 121, 202, 564, 668, 833, 852, 861, 862, 925, 1126, 1162, 1178, 1539.

**Elephantopus scaber L.**

Sp. pl. 2: 814 (1753).

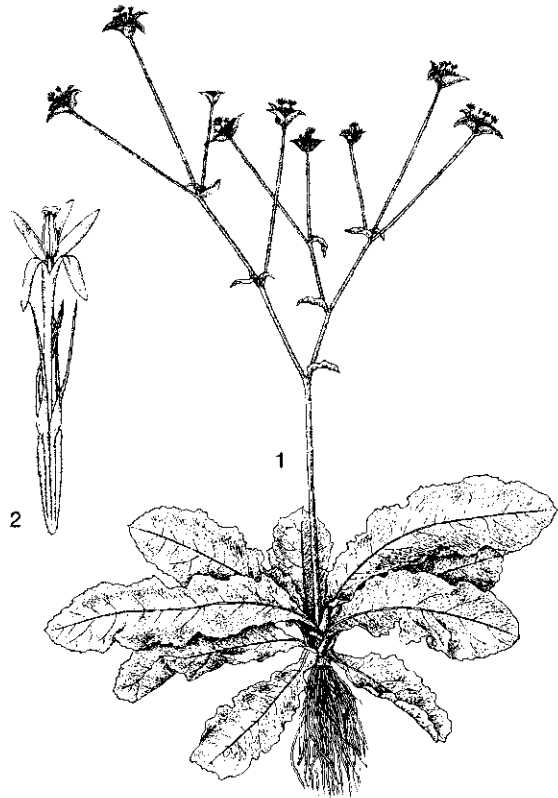
**Vernacular names** Prickly-leaved elephant's foot (En). Indonesia: tapak liman, tutup bumi (general), talpak tana (Madurese). Malaysia: tut-up bumi, tapak leman (Peninsular), pepalut (Sabah). Philippines: dila-dila, tabatabakohan (Tagalog), kabkabron (Iloko). Thailand: do mai ruu lom (central), kee fai nok khuun (Loei), naat me khlaen (Surat Thani). Vietnam: c[us]c chi thi[ee]n, c[or] l[uw][owx]i m[ef]o, dia d[ar]m th[ar]o.

**Distribution** Widespread in tropical America, Africa, Asia and Australia; throughout South-East Asia.

**Uses** *E. scaber* is widely used as a diuretic, febrifuge and emollient throughout South-East Asia. In Malaysia, a decoction of the leaves or roots is additionally used as a tonic and anthelmintic, and to treat coughs, asthma and venereal diseases. The fresh roots are prescribed to arrest vomiting, and the leaves are recommended for application to the abdomen to treat dropsy. In Indonesia, the roots, either pounded or in decoction, are also used as a remedy for leucorrhoea, anaemia and as a tonic during parturition, whereas the leaves are used as anthelmintic and aphrodisiac, and to treat cough, sprue and diarrhoea. In Indo-China, a decoction of the plant is used as tonic, diaphoretic and emmenagogue and to treat dyspepsia. In Thailand, besides being used as a diuretic and febrifuge, the roots are used to treat cough, malaria and parasites. In India, China, Africa and South and Central America, a decoction or infusion of the roots and leaves is used as an emollient, to treat diarrhoea, dysentery, pulmonary diseases, scabies, urethral discharges, epistaxis, jaundice, oedema and to relieve anuria and blennorrhoea.

**Observations** An erect herb up to 80 cm tall, stems rigid, appressed long-haired or scabrous; leaves in a radical rosette, if cauline much smaller, oblong-obovate to spatulate, 5–38 cm × 1–6 cm; glomerules terminal, generally long-peduncled, glomerule bracts generally longer than the involucre bracts; flowers with corolla 7–9 mm long, bluish or purplish, sometimes white; fruit about 4 mm long with pappus bristles equal and 4–6 mm long. *E. scaber* occurs in grasslands, wasteland, roadsides, along fields and in forest borders, up to 1500 m altitude.

**Selected sources** 27, 29, 97, 202, 244, 333, 350, 539, 580, 592, 815, 852, 861, 862, 1035, 1126, 1148, 1178, 1539.



*Elephantopus scaber* L. – 1, plant habit; 2, flower.

**Elephantopus spicatus Juss. ex Aublet**

Hist. pl. Guiane 2: 808 (1775).

**Synonyms** *Pseudelephantopus spicatus* (Juss. ex Aublet) Rohr (1792).

**Vernacular names** False elephant's foot (En). Philippines: dilang-aso (Tagalog), maratabako (Iloko), kalkalapikap (Bontok). Vietnam: ch[aa]n voi gi[es].

**Distribution** Native to tropical America; introduced in tropical Africa and Asia. In South-East Asia, reported very locally for Java and widespread for the Philippines; also in Vietnam, southern China and Taiwan.

**Uses** In the Philippines, the leaves are used topically to treat eczema, and as a vulnerary.

**Observations** An erect herb up to 120 cm tall, usually branched and with a strong taproot, stems slightly hairy to glabrous; leaves in a radical rosette and cauline, oblong-obovate to oblong-lanceolate, lower ones 5–25 cm × 1–7 cm, upper ones smaller; heads clustered in the axil of small cauline leaves, combined into a long, lax terminal spike, involucre bracts 2-seriate, the outer ones

shorter than inner ones; flowers with corolla about 7 mm long, white; fruit 6–7 mm long with pappus bristles unequal, the 2 largest sigmoid near the apex. *E. spicatus* is common in waste places in the Philippines.

**Selected sources** 97, 852, 861, 862, 1178.

Ng Lean Teik

### **Embelia** Burm.f.

Fl. indica; 62, t. 23 (1768).

MYRSINACEAE

*x* = unknown

**Major species** *Embelia ribes* Burm.f.

**Origin and geographic distribution** *Embelia* comprises approximately 130 species and occurs in Africa, Asia including Malesia, the western Pacific and Australia. Most species are found in Asia. In Indo-China 11 species have been found, in Peninsular Malaysia 17, in Sumatra 6, in Java 6, in Borneo more than 15 and in New Guinea 10. However, no recent overview is available for the Malesian region.

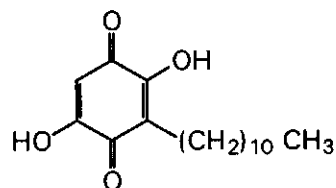
**Uses** Several *Embelia* species are well known for their anthelmintic and taeniafuge uses. The active principles primarily act as a purgative. *E. ribes* has been known since ancient times in Ayurvedic medicine under the Sanskrit name 'vidanga'. The dried fruit is considered anthelmintic, astringent, carminative, alterative and stimulant. In China, roots and leaves of *E. laeta* are used to treat dysentery, indigestion and eczema, while the fruits are used to treat anaemia, gum bleeding and anorexia. In eastern Africa, *Embelia* (e.g. *E. schimperi* Vatke) fruits either fresh or dried are a popular anthelmintic. The roots are used as a purgative and vermifuge. In Thailand, the roots of *E. ribes* are used as expectorant and in the treatment of internal inflammations. In Papua New Guinea, squeezed sap of *Embelia* stems and young leaves is drunk to treat malaria or as a general febrifuge. In East Kalimantan, the crushed fresh bark of *E. ribes* is used as a fish poison. In Java the young leaves, shoots and immature fruits of *E. ribes* are consumed cooked as a sour vegetable or condiment. *E. philippinensis* A. DC. is used in a similar way in the Philippines. The ripe sour-sweet fruits of both species are also eaten as a delicacy. The fruits of *E. coriacea* Wallich ex A. DC. are reported to be poisonous. When dissolved in alcohol, embelin (a major constituent of *Embelia*) can be used as a yellow dye for silk and wool.

**Production and international trade** Al-

though fruits of *Embelia* were traded from Malesia to Europe at the beginning of the 20th Century, at present no commercial trade is reported.

**Properties** Embelin (2,5-dihydroxy-3-undecyl-1,4-benzoquinone) is recognized as the main active principle of *Embelia* species. The fruits of *E. ribes* contain 2.5–3.1% embelin and about 1% of the sugar quercitol, those of *E. tsjeriam-cottam* about 1.6% embelin and no quercitol. The fruits of the African *E. schimperi* yield 4.8–7.5% embelin and about 1% quercitol. Embelin is also isolated from *E. robusta*. Embelin can be isolated as golden yellow needles, insoluble in water, but clearly soluble in ethanol, chloroform and benzene.

The best known pharmacological action of embelin is its effect on reproduction: male antifertility, anti-implantation and abortifacient activity has been shown in tests with various animals. Ethanolic (90%) extracts of *E. ribes* fruits administered orally to female rats at 250 mg/kg during early pregnancy showed 66% anti-implantation activity, and the abortifacient activity in late pregnancy was about 25%. A daily subcutaneous administration of embelin at a dose of 20 mg/kg body weight to male albino rats for 15 or 30 days inhibited the motility of epididymal sperm, fertility parameters (such as pregnancy attainment and litter size) and the activities of enzymes of glycolysis and energy metabolism. However, all changes were reversible, as seen after 15 and 30 days of recovery. Addition of embelin to epididymal sperm suspensions caused a dose and duration-dependent inhibition of spermatozoal motility and inhibited the activities of carbohydrate metabolism enzymes. Light and scanning electron microscopy showed that both in vitro and in vivo treatment with the substance causes profound morphological changes in spermatozoa such as decapitation of the spermatozoal head, discontinuity of the outer membranous sheath in the mid-piece and the tail region, and alteration in the shape of the cytoplasmic droplet in the tail. In another experiment, sixty 3-month-old pregnant mice were treated with a suspension of embelin (50 mg/kg body weight) in carboxymethylcellulose (CMC, 0.5 ml) from days 3



embelin



to 8, days 6 to 9 or days 10 to 16 of gestation. A control group was treated with distilled water and CMC (0.5 ml). In a second experiment, males and virgin females were treated with embelin (50 mg/kg body weight) for 15 and 30 days respectively. Treated males were than mated with untreated females, and treated females were mated with untreated males. A distilled water and CMC control was maintained during the experiment. Embelin treatment resulted in a significant increase in resorptions (maximum of 18.5% in females treated from days 3 to 8) and skeletal abnormalities (maximum of 26% in females treated from days 3 to 8). Furthermore, embelin treatment resulted in changes in the sex ratio of litters, with 132 females to 100 males occurring in females treated from days 3 to 8 compared with 96 females to 100 males in the distilled water control. Treatment with embelin prior to mating significantly increased the incidence of dead foetuses and increased the occurrence of physical and skeletal abnormalities and resorption. Histochemical and biochemical investigations of male rats, treated for 35 days with doses of 0.3–0.5 mg/kg body-weight (subcutaneously), revealed a picture of the testis with arrest of spermatogenesis, a marked diminution in the cell population and deformation of the seminiferous tubulus at all doses. Necrobiosis and nuclear degeneration of varying degrees were observed. The weight of testes, epididymis, vas deferens, seminal vesicle, coagulating gland, ventral prostate, dorso-lateral prostate, adrenal gland and levator and muscle were reduced considerably, just like the biochemical parameters in the tissues, such as glycogen in testis and fructose in seminal vesicle, prostate and coagulating glands. These observations may be due to a hormonal imbalance, caused by embelin; an anti-androgenic nature is suggested. A significant anti-oestrogenic and weak progestational activity was also found in female rats.

Oral administration of embelin (75 mg/kg body weight, daily for 15 and 30 days) to male rats caused a significant increase in the uptake of D-glucose, L-alanine, L-leucine and calcium in small intestine segments. Embelin also produced significant increases in intestinal brush border membrane-associated enzymes (invertase, lactase, maltase, alkaline phosphatase and leucine aminopeptidase) in both intestinal homogenates and partially purified brush border membrane preparations. Significant increases were also noted for microsomal glucose-6-phosphatase and cytosolic lactate dehydrogenase. Increases in brush

border membrane-associated total lipids, phospholipids, cholesterol, triacylglycerol, unesterified fatty acids and ganglioside sialic acid were seen but there was little change in the cholesterol-phospholipid molar ratio. All changes returned to control or near control levels following withdrawal of the drug.

Embelin (isolated from the fruits of *E. ribes*) and administered to male albino rats with fibrosarcoma at doses of 50 or 100 mg/kg, orally, for 20 days, prolonged survival time and induced significant tumour regression. To evaluate the possible mode of action of embelin, the anti-oxidant status of the drug was studied. Embelin exhibited significant free radical scavenging properties. The biological defence system constituting the superoxide dismutase, catalase, glutathione peroxidase, glutathione-S-transferase and glutathione was significantly enhanced, whereas lipid peroxidation was greatly restricted. In vivo studies in rats and mice indicated an analgesic effect of potassium embelate. It was found to be effective by oral, intramuscular, subcutaneous and intracerebro ventricular routes (doses ranged from 5–40 mg/kg), and the results compared well with morphine (a mainly  $\mu$  receptor agonist) as reference. Potassium embelate acts centrally to produce analgesia (binding studies indicate that mixed  $\mu$  and  $\kappa$  receptor sites may be involved), and its effect is not strongly antagonized by naloxone (a pure  $\mu$  receptor antagonist), indicating that other central sites are also of importance. Furthermore, there is no precipitation of abstinence syndrome as observed with morphine. A toxicity evaluation which included subacute, chronic, reproductive testing and teratological investigations in laboratory animals (mice, rats and monkeys; doses 10–100 mg/kg per day, orally) did not indicate adverse effects, suggesting the compound is safe.

'Pestoban' is a mixture of 3 plant extracts, only 2 of which (*Cedrus deodara* Loud. and *Azadirachta indica* A.H.L. Juss. oils) show toxicity against the mollusc species *Lymnaea acuminata* and *Indoplanorbis exustus*. The third plant extract, that of *E. ribes*, is non-toxic, but when all three are mixed, the toxicity of the mixture is about 100 times more active than either of the two toxic components. The toxic effect against the snail *Lymnaea acuminata*, an intermediate host for both *Fasciola hepatica* and *F. gigantica* is time- as well as dose-dependent. The molluscicidal activity is higher than synthetic molluscicides, and the dose levels used against the snail are safe for fish. 'Ectozee' is a herbal preparation containing the same

combination of plants, which is used against mites.

Application of a seed extract of *E. ribes* results in a high mortality of the nematodes *Meloidogyne incognita* and *Rotylenchulus reniformis*.

**Adulterations and substitutes** Dried fruits of *E. ribes* and *E. tsjeriam-cottam* have been used as an adulterant of black pepper (*Piper nigrum* L.).

**Description** Dioecious scandent shrubs or lianas up to 20 m long. Leaves alternate, distichous or spirally arranged, simple, entire to serrate, glabrous, dotted with red-brown or black glandular dots or streaks, petioled. Inflorescence axillary or terminal, fascicled or solitary, racemose or paniculate, sometimes subumbellate. Flowers functionally unisexual, small, 4–5(–6)-merous; calyx deeply lobed, lobes imbricate; petals free or slightly connate at base, imbricate or contorted, usually papillate inside and glabrous outside; stamens and staminodes inserted towards the base of the petals, filaments short to rather long, anthers dorsifixed; ovary superior, in male flowers minute, in female flowers globose or ovoid, few-ovuled, style short, persistent, stigma broad, disciform. Fruit a subglobose to ovoid drupe, usually drying black, 1-seeded. Seedling with epigeal germination; cotyledons emergent, leafy with finely toothed margins, hypocotyl elongated.

**Growth and development** The various *Embelia* species do not flower simultaneously, and fruit maturation takes several months. The juicy fruits are probably dispersed by birds.

**Other botanical information** The closely related genus *Grenacheria* differs primarily in the connate petals. No recent taxonomical revision of the genus *Embelia* is available and the existing revision from the beginning of the 20th Century is based on a limited number of specimens. Numerous species have been described with a high degree of endemism, and this causes considerable taxonomic difficulties. The species are rather difficult to distinguish in the field. For example *E. robusta* and *E. tsjeriam-cottam* are often confused.

**Ecology** *Embelia* can be found from sea-level up to 2000(–2800) m altitude. Some species prefer primary forest, but others are common along forest margins and in secondary forest.

**Propagation and planting** *Embelia* can be propagated by seed or cuttings. Most species are common in secondary vegetation and not propagated or planted deliberately. Occasionally some *Embelia* are grown on a small scale in home gardens.

**Harvesting** *Embelia* fruits are simply picked

when ripe. Young shoots to be used as a vegetable are harvested shortly before use or sale. For collecting the roots, plants are uprooted.

**Handling after harvest** Fruits of *Embelia* are either used fresh or dried for medicinal purposes. Sometimes the fruit pulp is removed by washing and threshing the fruits, and the seeds are dried and stored. Dried fruits and seeds guarantee a year-round supply in the market. Stems harvested to be used as a fish poison are simply bundled, and the bark is later pulverized at the spot to release the toxic agents.

**Genetic resources and breeding** Some species like *E. ribes* and *E. tsjeriam-cottam* have a wide distribution, but most *Embelia* species have a very local occurrence. The latter species are theoretically more vulnerable to extinction.

**Prospects** The application of embelin in herbal molluscicides and against mites appears promising. Other applications of embelin, e.g. as analgesic, antifertility or free radical scavenging agent deserve more attention.

**Literature** |1| Atal, C.K., Siddiqui, M.A., Zutshi, U., Amla, V., Johri, R.K., Rao, P.G. & Kour, S., 1984. Non-narcotic orally effective, centrally acting analgesic from an Ayurvedic drug. *Journal of Ethnopharmacology* 11: 309–317. |2| Chitra, M., Devi, C.S.S. & Sukumar, E.P., 1994. Protective action of embelin against lipid peroxidation on tumour bearing rats. *Fitoterapia* 65(4): 317–321. |3| Das, S.S. & Bhatia, B.B., 1994. Comparative therapeutic evaluation of Ectozee aerosol spray and Betnovate-N against mite causing canine dermatitis. *Indian Journal of Indigenous Medicines* 10(2): 9–10. |4| Gupta, S., Sanyal, S.N. & Kanwar, U., 1989. Antispermatic effect of embelin, a plant benzoquinone, on male albino rats in-vivo and in-vitro. *Contraception* 39(3): 307–320. |5| Gupta, S., Sanyal, S. & Kanwar, U., 1991. Effects of embelin, a male antifertility agent, on absorptive and digestive functions of rat intestine. *Journal of Ethnopharmacology* 33(3): 203–212. |6| Mez, C., 1902. *Embelia*. In: Engler, A. (Editor): *Das Pflanzenreich [The plant kingdom]. Heft 9 (IV 236). Wilhelm Engelmann, Leipzig, Germany.* pp. 295–332. |7| Singh, K., Singh, A. & Singh, D.K., 1995. Molluscicidal activity of different combinations of the plant products used in the molluscicide Pestoban. *Biological Agriculture & Horticulture* 12(3): 253–261. |8| Srinath, B.R., Vivekanandan, O.S., Shivakumar, K.R. & Rao, K.R.R., 1991. Foetotoxic and teratogenic effects of embelin - an abortifacient compound on mice. *Current Research - University of Agricultural Sciences (Ban-*

galore) 20(5): 91-93. |9| Varshney, M.D., Sharma, B.B. & Gupta, D.N., 1986. Antifertility screening of plants. Part II. Effect of ten indigenous plants on early and late pregnancy in albino rats. *Comparative Physiology and Ecology* 11(4): 183-189. |10| Zutshi, U., Johri, R.K. & Atal, C.K., 1989. Possible interaction of potassium embelate, a putative analgesic agent, with opiate receptors. *Indian Journal of Experimental Biology* 27(7): 656-657.

*Selection of species*

**Embelia laeta (L.) Mez.**

Engler, *Pflanzenr.*, Heft 9 (IV 236): 326 (1902).

**Synonyms** *Choripetalum benthamii* Hance (1853), *Embelia obovata* Hemsl. (1889).

**Vernacular names** Vietnam: th[uf]n m[ux]n, c[aa]y m[ux]n, d[aa]y ng[os], chua ng[us]t, chua m[es]lo.

**Distribution** Vietnam, southern China and Taiwan.

**Uses** The fruits are used as a vermifuge (taenifuge) in Vietnam. The sap of the leaves is reported to be taken orally against snake bites, whereas the leaves are applied as a poultice on the bite. In China, roots and leaves are used to treat dysentery, indigestion and eczema, and the fruits are used to treat anaemia, gum bleeding and anorexia.

**Observations** A small shrub 1-2 m tall; leaves obovate to oblong, 3.5-9 cm x 1.5-3 cm, chartaceous, base acute, apex obtuse, glabrous, petiole 3-10 mm long; inflorescence a lateral or axillary raceme, 6-10 mm long, few-flowered, with numerous bracts at the base; flowers 4-merous, yellowish-green. *E. laeta* generally occurs at higher elevations in mountainous regions. In Indo-China it flowers in December and mature fruits are present in April. It may well be grown at higher elevations in the Malesian region.

**Selected sources** 364, 849, 940, 1128, 1140.

**Embelia ribes Burm.f.**

*Fl. indica*: 62, t. 23 (1768).

**Synonyms** *Embelia garciniifolia* Wallich ex Miq. (1853).

**Vernacular names** Indonesia: kacembang (Sundanese). Malaysia: akar sulur kerang. Cambodia: chou pruc. Laos: reut jeum bang. Thailand: som kung (peninsular). Vietnam: th[uf]n m[ux]n, c[aa]y m[ux]n, d[aa]y ng[us]t.

**Distribution** From Sri Lanka and India eastward to Indo-China, Thailand and southern China

and southward to Peninsular Malaysia, Sumatra, Java and Borneo.

**Uses** The seeds or fruits are widely used as a vermifuge. An infusion of the roots is given to treat coughs and diarrhoea in Java. In East Kalimantan, the crushed fresh bark is used to repel leeches and as a fish poison. The young leaves, shoots and young fruits are consumed as a (cooked) vegetable or condiment. The ripe sour-sweet fruits are also eaten as a delicacy, mostly by children.

**Observations** A scandent shrub or liana up to 20 m long, young shoots densely pubescent; leaves lanceolate to oblanceolate, 3-9 cm x 1.5-3.5 cm, chartaceous, base round or acute, apex acuminate, glabrous, petiole 5-10 mm long; inflorescence a terminal panicle, 10-17 cm long; flowers (4-)5-merous, petals inside and outside pubescent, white. *E. ribes* occurs from sea-level up to 1500 m altitude, in forest or at forest borders. In Indo-China it flowers in February-March and mature fruits are present in March-October.



*Embelia ribes* Burm. f. - 1, flowering twig; 2, infructescence; 3, 4-merous flower; 4, 5-merous flower; 5, petal with stamen.

**Selected sources** 25, 97, 287, 565, 580, 687, 741, 829, 886, 940, 1035, 1066, 1128, 1140, 1156, 1361, 1364, 1512.

### ***Embelia robusta* Roxb.**

Hort. bengal.: 16 (1814).

**Synonyms** *Samara robusta* (Roxb.) Kurz (1877).

**Vernacular names** Vietnam: r[ef] d[oo]s[m].

**Distribution** From India eastward to Indo-China.

**Uses** In India the fruits are widely used as an anthelmintic.

**Observations** A scandent shrub or liana; leaves elliptical to obovate-elliptical, 4–9 cm × 2–5 cm, membranous, base rounded to cuneate, apex obtuse to shortly acuminate, initially pubescent below but soon glabrescent, petiole 4–7 mm long; inflorescence axillary, capituliform, 2–3 cm long; flowers 5-merous, anthers rotundate. *E. robusta* generally occurs at higher elevations. It may well be grown at higher elevations in the Malesian region.

**Selected sources** 202, 940, 1140.

### ***Embelia tsjeriam-cottam* (Roemer & Schultes) A. DC.**

Trans. Linn. Soc. 17: 131 (1834).

**Synonyms** *Embelia robusta* C.B. Clarke (1882) non Roxb.

**Distribution** From India and Sri Lanka eastward to Indo-China and Peninsular Malaysia.

**Uses** The dried fruits are used as a taeniafuge, antispasmodic and carminative. The bark of the root is reportedly used to treat toothache.

**Observations** A small shrub 1–2 m tall; leaves elliptical to obovate-elliptical, 6–9 cm × 3.5–5 cm, membranous, base rounded to shortly cuneate, apex shortly acuminate, petiole 8–12 mm long; inflorescence an axillary, cylindrical raceme, 3–6 cm long; flowers 5-merous, anthers acuminate or mucronate. *E. tsjeriam-cottam* generally occurs at lower elevations. In Indo-China it flowers in June–August.

**Selected sources** 287, 741, 940, 1140.

L.S.L. Chua & J.L.C.H. van Valkenburg

### ***Erythroxylum* P. Browne**

Civ. nat. hist. Jamaica: 278 (1756).

ERYTHROXYLACEAE

$x = 12$ ; *E. coca*:  $2n = 24$ , *E. novogranatense*:  $2n = 24$

**Major species** *Erythroxylum coca* Lamk, *E. novogranatense* (Morris) Hieron.

**Vernacular names** Coca (En, Fr).

**Origin and geographic distribution** *Erythroxylum*, which has about 250 species, is pantropical and its centre of diversity is in the Andes and the Amazon basin in South America. There are 6 indigenous species and 2 cultivated species in the Malesian region. The two species that are medicinally most important, *E. coca* and *E. novogranatense*, contain cocaine. They are native to South America, and were introduced into South-East Asia by the end of the 19th Century.

**Uses** The dried leaves of *Erythroxylum* spp. have been used in South America for at least 5000 years as a masticatory to allay hunger and fatigue, and as an element in religious rituals. Many medicinal applications have been recorded for coca in South America: infusions of coca leaves are used to treat indigestion and other stomach complaints, and altitude sickness. Coca is also reported to be effective against respiratory problems, teeth and gum complaints, malaria, eye irritations and sore throat, and it is valued as an aphrodisiac and a means of ensuring longevity. In some countries, e.g. Bolivia, the leaves are also used in infusions ('maté de coca') which are consumed like coffee or tea.

*E. coca* and *E. novogranatense* are a source of cocaine, which is used medicinally or as a narcotic. The therapeutic importance of cocaine has declined considerably. Cocaine (in its form as a hydrochloride) has long been used as a local anaesthetic in ear, nose and throat surgery. Although it is applied locally, systemic absorption may be marked, giving side effects (e.g. tachycardia, euphoria). It has been replaced by less toxic agents at present. Cocaine (hydrochloride) also used to be the sole active component in analgesic potions, or was added to oral solutions containing salts of morphine in order to potentiate the analgesic effect. These applications, however, are considered obsolete today. As a narcotic, cocaine is consumed in various ways. Coca paste, the initial product of extraction of the leaves, contains 40–70% cocaine (base), and is smoked. Cocaine (hydrochloride) is usually inhaled through the nose. 'Crack' or 'rock', a pure form of cocaine base, obtained by treating cocaine hydrochloride with bicarbonate, is smoked too ('freebasing').

The cola soft drink originally included coca extracts and caffeine-rich extracts from *Cola nitida* (Vent.) Schott & Endl., but nowadays coca leaves are only used to flavour the syrup after the co-

caine has been removed from them.

The leaves of *E. cuneatum* (Miq.) Kurz are reported to be used as fish poison in the Philippines (Luzon). *E. cuneatum* is also planted as an ornamental. The leaves of *E. ecarinatum* Burck are used medicinally, known as 'obat jaguar' in Sulawesi. In Papua New Guinea (Morobe Highlands), leaves are chewed and the sap ingested with traditional wood-ash salt against an upset stomach.

The wood of *Erythroxylum* has been used for fence posts and poles, flooring and sometimes for local house building, bridges, boat building and tool handles.

**Production and international trade** Coca is cultivated extensively in South America (Peru, Argentina, Bolivia, Colombia and Brazil), but also in Java, Sri Lanka, Taiwan, India and the United States. Few statistics are available on coca production. In South America, most of the coca leaves produced are consumed locally; very little is exported. Part of the coca leaves are used to make cocaine for illegal trafficking. Illegal imports of cocaine into the United States and Europe in 1988 were estimated at 100 t and 50–60 t, respectively.

**Properties** The most important constituent of *E. coca* and *E. novogranatense* is the tropane-ester alkaloid cocaine (methylbenzoyl ecgonine). The tropane part of the molecule, ecgonine, is biosynthetically derived from the amino acid ornithine. In the plant, cocaine occurs alongside other derivatives of tropine, e.g. cis- and trans-cinnamyl cocaine and tropa cocaine, and pyrrolidines e.g. hygrine, hygroline and cuscohygrine.

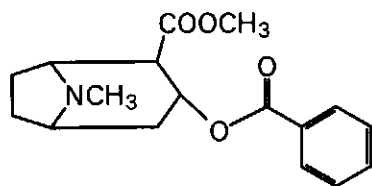
Cocaine and cinnamyl cocaine are considered to be the naturally occurring alkaloids of the coca plant, and have been identified in the roots, bark and leaves of several species and varieties. The younger leaves seem to accumulate cinnamyl cocaine as the main alkaloid, but the older leaves contain mainly cocaine. In an extensive study, the cocaine content in leaves of *E. coca* var. *coca* (30 samples) was found to range from 0.23–0.96%, with a mean of 0.63%, while the cocaine content in *E. coca* var. *ipadu* (6 samples) was lower: 0.11–0.41%, with an average of 0.25%. *E. novo-*

*granatense* var. *novogranatense* (3 samples) contained 0.55–0.93% cocaine, with an average of 0.77% and *E. novogranatense* var. *truxillense* (14 samples) 0.42–1.02%, with a mean of 0.72%. Cocaine (0.0003–0.3%) was detected in the leaves of 13 out of 29 wild *Erythroxylum* species, but no cocaine was detected in Old World species. The drug 'Cocae Folium' is also known to contain  $\alpha$ - and  $\gamma$ -truxilline and benzoyl ecgonine. These components are probably formed during harvesting and processing of coca leaves: truxillines are dimers of cinnamoyl cocaine, and hydrolysis of cocaine leads amongst others to benzoyl ecgonine.

Cocaine was first isolated around 1859. The compound displays several pharmacological activities, the best known being its local anaesthetic action. In this respect, cocaine acts by blocking  $\text{Na}^+$  ion channels in the neuron membranes, slowing down impulse conduction, and finally interrupting the propagation of action potentials. Because of cocaine's toxicity and addictive properties, a search for synthetic substitutes began. This is why the general molecular structure of cocaine can be found in several well-known local anaesthetics, e.g. procaine and tetracaine. Besides being a local anaesthetic, cocaine also acts as an adrenergic stimulant, by blocking the re-uptake of catecholamines (noradrenaline (= norepinephrine), dopamine) in both the peripheral and central nervous systems. Its euphoric properties are primarily due to inhibition of catecholamine uptake, particularly that of dopamine, at central nervous system synapses.

The chewing of coca leaves is a well-known practice in several South American countries. Alkalis added to the coca leaves facilitate the release of cocaine in its basic form. As a base, the ester is also volatile and can be smoked. Occasional use of cocaine results in feelings of mood elevation, vitality, mental clarity, sexual stimulation and reduction in appetite. However, chronic use can lead to anxiety, confusion, insomnia and impotence, and long-term heavy use can lead to paranoid psychosis. There are also indications that cocaine can be detrimental to fetal development. Regular use of cocaine finally leads to psychic dependence, but in contrast to heroin, not to physical dependence.

**Adulterations and substitutes** Although cocaine itself has never been synthesized, this has been done with similar compounds, of which procaine is the most important. Synthetic alkaloids with a similar action have almost completely replaced cocaine in medical use in the United States.



cocaine

**Description** Shrubs or small trees. Leaves simple, alternate, distichous, entire; stipules intrapetiolar, inserted semi-amplexicaulously, mostly entirely connate, rarely bifid, sometimes emarginate or 2-toothed at the apex, often bicarinate, long-persistent or early caducous, leaving a distinct scar, involute in bud, the margins leaving a subpermanent trace as 2 longitudinal lines on the upper leaf surface ('areolate'). Flowers in axils of leaves, solitary or in clusters, bisexual, 5-merous, actinomorphic, often heterodistylous, or even heterotristylous; pedicel slightly thickened, often only under the calyx, with 2 bracteoles at the base; calyx campanulate, persistent; petals free, alternating with the calyx lobes, caducous, quincuncial in bud, nearly always provided with an emarginate or 3-lobed ligule inserted on the apex of the claw; stamens in 2 whorls of 5, persistent, filaments towards the base connate into a staminal tube, anthers ellipsoid, basifixed, cordate at the base, 2-celled; ovary superior, (1-)3-celled, normally only 1 cell fertile, each cell with 1 ovule, ovules pendulous, anatropous, with a ventral raphe, styles 3, erect, free or partly connate or stigmas sessile. Fruit a drupe. Seed with or without endosperm, embryo oblong, erect, cotyledons flat to plano-convex, plumule absent, radicle distinct.

**Growth and development** *E. novogranatense* seed starts to germinate in 2-3 weeks after planting. Three months after germination the plants may have attained a height of 15 cm. The first harvest, usually 1-3 years after transplanting into the field, consists of the main shoot that is pruned to promote frame formation as in tea (*Camellia sinensis* (L.) Kuntze). Consecutive pruning is carried out when coca develops flower buds; this coincides with a maximum in harvestable leaves. Leaf harvesting brings *E. coca* plants from the vegetative into the reproductive development phase.

**Other botanical information** Cocaine-rich leaves are obtained from 4 taxa: *E. coca* var. *coca* (Bolivian or Huánoco coca), *E. coca* var. *ipadu* Plowman (Amazonian coca), *E. novogranatense* var. *novogranatense* (Colombian coca) and *E. novogranatense* var. *truxillense* (Rusby) Plowman (Trujillo coca). *E. coca* var. *coca* is considered the ancestor, while *E. novogranatense* var. *truxillense* is derived from it, and *E. novogranatense* var. *novogranatense* derived from *E. novogranatense* var. *truxillense*. Cultivation and selection has probably been the main selective force. *E. coca* var. *ipadu* is considered to be a cultivar selected

by man from *E. coca* var. *coca*. Wild populations of *E. coca* var. *coca* are found in the eastern Andes, but the other 3 taxa are only known as cultivated plants. Crosses between *E. coca* var. *coca* and *E. novogranatense* var. *novogranatense* have failed. Crosses between *E. coca* and *E. novogranatense* var. *truxillense* have been made, but gave abnormal, dwarfed progenies.

In the past *Erythroxylum* has been incorporated in different families, i.e. *Malpighiaceae*, *Linaceae* and *Erythroxylaceae*. In recent systems of classification it is widely accepted that the *Erythroxylaceae* differ from the *Malpighiaceae* by their un-winged fruits that do not split into 3 nut-like parts at maturity, and from the *Linaceae* by their ligulate petals and 3-locular ovary with only one fully developed locule.

**Ecology** *E. coca* var. *coca* is well adapted to the eastern Andes of Peru and Bolivia, an area of humid, tropical montane forest, whereas *E. coca* var. *ipadu* is cultivated in the lowland Amazonian basin. *E. novogranatense* is cultivated in drier regions in South America. However, *E. novogranatense* var. *novogranatense* is very adaptable to varying ecological conditions, and grows well in both humid and dry areas, and at low and higher altitudes. In Java, *E. novogranatense* has been cultivated from sea-level to 1000 m altitude, with best results at 400-600 m. In controlled environment studies, the optimum average daily temperature for leaf growth for both *E. coca* and *E. novogranatense* var. *novogranatense* was found to be around 27°C, whereas leaf growth was generally higher at photosynthetic photon flux densities of 250 or 400  $\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$  than at 155  $\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$ . Environmental effects on the cocaine concentration in the leaves were smaller, so that total cocaine production per plant was largely a function of leaf mass, with environmental conditions that stimulated leaf growth giving higher cocaine yields. Both species grow on soils with low pH, and a greenhouse study has shown that the optimum pH for biomass accumulation of *E. coca* and *E. novogranatense* is as low as 3.5 and 4.7-6.0, respectively.

**Propagation and planting** *E. novogranatense* var. *novogranatense*, *E. novogranatense* var. *truxillense* and *E. coca* var. *coca* have to be reproduced by seed, because vegetative propagation is difficult. However, *E. coca* var. *ipadu* does not produce seed and is propagated by stem cuttings. It cannot reproduce without human interference. Cultivated *E. novogranatense* var. *novogranatense* produces abundant seed and is easy to propagate.

Seed viability decreases rapidly. Germination percentages of *E. coca* and *E. novogranatense* seed were found to decrease from around 95% and 89% directly after harvesting to 29% and 0%, respectively, after 24 days' storage at 4°C. Coca seedlings are usually sown in shaded nurseries and transplanted to the field when they are about one year old and 20–25 cm tall. In the field, they are planted at a spacing of 1–2 m. The actual time of transplanting and the spacing of the plants varies with climatic factors and whether coca is interplanted or cultivated as a sole crop.

**In vitro production of active compounds** It is possible to obtain cocaine and other alkaloids from *E. coca* var. *coca* shoot tissue cultures. The levels of cocaine produced have been found to be 50% of that produced by the parent plant, but within the range of the species. Whereas in *Solanaceae* the tropane alkaloids are biosynthesized in the roots, in *E. coca* tropane alkaloids (e.g. cocaine) can be produced from rootless explants.

**Husbandry** In mountain areas, coca is often grown on terraced fields. The fields are often manured with ash or plant material to sustain production. Older plants are severely pruned when leaf production decreases, after which regrowth occurs, but coca plantings are usually renewed after 20 years. *E. coca* var. *ipadu* is grown by semi-nomadic Amazonian tribes in isolated small plots, and constitutes an important part of their shifting cultivation system.

**Diseases and pests** *Erythroxylum* spp. are susceptible to Fusarium wilt, caused by the host-specific fungus *Fusarium oxysporum*, which is considered a potential mycoherbicide to combat illegal coca production. Symptoms are vascular wilt and permanent defoliation.

**Harvesting** The first harvest of coca takes place at 1–3 years after transplanting. In Java, a first harvest can be expected within a year after transplanting. The leaves have to be stiff and easily detachable to be harvested. Leaves can be harvested every 50–60 days in the rainy season, but when it is drier, they are usually harvested every 3–4 months. The leaves should be pinched from the plant, not ripped off.

**Yield** Annual yields of 1.7–2.2 t dry coca leaves per ha have been reported. The highest yields are obtained between 3 and 10–15 years after transplanting. In the Asian coca plantations, yields are about 700 kg dry coca leaves per ha, with the leaves containing up to 1.4% cocaine.

**Handling after harvest** When the leaves of coca are used for cocaine production, they are dried

in the sun or artificially. Temperatures should not exceed 40°C, because the cocaine content of the leaves decreases at higher temperatures. The leaves are considered dry when they can be broken. In Asia, dried leaves are ground, pulverized and packed in plastic bags, while in South America they are pressed into bales of about 50 kg for transport. Leaves intended for chewing in South America must keep their form and colour and are dried more carefully.

**Genetic resources and breeding** The gene pool of cultivated *E. coca* has changed little over 5000 years, and wild or abandoned *E. coca* shows no morphological and genetic differences from cultivated *E. coca*. No germplasm collection and breeding programmes are known to exist.

**Prospects** The chewing of coca leaves is very important in Indian culture in parts of South America, and therefore coca will remain important there in the future. Elsewhere, the crop will only be important in regions where illegal production of narcotics is possible, because the world market for cocaine for medical use is limited, as synthetic agents have almost completely replaced the use of cocaine as a local anaesthetic.

**Literature** [1] Acock, M.C., Lydon, J., Johnson, E. & Collins, R., 1996. Effects of temperature and light levels on leaf yield and cocaine content in two *Erythroxylum* species. *Annals of Botany* 78(1): 49–53. [2] Bohm, B.A., Ganders, F.R. & Plowman, T., 1982. Biosystematics and evolution of cultivated coca (*Erythroxylaceae*). *Systematic Botany* 7(2): 121–133. [3] Chung, R.C.K., 1996. *Erythroxylaceae*. In: Soepadmo, E., Wong, K.M. & Saw, L.G. (Editors): *Tree Flora of Sabah and Sarawak*. Vol. 2. Forest Research Institute Malaysia, Sabah Forestry Department, and Sarawak Forestry Department, Kuala Lumpur, Malaysia. pp. 167–174. [4] De Jong, A.W.K., 1948. Coca. In: Van Hall, C.J.J. & Van de Koppel, C. (Editors): *De Landbouw in de Indische Archipel [The agriculture in the Indonesian Archipelago]*. Vol. 2a. Van Hoeve, 's-Gravenhage, the Netherlands. pp. 866–888. [5] Martin, R.T., 1970. The role of coca in the history, religion, and medicine of South American Indians. *Economic Botany* 24: 422–438. [6] Payens, J.P.D.W., 1958. *Erythroxylaceae*. In: van Steenis, C.G.G.J. (Editor): *Flora Malesiana*. Series 1, Vol. 5(4). Noordhoff-Kolff N.V., Djakarta, Indonesia. pp. 543–552. [7] Plowman, T., 1982. The identification of coca (*Erythroxylum* species): 1860–1910. *Botanical Journal of the Linnean Society* 84: 329–353. [8] Plowman, T. & Rivier, L., 1983. Cocaine and cinnamoylcocaine content of

*Erythroxylum* species. *Annals of Botany* 51: 641–659. |9| Schröder, R., 1991. Kaffee, Tee und Kardamom: Tropische Genussmittel und Gewürze; Geschichte, Verbreitung, Anbau, Ernte, Aufbereitung [Coffee, tea and cardamom: tropical stimulants and spices; history, distribution, cultivation, harvest, processing]. Verlag Eugen Ulmer, Stuttgart, Germany. pp. 111–115. |10| Simpson, B.B. & Conner-Ogorzaly, M., 1986. *Economic Botany: plants in our world*. McGraw-Hill Book Company, New York, United States. pp. 395–398.

#### *Selection of species*

#### ***Erythroxylum coca* Lamk**

Encycl. méth. Bot. 2: 393 (1786).

**Synonyms** *Erythroxylum peruvianum* Prescott (1847), *Erythroxylum bolivianum* Burck (1890).

**Vernacular names** Coca, Peru coca, Peruvian coca (En).

**Distribution** *E. coca* var. *coca* (Bolivian or Huánoco coca) is widely cultivated in the Andean region, where it locally also occurs wild. It is not easy to cultivate elsewhere, and it is little known in other parts of the world. In South-East Asia, it is only grown in botanical gardens, not as a crop. *E. coca* var. *ipadu* (Amazonian coca) is only found as cultivated plant in Amazonian lowland rain forest areas.

**Uses** *E. coca* is the most important commercial coca species, cultivated for the legal or illegal production of cocaine. The leaves are used as a masticatory by millions of Indians in South America.

**Observations** A shrub or small tree, with very prominent, sometimes warty lenticels on the branches; leaves mainly at the end of the twigs, early caducous, broad-elliptical, 3–8 cm × 2–4 cm, base cuneate, apex acuminate or rounded with a mucronate tip; flowers in clusters of 6–12, rarely more, in the axils of leaves or rammenta, pedicel 4–6 mm, calyx with a 0.2–1 mm long tube and 5 lobes, triangular-ovate, 1–2 mm long, acute, green, petals oblong, 4–4.5 cm × 2 mm, yellow or yellowish green; fruit oblong-ovoid, pointed, 7–10 mm × 3–4.5 mm, red. *E. coca* has heterodistylous flowers, and self-pollination or pollination between plants of the same flower type gives few seeds. In Bogor it produces abundant fruit but not much foliage.

**Selected sources** 193, 379, 432, 466, 549, 555, 682, 683, 684, 880, 1119, 1145, 1146, 1147, 1167, 1178, 1277, 1278, 1301.

#### ***Erythroxylum novogranatense* (Morris) Hieron.**

Bot. Jahrb. 20, Beibl. 49: 35 (1895).

**Synonyms** *Erythroxylum coca* Lamk var. *novogranatense* Morris (1889), *Erythroxylum coca* Lamk var. *spruceanum* Burck (1890), *Erythroxylum truxillense* Rusby (1900).

**Vernacular names** Java coca, Truxillo coca (En).

**Distribution** *E. novogranatense* var. *novogranatense* (Colombian coca) is native to Colombia and Venezuela, but its adaptability and easy propagation has led to a wide distribution over the Old and New World tropics. *E. novogranatense* was introduced in the Bogor Botanical Gardens in 1875, and by 1888 large quantities of seed were already being distributed. In South-East Asia, it has been grown in Peninsular Malaysia, western and eastern Java, northern Borneo, northern Sulawesi (Minahasa) and the Philippines (Luzon). *E. novogranatense* var. *truxillense* (Trujillo coca) is grown in arid areas in northern Peru. Neither of these varieties is known from wild populations.

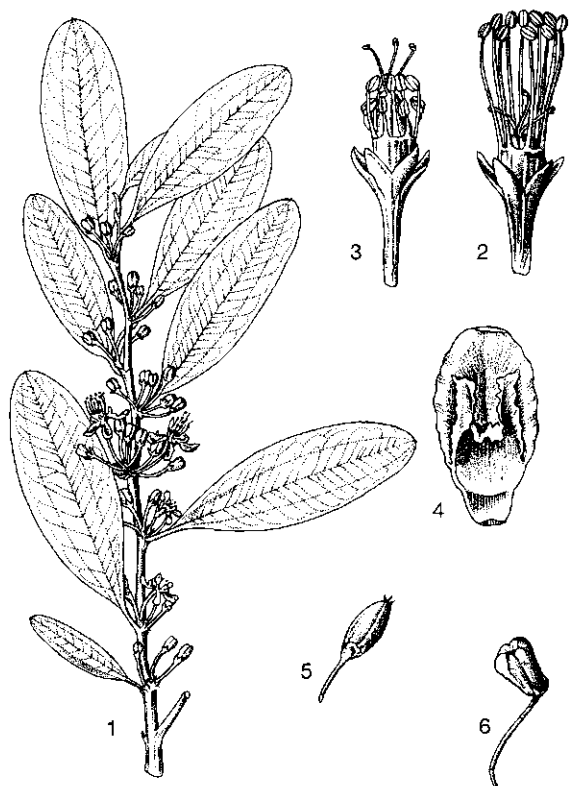
**Uses** The medicinal uses of *E. novogranatense* are similar to those of *E. coca*. It used to be grown as a hedge plant in Malesia, up to 750 m altitude, for its bushiness and the contrast of its light green foliage with the bright red berries, but its cultivation is now prohibited. Although of importance as a plantation crop for the Malesian region in the past, it has since long lost its importance. *E. novogranatense* var. *truxillense* is highly regarded for its flavour and cocaine content, and has long been used for coca-flavoured beverages.

**Observations** A shrub of 1–3 m tall, with minute lenticels on the branches; leaves abundant along the twigs, rather persistent, obovate-oblong, (2–)3–6(–7) cm × 1–3 cm, base attenuate, apex rounded or sometimes emarginate, always with a mucronate tip or notch; flowers in clusters of 1–4(–8), pedicel 5–10 mm, calyx with a 1–1.5 mm long tube and 5 lobes, triangular-ovate, 1.5–2 mm × 1 mm, acuminate, petals oblong, convex, (3.5–)4(–5) mm × 2 mm, white or greenish-white; fruit ellipsoid-oblong, red. *E. novogranatense* has heterodistylous flowers, but, in contrast to *E. coca*, at least the long-styled type is self-compatible.

**Selected sources** 193, 202, 284, 328, 466, 549, 555, 580, 682, 683, 684, 1119, 1145, 1146, 1147, 1167, 1277, 1278.

R.C.K. Chung & M. Brink





*Erythroxyllum novogranatense* (Morris) Hieron. — 1, flowering twig; 2, short-styled flower with petals removed; 3, long-styled flower with petals removed; 4, petal showing 3-lobed ligule-like appendage; 5, fresh fruit; 6, dried fruit.

## Euphorbia L.

Sp. pl. 1: 450 (1753); Gen. pl. ed. 5: 208 (1754).

### EUPHORBIACEAE

$x = 9, 10, 11$ ; *E. antiquorum*:  $2n = 60$ , *E. barnhartii*:  $2n = 40$ , *E. heterophylla*:  $2n = 54, 56$ , *E. hirta*:  $n = 8, 9, 10$ , *E. neriifolia*:  $2n = 60, 80, 90, 180$ , *E. prostrata*:  $n = 9, 2n = 20$ , *E. thymifolia*:  $n = 9$ , *E. tirucalli*:  $2n = 20$

**Major species** *Euphorbia antiquorum* L., *E. hirta* L., *E. thymifolia* L.

**Vernacular names** Spurge (En). Euphorbe (Fr).

**Origin and geographic distribution** *Euphorbia* comprises over 2000 species and occurs worldwide, though most species are found in tropical, subtropical and warm temperate regions. There are 35 species native to South-East Asia: Vietnam has 24 species, Thailand 25, Sumatra 6, Java 5,

Borneo 5, the Philippines 6, Sulawesi 5, the Lesser Sunda Islands 11, the Moluccas 7 and New Guinea 15. Australia has 45 species.

**Uses** The latex of many *Euphorbia* species is used medicinally as a purgative, has antidiarrhoeal and antibacterial properties, and is used to treat boils, warts, wounds and other skin disorders. It is also often used as a fish poison and has insecticidal and fungicidal properties. The woody central part of the stem of some succulent species is applied as a remedy for dysentery. The leaves of several *Euphorbia* species are used to treat asthma. Traditional pharmacies often include material from several *Euphorbia* species.

Some species are cultivated as ornamentals or living fences. The latex of some species can be converted into fuel and has been investigated as a possible energy source.

**Properties** Phytochemical investigations of the lipophilic constituents of *E. tirucalli* has revealed the presence of steroids (sitosterol, stigmasterol, campesterol) and fatty acids (palmitic acid, linoleic acid).

A lectin isolated from the latex of *E. neriifolia* agglutinated trypsinized human and rabbit erythrocytes. Untreated sheep erythrocytes did not agglutinate, but sialidase-treated sheep erythrocytes did. Galactose and sugars containing galactose inhibited the haemagglutination with increased  $\beta$ -anomeric specificity. The lectin possesses mitogenic activity with murine spleen lymphocytes but it does not inhibit protein synthesis in the rabbit reticulocyte lysate assay. The lyophilized aqueous extract of *E. hirta* has central analgesic properties. A dose of 20 mg/kg produced action against chemical stimuli, whereas one of 25 mg/kg produced action against thermic stimuli; the effects were inhibited by a pretreatment with naxolene, a specific morphinic antagonist compound. At the sedative doses of 100 and 400 mg/kg an antipyretic activity was obtained on the yeast-induced hyperthermia model. Significant and dose-dependent anti-inflammatory effects were also observed on an acute inflammatory process in a carrageenin-induced oedema test in rats from a dose of 100 mg/kg.

The ethyl acetate fraction of the ethanol extract of *E. prostrata* administered orally to rats at 200 mg/kg inhibited 76% of acute carrageenin-induced paw oedema. The ethyl acetate extract and a fraction (KSE-23) isolated chromatographically from it, showed significant anti-inflammatory activity when applied topically in a murine model of carrageenin-induced footpad oedema in mice. KSE-23

was found to be more potent than indomethacin given by the same route.

The extracts of *E. tirucalli* have markedly enhancing effects on the activation of latent Epstein-Barr virus (EBV) genomes in the EBV carrying lymphoblastoid cells and also on EBV-induced transformation of human lymphocytes. Soil and drinking water taken around the plants have the same enhancing effects and are a serious risk to humans in Africa. Various doses of powdered *E. prostrata* administered orally to male albino rabbits produced significant hypoglycaemic effects in normal rabbits, but had no effect in alloxan-diabetic rabbits. The methanol extract also decreased the blood glucose of normal rabbits. The methanol extract of *E. hirta* was effective against dysentery-causing *Shigella* spp. using the vero cell line, and it had no cytotoxic effects. This is attributable to quercitrin, a flavonoid, isolated from a lyophilized decoction of *E. hirta*. At doses of 50 mg/kg quercitrin is known to show preventive activity against diarrhoea induced by castor oil and prostaglandin E2 in mice, but not when magnesium sulphate is used as a cathartic agent. The aqueous extract of *E. hirta* has been found to strongly reduce the release of prostaglandins. Additionally, the extracts exerted an inhibitory effect on platelet aggregation. In organ bath tests with ileum preparations shikimic acid and choline extracted from the aerial parts of *E. hirta* had relaxing and contracting properties, respectively. The methanol extract of leaves and stems of *E. hirta* inhibited the activity of angiotensin-converting enzyme by 90% at 500 µg and 50% at 160 µg. The effect of the extract on thirst was examined in Wistar rats. The extract (10 mg/100 g, intraperitoneally) significantly decreased the amount of water the rats consumed. Extracts of whole plant material are also reported to have oestrogenic activity. Extracts of *E. hirta* showed anti-microbial activity against *Candida albicans*, *Escherichia coli*, and *Staphylococcus aureus*. Leaf extracts of *E. hirta* severely inhibited sporulation in the hyphomycete *Helminthosporium* sp. Finally, drying

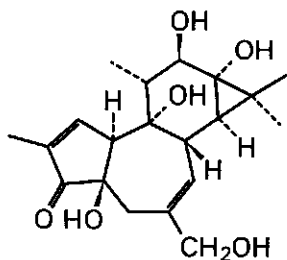
*E. hirta* prior to extraction considerably reduces the cytotoxic activity of certain of its extracts.

The latex of *Euphorbia* is often highly irritant to the eyes and must be washed away immediately. It can cause irritation and vesication from contact and emesis and purgation from ingestion. These effects are caused by a large number of different esters of the tetracyclic diterpenoid phorbol, many of which have also been shown to act as tumour promoters (cocarcinogens). The latex of *E. tirucalli* contains the irritant and/or cocarcinogenic constituents ingenane and tiglane-type diterpene esters derived from the parent alcohols ingenol, phorbol and resiniferonol. Furthermore, the latex is an emulsion of about 30% terpenes in water.

The latex hydrocarbon is largely a C<sub>30</sub> triterpenoid which can be cracked to make high octane gasoline. The gross energy value of *E. tirucalli* biomass is 17 600 kJ/kg. Biomass can be converted into gas, liquid fuels and solid fuels such as pellets, briquettes and charcoal.

Rubber production from the latex of *E. tirucalli* was investigated early in the 20th Century, but continuous latex production proved difficult and its high resin content limited the profitability of the rubber production.

**Description** Monoecious or rarely dioecious, annual, biennial or perennial herbs, shrubs or trees, sometimes succulent, armed or unarmed, with latex. Leaves alternate, opposite or verticillate, sometimes much reduced, simple, uniform or heterophyllous, margin entire, toothed or lobed; stipules present or absent, sometimes modified into spines. Flowers unisexual, one female flower together with several male flowers enclosed within a small, cupuliform, glanduliferous involucre, the whole structure termed a 'cyathium', functioning as a single, bisexual flower; cyathia solitary or combined into corymbs or cymes, occasionally unisexual; bracteoles usually ligulate and fringed. Male flowers reduced to single stamens; anthers with 2 cells. Female flowers pedicellate, reduced to single pistils or with a minute perianth; ovary superior, during maturation on an accrescent pedicel and often nodding outside the cyathial cup, 3-locular with 1 ovule per locule, styles 3, partly fused, entire to 2-fid. Fruit a smooth to tuberculate, sometimes fleshy capsule, splitting open elastically, first dehiscing septicidally and then loculicidally. Seeds with endosperm, with or without caruncle. seedling with epigeal germination; cotyledons leafy, usually elliptical to ovate, glabrous; hypocotyl elongated, glabrous, epicotyl absent or very short; first two leaves opposite.



phorbol

**Growth and development** In Java, *E. atoto*, *E. hirta*, *E. prostrata* and *E. thymifolia* flower throughout the year, *E. barnhartii* usually in September–November, whereas *E. tirucalli* rarely flowers in October but never sets fruit. In most *Euphorbia* the capsules dehisce with force and ejaculate their seeds which are thus dispersed over some distance. Ants may act as a dispersal agent of some *Euphorbia*, for example in *E. hirta* and *E. heterophylla*. *E. hirta* produces up to 3000 seeds per plant which show a high germination rate. *E. tirucalli* is one of the very few known plants combining a Crassulacean acid metabolism (CAM) pathway of its branches and twigs with the C<sub>3</sub> metabolism pathway of its leaves. Consequently it is very drought-resistant and very efficient in photosynthesis.

**Other botanical information** Some authors have separated *Euphorbia* into a number of distinct genera, usually based upon peculiarities of the cyathial structure. Others recognize these entities at infrageneric level. In South-East Asian and Australian literature one may, for example encounter the genus *Chamaesyce* next to *Euphorbia*; here, however, *Chamaesyce* is regarded as a subgenus.

Some of the American *Euphorbia* species that have been introduced into South-East Asia are sometimes referred to as members of the genus *Poinsettia*, or of the subgenus *Poinsettia*. All tree-like, succulent, spinose species belong to the subgenus *Euphorbia*. *E. cyathophora* and *E. heterophylla* are now regarded as two distinct species, but have not been treated as such in older literature. Therefore much of the older information cannot be referred to either of these species. Both species are strikingly variable in leaf shape. *E. buxoides* Radcl.-Sm., which closely resembles *E. plumerioides*, is widely planted as a hedge and boundary marker in the highlands throughout New Guinea. Its bark is chewed to induce vomiting and acts as a poison antidote. Leaves of *E. vachellii* Hook. & Arn. (syn. *E. serrulata* Reinw. ex Blume non Thuill.) are used internally to treat catarrh in Papua New Guinea.

**Ecology** The succulent, spinose *Euphorbia* species generally occur in dry places, on rocky or sandy soils, occasionally in dry forest. Most herbaceous *Euphorbia* species are commonly found in waste places and as a weed in fields and gardens.

**Propagation and planting** The weedy medicinal *Euphorbia* species produce seed in abundance and reproduce spontaneously. *E. tirucalli* can easily be propagated by stem cuttings, which greatly

facilitates its planting as a hedge. Cuttings can best be taken from older branches; they are left to dry for 1 day before planting. A density of 10 000–20 000 plants/ha is normal when grown as fuel crop.

**In vitro production of active compounds** Isolation and culturing of protoplasts of *E. tirucalli* has been successful. Callus tissue of *E. tirucalli* produces the 4,4'-dimethyl sterols euphol and tirucalol. Euphol is the principal terpene in the latex of the plant, whereas tirucalol is found in intact parental tissue. The highest yield of sterol has been obtained on a Murashige and Skoog medium supplemented by indole acetic acid (IAA) and kinetin; incorporation of squalene at 1.0 mg/l enhanced sterol production.

**Husbandry** The herbaceous medicinal *Euphorbia* species are in general weeds. *E. heterophylla* is a shade-tolerant, pantropical weed and as such its control is more important than its husbandry. It may be troublesome in crops like maize, cotton, cowpea and soya bean. Crops need to be kept free from *E. heterophylla*, especially in the early phases of development. Well-established *E. heterophylla* can depress yields greatly, for instance by as much as 75% in cowpea. Fresh *E. heterophylla* seed germinates readily under tropical conditions, but remains dormant under temperate circumstances and then both light and temperature influence dormancy breaking. *E. prostrata* shows strong allelopathic effects on a number of crops. Its aqueous extract, decaying residues and root exudates have been found to be inhibitory to several species. Under semi-arid conditions the regrowth of coppice of *E. tirucalli* is excellent.

**Harvesting** In general stems or whole plants of herbaceous *Euphorbia* are harvested to be used fresh. *E. tirucalli* can be coppiced at 20–30 cm height.

**Yield** When planted at a spacing of 1 m × 1 m *E. tirucalli* in Thailand produced 120 t/ha fresh material and 14 t/ha dry matter after 1 year, yielding 40–88 kg of crude oil, 135–213 kg of sugar and 1.8 t of bagasse. After 1.5 years with 6 trimmings a year, 148 t/ha of biomass (i.e. 17.5 t dry weight) could be harvested. It was calculated in Japan that 1–2 t of crude oil could be obtained per ha per year from *E. tirucalli*. A daily production of biogas from *E. tirucalli* of about 31 m<sup>3</sup>/ha (226 l/kg dry matter) is considered possible based on an annual production of 500 t biomass/ha by means of a two-phase methane fermentation process; a one-phase process yields 175–323 l of biogas per kg dry matter.

**Genetic resources and breeding** Given the common and widespread occurrence of most medicinal *Euphorbia* species and their weedy nature, the risk of genetic erosion appears very limited. The same holds for the succulent *Euphorbia* species planted as ornamentals. Neither germplasm collections nor breeding programmes are known of.

**Prospects** Phorbols and phorbol-esters are extremely harmful to the skin and mucous membranes, and also reported to be tumour promoters. Therefore, the medicinal prospects of many *Euphorbia* species are very limited. Acetylated esters of phorbols, however, may play a role in the search for possible anti-tumour compounds.

*E. tirucalli* may hold promise for energy production through biomass utilization or use of the crude oil extracted from the plants.

**Literature** |1| Calvin, M., 1987. Fuel oils from euphorbs and other plants. *Botanical Journal of the Linnean Society* 94: 97–110. |2| Galvez, J., Crespo, M.E., Jimenez, J., Suarez, A. & Zaruelo, A., 1993. Antidiarrhoeic activity of quercitrin in mice and rats. *Journal of Pharmacy and Pharmacology* 45(2): 157–159. |3| Hiermann, A. & Bucar, F., 1994. Influence of some traditional medicinal plants of Senegal on prostaglandin biosynthesis. *Journal of Ethnopharmacology* 42(2): 111–116. |4| Jantarawatit, S., Reutrakul, V. & Ratanabanangkoon, K., 1997. Estrogenic activity found in the herb *Euphorbia hirta* (nam nom rat chasee). *Mahidol University Annual Abstracts* 192. |5| Lanhers, M.C., Fleurentin, J., Dorfman, P., Mortier, F. & Pelt, J.M., 1991. Analgesic, antipyretic and anti-inflammatory properties of *Euphorbia hirta*. *Planta Medica* 57(3): 225–231. |6| Mizuno, F. et al., 1986. Epstein-Barr virus-enhancing plant promoters in East Africa. *AIDS Research* 2. Supplement 1: S151–S155. |7| Nguyen Nghia Thin, 1989. Useful plants of Euphorbiaceae in the flora of Vietnam. *Forestry Revue*, Hanoi 1989: 29–30. |8| Shingla, A.K. & Pathak, K., 1990. Topical antiinflammatory effects of *Euphorbia prostrata* on carrageenan-induced footpad oedema in mice. *Journal of Ethnopharmacology* 29(3): 291–294. |9| Van Damme, P., 1989. *Studie van Euphorbia tirucalli* L., morfologie, fysiologie, teeltvoorwaarden [Study of *Euphorbia tirucalli* L., morphology, physiology, agronomy]. Thesis, Gent University, Belgium. 375 pp. + appendices. |10| Wilson, A.K., 1981. *Euphorbia heterophylla*: a review of distribution, importance and control. *Tropical Pest Management* 27: 32–38.

### *Selection of species*

#### ***Euphorbia antiquorum* L.**

Sp. pl. 1: 450 (1753).

**Vernacular names** Malayan spurge tree (En). Euphorbe des anciens (Fr). Indonesia: sudu-sudu, susudu, susuru. Malaysia: sesudu, sudu-sudu (Peninsular). Cambodia: chan bat day. Laos: lep nguak. Thailand: kalam-phak (eastern), khia phaa (northern), salatdai paa (central). Vietnam: x[uw]low]ng r[oof]ng, x[uw]low]ng r[oof]ng claj]nh.

**Distribution** Southern India, Sri Lanka, Burma (Myanmar), Laos, Vietnam, Thailand and Peninsular Malaysia; locally cultivated within the area of natural occurrence, in Java and possibly also in the Philippines. Also sporadically cultivated as an ornamental or hedge plant in other tropical and subtropical regions, and as an indoor plant in temperate areas.

**Uses** The poisonous milky latex or other plant parts (e.g. root bark) are taken as a drastic purgative and induce vomiting. The latex is applied externally to swellings, boils, warts and other skin affections. When mixed with oil, it is a rubefacient embrocation for rheumatism. It may also be used to treat toothache, earache and asthma. The dried heartwood is an antipyretic and used in applications to treat toothache, and is used in Cambodia as a febrifuge and against dysentery. In India, the plant is employed for nervine diseases and dropsy. A saline extract of the stem shows antibiotic activity. Furthermore, the plant is used as a fish poison and shows insecticidal properties. *E. antiquorum* is often planted for ornamental purposes and as a fence. In Java, young twigs that have been properly boiled, soaked in water and covered with sugar, have been eaten as sweetmeats.

**Observations** A spiny, succulent shrub or small tree up to 6(–9) m tall, branches tufted, ascending, 3–5-ribbed, with 3–5 mm long, persistent spines on the exsculptate ribs, young branches constricted at the joints; leaves early caducous, obovate-spathulate or subrhomboid, 0.3–1.3 cm long, cuneate to attenuate at base, rounded to emarginate at apex, sessile; inflorescence borne on the exsculptate ribs, composed of 1–7 cyathia, peduncles short, rigid, 0–2 times forked, bracts scale-like, about 2.5 mm long, pale green; cyathia with 5 pale yellow glands, 4–4.5 mm wide, anthers dark red; capsule obtusely trigonous, 8–12 mm across, smooth. *E. antiquorum* is found in dry, open, evergreen forest, in scrubby vegetation, on rocky limestone hills and on sandy soils, up to 800 m altitude.

**Selected sources** 97, 202, 287, 580, 921, 1035, 1126, 1128, 1135, 1181, 1185.

### **Euphorbia atoto J.G. Forster**

Fl. ins. austr.: 36 (1786).

**Synonyms** *Euphorbia laevis* Poir. (1812), *Euphorbia halophila* Miq. (1852), *Chamaesyce atoto* (J.G. Forster) Croizat (1936).

**Vernacular names** Indonesia: jelutung laut (Java), susuan i lawanan (Sulawesi). Philippines: tairas (Filipino), lamingo (Bagobo). Thailand: ma phraao nok khao (south-western), nam nom raatchasee thale (peninsular). Vietnam: c[aa]y thu[oolc d]offi.

**Distribution** Indo-China, China, Japan, Thailand, throughout the Malesian region, northern Australia and Polynesia.

**Uses** In Indo-China the latex is used as an emmenagogue and an abortifacient. In New Caledonia, seawater in which the plant has been soaked and trampled is often used as a purgative.

**Observations** An unarmed, perennial, glabrous herb with prostrate to ascending stems; leaves opposite, slightly leathery, elliptical to ovate-oblong, 1.2–3 cm long, obliquely subcordate at base, acute at apex, margin entire, glaucous, shortly petiolate, stipules entire; bracts leaf-like; cyathia 1–3 together in terminal cymes, glands 4, transversely elliptical to oblong, yellow, with white appendages, anthers yellow; capsule obtusely trigonous, about 2.5 mm in diameter, smooth. *E. atoto* is a typical beach plant confined to sandy beaches and coral reefs.

**Selected sources** 97, 155, 865, 921, 1128, 1178, 1181, 1380.

### **Euphorbia barnhartii Croizat**

Euph. antiq. offic.: 54 (1934).

**Synonyms** *Euphorbia trigona* Roxb. (1832) non Miller.

**Vernacular names** Indonesia: sudu-sudu, susudu, susuru. Malaysia: sesudu, sudu-sudu (Peninsular). Philippines: sorog-sorog (Tagalog).

**Distribution** Native origin possibly Java; planted in Peninsular Malaysia, Java, the Philippines, possibly also elsewhere.

**Uses** The latex is used to treat earache, and has been reported as poisonous. Pounded leaves are applied as a poultice to boils. *E. barnhartii* is fairly commonly cultivated for ornamental purposes and as a hedge.

**Observations** A spiny, succulent shrub or small tree up to 5 m tall, main stem 4–5-ribbed, branches 3-ribbed, with 3–5 mm long, readily ca-

duous spines on the exsculptate ribs; leaves early caducous, oblong-obovate, 4–15(–30) cm long, apex rounded with a recurved mucro; inflorescence borne on the exsculptate ribs, composed of 3–7 cyathia on short, rigid, forked peduncles, bracts 2–7 mm long; cyathia with 5 shiny yellow glands, 3.5–4.5 mm broad, anthers red; capsule obtusely trigonous, ovoid, 4–7 mm in diameter, smooth. *E. barnhartii* is found as an escape from cultivation, e.g. on limestone and lava rocks, at low altitude.

**Selected sources** 97, 202, 296, 580, 933, 1126.

### **Euphorbia cyathophora Murray**

Comm. Göttingen 7: 81, t. 1 (1786).

**Synonyms** *Euphorbia heterophylla* L. var. *cyathophora* (Murray) Griseb. (1859), *Poinsettia cyathophora* (Murray) Klotzsch & Garcke (1859), *Poinsettia graminifolia* (Michx.) Millsp. (1909).

**Vernacular names** Painted leaf, red milkweed, wild poinsettia (En). Philippines: pintado (Tagalog). Vietnam: tr[aj]ng nguy[ee]n ghi ta.

**Distribution** Native to the southern United States, Mexico and perhaps also the Greater Antilles, but now cultivated and commonly escaping and naturalizing throughout the tropics. Within Malesia occurring in Peninsular Malaysia, Java, the Lesser Sunda Islands, the Philippines and Papua New Guinea.

**Uses** In Peninsular Malaysia, a decoction of the roots and bark is used to treat ague. In Mexico, the stem latex is applied against erysipelas. In the West Indies, the latex is applied to corns. In Guatemala, a decoction of the flowers is taken as a pectoral. In Central America, the roots are applied as an emetic and cathartic and administered in very small doses. In Brazil, the leaves are used to produce the red dye porcetin. *E. cyathophora* is sometimes planted for ornamental purposes.

**Observations** An annual or facultative perennial, unarmed herb up to 1.5 m tall; leaves alternate, ovate or lanceolate to fiddle-shaped, 4–10 cm × 1–5 cm, base cuneate to rounded, apex obtuse or acute, margin entire to serrulate or dentate, glossy green, lower surface pilose, petiole up to 1.7 cm long, glabrous or sparsely pilose; inflorescence a terminal, clustered cyme of cyathia, bracts similar to the leaves but progressively smaller, with a red blotch at base or entirely red; cyathia with 1(–2) peltate, funnel-shaped glands with an elliptical, 2 mm wide opening, anthers yellow; capsule deeply 3-lobed, 4–5 mm × 3.5–5 mm, smooth; seeds ovoid, sharply tuberculate. *E. cyathophora* is found in waste places and roadsides, up to 1800 m altitude.

**Selected sources** 97, 217, 372, 864, 865, 979, 1178, 1183, 1185, 1186, 1582.

### ***Euphorbia heterophylla* L.**

Sp. pl. 1: 453 (1753).

**Synonyms** *Euphorbia prunifolia* Jacq. (1798), *Poinsettia geniculata* (Ort.) Klotzsch & Garcke ex Klotzsch (1859), *Poinsettia heterophylla* (L.) Klotzsch & Garcke ex Klotzsch (1859), *Euphorbia taiwaniana* Ying (1987).

**Vernacular names** Japanese poinsettia, Mexican fireweed, mole plant (En). Malaysia: pekapar (Peninsular). Thailand: yaa yaang (general), bai taang dok, luuk khoei taai mae yai tham sop (Bangkok). Vietnam: c[or] m[ur].

**Distribution** Native to Central and South America, but nowadays naturalized throughout the tropics. Within Malesia not yet reported from the Philippines.

**Uses** *E. heterophylla* may cause poisoning in livestock. The young leaves are sometimes eaten as a vegetable, but may act as a laxative.

**Observations** An annual or facultative perennial, unarmed herb up to 110(-200) cm tall; leaves

alternate, lanceolate or ovate to distinctly fiddle-shaped, 3-14 cm × 0.5-7 cm, base cuneate to rounded, apex obtuse to slightly acuminate, margin entire to serrulate, dull green, glabrous to sparsely pilose above, pilose below, petiole up to 3(-6) cm long, pilose; inflorescence composed of densely clustered cyathia in axillary or terminal cymes, bracts similar to the leaves but progressively smaller and paler green, sometimes purple spotted; cyathia with 1(-2) peltate, funnel-shaped glands with a circular, 0.5-1.2 mm wide opening, anthers yellow; capsule deeply 3-lobed, 4-5.5 mm × 3.5-4.5 mm, smooth, puberulent; seeds truncate-ovoid, bluntly tuberculate. *E. heterophylla* is a noxious weed of e.g. cocoa, tea, rice and sugar cane, also found in gardens and waste places, and on alluvial soils and sandy beaches, up to 3000 m altitude.

**Selected sources** 97, 202, 217, 372, 638, 864, 865, 1182, 1183, 1186, 1380, 1386, 1425, 1582.

### ***Euphorbia hirta* L.**

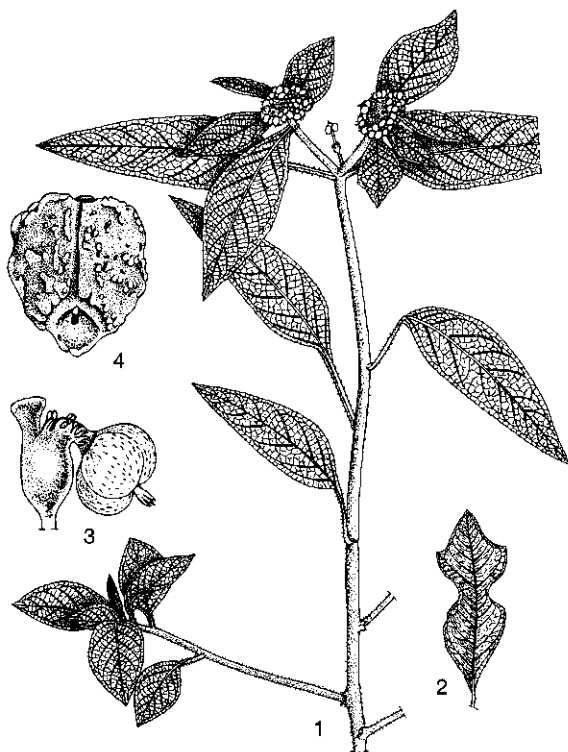
Sp. pl. 1: 454 (1753).

**Synonyms** *Euphorbia pilulifera* L. (1753), *Chamaesyce pilulifera* (L.) Small (1903), *Chamaesyce hirta* (L.) Millsp. (1909).

**Vernacular names** Asthma herb, hairy spurge, pill-bearing spurge (En). Euphorbe à fleurs en tête, euphorbe pilulifère (Fr). Indonesia: daun biji kacang (Malay, Moluccas), nanangkaan (Sundanese), patikan kebo (Javanese). Malaysia: ambin jantan, kelusan, keremak susu (Peninsular). Papua New Guinea: sip (Kurtatchi, Bougainville), kiki kana kuku (Gunantuna, New Britain). Philippines: botobotonis (Tagalog), gatas-gatas (Bisaya, Tagalog), maragatas (Iloko). Laos: mouk may, nom ra sa 'si, ung1 yang. Thailand: nam no raatchasee (central), yaa nam muek, yaa-lang-ueng (northern). Vietnam: c[or] s[uwx]ra, c[or] s[uwx]a l[ows]n l[as].

**Distribution** A pantropical weed of Central American origin; introduced into South-East Asia long ago and nowadays occurring throughout Malesia.

**Uses** The milky latex is cooling and is used as a remedy for conjunctivitis, ulcerated cornea and other eye complaints. It is also applied to cuts, sores and warts. It has a depressant action on the heart and respiration, and a decoction, infusion or tincture of the plant is used to treat asthma, chronic bronchial disorders, acute nasal catarrh, hay fever and emphysema. A decoction is also administered to allay convulsions, as an expectorant, and as a mouthwash to treat toothache. A



*Euphorbia heterophylla* L. - 1, plant habit; 2, leaf showing variability in shape; 3, cyathium; 4, seed.

tincture is considered useful in colic, dysentery, as a vermifuge and in diseases of the genito-urinary tract. The leaves are mixed with those of downy thorn apple (*Datura metel* L.) in preparing 'asthma cigarettes'. The plants are slightly narcotic and are reported as being haemostatic, sedative, and sudorific. They are also applied to stimulate milk secretion and to promote sweating. An infusion of the roots is taken to relieve headache, and heat exhaustion. The ground fruits are given to children as a laxative. In Java and India the tender shoots serve as famine food, raw or steamed, but these may cause intestinal complaints.

**Observations** An annual, unarmed, pilose herb up to 70 cm tall, stems sparingly branched near the base; leaves opposite, ovate to ovate-oblong, 1–5 cm × 0.3–2.5 cm, base obliquely cuneate to rounded, apex bluntly pointed, margin finely serrate, pale to dark green, often with purple spots, short-petioled, stipules free, subulate; inflorescence axillary or terminal, composed of peduncled, globose clusters of cyathia; cyathia with appressed-hairy involucre and 4 minute, green or

purplish glands bearing a narrow appendage, anthers yellow; capsule acutely 3-lobed, broadly ovoid-pyramidal, about 1 mm × 1.2 mm, appressed-hairy; seeds oblong, with slight transverse wrinkles. *E. hirta* is a weed of waste places and in crops, occurring up to 2000 m altitude.

**Selected sources** 97, 287, 380, 393, 465, 531, 581, 671, 716, 814, 865, 1035, 1126, 1128, 1135, 1178, 1181, 1182, 1183, 1184, 1185, 1186, 1380, 1386, 1525, 1527, 1572, 1578.

### ***Euphorbia neriifolia* L.**

Sp. pl. 1: 451 (1753).

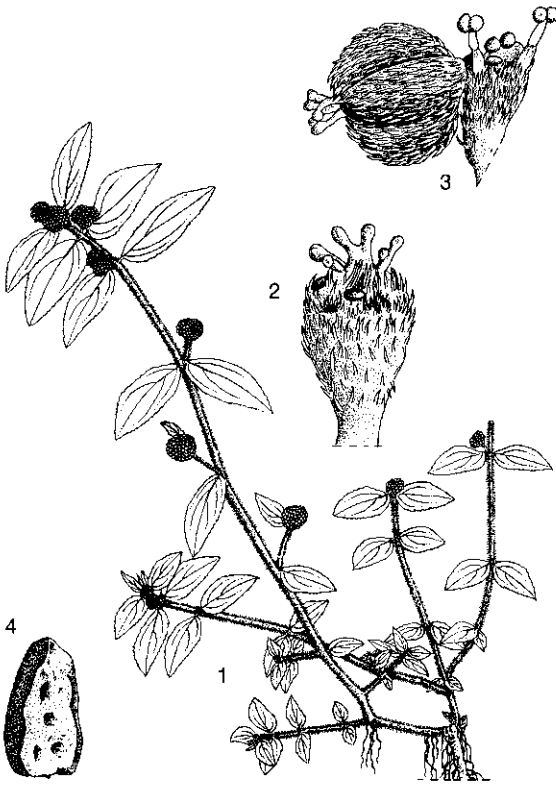
**Synonyms** *Euphorbia ligularia* Roxb. (1832).

**Vernacular names** Common milk hedge, hedge euphorbia, oleander spurge (En). Indonesia: sudu-sudu, susudu, susuru. Malaysia: sesudu, sudu-sudu (Peninsular). Philippines: bait (Pampangan, Tagalog), soro-soro (Tagalog), sorog-sorog (Pampangan, Tagalog). Thailand: som chao (central). Vietnam: x[uw]l[ow]ng r[oo]fng, x[uw]l[ow]ng r[oo]fng r[af]o, x[uw]l[ow]ng r[oo]fng ta.

**Distribution** Probably of South-Asian origin, but nowadays locally cultivated and naturalizing in Sri Lanka, India, Burma (Myanmar), Thailand and throughout the Malesian region except for Borneo; also occasionally cultivated in other tropical regions.

**Uses** In Malaysia and the Philippines, the latex from heated leaves is applied externally to relieve earache. The latex may also be used as a purgative, diuretic, vermifuge, and to treat asthma. In Guatemala, the latex is applied on haemorrhoids. In El Salvador, it is used to relieve sore throat and cracked lips, and also to cure gonorrhoea. In the Moluccas, the bark has been used as a strong purgative. The root is considered antiseptic. In India, the latex is used to remove warts and cutaneous eruptions. The leaves and roots have been used as a fish poison. *E. neriifolia* is fairly commonly planted as a live fence and for ornamental purposes. When boiled with syrup the leaves and slices of the branches can be made into sweetmeats. The nicely figured and aromatic wood is used for small objects such as kris handles. The leaves can be eaten, even raw. In India, the latex is smeared on cuts in *Borassus flabellifer* L. made by tappers, in order to prevent attack by red weevil.

**Observations** An armed, succulent shrub or small tree up to 8 m tall, branches obtusely 5-angular, with pairs of spines of 4–12 mm long arising from the ribs; leaves alternate, obovate to narrowly oblanceolate, (5–)15–30 cm × (1.5–)2–7.5 cm,



*Euphorbia hirta* L. – 1, plant habit; 2, young cyathium; 3, mature cyathium; 4, seed.

base cuneate to attenuate, apex rounded, margin entire, glabrous; inflorescence lateral, composed of 3-7 cyathia on short, rigid, forked peduncles, bracts ovate; cyathia with 5 oblong glands, 1.5-2 mm × 4-5 mm; capsule 10-12 mm in diameter, glabrous. *E. neritifolia* grows well in dry, often rocky places.

**Selected sources** 97, 202, 287, 580, 638, 979, 1126, 1128, 1135, 1178, 1181, 1183, 1184, 1185, 1186, 1311, 1380.

### ***Euphorbia plumerioides* Teijsm. ex Hassk.**

Hort. bogor. descr. 1: 29 (1858).

**Vernacular names** Papua New Guinea: noti (Koheno, North Solomons Province), Simbu (Anji, Enga), temp (Mt Hagen, Western Highlands).

**Distribution** Probably native to the Moluccas and New Guinea, but now cultivated and locally naturalized in Java, the Lesser Sunda Islands, the Philippines (Gola Island), the Moluccas and New Guinea.

**Uses** In Papua New Guinea, the latex is mixed with water and taken to induce vomiting in cases of poisoning or sickness due to sorcery. It is also used as a purgative, vermifuge and fish poison. *E. plumerioides* is often planted as an ornamental on cemeteries and as a hedge.

**Observations** An unarmed shrub up to 2.5 m tall, twigs thick, terete, glabrous; leaves semipersistent, alternate, crowded towards the twig apices, almost sessile, narrowly lanceolate to obovate-lanceolate, 5-14 cm × 1.2-3.5 cm, gradually narrowed at base, apex obtuse with a long mucro, glabrous, pale glaucous below, older ones red; inflorescence terminal, umbelliform, bracts ovate; cyathia with sordidly red involucre and 5 elliptical to orbicular, green or sometimes red-margined glands, anthers yellow; capsule 3-lobed, glabrous. *E. plumerioides* is very variable and has been subdivided into 4 varieties mainly based on leaf characters. It is found on littoral cliffs.

**Selected sources** 97, 597, 768, 1126, 1183, 1185, 1186.

### ***Euphorbia prostrata* Aiton**

Hort. kew. 2: 139 (1789).

**Synonyms** *Chamaesyce prostrata* (Aiton) Small (1903).

**Vernacular names** Trailing red spurge (En). Indonesia: gelang pasir, ki mules (Sundanese), patikan cina (Javanese). Vietnam: c[or] s[uw]x[la n[awf]m.

**Distribution** Native to the West Indies (Ja-

maica), but nowadays introduced and widely naturalized throughout the tropics and subtropics. In South-East Asia reported from Thailand, Java, the Philippines, Sulawesi, the Lesser Sunda Islands, the Moluccas and New Guinea, but probably occurring elsewhere as well.

**Uses** In traditional medicine it is not distinguished from *E. thymifolia*; hence for its uses see under that species.

**Observations** An annual, unarmed, prostrate, often purplish-tinged herb, branches up to 20 cm long, pilose above; leaves opposite, ovate to obovate, up to 0.8 cm × 0.5 cm, base obliquely rounded to truncate, apex rounded, margin serrulate to subentire, puberulous on both surfaces or only below, subsessile, stipules pilose, partly fused; inflorescence terminal and pseudoaxillary, composed of solitary cyathia, bracts absent; cyathia with 4 red glands with minute appendages, bracteoles hair-like; capsule acutely 3-lobed, exerted on a reflexed pedicel, about 1.3 mm × 1.3 mm, hairy; seeds with distinct transverse ridges and grooves. *E. prostrata* is closely related to and sometimes even, incorrectly, regarded as conspecific with *E. thymifolia*. It is found in disturbed places, gardens, fields and roadsides, usually on sandy or gravelly soils, up to 2000 m altitude.

**Selected sources** 40, 51, 97, 217, 580, 921, 1126, 1181, 1183, 1185, 1186, 1334, 1335.

### ***Euphorbia synadenium* Ridley**

Journ. Roy. As. Soc. Straits Br. 61: 36 (1912).

**Synonyms** *Euphorbia ridleyi* Croizat (1937).

**Vernacular names** Malaysia: pokok susu hutan, sesudu bukit, sesudu hutan (Peninsular). Thailand: rak luukmaa (peninsular).

**Distribution** Peninsular Thailand, Peninsular Malaysia; possibly also in India (Bengal).

**Uses** *E. synadenium* is used for poulticing burns.

**Observations** A slightly succulent, unarmed shrub up to 1.5 m tall, stem unbranched; leaves alternate, oblanceolate-spathulate, up to 20 cm long, apex blunt to subacute, petiole about 1.5 cm long; inflorescence composed of axillary, solitary, unisexual cyathia, bracts ovate, purple-pink; male cyathia with 10 lacinate, pink glands, female cyathia with green glands. *E. synadenium* is found locally in rocky, evergreen forest, up to 330 m altitude.

**Selected sources** 120, 202, 1181, 1227, 1380.



**Euphorbia thymifolia L.**

Sp. pl. 1: 454 (1753).

**Synonyms** *Chamaesyce thymifolia* (L.) Millsp. (1916).**Vernacular names** Euphorbe à feuilles de thym, rougette (Fr). Indonesia: gelang pasir, ki mules (Sundanese), patikan cina (Javanese). Malaysia: gelang susu, rumput barah, rumput janjot (Peninsular). Philippines: makikitot (Iloko). Laos: nhayang ung baynoy. Thailand: namnom raat-chasee lek (central). Vietnam: c[or]l s[uwx]a l[as] nh[or], c[or] s[uwx]a d[aas]t, nh[ar] m[uwj]c n[o]j].**Distribution** Throughout the Old World tropics, apparently not widespread in tropical East Africa, but advancing there; throughout the Malayan region.**Uses** *E. thymifolia* has stimulant, astringent, diuretic, anthelmintic and laxative properties, and shows antispasmodic and antifungal activity. It is used as a remedy in earache, eye infections, herpes, gonorrhoea and asthma. In India, *E. thymifolia* is administered to cure ringworm. A poultice is applied to wounds, skin complaints and dislocated bones. A decoction is used in cases of diarrhoea, dysentery, prolapsed rectum, and other abdominal troubles. In the Philippines, a poultice is applied for snake bites, whereas the latex is used to dissipate corneal opacity. In India, whole plants are used as a fish poison, whereas an essential oil can be extracted and applied as an insecticide.**Observations** An annual, unarmed, prostrate herb, branches up to 25 cm long, often reddish, subglabrous to pubescent especially on the upper side; leaves opposite, elliptical or ovate to obovate-oblong, 0.2–0.9 cm × 0.1–0.4 cm, base obliquely rounded to subcordate, apex subacute to rounded or truncate, margin crenate to serrate, glabrous above, sparsely pilose below, subsessile, stipules free, pilose; inflorescence terminal and pseudo-axillary, composed of solitary cyathia, bracts absent; cyathia with 4, minute, red glands bearing minute appendages, bracteoles hair-like; capsule 3-lobed, short-stalked and only barely protruding from the involucre, 1–1.5 mm in diameter, hirsute; seeds with shallow transverse ridges and grooves. *E. thymifolia* is a common weed of cultivated and waste ground, often on sandy or gravelly soils, up to 1650 m altitude.**Selected sources** 97, 202, 217, 287, 363, 531, 580, 1035, 1126, 1128, 1130, 1135, 1178, 1181, 1182, 1183, 1184, 1185, 1186, 1380, 1572.**Euphorbia tirucalli L.**

Sp. pl. 1: 452 (1753).

**Synonyms** *Euphorbia rhipsaloides* Lem. (1857), *Euphorbia media* N.E.Br. (1911), *Euphorbia scoparia* N.E.Br. (1911).**Vernacular names** Finger tree, pencil tree, rubber euphorbia (En). Euphorbe effilé, tirucalli (Fr). Indonesia: patah tulang (general), kayu urip, tikel balung (Javanese). Malaysia: kayu patah tulang, tulang-tulang, tentulang (Peninsular). Philippines: bali-bali (Panay Bisaya), suelda-consuelda (Bicol), suerda (Tagalog). Laos: 'khi<sup>2</sup> hai bai. Thailand: khia cheen, khia thian, phayaa rai bai (northern). Vietnam: x[uw][ow]ng c[as], c[aa]y x[uw][ow]ng kh[oo], san h[oo] xanh.**Distribution** Native to tropical Africa, but widely planted and naturalized throughout the tropics and subtropics. Within Malesia not yet reported from Borneo and New Guinea.**Uses** In traditional medicine, poultices from the stem or bark of *E. tirucalli* are applied to heal broken bones. The latex is used for similar purposes as that of *E. antiquorum* and *E. nerifolia*, but is poisonous, corrosive, and emetic. In Peninsular Malaysia, a poultice of the roots or stems has been applied to ulceration of the nose, haemorrhoids and swellings. Root scrapings, mixed with coconut oil, are given to cure stomach-ache. An extract of the plant shows antibiotic activity. *E. tirucalli* has also been used as a fish poison. It is widely planted as a hedge and for ornamental purposes, especially in dry regions. During the Second World War the latex was tested in South Africa as a rubber substitute, but it proved to be unstable. An oil obtained from the latex appeared useful for application in linoleum, oilskin and leather cloth industries. The white, close-grained and fairly hard wood is used for rafters, toys and veneer. It yields charcoal suitable for use in gunpowder.**Observations** An unarmed, succulent shrub or small tree up to 10(–15) m tall, branches often in whorls, terete, 5–8 mm in diameter, finely longitudinally striate; leaves alternate, early caducous, linear-lanceolate, 0.7–1.6 cm × 0.1–0.3 cm, narrowing at base, apex obtuse to subacute, glabrous throughout or puberulent below, sessile or subsessile, stipules minute, glandular; inflorescence on the stem apices and in bifurcations, generally composed of unisexual cyathia, bracts rounded, small; cyathia with 5 subglobose to transversely elliptical, bright yellow glands; capsule exerted on a tomentose pedicel, subglobose, 7–8 mm in diameter, glabrescent; seeds smooth, buff speckled with brown and with a dark brown ventral line. *E.*

*tirucalli* easily naturalizes in brushwood, open woodland and grassland, up to 2000 m altitude.

**Selected sources** 97, 134, 154, 202, 217, 287, 337, 453, 531, 580, 638, 647, 828, 864, 865, 960, 979, 1035, 1074, 1126, 1128, 1135, 1178, 1181, 1184, 1185, 1186, 1380, 1402, 1499, 1525, 1547, 1610.

Nguyen Nghia Thin & M.S.M. Sosef

### Eurycoma Jack

Mal. Misc. 2: 45 (1822).

SIMAROUBACEAE

x = unknown

**Major species** *Eurycoma apiculata* A.W. Bennett, *E. longifolia* Jack.

**Vernacular names** Brunei: langsia siam, tungkat ali, pasak bumi. Indonesia: pasak bumi (general), bidara laut, beseng (Sumatra). Malaysia: bedara merah, bedara putih, tongkat ali (Peninsular). Cambodia: antong sar, antoung sar. Laos: tho nan. Thailand: hae phan chan (northern), plaalai phuengk (central), phiak (peninsular). Vietnam: c[aa]y b[as] b[eej]nh.

**Origin and geographic distribution** *Eurycoma* is confined to tropical South-East Asia and consists of 3 species, with several subspecies. In the Malasian region 2 species are present: a widespread, rather variable species (*E. longifolia*), occurring from Burma (Myanmar) through Indo-China and Thailand, to Peninsular Malaysia, Sumatra, Borneo and the Philippines; and *E. apiculata*, which is confined to Peninsular Malaysia and Sumatra.

**Uses** Where *E. apiculata* and *E. longifolia* occur sympatrically, the uses are the same for both, and no actual distinction is made. The roots, in particular the bark, are used as a febrifuge, and also as a tonic after childbirth. In Malaysia pounded roots are applied externally as a poultice to treat headache, and also on wounds, ulcers and syphilitic sores. A decoction of the leaves is also sometimes used as a febrifuge. An infusion of the root bark of *E. longifolia* is used to treat stomachache and fever by the Kenyah Dayak in Borneo. In Kalimantan, Banjarese men drink a decoction of the root as an aphrodisiac, in peninsular Thailand it is used as a traditional remedy against malaria. 'Babi kurus' is the vernacular name used in Java for the imported drug.

In Vietnam *E. longifolia* is reputed to cure a variety of diseases ('c[aa]y b[as] b[eej]nh' means the plant for hundred diseases). The bark is pre-

scribed against indigestion, fever and lumbago, the fruits against dysentery. A decoction of the leaves is used as a wash to treat itch. In Cambodia, the root is employed as a vermifuge and as an antidote for intoxication, including drunkenness. A large enough dose of the bitter constituents will provoke vomiting, thereby acting as an antidote for poisoning. Furthermore, roots are used against malaria, as an antipyretic, anti-inflammatory and diaphoretic. The fruits are used against dysentery.

**Production and international trade** No statistics are available on the production and trade of *Eurycoma*. Some interinsular trade seems to exist.

**Properties** Antimalarial activities have been reported from *Eurycoma*. A semi-purified extract of the powdered root (mainly consisting of the quassinoids 13 $\beta$ ,18-dihydroeurycomanol, eurycomanol-2-O- $\beta$ -glucopyranoside, eurycomanol and eurycomanone) was evaluated in vitro using 6 chloroquine-resistant *Plasmodium falciparum* isolates. Results indicated complete inhibitions at 1.25–5  $\mu$ g/ml extract after 3 days post-treatment, and at 0.62  $\mu$ g/ml and 0.31  $\mu$ g/ml after 4 and 6 days post-treatment, respectively. Subsequently, the purified compounds performed well in an in vitro test with 9 *Plasmodium falciparum* isolates from patients infected with chloroquine-resistant malaria. Eurycomanol, eurycomanol-2-O- $\beta$ -glucopyranoside and 13 $\beta$ ,18-dihydroeurycomanol possessed antimalarial activities with IC<sub>50</sub> values of 1.231–4.899  $\mu$ M, 0.389–3.498  $\mu$ M and 0.504–2.343  $\mu$ M, respectively, compared with 0.323–0.774  $\mu$ M for chloroquine. Furthermore, the quassinoids eurycomanone, eurycomalactone, 6-hydroxy-5,6-dehydroeurycomalactone and 7-methoxy- $\beta$ -carboline-1-propionic acid (a  $\beta$ -carboline alkaloid), all isolated from the roots, also demonstrated significant antimalarial activity in vitro.

In general, several constituents isolated from *E. longifolia* possess cytotoxic activity. Quassinoids (eurycomalactone, 6 $\alpha$ -hydroxyeurycomalactone, 5,6-dehydroeurycomalactone, longilactone, 14,15 $\beta$ -dihydroxyklaineanone, 11-dehydroklaineanone) and tirucallane-type triterpenes (niloticin, dihydroniloticin, piscidinol, boujotinolone A, 3-episapelin A, melianone, hispidone) isolated from the wood have shown potent cytotoxicity against P388 and KB cell lines in vitro. Cathin-6-one alkaloids (biosynthetically derived from the amino acid tryptophan) were isolated from the root. 9-Methoxycanthin-6-one, 9-methoxy-canthin-6-one-N-oxide, 9-hydroxy-canthin-6-one and 9-hydroxy-canthin-

6-one-N-oxide were active against a panel of human and murine cancer cell lines *in vitro*: breast, colon, fibrosarcoma, lung, melanoma, KB and P388. Though several other cathin-6-one alkaloids (9,10-dimethoxy-, 10-hydroxy-9-methoxy-, 11-hydroxy-10-methoxy- and 5,9-dihydroxy-cathin-6-one and 9-methoxy-3-methyl-cathin-5,6-dione) are known to be present in the wood and bark, their cytotoxic effects have not been evaluated. Finally, the quassinoid eurycomanone is active against the cell lines mentioned above (except for P388) but, in addition, has significant activity against the KB-V1 (a cell line derived from KB that is resistant to several drugs).

A bioassay study of the trunk root without the root bark of *E. longifolia* led to the isolation of four quassinoids pasakbumin-A, -B, -C and -D. Both pasakbumin A (= eurycomanone) and pasakbumin B exhibited potent anti-ulcer activity on stomach ulcers *in vivo* in the rat, induced by indomethacin as well as by stress.

The effects of *E. longifolia* were studied in male rats. Doses of 200–800 mg/kg body weight twice daily for 10 days prior to the test intensified the orientation activities towards receptive females, supporting the aphrodisiac property of the plant.

**Adulterations and substitutes** *Strychnos* species are used for the same purpose as *Eurycoma*; they are also called 'bidara laut' in Indonesia.

**Description** Small trees or rarely shrubs up to 10 m tall, monoecious or dioecious, branches rather thick with large leaf scars. Leaves alternate, imparipinnate, usually multijugate, crowded at the tip of the branches; leaflets opposite or subopposite, slightly oblique, ovate-lanceolate to obovate-lanceolate, (nearly) sessile, with a conspicuous articulation at base, midrib slightly prominent on the upper surface, prominent beneath, secondary veins straight, ending in an intramarginal looped vein. Inflorescence an axillary panicle, mostly large and lax, puberulous. Flowers bisexual or unisexual, male flower always with a sterile pistil, female flower always with rather large but sterile stamens; calyx small, 5(–6)-lobed, lobes ovate to triangular, acute or blunt, longer than the tube; petals 5(–6), induplicate-valvate in bud, lanceolate or ovate to ovate-oblong; stamens 5(–6), alternating with petals, filaments narrowing to the top, alternating with 5(–6) small, entire, emarginate or cleft staminodes, sometimes a second row of staminodes present, stamens and staminodes sometimes connate with the petals; disk inconspicuous; carpels 5(–6), free, styles attached adaxially near the top and mutually connate or co-

herent, stigma peltate, 5(–6)-lobed, 1 ovule per carpel. Fruit consisting of up to 5 ellipsoid or ovoid drupes on a stalk of about 3 mm, with very thin exocarp, fleshy mesocarp and hard endocarp. Seeds exalbuminous, with 2 planoconvex cotyledons and a short plumule.

**Growth and development** *Eurycoma* can be found flowering and fruiting all year round, sometimes at an early age. Pollination is probably by insects, and the fleshy fruits are probably dispersed by birds.

**Other botanical information** *Eurycoma* is closely related to *Quassia*, another *Simaroubaceae* genus, containing similar bitter principles and also used medicinally in South-East Asia. *Eurycoma* is most often encountered as an unbranched treelet with an umbrella-like rosette of leaves. However, larger specimens with upright branches (see habit drawing) are also found but appear to be increasingly scarce in easily accessible areas.

**Ecology** *Eurycoma* is a common understorey plant occurring from beach forest to lower montane forest. *E. longifolia* rarely occurs up to 1000 m altitude; *E. apiculata* occurs from sea-level to about 1200 m altitude, being more common at higher elevations. *E. longifolia* shows a preference for acidic, leached, well-drained soils. Both *Eurycoma* species can be found in primary and secondary vegetation.

**Propagation and planting** It is assumed that *Eurycoma* can be propagated by seed.

**In vitro production of active compounds** Stem sections of *E. longifolia* cultured in Murashige and Skoog medium gave highest callus production when supplemented with 2.0 mg/l naphthalene acetic acid and 0.1 mg/l benzyladenine. Optimum growth of callus was obtained on medium with pH 6.0, adjusted before autoclaving. Cultures grown at 35°C produced the highest amount of biomass. At this temperature, light intensity of 610 lux was adequate and gave the highest callus yield. Callus growth was best at a photoperiod of 24 hours. The callus produced was yellowish, soft and loose. Successful production of plant biomass is the first necessary step towards the establishment of cell cultures for the *in vitro* production of secondary metabolites.

**Husbandry** In general, *Eurycoma* spp. are collected from the wild or seedlings are transplanted from the forest to the home garden.

**Harvesting** The small *Eurycoma* trees are usually simply uprooted, and the roots are collected and traded. Collecting only bark or leaves involves slightly less destructive harvesting methods.

**Handling after harvest** After harvesting, the roots, bark and leaves of *Eurycoma* are simply sold in the market. In the Kerayan area in East Kalimantan, the roots are sometimes modelled into cups and filled with water. After some time the water has taken up bitter constituents, and is drunk as a tonic.

**Genetic resources and breeding** There are no known germplasm collections of *Eurycoma*. As *Eurycoma* species occur in both primary and secondary forest and are in general common in their areas of distribution, the risk of genetic erosion seems limited. However, as collecting is primarily from the wild and the plants are often destroyed during collection, overexploitation can become a serious risk, especially in easily accessible areas.

**Prospects** The demonstrated antimalarial properties of *Eurycoma* against chloroquine-resistant *Plasmodium falciparum* strains deserves further attention. Given the increasing problem of the resistance to antimalarial agents, the development of alternative medicaments is of great importance to keep malaria under control. The crude extract of *Eurycoma* should be used with caution because of its toxic effect. Further research on the cytotoxic and anti-ulcer effects of isolated compounds seems desirable as well.

**Literature** |1| Ang, H.H., Chan, K.L. & Mak, J.W., 1995. In vitro antimalarial activity of quassinoids from *Eurycoma longifolia* against Malaysian chloroquine-resistant *Plasmodium falciparum* isolates. *Planta Medica* 61(2): 177–178. |2| Burkill, I.H., 1966. A dictionary of the economic products of the Malay Peninsula. 2nd edition. Vol. 1. Ministry of Agriculture and Co-operatives, Kuala Lumpur, Malaysia. pp. 1000–1002. |3| Darise, M., Kohda, H., Mizutani, K. & Tanaka, O., 1982. Eurycomanone and eurycomanol, quassinoids from the roots of *Eurycoma longifolia*. *Phytochemistry* 21(8): 2091–2093. |4| Itokawa, H., Kishi, E., Morita, H. & Takeya, K., 1992. Cytotoxic quassinoids and tirucallane-type triterpenes from the woods of *Eurycoma longifolia*. *Chemical and Pharmaceutical Bulletin* 40(4): 1053–1055. |5| Kardono, L.B.S., Angerhofer, C.K., Tsauri, S., Padmawinata, K., Pezzuto, J.M. & Kinghorn, A.D., 1991. Cytotoxic and antimalarial constituents of the roots of *Eurycoma longifolia*. *Journal of Natural Products (Lloydia)* 54(5): 1360–1367. |6| Mitsunaga, K., Koike, K., Tanaka, T., Ohkawa, Y., Kobayashi, Y., Sawaguchi, T. & Ohmoto, T., 1994. Canthin-6-one alkaloids from *Eurycoma longifolia*. *Phytochemistry* 35(3): 799–802. |7| Nguyen

Van Duong, 1993. Medicinal plants of Vietnam, Cambodia and Laos. Mekong Printing, Santa Ana, California, United States. pp. 393–394. |8| Nootboom, H.P., 1972. Simaroubaceae. In: van Steenis C.G.G.J. (General editor): *Flora Malesiana*. Series 1, Vol. 6. Wolters Noordhoff, Groningen, the Netherlands. pp. 203–206. |9| Singaram, N. & Teo, C.K.H., 1994. Factors affecting the biomass production of tongkat Ali (*Eurycoma longifolia*). *Malaysian Applied Biology* 22(2): 197–203. |10| Tada, H., Yasuda, F., Otani, K., Doteuchi, M., Ishihara, Y. & Shiro, M., 1991. New anti-ulcer quassinoids from *Eurycoma longifolia*. *European Journal of Medicinal Chemistry* 26(3): 345–350.

#### *Selection of species*

#### ***Eurycoma apiculata* A.W. Bennett**

Hook. f., *Fl. Brit. India* 1: 522 (1875).

**Distribution** Peninsular Malaysia and Sumatra.

**Uses** The most common application of *E. apiculata* is as a decoction of the roots as a febrifuge.

**Observations** A small tree up to 5 m tall; leaves up to 40 cm long, leaflets 8–14 cm × 2–4 cm, usually rather abruptly blunt-acuminate; petals linear, rarely lanceolate, 4–9 mm × 1–1.5 mm, puberulous, with glandular hairs outside, glabrous inside, styles very short, stigma 5-lobed; fruit 10–17(–20) mm × 5–12 mm. *E. apiculata* is common in the understorey of primary and secondary forest.

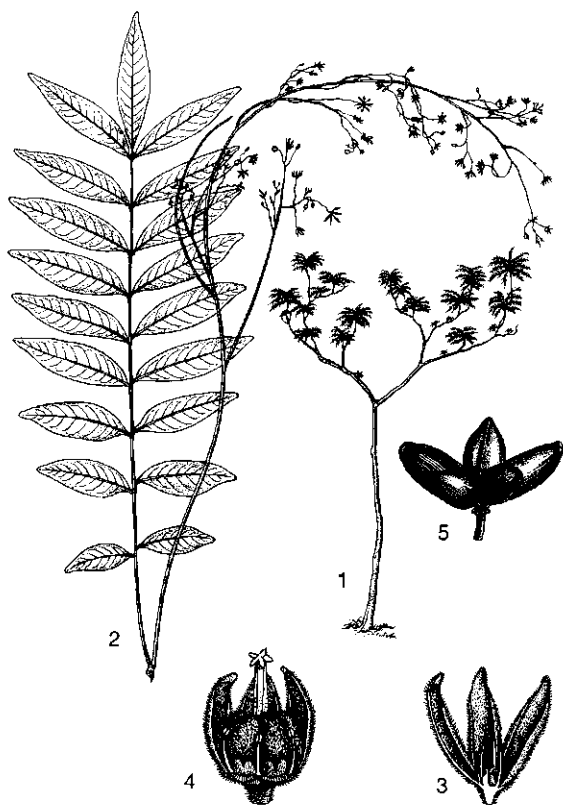
**Selected sources** 202, 1052.

#### ***Eurycoma longifolia* Jack**

Mal. Misc. 2: 45 (1822).

**Distribution** Southern Burma (Myanmar), Indo-China (Cambodia, Laos and Vietnam), Thailand, Peninsular Malaysia, Sumatra, Borneo and the Philippines (subsp. *eglandulosa* (Merr.) Nootboom only).

**Uses** A decoction of the root is a well-known febrifuge. A poultice of the pounded root is used on wounds, ulcers and sores. The bitter constituents of the roots will in a large enough dose provoke vomiting and are employed as such. In Brunei, a decoction of the root is drunk to relieve gastric pains, reduce high blood pressure and fever. The bark is used as a blood coagulant in complications during childbirth. Also in Brunei, the leaves are reportedly eaten raw to relieve stomach-ache. Furthermore, the roots are locally popular as an aphrodisiac.



*Eurycoma longifolia* Jack - 1, tree habit; 2, detached leaf and inflorescence; 3, male flower in section; 4, female flower with 2 petals removed; 5, fruit.

**Observations** A spindly unbranched androdioecious tree or shrub up to 10 m tall, or with a few upright branches, each crowned by an umbrella-like rosette of leaves; leaves up to 100 cm long, leaflets lanceolate to obovate-lanceolate, 5–20 cm × 1.5–6 cm; petals lanceolate to ovate or obovate-oblong, 4.5–5.5 mm × 2–3 mm, puberulous on both surfaces, styles rather long with a peltate 5(–6)-lobed stigma, elevated about 1 mm above the carpels; fruit 10–17(–20) mm × 5–12 mm. *E. longifolia* is common in the understorey of primary and secondary forest on a wide range of soils and is locally abundant.

**Selected sources** 68, 69, 70, 202, 228, 284, 350, 822, 829, 1035, 1052, 1126, 1287, 1383, 1509.

Tahan Uji

## *Fatoua villosa* (Thunb. ex Murray) Nakai

Bot. Mag. Tokyo 41: 516 (1927).

MORACEAE

$n = 13$

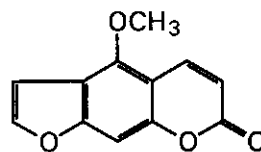
**Synonyms** *Fatoua pilosa* Gaudich. (1826), *Fatoua japonica* (Thunb. ex Murray) Blume (1856).

**Vernacular names** Indonesia: akar kuning (Malay, Timor), ranggitan (Javanese), utu guraci (Ternate). Philippines: sikir (general), malbasdamo (Tagalog), sarungkar-a-babassit (Iloko). Vietnam: d[aa]u b[li]s[ic]h, du[oo]s[li] c[or].

**Origin and geographic distribution** *Fatoua* comprises 2 species, one endemic to Madagascar, the other distributed from eastern Asia to Australia. *F. villosa* occurs from Japan and China to Vietnam, Taiwan, the Philippines, Sulawesi, Java, the Lesser Sunda Islands, the Moluccas, New Guinea, the Solomon Islands, New Caledonia and northern Australia. It has escaped from cultivation and naturalized in the United States, where it is likely to become a weed.

**Uses** In Indonesia, the ground yellow roots of *F. villosa*, known as 'greges otot', used to be smeared on the legs of children with weak legs. In the Philippines, a decoction of the roots is given against fevers and is effective for swollen gums when used as a gargle. An infusion of the roots is prescribed for irregular menstruation and as a diuretic. In Taiwan, the chewed leaf is considered a remedy against stomach-ache. In Indo-China, the crushed and roasted roots are used to prepare a depurative medicine for women after childbirth.

**Properties** The methanol root extract of *F. villosa* has been found to contain phototoxins that possess a UV-A light ( $\lambda$  320–380 nm) activated anti-microbial activity using *Escherichia coli* (ATCC 12407) as test organism. Subsequent high pressure liquid chromatography analysis revealed that the compound responsible was 5-methoxypsoralen (5-MOP or bergapten). 5-Methoxypsoralen belongs to the linear (or 6,7-) furanocoumarins, compounds which are known to have phototoxic activity. The isomeric angular (or 7,8-) furanocoumarins, however, appear to be inactive in



5-methoxypsoralen (bergapten)

this respect. Dermatitis may arise after plants containing linear furanocoumarins come into direct contact with the skin, if this is immediately followed by exposure to UV-A light, e.g. from the sun. The mechanism of photosensitization by linear furanocoumarins is based on interference with DNA base pairs. Energy provided by UV-A irradiation leads to the formation of additional products between the furanocoumarin and cytosine and thymine bases. This bridge-building inhibits the replication and transcription of DNA and, consequently, the synthesis of RNA and proteins and the occurrence of cell division.

Due to this mechanism of DNA synthesis inhibition, psoralens like 5- and 8-methoxypsoralen and the synthetic 4,5',8-trimethylpsoralen are used in therapy for the treatment of psoriasis. Psoriasis is a non-infectious skin disease, characterized by an abnormal production of the outermost layer of the skin, which forms scales and peels off, often in large amounts. Therapy might consist of application of psoralens orally or locally, followed by irradiation with UV-A light.

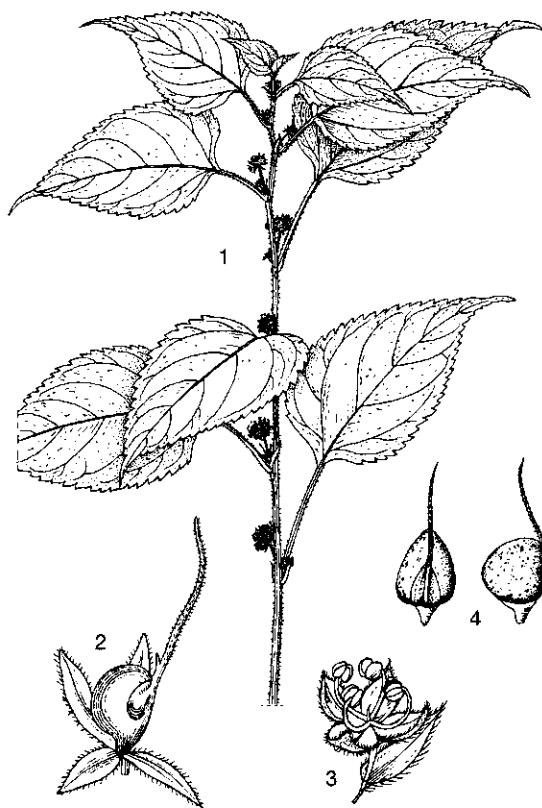
**Description** A monoecious, annual or perennial, ascending or erect, often half-woody herb up to 100 cm tall, without latex; stem with hooked hairs. Leaves alternate, simple, ovate to broadly ovate, 4–11 cm × 2–6 cm, cordate to cuneate at base, acute to acuminate at apex, margin dentate, hirsute, long-petioled; stipules free, lateral. Inflorescence an axillary, peduncled, bisexual, capitate cyme. Flowers unisexual, small, green, with valvate tepals; male flowers with 3–4 tepals, fused up to halfway, stamens 3–4, incurved in bud but exerted when mature, pistillode minute; female flowers sessile with 4 tepals, free or fused at base, ovary superior, 1-locular with a single ovule, style lateral, filiform. Fruit a warty, achene-like drupe, asymmetrically globular to ovoid, enclosed by the enlarged but not fleshy perianth. Seed with endosperm.

**Growth and development** Female flowers of *F. villosa* predominate in inflorescences positioned in the lower and middle parts of the stem, male ones in those of the upper parts.

**Other botanical information** Recently, some authors have recognized *F. villosa* and *F. pilosa* as distinct species, but it is doubtful whether this distinction is valid.

**Ecology** *F. villosa* occurs in dry thickets, grassy places, on walls, stony sites and cliffs at 0–1200 m altitude. It may form a carpet in light secondary forest.

**Genetic resources and breeding** *F. villosa* is



*Fatoua villosa* (Thunb. ex Murray) Nakai – 1, flowering stem; 2, female flower; 3, male flower; 4, fruits with perianth removed.

common in its area of distribution and is found in various anthropogenic habitats. The risk of genetic erosion appears to be limited, in view of its rather weedy nature.

**Prospects** The furanocoumarins present in the roots of *F. villosa* merit further research on its potential as a local or industrial source of psoralens for application in the treatment of psoriasis.

**Literature** [1] Backer, C.A. & Bakhuizen van den Brink Jr., R.C., 1965. Flora of Java. Vol. 2. Wolters-Noordhoff, Groningen, the Netherlands. p. 14. [2] Chew, W.-L., 1989. Moraceae. In: George, A.S. (Editor): Flora of Australia. Vol. 3. Australian Government Publishing Service, Canberra, Australia. pp. 15–68. [3] Heyne, K., 1927. De nuttige planten van Nederlands-Indië [The useful plants of the Dutch East Indies]. 2nd edition. 3 volumes. Departement van Landbouw, Nijverheid en Handel in Nederlandsch-Indië. p. 546. [4] Liao, J.-C., 1996. Moraceae. In: Huang, T.-C. (Editor): Flora of Taiwan. 2nd edition. Vol. 2. Editorial Committee

of the Flora of Taiwan, Taipei, Taiwan, Republic of China. pp. 136–195. |5| Perry, L.M., 1980. Medicinal plants of East and Southeast Asia. Attributed properties and uses. The MIT Press, Cambridge, Massachusetts, United States & London, United Kingdom. p. 271. |6| Quisumbing, E., 1978. Medicinal plants of the Philippines. Katha Publishing Co., Quezon City, the Philippines. pp. 230–231. |7| Rohwer, J.G., 1993. Moraceae. In: Kubitzki, K., Rohwer, J.G. & Bittrich, V. (Editors): The families and genera of vascular plants. Volume II. Springer Verlag, Berlin, Germany. pp. 438–453. |8| Samuelsson, G. (Editor), 1992. Drugs of natural origin, a textbook of pharmacognosy. Swedish Pharmaceutical Press, Stockholm, Sweden. pp. 96–99. |9| Swain, L.A. & Downum, K.R., 1990. Light-activated toxins of the Moraceae. *Biochemical Systematics and Ecology* 18: 153–156. |10| Yamazaki, T., 1982. The seed formation of *Fatoua villosa* (Moraceae). *Journal of Japanese Botany* 57: 358–365.

**Other selected sources** 190, 280, 478, 767, 1128.

M.S.M. Sosef & S.F.A.J. Horsten

## Ficus L.

Sp. pl. 2: 1059 (1753); Gen. pl. ed. 5: 482 (1754).

MORACEAE

$x = 13$ ;  $2n = 26$  for the vast majority of species (e.g. *F. hispida*, *F. religiosa*),  $2n = 52$  for few species

**Major species** *Ficus benghalensis* L., *F. hispida* L.f., *F. religiosa* L., *F. rumphii* Blume, *F. septica* Burm.f.

**Vernacular names** *Ficus*, fig (En). Figue (Fr). Indonesia: ara, bunut, karet. Malaysia: ara, ara kelumpang (stem figs), ara tanah (geocarpic figs), nunok (strangling figs, Dusun, Sarawak), giwit (geocarpic figs, Dusun, Sarawak). Philippines: balete (Filipino). Burma (Myanmar): nyaung. Laos: hai. Thailand: sai. Vietnam: chi da, [leef], sung.

**Origin and geographic distribution** *Ficus* comprises about 1000 species and occurs in all tropical and subtropical regions, with a few species in warm temperate areas. About half of the species occur in Malesia, which forms the main centre of speciation.

**Uses** The latex of many *Ficus* species is used medicinally, mainly to cover and cure wounds, boils and sores, but also as an antirheumatic, and it is swallowed to cure coughs and colds and to

treat diarrhoea. The bark of many species has astringent properties. In India, the dried bark of *F. benghalensis* and *F. religiosa* is used as an antidiabetic.

Several of the medicinally used *Ficus* species yield useful timber, used for e.g. temporary construction, interior work, concrete formwork, small domestic articles, fruit crates, low grade plywood, and firewood. The fruits of some species are edible but are generally not sought after or prized. Some *Ficus* species have poisonous fruits. The latex has been used as a wax in dyeing batik cloth. The latex is also used as birdlime and in Papua New Guinea for sealing leaks in canoes, whereas that of some species is highly toxic and applied as dart poison. The tough and fibrous bark of a few species is a well-known raw material for rough cordage and matting and formerly for clothing; it is still used for bow strings and fish nets. Young leaves of several species are eaten raw in salads or cooked with meat wrapped in them; the latter dish is considered a delicacy in the highlands of New Guinea. They have also been used as fodder, and leaves of other species are applied as sandpaper or to scour cooking pots. Several *Ficus* species are well-known for their ornamental value, planted as wayside trees or even grown commercially as pot plants in temperate regions.

*F. benjamina* L., *F. elastica* Roxb. ex Hornem., *F. minahassae* (Teysm. & de Vriese) Miq., *F. racemosa* L. and *F. retusa* L.f. are also used medicinally but have other primary uses.

**Properties** Bark extracts of various *Ficus* species (*F. benghalensis*, *F. racemosa*, *F. religiosa*, *F. rumphii*) display hypoglycaemic effects, which are probably mainly related to the presence of  $\beta$ -sitosterol and its related compounds. A bark extract of *F. benghalensis* showed in vitro hypoglycaemic activity (the lowering of blood glucose) in both normoglycaemic and moderately diabetic (streptozotocin-induced) rats after oral administration. The extract also enhanced serum insulin levels in both groups of animals. Furthermore, incubating of isolated islets of Langerhans from normal and diabetic animals with the extract stimulated insulin secretion, and reduced the insulinase activity of liver and kidneys. The alcohol extract of *F. racemosa* also showed hypoglycaemic effects in rats.

The stem bark of *F. benghalensis* contains  $\beta$ -sitosterol and its glucoside  $\beta$ -sitosterol-D-glucoside. Both compounds have also been isolated from other species: the former from *F. rumphii* and the latter from the bark of *F. religiosa*. Stigmasterol and

$\beta$ -sitosterol (both sterols) and lupeol (a related triterpene) have been isolated from the petroleum ether extract of trunk bark of *F. religiosa*.  $\beta$ -Sitosterol-D-glucoside produced a hypoglycaemic effect in rabbits, which compared favourably with tolbutamide as a positive control. Although the aglycone  $\beta$ -sitosterol is also active, the glucoside is more potent, probably because of its better water solubility and adsorption.

Also of interest besides the sterols are the flavonoids coumarin and related compounds isolated from *Ficus* species. A dimethoxy derivative of leucocyanidin-3-O- $\beta$ -galactosyl-cellobioside, isolated from the bark of *F. benghalensis* and administered orally (250 mg/kg) decreased blood glucose levels significantly in normal and moderately diabetic (alloxan-induced) rats, and increased serum insulin significantly in the latter group. Furthermore, during one month treatment of the diabetic rats orally (100 mg/kg), a significant decrease in blood and urine sugar was found, as compared with the diabetic controls. A dimethoxy ether of leucopelargonidin-3-O- $\alpha$ -L-rhamnoside (from the bark of *F. benghalensis*) has also been tested for antidiabetic effects. A medium effective dose (100 mg/kg) administered orally to normal and moderately diabetic (alloxan-induced) dogs, significantly reduced blood glucose and raised serum insulin. Acute and chronic administration in single doses of 0.2–1.8 g/kg to mice, and daily administration of 100–500 mg/kg to rats for a period of one month did not produce observable toxic effects. The antidiabetic effects e.g. low blood glucose levels, glucose tolerance and urinary sugars, were also found in normal and in moderately diabetic rats compared to glibenclamide as a positive control. Finally, in vitro studies showed that insulin secretion by pancreatic  $\beta$ -cells was greater in the presence of the leucopelargonidin derivative than in the presence of the leucocyanidin derivative.

An alcohol extract of the bark of *F. religiosa* showed parasympholytic effects and a protective action against acetylcholine and histamine-induced asthma in guinea-pigs. The extract also showed antiprotozoal, protease inhibitor and antiviral activity. Antiprotozoal activity is furthermore reported from the alcohol extract of *F. racemosa*.

A methanolic extract of the leaves of *F. septica* displayed intense antibacterial and antifungal activities. This activity is probably related to the presence of 2 indolizidine alkaloids, ficuseptine (4,6-bis-(4-methoxyphenyl)-1,2,3-trihydroindolizinium chloride) and antofine. A leaf extract of

*F. religiosa* showed antifungal properties against *Diplodia natalensis*, the agent of stem-end rot of mango fruits. Fruit extracts of *F. benjamina*, *F. benghalensis* and *F. religiosa* also had significant antibacterial activity, but no antifungal activity. Furthermore, the extracts of *F. benghalensis* and *F. religiosa* demonstrated activity in the brine shrimp assay (*Artemia salina*) which indicates toxicity, whereas *F. benjamina* showed no activity. All the fruit extracts exhibited antitumour activity in the potato disk bioassay, and none of the tested extracts showed any marked inhibition on the uptake of calcium into rat pituitary cells (GH-4C-1). A water extract of dried fruits of *F. benghalensis* exhibited anti-HIV activity.

The anthelmintic properties of several *Ficus* species (e.g. *F. pumila*) can be ascribed to the proteolytic enzyme ficin present in the latex. Excessive amounts of this substance are toxic to humans when administered orally or intravenously. Furthermore, ficin, whether fresh or dry, is highly irritant to the skin and eyes.

Seed extracts of *F. deltoidea* have been observed to agglutinate human erythrocytes (A, B, AB, O), and some strains of bacteria including *Chlamydia trachomatis*, a significant pathogen. Seed extracts of *F. racemosa* have been found to agglutinate white blood cells from patients with different types of leukaemia.

The popular pot plant *F. benjamina* may give rise to allergic reactions, e.g. conjunctivitis, rhinitis. Sensitization is believed to occur by inhalation of allergen-enriched dust emanating from the leaves. Phytochemical investigations of a petroleum ether extract from dried *F. hispida* bark yielded the acetates of n-triacontanol,  $\beta$ -amyrin and gluanol, whereas the leaves contain bergapten, psoralen (two furanocoumarins, which might give rise to phototoxicity),  $\beta$ -amyrin and  $\beta$ -sitosterol. The leaves of *F. pachyrrachis* contain 2 tetrahydrobenzylisoquinoline alkaloids, (-)-reticuline and (+)-norreticuline.

**Description** Evergreen or sometimes deciduous, woody epiphytic climbers or stranglers, creepers, shrubs or small to large trees up to 40(–50) m tall, or banyans, i.e. trees whose branches send down aerial roots that thicken ('pillar roots') and function as props; bole fairly straight in tree-like species, sometimes fluted, up to 100(–190) cm in diameter, sometimes heavily buttressed; bark surface smooth, often pale grey, sometimes whitish or brown, sometimes lenticellate, inner bark yellowish, exuding white or yellow latex. Leaves arranged spirally, alternate or opposite, simple to



palmately lobed, symmetrical to asymmetrical, dentate to entire, often with glands below in the axil of the lateral or basal veins or abaxial at the apex of the petiole; stipules free or connate. Inflorescence axillary or ramiflorous to cauliflorous, sometimes subterranean, solitary or clustered, monoecious or gynodioecious, with the flowers set inside an urn-shaped receptacle (syconium; a fig). Flowers unisexual; tepals 2–8, free or joined; stamens 1–7; ovary unilocular with a single ovule, style single. Inflorescence a subfleshy fig; individual fruit a drupelet. Seedling with epigeal germination; cotyledons emergent; hypocotyl elongated; all leaves arranged spirally.

**Growth and development** The strangling figs start as epiphytic plants and send down aerial roots that eventually form a false trunk composed of a trellis-work of interlacing and anastomosing roots around the trunk of the support tree. The roots of *F. religiosa*, however, penetrate inside the support trunk, eventually splitting it from within. Many species have more than one kind of leaf (heterophylly).

The symbiotic relation of figs with specialized wasps is well-known. Figs can only be pollinated by female agaonid wasps (*Hymenoptera*, *Chalcididae*, *Agaonidae*). The wasp species are highly species-specific. Fig species are divided into 2 groups: monoecious species and gynodioecious ones. In the former the wasps arrive when only female flowers are receptive. They enter the fig via the ostiole, a bract-covered apical pore. Once inside, they pollinate the female flowers and deposit their eggs in the ovaries. As style length varies greatly within these figs and because the wasp can only reach the ovary of short-styled flowers, only some of the flowers obtain an egg, while in others the seed develops. Male and female wasps emerge after a few weeks, and mating takes place within the fig. The females then emerge from the fig and, in so doing, pick up pollen from the newly mature anthers. On a single tree figs mature at the same time, while different trees of the same species flower out of synchrony, thus inducing cross-pollination. The gynodioecious fig species either bear hermaphrodite figs or figs with female flowers only. In hermaphrodite figs the styles are uniformly short. Wasps can oviposit every female flower and such trees rear the pollinators' offspring and function as pollen donors. All the flowers of female-flowered figs have long styles. The wasps can only deposit pollen, so such figs produce large amounts of seed.

During daytime the figs are eaten by birds (e.g. pi-

geons, hornbills, bulbuls, cassowaries), monkeys and squirrels. At night they are visited by bats and civet cats. Cauliflorous figs are eaten by deer and pigs. The latter also uproot the geocarpic figs. Even elephants, rhinoceros, tapir and wild cattle have been reported to feed on figs. As fig fruits are often available year-round, they constitute an extremely important forest food, a so-called keystone resource that sustains frugivorous animals at the famine period of the year when few species, if any, are fruiting.

**Other botanical information** In Asia and Australia the large genus *Ficus* has been subdivided into 4 subgenera, 14 sections and numerous smaller taxonomic groups.

**Ecology** *Ficus* species are common and form an important element of lowland rain forest, both as canopy and understorey trees. Most species prefer per-humid forest, but several are found in areas with a monsoon climate and in teak forest, including locations where the soil dries out. *Ficus* does not occur in mangrove vegetation but is often present in brackish swamps behind the mangrove. *Ficus* species are generally found below 1500 m altitude, some between 1500 and 2750 m or rarely up to 3200 m. Many are epiphytic and/or strangling.

**Propagation and planting** *Ficus* can be propagated from seed and vegetatively. Per kg there are about 2.1–2.5 million seeds of *F. benghalensis*. The drupelets are usually the unit of sowing. These cannot be stored without a serious decrease in viability. A 50% germination rate is achieved in 27–37 days in *F. deltoidea* and in 18–34 days in *F. microcarpa*. In India, pretreatment with hot water of 60°C for 10 minutes was found to increase the germination rate from 20% to 24% in *F. benghalensis* and from 19% to 28% in *F. racemosa*. The tiny seedlings are pricked out twice, first in clumps and later individually. The young seedlings are sensitive to excess of water. Both large and small cuttings are used in vegetative propagation, but small cuttings are less successful. Tissue culture and air layering can also be used for propagation of *Ficus*. Some species, e.g. *F. hispida*, easily regenerate naturally in abandoned fields.

**Yield** Information on yield of *Ficus* is rather scarce. Petrol and methanol extraction yielded a residue of 85 g from 1.7 kg powdered leaves of *F. septica*. Complete isolation yielded 500 mg of ficuseptine and 130 mg of antofine.

**Handling after harvest** The bark, roots or leaves of *Ficus* can be applied fresh or dried. The fresh products are used or sold shortly after har-

vesting. When dried in shade or sunlight the products can be stored for longer periods.

**Genetic resources and breeding** With the exception of some species that are widely planted (these include *F. benghalensis* and *F. benjamina*) there are no records of ex situ conservation of *Ficus*. Some of them have a high ritual or ornamental value and have been translocated within South-East Asia or even on a worldwide scale. As most *Ficus* species are fairly common and widespread, the risk of genetic erosion seems comparatively low. Breeding efforts are restricted to species with ornamental value, of commercial importance.

**Prospects** Some *Ficus* species are well-known medicinal plants which have been successfully used in the treatment of common illnesses. The promotion of traditionally used plant resources that are widely available or can easily be grown deserves attention from government extension workers, especially in rural areas. Ficin as found in the latex of *F. pumila* can be used for its anti-inflammatory properties. Its proteolytic activity makes it a potential meat tenderizer in industrial applications.

**Literature** |1| Achrekar, S., Kaklij, G.S., Pote, M.S. & Kelkar, S.M., 1991. Hypoglycemic activity of *Eugenia jambolana* and *Ficus benghalensis*: mechanism of action. In-Vivo 5(2): 143-147. |2| Agrawal, S. & Agarwal, S.S., 1990. Preliminary observations on leukaemia specific agglutinins from seeds. Indian Journal of Medicinal Research 92: 38-42. |3| Baumgartner, B., Erdelmeier, C.A.J., Wright, A.D., Rali, T. & Sticher, O., 1990. An antimicrobial alkaloid from *Ficus septica*. Phytochemistry 29(10): 3327-3330. |4| Berg, C.C., 1989. Classification and distribution of *Ficus*. Experientia 45: 605-611. |5| Boer, E. & Sosef, M.S.M., 1998. *Ficus* L. In: Sosef, M.S.M., Hong, L.T. & Prawirohatmodjo, S. (Editors): Plant Resources of South-East Asia No 5(3). Timber trees: Lesser-known timbers. Backhuys Publishers, Leiden, the Netherlands. pp. 232-238. |6| Bronstein, J.L. & McKey, D., 1989. The fig/pollinator mutualism: a model system for comparative biology. Experientia 45: 601-604. |7| Cherian, S., Kumar, R.V., Augusti, K.T. & Kidwai, J.R., 1992. Antidiabetic effect of a glycoside of pelargonidin isolated from the bark of *Ficus benghalensis* L. Indian Journal of Biochemistry and Biophysics 29: 380-382. |8| Hunter, J.B., Suresh, M.R., Keshvarz, E., Wenman, W.M. & Micetich, R.G., 1986. Purification of lectins from *Artocarpus altilis* and *Ficus deltoidea* by gel filtration fast protein liquid chromatography. Biochemi-

cal Archives 2(2): 99-106. |9| Mousa, O., Vuorela, P., Kiviranta, J., Wahab, S.A., Hiltunen, R. & Vuorela, H., 1994. Bioactivity of certain Egyptian *Ficus* species. Journal of Ethnopharmacology 41(1-2): 71-76. |10| Verheij, E.W.M. & Coronel, R.E. (Editors), 1991. Plant Resources of South-East Asia No 2. Edible fruits and nuts. Pudoc, Wageningen, the Netherlands. pp. 334-336.

#### *Selection of species*

#### ***Ficus adenosperma* Miq.**

Ann. Mus. Bot. Lugd.-Bat. 3: 233 (1867).

**Synonyms** *Ficus pauper* King (1888), *Ficus turbinata* Ridley (1916).

**Vernacular names** Indonesia: fangkis (Mabrat, Irian Jaya), nusu, tintinalino (Sulawesi). Papua New Guinea: simpia (Kabiufo, Eastern Highlands).

**Distribution** Sulawesi, the Moluccas, New Guinea, east to the Solomon Islands and Vanuatu, south to northern Australia.

**Uses** In Papua New Guinea, the latex from the leaves is applied to sores and scabies; fresh roots are chewed to treat malaria. The timber, which is of poor quality, has been used for house building.

**Observations** A small to medium-sized tree up to 20 m tall; leaves alternate, narrowly ovate to ovate or elliptical, 5-18 cm × 3-7.5 cm, base cuneate to subcordate, apex short acuminate to attenuate, margin entire, appressed hairy below when young, with 8-10 pairs of lateral veins, stipules 1-3 cm long; figs globose to pyriform, 1-1.8 cm in diameter, smooth to pustular, on a peduncle up to 1 cm long; male flowers in 1 row, with 4-5 tepals and 1 stamen, female flowers sessile, with 3-4 tepals. *F. adenosperma* is found in primary and secondary forest, up to 2500 m altitude, chiefly near rivers, often developing thickets on sandbanks and islands.

**Selected sources** 167, 248, 281, 597, 607, 1104.

#### ***Ficus ampelas* Burm.f.**

Fl. ind.: 226 (1768).

**Synonyms** *Ficus soronensis* King (1887), *Ficus kingiana* Hemsley (1897), *Ficus blepharosepala* Warb. (1905).

**Vernacular names** Indonesia: hampelas (Sundanese), rempelas (Javanese), pila (Moluccas). Philippines: upling-gubat (Tagalog).

**Distribution** From the Ryukyu Islands and Taiwan to the Philippines, Sumatra, Java, the Lesser Sunda Islands, Sulawesi, the Moluccas and

New Guinea including New Britain; possibly also in Borneo.

**Uses** The latex is taken internally to treat diarrhoea. In Indonesia it is used as a diuretic. In New Ireland, the latex of young leaves, mixed with water, is given to babies with mouth sores. The fruits are eaten raw or cooked. The leaves have been used for sandpapering.

**Observations** An evergreen, small to medium-sized tree up to 15(-20) m tall, bark smooth, greyish to brown; leaves distichous, narrowly elliptical, 4-15 cm × 1.5-5 cm, base cuneate, apex acute or acuminate, margin entire, with 3-6 pairs of lateral veins, scabrid, stipules up to 8 mm long; figs axillary and clustered on twigs, stipitate, subglobose, 7-10 mm in diameter, scabridulous, ripening to red or purple; flowers with 3-5 tepals, male flowers sessile, with 1 stamen, female flowers sessile or with a short stipe. *F. ampelas* is common in primary and secondary lowland forest.

**Selected sources** 202, 281, 580, 606, 856, 1126, 1274, 1384.

### **Ficus aurantiacea Griffith**

Not. pl. asiat. 4: 394 (1854).

**Synonyms** *Ficus callicarpa* Miq. (1867), *Ficus pomifera* Kurz (1873), *Ficus megacarpa* Merr. (1904).

**Vernacular names** Indonesia: oyod santenan (Javanese). Malaysia: akar jalar-jalar, akar palapala, tengkok biawak hitam (Peninsular). Vietnam: sung cam.

**Distribution** From Burma (Myanmar), the Nicobar Islands, Indo-China and Taiwan to Thailand and throughout the Malesian region except for New Guinea.

**Uses** In Sabah, leaf shoots are heated slowly over a fire and applied as a poultice on skin itches. In Peninsular Malaysia, the leaves are used to treat fever and toothache. The fruits are sometimes reported to be poisonous.

**Observations** A climber; leaves elliptical to obovate, 4.5-12 cm × 3-7.5 cm, base cuneate, apex rounded to slightly pointed, margin entire, with 3-6 pairs of veins and prominent reticulations below, stipules 3-13 mm long; figs cauliflorous, pear-shaped, 50 mm × 30 mm, glabrous or sparsely puberulous, ripening red to black with white spots; male flowers gamophyllous, female flowers with 3-4 tepals. *F. aurantiacea* is found in lowland to montane forest, up to 1700 m altitude, also in villages, coconut plantations and other anthropogenic habitats.

**Selected sources** 27, 202, 281, 284, 856, 1289.

### **Ficus bauerleni King**

Journ. Asiat. Soc. Bengal II, 55: 408 (1887).

**Synonyms** *Ficus mespiloides* King (1888), *Ficus hollrungii* Lauterb. & K. Schumann (1901), *Ficus laurentina* Diels (1935).

**Distribution** New Guinea, New Britain and the Solomon Islands.

**Uses** In Papua New Guinea, the latex of the inner bark is drunk to treat diarrhoea or dysentery.

**Observations** A large climber, stem red when cut, bark thick, corky; leaves elliptical, 10-25 cm × 4.5-10.5 cm, base subcordate to broadly cuneate, apex acuminate, margin entire, with 5-9 pairs of lateral veins, hairy on the veins below, stipules up to 12 mm long; figs axillary, solitary or paired, depressed globose, 23-30 mm in diameter, lanuginose, ripening rose-red. *F. bauerleni* occurs in lowland forest.

**Selected sources** 281, 609.

### **Ficus benghalensis L.**

Sp. pl. 2: 1059 (1753).

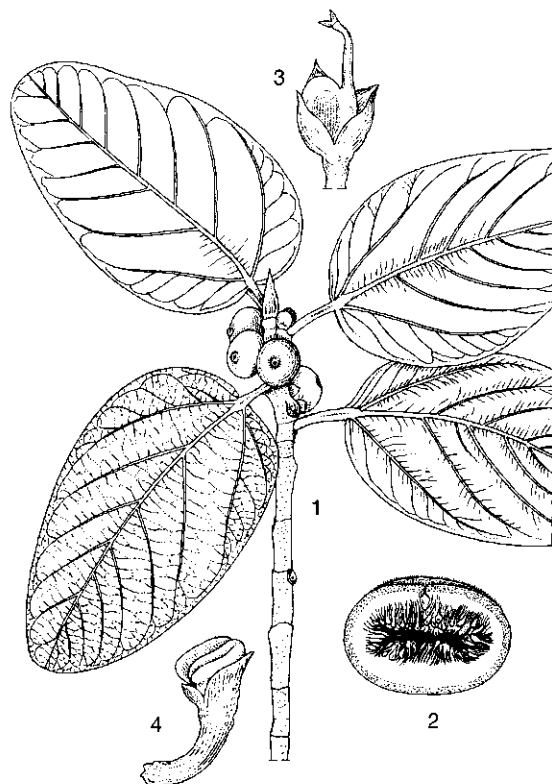
**Synonyms** *Ficus indica* L. (1753), *Ficus lasiophylla* Link (1822), *Ficus banyana* Oken (1841).

**Vernacular names** Banyan tree, Indian banyan (En). Indonesia: beringin India. Malaysia: banyan (general), ara tandok, bohdi (Peninsular). Burma (Myanmar): pyi-nyaung. Thailand: krang, ni khrot (central). Vietnam: da l[as] tr[of]n.

**Distribution** Originally from India and Pakistan but widely planted in Indo-China, Thailand and in the Malesian region and locally naturalized.

**Uses** The leaves are used to remedy dysentery and diarrhoea, and are applied to abscesses as a poultice to promote suppurations and discharge of pus. In a decoction with toasted rice, the leaves are used as a diaphoretic. The bark is tonic and diuretic, an infusion is antidiabetic and a decoction is used as an astringent in leucorrhoea. A decoction of root fibres is useful against gonorrhoea, whereas the tender ends of aerial roots are used for obstinate vomiting. An infusion of the twigs is good for haemoptysis. The milky latex is used against pains and fever, rheumatism and lumbago, toothache, and applied to cracked and inflamed soles. The concentrated latex plus fruit is aphrodisiac and used to treat spermatorrhoea and gonorrhoea. The fruit is tonic and has a cooling effect.

**Observations** A deciduous to evergreen, wide-spreading banyan up to 20(-25) m tall, with copious aerial roots, bark surface smooth, grey; leaves arranged spirally, ovate or broadly ovate to ellipti-



*Ficus benghalensis* L. - 1, fruiting twig; 2, halved fig; 3, female flower; 4, male flower.

cal, 10–30 cm × 7–20 cm, base cordate, apex blunt to rounded, margin entire, with 5–7 pairs of lateral veins, puberulous below, stipules 1.5–2.5 cm long; figs paired, sessile, globose to depressed globose, 15–25 mm in diameter, puberulous, orange to red or pinkish-red when ripe; male flowers many, shortly stipitate, with 2–3 tepals and 1 stamen, female flowers sessile, with 3–4 tepals. *F. benghalensis* occurs in evergreen to deciduous lowland forest.

**Selected sources** 9, 167, 248, 281, 284, 478, 795, 874, 921, 1115, 1178, 1191, 1289, 1404.

#### ***Ficus botryocarpa* Miq.**

Ann. Mus. Bot. Lugd.-Bat. 3: 233 (1867).

**Synonyms** *Ficus barnesii* Merr. (1904), *Ficus mindorensis* Merr. (1904), *Ficus linearifolia* Elmer (1907).

**Vernacular names** Indonesia: bali susuk, delah, tarera intalun (Sulawesi). Papua New Guinea: simbahu (Sui, Northern Province). Philippines: basikong (Manobo, Bagobo), daing-daing (Tagalog).

**Distribution** The Philippines, Sulawesi, the Moluccas, New Guinea and New Britain.

**Uses** In Papua New Guinea, the fruit latex is placed on a boil to effect healing. The leaves and fruits are edible.

**Observations** A small tree up to 14 m tall; leaves distichous or opposite, elliptical, 9–26 cm × 3–11 cm, base cuneate, apex shortly acuminate, margin entire, with 6–10 pairs of lateral veins, sparsely hairy below, stipules 1–2.5 cm long; figs cauliflorous, paired, subglobose, 15–27 mm in diameter, glabrescent, ripening yellowish-white; male flowers with 1 stamen, female flowers sessile or shortly stipitate. *F. botryocarpa* is found in primary and secondary lowland to montane forest, up to 1700 m altitude, often along rivers.

**Selected sources** 281, 597, 604, 1104.

#### ***Ficus calopilina* Diels**

Bot. Jahrb. Syst. 67: 212 (1935).

**Synonyms** *Ficus setistyla* Warb. (1905).

**Distribution** New Guinea.

**Uses** In Papua New Guinea, the fruit latex is used to cover sores; the sores are subsequently covered by a leaf of the same plant. The figs are edible but tasteless. The bark is used for twine.

**Observations** A small tree up to 15 m tall; leaves spirally arranged or decussate, elliptical to slightly obovate, 10–36 cm × 5.5–21 cm, base subcordate to subcuneate, apex acuminate, margin entire, with 6–9 pairs of lateral veins, hispid, stipules up to 30 mm long; figs axillary and cauliflorous, subglobose to pyriform, 30–60 mm in diameter, densely hairy but glabrescent, ripening yellow to brown; male flowers in 2–3 rings, with 1(–2) stamens, female flowers sessile or stipitate. *F. calopilina* is locally common in primary or secondary montane forest, at 1000–2400 m altitude, often along streams.

**Selected sources** 281, 610.

#### ***Ficus copiosa* Steud.**

Nomencl. bot., ed. 2, 1: 635 (1840).

**Synonyms** *Ficus magnifolia* F. v. Mueller (1863), *Ficus krausseana* Rechinger (1912), *Ficus longipedunculata* Rechinger (1912).

**Vernacular names** Indonesia: ampelas (Sulawesi), gohi (Halmahera), sosa kecil (Ternate). Papua New Guinea: kagua (Raluana, New Britain).

**Distribution** Sulawesi, the Moluccas, New Guinea, the Solomon Islands, northern Australia, Vanuatu, the Palau Islands and Yap.

**Uses** In Papua New Guinea, the unripe fruits

are chewed to relieve stomach-ache, and the fruit latex is applied to boils. Fresh leaves are used in the Trobriand Islands as a poison antidote; the roots and leaves are used to treat stomach-ache. In New Britain, massaging the stomach with crushed leaves is said to relieve stomach-ache. Young leaves and figs are eaten raw or cooked. The bark is used for clothing and ropes.

**Observations** A small to medium-sized tree up to 20 m tall, bole muriculate, bark surface brown; leaves alternate to decussate, ovate to obovate or obpentagonal, 10–35 cm × 5–18 cm, base subcordate to subcuneate, apex acute to acuminate, margin entire, with 6–10 pairs of lateral veins, hairy, stipules 1–2.5 cm long; figs axillary to cauliflorous, slightly depressed-globose, 25–60 mm in diameter; flowers with 4–7 tepals, male flowers with 1–2 stamens, female flowers long-stipitate. *F. copiosa* is found in both coastal and inland regions, in primary and secondary forest, up to 1700 m altitude. It is also grown in villages.

**Selected sources** 248, 281, 576, 603, 605, 610, 1104.

### **Ficus dammaropsis Diels**

Flora 128: 28, t. 2A, 2B (1933).

**Synonyms** *Dammaropsis kingiana* Warb. (1891).

**Distribution** New Guinea.

**Uses** In New Guinea, a daily drink of stem latex is said to relieve a severe cough. The figs are edible. Young leaves are used for wrapping meat and eaten as a vegetable. The bark fibre is used to make rope.

**Observations** A small tree up to 10 m tall; leaves arranged spirally, broadly elliptical, to 90 cm × 60 cm, base deeply cordate, apex shortly apiculate, margin sinuate, with 8–10 pairs of lateral veins, puberulous below, stipules 15–30 cm long; figs axillary, solitary, sessile, up to 60 mm in diameter, covered with large bracts; male flowers in many rows, with 2 stamens, female flowers sessile. *F. dammaropsis* occurs in montane forest, at 1000–2300 m altitude, on riverbanks and in clearings.

**Selected sources** 281, 607, 900, 1104.

### **Ficus deltoidea Jack**

Malayan Misc. 2: 71 (1822).

**Synonyms** *Ficus diversifolia* Blume (1825), *Ficus lutescens* Desf. (1829), *Ficus motleyana* Miq. (1867).

**Vernacular names** Rusty-leaved bush fig, mistletoe fig (En). Indonesia: tabat barito (gener-

al). Malaysia: ara burong, ara jelateh, ara tanah (Peninsular).

**Distribution** Thailand, Peninsular Malaysia, Sumatra, Java, Borneo, the Philippines (Palawan) and Sulawesi; introduced in Indo-China, India and Pakistan.

**Uses** In South Kalimantan, *F. deltoidea* is reported to be an effective remedy against leucorrhoea. As it promotes contraction of the vagina it is also considered an aphrodisiac. The latex may have been applied as fish poison. In Thailand, *F. deltoidea* is used as an ornamental. Various selections are commercially grown as pot plant in temperate regions.

**Observations** An evergreen epiphyte or small shrub up to 2 m tall; leaves obovate to elliptical or obdeltooid, 2.5–8 cm × 1.3–7.5 cm, base broadly cuneate, apex blunt to truncate or widely notched, rarely pointed, margin entire, white-spotted above, rusty or yellow-olive below, the midrib sometimes forked, stipules 8–12 mm long; figs axillary, solitary or paired, globose to oblong, 5–10 mm in diameter, ripening to orange or red; male flowers dispersed, with 2–3 free tepals and 2 stamens, female flowers sessile to subsessile, with 3–4 tepals. *F. deltoidea* is a variable species divided into many varieties. It is common in lowlands and mountains, up to 3200 m altitude, generally occurring as an epiphyte, but as a terrestrial bush on sandy shores and mountain tops and bogs.

**Selected sources** 202, 281, 282, 284, 478, 1230, 1289.

### **Ficus hispida L.f.**

Suppl. pl.: 442 (1781).

**Synonyms** *Ficus letaqui* Lév. & Van. (1910), *Ficus poilanei* Gagnep. (1927).

**Vernacular names** Rough-leaved stem fig (En). Indonesia: bisoro (Sundanese), luwing (Javanese), mongmong (Sumatra). Malaysia: ara bumbing, ara sinigai, ara seniah (Peninsular). Laos: dua<sup>1</sup> pong<sup>1</sup>. Thailand: duea plong (northern, central, peninsular), duea pong (Bangkok), madaea plong (central). Vietnam: ng[as]i.

**Distribution** From Sri Lanka and India to Indo-China, southern China, Thailand, the Andaman Islands, Peninsular Malaysia, Sumatra, Java, Borneo, Sulawesi, the Lesser Sunda Islands, New Guinea and northern Australia (Queensland).

**Uses** The immature fruits are considered tonic, galactagogue and emetic. The latex of the leaves is taken internally to treat fever, diarrhoea and to relieve painful urination; the latex of the bark is

regarded as an emetic. An extract of the bark is used in the treatment of jaundice, leprosy and anaemia. Boiled leaves are used to poultice boils and ulceration of the nose. The fruits are also eaten in curries, but are, however, likely to cause giddiness. Ripe fruits are made into a jam. The bark yields a rough fibre. The leaves may be used as fodder for cattle. Large cuttings have been used to establish live fences.

**Observations** An evergreen, small tree up to 15 m tall, bark smooth, grey; leaves often decussate, asymmetrical, pentagonal to oblong, 10–35 cm × 4–20 cm, base subcordate to broadly cuneate, apex acuminate, margin crenulate, with 5–10 pairs of veins and prominent reticulation below, hispid, stipules 1–2.5 cm long; figs on long twigs hanging from the trunk and main branches, obovoid, 25–40 mm in diameter, densely brown pubescent, pale or greenish-yellow when ripe; male flowers in 1–2 rows, with 1 stamen, female flowers sessile or stipitate. *F. hispida* is common in secondary lowland forest in per-humid to monsoon climates.

**Selected sources** 7, 202, 248, 281, 284, 287, 580, 766, 921, 1380, 1478, 1525, 1564.

### ***Ficus microcarpa* L.f.**

Suppl. pl.: 442 (1781).

**Synonyms** *Ficus cairnsii* Warb. (1905), *Ficus retusifomis* Lév. (1910), *Ficus retusa* auct. non L.f.

**Vernacular names** Chinese banyan, laurel fig, Malayan banyan (En). Indonesia: preh (Javanese). Malaysia: jawi jawi, jejawi (Peninsular). Papua New Guinea: magi (Barakau, Central Province). Philippines: baleteng-liitan (Filipino). Thailand: sai khao (Nakhon Si Thammarat), sai rayong (Nakhon Ratchasima), sai yoi bai thu (central). Vietnam: g[uw]f[a].

**Distribution** From Sri Lanka and India to Indo-China, southern China, the Ryukyu Islands, Thailand, and throughout Malesia towards the Solomon Islands, Australia, the Caroline and Marianas Islands, New Caledonia, the Loyalty Islands and Palau.

**Uses** The root, bark and leaf latex are used medicinally to treat wounds, headache and toothache. The bark and leaf latex is taken internally to treat colic and liver trouble. In the Admiralty Islands, patients with fever or headache perspire in the steam of boiling young leaves. The trees are often planted for shade and in cemeteries.

**Observations** An evergreen, small or medium-sized banyan up to 25 m tall, developing numerous slender aerial roots from the branches, bark

surface grey; leaves usually alternate, often asymmetrical, elliptical-obovate to elliptical-ovate, 3–12 cm × 1.5–9 cm, base cuneate, apex blunt or slightly pointed, margin entire, with 5–9 pairs of veins, usually glabrous, stipules 1–1.5 cm long; figs monoecious, axillary, paired, sessile, pyriform to subglobose, 8–12 mm in diameter, glabrous, ripening purple to black; male and female flowers sessile, with 3(–4) tepals, male flowers with 1 stamen. *F. microcarpa* grows in widely varying locations, from rocky sea coasts to limestone hills, and from swampy ground near the sea to montane forest.

**Selected sources** 167, 248, 281, 284, 478, 608, 856, 921, 1274, 1289, 1380.

### ***Ficus nasuta* Summerh.**

Hooker's Icon. Pl.: t. 3189 (1933).

**Distribution** New Guinea and the Solomon Islands.

**Uses** In Papua New Guinea, the bark latex is drunk by patients with asthma or other respiratory problems. Leaves eaten with salt are believed to increase the fertility of women.

**Observations** A large climber; leaves arranged spirally, ovate to suborbicular, 8–15 cm × 6–13.5 cm, base cordate to rounded, apex shortly acuminate, margin entire, with 4–5(–6) pairs of lateral veins, main veins appressed hairy below; figs axillary, depressed globose to subpyriform, to 35 mm in diameter, minutely brown velutinate, ripening dingy purple; flowers sessile. *F. nasuta* is locally common in lowland to montane forest, up to 1600 m altitude.

**Selected sources** 281, 609, 611.

### ***Ficus nodosa* Teijsm. & Binnend.**

Natuurk. Tijdschr. Ned. Ind. 29: 245 (1867).

**Synonyms** *Ficus du* Lauterb. & K. Schumann (1900).

**Vernacular names** Indonesia: laura (Moluccas). Papua New Guinea: kem kem (New Ireland).

**Distribution** The Moluccas, New Guinea, east to the Solomon Islands and south to northern Australia (Queensland).

**Uses** In New Britain, the leaves are applied externally as a styptic and antiseptic. The fibrous bark is used to make strings and clothing. The leaves and figs are edible.

**Observations** A medium-sized, buttressed tree up to 30 m tall, bark surface grey to reddish-brown, inner bark fibrous; leaves alternate, broadly ovate, 15–37 cm × 10–30 cm, base rounded to cordate, apex obtuse to acuminate, margin sinu-

ate, with 5-7 pairs of veins, glabrous, stipules silky; figs cauliflorous and ramiflorous, subglobose to subpyriform, 25-40 mm in diameter, densely lenticellate, ripening yellow to purple-brown; flowers with 3-4 lacinate-dentate tepals fused at base, male flowers in 3 rows, sessile, female flowers sessile or shortly stipitate. *F. nodosa* is found in lowland forest.

**Selected sources** 167, 248, 281, 576, 597, 1104.

**Ficus pachyrrachis Lauterb. & K. Schumann**

Fl. Schutzgeb. Südsee: 282 (1900).

**Synonyms** *Ficus grandis* King (1888) non Miq., *Ficus hypoglauca* Lauterb. & K. Schumann (1900), *Ficus pachythyrsa* Diels (1935).

**Vernacular names** Papua New Guinea: topu (Awala, Northern Province).

**Distribution** New Guinea.

**Uses** In Papua New Guinea, the latex is smeared onto an ulcer daily until cured. The leaves are edible, and the fibrous bark of saplings is used to make rope.

**Observations** A small to medium-sized tree up to 20 m tall; leaves arranged spirally or occasionally opposite, obovate to ovate-elliptical, 20-45 cm × 14-32 cm, base broadly cuneate to subcordate, apex shortly acuminate, margin denticulate, with 5-10 pairs of lateral veins, thinly hairy above, hispidulous velutinate below, stipules 2-4.5 cm long; figs on stout, leafless twigs, pyriform, 25-30 mm in diameter, villous but glabrescent; male flowers in 2-3 rows, subsessile, with 1 stamen, female flowers subsessile to stipitate, with cupular, short perianth. *F. pachyrrachis* is found in lowland to montane forest, up to 1500 m altitude.

**Selected sources** 281, 604, 723, 1104.

**Ficus pachystemon Warb.**

K. Schumann & Lauterb., Nachtr. Fl. Schutzgeb. Südsee: 242 (1905).

**Synonyms** *Ficus mangiferifolia* Lauterb. & K. Schumann (1901) non Griffith, *Ficus brassii* Summerh. (1929) non Sabine, *Ficus aechmophylla* Summerh. (1933).

**Vernacular names** Papua New Guinea: wawaina (Rabagi, New Britain).

**Distribution** New Guinea and the Bismarck Archipelago.

**Uses** In New Britain, leaves are chewed and swallowed to relieve diarrhoea. The bark is used for clothing.

**Observations** A shrub or small to medium-sized tree up to 20 m tall; leaves arranged spirally

to subdigitate, lanceolate to ovate-lanceolate, 9-21 cm × 1.8-6 cm, base cuneate, apex attenuate to subacute, margin entire, with (9-)11-19 pairs of lateral veins, glabrous, stipules 2-5.5 cm long; figs axillary, paired, sessile, subglobose to ellipsoid, 12-16 mm in diameter, glabrous, ripening through white, yellow, pinkish-orange to red; flowers with 3-4 tepals, male flowers dispersed, tepals free, stamen 1, female flowers sessile, tepals fused at base. *F. pachystemon* is found in lowland forest, commonly on river banks and in stony river beds.

**Selected sources** 281, 603.

**Ficus parietalis Blume**

Bijdr. fl. Ned. Ind. 9: 462 (1825).

**Synonyms** *Ficus cerasiformis* Desf. (1829), *Ficus grandifolia* Wallich ex Miq. (1848).

**Vernacular names** Indonesia: pelas kebo, seprah (Javanese). Malaysia: ara kesinai, ara landang puteh, sepedeh (Peninsular). Thailand: ma hai (Chiang Mai), maduea khon (Nakhon Si Thammarat).

**Distribution** Vietnam, Thailand, Peninsular Malaysia, Sumatra, Java, Borneo and the Philippines (Palawan).

**Uses** A decoction of the roots is used to treat stomach-ache.

**Observations** A climber or epiphytic shrub; leaves distichous, elliptical to oblong, 9-30 cm × 4-12 cm, base rounded to cuneate, apex long tipped, margin entire, with 2-6 pairs of lateral veins and prominent reticulation below, 3-veined at base, rusty pubescent below, stipules up to 8 mm long; figs solitary or paired, globose, 10 mm in diameter, hispidulous, ripening orange to reddish-brown; flowers with 4 tepals, male flowers with free or shortly connate tepals and 1 stamen, female flowers sessile or stipitate, tepals connate to halfway. *F. parietalis* is common in lowland forest, up to 1000 m altitude, often on riversides.

**Selected sources** 202, 281, 284, 1289, 1380, 1564.

**Ficus pumila L.**

Sp. pl. 2: 1060 (1753).

**Synonyms** *Ficus stipulata* Thunb. (1786), *F. scandens* Lamk (1788), *F. repens* Hort. var. *lutchuensis* Koidz. (1925).

**Vernacular names** Creeping fig, fig ivy (En). Indonesia: karet rambat. Thailand: madueo thao, lin suea. Vietnam: c[aa]y th[awf]n l[awf]n, cl[aa]y x[ooj]p x[ooj]p, bi l[eej].

**Distribution** Indigenous in Japan, China, the

Ryukyu Islands, Taiwan and northern Indo-China. At present widely cultivated as an ornamental and pot plant in tropical and subtropical regions.

**Uses** In folk medicine in Vietnam, the fruits and the leaves are considered to be tonic and are used in cases of impotence, lumbago and as a galactagogue. Furthermore, they are considered a treatment for rheumatism, anaemia, haematuria, chronic dysentery and haemorrhoids. Externally the leaves are applied to carbuncles. The latex is reported to have anthelmintic properties. In cases of dropsy the plant ash is rubbed on the body. *F. pumila* is widely cultivated to cover walls and rock faces; the colourful figs add further lustre to the attractive green foliage. Various selections are commercially grown as a pot plant in temperate regions.

**Observations** A prostrate or climbing shrub, reaching up to 10 m or more, creeping and clinging close to walls or tree trunks by means of numerous aerial rootlets, ultimate branches 30–80 cm long, erect; leaves dimorphous, two-ranked, on sterile branches ovate, 1.5–3 cm long and shortly petioled, on fertile branches oblong, 5–10 cm long and with long petioles; figs solitary in the axils of leaves, pyriform, 40–60 mm long, yellow-brown pilose when young, ripening glabrous, red to dark blue; male flowers in many rows, filling the distal half of the fig, stipitate with 2–3 stamens, female flowers sessile or shortly stipitate. *F. pumila* is an increasingly common feature in urban areas.

**Selected sources** 97, 281, 287, 364, 856, 900, 921, 1035, 1126, 1178, 1289.

### ***Ficus pungens* Reinw. ex Blume**

Bijdr. fl. Ned. Ind. 9: 478 (1825).

**Synonyms** *Ficus myriocarpa* Miq. (1867), *Ficus ovalifolia* Ridley (1916), *Ficus kalingaensis* Merr. (1922).

**Vernacular names** Indonesia: gososo (Ter-nate). Papua New Guinea: baguai (Harigen, Sepik), wopope (North Solomons Province), ohohone (Sui, Northern province). Philippines: Kalin-ga fig (En).

**Distribution** The Philippines, the Moluccas, New Guinea and New Britain.

**Uses** In Papua New Guinea, the root or leaf latex is swallowed to cure cough quickly. Heated leaves are applied externally to relieve body pains. In some reports the latex is reported to be very poisonous. The leaves are eaten cooked as a vegetable. The bark is used to make mats. The water that flows from a large root that has been cut obliquely can be drunk after boiling.

**Observations** A small to medium-sized tree up to 25 m tall, sometimes with short stilt roots, bark surface finely fissured, grey-brown; leaves arranged spirally, ovate to broadly ovate, 12–45 cm × 8–36 cm, base subcordate to subcuneate, apex with a short tip, margin serrate to denticulate, with 5–10 pairs of lateral veins, variably scabrid on both surfaces, stipules up to 7 cm long; figs on leafless twigs from the trunk and branches, paired, sessile, pyriform, 4–8 mm in diameter, puberulous but glabrescent, ripening red; flowers with 3–4 free tepals, male flowers in 1 row, sessile, with 1 stamen, female flowers sessile or shortly stipitate. *F. pungens* is locally common in primary and secondary lowland forest, up to 1700 m altitude.

**Selected sources** 281, 580, 604, 605, 1274, 1356.

### ***Ficus religiosa* L.**

Sp. pl. 2: 1059 (1753).

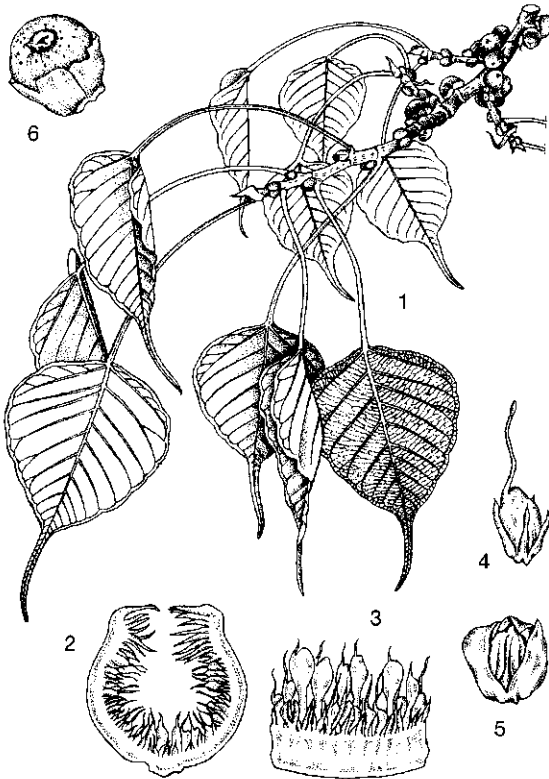
**Synonyms** *Ficus caudata* Stokes (1812), *Ficus superstitionosa* Link (1822), *Ficus peepul* Griffith (1854).

**Vernacular names** Bo tree, bodhi tree, pipal tree (En). Indonesia: bodhi. Cambodia: dom pur. Laos: pho. Thailand: pho see ma haa pho (central), yong (Shan-Mae Hong Son), salee (northern). Vietnam: c[aa]ly b[oo]f d[eef], c[aa]ly da, c[aa]ly da b[oo]f d[eef].

**Distribution** Originally from the Himalayas to southern China (Yunnan), Vietnam and northern Thailand; nowadays widely cultivated in the Malesian region but also in e.g. the Middle East, northern Africa and the United States.

**Uses** A decoction of the bark is used as skin wash to treat scabies, whereas the aerial roots are chewed by women to promote fertility. In India, an infusion of the bark is drunk as an antidiabetic and used externally against ulcers and skin diseases. The leaves and twigs are reputedly used against bites of venomous animals, as an astringent, anticonorrhoeal, laxative, aphrodisiac, and for the treatment of haemoptysis and fistula. Fresh sap from the leaves is used to cure diarrhoea, cholera and for wound healing. In Vietnam, the aerial roots are considered to be diuretic and used in ascites. The leaves and twigs are also applied as fodder. The fibrous bark is used to make paper. The fruits and tender leaf buds are edible though not tasty, and are considered to be cooling, alterative and laxative. The latex can be applied as birdlime. The tree is a host of the lac insect. The low-quality wood may be used for packing





*Ficus religiosa* L. – 1, fruiting twig; 2, fig in cross section; 3, part of receptacle with flowers; 4, female flower; 5, male flower; 6, infructescence.

cases and matches. The bark contains tannin which may be used to tan leather and for dyeing cloth. The tree is sacred to Hindus and Buddhists, and the trees which were brought to Sri Lanka in 245 B.C. are the oldest known trees in the world. It is regularly planted as a roadside tree.

**Observations** An evergreen or deciduous banyan or small to medium-sized tree up to 20 m tall, bark surface fissured, grey; leaves arranged spirally, ovate-cordate to ovate, 6–26 cm × 4–16 cm, base subcordate to truncate, apex caudate, margin often uneven or sinuous, with 6–9 pairs of lateral veins, glabrous, stipules up to 1.5 cm long; figs axillary, paired, sessile, subglobose, 10–15 mm in diameter, glabrous, ripening pink, purple or black; flowers with free tepals, male flowers in 1 row, sessile, with 2–3 tepals, female flowers sessile or stipitate, with 3–4(–5) tepals. *F. religiosa* occurs naturally in submontane forest.

**Selected sources** 54, 125, 202, 281, 284, 287, 478, 800, 900, 921, 1035, 1115, 1152, 1191, 1289, 1319, 1350, 1478, 1525.

### *Ficus rumphii* Blume

Bijdr. fl. Ned. Ind. 9: 437 (1825).

**Synonyms** *Ficus cordifolia* Roxb. (1832), *Ficus conciliorum* Oken (1841), *Ficus damit* Gagnep. (1927).

**Vernacular names** Mock bodh tree (En). Indonesia: ancak (Bali), bandira (Javanese), waringin jawa (Ambon). Thailand: pho kheek nok, pho prasaat, pho tua phuu (central). Vietnam: l[aa]m v[oof], da m[is]t.

**Distribution** India, Cocos Island, the Andaman and Nicobar Islands, Burma (Myanmar), Indo-China, Thailand, Peninsular Malaysia, Java, Sulawesi, the Lesser Sunda Islands and the Moluccas.

**Uses** The latex and fruits are emetic and anthelmintic, and used to treat itch. The latex is given internally as a vermifuge and for the relief of asthma. The tree is a host of the lac insect, and is also planted as a shade tree. The young leaves and ripe fruits are edible, raw or cooked. The leaves and twigs may be used as fodder for cattle and elephants. The bark yields a rough cordage. In India, the soft wood is used as fuel and for the production of charcoal.

**Observations** A deciduous, small to medium-sized tree or strangler up to 20 m tall, bole often fluted, bark surface flaky, silvery grey; leaves arranged spirally, ovate to broadly ovate, 7.5–17.5 cm × 5–12.5 cm, base cordate to rounded, apex acuminate, margin entire or wavy, palmately veined at base, with 4–6 pairs of lateral veins, glabrous, stipules 1–2.5 cm long; figs paired, sessile, globose, 10–15 mm in diameter, glabrous, whitish with dark spots, ripening pink or purple, then black; male flowers few, with 2(–3) free tepals and 1 stamen, female flowers sessile or shortly stipitate, with 3 fused tepals. *F. rumphii* is found on rocky coasts and is commonly planted.

**Selected sources** 202, 281, 284, 287, 478, 580, 900, 1289, 1380, 1564.

### *Ficus sagittata* J. König ex Vahl

Symb. bot. 1: 83 (1790).

**Synonyms** *Ficus ramentacea* Roxb. (1832), *Ficus crininervia* Miq. (1861), *Ficus ramosii* Merr. ex Sata (1944).

**Vernacular names** Indonesia: darandan, ham-pelas telpe (Sundanese), lawean (Javanese). Malaysia: akar beringin, sepedeh (Peninsular). Vietnam: sung d[af]u t[ee]n.

**Distribution** From north-eastern India (Sikkim, Assam), the Andaman Islands to Burma (Myanmar), Indo-China, southern China, Thailand

and throughout Malesia except for New Guinea, and in the Caroline Islands (Palau group).

**Uses** The leaves are said to have narcotic properties. *F. sagitata* is sometimes cultivated for its ornamental value.

**Observations** A large climber; leaves elliptical to ovate, 6–22 cm × 4–11.5 cm, base broadly cuneate to subcordate, apex acute, margin entire, 3-veined at base, main veins sunken above, with 6–8 pairs of lateral veins, glabrous, stipules up to 17 mm long; figs solitary or paired, subglobose, 14–18 mm in diameter, ripening red. *F. sagittata* is common in lowland and montane forest, up to 1500 m altitude.

**Selected sources** 202, 281, 1289, 1564.

### ***Ficus septica* Burm.f.**

Fl. ind.: 226 (1768).

**Synonyms** *Ficus hauili* Blanco (1837), *Ficus casearia* F. v. Mueller ex Benth. (1873), *Ficus kaukauensis* Hayata (1918).

**Vernacular names** Indonesia: awar-awar (general), ki ciyat (Sundanese), tagalolo (Sulawesi, Ternate). Papua New Guinea: omia (Kurereda, Northern Province), manibwohebwahe (Wagawaga, Milne Bay), bahuueru (Vanapa, Central Province). Philippines: hauili (Filipino), kauili (Tagalog), sio (Bikol).

**Distribution** The Ryukyu Islands, Taiwan, throughout Malesia except for Peninsular Malaysia, the Solomon Islands to Vanuatu and northern Australia (Queensland).

**Uses** In the Moluccas and New Guinea, the roots are chewed as an antidote, the latex of the leaves and fruits is used to produce purging, and the fruits are also emetic. In Papua New Guinea, the leaves are applied to cure colds, coughs, fever and fungal and bacterial diseases, whereas root scrapings or leaves have been mixed with water and drunk to cure dysentery or diarrhoea. The crushed root, mixed with coconut water, is drunk daily to treat urinary tract infections. In the Philippines, the leaves are applied for rheumatism and used as a sudorific to treat headache. The roots are used as a poultice in boils and a decoction is prescribed as diuretic. The latex is used to cure certain varieties of herpes, and wounds caused by poisonous fish. In Java, the dried leaves were formerly used as a substitute for opium or mixed with it.

**Observations** A small to medium-sized tree up to 25 m tall, bark surface pale grey or white; leaves alternate or decussate, elliptical, ovate or oblong, 10–28 cm × 4–13.5 cm, base cordate to cuneate, apex acuminate, margin entire, with

6–12 pairs of lateral veins, glabrous, stipules 1–4 cm long; figs axillary or cauliflorous, paired, obovoid to depressed subglobose, to 15 mm in diameter, ripening white to yellowish; male flowers with 1 stamen, female flowers with united tepals. *F. septica* is found in secondary rain forest and scrub vegetation on various soil types, up to 1800 m altitude.

**Selected sources** 137, 140, 202, 248, 281, 430, 580, 597, 603, 604, 606, 856, 1178.

### ***Ficus subcuneata* Miq.**

Ann. Mus. Bot. Lugd.-Bat. 3: 235 (1867).

**Synonyms** *Ficus stoechotricha* Diels (1935), *Ficus formosa* Summerh. (1941).

**Distribution** Sulawesi, the Moluccas, New Guinea and New Britain.

**Uses** In Papua New Guinea, the latex is applied directly to scabies or a skin rash.

**Observations** A small to medium-sized tree up to 30 m tall, bark surface greyish-brown to reddish or orange-brown, branches whorled; leaves arranged spirally, obovate, 12–20 cm × 4.5–9 cm, base broadly cuneate to subcordate, apex acuminate, margin entire, with (7–)9–13(–16) pairs of lateral veins, hairy below, stipules up to 22 mm long; figs axillary, solitary or occasionally paired, depressed globose, up to 40 mm in diameter, glabrescent, ripening scarlet; flowers with 4–6 free tepals, male flowers in 1 row, sessile, with 1 stamen, female flowers sessile. *F. subcuneata* is locally common in rain forest, often on riversides, from sea-level up to 1200 m altitude.

**Selected sources** 167, 281, 611.

### ***Ficus sublimbata* Corner**

Gard. Bull. Sing. 18: 50 (1960).

**Distribution** New Guinea.

**Uses** In Papua New Guinea, the latex from the leaves is applied directly onto a sore.

**Observations** A small tree up to 10 m tall; leaves arranged spirally to decussate, elliptical to obovate, 10–35 cm × 4–17 cm, base rounded to subcuneate, apex acuminate, margin entire to dentate or denticulate, with 4–8 pairs of lateral veins, scabrid on both sides, stipules 1.5–4 cm long; figs cauliflorous or ramiflorous, paired, slightly depressed globose, 18–25 mm in diameter, hairy but glabrescent, ripening red-brown to purple-red; male flowers in 2 rows, with 1 stamen, female flowers sessile or stipitate, perianth a short collar. *F. sublimbata* is locally common in montane forest and grassland, at 1500–2000 m altitude.

**Selected sources** 281, 611.

**Ficus wassa Roxb.**

Fl. ind., Carey ed. 3: 539 (1832).

**Synonyms** *Ficus eulampra* K. Schumann (1900), *Ficus rhodocarpa* Summerh. (1929), *Ficus nubigena* Diels (1935).

**Vernacular names** Indonesia: gohi, wassa, wassa laki-laki (Moluccas). Papua New Guinea: avavaia (Nangananga, New Britain), gabajekni (Nyamikum, Sepik).

**Distribution** The Lesser Sunda Islands, the Moluccas, New Guinea, the Solomon Islands and Vanuatu.

**Uses** In New Britain, the bark is scraped and chewed to give quick relief in dysentery. The leaves are used as sandpaper. Young leaves are eaten raw or cooked. In Papua New Guinea, the leaves and fruits are eaten raw or cooked, and the fibrous bark is used for men's head covering. The bark is also applied to make rope.

**Observations** A small to medium-sized tree up to 20 m tall, bark surface brown; leaves arranged spirally or occasionally opposite, elliptical to ovate, 9–20 cm × 3–8 cm, base cuneate, apex acuminate, margin entire to denticulate, with 4–7(–8) pairs of lateral veins, scabrid on both sides, stipules 6–13 mm long; figs axillary, ramiflorous or cauliflorous, subglobose, 12–16 mm in diameter, scabridulous, ripening yellow to red and purple; flowers sessile or shortly stipitate, with 4–6 free tepals, male flowers in 2–3 rows, stamens 1–2(–3). *F. wassa* is a forest species, generally preferring open places, and can be found from sea-level to 2000(–2600) m altitude.

**Selected sources** 281, 576, 580, 597, 603, 900, 1104.

J.P. Rojo, F.C. Pitargue & M.S.M. Sosef

**Gloriosa superba L.**

Sp. pl. 1: 305 (1753).

COLCHICACEAE

2n = 22, 66, 88

**Synonyms** *Gloriosa simplex* auct.

**Vernacular names** Flame lily, superb lily (En).

Indonesia: kembang sungsang (general), pacing tawa (Javanese), katongkat (Sundanese). Philippines: climbing lily, Turk's cap. Cambodia: var sleng dong dang. Laos: phan ma ha. Thailand: dong dueng (general), ma khaa kong (northern), waan kaam puu (central). Vietnam: c[aa]y ng[os]t ngh[ex]o, c[aa]y nh[us] nho[as]i.

**Origin and geographic distribution** Flame lily is widespread in the Old World tropics, occur-

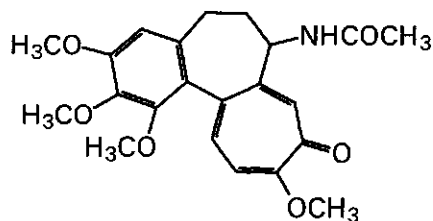
ring from tropical and southern Africa and Madagascar to India, Burma (Myanmar) and South-East Asia as far as Java and the Lesser Sunda Islands. It is not indigenous in Sumatra and Borneo. It is commonly grown as a garden ornamental in the tropics.

**Uses** The use of flame lily in folk medicine in South-East Asia is rather limited, probably due to its high toxicity. This may explain the reports of its use as poison in Cambodia and as suicidal agent in Burma (Myanmar). The tubers are applied in traditional Ayurvedic medicine in India. They are used as a tonic, cholagogue, antiperiodic, alterative, anti-arthritis, antileprotic, antihaemorrhoid and purgative. In East Africa the tubers have various applications in folk medicine but their main use exploits the poisonous constituents.

The selection known as *G. rothschildiana* O'Brien is cultivated in Europe and the United States for the production of cut flowers.

**Properties** Flame lily seeds are valued as a commercial source of colchicine, an amino alkaloid which is biosynthetically derived from the amino acids phenylalanine and tyrosine. The tubers also contain colchicine, but the content of the seeds is reported to be up to ten times higher, hence the importance attached to good seed set when the plant is grown for colchicine production. A report from Rwanda claims that the highest colchicine content is present in the young leaves.

In medicine, colchicine is used in the treatment of gout and rheumatism. At present it is the drug of choice for acute gout. It reduces the inflammatory reaction to urate crystals deposited in the joints. Its efficacy might be due to several actions, including decreased leucocyte mobility. The substance is not an analgesic, and has no effect on blood concentration, nor renal excretion of uric acid. Because of its highly toxic nature, colchicine should be used under supervision of a physician. Diarrhoea, nausea, vomiting and abdominal pains are often the first signs of poisoning. The diarrhoea may become severe and haemorrhagic, and can



colchicine

thus lead to metabolic acidosis, dehydration, hypotension and shock. A burning sensation in the throat, stomach and skin may also be an early sign of intoxication. Severe reactions include extensive vascular damage and acute renal toxicity with oliguria and haematuria. The patients may develop convulsions, delirium, muscle weakness, neuropathy and ascending paralysis of the central nervous system. In patients who have taken an overdose of *G. superba* bulbs, death occurs as a result of respiratory depression and cardiovascular collapse.

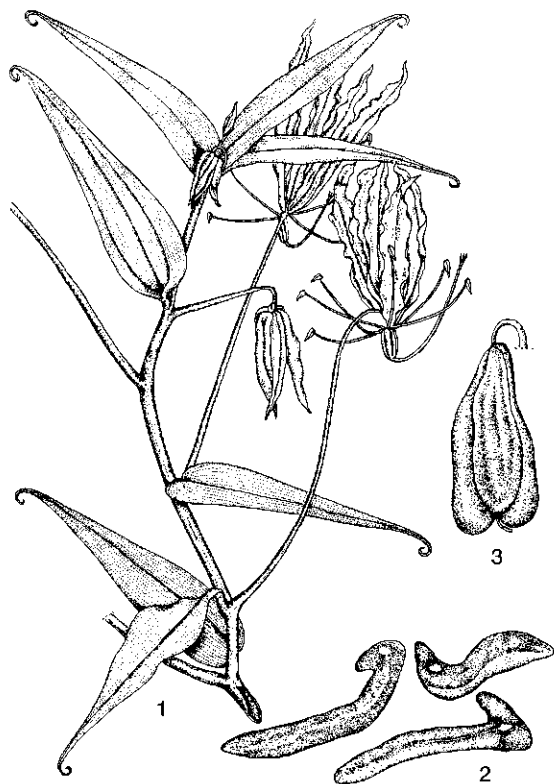
Colchicine inhibits cell division and is used in plant breeding to produce polyploidy, as it does not prevent chromosome division but inhibits formation of a mitotic spindle figure. Therefore no sister cells are formed. This so-called 'C-mitotic' activity of the alkaloid may arise from interaction with the disulphide bonds of the spindle protein, and from inhibition of the conversion of globular proteins to fibrous proteins. Once the treatment has stopped, however, the spindle figure forms again in the normal way. Colchicine also inhibits the division of animal cells, but it is too poisonous to be used to arrest tumour growth. A biosynthetic precursor of colchicine, demecolcine, has a wider margin of safety and is used for the treatment of myelogenous leukaemia and malignant lymphoma.

Extracts of the shoots and of the tubers of the plant show strong nematocidal activity, which can be largely attributed to colchicine. The chemical constituents of the tuber are known to be very poisonous to fish. Uterine stimulant properties are also reported.

Several other alkaloids have been isolated from tubers and seeds besides colchicine e.g. 1,2-didesmethyl colchicine, 2,3-didesmethyl colchicine, 3-desmethyl colchicine, N-formyl-N-deacetyl colchicine, N-deacetyl-2,3-didesmethyl colchicine, cornigerine, 2-desmethyl colchifoline and colchicoside.

**Adulterations and substitutes** The corms of *Colchicum autumnale* L. and *Iphigenia* (both also included in *Colchicaceae*) are traditional sources of colchicine. An increase in demand for colchicine stimulated the search for an alternative source, leading to *G. superba*.

**Description** A climbing or, less often, erect, glabrous herb, 3(-6) m long; tuber (corm) perennial, horizontal, roots fibrous; aerial stem annual, moderately branched. Leaves alternate, opposite or in whorls of 3(-4), sessile, lanceolate or ovate, entire, base obtuse, apex gradually narrowing in a coiled tendril (except for erect forms). Flowers



*Gloriosa superba* L. - 1, flowering stem; 2, tubers; 3, fruit.

solitary in the axils of leaves, bisexual; perianth segments 6, connate at the base, narrowly elliptical, 5-7(-9) cm × 1(-3) cm, with undulate margins, usually reflexed, yellow, red or 2-coloured; stamens 6, filaments spreading 2.5-5 cm long, anthers 0.7-1 cm long, dorsifixed; ovary superior, sessile, 3-celled, style filiform, bent basally almost at right angle, 3.5-5.5 cm long including short apical branches. Fruit a capsule, 4-10 cm × 1.5-2 cm. Seeds globose, about 5 mm in diameter, red or orange red, with a fleshy sarcotesta.

**Growth and development** The perennial organ of flame lily should be classified as a hypopodial tuber, mostly consisting of two metaphorous hypopodia with an apical bud enveloped by a prophyll, all covered in the remains of leaf bases. Two or more hypopodial tubers are left in the substrate after each growing season, while the previous season's tuber starts to shrivel. Plants propagated from seed take 3-4 years to bloom. Plants produced from tubers start flowering after 5 weeks, and continue flowering for about another 7 weeks. Development from visible flower bud to bloom

takes about 2 weeks and anthesis occurs 1 day later; anther dehiscence takes another day. The same branch flowers at 3-day intervals. Terminal flowers do not usually set fruit, but if they do only a few seeds are produced. Cross-pollination generally improves seed production. Pollination is probably by butterflies and sugar birds. Fruit is mature 6–10 weeks after pollination. The dimensions of the plants are strongly correlated with tuber weight, whereas the plants' dimensions determine the size of the next season's tuber.

**Other botanical information** *Gloriosa* has often been included in *Liliaceae*, but is nowadays considered as belonging to *Colchicinaeae*, differing from the bulbous *Liliaceae* s.s. in having a tuber or corm, i.e. a thickened underground stem part which is stoloniferous in *Gloriosa*. *G. superba* is very variable and is sometimes considered to consist of several species. Only one species, *G. superba* s.s., is indigenous to Malasia. Several selected forms in cultivation are traded under a 'specific rank', e.g. *G. rothschildiana*.

**Ecology** In general, flame lily prefers a pronounced monsoon climate, avoiding per-humid tropical areas. It is locally common in brushwood, hedges and open forest up to 300(–600) m altitude. It can be found flowering and fruiting throughout the year, although flowering is most abundant during the rainy season.

**Propagation and planting** Chemical scarification (e.g. with 1% hypochlorite) or removal of the sarcotesta reduces seed dormancy in flame lily from 6–9 months to about 4 months, and accelerates germination from 29–30 days to 11–15 days. Germination rates as high as 97% were reached for seeds incubated at 20–25°C for a period of 31 days. Higher temperatures have adverse effects.

Vegetative propagation by tubers is common practice, but not very suitable for the establishment of large plantings. The maximum number of daughter corms produced per plant is two. Separating the bilobed hypopodial tubers produces a higher percentage of flowering plants than leaving the tubers undivided (about 97% versus about 63%). Tuber dormancy can be overcome by soaking in continuously aerated water. Small tubers have been found to have a higher multiplication rate than bigger ones. Seed yield could be obtained from tubers heavier than 7 g.

Experiments with clonal propagation, using a Murashige and Skoog medium supplemented with 0.1 mg/l indole acetic acid have given promising results. However, the cloned plants did not flower at the first vegetative phase but produced small

tubers that could be used as vegetative propagules.

For small-scale plantings, tubers of 50–60 g are planted 30–45 cm apart in well-tilled soil at a depth of 6 cm in furrows 45–60 cm apart. A closer spacing gives a higher percentage of cross-pollination resulting in improved fruit set. The best seedling medium is a 1:1:2 mixture of soil, sand and compost. In Thailand, the optimum fertilizer (13-13-21) rate for optimal seedling growth and tuber weight was found to be about 250 kg/ha.

**In vitro production of active compounds** In vitro production of colchicine is feasible. Levels of colchicine extracted from *G. superba* callus, malformed roots, and entire plantlets show an increase that can be directly related to the amount of differentiation in culture. To derive levels of colchicine in vitro equal to those found in complete plants, entire plantlet regeneration is necessary.

**Husbandry** Although considerable information is available on the cultivation of flame lily under greenhouse conditions, the techniques used are not applicable to field conditions in tropical regions.

At the time of planting a dose of 40 kg N, 50 kg P<sub>2</sub>O<sub>5</sub> and 75 kg K<sub>2</sub>O per ha should be applied with a top dressing of 80 kg N per ha, 8 weeks after planting. The top dressing should coincide with staking of the growing vines. Irrigation is needed at dry weather during the initial stages of growth. Irrigation applied after flowering may cause the tubers to rot.

Poor fruit set resulting in low yield of seed is a problem in plantings in southern India. This may be attributed to inadequate pollination, which can be overcome by artificial hand pollination.

**Diseases and pests** Leaf blight (*Curvularia lunata*) and tuber rot (*Sclerotium* spp.) are two important fungal diseases of flame lily under per-humid situations. Lily caterpillar (*Polytela gloriosae*) and green caterpillar (*Pulsia chalcites*) attack foliage and flower buds.

**Harvesting** Mature fruits of flame lily are hand picked, and the tubers are dug out manually.

**Yield** In Tamil Nadu, India, small-scale plantings, raised from tubers, yield on average 250–300 kg of seed per ha from the second year onwards. In South Africa the seed production of 'wild-type' plants is positively correlated with height of the plant, and is on average 258 seeds per plant for plants 60–65 cm tall compared with about 30 seeds per plant for plants 30–40 cm tall.

**Handling after harvest** After harvesting, mature fruits of flame lily are left in the shade to dry

for 7–10 days. The fruits are then split open and the seeds removed, dried for a week in the shade and subsequently sun dried for another week.

**Genetic resources and breeding** *G. superba* has a wide natural distribution, and many selections are cultivated in the tropics as well as in greenhouses in temperate regions. Although local depletion of the resource may occur, the species is not threatened and the diversity still offers opportunities for further selection either for chemical constituents or as an ornamental.

**Prospects** Flame lily shows interesting potential as a source of colchicine, which is highly valued as a medicament for gout. As it is commonly grown as a garden ornamental in the tropics, there might be possibilities for the local production of drugs.

**Literature** [1] Chaudhuri, P.K. & Thakur, R.S., 1993. 1,2-didesmethylcolchicine a new alkaloid from *Gloriosa superba*. *Journal of Natural Products (Lloydia)* 56(7): 1174–1176. [2] Farooqi, A.A., Kumaraswamy, B.K., Bojappa, K.N., Pusalkar, V.R. & Gupta, R., 1993. Plantations of the clinically important *Gloriosa superba*. *Indian Horticulture* 37(4): 26–29. [3] Finnie, J.F. & Van Staden, J., 1994. *Gloriosa superba* L. (Flame Lily): Micro-propagation and in vitro production of colchicine. In: Bajaj, Y.P.S. (Editor): *Biotechnology in Agriculture and Forestry*. Vol. 26. Springer Verlag, Berlin, Heidelberg, New York, London, Paris, Tokyo. pp. 147–166. [4] Le Roux, L.G. & Robertse, P.J., 1994. Tuber ontogeny, morphology and vegetative reproduction of *Gloriosa superba* L. *South African Journal of Botany* 60(6): 321–324. [5] Le Roux, L.G. & Robertse, P.J., 1997. Aspects relating to seed production of *Gloriosa superba* L. *South African Journal of Botany* 63(4): 191–197. [6] Ntahomvukiye, D., Hakizimana, A., Nkiliza, J. & Van Puyvelde, L., 1984. Dosage de la colchicine dans le *Gloriosa simplex* L. (Liliacées) du Rwanda (Afrique Centrale) [Determination of colchicine in *Gloriosa simplex* L. (Liliaceae) from Rwanda (Central Africa)]. *Plantes Médicinales et Phytothérapie* 18(1): 24–27. [7] Pandey, R. & Haseeb, A., 1988. Studies in the toxicity of extracts of certain medicinal plants to rootknot nematode *Meloidogyne incognita* (Kifoid & White) Chitwood. *Indian Journal of Plant Pathology* 6(2): 184–186. [8] Pételot, A., 1954. Les plantes médicinales du Cambodge, du Laos et du Vietnam [The medicinal plants of Cambodia, Laos and Vietnam]. Vol. 3. *Archives des Recherches Agronomiques et Pastorales au Vietnam* No 22. Centre National de Recherches Scientifiques et Techniques, Saigon,

Vietnam. pp. 224–225. [9] Jessop, J.P., 1979. Liliaceae I. *Gloriosa*. In: van Steenis, C.G.G.J. (General editor): *Flora Malesiana*. Series 1, Vol. 9. Martinus Nijhoff/Dr. W. Junk Publishers, The Hague, Boston, London. pp. 193–195. [10] Watt, J.M. & Breyer-Brandwijk, M.G., 1962. *The medicinal and poisonous plants of Southern and Eastern Africa*. 2nd Edition. E. & S. Livingstone Ltd., Edinburgh, London, United Kingdom. pp. 700–707.

**Other selected sources** 277, 402, 893, 1035, 1058, 1287, 1447.

N. Bunyapraphatsara  
& J.L.C.H. van Valkenburg

## Heliotropium L.

Sp. pl. 1: 130 (1753); Gen. pl. ed. 5: 164 (1754).

BORAGINACEAE

$x = 7, 8, 9, 11, 13$ ; *H. curassavicum*:  $2n = 26, 52$ ,  
*H. indicum*:  $2n = 22$ , *H. ovalifolium*:  $2n = 22$

**Major species** *Heliotropium indicum* L.

**Vernacular names** Heliotrope (En). Hélioïtrope (Fr).

**Origin and geographic distribution** *Heliotropium* consists of about 250 species, and is distributed in tropical, subtropical and warm temperate regions of all continents. A total of 11 species are recorded for the Malesian region: 7 species can be considered indigenous and 1 naturalized; 3 species are known solely from cultivation, though they occasionally occur as a weed.

**Uses** The leaves of various *Heliotropium* species are generally applied in poultices throughout the Malesian region, and in other parts of the world. They are used to cure ulcers, wounds and local inflammations. A decoction of the leaves of *H. indicum* is reported to be applied in the treatment of urticaria. A decoction of various parts of the plants is used as an emmenagogue or even abortifacient in Indo-China and the Philippines. *H. arborescens* L. (synonym: *H. peruvianum* L.) is widely cultivated as an ornamental in tropical (e.g. Java), subtropical and temperate countries.

**Properties** Pyrrolizidine alkaloids are common constituents of various genera belonging to the *Boraginaceae* and *Compositae* and to the papilionoid genus *Crotalaria*. Biosynthetically, these alkaloids are derived from two molecules of the amino acid ornithine, which are utilized in the formation of the bicyclic pyrrolizidine skeleton (necine moiety). This basic skeleton seldom occurs in its free form, but is generally found as an ester

with rare monobasic or dibasic acids: the necic acids.

The interest in these compounds is mainly focused on their toxic effects in animals, livestock and sporadically in humans. Many pyrrolizidine alkaloids exhibit a pronounced hepatic toxicity. Toxic structures must have an 1,2-unsaturation in the pyrrolizidine ring, and an ester function on the side chain. Mammalian liver oxidases transform typical alkaloids into reactive pyrrole structures, which are potent alkylating agents and react with suitable cell nucleophiles, e.g. nucleic acids and proteins. Although the toxic effects of these metabolites are usually primarily seen in the liver, lung and/or other tissues may also be affected. In addition to the cytotoxic effects mentioned, mutagenic and carcinogenic activities of pyrrolizidine alkaloids have been reported in literature.

In general, animals will avoid eating plants containing pyrrolizidine alkaloids. However, in times of scarcity, and when fodder is contaminated, accidental consumption can lead to acute or chronic intoxication. In humans, generally only symptoms of chronic intoxication are observed, as a result of a prolonged use of herbal medicines consisting of plants containing pyrrolizidine alkaloids (e.g. *Senecio* and *Symphytum* species).

Indicine, indicine-N-oxide, acetyl-indicine, indicinine, heleurine, heliotrine, supinine, supinidine and lindelofidine are pyrrolizidine alkaloids isolated from *H. indicum*, that are all hepatotoxic. Furthermore, the literature contains reports of the presence of trachelanthamidine and retronecine in *H. indicum*. *H. curassavicum* contains trachelanthamidine, supinidine and retronecine. In all organs at all developmental stages, supinidine was the minor necine; trachelanthamidine was the dominant base in *H. curassavicum*, whereas retronecine was dominant in *H. indicum*. A larger proportion (17–65%) of retronecine has been found in the roots and generative parts of *H. curassavicum* than in the leaves and stems.

Indicine-N-oxide has shown significant activity against W-256 carcinoma in rats, and leukaemia L-121 in mice. The compound has also been administered as a brief infusion over 15 minutes to patients with solid tumours who had received prior chemotherapy. Dose-limiting effects were leukopenia and thrombocytopaenia, and the toxicity was cumulative with repeated doses. In general, the undesirable hepatotoxicity prevented any further development. In addition, anti-inflammatory and wound healing properties are reported.

Furthermore, phytochemical investigations have

revealed the presence of the biogenic amines putrescine, spermidine and spermine in leaves and inflorescence of *H. indicum*, and the sterols diosgenin, tigogenin, lanosterol and  $\beta$ -sitosterol in *H. scabrum*.

**Description** Annual or perennial herbs or subshrubs. Leaves alternate, simple, petiolate or sessile, stipules absent. Inflorescence usually a unilateral, scorpioid cyme, sometimes 2 cymes close together on dichotomous branches, or even numerous short cymes crowded together in a head-like manner, with or without bracts. Flowers actinomorphic, bisexual, 5-merous; calyx lobes almost free, unequal; corolla tubular, funnel-shaped or hypocrateriform, white, yellow or purple; stamens included in corolla tube, with very short filaments; ovary superior, 4-locular. Fruit undivided with 4 locules or by reduction 1 fertile locule, or separating in 2 bilocular pyrenes, most often these pyrenes again separating into 2 unilocular nutlets; nutlets smooth or sculptured. Seedling with epigeal germination; cotyledons leafy, glabrous, with rounded apex; hypocotyl elongated, densely hairy, epicotyl very short.

**Growth and development** *H. indicum* may flower throughout the year. The flowering season is very long, and new flowers develop apically within a cyme while mature nutlets are already present at the base of the inflorescence.

**Other botanical information** *Heliotropium* belongs to the subfamily *Heliotropioideae*, which also includes *Tournefortia*. Infrageneric classification suffers from the absence of a recent taxonomic revision covering Old World and New World species. The Malesian *Heliotropium* species belong to several sections.

**Ecology** *Heliotropium* occurs in very diverse habitats, though drier places are preferred in general. Some species are weeds, often introduced from the New World at an early date and now widespread in the palaeotropics.

**Harvesting** Plants of *Heliotropium* are harvested when fully grown, and can be used fresh or dried.

**Yield** *H. curassavicum* and *H. indicum* grown under greenhouse conditions showed the greatest accumulation of alkaloids after the beginning of flowering. Young leaves, young inflorescences and seedlings showed very high alkaloid levels. With ageing, the content of alkaloids in leaves decreased 20-fold. In both species the highest alkaloid content was found in the roots and inflorescence. These parts also exhibited the highest relative amounts of N-oxides ranging from 60–90%

the total alkaloid content. No significant age-dependent differences in N-oxides were found.

**Genetic resources and breeding** Most *Heliotropium* species are widespread and common weeds. Therefore the risk of genetic erosion seems slight. No breeding programmes are known to exist.

**Prospects** The possible applications in cancer therapy are limited by the toxic effects, in particular the hepatotoxic effects, of the pyrrolizidine alkaloids.

**Literature** |1| Birecka, H., Di Nolfo, T.E., Martin, W.B. & Frohlich, M.W., 1984. Polyamines and leaf senescence in pyrrolizidine alkaloid bearing *Heliotropium* plants. *Phytochemistry* 23: 991–997. |2| Bruneton, J., 1995. Pharmacognosy, phytochemistry, medicinal plants. Technique & Documentation Lavoisier, Paris, France. pp. 675–680. |3| Catalfamo, J.L., Martin, W.B. & Birecka, H., 1982. Accumulation of alkaloids and their necines in *Heliotropium curassavicum*, *H. spathulatum* and *H. indicum*. *Phytochemistry* 21(11): 2669–2675. |4| Dewick, P.M. (Editor), 1997. Medicinal natural products. John Wiley & Sons, Chichester, United Kingdom. pp. 283–285. |5| Gutierrez, H.G., 1982. An illustrated manual of Philippine materia medica. Vol. 2. National Research Council of the Philippines, Tagig, Metro Manila, the Philippines. pp. 442–444. |6| Holm, L.G., Plucknett, D.L., Pancho, J.V. & Herberger, J.P., 1977. The world's worst weeds. Distribution and biology. East-West Center. The University Press of Hawaii, Honolulu, United States. pp. 291–294. |7| Kugelman, M., Liu, W.C., Axelrod, M., McBride, T.J. & Rao, K.V., 1976. Indicine-N-oxide: the anti-tumor principle of *Heliotropium indicum*. *Lloydia* 39(2/3): 125–128. |8| Ohnuma, T., Sridhar, K.S., Ratner, L.H. & Holland, J.F., 1982. Phase I study of indicine N-oxide in patients with advanced cancer. *Cancer Treatment Report* 66(7): 1509–1515. |9| Riedl, H., 1997. Boraginaceae. In: Kalkman, C. et al. (Editors): *Flora Malesiana*. Series 1, Vol. 13. Rijksherbarium/Hortus Botanicus, Leiden, the Netherlands. pp. 43–144. |10| Van den Dungen, F.M., 1993. *Symphytum officinale* L. PhD-thesis Universiteit Utrecht, the Netherlands. 191 pp.

#### *Selection of species*

#### ***Heliotropium curassavicum* L.**

Sp. pl. 1: 130 (1753).

**Distribution** A native of the Americas from Patagonia to the United States and the West In-

dies; in Malesia occasionally occurring as a weed in Java and the Philippines.

**Uses** In the Americas the dried roots are ground to powder and applied to sores and wounds. In Curaçao a decoction of the plant is taken as a remedy for leucorrhoea. It is also taken as a substitute for *H. indicum*.

**Observations** An annual, prostrate to ascending, subsucculent herb; leaves oblong, spatulate to linear, 1–5 cm × 0.3–1 cm, glabrous, with short petiole; inflorescence a spike-like cyme, 3–10 cm long, dense, usually forked once, ebracteate; calyx glabrous, corolla 1–2.5(–3) cm long with 5 rounded lobes, white or bluish with a yellow centre, stigma an umbrella-shaped disk on a short style; fruit breaking up into 4 nutlets about 2 mm long, glabrous. In its native countries *H. curassavicum* is found in wet places, along the sea-shore and in similar habitats.

**Selected sources** 97, 221, 287, 921, 1229.

#### ***Heliotropium indicum* L.**

Sp. pl. 1: 130 (1753).

**Vernacular names** Indian heliotrope (En). Tournesol indien (Fr). Indonesia: buntut tikus (Malay), bandotan, gajahan (Javanese). Malaysia: rumput ekur kucing, rumput kala jenkeng, rumput oleh (Peninsular). Philippines: trompa ng elephante, buntut-leon (Tagalog, Bikol), kambra-kambra (Bisaya). Cambodia: promoi damrey, kantui damrey. Laos: nha nguong xang. Thailand: kuno kaa-mo (peninsular), yaa nguang chaang (general), yaa nguang chaang noi (northern). Vietnam: c[aa]y v[off]i voi.

**Distribution** Probably a native of tropical America, now widespread in all tropical regions of the world. *H. indicum* is a common weed throughout Malesia.

**Uses** *H. indicum* has been used on warts and in poultices since antiquity, to treat inflammatory tumours. In Indonesia a decoction of the leaves is used against thrush. In folk medicine in Indo-China the whole plant, either in decoction or as a poultice, is applied to treat inflammation, swelling, sprain, contusion, pharyngitis, abscesses and rheumatism. A poultice of the leaves is used in the treatment of herpes and rheumatism. In the Philippines, the roots are considered a good emmenagogue. An infusion of the leaves is used to wash wounds and sores. In Thailand, a decoction of aerial parts is applied as an antipyretic and anti-inflammatory, and the roots are used for eye diseases. In West Africa, a poultice is applied in the treatment of eczema and impetigo. In South



and Central America, the plant is used for similar purposes. In addition a decoction of the leaves is taken orally to cure dysentery and to treat haemorrhoids. The leaf juice is drunk to stop internal bleeding. An infusion is taken as a gargle to relieve sore throat.

**Observations** An annual herb, 15–60(–100) cm tall, stem simple or with a few branches, hairy; leaves ovate, (1.5–)2–10(–12) cm × 1–8(–9) cm, base truncate but narrowly long-decurrent, apex acute, with tubercles of mineralized cells and bristly hairs, petiole 1–9 cm long; inflorescence consisting of 1 to several spike-like cyme(s), elongated, 5–20 cm long, ebracteate; calyx with patent, bristly, white hairs, corolla salver-shaped, tube 3–4.5 mm long, lobes rounded, about 1 mm long, pale-violet, blue or white, apex of carpels strongly bidentate, strongly divergent at anthesis; fruit 2–3 mm long, fruit halves 2-celled, cells 2-locular, outer partition with one seed, inner one larger, empty. *H. indicum* is found in sunny places, on waste land, in periodically desiccating pools and

ditches and other anthropogenic habitats, in general up to 800 m altitude.

**Selected sources** 97, 190, 201, 202, 221, 287, 531, 580, 614, 787, 921, 979, 1035, 1072, 1126, 1128, 1178, 1229, 1299, 1386.

### *Heliotropium ovalifolium* Forssk.

Fl. Aegypt.-Arab.: 38 (1775).

**Distribution** Tropical Africa, Arabian Peninsula, India, Vietnam, Sumba, Timor and the Solomon Islands.

**Uses** In West and Central Africa, *H. ovalifolium* is used in the treatment of syphilis, both as a strong purgative and as a topical application to syphilitic ulcers. It is reported to have analgesic action and is applied in hot poultices to treat severe pain.

**Observations** A perennial herb, sometimes with woody base, up to 40 cm tall, much branched, covered by appressed short hairs; leaves obovate, 1–1.5 cm × 0.3–0.4 cm, tapering towards the base, apex obtuse, with silky hairs, petiole 0.1–0.5 cm long; inflorescence a spike-like cyme, up to 4 cm long, ebracteate, flowers variably densely arranged in two ranks; calyx densely covered by antrorse hairs, corolla funnel-shaped, 3 mm long, with dense antrorse hairs outside, lobes ovate-triangular to ovate, 0.8 mm long, white; fruit consisting of 4 nutlets, densely antrorse hairy. *H. ovalifolium* is found in valleys, on roadsides, in dried ponds and similar habitats.

**Selected sources** 201, 287, 921, 1229.

### *Heliotropium scabrum* Retz.

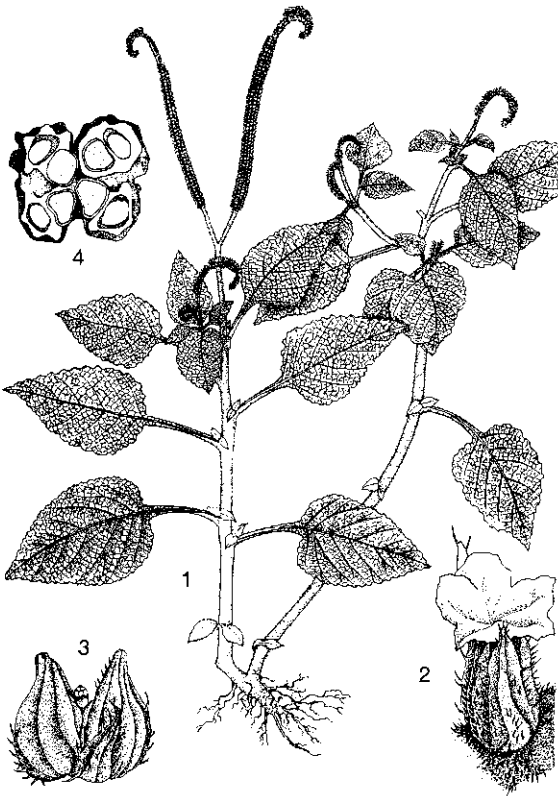
Observ. bot. 2: 8 (1781).

**Synonyms** *Heliotropium marifolium* Retz. (1781), *Heliotropium cyrtostachyum* Miq. (1856).

**Distribution** Pakistan, India, Sri Lanka, Cambodia, southern China and Malesia (Java and Flores).

**Uses** The alkaloids found in *H. scabrum* indicate that it can be used for the same purposes as some of the other *Heliotropium* species.

**Observations** A perennial, prostrate, sometimes erect herb, stems 5–30 cm long, branched from the base, covered in antrorsely appressed, white, bristly hairs; leaves linear to lanceolate-oblong, 0.5–2 cm × 0.1–0.5 cm, base cuneate or rounded, apex acute, margin revolute, with antrorsely appressed bristly hairs, petiole 0–3 mm long; inflorescence a subcapitate cyme at the end of stems and branches with leaf-like bracts or flowers single in axils of leaves; calyx covered in bristly hairs, accrescent, corolla funnel-shaped,



*Heliotropium indicum* L. – 1, plant habit; 2, flower; 3, bilobed fruit; 4, fruit in cross-section.

3–3.5 mm long, with wavy margins, white; fruit separating into 4 nutlets. *H. scabrum* occurs in dunes, but also in dry roadsides and lawns, and shady places, in general near the sea. However, in Sri Lanka it is reported to occur in foothills up to 1500 m altitude.

**Selected sources** 97, 921, 1229, 1369.

Wongsatit Chuakul, Noppamas  
Soonthornchareonnon & Promjit Saralamp

### **Holarrhena R. Br.**

Asclepiadeae: 51 (1810).

APOCYNACEAE

$x$  = unknown; *H. pubescens*:  $2n = 22$

**Major species** *Holarrhena pubescens* Wallich ex G. Don.

**Origin and geographic distribution** *Holarrhena* comprises 4 species and is found in tropical Africa, India, Sri Lanka, Burma (Myanmar), Indo-China, southernmost China, Thailand and northern Peninsular Malaysia.

**Uses** The stem bark of *Holarrhena* is a well-known medicine against amoebic dysentery, and has been used as such in India since antiquity. It has astringent, antidyenteric, anthelmintic, stomachic and febrifugal properties, and is also taken as a general tonic. The drug is usually administered as an extract or a decoction. The seeds are sometimes applied similarly; their oil is anthelmintic. Decoctions of the bark or leaves used to be applied in baths to cure scabies. The leaves are used in chronic bronchitis, and against boils, ulcers and haemorrhoids, sometimes as an anthelmintic. In Cambodia, the latex is used locally to treat conjunctivitis and in southern Vietnam as a vulnerary.

The pale yellow or pale pink wood is of low quality but is appreciated locally for small objects like picture frames, household utensils, carving and turnery articles, because of its fine grain. The wood ash has been used for dyeing and as a caustic to open abscesses. *H. pubescens* may prove useful for the reforestation of deforested land in comparatively dry regions, also because it is not readily browsed even by goats. It is also cultivated in India as an ornamental for its attractive flowers.

**Properties** The bark and seed of *H. pubescens* contain conessine, which is mainly responsible for the amoebicidal properties. This compound is the most important representative of a series of steroidal alkaloids that belong to the conanine type and that also occur in other *Holarrhena*

species, both Asian and African. In contrast, the steroidal alkaloids in the leaves are mostly 5 $\alpha$ -pregnane or pregn-5-ene derivatives with amino groups at the 3 or 20-positions. The fact that many of these substances have an oxygen or nitrogen function at position 18 has led to a detailed study of their chemistry aimed at using them as starting material for the partial synthesis of pharmaceutical steroids, such as adrenocortical hormones (corticosteroids) and sex hormones (e.g. oestrogens, progestogens and androgens). In all, about 40–45 steroidal alkaloids have been isolated from various parts of *H. pubescens*. The bark should be used with caution as an anti-amoebic, as one of the possible side effects is hypotension. Various aminoglyco-steroids are present in the leaves of *H. curtisii*, *H. mitis* (Vahl) R.Br. and *H. pubescens*. These substances contain an amino sugar, and in some of them e.g. holarosine and holacurtine, the steroidal part of the molecule is a cardenolide. In others, e.g. the holantosines, the oxygenated side-chain is of rather different structure. The 8 steroidal alkaloids isolated from the ethanolic extract of the leaves of *H. curtisii* all showed significant cytotoxic and leishmanicidal activity.

Apart from its anti-amoebic properties, conessine also possesses antitubercular activity in situ. It increases coronary outflow in isolated rabbit heart and induces narcosis in frogs. It also produces local anaesthesia in guinea-pigs, being about twice as active as cocaine, but causes tissue necrosis if injected subcutaneously. Furthermore, conessine is highly toxic to *Trichomonas intestinalis* and *T. vaginalis*.

The alcoholic extract of the fruit was found to show anti-cancer activity against human epidermoid carcinoma of the nasopharynx in tissue culture. The aqueous extract of the fruits exhibited hypoglycaemic effects in rats.

Chloroform and methanolic extracts of *H. pubescens* seed showed antibacterial activity against *Bacillus subtilis*, *Escherichia coli*, *Pseudomonas aeruginosa* and *Staphylococcus aureus*. Conessine from *H. pubescens* showed potential as a larval growth inhibitor, sterilant and antifeedant in laboratory tests on 4 insect species. The wood of *H. pubescens* contains 14% protein, 16% polyphenol, 5.4% oil and 4.8% hydrocarbons and due to its relatively high oil and hydrocarbon content it has been recommended as a renewable source of energy.

**Adulterations and substitutes** Seeds of *H. pubescens* are sometimes mixed with those of

*Wrightia tinctoria* R.Br., which do not have the same medicinal properties. However, they can be distinguished because *H. pubescens* seed is very bitter. Similarly, the bark of *W. tinctoria* is sold as a substitute for *H. pubescens*. The bark can be distinguished by a different thin layer chromatography pattern of the petroleum extract.

**Description** Evergreen or deciduous shrubs to medium-sized trees up to 25 m tall; bole short, up to 60 cm in diameter; bark flaking off irregularly, pale brownish or greyish, inner bark with abundant white latex; branchlets often pubescent. Leaves opposite or subopposite, simple, entire, pinnately veined, short petiolate; stipules intrapetiolar and often obscure or absent. Inflorescence terminal or seemingly axillary, cymose, many-flowered. Flowers bisexual, 5-merous, actinomorphic, fragrant; calyx persistent, sepals free or connate at the very base, valvate; corolla salver-shaped, white, tube cylindrical, pubescent at the throat, lobes in bud contorted and overlapping to the right; stamens included, inserted at the base of the corolla tube, filaments short, anthers mucronate; disk absent; ovary superior, with 2 carpels which are fused at base, ovules many, style 1, split at base, with an ovoid apex (stigma head). Fruit composed of two paired, slender follicles which split along a single line; wall thinly coriaceous. Seeds many, in 2 or more rows, linear to narrowly oblong, glabrous but with a dense and long tuft of hairs at the apex. Seedling with epigeal germination; cotyledons emergent, leafy; hypocotyl elongated; all leaves opposite.

**Growth and development** In India, early height increment of *H. pubescens* is low, only 10–15 cm/year under natural conditions. Its maximum annual diameter increment as observed in India is 0.7 cm, but is greatly reduced to 0.1 cm when it grows in shaded locations. In India, trees are leafless in December-January and new leaves appear after 2–3 months. Trees flower in (April-) May-June(-August), sometimes a second flush is produced in September-November. Fruits are full-grown in August-October but do not dehisce until February-April. The seeds are wind-dispersed.

**Other botanical information** *H. mitis*, an endemic from Sri Lanka, is also used medicinally for similar purposes. Information on the uses of *H. antidysenterica* from Sri Lanka probably refers to *H. mitis*.

**Ecology** *Holarrhena* is found in dry evergreen to dry deciduous forest, bamboo forest, scrub woodland and savannas, at 0–1500 m altitude. *H. pubescens* shows pioneer characteristics. In its

natural habitats in India, *H. pubescens* grows in areas with an absolute maximum temperature of 40–48°C and an absolute minimum temperature of 0–12°C, and an annual rainfall between 750 mm and 3800 mm.

**Propagation and planting** *Holarrhena* can be propagated by seed, either by direct sowing or by raising seedlings in the nursery. Fresh seed has a high germination rate, but seed loses its viability after having been stored for 1 year. *H. pubescens* has 32 000–35 000 seeds/kg. Seed usually germinates in 2–3 weeks. In a germination experiment in Malaysia seed of *H. curtisii* took 12–28 days to germinate. Tissue culture of *H. pubescens* in India proved possible and rooted plantlets were obtained from nodal explants of a 20-year-old tree cultured on Murashige and Skoog medium with added indole acetic acid at 1 mg/l.

**In vitro production of active compounds** In India a callus culture of *H. pubescens* yielded a maximum of 300 mg alkaloids per 100 g dry cells in 40 days; a suspension culture reached 130 mg per 100 g dry cells in 8 days. A modified Murashige and Skoog medium may result in an increased production up to 660 mg per 100 g dry cells, depending on nutrient concentration and source. The alkaloid production can be boosted to 110 mg per 100 g dry cell weight per day by adding cholesterol as a precursor. About 90% of the total alkaloids produced in cell culture was conessine.

**Husbandry** *H. pubescens* easily recovers from damage, coppices well and produces root suckers in abundance.

**Diseases and pests** The moth *Glyphodes latipennis* is one of the major pests on *H. pubescens* in Maharashtra State, India.

**Harvesting** In India, bark for medicinal use is collected from 8–12-year-old *H. pubescens* trees. The alkaloid content varies with the age of the tree and the season. The content in the bark is highest when new shoots are formed.

**Yield** Values for total alkaloid content may be as high as 4.3% for the stem bark of *H. pubescens*, but only 0.4% has also been reported for flowering plants. Leaves contain 1–1.6% and seeds 0.6–1.8% total alkaloids.

**Handling after harvest** The bark of *H. pubescens* is dried and sold in 1–7 mm thick pieces.

**Genetic resources and breeding** The Malaysian *Holarrhena* species are common in their areas of distribution and are also found in open anthropogenic habitats. Therefore the risk of genetic erosion seems limited. The considerable variation

in nature and content of alkaloids as found in market samples is probably also linked to non-seasonal differences. As both influence the therapeutic effect, breeding and selection may well improve the prospects of the crude drug.

**Prospects** The steroidal alkaloids of *Holarrhena* might be usable as starting material for the partial synthesis of steroids of pharmaceutical interest, so deserve further attention. Furthermore, the amoebicidal, bactericidal and insecticidal properties of the crude drug, and/or the main steroidal alkaloid conessine might be valuable alternatives e.g. in the treatment of amoebic dysentery.

**Literature** [1] Bisset, N.G., 1958. The occurrence of alkaloids in the Apocynaceae. *Annales Bogorienses* 3(1): 105–236. [2] Bisset, N.G., 1981. *Phytochemistry of Holarrhena* R.Br. Mededelingen Landbouwhogeschool Wageningen 81-2: 37. [3] Council of Scientific and Industrial Research, 1959. *The Wealth of India: a dictionary of Indian raw materials and industrial products*. Raw materials. Vol. 5. Publications and Information Directorate, New Delhi, India. pp. 103–207. [4] Dwivedi, R.K. & Tripathi, Y.C., 1991. Pharmacognostical, phytochemical and biological studies on *Holarrhena antidysenterica* Wall.: a review (part-1). *New Agriculturist* 1(2): 209–212. [5] de Kruijff, A.P.M., 1981. A revision of *Holarrhena* R.Br. (Apocynaceae). *Mededelingen Landbouwhogeschool Wageningen* 81-2: 1–36. [6] Kaul, M.K. & Atal, C.K., 1983. Studies on *Holarrhena antidysenterica* Wall.: 1 Botany, medico-ethnobotany and distribution. *Journal of Ethnopharmacology* 8(3): 349–356. [7] Nguyen Van Duong, 1993. Medicinal plants of Vietnam, Cambodia and Laos. Mekong Printing, Santa Ana, California, United States. pp. 58–59. [8] Panda, A.K., Bisaria, V.S., Mishra, S. & Bhojwani, S.S., 1991. Cell culture of *Holarrhena antidysenterica*: growth and alkaloid production. *Phytochemistry* 30(3): 833–836. [9] Pételot, A., 1953. *Les plantes médicinales du Cambodge, du Laos et du Vietnam* [The medicinal plants of Cambodia, Laos and Vietnam]. Vol. 2. Centre National de Recherches Scientifiques et Techniques, Saigon, Vietnam. pp. 125–129. [10] Thappa, R.K., Tikku, K., Bhaskar, P.S., Vaid, R.M. & Bhutani, K.K., 1989. Conessine as a larval growth inhibitor, sterilant, and antifeedant from *Holarrhena antidysenterica* Wall. *Insect Science and its Application* 10(2): 149–155.

### *Selection of species*

#### ***Holarrhena curtisii* King & Gamble**

Journ. Straits Branch Roy. Asiat. Soc. 74: 446 (1908).

**Synonyms** *Holarrhena densiflora* Ridley (1911), *Holarrhena latifolia* Ridley (1911), *Holarrhena pulcherrima* Ridley (1911).

**Vernacular names** Cambodia: doeum tuk das. Laos: mak mouk kuay, mouk noy<sup>2</sup>. Thailand: hatsakhun thet (Phangnga), mok noi (northern), phut nam (Surat Thani). Vietnam: h[oof] li[ee]n l[as] nh[or], m[uws]c hoa tr[aws]ng.

**Distribution** Indo-China, Thailand and northern Peninsular Malaysia.

**Uses** The bark and roots have been used to treat dysentery.

**Observations** A small shrub up to 2.5 m tall, branches pale grey to dark grey, often lenticellate or white-dotted; leaves narrowly elliptical or elliptical to obovate, 2.3–10 cm × 1.2–4.5 cm, apex rounded to emarginate, with 9–15 pairs of lateral veins, chartaceous when dry, petiole up to 6 mm long; corolla tube 12–22 mm long, the lobes 10–23 mm long; fruit carpels erect, at least when young, 11–33 cm × 3–8 mm. *H. curtisii* occurs in secondary rain forest, mixed deciduous forest, bamboo forest, scrub woodland, savanna and mangrove swamps, at 0–400 m altitude.

**Selected sources** 161, 330, 698, 1020, 1022, 1128, 1227, 1380.

#### ***Holarrhena pubescens* Wallich ex G.**

##### **Don**

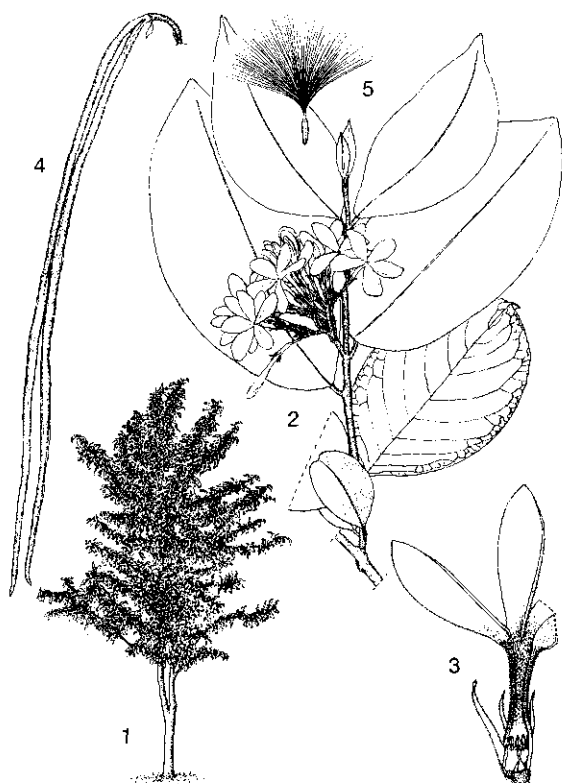
Gen. hist. 4: 78 (1837).

**Synonyms** *Holarrhena antidysenterica* (L.) Wallich ex A.DC. (1844), *Holarrhena malaccensis* Wight (1848), *Holarrhena macrocarpa* (Hassk.) Villar (1880).

**Vernacular names** *Holarrhena* (En). Ecorce de conessie (Fr). Cambodia: khlang kong, tuk das khla. Laos: mouk nhai<sup>2</sup>. Thailand: mok thung (northern), mok yai (central), so-thue (Karen-Mae Hong Son). Vietnam: m[oo]c hoa tr[aws]ng, h[oof] li[ee]n l[as] to.

**Distribution** Tropical Africa, India, Burma (Myanmar), Indo-China, southern China (Yunnan) and Thailand.

**Uses** The most important medicinal application of this species is for amoebic dysentery, for which especially the bark and occasionally also the seeds are used. The wood is used for small implements and as fuel wood, and the tree is also planted for its ornamental value.



*Holarrhena pubescens* Wallich ex G. Don - 1, tree habit; 2, flowering twig; 3, opened flower; 4, fruit; 5, seed.

**Observations** A shrub to medium-sized tree up to 18 m tall, bole up to 25 cm in diameter, bark surface rough and corky, longitudinally fissured, pale to dark grey; leaves ovate to elliptical or narrowly so, 1.7–19.5 cm × 1.3–11 cm, apex acute to acuminate, with 5–25 pairs of lateral veins, papery when dry, petiole up to 12 mm long; corolla tube 9–19 mm long, the lobes 10–24(–30) mm long; fruit carpels pendulous, 20–37.5 cm × 2–9 mm. *H. pubescens* is found in open places in evergreen rain forest, mixed deciduous forest, bamboo forest, scrub woodland and savanna, at 0–1500 m altitude. It is often found gregarious, on clay, laterite or sand, but also on rocky outcrops.

**Selected sources** 138, 149, 150, 158, 161, 202, 239, 287, 318, 330, 364, 402, 480, 675, 689, 725, 741, 904, 1035, 1109, 1110, 1111, 1113, 1128, 1227, 1277, 1287, 1348, 1349, 1352, 1380, 1478, 1538.

Wongsatit Chuakul, Noppamas  
Soonthornchareonnon & Promjit Saralamp

## Hydnocarpus Gaertner

Fruct. sem. pl. 1: 288, t. 60, f. 3 (1788).

FLACOURTIACEAE

$x = 12$ ; *H. alcalae*, *H. anthelmintica*, *H. kurzii*:  
 $2n = 24$

**Major species** *Hydnocarpus alcalae* C.DC., *H. anthelmintica* Pierre ex Lanessan, *H. kurzii* (King) Warb.

**Vernacular names** Malaysia: setumpol (Peninsular), karpus (Sabah). Burma (Myanmar): kalaw. Thailand: krabao. Vietnam: phong t[uw]r.

**Origin and geographic distribution** *Hydnocarpus* comprises about 40 species occurring in south-western India, Sri Lanka, Burma (Myanmar), Indo-China, southern China, Thailand (7 species), Peninsular Malaysia (12), Sumatra (12), Java (2), Borneo (17), the Philippines (5) and Sulawesi (2). *H. kurzii* (from Burma (Myanmar)), *H. alcalae* (from the Philippines) and *H. anthelmintica* (from Indo-China) used to be cultivated but with the advent of synthetic leprosy drugs in the 1960s, this cultivation has lost its importance.

**Uses** Seeds of many *Hydnocarpus* species (notably *H. anthelmintica* and *H. kurzii*) yield an oil that has been well known as a cure for leprosy and skin diseases since antiquity. The oil has also been recommended as a topical application to treat rheumatism, sprains and bruises, sciatica and chest complaints, and for dressing wounds. Major sources are *H. kurzii* from Burma (Myanmar), known as 'chaulmoogra' oil, and *H. laurifolia* (Dennst.) Sleumer (synonyms: *H. pentandrus* (Ham.) Oken, *H. wightiana* Blume) from south-western India, known as 'moratti' or 'marotti' oil. The major source of *Hydnocarpus* oil in China, where it is known as 'lukrabo' or 'krabao' oil, is *H. anthelmintica* seed from Indo-China. In Cambodia, this oil has also been used for illumination and it has been used to make soap. *H. venenata* Gaertner from Sri Lanka known as 'makulu' is used medicinally to treat leprosy but also as a fish poison. The seed of many *Hydnocarpus* species can be used as a fish poison, similar to *Pangium edule* Reinw. The oil from *H. kurzii* seed has been used to treat saddle-sores, and for liniment in veterinary practice.

The fibrous bark of *H. anthelmintica* is made into cordage, whereas the pulp of the fruits is edible. The wood of *Hydnocarpus* is used for local house building (poles), temporary heavy construction, posts, fences, interior finishing, panelling and door and window frames.

**Production and international trade** In for-

mer times seeds or seed oil of *Hydnocarpus* were traded from India and Indo-China to the Malasian region, China, Hawaii and Europe. However, at present no information on trade is available.

**Properties** Cyclopentenylglycine and cyclopentenyl fatty acids are found in *Hydnocarpus* seed. The Malasian species *H. alcalae*, *H. cauliflora* Merr., *H. subfalcata* Merr. and *H. woodii* Merr. contain high concentrations of the cyclopentenyl fatty acid chaulmoogric acid and of hydnocarpic acid, whereas the seeds of many other species contain glycosides which, upon hydrolysis, discharge the highly toxic hydrocyanic acid. The percentages of individual fatty acids for the seed oils of *H. kurzii* and *H. laurifolia* respectively have been found to be (in %): hydnocarpic acid 23.0 and 33.9, chaulmoogric acid 29.6 and 35.0, gorlic acid 25.1 and 12.8, lower cyclic homologues 0.3 and 4.6, myristic acid 0.6 and 0.8, palmitic acid 8.4 and 5.6, stearic acid zero and 0.6, palmitoleic acid 6.0 and 1.3, oleic acid 5.4 and 3.6 and linoleic acid 1.6 and 1.8. Very pure hydnocarpic acid, chaulmoogric acid and gorlic acid have been prepared from the seed oil of *H. laurifolia*. The isolated acids were used as starting materials to synthesize the corresponding cyclopentenyl alkylmethane sulphonates (mesylates), cyclopentenyl alkanes, cyclopentenyl nitriles, cyclopentenyl alcohols, 1-0-cyclopentenyl and 1,2-0-cyclopentenyl alkylglyceryl ethers. Pure homohydnocarpic acid and homochaulmoogric acid could be obtained, as well as pure hormelic acid. The purity of the fatty products obtained was assessed using chromatographic and spectroscopic techniques and their physicochemical constants were determined. These products may find uses as potential pheromones or as chemotherapeutics against certain mycobacteria.

The oil from *H. kurzii* is active against *Mycobacterium leprae*; best results have been obtained by administering ethyl esters of the fatty acids (ethyl chaulmoograte) in combination with sulphone drugs.

*Hydnocarpus* oil alone and mixed with dapsone fed to mice infected with *M. leprae* inhibited the growth of the leprosy bacilli. Dapsone and oil combined had an additive inhibitory effect on the growth of the bacilli.

Flavonolignans isolated from *H. laurifolia* seed, namely hydnowightin, hydnocarpin, and neohydnocarpin, have demonstrated potent hypolipidemic activity in mice, lowering both serum cholesterol and triglyceride levels. Hydnowightin demonstrated the best lipid-lowering effect of the three compounds. Good anti-inflammatory and

antineoplastic activity has been demonstrated for hydnocarpin in mice in vivo. The other two derivatives were not as active in these screens. All three compounds were moderately active against murine L-1210 leukaemia growth and demonstrated good activity against the growth of human KB nasopharynx, colon adenocarcinoma, osteosarcoma, and HeLa-S3 uterine growth. Hydnocarpin was the only compound of the three which was active against glioma growth. Hydnocarpin and neohydnocarpin demonstrated significant activity against Tmolt3 leukaemia cell growth.

In India, a pomace of *H. laurifolia* was found to be nematocidal. Aqueous extract showed greater nematocidal activity than the steam distillate. The nematocidal property is not adversely affected either by boiling or by change in pH (4-10).

A petroleum ether extract of seed of *H. laurifolia* at up to 1000 ppm was moderately active as an antifeedant against 4th-instar larvae of the noctuid *Spodoptera litura*. Positive activity was correlated with the percentage of linoleic acid and oleic acid in the seed oil.

**Description** Evergreen, dioecious or occasionally monoecious shrubs or small to medium-sized or rarely large trees up to 25(-50) m tall; bark surface usually smooth, sometimes cracking and scaly. Leaves alternate, simple, entire or serrate, variously asymmetrical at base; petiole thickened at apex; stipules early caducous. Flowers unisexual, 4-5-merous; sepals (3-)4-5, rarely 7-11, free or rarely slightly connate at base, imbricate; petals 4-5, rarely up to 14, with an in general densely pilose scale at base inside; male flowers in an axillary cyme or rarely in a raceme-like cauliflorous or ramiflorous panicle, with 5-many stamens; female flowers 1-3 together, with superior ovary, unilocular with many ovules, stigma sessile and with 3-5 spreading branches, often shortly bifid. Fruit an indehiscent, globose to obovoid drupe. Seeds closely packed, with membranous aril, endosperm albuminous-oily. Seedling with epigeal germination; cotyledons emergent or not, leafy; hypocotyl elongated; all leaves arranged spirally, conduplicate.

**Growth and development** Flowering in *Hydnocarpus* is usually once a year, but the period differs per region. Fruit takes rather long to develop, for instance about 7-8 months for *H. anthelmintica* and *H. woodii*. The fruits are probably dispersed by animals, but there are no reports of this.

**Other botanical information** There is widespread confusion on another *Flacourtiaceae*

species *Gynocardia odorata* R.Br. that is reputed to yield seed oil comparable to *Hydnocarpus*. Unlike real *Hydnocarpus* oil, this oil is neither optically active, nor does it have any therapeutic activity.

**Ecology** Most *Hydnocarpus* species are found scattered in primary rain forest, in well-drained, flat locations or on hillsides, on sandy or clayey soils, up to 1000(-1800) m altitude, occasionally in beach forest or on rocky outcrops. The medicinally important species prefer well-drained, sandy, alluvial flats and floodplains along rivers, or at least moist but well-drained surroundings.

**Propagation and planting** *Hydnocarpus* is usually propagated by seed. Seeds are separated from the fruit pulp by washing. Seed of *H. kunstleri* (King) Warb. germinated for 50% in 4-8 months and that of *H. woodii* for about 50% in 5 months to over 2 years. Under natural conditions, seed germinates during the rains shortly after falling to the ground. Seedlings and saplings should be grown under shade.

**Harvesting** Traditionally, fallen fruits of *Hydnocarpus* were simply collected in the forest, giving often rise to a mixture of species being collected, as several *Hydnocarpus* contain chaulmoogric acid and hydnocarpic acid. The result was a very variable raw product. Accessibility of the floodplains at the time of harvesting was often difficult.

**Handling after harvest** Seeds tend to go rancid rather quickly. Therefore ripe fruits should be opened and the fruit pulp and aril removed from the seeds. Seeds are then washed and dried in the sun. Sun-dried seeds, either whole or broken, are cold-pressed or pressed under concurrent heating to extract the oil. The chemical composition of the end-product of these methods varies.

**Genetic resources and breeding** With the exception of *H. heterophylla* Blume, *H. kunstleri* and *H. woodii*, most *Hydnocarpus* have a limited geographical distribution. Together with the often low density at which species are present, this means there is a serious risk of genetic erosion. Because of their importance as leprosy drugs, many *Hydnocarpus* species were planted in botanical gardens in the Malesian region, South America and Central Africa, and some of them may still be present.

**Prospects** The use of seed oil of *Hydnocarpus* to treat leprosy has been replaced by synthetic drugs. However, with the renewed interest in medicines of plant origin, investigations of the oils and extracts of *Hydnocarpus* seem worthwhile, es-

pecially for the treatment of various skin diseases. The cyclopentenyl fatty acids and their synthesized derivatives are of interest as antibacterial compounds, whereas the flavonolignans may have potential as anti-inflammatory and anti-cancer agents.

**Literature** |1| Abdel-Moety, E.M., 1989. New products from lipids, cyclopentenyl fatty acids as starting materials. *Grasas Aceites* 40(1): 15-21. |2| Burkill, I.H., 1966. A dictionary of the economic products of the Malay Peninsula. 2nd edition. Vol. 1. Ministry of Agriculture and Co-operatives, Kuala Lumpur, Malaysia. pp. 1224-1229. |3| Kumar, B.H. & Thakur, S.S., 1988. Certain non-edible seed oils as feeding deterrents against *Spodoptera litura* Fb. *Journal of the Oil Technologists' Association of India* 20(3): 63-65. |4| Ong, H.C., 1998. *Hydnocarpus* Gaertn. In: Sosef, M.S.M., Hong, L.T. & Prawirohatmodjo, S. (Editors): *Plant Resources of South-East Asia No 5(3)*. Lesser-known timbers. Backhuys Publishers, Leiden, the Netherlands. pp. 296-298. |5| Pillai, S.N. & Desai, M.V., 1975. Antihelminthic property of 'Marotti' cake (*Hydnocarpus laurifolia*). *Pesticides (India)* 9(4): 37-39. |6| Sengupta, A., Gupta, J.K., Dutta, J. & Ghosh, A., 1973. The component fatty acids of chaulmoogra oil. *Journal of the Science of Food and Agriculture* 24(6): 669-674. |7| Sharma, D.K. & Hall, I.H., 1991. Hypolipidemic, anti-inflammatory, and antineoplastic activity and cytotoxicity of flavonolignans isolated from *Hydnocarpus wightiana* seeds. *Journal of Natural Products* 54(5): 1298-1302. |8| Sleumer, H., 1938. *Monographie der Gattung Hydnocarpus Gaertner* [Monograph of the genus *Hydnocarpus* Gaertner]. *Botanische Jahrbucher* 69: 1-94. |9| Sleumer, H., 1985. The Flacourtiaceae of Thailand. *Blumea* 30: 217-250. |10| Quisumbing, E., 1978. *Medicinal plants of the Philippines*. Katha publishing Co., Quezon City, the Philippines. pp. 628-629.

#### *Selection of species*

#### **Hydnocarpus alcalae C.DC.**

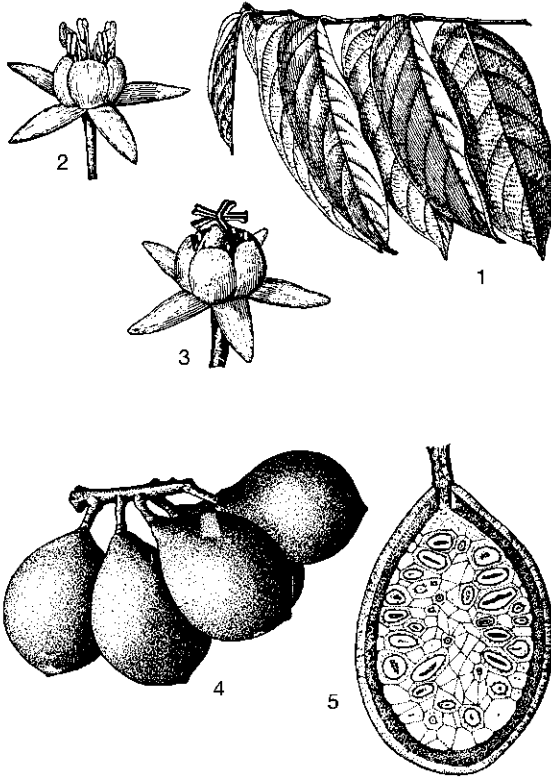
*Philipp. Journ. Sci., Bot.* 11: 37 (1916).

**Vernacular names** Philippines: dudoa (Bikol).

**Distribution** Philippines (southern Luzon), but planted and naturalized elsewhere in Malesia.

**Uses** The seed oil has been extensively used in the treatment of leprosy.

**Observations** A small tree or shrub, 4-7 m tall, branches glabrous; leaves ovate-oblong, 15-25 cm x 7-11 cm, subcoriaceous, base very asymmetrical,



*Hydnocarpus alcalae* C. DC. - 1, leafy twig; 2, male flower; 3, female flower; 4, fruits; 5, fruit in longitudinal section.

apex acuminate; inflorescence a pseudo-racemose panicle, 15-30(-60) cm long, male inflorescences in fascicles of 3-4, female inflorescences solitary, cauliflorous or ramiflorous; flowers 5-merous, greenish white or cream; fruit obovoid, pendent, 15-31 cm × 8-15(-20) cm, dark brown with purple and greenish spots, exocarp 0.3-0.7 mm thick, mesocarp stony, 6 mm thick, endocarp soft, 7 mm thick, with 80-110 seeds; seeds subglobular or ellipsoidal, 3-5 cm × 2.5-3 cm, with a pale yellow aril, embedded in a fragrant, astringent pulp. *H. alcalae* occurs naturally in moist ravines.

**Selected sources** 190, 332, 1126, 1178, 1376, 1377.

### *Hydnocarpus anthelmintica* Pierre ex Lanessan

Pl. util. colon. franç.: 303 (1896).

**Synonyms** *Hydnocarpus alpina* Wight var. *elongata* Boerl. (1899), *Hydnocarpus alpina* Wight var. *macrocarpa* Boerl. (1899).

**Vernacular names** Sausage tree (En). Chaul-

moogra (Fr). Burma (Myanmar): kalaw-wa. Cambodia: krabao (Kompong Thom), krabaou thom (Pursat), krabaou phlae thom (general). Laos: ka bao. Thailand: krabao (general), krabao yai (central), bao (peninsular). Vietnam: ch[uf]m bao l[ows]n, d[aj]i phong t[luwr], l[oj] n[oof]i.

**Distribution** Burma (Myanmar), Indo-China and Thailand; also cultivated within this region for its medicinal seeds.

**Uses** The seeds are used as a vermifuge in Thailand and Vietnam. However, the use of the seed oil in the treatment of leprosy and other skin complaints is more important. This species is reported to be the source of almost all *Hydnocarpus* seed used in China.

**Observations** A dioecious, small to medium-sized tree, 10-20(-30) m tall, outer bark lenticellate, greyish-black, inner bark brown; leaves ovate-lanceolate to ovate-oblong, 10-33 cm × 3-7 cm, coriaceous, glabrous, base cuneate to rounded-obtuse, usually asymmetrical, apex gradually long attenuate; inflorescence a 2-3-branched cyme, few-flowered; flowers 5-merous, white or greenish, scented, female flowers with 5 fusiform staminoes; fruit globose, 8-12 cm in diameter, brownish tomentulose, exocarp 0.2 mm thick, mesocarp yellowish, very hard; seeds 30-50(-100) per fruit, ovoid, compressed, 1.5-1.8 cm × 1-1.5 cm. *H. anthelmintica* occurs in evergreen forest and is locally common along river banks, at elevations up to 1300 m. It may well be grown in the Malesian region.

**Selected sources** 202, 260, 293, 294, 364, 843, 900, 1035, 1126, 1128, 1287, 1376, 1378, 1392.

### *Hydnocarpus kurzii* (King) Warb.

Engl. & Prantl, Nat. Pflanzenfam. 3, 6a: 21 (1893).

**Synonyms** *Taraktogenos kurzii* auct. non King s.s.

**Vernacular names** Malaysia: kulau (Peninsular). Burma (Myanmar): kalaw. Thailand: ngaa yoi, ma duuk.

**Distribution** India, Burma (Myanmar), Thailand and Peninsular Malaysia.

**Uses** The oil extracted from the seeds is used in the treatment of leprosy. It is the major source of chaulmoogric acid. It is advised not to eat pigs nor fish that have been feeding on the seeds, as their flesh induces nausea and vomiting. A decoction of the bark is drunk for internal disorders and skin diseases in Burma (Myanmar). The roots as well as the seeds are used by Thai traditional doctors for the treatment of skin diseases.



**Observations** A dioecious tree, 8–20(–30) m tall, bark smooth, grey, young branchlets fulvous pubescent, rather soon glabrescent; leaves lanceolate-oblong, entire, 15–22(–32) cm × 4–6.5 cm, subcoriaceous, glabrous, base cuneate, symmetrical, apex subacuminate; inflorescence a short pedunculate cyme 5–7(–9)-flowered; flowers whitish with unpleasant odour, male flowers with 4 sepals and 8 petals, stamens (15–)18–25(–30), female flowers with 10–16 staminodes; fruit globose, (5–)8–10 cm in diameter, rugose, exocarp fibrose-cancellate, 3–4(–6) mm thick; seeds angular-ovoid, 3 cm × 1.5 cm. Two subspecies have been distinguished: subsp. *australis* Sleum. occurring in southern Burma (Myanmar), Thailand and Peninsular Malaysia, and subsp. *kurzii* from India and northern Burma (Myanmar). Subsp. *australis* differs from subsp. *kurzii* in the thicker exocarp and the venation of the leaves. *H. kurzii* occurs scattered in evergreen forest at 600–1800 m altitude.

**Selected sources** 202, 260, 580, 843, 900, 1126, 1308, 1376, 1377, 1378, 1564.

Khozirah Shaari & L.S.L. Chua

## Hypericum L.

Sp. pl. 2: 783 (1753); Gen. pl. ed. 5: 341 (1754).

GUTIFERAE

$x = (6), 7, 8, 9, 10, 11, 12$

**Major species** *Hypericum japonicum* Thunb. ex Murray, *H. monogynum* L.

**Vernacular names** St John's wort (En). Millepertuis (Fr).

**Origin and geographic distribution** *Hypericum* comprises about 400 species and can be considered cosmopolitan, occurring in all continents except for arctic and desert areas and most of the lowland tropics. There are 15 species of *Hypericum* in Malesia, but the genus is absent from the Moluccas. The genus is also present in Indo-China and Thailand and rare in Australia, with only 2 species.

**Uses** All *Hypericum* species are said to yield essential oils and to be medicinally interesting to some extent. Their medicinal use has been basically the same throughout the world, as they have been used internally against rheumatism, jaundice, oedema and gastric catarrh, and externally to treat wounds and bruises. These effects of *Hypericum* species are regarded as based on their astringent, diuretic, and haemostatic activities. Though there are 15 Malesian species, specific re-

ports of medicinal use are only found for *H. gramineum*, *H. japonicum*, *H. monogynum*, *H. papuanum* and *H. uralum*.

Reports of medicinal use also exist for the non-Malesian *H. ascyron* L., *H. patulum* Thunb. ex Murray and *H. sampsonii* Hance. In Indo-China, the seeds of *H. patulum* are applied to treat dog bites and bee stings. *H. sampsonii* is reported to be used as a vulnerary in northern Vietnam. The fruits and seeds of *H. ascyron* are applied to treat skin problems, gonorrhoea and dysmenorrhoea in Indo-China and China. In Chinese medicine, *H. ascyron*, *H. japonicum* and *H. sampsonii* are mentioned as antihaemorrhagic, and the latter two as vulneraries in the treatment of scrofula and contusions.

A well-known medicinal species in Europe is *H. perforatum* L., which does not occur in South-East Asia. *H. perforatum* was already recognized as a medicinal plant by the ancient Greek writers Dioscorides and Hippocrates. Nowadays, drugs based on the plant are used to treat skin and mouth problems and as an antidepressant.

*Hypericum* species are often grown as ornamentals.

**Properties** The most characteristic compounds of *Hypericum* are xanthonoid pigments and anthraquinonoids (naphthodianthrone, hydroxyanthraquinones), such as hypericin and pseudohypericin. Also present is an essential oil containing considerable amounts of aliphatic compounds such as 2-methyloctane, nonane, undecane, octanal and decanal. Other constituents include some common phenolic components (catechins, leucoanthocyanins, chlorogenic acids, condensed tannins) and flavonoids (quercetin glycosides: rutin, quercitrin, hyperin).

Hypericin and pseudohypericin are present in *H. perforatum* and many other species, but are not found in the South-East Asian *H. ascyron*, *H. japonicum* and *H. monogynum*, while results are not clear for *H. patulum*. The pharmacology of hypericin and pseudohypericin has been well investigated. Strong antiviral properties against retroviruses, such as the HIV-1, influenza and cytomegalo viruses has been shown, with no toxic side-effects at therapeutic doses. In this respect, hypericin seems to be more potent than pseudohypericin and clinical trials have been initiated on this compound. At molecular level, hypericin and pseudohypericin specifically inhibit protein kinase C (PKC), having IC<sub>50</sub> levels of respectively 1.7 µg/ml and 15 µg/ml. Thus, the anti-retroviral activity could be attributable to phosphorylation re-

actions being inhibited by the PKC occurring during viral infection of cells. In addition to the antiviral activity, antiproliferative activity against mammalian cells (possibly due to PKC inhibition) has been reported in the literature.

The ingestion of hypericin or pseudohypericin (or their biosynthetic precursors proto-hypericin/proto-pseudohypericin) and subsequent exposure to UV light ( $\lambda$  320 nm) may cause photodermatitis. Therefore, exposure to the sun must be avoided after using *Hypericum*-based drugs. In animals, this syndrome is called 'hypericism'. Symptoms are swelling of the face, itches, loss of hair, appearance of sores and eventually apoplexy and death.

In Europe, extracts of *H. perforatum* are well known as an antidepressant. In several clinical trials of mild and moderate depressions they have been reported to be as effective as standard medications (e.g. using tricyclic antidepressants), but with far fewer side-effects. An antidepressant activity of *H. patulum* has also been reported in the literature. It remains unclear whether the antidepressant activity is related to the content of hypericin/pseudohypericin. Experiments with purified substances have not given unambiguous results, and tentative investigations of the extracts indicate that other compounds (e.g. flavones) might also play a role.

Several antimicrobial phloroglucinol derivatives (saroaspidin A, B and C, sarothralin A, B, C and D, sarothralin, and sarothralin G) have been isolated from *H. japonicum*. Of these compounds, sarothralin and sarothralin G have the strongest activity against gram-positive bacteria e.g. *Staphylococcus aureus*, *Bacillus cereus*, and *Nocardia gardneri*. In addition to the phloroglucinols, lactones and flavonoids have also been isolated. Furthermore, the phloroglucinol derivative japonicine A (from *H. japonicum*) is reported to show antimalarial activity in mice. Acetone extracts of the leaves of *H. papuanum* showed antibacterial activity against *Staphylococcus aureus*.

Aqueous plant extracts of *H. japonicum* (10 mg/ml) have been found to have strong in vitro antiviral activity against the herpes simplex virus HSV-II, whether administered simultaneously (simultaneous addition of extract and virus to the cell bottle) or therapeutically (virus inoculated into the cell bottle, later followed by addition of the extract). Methanolic plant extracts of *H. uralum* have shown antiviral activity against 3 mammalian viruses: herpes simplex, Sindbis and polio. The activity was not enhanced by UV light, which suggests that antiviral compounds other than the

naphthodianthrones (hypericin, pseudohypericin) are present.

**Description** Herbs or shrubs, sometimes small trees, usually glabrous; branchlets terete or 2-4-lined or -angled. Leaves opposite or rarely whorled, simple and entire, with translucent glands and/or black or red glands, sessile or shortly petiolate. Inflorescence a terminal dichasium or monochasium. Flowers bisexual, (4-)5-merous; sepals quincuncial or rarely decussate, coriaceous to chartaceous, glandular; petals yellow to orange, sometimes tinged or veined red, glandular, glabrous, persistent or deciduous; stamens in epipetalous fascicles, free or variously united, glabrous, persistent or deciduous, each fascicle with up to 70 stamens, filaments yellow, slender, anthers short, oblong, yellow or reddish, dorsifixed or apparently basifixed; ovary superior, 3-5-celled or 1-celled with (2-)3-5 parietal or axile placentas, ovules 2-many on each placenta, styles (2-)3-5, free to united, slender, stigmas small. Fruit a (2-)3-5-valved capsule. Seeds 1-many on each placenta, curved cylindrical to ellipsoid, sometimes carinate or winged; embryo cylindrical, straight or curved, with cotyledons usually shorter than hypocotyl. Seedling with epigeal germination; cotyledons leafy, sessile, glabrous; hypocotyl short, epicotyl very short.

**Growth and development** *Hypericum* includes species with very different habits, from herbs to small trees. The arrangement of stamens in the flowers, with the innermost shorter and not attaining the level of the stigmas favours cross-pollination. The flowers are typically visited by less-specialized insects such as *Diptera*. Usually seeds are shed by septical dehiscence of the capsule. Seed from *Hypericum* growing in damp or marshy locations may possibly adhere to the feet and feathers of wading birds and waterfowl and be dispersed thus.

**Other botanical information** *Hypericum* is closely related to *Cratoxylum* (from a different tribe). The tribes *Vismieae*, *Cratoxyleae* and *Hypericeae* together constitute the subfamily *Hypericoideae*. As this subfamily forms a natural group, in the past it was often given family ranking (*Hypericaceae*), but nowadays it is usually classified within the large family *Guttiferae* (*Clusiaceae*). *Hypericum* has been further divided into no fewer than 30 sections.

**Ecology** In the tropics, *Hypericum* is generally a high-altitude genus, though some species are sometimes found at low elevations. In temperate regions, it is found in widely varying conditions,

but never in very arid habitats. In South-East Asia, *Hypericum* is found in forest margins, grassland, marshes or among rocks. It occurs from sea-level in Sumatra up to 3400 m in New Guinea, but is rarely found in the lowlands.

**Propagation and planting** *Hypericum* can be propagated by seed, cuttings, division or suckers. *H. monogynum* is propagated by division. Light is essential for germination of seed.

**Handling after harvest** Whole plants can be used fresh or dried.

**Genetic resources and breeding** Wild *Hypericum* hybrids have been found, but only between closely related species. Artificial hybrids are always sterile and usually weak, though some crosses have thrived.

**Prospects** *Hypericum* extracts have shown strong antiviral, antidepressant and antimicrobial activity, with limited toxic side-effects. Because of these promising medicinal properties, *Hypericum* may become important in the future. However, chemical and medicinal properties vary between species and there is very little information on the presence of active compounds in South-East Asian *Hypericum* species.

**Literature** |1| Baureithel, K.H., Buter, K.B., Engesser, A., Burkard, W. & Schaffner, W., 1997. Inhibition of benzodiazepine binding in vitro by amentoflavone, a constituent of various species of *Hypericum*. *Pharmaceutical Acta Helvetica* 72(3): 153–157. |2| Hegnauer, R., 1966. *Chemotaxonomie der Pflanzen: eine Übersicht über die Verbreitung und die systematische Bedeutung der Pflanzenstoffe* [Chemotaxonomy of plants: an overview of the distribution and the systematic importance of plant substances]. Vol. 4. Birkhäuser Verlag, Basel, Switzerland. pp. 223–226. |3| Ishiguro, K., Nagata, S., Fukumoto, H., Yamaki, M. & Isoi, K., 1994. Phloroglucinol derivatives from *Hypericum japonicum*. *Phytochemistry* 35(2): 469–471. |4| Ishiguro, K., Yamaki, M., Kashihara, M., Takagi, S. & Isoi, K., 1990. Sarothralin G: a new antimicrobial compound from *Hypericum japonicum*. *Planta Medica* 56(3): 274–276. |5| Mathis, C. & Ourisson, G., 1963. Étude chimio-taxonomique du genre *Hypericum*. I. Répartition de l'hypericine [Chemo-taxonomical study of the genus *Hypericum*. I. Distribution of hypericine]. *Phytochemistry* 2: 157–171. |6| Meruelo, D., Lavie, G. & Lavie, D., 1988. Therapeutic agents with dramatic antiretroviral activity and little toxicity at effective doses: aromatic polycyclic diones hypericin and pseudohypericin. *Proceedings of the National Academy of Science, United States* 85(14):

5230–5234. |7| Robson, N.K.B., 1974. *Hypericum*. In: van Steenis, C.G.G.J. (General editor): *Flora Malesiana*. Series 1, Vol. 8. Sijthoff & Noordhoff International Publishers, Alphen aan den Rijn, the Netherlands. pp. 14–29. |8| Robson, N.K.B., 1980. *Hypericum* L. In: Townsend, C.C. & Guest, E.R. (Editors): *Flora of Iraq*. Vol. 4. Ministry of Agriculture and Agrarian Reform, Bagdad, Iraq. pp. 363–381. |9| Robson, N.K.B., 1996. *Hypericum*. In: Huang, T.-C. (Editor): *Flora of Taiwan*, 2nd edition. Vol. 2. Editorial Committee of the Flora of Taiwan, Taipei, Taiwan, Republic of China. pp. 698–714. |10| Taylor, R.S.L., Manandhar, N.P., Hudson, J.B. & Towers, G.H.N., 1996. Antiviral activities of Nepalese medicinal plants. *Journal of Ethnopharmacology* 52(3): 157–163.

#### *Selection of species*

#### ***Hypericum gramineum* G. Forster**

Fl. ins. austr. prodr.: 53 (1786).

**Synonyms** *Hypericum involutum* (Labill.) Choisy (1821).

**Distribution** From India, Vietnam, Taiwan, and China (Yunnan), to Australia, New Zealand and New Caledonia. In Malesia only in New Guinea.

**Uses** In Papua New Guinea, Simbu province, the leaves are chewed with traditional ash salt daily to treat malarial fever.

**Observations** Perennial or annual herb, (3–)5–72 cm tall, with erect or decumbent stems, branching strictly from the base or unbranched, not rooting; leaves usually lanceolate to linear or oblong, 4–25 mm × 1–8 mm, with 1–3 basal veins, without reticulate venation, laminar glands pale, sessile; inflorescence lax, 1–30-flowered; flowers (5–)6–12(–15) mm in diameter, sepals 2.8–7.5(–9) mm × 0.8–2 mm, free, 3–5 veined, often with prominent midrib, laminar glands pale, petals 5–10 mm × 2–5 mm, persistent, pale yellow to orange, without glands, stamens 30–50, ovary 1-celled, placentas 3, parietal, styles divergent; fruit narrowly ovoid to cylindrical; seeds minute, cylindrical, longitudinally ribbed with striae, not carinate. *H. gramineum* is found in humid to dry localities, but always in open and well-drained habitats, from sea-level to 2600 m altitude.

**Selected sources** 600, 1238, 1243.

**Hypericum japonicum Thunb. ex Murray**

Syst. veg. ed. 14: 702 (1784).

**Synonyms** *Hypericum pusillum* Choisy (1821), *Hypericum mutilum* Maxim. (1881) non L., *Sarothra japonica* (Thunb. ex Murray) Y. Kimura (1951).

**Vernacular names** Papua New Guinea: ngotokong (Mt Hagen, Western Highlands). Vietnam: n[ojc] s[owr]i c[or] ban, di[eef]n c[ow] ho[af]ng.

**Distribution** From Japan, South Korea and south-eastern China, Nepal, India and Sri Lanka to Australia, New-Zealand and Hawaii. *H. japonicum* occurs throughout Malesia, Indo-China and Thailand.

**Uses** Generally, *H. japonicum* is thought to have astringent and alterative action, and it is used externally to treat swellings, abscesses, scrofula and fungal skin diseases. In Malaysia, it is applied externally to treat wounds. In Papua New Guinea, crushed plants are reported to be used internally against malaria, together with ginger and

ash salt. In Vietnam, *H. japonicum* is used internally as stomachic and externally as vulnerary on wounds, leech and snake bites, and to treat caries and bad breath. In Chinese medicine, it is applied as vulnerary on wounds and leech bites, and to treat bacterial diseases, hepatitis and tumours.

**Observations** An extremely variable annual herb, 2–50 cm tall, with erect to decumbent or prostrate stems rooting at the base; leaves usually ovate or ovate-triangular to oblong or elliptical, 2–18 mm × 1–10 mm, chartaceous, lower side sometimes glaucous, with 1–7 basal veins, usually without reticulate venation, laminar glands pale, sessile; inflorescence lax, 1–30-flowered; flowers 4–8 mm in diameter, sepals 2–5.5 mm × 0.5–2 mm, free, 3–5-veined, often with prominent midrib, laminar glands pale, petals 1.7–5 mm × 0.8–1.8 mm, persistent, pale yellow to orange, without glands, stamens 5–30, in 5 irregular groups, ovary 1-celled, placentas 2–3, parietal, styles divergent, broadening to capitate stigmas; fruit cylindrical to globose, 2–6 mm long; seeds minute, cylindrical, longitudinally ribbed with striae, not carinate. *H. japonicum* is found in wet or marshy to dry localities, but always in exposed places, from sea-level up to 3400 m altitude.

**Selected sources** 97, 202, 456, 597, 655, 656, 657, 658, 1035, 1126, 1128, 1238, 1239, 1240, 1241, 1242, 1243, 1476, 1513, 1655.

**Hypericum monogynum L.**

Sp. pl. ed. 2: 1107 (1763).

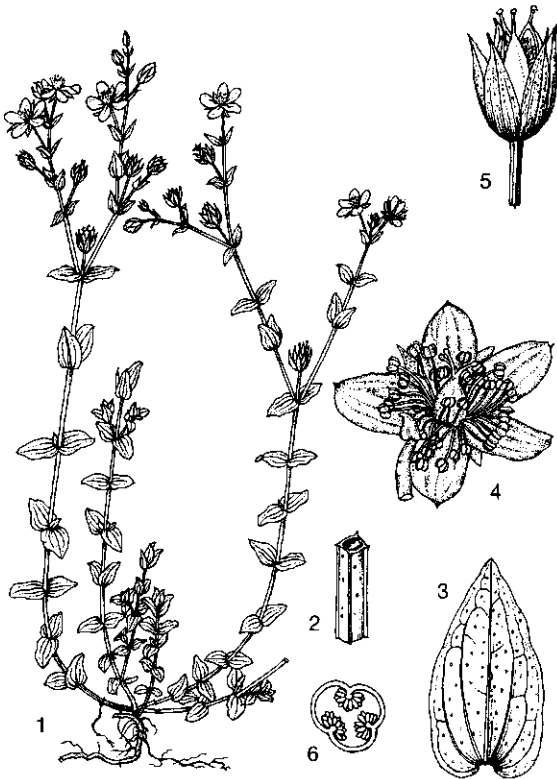
**Synonyms** *Hypericum chinense* L. (1759) non Osbeck.

**Vernacular names** Vietnam: ban.

**Distribution** Native to south-eastern China and Taiwan, now cultivated in many parts of the world, including Java and Sulawesi.

**Uses** In Indo-China, the green twigs and leaves are made into a paste, which is used to treat dog bites and bee stings. In India, *H. monogynum* is considered astringent and alterative and it is used to treat diarrhoea and vomiting.

**Observations** A bushy shrub, 50–130 cm tall, with spreading branches; leaves elliptical or oblong to oblanceolate, 2–4.5 cm × 1–1.8 cm, lower side paler, with 4–6 main lateral veins and dense reticulate venation, glands pale, sessile or petiole; inflorescence corymbose, 1–15-flowered; flowers 3–5 cm in diameter, sepals 4.5–10 mm × 1.5–3 mm, free, glands all pale, petals 20–30 mm × 12.5–15 mm, caducous, golden yellow to lemon yellow, glands pale, stamens in 5 fascicles, each with 25–35 stamens, caducous, ovary 5-celled, pla-



*Hypericum japonicum* Thunb. ex Murray - 1, plant habit; 2, detail of stem; 3, leaf; 4, flower; 5, dehiscent capsule; 6, ovary in cross-section.

centas 5, axile, styles 5, united almost to the apex, stigmas small; fruit broadly ovoid or ovoid-conical to subglobose, 6–10 mm long; seeds minute, cylindrical, curved, narrowly carinate, shallowly linear-reticulate to linear-foveolate. In Malesia, *H. monogynum* is only found as a cultivated ornamental, up to 1800 m altitude.

**Selected sources** 287, 1128, 1238, 1239, 1240, 1241.

### **Hypericum papuanum** Ridley

Trans. Linn. Soc. II, Bot. 9: 19 (1916).

**Synonyms** *Hypericum helwigii* Laut. (1922), *Hypericum habbemense* A.C. Smith (1941).

**Vernacular names** Papua New Guinea: enaime (Tauade, Central Province).

**Distribution** New Guinea, where it is widespread in the mountainous regions.

**Uses** The leaves are reported to be used in New Guinea to treat sores.

**Observations** A very variable woody herb or shrub, 10–130 cm tall, branches often creeping and rooting at the base; leaves narrowly ovate to elliptical or subcircular, 0.6–2.5 cm × 0.3–1.7 cm, concolorous, with 4–6 main lateral veins and reticulate venation, laminar glands pale, intramarginal glands pale or (partly) black, sessile; inflorescence 1-flowered; flowers 1.8–2 cm in diameter, sepals 3–8 mm × 1–3.5 mm, free, foliaceous, laminar glands pale, submarginal glands black or absent, petals 9–15 mm × 4–9 mm, persistent, bright yellow, laminar glands pale, marginal glands black or absent, stamens (15–)25–40(–50), not clearly in fascicles, persistent, ovary ovoid, placentas 3(–5), parietal, styles 3(–5), divergent, stigmas capitate; fruit ovoid to ellipsoid, 5–10 mm long, longitudinally striped; seeds minute, cylindrical or subcylindrical, scarcely carinate, densely linear-foveolate to scalariform. *H. papuanum* is found in wet to dry alpine grasslands, bogs, and screes at 1800–3800 m altitude.

**Selected sources** 597, 827, 1238, 1239, 1240, 1241.

### **Hypericum uralum** Buch.-Ham. ex D. Don

Bot. Mag.: t. 2375 (1823).

**Synonyms** *Norysca urala* (Buch.-Ham. ex D. Don) K. Koch (1853), *Hypericum patulum* Thunb. var. *uralum* (Buch.-Ham. ex D. Don) Koehne (1893).

**Distribution** Nepal, northern India, Burma (Myanmar), China, Thailand and northern Sumatra.

**Uses** No reports have been found of medicinal use in South-East Asia, but in Nepal the root juice is used against fever.

**Observations** A shrub, 120–180 cm tall, branches arching, sometimes frondose; leaves lanceolate to ovate, 1–4.5 cm × 0.5–2.5 cm, lower side very glaucous, with 3 main lateral veins and pale glands, subsessile or petiolate; inflorescence corymbose, racemiform, 1–3(–10)-flowered; flowers 1.5–3 cm in diameter, sepals 3.5–9 mm × (1–)2–6.5 mm, free, glands pale, petals 9–18 mm × 5–12 mm, caducous, bright yellow to golden yellow, glands pale, stamens in 5 fascicles, each with 40–60 stamens, caducous, ovary 5-celled, placentas 5, axile, styles 5, free, stigmas narrowly capitate; fruit subglobose to globose, 7–11 mm long; seeds minute, cylindrical-ellipsoid, not curved, slightly carinate, shallowly linear-reticulate. The Sumatran form tends to be more luxuriant and to have larger flowers than plants from elsewhere. *H. uralum* is found in grassy or rocky slopes, in pastures, thickets, open woodland and montane forest at 1700–3300 m altitude.

**Selected sources** 1238, 1239, 1240, 1241, 1442.

M. Brink

## **Imperata** Cirillo

Pl. rar. neap. 2: 26, t. 11 (1792).

GRAMINEAE

$x = 5$ ; *I. cylindrica*:  $2n = 20$

**Major species** *Imperata conferta* (J.S. Presl) Ohwi, *I. cylindrica* (L.) Raeuschel.

**Vernacular names** Cogon grass (En). Indonesia: alang-alang. Philippines: kogon.

**Origin and geographic distribution** *Imperata* comprises about 8 species, occurring in tropical and subtropical regions. *I. conferta* and *I. cylindrica* are found throughout the Malesian region.

**Uses** The uses (including medicinal) of *I. cylindrica* and *I. conferta* are very similar and both are in general used indiscriminately. *I. cylindrica* is reported to be used in various countries in South-East Asia for a wide range of medical problems such as fever, nausea, dropsy, jaundice, asthma, haematuria, influenza, internal haemorrhages, nose bleeding, cough and kidney diseases. A decoction of the rhizome is commonly applied to purify the blood, as a diuretic and in the treatment of diarrhoea and dysentery.

Both *Imperata* species are an important source of roofing material, readily available and durable. When young, they may be used as fodder for rumi-

nants. Both species have been used for making paper. The aggressive rhizomatous growth makes them suitable for erosion control and stabilizing slopes, despite their weedy characteristics.

**Production and international trade** *Imperata* is used only locally and has no importance in international markets.

**Properties** The rhizomes of *I. cylindrica* contain the biphenyl ethers cylindol A and B, the phenolic compound imperanene, the sesquiterpenoid cylindrene, and the lignans graminone A and B. Cylindol A has shown 5-lipoxygenase activity, which may be partly involved in the reputed anti-inflammatory activity of *I. cylindrica*. Imperanene has shown inhibitory activity against rabbit platelet aggregation. Cylindrene and graminone B have shown inhibitory effects on rabbit aorta contraction. *I. cylindrica* has also been found to contain 5-hydroxytryptamine (serotonin) and the triterpenoids arundoin, cylindrin, fernenol, isoarborinol and aimiarenol. The rhizome has been found to contain 19% sugars (saccharose, dextrose, fructose, xylose) and various acids (including malic, citric, tartaric, chlorogenic, coumaric and oxalic acid). An aqueous extract of stem and leaves is reported to have shown some antitumour activity against sarcoma 180 and adenocarcinoma 75 in mice. Although *I. cylindrica* is widely considered to have diuretic properties, this could not be confirmed in a recent double blind study. Besides the above pharmacological activities, antiviral, antihepatotoxic, antihistamine and larvicidal activities are also reported.

**Description** Rhizomatous, herbaceous, unbranched perennials up to 150(–300) cm tall, with deep, branched, fleshy rhizomes; culms with solid internodes. Leaves mostly basal, leaf sheath non-auriculate, with a fringed membranous ligule; leaves sometimes pseudopetiolate, without cross venation, persistent. Inflorescence a spiciform panicle, cylindrical; spikelets paired, homologous, bisexual, 3–6 mm long, 2-flowered, at the base with a dense whorl of silky white hairs, unequally pedicellate in each pair; glumes 2, more or less equal; lower floret reduced to a hyaline lemma; upper floret fertile, stamens 1–2, ovary glabrous. Fruit an ellipsoidal caryopsis, about 1 mm long, brown.

**Growth and development** *I. cylindrica* is a prolific seed producer and the light seeds are readily dispersed by wind. There is no dormancy and seeds may give 95% germination within one week after fruit maturation. The optimum temperature for growth is about 30°C with negligible growth at 20°C and 40°C. Some individual plants flower fre-

quently, some never flower, and others are intermediate. The rhizomes are highly competitive and penetrate the roots of other plants, causing rot or death. In a controlled greenhouse experiment young plants initiated new rhizomes between the third and fourth leaf-stages. Rhizome growth is determinate, with the apical bud forming a shoot and sub-apical buds forming rhizome branches. Under favourable conditions apical and sub-apical buds develop simultaneously, but under stress the growth of the apical bud is favoured. Roots and buds develop at the distal nodes of the young rhizome long after the rhizome has been formed. Flowering is in general promoted by burning, and follows a few weeks after the fire.

**Other botanical information** *Imperata* is divided into two sections, based on the number of stamens: section *Imperata* (2 stamens) with only 1 species (*I. cylindrica*), and section *Eriopogon* Endl. (1 stamen). A number of varieties have been described for *I. cylindrica*, often pertaining to geographical regions. However, as these varieties more or less intergrade, the distinction is at present generally ignored.

**Ecology** *I. cylindrica* is often found in areas receiving more than 1000 mm rainfall/year, but has been recorded in sites receiving 500–5000 mm annual rainfall. It can withstand waterlogging but not continuous flooding. It grows at altitudes from sea-level up to 2000 m in several countries and has been recorded at 2700 m in Indonesia. Whereas *I. cylindrica* may have originally been restricted to infertile and acid soils in the tropics, it has become widespread through man's intervention, particularly following slashing and burning of forest lands. Its resistance to burning is associated with its vigorous underground rhizomes, but seedlings also establish after burning. It is found in a wide range of habitats, including the dry sand dunes of seashores and deserts, as well as swamps and river valleys. It grows in grassland, cultivated areas, and plantations. It quickly invades abandoned farmland and occurs on railway and highway embankments and in deforested areas. It is regarded as a light-loving plant and will not persist under heavy shade in plantations. Although it grows in a wide range of soil types of widely differing fertility, it grows most vigorously in wet soil of reasonable fertility. It has been reported to grow on soils with pH 4.0–7.5. It can even tolerate very hot, steamy and sulphurous conditions near an active volcanic fumarole or vent. *I. cylindrica* is reported to have allelopathic properties that adversely affect the growth of other plants.

**Propagation and planting** *Imperata* is seldom propagated deliberately, but spreads by rhizomes and seed. Rhizomes accidentally cut by cultivation can reestablish from pieces with as few as 2 nodes.

**Husbandry** *Imperata* is favoured by burning, which can lead to virtually monospecific swards. If it is to be used for thatching, swards are left ungrazed until after the mature growth has been removed. It can be eliminated by heavy continuous grazing; vigour will be reduced by close, frequent cutting and rhizomes can be destroyed by frequent intensive cultivation. It has been suggested that *I. cylindrica* that is used regularly for grazing should be grazed rotationally when 15–25 cm high. Because of its aggressiveness and low quality as forage, *I. cylindrica* is widely regarded as a weed. It is considered to be one of the most pernicious weeds in South-East Asia. In Indonesia, it is considered to be one of the three most important weeds (the others being *Echinochloa crus-galli* (L.) P. Beauv. and *Monochoria vaginalis* (N.L. Burmann) Kunth), and to be the single most important weed in perennial crops.

**Harvesting** *Imperata* is either grazed when young or cut for thatch when mature. To collect the medicinally important rhizomes, plants are simply uprooted.

**Yield** Reported above-ground dry matter yields of *I. cylindrica* are 2–11 t/ha per year. In Indonesia per ha production was found to be 11 t of leaves and 7 t of rhizomes, with on average 4.5 million shoots.

**Genetic resources and breeding** *Imperata* is widespread and common throughout Malesia, in general favoured by human activities, and therefore certainly not endangered. There are no known breeding programmes on *Imperata*.

**Prospects** Although *Imperata* is widely considered a serious weed, it can be useful for various purposes, especially to provide fodder when young and thatching material when mature. A broad range of medicinal uses has been reported, but most claims still need to be validated, though compounds that have shown vasodilative activity, anti-inflammatory properties or inhibition of platelet aggregation have been isolated.

**Literature** |1| Aguilar, N.O., 1992. *Imperata cylindrica* (L.) Raeuschel. In: 't Mannelje, L. & Jones, R.M. (Editors): Plant Resources of South-East Asia No 4. Forages. Pudoc Scientific Publishers, Wageningen, the Netherlands. pp. 140–142. |2| Falvey, J.L., 1981. *Imperata cylindrica* and animal production in South-East Asia: a review.

Tropical Grasslands 15: 52–56. |3| Holm, L.G., Plucknett, D.L., Pancho, J.V. & Herberger, J.P., 1977. The world's worst weeds. Distribution and biology. East-West Center. The University Press of Hawaii, Honolulu, United States. pp. 62–71. |4| MacDicken, K.G., Hairiah, K., Otsamo, A., Duguma, B. & Majid, N.M., 1997. Shade-based control of *Imperata cylindrica*: tree fallows and cover crops. *Agroforestry Systems* 36: 131–149. |5| Matsunaga, K., Ikeda, M., Shibuya, M. & Ohizumi, Y., 1994. Cylindol, a novel biphenyl ether with 5-lipoxygenase inhibitory activity, and a related compound from *Imperata cylindrica*. *Journal of Natural Products* 57(9): 1290–1293. |6| Matsunaga, K., Shibuya, M. & Ohizumi, Y., 1994. Cylindrene, a novel sesquiterpenoid from *Imperata cylindrica* with inhibitory activity on contractions of vascular smooth muscle. *Journal of Natural Products* 57(8): 1183–1184. |7| Matsunaga, K., Shibuya, M. & Ohizumi, Y., 1994. Graminone B, a novel lignan with vasodilative activity from *Imperata cylindrica*. *Journal of Natural Products* 57(12): 1734–1736. |8| Matsunaga, K., Shibuya, M. & Ohizumi, Y., 1995. Imperanene, a novel phenolic compound with platelet aggregation inhibitory activity from *Imperata cylindrica*. *Journal of Natural Products* 58(1): 138–139. |9| Terry, P.J., Adjers, G., Akobundu, I.O., Anoka, A.U., Drilling, M.E., Tjitrosemito, S. & Utomo, M., 1997. Herbicides and mechanical control of *Imperata cylindrica* as a first step in grassland rehabilitation. *Agroforestry Systems* 36: 151–179. |10| Watson, L. & Dallwitz, M.J., 1992. The grass genera of the world. C.A.B. International, Wallingford, United Kingdom. pp. 470–471.

#### *Selection of species*

#### ***Imperata conferta* (J.S. Presl) Ohwi**

Bot. Mag., Tokyo 55: 549 (1941).

**Synonyms** *Saccharum confertum* J.S. Presl (1830), *Imperata exaltata* (Roxb.) Brongn. (1831).

**Vernacular names** Malaysia: lalang jawa. Philippines: kogon-lake (Tagalog), gogon (Bikol), kogon (Tagalog, Bisaya, Sulu). Vietnam: c[or] lau.

**Distribution** *I. conferta* is found throughout South-East Asia in habitats similar to *I. cylindrica*, but it is less common.

**Uses** In the Philippines a decoction of the rhizomes is drunk against diarrhoea caused by indigestion and against gonorrhoea. The leaves are often used for thatching and the young shoots are grazed. The plants are sometimes used to make

hats and mats. Experiments have shown that they can be used to make good paper.

**Observations** A rhizomatous, herbaceous perennial up to 200 cm tall, culms glabrous or with a few hairs below the nodes; leaf sheaths of basal leaves glabrous, leaf blade 30–80 cm × 0.7–2 cm, scabrid, glabrous; inflorescence a spiciform panicle 25–50 cm long, floret with upper glume 3–4-veined, stamen 1. *I. conferta* is found in a wide range of open, disturbed anthropogenic habitats from sea-level up to 1000 m altitude, but in the Philippines in general preferring forest clearings at higher elevations.

**Selected sources** 97, 172, 190, 202, 577, 580, 630, 1178, 1386.

### *Imperata cylindrica* (L.) Raeuschel

Nomencl. bot., ed. 3: 10 (1797).

**Synonyms** *Lagurus cylindricus* L. (1759), *Imperata arundinacea* Cirillo (1792).

**Vernacular names** Cogon grass, satintail (En). Paillotte (Fr). Indonesia: alang-alang, ilalang, lalang (general). Malaysia: lalang, alang-alang. Papua New Guinea: kunai (Pidgin), kurukuru (Barakau, Central Province). Philippines: kogon (Tagalog), gogon (Bikol), bulum (Ifugao). Burma (Myanmar): kyet-meí. Cambodia: sbō:w. Laos: hn-ha:z kh'a:. Thailand: ya-kha, laa laeng, koe hee (Karen, Mae Hong Son). Vietnam: c[or] tranh.

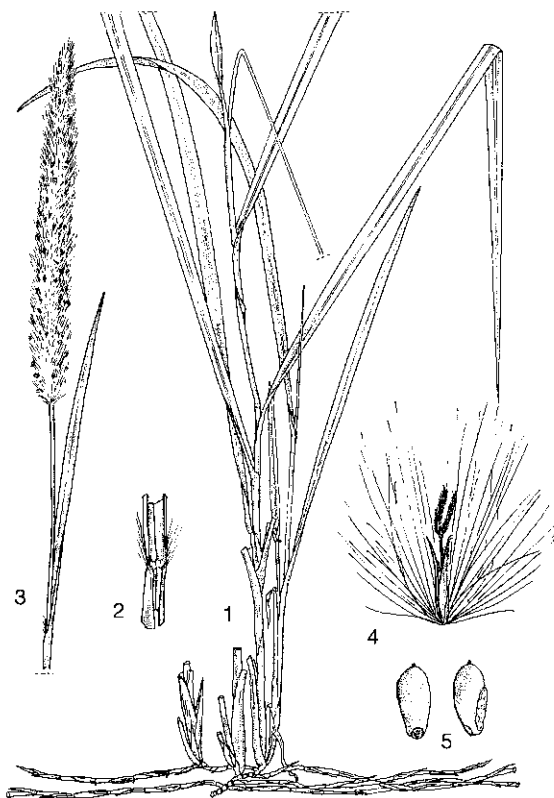
**Distribution** Widely distributed throughout the tropics and subtropics of Africa, the Indian subcontinent, South-East Asia and Australia; occurring to a lesser extent in North, Central and South America, and also occurring in warm temperate areas, being recorded at latitudes of 45° in New Zealand and Japan.

**Uses** Rhizome decoctions of *I. cylindrica* are used as a diuretic in Indonesia, Malaysia, Thailand, Indo-China and China. A rhizome decoction is used to treat dysentery in the Philippines and to treat diarrhoea and gonorrhoea in Malaysia. In Malaysia and Papua New Guinea, rhizome decoctions are used to purify the blood. Furthermore, *I. cylindrica* is administered in the treatment of nose, kidney and bladder complaints in Papua New Guinea, and the shoots are chewed to treat diarrhoea. In Brunei, a root decoction is used as febrifuge. In Indo-China and by Chinese throughout the South-East Asian region the rhizomes are considered haemostatic and cooling and are prescribed as a general tonic as well as in cases of acute nephritis, hypertension, epistaxis and haemoptysis. The flowers or sprouts are reported to be thirst-relieving and digestive, and are used

in the treatment of haemorrhages and wounds. In the Philippines the fruiting spikes are considered a vulnerary and sedative. The rhizomes contain a fair amount of starch and a kind of beer can be made from them. The leaves are often used for thatching and the young shoots are grazed. The plants are sometimes used to make hats and mats. Experiments have shown that they can be used to make good paper.

**Observations** A rhizomatous herbaceous perennial up to 120(–300) cm tall, culms below the nodes usually with a crown of long slender hairs; leaf sheaths of basal leaves coriaceous, glabrous or finely hairy, leaf blade 12–80 cm × 0.5–2 cm; inflorescence a spiciform panicle 10–30 cm long, floret with upper glume 4–5-veined, stamens 2. *I. cylindrica* is found in a wide range of open anthropogenic habitats, preferring well-aerated soils, from sea-level up to 2700 m altitude.

**Selected sources** 89, 97, 172, 190, 202, 287, 332, 362, 401, 404, 474, 572, 577, 580, 597, 614,



*Imperata cylindrica* (L.) Raeuschel – 1, plant habit; 2, ligule; 3, inflorescence; 4, spikelet; 5, caryopsis in front and side view.



630, 1035, 1043, 1126, 1178, 1375, 1385, 1386, 1571.

Juliana Jonathan & Bambang P.J. Hariadi

## ***Ixora* L.**

Sp. pl. 1: 110 (1753); Gen. pl. ed. 5: 48 (1754).

### RUBIACEAE

$x = 11$ ; *I. chinensis*, *I. coccinea*, *I. javanica*, *I. nigricans*:  $2n = 22$

**Major species** *Ixora chinensis* Lamk, *I. coccinea* L., *I. javanica* (Blume) DC., *I. longifolia* J.E. Smith, *I. nigricans* R.Br. ex Wight & Arn.

**Vernacular names** Indonesia: soka (general), ki soka (Sundanese), areng-arengan (Javanese). Malaysia: pechah priok, jarum-jarum, todong perik (Peninsular). Philippines: santan (Tagalog). Burma (Myanmar): ponna, pan. Cambodia: chann tanea, tè prey. Thailand: kheme (general). Vietnam: d[ow]n, b[oo]ng trang.

**Origin and geographic distribution** *Ixora* consists of about 400 species, and is distributed throughout the tropics. The Indo-Malesian region is richest in species. In Malesia about 160 species occur; the highest number of species (about 65) is found in Borneo, most of them endemic.

**Uses** Several *Ixora* species are used in traditional medicine, e.g. as an astringent and to treat dysentery and tuberculosis. The use in China and India is widespread. An infusion of the leaves or flowers of several species is administered to treat fever, headache and colic. A decoction of the roots is used as a sedative; the roots are believed to be more potent. The internal application is based on stomachic and antiseptic properties, while external applications are based on astringent and antiseptic properties.

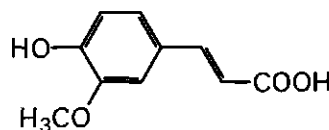
*Ixora* species are well known as ornamentals (e.g. *I. chinensis*, *I. coccinea*, *I. javanica*), and are commonly planted in gardens, parks and along roadsides. The fruits of *I. philippinensis* Merr. are edible. The wood of *Ixora* is occasionally used, often for implements and comparatively small objects; only a few species reach timber size.

**Properties** In a modified tumour promotion test, complete inhibition of all kinds of tumours was exhibited by decoctions of flowers of *I. coccinea* and *I. chinensis*. The antitumour factor from *I. javanica* flowers showed broad activity against transplantable solid tumours in mice by inhibiting the growth of tumours and arresting the growth of already formed tumours; it showed lesser activity against ascites tumours. In vitro studies showed

50% cytotoxicity to Dalton's lymphoma and Ehrlich ascites tumour cells at concentrations of 12 µg and 65 µg, respectively, with no activity against normal lymphocytes but preferential activity against lymphocytes derived from leukaemia patients and K 562 suspension cell culture. Topical application of 100 mg/kg body weight of *I. javanica* flower extract inhibited the growth and delayed the onset of papilloma formation in mice initiated with 7,12-dimethylbenz- $\alpha$ -anthracene (DMBA) and promoted by using croton oil. When administered orally at the same dose the extract significantly inhibited the growth of soft tissue fibrosarcomas induced by subcutaneously injected 20-methylcholanthrene. Oral administration of 200 mg/kg of the extract inhibited the growth of intraperitoneally transplanted sarcoma-180 and Ehrlich ascites carcinoma tumours and also showed an increase in the life span of the treated mice. Toxicity studies showed that the blood urea nitrogen levels were elevated after treatment. Furthermore, tritiated thymidine incorporation studies indicated that the mechanism of action of the factor is at the site of DNA synthesis. The purified fractions contained ferulic acid, pyrocatechuic acid and caffeic acid. The compounds responsible for the inhibitory effects on tumour growth were identified as ferulic acid (4-hydroxy-3-methoxycinnamic acid) and its structural isomer, 3-hydroxy-4-methoxycinnamic acid.

Antimutagenicity tests with *I. coccinea* by the Rec-Assay and the Micronucleus Test revealed that the crude alcoholic extract and the ethyl acetate fraction showed antimutagenic activity. Fractions obtained from the ethyl acetate extract were found to be antimutagenic against a known carcinogen, 4-nitroquinoline, in two *Bacillus subtilis* strains. After purification the antimutagenic fraction was identified by spectroscopic methods as ursolic acid. The activity of the isolated compound was confirmed by the Rec-Assay.

The saponifiable fraction of the petroleum ether extract of *I. coccinea* root was found to have anti-inflammatory activity in carrageen-induced paw oedema in albino rats. The ethanol (50%) extract of the aerial parts potentiates barbiturate activity and causes semen coagulation. The same type of



ferulic acid

extract of aerial parts of *I. nigricans* showed antiviral and hypothermic activities.

The roots of *I. coccinea* contain an acrid aromatic oil, tannin, fatty acids, and a white crystalline substance. The root bark contains  $\delta$ -9,11-octadecadienoic acid, mannitol and myristic acid. The flowers have a yellow colouring matter related to quercitrin, an astringent principle, wax, and a neutral crystalline substance. The roots of *I. chinensis* also contain an iridoid derivative called ixoside (1,8-dehydroxyforsythide).

**Description** Shrubs to small or sometimes medium-sized trees up to 25 m tall; bark surface smooth, lenticellate, fissured or scaly, greyish-brown; twigs terete, often with series of rather close, leafless nodes especially at branching points. Leaves opposite or sometimes in whorls of 3, simple, entire, broadly elliptical to linear, herbaceous to coriaceous, base usually acute or cuneate, apex obtuse to acute, acuminate or caudate, almost always glabrous; petiole usually present, concave or canaliculate; stipules interpetiolate, connate at base, distinctly cuspidate or with a long, stiff, needle-like extension at the tips. Inflorescence a terminal corymb or corymbose panicle, peduncle short with an erect inflorescence or longer with a nodding or pendulous inflorescence, often puberulous or pubescent, usually 45–300-flowered. Flowers often 3 together, bisexual, 4-merous, fragrant or not, protandrous; calyx often divided to the base; corolla with a cylindrical tube, lobes contorted in bud, spreading and flat or reflexed in the open flower, often white but sometimes pink, yellow or red; stamens inserted at corolla throat, with short filaments, anthers dorsifixed, sagittate and reflexed, usually yellow; disk annular; ovary inferior, 2(–3)-locular with 1 ovule per cell, style filiform, slightly exerted from the corolla tube, the exerted part not longer than the corolla lobes, stigma 2-lobed, the lobes linear and recurved. Fruit a globose to 2-lobed drupe, 5–15 mm in diameter, ripening red to black, with 1–2 pyrenes; pyrenes 1-seeded, thin-walled, plano-convex, with a round excavation inside. Seed with thin testa, endosperm entire. Seedling with epigeal germination; cotyledons leafy, green.

**Growth and development** Malesian *Ixora* species are generally evergreen. Flowering is seasonal but cultivars may bloom throughout the year. The flowers are mainly pollinated by moths and butterflies probing for the nectar at the corolla base, but honey-suckers may also visit the flowers, particularly the reddish ones. The seeds are probably dispersed by fruit-eating birds.

**Other botanical information** As a rule the Malesian *Ixora* species have a rather local distribution. *I. nigricans* is an exception, ranging from India to Bali. Many of the species in Java are endemic, as are most of the Bornean species. The New Guinean species are all endemic.

*Pavetta* closely resembles *Ixora*, but can be distinguished by its long-exserted style with coherent stigmas, whereas the anthers are conspicuously twisted. The differences between the widely cultivated *I. chinensis* and *I. coccinea* are sometimes obscure as a result of selection for rare or extreme forms. Accidental or deliberate hybridization appears to occur.

**Ecology** *Ixora* species are usually confined to lowland and lower montane forest up to 1700 m altitude. Some species are also found in swampy locations in the vicinity of rivers or occasionally in rice fields (e.g. *I. grandifolia*).

**Propagation and planting** *Ixora* may be propagated by seed, although ornamental species are usually propagated by cuttings. Both seeds and sown fruits of *I. lobbii* have been found to have about 25% germination in 1–3 months. Treating *I. coccinea* cuttings of 15 cm long by dipping them in indole butyric acid at 2000 ppm for 10 seconds, gave a rooting success of 87%, the development of a high number of primary roots and a survival of 96%. This was considerably better than the performance of untreated cuttings: only 40% rooting success and 67% survival.

**Genetic resources and breeding** The high incidence of endemism in *Ixora* may increase the risk of genetic erosion. However, the species with a reported medicinal use have a relatively large area of natural distribution or are widely cultivated.

**Prospects** The reported antitumour and antimutagenic activities of *Ixora* may justify more research, which might result in future applications in modern medicine. Moreover, several species are attractive ornamentals.

**Literature** [1] Bremekamp, C.E.B., 1937. The Malaysian species of the genus *Ixora* (Rub.). Contributions à l'étude de la flore des Indes Néerlandaises XXXIV. Bulletin du Jardin Botanique de Buitenzorg, Série III, 14: 197–367. [2] Burkill, I.H., 1966. A dictionary of the economic products of the Malay Peninsula. Revised edition, vol. 2. Ministry of Agriculture and Co-operatives, Kuala Lumpur, Malaysia. pp. 1280–1283. [3] Fosberg, F.R. & Sachet, H.H., 1989. Three cultivated *Ixoras* (Rubiaceae). *Baileya* 23(2): 74–85. [4] Gupta, V.N. & Kher, M.A., 1989. A note on the effect of root

promoting hormones on rooting of *Ixora coccinea* L. by tip cuttings under intermittent mist. *Progressive Horticulture* 21(1-2): 138-140. |5| Nair, S.C. & Panikkar, K.R., 1990. Antitumour principles from *Ixora javanica*. *Cancer Letters* 49(2): 121-126. |6| Nair, S.C., Panikkar, B., Akamanchi, K.F. & Panikkar, K.R., 1991. Inhibitory effects of *Ixora javanica* extract on skin chemical carcinogenesis in mice and its antitumour activity. *Cancer Letters* 60(3): 253-258. |7| Padmaja, V., Sudhakaran Nair, C.R., Velayudha Panicker, P. & Hisham, A., 1993. Anti-inflammatory activity of the saponifiable fraction of the petroleum ether extract of the root of *Ixora coccinea* Linn. *Indian Journal of Pharmaceutical Sciences* 55(1): 28-32. |8| Panlilio, B., Aguinaldo, A., Yamauchi, T., Abe, F., Lim-Sylianco, C.Y. & Guevara, B., 1992. An antimutagenic constituent from *Ixora coccinea* Linn. (Rubiaceae). Paper presented at 7th Asian Symposium on Medicinal Plants, Spices, and other Natural products (ASOMPS VII) February 2-7, 1992, Manila, the Philippines. |9| Quisumbing, E., 1978. *Medicinal plants of the Philippines*. Katha publishing Co., Quezon City, the Philippines. pp. 912-913. |10| Serrame, E. & Lim-Sylianco, C.Y., 1995. Anti-tumor promoting activity of decoctions and expressed juices from Philippine medicinal plants. *Philippine Journal of Science* 124(3): 275-281.

#### *Selection of species*

### ***Ixora chinensis* Lamk**

Encycl. 3: 344 (1789).

**Synonyms** *Ixora stricta* Roxb. (1820).

**Vernacular names** Chinese *ixora* (En). Indonesia: kembang soka, siantan. Malaysia: jarum-jarum merah, pechah priok, siantan hutan (Peninsular). Philippines: santan (Tagalog, Bikol), santan-pula, santan-tsina (Tagalog). Cambodia: kam rontea. Vietnam: d[ow]n d[or], b[oo]ng trang d[or].

**Distribution** Southern Burma (Myanmar), Vietnam, Peninsular Malaysia, Borneo; cultivated in Java, the Philippines and elsewhere.

**Uses** In Malaysia a decoction of the root is used after childbirth. In the Philippines an infusion of the fresh flowers is said to be a remedy against incipient tuberculosis and haemorrhage. An infusion of leaves or flowers is used against headache. In Indonesia, a decoction of the roots is used against bronchial disorders; a decoction of the flowers is prescribed in amenorrhoea and hyper-

tension. *I. chinensis* is widely cultivated as an ornamental.

**Observations** A shrub with many stems, up to 2 m tall; leaves obovate-oblong, 6-10 cm × 2.5-5 cm, coriaceous, base rounded, cordate or sometimes obtuse, apex obtuse, petiole short, stipules long-awned; branchlets of inflorescence opposite, red; flowers with corolla tube 3-3.5 cm long, lobes circular-obovate, broadly rounded at apex, 6 mm × 6 mm, orange-red or white (cultivated plants only), not fragrant; fruit globose, black. *I. chinensis* is reportedly common on river banks in Peninsular Malaysia.

**Selected sources** 97, 182, 202, 427, 625, 768, 1126, 1178, 1227, 1572, 1591.

### ***Ixora coccinea* L.**

Sp. pl. 1: 110 (1753).

**Synonyms** *Ixora montana* Lour. (1790), *Ixora grandiflora* Loddiges (1819).

**Vernacular names** Red *ixora* (En). Indonesia: soka merah (general), kembang santen merah (Malay), soka beureum (Sundanese). Philippines: santan-pula, santan (Tagalog), tangpupo (Bisaya). Cambodia: kam ron tea. Thailand: khem baan, khem nuu (Bangkok), khem farang (central). Vietnam: b[oo]ng trang d[or], d[ow]n d[or].

**Distribution** Native in India, widely cultivated in Indonesia, Malaysia, the Philippines, Vietnam, Cambodia, Laos and Thailand.

**Uses** In the Philippines a decoction of the roots is used as a sedative in the treatment of nausea, hiccups and loss of appetite. The flowers are used in the treatment of dysentery, leucorrhoea and dysmenorrhoea, and a decoction of the flowers is prescribed to treat haemoptysis and catarrhal bronchitis. In Indo-China a decoction or infusion of the roots is administered to clear the urine. A decoction of the root is used in folk medicine as an analgesic, sedative, diuretic and antidiarrhoeic; the flowers have the same but weaker properties. In Thailand, the roots or flowers are used as anti-inflammatory and antidiarrhoeal drugs, astringent, tranquilizer and appetite stimulant. The flowers are also used as cholagogue and stimulant of digestive enzyme secretion. In India the roots are reported to possess sedative and stomachic properties and are used against hiccups, fever, gonorrhoea, loss of appetite, diarrhoea and dysentery. They are reported to stimulate gastric secretions and bile and to provide relief in abdominal pains. The roots possess astringent and antiseptic properties and are applied to sores and chronic ulcers, and also to treat headache. A decoction of the



*Ixora coccinea* L. - 1, flowering twig; 2, flower; 3, flower as seen from above.

flowers or the bark is employed as a lotion against eye troubles, sores and ulcers. The leaves are used to treat diarrhoea. *I. coccinea* is widely cultivated as an ornamental.

**Observations** A shrub with many stems, up to 3 m tall, glabrous; leaves ovate to oblong or obovate, 3.5–10 cm × 2–5 cm, coriaceous, base subcordate or rounded, apex obtuse or slightly acuminate, mucronate, with 8–15 secondary veins, petiole absent or short, stipules long-awned; inflorescence sessile, densely corymbose-shaped; flowers with triangular calyx lobes, about 3 mm long, acute, red, corolla tube 3–4.5 cm long, lobes lanceolate or ovate-lanceolate, 1–1.5 cm long, acute, orange to scarlet or white, yellow or pink (mostly in cultivated plants), not fragrant, style 3–4 mm exserted, red; fruit globose, about the size of a pea, reddish, fleshy. *I. coccinea* is cultivated in lowland areas but also at higher elevations.

**Selected sources** 97, 182, 202, 288, 332, 427, 531, 580, 768, 1035, 1126, 1128, 1178, 1227, 1310, 1591.

### *Ixora grandifolia* Zoll. & Moritzi

Syst. Verz.: 65 (1846).

**Synonyms** *Ixora crassifolia* Ridley (1918), *Ixora ridleyi* Bremek. (1937).

**Vernacular names** Pink river ixora (En). Indonesia: sikatan (Javanese), ki soka (Sundanese). Malaysia: jarum hutan, segading jantan, kelat tandok (Peninsular). Thailand: khem yai (Bangkok, peninsular).

**Distribution** Sri Lanka, Burma (Myanmar), Indo-China, Thailand, Peninsular Malaysia, Singapore, Sumatra, Bangka, Java, Madura and Borneo.

**Uses** In Malaysia a decoction of the root is used to treat ague and colic. The leaves are eaten before childbirth, at the commencement of labour and are considered to make delivery easier. An infusion of leaves is drunk against stomach-ache.

**Observations** A shrub or small to medium-sized tree up to 18 m tall, bark smooth to lenticellate, fissured or scaly, grey-brown; leaves elliptical, ovate or obovate, 10–32 cm × 4–18 cm, thickly coriaceous, glabrous, base acute, rounded or cordate, apex acute or blunt, with 6–16 secondary veins, petiole 0.5–3.5 cm long, stipules broadly triangular, apiculate; inflorescence subsessile, with 3 main branches, erect, spreading, up to 5 cm long, branches not jointed; flowers long-pedicellate, calyx tube 0.5–1 mm long, lobes triangular and less than 0.5 mm long, corolla tube 0.5–3 cm long, lobes 2.5–6 mm × 1.5–2 mm, white sometimes pink-tipped, fragrant, anthers grey; fruit globose or strongly 2-lobed, up to 12 mm wide, red turning black. *I. grandifolia* occurs in both lowland and hill forest, also on swampy ground.

**Selected sources** 97, 182, 202, 288, 768, 1126, 1227, 1591.

### *Ixora javanica* (Blume) DC.

Prodr. 4: 487 (1830).

**Synonyms** *Ixora amoena* Wallich ex G. Don (1834).

**Vernacular names** Javanese ixora (En). Thailand: khem thong (Chumphon).

**Distribution** Southern Burma (Myanmar), Thailand, Peninsular Malaysia, Sumatra, Java and Borneo.

**Uses** There are no medicinal uses reported for *I. javanica*, but extracts showed promising antitumour activity. It is cultivated as an ornamental.

**Observations** A shrub, 3–5 m tall; leaves elliptical, oblong or oblong-ovate, 7.5–17 cm × 2.5–7 cm, herbaceous, base acute, apex acuminate, with 9–10 secondary veins, petiole 3–6 mm long, stip-

ules long-awned; inflorescence loose, short-haired, peduncle 1–4 cm long; flowers with calyx tube 0.2 mm long, lobes ovate, 0.4 mm long, corolla tube 2.5–3.5 cm long, lobes ovate, obtuse or rounded, 6–8 mm long, orange-red sometimes pink or yellow, not fragrant, anthers pale orange, style 5 mm long slightly exserted; fruit about the size of a pea. *I. javanica* is common in evergreen forest on fertile soils in Java.

**Selected sources** 97, 182, 768, 1001, 1002, 1591.

### ***Ixora lobbii* Loudon**

Encycl. pl. new edition: 1543 (1855).

**Vernacular names** Brunei: petagar mangas. Malaysia: bunga selang, pechah priok, kramat hujan (Peninsular). Thailand: khem daeng (Yala).

**Distribution** Thailand and Peninsular Malaysia.

**Uses** In Malaysia a decoction of the root is given during and after childbirth, and a poultice is made from it against headache. In Brunei, a decoction of the root of a plant identified as *I. lobbii* is taken as a tonic, or used as a herbal bath.

**Observations** A shrub up to 2 m tall; leaves entire, oblong to lanceolate, 10–20 cm × 2–4.8 cm, subcoriaceous, base cuneate, apex caudate or mucronate, with 15–25 secondary veins, petiole short, 2–4 mm long, stipules triangular, acuminate; inflorescence a subsessile loose corymb with about 50 flowers; flowers with red pedicel, calyx cylindrical campanulate, corolla tube 3–4 cm long, lobes 9 mm × 3–4 mm, acute, bright orange-red, not fragrant; fruit black. *I. lobbii* is a common shrub in forest throughout Peninsular Malaysia and Thailand.

**Selected sources** 182, 202, 288, 964, 1126, 1227.

### ***Ixora longifolia* J.E. Smith**

Rees, Cycl. XIX: n. 3 (1811).

**Synonyms** *Ixora amboinica* (Blume) DC. (1830), *Ixora fulgens* auct. non Roxb.

**Vernacular names** Indonesia: jarong-jarong (Moluccas).

**Distribution** The Moluccas.

**Uses** The roots have been reported long ago to be used against pain in the side in the Moluccas, both internally and externally; chewing the roots has been reported to ease toothache.

**Observations** A shrub 1.5–3 m tall; leaves lanceolate or oblong-lanceolate, 15–30 cm × 9–11 cm, herbaceous or subcoriaceous, base rounded or subacute, apex acuminate, with about 13 sec-

ondary veins, petiole 1–1.5 cm long, stipules broadly triangular, shortly awned; inflorescence loose, shortly pubescent, peduncle 3 mm long, with up to 100 flowers; flowers with calyx tube 0.5 mm long, lobes broadly ovate and 0.5 mm long, corolla tube 4 cm long, lobes about 1 cm long, acute, red; fruit red turning black at maturity. *I. longifolia* is found in abandoned fields and dense scrub up to the beach, but it is less common at higher elevations.

**Selected sources** 97, 182, 580, 1126, 1227, 1265.

### ***Ixora nigricans* R.Br. ex Wight & Arn.**

Prodr. fl. Ind. orient. 1: 428 (1834).

**Vernacular names** Thailand: khem tuut maa (Sukhothai), khem nam, khem phuut maa (Surat Thani, Yala). Vietnam: b[oo]ng trang tr[aws]ng, d[ow]n tr[aws]ng.

**Distribution** India, Burma (Myanmar), Vietnam, Thailand, Peninsular Malaysia, Sumatra, Java, Bali.

**Uses** In Vietnam *I. nigricans* is used similar to *I. coccinea*. The leaves are reported to be used in India for antidiarrhetic purposes.

**Observations** A shrub or small tree up to 5 m tall; leaves elliptical, obovate or oblong, 7–12.5 (–18) cm × 4–6.5 cm, herbaceous, base acute, apex acutely acuminate, with 7–9 secondary veins, dark green above, pale green below, turning black when dried, petiole about 5 mm long, stipules subtruncate with a very long awn; inflorescence loose, subpaniculiform, peduncle 2–4.5 cm long, often nodding, with 100–200 flowers; flowers with pedicel 0.5–2 mm long, calyx lobes oblong-triangular, about 1.2 mm long, corolla tube 8–12 mm long, glabrous, lobes 6–7 mm long, acute, white, fragrant, style about 6 mm long exserted, glabrous, anthers 4–6 mm long, violet; fruit globose, black. *I. nigricans* is found in evergreen forest on fertile soils from sea-level up to 800 m altitude.

**Selected sources** 97, 182, 288, 768, 1035, 1126, 1128, 1591.

M.C. Ysrael & J.L.C.H. van Valkenburg

## **Jasminum L.**

Sp. pl. 1: 7 (1753); Gen. pl. ed. 5: 7 (1754).

OLEACEAE

$x = 13$ ; *J. multiflorum*, *J. sambac*:  $2n = 26, 39$

**Major species** *Jasminum elongatum* (Bergius) Willd., *J. multiflorum* (Burm.f.) Andr., *J. sambac* (L.) Aiton.

**Vernacular names** Jasmine (En). Jasmin (Fr).

**Origin and geographic distribution** *Jasminum* consists of about 200 species, with approximately 90 species occurring in the Old World tropics. It comprises 52 indigenous species in the Malesian region. *J. multiflorum* and *J. sambac* have been cultivated since antiquity.

**Uses** The flowers and leaves of several *Jasminum* species are used as a lactifuge in the Malesian region. Either the bruised flowers or a poultice of the leaves are applied to the breast. The flowers are also considered cooling and are soaked in water overnight; the fluid is used as a face wash. Poultices of the leaves of several species are used in the treatment of ulcers or skin complaints in general. The leaves are also employed as a mouthwash for inflamed gums and ulceration of the mucous membranes. Extracts of the roots of several species are used as febrifuges. A tincture made from the root of *J. sambac* is said to have very strong sedative, anaesthetic and vulnerary properties. Roots are used as poultices to treat sprains and fractures. A decoction of the roots or an infusion of the flowers is employed in pulmonary catarrh, bronchitis, and also asthma.

The flowers of *J. grandiflorum* L. and *J. sambac* are the traditional flavouring agents for jasmine tea. *J. grandiflorum* is widely cultivated in the tropics and subtropics, particularly for its essential oil, but also for its ornamental and medicinal properties.

**Production and international trade** In South-East Asia, the flowers of *J. grandiflorum* and *J. sambac* are produced for the local market only and no export data are available. In India, *J. sambac* is commercially cultivated for its essential oil. *J. grandiflorum* is commercially cultivated in Algeria, Morocco, Italy, Spain, Egypt, India and China for essential oil.

**Properties** The fresh leaves and flowers of *J. multiflorum* have been reported to contain 4 secoiridoid lactones: jasmolactones A, B, C, and D. They all contain a bicyclic 2-oxo-oxepano (4,5-C) pyran ring system. The jasmolactones B and D have been found to possess coronary vasodilating and cardiotropic activities. Furthermore, five 10-hydroxyoleoside-type secoiridoid glycosides (probably derived from secologanin) have been isolated: 10-hydroxyoleuropein, 10-hydroxyligustroside, multifloroside, multiroside and 10-hydroxyoleoside-11-methylester. Two of them showed cardiotropic and coronary-dilating activities on preparations from isolated guinea-pig organs: multifloroside ( $\geq 1.5 \mu\text{M}$  and  $\geq 3.7 \mu\text{M}$ , respective-

ly) and 10-hydroxyoleuropein (both  $\geq 9 \mu\text{M}$ ).

Direct contact with bruised flowers of *J. multiflorum* in mice suppressed milk production and caused involution of the mammary gland. Exposure to the smell of the flowers alone produced similar, but less marked changes. In humans, the fresh flowers placed on the breasts of women who had just delivered, showed antilactagogue and prolactin-inhibition activities.

An extract of the young shoots of *J. subtriplinerve* inhibited the growth of *Staphylococcus aureus* and *S. haemolyticus*. The whole plant, excluding the root, of *J. sambac* showed hypotensive activity and depressant activity on the central nervous system of mice.

Several terpenes (e.g. caryophyllene, menthene, jasminin and jasmone) and flavonoids (quercetin, rutin and isoquercetin) have been isolated from *J. sambac* flowers. The characteristics of the concrete of *J. sambac* flowers are: specific gravity (30°C) 0.8794, refractive index 1.4665 and melting point 46°C.

**Adulterations and substitutes** Jasmine absolute is frequently adulterated with synthetics, inferior quality absolutes and non-jasmine material. No substitute is able to duplicate the original accurately.

**Description** Scandent shrubs or climbers. Leaves opposite (rarely alternate), seemingly simple (1-foliolate) or 3-9-foliolate, leaflets entire, stipules absent. Inflorescence a terminal or axillary 2-3-chotomous cyme or flowers solitary. Flowers bisexual, regular, usually heterodistylous, fragrant; calyx short, funnel- or bell-shaped with 4-10 linear lobes; corolla with a long narrow cylindrical tube and 4-14 imbricate lobes, white or yellow, outside often tinged with violet or almost entirely violet; stamens 2, inserted on the corolla tube, included or almost so, with short filaments, anthers large, ovoid or oblong, connective apiculate; ovary superior, 2-locular with 2 ovules per cell, style filiform, stigma oblong, 2-lobed. Fruit a 2-lobed berry with 2 seeds, or by abortion entire and 1-seeded. Seeds exalbuminous. Seedling with epigeal germination, paracotyledons thin, green, leaf-like and relatively long persistent.

**Growth and development** In India *J. sambac* grown from layers will start flowering in the second year, but commercial harvest only commences in the third year, and best yields are obtained from the fifth year onwards. Profitable yields are obtained for up to 20-25 years. In India the flowering season lasts from March until September, with flowering peaking in April-July. In Java and

Thailand flowering is more or less throughout the year with the highest production in November–December, whereas in Malaysia, *J. sambac* flowers throughout the year but fewer flowers are produced in the rainy season. Dispersal of seed is most likely by birds that eat the fruits.

**Other botanical information** Within the *Oleaceae*, *Jasminum* is placed in the subfamily *Jasminoideae* and the tribe *Jasmineae* together with *Menodora* from tropical and subtropical America and southern Africa. The common jasmine (*J. officinale* L.) is a native of the Sino-Himalayan region, but cultivated worldwide in many temperate regions; single-flowered and double-flowered forms exist. In Europe, common jasmine is cultivated commercially for perfume. In subtropical and tropical regions *J. grandiflorum* L. is widely cultivated for its essential oil and for its fresh flowers; in China and Java the flowers are used to flavour tea. In South-East Asia *J. sambac* is more important for its fresh flowers and essential oil.

**Ecology** *Jasminum* species occur from seashore habitats to high mountain tops and limestone locations, but most species prefer lowland and lower montane conditions. They are frequently found on forest edges as they do not tolerate deep shade. In general the Malesian species prefer well-drained light loams rich in organic matter with a plentiful supply of moisture. Though species may be cultivated in a wide range of soils, preference is given to slightly sandy loams which are easy to work.

**Propagation and planting** Fruits and seeds are rarely produced by the ornamental *Jasminum*, and propagation is therefore almost exclusively by stem cuttings. The best time for vegetative propagation is during the rainy season when production of flowers tends to be lower and vegetative growth more prolific. Propagation of *J. sambac* is most successful when using hardwood cuttings with 5–6 buds, with sand as growing medium. In India layering of *J. sambac* is commonly practised; a small strip of bark is removed from the branch, which is bent down and partly buried to promote rooting. Propagation from leaf cuttings, consisting of a mature leaf, petiole and an axillary bud, is also possible. The cutting is soaked for about 24 hours in 20 ppm naphthalene acetic acid and then half immersed in the planting medium. After 1 month the roots have developed well; shoot formation follows 2–4 weeks later. In India plants are spaced at 2 m × 2 m giving 2500 plants/ha. Production of flowers increases by 170% if the density is increased to 10 000 plants/ha.

**Husbandry** In commercial cultivation *J. sambac* is trained into bushes. In home gardens it is often trained on arbours. Pruning of *J. sambac* consists of heading back the bushes to about one third of the length of shoots, and thinning unwanted shoots or runners. Pruning should be done during the resting period of the crop and before new flushes emerge. When production of foliage is too prolific, some of the foliage should be selectively removed to promote flowering.

To obtain optimal flower production, *J. sambac* should be grown in full sun, with a regular supply of water and with farm manure applied twice a year during active vegetative growth. Application of nitrogen significantly increases flower production in *J. sambac*. Flowering can be regulated by applying cycocel (chlormequat) after moderate pruning; this shortens time to flowering, prolongs flowering and increases flower yields.

**Diseases and pests** The bud worm *Hendecasis duplifacialis* is a pest of some consequence in *J. sambac* plantings.

**Harvesting** Flowers open and are most scented at dusk. The content of essential oils increases considerably at the time of opening of the flowers. Flowers should preferably be harvested in the early morning. However, flower buds for garlands are usually picked in the late afternoon in India. In general, picked flower buds open 7–9 hours after harvesting. In Malaysia, flower buds are gathered in the morning for sale the same day as garlands or to be put in the hair. Strings of threaded flowers are sold per 'hasta' (measurement from elbow to finger tips). For home use, buds are picked in the evening when they are about to burst.

**Yield** In India the annual production of *J. sambac* flowers amounts to 10 t/ha, yielding 1.2–12 kg concrete. In Indonesia (Java) the monthly flower production per ha is 600 kg for November–December, 90–150 kg for January–June and 90 kg for July–October. Production figures per ha per day for *J. grandiflorum* in Java range from 50 kg in the rainy season to 15 kg in the dry season.

**Handling after harvest** In Malesia the flowers of *J. sambac* are picked as fully developed buds. These are brought to the market as quickly as possible to maintain good quality, i.e. a fresh white colour without blemishes. The quality of the flowers can be improved by ice cooling at the time of harvesting and hydro cooling before packing. Flowerbuds can be stored in polyethylene bags at 10°C for up to 4 days without affecting quality or shelf life.

Perfume can be extracted by enfleurage using

sesame oil as fragrance receptor, but at present jasmine flowers are usually extracted by solvents (e.g. petroleum ether, hexane or liquid carbondioxide) to obtain concrete which is subsequently distilled to produce jasmine absolute.

**Genetic resources and breeding** Few *Jasminum* species are widespread and common e.g. *J. elongatum*. Most species are endemic, sometimes with a very local distribution. The latter are more at risk in view of the continuing forest conversion. Although a considerable number of species are adapted to forest fringes, they do not adapt well in secondary forest settings.

**Prospects** The medicinal potential of the cardiotonic and coronary-dilating activities of the secoiridoids in leaves and flowers deserves further attention. The production of flowers for local consumption will remain an interesting market for small-scale growers. The prospects of plantations of *J. sambac* for production of the essential oil are limited, in view of the technology required and the world market prices.

**Literature** [1] Abdullah, T.L., Ahmad, S.H. & Rejab, N.A., 1993. Determination of floral stages and packaging methods for prolonged storage of *Jasminum multiflorum*. In: Palevitch, D., Simon, J.E. & Mathé, A. (Editors): First world congress on medicinal and aromatic plants for human welfare WOCMAP: raw material production, product introduction. *Acta Horticulturae* 331: 325-329. [2] Abraham, M., Devi, N.S. & Sheela, R., 1979. Inhibiting effect of jasmine flowers on lactation. *Indian Journal of Medical Research* 69: 88-92. [3] Casyao, J.M., 1992. The botany of sampaguita, its production and potentials in industry: an overview. *Philippine Technology Journal* 17(2): 1-12. [4] Jonard, R., 1989. *Jasminum* spp. (Jasmine). Micropropagation and the production of essential oils. In: Bajaj, Y.P.S. (Editor): *Biotechnology in agriculture and forestry*. Vol. 7. Medicinal and aromatic plants 2. Springer Verlag, Berlin, Heidelberg, New York, London, Paris, Tokyo. pp. 315-331. [5] Kiew, R., 1994. Name changes for Malaysian plants. *Begonia wrayi* (Begoniaceae) and *Jasminum aemulum* (Oleaceae). *Malayan Nature Journal* 47(3): 311-317. [6] Kiew, R., 1994. Checklist of *Jasminum* (Oleaceae) in Malesia. *Sandakania* 5: 1-14. [7] Kirtikar, K.R. & Basu, B.D., 1935. In: Blatter, E., Caius, J.F. & Mhaskar, K.S. (Editors): *Indian medicinal plants*. 2nd edition. L.H. Basu, Allahabad, India. pp. 1514-1526. [8] Nambisan, K.M.P. & Krishnan, 1980. Gundumalli - the aromatic jasmine. *Indian Horticulture* 24(4): 23-24. [9] Surjosubandrio, S., 1959. Melati

[Jasmine]. *Madjalah Berkala Pertanian* 10: 236-244. [10] Tobroni, M., 1981. Tanaman melati di Jawa Tengah dan Yogyakarta [*Jasminum* cultivation in Central Java and Yogyakarta]. *Warta Balai Penelitian Teh dan Kina* 7(3/4): 343-353.

#### *Selection of species*

#### ***Jasminum elongatum* (Bergius) Willd.**

Sp. pl. 1: 37 (1797).

**Synonyms** *Jasminum aemulum* R.Br (1810), *Jasminum bifarium* Wallich ex G. Don (1837), *Jasminum pubescens* sensu Backer (1931) non (Retz.) Willd., *Jasminum multiflorum* sensu Bakhuizen f. (1950) non (Burm. f.) Andr.

**Vernacular names** Indonesia: pancasuda (Javanese), malati leuweung (Sundanese), gambir hutan (general). Malaysia: melor hutan, pekan hutan, pekan jantan (Peninsular). Philippines: manol (Central Bisaya), sampagitang-gubat (Tagalog). Thailand: mali som, malulee (Bangkok), mali luei (central).

**Distribution** South-East Asia, occurring from India to Australia, and to southern China. Common throughout the Malesian region, and present on almost all islands.

**Uses** In Malaysia a decoction of the root is administered after childbirth and as a febrifuge, and an infusion against yaws. A poultice of leaves is applied to relieve headache and vertigo, and ague in children. In the Philippines, a decoction of the root is used externally for scurvy, and as a gargle for inflamed gums. In Indonesia the leaves are used in a mixture to reduce fever. A decoction of the leaves can be used to cure intestinal complaints and kidney stones. An infusion of the leaves can be used to treat catarrh of the bladder.

**Observations** A slender, erect or climbing shrub up to 2 m tall, glabrous or subpubescent, with distinctly annular nodes and short opposite branches; leaves 1-foliolate, ovate to ovate-oblong, 3-10 cm × 1.5-5 cm, chartaceous, base rounded or truncate, apex acuminate, round or truncate, with pinnate venation, veins glabrous or midrib pubescent, petiole 3-7(-10) mm long; inflorescence a sub-capitate cyme with (1-)6-9(-15) flowers, sessile, supported by foliaceous bracts; flowers with a campanulate calyx tube 1.5-2 mm long, with 5-7 subulate teeth 1-4(-7) mm long, usually pubescent, corolla tube 10-25 mm long with 6-9 narrow ovate-oblong lobes 6-15 mm × 2-4 mm, apiculate, white; fruit a 2-lobed berry, black. *J. elongatum* is very common in open or disturbed



primary and secondary forest and forest fringes from sea-level up to about 1500 m altitude and occasionally up to 3000 m, e.g. on Mount Kinabalu.

**Selected sources** 92, 97, 202, 271, 506, 732, 750, 1126, 1178, 1227.

***Jasminum multiflorum* (Burm.f.) Andr.**

Bot. repos. 8: t. 496 (1807).

**Synonym** *Jasminum pubescens* (Retz.) Willd. (1797).

**Vernacular names** Star jasmine (En). Philippines: sampagitang-sunsong (Tagalog). Vietnam: nh[af]i nhi[eef]u hoa.

**Distribution** A native of India and cultivated in western Malesia.

**Uses** In India the root is reportedly used as an emmenagogue or emetic. The flowers are applied as a lactifuge. A poultice of the leaves is used to treat indolent ulcers. In Malaysia *J. multiflorum* is grown in gardens to supply flowers for home Hindu altars.

**Observations** A robust bushy climber up to 5 m long, young shoots densely pubescent-tomentose; leaves broadly ovate, 4–7 cm × 1.5–4 cm, base rounded or often cordate, apex acute or often mucronate, pubescent beneath especially at the veins, with 4–6 pairs of secondary veins, slightly bullate, petiole 6–10 mm long; inflorescence a compact, cymose panicle, terminal or at the extremities of short axillary branches, with up to 40 flowers, supported by large, ovate, acute foliaceous bracts; flowers with a 13–16 mm long calyx tube, lobes 7–10(–13) mm long, densely tomentose, corolla tube (15–)20–22 mm long, glabrous, with yellowish-green eye, with 6–9 lobes opening horizontally, recurved at the tip, up to 17 mm × 7–8 mm, overlapping at the base, slightly fragrant; fruit a globose berry, black, surrounded by the suberect calyx lobes.

**Selected sources** 460, 580, 731, 732, 741, 900, 921, 1021, 1178.

***Jasminum sambac* (L.) Aiton**

Hort. Kew. 1: 8 (1789).

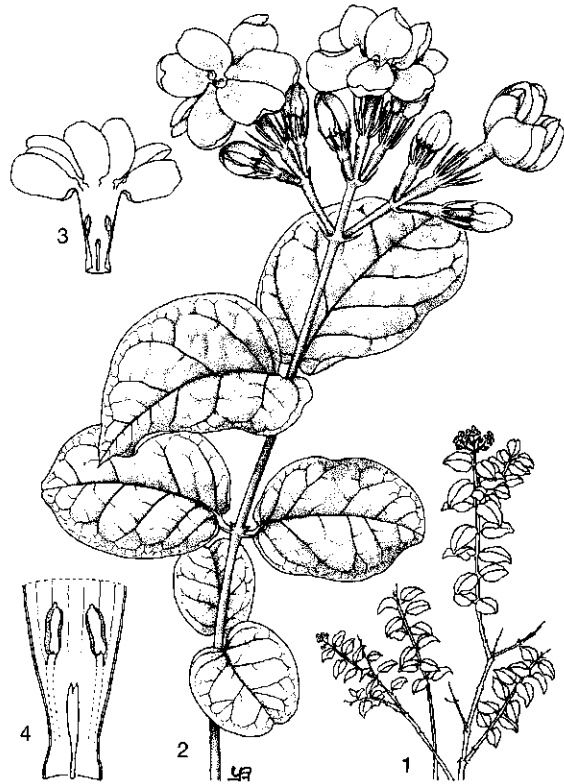
**Synonyms** *Nyctanthes sambac* L. (1753).

**Vernacular names** Arabian jasmine (En). Jamin d'arabie (Fr). Indonesia: melati (general), menur (Javanese). Malaysia: melor (Peninsular). Philippines: manul (Bisaya), sampaguita (Tagalog), kimpupot (Tagalog, Pampanga). Cambodia: molih (Chinese). Thailand: khao taek (Mae Hong Son), tiamuun (Chiang Mai), mali son. Vietnam: l[af]i, hoa nh[af]i.

**Distribution** *J. sambac* probably originated in

India and was brought to Malaysia and Java around the 3rd Century; since then widely cultivated throughout the Malesian region for its heavily scented flowers.

**Uses** The leaves are more medicinal than the flowers. A decoction is used internally against fever. A poultice of the leaves is applied to treat skin complaints and wounds in Malaysia. In India, Indonesia, Malaysia and the Philippines the bruised leaves or flowers are applied as a poultice to the breast of women as a lactifuge. An infusion of the flowers is applied to the eyelids as a decongestant. Besides the above mentioned uses, in Thailand the leaves are used as an astringent and antiamebic. The root is given fresh to treat venereal diseases in Malaysia and to treat fever in Indonesia. A tincture made from the root is said to have very strong sedative, anaesthetic and vulnerary properties. Roots are used as poultices for sprains and fractures. A decoction of the roots or an infusion of the flowers is employed in pulmonary catarrh, bronchitis, and also asthma. The



*Jasminum sambac* (L.) Aiton – 1, plant habit; 2, flowering twig; 3, opened corolla; 4, detail of stamens and style.

stems are employed as an antipyretic and in the treatment of abscesses. The flowers are widely used for their scent and their cooling effect, either directly or in perfumes. In China and Java flowers are used to flavour jasmine tea. In India, *J. sambac* is commercially cultivated for its essential oil.

**Observations** A shrub, untidy (straggling) climbing or lax when young and rooting at the nodes or ascending, up to 3 m tall; leaves all 1-foliolate, ovate, 2.5–9 cm × 2–6.5 cm, thin, base subcordate to obtuse or cuneate, apex obtuse or acuminate, margins subundulate, glabrous or finely pubescent on the main veins, with several sunken and bearded vein-axils beneath; inflorescence a 3-flowered cyme or a many-flowered compact cluster; flowers single or double (in cultivated varieties), with 7–10 calyx segments, 2.5–7 mm long, finely pubescent, corolla tube 7–15 mm long, with 5–many lobes, oval or oblong, 8–15 mm long, mostly white, heavily fragrant; fruit a black berry, surrounded by the calyx. *J. sambac* is widely planted and occurring from sea-level up to 800 m altitude. Several double-flowered varieties are recognized, none of which produce fruit.

**Selected sources** 97, 202, 219, 220, 271, 332, 460, 505, 580, 741, 900, 1021, 1035, 1126, 1128, 1178, 1287, 1571.

### **Jasminum subtriplinerne Blume**

Mus. bot. 1 : 272 (1851).

**Vernacular names** Vietnam: d[aa]jy v[awf]ng, ch[ef] v[awf]ng, d[aa]jy c[aa]r m v[aa]n.

**Distribution** Northern India, Vietnam, Laos and Cambodia.

**Uses** In Vietnam an infusion of the leaves is administered to women after childbirth. The dried young shoots are used in the treatment of lymphadenopathy, metritis, galactophoritis, leucorrhoea, rheumatism, ostalgia, impetigo, dysmenorrhoea and haematometra. A decoction of the fresh leaves is used to wash wounds and against skin problems; a poultice is used to treat ulcers and mastitis. In Laos, an extract of the pounded root is used to treat quotidian fever. The stem is used for wickerwork and binding.

**Observations** A straggling shrub, lax or ascendant, with slender branches up to 20 m long, glabrous; leaves 1-foliolate, elliptical-lanceolate, 4–8 cm × 2–5 cm, with a round to obtuse base, apex acuminate, 3-veined, petiole 3–12 mm long, glabrous, articulate; inflorescence an axillary or terminal cyme with up to 9 flowers; flowers with a calyx tube 3 mm long, glabrous, calyx lobes 9, linear, up to 9 mm long, corolla tube 16–18 mm long,

with 9 oblong and subobtusate lobes, 15 mm × 3–4 mm, with 3 veins, white, fragrant, anthers 5 mm long, not exerted, style short; fruit a subglobose 1-seeded berry, 7–8 mm in diameter. *J. subtriplinerne* occurs in rather humid forested areas. It may well be grown in similar habitats in the Malesiana region.

**Selected sources** 271, 363, 460, 1035, 1126, 1128.

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### **Jatropha L.**

Sp. pl. 2: 1006 (1753); Gen. pl. ed. 5: 437 (1754).

EUPHORBIACEAE

$x = 11$ ; *J. curcas*:  $2n = 22, 44$ , *J. gossypifolia*, *J. multifida*:  $2n = 22$

**Major species** *Jatropha curcas* L., *J. gossypifolia* L., *J. multifida* L.

**Vernacular names** *Jatropha*, physic nut (En). Indonesia: jarak (general), balacai (Moluccas). Malaysia: jarak (Peninsular). Philippines: tagumbau (Iloko), tuba (Bikol, Tagalog). Laos: nhao.

**Origin and geographic distribution** *Jatropha* comprises about 175 species and is found from warm temperate North America to Central America, the West Indies and drier regions of South America, and furthermore in drier regions of tropical Africa, in South Africa and in India. Several *Jatropha* species are widely cultivated in the tropics and these can be encountered throughout the Malesian region, sometimes as escapes. The centre of origin of *J. curcas* probably lies in Mexico and Central America. It was brought to the Cape Verde Islands and elsewhere in the Old World long ago by the Portuguese. It was present in the Philippines before 1750.

**Uses** The seed oil of *Jatropha* is known as cathartic, although its applications often lead to strong irritation of the gastro-intestinal tract or even poisoning. Further applications include the use, both internally and externally, as an abortifacient. Whole seeds are reported as being cathartic, as well as anthelmintic, and they are used in the treatment of e.g. gout and skin diseases. However, because they contain toxic diterpenes, it is considered unsafe and inadvisable to use seeds and seed oil for these indications. The latex of several *Jatropha* species have a long-standing reputation for healing wounds, as a haemostatic, and in the treatment of various skin problems like eczema and dermatomycosis. It is applied externally in

the treatment of infected wounds, ulcers, cuts, abrasions, ringworm and scabies. Upon drying, the initially viscous juice dries, forming an airtight film, resembling that produced by collodium. Leaves of *J. curcas*, *J. gossypifolia* and *J. multifida* are used as a purgative, as a cure for eczema and itches, as an antiparasitic to scabies, and as a rubefacient to treat rheumatism. Decoctions of the roots are a cure for indigestion (*J. multifida*) and diarrhoea (*J. curcas*). Fresh stems of *J. curcas* are used as toothbrushes, to strengthen the gums, and to cure bleeding, spongy gums or gum boils.

*J. curcas* is widely cultivated in the tropics as a living fence. The seed oil is used for the manufacture of candles and soap, for lamp and motor oil and as fuel for cooking, whereas the seed-cake is applied as fertilizer. *J. multifida* is widely grown for its ornamental foliage and flowers. The seed oil is an ingredient of hair conditioners. *J. curcas* and *J. multifida* might also be used as fish poison.

**Properties** The fatty acid composition of the seed oil of *J. curcas* has been analysed: palmitic acid, stearic acid, oleic acid and linoleic acid occur in large quantities. The toxic principles of the oil have been identified as esters of the diterpene 12-deoxy-16-hydroxyphorbol. Their irritant properties have been evaluated in the mouse irritation test, and their co-carcinogenic properties by their enhancing effects on Epstein-Barr virus-induced transformation of human lymphocytes and induction of skin tumours after initiation with 7,12-dimethylbenz[a]anthracene. The toxic properties of entire *J. curcas* seeds in animals and man might not only be attributable to the seed oil and its phorbol esters, but also in part to a toxic protein fraction, sometimes referred to as 'curcin'. More purified proteins from this fraction have been shown in vitro to inhibit protein synthesis in rabbit reticulocytes and to agglutinate human erythrocytes.

Investigations to test the hypothesis that proteolytic activity might be responsible for some of the therapeutic effects (e.g. healing wounds, haemostatic) of *J. curcas* latex resulted in the protease curcain being isolated. Curcain is a protein with an average molecular weight of 22 000. Since successful treatment of wounds by the application of various proteolytic enzymes is reported, the wound-healing properties of this protein were investigated in a mouse model. It was shown that healing of wounds by curcain in a hydrophilic ointment (0.5–1%) was better than observed for nitrofurazone.

Focused phytochemical investigation has revealed

the presence of two cyclic peptides, curcacycline A and B, in the latex of *J. curcas*. So far, it seems that they may be restricted to *Jatropha*. The primary structure of curcacycline A appeared to be cyclo-gly-leu-leu-gly-thr-val-leu-leu. Curcacycline B was shown to be a cyclic heptapeptide, containing 1 val, 1 ser, 2 asn, 1 phe and 2 trp residues. All residues in both peptides were found to have the L-configuration. Curcacycline A showed a moderate dose-dependent inhibition of human T cell proliferation. No direct cytotoxic effects as measured by trypan blue dye exclusion were observed.

The leaves of *J. curcas* have a potent cardiovascular action, and are a possible source of an anti-arrhythmic ( $\beta$  blocker) agent. Experiments on guinea-pigs showed decreased heart force (negative inotropic), decreased heart rate (negative chronotropic) and blocking of the isoprenaline stimulation response on the auricle, which is somewhat similar to that of a  $\beta$  blocker. The methanol extract of the leaves also showed antispasmodic activity against KCl and acetylcholine. Steroids (stigmasterol,  $\beta$ -sitosterol,  $\beta$ -sitosterol- $\beta$ -D-glucoside) and flavonoids have been found to be present in *J. curcas* too.

In Costa Rica and on Aruba and Curaçao, a decoction of the whole plant or the stems of *J. gossypifolia* has a reputation in the treatment of cancer. On the other hand there is a high incidence of oesophageal cancer on Curaçao, which is possibly due to co-carcinogenic esters of 12-deoxy-16-hydroxyphorbol occurring in the seed oil. In a search for tumour inhibitors of plant origin, it was observed that an alcoholic root extract of *J. gossypifolia* showed significant inhibitory activity in standard animal systems. This finding led to the isolation of jatrophone and the related 2 $\alpha$ -hydroxyjatrophone, 2 $\beta$ -hydroxyjatrophone and 2 $\beta$ -hydroxy-5,6-isojatrophone. Evaluation of the anti-neoplastic activity in the P-388 lymphocytic leukaemia test in vitro and in vivo and Eagle's carcinoma of the nasopharynx in vitro revealed all components to be very active, except for 2 $\beta$ -hydroxy-5,6-isojatrophone showing in vitro, but lacking in vivo activity. In addition, it was found that jatrophone had direct inhibitory effects on contractions of cardiac and smooth muscle preparations, which were typically non-competitive in nature. The action of jatrophone may involve more than one mechanism; this might be related to alterations in Ca<sup>2+</sup> handling by cells.

Much effort has been invested in evaluating the molluscicidal properties of *J. gossypifolia* preparations, because snails are important as interme-

diate hosts in the spread of several infectious diseases (schistosomiasis, trematodiasis). Both the latex and a methanolic seed extract of *J. gossypifolia* have been investigated. The extract was found active against the snail *Bulinus globosus*, but was considered of little value, because of the large quantity required for lethal concentrations and the rapid inactivation in the environment under field conditions. The latex, however, was shown to be strongly active against the snail *Lymnaea acuminata* in its aquatic environment. Toxicity of the latex is partly due to inhibitory effects of its constituents on acetylcholinesterase, and on acid and alkaline phosphatases in the snail. The latex of *J. gossypifolia* furthermore contains cyclic peptides. Cyclogossine A and B were isolated from latex collected from plants growing in Indonesia, and their primary structures were determined as cyclo-leu-ala-thr-trp-leu-gly-val, and cyclo-ala-ile-leu-gly-gly-trp-leu-ala, respectively. Latex from plants growing in Aruba also yielded two cyclic peptides: on the basis of preliminary analysis one was tentatively identified as cyclogossine B, the other was shown to be different from cyclogossine A and B and represents another novel compound named cyclogossine C. All residues were determined to have the L-configuration.

In addition to yielding the compounds mentioned above, phytochemical investigations of *J. gossypifolia* have revealed the presence of lignans in the light petroleum extract of stem, root and seeds, e.g. gadain and prasanthaline, alkaloids in the latex, and flavonoids, i.e. apigenin, vitexin and isovitexin and triterpenes in the ethanolic leaf extract.

The seed oil of *J. multifida*, sometimes used as a purgative, may cause strong irritation and poisoning. The toxic principles have been shown to be esters of the diterpene 16-hydroxyphorbol, known for their irritant and co-carcinogenic properties. Researchers seeking a molecular explanation for some therapeutical effects of *J. multifida* claimed (e.g. treatment of infected wounds, ulcers) subjected the latex to activity-guided isolation procedures. These led to the isolation of, amongst others, 2 cyclic peptides: labaditin and biobollein, with primary structures cyclo-ala-gly-val-trp-thr-val-trp-gly-thr-ile and cyclo-trp-ala-ala-ser-ile-leu-gly-leu-gly, respectively. All residues in both peptides were determined to have the L-configuration. Labaditin and biobollein were found to selectively inhibit classical pathway complement activity; no significant effects on the alternative path-

way or terminal route were observed. More detailed mechanistic studies revealed that their effects were caused by the activation (consumption) of complement rather than a direct inactivation of participating components. Complement component C1 was shown to be the primary target, not C2 or C4. Biobollein was furthermore studied for antigenicity in an in vivo model using mice for delayed-type hypersensitivity. The results indicated that the cyclic peptide is not immunogenic in this model.

Besides yielding the cyclic peptides, the procedure led to the isolation of multifidol and multifidol glucoside, 2 acylphloroglucinols which inhibit the luminol-dependent chemiluminescence mediated by reactive oxygen species produced by activated human polymorpho-nuclear leukocytes, and a proanthocyanidin which inhibits activation of the classical complement pathway through  $\text{Ca}^{2+}$  chelation. The presence of minor amounts of (+)-catechin and (-)-epicatechin was also established; both catechins were found to have an inhibitory effect in the chemiluminescence assay. Furthermore, phytochemical investigations revealed the presence of multifidin (a cyanoglucoside) in the latex, and in the tannins (6–14%).

Finally, *J. curcas* preparations (seeds, seed oil, and seed extracts) are reported to have insecticidal properties.

The energy value of the seed oil is 39–41 MJ/kg for *J. curcas*, 42 MJ/kg for *J. gossypifolia* and 57 MJ/kg for *J. multifida*. Seed oil from *J. curcas* can be used in diesel engines, but it appears to be advisable to make the oil less viscous by trans-esterification.

**Description** Monoecious or rarely dioecious trees, shrubs or herbs with the stem arising from a thick, perennial rootstock, sometimes succulent, with yellow to red latex; indumentum simple, sometimes glandular. Leaves alternate, simple to palmately lobed, less often pinnately lobed, margin entire to serrate, veins looping (brochidodromous); stipules simple or branched, sometimes spiny. Inflorescence terminal or axillary, often corymb-shaped, dichotomously cymose, with a solitary female flower terminating each major axis, lateral cymules male. Flowers unisexual or rarely bisexual, actinomorphic, with (4–)5(–6) sepals often connate at base, and 5 petals which are free or coherent and white, green, yellow-green, yellow-brown or red, or rarely without petals. Male flowers with disk entire or composed of 5 free glands; stamens (6–)8–10, rarely more, commonly arranged in 2 distinct whorls, filaments

free to fused into a column, anthers longitudinally dehiscent; pistillode absent. Female flowers with disk annular, 5-lobed or composed of 5 free glands; ovary superior, (1)–3(–5)-locular with 1 ovule per locule, styles free or connate at base; staminodes sometimes present. Fruit a somewhat fleshy to dry capsule, explosively to tardily dehiscent into 2-valved cocci. Seeds spherical to ovoid or oblong, carunculate, plain or variously mottled; testa crustaceous. Seedling with epigeal germination; hypocotyl elongated; cotyledons emergent or occasionally not emergent (e.g. *J. multifida*), petiolate to sessile; first 2 leaves alternate or less often opposite to subopposite (e.g. *J. multifida*), subsequent ones alternate.

**Growth and development** Growth of *J. curcas* is intermittent, and is regulated by fluctuations in rainfall, temperature and light. Flowers of *Jatropha* are generally insect-pollinated. They produce nectar and may be scented. In subgenus *Jatropha* the nectaries are usually exposed and accessible to flies, wasps and other insects, whereas those in subgenus *Curcas* are hidden in a tube and only accessible to insects with a long proboscis or tongue, such as butterflies. In inflorescences of *J. curcas* the female flowers open one or two days before the male ones or at the same time as the earliest males. Male flowers last only one day. The sweet, heavy perfume at night and greenish-yellow flowers of *J. curcas* suggest that this species is pollinated by moths. Its occasionally bisexual flowers are self-compatible; seed never sets in indoor cultivation unless the flowers are pollinated by hand. In Thailand, *J. curcas* flowers in November and May. The explosively dehiscent capsules of some *Jatropha* species disperse seed over some distance. Ants are presumably another dispersal vector as they may be attracted by the fleshy caruncle. Mycorrhizae have been observed on *J. curcas*. They promote growth, especially where phosphate is limiting.

**Other botanical information** *Jatropha* belongs to the tribe *Jatrophae* of the subfamily *Crotonoideae*. It has been subdivided into 2 subgenera, subgenus *Jatropha* and subgenus *Curcas*, and these again into several sections and subsections. *J. gossypifolia*, and *J. multifida* belong to subgenus *Jatropha*, *J. curcas* belongs to subgenus *Curcas*.

**Ecology** Most *Jatropha* species are well-adapted to arid and semi-arid conditions and generally occur in seasonally dry areas. They grow in grass savanna, scrub vegetation, and other open vegetation. *J. curcas* is most successful in drier tropical

regions with an average annual rainfall of 300–1000 mm, but it has been reported from an area with 2380 mm of rain annually. In its centre of origin it is found in regions with average annual temperatures of 20°C to 28°C, but it does withstand slight frost.

**Propagation and planting** *J. curcas* can be easily propagated by stem cuttings 45–100 cm long and development is more rapid than from seed. Stem cuttings root readily in well-aerated rooting media without rooting hormones. Cuttings 30 cm long developed more roots and their survival rate was higher than cuttings 15 cm long. In one kg there are 1700–2400 seeds of *J. curcas*. Soaking seed overnight improves the germination of *J. curcas*. The seed takes about 10 days to germinate. Both direct seeding and sowing in pots are feasible. Explants from hypocotyls, petioles and leaves effectively produced plants by tissue culture techniques. In heavy soils, root formation of *J. curcas* is reduced. In Thailand, a spacing of 2 m × 2 m showed best vegetative development and the highest first seed yield of nearly 800 kg/ha from plants 13–14-months old. Spacings applied for *J. curcas* are 0.5–1.5 m × 1–2 m (rainfed) to 2 m × 2 m (irrigated) when seeding directly, 1–3 m when seedlings are planted, and 2–3 m × 1.5–3 m when cuttings are planted.

**Husbandry** *J. curcas* cuttings start producing seed 4–5 months after planting. Plants from cuttings are more short-lived and less drought- and disease-resistant than those raised from seed. In north-eastern Thailand, seedling plants 4–5 months after sowing are 1 m tall and start flowering. Plants from cuttings attain 2 m height in 8 months. The mean plant height and mean stem diameter of 11 provenances of *J. curcas* tested in Senegal were 150 cm and 7.5 cm, respectively, 15 months after planting. As plants are not browsed and grow into dense bushes they are favoured for living fences and can also be coppiced. *J. gossypifolia* is a serious weed in pastures in Papua New Guinea.

**Diseases and pests** Although a number of diseases and pests have been observed in different regions in *J. curcas*, none of them causes serious problems.

**Harvesting** Fruits of *J. curcas* which turn yellow on maturity can be easily plucked. Harvesting is often during the dry season and thus it does not interfere with other agricultural activities.

**Yield** Annual seed yield of *J. curcas* in Cape Verde is 400–1200 kg/ha; in Thailand average annual yields of 2150 kg/ha have been obtained. In

the Philippines, 5-year-old plants produce 4–6 kg of dry seed.

**Handling after harvest** Fruits are harvested and allowed to dry and dehisce spontaneously. Seeds are then dried, roasted and ground. The ground seed is boiled in water and the oil can be collected from the surface. When oil is extracted by pressing about 50% of the seed weight remains as seed-cake. This residue contains toxic compounds and cannot be used as animal feed. Biogas has been produced on an experimental scale, using an anaerobic filter; the daily yield was 3.5 m<sup>3</sup> gas from 13 kg seed-cake.

**Genetic resources and breeding** There are only four records of systematic provenance trials on *J. curcas*. *J. curcas* germplasm is maintained in three institutions: Centro Agronomico Tropical de Investigacion y Ensenanza (CATIE) in Costa Rica, the Centre National de Semences Forestières (CNSF) in Burkina Faso and Instituto Nacional de Investigacao e Desenvolvimento Agrario (INIDA), Cape Verde.

**Prospects** The seed or the seed oil of *Jatropha* should not be used as cathartic, because they contain toxic diterpenes, with irritant and co-carcinogenic properties. The latex of several *Jatropha* species has a long-standing reputation for healing wounds, as a haemostatic, and in the treatment of various skin problems like eczema and dermatomycosis. Several constituents that display interesting pharmacological activities have been isolated. The latex seems to be an interesting topical application for use in rural communities. *J. curcas* is well adapted to marginal areas with poor soils and low rainfall, where it grows without competing with annual food crops. Of its many uses, the combination of erosion control and oil production seems particularly promising.

**Literature** |1| Adolf, W., Opferkuch, H.J. & Hecker, E., 1984. Irritant phorbol derivatives from four *Jatropha* species. *Phytochemistry* 23: 129–132. |2| Council of Scientific and Industrial Research, 1959. The wealth of India: a dictionary of Indian raw materials & industrial products. Volume 5. Publications and Information Directorate, New Delhi, India. pp. 293–297. |3| Dehgan, B. & Webster, G.L., 1979. Morphology and infra-generic relationships of the genus *Jatropha* (Euphorbiaceae). University of California Publications in Botany 74. University of California Press, Berkeley & Los Angeles, United States. 73 pp. & 33 plates. |4| Heller, J., 1996. Physic nut. *Jatropha curcas* L. Promoting the conservation and use of underutilized and neglected crops 1. Institute of

Plant Genetics and Crop Plant Research, Gatersleben, Germany & International Plant Genetic Resources Institute, Rome, Italy. 66 pp. |5| Horsten, S.F.A.J., 1995. Cyclic peptides in the genus *Jatropha* (Euphorbiaceae). Thesis, Utrecht University, the Netherlands. 239 pp. |6| Kosasi, S., 1990. Immunomodulators from the latex of *Jatropha multifida* L. Thesis, Utrecht University, the Netherlands. 239 pp. |7| Mizuno, F., Koizumi, S., Osato, T., Kokwaro, J.O. & Ito, Y., 1983. Chinese and African Euphorbiaceae plant extracts: markedly enhancing effect on Epstein-Barr virus induced transformation. *Cancer Letters* 19: 199–205. |8| Nath, L.K., 1992. Wound healing response of the proteolytic enzyme curcain. *Indian Journal of Pharmacology* 24: 114–115. |9| Singh, A. & Agarwal, R.A., 1992. Toxicity of the latex of Euphorbiaceae. Effect on acid and alkaline phosphatases of the snail *Lymnaea acuminata*. *Biological Agriculture and Horticulture* 8: 211–219. |10| Taylor, M.D., Smith, A.B., Furst, G.T., Gunasekara, S.P., Bevelle, C.A., Cordell, G.A., Farnsworth, N.R., Kupchan, S.M., Uchida, H., Branfman, A.R., Daily, R.G. & Sneden, A.T., 1983. New antileukemic jatrophone derivatives from *Jatropha gossypifolia*. *Journal of the American Chemical Society* 105: 3177–3183.

#### *Selection of species*

#### ***Jatropha curcas* L.**

Sp. pl. 2: 1006 (1753).

**Synonyms** *Curcas purgans* Medik. (1771), *Curcas indica* A. Rich. (1853), *Jatropha afrocurcas* Pax (1909).

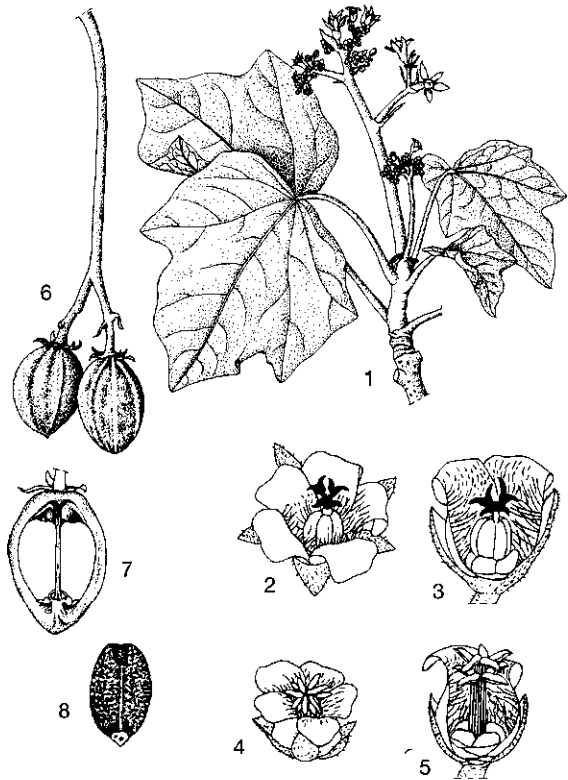
**Vernacular names** Physic nut, purging nut (En). Poughère, pignon d'Inde (Fr). Indonesia: jarak kosta jarak pagar (general), balacai (Moluccas). Malaysia: jarak belanda, jarak keling, jarak pagar (Peninsular). Papua New Guinea: kadel, lam (Gunantuna, New Britain). Philippines: tagumbau-na-purau (Iloko), tuba (Igorot, Bikol, Tagalog), tubang-bakod (Tagalog). Cambodia: kuang, lohong. Laos: nhao. Thailand: ma yao (northern), sabuu dam, salot paa (central). Vietnam: d[aa]f[u] m[ef], ba d[aa]j[u] nam.

**Distribution** *J. curcas* probably originated from Mexico and Central America, but it was introduced long ago in all tropical regions and some subtropical regions like Florida and South Africa. It is cultivated throughout the Malesian region, though especially in the drier areas.

**Uses** The seed oil is possibly the best known

product of *J. curcas* applied as a cathartic, although application often leads to poisoning. Seeds themselves are also used as a cathartic, as well as an anthelmintic, and in the treatment of gout and skin diseases. They are often a source of poisoning, both in animals and humans. The latex is used as a vulnerary by the Malays. It is used to treat ear disease, toothache, eczema and scabies in Indonesia, as a styptic in India, and in Cambodia it is applied to sores and ulcers. The fresh, viscous juice flowing from the leaf stalks or stems is employed to arrest bleeding, and to treat ulcers, cuts and abrasions. It is said to promote healing by coagulating the blood and forming an airtight film when dry, resembling that produced by colloidum. Furthermore, it is a successful local remedy for ringworm. Decoctions of leaves or roots are a good cure for diarrhoea and to treat polyuria, whereas a decoction of the leaves is also employed as a cough remedy. Leaves are applied to wounds and itches (Cambodia), as an antiparasitic to scabies, and as a rubefacient to treat paralysis and rheumatism (Indonesia). In India, crushed leaves are applied as a cataplasm to swollen breasts, and as a lactagogue. The bark is bruised and placed in the mouth as a cure for the bites of snakes or other animals. The bark is also used as a poultice for sprains and dislocations. In Goa, the root bark is applied externally for rheumatism. The fresh stems are used as toothbrushes, to strengthen the gums, and to cure bleeding, spongy gums or gum boils. The juice may kill fish, and can also be applied for stupefying them while hunting (the Philippines). In arid and semi-arid regions *J. curcas* is commonly planted as living fence and for erosion control. A dark blue dye from the bark has been used in the Philippines for colouring cloth, fishing nets and lines. The oil is used for the manufacture of candles and soap and as fuel for cooking, whereas the seed-cake is applied as fertilizer.

**Observations** A somewhat succulent shrub or small tree up to 5(-8) m tall with pink latex, bark smooth, shiny, greenish-brown or yellowish-grey, peeling off in papery scales; leaf blade broadly ovate in outline, usually shallowly (3-)5(-7)-lobed or occasionally not lobed, 7-14(-18) cm x 5.5-14 (-18) cm, shallowly to deeply cordate at base, sparsely puberulous along the veins below at first, otherwise glabrous, petiole (3-)10-15(-20) cm long, glabrous; inflorescence subcorymbose, peduncle up to 5(-7) cm long; male flowers with ovate calyx lobes about 2 mm long, petals fused in lower half, about 3 mm long, greenish-yellow, sta-



*Jatropha curcas* L. - 1, flowering stem; 2, female flower; 3, opened female flower; 4, male flower; 5, opened male flower; 6, fruits; 7, fruit in longitudinal section; 8, seed.

mens 10, in two whorls of 5; female flowers with about 4 mm long calyx lobes, petals about 6 mm long, staminodes present, stigmas bifid; fruit broadly ellipsoid, 2.5-3 cm x 2 cm; seeds about 1.7 cm long, black, with minute caruncle. *J. curcas* frequently escapes from cultivation and may become naturalized. It grows on well-drained, well-aerated soils and is well-adapted to low fertility. It may be found on rocky slopes, dry river beds and similar habitats, from sea-level up to 1700 m altitude.

**Selected sources** 122, 190, 202, 284, 287, 332, 338, 418, 573, 580, 690, 899, 984, 1010, 1128, 1135, 1178, 1187, 1380, 1400, 1409, 1500, 1525, 1571.

***Jatropha gossypifolia* L.**

Sp. pl. 2: 1006 (1753).

**Synonyms** *Jatropha elegans* (Pohl) Klotzsch (1853).

**Vernacular names** Cotton-leaved physic nut

(En). Indonesia: jarak kosta merah (general), jarak ulung (Lampung, Sumatra), kaleke bacu (Kangean). Malaysia: jarak beremah, jarak hitam, jarak merah (Peninsular). Philippines: lansilansinaan (Tagalog), tagumbau-a-nalabaga (Iloko), tuba-tuba (Panay Bisaya, Cebu Bisaya). Laos: nhao luat. Thailand: sabuu daeng (central), sabu lueat, salot daeng (peninsular). Vietnam: d[aa]f[u lai ti[as].

**Distribution** *J. gossypifolia* is native to tropical America from Mexico to Paraguay and the West Indies. It has been introduced as an ornamental elsewhere in tropical America and many parts of the Old World tropics, especially West Africa and Malesia, and regularly escapes from cultivation.

**Uses** Seeds of *J. gossypifolia* are used as a cathartic. The Madurese use them after roasting, as a purgative for adults. The seed oil may be useful in treating leprosy. The latex from the leaves is applied on the tongue of babies to cure sores. In general, the latex is used externally to cure ulcers. It also has molluscicidal properties. In the Philippines, a cataplasm of fresh leaves is applied to swollen breasts. Leaves are also employed as a febrifuge in intermittent fevers, or boiled and used in a bath to cure fever. A decoction of the leaves is given as a purgative, as a stomachic, in the treatment of ear and venereal diseases, and as a blood purifier. Externally, leaves are applied to boils and carbuncles, eczema and itches. The roots are employed against leprosy, and a decoction of the bark is used as an emmenagogue.

**Observations** A small, somewhat succulent shrub up to 3 m tall; leaf blade deeply 3-5-lobed, 6-20 cm × 7-22 cm, base cordate, lobes broadly ovate to obovate, margin denticulate, with coarsely stipitate glandular hairs, petiole (2.5-4-8(-14) cm long, sparsely to densely set with coarse glandular hairs; inflorescence opposite a leaf, cymose, glandular hairy; male flowers with elliptical-lanceolate to ovate, about 2.5 mm long calyx lobes, petals free, about 3.5 mm long, purplish-red, stamens 8(-12), outer ones shorter than inner ones, filaments partly united; female flowers with calyx and petals as in male flower but twice as large, staminodes absent, stigmas capitate, bifid; fruit 3-lobed, roundish, about 1 cm in diameter; seeds about 7.5 mm long, pale greyish-brown, with a multifid caruncle. *J. gossypifolia* may be locally common in drier regions along roads, on waste places, in lawns, scrub vegetation, generally at low altitudes. In Asia all plants are referred to as var. *elegans* (Pohl) Muell. Arg. which differs from

var. *gossypifolia* in having denticulate and glandular-ciliate leaf margins.

**Selected sources** 83, 97, 122, 202, 284, 287, 332, 338, 618, 1128, 1135, 1178, 1187, 1380.

### ***Jatropha multifida* L.**

Sp. pl. 2: 1006 (1753).

**Synonyms** *Adenoropium multifidum* (L.) Pohl (1827), *Jatropha janipha* Blanco (1837).

**Vernacular names** Coral plant, French physic nut, Spanish physic nut (En). Arbre corail, médicinier d'Espagne, noisetier purgatif (Fr). Indonesia: jarak cina (Javanese), jarak gurita (Sundanese), balacai batai (Ternate). Philippines: mana (Filipino), tubang amerikano (Bikol). Cambodia: lohong khvang kraham. Thailand: fin ton, malako farang (Bangkok), ma hung daeng (north-ern). Vietnam: d[aa]f[u m[ef] d[or], b[aj]ch ph[u]j t[uwr].

**Distribution** *J. multifida* is native to tropical America from Mexico to Paraguay; it was introduced as an ornamental into the Old World tropics long ago. Within Malesia it is cultivated at least in Peninsular Malaysia, Java, the Moluccas and the Philippines.

**Uses** The seed oil of *J. multifida* is sometimes used as a cathartic, although it may cause strong irritation and even poisoning. The oil is applied both internally and externally as an abortifacient. Seeds are used fresh as a purgative and emetic. The latex is used externally in the treatment of infected wounds, ulcers, skin infections and scabies. In Indonesia one of its local names is 'yodium' ('iodine'), which reflects the popular use as a wound-healing remedy. In Indo-China, dried roots are given as a decoction against indigestion and colic; they are also prescribed as a tonic to treat orchitis and oedemas. The leaves are used as a purgative, and in the treatment of dysentery and scabies. *J. multifida* is widely grown, often in hedges, for its ornamental foliage and flowers. The tuberous roots can be eaten after roasting. The plant may also be used as a fish poison.

**Observations** A glabrous, slightly succulent shrub or small tree up to 6 m tall with white latex, roots tuberous; leaf blade peltate, broadly ovate-orbicular, 15-35 cm in diameter, palmatipartite with (9-)11(-12) segments, segments simple to deeply pinnatipartite, petiole 10-25(-35) cm long; inflorescence tightly corymb-shaped, peduncle up to 23 cm long; male flowers with broadly ovate calyx lobes, about 1 mm long, petals free, about 5 mm long, scarlet, stamens 8, filaments free; female flowers with petals up to 9 mm long, stigmas



capitate, bilobed; fruit tardily dehiscent to subdrupaceous, broadly obovoid, about 3 cm long, 3-lobed with keeled lobes, yellow when mature; seeds 1.7–2 cm long, buff, mottled brownish, with a small caruncle. *J. multifida* occasionally escapes from cultivation into grassland and thickets, generally at low altitudes.

**Selected sources** 97, 122, 202, 287, 332, 338, 771, 772, 1035, 1128, 1178, 1187, 1380, 1501.

S. Susiarti, E. Munawaroh & S.F.A.J. Horsten

## Justicia L.

Sp. pl. 1: 15 (1753); Gen. pl. ed. 5: 10 (1754).

ACANTHACEAE

$x = 7, 8, 9, 11, 13, 14, 15, 16, 17, 18$ ; *J. adhatoda*:  $2n = 30, 34, 40, 46, 50, 56, 58$ , *J. gendarussa*:  $2n = 28, 30, 32$ , *J. procumbens*:  $2n = 18, 28, 36$

**Major species** *Justicia adhatoda* L., *J. gendarussa* Burm.f., *J. procumbens* L.

**Origin and geographic distribution** *Justicia* is a large genus with somewhere between 300 and 600 species. It is distributed in all tropical and subtropical regions, with some species extending into temperate regions (e.g. in China and Japan).

**Uses** Leaf and root extracts of *Justicia* are commonly used in traditional medicine throughout South-East Asia, India and China, particularly for treating bronchitis, asthma, cough, fever and jaundice. Fresh leaves are often applied as a topical, poultice or lotion to treat swellings, skin eruptions, rheumatism and as a sedative. In India, *J. adhatoda* has a considerable reputation for its anti-allergic and anti-asthmatic properties.

*J. adhatoda* and *J. gendarussa* are often planted in hedges or as ornamental; the wood of the former is used as firewood.

**Properties** Several lignans (e.g. justicidins A–H, diphyllin, diphyllin apioside, diphyllin apioside-5-acetate, justicidiniosides A–C, neojusticin A–B, taiwanin E, taiwanin E methyl ether) have been identified from *J. procumbens*. Neojusticin A and B, taiwanin E methyl ether and taiwanin E significantly inhibit platelet aggregation. The methanolic extract of the herb has been found to have significant inhibitory activity in vivo against P-388 lymphocytic leukaemia in mice, as well as in vitro cytotoxicity in the 9-KB (human nasopharyngeal carcinoma) cell culture assay. Justicidin A and diphyllin were demonstrated to be the active compounds. Justicidin A and B, diphyllin, diphyllin apioside and diphyllin apioside-5-acetate showed strong antiviral activity against vesicular

stomatitis virus and low cytotoxicity against cultured rabbit lung cells. Justicidin A and B caused respectively 100% and 90% mortality of fourth-instar larvae of *Bombyx mori* after 6 days of feeding at 20 ppm. Several justicidins have piscicidal activity. Methods for synthesis of justicidin B and E and diphyllin have been described. The lignan justisolin and the lignan glucoside simplexoside, exhibiting growth regulatory properties in plants, have also been isolated from *J. procumbens*. Simplexoside produced a weak depressant action on the central nervous system in mice and rats, whereas the free lignans produced a stimulant action on the central nervous system.

The alkaloids found in *J. adhatoda* include vasicine, vasicinone, vasicinine, vasicinolone, adhatodane, adhatodine, adhavasacinone and anisotine. The yield of vasicine in the leaves ranges from 0.5–2% on dry weight basis. Comprehensive pharmacological investigations have been reported on drugs from *J. adhatoda*. A combination of the two alkaloids vasicine and vasicinone showed bronchodilator activity comparable to that of theophylline and greater than that achieved with each alkaloid separately. The mechanism of action is thought to be anticholinergic. A hitherto unidentified alkaloid showed pronounced protection against allergen-induced bronchial obstruction in guinea-pigs (10 mg/ml aerosol). Vasicine exhibits strong respiratory stimulant activity, moderate hypotensive activity and cardiac-depressant effect; vasicinone does not have these activities. Vasicine has been reported to lower blood pressure. The benzylamines bromhexine and ambroxol, semi-synthetic derivatives of vasicine and widely used as mucolytics, have a pH-dependent growth-inhibiting effect on *Mycobacterium tuberculosis*; they have a potentially useful adjunctive function in the therapy of tuberculosis. The leaf juice of *J. adhatoda* showed activity against some strains of e.g. *Bacillus subtilis*, *Staphylococcus epidermidis* and *Salmonella typhosa*. In rats fed for 10 days following insemination a leaf extract was 100% abortive at a dose equivalent to 175 mg/kg of starting dry material. The alcohol extract of dried leaves showed atropine-like activity due to the presence of a non-nitrogenous and non-toxic principle, vasakin. Vasakin exhibits hypoglycaemic activity, but is less potent than tolbutamide. The oil, which is present in small amounts e.g. in the root, showed slight insecticidal activity against the stored-grain pests *Bruchus chinensis*, *Sitophilus oryzae*, *Rhizopertha dominica*, *Stegobium paniceum* and *Sitotroga cerealella*, but it displayed su-

perior activity against freshly moulted fifth-instar nymphs of the red cotton bug *Dysdercus koenigii*. The leaf powder did not afford protection against attack of *Callosobruchus chinensis* on stored pulses. Larvae of the noctuid *Spodoptera littoralis* fed with *J. adhatoda* leaves showed high mortality. Powdered preparations of the leaves combined with burning charcoal to produce smoke repelled the mosquitoes *Armigeres subalbatus* and *Culex quinquefasciatus*. A 10% alcoholic water extract was effective in decreasing the severity of powdery mildew (caused by *Phyllactinia corylea*), leaf spot (caused by *Pseudocercospora mori*) and leaf rust (caused by *Cerotelium fici*) in mulberry (*Morus* spp.). Chromatographic analysis revealed the presence of caffeic, ferulic, vanillic, p-coumaric, p-hydroxybenzoic and tannic phenolic acids in aqueous extracts of *J. adhatoda* for which allelopathic activities are known.

Four simple o-disubstituted aromatic amines have been isolated from the leaves of *J. gendarussa* and characterized as 2-amino benzyl alcohol, 2-(2'-amino-benzylamino) benzyl alcohol and their respective O-methyl ethers.  $\beta$ -Sitosterol (a phytosterol) has also been reported. A leaf infusion of *J. gendarussa* showed analgesic effect in tests with mice. Fungicidal activity was reported for a crude extract; the extract was quite effective against *Rhizoctonia*-like isolates. Aqueous extracts significantly inhibited aflatoxin production by *Aspergillus parasiticus* in rice, wheat, maize and groundnut.

Justicidin B has also been isolated from the Central and South American *J. pectoralis* Jacq., which is considered as wound-healing in Jamaica. It also contains coumarin and umbelliferone, which relax smooth muscles and is used as snuff. The Mexican *J. spicigera* Schltdl. showed high trophozoite mortality and had a clear in vitro anti-giardiasis effect, better than tinidazol commonly used in the treatment of giardiasis. In animal tests, extracts from the South American *J. cydoniifolia* (Nees) Lindau showed anti-inflammatory activity equipotent to aspirin. The alkaloid 7-methoxyvasicine exhibited anti-inflammatory properties.

**Adulterations and substitutes** In India, leaves of *J. adhatoda* are sometimes adulterated with those of *Ailanthus excelsa* Roxb.

**Description** Erect shrubs or herbs, sometimes procumbent or ascending. Leaves opposite-decussate, simple, ovate to elliptical or linear-lanceolate, usually entire, often with cystoliths, petiole, exstipulate. Inflorescence a terminal or axillary congested spike or panicle, sometimes a thyr-

soid cyme or flowers solitary; bracts small and linear or subulate to large and foliaceous, bracteoles 2, linear-lanceolate, occasionally absent. Flowers sessile or pedicellate, bisexual, zygomorphic, white, pink or purple; calyx deeply 4-5-lobed, persistent; corolla with often narrow, funnel-shaped, straight or incurved tube, bilabiate, upper lip narrow, entire to bifid, lower lip broad, patent or recurved, 3-lobed; stamens 2, slightly exerted, with filiform filaments attached near the middle of the tube, anthers dorsifixed, with 2 usually unequal thecae, one or both thecae apiculate or calcarate; ovary superior, 2-locular, each locule with 2 ovules, style simple, filiform, with minute unequally 2-lobed stigma. Fruit an oblong, ovoid or obovoid-clavate capsule, distinctly stiped, bivalved and elastically and loculicidally dehiscent from apex downwards, usually 4-seeded. Seeds orbicular-lenticular, compressed, rough, tuberculate or smooth, on curved retinacula.

**Growth and development** The flowers of *Justicia* are insect-pollinated, mainly by bees, butterflies and flies. *J. adhatoda* and *J. gendarussa* rarely produce fruits in Java, but the latter apparently fruits regularly in the Philippines.

**Other botanical information** Taxonomical studies on *Justicia* have followed one of two divergent trends, resulting either in the recognition of a large number of small segregate genera, or in the adoption of a very broad definition of *Justicia*. Here, the concept of *Justicia* sensu lato is adopted. As a result, each species treated here is found in the literature under several names: *J. adhatoda* is often considered as belonging to the split genus *Adhatoda*, *J. gendarussa* to *Gendarussa*, and *J. procumbens* to *Rostellaria* or *Rostellularia*. *Justicia* is related to *Rungia*, but differs in the usually not rising placenta; when it does rise (e.g. in *J. gendarussa*), then both bracts at each node are fertile.

**Ecology** In South-East Asia, *J. procumbens* occurs in open locations, such as grasslands and roadsides, whereas *J. gendarussa* prefers more shady places such as forest, forest borders or along streams.

**Propagation and planting** Cuttings taken from terminal shoots of *J. gendarussa* root easily when planted in sand, particularly when treated with auxin (e.g. naphthalene acetic acid or indole butyric acid). Cuttings taken from lateral shoots by systematically decapitating apical meristems of growing shoots, rooted earlier and produced more roots than normal cuttings. Rooting of *J. adhatoda* stem cuttings in India is usually satisfac-

tory, with a maximum success of 90%. *J. procumbens* is a short-living herb which reproduces freely by seed.

**Husbandry** *Justicia* is rarely planted for medicinal purposes. In India, *J. adhatoda* is reported to be suitable for intercropping with poplar (*Populus* sp.).

**Diseases and pests** *Fusarium* spp. may attack *J. gendarussa*. A rust disease caused by *Puccinia thwaitesii* is reported from Malaysia for *J. gendarussa*, resulting in premature drop of infected leaves. A graft-transmissible disease, possibly of mycoplasma origin, and resulting in witches' broom symptoms has been described from India. In India, *J. adhatoda* is host for root-knot nematodes (*Meloidogyne javanica* and *M. incognita*) and for fungi such as *Phyllosticta ribiseda* and *Phomopsis pustulata*. The most common fungal disease is greasy rust caused by *Chnoospora butleri*.

**Genetic resources and breeding** All *Justicia* species described here have a large area of distribution, either naturally or as a result of cultivation, and do not seem to be at risk of genetic erosion. However, as some *Justicia* species rarely fruit in at least part of Malesia (e.g. *J. adhatoda* and *J. gendarussa* in Java) and are consequently propagated vegetatively, the genetic base is probably comparatively small in this region. There is no known selection and breeding work for medicinal purposes.

**Prospects** The known active compounds of *Justicia* are quite different for each species, but all species described here deserve special attention. The alkaloids and their derivatives from *J. adhatoda* show promising activity as bronchodilator, and have a potentially adjunctive function in the treatment of tuberculosis. Vasicine is reported as a promising uterotonic abortifacient, and may prove useful for the control of postpartum haemorrhage. It acts as a cholagogue and may be employed in some types of jaundice. Moreover, the insecticidal activity suggests some promise against insect pests of crops and mosquitoes transferring malaria. Some lignans from *J. procumbens* showed antitumour and antiviral activity, but more research is needed to establish their value for modern medicine. The fungicidal activity of *J. gendarussa* extracts warrants more research.

**Literature** [1] Asano, J., Chiba, K., Tada, M. & Yoshii, T., 1996. Antiviral activity of lignans and their glycosides from *Justicia procumbens*. *Phytochemistry* 42(3): 713-717. [2] Council of Scientific and Industrial Research (various editors), 1985.

The wealth of India. Revised Edition. Vol. 1. Publications and Information Directorate, New Delhi, India. pp. 76-79. [3] Ferdous, A.J., Islam, S.N., Ahsan, M. & Faroque, A.B.M., 1990. Antibacterial activity of the leaves of *Adhatoda vasica*, *Calotropis gigantea*, *Nerium odorum* and *Ocimum sanctum*. *Bangladesh Journal of Botany* 19(2): 227-229. [4] Fukamiya, N. & Lee, K.-H., 1986. Antitumor agents, 81. Justicidin-A and diphyllin, two cytotoxic principles from *Justicia procumbens*. *Journal of Natural Products* 49(2): 348-350. [5] Graham, V.A.W., 1988. Delimitation and infrageneric classification of *Justicia* (Acanthaceae). *Kew Bulletin* 43(4): 551-624. [6] Grange, J.M. & Snell, N.J., 1996. Activity of bromhexine and ambroxol, semi-synthetic derivatives of vasicine, against *Mycobacterium tuberculosis* in vitro. *Journal of Ethnopharmacology* 50(1): 49-53. [7] Hansen, B., 1989. Notes on Southeast Asian Acanthaceae 1. *Nordic Journal of Botany* 9(2): 209-216. [8] Malik, K.A. & Ghafoor, A., 1988. Acanthaceae. In: Nasir, E. & Ali, S.I. (Editors): *Flora of West Pakistan*. No 188. Department of Botany, University of Karachi and National Herbarium, Pakistan Agricultural Research Council, Islamabad, Pakistan. pp. 29-44. [9] Okigawa, M., Maeda, T. & Kawano, N., 1970. The isolation and structure of 3 new lignans from *Justicia procumbens* var. *leucantha*. *Tetrahedron* 26(18): 4301-4305. [10] Wagner, H., 1989. Search for new plant constituents with potential antiphlogistic and antiallergic activity. *Planta Medica* 55(3): 235-241.

#### *Selection of species*

#### **Justicia adhatoda L.**

Sp. pl. 1: 15 (1753).

**Synonyms** *Adhatoda zeylanica* Medic. (1790), *Adhatoda vasica* Nees (1832).

**Vernacular names** Malabar nut tree (En). Thailand: kra niat (peninsular), kulaa khao (northern), saniat moraa (central). Vietnam: xu[aa]n ti[ees]t.

**Distribution** Possibly a native of India, but now much cultivated, e.g. in Java and Thailand.

**Uses** The roots and leaves are widely used in the Ayurvedic and Unani systems of medicine in India and Thailand for treating bronchitis, asthma, fever and jaundice. *J. adhatoda* is often planted in hedges, and the wood is used as firewood. In India, it is also grown as a green manure in rice, tobacco and tea. Boiled tender shoots are eaten with salt locally in India.

**Observations** An erect, much branched shrub up to 4(–6) m tall, stems quadrangular to nearly terete; leaves lanceolate-elliptical, 8–25 cm × 2.5–8 cm, with (1–)1.5–3.5(–4.5) cm long petiole; flowers in spikes with leafy, broadly elliptical, 1.5–4 cm long bracts, 3–4.5 cm long, white; fruit broadly clavate, about 2.5 cm long, pubescent. *J. adhatoda* is planted in Java up to 1400 m altitude.

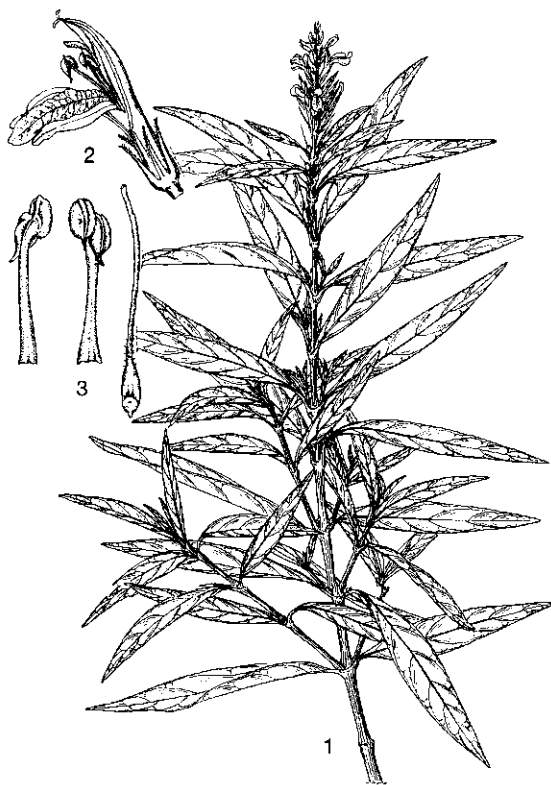
**Selected sources** 97, 226, 288, 369, 410, 502, 1012, 1013, 1126, 1535.

### ***Justicia gendarussa* Burm.f.**

Fl. indica: 10 (1768).

**Synonyms** *Gendarussa vulgaris* Nees (1832).

**Vernacular names** Indonesia: gandarusa (general), besi-besi (Aceh), kawo (Seram). Malaysia: gandarusa, temenggong melela, urat sugi (Peninsular). Philippines: kapanitulot (Tagalog), bunlao (Bisaya), tagpayan (Iloko). Thailand: Chiang phraa mon (central), pong dam (Trat), kraduuk kaidam (northern). Vietnam: t[aa]f[n] c[uw]r[u], thu[oo]s[c] tr[aw]j[c], t[aa]f[n] g[iao].



*Justicia gendarussa* Burm. f. – 1, flowering twig; 2, flower; 3, stamens and pistil.

**Distribution** Pakistan, India, Sri Lanka, Indo-China, China, Thailand, Peninsular Malaysia, Java, the Moluccas and the Philippines; possibly a native of China, but now much cultivated and naturalized.

**Uses** An extract of the leaves or young shoots is used as an emetic in coughs and asthma in the Philippines, whereas fresh leaves are applied as topical to cure oedema of beri-beri and rheumatism. A decoction of the leaves is used for bathing during childbirth. In Malaysia, the leaves are much applied for poulticing to treat headache and pains, as a lotion to treat swellings and rheumatism, and in a bath after confinement, the roots for treating thrush and cough. The leaves are also used in preparations to treat gonorrhoea, amenorrhoea and malaria. In Indonesia, the leaves are used to treat headache, rheumatism and pain. In Vietnam, the leaves are applied externally, as a poultice, decoction or tincture, to treat rheumatic arthritis and swellings. In Thailand, the roots are used against diuresis, diarrhoea and as antivenin; the bark is used as antipyretic, anticough, diuretic and anti-amoebic, in the treatment of wounds and allergy; the leaves are taken internally against cough, fever and as a cardiotonic, and used externally to treat inflammation, wounds and allergy. Numerous medicinal uses are recorded from India and China; the roots are used to treat rheumatism, dysuria, fever, carbuncles, jaundice and diarrhoea, the leaves as a diaphoretic and febrifuge and to treat lumbago, amenorrhoea, swellings, coughs, asthma, colics, eczema, cephalalgia, hemiplegia, facial paralysis, earache and hemicrania, and the bark as emetic. Magical uses are reported for Indonesia, Malaysia and the Philippines. *J. gendarussa* is cultivated as an ornamental, and is often used for living fences.

**Observations** An undershrub up to 150 cm tall, stems terete, young twigs usually dark purple; leaves linear-lanceolate, 5–20 cm × 1–3.5 cm, with up to 1 cm long petiole; inflorescence a spike, bracts lanceolate, about 4 mm long; flowers 1.5–2 cm long, white with purplish streaks and spots inside; fruit clavate to ellipsoid, about 1.3 cm long, glabrous. *J. gendarussa* is cultivated, and naturalized in forest, forest borders and on river banks in Java, up to 1500 m altitude. In the Philippines, it often grows along streams in primary and secondary forest.

**Selected sources** 97, 202, 332, 580, 909, 1012, 1035, 1126, 1178, 1366, 1572.

**Justicia procumbens L.**

Sp. pl. 1: 15 (1753).

**Synonyms** *Justicia japonica* Thunb. (1784), *Justicia simplex* D. Don (1825), *Rostellularia procumbens* (L.) Nees (1832).

**Vernacular names** Vietnam: t[uw][ows]s[af]ng.

**Distribution** Pakistan, India, Sri Lanka, Burma (Myanmar), Indo-China, China, Taiwan, Japan, Peninsular Malaysia, the Philippines, Timor, possibly Australia; probably also Java.

**Uses** In the Philippines, the leaves are used externally as an astringent to cure certain eruptions of the skin. In India, the herb is considered as an alterative, expectorant, laxative and diuretic, and an infusion or decoction is used to treat asthma, coughs, rheumatism, backache and flatulence. The juice from the leaves is used to treat ophthalmia. In Chinese medicine, whole plants are used to treat fever, pain due to pharyngo-laryngeal swelling and cancer.

**Observations** A herb up to 50 cm tall, with erect or procumbent to ascending, quadrangular stems, often diffusely branched; leaves elliptical-ovate to lanceolate-elliptical, 1–5 cm × 0.5–2.5 cm, with up to 1(–1.5) cm long petiole; inflorescence a spike; bracts obovate to elliptical-ovate or linear-lanceolate, 4–5 mm long; flowers 8–10 mm long, pink or purplish-pink; fruit oblong, 3–5 mm long, hairy to glabrescent. *J. procumbens* and its related species are in need of a thorough taxonomical revision covering the complete area of distribution. Several closely related species are reported from Malesia and Australia. These might be conspecific. *J. procumbens* occurs in grasslands and roadsides, in Japan also in forest, up to 1600 m altitude.

**Selected sources** 77, 202, 241, 440, 623, 1012, 1079, 1126, 1178.

H. Sangat-Roemantyo

**Kaempferia L.**

Sp. pl. 1: 2 (1753); Gen. pl. ed. 5: 3 (1754).

ZINGIBERACEAE

$x = 11, 12, 13, 14$ ; *K. angustifolia*:  $2n = 36$ , *K. galanga*:  $2n = 54$ , *K. rotunda*:  $2n = 44, 45, 54$

**Major species** *Kaempferia galanga* L., *K. rotunda* L.

**Vernacular names** Galangal, kaempferia (En).

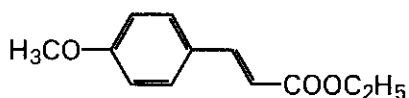
**Origin and geographic distribution** *Kaempferia* comprises about 40 species and occurs in India and Sri Lanka to Indo-China, Thailand and

Malesia east to the Moluccas. Within Malesia some 7 species are present; Thailand has 15 species.

**Uses** Rhizomes of various *Kaempferia* species are credited with stimulant, stomachic, carminative and similar properties, and are also used throughout South-East Asia to relieve headache. They are commonly applied in the treatment of abdominal disorders, apparently for their astringent properties. In Peninsular Malaysia and Indonesia the leaves and rhizomes are chewed as an expectorant for coughs and sore throat, or pounded and used in poultices or lotions; they are often an ingredient of children's medicines and tonics. In Indonesia, rhizomes of *Kaempferia* are a common ingredient of a multitude of 'jamus'. Rhizomes are applied externally as a 'drawing' agent on boils and, mixed with oil, as a cicatrizant. Leaves and rhizomes of *K. galanga* are applied externally as a poultice to treat abdominal pains, and as an embrocation or sudorific to treat swellings, inflammations, and muscular rheumatism. Pounded rhizomes are also applied to traumatic injuries and nose bleeding. Mixed with rice flour, they are used in 'beras kencur', a refreshing drink used as a sudorific. In the Philippines, the whole plant is applied as a remedy for cold. A decoction of rhizomes is a tonic, carminative, gargle, a remedy for dyspepsia and malarial chills, and given after childbirth. Rhizomes of *K. rotunda* and *K. angustifolia* are considered cooling. Rhizomes of *K. galanga* are well-known in Chinese medicine as a remedy for toothache or a wash to treat dandruff or scabs on the head.

Rhizomes of *K. galanga* and *K. rotunda* are also widely known as a flavouring for various dishes and rice; in Indonesia, leaves of *K. galanga* are used in a similar way. Rhizomes are also used in perfumery and as a means to preserve cloth from insects. In Indonesia, leaves of *K. galanga* and *K. rotunda* are eaten raw in 'lalab' or cooked as a vegetable. Dried and ground rhizomes of *K. galanga* are also an ingredient of cosmetics, especially powders. Furthermore, rhizomes of *K. galanga* are used as an ingredient of a fermentation agent and in the production of 'kretek' cigarettes. *K. rotunda*, and several other *Kaempferia* species, are often cultivated as an ornamental; the flowers of *K. rotunda* are fragrant. In Vietnam, rhizomes of *K. angustifolia* are chewed as a masticatory together with betel nuts.

**Production and international trade** The estimated area planted with *K. galanga* in Boyolali (Central Java) and West Pasaman (West Suma-



ethyl-p-methoxy-trans-cinnamate

tra) in Indonesia is about 2000 ha, yielding at least 20 000 t of fresh rhizomes annually.

**Properties** The roots of *K. galanga* contain 2.4–3.9% of an essential oil whose main constituents are about 25% ethyl cinnamate (cinnamic acid ethyl ester) and up to 30% methyl-p-methoxycinnamate (p-methoxycinnamic acid methyl ester).

The chloroform extract of *K. galanga* rhizome has been found to inhibit tonic contractions in the rat aorta induced by high  $K^+$  concentrations or phenylephrine, in a dose-dependent manner. It was suggested that the chloroform extract inhibits vascular smooth muscle contraction by inhibiting  $Ca^{2+}$  influx through both voltage and receptor-operated non-selective cation channels, and  $Ca^{2+}$  sensitivity of contractile elements. Ethyl-p-methoxy-trans-cinnamate isolated from *K. galanga* rhizomes, acted as a mono amine oxidase-inhibitor; such inhibitors are sometimes used as drugs to treat depression. In experiments on guineapigs, ethyl-p-methoxy-trans-cinnamate caused trachea and tracheal branches to dilate. This finding supports its traditional use in the treatment of asthma.

Furthermore, the ethanol extract of *K. galanga* showed cytotoxicity against human cervical carcinoma (HeLa) cells ( $CD_{50}$  10–30  $\mu\text{g/ml}$ ). *K. galanga* extracts showed amoebicidal activity in vitro against 3 *Acanthamoeba* species, the causative agents of granulomatous amoebic encephalitis and amoebic keratitis.

The larvicidal principles obtained from the methanolic extract of rhizomes of *K. galanga* are ethyl cinnamate, ethyl-p-methoxycinnamate, and p-methoxycinnamic acid. Two active metabolites, benzyl benzoate and the cyclohexane derivative crotexoxide, have been isolated from the rhizome of *K. rotunda*. Crotexoxide was active in the chronic feeding bioassay of larvae of *Spodoptera littoralis* ( $LC_{50}$  5.6  $\mu\text{g/cm}$ ). Finally, *K. galanga* also exhibits antifungal activity.

**Description** Perennial, rhizomatous, short-stemmed herbs; rhizome sympodial, tuberous or cylindrical, fleshy; roots often bearing small tubers. Leaves usually few, distichous, simple, lower ones sheathing; sheath auriculate at apex; ligule small or absent; petiole short; blade usually broad,

with fan-like venation. Inflorescence terminal; peduncle usually enclosed by the imbricating leaf-sheaths. Flowers arranged spirally on a discoid receptacle (the reduced rachis), solitary in the axil of a bract, bisexual, zygomorphic; bracteoles 1 or 2, small, 2-topped to divided almost to the base; calyx tubular, split for a short distance, unequally toothed, usually much shorter than the corolla tube; corolla tubular at base, with 3 subequal, usually linear lobes; staminodes 3, petaloid, the anterior one, called labellum, nearly flat, deeply bilobed, usually white or lilac, sometimes with a different colour towards the base, the 2 lateral ones spreading, often similar to the halves of the labellum; fertile stamen 1, filament very short or absent, anther not or hardly exerted, thecae parallel, dehiscing with longitudinal slits, connective (also called anther-crest) usually large, entire or lobed, often reflexed and filling the throat of the flower but not enfolding the style; ovary inferior, 3-locular with axillary placentation and few to many ovules, style 1, filiform with funnel-shaped stigma, held between the thecae. Fruit a thin-walled, dehiscent capsule. Seeds few to many, ellipsoidal to nearly globular, with a lacerate aril.

**Growth and development** In Java, *K. angustifolia* flowers from October to January and in April, *K. galanga* in November and December, and *K. rotunda* in April and September to November. *K. galanga* can be induced to flower by a period of drought. Each flower opens for only one day. Pollination is by ants, flies and bees. Fruits are seldom formed.

**Other botanical information** *Kaempferia* belongs to the tribe *Hedychieae* and is closely related to *Boesenbergia*, *Scaphochlamys* and *Haplochorema*, from which it differs by its variably deeply 2-lobed bracteoles. All African species formerly included in *Kaempferia* have been attributed to a distinct genus called *Siphonochilus*. They differ amongst others in the position of their lateral inflorescences, absence of bracteoles and structure of the labellum.

In *Kaempferia* flowers the 'petal-like' organs, the labellum and 2 lateral staminodes, are modified stamens that function as petals, a situation called homeosis. There seem to be two forms within *K. galanga*, a broad-leaved one and a narrow-leaved one. Both have been reported from Cambodia, Thailand and Java.

About 10 cultivars of *K. galanga* are currently distinguished in Indonesia with 'Cileungsi Besar' the highest yielder.

**Ecology** Most *Kaempferia* species occur natu-

rally in open forest, forest margins, and teak forest, at low altitudes. They easily escape from cultivation and can then be found in grassy waysides, lawns and waste places. They occur in both ever-wet and seasonal climates. *K. galanga* thrives best up to 1000 m altitude, in areas with an annual rainfall of 2000–4000 mm, but waterlogging causes damage. It can grow on various soils, from heavy vertisols to latosols and andosols, with clayey to silt-loamy texture, but it develops poorly on acid soils. *K. galanga* and *K. rotunda* grow best in fertile garden soil in shaded conditions.

**Propagation and planting** *Kaempferia* can be propagated by rhizome pieces 2.5–4 cm long, with at least 2–3 buds; both the younger and older parts of rhizomes may be used. Rhizome cuttings may be kept for 1–2 weeks under dry, shaded conditions prior to planting at the beginning of the rainy season. *K. galanga* and *K. rotunda* can also be propagated by tissue culture using rhizome explants with vegetative buds. In Central Java *K. galanga* is usually planted in furrows 7.5–10 cm deep at 10–15 cm spacing. In West Sumatra it is planted in separate planting holes 5–7.5 cm deep at a spacing of 25–60 cm. In Java *K. galanga* is usually intercropped with groundnut and rice.

**Husbandry** In *K. galanga* application of 15 t/ha manure proved to be most effective in increasing rhizome yield; higher doses, however, promoted the development of aerial parts only. It loses its leaves towards the east monsoon period and it is recommended to dig up the rhizome then to prevent fungal decay.

**Diseases and pests** *Pythium* sp. has been observed in Central Java to cause rhizome soft rot of *K. galanga*, particularly under poor drainage conditions.

**Harvesting** In Central Java *K. galanga* is harvested 6–18 months after planting, in West Sumatra after 18–24 months.

**Yield** Fresh rhizome yields of *K. galanga* in Central Java are 6–15 t/ha, compared with 12–20 t/ha in West Sumatra. Yields as low as 1.2 t/ha have also been reported.

**Handling after harvest** Rhizomes of *K. galanga* can be washed, sliced and dried for storage.

**Genetic resources and breeding** There are no germplasm collections and breeding programmes of *Kaempferia*.

**Prospects** In Indonesia most rhizomes of *K. galanga* are obtained from the wild. Since it is expected that *K. galanga* will be increasingly used in the rapidly developing Indonesian industries of traditional medicine and cosmetics, cultural prac-

tices need to be developed to meet this demand. In Thailand, *K. galanga* rhizomes are collected from the wild and from small-scale cultivation and exported to China. Further agronomic research may lead to market expansion.

**Literature** |1| Holttum, R.E., 1950. The Zingiberaceae of the Malay Peninsula. Gardens' Bulletin, Singapore 13: 117–123. |2| Juhaeti, T., Chairul & Harapini, M., 1992. Pertumbuhan, produksi dan kandungan komponen utama kencur yang ditanam pada musim kemarau [Growth, production and major constituents of the rhizomes of kencur grown during the dry season]. In: Nasution, R.E. et al. (Editors): Prosiding Seminar Hasil Penelitian dan Pengembangan Sumber Daya Hayati 1991/1992, Bogor, 6 Mei 1992 [Proceedings of the Seminar on Results of Research and Development in Biological Resources 1991/1992, Bogor, 6 May 1992]. Bogor, Puslitbang Biologi-LIPI. pp. 354–359. |3| Kiuchi, F., Nakamura, N., Tsuda, Y., Kondo, K. & Yoshimura, H., 1988. Studies on crude drugs effective on visceral larva migrans II. Larvicidal principles in *Kaempferia* Rhizoma. Chemical and Pharmaceutical Bulletin 36(1): 412–415. |4| Mackeen, M.M. et al., 1997. Antimicrobial and cytotoxic properties of some Malaysian traditional vegetables (ulam). International Journal of Pharmacognosy 35(3): 174–178. |5| Mustafa, M.R., Mustafa, A.M. & Hashim, S., 1996. Vasorelaxant effects of the chloroform extract of *Kaempferia galanga* on smooth muscles of the rat aorta. Asia Pacific Journal of Pharmacology 11(3–4): 97–101. |6| Noro, T. et al., 1983. Mono amine oxidase inhibitor from the rhizomes of *Kaempferia galanga*. Chemical and Pharmaceutical Bulletin 31(8): 2708–2711. |7| Nugroho, B.W., Schwarz, B., Wray, V. & Proksch, P., 1996. Insecticidal constituents from rhizomes of *Zingiber cassumunar* and *Kaempferia rotunda*. Phytochemistry 41(1): 129–132. |8| Perry, L.M., 1980. Medicinal plants of East and Southeast Asia. Attributed properties and uses. The MIT Press. Cambridge, Massachusetts, United States & London, United Kingdom. p. 442. |9| Sadikun, A., Asmawi, A.Z., Ean, U.L. & Rahman, A.A., 1987. The effect of *Kaempferia galanga* rhizome extract on superfused guinea pig tracheal chain and ileum preparation and mice hind paw edema. Proceedings of the 4th Annual Seminar of the Natural Products Group. pp. 2–12. |10| Sudiarto, 1989. *Kaempferia galanga* L. in Central Java and West Sumatra. In: Siemonsma, J.S. & Wulijarni-Soetjipto, N. (Editors): Plant Resources of South-East Asia. Proceedings of the first PROSEA International Sym-

posium, May 22–25, 1989, Jakarta, Indonesia. P. doc, Wageningen, the Netherlands. pp. 306–308.

*Selection of species*

***Kaempferia angustifolia* Roscoe**

Trans. Linn. Soc. London 8: 351 (1807).

**Synonyms** *Kaempferia roxburghiana* Schult. (1822), *Kaempferia undulata* Teijsm. & Binnend. (1855) non Link, *Kaempferia gilbertii* W. Bull. (1882).

**Vernacular names** Indonesia: kunci kunot (Sundanese), kunci menir (Malay, Jakarta), kunci pepet (Javanese). Laos: 'van<sup>2</sup> toup 'moup. Thailand: thao nhang hang, prab samut. Vietnam: thi[ee]n li[ee]n li[ee]n li[as] h[e]p.

**Distribution** The eastern Himalayas, Laos, Vietnam, Thailand and Java; also cultivated in Java.

**Uses** The small roots and tubers of *K. angustifolia* have astringent properties and are used against dysentery and diarrhoea, but also to treat coughs; the watery main rhizome is considered cooling.

**Observations** A small herb; leaves 2–8, glabrous, sheaths 3–4 cm long, blade linear-lanceolate to oblong-elliptical, 7–20 cm × 1–10 cm; inflorescence emerging from between the leaves, sessile, with 10–12 flower buds, only a few of which develop; calyx 3–5 cm long, white, corolla white, tube up to 8.5 cm long, lobes up to 3.5 cm long, labellum obovate, 20–30 mm × 10–25 mm, incised to about one third, purple, other staminodes lanceolate to oblong-obovate, 18–37 mm long, white, fertile stamen about 10 mm long, connective shortly bilobed with acute lobes. *K. angustifolia* is found in teak forest, lowland rice, and on calcareous marl, up to 150 m altitude.

**Selected sources** 97, 455, 580, 1128, 1372, 1495, 1496.

***Kaempferia galanga* L.**

Sp. pl. 1: 2 (1753).

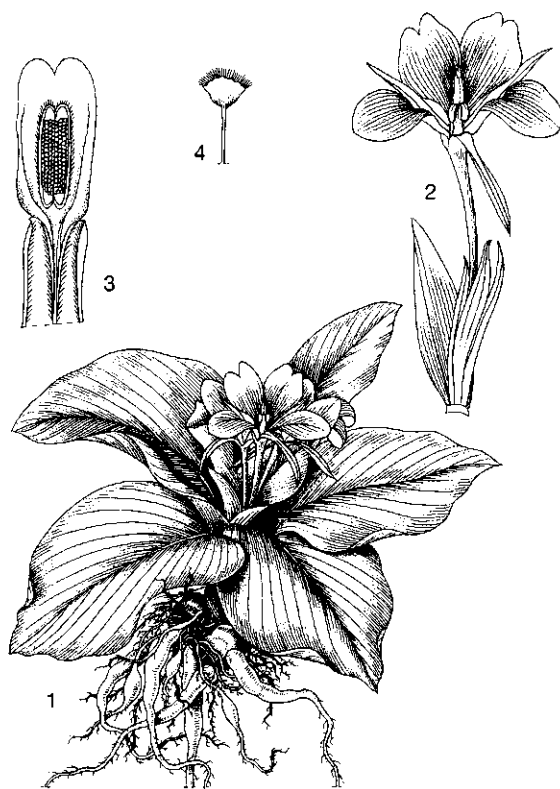
**Vernacular names** East-Indian galangal (En). Indonesia: kencur, cekur (general), bataka (North Sulawesi, Ternate, Tidore). Malaysia: cekur, cekur Jawa, cengkur (Peninsular). Philippines: gisol (general), disok (Iloko), dusol (Tagalog). Laos: 'van<sup>2</sup> 'hom. Thailand: hom proh (central), waan hom, waan teen din (northern). Vietnam: dia li[ee]n, s[ow]n nai, tam n[aj]i.

**Distribution** Possibly native only to India, where it is widespread, cultivated throughout

South-East Asia, including southern China, in Malesia east to the Moluccas, possibly also introduced in northern Australia.

**Uses** In Malaysia, rhizomes are used in traditional medicine for the treatment of high blood pressure, swellings, ulcers, sprains and asthma. Leaves and rhizomes are chewed to treat coughs and sore throat; they are also pounded and used in poultices and lotions applied to relieve many ailments. Rhizomes are an ingredient of post partum medicine and to treat common cold. Leaves and rhizomes are eaten fresh as a vegetable and used in cosmetic powder and as a food flavouring agent. In Indonesia, *K. galanga* is used in a similar way, to treat swellings, muscular rheumatism, wounds and as an antidote. It is a common ingredient of 'jamus'. In the Philippines, the whole plant is used as a remedy for common cold. Rhizomes are used to treat headache, dyspepsia and malarial chills. Rhizomes and leaves are used as a flavouring in food, rhizomes also in perfumery. The leaves are used as vegetable.

**Observations** A small herb; leaves usually



*Kaempferia galanga* L. – 1, plant habit; 2, flower; 3, apex of fertile stamen; 4, style apex.



2-3(-5), sheaths 1.5-5 cm long, blade often horizontal and appressed to the soil, broadly elliptical to suborbicular, 6-15 cm × (2-)5-10 cm, acuminate, glabrous above, arachnoid-hairy below; inflorescence emerging from between the leaves, sessile, 4-12(-15)-flowered; calyx 2-3 cm long, corolla white, tube 2.5-5 cm long, lobes 1.5-3 cm long, labellum broadly obovate, divided to about halfway or more, white or pale purple with violet to purple spots at base, each lateral lobe about 2-2.5 cm × 1.5-2 cm, other staminodes oblong-obovate to oblanceolate, 1.5-3 cm long, white, fertile stamen 10-13 mm long, connective deeply bilobed with reflexed lobes. *K. galanga* thrives best in slightly shaded places such as open forest, forest edges, and bamboo forest, on various soils, up to 1000 m altitude.

**Selected sources** 97, 190, 202, 261, 287, 350, 363, 455, 580, 615, 875, 1035, 1066, 1126, 1128, 1178, 1287, 1372, 1406, 1496.

### **Kaempferia rotunda L.**

Sp. pl. 1: 3 (1753).

**Synonyms** *Kaempferia longa* Jacq. (1798).

**Vernacular names** Round-rooted galangal (En). Indonesia: kunci pepet, kunir putih (Javanese), temu rapet (eastern Sumatra). Malaysia: kencur, kunyit putih, temu putih (Peninsular). Philippines: gisol na bilog (general). Thailand: waan nonlap (Chiang Mai), waan haa non (Ratchaburi), ueang din (northern). Vietnam: c[aa]r m dia la, ng[ar]i m[as]u.

**Distribution** Possibly native to Indo-China, but nowadays cultivated almost throughout tropical Asia, mainly as an ornamental but especially in South-East Asia also for medicinal purposes; regularly escaping from cultivation.

**Uses** In Indonesia, rhizomes are used to treat abdominal illness; the watery little corms are considered cooling. In the Philippines, rhizomes are used internally to treat gastric complaints, and externally, mixed with oil, as a cicatrizant. They are also used in perfumery and as a means to preserve cloth from insects. Leaves and rhizomes are eaten fresh or cooked as a vegetable and used in cosmetic powder and as a food flavouring agent.

**Observations** A small herb; leaves (2-)3-5, erect, petiolate, sheaths 7-24 cm long, blade oblong-lanceolate to elliptical, (7-)12-25(-36) cm × 4-7(-11) cm, gradually acuminate, glabrous above, puberulous below, often flamed or marked; inflorescence appearing before the leaves on stems with rudimentary leaves, on a well-developed peduncle, 4-16-flowered; calyx 3-7 cm long, white or

greenish, corolla white, interruptedly striped-punctate, tube 3.5-7 cm long, lobes 3.5-7 cm long, labellum obcordate, divided halfway or further, 4-7 cm × 2-4 cm, purple with yellowish midrib, other staminodes elliptical to linear, 3-5 cm long, white or lilac, fertile stamen 0.8-2.5 cm long, connective 2-4-lobed. *K. rotunda* grows well in teak forest, open lower montane forest, old bamboo forest, but also in open grassland, up to 1300 m altitude.

**Selected sources** 97, 202, 455, 580, 615, 875, 1066, 1126, 1128, 1178, 1372, 1496.

Halijah Ibrahim

### **Kalanchoe Adans.**

Fam. pl. 2: 248 (1763).

CRASSULACEAE

$x = 17, 18$ ; *K. laciniata*:  $2n = 34, 68$

**Major species** *Kalanchoe ceratophylla* Haw., *K. crenata* (Andrews) Haw.

**Vernacular names** Malaysia: sedingin, seringin (Peninsular).

**Origin and geographic distribution** *Kalanchoe* includes about 60 species, mainly from the Old World tropics and South Africa. Only a few species occur in South-East Asia. *K. crenata* is probably native to Africa, but since long naturalized in the drier parts of South-East Asia. *K. ceratophylla* is native to continental Asia and commonly cultivated in Malesia. *K. laciniata* is indigenous in Africa, southern India and Thailand.

**Uses** The uses of the various *Kalanchoe* species generally involve their antiseptic and cooling properties. Crushed leaves of *K. crenata* are applied to ulcers in the Philippines and Indo-China. In Taiwan, a decoction of *K. crenata* is employed as an eyewash and a styptic for contusions. In East and West Africa the juice of the leaves is applied to septic wounds, or is used to relieve headache. The roots and leaves are said to be toxic and are used as an abortifacient in some parts of East Africa.

The leaves of *K. ceratophylla* have styptic, astringent, antiseptic and cooling properties. In the Philippines, the pulped leaves are used to treat chronic ulcers and headache. In Indo-China, the leaves are used as topicals for ulcers. In Malaysia, the leaves are applied as a poultice on the chest for coughs and cold. In India, poulticed leaves of *K. ceratophylla* or their juice are applied to bruises and contusions to allay inflammation and prevent discoloration. The leaves are also used as a

styptic to cuts, abrasions and other wounds, and applied to insect bites. The plant is used internally to treat diarrhoea, dysentery, lithiasis, cholera and phthisis. In Malaysia, twigs are placed in houses to attract good spirits and drive away evil ones, and to ward off diseases, such as cholera. *K. laciniata* probably has similar uses to *K. ceratophylla*.

*K. ceratophylla*, *K. crenata* and *K. laciniata* are all grown as ornamentals too. Many other *Kalanchoe* species are also used as ornamentals, e.g. *K. blossfeldiana* Poelln., a well-known pot plant in Europe and the United States. In South-East Asia, *K. blossfeldiana* plants are sold as 'flower of longevity' in spring festivals.

**Properties** The leaves and flowers of *K. crenata* are reported to contain several flavonoids (quercetin, kaempferol, patuletin) and their glycosides (quercetin-3-O-glucoside-7-O-rhamnoside, kaempferol-3-O-rhamnoside, patuletin-3,7-di-O-rhamnoside). Several sterols (sitosterol, stigmasterol, campesterol) and triterpenoids (friedelin, taraxerol, glutinol) have also been isolated from the flowers of *K. crenata*.

Little is known about the biological activities of the *Kalanchoe* species described or their constituents. Kalambrosides A-C (patuletin-acetyldi-O-rhamnosides), isolated from *K. brasiliensis* Camb., were found to show potent inhibitory effects on PHA-induced human lymphocyte proliferation in vitro. In this assay the structurally related, but non-acetylated patuletin-3,7-di-O-rhamnoside (also isolated from *K. crenata*) was devoid of any activity. The ethanol extract of *K. laciniata* has also been found to exhibit cytotoxic effect against CA-9 KB cells. Furthermore, *K. crenata* is reported to cause poisoning in sheep, but the toxic principle is not known. Cotyledonosis (or 'krumpsiek') is a neurotoxic syndrome in animals and, like cardiac glycoside poisoning can be caused by *Crassulaceae*-bufadienolides e.g. isolated from *K. lanceolata* (Forssk.) Pers. (3-O-acetylhellebrigenin, lanceotoxin A, lanceotoxin B) or from several *Bryophyllum* species.

**Adulterations and substitutes** *Bryophyllum pinnatum* (Lamk) Oken has comparable medicinal uses.

**Description** Shrubs or shrublets up to 2 m tall, with branches spreading, fleshy but somewhat woody towards base, often regenerating from base. Leaves opposite, simple, from entire to crenate and pinnatifid, free or slightly fused at base, fleshy, persistent or deciduous. Inflorescence terminal, usually consisting of many-flowered corym-

bose or paniculate cymes, peduncle present or absent with gradual transition from leaves to shorter bracts below the flowers. Flowers bisexual, spreading or stiffly erect, 4-merous; calyx 4-partite; corolla with petals fused into a tube longer than the lobes; stamens 8 in 2 whorls, filaments glabrous and fused to corolla tube at about the middle, anthers usually included, with terminal appendage; ovary superior, consisting of 4 free carpels, gradually constricted into styles and with terminal stigmas. Fruit a many-seeded follicle. Seeds ellipsoidal, with a constriction and abruptly widening at blunt proximal end.

**Growth and development** All *Kalanchoe* species have the Crassulacean Acid Metabolism (CAM) pathway, enabling them to fix CO<sub>2</sub> at night and to photosynthesize with closed stomata during daytime, to minimize water loss. *K. crenata* flowers and fruits freely in South-East Asia.

**Other botanical information** In the past *Kalanchoe* was enlarged to incorporate other closely related genera, and then split up again. Some taxonomists include *Bryophyllum* in the genus, but here *Bryophyllum* is considered to be a separate genus. A recent taxonomical publication has resulted in nomenclatural changes affecting South-East Asian species. One such change is that the widespread Malaysian species known under the name *K. laciniata* is now called *K. ceratophylla*; the true *K. laciniata* is indigenous in India and Thailand. The Malaysian species known as *K. integra* is now called *K. crenata*.

**Ecology** *Kalanchoe* species are mainly found in dry rocky or sandy, sunny or slightly shaded locations. *K. ceratophylla* is grown in gardens or as a pot plant in lowland regions. *K. crenata* is found up to an altitude of 2000 m in sunny, stony or rocky sites, even on almost bare rocks. It is locally abundant in light *Casuarina* forest.

**Propagation and planting** *Kalanchoe* plants are easy to propagate from seed and cuttings.

**Diseases and pests** In Japan, *K. laciniata* is reported to be susceptible to grey mould caused by *Botrytis cinerea*.

**Harvesting** The leaves are plucked and used fresh.

**Genetic resources and breeding** The *Kalanchoe* species used medicinally are grown as ornamentals and, in general, the harvested material originates from such plants. The natural resource as such is therefore not directly threatened in the Malaysian region.

**Prospects** *Kalanchoe* species will remain of some importance for home consumption, as a

readily available traditional antiseptic. It is unlikely that this importance will increase in the near future.

**Literature** |1| Backer, C.A., 1951. Crassulaceae. In: van Steenis, C.G.G.J. (General editor): *Flora Malesiana*. Series 1, Vol. 4. Noordhoff-Kolff, Djakarta, Indonesia. pp. 197–202. |2| Burkill, I.H., 1966. A dictionary of the economic products of the Malay Peninsula. Revised reprint. Vol. 2. Ministry of Agriculture and Cooperatives, Kuala Lumpur, Malaysia. pp. 1297–1298. |3| Gaiad, K.N., Singla, A.K., Boar, R.B. & Copsey, D.B., 1976. Triterpenoids and sterols of *Kalanchoe spathulata*. *Phytochemistry* 15: 1999–2000. |4| Gaiad, K.N., Singla, A.K. & Wallace, J.W., 1981. Flavonoid glycosides of *Kalanchoe spathulata*. *Phytochemistry* 20: 530–531. |5| Perry, L.M., 1980. Medicinal plants of East and Southeast Asia: Attributed properties and uses. The MIT Press, Cambridge, Massachusetts, United States & London, United Kingdom. p. 110. |6| Pételot, A., 1952. Les plantes médicinales du Cambodge, du Laos et du Vietnam [The medicinal plants of Cambodia, Laos and Vietnam]. Vol. 1. Centre National de Recherches Scientifiques et Techniques, Saigon, Vietnam. pp. 309–310. |7| Quisumbing, E., 1978. Medicinal plants of the Philippines. Katha Publishing Co., Quezon City, the Philippines. pp. 349–352. |8| Takeuchi, J., Horie, H. & Hirano, T., 1995. Grey mould of some garden plants is caused by *Botrytis cinerea* Persoon. *Proceedings of the Kanto-Tosan Plant Protection Society* 42: 105–107. |9| Wickens, G.E., 1982. Miscellaneous notes on *Crassula*, *Bryophyllum* and *Kalanchoe*. *Studies in the Crassulaceae for the Flora of Tropical East Africa*: 3. *Kew Bulletin* 36(4): 665–674. |10| Wickens, G.E., 1987. Crassulaceae. In: Polhill, R.M. (Editor): *Flora of Tropical East Africa*. A.A. Balkema, Rotterdam, Boston. pp. 30–58.

#### *Selection of species*

#### ***Kalanchoe ceratophylla* Haw.**

Rev. pl. succ.: 23 (1821).

**Synonyms** *Bryophyllum serrata* Blanco (1845), *Kalanchoe laciniata* auct. non (L.) DC.

**Vernacular names** Malaysia: sedingin, setawar kampong, chakar bebek. Philippines: siempreviva (Sp, Tagalog). Thailand: khong saamyaan (central), hom haem (northern). Vietnam: s[uw]f[ng] h[uw][ow]u, c[af] kheo, tr[uw][ow]f[ng] sanh r[as]ch l[as].

**Distribution** From the Himalayas, through As-

sam (India) and Burma (Myanmar) to southern China (Yunnan), Taiwan, peninsular Thailand and Peninsular Malaysia. Cultivated in Indonesia, Malaysia, the Philippines, Singapore, Thailand and Vietnam; also cultivated in Africa, South and Central America.

**Uses** The leaves or the juice pressed from the leaves of *K. ceratophylla* have widespread application in the topical treatment of ulcers and to relieve headache.

**Observations** An erect, unbranched or sparingly branched shrub, 30–125 cm tall, stems terete, lower internodes short, intermediate and higher ones gradually becoming longer; leaves numerous, very variable, the lowest simple, ovate and undulate-dentate, median ones deeply pinnatifid or bipinnatifid, 8–15 cm long, pale glaucous-green, tinged with purple when young, petiolate, upper leaves much smaller, narrow, often almost entire; inflorescence 10–30 cm long, glabrous; flowers with calyx variable in size, 4–10 mm long, segments erect or erecto-patent, ovate-lanceolate, acute, corolla salver-shaped, tube distinctly widened downwards, about 1.3 cm long, green at the base, yellowish upwards, lobes 4, widely patent, bright yellow, ovate or ovate-oblong, acute, about 1 cm long, anthers slightly exserted, carpels lanceolate, 5–6 mm long, glabrous, green, styles glabrous, 2–4 mm long. *K. ceratophylla* is found in gardens at lower elevations, and commonly grown as a pot plant.

**Selected sources** 95, 97, 202, 979, 1126, 1128, 1178, 1476, 1568.

#### ***Kalanchoe crenata* (Andrews) Haw.**

Syn. pl. succ.: 109 (1812).

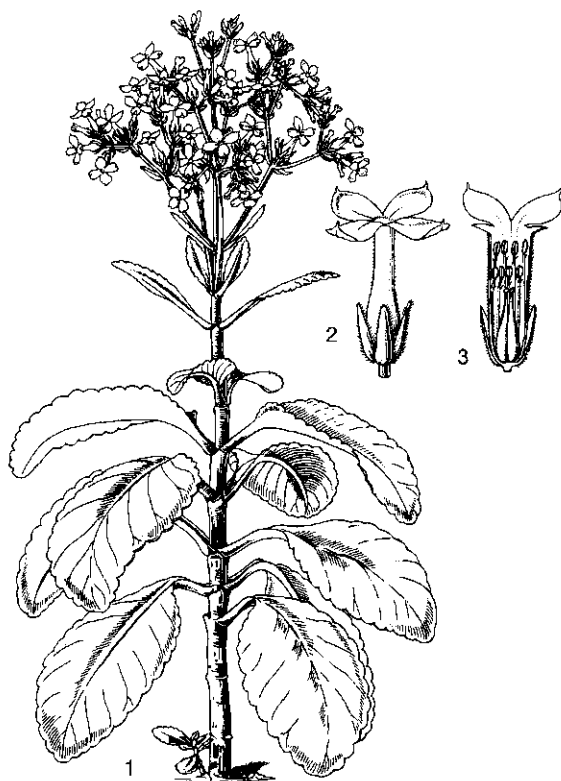
**Synonyms** *Cotyledon crenata* (Andrews) Vent. (1804), *Kalanchoe spathulata* DC. (1811), *Kalanchoe integra* auct. non (Medic.) O. Kuntze.

**Vernacular names** Indonesia: kayu urip (Javanese), buntiris (Sundanese), tampu taura (southern Sulawesi). Thailand: kong saamyaan, thong saamyaan (central). Vietnam: tr[uw][ow]f[ng] sinh l[as] to.

**Distribution** Probably native to tropical and South Africa, naturalized in Brail and South and South-East Asia. In Java mostly found in the east, on the hot and dry eastern slopes of the mountains.

**Uses** The juice of the leaves is generally used for its antiseptic, anti-inflammatory and counter-irritant properties.

**Observations** An erect or ascending shrub, 30–200 cm tall, usually unbranched, with a strong



*Kalanchoe crenata* (Andrews) Haw. - 1, plant habit; 2, flower; 3, flower in longitudinal section.

taproot, stems terete or obtusely quadrangular; leaves ovate or obovate-oblong to spatulate, 4–30 cm × 2.5–20 cm, base cuneate, apex obtuse, thickly coriaceous, margins crenate, pale green, glaucous or variably tinged with purple, petiolate; inflorescence up to 30 cm long; flowers with calyx lobes fused basally, 4–12 mm × 3–4 mm, acute to attenuate, corolla tube 8–16 mm long, distinctly widened downwards, (orange-)yellow above the greenish base, corolla lobes oblong-lanceolate to elliptical, (4–)8–12 mm × (2–)3–4(–5) mm, patent or subreflexed, after anthesis erect and twisted together, mucronate, anthers either all included or those of the upper series slightly exserted, carpels free or subconnate at the base, glabrous. *K. crenata* is a very variable, and widespread species known under a multitude of synonyms. It can be found in a wide range of open, dry, stony habitats up to 2000 m altitude, sometimes abundant in light *Casuarina* forest.

**Selected sources** 66, 67, 95, 97, 202, 286, 1126, 1128, 1178, 1179, 1476, 1523, 1536, 1569.

### *Kalanchoe laciniata* (L.) DC.

Hist. pl. Grass. 2: 100 (1802).

**Synonyms** *Cotyledon laciniata* L. (1753), *Kalanchoe craibii* Raymond-Hamet (1914).

**Vernacular names** Thailand: khong saamy-aan, thong saamyaaan (Bangkok).

**Distribution** Southern India and Thailand, South and East Africa and the Arabian peninsula. In Malasia locally cultivated, but not found wild.

**Uses** In view of the considerable overlap in chemical constituents found in *Kalanchoe* and the morphological similarity with *K. ceratophylla* which makes it difficult to distinguish between the 2 species, it is presumed that this species is used for the same purposes as *K. ceratophylla*.

**Observations** An erect shrub, 40–100(–150) cm tall, usually unbranched, stems terete, glabrous; leaves numerous, undivided or 3-lobed to seemingly compound, median leaves mostly 3-foliolate or 3-ternate, with linear-lanceolate leaflets, terminal leaflets 2–5(–8) cm long, with a crenate to dentate margin, green, petiolate; inflorescence an oblong cyme, glandular-pubescent; flowers with a subcampanulate calyx, 2.5–9 mm long, green, with lanceolate lobes, corolla salver-shaped, tube distinctly widened downwards, about 1 cm long, lobes 4, ovate-lanceolate to oblong, about 5–10 mm long, acute, cream to yellow, anthers included within the tube, carpels ovate-lanceolate, up to 9 mm long, glabrous, green. *K. laciniata* is found in open, rather dry, stony, habitats at 400–2000 m altitude.

**Selected sources** 1179, 1568, 1569.

Wardah & J.L.C.H. van Valkenburg

### *Lantana* L.

Sp. pl. 2: 626 (1753); Gen. pl. ed. 5: 275 (1754).

VERBENACEAE

$x = 11$ ; *L. camara*:  $2n = 22, 33, 44, 55, 66$ ; *L. trifolia*:  $2n = 48$

**Major species** *Lantana camara* L., *L. trifolia* L.

**Vernacular names** Sage, wild sage (En).

**Origin and geographic distribution** *Lantana* consists of approximately 150 species, and is native to tropical and subtropical America, the West Indies and Africa. Several species are well-known ornamentals, some are noxious weeds and have spread throughout the tropics and subtropics. *L. camara* and *L. trifolia* were introduced in Malasia in the 19th Century.

**Uses** In Indonesia, Malaysia and Thailand *L. camara* is assumed to have antiseptic properties, and so pounded leaves are applied to cuts and ul-

cers; a decoction of the leaves is used for the same purpose in the Philippines. In Java, the pounded leaves are applied to swellings to make them disappear; also a lotion or fomentation is made from them to treat rheumatism. A decoction of the leaves is used to treat constipation or as an emetic. In Indonesia and the Philippines a decoction or infusion of the leaves and flowers is used as a febrifuge, a diaphoretic and stimulant, and to relieve catarrh and bronchitis. A decoction of the fresh roots is used as a gargle to treat toothache in the Philippines and Thailand, and in Indonesia as a remedy for gonorrhoea and leucorrhoea. The ripe fruits are widely eaten by children.

In Central America a decoction is taken as a stomachic and remedy for rheumatism. A strong decoction is taken as an antidote for snakebites, whereas the crushed leaves are poulticed on the wound. A decoction of leaves or flowers is considered a remedy for colds or fever and employed for its diuretic and sudorific properties and as an emmenagogue. It is sometimes taken as a tonic and to treat hypertension in Costa Rica.

In Malaysia, *L. trifolia* is reported to be used in a similar way as *L. camara*. In Cuba, a decoction of the plant is applied externally to relieve rheumatism and is used as an eyewash in ophthalmia. In Colombia a decoction of the plant is used as an emmenagogue and sudorific. In Burundi the plant is used as a traditional remedy against gonorrhoea and applied to treat theileriasis in livestock. In general, *L. camara* is widely grown for its colourful flowers; several forms and varieties are in cultivation. Despite its weedy nature it can be used to stabilize slopes for erosion control and as an undemanding hedge in nurseries.

**Properties** The leaves of *L. camara* contain an essential oil, which is rich in sesquiterpenes. Furthermore, 6 compounds have been isolated and identified on the basis of chemical and spectral analysis: oleanonic acid, lantadene A, lantadene B, lantanilic acid, icterogenin and camaroside (4',5'-dihydroxy-3,7-dimethoxyflavone-4'-O- $\beta$ -D-glucopyranoside). Lantadene C (22- $\beta$ -2-methylbutanoyloxy-3-oxo-olean-12-en-18-oic acid), also isolated from the leaves of *L. camara* has been found to be identical with dihydrolantandene A. Lantadene C resembles lantadene A in the pentacyclic part of the molecule, but differs in the side chain region. Atom C-34 is *cis* to C-35 in lantadene C, but is *trans* in lantadene A. Semisynthetic lantadene C has been prepared by catalytic hydrogenation of lantadene A: it appears in two forms, crystalline and amorphous. Furthermore, the two

(pentacyclic triterpene acid) isomers lantadene A and lantadene B are the causal agents of most of the toxic effects.

Six oligosaccharides and six iridoid glucosides isolated from the ethanolic extract of *L. camara* roots have been identified as stachyose, verbascose, ajugose, verbascotetracoside, lantanose A, lantanose B, and the glucosides theveside, 8-epiloganin, shanzhsid methyl ester, theviridoside, lamiridoside and geniposide.

Eight triterpenoids have been isolated from the roots of *L. camara*. On the basis of their chemical properties and spectral data, they have been identified as lantanolic acid, 22  $\beta$ -O-angeloyl-lantanolic acid, oleanolic acid, 22  $\beta$ -O-angeloyl-oleanolic acid, 22  $\beta$ -O-seneciyl-oleanolic acid, 22  $\beta$ -hydroxy-oleanolic acid, 19  $\alpha$ -hydroxy-ursolic acid and 3  $\beta$ -isovaleroyl-19  $\alpha$ -hydroxy-ursolic acid (lantaiursolic acid). A bioactive triterpene, 22  $\beta$ -acetoxylantolic acid, was also isolated from *L. camara*; it showed antimicrobial activity against *Staphylococcus aureus* and *Salmonella typhi*, as well as antimutagenic activity.

The extract of root-bark of *L. camara* showed *in vitro* antimalarial activity in a test using the K1 strain of *Plasmodium falciparum* that is resistant to several drugs. *L. camara* leaves have been shown to contain water-soluble factors which caused isolated guinea-pig ileum to contract. The inhibitory effects of lantadenes and related triterpenoids from *L. camara* on the Epstein-Barr virus suggest potential against tumours. Verbascoside isolated from *L. camara* has *in vitro* antitumour activity, possibly at least partly due to inhibition of protein kinase C.

Potent inhibitors of human thrombin were demonstrated to be present in methanolic extracts from *L. camara* leaves. These were shown to be 5,5-trans-fused cyclic lactone-containing triterpenes. A methanol extract (0.05  $\mu$ g/ml) of aerial parts of *L. trifolia* produced bronchodilation of isolated guinea-pig trachea comparable with that of salbutamol (0.05  $\mu$ g/ml). The extract (0.1  $\mu$ g/ml) reduced bronchoconstriction of isolated guinea-pig trachea induced by histamine, 5-hydroxytryptamine (serotonin) or acetylcholine. Physostigmine (2-4  $\mu$ g/ml) failed to inhibit neuromuscular blocking activity of the extract (9 mg/ml) on rat phrenic nerve diaphragm. Using a disk diffusion method a methanolic extract of the leaves was found active against *Neisseria gonorrhoea* and *N. meningitidis*. The active component is probably the flavonoid umuhengerin (5-hydroxy-6,7,3',4',5'-pentamethoxyflavone).

All parts of *Lantana* except the flowers are toxic to livestock. *Lantana* poisoning in cattle, sheep, buffalo and guinea-pigs causes obstructive jaundice, photosensitization and raises serum glutaminoxaloacetic transaminase activity. The symptoms can be reproduced in sheep by administering purified lantadene A. The organs most affected during *Lantana* poisoning are the liver and kidneys. Intoxicating guinea-pigs with *L. camara* leads to marked alterations in major tissue constituents in these organs. Hepatic and renal xanthine oxidase activity is also elevated during poisoning. *Lantana* toxicity is manifested in three phases: the release and absorption of toxins in the gastro-intestinal tract, the hepatic phase resulting in cholestasis, hyperbilirubinaemia, hyperphylloerythrinaemia, and finally the tissue phase in which cell injury results from the accumulation of bilirubin and phylloerythrin. Thus, therapeutic measures should be aimed at arresting one or more of these phases. No antidote is available against the toxic section of *L. camara*. Symptomatic treatments have been proposed, with limited success. Unlike lantadene A (with a toxic and non-toxic form), both the crystalline and amorphous forms of lantadene C elicited strong hepatotoxic response in guinea-pigs associated with decrease in faecal output, feed intake, hepatomegaly, hepatic injury at the cellular and subcellular level, and increase in plasma bilirubin, and acid phosphatase activity. All the clinical signs, hepatic lesions, and changes in blood plasma typified *Lantana* toxicity. Water extracts of leaves of *L. camara* showed antifungal activity against rice blast (*Piricularia oryzae*) and brown spot of rice (*Helminthosporium oryzae*) in vitro, and antibiotic activity against gram-positive bacteria.

The crude extract of *L. camara* flowers is toxic against cotton stainer (*Dysdercus cingulatus*), housefly (*Musca domestica*) and corn weevil (*Sitophilus zeamais*). The oil from *L. camara* flowers is toxic to cotton stainer, housefly, corn weevil, black army worm (*Spodoptera exempta*) and lesser grain borer (*Rhizopertha dominica*). Flower extracts of *L. camara* showed a repellent effect against *Aedes* mosquitoes.

**Adulterations and substitutes** Lantadene A is identical to rehmanic acid isolated from *Lippia rehmanii* Pears.

**Description** Herbs or shrubs, sometimes subscandent, usually subscabrous and hirtous-pubescent or tomentose with simple hairs. Leaves opposite, sometimes in whorls of 3, simple, dentate, often rugose, petiole usually present; stipules ab-

sent. Inflorescence a dense cylindrical spike or contracted to form heads, usually axillary, pedunculate. Flowers bisexual, sessile, borne in the axil of solitary bracts, which are often ovate, acuminate, and subimbricate or spreading; calyx small, membranous, truncate or sinuate-dentate; corolla with a cylindrical, slender tube, actinomorphic or obscurely 2-lipped, 4-5-fid, lobes broadly obtuse or retuse, spreading, red, yellow or white often fading to other colours; stamens 4, didynamous, inserted at about the middle of the corolla tube, included, anthers ovate; ovary superior, 2-locular with 1 ovule in each cell, style usually short, stigma rather thick, oblique or sublateral. Fruit drupaceous, the exocarp subfleshy, endocarp hard, 2-celled or splitting in 2 parts. Seeds exalbuminous.

**Growth and development** The *Lantana* species in Malesia are evergreen and flower throughout the year. The flowers are mainly pollinated by moths and butterflies. The seeds are dispersed by fruit-eating birds.

**Other botanical information** *Lantana* is closely related to *Lippia* and *Verbena* and placed in the subfamily *Verbenoideae*. Various authors disagree whether the observed variation in growth habit, armature of the branches, indumentum of the leaves and changes in flower colour during anthesis of *L. camara* and *L. trifolia* is sufficient support for a further subdivision of these taxa in varieties or formae.

**Ecology** *Lantana* prefers rather open not too moist habitats, and occurs naturally from latitude 45°N to 45°S. As open habitats are often man-induced, some species have spread as a weed in cropped land and infested abandoned fields and pastures. *L. camara* is somewhat shade-tolerant and can become the dominant understorey in open forests or in tropical tree crops. Moderately fertile and well-drained soils are favoured.

**Propagation and planting** *L. camara* can be grown from seed but can also easily be propagated from cuttings 7.5 cm long.

**Husbandry** When grown as a pot plant, *L. camara* can easily be pruned into a desirable shape.

**Diseases and pests** *L. camara* often poses a serious problem in plantation crops and pastures. Conventional control methods such as burning, slashing and digging result in the regrowth of even more shoots. Therefore great effort has been put in finding methods for biological control. The most important biological control today is by the *Lantana* defoliator caterpillar *Hypena strigata*.

**Genetic resources and breeding** As both Malaysian *Lantana* species have a pantropical distribution as a weed, there is no risk of genetic erosion. Furthermore, both species are grown as ornamental.

**Prospects** The reported antitumour, antibiotic and bronchodilatory activities of *Lantana* seem to justify more research, which might result in future applications in modern medicine. The potential as a biological insecticide also deserves attention. Applications for erosion control should be limited to areas where livestock numbers are low. *L. camara* in particular is a popular pot plant with a wide range of cultivars.

**Literature** |1| Achola, K.J. & Munenge, R.W., 1996. Pharmacological activities of *Lantana trifolia* on isolated guinea pig trachea and rat phrenic nerve diaphragm. *International Journal of Pharmacognosy* 34(4): 273–276. |2| Burkill, I.H., 1966. A dictionary of the economic products of the Malay Peninsula. 2nd edition. Vol. 2. Ministry of Agriculture and Co-operatives, Kuala Lumpur, Malaysia. pp. 1280–1283. |3| Dharma, A.P., 1981. *Indonesische geneeskrachtige planten* [Indonesian medicinal plants]. De Driehoek, Amsterdam, the Netherlands. pp. 141–142. |4| Holm, L.G., Plucknett, D.L., Pancho, J.V. & Herberger, J.P., 1977. The world's worst weeds. Distribution and biology. East-West Center. The University Press of Hawaii, Honolulu, United States. pp. 299–302. |5| Morton, J.S., 1981. *Atlas of medicinal plants of middle America. Bahamas to Yucatan*. Charles C. Thomas Publisher, Springfield, Illinois, United States. pp. 739–741, 744–745. |6| Nguyen Van Duong, 1993. *Medicinal Plants of Vietnam, Cambodia and Laos*. Mekong Printing, Santa Ana, California, United States. p. 424. |7| Pan, W.D., Mai, L.T., Li, Y.J., Xu, X.L. & Yu, D.Q., 1993. Studies on the chemical constituents of the leaves of *Lantana camara*. *Yao Hsueh Hsueh Pao* 28(1): 35–39. (in Chinese) |8| Quisumbing, E., 1978. *Medicinal plants of the Philippines*. Katha publishing Co., Quezon City, the Philippines. pp. 795–797. |9| Rwangabo, P.C., Claeys, M., Pieters, L., Corthout, J., Berghe, D.A.V. & Vlietinck, A.J., 1988. Umuhengerin, a new antimicrobially active flavonoid from *Lantana trifolia*. *Journal of Natural Products* 51(5): 966–968. |10| Sharma, O.P., Makkar, H.P., Dawra, R.K. & Negi, S.S., 1981. A review of the toxicity of *Lantana camara* (Linn) in animals. *Clinical Toxicology* 18(9): 1077–1094.

### *Selection of species*

#### ***Lantana camara* L.**

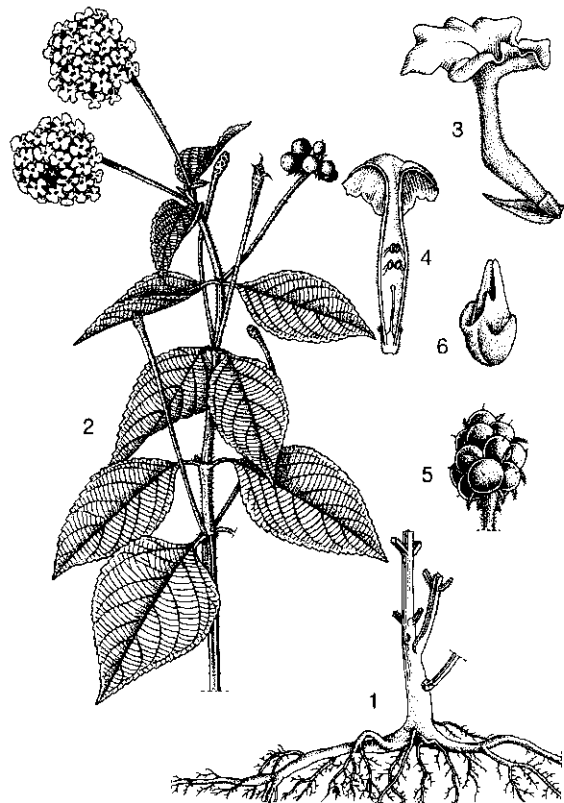
Sp. pl. 2: 627 (1753).

**Synonyms** *Lantana aculeata* L. (1753).

**Vernacular names** Sage, wild sage (En). Indonesia: kembang telek, tembelekan (Javanese), saliar (Sundanese). Malaysia: bunga pagar, bunga tahi ayam, jebat harimau (Peninsular). Philippines: koronitas, kantutay (Tagalog), baho-baho (Bisaya). Thailand: kaam kung (Mae Hong Son), khee kae (Pranchin Buri), yeesun (Trang). Vietnam: c[aa]y tr[aa]m [oor]i, c[aa]y b[oo]ng [oor]i, c[aa]y t[ua]w[s] qu[is].

**Distribution** Native to tropical America, but introduced and naturalized throughout the tropics and subtropics.

**Uses** In South-East Asia the leaves (and sometimes the wet, ground roots) are applied to cuts, ulcers, swellings and to treat rheumatism, a decoction of the leaves and flowers is used to treat



*Lantana camara* L. – 1, root system; 2, flowering and fruiting twig; 3, flower with bract; 4, flower in longitudinal section; 5, infructescence; 6, seed.

constipation, as a febrifuge, diaphoretic and stimulant, and to relieve catarrh and bronchitis. A decoction of the roots is used to treat toothache, headache, inflammation, gonorrhoea and leucorrhoea. *L. camara* is used as an ornamental and in hedges.

**Observations** An erect or subscandent much-branched shrub, up to 5 m tall, stems square or 3-angled, often bearing hooked prickles, highly aromatic; leaves opposite or rarely in whorls of 3, ovate to oblong-ovate, 5–8 cm × 3–5.5 cm, petiole 1.5–3 cm long; inflorescence flat or hemispherical subcapitate; flowers with corolla tube extending to 12 mm long during anthesis, often slightly curved, orange-yellow or orange to pink, white, or variegated, changing to red or scarlet; fruit a globose glossy drupe, deep blue when ripe. *L. camara* occurs from sea-level to 1700 m altitude in relatively open and disturbed, not too moist habitats. It is mainly a weed of plantation crops and pastures. Due to the toxicity of leaves and seeds it is a serious threat to sheep and cattle.

**Selected sources** 92, 97, 129, 202, 332, 350, 468, 578, 650, 968, 979, 1035, 1065, 1089, 1106, 1107, 1108, 1126, 1128, 1178, 1213, 1321, 1322, 1323, 1558, 1561, 1572.

### **Lantana trifolia L.**

Sp. pl. 2: 626 (1753).

**Vernacular names** Indonesia: tahi ayam (general), puyengan (Javanese), mandakaki (Sundanese). Malaysia: bunga pagar putih (Peninsular). Thailand: phakaa krong.

**Distribution** Native to the West Indies and South America, common in tropical Africa and introduced and naturalized in India, Peninsular Malaysia and Java.

**Uses** In Malaysia and South America *L. trifolia* is used in a similar way as *L. camara*. In Burundi it is also applied to treat theileriasis in livestock.

**Observations** An erect shrub up to 3 m tall, stems angular, with white hairs but not prickly, faintly aromatic; leaves mostly in whorls of 3, sometimes opposite or 4-whorled, oblong-lanceolate to elliptical-lanceolate, 5–12 cm × 2–6 cm, petiole 5–12 cm long; inflorescence spicate, subcapitate when young, later elongated to 5 cm long; flowers with a corolla tube extending to 7 mm long during anthesis, pink, lavender, purple or white; fruit a sweet, juicy drupe, globose or obovate, purplish-red or white. *L. trifolia* occurs in open disturbed habitats from sea-level to 800 m altitude, preferring dry thickets.

**Selected sources** 92, 97, 101, 202, 968, 979, 1267, 1505.

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### **Melochia corchorifolia L.**

Sp. pl. 2: 675 (1753).

STERCULIACEAE

2n = 46

**Synonyms** *Melochia concatenata* L. (1753).

**Vernacular names** Chocolate-weed, redweed (En). Indonesia: orang-aring (general), jaring (Sundanese), gendiran (Javanese). Malaysia: lemak ketam, lemak kepiting, bayam rusa (Peninsular). Philippines: bankalanan (Iloko), kalingan (Panay Bisaya). Thailand: khaang paak put (Chi-ang Mai), sa aeng bai mon (Chai Nat), seng lek (Ang Thong). Vietnam: tr[uws]ng cua, v[ai] gi[aa]s[y].

**Origin and geographic distribution** *Melochia* consists of approximately 55 species and is largely confined to the tropics, although some species reach subtropical regions. The greatest diversity in species is found in Central and South America. Only 2 or 3 species are native to South-East Asia. *M. corchorifolia* is a weed throughout the tropics and subtropics, including South-East Asia. It originates from the Old World tropics and has been introduced in the Americas.

**Uses** Traditional uses of *M. corchorifolia* in South-East Asia are only reported for Malaysia. The leaves are used for poulticing sores and swellings of the abdomen, and the sap is applied as an antidote to wounds caused by arrows poisoned with *Antiaris toxicaria* Lesch. Leaves and roots are used for poulticing in cases of smallpox. A decoction of the leaves and roots is used internally to treat dysentery, and a decoction of the leaves to stop vomiting. A leaf decoction is prescribed in a compound mixture against urinary disorders. A decoction of the plant is applied in folk medicine in India as a cure for abdominal swelling, dysentery and snake bites.

In Papua New Guinea, the leaves of an unidentified *Melochia* species are applied to the forehead to treat headache, and the fruit is eaten. The leaves of *M. corchorifolia* are sometimes eaten in Indo-China and India. The plant yields a beautifully silvery-white, fine and strong fibre, but in too small quantity to be important.

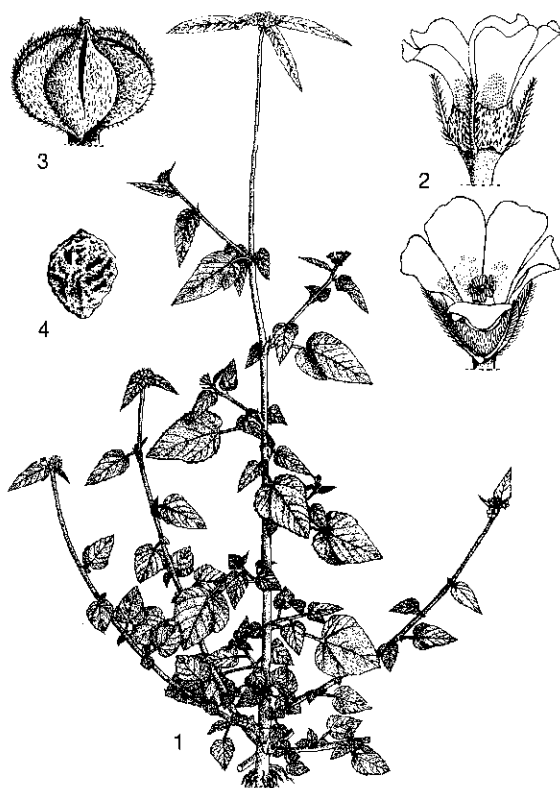
**Properties** A phytochemical investigation of extracts from the dried, powdered aerial parts of



*M. corchorifolia* revealed the presence of the triterpenes friedelin, friedelinol and  $\beta$ -amyrin, the sterol  $\beta$ -sitosterol together with its stearate and  $\beta$ -D-glucoside, the aliphatic compounds ethylstearate, tetratriacontanol, nonacosylnon-4-enoate, 24-ethyl-2-methyltritetracont-1-ene-3,23-diol and 27-methyloctacosane-1,3-diol and the flavonoids vitexin and robunin. Furthermore several alkaloids have also been reported in similarly prepared extracts: franganine, frangufoline adouetine-y' and melofoline (cyclopeptide alkaloids), melochicorine (a pseudo-oxindole alkaloid) and 6-methoxy-3-propenyl-2-pyridine carboxylic acid (a pyridine alkaloid). The latter compound may be of significance, since related pyridine derivatives (e.g. pyridoxine, 4'-methoxypyridoxine, nicotinic acid) are physiologically active. The flavonol glycosides hibifolin, triflin and melocorin have been isolated from the leaves.

The main alkaloid in *M. pyramidata* L., American in origin but naturalized in many tropical and subtropical regions including South-East Asia, is (-)-(R)-melochinine. This compound has been shown to produce paralysis, bradypnea, bradycardia and hypotension in laboratory animals, and ingestion of plant material by cattle may cause paralysis. The mechanism of action of this alkaloid can be described as in general non-specific. It may be partly explained by an unspecific interaction with membranes, partially responsible for a calcium-antagonistic effect. Unlike its structural analogue piercidin A, a well-known inhibitor of the mitochondrial respiratory chain, melochinine does not show insecticidal activity.

**Description** A perennial herb or subshrub up to 130 cm tall, erect or spreading and often widely branched, with tough bark. Leaves arranged spirally, simple, triangular or broadly ovate to lanceolate, lower leaves often slightly 3-lobed, 1-9 cm  $\times$  0.5-5 cm, margin crenate-serrate, 3-veined or 5-veined from the base, pubescent on the veins, green or with purplish tinge; petiole 0.3-4.5 cm long, sparsely pubescent to subvillose; stipules about 5 mm long, present on young twigs. Inflorescence an axillary or terminal head-like cyme, rarely less compact, subtended by 1-4 leaves with their stipules forming a kind of involucre, many-flowered. Flowers with pedicel bearing 3-4 pilose bracteoles at apex, bisexual, actinomorphic, 5(-7)-merous; calyx campanulate, about 2.5 mm long, with teeth much shorter than tube, pubescent; petals obovate-spatulate, 4-7 mm long, lilac or white with a yellow spot at base, soon withering; stamens opposite the petals, filaments connate to



*Melochia corchorifolia* L. - 1, flowering plant; 2, flowers viewed from different angles; 3, fruit; 4, seed.

halfway or more, anthers broad, 2-lobed; ovary superior, 5-celled, densely pilose, each cell with 1-2 ovules, styles 5, united at base. Fruit a small globose capsule, 3.5-5 mm in diameter, green, whitish, pink to purplish-black, loculicidal but valves easily septicial dehiscent, each cell 1-2-seeded. Seeds small, wingless; endosperm abundant; embryo straight, cotyledons flattened. Seedling with epigeal germination; cotyledons suborbicular, foliaceous.

**Growth and development** *M. corchorifolia* is reported to flower and fruit all year round. The flowers are probably pollinated by small insects.

**Other botanical information** *M. corchorifolia* is extremely variable morphologically and numerous varieties have been distinguished.

Roots of *M. tomentosa* L. have been used in Curaçao to relieve throat inflammation. However, the root extract was reported to be tumorigenic. *M. umbellata* (Houtt.) Stapf is a small tree indigenous from India to New Guinea for which no medicinal uses have been recorded.

**Ecology** *M. corchorifolia* is a common weed in many regions in sunny or slightly shaded, usually humid localities, at watersides and in fields, waste places and open forest, up to 700 m altitude in Java. Although it is adapted to xerophytic conditions, *M. corchorifolia* has retained its ability to grow in mesophytic and hydrophytic habitats. In the Philippines, it is reported as one of the dominant weeds in upland rice, together with *Echinochloa colona* (L.) Link. In Thailand and Indonesia it is also a weed in lowland rice, moreover it is also recorded as such in soya bean.

**Propagation and planting** Scarification of seed improves germination considerably. Scarified seed germinates best at a temperature of 35–40°C. Seed buried to a depth of 1–5 cm gave a germination rate of 80–90% after 7 days; when planted at the soil surface or deeper than 8 cm the seed did not germinate.

**Genetic resources and breeding** Since *M. corchorifolia* shows a very extensive geographical distribution, occurring in anthropogenic habitats, it is not liable to genetic erosion.

**Prospects** Although some research has been done on the phytochemistry, very little is known about the pharmacological properties and activity of *M. corchorifolia* and its compounds. The fact that tumorigenic and toxic activity has been reported from some other *Melochia* spp. should lead to caution in using *M. corchorifolia* in phytotherapy as so little is known about its biological activity.

**Literature** [1] Bhakuni, R.S., Shukla, Y.N. & Thakur, R.S., 1986. 6-methoxy-3-propenyl-2-pyridine carboxylic acid: a new pyridine alkaloid from *Melochia corchorifolia*. *Chemistry and Industry* 1986(13): 464. [2] Bhakuni, R.S., Shukla, Y.N. & Thakur, R.S., 1987. Chemical constituents of *Melochia corchorifolia* Linn. *Indian Journal of Chemistry, section B, Organic Chemistry including Medicinal Chemistry*, 26(12): 1161–1164. [3] Bhakuni, R.S., Shukla, Y.N. & Thakur, R.S., 1987. Cyclopeptide alkaloids from *Melochia corchorifolia*. *Phytochemistry* 26(1): 324–325. [4] Bhakuni, R.S., Shukla, Y.N. & Thakur, R.S., 1991. Melochicine, a pseudooxindole alkaloid from *Melochia corchorifolia*. *Phytochemistry* 30(9): 3159–3160. [5] Burkill, I.H., 1966. A dictionary of the economic products of the Malay Peninsula. Revised reprint. Vol. 2. Ministry of Agriculture and Cooperatives, Kuala Lumpur, Malaysia. p. 1472. [6] Eastin, E.F., 1983. Redweed, *Melochia corchorifolia*, germination as influenced by scarification, temperature and seeding depth. *Weed Sci-*

*ence* 31(2): 229–231. [7] Goldberg, A., 1967. The genus *Melochia* L. (Sterculiaceae). *Contributions from the United States National Herbarium* 34(5): 191–363. [8] Li, H.-L. & Lo, H.-C., 1993. Sterculiaceae. In: Huang, T.-C. (Editor): *Flora of Taiwan*. 2nd edition. Vol. 3. Editorial Committee of the Flora of Taiwan, Taipei, Taiwan, Republic of China. p. 764. [9] Nair, A.G.R., Ramesh, P. & Subramanian, S.S., 1977. Melocorin, a new herbacetin glucuronide from *Melochia corchorifolia*. *Indian Journal of Chemistry, section B, Organic Chemistry including Medicinal Chemistry*, 15(11): 1045. [10] Soerjani, M., Kostermans, A.J.G.H. & Tjitrosoepomo, G., 1987. *Weeds of rice in Indonesia*. Balai Pustaka, Jakarta, Indonesia. pp. 554–555, 640–641.

**Other selected sources** 97, 185, 1126, 1178.

M.C. Ysrael

## **Mentha arvensis L.**

Sp. pl. 2: 577 (1753).

LABIATAE

2n = 36, 72, 96

**Vernacular names** Mint, field mint (En). Baume des champs (Fr). Indonesia: daun poko (general), bijanggut (Sundanese), janggot (Javanese). Malaysia: pohok. Philippines: polios (Sp, Tagalog). Singapore: pokok kepari. Cambodia: chi poho. Thailand: min indoneesia (Bangkok). Vietnam: b[aj]c h[af], b[aj]c h[af] nam.

**Origin and geographic distribution** *Mentha* includes about 30 species and numerous hybrids, and mainly occurs in the temperate parts of the northern hemisphere; the centres of diversity are located in Europe and northern and central Asia. Mint has been known as a kitchen herb and medicinal plant since ancient times. It has been found in pyramids in Egypt and is described in old Chinese literature. Only *M. arvensis* occurs in South-East Asia, but some other species and hybrids are cultivated in kitchen gardens and in pots.

*M. arvensis* is native to Europe and northern and central Asia and is cultivated in many parts of the world. Var. *arvensis* has also been introduced in Malesia from Europe. The Spaniards introduced it in the Philippines, where its cultivation is now widespread, but where it does not flower; it is also cultivated as a pot-herb in Peninsular Malaysia and Singapore. It is cultivated throughout Vietnam, whereas wild forms of *M. arvensis* are also found in northern Vietnam. Var. *javanica* (Blume) Hook.f. (synonym: *Mentha javanica* Blume) occurs

in Sri Lanka and the Malesian area (Peninsular Malaysia, Sumatra, Java, Timor, north-eastern Sulawesi, Banda in the Moluccas and the Philippines). It is often cultivated.

Var. *piperascens* Malinv. ex Holmes (Japanese mint) has been introduced into northern Thailand where since 1973 it has been grown on a commercial scale, mainly in Nan Province, for the extraction of mint oil.

**Uses** Most *Mentha* species are very fragrant and used as a condiment, medicinal plant and as a source of essential oil. The most important use of mints today on a world scale, is as a source of essential oil, which is used in medicinal preparations (ointments, itch-relieving creams, cough syrups, cough lozenges, tablets), as a flavouring agent in toothpastes, mouthwashes, confectionery, candies, chewing gums, beverages and cigarettes, and in the perfume industry (for lotions, soap and cologne). In South-East Asia, however, mints are mainly used to flavour food and for medicinal purposes. However, commercial mint oil production is developing as well, e.g. in Thailand and Indonesia (West Java).

Leaves, whole plants and the oil extracted from *M. arvensis* are all reported as having medicinal properties. They are used as a carminative, stomachic, antispasmodic, stimulant, sedative, sudorific, emmenagogue, astringent (externally) and refrigerant (externally) all over the world. They are administered internally to treat indigestion, flatulence, gastro-intestinal atony, colic and diarrhoea, or externally in the treatment of colds, influenza, fever, sinusitis, nose and throat complaints (all e.g. as nasal drops), headache, facial neuralgia and insect stings (e.g. as rubefacient). In Indonesia, pounded leaves are used externally against headache, and an infusion of the leaves as sudorific and expectorant to treat cough, as a carminative, and as antispasmodic in gastro-enteritis. Although the main use in the Philippines is reportedly as a culinary herb, an infusion of the leafy stems is also used as carminative, and pounded leaves are used to treat insect stings. Mint leaves in the form of tea or tablets are used in the Philippines as an analgesic, particularly in dental surgery. In Thailand, *Mentha* is widely used as a culinary herb but also as a medicine for its carminative, stomachic and expectorant properties.

Compared with other *Mentha* spp., South-East Asian mint is usually mild in flavour and the young leaves are also eaten raw as a side dish; except those of Japanese mint (*M. arvensis* var. *piperascens*), which are rich in menthol.

**Production and international trade** World production of *Mentha* oil is estimated at 6500–8000 t/year. Japanese mint (*M. arvensis* var. *piperascens*) contributes the greatest part, with 4000–4500 t/year (and a value of about US\$ 43 million/year) in the period 1990–1995. The main producing countries are China, Brazil and India. Small amounts of mint oil are produced in Thailand and Vietnam; the current production in Vietnam is about 100 t/year, and the production in Thailand in 1975 was estimated at about 15 t. The world production of peppermint oil is 2000–2500 t/year, with the United States as the main producer. World production of spearmint oil is about 1000 t/year.

**Properties** Mint oil is a complex mixture of numerous constituents. In general, the different *Mentha* species contain characteristic monoterpenes as main components, but several species also have divergent chemotypes. The characteristic 'cool taste' is due to (-)-menthol ((1R,3R,4S)-menthol), a monoterpenoid alcohol; other optically active isomers of menthol (e.g. (1R,3S,4S)-neomenthol) do not have the same organoleptic characteristics.

Mint oil is obtained from the flowering tops of Japanese mint (*M. arvensis* var. *piperascens*); leaves can contain over 5% (in general 1–1.8%), but at least 0.8% V/w of essential oil on a moisture-free basis according to the Chinese Pharmacopoeia. Japanese mint oil can contain as much as 92.5% (-)-menthol. Very slow cooling of the essential oil (e.g. from 35°C to 5°C at 2°C/day) induces part of the menthol to crystallize. An additional quantity of menthol can be recovered by saponifying (-)-menthyl acetate and by hydrogenating (-)-menthone.

After some of the menthol has been removed, the average composition of the essential oil (referred to as 'rectified' or more often 'dementholized') used for e.g. pharmaceutical or cosmetic products is 30–45% (-)-[1R,3R,4S]-menthol, 17–35% (-)-[1R,4S]-menthone, 5–13% (+)-[1R,1R]-isomenthone, 2–7% menthyl acetate (mainly (-)- or (1R,3R,4S)-), 1.5–7% limonene and 2.5–4% (+)-[1R,3S,4S]-neomenthol. About 30 minor terpenoid constituents have also been identified. They include piperitone (0.5–4%), pulegone (0.2–3.5%),  $\beta$ -caryophyllene (2–5%),  $\beta$ -caryophyllene-epoxide (0.5–2%),  $\alpha$ -pinene (2–4%),  $\beta$ -pinene (2–4%), germacrene D (0.1–1.3%), 1,8-cineole (<1%), linalool (<1%), menthofuran (<1%) and camphene (<1%). The oil of *M. arvensis* var. *javanica* from Java has been reported as bitter with a low menthol con-

tent and a high pulegone content, but with an agreeable aromatic odour.

The pharmacology of *Mentha* drugs has not been explored much. Mint oil showed a significant in vitro spasmolytic activity on isolated guinea-pig ileum. It is possible that this activity is linked to an inhibition of calcium entry into the cells. The usefulness of the essential oil in cases of 'irritable colon' has been demonstrated in some studies, but in others there was no improvement. Although the oil is the major constituent of several over-the-counter remedies for symptoms of irritable bowel syndrome (a common disorder with a psychophysiological basis), its role in the symptomatic treatment has so far not been established beyond reasonable doubt. Oil from Indian mint showed strong in vitro fungitoxic activity against the dermatophytes *Trichophyton rubrum* and *Microsporum gypseum*, and also against *Aspergillus* spp. The oil was able to cure experimentally induced ringworm in guinea-pigs within 2 weeks. Antibacterial activity of mint oil was also demonstrated in vitro and in foods to which the oil had been added; the growth of *Salmonella* and *Listeria* bacteria was inhibited. In tests in Taiwan an aqueous extract of *M. arvensis* markedly inhibited the growth of both gram-positive and gram-negative bacteria too.

Mint oil is not without toxicity. High doses of menthol are toxic; there is a risk of spasm of the glottis (asphyxia) in young children and asthmatic patients.

The traditional use of *M. arvensis* as carminative and stomachic has been verified by pharmacological evidence of the carminative effect of the essential oil and antispasmodic and choloretic activity. In tests with rats in India a 50% ethanolic extract of mint leaves has been found to reduce the fructose synthesis in seminal vesicles: male sterility occurred until 30 days after the last treatment. Subcutaneous administration to rats in early pregnancy caused a significant number of abortions.

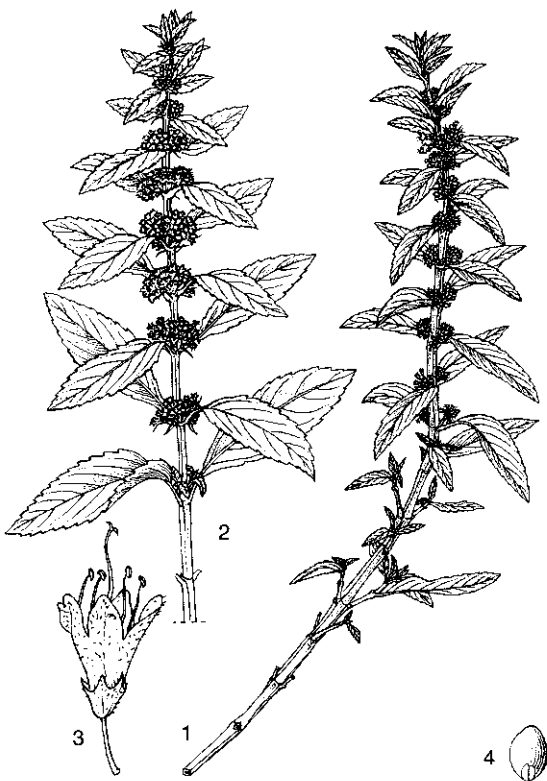
Peppermint oil showed anti-inflammatory effect in xylene-induced ear oedema in mice and in a cotton pellet granuloma test in rats. Moreover, it showed anti-nociceptive effect against acetic acid-induced writhing and hot plate-induced thermal stimulation in mice. Tablets of 'yerba buena' (*M. xcordifolia* Opiz ex Fresen) tested in the Philippines did not possess direct DNA damaging capacity. They were not mutagenic before and after metabolic activation and they did not possess chromosome-breaking effects. They exhibited an-

timutagenic effects against dimethylnitrosamine, mitomycin C, N-nitrosopyrrolidine and tetracycline; they reduced the formation of micronucleated polychromatic erythrocytes induced by these compounds.

Mint oil was found to be effective as a fumigant against rice weevil (*Sitophilus oryzae*) in stored sorghum grain, without affecting the germination capacity of the grain. However, it did affect the taste of boiled sorghum.

**Adulterations and substitutes** Several other *Labiatae* have essential oil with similar or related compounds and with similar applications, i.e. mainly for minor digestive disturbances, in dermatology and hygiene, and in perfumery, cosmetology and confectionery. Examples include the genera *Lavandula*, *Ocimum* and *Salvia*.

**Description** An aromatic, stoloniferous herb up to 60 cm tall, often rooting on lowest stem parts; stem prostrate, quadrangular, pubescent with appressed hairs. Leaves decussately opposite, simple, lanceolate to broadly lanceolate, 2.5–4.5(–7) cm × 1–2.5(–3) cm, long-cuneate at base, acute at



*Mentha arvensis* L. - 1, flowering plant; 2, upper part of flowering stem; 3, flower; 4, nutlet.

apex, serrate but entire at base, membranaceous, sparingly hairy above, glabrous beneath; petiole 0.5–1 cm long; stipules absent. Inflorescence consisting of short, fascicled cymes in the axils of the leaves forming a verticillaster; bracts linear or subulate. Flowers small, bisexual; pedicel 2–2.5 mm long; calyx tubular-campanulate, 2–3 mm long, 5-toothed, appressed hairy; corolla funnel-shaped, 4.5–5 mm long, 4-lobed, faintly 2-lipped due to the broader, emarginate upper lobe, puberulent outside, violet or lilac; stamens 4, slightly didynamous, filaments inserted on the corolla tube, erect, anthers 2-celled; disk entire and uniform; ovary superior, consisting of 2 carpels each of which is 2-celled, style with 2 short branches. Fruit consisting of 4 dry 1-seeded schizocarpous nutlets enclosed in the persistent calyx; nutlets ellipsoid, about 1 mm long, finely granular, often pointed at apex and with a large lateral scar below. Seedling with epigeal germination; cotyledons petiolate, circular-oblong, rounded to truncate at apex; hypocotyl elongate, epicotyl nearly absent.

**Growth and development** In Vietnam, new shoots of *M. arvensis* normally arise from cuttings within 5–10 days after planting. Flowering starts 80–100 days after planting, and although plants usually blossom profusely they rarely set seed. Except for *M. arvensis*, all *Mentha* spp. in the Malesian region do not flower, or flower rarely.

**Other botanical information** *Mentha* is taxonomically rather complex. Identification is often difficult since, apart from their phenotypic plasticity and genetic variability, most species hybridize easily with each other; polyploidy is common. *M. arvensis* is a complex species with a very large area of distribution, which has resulted in a confusing infraspecific taxonomy. The widespread var. *javanica* is reported as possibly indigenous to the Malesian region. Var. *javanica* differs from var. *arvensis* mainly in the almost filiform or mucronate calyx teeth, which are separated by wide bays; the calyx teeth of var. *arvensis* are triangular without distinct mucronate apices. The assumption that var. *javanica* is of hybrid origin and was originally imported from Europe, just like var. *arvensis*, has not been proved. The relationship between *M. arvensis* var. *javanica* and other taxa such as *M. haplocalyx* Briq. (synonym: *M. arvensis* subsp. *haplocalyx* (Briq.) Briq.), *M. canadensis* L. and *M. arvensis* var. *piperascens* Malinv. ex Holmes is still unclear, but all these are at least closely related.

A hybrid, named *M. xcordifolia* Opiz ex Fresen, is

also cultivated in Java, the Philippines and Thailand. The correct name might be *M. xvillosa* Huds., which is a hybrid between *M. spicata* L. and *M. suaveolens* Ehrh. It originates from northern temperate regions, never flowers and is propagated vegetatively. It can be distinguished from *M. arvensis* by its rounded to truncate or shallowly cordate leaf bases and less hairy stems. *M. xcordifolia* is commonly cultivated for medicinal purposes and for use as flavouring throughout the Philippines and Thailand. It is likely that this taxon has been often confused with varieties of *M. arvensis*, particularly in the Philippines where they all appear to occur.

*M. pulegium* L. (pennyroyal), also a native of northern temperate regions, is cultivated locally in gardens in Java as a culinary herb. *M. aquatica* L. (water mint) is cultivated in Vietnam and Thailand. *M. xpiperita* L. (peppermint, a hybrid between *M. aquatica* and *M. spicata*) is cultivated for its essential oil throughout the temperate regions of the world; it has been introduced very locally in Indonesia (Java, Timor). *M. xrotundifolia* (L.) Huds. (apple-mint, a hybrid between *M. longifolia* L. and *M. suaveolens*) is cultivated and naturalized in Vietnam. *M. spicata* L. (spearmint) is a well-known medicinal plant in India, used to treat fever, bronchitis and aphthae; it is not reported for South-East Asia.

**Ecology** Most *Mentha* spp. originate from temperate regions and grow best under cool conditions. They are probably quantitative long-day plants and usually do not flower in tropical regions, although some species have been found flowering at higher altitude. *M. arvensis* var. *javanica* occurs in open, mostly humid localities, e.g. in borders of rice fields, at 150–1200 m altitude. It is found flowering throughout the year. At least some cultivars of Japanese mint (*M. arvensis* var. *piperascens*) are better adapted to tropical climates. In greenhouse experiments, this species even came into flower under the minimum photoperiod of 10 hours, which is much less than for other species such as peppermint (*M. xpiperita*) and spearmint (*M. spicata*). In northern Thailand, cv. So Wo 1 of Japanese mint outyielded spearmint and peppermint production (in both dry matter and mint oil production) in the lowlands, but in the highlands other cultivars of Japanese mint and a cultivar of spearmint performed better. Unfavourable climatic conditions can cause excessive levels of undesirable compounds or low levels of desirable compounds in the oil of commercially grown mint taxa, resulting in oil of poor quality.

In peppermint (*M. x piperita*), for instance, it is known that long days and cool nights lead to higher yields of oil and to an increase in the menthofuran level, and that cold nights favour the formation of menthol. Japanese mint can thrive in the Philippines in well-watered, well-drained soils in the dry season (September to April). Mints grow best in deep soils rich in humus, with a pH of 6–7.5.

**Propagation and planting** Mints are usually propagated vegetatively by stolons. Cuttings should preferably be taken from one-year-old plants, and cut into pieces of 4–5 cm long with 3–4 nodes. Japanese mint is usually planted in northern Vietnam in January–April (spring). The cuttings are planted 12–20 cm apart in shallow furrows 7–10 cm deep and 45–60 cm apart. For planting one ha, 400–450 kg of stolons are required.

Tissue culture of mints is possible. Callus cultures of Japanese mint have been established in India, using standard medium with some modifications. The plantlets produced were transplanted to soil and had a high rate of survival. Before planting, the land should be cleaned as thoroughly as possible. In Thailand, Japanese mint is intercropped with teak (*Tectona grandis* L.f.) seedlings.

The crop responds favourably to the application of 20–30 t/ha of manure or compost to the field before planting. Green manuring may also be practised.

**In vitro production of active compounds** A suspension culture of *Mentha* has been established from callus formed on the tips of young shoots. Cells were grown in B-5 liquid medium supplemented with 1 mg/l 2,4-dichlorophenoxy acetic acid in the dark, and subcultured at intervals of 2 weeks. The cell suspension has been maintained for up to 4 years. The culture released a large amount of extracellular polysaccharides.

**Husbandry** Although a peppermint crop can be ratooned for 3 years, Japanese mint is usually planted every year. Weeding should be carried out regularly. When plants are grown for oil production it is especially important to avoid contamination with weeds, because of the risk of off-flavours. The crop should be watered liberally to support the profuse vegetative growth. Application of fertilizers (up to 160 kg N and 60 kg P<sub>2</sub>O<sub>5</sub> per ha under northern Vietnamese conditions) increases crop and essential oil yields.

**Diseases and pests** Mint crops are affected by a number of fungal diseases, particularly when grown in regions with a warm and humid climate. Severe leaf shedding is caused by the mint rust

*Puccinia menthae* and *Fusarium* spp. Powdery mildew caused by *Erysiphe cichoracearum* appears to be the most serious problem in India. *Verticillium* wilt is the major disease in peppermint and spearmint crops in the United States. Collar rot developed in an experimental plantation of Japanese mint in Papua New Guinea, resulting in the wilting and death of shoots; it was caused by *Marasmiellus epochnous*. Mints are reported to be attacked by a large number of insect pests, the most serious of which are moths.

**Harvesting** In Vietnam, 2–3 harvests per year can be obtained from Japanese mint. The first harvest is 100–130 days after planting, when the lower leaves turn yellow and the crop is flowering, the second harvest is carried out 80–100 days after the first, and the third 80–90 days after the second. If harvesting is delayed and leaves start falling, oil yields will be lower. The oil content decreases rapidly after the full bloom stage.

**Yield** A good crop of Japanese mint may produce 40–45 t/ha of fresh material in 2 cuttings. Fresh plant material contains 0.4–0.8% essential oil, but the oil yield and quality largely depend on field conditions, plant age, and the presence of weeds, diseases and pests. In India the highest yields of fresh mint (53 t/ha) and oil (270 kg/ha) were obtained when the crop was planted at 60 cm row spacing and fertilized with 240 kg N/ha.

The yield of oil from *M. arvensis* cv. Jombang in Indonesia is reported at 30.5 l/ha.

In India, tetraploid plants of Japanese mint contain more oil than diploid ones. However, the menthol content of oil from tetraploid Japanese mint was lower and the menthone content higher. Moreover, it was inferior in growth habit and more susceptible to diseases, which made the quality and total yield of oil lower compared with diploid plants.

In the Philippines, oil yield of *M. x cordifolia* reaches a peak in the period March–June, with a secondary peak in December. Heavy rains in July–September depress growth and oil yield.

**Handling after harvest** Mint oil is obtained by distillation of fresh or slightly dried plant material. The crop should not be excessively dried in the sun. To obtain good quality oil, the herb is distilled as rapidly as possible to prevent hydrolysis of esters and alteration of other constituents by excessively long exposure to steam. It usually takes 1.5–2 hours to complete the process of distillation in a well-designed still with good steam pressure. In the case of small-scale farming a direct-fired still can be used, but the oil yield de-

pends on the efficiency of distillation equipment.

**Genetic resources and breeding** In recent years, *Mentha* germplasm has been collected in Russia, Ukraine, China, Japan, India, United States and Brazil. In India, breeding work is done at the Central Institute of Medicinal and Aromatic Plants in Lucknow, with the aim of developing cultivars that combine excellent oil quality and a wide range of maturity dates with resistance against diseases.

**Prospects** Successful mint cultivation in South-East Asia partly depends on the availability and maintenance of certified plant material. However, the climatological conditions are a serious drawback for commercial mint growing. Some regions at the edge of South-East Asia, such as northern Vietnam and northern Thailand, may have promising prospects for large-scale production of good-quality mint oil. In the Malesian region, the role of mint will probably remain limited to the use of imported mint oil and to grow plants as a pot herb or occasionally as a medicinal herb. However, when better adapted cultivars become available, which do not suffer a decrease in oil quality in tropical conditions, there might also be a future for commercial mint oil production in Malesia. Recent developments are encouraging.

**Literature** |1| Bruneton, J., 1995. Pharmacognosy, phytochemistry, medicinal plants. Lavoisier Publishing, Paris, France. pp. 431–437. |2| Cantoria, M., 1980. Physiology and biochemistry of the volatile oils of *Mentha* species (Fam. Labiatae) grown in the Philippines. Transactions of the National Academy of Science and Technology 2: 163–190. |3| Chomchalow, N., Buranasilpin, P., Pichitakul, N., Euraree, A. & Pangspa, A., 1976. Hill-tribe mint production and processing. Thai Journal of Agricultural Sciences 9(3): 127–144. |4| Do Tat Loi, 1995. Medicinal plants and drugs of Vietnam. Science and Technics Publishing House, Hanoi, Vietnam. pp. 747–750 (in Vietnamese). |5| Ellis, N.K., 1968. Production of essential oil from mint. In: Essential oils production in developing countries. Papers presented at the conference held in London, 15–19 May 1967. Tropical Products Institute, London, United Kingdom. pp. 1–10. |6| Keng, H., 1978. Labiatae. In: van Steenis, C.G.G.J. (General editor): Flora Malesiana. Series 1, Vol. 8. Sijthoff & Noordhoff International Publishers, Alphen aan den Rijn, the Netherlands. pp. 343–344. |7| Kishore, N., Mishra, A.K. & Chansouria, J.P., 1993. Fungitoxicity of essential oils against dermatophytes. Mycoses 36(5–6): 211–215. |8| Kykreja, A.K., Phawan, O.P., Ahuja, P.S.,

Sharma, S. & Mathus, A.K., 1992. Genetic improvement of mints: On the qualitative traits of essential oil of in-vitro derived clones of Japanese mint (*Mentha arvensis* L. var. *piperascens* Holmes). Journal of Essential Oil Research 4(6): 623–629. |9| Lim-Sylianco, C.Y., Blanco, F.R.B. & Lim, C.M., 1986. Mutagenicity, clastogenicity and antimutagenicity of medicinal plant tablets produced by the NSTA pilot plant. I. Yerba buena tablets. Philippine Journal of Science 115(4): 299–305. |10| Visuttipitakul, S., Britten, E.J. & Chaimongkol, S., 1990. Comparative oil production of varieties of Japanese mint, spearmint and peppermint at low and medium elevations in the tropics. Thai Journal of Agricultural Science 23(3): 233–241.

**Other selected sources** 79, 178, 179, 190, 202, 211, 212, 231, 332, 350, 382, 431, 513, 526, 534, 549, 580, 803, 858, 900, 1035, 1126, 1142, 1178, 1387, 1482, 1508, 1534.

La Dinh Moi

### **Mimosa pudica L.**

Sp. pl. 1: 518 (1753).

LEGUMINOSAE

2n = 52

**Vernacular names** Sensitive plant (En). Sensitive (Fr). Brunei: puteri malu, rumput malu, sopan malu (Malay). Indonesia: putri malu (general), jukut riyud (Sundanese), pis kucing (Javanese). Malaysia: memalu, malu-malu. Papua New Guinea: matmat (Gunantuna, New Britain). Philippines: makahiya (Tagalog), torog-torog (Bikol), babain (Ilokano). Cambodia: smau bânla, bânkráp. Laos: f'a:z langab, th'ûb nhub. Thailand: ka-ngap (peninsular), maiyaraap (central), yaa pan yot (northern). Vietnam: c[aa]y m[aws]c c[owr], c[aa]y x[aa]s[u] h[oor], c[aa]y tr[inh] n[uwx].

**Origin and geographic distribution** Sensitive plant probably originated in South America but is now pantropical. It occurs commonly throughout South-East Asia, usually along roadsides and on wasteland.

**Uses** In Indonesia, Malaysia and Thailand, sensitive plant was traditionally used to treat insomnia. Twigs were placed under the sleeping mats of children, and they were also used as a decoction to prepare a bath for children with sleeping problems. This use was probably based on signature: the leaves 'go to sleep' in the evening. At the beginning of the 20th Century, twigs were being sold for this purpose in Java, but there is no recent

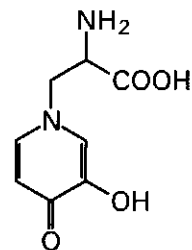
confirmation of this traditional use. In Vietnam, however, the leaves are considered in folk medicine to be sedative and hypnotic, and an infusion of the leaves is still regarded beneficial for patients suffering from insomnia. Test with mice seem to confirm this activity.

Other uses reported are the treatment of haematuria (all parts of the plant, Indonesia and Thailand) and as a poultice to treat swellings (pounded leaves, Indonesia, Malaysia and Thailand). In Brunei, a root decoction is drunk to relieve asthma and diarrhoea. The most extensive use of the plant in South-East Asia is in the Philippines, where a decoction of the entire plant is considered anti-asthmatic and a root decoction is given as a diuretic and also to treat dysmenorrhoea. In Thailand, the whole plant is used as a diuretic. The leaves are used externally to treat dermatitis, wounds and ulcers. The roots are traditionally used in Vietnam to treat arthritis. In India, a root decoction is used to treat urinary complaints and as an aphrodisiac, whereas it is considered emetic in South America and is also used there to treat diarrhoea and dysentery. In India, crushed leaves are applied as an emollient to glandular swellings, and an infusion of the leaves is used in Vietnam to treat febrile stiffness. In New Britain, roots and leaves have been used externally to treat swollen testicles. In Indo-China seeds are considered to be emetic; in India they are also used to treat sore throat and hoarseness. In the traditional pharmacopoeia of La Réunion, the stems, leaves and roots are mentioned as a calming remedy against insomnia, spasms and convulsions of children.

Sensitive plant contains tannins that can be used in the production of leather. Young stems and leaves are useful as forage. When the prickles on the stem and the fruits become too hard, they can cause intestinal inflammation in animals. Moreover, the mimosine present in the plant may cause poisoning. Sensitive plant is often considered as a noxious weed. In areas where other leguminous plants establish with difficulty, it can be of use as a cover crop or green manure. In Thailand, it is used as ground cover on road verges.

**Production and international trade** Sensitive plant is harvested in small amounts for personal use, and is only very rarely traded.

**Properties** Sensitive plant contains mimosine (N-(3-alanyl)-3-hydroxy-4-pyridone), an amino acid which is biosynthetically derived from lysine. This compound is reported as being toxic to several animal species, including pigs and rabbits, and to a lesser extent, ruminants such as cattle. Mi-



mimosine

mosine is reported to have depilatory properties, and prolonged use may lead to alopecia. The intoxication also manifests itself by loss of appetite and weight, and retarded growth. These symptoms are accompanied by an enlargement of the thyroid gland, and lowered serum thyroid hormone levels. Mimosine itself inhibits the synthesis of proteins and nucleic acids, but is not known as a goitrogen. Gut bacteria, however, transform mimosine enzymatically into 3-hydroxy-4(1H)-pyridone (= 3,4-dihydroxypyridine, DHP), which is a potent goitrogen able to cause hypo-thyroidism. Oral application of 1 ml of a DHP solution (0.25 mmol) in the rat has been found to significantly inhibit  $^{125}\text{I}$  uptake by the thyroid gland. Bacteria able to further metabolize DHP into non-goitrogenic compounds have been isolated from the gut of various animals, including Indonesian goats. When these bacteria were transferred into Australian cattle, the cattle were able to feed on mimosine-containing forage without suffering toxicological reactions.

Mimosine has been demonstrated to reversibly block cell cycle progression in mammalian cells in culture. It also, through iron chelation, blocks cell cycle progression in asynchronous human breast cancer cells. In addition to mimosine, two C-glycosylflavones have been isolated from the aerial parts of the plant, and identified as 2'-O-rhamnosyl-orientin and 2'-O-rhamnosyl-isoorientin.

In Vietnam, a significant hypotensive effect has been reported in experiments with dogs injected with a 10% infusion of leaves. Sedative, anti-inflammatory, anti-implantation and anti-arthritic effects have also been reported. Tests with mice showed prolongation of the time spent sleeping. An alcoholic extract, a petroleum-water extract and quaternary alkaloids isolated from whole dried plants in Thailand lowered blood sugar in diabetic rats, beginning in the second hour after single oral application and reaching a maximum after 6 hours; no abnormal symptoms were observed. The total alkaloidal extract of the roots



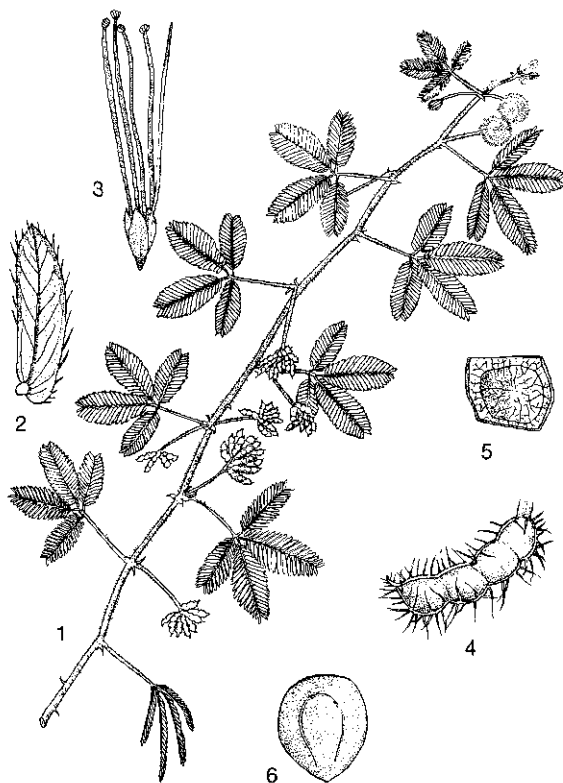
was found to be antagonistic to both acetylcholine and histamine on isolated guinea-pig ileum. In tests with rats in India extracts were not effective in preventing bladder stone deposition or in dissolving preformed stones. The methanol extract of aerial parts showed diuretic activity and some protection against the injurious effect of radiations.

In tests in Nigeria, the extract exhibited antimicrobial activity against *Vibrio cholerae*, but the potential in the control of cholera needs to be determined. An in vitro bioassay of an aqueous methanol extract of green leaves in Jamaica showed that the extract inactivated 50% of the filariform larvae of the nematode *Strongyloides stercoralis* (causing strongyloidiasis) in less than 1 hour. The root extract is also nematocidal. At a concentration of 300 ppm it completely inhibited egg hatch of *Meloidogyne incognita*, and also significantly affected the infectivity and development of larvae. A crude ethanol extract of the leaves was found to have insecticidal activity against adult *Tribolium confusum* when sprayed as a 10% concentrate; it killed 60% of the beetles.

Very little is known about the forage quality of sensitive plant. It is likely that the quality of any sample would vary considerably with the leaf/stem ratio.

**Adulterations and substitutes** Mimosine and its optically inactive form leucaenine are also known from *Leucaena leucocephala* (Lamk) de Wit.

**Description** An annual or perennial herb, sometimes woody at base and then subshrubby, up to 1(-1.5) m tall, often prostrate or straggling; stem usually sparsely armed with recurved prickles up to 5 mm long, glabrous to densely hispid. Leaves alternate, bipinnate, unarmed, sensitive; petiole (1.5-)3-5.5 cm long, hispid, rachis very short, giving the two pairs of pinnae a subdigitate position; stipules caducous; leaflets 10-26 pairs per pinna, oblong to subfalcate, 0.6-1.5 cm x 0.1-0.3 cm, margins setulose. Inflorescence an axillary globose head, about 1 cm in diameter, 1-2(-5) together per axil; peduncle up to 4 cm long. Flowers bisexual, 4-merous, sessile, lilac, pink or blue-purple; calyx inconspicuous, about 0.1 mm long; corolla narrowly campanulate, 2-2.5 mm long, with obtuse to rounded lobes; stamens free, much longer than corolla; ovary superior, glabrous, style long and slender. Fruit a flattened oblong pod, 1-1.8 cm x 0.3-0.5 cm, several together in a cluster, densely setose, prickly on margins, consisting of 3-5 1-seeded joints which break



*Mimosa pudica* L. - 1, flowering and fruiting stem; 2, leaflet; 3, flower; 4, pod; 5, one-seeded pod segment; 6, seed.

away from the persistent sutures. Seeds suborbicular to broadly ellipsoidal, flattened, 2.5-3 mm long, pale brown, surface finely granular. Seedling with epigeal germination, smelling like garlic; cotyledons ovate, sagitate at base, truncate to emarginate at apex, glabrous; hypocotyl up to 15 mm long, hairy, epicotyl absent; first leaf solitary, with 3 pairs of leaflets.

**Growth and development** Sensitive plant flowers throughout the year and it can complete its life cycle in 3 months. The leaves are extremely sensitive to the touch. At nightfall the leaflets fold up and the rachises bend down.

**Other botanical information** Within the subfamily *Mimosoideae*, *Mimosa* is classified in the tribe *Mimoseae*. It is a large genus of about 400 species mainly occurring in tropical America.

Four varieties of *M. pudica* are distinguished: var. *pudica* (only known from the sterile type specimen, no distinction possible); var. *hispida* Brenan (corolla in bud densely grey puberulous; heads in bud densely bristly; stipules 8-14 mm long); var.

*tetrandra* (Humb. & Bonpl. ex Willd.) DC., synonym: *Mimosa tetrandra* Humb. & Bonpl. ex Willd. (corolla in bud densely grey puberulous; heads in bud sometimes sparsely bristly; stipules 4–8 mm long); var. *unijuga* (Duchass. & Walp.) Griseb., synonym: *Mimosa unijuga* Duchass. & Walp. (corolla in bud glabrous; heads in bud not bristly; stipules 4–8 mm long). The three latter varieties are all pantropical and occur throughout South-East Asia.

**Ecology** Sensitive plant is common in wasteland, disturbed areas and overgrazed sites with moderately to poorly fertile soils. It occurs in the humid tropics and tolerates waterlogging but is not well adapted to the seasonally dry tropics. In the Malesian region, sensitive plant is found up to 1000 m altitude. It is regarded as a weed in upland field crops, in rainfed wetland rice and in plantation crops where it is reasonably tolerant of shade. It occurs over a wide range of soils, but iron chlorosis has been noted on coralline soils with a high pH.

**Propagation and planting** The persistence and spread of sensitive plant is aided by its prolific seed set. There are approximately 110 seeds/g.

**Husbandry** Sensitive plant is not sown deliberately, nor is its establishment or spread promoted. On the contrary, it is usually regarded as a noxious weed. It tends to invade pastures of declining soil fertility and is less common where soil fertility is good and pastures are not overgrazed. Sensitive plant has been eliminated from pastures where soil fertility is very poor and grazing pressure is very high. It grows with a wide range of grasses, including signal grass (*Brachiaria decumbens* Stapf) provided that grass is not too vigorous. It can hamper the establishment of improved species. Sensitive plant is more readily accepted if it is grazed continuously rather than rotationally. Observations suggest that it may at times stimulate growth of associated grasses.

**Diseases and pests** *M. pudica* plants in Bogor (Indonesia) are often found attacked by a fungus (*Ramularia mimosae*) which grows on the upper surface of the leaflets, forming irregular white spots. This fungus originates from tropical America and is also reported on *M. pudica* in India.

**Harvesting** Sensitive plant is harvested from populations in the wild.

**Handling after harvest** In the Philippines, harvested plants intended for preparing an ointment or cream for external use against skin complaints are air dried, cut into small pieces and ground in a grinding machine. The powdered

plant is subsequently submerged in 80% ethyl alcohol, and the mixture heated for one hour. The extract obtained is a yellowish-green syrupy substance with a bitter taste. Powdered dried roots should be stored in airtight containers, preferably in sterilized bottles.

**Genetic resources and breeding** Being a common weed, sensitive plant is not at risk of genetic erosion. There are no known germplasm collections of sensitive plant, nor breeding programmes.

**Prospects** Very little research has been done on the medicinal properties of sensitive plant. The scarce information from research suggests interesting possibilities as an anti-microbial, nematocidal and insecticidal agent, but the true value of the plant should be established by further investigations. Other attributed properties, such as the sedative activity, seem to have some scientific basis, but the indications are still vague.

Although sensitive plant can provide useful forage, it will continue to be primarily regarded as a weed. Although more appropriate management may promote its use, it would be preferable to grow forages that are more readily accepted by grazing animals or more suited to cut-and-carry feeding systems.

**Literature** |1| Bruneton, J., 1995. Pharmacognosy, phytochemistry, medicinal plants. Technique & Documentation Lavoisier, Paris, France. p. 164. |2| Dechatiwong Na, Ayutthaya, K., Thap-phayutphichan, P., Hinchiraran, T., Rattanaraphi, S. & Phidet, P., 1988. Decreasing blood sugar property and toxicity of *Mimosa pudica*. Warasan Pheasat Witthaya [Thai Journal of Pharmacology] 10: 33–43 (in Thai). |3| de Padua, L.S., Lugod, G.C. & Pancho, J.V., 1977. Handbook on Philippine medicinal plants. Vol. 1. Documentation and Information Section, Office of the Director of Research, University of the Philippines at Los Baños, the Philippines. p. 39. |4| Englert, J., Jiang, Y., Cabalion, P., Oulad-Ali, A. & Anton, R., 1994. C-glycosylflavones from aerial parts of *Mimosa pudica*. *Planta Medica* 60(2): 194. |5| Jones, R.M. & Aguilar, N.O., 1992. In: 't Mannetje, L. & Jones, R.M. (Editors): Plant Resources of South-East Asia No 4. Forages. Pudoc Scientific Publishers, Wageningen, the Netherlands. pp. 167–169. |6| Le Thi Hoan & Davide, R.G., 1979. Nematicidal properties of root extracts of seventeen plant species on *Meloidogyne incognita*. *Philippine Agriculturist* 62(4): 285–295. |7| Nguyen Van Duong, 1993. Medicinal plants of Vietnam, Cambodia and Laos. Mekong Printing, Santa Ana, California,

United States. pp. 267–268. |8| Nielsen, I.C., 1992. Mimosaceae (Leguminosae - Mimosoideae). In: de Wilde, W.J.J.O., Nooteboom, H.P. & Kalkman, C. (Editors): Flora Malesiana. Series 1, Vol. 11(1). Foundation Flora Malesiana, Leiden, the Netherlands. pp. 183–186. |9| Quashem, K.M.A., Hasan, Q. & Ahmed, S.U., 1977. Antagonism of acetylcholine and histamine hydrochloride by the total extract of *Mimosa pudica* Linn. on the isolated ileum of guinea pigs. Bangladesh Journal of Agricultural Sciences 4(1): 69–74. |10| Robinson, R.D., Williams, L.A.D., Lindo, J.F., Terry, S.I. & Mansingh, A., 1990. Inactivation of *Strongyloides stercoralis* filariform larvae in-vitro by six Jamaican plant extracts and three commercial anthelmintics. West Indian Medical Journal 39(4): 213–217.

**Other selected sources** 202, 306, 569, 580, 597, 691, 788, 1126, 1177, 1178, 1214, 1434, 1550, 1579.

Erlin Rachman

## Momordica L.

Sp. pl. 2: 1009 (1753); Gen. pl. ed. 5: 440 (1754).

CUCURBITACEAE

$x = 7, 11, 14$ ; *M. charantia*:  $2n = 22$ , *M. cochinchinensis*:  $2n = 28$

**Major species** *Momordica charantia* L., *M. cochinchinensis* (Lour.) Spreng.

**Vernacular names** *Momordica* (En).

**Origin and geographic distribution** *Momordica* comprises some 45 species and is confined to the Old World tropics, except for the few species introduced into the New World tropics. The majority of species are found in the warmer parts of Africa; Asia harbours only 5–7 species, 3 of which are reported from the Malesian region. *M. charantia* was probably first domesticated in eastern India and southern China, and then taken to other regions in tropical Asia and Africa from which it occasionally naturalizes. It is thought to have been introduced into Brazil from Africa with the slave trade, and that bird dispersal of the seeds accounts for its spread within South America. *M. cochinchinensis* occurs wild and cultivated from India to Japan and throughout Malasia.

**Uses** A decoction of the root, stem, leaves and fruit of *M. charantia* may be used as a febrifuge. Most plant parts act as a laxative. Juice from various plant parts is used externally to treat skin disorders, abscesses and burns, and also as a cure for

children with diarrhoea and stomach-ache. Leaf juice has been applied as a gargle against sprue, to treat jaundice, and for 'female disorders', whereas the flowers are part of a mixture to treat asthma. In Peninsular Malaysia, a decoction of the leaves has been used as an abortifacient. In Indonesia, it is considered to be appetizing, depurative, mildly laxative, useful in treating liver diseases and biliousness; it is also used as a vermifuge to treat pinworms. A little leaf juice is given orally to new-born babies to cleanse the stomach and bowels. Fruits are considered tonic, stomachic, carminative and cooling, and are applied in the treatment of inflammation, rheumatism, gout, pruritus, dermatitis and liver and spleen diseases. The fruits, leaves and roots have long been used in India and Puerto Rico as a folk medicine for diabetes mellitus, though large doses are toxic. In the Philippines, the fruit and young shoots, either in the form of a decoction or as tablets, are used for mild non-insulin dependent diabetes mellitus.

Seeds of *M. cochinchinensis* are used in local medicine in Burma (Myanmar), Thailand and the Philippines to treat chest complaints, whereas in China and Peninsular Malaysia they are a remedy for abdominal pains, dysentery, mesenteric enlargements, obstructions of liver and spleen and haemorrhoids. They are further used to treat chronic malaria, and after being ground and soaked in alcohol or water they are applied externally to wounds, bruises, burns, skin trouble, ulcers, breast cancer, abscesses, mumps and lumbago. Seeds are indicated as cooling, resolvent, laxative and poisonous. The root is used as an expectorant. Roots and leaves have been reported to be useful in the treatment of oedema of the legs, a kind of rheumatism.

The immature fruits of *M. charantia* and *M. cochinchinensis* are a well-known vegetable, whereas leaves and flowers are also eaten as a vegetable or flavouring agent.

**Production and international trade** *Momordica* fruits or any of its derived products for medicinal applications are only traded locally.

**Properties** The seeds and the fruit wall of *M. charantia* are reported to contain a resin, a saponin glycoside of the cucurbitacin type, and alkaloids that may cause vomiting and diarrhoea. Furthermore, several proteins that display a variety of pharmacological effects can be isolated from *Momordica*.

The proteins  $\alpha$ -momorcharin and  $\beta$ -momorcharin, from seeds of *M. charantia*, have been found to show a hepatotoxic effect on isolated rat hepato-

cytes. Several immunotoxins were prepared by linking the type 1 ribosome-inactivating protein momordin I to antibodies specific to various cell lines, e.g. bladder carcinoma antibody, CD5-, and CD22-monoclonal antibodies. Treatment with these immunotoxins significantly inhibits tumour development in vitro, e.g. CD5- or CD22-expressing cell lines, with  $IC_{50}$  values generally at picomolar scale. The treatment alone, or in combination with a general cytostatic significantly inhibits tumour development in vivo, e.g. in mice with transplanted CD22-expressing cells.

The glycoprotein momorcochin-S, purified from the seeds of *M. cochinchinensis*, shows ribosome-inactivating effects. The protein was linked to a monoclonal antibody (8A) against human plasma cells, and the resulting immunotoxin was found selectively toxic to the target cells. Furthermore,  $\tau$ -momorcharin, a small ribosome-inactivating protein, can inhibit the protein synthesis in the rabbit reticulocyte cell-free system with an  $ID_{50}$  of 55 nM.

Momordin-folate, the conjugate of folic acid with the cytotoxic protein momordin, can selectively kill HeLa and KB cells, two malignant human cell lines, in co-cultures with W138 and Hs67 cells, two normal human cell types. The in vivo antitumour activity of a crude extract from *M. charantia* was significant for several types of tumour cells in mice at an optimum dose of 8  $\mu$ g protein administered biweekly and intraperitoneally. It is thought that in vivo enhancement of immune functions may contribute to the antitumour effects of the *M. charantia* extract.

Expressed juices from *M. charantia* fruits appreciably reduced the incidence of skin tumours in mice initiated by dimethylbenz[a]anthracene and promoted by croton oil. The extracts of the peel, pulp, seed and whole fruit of *M. charantia* exhibited marked anti-carcinogenic activity against mouse skin papilloma genesis when applied topically at 100  $\mu$ m<sup>2</sup>/animal. In vitro and in vivo tests with aqueous and ethanolic extracts of *M. cochinchinensis* also showed marked antitumour activity.

MAP30, an antiviral protein (30 kDa) from *M. charantia*, may regulate Herpes simplex virus (HSV) replication in concert with dexamethasone and indomethacin, which are inhibitors of prostaglandin synthesis.  $EC_{50}$  values for MAP30 were 0.1  $\mu$ M for HSV-2 and 0.3  $\mu$ M for HSV-1 in human lung WI-38 fibroblasts. MAP30 is also capable of inhibiting infection of HIV-1 in T lymphocytes and monocytes, as well as replication of the

virus in infected cells. It was found not toxic to normal uninfected cells; the peptide is probably unable to enter healthy cells. It exhibits a dose-dependent inhibition of integration of viral DNA into the host chromosomes (HIV-1 integrase), which is a vital step in the replicative cycle of the AIDS virus.

*M. charantia* trypsin inhibitor-II prolonged the prothrombin time of human plasma. Furthermore, a glycoprotein isolated from fresh tuberous roots of *M. cochinchinensis* was capable of inducing mid-term abortion in mice.

Acylglucosylsterols isolated from the green fruits of *M. charantia* showed antimutagenic activity and reduced the number of micronucleated polychromatic erythrocytes induced by the well-known mutagen mitomycin C by about 80% in mice, at a dosage range of 12.5–50  $\mu$ g extract/g. Powder and defatted extract from *M. charantia* leaves reduced the genotoxic activity of dimethylnitrosamine, methylmethanesulphonate and tetracycline as shown by the reduction of chromosome-breaking effects. *M. charantia* has also been screened for its genotoxic activity using a plate incorporation assay involving *Aspergillus nidulans*. The aqueous extract of *M. charantia* leaves resulted in a significant increase in the frequency of segregant sectors per colony.

Bitter gourd (*M. charantia*) is often used in folk medicine to treat diabetes. However, its hypoglycaemic activity seems contradictory. The hypoglycaemic activity of bitter gourd in experimental animals is also contradictory. The pulp juice of *M. charantia* lowered fasting blood glucose levels in normal rats; the effect was more pronounced using saponin-free methanol extract of the pulp juice. The hypoglycaemic effect was also significant in normal rats fed glucose 45 minutes after the extract had been administered. In the insulin-dependent diabetes mellitus model rats, the pulp juice had no significant effect on blood glucose levels in the fasting, and postprandial states. In the non-insulin dependent model rats, however, the saponin-free methanol extract of juice produced a significant hypoglycaemic effect in both these states. Seed and whole plant extracts showed a small but consistent tendency to increase blood glucose levels in the normal rats. In normal mice, an aqueous extract of bitter gourd lowered the glycaemic response to both oral and intraperitoneal glucose, without altering the insulin response. This aqueous extract and the residue after alkaline chloroform extraction had reduced the hyperglycaemia in diabetic mice after 1 hour. The re-

sults suggest that bitter gourd extracts administered orally lower glucose concentrations independently of intestinal glucose absorption and involve an extrapancreatic effect. Two glycosides isolated from the tuber of *M. cochinchinensis* containing oleanolic acid as aglycone showed hypoglycaemic activity in streptozotocin-induced diabetic rats at a dose of 25 mg/kg; intraperitoneal administrations showed higher activity than oral administration. The alcoholic extract of *M. charantia* administered orally to female Wistar rats at 500 mg/kg reduced glucose levels by 10–16% and 6% after 1 and 2 hours, respectively, in normal rats and by 26% after 3.5 hours in streptozotocin-induced diabetic rats. The extract increased the rate of glycogen synthesis from 14C-glucose in the liver of normally-fed rats by 4–5 times, suggesting that the extract may act at least in part by enhancing glucose utilization in the liver. In normal mice, intraperitoneal administration of a concentrated aqueous extract of bitter gourd improved glucose tolerance after 8 hours, and in streptozotocin-induced diabetic mice the level of hyperglycaemia was reduced by 50% after 5 hours. The bitter gourd extracts did not significantly alter plasma insulin concentrations, suggesting that they may exert an extrapancreatic effect to promote glucose disposal. Oral administration of the aqueous extract (0.5 g/kg) of bitter gourd reduced the fasting glucose levels of hyperglycaemic and normoglycaemic mice; the ethanol extract had no significant effect on glucose levels. The water extract did not improve the tolerance of mice to oral glucose.

In conclusion, a significant number of studies have established the hypoglycaemic activity of bitter gourd; its effect appears to be more acute and transient than cumulative. The fresh aqueous extract of the whole fruit is more effective than dried powder, or dietary consumption. Some studies found that the seed also contained hypoglycaemic principles. In most of the cases where hypoglycaemic activity could not be demonstrated, normoglycaemic animals were experimented upon. The mechanism of hypoglycaemic activity remains unclear. If the hypoglycaemic action of bitter gourd is mediated through its effect on glucose absorption and alterations to the activities of enzymes involved in glucose metabolism, it would be beneficial to both insulin-dependent as well as non-insulin-dependent diabetics. If, however, it has an insulin secretagogue effect, only non-insulin-dependent diabetics would benefit from it.

Nine diabetic patients underwent 3 glucose toler-

ance tests with 50 g of glucose, 50 ml bitter gourd juice, and a test after 8–11 weeks of consuming fried bitter gourd (250 g) daily. The results indicated that fresh bitter gourd juice brought about a significant reduction in plasma glucose concentration, and an improvement in the response to an oral glucose load. The effect of fried bitter gourd was not so pronounced, although it was significant. A cumulative and gradual hypoglycaemic effect was found in diabetic patients using the aqueous extract at the end of a 3-week trial. When 8 non-insulin dependent diabetes mellitus patients consumed bitter gourd cooked and then fried in oil along with their regular meal, their plasma glucose levels were significantly reduced after 1 hour as compared to those whose meal was not supplemented with bitter gourd. However, improvement in glucose tolerance was not significant. Contradictory to these findings, however, is a study in which bitter gourd in the form of fresh juice, dried powder or the powder given as a tablet did not have any beneficial influence on diabetic patients.

No antimalarial activity could be demonstrated in mice infected with *Plasmodium berghei* when extracts of *M. charantia* were administered orally at 1 g/kg for 5 consecutive days. *M. charantia* fruits and seeds contained components that inhibited hormone-induced lipolysis in isolated rat adipocytes. The haemolytic activity of a fraction obtained from fresh tubers of *M. cochinchinensis* can be attributed to a sterol-glycoside. The crude saponin (4–6% in dry roots) obtained from *M. cochinchinensis* also showed haemolytic activity.

Extracts of *M. charantia* were effective in treating *Ascaridia galli* worms in birds. Oral administration of *M. charantia* extract containing 100 mg iron was as effective as a commercial preparation to prevent anaemia in piglets. Chitinase isolated from *M. charantia* fruits may be strongly bacteriostatic. Pollen from *M. charantia* completely inhibited spore germination of the pathogenic fungi *Cochliobolus lunatus*, *Cylindrocarpon lichenicola*, *Fusarium solani* and *Myrothecium leucotrichum*.

Momordicines I and II were isolated from dried leaves of *M. charantia*. Momordicine I at 0.5 mg/ml and 1 mg/ml showed about 33% and 59% inhibition of *Colletotrichum gloeosporioides*, respectively, whereas momordicine II at 0.25 mg/ml and 0.5 mg/ml showed about 17% and 23% inhibition, respectively. Against *Cladosporium cucumerinum*, momordicine I exhibited activity at 0.5 mg/ml, whereas momordicine II did not show any activity up to 1 mg/ml. It was suggested that the

difference in activity between the 2 compounds could be due to the higher lipophilicity of the aglycone. The leaf extracts were also effective against *Botryodiplodia theobromae*, *Curvularia lunata*, *Phytophthora colocasia* and *Sclerotium rolfsii*. The active constituents of the leaves of *M. charantia* were extracted by cold maceration using 95% ethanol. This extract has antimicrobial activity against *Escherichia coli*, *Salmonella paratyphi* and *Shigella dysenteriae*. Seed extracts of *M. charantia* resulted in high mortality of the nematodes *Meloidogyne incognita* and *Rotylenchulus reniformis*. The petroleum ether extract of *M. charantia* was active against the bean weevil *Callosobruchus chinensis*.

**Adulterations and substitutes** Extensive studies on immunotoxic and anti-HIV potential of proteins from *Momordica* may lead to substitution for *Trichosanthes* and vice versa.

**Description** Monoecious or dioecious, annual or perennial herbs, with climbing or trailing stems. Tendrils lateral, one at each node, simple or bifid. Leaves alternate, blade simple or palmately lobed to 3–7(–15)-foliolate, petiolate, exstipulate. Inflorescence axillary. Flowers unisexual, actinomorphic, often subtended by a conspicuous bract. Male flowers solitary, umbellate or in short racemes or fascicles; hypanthium shallow; calyx 5-lobed; petals 5, free, white to yellow, 1–3 of them with a scale at base; stamens 3, inserted towards the base of the hypanthium, 2 bilocular, 1 unilocular, locules curved or flexuose; pistillode absent. Female flowers solitary; perianth usually similar to the male flowers; staminodes 3; ovary inferior, oblong to fusiform, 3-locular with many horizontal ovules, stigmas 3, entire to 2-lobed. Fruit a berry (pepo), ovoid-ellipsoid to elongate-fusiform, fleshy, ornamented with tubercles, spines, wings or ridges, indehiscent or dehiscent by 3 valves and exposing the seeds enveloped in pulp. Seeds arillate, usually compressed, with sculptured testa and grooved margins.

**Growth and development** Flowering of *M. charantia* starts within 2 months from sowing, that of *M. cochinchinensis* after about 2 months. Flowers of *M. charantia* start opening early in the morning. Anthers dehisce about 2 hours before anthesis. Flowers of *Momordica* are pollinated by insects, especially bees. Indehiscent *Momordica* fruits may be shattered and eaten by large birds or mammals. Seeds within dehiscent fruits of *M. charantia* strongly contrast with the large red aril and are thus easily spotted by birds who eat and disperse them. At higher latitudes, plants of *M.*

*cochinchinensis* remain dormant in winter and regrow from the tuberous root in spring. They fruit mainly during the rainy season.

**Other botanical information** *Momordica* belongs to the tribe *Joliffieae* of the subfamily *Cucurbitaceae*. The tribe contains the least specialized genera of the *Cucurbitaceae*. *Momordica* is closely related to *Thladiantha*, but the latter has 5 stamens or staminodes and straight or only slightly curved anther locules.

**Ecology** *M. charantia* grows well in tropical and subtropical climates. It is adapted to a wide range of environments and can be grown year-round, but is usually cultivated during the warmer season, up to an altitude of 1500 m. It is sensitive to waterlogging. It tolerates a wide range of soils but it thrives in a well-drained sandy loam, rich in organic matter. *M. cochinchinensis* prefers a warm humid climate with temperatures ranging from 20–35°C and an average rainfall of 1500–2500 mm. It does not tolerate waterlogging and grows well in fertile, well-drained, sandy loams with pH near neutral.

**Propagation and planting** *M. charantia* is most commonly propagated by seed. Pre-germinated seed results in even establishment. Optimum plant density differs per cultivar, but ranges from 6500–11 000 plants per ha. *M. cochinchinensis* is mainly propagated by its tuberous roots. Since it is dioecious, tubers from male and female plants should be planted together. About 50 000 sprouted tubers per ha are required, but in India a spacing of 1.5 m × 2.5 m is adopted. Mean germination of the seeds of *M. cochinchinensis* is 50%, and germination may take up to 1 year; cuttings root for about 80%.

**In vitro production of active compounds** The antiviral protein MAP30, from *M. charantia*, has been cloned and expressed. It has similar anti-HIV, anti-viral and anti-tumour activity as the natural MAP30.

**Husbandry** Fertilization and furrow irrigation if necessary are important cropping techniques when growing *M. charantia* on trellises. Wild *M. charantia* can become a troublesome weed in large-scale plantations of e.g. rubber and oil palm in Indonesia and possibly in other South-East Asian countries too.

**Diseases and pests** Serious diseases of bitter gourd are *Cercospora* leaf spot, downy mildew (caused by *Pseudoperonospora cubensis*) and bacterial wilt (caused by *Pseudomonas solanacearum*). Fruit fly (*Dacus cucurbitae*) is the most destructive insect pest of bitter gourd, whereas

root-knot nematodes (*Meloidogyne incognita*) also attack the crop.

**Harvesting** *M. charantia* usually takes 15–20 days after fruit set to mature, whereas *M. cochinchinensis* fruits are harvested when they turn yellow or red.

**Yield** A fruit yield of 20–30 t/ha is considered satisfactory for *M. charantia*. Some F<sub>1</sub> hybrids yield up to 40 t/ha. The number of fruits per plant may reach 20–25 during the cropping period. *M. cochinchinensis* may yield 30–60 fruits per plant, each weighing 1–3 kg. In Japan, the yield of dry roots of 5-year-old plants of *M. cochinchinensis* was about 10 t/ha.

**Handling after harvest** Mature fruits are split and seeds and fruit pulp are separated. Fruit pulp is dried at low temperatures, and an oil can be extracted. This oil, which is rich in  $\beta$ -carotene, is used to treat rachitis, xerophthalmia and night blindness. Seeds are dried in the sun or in ovens. The oil that can be extracted from the kernel is used to treat skin disorders. Roots are washed and dried in the sun or oven, and stored for later use.

**Genetic resources and breeding** The world collection of *Momordica* germplasm is held at NBPGR, New Delhi, India. In South-East Asia, collections are available in the Philippines (NPGRL-IPB, Los Baños) and in Thailand (Department of Horticulture, Kasetsart University, Bangkok). Elsewhere, collections are held in several institutes in India, South Africa, Taiwan and the United States. In many South-East Asian countries, commercial F<sub>1</sub> hybrids often twice as productive as the traditional open-pollinated cultivars, have been released.

**Prospects** Since the results with *M. charantia* in the treatment of diabetes are still somewhat contradictory, more research needs to be done on its hypoglycaemic activity. Furthermore, several compounds from *Momordica* show interesting pharmacological activities, e.g. immunotoxic and anti-HIV, which merit further research, and may have potential in the development of future medicines.

**Literature** [1] Ali, L., Khan, A.K.A., Mamun, M.I.R., Mosihuaman, M., Nahar, N., Alam, M.N. & Rokeya, B., 1993. Studies on hypoglycemic effects of fruit pulp, seed, and whole plant of *Momordica charantia* on normal and diabetic model rats. *Planta Medica* 59(5): 408–412. [2] Bolognesi, A., Barbieri, L., Carnicelli, D., Abbondanza, A., Cenini, P., Falasca, A.I., Dinota, A. & Stirpe, F., 1989. Purification and properties of a new ribosome-inactivating protein with RNA N-glycosi-

dase activity suitable for immunotoxin preparation from the seeds of *Momordica cochinchinensis*. *Biochimica et Biophysica Acta* 993(2–3): 287–292. [3] Bolognesi, A., Tazzari, P.L., Olivieri, F., Polito, L., Lemoli, R., Terenzi, A., Pasqualucci, L., Falini, B. & Stirpe, F., 1998. Evaluation of immunotoxins containing single-chain ribosome-inactivating proteins and an anti-CD22 monoclonal antibody (OM124): in vitro and in vivo studies. *British Journal of Haematology* 101(1): 179–188. [4] Bourinbaier, A.S. & Lee-Huang, S., 1995. The activity of plant-derived antiretroviral proteins MAP30 and GAP31 against Herpes simplex virus infection in vitro. *Biochemical and Biophysical Research Communications* 219(3): 923–929. [5] Cakici, I., Hurmoglu, C., Tunctan, B., Abacioglu, N., Kanik, I. & Sener, B., 1994. Hypoglycaemic effect of *Momordica charantia* extracts in normoglycaemic or cyproheptadine-induced hyperglycaemic mice. *Journal of Ethnopharmacology* 44(2): 117–121. [6] Guevara, A.P., Lim-Sylianco, C., Dayrit, F. & Finch, P., 1990. Antimutagens from *Momordica charantia*. *Mutation Research* 230(2): 121–126. [7] Lee-Huang, S., Huang, P.L., Huang, P.L., Bourinbaier, A.S., Chen, H.C. & Kung, H.F., 1995. Inhibition of the integrase of human immunodeficiency virus (HIV) type 1 by anti-HIV plant proteins MAP30 and GAP31. *Proceedings of the National Academy of Sciences* 92(19): 8818–8822. [8] Platel, K. & Srinivasan, K., 1997. Plant foods in the management of diabetes mellitus: vegetables as potential hypoglycaemic agents. *Nahrung* 41(2): 68–74. [9] Reyes, M.E.C., Gilde-macher, B.H. & Jansen, G.J., 1993. *Momordica L.* In: Siemonsma, J.S. & Kasem Piluek (Editors): *Plant Resources of South-East Asia No 8. Vegetables*. Pudoc Scientific Publishers, Wageningen, the Netherlands. pp. 206–210. [10] Shubhashish Sarkar, Maddali Pranava & Marita, A.R., 1996. Demonstration of the hypoglycemic action of *Momordica charantia* in a validated animal model of diabetes. *Pharmacological Research* 33(1): 1–4.

#### *Selection of species*

#### **Momordica charantia L.**

Sp. pl. 2: 1009 (1753).

**Synonyms** *Momordica indica* L. (1754), *Momordica elegans* Salisb. (1796), *Momordica chinensis* Spreng. (1891).

**Vernacular names** Bitter gourd, bitter cucumber, balsam pear (En). Bitter melon (Am). Margose, paroka (Fr). Indonesia: paria (general), pare

(Javanese), papari (Moluccas). Malaysia: peria, peria laut, periok. Philippines: ampalaya (Tagalog), paria (Ilocano), palia (Bisaya). Cambodia: mreah. Laos: 'hail, 'phak 'ha, sai1. Thailand: mara (general), maha (northern), phakha (northeastern). Vietnam: m[uw] [ows]p d[aws]ng, m[uw] [ows]p m[ur] (northern), kh[oor] qua (south).

**Distribution** *M. charantia* was possibly first domesticated in eastern India and southern China. It now has a pantropical distribution, with wild and cultivated populations.

**Uses** See under genus treatment for the numerous medicinal uses. The immature fruits are a well-known vegetable. The pulpy arils can be eaten as a sweet. The seed mass of the ripe fruit can be used as a condiment. *M. charantia* is occasionally planted as an ornamental.

**Observations** A monoecious, annual vine up to 5 m long, stem 5-ridged, tendrils simple; leaf blade broadly ovate to suborbicular or ovate-reniform in outline, 2.5–10 cm × 3–12.5 cm, deeply palmately

(3–)5(–9)-lobed, deeply cordate at base, lobes obovate and sinuate-lobulate or sinuate-toothed, glabrous or sparsely pubescent; flowers solitary, 2–3.5 cm in diameter, yellow; male flowers on a 0.5–3 cm long peduncle bearing an apical bract of up to 2.2 cm long, pedicel 2–5.5 cm long; female flowers on a 0.2–5 cm long peduncle bearing an apical bract of up to 1.2 cm long, pedicel 1–10 cm long; fruit 3–11(–45) cm × 2–4(–8) cm, irregularly warty, orange, dehiscing; seeds 8–16 mm × 4–10 mm × 2.5–3.5 mm, brown, testa ornamented. *M. charantia* is found in lowland rain forest, riverine forest, thickets, hedges, waste places and roadsides and may be locally abundant.

**Selected sources** 48, 97, 103, 109, 110, 136, 170, 176, 177, 209, 234, 257, 287, 324, 340, 470, 498, 515, 563, 680, 707, 721, 808, 830, 835, 836, 847, 886, 946, 1024, 1027, 1035, 1086, 1121, 1128, 1134, 1143, 1144, 1153, 1164, 1178, 1196, 1216, 1341, 1362, 1443, 1485, 1540, 1589.

### *Momordica cochinchinensis* (Lour.) Spreng.

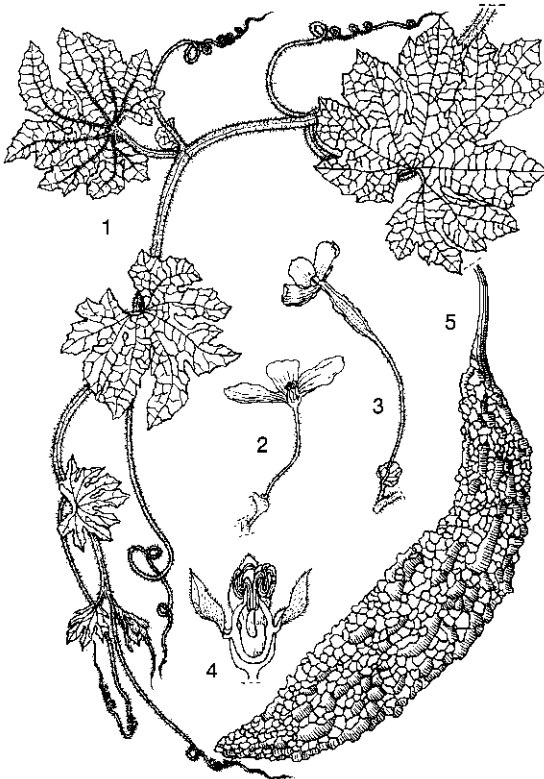
Syst. veg. 3: 14 (1826).

**Synonyms** *Muricia cochinchinensis* Lour. (1790), *Momordica mixta* Roxb. (1832), *Momordica meloniflora* Hand.-Mazz. (1921).

**Vernacular names** Sweet gourd, spiny bitter cucumber, giant spine gourd (En). Muricie (Fr). Indonesia: pupia (Malay, Moluccas), torobuk, toropu (North Halmahera). Malaysia: teruah. Philippines: buyok-buyok (Tagalog), paruk-paruk (Ilocano), taboguak (Bikol). Cambodia: makkao. Laos: khua mak 'khao', cup 'khoa' 'nhai'. Thailand: fak-khao (central), phak-khao (northern), khika-khrua (southern). Vietnam: d[aa]y g[aa]s (general), tr[as]i g[aa]s, m[oo]c m[ees]t t[ur].

**Distribution** *M. cochinchinensis* is found wild and cultivated from India to Indo-China, China, Japan, Taiwan, Thailand, scattered throughout the Malesian region (reported from Peninsular Malaysia, the Philippines, Sulawesi, Bali and the Moluccas, but probably present elsewhere as well) and in northern Australia (Cape York peninsula).

**Uses** See under genus treatment for the numerous medicinal uses. The immature fruits are a well-known vegetable. The seeds contain an oil which is used as an illuminant in Indo-China and may be applied in the formulation of paint and varnishes. The roots froth in water and may be used as a soap and to kill head lice. In Vietnam the aril of the seeds is used as a colouring agent for rice, called 'steamed momordica glutinous rice'.



*Momordica charantia* L. – 1, leafy shoot; 2, male flower in longitudinal section; 3, female flower in longitudinal section; 4, male flower in cross section with petals removed; 5, fruit.



**Observations** A dioecious, perennial vine arising from a tuberous root; stem robust, angular, tendrils simple; leaf blade suborbicular in outline, 12–20 cm in diameter, deeply palmately 3(–5)-lobed, cordate and with some glands at base, lobes subovate with entire or subdentate margins, glabrous, petiole with 2–5 glands near the middle; flowers solitary, about 8 cm in diameter, yellow, but blackish at base inside; male flowers on a peduncle 5–30 cm long bearing an apical, suborbicular, sessile bract, 3–4 cm × 4–5 cm, pedicel 3–10 mm long; female flowers similar but with a smaller bract; fruit 10–20 cm × 6–10 cm, yellow, turning red at maturity, densely covered with small tubercles; seeds about 25 mm × 20 mm × 5 mm, brown, testa sculptured. *M. cochinchinensis* is locally abundant in forest margins, along rivers, in open places, and in disturbed locations, at low altitudes.

**Selected sources** 169, 287, 363, 721, 776, 1000, 1024, 1026, 1035, 1128, 1178, 1216, 1223, 1443, 1629.

Nguyen Huu Hien & Sri Hayati Widodo

## Morus L.

Sp. pl. 2: 986 (1753); Gen. pl. ed. 5: 424 (1754).

MORACEAE

$x = 14$ ; *M. alba*:  $n = 14$ , *M. australis*:  $n = 14$ , *M. nigra*:  $2n = 89$ –106, 154, 308

**Major species** *Morus alba* L., *M. nigra* L.

**Vernacular names** Mulberry (En). Mûrier (Fr). Moral, morera (Sp). Indonesia: murbei (general). Malaysia: bebesaran (general). Philippines: amoras (Filipino). Cambodia: moon. Laos: mon. Thailand: mon. Vietnam: d[aa]u t[awf]m, d[aa]u t[af]lu, t[awf]m tang.

**Origin and geographic distribution** *Morus* comprises 10–15 species and is distributed in all tropical and temperate regions; in the tropics, mainly in montane habitats. Only one species (*M. macroura* Miq.) is native to the Malesian region; several others have been introduced and have occasionally naturalized.

**Uses** In most parts of South-East Asia it seems to be more important to cultivate mulberry trees (most often *M. alba*) for their leaves, which are used to rear silkworms (*Bombyx mori*), than for their medicinal application. Trials have been done on the species discussed below and (in Sumatra and Java) on *M. cathayana* Hemsl. and *M. latifolia* Poir. to ascertain their suitability for raising silkworms. Furthermore, the tasty fruits are high-

ly valued and made into juice, wine, jam, etc. In India the fruits are also used as a dye.

In general, the root bark, twigs and fruits are used as restorative, tonic, pectoral, diuretic, and are prescribed to treat cough, asthma, phthisis, and other chest complaints, dropsy and rheumatism. The leaves are depurative, cooling and resolvent.

The syrup made of fresh *Morus* fruits is used as a refrigerant in fevers and as an expectorant in coughs and sore throats. A drink made of it has similar applications. A decoction of the leaves of *Morus* is used for its blood-purifying properties as a febrifuge, diuretic and galactagogue. The leaves, bruised or withered over a fire and covered in coconut oil, are used to cover wounds and against insect bites, apparently for their anti-inflammatory properties. The root-bark is used in various applications for its general restorative properties, as a remedy against toothache and as emmenagogue, and furthermore in Vietnam for similar applications as the leaves, as diuretic, antitussive and expectorant and prescribed in oedema, high blood pressure, cough, bronchitis and asthma. The bark of *Morus* is used as a purgative and vermifuge. Root-bark, leaves and fruits of various *Morus* species are considered a remedy for diabetes. The root-bark of *M. alba* and possibly other *Morus* species is also known as the oriental drug 'sohakuhi', which has long been used for anti-inflammatory, diuretic, antitussive and antipyretic purposes in oriental medicine.

*M. alba* yields an attractive wood, and is also used in agroforestry. It is occasionally planted as a roadside tree. Its fibrous bark has been used to make paper. *M. macroura* wood yields a medium-quality firewood.

**Properties** Preliminary pharmacological investigations of n-butanol and water-soluble fractions of *M. alba* root showed cathartic, analgesic, diuretic, antitussive, antioedemic, sedative, anticonvulsant and hypotensive actions in mice, rats, guinea-pigs and dogs. These experimental results seem to show some correlation with the traditional clinical applications in Chinese medicine.

The anti-inflammatory activity of the methanol extract of *M. alba* root has been studied in rats using the following methods: rat paw oedema, inflammatory exudation, carrageenin-induced pleurisy, cotton pellet granuloma and chronic experimental arthritis. The extract has been found to be effective in carrageenin-induced oedema and this was not due to a counter-irritant effect. It was also effective against mediator-induced (histamine, serotonin, bradykinin) oedema; it reduced

the intensity of peritoneal inflammation and also inhibited the migration of leukocytes, suggesting an anti-exudative effect. The extract reduced the formation of granulation tissue and inhibited experimental arthritis, suggesting its effect on proliferative phases of inflammation and in arthritic conditions. Antipyretic studies revealed its potential to reduce body temperature in pyretic rats. The extract further possessed analgesic activity. A series of flavone derivatives have been isolated from the root-bark of *M. alba*: mulberrin, mulberochromene, cyclomulberrin, cyclomulberochromene, mulbeeranol, and phenolic compounds albactalol, albanol A and albanol B. The flavone morin (2',3,4',5,7-pentahydroxyflavone, an isomer of quercetin) has been identified in the heartwood. Morin shows anti-angiotensin properties, with activity on blood pressure and isolated tissues of the rat. The latter effect of morin seems to be a direct action on the muscle-relaxing system. Morusinol, an isoprene-substituted flavone is found in the root-bark of *M. alba* together with a number of related compounds. In general these flavones are known to have some anti-tumour activity.

Eighteen N-containing sugars have been isolated from the roots of *M. alba*, including seven that were isolated from the leaves of *M. bombycis* Koidz. These N-containing sugars are: 1-deoxynojirimycin, N-methyl-1-deoxynojirimycin, fagomine, 3-epi-fagomine, 1,4-dideoxy-1,4-imino-D-arabinitol, 1,4-dideoxy-1,4-imino-D-ribitol, calystegin B<sub>2</sub> (= 1 $\alpha$ ,2 $\beta$ ,3 $\alpha$ ,4 $\beta$ -tetrahydroxy-nor-tropane), calystegin C<sub>1</sub> (=1 $\alpha$ ,2 $\beta$ ,3 $\alpha$ ,4 $\beta$ ,6 $\alpha$ -pentahydroxy-nor-tropane), 1,4-dideoxy-1,4-imino-(2-O- $\beta$ -D-glucopyranosyl)-D-arabinitol, and nine glycosides of 1-deoxynojirimycin. These glycosides consist of 2-O- and 6-O- $\alpha$ -D-galactopyranosyl-1-deoxynojirimycins, 2-O-, 3-O- and 4-O- $\alpha$ -D-glucopyranosyl-1-deoxynojirimycins and 2-O-, 3-O-, 4-O- and 6-O- $\beta$ -D-glucopyranosyl-1-deoxynojirimycins. Due to the presence of nitrogen in the molecules, some authors also refer to these compounds as alkaloids instead of N-containing sugars; this makes 3-epi-fagomine a new member of the polyhydroxylated piperidine alkaloids. Furthermore, the isolation of 1,4-dideoxy-1,4-imino-D-ribitol is the first report of its natural occurrence.

It has recently been found that the polyhydroxy-nor-tropane alkaloids possess potent glycosidase-inhibiting activities. Inhibition of glycosidases might be part of the mechanism involved in anti-hyperglycaemic effects. Calystegin A<sub>3</sub> (from *Calystegia sepium* (L.) R. Br., Convolvulaceae) is an example of a trihydroxy-nor-tropane, and the ca-

lystegins B<sub>1</sub> (from *C. sepium*) and B<sub>2</sub> (from *C. sepium* and *M. alba*) are examples of tetrahydroxy-nor-tropanes. Calystegin C<sub>1</sub> (*M. alba*) is a new member of the calystegins, the first naturally occurring pentahydroxy-nor-tropane. The inhibitory activities of 3-epi-fagomine, calystegin B<sub>2</sub>, calystegin C<sub>1</sub>, and four glycosides of 1-deoxynojirimycin have been investigated against rat digestive glycosidases and various commercially available glycosidases. Calystegin C<sub>1</sub> was found to be a particularly potent inhibitor of rat digestive glycosidase. The  $\beta$ -D-glucoside of 1,4-dideoxy-1,4-imino-D-arabinitol, which is known to be a potent inhibitor of yeast  $\alpha$ -glucosidase and mouse intestinal isomaltase, completely lost inhibitory activity against rat glycosidases. When compared with the commercially available glycosidases, calystegin B<sub>2</sub> performed better than calystegin C<sub>1</sub>.

Diabetic patients experience xerostomia, a feeling of thirst and the need to frequently drink water. This syndrome is believed to be partially related to a reduced flow rate of saliva and an accompanying decrease in salivary protein components. Furthermore, many of these salivary protein constituents possess local antimicrobial properties and promote local wound healing and epidermal tissue generation. Hot water extracts and six N-containing sugars derived from *M. alba* leaves, have been investigated on pilocarpine-induced saliva secretion in streptozocin-induced diabetic mice. Hot water extracts (100 and 200 mg/kg, intraperitoneal) significantly potentiated the pilocarpine-induced salivary flow, but not the protein content. The N-containing sugars (37.5–300  $\mu$ mol/kg) potentiated the saliva secretion, and the order of potency was 1,4-dideoxy-1,4-imino-D-arabinitol > fagomine > 2-O- $\alpha$ -D-galactopyranosyl-1-deoxynojirimycin. Only fagomine significantly increased the protein content in the saliva.

An aqueous methanol extract of the root of *M. alba* showed a hypoglycaemic effect on intraperitoneal administration in normal mice. The active component was found to be a glycoprotein called moran A. A dose-dependent activity for the purified compound was furthermore established on intraperitoneal injection in both normal and alloxan-induced diabetic mice, 7 and 24 hours after administration. The hypoglycaemic activity is also found in an ethanol extract of the leaves of *M. alba*. The aqueous extract of the leaves of *M. alba* also exhibited uterine stimulant and estrogenic activity. Clinical study of aqueous extracts of the fruits showed laxative, central nervous system depressant and cholecystokinin receptor binding ac-

tivities. Hypoglycaemic activity in humans is not only for *M. alba* but also for orally administered *M. nigra* leaf extracts.

Morusin and kuwanon C, isolated from root bark of *M. australis* showed significant effects in platelet aggregation assays. An aqueous extract of *M. australis* showed significant antibacterial activity against *Streptococcus mutans* with a minimal inhibitory activity of less than 7.8 mg/ml. Small pieces of *M. alba* leaves inoculated with *Fusarium solani* f. sp. *mori* produced new antifungal substances. These substances can be extracted by methanol, ethanol or acetone but not with chloroform or water. An acetone extract of the non-infected leaves showed antibacterial activity against *Staphylococcus* species.

**Description** Dioecious or monoecious shrubs or trees up to 35 m tall; bark surface fissured, exuding white or yellowish-white latex. Leaves alternate, simple or 3-5-lobed, dentate, palmately 3-5-veined; stipules lateral, caducous. Inflorescence axillary, spicate. Flowers unisexual, small, with 4, free or basally united, imbricate tepals; male flowers in long catkin-like inflorescences, with 4 exerted stamens and top-shaped pistillode; female flowers in short to capitate inflorescences, tepals accrescent and succulent in fruit, ovary included, 1(-2)-locular with a single ovule, style 2-partite, staminodes absent. Fruit a juicy syncarp, composed of many achenes enclosed in the succulent tepals; endocarp woody. Seed subglobose, with endosperm. Seedling with epigeal germination; cotyledons emergent; hypocotyl elongated; first leaves often palmately or pinnately lobed.

**Growth and development** Growth of *M. alba* is initially rapid but slows down abruptly after about 10 years. Pollination is probably by wind. *Morus* fruits may be set without pollination and are eaten by birds, the seeds defecated and thus dispersed; dispersal by water is also known. Vesicular-arbuscular mycorrhizae are present in *M. alba*, *M. australis* and *M. nigra* but the degree of colonization varies.

**Other botanical information** It are the light-coloured buds of *M. alba* that account for its name rather than the colour of its fruit, which can be almost any colour including white, lavender, red and black. The leaves may be entire to variously lobed on the same plant.

The number of *Morus* species is often greatly overestimated because many cultivated forms that have arisen through hybridization have been described as species. *M. bombycis* Koidz. is sometimes thought to be conspecific with *M. australis*.

**Ecology** Most mulberry species occur in warm temperate to subtropical regions, and in tropical highland areas. In temperate regions white mulberry is the most frost-hardy *Morus*, though some clones are damaged at 6°C whereas others can stand temperatures of -40°C. In India, white mulberry has invaded irrigated plains very rapidly. Although black mulberry seems to be much less cold-tolerant, this may vary among clones, with absolute minimum temperatures between -18°C and -12°C. In general, *Morus* prefers well-drained, loamy soils, but white mulberry, for example, is quite tolerant of drought and poor soils.

**Propagation and planting** *Morus* can be propagated by seed and by cuttings, grafting and air-layering. Hardwood, softwood and root cuttings can all be used for vegetative propagation. Seeds are extremely small and the 1000-seed weight is 2.2-2.3 g. In India, *M. alba* is raised from seed which germinates in 9-14 days. Seedlings are pricked out when 10-15 cm tall. All but a few terminal leaves are stripped ('striplings') before seedlings are planted during the cold season or at the beginning of the rains. Cuttings with 3-4 buds are used for the production of leaves to feed silkworms. They are buried for 15-20 cm of their length, including 2 buds. The rooted cuttings are planted out in the field after two months. Spacing depends on the method of harvesting leaves and is variable, being (0.5-0.8(-1.2) m × (1.5-1.8-2.0(-2.5) m. When planted in paired rows, the distance between pairs is about 1.8 m, about 0.6 m within one pair, with 0.5 m in the row. Grafted *M. alba* gives higher leaf yields than plants raised from seedlings or cuttings, resulting in silkworm cocoons of superior quality. Experimental micropropagation was quite successful in India where shoot tip and nodal explants from a 12-year-old black mulberry tree were multiplied. Nodal explants from seedlings of white mulberry raised in vitro all rooted and were successfully transferred to a 1:1 sand-vermiculite mixture.

**In vitro production of active compounds** *M. alba* callus cultures from the leaves give rise to cell lines, which are reported to produce a variety of compounds e.g. prenylated chalcones and phytoosteroids. The yield of these Diels-Alder adducts is higher than in the intact plant.

**Husbandry** *Morus* trees are pruned to maintain an adequate framework for effective branching. *M. alba* is grown as a tree, a bush or as an espalier aiming at optimum leaf production. Low cutting ('bush type') is generally practised, but se-

mi-low and medium cutting (at 1 m height) are also known. When grown as a bush, white mulberry in India is productive for about 15 years after which period the crop is replaced. Black mulberry is pruned to a height of 1 m during the resting period; this promotes vigorous growth and the production of large-sized fruits.

**Diseases and pests** The major diseases of *Morus* as observed in India are: powdery mildew (*Phyllactinia corylea*, also found in Indonesia), leaf spot (*Cercospora moricola*, *Pseudocercospora mori*) and leaf rust (*Cerotelium fici*). The stem borer *Batocera rufomaculata* is an important pest. In Thailand, the nematode *Hoplolaimus seinhorsti* is often found in *M. alba* plantations and probably associated with a root-rot disease which is widespread in the north-eastern part of the country.

**Harvesting** Mature leaves of *M. alba* are harvested either by plucking and leaving a few terminal leaves, or by pruning, coppicing or pollarding. For harvesting there seems to be no standard procedure with regard to frequency and cutting height. In a pruning experiment with *M. nigra* in Indonesia neither total leaf production nor the chemical composition of the leaves was affected by the height of pruning (20, 70 and 120 cm) and the frequency (3-4 months). The roots are simply dug up.

**Yield** An annual yield of 25 t fresh leaves per ha is considered acceptable for *M. alba*.

**Handling after harvest** Harvested roots of *Morus* are washed and the rough part is scraped off. Then a longitudinal incision is made, the bark pounded lightly and separated from the wood, tied in bundles and dried in the sun. Harvested leaves for feeding silkworms are stored in loose heaps in cool rooms and heating, fermentation and drying-out are prevented.

**Genetic resources and breeding** There are over 1000 races of *M. alba*, mainly to produce fodder for silkworms. In India, the techniques to induce tetraploidy in *M. alba* have recently been standardized and diploids, autotriploids and tetraploids are being studied, to assess their relative advantages.

**Prospects** The hypoglycaemic properties of *Morus* deserve further attention for their possible application in diabetes. Furthermore, the anti-inflammatory properties and the wide range of traditional clinical applications warrant continuing interest in *Morus*.

**Literature** |1| Asano, N., Oseki, K., Tomioka, E. & Matsui, K., 1994. N-containing sugars from

*Morus alba* and their glycosidase inhibitory activities. *Carbohydrate Research* 259(2): 243-255. |2| Boer, E. & Sosef, M.S.M., 1998. *Morus L.* In: Sosef, M.S.M., Hong, L.T. & Prawirohatmodjo, S. (Editors): *Plant Resources of South-East Asia No 5(3). Timber trees: Lesser-known timbers*. Backhuys Publishers, Leiden, the Netherlands. pp. 387-389. |3| Chatterjee, G.K., Burman, T.K., Nagchaudhuri, A.K. & Pal, S.P., 1983. Antiinflammatory and antipyretic activities of *Morus indica*. *Planta Medica* 48(2): 116-119. |4| Chen, F., Nakashima, N., Kimura, I., Kimura, M., Asano, N. & Koya, S., 1995. Potentiating effects on pilocarpine-induced saliva secretion, by extracts and N-containing sugars derived from mulberry leaves, in streptozocin-diabetic mice. *Biological and Pharmaceutical Bulletin* 18(12): 1676-1680. |5| Council of Scientific and Industrial Research, 1962. *The wealth of India: a dictionary of Indian raw materials & industrial products*. Vol. 6: 429-439. |6| Ghafoor, A., 1985. *Moraceae*. In: Nasir, E. & Ali, S.I. (Editors): *Flora of Pakistan*. No. 171. Shamim Printing Press, Karachi, Pakistan. 54 pp. |7| Konno, C., Oshima, Y. & Hikino, H., 1977. Morusinol, isoprenoid flavone from *Morus* root barks. *Planta Medica* 42(2): 118-124. |8| Lim, S.H., Kim, Y.T., Lee, S.P., Rhee, I.J., Lim, J.S. & Lim, B.H., 1990. Sericulture training manual. *FAO Agricultural Service Bulletin* 80. Food and Agriculture Organization of the United Nations, Rome, Italy. 115 pp. |9| Samsijah & Sudrajat, 1975. *Hama dan penyakit tanaman murbei [Pests and diseases of mulberry plants]*. Laporan No 197. Lembaga Penelitian Hutan, Bogor, Indonesia. 67 pp. |10| Siddiqui, H.H., Malhotra, N.K. & Ramaswamy, A.S., 1975. Antiangiotensin activity of morin on the blood pressure and isolated tissues of the rat. *Journal of Research on Indian Medicine* 10(4): 120.

#### *Selection of species*

#### **Morus alba L.**

Sp. pl. 2: 986 (1753).

**Synonyms** *Morus indica* L. (1753), *Morus atropurpurea* Roxb. (1832), *Morus morettiana* Jacq. ex Burr. (1873).

**Vernacular names** White mulberry (En). Mûrier blanc (Fr). Moral blanco, morera blanca (Sp). Indonesia: murbei (general), besaran (Javanese). Philippines: amoras (Filipino), amingit (Igorot), mora (Ibanag). Burma (Myanmar): posa. Cambodia: mon. Vietnam: d[aa]ju t[aa]f[m], tang.

**Distribution** Native of China, now widely cultivated in temperate and tropical regions; in Malasia occasionally naturalized, i.e. in the Philippines (Batan Island and Cagayan Province) where it was introduced in 1780.

**Uses** Most of the medicinal applications of *M. alba* are reported from China. Elsewhere it seems to be more often cultivated for its edible fruits and leaves that are fed to silkworms. In Vietnam the root bark is used as a diuretic, antitussive and expectorant and prescribed in oedema, high blood pressure, cough, bronchitis and asthma.

**Observations** A small to medium-sized tree up to 15(-20) m tall, bole up to 70 cm in diameter, bark surface dark grey-brown, with horizontal lenticels; leaves ovate to broadly ovate, 5-16 cm × 4-12 cm, rounded to shallowly cordate at base, acute to acuminate at apex, pubescent on the main veins, with a slender, 1-3.5 cm long petiole; male spikes 1-1.5(-2) cm long, female spikes ovoid, 0.5-1.3 cm long; syncarp ovoid, 1.5-2.5 cm long. The native and subspontaneous habitats of

*M. alba* are generally moist places in mountains and thickets along rivers. In subtropical or dry tropical regions, *M. alba* can be cultivated at 0-3500 m altitude, but in the humid tropics it does not produce good fruit when planted at sea-level.

**Selected sources** 78, 175, 190, 202, 244, 335, 350, 396, 478, 540, 580, 584, 717, 900, 1035, 1048, 1128, 1178, 1212, 1252, 1276, 1316, 1317, 1336, 1345, 1392, 1405, 1470, 1478, 1613.

### *Morus australis* Poir.

Lamk, Encycl. 4: 380 (1797).

**Synonyms** *Morus acidosa* Griffith (1854), *Morus cavaleriei* H. Lév. (1911), *Morus inusitata* H. Lév. (1914).

**Vernacular names** Korean mulberry (En). Mûrier (Fr). Vietnam: d[aa]ju t[awf]m, d[aa]ju t[af]u, d[aa]u ta.

**Distribution** Originally from China, Korea, Japan and Taiwan; cultivated in Indo-China, the Philippines and Java, occasionally naturalized.

**Uses** In Java *M. australis* is used to feed silkworms; medicinal applications are mainly reported from its native region. In Vietnam a decoction of the leaves is used in the treatment of coughs and colds, and insomnia. A decoction of the root-bark is used against asthma, rheumatism and oliguria.

**Observations** A large shrub or small tree up to 10 m tall; leaves ovate to broadly ovate, 5-20 cm × 3-12 cm, rounded to shallowly cordate at base, long acuminate at apex, nearly glabrous to soft-hairy below, with a 1-4 cm long petiole; male spikes 1.5-3 cm long, female spikes erect or patent, broadly oblong, 1-2 cm long; syncarp oblong to ellipsoid, 1.5-3.5 cm long. In its natural habitats, *M. australis* is fairly common at low and moderate altitudes.

**Selected sources** 97, 364, 753, 856, 900, 1126, 1252, 1276.

### *Morus nigra* L.

Sp. pl. 2: 986 (1753).

**Synonyms** *Morus laciniata* Miller (1768), *Morus scabra* Moretti (1841).

**Vernacular names** Black mulberry, common mulberry, sycamine (En). Mûrier noire (Fr). Moral negro, morera negra (Sp). Vietnam: d[aa]ju g[ur]a den.

**Distribution** Originating from western Asia, but much cultivated in that region and the Mediterranean since ancient times. Nowadays cultivated and occasionally naturalized in most



*Morus alba* L. - 1, fruiting twig; 2, infructescence; 3, female flower; 4, male inflorescence; 5, male flower.

tropical and temperate regions, though in the tropics only at higher elevations. Only occasionally planted in Malesia.

**Uses** *M. nigra* is most commonly cultivated for its tasty fruits, which are also applied medicinally. The leaves are fed to silkworms, but generally considered inferior to those of the white mulberry.

**Observations** A small to fairly large tree up to 35 m tall; leaves broadly ovate, 5–16 cm × 5–16 cm, deeply cordate at base, shortly and bluntly acuminate at apex, rough above, pubescent below, with a striate, 2–3.5 cm long petiole; male spikes 1.5–2.5 cm long, female spikes ovoid, 1–2 cm long; syncarp ovoid, 1.5–2.5 cm long. *M. nigra* is cultivated in humid regions, up to 2000 m altitude.

**Selected sources** 131, 478, 580, 900, 1178, 1252, 1275, 1276, 1470, 1521.

D.S. Alonzo

## Oldenlandia L.

Sp. pl. 1: 119; Gen. pl. ed. 5: 55 (1754).

RUBIACEAE

$x = 9$ ; *O. affinis*:  $2n = 18$ , *O. brachypoda*:  $2n = 32$ , 54, *O. corymbosa*:  $2n = 18$ , 36, 54, *O. herbacea*:  $2n = 18$ , 36

**Major species** *Oldenlandia brachypoda* DC., *O. capitellata* (Wallich ex G. Don) O. Kuntze, *O. corymbosa* L.

**Origin and geographic distribution** *Oldenlandia* consists of approximately 100 species (but estimates of up to 300 species have also been made), and is distributed in all tropical and subtropical regions.

**Uses** In many areas whole *Oldenlandia* plants are used internally in a decoction to treat fever, stomach disorders and diarrhoea. Externally they are commonly used pounded in poultices to treat ulcers, wounds, bruises, snake bites, insect stings, broken bones, rheumatism, lumbago and ague. In traditional medicine in India, *O. corymbosa* is used in the treatment of jaundice and gonorrhoea. The aerial parts of *O. brachypoda* and *O. corymbosa* are used in Chinese medicine to treat tumours in liver, lungs and rectum, and as an antiphlogistic.

In India, the bark of the roots of *O. umbellata* L. was much used to prepare a red dye before the large-scale production of synthetic dyes started at the end of the 19th Century. A decoction of its leaves and bark is considered expectorant and is

prescribed in cases of bronchial catarrh, bronchitis and asthma, and is also used as a wash for poisonous bites.

**Production and international trade** *Oldenlandia* is usually collected in the wild, and because it often acts as a weed, it is usually available in sufficient amounts for private use. There is no real international trade, although small amounts of dried plants are exported from China.

**Properties** Tests with *O. brachypoda* showed inhibition on aflatoxin B1-induced mutagenesis using *Salmonella typhimurium* TA 100 as the bacterial tester strain and rat liver supernatant as the activation system. This suggests that it possesses mutagenic activity and possibly cancer chemopreventive properties. In tests with mice in Indonesia, *O. corymbosa* leaves caused humoral immune response stimulation and suppression on the phagocytosis system. The water-soluble fraction administered intraperitoneally had no influence on cellular immune responses, but the residue fraction given orally caused stimulation. *O. brachypoda* markedly stimulated murine spleen cells to proliferate in in vitro studies. This suggests that it has immunomodulatory activity. Nine iridoid glucosides have been isolated from the aerial parts of *O. corymbosa*, among which asperuloside, asperulosidic acid and scandoside methyl ester. Some iridoids are known to have antiphlogistic activity.

**Description** Annual herbs or sometimes herbaceous perennials; stem erect or procumbent. Leaves opposite, simple and entire, sessile or petiolate; stipules interpetiolar and adnate to leaf-bases. Inflorescence terminal or axillary, cymose and paniculate or corymbose, sometimes flowers in fascicles or solitary. Flowers bisexual, protandrous, small, 4-merous, homostylous or heterostylous; calyx with turbinate, globose or oblong tube, and distinct lobes; corolla hypocrateriform or narrowly infundibular, with valvate lobes, white, pale mauve, blue or pink; stamens with filaments attached at the corolla tube between the lobes, anthers dorsifixed; ovary inferior, 2-locular, with 2-many ovules, style terete or filiform, stigma usually bifid but sometimes capitate. Fruit a crustaceous capsule, loculicidally dehiscent at apex, usually many-seeded. Seeds obconical, menisoid, scutelliform, cerebriform or obovoid, with variously patterned surface. Seedling with epigeal germination; cotyledons very small, ovate.

**Growth and development** Most *Oldenlandia* species are annual and complete their lifespan from seed to seed in a short period, which is in ac-

cordance with their status as weed. Some species are perennial, e.g. *O. capitellata*.

**Other botanical information** *Oldenlandia* is often included in *Hedyotis*. However, recent studies in northern America, Australia and India have shown that there are differences in habit and morphology of inflorescences, flowers, fruits and seeds, as well as in cytology, embryology and palynology, so *Oldenlandia* has been kept separate here.

Several *Oldenlandia* species are difficult to distinguish and often confused. The correct name for the taxon usually referred to as *O. diffusa* is *O. brachypoda* DC., although some caution is needed as the 'true' *O. diffusa* (Willd.) Roxb. is also a widespread weed.

**Ecology** *Oldenlandia* often occurs in disturbed locations such as grasslands and roadsides. Several species are common weeds in fields; *O. corymbosa* is particularly common in e.g. cassava, pineapple and maize throughout the tropics.

**Propagation and planting** In *O. corymbosa* three plant types have been determined: a type in which almost all seeds are non-dormant and germinate immediately in light at 35–40°C, a type in which almost all seeds are dormant and need a pre-germination treatment at 12°C for 7 days, and a type in which approximately 50% of the seeds are dormant and the remainder are non-dormant.

Tests with the Indian species *O. umbellata* showed promising results for in vitro propagation. Direct somatic embryogenesis and subsequent plant formation were achieved by using leaves, pieces of stems, axillary buds and developing fruits. The best results were obtained in half-strength Murashige and Skoog basal medium containing only 12% of the usual N content, supplemented with 0.2 mg/l benzyladenine, 0.1% ascorbic acid and 1.5% sucrose under diffuse light (300 lux). Rooting was achieved in the same medium, but with naphthalene-acetic acid instead of benzyladenine.

**Harvesting** For personal use, whole plants of *Oldenlandia* are usually harvested in small amounts, when needed.

**Handling after harvest** Freshly harvested *Oldenlandia* plants are often used immediately, but sometimes they are dried and stored for later use or trade.

**Genetic resources and breeding** In South-East Asia the medicinally used *Oldenlandia* species are common weeds and widely distributed, so there is no danger of genetic erosion.

**Prospects** Several tests on the mutagenic activity and possible cancer chemopreventive properties of *Oldenlandia* have given positive results; animal tests on immune response stimulation and suppression on the phagocytosis system have also been promising. The antiphlogistic properties attributed to *Oldenlandia* are interesting. More research is needed to establish the definite medicinal value of these plants.

Taxonomic research on *Oldenlandia* and related genera on a worldwide basis is desirable. Species and genus concepts and delimitations differ greatly, resulting in much confusion in floras of the various regions.

**Literature** [1] Corbineau, F. & Côme, D., 1981. Some particularities of the germination of *Oldenlandia corymbosa* L. seeds (tropical Rubiaceae). *Israel Journal of Botany* 29: 157–167. [2] Halford, D.A., 1992. Review of the genus *Oldenlandia* L. (Rubiaceae) and related genera in Australia. *Austrobaileya* 3(4): 683–722. [3] Huang, J.-T., 1981. Neue Iridoide aus *Oldenlandia diffusa* Roxb. [New iridoids from *Oldenlandia diffusa* Roxb.]. *Archiv der Pharmazie* 314(10): 831–836. [4] Otsuka, H., Yoshimura, K., Yamasaki, K. & Cantoria, M.C., 1991. Isolation of 10-O-acyl iridoid glucosides from a Philippine medicinal plant, *Oldenlandia corymbosa* L. (Rubiaceae). *Chemical and Pharmaceutical Bulletin* 39(8): 2049–2052. [5] Quisumbing, E., 1978. Medicinal plants of the Philippines. Katha Publishing Co., Quezon City, the Philippines. pp. 920–922. [6] Rao, G.P. & Bahadur, B., 1990. Somatic embryogenesis and plant regeneration in self incompatible *Oldenlandia umbellata* L. (Rubiaceae). *Phytomorphology* 40(1–2): 95–101. [7] Sivarajan, V.V. & Biju, S.D., 1990. Taxonomic and nomenclatural notes on the *Hedyotis corymbosa-diffusa* complex (Rubiaceae) in India. *Taxon* 39(4): 665–674. [8] Sutarjadi, Santosa, M.H., Bendryman & Dyatmiko, W., 1991. Immunomodulatory activity of *Piper betle*, *Zingiber aromatica*, *Andrographis paniculata*, *Allium sativum*, and *Oldenlandia corymbosa* grown in Indonesia. *Planta Medica* 57, Supplement Issue 2: A136. [9] Wong, B.Y.Y., Lau, B.H.S., Tadi, P.P. & Teel, R.W., 1992. Chinese medicinal herbs modulate mutagenesis, DNA binding and metabolism of aflatoxin B1. *Mutation Research* 279(3): 209–216. [10] Wong, B.Y.Y., Lau, B.H.S., Yamasaki, T. & Teel, R.W., 1993. Modulation of cytochrome P-450IA1-mediated mutagenicity DNA binding and metabolism of benzo-a-pyrene by Chinese medicinal herbs. *Cancer Letters* 68(1): 75–82.

*Selection of species***Oldenlandia affinis (Roemer & Schultes) DC.**

Prodr. 4: 428 (1830).

**Synonyms** *Hedyotis affinis* Roemer & Schultes (1818), *Hedyotis dichotoma* Heyne ex Roth (1821), *Oldenlandia dichotoma* (Heyne ex Roth) Hook.f. (1880).

**Vernacular names** Thailand: ya thopthaep (Tak).

**Distribution** Tropical Africa, Madagascar, the Comoro Islands, India, Thailand, Peninsular Malaysia and Singapore.

**Uses** *O. affinis* was reported to be used medicinally in Peninsular Malaysia, without specification.

**Observations** A slender much-branched annual or perennial herb up to 50(-120) cm long; leaves narrowly oblong-lanceolate to linear, up to 3(-6) cm long; flowers in a lax paniculate cyme, dichotomously branched, corolla with spreading lobes, bluish or pinkish-white; fruit subglobular. *O. affinis* occurs in Malaysia in dry, often sandy locations, usually near the sea; in India it is commonly found on slopes of hills up to 1200 m altitude.

**Selected sources** 202, 921.

**Oldenlandia biflora L.**

Sp. pl. 1: 119 (1753).

**Synonyms** *Hedyotis biflora* (L.) Lamk (1792).

**Vernacular names** Philippines: pisek (Ivatan), dalumpang (Subanun), palarapdap (Samar-Leyte Bisaya). Thailand: phak khuang, mak dip nam khang, sadao din (Bangkok). Vietnam: an d[ieef]n hai hoa, mai h[oo]f]ng.

**Distribution** From Sri Lanka, India and Indo-China to Samoa and eastern Australia; throughout Malesia.

**Uses** In the Philippines, the plants are pounded and applied to wounds, and a decoction is administered internally to cure diarrhoea. Besides these uses, traditional doctors in Thailand also use the whole plant for its anti-amoebic and antipyretic properties. In India, it is used to treat fever and stomach complaints. In Vietnam, it is applied to treat snake bites.

**Observations** An erect or prostrate to decumbent, divaricately branched annual herb up to 50 cm long, branches obtusely or acutely 4-angled; leaves oblong, elliptical to narrowly elliptical, up to 8 cm long; flowers in 3-7-flowered cymes, corolla with geniculate lobes, white or occasionally pale

blue; fruit turbinate-obovoid; seeds transversely ellipsoid. *O. biflora* occurs in sunny or slightly shaded locations, often as a weed, in fields, gardens, roadsides, river banks and open spots in forest up to 300 m altitude.

**Selected sources** 97, 537, 921, 1178.

**Oldenlandia brachypoda DC.**

Prodr. 4: 424 (1830).

**Synonyms** *Hedyotis brachypoda* (DC.) Sivar. & Biju (1990), *Hedyotis diffusa* auct. non Willd., *Oldenlandia diffusa* auct. non (Willd.) Roxb.

**Vernacular names** Malaysia: bunga chakar ayam, rumput angga (Peninsular). Philippines: ulasiman-kalat (Tagalog), daniri (Bisaya). Thailand: hom chaeo naa (Khon Kaen).

**Distribution** From Sri Lanka, India, southern China and Japan to Malesia; also in tropical America.

**Uses** In Malaysia, the plant is used for poulticing, apparently against lumbago. In India, a decoction of the whole plant is used to treat biliousness, fever and gonorrhoea.

**Observations** A prostrate annual herb up to 45 cm long, usually branched from the base, branches terete; leaves linear-lanceolate, up to 4 cm long; flowers axillary, solitary, sometimes 2-3 together, corolla white to purplish; fruit subglobose; seeds reticulate. *O. brachypoda* usually occurs in humid or swampy locations such as rice fields and along streams, up to 1100(-1500) m altitude. It has been very often confused with *O. diffusa* (Willd.) Roxb., which is also a common weed, and which possibly has comparable uses and properties.

**Selected sources** 97, 202, 627, 921, 1178, 1373, 1587, 1588, 1637.

**Oldenlandia capitellata (Wallich ex G. Don) O. Kuntze**

Revis. gen. pl.: 292 (1891).

**Synonyms** *Hedyotis capitellata* Wallich ex G. Don (1834), *Oldenlandia recurva* (Korth.) Miq. (1859).

**Vernacular names** Indonesia: akar kemenyan hantu (Sumatra). Malaysia: seketan, akar patah bubul, akar patah gogoh (Peninsular). Thailand: kamlang hualamaan (Chiang Mai), tong haeng (peninsular), yan bueang thuai (Narathiwat). Vietnam: c[aa]y lo[es]t m[oo]f]m, c[aa]y d[aj] c[aa]r]m.

**Distribution** Eastern India, Indo-China, southern China, Burma (Myanmar), Thailand, Peninsular Malaysia, Singapore, Sumatra, Borneo; once collected in Java.



**Uses** Leaves are used in Malaysia for poulticing to treat snake bites, broken bones, bruises, rheumatism, lumbago and ague, and internally for treating kidney complaints. A decoction of the root is administered to women after childbirth, and also used against constipation, indigestion, gastric vertigo and dysentery. In Vietnam, leaves and young stems are used in decoction to treat ulcers in the mouth, glossitis, pharyngitis and peptic ulcers.

**Observations** A sprawling perennial herb up to 2 m long, branches 4-angled or terete; leaves oblong to lanceolate, up to 11(-15) cm long; flowers in a paniculate inflorescence composed of peduncled heads, corolla cream-coloured; fruit obovoid. *O. capitellata* is a common climber over bushes and hedges up to 1200 m altitude.

**Selected sources** 97, 202, 580, 1035.

### **Oldenlandia corymbosa L.**

Sp. pl. 1: 119 (1753).

**Synonyms** *Hedyotis corymbosa* (L.) Lamk (1792).

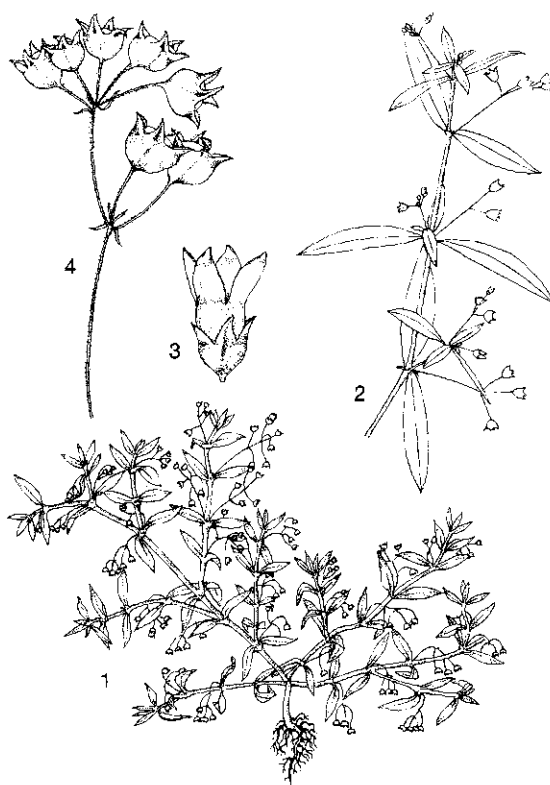
**Vernacular names** Malaysia: siku-siku, siku dengan, pokok telur belangkas (Peninsular). Philippines: malaulasiman, ulasiman-aso (Tagalog). Thailand: yaa linnguu (Bangkok). Vietnam: l[uw][owx]i r[aws]n, c[os]c m[awr]n.

**Distribution** Probably native to Africa and India, but now with pantropical distribution; throughout Malesia.

**Uses** The leaves are commonly used for poulticing to treat sores and sore eyes. The entire plant is used in decoction as a febrifuge and stomachic. In Indo-China, it is also used as antirheumatic. In India, the plant is a common ingredient in mixtures used internally to treat fever and as a tonic. It is also used to treat jaundice. The roots are reported to have vermifuge properties.

**Observations** A prostrate to decumbent, divaricately branched annual herb up to 60 cm long, branches 4-angled; leaves narrowly elliptical to linear-lanceolate, up to 3(-5) cm long; flowers in a (1-)3-8-flowered umbel-like corymb, corolla with spreading lobes, white or pinkish; fruit depressed obovoid or broadly obovoid; seeds obconical to depressed obconical, laterally compressed. *O. corymbosa* is a weed in fields, roadsides, lawns and gardens, preferably in not to wet, sunny, stony locations, usually up to 800 m altitude, but sometimes up to 1500 m.

**Selected sources** 97, 202, 279, 537, 921, 1035, 1094, 1178, 1373, 1423.



*Oldenlandia corymbosa* L. - 1, plant habit; 2, flowering and fruiting stem; 3, flower; 4, infructescence.

### **Oldenlandia herbacea (L.) Roxb.**

Fl. ind. 1: 445 (1820).

**Synonyms** *Hedyotis herbacea* L. (1753).

**Vernacular names** Vietnam: an di[ee]n c[or].

**Distribution** Tropical and subtropical Africa and Asia, including the whole of Malesia.

**Uses** *O. herbacea* has been reported to be used medicinally in Peninsular Malaysia, without specification.

**Observations** An erect or ascending slender annual or perennial herb up to 50(-60) cm long, usually much-branched, branches 4-winged; leaves linear-lanceolate to linear, up to 3(-5.5) cm long; flowers axillary, solitary, sometimes 2(-4) together, corolla white to purplish; fruit subglobular, small, about 2.5 mm long; seeds ovoid to ellipsoid, reticulate. *O. herbacea* occurs in open places such as fields, grasslands and roadsides, often on stony or sandy soils, up to 1500 m altitude.

**Selected sources** 97, 202, 921.

N.O. Aguilar & R.H.M.J. Lemmens

**Orthosiphon aristatus (Blume) Miq.**

Fl. Ind. Bat. 2: 943 (1858).

LABIATAE

2n = 48

**Synonyms** *Orthosiphon stamineus* Benth. (1831), *Orthosiphon grandiflorum* auct. non Terac., *Orthosiphon spicatus* auct. non Benth.

**Vernacular names** Java tea (En). Thé de Java (Fr). Indonesia: kumis kucing (general), kumis ucing (Sundanese), remuk jung (Javanese). Malaysia: kumis kucing. Philippines: balbas-pusa (Tagalog), kabling-gubat. Cambodia: kapen prey. Laos: hnwàd mēew. Thailand: yaa nuat maeo. Vietnam: r[aa]u m[ef]o.

**Origin and geographic distribution** Java tea is distributed from India, Indo-China and Thailand, through Malesia to tropical Australia. As a wild plant, it occurs throughout Malesia, but is apparently rare in Borneo, Sulawesi and the Moluccas. It is now grown in South-East Asia (in Java since 1928), Africa, Georgia (Caucasus) and Cuba.

**Uses** In Malesia, Thailand and Vietnam the leaves are used as a diuretic in teas and infusions against various kidney complaints and illnesses, renal calculi, phosphaturic catarrh of the bladder, gout, and also, in combination with other drugs, to stimulate the kidneys and as a medicine for nephritis, gallstones and diabetes. For these purposes, Java tea is sometimes mixed with the leaves of *Sonchus* or *Barleria*.

In Europe, the therapeutic indications for *Orthosiphon* are: diuresis and irrigation of the urinary tract, especially in cases of inflammation and renal gravel, and as an adjuvant in treatment of bacterial infections of the urinary tract. The drug is normally taken as an infusion of 2–3 g dried material in 150 ml water, 2–3 times per day. *Orthosiphon* is also included in the form of an extract in instant preparations, which are used accordingly. In Indonesia, it is used to treat jaundice in a mixture with leaves of *Blumea balsamifera* (L.) DC. and *Phyllanthus fraternus* Webster and rhizomes of *Curcuma xanthorrhiza* Roxb., and to treat diabetes together with the leaves of *Andrographis paniculata* (Burm.f.) Nees. In mixtures with leaves of other plants it is also used against gout, rheumatism and arteriosclerosis. *Orthosiphon* preparations are on the market in Indonesia as capsules, pure or in combination with other ingredients. The crude herb is said to cause vomiting. In gardens the plant is also cultivated as an ornamental.

**Production and international trade** Indonesia is the main producing country (Java, Sumatra, North Sulawesi). Before the Second World War about 80 t/year of dried leaves was exported to the Netherlands, Germany, France, Japan and the United States. After the war interest waned because more modern diuretics became available. However, Indonesian exports to Europe and other parts of the world are again substantial. In the period 1991–1995 an average of 170 t/year of dried leaves was exported. The average value in 1995 was US\$ 1.3/kg. The main importing country is Germany.

**Properties** 'Orthosiphonis Folium' or Java tea consists of the dried leaves and stem tips of *O. aristatus* collected shortly before flowering. It contains up to 12% minerals with a high proportion of potassium (600–700 mg per 100 g fresh leaf), approximately 0.2% lipophilic flavones including sinensetin, flavonol glycosides, caffeic acid derivatives (mainly rosmarinic acid and 2,3-dicaffeoyltartaric acid), inositol, phytosterols ( $\beta$ -sitosterol), saponins and up to 0.7% of essential oil. Analysis revealed caffeic acid derivatives like rosmarinic acid (and 2,3-dicaffeoyltartrate) to be predominant components in a hot water extract prepared comparable to that of a herbal tea.

Various tests have been performed to demonstrate the diuretic activity of *Orthosiphon* extracts both in animals and man, and to establish the source of this effect. Diuretic effects were observed in rabbits, dogs and rats; oral application of 750 mg/kg body weight lyophilized aqueous *Orthosiphon* extract enhanced ion excretion ( $K^+$ ,  $Na^+$ ,  $Cl^-$ ) in rats, whereas no increase in urine output was observed. Similar effects were reported in man, such as increased diuresis and elimination of chlorides and urea. In a placebo-controlled double-blind crossover study with 40 volunteers, however, no influence on urine output or  $Na^+/K^+$  excretion was recorded with a daily dose of 600 ml of an infusion equivalent to 10 g of dried leaves.

Although the possible diuretic compounds of *Orthosiphon* extracts are not yet known, it has been postulated that the effects could be partially due to the high content of potassium in the leaves and the presence of inositol (and possibly saponins), as well as to the isolated flavones sinensetin and 3'-hydroxy-5,6,7,4'-tetramethoxyflavone which exhibited a diuretic activity in rats after intravenous administration of 10 mg/kg body weight. The total activity of *Orthosiphon* leaves should not be attributed to these flavones, however, since it has also been shown that only minute amounts of

these lipophilic compounds are extracted by (hot) water when an aqueous infusion is prepared. The highest content of sinensetin (up to almost 0.4%) was found in old leaves of forms with bluish-violet flowers, and the lowest content (about 0.1%) in young leaves of forms with white flowers.

In tests with healthy volunteers in Thailand, *Orthosiphon* extracts increased excretion of citrate and oxalate. Although a higher level of oxalate may increase the risk of kidney stones, the increased citrate output helps prevent stone formation.

It has been demonstrated that Java tea has antimicrobial properties. In *in vitro* tests, aqueous extracts markedly inhibited the growth of both gram-positive and gram-negative bacteria. Saponins may play a role in bacteriostatic activity *in vitro*. Caffeic acid derivatives (which represent as much as 95% of the phenolic substances present in a hot water extract) may also be responsible for the antibiotic activity.

The lipophilic flavonoids present in *O. aristatus*, of which sinensetin and tetramethylscutellarein are the most abundant, have shown inhibitory effect against Ehrlich ascites tumour cells *in vitro*. Additionally, the lipophilic flavonoids may be partially responsible for anti-inflammatory effects, since flavonoids are inhibitors of cyclo-oxygenase and lipoxygenase.

Doses of less than 1 g/kg body weight have been found to be lethal for rats and mice after intraperitoneal injection, but no injurious effects were found after feeding up to 5 g/kg body weight orally.

In patients, increased choleresis and cholekinesis have been reported, together with an antibacterial action in cholecystitis and cholangitis, after oral administration of a Java tea extract. However, *in vivo* studies in rats with the isolated flavones sinensetin and 3'-hydroxy-5,6,7,4'-tetramethoxyflavone administered intravenously at a dose of 10 mg/kg body weight did not confirm these findings.

**Adulterations and substitutes** It is reported that Java tea is adulterated with leaves of *Ageratina riparia* (Regel) R.M. King & H. Robinson (*Compositae*). Among the many South-East Asian plants with diuretic activity are *Clerodendrum*, *Desmodium*, *Ludwigia* and *Sida* species. Species which, like Java tea, combine diuretic activity and antimicrobial activity, belong to, among others, the genera *Artemisia*, *Cassia*, *Elephantopus*, *Phyllanthus* and *Plantago*. *Acorus calamus* L. and *Mentha arvensis* L. are examples of species important for their antibacterial activity.

**Description** A perennial herb, 25–200 cm tall, with quadrangular, poorly ramified, ascending stem. Leaves decussately opposite, ovate or rhombic, 2–9(–12) cm × 1.5–5 cm, cuneate at base, acute or acuminate at apex, serrate, glabrous or minutely pubescent, glandular-punctate; petiole 0.5–2 (–4.5) cm long; stipules absent. Inflorescence an opposed cyme arranged in terminal racemes, 7–29 cm long. Flowers pedicellate; calyx 2.5–4.5 mm long (up to 12 mm in fruit), bilabiate, gland-dotted; corolla 10–20 mm long, tubular, bilabiate, white or (pale) lilac; stamens 4, long-protruding from the corolla tube; ovary superior, style long-protruding, slender, with enlarged, club-shaped and shallowly cleft stigma. Fruit splitting into 4 oblong-ovoid nutlets, 1.5–2 mm long, brownish, rugose.

**Growth and development** The flowers are sometimes cleistogamous, in which case the corolla is hidden in the calyx. The ovary is normal and develops into normal nutlets.

**Other botanical information** *Orthosiphon* comprises approximately 40 species with an Old



*Orthosiphon aristatus* (Blume) Miq. – 1, flowering stem; 2, flower; 3, fruiting calyx and nutlet.

World distribution. In Malesia, only 2 species occur. The wild relative *Orthosiphon thymiflorus* (Roth) v.d. Sleesen is rare in Malesia (central and eastern Java), more common from India and Sri Lanka to Indo-China. Three cultivars of *O. aristatus* are distinguished: one with bluish-violet and two with white flowers. The white-flowered cultivar with reddish stems, petioles and leaf veins appears to possess the best diuretic qualities.

**Ecology** Java tea occurs in the wild in thickets, regrowths, grasslands and along forest borders and roadsides, often in shaded not too dry localities, but also in sunny places, up to 1000 m altitude.

**Propagation and planting** Propagation is by stem cuttings, 15–20 cm long, which have some buds. Cuttings are usually planted in shade, with 40–60 cm between plants and rows. Often 4–6 cuttings are placed in one hole. Direct planting in the field or in the backyard, as is most common, can be done all the year round, but the usual time of planting is at the beginning of the rainy season. For plantations, planting in a nursery for a period of 45 days with the cuttings placed vertically with only one bud visible is preferred.

**In vitro production of active compounds** Cell suspension cultures of *O. aristatus* have been shown to accumulate rosmarinic acid. The accumulation of rosmarinic acid in the cell suspension cultures could be increased by adding yeast extract to the culture medium.

**Husbandry** Regular weeding is necessary. Inflorescences should be removed. Manuring is advantageous; the standard application per ha is of 200 kg triple superphosphate, 100 kg potassium salt and 15 t manure. It is advised to add a nitrogenous fertilizer at the rate of 100 kg/ha after each harvest.

**Diseases and pests** Fungal diseases that have been reported to cause losses in Java tea are *Botrytis cinerea*, *Corticium rolfsii*, *Moniliopsis aderholdii* and *Pythium debaryanum*. Nematodes may cause galls to develop on the roots, but no standard treatment is practised to limit their damage. Insecticides are usually applied to the planting holes in order to prevent termite attack.

**Harvesting** Harvest usually starts 8–10 weeks after planting, at the beginning of flowering. Every 2–3 weeks the upper 4–10 leaves of shoots are plucked by hand.

**Yield** Annual yields of dry leaves amount to 1500 kg/ha.

**Handling after harvest** Smallholders usually sun-dry leaves. In estate farming artificial drying

is practised. To obtain a high-quality product, the leaves are first withered in the air, and then dried at 45–50°C. Dried leaves of good quality are green (a blackish colour is due to overheating or contact with metal containers), have a good aroma, a moisture content below 14%, a bitter taste, an ash content of about 10%, a contamination content of less than 2%, and do not contain insects or fungi. Properly dried leaves should be pressed as soon as possible to prevent moisture uptake. They are packed in ordinary tea chests, each containing up to 50 kg of leaves; aluminium foil is used to prevent moisture uptake.

**Genetic resources and breeding** As no germplasm collections exist, plant material should be collected from all growing regions.

**Prospects** Java tea shows promising medicinal properties. More research is needed to confirm the activities reported and to determine and isolate the active constituents. For a good quality product a survey on market requirements and potential for expansion is needed.

**Literature** |1| Acosta, L., Lerch, G. & Sklizkov, V., 1985. Algunos aspectos fitotécnicos en la introducción a cultivo de *Orthosiphon stamineus* en Cuba [Some phytotechnical aspects on the introduction of the cultivation of *Orthosiphon stamineus* in Cuba]. Boletín de reseñas: Plantas medicinales 14. 22 pp. |2| Chen, C.P., Lin, C.C. & Namba, T., 1989. Screening of Taiwanese crude drugs for antibacterial activity against *Streptococcus mutans*. Journal of Ethnopharmacology 27(3): 285–296. |3| Dat, D.D., et al., 1992. Studies on the individual and combined diuretic effects of four Vietnamese traditional herbal remedies: *Zea mays*, *Imperata cylindrica*, *Plantago major* and *Orthosiphon stamineus*. Journal of Ethnopharmacology 36(3): 225–231. |4| Englert, J. & Harnischfeger, G., 1992. Diuretic action of aqueous *Orthosiphon* extract in rats. Planta Medica 58(3): 237–238. |5| Keng, H., 1978. Labiatae. In: van Steenis, C.G.G.J. (General editor): Flora Malesiana. Series 1, Vol. 8. Sijthoff & Noordhoff International Publishers, Alphen aan den Rijn, the Netherlands. pp. 379–382. |6| Malterud, K.E., Hanche-Olsen, I.M. & Smith-Kielland, I., 1989. Flavonoids from *Orthosiphon spicatus*. Planta Medica 55(6): 569–570. |7| Nirdnoy, M., 1987. The effects of roselle and *Orthosiphon* on urinary uric acid and citrate excretion. Proceedings of the symposium on research and development of medicinal drugs for tropical diseases, 26–27 February 1987, Mahidol University, Bangkok, Thailand. p. 57 (in Thai). |8| Schut, G.A. & Zwaving, J.H., 1993.

Pharmacological investigation of some lipophilic flavonoids from *Orthosiphon aristatus*. *Fitoterapia* 64(2): 99–102. |9| Sumaryono, W., Proksch, P., Hartmann, T., Nimtz, M. & Wray, V., 1991. Induction of rosmarinic acid accumulation in cell suspension cultures of *Orthosiphon aristatus* after treatment with yeast extract. *Phytochemistry* 30(10): 3267–3271. |10| Wongkrachang, Y., Wongrattanasathit, T., Athisuk, K., Chai-at, P. & Phungwicha, P., 1990. Toxicity study of *Orthosiphon grandiflorus*. *Warasan Phesat Witthaya* 12(2): 63–69 (in Thai).

**Other selected sources** 243, 332, 350, 399, 672, 1035, 1040, 1126, 1178, 1502, 1566, 1572.

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### ***Oxalis corniculata* L.**

Sp. pl. 1: 435 (1753).

OXALIDACEAE

$2n = 24, 28, 44, 48$

**Synonyms** *Oxalis repens* Thunb. (1781), *Oxalis javanica* Blume (1825).

**Vernacular names** Indian sorrel, creeping wood-sorrel, procumbent yellow sorrel (En). Indonesia: daun asem kecil (Sumatra), calingcing (Sundanese), semangnen (Javanese). Malaysia: sikap dada (Peninsular). Papua New Guinea: kokavu (Kami, Eastern Highlands), akler (New Britain). Philippines: taingan-daga (Tagalog), marasiksik (Iloko), daraisig (Bikol). Singapore: sikap dada. Burma (Myanmar): hmô-gyin. Cambodia: chantoe phnom kok. Laos: som ten kalm. Thailand: phak waen (central), som sangka (Chiang Mai), som din (Mae Hong Son). Vietnam: me d[aas]t, chua me ba ch[if]a, toan t[uw]ow[ng] th[ar]o.

**Origin and geographic distribution** *Oxalis* consists of about 700 species and has its centres of diversity in South America and South Africa. Only a few species occur in South-East Asia; in Malesia there are 3 native species and 5 introduced ones, 4 of which have naturalized, sometimes becoming weeds. *O. corniculata* is a cosmopolitan weed of unknown origin, although a southern European origin has been postulated. It occurs throughout South-East Asia, but is scarce in Peninsular Malaysia, Borneo and Sulawesi.

**Uses** *O. corniculata* is considered a good medicine against scurvy, e.g. in the Philippines, India and China. The juice of the leaves is used for cleansing wounds and to treat itch, burns, sores,

insect and scorpion stings, and for removing warts. A poultice of the leaves relieves pain due to swellings. The leaves are taken as a good remedy in cases of intestinal complaints, and are applied externally to treat miliaria. A decoction of the leaves is sometimes used as a gargle against sore mouth and gums, and to treat cough, fever and dysentery. The whole plant is used as a diuretic and to treat opacity of the cornea. In Thailand, the aerial parts are used as an emmenagogue and to treat cough. In Korea, the entire plant is used in the treatment of dysentery, haemorrhoids and as an antidote. In China, it is considered to be cooling, emmenagogic, anthelmintic and lithotriptic. In Papua New Guinea and East Africa, the pounded plant is applied to remove ringworm.

*O. corniculata* is sometimes used to remove rust and ink stains, and to scour copper, due to the presence of oxalates (mainly potassium) and their ability to form metal complexes. The use as an antidote to mercury or arsenic poisoning is also sometimes reported. Small amounts of leaves have been added to food as a seasoning, but larger amounts can be toxic. Although it is usually a serious weed, it may sometimes serve as a valuable cover crop in preventing soil erosion in perennial crops such as tea and coconut.

**Production and international trade** *O. corniculata* is only used locally and has no importance on the market.

**Properties** *Oxalis* derives its acid taste from the presence of oxalates (mainly in the form of soluble potassium salt). Oxalates are toxic in high doses. They combine with serum calcium to form insoluble calcium oxalate. The reduction in available calcium leads to violent muscular stimulation with convulsions and collapse. In cases of poisoning, calcium oxalate crystals are also found in the renal tubulus; acute renal failure may occur from blockage of the renal tubuli. The principal manifestation of oxalate poisoning is anuria. In laboratory animals, the symptoms of acute poisoning are enlarged, pale kidneys, loss of control of fore or hind quarters and tetany.

The leaves contain  $\beta$ -carotene, and the C-glycosyl flavonoids vitexin, isovitexin and vitexin-2'-O- $\beta$ -D-glucoside. On extraction with chloroform-methanol young leaves yielded 1.5% lipids (on dry weight basis). These leaf lipids are a good source of essential fatty acids and also of  $\beta$ -tocopherols.

Aqueous leaf and root extracts have brought about significant reduction in seedling growth of e.g. wheat; the allelopathic activity is due to oxalic acid and fumaric acid. Tests in India showed that

powdery mildew (caused by *Phyllactinia corylea*) and leaf rust (caused by *Pseudocercospora mori*) of mulberry (*Morus* spp.) were minimized by a 10% alcoholic water extract of fresh *O. corniculata* plants. The extract also has insecticidal property.

**Description** A perennial, creeping or ascending small herb, with stems up to 50 cm long, rooting at the nodes; main root fibrous, in old plants sometimes developing into a thickened and woody rootstock; stems several from the main root, branching, puberulous. Leaves alternate, digitately 3-foliolate, petiole 1–5.5(–10) cm long, stipules small, up to 3 mm long; leaflets broadly obcordate to elliptical-obcordate, 4–20(–25) mm × 5–18(–25) mm, pubescent to sericeous beneath. Inflorescence cymose to pseudo-umbellate, pedunculate, 1–6 (–8)-flowered; bracts 2–several, subopposite to whorled. Flowers bisexual, actinomorphic, 5-merous, usually homostylous with mid-styled form, rarely with long-styled form; pedicel up to 20 mm long, articulate at base and beneath the calyx;

sepals imbricate, short connate at base, lanceolate, 2–6 mm × 0.5–2 mm, sparsely puberulous to sericeous; petals contorted, coherent above the claw, spatulate-oblong to spatulate-lanceolate, 3.5–10 mm × 1–7 mm, glabrous, yellow; stamens 10, with 5 longer and 5 shorter filaments connate at base, the shorter rarely with rudimentary anthers; ovary superior, 5-celled, styles 5, free, with small, cylindrical stigmas. Fruit a linear-cylindrical, sometimes ellipsoid, pentagonal capsule, 9–20(–24) mm × 2–4 mm, pubescent, green, opening by longitudinal loculicidal slits, many-seeded. Seeds flattened-ovoid, about 1 mm long, reddish-brown; testa with about 3 regular rows of 7–10 transversally connected rows of ridges; aril bivalved, membranous, white. Seedling with epigeal germination; cotyledons leafy, with cuneate base and rounded apex, glabrous; hypocotyl elongated, epicotyl absent.

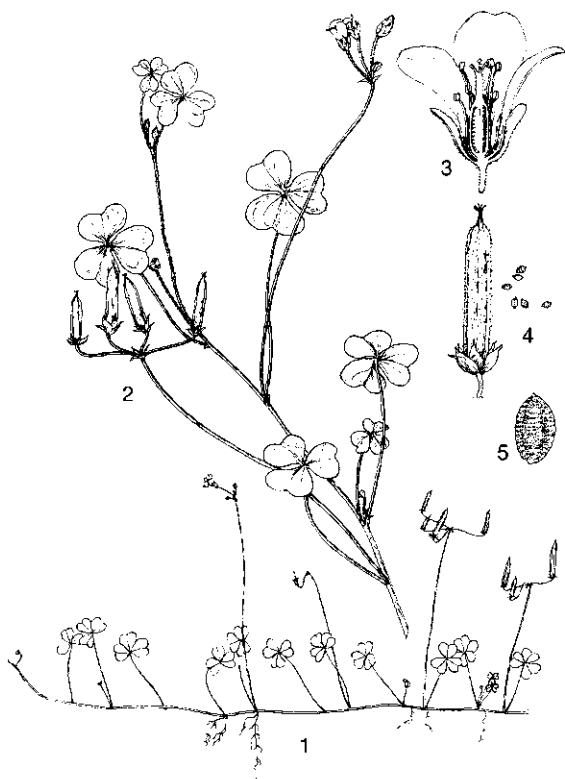
**Growth and development** In the evening the leaves of *O. corniculata* fold down around the leaf stalk. Plants usually flower throughout the year in the tropics. It is reported to be a facultative self-pollinator. Plants may produce seed about 20 weeks after germination. The seeds are forcefully ejected from the fruit in dry weather by the elastic aril.

**Other botanical information** Some other *Oxalis* species are occasionally used for medicinal purposes, e.g. *O. magellanica* J.G. Forster to aid conception in Papua New Guinea. *O. corymbosa* DC. is also used medicinally, e.g. in Vietnam and India, but its main use is as a culinary herb.

**Ecology** *O. corniculata* is a common weed in gardens, fields, grassland, in roadsides, on river banks and on walls, up to 3000 m altitude. It thrives best in warm, moist, freely drained and fertile soils with a pH of 3.5–6.5. It is commonly found in vegetables, groundnut, maize, soya bean and root crops in the Philippines, and is a weed in tea and rice in Indonesia. In temperate regions, *O. corniculata* is mainly a problem in greenhouses, but sometimes escapes to establish locally outdoor populations. Its prostrate habit helps to protect it from being mowed in lawns.

**Propagation and planting** A single plant of *O. corniculata* may produce up to 26 000 seeds. Seed viability is approximately 95%, and seeds require low levels of light for germination. When stored under dry conditions, they may retain germination rate of over 80% until 8 months, and of 50% after 15 months.

**Diseases and pests** *O. corniculata* may act as a host for several pathogenic organisms, e.g. *Botry-*



*Oxalis corniculata* L. – 1, creeping stem with flowers and fruits; 2, part of stem with flowers and fruits; 3, flower in longitudinal section; 4, fruit with seeds; 5, seed.

*tis*, *Fusarium*, *Microsphaera* and *Puccinia* species, and for organisms feeding on crops, e.g. *Meloidogyne* nematodes and insects of the genera *Ovarus* and *Argynnis*.

**Harvesting** *O. corniculata* is common around houses and on agricultural land. It is collected whenever the need arises.

**Genetic resources and breeding** *O. corniculata* is extremely widespread and occurs in anthropogenic habitats, making it not at risk of genetic erosion. There is even much effort to eradicate it as a noxious weed, but it seems well able to survive since it is often resistant to hormone-type herbicides.

**Prospects** Since hardly any information is available on the pharmacological activity of *O. corniculata* and its compounds it is impossible to indicate its future importance as a medicinal plant in modern phytotherapy.

**Literature** |1| de Padua, L.S. & Pancho, J.V., 1983. Handbook on Philippine medicinal plants. Vol. 4. University of the Philippines at Los Baños, the Philippines. p. 43. |2| Dharma, A.P., 1981. Indonesische geneskrachtige planten [Indonesian medicinal plants]. De Driehoek, Amsterdam, the Netherlands. p. 41. |3| Goh, S.H., Chuah, C.H., Mok, J.S.L. & Soepadmo, E., 1994. Malaysian medicinal plants for the treatment of cardiovascular diseases. Penerbit Universiti Malaya, Kuala Lumpur, Malaysia. pp. 111-112. |4| Gunasegaran, R., 1992. Flavonoids and anthocyanins of three Oxalidaceae. *Fitoterapia* 63(1): 89-90. |5| Holm, L.G., Plucknett, D.L., Pancho J.V. & Herberger, J.P., 1977. The world's worst weeds, distribution and biology. East-West Center, The University Press of Hawaii, United States. pp. 343-347. |6| Nguyen Van Duong, 1993. Medicinal plants of Vietnam, Cambodia and Laos. Mekong Printing, Santa Ana, California, United States. p. 299. |7| Perry, L.M., 1980. Medicinal plants of East and Southeast Asia. Attributed properties and uses. The MIT Press, Cambridge, Massachusetts, United States & London, United Kingdom. p. 135. |8| Quisumbing, E., 1978. Medicinal plants of the Philippines. Katha Publishing Co., Quezon City, the Philippines. pp. 442-443. |9| Sridhar, R. & Lakshminarayana, G., 1993. Lipid classes, fatty acids, and tocopherols of leaves of six edible plant species. *Journal of Agricultural and Food Chemistry* 41(1): 61-63. |10| Veldkamp, J.F., 1971. Oxalidaceae. In: van Steenis, C.G.G.J. (General editor): *Flora Malesiana*. Series 1, Vol. 7. Noordhoff International Publishing, Leyden, the Netherlands. pp. 155-156.

**Other selected sources** 202, 371, 580, 676, 1515, 1572.

R.C.K. Chung

## Papaver L.

Sp. pl. 1: 506 (1753); Gen. pl. ed. 5: 224 (1754).

PAPAYERACEAE

$x = 7, 14$ ; *P. rhoeas*:  $2n = 14$  *P. somniferum*:  $2n = 20, 22, 36, 44$

**Major species** *Papaver somniferum* L.

**Vernacular names** Poppy (En).

**Origin and geographic distribution** *Papaver* consists of about 100 species, most of which are found in the Mediterranean region. The most important species is *P. somniferum* (opium poppy), the source of opium. The cultivation of *Papaver* for medicinal purposes dates to about 2000 BC in Mesopotamia. The cultivation of *P. somniferum* has been prohibited in many countries due to the misuse of opium and its deleterious effects on human health.

**Uses** About 2350 years ago, the Greek physician Hippocrates mentioned poppy juice ('opium') as a cathartic, hypnotic, narcotic and styptic. *P. somniferum* is the only source of opium, which is the dried, alkaloid-containing latex obtained from the immature capsules of the plant. When the surface of these unripe fruits is damaged, e.g. by making incisions with a specially designed knife, latex oozes out in the form of white droplets which quickly turn brown and become semi-solid. Commercial opium is used medicinally, as a starting material for the extraction of several medicinally important alkaloids, or as a narcotic.

The only *Papaver* species with medicinal use in South-East Asia are *P. somniferum* and *P. rhoeas*. The dried fruit without the seed of *P. somniferum* is the 'anh tuc xac' of the traditional Vietnamese Pharmacopoeia, used to treat cough and diarrhoea. Dried fruits have also been used to treat cough in Europe and diarrhoea and cough in China. In northern Thailand, as well as in many other countries worldwide, opium is used to treat pain, diarrhoea, cough and insomnia, or as a narcotic. It is sometimes taken orally, but also often smoked, which is habit-forming and may subvert the personality. The major role of opium in medicinal practice has been for its astringent properties. Standardized opium tincture and camphorated opium are prescribed for severe diarrhoea and dysentery, as well as for pain, cough and nausea. Although still present in official pharmacopoeias

of some countries, the use of opium or opium preparations has largely been superseded by its purified, extracted alkaloids, mainly morphine, codeine and noscapine (= narcotine).

The flowers and roots of *P. rhoeas* are recommended in China for treating jaundice and in Japan and the Philippines an infusion of the flowers may be used as a gargle or ingested as a bechic. The petals have a reputation as having mild sedative, antitussive, anodyne and sudorific properties. Infusions or syrups made from the petals have been used as a red colourant in pharmacy; however, they act as litmus, turning red with acid and blue with base.

*P. somniferum* is also a source of edible seeds and seed oil. The seeds are used in food, for instance in cakes and on bread. They are also used in bird feed. Poppy seed oil is mainly used as an edible oil, but also for the production of paint and soap. Poppy-seed cake or meal left after extraction of the oil is sweet and nutritious and used alone or mixed with other feeds, suitable as food for cattle and other animals. However, its use as animal feed has sometimes resulted in poisoning due to the presence of alkaloids arising from contamination of the seed with particles of the capsule. Some tribes in South-East Asia eat the young leaves of *P. somniferum*.

In Java, several *Papaver* species (e.g. *P. nudicaule* L., *P. rhoeas* and *P. somniferum*) used to be grown as ornamentals. They have bright flowers, ranging from white to almost black, and through various shades of yellow, pink, orange, scarlet and crimson.

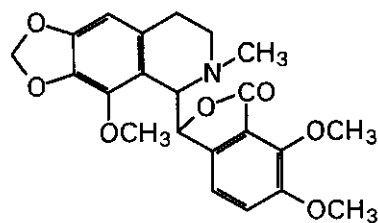
**Production and international trade** *P. somniferum* is cultivated for the production of opium, straw for extraction of alkaloids, and/or seeds and seed oil. Poppy straw consists of the upper stalks with crushed capsules. The legal production of opium is limited to India, but extensive illegal opium production is found in the 'Golden Triangle' (the border region of Thailand, Burma (Myanmar) and Laos), the 'Golden Crescent' (Pakistan, Afghanistan, Iran), Lebanon and Mexico. Total legal opium production in 1983 was 1000 t, produced in India on about 32 000 ha. The opium-producing areas in Burma (Myanmar), Thailand and Laos were estimated at respectively 60 000 ha, 3500 ha and 3500 ha. Opium production estimates for 1989 were for Burma (Myanmar) 2000 t, for Afghanistan 800 t, for Laos 400 t, for Pakistan 130 t, and for Thailand 50 t.

Production for straw is mainly found in the more temperate zones, being for Spain 13 700 t, for

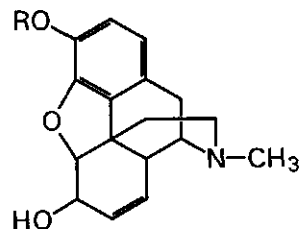
France 3700 t, for Australia 1180 t and for Turkey 940 t in 1980.

**Properties** A large number of alkaloids have been isolated from *Papaver*. At least 25 alkaloids that are biosynthetically derived from the amino acid tyrosine have been isolated from opium. The most important are morphine (5–25%), codeine (0.2–3%), thebaine (0.2–5%), noscapine (2–10%), narceine (0.1–0.7%) and papaverine (0.5–3%). Biosynthetic routes are extremely complex within the family of *P. somniferum* alkaloids, thus several subgroups of related compounds can be recognized. Of the 6 main alkaloids, morphine, codeine and thebaine belong to the phenanthrene (or morphinane) type, noscapine and narceine belong to the phthalide-isoquinoline type, and papaverine to the benzyl-isoquinoline type. Other alkaloids include aporeine, codamine, cryptopine, gnoscopine, hydrocotarnine, lanthopine, laudanidine, laudanine, laudanosine, meconidine, narcotoline, neopine, oxynarcotine, papaveramine, porphyroxine, protopine, reticuline, rhoeadine and xanthaline. Many of these opium alkaloids are not only present in the fruit but also in other parts of *P. somniferum*.

In opium, the alkaloids are largely present as salts of organic acids, such as meconic or lactic acid. The drug also contains sugars, salts (e.g. sulphates), free organic acids such as meconic, lactic, fumaric and oxalacetic acid, albuminous substances, colouring matters and water. Most of the world legal opium production is used to obtain morphine, codeine and noscapine, which are medi-



noscapine



morphine (R = H) and codeine (R = CH<sub>3</sub>)



cinally the most important. A large part of the isolated morphine is converted into codeine by chemical semisynthesis, since there is much greater demand for this compound. Morphine can also be converted into heroin by addition of acetyl groups; this substance is very addictive.

Morphine (named after Morpheus, the greek god of dreams) as a pure substance is mostly applied as one of its salts (e.g. HCl or sulphate). It is a very strong analgesic, acting on the central nervous system and widely used to relieve pain. Furthermore, morphine has digestive effects leading to constipation, and it is antidiuretic. At high doses it causes depression of respiration, bradycardia, vasodilatation and hypotension, but the best known side effects are the development of psychic and physical dependence. Morphine acts by interaction with specialized receptors, called opioid receptors, of which several types (e.g.  $\mu$ ,  $\kappa$ ,  $\sigma$ ,  $\delta$ ) are recognized. There is also evidence of the existence of several subtypes.  $\mu$ -Receptor activation in the brain is presumed to be responsible for the strong central analgesic effect.  $\kappa$ -receptors in the brain and spinal cord also appear capable of producing analgesia, particularly in the spine. Most of the psychomimetic effects (e.g. hallucinations) are apparently mediated by  $\sigma$ -receptors. The  $\delta$ -receptor is presumed to be the primary receptor for endogenous opioid pentapeptides known as enkephalins. These enkephalins, together with the other endogenous opioid peptide families endorphines and dynorphines, and the opioid receptors, are believed to be responsible for the modulation of nociception, especially in cases of pain. Endogenous opioid peptides regulate respiration ( $\mu$ -receptor); respiratory depression is therefore a very significant side effect of morphine, even at therapeutic levels. Some endogenous opioid peptides and receptors also exist in tissues and organs other than the brain: the gastro-intestinal tract is another principal site of action ( $\mu$ -,  $\kappa$ -receptor; decreasing of motility).

Heroin is the diacetyl ester of morphine. The effects of heroin are similar to those of morphine, but it is more addictive and causes disruptions of blood flow, infections and collapsed blood veins. The manufacture and possession of heroin is illegal in most countries.

Codeine is the most widely used opium alkaloid, which also has some analgesic properties of its own. However, it is known to enhance the effects of other (non-opiate) analgesics, and thus it is often given in combination with e.g. paracetamol (acetaminophen). Codeine acts like morphine, but

it is far less narcotic, euphoric or constipating. It has strong antitussive properties, and it is often employed in cough medicines and decongestants.

Thebaine is more a convulsant than a narcotic. It may cause tetanic spasms, which do resemble those caused by strychnine. The compound has been used as a narcotic antagonist in the treatment of heroin addiction, although better alternatives (e.g. naloxone) are available at present. Noscapine (narcotine) is not an analgesic, but a moderately strong antitussive. Like codeine, it is often used in cough medicines, especially for children. Papaverine has very little narcotic or analgesic action, but relaxes the involuntary muscles of the body and increases the blood flow by its effects on heart and blood vessels.

The leaves, stems and roots of *P. rhoeas* contain coptisine and protopine, whereas the capsules contain rhoeadine, rhoeagenine and rhoearubine. The pharmacology of the main alkaloid, rhoeadine (a tetrahydrobenzazepine) is unknown, but closely related derivatives are dopaminergic antagonists and neuroleptics. The presence of *P. rhoeas* in hay can be dangerous to animals.

The seeds of *Papaver* are rich in oil, carbohydrates, proteins and calcium. The oil content of 10 different *Papaver* species was found to range from 35% to 48%, the protein content from 21% to 28%. The fatty acids in poppy-seed oil are linoleic acid (65%), oleic acid (25%) and saturated acids (6-10%).

**Adulterations and substitutes** An alternative source of codeine is *P. bracteatum* Lindley, which grows wild in mountainous regions in western Asia. It contains thebaine, but no morphine or codeine. Thebaine can easily be converted into codeine and morphine by chemical methods, but is not readily converted into heroin.

Synthetic derivatives of morphine with fewer side effects have been developed, such as meperidine, methadone and fentanyl. However, the isolation of morphine derivatives from natural sources still seems easier and cheaper than synthetic production.

In Western medicine, preparations from hemp (*Cannabis sativa* L.) were extensively used between the middle of the 19th Century and the Second World War as a milder and less dangerous analgesic than opium.

**Description** Herbs or sometimes shrubs up to 150 cm tall, smooth or hispid, pale green or glaucous, with bitter, milky latex. Leaves arranged spirally or in a radical rosette, often deeply incised. Flowers solitary, bisexual, long pedicelled;

sepals 2-3, free, deciduous when the flower expands; petals 4-6, corrugate in bud, fugacious; stamens numerous, much shorter than corolla; ovary superior, with 4-18 parietal, much intruded placentas and many ovules, stigma sessile, peltate, rather deeply lobed, lobes as many as placentas, opposite to them, radiating. Fruit a capsule, globose, urceolate to broadly ovoid, campanulate or cylindrical-clavate, stalk-like contracted at the base, under the stigmatic shield dehiscing by transverse pores (arising by the recurving of short valves). Seeds numerous, globose to slightly reniform, reticulate. Seedling with sessile or shortly petioled, linear cotyledons, base cuneate; hypocotyl elongate, epicotyl absent.

**Growth and development** Most *Papaver* species are annuals or biennials, including *P. rhoeas* and *P. somniferum*. The seed of *P. somniferum* germinates in 10-15 days. The minimum temperature for germination is around 3°C. The seedling grows slowly; it takes a month to reach the rosette stage. After that, stem differentiation occurs and growth is more rapid. Reproductive buds appear at about one month from the rosette stage. About a week before flowering, the pedicel bends, but it unbends when the flower opens. Flowering occurs about 90-100 days after sowing; the petals are shed within 3 days. After fertilization, the fruit grows fast, reaching its final size within 2 weeks. In another 10-15 days, the fruits become ready for lancing and latex collection. The seeds reach maturity 2 weeks after that.

**Other botanical information** *Papaver* spp. have been grouped in different sections, but no consensus exists. Neither is there agreement on the relationship between *P. somniferum* and *P. setigerum* DC., which some taxonomists separate, whereas others lump them into one very variable species. Sometimes 3 subspecies are recognized: *P. somniferum* subsp. *somniferum* (opium poppy) cultivated and escaped throughout most of Europe, *P. somniferum* subsp. *songaricum* Basil. cultivated in the Balkans and Asia, and *P. somniferum* subsp. *setigerum* (DC.) Corb. (setaceous poppy) probably native in the western Mediterranean region and a weed elsewhere.

**Ecology** *Papaver* is found in the relatively dry temperate and subtropical parts of the Old World and does not tolerate tropical lowland conditions. In the 'Golden Triangle', opium poppy only grows well when grown above 850 m altitude, and the best opium is obtained from above 1000 m altitude. *P. somniferum* is a long-day plant. Both *P. somniferum* and *P. rhoeas* are adapted to open or

disturbed habitats, and commonly found in association with cereal cropping. Regions where it may rain during the latex stage of the crop are not suitable for latex production, because exuded latex may be washed away. Windy areas should be avoided, to prevent lodging of the heavy-capsuled plants. Opium poppy can be grown on a relatively wide range of soils.

**Propagation and planting** Most *Papaver* species are propagated by seed. Seed is broadcast (3-4 kg/ha) and plants are thinned later to a distance of 20-25 cm when about 10 cm tall. Seedlings resent transplanting. In Tasmania, poppy is sown in rows with 100-150 plants/m<sup>2</sup>. Some hill tribes in the 'Golden Triangle' practise staggered planting, to spread the harvest over a longer period, but others sow their fields in one go. Seed of *P. somniferum* is very small, with a 1000-seed weight of less than 0.2 g; the colour varies from yellowish-white to purplish-black.

**In vitro production of active compounds** Tissue culture of both *P. somniferum* and *P. bracteatum* is possible and may serve as a future technique in commercial production of codeine.

**Husbandry** In northern Thailand, *P. somniferum* is usually sown in September-October after maize on recently cleared land. In India, *P. somniferum* is sown in November. *P. somniferum* has high nutrient requirements. It benefits from phosphate fertilization during early growth, whereas later on nitrogen fertilization, at 100-140 kg/ha, will help to increase opium yield and morphine content. Under Tasmanian conditions, irrigation can have a beneficial effect. The crop especially needs adequate water at the start of flowering. Because of the heavy capsules, lodging can be a serious problem at higher plant densities.

**Diseases and pests** In general, *P. somniferum* is not regarded as very susceptible to diseases and pests. It is reported to be susceptible to downy mildew (*Peronospora arborescens*), leaf blight (*Helminthosporium* spp.), root rot (*Rhizoctonia* sp.) and thread mould (*Dactylium roseum*). Insect pests include *Agrotis suffusa* (opium cut worm), *Agrotis epsilon*, *Aphis fabae*, *Franklinella* sp., *Heliothis armigera*, *Myzus persicae*, *Nephotettix* sp., *Pachycephus smyrnensis* and *Phytomyza horticola*.

**Harvesting** Latex is collected before the capsules are mature and dry. They are carefully incised; the depth of the incision is very important. It must be deep enough to reach the laticiferous ducts, but not so deep that it cuts into the endocarp, otherwise the latex flows inside the capsule.

Lancing or scarification is done in the afternoon. The exuding latex is then left to coagulate overnight, during which time it will become darker and thicker. The next morning it is collected with a special scraper and air dried. Opium collection is a labour-intensive activity. In northern Thailand, the capsules are incised when they change colour from green to slightly grey-green. The harvest period lasts from a few days up to a week after the last petals have dropped from a flower. Tapping is carried out with a special knife, consisting of several sharp blades bound together so that one stroke makes several incisions on the capsule, about 1 mm apart. The lancing and collection is repeated 3–4 times for each capsule in as many days. The capsules are then allowed to mature, and those from the most robust plants are harvested to obtain seed for the next year. The hill tribes of the 'Golden Triangle' do not use the seed for food or oil. When the crop is grown for straw extraction, the fruits are harvested after maturity with 10–30 cm of the pedicel attached. The seeds are separated and the straw is used for extraction of alkaloids. The crop can also be harvested about 3 weeks before flowering, giving 'green poppy', which is rich in alkaloids but has to be dried quickly.

When grown for seed production, the crop is harvested when the capsules turn yellow-green and the seeds inside rattle.

**Yield** Yield levels in northern Thailand depend on how frequently poppy has been grown on a given field. In the first year, the yield is high, but it drops the second year, and it is usually not worth planting after the third year. The yield is also influenced by the number and mode of lancing practices. The first lancing gives raw opium with a higher morphine content than subsequent lancing; thus, the raw opium obtained from different lancing is stored separately. Terminal capsules (each plant bears 5–8 capsules) yield opium of considerably higher morphine content than the lateral ones. In India, average yield of raw opium is around 30 kg/ha, but yields as high as 60 kg/ha have been reported. When poppy is grown for seed, seed yields are usually between 1.2 and 1.8 t/ha. High commercial yields are obtained in Tasmania, where average dry matter yield of poppy heads (capsules and seeds) is 2.5 t/ha. In Tasmanian field experiments, 3–4 t/ha of poppy heads have been obtained, and morphine yields of 18–30 kg/ha.

**Handling after harvest** In northern Thailand, the harvested opium is carefully packed in leaves

before marketing. In India, the raw opium is stored in metal or earthen pots (perforated at the bottom or placed at a tilt to allow the moisture to drain off) and turned over every 10 days to give it a uniform consistency. After that, it is sun-dried and packed in sacks or jars.

Opium requires careful storage as morphine is quickly decomposed by enzyme activity on exposure to air at 15.5°C. However, storing in sealed containers at 36–37°C gave little loss. Under moist conditions, opium may become covered with mould but this does not seem to affect its morphine content. The total alkaloid content of opium may vary from 5–25%. The opium alkaloids (in particular, morphine and codeine) and their salts may be extracted from either raw opium or poppy straw, the process involving concentration of aqueous extracts to a syrupy consistency, followed by precipitation of alkaloids from concentrated extracts.

When poppies are grown for straw production, the seeds are separated from the capsules and pedicels, which are used for direct alkaloid extraction. Oil is extracted from the seeds by pressing or by solvent extraction of crushed seed.

**Genetic resources and breeding** Cross-pollination is common in *P. somniferum*, and a crop can contain a wide range of forms. Breeding has mainly been carried out in Europe. Until the 1970s, European breeding work was aimed at increased seed and oil yields as well as high alkaloid contents. The increasing abuse of opiates as heroin has stimulated attempts to reduce illicit traffic in opium by banning cultivation of the opium poppy, and to search for *Papaver* spp. which do not contain morphine. One of these is *P. bracteatum*, which lacks an enzyme for the demethylation of thebaine. It produces this alkaloid instead of morphine and codeine, and its content can reach up to 3.5% in the dried capsules. The amount is sufficient for economic production on a large scale; chemical demethylation of thebaine to codeine is relatively simple, demethylation of codeine to morphine, however, is not easy. For *P. somniferum*, breeding in more recent years has been aimed at creating types with a low morphine content. *P. orientale* L. and *P. bracteatum* have been crossed to develop perennial types containing thebaine but no morphine, but the resulting progenies were sterile.

Many varieties, strains and hybrids of ornamental poppy with single and double flowers and some with fringed petals have been raised.

**Prospects** *P. somniferum* as a medicinal crop

has generally declined in importance due to its narcotic reputation and the strict regulations imposed on any such ventures. Only the production of limited quantities of drug for legitimate uses is allowed. This situation is likely to continue for the foreseeable future. Nevertheless, in South-East Asia illegal planting still exists, especially in northern Thailand, Laos and Burma (Myanmar). Opium poppy is a popular cash crop for the hill tribes in these areas. In certain areas, activities may also include illegal processing into morphine and heroin. However, there has recently been a drop in opium production in northern Thailand, mainly due to the efforts of the government to suppress opium poppy cultivation and to develop the inaccessible poppy-growing areas. Cultivation as an ornamental and for the production of edible seeds and seed oil may hold out some prospects but should be monitored closely to prevent abuse. Investigation into the potential of poppy flowers for the production of colouring agent (dye) may be worthwhile.

**Literature** [1] Anderson, E.F., 1993. Plants and people of the Golden Triangle. Ethnobotany of the hill tribes of Northern Thailand. Dioscorides Press, Portland, United States. 279 pp. [2] Broszat, W., 1992. Der Mohn (Papaver somniferum L.): Anbau und Markt einer wiederentdeckten Kulturpflanze [Poppy (Papaver somniferum L.): cultivation and market of a rediscovered crop]. Der Tropenlandwirt (Journal of Agriculture in the Tropics and Subtropics), Beiheft No 47. Deutsches Institut für Tropische und Subtropische Landwirtschaft, Witzenhausen, Germany. 170 pp. [3] Bruneton, J., 1995. Pharmacognosy, phytochemistry, medicinal plants. Lavoisier Publishing, Paris, France. pp. 749–770. [4] Council of Scientific and Industrial Research, 1966. The Wealth of India: a dictionary of Indian raw materials and industrial products. Raw materials. Vol. 7. Publications and Information Directorate, New Delhi, India. pp. 231–248. [5] Duke, J.A., 1973. Utilization of papaver. Economic Botany 27: 390–400. [6] Godin, V.J. & Spensley, P.C., 1971. Crop and Products Digests 1. Oils and oilseeds. The Tropical Products Institute, London, United Kingdom. pp. 111–115. [7] Husain, A. & Sharma, J.R., 1983. The opium poppy. Medicinal and aromatic plants series No 1. Central Institute of Medicinal and Aromatic Plants, Lucknow, India. 167 pp. [8] Kapoor, L.D., 1995. Opium poppy: botany, chemistry, and pharmacology. Food Products Press, New York, United States. 326 pp. [9] Merlin, M.D., 1984. On the trail of the ancient opium poppy.

Fairleigh Dickinson University Press, Rutherford, Madison, Teaneck, United States. 324 pp. [10] Salzer, W., 1994. Entwicklung statt Opium: Anmerkungen zum Mohnersatzkulturenanbau in Nord-Thailand [Development instead of opium: observations on the cultivation of substitute crops in northern Thailand]. Entwicklung und ländlicher Raum 28(4): 16–20.

#### *Selection of species*

#### **Papaver rhoeas L.**

Sp. pl. 1: 507 (1753).

**Vernacular names** Corn poppy, red poppy (En). Pavot, pavot rouge, coquelicot (Fr). Vietnam: h[oo]fng anh.

**Distribution** The origin of *P. rhoeas* is still unknown. Nowadays, it is common all over Europe, mainly along country roads and on wasteland. It is sometimes grown as an ornamental in the Malesian region.

**Uses** In the Malesian region, the flowers are used for their mild sedative, antitussive, anodyne and sudorific properties.

**Observations** An erect annual herb, 30–80 cm tall; medium and superior leaves petioled or sessile from a narrowed, non-amplexicaul base, deeply pinnatifid bipinnatifid, pale green, hispid; pedicel usually with patent to erecto-patent, but sometimes appressed bristles, petals 3–5 cm long, bright or pale red to bluish, occasionally white-margined or entirely white, often with a black basal blotch, flowers sometimes double, filaments not widened at the top, stigma broader than top of fruit, stigmatic rays mostly 8–13, black-purple; fruit campanulate; seeds dark brown.

**Selected sources** 97, 193, 287, 376, 931, 1126.

#### **Papaver somniferum L.**

Sp. pl. 1: 508 (1753).

**Synonyms** *Papaver officinale* C.C. Gmelin (1806), *Papaver hortense* Hussenot (1835), *Papaver setigerum* DC. (1893).

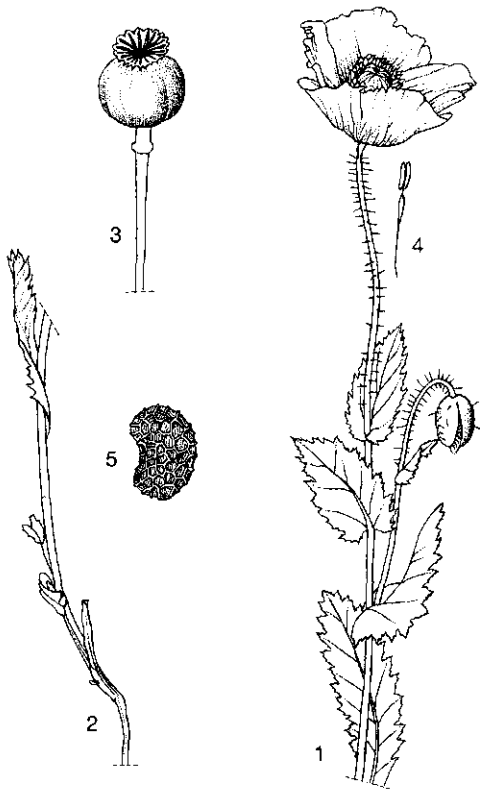
**Vernacular names** Opium poppy, mawseed (En). Pavot officinale (Fr). Thailand: ya pi (Akha), ya fin (Lahu). Vietnam: c[aa]y thu[oos]c phi[eej]n, c[aa]y anh t[us]c.

**Distribution** The origin of *P. somniferum* is not clear; suggestions range from south-western Europe to central Asia. No truly wild populations of *P. somniferum* have been found. The main areas of cultivation are in India, China, Turkey and the Balkans. In South-East Asia, it is grown in the

'Golden Triangle' (Thailand, Burma (Myanmar), Laos) and Vietnam.

**Uses** Opium is traditionally used for its astringent properties in the treatment of coughs and diarrhoea, and also to relieve pain and to treat insomnia.

**Observations** An annual erect herb, 50–150 cm tall; medium and superior leaves sessile with an amplexicaul base, varying from obovate to ovate-oblong, coarsely crenate-dentate, not deeply incised, strikingly glaucous, glabrous; pedicel glabrous or hispid, petals 3.5–8 cm long, lilac with darker base, white (with or without a dark basal blotch), red (with or without a dark basal blotch) or variously marked, entire or variably deeply incised, flowers often double, stigma much broader than top of the fruit, mostly 8–15-rayed; fruit a globose capsule, 5–10 cm wide, with copious latex; seeds blue, black, yellow or white. *P. somniferum* is an extremely variable and complex species. There is still disagreement about its intraspecific classification. It does not thrive in the per-humid tropical lowlands.



*Papaver somniferum* L. - 1, flowering stem; 2, stem base; 3, fruit; 4, stamen; 5, seed.

**Selected sources** 62, 63, 64, 97, 189, 193, 202, 265, 266, 287, 387, 555, 701, 813, 823, 900, 931, 1035, 1126, 1167, 1277, 1296, 1304, 1457.

Khozirah Shaari & M. Brink

### *Peperomia pellucida* (L.) Kunth

Humb., Bonpl. & Kunth, Nov. gen. sp. 1: 64 (1816).

PIPERACEAE

$2n = 44$

**Synonyms** *Piper pellucidum* L. (1753), *Piper exiguum* Blume (1826).

**Vernacular names** Indonesia: ketumpangan air (Malay, Sumatra), sasaladaan (Sundanese), suruh-suruhan (Javanese). Malaysia: ketumpangan air (Malay, Peninsular). Philippines: ulasiman-bato (Tagalog), olasiman-ihalas (Cebu Bissaya), tangon-tangon (Bikol). Thailand: chaa kruut (peninsular), phak krasang (central), phak haak kluai (northern). Vietnam: rau c[af]ng cua.

**Origin and geographic distribution** *Peperomia* comprises over 1000 species. Its main centre of distribution is in Central and South America. In South-East Asia, 50–90 species occur. *P. pellucida* is native to South America but has naturalized widely in the Old World tropics. It is common in South-East Asia, occurring throughout the region.

**Uses** In the Philippines, the whole plant of *P. pellucida* is used as a warm poultice to treat abscesses, boils and pimples. An infusion or decoction is used against gout, kidney troubles and rheumatic pain, and externally as a rinse for complexion problems. In Peninsular Malaysia, the plant is boiled and the water drunk to relieve rheumatism and fatigue. In Java, the juice of the leaves is prescribed for colic and abdominal pains, and the bruised leaves are applied to the temples to treat headache. The plant is eaten as a salad. In West Africa, it is similarly eaten as a vegetable, and used in local medicine to treat convulsions. In Central and South America, it is widely used in local medicine in similar applications as described for the Philippines. It is recommended to eat *P. pellucida* as a culinary herb because it has an aromatic taste and stimulates appetite and digestion.

**Properties** *P. pellucida* contains an essential oil with apiole (a phenylpropane derivative) as the main component. Further components are the related 2,4,5-trimethoxystyrene, caryophyllene (a sesquiterpenoid), and an (unidentified) sesquiter-

pene alcohol. Apiole affects the kidney parenchyma; there are reports in the literature of kidney and liver damage due to apiole. The known strong diuretic activity is also probably toxic. In pure form the compound stimulates the uterus, provoking menstruation; its misuse as an abortifacient which can lead to serious intoxications is also known.

The essential oil from the whole plant showed fungicidal activity at a minimum inhibitory concentration of 2000 ppm, with a wide range of toxicity and quick killing activity. It was thermostable, remained toxic for at least 150 days, was non-phytotoxic and non-systemic. The oil is antagonistic to the growth of *Helminthosporium oryzae* (brown spot) in rice.

An ethyl acetate extract of the air-dried plants showed antibacterial activity against *Bacillus subtilis*, *Pseudomonas aeruginosa* and *Staphylococcus aureus*. The antibacterial activity was more potent than the penicillin standard used. The water extract of leaves showed antimutagenic activity.



*Peperomia pellucida* (L.) Kunth - 1, flowering plant; 2, part of infructescence.

Several flavonoids have been isolated from the plant, including acacetin, apigenin, pellucidatin, pellucidatin-8-neohesperidoside and isovitexin.

**Adulterations and substitutes** Apiole, the main constituent of the essential oil, is better known from the root of parsley (*Petroselinum crispum* (Miller) Nyman ex A.W. Hill).

**Description** A small fleshy herb, up to 30 cm tall; stems initially erect, becoming decumbent, rooting at nodes, glabrous, internodes up to 5 cm long and 2 mm in diameter. Leaves spirally arranged, simple, ovate elliptical to broadly ovate or almost triangular, 2.5–3.5 cm × 2–3 cm, entire, membranous when dry, 5-veined, base rounded to truncate, apex acute; petiole up to 20 mm long and about 1 mm in diameter, glabrous; stipules absent. Inflorescence a terminal or axillary spike, solitary, glabrous, peduncle 0.5–1.5 cm long and about 0.5 mm in diameter, fertile axis 2–5 cm long and about 0.5 mm in diameter. Flowers bisexual, sessile, not sunken into axis, spaced 0.4–1 mm apart; floral bracts rounded, 0.3–0.4 mm × 0.2–0.3 mm; perianth absent; stamens 2, anthers oblong, about 0.1 mm × 0.1 mm; ovary superior, rounded-oblong, about 0.3 mm × 0.3 mm, 1-locular. Fruit drupe-like, subglobose, 0.5–1 mm in diameter, sticky, papillate, 1-seeded.

**Growth and development** *P. pellucida* is presumed to be an annual but its lifespan is unknown. The profusely produced seed is probably dispersed by rain wash and more widely by people as a contaminant in soil. Plant growth is fast under moist conditions.

**Other botanical information** Some botanists separate *Peperomia* from the *Piperaceae* into the *Peperomiaceae*, on the basis of the absence of stipules, the presence of two stamens and on differences in pollen morphology and anatomy.

Although there is a recent revision for Australia (and Africa), no overall revision of *Peperomia* is available for South-East Asia. It is very difficult to compare the names and species for the Philippines, New Guinea, Peninsular Malaysia and Java from the literature, and no botanical information is available for other Malesian regions. *P. tetraphylla* (J.G. Forster) Hook. & Arn. is another pantropical representative that occurs in South-East Asia. Although it is used in local medicine in South America, there are no reports of its medicinal use in South-East Asia.

**Ecology** *P. pellucida* is mostly found in disturbed habitats up to 1000 m altitude. It is a common and widespread weed, frequent in gardens and cultivated areas that are damp and lightly

shaded, particularly common on damp hard surfaces such as walls, roofs, steep gullies, and in flower pots. Under certain growing conditions impoverished types that were previously described as separate species can develop.

**Propagation and planting** *P. pellucida* produces seed in abundance. It is not cultivated in South-East Asia. In West Africa, however, it is cultivated from seed and reputed for its fast growth.

**Harvesting** *P. pellucida* is collected fresh whenever the need arises.

**Genetic resources and breeding** Since *P. pellucida* is a common pantropical weed the risk of genetic erosion is minimal.

**Prospects** *P. pellucida* is easy to grow, even in urban settings, and is a nutritious and savoury vegetable. Its fungicidal properties deserve further research for low-budget applications. The significant antibacterial activity of the ethyl acetate extract suggests a potential as a broad-spectrum antibiotic.

**Literature** |1| de Padua, L.S. & Pancho, J.V., 1983. Handbook on Philippine medicinal plants. Vol. 4. University of the Philippines, Los Baños, the Philippines. p. 45. |2| Düll, R., 1973. Die Peperomia-Arten Afrikas [The Peperomia species of Africa]. Botanische Jahrbücher für Systematik, Pflanzengeschichte und Pflanzengeographie 93: 56–129. |3| Forster, P.I., 1993. A taxonomic revision of the genus Peperomia Ruiz & Pav. (Piperaceae) in mainland Australia. Austrobaileya 4(1): 93–104. |4| Jose, J., Thoppil, J.E. & Mathew, L., 1992. Chromosome complement studies in five species of Peperomia Ruiz and Pav. Cytologia 57(2): 227–229. |5| Morton, J.S., 1981. Atlas of medicinal plants of middle America. Bahamas to Yucatan. Charles C. Thomas Publisher, Springfield, Illinois, United States. pp. 120–121. |6| Nguyen Van Duong, 1993. Medicinal plants of Vietnam, Cambodia and Laos. Mekong Printing, Santa Ana, California, United States. p. 331. |7| Poscidio, G.N., Garcia, E.A. & Bojo, A.C., 1993. Antibacterial activity of Peperomia pellucida (L.) HBK, family Piperaceae. Philippine Journal of Biotechnology 4(2): 199. |8| Quisumbing, E., 1978. Medicinal plants of the Philippines. Katha publishing Co., Quezon City, the Philippines. pp. 210–211. |9| Singh, A.K., Dikshit, A. & Dixit, S.N., 1983. Antifungal studies of Peperomia pellucida. Beitrage zur Biologie der Pflanzen 58(3): 357–368. |10| van der Zon, A.P.H. & Grubben, G.J.H., 1976. Les légumes-feuilles spontanés et cultivés du Sud-Dahomey [Wild and cultivated leaf vegetables of

South Dahomey]. Communications 65. Koninklijk Instituut voor de Tropen, Amsterdam, the Netherlands. p. 84.

**Other selected sources** 97, 202, 572, 580, 1227, 1570, 1647, 1648.

R. Kiew

## Phyllanthus L.

Sp. pl. 2: 981 (1753); Gen. pl. ed. 5: 422 (1754).

EUPHORBACEAE

$x = 6, 7, 8, 9, 13, 15$ , most commonly 13; *P. acidus*:  $2n = 26$ , *P. amarus*:  $2n = 26, 52$ , *P. emblica*:  $2n = 98, 104$ , *P. maderaspatensis*:  $2n = 26, 52$ , *P. pulcher*:  $2n = 78$ , *P. reticulatus*:  $2n = 26$ , *P. simplex*:  $2n = 26$ , *P. urinaria*:  $2n = 52$

**Major species** *Phyllanthus acidus* (L.) Skeels, *P. amarus* Schum., *P. emblica* L., *P. reticulatus* Poiret.

**Vernacular names** Phyllanthus, seaside laurel (En). Phyllanthe, bois à enivrer, bois de gaulettes (Fr). Vietnam: ph[ef]n den, me r[uw]ng.

**Origin and geographic distribution** *Phyllanthus* comprises over 700 species and has a pantropical distribution. It is primarily an Asiatic genus, with only about 100 species being native to Africa and some 200 to the New World. Most of the New World species are found in the West Indian region. Southern Brazil appears to be another important centre of speciation. The most important centres of diversity in the Old World are located in India, Indo-China, the Philippines and New Guinea. Within Malesia about 100 species occur.

**Uses** The medicinal applications of *Phyllanthus* have a long history. Several *Phyllanthus* species have been used in India for about 2000 years in the Ayurveda, Unani and Siddha systems of medicine, especially for the treatment of jaundice. Although the species may vary locally, *Phyllanthus* is used in virtually the whole of South-East Asia, the Pacific, East, West and Central Africa, the Caribbean and South America. Generally *Phyllanthus* has aphrodisiac, diuretic and purgative properties, and is used in the treatment of chest complaints, conjunctivitis, cough, diabetes, diarrhoea, oedema, fevers, hepatitis, nephritis, ophthalmic diseases, smallpox and venereal diseases. Some of the medicinal *Phyllanthus* species are also used for dye and tanning purposes (e.g. *P. emblica*, *P. reticulatus*), as edible fruits (*P. acidus*, *P. emblica*), and as ornamentals (*P. pulcher*). The wood of *Phyllanthus* (*P. acidus*, *P. emblica*, *P.*

*reticulatus*) can be used for utensils and other small objects, and as firewood.

Crushed young leaves of *P. gomphocarpus* Hook.f. are used in Indonesia to heal wounds. The roots of *P. elegans* Wallich are used in Indo-China as a febrifuge; the leaves are given to children with a coated tongue.

**Production and international trade** In India, *P. amarus*, *P. debilis* and *P. fraternus* Webster are generally collected from natural stands, often on well-drained, cultivated ground in the rainy season. *P. amarus* is sometimes also cultivated in small plots and sold on local markets. Fruits of *P. emblica* are gathered from wild and garden trees and used for local consumption or sold on the market. In India, preserves made from these fruits are manufactured and marketed on a large scale.

**Properties** The latex of many members of the *Euphorbiaceae* is known for its toxicity; it causes inflammations of the skin and mucous membranes, conjunctivitis and sometimes even blindness. The latex of *Phyllanthus* can also cause allergic reactions. Several *Phyllanthus* species are poisonous to livestock or fish and can be allelopathic to desirable forage plants. Various groups of medicinally interesting compounds are present in *Phyllanthus*, including triterpenoids, flavonoids, tannins, alkaloids and phenolic acids. Triterpenoids have been isolated from *P. acidus* (phyllanthol,  $\beta$ -amyirin), *P. discoides* Muell. Arg. (betulinic acid), *P. emblica* (lupeol, lupenone), *P. reticulatus* (friedelin,  $\beta$ -sitosterol, betulinic acid, glochidonol, friedelan-3- $\beta$ -ol, 21 $\alpha$ -hydroxyfriedelan-3-one, 21 $\alpha$ -hydroxyfriedel-4(23)-en-3-one) and *P. urinaria* ( $\beta$ -amyirin,  $\beta$ -sitosterol, triacontanol). Flavonoids have been found in *P. amarus* (quercetin-3-O-glucoside, rutin), *P. emblica* (quercetin, kaempferol, kaempferol-3-O-glucoside (= astragalinalin)) and *P. urinaria* (quercetin, kaempferol, rutin). Tannins have been isolated from *P. amarus* (mainly geraniin, amariin and galocatechin) and *P. emblica* (e.g. phyllembin, gallotannin as 1,2,3-trigalloylglucose and the ellagitannins terchebin, corilagin, chebulagic acid and chebulinic acid). Alkaloids, mainly of the quinolizidine type, have been reported for *P. amarus* (phyllantine, securinine, norsecurinine, isobubbialine, epibubbialine) and *P. discoides* (allosecurine, phyllantidine, phyllantine). The quinolizidine alkaloid phyllantine (= methoxy-securinine) must not be confused with phyllanthin and hypophyllanthin, which are lignans isolated from e.g. *P. amarus* and *P. urinaria*. Furthermore, a range of phenolic acids has been

isolated from *P. acidus*, *P. emblica*, *P. maderaspatensis*, *P. reticulatus* and *P. simplex*.

*P. amarus*, *P. debilis* and *P. urinaria* have all been found to inhibit DNA polymerase of the hepatitis B virus and other hepatitis-DNA-viruses, such as the woodchuck hepatitis virus (WHV). Lesser known species of the same subgenus *Phyllanthus* also showed this effect, but species from other subgenera generally show much weaker inhibitory activity. However, intraspecific differences make it difficult to compare the activity of the different species. In some other studies, *P. urinaria* extracts showed inhibitive effects on duck hepatitis B virus (DHBV) polymerase, whereas *P. amarus* and *P. maderaspatensis* extracts were found to lack antiviral activity against DHBV.

*P. amarus* possesses in vitro and possibly in vivo activity against hepatitis B virus (HBV) and related viruses, together with in vitro activity against the enzyme reverse-transcriptase of retroviruses and possibly in vivo activity against retroviruses. The efficacy of *P. amarus* preparations in treating hepatitis B carriers is unclear: both success and failure have been reported.

*P. amarus* has also shown antibacterial and antifungal activity. Whole plant extracts of *P. amarus* reduce digestive tract motility, delay gastric emptying in rats, and cause relaxation of isolated rat fundus and ileum. This confirms the use of *P. amarus* in the treatment of diarrhoea and other gastro-intestinal disorders in Nigeria. Whole plant extracts of *P. amarus* also have diuretic, hypotensive and hypoglycaemic effects in humans. An alcohol extract of *P. amarus* has been found to induce declining fertility in male mice.

*P. amarus* contains the lignans phyllanthin and hypophyllanthin, which have shown endothelin antagonistic effects. *P. debilis* and *P. urinaria* also contain phyllanthin. Furthermore, phyllanthin and hypophyllanthin have been reported to have protective activity in rat hepatocytes against cytotoxicity induced by carbon tetrachloride and galactosamine, and it has been suggested that phyllanthin is responsible for antigenotoxic effects reported for *P. amarus* extracts. But phyllanthin has also been reported to be toxic to the nervous system and liver.

A range of hydrolysable tannins isolated from *P. amarus* has shown inhibitory activity on signal-regulated protein kinases. Aqueous extracts of *P. debilis* leaves have shown immunomodulatory activity, whereas leaf decoctions have a high potassium content, producing diuretic effects.

The triterpenoid triacontanol isolated from *P. uri-*



*itaria* has hepatoprotective properties, e.g. against galactosamine-induced cytotoxicity in rat hepatocytes. This may, at least partly, confirm the hepatoprotective action shown by alcohol extracts of *P. urinaria* in rats. Furthermore, hydroalcoholic extracts of *P. urinaria* have shown analgesic effects against formalin-induced and capsaicin-induced pain in mice (antinociceptive activity), and methanol extracts have demonstrated hypoglycaemic activity in diabetic rats. *P. urinaria* plant extracts exhibited some antibiotic activity against *Staphylococcus*, *Escherichia coli*, *Salmonella typhi*, *Vibrio cholerae* and *Shigella dysenteriae* bacteria.

The fruit of *P. acidus* is very acidic, and, similar to lemon or grapefruit, contains 40 mg/100 g ascorbic acid (vitamin C). The root-bark of *P. acidus* contains saponins, gallic acid and tannins.

*P. emblica* fruits are also highly acidic and contain much vitamin C, with reported contents ranging from 470–1810 mg/100 g. The fruits also contain trigalloylglucose, ellagic acid, corilagin, terchebin, phyllembin, phyllembic acid and emblicol. They have strong antibacterial, antifungal and antioxidant properties. Aqueous extracts of *P. emblica* fruit have been found to antagonize the toxic effects of Cs<sup>+</sup>, Zn<sup>2+</sup> and metanil yellow similarly to the equivalent amount of vitamin C, but to be more effective against the effects of ethylparathion, Pb<sup>2+</sup>, Al<sup>3+</sup> and Ni<sup>2+</sup> than the equivalent amount of vitamin C alone. These results imply that vitamin C is an important active compound of *P. emblica* fruits, but that they contain other active compounds as well. Gallic acid and vitamin C both have antioxidant, antibacterial and chelating properties, whereas tannins protect vitamin C from oxidation. The antioxidant activity of *P. emblica* fruits may also be due to the presence of hydrolysable tannins itself, such as emblicanin A and B, punigluconin and pedunculagin.

Phyllembin has been reported to potentiate the action of adrenaline, to have a mild depressant action on the central nervous system and to have spasmolytic properties. The administration of fruit extracts to mice reduced the cytotoxic effects of the carcinogen 3,4-benzo(a)pyrene and aqueous fruit extracts provided protection against chromosome damage in *Allium sativum* L. root tips induced by X-radiation: these effects were attributed to the antioxidation system of vitamin C, gallic acid, reducing sugars and tannins. Powdered *P. emblica* fruits have been found to enhance natural killer cell activity and antibody-dependent cellular toxicity in mice with Dalton's lymphoma as-

cites tumour. The fruit juice reduced cholesterol levels in humans and in rabbits fed cholesterol.

Further research has shown that methanol extracts of the fruits have a potent inhibitory activity on Human Immunodeficiency Virus (HIV) reverse transcriptase, which may be exploited in the prophylaxis and intervention of AIDS. The most active compound isolated from the methanol extracts was putranjivain A, whereas 1,6-di-O-galloyl- $\beta$ -D-glucose and digallic acid showed weak inhibitory activity. An alcoholic extract of *P. emblica* fruits, and quercetin isolated from the extract have shown in vivo hepatoprotective activity in rats and mice. Seeds of *P. emblica* contain about 16% oil, with linoleic acid (44%), oleic acid (28.4%), linolenic acid (8.8%), stearic acid (2.2%), palmitic acid (3.0%) and myristic acid (1.0%).

Leaves of *P. emblica* contain ellagic acid, kaempferol, kaempferol-3-glycoside and amlaic acid, stems and leaves lupeol,  $\beta$ -sitosterol and ellagic acid, and the bark lupeol and (+)-leucodelphinidin. Various *P. emblica* leaf extracts have shown inhibitory activity on human polymorphonuclear leukocytes and platelets, which at least partly confirms the anti-inflammatory and antipyretic properties of *P. emblica* leaves. A large part of the medicinal uses of *P. emblica* is related to the astringent action of tannins. Though short-term effects may be beneficial, the frequent systemic use of tannins might be dangerous, because of their antinutrient effects.

Finally, extracts of *P. reticulatus*, especially aqueous extracts of the leaves, have shown in vitro activity against *Plasmodium falciparum*.

**Description** Monoecious or dioecious herbs, shrubs or trees of various habit; branching either unspecialized (the phyllotaxy spiral or distichous) or phyllanthoid, i.e. the spiralled leaves on the main axes reduced to cataphylls which subtend a deciduous branchlet with distichous leaves, the latter resembling a compound leaf; indumentum present or absent, simple or rarely dendritic. Leaves alternate, simple, entire, shortly petiolate; stipules present, those of cataphylls larger than those of foliage leaves. Inflorescence axillary, composed of a solitary flower or of a vestigial, bisexual or unisexual cyme, these cymes occasionally aggregated into thyrses. Flowers unisexual, small, apetalous; calyx 4–6-lobed, the lobes imbricate in bud; disk nearly always present. Male flowers with 2–6(–15) stamens, filaments free or connate, anthers free or connate (and forming a 'synandrium'), extrorse; pistillode absent. Female flowers usually without staminodes; ovary superior, ses-

sile or stipitate, 3(–12)-locular with 2 ovules in each cell, styles 3 or rarely more, free or connate at base, bifid, multifid, or dilated into an entire or lacerate stigma. Fruit usually a dehiscent capsule, less commonly baccate or drupaceous. Seeds usually triquetrous, smooth or variously ornamented, testa usually thinly crustaceous; endosperm present. Seedling with epigeal germination; cotyledons emergent, leafy, persistent; hypocotyl elongated, often first curved in a loop above the soil; all leaves alternate, sometimes the first few scale-like, in species showing phyllanthoid branching a few normal leaves are generally produced, followed by cataphylls.

**Growth and development** *Phyllanthus* seedlings are of the *Macaranga* seedling type and develop according to Cook's architectural model, which is determined by continuous growth of a monopodial trunk with phyllomorphic branches. Two types of shoots are produced: indeterminate and determinate. The indeterminate shoots provide annual extension growth, but do not flower and do not abscise. The determinate shoots are often short, bear flowers, and abscise.

In Java, all medicinal *Phyllanthus* species flower throughout the year, except for *P. emblica*, which flowers in August. In India, the fruits of *P. emblica* remain dormant throughout the summer and begin to develop as the monsoon starts.

The seeds of the often explosively dehiscing fruits are ejected and thus dispersed over some distance. Under experimental conditions in Florida *P. amarus* reached maximum height at 5–7 months after sowing, and senescence began after 7 months. *P. acidus* trees start producing a substantial crop after 4 years. *P. emblica* seedlings start bearing after about 8 years, whereas budded trees take about 6 years.

**Other botanical information** *Phyllanthus* belongs to the subfamily *Phyllanthoideae* and is related to *Breynia*, *Glochidion*, *Phyllanthodendron* and *Sauropus* which all show phyllanthoid branching. The genera *Cicca* and *Emblia* are incorporated in *Phyllanthus*, although occasionally still treated as being distinct. *Phyllanthus* has been subdivided into 10 subgenera which are sometimes further divided into a number of sections.

The species *P. simplex* and *P. virgatus* P. Forst. are sometimes regarded as a single species, *P. virgatus* sensu lato, but the mainland populations (*P. simplex*) appear separable from those in the Pacific (*P. virgatus* sensu stricto) although there are records of intermediate specimens. Another taxo-

nomic problem concerns *P. niruri* L. Asiatic specimens formerly assigned to this species actually belong to either *P. amarus*, *P. debilis* or *P. fraternus* Webster, whereas true *P. niruri* is restricted to the West Indies. Much of the research performed with '*P. niruri*' in Asia, mainly in India, thus needs botanical correction.

**Ecology** *P. amarus* (often reported as *P. niruri*) is a troublesome weed in pulses, soya bean, groundnut, cereals, sugar cane, cassava, taro, sesame, sunflower, and cotton. Less important weeds are *P. debilis*, *P. maderaspatensis*, *P. simplex* and *P. urinaria*, outside South-East Asia also *P. fraternus*. The species belonging to the subgenera *Phyllanthus* (e.g. *P. amarus*, *P. debilis*, *P. emblica*, *P. urinaria*) and *Isocladus* (e.g. *P. maderaspatensis*) show a marked preference for calcareous sites in humid tropical areas. In China, warm, well-drained sandy soils and fertilization with N and K are recommended for cultivated *P. urinaria*. *P. urinaria* is more drought-sensitive than *P. amarus* and *P. debilis* and is generally found in wetter sites. *P. amarus* grown in greenhouses at a temperature of 15°C showed much less inhibitory activity on the DNA polymerase of woodchuck hepatitis virus (WHV) than when grown at 25°C. *P. emblica* is a short-day plant, requiring photoperiods of 12–13.5 hours for flowering. It can be grown in both light and heavy soils, but it prefers well-drained, fertile loamy soils. Seeds of the herbaceous *P. debilis*, *P. simplex* and *P. urinaria* are dispersed by water and animals.

**Propagation and planting** Seed of *P. amarus* and *P. urinaria* requires light to germinate. *P. urinaria* seed showed adequate germination at temperatures of 25–35°C, but germination was poor at 20°C or 40°C. Germination of *P. urinaria* seed is poor under moisture stress conditions too (osmotic potential higher than 300 kPa). A 1000-seed weight of 150 mg has been reported for *P. amarus* seed harvested in Puerto Rico. There is some speculation that seed production in *P. amarus* may be parthenocarpic. *P. acidus* is generally grown from seed, but vegetative propagation such as budding, greenwood cuttings or air layering can also be used, whereas *P. emblica* may also be propagated through seed and vegetative methods (budding, grafting, cutting, root sprouting). Inarching is also possible, but gives only limited success. *P. emblica* has been successfully propagated in vitro. In India, *P. emblica* is often grown at spacings of 9–12 m × 9–12 m, *P. acidus* in Indonesia at 8 m × 8 m.

**In vitro production of active compounds** Callus can be induced from stem or phyllanthoid

branch pieces of *P. amarus* and *P. urinaria*, but callus extracts showed less activity against viral DNA polymerase and reverse transcriptase than extracts from field-grown plants. Phyllembin present in extracts of in vitro *P. emblica* tissue cultures showed antimicrobial activity against bacteria (*Escherichia coli*, *Staphylococcus aureus*, *Salmonella typhosa*) and a fungus (*Candida albicans*).

**Husbandry** Most material used for medicinal purposes is collected from the wild. Though *P. amarus* is also cultivated in India, there is no information on any specific cultivation measures. Soil fertility and soil moisture experiments in the United States affected morphology and yield of *P. debilis* and *P. urinaria*, with plants being shorter, more branched and with a higher dry weight under favourable conditions. Differences in soil fertility and soil moisture generally did not affect these species' in vitro inhibitory activity on WHV viral DNA polymerase. In *P. amarus*, maximum dry weight is obtained in wet neutral soils, but differences in soil moisture, pH and Ca content did not affect the inhibitory activity of plant extracts. These findings imply that plant yields can be maximized through cultivation measures, without affecting the antiviral activity of plant material.

Where *P. urinaria* is not desired, it may be controlled by mulching. Young plantations of *P. emblica* need regular weeding. Established *P. emblica* trees do not need frequent irrigation, which makes them well suited for drier regions. However, the crop benefits from 2-3 irrigations at full bloom stage and fruit set. Irrigation is not beneficial during fruit dormancy. *P. emblica* does not need regular pruning, but pruning in the early years promotes the development of a proper shape and a strong frame.

**Diseases and pests** In India, *P. simplex* is infected by mildew (*Erysiphe cichoracearum*), which appears as small, circular, whitish powdery spots on leaves, petioles and stems.

Caterpillars of *Parallelia absentimacula* and *P. joviana* feed on *Phyllanthus acidus* in Indonesia. The main pest of *P. emblica* in India is the bark-eating caterpillar *Indarbela* sp., which tunnels into the trunk and branches. *P. urinaria* weed in Indian rice fields is infested with the rice root-knot nematode (*Meloidogyne graminicola*).

**Yield** Since most *Phyllanthus* products are collected from the wild, yield data are scarce. Under experimental conditions in Florida, 6-7 months old *P. amarus* reached an average dry weight of about 40 g/plant when harvested in July or Au-

gust. Annual fruit yields of *P. emblica* vary considerably, depending on cultivar, plant age and management: from 15 kg per tree for wild trees to 25-200 kg per tree for some cultivars.

**Handling after harvest** Sun-drying of *P. emblica* fruits leads to loss of vitamin C, but this is not the case when fruits are vacuum-dried. Once dried, the vitamin C is very stable, even when stored for prolonged periods; this has been attributed to tannins and polyphenols retarding oxidation. Vitamin-rich syrups and concentrates have been prepared from *P. emblica* fruits in India.

**Genetic resources and breeding** There is rich genetic diversity in cultivated and wild relatives of *P. emblica* in India, but genetic erosion is severe because of deforestation and the use of only a few popular cultivars. Germplasm survey, collection and evaluation work has been done and accessions are maintained at Narendra Dava University of Agriculture and Technology in Faizabad (India). Other accessions are reported to be kept in Havana (Cuba), Uttar Pradesh (India), Udaipur (India), Bangkok (Thailand) and Miami (United States). No breeding has been done on other *Phyllanthus*, except for *P. emblica*.

**Prospects** *Phyllanthus* seems under-exploited. A considerable number of its species have been used against jaundice for a long time. More research is needed to ascertain its activity against the hepatitis virus and other viruses and to obtain the necessary information on possible side-effects. *Phyllanthus* has potential beneficial therapeutic action in the management of hepatitis B, nephrolithiasis and in painful disorders (as antinociceptive agent). However, well-controlled, double-blind clinical trials are lacking. The possible anti-AIDS activity deserves further research. Some *Phyllanthus* also produce edible fruits, others can be used for tanning and dyeing. The fruits of *P. emblica* are somewhat under-valued. Not only are they highly nutritious and contain much vitamin C, but they also have many medicinal properties. The ease of long-distance transportation and the stability of the vitamin C make *P. emblica* interesting for commercial exploitation.

**Literature** [1] Calixto, J.B., Santos, A.R., Cechinel-Filho, V. & Yunes, R.A., 1998. A review of the plants of the genus *Phyllanthus*: their chemistry, pharmacology, and therapeutic potential. *Medicinal Research Reviews* 18(4): 225-258. [2] Doshi, J.C., Vaidya, A.B., Antarkar, D.S., Deolalikar, R. & Antani, D.H., 1994. A two-stage clinical trial of *Phyllanthus amarus* in hepatitis B carriers: failure to eradicate the surface antigen. In-

dian Journal of Gastroenterology 13(1): 7-8. |3| Hussain, R.A., Dickey, J.K., Rosser, M.P., Matson, J.A., Kozlowski, M.R., Brittain, R.J., Webb, M.L., Rose, P.M. & Fernandes, P., 1995. A novel class of non-peptidic endothelin antagonists isolated from the medicinal herb *Phyllanthus niruri*. Journal of Natural Products 58(10): 1515-1520. |4| Ihantola-Vormisto, A., Summanen, J., Kankaanranta, H., Vuorela, H., Asmawi, Z.M. & Moilanen, E., 1997. Anti-inflammatory activity of extracts from leaves of *Phyllanthus emblica*. Planta Medica 63(6): 518-524. |5| Mitra, R.L. & Jain, S.K., 1985. Concept of *Phyllanthus niruri* (Euphorbiaceae) in Indian floras. Bulletin of the Botanical Survey of India 27(1-4): 161-176. |6| Ott, M., Thyagarajan, S.P. & Gupta, S., 1997. *Phyllanthus amarus* suppresses hepatitis B virus by interrupting interactions between HBV enhancer I and cellular transcription factors. European Journal of Clinical Investigation 27(11): 908-915. |7| Rizk, A.M., 1987. The chemical constituents and economic plants of the Euphorbiaceae. Botanical Journal of the Linnean Society 94: 293-326. |8| Suresh, K. & Vasudevan, D.M., 1994. Augmentation of murine natural killer cell and antibody dependent cellular cytotoxicity activities by *Phyllanthus emblica*, a new immunomodulator. Journal of Ethnopharmacology 44(1): 55-60. |9| Unander, D.W., 1991. Callus induction in *Phyllanthus* species and inhibition of viral DNA polymerase and reverse transcriptase by callus extracts. Plant Cell Reports 10: 461-466. |10| Unander, D.W., Bryan, H.H., Lance, C.J. & McMillan Jr., R.T., 1995. Factors affecting germination and stand establishment of *Phyllanthus amarus* (Euphorbiaceae). Economic Botany 49(1): 49-55.

#### *Selection of species*

#### ***Phyllanthus acidus* (L.) Skeels**

U.S. Dept. Agric. Bur. Pl. Industry Bull. 148: 17 (1909).

**Synonyms** *Phyllanthus acidissimus* (Blanco) Muell. Arg. (1863), *Phyllanthus distichus* (L.) Muell. Arg. (1866), *Cicca acida* (L.) Merr. (1917).

**Vernacular names** Otaheite gooseberry, Malay gooseberry, country gooseberry (En). Cerisier de Tahiti (Fr). Indonesia: ceremai, cerme (general), caramele (southern Sulawesi). Malaysia: chermay, chermala, kemangur. Philippines: iba (Tagalog), bangkiling (Bisaya, Tagalog), karmay (Ilokano). Burma (Myanmar): thinbozihpyoo. Cambodia: kântûtët, kântouot srôk. Laos: nhôm

baanz, mak nhom, nhom ban<sup>2</sup>. Thailand: ma yom (general). Vietnam: ch[uf]m ru[ooj]t, t[aa]f]m ru[ooj]t.

**Distribution** *P. acidus* is probably native to the coastal region of north-eastern Brazil, but since time immemorial it has been cultivated, mainly as a fruit tree, in tropical Asia from India to Malesia and Polynesia, and on all larger islands of the West Indies. Within Malesia it has not yet been reported from New Guinea.

**Uses** The latex is credited with emetic and purgative activity. In Indonesia, the bark is heated with coconut oil and spread on eruptions on feet and hands. In Java, an infusion of the root is taken to alleviate asthma. In Borneo, roots are used in the treatment of psoriasis of the feet. Although the roots are weakly poisonous, in Malaysia they used to be boiled and the vapour inhaled to relieve cough and headache. In the Philippines, leaf decoctions are applied to urticaria, and a decoction of the bark is used to treat bronchial catarrh. In Burma (Myanmar), the fruit is used as a laxative. In India, the fruits are taken as a liver tonic to enrich the blood. The juice of the root bark is reported to have been employed in criminal poisonings. The fruit flesh is added to many dishes in Indonesia as a flavouring. In the Philippines, the fruit juice is used to make cold drinks and the fruit to make vinegar. In Malaysia, ripe and unripe fruits are served as a relish, syrup or sweet preserve. The fruits are also combined with other fruits in making chutney or jam, because of their setting properties. Young leaves are cooked as a vegetable in Indonesia, Thailand and India. The wood is fairly hard, strong, tough and durable if seasoned. The bark has limited use in India as a tanning agent.

**Observations** A monoecious, small, glabrous tree up to 10 m tall with phyllanthoid branching, bark rough, grey, with prominent lenticels; cataphylls not persistent, blackish-brown, their stipules triangular-ovate; deciduous branchlets ascending, (20-)25-52 cm long, with 25-40 leaves; leaves broadly ovate to ovate-lanceolate, (4-)5-9 cm × (2-)2.5-4.5 cm, base obtuse to rounded, apex acute, petiole 2.5-4 mm long, stipules triangular-acuminate; flowers in dense, cushion-shaped cymes at the nodes of leafless branchlets on older wood, and usually also on proximal branchlets of current year's growth, pale green to reddish; male flowers 4-merous, filaments and anthers free, dehiscent vertically; female flowers on a stout pedicel, 4-merous, disk deeply lobed or split, styles connate, deeply bifid, staminodes present; fruit

drupaceous, obovate, 1–1.5 cm × (1.2–)1.5–2(–2.5) cm when fresh, shallowly 6- or 8-lobed, greenish-yellow to creamy-white; seeds smooth. In north-eastern Brazil *P. acidus* has been found in coastal forest. In South-East Asia it is cultivated on humid sites, up to 1000 m altitude.

**Selected sources** 97, 190, 202, 287, 580, 641, 697, 900, 980, 1035, 1126, 1128, 1178, 1211, 1212, 1476, 1521, 1555.

**Phyllanthus amarus Schum.**

Kongel. Danske Vidensk. Selsk. Skr., Naturvidensk. Math. Afh. 4: 195 (1829).

**Synonyms** *Phyllanthus swartzii* Kostel. (1836), *Phyllanthus nanus* Hook.f. (1887), *Phyllanthus niruri* auct. non L.

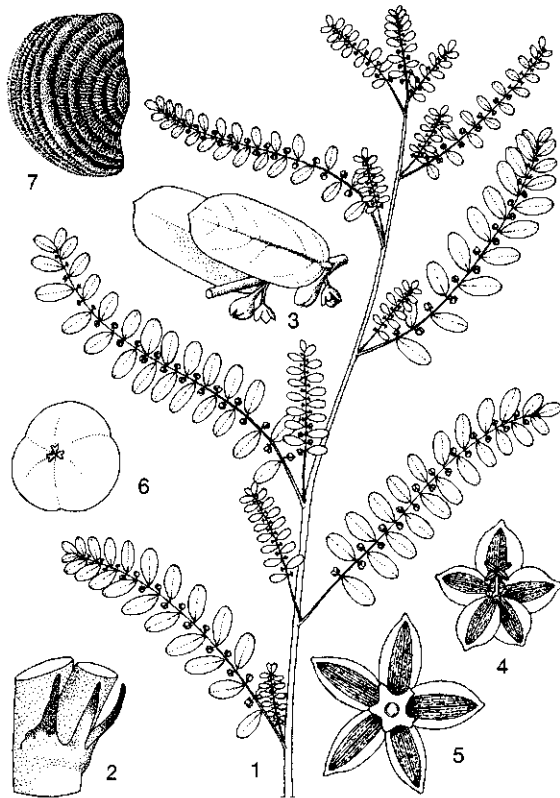
**Vernacular names** Herbe au chagrin (Fr). Indonesia: memeniran (Sundanese), meniran (Javanese), gosau ma dungi (Ternate). Malaysia: dukong anak, dukong-dukong anak, rami buah (Peninsular). Papua New Guinea: manjinimbi (Kanganaman, Sepik). Philippines: kurukalungai (Bikol), sampasampalukan (Tagalog), san pedro (Bisaya). Cambodia: préak phlè (Battambang). Thailand: luuk tai bai (central), ma khaam pom din (northern), yaa tai bai (general). Vietnam: c[aa]y ch[os] d[er], di[ee]p h[aj] ch[aa]u.

**Distribution** *P. amarus* is native to the Americas, but now a pantropical weed. It is found throughout Malesia, though not yet reported from Sulawesi.

**Uses** From Hainan to Indonesia a decoction or tea is drunk as a diuretic to treat kidney and liver trouble, colic and venereal diseases. It is credited with expectorant (children's coughs), febrifuge, emmenagogue and antidiarrhetic properties. The pounded plants are applied externally on contusions and skin complaints. A decoction of the whole plant is used as a stomach tonic. In Papua New Guinea, a cooled tisane of the whole plant is used to treat headache or migraine. In India, leaves and fruits of *P. amarus* are ground into a paste with buttermilk, garlic and peppers, and given orally for seven days to treat jaundice. According to Ayurvedic medicine *P. amarus* has astringent, deobstruent and antiseptic properties, and is furthermore used to treat dyspepsia, dysentery, dropsy, diseases of the urogenital system, gonorrhoea and diabetes. In the form of a poultice with rice water, *P. amarus* is used on oedematous swellings and ulcers. South-American uses include the treatment of malaria, kidney and bladder stones and urinary disorders in general, whereas the plant is also reported to induce abor-

tion. Traditional healers in Tanga (north-eastern Tanzania) use an aqueous extract of aerial parts of *P. amarus* for the management of diabetes mellitus that is not insulin-dependent. In Nigeria, an aqueous extract of dried plant material is used against diarrhoea. Leaves are chewed against persistent coughs and used to soothe stomach-ache. In the West Indies, *P. amarus* is applied to treat childhood intestinal worms and on the Cook Islands (Rarotonga) to treat sarache.

**Observations** A monoecious, annual, erect, glabrous herb up to 60 cm tall with phyllanthoid branching; cataphylls subulate, with triangular stipules often turning black; deciduous branchlets 4–12 cm long, with about 15–30 leaves; leaves sessile, elliptical-oblong, 5–11 mm × 3–6 mm, obtuse to rounded at base, obtuse or rounded and often apiculate at apex, stipules ovate-lanceolate to lanceolate; proximal (1–)2 axils of deciduous branchlets with cymules of (1–)2 male flowers, all



*Phyllanthus amarus* Schum. - 1, plant habit; 2, cataphyll and stipules; 3, leaves and inflorescences; 4, male flower; 5, female flower; 6, fruit; 7, seed.

succeeding axils consisting of 1 male and 1 female flower; flowers pale green, often flushed with red, with 5(–6) calyx lobes with scarious margins; male flowers with 5 disk segments, stamens (2–)3, filaments completely connate, anthers free, dehiscing obliquely to horizontally; female flowers shortly pedicellate, disk deeply 5-lobed to lacinate, styles free, very shallowly bifid; fruit an oblate capsule, 1.8–2.5 mm in diameter, obtusely trigonous, smooth; seeds longitudinally ribbed on the back. *P. amarus* is the commonest species of the genus and occurs as a weed of open ground, waste places, grassy scrub and dry deciduous forest, usually on humid, sandy soils, up to 1000 m altitude.

**Selected sources** 97, 192, 202, 225, 580, 597, 838, 949, 982, 1035, 1126, 1128, 1135, 1178, 1193, 1380, 1555, 1556.

### **Phyllanthus debilis Klein ex Willd.**

Sp. pl. 4: 582 (1805).

**Synonyms** *Phyllanthus niruri* L. var. *debilis* (Klein ex Willd.) Muell. Arg. (1863), *Phyllanthus niruri* L. var. *javanicus* Muell. Arg. (1863), *Phyllanthus niruri* auct. non L.

**Vernacular names** Lagoon spurge (En). Indonesia: meniran (Javanese), memeniran (Sundanese). Vietnam: di[ee]p h[a]j ch[aa]u y[ees]u.

**Distribution** *P. debilis* is probably native to southern India and Sri Lanka; introduced into Vietnam, Thailand, Peninsular Malaysia, Singapore, Indonesia (throughout), Papua New Guinea, the Pacific Islands, and the West Indies.

**Uses** The uses of *P. debilis* are similar to those of *P. amarus*, with which it may have often been confused.

**Observations** A monoecious, annual, erect, glabrous herb up to 100 cm tall with phyllanthoid branching; cataphylls narrowly lanceolate with triangular-lanceolate to triangular stipules; deciduous branchlets 4–10(–12) cm long, with 15–35 leaves; leaves sessile, narrowly elliptical to narrowly elliptical-ovate, (5–)8–20 mm × 2.5–5 mm, cuneate at base, acute at apex, stipules lanceolate, acuminate; proximal 1–4 axils of deciduous branchlets with cymules of 2–4 male flowers, distal axils with solitary female flowers, sometimes accompanied by a few male ones; male flowers with 6 calyx lobes, disk with 6 segments, stamens 3, filaments completely connate, anthers free or partially connate, dehiscing horizontally; female flowers pedicellate, with 6 calyx lobes with wide scarious margin, disk entire to shallowly 6-lobed, styles free, appressed to the ovary, bifid to

about the middle; fruit an oblate capsule about 2–2.5 mm in diameter, smooth; seeds radially and tangentially with longitudinal ribs and fine striae on the back. *P. debilis* is locally an abundant weed of ruderal places, gardens, grassland and upland rice fields, preferring fertile soils, up to 2000 m altitude.

**Selected sources** 33, 102, 192, 1135, 1386, 1555, 1556.

### **Phyllanthus emblica L.**

Sp. pl. 2: 982 (1753).

**Synonyms** *Emblica grandis* Gaertner (1790), *Emblica officinalis* Gaertner (1790), *Emblica arborea* Raf. (1838).

**Vernacular names** Aonla, emblic myrobalan, Indian gooseberry (En). Embligue officinale, grosseillier de Ceylan, myrobalan emblic (Fr). Indonesia: malaka (general), kemloko (Javanese), ki malaka (Sundanese). Malaysia: laka, melaka (Peninsular). Philippines: nelli. Burma (Myanmar): ta-sha-pen. Cambodia: karn lam, kam lam ko, kântût préi. Laos: khaam poomz, mak kham pom. Thailand: ma-khaam pom (general), kan-tot (south-eastern), kam thuat (south-western). Vietnam: me r[uw]f[ng], chu me (northern), b[oo]ng ng[os]t (southern).

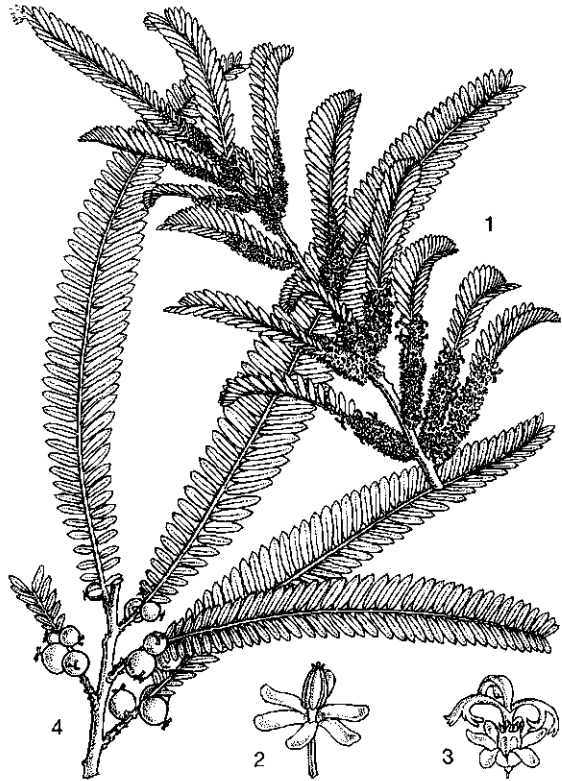
**Distribution** *P. emblica* is native to tropical South-East Asia, from India, Nepal and Pakistan to Burma (Myanmar), Indo-China, southern China, Thailand, Peninsular Malaysia, Sumatra, Borneo, Java, the Lesser Sunda Islands and the Moluccas. It is widely cultivated for its fruits throughout its natural area of distribution, in India since time immemorial, and also in the West Indies, Japan, the Mascarene Islands and Sri Lanka, where it is doubtfully native. For several centuries only its fruits were known; they were used medicinally. In 1901 seeds of *P. emblica* were distributed to early settlers in Florida and to public gardens and experimental stations in e.g. the West Indies, Hawaii and the Philippines. In 1982 seeds were sent to Australia.

**Uses** All plant parts are applied medicinally. A decoction of the dried fruits is used in Indonesia to treat bloody diarrhoea. Fruit pulp is smeared on the head to dispel headache and dizziness, caused by excessive heat and fever. In Burma (Myanmar) and Thailand, the fruit juice is used as a laxative and for the treatment of inflamed eyes. In Indo-China, fruits are used to treat diarrhoea, and fruit juice is administered to treat colic and other abdominal disorders. In India, the fruit is valued as an antiscorbutic, refrigerant, diuretic, laxative

and antibiotic, and considered useful in the treatment of haemorrhages, diarrhoea, dysentery, anaemia, jaundice, dyspepsia, diabetes, fever, bronchitis and cough. It is used as an ingredient for several medicines of the indigenous Ayurvedic system. Leaf decoctions are used in Malaysia as a febrifuge, and in Thailand for skin diseases. Seeds are applied in India against asthma, bronchitis, and biliousness, whereas flowers are credited with refrigerant and aperient properties. The bark is used in India for the treatment of diarrhoea or as a stomachic for elephants. The root bark is used in Burma (Myanmar) as an astringent and in India, mixed with honey, to treat inflammation in the mouth. In Malaysia, Thailand and to a greater extent India the fruits are a delicacy, eaten raw or preserved. The Akha in northern Thailand use the fruit as a masticatory and to blacken the teeth. Dried fruits are sometimes used as a shampoo. Fruits, leaves and bark are used for tanning and dyeing. Leaves and fruits are used for animal fodder, whereas leaves can also be applied as green manure. Although the wood may warp and split, it is used for the construction of furniture and implements; it is very durable when submerged. Finally, the wood is suitable as firewood and produces charcoal of good quality.

**Observations** A monoecious, small to medium-sized tree up to 15(–25) m tall with phyllanthoid branching, bole often crooked and gnarled, up to 35 cm in diameter, bark thin, grey, smooth, flaking; cataphylls inconspicuous, scarious, their stipules triangular-ovate; deciduous branchlets (5–)10–25(–30) cm long, with (15–)30–100(–150) leaves; leaves subsessile, narrowly oblong, 12–20 mm × 2–5 mm, slightly oblique and subcordate at base, margin and tip almost inflexed, stipules triangular; proximal axils of deciduous branchlets with reduced leaves and cymules of male flowers followed by cymules of 1–2 female flowers surrounded by several male ones, distal part sterile or rarely reduced; flowers pale green, with 6 calyxlobes; male flowers with 6 disk segments, stamens 3, filaments entirely connate, anthers free, minutely apiculate, dehiscing vertically; female flowers subsessile, with cup-shaped, 6-ribbed disk enclosing the ovary, styles shortly connate, mostly twice bifid; fruit drupaceous, tardily dehiscent, depressed globose, in wild plants 13–25 mm × 23–30 mm, in cultivated ones up to 42 mm in diameter, pale green becoming yellowish-white, smooth; seeds smooth. *P. emblica* is most common in dry, semi-deciduous forest, at elevations up to 1800 m.

**Selected sources** 64, 97, 105, 190, 202, 264,



*Phyllanthus emblica* L. – 1, flowering twig; 2, male flower; 3, female flower; 4, fruiting twig.

284, 287, 580, 842, 900, 980, 1126, 1128, 1135, 1211, 1212, 1257, 1380, 1476, 1555.

***Phyllanthus maderaspatensis* L.**

Sp. pl. 2: 982 (1753).

**Synonyms** *Phyllanthus venosus* A. Rich. (1851), *Phyllanthus vaccinioides* Klotzsch (1861), *Phyllanthus gueinzii* Muell. Arg. (1863).

**Distribution** Tropical Africa to India and Sri Lanka, East Java and Australia.

**Uses** In India, a leaf infusion is used to treat headache, and seeds are credited with laxative, carminative and diuretic properties. Powder from dried plant material mixed with cow milk is given orally for eight days to treat jaundice.

**Observations** A monoecious, annual or perennial, erect to spreading, unbranched to much branched, glabrous herb up to 90(–120) cm tall with unspecialized branching; leaves arranged spirally, linear to oblanceolate, (5–)10–30(–60) mm × (1–)2–7(–17) mm, cuneate to broadly cuneate at base, acute to rounded at apex, on a petiole about 1 mm long, with ovate-lanceolate stip-

ules; proximal axils of branches with solitary female flowers, distal ones with 1–4 male flowers and a single female one; male flowers with 6 calyx lobes, yellowish-green or whitish, disk segments 6, stamens 3, filaments partly united, anthers free, vertically dehiscent; female flowers pedicellate, with 6 calyx lobes, dark green, sometimes flushed with red or pink, margins white, disk with 6 free segments, styles free, shortly bifid; fruit an oblate capsule, about 3 mm in diameter, smooth; seeds with longitudinal rows of tubercles on the back. *P. maderaspatensis* is found in deciduous woodland, wooded savanna, beaches, dunes, also along streams and ponds, in cultivated and disturbed places, on a wide variety of soils, up to 1850 m altitude.

**Selected sources** 92, 97, 287, 1135, 1187, 1193.

### ***Phyllanthus oxyphyllus* Miq.**

Fl. Ind. Bat., Suppl.: 179, 448 (1860).

**Synonyms** *Phyllanthus frondosus* Wallich ex Muell. Arg. (1863), *Phyllanthus hasskarlianus* Muell. Arg. (1863), *Phyllanthus kunstleri* Hook.f. (1887).

**Vernacular names** Piggyback tree (En). Malaysia: asin-asin, cherek hantu, meroyan puteri (Peninsular). Thailand: yaai chuung laan, yaai theep laan, yom hin (peninsular).

**Distribution** Peninsular Thailand, Peninsular Malaysia and Sumatra.

**Uses** In Peninsular Malaysia, a decoction of the leaves may be applied to cure fever, and is given after childbirth as a protective medicine. It is also a diuretic and diaphoretic in treating gonorrhoea.

**Observations** An evergreen, glabrous shrub or small tree up to 3 m tall with phyllanthoid branching, bole crooked; deciduous branchlets 15–42 cm long; leaves subsessile, oblong to oblong-lanceolate, 2–7 cm × 1–2 cm, cuneate to slightly cordate at base, apex long tapering, stipules minute, narrowly triangular; cymules borne directly in the leaf axils; male flowers in contracted cymules, calyx lobes 5–6, stamens 3, filaments almost connate, anthers apiculate; female flowers subsessile, solitary or accompanied by male ones, calyx lobes 5–6, styles free or connate below, bifid; fruit a subglobose capsule, 3-lobed, about 7 mm in diameter, red. *P. oxyphyllus* is found in evergreen, primary or secondary rain forest, in lowland and mountains up to 1200 m altitude, sometimes on rocky places, rarely on limestone.

**Selected sources** 32, 34, 202, 284, 1126, 1227, 1564.

### ***Phyllanthus pulcher* Wallich ex Muell. Arg.**

Linnaea 32: 49 (1863).

**Synonyms** *Diasperus pulcher* (Wallich ex Muell. Arg.) Kuntze (1891).

**Vernacular names** Malaysia: kelurut tanjong, naga buana, semelit patong. Thailand: kaang plaa (peninsular), trueng baa daan (south-western), waan thoraanee saan (Bangkok). Vietnam: me l[as] l[eej]ch.

**Distribution** *P. pulcher* is found from Burma (Myanmar) and Indo-China towards Thailand, Peninsular Malaysia, Sumatra, Borneo, Java and the Lesser Sunda Islands; cultivated in Sri Lanka, Tanzania and the West Indies; occasionally escaping.

**Uses** In Peninsular Malaysia, a decoction is drunk to relieve stomach-ache, or is used as an eyewash. Poultices are applied to the skin to treat boils, gumboils, swellings and itch, to the nose to treat ulceration, and to the abdomen to treat fever or, in children, kidney trouble. Leaves may be applied to the gums to treat toothache. *P. pulcher* is also cultivated for ornamental purposes.

**Observations** A monoecious, small shrub up to 1.5 m tall with phyllanthoid branching, younger branches with dendritic hairs; cataphylls persistent, with triangular-lanceolate stipules; deciduous branchlets 10–15(–18) cm long, with about 15–30 leaves; leaves oblong to elliptical or elliptical-ovate, 18–28 mm × 8–14 mm, strongly asymmetrical at base, apex abruptly pointed, very shortly petiolate, stipules persistent, triangular-lanceolate; proximal axils of deciduous branchlets with cymules of male flowers, distal ones with solitary female ones; male flowers with 4 calyx lobes, pale green with a dark red base and pale, fimbriate margin, disk segments 4, subpetaloid, stamens 2, filaments connate into a very short column, anthers fused, dehiscent horizontally, rudimentary pistil present; female flowers with 6 calyx lobes similar to those of the male flower but larger, disk cup-shaped and enclosing the base of the ovary, styles free, bifid nearly to the base; fruit a subglobose capsule, smooth, about 3 mm in diameter, pale brown, often not developed. *P. pulcher* may be locally common and is found as an invasive plant in old forest clearings, or as a weed in fruit gardens and along rivers, but also scattered in evergreen forest, up to 700 m altitude.

**Selected sources** 32, 97, 202, 1126, 1135, 1187, 1380, 1555, 1564.



**Phyllanthus reticulatus Poiret**

Lamk, Encycl. 5: 298 (1804).

**Synonyms** *Phyllanthus microcarpus* (Benth.) Muell. Arg. (1863), *Phyllanthus dalbergioides* Wallich ex J.J. Smith (1910), *Phyllanthus erythrocarpus* Ridley (1923).

**Vernacular names** Indonesia: congcong belut, trembilu (Javanese), wawulitan (Sundanese). Malaysia: kayu darah belut, tampal besi.

Philippines: malatinta (Tagalog), matang-buiud (Bikol), sungot-olang (Bisaya). Cambodia: prâ-peénh chhmôól. Laos: am ai<sup>2</sup>, kang<sup>2</sup> pa. Thailand: kaang plaa khrua (general), mat kham (northern), am aai (eastern). Vietnam: ph[ef]n den, c[aa]ly n[oox].

**Distribution** *P. reticulatus* is widespread in the Old World tropics, from tropical Africa to Pakistan, India, Sri Lanka, Burma (Myanmar), Indo-China, southern China, Thailand and throughout the Malesian region towards northern Australia (Queensland); it has been introduced into the West Indies.

**Uses** In Peninsular Malaysia, stems and leaves of what is probably *P. reticulatus* are rubbed onto the chest to alleviate asthma, whereas a decoction of leaves is used to treat a sore throat. In the Philippines, a decoction of the leaves or bark is used for its diuretic, alterative, depurative, refrigerant and odontalgic properties, and the leaves can be applied to the abdomen as a remedy for pinworms. An infusion of the bark is a cure for dysentery and an infusion of the roots for asthma. In Indo-China, *P. reticulatus* is used to treat smallpox and syphilis. In southern Africa, dried and powdered leaves are dusted over wounds to aid the healing process. Ink is prepared from the ripe fruits in the Philippines, whereas in Indonesia a decoction of stems and leaves used to be used to dye cotton black. It is also used as a mordant. In India, the root is reported to produce a red dye. The wood is sometimes used to make utensils.

**Observations** A monoecious, glabrous to pubescent, bushy shrub or small tree up to 5(-18) m tall with disagreeable scent, with phyllanthoid branching, bark rough, brown or grey, cataphylls lanceolate, with triangular stipules; deciduous branchlets steeply ascending, (8.5-)10-20(-25) cm long, with (10-)13-20(-25) leaves; leaves elliptical to elliptical-ovate or elliptical-obovate, 10-50 mm × 5-27 mm, cuneate to rounded at base, apex obtuse to emarginate, shortly petiolate, stipules lanceolate; cymules usually axillary or sometimes on leafless shoots and resembling a raceme, usual-

ly bisexual and composed of 1(-2) female flowers and up to 8 male ones; flowers with 5-6 calyx lobes and 5(-6) disk segments; male flowers with 5-6 stamens, in two sets, one with longer filaments fused into a central column and one with shorter, free filaments, anthers free, dehiscing longitudinally; female flowers on a slender pedicel, styles bifid, free but connivent over the top of the ovary; fruit a berry, globose or oblate, 4-6 mm in diameter, smooth, blueish-black when ripe; seeds nearly smooth. *P. reticulatus* is a variable and weedy species of secondary vegetation, mixed evergreen forest, scrub and hedges, frequently along watercourses, up to 1000 m altitude, in India up to 2000 m.

**Selected sources** 97, 190, 202, 580, 842, 1035, 1126, 1128, 1135, 1178, 1187, 1380, 1476, 1525, 1555, 1564.

**Phyllanthus simplex Retz.**

Observ. bot. 5: 29 (1788).

**Vernacular names** Indonesia: sahakepo (Minahassa, Sulawesi). Philippines: kaya-an, kayut-bulang (Bagobo). Laos: ket 'hoy, 'khi<sup>2</sup> doy<sup>2</sup>. Thailand: khaang amphai (northern), luuk tai bai (central), phaeng kham hoi (eastern). Vietnam: v[aa]rly [oos]c.

**Distribution** *P. simplex* is found from India and Sri Lanka to Indo-China, southern China, Thailand, and throughout the Malesian region.

**Uses** In the Philippines, leaf juice is used as an eyewash for inflamed eyes. In India, *P. simplex* is credited with antiseptic properties. Leaves are crushed and mixed with buttermilk to make a lotion against itching, and root preparations are externally applied to mammary abscesses.

**Observations** A monoecious, annual or perennial, erect to prostrate, glabrous herb up to 50 cm tall with unspecialized branching, branchlets compressed, narrowly wing-angled; leaves distichous on the main stem, narrowly to broadly oblong-lanceolate, 5-32 mm × 2-9 mm, obtuse to rounded at base, apex obtuse, margin often purplish, sessile, stipules broadly ovate; male flowers in axillary glomerules of 2-4, with (5-)6 calyx lobes, disk segments 6, stamens 3, free, anthers dehiscing horizontally; female flowers solitary in leaf axils, long-pedicellate, with 6 calyx lobes, disk shallowly cupular, entire to subentire, styles free, bifid down to the base; fruit a depressed globose capsule, 2.5-3.5 mm in diameter, papillate-verruculose; seeds verruculose. *P. simplex* is a weed of roadsides, grassy places, arable land and upland rice fields, up to 750 m altitude.

**Selected sources** 97, 287, 580, 1126, 1128, 1135, 1178, 1188, 1380, 1386, 1525, 1556.

### **Phyllanthus urinaria L.**

Sp. pl. 2: 982 (1753).

**Synonyms** *Phyllanthus lepidocarpus* Siebold & Zucc. (1843), *Phyllanthus leprocarpus* Wight (1852), *Phyllanthus verrucosus* Elmer (1915).

**Vernacular names** Leaf flower (Am). Kikilé, petit tamarind rouge, urinaire de Malabar (Fr). Indonesia: memeniran (Sundanese), meniran (Javanese), gosau ma dungi roriha (Ternate). Malaysia: amin buah, dukong anak, keman jolok (Peninsular). Philippines: laiolaioan (Bikol), ibai-ba-an (Tagalog), takumtakum (Bisaya). Cambodia: prak phlè. Laos: khao ham, khao ham 'sano khok. Thailand: ma khaam pom din (northern), maak khai lang (north-eastern), yaa tai bai (central, peninsular). Vietnam: ch[os] d[er] r[aw]ng c[uw]a, di[eej]p h[aj] ch[aa]u.

**Distribution** *P. urinaria* is native to the Asian tropics, but was introduced into America and Africa and is nowadays an almost pantropical weed; throughout the Malesian region.

**Uses** *P. urinaria* has the same uses as *P. amarus* in South-East Asia, but *P. amarus* is generally preferred. In Malaysia, the juice is also used to clean a child's tongue and to stimulate a child's appetite. In Papua New Guinea, a decoction is used as a febrifuge. In Brunei, a leaf poultice is applied, with coconut milk, to smallpox. In Cambodia, it is used against malaria. In the Pacific, *P. urinaria* is known as an emmenagogue and abortifacient. In Guam, a decoction is employed to treat dysentery and in the Solomon islands the leaves are used to relieve pain in the chest. In India, *P. urinaria* is considered a very good diuretic.

**Observations** A monoecious, generally annual, usually erect herb up to 80 cm tall with phyllanthoid branching, cataphylls scarious, their stipules ovate-lanceolate and conspicuously auriculate; deciduous branchlets (3-)5-13 cm long with 10-42 leaves; leaves oblong or elliptical-oblong to elliptical-obovate, 4-20(-25) mm × (1-)3-6(-9) mm, obtuse to rounded and sometimes slightly unequal at base, apex rounded to obtuse and often apiculate, subsessile, stipules unequal, triangular-lanceolate; proximal 5-20 nodes of deciduous branchlets with solitary female flowers, succeeding nodes bearing monochasia of 5-7 male flowers; male flowers with 6 calyx lobes, disk segments 6, stamens 3, filaments connate, anthers free, dehiscing vertically; female flowers subsessile, calyx

lobes 6, disk cup-shaped, margin sometimes crenulate, styles fused at base into a triangular plate; fruit a globular capsule, about 2 mm in diameter, usually rugose; seeds with sharp transverse ridges on the back and sides. *P. urinaria* is highly variable and two subspecies have been recognized. The rare subspecies *nudicarpus* Rossignol & Haicour has creeping branches that root on the nodes and smooth-skinned capsules; within Malesia it is found in the Philippines. *P. urinaria* is a common weed of waste places, clearings, gardens, along paths, but is also found in evergreen forest and bamboo forest. It grows on well-drained, fertile, sandy soils, sometimes on limestone, often in humid places or even in marshy ground, up to 1500 m altitude.

**Selected sources** 97, 202, 536, 580, 597, 965, 1035, 1126, 1128, 1135, 1204, 1255, 1380, 1386, 1476, 1525, 1555, 1560.

F.L. van Holthoorn

### **Phytolacca L.**

Sp. pl. 1: 441 (1753); Gen. pl. ed. 5: 200 (1754).

PHYTOLACCACEAE

$x = 9$ ; *P. acinosa*:  $n = 9$ ,  $2n = 36$ , 72, *P. americana*:  $n = 9$ ,  $2n = 36$ , *P. dodecandra*:  $2n = 36$ , *P. octandra*:  $2n = 36$

**Major species** *Phytolacca acinosa* Roxb., *P. americana* L.

**Vernacular names** Pokeweed (En). Vietnam: th[uw]lowng l[u]j.c.

**Origin and geographic distribution** *Phytolacca* consists of about 25 species, most of which are native to the tropical and subtropical regions of South and Central America, with a few species in Africa, Madagascar and Asia. No species occur naturally in South-East Asia, but a few have been introduced and these have sometimes naturalized.

**Uses** In traditional medicine in Vietnam the roots of *P. acinosa* are considered to be diuretic and antiphlogistic, and are used internally (as a decoction) against dropsy, ascites, oedemas and pleuritis. Externally they are used against pharyngitis, boils and swellings. Outside South-East Asia the uses of roots of *P. acinosa* and *P. americana* (e.g. in Chinese medicine) are, in addition to those already mentioned, antiparasitic, laxative and antirheumatic, and against apoplexy, tumours and bronchitis.

Berries of the African *P. dodecandra* are a potent molluscicide, and can be used to control bilharzia-transmitting snails. They are also used as a soap

substitute in Ethiopia. In Africa, parts of the plant are used for various medicinal purposes.

The young shoots and leaves of several *Phytolacca* species, when cooked, are used as a vegetable. After their toxic constituents have been removed, the red fruits of *P. americana* can be used to colour wine and foods. Some species, particularly *P. americana*, are common ornamentals in temperate climates. They are sometimes weeds.

**Production and international trade** To date, *Phytolacca* has not been important as a medicinal plant in South-East Asia except in countries in Indo-China, where it is sometimes cultivated as a medicinal plant in pots or home gardens. Plant parts or products are not traded commercially.

Small areas of *P. dodecandra* are grown in Ethiopia, Swaziland, Zambia and Zimbabwe. Experimental trials for its introduction in Indonesia are being started.

**Properties** *Phytolacca* leaves, fruits and roots contain numerous triterpenoids, e.g. phytolaccagenin, isophytolaccagenin, phytolaccagenic acid, isophytolaccinic acid, phytolaccanol, acetylaleuritic acid, acinospesigenin, acinosolic acid, esculentic acid, esculentagenic acid, esculentagenin, jaligonic acid and spergulagenic acid.

On a dry weight basis the pericarp of *P. dodecandra* berries contains 25% bidesmosidic saponins with an oleanolic-acid aglycon; these saponins have molluscicidal properties. At concentrations well below 75 mg/kg, and after a certain period of exposure, fish and snails are killed. Other water animals such as insect larvae and tadpoles, are not affected at the concentrations that kill fish and snails. Tests showed that *P. dodecandra* berries have a potential use against *Schistosoma* larval stages in fresh water in schistosomiasis control programmes; they have cercariacidal and miracidicidal properties.

Anti-inflammatory activity is attributed to the triterpenoid saponins, such as phytolaccoside and esculentoside, which are common e.g. in the roots. The inhibition of antibody production may partially explain the anti-inflammatory effect. Fungistatic properties have also been attributed to the saponins (e.g. yiamoloside from *P. octandra*). The bark and the roots of several species (e.g. *P. dodecandra*) are poisonous for people and animals. The toxicity may manifest in hallucinations. Lethal poisoning of horses caused by *Phytolacca* roots has been reported. Patients using Chinese drugs containing *Phytolacca* have also been poisoned. Aerial parts of *P. americana* have been found to have antigalactagogic effects in cattle. The roots of *P.*

*acinosa* and *P. americana* have hypotensive properties; they contain the hypotensive agents histamine and gamma-aminobutyric acid. When ingested, leaves of *P. americana* typically produce self-limited but severe gastro-enteritis, characterized by intense vomiting and frothy diarrhoea.

Tests with mice suggest that *P. acinosa* polysaccharides augment the immunological functions in vivo and inhibit tumour growth; the antitumour effect may be mainly related to the augmenting effect on macrophages in the mice. Leaf and seed extracts of several species have shown antiviral activity in tests, e.g. against golden mosaic virus and tobacco mosaic virus in beans and tobacco, and against sugar cane mosaic virus and cucumber green mottle mosaic virus, and also against viruses in animal cells. The complete amino acid sequence of antiviral protein from *P. americana* seeds has been determined. The proteins are ribosome-inactivating. The lectins from the roots of *P. octandra* are mitogenic for unseparated human peripheral blood lymphocytes and stimulate plasma cell formation. Lectins from the roots of *P. americana* are called pokeweed mitogen (PWM). They are haemagglutinating (tested with human blood group A erythrocytes) and mitogenic (determined by <sup>3</sup>H-thymidine incorporation in lymphocyte cultures). Pokeweed mitogen plays an important role in fundamental leukocyte research.

The roots, leaves, and particularly, seeds of *P. acinosa* have abortifacient activity in mice. Extracts of *P. americana* have lysozyme activity. Fruits of *P. acinosa* and *P. americana* contain betalains such as humilixanthin. Lectins from *P. americana* have insecticidal properties. The neo-lignans americanol and isoamericanol from the seeds of the latter species showed neurotrophic properties in in vitro tests with rat cells. *P. americana* shows allelopathic activity: it may inhibit seed germination and seedling growth of crops like lettuce, sesame and cucumber.

The berries of *P. americana* contain the red betacyanin pigment phytolaccanin, which is identical to betanin from beetroot. The toxic saponins must be removed before this pigment is used as a food colourant. The colourant can be used in foods with a wide range of acidity, since it is not influenced by pH; however, its stability at room temperature is poor, so it is recommended for refrigerated and frozen foods.

The leaves of *P. acinosa* are reported to be a rich source of iron, phosphorus and calcium; fresh samples contain 84% water and 2.4% ash.

**Adulterations and substitutes** In Japan,

roots of *Cynanchum caudatum* Maxim. are used as a substitute for *Phytolacca* roots; they are also diuretic. *Hibiscus sabdariffa* L., *Tamarindus indica* L. and *Tacca leontopetaloides* (L.) O. Kuntze show toxic activity against snails transferring the parasitic trematode *Schistosoma mansoni* (causing bilharzia), which is comparable to *Phytolacca dodecandra*.

**Description** Erect or scandent perennial herbs, sometimes shrubs and rarely trees, usually glabrous except for the often papillate or short-haired main axis of the inflorescence; stems up to 3 m long, but up to 10 m when scandent, often angular; roots often long and fleshy or tuberous. Leaves alternate, simple and entire, usually ovate-oblong to oblong-lanceolate, herbaceous, acute at both ends, petiolate; stipules absent. Inflorescence terminal or pseudolateral, racemose, often long, sometimes spiciform. Flowers usually bisexual, but sometimes functionally male or female, actinomorphic, pedicel often short; perianth 5-partite, herbaceous, greenish, whitish or pinkish during anthesis and dark red under the ripe fruit, tepals free, equal or slightly unequal, ovate or obovate to lanceolate, persistent, spreading during anthesis, later often reflexed; stamens (6–)7–22(–33), in 1 or 2 series, inserted on the outer margin of the disk, sometimes also partly on the underside of the disk, filaments usually free, filiform-subulate, anthers dorsifixed, bilobed at both ends; carpels (3–)5–10(–16), whorled, laterally connate into a superior depressed-globose ovary, or almost free (*P. acinosa* and *P. dodecandra*), ovules solitary and basal in each carpel, styles terminal on the inner angle of the carpels, subulate, short, erect or recurved. Fruit a depressed-globose berry, longitudinally 5–10-furrowed, or apocarp, juicy, orange or dark red to black, usually 5–10-seeded. Seeds strongly laterally compressed, oval to lenticular, inaequilateral at base, shining black; embryo large, peripheric, enclosing the endosperm.

**Growth and development** In Ethiopia *P. dodecandra* flowers and fruits throughout the year, but with a peak during the dry season. Plants in the shade often do not flower. Pollination is effected by insects such as ants, flies and spider wasps. *Phytolacca* fruits are often eaten by birds which disperse the seed. Sometimes monkeys also eat the fruits.

**Other botanical information** *Phytolacca* belongs to the subfamily *Phytolacchoideae*, together with the genera *Anisomeria* and *Ercilla*, which are both restricted to South America.

*Phytolacca* species are often difficult to distinguish. Many of the morphological characters appear to be under weak genetic control. Moreover, many species hybridize readily, thus obscuring the characters by which they are recognized. Three highly molluscicidal and productive cultivars of *P. dodecandra* have been developed and are now in production in eastern and southern Africa.

**Ecology** In South-East Asia, *Phytolacca* species are found in open forest and forest borders, roadsides, along watercourses and in waste places, up to 1700 m altitude in Java. They occur very locally in South-East Asia, but are common in some locations. *P. dodecandra* usually occurs in Africa at altitudes above 1000 m, with an annual rainfall of about 1400 mm and a distinct dry period.

**Propagation and planting** Mass multiplication is done by non-woody stem cuttings. It is advantageous to use a 50–75 mg/kg  $\alpha$ -naphthalene acetic acid solution as root-promoting substance, and a slightly acid soil medium. After 6–8 weeks rooted cuttings can be planted in the field, usually at 1–3 m  $\times$  2–3 m. Propagation by seed is only appropriate for selection purposes. Soaking seed of *P. americana* in e.g. concentrated  $H_2SO_4$  prior to sowing improves germination rate. Seed of *P. dodecandra* takes about 14 days to germinate.

**In vitro production of active compounds** Betacyanins have been produced in cell cultures of *P. americana* initiated from stem explants. The cells were maintained in Schenk-Hildebrandt medium. The suspension was subcultured every week in darkness at 25°C, and calluses were subcultured every 3 weeks. Whereas in fruits prebetanin (betanin 6'-O-sulphate) and its isoform predominate, in the cell culture feruloylated derivatives occur as the major components. Callus cultures of *P. americana* can be stored at 4°C for at least 3 months, but betalain production of cultures that have been stored is inferior. A dual culture consisting of callus of *P. americana* and the fungus *Botrytis fabae* showed marked fungicidal activity to *Cladosporium herbarum*. The main active constituent of this extract was identified as phytolaccoside B.

**Husbandry** Plantings of *P. dodecandra* must be shaded in the first weeks. Occasional watering and weeding are important until the crop has become established.

**Diseases and pests** *P. dodecandra* plantings may be attacked by leaf and stem borers (*Gitona* spp.), so far the only serious insect pests. Precautions have to be taken against soilborne insect larvae.

**Harvesting** The content of saponins of *P. dodecandra* berries varies seasonally: berries harvested during the dry season just before the onset of the rains have the highest content. Berries possess the highest molluscicidal potency when fully developed but still unripe. Complete fruiting racemes are collected and dried in the open under shade.

**Yield** About 1000 kg dry fruits of *P. dodecandra* can be obtained per ha per year, i.e. about 250 g of dry fruits per plant annually.

**Handling after harvest** Dry fruits of *P. dodecandra* can be stored for many years without losing molluscicidal activity. While grinding the berries care should be taken to avoid contact with the dust because it irritates the mucous membranes.

**Genetic resources and breeding** *Phytolacca* occurs in South-East Asia only in cultivation or as an escape. The species highlighted here have a wide distribution and are rather commonly cultivated and naturalized outside South-East Asia. With the exception of *P. dodecandra* and some ornamentals (particularly *P. americana*) there has been no serious selection and breeding.

The world germplasm collection of *P. dodecandra* covers all the highland areas in Africa between 20°N and 30°S. There is abundant morphological variation. In Ethiopia, the Institute of Pathobiology in Addis Ababa has a collection.

**Prospects** Although *Phytolacca* species are little known in South-East Asia, they might be promising for planting. The medicinal properties are interesting and comparatively well documented and, moreover, the plants also have molluscicidal, insecticidal and possibly fungicidal properties, and can be used as a vegetable and ornamental. Undoubtedly, several *Phytolacca* species are able to grow well in South-East Asia, particularly at higher elevations. However, more research is needed on planting requirements and uses, to realize the potential of these plant resources.

The use of *P. dodecandra* as a molluscicide might be limited, since the frequent applications required to ensure that treated waters remain clear of snails might also drastically reduce the fish population. However, as infected snails only occur at locations heavily frequented by people, berry suspensions can be applied locally. A few square metres of cultivated plants will enable people to treat their snail-infested watersides themselves, preferably during the dry season.

**Literature** |1| Bodger, M.P., McGiven, A.R. & Fitzgerald, P.H., 1979. Mitogenic proteins of poke-

weed - part 1: purification, characterization and mitogenic activity of 2 proteins from pokeweed (*Phytolacca octandra*). *Immunology* 37(4): 785-792. |2| Fukuyama, Y., Hasegawa, T., Toda, M., Kodama, M. & Okazaki, H., 1992. Structures of americanol A and isoamericanol A having neurotrophic properties from the seeds of *Phytolacca americana*. *Chemical and Pharmaceutical Bulletin* 40(1): 252-254. |3| Hamilton, R.J., Shih, R.D. & Hoffman, R.S., 1995. Mobitz type I heart block after pokeweed ingestion. *Veterinary and Human Toxicology* 37(1): 66-67. |4| Kobayashi, A., Hagi-hara, K., Kajiyama, S., Kanzaki, H. & Kawazu, K., 1995. Antifungal compounds induced in the dual culture with *Phytolacca americana* callus and *Botrytis fabae*. *Zeitschrift für Naturforschung, Section C, Biosciences* 50(5-6): 398-402. |5| Lugt, Ch.B., 1989. *Phytolacca dodecandra* L'Hér. In: Westphal, E. & Jansen, P.C.M. (Editors): *Plant Resources of South-East Asia. A selection*. Pudoc, Wageningen, the Netherlands. pp. 224-225. |6| Nguyen Van Duong, 1993. *Medicinal plants of Vietnam, Cambodia and Laos*. Mekong Printing, Santa Ana, California, United States. 528 pp. |7| Nowicke, J.W., 1969. Palynotaxonomic study of the Phytolaccaceae. *Annals of the Missouri Botanical Garden* 55(3): 294-364. |8| Wang, H.B., Zheng, Q.Y., Qian, D.H., Fang, J. & Ju, D.W., 1993. Effects of *Phytolacca acinosa* polysaccharides I on immune function in mice. *Acta Pharmacologica Sinica* 14(3): 243-246. |9| Yeung, H.W., Feng, Z., Li, W.W., Cheung, W.K. & Ng, T.B., 1987. Abortifacient activity in leaves, roots and seeds of *Phytolacca acinosa*. *Journal of Ethnopharmacology* 21(1): 31-36. |10| Zhu, X. & Hu, Z., 1989. Preparation of the antiviral protein from pokeweed seeds and assay of its toxicity. *Acta Botanica Yunnanica* 11(4): 440-448.

#### *Selection of species*

#### **Phytolacca acinosa Roxb.**

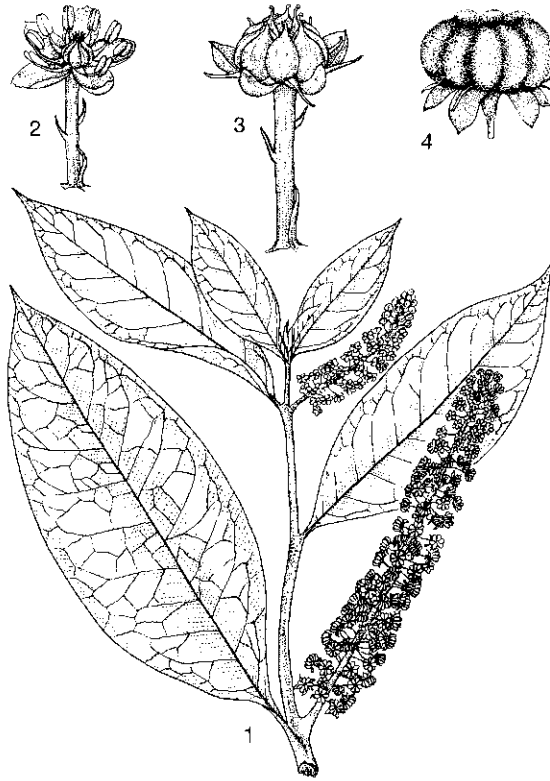
Hort. bengal.: 35 (1814).

**Synonyms** *Phytolacca esculenta* van Houtte (1848).

**Vernacular names** Indian poke (En). Vietnam: th[uw][ow]ng l[u]c nh[or].

**Distribution** Pakistan, India, Nepal, Bhutan, China, Taiwan, Korea and Japan; introduced in Vietnam, occasionally cultivated elsewhere, e.g. in the Philippines.

**Uses** The roots are used in traditional medicine in Vietnam, both internally and externally; they



*Phytolacca acinosa* Roxb. - 1, flowering and fruiting stem; 2, flower; 3, young fruit; 4, mature fruit.

are commonly used in Chinese medicine. In Japan, the entire plant is used as a diuretic, while in India, it is used to alleviate body pain. Young leaves are cooked and eaten as a vegetable. *P. acinosa* is sometimes cultivated as an ornamental.

**Observations** A perennial herb up to 2(-3) m tall, often with purplish stems; leaves elliptical to ovate, up to 35 cm × 16 cm; flowers generally shorter than leaves, bisexual, stamens 7-15 (often 10), sometimes in 2 whorls, carpels 6-9 (often 8), free in fruit. The roots of *P. acinosa* taste bitter-sour and pungent.

**Selected sources** 190, 287, 452, 816, 900, 1035, 1057, 1356, 1545, 1627, 1631, 1632, 1660.

### *Phytolacca americana* L.

Sp. pl. 1: 441 (1753).

**Synonyms** *Phytolacca decandra* L. (1763).

**Vernacular names** Pokeweed (En). Raisin d'Amérique, morelle à grappe (Fr). Laos: kub nyuj (Hmong). Vietnam: th[uw]low]ng l[u]c.

**Distribution** Originating from North America, now cultivated worldwide and sometimes natural-

ized; in South-East Asia particularly in Indo-China.

**Uses** The roots in particular have numerous medicinal uses in Indo-China; they are also used in Chinese medicine. Young shoots are cooked and eaten as a vegetable. The red fruits can be used to colour wine and foods, but the toxic substances must be removed. *P. americana* is a common ornamental in temperate climates.

**Observations** A perennial branched herb up to 3 m tall, often with purplish stems, roots tuberous; leaves elliptical to ovate-lanceolate, up to 30(-40) cm × 12 cm; flowers in racemes generally longer than leaves, bisexual, stamens about 10, in 1 whorl, carpels about 10, united in fruit. *P. americana* occurs in Laos up to 2000 m altitude. In Indo-China it is a relic of former cultivation for the dye from the fruits.

**Selected sources** 80, 298, 390, 424, 425, 436, 450, 507, 538, 590, 703, 755, 797, 816, 900, 1035, 1057, 1298, 1356, 1660.

### *Phytolacca dodecandra* L'Hér.

Stirp. nov. 6: 143, pl. 69 (1791).

**Synonyms** *Phytolacca abyssinica* Hoffm. (1796).

**Vernacular names** Endod, soap berry (En).

**Distribution** Central and southern Africa and Madagascar.

**Uses** The berries are used to control bilharzia-transmitting snails; they are also used as a soap substitute. Parts of the plant have various medicinal uses, e.g. laxative, anthelmintic and in the treatment of respiratory problems.

**Observations** A dioecious scandent shrub with branches up to 10 m long; leaves usually ovate, up to 15 cm × 10 cm; flowers in racemes generally longer than leaves, unisexual, stamens 8-15(-20), in 2 whorls, carpels (3-)5(-8), free in fruit.

**Selected sources** 157, 670, 841, 877, 1014, 1057, 1563.

### *Phytolacca octandra* L.

Sp. pl. ed. 2, 1: 631 (1762).

**Vernacular names** Inkweed, dyeberry (En).

**Distribution** Probably native to tropical America, but now pantropical; in South-East Asia very locally naturalized in northern Sumatra and western Java.

**Uses** The roots have fungistatic and medicinal properties. Young sprouts and leaves can be used as a vegetable.

**Observations** A perennial branched herb up to 60(-200) cm tall; leaves oblong-lanceolate to

ovate-lanceolate, up to 15(-22) cm × 6(-7.5) cm; flowers in spiciform racemes shorter to slightly longer than leaves, bisexual, stamens 8-15(-20), in 1 whorl, carpels 7-9(-10), united in fruit. In Malesia, *P. octandra* occurs at about 1700 m altitude.

**Selected sources** 96, 165, 166, 976, 1057, 1356.  
Razali Yusuf

## Plantago L.

Sp. pl. 1: 112 (1753); Gen. pl. ed. 5: 52 (1754).

### PLANTAGINACEAE

$x = 4, 5$ ; *P. afra*:  $2n = 12$ , *P. asiatica*:  $2n = 12, 24, 36$ , *P. lanceolata*:  $2n = 12 + 0-1B, 24$ , *P. major*:  $2n = 12, 24$ , *P. ovata*:  $2n = 8, 16$

**Major species** *Plantago major* L.

**Vernacular names** Plantain (En; a confusing name as in the tropics it is mainly used for the cooking banana).

**Origin and geographic distribution** *Plantago* consists of approximately 250 species and is cosmopolitan except the polar regions. It is essentially temperate in its natural distribution. Some species, particularly *P. major* and *P. lanceolata*, are extremely widespread, throughout temperate regions and also penetrating into tropical highland regions. *P. major* is the most widespread species in South-East Asia, and *P. lanceolata* is only very locally naturalized.

**Uses** The seed or seed coat of e.g. *P. afra* and *P. ovata* are widely used as laxative, but also to treat dysentery of amoebic and bacillary origin and diarrhoea. Products containing seed preparations are taken by many people worldwide to control bowel function: to treat habitual constipation, as supportive therapy in diarrhoea and with irritable bowel syndrome. Seed husks ('Psyllium husk') have become very popular in breakfast foods in the United States. They may also be a useful supplement in weight control diets by affecting fat intake and effecting a subjective feeling of fullness. The seeds of *P. asiatica* and *P. major* are considered as expectorant, diuretic and antimicrobial in Chinese and Vietnamese medicine.

In China and Japan, the aerial parts of these plants (known as 'Plantago herba') are commonly used to treat coughs, bronchitis and asthma. *Plantago* leaves are used traditionally topically as an emollient and itch-relieving treatment against dermatological complaints and eye irritation. A poultice of 10% of dried and pulverized leaves of *P. major* is used to treat eczema, but juice from

pressed fresh leaves, extracts in 96% alcohol and maceration products are also used. In modern phytotherapy aerial parts or leaves of *P. major* ('Plantago major herba') and *P. lanceolata* ('Plantago lanceolatae herba/fohium') are used to alleviate irritation in catarrh of the upper respiratory tract. Macerates, fluid extracts, syrups and expressed juice from the fresh plant are all used for treating inflammation of the mouth and throat, and externally for inflamed skin.

The seed mucilage is also used in cosmetics (e.g. in lotions and hair wave sets) and as a basic stabilizer in the ice-cream industry. It is also used in the preparation of chocolate; it is an excellent thickener. The seeds can be used as a source of a low-cost gelling agent for tissue culture. The quality is reasonably comparable with that of agar, and the costs are only about 10% of those for agar.

Leaves are edible and sometimes eaten as vegetable. In some areas, *P. lanceolata* is considered a good fodder; it is reported to be a good source of calcium, potassium, sodium, cobalt, phosphorus and chlorine.

**Production and international trade** India is an important exporter of seed husks (mainly of *P. ovata* and to a lesser extent of *P. afra*), especially to the United States, where annual imports were worth about US\$ 3.5 million in the late 1980s. It has been estimated laxatives containing *Plantago* are daily used by about 4 million Americans.

**Properties** The amount of mucilage in the seeds can be substantial (up to 30% in *P. ovata*). Being located only in the epidermis of the testa, it mainly consists (up to 85%) of a water-soluble polysaccharide fraction (arabinoxylan) in which D-xylose is the main constituent. The backbone is a xylan polymer with 1-3 and 1-4 linkages and no apparent regularity in their distribution, with the xylose monomers substituted on C-2 or C-3 by L-arabinose, D-xylose and  $\alpha$ -D-galacturonyl-L-rhamnose. The quality of the mucilage is evaluated by measuring its swelling index, which should exceed 9. *P. lanceolata* and *P. major* provide a similar mucilage which is rich in D-galactose and L-arabinose and contains nearly 40% uronic acids. On hydrolysis the mucilage of *P. afra* yields D-xylose (about 70%), L-arabinose (about 10%),  $\alpha$ -D-galacturonyl-(1-4)-L-xylose and D-galactose; the swelling index must be at least 10. The seeds also contain 5-10% lipids with unsaturated fatty acids, sterols, 15-18% proteins, traces of cyclopentopyridine-type alkaloids and the iridoid aucubin. An isomer of ricinoleic acid,  $\beta$ -hydroxyolefinic acid 9-hydroxy-cis-11-octadecenoic acid, has been

found as a minor constituent (1.5%) of the seed oil of *P. major*.

The seeds owe their laxative properties to the very hydrophilic polysaccharides. The action is purely mechanical and linked to the mucilage taken together with abundant fluid; the polysaccharide macromolecules absorb much water and form a gel that increases stool bulk, stimulates peristalsis and facilitates bowel movements. The effect has been confirmed by several clinical studies. In double-blind, placebo-controlled studies with patients suffering from chronic constipation, administration of *Plantago* seeds (or preparations containing seed testa) showed good results, increasing the frequency and decreasing the consistency of stools. No adverse effects were observed and, notably, no flatulence occurred, as often seen in patients taking bran. On the other hand, the mucilage can also be used as supportive therapy in diarrhoea: by absorbing water, the transit period of the bowel contents is extended. The mucilage has also frequently been reported to lower blood sugar and cholesterol, similar to the activity following the administration of galactomannans or pectins, although in general these effects are only very slight and often not demonstrated in clinical studies. In some tests, however, it was demonstrated that *P. afra* mucilage is useful as an adjunct to dietary therapy in patients with type II diabetes. A test in which patients were treated with a commercially available preparation of *P. ovata* showed positive effects on internal bleeding haemorrhoids. *P. ovata* seeds might be as effective as mesalamine to maintain remission in ulcerative colitis.

*P. major* seeds, administered orally, showed a significant haemostatic activity in the treatment of menorrhagia in a preliminary clinical study in India. When tested under standardized conditions in a placebo-controlled double-blind crossover model in Vietnam, no influence of the drug was recorded on the urine output and sodium excretion.

The reticuloendothelial system-potentiating and alkaline phosphatase-inducing activities of the mucilage from *P. asiatica* have been found to be markedly enhanced when the mucilage was de-O-acetylated. The deacetylated product showed considerable anti-complementary activity as well as considerable hypoglycaemic activity on administration to mice.

Administration of the seeds has no serious side-effects; only a few exceptional cases of allergic reactions have been documented. However, the husks are known to elicit respiratory allergic reactions

after inhalation or ingestion by sensitized individuals. Immunological, biochemical and microscopic findings suggest that other contaminating seed components are primarily responsible for the allergenicity of commercial-grade *Plantago* husk powder rather than the husk itself. The preparations are contra-indicated if there is pyloric stenosis and abnormal narrowing of the gastro-intestinal tract, and must be used with care, to avoid product stagnation in the oesophagus in the case of bedridden patients with megacolon by alteration of colon motility, and in patients with diabetes that is difficult to control.

Leaves of *P. major* contain iridoids and phenols: flavonoids, phenolic acids and phenylpropanoic esters of glycosides (verbascoside, plantamajoside). The iridoid glucoside majoroside has also been isolated from *P. major*, along with aucubin and catalpol. *P. lanceolata* leaves also contain iridoids (1.9–2.4%) such as aucubin, catalpol and asperuloside, flavonoids and phenolic acids. Aucubin glycoside can be detected by thin-layer chromatography of a methanol extract. It can also be quantified by high pressure liquid chromatography. The proposed levels for the French pharmacopoeia are 0.5% for *P. major* and 1.0% for *P. lanceolata*. The content of aucubin in *P. asiatica* is reported in Vietnam to decrease gradually from the roots to the flowers and leaves. The drying temperature affects the content of aucubin; plant material dried at 80°C is reported to contain higher concentrations of aucubin than plant material dried at 40°C. Analysis of dried samples sold on markets in Vietnam showed that aucubin was absent, but that the allantoin content was often high (up to 80%) which makes the drug effective for the treatment of burns and stomach ulcers. The iridoids have a potential role in anti-inflammatory activity. Aucubigenin liberated from aucubin by glycosidases present possesses antibacterial activity. 3,4-Dihydroxyphenethyl alcohol-6-O-caffeoyl- $\beta$ -D-glucoside, the phenylethanoid glycoside plantasioside, acteoside (verbascoside), plantaginin, plantamajoside and the phenylpropanoid glycoside hellioside have been isolated from aerial parts of *P. asiatica*. Acteoside showed high inhibition of lens aldose reductase. Plantamajoside and hellioside showed high inhibition of cyclic AMP phosphodiesterase and 5-lipoxygenase, which might have some correlation with the therapeutic effect of the herb as anti-inflammatory and anti-asthmatic. Acteoside and plantamajoside, which have also been found in *P. lanceolata*, showed inhibitory effects on arachidonic acid-induced ear oedema in



mice. A hot-water extract of the whole plant of *P. major* exhibits diuretic activity and dissolves kidney stones. A chromatographic fraction of dried leaves was found to promote wound healing. Aqueous extracts of *P. lanceolata* showed immunomodulatory effects; in tests they stimulated the production of anti-SRBC (IgG) antibodies in mice and stimulated the release of angiogenic factors by mouse spleen cells and human mononuclear blood cells, and in vitro a *P. lanceolata* polysaccharide fraction showed an increase in phagocytosis of granulocytes. Liquid *P. lanceolata* preparations are said to have hepatoprotective (chloroform,  $\alpha$ -amanitin) activity, and might also offer protection against adverse effects of cytostatic agents (e.g. 5-fluorouracil). In vitro experiments showed a mortality of 76% of the zooflagellate *Giardia duodenalis* when treated with a *P. major* extract.

In the United States, interplanting peach trees with *P. lanceolata* reduced nematode numbers (*Criconebella xenoplax*), but not to acceptable levels to justify commercial control. In tests with germinating lettuce seeds, aqueous extracts of *P. lanceolata* and *P. major* showed allelopathic activity.

Pollen, particularly that of *P. lanceolata*, may cause allergic reactions in sensitive persons.

**Adulterations and substitutes** The seed of flax (*Linum usitatissimum* L.) also contains mucilage which can be used as 'bulk laxative' just like that of *Plantago* seeds. All parts of some *Malvaceae* such as the European *Althaea officinalis* L. and *Malva sylvestris* L. contain mucilage which is used orally in the adjunctive therapy of the painful component of spasmodic colitis and for symptomatic treatment of cough, and topically as emollient and to treat itch. In India, *Plantago* seeds are frequently mixed with those of *Salvia aegyptiaca* L., which also yield copious mucilage.

Iridoid glycosides are fairly common in dicotyledonous Angiosperms. They are, for instance, present in *Scrophulariaceae* such as *Verbascum* species, which are used in phytotherapy for similar purposes as *Plantago*. Aucubin is also present in the genera *Aucuba* (*Cornaceae*) and *Garrya* (*Garryaceae*).

**Description** Annual or perennial herbs up to 50(–80) cm tall, stemless or with branched stem. Leaves in basal rosettes or opposite, with distinct, parallel veins, usually distinctly petiolate when in rosettes and without distinct petiole when inserted on stems; stipules absent. Inflorescence a pedunculate, bracteate spike. Flowers actinomor-

phic, usually bisexual, 4-merous; sepals connate at base or free, equal or nearly so, imbricate, scarious, persistent; corolla gamopetalous, usually with patent or deflexed imbricate lobes, scarious, persistent; stamens inserted on corolla tube, alternating with corolla lobes, exerted, anthers conspicuous; ovary superior, 2–4-locular, style 1 with a stout pilose stigma, protruding. Fruit a circumscissile capsule, few- to many-seeded. Seeds with endosperm and straight embryo; testa thin, often mucilaginous when wet. Seedling with epigeal germination; cotyledon sessile or shortly petiolate, sheathed at base; hypocotyl elongate, epicotyl absent; first leaves alternate or opposite (in species developing stems).

**Growth and development** In tropical climates *Plantago* may flower all year round, and a life cycle may be accomplished in 6 weeks. The flowers remain functionally female much longer than functionally male. The stigma is already protruding and receptive when other flower parts are still in bud, whereas it can still be functional after the stamens have withered. *P. lanceolata* is gynodioecious: specimens can be found with either bisexual flowers or only female ones. In *P. lanceolata* cross-pollination is the rule, whereas in *P. major* self-pollination is common. The flowers are wind-pollinated.

The seeds are already ripe 2–3 weeks after fertilization. Seeds readily adhere to animals or people thus facilitating their dispersal. They can also be transported by water. Perennial *Plantago* species may live for up to 15 years; in cultivation in Turkmenistan they may last up to 8 years.

**Other botanical information** *P. major* and *P. asiatica* are probably often confused as they are difficult to distinguish. However, they are reported to be clearly distinguishable in Japan by anatomical characters of seeds and leaves. Some authors consider both species to be conspecific.

*P. afra* is closely related to *P. arenaria* Waldst. & Kit. (synonym: *P. indica* L.), which in southern Europe is a source of mucilage with medicinal value too. Some cultivars of *P. afra* and *P. ovata* were developed in India in the late 1980s.

*P. ovata* and *P. afra* seem to be occasionally confused in the literature on cultivated *Plantago* in India, although they can easily be distinguished by leaves in rosettes (*P. ovata*) or on distinct, much-branched stems (*P. afra*).

**Ecology** *P. lanceolata* and *P. major* occur in areas disturbed by man. Because of their morphology they are well adapted to withstand the trampling of livestock and humans. Once established

in fields, they can become a noxious weed in e.g. coffee, onions, cotton and, in particular, in cereals like rice.

In general, cool and dry weather is favourable to crops of *P. afra* and *P. ovata*; they require dry weather from flowering until seed maturity (about 2.5 weeks). *P. major* is more tolerant of compacted soils than *P. lanceolata*, so the former appears frequently in almost pure stands on the edges of paths, whereas the latter often occurs in grasslands. *P. major* is more tolerant of waterlogging, whereas *P. lanceolata* is more drought tolerant. *P. afra* and *P. ovata* tolerate dry, sandy soils extremely well. They are grown in India on medium to poor sandy soils; however, they grow best on rich, well-drained loamy soils.

**Propagation and planting** *P. major* reproduces mainly by seed. A seed production of up to 14 000 seeds per plant has been reported for this species. *Plantago* may be multiplied by seed or vegetative means. For large-scale seed production of *P. ovata*, propagation is by seed. The seed is broadcast or drilled in rows 30 cm apart, so 6–13 kg/ha of seed is needed. *P. lanceolata* may also be propagated by new buds arising on the thick underground stem. The 1000-seed weight of *P. afra* is 1.1–1.2 g. Plant spacing in experimental plantings of *P. afra* in Thailand was 25 cm × 10–30 cm. Interplanting of *P. ovata* with poplar (*Populus* spp.) has shown good results in India. Seeds may exhibit dormancy, which can be broken by several months of dry storage at room temperature or by a few weeks at 5°C. Seed 1–5 years old shows better germination rates than fresh seed. Seed has remained viable in the soil for over 60 years.

**Husbandry** In India, *P. ovata* as a crop is irrigated. Weeding is carried out when the crop is about 6 weeks old.

**Diseases and pests** *P. lanceolata* may serve as a host for *Pseudomonas viridiflava* and *P. syri-nage* which can cause a serious bacterial disease in tomato. In India, *P. major* has been reported a host for *Meloidogyne hapla* which causes root galls in Irish potato. *Meloidogyne* species have also been identified from *P. lanceolata*. Larvae of the fruit weevil *Naupactus xanthographus*, a pest of several fruit trees, have been observed on *P. major*. *P. ovata* has been reported to be a host of the coriander aphid (*Hyadaphis coriandri*) which can be a pest of coriander, fennel, dill and celery in India.

**Harvesting** In Thailand, harvesting *P. afra* seeds 17 days after the first inflorescence was mature gave the highest seed yield. This means that

the crop can already be harvested about 10 weeks after sowing. In western India, *P. ovata* is harvested after 3–4 months. Plants are cut about 15 cm above the ground early in the morning to prevent seed shedding.

Care should be taken when collecting *P. major* from the wild for medicinal purposes, since plants may contain high concentrations of heavy metals like lead and cadmium as they often grow along the roads.

**Yield** The seed yield of *P. afra* in experimental plantings in Thailand was estimated at about 1150 kg/ha. The seed yield of *P. ovata* in India is 500–1100 kg/ha. The yield of husks is approximately 25% of the seed weight.

**Handling after harvest** Plants harvested for seed are threshed, winnowed and sieved until the seed is clean. The husk is removed by mechanical milling and subsequent winnowing and sieving.

**Genetic resources and breeding** The *Plantago* species treated here are common in anthropogenic habitats, and there is no reason to consider them as liable to genetic erosion. Except for *P. major* and *P. lanceolata*, the *Plantago* species highlighted here are not yet found in Malesia. This implies that the genetic variability of *Plantago* is limited in the region.

**Prospects** The uses of *Plantago* orally to treat digestive and bronchial disorders and topically to treat skin disorders are very widespread. Modern research seems to confirm the activity of the drug, although information is far from complete. This, plus the ease of cultivation, seems to justify advocating planting *Plantago* for medicinal purposes. However, more research is needed to achieve a proper standardization of the drug, and to establish the potential in the Malesian region of *Plantago* species that are currently cultivated in neighbouring countries like Thailand. Leaves of *P. asiatica* and *P. lanceolata* have been recorded as a promising vegetable in regions with malnutrition.

**Literature** |1| Bruneton, J., 1995. Pharmacognosy, phytochemistry, medicinal plants. Technique & Documentation Lavoisier, Paris, France. pp. 97–100. |2| Chanprasert, W., Paisooksantivatana, Y., Lersrutaiyotin, R., Phoolkets, U., Satakun, D. & Thapatat, Y., 1994. Effect of planting date, plant spacing, and harvesting date on seed yield and quality of *Plantago psyllium* L. Thai Journal of Phytopharmacy 1(2): 29–37. |3| Council of Scientific and Industrial Research, 1969. The wealth of India. Vol. 8. Publications and Information Directorate, New Delhi, India. pp. 146–154. |4| Dat, D.D. et al., 1992. Studies on the

individual and combined diuretic effects of four Vietnamese traditional herbal remedies: *Zea mays*, *Imperata cylindrica*, *Plantago major* and *Orthosiphon stamineus*. *Journal of Ethnopharmacology* 36(3): 225–231. |5| Gupta, R., 1990. Indian herbs and herbal raw material - time-tested remedies for all ailments. *Indian Horticulture* 34(4): 39–41. |6| Holm, L.G., Plucknett, D.L., Pancho, J.V. & Herberger, J.P., 1977. The world's worst weeds. Distribution and biology. East-West Centre. University Press, Hawaii. pp. 385–393. |7| Kazmi, S.M.A., 1974. *Plantaginaceae*. In: Nasir, E. & Ali, S.I. (Editors): *Flora of West Pakistan*. No 62. Department of Botany, University of Karachi and National Herbarium, Pakistan Agricultural Research Council, Islamabad, Pakistan. 21 pp. |8| Ravn, H., Nishibe, S., Sasahara, M. & Xuebo, L., 1990. Phenolic compounds from *Plantago asiatica*. *Phytochemistry* 29(11): 3627–3631. |9| Sas, A.C., 1984. Plants and health. Eastern Publishing Association, Manila, the Philippines. p. 181. |10| Tomoda, M., Takada, K., Shimizu, N., Gonda, R. & Ohara, N., 1991. Reticuloendothelial system-potentiating and alkaline phosphatase-inducing activities of *Plantago-mucilage A*, the main mucilage from the seed of *Plantago asiatica*, and its five modification products. *Chemical and Pharmaceutical Bulletin* 39(8): 2068–2071.

#### *Selection of species*

#### **Plantago afra L.**

Sp. pl. ed. 2: 168 (1762).

**Synonyms** *Plantago psyllium* auct. non L. (1753).

**Vernacular names** *Psyllium*, black *psyllium* (En).

**Distribution** Southern Europe, northern Africa, western Asia to Afghanistan and Pakistan; cultivated in India and locally introduced in South-East Asia as medicinal plant, e.g. in Thailand.

**Uses** The mucilage obtained from the seeds is used as laxative.

**Observations** A small annual herb up to 50(–80) cm tall, with well developed erect to slightly ascending stems, upper part of stems with glandular hairs; leaves opposite, linear-lanceolate to linear, 3–8 cm × 0.1–0.3(–0.4) cm, entire or distantly dentate, sparsely covered with short glandular hairs; spike 0.5–1.5 cm long, dense, bracts ovate-lanceolate to lanceolate, 3–8 mm long; fruit about 2 mm long, 2-seeded; seeds narrowly oblong

or narrowly ellipsoidal, 2–3 mm long, smooth and reddish-brown. *P. afra* occurs in dry, open and usually sandy locations.

**Selected sources** 193, 237, 287, 525, 549, 1012, 1249, 1566.

#### **Plantago asiatica L.**

Sp. pl. 1: 113 (1753).

**Synonyms** *Plantago major* L. var. *asiatica* (L.) Decne.

**Vernacular names** Cambodia: slap chravea. Vietnam: m[ax] d[eeff], xa ti[eeff]n.

**Distribution** India, Indo-China, China, Taiwan, Korea, Japan and Siberia.

**Uses** In Indo-China, the seeds are used as diuretic, whereas the whole plant is used to treat coughs and bronchitis. The leaves are applied as a poultice to treat boils and furuncles. The leaves are sometimes consumed as a vegetable.

**Observations** A small perennial herb up to 20 cm tall; leaves in a rosette, broadly ovate to ovate or elliptical-ovate, 5–15 cm × 3–8 cm, entire or obscurely dentate, glabrous or nearly so; spike 2–10 cm long, variably densely flowered, bracts ovate, about 1.5 mm long; fruit 3–4 mm long, 4–6(–12)-seeded; seeds ellipsoidal and plano-convex, (1–)1.5–2 mm long, blackish-brown. *P. asiatica* is closely related to *P. major* and often considered to be conspecific. It may differ in its thinner and slightly broader leaves and on average fewer and larger seeds; however, there is much overlap with the variable *P. major*. It is common in grasslands, along roads and in waste places, especially on poorly drained soils.

**Selected sources** 287, 826, 851, 1035, 1042, 1208, 1439, 1464.

#### **Plantago lanceolata L.**

Sp. pl. 1: 113 (1753).

**Vernacular names** Buckhorn plantain, ribwort (En). Plantain lancéolé, herbe-à-cinq-côtes, bonne femme (Fr). Philippines: lanting-haba (Tagalog). Thailand: phak kaat nam, mo noi (Bangkok), yaa enyuet (northern).

**Distribution** Originally from Europe and western Asia, *P. lanceolata* is now cosmopolitan, including some tropical highland regions.

**Uses** The leaves are applied to wounds, skin inflammations and sores. In India, the seeds are used with sugar as a purgative and haemostatic. The plant is also used in traditional medicine in India to treat headache.

**Observations** A small perennial herb up to 30(–60) cm tall, with a short thick rootstock;

leaves in several rosettes, narrowly lanceolate to linear-lanceolate or narrowly elliptical, (2-)10-25(-40) cm × (0.5-)1-3(-5) cm, entire or remotely and shallowly denticulate, subglabrous or appressed pubescent to villous; spike 0.5-5(-8) cm long, very dense, bracts ovate, about 3 mm long; fruit 3-4 mm long, 2-seeded; seeds boat-shaped, about 2 mm long, smooth, brown. *P. lanceolata* is extremely variable, but much of the variation reflects differences in habitat (e.g. hairy plants in more dry habitats). It is very local in eastern Java (Ijen plateau, in fields and roadsides at about 1500 m altitude) and the Philippines (Luzon, in gardens at about 2200 m altitude).

**Selected sources** 97, 193, 287, 549, 614, 1012, 1178, 1403, 1565, 1566.

### ***Plantago major* L.**

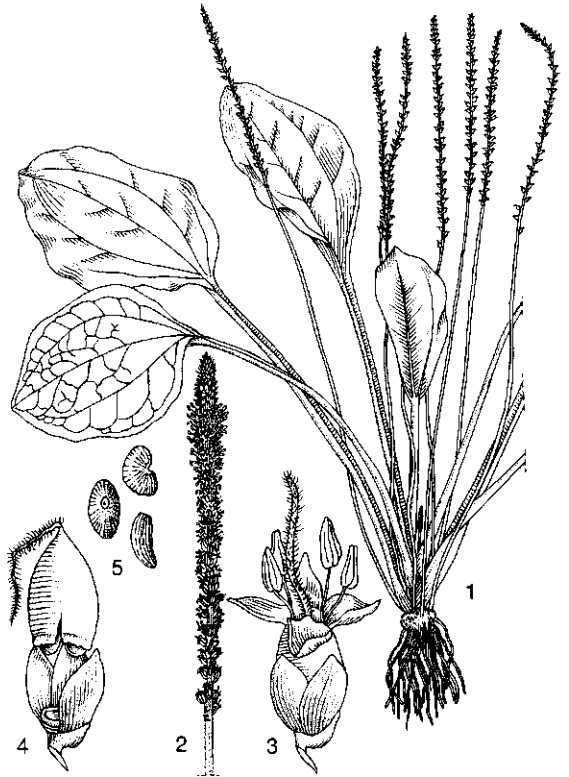
Sp. pl. 1: 112 (1753).

**Vernacular names** Great plantain, waybread, nipple grass (En). Grand plantain, plantain majeur (Fr). Indonesia: daun sendok, daun urat (general), ki urat (Sundanese). Malaysia: ekor anjing (general). Philippines: lanting, lantin, lanting haba (Tagalog), llantin (Spanish).

**Distribution** *P. major* is cosmopolitan; in the tropics it is most common in mountainous regions.

**Uses** The seeds have a great reputation as a remedy for dysentery and diarrhoea. They are considered pectoral, demulcent, quieting, antirheumatic and tonic, and to conduce fertility. In Indonesia, an extract of the whole plant is used as diuretic in cases of renal calculi, often in a mixture with 5 parts of *Clerodendrum*, and also to treat diabetes and skin diseases. Leaves are used to heal wounds and to treat pimples. A decoction of the plant is used in Japan to treat asthma and cough, and of the leaves in the Philippines as emollient. In Thailand, the whole plant or leaves of *P. major* are used as diuretic and antipyretic. The seeds are used as laxative, anti-inflammatory and carminative. The uses of the leaves as diuretic, astringent, and to treat wounds, insect stings and skin diseases are widespread all over the world. Other applications are against malaria (fresh leaf juice or decoction of whole plant), earache (leaves), dysentery (decoction of leaves), burns, contusions and ulcers of the mouth (decoction of leaves), gonorrhoea (decoction of leaves) and as eyewash (decoction of leaves) and mouthwash against inflammation of gums (decoction of leaves).

**Observations** A small perennial herb up to 30(-70) cm tall, with numerous fibrous and



*Plantago major* L. - 1, fruiting plant; 2, inflorescence; 3, flower; 4, dehisced fruit; 5, seeds.

whitish roots; leaves in one or few rosettes, ovate to elliptical, (1.5-)5-30(-40) cm × (0.5-)3-10(-15) cm, entire or irregularly dentate, glabrous or nearly so; spike 5-20(-35) cm long, densely to rather laxly flowered, bracts ovate, 1-2 mm long; fruit 2-4 mm long, (4-)6-34-seeded; seeds ellipsoidal or ellipsoidal-trigonous, 1-1.5 mm long, dark brown to dull black. *P. major* is a variable species in which several subspecies and varieties have been described. However, the different types are often connected by a series of intermediates. It is common in open grasslands and along roads, particularly on more fertile and compact soils, from sea-level (but usually above 700 m) up to 3300 m altitude.

**Selected sources** 30, 97, 190, 193, 202, 287, 317, 332, 350, 545, 549, 580, 614, 851, 1012, 1151, 1178, 1287, 1439, 1566, 1571.

### ***Plantago ovata* Forssk.**

Fl. aegypt.-arab.: 31 (1775).

**Vernacular names** Flea seed, blond psyllium (En).

**Distribution** South-eastern Spain, northern Africa, western Asia to Uzbekistan, Afghanistan and Pakistan; much cultivated in India and locally introduced in South-East Asia as medicinal plant, e.g. in Thailand.

**Uses** The mucilage obtained from the seeds is widely used as laxative. It can also be used as a gelling agent for tissue culture.

**Observations** A small annual or sometimes perennial herb up to 15(-20) cm tall; leaves in one or few rosettes, linear-lanceolate to linear, 2-12 cm  $\times$  0.1-0.8 cm, entire or distantly denticulate, sparsely to densely villous-lanate; spike 0.5-3.5 cm long, dense, bracts suborbicular to ovate, about 3 mm long; fruit about 3 mm long, 2-seeded; seeds boat-shaped, 2-2.5 mm long, smooth and yellowish-brown to pale greyish-pink. *P. ovata* occurs in dry regions, and is cultivated on poor to moderately fertile sandy soils.

**Selected sources** 147, 193, 287, 411, 525, 549, 1012, 1125, 1566.

Lilis Pangemanan

## Plectranthus L'Hér.

Stirp. nov.: 84, pl. 41, 42 (1788).

LABIATAE

$x = 12, 14, 15, 16, 17$ ; *P. amboinicus*:  $2n = 28, 32, 34, 68, 112$ , *P. barbatus*:  $2n = 28, 30, 32, 34$ , *P. scutellarioides*:  $2n = 16, 24, 30, 36, 40, 48, 54, 72$

**Major species** *Plectranthus amboinicus* (Lour.) Spreng., *P. barbatus* Andrews, *P. scutellarioides* (L.) R.Br.

**Origin and geographic distribution** *Plectranthus* comprises approximately 200 species and is distributed in the tropical and subtropical regions of the Old World. The greatest diversity in species is found in Africa. Several species have been introduced and cultivated outside their natural areas of distribution long ago (also in tropical America) and have naturalized, and it is sometimes very difficult or even impossible to deduce their origin. About 15 species occur in the Malaysian region.

**Uses** The most common medicinal uses of *Plectranthus* in South-East Asia are externally for healing wounds, sores, swellings, burns, insect stings, aphtha and haemorrhoids, and internally to treat asthma, bronchitis, cough, dyspepsia, diarrhoea, and as an analgesic. In the African and South American tropics, the uses in local medicine are similar. In the Ayurvedic healing system in India, *Plectranthus* leaves are used to treat asth-

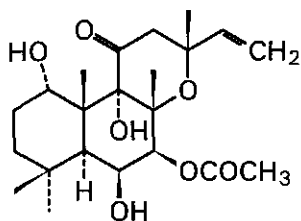
ma, chronic cough, strangury, calculus, gonorrhoea, piles, fever, epilepsy, heart diseases, abdominal colic, dyspepsia, respiratory problems and disorders of the nervous system such as insomnia and convulsions.

In Vanuatu, *P. amboinicus* and *P. scutellarioides* have been suggested to be useful to protect *Cordia alliodora* (Ruiz & Pavon) Oken plantations from *Phellinus noxius* attack. Planting *P. scutellarioides* around taro (*Colocasia esculenta* (L.) Schott) plantations in Samoa controlled the pests *Spodoptera litura* (cluster caterpillar) and *Tarophagus proserpina* (planthopper). The leaves of several *Plectranthus* species and the tuberous roots of *P. barbatus* are used as a spice or condiment. Forms with variegated and often purplish leaves (e.g. of *P. scutellarioides*) are cultivated as ornamental.

**Properties** Steam distillation, hexane extraction and supercritical CO<sub>2</sub> extraction of *P. amboinicus* leaves resulted in respectively 0.5%, 6.5% and 1.5% volatile compounds. Approximately 30 components were identified, with carvacrol (50-90%) as the main constituent. In another experiment, stems and leaves yielded 0.07% of a yellow essential oil after hydrodistillation, in which over 30 different compounds have been identified, with monoterpene hydrocarbons forming the major part (53%). The major components were 3-carene (16%),  $\gamma$ -terpinene (12%), camphor (12%) and carvacrol (13%). The antiseptic activity of *P. amboinicus* has been attributed to the presence of phenolic compounds such as carvacrol in the essential oil. The essential oil has antibiotic activity against numerous gram-positive and gram-negative bacteria. Biochemical studies of the leaf extract revealed the presence of hexacosanol,  $\beta$ -sitosterol, oleanolic acid, betulin and other triterpenoids, whereas the flavones salvigenin, 6-methoxygenkwanin, quercetin, chrysoeriol, luteolin and apigenin, the flavanone eriodyctiol and the flavanonol taxifolin were isolated from leaves of South American origin.

In vitro tests in Burma (Myanmar) with isolated trachea, intestine and uterus segments of guinea-pigs and rats showed that *P. amboinicus* extracts inhibited the contractions of the smooth muscles induced by carbachol, histamine and 5-hydroxytyptamine. At its minimum inhibitory concentration as antimicrobial the extract showed toxicity in the brine shrimp bioassay.

Several abietane diterpenes have been isolated from the leaves of *P. barbatus* grown in Brazil, but the leaves of Kenyan plants afforded highly unsat-



forskolin

urated rearranged abietanes, and the roots of Indian plants furnished polyhydroxylated labdane diterpenes. The steroid stigmasterol has also been isolated. *P. barbatus* contains several diterpenes whose basic skeleton is 11-oxo-nanoyl oxide (8,13-epoxy-labd-14-en-11-one), with forskolin as the chief constituent. Forskolin was first isolated as the active component from the ayurvedic plant *P. barbatus* (synonym *Coleus forskohlii*) by 2 separate research groups almost simultaneously. This led to some confusion in the literature on the name of this compound (forskolin or coleonol) and its absolute structure (does the 7-acetoxy group of the molecule have the  $\beta$ -configuration or  $\alpha$ -configuration?). Later, the identity of the two molecules was demonstrated unambiguously to correspond to the structure given to forskolin. The compound has not been found in *P. amboinicus* and *P. scutellarioides*.

Forskolin has numerous pharmacological actions. The compound has a positive inotropic action on the myocardium, and it exerts an antihypertensive activity by decreasing peripheral vascular resistance. From animal tests it has been concluded that forskolin affects blood flow and platelet parameters favourably in cases of occlusive arterial disease and reconstructive arterial surgery. Preliminary studies in humans have shown that forskolin does indeed increase the contractility of the myocardium, without increasing oxygen consumption, and that it is a vasodilator. It also possesses bronchodilating properties, causes a substantial and lasting decrease in intra-ocular pressure, and has an immunostimulant effect. Forskolin has been demonstrated to strongly inhibit the aggregation of human platelets induced by melanoma cells, showing a potential as an agent to prevent cancer metastasis. An *in vitro* study involving pre-injection of mice with forskolin at a dose of 82  $\mu\text{g}/\text{mouse}$ , followed by tail-vein injection of cultured B16-F10 cells, reduced tumour colonization in the lungs by more than 70%. Giving forskolin orally to alloxan dia-

betic rats caused 37% increase in blood glucose level compared with alloxan diabetic controls, whereas feeding it for 7 days to normal rats raised blood glucose, serum insulin, glucagon and free fatty acid levels, with a corresponding increase in glucose-6-phosphatase activity and depletion of liver glycogen. Forskolin has also shown effects on the thyroid gland (increased secretion, iodine incorporation), the adrenal glands (increased steroid genesis) and the pituitary gland (increased ACTH release).

Much research has been done on the mechanism of action. Forskolin has been found to act by activating membrane-bound adenylate cyclase to cause an increase in cellular cyclic AMP (cAMP) levels. The exact site of action is a direct activation of the catalytic unit of the enzyme. The increase of cAMP e.g. in heart muscle is known to increase its contractility (due to opening of the slow  $\text{Ca}^{2+}$  channels, thus leading to a rise in intracellular calcium). In addition, increase of cAMP in the smooth muscle causes relaxation. This mechanism is probably responsible for the cardiovascular and vascular effects of forskolin. Furthermore, cAMP acts as a second messenger in many receptor-mediated signal transduction systems, e.g. that of the  $\beta$ -adrenergic receptor. Several hormone receptors also regulate their actions via adenylate cyclase reactions, which accounts for many of the effects of forskolin on the hormonal system.

The compound barbatusol has been isolated from Brazilian *P. barbatus* plants. Given intravenously at 3 mg/kg it induced potent lowering of blood pressure associated with discrete bradycardia in rats. The cardioactive dichloromethane crude stem extract of *P. barbatus* yields 20-deoxocarnosol (a phenolic diterpene with an abietane skeleton) and cariocol. The alcoholic extract of *P. barbatus* has been found to inhibit passive cutaneous anaphylaxis in mouse and rat. The extract showed highly significant antisecretory activity against *Escherichia coli* enterotoxin-induced secretory responses in rabbit and guinea-pig ileal loop models. Tests with mice in Brazil showed that water extracts of *P. barbatus* produced mild stimulation of the central nervous system, increased intestinal movements and reduced gastric secretion, indicating an antidyseptic activity and protective activity against gastric ulcers induced by stress. A leaf extract was found active in the *in vivo* test against Ehrlich's ascites tumour in mice. The diterpenoids barbatusin, cyclobutatusin and 3 $\beta$ -hydroxy-3-deoxybarbatusin have been isolated and identified; barbatusin was the major com-

pound. Tumour-inhibitory tests against Lewis lung carcinoma and lymphocytic leukaemia P388 in mice indicated significant inhibitory activity for barbatusin at doses of 200 mg/kg and 400 mg/kg, respectively.

The caffeic acid ester rosmarinic acid has been isolated from *P. scutellarioides*. This compound, which is one of the most abundant caffeic acid esters occurring in plants, is of pharmaceutical interest because of its anti-inflammatory, antiviral, antibacterial and antioxidant properties. A mixture of 6 n-saturated hydrocarbons was isolated from *P. scutellarioides* leaves. The sterol mixture isolated was shown to consist of 4 sterols, with  $\beta$ -sitosterol and stigmasterol as major components. In tests with mice in the Philippines, the leaf juice of *P. scutellarioides* completely inhibited the formation of all types of tumours initiated by dimethylbenzanthracene and promoted by croton oil; 67% of the mice in the control group developed skin tumours, 50% liver tumours and 33% colon tumours. The crude extract showed antibacterial activity and inhibited the growth of Ehrlich ascites tumour cells.

A crude water extract of *P. scutellarioides* showed in vitro fungicidal activity against *Cercospora cruenta*, the pathogen of leafspot of mung bean, and against *Helminthosporium* spp. A crude leaf extract of *P. barbatus* was found to have repellent action on the larvae of the cigarette beetle (*Lasioderma serricorne*), whereas the oil and powder of *P. amboinicus* significantly protected stored maize, sorghum and mung bean seeds from the attacks of the insect pests *Sitophilus zeamais*, *Rhizopertha dominica* and *Callosobruchus chinensis* but reduced germination of mung bean seeds. The oil from *P. amboinicus* has also been found to be topically toxic to red flour beetle, common cutworm, corn weevil, lesser grain borer, black armyworm and housefly, and caused 100% mortality in young *Pomacea* snails at 10–20 ppm in tests in the Philippines.

**Adulterations and substitutes** The phenolic acid rosmarinic acid is also found in many other *Labiatae* (e.g. in *Orthosiphon aristatus* (Blume) Miq., *Rosmarinus* spp. and *Salvia* spp.) and in *Boraginaceae*. Diterpenes are commonly found in *Labiatae*, but they have limited therapeutic applications in pure form. However, some of them have therapeutic potential, just like forskolin from *P. barbatus*, whereas several diterpene-containing drugs are ingredients of phytotherapeutic products or allopathic proprietary drugs. Examples are compounds or drugs from *Ballota*, *Marrubium*,

*Rabdosia*, *Salvia*, *Sideritis* and *Teucrium* species.

**Description** Herbs or undershrubs, aromatic, sometimes with tuberous roots. Leaves opposite, simple, margin serrate, crenate or dentate, petiole, exstipulate. Inflorescence a lax or dense cyme or verticillaster, arranged collectively in a terminal or axillary spurious spike, raceme or panicle, 6-many-flowered. Flowers bisexual, zygomorphic; calyx tubular or campanulate, straight or declinate, with 5-toothed limb, usually 2-lipped with larger and broader upper lip, but teeth sometimes subequal, often accrescent; corolla with long or short tube, decurved or straight, sometimes with a spur or angle on the upper side, limb 2-lipped with short 3–4-fid, recurved upper lip and entire or notched, boat-shaped lower lip, whitish, bluish or purplish; stamens 4, declinate, filaments free or connate at base into a sheath around the style or adnate to the corolla tube but free from each other, anther cells usually confluent; disk prominent, produced anteriorly, about as long as the ovary; ovary superior, style briefly 2-fid. Fruit splitting into 4 orbicular, or occasionally oblong or ovoid nutlets, these smooth, granulate or punctate, 1-seeded.

**Growth and development** *Plectranthus* usually flowers throughout the year. *P. amboinicus* rarely flowers in Malesia. The flowers are insect-pollinated.

**Other botanical information** *Coleus* is often considered as distinct from *Plectranthus* sensu stricto, mainly on the basis of the fused bases of the filaments. However, this is a variable and unreliable character, and in most modern treatments *Coleus* is considered as a synonym of *Plectranthus*. *Solenostemon* is sometimes also kept apart because of the distinctly 2-lipped calyx; however, *Solenostemon* is connected by intermediate structures to *Plectranthus* s.s. (with calyx segments about equal). Here, one large genus, *Plectranthus* sensu lato, including *Coleus* and *Solenostemon* has been assumed, although there is still disagreement about generic delimitation in the complex.

**Ecology** *Plectranthus* species are usually found in open locations, on waste places, roadsides, river banks and thickets, and along cropped fields, but *P. scutellarioides* also occurs in shaded locations and forest vegetation. *P. barbatus* can more readily withstand extended periods of drought than the other species.

**Propagation and planting** *Plectranthus* is usually propagated by stem cuttings, which root readily. Usually, cuttings of 15–20 cm long are

taken from the end of young stems and planted at a spacing of 40 cm × 40 cm. Soaking *P. scutellarioides* cuttings in placobutrazol (up to 25 mg/l) for 1.5–3 days increased the number of roots formed, but strongly inhibited shoot growth for a period of up to 10 weeks.

#### **In vitro production of active compounds**

Cell suspension cultures of *P. scutellarioides* have been established, producing high amounts of rosmarinic acid. In the growth phase, the cell suspension cultures are maintained in bioreactors in modified B5-medium with 2% sucrose, and later the cell mass is diluted into a production medium consisting of a 4–5% sucrose solution, where the cells start to accumulate rosmarinic acid, but grow only slowly. Rosmarinic acid starts to accumulate at the end of the growth phase and continues for only 5 days of the culture period. The accumulation of up to 21% of the cell dry weight as rosmarinic acid makes cell suspension cultures of *P. scutellarioides* among the highest-producing plant cell cultures with respect to secondary product formation.

The synthetic purine derivative 1-(6-puriny)-2,5-dimethylpyrrole has a stimulatory effect on the callus, which is pale green and fluffy and considered suitable for establishing cell suspension cultures; the stimulatory effect is better than that of kinetin. A biosynthetic pathway for rosmarinic acid has been deduced from studies of the enzymes detectable in the cell suspension cultures. Eight enzymatic activities are involved in the transformation of the precursors phenylalanine and tyrosine to the end product rosmarinic acid. Continuous permeabilization of preconditioned cells with dimethyl sulphoxide showed an effective strategy for the enhanced release of rosmarinic acid while preserving cell viability. Product release peaked at 0.5% dimethyl sulphoxide.

Sucrose has a greater stimulative effect on growth and rosmarinic acid accumulation of the culture than glucose and fructose. The rosmarinic acid content in normal growth medium with 2% sucrose is similar to the level in the whole plant (about 2.5% of the cell dry weight). Rosmarinic acid contents of about 20% of the cell dry weight have been found in suspension cultures grown in medium with 4% sucrose. Low phosphate concentrations in the medium result in an increased rosmarinic acid accumulation. Callus and suspension cultures have been cryopreserved and successfully stored for periods up to 15 months, with the best results obtained when using cells from the early growth period.

**Husbandry** Most *Plectranthus* species are easy to cultivate. Seedlings of *P. barbatus* grow well when transplanted in sandy soils.

**Diseases and pests** In India, *P. amboinicus* is recorded as a host of the root-knot nematode *Meloidogyne incognita*, whereas *P. barbatus* is highly susceptible to *Meloidogyne incognita* and *M. javanica*.

**Yield** In the United States sixty *P. barbatus* plants grown from seed yielded 430 g of dried tuberous roots after 9 months, whereas another sixty plants grown from stem cuttings yielded 730 g of dried tuberous roots. About 2 g of forskolin was isolated from 430 g dried tuberous roots.

**Genetic resources and breeding** Most *Plectranthus* species are widespread and common in habitats that are not at risk. All the species described here (except for *P. congestus*) are also commonly planted. This means they are not readily liable to genetic erosion. No selection and breeding is known of, but future work might focus on obtaining large amounts of compounds with interesting medicinal properties. Chemical studies of plant material from different regions showed considerable differences in compounds isolated, providing a basis for selection.

Several other *Plectranthus* species seem to be local endemics which could easily become endangered, e.g. *P. kunstleri* Prain in Peninsular Malaysia, *P. petraeus* Back. ex Adelb. and *P. steenisii* H. Keng in Java, and *P. apoensis* (Elmer) H. Keng, *P. merrillii* H. Keng and *P. sparsiflorus* (Elmer) H. Keng in the Philippines.

**Prospects** *Plectranthus* is extremely interesting medicinally. The efficacy of many of the traditional applications is supported by modern research. Forskolin is thus an example of a new pharmacologically active natural product with a unique mode of action. It is useful not only as a tool in pharmacology but also for the development of, for example, anti-hypertensive or cardioactive drugs based on a novel mechanism of action. The antitumour, antimicrobial and anti-allergenic effects of *Plectranthus* compounds also deserve further attention. The wide distribution and ease of cultivation make some species ideal for commercial exploitation. Although *P. barbatus* is neither indigenous to nor currently planted in South-East Asia, it seems worthwhile to establish experimental plantings for this promising species in the drier regions.

The presence of  $\beta$ -sitosterol and stigmasterol is interesting too. The former is employed in hypercholesterolaemia, and both compounds can be used as



starting material for commercial semisynthesis of steroid hormones, such as corticosterone and antifertility hormones. Generally easy to propagate, *Plectranthus* might be developed as a commercial source of sterols of medicinal and economic importance.

A thorough taxonomical study covering the whole genus complex on a worldwide scale is desirable.

**Literature** |1| Garcia, L.L., Takahashi, M. & Sato, T., 1978. Phytochemical investigation of *Coleus blumei* Benth. II. Identification of the sterol and hydrocarbon constituents. *Philippine Journal of Science* 107(1-2): 95-102. |2| Gupta, S., Yadava, J.N.S. & Tandon, J.S., 1993. Antisecretory antidiarrhoeal activity of Indian medicinal plants against *Escherichia coli* enterotoxin-induced secretion in rabbit and guinea pig ileal loop models. *International Journal of Pharmacognosy* 31(3): 198-204. |3| Kelecom, A., 1983. Isolation, structure determination, and absolute configuration of barbatusol, a new bioactive diterpene with a rearranged abietane skeleton from the labiate *Coleus barbatus*. *Tetrahedron* 39(21): 3603-3608. |4| Keng, H., 1978. Labiatae. In: van Steenis, C.G.G.J. (General editor): *Flora Malesiana*. Series 1, Vol. 8. Sijthoff & Noordhoff International Publishers, Alphen aan den Rijn, the Netherlands. pp. 382-394. |5| Petersen, M., Szabo, E., Meinhard, J., Karwatzki, B., Gertlowski, C., Kempin, B. & Fuss, E., 1995. Biosynthesis and accumulation of rosmarinic acid in suspension cultures of *Coleus blumei*. *Plant Cell, Tissue and Organ Culture* 43(2): 89-92. |6| Pino, J.A., Garcia, J. & Martinez, M.A., 1996. Comparative chemical composition of the volatiles of *Coleus aromaticus* by steam distillation, solvent extraction and supercritical carbon dioxide extraction. *Journal of Essential Oil Research* 8(4): 373-375. |7| Serrame, E. & Lim-Sylianco, C.Y., 1995. Anti-tumor activity of decoctions and expressed juices from Philippine medicinal plants. *Philippine Journal of Science* 124(3): 275-281. |8| Valdés III, L.J., Mislankar, S.G. & Paul, A.G., 1987. *Coleus barbatus* (*C. forskohlii*) (Lamiaceae) and the potential new drug forskolin (coleonol). *Economic Botany* 41(4): 474-483. |9| Vera, R., Mondon, J.M. & Pieribattesti, J.C., 1993. Chemical composition of the essential oil and aqueous extract of *Plectranthus amboinicus*. *Planta Medica* 59(2): 182-183. |10| Zelnik, R., Lavie, D., Levy, E.C., Wang, A.H.J. & Paul, I.C., 1977. Barbatusin and cyclobutatusin, two novel diterpenoids from *Coleus barbatus* Benth. *Tetrahedron* 33(12): 1457-1467.

### *Selection of species*

#### ***Plectranthus amboinicus* (Lour.) Spreng.**

Syst. veg. 2: 690 (1825).

**Synonyms** *Coleus amboinicus* Lour. (1790), *Coleus aromaticus* Benth. (1830), *Coleus carnosus* Hassk. (1842).

**Vernacular names** Country borage, Indian borage (En). Indonesia: daun jinten (general), ajeran (Sundanese), daun kucing (Javanese). Malaysia: bangun-bangun, membangun (Peninsular). Philippines: oregano (Sp), suganda (Tagalog), latai (Subanun). Cambodia: sak dam ray. Thailand: niam hu suea (central), hom duan luang, hom duan hu suea (northern). Vietnam: rau t[aa]f[n], rau t[aa]f[n] d[aa]f[y] l[as], h[us]ng chanh.

**Distribution** Almost pantropical nowadays, assumed to be of Indonesian origin, but possibly originally from Africa; also much planted throughout Malaysia.

**Uses** *P. amboinicus* is considered as wound-healing in Indonesia, and is used to treat sores, fever (externally and internally), asthma and cough (juice or decoction of the leaves), headache (externally) and aphtha (chewing). In Malaysia, a decoction of the leaves is given after childbirth, and the juice to treat cough. Macerated fresh leaves are applied externally to burns and stings of centipedes and scorpions in the Philippines, and also to treat headache, and an infusion of the leaves as a carminative, and to treat dyspepsia and asthma. In Thailand, it is used to treat wounds and to alleviate cough. In Papua New Guinea, cuts, sores and scabies are treated with the sap squeezed from heated leaves. In pharmacy in Indonesia the leaves are known as 'Folia Colei'; they are commonly used to treat thrush and aphtha. In Brazil, *P. amboinicus* is used to treat leishmanial ulcers. In Indo-China, it is used to treat asthma, bronchitis and insect stings, and as a pectoral and vulnerary. In India, it is applied to treat complaints of the genito-urinary system and colics. In Réunion, the leaf juice is used for curing wounds, and an infusion is said to possess anti-influenza properties. The leaves are used in Indonesia and the Philippines as a spice to give fragrance to dishes, and they are also rubbed on the hair and clothes for their scent. In Vietnam, the leaves are often used as a condiment in a popular sour soup, and also in meat dishes and stews, and in India they are eaten raw with bread and butter.

**Observations** A perennial or semi-shrubby, variably succulent herb up to 100(-120) cm tall,

non-tuberous; leaves broadly ovate, suborbicular or reniform, (3–)5–7(–10) cm × (2.5–)4–6(–8.5) cm, thick and fleshy; flowers in dense verticillasters disposed in terminal spike-like inflorescences, calyx subequally 5-toothed, corolla 8–12 mm long, pinkish-lilac to blue; nutlets flattened-globose, about 0.7 mm long, smooth, pale brown. *P. amboinicus* occurs on roadsides, waste places and on river banks up to 1500 m altitude.

**Selected sources** 202, 292, 332, 350, 580, 720, 981, 1035, 1126, 1139, 1178, 1518.

### **Plectranthus barbatus Andrews**

Bot. repos. 9: pl. 594 (1810).

**Synonyms** *Coleus barbatus* (Andrews) Benth. (1831), *Coleus forskohlii* (Willd.) Briq. (1897) non Vahl (1790).

**Distribution** Nepal, Bhutan, India and Sri Lanka; introduced into Madagascar, East Africa and western Asia, and also planted in the Neotropics.

**Uses** In traditional medicine in Brazil, *P. barbatus* is commonly used as an analgesic and to cure liver and stomach diseases. In India it is considered to have anti-allergenic activity. The tuberous roots are eaten as a condiment in India, where they are prepared as a pickle.

**Observations** A perennial semi-succulent herb up to 75 cm tall, with generally decumbent stem rooting at lower nodes and thick tuberous roots; leaves ovate-oblong, 3–9 cm × 2–5 cm, thick; flowers in lax verticillasters disposed in terminal raceme-like inflorescences, calyx 2-lipped, corolla 13–17 mm long, deep purplish-blue; nutlets globose, about 1.2 mm long, granulate, blackish-brown. *P. barbatus* occurs in open, semi-arid areas, on waste places and often on shallow, moist soil among rocks on slopes, up to 1500 m altitude.

**Selected sources** 58, 193, 292, 524, 529, 718, 719, 1277, 1492, 1573, 1646.

### **Plectranthus congestus R.Br.**

Prodr.: 506 (1810).

**Vernacular names** Papua New Guinea: ragi, magwu (Nyamikum, Sepik).

**Distribution** Timor, eastern New Guinea and northern Australia.

**Uses** The sap from crushed leaves has been reported to be applied to sores and scabies in Papua New Guinea.

**Observations** An annual herb, up to 150 cm tall; leaves ovate to elliptical, 2–6 cm × 1.5–4 cm, membranaceous; flowers in dense verticillasters forming false spikes and disposed in terminal pan-

icles, calyx 2-lipped, corolla about 6 mm long, pale blue or lilac; nutlets flattened-globose, about 0.8 mm long, gland-dotted. *P. congestus* occurs in open places, thickets, savanna and on coastal rocks, up to 1500 m altitude.

**Selected sources** 597, 720.

### **Plectranthus scutellarioides (L.) R.Br.**

Prodr.: 506 (1810).

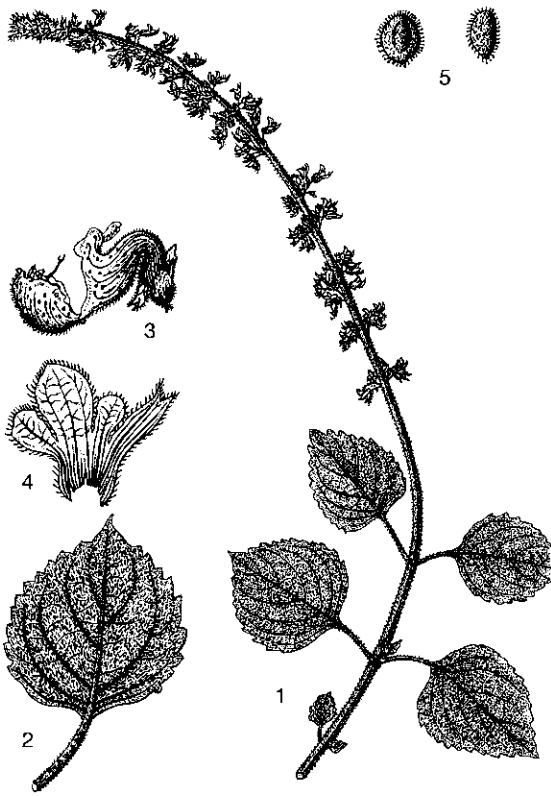
**Synonyms** *Coleus atropurpureus* Benth. (1830), *Coleus scutellarioides* (L.) Benth. (1830), *Coleus blumei* Benth. (1832), *Solenostemon scutellarioides* (L.) Codd (1975).

**Vernacular names** Painted nettle (En). Indonesia: jawer kotok (Sundanese), kentangan (Javanese), adang-adang (Palembang, Sumatra). Malaysia: daun ati-ati, ati-ati merah, ati-ati besar (Peninsular). Papua New Guinea: jangata (Morawaka, Eastern Highlands), jeune (Agenehemo, Northern Province), okavu (Kami, Eastern Highlands). Philippines: badiara, malaina, mayana (general). Thailand: ruese phasom laeo (central), waan lueat haeng (Chiang Mai). Vietnam: t[aa]y.

**Distribution** India, Burma (Myanmar), Indo-China, southern China, Taiwan, Thailand, throughout Malesia, the Solomon Islands, northern Australia and Polynesia; often cultivated, also outside this region.

**Uses** The roots of *P. scutellarioides* are used internally in the Moluccas to treat diarrhoea and colic, and the leaves as anthelmintic and to treat urinary complaints, whereas sap is squeezed into the eye in the case of eye injury, and rubbed on swellings. Elsewhere in Indonesia the sap or a decoction is used as an abortivum and emmenagogue, and to treat haemorrhoids, inflamed eyes and boils. In Malaysia, a decoction of the leaves is used to stimulate digestion, as a sedative, to treat dyspepsia and congestion of the liver, and externally against swellings, smallpox and ophthalmia. Fresh leaves are applied in the Philippines externally or in cataplasm to bruises and contusions, and to treat headache. In Papua New Guinea, young leaves are baked and squeezed whilst hot onto fresh cuts and sores. *P. scutellarioides* is commonly cultivated for its ornamental purplish foliage.

**Observations** An erect or ascending, branched perennial herb up to 150 cm tall, non-tuberous; leaves generally ovate, 1–15 cm × 1–10 cm, membranaceous; flowers in lax verticillasters or in irregularly branched cymes disposed in simple or branched thyrses, calyx 2-lipped, corolla about



*Plactranthus scutellarioides* (L.) R. Br. — 1, flowering stem; 2, leaf; 3, flower; 4, opened calyx; 5, nutlets.

8–13(–18) mm long, blue or violet with whitish tube; nutlets broadly ovoid or globose, 1–1.2 mm long, shining, brown. *P. scutellarioides* occurs in all kinds of habitats, from rain forest to cropped fields and thickets, and from the lowland to 2900 m altitude.

**Selected sources** 140, 202, 292, 332, 471, 472, 477, 580, 628, 720, 1114, 1126, 1129, 1178, 1310.

Mulyati Rahayu

## Plumbago L.

Sp. pl. 1: 151 (1753); Gen. pl. ed. 5: 75 (1754).

PLUMBAGINACEAE

$x$  = unknown; *P. indica*:  $2n = 14$ , *P. zeylanica*:  $2n = 28$

**Major species** *Plumbago indica* L., *P. zeylanica* L.

**Vernacular names** Leadwort (En). Vietnam: du[oo]i c[oo]ng.

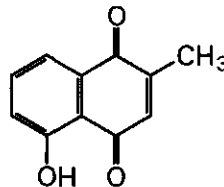
**Origin and geographic distribution** Plum-

*bago* consists of about 24 species from tropical and warm temperate regions. For the Malesian region 4 species are reported, 2 of which are indigenous to the region.

**Uses** *P. indica* and *P. zeylanica* are widely considered a vesicant and abortifacient, and are further used in the treatment of rheumatism and skin problems. Although both are generally reported to have similar applications, *P. zeylanica* is said to be milder and less dangerous than *P. indica*. Some *Plumbago* species are widely cultivated ornamentals in tropical and subtropical regions. In the Malesian region, *P. aphylla* Bojer ex. Boiss. originating from Madagascar and *P. auriculata* Lamk (synonym: *P. capensis* Thunb.) from southern Africa are planted as ornamentals.

**Properties** Dried roots of *Plumbago* are often the basis of the drugs used in traditional medicine and are found as such in the market. They are traded in pieces of usually less than 1.3 cm thick, with a shrivelled, yellowish to reddish-brown bark. The activity of preparations of *P. indica* and *P. zeylanica* can be largely attributed to the presence of plumbagin (2-methyl-5-hydroxy-1,4-naphthoquinone), which is mainly extracted from the roots and is only found in *Plumbaginaceae*. The *Plumbago* species from which plumbagin has been isolated not only include *P. indica* and *P. zeylanica*, but also *P. auriculata*, *P. caerulea* Humb., Bonpl. & Kunth, *P. europaea* L., *P. pearsonii* L. Bolus, *P. pulchella* Boiss. and *P. scandens* L. Analysis of dried and powdered *P. zeylanica* roots from Sri Lanka revealed the presence of naphthoquinone derivatives such as plumbagin (0.036%), isoshinanolone (0.035%), droserone (0.0013%) and 1,2(3)-tetrahydro-3,3'-biplumbagin (0.005%), and the steroid sitosterol (0.08%).

Plumbagin has been reported to have anti-implantation and abortifacient activity in rats, without having teratogenic effects, whereas it produces testicular lesions and testis weight reduction in dogs. It has shown antigonadotropic activity in rats, causing a decrease in weight of ovaries, and blocking the effect of applied gonadotrophin. It has also shown antiprogestational activity in rats.



plumbagin

Furthermore, in lower concentrations, plumbagin has an antimitotic activity comparable to that of colchicine. In larger doses, plumbagin also has nucleotoxic and cytotoxic effects. It has also been found that *P. indica* extracts have inhibitory activity on indirect mutagens and are not mutagenic themselves.

In small doses, plumbagin stimulates the central nervous system of frogs, mice and rabbits, whereas larger doses lead to convulsions and paralysis. In rabbits, it caused a decreased respiration and blood pressure, but no diuresis. In frogs, it paralysed muscular tissue and caused dilation of perfused blood vessels. Furthermore, it seems that plumbagin is a strong irritant and/or hepatotoxic. Plumbagin has shown antibacterial activity against both gram-positive (e.g. *Staphylococcus*, *Streptococcus*, *Pneumococcus* spp.) and gram-negative (e.g. *Salmonella*) bacteria, whereas it is also active against certain fungi (*Trichophyton*, *Epidermophyton* and *Microsporium* spp.) and protozoa (*Leishmania*). It has been found to prevent *Escherichia coli* and *Staphylococcus aureus* developing resistance to antibiotics and it eliminated multidrug-resistant plasmids from *E. coli* strains, resulting in loss of resistance to the antibiotics tested. In low concentrations, it augments the bacterial activity of mouse macrophages against *Staphylococcus aureus*, whereas at higher concentrations it has shown inhibitory effects. Furthermore, plumbagin shows antitumour activity in mice, especially in combination with gamma radiation.

Plumbagin also has insecticidal activity. It has shown strong insect antifeedant activity against larvae of army worms (*Spodoptera exempta*, *S. littoralis*), toxicity to nymphs of red cotton bug or cotton stainer (*Dysdercus koeningii*) and mosquito larvae (*Culex quinquefasciatus*), whereas it retarded growth, delayed metamorphosis and reduced fertility in housefly (*Musca domestica*). It also produced morphogenetic effects in *Dysdercus koeningii* and various other insects. These effects are probably due to its interference with the neuro-endocrine system and the system which regulates metamorphosis.

Besides plumbagin, phytochemical analysis has revealed the presence of other naphthoquinones and their derivatives in the roots of *P. zeylanica* including 3-chloroplumbagin and 2-methylnaph-tazarin, the plumbagin dimers 3,3'-biplumbagin, 3,6'-biplumbagin (= chitranone), 6,6'-biplumbagin (= elliptinone), maritinone, methylene-3,3'-dip-lumbagin, zeylanone and isozeylanone, and the

plumbagin trimer plumbazeylanone. Further compounds include: nonyl-nonanoate and nonyl-8-methyl-dodec-7-enoate (long-chain aliphatic esters), benzyl-2,5-dihydroxy-6-methoxybenzoate and 2,2-dimethyl-5-hydroxy-6-acetyl-chromene (aromatic derivatives), steroids (e.g. stigmasterol, stigmasterol acetate, sitosterone) and triterpenes (lupeol, lupeol acetate, lupanone, friedelinol). The leaves of *P. zeylanica* are reported to contain leucodelphinidin and quercetin-3-rhamnoside, and the petals azaleatin-3-rhamnoside. Compounds isolated from the aerial part of *P. indica* include 6-hydroxyplumbagin, plumbaginol (a flavonol), leucodelphinidin and steroids (e.g.  $\beta$ -sitosterol, stigmasterol, campesterol). The petals are reported to contain delphinidin, cyanidin and pelargonidin-3-rhamnosides, kaempferol-3-rhamnoside, galloyl-glucose and digalloylglucose.

**Adulterations and substitutes** *P. indica* and *P. zeylanica* are sometimes used as a substitute for *Rauvolfia serpentina* (L.) Benth. ex Kurz. Different naphthoquinone derivatives have been prepared from plumbagin, with halogenated plumbagin showing stronger ichthyotoxicity than plumbagin itself.

**Description** Shrubs or perennial herbs, rarely annual, often straggling or subscandent. Leaves alternate, simple, entire, older ones often pale lepidote underneath as a result of carbonate exudations, or reduced in flowering stems, petiole often semi-amplexicaulous auriculate; stipules absent. Inflorescence composed of 1-flowered spikelets grouped in a terminal raceme or spike, often united in a leafy panicle. Flowers subtended by 2-3 bracts, 5-merous; calyx tubular, 5-ribbed, often with sessile or stalked glands outside, teeth erect, not enlarged in fruit; corolla funnel-shaped, lobes spreading, blue, red, white or violet; stamens 5, free, filaments broadened at the base; ovary superior, 1-locular, style 1, stigma lobes 5. Fruit a 1-seeded capsule, included in the calyx and corolla, often splitting from the base with 5 valves. Seed long, narrowly ovoid, slightly flattened, tapering towards apex, dark brown or black, surface colliculate, hilum small, oval, in a longitudinal depression.

**Growth and development** In South-East Asia, both *P. indica* and *P. zeylanica* flower throughout the year, which explains their success as ornamentals. No fruit of *P. indica* has ever been found. The fruits of *P. zeylanica* are easily dispersed because of the sticky glands on the persistent calyx.

**Other botanical information** *Plumbago* is

placed together with *Ceratostigma* in the subfamily *Plumbaginoideae*, characterized by inflorescences consisting of spikes, racemes or heads, as opposed to the other subfamily *Staticoideae* (inflorescence a thyrse with cincinnate partial inflorescences) comprising e.g. *Armeria* and *Limonium* which are both mainly saltmarsh and maritime genera. *P. europaea* from the Mediterranean region and West Asia used to be used in traditional medicine to treat eye diseases. One of the side-effects of this rather poisonous plant is a discoloration of the skin resembling the colour of lead, from which the Latin name *Plumbago* and the popular name leadwort are derived.

**Ecology** In general, *Plumbago* prefers semi-arid conditions. *P. indica* is reported to be a short-day plant. It prefers rich, moist and well-drained soils, with a pH between 5.5 and 6.0, whereas a pH below 5.0 or above 7.0 may lead to stunted growth. In Indian experiments with *P. zeylanica*, the largest and heaviest roots were obtained from plants grown in loamy soils.

**Propagation and planting** Shrubby *Plumbago* species are usually propagated through cuttings. *P. indica* is propagated through offsets and root cuttings. In vitro micropropagation from stem segments, nodes and leaves is also possible. However, in vitro plants may not transplant successfully into a soil medium.

**In vitro production of active compounds** *P. zeylanica* cell strains derived from stem explants and grown in vitro have been found to have a different potential of plumbagin synthesis.

**Handling after harvest** Leaf poultices of *P. zeylanica* can be dried and stored for several months without losing their vesicant properties. However, dried roots of *P. indica* and *P. zeylanica* contain less plumbagin and show less activity than fresh ones.

**Genetic resources and breeding** In view of their wide distribution, neither *P. indica* nor *P. zeylanica* appear to be endangered. No breeding programmes are known to exist.

**Prospects** Although plumbagin may have medicinal potential, e.g. for its antimicrobial and anti-tumour activity, the use of plumbagin or plumbagin-containing plant material as medicine for humans is not devoid of danger. Plumbagin may have potential as a compound in synthetic insecticides.

**Literature** [1] Devi, P.U., Solomon, F.E. & Sharada, A.C., 1994. In vivo tumor inhibitory and radiosensitizing effects of an Indian medicinal plant, *Plumbago rosea* on experimental mouse tu-

mors. *Indian Journal of Experimental Biology* 32(8): 523-528. [2] Dhar, S.K. & Rao, P.G., 1995. Hormonal profile of plumbagin. *Fitoterapia* 66(5): 442-446. [3] Gunaherath, G.M.K.B., Gunatilaka, A.A.L. & Thomson, R.H., 1988. Studies on medicinal and related plants of Sri Lanka. Part 18. Structure of a new naphthoquinone from *Plumbago zeylanica*. *Journal of the Chemical Society. Perkin Transactions 1, Organic and Bio-organic Chemistry* 1988: 407-410. [4] Gupta, M.M., Verma, R.K. & Gupta, A.P., 1995. A chemical investigation of *Plumbago zeylanica*. *Current Research on Medicinal and Aromatic Plants* 17: 161-164. [5] Harborne, J.B., 1967. Comparative biochemistry of the flavonoids - IV. Correlations between chemistry, pollen morphology and systematics in the family *Plumbaginaceae*. *Phytochemistry* 6: 1415-1428. [6] Heble, M.R., Narayanaswamy, S. & Chadha, M.S., 1974. Tissue differentiation and plumbagin synthesis in variant cell strains of *Plumbago zeylanica* L. in vitro. *Plant Science Letters* 2: 405-409. [7] Karnick, C.R., Tiwari, K.C. & Majumber, R., 1982. Cultivation trials, pharmacognosy and ethnobotanical investigations of *Plumbago zeylanica* L. (*Chitraka*) of the Indian system of medicine. *International Journal of Crude Drugs Research* 20: 193-199. [8] Valsaraj, R., Pushpangadan, P., Smitt, U.W., Adersen, A. & Nyman, U., 1997. Antimicrobial screening of selected medicinal plants from India. *Journal of Ethnopharmacology* 58: 75-83. [9] van Steenis, C.G.G.J., 1949. *Plumbaginaceae*. In: van Steenis, C.G.G.J. (General editor): *Flora Malesiana*. Series 1, Vol. 4. Noordhoff-Kolff N.V., Djakarta, Indonesia. pp. 107-112. [10] Wilmot-Dear, C.M., 1976. *Plumbaginaceae*. In: Polhill, R.M. (Editor): *Flora of Tropical East Africa*. Crown Agents for Oversea Governments and Administrations, London, United Kingdom. 12 pp.

#### *Selection of species*

#### **Plumbago indica L.**

Stickm. Herb. Amb.: 24 (1754).

**Synonyms** *Plumbago rosea* L. (1762).

**Vernacular names** Rosy-flowered leadwort, officinal leadwort (En). Indonesia: akar binasa (Malay, Moluccas), ceraka merah (Malay), mehulatu (Ambon). Malaysia: cheraka merah, setaka. Philippines: laurel (Bikol, Tagalog), pampasapit (Tagalog). Burma (Myanmar): chuvondacovailie, kanchopni, kenkhyokeni. Thailand: chettamun phloeng daeng (central), pit piu daeng (northern),

fai tai din (peninsular). Vietnam: du[oo]i c[oo]ng, x[is]ch hoa x[af], du[oo]i c[oo]ng hoa d[or].

**Distribution** Tropical Africa, tropical Asia and the Pacific region. Common throughout South-East Asia but not reported for New Guinea and Borneo; reports for Peninsular Malaysia are contradictory. Widely cultivated in other tropical and subtropical regions.

**Uses** In Indonesia, a poultice of the roots is used as a remedy for rheumatism, paralysis, tumours, toothache (as a counter-irritant) and swollen glands. The root-bark is used as a vesicant. To this end, the fresh root is cut into very thin slices, which are tied on the skin. Similarly these slices may also be applied to the forehead against headache. The leaves are also used in the treatment of rheumatism and headache. Locally, *P. indica* is used as an abortifacient, either internally by topical application for its vesicant properties, or by chewing the roots for some time. In veterinary medicine, it is given as a vermifuge to horses only. In Malaysia, leaves and roots are externally applied in the treatment of rheumatism, glandu-

lar swellings and leprosy. In the Philippines, the bark is used as vesicant and the roots are employed in poultices to treat headache. The bark is also said to be an antidyspeptic. In Thailand, the dried root is credited with emmenagogue, stomachic and carminative activities, and it is reported to be used in the treatment of haemorrhoids, as an abortifacient and as a means to purify the blood and stimulate digestion. *P. indica* is widely cultivated for its ornamental value.

**Observations** A shrub up to 1.5 m tall, branched from the base, stems drooping, sometimes rooting; leaves oblong, 5–15 cm × 2–8 cm, petiole not auriculate; inflorescence a rather sparsely flowered spike, not corymbose, rachis glabrous, 10–30 cm long; flowers with calyx about 1 cm long, covered in glands, red, corolla tube 2.5–4 cm long, lobes 2–3 cm in diameter, distinctly mucronate, red; fruit unknown. *P. indica* is found in the vicinity of (former) anthropogenic localities, locally semi-spontaneous, often persistent in abandoned cultivation, also in teak forest, up to 1000 m elevation.

**Selected sources** 92, 97, 190, 193, 202, 252, 287, 350, 354, 355, 556, 572, 580, 754, 793, 900, 1035, 1044, 1068, 1122, 1128, 1159, 1178, 1254, 1284, 1285, 1287, 1476, 1507.

### *Plumbago zeylanica* L.

Sp. pl. 1: 151 (1753).

**Vernacular names** Ceylon leadwort, white-flowered leadwort (En). Indonesia: daun encok (general), bama (Javanese, Balinese), ki encok (Sundanese). Malaysia: jarak, cheraka. Philippines: bangbang (Ilokano), sangdikit (Tagalog), talankan (Ilokano). Laos: pit pi' khao. Thailand: chettamun phloeng khaoo (central), pit piu khaoo (northern). Vietnam: du[oo]i c[oo]ng tr[aws]ng, b[a]ch hoa x[af], du[oo]i c[oo]ng hoa tr[aws]ng.

**Distribution** Tropical Africa, tropical Asia and the Pacific region. Common throughout South-East Asia, although not reported for Borneo and the Moluccas.

**Uses** In Indonesia, crushed leaves are applied as a poultice to treat rheumatism and headache (never longer than a few minutes behind the ears). The crushed leaves may also be used as a substitute for cantharides ('Spanish fly'); though more painful they have the advantage of not irritating the uro-genital organs. In Bali, *P. zeylanica* is used to treat ringworm. Roots of *P. zeylanica* are also used internally, for their vesicant properties, as an abortifacient. In Malaysia, a decoction of the plant is reported to be taken during the first three



*Plumbago indica* L. – 1, plant habit; 2, roots; 3, flower.

days after childbirth, probably for its tonic properties. Furthermore, chewing the leaves or a decoction is used as an emmenagogue, but may well act as an abortifacient in early pregnancy. In the Philippines, an infusion of the roots is used against itch. The pounded roots are applied externally as a vesicant, and root decoctions are used to treat scabies, whereas the roots are also said to be ecibolic. In Indo-China, the roots and leaves are considered rubefacient and vesicant, and they are used as a poultice against rheumatism. Leaf and root infusions are applied as a wash to treat itch and skin diseases. In Java, *P. zeylanica* is planted in living fences.

**Observations** A creeping herb or scandent shrub up to 2.5 m tall, laxly branched; leaves ovate, ovate-lanceolate, elliptical or oblong, 2.5–13 cm × 1–6 cm, base cuneate, apex acute or mucronate, glabrous, often with white waxy dots below, petiole 2–12 mm long, base amplexicaul, sometimes auriculate; inflorescence a 6–30 cm long raceme, rachis with sessile glands; flowers with a green glabrous calyx, covered in stalked glands, corolla tube 1.7–2.6 cm long, lobes 0.6–1 cm × 0.3–0.5 cm, white, scented, anthers purple, ovary and style glabrous. Seed 6 mm × 2 mm, dark brown. In general preferring a dry season climate, *P. zeylanica* is almost exclusively found in rather open anthropogenic habitats, e.g. savanna, forest edges and fallow fields, up to 1000 m altitude.

**Selected sources** 2, 92, 97, 190, 193, 202, 287, 350, 381, 516, 518, 519, 520, 521, 572, 580, 715, 754, 785, 786, 807, 900, 921, 1035, 1122, 1126, 1128, 1154, 1178, 1200, 1210, 1280, 1281, 1293, 1353, 1476, 1507, 1581.

Wongsatit Chuakul, Noppamas  
Soonthornchareonnon & Promjit Saralamp

## Premna L.

Mant. pl. 2: 154, 252 (1771).

VERBENACEAE

*x* = unknown

**Major species** *Premna herbacea* Roxb., *P. odorata* Blanco, *P. serratifolia* L., *P. tomentosa* Willd.

**Origin and geographic distribution** The number of species in *Premna* varies from 50–200, depending on literature sources. They are widely distributed in the African, Asian and Australian tropics and subtropics. The main distribution area extends from India, Indo-China, China and Japan, through Thailand and the Malesian region to northern Australia. *P. serratifolia* has the largest

area of distribution, extending from eastern Africa to the whole of southern and eastern Asia (including Malesia) and northern Australia.

**Uses** Infusions or extracts of the leaves and roots of *Premna* spp. are widely used in South-East Asian traditional medicine as a diuretic to treat dropsy, as a carminative and stomachic, to treat diarrhoea, bronchial affections, rheumatism and headache, and as a febrifuge and tonic (e.g. after childbirth). In the Philippines, a decoction of the leaves of *P. odorata* is used for loosening phlegm and as a cough remedy.

The timber is sometimes used if trees have reached larger dimensions (particularly *P. tomentosa*). Some *Premna* spp. are used in hedges, occasionally also as ornamentals. Ripe fruits and cooked leaves are occasionally eaten.

**Production and international trade** Plant parts of *Premna* are only collected for local use and are not traded on a larger scale.

**Properties** Premnazole (an isoxazol alkaloid) has been isolated from the leaves of *P. serratifolia*. This compound has anti-inflammatory activity. In rats, it has reduced granuloma formation with an activity comparable to phenyl butazone and has lowered the weight of adrenal glands and their ascorbic acid content. In the same experiment, the enzyme activities of acid phosphatase, glutamate pyruvate transaminase (GPT, ALAT) and glutamate oxaloacetate transaminase (GOT, ASAT) were reduced in serum and liver, whereas the protein content was lowered in serum. The terpenoid compound  $\beta$ -sitosterol and the flavonoid luteolin have also been isolated from the leaves of *P. serratifolia*. The major alkaloid in the stem bark of *P. serratifolia* was found to be aphelandrine (a spermine alkaloid) which occurs together with two other alkaloid compounds. Several sesquiterpenoids and diterpenoids have been isolated from the root bark, e.g. caryophyllen-3-one, premnenol. Phytochemical investigations of the leaves of *P. odorata* (methanol extract) have revealed the presence of six iridoid-glycoside-caffeoyl esters, two of them having a monoacyl-rhamnopyranosyl-catapol structure and four being diacyl-rhamnopyranosyl-catapols (named premnosides A–D). Premnaodoroside A–C (acyclic monoterpenediol-iridoidglucoside diesters verbascoside, decaffeoyl verbascoside and isoverbascoside) were also isolated from the same extract.

The leaves of *P. tomentosa* have been shown to contain limonene, caryophyllene and other di- and sesquiterpenoids. The flavone 6,8-di-C-glycoside has been isolated from the heartwood. Root nod-

ules of *P. herbacea* have been reported to contain a diterpenoid quinone, methide. The heartwood yields the flavonoid vicenin 3.

Several African *Premna* species (e.g. *P. schimperi* Engl., *P. oligotricha* Baker) contain diterpenes, which showed antibacterial and cytotoxic effects in vitro.

An extract of *P. serratifolia* showed antifeedant and anti-ovipositional effects on *Callosobruchus chinensis*, which is a pest of stored crop products. However, the action persisted for no longer than one week. The water extract of the stem bark exhibits myocardial depressant, skeleton and smooth muscle stimulant, uterine stimulant and hypoglycaemic activity.

**Description** Perennial herbs or undershrubs, shrubs (sometimes lianescent) to small or medium-sized trees up to 20 m tall; bole up to 30(-50) cm in diameter, often with fissured flaky bark. Leaves decussately opposite, simple, entire or serrate, crenate or dentate in upper half, with reticulate tertiary venation, with foetid smell when crushed, petiolate or sessile, exstipulate. Inflorescence terminal, cymose, compound and often much branched, pedunculate; bracts small. Flowers small, bisexual, zygomorphic, usually white, greenish-white or yellowish, pedicellate or subsessile; calyx tubular or somewhat campanulate, variously toothed or truncate, often almost 2-lipped with one lip entire or 2-toothed, the other entire or 3-toothed, persistent; corolla tubular below with usually densely villous tube in upper half, almost 2-lipped above, usually with entire upper lip and 3-lobed lower lip; stamens 4, didynamous, inserted at about the middle of the corolla tube, filaments filiform, anthers dorsifixed; ovary superior, 4-locular, style filiform, with 2 short stigmatic lobes. Fruit a small globose or obovoid succulent drupe with a hard 4-celled kernel, green but usually turning dark purple or black at maturity, 1-4-seeded. Seeds without endosperm. Seedling with epigeal germination; cotyledons thin-coriaceous, hairy; hypocotyl elongate, hairy, epicotyl present.

**Growth and development** Many *Premna* species flower all year round. Fruits mature a few months after flowering.

**Other botanical information** *Premna* is closely related to *Gmelina*, but it can easily be distinguished by its short and cylindrical corolla tube; in *Gmelina* the corolla tube is long and ampliate above. *Vitex* is another closely related genus.

*Pygmaepremna* is often kept separate from *Premna*, but the only reliable differential charac-

ter is the dwarf habit, which does not seem to justify a distinct generic status.

Some *Premna* spp. have minor or obscure medicinal uses. In Peninsular Malaysia, leaves of *P. divaricata* Wallich ex Schauer have been used to treat headache and colds. *P. parasitica* Blume has been used in Indonesia after childbirth and as fish poison and in Peninsular Malaysia as febrifuge. *P. cordifolia* Roxb. has been used to treat shortness of breath and as febrifuge. The leaves of *P. nauseosa* Blanco have been used in the Philippines to treat stomach-ache.

**Ecology** *Premna* species usually occur in the lowland up to 1000 m altitude, in secondary forest, open forest, brushwood and hedges. *P. serratifolia* often grows near the sea. *P. herbacea*, which is a subshrub, often occurs in grasslands exposed to periodic fires.

**Propagation and planting** *Premna* is rarely planted and, if so, it is probably propagated by seed.

**Harvesting** The plant parts of *Premna* like leaves and roots needed to make medicinal preparations are collected from the wild.

**Handling after harvest** The collected leaves and roots are washed and dried in the sun. An infusion or extract is made when needed.

**Genetic resources and breeding** The *Premna* spp. treated here are usually common in secondary forest, scrub vegetation and sometimes even in grassland, and are not easily liable to genetic erosion. *P. odorata* is endemic to the Philippines, but occurs there abundantly and is not threatened; *P. cumingiana* and *P. peekelii* also have limited areas of distribution but do not seem to be immediately endangered. However, several species in South-East Asia appear to be narrow endemics and might easily be endangered.

**Prospects** Very little pharmacological research has been done, and little is known about active compounds of *Premna* species. There is so little information available that no statements about prospects in modern medicine are warranted. However, the few studies done indicate that the therapeutic properties attributed to several *Premna* species may have a scientific basis.

**Literature** |1| Barik, B.R., Bhowmik, T., Dey, A.K., Patra, A., Chatterjee, A., Joy, S., Susan, T., Alam, M. & Kundu, A.B., 1992. Premnazole, an isoxazole alkaloid of *Premna integrifolia* and *Gmelina arborea* with anti-inflammatory activity. *Fitoterapia* 63(4): 295-299. |2| Dasgupta, B., Sinha, N.K., Pandey, V.B. & Ray, A.B., 1984. Major alkaloid and flavonoid of *Premna integrifolia*.



Planta Medica 50(3): 281. |3| de Padua, L.S., Ludwig, G.C. & Pancho, J.V., 1978. Handbook on Philippine medicinal plants. Vol. 2. Documentation and Information Section, Office of the Director of Research, University of the Philippines at Los Baños, the Philippines. p. 49. |4| Habtemariam, S., 1995. Cytotoxicity of diterpenes from *Premna schimperi* and *Premna oligotricha*. Planta Medica 61: 368–369. |5| Holdsworth, D.K., 1977. Medicinal plants of Papua New Guinea. South Pacific Commission, Noumea, New Caledonia. p. 51. |6| Munir, A.A., 1984. A taxonomic revision of the genus *Premna* L. (Verbenaceae) in Australia. Journal of the Adelaide Botanic Gardens 7(1): 1–44. |7| Nguyen Van Duong, 1993. Medicinal plants of Vietnam, Cambodia and Laos. Mekong Printing, Santa Ana, California, United States. p. 426. |8| Otsuka, H., Kubo, N., Yamasaki, K. & Padolina, W.G., 1989. Premnosides A–D: diacyl 6-O- $\alpha$ -L-rhamnopyranosylcatalpols from *Premna odorata*. Phytochemistry 28(11): 3063–3067. |9| Otsuka, H., Kashima, N., Hayashi, T., Kubo, N., Yamasaki, K. & Padolina, W.G., 1992. Premnaodorosides A, B and C, iridoid glucoside diesters of an acyclic monoterpenediol from leaves of *Premna odorata*. Phytochemistry 31(9): 3129–3133. |10| Quisumbing, E., 1978. Medicinal plants of the Philippines. Katha Publishing Co., Quezon City, the Philippines. pp. 798–801.

#### Selection of species

#### *Premna cumingiana* Schauer

DC., Prodr. 11: 634 (1847).

**Synonyms** *Premna cardiophylla* Schauer (1847).

**Vernacular names** Philippines: magilik (Tagalog), manaba (Bikol, Bukidnon), banaba (Ibanag).

**Distribution** The Philippines and Sulawesi.

**Uses** An infusion of the leaves is reported to be used in the Philippines as a diuretic to treat dropsy.

**Observations** A small tree up to at least 8 m tall; leaves broadly ovate to subrotundate, 10.5–30(–42) cm  $\times$  8–20(–25) cm, densely tomentose with branched or stellate hairs, particularly beneath, petiolate; flowers with white corolla; fruit subglobose, about 5 mm long. *P. cumingiana* occurs mainly in secondary forest up to 1000 m altitude.

**Selected sources** 1178.

#### *Premna herbacea* Roxb.

Fl. Ind. (Carey ed.) 3: 80 (1832).

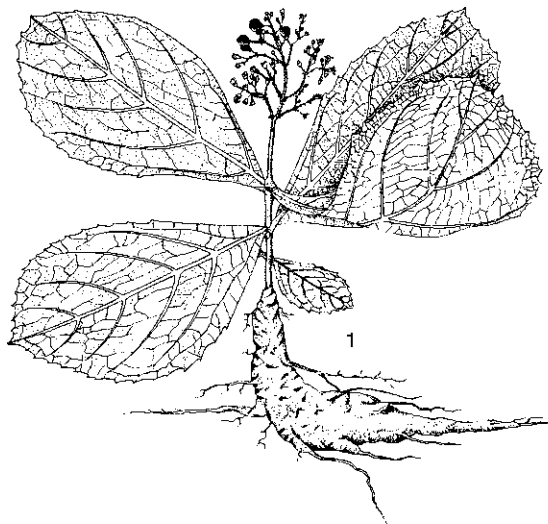
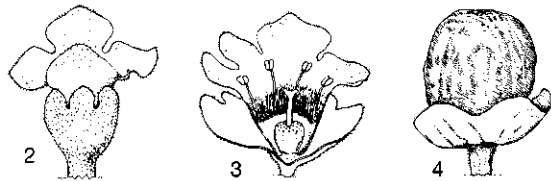
**Synonyms** *Pygmaeopremna humilis* Merr. (1910), *Pygmaeopremna herbacea* (Roxb.) Moldenke (1941).

**Vernacular names** Thailand: khang hua lek (Chiang Mai), phaen din yen (Chiang Rai), som kang (Prachin Buri). Vietnam: c[as]ch c[or].

**Distribution** From Pakistan, Nepal and India, through Burma (Myanmar), Indo-China, southern China, Thailand, the Malesian region (the Philippines, the Lesser Sunda Islands (Sumba), Sulawesi, New Guinea) to Australia; cultivated as a medicinal plant in India and Sri Lanka.

**Uses** The juice from roots and rhizomes is used in India to treat dropsy, cough, asthma, fever, rheumatism and cholera. Ripe fruits are occasionally eaten in northern India.

**Observations** A low-growing perennial herb or a dwarf undershrub up to 15(–30) cm tall, most of the stem underground with creeping woody rhizome, the aboveground part slender and simple or with single dichotomous branching; leaves obo-



*Premna herbacea* Roxb. – 1, plant habit; 2, flower; 3, opened flower; 4, fruit.

vate, (2.5-)5-12(-15) cm × (2-)3-7(-10) cm, dentate or crenate-undulate in the upper half, subglabrous, sessile or subsessile, lowest pair lying flat on the ground; flowers with pedicel 1.5-2 mm long, corolla white or greenish-white; fruit obovoid-globose, 5-8(-10) mm long, green turning black. *P. herbacea* is particularly characteristic of open grasslands exposed to periodic fire.

**Selected sources** 972, 991.

### ***Premna odorata* Blanco**

Fl. Filip.: 488 (1837).

**Synonyms** *Premna curranii* H.J. Lam (1919), *Premna benthamiana* Domin (1929), *Premna inaequilateralis* E. Beer & H.J. Lam (1936).

**Vernacular names** Philippines: alagao (general), agdau (Pangasinan), anobran (Iloko).

**Distribution** From Nepal, India, Burma (Myanmar), Indo-China, China, Taiwan and Japan, through Thailand and probably the whole of Malesia, to Australia; sometimes planted (e.g. in the Philippines, Java, India and Florida).

**Uses** The leaves are used as diuretic; a decoction with sugar and a little 'calamansi' (*Citrofortunella microcarpa* (Bunge) Wijnands) juice is taken to treat cough, and an infusion is considered carminative and useful to treat beri-beri. A decoction of leaves or flowers is considered to be a febrifuge and is also used against abdominal pains and dysentery. Masticating the roots is prescribed against cardiac troubles.

**Observations** A shrub or small tree up to 10(-12) m tall, with bole up to 25(-30) cm in diameter, bark finely flaky fissured; leaves ovate to ovate-rotundate, sometimes ovate-lanceolate, (5-)6.5-16(-20) cm × 4-10(-13.5) cm, mostly entire, sometimes serrate-denticulate in upper half, pubescent all over, especially beneath, petiolate; flowers with pedicel 0.5-1.5 mm long, corolla greenish-white or pinkish-white; fruit obovoid, 3-5 mm long, green turning black. *P. odorata* has been much confused with *P. pubescens* Blume, which is a liana or climbing shrub. *P. odorata* is common in thickets and secondary forest at low altitudes.

**Selected sources** 327, 332, 991, 1091, 1092, 1093, 1178.

### ***Premna peekelii* H.J. Lam**

Verben. Malay. Archip.: 115 (1919).

**Vernacular names** Papua New Guinea: kua (New Britain).

**Distribution** New Britain.

**Uses** In Papua New Guinea, young leaves are

rubbed on the cheek of patients with mumps.

**Observations** A small tree up to 6 m tall; leaves broadly ovate to rotundate, 9-22 cm × 6-16 cm, entire or crenate-toothed, densely tomentose beneath, petiolate; fruit globose, 4-6 mm long. *P. peekelii* is common in forest and scrub vegetation.

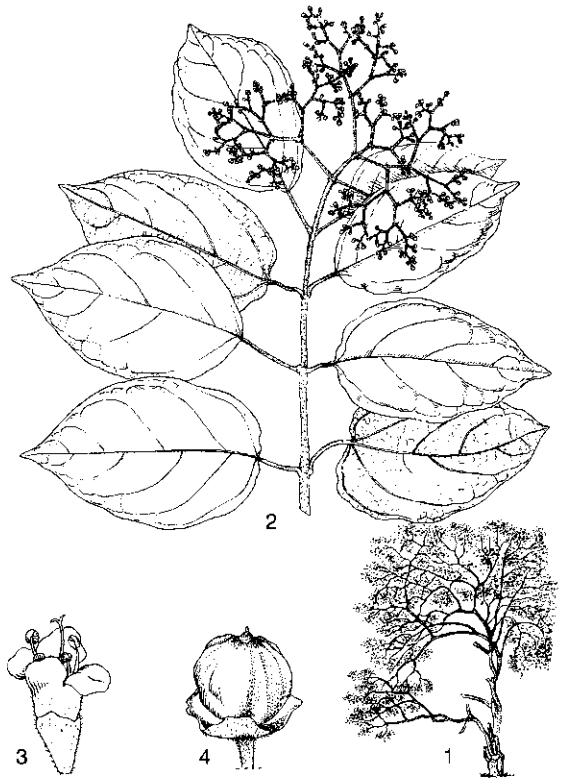
**Selected sources** 597.

### ***Premna serratifolia* L.**

Mant. pl. 2: 253 (1771).

**Synonyms** *Premna integrifolia* L. (1771), *Premna corymbosa* Rottl. & Willd. (1803), *Premna obtusifolia* R.Br. (1810), *Premna foetida* Reinw. ex Blume (1826).

**Vernacular names** Indonesia: ki pahang (Sundanese), singkil alas (Javanese), daun kambing (Moluccas). Malaysia: buas-buas, pokok buru hantu (Peninsular). Papua New Guinea: kalokalo (Sipupu, Milne Bay), ninggrp (Yenchimangua, Sepik), lavakaliu (New Ireland). Thailand: cha lueat (Trat), man kai (Lampang), akkhe thawaan thale (peninsular). Vietnam: c[aa]y c[as]ch, v[o]ng c[as]ch.



*Premna serratifolia* L. - 1, tree habit; 2, flowering twig; 3, flower; 4, fruit.

**Distribution** Madagascar, Mauritius, East Africa, and from India, Bangladesh, Indo-China, China, Taiwan and Japan, through Thailand and the whole Malesian area (at least reported from Peninsular Malaysia, Java, the Moluccas, New Guinea and the Philippines) to northern Australia, Polynesia and Melanesia.

**Uses** The leaves and roots are used in traditional medicine in Indo-China as a diuretic, stomachic and febrifuge. The leaves are employed as a galactagogue in India and Indonesia, and also to treat rheumatic arthritis, colic and flatulence in India. A decoction of roots and leaves is used as a febrifuge in Peninsular Malaysia. Extracts of the leaves are used to treat cough, headache and fever in Papua New Guinea. A tea made from the boiled bark is used in Guam to treat neuralgia. The wood is used for implements and paddles, the bark as binding material. Cooked leaves are eaten as vegetable. The plant is used locally in hedges.

**Observations** A shrub or small tree up to 10 m tall, bole up to 30 cm in diameter, much-branched and sometimes spiny, bark fissured-flaky, brownish-grey; leaves broadly ovate, obovate to suborbicular, sometimes oblong, (2-)8-15(-21) cm × (1-)5-10(-16) cm, mostly entire, sometimes serrate, crenate or dentate, glabrous, petiolate; flowers with pedicel 0.5-1 mm long, corolla greenish-white; fruit obovoid-globose, 3-6 mm long, green turning black. *P. serratifolia* is extremely variable, particularly the leaves and calyces. It grows in brushwood and hedges, often near the sea.

**Selected sources** 97, 126, 202, 316, 359, 580, 597, 972, 991, 1035.

### ***Premna tomentosa* Willd.**

Sp. pl. 3(1): 314 (1800).

**Synonyms** *Premna pyramidata* Wallich (1827), *Premna flavida* Miq. (1861).

**Vernacular names** Indonesia: bulang (Javanese, Sundanese), leban capo, bebulang handak (Sumatra). Malaysia: bebuas, sarang burong, tembaroh (Peninsular). Papua New Guinea: garogira (Papa, Central Province). Thailand: kapiat (Prachuap Khiri Khan), sak khe kai (northern), saam paang (Chanthaburi).

**Distribution** Nepal, India, Sri Lanka, Burma (Myanmar), Indo-China, Thailand and the Malesian region (at least Peninsular Malaysia, Sumatra, Java and Timor).

**Uses** A decoction of the roots and leaves has been used in Malaysia as a tonic in mixtures after childbirth. The bark was used in Indonesia to treat diarrhoea. In Papua New Guinea, the leaves

are crushed and applied to sores. The leaves are reputed to have diuretic properties, and they are used externally to treat dropsy. In Thailand, the dried entire plant is used to soothe skin irritation caused by caterpillars. The leaves have been used in veterinary medicine against maggots in wounds. The timber is used for house building and furniture, and also for carving and turnery.

**Observations** A shrub or small to medium-sized tree up to 20 m tall, with bole up to 30(-50) cm in diameter, often crooked and fluted at base, bark fissured or striate and shaggy, grey or yellowish to pale brown or pale rusty; indumentum consisting of branched or stellate hairs; leaves ovate or ovate-orbicular to ovate-oblong, (3-)10-35 cm × (2.5-)6-22 cm, entire, densely tomentose beneath, petiolate; flowers with pedicel 0.5-1 mm long, corolla white, greenish-white or yellowish; fruit obovoid-globose, 3-6 mm long, green turning black. *P. tomentosa* closely resembles *P. odorata*, but differs in the branched or stellate hairs. It occurs frequently in open forest and deciduous forest, in Java up to 800 m altitude.

**Selected sources** 97, 202, 580, 972.

L.B. Cardenas

### ***Pueraria montana* (Lour.) Merr.**

Trans. Amer. Philos. Soc. n.s. 24(2): 210 (1935).

LEGUMINOSAE

2n = 22, 24

**Synonyms** Var. *lobata*: *Pueraria thunbergiana* (Sieb. & Zucc.) Benth. (1867), *Pueraria lobata* (Willd.) Ohwi (1947).

**Vernacular names** Kudzu, Japanese arrowroot (En). Koudzou (Fr). Indonesia: bitok, tobi (Madurese), tebi (Kangean). Philippines: baai (Igorot), tahaunon (Manobo). Laos: chũa tau kung, khauz pièd (northern). Thailand: tamyakhrua, phakpeetpe. Vietnam: s[aw]n d[aa]ly, cl[as]t c[aw]n.

**Origin and geographic distribution** *Pueraria* consists of 16 species of Asian origin. *P. montana* has a large area of distribution, ranging from eastern India, Burma (Myanmar), Indo-China, China, Korea and Japan, through Thailand and the Malesian region, to the Pacific islands and northern Australia. It was successfully introduced into South America and the southern United States, but did not become established in Africa. The commonest variety in the Malesian region is var. *lobata* (Willd.) v.d. Maesen & Almeida; it has been reported in Peninsular Malaysia, the Philip-

pinus, the Lesser Sunda Islands, the Moluccas and Papua New Guinea.

**Uses** In Chinese medicine the tuber of kudzu is known as 'Radix Puerariae', and it is one of the most important crude drugs. Tea from the tubers is used in China and Indo-China against colds, fever, influenza, diarrhoea, dysentery and hangovers. The flower buds are used as a diaphoretic and febrifuge. In China, its clinical use for various diseases in internal medicine, surgery, pediatrics and dermatology has been reported. The most important efficacy is for arrhythmia. The starch from the tuber is used medicinally in Japan in soup or tea to restore intestinal and digestive disorders. The extract is effective in lessening alcohol intoxication.

Kudzu tuber is esteemed for its fine starch, used especially in China, Japan and Papua New Guinea for sauces, soups, jelled salads, noodles, porridges, jelly puddings, confectionary and beverages. Elsewhere in South-East Asia the tubers are used in times of famine. The stem fibres are used for binding (ropes), weaving (clothes, fishing lines, baskets) and for paper production. The young leaves, shoots and flowers may be consumed as a vegetable. Kudzu is excellent for fodder and silage, if mixed with grass. It is effective for erosion control, provided its growth is controlled well. Its aggressive growth may lead to entire forests being covered and trees dying, as has been experienced in the United States. Kudzu is a good shade plant and also popular as an ornamental climber with fragrant flowers.

**Production and international trade** Japan produces over 300 t/year of kudzu tubers.

**Properties** *P. montana* var. *lobata* has a high flavonoid content. In a methanol extract of the tuber 7 isoflavones were identified and quantified: puerarin (160 mg/g extract), daidzin (22 mg/g), genistin (3.7 mg/g), daidzein (2.6 mg/g), daidzein-4',7-diglucoside (1.2 mg/g), genistein (0.2 mg/g) and formononetin (0.2 mg/g).

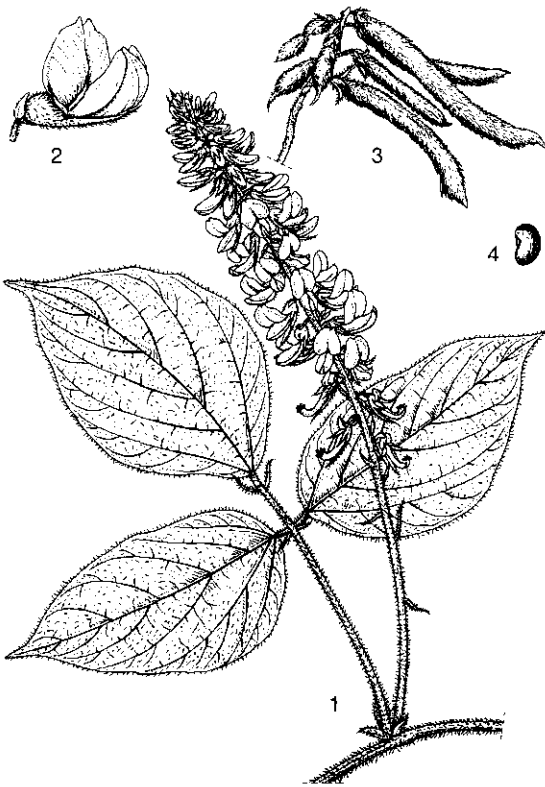
In tests with rats the plant showed antipyretic and anti-myocardial ischemia effects. The isoflavonoid glycosides have antioxidant, anti-hepatotoxic activity and also hypotensive effect, with excellent clinical results in the treatment of hypertension. Chinese pharmacologists have reported that the isoflavonoids stimulate cerebral and coronary blood circulation. Daidzein has been found to show a papaverine-like musculotropic action. The spasmolytic activity of daidzein has been proved using excised small intestine of mice. Puerarin acts as a  $\beta$ -adrenoreceptor antagonist in iso-

lated arteries and veins. The extract showed antidiaprotropic activity (suppression of ethanol intake) in golden hamsters, for which daidzin is the major active principle. This activity of the extract is greater than that of pure daidzin, and it seems that additional constituents in the methanol extract assist uptake of daidzin. The bioavailability of daidzin in the crude extract is about 10 times greater than that of the pure compound. In tests with rats, daidzin was efficacious in lowering blood alcohol levels and shortened sleeping time induced by alcohol ingestion. Daidzein and puerarin have also been effective in suppressing voluntary alcohol consumption by rats, but induced increased water intake. The compounds did not affect the activities of liver alcohol dehydrogenase and aldehyde dehydrogenase, from which it appears that the reversal of alcohol preference produced by these compounds may be mediated via the central nervous system. Antifebrile activity of the extract has been demonstrated in Vietnam. An aqueous extract of flowers and tubers showed wormicidal effect on *Clonorchis sinensis*.

Acute toxicity of each fraction of the tuber is very weak. High-performance liquid chromatography methods have been established for the determination of puerarin for the quality control of Chinese medicinal preparations.

Per 100 g, cooked leaves contain approximately water 89 g, protein 0.4 g, fat 0.1 g, carbohydrates 9.7 g, fibre 7.7 g and ash 0.8 g. Kudzu is nearly as nutritious as alfalfa (*Medicago sativa* L.) and is palatable to all types of livestock. The green forage contains 14.5–20% crude protein, 2–3.5% fat, 27–36% crude fibre and 7–8.5% ash on dry weight basis.

**Description** A perennial, woody climber, with stems up to 30 m long and up to 10 cm in diameter, initially grey to brown pubescent, later glabrescent, and with very large oblong tubers up to 2 m long and up to 45 cm in diameter. Leaves alternate, pinnately 3-foliolate; petiole 8–13(–21) cm long, rachis 1.5–7 cm long, both grey to golden-brown hairy, stipules peltate, up to 1.5(–2.5) cm long; leaflets ovate to orbicular, 8–26 cm  $\times$  5–22 cm, lateral leaflets oblique and often somewhat smaller than terminal leaflet, entire to 3-lobed, thinly appressed pubescent, petiolules 4–10 mm, stipules linear to lanceolate, up to 2(–3) cm long. Inflorescence a usually unbranched elongated pseudoraceme up to 35 cm long, with 3 flowers per node, bracts up to 10 mm long, early caducous, bracteoles up to 5 mm long, fairly persistent. Flowers bisexual, short-pedicelled; calyx campan-



*Pueraria montana* (Lour.) Merr. - 1, part of flowering stem; 2, flower; 3, part of infructescence; 4, seed.

ulate with 5 unequal teeth, tube 3–5 mm long, teeth 4–9 mm long; corolla papilionaceous, petals up to 2.5 cm long, purplish to blue or pink, often with a yellow or green spot on vexillum; stamens 10, monadelphous or with one free stamen; ovary superior, elongated, 1-celled. Fruit a flattened oblong pod, 4–13 cm × 0.5–1.5 cm, straight to falcate, with golden-brown hairs, 5–15-seeded. Seeds flattened ovoid, 4–5 mm × 4 mm × 2 mm, red-brown with black mosaic. Seedling with epigeal germination; first 2 leaves simple and opposite.

**Growth and development** Kudzu may grow 35 m or more in a single season. Bees have been reported to act as pollinators, and kudzu is said to be cross-pollinated. Outside its native area of distribution seed set is often poor.

**Other botanical information** Three varieties are distinguished within *P. montana*, of which var. *lobata* (Willd.) v.d. Maesen & Almeida is particularly common in the Malesian region. This variety is often considered as a distinct species: *P. lobata* (Willd.) Ohwi. The other two varieties, var.

*montana* and var. *chinensis* (Ohwi) v.d. Maesen & Almeida (synonym: *P. lobata* (Willd.) Ohwi var. *thomsonii* (Benth.) v.d. Maesen), are mainly restricted to mainland Asia, although both have been reported from the Philippines. The main distinguishing characteristics are flower size, leaflet form and fruit size.

Extracts from *P. tuberosa* (Roxb. ex Willd.) DC., which does not occur in South-East Asia, showed anti-implantation activity in female rats. Its tubers are used in local medicine in Nepal, Pakistan and India, e.g. against renal complaints, as a febrifuge, as a cataplasm to cure swellings of joints and as a galactagogue; they are also used as a fish poison. In Thailand, the tubers of *P. candollei* Grah. ex Benth. var. *mirifica* (Airy Shaw & Suvat.) Niyomdham (synonym: *P. mirifica* Airy Shaw & Suvat.) are used as a tonic and aphrodisiac, to treat mammary gland expansion and for their oestrogenic effect. Many flavonoids (daidzein, daidzin, genistein, genistin, kwakhurin, mirificin, miroestrol) as well as coumarins (columestrol, mirificoumestan and mirificoumestan hydrate) have been isolated. Pharmacological studies have shown oestrogenic, anti-implantation, abortifacient, antifertility, antispermatogenic and hypercalcaemic effects. Studies on the effects on birds found accelerated growth but inhibited egg-laying. The fertility in both male and female mice was effectively controlled by an aqueous extract of leaves, whereas the extract could effectively interrupt pregnancy.

**Ecology** *P. montana* occurs in thickets, forests, roadsides, pastures and hedges, and is common in the lowlands but is found up to 2000 m altitude. It grows on a wide range of soils, but does not grow well on poor sandy soils and poorly drained heavy clays. It is intolerant of waterlogging, and grows best on well-drained fertile loams. Kudzu is drought resistant because of its deep roots.

**Propagation and planting** Kudzu is propagated by seed, except in regions outside its native area where propagation is mainly by planting young stem cuttings almost horizontally. The very hard seed coat should be scarified with acid or by mechanical means before planting; a germination rate of 70% is considered excellent. Seeds are planted in a nursery in rows 1 m apart and 0.5–1.5 cm deep, and should be inoculated with the cowpea type of rhizobia. Seedlings are transplanted into the field after about 4 months when they have developed 4–6 leaves. Seed may be sown directly into the field, in rows 2 m apart; 1 kg of seed per ha is needed.

**In vitro production of active compounds**

The biosynthesis of isoflavonoids in elicitor-treated cell suspension cultures of kudzu has been studied at the enzyme level. The main secondary metabolites produced by cell cultures are daidzin and puerarin. Addition of yeast extract to the cell culture stimulates the accumulation of isoflavones and daidzein-dimers.

**Husbandry** At planting, kudzu is fertilized. Plantings should be kept free of weeds during the first year.

**Diseases and pests** Fungal diseases in kudzu include leaf-spot (caused by *Alternaria* spp.), anthracnose (caused by *Colletotrichum* sp.), stem rot (caused by *Fusarium* sp.), and damping-off (caused by *Pellicularia solani*), whereas bacteria (*Pseudomonas* spp.) may cause blight. Nematodes (mainly *Meloidogyne* spp.) have been reported to attack the roots. Velvetbean caterpillars (*Anticarsia gemmatilis*) eat the leaves.

**Harvesting** Tubers of kudzu can be harvested about 1 year after planting the cuttings. If left longer in the soil they can become very large. For fodder production, the first harvest is possible in the second year, but full production is reached from the third year onwards.

**Yield** Tubers of kudzu may weigh up to 180 kg when old. A forage yield of 5 t/ha can be expected from good stands on fertile soil.

**Handling after harvest** A good method of preparing a decoction from kudzu is reported from China: the tuber is cut into slices of 4–7 mm, water is added at 12–15 times the weight of the tuber, and the mixture is decocted for 30 minutes.

**Genetic resources and breeding** Thanks to its large area of distribution, kudzu is not at risk of genetic erosion. Few cultivars have been developed, and breeding activities have mainly focused on its value as a forage. In Puerto Rico, crosses have been made between *P. montana* and *P. phaseoloides* (Roxb.) Benth. in order to combine the more vigorous growth of the former with the better adaptation to tropical conditions of the latter.

**Prospects** In China, the wide application of *P. montana* var. *lobata* has been recommended because the flavonoid fraction increases the blood flow to the brain and heart, decreases oxygen consumption by the myocardium and exerts spasmolytic activity. Moreover, clinical trials have shown the applicability of isoflavonoids in the treatment of hypertension, angina pectoris, myocardial infarction and migraines. Furthermore, kudzu is a valuable forage, yields useful fibre from

the stems and starch from the roots, and can be used for erosion control and soil improvement. Its occasional behaviour as a weed which is difficult to eradicate acts as a brake on its wide application for planting as a multipurpose crop.

**Literature** [1] Bruneton, J., 1995. Pharmacognosy, phytochemistry, medicinal plants. Technique & Documentation Lavoisier, Paris, France. p. 298. [2] Duke, J.A., 1981. Handbook of legumes of world economic importance. Plenum Press, New York and London. pp. 211–214. [3] Groen, L.E., Siemonsma, J.S. & Jansen, P.C.M., 1996. *Pueraria lobata* (Willd.) Ohwi. In: Flach, M. & Rumawas, F. (Editors): Plant Resources of South-East Asia No 9: Plants yielding non-seed carbohydrates. Backhuys Publishers, Leiden, the Netherlands. pp. 178–179. [4] Harada, M. & Ueno, K., 1975. Pharmacological studies on *Pueraria* root 1. Fractional extraction of *Pueraria* root and identification of its pharmacological effects. Chemical and Pharmaceutical Bulletin 23(8): 1798–1805. [5] Keung, W.M., Lazo, O., Kunze, L. & Vallee, B.L., 1996. Potentiation of the bioavailability of daidzin by an extract of *Radix puerariae*. Proceedings of the National Academy of Sciences of the United States of America 93(9): 4284–4288. [6] Nguyen Van Duong, 1993. Medicinal plants of Vietnam, Cambodia and Laos. Mekong Printing, Santa Ana, California, United States. pp. 320–321. [7] Ohshima, Y., Okuyama, T., Takahashi, K., Takizawa, T. & Shibata, S., 1988. Isolation and high performance liquid chromatography (HPLC) of isoflavonoids from the *Pueraria* root. *Planta Medica* 54(3): 250–254. [8] Quisumbing, E., 1978. Medicinal plants of the Philippines. Katha Publishing Co., Quezon City, the Philippines. pp. 427–428. [9] van der Maesen, L.J.G., 1985. Revision of the genus *Pueraria* DC. with some notes on *Teyleria* Backer (Leguminosae). Wageningen Agricultural University Papers 85-1: 1–132. [10] van der Maesen, L.J.G., 1994. *Pueraria*, the kudzu and its relatives, an update of the taxonomy. In: Sørensen, M. (Editor): Proceedings of the First International Symposium on Tuberous Legumes, Guadeloupe, F.W.I., 21–24 April 1992. Jordbrugsforlaget, København. pp. 55–86.

**Other selected sources** 202, 580.

Praptiwi

## Quisqualis L.

Sp. pl., ed. 2, 1: 556 (1762).

COMBRETACEAE

$x$  = unknown; *Q. conferta*:  $2n = 26$ , *Q. indica*:  $2n = 22, 24, 26$

**Major species** *Quisqualis indica* L.

**Vernacular names** Indonesia: udani. Malaysia: akar dani, selimpas (Peninsular). Philippines: niyog-niyogan. Thailand: lep mue naang. Vietnam: qu[ar] giun.

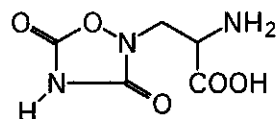
**Origin and geographic distribution** *Quisqualis* has about 14 species, 6 in tropical and South Africa, and 8 in tropical Asia of which *Q. indica* is now commonly cultivated throughout the tropics. In Malesia there are 4 species.

**Uses** The bitter half-ripe fruits and seeds of *Q. indica* are widely known as being anthelmintic and are extensively used as such, usually in decoction, particularly to treat ascariids. In large doses they cause nausea, vomiting, hiccough and even unconsciousness. The seed of the dried ripe fruit is used preferably to reduce the incidence of vomiting, but roots (*Q. conferta*, *Q. indica*) and decoctions of the leaves (*Q. conferta*) are also used as a vermifuge. Although the seeds are often applied to stop diarrhoea, an oil extracted from the seed has purgative properties. Leaf juice or seeds, macerated in oil, are applied externally to treat boils, ulcers and parasitic skin infections. Various preparations of the plant are applied both externally and internally for pain relief. In the Philippines, *Quisqualis* is used as a remedy for coughs, and the fruits and seeds are applied to alleviate nephritis. In Vietnam, the root is used in decoction to treat rheumatism. In Papua New Guinea, plants are eaten daily by men and women as a method of birth control.

*Q. indica* is widely cultivated as an ornamental climber, planted in hedges. In Indonesia, very young shoots are eaten raw or steamed. In West Africa, the long, flexible stems are used for basketry, fish weir and fish traps.

**Production and international trade** The dried fruits of *Q. indica* can be found in small drug stores and Chinese pharmacies throughout Malesia. Most of the fruits on sale in Malaysia and Thailand are imported from China.

**Properties** The seeds of *Q. indica* contain L-quisqualic acid (=  $\beta$ -(3,5-dioxo-1,2,4-oxodiazolidin-2-yl)-L-alanine or (S)- $\alpha$ -amino-3,5-dioxo-1,2,4-oxodiazolidine-2-propanoic acid), which exhibits marked anthelmintic activities. This active principle somewhat resembles the actions of the an-



quisqualic acid

thelmintic  $\alpha$ -santonin (e.g. from *Artemisia maritima* L.); in China, *Quisqualis* seeds are used as a substitute for  $\alpha$ -santonin as drug. In screening tests, parts of the fruit, e.g. the gum isolated from it, have failed to exhibit anthelmintic activity.

Furthermore, quisqualic acid has shown excitatory effects on cultured neurons, and in a variety of animal models (e.g. snails, chicks, mice and rats). It causes various types of limbic seizures and neuronal necrosis. Thus, in neuropharmacology this compound is known as one of the excitatory amino acids (EAAs) and, besides the study of the neurological effects mentioned, is used to identify a specific set of EAA receptors, in the case of quisqualic acid known as quisqualate receptors.

Pharmacological investigation of the chloroform fraction of a hot aqueous water extract of *Q. indica* showed that this fraction inhibits cyclic AMP phosphodiesterase by about 80%.

*Q. indica* extracts showed a mild repellent effect to the oviposition of oriental fruit fly (*Darcus dorsalis*). The aqueous extract of *Q. indica* showed antifungal properties against *Drechslera oryzae* through inhibition of germ tube elongation. The extract of the seeds of *Q. indica* had anticoccidial effect against *Eimeria tenella* in chicken.

**Description** Woody climbers or occasionally scrambling shrubs. Leaves opposite or subopposite, simple, margin entire, with domatia on the lower surface; base of petiole persisting as spines after leaf-fall. Flowers in elongated, usually unbranched, terminal or axillary, bracteate spikes, bisexual, actinomorphic or slightly zygomorphic, 5-merous; receptacle (calyx tube) hairy to nearly glabrous, divided into a lower part surrounding the ovary and a tubular upper part terminating in the calyx lobes, upper receptacle caducous, calyx lobes triangular; petals inserted on the receptacle tube and much larger than the calyx lobes; stamens 10, in 2 rows, inserted inside and near the mouth of the upper receptacle, not or scarcely exerted; disk narrowly tubular or absent; ovary inferior, 1-locular with 2-4 ovules, style adnate to the upper receptacle for at least half the length of the latter. Fruit a dry pseudocarp, 5-winged or -ridged, dehiscent or indehiscent, 1-seeded. Seed without endosperm.

**Growth and development** The lianescent stem of *Q. indica* twines to the left. Plants maintain themselves by root suckers and stooling. The flowers, which open at dusk, are initially white but gradually turn red during the next day. In the meantime the orientation of the flower changes from obliquely upwards or horizontal to pendulous. At night the white flowers are visited by hawk moths, during the day the pink and red flowers are visited by a wide range of pollinators such as solitary bees, honey bees, flies and sunbirds. Each flower lasts 3 days; the largest amount of nectar is present at the morning of the first day. In Vietnam, ripe fruits of *Q. indica* are available in August and September. Fruiting plants are rare in many localities. The fruits of *Quisqualis* are buoyant in both fresh water and seawater, and are thus dispersed.

**Other botanical information** *Quisqualis* is closely related to *Combretum* and the most recent view is that both should be united. *Quisqualis* was considered distinct from *Combretum* by having a dehiscent fruit and by the long tubular upper receptacle. These characters proved to be unreliable for the separation of the two genera. Another proposed delimitation based on the insertion of the style in the upper receptacle (adnate to the wall in *Quisqualis*, free in *Combretum*) and the length of the stamens (exserted in *Combretum*, not exserted in *Quisqualis*) also proved untenable. The junction of the two genera seems obvious, but has not yet been put into practice in literature. The name *Combretum* has priority over *Quisqualis*.

**Ecology** *Q. conferta* occurs in margins of primary forest at low altitudes. *Q. indica* is occasionally found in the same habitat, but more often in more disturbed habitats such as secondary forest, thickets, along streams, and even as a weed along roadsides, on waste places, in rice fields and along railway tracks. It occurs from sea-level up to 300 m altitude, preferably in full sunlight, on a wide range of soils, but preferably on well-drained soils.

**Propagation and planting** *Q. indica* can be propagated by leafless stem cuttings with at least 3 nodes; after 1 month an adequate root system has developed. Cuttings in coarse sand show about 55% rooting success. Good results have also been obtained using the tips of twigs placed in a mist bed after their leaves have been cut in half. *Q. indica* is also propagated by air layering and root division. Propagation from seed is easy, but fruits and seeds are seldom formed. In the Philippines, *Quisqualis* is planted at a spacing of 2–3 m × 4 m with 1.5 m high trellises along the rows.

**Diseases and pests** A leaf spot disease probably caused by a fungus has been very destructive in the Philippines in both the wet and dry seasons; older lesions are mostly circular and dark with distinct chlorotic halos. *Cercospora* leaf spot also causes irregular dark brown spots on the upper leaf surface, in which a clear whitish centre develops. In the Philippines, mites, several lepidopterous insects and also scarabaeid beetles have severely infested *Q. indica*. The bag-worm moth *Eumeta crameri* has become a serious pest on ornamental plants in Dacca, Bangladesh. In India, larvae of the moth *Othreis homaena* caused heavy defoliation of *Q. indica*. The nematode *Xiphinema americanum* was found infecting *Q. indica* and was associated with swollen root-tips.

**Harvesting** Mature fruits are ready for picking when they have turned golden-yellow.

**Handling after harvest** Fruits can be collected half-ripe, when they are still bitter, pulped in water and the liquid can be taken internally as a drug. Mature fruits are air-dried for a month in shallow containers with screened bottoms and stirred constantly to attain a moisture content of less than 10%. Immature fruits should be checked and any containing small holes, which are caused by oviposition of *Ephestia* sp., must be discarded as these will contaminate all fruits within 6 months after storage. Dried fruits can be stored for up to 1 year, but the effect of storage on the quisqualic acid content is not yet known.

**Genetic resources and breeding** No information is available on germplasm collections and breeding programmes.

**Prospects** The prospects for *Q. indica* seeds as an anthelmintic are limited, due to the toxic side-effects of quisqualic acid. The cultivation of *Q. indica* for its anthelmintic properties is being encouraged in the Philippines. However, it is not yet widely planted, possibly due to its toxic side-effects. Moreover, reports from Thailand state that fruiting is not common. Quisqualic acid is of interest for neuropharmacological applications, but there will be no need for large-scale production.

**Literature** [1] Council of Scientific and Industrial Research, 1969. The Wealth of India: a dictionary of Indian raw materials and industrial products. Raw materials. Vol. 8. Publications and Information Directorate, New Delhi, India. pp. 357–358. [2] Eisikowitch, D. & Rotem, R., 1987. Flower orientation and color change in *Quisqualis indica* and their possible role in pollinator partitioning. Botanical Gazette 148: 175–179. [3] Fang, S.D., Xu, R.S. & Gao, Y.S., 1981. Some recent ad-



vances in the chemical studies of Chinese herbal medicine. *American Journal of Botany* 68(2): 300–303. |4| Gutierrez, H.C., 1980. An illustrated manual of Philippine materia medica. Vol. 1. National Research Council of the Philippines, Bicutan, Tagig, Metro Manila, the Philippines. pp. 154–156. |5| Jongkind, C.C.H., 1991. *Novitates gabonenses* 6. Some critical observations on Combretum versus Quisqualis (Combretaceae) and description of two new species of Combretum. *Bulletin du Muséum Nationale d'Histoire Naturelle*, Paris, 4e sér., 12, section B, Adansonia: 275–280. |6| Kim Byeung Gie, 1995. Anticoccidial effect of herb extracts against *Eimeria tenella*. *Korean Journal of Veterinary Clinical Medicine* 12(1): 123–128. |7| Kyi Thein et al., 1995. Preliminary screening of medicinal plants for biological activity based on inhibition of cyclic AMP phosphodiesterase. *International Journal of Pharmacognosy* 33(4): 330–333. |8| Lecompte, O., 1969. Combretaceae. In: Aubréville, A. (Editor): *Flore du Cambodge, du Laos et du Vietnam* [Flora of Cambodia, Laos and Vietnam]. Vol. 10. Muséum National d'Histoire Naturelle, Paris, France. pp. 22–31. |9| Quintana, E.G., Salduluz, J.D., Batoon, M.P. & Generalao, M.L., 1982. Agricultural production of selected medicinal plants: propagation to postharvest handling. PCARRD [Philippine Council for Agriculture, Forestry, and Natural Resources Research and Development] *Monitor* 10(4): 8–10. |10| Shinazaki, H., Hirate, K. & Ishida, M., 1987. Modification of drug-induced tremor by systematic administration of kainic acid and quisqualic acid in mice. *Neuropharmacology* 26(1): 9–18.

#### *Selection of species*

#### **Quisqualis conferta (Jack) Exell**

*Journ. Bot.* 69: 122 (1931).

**Synonyms** *Quisqualis densiflora* Wallich ex Miq. (1855), *Quisqualis prostrata* Craib (1926), *Quisqualis thorelli* Exell (1931).

**Vernacular names** Malaysia: akar dani, selimpas, sumang (Peninsular). Thailand: lep mue naang (Chumphon). Vietnam: l[af]ng nhi[eef]u hoa, d[aa]y giun nh[or].

**Distribution** Indo-China, Thailand, Peninsular Malaysia and possibly in Sumatra.

**Uses** A decoction of the leaves or juice from the pounded roots is used as a vermifuge.

**Observations** A prostrate shrub or woody climber, young branches appressed pubescent or

tomentulose; leaves 5–16 cm × 2–6 cm; upper receptacle 18–25 mm long, calyx lobes with recurved filiform tips, 2–3 mm long, petals 3–4 mm long; fruit 2–2.5 cm × 1–1.8 cm. *Q. conferta* is found in margins of primary forest at low altitude.

**Selected sources** 202, 403, 831, 1126, 1128, 1380.

#### **Quisqualis indica L.**

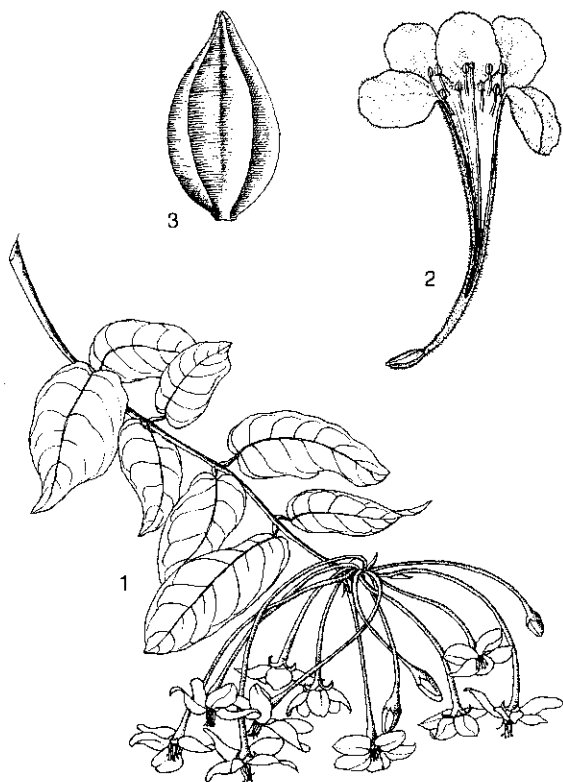
Sp. pl. ed. 2, 1: 556 (1762).

**Synonyms** *Quisqualis glabra* Burm.f. (1768), *Quisqualis pubescens* Burm.f. (1768), *Quisqualis spinosa* Blanco (1845).

**Vernacular names** Burma creeper, Chinese honeysuckle, Rangoon creeper (En). Indonesia: udani (Sumatra), bidani (Sundanese), cekluk (Javanese). Malaysia: akar dani, selimpas, udani (Peninsular). Philippines: niyog-niyogan (general, Tagalog), balitadham (Bisaya), tartaraok (Ilokan). Cambodia: dong preah phnom, vor romiet nhi. Laos: dok ung, khua hung, 'sa mang'. Thailand: cha mang (northern), thai-mong (Karen-Mae Hong Son), lep mue naang (central, peninsular). Vietnam: d[aa]y giun, qu[ar] giun, s[uwr] qu[aa]n.

**Distribution** Probably native to the Asian tropics and occurring throughout the Malesian region. Nowadays widely cultivated, mainly as an ornamental hedge plant, throughout the tropics. There is still doubt whether it is indigenous to east tropical Africa or was introduced there long ago.

**Uses** The fruits and seeds of *Q. indica* are well-known throughout Malesia as an anthelmintic, particularly to treat ascarids. In large doses they cause nausea, vomiting, hiccough and even unconsciousness. In Indonesia, a decoction of the root, seed or fruit is used as a vermifuge. In the Philippines, *Q. indica* is used as a bechic or pectoral, the fruits and seeds to alleviate nephritis, and the seeds as anthelmintic. In Peninsular Malaysia, a decoction of the seeds is given to children to stop diarrhoea, the juice of the leaves is considered a remedy for boils and ulcers and the leaves are applied to the head to relieve ache caused by jungle fever. In Papua New Guinea, plants are eaten daily by men and women as a method of birth control. In Vietnam, the root is used to treat rheumatism and a concentrated decoction of the fruit is used as a gargle effective against toothache. In China, seeds macerated in oil are applied to parasitic skin diseases, boils or sores on children's faces, and when roasted are given to treat diarrhoea and fever. In Mongolia, the seeds and fruit are reported to sustain the spleen and cause obstructions to disappear.



*Quisqualis indica* L. – 1, flowering twig; 2, opened flower; 3, fruit.

**Observations** A woody climber, young branchlets tomentose to sparsely pubescent; leaves 5–18.5 cm × 2.5–9 cm; upper receptacle 5–8 cm long, calyx lobes deltoid to triangular, 1–2 mm long, petals 10–20 mm long; fruit 2.5–4 cm × 0.7–1.3 cm. *Q. indica* occurs in forest margins of undisturbed forest to very disturbed places, at low altitudes. *Q. indica* is highly variable; in Indo-China 3 varieties have been recognized on the basis of the bract size and fruit shape: var. *indica*, var. *pierrei* (Gagnep.) O. Lecompte (synonym: *Q. pierrei* Gagnep.), and var. *villosa* (Roxb. ex DC.) Kurz. Elsewhere still other varieties have been distinguished.

**Selected sources** 55, 61, 75, 186, 190, 202, 245, 287, 321, 332, 350, 358, 363, 368, 391, 403, 405, 454, 464, 467, 469, 531, 580, 597, 694, 728, 734, 802, 831, 913, 922, 1003, 1035, 1126, 1128, 1176, 1178, 1287, 1303, 1333, 1359, 1508, 1567, 1586, 1598, 1639.

N.O. Aguilar

## *Rauwolfia* L.

Sp. pl. 1: 208 (1753); Gen. pl. ed. 5: 98 (1754).

APOCYNACEAE

$x = 11$ ; *R. serpentina*:  $2n = 22, 44$ , *R. sumatrana*:  $n = 88$ , *R. verticillata*:  $2n = 22$

**Major species** *Rauwolfia serpentina* (L.) Benth. ex Kurz, *R. verticillata* (Lour.) Baillon.

**Vernacular names** Snakewood (En). Indonesia: laméh. Philippines: sibakong. Vietnam: c[aa]y ba g[a]lc.

**Origin and geographic distribution** The number of species in *Rauwolfia* varies from 60–125, depending on literature sources. The genus is pantropical. Ten species occur in Africa and Madagascar, about 20 in tropical America, and the remainder in Asia, Australia and the Pacific. Nine species have been distinguished in the Malasian region, but as a result of new taxonomic studies this number might be reduced to 5.

**Uses** *Rauwolfia* has been used in Ayurvedic medicine in India since ancient times to treat snake bites, mental diseases and epilepsy, and is still important. In Vietnam it is considered to be one of the most useful and effective medicinal plants. Its main use is as an anti-hypertensive agent and as tranquilizer. An extract of the root rind is considered as a highly effective remedy against high blood-pressure and to relieve the central nervous system. Besides this, it is also used to treat dysentery, diarrhoea, liver diseases, psychoses, insanity, epilepsy and snake bites and/or scorpion stings, and to stimulate uterine contraction and to promote the expulsion of the foetus. In a mixture with other plants it is sometimes used to treat cholera and fever. The leaf juice is applied locally against opacity of the cornea and to treat wounds and itch. The drug is also considered as an anthelmintic and tonic.

*Rauwolfia* species are used for the extraction of alkaloids, principally reserpine, but also ajmaline and ajmalicine. Reserpine is marketed for the treatment of (mild to moderate) arterial hypertension. Combination with a diuretic is classical in order to control the water and  $\text{Na}^+$  retention that is a side effect of reserpine. The drug has been much used since the 1960s, however, because of its narrow therapeutic index (i.e. the difference between therapeutic and toxic doses is very small) and its major side effect (depression), reserpine is currently of secondary interest. More manageable compounds have become available for therapy. In psychiatric therapy, reserpine has also been replaced by synthetic products (e.g.

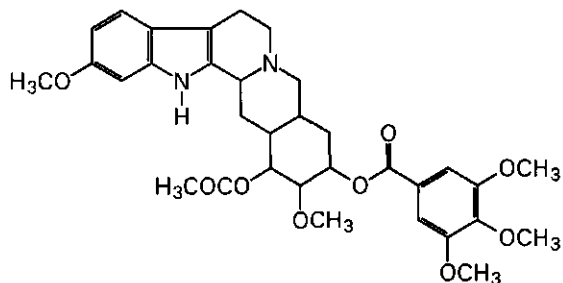
phenothiazines and butyrofenon derivatives).

Ajmalicine is an ingredient of proprietary products used to treat the psychological and behavioral problems of senility, sensory problems, cerebrovascular accidents, cranial traumas and their neurological sequelae. Another alkaloid, ajmaline is used as a remedy for heart arrhythmias in distinct cases (class 1A agent).

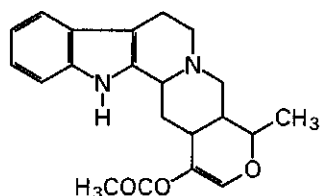
The lightweight wood of some *Rauwolfia* spp. which can reach the size of a tree is sometimes used for small objects such as knife handles.

**Production and international trade** *Rauwolfia* roots and their preparations are used and traded in many countries and are important on the international market. Thailand appears to be the main exporter, with amounts of about 100 t/year in the second half of the 1970s, whereas Nepal exported some 30 t/year in the same period. India was a large supplier of *R. serpentina* roots before 1969 (on average 40 t/year), but then the export was banned by the government to help develop an extraction industry. In 1982, the price of powdered *R. serpentina* roots was US\$ 9/kg and of reserpine US\$ 0.4/g.

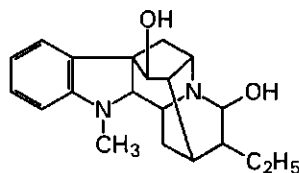
**Properties** Commercial samples of *R. serpentina* drugs consist of tortuous pieces of root of up to 15 cm long and 2 cm in diameter; they are yellowish and of low density. Although at present the botanical name is *Rauwolfia*, the crude drug is called *Rauwolfia* (*Rauwolfia Radix*). The total alkaloid content is 0.5–3%, and over 50 different alkaloids have been demonstrated, most of them indole-type, derived from tryptophan. The alkaloids can further be classified into 4 main groups (typical analysis in parentheses): (1) yohimbane-type derivatives, e.g. reserpine (0.14%), reserpinine (rescinnamine, 0.015%), isorauhimbine (3-epirauhimbine, 0.08%), (-)-corynanthine (rauhimbine, 0.03%), deserpidine, yohimbine and corynantheine; (2) heteroyohimbane derivatives, e.g. serpentinine (serpentinine, 0.13%), serpentine (0.08%), raubasine (ajmalicine, 0.02%), reserpiline and alstonine; (3) sarpagane derivatives, e.g. sarpagine (raupine, 0.02%); and (4) dihydro-indole (ajmalane) derivatives, e.g. ajmaline (0.1%). Standardized *Rauwolfia* powder has a total content of 0.15–0.20% alkaloids of the reserpin-rescinnamine group, calculated as reserpine, and analysed according to the USP XXII. Furthermore, several spectrophotometric and extraction-photometric methods have been developed for determining *Rauwolfia* alkaloids, e.g. reserpine, serpentine and ajmaline. These methods are comparatively simple and accurate.



reserpine



raubasine (ajmalicine)



ajmaline

Of the *Rauwolfia* alkaloids, 5 are used in medicine: reserpine, rescinnamine, deserpidine, raubasine (ajmalicine) and ajmaline. There are several patented methods available for the extraction of the main component reserpine.

Reserpine is a sympatholytic agent acting indirectly on the peripheral (noradrenergic) and central (noradrenergic and serotonergic) nerve terminals. By inhibiting the  $Mg^{2+}$  and ATP dependent transport of e.g. noradrenaline (norepinephrine) it amplifies the breakdown of this substance by monoamino oxidases and catecholamine transferases. The resulting depletion of catecholamines on the peripheral level induces a lasting drop in blood pressure and heart rate. Sedative and neuroleptic activity is caused by central neurotransmitter depletion. Contra-indications for using reserpine are depression, combination with monoamino-oxidase inhibitors or laevodopa, peptic ulcer, and hypersensitivity to the alkaloid. Side effects of the medication include drowsiness, nasal congestion, salivary and gastric hypersecretion, paradoxical anxiety, depression and retention of

water and Na<sup>+</sup> (the latter may be overcome by co-administration of a diuretic). Overdose may cause respiratory depression, bradycardia, hypotension, confusion, tremors, myosis, convulsions and gastro-intestinal distress. Oral administration of *R. serpentina* extract has caused blood and urine glucose to decrease in cats and diabetic patients. Reserpine has been shown to enhance the hypoglycaemic effect of insulin and the hyperglycaemic effect of adrenalin, and has inhibited the physiological hyperglycaemic response in diabetic patients. Reserpine did not show genotoxicity and was unable to induce reverse mutation and recombinational mitotic events (crossing-over and gene conversion) in yeast strains.

Rescinnamine (reserpinine) is a reserpine analogue containing a trimethoxycinnamic rather than a trimethoxybenzoic-acid residue. Deserpidine (11-demethoxyreserpine) lacks a methoxy function at position 11 of the reserpine molecule. Both alkaloids have the same effects as reserpine, and can be used to treat the same conditions, although their side effects are reported to be less pronounced.

Raubasine (ajmalicine) is an  $\alpha$ -adrenergic blocking spasmolytic which, at high doses, reverses the effects of adrenalin (epinephrine), and moderates the activity of the vasomotor centres, especially in the brain stem. It causes a transient increase of the blood flow to the brain, and is slightly anxiolytic.

Ajmaline is an anti-arrhythmic (class 1A group), which substantially decreases the rate of depolarization of atrial and ventricular cells. Its toxicity has limited its uses to some specialized cases.

The alkaloids reserpine, spagatrine and verticillatine have been reported in *R. verticillata*. Spagatrine has been shown to be an  $\alpha$ -adrenergic blocker, whereas verticillatine exhibited ganglionic blocking activity. Clinical application of verticillatine showed significant therapeutic effect in treating severe cases of hypertension, with few side effects.

*R. serpentina* powder at a concentration of 0.25% has been found to be very effective in protecting grain against *Rhizopertha dominica*. The roots also exhibit plant growth inhibition and uterine stimulant, antihyperglycaemic, and dopamine receptor blocking activity. The leaf extract showed fungitoxicity; it reduced growth of *Sclerotium rolfsii* significantly. An extract of *R. serpentina* markedly decreased the number of local lesions and systemic infection caused by brinjal necrotic mosaic virus on aubergines.

**Adulterations and substitutes** *Apocynaceae* other than *Rauwolfia*, for example *Catharanthus roseus* (L.) G. Don, have similar or related alkaloids with similar applications.

**Description** Shrubs or small to medium-sized trees up to 30 m tall, often candelabra-shaped, with latex in branchlets but not in bark; bark smooth, rough, fissured or scaly. Leaves verticillate in whorls of 3-4(-5), rarely opposite, simple and entire, short-petioled with axillary glands on the petiole, exstipulate. Inflorescence a terminal, peduncled cyme, sometimes seemingly axillary. Flowers actinomorphic, bisexual, small, 5-merous; calyx deeply divided, with overlapping lobes; corolla salver-shaped, with cylindrical or campanulate tube, sometimes ventricose in or above the middle, with long hairs inside in the upper half, throat usually constricted, often hairy, lobes twisted to the left in the bud, white or greenish-white to pink, tube often reddish outside; stamens inserted in the widening of the corolla tube, alternating with corolla lobes, filaments very short, anthers not or hardly exerted, free from stigma, medifixed, acute; disk annular or cupular; ovary, superior, with 2 carpels which are free or connate, style 1, glabrous, stigma provided at the base with a collar and with a bifid apical cusp. Fruit consisting either of 2 free drupelets or a single, entire or bilobed drupe with 1-2 tuberculate pyrenes, usually about 1 cm in diameter, ripening blackish. Seeds laterally compressed, obliquely ovate or elliptical, with a large embryo; endosperm fairly abundant. Seedling with epigeal germination; cotyledons leafy, green; first 2-3 nodes with decussate leaves, subsequent leaves whorled.

**Growth and development** Ramification in *Rauwolfia* is determined by the verticillate leaves; branches terminate in 2-5 elements, consisting of branchlets or inflorescences. The number of elements is the same as the number of leaves in the whorl. This results in an umbellate ramification and a candelabra-shaped habit.

*R. serpentina* and *R. verticillata* flower throughout the year in Peninsular Malaysia. The flowers are pollinated by insects like small bees and flies.

**Other botanical information** Full-grown flowers and ripe fruits are indispensable for a reliable identification of *Rauwolfia*. *R. javanica*, *R. reflexa* and *R. sumatrana* are difficult to distinguish, and might be considered as one polymorphous species.

*Rauwolfia* resembles *Ervatamia* and *Kopsia*, but *Ervatamia* differs in its opposite leaves and seeds with red or orange sarcotesta, and *Kopsia* in its

opposite leaves, corolla segments overlapping to the right and disk consisting of 2 scales alternating with the carpels.

The roots or root rind of *R. cambodiana* Pierre ex Pitard, *R. chaudocensis* Pierre ex Pitard, *R. indochinensis* Pichon and *R. vietnamensis* Ly are used in Vietnam to treat high blood pressure, and sometimes also to treat dysentery and as antibiotic and antiseptic. *R. vomitoria* Afzel. is an important species from West and Central Africa with high alkaloid concentration. *R. tetraphylla* L. is from tropical America. Both species are used for industrial alkaloid extraction; *R. tetraphylla* is cultivated very locally in gardens in India, Vietnam and China, *R. vomitoria* in Vietnam.

**Ecology** *Rauvolfia* species occur scattered in forest, often in secondary forest, and scrub vegetation. Several tolerate shade well, but some (e.g. *R. serpentina*) occur especially in more open places, e.g. in forest edges and along rivers. Slightly acid soils (pH 5–6.5) are favourable, but some *Rauvolfia* spp. also grow well on limestone soil. Experiments with *R. serpentina* grown in different soil media in the Philippines showed no significant difference in root production.

**Propagation and planting** *R. serpentina* is usually propagated by seed, although stem and root cuttings can also be used. The germination rate can be very low, therefore it is recommended to select mature and heavy seeds and to sow them within 6 months after ripening. In Vietnam, fruits are soaked in water for 12 hours, they are then crushed and the seeds are cleaned and subsequently soaked in warm water (40–45°C) for a further 12 hours. The seeds are sown in nursery beds, and start germinating within 3 weeks. Seedlings 10–12 cm tall are usually transplanted during the rainy season at a planting distance of 45 cm × 30 cm. A planting distance of 50 cm × 50 cm is recommended for *R. verticillata*. Propagation of *R. serpentina* in India gave a success rate of 40–65% from stem cuttings, 50–80% from root cuttings; a success rate of up to 90% has been obtained in Vietnam under careful management.

Successful in vitro propagation techniques have been developed for *R. serpentina*. Sterilized shoot tips can be cultured in Murashige and Skoog medium containing 3% sucrose. Best shoot response was obtained in the presence of 0.5 mg/l naphthalene acetic acid and 2 mg/l benzyladenine, with 15–20 shoots arising from one shoot tip. For rooting it is essential to replace benzyladenine by kinetin. When the resulting plantlets were planted in the field the survival rate was 60% and the

plantlets were cytologically stable. Plantlets have also been successfully regenerated from shoot cultures of *R. serpentina* initiated from auxiliary meristems on medium containing benzyladenine (4.5 µM) and naphthalene acetic acid (0.5 µM). Rooting was initiated in White's basal medium supplemented with 0.5 µM naphthalene acetic acid. The resulting plants were similar to normal ones in their morphological characteristics and chemical constitution, but produced more biomass.

Nodal cultures of *R. serpentina* could be maintained for 9 months at 25°C on a standard Murashige and Skoog medium. Low-temperature incubation of in vitro cultures appeared highly promising because cultures exhibited normal health even after 15 months of storage at 15°C. Temperatures of 5–10°C were found to be deleterious to the growth of the cultures.

The total indole alkaloid content of *R. serpentina* roots from plants regenerated from stem and root callus was slightly higher than in the parental stock, but the content of ajmaline, serpentine and reserpine was lower.

**In vitro production of active compounds** Cell suspension cultures of *R. serpentina* have proved to be an excellent source of the enzymes involved in the biosynthesis of the alkaloids of the ajmalane and sarpagane class. Moreover, cell suspension culture is one of the most efficient methods for indole alkaloid formation. So far, about 30 different indole alkaloids have been isolated and identified from cultivated *Rauvolfia* cells. Alkaloid production under optimum conditions ranges from micrograms to grams per litre medium (e.g. yields of up to 1.6 g/l of raucaffricine have been obtained). A cell suspension culture of *R. serpentina* continuously treated with hydroquinone produced up to 18 g/l of p-hydroxyphenyl-O-β-D-glucoside (arbutin), which is the highest transformation rate ever observed with a plant cell culture system for a single natural product.

Raucaffricine (vomilenine-galactoside) has been shown to be the major indole alkaloid of cell suspension cultures of *R. serpentina* grown in alkaloid production medium. This compound is converted by an enzyme to its aglycon vomilenine, which has a key function in the biosynthesis of ajmaline. A higher content of intracellular indole alkaloids of the ajmaline type was found in a transgenic hairy root culture in liquid Murashige and Skoog medium than in the leaves and roots of the intact plant. It has also been found that phytoosterols, including stigmasterol, β-sitosterol and

cholesterol, are produced in callus tissue cultures. It has been demonstrated that there are significant genome rearrangements in *R. serpentina* callus cultivated for a long time. These can occur at early stages of cultivation, but the process of subsequent subculturing in vitro leads to more significant genome changes. A highly productive cell line was found to be a partially synchronized myxoploid stable cell population in which tetraploid and multiploid cells and cells with a low level of structural mutations of the chromosomes predominated. The content of indole alkaloids in the tissue was positively correlated with the increase in the frequency of tetraploid and multiploid cells.

Hybrid cell suspension cultures have also been generated (e.g. from *R. serpentina* and *Rhazya stricta* Decne., and from *R. serpentina* and *Vinca minor* L.). Alkaloids not previously detected in the parental cell cultures may be formed in the hybrid culture, and in a cell line of a *R. serpentina* and *Vinca minor* hybrid a 10-fold increase in raucaffricine accumulation was observed relative to the parental *Rauwolfia* strain.

**Husbandry** Regular weeding and hoeing (2–3 times during the growing season in India) is needed to maintain satisfactory development of the roots. A top dressing of ammonium sulphate given after weeding will promote the development of a vigorous stand. *R. verticillata* grows more vigorously than *R. serpentina* and will soon shade out weeds. In Malaysia, *R. serpentina* has been found very sensitive to magnesium deficiency, causing severe chlorosis; application of moderate amounts of potash fertilizers is therefore recommended. In India, application of farmyard manure and compost showed a beneficial effect on the growth of *R. serpentina*.

**Diseases and pests** In India, wilting caused by *Fusarium* is the most serious disease in cultivated *R. serpentina*, followed by *Alternaria* leaf blight, powdery mildew, mosaic virus disease and root-knot disease, resulting in stunted growth. Leaf spot and blight caused by *Rhizoctonia solani* result in premature defoliation in the rainy season; weeding up to a distance of 45 cm from the plants effectively controlled the pathogen, as weeds serve as host. Young branches are sometimes infected with green bugs.

**Harvesting** The roots of *R. serpentina* are usually harvested from the wild, although it is cultivated to a limited extent, e.g. in India. When cropped, the roots are harvested after 1.5–3 years. Care should be taken to keep the root bark intact

as it has a high alkaloid content. In Vietnam, it is recommended not to gather the roots of *R. verticillata* in spring because the concentration of active compounds is low then. Preferably, only roots with a diameter over 3 cm should be collected, leaving small-diameter roots for a next harvest.

**Yield** The average yield in *R. serpentina* plantations in India is 2 t/ha when roots are harvested 15 months after planting, and 36 t/ha when harvested after 3 years. However, harvesting after 15 months was found to be most lucrative. Under favourable soil conditions in Vietnam, 3-year-old plants produced on average 50 g of dried root rind. In a small-scale experiment in Peninsular Malaysia, the average yield of dried root per *R. verticillata* plant was 200 g 1.5 years after planting, 600 g after 2 years and 930 g after 3 years.

**Handling after harvest** Roots of *Rauwolfia* are usually cut into pieces 10–15 cm long. To prevent fungus infection they should be dried before storage to approximately 8% moisture content. Dried roots are usually packed in jute bags or bales, smaller quantities for regional markets are packed in polyethylene bags of 2–5 kg. Material to be packed in polyethylene bags must be adequately dried.

**Genetic resources and breeding** Natural populations of *R. serpentina* have been overexploited in many regions, particularly because roots are usually harvested, which destroys the plants. The species may now be extinct in Sri Lanka, due to intense exploitation, and in India and Java it has become rare. Several *Rauwolfia* species are threatened with extinction in Vietnam, e.g. *R. chaudocensis*, *R. indochinensis* and *R. vietnemensis*. These valuable plant resources must be protected by ex situ and in situ conservation. There are no known germplasm collections.

**Prospects** The use of *Rauwolfia* in galenic preparations and the use of reserpine have declined significantly in recent decades in industrialized countries because of their strong side effects and the availability of more effective alternatives. Moreover, reserpine has been suspected of having carcinogenic effects (causing breast neoplasm) and, although this effect has not been confirmed, its use has declined considerably and is unlikely to regain its former level. In developing countries, products based on *Rauwolfia* are still in demand owing to their easy availability and comparatively low prices, but it is expected that in the South-East Asian countries they will be gradually replaced by modern alternatives. As long ago as 1958, commercial cultivation of *Rauwolfia* was

considered uneconomic in Malaysia, mainly because of the apparently adequate stocks of Indian origin, although trials had shown that it could be brought into cultivation at short notice. However, continued research might reveal new possibilities.

**Literature** |1| Allen, E.F., 1958. Notes on the cultivation of *Rauwolfia* in Malaya. *Malayan Agricultural Journal* 41(2): 100-105. |2| Banerjee, N. & Sharma, A.K., 1983. Cytotaxonomy, tissue culture and alkaloids of *Rauwolfia* L. *Nucleus* 26(3): 197-207. |3| International Trade Centre UNCTAD/GATT, 1982. Markets for selected medicinal plants and their derivatives. Geneva, Switzerland. pp. 106-112. |4| Markgraf, F., 1984. *Flora Malesianae Praecursores* LXIV. Apocynaceae VI. *Rauwolfia*. *Blumea* 30: 157-167. |5| Mukhopadhyay, S., Mukhopadhyay, M.J. & Sharma, A.K., 1991. In vitro multiplication and regeneration of cytologically stable plants of *Rauwolfia serpentina* Benth. through shoot tip culture. *Nucleus* 34(3): 170-173. |6| Roja, G. & Heble, M.R., 1996. Indole alkaloids in clonal propagules of *Rauwolfia serpentina* Benth. ex Kurz. *Plant Cell, Tissue and Organ Culture* 44(2): 111-115. |7| Ruyter, C.M., Akram, M., Illahi, I. & Stöckigt, J., 1991. Investigation of the alkaloid content of *Rauwolfia serpentina* roots from regenerated plants. *Planta Medica* 57(4): 328-330. |8| Sarin, Y.K., 1982. Cultivation and utilization of *Rauwolfia serpentina* in India. In: Atal, C.K. & Kapur, B.M. (Editors): *Cultivation and utilization of medicinal plants*. Regional Research Laboratory, Council of Scientific and Industrial Research, Jammu-Tawi, India. pp. 288-294. |9| Schütte, H.-R., 1991. Secondary plant substances: monoterpene indole alkaloids. *Progress in Botany* 52: 84-96. |10| Sharma, N. & Chandel, K.P.S., 1992. Low-temperature storage of *Rauwolfia serpentina* Benth. ex Kurz: An endangered, endemic medicinal plant. *Plant Cell Reports* 11(4): 200-203.

#### *Selection of species*

#### ***Rauwolfia amsoniifolia* DC.**

*Prodr.* 8: 338 (1844).

**Vernacular names** Indonesia: parempasa, pamedang (Sulawesi). Philippines: sibakong (Tagalog), banogan (Panay Bisaya), maladita (Bikol, Bukidnon).

**Distribution** The Philippines, Timor, southern Sulawesi, southern Moluccas (Tanimbar Islands and Kai Islands).

**Uses** In the Philippines, a decoction of the bark

is used as a stomachic and young leaves are used to treat stomach disorders in babies. In Indonesia, a decoction of the leaves is used as a laxative and febrifuge, and to stimulate delivery, the bark to treat framboesia.

**Observations** A shrub or small tree up to 15 m tall; leaves verticillate, lanceolate, 6-12 cm × 1.5-3 cm; flowers with campanulate-infundibuliform corolla tube at most twice the length of the calyx; fruit nearly obreniform. *R. amsoniifolia* occurs in lowland rain forest and in secondary thickets.

**Selected sources** 332, 580, 905, 1178.

#### ***Rauwolfia javanica* Koord. & Valeton**

*Bijdr.* 1: 191 (1894).

**Vernacular names** Indonesia: lameh, lameh utan (Sumatra).

**Distribution** Sumatra, Java and the Lesser Sunda Islands.

**Uses** Pounded leaves are used externally to treat wounds.

**Observations** A small to medium-sized tree up to 22(-30) m tall with bole up to 65 cm in diameter; leaves 3-4-verticillate, lanceolate, 14-30 cm × 3.5-6.5 cm, petiole rather thick, up to 1 cm long; flowers with campanulate-infundibuliform corolla tube at most twice the length of the calyx; fruit obversely trapezoid. *R. javanica* is closely related to *R. reflexa* and *R. sumatrana*. It occurs in lowland to lower montane rain forest up to 1800 m altitude, sometimes in open places.

**Selected sources** 97, 580, 905.

#### ***Rauwolfia reflexa* Teijsm. & Binnend.**

*Nat. Tijds. Ned. Ind.* 3: 329 (1852).

**Vernacular names** Indonesia: ki benteli (Sundanese), lameh, lameh utan (Java).

**Distribution** Java and the Lesser Sunda Islands.

**Uses** The bark is reported to serve as a laxative.

**Observations** A small tree up to 15 m tall with bole up to 25 cm in diameter; leaves 3-4-verticillate, obovate-elliptical, 6-21 cm × 3.5-6.5 cm, petiole thin, up to 1.5(-2.5) cm long; flowers with campanulate-infundibuliform corolla tube 2-3 times the length of the calyx; fruit subglobose with a broad obtuse base. *R. reflexa* is closely related to *R. javanica* and *R. sumatrana*. It occurs in lowland rain forest, also in forest edges, open places and beach forest, up to 1000 m altitude.

**Selected sources** 97, 580, 905.

***Rauvolfia serpentina* (L.) Benth. ex Kurz**

For. Fl. Burma 2: 171 (1877).

**Synonyms** *Ophioxylon serpentinum* L. (1753).

**Vernacular names** Indonesia: pule pandak (Javanese). Thailand: khem daeng (northern), ra yom (central), ka yom (peninsular). Vietnam: ba g[aj]c [aas]n d[ooj], ba g[aj]c hoa d[or], ba g[aj]c thu[oo]c.

**Distribution** India, Sri Lanka, Indo-China, southern China (Yunnan), Thailand, northern Peninsular Malaysia, Java and the Lesser Sunda Islands (Flores, Timor); cultivated in Pakistan, Nepal, India, Java, Ambon, Vietnam, southern China and Georgia.

**Uses** An extract of the root rind is considered as a highly effective remedy against high blood pressure and to relieve the central nervous system. Besides this, it is also used to treat dysentery, diarrhoea, psychoses, insanity, epilepsy and snake bites, and to stimulate uterine contraction and to promote the expulsion of the foetus. In Thailand,

besides these uses, the roots are used to enhance appetite and as a galactagogue. In a mixture with other plants, *R. serpentina* is also used to treat cholera and fever. The leaf juice is applied against opacity of the cornea and to treat wounds and itch. The root is also used as a vermifuge in veterinary medicine.

**Observations** A small shrub up to 0.6(-1) m tall, with prominent tuberous usually unbranched root and usually unbranched slender stem; leaves opposite or 3(-5)-verticillate, oblanceolate or obovate, 7-16 cm × 3-9 cm, petiole up to 1.5 cm long; flowers with narrowly cylindrical tube much longer than calyx; fruit consisting of 1-2 globose drupelets connate at base. *R. serpentina* occurs in sunny or shaded places in well-drained rain forest and secondary thickets up to 2100 m altitude, sometimes as a weed in sugar cane fields.

**Selected sources** 49, 78, 97, 118, 193, 476, 549, 580, 652, 796, 879, 905, 987, 1126, 1253, 1266, 1277, 1302, 1320.

***Rauvolfia sumatrana* Jack**

Mal. Misc. 1(5): 22 (1820).

**Synonyms** *Cyrtosiphonia madurensis* Teijsm. & Binnend. (1823), *Cyrtosiphonia sumatrana* (Jack) Miq. (1856), *Rauvolfia madurensis* (Teijsm. & Binnend.) Boerl. (1899).

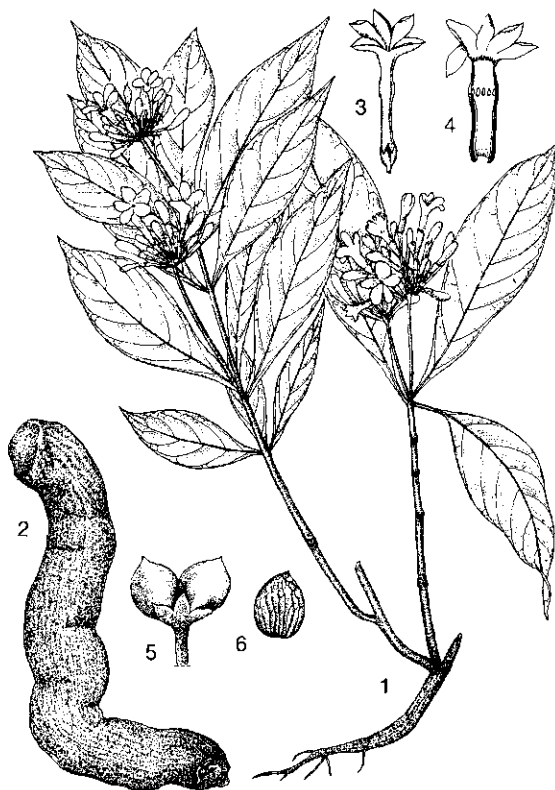
**Vernacular names** Indonesia: lame lalaki (Sundanese), polay lakek (Madurese), tampa badak (Sumatra). Malaysia: pulai pipit, pelir kambing (Peninsular). Thailand: teenpet lek (Surat Thani), ra yom teenpet (Bangkok).

**Distribution** The Andaman Islands, Burma (Myanmar), Thailand, Peninsular Malaysia, Sumatra, Java, the Lesser Sunda Islands, Borneo, Sulawesi, the Moluccas, the Aru Islands and the Philippines.

**Uses** The bark is used to treat dysentery. In the Philippines, the dried bark is reported as an anti-malarial. The lightweight wood is sometimes used for planks and small objects such as knife handles.

**Observations** A small to medium-sized tree up to 20 m tall; leaves 3-5-verticillate, elliptical-obovate to elliptical, 7-26 cm × 3-5 cm, petiole thin, up to 3 cm long; flowers with campanulate-infundibuliform corolla tube 2-3 times the length of the calyx; fruit subglobose with a broad obtuse base. *R. sumatrana* is closely related to *R. javanica* and *R. reflexa*. It occurs in lowland dipterocarp forest and teak forest, sometimes also in forest edges and secondary vegetation, up to 1400 m altitude.

**Selected sources** 97, 118, 202, 580, 905, 1564.



*Rauvolfia serpentina* (L.) Benth. ex Kurz - 1, plant habit; 2, piece of root; 3, flower; 4, opened corolla; 5, fruit; 6, stone.



**Rauvolfia verticillata (Lour.) Baillon**

Bull. Soc. Linn. Paris 1: 768 (1888).

**Synonyms** *Dissolena verticillata* Lour. (1790), *Rauvolfia chinensis* (Spreng.) Hemsl. (1889), *Rauvolfia perakensis* King & Gamble (1907).

**Vernacular names** Indonesia: salung-salung (Sumatra). Malaysia: pokok batu pelir kambing (Peninsular). Thailand: cheepuk (northern), chaek (Trang), yaa kae haak khom (Chiang Mai). Vietnam: ba g[aj]c v[of]ng, t[is]ch t[ee]n, sam t[oo].

**Distribution** India, Sri Lanka, Burma (Myanmar), Indo-China, southern China, Taiwan, Thailand, Peninsular Malaysia, Sumatra, Java, Lombok, Borneo and the Philippines (Luzon).

**Uses** The root is valued in Indo-Chinese and Chinese medicine as hypertensive and sedative. Fresh leaves are applied externally to treat snake bites, wounds and inflamed eyes.

**Observations** A shrub up to 3 m tall; leaves (2–)3-verticillate, elliptical, (8–)10–20 cm × (2–)4–6 cm, petiole up to 1.5 cm long; flowers with narrow cylindrical corolla tube much longer than calyx; fruit consisting of 1–2 elliptical drupelets. *R. verticillata* occurs in lowland to montane rain forest and monsoon forest, up to 1700 m altitude, often in open places in hills and mountains, e.g. along rivers, near villages and rice fields.

**Selected sources** 49, 118, 202, 850, 879, 905, 1035, 1564.

Tran Dinh Ly & Pham Duy Mai

**Rhinacanthus nasutus (L.) Kurz**

Journ. Asiat. Soc. Bengal 39, pt. 2, Nat. Hist.: 79 (1870).

ACANTHACEAE

2n = 30

**Synonyms** *Rhinacanthus communis* Nees (1832).

**Vernacular names** Indonesia: daun burung (Malay, Ambon), tarebak (Sundanese), tereba jepang (Malay, Jakarta). Philippines: parajito (Spanish), ibon-ibonan, tagak-tagak (Tagalog). Burma (Myanmar): anitia. Laos: thong kan<sup>2</sup> sang. Thailand: thong khan chang, thong phan chang, yaa man kai (central). Vietnam: bl[aj]ch h[aj]c, ki[ees]n c[of].

**Origin and geographic distribution** *R. nasutus* is probably native to Sri Lanka, India, Indo-China and southern China, but has been introduced long ago in Madagascar, tropical East Africa, Thailand and the Malesian region (Peninsular Malaysia, Java, the Moluccas, the Philip-

pinas) where it is now widely naturalized and often common.

**Uses** The roots and leaves of *R. nasutus* are applied externally as a remedy for certain skin disorders such as ringworm, eczema, scurf and herpes. They are either soaked in vinegar or alcohol, pounded with lemon or tamarind, or made into a decoction. In Peninsular Malaysia, they are prepared with sulphur and benzoin or vaseline. In Thailand the leaves may be pounded with alcohol, lemon and tamarind juice. The resulting extract is applied on the infected skin. In Vietnam, an infusion of *R. nasutus* has a reputation in folk medicine for the treatment of hypertension. In China, the stem and leaves are also applied to treat ringworm infections, as well as in early stages of tuberculosis. When applied internally the leaf is used as an antipyretic, antihypertensive, anti-inflammatory and detoxicant, and against snake venom. *R. nasutus* is considered to be aphrodisiac. In Thailand, anti-cancer activity has been reported.

*R. nasutus* is also regularly planted as a hedge plant and has been applied for erosion control in road construction. Moreover, in Thailand it is planted for its ornamental value. In Madagascar, the seeds are used for scenting clothes.

**Production and international trade** The roots of *R. nasutus* used to be imported in Europe from China under the name 'tong pamg chong'.

**Properties** *R. nasutus* is reported to contain several naphthoquinones: rhinacanthin-A and -B have been isolated from roots collected in Thailand. Bioassay-directed fractionation showed significant cytotoxicity for the latter compound in the Kenacid Blue (KB) cell line with ED<sub>50</sub> values of 3 µg/ml. The naphthoquinones rhinacanthin-C and -D, isolated from the aerial parts, exhibited potent inhibitory activity against human and murine strains of human cytomegalovirus (CMV) in vitro (ED<sub>50</sub> values of 0.02 and 0.22 µg/ml, respectively). Several other naphthoquinones have been isolated. Most of these showed significant cytotoxicity, particularly rhinacanthin-D, -H, -K, -M and -Q, whereas they also showed inhibition of rabbit platelet aggregation.

From the methanol extract of stems and leaves of *R. nasutus* collected in south-eastern Thailand, an antifungal naphthoquinone was isolated with an ED<sub>50</sub> value of 0.4 ppm on spore germination of *Pyricularia oryzae* (causing rice blast disease) and with 82% inhibition at 100 ppm. Originally the chemical structure of this naphthoquinone was thought to be 3,4-dihydro-3,3-dimethyl-2H-naphtho[2,3-b]pyran-5,10-dione, a p-quinone,

which is very similar to the structures of the rhinacanthins. Recently however, this structure was revised into 3,4-dihydro-3,3-dimethyl-2H-naphtho[1,2-b]pyran-5,6-dione, an o-quinone, also known as rhinacanthone.

Furthermore, two lignans, rhinacanthin-E and -F isolated from the aerial parts, exhibited significant antiviral activity against influenza virus type A.

A 95% alcohol extract of the aerial parts exhibited antimicrobial activity against *Staphylococcus aureus* at a dose of 100 mg/disk. The chloroform and alcohol extracts of aerial parts show antifungal activity against *Epidermophyton floccosum*, *Microsporium gypseum* and *Trichophyton rubrum*. The crude ethanol extract proved to possess relatively high acaricidal activity (71–85% mortality) tested in vitro on cattle ticks.

The aqueous extract of *R. nasutus* has a high anti-alkylating effect against ethyl methane sulphonate; anti-alkylating substances are associated with anti-carcinogenic activity.

Finally, phytochemical investigations have furthermore revealed the presence of flavonoids (luteol) and phytosterols ( $\beta$ -sitosterol, stigmasterol and their glucosides).

**Description** An erect, branched shrub up to 2(–3) m tall; stems obtusely quadrangular, puberulent when young. Leaves opposite, simple, ovate to lanceolate or elliptical, 3–10 cm  $\times$  1–5 cm, base acute to attenuate, margin entire, apex acute, puberulent; petiole 0.5–2 cm long; stipules absent. Inflorescence an axillary, peduncled, lax cyme often combined into a leafy, terminal panicle, densely appressed pubescent. Flowers subsessile; calyx with 5 narrow lobes which are shortly connate at base, 5–6 mm long; corolla 2-lipped, tube narrowly cylindrical, green, upper lip with 2 teeth, 8–10 mm  $\times$  2–3 mm, white, lower lip with 3 large lobes, the central one 10–14 mm  $\times$  9–13 mm, white with red markings at base; stamens 2, inserted near the corolla tube apex, anther cells inserted at unequal level; disk present; ovary superior, 2-locular with 2 ovules in each cell, style 1, with a 2-fid stigma. Fruit a clavate, loculicidal, puberulous capsule, 17–25 mm long, basal part sterile. Seeds held up on well-developed hooks (retinacula), orbicular, flat, pubescent.

**Growth and development** In Java *R. nasutus* flowers throughout the year, but in the Philippines it flowers only in December–March. Pollination is by insects. Fruits apparently do not ripen in Java. The seeds are ejected from the capsule by the retinacula.



*Rhinacanthus nasutus* (L.) Kurz – 1, flowering twig; 2, upper part of flower; 3, fruit.

**Other botanical information** *Rhinacanthus* comprises about 10–15 species. It belongs to the tribe *Justicieae* of the subfamily *Acanthoideae*.

**Ecology** *R. nasutus* is found in thickets, hedges and waste places up to 750 m altitude. It thrives best on moist, well-drained soils, but it is also found in much drier habitats such as rock crevices.

**Propagation and planting** *R. nasutus* can easily be propagated by stem cuttings. Cuttings of about 10 cm, comprising 2–3 nodes, are planted under partial shade during the rainy season. Propagation by in vitro culture is another option. To obtain complete callus initiation and highest callus production in stem explants of *R. nasutus*, the Murashige and Skoog basal medium should be supplemented with the growth regulators 2,4-dichlorophenoxyacetic acid (2,4-D; 1.0 mg/l) and kinetin (1.0 mg/l).

**Harvesting** Mature leaves are hand picked when required, preferably shortly before flowering.

**Yield** Rhinacanthone has antifungal properties but occurs only at low concentration in *R. nasutus*.

Methyl 1-methoxy-2-naphthoate is one of several precursors also isolated from *R. nasutus*. This precursor can easily be transformed to rhinacanthone, thereby increasing the yield to 30%.

**Prospects** The naphthoquinones and lignans from *R. nasutus* display some very interesting effects, e.g. cytotoxic, antifungal or antiviral, which merit further attention.

**Literature** [1] Achararit, C., Panyayong, W. & Ruchatakumut, E., 1983. Inhibitory action of some Thai herbs to fungi. Undergraduate Special Project Report 1983. Faculty of Pharmacy, Mahidol University, Bangkok, Thailand. 13 pp. [2] Farnsworth, N.R. & Bunyapraphatsara, N., 1992. Thai medicinal plants. Prachachon Co. Ltd., Bangkok, Thailand. pp. 216-217. [3] Greshoff, M., 1894. Schetsen nuttige Indische planten [Sketches of useful Indonesian plants]. Series 1 (1-50). Extra Bulletin van het Koloniaal Museum. J.H. De Bussy, Amsterdam, the Netherlands. pp. 41-42. [4] Kernan, M.R. et al., 1997. Two new lignans with activity against influenza virus from the medicinal plant *Rhinacanthus nasutus*. *Journal of Natural Products* 60(6): 635-637. [5] Kodama, O., Ichikawa, H., Akatsuka, T., Santisopasri, V., Kato, A. & Hayashi, Y., 1993. Isolation and identification of an antifungal naphthopyran derivative from *Rhinacanthus nasutus*. *Journal of Natural Products* 56(2): 292-294. [6] Kuwahara, S., Nemot, A. & Hiramoto, A., 1991. Synthesis of an antifungal naphthopyran derivative isolated from *Rhinacanthus nasutus*, Acanthaceae. *Agricultural and Biological Chemistry* 55(11): 2909-2912. [7] Nguyen Van Duong, 1993. Medicinal plants of Vietnam, Cambodia and Laos. Mekong Printing, Santa Ana, California, United States. pp. 35-36. [8] Sendl, A. et al., 1996. Two new naphthoquinones with antiviral activity from *Rhinacanthus nasutus*. *Journal of Natural Products* 59(8): 808-811. [9] Sinchaisri, P., 1989. Phon khong suan sakat khong phut totan mateng thimi, to alkylating activity khong ethyl methane sulphonate [Effects of solvent fractions of some anti-carcinogenic plants on the alkylating activity of ethyl methane sulphonate]. *Warasan Wicha kan kaset* 7(1-3): 27-33. [10] Wu, T.S., Tien, H.J., Yeh, M.Y. & Lee, K.H., 1988. Isolation and cytotoxicity of rhinacanthin A and B, two naphthoquinones from *Rhinacanthus nasutus*. *Phytochemistry* 27(12): 3787-3788.

**Other selected sources** 88, 97, 142, 190, 202, 287, 350, 580, 741, 801, 921, 933, 1126, 1128, 1178, 1287, 1380, 1508, 1525, 1530, 1600.

Wongsatit Chuakul, Noppamas  
Soonthornchareonnon & Promjit Saralamp

## Schefflera J.R. Forster & J.G. Forster

Char. gen. pl.: 45, t. 23 (1775).

ARALIACEAE

$x = 12$ ; *S. elliptica*:  $n = 24$

**Major species** *Schefflera elliptica* (Blume) Harms, *S. heptaphylla* (L.) Frodin.

**Vernacular names** Schefflera (En, Fr).

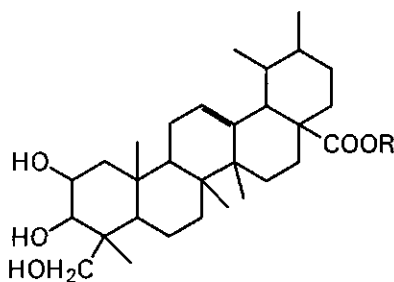
**Origin and geographic distribution** *Schefflera* probably comprises over 400 species and is widely distributed in the tropics and subtropics. In Asia it occurs from Sri Lanka north to the Himalayas and Japan, in Indo-China, Thailand and throughout the Malesian region towards northern Australia, New Zealand and east to Samoa in the Pacific.

**Uses** The leaves and bark of several *Schefflera* species are used as a remedy for cough and as a diuretic and tonic. The leaves show astringent properties and are, for example, given to women after childbirth. A decoction of the leaves of *S. elliptica* has been proved to be an effective antiscorbutic. The resin has been applied as a vulnerary. In Thailand, an infusion of the leaves is used to relieve asthma.

A great number of other *Schefflera* species are popular pot plants of considerable economic importance worldwide. In Vietnam, *S. leucantha* R. Vig. and *S. heptaphylla* are reported to be cultivated as ornamentals and pot plants. The wood of *S. heptaphylla* is soft, light and easy to work, and can be used for paper, musical instruments and matchboxes. The leaves and young branches are used as green manure.

**Production and international trade** Medicinal products from *Schefflera* are only used and traded on a local scale.

**Properties** Phytochemical investigations have revealed the presence of terpenoid saponins in the leaves and bark of *S. heptaphylla*. Asiaticoside and its aglycone asiatic acid have been isolated



asiatic acid (R = H) and asiaticoside (R = glucose-glucose-rhamnose)

from the bark, together with cauloside D and several new related triterpene saponin glycosides. In general, the glycosides mentioned can be divided into two series of six corresponding ursene and oleanene glycosides, all of which have the same triose moiety in the C-28 position. This is why the names scheffursosides A-F and scheffoleosides A-F have been proposed for these corresponding pairs of glycosides, respectively (asiaticoside = scheffursoside A, cauloside D = scheffoleoside C). From the leaves of *S. heptaphylla*, 3-epi-betulinic acid and some of its (acetylated) glycosides (3,28-bidesmosidic saponins), oleanonic acid and other closely related triterpene saponins have been isolated. A betulinic acid glycoside has also been isolated from *S. elliptica*; moreover, its leaves also contain saponins.

Asiaticoside and asiatic acid are known to have several pharmacological effects, mainly involved in wound healing. There are literature reports of stimulation of human fibroblast collagen I synthesis (in vitro), just as bacteriostatic actions in tuberculosis models.

An aqueous extract from the leaves of *S. elliptica* showed bronchodilator activity on the isolated smooth muscle preparation of the respiratory tract of guinea-pigs (constrictions induced by addition of histamine). The effect was similar to other well known bronchodilators such as terbutaline and theophylline (as aminophylline). Therefore, the extract appears promising for the prevention and relief of asthmatic attacks, although the exact nature of the components involved in the action remains to be investigated. Furthermore, a leaf extract of *S. elliptica* showed a dose-related positive inotropic action on an atria preparation of rats.

Finally, saponins from the leaves of *S. leucantha* were found to have bronchodilator action.

**Adulterations and substitutes** Asiatic acid and asiaticoside have also been reported in extracts of *Centella asiatica* (L.) Urb., which is a pantropical species extending into some subtropical regions. Asiatic acid has also been found in ether extracts from the wood of *Terminalia brassii* Exell and *T. complanata* K. Schumann.

**Description** Evergreen or deciduous trees, shrubs and woody climbers, often epiphytic; twigs with stellate hairs or glabrous; leaves and bark fragrant when crushed. Leaves arranged spirally, palmately compound, rarely unifoliate or 2-3 times palmately compound; stipules often elongate, connate and intrapetiolar; leaflets usually entire or occasionally coarsely toothed. Flowers in

umbellules or small heads arranged in a panicle or rarely in a raceme or spike, bisexual, actinomorphic; pedicel not articulated; calyx rim-like, undulate or 5-toothed; petals 5 or more, valvate in bud; stamens usually 5, sometimes up to 12 or rarely more; disk epigynous; ovary inferior or half-inferior, 5-12-locular or rarely more, with a single ovule in each cell, styles as many as locules, united or free or absent and then the stigmas sessile. Fruit a smooth or slightly ribbed drupe, fleshy or dry, dark red or black when mature; pyrenes compressed.

**Growth and development** Many *Schefflera* species develop according to the architectural tree model of Leeuwenberg, in which 2 or more orthotropic modules develop below an inflorescence and these are equivalent and determinate by terminal flowering. Development according to the architectural tree model of Tomlinson, which is characterized by the repeated development of equivalent orthotropic modules in the form of basal branches, has been reported for an unidentified *Schefflera* species. Inflorescences may be terminal or lateral, growth of the modules is usually continuous, sometimes rhythmic. Several species develop strangling roots whereas others are unbranched treelets. Flowers are pollinated by insects.

**Other botanical information** *Schefflera* includes several previously recognized genera, the most important being *Agalma*, *Brassaia*, *Heptapleurum*, *Paratropia*, *Scheffleropsis*, *Sciadophyllum* and *Tupidanthus*, although some authors still prefer to distinguish some or all of these separately. *Schefflera* includes several complexes in which species boundaries are still unclear; the Malesian species are in need of a thorough taxonomic revision. In the light of recent taxonomical insight, the species known almost universally since the 1890s as *Schefflera octophylla* (Lour.) Harms should be called *S. heptaphylla* (L.) Frodin. It is a renowned medicinal plant from Indo-China, southern China, Taiwan and the Ryukyu Islands.

**Ecology** Most medicinal *Schefflera* are found in the understorey of primary lowland rain forest, occasionally also in secondary forest and thickets. Individual species may be found on limestone hills, and up to 2500 m altitude.

**Propagation and planting** *Schefflera* can be propagated from ripe seed sown under humid conditions. Alternatively, propagation by air-layering, stem and softwood cuttings are common practice for the popular ornamental *Schefflera*.

**Harvesting** In Vietnam, the bark of stem and

root of *Schefflera* are collected all year round, but the best time for harvesting is autumn or just before flowering.

**Handling after harvest** The stem bark of *Schefflera* is scraped off and the root-bark washed to remove the outer layer. The bark is then wrapped for 24–48 hours to develop the aroma as a result of fermentation, and dried in the shade.

**Genetic resources and breeding** *Schefflera* species confined to primary forest habitats and those with limited geographical distribution are potentially threatened by ongoing deforestation in South-East Asia. *S. elliptica* is widespread and commonly found in a wide range of habitats, including disturbed anthropogenic vegetation, and is less likely threatened.

**Prospects** *Schefflera*'s wound-healing properties and soothing effect on itching skin are probably related to the presence of the triterpenes asiatic acid and asiaticoside. The pharmacological effects of *Schefflera* triterpenes in general deserve further attention.

**Literature** [1] Frodin, D.G., 1978. *Schefflera*. In: Ng, F.S.P. (Editor): Tree flora of Malaya. Vol. 3. Malayan Forest Records No 26. Longman Malaysia Sdn. Berhad, Kuala Lumpur & Petaling Jaya, Malaysia. pp. 25–32. [2] Frodin, D.G., 1986. Studies in *Schefflera* (Araliaceae), II. Northern Luzon (Philippines) species of the Heptapleurum group. Proceedings of the Academy of Natural Sciences of Philadelphia 138: 403–425. [3] Frodin, D.G., 1990. Studies in *Schefflera* (Araliaceae), IV. The identity of *Vitis heptaphylla* L., a long-misplaced Linnaean ivy tree. Botanical Journal of the Linnean Society 104: 309–424. [4] Maeda, C., Ohtani, K., Kasai, R., Yamasaki, K., Nguyen, M.D., Nguyen, T.N. & Nguyen, K.Q., 1994. Oleanane and ursane glycosides from *Schefflera octophylla*. Phytochemistry 37(4): 1131–1137. [5] Quisumbing, E., 1978. Medicinal plants of the Philippines. Katha Publishing Co., Quezon City, the Philippines. pp. 678–681. [6] Satayavivad, J., Bunyapraphatsara, N. & Saivises, R., 1980. Pharmacological and toxicological studies to the constituents of *Schefflera venulosa* (Araliaceae). In: Department of Chemistry, Faculty of Science, Mahidol University, Bangkok: 4th Asian symposium on medicinal plants and spices (Abstracts). Mahidol University, Bangkok, Thailand. p. 47. [7] Shang, C.B., 1984. Le genre *Schefflera* (Araliacées) en Chine et en Indochine [The genus *Schefflera* (Araliaceae) in China and Indo-China]. Candollea 39: 453–486. [8] Taesotikul, T., Panthong, A. & Kanjanapothi, D., 1980. Bronchodilator ac-

tivity of *Schefflera venulosa* (family Araliaceae): preliminary investigation. In: Department of Chemistry, Faculty of Science, Mahidol University, Bangkok: 4th Asian symposium on medicinal plants and spices (Abstracts). Mahidol University, Bangkok, Thailand. p. 46. [9] Tomlinson, P.B. & Zimmermann, M.H. (Editors), 1978. Tropical trees as living systems. The proceedings of the fourth Cabot symposium held at Harvard Forest, Peterham, Massachusetts, on April 26–30, 1976. Cambridge University Press, Cambridge, London, New York, Melbourne. pp. 223–231, 269–284. [10] Vu Van Dung et al., 1996. Vietnam forest trees. Agricultural Publishing House, Hanoi, Vietnam. p. 64.

#### *Selection of species*

#### ***Schefflera blancoi* Merr.**

Philipp. Journ. Sci. 1, Suppl.: 109 (1906).

**Synonyms** *Cephaloschefflera blancoi* (Merr.) Merr. (1923).

**Vernacular names** Philippines: sainat (Filipino), abkal (Igorot), sagaba (Iloko).

**Distribution** The Philippines.

**Uses** *S. blanco* is reputed to be used as a fish poison.

**Observations** A large climber; leaves with 10 leaflets, petiole about 1/3 of the length of the leaf, leaflets lanceolate, entire; flowers in rounded umbellules, 4–6-merous, stigma sessile. *S. blancoi* is commonly found on exposed ridges in mossy forests at 700–1500 m altitude.

**Selected sources** 190, 935.

#### ***Schefflera caudata* (S. Vidal) Merr. & Rolfe**

Philipp. Journ. Sci. 3, Bot.: 118 (1908).

**Synonyms** *Heptapleurum caudatum* S. Vidal (1885), *Schefflera acuminatissima* Merr. (1906), *Schefflera piperioidea* Elmer (1908).

**Vernacular names** Philippines: himainat (Filipino), lima-lima (Tagalog, Bisaya).

**Distribution** The Philippines (Luzon).

**Uses** A decoction of *S. caudata* is given as a tonic to women after childbirth.

**Observations** An epiphytic vine; leaves few, crowded towards the ends of twigs, 3-foliolate, leaflets elliptical, over 15 cm long, pointed at both ends; inflorescence terminal, with 1–5 main branches along which the rounded umbellules are racemosely displayed. The formerly recognized species *S. piperioidea* is now regarded as a variety: *S. caudata* var. *piperioidea* (Elmer) Frodin. *S. cau-*

*data* is found in primary forest at low to medium altitude.

**Selected sources** 190, 935, 936, 1178.

***Schefflera cumingii* (Seem.) Harms**

Engl. & Prantl, Nat. Pflanzenfam. 3(8): 39 (1894).

**Synonyms** *Heptapleurum cumingii* Seem. (1865).

**Vernacular names** Philippines: kalkugamat (Filipino), kolokagama (Negrito).

**Distribution** The Philippines (Luzon).

**Uses** *S. cumingii* is reputed to be used to cure stomach troubles.

**Observations** A small shrub or scrambler or climber; leaves palmately 3–5-foliolate, petiole longer than the petiolules, leaflets thin, elliptical to ovate-elliptical, 10–15 cm long, pointed at two ends, without an intramarginal vein; main inflorescence axis shorter than its branches; flowers in umbellules; fruit obovoid, 5-locular. *S. cumingii* is found in primary forest at low altitudes; most localities have a relatively high and consistent rainfall.

**Selected sources** 190, 434, 936, 1126, 1178.

***Schefflera elliptica* (Blume) Harms**

Engl. & Prantl, Nat. Pflanzenfam. 3(8): 39 (1894).

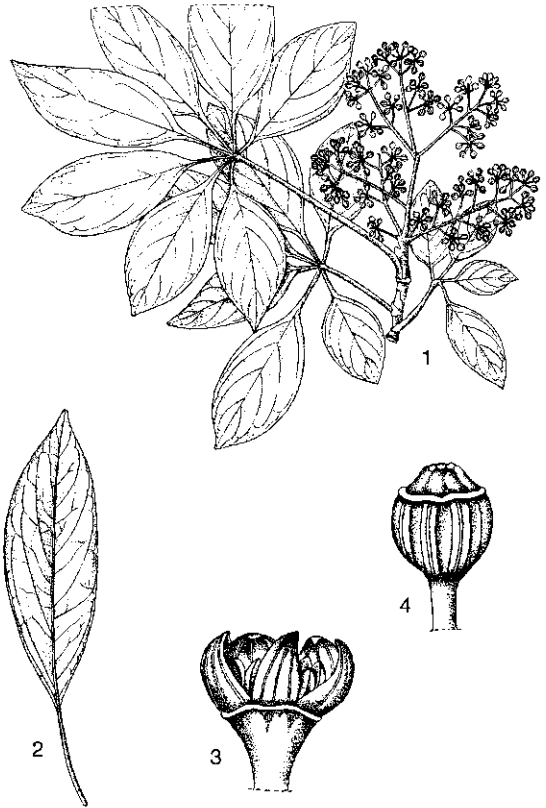
**Synonyms** *Schefflera venulosa* (Wight & Arn.) Harms (1894), *Schefflera odorata* (Blanco) Merr. & Rolfe (1908), *Schefflera bengalensis* Gamble (1919).

**Vernacular names** Indonesia: panakomo (Timor), putiana ma gitipi (Halmahera), tanganan (Javanese). Malaysia: ara bebari, chenamah gajah (Peninsular). Philippines: lima-lima (Filipino), arasagat (Iloko), galamai-amo (Tagalog). Laos: lep mu nang, pha nha hay, tang<sup>2</sup> to<sup>1</sup>. Thailand: mue phranaaraai (Trat), nuat plaa muek khao (Chiang Mai), waan oi chaang (Loei). Vietnam: d[as]ng nhi[ee]f[u g[aa]n, d[as]ng thu[oo]n, ch[aa]n chim leo.

**Distribution** From India to Indo-China, southern China, Thailand and throughout the Malesian region except for New Guinea.

**Uses** The bark is employed as a bechic, the resin as a vulnerary. A decoction of the leaves is an effective antiscorbutic and may also be used in aromatic baths. The wood has been chewed to relieve toothache. In India, the roots mixed with rice are eaten to cure dropsy.

**Observations** An epiphytic or terrestrial climber or shrub up to 10 m tall; leaves palmately



*Schefflera elliptica* (Blume) Harms – 1, flowering twig; 2, narrower leaflet; 3, flower; 4, fruit.

(4–)5–7-foliolate, petiole 10–12 cm long, leaflets ovate-elliptical to obovate-elliptical, 7–18 cm × 3–10 cm, apex acute or obtuse, entire, leathery, glabrous, petiolules 1.5–6 cm long; inflorescence with some branches as long as or shorter than the main axis, glabrescent, 10–20 cm long; flowers 5-merous, very small, in 10-flowered umbellules which are arranged racemously along the branches; fruit globose to ovoid, usually 5–6-locular, yellow or orange becoming black. *S. elliptica* is common in secondary forest and thickets, often along rivers and also frequent along the coast and in mangrove vegetation, up to 2500 m altitude.

**Selected sources** 97, 190, 202, 287, 434, 580, 1126, 1291, 1314, 1380, 1435, 1476, 1525, 1526, 1564.

***Schefflera elliptifoliola* Merr.**

Philipp. Journ. Sci. 13, Sect. C, Bot.: 320 (1918).

**Vernacular names** Philippines: galamai (Filipino).

**Distribution** The Philippines.

**Uses** A decoction of the leaves is used as a tonic by women after childbirth.

**Observations** An epiphytic shrub or woody climber; leaves palmately 9–11-foliolate, petiole 13–18 cm long, clasping the stem, leaflets elliptical to obovate-elliptical, 7–15(–25) cm × 4.5–7 cm, rounded at base, pointed at apex, entire, glabrous, on 3–5.5 cm long petiolules; inflorescence up to 20 cm long, primary branches 15–20; flowers 8–12 together in rounded umbellules, 5-merous, 8–12 mm long; fruit ellipsoid, 3–4 mm long. *S. elliptifoliola* is found in damp forest.

**Selected sources** 190, 934, 935, 1126, 1178.

### *Schefflera heptaphylla* (L.) Frodin

Bot. Journ. Linn. Soc. 140: 314 (1990).

**Synonyms** *Vitis heptaphylla* L. (1771), *Schefflera octophylla* (Lour.) Harms (1894).

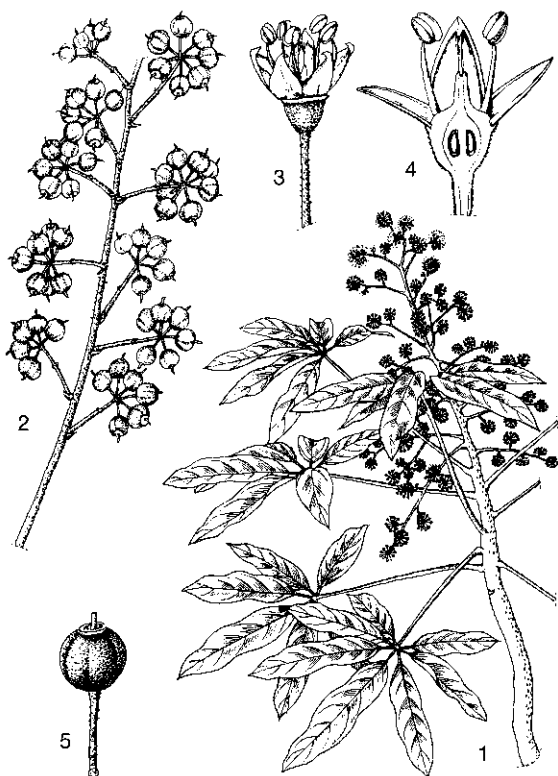
**Vernacular names** Laos: ko tan. Vietnam: ch[aa]n chim, nam s[aa]m.

**Distribution** Burma (Myanmar), Thailand, Indo-China, the Philippines (Batan Island), southern China, Taiwan, the Ryukyu Islands and southernmost Japan.

**Uses** The bark is widely used in folk medicine for its diuretic properties and as a tonic. The ashes are sometimes used to treat dropsy. In Hong Kong the fresh branchlets are used as a wash to soothe the itching of the skin.

**Observations** A small to medium-sized, semi-deciduous or evergreen tree up to 25 m tall, bole up to 80 cm in diameter; leaves palmately 6–8(–11)-foliolate, polymorphic, petiole 8–35 cm long, leaflets elliptical to ovate-elliptical, 7–20 cm × 3–6 cm, base attenuate, apex narrowly pointed, margin entire, glabrous, petiolules unequal, 1–5 cm long; inflorescence a well-developed panicle with hairy branches; flowers in many-flowered umbellules or sometimes solitary at the top of secondary axes; flowers 5-merous, ovary 5–8(–10)-locular; fruit globular, 3–4 mm in diameter, black. *S. heptaphylla* is found in relatively open forest and forest edges. In southernmost Japan it occurs near sea-level; in the Ryukyu Islands up to 600 m elevation. Southward in the tropics its maximum altitude rises to 1200(–1400) m, or it even becomes entirely montane. Its distribution corresponds with the 20°C average January isotherm. Exploited from wild sources as well as from cultivation, this species can probably be grown easily at higher elevations in the Malesian region.

**Selected sources** 363, 435, 884, 1035, 1070, 1126, 1128, 1314, 1416, 1417, 1418, 1419, 1526.



*Schefflera heptaphylla* (L.) Frodin – 1, flowering twig; 2, part of infructescence; 3, flower; 4, flower in longitudinal section; 5, fruit.

### *Schefflera heterophylla* (Wallich ex G. Don) Harms

Engl. & Prantl, Nat. Pflanzenfam. 3(8): 38 (1894).

**Synonyms** *Heptapleurum junghuhniana* (Miq.) Seem. (1865), *Schefflera junghuhniana* (Miq.) Harms (1894), *Schefflera curtisii* (King) Ridley (1922).

**Vernacular names** Malaysia: akar chabang lima, seregang (Peninsular). Thailand: khan haam suea (Nakhon Si Thammarat).

**Distribution** Thailand, Peninsular Malaysia and Java.

**Uses** A decoction of the leaves is given after childbirth.

**Observations** A large scrambling shrub; leaves 2–3 times palmately 3–5-foliolate, petiole 9–20 cm long, leaflets ovate to elliptical, 3.5–15 cm × 2–4.5 cm, oblique at base, long acuminate at apex, entire or with some coarse teeth above the middle, stellate-hairy when young; inflorescence branches about as long as the main axis, up to 30 cm long;

flowers 8–14 in each umbellule, 5–6-merous. *S. heterophylla* occurs in lowland to lower montane primary forest, up to 1700 m altitude.

**Selected sources** 97, 202, 1126, 1380, 1564.

***Schefflera insularum* (Seem.) Harms**

Engl. & Prantl, Nat. Pflanzenfam. 3(8): 39 (1894).

**Synonyms** *Heptapleurum insularum* Seem. (1865), *Schefflera mindanaensis* Merr. (1908).

**Vernacular names** Philippines: galamai-amo (Filipino), kalangkang (Panay Bisaya), pararan (Bagobo).

**Distribution** The Philippines.

**Uses** The juice of pounded fresh leaves is used as a purgative.

**Observations** A woody climber; leaves palmately 7–9-foliolate, petiole about 20 cm long, leaflets oblong to oblong-elliptical, 10–24 cm × 3–8.5 cm, base cuneate, often slightly unequal, apex narrowly pointed, margin toothed in the upper half, glabrous, on 2.5–7 cm long petiolules; inflorescence 15–30 cm long, with only 1–3 branches of up to 20 cm long; flowers 9-merous, in small, 3–6-flowered umbellules, ovary 9–11-locular; fruit ovoid, 7–9 mm in diameter, orange. *S. insularum* is common in primary forest, often along streams, at low altitudes.

**Selected sources** 190, 932, 935, 1126, 1178.

***Schefflera oxyphylla* (Miq.) R. Vig.**

Ann. Sci. Nat., Bot. IX, 9: 355 (1909).

**Synonyms** *Schefflera subracemosa* R. Vig. (1909), *Schefflera subulata* (Miq.) R. Vig. (1909), *Schefflera klossii* Ridley (1920).

**Vernacular names** Malaysia: akar pesat badak, akar sepakan, akar sesudu (Peninsular).

**Distribution** Thailand, Peninsular Malaysia, Sumatra and Borneo.

**Uses** A decoction of the roots has been given as a sedative to calm frightened children and has been used externally against fevers.

**Observations** An epiphytic or terrestrial creeper or climber; leaves 3–5-foliolate, leaflets usually with 1–2 steeply ascending basal lateral veins; inflorescence branches spike-like, few-branched; flowers in very short-stalked umbellules. *S. oxyphylla* is a variable, common species occurring in lowland and hill forest, occasionally in submontane rain forest, sometimes on limestone, up to 1200 m altitude.

**Selected sources** 202, 1126, 1564.

***Schefflera simulans* Craib**

Kew Bull.: 421 (1930).

**Synonyms** *Schefflera affinis* (King) R. Vig. (1909) non Baillon.

**Vernacular names** Malaysia: bekak rengat, pokok bajang beranak (Peninsular).

**Distribution** Thailand and Peninsular Malaysia.

**Uses** A decoction of the leaves has been used to cure stomach trouble, and that of the root and leaves has been applied as a protective medicine after childbirth.

**Observations** A terrestrial or epiphytic shrub up to 3 m tall; leaves palmately 5–7-foliolate, petiole 6.5–9 cm long, leaflets oblong-oblong-ovate, 7–11 cm × 2–3.7 cm, base cuneate and unequal, apex acuminate, glabrous, petiolules 1.5–3.5 cm long; inflorescence 10 cm long, 15–20 cm wide, hairy but glabrescent; flowers about 10 per umbellule, 6–7-merous. *S. simulans* occurs locally in montane forest at 1300–1650 m altitude.

**Selected sources** 202, 291, 1564.

***Schefflera trifoliata* Merr. & Rolfe**

Philipp. Journ. Sci. 3, sect. C, Bot.: 118 (1908).

**Vernacular names** Philippines: sinat (Filipino), gauai-gauai, himainat (Tagalog).

**Distribution** The Philippines.

**Uses** Crushed leaves, with or without oil, are applied externally against tympanites of children. Internally, the leaves are given to women after childbirth and to treat irregular menstruation.

**Observations** A woody vine; leaves 3-foliolate, petiole 3–8 cm long, leaflets oblong to oblong-ovate, apex caudate-acuminate, entire, petiolules 1.5–4 cm long; inflorescence 20–25 cm long, with few branches; flowers many, in 3–6-flowered umbellules, pedicels 4–5 mm long; fruit oblong, 6 mm × 3 mm, 5-locular. *S. trifoliata* is found in primary forest at low and medium altitudes.

**Selected sources** 190, 935, 936, 1126, 1178.

Nguyen Tap & M.S.M. Sosef

***Scutellaria* L.**

Sp. pl. 2: 598 (1753); Gen. pl. ed. 5: 260 (1754).

LABIATAE

$x$  = unknown; *S. discolor*:  $2n = 24, 26$

**Major species** *Scutellaria javanica* Jungh.

**Vernacular names** Skullcap (En). Scutellaire (Fr). Vietnam: thu[aax]n.

**Origin and geographic distribution** *Scutellaria*, with 360 species, is almost cosmopolitan.



Originally it was absent only from the Amazon Basin, lowland tropical Africa, South Africa, deserts of Central Asia, most of the Pacific Islands and New Zealand, and north of the Arctic circle. The centres of diversity are found in the mountain regions of Central Asia and China, which are also considered to be the regions where *Scutellaria* originated; only 3 species are native in Malesia.

**Uses** A multitude of *Scutellaria* species are used internally in the form of infusions and decoctions to alleviate stomach complaints and as a diuretic and antipyretic. Various species are used externally as a decoction or poultice to treat fungal skin infections as well as boils or scabies.

Outside the Malesian region *S. baicalensis* Georgi is an important medicinal plant, cultivated extensively for its roots in China, Korea and Japan. The roots are used in traditional Vietnamese medicine as a general tonic to balance the body, to treat bacterial infections of the respiratory and gastrointestinal tracts, and this species has been successfully grown in northern Vietnam.

**Properties** *Scutellaria* species, like many other *Labiatae*, contain an essential oil. The constituents of this oil are referred to as belonging to the monoterpenoids, sesquiterpenoids, phenylpropane derivatives or iridoid glycosides without further specification. *Scutellaria* species are also reported to contain diterpenes (in aerial parts e.g. of the neoclerodane type), large amounts of triterpenes, phytosterols and phenolic constituents (e.g. caffeic acid, flavonoids).

The presence and biological activities of flavonoids in *Scutellaria* have been particularly well studied. The aerial parts of *S. discolor* were found to contain chrysin, chrysin-7-O-glucuronide, apigenin, luteolin, wogonin, 5,7-dihydroxy-8,2'-dimethoxyflavone, 5,7,8-trihydroxyflavone-8-O- $\beta$ D-glucuronide, 5,7,2',6'-tetrahydroxy-8-methoxyflavone-2'-O- $\beta$ D-(2-O-caffeoyl)-glucoside, 5,7-dihydroxy-8,2'6'-trimethoxyflavone and 5,7,4'-trihydroxy-8-methoxyflavone. The latter two compounds are also present in the roots of *S. discolor*, together with pinocembrin, wogonin, wogonin-7-O-glucuronide, norwogonin, norwogonin-7-O-glucuronide, 7-hydroxy-5,8-dimethoxyflavone, 7-hydroxy-5,8,2'-trimethoxyflavone, 5,7-dihydroxy-8,2'-dimethoxyflavone, 5,7,2'-trihydroxy-8-methoxyflavone, 5,7,2'-trihydroxy-8,6'-dimethoxyflavone, 2(S)-5,7-dihydroxy-8,2'-dimethoxyflavanone, 2(S)-7-hydroxy-5,8,2'-trimethoxyflavanone, 5,2'-dihydroxy-7,8,6'-trimethoxyflavanone, 5,2'-dihydroxy-6,8,6'-trimethoxyflavanone and 2'4'-dihydroxy-2,3',6'-trimethoxy-chalcone.

The flavonoids scutellarin, found in *S. baicalensis* and *S. javanica*, and baicalein (= 5,6,7-trihydroxyflavone) inhibit the activity of partially purified rat brain protein kinase C. Various flavonoids present in *S. baicalensis* also show inhibitory effects against a considerable number of viruses: e.g. baicalin (= 5,6-dihydroxyflavone-7-O- $\beta$ D-glucuronide) against the human T cell leukaemia virus type 1 and the human immunodeficiency virus (HIV-1), 5,7,2'-trihydroxy- and 5,7,2',3'-tetrahydroxyflavone against the Epstein-Barr virus, and 5,7,4'-trihydroxy-8-methoxyflavone against A-H3N2 subtype and B-subtype of the influenza virus. 2(S)-5,7,2',6'-Tetrahydroxyflavanone showed a remarkable antibacterial activity against e.g. *Escherichia coli*, *Bacillus subtilis* and *Staphylococcus aureus*.

Furthermore, baicalein shows antiproliferative activity in cultured rabbit vascular muscle cells, and lipoxygenase-inhibitory activity. It may be useful as a template for the development of drugs to prevent the pathological changes of atherosclerosis and restenosis. The pharmacological action of baicalein may be partially attributed to its free radical scavenging activity. Other pharmacological activities of the flavonoids include: anti-inflammatory activity of baicalein in the rat adjuvant arthritis model, inhibition of LPS-induced IL-1 production by baicalin, baicalein and wogonin, and inhibition by baicalein of leukotriene C-4 biosynthesis by rat resident peritoneal macrophages. Another effect of the flavanoids is the inhibition of the release of slow reacting substances of anaphylaxis (SRS-A) from sensitized guinea-pig lungs after antigen challenge.

**Description** Perennial or rarely annual herbs or small shrubs; stems prostrate to erect, often 4-angled. Leaves opposite, simple, margin entire to pinnatifid, petiolate to sessile, exstipulate. Flowers solitary, opposite or in small false whorls, in the axil of leaves or bracts, in terminal or axillary, 1-sided or all-sided racemes or spikes; calyx with a short tube, 2-lipped, the lips entire, upper lip deciduous, with a large shield- or pouch-like appendage or rarely both lips expanded to form a bladder-like structure; corolla 2-lipped, with a usually long tube which is bent upwards at base or bent distally or both, upper lip hooded, entire or notched, lower lip 3-lobed; stamens 4, inserted on the corolla tube, didynamous, anterior pair longest, anthers of the anterior pair 1-celled, those of the posterior pair 2-celled; disk tubular; ovary superior, on a short gynophore, 2-carpellate but 4-locular with a single ovule in each cell, style with

a 2-fid stigma. Fruit consisting of 4 dry, ovoid to globose nutlets. Seeds without endosperm. Seedling with epigeal germination; cotyledons free, leafy; hypocotyl elongated; all leaves opposite.

**Growth and development** Malesian *Scutellaria* species have been found flowering throughout the year. In Taiwan *S. indica* has been observed flowering and fruiting from September to May. Pollination is by insects, mainly bees. The nutlets are simply shed when the wind tosses the inflorescence to and fro, without a distinct dispersal mechanism.

**Other botanical information** *Scutellaria* takes a rather isolated position within the *Labiatae*, being the only genus of the subfamily *Scutellarioideae*. Its subdivision into subgenera and sections has been subject to much debate, but the most recent view recognizes 2 subgenera, *Scutellaria* and *Apeltanthus*, and 7 sections. All Malesian species belong to the subgenus *Scutellaria* and section *Scutellaria* which harbours about 240 species.

**Ecology** *Scutellaria* species generally occur on grassy plains, along forest tracks and streams, in forest edges, but also in open primary forest and savanna forest, in Malesia from the lowland up to 2400(–3300) m altitude.

**In vitro production of active compounds**

Research on in vitro production of flavonoids in *Scutellaria* is restricted to *S. baicalensis*. Cell suspension culture as well as stem callus culture yielded a range of flavonoids, with the major constituents being baicalin and wogonin-7-O-glucuronic acid.

**Harvesting** *Scutellaria* plants are uprooted to collect the roots.

**Handling after harvest** The roots are washed and dried before being stored.

**Prospects** The flavonoids isolated from various *Scutellaria* species show a broad range of interesting pharmacological effects that merit further research, e.g. the inhibition of several pathological viruses, activities on the immune system (including inhibition of immune factors) and free radical scavenging activity.

**Literature** [1] Cantino, P.D. & Sanders, R.W., 1986. Subfamilial classification of *Labiatae*. *Systematic Botany* 11: 163–185. [2] Hamada, H., Hiramatsu, M., Edamatsu, R. & Mori, A., 1993. Free radical scavenging action of baicalein. *Archives of Biochemistry and Biophysics* 306(1): 261–266. [3] Huang, H.C., Wang, H.R. & Hsieh, L.M., 1994. Antiproliferative effect of baicalein, a flavonoid

from a Chinese herb, on vascular smooth muscle cell. *European Journal of Pharmacology* 251(1): 91–93. [4] Keng, H., 1978. *Labiatae*. In: van Steenis, C.G.G.J. (General editor): *Flora Malesiana*. Series 1, Vol. 8. Sijthoff & Noordhoff International Publishers, Alphen aan den Rijn, the Netherlands. pp. 323–326. [5] Konoshima, T. et al., 1992. Studies on inhibitors of skin tumor promotion XI. Inhibitory effects of flavonoids from *Scutellaria baicalensis* on Epstein-Barr virus activation and their anti-tumor-promoting activities. *Chemical and Pharmaceutical Bulletin* 40(2): 531–533. [6] Nguyen Van Duong, 1993. *Medicinal plants of Vietnam, Cambodia and Laos*. Mekong Printing, Santa Ana, California, United States. pp. 215–216. [7] Paton, A., 1990. A global taxonomic investigation of *Scutellaria* (*Labiatae*). *Kew Bulletin* 45: 399–450. [8] Perry, L.M., 1980. *Medicinal plants of East and Southeast Asia*. Attributed properties and uses. The MIT Press, Cambridge, Massachusetts, United States & London, United Kingdom. p. 194. [9] Tomimori, T., Miyaichi, Y., Imoto, Y., Kiu, H. & Namba, T., 1988. Studies on the Nepalese crude drugs XI. On the flavonoid constituents of the aerial parts of *Scutellaria discolor* Colebr. *Chemical and Pharmaceutical Bulletin (Tokyo)* 36(9): 3654–3658. [10] Xu, G., Zhang, L.P., Shen, H.F. & Hu, C.Q., 1993. Inhibition of protein kinase C by scutellarein and its analogues. *Acta Academiae Medicinae Shanghai* 20(3): 187–191.

*Selection of species*

**Scutellaria discolor Wallich ex Benth.**

Wallich, *Pl. asiat. rar.* 1: 66 (1830).

**Synonyms** *Scutellaria cyrtopoda* Miq. (1859), *Scutellaria heteropoda* Miq. (1859), *Scutellaria zollingeriana* Briq. (1898).

**Vernacular names** Indonesia: jawer kotok (Sundanese), amperu lemah (Javanese), daun kukur (Moluccas). Malaysia: nilam bukit (Peninsular), toma (Sakai, Peninsular). Vietnam: thu[aax]n nh[i]e[ef]u m[af]u.

**Distribution** From northern India and Nepal to Burma (Myanmar), Indo-China, southern China, Thailand, Peninsular Malaysia, Java, the Lesser Sunda Islands, the Moluccas and New Guinea.

**Uses** In Java, *S. discolor* has been used to treat pain in the loins. In China, it is applied as a folk remedy for colds, fever, sore throat, and enteritis.

**Observations** A small herb up to 50(–100) cm

tall, stems hirsute, usually simple; leaves broadly elliptical to rounded or rarely ovate, (3.5-)4-6(-11) cm × (2-)2.5-5(-10) cm, base rounded to cordate, margin coarsely crenate, sparsely pubescent to hirsute; bracts 1-3 mm long; flowers in small false whorls of 2-4(-5), in a simple, terminal, all-sided raceme of 5-24 cm long, glandular pubescent, corolla blue or pale blue to purple-violet, 9-12 mm long. *S. discolor* is subdivided into three varieties: var. *hirta* Handel-Mazzetti occurring in Yunnan (China), var. *cyrtopoda* (Miq.) Adelb. found at 1600-3200 m altitude in Java, and var. *discolor* occurring throughout the range of *S. discolor*. It is found in grassland along streams, shady and moist places in rain forest, in Irian Jaya in oak forest, in Timor on limestone in *Podocarpus* forest, up to 2400(-3200) m altitude.

**Selected sources** 202, 287, 580, 720, 854, 1118, 1126, 1461, 1462, 1463, 1476.

### *Scutellaria indica* L.

Sp. pl. 2: 600 (1753).

**Synonyms** *Scutellaria copelandii* Merr. (1912).

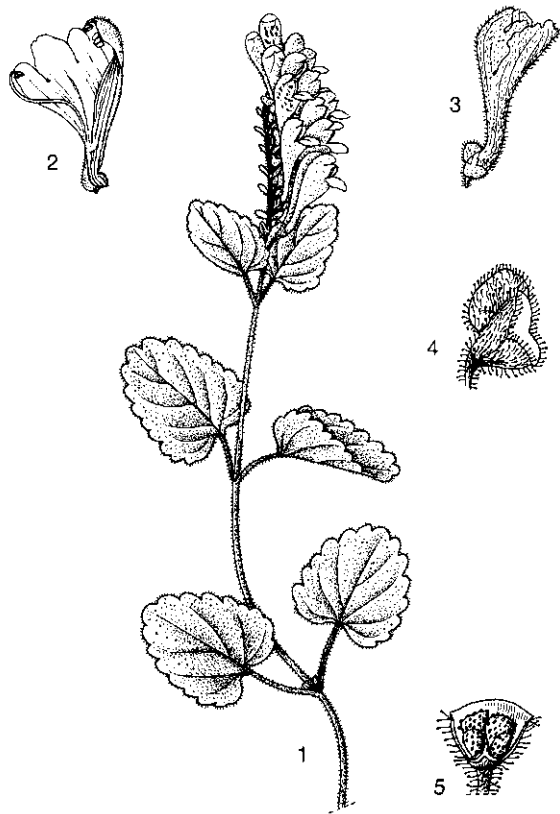
**Vernacular names** Indonesia: bangun bangun batu (Batak), daun kukuran (Java). Philippines: banod (Bagobo). Vietnam: thu[aax]n [aas]n d[ooj].

**Distribution** From India to Indo-China, China, Japan, Taiwan, the Philippines, Thailand, Sumatra, West Java, the Lesser Sunda Islands, Sulawesi, the Moluccas and New Guinea.

**Uses** In China, a decoction of *S. indica* is used as a folk remedy for traumatic injuries, whereas a poultice is applied to skin affected by fungal diseases. It is also carminative, tonic and resolves blood clots. In Indo-China its roots are regarded as febrifuge and also recommended for skin diseases such as scabies and boils.

**Observations** A small herb up to 30 cm tall, stems prostrate to ascending, often simple; leaves broadly ovate to rounded or reniform, 1.5-2(-3) cm × 1.5-2 cm, base cordate to truncate or rarely cuneate, margin crenate, appressed hirsute on both surfaces; bracts 2-3 mm long; flowers opposite, in a simple, terminal, 1-sided raceme of 2-8(-12) cm long, hirsute to puberulent, corolla pale to deep purple, (10-)15-20 mm long. *S. indica* has been subdivided into several varieties, but within Malesia only var. *indica* occurs. It is found infrequently on cliffs and boulders along streams, grassy open plains, along tracks in secondary forest, up to 2300 m altitude.

**Selected sources** 624, 720, 854, 1118, 1126, 1476.



*Scutellaria indica* L. - 1, flowering stem; 2, opened corolla; 3, flower; 4, calyx with scutellum; 5, nuts.

### *Scutellaria javanica* Jungh.

Java 1, ed. 2: 661 (1853).

**Synonyms** *Scutellaria horsfieldiana* Miq. (1859), *Scutellaria sumatrana* Miq. (1859), *Scutellaria russeliaefolia* Vatke (1872), *Scutellaria luzonica* Rolfe (1884).

**Vernacular names** Indonesia: kapunten, upar upar (Javanese), bangun bangun na gerger (Karo-Batak). Philippines: lupingan, sidit (Igorot). Vietnam: thu[aax]n java.

**Distribution** Indo-China, the Philippines, Sumatra, Java, the Lesser Sunda Islands, Sulawesi, the Moluccas and New Guinea.

**Uses** In the Philippines, a decoction of *S. javanica* is used to cure stomach pains.

**Observations** A herb or slender shrub up to 1 m tall, stems procumbent to erect, branched; leaves lanceolate to broadly ovate, 0.5-5(-10) cm × 0.3-2.5(-3.5) cm, base slightly cuneate or rounded to subcordate, margin usually remotely crenate or dentate, puberulent on both surfaces; bracts

(2-)3-6 mm long; flowers opposite, in a simple, terminal, 1-sided raceme of (2-)4-10(-20) cm long, hirsute to puberulent, corolla blue or white, 14-16 mm long. *S. javanica* has been subdivided into 4 varieties, mainly on characters concerning the leaves. One of these, var. *luzonica* (Rolfe) H. Keng is sometimes regarded as a distinct species. It is found in open, primary or secondary, lowland to montane forest, coffee estates, savanna forest, in ravines, on ridges in mossy forest, on various soils including peat overlying sand, up to 2850(-3300) m altitude.

**Selected sources** 190, 720, 854, 1118, 1126, 1178.

H.P. Hernandez

### Senna Miller

Gard. Dict. abr. ed. 4, vol. 3 (1754).

LEGUMINOSAE

$x = 11, 12, 13, 14$ ; *S. alata*:  $2n = 28$ , *S. garrettiana*:  $2n = 28$ , *S. sophera*:  $2n = 28$

**Major species** *Senna alata* (L.) Roxb., *S. sophera* (L.) Roxb., *S. tora* (L.) Roxb.

**Vernacular names** Malaysia: bebusok. Cambodia: angkanh. Laos: khi lek. Vietnam: mu[oo]f[ng].

**Origin and geographic distribution** *Senna* comprises about 260 species and has a pantropical distribution; a few species extend to temperate regions. Tropical Asia has far fewer *Senna* species than tropical America, Africa and Australia. Only about 7 species occur naturally in tropical Asia, and only about 5 in the Malesian region, including *S. tora*. Approximately 10 species have been introduced in Malasia and have become naturalized or even weedy (e.g. *S. alata* and *S. sophera*) others are planted as ornamentals and are only rarely found as escapes. *S. garrettiana* (Craib) Irwin & Barneby, which is an important medicinal species, is endemic to Indo-China and northern Thailand.

**Uses** The main medicinal uses of *Senna* in South-East Asia are in the treatment of skin problems and as a laxative or purgative. Skin problems treated with *Senna* include ringworm (e.g. *S. alata*, *S. garrettiana*, *S. hirsuta* (L.) Irwin & Barneby, *S. occidentalis* (L.) Link, *S. sophera* and *S. tora*), scabies (e.g. *S. alata*, *S. timoriensis* (DC.) Irwin & Barneby), eczema (*S. hirsuta*, *S. occidentalis*) and itching (*S. alata*, *S. timoriensis*). Laxative properties have been reported for *S. alata*, *S. garrettiana*, *S. obtusifolia* (L.) Irwin & Barneby, *S. occidentalis*, *S. siamea* (Lamk) Irwin & Barne-

by, *S. surattensis* (Burm. f.) Irwin & Barneby and *S. tora*.

The wood of *S. alata*, *S. garrettiana* and *S. siamea* is included in recipes for decoctions to treat liver problems, urticaria, rhinitis and loss of appetite caused by gastro-intestinal problems. The heartwood of *S. garrettiana* is the Thai drug 'Sae mae sarn', which is used as a mild purgative. *S. siamea* and *S. timoriensis* are used as vermifuge. In Indonesia, a decoction of young *S. siamea* leaves has been suggested for the treatment of malaria. In Burma (Myanmar), leaves, flowers and fruits of *S. siamea* are ingredients of a broth which is used as a tonic and to treat stomach problems. In Thailand, the heartwood of *S. siamea* is considered as a tranquilizer, antipyretic, and used in the treatment of venereal diseases; the leaves are used in the treatment of leucorrhoea; antihypertensive and antipyretic properties are ascribed to the flowers. The leaves of *S. obtusifolia* are used against vomiting and stomach-ache. The leaves of *S. occidentalis* are used in cases of toothache and headache. The seeds act as an emeto-cathartic and in the Philippines are used to treat fever. A decoction of the roots of *S. surattensis* is used against gonorrhoea, a decoction of the leaves is used against dysentery.

The bark of *S. alata* contains tanning material; the seeds are a promising source of gums. The bark of *S. auriculata* (L.) Roxb. is used as a source of tannin, but the astringent properties are also of medicinal importance. Toasted leaves of *S. alata* and the seeds of *S. tora* are sometimes used as a coffee substitute. The young pods of *S. alata* and the young leaves of *S. tora* may, in small quantities, be eaten as a vegetable. In West Africa, the roots of *S. tora* are used for tattoos or tribal markings.

**Properties** Many of the pharmacological effects of *Senna* species can be attributed to the presence of anthraquinone derivatives. The basic structure of these compounds is the 9,10-anthraquinone. They differ in the arrangement of the attached substituents. The derivatives may occur in various oxidation stages; anthraquinones can be reduced to anthrones, which may be oxidized to dianthrones. Dianthrones, on the other hand, can be reduced back to anthrones, which may oxidize to anthraquinones relatively easily.

Anthraquinone and dianthrone drugs are used as laxatives. The presence of sugar in the molecule is a prerequisite for their pharmacological action; it enhances their solubility in water, thus facilitating their transport to the site of action (the colon).

Bacteria in the colon hydrolyse the glycosides and dianthrones to anthraquinones, a reaction which is immediately followed by the local reduction of the anthraquinones to their corresponding anthrones. The latter compounds act directly on the large intestine, to stimulate peristalsis. However, laxative drugs containing anthraquinone derivatives should be used with caution, as daily and prolonged use can lead to dependence and 'cathartic colon'.

In anthrones, the explicit chemical structure is very reactive. An imperfectly understood mechanism enables these compounds to completely inhibit cell growth and thymidine incorporation in human cultured cells, and inhibit both DNA replication and repair synthesis. Because of these effects on the cell cycle, anthrones may be used as topical agents in the treatment of psoriasis. Their (topical) use in the treatment of skin infections, and their fungicidal properties (treatment of ringworm) have also been reported in the literature. Furthermore, their reactivity makes these compounds very irritant to the eyes and mucous membranes; they should never be used systemically.

Extensive phytochemical investigations have revealed much information about the constituents of several *Senna* species. Components isolated from the leaves of *S. alata* include the anthraquinone derivatives rhein (cassic acid), rhein-anthrone and aloe-emodin-anthrone, and flavonoids (e.g. kaempferol, glycosides). The yellow phenolic pigment cassiaxanthone has been isolated from the roots. *S. sophera* has been reported to contain the anthraquinones chrysophanol (chrysophanic acid) and emodin.

The reported constituents from *S. tora* seeds include anthraquinone derivatives (emodin, physcion, chrysophanol (chrysophanic acid), chrysophanol-triglucoside, chrysophanol-tetraglucoside, chryso-obtusin, aurantio-obtusin and obtusifolin-glucoside), naphtho- $\gamma$ -pyrones (cassiaside, rubrofusarin-gentobioside, rubrofusarin-glycoside), toralactone-gentobioside (a naphtho- $\alpha$ -pyrone), cassitoroside (a naphthalene glycoside) and  $\beta$ -sitosterol (a sterol). *S. tora* also contains the flavonoid glycoside kaempferol-3-sophoroside. Anthraquinone derivatives (chrysophanol (chrysophanic acid), chrysophanol benzanthrone, chrysophanol dianthrone, cassialoin, (-)-11-deoxyaloin), bibenzyl derivatives, flavonoids, stilbene derivatives and polyphenolic compounds (cassigarol A-G, scirpusin B) have been isolated from the heartwood of *S. garttiana*. Seeds of several *Senna* species (including *S. alata*, *S. hirsuta*, *S. occiden-*

*talis*, *S. siamea* and *S. tora*) are reported to contain the enzyme urease.

Some other pharmacological activities of anthraquinone derivatives and other *Senna* constituents in addition to the purgative effect are mentioned in the literature. In an assay with *Salmonella typhimurium* the methanol extract of *S. tora* seeds showed antimutagenic activity against aflatoxin B1. The numbers of revertants per plate decreased significantly when this extract was added to the assay system. The extract was not able to inhibit the direct-acting mutagen N-methyl-N'-nitro-N-nitrosoguanidine; this suggests that it may prevent the metabolic activation of aflatoxin B1 or scavenge the electrophilic intermediate capable of inducing mutations. Activity guided isolation yielded the anthraquinones chrysophanol (chrysophanic acid), chryso-obtusin and aurantio-obtusin, and the naphtho- $\gamma$ -pyrones cassiaside and rubrofusarin-gentobioside as pure compounds. All of these demonstrated significant antimutagenic activity.

*S. alata* extracts have shown antibacterial and antifungal properties (e.g. against *Pityriasis versicolor* in humans) and anti-tumour activity; they might be useful in the treatment of opportunistic infections in AIDS patients. Leaf extracts of *S. tora* also have antifungal activity. The major antifungal principle has been shown to be chrysophanol-anthrone (chrysophanic acid-anthrone), which is also present in the seeds.

Furthermore, the polyphenol cassigarol A (from *S. garrettiana*) inhibits H<sup>+</sup>/K<sup>+</sup>-ATPase, resulting in a reduced gastric acid secretion. The flavonoid glycoside kaempferol-3-sophoroside (from *S. tora*) has analgesic activity, as has been shown by intraperitoneal injections in mice and rats. Naphtho-pyrone glycosides in *S. tora* seeds have been found to protect the liver against galactosamine damage. The leaves of *S. siamea* exhibits central nervous system depressant effects, and the compound responsible for the activity is barakol.

Raw *S. tora* seeds have furthermore been found to be highly toxic in experiments with rats; pigs fed on leaves and pods of *S. siamea* in the Philippines died.

**Adulterations and substitutes** It is reported that in China *S. occidentalis*, *S. sophera*, *S. tora* and *Chamaecrista mimosoides* (L.) Greene are used medicinally almost without distinction. Anthraquinone glycosides and sennosides are also found in *Cassia* species, which are also used for their laxative and purgative properties.

**Description** Herbs, shrubs or small to medium-

sized trees up to 20(–30) m tall; bole usually short, up to 50 cm in diameter, bark surface smooth, greyish. Leaves alternate, paripinnate with up to 24(–40) pairs of leaflets, sometimes with extra-floral nectaries; stipules small, usually caducous. Inflorescence an axillary raceme, often becoming corymbose-paniculate towards the tips of branchlets, 1–many-flowered. Flowers bisexual, 5-merous; sepals imbricate, obtuse at apex; corolla with subequal to heteromorphic petals, yellow; androecium basically 10-merous, filaments all straight and never more than twice as long as anthers, accrescent towards the abaxial side of the flower; ovary superior, linear and curved. Fruit a stipitate, often strap-shaped, terete to compressed, indehiscent or inertly dehiscent pod, usually with septae between the numerous seeds. Seeds with distinct areole. Seedling with epigeal germination; cotyledons emergent, semi-fleshy.

**Growth and development** *S. alata* has Scarone's architectural model: an indeterminate trunk with tiers of orthotropic branches, which branch sympodially because they have terminal inflorescences. Most of the medicinally important *Senna* species are reported to flower and fruit throughout the year, though fluctuations occur in pronounced monsoon climates. Growth of *S. tora* is affected by photoperiod, and pods are only produced when plants receive 8–11 hours of light. Flowers are produced at 6–12 hours of light.

**Other botanical information** Until the beginning of the 1980s *Cassia* was considered to be a very large genus of over 500 species, but then it was split into 3 genera: *Cassia* sensu stricto, *Senna* and *Chamaecrista*. *Cassia* now has only about 30 species, whereas *Senna* and *Chamaecrista* comprise about equal numbers of species (about 260 and 270 respectively). *S. tora* is closely related to *S. obtusifolia*, which is similar in appearance, but *S. obtusifolia* can easily be recognized by its longer pedicels. *S. sophera* is closely related to *S. occidentalis*, and the two are often confused.

**Ecology** In South-East Asia, *S. alata*, *S. sophera* and *S. tora* are found in a multitude of habitats, but preferably in disturbed, rather open (anthropogenic) vegetation: roadsides, river banks, rain forest edges, lake shores, margins of ponds and ditches, in open forest and wet areas, in orchards and around villages. *S. alata* is found up to 1400(–2100) m altitude but is more common at lower elevations. The other species are only found at lower elevations: *S. sophera* up to 400 m and *S. tora* up to 1000 m altitude. *S. alata* and *S. tora* are reported to tolerate an annual rainfall from 600

mm to 4300 mm and average yearly temperatures of 15–30°C. Soils should retain moisture adequately, although *S. tora* is tolerant of considerable drought, whereas the pH may range between 4.3–8.0.

**Propagation and planting** *Senna* can be propagated by seed. Propagation by stem and root cuttings has not been successful. *S. tora* has hard seeds and mechanical and acid scarification are equally effective in breaking dormancy. *S. tora* germinates in continuous light or darkness and under alternating light conditions. No germination occurs below 13°C or above 40°C. Seed stored at 15–20°C germinates when scarified and placed in a 30°C environment. The stage of maturity at harvest affects seed viability and germination. In dry storage seed loses viability somewhat rapidly (germination 22% after 3 years). Germination is optimal when soil moisture is 75% of field capacity. Shoot and root dry matter production diminish at densities above 20 plants/m<sup>2</sup>. The number of flowers and fruits per plant drop at densities between 10 and 20 plants/m<sup>2</sup>. The average number of seeds/m<sup>2</sup> increases from about 40 (1 plant/m<sup>2</sup>) to a maximum of about 570 (17 plants/m<sup>2</sup>); it drops to about 145 at a density of 40 plants/m<sup>2</sup>.

**In vitro production of active compounds** Anthraquinones have been obtained from plant cell cultures of *S. tora*, with yields of 0.33% of the fresh weight.

**Diseases and pests** In Indonesia, brown leaf blight caused by *Cercospora* sp. is reported on *S. alata*. In Uganda, common bean mosaic necrosis virus has been isolated from *S. sophera*. In India, *S. tora* is reported to be attacked by the defoliator *Myllocerus viridanus* and the bruchids *Callosobruchus chinensis*, *Bruchidius cassiae* and *Caryedon lineatonota*. The seed bruchid *Sennius instabilis* has been suggested as a potential biological agent for controlling *S. tora* in pastures.

**Harvesting** The leaves of *S. alata* are harvested when needed. The active constituents are probably most abundant prior to flowering, which is why the leaves are preferably collected at that time. The pods of *S. tora* are collected when mature.

**Handling after harvest** After harvesting, *S. alata* leaves are dried and sometimes stored in containers until needed. The mature pods of *S. tora* are sun-dried; prior to use the seeds are removed and roasted.

**Genetic resources and breeding** *S. alata*, *S. sophera* and *S. tora* are widely found wild and cultivated in and outside South-East Asia, and they

are neither endangered nor liable to genetic erosion.

**Prospects** As *Senna* species have various medicinal properties, ornamental value, and are used for various other purposes, they are true multi-purpose plants. The antifungal and anti-tumour properties seem to justify more research.

**Literature** [1] Baba, K., Kido, T., Taniguchi, M. & Kozawa, M., 1994. Stilbenoids from *Cassia garrettiana*. *Phytochemistry* 36(6): 1509–1513. [2] Boer, E. & Lemmens, R.H.M.J., 1998. *Senna* Mill. In: Sosef, M.S.M., Hong, L.T. & Prawirohatmodjo, S. (Editors): *Plant Resources of South-East Asia No 5(3). Timber trees: Lesser-known timbers*. Backhuys Publishers, Leiden, the Netherlands. pp. 522–524. [3] Bruneton, J., 1995. *Pharmacognosy, phytochemistry, medicinal plants*. Lavoisier Publishing, Paris, France. pp. 349–366. [4] Bulyalert, D., 1993. Effect of barakel on the central nervous system: qualitative analysis of BEG in the rat. *Chiangmai Medical Bulletin* 32(4): 191–196. [5] Burkill, I.H., 1966. *A dictionary of the economic products of the Malay Peninsula*. Revised reprint. Vol. 1. Ministry of Agriculture and Cooperatives, Kuala Lumpur, Malaysia. pp. 478–488. [6] Faridah Hanum, I. & van der Maesen, L.J.G. (Editors), 1997. *Plant Resources of South-East Asia No 11. Auxiliary plants*. Backhuys Publishers, Leiden, the Netherlands. pp. 229–236, 294–295. [7] Hata, K., Baba, K. & Kozawa, M., 1979. Chemical studies on the heartwood of *Cassia garrettiana* Craib. II. Nonanthraquinonic constituents. *Chemical and Pharmaceutical Bulletin* 27(4): 984–989. [8] Holm, J., Doll, J. & Holm, E., 1997. *World weeds: natural histories and distribution*. Wiley, New York, United States. pp. 158–171. [9] Larsen, K. & Ding Hou, 1996. *Senna*. In: Kalkman, C., Kurkop, D.W., Nootboom, H.P., Stevens, P.F. & de Wilde, W.J.J.O. (Editors): *Flora Malesiana. Series 1, Vol. 12(2)*. Rijksherbarium/Hortus Botanicus, Leiden University, the Netherlands. pp. 673–691. [10] Nguyen Van Duong, 1993. *Medicinal plants of Vietnam, Cambodia and Laos*. Mekong printing, Santa Ana, California, United States. pp. 92–96.

*Selection of species*

***Senna alata* (L.) Roxb.**

Fl. ind., ed. 2, 2: 349 (1832).

**Synonyms** *Cassia alata* L. (1753).

**Vernacular names** Ringworm bush, seven golden candlesticks (En). Dartrier (Fr). Brunei:

raun suluk (Dusun), paa-ul, tarump (Malay). Indonesia: daun kupang, ketepeng (Malay, Manado), ketepeng kebo (Javanese). Malaysia: daun kurap, gelenggang, ludanggan (Peninsular). Papua New Guinea: kabaiura (Harigen, Sepik), levoanna (Gaire and Tubusereia, Central Province), Orere (Awala, Northern Province). Philippines: andadasi (Iloko), katanda (Tagalog), palochina (Bisaya). Cambodia: dang het, dang het khmoch. Laos: khi let ban. Thailand: kheekhaak (northern), chumhet thet (central, peninsular), chumhet yai (central). Vietnam: mu[oof]ng l[as]c, mu[oof]ng tr[aa]u.

**Distribution** Native to South America, now pantropical; abundantly naturalized in South-East Asia, and occasionally planted throughout the region for medicinal and ornamental purposes.

**Uses** *S. alata* is used against ringworm and various other skin diseases, such as scabies. Ringworm is treated by externally applying the leaves, which might be pounded first, the sap of the leaves, the roots, or the pods. *S. alata* is also used as a laxative or purgative. For laxative purposes, a decoction of the leaves, the flowers, the roots or the wood may be used. In India, the leaves are taken as a purgative, while leaf decoctions are used as an expectorant in bronchitis and dyspnea, as an astringent, a mouthwash and a wash in cases of eczema. *S. alata* is sometimes a weed in pastures; it is not eaten by livestock and sometimes even reported to be poisonous.

**Observations** A shrub, 1–2(–5) m tall, with thick branches, pubescent; leaves with 8–20 pairs of leaflets, petiole 2–3 cm long, rachis 30–60 cm long, leaflets oblong-elliptical, 5–15 cm × 3–7 cm, obtuse at the ends, glabrous; inflorescence an axillary raceme, robust, dense, 20–50 cm long, many-flowered; flowers with oblong sepals, 10–20 mm × 6–7 mm, orange-yellow, petals ovate-orbicular, 16–24 mm × 10–15 mm, bright yellow, 2 large stamens with stout filaments 4 mm long and anthers 12–13 mm long, 4 small stamens with filaments 2 mm long and anthers 4–5 mm long, 3–4 staminodes, ovary puberulous, sessile, with many (up to 58) ovules, style filiform, stigma small; fruit tetragonal, 10–15 cm × 1.5–8 cm, winged, wings 4–8 mm, black, glabrous, up to 50-seeded; seeds quadrangular, flattened, 7–8 mm × 5–8 mm, shiny. *S. alata* occurs up to 2100 m altitude in New Guinea but is most abundant below 500 m altitude. It has a wide ecological amplitude, but prefers rather open, not too dry habitats.

**Selected sources** 97, 190, 202, 287, 295, 308, 336, 350, 357, 364, 402, 409, 522, 553, 580, 597,

653, 817, 829, 1035, 1105, 1126, 1128, 1178, 1220, 1277, 1287, 1520.

***Senna sophera* (L.) Roxb.**

Fl. ind., ed. 2, 2: 347 (1832).

**Synonyms** *Cassia sophera* L. (1753).

**Vernacular names** Indonesia: enceng-enceng (Javanese). Philippines: andadasi (Iloko), tambalisa (Tagalog). Laos: ngot. Thailand: phak khlet (Bangkok), phak waan baan (central). Vietnam: mu[oof] ng[os]t, mu[oof]ng h[of]e.

**Distribution** Originating from the New World tropics, but now pantropical.

**Uses** In Indonesia, extracts of all plant parts are used to treat epilepsy. In the Philippines, the seeds are used to treat fever. In India, the juice of the leaves is applied against ringworm, while it is also employed as an expectorant, an anthelmintic and as a remedy for rheumatic and inflammatory fevers. Besides these applications, in Thailand the leaves are used for wound healing and as an antipyretic.

**Observations** An erect shrub, 1-2(-3) m tall, almost glabrous; leaves with 4-10 pairs of leaflets, petiole 3-5 cm long, with a thin, subulate or clavate gland 5-10 mm above the petiole joint, leaflets lanceolate, 2-5(-8) cm × 1-2 cm, upper leaflets largest, base rounded, apex acute; inflorescence an axillary corymb, few-flowered; flowers with ovate sepals 5 mm long, petals obovate, 10-14 mm × 6-8 mm, yellow, 2 longer stamens with filaments 5-7 mm long and anthers 5-6 mm long, 4 shorter stamens with filaments 2 mm long and anthers 5 mm long, 3-4 staminodes, ovary pubescent, style thin, glabrous, stigma slightly dilated, strongly incurved; fruit 6-10 cm × 0.5-1 cm; seeds 30-40 per pod, ovoid, compressed, 3-4 mm long. *S. sophera* occurs in secondary habitats such as roadsides and wast places at lower elevations.

**Selected sources** 97, 190, 336, 357, 580, 653, 817, 1178.

***Senna tora* (L.) Roxb.**

Fl. ind., ed. 2, 2: 340 (1832).

**Synonyms** *Cassia tora* L. (1753), *Cassia borneensis* Miq. (1850).

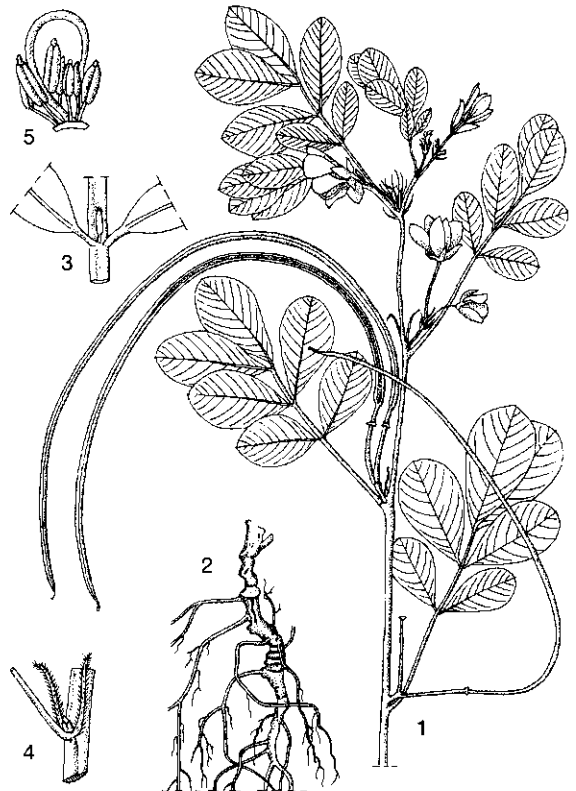
**Vernacular names** Foetid cassia, paniced milkwort (En). Casse fétide (Fr). Indonesia: gelenggang kecil, ketepeng kecil (Malay), katepingleutik (Sundanese). Malaysia: gelenggang kechil, gelenggang padang. Philippines: katanda, balatong-aso (Tagalog). Cambodia: d[aa]ng'hët khmaoch ni. Laos: nha leung meum, nha lap mun. Thailand: chum het tai, chum het lek (central),

lapmuen noi (northern). Vietnam: th[ar]o quy[ees]t minh, mu[oof]ng h[oo]i, mu[oof]ng ng[ur].

**Distribution** The origin of *S. tora* is unknown, but nowadays it is only found in the Old World tropics.

**Uses** A decoction of the leaves may be used as a purgative, vermifuge or to treat cough. The leaves and seeds are used in the treatment of ringworm and scabies, as a diuretic and an antipyretic. In Indonesia and the Philippines, pounded leaves are smeared on the head of restless children. In Vietnam, seeds are used as a laxative, and roasted seeds are employed to treat insomnia, hypertension and ophthalmia. In India, pounded seeds are used to treat itching. In Chinese medicine, *S. tora* is used to improve vision. Cattle do not eat the green plant, but do eat silaged plants and dry pods.

**Observations** A foetid smelling herb or under-



*Senna tora* (L.) Roxb. - 1, twig with flowers and fruits; 2, root system; 3, detail of gland on leaf rachis; 4, detail of stipules; 5, detail of stamens and pistil.



shrub, up to 1 m tall, almost glabrous, with a stout taproot; leaves with 3 pairs of leaflets, petiole 1–4 cm long, rachis 2–3 cm long, with a subulate, 2 mm long gland between the two lowest pairs of leaflets, leaflets obovate, 2–5 cm × 1.5–2.5 cm, upper leaflets largest, thin, base cuneate to rounded, apex rounded; inflorescence a short axillary raceme, 1–2(–3)-flowered; flowers with ovate sepals, 5 mm × 2–4 mm, petals obovate, unequal, up to 10 mm × 6 mm, yellow, stamens 7, almost equal, filaments 1.5–2 mm long, 3 larger anthers, 4 smaller ones 1.5–2.5 mm long, staminodes absent, ovary densely pubescent, style glabrous, stigma truncate; fruit linear, terete, 10–15 cm × 0.5 cm; seeds 20–30 per pod, rhomboidal, 5 mm × 3 mm, glossy, with an areole (1.5–2 mm wide) covering part of the seed surface. *S. tora* is common throughout South-East Asia in anthropogenic habitats at lower altitudes. It is reported to be an important weed in pastures and cropped fields in South-East Asia, partly because of its high seed production.

**Selected sources** 8, 97, 190, 202, 253, 254, 272, 336, 357, 364, 402, 429, 503, 553, 580, 817, 986, 988, 1035, 1126, 1178, 1189, 1197, 1220, 1277, 1287, 1520, 1594.

Anny Victor Toruan-Purba

## Smilax L.

Sp. pl. 2: 1028 (1753); Gen. pl. ed. 5: 455 (1754).

SMILACACEAE

$x = 16$ ; *S. bracteata*:  $2n = 32$ , *S. china*:  $2n = 30$ , 60, 90, *S. leucophylla*:  $2n = 32$ , *S. zeylanica*:  $2n = 32$

**Major species** *Smilax china* L., *S. glabra* Wallich ex Roxb., *S. leucophylla* Blume.

**Vernacular names** Indonesia: gadung cina. Malaysia: akar gadong. Philippines: sarsaparilla, banag. Cambodia: voë pâprâ:hs. Laos: khua:ng. Thailand: khueang. Vietnam: d[aa]ly kim cang.

**Origin and geographic distribution** *Smilax* contains approximately 200 species which are distributed in tropical and subtropical regions, with some herbaceous species extending their range into the temperate regions of North America, Europe and Asia.

The greatest diversity is found in eastern Asia. In Indo-China 27 species have been found, in Thailand 24 species, and the number of species is probably slightly less in Malasia.

**Uses** In South-East Asia, decoctions of *Smilax* roots and rhizomes are rather commonly used in-

ternally to treat syphilis, gonorrhoea, rheumatism and coughs and as a tonic (e.g. after childbirth) and aphrodisiac, and externally to treat skin diseases including psoriasis, wounds, inflammations, swellings, ulcers and boils. Leaf tops have been reported to be eaten with food as contraceptive in Papua New Guinea. A decoction of the leaves is used as a purgative to expel worms and reduce swelling of the stomach in the Central Province in Papua New Guinea. The rhizomes of *S. calophylla* and *S. myosotiflora* are reputed to be used in Peninsular Malaysia as an aphrodisiac. *Smilax* species, particularly *S. china*, are used in anti-cancer drugs in Chinese medicine, and have been reputed to be effective as diuretic and anti-inflammatory agents. A decoction of *S. glabra* rhizomes is used in China in orally administered mixtures of medicinal plants for the treatment of psoriasis, and in Vietnam as an antiphlogistic to treat rheumatic arthritis, psoriasis and inflammations, and as a tonic. In Australia, an infusion of the leaves of *S. australis* R.Br. and *S. glycopylla* Smith has been used medicinally. In Guatemala and other Central American and West Indian countries, *S. regelii* Killip & C. Morton ('Honduras-sarsaparilla') and *S. officinalis* Kunth are used to treat dermatophyte infections and rheumatism. *S. aristolochiifolia* Miller from Mexico, known as 'Veracruz-sarsaparilla', is reputed as a diuretic, increasing both volume and chloride and uric acid concentration of the urine. *S. papyracea* Duhamel from Mexico, Brazil and Guyana is also known as 'sarsaparilla'. In Saudi Arabia *S. sarsaparilla* L. is used for the treatment of rheumatism, arthritis, gout and other forms of inflammation. The roots of *S. zeylanica* and *S. wightii* A.DC. are used medicinally in India against amongst others, rheumatism, urinary complaints and venereal diseases.

Thickened roots of *S. china* can be used as bait for subterranean termites, in conjunction with insecticides. The stems are sometimes used as cane to make baskets. The edible but acid fruits of *S. macrocarpa* Blume are commonly collected from the wild in Java, eaten fresh, as an ingredient of fruit salad or preserved in syrup. Young shoots and leaves of *S. leucophylla* and rhizomes of *S. megacarpa* are edible.

**Production and international trade** The rhizomes of some *Smilax* spp. are much sought after and are exported in fairly large quantities; for example, rhizomes of *S. glabra* in Laos. Roots of *S. china* and *S. glabra* are mainly imported from China and sold as herbal medicine in Chinese

pharmacies throughout South-East Asia.

**Properties** Roots and rhizomes of *Smilax* species are a well known source of steroidal saponins (1–3%): mainly glycosides of the furostanol and spirostanol aglycones sarsapogenin, smilagenin, tigogenin, neotigogenin, diosgenin and yamogenin. In the plant, the bis-desmosidic glycoside form of these aglycones seems to be the genuine component. Several steroidal saponins have been isolated from *S. china* rhizomes; they include dioscin, protodioscin, methylprotodioscin, gracillin and methylprotogrillin. The aglycones of these saponins are of great industrial interest as starting material for steroid hormone semisynthesis in the production of corticosteroids and sex hormones (androgens, oral contraceptives). The spirostanol saponins dioscin and gracillin are reported to have antimutagenic activity, but their similar furostanol glycosides have not shown this activity. Hot water extract of *S. china* shows histamine release inhibition and cytotoxic activities. Methanol extract brings about uterine relaxation and decreases barbiturate sleeping time.

The flavonoids astilbin, taxifolin and engeletin have been isolated from the dried rhizomes of *S. glabra*, together with some organic acids (ferulic acid, 3-O-caffeoylshikimic acid) and  $\beta$ -sitosterol. Quercetin and kaempferol were isolated from leaf extracts of the same plant after acid hydrolysis. A methanol extract of *S. glabra* rhizomes showed hypoglycaemic effects in normal and diabetic mice. *S. glabra* extracts showed anthelmintic activity against the trematode *Clonorchis sinensis*. The extracts also exhibit antiviral and anti-inflammatory activities. Extracts of *S. china* have been demonstrated to inhibit the mutagenicity of benzo(a)pyrene completely. Tests in Saudi Arabia showed significant inhibition of carrageenan-induced inflammation in rats when an oral dose of 500 mg/kg of *S. sarsaparilla* extract was applied; this extract also inhibited cotton pellet-induced exudation. In tests in Guatemala, extracts of *S. lundellii* Killip & C. Morton showed activity against *Candida albicans*, a parasitic imperfect fungus causing thrush.

In tests in China, a mixture of medicinal plants including a decoction of *S. glabra* rhizomes showed a lower relapse rate and longer remission in comparison with aminopterin therapy and the topical application of psoriasis to treat psoriasis.

**Adulterations and substitutes** The nodular growths found on the roots of fir (*Abies*) trees and produced by a fungus, and similar growths on *Liq-*

*uidambar* roots are sometimes sold as a substitute for China root (from *S. china*). Saponins of the same type are found in various *Yucca* species; other saponins which are interesting as starting material for steroid hormone semisynthesis include diosgenin from *Dioscorea* spp. and hecogenin from *Agave* spp.

**Description** Dioecious climbers up to 20 m long with woody or herbaceous stems up to 12 mm in diameter, or becoming shrubby with suberect stems; stem and branches often prickly; rhizome and/or roots often thickened. Leaves alternate, simple and entire, mostly ovate-orbicular, ovate to lanceolate, with 3–7 veins connected by reticulate lateral veinlets; petiole sheathing in lower part with a pair of wings, often with tendrils arising immediately above the apex of the sheath. Inflorescence an axillary umbel or an umbellate raceme whose main axis bears a prophyll at the base. Flowers unisexual, small, usually greenish or greenish-white; perianth with 6 free, usually recurved or patent tepals, inner tepals much narrower than outer ones; male flowers with 6(–18) free or almost connate stamens having unilocular anthers; female flowers with a superior, 3-locular ovary, style very short and with 3 elongate, recurved stigmas, 3–6 needle-like staminodes present. Fruit a globose berry, usually containing 1–2 seeds.

**Growth and development** In Java, most *Smilax* spp. flower in the period from April to September and sometimes also January–February. Pollination is by insects.

**Other botanical information** *Smilax* has often been treated as a genus of the family *Liliaceae*, but is nowadays considered as a separate family *Smilacaceae*, together with *Heterosmilax*, which differs in its connate perianth segments and 3 stamens (rarely up to 12) and occurs from India to Japan, and *Rhipogonum*, which differs by not having sheaths and tendrils on the petioles, or bilocular anthers, and occurs in New Guinea, Australia and New Zealand. Chromosome features support the removal from the Liliaceous assemblage.

**Ecology** *Smilax* species are often common climbers in evergreen lowland and lower montane forest up to 2400 m altitude. Several species are frequently found in scrub vegetation, open forest and shrub savanna, often on stony soils.

**Propagation and planting** Tests with seed of the Australian *S. australis* showed a germination rate of only 10–20% after 12 months. Adventitious roots are formed in some species (e.g. *S. zeylani-*

ca), which might enhance modes of vegetative propagation. Cuttings are a successful method of propagation. Mature 3-leaved stem sections treated with rooting powder and placed in pots with 10 cm diameter sealed in plastic bags and with a 1:1 sand/peat mixture showed callus tissue formation in the leaf axils within one month, when kept at 25°C and in 70% shade. The aerial calli developed roots and shoot buds after a further 3–4 weeks, and about 60% of the cuttings established successfully.

**Harvesting** In China, the tuberous rhizomes of *S. china* are dug out and dried for the market.

**Handling after harvest** In China, the rhizomes are often peeled after drying. Usually the crude drug is extracted with boiling water for 2 hours, to prepare the drug for oral application.

**Genetic resources and breeding** Those *Smilax* spp. that are generally used medicinally are widespread and common and seem not to be easily endangered. Despite the multiple uses, no serious attempts have been made either to conserve the genetic variation, or to domesticate these species.

**Prospects** The medicinal uses of *Smilax* are more or less the same world-wide, which may indicate a certain consensus on activity of the drug. *Smilax* has good prospects as starting material for steroid hormone semisynthesis. Research on properties has been carried out mainly on Chinese and Japanese *Smilax* species, but little is known about those found in Malesia. The taxonomy of Malesian species is also very incompletely investigated, and this hampers the interpretation of findings in literature.

**Literature** [1] Chang, C.L., Huang, C.Y., Yao, Y.C., Ch'ien, W.Y. & Chao, P.N., 1974. 200 cases of psoriasis treated with traditional Chinese medicine. *Chinese Medical Journal* 4: 205–207. [2] Chien, N.Q. & Adam, G., 1979. Über die Inhaltsstoffe von *Smilax glabra* Roxb. [The constituents of *Smilax glabra* Roxb.]. *Pharmazie* 34(12): 841–843. [3] Koyama, T., 1975. Smilacaceae. In: Smitinand, T. & Larsen, K. (Editors): *Flora of Thailand*. Vol. 2. The Forest Herbarium, Royal Forest Department, Bangkok, Thailand. pp. 211–250. [4] Koyama, T., 1983. Smilacaceae. In: Tirel, C. (Editor): *Flore du Cambodge, du Laos et du Vietnam*. Vol. 20. Muséum National d'Histoire Naturelle, Paris, France. pp. 69–124. [5] Lee, H. & Lin, J.Y., 1988. Antimutagenic activity of extracts from anticancer drugs in Chinese medicine. *Mutation Research* 204(2): 229–234. [6] Quisumbing, E., 1978. *Medicinal plants of the Philippines*. Katha Publishing Co., Quezon City, the Philippines. pp.

168–170. [7] Rhee, J.K., Woo, K.J., Baek, B.K. & Ahn, B.J., 1982. Screening of the wormicidal Chinese raw drugs on *Clonorchis sinensis*. *American Journal of Chinese Medicine* 9(4): 277–284. [8] Sashida, Y., Kubo, S., Mimaki, Y., Nikaido, T. & Ohmoto, T., 1992. Steroidal saponins from *Smilax riparia* and *S. china*. *Phytochemistry* 31(7): 2439–2443. [9] Ungson, L.B. & Sastrapradja, S., 1976. Variation in *Smilax* species of Java. *Biotrop Bulletin* No 12. 24 pp. [10] Yi, Y.J., Cao, Z.Z., Yang, W.H., Hong, W.Q., Cao, Y. & Leng, Z.K., 1995. Chemical studies of *Smilax glabra* (III). Isolation and identification of smiglanin from *Smilax glabra*. *Acta Pharmaceutica Sinica* 30(9): 718–720.

#### *Selection of species*

#### **Smilax blumei A.DC.**

Monogr. phan. 1: 202 (1878).

**Distribution** The Andaman Islands, peninsular Thailand, Peninsular Malaysia, Java, New Guinea and north-eastern Australia.

**Uses** *S. blumei* is probably used for similar medicinal purposes as *S. leucophylla*. The roots are boiled by the Kedayan people in Sarawak to make a drink for treating rheumatism.

**Observations** A climber up to 20 m long with sparsely prickly stem and branches; leaves ovate-elliptical, up to 25 cm long, petiole up to 3 cm long, wings of petiolar sheaths distinct, tendrils present; inflorescence with (1–)3–10 umbels; fruit about 8–12 mm in diameter, dark purple to black at maturity. *S. blumei* occurs in evergreen forest up to 1500 m altitude. It has often been confused with *S. leucophylla*, which can be distinguished by its glaucous leaf undersurface and thicker leaves.

**Selected sources** 278, 779, 780, 1227.

#### **Smilax bracteata K. Presl**

Reliq. haenk. 1: 131 (1827).

**Vernacular names** Philippines: banag (general), kamagsa-obat (Tagalog), banagan (Bisaya). Cambodia: voë pâprohs dâm ròi (Pursat). Laos: khua:ng khua (Xieng Khouang), khua:ng th'ò:n (Vientiane). Thailand: khueang thon (Loei). Vietnam: c[aa]j m k[ee]j ch.

**Distribution** From southern Japan, through the Ryukyu Islands and Taiwan to Cambodia, Laos, Vietnam, Thailand, Peninsular Malaysia, Java, Borneo and the Philippines.

**Uses** In the Philippines, a decoction of the rhizomes is used as an emmenagogue; it is considered as depurative.

**Observations** A climber up to 6 m long with smooth or sparingly prickly, sometimes verruculose stem and branches; leaves broadly elliptical to broadly ovate-elliptical or lanceolate, up to 22 cm long, petiole up to 3 cm long, wings of petiolar sheaths weakly developed, tendrils up to 20 cm long; inflorescence with (2-)3-7 umbels; fruit 5-12 mm in diameter, dirty yellow-brown turning purple-brown or shiny black at maturity; seeds dark red. Two subspecies have been distinguished: subsp. *bracteata* (synonym: *S. stenopetala* A. Gray) with usually smooth or nearly smooth stem and branches, and subsp. *verruculosa* (Merr.) T. Koyama (synonyms: *S. odoratissima* Blume, *S. verruculosa* Merr.) with densely verruculose and usually sparingly prickly stem and branches; the latter subspecies occurs in Taiwan, northern Thailand, the Philippines, Java and Borneo. *S. bracteata* occurs in evergreen or lower montane forest up to 2400 m altitude.

**Selected sources** 97, 190, 779, 780, 781, 1178, 1491.

### ***Smilax calophylla* Wallich ex A.DC.**

Monogr. phan. 1: 60 (1878).

**Vernacular names** Malaysia: dawai-dawai, sedawai, akar kancil (Peninsular).

**Distribution** Peninsular Thailand, Peninsular Malaysia, Sumatra, New Guinea and north-eastern Australia.

**Uses** Rhizomes are used in Malaysia to treat gonorrhoea and as an aphrodisiac and tonic. A decoction is administered after childbirth. Leaves are smeared with coconut oil, heated and applied to swellings.

**Observations** An erect to straggling slender shrub up to 2.5 m tall with smooth stems and branches; leaves elliptical to lanceolate-elliptical or narrowly ovate, up to 20 cm long, petiole up to 2 cm long, wings of petiolar sheaths not well developed, tendrils absent; inflorescence with 2-7 umbels; fruit 7-9 mm in diameter, reddish-brown at maturity. *S. calophylla* occurs in evergreen forest up to 1600 m altitude.

**Selected sources** 202, 779, 780, 1227.

### ***Smilax china* L.**

Sp. pl. 2: 1029 (1753).

**Vernacular names** China root, Chinese sarsaparilla (En). Racine de Chine, squine (Fr). Indonesia: gadung china (general), peundang (Aceh), ghadhung tambha (Madurese). Malaysia: gadong china, gadong saberang, akar restong. Philippines: sarsaparillang-china (Tagalog), buanal

(Igorot), palipit (Bontok). Vietnam: kim chang trung qu[oo]sle.

**Distribution** From Japan and southern China, through the Ryukyu Islands to Laos, Vietnam, northern Thailand, Burma (Myanmar) and the Philippines. *S. china* has long been cultivated in China and Japan as a medicinal plant. The rhizomes are imported in Indonesia and Malaysia.

**Uses** Rhizomes had a long-standing reputation as a remedy for syphilis, complaints of the genitourinary system, rheumatism and skin diseases. *S. china* is used in anticancer drugs in Chinese medicine. In Malaysia, the imported rhizomes are used against syphilis and gonorrhoea, as a tonic and after childbirth; in Indonesia against syphilis and framboesia; in the Philippines against herpes, syphilis, chronic rheumatism, skin diseases and asthma.

**Observations** A climber up to 5 m long with smooth or sparingly prickly stem and branches; leaves ovate-orbicular, broadly elliptical to ovate-elliptical or narrowly elliptical, up to 12 cm long, petiole up to 1.5 cm long, wings of petiolar sheaths



*Smilax china* L. - 1, rhizome; 2, part of flowering stem; 3, male flower.

rather weakly developed, tendrils up to 15 cm long; umbels solitary; fruit 10–12 mm in diameter, red at maturity; seeds red-purple. In China, two types can be distinguished, one with big berries and the other with small berries; these types can be distinguished by pyrolysis-high resolution gas chromatography. In tropical Asia *S. china* occurs only in scrub vegetation and open forest in mountains above 1000 m altitude, but in more temperate regions it also grows in the lowland; in the Philippines it is found in moss forest at 1600–2400 m altitude.

**Selected sources** 190, 202, 549, 580, 735, 779, 780, 781, 832, 1178, 1288, 1570.

### *Smilax corbularia* Kunth

Enum. pl. 5: 262 (1850).

**Vernacular names** Cambodia: voë chon (Koh Kong). Laos: khuang khau (Xieng Khouang). Thailand: hua khaao-yen wok, hua khaao-yen nuea (northern). Vietnam: d[aa]y g[aj]o (Thanh Hoa), d[aa]y kim cang (Thanh Hoa, Quang Nam-Da Nang), t[aa]f[u] mu[oos]i (Ha Tuyen).

**Distribution** From Burma (Myanmar) and southern China through Indo-China and Thailand, to Peninsular Malaysia and Borneo.

**Uses** In Thailand, rhizomes are used for the treatment of gonorrhoea, urinary infections, inflammation, skin diseases and as diuretic. Leaves are used as antipyretic. A decoction of the rhizome is applied to wounds in Cambodia. A decoction of the stems is used to stimulate the appetite in Vietnam, and in Laos the leaves are used to prepare a popular drink.

**Observations** A climber up to 4 m long with smooth stem and branches; leaves elliptical to ovate or broadly ovate, up to 10 cm long, petiole up to 1.5 cm long, wings of petiolar sheaths rather weakly developed, tendrils up to 9 cm long; umbels solitary, sometimes up to 3 together; fruit 6–8 mm in diameter, purple-black at maturity. Two subspecies are distinguished: subsp. *synandra* (Gagnepain) T. Koyama (synonym: *S. synandra* Gagnepain) and subsp. *corbularia* (synonyms: *S. balansaeana* H. Bon ex Gagnepain, *S. hypoglauca* Benth., *S. peguana* A.DC.). The former differs from the latter in the partly connate perianth and stamens, and tetragonous stems, and it is found in China, Vietnam and north-eastern Thailand. *S. corbularia* occurs in evergreen and lower montane forest up to 2000 m altitude, in Peninsular Malaysia on exposed rocks in the mountains.

**Selected sources** 779, 780, 781, 1227.

### *Smilax glabra* Wallich ex Roxb.

Fl. ind., ed. 1832, 3: 792 (1832).

**Vernacular names** Cambodia: p[ap]r[aa]:hs (Koh Kong), voë sr[aa]:m (Kampot). Laos: ya: hu:a (general), khuang la:y (Khammouane). Thailand: yaa hua (north-eastern). Vietnam: th[oor] ph[u]c linh (general), d[aa]y ch[aws]it (Vinh Phu), d[aa]y khum (Phu Khanh).

**Distribution** India, Burma (Myanmar), Cambodia, Laos, Vietnam, Thailand, central and southern China and Taiwan.

**Uses** A decoction of rhizomes is used in China in orally administered mixtures of medicinal plants for the treatment of psoriasis, and as anthelmintic, and in Vietnam as antiphlogistic to treat rheumatism and inflammations.

**Observations** A climber with smooth stem and branches; leaves lanceolate to lanceolate-elliptical or narrowly ovate, up to 18 cm long, petiole up to 3 cm long, wings of petiolar sheaths weakly developed, tendrils up to 15 cm long; umbels solitary;



*Smilax glabra* Wallich ex Roxb. - 1, rhizome; 2, stem with male inflorescences; 3, stem with infructescences; 4, female flower; 5, male flower.

fruit 5–8 mm in diameter, blue-black at maturity. In Thailand and Indo-China, *S. glabra* occurs in evergreen forest and shrub savanna at 300–1500 m altitude, often on stony soils.

**Selected sources** 235, 249, 449, 549, 779, 780, 781, 1035, 1128, 1224, 1633.

### ***Smilax lanceifolia* Roxb.**

Fl. ind., ed. 1832, 3: 792 ('*lanceaefolia*', 1832).

**Vernacular names** Thailand: dao, naam dao (northern), thao yang dong (south-eastern). Vietnam: d[aa]y kim cang.

**Distribution** From the Indian Himalayas, south-eastern China and Taiwan to Burma (Myanmar), Cambodia, Laos, Vietnam, Thailand, Peninsular Malaysia, Sumatra, Java, Borneo and the Philippines.

**Uses** Leaves and fruits are used in traditional medicine in Vietnam. A decoction of the roots is used against syphilis and rheumatism. The fruits are edible.

**Observations** A climber up to 5 m long with smooth or sometimes sparsely prickly stem and branches; leaves broadly lanceolate to elliptical or ovate-oblong, up to 15 cm long, petiole up to 2.5 cm long, wings of petiolar sheaths weakly developed, tendrils up to 20 cm long; umbels usually solitary; fruit 5–7 mm in diameter, yellowish-red-dish at maturity. Two subspecies are distinguished: subsp. *reflexa* (Norton) T. Koyama (synonym: *S. chapaensis* Gagnepain) and subsp. *lanceifolia* (synonyms: *S. laevis* Wallich ex A.DC., *S. micropoda* A.DC., *S. opaca* (A.DC.) Norton). The former differs from the latter in its verrucose and often also prickly stem and branches, and is found in China and northern Vietnam. *S. lanceifolia* is a common climber in evergreen and lower montane forest and shrub savanna at 500–2000 m altitude.

**Selected sources** 779, 780, 781, 1227.

### ***Smilax leucophylla* Blume**

Enum. pl. Javae 1: 18 (1827).

**Vernacular names** Indonesia: canar bokor (Sundanese). Philippines: sarsaparillang-puti (Tagalog), banag (Tagbanua), kaguno (Negrito). Papua New Guinea: wanabekira.

**Distribution** Peninsular Malaysia, Sumatra, Java, Borneo and the Philippines to New Guinea and northern Australia (once collected).

**Uses** Pounded rhizomes and leaves are applied to ulcerations of the nose in Malaysia. In the Philippines, the rhizomes are considered as a purifier of the blood and used in cases of syphilis, rheumatism and skin diseases. In Papua New

Guinea, the sap from the top shoot is sucked as a poison antidote. Young shoots and leaves are edible.

**Observations** A climber up to 20 m long with sparsely prickly stem and branches; leaves broadly ovate to ovate-oblong, up to 32 cm long, glaucous beneath, petiole up to 4 cm long, wings of petiolar sheaths distinct, tendrils present; inflorescence with 2–6 umbels; fruit about 10 mm in diameter, black at maturity. *S. leucophylla* occurs in mixed forest and teak forest, in Java up to 1100 m altitude. It has been much confused with *S. blumei*, which can be distinguished by its non-glaucous leaf undersurface and thinner leaves.

**Selected sources** 97, 190, 202, 278, 580, 601, 608, 779, 1178, 1227, 1356, 1491.

### ***Smilax luzonensis* K. Presl**

Reliq. haenk. 1: 131 (1827).

**Synonyms** *Smilax helferi* A.DC. (1878).

**Vernacular names** Malaysia: akar banar, akar gadung tikus, banar babi (Peninsular). Cambodia: voë bâng-he:t khmô:t (Ratanakiri). Thailand: khueang (eastern), yaan thaat, faa laep (peninsular). Vietnam: d[aa]y kim cang.

**Distribution** India, Burma (Myanmar), Cambodia, Laos, Vietnam, Thailand, Peninsular Malaysia, Sumatra, Java and the Philippines.

**Uses** Pounded rhizomes are used for poulticing boils in Malaysia, often together with leaves of *Macaranga triloba* (Blume) Muell.-Arg.

**Observations** A climber up to 5 m long with smooth or sparsely prickly stem and branches; leaves broadly elliptical to elliptical, sometimes lanceolate-elliptical, up to 15 cm long, petiole up to 2 cm long, wings of petiolar sheaths weakly developed, tendrils up to 12 cm long; inflorescence with 1–3 umbels; fruit 5–6 mm in diameter, dirty yellow at maturity; seeds dark red. *S. luzonensis* is locally frequent in evergreen forest up to 1000 m altitude, in Malesia also in bushes in open country.

**Selected sources** 202, 779, 780, 1227.

### ***Smilax megacarpa* A.DC. & C.DC.**

Monogr. phan. 1: 186 (1878).

**Vernacular names** Malaysia: akar banar, akar rebanar, akar lampu bukit (Peninsular). Laos: kh'u:a hmu: 'wa:k, kê:ng hmu: (Louang Prabang), khu:ng th'ô:n (Vientiane). Vietnam: cam [is]ch (Quang Nam-Da Nang), kim cang (Sông Bé), d[aa]y man [ees]t (Quang Nam-Da Nang).

**Distribution** India, Burma (Myanmar), Cambodia, Laos, Vietnam, Hainan, Thailand, Peninsu-

lar Malaysia and Singapore; possibly also Java, Borneo and the Moluccas.

**Uses** *S. megacarpa* is used in local medicine in Laos; it plays a role in medicines given after childbirth and against cough. Rhizomes are eaten in Peninsular Malaysia.

**Observations** A climber up to 6 m long with smooth stem and branches; leaves broadly elliptical to broadly ovate or ovate, up to 27 cm long, petiole up to 5 cm long, wings of petiolar sheaths rather weakly developed, tendrils up to 25 cm long; inflorescence with 2–3 umbels; fruit 15–20 mm in diameter, dark red or purple at maturity. *S. megacarpa* occurs locally frequent in evergreen forest up to 1200 m altitude, in Malesia also in bushes in open country.

**Selected sources** 202, 779, 780, 1227, 1356.

### **Smilax myosotiflora A.DC.**

Monogr. phan. 1: 65 (1878).

**Vernacular names** Malaysia: akar ali, akar tanding, akar dedingin (Peninsular).

**Distribution** Peninsular Thailand, Peninsular Malaysia and Singapore.

**Uses** Rhizomes are used in Malaysia as an aphrodisiac and leaves and fruits to cure syphilis.

**Observations** A herbaceous climber with slender smooth stem and branches; leaves broadly elliptical to lanceolate-elliptical, up to 15 cm long, petiole up to 2.5 cm long, wings of petiolar sheaths weakly developed, tendrils up to 13 cm long; umbels solitary; fruit 7–9 mm in diameter. *S. myosotiflora* occurs in evergreen forest, often climbing on bushes and low trees, up to 1400 m altitude.

**Selected sources** 202, 779, 780, 1227.

### **Smilax verticalis Gagnepain**

Bull. Soc. Bot. Fr. 81: 74 (1934).

**Synonyms** *Smilax simulans* T. Koyama (1967).

**Vernacular names** Cambodia: voë mē:m thnam chön (Koh Kong), voë pāpāhs (Pursat). Laos: kh'a:ng lua:yx, khua:ng th'ō:n (Vientiane), khua:ng no:yz (Savannakhet). Thailand: khrua daao (northern). Vietnam: kim chang d[uws]ng.

**Distribution** Cambodia, Laos, Vietnam and Thailand.

**Uses** Rhizomes and leaves are used in local medicine in Cambodia and Laos.

**Observations** A sub-erect to straggling slender shrub up to 2 m tall, somewhat tufted at base with smooth or sparsely prickly stem and branches; leaves elliptical to ovate-elliptical, ovate or lanceolate-oblong, up to 15(–18) cm long, petiole up to 2 cm long, wings of petiolar sheaths not well devel-

oped, tendrils absent or short; inflorescence with 1–2 umbels; fruit 8 mm in diameter, reddish at maturity. *S. verticalis* occurs in shrub savanna and open forest up to 1000 m altitude.

**Selected sources** 780.

### **Smilax zeylanica L.**

Sp. pl. 2: 1029 (1753).

**Vernacular names** Indonesia: kayu cina utan, saihe maruani, asaihe tuni (Moluccas).

**Distribution** Sri Lanka, India, Burma (Myanmar), northern Thailand and Java, possibly also Sulawesi, the Moluccas and New Guinea.

**Uses** *S. zeylanica* is used as a medicinal plant in India, and possibly as a substitute for *S. china* in the Moluccas.

**Observations** A climber up to 6 m long with smooth or sparingly prickly stem and branches; leaves elliptical to ovate, oblong or lanceolate, sometimes ovate-orbicular, up to 24 cm long, petiole up to 3 cm long, wings of petiolar sheaths rather weakly developed, tendrils up to 20 cm long; inflorescence with 1–5 umbels; fruit 6–10 mm in diameter, dirty yellow to shiny black at maturity; seeds brown. Two subspecies are distinguished: subsp. *hemsleyana* (Craib) T. Koyama (synonym: *S. hemsleyana* Craib) and subsp. *zeylanica*. The former differs from the latter in its solitary umbels and comparatively broader leaves, and is only known from eastern India and northern Thailand. *S. zeylanica* occurs in primary and secondary forest, teak forest, brushwood and bamboo thickets, in Java up to 1600 m altitude. *S. celebica* Blume, a decoction of whose leaves has been reported to be used in the Moluccas as tonic after childbirth, might be merely a form of *S. zeylanica* differing only in the leaves.

**Selected sources** 97, 140, 580, 779, 780, 1491.

Stephen P. Teo

### **Solanum L.**

Sp. pl. 1: 184 (1753); Gen. pl. ed. 5: 85 (1754).

SOLANACEAE

$x = 12$ ; *S. capsicoides*:  $2n = 24$ , *S. erianthum*:  $2n = 24$ , *S. mammosum*:  $n = 11$ ,  $12$ , *S. nigrum*:  $2n = 72$ , *S. trilobatum*:  $2n = 24$

**Major species** *Solanum erianthum* D. Don, *S. nigrum* L.

**Vernacular names** Solanum, nightshade (En). Solanum, morelle (Fr).

**Origin and geographic distribution** Estimates differ greatly, but *Solanum* probably com-

prises some 1000–1100 species. It is cosmopolitan, except in boreal, alpine and aquatic habitats. The principal centre of diversity is located in Central and South America, with secondary centres in Africa and Australia. With a total of 60 wild and cultivated species, New Guinea is probably the area in Malesia with the most species; by comparison, Peninsular Malaysia has about 15 species and Java and the Philippines about 25 species each.

**Uses** *Solanum* is used to cure digestive and intestinal problems, including stomach-ache, diarrhoea, piles and dysentery, and for various skin problems such as sores, boils, cuts, wounds and bruises. Many species are also employed to treat fever and malaria, headache and rheumatism. Some are considered to be stimulants whereas others have sedative properties. Furthermore, *Solanum* is frequently used for various diseases of the respiratory tract, such as coughs, sore throat, bronchitis and asthma. Finally, many species are applied to treat urinary problems.

*Solanum* shows insecticidal and fungicidal properties. The leaves and stems of many species are often cooked or steamed and eaten as a vegetable. The unripe fruits are eaten in curries, whereas the ripe ones of some *Solanum* species are edible either cooked or raw. Caution must be taken when eating *Solanum*, as several species are poisonous.

**Production and international trade** The fruits of *S. sanitwongsei* are on sale in Bangkok markets, and have been cultivated commercially in Manila.

**Properties** The steroidal alkaloids of the *Solanaceae* occur as glycosides (hence glycoalkaloids). The known steroidal alkaloids are based on a C<sub>27</sub> cholestane skeleton and can be divided into 5 groups: solanidanes, spirosolanes, 22,26-epimincholestane,  $\alpha$ -epiminocyclohemiketal and 3-aminospirostane. The first 2 groups have attracted the most research. The solanidanes, with important alkaloids such as solanidine, leptidine and demissidine, are characterized by an indolizidine moiety. Important spirosolane alkaloids include solasodine, tomatidine and tomatidenol. The spirosolanes are structurally similar to saponins of the diosgenin type, except that the oxygen in the spiro-alketal pattern has been replaced by nitrogen.

Solanidine is present in *S. capsicoides*, whereas solasodine is found in *S. erianthum*, *S. nigrum* and *S. trilobatum*. The total alkaloid content of air-dried leaves and fruits is respectively 0.26% and 0.14% for *S. capsicoides*, 0.37% and 0.39% for

*S. erianthum*, 0.43% and 0.10% for *S. nigrum* and 0.36% and 0.96% for *S. trilobatum*. The solasodine content in *Solanum* fruits from Indian samples is 0.01–0.70% in *S. erianthum* and 0–0.28% in *S. nigrum*, whereas the total glycoalkaloid content in fruits of *S. trilobatum* has been found to be 3.5%. Leaf samples of *S. erianthum* from Vietnam contained 0.26% solasodine, 0.05% tomatidine and 0.01% solaverbascine. *S. erianthum* also has steroidal saponins and free genins. As well as solasodine, *S. nigrum* contains the sapogenins diosgenin and tigogenin. The unripe berries have 0.68% solasodine, 0.19% diosgenin and 0.15% tigogenin, whereas leaves contain 1.28% tigogenin. Glycoalkaloids are toxic to animals when injected. Like saponins, they are surface-active and haemolytic, and possess antifungal and cytostatic properties. *Solanum* steroidal glycoalkaloids affect the body mainly in two ways: the intact glycoalkaloid is an irritant, whereas the steroidal alkaline affects the nervous system. The pharmacological effects of these compounds may possibly be attributable to the ability of the steroidal glycoalkaloids to impair the functioning of membranes of strategic muscle and nerve cells in mammals. Evidence to support this is the toxicity of alkaloids to plants and fungi where complexation has been demonstrated with membrane sterols; it has been shown that the alkaloid sensitivity in *Pythium* and *Phytophthora* increases when these fungi are grown on a medium containing sterol and incorporate the sterol into their membranes. However, aglycones also have a still unexplained deleterious effect on certain tissues and organisms, and other mechanisms may be involved in the toxicity of *Solanum* alkaloids. Furthermore, the toxicity of the alkaloids is highly pH-dependent: lower pH levels greatly reduce toxicity.

The physiological and pathological effects of steroidal glycoalkaloids on mammals are numerous. Solanine has been observed to depress the central nervous system in rabbits; solanine, chacoine and tomatine have been observed to induce tachycardia in rabbits and rats; chacoine, tomatine and solanine have been observed to induce tachypnea or bradypnea in rabbits. Other reported effects are hypotension induced by tomatine in rats and rabbits, positive inotropic (cardiotonic) action by tomatine, chacoine, solanine, demissidine, commersonine and solanidine in frogs, hyperglycaemia induced by solanine in rats, haemolysis and haemorrhage induced by tomatine in rats, inhibition of plasma cholinesterase by solanine, solanidine, tomatine and demissidine in humans



and embryotoxicity induced by solanine in rats and mice.

The powdered aerial parts of *S. nigrum* and its methanolic extract significantly reduced gastric ulcer formation in rats. The activity may be due to inhibition of acid and pepsin secretions and/or their in vitro ability to bind these; inhibition of the acid production alone by cimetidine did not decrease ulcer formation. 'Sobatum' is the partially purified component of *S. trilobatum* obtained from the 75:25 petroleum ether/ethyl acetate extract. It has been found to be cytotoxic in Dalton's Lymphoma ascites, Ehrlich ascites cell lines and tissue culture cells (L929 and Vero). 'Sobatum' significantly inhibited peritoneal tumours induced by Dalton's Lymphoma ascites and Ehrlich ascites tumour cells. It was also found to reduce solid tumour growth in mice, when given either simultaneously or prophylactically, and is more active in simultaneous administration in Ehrlich ascites cell lines. It was found that 'Sobatum' was more active against Ehrlich ascites-induced solid tumour than Dalton's Lymphoma ascites-induced solid tumours. It has been experimentally proven that 'Sobatum' has the ability to retard the development of solid tumours and 7,12-dimethylbenz(a)anthracene-induced carcinogenesis. In another experiment with mice, 'Sobatum' was administered intraperitoneally. It failed to influence the induction of micronuclei in bone marrow erythrocytes of mice 24 hours and 72 hours after the second administration, thereby demonstrating that 'Sobatum' has no cytogenic toxic potential. The rare sterol carpesterol isolated from *S. trilobatum* (also found in *S. torvum* Swartz) has anti-inflammatory activity on carrageenin-induced mouse paw oedema; it proved as effective as hydrocortisone and withaferin A (from *Withania somnifera* (L.) Dunal). The alcoholic extract of *S. nigrum* berries (100–400 mg/kg) showed significant inhibition of carrageenin-induced oedema in albino rats. The aqueous leaf extract of *S. erianthum* did not produce any significant suppression of *Plasmodium berghei* infection in mice.

A large number of fungi are inhibited in growth and development by steroidal alkaloids such as tomatine, solanine, and chaconine. An alcoholic extract of leaves of *S. nigrum* is active against *Staphylococcus aureus* and *Escherichia coli*. Leaf extracts of *S. nigrum* inhibited lesion production in leaves in response to the tobacco mosaic virus. The flavonoid-rich extract of *S. erianthum* possesses antibacterial and antifungal activity. Gram-positive bacteria are inhibited, but gram-

negative ones are not, whereas the flavonoids have been found to be toxic to the fungi *Aspergillus flavus* and *Candida albicans*. Some other pharmacological activities of *Solanum* include antispasmodic, hypotensive, hypocholesterolaemic and anti-HIV-1 activity induced by *S. nigrum* in mammals, and insecticidal and molluscicidal activity of *S. nigrum* and of *S. mammosum* L.

*Solanum* steroidal alkaloids are useful in industry as steroid precursors. Solasodine is a nitrogen analogue of diosgenin, a compound often used as raw material for the production of medicinal steroids. The synthetic steroids have three main applications in medicine: as anti-inflammatory corticosteroids, as contraceptive sex steroids and as anabolic steroids.

**Adulterations and substitutes** Steroidal alkaloids (e.g. diosgenin and tigogenin) are also found in *Dioscorea* and *Smilax* species; these are also used as starting material for steroid hormone semisynthesis.

**Description** Annual or perennial, erect or ascending, unarmed or spiny herbs, shrubs or rarely small trees, with simple, branched, stellate or glandular hairs. Leaves alternate or rarely subopposite, simple and entire to lobed, pinnatisect or imparipinnate, petiolate, exstipulate. Inflorescence a terminal cyme but usually appearing lateral by growth of a lateral bud and extra-axillary cyme, appearing racemose, umbellate or paniculate or rarely reduced to a solitary flower. Flowers regular, bisexual or rarely andromonoecious; calyx campanulate, rotate or cupular, (4–)5(–10)-lobed; corolla stellate, rotate or campanulate, (4–)5(–10)-lobed, white, violet, purple or blue; stamens (4–)5, inserted on the corolla throat, alternating with the corolla lobes, anthers often connivent, opening by terminal pores or slits; ovary superior, 2(–4)-locular with many ovules in each cell, style simple, stigma capitate or bifid. Fruit a usually globose berry, with a persistent and sometimes enlarged calyx, few to many-seeded. Seeds orbicular to subreniform, compressed, often minutely pitted or reticulate. Seedling with epigeal germination; cotyledons emergent, ovate to linear-lanceolate; first leaves usually entire.

**Growth and development** Most *Solanum* species flower and fruit almost throughout the year. Flowers are pollinated by insects. Fruit maturation takes about 2–3 months. The fruits are eaten by birds and mammals which disperse the seeds.

**Other botanical information** *Solanum* has been subdivided into 7 subgenera and numerous

sections and series. *S. nigrum*, being the type-species of the genus, belongs to subgenus *Solanum*, *S. erianthum* to subgenus *Brevantherum* and the other four species treated below to subgenus *Leptostemon* which also includes *S. melongena* L. and *S. mammosum* L., species well-known for their fruits. It is in South-East Asia that the taxonomy of *Solanum* is least known: a thorough taxonomic revision is urgently needed. Adding to the taxonomic confusion is the fact that *S. erianthum* has been extensively referred to as *S. verbascifolium* L., which actually proved to be identical with a South American species. Moreover, in South-East Asian literature, *S. capsicoides* has often been misinterpreted as *S. aculeatissimum* Jacq. Furthermore, *S. nigrum* belongs to a complex of very similar species (including e.g. *S. americanum* Miller and *S. villosum* Miller), which are sometimes regarded as mere forms of a single variable species. *S. nigrum* arose from hybridization of the diploid *S. americanum* and the tetraploid *S. villosum*. *S. villosum* is distinguished by its yellow, orange or red berries. *S. americanum* and *S. nigrum* are much more difficult to distinguish. The former has usually umbellate inflorescences, anthers 1.0–2.0 mm long, a glossy, purplish-black berry with generally 50–80 seeds and an enlarged calyx with reflexed lobes in fruit. For the latter, see below. *S. mammosum*, originally from Central and South America but introduced in the Malesian region for its decorative fruits, has poisonous fruits which are occasionally used as an insecticide against cockroaches and caterpillars. *S. procumbens* Lour., *S. spirale* Roxb. and *S. surattense* Burm.f. (synonym: *S. xanthocarpum* Schrad. & J.C. Wendl.) are used in folk medicine in Indo-China. *S. aviculare* J.G. Forster, *S. laciniatum* Aiton and *S. khasianum* C.B. Clarke are a reputed rich source of solasodine. Originating from outside the Malesian region, they have received special attention in breeding programmes for cultivation at higher elevation in Java. In particular *S. khasianum* merits further research.

**Ecology** Most medicinal species of *Solanum* are weeds of gardens, fields and waste places, occurring in sunny or slightly shaded sites at low to medium altitudes, but *S. nigrum* is found up to 3100 m.

**Propagation and planting** Those *Solanum* cultivated as vegetables in South-East Asia can be propagated by seed, shoot cuttings and by division of rooted shoots.

**In vitro production of active compounds**

Diosgenin and solasodine have been isolated from 6-month-old callus of *S. erianthum*; the undifferentiated callus tissue was established from sterilized seeds on Murashige and Skoog's revised medium. Blue light stimulated solasodine synthesis and green light stimulated diosgenin synthesis in the callus. Optimal growth was reached after 6 weeks when the dry weight of the tissue had increased 6.6-fold. After 6 weeks only about 147 µg diosgenin and 47 µg solasodine had been produced per g of dry-weight tissue; this is very little.

**Diseases and pests** In West Java *S. nigrum* grown as a leaf vegetable is fairly free from diseases and pests.

**Harvesting** Leaves, stems, berries and roots of *Solanum* are harvested for medicinal use. When used as a vegetable, leaves and young shoots of *S. nigrum* are hand-picked.

**Yield** In many species the steroidal alkaloid and sapogenin content decline as the fruit ripens. Leaf alkaloid and sapogenin content also decline with age. In India a method has been developed to estimate solasodine content in *S. nigrum* leaves 3 months after sowing by estimating their N content. At this stage of growth a top dressing or foliar sprays may be applied to increase solasodine yield.

**Handling after harvest** In general, *Solanum* fruits are used fresh. The other harvested parts are dried and can be stored in sealed containers, preferably in a cool well-aerated room.

**Genetic resources and breeding** *S. capsicoides* is resistant to *Pseudomonas* and is a non-host for potato cyst nematodes. There are no records of medicinally important *Solanum* in germplasm collections. In view of their weedy nature the risk of genetic erosion seems to be rather limited. The great variation in alkaloid content within species may offer possibilities for selection. However, the alkaloid content also varies substantially as a result of ecological conditions, drying and storage.

**Prospects** Several *Solanum* species containing the spirostanol alkaloids solasodine and tomatidine are considered promising as alternatives to *Dioscorea* species as a source of raw material for steroid production, including 16 dehydropregnelone acetate. Hormonal derivatives of this steroid are used as active ingredients of the oral contraceptive pill. The ability of *S. trilobatum* components to retard the development of solid tumours and to act as anti-inflammatory deserves further research.

**Literature** |1 Akhtar, M.S. & Munir, M., 1989.

Evaluation of the gastric antiulcerogenic effect of *Solanum nigrum*, *Brassica oleracea* and *Ocimum basilicum* in rats. *Journal of Ethnopharmacology* 27(1-2): 163-176. |2| Bhattacharya, T.K., Ghosh, M.N. & Subramanian, S.S., 1980. A note on anti-inflammatory activity of carpesterol. *Fitoterapia* 51(5): 265-268. |3| Blomqvist, M.M., 1997. Taxonomy and uses of medicinally important species in the genera *Datura* L. and *Solanum* L. (Solanaceae) in South-East Asia. Unpublished MSc. thesis, Department of Plant Taxonomy, Wageningen Agricultural University, the Netherlands. 132 pp. |4| Council of Scientific and Industrial Research, 1972. The wealth of India: a dictionary of Indian raw materials and industrial products. Vol. 9. Publications and Information Directorate, New Delhi, India. pp. 378-429. |5| Mohanan, P.V. & Devi, K.S., 1997. Effect of *Sobatum* on tumour development and chemically induced carcinogenesis. *Cancer Letter* 112(2): 219-223. |6| Mohanan, P.V., Rathinam, K. & Devi, K.S., 1996. Lack of micronucleus induction by '*Sobatum*' in bone marrow erythrocytes of Swiss mice. *Mutation Research* 361(1): 23-27. |7| Perry, L.M., 1980. Medicinal plants of East and Southeast Asia. Attributed properties and uses. The MIT Press, Cambridge, Massachusetts, United States & London, United Kingdom. pp. 394-396. |8| Roddick, J.G., 1986. Steroidal alkaloids of the Solanaceae. In: d'Arcy, W.G. (Editor): *Solanaceae: biology and systematics*. Columbia University Press, New York, United States. pp. 201-222. |9| Roddick, J.G., 1991. The importance of the Solanaceae in medicine and drug therapy. In: Hawkes, J.G., Lester, R.N., Nee, M. & Estrada, N. (Editors): *Solanaceae III: taxonomy, chemistry, evolution*. Royal Botanic Gardens Kew and Linnean Society of London, United Kingdom. pp. 7-23. |10| Siemonsma, J.S. & Kasem Piluek (Editors), 1993. *Plant Resources of South-East Asia No 8. Vegetables*. Pudoc Scientific Publishers, Wageningen, the Netherlands. pp. 249-260.

#### *Selection of species*

#### ***Solanum capsicoides* All.**

Auct. synop. meth. stirp. hort. reg. Taurenensis: 12 (1773).

**Synonyms** *Solanum ciliatum* Lamk (1794), *Solanum ciliare* Willd. (1809), *Solanum aculeatis-simum* auct. non Jacq.

**Vernacular names** Indonesia: terong kori, terong tenang (Sundanese). Malaysia: terong

asam hutan, terong perat, terong puyoh (Peninsular). Thailand: khuea hin (peninsular), ma khuea kham (northern), ma khuea khuen (Chiang Mai).

**Distribution** Originally from coastal Brazil, but now commonly naturalized in western Africa, the Caribbean, Central America, Mexico, Florida, Hawaii, the Ryukyu Islands, Taiwan, China, India, Sri Lanka and in South-East Asia in Thailand, Peninsular Malaysia, Sumatra and Java; to be expected elsewhere.

**Uses** The fruits, charred and pounded in oil, are used to treat skin complaints. In Peninsular Malaysia, the pounded roots have been applied to the gums against toothache. The smoke of dried, pounded and burned seeds has been inhaled to cure an ulcerated nose. The fruits are edible when roasted or cooked in curry. *S. capsicoides* is sometimes cultivated as an ornamental for its decorative fruits.

**Observations** A perennial, slightly woody, prickly herb up to 100 cm tall, stem and leaves glabrous or sparsely pilose with simple hairs; leaves broadly ovate to ovate-cordate, pinnately 5-7-lobed, 4.5-18 cm × 4-12 cm, cordate at base, apex acute; inflorescence extra-axillary, racemose, 1-7-flowered, on a short peduncle; calyx campanulate, 4-5 mm long, shortly glandular hairy, corolla rotate, about 3 cm in diameter, white, lobes lanceolate, anthers narrowed towards the apex, about 6 mm long, with apical pores; fruit solitary or sometimes 2-3 together, globose to depressed globose, 2.5-3 cm in diameter, glabrous, orange-red when mature, calyx slightly enlarged; seeds strongly flattened, winged all around, 4-6 mm in diameter. *S. capsicoides* is a common weed of slightly shaded places, along hedges, sugar-cane fields and roadsides up to 1500 m altitude.

**Selected sources** 97, 202, 287, 320, 580, 873, 1016, 1017, 1380, 1652.

#### ***Solanum erianthum* D. Don**

Prodr. fl. nepal. 96 (1825).

**Synonyms** *Solanum mauritianum* Blanco (1837) non Scop., *Solanum verbascifolium* auct. non L.

**Vernacular names** Potato tree, tobacco tree, tropillo (En). Indonesia: daun salawar, tembako utan (Malay, Moluccas), teter (Javanese, Sundanese). Malaysia: daun telinga kerbau, terong belah, terong raya (Peninsular). Papua New Guinea: kumboomba (Lesu, New Ireland), epiap (Gunantuna, New Britain). Philippines: malatalong (Tagalog), liangkag (Bukidnon), ungali (Bisaya). Laos: sang mong peng, sang mou. Thai-

land: dap yaang (central), faa paeng (northern), khaa taai (peninsular). Vietnam: c[af] h[oo]i, c[aa]y la, la r[uw]ng.

**Distribution** Originally from the West Indies, Central America and Mexico, but now an almost pantropical weed, although hardly penetrating South America. Probably introduced into the Philippines by the Spanish in the 16th Century, from where it has spread throughout the Malesian archipelago and to mainland Asia and Australia.

**Uses** The leaves act as an abortifacient and are considered a potent medicine for expelling all impurities through the urine, and in particular to treat leucorrhoea. Pounded leaves are poulticed to treat piles, haemorrhoids and scrofula. Heated leaves are applied as a cream to the forehead against headache. A decoction of the leaves is drunk against vertigo; an infusion of the plant is used for a bath after childbirth. A decoction from the roots is applied to treat violent pains all over the body or to relieve digestive troubles; it is also given to treat dysentery, diarrhoea and fever. In



*Solanum erianthum* D. Don - 1, flowering stem; 2, flower; 3, infructescence.

Papua New Guinea, the plant is used internally to treat stomach-ache and is applied externally to skin irritations and rashes. In the Solomon Islands, leaf juice is used as a rinse for sores in the mouth. *S. erianthum* is considered poisonous to livestock. The root bark is poisonous and can be used as an antiphlogistic and against arthritis. The fruits can be eaten when cooked.

In the Philippines, the velvety leaves are used to remove grease from dishes. *S. erianthum* is considered suitable as a shade plant for coffee.

**Observations** An unarmed shrub or small tree up to 4(-10) m tall with a dense indumentum of soft stellate hairs, stem up to 20 cm in diameter; leaves simple, ovate-elliptical, (7-)10-20(-29) cm x 3.5-15 cm, margin entire or slightly wavy, base rounded to cuneate, apex acute to acuminate; inflorescence appearing terminal, a compound cyme; calyx campanulate, 5 mm long, lobes ovate, corolla stellate, about 1.5 cm in diameter, white, anthers oblong, about 2 mm long, opening with apical pores, ovary densely pubescent, style 4-6 mm long, glabrous; fruit globose, 8-12 mm in diameter, pubescent, dull yellow when ripe; seeds many, compressed, 1-2 mm in diameter. *S. erianthum* is fairly common, occurring scattered in sunny or slightly shaded places, in brushwood, roadsides, field edges, on waste ground and in forest edges, up to 1500 m altitude.

**Selected sources** 13, 78, 97, 127, 164, 202, 287, 580, 665, 666, 873, 889, 1035, 1126, 1178, 1250, 1251, 1380, 1433, 1525, 1652.

### *Solanum nigrum* L.

Sp. pl. 1: 186 (1753).

**Synonyms** *Solanum schultesii* Opiz (1843).

**Vernacular names** Black nightshade, common nightshade, garden nightshade (En). Herbe à calalou, morelle noire (Fr). Indonesia: ranti (Javanese), leunca (Sundanese), bobose (Ternate). Malaysia: ranti, terong meranti, terong parachit (Peninsular). Papua New Guinea: takuta (Wapenamanda, West Sepik). Philippines: konti (Filipino), anti (Bontok, Tagalog), kuti (Bikol). Thailand: ma waeng nok (southern), ya tom tok (northern), kha om (Prachuap Khiri Khan). Vietnam: c[af] n[us]t [as]o, gia c[aa]f[u], lu lu d[uw]j[c].

**Distribution** Native to Europe and western Asia, introduced in North America, Africa, Asia and Australia; probably fairly widely distributed throughout the Malesian region.

**Uses** The medicinal use of *S. nigrum* goes back more than 2000 years. The plants are used as an emollient and antalgic in itching, burns and neu-

ralgic pains, and are also considered expectorant and laxative. The leaves are said to have sedative and healing properties and are applied to cuts and ulcers. A decoction of the leaves is used to treat yaws. The fruit is considered to be a cure for diabetes. In Papua New Guinea, cooked leaves and stems are given to infants suffering from diarrhoea. In China, cooked young shoots are considered to be corrective and cooling, to increase the virility of men and to benefit menstrual disorders. A decoction of the leaves or seeds is used to treat wounds, cancerous sores and as an astringent. Diuretic properties are also attributed to the plant. The leaf juice is used against pain caused by an inflammation in the kidneys and bladder and by virulent gonorrhoea. In India, the leaves are used to treat inflammations on any part of the body, rheumatic and gouty joints and skin diseases. An extract of the leaves and stem is considered useful to treat dropsy, heart diseases, piles, gonorrhoea,

fevers, eye diseases and the chronic enlargement of liver and spleen. Young shoots are eaten as a vegetable either raw or cooked. The ripe fruits have been used for jam-making and in pies, but caution should be taken as unripe fruits are certainly poisonous and the fruits of European *S. nigrum* and possibly of other *Solanum* species are thought to be poisonous at all stages of their development.

**Observations** An annual or perennial unarmed herb up to 1 m tall, indumentum of simple eglandular and glandular hairs; leaves ovate, 4–10 cm × 2–7 cm, simple, margin entire to bluntly toothed, base cuneate, apex obtuse; inflorescence extra-axillary, a short raceme of (3–)4–8(–12) flowers; calyx campanulate, lobes up to 1 mm long, corolla stellate, 8–10 mm in diameter, white or rarely tinged with purple, anthers oblong, 2–3.5 mm long, ovary glabrous, style 5–6 mm long, stigma capitate; fruit globose to ellipsoid, 6–8(–10) mm in diameter, 2-locular, dull or somewhat shiny black or purplish-black, calyx not enlarged and with appressed or scarcely reflexed lobes; seeds (15–)25–35(–45) per fruit, 1.8–2.2 mm long. *S. nigrum* is common in open and disturbed places, in full sunshine or slight shade, also in light forest, up to 3100 m altitude.

**Selected sources** 21, 42, 78, 97, 164, 190, 202, 214, 287, 349, 375, 566, 749, 874, 999, 1035, 1087, 1178, 1235, 1244, 1245, 1260, 1297, 1356, 1380, 1432, 1652.

### *Solanum sanitwongsei* Craib

Kew Bull.: 246 (1928).

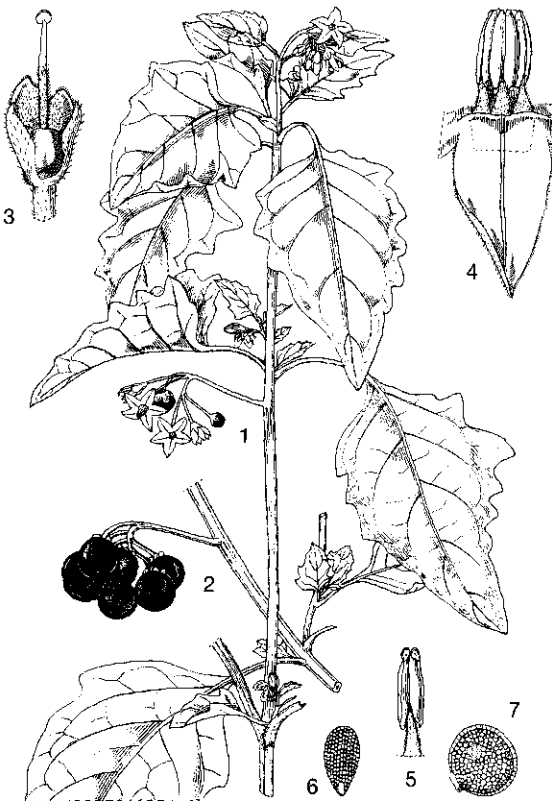
**Vernacular names** Philippines: talong-siam (Filipino). Thailand: ma waeng khrua (central).

**Distribution** Thailand; cultivated in the Philippines.

**Uses** The fruits are effective against diabetes and even seem to be capable of curing it. They are also expectorant and diuretic.

**Observations** A small, stellately hairy shrub up to 1 m tall, leaves oblong-ovate, 5–9 cm × 3.5–8 cm, with 2–3 lobes on either side, base cordate to cuneate, apex obtuse; inflorescence an extra-axillary, short raceme on a short peduncle; sepals 5, connate up to halfway, corolla stellate, about 2 cm in diameter, purple, anthers 6 mm long, ovary glabrous except for the apex, stigma subcapitate; fruit subglobose, 1 cm in diameter, glabrous, orange when mature; seeds abundantly, minutely dotted, about 2 mm in diameter.

**Selected sources** 290, 1126, 1178, 1287.



*Solanum nigrum* L. - 1, flowering stem; 2, part of fruiting stem; 3, opened calyx and gynoecium; 4, petal and stamens; 5, stamen; 6, seed from below; 7, seed in side view.

**Solanum sarmentosum** Nees

Trans. Linn. Soc. 17: 58 (1837).

**Synonyms** *Solanum maingayi* O.Kuntze (1891).**Vernacular names** Malaysia: terong pipit, terong puyoh, terong tikus (Peninsular).**Distribution** Peninsular Malaysia and Singapore.**Uses** The leaf juice is drunk as a remedy for fever. The seeds are used as a vermifuge.**Observations** An armed, straggling shrub; leaves elliptical, 5–10 cm × 2–5 cm, lobed, almost glabrous above, sparsely hairy below; inflorescence composed of 1–2 flowers on a peduncle of about 2.5 cm long; calyx funnel-shaped, thorny, corolla about 2 cm in diameter, blue; fruit about 12 mm in diameter, glabrous, orange when mature, seated on an enlarged calyx. *S. sarmentosum* occurs as a weed in gardens and ruderal places.**Selected sources** 202, 1126, 1227.**Solanum trilobatum** L.

Sp. pl. 1: 188 (1753).

**Synonyms** *Solanum acetosaefolium* Lamk (1794), *Solanum canaranum* Miq. ex C.B. Clarke (1883).**Vernacular names** Thailand: khwaeng khia (Tak), ma waeng khrua (central). Vietnam: c[af]ba th[uf]y.**Distribution** India, Vietnam, Thailand and Peninsular Malaysia.**Uses** In India, the bitter roots and young shoots have been given in the form of an electuary, a decoction or a powder for consumption. The medicine is mainly used for asthma, chronic febrile affections and difficult parturition. The fruit is edible, and the leaves are eaten cooked as a vegetable.**Observations** A nearly glabrous, thorny herb, slightly woody at base; leaves broadly elliptical to broadly ovate, 4–7.5 cm × 2.5–4 cm, sinuate-lobed, base rounded to slightly cordate, apex rounded; inflorescence extra-axillary, composed of a few-flowered cyme or a 6–10-flowered raceme; calyx campanulate, 2–3 mm long, glabrous, not enlarged in fruit, corolla stellate, 2.5–3.5 cm in diameter, blue, anthers 5, 7–8 mm long; fruit about 1 cm in diameter, purple to blackish; seeds about 3 mm in diameter. *S. trilobatum* is found as a weed in gardens and on waste ground. In India it is reported to grow in tidal swamps.**Selected sources** 78, 97, 148, 171, 202, 287, 962, 963, 1227, 1380.

M.M. Blomqvist &amp; Nguyen Tien Ban

**Sophora tomentosa** L.

Sp. pl. 1: 373 (1753).

LEGUMINOSAE

 $x = 9; 2n = 18$ **Synonyms** *Sophora heptaphylla* L. (1754), *Sophora crassifolia* J. St.-Hil. (1806), *Sophora havanensis* Jacq. (1860).**Vernacular names** Seacoast laburnum, silver bush (En). Indonesia: kayu penawar (Javanese), ki ucing (Sundanese), upas biji (Malay, Moluccas). Malaysia: pelochok, pelotok (Peninsular). Philippines: tambalisa (Filipino), mangguiau (Bikol, Tagalog), pangalangan (Tagalog, Bisaya). Cambodia: sândaæk préi srâmot. Thailand: saaraphat phit (Phetchaburi). Vietnam: (c[aa]y) chu[oox]i h[ooj]t, h[of]e l[oo]ng.**Origin and geographic distribution** *S. tomentosa* has a pantropical, coastal distribution. It is common throughout the South-East Asian region, occurring north to China and the Ryukyu Islands, south to eastern Australia and east into Polynesia.**Uses** The seed oil of *S. tomentosa* is reputed to be a good expectorant; in the Philippines it is applied externally to soothe painful bones. A decoction of the seeds and roots is given in bilious disorders. A decoction of the seeds, roots or bark has been used against cholera. The pounded seeds are used to cure colic and dysentery. Smaller doses are applied as an antidote in food poisoning and stings of poisonous fish, as well as to treat haemoptysis, painful urination and gonorrhoea. The root-bark is also used to treat such cases of poisoning. The Thai name 'saaraphat phit' means various poisons, probably referring to its use as an antidote. In East Africa *S. tomentosa* is used as a fish poison.The timber of *S. tomentosa* is hard and heavy and used for small objects. It is occasionally grown as an ornamental in gardens close to the sea.**Properties** The bitter seeds of *S. tomentosa* have astringent, febrifugal, stomachic and dangerous emetocathartic properties. The leaves are also strongly emetocathartic and toxic in large doses. *Sophora* species are relatively rich in alkaloids: these can be found in leaves, roots and seeds. Some of these alkaloids seem to be common, whereas others appear to be confined to a single or a limited number of species. Detailed phytochemical investigations have led to the isolation of (+)-matrine, (+)-matrine-N-oxide, (+)-sophocarpine-N-oxide, (-)-anagyryne, (-)-baptifoline, (-)-cytisine, (-)-N-methylcytisine, (-)-formylcytisine, (-)-

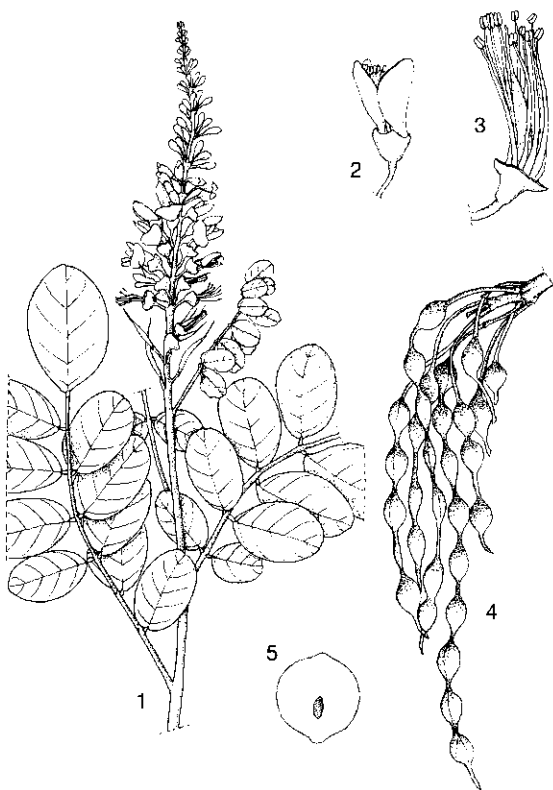
N-acetylcytisine, (+)-ammodendrine, (-)-epiamprolobine, (+)-epiamprolobine-N-oxide and 5-(3'-methoxycarbonylbutyroyl)aminomethyl-trans-quinolizidine from the various (fresh) parts of *S. tomentosa*. All compounds belong to the quinolizidine group of alkaloids (sometimes also called lupin alkaloids), which are biosynthesized from the amino acid lysine. The amounts of this amino acid in the leaves, stems, immature pods and immature seeds are respectively 0.15%, 0.22%, 0.37% and 0.64% of the fresh weight, with (+)-epiamprolobine-N-oxide, (+)-matrine-N-oxide, (+)-matrine and (-)-cytisine being the most abundant alkaloids. Together with (-)-N-methylcytisine, the latter three compounds are also found in *S. flavescens* Aiton from China, a species commonly traded in Chinese pharmacies throughout Malaysia.

The pharmacological actions of several of the alkaloids have been quite well investigated. Dried seed of *S. tomentosa* contains up to 2% (-)-cytisine (also known as sophorine or ulexine), which resembles the alkaloid nicotine in its action as a neurotransmitter, but is much less toxic as a poison. It also has insecticidal properties. The effects of (-)-N-methylcytisine and (+)-matrine isolated from *S. flavescens* on the motility of parasitic helminths and mouse ileum preparations have been experimentally tested. The two alkaloids acted antagonistically on all the preparations: (-)-N-methylcytisine acted spastically and (+)-matrine paralytically. From the results on interactions between these alkaloids and known neuropharmacological agents, it is suggested that the effects of both alkaloids are elicited through a neuropharmacological mechanism in parasitic helminths and host tissue. (+)-Matrine also has anti-inflammatory properties as well as anti-ulcer properties; this was demonstrated in an experiment in fundus strips of rats. (+)-Matrine isolated from the root of *S. subprostrata* Chun & Chen showed significant inhibition of ocular inflammation induced by lens proteins. Unlike corticosteroids, (+)-matrine did not facilitate the intra-ocular pressure recovery in rabbit eyes nor did it change the electrical potential difference across rabbit iris-ciliary body. These results indicate that (+)-matrine could become a safer ocular anti-inflammatory agent than corticosteroids. Furthermore, (+)-matrine was found to markedly increase the reaction time of a mouse placed on a hot plate; the results indicate that this component could be used as an analgesic as well. The effect of (+)-matrine, from *S. flavescens*, on glutamate-

induced responses was investigated using electrophysiological techniques at the crayfish neuromuscular junction. At concentrations greater than 0.1 mM, (+)-matrine depressed both glutamate-induced responses and neurally evoked excitatory junctional potentials. In this assay (+)-matrine-N-oxide (= oxymatrine) also shows activity, although far less powerfully than (+)-matrine. Finally, (+)-matrine-N-oxide (oxymatrine) has a biphasic effect on cultures of rat myocytes: slowing the beating rate by  $\alpha$ -adrenoreceptor stimulation (concentrations at 50  $\mu\text{mol/l}$ ) and accelerating it by  $\beta$ -adrenoreceptor stimulation (250  $\mu\text{mol/l}$  concentration).

Five isoprenylated flavonoids have been isolated from the aerial parts of *S. tomentosa*: sophoraisoflavanone A, sophoraflavanone B, sophoronol, isosophoranone and isobavachin. The structurally closely related flavonoid vexibiol, from *S. flavescens*, is known to exhibit anti-ulcer effects.

**Adulterations and substitutes** (-)-Cytisine (also known as sophorine or ulexine) as found in



*Sophora tomentosa* L. - 1, flowering twig; 2, flower; 3, flower with petals removed; 4, infructescence; 5, seed.

several *Sophora* species is also found in *Laburnum anagyroides* Medik. and *Ulex europaeus* L.

**Description** An evergreen shrub or small tree up to 7 m tall; bole branchless for up to 3 m, up to 45 cm in diameter; young twigs, leaf rachises, lower surface of leaflets, inflorescences, calyces and pods densely puberulous or tomentose. Leaves arranged spirally, imparipinnate, up to 30 cm long, with 9–23 leaflets, stipules absent; leaflets opposite, broadly elliptical to suborbicular, up to 4(–5) cm × 3 cm, apex rounded to slightly emarginate. Flowers in a terminal raceme up to 25 cm long, papilionaceous; bracteoles absent; pedicel 5–10 mm long; calyx campanulate, with 5 shallow teeth; petals yellow, standard 14–20 mm × 11–14 mm, wings and keel of equal length; stamens 10, free; ovary superior, densely appressed pubescent. Fruit a 3–10-seeded pod, 5–19 cm long, markedly constricted between the seeds. Seeds subglobose, 6–7 mm long, brownish. Seedling with hypogeal germination; cotyledons not emergent; hypocotyl elongated; epicotyl with a few scale leaves, followed by simple, then 2-foliolate, 3-foliolate, 5-foliolate, etc. leaves.

**Growth and development** In Indo-China, *S. tomentosa* flowers in December–January, in Malasia from October to June. Fruits mature in about 3–4 months and are dispersed by sea currents. Seeds are able to float for at least 3 months and in this period the germination capacity is not affected. However, germination significantly dropped following immersion in sea water. *S. tomentosa* has been observed nodulating in Hawaii.

**Other botanical information** *Sophora* comprises about 50 species and belongs to the tribe *Sophoreae* of the subfamily *Papilionoideae*. *S. tomentosa* has been subdivided into several subspecies; within Malasia only subsp. *tomentosa* occurs. The correct name for the well-known *Sophora japonica* L., which is used medicinally, as a dye and as an ornamental, is *Styphnolobium japonicum* (L.) Schott. The segregation of the small genus *Styphnolobium* from *Sophora* is supported by morphological evidence, a deviating somatic chromosome number and DNA research.

**Ecology** *S. tomentosa* is a coastal species, growing on sandy foreshores or open grassland along the beach, often locally abundant. It is more or less a pioneer species.

**Propagation and planting** In a germination experiment in Malaysia, seeds of *S. tomentosa* germinated in 0.5–11 months.

**Genetic resources and breeding** The risk of genetic erosion or depletion of *S. tomentosa* seems

rather limited, since it is pantropical and locally common.

**Prospects** The pharmacological properties from alkaloids such as (–)-cytisine, (–)-N-methylcytisine, (+)-matrine and (+)-matrine-N-oxide might be of interest (e.g. as lead compounds) in the research and development of new pharmaceuticals.

**Literature** |1| Chuang, C.Y., Xiao, J.G. & Chiou, G.C., 1987. Ocular anti-inflammatory actions of matrine. *Journal of Ocular Pharmacology* 3(2): 129–134. |2| Council of Scientific and Industrial Research, 1972. The wealth of India: a dictionary of Indian raw materials & industrial products. Volume IX. New Delhi, India. pp. 434–435. |3| Ishida, M. & Shinozaki, H., 1984. Glutamate inhibitory action of matrine at the crayfish neuromuscular junction. *British Journal of Pharmacology* 82(2): 523–531. |4| Kinoshita, T., Ichinose, K., Takahashi, C., Ho, F.C., Wu, J.B. & Sankawa, U., 1990. Chemical studies on *Sophora tomentosa*: the isolation of a new class of isoflavonoid. *Chemical and Pharmaceutical Bulletin* 38(10): 2756–2759. |5| Komatsu, M., Yokoe, I. & Shirataki, Y., 1978. Studies on the constituents of *Sophora* species. XI–II. Constituents of the aerial parts of *Sophora tomentosa* L. *Chemical and Pharmaceutical Bulletin (Tokyo)* 26(12): 3863–3870. |6| Murakoshi, I., Kidoguchi, E., Nakamura, M., Haginiwa, J., Ohmiya, S., Higashiyama, K. & Otomasa, H., 1981. (–)-Epilamprolobine and its N-oxide, lupin alkaloids from *Sophora tomentosa*. *Phytochemistry* 20(7): 1725–1730. |7| Niyomdham, C., 1980. Preliminary revision of Tribe Sophoreae (Leguminosae – Faboideae) in Thailand: *Ormosia* Jacks. and *Sophora* Linn. *Thai Forest Bulletin, Botany* 13: 1–22. |8| Perry, L.M., 1980. Medicinal plants of East and Southeast Asia. Attributed properties and uses. The MIT Press, Cambridge, Massachusetts, United States & London, United Kingdom. pp. 226–227. |9| van Steenis, C.G.G.J., 1948. Malaysian species of *Sophora* (Legum.). *Bulletin of the Botanic Gardens Buitenzorg, Ser. III, Vol. 17(4)*: 421–428. |10| Yamazaki, M. & Arai, A., 1985. On the contractile response of fundus strip from rats to matrine, an alkaloidal component of *Sophora flavescens*. *Journal of Pharmacobiodynamics* 8(7): 513–517.

**Other selected sources** 50, 97, 202, 284, 334, 487, 580, 855, 1006, 1020, 1022, 1035, 1036, 1039, 1128, 1178, 1444, 1480, 1608, 1609.

J.L.C.H. van Valkenburg



## Stephania Lour.

Fl. cochinch.: 608 (1790).

MENISPERMACEAE

$x = 11, 12, 13$ ; *S. japonica*:  $2n = 22, 24$

**Major species** *Stephania japonica* (Thunb.) Miers, *S. sinica* Diels.

**Vernacular names** Vietnam: b[if]nh v[oo]i (general).

**Origin and geographic distribution** *Stephania* consists of approximately 45 species and occurs in tropical and subtropical regions and in the warmer parts of temperate regions of the Old World. In Malesia, 12 or perhaps 13 species have been found, some of which have very wide distribution, particularly *S. japonica*.

**Uses** Many *Stephania* species are used in local medicine against a variety of complaints. In the Philippines, *S. japonica* is known as a cure for itches; in Papua New Guinea, *Stephania* roots and juice are applied to sores, cuts and stings. In India, the roots are used in the treatment of diarrhoea, dyspepsia, urinary diseases and heart ailments. Roots of *Stephania* spp. are employed in India and Indo-China in the treatment of pulmonary tuberculosis, asthma, dysentery, fever, hypoglycaemia and intestinal complaints (decoction, reported dose 4–5 g/day). Stems are used to treat ascariasis and dysmenorrhoea, leaves to treat indigestion, wounds, headache, sore breasts, dysuria, oliguria and oedema (decoction, reported dose 6–12 g/day), and flowers to treat leprosy. Root extracts of *S. glabra* (Roxb.) Miers have a long-standing reputation in India as an anti-dysenteric, antipyretic and antiasthmatic. In Thailand, *S. venosa* is commonly known as 'blood-soap', due to the red colour of its latex. The plant is often employed as a bitter tonic. The roots of *S. pierrei* Diels are used in Thai folk medicine as a skeletal muscle relaxant and also as an analgesic and tonic. A number of *Stephania* species are well known in traditional Chinese medicine; the most important are *S. tetrandra* S. Moore (analgesic, diuretic and antihypertensive) and *S. cepharantha* Hayata (analgesic, diuretic and tuberculostatic). In Africa (e.g. in Nigeria), *Stephania* extracts are also used medicinally.

Drugs from the tuberous roots are usually administered in the form of a decoction, as tincture or as a tablet with high alkaloid content. In Vietnam, for instance, (-)-tetrahydropalmatine is prescribed to treat neurasthenia and psychoses in the form of a 0.05 g tablet with a daily dose of 1–3 tablets.

*S. tetrandra* has been used in slimming regimens

because it eliminates oedema and 'stress-related water retention' but some cases of rapidly progressive fibrosing interstitial nephritis have been recorded in young women following this slimming regimen. However, the renal failure was not caused by the *Stephania* ingredient, since thorough investigations showed that *Stephania* had been confused with a species of *Aristolochia*, but these cases demonstrate how important it is to properly identify natural products to be used as medicines.

The leafy branches of *S. japonica* are sometimes used as forage.

**Production and international trade** *Stephania* species are only used on a local scale. However, recently the use of tuberous *Stephania* has gained importance, and in Vietnam they are much traded and also exported unregistered.

**Properties** Numerous alkaloids have been isolated from the tuberous roots and stems of *S. japonica*. Epistephanine and hypoeptephanine (major components), stepholine, stebisimine and insularine are alkaloids of the bisbenzylisoquinoline type. Stepinonine is a related alkaloid with a benzazepine structural element. A series of alkaloids derived from hasubanan (a structural isomer of morphinan) have also been isolated from the roots and stems of *S. japonica*: hasubanonine, metaphanin, homostephanoline, prometaphanine, stephamiersine, epistephamiersine, oxostephamiersine, stephasunoline, oxoprometaphanine, oxohasubanonine, oxoepistephamiersine and oxostephasunoline. Various other alkaloids have been isolated from the roots and/or stems, including stephadiamine (norhasubanan type), oxostephanine, lanuginosine and magnoflorine (aporphine type), cyclanoline and steponine (tetrahydroprotoberberine type), and protostephanine (bibenz[d,f]azonine type). The composition of alkaloid constituents seems to differ somewhat between plants collected in different habitats. The alkaloids oxostephamiersine and oxoprometaphanine (hasubanan type) and stebisimine (bisbenzylisoquinoline type) have been isolated from the leaves of *S. japonica*, and stephabenine, oxostephabenine and prostephanaberrine (all hasubanan type) from the fruits. Oxostephabenine can be converted by alkaline hydrolysis into N,O-dimethyloxostephine, whereas prostephanaberrine is converted into stephanaberrine upon treatment with aqueous HCl.

Four alkaloids have been isolated from the aerial parts of *S. japonica* var. *discolor*: (+)-epistephanine (bisbenzylisoquinoline type), magnoflorine

(aporphine type), aknadinine (= 4-demethylhasubanonine) and hernandifoline (hasubanan type).

(+)-Epistephanine possesses significant adrenergic neurone blocking activity. The compound acts like guanethidine, by selectively blocking the responses to sympathetic nerve stimulation without affecting the responses to the receptor agonist adrenaline. Parallel dose response curves suggest that both components act by the same mechanism, although (+)-epistephanine is approximately one tenth as potent as guanethidine, and the onset of action was slower and duration shorter than that of the latter. Finally, the bisbenzylisoquinoline alkaloid insularine is reported to have a curare-like activity.

Four isoquinoline alkaloids with (-)-tetrahydropalmatine (tetrahydroprotoberberine type) as the major alkaloid have been isolated from the tuberous roots of plants identified as *S. rotunda* Lour. (its status is uncertain); cepharamine, a hasubanan alkaloid, has been isolated from the leaves and stems. The alkaloidal content of the aerial parts of the plant was found to be quite different from that of the tuberous roots.

Investigations of the roots of *S. glabra* resulted in the isolation of two bisbenzylisoquinoline alkaloids ((-)-cycleanine and (-)-N-desmethylecycleanine), five tetrahydroprotoberberine alkaloids ((-)-capaurine, (-)-corynoxidine, (-)-tetrahydropalmatine, (-)-corydalmine and (-)-stepholidine), two aporphine alkaloids ((+)-stepharine and (+)-pronuciferine), and five quaternary protoberberine alkaloids (palmatine, palmatrubine, dehydrocorydalmine, jatrorrhizine and stepharanine).

Several alkaloids isolated from plants identified as *S. rotunda* or *S. glabra* possess pharmacological activity. (-)-Cycleanine has shown significant inhibition of nitric oxide production in vitro, and reduced the level of tumour necrosis factor in vivo, using a mouse model for fulminant hepatitis.

(-)-Stepholidine has shown sedative and antispastic effects in experiments with animals. Detailed pharmacological investigations revealed the compound to be a dopamine (DA<sub>2</sub>-subtype) antagonist in normal rats. Under certain experimental conditions (in 6-hydroxydopamine lesioned rats), however, dopamine (DA<sub>1</sub>-subtype) agonistic effects could be demonstrated. Further investigations on the calcium metabolism suggest that (-)-stepholidine may modulate its pharmacological actions by altering Ca<sup>2+</sup> regulating processes in the central dopaminergic nervous system.

(-)-Tetrahydropalmatine has been found to show antispasmodic (especially on the gastro-intestinal

tract), sedative and cardiotoxic activity. Investigations indicate the component to be a dopamine (DA<sub>2</sub>-subtype) antagonist. The interaction also has stereoselectivity; (-)-tetrahydropalmatine is a DA<sub>2</sub> antagonist whereas (+)-tetrahydropalmatine seems not to be.

The identity of *S. erecta* Craib is somewhat controversial; it is usually considered a synonym of *S. pierrei*. Comparison of the alkaloid content of both species indicated that different types of isoquinoline alkaloids were produced. Bisbenzylisoquinolines were the major alkaloids of *S. erecta*, whereas aporphine and tetrahydroprotoberberine type alkaloids represented the major constituents of *S. pierrei*. Isolation procedures guided by biological activity resulted in the isolation and identification of 23 isoquinolines from the latter species, which were subsequently tested for antimalarial and cytotoxic effects. Only the aporphine type alkaloid (-)-asimilobine demonstrated appreciable antimalarial activity together with a lack of cytotoxicity and may thus provide a good starting point for further development of more potent analogues. However, (-)-cycleanine also showed selective antiplasmodial activity.

At least 23 alkaloids have been identified in *S. venosa*, belonging to the protoberberine or (oxo)-aporphine type. The major alkaloids found in the roots were (+)-stepharine (aporphine type) and (-)-crebanine (aporphine type), each about 5% by weight.

*S. tetrandra* and *S. cepharantha* are well known in traditional Chinese medicine. The roots of *S. tetrandra* are a rich source of bisbenzylisoquinolines, in which S,S-tetrandrine is the predominant (0.7–1.3%) alkaloid. S,S-Tetrandrine has interesting pharmacological properties, including Ca<sup>2+</sup> channel blocking, anti-inflammatory and immunosuppressive activities (See under *Cyclea* for more detailed information). Cepharanthine, a bisbenzylisoquinoline alkaloid found in the roots of *S. cepharantha*, has been found to decrease leucopenia due to the use of antineoplastic agents, to inhibit collagen-induced blood platelet aggregation and to counteract the development of experimental silicosis. Cepharanthine has also been reported from dried roots of *S. pierrei* (about 1%); it is considered to be an immunostimulant and to decrease effectively the side effects of anti-cancer drugs, as well as to inhibit the growth of tuberculous bacteria. In in vitro experiments, cepharanthine inhibited proliferation of cancer cells by inducing apoptosis, and it was a highly potent inhibitor of HIV-1 replication.

Extracts from the leaves of *S. japonica* showed mild insecticidal properties against fruit flies in Thailand.

**Adulterations and substitutes** Other plants that contain isoquinoline alkaloids and are used in Chinese medicine include *Berberis*, *Coptis*, *Corydalis* and *Cyclea* species and several *Menispermaceae* not mentioned here.

**Description** Mostly slender dioecious climbers up to 20 m long; stem herbaceous or woody, usually glabrous but sometimes puberulous; roots often tuberous. Leaves arranged spirally, simple and entire, peltate, palmately veined, petiole usually geniculate at base; stipules absent. Inflorescence axillary or cauliflorous, usually composed of peduncled umbelliform, solitary or racemose cymes, sometimes flowers in capitula. Flowers unisexual, calyx with 1–8 free imbricate sepals, corolla with 2–4 free petals; male flowers with stamens fused into a peltate synandrium with 4–8 anther cells; female flowers with 1 carpel having a short-lobed or divaricately lacinate stigma. Fruit an obovoid drupe with style scar near base; endocarp bony, dorsally ornamented with a horseshoe-shaped band of 2 or 4 longitudinal rows of processes or transverse ridges. Seed horseshoe-shaped; embryo with cotyledons equalling the radicle, embedded in endosperm.

**Growth and development** Some *Stephania* species can develop 2–3 tuberous roots under favourable conditions, but usually there is only one. Several species are deciduous in the dry or cold season. Flowers are pollinated by small flies, bees, and possibly also small beetles and moths. The leaves of *S. japonica* produce a fragrance and may also play a role in attracting insects. The fruits may be dispersed by water.

**Other botanical information** *Stephania* belongs to the tribe *Menispermeae* and is most closely related to *Cissampelos* and *Cyclea*. It can be recognized by the inflorescences composed of umbelliform cymes or disciform capitula.

The taxonomy and nomenclature of the Asian *Stephania* species is still very complex. In Vietnam many species are exploited under a single common name bí[í]nh v[oo]i. The correctness of the name *Stephania rotunda* Lour. is still being debated; this name has often been applied to e.g. *S. glabra* (Roxb.) Miers, *S. venosa* (Blume) Spreng. and *S. pierreii* Diels. On nomenclatural grounds, these couplings are difficult to approve, since the *S. rotunda* Lour. is the oldest name. A large number of species not occurring in the Malesian region are used in Chinese and Indo-Chinese medicine, e.g.

*S. cepharantha*, *S. kwangsiensis* H.S. Lo, *S. longa* Lour., *S. pierreii* and *S. tetrandra*.

**Ecology** *Stephania* species occur in primary and secondary forest, regrowths, hedges, thickets and on river banks, sometimes on limestone and along the seashore, up to 2000(–2700) m altitude. Most species are slightly ombrophilous and hygrophilous, especially during the seedling period.

**Propagation and planting** *Stephania* is rarely planted. Cultivation trials have been conducted in Vietnam (1983–1984) and Georgia (1965–1969). Plants were propagated from root cuttings and seed, and planted in the fields at a spacing of 70 cm × 20 cm.

**In vitro production of active compounds** Cell suspension cultures of *S. glabra* have been studied in Russia, using a Murashige and Skoog medium supplemented with saccharose (30 g/l), vitamins, mezoinozite, 2,4-D (1 mg/l) and kinetine (0.1 mg/l). Eleven alkaloids were found in the culture, with no less than 0.3–0.8% stepharine, which was not detectable in plants grown in Georgia. (–)-Tetrahydropalmatine, which is the main alkaloid in the roots, was not detectable in the culture.

**Husbandry** In trials in Georgia, a combination of organic and chemical fertilizers (5 t of organic fertilizer, 150 kg N, 200 kg P and 120 kg K per ha) increased yields of tuberous roots by 27%.

**Diseases and pests** Three nematode species have been isolated from *S. japonica* in India.

**Harvesting** In Vietnam, it is recommended to harvest only tuberous roots in the weight class of 800–1000 g. This ensures a good quality of the product and a fair rate of regeneration.

**Yield** The yield per ha of fresh tuberous roots in Georgia was 11–13 t for 1-year-old plants, 28–35 t for 2-year-old plants and 42–44 t at the age of 3 years.

**Handling after harvest** The harvested roots are sliced and dried in the sun or in ovens at not too high temperature. The quality of the product varies considerably; for instance, the content of (–)-tetrahydropalmatine may vary from 0.3–3.6% of the dry tuberous root in *S. glabra*.

**Genetic resources and breeding** The *Stephania* species described here are widely distributed, locally rather common, and occur particularly in disturbed forest. They seem to be not very liable to genetic erosion. However, several species are rare and threatened in Indo-China and China, partly because of overcollecting. Conservation and planting of these species is essential to counteract genetic erosion which is serious, since the drugs have become popular in recent years.

**Prospects** The alkaloids which are particularly present in the tuberous roots of *Stephania* have interesting properties, as is the case in many other *Menispermaceae*. The roots are used in traditional medicine in different parts of the world for similar purposes, which also seems to confirm their effectiveness.

**Literature** |1| Bruneton, J., 1995. Pharmacognosy, phytochemistry, medicinal plants. Lavoisier Publishing, Paris, France. p. 748. |2| Forman, L.L., 1986. Menispermaceae. In: van Steenis, C.G.G.J. & de Wilde, W.J.J.O. (General editors): Flora Malesiana. Series 1, Vol. 10. Kluwer Academic Publishers, Dordrecht, the Netherlands. pp. 243–253. |3| Forman, L.L., 1991. Menispermaceae. In: Smitinand, T. & Larsen, K. (Editors): Flora of Thailand. Vol. 5(3). The Forest Herbarium, Royal Forest Department, Bangkok, Thailand. pp. 311–323. |4| Kozuka, M., Miyaji, K., Sawada, T. & Tomita, M., 1985. A major alkaloid of the leaves and stems of *Stephania rotunda*. Journal of Natural Products 48(2): 341–342. |5| Matsui, M., Kabashima, T., Ishida, K., Takebayashi, T. & Watanabe, Y., 1982. Alkaloids of the leaves of *Stephania japonica* (Japan). Journal of Natural Products 45(4): 497–500. |6| Matsui, M. & Yamamura, Y., 1986. Alkaloids from the fruits of *Stephania japonica*, part 3. Structures of prostephaneberrine and stephanaberrine, two new hasubanan alkaloids. Journal of Natural Products 49(4): 588–592. |7| Nguyen Van Duong, 1993. Medicinal plants of Vietnam, Cambodia and Laos. Mekong Printing, Santa Ana, California, United States. pp. 260–261. |8| Patra, A., Ghosh, A. & Mitra, A.K., 1980. Alkaloids of *Stephania glabra*. Planta Medica 40(4): 333–336. |9| Taga, T., Akimoto, N. & Ibuka, T., 1984. Stephadamine, a new skeletal alkaloid from *Stephania japonica*: the first example of a C-norhasubanan alkaloid. Chemical and Pharmaceutical Bulletin 32(10): 4223–4225. |10| Vanherweghem, J.L. et al., 1993. Rapidly progressive interstitial renal fibrosis in young women and association with slimming regimen including Chinese herbs. Lancet (North American Edition) 341 (8842): 387–391.

#### *Selection of species*

#### ***Stephania capitata* (Blume) Sprengel**

Syst. veg. 4(2): 316 (1827).

**Vernacular names** Indonesia: areuy geureung, areuy camcau minyak (Sundanese), sumpat kendi (Javanese).

**Distribution** Peninsular Thailand, Peninsular Malaysia, Sumatra, Java, Bali and Borneo.

**Uses** The leaves have been reported to be used in Java as a substitute for those of *Cyclea barbata* Miers to prepare 'cincau', which is used as a refreshment and as a medicine against stomach complaints.

**Observations** A slender woody climber up to 15 m long, root tuberous and fusiform; leaves lanceolate to broadly ovate, 6–17 cm × 2.5–10.5(–14) cm; flowers in peduncled capitula on a short axillary axis, sessile; fruit on a slender stalk, red, endocarp dorsally with 4 rows of capitate projections. *S. capitata* occurs scattered in forest on plains and mountains up to 2000 m altitude.

**Selected sources** 202, 421, 423, 580.

#### ***Stephania japonica* (Thunb.) Miers**

Ann. Mag. Nat. Hist. ser. 3, 18: 14 (1866).

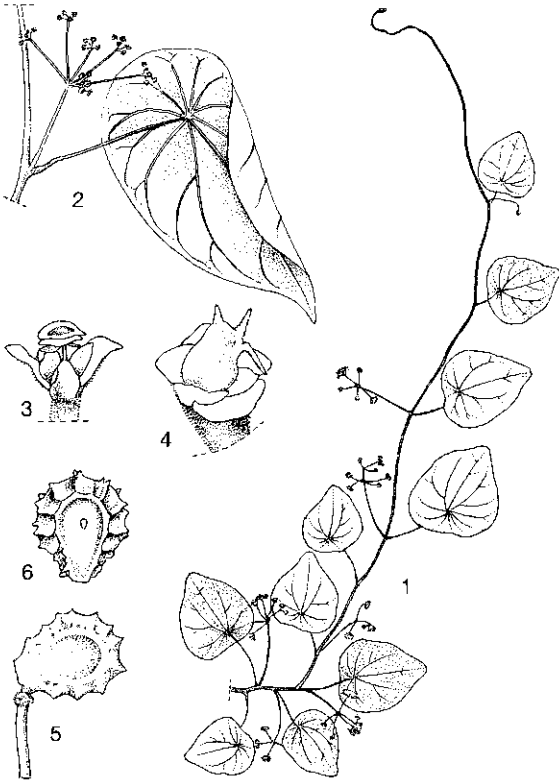
**Vernacular names** Indonesia: areuy geureung (Sundanese), kepleng (Javanese), ginato bobudo (Moluccas). Philippines: malabuta (Igorot), maratugi (Iloko), kuren (Ibanag). Thailand: kon pit (central), pang pon (northern), tap tao (peninsular). Vietnam: thi[ee]n kim d[aw]fng, d[aa]y l[ox]i ti[ee]fn.

**Distribution** Nepal, India, Burma (Myanmar), Indo-China, southern China, Taiwan, Japan, Thailand, throughout Malesia, northern and eastern Australia and Polynesia.

**Uses** The tuberous root is used in local medicine to treat dysentery, stomach-ache, fever, urinary disorders, hepatitis, inflammation and itch. Crushed leaves in water are applied to breast infections.

**Observations** A slender climber up to 10 m long, root tuberous, stem herbaceous or thinly woody; leaves broadly triangular-ovate to ovate, (4–)6–12(–17) cm × 4–10(–14) cm; flowers in an axillary compound umbelliform cyme, sessile or subsessile; fruit sessile or subsessile, red, endocarp dorsally with 4 rows of processes. *S. japonica* is subdivided into 3 varieties based on the hairiness of lower leaf surface and inflorescence: var. *japonica*, var. *timoriensis* (DC.) Forman (synonym: *S. forsteri* (DC.) A. Gray) and var. *discolor* (Miq.) Forman (synonym: *S. hernandiifolia* (Willd.) Walp.). *S. japonica* occurs in secondary forest, re-growth, hedges, thickets and on river banks, also in *Nypa* swamp, up to 2000 m altitude.

**Selected sources** 190, 202, 421, 423, 580, 642, 643, 798, 915, 916, 917, 918, 919, 1031, 1035, 1178, 1209, 1436, 1611.



*Stephania japonica* (Thunb.) Miers - 1, flowering twig; 2, part of stem with leaf and inflorescence; 3, male flower; 4, female flower; 5, fruit; 6, endocarp.

### *Stephania sinica* Diels

Engl., Pflanzenr. IV 94 (Heft 46): 272 (1910).

**Vernacular names** Vietnam: b[if]nh v[oo]i, c[ur] m[oo]t, c[af] t[of]m.

**Distribution** Northern Vietnam, Laos and southern China.

**Uses** The tuberous roots are used as a remedy for neural anodyne, to treat insomnia, asthma, dysentery, acute stomach trouble and sore throat. The pounded fresh tuberous root is applied as a poultice on pimples and wounds.

**Observations** A twining climber, stem glabrous, root tuberous, up to 70 kg; leaves suborbicular, herbaceous-membranous, round or nearly so at the base, apex acuminate, palmately 10-11-veined; flowers in an axillary compound umbelliform cyme, orange pink; fruit globose compressed, up to 6.5 mm in diameter, endocarp with 4 transverse dorsal ribs.

**Selected sources** 1031, 1130.

### *Stephania venosa* (Blume) Spreng.

Syst. veg. 4(2): 316 (1827).

**Vernacular names** Indonesia: gorong bodas (Sundanese). Thailand: plao lueat khruua (northern), kling klang dong (south-western), boraphet yang daeng (peninsular). Vietnam: l[ox]i ti[ee]n d[or].

**Distribution** The Andaman Islands, Vietnam, southern China, Thailand, northern Peninsular Malaysia, northern Sumatra, Java, Borneo (Sabah), south-western Sulawesi and the Philippines (Luzon).

**Uses** *S. venosa* is cultivated for medicinal purposes in Thailand. The red juice is used for tattooing by forest tribes in Thailand.

**Observations** A slender climber up to 20 m long, root tuberous, leafy stem herbaceous, containing red sap; leaves broadly triangular-ovate, 6-20 cm x 6-20 cm; flowers in an axillary umbelliform cyme, pedicellate; fruit stalked, red, endocarp with 4 dorsal rows of papilliform processes. *S. venosa* occurs scattered, but locally common, in forest on hillsides, plains and mountains, sometimes on limestone, up to 1600 m altitude.

**Selected sources** 238, 421, 423, 1132.

Nguyen Tien Ban, Bui Thi Bang,  
Nguyen Tap & Nguyen Chieu

### *Strychnos* L.

Sp. pl. 1: 189 (1753); Gen. pl. ed. 5: 86 (1754).

LOGANIACEAE

$x =$  unknown; *S. ignatii*:  $2n = 44$ , *S. nux-vomica*:  $2n = 44$

**Major species** *Strychnos ignatii* Bergius, *S. nux-vomica* L.

**Vernacular names** Malaysia: akar ipoh.

**Origin and geographic distribution** *Strychnos* consists of 150-200 species and is distributed throughout the tropics and subtropics. Approximately 25 species occur in the Malesian region.

**Uses** The seeds, bark, wood and roots (e.g. of *S. ignatii* and *S. nux-vomica*) had or have numerous applications in traditional medicine. *Strychnos* was already being used medicinally in China in the 14th Century. It has been prescribed as a stomachic, febrifuge, vermifuge, anticholeric and tonic and to treat sores, wounds, eczema and snake bites in Indonesia and the Philippines. In India, the seeds have been used to obstinate vomiting, to treat cholera, diarrhoea, asthma, dropsy, rheumatism, paralytic and neuralgic affections, spermatorrhoea, epilepsy, diabetes, anaemia and

chlorosis, and as a tonic, febrifuge and vermifuge, and to treat alopecia. In Australia, the fruit pulp of *S. lucida* has been used to treat a variety of skin complaints. *S. wallichiana* Steudel ex DC. (synonym: *S. gauthieriana* Pierre ex Dop p.p.) is used in traditional medicine in Vietnam to treat rabies, leprosy, and as an aphrodisiac. *S. potatorum* L. is used in both the Ayurvedic and Yunani systems of medicine in India where its bark is used to adulterate *S. nux-vomica* bark and its seeds are used as a natural coagulant to purify drinking water. Several *Strychnos* species are used medicinally in Africa and South America.

*Strychnos* has also been used as an ingredient in tonic preparations, but these are no longer used in modern therapy, except for a few rare proprietary products based on 'nux vomica' tincture (from *S. nux-vomica*) or Saint Ignatius beans (seeds of *S. ignatii*).

In general, all preparations containing *Strychnos* should be used in small doses, and with the greatest caution, because of their toxicity.

Strychnine was formerly used mainly to poison rodents. The drug was introduced in Europe as long ago as the 16th Century to eliminate pests.

*S. axillaris* Colebr. has apparently been used in Peninsular Malaysia in the preparation of arrow poison; its leaves have been used in India as a suppurative and the seeds internally as a febrifuge. The root bark of *S. vanprukii* Craib has been reportedly used to prepare an arrow poison in Peninsular Malaysia; *S. rufa* C.B. Clarke has been used similarly.

In Borneo and other regions *Strychnos* seeds, bark and roots are used for making dart poison for blowpipes, usually in combination with the sap of *Antiaris toxicaria* Lesch. In South America, *Strychnos* (mainly *S. castelnaei* Wedd., *S. guianensis* (Aubl.) Mart. and *S. toxifera* Schomb. ex Benth.) is used to prepare the so-called gourd or calabash curare. Tube (or bamboo) curare is made from lianescent *Chondrodendron* species (*C. tomentosum* Ruiz & Pavon). Both curares, alone or as a mixture, are used as arrow poison.

The stems of lianescent species are used for rough cordage. The wood of species with a tree habit is occasionally used for implements and cabinet work.

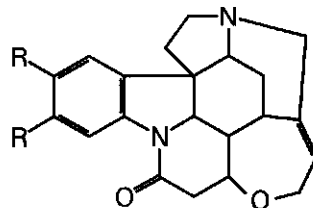
**Production and international trade** The annual production of *S. nux-vomica* seed in India was estimated at 2000 t at the beginning of the 1970s. In the period 1965–1971 the average production of strychnine and brucine from *S. nux-vomica* seeds in India was 18 000 kg/year. Most of

the alkaloids are exported to the United States and Europe.

**Properties** The drug usually consists of dried seeds. The seeds of *S. nux-vomica* contain 1–3% total alkaloids (of the indole type; biosynthetically derived from tryptophan), chiefly represented by strychnine (1.1–1.5(–2.3%)) and its dimethoxylated derivative brucine (1.1–2.1(–3.6%)). Bark, wood, roots and flowers also contain these compounds, but with strychnine in smaller amounts. Wood collected from *S. lucida* in Java during the rainy season showed a higher alkaloid content than that of wood harvested in the dry period of the year. Several minor alkaloids, with closely related structures, have also been isolated from *S. nux-vomica* seeds (total concentration up to 1%): 12-hydroxy-strychnine, 15-hydroxy-strychnine,  $\alpha$ -colubrine,  $\beta$ -colubrine, icajine, 11-methoxy-icajine, novacine, vomicine, pseudostrychnine, pseudo-brucine, pseudo- $\alpha$ -colubrine, pseudo- $\beta$ -colubrine, N-methyl-sec-pseudo- $\beta$ -colubrine and isostrychnine.

The seeds of *S. ignatii* contain 1.5–2% of strychnine, 0.5% of brucine and the minor related alkaloids 12-hydroxystrychnine,  $\alpha$ -colubrine, icajine, novacine and vomicine. Small amounts of berberine (an isoquinoline alkaloid) have also been reported. The major alkaloid in the wood of *S. lucida* is brucine. The bis-indole alkaloid longicaudatine has been isolated from several *Strychnos* spp., including *S. ignatii* and *S. nux-vomica*. This compound has strong reserpine-like activity.

Strychnine is a very toxic alkaloid. The lethal dose in adult humans may start at about 0.4 mg/kg. In small doses the compound produces excitation of all parts of the central nervous system. It acts as a competitive inhibitor of the neurotransmitter glycine at its receptor-binding site located at the Renshaw cells in the spinal cord. By binding, strychnine blocks the normal inhibitory action of glycine and the Renshaw cells on the motor-neurons, thus leading to a spread of motor-cell stimulation. Intoxication may cause anxiety, increased sensitivity to noise and light and periodic convul-



strychnine (R = H) and brucine (R = OCH<sub>3</sub>)

sive attacks: all the muscles contract, forcing the patient into a position, with the back arched and resting only on the head and heels. Death may occur by asphyxia following the contraction of the diaphragm. Brucine is less active as a poison; about 50–100 less than a comparable dose of strychnine.

Strychnine and brucine can be distinguished by thin-layer chromatography of e.g. a macerate of the simplex in 70% ethanol. Quantification can be achieved by spectrophotometry on an alkaloid extract; the difference between the absorbances at 258 nm ( $\lambda^{\max}$  of strychnine) and 300 nm ( $\lambda^{\max}$  of brucine) is taken into account. Reversed-phase HPLC procedures for qualitative and quantitative analysis, and more recently a capillary zone electrophoresis method for quantitative estimation of strychnine and brucine in *S. nux-vomica* seeds are also available. According to the Netherlands Pharmacopoeia VI, for the drug to be of good quality the strychnine concentration should not be less than 1%.

Some alkaloids from African *Strychnos* species have shown potential as anti-cancer agents in animal tests, and also anti-amoebic and anti-plasmodial activity in mice. Alkaloids isolated from South American *Strychnos* have demonstrated a wide anti-microbial spectrum.

Galactomannans and galactans have been demonstrated in the seeds of *S. nux-vomica*. These polysaccharides, which have coagulant properties, are also present in the seeds of the Indian *S. potatorum*. The seed extract is efficient in the coagulation-flocculation of hydrophobic colloids (such as a clay suspension), but it is a poor flocculant in the case of hydrophilic colloids (such as bacteria). The seed extract of *S. nux-vomica* exhibits analgesic, anti-ulcer, cytotoxin and uterine stimulant activity.

Aqueous and ethanolic extracts of *Strychnos* leaves showed antifungal activity against several pathogens of rice in India.

**Adulterations and substitutes** As well as occurring in *Loganiaceae*, monoterpenoid indole alkaloids also occur particularly in *Rubiaceae* (e.g. *Cinchona*, *Uncaria*) and *Apocynaceae* (e.g. *Catharanthus*, *Rauwolfia*, *Voacanga*). Several species from these families and genera have uses comparable to *Strychnos*.

**Description** Lianas or sometimes shrubs (often scrambling) to treelets; stem usually with axillary simple or double tendrils, sometimes with axillary thorns. Leaves opposite, simple and entire, and except for the midrib having 1–2(–3) pairs of near-

ly equally strongly developed basal veins which do not fully reach the leaf apex; petiole mostly inserted upon distinct leaf-cushions; stipules reduced to a mostly ciliate and straight rim connecting the leaf bases. Inflorescence terminal or axillary, thyrsoid; bracts scale-like. Flowers bisexual, actinomorphic, (4–)5-merous; calyx divided nearly to the base, lobes ciliate on the margins, brown; corolla gamopetalous, rotate to salver-shaped, sometimes thickened towards the lobes, white to yellowish or greenish, lobes valvate in bud, spreading to reflexed, tube usually densely papillose outside, variably hairy inside; stamens inserted on the corolla tube alternating with the lobes, exerted, anthers basifixed, longitudinally dehiscent, introrse; ovary superior, 2-celled, style cylindrical, stigma faintly 2-lobed. Fruit a globose or ellipsoid berry, with hard shell, smooth or minutely warty, glabrous, orange to red when ripe, with fleshy, usually orange pulp, 1–many-seeded. Seeds lenticular, orbicular to ellipsoid, often convex on one side and concave on the other side, with a silky or felty testa or glabrous; endosperm copious, bony; embryo minute, straight, with small cotyledons. Seedling with epigeal germination; cotyledons emergent, thin and leaf-like, long persistent; hypocotyl long.

**Growth and development** The flowers of *Strychnos* are pollinated by insects. Mammals (e.g. monkeys and civet-cats) and birds digest the fruit pulp and disperse the seeds.

**Other botanical information** Flowers are essential when identifying *Strychnos* species. Fruits and seeds are also characteristic features of *Strychnos* species and should be kept in spirit (when collecting herbarium material) to preserve important characters that will disappear upon drying.

**Ecology** Most *Strychnos* are large lianas of the forest, but a few are scrambling to erect shrubs or small to medium-sized trees occurring in more open habitats (e.g. *S. lucida*).

**Propagation and planting** In India, *S. nux-vomica* plants have been successfully regenerated from hypocotyl tissue. Seeds were germinated on Murashige and Skoog medium supplemented with 1 mg/l gibberellic acid, and hypocotyls excised from the resulting seedlings were inoculated on Murashige and Skoog medium. The optimal growth of plantlets occurred when the medium was supplemented with 2 mg/l kinetin and 1 mg/l naphthalene acetic acid. In vitro multiplication was also successful using nodal explants excised from healthy mature trees.

**Diseases and pests** Leaf galls induced by the jumping plant louse *Diaphorina truncata* have been reported for *S. nux-vomica* in India, resulting in abscission of the leaves.

**Harvesting** Fruits of *S. nux-vomica* are usually gathered from the trees. Fallen fruits are considered to be of inferior quality.

**Handling after harvest** After collecting, seeds of *Strychnos* are cleaned and dried in the sun. Dried seeds can be stored in jute bags for a long period without any loss of alkaloidal content, but they should be kept in dry conditions to prevent deterioration by fungi such as *Aspergillus* and *Penicillium* species. Seeds are further processed in factories to obtain strychnine and brucine.

In traditional Chinese medicine, seeds of *S. nux-vomica* are usually processed to reduce their toxicity. Sand or sesame oil is heated in an iron pan up to 235°C and seeds are parched for about 3 minutes. This significantly reduces the contents of strychnine, brucine and  $\beta$ -colubrine and increases the amounts of isostrychnine, isobrucine, strychnine N-oxide and brucine N-oxide.

**Genetic resources and breeding** The *Strychnos* species mentioned have a large area of distribution and are, at least locally, common. They are too little used to be threatened at present. However, forest destruction may easily endanger *Strychnos* spp. with a narrow distribution. *S. nux-vomica* is widespread in India, Indo-China and Thailand, but is rare in the Malesian region. There are no records of *Strychnos* in germplasm collections.

**Prospects** The importance of *Strychnos* as a medicinal plant and vermin destroyer has diminished in recent years. It is now hardly used in modern phytotherapy. Some of the alkaloids present in the plants may play a role in the development of new anti-cancer or antimalarial drugs, but research is needed to establish the possibilities.

**Literature** [1] Bisset, N.G., 1974. The Asian species of *Strychnos*. Part III. The ethnobotany. *Lloydia* 37(1): 62–107. [2] Bisset, N.G. & Phillipson, J.D., 1976. The Asian species of *Strychnos*. Part IV. The alkaloids. *Lloydia* 39(5): 263–325. [3] Bruneton, J., 1995. Pharmacognosy, phytochemistry, medicinal plants. Lavoisier Publishing, Paris, France. pp. 829–830. [4] Cai, B.-C., Hattori, M. & Namba, T., 1990. Processing of nux vomica. II. Changes in alkaloid composition of the seeds of *Strychnos nux-vomica* on traditional drug-processing. *Chemical and Pharmaceutical Bulletin* 38(5): 1295–1298. [5] Corsaro, M.M., Giudicianni,

I., Lanzetta, R., Marciano, C.E., Monaco, P. & Parrilli, M., 1995. Polysaccharides from seeds of *Strychnos* species. *Phytochemistry* 39(6): 1377–1380. [6] Council of Scientific and Industrial Research, 1976. The wealth of India: a dictionary of Indian raw materials & industrial products. Vol. 10. New Delhi, India. pp. 61–68. [7] Kopp, B., Bauer, W.P. & Bernkop-Schnürch, A., 1992. Analysis of some Malaysian dart poisons. *Journal of Ethnopharmacology* 36(1): 57–62. [8] Kumar, A. & Datta, S.K., 1989. Plantlet regeneration from hypocotyl tissue of *Strychnos nux-vomica* Linn. *Current Science* 58(14): 812–813. [9] Leenhouts, P.W., 1962. Loganiaceae. In: van Steenis, C.G.G.J. (General editor): *Flora Malesiana*. Series 1, Vol. 6. Wolters-Noordhoff Publishing, Groningen, the Netherlands. pp. 343–361. [10] Massiot, G. et al., 1983. Occurrence of longicaudatine, a new type of bis-indole base and bisnor-C alkaloid H in *Strychnos* spp. *Journal of Organic Chemistry* 48(11): 1869–1872.

#### *Selection of species*

### ***Strychnos ignatii* Bergius**

Mater. med. 1: 146 (1778).

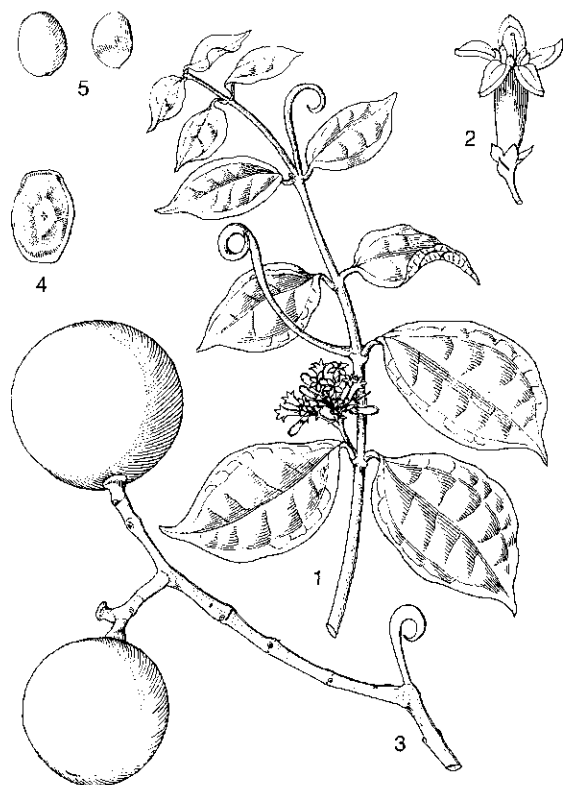
**Synonyms** *Strychnos tieute* Lesch. (1810), *Strychnos ovalifolia* Wallich ex G. Don (1837), *Strychnos beccarii* Gilg (1897).

**Vernacular names** Saint Ignatius bean (En). Fèves de Saint-Ignace (Fr). Pepita de San Ignacio (Sp). Indonesia: pokru (Sundanese), cetek (Javanese), bina (Kalimantan). Malaysia: akar ipoh, belai hitam, ipoh akar besar (Peninsular). Philippines: katbalonga (Tagalog), igasud (Bisaya), pepita-sa-katbalogan (Tagalog, Bisaya, Pampango/Kapampangan). Thailand: phayaa mue lek (Krabi). Vietnam: lo[af]ng n[af]n.

**Distribution** Indo-China, Hainan, Thailand, Peninsular Malaysia, Java, Borneo and the south-eastern Philippines (Biliran, Samar, Leyte and Mindanao).

**Uses** The seeds and bark have been prescribed as a stomachic, febrifuge, anticholeric and tonic in the Philippines. In Thailand, the seeds, roots and wood are used as a stomachic, febrifuge and to treat malaria. Leaves are used to treat diabetes. In India, the seeds have been used to obstinate vomiting, to treat cholera, asthma, dropsy and rheumatism, and as a tonic and vermifuge. The seeds are commercially traded as a source of strychnine. The bark and the seeds are used to treat stomach-ache in Vietnam. The roots are





*Strychnos ignatii* Bergius - 1, flowering twig; 2, flower; 3, twig with fruits; 4, lenticular-shaped seed; 5, irregularly shaped seeds.

used in Indonesia and Malaysia as arrow poison, and sometimes as a fish poison.

**Observations** A liana up to 35 m long, with stem up to 5 cm in diameter, bark densely and finely lenticellate, brown or grey, tendrils simple; leaves ovate or elliptical to lanceolate, 4-18(-22) cm × 2-9(-12) cm, petiole 5-10 mm long; inflorescence axillary, 10-20-flowered; corolla 7-17 mm long, tube about 3 times longer than lobes, with some long woolly hairs inside in the lower half; fruit globose, 4-12 cm in diameter, about 10-seeded; seeds lenticular to ellipsoid or orbicular, 20-35 mm × 16-20 mm × 8-9 mm, silky to felty, or irregularly shaped like castor beans, about 20 mm × 10 mm × 10 mm and rough but glabrous. *S. ignatii* is usually found in dense forest, often in dipterocarp forest on sandy soil or in mixed rain forest, sometimes on river banks, up to 1500 m altitude.

**Selected sources** 160, 163, 202, 287, 580, 769, 839, 911, 1126, 1178, 1460, 1475.

### *Strychnos lucida* R.Br.

Prodr.: 469 (1810).

**Synonyms** *Strychnos ligustrina* Blume (1836).

**Vernacular names** Indonesia: bidara laut (general), dara laut (Javanese), kayu ular (Sumatra, Timor). Thailand: phayaa mue lek, phayaa muun lek (central), sieo duuk (northern). Vietnam: m[ax] ti[eef] ll[as]ng.

**Distribution** Indo-China, Thailand, eastern Java, the Lesser Sunda Islands, the southern Moluccas and northern Australia.

**Uses** The bark, wood and roots are used in traditional medicine in Indonesia to treat fever, snake bites, sores, wounds, eczema, and as stomachic and vermifuge. Australian aborigines apply the fruit pulp to the skin to treat skin complaints such as scabies, rashes, burns, leprosy, sores and cuts. The roots are used to treat diabetes. Leaves and fruits are used as a fish poison in Australia.

**Observations** A shrub or small deciduous tree up to 12 m tall, bole often crooked and up to 25 cm in diameter, spiny when young, branches densely and finely lenticellate, grey, tendrils absent; leaves ovate or elliptical to suborbicular, 2.5-10 cm × 1.5-6 cm, petiole 2-4 mm long; inflorescence terminal, c. 10-flowered; corolla 10-15 mm long, tube about 3 times as long as lobes, sometimes with a few woolly hairs inside; fruit globose, 2-2.5 cm in diameter, 2-3-seeded; seeds nearly disk-shaped, 12-15 mm × 10-12 mm × 2.5-5 mm, densely short pubescent. *S. lucida* occurs in regions with a monsoon climate, in teak forest and other dryland forest, often in secondary forest but also in scrubs and savannas, up to 200(-400) m altitude; in Thailand also on limestone hills.

**Selected sources** 160, 163, 202, 276, 580, 839, 957, 1126.

### *Strychnos minor* Dennst.

Schlüss. Hortus malab.: 33 (1818).

**Synonyms** *Strychnos laurina* Wallich ex DC. (1845), *Strychnos multiflora* Benth. (1856), *Strychnos colubrina* auct. non L.

**Vernacular names** Indonesia: ranosandang (Sulawesi), ipu tanah (Kalimantan), wale ammelum (Moluccas). Malaysia: lengkoyan, semiyo akar (Peninsular). Philippines: bukuan (Ibanag, Negrito), pamulaklakin (Tagalog), bugahin (Bisaya). Thailand: tum kaa khao, tum kaa daeng (Lampang). Vietnam: kim lu[oo]ng (Biên Hoa), thu[oor]c m[oj]i (Tây Ninh).

**Distribution** From India, Sri Lanka and Burma (Myanmar), through Indo-China (southern Vietnam), Thailand and the whole of Malesia (ex-

cept Java), to the Solomon Islands and northern Australia.

**Uses** The wood, bark and roots are used medicinally; a decoction is prescribed in the Philippines as an emmenagogue and to treat throat complaints, and it is also used as arrow poison. In Thailand, the stems are used for their carminative, antipyretic and stomachic activity. The stems are used for tying.

**Observations** A liana up to 12 m long, with bark sometimes densely lenticellate, grey, tendrils double; leaves ovate or suborbicular to lanceolate, 8–25 cm × 3–12 cm, petiole 3–15 mm long; inflorescence axillary and/or terminal, many-flowered; corolla 3–6.5 mm long, tube about as long as lobes, inside in the upper half with woolly hairs; fruit globose, (1–)2–3 cm in diameter, sometimes ellipsoid and 3.5 cm long, about 1–8-seeded; seeds lenticular, 12–15 mm × 10–12 mm × 2–4 mm, minutely densely tomentose. *S. minor* occurs in primary and secondary forest, in New Guinea also in *Araucaria* forest, up to 1850 m altitude.

**Selected sources** 160, 163, 202, 276, 287, 839, 1126, 1178, 1460, 1475.

### ***Strychnos nux-vomica* L.**

Sp. pl. 1: 189 (1753).

**Vernacular names** Nux-vomica tree, strychnine plant (En). Cambodia: slaêng, slaêng thom. Laos: 'sêng bua' (Vientiane). Thailand: krachee, tuumka daeng (central), salaeng thom (Nakhon Ratchasima). Vietnam: c[ur] chi (general), c[oo] chi (Khanh Hoa), m[ax] ti[eff]n.

**Distribution** India, Bangladesh, Sri Lanka, Burma (Myanmar), Indo-China (Cambodia, Laos, southern Vietnam), Thailand and northern Peninsular Malaysia; introduced and locally naturalized in the Philippines (Mindoro).

**Uses** The seeds have been used in traditional medicine in many regions as a nervine, stomachic, tonic, aphrodisiac and respiratory and cardiac stimulant, the bark as tonic and febrifuge. The seeds are used in Vietnam to treat rheumatic arthritis and paralysis. In Thailand the leaves are used in the treatment of skin diseases. In India, they are also used as mild irritating rubefacient product to treat alopecia by intensifying the capillary blood flow and keeping the scalp and hair follicles active. In traditional Chinese medicine the dried seeds have been used for promoting blood circulation, alleviating blood stasis and relieving pain. The seeds constitute a major source of strychnine, and the plant is cultivated for this purpose. The wood is used in India for agricultur-

al implements and tool handles, cart wheels and fancy cabinet work.

**Observations** A small to medium-sized tree up to 25 m tall, bole up to 100 cm in diameter, sometimes a liana, branches not rough, yellowish-grey, sometimes with axillary thorns, tendrils absent; leaves broadly ovate to elliptical or suborbicular, 5–18 cm × 4–12.5 cm, petiole 5–12 mm long; inflorescence terminal on short axillary branchlets with usually one pair of leaves, fairly many-flowered; corolla 10–13 mm long, tube about 3 times longer than lobes, sparsely woolly hairy in lower half inside; fruit globose, 2.5–4(–6) cm in diameter, 1–4-seeded; seeds lenticular, orbicular to elliptical, 20–23 mm × 18–20 mm × 4 mm, densely sericeous. *S. nux-vomica* occurs often at the edge of dense forest, on river banks and along the shore, on loamy or loamy-sandy soil.

**Selected sources** 107, 160, 163, 193, 202, 208, 285, 287, 332, 580, 790, 839, 911, 1035, 1126, 1178, 1460, 1475.

Purwaningsih

### ***Styphnolobium japonicum* (L.) Schott**

Wiener Zeitschr. Kunst. 3: 844 (1830).

LEGUMINOSAE

2n = 28

**Synonyms** *Sophora japonica* L. (1767).

**Vernacular names** Japanese pagoda tree, Chinese scholar tree, umbrella tree (En). Indonesia: sari cina, sari kuning. Vietnam: c[aa]y h[off]e.

**Origin and geographic distribution** *S. japonicum* is native to central and northern China and Korea, but now widely cultivated in temperate and subtropical regions of the world, in South-East Asia at least in Vietnam and Thailand.

**Uses** The flower buds of *S. japonicum* are astringent and possess styptic properties. They are a reputed remedy for the prevention of various types of haemorrhages (e.g. haemoptysis, epistaxis, metrorrhagia), haemorrhoids, and are useful for the treatment of hypertension. Flower buds and young pods are an important source of rutin, which has 'vitamin P'-like properties and is used in the treatment of conditions characterized by increased capillary permeability and fragility. An extract from the pods is toxic. *S. japonicum* is also reputed to show oestrogenic activity.

The flower buds and occasionally also the pods are used for dyeing yellow or granite-grey. The dye was mainly used to dye silk. Mixed with indigo, it gives a green colour. *S. japonicum* is commonly





*Styphnolobium japonicum* (L.) Schott - 1, flowering twig; 2, flower; 3, stamens and ovary; 4, pod.

nate, 15–25 cm long; stipules early caducous; leaflets alternate to subopposite, 7–17, elliptical to ovate-lanceolate, 1.5–6 cm × 1–2.5 cm, acute or sometimes obtuse at apex, mucronate, glabrous or sparsely hairy above, short-haired below, shortly petiolulate; stipellae absent or small and setaceous. Flowers in a terminal, 15–35 cm long panicle, papilionaceous; bracteoles present; calyx 3–4 mm long, teeth shorter than the tube; corolla yellowish-white or greenish-white, standard 12–15 mm long; stamens 10, filaments joined near the base; ovary superior, pilose. Fruit an indehiscent pod, 3–12 cm × 7–12 mm, constricted between the seeds, stipitate, glabrous, beaked, 1–8-seeded. Seeds 8 mm × 4–5 mm, yellowish-brown. Seedling with epigeal germination; cotyledons emergent; first few leaves imparipinnate or sometimes 1- or 3-foliolate.

**Growth and development** Seeds of *S. japonicum* germinate quickly, usually within about 4 days. The tree starts flowering when about 3–4 years old, though sometimes only when about 30 years old. In Vietnam, trees flower from May to

August and fruit from September to November. In China, flowering is in August–September, fruiting in October–November.

**Other botanical information** *Styphnolobium* is a small genus of 9 species occurring in North and Central America, with *S. japonicum* as the only Asian representative. The segregation of *Styphnolobium* from the closely related genus *Sophora* is supported by morphological characters (flowers with bracteoles, pods indehiscent), a different somatic chromosome number and the results of DNA studies. The DNA studies even indicate that the two genera are not closely related. Several varieties have been distinguished within *S. japonicum*, the status of which is still unclear, and several cultivars are grown as ornamentals.

**Ecology** *S. japonicum* is well adapted to dry weather conditions and to a great variety of soils, but thrives best in well-drained, sandy loam. Being native in temperate and subtropical regions, cultivation in the tropics is only possible in drier regions or at higher altitudes. Under temperate conditions *S. japonicum* is tolerant of heat and drought, whereas it also tolerates severe frost (up to  $-25^{\circ}\text{C}$ ) except when young.

**Propagation and planting** *S. japonicum* can be propagated by seed, which should be scarified or treated with hot water and soaked before sowing. Under temperate conditions the seed germinates well and within a few days. Grafting, layering, greenwood and root cuttings are used for ornamental cultivars. Trees can be coppiced successfully.

**Diseases and pests** Among the reported diseases and pests of *S. japonicum* are fungi such as *Uromyces truncicola* which cause a canker disease in seedlings, and leafminers such as *Odontota dorsalis*, and aphids (*Aphis* spp.).

**Harvesting** Flower buds, inflorescences and young fruits of *S. japonicum* are hand picked. Inflorescences are harvested when flowers begin to open. Young stems can simply be cut as routine under coppice management.

**Yield** Analyses of bark, leaves, flower buds, flowers and fruits of *S. japonicum* revealed that the highest rutin content (24–37%) was found in young flower buds and the lowest (1.5–3%) in the bark. In view of the total volume and weight of harvestable plant parts it is recommended to use the flower buds, flowers and young fruits as a source of rutin. In the temperate zones *S. japonicum* is considered fast-growing.

**Handling after harvest** After harvesting, the inflorescences of *S. japonicum* are dried, and

rachises and pedicels discarded. Rutin is extracted by subsequent boiling in water and crystallization upon cooling, followed by recrystallization from ethanol.

**Genetic resources and breeding** Various ornamental cultivars of *S. japonicum* are available, mainly in western Europe, e.g. weeping, dwarf and variegated forms. Breeding efforts should be directed towards plants flowering at a relatively early age.

**Prospects** *S. japonicum* is an important industrial source of rutin. Research in further applications in the treatment of capillary and venous disorders deserves more attention.

**Literature** |1| Balbaa, S.I., Zaki, A.Y. & El-Shamy, A.M., 1974. Qualitative and quantitative study of the flavonoid content of different organs of *Sophora japonica* at different stages of development. *Planta Medica* 25: 325-330. |2| Doan Thi Nhu, Nguyen Thuong Thuc, Do Huy Bich & Vu Thuy Huyen (Editors), 1991. Les plantes médicinales au Vietnam (livre 1) [The medicinal plants of Vietnam (volume 1)]. *Médecine traditionnelle et pharmacopée*. Agence de coopération Culturelle et Technique, Paris, France. p. 76. |3| González, G., Alzuetta, C., Barro, C. & Salvador, A., 1988. Yield and composition of protein concentrate, press cake, green juice and solubles concentrate from wet fractionation of *Sophora japonica* L. foliage. *Animal Feed Science and Technology* 20: 177-188. |4| Ishida, H., Umino, T., Tsuji, K. & Kosuge, T., 1989. Studies on the antihemostatic substances in herbs classified as hemostatics in traditional Chinese medicine. I. On the antihemostatic principles in *Sophora japonica* L. *Chemical and Pharmaceutical Bulletin* 37(6): 1616-1618. |5| Palomino, G., Martínez, P., Bernal, C. & Sousa, S.M., 1993. Diferencias cromosómicas entre algunas especies de los generos *Sophora* L. y *Styphnolobium* Schott [Chromosomal differences between some species of the genera *Sophora* L. and *Styphnolobium* Schott]. *Annals of the Missouri Botanic Garden* 80: 284-290. |6| Perry, L.M., 1980. Medicinal plants of East and Southeast Asia. Attributed properties and uses. The MIT Press, Cambridge, Massachusetts, United States & London, United Kingdom. pp. 226-227. |7| Poretz, R.D. & Barth, R.F., 1976. Studies on the interaction of the *Sophora japonica* lectin and concanavalin A with erythrocytes and lymphocytes. *Immunology* 31(2): 187-194. |8| Sangat-Roemantyo, H. & Wirdateti, 1991. *Sophora japonica* L. In: Lemmens, R.H.M.J. & Wulijarni-Soetjipto, N. (Editors): *Plant Resources of South-East Asia No 3. Dye and tan-*

*nin*-producing plants. Pudoc, Wageningen, the Netherlands. pp. 113-115. |9| Sousa, S.M. & Rudd, V.E., 1993. Revision del genero *Styphnolobium* (Leguminosae: Papilionoideae: Sophoreae) [Revision of the genus *Styphnolobium* (Leguminosae: Papilionoideae: Sophoreae)]. *Annals of the Missouri Botanic Garden* 80: 270-283. |10| Ueno, M., Ogawa, H., Matsumoto, I. & Seno, N., 1991. A novel mannose-specific and sugar aggregatable lectin from the bark of the Japanese pagoda tree (*Sophora japonica*). *Journal of Biological Chemistry* 266: 3146-3153.

**Other selected sources** 193, 202, 287, 360, 710, 806, 871, 997, 1035, 1036, 1045, 1128, 1277, 1283, 1468, 1480, 1554, 1634.

Tran Cong Khanh

### **Taraxacum officinale Weber ex F.H. Wigg.**

Prim. fl. holsat.: 56 (1780).

COMPOSITAE

$2n = 16, 24, 32, 40, 48$

**Vernacular names** Dandelion (En). Dent de lion, pissenlit (Fr). Indonesia: jombang (Java). Vietnam: b[oof] c[oo]ng anh.

**Origin and geographic distribution** Dandelion is native to Europe and continental temperate Asia south to the Himalayas, but now distributed almost all over the world. In the Malesian region it has been introduced and naturalized in Peninsular Malaysia, West Java and the Philippines (Benguet Province). It is sometimes cultivated as a vegetable or for medicinal applications.

**Uses** Infusions or decoctions of dried roots, leaves or simply the entire plant of dandelion are widely used as a general tonic, anti-inflammatory, depurative, cholagogue, diuretic, mild laxative, and for kidney and liver disorders. Infusions are also recommended in the treatment of skin problems, such as acne, eczema, psoriasis and even for arthritic and rheumatic complaints. Externally the latex is applied to boils and other skin infections or applied as a poultice on inflamed wounds. In South-East Asia, dandelion is a fairly recently introduced weed, so traditional uses are very limited. In Indo-China it is used as a diuretic and cholagogue. In India, the roots are applied as a tonic, diuretic, mild laxative, and chiefly used in kidney and liver disorders. In China, the leaves are prescribed internally as a bitter depurant, in the treatment of breast and lung tumours, mastitis, abscesses, jaundice, and urinary tract infec-

tions; and externally to treat snake bites. In Europe and North America, the leaves and roots, fresh or dried, are used as a mild laxative, a diuretic and for the treatment of high blood pressure by reducing the volume of fluid in the body. The roots accelerate steady elimination of toxins, by working principally on liver and gall bladder to help remove waste products, and simultaneously stimulating the kidneys to remove toxins in the urine. The leaves or roots may also help to prevent or even dissolve gallstones. A decoction of the roots is used as an antidiabetic.

The leaves are also eaten as a vegetable. When grown without light (artificially or when covered with earth) the pale leaves are more brittle and taste better. The young and unopened flower heads can be used as capers. In North Africa the leaves are used as a seasoning. The bitter leaves are also applied in wines, beers and non-alcoholic drinks. The ground roots are used as a substitute for coffee. In spring the flowers contain much nectar and are locally important for the production of honey. Formerly, dandelions were cultivated in Japan for ornamental purposes.

**Production and international trade** In Germany, France and the United States dandelion is comparatively often cultivated as a vegetable. It is also cultivated in India, where it is a popular remedy for liver complaints. However, no statistics are available on production and trade.

**Properties** Phytochemical analysis has revealed chicoric acid and monocaffeoyltartaric acid to be the major phenolic constituents of flowers, roots, leaves and involucre bracts of *T. officinale*. These compounds are also the main phenolic constituents of some common dandelion preparations, e.g. dandelion tea, root coffee and root capsules. Furthermore, the presence of sesquiterpene lactones (germacranolide type, as glucosides), triterpenes (e.g. cycloartenol) and flavonoids (apigenin-7-glucoside, luteolin-7-glucoside) in the leaves is reported in literature.

As with many *Compositae*, the roots of dandelion also have a high content of inulin, a polysaccharide based on fructose. This compound serves as a food reserve and can reach levels as high as 25% in autumn in the temperate zones. Inulin can be used as a sugar substitute, which is of interest for diabetic patients. Furthermore, the hypoglycaemic activity of this compound is sometimes mentioned in literature, though many reports are not conclusive in this respect. On the other hand, hypoglycaemic activity of dandelion preparations have been observed in various animal models: the

50% ethanol-water extract of the entire plant at a dose of 250 mg/kg orally in rats, dried entire plants at doses of 1-2 g/kg administered intragastrically to rabbits and a water extract of dried roots at a dose of 25 mg/kg administered intragastrically to mice all showed hypoglycaemic activities.

The high potassium content of dandelion, especially in the leaves (up to 4.5% of the dry weight) is considered to be responsible for the well-known diuretic activity, which has been confirmed in various animal models. The ethanol (30%) extract of dandelion roots, administered orally at a dose of 0.1 ml/kg in male rats, shows diuretic activity. In experiments with mice and rats the diuretic and saluretic indices of a fluid extract of dandelion, corresponding to approximately 8 g dried aerial parts/kg body weight, were comparable to those of furosemide (80 mg/kg body weight) a well-known diuretic. The high potassium content ensures that potassium eliminated in the urine is replaced. Furthermore, in dogs, the volume of bile doubled when a decoction of fresh leaves (equivalent to 5 g of dried plant material) was administered intravenously. In rats, a choleric effect was observed following administration of a 5% dandelion extract (2 ml) by means of a cannula, and in another experiment, an alcoholic extract of the whole plant administered to rats gave a 40% increase in bile secretion.

The anti-inflammatory activity of dandelion has been investigated in several animal models. A methanol extract of dandelion leaves, at a dose of 2.0 mg/ear applied externally, reduced swelling and inflammation in mice with 12-*O*-tetradecanoylphorbol-13-acetate (TPA)-induced ear inflammations. Furthermore, a 95% ethanol extract of dried dandelion leaves, administered intraperitoneally in rats with carrageenan-induced pedal oedema at a dose of 0.1 g/kg showed anti-inflammatory activity. Finally, the 80% ethanol extract of dried dandelion roots administered by gastric intubation at a dose of 100 mg/kg in male rats showed 25% inhibition of oedema in a carrageenan-induced pedal oedema model, in comparison with 45% inhibition of indomethacin (5 mg/kg) in the same experiment.

Other pharmacological activities include: a dose-dependent inhibition of the ADP-induced aggregation of human platelets by an ethanolic extract of the roots of *T. officinale*. A bioguided purification revealed activity in two fractions: one containing low-molecular weight polysaccharides, and one fraction characterized by the presence of triter-

penes/steroids. A 95% ethanol extract of dried dandelion leaves administered to mice intragastrically at a dose of 1.0 g/kg, and intraperitoneally at a dose of 0.1 g/kg, exhibited analgesic activity in both the phenylquinone-induced writhing and the hot plate models. The water extract of dandelion roots administered intragastrically at a dose of 2 g/kg in rats with ETOH-HCl-induced ulcerations showed a strong anti-ulcer activity. However, the methanol extract at the same dose shows only weak activity. The hot water extract of dried dandelion aerial parts given intraperitoneally at doses of 30–40 mg/kg exhibited antitumour activities against CA-C3H/HE-MM46 and fibrosarcoma METH-1 in mice.

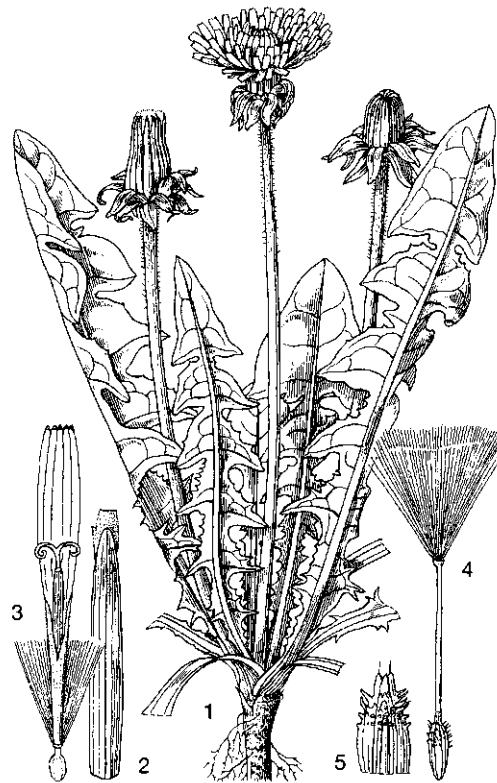
The 70% ethanol extract of dried dandelion aerial parts at concentrations of 2, 10 and 50% showed acaricide activity against *Tetranychus urticae*, with percentages of inhibition being respectively 57, 90 and 100%. Both ether and methanol extracts of fresh aerial parts used at undiluted concentrations in the disk method showed antifungal activity against *Aspergillus flavus*. Furthermore, the tincture (10%) of dried leaves at a concentration of 0.1 ml/disk showed antibacterial activity against *Bacillus subtilis*.

Lettucenin A is a sesquiterpenoid phytoalexin produced by a number of *Compositae* including *T. officinale* to protect themselves against microorganisms. In a liquid cell culture 16.4 µg/g fresh weight inhibits the growth of the fungus *Cladosporium herbarum*; the lettukenin A production peaked only 2–6 hours after infection.

An analysis of leaves and flowers from Pakistan gave per 100 g edible portion: water 88.8 g, protein 3.6 g, ether extracts 1.6 g, total carbohydrates 3.7 g, fibre 0.4 g, and ash 2.3 g, phosphorus 59 mg, calcium 474 mg, iron 3.3 mg and vitamin C 73 mg. In vitro dry-matter digestibility is about 80%. The latex from dandelion may cause skin irritation. The sugar content of the nectar is around 50%. The sugars are mainly glucose and fructose, and a smaller proportion of sucrose. The pollen contains about 15% of protein.

**Adulterations and substitutes** Roots of *Cichorium intybus* L. are used as a substitute for the roots of dandelion. They are also rich in inulin, and bitter due to sesquiterpenoid lactones.

**Description** A perennial, stemless herb up to 30(–50) cm tall, with a long taproot and latex in all parts. Leaves arranged spirally in a radical rosette, oblanceolate to narrowly spatulate, 4–35 cm × 0.7–10 cm, very variably and irregularly pinnatolobed to pinnatipartite, variably hairy or



*Taraxacum officinale* Weber ex F.H. Wigg. – 1, plant habit; 2, inner involucre bract; 3, flower; 4, fruit; 5, upper part of achene.

rarely completely glabrous, almost distinctly petiolate or narrowly tapering into a winged petiole, petiole green or pink to purplish. Inflorescence an axillary head, 1–25 per plant, peduncle simple, hollow, leafless, (3–)3.5–5(–6.5) cm in diameter, outer involucre bracts many-seriate, patent to recurved, ovate to linear-lanceolate, unequal, without 'horns' (thickened and/or clawed apices), inner involucre bracts 1-seriate, erect, oblong, receptacle flat, naked. Flowers many, all ligulate; corolla yellow, but often with a purple line outside; stamens 5, anthers fused into a tube, sagittate at base; ovary inferior, with a single ovule; style 1, greenish or yellowish to black, stigmas 2, spreading. Fruit an achene, narrowly obovoid, about 3 mm long, ribbed, greenish to straw-coloured or brownish, the upper third minutely spiny, abruptly contracted into a 6–12 mm long beak which is crowned by spreading, scabrid, white pappus hairs. Seedling with epigeal germination; cotyledons free, leafy, obspathulate, sheathed at base; epicotyl absent; all leaves alternate.

**Growth and development** Both sexually and asexually reproducing populations of *T. officinale* exist. The sexually reproducing plants are generally obligate cross-pollinators, although exceptions have been reported. The self-compatibility is hereditary. The asexual reproduction is known as agamospermy, and functions only in polyploid plants. It may be obligatory or facultative, even within a single head.

The flowers produce nectar and pollen that attract insects, mainly bees, which pollinate the flowers. Most of the orange-coloured pollen is released between 10–12 h in the morning, and some 20–75% is sterile. Wind can disperse the seeds over long distances by means of their 'parachute'-like papus.

**Other botanical information** *Taraxacum* is closely related to the genus *Crepis*, and belongs to the tribe *Lactuceae*. It has been subdivided into some 40 sections. *T. officinale* sensu lato forms a large and highly variable polyploid complex. The agamospermous reproduction results in a high number of uniform 'clones', which have often been described as microspecies or agamospecies. *T. officinale* sensu stricto belongs to section *Ruderalia* J. Kirschn., H. Ollg. & Stepanek (synonym: section *Vulgaria* Dahlst. p.p.) with about 1000 microspecies. Some authors prefer to distinguish many taxa at subspecific rank while recognizing only a few species within *Taraxacum*. The section *Ruderalia* is likely to be of recent origin, as it harbours the more advanced microspecies. These are often polyploid and tend to possess more satellite chromosomes than generally diploid primitive microspecies. The length of the beak on the achene generally increases with increasing ploidy level.

The microspecies *T. javanicum* v. Soest has been described from Java, and belongs to *T. officinale* sensu lato. Recently, another microspecies, *T. indonesicum* Sonck has been described from West Java. Although the two are clearly distinct to specialists, it is likely that both are used in a similar way by local people.

**Ecology** Being weedy in nature, *T. officinale* is most often found in ruderal places, along roads and fields and in grassland. In tropical regions it occurs only at higher elevations, in Malesia at 1200–1500 m altitude. It occurs on various soils, from sandy dunes to thick clay, and from dry to wet, sometimes even brackish habitats, though it seems to grow best on fertile sandy or loamy soils.

**Propagation and planting** Propagation of dandelion is by seed or by division. In Europe about 60% of all achenes germinate in the year of

production; about 30% in the next year and about 5% in the year thereafter. A neglectable percentage germinates after 5 years. The viability of fresh achenes is 70–100%, but drops rapidly when stored dry at 20°C; cool and dry storage does not cause a rapid decrease in viability. Achenes germinate best at temperatures of 20–25°C, with a daily fluctuation of about 5°C. They should not be sown deeper than 1 cm as this will affect fast and uniform emergence.

**In vitro production of active compounds** Undifferentiated cultured cells of dandelion produce oleanolic and ursolic acids as major triterpenoids, in addition to triterpenols composed mainly of  $\alpha$ - and  $\beta$ -amyrins. Regenerated and wild plants contain additional triterpenols (taraxasterol and lupeol), but negligible quantities of triterpene acids. Squalene synthase activity has been detected in the microsomal fractions of suspension-cultured cells of *T. officinale*, which produce cycloartane (involved in phytosterol biosynthesis) and other triterpenoids e.g. oleanane and ursane.

**Husbandry** Injured roots or small parts of roots of dandelion can regenerate and develop new rosettes. Therefore mechanical control of *T. officinale* as a weed is not effective. In Canada, ethalfluralin proved an effective herbicide for dandelion cultivation with no residues in the roots.

**Diseases and pests** Diseases observed in dandelion in the temperate zones include *Agrobacterium tumefaciens*, and fungi like *Synchytrium taraxaci*, *Bremia lactucae*, *Protomyces pachydermus*, *Sphaerotheca fuliginea*, *Puccinia* spp., *Ramularia taraxaci* and *Septoria taraxaci*. Pests include the nematodes *Ditylenchus dipsaci* and *Meloidogyne hapla* and the beetle *Ceutorhynchus punctiger*, whereas various other insects, spiders, snails, birds and mammals feed on dandelion in some way or another.

**Harvesting** Dandelion roots are harvested at the end of the growing season, when inulin contents are highest.

**Yield** In India, cultivated dandelion yields about 1650 kg of roots per ha. In Canada, the dry matter production of roots was about 2300 kg/ha, i.e. 19% higher when planted at a high density of over 114 000 plants/ha compared to a lower density of about 89 000 plants/ha. Average root production per dandelion plant with the flower buds removed was 40 g when grown in a nutrient solution and 30 g when grown on a peat substrate with organic fertilizer.

**Handling after harvest** In India, roots of dan-



dellion are washed, dried and subsequently stored in containers to which a few drops of carbon tetrachloride have been added as preservative.

**Genetic resources and breeding** *T. officinale* is such a widespread weed that its genetic basis does not seem to be at risk; this is not the case for some of its microspecies. Breeding efforts have been directed to a more palatable crispy vegetable rather than to its medicinal properties.

**Prospects** As a result of the quite well investigated diuretic properties, dandelion preparations could be used as an adjunct to treatments where enhanced urinary output is desirable, for example, the prevention of renal gravel or frequently returning uncomplicated urinary tract infections. Other pharmacological actions (e.g. on the biliary excretion) are also interesting, but merit further research.

**Literature** |1| Akashi, T., Furuno, T., Takahashi, T. & Ayabe, S., 1994. Biosynthesis of triterpenoids in cultured cells, and regenerated and wild plant organs of *Taraxacum officinale*. *Phytochemistry* 36(2): 303-308. |2| Baba, K., Abe, S. & Mizuno, D., 1981. Antitumor activity of hot water extract dandelion, *Taraxacum officinale* - Correlation between antitumor activity and timing of administration. *Yakugaku Zasshi* 10(1): 538-543. |3| Doll, R., 1982. Grundriss der Evolution der Gattung *Taraxacum* Linn. [Review of the evolution of the genus *Taraxacum* Linn.]. *Feddes Repertorium* 93: 481-624. |4| Muto, Y., et al., 1994. Studies on antiulcer agents. 1. The effects of various methanol and aqueous extracts of crude drugs on antiulcer activity. *Yakugaku Zasshi* 114(2): 980-994. |5| Rácz-Kotilla, E., Rácz, G. & Solomon, A., 1974. The action of *T. officinale* extracts on the body weight and diuresis of laboratory animals. *Planta Medica* 25: 212-217. |6| Richards, A.J., 1973. The origin of *Taraxacum* agamospecies. *Botanical Journal of the Linnean Society* 66: 189-211. |7| Sterk, A.A., 1987. *Paardebloemen: planten zonder vader* [Dandelions: plants without a father]. Koninklijke Nederlandse Natuurhistorische Vereniging, Utrecht, the Netherlands. 348 pp. |8| Swanston-Flatt, S.K., Day, C., Flatt, P.R., Gould, B.J. & Bailey, C.J., 1989. Glycaemic effects of traditional European plant treatments for diabetes. Studies in normal and streptozotocin diabetic mice. *Diabetes Research* 10(2): 69-73. |9| Tita, B. et al., 1993. *Taraxacum officinale* W.: Pharmacological effect of ethanol extract. *Pharmacology Research* 27(1): 23-24. |10| Yasukawa, K., Yamaguchi, A., Arita, J., Sakurai, S., Ikeda, A. & Takido, M., 1993. Inhibitory effect of edible

plant extracts on 12-0-tetradecanoylphorbol-13-acetate-induced ear oedema in mice. *Phytotherapy Research* 7(2): 185-189.

**Other selected sources** 41, 97, 184, 193, 202, 207, 287, 349, 365, 378, 400, 542, 571, 678, 740, 762, 844, 845, 852, 910, 945, 961, 989, 1018, 1019, 1035, 1066, 1126, 1178, 1225, 1306, 1356, 1391, 1577.

Wongsatit Chuakul

## Tinospora Miers

*Ann. Mag. Nat. Hist. ser. 2, 7: 35 (1851).*

MENISPERMACEAE

$x =$  unknown; *T. cordifolia*:  $2n = 24, 26$

**Major species** *Tinospora crispa* (L.) Hook.f. & Thomson, *T. glabra* (Burm.f.) Merr.

**Vernacular names** Philippines: makabuhay (general).

**Origin and geographic distribution** *Tinospora* consists of 33 species, which occur in the tropical and subtropical parts of the Old World: 7 species in tropical Africa, 2 in Madagascar, and 24 in Asia extending to Australia and the Pacific. In Malesia, 14 species have been found, but most of them are known from few collections; only *T. crispa* and *T. glabra* are widespread. *T. crispa* is also cultivated as a medicinal plant, e.g. in Thailand, Sri Lanka and India.

**Uses** Throughout most of South-East Asia, *Tinospora* species are a widely acclaimed source of remedies for many different complaints. Much of their reputation is probably due to influences from Chinese traditional medicine, where for instance stems and leaves of *T. crispa* are valued for their anti-inflammatory (antirheumatic), febrifugal, antimalarial and antibacterial properties. In Indonesia, Malaysia, Thailand and the Philippines, stems (infusions, decoctions) of *T. crispa* are considered to be effective in the treatment of skin complaints (external as antiseptic, antiparasitic, and for treating wounds and itches), stomach complaints (ulcers, as appetizer, tonic), diarrhoea, fevers (e.g. malaria, smallpox), diabetes mellitus, cholera (whole plant), jaundice, and as a vermifuge (also in children). In veterinary medicine, *T. crispa* is applied as veterinary tonic; powdered stems are used to fatten horses and cattle by stimulating their appetite.

*T. glabra* has similar uses as *T. crispa* although the latter is thought to be more effective, but is recommended particularly for dealing with wounds, scabies and tropical ulcers.

It is perhaps in the Indian subcontinent where the widest use is made of *Tinospora*. The main species is *T. cordifolia* (Willd.) Miers, which does not occur in Malasia. In present-day Ayurvedic medicine in India, all parts of the plant find wide use for their general tonic, anti-inflammatory, anti-arthritic, anti-allergic, antimalarial, antidiabetic, hepato-protective and aphrodisiac properties. Categorized as 'rasayana' in Ayurveda, it is also used for its general adaptogenic and pro-host immunomodulatory activity in fighting infections. *T. cordifolia* is thus claimed to be useful in e.g. skin diseases, jaundice, diabetes, anaemia, fever and rheumatism. In fact the plant is part of almost all decoctions mentioned in Ayurvedic textbooks for use in joint diseases. The starch from the stems and roots is used as a nutrient in chronic diarrhoea and dysentery. Juice of the fresh plant is a powerful diuretic (urinary diseases), and also used in gonorrhoea with advantage. Besides its anti-malarial activity, the root is known for its anti-stress and antileprotic activities. Finally, *T. cordifolia* is also used as veterinary medicine.

A tincture from the stem of *T. sinensis* (Lour.) Merr. (synonyms: *T. malabarica* (Lamk.) Hook.f. & Thomson, *T. tomentosa* (Colebr.) Hook.f. & Thomson) has considerable reputation in Indo-China for treating arthritis and chronic rheumatism. There are also reports suggesting the efficacy of the roots, stems and leaves in conditions like cough, wound healing, malaria, skin complaints and allergic disorders.

**Production and international trade** Stem parts or powdered stems can be found frequently on local markets, but are not traded internationally.

**Properties** A number of chemical constituents have already been isolated from different parts of *T. crispa*, e.g. diterpenes, alkaloids and flavonoids. Most extensively investigated are a series of furano-diterpene (glycoside) compounds: tinotufolin A-F (leaves), borapetoside A-H (glycosides, stems), borapetol A, B (stems, aglycones of borapetoside A, B) and the bitter tinocrisposide (glycoside, stems). In addition to these furano-diterpenes, a series of clerodane-diterpene glycosides has also been isolated from the stems and named rumphioside A, B, C, C-1, Ac-D, E and F.

Amongst the alkaloids isolated from *T. crispa* are the well known protoberberine type alkaloids palmatine (stems), berberine (stems, aerial roots) and the aporphine type alkaloid tembetarine (stems, aerial roots). From the methanolic stem extract three further N-acyl-aporphine type alka-

loids have been isolated and their structures elucidated: (-)-N-formyl-annonaine, (-)-N-formyl-nornuciferine (= tinocrispicine) and (-)-N-acetyl-nornuciferine. Of these isolated alkaloids, berberine and its salts are known to have spasmolytic, antibacterial and in some degree antifungal and anti-protozoal activity.

Further constituents from *T. crispa* include: N-trans and N-cis feruloyl tyramine (stems, phenolic acid amides), tinotuberide (stems, phenolic glucoside), and the flavone-O-glycosides (from stems): luteolin-4'-methylether-7-glucoside, genkwanin-7-glucoside, luteoline-4'-methylether-3'-glucoside, diosmetin and genkwanin.

An aqueous extract from *T. crispa* stems showed lowering of blood glucose levels and stimulated insulin release in moderate alloxan-diabetic rats. The dose administered orally (4 g/l in drinking water) and the method of preparation of the extract were comparable to those used by diabetics in Malaysia. After two weeks of treatment, the rats also showed an improvement in the glucose tolerance test; there were no effects in normal and severe alloxan-diabetic rats. These results suggest that the hypoglycaemic effect observed is due to stimulation of insulin release, rather than some extra-pancreatic action. This insulinotropic activity was also observed after intravenous injection of 50 mg/kg of the extract in normal rats.

Toxicity of *Tinospora* extracts proved to be low in tests with rats (LD<sub>50</sub> values of over 5 g/kg orally, 3 g/kg dermally). *Tinospora* extracts in high concentrations can cause infection in the liver and follicular atresia in mice, but the kidneys are not much affected. Crude extracts showed protective effects in mice with experimental urinary tract infection caused by *Pseudomonas aeruginosa*.

A number of compounds have been isolated from the stems of *T. cordifolia*, including diterpenes and alkaloids. The diterpenes comprise the major group of components isolated: the norditerpene-furan-glycosides cordifoliside A-E, the clerodane-diterpenes tinosponone, tinosporaside (= tinosponone-glycoside) and tinocordioside (glycoside) and the clerodane-furano-diterpenes cordioside (glycoside), together with 4 other furano-diterpene compounds: colombin, one being an epimer of 6-hydroxy-arcangelisin, and two being a set of optical isomers (not yet named). The biosynthesis of the clerodane-furano-diterpene skeleton has also been investigated.

Amongst the alkaloids isolated from *T. cordifolia* are magnoflorine and tembetarine (stems, aporphine type alkaloids) and jatrorrhizine (roots, pro-

toberberine type alkaloid). Syringin is a phenolic component, isolated from the fresh stems. Considerable efforts have been made to investigate the biological effects of *T. cordifolia*. An aqueous, alcoholic and chloroform extract of the leaves after oral application exerted a significant hypoglycaemic effect in both normal and alloxan-diabetic rabbits. The reaction in alloxan-diabetic animals, in which almost all pancreatic  $\beta$ -cells have been destroyed, appears to be like a direct effect, probably by a mechanism similar to insulin. However, the stronger effect in normo-glycaemic rabbits suggests that the mechanism of action consists not only of this direct insulin-like effect, but that there is also an indirect action by stimulating the insulin release from pancreatic  $\beta$ -cells. Besides the hypoglycaemic activity, no effects were found on the blood lipid levels, and acute toxicity studies did not reveal visible signs and symptoms of toxicity. Further investigations in albino rats and different groups of rabbits confirm the hypoglycaemic effect and the proposed mechanism of action.

Several studies have focused on the immunotherapeutic effects. Pretreatment (oral) with an aqueous extract strongly reduced the mortality in a mouse model for *E. coli* peritonitis. This was associated with significantly improved bacterial clearance and improved phagocytic capacities of neutrophils in the group treated with *T. cordifolia*. *T. cordifolia* extract itself did not possess in vitro bactericidal activity. These results were confirmed in experiments with wistar rats in which abdominal sepsis was induced by caecal ligation. Both in rat and man, oral application of a *T. cordifolia* extract improved the surgical outcome in patients with obstructive jaundice, in which sepsis initiated by bacteria in the bile at the time of biliary tract surgery comprises a major risk. In the group receiving the extract, the phagocytic and killing capacities of neutrophils improved; thus *T. cordifolia* appears to act by strengthening host defences.

Further investigations included antileishmanial, anti-stress, antipyretic and diuretic effects. Leishmaniasis, commonly known as 'kala-azar', causes great mortality in tropical and subtropical regions of the world. The antileishmanial effect was evaluated in infected golden hamsters. A 50% ethanol extract of *T. cordifolia* stems showed significant inhibition of multiplication of parasites, and increased survival periods. The anti-stress activity was studied by investigating the effects of treatment on brain norepinephrine, dopamine and

serotonine levels in stressed rats. The ethanol extract, prepared from roots previously extracted with petrol (60-80) and chloroform, and given orally after drying, was found to possess normalizing activity against stress-induced changes on brain neurotransmitter levels. The antipyretic effect was investigated in rats, and yeast-induced pyrexia was used to screen the extracts. The antipyretic action was clearly significant for a *T. cordifolia* ethanol extract (whole plant, given orally) and appeared to be comparable to that of the aspirin control. Finally, *T. cordifolia* extract showed mild diuresis and a significant increase in the excretion of electrolytes in rats after oral application.

*Tinospora* extracts are toxic to brown planthoppers (*Nilaparvata lugens*) and green leafhoppers (*Nephotettis virescens*), which are common pests in rice in the Philippines. Soaking the roots of rice seedlings in aqueous *Tinospora* extract in the field is effective in controlling the major rice pests, and broadcasting the ground stems on the seed-beds 10 days after sowing is also applied successfully. The extracts can control the diamondback moth (*Plutella xylostella*); the effectivity is comparable to malathion, and the extracts show antifeedant effects on the insect. In tests in Thailand, *T. crispa* extract showed mild repellency to the oviposition of the oriental fruit fly (*Dacus dorsalis*). It is also effective in controlling borers on maize, and bollworms (*Helicoverpa armigera*) in cotton. The extract showed high toxicity to fish in experiments in Malaysia. In vitro tests in the Philippines using mutant strains of *Bacillus subtilis* and *Salmonella typhimurium* showed that *Tinospora* extracts do not contain direct mutagens.

**Adulterations and substitutes** Other *Menispermaceae* (e.g. *Cyclea* and *Stephania* species) have similar or related alkaloids and have similar applications. Other insecticides of plant origin used in South-East Asia are present in the roots of *Derris* and *Lonchocarpus* spp., in seeds of *Croton tiglium* L., in leaves of *Azadirachta indica* A.H.L. Juss., *Nicotiana tabacum* L. and *Vitex negundo* L., in leaves, fruits and bark of *Melia azedarach* L., in whole plants of *Tanacetum cinerariifolium* (Trev.) Schultz-Bip., *Lantana* spp. and *Tagetes* spp., and in leaves, roots and seeds of *Annona squamosa* L.

**Description** Dioecious woody climbers up to 15 m long, sometimes scandent shrubs, usually entirely glabrous; stem woody, with bark often becoming detached on drying, striate when young, becoming tuberculate or warty with raised lenticels, usually glabrous but sometimes puberu-

lous, sometimes producing very long filiform aerial roots. Leaves arranged spirally, simple and entire (occasionally dentate or 3-lobed), often cordate, palmately veined, sometimes domatia present in axils of veins beneath, petiole swollen and geniculate at base; stipules absent. Inflorescence axillary or cauliflorous, thyrsoid, pseudopaniculate, pseudoracemose or pseudospicate. Flowers unisexual, 6-merous; sepals usually free, outer 3 usually smaller, elliptical, imbricate; petals free, often broadly cuneate-ovate with the lateral edges inrolled, usually fleshy and often glandular-papillose externally towards the base; male flowers with 6 free stamens; female flowers with 3 curved-ellipsoid carpels having short-lobed, reflexed stigmas, and 6 subulate staminodes. Fruit a usually ellipsoidal drupe with terminal style scar, borne on a short or columnar carpophore; endocarp bony, dorsally convex and often verrucose or tuberculate, ventrally with central aperture or with shallow longitudinal groove. Seed with usually ruminate endosperm.

**Growth and development** *T. crispa* usually flowers when leafless, in Thailand in January to March. The scented flowers are pollinated by insects such as small flies and bees, and possibly also small beetles and moths. Female plants, with or without fruits, have rarely been collected.

**Other botanical information** *Tinospora* belongs to the tribe *Tinosporeae*, characterized by the drupe with terminal style scar, and embryo with foliaceous cotyledons, together with *Chlaenandra*, *Fibraurea*, *Parabaena* and *Tinomiscium*. *T. glabra* has been much confused with *T. crispa*. Information on medicinal and chemical properties of *T. crispa* (or its synonym *T. rumphii*) given in literature for the Philippines partly refers to *T. glabra*.

**Ecology** *Tinospora* occurs in forest, thickets and hedges up to 1000 m altitude. It has a very strong capacity of regeneration and is often very abundant in secondary regrowth after disturbance of the natural forest. In primary forest it occurs much more scattered. *T. glabra* can also be common in littoral forest and on limestone.

**Propagation and planting** Stems of *T. crispa* remain viable when cut into pieces because the dried sap effectively seals the cut ends. If kept in a closed box they can still be viable after one year. Apparently *T. crispa* is mainly propagated vegetatively.

**In vitro production of active compounds** Experiments on the in vitro culture of *Tinospora* root segments showed promising results, with

good callus formation and a high content of therapeutic substances.

**Diseases and pests** Larvae of the noctuid moth *Othreis fullonia* feed mainly on *Tinospora* leaves. The adult moth can cause considerable damage to commercial fruit tree plantations, especially of longan (*Dimocarpus longan* Lour.) and citrus in Thailand, by piercing the skin of the fruits. The destruction of natural forest and the increasing area of secondary vegetation, preferred by *Tinospora*, promote the spread of the pest.

In Pakistan, *T. cordifolia* showed effective resistance to the root-knot nematodes *Meloidogyne incognita* and *M. javanica*.

**Handling after harvest** Under ambient and refrigerated conditions stored crude stem extracts maintain their pesticidal activity during 3 months.

**Genetic resources and breeding** *T. crispa* and *T. glabra* are widely distributed and will not easily become endangered, since they prefer sites with secondary vegetation and show strong regeneration capacity. In India, *T. cordifolia* is a popular medicinal plant and is locally endangered because of extensive collecting from the wild.

**Prospects** *T. crispa* and the non-Malesian *T. cordifolia* are well investigated medicinal plants with numerous interesting properties which have been confirmed by modern research. It is expected that they will keep their important role in local medicine in South-East Asia, whereas they have good prospects to play a more prominent role in modern medicines in other parts of the world. The need for safer pesticides lead to a revival of interest in pesticides of plant origin which can be used in integrated pest management. In the Philippines, *Tinospora* extracts have been evaluated in the field and proved effective.

**Literature** [1] Acevedo, R.A., Santos, A.C. & Pabatao, P., 1970. A diterpene from *Tinospora rumphii* Boerl. Philippine Journal of Science 97(3): 269-275. [2] Bänziger, H., 1982. Fruit-piercing moths (Lep., Noctuidae) in Thailand: a general survey and some new perspectives. Mitteilungen der Schweizerischen Entomologischen Gesellschaft 55(3-4): 213-240. [3] Comley, J.C.W., Titanji, V.P.K., Ayafor, J.F. & Singh, V.K., 1990. In vitro antifilarial activity of some medicinal plants. Acta Leidensia 59(1-2): 361-363. [4] del Fierro, R.S., 1983. Studies on the mutagenicity, clastogenicity and antimutagenicity potential of *Tinospora rumphii* Boerlage (Family Menispermaceae). Philippine Scientist 20: 1-10. [5] Forman, L.L., 1986. Menispermaceae. In: van Steenis,

C.G.G.J. & de Wilde, W.J.J.O. (General editors): Flora Malesiana. Series 1, Vol. 10. Kluwer Academic Publishers, Dordrecht, the Netherlands. pp. 188–201. |6| Forman, L.L., 1991. Menispermaceae. In: Smitinand, T. & Larsen, K. (Editors): Flora of Thailand. Vol. 5(3). The Forest Herbarium, Royal Forest Department, Bangkok, Thailand. pp. 359–365. |7| Noor, H. & Ashcroft, S.J.H., 1989. Antidiabetic effects of *Tinospora crispa* in rats. Journal of Ethnopharmacology 27(1–2): 149–161. |8| Noor, H., Hammonds, P., Sutton, R. & Ashcroft, S.J.H., 1989. The hypoglycemic and insulinotropic activity of *Tinospora crispa* studies with human and rat islets and HIT-T15 B cells. Diabetologia 32(6): 354–359. |9| Pachaly, P. & Adnan, A.Z., 1989. The structure of tinocrisposide, a bitter furanoditerpene glucoside from *Tinospora crispa*. Planta Medica 55(7): 632. |10| Wadood, N., Wadood, A. & Shah, S.A.W., 1992. Effects of *Tinospora cordifolia* on blood glucose and total lipid levels of normal and alloxan-diabetic rabbits. Planta Medica 58(2): 131–136.

#### Selection of species

### *Tinospora crispa* (L.) Hook.f. & Thomson

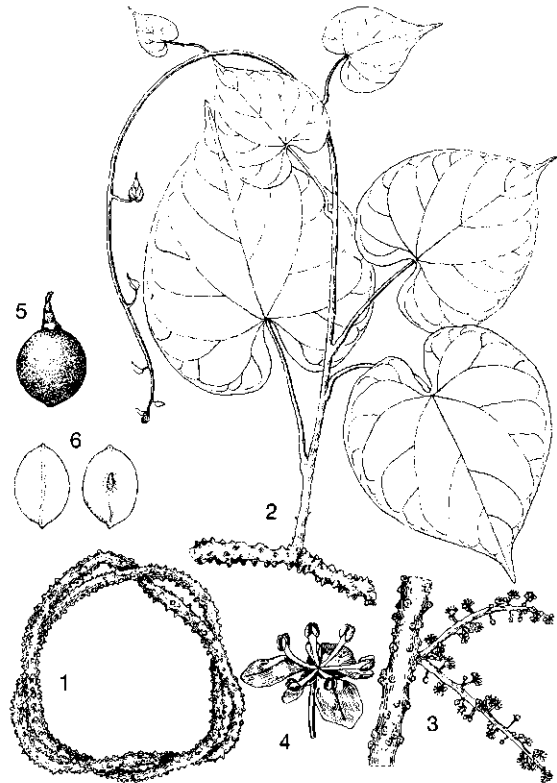
Fl. ind. 1: 183 (1855).

**Synonyms** *Tinospora rumphii* Boerl. (1901), *Tinospora tuberculata* (Lamk) Beumée ex K. Heyne (1927).

**Vernacular names** Indonesia: andawali (Sundanese), brotowali, putrowali (Javanese). Malaysia: akar putarwali, petawali (Peninsular), daun akar wali (Sarawak). Philippines: makabuhay, meliburigan (Mindanao), paliaban (Bisaya), panyawan vine (Visayas). Cambodia: bandaul pech. Laos: khua kao ho. Thailand: boraphet (central). Vietnam: d[aa]ly th[aa]n th[oo]ng, d[aa]ly k[ys]ninh, d[aa]ly c[os]c.

**Distribution** India, Burma (Myanmar), Cambodia, Laos, Vietnam, southern China (Yunnan), Thailand, Peninsular Malaysia, Singapore, Indonesia (Java, Sumbawa) and the Philippines (Luzon, Mindoro, Mindanao); cultivated as a medicinal plant, e.g. in Thailand, Sri Lanka and India.

**Uses** An infusion of the stem is drunk in Malaysia and Indonesia as a vermifuge and of the whole plant to treat cholera; it is also used to treat diabetes mellitus. Externally it is applied against scabies and to heal wounds. In Brunei, the plant is used in the treatment of high blood pressure, diabetes and to relieve abdominal pains. In Indo-



*Tinospora crispa* (L.) Hook. f. & Thomson – 1, part of stem as found in the market; 2, part of stem with leafy shoot; 3, part of stem with male inflorescence; 4, male flower; 5, fruit; 6, endocarp in dorsal and ventral view.

China, an infusion of the stem is drunk to treat fever (also when caused by malaria) and jaundice. Powdered stems are used to fatten horses and cattle by stimulating their appetite. *T. crispa* is a very commonly used medicinal plant in the Philippines. In Thailand, an infusion from the stem is used to treat jaundice, cholera, malaria, and against worms in children.

**Observations** A woody climber up to 15 m long, older stems very prominently tuberculate and producing very long filiform aerial roots; leaves broadly ovate to orbicular, 7–14(–25) cm × 6–12(–24) cm, without domatia; inflorescences appearing when plant is leafless; flowers usually with 3 petals; fruit ellipsoidal, about 2 cm long, orange. *T. crispa* occurs in primary rain forest and mixed deciduous forest, but can be very common in secondary vegetation after logging and in hedges, up to 1000 m altitude. The stem contains an exceedingly bitter milky sap.

**Selected sources** 6, 75, 123, 202, 213, 331, 332, 339, 350, 421, 423, 444, 445, 446, 447, 448, 580, 908, 965, 975, 1035, 1049, 1050, 1051, 1100, 1101, 1102, 1103, 1178, 1190, 1388, 1486, 1572.

### ***Tinospora glabra* (Burm.f.) Merr.**

Journ. Arn. Arb. 19: 340 (1938).

**Synonyms** *Tinospora reticulata* Miers (1864), *Tinospora coriacea* (Blume) Beumée ex K. Heyne (1927).

**Vernacular names** Indonesia: pancasona (Sundanese), tajungan (Javanese), wase wages (Flores). Philippines: makabuhay (Luzon, Mindoro), papaitan (Palawan), sangawnaw (Mindanao).

**Distribution** The Andaman Islands, Hainan, throughout Malesia (possibly except Sulawesi) and the Solomon Islands.

**Uses** Burnt leaves are used in the Philippines to treat pinworms, and ground bark is applied to sore breasts of nursing mothers. *T. glabra* has much more medicinal applications, but the exact usage is unclear because of confusion with *T. crispa*; the latter is said to be more effective. *T. glabra* is also used for baiting wild pigs by mixing sliced roots with sweet potato (*Ipomoea batatas* (L.) Lamk).

**Observations** A woody climber, older stems becoming warty and finally developing a smooth, thin papery bark often becoming detached on drying; leaves oblong-ovate or narrowly to broadly ovate, 7–12(–15) cm × 5–9(–13) cm, domatia usually present in basal vein-axils; inflorescences appearing together with leaves; flowers with 6 petals; fruit ellipsoidal or subglobular, about 1 cm long, red. *T. glabra* often occurs in littoral rain forest and in forest dominated by *Casuarina equisetifolia* L., in mangrove vegetation and on sandy beaches, but also inland in disturbed forest and thickets, up to 500 m altitude. It is very frequently found on limestone and on black soils under seasonal conditions.

**Selected sources** 421, 580, 1178.

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### ***Trichosanthes* L.**

Sp. pl. 2: 1008 (1753); Gen. pl. ed. 5: 439 (1754).

CUCURBITACEAE

$x = 11$ ; *T. cucumerina*:  $2n = 22$ , *T. tricuspidata*:  $2n = 44$

**Major species** *Trichosanthes kirilowii* Maxim.

**Vernacular names** Malaysia: timun dendang, timun gagak (Peninsular).

**Origin and geographic distribution** *Trichosanthes* comprises some 40 species and is found from Pakistan, India and Sri Lanka to the Himalayas, Burma (Myanmar), Indo-China, China, Japan, Thailand, throughout Malesia, towards northern and eastern Australia and into the Pacific east to Fiji. Some 15 species are present within the Malesian region. Fossil evidence proves the presence of *Trichosanthes* in Eurasia during the Miocene and Eocene.

**Uses** Various *Trichosanthes* species are used for a wide array of medicinal purposes. The best known of these include application as a cooling agent, as diuretic, as galactagogue, as a hypoglycaemic, and in the treatment of various skin complaints. The fruits of several *Trichosanthes* species, e.g. those of *T. ovigera* and of cultivated forms of *T. cucumerina* are eaten as a vegetable.

**Production and international trade** Snake gourd (*T. cucumerina*) is mainly grown in home gardens for own consumption or for the local market. The wild *Trichosanthes* are collected and consumed locally only.

**Properties** The importance of *T. kirilowii* has increased due to the discovery of its activity against human immunodeficiency virus (HIV). The active proteins, trichosanthin (a mixture of 4–5 antigenic proteins), and TAP-29 (*Trichosanthes* anti-HIV protein, a 29 kDa protein), have been isolated from the roots. Both proteins exhibit anti-HIV activity in a similar way, as measured by assays for syncytium formation, p24 expression, and HIV reverse transcriptase activity. However, they differ significantly in cytotoxicity: trichosanthin demonstrates a dose-dependent toxic effect on host cells, but TAP-29 does not.

Furthermore, trichosanthin is reported to inactivate eukaryotic ribosomes via its N-glycosidase activity; it is also a potent inhibitor of protein synthesis in a reticulocyte-lysate assay. Two other proteins, karasurin-B and karasurin-C have also been isolated from the tubers of *T. kirilowii* var. *japonica*. Both karasurins had strong ribosome-inactivating activities, revealed by in vitro inhibition of translation in the rabbit reticulocyte system.

The traditional use of *T. kirilowii* as an abortifacient has led to extensive studies. In a study in which the water extract of the roots at a dose of 0.2 mg/person was administered intramuscularly to 2500 pregnant patients, the percentage of labour-

induction was 96% in late-term pregnancy, and 71% in mid-term pregnancy. Drug recipes containing whole roots of *T. kirilowii* were also found to be active. However, precautions are recommended, because of the risk of death as a complication. The compounds responsible for the abortifacient activity are trichosanthin,  $\beta$ -trichosanthin,  $\alpha$ -kirilowin and  $\beta$ -kirilowin. Furthermore,  $\beta$ -trichosanthin isolated from the tubers of *T. ovigera*, was found to be about twice as potent as trichosanthin from *T. kirilowii* in inducing mid-term abortion in mice.

The proteins  $\alpha$ -kirilowin and  $\beta$ -kirilowin, isolated from the seeds of *T. kirilowii*, have very similar biological activities as the other trichosanthins. They were found to inhibit protein synthesis in a cell-free system, to suppress [ $^3$ H]-thymidine incorporation into mouse melanoma cells, and to induce abortion in mice.  $\beta$ -Kirilowin, with an  $ID_{50}$  of about 1.8 ng/ml, inhibits the cell-free translation system.

A type-1 ribosome-inactivation protein, designated as TK-35, has been purified from suspension cultures of stem sections of *T. kirilowii* transformed by *Agrobacterium rhizogenes*. In a protein translation inhibition assay, TK-35 had an  $IC_{50}$  value of 2.45 nM and was able to release the rRNA N-glycosidase diagnostic fragment from rabbit reticulocytes.

The polysaccharide fraction from the rhizomes of *T. kirilowii* showed marked anti-tumour and cytotoxic activities together with immunopotentiating effects. The latter effects were evidenced by an increase in the number of circulating leucocytes and peritoneal exudate cells, and recovery from reduced antibody formation in mice.

In vitro cytotoxicity tests with trichosanthin showed that it selectively injured choriocarcinoma and melanoma cells. Under experimental conditions, the marked decrease in secretion of human chorionic gonadotropin and progesterone by choriocarcinoma cells after treatment with the proteins could be mainly attributed to the loss of cells. A structural, electrophoretic, variant of karasurin isolated from *T. kirilowii* roots strongly inhibited the growth of BeWo cells (a human choriocarcinoma cell line) in vitro.

Furthermore, human peripheral blood-derived monocytes and macrophages were found highly sensitive to trichosanthin; the mixture suppressed lymphocyte proliferation ( $ID_{50}$  about 1.7  $\mu$ g/ml). Human T and macrophage cell-lines were more sensitive ( $ID_{50}$  < 0.9  $\mu$ g/ml) to trichosanthin compared with B and myeloid cell-lines. These find-

ings suggest that this selective cytotoxicity towards human macrophages and/or monocytes may be implicated in trichosanthin's anti-HIV activity and, furthermore, that the selective killing of leukaemia-lymphoma cells by trichosanthin merits evaluation for possible use to treat some forms of lymphoma and leukaemia.

At non-cytotoxic concentrations (10–1000 ng/ml for splenocytes, and 10–100 ng/ml for macrophages), karasurin-A from the roots of *T. kirilowii* inhibited the lymphocyte proliferation induced by lipopolysaccharide, concanavalin A or phytohaemagglutinin, and nitric oxide production induced by lipopolysaccharide. It has also been suggested that karasurin-A has immunosuppressive activity in vitro. Trichosanthin increased the secretion of the enzymes glutamate-pyruvate-transaminase, lactate dehydrogenase and isocitrate dehydrogenase by isolated rat hepatocytes into the culture medium. This hepatotoxic effect appeared to be concentration-dependent. Trichosanthin furthermore produced adverse effects on prenatal development of mice, both in vitro and in vivo.

$\alpha$ -Trichosanthin, isolated from fresh tubers of *T. kirilowii*, showed no lipogenic activity in rat cells, nor did it affect the fasting plasma-glucose levels in mice, or testosterone and corticosterone production in isolated rat cells.

The immunotoxin trichokirin conjugated to a monoclonal antibody directed against the Thy 1.2 antigen, selectively killed leukaemia cells expressing this Thy 1.2 antigen. In in vivo application it is reported to be more advantageous than ricin A-chain immunotoxins. 2-Iminothiolane-trichosanthin conjugated to Hepama-1, which is a monoclonal antibody directed against human hepatoma, proved to be a potent and quite specific agent against hepatoma.

Bryonolic acid isolated from transformed cultures of hairy roots of *T. kirilowii* var. *japonica* exhibited cytotoxic effects against human and animal tumour cell-lines in vitro, which were independent of the cell type. Normal cells, such as rat hepatocytes, were less sensitive to bryonolic acid than tumour cells. A so-called DNA ladder was detected in bryonolic acid-treated HL-60RG cells, indicating that apoptosis may be the cause of cell death triggered by bryonolic acid.

Anti-inflammatory activity of *T. kirilowii* was investigated against ear inflammation in mice induced by tetradecanoylphorbol-13-acetate (TPA). The active constituents, isolated from the seeds, were identified as 3-epikarounidiol (= D:C-friedo-

oleana-7,9(11)-diene-3 $\beta$ ,29-diol), 7-oxoisomultiflorenol (= 7-oxo-D:C-friedo-olean-8-en-3 $\beta$ -ol), 3-epibryonolol (= D:C-friedo-olean-8-ene-3 $\alpha$ ,29-diol), and 7-oxo-10 $\alpha$ -cucurbitadienol and its acetyl- and 24-dihydro-derivatives karounidiol (= D:C-friedo-oleana-7,9(11)-diene-3 $\alpha$ ,29-diol) and 7-oxodihydrokarounidiol (= 7-oxo-D:C-friedo-olean-8-ene-3 $\alpha$ ,29-diol) which are all triterpenes. Furthermore, at 2 mmol/mouse, karounidiol markedly suppressed the promoting effect of TPA (1 mg/mouse) on skin tumour formation in mice following initiation with 7,12-dimethylbenz[a]anthracene (50 mg/mouse).

The 50% ethanol extracts of the whole fruit, as well as of the seeds of *T. kirilowii* administered orally, exhibited anti-inflammatory and analgesic activity. The anti-inflammatory activity was investigated against vascular permeability in mice induced by acetic acid, carrageenin-induced oedema, and granuloma formation in rats induced by cotton pellets; the analgesic activity against writhing symptoms in mice.

Seed extracts of many *Trichosanthes* species show potent haemagglutinating activity. Data on haemagglutination inhibition show that Me- $\beta$ D-galactose is the best monosaccharide inhibitor of the galactose-specific lectin present in the seeds of cultivated *T. cucumerina*. A lectin isolated from the root tuber of *T. kirilowii* agglutinated rabbit erythrocytes. Studies of carbohydrate-binding specificity demonstrated that agglutination was strongly inhibited by lactose and D-galactose. The galactose-binding lectin from root tubers of *T. kirilowii* stimulated the incorporation of D-(3-<sup>3</sup>H)-glucose into lipids in rat epididymal adipocytes, but did not inhibit lipolysis.

A decoction of dried tubers of *T. kirilowii* exhibited antihyperglycaemic activity. The ethanolic extract of *T. cucumerina*, when administered orally to rats in 250 mg/kg doses failed to lower blood sugar levels, or to depress the peak value after a glucose load. Five glycans, trichosans A, B, C, D and E, isolated from *T. kirilowii* roots, showed hypoglycaemic actions in normal mice. The main glycan, trichosan A, also exhibited activity in alloxan-induced hyperglycaemic mice.

The hypolipidemic activity of the pectin from cultivated *T. cucumerina* was investigated by feeding male Sprague-Dawley rats a diet containing 5% pectin. Levels of serum cholesterol, phospholipids in the liver and fatty acids in the blood fell significantly.

The chloroform extract of roots of *T. cucumerina* showed significant antibacterial activity against

*Pseudomonas aeruginosa*; its activity against *Staphylococcus aureus*, however, was not significant. Seeds extracts of cultivated *T. cucumerina* resulted in high mortality of the nematodes *Meloidogyne incognita* and *Rotylenchulus reniformis*.

**Adulterations and substitutes** *Momordica* may be a potential substitute for *Trichosanthes*, due to quite similar properties, such as antihyperglycaemic and cytotoxic activities.

**Description** Annual or perennial, monoecious or dioecious, climbing or trailing herbs. Tendrils adjacent to the petiole insertion, simple or 2–5-fid. Leaves alternate, petiolate; blade simple and unlobed to palmately 3–9-lobed or rarely palmately compound; stipules absent. Flowers axillary, unisexual, actinomorphic; calyx with a long, narrow tube and 5 entire to dentate lobes; corolla funnel-shaped, white or greenish-white, deeply 5-lobed, lobes fimbriate with long hairs. Male flowers in usually bracteate racemes or rarely solitary; stamens 3, inserted on the corolla tube, filaments short, free, anthers free or united, one 1-locular and two 2-locular; pistillode 3-parted, filiform. Female flowers solitary or rarely in racemes; staminodes absent; ovary inferior, 3-carpellate but 1-celled with 3 placentas and many ovules, style 1, with 3, entire to bifid stigmas. Fruit a fleshy, indehiscent berry (pepo), globose to long spindle-shaped. Seeds often compressed and elongated.

**Growth and development** Flowering of *T. cucumerina* commences about 5 weeks after sowing. The flowers open in the evening or early morning. Anthers shed their pollen several days before complete flower anthesis; stigmas are receptive from a few hours before anthesis to a few hours after. *Trichosanthes* flowers are pollinated by insects. The often brightly coloured fruits are eaten by monkeys but especially by large crows which thus disperse the seeds.

**Other botanical information** Within the subfamily Cucurbitaceae, *Trichosanthes* belongs to the tribe Trichosantheae, an Old World tribe of 10 genera characterized by elongated hypanthia in both male and female flowers. *Trichosanthes* is still rather poorly known taxonomically, but a revision is in preparation. Reports from the Malaysian region of *T. bracteata* (Lamk) Voigt are erroneous and may concern three widespread, related species: *T. pubera* Blume, *T. quinquangulata* and *T. tricuspidata*. True *T. bracteata* is from continental Asia.

**Ecology** Most *Trichosanthes* species are



climbers of open forest, forest margins and thickets, at low to medium altitudes. They are fairly drought resistant, although *T. cucumerina* does not tolerate dry soil.

**Propagation and planting** *T. cucumerina* is propagated by seed, requiring 4–6 kg/ha. Seed can be sown in a nursery and seedlings transplanted at the 2-true-leaf stage but usually the seed is directly sown in planting holes or on ridges 1–1.5 m apart, with 60–75 cm between plants. Seeds are soaked in water to hasten germination; under favourable conditions germination takes place within a week. Tuber-bearing *Trichosanthes* has been propagated vegetatively, but no details are available.

**In vitro production of active compounds** In vitro culture hairy roots of *T. kirilowii* were successfully induced by infection with *Agrobacterium rhizogenes* strain R1601, yielding 8.16 mg trichosanthin per g fresh weight. A recombinant *T. kirilowii* trypsin inhibitor analogue with the same activity as the natural one has been successfully synthesized, yielding 2 mg/l. Recombinant  $\alpha$ -trichosanthin with concentration-dependent inhibition of protein synthesis in vitro has been synthesized.

**Husbandry** For fruit production (e.g. *T. cucumerina*) plants are trellised or otherwise supported so that the fruits can hang down.

**Diseases and pests** The most serious diseases of *T. cucumerina* are downy mildew (*Pseudoperonospora cubensis*) and anthracnose (*Colletotrichum lagenarium*). Repeated spraying with fungicides, e.g. maneb, can control both diseases. The major pests of *T. cucumerina* are leaf beetles (*Aulacophora vinula*, *Copa occidentalis* and *Lagria villosa*) and root-knot nematodes (*Meloidogyne* spp.).

**Harvesting** For consumption as vegetable, fruits are picked 12–20 days after fruit set. For seed production, fruits are harvested when fully ripe and have attained full size.

**Yield** From dried tubers of *T. kirilowii* 0.16% of pure trichosanthin has been obtained after cation-exchange perfusion chromatography for only 10 minutes.

**Genetic resources and breeding** Germplasm collections of *T. cucumerina* are available in the Philippines (NPGRL-IPB, Los Baños), India (Kerala Agricultural University, Trichur, Kerala), Nigeria (NACGRAB, Ibadan), Russia (the Vavilov Institute of Plant Industries, St Petersburg) and the United States (Department of Horticultural Sciences, Cornell University, New York).

**Prospects** Several compounds from *Trichosanthes* show interesting pharmacological activities, e.g. cytotoxic (anti-tumour), anti-inflammatory and anti-HIV. As lead-compounds, they may have potential in research and development of future medicines. Although extracts of the roots of *T. kirilowii* are reported to be abortifacient, this activity must be considered too toxic to be applied, as death may occur as a complication.

**Literature** |1| Akihisa, T., Yasukawa, K., Kimura, Y., Takido, M., Kokke, W.C.M.C. & Tamura, T., 1994. Five D:C-friedo-oleanane triterpenes from the seeds of *Trichosanthes kirilowii* Maxim. and their anti-inflammatory effects. *Chemical and Pharmaceutical Bulletin* 42(5): 1101–1105. |2| Chan, W.Y., Ng, T.B., Wu, P.J. & Yeung, H.W., 1993. Developmental toxicity and teratogenicity of trichosanthin, a ribosome-inactivating protein in mice. *Teratogenesis, Carcinogenesis and Mutagenesis* 13(2): 47–57. |3| Gildemacher, B.H., Jansen, G.J. & Chayamarit, K., 1993. *Trichosanthes* L. In: Siemonsma, J.S. & Kasem Piluek (Editors): *Plant Resources of South-East Asia No 8. Vegetables*. Pudoc Scientific Publishers, Wageningen, the Netherlands. pp. 271–274. |4| Kumagai, M.H. et al., 1993. Rapid, high-level expression of biologically active  $\alpha$ -trichosanthin in transfected plants by an RNA viral vector. *Proceedings of the National Academy of Sciences* 90(2): 427–430. |5| Lee-Huang, S. et al., 1991. TAP 29: an antihuman immunodeficiency virus protein from *Trichosanthes kirilowii* that is nontoxic to intact cells. *Proceedings of the National Academy of Sciences* 88(1b): 6570–6574. |6| Ng, T.B., Chan, W.Y. & Yeung, H.W., 1992. Proteins with abortifacient ribosome inactivating immunomodulatory antitumor and anti AIDS activities from Cucurbitaceae plants. *General Pharmacology* 23(4): 575–590. |7| Qiu, D.Y., Zhu, G. & Zhu, Z.Q., 1996. Study on production of trichosanthin from the hairy roots of *Trichosanthes kirilowii* Maxim. *Acta Botanica Sinica* 38(6): 439–443. |8| Rugayah & de Wilde, W.J.J.O., 1997. *Trichosanthes* L. (Cucurbitaceae) in Java. *Blumea* 42(2): 471–482. |9| Shaw, P.C., Chan, W.L., Yeung, H.W. & Ng, T.B., 1994. Trichosanthin – A protein with multiple pharmacological properties. *Life Sciences* 55(4): 253–262. |10| Tsao, S.W., Ng, T.B. & Yeung, H.W., 1990. Toxicities of trichosanthin and  $\alpha$ -momorcharin, abortifacient proteins from Chinese medicinal plants, on cultured tumor cell lines. *Toxicol* 28(10): 1183–1192.

*Selection of species***Trichosanthes borneensis Cogn.**

A. DC. & C. DC., Monogr. phan. 3: 369 (1881).

**Synonyms** *Trichosanthes grandibracteata* Kurz (1877), *Trichosanthes wallichiana* Ridley non (Ser.) Wight.

**Vernacular names** Malaysia: akar timun gagak, daun jari buaya, labu ayer hutan (Peninsular).

**Distribution** Thailand, Peninsular Malaysia, Sumatra and Borneo.

**Uses** In Peninsular Malaysia, crushed fruits have been applied to the head to relieve headache; they are reported to be poisonous. The leaves are applied to the abdomen after miscarriage.

**Observations** A vine with slightly grooved, subglabrous stem; leaves suborbicular, 3–5-lobed, 8–10 cm × 6–10 cm, base cordate, lobes oblong to triangular, acute, glabrous to sparsely hirsute above, glabrescent below; corolla about 7 cm in diameter; male inflorescence 15–35 cm long, bracts oblong-ovate, 2–3 cm long, laciniate; female flowers solitary; fruit ellipsoidal to ovoid, 5–8 cm long, red; seeds angular. *T. borneensis* is reported from forests.

**Selected sources** 202, 457, 1126, 1227.

**Trichosanthes cucumerina L.**

Sp. pl. 2: 1008 (1753).

**Synonyms** *Trichosanthes anguina* L. (1753), *Trichosanthes pedatifolia* Miq. (1856), *Trichosanthes reniformis* Miq. (1856).

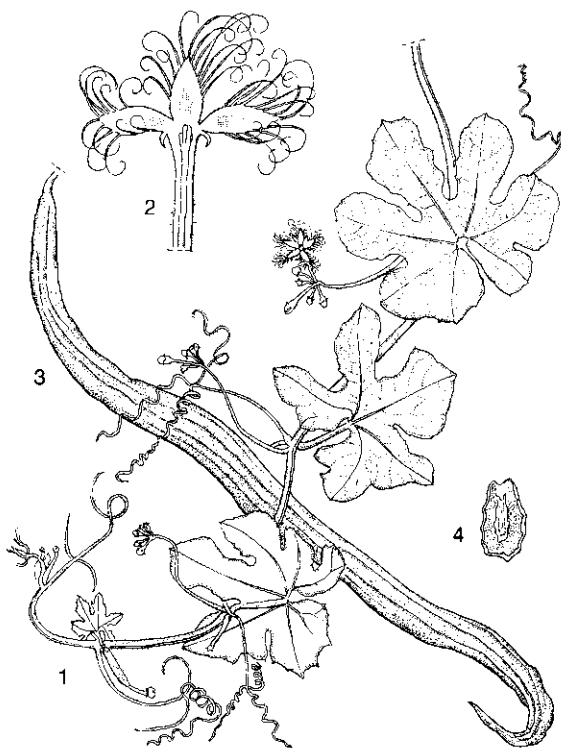
**Vernacular names** Snake gourd (En). Patole, serpent végétal (Fr). Indonesia: paria belut (general), petola ular (Malay, Moluccas), pare welut (Javanese). Malaysia: ketola ular, timun bengkok, petola ular (Peninsular). Philippines: melon-daga, pakupis, tabubok (Tagalog). Laos: ngoo ngèewz. Thailand: buap ngu, nom phichit (central), ma noi (northern). Vietnam: d[aa]y na t[aa]y, d[uw]a n[us]i, m[uw] [ows]p t[aa]y.

**Distribution** From India and Sri Lanka to Indo-China, southern China, Thailand and throughout Malesia towards northern Australia; cultivated in this area and elsewhere.

**Uses** Fruits of wild plants are used as a purgative and a vermifuge. In Indonesia, the inner pulp of the fruit is made into a syrup to treat cough. Immature fruits of cultivated forms are eaten boiled as a vegetable or in curries. Young shoots and leaves are also edible. The fruits become inedible upon ripening. In West Africa, the red fruit pulp is used as a kind of cheap tomato paste.

Fruits of the wild forms are very bitter and inedible.

**Observations** A monoecious, annual vine up to 6 m long, stem 4-angular, sparsely pubescent or glabrescent; leaves suborbicular or slightly kidney-shaped to broadly ovate, shallowly to deeply 3–7-lobed, 7–10 cm × 6–13 cm (in cultivated forms up to 25 cm across), base cordate, lobes rounded to obtuse, sparsely hirsute to almost glabrous on both sides; corolla about 3 cm in diameter; male flowers in lax, 3–25 cm long racemes, bracts small, up to 2 mm long; female flowers solitary or rarely in pairs; fruit ovoid, 4.5–7.5 cm × 3–4 cm, ripening to yellow or orange (in cultivated forms fruit linear, up to 100 cm long); seeds ovate-truncate, compressed. The cultivated forms are traditionally distinguished as a species (*T. anguina* L.), subspecies (subsp. *anguina* (L.) Greb.) or variety (var. *anguina* (L.) Haines), though it seems more appropriate to classify these in a cultivar group (e.g. cv. group Snake Gourd). Wild *T. cucumerina* is found in open forest, forest margins and scrub vegetation, up to 1000–1500 m altitude.



*Trichosanthes cucumerina* L. – 1, flowering shoot; 2, top of female flower in longitudinal section; 3, fruit of cultivated plant; 4, seed.

**Selected sources** 97, 233, 486, 721, 759, 763, 792, 886, 1128, 1134, 1178, 1263, 1313, 1443.

**Trichosanthes kirilowii Maxim.**

Prim. fl. amur.: 482 (1859).

**Synonyms** *Trichosanthes quadricirra* Miq. (1865).

**Vernacular names** Vietnam: qua l[aa]u, qua l[aa]u nh[aa]n, thao ca.

**Distribution** Korea, China, Japan and Vietnam (especially the northern part); possibly also in Taiwan and Laos.

**Uses** *T. kirilowii* is a well-known medicinal plant. A decoction of its tuberous roots and seeds is administered as thirst-quenching, bechic, expectorant, febrifuge, diuretic and galactagogue. Starch from the roots is spread on ulcers, wounds, irritation from perspiration, chicken pox and other skin diseases. A decoction of the root, fruit rind and seeds is used as an anti-emetic and a diuretic to regulate the excretory system and to relieve constipation. Furthermore, the roots are reported

as a cooling agent, a depurative, a sialagogue, and a maturative, useful in treating sunstroke, sore throat, jaundice and large boils. In Peninsular Malaysia, Chinese used to import seeds and fruit rind for use as a tonic and an astringent.

**Observations** A dioecious, perennial vine up to 10 m long, stem angular, sparsely pubescent; leaves broadly ovate to orbicular in outline, 3-5(-7)-lobed, 10-12 cm × 10-12 cm, base cordate, lobes mucronate, shortly pilose above, glabrescent below; corolla up to 7 cm in diameter; male flowers in 10-20 cm long racemes, bracts obovate-rhombic, about 1.5-2.5 cm long, coarsely toothed; female flowers solitary; fruit ovoid-globose to ellipsoidal, up to 10 cm long, yellow to orange-red; seeds oblong to ovate-orbicular, compressed. Plants from Japan have been distinguished as var. *japonica* (Miq.) Kitam (synonym: *Trichosanthes japonica* (Miq.) Regel).

The plants formerly identified as *T. kirilowii* from Taiwan have now been distinguished as a separate species: *T. rosthornii* Harms; whether this concerns a truly distinct species or a mere form of *T. kirilowii* remains as yet uncertain. *T. kirilowii* is found in open forest, scrub vegetation and grassy places, up to 2000 m altitude.

**Selected sources** 43, 44, 146, 202, 218, 230, 247, 267, 366, 367, 586, 626, 721, 764, 765, 789, 837, 870, 902, 939, 1024, 1025, 1027, 1028, 1035, 1099, 1173, 1325, 1331, 1332, 1441, 1445, 1472, 1479, 1546, 1592, 1599, 1625, 1628, 1630, 1656, 1657.

**Trichosanthes ovigera Blume**

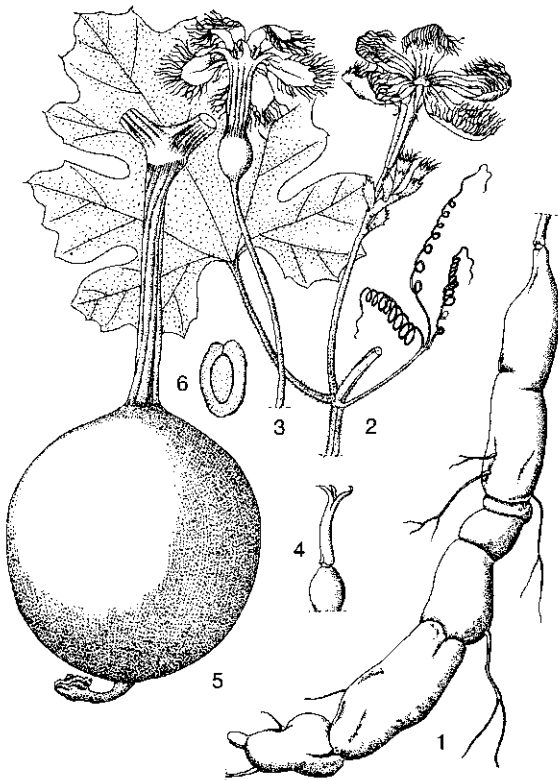
Bijdr. fl. Ned. Ind.: 934 (1826).

**Synonyms** *Trichosanthes horsfieldii* Miq. (1856), *Trichosanthes cucumeroides* (Ser.) Maxim. ex Fr. & Sav. (1875), *Trichosanthes himalensis* C.B. Clarke (1879).

**Vernacular names** Indonesia: areuy tiwuk (Sundanese). Vietnam: hoa b[as]t.

**Distribution** From the Himalayas to China, Taiwan, Vietnam, Thailand, Sumatra, Java, the Lesser Sunda Islands and north-eastern Australia.

**Uses** In China, Japan and Taiwan the root is regarded as a good remedy for intestinal parasites, jaundice and bloody stools. It is also considered to be a diuretic and galactagogue. In eastern China, it is a substitute for *T. kirilowii*. In Taiwan, the root and seeds are applied to soothe the throat and lungs and to prevent inflammation, and are also considered to be a bechic, expectorant and thirst quenching. Boiled fruits are eaten as a side



*Trichosanthes kirilowii* Maxim. - 1, main root; 2, twig with male inflorescence; 3, female flower; 4, ovary and styles; 5, fruit; 6, seed.

dish with rice. In China and Japan the starch of the tubers is sometimes extracted.

**Observations** A dioecious, perennial vine, stem angular, sparsely puberulous to glabrescent; leaves broadly ovate to suborbicular in outline, unlobed to deeply 3–5-lobed, 7–15 cm × 6–15 cm, base deeply cordate, lobes ovate to obovate, obtuse to acute, mucronate, denticulate, scabrid above, densely pubescent to velvety below; corolla up to 5 cm in diameter; male flowers in 6–20 cm long racemes, bracts narrowly obovate, 5–10 mm long, dentate; female flowers solitary; fruit ovoid to ellipsoidal, 8–10 cm × 2.5–3 cm, glabrous; seeds broadly ovate, cylindrical. *T. ovigera* is found in light forests, forest margins, thickets and roadsides, up to 1600 m altitude.

**Selected sources** 97, 486, 721, 870, 1024, 1126, 1263, 1443.

### **Trichosanthes quinquangulata A. Gray**

U.S. Expl. Exped., Bot., Phan. 1: 645 (1854).

**Vernacular names** Indonesia: kalayar (Sundanese). Philippines: kalanum-uak (Bisaya), katinbau (Iloko), patolang-gubat (Tagalog).

**Distribution** Laos, Vietnam, Thailand, Peninsular Malaysia, Sumatra, Java, Borneo and the Philippines.

**Uses** In the Philippines, the cooked, powdered seeds are applied to itch and mixed with wine to treat stomach-ache. Some of the uses reported below for *T. tricuspidata* may actually refer to *T. quinquangulata*, due to the confusion of species.

**Observations** A largely monoecious, perennial vine up to 6 m long, stem angular, glabrous; leaves roundish in outline, shallowly or sometimes deeply 5–7-lobed, up to 20 cm in diameter, base cordate, lobes acute, glabrous or glabrescent; corolla 7–9 cm in diameter; male inflorescence 15–20 cm long, bracts elliptical to obovate-lanceolate, 15–20 mm long, subentire; female flowers solitary; fruit globose, 5–10 cm in diameter, red; seeds elliptical with obtusely acuminate apex, compressed. *T. quinquangulata* occurs in thickets, old clearings and rocky places up to 1500 m altitude.

**Selected sources** 190, 457, 721, 1126, 1178, 1263.

### **Trichosanthes tricuspidata Lour.**

Fl. cochinch. 2: 588 (1790).

**Synonyms** *Trichosanthes tricuspis* Miq. (1856).

**Vernacular names** Indonesia: kalayar (Sundanese). Thailand: kradueng chang phueak (Prachuap Khitikan), kheekaa khom (Phangnga),

matuum kaa (Nakhon Ratchasima). Vietnam: l[aa]u s[as]c.

**Distribution** Indo-China, Thailand, Peninsular Malaysia, Sumatra, Java, Borneo and the Philippines.

**Uses** The fruits are strongly purgative and emetic. In Peninsular Malaysia and Borneo, the leaves are used to poultice boils. In Indonesia, the leaf juice is drunk by children to treat diarrhoea. However, in Borneo leaves and shoots are reported strongly purgative and emetic. The crushed and fermented fruit is eaten as a condiment. Due to confusion with other species, some of these uses may actually refer to *T. quinquangulata*, *T. pubera* or even the extra-Malesian species *T. bracteata*.

**Observations** A dioecious, perennial vine up to 6 m long, stem sharply angular, almost glabrous; leaves broadly ovate in outline, 3(–5)-lobed or rarely entire, 10–12 cm in diameter, base deeply cordate, lobes acute, glabrescent; corolla about 8 cm in diameter; male inflorescence 10–18 cm long, bracts broadly elliptical to broadly elliptical-obovate, about 15 mm × 25 mm, dentate; female flowers solitary; fruit ovoid, red, 6–9 cm long; seeds obovate, compressed. *T. tricuspidata* is found in thickets and open places.

**Selected sources** 457, 580, 721, 829, 1126, 1263.

### **Trichosanthes villosa Blume**

Bijdr. fl. Ned. Ind.: 934 (1826).

**Synonyms** *Trichosanthes kerrii* Craib (1914).

**Vernacular names** Indonesia: areuy baduyut, waluh leuweung (Sundanese). Philippines: kandelamo (Bukidnon). Laos: cho' tau quan. Vietnam: d[aa]y d[or] m[or].

**Distribution** Laos, Vietnam, Thailand, Peninsular Malaysia, Sumatra, Java, Borneo and the Philippines.

**Uses** Juice from the plant is drunk to treat diarrhoea when the stool is white. Crushed leaves have been smeared on the body to reduce fever, and also to alleviate the pain of swollen legs of women during pregnancy. Young fruits are edible when boiled.

**Observations** A dioecious, perennial vine of up to 10 m long, stem slightly grooved, velutinous; leaves broadly ovate, entire to slightly 3-lobed, 10–16 cm × 5–18 cm, base cordate, lobes acute, pubescent above, velutinous below; corolla about 6 cm in diameter; male inflorescence 10–20 cm long, bracts elliptical, 3–4 cm long, entire or nearly so, acute; female flowers solitary; fruit globose to el-

lipoid-globose, 8–13 cm in diameter, bright yellow to reddish-brown or red; seeds obovate with a truncate apex, compressed. *T. villosa* is found in brushwood and forest edges up to 700 m altitude.

**Selected sources** 97, 457, 486, 580, 721, 935, 1263.

### **Trichosanthes wawrae Cogn.**

A. DC. & C. DC., Monogr. phan. 3: 384 (1881).

**Synonyms** *Trichosanthes trifolia* auct. non (L.) Blume.

**Vernacular names** Malaysia: akar tiga cabang (Peninsular). Thailand: kheekaa din (peninsular).

**Distribution** Peninsular Malaysia, Singapore, Sumatra and Java: possibly also in peninsular Thailand.

**Uses** In Peninsular Malaysia, pounded leaves have been applied to the abdomen to relieve abdominal pains. In Indonesia, the leaf juice has been used to treat paralysis and oedema.

**Observations** A dioecious, perennial vine up to 10(–15) m long, stem angular, glabrous; leaves 3-foliolate with elliptical to ovate-elliptical, acuminate, serrate-dentate, glabrous leaflets, the central one 6–12.5 cm × 2–5 cm, lateral ones very unequal; corolla lobes about 1 cm long (excluding fringe); male inflorescence 10–16 cm long, bracts elliptical, distinctly dentate; female flowers solitary; fruit broadly ellipsoidal, 8–10 cm × 6–7 cm, red; seeds elliptical-obovate, truncate at base, compressed. *T. wawrae* is found in brushwood and forest, up to 1000 m altitude. It has often erroneously been treated as *T. trifolia* (L.) Blume, but that name actually refers to a *Momordica* species.

**Selected sources** 97, 202, 1126, 1227, 1263, 1380.

M.S.M. Sosef, E. Boer & N. Bunyapraphatsara

### **Verbena officinalis L.**

Sp. pl. 1: 20 (1753).

VERBENACEAE

2n = 12, 14

**Vernacular names** Vervain, holy wort (En). Herbe sacrée, verveine officinale (Fr). Malaysia: marphin choo. Philippines: verbena. Thailand: nang dong laang. Vietnam: c[or] roi ng[uw]ja, m[ax] tilee[n] th[ar]o.

**Origin and geographic distribution** *V. officinalis* is a species of temperate and subtropical regions, probably originating from the Mediterranean region. It is found at both low and higher

elevations in South-East Asia, e.g. in Java, New Guinea, Luzon, northern Thailand and Vietnam. It was introduced for its ornamental value or traditional use in folk medicine in Europe. In general it has become a weed as a follower of cultivation.

**Uses** The aerial parts of *V. officinalis* are commonly used in European traditional medicine as digestive aid and mild diuretic, to stimulate the renal excretion of water. Furthermore, it is considered a tonic, galactagogue, emmenagogue, purgative, febrifugal, diaphoretic, astringent, anthelmintic, antihæmorrhagic, antispasmodic and antiscorbutic. Externally, it is applied as a gargle to treat throat problems and stomatitis, and as a compress or poultice against ulcers, cuts, contusions, piles and headache. Topical application as an emollient also includes the relief of itching in cases of skin disorders, sunburn and burns. In Europe, reported indications for its use are treatment of jaundice, chlorosis, dropsy, gout, kidney and bladder stones, rheumatism, hæmaturia, fever, neuralgia and ophthalmia.

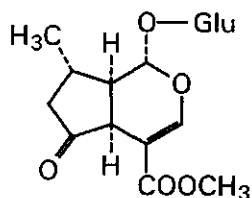
In Indo-China, China, Taiwan and Korea, *V. officinalis* is used internally against colds, fever, inflammations, digestive and intestinal complaints, uterine problems and disorders in the urinary tract. It is also taken after parturition, as a depurative and to help to remove the placenta, in cases of oedema, anaemia, tympanitis, congestion and as antidote after insect bites. Externally, it is used as a poultice or wash to treat skin diseases, wounds, abscesses and tumours. In Vietnam, a decoction of *V. officinalis* is used against dropsy and a poultice against tumours on the scrotum, whereas it is also considered to be useful to regulate menstruation. In Thailand, *V. officinalis* is used in the treatment of liver and gall bladder complaints, colds, fever, bronchitis and mental disorders, whereas the leaves are applied externally to treat rheumatism, wounds and eczemas. In India, the fresh leaves are used as a febrifuge, tonic and as a rubefacient to treat rheumatism or other diseases of the joints, whereas the root is believed to be useful in the treatment of scrofula and snake bite.

*V. officinalis* is applied as an insecticide. Other *Verbena* species like *V. laciniata* (L.) Briq., *V. rigida* Spreng. and *V. xhybrida* Voss are cultivated in Java as ornamentals, but *V. bonariensis* L. is naturalized and found in fields, grassland, plantations and roadsides. Medicinal uses have been reported for *V. rigida* in Africa, where root decoctions are used against heartburn and colic and for

*V. bonariensis* in Brazil, where it is used to treat fevers and catarrh.

**Properties** *V. officinalis* has a bitter taste. It is known to contain iridoids and their glycosides, such as hastatoside, verbenalin (= verbanalloside, cornin) and verbenin (= aucubin), and phenylpropanoid glycosides e.g. verbascoside (= acetoside) and eukovoside (= leucosceptosid-A). Further phytochemical investigations have also revealed the presence of phytoestrogens and phytoprogestins, flavonoids, saponins, tannins, mucilage, adenosine and  $\beta$ -carotene. *V. officinalis* leaves from Morocco were found to yield an essential oil containing more than 40 compounds, the predominant ones being spathulenol (10.8%), limonene (7.5%), 1,8-cineole (7.5%), caryophyllene-epoxide (7.3%), ar-curcumene (6.0%), geranial (3.3%), neral (2.5%) and  $\alpha$ -terpineol. Verbenalin (= verbanalloside), a monoglycoside of the iridoid verbenalol, is present in all plant parts. It has been shown that iridoid biosynthesis efficiency in *V. officinalis* decreases during the flowering period. Furthermore, verbenalin is identical to cornin, which has been isolated from *Cornus florida* L. Verbenalin is non-toxic and has parasympathomimetic properties. It stimulates the uterus in mammals, giving an increased tonus and stronger contractions. In dogs it slightly retards cardiac movements, reduces blood pressure and respiration, causes vasodilation in the kidneys and gives increased and more regular intestinal movements. In rabbits, it has shown antithermic activity, and it stimulates smooth muscle preparations of isolated rabbit intestines. In frogs, verbenalin in large doses is reported to stimulate the motoric activities of the central nervous system, causing stupor, convulsions and paralysis.

Verbenin (= aucubin), another iridoid glycoside, is known to have galactagogue properties. When given to lactating animals it increases milk secretion. The phenylpropanoid verbascoside (= acetoside) is reported to act as an agonist to the antitremor action of laevodopa, and furthermore as an antihypertensive and analgesic.

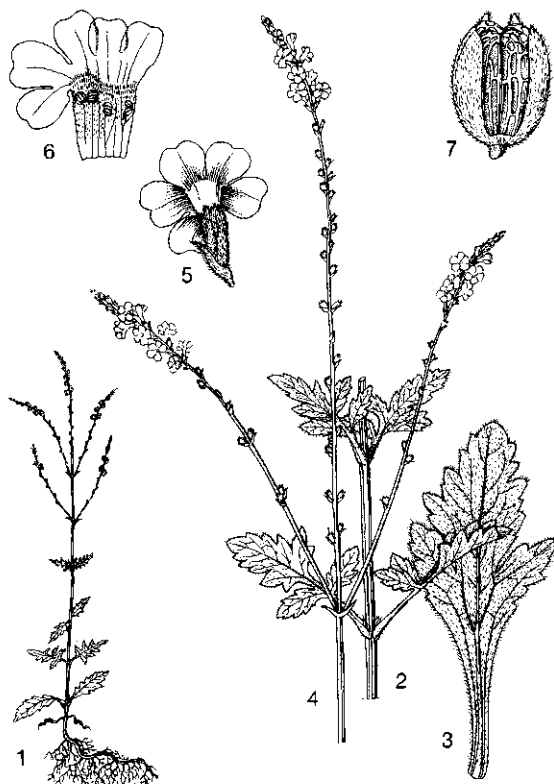


verbenalin (Glu = glucose)

Pharmacological properties attributed to the plant as a whole include anti-microbial activity and antiviral activity against the murine cytomegalovirus. Its usefulness in the prevention and treatment of kidney-stone formation might be attributed to its disinfectant action and perhaps to the presence of saponins.

Finally, *V. officinalis* is suspected of being poisonous to cattle, *V. rigida* of causing irritation in livestock and *V. bonariensis* of causing abortion in cows.

**Description** A perennial herb up to 100 cm tall; stem erect or decumbent at base, shallowly furrowed, glabrous or sparingly pubescent, tough. Leaves opposite, ovate-oblong in outline, pinnatifid to pinnately divided, 2.5–8 cm  $\times$  0.8–5 cm, base attenuate, apex acuminate, sessile by a narrowed base. Inflorescence a spike, lax at anthesis, 5–20 cm long, solitary or combined into a lax compound inflorescence. Flowers with a tubular calyx 2.5–3 mm long at anthesis, 5-toothed, densely glandular



*Verbena officinalis* L. - 1, plant habit; 2, part of stem; 3, leaf; 4, inflorescence; 5, back of flower with bracteole; 6, corolla opened out; 7, fruit with part of calyx removed.

pubescent, scarcely longer than the fruit; corolla tube 3–4 mm long, corolla limb 5-lobed, 3–5 mm in diameter, pale lilac; stamens 4, inserted on the corolla tube; ovary superior, 4-celled, cells 1-ovuled, style short. Fruit breaking up into 4 closed cocci; cocci about 2 mm long. Seedling with epigeal germination; cotyledons herbaceous, glabrous; hypocotyl elongated, epicotyl present.

**Other botanical information** *Verbena* consists of about 200(–450) species, almost exclusively originating from temperate and tropical regions of the Americas. Only 2 species can be considered as originating from Europe or the Near East. A considerable number have been introduced in other parts of the world, mostly as garden ornamentals, and sometimes have become naturalized or even weedy. In North America *V. hallei* Small, indigenous to the south-eastern United States, is now considered as a subspecies of *V. officinalis* (*V. officinalis* L. subsp. *hallei* (Small) Barber). However, *V. officinalis* L. subsp. *officinalis* has been introduced from Europe and is now common in the eastern Atlantic part of the United States. In East Africa, the morphological distinctness found is given a taxonomic rank as *V. officinalis* subsp. *africana* R. Fernandes & Verdcourt.

**Ecology** *V. officinalis* seeds germinate only when mean temperatures are above 14°C, with day temperatures higher than 19°C. Flowering is promoted at temperatures of 16°C or higher. *Verbena* spp. do not tolerate high relative humidity.

**Propagation and planting** *V. officinalis* can be propagated through seed, cuttings and root-stock division.

**Harvesting** Aerial parts are collected before anthesis.

**Handling after harvest** After collecting, aerial parts and roots can be dried for storage and future use.

**Prospects** Although *V. officinalis* is credited with a wide range of medicinal properties, most of these remain to be scientifically confirmed. It might be useful as a remedy for fever and common colds, and to prevent and treat urolithiasis, but more effective and equally innocuous substances are already available.

**Literature** [1] Chalchat, J.C. & Garry, R.P., 1996. Chemical composition of the leaf oil of *Verbena officinalis* L. *Journal of Essential Oil Research* 8: 419–420. [2] Chuakul, W., Saralamp, P., Paonil, V. & Temsiririkkul, R., 1996. Samunpri Puenban Lanna [Medicinal plants in the northern part of Thailand]. Department of Pharmaceutical Botany, Faculty of Pharmacy, Mahidol University.

Amarin Printing and Publishing Public Co. Ltd., Bangkok, Thailand. p. 222 [3] Garnier, G., Bézanger-Beauquesne, L. & Debraux, G., 1961. *Ressources médicinales de la flore française* [Medicinal resources of the French flora]. Vol. 2. Vigot Frères, Paris, France. pp. 1146–1148. [4] Lahloub, M.F., 1986. Phenylpropanoid and iridoid glycosides from the Egyptian *Verbena officinalis*. *Planta Medica* 52: 47. [5] Moldenke, H.N. & Moldenke, A.L., 1983. *Verbena*. In: Dassanayake, M.D. & Fosberg, F.R. (Editors): *A revised handbook to the flora of Ceylon*. Vol. 4. Amerind Publishing Co., New Delhi, India. pp. 198–211. [6] Newall, C.A., Anderson, L.A. & Phillipson, J.D. (Editors), 1996. *Herbal Medicines*. The Pharmaceutical Press, London, United Kingdom. p. 263. [7] Oliver-Bever, B., 1986. *Medicinal plants in tropical West Africa*. Cambridge University Press, Cambridge, United Kingdom. p. 239. [8] Perry, L.M., 1980. *Medicinal plants of East and Southeast Asia*. Attributed properties and uses. The MIT Press, Cambridge, Massachusetts, United States & London, United Kingdom. p. 430. [9] Pételot, A., 1953. *Les plantes médicinales du Cambodge, du Laos et du Vietnam* [The medicinal plants of Cambodia, Laos and Vietnam]. Vol. 2. Centre National des Recherches Scientifiques et Techniques, Saigon, Vietnam. p. 243. [10] Quisumbing, E., 1978. *Medicinal plants of the Philippines*. Katha Publishing Co., Quezon City, the Philippines. pp. 805–806.

**Other selected sources** 97, 124, 135, 151, 193, 195, 215, 258, 287, 309, 428, 504, 508, 548, 572, 646, 900, 950, 1035, 1217, 1233, 1476, 1554, 1583, 1596, 1645.

Wongsatit Chuakul, Noppamas  
Soonthornchareonnon & Promjit Saralamp

## Vernonia Schreber

Gen. pl. 2: 541 (1791).

COMPOSITAE

$x = 9, 10$ ; *V. anthelmintica*:  $2n = 20$ , *V. cinerea*:  $2n = 18$ , *V. elaeagnifolia*:  $2n = 40$

**Major species** *Vernonia cinerea* (L.) Less., *V. patula* (Dryander) Merr.

**Origin and geographic distribution** *Vernonia* comprises about 1000 species occurring in tropical, subtropical and temperate regions of America, Africa and Asia with its main centre of diversity in the Neotropics. About 35 species occur in the Malesian region, most of which are herbs, shrubs or climbers; only two species are trees.

**Uses** All *Vernonia* species have in common that

bitter constituents can be found in almost all plant parts. A multitude of medicinal properties are ascribed to these bitter constituents in particular. Ground leaves or a poultice of leaves of *V. cinerea* are a remedy for headache and when mixed with a little lime they are also applied to dress wounds. The leaves, either ground or as a decoction, are also used against skin diseases. In Indo-China and the Philippines, a decoction of the root is administered against diarrhoea and stomach-ache. In the Philippines, an infusion of the plant is used in the treatment of cough. In the Moluccas, the root is applied as a cough medicine and the juice of the whole plant to promote parturition. In India the flowers are administered for conjunctivitis and the root is given in cases of dropsy, whereas the seeds are also employed as an anthelmintic and alexipharmic. A decoction of *V. patula* is used as a general tonic, febrifuge and to treat diarrhoea.

*V. anthelmintica* (L.) Willd., occurring in India, Burma (Myanmar) and Laos, but not in the Malayan region, is applied as a vermifuge, and has a reputation for curing leprosy and skin diseases. It is also used as insecticide and insect repellent. The seeds, with a high content of epoxy acid, have been traded to Java as 'kursani'; in high dosage they are claimed to be an abortivum. *V. elaeagnifolia* DC., from Burma (Myanmar), northern Thailand and Indo-China, is reported to be used in a mixture with tobacco. The dried and finely chopped wood is smoked in the form of cigarettes to treat ulcerations of the nose. It is also an ingredient of an infusion to relieve cough. *V. arborea* Buch.-Ham., a common tree of secondary forest, has some medicinal applications as well. An infusion of the root, or a decoction of the bark together with other ingredients, may be given against fever. In southern Sumatra, the bark was chewed at the first signs of sprue.

In Africa, the leaves and roots of *V. amygdalina* Delile are widely applied in folk medicine. A decoction is taken as a febrifuge and to relieve abdominal pains. In East Africa, the leaves or roots of various *Vernonia* species, either chewed or as a decoction, are used to cure stomach-ache. Another general application of the leaves, as an infusion or decoction, is in the treatment of coughs, and as a poultice on wounds and sores, either fresh, cooked or pounded.

The young shoots of *V. cinerea* and *V. patula* are consumed as a cooked vegetable in Java.

**Properties** As with many other *Compositae*, *Vernonia* is rich in sesquiterpene lactones. The

presence of common compounds such as triterpenes, sterols and flavone glycosides in Old World *Vernonia* is reported in literature.

The methanol extract of the whole plant of *V. cinerea* showed significant diuretic activity in rats at doses of 300, 700 and 1000 mg/kg body weight. Activity could be concentrated in the water-soluble fraction of the extract: the freshly prepared water-soluble fraction, representing 300 mg/kg body weight of the dried methanol extract was found to be effective. The effect was not linked to a certain compound or group of compounds; general chemical analysis of this fraction showed the presence of tannins, sugars, flavonoids and glycosides. Furthermore, an aqueous ethanol (50%) extract of the whole plant showed anti-cancer activity against Sarcoma 180 in mice; the maximum tolerated dose was found to be 500 mg/kg body weight.

Vernonin is a triterpenoid isolated from *V. cinerea*. When injected intravenously in dogs it produces hypotension and an action on the heart comparable to that of digitalin, but in general the compound is much less toxic. Other triterpenes isolated from the roots of *V. cinerea* are 3 $\beta$ -acetoxy-urs-19-ene and lupeol acetate. When *V. cinerea* flowers were screened for insecticidal principles, six pyrethrins (pyrethrin I, cinerin I, jasmolin I, pyrethrin II, cinerin II and jasmolin II) were identified by thin layer chromatography.

The anti-inflammatory effect of the alcoholic extract of *V. cinerea* flowers was tested in adjuvant-induced arthritic rats. It was concluded that the extract, administered orally at 100 mg/kg, contained as yet unidentified anti-inflammatory principles. The petroleum ether and chloroform extracts of *V. patula* showed significant in vitro activity against lymphoid leukaemia L1210 and lymphocytic P388 tumour cell lines. A partially purified fraction was obtained from the chloroform extract, which was significantly active against these tumour cells in in vitro and in vivo models. Preliminary investigations by column chromatography of both petroleum ether and chloroform extract yielded 4 terpenes (2 identified as  $\alpha$ -amyrin and its acetate; both inactive in anti-cancer tests) and 1 sterol ( $\beta$ -sitosterol).

Vernolepin, a sesquiterpene dilactone isolated from *V. amygdalina*, shows platelet anti-aggregating properties. It has a stabilizing effect during freeze-thawing of platelets; it inhibits platelet aggregation induced by arachidonic acid, ADP and collagen, and interferes with ATP release. Electron microscopy shows protection of platelets



against adhesion together with a disaggregating effect. All these activities are time-dependent; a steep dose-response relationship is seen. Furthermore, vernolepin has antitumour effects in vitro, just like vernodalin and vernomygdin, two more cytotoxic sesquiterpene lactones isolated from *V. amygdalina*.

Anti-leishmanial activity of chloroform and methanol extracts of *V. amygdalina* has been assessed in vitro on *Leishmania aethiopica*. Amastigotes were more sensitive to *V. amygdalina* than promastigotes. The chloroform extract had a stronger parasitocidal activity (with median effective doses (ED<sub>50</sub>) of 18.5 µg/ml for promastigotes and 13.3 µg/ml for amastigotes), than the methanol extract (with ED<sub>50</sub> of 74.4 µg/ml and 45.8 µg/ml, respectively). Cytotoxicity caused by *V. amygdalina* to host cells, the human leukaemia monocyte THP-1 cell line, as determined by the methyl tetrazolium assay, resulted in a median lethal dose (LD<sub>50</sub>) of 19.6 µg/ml for the chloroform extract and 243.4 µg/ml for the methanol extract. In comparison, the ED<sub>50</sub> and LD<sub>50</sub> of pentamidine, a standard anti-leishmanial drug, were 0.5 µg/ml and 1.4 µg/ml respectively. These results indicate that *V. amygdalina* displays potent anti-leishmanial activities and warrants further investigation. Organic solvent extracts of leaves of *V. amygdalina* have inhibitory activity for His<sup>r</sup> to His<sup>s</sup> reverse-mutations induced by ethyl methane sulphonate acting on *Salmonella typhimurium* TA100. The concentrated ethyl acetate, methanol and petroleum ether extracts were heat-stable when dissolved in dimethyl sulphoxide.

Phytochemical investigations of *V. anthelmintica* revealed the presence of a novel 4 $\alpha$ -methylsterol, being 4 $\alpha$ -methyl-5 $\alpha$ -stigmasta-8,14,24,24'-Z-trien-3 $\beta$ -ol (= 4 $\alpha$ -methylvernosterol). The 4-demethylsterol and 4,4-dimethylsterol fractions from the seeds were also investigated. The 4-demethylsterol fraction contained vernosterol and avenasterol as the dominant sterols. 4 $\alpha$ -Methylvernosterol is suggested to be the possible intermediate in the biosynthesis of vernosterol in *V. anthelmintica* seeds.

Clinical tests of the pollen antigens from *V. cinerea* revealed that they are common allergens causing respiratory tract allergy. The pollen showed allergic manifestations in very low concentrations, were heat stable and non-dialysable, but lost their allergenic property on incubation with trypsin and chymotrypsin.

**Adulterations and substitutes** Sesquiterpene lactones of the germacranoline type are also found

in *Elephantopus*, a closely related genus belonging to the tribe *Vernonieae*.

**Description** Evergreen herbs, shrubs, climbers or rarely small to medium-sized trees up to 30(–40) m tall. Leaves arranged spirally, simple, margin entire, glandular below, petiolate, stipules absent. Inflorescence terminal or in the upper leaf axils, consisting of widely branched panicles of heads; head with many white, pinkish or purple flowers which are slightly to much longer than the turbinate to campanulate involucre; receptacle alveolate, naked. Flowers bisexual, 5-merous, pappus present, corolla tubular with a campanulate to funnel-shaped limb; anthers fused, with sagittate base; ovary inferior, 1-celled and 1-ovulate, style split into 2 stigmas. Fruit a faintly to prominently ribbed, cylindrical to many-angled achene; pappus hairs usually 2- or rarely 1-seriate, scabrous. Seedling with epigeal germination; cotyledons leafy, glabrous but upper surface glandular dotted, apex truncate to shallowly emarginate; hypocotyl up to 5 mm long, epicotyl very short; first two leaves opposite, subsequent ones alternate.

**Growth and development** Light enhances germination of *V. cinerea* and seeds germinate over a 30-day period at 25°C. Germination decreases gradually from about 60% on the soil surface to less than 10% at 4 cm depth, and seedlings do not emerge from a depth over 1 cm. The pappus on the fruits enhances dispersal by wind and animals.

**Other botanical information** The Old World *Vernonia* species are sometimes placed in the subgenus *Orbisvestus*, whereas the New World species are placed in the subgenus *Vernonia*. This subdivision is supported by differences in chromosome numbers and sesquiterpene lactones. Several attempts have been made to refine the genus *Vernonia* by segregating genera and delimiting sections. However, at present no consensus has been reached by those working on the genus.

**Ecology** *Vernonia* occurs mostly in sunny or slightly shaded habitats, in general corresponding with young secondary vegetation, wasteland and other anthropogenic habitats, from sea-level to 1400 m altitude.

**Propagation and planting** *Vernonia* can be easily propagated by seed.

**Diseases and pests** *V. cinerea* can be the host of tobacco leaf curl virus, powdery mildew (*Erysiphe cichoracearum*), root-knot nematodes, and cotton bollworm.

**Harvesting** When grown as a vegetable, *Ver-*

*nonia* is harvested when still young. For medicinal purposes the leaves are picked or the roots are collected.

**Genetic resources and breeding** The *Vernonia* species of medicinal importance have a wide distribution and are common weeds in anthropogenic habitats. Therefore the risk of genetic erosion seems limited.

**Prospects** Biological activities of extracts of *Vernonia*, and their isolated, purified compounds (e.g. the sesquiterpene lactones) show considerable potential in the treatment of e.g. tumours or leishmaniasis. The importance of these indications merits further research. Furthermore, the medicinal *Vernonia* have potential to be grown as a plantation crop.

**Literature** |1| Akihisa, T., Hayashi, Y., Patterson, G.W., Shimizu, N. & Tamura, T., 1992. 4 $\alpha$ -Methylvernosterol and other sterols from *Vernonia anthelmintica* seeds. *Phytochemistry* 31(5): 1759-1763. |2| Burkill, I.H., 1966. A dictionary of the economic products of the Malay Peninsula. 2nd edition. Vol. 2. Ministry of Agriculture and Co-operatives, Kuala Lumpur, Malaysia. pp. 2266-2268. |3| Harborne, J.B. & Williams, C.A., 1977. Vernonieae - chemical review. In: Heywood, V.H., Harborne, J.B. & Turner, B.L. (Editors): The biology and chemistry of Compositae. Vol. 1. Academic Press, London, New York, San Francisco. pp. 523-537. |4| Herrera, C.L., Chanco, G.L. & Sison, F.M., 1980. Chemical and pharmacological studies on *Vernonia patula* (Dry.) Merr. and *Vernonia cinerea* (Linn.) Less. In: 4th Asian Symposium on Medicinal Plants and Spices: Abstracts. Department of Chemistry, Faculty of Science, Mahidol University, Bangkok, Thailand. p. 249. |5| Holm, J., Doll, J. & Holm, E., 1997. World weeds: natural histories and distribution. Wiley, New York, United States. pp. 903-906. |6| Isawumi, M.A., 1995. Notes on *Vernonia* (Vernonieae: Compositae) in West Africa. In: Hind, D.J.N., Jeffrey, C. & Pope, G.V. (Editors): Advances in Compositae systematics. Royal Botanic Gardens, Kew, United Kingdom. pp. 51-106. |7| Koster, J.T., 1935. The Compositae of the Malay Archipelago. I. Vernonieae and Eupatorieae. *Blumea* 1: 351-536. |8| Obaseiki-Ebor, E.E., Odukoya, K., Telikepalli, H., Mitscher, L.A. & Shankel, D.M., 1993. Antimutagenic activity of extracts of leaves of four common edible vegetable plants in Nigeria (West Africa). *Mutation Research* 302(2): 109-117. |9| Oliver-Bever, B., 1986. Medicinal plants in tropical West Africa. Cambridge University Press, Cambridge, New York, Melbourne. p. 28. |10| Tadesse, A., Ge-

bre-Hiwot, A., Asres, K., Djote, M. & Frommel, D., 1993. The in vitro activity of *Vernonia amygdalina* on *Leishmania aethiopica*. *Ethiopian Medical Journal* 31(3): 183-189.

#### *Selection of species*

#### ***Vernonia cinerea* (L.) Less.**

Linnaea 4: 291 (1829).

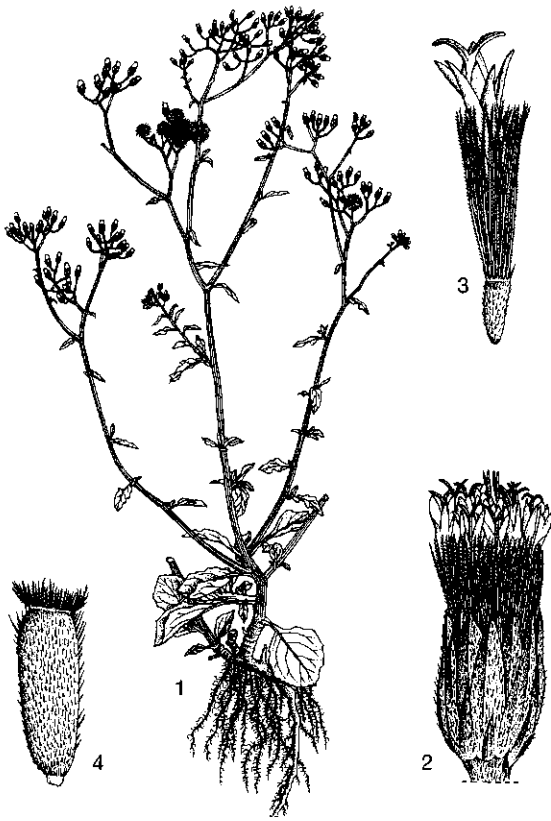
**Synonyms** *Conyza cinerea* L. (1753), *Vernonia leptophylla* DC. (1836).

**Vernacular names** *Vernonia* (En). Little ironweed (Am). Indonesia: maryuna (Javanese), sasawi langit (Sundanese), buyung-buyung (Malay, Moluccas). Malaysia: rumput tahi babi, tambak-tambak (Peninsular). Philippines: kolong-kugon (Bisaya), agas-moro (Ilokano), bulak-manok (Tagalog). Thailand: kaan thuup (south-eastern), yaa dok khaao (central), yaa saam wan (northern). Vietnam: b[aa]c d[aaf]u, d[aj] h[uw]l[ow]ng ng[uw]u.

**Distribution** Originally from the Old World, but now a pantropical weed. In Malesia, it is common throughout the region.

**Uses** Ground leaves or a poultice of leaves is a well-known remedy for headache and, when mixed with a little lime, they are also used to dress wounds. The leaves, either ground or as a decoction, are also applied against skin diseases. In Indo-China and the Philippines, a decoction of the root is used against diarrhoea and stomachache. An infusion of the plant is administered in the treatment of cough. In Thailand, the leaves are used in the treatment of asthma and bronchitis. The young shoots are eaten as a cooked vegetable in Java.

**Observations** An erect herb, 15-80 cm tall, sometimes branched in the upper part, roots crowded, short; leaves variable, lower ones 3-9 cm  $\times$  1.5-3.5 cm, upper ones 1-7 cm  $\times$  0.3-1.5 cm; inflorescence a terminal panicle, compound and corymbose, 5-35 cm  $\times$  5-15 cm, consisting of numerous heads; head turbinate, 20-25-flowered, involucre 3-4-seriate, much shorter than the head, 4-5 mm long, tomentose, corolla 3.5-4.5 mm long, tubular; fruit 1.5-2(-2.5) mm long, cylindrical, faintly ribbed, appressed whitish pubescent, densely brownish glandular, pappus biseriate, outer elements shorter and broader than inner ones, inner setae 4-5 mm long, caducous, white. *V. cinerea* is very polymorphic, and may be divided into several varieties. It flowers throughout the year and grows on a wide range of soils in rather



*Vernonia cinerea* (L.) Less. - 1, flowering plant; 2, flower head; 3, flower; 4, achene without inner pappus row.

open habitats, from sea-level to 1300 m altitude. **Selected sources** 92, 97, 202, 287, 332, 458, 531, 580, 613, 654, 741, 758, 774, 821, 953, 1085, 1126, 1128, 1178, 1386.

***Vernonia patula* (Dryander) Merr.**

Philipp. Journ. Sci., Bot. 3: 439 (1908).

**Synonyms** *Vernonia chinensis* Less. (1831).

**Vernacular names** Indonesia: yawun (Javanese), sarap (Madurese). Malaysia: ruku gajah, perasi putih (Peninsular). Vietnam: b[aj]c d[aa]f[u].

**Distribution** Common in north-east India, Burma (Myanmar), Indo-China, southern China, Japan and throughout Malesia.

**Uses** In Malaysia a decoction of roots and leaves is applied to treat colds and fevers. In Indo-China the plant is used as a tonic and against diarrhoea. The young shoots are consumed as a vegetable in Java.

**Observations** An almost shrubby herb, 20-70 cm tall, stem much branched, roots crowded;

leaves very variable, lower ones 2.5-10.5 cm × 1-4.5 cm, upper ones up to 1.5 cm × 0.5 cm; inflorescence a widely branched panicle or corymb consisting of many heads; head campanulate, 75-100-flowered, involucre 4-seriate, almost as long as the head, 4-6 mm long, tomentose, corolla 6-7 mm long, tubular; fruit 1-1.5 mm long, 4-6-angular, with prominent ribs, glabrous, glandular, pappus uniseriate, 2 mm long, caducous, setaceous, white. *V. patula* is polymorphic, and may be divided into several varieties. It flowers throughout the year and grows on a wide range of soils in rather open habitats, from sea-level to 1400 m altitude.

**Selected sources** 92, 97, 202, 287, 458, 580, 774, 1126, 1128.

B. Ibnu Utomo & J.L.C.H. van Valkenburg

**Vitex L.**

Sp. pl. 2: 638 ('938'; 1753); Gen. pl. ed. 5: 285 (1754).

VERBENACEAE

$x = 6, 8$ ; *V. negundo*:  $2n = 24, 26, 32, 34$ , *V. trifolia*:  $2n = 26, 32, 34$

**Major species** *Vitex negundo* L., *V. trifolia* L.

**Vernacular names** *Vitex* (En). Indonesia: le-gundi (general). Malaysia: laggundi (Peninsular), leban (Peninsular, Sarawak), kulim papa (Sabah). Papua New Guinea: garamut. Philippines: lagundi. Burma (Myanmar): kyetyo. Laos: 'sa 'khang', tin nok. Thailand: tinnok. Vietnam: b[if]n[h] linh.

**Origin and geographic distribution** *Vitex* comprises about 150 species and occurs throughout the tropics and subtropics with a few species extending to temperate regions. About 30 species occur in the Malesian region. The most important medicinal species, *V. negundo* and *V. trifolia*, are widely cultivated not only for their medicinal properties but also as ornamental and hedge plants, and have sometimes naturalized.

**Uses** The bark, leaves, flowers, fruits or roots of various *Vitex* species are used as a general tonic, anthelmintic and in the treatment of gastro-intestinal disorders. The therapeutical applications of *V. negundo* and *V. trifolia* are very similar, although in various countries different parts of the plant are used for the same ailments. General applications are as an anodyne, febrifuge, expectorant and diuretic. A poultice of the leaves is applied in rheumatic pains, inflammations and sprains. An infusion of the leaves is used in the treatment of dermatitis and eczema. The leaves are traditionally placed between books and clothes

as well as in rice stores to ward off insects. *V. altissima* L.f., *V. parviflora* A.L. Juss. and *V. pinnata* L. are also used in traditional medicine in South-East Asia but their primary use is for their timber. In the Philippines *V. negundo* is applied in veterinary medicine to treat internal parasites.

**Production and international trade** In the Philippines *V. negundo* is grown as one of 3 major species on government plantations totalling 3 ha, with an annual yield of 600 kg of powdered drugs. Private plantations have been established alongside these projects.

**Properties** In experiments on cat trachea leaf extracts of *V. negundo* showed bronchial-relaxing properties. The active constituents are probably the flavonoids casticin, chrysophenol D, luteolin and isoorientin, and p-hydroxybenzoic acid.

A double-blind placebo-controlled trial was carried out to investigate the effects of lagundi (*V. negundo*) tablets in the control of non-bacterial cough in children: the clinical response was assessed subjectively and objectively and the pulmonary function was also measured. It was found that the *V. negundo* tablets significantly improved the subjective assessments of frequency of cough and colour of sputum in children older than 7 years. In younger children, they were no better than the placebo in terms of the subjective clinical parameters used. No side-effects were reported and the tablet was acceptable in its taste and smell.

An ethanol extract (50%) of *V. trifolia* shows antispasmodic and antihistamine properties. The essential oil contains several terpenes, including cineol, terpineol and  $\alpha$ -pinene.

An ethanol extract and a cold aqueous infusion of leaves of *V. leucoxydon* L.f., an Indian species, were evaluated in a battery of tests to define its activity profile. The cold aqueous solution depressed SMA, antagonized d-amphetamine stereotypy and oxotremorine tremors, shortened the duration of mice immobility in a behavioural 'despair' test and lowered serum total cholesterol level. The ethanol extract showed significant inhibition of carrageenin-induced paw oedema and granulation tissue formation in rats. Suppression of acetic acid writhing was observed with both the ethanol extract and the aqueous solution. The LD<sub>50</sub> value of the ethanol extract was over 3000 mg/kg (intraperitoneal) and that of the aqueous solution 800–1200 mg/kg.

The antifungal properties of leaf extracts of *V. negundo* have been confirmed experimentally both in vitro and in vivo. Antiviral properties against

several viral diseases in rice have also been experimentally confirmed.

The ability of the flavonoid-rich fraction (5,7,3'-trihydroxy,6,8,4'-trimethoxy flavones) of *V. negundo* seeds to antagonize the androgen action of exogenous testosterone propionate on the male reproductive system has been confirmed in experiments with castrated prepubertal and intact adult dogs. Similar effects were observed in experiments with male rats. An alcohol extract from *V. negundo* seeds obtained by cold maceration was found effective as a hepatoprotective against liver damage induced by carbon tetrachloride.

A crude drug extract of *V. trifolia* leaves mediated a significant increase in lifespan in mice bearing sarcoma 180 cells, indicating potential antitumour activity. Similarly, expressed leaf juice from *V. negundo* showed antitumour activity in mice. The incidence of skin tumours initiated by dimethylbenzanthracene and promoted by croton oil was reduced appreciably.

Ecdysteroids are found in the bark of several *Vitex* species, e.g. *V. glabrata* and *V. pinnata*. The bark of these species is used as an anthelmintic and a remedy for gastro-intestinal disorders.

Leaf extracts of *V. negundo* show promise as a pesticide for integrated pest management. The insecticidal properties include effects on *Culex* and *Anopheles*, well-known vectors of malaria. Furthermore, dried and pulverized leaves or leaf extracts can be successfully applied in pest management during storage of potatoes, cereals and pulses. Oils from the leaves of *V. negundo* and *V. trifolia* show considerable mosquito repellent activity. The active principle in the leaves of *V. trifolia* has been identified as rotundinal, a cycloterpene aldehyde.

**Description** Evergreen or deciduous shrubs or small to medium-sized trees, sometimes large, up to 45 m tall; bole crooked to straight, up to 125(–200) cm in diameter, usually without buttresses but sometimes with distinct buttresses, often deeply fluted; bark surface rather smooth to shallowly fissured or flaky, pale grey to pale yellowish-brown, inner bark pale yellow to bright orange; crown often spreading. Leaves opposite or in whorls of 3, palmately compound with 3–7(–9) leaflets, rarely reduced to 1 leaflet, without stipules; leaflets entire, dentate or lobed. Inflorescence terminal or axillary, cymose, the cymes sessile or pedunculate, solitary or arranged in racemes, thyrses or panicles. Flowers bisexual; calyx campanulate to tubular, 5-lobed to truncate; corolla usually with a short tube, 2-lipped, upper

lip 2-fid, lower lip 3-fid, often pubescent outside, white to blue or violet or rarely yellowish; stamens 4, didynamous, inserted on the corolla tube, exerted; ovary superior, usually first 2-locular and later 4-locular with a single ovule in each cell, style 1, filiform, stigma bifid. Fruit a juicy or dry drupe, seated on the often enlarged calyx, generally with one 4-seeded pyrene. Seeds obovoid or oblong, without endosperm. Seedling with epigeal germination; cotyledons emergent, leafy; hypocotyl elongated; leaves opposite, duplicate, first ones simple and with toothed margins.

**Growth and development** Young *Vitex* trees grow rather slowly. *V. glabrata* trees showed a mean annual diameter increment of 0.8 cm in Burma (Myanmar), and *V. quinata* trees 1.2 cm in Java. One-year-old seedlings of the latter may reach 2 m in height in Java, and the mean annual height increment for the first 15 years after planting is 1 m. Usually flowering is during the rainy season and fruits ripen within a few months. In Java, trees of *V. quinata* do not start flowering until 11–12 years old. In the Philippines, *V. negundo* and *V. trifolia* flower year-round. In India, *V. negundo* flowers twice a year, once in July–November and again in March–May. Its flowers open from 8.30–13.00 h and are visited by various insects, some of which puncture the unopened buds to collect nectar. The flowers last for only 24 hours.

**Other botanical information** *Vitex* is generally placed in the subfamily *Viticoideae* and is probably most closely related to *Premna* which has simple leaves and very small flowers. *Vitex* might be confused with *Teijsmanniodendron* which differs, however, in the swollen apex of petioles and petiolules and the 2-celled ovary and fruit. Numerous subspecific taxa that are mainly varieties and formae have been described within many species of *Vitex*, but the usefulness of distinguishing them is questionable. *V. negundo* closely resembles *V. trifolia* but can be distinguished by its long-petioluled median leaflet and 3–5 leaflets.

**Ecology** Most *Vitex* occur in comparatively dry regions with a prominent dry season, often in lowland deciduous forest on rocky ground, on grassy slopes and on dry limestone soils, but sometimes also in littoral rain forest or in hill forest, occasionally up to 2000 m altitude.

**Propagation and planting** *Vitex* can be propagated by seed, but germination of *V. trifolia* seed is reported to be difficult. *V. quinata* has 7800–8200 dry fruits per kg. *V. negundo* can be propagated by cuttings 20 cm long which had 60% rooting success without the use of growth hor-

mones; soaking the cuttings in naphthalene acetic acid at a concentration of 0.1–1.0 mg/l increased rooting to almost 100%. In the Philippines lagundi (*V. negundo*) for medicinal purposes is planted at a spacing of 2 m × 4 m. When nursery stock is transplanted, 25 g of ammonium sulphate is applied directly to the planting hole. In India, *V. negundo* has been planted at an experimental scale for the production of firewood with 25 000 plants/ha. Planting should be done early in the rainy season. Generally *V. quinata* is planted at 1 m × 3 m in Java, occasionally at 1 m × 1–2 m on very fertile soils.

**In vitro production of active compounds** Tissue culture of *V. negundo* in the Philippines produced callus with a high content of therapeutic substances, in particular fats.

**Husbandry** *V. negundo* readily produces root suckers and for maximum biomass production can be managed as coppice. Its twigs are usually not browsed. It grows best in full sun and the content of therapeutically active essential oils is highest under these circumstances. Optimum growth is achieved by applying NPK 14-14-14 at a dosage of 20 g/plant.

**Diseases and pests** In the Philippines, circular leaf spot caused by *Corynespora* sp. severely damages *V. negundo* plantations locally, especially in the rainy season. *Cercospora* leaf spot and leaf blight caused by an unknown agent have also been observed. Thrips, lepidopterous defoliators and leaffolders are economically important pests of *V. negundo*. In Java, *Cromerus kalshoveni*, a top-sucking insect, has been observed on *V. quinata*.

**Harvesting** In the Philippines *V. negundo* is harvested 3–9 times a year. Leaves of flowering plants contain significantly more essential oil than those of non-flowering plants. Only mature and healthy leaves are harvested; diseased and senescent ones are discarded.

**Yield** Steam distillation of *V. negundo* leaves gave a yield of 0.4% of essential oil, principally  $\alpha$ -pinene, camphene, caryophyllene and citral.

**Handling after harvest** The leaves of *V. negundo* may be dried and pulverized and as such applied in tablets and capsules, whereas the leaf extract is used in syrups. The fruits can also be dried and stored for longer periods. In the Philippines harvested leaves are air-dried and stirred constantly in containers with screened bottoms. The desired moisture content of 10% is attained within 3–4 days during the dry season and within 2 weeks during the wet season. Storing dried leaves for 2 months in sealed, clear polyethylene

bags under ambient conditions does not significantly reduce essential oil content.

**Genetic resources and breeding** Most *Vitex* species do not seem to be liable to genetic erosion as they are widespread and generally regenerate easily and abundantly after disturbance of the forest. The medicinally important *V. negundo* and *V. trifolia* are widely planted in hedges and for ornamental purposes. *V. negundo* is most probably an outbreeding species.

**Prospects** The application of *V. negundo* as a general tonic in syrups and tablets deserves further attention. The antitumour activity as observed in vitro seems promising. *V. negundo* and *V. trifolia* show considerable potential as a botanical pesticide that may be applied both indoors and outdoors.

**Literature** |1| Avadhoot, Y. & Rana, A.C., 1991. Hepatoprotective effect of *Vitex negundo* against carbon tetrachloride-induced liver damage. *Archives of Pharmacological Research* 14(1): 96-98. |2| Balboa, J.G. & Lim-Sylianco, C.Y., 1995. Effect of some medicinal plants on skin tumor promotion. *Philippine Journal of Science* 124(2): 203-207. |3| Bhargava S.K., 1989. Antiandrogenic effects of a flavonoid-rich fraction of *Vitex negundo* seeds, a histological and biochemical study in dogs. *Journal of Ethnopharmacology* 27(3): 327-340. |4| Dayrit, F.M., Lapid, M.R.G., Cagampang, J.V. & Lagurin, L.G., 1987. Phytochemical studies on the leaves of *Vitex negundo* L. ('lagundi'). I. Investigations of the bronchial relaxing constituents. *Philippine Journal of Science* 116(4): 403-410. |5| Lam, H.J., 1919. The Verbenaceae of the Malayan Archipelago. M. de Waal, Groningen, the Netherlands. pp. 164-214. |6| Makwana, H.G., Ravishankar, B., Shukla, V.J., Nair, R.B., Vijayan, N.P., Sasikala, C.K., Saraswathy, V.N. & Bhatt, S.V., 1994. General pharmacology of *Vitex leucoxylo* Linn leaves. *Indian Journal of Physiology and Pharmacology* 38(2): 95-100. |7| Moldenke, H.N. & Moldenke, A.L., 1983. Verbenaceae. In: Das-sanayake, M.D. & Fosberg, F.R. (Editors): A revised handbook to the flora of Ceylon. Amerind Publishing Co., New Delhi, India. pp. 348-388. |8| Suksamrarn, A. & Sommechai, C., 1993. Ecdysteroids from *Vitex pinnata*. *Phytochemistry* 32(2): 303-306. |9| Sunarno, B., Lemmens, R.H.M.J. & Ani binti Sulaiman, 1995. *Vitex* L. In: Lemmens, R.H.M.J., Soerianegara, I. & Wong, W.C. (Editors): *Plant Resources of South-East Asia No 5(2)*. Timber trees: Minor commercial timbers. Backhuys Publishers, Leiden, the Netherlands. pp. 502-509. |10| Sundarrao, K., Burrows, I., Kuduk,

M., Yi, Y.D., Chung, M.H., Suh, N.J. & Chang, I.M., 1993. Preliminary screening of antibacterial and antitumor activities of Papua New Guinean native medicinal plants. *International Journal of Pharmacognosy* 31(1): 3-6.

#### *Selection of species*

#### **Vitex glabrata R.Br.**

Prodr.: 512 (1810).

**Synonyms** *Vitex minahassae* Koord. (1898), *Vitex helogiton* K. Schumann (1905), *Vitex pentaphylla* Merr. (1909).

**Vernacular names** Indonesia: bihbul (Sundanese), gentileng, ketileng (Javanese). Philippines: bongoo (general), ampapalut (Balabac), talang-pulo (Camarines). Cambodia: popoul ach, popul tuk. Laos: 'khi<sup>2</sup> hen, 'khi<sup>2</sup> nok, tin nok. Thailand: khainao (general), khee hen (north-eastern), khom khwaan (central). Vietnam: ma, b[if]nh linh nh[awx]n.

**Distribution** India, Burma (Myanmar), Indo-China, Thailand, Peninsular Malaysia (rare), Java, the Philippines, Sulawesi, Timor, New Guinea and northern Australia.

**Uses** The root and bark are astringent. The bark is used as an anthelmintic and as a remedy for gastro-intestinal disorders. The fruit and bark are applied as a component of masticatories. The timber is used for house construction, furniture, cart-wheels and oars. The fruits are edible. In India (Assam), *V. glabrata* has been recommended for planting in tea plantations for shade and wood.

**Observations** A medium-sized tree up to 25 m tall, bole branchless for up to 15 m, up to 125 cm in diameter, bark surface smooth, ash-grey; leaflets 3-5(-6), elliptical-oblong to elliptical-obovate, glabrous above, except for the larger veins, glabrous to pubescent below, especially in the primary vein axils, petioluled, median leaflet 11-31 cm × 4-13.5 cm, with 12-17 pairs of lateral veins; cymes axillary, solitary, lax, 7-22 cm long; calyx 3-4 mm long, with 5 small teeth, corolla yellowish-white, tube villous inside, median segment of lower lip blue-violet; fruit ovoid or obovoid, 1.2-2.5 cm long, purplish-black when mature. *V. glabrata* occurs in mixed evergreen forest, often along forest edges, sometimes also in deciduous forest or grassland, up to 1000 m altitude. In Burma (Myanmar) it is often associated with teak (*Tectona grandis* L.f.).

**Selected sources** 97, 287, 580, 809, 810, 969, 1067, 1128, 1215, 1413, 1562.

**Vitex negundo L.**

Sp. pl. 2: 638 ('938'; 1753).

**Synonyms** *Vitex incisa* Lamk (1786), *Vitex paniculata* Lamk (1786), *Vitex leucoxyton* Blanco (1837).

**Vernacular names** Five-leaved chaste tree, horseshoe vitex (En). Indonesia: ai tuban (Ambon), lagundi laut laki-laki (Malay, Moluccas). Malaysia: lagundi, lemuning, lenggundi (Peninsular). Papua New Guinea: paparau (Kurtachi, Bougainville). Philippines: lagundi (Filipino), dangla (Iloko). Cambodia: trasiet. Thailand: khon thee khamoa (central), ku-no-kaa-mo (Malay, Pattani), kuuning (Malay, Narathiwat). Vietnam: ng[ux] tr[ar]o.

**Distribution** Eastern Africa and Madagascar to Iran, Afghanistan, Pakistan, India, Sri Lanka, Burma (Myanmar), Indo-China, China, Japan, Taiwan, Thailand, throughout the Malesian region, east to the Palau Islands, the Caroline Islands and the Mariana Islands. *V. negundo* is widely cultivated in Europe, Asia, North America and the West Indies, and has a tendency to escape and naturalize.

**Uses** Roots and leaves are applied in various manners as an anodyne, bitter tonic, expectorant and diuretic. As a febrifuge, the leaves can be employed as a poultice or in decoction. A decoction of the leaves is applied to wounds and ulcers, for aromatic baths, and internally as galactagogue, emmenagogue, antigestralgic, and against flatulence. In the Philippines the seeds are boiled in water and eaten, or the water is taken internally, to prevent the spread of toxins from poisonous bites of animals. Syrup, tablets and capsules prepared from leaves and flowering tops are given for coughs, colds, fever and asthma. In Papua New Guinea, sap from crushed heated leaves is diluted with water to treat coughs and sore throat. In India, the flowers are used in the treatment of diarrhoea, cholera and liver disorders, apparently for their astringent properties. *V. negundo* is often planted as a hedge or for ornamental purposes. The twigs are used for wattle-work and rough baskets.

**Observations** A deciduous shrub or small tree up to 8 m tall, bark surface slightly rough, peeling off in papery flakes, pale reddish-brown; leaflets 3-5, narrowly elliptical to ovate-lanceolate, minutely puberulous or glabrous above, densely tomentose or puberulent below, median leaflet 5-15 cm x 1-4 cm, with 2-12(-18) pairs of lateral veins, median leaflet on a 1-2.5 cm long petiolule, lateral ones sometimes sessile; cymes arranged in



*Vitex negundo* L. - 1, flowering twig; 2, flower; 3, opened corolla; 4, calyx and style; 5, fruit.

panicles which are terminal and axillary in the upper leaf axils; calyx 1-2 mm long, shortly 5-toothed, corolla blue-violet, villous inside; fruit globose to broadly ovoid, 3-6 mm long, purple or black when mature. *V. negundo* is very variable and many varieties and formae have been distinguished. It is often found gregariously in humid places or along watercourses, in waste places, thickets and mixed open forest, up to 1700 m altitude.

**Selected sources** 35, 84, 97, 100, 110, 145, 190, 202, 206, 213, 284, 287, 307, 312, 325, 327, 332, 358, 415, 567, 580, 597, 709, 809, 810, 890, 894, 921, 942, 943, 970, 971, 972, 1035, 1126, 1128, 1171, 1176, 1178, 1195, 1287, 1307, 1380, 1478, 1524, 1525, 1564.

**Vitex quinata (Lour.) F.N. Williams**

Bull. Herb. Boiss. 2, 5: 431 (1905).

**Synonyms** *Vitex heterophylla* Roxb. (1832), *Vitex sumatrana* Miq. (1861), *Vitex celebica* Koord. (1898).

**Vernacular names** Orange-barked vitex (En).

Indonesia: ketileng (Javanese), ki bangbara (Sundanese), gofasa (Malay, northern Sulawesi). Malaysia: leban bunga, leban tandok, merboh (Peninsular). Philippines: kalipapa (general). Burma (Myanmar): kyetyo-po. Thailand: ee pae, makhang, maak lek maak noi (north-eastern). Vietnam: c[aa]y m[aj]n kinh.

**Distribution** India, the Andaman Islands, Burma (Myanmar), Indo-China, southern China, Thailand, Peninsular Malaysia, Sumatra, Java, the Philippines, Sulawesi, Timor and the Moluccas.

**Uses** The bark is used as a tonic and as a stomachic; an infusion of it is drunk to stimulate the appetite. The timber is used for house construction and boat building.

**Observations** A medium-sized to large tree up to 45 m tall, bole up to 100(-150) cm in diameter, often fluted, bark surface shallowly fissured, pale grey, inner bark bright orange; leaflets (3-)5, lanceolate to ovate, glabrous or pubescent on the veins only, petioluled, median leaflet 5-13 cm × 2.5-6 cm, with 9-12 pairs of lateral veins; cymes arranged in large panicles which are terminal or axillary in the upper leaf axils; calyx 3-4 mm long, almost truncate to minutely 5-toothed, corolla pale yellowish to purplish, glabrous to sparsely pubescent inside; fruit subglobose to pear-shaped, 6-12 mm long, blackish when mature. *V. quinata* occurs scattered in forest up to 1400 m altitude.

**Selected sources** 97, 284, 554, 580, 673, 809, 810, 992, 1035, 1067, 1128, 1413, 1564.

### **Vitex trifolia L.**

Sp. pl. 2: 638 ('938'; 1753).

**Synonyms** *Vitex rotundifolia* L.f. (1781), *Vitex repens* Blanco (1837), *Vitex lagundi* Ridley (1906).

**Vernacular names** Common blue vitex, hand of Mary, three-leaved chaste tree (En). Indonesia: legundi (Javanese), galumi (Sumbawa), lagundi (Sumatra). Malaysia: lagundi, lenggundi, muning (Peninsular). Papua New Guinea: pitipitikoto (Gunantuna, New Britain). Philippines: lagunding-dagat (Filipino), dangla (Iloko), tigau (Sulu). Laos: kok pa pay, 'phi 'sua<sup>2</sup> noy<sup>2</sup>. Thailand: khon dinso, khon thiso (central), phae suea noi (north-eastern). Vietnam: d[e]n ba l[as], m[aj]n kinh, quan [aa]m.

**Distribution** From southern Africa, Madagascar and Mauritius to Afghanistan, India, Sri Lanka, Burma (Myanmar), Indo-China, southern China, Japan, Thailand, throughout the Malesian region, south to northern Australia and east to New Caledonia.

**Uses** The uses of *V. trifolia* are very similar to *V. negundo*. A poultice of leaves is used to treat rheumatism, contusions, swollen testicles, and as a discutient in sprains. An infusion of the boiled roots is regarded as diaphoretic and diuretic, and is widely drunk in cases of fever and after childbirth. In Malaysia, various parts of the plants are considered a panacea for a wide variety of illnesses ranging from headache to tuberculosis. In Indonesia, the leaves are used in medicinal baths and a tincture or decoction of them for intestinal complaints, whereas the fruits are used as an anthelmintic. In the Bismarck Archipelago, the sap from crushed heated leaves is diluted with water and drunk to relieve headaches. In Vietnam, a decoction of dried fruits is given in the treatment of common cold, headache, watery eyes and mastitis. In Thailand, the fruits are used to treat asthmatic cough and haemorrhoids, and the root is applied in the treatment of liver diseases. *V. trifolia* is often used as a hedge plant, although it may trigger various allergic reactions (sneezing, respiratory problems, dizziness, headache, nausea) to people trimming or pruning such hedges.

**Observations** A shrub up to 6 m tall; leaflets (1-)3, glabrous above (except for the midrib), densely greyish puberulous below, median leaflet oblong-elliptical to obovate, 2.5-9.5 cm × 1.5-4 cm, with 6-13 pairs of lateral veins, on a 1-6 mm long petiolule, lateral leaflets sessile or subsessile; cymes terminal and axillary, arranged in panicles; calyx 3-5 mm long, obscurely 2-lipped, with 5 small teeth, corolla blue to purple or violet, throat villous inside; fruit globose to ovoid, 5-6 mm long, black or bluish-black when mature. *V. trifolia* is found in teak forest, secondary forest and thickets up to 1100 m altitude, but also in mangrove forest and along the shore. The phenotypical variation observed between these habitats is given specific or subspecific rank by various authors.

**Selected sources** 97, 190, 202, 284, 364, 580, 597, 809, 810, 921, 967, 972, 993, 1035, 1128, 1178, 1287, 1380, 1412, 1415, 1508, 1525, 1553, 1564, 1570.

E.P. Capareda



## Literature

1. Abburra, R.E., Zygodlo, J.A. & Guzman, C.A., 1992. Fatty acids variation in Sapindaceae. *Biochemical Systematics and Ecology* 20(5): 469–471.
2. Abdul, K.M. & Ramchender, R.P., 1995. Modulatory effect of plumbagin (5-hydroxy-2-methyl-1,4-naphthoquinone) on macrophage functions in BALB/c mice. I. Potentiation of macrophage bactericidal activity. *Immunopharmacology* 30(3): 231–236.
3. Abdullah, T.L., Ahmad, S.H. & Rejab, N.A., 1993. Determination of floral stages and packaging methods for prolonged storage of *Jasminum multiflorum*. In: Palevitch, D., Simon, J.E. & Mathé, A. (Editors): Raw material production, product introduction. Proceedings of the First World Congress on Medicinal and Aromatic Plants for Human Welfare, WOCMAP. Maastricht, Netherlands, July 19–25, 1992. *Acta Horticulturae* 331: 325–329.
4. Abraham, M., Devi, N.S. & Sheela, R., 1979. Inhibiting effect of jasmine flowers on lactation. *Indian Journal of Medical Research* 69: 88–92.
5. Abrams, G.A. & Fallon, M.B., 1998. Treatment of hepatopulmonary syndrome with *Allium sativum* L. (garlic): a pilot trial. *Journal of Clinical Gastroenterology* 27(3): 232–235.
6. Acevedo, R.A., Santos, A.C. & Pabatao, P., 1970. A diterpene from *Tinospora rumphii* Boerl. *Philippine Journal of Science* 97(3): 269–275.
7. Acharya, B.M. & Kumer, K.A., 1984. Chemical examination of the bark of *Ficus hispida* Linn. *Current Science, India* 53(19): 1034–1035.
8. Acharya, T.K. & Chatterjee, I.B., 1975. Isolation of chrysophanic acid-9-anthrone, the major antifungal principle of *Cassia tora*. *Lloydia* 38(3): 218–220.
9. Achrekar, S., Kaklij, G.S., Pote, M.S. & Kelkar, S.M., 1991. Hypoglycemic activity of *Eugenia jambolana* and *Ficus bengalensis*: mechanism of action. *In-Vivo* 5(2): 143–147.
10. Acton, N., Karle, J.M. & Miller, R.E., 1993. Synthesis and antimalarial activity of some 9-substituted artemisinin derivatives. *Journal of Medicinal Chemistry* 36: 2552–2557.
11. Acton, N. & Klayman, D.L., 1985. Artemisitene, a new sesquiterpene lactone endoperoxide from *Artemisia annua*. *Planta Medica* 51: 441–442.
12. Acton, N. & Klayman, D.L., 1987. Conversion of artemisinin to iso-artemisitenone and to 9-epi-artemisinin. *Planta Medica* 53: 266–268.
13. Adam, G., Huong, H.T. & Khoi, N.H., 1979. The constituents of the Vietnamese drug plant *Solanum verbascifolium* L. *Planta Medica* 36: 238–239.
14. Addy, M.E. & Awumey, E.M., 1984. Effects of the extracts of *Desmodium adscendens* on anaphylaxis. *Journal of Ethnopharmacology* 11(3): 283–292.
15. Addy, M.E. & Burka, J.F., 1988. Effect of *Desmodium adscendens* fractions

- on antigen- and arachidonic acid-induced contractions of guinea pig airways. *Canadian Journal of Physiology and Pharmacology* 66(6): 820–825.
16. Addy, M.E. & Burka, J.F., 1990. Effect of *Desmodium adscendens* fraction 3 on contractions of respiratory smooth muscle. *Journal of Ethnopharmacology* 29(3): 325–335.
  17. Addy, M.E. & Dzandu, W.K., 1986. Dose-response effects of *Desmodium adscendens* aqueous extract on histamine response, content and anaphylactic reactions in the guinea pig. *Journal of Ethnopharmacology* 18(1): 13–20.
  18. Adesogan, E.K. & Okunade, A.L., 1979. A new flavone from *Ageratum conyzoides*. *Phytochemistry* 18(11): 1863–1864.
  19. Adinarayana, D., Narayana Rao, K. & Sarada, M., 1987. A new aromatic nitrosulfone from *Cardiospermum corindum*. *Journal of Natural Products* 50(4): 620–622.
  20. Adinarayana, D. & Syamasundar, K.V., 1982. Occurrence of a rare dihydroxyflavone, 2'-O-glucosylvitexin in *Desmodium triflorum*. *Current Science* 51(19): 936–937.
  21. Afaq, S.H. & Khan, M.M.A., 1994. Predicting solasodine content of *Solanum nigrum* – a statistical model. In: *Ethnobiology in human welfare: abstracts of the Fourth International Congress of Ethnobiology*, Lucknow, Uttar Pradesh, India, 17–21 November, 1994. National Botanical Research Institute, Lucknow, India. p. 97.
  22. Afriastini, J.J., 1986. Bertanam kencur [Growing kencur]. P.T. Penebar Swadaya, Jakarta, Indonesia. 20 pp.
  23. Afsharypuor, S., Mostajeran, A. & Mokhtary, R., 1995. Variation of scopolamine and atropine in different parts of *Datura metel* during development. *Planta Medica* 61(4): 383–384.
  24. Agarwal, R., Singh, R., Siddiqui, I.R. & Singh, J. 1995. Triterpenoid and prenylated phenol glycosides from *Blumea lacera*. *Phytochemistry* 38(4): 935–938.
  25. Agrawal, S., Chauhan, S. & Mathur, R., 1986. Antifertility effects of embelin in male rats. *Andrologia* 18(2): 125–131.
  26. Ahammed, J. & Paria, N., 1996. Systematic value of seedling morphology in some Indian Asteraceae. *Acta Botanica Indica* 24: 49–55.
  27. Ahmad, F.B. & Holdsworth, D.K., 1994. Medicinal plants of Sabah, Malaysia. Part II. The Muruts. *International Journal of Pharmacognosy* 32(4): 378–383.
  28. Ahmad, F.B. & Holdsworth, D.K., 1994. Medicinal plants of Sarawak, Malaysia. Part I. The Kedayans. *International Journal of Pharmacognosy* 32(4): 384–387.
  29. Ahmad, F.B. & Holdsworth, D.K., 1995. Medicinal plants of Sabah, Malaysia. Part III. The Rungus people of Kudat. *International Journal of Pharmacognosy* 33(3): 262–264.
  30. Ahmad, M.S., Ahmad, M.U. & Osman, S.M., 1980. A new hydroxyolefinic acid from *Plantago major* seed oil. *Phytochemistry* 19(10): 2137–2139.
  31. Ahmed, S.M. & Ahamad, A., 1992. Efficacy of some indigenous plants as pulse protectants against *Callosobruchus chinensis* L. infestation. *International Pest Control* 34(2): 54–56.
  32. Airy Shaw, H.K., 1972. The Euphorbiaceae of Siam. *Kew Bulletin* 26: 191–363.

33. Airy Shaw, H.K., 1980. The Euphorbiaceae of New Guinea. Kew Bulletin Additional Series VIII. Her Majesty's Stationery Office, London. 243 pp.
34. Airy Shaw, H.K., 1981. The Euphorbiaceae of Sumatra. Kew Bulletin 36: 239-374.
35. Aiyanathan, K.E.A. & Narayanasamy, P., 1988. Effect of antiviral principles on rice tungro virus infection. Indian Journal of Virology 4(1-2): 97-99.
36. Akao, T., Che, Q.M., Kobashi, K., Hattori, M. & Namba, T., 1996. A purgative action of barbaloin is induced by Eubacterium sp. strain BAR, a human intestinal anaerobe, capable of transforming barbaloin to aloë-emodin anthrone. Biological and Pharmaceutical Bulletin 19(1): 136-138.
37. Akbarsha, M.A., Manivannan, B., Hamid, K.S. & Vijayan, B., 1990. Anti-fertility effect of *Andrographis paniculata* (Nees) in male albino rats. Indian Journal of Experimental Biology 28(5): 421-426.
38. Akhila, A., Rani, K. & Thakur, R.S., 1990. Biosynthesis of artemisinic acid in *Artemisia annua*. Phytochemistry 29: 2129-2132.
39. Akhila, A., Thakur, R.S. & Popli, S., 1987. Biosynthesis of artemisinin in *Artemisia annua*. Phytochemistry 26: 1927-1930.
40. Akhtar, M.S., Khan, Q.M. & Khaliq, T., 1984. Effects of *Euphorbia prostrata* and *Fumaria parviflora* in normoglycaemic and alloxan-treated hyperglycaemic rabbits. Planta Medica 50(2): 138-142.
41. Akhtar, M.S., Khan, Q.M. & Khaliq, T., 1985. Effects of *Portulaca oleracea* (Kulfa) and *Taraxacum officinale* (Dhudhal) in normoglycaemic and alloxan-treated hyperglycaemic rabbits. Journal of Pakistan Medical Association 35: 207-210.
42. Akhtar, M.S. & Munir, M., 1989. Evaluation of the gastric antiulcerogenic effect of *Solanum nigrum*, *Brassica oleracea* and *Ocimum basilicum* in rats. Journal of Ethnopharmacology 27(1-2): 163-176.
43. Akihisa, T., Yasukawa, K., Kimura, Y., Takido, M., Kokke, W.C.M.C. & Tamura, T., 1994a. Five D:C-friedo-oleanane triterpenes from the seeds of *Trichosanthes kirilowii* Maxim. and their anti-inflammatory effects. Chemical and Pharmaceutical Bulletin 42(5): 1101-1105.
44. Akihisa, T., Yasukawa, K., Kimura, Y., Takido, M., Kokke, W.C.M.C. & Tamura, T., 1994b. 7-Oxo-10- $\alpha$ -cucurbitadienol from the seeds of *Trichosanthes kirilowii* and its anti-inflammatory effect. Phytochemistry 36(1): 153-157.
45. Alagesaboopathi, C. & Balu, S., 1996. Seed germination studies on *Andrographis paniculata* Nees. Journal of Living World 3(1): 46-49.
46. Alam, M.K., 1992. Medical ethnobotany of the Marma tribe of Bangladesh. Economic Botany 46(3): 330-335.
47. Alexandre, M.A.V., Noronha, A.B. & Vicente, M., 1987. Acao de inibidores naturais sobre duas viroses do feijoeiro: mosaico dourado e mosaico do fumo 'strain' adaptado as leguminosas [Action of natural inhibitors on two virus diseases of bean, bean golden mosaic virus and tobacco mosaic virus legume strain]. Fitopatologia Brasileira 12(3): 202-205.
48. Ali, L., Kahna, A.K., Mamun, M.I., Mosihuaman, M., Nahar, N., Nur-e-Alam, M. & Rokeya, B., 1993. Studies on hypoglycemic effects of fruit pulp, seed, and whole plant of *Momordica charantia* on normal and diabetic model rats. Planta Medica 59(5): 408-412.
49. Allen, E.F., 1958. Notes on the cultivation of *Rauwolfia* in Malaya. Malayan Agricultural Journal 41(2): 100-105.

50. Allen, O.N. & Allen, E.K., 1981. The Leguminosae. A source book of characteristics, uses and nodulation. MacMillan Publishers, London, United Kingdom. 812 pp.
51. Alsaadawi, I.S., Sakeri, F.A.K. & Al-Dulaimy, S.M., 1990. Allelopathic inhibition of *Cynodon dactylon* (L.) Pers. and other plant species by *Euphorbia prostrata* L. *Journal of Chemical Ecology* 16(9): 2747–2754.
52. Alvarez, L., Marquina, S., Villareal, M.L., Alonso, D., Arranda, E. & Delgado, G., 1996. Bioactive polyacetylenes from *Bidens pilosa*. *Planta Medica* 62(4): 355–357.
53. Amaral, A. & Takaki, M., 1993. Weed seed germination. III. *Bidens pilosa* L. *Arquivos de Biologia e Tecnologia (Curitiba)* 36(2): 401–408.
54. Ambike, S.H. & Rajarama Rao, E.R., 1967. Studies on a phytosterolin from the bark of *Ficus religiosa*. *Indian Journal of Pharmacy* 29: 91–94.
55. Ameen, M.U. & Sultana, P., 1977. Biology of the bag-worm moth *Eumeta crameri* Westwood (Lepidoptera: Psychidae) from Dacca, Bangladesh. *Journal of Natural History* 11(1): 17–24.
56. Ameen Mahmudul & Shahjahan Reza Md, 1987. Lethal effect of *Derris elliptica* (Benth.) root on the catfish (*Heteropneustes fossilis* Bloch). *Bangladesh Journal of Agriculture* 12(1): 19–26.
57. Amin, K.M.Y., Ahmed, S. & Khan, N.A., 1994. Anti-nephrotic syndrome ethnic drug Bishiri Booti (*Aerva lanata*) – experimental study of relevant pharmacological actions. *Ethnobiology in human welfare: abstracts of the Fourth International Congress of Ethnobiology, Lucknow, Uttar Pradesh, India, 17–21 November, 1994. National Botanical Research Institute, Lucknow, India. p. 94.*
58. Ammon, H.P.T. & Müller, A.B., 1985. Forskolin: From an Ayurvedic remedy to a modern agent. *Planta Medica* 51: 473–477.
59. Ammon, H.P.T., Safayhi, H., Mack, T. & Sabieraj, J., 1993. Mechanism of anti-inflammatory actions of curcumin and boswellic acids. *Journal of Ethnopharmacology* 38(2–3): 113–119.
60. Ammon, H.P.T. & Wahl, M.A., 1991. Pharmacology of *Curcuma longa*. *Planta Medica* 57(1): 1–7.
61. Anand, H., Roberts, P.J. & Lopez-Colome, A.M., 1985. Excitatory amino acids in the chick retina: possible involvement of cyclic guanosine monophosphate. *Neuroscience Letters* 58(1): 31–36.
62. Anderson, E.F., 1986. Ethnobotany of hill tribes of Northern Thailand. I. Medicinal plants of Akha. *Economic Botany* 40(1): 38–53.
63. Anderson, E.F., 1986. Ethnobotany of hill tribes of Northern Thailand. II. Lahu medicinal plants. *Economic Botany* 40(4): 442–450.
64. Anderson, E.F., 1993. Plants and people of the Golden Triangle. *Ethnobotany of the hill tribes of Northern Thailand. Dioscorides Press, Portland, United States. 279 pp.*
65. Anderson, J.D., Mandava, N. & Garrett, S., 1975. Inhibition of hormone-induced ethylene synthesis by the indole plant-growth inhibitor from *Abrus precatorius* seeds. *Plant and Cell Physiology* 16(2): 233–236.
66. Anderson, L.A., Schultz, R.A., Joubert, J.P., Prozesky, L., Kellerman, T.S., Erasmus, G.L. & Procos, J., 1983. Krimpsieke and acute cardiac glycoside poisoning in sheep caused by bufadienolides from the plant *Kalanchoe lanceolata* (Forssk.) Pers. *Onderstepoort Journal of Veterinary Research*

- 50: 295-300.
67. Anderson, L.A., Steyn, P.S. & Heerden, F.R., 1984. The characterization of two novel bufadienolides from *Kalanchoe lanceolata* (Forssk.) Pers. *Journal of the Chemical Society (Perkin Transactions I)*: 1573-1575.
  68. Ang, H.H., Chan, K.L. & Mak, J.W., 1995. Effect of 7-day daily replacement of culture medium containing *Eurycoma longifolia* Jack constituents on the Malaysian *Plasmodium falciparum* isolates. *Journal of Ethnopharmacology* 49: 171-175.
  69. Ang, H.H. & Sim, M.K., 1998a. *Eurycoma longifolia* Jack and orientation activities in sexually experienced male rats. *Biological and Pharmaceutical Bulletin* 21(2): 153-155.
  70. Ang, H.H. & Sim, M.K., 1998b. *Eurycoma longifolia* increases sexual motivation in sexually naive rats. *Archives of Pharmaceutical Research* 21(6): 779-781.
  71. Anjani, K., 1990. Regeneration of the forest tree *Strychnos nux-vomica* L. through tissue culture. *Indian Journal of Forestry* 13(3): 263-265.
  72. Anwar Masood, Sultanul Haq & Saxena, S.K., 1985. Effect of some plant extracts on the larval hatching of *Meloidogyne incognita* (Kofoid & White, 1919) Chitwood, 1949-III. *Indian Forester* 111(10): 841-845.
  73. Apisariyakul, A., Vanittanakom, N. & Buddhasukh, D., 1995. Antifungal activity of turmeric oil extracted from *Curcuma longa* (Zingiberaceae). *Journal of Ethnopharmacology* 49(3): 163-169.
  74. Arano, H. & Saito, H., 1979. Cytological studies in family Umbelliferae 4. Karyotypes in genus *Angelica* 2. *Kromosomo (Tokyo)* 2(15-16): 417-426.
  75. Areekul, S. & Sinchaisri, P., 1988. Effects of Thai plant extracts on the oriental fruit fly (*Darcus dorsalis*) II. Repellency test. *Witthayasan Kasetart Sakha Witthayasat* 22(1): 56-61. (in Thai)
  76. Ariga, T. & Kase, H., 1986. Composition of essential oils of the genus *Allium* and their inhibitory effect on platelet aggregation. *Bulletin of the College of Agriculture and Veterinary Medicine of Nihon University* 43: 170-175. (in Japanese)
  77. Asano, J., Chiba, K., Tada, M. & Yoshii, T., 1996. Antiviral activity of lignans and their glycosides from *Justicia procumbens*. *Phytochemistry* 42(3): 713-717.
  78. Atal, C.K. & Kapur, B.M. (Editors), 1982. Cultivation and utilization of medicinal plants. Regional Research Laboratory, Council of Scientific and Industrial Research, Jammu-Tawi, India. 877 pp.
  79. Atta, A.H. & Alkofahi, A., 1998. Anti-nociceptive and anti-inflammatory effects of some Jordanian medicinal plant extracts. *Journal of Ethnopharmacology* 60(2): 117-124.
  80. Audy, P., le Quere, D., Leclerc, D. & Asselin, A., 1990. Electrophoretic forms of lysozyme activity in various plant species. *Phytochemistry* 29(4): 1143-1159.
  81. Augusti, K.T., Daniel, R.S., Cherian, S., Sheela, C.G. & Nair, C.R., 1994. Effect of leucopelargonin derivative from *Ficus bengalensis* Linn. on diabetic dogs. *Indian Journal of Medical Research* 99: 82-86.
  82. Augusti, K.T. & Sheela, C.G., 1996. Antiperoxide effect of S-allyl cysteine sulfoxide, an insulin secretagogue, in diabetic rats. *Experientia* 52(2): 115-120.

83. Auvin-Guette, C., Baraguey, C., Blond, A., Pousset, J.L. & Bodo, B., 1997. Cyclogossine B, a cyclic octapeptide from *Jatropha gossypifolia*. *Journal of Natural Products* 60: 1155–1157.
84. Avadhoot, Y. & Rana, A.C., 1991. Hepatoprotective effect of *Vitex negundo* against carbon tetrachloride-induced liver damage. *Archives of Pharmacological Research* 14(1): 96–98.
85. Avalos, A.A., Diaz, M.Q. & Guerrero, M.C., 1984. Influence of extracts from leaves and stem of *Bidens pilosa* on experimental ulcerogenesis in rats. *Revista Farmaceutica de Cuba* 18(2): 143–150.
86. Avé, W. & Sunito, S., 1990. Medicinal plants of Siberut. WWF International, Gland, Switzerland. 186 pp.
87. Avery, A.G., Satina, S. & Rietsema, J. (Editors), 1959. *Blakeslee: the genus Datura*. The Ronald Press Company, New York, United States. 289 pp.
88. Avirutnant, W. & Pongpan, A., 1983. The antimicrobial activity of some Thai flowers and plants. *mahidol University Journal of Pharmaceutical Sciences* 10(3): 81–86.
89. Ayeni, A.O., 1985. Observations on the vegetative growth pattern of speargrass (*Imperata cylindrica* (L.) Beauv.). *Agriculture, Ecosystems and Environment* 13: 301–307.
90. Azuine, M.A. & Bhide, S.V., 1994. Adjuvant chemoprevention of experimental cancer: catechin and dietary turmeric in forestomach and oral cancer models. *Journal of Ethnopharmacology* 44(3): 211–217.
91. Babeley, G.S. & Kandya, A.K., 1988. On finding out some suitable pretreatments for *Cassia fistula* Linn. seeds. *Journal of Tropical Forestry* 4(2): 147–154.
92. Backer, C.A., 1928–1934. *Onkruidflora der Javasche suikerrietgronden*. Handboek ten dienste van de suikerriet-cultuur en de rietsuiker-fabricage op Java [Weed flora of Javanese sugar-cane fields. Handbook for the cultivation of sugar-cane and manufacturing of cane-sugar in Java]. Vereeniging het Proefstation voor de Java-Suikerindustrie, Pasuruan, Indonesia. 4 volumes. 970 pp.
93. Backer, C.A., 1949. *Amaranthaceae*. In: van Steenis, C.G.G.J. (General editor): *Flora Malesiana*. Series 1, Vol. 4. Noordhoff-Kolff, Djakarta, Indonesia. pp. 69–98.
94. Backer, C.A., 1951. *Cannabaceae*. In: van Steenis, C.G.G.J. (General editor): *Flora Malesiana*. Series 1, Vol. 4. Noordhoff-Kolff, Djakarta, Indonesia. pp. 222–223.
95. Backer, C.A., 1951. *Crassulaceae*. In: van Steenis, C.G.G.J. (General editor): *Flora Malesiana*. Series 1, Vol. 4. Noordhoff-Kolff, Djakarta, Indonesia. pp. 197–202.
96. Backer, C.A., 1951. *Phytolaccaceae*. In: van Steenis, C.G.G.J. (General editor): *Flora Malesiana*. Series 1, Vol. 4. Noordhoff-Kolff, Djakarta, Indonesia. pp. 228–232.
97. Backer, C.A. & Bakhuizen van den Brink Jr, R.C., 1963–1968. *Flora of Java*. 3 volumes. Noordhoff, Groningen, the Netherlands. Vol. 1 (1963) 647 pp., Vol. 2 (1965) 641 pp., Vol. 3 (1968) 761 pp.
98. Baczynsky, W.O.T. & Zimmerman, A.M., 1983. Effects of  $\Delta^9$ -tetrahydrocannabinol, cannabinol and cannabidiol on the immune system in mice. *Pharmacology* 26: 1–11.

99. Badan Litbang Pertanian & APPPI, 1995. Petunjuk kultur teknis tanaman kina [Technical instructions for the planting of kina]. Jakarta, Indonesia. 143 pp.
100. Badola, H.K. & Badoni, A.K., 1990. Effect of Stik and GA3 on vegetative propagation in stem-cuttings of *Vitex negundo* Linn. during winter. *Indian Forester* 116(12): 980-983.
101. Baerts, M. & Lehmann, J. 1991. Plantes medicinales veterinaires de la region des Cretes Zaire-Nil au Burundi. [Veterinary medicinal plants of the region of Cretes Zaire-Nil in Burundi]. *Annalen Economische Wetenschappen-Koninklijk Museum voor Midden-Afrika* 21. 133 pp.
102. Bagchi, G.D., Srivastava, G.N. & Singh, S.C., 1992. Distinguishing features of medicinal herbaceous species of *Phyllanthus* occurring in Lucknow district (U.P.) India. *International Journal of Pharmacognosy* 30(3): 161-168.
103. Bailey, C.J., Day, C., Turner, S.L. & Leatherdale, B.A., 1985. Cerasee, a traditional treatment for diabetes. *Studies in normal and streptozotocin diabetic mice*. *Diabetes Research* 2(2): 81-84.
104. Bailey, L.H., 1947. *The standard cyclopedia of horticulture*. 3 volumes. MacMillan, New York, United States. 3639 pp.
105. Bajpai, P.N. & Shukla, H.S., 1990. Aonla. In: Bose, T.K. & Mitra, S.K. (Editors): *Fruits: tropical and subtropical*. Naya Prokash, Calcutta, India. pp. 757-767.
106. Balagtas-Burow, G.E., Moroney, J.V. & Longstreth, D.J., 1993. Growth and osmotic adjustment of cultured suspension cells from *Alternanthera philoxeroides* (Mart.) Griseb. after an abrupt increase in salinity. *Journal of Experimental Botany* 44(260): 673-679.
107. Balakrishna, P. & Raman, A., 1992. Cecidogenesis of leaf galls of *Strychnos nux-vomica* (Loganiaceae) induced by the jumping plant louse species *Diaphorina truncata* (Homoptera: Psylloidea: Psyllidae). *Entomologia Generalis* 17(4): 285-292.
108. Balakrishnan, N.P. & Bhargava, N., 1984. The genus *Curcuma* L. (Zingiberaceae) on Andaman and Nicobar Islands. *Journal of the Bombay Natural History Society* 81: 510-514.
109. Balboa, J.G. & Lim-Sylianco, C.Y., 1992. Antigenotoxic effects of drug preparation of akapulko and ampalaya. *Philippine Journal of Science* 121(4): 399-411.
110. Balboa, J.G. & Lim-Sylianco, C.Y., 1995. Effect of some medicinal plants on skin tumor promotion. *Philippine Journal of Science* 124(2): 203-207.
111. Ballard, R., 1986. *Bidens pilosa* complex (Asteraceae) in North and Central America. *American Journal of Botany* 73: 1452-1465.
112. Ballica, R., Ryu, D.D.Y. & Furusaki, S., 1994. Tropane alkaloid production from *Datura stramonium*: an integrated approach to bioprocess optimization of plant cell cultivation. In: Ryu, D.D.Y. (Editor): *Advances in plant biotechnology. Studies in plant science* 4. Elsevier Science, Amsterdam, the Netherlands. pp. 221-254.
113. Balick, M.J., Elisabetsky, E. & Laird, S.A. (Editors), 1996. *Medicinal resources of the tropical forest. Biodiversity and its importance to human health*. Columbia University Press, New York, United States. 440 pp.
114. Balu, S. & Alagesaboopathi, C., 1996. Introduction of mass-multiplication

- of *Andrographis paniculata* Nees by stem cuttings in Shevaroy Hills, Tamil Nadu. *Journal of Economic and Taxonomic Botany* 20(1): 107–109.
115. Balun, L. & Holdsworth, D.K., 1988. Ethnomedicine of the Gulf Province of Papua New Guinea. Part 1: The mountains around Kanabea and Kaintiba. *International Journal of Crude Drug Research* 26(1): 51–55.
  116. Bancilhon, L., 1971. Contribution à l'étude taxonomique de genre *Phyllanthus* (Euphorbiacées) [Contribution to the taxonomic study of the genus *Phyllanthus* (Euphorbiaceae)]. *Boissiera* 18: 1–81, 22 plates.
  117. Bandara, B.M.R. et al., 1989. Antifungal activity of some medicinal plants of Sri Lanka. *Journal of the National Science Council of Sri Lanka* 17(1): 1–13.
  118. Banerjee, N. & Sharma, A.K., 1983. Cytotaxonomy, tissue culture and alkaloids of *Rauwolfia* L. *Nucleus* 26(3): 197–207.
  119. Banerjee, P.K. & Ghosal, S., 1969. Simple indole bases of *Desmodium gangeticum* (Leguminosae). *Australian Journal of Chemistry* 22: 275–277.
  120. Banerjee, R.N. & Pal, D.C., 1984. Some neglected plants of West-Bengal, India. *Journal of Economic and Taxonomic Botany* 5: 905–906.
  121. Banerjee, S., Schmeda-Hirschmann, G., Castro, V., Schuster, A., Jakupovic, J. & Bohlmann, F., 1986. Further sesquiterpene lactones from *Elephantopus mollis* and *Centratherum punctatum*. *Planta Medica* 52: 29–32.
  122. Banerji, R. et al., 1985. *Jatropha* seed oils for energy. *Biomass* 8(4): 277–282.
  123. Bänziger, H., 1982. Fruit-piercing moths (Lep., Noctuidae) in Thailand: a general survey and some new perspectives. *Mitteilungen der Schweizerischen Entomologischen Gesellschaft* 55(3–4): 213–240.
  124. Barber, S.C., 1982. Taxonomic studies in the *Verbena stricta* complex (Verbenaceae). *Systematic Botany* 7(4): 433–456.
  125. Barbind, R.P., Waghmare, P.S. & Patel, P.M., 1994. Nutritive value of some top feeds and cultivated fodders. *Indian Journal of Animal Nutrition* 11(1): 59–61.
  126. Barik, B.R., Bhowmik, T., Dey, A.K., Patra, A., Chatterjee, A., Joy, S., Susan, T., Alam, M. & Kundu, A.B., 1992. Premnazole, an isoxazole alkaloid of *Premna integrifolia* and *Gmelina arborea* with anti-inflammatory activity. *Fitoterapia* 63(4): 295–299.
  127. Barnabas, C.G.G. & Nagarajan, S., 1988. Antimicrobial activity of flavonoids of some medicinal plants. *Fitoterapia* 59(6): 508–510.
  128. Barnett, O.W. & Alper, M., 1977. Characterization of *Iris fulva* mosaic virus. *Phytopathology* 67(4): 448–454.
  129. Barre, J.T. et al., 1997. A bioactive triterpene from *Lantana camara*. *Phytochemistry* 45(2): 321–324.
  130. Barri, M.E.S., El-Dirdiri, N.I., Abu-Damir, H. & Idris, O.F., 1990. Toxicity of *Abrus precatorius* in Nubian goats. *Veterinary and Human Toxicology* 32(6): 541–545.
  131. Bart, C., 1932. Action hypoglycémiant des feuilles de mûrier [Hypoglycaemic action of mulberry leaves]. *Comptes Rendus Hebdomadaires des Séances et Mémoires de la Société de Biologie et de ses Filiales* 109: 897–899.
  132. Barthakur, N.N. & Arnold, N.P., 1991. Chemical analysis of the emblic



- (*Phyllanthus emblica* L.) and its potential as a food source. *Scientia Horticulturae* 47(1-2): 99-105.
133. Baruah, A., Sheikh, M.S. & Kalita, J., 1993. An investigation on the ecology of *Catopsila pyranthe* Linn. (Lepidoptera - Pieridae). *Journal of the Assam Science Society* 35(1): 23-34.
  134. Baslas, R.K. & Gupta, N.C., 1983. Chemical investigation on Indian medicinal plant possessing anticancer activity: roots of *Euphorbia tirucalli* Linn. *Journal of the Indian Chemical Society* 60(5): 506-508.
  135. Batarda Fernandes, R. & Verdcourt, B., 1989. A new African subspecies of *Verbena officinalis* L. *Boletim da Sociedade Broteriana* 62: 305-310.
  136. Battelli, M.G., Polito, L., Bolognesi, A., Lafleur, L., Fradet, Y. & Stirpe, F., 1996. Toxicity of ribosome-inactivating proteins-containing immunotoxins to a human bladder carcinoma cell line. *International Journal of Cancer* 65(4): 485-490.
  137. Baumgartner, B., Erdelmeier, C.A.J., Wright, A.D., Rali, T. & Sticher, O., 1990. An antimicrobial alkaloid from *Ficus septica*. *Phytochemistry* 29(10): 3327-3330.
  138. Begum, S., Usmani, S.B., Siddiqui, B.S. & Siddiqui, S., 1994. Alkaloids from the bark of *Holarrhena pubescens*. *Phytochemistry* 36(6): 1537-1541.
  139. Bélanger, A. & Dextraze, L., 1993. Variability of chamazulene within *Achillea millefolium*. In: Bernáth, J., Craker, L.E. & Levy, A. (Editors): *Botanical aspects, genetic resources, genetics, breeding, biotechnology. Proceedings of the First World Congress on Medicinal and Aromatic Plants for Human Welfare, WOCMAP. Maastricht, the Netherlands, July 19-25, 1992. Acta Horticulturae* 330: 141-146.
  140. Bell, J.M. & van Houten, A.S., 1993. The medicinal plants of Central Seram. In: Edwards, I.D., MacDonald, A.A. & Proctor, J. (Editors): *The natural history of Seram. Intercept, Andover, United Kingdom. pp. 207-230.*
  141. Beltran, I.C. & Kiew, K.Y., 1984. Cytotaxonomic studies in the Zingiberaceae. *Notes from the Royal Botanic Garden Edinburgh* 41: 541-559.
  142. Benoist, R., 1935. *Acanthacées [Acanthaceae]*. In: Gagnepain, F. (Editor): *Flore générale de l'Indo-Chine [General flora of Indo-China]. Vol. 4. Masson & Cie, Paris, France. pp. 610-772.*
  143. Beppu, H., Nagamura, Y. & Fujita, K., 1993. Hypoglycaemic and antidiabetic effects in mice of *Aloe arborescens* Miller var. *natalensis* Berger. *Phytotherapy Research* 7: S37-42.
  144. Berg, C.C., 1989. Classification and distribution of *Ficus*. *Experientia* 45: 605-611.
  145. Bhargava, S.K., 1989. Antiandrogenic effects of a flavonoid-rich fraction of *Vitex negundo* seeds: a histological and biochemical study in dogs. *Journal of Ethnopharmacology* 27(3): 327-340.
  146. Bhatia, N., McDonald, K.A., Jackman, A.P. & Dandekar, A.M., 1996. A simplified procedure for the purification of trichosanthin (a type 1 ribosome inactivating protein) from *Trichosanthes kirilowii* root tubers. *Protein Expression and Purification* 7(2): 143-146.
  147. Bhattacharya, P., Dey, S. & Bhattacharya, B.C., 1994. Use of low-cost gelling agents and support matrices for industrial scale plant tissue cul-

- ture. *Plant Cell, Tissue and Organ Culture* 37(1): 15–23.
148. Bhattacharya, T.K., Ghosh, M.N. & Subramanian, S.S., 1980. A note on anti-inflammatory activity of carpesterol. *Fitoterapia* 51(5): 265–268.
  149. Bhutani, K.K., Ali, M., Sharma, S.R., Vaid, R.M., Gupta, D.K., 1988. Three new steroidal alkaloids from the bark of *Holarrhena antidysenterica*. *Phytochemistry* 27(3): 925–928.
  150. Bhutani, K.K., Vaid, R.M., Ali, M., Kapoor, R., Soodan, S.R. & Kumar, D., 1990. Steroidal alkaloids from *Holarrhena antidysenterica*. *Phytochemistry* 29(3): 969–972.
  151. Bianco, A., Guiso, M. & Passacantilli, P., 1984. Iridoid and phenylpropanoid glycosides from new sources. *Journal of Natural Products* 47(5): 901–902.
  152. Bierer, D.E., Carlson, Th.J. & King, S.R., 1996. Shaman Pharmaceuticals: Integrating indigenous knowledge, tropical medicinal plants, medicine, modern science and reciprocity into a novel drug discovery approach. *Network Science* 2(5): <http://www.netsci.org/Science/Special/feature11.html>.
  153. Bierhaus, A., Zhang, Y., Quehenberger, P., Luther, T., Hasse, M., Müller, M., Mackman, N., Ziegler, R. & Nawroth, P.P., 1997. The dietary pigment curcumin reduces endothelial tissue factor gene expression by inhibiting binding of AP-1 to DNA and activation of NF-kappa B. *Thrombosis and Haemostasis* 77(4): 772–782.
  154. Biesboer, D.D. & Mahlberg, P.G., 1984. The effect of medium modification and selected precursors on sterol production by short-term callus cultures of *Euphorbia tirucalli*. *Journal of Natural Products* 42(6): 648–657.
  155. Binojkumar, M.S. & Balakrishnan, N.P., 1993. Notes on *Euphorbia atoto* G. Forster and its allied species (Euphorbiaceae). *Rheedea* 3(2): 113–116.
  156. Bioka, D. & Mabika, A., 1993. Effet analgesique d'un extrait brut d'*Ageratum conyzoides* chez le rat [Analgesic effect of a crude extract from *Ageratum conyzoides* on the rat]. In: Schilcher, H., Phillipson, J.D. & Loew, D. (Editors): *Pharmacology, phytotherapy, human welfare, regional aspects. Proceedings of the First World Congress on Medicinal and Aromatic Plants for Human Welfare, WOCMAP. Maastricht, the Netherlands, July 19–25, 1992. Acta Horticulturae* 332: 171–176.
  157. Birrie, H., Balcha, F., Erko, B., Bezuneh, A. & Gameda, N., 1998. Investigation into the cercariacidal and miracidicidal properties of endod (*Phytolacca dodecandra*) berries (type 44). *East African medicinal Journal* 75(5): 311–314.
  158. Bisset, N.G., 1958. The occurrence of alkaloids in the Apocynaceae. *Annales Bogorienses* 3(1): 105–236.
  159. Bisset, N.G., 1962. Cardiac glycosides. Part VI. Moraceae: The genus *Antiaris* Lesch. *Planta Medica* 10: 143–151.
  160. Bisset, N.G., 1974. The Asian species of *Strychnos*. Part III. The ethnobotany. *Lloydia* 37(1): 62–107.
  161. Bisset, N.G., 1981. Phytochemistry of *Holarrhena* R. Br. *Mededelingen Landbouwhogeschool Wageningen* 81-2: 37.
  162. Bisset, N.G., Baser, H.C., Phillipson, J.D., Bohlin, L. & Sandberg, F., 1977. Muscle-relaxant activity in Asian *Strychnos* species. A re-examination of two Western Malaysian dart poisons. *Lloydia* 40(6): 546–560.
  163. Bisset, N.G. & Phillipson, J.D., 1976. The Asian species of *Strychnos*. Part

- IV. The alkaloids. *Lloydia* 39(5): 263–325.
164. Blomqvist, M.M., 1997. Taxonomy and uses of medicinally important species in the genera *Datura* L. and *Solanum* L. (Solanaceae) in South-East Asia. Unpublished MSc thesis, Department of Plant Taxonomy, Wageningen Agricultural University, the Netherlands. 132 pp.
  165. Bodger, M.P., McGiven, A.R. & Fitzgerald, P.H., 1979. Mitogenic proteins of pokeweed – part 1: purification, characterization and mitogenic activity of 2 proteins from pokeweed (*Phytolacca octandra*). *Immunology* 37(4): 785–792.
  166. Bodger, M.P., McGiven, A.R. & Fitzgerald, P.H., 1979. Mitogenic proteins of pokeweed – part 2: the differentiation of human peripheral blood bone marrow derived lymphocytes stimulated with purified pokeweed mitogens PO-2 and PO-3 from pokeweed (*Phytolacca octandra*). *Immunology* 37(4): 793–800.
  167. Boer, E. & Sosef, M.S.M., 1998. *Ficus* L. In: Sosef, M.S.M., Hong, L.T. & Prawirohatmodjo, S. (Editors): *Plant Resources of South-East Asia No 5(3). Timber trees: Lesser-known timbers*. Backhuys Publishers, Leiden, the Netherlands. pp. 232–238.
  168. Bohlmann, F., 1990. Chemistry of the Heliantheae (Compositae). *Plant Systematics and Evolution Suppl.* 47: 67–75.
  169. Bolognesi, A., Barbieri, L., Carnicelli, D., Abbondanza, A., Cenini, P., Falasca, A.I., Dinota, A. & Stirpe, F., 1989. Purification and properties of a new ribosome-inactivating protein with RNA N-glycosidase activity suitable for immunotoxin preparation from the seeds of *Momordica cochinchinensis*. *Biochimica et Biophysica Acta* 993(2–3): 287–292.
  170. Bolognesi, A., Tazzari, P.L., Olivieri, F., Polito, L., Lemoli, R., Terenzi, A., Pasqualucci, L., Falini, B. & Stirpe, F., 1998. Evaluation of immunotoxins containing single-chain ribosome-inactivating proteins and an anti-CD22 monoclonal antibody (OM124): in vitro and in vivo studies. *British Journal of Haematology* 101(1): 179–188.
  171. Bonati, G., 1915–1927. Solanacées [Solanaceae]. In: Gagnepain, F. (Editor): *Flore générale de l'Indo-Chine [General flora of Indo-China]*. Vol. 4. Masson & Cie, Paris, France. pp. 313–341.
  172. Bor, N.L., 1960. *The grasses of Burma, Ceylon, India and Pakistan*. Pergamon Press, Oxford, United Kingdom. 767 pp.
  173. Borhanuddin, M., Shamsuzzoha, M & Hussain, A.H., 1994. Hypoglycaemic effects of *Andrographis paniculata* Nees on non-diabetic rabbits. *Bulletin of the Bangladesh Medical Research Council* 20(1): 24–26.
  174. Borthwick, H.A. & Scully, N.J., 1954. Photoperiodic responses of hemp. *Botanical Gazette* 116: 14–29.
  175. Bose, P.C., Majundar, S.K. & Sengupta, K., 1990. Sandy-loam soils of Andamans are suitable for mulberry cultivation. *Indian Farming* 39(11): 8–9.
  176. Bourinbaiar, A.S. & Lee-Huang, S., 1995. The activity of plant-derived antiretroviral proteins MAP30 and GAP31 against Herpes simplex virus infection in vitro. *Biochemical and Biophysical Research Communications* 219(3): 923–929.
  177. Bourinbaiar, A.S. & Lee-Huang, S., 1995. Potentiation of anti-HIV activity of anti-inflammatory drugs, dexamethasone and indomethacin, by

- MAP30, the antiviral agent from bitter melon. *Biochemical and Biophysical Research Communications* (March 17)
178. Bradu, B.L., Agarwal, S.G., Vashist, V.N. & Atal, C.K., 1971. Comparative performance of diploid and tetraploid *Mentha arvensis* and evaluation of their oils. *Planta Medica* 20(3): 219-222.
179. Brain, B.B. & Smith, R.C.G., 1980. Potential for mint production in Australia: a preliminary study. *The Journal of the Australian Institute of Agricultural Science* 46: 207-213.
180. Brandao, M.G.L., Krettli, A.U., Soares, L.S.R., Nery, C.G.C. & Marinuzzi, H.C., 1997. Antimalarial activity of extracts and fractions from *Bidens pilosa* and other *Bidens* species (Asteraceae) correlated with the presence of acetylene and flavonoid compounds. *Journal of Ethnopharmacology* 57: 131-138.
181. Braza, R.D., 1990. Psyllids on nitrogen fixing trees in the Philippines. *Nitrogen Fixing Tree Research Reports* 8: 62-63.
182. Bremekamp, C.E.B., 1937. The Malaysian species of the genus *Ixora* (Rub.). *Contributions à l'étude de la flore des Indes Néerlandaises XXXIV*. *Bulletin du Jardin Botanique de Buitenzorg, Série III*, 14: 197-367.
183. Bremekamp, C.E.B., 1948. Notes on the Acanthaceae of Java. *Verhandelingen der Koninklijke Nederlandsche Akademie van Wetenschappen, afd. Natuurkunde, 2nd sect.*, 45(2): 1-78.
184. Bremer, K., 1994. *Asteraceae, cladistics & classification*. Timber Press, Portland, Oregon, United States. 752 pp.
185. Breuer, H., Rangel, M. & Medina, E., 1982. Pharmacological properties of melochinine, an alkaloid producing Central American cattle paralysis. *Toxicology* 25: 223-242.
186. Brill, H.C. & Wells, A.H., 1917. The physiological active constituents of certain Philippine medicinal plants. II. *Philippine Journal of Science* 12: 16-95.
187. Bronstein, J.L. & McKey, D., 1989. The fig/pollinator mutualism: A model system for comparative biology. *Experientia* 45: 601-604.
188. Brook, M.G., 1988. Effect of *Phyllanthus amarus* on chronic carriers of hepatitis B virus. *The Lancet* 332: 1017-1018.
189. Broszat, W., 1992. *Der Mohn (Papaver somniferum L.): Anbau und Markt einer wiederentdeckten Kulturpflanze [The poppy (Papaver somniferum L.): cultivation and market of a rediscovered crop]*. *Der Tropenlandwirt, Beiheft 47*. Deutsches Institut für Tropische und Subtropische Landwirtschaft, Witzenhausen, Germany. 170 pp.
190. Brown, W.H., 1951-1957. *Useful plants of the Philippines*. Reprint of the 1941-1943 edition. 3 volumes. Technical Bulletin 10. Department of Agriculture and Natural Resources. Bureau of Printing, Manila, the Philippines. Vol. 1 (1951) 590 pp., Vol. 2 (1954) 513 pp., Vol. 3 (1957) 507 pp.
191. Bruchhausen, F. et al. (Editors), 1994. *Curcuma*. *Hager's Handbuch der Pharmazeutischen Praxis [Hager's Handbook of pharmacological practice]*. 5th Edition. Springer Verlag, Berlin, Germany. pp. 1084-1102.
192. Brunel, J.F. & Roux, J., 1984. South-East Asian *Phyllanthus* II. Some *Phyllanthus* of subsect. *Swartziani*. *Nordic Journal of Botany* 4(4): 469-473.
193. Bruneton, J., 1995. *Pharmacognosy, phytochemistry, medicinal plants*.

- Technique & Documentation Lavoisier, Paris, France. 915 pp.
194. Bucher, M., Brandle, R. & Kuhlemeier, C., 1996. Glycolytic gene expression in amphibious *Acorus calamus* L. under natural conditions. *Plant and Soil* 178(1): 75–82.
  195. Büchi, G. & Manning, R.E., 1962. Constitution of verbenalin. *Tetrahedron Letters* 18: 1049–1059.
  196. Budavari, S. (Editor), 1989. *The Merck Index, an Encyclopedia of Chemicals, Drugs and Biologicals*. 11<sup>th</sup> Edition. Merck & Co. Inc. Rahway, New Jersey, United States.
  197. Bugge, G., 1991. Untersuchungen der Sippen des *Achillea millefolium* Komplexes auf Azulengehalt und Ploidiegrad [Investigations of the taxa of the *Achillea millefolium* complex on the content of azulene and ploidy level]. *Angewandte Botanik* 65(5–6): 331–339.
  198. Bunyapraphatsara, N., Jirakulchaiwong, S., Thirawarapan, S. & Manonukul, J., 1995. The efficacy of Aloe vera cream in the treatment of first, second and third degree burns in mice. *Phytomedicine* 2(3): 247–251.
  199. Bunyapraphatsara, N., Yongchaiyudha, S., Rungpitarangsi, V. & Chokechajaroenporn, A., 1996. Antidiabetic activity of Aloe vera L. juice. II. Clinical trial in diabetes mellitus patients in combination with glibenclamide. *Phytomedicine* 3(3): 245–248.
  200. Burgos, R.A., Caballero, E.E., Sanche, N.S., Schroeder, R.A., Wikman, G.K. & Hancke, J.L., 1997. Testicular toxicity assessment of *Andrographis paniculata* dried extract in rats. *Journal of Ethnopharmacology* 58(3): 219–224.
  201. Burkill, H.M., 1985–1997. *The useful plants of West tropical Africa*. Edition 2. 5 volumes. Royal Botanic Gardens Kew, United Kingdom. Vol. 1 (1985), Families A–D, 960 pp.; Vol. 2 (1994), Families E–I, 636 pp.; Vol. 3 (1995), Families J–L, 857 pp.; Vol. 4 (1997), Families M–R, 969 pp.
  202. Burkill, I.H., 1966. *A dictionary of the economic products of the Malay Peninsula*. Revised reprint. 2 volumes. Ministry of Agriculture and Co-operatives, Kuala Lumpur, Malaysia. Vol. 1 (A–H) pp. 1–1240. Vol. 2 (I–Z) pp. 1241–2444.
  203. Burtt, B.L., 1977. *Curcuma zedoaria*. *Gardens' Bulletin*, Singapore 30: 59–62.
  204. Busciglio, J.A., 1988. Anti-inflammatory composition containing lidocaine and disphenhydramine. U.S. Patent 4, 748, 022 (Cl. 424-1951; A61k 35/78), 31 May 1988.
  205. Buijsen, J.R.M., 1993. *Alliaceae*. In: Kalkman, C., Kirkup, D.W., Nooteboom, H.P., Stevens, P.F. & de Wilde, W.J.J.O. (Editors): *Flora Malesiana*. Series 1, Vol. 11. Rijksherbarium/Hortus Botanicus, Leiden, the Netherlands. pp. 375–384.
  206. Byragi Reddy, T. & Subba Reddi, C., 1994. Pollination ecology of *Vitex negundo* (Verbenaceae). *Proceedings of the Indian National Science Academy, Part B, Biological Sciences* 60: 57–66.
  207. Caceres, A., Giron, L.M., Alvarado, S.R. & Torres, M.F., 1987. Screening of antimicrobial activity of plants popularly used in Guatemala for the treatment of urinary ailments in Guatemala. *Journal of Ethnopharmacology* 20(3): 223–237.
  208. Cai, B.-C., Hattori, M. & Namba, T., 1990. Processing of *nux vomica*. II.

- Changes in alkaloid composition of the seeds of *Strychnos nux-vomica* on traditional drug-processing. *Chemical and Pharmaceutical Bulletin* 38(5): 1295–1298.
209. Cakici, I., Hurmoglu, C., Tunctan, B., Abacioglu, N., Kanik, I. & Sener, B., 1994. Hypoglycaemic effect of *Momordica charantia* extracts in normoglycaemic or cyproheptadine-induced hyperglycaemic mice. *Journal of Ethnopharmacology* 44(2): 117–121.
210. Calvin, M., 1987. Fuel oils from euphorbs and other plants. *Botanical Journal of the Linnean Society* 94: 97–110.
211. Cantoria, M., 1974. Studies on the physiology of Philippine mint (*Mentha cordifolia* Opiz). III. Variation in oil yield. *Philippine Journal of Science* 103(1): 67–77.
212. Cantoria, M., 1980. Studies on the growth and development of *Mentha* and *Datura* in the Philippines. *NRCP Bulletin No 9*: 2–10.
213. Cardenas, L.B. & Quimado, M.O., 1986. Tissue culture of medicinal plants: an investment for the future. *Research at Los Baños (Philippines)* 5(2): 14–15.
214. Carle, T., 1981. Investigations on the content of steroidal alkaloids and saponin within *Solanum* sect. *Solanum* (= sect. *Morella*) (*Solanaceae*). *Plant Systematics and Evolution* 138: 61–71.
215. Carnat, A., Carnat, A.P., Chavignon, O., Heitz, A., Wylde, R. & Lamaison, J.L., 1995. Luteolin 7-diglucuronide, the major flavonoid compound from *Aloysia triphylla* and *Verbena officinalis*. *Planta Medica* 61: 490.
216. Carter, S., 1994. *Aloaceae*. In: Polhill, R.M. (Editor): *Flora of Tropical East Africa*. A.A. Balkema, Rotterdam, the Netherlands. 60 pp.
217. Carter, S. & Radcliffe-Smith, A., 1988. *Euphorbiaceae* (Part 2). In: Polhill, R.M. (Editor): *Flora of Tropical East Africa*. A.A. Balkema, Rotterdam, the Netherlands. pp. 409–576.
218. Casellas, P., Dussossoy, D., Falasca, A.I., Barbieri, L., Guillemot, J.C., Ferrara, P., Bolognesi, A., Cenini, P. & Stirpe, F., 1988. Trichokirin, a ribosome-inactivating protein from the seeds of *Trichosanthes kirilowii* Maximowicz. Purification, partial characterization and use for preparation of immunotoxins. *European Journal of Biochemistry* 176(3): 581–588.
219. Casyao, J.M., 1992. The botany of sampaguita, its production and potentials in industry: an overview. *Philippine Technology Journal* 17(2): 1–12.
220. Casyao, J.M. & Medina, F.I.S., 1990. The detached leaf propagation of sampaguita (*Jasminum sambac* (L.) Ait.). *Philippine Technology Journal* 15(2): 43–52.
221. Catalfamo, J.L., Martin Jr, W.B. & Birecka, H., 1982. Accumulation of alkaloids and their necines in *Heliotropium curassavicum*, *H. spathulatum* and *H. indicum*. *Phytochemistry* 21(11): 2669–2675.
222. Cavallini, A., Natali, L. & Castorena Sanchez, I., 1991. *Aloe barbadensis* Mill. (= *A. vera* L.). In: Bajaj, Y.P.S. (Editor): *Biotechnology in agriculture and forestry*. Vol. 15. *Medicinal and aromatic plants III*. Springer Verlag, Berlin, Germany. pp. 95–106.
223. Cera, L.M., Hegggers, J.P., Robson, M.C. & Hagstrom, W.J., 1980. The therapeutic efficacy of *Aloe vera* cream (Dermaide Aloe) in thermal injuries: two case reports. *Journal of the American Animal Hospital Association* 16(5): 768–772.

224. Chagnon, M., 1984. General pharmacologic inventory of Rwandese medicinal plants. *Journal of Ethnopharmacology* 12(3): 239–251.
225. Chainuvati, P., Bunyapraphatsara, N., Luengrojanakul, P. & Damrongsak, C., 1994. Effect of *Phyllanthus amarus* Schum. et Thonn. on Thai chronic carriers of hepatitis B virus. *Thai Journal of Phytopharmacy* 1(2): 13–16.
226. Chakravarty, A.K., Dastidar, P.P.G. & Pakrashi, S.C., 1982. Simple aromatic amines from *Justicia gendarussa*.  $^{13}\text{C}$  NMR spectra of the bases and their analogues. *Tetrahedron* 38(12): 1797–1802.
227. Chalchat, J.C. & Garry, R.P., 1996. Chemical composition of the leaf oil of *Verbena officinalis* L. *Journal of Essential Oil Research* 8: 419–420.
228. Chan, K.L., O'Neill, M.J., Phillipson, J.D. & Warhurst, D.W., 1986. Plants as sources of antimalarial drugs: *Eurycoma longifolia*. *Planta Medica* 52: 105–107.
229. Chan, M.M.Y & Fong, D., 1994. Anti-inflammatory and cancer-preventive immunomodulation through diet: effects of curcumin on T lymphocytes. *American Chemical Society Symposium Series*. American Chemical Society, Washington, D.C., United States. pp. 222–230.
230. Chan, W.Y., Ng, T.B., Wu, P.J. & Yeung, H.W., 1993. Developmental toxicity and teratogenicity of trichosanthin, a ribosome-inactivating protein in mice. *Teratogenesis, Carcinogenesis and Mutagenesis* 13(2): 47–57.
231. Chanda Bhaumik (Modal) & Datta, P.C., 1989. Development of Japanese mint tissue culture method. *Indian Perfumer* 33(3): 165–168.
232. Chandra, S., Shahi, A.K., Dutt, P. & Tawa, A., 1996. Essential oil composition of *Ageratum houstonianum* Mill. from Jammu region of India. *Journal of Essential Oil Research* 8(2): 129–134.
233. Chandrasekar, B., Mukherjee, B. & Mukherjee, S.K., 1989. Blood sugar lowering potentiality of selected Cucurbitaceae plants of Indian origin. *Indian Journal of Medical Research* 90: 300–305.
234. Chandravadana, M.V., Nidiry, E.S.J. & Venkateshwarlu, G., 1997. Antifungal activity of momordicines from *Momordica charantia*. *Fitoterapia* 68(4): 383–384.
235. Chang, C.L., Huang, C.Y., Yao, Y.C., Ch'ien, W.Y. & Chao, P.N., 1974. 200 cases of psoriasis treated with traditional Chinese medicine. *Chinese Medical Journal* 4: 205–207.
236. Chang, R.S. et al., 1991. Dehydroandrographolide succinic acid monoester as an inhibitor against the human immunodeficiency virus. *Proceedings of the Society for Experimental Biology and Medicine* 197(1): 59–66.
237. Chanprasert, W., Paisooksantivatana, Y., Lersrutaiyotin, R., Phoolkets, U., Satakhun, D. & Thapatat, Y., 1994. Effect of planting date, plant spacing, and harvesting date on seed yield and quality of *Plantago psyllium* L. *Thai Journal of Phytopharmacy* 1(2): 29–37.
238. Charles, B. et al., 1987. Some unusual proaporphine and aporphine alkaloids from *Stephania venosa*. *Journal of Natural Products* 50: 1113–1117.
239. Chaturvedi, G.N. & Singh, K.P., 1983. Side effects of a traditional indigenious drug – kutaja, *Holarrhena antidysenterica*. *Indian Journal of Physiology and Pharmacology* 27(3): 355–356.
240. Chatzopoulou, P., Katsiotis, S.T. & Baerheim Svendsen, A., 1992. An ascaridole containing essential oil of the *Achillea millefolium* L. complex

- growing wild in northern Greece. *Journal of Essential Oil Research* 4(5): 457–459.
241. Chen, C.C., Hsin, W.C., Ko, F.N., Huang, Y.L., Ou, J.C. & Teng, C.M., 1996. Antiplatelet aryl-naphthalide lignans from *Justicia procumbens*. *Journal of Natural Products* 59(12): 1149–1150.
242. Chen, C.-H., 1993. Sapindaceae. In: Huang, T.-C. (Editor): *Flora of Taiwan*. 2nd Edition. Vol. 3. Editorial Committee of the Flora of Taiwan, Taipei, Taiwan, Republic of China. pp. 599–608.
243. Chen, C.P., Lin, C.C. & Namba, T., 1987. Development of natural crude drug resources from Taiwan VI. In-vitro studies of the inhibitory effect of 12 microorganisms. *Shoyakugaku Zasshi* 41(3): 215–225. (in Japanese)
244. Chen, C.P., Lin, C.C. & Namba, T., 1989. Screening of Taiwanese crude drugs for antibacterial activity against *Streptococcus mutans*. *Journal of Ethnopharmacology* 27(3): 285–296.
245. Chen, K.K. & Hou, H., 1926. The alleged anthelmintic properties of *Quisqualis indica* with case reports. *Annual Journal of Medical Science* 172(1): 113–116.
246. Chen, P.K. & Leather, G.R., 1990. Plant growth regulatory activities of artemisinin and its related compounds. *Journal of Chemical Ecology* 16(6): 1867–1876.
247. Chen, X.M., Qian, Y.W., Chi, C.W., Gan, K.D., Hang, M.F. & Chen, C.O., 1992. Chemical synthesis, molecular cloning, and expression of the gene coding for the *Trichosanthes* trypsin inhibitor, a squash family inhibitor. *Journal of Biochemistry* 112(1): 45–51.
248. Chew, W.-L., 1989. Moraceae. In: George, A.S. (Editor): *Flora of Australia*. Vol. 3. Hamamelidales to Casuarinales. Australian Government Publishing Service, Canberra, Australia. pp. 15–68.
249. Chien, N.Q. & Adam, G., 1979. Über die Inhaltstoffe von *Smilax glabra* Roxb. [The constituents of *Smilax glabra* Roxb.]. *Pharmazie* 34(12): 841–843.
250. Chithra, P., Sajithlal, G.B. & Chandrakasan, G., 1998a. Influence of *Aloe vera* on the glycosaminoglycans in the matrix of healing dermal wounds in rats. *Journal of Ethnopharmacology* 59(3): 179–186.
251. Chithra, P., Sajithlal, G.B. & Chandrakasan, G., 1998b. Influence of *Aloe vera* on the healing of dermal wounds in diabetic rats. *Journal of Ethnopharmacology* 59(3): 195–201.
252. Chockalingam, S., Thenmozhi, S. & Nalina Sundari, M.S., 1990. Larvicidal activity of different products against mosquito larvae. *Journal of Environmental Biology* 11(2): 101–104.
253. Choi, J.S., Jung, J.H., Hee, J.L., Lee, H.J. & Kang, S.S., 1995. A naphthalene glycoside from *Cassia tora*. *Phytochemistry* 40(3): 997–999.
254. Choi, J.S., Lee, H.J., Park, K.Y., Ha, J.O. & Kang, S.S., 1997. In vitro antimutagenic effects of anthraquinone aglycones and naphthopyrone glycosides from *Cassia tora*. *Planta Medica* 63(1): 11–14.
255. Choi, Y.H., Hussain, R.A., Pezzuto, J.M., Kinghorn, A.D. & Morton, J.F., 1989. Abrusosides A–D, four novel sweet-tasting triterpene glycosides from the leaves of *Abrus precatorius*. *Journal of Natural Products* 52(5): 1118–1127.
256. Chomchalow, N. & Henle, H.V. (Editors), 1993. *Medicinal and aromatic*



- plants in Asia. Breeding and improvement. RAPA Publication 1993/19. Science Publishers, Lebanon, United States. 196 pp.
257. Choomsai, A., 1982. Bitter gourd (*Momordica charantia* Linn.) in Thailand. Newsletter Regional Committee for Southeast Asia, IBPGR 6(1): 4.
258. Chopra, R.N., Badhwar, R.L. & Ghosh, S., 1965. Poisonous plants of India. 2 volumes. Indian Council of Agricultural Research, New Delhi, India. 972 pp.
259. Christensen, L.P., Lam, J. & Thomasen, T., 1990. Chalcone and other constituents of *Bidens tripartita*. *Phytochemistry* 29(10): 3155–3156.
260. Christie, W.W., Brechany, E.Y. & Shukla, V.K., 1989. Analysis of seed oils containing cyclopentenyl fatty acids by combined chromatographic procedures. *Lipids* 24: 116–120, 564.
261. Chu, D.M., Miles, H., Toney, D., Nguyen, C. & Marciano-Cabral, F., 1998. Amoebicidal activity of plant extracts from Southeast Asia on *Acanthamoeba* spp. *Parasitology Research* 84(9): 746–752.
262. Chuakul, W., Saralamp, P., Jiratchariyakul, W. & Supratrarawanich, P., 1994. Specification of Thai medicinal plant: *Abrus fruticulosus* Wall. ex Wight & Arn. *Thai Journal of Phytopharmacy* 1(1): 17–25.
263. Chuakul, W., Saralamp, P., Paonil, V. & Temsiririrkkul, R., 1996. *Samunpri puenban lanna*. [Medicinal plants in the northern part of Thailand]. Department of Pharmaceutical Botany, Faculty of Pharmacy. Mahidol University. Amarin Printing and Publishing Public Co. Ltd., Bangkok, Thailand. 264 pp.
264. Chundawat, B.S., 1990. Arid fruit culture. Oxford & IBH Publishing Co., New Delhi, India. 208 pp.
265. Chung, B., 1990. Effects of plant population density and rectangularity on the growth and yield of poppies (*Papaver somniferum*). *Journal of Agricultural Science (Cambridge)* 115(2): 239–245.
266. Chung, B., 1992. The effects of plant density and irrigation on the lodging, yield and yield components of poppies (*Papaver somniferum* L.). *Acta Horticulturae* 306: 458–465.
267. Chung, Y.B., Lee, C.C., Park, S.W. & Lee, C.K., 1990. Studies on antitumour and immunopotentiating activities of polysaccharides from *Trichosanthes* rhizome. *Archives of Pharmacological Research* 13(3): 285–288.
268. Citores, L. et al., 1993. Molecular mechanism of inhibition of mammalian protein synthesis by some four-chain agglutinins. Proposal of an extended classification of plant ribosome-inactivating proteins rRNA N glucosidases. *FEBS (Federation of European Biochemical Societies) Letters* 329(1–2): 59–62.
269. Claeson, P., Panthong, A., Tuchinda, P., Reutrakul, V., Kanjanapathi, D., Taylor, W.C. & Santisuk, T., 1993. Three non-phenolic diarylheptanoids with anti-inflammatory activity from *Curcuma xanthorrhiza*. *Planta Medica* 59(5): 451–454.
270. Claeson, P., Pongprayoon, U., Sematong, T., Tuchinda, P., Reutrakul, V., Soonthorsaratune, P. & Taylor, W.C., 1996. Non-phenolic linear diarylheptanoids from *Curcuma xanthorrhiza*: a novel type of topical anti-inflammatory agents: structure-activity relationship. *Planta Medica* 62(3): 236–240.
271. Clarke, C.B., 1882. Oleaceae. In: Hooker, J.D. (Editor): *The Flora of*

- British India. Periodical Expert Book Agency, New Delhi, India. Vol. 3, part 9. pp. 590–618.
272. Cock, M.J.W. & Evans, H.C., 1984. Possibilities for biological control of *Cassia tora* and *C. obtusifolia*. *Tropical Pest Management* 30(4): 339–350.
273. Colasanti, B.K., Craig, C.R. & Allara, R.D., 1984. Intraocular pressure, ocular toxicity and neurotoxicity after administration of cannabinal or cannabigerol. *Experimental Eye Research* 39: 251–259.
274. Colegate, S.M. & Molyneux, R.J., 1993. Bioactive natural products – Detection, isolation and structural determination. CRC Press, Boca Raton, Florida, United States.
275. Collin, C.F. & Collin, C., 1935. Roentgen dermatitis treated with fresh whole leaf of *Aloe vera*. *American Journal of Roentgenology, Radium Therapy and Nuclear Medicine* 33: 396–397.
276. Conn, B.J. & Brown, E.A., 1993. Notes on *Strychnos* L. (Loganiaceae) in Australia. *Australian Systematic Botany* 6: 309–319.
277. Conran, J.G., 1987. *Gloriosa*. In: George, A.S. (Editor): *Flora of Australia*. Vol. 45. Hydatellaceae to Liliaceae. Australian Government Publishing Service, Canberra, Australia. pp. 412.
278. Conran, J.G. & Clifford, H.T., 1986. Smilacaceae. In: George, A.S. (Editor): *Flora of Australia*. Vol. 46. Iridaceae to Dioscoreaceae. Australian Government Publishing Service, Canberra, Australia. pp. 180–196.
279. Corbineau, F. & Côme, D., 1981. Some particularities of the germination of *Oldenlandia corymbosa* L. seeds (tropical Rubiaceae). *Israel Journal of Botany* 29: 157–167.
280. Corner, E.J.H., 1962. The classification of Moraceae. *Gardens' Bulletin, Singapore* 19: 187–252.
281. Corner, E.J.H., 1965. Check-list of *Ficus* in Asia and Australia. *Gardens' Bulletin, Singapore* 21: 1–186.
282. Corner, E.J.H., 1969. The complex of *Ficus deltoidea*. *Philosophical Transactions of the Royal Society of London, B*, 256: 281–317.
283. Corner, E.J.H., 1981. Moraceae. In: Dassanayake, M.D. & Fosberg, F.R. (Editors): *A revised handbook to the flora of Ceylon*. Vol. 3. Amerind Publishing Co. Pvt. Ltd., New Delhi, India. pp. 213–292.
284. Corner, E.J.H., 1988. *Wayside trees of Malaya*. 3rd Edition. 2 volumes. The Malayan Nature Society, Kuala Lumpur, Malaysia. 774 pp.
285. Corsaro, M.M., Giudicianni, I., Lanzetta, R., Marciano, C.E., Monaco, P. & Parrilli, M., 1995. Polysaccharides from seeds of *Strychnos* species. *Phytochemistry* 39(6): 1377–1380.
286. Costa, S.S., Jossang, A., Bodo, B., Souza, M.L. & Moraes, V.L., 1994. Palutelin acetyl rhamnosides from *Kalanchoe brasiliensis* as inhibitors of human lymphocyte proliferative activity. *Journal of Natural Products* 57(11): 1503–1510.
287. Council of Scientific and Industrial Research, 1948–1976. *The wealth of India: a dictionary of Indian raw materials & industrial products*. 11 volumes. Publications and Information Directorate, New Delhi, India.
288. Council of Scientific and Industrial Research, 1985. *The wealth of India: a dictionary of Indian raw materials & industrial products*. Revised Edition. Vol. 1. Publications and Information Directorate, New Delhi, India. 513 pp.

289. Council of Scientific and Industrial Research, 1988. The wealth of India: a dictionary of Indian raw materials & industrial products. Revised Edition. Vol. 2B. Publications and Information Directorate, New Delhi, India. xlii + 350 + 90 pp.
290. Craib, W.G., 1928. XXXIV – Contributions to the flora of Siam. *Additamentum XXV*. Kew Bulletin 1928: 234–246.
291. Craib, W.G., 1930. XLVII – Contributions to the flora of Siam. *Additamentum XXIX*. Kew Bulletin 1930: 405–427.
292. Cramer, L.H., 1981. Lamiaceae (Labiatae). In: Dassanayake, M.D. & Fosberg, F.R. (Editors): A revised handbook to the flora of Ceylon. Vol. 3. Amerind Publishing Co., New Delhi, India. pp. 108–194.
293. Cramer, U., Rehfeldt, A.G. & Spener, F., 1980. Isolation and biosynthesis of cyclopentenyl glycine, a novel nonproteinogenic amino-acid in Flacourtiaceae. *Biochemistry* 19(13): 3074–3080.
294. Cramer, U. & Spener, F., 1977. Biosynthesis of cyclopentenyl fatty acids. Cyclopentenylglycine, a non-proteinogenic amino acid as precursor of cyclic fatty acids in Flacourtiaceae. *European Journal of Biochemistry* 74(3): 495–500.
295. Crockett, C.O., Guede-Guina, F., Pugh, D., Vangah-Manda, M., Robinson, T.J., Olubadewo, J.O. & Ochillo, R.F., 1992. Cassia alata and the preclinical search for therapeutic agents for the treatment of opportunistic infections in AIDS patients. *Cellular and Molecular Biology (Oxford)* 38(5): 505–511.
296. Croizat, L., 1934. De euphorbio antiquorum atque officinarum [A study of succulent Euphorbiae long known in cultivation]. New York, United States. 127 pp.
297. Currò, P., Micali, G. & Lanuzza, F., 1987. Determination of  $\beta$ -asarone, safrole, isosafrole and anethole in alcoholic drinks by high-performance liquid chromatography. *Journal of Chromatography* 404: 273–278.
298. Czaplá, T.H. & Lang, B.A., 1990. Effect of plant lectins on the larval development of European corn borer (Lepidoptera, Pyralidae) and southern corn rootworm (Coleoptera, Chrysomelidae). *Journal of Economic Entomology* 83(6): 2480–2485.
299. Czygan, F.C., 1995. Catharanthus roseus (L.) G. Don – Das Madagaskar-Immergrün [Madagascar periwinkle]. *Zeitschrift für Phytotherapie* 16: 178–186.
300. D'Arcy, W.G., 1972. Solanaceae studies II: Typification of subdivisions of Solanum. *Annals of the Missouri Botanical Garden* 59: 262–278.
301. D'Silva, I., Vaidyanathan, C.S. & Podder, S.K., 1993. Ribosome-inactivating proteins and agglutinins from callus and suspension cultures of *Ricinus communis* L. and *Abrus precatorius* L. *Plant Science (Limerick)* 94(1–2): 161–172.
302. Da Silva, S.A.G., Costa, S.S., Mendonca, S.C.F., Silva, E.M., Moraes, V.L.G. & Rossi-Bergmann, B., 1995. Therapeutic effect of oral *Kalanchoe pinnata* leaf extract in murine leishmaniasis. *Acta Tropica* 60(3): 201–210.
303. Dacanay, E.P., Laurel, O. & Manalo, J.B., 1972. Clinical evaluation of National Institute of Science and Technology. Produced allergenic extracts part 2. Hypo sensitization injection treatment with pollen extracts. *Philippine Journal of Science* 101(1–2): 15–30.

304. Dahl, T.A., Bilski, P., Reszka, K.J. & Chignell, C.F., 1994. Photocytotoxicity of curcumin. *Photochemistry and Photobiology* 59(3): 290-294.
305. Dahmen, K., Pachaly, P. & Zymalkowski, F., 1977. Alkaloide aus der thailändischen Menispermaceen-Droge Krung Kha Mao (*Cyclea barbata*), 5. Mitt. Isolierung und Strukturaufklärung weiterer Bisbenzylisochinolin-Alkaloide [Alkaloids from the Thai Menispermaceae drug krung kha mao (*Cyclea barbata*), part 5. Isolation and structural elucidation of further bisbenzylisoquinoline alkaloids]. *Archiv der Pharmazie (Weinheim)* 310(2): 95-102.
306. Dam Trung Bao et al., 1984. Study on selen-rich medicinal plants used in Vietnamese traditional medicine. *Tap chi Duoc hoc* 4: 10-14. (in Vietnamese)
307. Damayanti, M., Susheela, K. & Sharma, G. J., 1996. Effect of plant extracts and systemic fungicide on the pineapple fruit-rotting fungus, *Ceratocystis paradoxa*. *Cytobios* 86(346): 155-165.
308. Damodaran, S. & Venkataraman, S., 1994. A study on the therapeutic efficacy of *Cassia alata*, Linn. leaf extract against *Pityriasis versicolor*. *Journal of Ethnopharmacology* 42(1): 19-23.
309. Damtoft, S., Godthjaelpsen, L., Jensen, S.R. & Nielsen, B.J., 1983. Age-dependent variations of the efficiency of iridoid biosynthesis in *Verbena officinalis*. *Phytochemistry* 22(11): 2614-2615.
310. Dan Thi Mai et al., 1982. Cao long *Curcuma xanthorrhiza* (Zingiberaceae) lam ha cholesterol huyet thuc nghiem [Liquid balm of *Curcuma xanthorrhiza* (Zingiberaceae) can reduce blood cholesterol]. *Tap chi Duoc hoc* 2: 10-12. (in Vietnamese)
311. Dandekar, U.P., Chandra, R.S., Dalvi, S.S., Joshi, M.V., Gokhale, P.C., Sharma, A.V., Shah, P.U. & Kshirsagar, N.A., 1992. Analysis of a clinically important interaction between phenytoin and Shankhapushpi, an Ayurvedic preparation. *Journal of Ethnopharmacology* 35(3): 285-288.
312. Dans, L., 1988. Herbal medicine and drug substitution: report on lagundi, yerba buena and akapulko. Vol. 2, University of the Philippines at Manila (UPM), Manila, Philippines. 96 pp.
313. Dao Lan Phuong, 1990. Chemical composition of Vietnamese *Amomum xanthioides* essential oil. *Tap chi Duoc hoc* 1: 17-19. (in Vietnamese)
314. Darwis SN, Madjo Indo, A.B.D. & Hasiyah, S., 1991. Tumbuhan obat famili Zingiberaceae [Medicinal plants of the Zingiberaceae]. Seri Pengembangan No. 17. Pusat Penelitian dan Pengembangan Tanaman Industri, Bogor, Indonesia. 103 pp.
315. Das, A. & Mallick, R., 1991. Correlation between genomic diversity and asiaticoside content in *Centella asiatica* (L.) Urban. *Botanical Bulletin of Academia Sinica* 32(1): 1-8.
316. Dasgupta, B., Sinha, N.K., Pandey, V.B. & Ray, A.B., 1984. Major alkaloid and flavonoid of *Premna integrifolia*. *Planta Medica* 50 (3): 281.
317. Dat, D.D., Ham, N.N., Khac, D.H., Lam, N.T., Son, P.T., Van Dau, N., Grabe, M., Johansson, R., Lindgren, G. & Stjernstrom, N.E., 1992. Studies on the individual and combined diuretic effects of four Vietnamese traditional herbal remedies: *Zea mays*, *Imperata cylindrica*, *Plantago major* and *Orthosiphon stamineus*. *Journal of Ethnopharmacology* 36(3): 225-231.

318. Datta, K. & Datta, S.K., 1984. Auxin induced clonal multiplication of *Holarrhena antidysenterica* by tissue culture. *Journal of Tree Sciences* 3(1-2): 45-52.
319. Daulatabad, C.D., Hosamani, K.M. & Alagawadi, K.R., 1987. Epoxy and cyclopropanoid fatty acids in cassia seed oils. *Journal of the Oil Technologists' Association of India* 19(2): 41-42.
320. Daunay, M.-C., Lester, R.N. & Laterrot, H., 1991. The use of wild species for the genetic improvement of brinjal egg-plant (*Solanum melongena*) and tomato (*Lycopersicon esculentum*). In: Hawkes, J.G., Lester, R.N., Nee, M. & Estrada, N. (Editors): *Solanaceae III: Taxonomy, chemistry, evolution*. The Royal Botanic Gardens, Kew, Richmond & The Linnean Society of London, London, United Kingdom. pp. 389-412.
321. Davenport, G.J., 1918. *Quisqualis indica*, a substitute for santonin. *China Medical Journal (Shanghai)* 32: 133.
322. Davis, R.H., Kabbani, J.M. & Maro, N.P., 1987. Aloe vera and wound healing. *Journal of the American Podiatric Medical Association* 77(4): 165-169.
323. Davis, R.H., Leitner, M.G., Russo, J.M. & Byrne, M.E., 1989. Wound healing, oral and topical activity of Aloe vera. *Journal of the American Podiatric Medical Association* 79(11): 559-562.
324. Day, C., Cartwright, T., Provost, J. & Bailey, C.J., 1990. Hypoglycaemic effect of *Momordica charantia* extracts. *Planta Medica* 56(5): 426-429.
325. Dayrit, F.M., Lapid, M.R.G., Cagampang, J.V. & Lagurin, L.G., 1987. Phytochemical studies on the leaves of *Vitex negundo*, L. ('lagundi'). I. Investigations of the bronchial relaxing constituents. *Philippine Journal of Science* 116(4): 403-410.
326. de Guzman, C.C. & Siemonsma, J.S. (Editors), [in preparation]. *Plant Resources of South-East Asia No 13. Spices*. Backhuys Publishers, Leiden, the Netherlands.
327. de Guzman, E.D., Umali, R.M. & Sotalbo, E.D., 1986. Guide to the Philippine flora and fauna. Vol. 3: Dipterocarps, non-dipterocarps. Natural Resources Management Centre, Ministry of Natural Resources, Quezon City & University of the Philippines, Los Baños, the Philippines. 414 pp.
328. de Jong, A.W.K., 1948. Coca. In: van Hall, C.J.J. & van de Koppel, C. (Editors). *De Landbouw in de Indische Archipel [The agriculture in the Indonesian Archipelago]*. Vol. 2a. W. van Hoeve, 's-Gravenhage, the Netherlands. pp. 866-888.
329. de Jong, A.W.K., 1948. Temoe lawak (*Curcuma xanthorrhiza* Roxb.). In: van Hall, C.J.J. & van de Koppel, C. (Editors): *De landbouw in de Indische Archipel [Agriculture in the Indonesian Archipelago]*. Vol. 2a. W. van Hoeve, 's-Gravenhage, the Netherlands. pp. 896-901.
330. de Kruif, A.P.M., 1981. A revision of *Holarrhena* R. Br. (Apocynaceae). *Mededelingen Landbouwhogeschool Wageningen* 81-2: 1-36.
331. de Leon, M.R. & Ocampo, V.R., 1989. Acute toxicity of makabuhai (*Tinospora rumphii* Boerlage) in laboratory rats. Pest Control Council of the Philippines, Baguio City, the Philippines. 1 p.
332. de Padua, L.S., Lugod, G.C. & Pancho, J.V., 1977-1983. *Handbook on Philippine medicinal plants*. 4 volumes. Documentation and Information

- Section, Office of the Director of Research, University of the Philippines at Los Baños, the Philippines.
333. de Silva, L.B., Herath, W.H.M.W., Jennings, R.C., Mahendran, M. & Wannigama, G.E., 1982. A new sesquiterpene lactone from *Elephantopus scaber*. *Phytochemistry* 21(5): 1173–1175.
334. de Vogel, E.F., 1980. Seedlings of dicotyledons. Structure, development, types. Descriptions of 150 woody Malesian taxa. Pudoc, Wageningen, the Netherlands. 465 pp.
335. de Vries, C.A., 1974. Sericulture. *Tropical Abstracts* 29: 633–642.
336. de Wit, H.C.D., 1956. A revision of the genus *Cassia* (Caesalp.) as occurring in Malaysia. *Webbia* 11: 197–292.
337. Declerck, M., Smets, P.H., Smets, J. & Roman, J., 1985. Euphorbia project: renewable energy production through the cultivation and processing of semi arid land biomass in Kenya. In: Palz, W., Coombs, J. & Hall, D.O., (Editors): *Energy from biomass, 3rd EC conference. Proceeding of the International Conference on Biomass held in Venice, Italy, 25–29 March 1985.* Elsevier Applied Science Publishers, London, United Kingdom. pp. 310–314.
338. Dehgan, B. & Webster, G.L., 1979. Morphology and infrageneric relationships of the genus *Jatropha* (Euphorbiaceae). University of California Publications in Botany 74. University of California Press, Berkeley & Los Angeles, United States. 73 pp. & 33 plates.
339. del Fierro, R.S., 1983. Studies on the mutagenicity, clastogenicity and antimutagenicity potential of *Tinospora rumphii* Boerlage (Family Menispermaceae). *Philippine Scientist* 20: 1–10.
340. del Rosario, C.E.S. & Molato, A.P., 1994. Evaluation of plant extracts as fungicidal material against major fungal diseases of root crop. Integrated pest management: learning from experience. Pest Management Council of the Philippines (PMCP), College, Laguna, the Philippines.
341. delle Monache, G., Botta, B., Vinciguerra, V., de Mello, J.F. & de Andrade Chiappeta, A., 1996. Antimicrobial isoflavanones from *Desmodium canum*. *Phytochemistry* 41(2): 537–544.
342. Demeyer, K. & Dejaegere, R., 1993. Influence of nitrogen on the alkaloid content of *Datura stramonium*. In: Palevitch, D., Simon, J.E. & Mathé, A. (Editors): *Raw material production, product introduction. Proceedings of the First World Congress on Medicinal and Aromatic Plants for Human Welfare, WOCMAP. Maastricht, the Netherlands, July 19–25, 1992.* *Acta Horticulturae* 331: 35–37.
343. Demeyer, K., Vanhaste, F., van de Velde, H. & Dejaegere, R., 1990. Introductory study for the optimization of growth and alkaloid production by cell cultures of *Datura stramonium* L. *Acta Horticulturae* 306: 210–218.
344. Deng, Q.Y., Ding, C.M., Zhang, W.H. & Lin, Y.C., 1996. Studies on the flavonoid constituents in *Blumea balsamifera*. *Ropuxue Zasshi* 13(5): 447–452. (in Japanese)
345. Desai, A.C. & Bhide, M.B., 1977. Hydnocarpus oil as an antileprotic agent in footpad technique. *Leprosy in India* 49(3): 360–363.
346. Dethier, M., Cordier, Y. & Demeyer, K., 1993. Cultivation of *Datura* species for scopolamine and hyoscyamine production in Burundi. In:

- Palevitch, D., Simon, J.E. & Mathé, A. (Editors): Raw material production, product introduction. Proceedings of the First World Congress on Medicinal and Aromatic Plants for Human Welfare, WOCMAP. Maas-tricht, the Netherlands, 19-25 July 1992. *Acta Horticulturae* 331: 39-48.
347. Dethier, M. & de Luca, V., 1993. Partial purification of an N-methyltrans-ferase involved in vindoline biosynthesis in *Catharanthus roseus*. *Phyto-chemistry* 32(3): 673-678.
348. Dey, D. & Das, M.N., 1988. Pharmacognosy of antidysenteric drugs of Indian medicine. *Acta Botanica Indica* 16(2): 216-226.
349. Dhar, M.L., Dhar, M.M., Dhawan, B.N., Mehrotra, B.N. & Ray, C., 1968. Screening of Indian plants for biological activity : Part I. *Indian Journal of Experimental Biology* 6: 232-247, 414.
350. Dharma, A.P., 1981. Indonesische geneeskrachtige planten [Indonesian medicinal plants]. De Driehoek, Amsterdam, the Netherlands. 168 pp.
351. Dhir, H., Agarwal, K., Sharma, A. & Talukder, G., 1991. Modifying role of *Phyllanthus emblica* and ascorbic acid against nickel clastogenicity in mice. *Cancer Letters*: 59(1): 9-18.
352. Dhir, H., Roy, A.K. & Sharma, A., 1993. Relative efficiency of *Phyllanthus emblica* fruit extract and ascorbic acid in modifying lead and aluminium-induced sister-chromatid exchanges in mouse bone marrow. *Environmental and Molecular Mutagenesis* 21(3): 229-236.
353. Dimetry, N.Z., El-Gengaihi, S., Reda, A.S. & Amer, S.A.A., 1992. Biol-ogical effects of some isolated *Abrus precatorius* L. alkaloids towards *Tetranychus urticae* Koch. *Anzeiger für Schädlingkunde, Pflanzen-schutz, Umweltschutz* 65(5): 99-101.
354. Dinda, B. & Chel, G., 1992. 6-Hydroxyplumbagin, a naphthoquinone from *Plumbago indica*. *Phytochemistry* 31(10): 3652-3653.
355. Dinda, B., Chel, G. & Achari, B., 1994. A dihydroflavonol from *Plumbago indica*. *Phytochemistry* 35(4): 1083-1084.
356. Ding Hou, 1984. Aristolochiaceae. In: van Steenis, C.G.G.J. & de Wilde, W.J.J.O. (General editors): *Flora Malesiana*. Series 1, Vol. 10. Kluwer Academic Publishers, Dordrecht, the Netherlands. pp. 53-108.
357. Ding Hou, Larsen, K. & Larsen, S.S., 1996. Caesalpiniaceae (Legumi-nosae-Caesalpinioideae). In: Kalkman, C., Kirkup, D.W., Nootboom, H.P., Stevens, P.F. & de Wilde, W.J.J.O. (Editors): *Flora Malesiana*. Series 1, Vol. 12(2). Rijksherbarium/Hortus Botanicus, Leiden University, the Netherlands. pp. 409-730.
358. Divinagracia, G.G. & Ros, L.B., 1985. Note: diseases of selected medicinal plants in the Philippines. *Philippine Agriculturist* 68(2): 297-308.
359. Dixit, O.P. & Saxena, R.C., 1990. Insecticidal action of *Premna integrifo-lia* against *Callosobruchus chinensis* (Coleoptera: Bruchidae). *Pesticides* 24(1): 29-31.
360. Djordjevic, S. & Gorunovic, M., 1991. Etude de *Sophora japonica* L. comme source d'obtention du rutoside [A study of *Sophora japonica* L. as a source of rutoside]. *Herba Hungarica* 30(1-2): 11-15.
361. Do Tat Loi, 1995. Medicinal plants and traditional remedies in Vietnam. 7th Edition. Science and Technics Publishing House, Hanoi, Vietnam. 1485 pp. (in Vietnamese)
362. Doan, D.D., Nguyen, N.H., Doan, H.K., Nguyen, T.L., Phan, T.S., Van-

- Dau, N., Grabe, M., Johansson, R., Lindgren, G. & Stjernstrom, N.E., 1992. Studies on the individual and combined diuretic effects of four Vietnamese traditional herbal remedies (*Zea mays*, *Imperata cylindrica*, *Plantago major* and *Orthosiphon stamineus*). *Journal of Ethnopharmacology* 36(3): 225-231.
363. Doan Thi Nhu, Nguyen Thuong Thuc, Do Huy Bich & Vu Thuy Huyen (Editors), 1991. Les plants médicinales au Vietnam. Livre 1. Médecine traditionnelle et pharmacopée [The medicinal plants of Vietnam. Volume 1. Traditional medicine and pharmacopoeia]. Agence de coopération Culturelle et Technique, Paris, France. 201 pp.
364. Doan Thi Nhu, Do Huy Bich, Pham Kim Man, Nguyen Thuong Thuc, Bui Xuan Chuong & Pham Duy Mai (Editors), 1993. Les plants médicinales au Vietnam. Livre 2. Médecine traditionnelle et pharmacopée [The medicinal plants of Vietnam. Volume 2. Traditional medicine and pharmacopoeia]. Agence de coopération Culturelle et Technique, Paris, France. 189 pp.
365. Doll, R., 1974. Die Gattung *Taraxacum* [The genus *Taraxacum*]. A. Ziemsen Verlag, Wittenberg, Germany. 158 pp.
366. Dong, T.X. et al., 1993. Investigation of hemagglutinating activity in seeds of various *Trichosanthes* species family Cucurbitaceae and comparison of lectins isolated from seeds and tubers of *Trichosanthes kirilowii*. *International Journal of Biochemistry* 25(3): 411-414.
367. Dong, T.X., Ng, T.B., Yeung, H.W. & Wong, R.N.S., 1994. Isolation and characterization of a novel ribosome-inactivating protein  $\beta$ -kirilowin, from the seeds to *Trichosanthes kirilowii*. *Biochemical and Biophysical Research Communications* 199(1): 387-393.
368. Donzanti, B.A. & Uretsky, N.J., 1983. Effects of excitatory amino acids on locomotor activity after bilateral microinjection into the rat nucleus accumbens possible dependence on dopaminergic mechanisms. *Neuropharmacology* 22(8): 971-981.
369. Dorsch, W., Dumoulin, S., Stuppner, H. & Wagner, H., 1990. Screening of drugs of traditional medicine for new antiasthmatic active principles. *Planta Medica* 56(6): 683.
370. Doshi, J.C., Vaidya, A.B., Antarkar, D.S., Deolalikar, R. & Antani, D.H., 1994. A two-stage clinical trial of *Phyllanthus amarus* in hepatitis B carriers: failure to eradicate the surface antigen. *Indian Journal of Gastroenterology* 13(1): 7-8.
371. Dreisbach, R.H., 1983. Handbook of poisoning. 11<sup>th</sup> Edition. Lange Medical Publications, Los Altos, California, United States. 632 pp.
372. Dressler, R.L., 1962. A synopsis of *Poinsettia* (Euphorbiaceae). *Annals of the Missouri Botanical Garden* 48: 329-341.
373. Dubey, N.K. & Kishore, N., 1987. Fungitoxicity of some higher plants and synergistic activity of their essential oils. *Tropical Science* 27(1): 23-27.
374. Dubey, N.K., Kishore, N., Srivastava, O.P., Dikshit, A. & Singh, S.K., 1983. Fungitoxicity of some higher plants against *Rhizoctonia solani*. *Plant and Soil* 72(1): 91-94.
375. Dugan, G.H. & Gumbmann, M.R., 1990. Toxicological evaluation of sicklepod and black nightshade seeds in short term feeding studies in rats. *Food and Chemical Toxicology* 28(2): 101-107.



376. Duke, J.A., 1973. Utilization of papaver. *Economic Botany* 27: 390–400.
377. Duke, J.A., 1981. Handbook of legumes of world economic importance. Plenum Press, New York, United States & London, United Kingdom. 345 pp.
378. Duke, J.A., 1985. Handbook of medicinal herbs. CRC Press, Inc., Boca Raton, Florida, United States. 677 pp.
379. Duke, J.A., Aulik, D. & Plowman, T., 1975. Nutritional value of coca. *Botanical Museum Leaflets, Harvard University* 24(6): 113–119.
380. Dulawan, M.J.K. & Soriano, N.R., 1991. Antibacterial property of gatas-gatas plant. BSc thesis pharmacy. Manila Central University, the Philippines.
381. Durga, R., Sridhar, P. & Polasa, H., 1990. Effects of plumbagin on antibiotic resistance in bacteria. *Indian Journal of Medical Research* 91: 18–20.
382. Duriyaprapan, S. & Britten, E.J., 1986. The effect of photoperiod on flowering and oil production of Japanese mint under controlled environmental conditions. *Thai Journal of Agricultural Science* 19(4): 313–320.
383. Durodola, J.I., 1977. Antibacterial property of crude extracts from a herbal wound healing remedy – *Ageratum conyzoides*. *Planta Medica* 32(4): 388–390.
384. Dutta, S.C., Saha, B.N., Pathak, M.G. & Mathur, R.K., 1989. Essential oil of *Blumea lanceolaria* (Roxb.) Druce. *Indian Perfumer* 33(1): 38–39.
385. Dutta, T.R., Ahmed, R., Abbas, S.R. & Rao, M.K.V., 1985. Plants used by Andaman aborigines in gathering rock-bee honey. *Economic Botany* 39(2): 130–138.
386. Duvall, M.R., Learn, G.H., Eguiarte, L.E. & Clegg, M.T., 1993. Phylogenetic analysis of *rbcL* sequences identifies *Acorus calamus* as the primal extant monocotyledon. *Proceedings of the National Academy of Sciences of the United States of America* 90(10): 4641–4644.
387. Dwijendra Singh & Tripathi A.K., 1984. Insect-pest-complex of opium poppy in India and their control. *Pesticides* 18(4): 33–34.
388. Dwivedi, R.K. & Tripathi, Y.C., 1991. Pharmacognostical, phytochemical and biological studies on *Holarrhena antidysenterica* Wall: a review (part-1). *New Agriculturist* 1(2): 209–212.
389. Dy Phon, P., Ohashi, H. & Vidal, J.E., 1994. Légumineuses – Desmodiées [Leguminosae (Fabaceae) Papilionoideae – Desmodieae]. In: Lescot, M., Vidal, J.E. & Vidal, Y. (Editors): *Flore du Cambodge, du Laos et du Viêt Nam* [Flora of Cambodia, Laos and Vietnam]. Vol. 27. Muséum National d'Histoire Naturelle, Paris, France. 154 pp.
390. Edwards, M.E., Harris, E.M., Wagner, F.H., Cross, M.C. & Miller, G.S., 1988. Seed germination of American pokeweed (*Phytolacca americana*). *American Journal of Botany* 75(12): 1794–1802.
391. Eisikowitch, D. & Rotem, R., 1987. Flower orientation and color change in *Quisqualis indica* and their possible role in pollinator partitioning. *Botanical Gazette* 148: 175–179.
392. El-Mekkawy, S., Meselhy, M.R., Kusumoto, I.T., Kadota, S., Hattori, M. & Namba, T., 1995. Inhibitory effects of Egyptian folk medicines on human immunodeficiency virus (HIV) reverse transcriptase. *Chemical and Pharmaceutical Bulletin* 43(4): 641–648.
393. El-Naggar, L. et al., 1978. A note on the isolation and identification of two pharmacologically active constituents of *Euphorbia pilulifera*. *Lloydia*

- 41(1): 73–75.
394. El-Sohly, H., Croom, E.M., El-Feraly, F.S. & El-Sherei, 1990. A large scale extraction technique of artemisinin from *Artemisia annua*. *Journal of Natural Products* 53: 1560–1564.
395. Emboden, W.A., 1980. *Centella asiatica*: elixir of life? *Pacific Horticulture* 41(3): 16–19.
396. Emmanuel, C.J.S.K., Kapoor, M.L. & Sharma, V.K., 1992. Three decades of forest genetics and tree improvement. *Indian Forester* 118: 489–500.
397. Emmyzar, Karawati, E. & Taryono, 1989. Pola tanam tumpang sari *Angelica acutiloba* dan sayuran [Intercropping *Angelica acutiloba* with vegetables]. *Buletin Penelitian Tanaman Rempah dan Obat* 4(2): 70–74.
398. Emmyzar & Rahmat, E.M., 1993. Budidaya dan peluang pengembangan saga manis [Cultural practices and development opportunity for *Abrus precatorius*]. *Warta Tumbuhan Obat Indonesia* 2(2): 30–31.
399. European Scientific Cooperative on Phytotherapy (ESCO), 1996. Monographs on the medicinal uses of plant drugs. Fascicule 1. Monograph on *Orthosiphonis folium*. ESCO, European Scientific Cooperative on Phytotherapy, Exeter, Devon, United Kingdom.
400. European Scientific Cooperative on Phytotherapy (ESCO), 1997. Monographs on the medicinal uses of plant drugs. Fascicule 2. Monograph on *Taraxaci folium*. Monograph on *Taraxaci radix*. ESCO, European Scientific Cooperative on Phytotherapy, Exeter, Devon, United Kingdom.
401. Eussen, J.H.H., 1978. Studies on the tropical weed *Imperata cylindrica* (L.) Beauv. var. *major*. Thesis, Rijksuniversiteit Utrecht, the Netherlands. 140 pp.
402. Evans, W.C. (Editor), 1989. Trease and Evans' pharmacognosy. 13th Edition. Baillière Tindall, London, United Kingdom. 832 pp.
403. Exell, A.W., 1954. Combretaceae. In: van Steenis, C.C.C.J. (General editor): *Flora Malesiana*. Series 1, Vol. 4. Noodhoff-Kolff N.V., Djakarta, Indonesia. pp. 533–589.
404. Falvey, J.L., 1981. *Imperata cylindrica* and animal production in South-east Asia: a review. *Tropical Grasslands* 15: 52–56.
405. Fang, S.D., Xu, R.S. & Gao, Y.S., 1981. Some recent advances in the chemical studies of Chinese herbal medicine. *American Journal of Botany* 68(2): 300–303.
406. FAO, 1994. FAO production yearbook 1993. Vol. 47. Food and Agriculture Organization, Rome, Italy.
407. Faridah Hanum, I. & van der Maesen, L.J.G. (Editors), 1997. Plant Resources of South-East Asia No 11. Auxiliary plants. Backhuys Publishers, Leiden, the Netherlands. 389 pp.
408. Farkas, A., 1963. Topical medicament containing Aloe polyuronides for treatment of wounds and burns. U.S. Patent 3, 360, 510 (Cl. 107-58 ) Sept 10, 1963.
409. Farooqi, M.I.H., Kapoor, V.P. & Islam, G., 1978. Seeds of genus *Cassia* as possible sources of industrial gums. *Indian Forester* 104(11): 729–733.
410. Ferdous, A.J, Islam, S.N., Ahsan, M. & Faroque, A.B.M., 1990. Antibacterial activity of the leaves of *Adhatoda vasica*, *Calotropis gigantea*, *Nerium odorum* and *Ocimum sanctum*. *Bangladesh Journal of Botany* 19(2): 227–229.

411. Fernandez-Banares, F. et al., 1999. Randomized clinical trial of *Plantago ovata* seeds (dietary fibre) as compared with mesalamine in maintaining remission in ulcerative colitis. Spanish group for the study of Crohn's Disease and Ulcerative Colitis (GETECCU). *American journal of gastroenterology* 94(2): 427-433.
412. Ferrara, L., Schettino, O. & Montesano, D., 1996. Triterpenoids from *Cardiospermum halicacabum* L. *Phytotherapy Research* 10 (supp.1): S192-S194.
413. Ferreira, L.A.F. et al., 1992. Antivenom and biological effects of ar-turmerone isolated from *Curcuma longa* (Zingiberaceae). *Toxicon* 30(10): 1211-1218.
414. Flach, M. & Rumawas, F. (Editors), 1996. *Plant Resources of South-East Asia No 9. Plants yielding non-seed carbohydrates*. Backhuys Publishers, Leiden, the Netherlands. 239 pp.
415. Florentino, C., Ocampo, P.P. & Ocampo, V.R., 1993. Effects of lagundi (*Vitex negundo* L.) crude extracts on some reproductive parameters of adult male albino rats. In: *Proceedings of the twenty-fourth Annual Scientific Meeting of the Pest Management Council of the Philippines*. College, Laguna, the Philippines. p. 43.
416. Flores, E.M., Rivera, D.I. & Vasquez, N.M., 1986. Germinacion y desarrollo de la plantula de *Cassia grandis* L. (Caesalpinioideae) [Germination and development of the seedling of *Cassia grandis* L. (Caesalpinioideae)]. *Revista de Biologia Tropical* 34(2): 289-296.
417. Fodstad, O., Johannessen, J.V., Schjerven, L. & Pihl, A., 1979. Toxicity of abrin and ricin in mice and dogs. *Journal of Toxicology and Environmental Health* 5(6): 1073-1084.
418. Fojas, F.R., Garcia, L.L., Venzon, E.L., Sison, F.M., Villanueva, B.A., Fojas, A.J. & Llave, I., 1986. Pharmacological studies on *Jatropha curcas* as a possible source of anti-arrhythmic (beta-blocker) agent. *The Philippine Journal of Science* 115: 317-328.
419. Foo, L.Y., 1993. Amariin, a di-dehydrohexahydroxydiphenoyl hydrolysable tannin from *Phyllanthus amarus*. *Phytochemistry* 33(2): 487-491.
420. Foo, L.Y. & Wong, H., 1992. Phyllanthusiin D, an unusual hydrolysable tannin from *Phyllanthus amarus*. *Phytochemistry* 31(2): 711-713.
421. Forman, L.L. 1986. Menispermaceae. In: van Steenis, C.G.G.J. & de Wilde, W.J.J.O. (General editors): *Flora Malesiana*. Series 1, Vol. 10. Kluwer Academic Publishers, Dordrecht, the Netherlands. pp. 157-253.
422. Forman, L.L., 1988. A synopsis of Thai Menispermaceae. *Kew Bulletin* 43(2): 369-407.
423. Forman, L.L., 1991. Menispermaceae. In: Smitinand, T. & Larsen, K. (Editors): *Flora of Thailand*. Vol. 5(3). The Forest Herbarium, Royal Forest Department, Bangkok, Thailand. pp. 325-331.
424. Forni, E., Trifilo, A. & Polesello, A., 1983. Researches on the utilization of the pigment from *Phytolacca decandra* L. as a food colorant: part 1 - preparation of an extract free from toxic substances. *Food Chemistry* 10(1): 35-46.
425. Forni, E., Trifilo, A. & Polesello, A., 1984. Researches on the utilization of the pigment from *Phytolacca decandra* L. as a food colorant: part 2 - tests on pigmenting power and stability of phytolaccanin in model solutions.

- Food Chemistry 13(2): 149-160.
426. Forster, P.I. & Clifford, H.T., 1986. Aloeaceae. Flora of Australia 46: 66-70.
427. Fosberg, F.R. & Sachet, H.H., 1989. Three cultivated *Ixoras* (Rubiaceae). *Baileya* 23(2): 74-85.
428. Fournier, P., 1948. Le livre des plantes médicinales et vénéneuses de France [The book on medicinal and poisonous plants of France]. Vol. 3. Paul Lechevalier, Paris, France. 636 pp.
429. Fowler, M.W., 1981. Plant cell biotechnology to produce desirable substances. *Chemistry and Industry* 7: 229-233.
430. Fox, R.B., 1952. The Pinatubo Negritos: their useful plants and material culture. *Philippine Journal of Science* 81(3-4): 173-391.
431. Franz, Ch., 1993. Genetics. In: Hay, R.K.M. & Waterman, P.G. (Editors): Volatile oil crops. Longman, Burnt Mill, Harlow, Essex, United Kingdom. pp. 63-96.
432. Fravel, D.R., Stosz, S.K. & Larkin, R.P., 1996. Effect of temperature, soil type, and matric potential on proliferation and survival of *Fusarium oxysporum* f.sp. *erythroxyli* from *Erythroxylum coca*. *Phytopathology* 86(3): 236-240.
433. Freiburghaus, F., Kaminsky, R., Nkunya, M.H.H. & Brun, R., 1996. Evaluation of African medicinal plants for their in vitro trypanocidal activity. *Journal of Ethnopharmacology* 55: 1-11.
434. Frodin, D.G., 1986. Studies in *Schefflera* (Araliaceae), II. Northern Luzon (Philippines) species of the *Heptapleurum* group. *Proceedings of the Academy of Natural Sciences of Philadelphia* 138: 403-425.
435. Frodin, D.G., 1990. Studies in *Schefflera* (Araliaceae), IV. The identity of *Vitis heptaphylla* L., a long-misplaced Linnean ivy tree. *Botanical Journal of the Linnean Society* 104: 309-424.
436. Fujii, Y., Furukawa, M., Hayakawa, Y., Sugahara, K. & Shibuya, T., 1991. Survey of Japanese medicinal plants for the detection of allelopathic properties. *Weed Research (Tokyo)* 36(1): 36-42.
437. Fujimoto, Y., Soemartono, A. & Sumatra, M., 1988. Sesquiterpenes from *Blumea balsamifera*. *Phytochemistry* 27: 1109-1111.
438. Fujimoto, Y., Suzuki, Y., Kanaiwa, T., Amiya, T., Hoshi, K. & Fujino, S., 1983. Studies on the Indonesian *Antiaris toxicaria* sap. *Journal of Pharmacobiodynamics* 6(2): 128-135.
439. Fujita, K., Teradaira, R. & Nagatsu, T., 1976. Bradykinase activity of aloe extract. *Biochemical Pharmacology* 25(2): 205.
440. Fukamiya, N. & Lee, K.H., 1986. Antitumor agents, 81. Justicidin-A and diphyllin, two cytotoxic principles from *Justicia procumbens*. *Journal of Natural Products* 49(2): 348-350.
441. Fukamiya, N., Okano, M., Aratani, T., Negoro, K., Lin, Y.M. & Lee, K.H., 1987. Antitumor agents, 87. Cytotoxic antileukemic canthin-6-one alkaloids from *Brucea antidysenterica* and the structure activity relationships of their related derivatives. *Planta Medica* 53(2): 140-143.
442. Fukamiya, N., Okano, M., Aratani, T., Negoro, K., McPhail, A.T., Ju-ichi, M. & Lee, K.H., 1986. Antitumor agents, 79. Cytotoxic antileukemic alkaloids from *Brucea antidysenterica*. *Journal of Natural Products* 49(3): 428-434.

443. Fukamiya, N., Okano, M., Tagahara, K., Aratani, T., Muramoto, Y. & Lee, K.H., 1987. Antitumor agents, 90. Bruceantinoside C, a new cytotoxic quassinoid glycoside from *Brucea antidysenterica*. *Journal of Natural Products* 50(6): 1075-1079.
444. Fukuda, N., Nakamura, M., Yonemitsu, M., Kimura, T., Isobe, R. & Komori, T., 1993. Studies on the constituents of the stems of *Tinospora tuberculata* 1. Isolation and structure elucidation of two new furanoid diterpenes tinotufolin A and B. *Liebigs Annalen der Chemie* 10(3): 325-327.
445. Fukuda, N., Yonemitsu, M. & Kimura, T., 1983. The constituents of the stems of *Tinospora tuberculata* 1. N-trans feruloyl tyramine and N-cis feruloyl tyramine and a new phenolic glucoside tintotuberide. *Chemical and Pharmaceutical Bulletin* 31(1): 156-161.
446. Fukuda, N., Yonemitsu, M. & Kimura, T., 1986. Studies on the constituents of the stems of *Tinospora tuberculata* 3. New diterpenoids borapetoside B and borapetol B. *Chemical and Pharmaceutical Bulletin* 34(7): 2868-2872.
447. Fukuda, N., Yonemitsu, M. & Kimura, T., 1993. Studies on the constituents of the stems of *Tinospora tuberculata* 4. Isolation and structure elucidation of the five new furanoid diterpene glycosides borapetoside C-G. *Liebigs Annalen der Chemie* 0(5): 491-495.
448. Fukuda, N., Yonemitsu, M., Kimura, T., Hachiyama, S., Miyahara, K. & Kawasaki, T., 1985. Studies on the constituents of the stems of *Tinospora tuberculata* 2. New diterpenoids borapetoside A and borapetol A. *Chemical and Pharmaceutical Bulletin* 33(10): 4438-4444.
449. Fukunaga, T., Miura, T., Furuta, K. & Kato, A., 1997. Hypoglycemic effect of the rhizomes of *Smilax glabra* in normal and diabetic mice. *Biological and Pharmaceutical Bulletin* 20(1): 44-46.
450. Fukuyama, Y., Hasegawa, T., Toda, M., Kodama, M. & Okazaki, H., 1992. Structures of americanol A and isoamericanol A having neurotrophic properties from the seeds of *Phytolacca americana*. *Chemical and Pharmaceutical Bulletin* 40(1): 252-254.
451. Fullas, F., Choi, Y.-H., Kinghorn, A.D. & Bunyapraphatsara, N., 1990. Sweet-tasting triterpene glycoside constituents of *Abrus fruticulosus*. *Planta Medica* 56(3): 332-333.
452. Funayama, S. & Hikino, H., 1979. Hypotensive principles of *Phytolacca* roots. *Journal of Natural Products* 42(6): 672-674.
453. Furstenberger, G. & Hecker, E., 1986. On the active principles of the Euphorbiaceae, XII. Highly unsaturated irritant diterpene esters from *Euphorbia tirucalli* originating from Madagascar. *Journal of Natural Products* 49(3): 386-397.
454. Gabriel, B.P. & Balatibat, J.B., 1985. Preliminary studies on arthropod pests of some medicinal plants. In: *Pest Control Council of the Philippines (Editor): Proceedings of the thirteenth anniversary and annual convention of the Pest Control Council of the Philippines*. College, Laguna, the Philippines. p. 67.
455. Gagnepain, F. 1908. Zingibéracées [Zingiberaceae]. In: *Gagnepain, F. (Editor): Flore générale de l'Indo-Chine [General flora of Indo-China]*. Vol. 6. Masson & Cie, Paris, France. pp. 25-121.
456. Gagnepain, F., 1920. Hypéricacées [Hypericaceae]. In: *Lecomte (Editor):*

- Flore générale de l'Indo-Chine. [General flora of Indo-China]. Vol. 1. Masson & Cie, Paris, France. pp. 284–287.
457. Gagnepain, F., 1921. Cucurbitacées [Cucurbitaceae]. In: Gagnepain, F. (Editor): Flore générale de l'Indo-Chine [General flora of Indo-China]. Vol. 2. Masson & Cie, Paris, France. pp. 1030–1095.
458. Gagnepain, F., 1924. Composées [Compositae]. In: Gagnepain, F. (Editor): Flore générale de l'Indo-Chine [General flora of Indo-China]. Vol. 3. Masson & Cie, Paris, France. pp. 448–663.
459. Gagnepain, F., 1928–1929. Moracées [Moraceae]. In: Gagnepain, F. (Editor): Flore générale de l'Indo-Chine [General flora of Indo-China]. Vol. 5. Masson & Cie, Paris, France. pp. 694–828.
460. Gagnepain, F., 1933. Oléacées [Oleaceae]. In: Gagnepain, F. (Editor): Flore general de l'Indo-Chine. [General flora of Indo-China]. Vol. 3 part 8. Masson et Cie, Paris, France. pp. 1034–1084.
461. Gaiind, K.N. & Gupta, R.L., 1971. Flavonoid glycosides from *Kalanchoe pinnata*. *Planta Medica* 20(4): 368–373.
462. Gaiind, K.N. & Gupta, R.L., 1972. Alkanes, alkanols, triterpenes and sterols of *Kalanchoe pinnata*. *Phytochemistry* 11: 1500–1502.
463. Galil, J., 1984. *Ficus religiosa*, the tree splitter. *Botanical Journal of the Linnean Society* 88: 185–204.
464. Gallo, V., Suergiu, R., Giovannini, C. & Levi, G., 1987. Glutamate receptor subtypes in cultured cerebella neurons: modulation glutamate and gamma-aminobutyric acid release. *Journal of Neurochemistry* 49(6): 1801–1809.
465. Galvez, J., Crespo, M.E., Jimenez, J., Suarez, A. & Zaruelo, A., 1993. Antidiarrhoeic activity of quercitrin in mice and rats. *Journal of Pharmacy and Pharmacology* 45(2): 157–159.
466. Ganders, F.R., 1979. Heterostyly in *Erythroxylum coca* (Erythroxylaceae). *Botanical Journal of the Linnean Society* 78: 11–20.
467. Ganesan, T., 1994. Antifungal properties of wild plants. *Advances in Plant Sciences* 7(1): 185–187.
468. Ganguly, L.K., 1994. Fungitoxic effect of certain plant extracts against rice blast and brown spot pathogen. *Environment and Ecology* 12(3): 731–733.
469. Ganguly, S. & Khan, E., 1991. Association of *Xiphinema americanum* (Nematoda: Dorylaimida) with root-tip galls of rangoon creeper (*Quisqualis indica* Linn.). *Current Nematology* 2(2): 183–184.
470. Garcia Jr., J.R., 1990. Bioassay of five botanical materials against the bean weevil, *Callosobruchus chinensis* (L.), on mungbean (*Vigna radiata* (L.) Wilczek). University of the Philippines at Los Baños, College, Laguna, the Philippines. 67 pp.
471. Garcia, L.L., Cosme, L.L., Peralta, H.R. & Garcia, B.M., 1973. Phytochemical investigation of *Coleus blumei* Benth. I. Preliminary studies of the leaves. *Philippine Journal of Science* 102(1–2): 1–12.
472. Garcia, L.L., Takahashi, M. & Sato, T., 1978. Phytochemical investigation of *Coleus blumei* Benth. II. Identification of the sterol and hydrocarbon constituents. *Philippine Journal of Science* 107(1–2): 95–102.
473. Garnier, G., Bézanger-Beauquesne, L. & Debraux, G., 1961. Ressources médicinales de la flore Française [Medicinal resources of the French flo-

- ra]. 2 volumes. Vigot Frères, Paris, France. 1511 pp.
474. Garrity, D.P., 1997. Agroforestry innovations for Imperata grassland rehabilitation (Special Issue). *Agroforestry Systems* 36 (1-3): 263-274.
475. Geissberger, P. & Sequin, U., 1991. Constituents of *Bidens pilosa* L.: do the components found so far explain the use of this plant in traditional medicine? *Acta Tropica* 48(4): 251-261.
476. Generalao, M.L., 1977. Root production of *Rauwolfia* (*Rauwolfia*) *serpentina* using different soil media. *Sylvatrop* 2(1): 45-48.
477. Gertlowski, C. & Petersen, M., 1993. Influence of the carbon source on growth and rosmarinic acid production in suspension cultures of *Coleus blumei*. *Plant Cell, Tissue and Organ Culture* 34(2): 183-190.
478. Ghafoor, A., 1985. Moraceae. In: Nasir, E. & Ali, S.I. (Editors): *Flora of Pakistan* No 171. National Herbarium (Stewart Collection), Pakistan Agricultural Research Council, Islamabad, Pakistan. 54 pp.
479. Ghazala Nasim, 1991. Vesicular arbuscular mycorrhizae in two *Curcuma* species (*C. zedoaria* and *C. longa*) of medicinal importance. *Pakistan Journal of Forestry* 41(4): 194-201.
480. Ghorpade, B.R. & Patil, S.P., 1989. Bionomics of *Glyphodes laticostalis* Guenee as one of the most important pests of forest plantations in the Konkan region of Maharashtra State (India). *Indian Journal of Forestry* 129(3): 241-242.
481. Ghosal, S. & Banerjee, P.K., 1969. Alkaloids of the roots of *Desmodium gangeticum*. *Australian Journal of Chemistry* 22: 2029-2031.
482. Ghosal, S., Srivastava, R.S., Banerjee, P.K. & Dutta, S.K., 1971. Alkaloids of *Desmodium triflorum*. *Phytochemistry* 10: 3312-3313.
483. Ghosal, S., Srivastava, R.S., Bhattacharya, S.K. & Debnath, P.K., 1973. *Desmodium* alkaloids. IV: chemical and pharmacological evaluation of *D. triflorum*. *Planta Medica* 23(4): 321-329.
484. Ghosal, S., Tripathi, V.K. & Chauhan, S., 1996. Active constituents of *Emblica officinalis*: Part 1 - The chemistry and antioxidative effects of two new hydrolysable tannins, emblicanin A and B. *Indian Journal of Chemistry. Section B, Organic including Medicinal* 35(9): 941-948.
485. Ghosh, A., Sharma, A. & Talukder, G., 1992. Relative protection given by extract of *Phyllanthus emblica* fruit and an equivalent amount of vitamin C against a known clastogen - caesium chloride. *Food and Chemical Toxicology* 30(10): 865-869.
486. Gildemacher, B.H., Jansen, G.J. & Chayamarit, K., 1993. *Trichosanthes* L. In: Siemonsma, J.S. & Kasem Piluek (Editors): *Plant Resources of South East Asia* No 8. Vegetables. Pudoc Scientific Publishers, Wageningen, the Netherlands. pp. 271-274.
487. Gillett, J.B., Polhill, R.M. & Verdcourt, B., 1971. Leguminosae (Part 3) subfamily Papilionoideae. In: Milne-Redhead, E. & Polhill, R.M. (Editors): *Flora of tropical East Africa*. Crown Agents for Oversea Governments and Administrations, London, United Kingdom. 501 pp.
488. Gillin, F.D., Reiner, D.S. & Suffness, M., 1982. Bruceantin, a potent amoebicide from a plant, *Brucea antidysenterica*. *Antimicrobial Agents and Chemotherapy* 22(2): 342-345.
489. Giri, A.K. & Banerjee, T.S., 1986. Antagonistic activity of herbal drug (*Phyllanthus emblica*) on cytological effects of environmental chemicals on

- mammalian cells. *Cytologia* 51: 375–380.
490. Goda, Y., Shibuya, M. & Sankawa, U., 1987. Inhibitors of the arachidonate cascade from *Allium chinense* and their effect on in vitro platelet aggregation. *Chemical and Pharmaceutical Bulletin* 35(7): 2668–2674.
491. Godwin, H., 1967. The ancient cultivation of hemp. *Antiquity* 41: 42–50.
492. Goel, B. & Kumar, A., 1989. Composition of uncommon foods. *Journal of Food Science and Technology* 26(1): 44–45.
493. Goepel, C., Yupraphat, T., Pachaly, P. & Zymalkowski, F., 1974. Alkaloide aus der thailändischen Menispermaceen-Droge Krung Kha Mao (*Cyclea barbata*), 4. Mitt. Isolierung und Strukturaufklärung zweier Berbamin-Alkaloide [Alkaloids from the Thai Menispermaceae drug krung kha mao (*Cyclea barbata*), part 4. Isolation and structural elucidation of two berbamine alkaloids]. *Planta Medica* 26(1): 94–97.
494. Gomes, C.M.R. et al., 1981. Systematic significance of flavonoids in *Derris* and *Lonchocarpus*. *Biochemical Systematics and Ecology* 9: 129–147.
495. Gonzalez, A.G., Aguiar, Z.E., Grillo, T.A., Luis, J.G., Rivera, A. & Calle, J., 1991a. Chromenes from *Ageratum conyzoides*. *Phytochemistry* 30(4): 1137–1139.
496. Gonzalez, A.G., Aguiar, Z.E., Grillo, T.A., Luis, J.G., Rivera, A. & Calle, J., 1991b. Methoxyflavones from *Ageratum conyzoides*. *Phytochemistry* 30(4): 1269–1271.
497. Goonaratna, C., Thabrew, I. & Wijewardena, K., 1993. Does *Aerva lanata* have diuretic properties? *Indian Journal of Physiology and Pharmacology* 37(2): 135–137.
498. Gouriath, A. & Manoharachary, C., 1989. Effect of pollen on four pathogenic fungi. *Indian Phytopathology* 41(1): 120–122.
499. Gowda, V.N., Gowda, J.V.N. & Vajranabaiyah, S.N., 1991. Regulation of flowering in gundumallige (*Jasminum sambac* Ait.) by foliar application of cycocel. *Current Research University of Agricultural Sciences (Bangalore)* 20(12): 256.
500. Gowrishanker, B. & Vivekanandan, O.S., 1994. In vivo studies of a crude extract of *Phyllanthus amarus* L. in modifying the genotoxicity induced in *Vicia faba* L. by tannery effluents. *Mutation Research* 322(3): 185–192.
501. Gramiccia, G., 1987. Notes on the early history of cinchona plantations. *Acta Leidensia* 55: 5–13.
502. Grange, J.M. & Snell, N.J., 1996. Activity of bromhexine and ambroxol, semi-synthetic derivatives of vasicine, against *Mycobacterium tuberculosis* in vitro. *Journal of Ethnopharmacology* 50(1): 49–53.
503. Grant, G., More, L.J., McKenzie, N.H., Dorward, P.M., Buchan, W.C., Telek, L. & Pusztai, A., 1995. Nutritional and haemagglutination properties of several tropical seeds. *Journal of Agricultural Science* 124(3): 437–445.
504. Grases, F., Melero, G., Costa-Bauza, A., Prieto, R. & March, J.G., 1994. Urolithiasis and phytotherapy. *International Urology and Nephrology* 26(5): 507–511.
505. Green, P.S., 1966. Studies in the genus *Jasminum* III. The species in cultivation in North America. *Baileya* 13(4): 137–172.
506. Green, P.S., 1995. New species and combinations in *Jasminum* especially from Thailand. Studies in the genus *Jasminum* (Oleaceae): XIV. Kew



- Bulletin 50(3): 567-580.
507. Griess, D., Enjalbert, F. & Rech, J., 1994. Diagnostic d'une intoxication par *Phytolacca decandra* L. chez le cheval [Diagnosis of poisoning by *Phytolacca decandra* in horses]. *Revue de Médecine Vétérinaire* 145(2): 133-139.
  508. Grieve, M., 1994. A modern herbal. Tiger Books International, London, United Kingdom. 912 pp.
  509. Grimaudo, S., Tolomeo, M., Gancitano, R.A., d'Alessandro, N. & Aiello, E., 1997. Effects of highly purified anthraquinoid compounds from *Aloe vera* on sensitive and multidrug resistant leukemia cells. *Oncology Reports* 4(2): 341-343.
  510. Grindlay, D. & Reynolds, T., 1986. The *Aloe vera* phenomenon: a review of the properties and modern uses of the leaf parenchyma gel. *Journal of Ethnopharmacology* 16(2-3): 117-151.
  511. Gringauz, A., 1997. Introduction to medicinal chemistry - How drugs act and why. Wiley-VCH, New York, United States.
  512. Gubernator, K. & Böhm, H.J. (Editors), 1998. Structure-based ligand design. In: Manngold, R., Kubinyi, H. & Timmerman, H. (Editors): *Methods and principles in medicinal chemistry*. Vol. 6. Wiley-VCH, Weinheim, Germany.
  513. Guenther, E., 1949. The essential oils. Vol. 3. Van Nostrand, New York, United States. 777 pp.
  514. Guerrero, R.D., Guerrero, C.A. & Garcia, L.L., 1990. Use of indigenous plants as sources of fish toxicants for pond management in the Philippines. *Philippine Technology Journal* 15(2): 15-17.
  515. Guevara, A.P., Lim-Sylianco, C., Dayrit, F. & Finch, P., 1990. Antimutagens from *Momordica charantia*. *Mutation Research* 230(2): 121-126.
  516. Gujar, G.T. & Mehrotra, K.N., 1988. Toxicity and morphogenetic effects of plumbagin on *Dysdercus koeningii* F. (Het., Pyrrhocoridae). *Journal of Applied Entomology* 105: 466-470.
  517. Gulati, R.K., Agarwal, S. & Agrawal, S.S., 1995. Hepatoprotective studies on *Phyllanthus emblica* Linn. and quercetin. *Indian Journal of Experimental Biology* 33(4): 261-268.
  518. Gunaherath, G.M.K.B., Gunatilaka, A.A.L., Cox, P.J., Howie, R.A. & Thomson, R.H., 1988b. A revised structure for plumbazeylanone. *Tetrahedron Letters* 29(6): 719-720.
  519. Gunaherath, G.M.K.B., Gunatilaka, A.A.L., Sultanbawa, M.U.S. & Balasubramaniam, S., 1983. 1,2(3)-Tetrahydro-3,3'-biplumbagin: a naphthalenone and other constituents from *Plumbago zeylanica*. *Phytochemistry* 22(5): 1245-1247.
  520. Gunaherath, G.M.K.B., Gunatilaka, A.A.L. & Thomson, R.H., 1984. Structure of plumbazeylanone: a novel trimer of plumbagin from *Plumbago zeylanica*. *Tetrahedron Letters* 25(42): 4801-4804.
  521. Gunaherath, G.M.K.B., Gunatilaka, A.A.L. & Thomson, R.H., 1988. Studies on medicinal and related plants of Sri Lanka. Part 18. Structure of a new naphthoquinone from *Plumbago zeylanica*. *Journal of the Chemical Society. Perkin Transactions 1, Organic and bio-organic chemistry* 1988: 407-410.

522. Gupta, D. & Singh, J., 1991. Flavonoid glycosides from *Cassia alata*. *Phytochemistry* 30(8): 2761–2763.
523. Gupta, P., Patni, V. & Kant, U., 1994. In vitro shoot differentiation in *Emblca officinalis* Gaertn. *Journal of Phytological Research* 7(2): 171–172.
524. Gupta, P.P., Srimal, R.C. & Tandon, J.S., 1993. Antiallergic activity of some traditional Indian medicinal plants. *International Journal of Pharmacognosy* 31(1): 15–18.
525. Gupta, R., 1990. Indian herbs and herbal raw material – time-tested remedies for all ailments. *Indian Horticulture* 34(4): 39–41.
526. Gupta, R., 1991. Agrotechnology of medicinal plants. In: Wijesekera, R.O.B. (Editor): *The medicinal plant industry*. CRC Press, Boca Raton, United States. pp. 43–57.
527. Gupta, S., Choudry, M.A., Yadava, J.N.S., Srivastava, V. & Tandon, J.S., 1990. Antidiarrheal activity of diterpenes of *Andrographis paniculata* (kal-megh) against *Escherichia coli* enterotoxin in in vivo models. *International Journal of Crude Drug Research* 28: 273–283.
528. Gupta, S., Prabhakar, V.S. & Madan, C.L., 1973. The distribution of total alkaloids and major components in the different organs of *Datura metel* var. *fastuosa* at various stages of growth. *Planta Medica* 23(4): 370–376.
529. Gupta, S., Yadava, J.N.S. & Tandon, J.S., 1993. Antisecretory antidiarrhoeal activity of Indian medicinal plants against *Escherichia coli* enterotoxin-induced secretion in rabbit and guinea pig ileal loop models. *International Journal of Pharmacognosy* 31(3): 198–204.
530. Gupta, S.C., Khanolkar, U.M., Koul, O. & Saxena, B.N., 1977. Pyrethrin synergistic activity by the essential oils of a few *Blumea* species. *Current Science* 46(9): 304–305.
531. Gutierrez, H.G., 1980–1982. An illustrated manual of Philippine materia medica. 2 volumes. Natural Research Council of the Philippines, Tagig, Metro Manila, the Philippines. Vol. 1 (1980) pp. 1–234, Vol. 2 (1982) pp. 235–485.
532. Habtemariam, S., 1995. Cytotoxicity of diterpenes from *Premna schimperi* and *Premna oligotricha*. *Planta Medica* 61: 368–369.
533. Hacker, J.B., 1990. A guide to herbaceous and shrub legumes of Queensland. University of Queensland Press, St Lucia, Australia. 351 pp.
534. Hadipoentyanti, E., 1990. Analisis stabilitas hasil dan mutu minyak *Mentha* spp. [Analysis of oil yield stability and quality of *Mentha* spp.]. *Pemberitaan Penelitian Tanaman Industri* 16(1): 18–23.
535. Hadisoesilo, S., Hartojo & Sudradjat, 1980. Pohon murbei (*Morus macroura* Linn.) sebagai kayu bakar [Mulberry trees (*Morus macroura* Linn.) as fuel wood]. Laporan No 343. Lembaga Penelitian Hutan, Bogor, Indonesia. 12 pp.
536. Haicour, R., 1983. La variabilité de la compatibilité génétique entre divers taxons de *Phyllanthus urinaria* L. (Euphorbiaceae): Mise en évidence et perspectives ouvertes par son analyse [Variation in genetic compatibility between various taxa of *Phyllanthus urinaria* L. (Euphorbiaceae): clarification and open perspectives by its analysis]. *Bulletin de la Société Botanique de France*, 130, *Lettres botaniques*: 207–226.
537. Halford, D.A., 1992. Review of the genus *Oldenlandia* L. (Rubiaceae) and

- related genera in Australia. *Austrobaileya* 3(4): 683-722.
538. Hamilton, R.J., Shih, R.D. & Hoffman, R.S., 1995. Mobitz type I heart block after pokeweed ingestion. *Veterinary and Human Toxicology* 37(1): 66-67.
539. Hammer, M.L.A. & Johns, E.A., 1993. Tapping an Amazonian plethora: four medicinal plants of Macajó Island, Pará (Brazil). *Journal of Ethnopharmacology* 40: 53-75.
540. Han, G.Q., Pau, J.X., Li, C.L. & Tu, F., 1991. The screening of Chinese traditional drugs by biological assay and the isolation of some active components. *International Journal of Chinese Medicine* 16(1): 1-17.
541. Hanan, A., 1996. Beberapa catatan tentang sambiloto [Some notes on sambiloto (*Andrographis paniculata* (Burm.f.) Nees)]. *Warta Tumbuhan Obat Indonesia* 3(1): 19-20.
542. Hanawa, F., Kanauchi, M., Tahara, S. & Mizutani, J., 1995. Lettucenin A as a phytoalexin and its elicitation in dandelion cell cultures. *Journal of the Faculty of Agriculture, Hokkaido University* 66(2): 151-162.
543. Handa, S.S. & Sharma, A., 1990a. Hepatoprotective activity of andrographolide from *Andrographis paniculata* against carbontetrachloride. *Indian Journal of Medical Research* 92: 276-283.
544. Handa, S.S. & Sharma, A., 1990b. Hepatoprotective activity of andrographolide against galactosamine & paracetamol intoxication in rats. *Indian Journal of Medical Research* 92: 284-292.
545. Handjieva, N., Spassov, S., Bodurova, G., Saadi, H., Popov, S., Pureb, O. & Zamjansan, J., 1991. Majoroside, an iridoid glucoside from *Plantago major*. *Phytochemistry* 30(4): 1317-1318.
546. Hano, Y., Mitsui, P. & Nomura, T., 1990a. Two new prenylaurones, antiarones A and B, from the root bark of *Antiaris toxicaria* Lesch. *Heterocycles* 30(2): 1023-1030.
547. Hano, Y., Mitsui, P. & Nomura, T., 1990b. Seven prenylphenols, antiarones C, D, E, F, G, H and I from the root bark of *Antiaris toxicaria* Lesch. *Heterocycles* 31(7): 1315-1324.
548. Hänsel, R. & Kallmann, S., 1986. Identitätsprüfung von *Verbenae Herba*: verbascosid als Leitstoff. [Proof of identity of *Verbenae herba*: verbascoside as the main constituent]. *Archiv der Pharmazie* 319(3): 227-230.
549. Hänsel, R. et al. (Editors), 1992. *Hagers Handbuch der Pharmazeutische Praxis* [Hagers handbook of the practice of pharmacology]. Springer Verlag, Berlin, Germany. 1209 pp.
550. Hansen, B., 1985. Notes on *Andrographis* and *Gymnostachyum* (Acanthaceae). *Nordic Journal of Botany* 5: 353-356.
551. Hansen, K., Nyman, U., Smitt, U.W., Adersen, A., Gudiksen, L., Rajasekharan, S & Pushpangadan, P., 1995. In vitro screening of traditional medicines for anti-hypertensive effect based on inhibition of the angiotensin converting enzyme (ACE). *Journal of Ethnopharmacology* 48(1): 43-51.
552. Harada, S., Otani, H., Maeda, S., Kai, Y., Kasai, N. & Kurihara, Y., 1994. Crystallization and preliminary X-ray diffraction studies of curculin. *Journal of Molecular Biology* 238: 286-287.
553. Harborne, J.B., Boulter, D. & Turner, B.L. (Editors), 1971. *Chemotaxonomy of the Leguminosae*. Academic Press, London, United Kingdom &

- New York, United States. 612 pp.
554. Hardjowasono, M.S., 1942. Gewicht en volume van verschillende vruchten en zaadsoorten [Weight and volume of fruits and seeds]. Korte Mededelingen No 20. Bosbouwproefstation, Buitenzorg, Nederlandsch-Indië. 172 pp.
555. Hardman, J.G., Limbird, L.E., Molinoff, P.B., Ruddon, R.W. & Goodman Gilman, A. (Editors), 1996. Goodman & Gilman's the pharmacological basis of therapeutics. 9<sup>th</sup> Edition. McGraw-Hill, New York, United States. 1905 pp.
556. Harikrishnan, K.N. & Hariharan, M., 1996. Direct shoot regeneration from nodal explants of *Plumbago rosea* Linn. – a medicinal plant. *Phytomorphology* 46(1): 53–58.
557. Hasanah, M., Rachmat, E.M. & Wahab, M.I., 1993. Studi pemecahan dormansi pada benih saga, *Abrus precatorius* L. [A study on seed dormancy breaking in saga, *Abrus precatorius* L.]. *Warta Tumbuhan Obat Indonesia* 2(2): 23–25.
558. Hase, K. et al., 1996. Hepatoprotective effects of traditional medicines. Isolation of the active constituent from seeds of *Celosia argentea*. *Phytotherapy Research* 10(5): 387–392.
559. Hassig, C.A., Tong, J.K. & Schreiber, S.L., 1997. Fiber-derived butyrate and the prevention of colon cancer. *Chemistry and Biology* 4(11): 783–789.
560. Hausen, B.M., 1993. *Centella asiatica* (Indian pennywort), an effective therapeutic but a weak sensitizer. *Contact Dermatitis* 29(4): 175–179.
561. Hawkes, J.G., Lester, R.N. & Skelding, A.D. (Editors), 1979. The biology and taxonomy of the Solanaceae. Published for the Linnean Society of London by Academic Press, Dorset, United Kingdom. 738 pp.
562. Hawkes, J.G., Lester, R.N., Nee, M. & Estrada, N. (Editors), 1991. The Solanaceae III: Taxonomy, chemistry, evolution. The Royal Botanic Gardens, Kew & The Linnean Society of London, London, United Kingdom. 483 pp.
563. Hayashi, K., Takehisa, T., Hamato, N., Takano, R., Hara, S., Miyata, T. & Kato, H., 1994. Inhibition of serine proteases of the blood coagulation system by squash family protease inhibitors. *Journal of Biochemistry* 116(5): 1013–1018.
564. Hayashi, T., Koyama, J., McPhail, A.T. & Lee, K.-H., 1987. Structure and absolute stereochemistry of tomenphantopin-A and -B, two cytotoxic sesquiterpene lactones from *Elephantopus tomentosus*. *Phytochemistry* 26(4): 1065–1068.
565. Hazarika, R.A., Deka, D.K., Phukan, S.C. & Saikia, P.K., 1995. Sarcoptic mange in buffalo calves and treatment with Pestoban. *Journal of Veterinary Parasitology* 9(2): 143–145.
566. Heal, R.E., Rogers, E.F, Wallace, R.T. & Starnes, O., 1950. A survey of plants for insecticidal activity. *Lloydia* 13: 89–62.
567. Hebbalkar, D.S., Hebbalkar, G.D., Sharma, R.N., Joshi, V.S. & Bhat, V.S., 1992. Mosquito repellent activity of oils from *Vitex negundo* Linn. leaves. *Indian Journal of Medical Research* (95): 200–203.
568. Hedge, R., Maiti, T.K. & Podder, S.K., 1991. Purification and characterization of three toxins and two agglutinins from *Abrus precatorius* seed by using lactamyl-sepharose affinity chromatography. *Analytical Biochemistry* 194: 101–109.

569. Hegarty, M.P., Lee, C.P., Christie, G.S., Court, R.D. & Haydock, K.P., 1979. The goitrogen 3-hydroxy-4(1H)-pyridone, a ruminal metabolite from *Leucaena leucocephala*: effects in mice and rats. *Australian Journal of Biological Sciences* 32: 27–40.
570. Heggers, J.P., Pelley, R.P. & Robson, M.C., 1993. Beneficial effect of Aloe in wound healing. *Phytotherapy Research* 7: S48–55.
571. Hegi, G., 1987. *Illustrierte flora von Mittel-Europa* [Illustrated flora of Central Europe]. 2nd Edition improved by G. Wagenitz. Vol. VI, Part 4, Compositae II: *Matricaria* – *Hieracium*. Verlag Paul Parey, Berlin & Hamburg, Germany. 1483 pp.
572. Hegnauer, R., 1962–1997. *Chemotaxonomie der Pflanzen* [Chemotaxonomy of plants]. 11 volumes. Birkhäuser Verlag, Basel, Switzerland.
573. Heller, J., 1996. Physic nut. *Jatropha curcas* L. Promoting the conservation and use of underutilized and neglected crops 1. Institute of Plant Genetics and Crop Plant Research, Gatersleben, Germany & International Plant Genetic Resources Institute, Rome, Italy. 66 pp.
574. Hema, D., Tidjani, M., Bassene, E., Pousset, J.L. & Giono-Barber, H., 1986. Plantes médicinales africaines XXIV. Etude de l'action anti-inflammatoire de *Bryophyllum pinnatum* (Crassulacées) [African medicinal plants XXIV. Study of the anti-inflammatory action of *Bryophyllum pinnatum* (Crassulaceae)]. *Plantes Médicinales et Phytothérapie* 20(3): 231–235.
575. Hemalatha, A., Nageswar, G. & Radhakrishnaiah, M., 1992. Phenolic acid profile of *Phyllanthus* s.l. (Euphorbiaceae). *Feddes Repertorium* 103(1–2): 83–85.
576. Henderson, C.P. & Hancock, I.R., 1988. *A guide to the useful plants of Solomon Islands*. Ministry of Agriculture and Lands, Honaira, Solomon Islands. 481 pp.
577. Henty, E.E., 1969. *A manual to the grasses of New Guinea*. Botany Bulletin No. 1. Division of Botany, Department of Forests, Lae, Territory of Papua and New Guinea. 215 pp.
578. Herbert, J.M., Maffrand, J.P., Taoubi, K., Augereau, J.M., Fouraste, I. & Gleye, J., 1991. Verbascoside isolated from *Lantana camara*, an inhibitor of protein kinase C. *Journal of Natural Products* 54(6): 1595–1600.
579. Hewson, H.J., 1985. Simaroubaceae. In: George, A.S. (Editor): *Flora of Australia*. Vol. 25. Melianthaceae to Simaroubaceae. Australian Government Publishing Service, Canberra, Australia. pp. 188–197.
580. Heyne, K., 1950. *De nuttige planten van Indonesië* [The useful plants of Indonesia]. 3rd Edition. 2 volumes. W. van Hoeve, 's-Gravenhage, the Netherlands/Bandung, Indonesia. 1660 + CCXLI pp.
581. Hiermann, A. & Bucar, F., 1994. Influence of some traditional medicinal plants of Senegal on prostaglandin biosynthesis. *Journal of Ethnopharmacology* 42(2): 111–116.
582. Higashino, H., Suzuki, A., Tanaka, Y. & Pootakham, K., 1992. Hypoglycemic effects of Siamese *Momordica charantia* and *Phyllanthus urinaria* extracts in streptozotocin-induced diabetic rats (the 1st report). *Nippon Yakurigaku Zasshi* 100(5): 415–421. (in Japanese)
583. Hikino, H., 1985. Antihepatotoxic activity of crude drugs. *Yakugaku Zasshi* 105(2): 109–118. (in Japanese)

584. Hikino, H., Mizuno, T., Oshima, Y. & Konno, C., 1985. Isolation and hypoglycemic activity of moran A, a glycoprotein of *Morus alba* root barks. *Planta Medica* 50(2): 159–160.
585. Hikino, H., Teruraki, S. & Kawanishi, H., 1986. Method of treating hypoglycemia using aloes polysaccharides. U.S. Patent 4, 598, 069.
586. Hikino, H., Yoshiawa, M., Suuji, Y., Oshima, Y. & Konno, C., 1989. Isolation and hypoglycemic activity of trichosans A, B, C, D, and E: glycans of *Trichosanthes kirilowii* roots. *Planta Medica* 55(4): 349–350.
587. Hikino, Y., Takashi, M., Murakami, M., Konno, C., Mirin, Y., Karikura, M., & Hayashi, T., 1986. Isolation and hypoglycemic activity of arboran A and B, glycans of *Aloe arborescens* var. *natalensis* leaves. *International Journal of Crude Drug Research* 24(4): 183–186.
588. Hilton, M.G. & Rhodes, M.J.C., 1993. Factors affecting the growth and hyoscyamine production during batch culture of transformed roots of *Datura stramonium*. *Planta Medica* 59(4): 340–344.
589. Hipkin, C.R., 1991. Phytophotodermatitis, a botanical view. *Lancet* 338(8771): 892.
590. Hiraoka, N. & Kodama, T., 1984. Effects of non-frozen cold storage on the growth, organogenesis and secondary metabolism of callus cultures. *Plant Cell, Tissue and Organ Culture* 3(4): 349–357.
591. Hirayama, H., Wang, Z., Nishi, K., Ogawa, A., Ishimatu, T., Ueda, S., Kubo, T. & Nohara, T., 1993. Effect of *Desmodium styracifolium* triterpenoid on calcium oxalate renal stones. *British Journal of Urology* 71(2): 143–147.
592. Hisham, A., Pieters, L., Claeys, M., Dommissie, R., Vanden Berghe, D. & Vlietinck, A., 1992. Guaianolide glucosides from *Elephantopus scaber*. *Planta Medica* 58(5): 474–475.
593. Ho, C.S., Wong, Y.H. & Chiu, K.W., 1989. The hypotensive action of *Desmodium styracifolium* and *Clematis chinensis*. *American Journal of Chinese Medicine* 17(3–4): 189–202.
594. Ho, L.M., Cheong, I., & Jalil, H.A., 1996. Rhabdomyolysis and acute renal failure following blowpipe dart poisoning. *Nephron* 72(4): 676–678.
595. Hodge, W.H., 1953. The drug aloes of commerce, with special reference to the Cape species. *Economic Botany* 7: 99–129.
596. Hoffman, M.T., 1988. The pollination ecology of *Aloe ferox* Mill. *South African Journal of Botany* 54(4): 345–350.
597. Holdsworth, D.K., 1977. Medicinal plants of Papua New Guinea. Technical Paper No 175. South Pacific Commission, Noumea, New Caledonia. 123 pp.
598. Holdsworth, D.K., 1987. Medicinal plants of the Central Province of Papua New Guinea. Part. IV. The Goilala Mountain People. *International Journal of Pharmacognosy* 25: 231–235.
599. Holdsworth, D.K., 1987. Medicinal plants of the Morobe Province, Papua New Guinea. Part V. The Upper Watut. *International Journal of Pharmacognosy* 25: 225–230.
600. Holdsworth, D.K., 1989. High altitude medicinal plants of Papua New Guinea. *International Journal of Crude Drug Research* 27(2): 95–100.
601. Holdsworth, D.K., 1991. Medicinal plants of the Central Province of Papua New Guinea. Part V. Coastal villages to the West and East of Port

- Moresby. *International Journal of Pharmacognosy* 29(3): 231–236.
602. Holdsworth, D.K., 1991. Traditional medicinal plants of Brunei Darussalam. Part I. Bukit Udal. *International Journal of Pharmacognosy* 29(4): 245–250.
603. Holdsworth, D.K., 1992. Medicinal plants of the Gazelle Peninsula, New Britain Island, Papua New Guinea. Part I. *International Journal of Pharmacognosy* 30: 185–190.
604. Holdsworth, D.K., 1993. Medicinal plants of the Oro (Northern) Province of Papua New Guinea. *International Journal of Pharmacognosy* 31: 23–28.
605. Holdsworth, D.K. & Balun, L., 1992. Medicinal plants of the East and West Sepik Provinces, Papua New Guinea. *International Journal of Pharmacognosy* 30: 218–222.
606. Holdsworth, D.K., Gideon, O. & Pilokos, B., 1989. Traditional medicine of New Ireland, Papua New Guinea. Part III. Konos, Central New Ireland. *International Journal of Crude Drug Research* 27: 55–61.
607. Holdsworth, D.K. & Kerenga, K., 1987. Medicinal plants of the Western Highlands, Papua New Guinea. *International Journal of Crude Drug Research* 25: 171–176.
608. Holdsworth, D.K. & Lacanienta, E., 1981. Traditional medicinal plants of the Central Province of Papua New Guinea. Part II. *Quarterly Journal of Crude Drug Research* 19(4): 155–167.
609. Holdsworth, D.K. & Mahana, P., 1983. Traditional medicinal plants of the Huon Peninsula, Morobe Province, Papua New Guinea. *International Journal of Crude Drug Research* 21: 121–133.
610. Holdsworth, D.K. & Rali, T., 1989. A survey of medicinal plants of the Southern Highlands, Papua New Guinea. *International Journal of Crude Drug Research* 27: 1–8.
611. Holdsworth, D.K. & Sakulas, H., 1986. Medicinal plants of the Morobe Province. Part II. The Aseki Valley. *International Journal of Crude Drug Research* 24: 31–40.
612. Holdsworth, D.K. & Sakulas, H., 1992. High altitude medicinal plants of Papua New Guinea. Part II. Mount Wilhelm, Simbu Province. *International Journal of Pharmacognosy* 30(1): 1–4.
613. Holm, J., Doll, J. & Holm, E., 1997. *World weeds: natural histories and distribution*. Wiley, New York, United States. 1129 pp.
614. Holm, L.G., Plucknett, D.L., Pancho, J.V. & Herberger, J.P., 1977. *The world's worst weeds. Distribution and biology*. East-West Center, the University Press of Hawaii, Honolulu, United States. 609 pp.
615. Holttum, R.E., 1950. The Zingiberaceae of the Malay Peninsula. *Gardens' Bulletin, Singapore* 13(1): 1–249.
616. Horie, T., Tominaga, H. & Kawamura, Y., 1993. Revised structure of a natural flavone from *Ageratum conyzoides*. *Phytochemistry* 32(4): 1076–1077.
617. Horsten, S.F.A.J., 1995. *Cyclic peptides in the genus Jatropha (Euphorbiaceae)*. Thesis, Utrecht University, the Netherlands. 239 pp.
618. Horsten, S.F.A.J., van den Berg, A.J.J., Kettenes-van den Bosch, Leeflang, B.R. & Labadie, R.P., 1996. Cyclogossine A: A novel cyclic heptapeptide isolated from the latex of *Jatropha gossypifolia*. *Planta Medica* 62: 46–50.

619. Houck, D.F. & Rieseberg, L.H., 1983. Hormonal regulation of epiphyllous bud release and development in *Bryophyllum calycinum*. *American Journal of Botany* 70(6): 912-915.
620. Houghton, P.J., Woldemariam, T.Z., o'Shea, S. & Thyagarajan, S.P., 1996. Two securinoga-type alkaloids from *Phyllanthus amarus*. *Phytochemistry* 43(3): 715-717.
621. Howlett, A.C., 1987. Cannabinoid inhibition of adenylate cyclase. *Neuropharmacology* 26: 507-512.
622. Hsieh, C.-F., 1978. Hypoxidaceae. In: Li, H.-L., Liu, T.-S., Huang, T.-C., Koyama, T. & DeVol, C.E. (Editors): *Flora of Taiwan*. Vol. 5. Epoch Publishing Co., Taipei, Taiwan, Republic of China. pp. 95-98.
623. Hsieh, C.-F. & Huang, T.-C., 1978. Acanthaceae. Li, H.-L., Liu, T.-S., Huang, T.-C., Koyama, T. & DeVol, C.E. (Editors): *Flora of Taiwan*. Vol. 4. Epoch Publishing Co., Taipei, Taiwan, Republic of China. pp. 620-663.
624. Hsieh, T.-H. & Huang, T.-C., 1995. Notes on the flora of Taiwan (20) - *Scutellaria* (Lamiaceae) in Taiwan. *Taiwania* 40: 35-56.
625. Hsu, H.-J., Chen, Y.-P. & Hong, M., 1982. The chemical constituents of oriental herbs. Vol. 1. Oriental Healing Arts Institute, Scotts Valley, California, United States. 619 pp.
626. Huang, B.L., 1982. Comparison of four methods of midterm termination of pregnancy and their maternal influences. *Tienchin I Yao* 10(5): 284-287.
627. Huang, J.-T., 1981. Neue Iridoide aus *Oldenlandia diffusa* Roxb. [New iridoids from *Oldenlandia diffusa* Roxb.]. *Archiv der Pharmazie* 314(10): 831-836.
628. Huang, T.-C. & Cheng, W.-T., 1978. Labiatae. In: Li, H.-L., Liu, T.-S., Huang, T.-C., Koyama, T. & DeVol, C.E. (Editors): *Flora of Taiwan*. Vol. 4. Epoch Publishing Co., Taipei, Taiwan, Republic of China. pp. 439-529.
629. Huang, T.-C. & Ohashi, H., 1993. Leguminosae. In: Huang, T.-C. (Editor): *Flora of Taiwan*. 2nd Edition. Vol. 3. Editorial Committee of the Flora of Taiwan, Taipei, Taiwan, Republic of China. pp 160-395.
630. Hubbard, C.E., 1944. Taxonomy, description and distribution of species and varieties. In: *Imperata cylindrica*. Taxonomy, distribution, economic significance and control. Imperial Agricultural Bureaux Joint Publication No. 7. Imperial Forestry Bureau, Oxford & Imperial Bureau of Pastures and Forage Crops, Aberystwyth, United Kingdom. pp. 5-13.
631. Hui, W.H. & Sung, M.L., 1968. An examination of the Euphorbiaceae of Hong Kong II. The occurrence of epitaraxerol and other triterpenoids. *Australian Journal of Chemistry* 21: 2137-2140.
632. Hui, W.H., Li, M.M. & Wong, K.M., 1976. A new compound, 21 $\alpha$ -hydroxyfriedel-4(23)-en-3-one and other triterpenoids from *Phyllanthus reticulatus*. *Phytochemistry* 15: 797-798.
633. Hung, C.H., Lee, M.C., Lee, T.C. & Lin, J.Y., 1993. Primary structure of three distinct isoabrinins determined by cDNA sequencing: conservation and significance. *Journal of Molecular Biology* 229(1): 263-267.
634. Hung, D.T., Jamison, T.F. & Schreiber, S.L., 1996. Understanding and controlling the cell cycle with natural products. *Chemistry and Biology* 3(8): 623-639.
635. Husain, A. & Sharma, J.R., 1983. The opium poppy. Medicinal and aromatic plants series no.1. Central Institute of Medicinal and Aromatic



- Plants, Lucknow, India. 167 pp.
636. Hussain, R.A., Dickey, J.K., Rosser, M.P., Matson, J.A., Kozlowski, M.R., Brittain, R.J., Webb, M.L., Rose, P.M. & Fernandes, P., 1995. A novel class of non-peptidic endothelin antagonists isolated from the medicinal herb *Phyllanthus niruri*. *Journal of Natural Products* 58(10): 1515–1520.
  637. Hussain, S.S. & Siddiqui, M.O., 1995. Specific activity and isozyme pattern of ATPase of *Blumea* species. *Acta Botanica Indica* 23(2): 215–217.
  638. Huxley, A., Griffiths, M. & Levy, M., 1992. The new Royal Horticultural Society dictionary of gardening. 4 volumes. The MacMillan Press Ltd., London, United Kingdom. 3353 pp.
  639. Hwang, W.I., Cha, S. & Lee, S.Y., 1980. Extraction of anti cancer components from Korean medicinal plants and the determination of their cytotoxic activities on the cancer cells. *Korean Biochemical Journal* 13(1): 25–40.
  640. Ibnu Utomo W., B., 1998. *Cassia L.* In: Sosef, M.S.M., Hong, L.T. & Prawirohatmodjo, S. (Editors): *Plant Resources of South-East Asia* 5(3). Timber trees: Lesser-known timbers. Backhuys Publishers, Leiden, the Netherlands. pp. 144–146.
  641. IBPGR, 1986. Genetic resources of tropical and subtropical fruits and nuts (excluding *Musa*). International Board of Plant Genetic Resources, Rome, Italy. 162 pp.
  642. Ibuka, T. & Kitano, M., 1967. Studies on the alkaloids of menispermaceous plants. Alkaloids of *Stephania japonica* Miers. 18. Structure of hasubanone (Suppl. 1). *Chemical and Pharmaceutical Bulletin* 15(11): 1809–1810.
  643. Ibuka, T. & Kitano, M., 1967. Studies on the alkaloids of menispermaceous plants. CCXXXVII. Alkaloids of *Stephania japonica* Miers. (Suppl. 17). Structure of homostephanoline. (3). *Chemical and Pharmaceutical Bulletin* 15(12): 1939–1943.
  644. Ihanola-Vormisto, A., Summanen, J., Kankaanranta, H., Vuorela, H., Asmawi, Z.M. & Moilanen, E., 1997. Anti-inflammatory activity of extracts from leaves of *Phyllanthus emblica*. *Planta Medica* 63(6): 518–524.
  645. Ikenaga, T., Hizako, M., Tajima, M. & Nakashima, K., 1994. Production of choleric substances in the capitulum, leaf and stem of *Artemisia capillaris* during the plant growth cycle. *Biological and Pharmaceutical Bulletin* 17(1): 150–151.
  646. Ikram, M. & Inamul, H., 1980. Screening of medicinal plants for antimicrobial activity. Part II. *Fitoterapia* 51(6): 281–284.
  647. Imai, S. et al., 1994. African Burkitt's lymphoma: A plant, *Euphorbia tirucalli*, reduces Epstein-Barr virus-specific cellular immunity. *Anticancer Research* 14(3A): 933–936.
  648. Imamura, K., Fukamiya, N., Nakamura, M., Okano, M., Tagahara, K. & Lee, K.H., 1995. Bruceanols G and H cytotoxic quassinoids from *Brucea antidysenterica*. *Journal of Natural Products* 58(12): 1915–1919.
  649. Imamura, K., Fukamiya, N., Okano, M., Tagahara, K. & Lee, K.H., 1993. Bruceanols D, E, and F three new cytotoxic quassinoids from *Brucea antidysenterica*. *Journal of Natural Products* 56(12): 2091–2097.
  650. Inada, A., Nakanishi, T., Tokuda, H., Nishino, H., Iwashima, A. & Sharma, O.P., 1995. Inhibitory effects of lantadenes and related triter-

- penoids on Epstein-Barr virus activation. *Planta Medica* 61(6): 558–559.
651. Ines, M.C., 1990. Effects of different phyto-repellents against insect pests of stored garlic. In: (Report on the) Annual Convention of the Pest Control Council of the Philippines, Bacolod City, the Philippines, 7–10 May, 1990.
652. International Trade Centre UNCTAD/GATT, 1982. Markets for selected medicinal plants and their derivatives. Geneva, Switzerland. 206 pp.
653. Irwin, H.S. & Barneby, R.C., 1982. The American Cassiinae. A synoptical revision of Leguminosae tribe Cassieae subtribe Cassiinae in the New World. *Memoirs of the New York Botanical Garden* 35(2): 64–635.
654. Isawumi, M.A., 1995. Notes on *Vernonia* (Vernonieae: Compositae) in West Africa. In: Hind, D.J.N., Jeffrey, C. & Pope, G.V. (Editors): *Advances in Compositae systematics*. Royal Botanic Gardens, Kew, United Kingdom. pp. 51–106.
655. Ishiguro, K., Nagata, S., Fukumoto, H., Yamaki, M. & Isoi, K., 1994. Phloroglucinol derivatives from *Hypericum japonicum*. *Phytochemistry* 35(2): 469–471.
656. Ishiguro, K., Yamaki, M., Kashihara, M. & Takagi, S., 1986. Sarothralen A and B, new antibiotic compounds from *Hypericum japonicum*. *Planta Medica* 52(4): 288–290.
657. Ishiguro, K., Yamaki, M., Kashihara, M. & Takagi, S., 1987. Saroaspidin A, B and C: additional antibiotic compounds from *Hypericum japonicum*. *Planta Medica* 53(5): 415–417.
658. Ishiguro, K., Yamaki, M., Kashihara, M., Takagi, S. & Isoi, K., 1990. Sarothralin G: a new antimicrobial compound from *Hypericum japonicum*. *Planta Medica* 56(3): 274–276.
659. Ishii, Y., Tanizawa, H. & Takino, Y., 1994. Studies of aloe. IV. Mechanism of cathartic effect (3). *Biological and Pharmaceutical Bulletin* 17(4): 495–497.
660. Itokawa, H., Hirayama, F., Funakoshi, K. & Takeya, K., 1992. Studies on the antitumor bisabolane sesquiterpenoids isolated from *Curcuma xanthorrhiza*. *Chemical and Pharmaceutical Bulletin* 33(8): 3488–3492.
661. Iwu, M.M., Jackson, J.E., Tally, J.D. & Klayman, D.L., 1992. Evaluation of plant extracts for antileishmanial activity using a mechanism-based radiorespirometric microtechnique (RAM). *Planta Medica* 58: 436–441.
662. Jacob, A., Pandey, M., Kapoor, S. & Saroja, R., 1988. Effect of the Indian gooseberry (amla) on serum cholesterol levels in men aged 35–55 years. *European Journal of Clinical Nutrition* 42: 939–944.
663. Jager, A.K., Hutchings, A. & Van Staden, J., 1996. Screening of Zulu medicinal plants for prostaglandin-synthesis inhibitors. *Journal of Ethnopharmacology* 52(2): 95–100.
664. Jain, P. & Sahu, T.R., 1993. An ethnobotanical study of Noradehi Sanctuary Park of Madhya Pradesh, India: native plant remedies for scorpion sting and snake bite. *Journal of Economic and Taxonomic Botany* 17(2): 315–328.
665. Jain, S.C. & Sahoo, S., 1981. Isolation and characterization of steroidal sapogenins and glyco alkaloids from tissue cultures of *Solanum verbascifolium*. *Chemical and Pharmaceutical Bulletin* 29(6): 1765–1767.
666. Jain, S.C., Sahoo, S.L. & Vijyvergia, R., 1995. Influence of light on growth and production of steroids and glycoalkaloids in *Solanum* species in vivo

- and in vitro. *Indian Journal of Pharmaceutical Sciences* 57(3): 100–101.
667. Jain, S.P. & Puri, H.S., 1984. Ethnomedicinal plants of Jaunsar Bawar hills, Uttar Pradesh, India. *Journal of Ethnopharmacology* 12(2): 213–222.
668. Jakupovic, J., Jia, Y., Zdero, C., Warning, U., Bohlmann, F. & Jones, S.B., 1987. Germacranolides from *Elephantopus* spp. *Phytochemistry* 26(5): 1467–1469.
669. Jamieson, G.I., 1984. Aloe vera (*Aloe barbadensis* Mill.). *Queensland Agricultural Journal* 110(4): 220.
670. Jansen, P.C.M., 1981. Spices, condiments and medicinal plants in Ethiopia, their taxonomy and agricultural significance. Pudoc, Wageningen, the Netherlands. 327 pp.
671. Jantarawatit, S., Reutrakul, V. & Ratanabanangkoon, K., 1997. Estrogenic activity found in the herb *Euphorbia hirta* (nam nom rat chasee). *Mahidol University Annual Abstracts* 192, Bangkok, Thailand.
672. Januwati, M., Sudiarto & Emmyzar, 1989. Kumis kucing, pule pandak, and touki [*Ortosiphon aristatus*, *Rauvolfia serpentina*, and *Angelica acutiloba*]. *Edisi Khusus Penelitian Tanaman Rempah dan Obat* 5(1): 24–31.
673. Japing, H.W. & Oey Djoen Seng, 1936. Cultuurproeven met wildhoutsoorten in Gadoengan – met overzicht van de literatuur betreffende deze soorten [Trial plantations of non-teak wood species in East Java – with survey of literature about these species]. *Korte Mededeelingen* No 55, part I–VI. Boschbouwproefstation, Buitenzorg, Nederlandsch-Indië. 270 pp.
674. Jaquat, C., 1990. Plants from the markets of Thailand. Editions Duang Kamol, Bangkok, Thailand.
675. Jayaswal, S.B., 1977. *Wrightia tomentosa*, a substitute for *Holarrhena antidysenterica*. *Indian Journal of Pharmacy* 39(2): 37–39.
676. Jayaweera, D.M.A., 1982. Medicinal plants (indigenous and exotic) used in Ceylon. Part IV. National Science Council of Sri Lanka, Colombo.
677. Jeffrey, C., 1980. A review of the Cucurbitaceae. *Botanical Journal of the Linnean Society* 81: 233–247.
678. Jenniskens, M.-J.P.J., 1984. Self-compatibility in diploid plants of *Taraxacum* section *Taraxacum*. *Acta Botanica Neerlandica* 33: 71–80.
679. Jeppe, B., 1974. South African aloes. 2nd Edition. Purnell, Cape Town, South Africa. 150 pp.
680. Jilka, C., Strifler, B., Fortner, G.W., Hays, E.F. & Takemoto, D.J., 1983. In vivo antitumor activity of the bitter melon (*Momordica charantia*). *Cancer Research* 43(11): 5151–5155.
681. Jitoe, A. et al., 1992. Antioxidant activity of tropical ginger extracts and analysis of the contained curcuminoids. *Journal of Agricultural and Food Chemistry* 40(8): 1337–1340.
682. Johnson, E.L., 1989. Seed viability of two *Erythroxylum* species stored at 4°C. *Planta Medica* 55(7): 691.
683. Johnson, E.L., 1996. Alkaloid content in *Erythroxylum coca* tissue during reproductive development. *Phytochemistry* 42(1): 35–38.
684. Johnson, E.L. & Fox, C.D., 1996. Biomass accumulation and alkaloid content in leaves of *Erythroxylum coca* and *Erythroxylum novogranatense* var. *novogranatense* grown in soil with varying pH. *Journal of Plant Physiology* 149(3–4): 444–450.

685. Johnson, M.F., 1971. A monograph of the genus *Ageratum* L. (Compositae-Eupatorieae). *Annals of the Missouri Botanical Garden* 58(1): 6-88.
686. Johnston, I.M., 1951. Studies in the Boraginaceae XX. Carmona. *Journal of the Arnold Arboretum* 32: 16-19.
687. Johri, R.K., Dhar, S.K., Pahwa, G.S., Sharma, S.C., Kaul, J.L. & Zutshi, U., 1990. Toxicity studies with potassium embelate, a new analgesic compound. *Indian Journal of Experimental Biology* 28: 213-217.
688. Jolin, D. & Torquebiau, E., 1992. Large cuttings: a jump start for tree planting. *Agroforestry Today* 4(4): 15-16.
689. Jolly, C.I. & Mechery, N.R., 1996. Comparative pharmacognostical, physicochemical and antibacterial studies on seeds of *Holarrhena antidysenterica* Wall and *Wrightia tinctoria* R. Br. *Indian Journal of Pharmaceutical Sciences* 58(2): 51-54.
690. Jones, N. & Miller, J.H., 1992. *Jatropha curcas*. A multipurpose species for problematic sites. Land Resources Series No 1. Agriculture Division, Asia Technical Department, The World Bank, Washington DC, United States. 12 pp. & 6 annexes.
691. Jones, R.J. & Megarrity, R.G., 1986. Successful transfer of DHP-degrading bacteria from Hawaiian goats to Australian ruminants to overcome the toxicity of *Leucaena*. *Australian Veterinary Journal* 63: 259-262.
692. Joshi, K.C., Meshram, P.B., Sambath, S., Kiran, U., Humane, S. & Kharkwal, G.N., 1992. Insect pests of some medicinal plants in Madhya Pradesh. *Indian Journal of Forestry* 15(1): 17-26.
693. Juneja, R.C., Nayyar, V.L. & Mukerji, K.G., 1976. Further additions to plant diseases of Delhi. *Angewandte Botanik* 50(1-2): 43-47.
694. Kajima, M., Tanaka, T. & Yonemasu, Y., 1987. Epileptogenic properties of quisqualic acid: microinjection into the unilateral amygdala in cats. *No To Shinkei* 39(10): 971-976. (in Japanese)
695. Kaleysa-Raj, R., 1975. Screening of indigenous plants for anthelmintic action against human *Ascaris lumbricoides*. Part 1. *Indian Journal of Physiological Pharmacology* 19: 47-49.
696. Kalpana Dixit, Shukla, H.S. & Dubey, P., 1986. Fungitoxic properties of some seedling extracts. *National Academy of Science Letters* 9(8): 219-221.
697. Kalshoven, L.G.E., 1950-1951. De plagen van cultuurgewassen in Indonesië [The pests of crops in Indonesia]. 2 volumes. van Hoeve, 's-Gravenhage, the Netherlands.
698. Kam, T.S., Sim, K.M., Koyano, T., Toyoshima, M., Hayashi, M. & Komiya, K., 1998. Cytotoxic and leishmanial aminoglycosteroids and aminosteroids from *Holarrhena curtisii*. *Journal of Natural Products* 61(11): 1332-1336.
699. Kamchonwongpaisan, S., Nilanonta, C., Tarnchompoo, B., Thebtaranonth, C., Thebtaranonth, Y., Yuthavong, Y., Kongsaree, P. & Clardy, J., 1995. An antimalarial peroxide from *Amomum krervanh* Pierre. *Tetrahedron Letters* 36(11): 1821-1824.
700. Kamilya, P. & Paria, N., 1994. Seedling morphology of some Indian species of *Jatropha* and its implications in taxonomy. *Acta Botanica Indica* 22: 251-256.

701. Kamo, K.K. & Mahlberg, P.G., 1988. Morphinan alkaloids: biosynthesis in plant (*Papaver* spp.) tissue cultures. In: Bajaj, Y.P.S. (Editor): *Biotechnology in agriculture and forestry 4. Medicinal and aromatic plants I*. Springer Verlag, Berlin, Germany. pp. 251–263.
702. Kamphukun, M., Stienswat, W. & Aiumtanaporn, R., 1984. Kan sukra raya pluk lae chuang wela kan kepkieo phayaraibai [Investigation on plant density and harvesting time of *Euphorbia tirucalli*]. Department of Horticulture, Faculty of Agriculture, Kasetsart University, Bangkok, Thailand. 19 pp.
703. Kang, S.S. & Woo, W.S., 1991. Phytolaccoside I, a new saponin from *Phytolacca americana*. *Fitoterapia* 62(6): 532–533.
704. Kanis, A., 1972. A review of the Amaranthaceae in Papuasia. *Contributions from Herbarium Australiense* No 1: 1–8.
705. Kao, M.-T., 1996. Umbelliferae. In: Huang, T.-C. (Editor): *Flora of Taiwan*. 2nd Edition. Vol. 3. Editorial Committee of the Flora of Taiwan, Taipei, Taiwan, Republic of China. pp. 1010–1045.
706. Karnick, C.R., 1978. Responses of lunar-phases on the growth of *Abrus precatorius* Linn. (Leguminosae) and its effects as crude drug on diseases. *Acta Horticulturae* 73: 239–247.
707. Karunanayake, E.H., Welihinda, J., Sirimanne, S.R. & Sinnadorai, G., 1984. Oral hypoglycaemic activity of some medicinal plants of Sri Lanka. *Journal of Ethnopharmacology* 11(2): 223–231.
708. Kasahara, S. & Hemmi, S. (Editors), 1995. *Medicinal herb index in Indonesia*. 2nd Edition. P.T. Eisai Indonesia, Jakarta, Indonesia. 453 pp.
709. Kashyap, N.P., Bhagat, R.M., Sharma, D.C. & Suri, S.M., 1992. Efficacy of some useful plant leaves for the control of potato tuber moth, *Phthorimaea operculella* Zell. in stores. *Journal of Entomological Research* 16(3): 223–227.
710. Kass, E. & Wink, M., 1997. Phylogenetic relationships in the Papilionoideae (family Leguminosae) based on nucleotide sequences of cpDNA (*rbcL*) and ncDNA (ITS 1 and 2). *Molecular Phylogenetics and Evolution* 8(1): 65–88.
711. Kato, S., Awaya, K., Nakatsukasa, Y. & Okuda, Y., 1981. Blastogenic property of lectin from jequirity beans (*Abrus precatorius*). *Bulletin of the Yamaguchi Medical School* 28(1–2): 27–34.
712. Kaufman, S.J. & McPherson, A., 1975. Abrin and hurin: two new lymphocyte mitogens. *Cell* 4(3): 263–268.
713. Kaul, M.L.H. & Neelangini, 1989. Male sterility in diploid *Ageratum conyzoides* L. *Cytologia* 54(3): 445–448.
714. Kaushik, P. & Khanna, P., 1992. Insecticidal substances from in vivo and in vitro tissue culture of *Abrus precatorius* L. *Advances in Plant Sciences* 5(2): 464–469.
715. Kavimani, S., Ilango, R., Madheswaran, M., Jayakar, B., Gupta, M. & Majumdar, U.K., 1996. Antitumour activity of plumbagin against Dalton's ascitic lymphoma. *Indian Journal of Pharmaceutical Sciences* 58(5): 194–196.
716. Kazmi, S.M., Trivedi, V.B. & Kazmi, S.N., 1981. Certain weeds of central India and their antimicrobial properties. In: *Proceedings of the Eighth Asian-Pacific Weed Science Society Conference*. Asian-Pacific Weed

- Science Society, Bangalore, India. pp. 187–189.
717. Keereewan, S. & Leeprasert, P., 1975. Distribution of plant parasitic nematodes of mulberry in Thailand. Technical Bulletin No 28. Plant Protection Service, Department of Agriculture, Bangkok, Thailand. 18 pp.
718. Kelecom, A., 1983. Isolation, structure determination, and absolute configuration of barbatusol, a new bioactive diterpene with a rearranged abietane skeleton from the labiate *Coleus barbatus*. *Tetrahedron* 39(21): 3603–3608.
719. Kelecom, A., 1984. An abietane diterpene from the labiate *Coleus barbatus*. *Phytochemistry* 23(8): 1677–1679.
720. Keng, H., 1978. Labiatae. In: van Steenis, C.G.G.J. (General editor): *Flora Malesiana*. Series 1, Vol. 8. Sijthoff & Noordhoff International Publishers, Alphen aan den Rijn, the Netherlands. pp. 301–394.
721. Keraudren-Aymonin, M., 1975. Cucurbitacées [Cucurbitaceae]. In: Vidal, J.E. & Galibert, Y. (Editors): *Flore du Cambodge, du Laos et du Vietnam* [Flora of Cambodia, Laos and Vietnam]. Vol. 15. Muséum National d'Histoire Naturelle, Paris, France. 123 pp.
722. Kern, J.H., 1974. Cyperaceae. In: van Steenis, C.G.G.J. (General editor): *Flora Malesiana*. Series 1, Vol. 7. Noordhoff, Leyden, the Netherlands. pp. 435–753.
723. Khan, I.A., Rali, T. & Sticher, O., 1993. Alkaloids from *Ficus pachyrhachis*. *Planta Medica* 59(3): 286.
724. Khan, M.M.A.A., Jain, D.C., Bhakuni, R.S., Mohd Zaim & Thakur, R.S., 1991. Occurrence of some antiviral sterols in *Artemisia annua*. *Plant Science (Limerick)* 75(2): 161–165.
725. Khan, P.S.H., 1987. Comparative seed structure of medicinally important *Holarrhena antidysenterica* (Roth.) A.DC. and its adulterant, *Wrightia tinctoria* R.Br (Apocynaceae). *International Journal of Crude Drug Research* 25(2): 81–86.
726. Khanna, K. & Arora, J.S., 1984. Note on propagation of *Cassia fistula* (amaltas) by cutting and layering. *Indian Journal of Horticulture* 41(1–2): 137–138.
727. Khanna, P., 1982. Tissue cultures and useful metabolites: a review of thirtyeight plant species. In: Atal, C.L. & Kapur, B.M. (Editors): *Cultivation and utilization of medicinal plants*. Regional Research Laboratory, Council of Scientific and Industrial Research, Jammu-Tawi, India. pp. 824–851.
728. Kiang, P.C., 1923. Chinese drugs of therapeutic value to western physicians. *China Medical Journal (Shanghai)* 37: 742–746.
729. Kiesewetter, H., Jung, F., Pindur, G., Jung, E.M., Mrowietz, C. & Wenzel, E., 1991. Effect of garlic on thrombocyte aggregation, microcirculation, and other risk factors. *International Journal of Clinical Pharmacology, Therapy and Toxicology* 29: 151.
730. Kiew, K.Y., 1980. Taxonomic studies in the genus *Kaempferia* (Zingiberaceae). Notes from the Royal Botanic Garden Edinburgh 38: 1–12.
731. Kiew, R., 1994. Checklist of *Jasminum* (Oleaceae) in Malesia. *Sandakania* 5: 1–14.
732. Kiew, R., 1994. Name changes for Malaysian plants. *Begonia wrayi* (Begoniaceae) and *Jasminum aemulum* (Oleaceae). *Malayan Nature*

- Journal 47(3): 311-317.
733. Kightlinger, L.K., Seed, J.R. & Kightlinger, M.B., 1996. *Ascaris lumbricoides* aggregation in relation to child growth status, delayed cutaneous hypersensitivity, and plant anthelmintic use in Madagascar. *Journal of Parasitology* 82(1): 25-33.
734. Kim Byeung Gie, 1995. Anticoccidial effect of herb extracts against *Eimeria tenella*. *Korean Journal of Veterinary Clinical Medicine* 12(1): 123-128.
735. Kim, S.W., Son, K.H. & Chung, K.C., 1989. Mutagenic effect of steroidal saponins from *Smilax china* rhizomes. *Journal of the Pharmaceutical Society of Korea* 33(5): 285-289 (in Korean).
736. Kimura, M., Sumizawa, T. & Funatsu, G., 1993. The complete amino acid sequences of the B-chains of Abrin-a and Abrin-b, toxic proteins from the seeds of *Abrus precatorius*. *Bioscience, Biotechnology and Biochemistry* 57: 166-169.
737. Kimura, Y., Okuda, H., Okuda, T., Hatano, T., Agata, I. & Arichi, S., 1985. Studies on the activities of tannins and related compounds from medicinal plants and drugs. VII. Effects of extracts of leaves of *Artemisia* species, and caffeic acid and chlorogenic acid on lipid metabolic injury in rats fed peroxidized oil. *Chemical and Pharmaceutical Bulletin* 33(5): 2028-2034.
738. Kinamore, P.A., Jaeger, R.W. & de Castro, F.J., 1980. *Abrus* and *Ricinus* ingestion: management of three cases. *Clinical Toxicology* 17(3): 401-405.
739. Kirchoff, B.K., 1991. Homeosis in the flowers of the Zingiberaceae. *American Journal of Botany* 78(6): 833-837.
740. Kirschner, J. & Stepanek, J., 1987. Again on the sections in *Taraxacum* (Cichoriaceae) (Studies in *Taraxacum* 6). *Taxon* 36: 608-617.
741. Kirtikar, K.R. & Basu, B.D., 1935. Indian medicinal plants. 2nd Edition, edited by Blatter, E., Caius, J.F. & Mhaskar, K.S. L.H. Basu, Allahabad, India. 4 volumes xxxvii + 2793 pp.
742. Kishore, N., Dubey, N.K. & Mishra, A.K., 1993. Efficacy of some essential oils against fungi causing deterioration of *Triticum aestivum* during storage. *Indian Journal of Microbiology* 33(4): 277-280.
743. Kiso, Y., Ogasawara, S., Hirota, K., Watanabe, N., Oshima, Y., Konno, C. & Hikino, H., 1984. Antihepatotoxic principles of *Artemisia capillaris* buds. *Planta Medica* 50(1): 81-85.
744. Kiso, Y., Sasaki, K., Oshima, Y. & Hikino, H., 1982. Structure of arcapillin, an antihepatotoxic principle of *Artemisia cappillaris* herbs. *Heterocycles* 19: 1615-1617.
745. Kiso, Y., Suzuki, Y., Konno, C., Hikino, H., Hashimoto, I. & Yagi, Y., 1982. Application of carbon tetrachloride induced liver lesion in mice for the screening of liver protective crude drugs. *Shoyakugaku Zasshi* 36(3): 238-244. (in Japanese)
746. Kiyohara, H., Cyong, J.C. & Yamada, H., 1989. Relationship between structure and activity of an anti-complementary arabinogalactan from the roots of *Angelica acutiloba* Kitagawa. *Carbohydrate Research* 31(193): 193-200.
747. Kiyohara, H., Yamada, H., Cyong, J.C. & Otsuka, Y., 1986. Studies on polysaccharides from *Angelica acutiloba*. V. Molecular aggregation and anti-complementary activity of arabinogalactan from *Angelica acutiloba*.

- Journal of Pharmacobiodynamics 9(4): 339–346.
748. Klayman, D.L. et al., 1984. Isolation of artemisinin (qinghaosu) from *Artemisia annua* growing in the United States. *Journal of Natural Products* 47: 715–717.
749. Kloos, H., Thongo, F.W., Ouma, J.H. & Bulterworth, A.E., 1987. Preliminary evaluation of some wild and cultivated plants for snail control in Machakos district, Kenya. *Journal of Tropical Medical Hygiene* 90(4): 197–204.
750. Kloppenburg-Versteegh, J., 1934. Wenken en raadgevingen betreffende het gebruik van Indische planten, vruchten, enz. [Hints and recommendations on the use of East-Indies plants, fruits etc.] G.T.C. van Dorp, Semarang, Surabaya, Bandung, Dutch East Indies. (5th Edition. 1978. Servire, Katwijk aan Zee, the Netherlands). 364 pp.
751. Klughardt, G. & Zymalkowski, F., 1982. Magnoflorin und Protoquercit als Inhaltsstoffen von *Cyclea barbata* Miers [Magnoflorine and protoquercitol as constituents of *Cyclea barbata* Miers]. *Archiv der Pharmazie (Weinheim)* 315(1): 7–11.
752. Knaap-van Meeuwen, M.S., 1962. Preliminary revisions of some genera of Malaysian Papilionaceae V. A census of the genus *Desmodium*. *Reinwardtia* 6(3): 239–276.
753. Ko, H.H., Yu, S.M., Fo, F.N., Teng, C.M. & Lin, C.N., 1997. Bioactive constituents of *Morus australis* and *Broussonetia papyrifera*. *Journal of Natural Products* 60(10): 1008–1011.
754. Ko, K., 1932. The pharmacological action of plumbagin. *Japanese Journal of Medical Science IV, Pharmacology* 6: 259–286.
755. Kobayashi, A., Hagihara, K., Kajiyama, S., Kanzaki, H. & Kawazu, K., 1995. Antifungal compounds induced in the dual culture with *Phytolacca americana* callus and *Botrytis fabae*. *Zeitschrift für Naturforschung, Section C, Biosciences* 50(5–6): 398–402.
756. Kodama, T., Yamakawa, T. & Minoda, Y., 1980. Rotenoid biosynthesis by tissue culture of *Derris elliptica*. *Agricultural and Biological Chemistry* 44(10): 2387–2390.
757. Kokate, C.K., Tipnis, H.P. & Gonsalves, L.X., 1980. Anti-insect and juvenile hormone mimicking activities of essential oils of *Adhatoda vasica*, *Piper longum* and *Cyperus rotundus*. In: 4th Symposium on Medicinal Plants and Spices, 15–19 September 1980, Mahidol University, Bangkok, Thailand. p. 154 (summary only).
758. Kokwaro, J.O., 1976. Medicinal plants of East Africa. East African Literature Bureau, Nairobi, Kampala, Dar es Salaam. 384 pp.
759. Kolte, R.M., Bisan, V.V., Kalorey, D.R. & Harne, S.D., 1996. Chloroform extract of roots of *Trichosanthes cucumerina* Linn. as antibacterial against *Staphylococcus aureus* and *Pseudomonas aeruginosa*. *Indian Journal of Veterinary Research* 5(1): 35–38.
760. Komai, K. & Tang, C.S., 1989. Chemical constituents and inhibitory activities of essential oils from *Cyperus brevifolius* and *Cyperus kyllingia*. *Journal of Chemical Ecology* 15(8): 2171–2176.
761. Komai, K., Tang, C.S. & Nishimoto, R.K., 1991. Chemotypes of *Cyperus rotundus* in Pacific rim and basin: distribution and inhibitory activities of their essential oils. *Journal of Chemical Ecology* 17(1): 1–8.



762. Komine, H., Takahashi, T. & Ayabe, S.I., 1996. Properties and partial purification of squalene synthase from cultured cells of dandelion. *Phytochemistry* 42(2): 405–409.
763. Konath, S.S., Nadimpalli, S.K. & Swamy, M.J., 1996. Purification in high yield and characterisation of the galactose-specific lectin from the seeds of snake gourd (*Trichosanthes anguina*). *Biochemistry and Molecular Biology International* 39(2): 243–252.
764. Kondo, T., Inoue, M., Mizukami, H. & Ogihara, Y., 1995. Cytotoxic activity of bryonolic acid isolated from transformed hairy roots of *Trichosanthes kirilowii* var. *japonica*. *Biological and Pharmaceutical Bulletin* 18(5): 726–729.
765. Kondo, T., Mizukami, H., Takeda, T. & Ogihara, Y., 1996. Amino acid sequences and ribosome-inactivating activities of karasurin-B and karasurin-C. *Biological and Pharmaceutical Bulletin* 19(11): 1485–1489.
766. Koorders, S.H., 1901. Kleine schetsen van merkwaardige javaansche planten. Schetsen No 11–14 [Short descriptions of peculiar Javanese plants. Description No 11–14]. *Teijsmannia* 11: 558–577.
767. Koorders, S.H., 1912. *Exkursionsflora von Java* [Excursion flora of Java]. Vol. 2. Gustav Fischer, Jena, Germany. 742 pp.
768. Koorders, S.H. & Valetton, T., 1894–1915. *Bijdrage tot de kennis der boomsoorten van Java* [Contribution to the knowledge of the tree species of Java]. 13 parts. G. Kolff & Co., Batavia, Dutch East Indies, 's-Gravenhage, the Netherlands.
769. Kopp, B., Bauer, W.P. & Bernkop-Schnürch, A., 1992. Analysis of some Malaysian dart poisons. *Journal of Ethnopharmacology* 36(1): 57–62.
770. Kosasi, S., 't Hart, L.A., van Dijk, H. & Labadie, R.P., 1989. Inhibitory activity of *Jatropha multifida* latex on classical complement pathway activity in human serum mediated by a calcium binding proanthocyanidin. *Journal of Ethnopharmacology* 27: 81–89.
771. Kosasi, S., van der Sluis, W.G., Boelens, R., 't Hart, L.A. & Labadie, R.P., 1989. Labaditin, a novel cyclic peptide from the latex of *Jatropha multifida* L. *FEBS Letters* 256: 91–96.
772. Kosasi, S., van der Sluis, W.G. & Labadie, R.P., 1989. Multifidol and Multifidol glucoside from the latex of *Jatropha multifida*. *Phytochemistry* 28: 2439–2441.
773. Koseki, I., Simoni, I.C., Nakamura, I.T., Noronha, A.B. & Costa, S.S., 1990. Antiviral activity of plant extracts against aphthovirus, pseudorabies virus and pestivirus in cell cultures. *Microbios Letters* 44(173): 19–30.
774. Koster, J.T., 1935. *The Compositae of the Malay Archipelago. I. Vernoniaeae and Eupatorieae*. *Blumea* 1: 351–536.
775. Koster, J.T., 1972. *The Compositae of New Guinea III*. *Blumea* 20: 13–226.
776. Kosuge, T., 1985. Studies on antitumour activities and antitumour principles of Chinese herbs. *Yakugaku Zasshi* 105(8): 791–795. (in Japanese)
777. Kosuge, T., Ishida, H., Yamazaki, H. & Ishii, M., 1984. Studies on active substances in the herbs used for oketsu, blood coagulation, in Chinese medicine. I. On antiagulative activities of the herbs for oketsu. *Yakugaku Zasshi* 104(10): 1050–1053. (in Japanese)

778. Kosuge, T., Yokota, M., Sugiyama, K., Yamamoto, T., Ni, M.Y. & Yan, S.C., 1985. Studies on antitumour activities and antitumour principles of Chinese herbs. *Yakugaku Zasshi* 105(8): 791-795. (in Japanese)
779. Koyama, T., 1960. Materials toward a monograph of the genus *Smilax*. *Quarterly Journal of the Taiwan Museum* 13: 1-62.
780. Koyama, T., 1975. Smilacaceae. In: Smitinand, T. & Larsen, K. (Editors): *Flora of Thailand*. Vol. 2. The Forest Herbarium, Royal Forest Department, Bangkok, Thailand. pp. 211-250.
781. Koyama, T., 1978. Smilacaceae. In: Li, H.-L., Liu, T.-S., Huang, T.-C., Koyama, T. & DeVol, C.E. (Editors): *Flora of Taiwan*. Vol. 5. Epoch Publishing Co., Taipei, Taiwan, Republic of China. pp. 110-137.
782. Krijgsman, B.J., 1948. De insecticide werking van derris (literatuurstudie) [The insecticide activity of derris (literature study)]. Rapport T.A. 262. Algemene Technische Afdeling TNO, The Hague, the Netherlands. 158 pp. (mimeographed).
783. Krishnakumar, A., Sivaramkrishnan, V.M. & Sivaswamy, S.N., 1991. Inhibition of nitrosation reaction by some spices/leafy vegetables. *Advances in Plant Sciences* 4(1): 189-193.
784. Krogsgaard-Larsen, P. & Bundgaard, H. (Editors), 1991. A textbook of drug design and development. Harwood Academic Publishers, Chur, Switzerland.
785. Kubo, I., Taniguchi, M., Chapya, A. & Tsujimoto, K., 1980. An insect antifeedant and antimicrobial agent from *Plumbago capensis*. *Planta Medica*, Supplement: 185-187.
786. Kubo, I., Uchida, M. & Klocke, J.A., 1983. An insect ecdysis inhibitor from the African medicinal plant, *Plumbago capensis* (Plumbaginaceae); a naturally occurring chitin synthetase inhibitor. *Agricultural and Biological Chemistry* 47(4): 911-913.
787. Kugelman, M., Liu, W.C., Axelrod, M., McBride, T.J. & Rao, K.V., 1976. Indicin-N-oxide: the anti-tumor principle of *Heliotropium indicum*. *Lloydia* 39(2-3): 125-128.
788. Kulp, K.S. & Vulliet, P.R., 1996. Mimosine blocks cell cycle progression by chelating iron in asynchronous human breast cancer cells. *Toxicology and Applied Pharmacology* 139(2): 356-364.
789. Kumagai, M.H., Turpen, T.H., Weinzettl, N., Della-Cioppa, G., Turpen, A.M., Donson, J., Hilf, M.E., Granthma, G.L., Dawson, W.O., Chow, T.P., Piatak Jr., M. & Grill, L.K., 1993. Rapid, high-level expression of biologically active  $\alpha$ -trichosanthin in transfected plants by an RNA viral vector. *Proceedings of the National Academy of Sciences* 90(2): 427-430.
790. Kumar, A. & Datta, S.K., 1989. Plantlet regeneration from hypocotyl tissue of *Strychnos nux-vomica* Linn. *Current Science* 58(14): 812-813.
791. Kumar, B.H. & Thakur, S.S., 1988. Certain non-edible seed oils as feeding deterrents against *Spodoptera litura* Fb. *Journal of the Oil Technologists' Association of India* 20(3): 63-65.
792. Kumar, C.P. et al., 1997. A comparative study on the hypolipidemic activity of eleven different pectins. *Journal of Food Science and Technology* 34(2): 103-107.
793. Kumar, K.S. & Bhavanandan, K.V., 1988. Micropropagation of *Plumbago rosea* Linn. *Plant Cell, Tissue and Organ Culture* 15: 275-278.

794. Kumar, M. & Berwal, J.S., 1998. Sensivity of food pathogens to garlic (*Allium sativum*). *Journal of Applied Microbiology* 84(2): 213-215.
795. Kumar, R.V. & Augusti, K.T., 1989. Antidiabetic effect of a leucocyanidin derivative isolated from the bark of *Ficus bengalensis* L. *Indian Journal of Biochemistry and Biophysics* 26: 400-404.
796. Kunakh, V.A., Kaukhova, I.E., Alpatova, L.K. & Vollosovich, A.G., 1982. Peculiarities of the behavior of cells in *Rauwolfia serpentina* Benth. tissue culture. *Cytology and Genetics* 16(5): 6-10.
797. Kung, S., Kimura, M. & Funatsu, G., 1990. The complete amino acid sequence of antiviral protein from the seeds of pokeweed (*Phytolacca americana*). *Agricultural and Biological Chemistry* 54(12): 3301-3318.
798. Kupchan, S.M., Suffness, M.I., White, D.N., McPhail, A.T. & Sim, G.A., 1968. The isolation and structural elucidation of 4-dimethylhasubanone, a new alkaloid from *Stephania hernandifolia*. *Journal of Organic Chemistry* 33(12): 4529-4532.
799. Kuroda, M., Mimaki, Y., Kameyama, A., Sashida, Y. & Nikaido, T., 1995. Steroidal saponins from *Allium chinense* and their inhibitory activities on cyclic AMP phosphodiesterase and Na<sup>+</sup>/K<sup>+</sup> ATPase. *Phytochemistry* 40(4): 1071-1076.
800. Kusumoto, I.T., Nakabayashi, T., Kida, H., Miyashiro, H., Hattori, M., Namba, T. & Shimothno, K., 1995. Screening of various plant extracts used in ayurvedic medicine for inhibitory effects on human immuno deficiency virus Type 1 (HIV-1) protease. *Phytotherapy Research* 9(3): 180-184.
801. Kuwahara, S., Awai, N. & Kodama, O., 1995. A revised structure for rhinocanthone. *Journal of Natural Products* 58(9): 1455-1458.
802. Kyi Thein et al., 1995. Preliminary screening of medicinal plants for biological activity based on inhibition of cyclic AMP phosphodiesterase. *International Journal of Pharmacognosy* 33(4): 330-333.
803. La Dinh Moi et al., 1988. First results of research on selection and acclimatization of the two mint (*Mentha arvensis* L.) cultivars NV-74 and NV-76. *Proceedings of the Seminar on Technology of Essential oils* (6-9 December, 1988), Hanoi. pp. 67-77. (in Vietnamese)
804. Laakso, I., Seppanen-Laakso, T. & Hiltunen, R., 1989. Composition of the essential oil of *Blumea lacera* DC. (Asteraceae) leaves from Nigeria. *Flavour and Fragrance Journal* 4(2): 73-75.
805. Lagrota, M.H.C., Wigg, M.D., Miranda, M.M.F.S., Santos, M.G.M. & Costa, S.S., 1995. Inhibition of herpes simplex virus replication by different extracts of Caryophyllales. *Biomedical Letters* 51(202): 127-135.
806. Laitinen, L., Juusela, H. & Virtanen, I., 1990. Binding of the blood group-reactive lectins to human adult kidney specimens. *Anatomical Record* 226(1): 10-17.
807. Lakshmi, V.V., Padma, S. & Polasa, H., 1987. Elimination of multi-drug-resistant plasmid in bacteria by plumbagin, a compound derived from a plant. *Current Microbiology* 16: 159-161.
808. Lal, J., Chandra, S., Raviprakash, V. & Sabir, M., 1976. In vitro anthelmintic action of some indigenous medicinal plants on *Ascaridia galli* worms. *Indian Journal of Physiology and Pharmacology* 20(2): 64-68.
809. Lam, H.J., 1919. *The Verbenaceae of the Malayan Archipelago*. M. de

- Waal, Groningen, the Netherlands. 370 pp.
810. Lam, H.J. & Bakhuizen van den Brink, R.C., 1921. Revision of the Verbenaceae of the Dutch East-Indies and surrounding countries. Bulletin du Jardin Botanique de Buitenzorg, Série III, 3: 1–116.
811. Lampe, F.K., 1976. Changes in therapy in Abrus and Ricinus poisoning suggested by recent studies in their mechanism of toxicity. *Clinical Toxicology* 9(1): 21.
812. Lange, D. & Schippmann, U., 1997. Trade survey of medicinal plants in Germany. A contribution to international plant species conservation. Bundesamt für Naturschutz, Bonn, Germany. 128 pp. + annexes.
813. Langer, R.H.M. & Hill, G.D., 1982. Agricultural plants. Cambridge University Press, Cambridge, United Kingdom. 344 pp.
814. Lanhers, M.C., Fleurentin, J., Dorfman, P., Mortier, F. & Pelt, J.M., 1991. Analgesic, antipyretic and anti-inflammatory properties of *Euphorbia hirta*. *Planta Medica* 57(3): 225–231.
815. Laranja, S.M.R., Bergamachi, C.M. & Schor, N., 1991. Evaluation of the acute administration of natural products with potential diuretic effects, in humans. *Memoires del Instituto Oswaldo Cruz (Rio de Janeiro)* 86, Suppl. II: 237–240.
816. Larsen, K., 1989. Caryophyllales. In: Lescot, M. (Editor): *Flore du Cambodge, du Laos et du Viêt Nam [Flora of Cambodia, Laos and Vietnam]*. Vol. 24. Muséum National d'Histoire Naturelle, Paris, France. 144 pp.
817. Larsen, K., Larsen, S.S. & Vidal, J.E., 1980. Légumineuses-Caesalpinioïdées [Leguminosae-Caesalpinioideae]. In: Vidal, J.E. & Vidal, Y. (Editors): *Flore du Cambodge, du Laos et du Viêt Nam [Flora of Cambodia, Laos and Vietnam]*. Vol. 18. Muséum National d'Histoire Naturelle, Paris, France. 227 pp.
818. Lasure, A., Van Poel, B., De Clarck, L.S., Bridts, C.H., Stevens, W.J., Rwangabo, P.C., Pieters, L. & Vlietinck, A.J., 1995. Screening of Rwandese plant extracts for their influence on lymphocyte proliferation. *Phytomedicine* 1(4): 303–307.
819. Latha, P. & Govindasamy, S., 1997. Gangetin – a reproductive inhibitor in male rats. *Phytotherapy Research* 11(6): 466–468.
820. Latha, P., Govindasamy, S. & Balakrishna, K., 1997. Effect of gangetin on fertility of male rats. *Phytotherapy Research* 11(5): 372–375.
821. Latha, R.M., Geetha, T. & Varalakshmi, P., 1998. Effect of *Vernonia cinerea* Less. flower extract in adjuvant-induced arthritis. *General Pharmacology* 31(4): 601–606.
822. Latiff, A., 1991. Plant resources for natural products: an ethnobotanical perspective. In: Shaari, K., Kadir, A.A. & Ali, A.R.H. (Editors): *Medicinal products from tropical rain forests. Proceedings of the conference, Forest Research Institute Malaysia, May 13–15, 1991. Forest Research Institute Malaysia, Kepong, Selangor, Malaysia.* pp. 1–12.
823. Laughlin, J.C., 1992. Evaluation of six medicinal crop plants in Tasmania, Australia. *Acta Horticulturae* 306: 100–104.
824. Lauzac-Marchal, M., 1974. Taxonomie végétale – Réhabilitation du genre *Bryophyllum* Salisb. (Crassulacées, Kalanchoïdées). [Plant taxonomy – Rehabilitation of the genus *Bryophyllum* Salisb. (Crassulaceae, Kalanchoideae)]. *Comptes rendus hebdomadaires des séances de l'Académie des*

- Sciences Paris. Série D. Sciences naturelles 278: 2505-2508.
825. Lawson, L.D., Wang, Z.-Y.J. & Hughes, B.G., 1991. Identification and HPLC quantitation of the sulfides and dialk(en)yl thiosulfates in commercial garlic products. *Planta Medica* 57: 363-370.
826. Le Van Truyen et al., 1983. Contribution to the study on galenical dosage forms of *Plantago major* var. *asiatica* Descaine. *Tap chi Duoc hoc* 6: 15-18. (in Vietnamese)
827. Leach, A.J., Leach, D.N. & Leach, G.J., 1988. Antibacterial activity of some medicinal plants of Papua New Guinea. *Science in New Guinea* 14(1): 1-7.
828. Leach, L.C., 1973. *Euphorbia tirucalli* L.: its typification, synonymy and relationships, with notes on 'Almeidina' and 'Cassoneira'. *Kirkia* 9: 69-86.
829. Leaman, D.J., Yusuf, R. & Sangat-Roemantyo, H., 1991. Kenyah Dayak forest medicines. Prospects for development and implications for conservation. Report for the World Wide Fund for Nature, Indonesia Programme. 34 pp. + appendices.
830. Leamon, C.P. & Low, P.S., 1994. Selective targeting of malignant cells with cytotoxin-folate conjugates. *Journal of Drug Target* 2(2): 101-112.
831. Lecompte, O., 1969. Combretaceae. In: Tardieu-Blot, M.-L. (Editor): *Flore du Cambodge, du Laos et du Vietnam* [Flora of Cambodia, Laos and Vietnam]. Vol. 10. Muséum National d'Histoire Naturelle, Paris, France. 119 pp.
832. Lee, H. & Lin, J.Y., 1988. Antimutagenic activity of extracts from anti-cancer drugs in Chinese medicine. *Mutation Research* 204(2): 229-234.
833. Lee, K.-H., Ibuka, T., Furukawa, H., Kozuka, M., Wu, R.Y., Hall, I.H. & Huang, H.-C., 1980. Antitumor agents. XXXVIII: Isolation and structural elucidation of novel germacranolides and triterpenes from *Elephantopus mollis*. *Journal of Pharmaceutical Sciences* 69(9): 1050-1056.
834. Lee, K.H., Hayashi, N., Okano, M., Nozaki, H. & Ju-Ichi, M., 1984. Antitumor agents, 65. Brusatol and cleomiscosin-A, antileukemic principles from *Brucea javanica*. *Journal of Natural Products* 47(3): 550-551.
835. Lee-Huang, S., Huang, P.L., Chen, H.C., Huang, P.L., Bourinbaiar, A.S., Huang, H.I. & Kung, H.F., 1995. Anti-HIV and anti-tumor activities of recombinant MAP30 from bitter melon. *Gene* 161(2): 151-156.
836. Lee-Huang, S., Huang, P.L., Huang, P.L., Bourinbaiar, A.S., Chen, H.C. & Kung, H.F., 1995. Inhibition of the integrase of human immunodeficiency virus (HIV) type 1 by anti-HIV plant proteins MAP30 and GAP31. *Proceedings of the National Academy of Sciences* 92(19): 8818-8822.
837. Lee-Huang, S., Huang, P.L., Kung, H.F., Li, B.Q., Huang, P.L., Huang, P., Huang, H.I. & Chen, H.C., 1991. TAP 29: an anti-human immunodeficiency virus protein from *Trichosanthes kirilowii* that is nontoxic to intact cells. *Proceedings of the National Academy of Sciences* 88(15): 6570-6574.
838. Leelarasamee, A., Trakulsomboon, S., Maunwongyathi, P., Somabandhu, A., Pidetcha, P., Matrakool, B., Lebnak, T., Ridthimat, W. & Chandanayingyong, D., 1990. Failure of *Phyllanthus amarus* to eradicate hepatitis B surface antigen from symptomless carriers. *The Lancet* 335(8705): 1600-1601.
839. Leenhouts, P.W., 1962. Loganiaceae. In: van Steenis, C.G.G.J. (General editor): *Flora Malesiana*. Series 1, Vol. 6. Wolters-Noordhoff Publishing,

- Groningen, the Netherlands. pp. 293–387.
840. Leeuwenberg, A.J.M. (Compiler), 1987. Medicinal and poisonous plants of the tropics. Proceedings of Symposium 5-35 of the 14th International Botanical Congress, Berlin, 24 July–1 August 1987. Pudoc, Wageningen, the Netherlands. 152 pp.
841. Lemma, A., 1970. Laboratory and field evaluation of the molluscicidal properties of *Phytolacca dodecandra*. Bulletin of the World Health Organization 42: 597–612.
842. Lemmens, R.H.M.J. & Wulijarni-Soetjipto, N. (Editors), 1991. Plant Resources of South-East Asia No 3. Dye and tannin-producing plants. Pudoc, Wageningen, the Netherlands. 196 pp.
843. Lescot, M., 1970. Flacourtiaceae. In: Tardieu-Blot, M.-L. (Editor): Flore du Cambodge, du Laos et du Viêt Nam [Flora of Cambodia, Laos and Vietnam]. Vol. 11. Muséum National d'Histoire Naturelle, Paris, France. pp. 3–98.
844. Leslie, G.B., 1978. A pharmacometric evaluation of nine bio-strath herbal remedies. Medita 810: 3–19.
845. Letchamo, W. & Gosselin, A., 1996. Light, temperature and duration of storage govern the germination and emergence of *Taraxacum officinale* seed. Journal of Horticultural Science 71(3): 373–377.
846. Letourneux, C., 1957. Tree planting practices in tropical Asia. FAO Forestry Development Paper No 11. Food and Agriculture Organization, Rome, Italy. 172 pp.
847. Lewis, W.H. & Elwin-Lewis, M.P.F., 1977. Medical Botany. Plants affecting man's health. John Wiley & Sons, New York, United States. 515 pp.
848. Li, H.-L., 1942. The Araliaceae of China. Sargentia 2. The Arnold Arboretum of Harvard University, Jamaica Plain, United States. 134 pp.
849. Li, H.-L., 1978. Myrsinaceae. In: Li, H.-L., Liu, T.-S., Huang, T.-C., Koyama, T. & DeVol, C.E. (Editors): Flora of Taiwan. Vol. 4. Epoch Publishing Co., Taipei, Taiwan, Republic of China. pp. 47–67.
850. Li, H.-L., 1978. Apocynaceae. In: Li, H.-L., Liu, T.-S., Huang, T.-C., Koyama, T. & DeVol, C.E. (Editors): Flora of Taiwan. Vol. 4. Epoch Publishing Co., Taipei, Taiwan, Republic of China. pp. 202–221.
851. Li, H.-L., 1978. Plantaginaceae. In: Li, H.-L., Liu, T.-S., Huang, T.-C., Koyama, T. & DeVol, C.E. (Editors): Flora of Taiwan. Vol. 4. Epoch Publishing Co., Taipei, Taiwan, Republic of China. pp. 701–703.
852. Li, H.-L., 1978. Compositae. In: Li, H.-L., Liu, T.-S., Huang, T.-C., Koyama, T. & DeVol, C.E. (Editors): Flora of Taiwan. Vol. 4. Epoch Publishing Co., Taipei, Taiwan, Republic of China. pp. 768–965.
853. Li, H.-L. & Lo, H.-C., 1993. Sterculiaceae. In: Huang, T.-C. (Editor): Flora of Taiwan. 2nd Edition. Vol. 3. Editorial Committee of the Flora of Taiwan, Taipei, Taiwan, Republic of China. pp. 756–771.
854. Li, H.-W. & Hedge, I.C., 1994. Lamiaceae (Labiatae). In: Wu, Z.-Y. & Raven, P.H. (Editors): Flora of China. Vol. 17. Science Press, Beijing, China & Missouri Botanical Garden, St. Louis, United States. pp. 50–299.
855. Li, R.S., Yu, C.L. & Jin, X.Z., 1989. Effect of oxymatrine on beating of cultured myocardial cells in vitro. Chung Kuo Yao Li Hsueh Pao 10(6): 530–532.
856. Liao, J.-C., 1996. Moraceae. In: Huang, T.-C. (Editor): Flora of Taiwan.

- 2nd Edition. Vol. 2. Editorial Committee of the Flora of Taiwan, Taipei, Taiwan, Republic of China. pp. 136–195.
857. Likhitwitayawuid, K., Angerhofer, C., Cordell, G.A. & Pezzuto, J.M., 1993. Cytotoxic and antimalarial bisbenzylisoquinoline alkaloids from *Stephania erecta*. *Journal of Natural Products* 56: 30–38.
858. Lim-Sylianco, C.Y., Concha, J.A., Jocano, A.P. & Lim, C.M., 1986. Antimutagenic effects of eighteen Philippine plants. *Philippine Journal of Science* 115(4): 293–298.
859. Lim-Sylianco, C.Y., Concha, J.A., San Augustin, J., Panizares, I. & Pablo, C., 1980. Mutagenicity and clastogenicity potential of decoctions and infusions from Philippine medicinal plants. *Bulletin of the Philippine Biochemical Society* 3: 54–65.
860. Lin, C.C., Chen, J.Y. & Namba, T., 1986. Development of natural crude drug resources from Taiwan IV. Pharmacognostical studies on the Chinese crude drug han-lian-cao. *Shoyakugaku Zasshi* 40(4): 357–366. (in Japanese)
861. Lin, C.C., Tsai, C.C. & Yen, M.H., 1995. The evaluation of hepatoprotective effects of Taiwan folk medicine "Teng-Khia-U". *Journal of Ethnopharmacology* 45(2): 113–123.
862. Lin, C.C., Yen, M.H. & Chiu, H.F., 1991. The pharmacological and pathological studies on Taiwan folk medicine VI. The effects of *Elephantopus scaber* ssp. *oblanceolata*, *Elephantopus mollis* and *Pseudoelephantopus spicatus*. *American Journal of Chinese medicine* 19(1): 41–50.
863. Lin, L.Z., Shieh, H.L., Angerhofer, C.K., Pezzuto, J.M., Cordell, G.A., Xue, L., Johnson, M.E. & Ruangrunsi, N., 1993. Cytotoxic and antimalarial bisbenzylisoquinoline alkaloids from *Cyclea barbata*. *Journal of Natural Products* 56: 22–29.
864. Lin, S.-C. & Hsieh, C.-F., 1991. A taxonomic study of the genus *Euphorbia* L. (Euphorbiaceae) in Taiwan. *Taiwania* 36: 57–79.
865. Lin, S.-C. & Hsieh, C.-F., 1993. *Euphorbia*. In: Huang, T.-C. (Editor): *Flora of Taiwan*, 2nd Edition. Vol. 3. Editorial Committee of the Flora of Taiwan, Taipei, Taiwan, Republic of China. pp. 456–469.
866. Lin, S.C. et al., 1995. Protective and therapeutic effects of *Curcuma xanthorrhiza* on hepatotoxin-induced liver damage. *American Journal of Chinese Medicine* 23(3–4): 243–254.
867. Lin, S.C., Teng, C.W., Lin, C.C., Lin, Y.H. & Supriyatna, S., 1996. Protective and therapeutic effect of the Indonesian medicinal herb *Curcuma xanthorrhiza* on  $\beta$ -D-galactosamine-induced liver damage. *Phytotherapy Research* 10(2): 131–135.
868. Lin, T.-S. & Kao, M.-T., 1996. *Amaranthaceae*. In: Huang, T.-C. (Editor): *Flora of Taiwan*, 2nd Edition. Vol. 2. Epoch Publishing Co., Taipei, Taiwan, Republic of China. pp. 388–409.
869. Lissoni, P., Giani, L., Zerbini, S., Trabattoni, P. & Rovelli, F., 1998. Biotherapy with the pineal immunomodulating hormone melatonin versus melatonin plus *Aloe vera* in untreatable advanced solid neoplasms. *Natural Immunity* 16(1): 27–33.
870. Liu, H.-Y., 1993. *Cucurbitaceae*. In: Huang, T.-C. (Editor). *Flora of Taiwan*, 2nd Edition. Vol. 3. Editorial Committee of the Flora of Taiwan, Taipei, Taiwan, Republic of China. pp. 855–871.

871. Liu, I.M. & Sheu, S.J., 1989. Analysis and processing of Chinese herbal drugs. VIII: The study of sophorae floe. *American Journal of Chinese Medicine* 17(3-4): 179-187.
872. Liu, K.C.-S.C., Yang, S.-L., Roberts, M.F., Elford, B.C. & Phillipson, J.D., 1992. Antimalarial activity of *Artemisia annua* flavonoids from whole plants and cell cultures. *Plant Cell Reports* 11(12): 637-640.
873. Liu, Y.-C. & Ou, C.-H., 1978. Solanaceae. In: Li, H.-L., Liu, T.-C., Huang, T.-C., Koyama, T. & DeVol, C.E. (Editors): *Flora of Taiwan*. Vol. 4. Epoch Publishing Co., Taipei, Taiwan, Republic of China. pp. 530-550.
874. Locher, C.P., Witrouw, M., De Bethune, M.P., Burch, M.T., Mover, H.F., Davis, H., Lasure, A., Pauwels, R., De Clerq, E. & Elietinck, A.J., 1996. Antiviral activity of Hawaiian medicinal plants against human immunodeficiency Type 1 (HIV-1). *Phytomedicine* 2(3): 259-264.
875. Lock, J.M., 1985. Zingiberaceae. In: Polhill, R.M. (Editor): *Flora of Tropical East Africa*. A.A. Balkema, Rotterdam, the Netherlands & Boston, United States. 37 pp.
876. Lu, C.F., Yang, L.L. & Yen, K.Y., 1992. Immunomodulative effects of Taiwan herbs. *Journal of the Chinese Society of Veterinary Science* 18(4): 233-242. (in Chinese)
877. Lugt, Ch.B., 1986. *Phytolacca dodecandra* berries as a means of controlling bilharzia transmitting snails. 3rd Edition. Bulletin 312, Royal Tropical Institute, Amsterdam, the Netherlands. 61 pp.
878. Luo, X.-D. & Shen, C.-C., 1987. The chemistry, pharmacology and clinical applications of quinghaosu (artemisinin) and its derivatives. *Medical Research Reviews* 7: 29-52.
879. Ly, T.D., 1986. Die Familie Apocynaceae Juss. in Vietnam. Teil 2: Spezieller Teil (1) [The family Apocynaceae Juss. in Vietnam. Part 2: special part (1)]. *Feddes Repertorium* 97(7-8): 405-468.
880. Lydon, J., Zimmerman, R.H., Fordham, I.M. & Lusby, W.R., 1993. Tissue culture and alkaloid production of *Erythroxylum coca* var. *coca*. *Journal of Herbs, Spices & Medicinal Plants* 2(1): 3-14.
881. Mabberley, D.J., 1997. *The Plant-Book*. A portable dictionary of the vascular plants. 2nd Edition. Cambridge University Press, Cambridge, United Kingdom. 858 pp.
882. Madav, S., Tripathi, H.C., Tandan & Mishra, S.K., 1995. Analgesic, antipyretic and antiulcerogenic effects of andrographolide. *Indian Journal of Pharmaceutical Studies* 57(3): 121-125.
883. Mader, F.H., 1990. Treatment of hyperlipidaemic with garlic-powder tablets. *Arzneimittel Forschung* 40: 1111-1116.
884. Maeda, C., Ohtani, K., Kasai, R., Yamasaki, K., Nguyen, M.D., Nguyen, T.N. & Nguyen, K.Q., 1994. Oleanane and ursane glycosides from *Schefflera octophylla*. *Phytochemistry* 37(4): 1131-1137.
885. Maghirang, R.G., 1985. Performance of some medicinal plants under varying degrees of shade and fertilizer levels. Terminal Report of NSTA-UPLB Project 7716-1AG, January 1978-June 1985. Philippine Council for Health and Resources Development, Manila, Philippines. 56 pp.
886. Mahmood, I., Masood, A., Saxena, S.K. & Husain, S.I., 1979. Effect of some plant extracts on the mortality of *Meloidogyne incognita* and *Rotylenchulus reniformis*. *Acta Botanica Indica* 7(2): 129-132.



887. Maini, P.N. & Morallo-Rejesus, B., 1993. Molluscicidal activity of *Derris elliptica* (Fam. Leguminosae). *Philippine Journal of Science* 122(1): 61-75.
888. Majumdar, N.C. & Banerjee, R.N., 1976. The distribution and economic uses of *Alternanthera philoxeroides* (Mart.) Griseb., an exotic aquatic herb in India and Burma. *Bulletin of the Botanical Society of Bengal* 30: 147-148.
889. Makinde, J.M., Obih, P.O. & Jimoh, A.A., 1987. Effect of *Solanum erianthum* aqueous leaf extract on *Plasmodium berghei berghei* in mice. *African Journal of Medicine and Medical Sciences* 16(4): 193-196.
890. Makwana, H.G., Ravishankar, B., Shukla, V.J., Nair, R.B., Vijayan, N.P., Sasikala, C.K., Saraswathy, V.N. & Bhatt, S.V., 1994. General pharmacology of *Vitex leucoxydon* Linn. leaves. *Indian Journal of Physiology and Pharmacology* 38(2): 95-100.
891. Mallabaev, A., Rakhimov, D.A. & Murdakhaev Yu, M., 1989. Carbohydrates of *Aerva lanata*. *Chemistry of Natural Compounds* 25(3): 369-370.
892. Malone, M.H., 1981. The pharmacological evaluation of natural products - general and specific approaches to screening ethnopharmaceuticals. In: *Proceedings of the 7th symposium for Pharmacognosy, October 10, 1980, Vrije Universiteit Brussel, Brussels, Belgium.* pp. 3-28.
893. Mamatha, H., Farooqi, A.A., Joshi, S.S. & Prasad, T.G., 1993. Pollen studies in *Gloriosa superba* Linn. In: *Palevitch, D., Simon, J.E. & Mathé, A. (Editors): Raw material production, product introduction. Proceedings of the First World Congress on Medicinal and Aromatic Plants for Human Welfare, WOCMAP. Maastricht, the Netherlands, 19-25 July 1992. Acta Horticulturae* 331: 371-376.
894. Manalo, J.B., 1982. A study of lagundi oil: the essential oil from *Vitex negundo* Linn. growing in the Philippines. *Philippine Journal of Science* 111(3-4): 79-97.
895. Manandhar, N.P., 1991. Medicinal plant-lore of Tamang tribe of Kabhrepalanchok District, Nepal. *Economic Botany* 45: 58-71.
896. Mandava, N., Anderson, J.D. & Dutky, S.R., 1974. Indole plant-growth inhibitor from *Abrus precatorius* seeds. *Phytochemistry* 13(12): 2853-2856.
897. Mangaly, J. K. & Sabu, M., 1993. A taxonomic revision of the South Indian species of *Curcuma* Linn. (Zingiberaceae). *Rheedea* 3(2): 139-171.
898. Mangenot, G., Bancilhon, L. & Mangenot, S., 1977. Caryologie du genre *Phyllanthus* (Euphorbiaceae, Phyllanthoideae) [Karyology of the genus *Phyllanthus* (Euphorbiaceae, Phyllanthoideae)]. *Annales des Sciences Naturelles, Botanique, Paris, 12e Série*, 18: 71-116.
899. Manipula, E.M., 1986. Tuba-tuba: a hydrocarbon and medicinal plant. *Canopy International* 12(4): 12.
900. Mansfeld, R., 1986. *Verzeichnis landwirtschaftlicher und gaertnerischer Kulturpflanzen (ohne Zierpflanzen)* [Register of agricultural and horticultural plants (withouth ornamentals)]. 2nd Edition, revised by J. Schultze-Motel. 4 volumes. Springer Verlag, Berlin, Germany. 1998 pp.
901. Mansor, P., 1988. Ula-ulam tradisional Malaysia [Traditional vegetables in Malaysia]. *Teknologi Sayur-sayuran* 4: 1-5.
902. Maraganore, J.M., Joseph, M. & Bailey, M.C., 1987. Purification and characterization of trichosanthin. Homology to the ricin A chain and implica-

- tions as to mechanism of abortifacient activity. *Journal of Biological Chemistry* 262(24): 11628–11633.
903. Marco, J.A., Sanz, J.F., Bea, J.F. & Barbera, O., 1990. Phenolic constituents from *Artemisia annua*. *Pharmazie* 45: 382–383.
904. Marimuthu, S., Subramanian, R.B., Kothari, I.L. & Inamdar, J.A., 1989. Laticiferous taxa as a source of energy and hydrocarbon. *Economic Botany* 43(2): 255–261.
905. Markgraf, F., 1984. *Florae Malesianae Praecursores* LXIV. Apocynaceae VI. *Rauvolfia*. *Blumea* 30: 157–167.
906. Martin, G.J., 1995. *Ethnobotany. A methods manual*. Chapman & Hall, London, United Kingdom. 268 pp.
907. Martin, H.-J., Pachaly, P. & Zymalkowski, F., 1977. Alkaloide aus der thailändischen Menispermaceen-Droge Krung Kha Mao (*Cyclea barbata*), 6. Mitt. Isolierung und Strukturaufklärung quartärer Tetrahydroprotoberberin-Alkaloide [Alkaloids from the Thai Menispermaceae drug krung kha mao (*Cyclea barbata*), part 6. Isolation and structural elucidation of quaternary tetrahydroprotoberberine alkaloids]. *Archiv der Pharmazie (Weinheim)* 310(4): 314–319.
908. Martin, T.S., Ohtani, K., Kasai, R. & Yamasaki, K., 1995. Clerodane diterpene glucosides from *Tinospora rumphii*. *Phytochemistry* 40(6): 1729–1736.
909. Martono, S. & Udayati, 1975. Efek analgetika dari infus daun *Gendarussa vulgaris* Nees pada mencit (penelitian pendahuluan) [Analgetic effect of *Gendarussa vulgaris* Nees leaf infusion on mice (A preliminary study)]. Thesis, Faculty of Pharmacy, Gadjah Mada University, Yogyakarta, Indonesia.
910. Mascolo, N., Autore, G., Capasso, F., Menghini, A. & Fasculo, M.P., 1987. Biological screening of Italian medicinal plants for antiinflammatory activity. *Phytotherapy Research* 11: 28–31.
911. Massiot, G. et al., 1983. Occurrence of longicaudatine, a new type of bisindole base and bisnor-C alkaloid H in *Strychnos* spp. *Journal of Organic Chemistry* 48(11): 1869–1872.
912. Masuda, T., Isobe, J., Jitoe, A. & Nakatani, N., 1992. Antioxidative curcuminoids from rhizomes of *Curcuma xanthorrhiza*. *Phytochemistry* 31(10): 3645–3647.
913. Mathad, M.C. & Nalwadi, U.G., 1989. Rooting ability of some important ornamental climbers. *South Indian Horticulture* 37(5): 307–308.
914. Mathur, R., Sharma, A., Dixit, V.P. & Varma, M., 1996. Hypolipidaemic effect of fruit juice of *Emblica officinalis* in cholesterol-fed rabbits. *Journal of Ethnopharmacology* 50(2): 61–68.
915. Matsui, M., Kabashima, T., Ishida, K., Takebayashi, T. & Watanabe, Y., 1982. Alkaloids of the leaves of *Stephania japonica* (Japan). *Journal of Natural Products* 45(4): 497–500.
916. Matsui, M., Uchida, M., Usuki, I., Saionji, Y., Murata, H. & Watanabe, Y., 1979. Alkaloids of *Stephania japonica* var. *australis*. *Phytochemistry* 18(6): 1087–1088.
917. Matsui, M. & Watanabe, Y., 1984. Structure of oxostephasunoline, a new hasubanalactam alkaloid from *Stephania japonica*. *Journal of Natural Products* 47: 465–469.

918. Matsui, M. & Yamamura, Y., 1986. Alkaloids from the fruits of *Stephania japonica*, part 3. Structures of prostephanaberrine and stephanaberrine, two new hasubanan alkaloids. *Journal of Natural Products* 49(4): 588–592.
919. Matsui, M., Yamamura, Y., Takebayashi, T., Iwaki, K., Takami, Y., Kunitake, K., Koga, F., Urasaki, S. & Watanabe, Y., 1984. Oxoepestephamiersine, a new hasubanalactam alkaloid from *Stephania japonica*. *Journal of Natural Products* 47(5): 858–861.
920. Matthew, K.M., 1977. Reproductive biology of *Bidens pilosa* L. (Compositae). *Current Science* 46(7): 238–239.
921. Matthew, K.M., 1981–1988. The flora of the Tamilnadu Carnatic. 4 volumes. The Rapinat Herbarium, St. Joseph's College, Tiruchirapalli, India.
922. McCaslin, P.P. & Morgan, W.W., 1987. Cultured cerebella cells as an in vitro model of excitatory amino acid receptor function. *Brain Research* 417(2): 380–384.
923. McKenzie, R.A. & Dunster, P.J., 1986. Hearts and flowers: Bryophyllum poisoning of cattle. *Australian Veterinary Journal* 63(7): 222–227.
924. McKenzie, R.A., Franke, F.P. & Dunster, P.J., 1987. The toxicity to cattle and bufadienolide content of six Bryophyllum species. *Australian Veterinary Journal* 64(10): 298–301.
925. McPhail, A.T., Onan, K.D., Lee, K.-H., Ibuka, T., Kozuka, M., Shingu, T. & Huang, H.-C., 1974. Structure and stereochemistry of the epoxide of phantomolin, a novel cytotoxic sesquiterpene lactone from *Elephantopus mollis*. *Tetrahedron Letters* 32: 2739–2741.
926. Melchior, J., Palm, S. & Wikham, G., 1997. Controlled clinical study of standardized *Andrographis paniculata* extract in common cold – a pilot trial. *Phytomedicine* 3(4): 315–318.
927. Mendelsohn, R. & Balick, M.J., 1995. The value of undiscovered pharmaceuticals in tropical forests. *Economic Botany* 49(2): 223–228.
928. Mensah, M., Sarpong, K., Baser, K.H.C. & Ozek, T., 1993. The essential oil of *Ageratum conyzoides* L. from Ghana. *Journal of Essential Oil Research* 5(1): 113–115.
929. Menut, C., Lamaty, G., Zollo, P.H.A., Kuate, J.R. & Bessiere, J.M., 1993. Aromatic plants of tropical central Africa. Part X. Chemical composition of the essential oils of *Ageratum houstonianum* Mill. and *Ageratum conyzoides* L. from Cameroon. *Flavour and Fragrance Journal* 8(1): 1–4.
930. Mercado, B.L., 1979. A monograph on *Cyperus rotundus* L. Biotrop Bulletin No 15. SEAMEO Regional Center for Tropical Biology, Bogor, Indonesia. 63 pp.
931. Merlin, M.D., 1984. On the trail of the ancient opium poppy. Fairleigh Dickinson University Press, Rutherford, Madison, Teaneck, United States. 324 pp.
932. Merrill, E.D., 1908. New Philippine plants from the collections of Mary Strong Clemens, I. *The Philippine Journal of Science* 3, sect. C, Botany: 129–165.
933. Merrill, E.D., 1912. A flora of Manila. Department of the Interior, Bureau of Science, Manila, the Philippines. 491 pp.
934. Merrill, E.D., 1918. New or noteworthy Philippine plants, XIV. *The Philippine Journal of Science* 13, sect. C, Botany: 263–333.

935. Merrill, E.D., 1923–1926. An enumeration of Philippine flowering plants. 4 volumes. Bureau of Printing, Manila, the Philippines.
936. Merrill, E.D. & Rolfe, R.A., 1908. Notes on Philippine Botany. The Philippine Journal of Science 3, sect. C, Botany: 95–127.
937. Mesfin, T., 1984. The genus *Bidens* (Compositae) in NE tropical Africa. *Symbolae Botanicae Upsalienses* 24(1): 1–138.
938. Mesfin, T., 1993. An account of *Bidens* (Compositae: Heliantheae) for Africa. *Kew Bulletin* 48: 437–516.
939. Meyer, F.G. & Walker, E.H., 1965. Flora of Japan. Smithsonian Institution, Washington D.C., United States. 1067 pp.
940. Mez, C., 1902. Myrsinaceae. In: Engler, A. (Editor): *Das Pflanzenreich*. Wilhelm Engelmann, Leipzig, Germany. Vol. 4, (fam.) 236 (Heft 9). 437 pp.
941. Mi, Z., Chen, H.S., Zhang, X.T., Shao, X.W., Li, Z. & Wu, X.M., 1995. Duck hepatitis B virus model for screening of antiviral agents from medicinal herbs. *Chinese Medical Journal* 108(9): 660–664.
942. Mia, M.D., Kabir, K.H. & Ahmed, A., 1985. Efficacy of some indigenous plant materials as repellents to *Sitophilus oryzae* on stored maize. *Bangladesh Journal of Agriculture* 10(3): 55–58.
943. Miah, M.R.U., Rahman, N.H., Sufia Begum, Islam, B.N. & Sutradhar, G.N.C., 1996. Application of leaf powders and oils as a protectant of lentil seeds against *Callosobruchus chinensis* Linn. *Bangladesh Journal of Scientific and Industrial Research* 31(3): 137–142.
944. Michaelis, K., Vostrowsky, O., Paulini, H., Zintl, R. & Knobloch, K., 1982. Das ätherische Öl aus Blüten von *Artemisia vulgaris* L. [Essential oil from flowers of *Artemisia vulgaris*]. *Zeitschrift für Naturforschung, Section C*, 37(3–4): 152–158.
945. Michaud, M.H. et al., 1993. Effect of herbicide and two plant densities on the yield of medicinal plants grown in Quebec (Canada). In: Palevitch, D., Simon, J.E. & Mathé, A. (Editors): *Raw material production, product introduction*. Proceedings of the First World Congress on Medicinal and Aromatic Plants for Human Welfare, WOCMAP. Maastricht, the Netherlands, 19–25 July 1992. *Acta Horticulturae* 331: 311–318.
946. Micu Jr., E.D., 1985. The potential use of ampalaya (*Momordica charantia* Linn.), malunggay (*Moringa oleifera* Lam.), and kolasiman (*Portulaca oleracea* Linn.) extracts in the prevention of piglet anemia. Thesis, University of the Philippines, Los Baños, College, Laguna, the Philippines. 56 pp.
947. Mihan, P.A. & Belanger, J.O., 1974. Propagation of *Belamcanda chinensis* by seed. *Plant Propagator* 20(3): 24.
948. Milhet, Y., Ferron, F. & Costes, C., 1978. Quelques résultats sur la physiologie d'*Abrus precatorius* L. [Some findings on the physiology of *Abrus precatorius*]. *Plantes Médicinales et Phytothérapie* 12(2): 151–156.
949. Milliken, W., 1997. Traditional anti-malarial medicine in Roraima, Brazil. *Economic Botany* 51(3): 212–237.
950. Milz, S. & Rimpler, H., 1979. Verbreitung von Iridoiden in der Gattung *Verbena* und einigen anderen *Verbenoideae* [Iridoids in *Verbena* and some other *Verbenoideae*]. *Zeitschrift für Naturforschung, Section C, Biosciences* 34: 319–329.

951. Mirvish, S.S., Salmasi, S., Lawson, T.A., Pour, P. & Sutherland, D., 1985. Test of catechol, tannic acid, *Bidens pilosa*, croton oil, and phorbol for co-carcinogenesis of esophageal tumors induced in rats by methyl-n-amyl-nitrosamine. *Journal of the National Cancer Institute* 74(6): 1283-1290.
952. Misra, P., Pal, N.L., Guru, P.Y., Katiyar, J.C., Srivastava, V. & Tandon, J.S., 1992. Antimalarial activity of *Andrographis paniculata* (kalmegh) against *Plasmodium berghei* NK 65 in *Mastomys natalensis*. *International Journal of Pharmacognosy* 30(4): 263-274.
953. Misra, T.N., Singh, R.S., Srivastava, R., Pandey, H.S., Prasad, C. & Singh, S.A., 1993. A new triterpenoid from *Vernonia cinerea*. *Planta Medica* 59(5): 458-460.
954. Misra, T.N., Singh, R.S., Upadhyay, J. & Tripathi, D.N.M., 1984. Aliphatic hydroxy-ketones from *Curculigo orchoides* rhizomes. *Phytochemistry* 23: 1643-1645.
955. Mitchell, J. & Rook, A., 1979. *Botanical dermatology. Plants and plant products injurious to the skin.* Greengrass, Vancouver, Canada. 787 pp.
956. Mitra, R.L. & Jain, S.K., 1985. Concept of *Phyllanthus niruri* (Euphorbiaceae) in Indian floras. *Bulletin of the Botanical Survey of India* 27(1-4): 161-176.
957. Mitsunaga, K., Koike, K., Fukuda, H., Ishii, K. & Ohmoto, T., 1991. Ligustrinoid, a new bisiridoid glucoside from *Strychnos ligustrina*. *Chemical and Pharmaceutical Bulletin* 39(10): 2737-2738.
958. Miura, Y., Fukui, H. & Tabata, M., 1988. Reduced inhomogeneity of *Angelica acutiloba* plants propagated clonally through somatic embryoids. *Planta Medica* 54(1): 79-81.
959. Miyazawa, M. & Kameoka, H., 1977. The essential oil of *Artemisia capillaris*. *Phytochemistry* 16(7): 1054-1057.
960. Mizuno, F. et al., 1986. Epstein-Barr virus-enhancing plant promoters in East Africa. *AIDS Research* 2, supplement 1: S151-S155.
961. Mogie, M., & Richards, A.J., 1983. Satellited chromosomes, systematics and phylogeny in *Taraxacum* (Asteraceae). *Plant Systematics and Evolution* 141: 219-229.
962. Mohanan, P.V. & Devi, K.S., 1997. Effect of *Sobatum* on tumour development and chemically induced carcinogenesis. *Cancer Letter* 112(2): 219-223.
963. Mohanan, P.V., Rathinam, K. & Devi, K.S., 1996. Lack of micronucleus induction by 'Sobatum' in bone marrow erythrocytes of Swiss mice. *Mutation Research* 361(1): 23-27.
964. Mohiddin, M.Y., Wong Chin & Holdsworth, D.K., 1991. Traditional medicinal plants of Brunei Darussalam. Part II. Sengkurong. *International Journal of Pharmacognosy* 29(4): 252-258.
965. Mohiddin, M.Y., Wong Chin & Holdsworth, D.K., 1992. Traditional medicinal plants of Brunei Darussalam. Part III. Sengkurong. *International Journal of Pharmacognosy* 30(2): 105-108.
966. Mohiuddin, S., Khan, M.A., Qureshi, R.A., Kapadia, Z. & Qureshi, S.A., 1990. *Ageratum houstonianum*, a plant with insectistatic potential. *Karachi University Journal of Science* 18(1-2): 159-164.
967. Mokkhasmit, M., Ngarmwathana, W., Savasdimongkol, K. & Permpiphat, U, 1971. Pharmacological evaluation of Thai medicinal plants (con-

- tinued). Journal of the Medical Association of Thailand 54(7): 490-504.
968. Moldenke, H.N., 1940. Verbenaceae. In: Pulle, A. (Editor): Flora of Suriname. Vol 4(2). Koninklijke Vereniging Koloniaal Instituut, Amsterdam, the Netherlands. Mededeeling No 30. Afdeling Handelsmuseum No 11. pp. 257-321.
969. Moldenke, H.N., 1956. Materials towards a monograph of the genus Vitex. V. Phytologia 5(8): 343-393.
970. Moldenke, H.N., 1957. Materials towards a monograph of the genus Vitex. VII. Phytologia 5(10): 465-507.
971. Moldenke, H.N., 1957. Materials towards a monograph of the genus Vitex. VIII. Phytologia 6(1): 13-64.
972. Moldenke, H.N. & Moldenke, A.L., 1983. Verbenaceae. In: Dassanayake, M.D. & Fosberg, F.R. (Editors): A revised handbook to the flora of Ceylon. Vol. 4. Amerind Publishing Co., New Delhi, India. pp. 196-487.
973. Monif, T., Malhotra, A.K. & Kapoor, V.P., 1992. Cassia fistula seed galactomannan: potential binding agent for pharmaceutical formulation. Indian Journal of Pharmaceutical Sciences 54(6): 234-240.
974. Morallo-Rejesus, B., 1986. Botanical insecticides against the diamondback moth. In: Diamondback moth management. Proceedings of the first international workshop, Tainan, Taiwan, 11-15 March, 1985. pp. 241-255.
975. Morallo-Rejesus, B., 1987. Botanical pest control research in the Philippines. Philippine Entomologist 7(1): 1-30.
976. Moreno, M. & Rodriguez, V.M., 1981. Yiamoloxide B, a fungistatic saponin of *Phytolacca octandra*. Phytochemistry 20(6): 1446-1447.
977. Morimitsu, Y. & Kawakishi, S., 1990. Inhibitors of platelet aggregation from onion. Phytochemistry 29: 3435-3439.
978. Mortada, A., Hegazy, M.A., Hegazy, M.R. & Helal, M., 1976. Use of aloe extracts in the treatment of experimental corneal ulcers. Ceskoslovenska Oftalmologie 32: 424-427.
979. Morton, J.F., 1981. Atlas of medicinal plants of Middle America. Bahamas to Yucatan. Charles C. Thomas, Springfield, Illinois, United States. 1420 pp.
980. Morton, J.F., 1987. Fruits of warm climates. J.F. Morton, Miami, United States. 505 pp.
981. Morton, J.F., 1992. Country borage (*Coleus amboinicus* Lour.): a potent flavoring and medicinal plant. Journal of Herbs, Spices and Medicinal Plants 1(1-2): 77-90.
982. Moshi, M.J., Uiso, F.C., Mahunnah, R.L.A., Malele, S.R. & Swai, A.B.M., 1997. A study of the effect of *P. amarus* extracts on blood glucose in rabbits. International Journal of Pharmacology 35(3): 167-173.
983. Mousa, O., Vuorela, P., Kiviranta, J., Wahab, S.A., Hiltunen, R. & Vuorela, H., 1994. Bioactivity of certain Egyptian *Ficus* species. Journal of Ethnopharmacology 41(1-2): 71-76.
984. Muench, E. & Kiefer, J., 1989. Die Purgiernuss (*Jatropha curcas* L.): Mehrzweckpflanze als Kraftstoffquelle der Zukunft [*Jatropha curcas* L.: a multi-purpose plant as fuel resource of the future]. Schriftenreihe der GTZ No 209. Deutsche Gesellschaft für Technische Zusammenarbeit, Eschborn, Germany. 32 pp.
985. Muhammad, H., Iskandar, M. & Pitono, J., 1996. Studi perbanyakan veg-

- etatif pada tanaman sambiloto [A study on vegetative propagation of *Andrographis paniculata* Nees plants]. *Warta Tumbuhan Obat Indonesia* 3(1): 35-37.
986. Mukherjee, P.K., Saha, K., Saha B.P., Pal, M. & Das, J., 1996. Antifungal activities of the leaf extract of *Cassia tora* Linn. (Fam. Leguminosae). *Phytotherapy Research* 10(6): 521-522.
987. Mukhopadhyay, S., Mukhopadhyay, M.J. & Sharma, A.K., 1991. In vitro multiplication and regeneration of cytologically stable plants of *Rauwolfia serpentina* Benth. through shoot tip culture. *Nucleus* 34(3): 170-173.
988. Mukhtar, A., 1989. Feeding diversity of *Myloccerus viridanus* Fab. (Coleoptera: Curculionidae) from south India. *Indian Forester* 115(11): 832-838.
989. Muller, F.M., 1978. Seedlings of the North-West European lowland. Dr W. Junk B.V. Publishers, The Hague, the Netherlands & Pudoc, Wageningen, the Netherlands. 654 pp.
990. Mulyono, E., Marwati, T. & Yuliani, S., 1994. Kencur [*Kaempferia galanga*]. *Edisi Khusus Penelitian Tanaman Rempah dan Obat* 10(2): 23-33.
991. Munir, A.A., 1984. A taxonomic revision of the genus *Premna* L. (Verbenaceae) in Australia. *Journal of the Adelaide Botanic Gardens* 7(1): 1-44.
992. Munir, A.A., 1985. A taxonomic revision of the genus *Viticipremna* H.J. Lam (Verbenaceae). *Journal of the Adelaide Botanical Garden* 7(2): 181-200.
993. Munir, A.A., 1987. A taxonomic revision of the genus *Vitex* L. Verbenaceae in Australia. *Journal of the Adelaide Botanical Garden* 10(1): 31-80.
994. Munshi, A., Mehrotra, R., Ramesh, R. & Panda, S.K., 1993. Evaluation of antihepadnavirus activity of *Phyllanthus amarus* and *Phyllanthus maderaspatensis* in duck hepatitis B virus carrier Peking ducks. *Journal of Medical Virology* 41(4): 275-281.
995. Murali, T.P. & Gowda J.V.N., 1987. Effect of cycocel and maleic hydrazide on growth and flowering in kakada (*Jasminum multiflorum* Andr.). *Indian Perfumer* 31(3): 215-218.
996. Murdiati, T. & Stoltz, D.R., 1987. Investigation of suspected plant poisoning of North Sumatran cattle. *Penyakit Hewan* 19: 101-105.
997. Muto, Y., Ichikawa, H., Kitagawa, O., Kumagai, K., Watanabe, M., Ogawa, E., Seiki, M., Shirataki, Y., Yokoe, I. & Komatsu, M., 1994. Studies on antiulcer agents. I. The effects of various methanol and aqueous extracts of crude drugs on antiulcer activity. *Yakugaku Zasshi* 114(2): 980-994. (in Japanese)
998. N'Dounga, M. Balansard, G., Babadjamian, A., David, P.T. & Gasqvét, M., 1983. Study on *Bidens pilosa* L. Identification and antiparasitic activity of 1-phenyl-1,3,5-heptatriyne. *Plantes Médicinales et Phytothérapie* 17: 64-75.
999. Nadeem, M. & Hussain, S.J., 1996. Anti-inflammatory activity of *Solanum nigrum* Linn. berries. *Hamdard Medicus* 39(4): 25-27.
1000. Nagao, Y., Fujioka, S., Tsukui, M. & Matsuoka, T., 1977. Studies on breeding and cultivation of medicinal plants. Part 2. The cultivation and the quality of *Momordica cochinchinensis*. *Journal of the Takeda Re-*

- search Laboratory 36(3-4): 193-199.
1001. Nair, S.C. & Panikkar, K.R., 1990. Antitumour principles from *Ixora javanica*. *Cancer Letters* 49(2): 121-126.
  1002. Nair, S.C., Panikkar, B., Akamanchi, K.F. & Panikkar, K.R., 1991. Inhibitory effects of *Ixora javanica* extract on skin chemical carcinogenesis in mice and its antitumour activity. *Cancer Letters* 60(3): 253-258.
  1003. Nakajima, T., Nonoto, K., Ohfuné, Y., Shiratori, Y., Takemoto, T., Takeuchi, H. & Watanabe, K., 1985. Effects of glutamic acid analogs on identifiable neurons, sensitive to beta-hydroxy-L-glutamic acid, of an African giant snail (*Achatina fulica* Ferrusac). *British Journal of Pharmacology* 13(7): 665-672.
  1004. Nakajo, S., Akabane, T., Nakaya, K., Nakamura, Y. & Kurihara, Y., 1992. An enzyme immunoassay and immunoblot analysis for curculin, a new type of taste-modifying protein. *Biochemica et Biophysica Acta* 1118: 293-297.
  1005. Nakamura, K., Tsuchiya, S., Sugimoto, Y., Sugimura, Y. & Yamada, Y., 1992. Histamine release inhibition activity of bisbenzylisoquinoline alkaloids. *Planta Medica* 58: 505-508.
  1006. Nakanishi, H., 1989. Dispersal ecology of the maritime plants in the Ryukyu Islands, Japan. *Ecological Research* 3(2): 163-173.
  1007. Nandi, P., Talukder, G. & Sharma, A., 1997. Dietary chemoprevention of clastogenic effects of 3,4-benzo(a)pyrene by *Emblica officinalis* Gaertn. fruit extract. *British Journal of Cancer* 76(10): 1279-1283.
  1008. Nano, G.M., Bicchi, C., Frattini, C. & Gallino, M., 1976. On the composition of some oils from *Artemisia vulgaris*. *Planta Medica* 30(3): 211-215.
  1009. Narayana Rao, M., Krupadanam, D.G.L. & Srimannarayana, G., 1994. Four isoflavones and two 3-aryl coumarins from the stems of *Derris scandens*. *Phytochemistry* 37: 267-269.
  1010. Narin Sombunsan & Watna Stienswat, 1983. Phon khong hormone IBA to kan koet rak nai king pakcham sabu dam [Effect of IBA on root formation of stem-cutting of Purging nut (*Jatropha curcas* L.)]. Department of Horticulture, Faculty of Agriculture, Kasetsart University, Bangkok, Thailand. 19 pp.
  1011. Naseema, A. & Wilson, K.I., 1990. New records on fungi on some medicinal plants from India. *Indian Phytopathology* 43(4): 595.
  1012. Nasir, E. & Ali, S.I. (Editors), 1970-1988. *Flora of West Pakistan*. No 1-188. Department of Botany, University of Karachi and National Herbarium, Pakistan Agricultural Research Council, Islamabad, Pakistan.
  1013. Nath, D., Sethi, N., Singh, R.K. & Jain, A.K., 1992. Commonly used Indian abortifacient plants with special reference to their teratologic effects in rats. *Journal of Ethnopharmacology* 36(2): 147-154.
  1014. Ndamba, J., Lemmich, E. & Molgaard, P., 1994. Investigation of the diurnal, ontogenetic and seasonal variation in the molluscicidal saponin content of *Phytolacca dodecandra* aqueous berry extracts. *Phytochemistry* 35(1): 95-99.
  1015. Ndamba, J., Nyazema, N.Z., Makaza, N., Anderson, C. & Kaondera, K.C., 1994. Traditional herbal remedies used for the treatment of urinary schistosomiasis in Zimbabwe. *Journal of Ethnopharmacology* 42(2): 125-132.



1016. Nee, M., 1979. Patterns in biogeography in *Solanum*, section *Acanthophora*. In: Hawkes, J.G., Lester, R.N. & Skelding, A.D. (Editors): The biology and taxonomy of the Solanaceae. Linnean Society Symposium Series 7. Academic Press, London, United Kingdom. p. 569–580.
1017. Nee, M., 1991. Synopsis of *Solanum* section *Acanthophora*: a group of interest for glycoalkaloids. In: Hawkes, J.G., Lester, R.N., Nee, M. & Estrada, N. (Editors): *Solanaceae III: Taxonomy, chemistry, evolution*. The Royal Botanic Gardens, Kew & the Linnean Society of London, London, United Kingdom. pp. 257–266.
1018. Neef, H., Cilli, F., Declerck, P.J. & Laekeman, G., 1996. Platelet anti-aggregating activity of *Taraxacum officinale* Weber. *Phytotherapy Research* 10, Supplement 1: s138–s140.
1019. Neef, H., De Clerq, P. & Laekeman, G., 1993. Hypoglycemic activity of selected European plants. *Pharmacology, World & Science* 156: H 11.
1020. Ng, F.S.P., 1991–1992. Manual of forest fruits, seeds and seedlings. 2 volumes. Malayan Forest Record No 34. Forest Research Institute Malaysia, Kepong, Malaysia. 997 pp.
1021. Ng, F.S.P., 1992. Guide to garden plants 10. *Nature Malaysiana* 17(1): 10–17.
1022. Ng, F.S.P. & Mat Asri Ngah Sanah, 1991. Germination and seedling records. Research Pamphlet No 108. Forest Research Institute Malaysia, Kepong, Malaysia. 191 pp.
1023. Ng, J., 1982. In-vitro pollen germination of *Cassia fistula* L. *Gardens' Bulletin, Singapore* 34(2): 239–242.
1024. Ng, T.B., Chan, W.Y. & Yeung, H.W., 1992. Proteins with abortifacient ribosome inactivating immunomodulatory antitumor and anti-AIDS activities from Cucurbitaceae plants. *General Pharmacology* 23(4): 575–590.
1025. Ng, T.B., Hon, W.K., Lo, L.H., Li, W.W. & Yeung, H.W., 1986. Effects of amomorpharin,  $\beta$ -momorpharin and  $\alpha$ -trichosanthin on lipogenesis and testicular and adrenal steroidogenesis in vitro and plasma-glucose levels in vivo. *Journal of Ethnopharmacology* 18(1): 45–53.
1026. Ng, T.B., Li, W.W. & Yeung, H.W., 1986. A steryl glycoside fraction with hemolytic activity from tubers of *Momordica cochinchinensis*. *Journal of Ethnopharmacology* 18(1): 55–62.
1027. Ng, T.B., Liu, W.K., Tsao, S.W. & Yeung, H.W., 1994. Effect of trochocanthin and momorpharins on isolated rat hepatocytes. *Journal of Ethnopharmacology* 43(2): 81–87.
1028. Ng, T.B., Wong, C.M., Li, W.W. & Yeung, H.W., 1985. Effect of Trichosanthin kirilowii lectin on lipolysis and lipogenesis in isolated rat and hamster adipocytes. *Journal of Ethnopharmacology* 14(1): 93–98.
1029. Nguyen Nghia Thin, 1989. Useful plants of Euphorbiaceae in the flora of Vietnam. *Forestry Revue, Hanoi* 1989: 29–30.
1030. Nguyen Nghia Thin, 1995. Euphorbiaceae of Vietnam. Agriculture Publishing House, Hanoi, Vietnam. 50 pp.
1031. Nguyen Tien Ban, 1995. The family Menispermaceae in the flora of Vietnam. *Journal of Biology* 17(4): 61–67. (in Vietnamese)
1032. Nguyen Tien Ban et al., 1990. Biological study on *Artemisia annua* L.: creating material for antimalarial medicaments. Scientific Report,

- Institute of Ecology and Biological Resources, Hanoi, Vietnam. 48 pp. (in Vietnamese)
1033. Nguyen Tien Ban, Le Kim Bien & Vu Xuan Phuong, 1990. The discovery and experimental cultivation of *Artemisia annua* L. for medicaments against malaria. In: Selected collection of scientific reports on ecology and biological resources. Science and Technics Publishing House, Hanoi, Vietnam. pp. 213–218. (in Vietnamese)
1034. Nguyen Van Dan & Pham Truong Thi Tho, 1973. Contribution to the study of *Ageratum conyzoides*. *Thong bao Duoc lieu* 20: 9–19. (in Vietnamese)
1035. Nguyen Van Duong, 1993. Medicinal plants of Vietnam, Cambodia and Laos. Mekong Printing, Santa Ana, California, United States. 528 pp.
1036. Nguyen Van Thuan, Dy Phon, P. & Niyomdhan, C., 1987. Légumineuses-Papilionoïdées [Leguminosae-Papilionoideae]. In: Lescot, M. (Editor): Flore du Cambodge, du Laos et du Viêt Nam [Flora of Cambodia, Laos and Vietnam]. Vol. 23. Muséum National d'Histoire Naturelle, Paris, France. 258 pp.
1037. Nguyen Xuan Dung & Do Tat Loi, 1991. Selection of traditional medicines for study. *Journal of Ethnopharmacology* 32(1–3): 57–70.
1038. Nguyen Xuan Dung, Do Tat Loi, Do Tat Hung & Leclercq, P.A., 1991. Chemical composition of the oil of *Blumea lanceolaria* (Roxb.) Druce from Vietnam. *Journal of Essential Oil Research* 3(4): 285–286.
1039. Nhatiuk, S.H., 1979. A survey of germination of seeds from some vascular plants found on Aldabra Atoll. *Journal of Biogeography* 6(2): 105–114.
1040. Nirdnoy, M. & Muangman, V., 1991. Effects of *Folia Orthosiphonis* on urinary stone promoters and inhibitors. *Journal of Medical Association of Thailand* 74(6): 318–321.
1041. Nirmala Mary, T. & Aruna Prabha, S., 1980. Ecology of *Aerva lanata* Juss. *Geobios* 7(6): 285–286.
1042. Nishibe, S., Tamayama, Y., Sasahara, M. & Andary, C., 1995. A phenylethanoid glycoside from *Plantago asiatica*. *Phytochemistry* 38(3): 741–743.
1043. Nishimoto, K., Ito, M., Natori, S. & Ohmoto, T., 1968. The structures of arundoin, cylindrin and fernenol. Triterpenoids of fernane and arborane groups of *Imperata cylindrica* var. *koenigii*. *Tetrahedron* 24: 735–752.
1044. Nitsch, C. & Nitsch, J.P., 1969. Floral induction in a short-day plant, *Plumbago indica* L., by 2-chloroethanephosphonic acid. *Plant Physiology* 44: 1747–1748.
1045. Niyomdham, C., 1980. Preliminary revision of tribe Sophoreae (Leguminosae-Faboideae) in Thailand: *Ormosia* Jacks. and *Sophora* Linn. *Thai Forest Bulletin, Botany* 13: 1–22.
1046. Nok, A.J., Williams, S. & Onyenekwe, P.C., 1996. *Allium sativum*-induced death of African trypanosomes. *Parasitology Research* 82(7): 634–637.
1047. Nomura, M., Hida, T., Miyamoto, K., Ohshima, T., Hayashi, H. & Sawanishi, H., 1995. Synergistic effects of inchin-ko-to (yin-chen-hao-tang) and ursodeoxycholic acid on cholestasis caused by alpha-naphthyl isothiocyanate. *Phytotherapy Research* 9(8): 563–566.
1048. Nomura, T. & Hano, J., 1994. Isoprenoid-substituted phenolic com-

- pounds of moraceous plants. Natural Products Report. pp. 205–218.
1049. Noor, H. & Ashcroft, S.J.H., 1989. Antidiabetic effects of *Tinospora crispa* in rats. *Journal of Ethnopharmacology* 27(1-2): 149–161.
1050. Noor, H., Hammonds, P. & Ashcroft, S.J.H., 1988. An aqueous extract from *Tinospora crispa* lowers blood glucose levels in alloxan-diabetic rats and stimulates insulin release in rat islets and man. *Diabetologia* 31(7): 526A–527A.
1051. Noor, H., Hammonds, P., Sutton, R. & Ashcroft, S.J.H., 1989. The hypoglycemic and insulinotropic activity of *Tinospora crispa* studies with human and rat islets and HIT-T15 B cells. *Diabetologia* 32(6): 354–359.
1052. Nooteboom, H.P., 1972. Simaroubaceae. In: van Steenis C.G.G.J. (General editor): *Flora Malesiana*. Series 1, Vol. 6. Wolters-Noordhoff, Groningen, the Netherlands. pp. 193–226.
1053. Nooteboom, H.P., 1981. Simaroubaceae. In: Smitinand, T. & Larsen, K. (Editors): *Flora of Thailand*. The Forest Herbarium, Royal Forest Department, Bangkok, Thailand. pp. 439–447.
1054. Noro, T. et al., 1983. Mono amine oxidase inhibitor from the rhizomes of *Kaempferia galanga*. *Chemical and Pharmaceutical Bulletin* 31(8): 2708–2711.
1055. Noronha, A.B., Amelia, M., Alexandre, V., De Gaetano, R. & Vicente, M., 1993. Protection against tobacco mosaic virus induced by some Caryophyllales plant extracts. *Microbios* 73(294): 75–80.
1056. Noronha, A.B., Gil, V.L., Vicente, M. & Goncalves, A.L., 1983. Occurrence of plant virus inhibitors in five species of Caryophyllales. II. *Alternanthera amoena*, *A. brasiliensis*, *A. philoxeroides*, *Iresine herbstii* and *Talinum paniculatum*. *Fitopatologia Brasileira* 8(2): 317–323.
1057. Nowicke, J.W., 1969. Palynotaxonomic study of the Phytolaccaceae. *Annals of the Missouri Botanical Garden* 55(3): 294–364.
1058. Nuntira Mekaroonkamol, 1990. Kan charoen toep to lae kan hai phonphalit khong dong dung [Growth and yield performance of gloriosa lily (*Gloriosa superba* Linn.)]. Bangkok, Thailand. 88 pp.
1059. Nurawan, A., Hadad, E.A. & Sitepu, D., 1993. Penanggulangan penyakit penting pada tanaman tempuyung dan saga manis [Control of main diseases of *Sonchus arvensis* and *Abrus precatorius*]. *Warta Tumbuhan Obat Indonesia* 2(2): 21–22.
1060. Nyazema, N.Z., Ndamba, J., Anderson, C., Makaza, N. & Kaondera, K.C., 1994. The doctrine of signatures or similitudes: a comparison of the efficacy of praziquantel and traditional herbal remedies used for the treatment of urinary schistosomiasis in Zimbabwe. *International Journal of Pharmacognosy* 32(2): 142–148.
1061. O'Neill, M.J., Bray, D.H., Boardman, P., Phillipson, J.D. & Warhurst, D.C., 1985. Plants as sources of antimalarial drugs. Part 1. In vitro test method for the evaluation of crude extracts from plants. *Planta Medica* 51(5): 394–398.
1062. Obara, Y., Matsubara, H. & Munakata, K., 1976. Isolation and identification of tubaic acid and  $\beta$ -tubaic acid from *Derris* roots. *Agricultural and Biological Chemistry* 40(6): 1245–1246.
1063. Obaseiki-Ebor, E.E., Odukoya, K., Telikeypalli, H., Mitscher, L.A. & Shankel, D.M., 1993. Antimutagenic activity of extracts of leaves of four

- common edible vegetative plants in Nigeria (West Africa). *Mutation Research* 302(2): 109–117.
1064. Obasi, B.N.B., Igboechi, C.A., Anuforo, D.C. & Aimufua, K.N., 1993. Effects of extracts of *Newbouldia laevis*, *Psidium guajava* and *Phyllanthus amarus* on gastrointestinal tract. *Fitoterapia* 64(3): 235–238.
1065. Occhiuto, F., Circosta, C. & Costa de Pasquale, R., 1989. Studies on some medicinal plants of Senegal: effects on isolated guinea pig ileum. *Journal of Ethnopharmacology* 26(2): 205–210.
1066. Ochse, J.J. & Bakhuizen van den Brink, R.C., 1980. *Vegetables of the Dutch East Indies*. 3rd English edition (translation of 'Indische groenten', 1931). Asher & Co., Amsterdam, the Netherlands. 1061 pp.
1067. Oey Djoen Seng, 1951. *De soortelijke gewichten van Indonesische houtsoorten en hun betekenis voor de praktijk* [Specific gravity of Indonesian woods and its significance for practical use]. Rapport No 46. Bosbouwproefstation, Bogor, Indonesia. 183 pp.
1068. Ogihara, K., Yamashio, R., Higa, M. & Yogi, S., 1997. Preparation of naphthoquinone derivatives from plumbagin and their ichthyotoxicity. *Chemical and Pharmaceutical Bulletin* 45(3): 437–445.
1069. Ohashi, H., 1973. *The Asiatic species of Desmodium and its allied genera (Leguminosae)*. Ginkgoana No 1. Academia Scientific Books, Tokyo, Japan. 318 pp.
1070. Ohashi, H., 1993. *Araliaceae*. In: Huang, T.-C. (Editor): *Flora of Taiwan*. 2nd Edition. Vol. 3. Editorial Committee of the Flora of Taiwan, Taipei, Taiwan, Republic of China. pp. 986–1009.
1071. Ohga, Y., Ono, M. & Furuno, K., 1989. The multiplication method of medical crops by tissue culture: Embryoid formation and plant regeneration. *Bulletin of the Fukuoka Agricultural Research Center, Series A. Crop (Japan)* 9: 75–78.
1072. Ohnuma, T., Sridhar, K.S., Ratner, L.H. & Holland, J.F., 1982. Phase I study of indicine N-oxide in patients with advanced cancer. *Cancer Treatment Report* 66(7): 1509–1515.
1073. Ohta, T., Kuhr, R.J. & Bowers, W.S., 1977. Radiosynthesis and metabolism of the insect antijuvenile hormone precocene II. *Journal of Agricultural and Food Chemistry* 25(3): 478–481.
1074. Ohyama, K., Misawa, N., Yamano, Y. & Komano, T., 1984. Protoplast isolation from *Euphorbia tirucalli* L. cell suspension cultures and sustained cell division. *Zeitschrift für Pflanzenphysiologie* 113(4): 367–370.
1075. Okada, Y., Miyauchi, N., Suzuki, K., Kobayashi, T., Tsutsui, C., Mayuzumi, K., Nishibe, S. & Okuyama, T., 1995. Search for naturally occurring substances to prevent the complications of diabetes. II. Inhibitory effect of coumarin and flavonoid derivatives on bovine lens aldose reductase and rabbit platelet aggregation. *Chemical and Pharmaceutical Bulletin* 43(8): 1385–1387.
1076. Okano, M., Fukamiya, N., Aratani, T., Juichi, M. & Lee, K.H., 1985. Antitumor agents, 74. Bruceanol-A and -B, two new antileukemic quassinoids from *Brucea antidysenterica*. *Journal of Natural Products* 48(6): 972–975.
1077. Okano, M., Fukamiya, N., Toyota, T., Tagahara, K. & Lee, K.H., 1989. Antitumor agents, 104. Isolation of yadanziosides M and P from *Brucea*

- antidysenterica and identification of bruceantinoside B as a mixture of yadanzioside P and bruceantinoside C. *Journal of Natural Products* 52(2): 398–401.
1078. Okano, M., Lee, K.H. & Hall, I.H., 1981. Antitumor agents. 39. Bruceantinoside-A and -B, novel antileukemic quassinoid glucosides from *Brucea antidysenterica*. *Journal of Natural Products* 44(4): 470–474.
1079. Okigawa, M., Maeda, T. & Kawano, N., 1970. The isolation and structure of 3 new lignans from *Justicia procumbens* var. *leucantha*. *Tetrahedron* 26(18): 4301–4305.
1080. Okuyama, E., Gao, L.H. & Yamazaki, M., 1990. Studies on pharmacologically active principles from Indonesian crude drugs. III. Toxic components from *Brucea javanica* (L.) Merr. *Yakugaku Zasshi* 110(11): 834–838. (in Japanese)
1081. Okuyama, T., Fujita, K., Shibata, S., Hoson, M., Kawada, T., Masaki, M. & Yamate, N., 1989. Effects of Chinese drugs 'xiebai' and 'dasuan' on human platelet aggregation (*Allium bakeri*, *A. sativum*). *Planta Medica* 55(3): 242–244.
1082. Okuyama, T., Shibata, S., Hoson, M., Kawada, T., Osada, H. & Noguchi, T., 1986. Effect of oriental plant drugs on platelet aggregation. III. Effect of Chinese drug 'xiebai' on human platelet aggregation. *Planta Medica* 52(3): 171–175.
1083. Oladiran, A.O. & Gana, R.W., 1991. Leaf spot of *Alternanthera sessilis* and its implications for food crops. *Journal of Phytopathology* 133(2): 169–174.
1084. Oldfield, S. (Compiler), 1997. Cactus and Succulent Plants – status survey and conservation action plan. IUCN/SSC Cactus and Succulent Specialist Group, IUW, Gland, Switzerland & Cambridge, United Kingdom. 222 pp.
1085. Oliver-Bever, B., 1986. Medicinal plants in tropical West Africa. Cambridge University Press, Cambridge, United Kingdom. 375 pp.
1086. Omeregbe, R.E., Ikuebe, O.M. & Ihimire, I.G., 1996. Antimicrobial activity of some medicinal plants extracts on *Escherichia coli*, *Salmonella paratyphi* and *Shigella dysenteriae*. *African Journal of Medicine and Medical Science* 25(4): 373–375.
1087. Omta, S.W.F. & Fortuin, F.T.J.M., 1978. The cultivation of *Solanum nigrum* L. as a leaf and fruit vegetable in the home gardens of West Java. Mimeographed research report. Institute of Ecology, Pajajaran University, Bandung, Indonesia & Department of Plant Physiology, State University, Groningen, the Netherlands. 70 pp.
1088. Omulokoli, E., Khan, B. & Chabra, S.C., 1997. Antiplasmodial activity of four Kenyan medicinal plants. *Journal of Ethnopharmacology* 56: 133–137.
1089. O'Neill, M.J. et al., 1998. Isolation of translactone-containing triterpenes with thrombin inhibitory activities from the leaves of *Lantana camara*. *Journal of Natural Products* 61(11): 1328–1331.
1090. Ong, H.C., 1990. Additional medicinal uses of *Ageratum conyzoides* L. *Wallaceana* 62–63: 4–5.
1091. Otsuka, H., Kashima, N., Hayashi, T., Kubo, N., Yamasaki, K. & Padolina, W.G., 1992. Premnaodorosides A, B and C, iridoid glucoside di-

- esters of an acyclic monoterpenediol from leaves of *Premna odorata*. *Phytochemistry* 31(9): 3129–3133.
1092. Otsuka, H., Kubo, N., Yamasaki, K. & Padolina, W.G., 1989. Premnosides A–D: diacyl 6-O- $\alpha$ -L-rhamnopyranosylcatalpols from *Premna odorata*. *Phytochemistry* 28(11): 3063–3067.
1093. Otsuka, H., Kubo, N., Yamasaki, K. & Padolina, W.G., 1989. Two iridoid glycoside caffeoyl esters from *Premna odorata*. *Phytochemistry* 28: 513–515.
1094. Otsuka, H., Yoshimura, K., Yamasaki, K. & Cantoria, M.C., 1991. Isolation of 10-O-acyl iridoid glucosides from a Philippine medicinal plant, *Oldenlandia corymbosa* L. (Rubiaceae). *Chemical and Pharmaceutical Bulletin* 39(8): 2049–2052.
1095. Ott, M., Thyagarajan, S.P. & Gupta, S., 1997. *Phyllanthus amarus* suppresses hepatitis B virus by interrupting interactions between HBV enhancer I and cellular transcription factors. *European Journal of Clinical Investigation* 27(11): 908–915.
1096. Ouyang, Y., Koike, K. & Ohmoto, T. 1994. Canthin-6-one alkaloids from *Brucea mollis* var. *tonkinensis*. *Phytochemistry* 36(6): 1543–1546.
1097. Ozaki, Y., 1990. Anti-inflammatory effect of *Curcuma xanthorrhiza* Roxb. and its active principles. *Chemical and Pharmaceutical Bulletin* 38(4): 1045–1048.
1098. Ozaki, Y. & Liang, O.B., 1988. Cholagogic action of the essential oil obtained from *Curcuma xanthorrhiza* Roxb. *Shoyakugaku Zasshi* 42(4): 257–263. (in Japanese)
1099. Ozaki, Y., Xing, L. & Satake, M., 1996. Antiinflammatory effect of *Trichosanthes kirilowii* Maxim. and its effective parts. *Biological and Pharmaceutical Bulletin* 19(8): 1046–1048.
1100. Pachaly, P. & Adnan, A.Z., 1989. The structure of tinocrisposide, a bitter furanoditerpene glucoside from *Tinospora crispa*. *Planta Medica* 55(7): 632.
1101. Pachaly, P. & Adnan, A.Z., 1992. Tinocrisposide, a new bitter furanoditerpene glycoside from *Tinospora crispa* Miers. *Archiv der Pharmazie (Weinheim)* 325(11): 705–708.
1102. Pachaly, P., Adnan, A.Z. & Will, G., 1989. N-formyl and N-acetylporphine alkaloids of *Tinospora crispa*. *Planta Medica* 55(1): 115–116.
1103. Pachaly, P., Adnan, A.Z. & Will, G., 1992. NMR assignments of N-acetylporphine alkaloids from *Tinospora crispa*. *Planta Medica* 58(2): 184–187.
1104. Pajmans, K. (Editor), 1976. *New Guinea vegetation*. Elsevier Scientific Publishing Company, Amsterdam, the Netherlands. 209 pp.
1105. Palanichamy, S. & Nagarajan, S., 1990. Analgesic activity of *Cassia alata* leaf extract and kaempferol-3-O-sophoroside. *Journal of Ethnopharmacology* 29(1): 73–78.
1106. Pan, W.D., Li, Y.J., Mai, L.T., Ohtani, K., Kasai, R. & Tanaka, O., 1992. Studies on chemical constituents of the roots of *Lantana camara*. *Yao Hsueh Hsueh Pao* 27(7): 515–521. (in Chinese)
1107. Pan, W.D., Li, Y.J., Mai, L.T., Ohtani, K.H., Kasai, R.T., Tanaka, O. & Yu, D.Q., 1993. Studies on triterpenoid constituents of the roots of *Lantana camara*. *Yao Hsueh Hsueh Pao* 28(1): 40–44. (in Chinese)

1108. Pan, W.D., Mai, L.T., Li, Y.J., Xu, X.L. & Yu, D.Q., 1993. Studies on the chemical constituents of the leaves of *Lantana camara*. Yao Hsueh Hsueh Pao 28(1): 35–39. (in Chinese)
1109. Panda, A.K., Bisaria, V.S. & Mishra, S., 1992. Alkaloid production by plant cell cultures of *Holarrhena antidysenterica*. II. Effect of precursor feeding and cultivation. Biotechnology and Bioengineering 39(10): 1052–1057.
1110. Panda, A.K., Bisaria, V.S., Mishra, S. & Bhojwani, S.S., 1991. Cell culture of *Holarrhena antidysenterica*: growth and alkaloid production. Phytochemistry 30(3): 833–836.
1111. Panda, A.K., Mishra, S. & Bisaria, V.S., 1992. Alkaloid production by plant cell suspension culture of *Holarrhena antidysenterica*. I. Effect of major nutrients. Biotechnology and Bioengineering 39(10): 1043–1051.
1112. Pandii, C. et al., 1993. Insecticidal constituents from four species of the Zingiberaceae. Phytochemistry 34(2): 415–419.
1113. Panigrahi, G., 1987. *Echites antidysenterica* (L.) Roxb. ex Fleming and *Holarrhena antidysenterica* (L.) Wall., validly published synonyms of *Wallida antidysenterica*. Taxon 36: 464–467.
1114. Park, C.-H. & Martinez, B.C., 1992. Enhanced release of rosmarinic acid from *Coleus blumei* permeabilized by dimethyl sulfoxide (DMSO) while preserving cell viability and growth. Biotechnology and Bioengineering 40(4): 459–464.
1115. Patel, G.N., 1986. Ayurveda: The traditional medicine in India. In: Steiner, R.P. (Editor): Folk medicine, the art and science. American Chemical Society, Washington D.C., United States.
1116. Patena, L.F., Pelegrina, W.R. & Barba, R.C., 1992. Rotenone production in in vitro culture of *Derris elliptica*. Philippine Journal of Crop Science 17, Suppl. No 1: S51.
1117. Pathak, R.K., Singh, I.S. & Dwivedi, R., 1987. You can grow aonla on usar soils. Indian Farming 37(2): 9–11.
1118. Paton, A., 1990. The phytogeography of *Scutellaria* L. Notes from the Royal Botanic Garden Edinburgh 46: 345–359.
1119. Payens, J.P.D.W., 1958. Erythroxylaceae. In: van Steenis, C.G.G.J. (General editor). Flora Malesiana. Series 1, Vol. 5(4). Noordhoff-Kolff N.V., Djakarta, Indonesia. pp. 543–552.
1120. Pedley, L. & Rudd, V.E., 1996. Fabaceae (Leguminosae) subfamily Faboideae (Papilionoideae) tribe Desmodieae. In: Dassanayake, M.D. & Clayton, W. D. (Editors): A revised handbook to the flora of Ceylon. Vol. 10. A.A. Balkema, Rotterdam, the Netherlands. pp. 149–198.
1121. Pei, Y., Zhang, Z.S., Xia, Y.X. & Song, S.Q., 1993. Purification of chitinase from *Momordica charantia* L. and its properties. Acta Botanica Sinica 35(6): 486–489.
1122. Pellegrin, F., 1930. Plumbaginacées [Plumbaginaceae]. In: Gagnepain, F. (Editor): Flore générale de l'Indo-Chine [General flora of Indo-China]. Vol. 3. Masson & Cie, Paris, France. pp. 748–753.
1123. Peng, J.-P., Yao, X.-S., Tezuka, Y. & Kikuchi, T., 1996. Furostanol glycosides from bulbs of *Allium chinense*. Phytochemistry 41(1): 283–285.
1124. Perez, R.M., Ocegueda, A., Munoz, J.L., Avila, J.G. & Morrow, W.W., 1984. A study of the hypoglycemic effect of some Mexican plants. Journal

- of Ethnopharmacology 12(3): 253–262.
1125. Perez-Miranda, M., Gomez-Cedenilla, A., Leon-Colombo, T., Pajares, J. & Mate-Jimenez, J., 1996. Effect of fiber supplements on internal bleeding hemorrhoids. *Hepato-Gastroenterology* 43(12): 1504–1507.
1126. Perry, L.M., 1980. Medicinal plants of East and Southeast Asia. Attributed properties and uses. MIT Press, Cambridge, Massachusetts, United States & London, United Kingdom. 620 pp.
1127. Pervykh, L.N., Karasartov, B.S. & Zapesochnaya, G.G., 1992. A study of the herb *Aerva lanata* IV. Flavonoid glycosides. *Chemistry of Natural Compounds* 28(5): 509–510.
1128. Pételot, A., 1952–1954. Les plantes médicinales du Cambodge, du Laos et du Vietnam [The medicinal plants of Cambodia, Laos and Vietnam]. 4 volumes. Centre National de Recherches Scientifiques et Techniques, Saigon, Vietnam.
1129. Petersen, M., Szabo, E., Meinhard, J., Karwatzki, B., Gertlowski, C., Kempin, B. & Fuss, E., 1995. Biosynthesis and accumulation of rosmarinic acid in suspension cultures of *Coleus blumei*. *Plant Cell, Tissue and Organ Culture* 43(2): 89–92.
1130. Pham Hoang Ho, 1991–1993. An illustrated flora of Vietnam. 3 volumes. Mekong Publisher, Montreal, Canada.
1131. Pham Truong Thi Tho & Nguyen Van Dan, 1976. Contribution to the study of *Ageratum conyzoides*. Second Communication: ageratochromene and dimethoxy-ageratochromene – the two main compounds of *A. conyzoides* essential oil. *Thong bao Duoc lieu* 8(3): 107–114. (in Vietnamese)
1132. Pharadai, K. et al., 1985. (–)-O-Acetylsukhodianine and oxostephanosine: two new aporphinoids from *Stephania venosa*. *Journal of Natural Products* 48: 658–659.
1133. Phengklai, C., 1975. Studies in flora of Thailand. *Thai Forest Bulletin (Botany)* 9: 1–11.
1134. Philcox, D., 1997. Cucurbitaceae. In: Dassanayake, M.D. & Clayton, W.D. (Editors): A revised handbook to the flora of Ceylon. Vol. 11. A.A. Balkema, Rotterdam, the Netherlands. pp. 8–46.
1135. Philcox, D., 1997. Euphorbiaceae. In: Dassanayake, M.D. & Clayton, W.D. (Editors): A revised handbook to the flora of Ceylon. Vol. 11. A.A. Balkema, Rotterdam, the Netherlands. pp. 80–283.
1136. Philipson, W.R., 1978. Araliaceae: growth forms and shoot morphology. In: Tomlinson, P.B. & Zimmermann, M.H. (Editors): Tropical trees as living systems. Proceedings of the Fourth Cabot Symposium held at Harvard Forest, Petersham, Massachusetts on April 26–30, 1976. Cambridge University Press, Cambridge, United Kingdom. pp. 269–284.
1137. Phuphathanaphong, L., 1987. Aristolochiaceae. In: Smitinand, T. & Larsen, K. (Editors): Flora of Thailand. Vol. 5, Part 1. The Forest Herbarium, Royal Forest Department, Bangkok, Thailand. pp. 1–31.
1138. Pillai, S.N. & Desai, M.V., 1975. Anthelmintic property of 'Marotti' cake (*Hydnocarpus laurifolia*). *Pesticides (India)* 9(4): 37–39.
1139. Pino, J.A., Garcia, J. & Martinez, M.A., 1996. Comparative chemical composition of the volatiles of *Coleus aromaticus* by steam distillation, solvent extraction and supercritical carbon dioxide extraction. *Journal of Essential Oil Research* 8(4): 373–375.



1140. Pitard, J., 1930. Myrsinacées [Myrsinaceae]. In: Gagnepain, F. (Editor): Flore générale de L'Indo-Chine [General flora of Indo-China]. Vol. 3(6). Masson & Cie, Paris, France. pp. 765–877.
1141. Pitono, J., Januwati, M. & Ngadimin, 1996. Pengaruh naungan terhadap pertumbuhan dan produksi sambiloto. [The effect of shading on the growth and yield of *Andrographis paniculata* Nees]. *Warta Tumbuhan Obat Indonesia* 3(1): 39–40.
1142. Pittler, M.H. & Ernst, E., 1998. Peppermint oil for irritable bowel syndrome: a critical review and metaanalysis. *American Journal of Gastroenterology* 93(7): 1131–1135.
1143. Platel, K. & Srinivasan, K., 1995. Effect of dietary intake of freeze dried bitter gourd (*Momordica charantia*) in streptozotocin induced diabetic rats. *Nahrung* 39(4): 262–268.
1144. Platel, K. & Srinivasan, K., 1997. Plant foods in the management of Diabetes mellitus: vegetables as potential hypoglycaemic agents. *Nahrung* 41(2): 68–74.
1145. Plowman, T., 1982. The identification of coca (*Erythroxylum* species): 1860–1910. *Botanical Journal of the Linnean Society* 84: 329–353.
1146. Plowman, T. & Rivier, L., 1983. Cocaine and cinnamoylcocaine content of *Erythroxylum* species. *Annals of Botany* 51: 641–659.
1147. Plowman, T., Rudenberg, L. & Greene, C.W., 1978. Chromosome numbers in neotropical *Erythroxylum* (*Erythroxylaceae*). *Botanical Museum Leaflets* 26(5): 203–209.
1148. Poli, A., Nicolau, M., Simoes, C.M.O., Nicolau, R.M.R. & Zanin, M., 1992. Preliminary pharmacologic evaluation of crude whole plant extracts of *Elephantopus scaber*. Part I: in vivo studies. *Journal of Ethnopharmacology* 37(1): 71–76.
1149. Pollack, Y., Segal, R. & Golenser, J., 1990. The effect of ascaridole on the in vitro development of *Plasmodium falciparum*. *Parasitology Research* 76: 570–572.
1150. Polya, G.M., Wang, B.H. & Foo, L.Y., 1995. Inhibition of signal-regulated protein kinases by plant-derived hydrolysable tannins. *Phytochemistry* 38(2): 307–314.
1151. Ponce-Macotela, M., Navarro-Alegria, I., Martinez-Gordillo, M.N. & Alvarez-Chacon, R., 1994. Efecto anti-giardiasico in vitro de 14 extractos de plantas [Antigiardiasic activity of 14 plant extracts in vitro]. *Revista de Investigacion Clinica* 46(5): 343–347.
1152. Popham, S., 1987. *Ficus religiosa* L. *The Bulletin - Pacific Tropical Botanical Garden* 17(1): 42–45.
1153. Porro, G., Bolognesi, A., Caretto, P., Cromo, G., Lento, P., Mistza, G., Sciumbata, T., Stirpe, F. & Modena, D., 1993. In vitro and in vivo properties of an anti-CD5-momordin immunotoxin on normal and neoplastic T lymphocytes. *Cancer Immunology and Immunotherapy* 36(5): 346–350.
1154. Prakash, A. & Rao, J., 1997. Botanical pesticides in agriculture. CRC Press, Boca Raton, United States. 461 pp.
1155. Prakash, A., Satyan, K.S., Wahi, S.P. & Singh, R.P., 1995. Comparative hepatoprotective activity of three *Phyllanthus* species, *P. urinaria*, *P. niruri* and *P. simplex*, on carbon tetrachloride induced liver injury in the rat. *Phytotherapy Research* 9(8): 594–596.

1156. Prakash, A.O., 1976. Antifertility investigation on embelin. *Planta Medica* 41: 259-266.
1157. Prana, M.S., 1983. Pembungaan beberapa jenis temu (*Curcuma* spp.) [Flowering of some species of temu (*Curcuma* spp.)]. *Pemberitaan Lembaga Penelitian Tanaman Industri* 8(4-6): 33-36.
1158. Pranata, R.I., 1984. Possibility of using turmeric (*Curcuma longa* L.) for controlling storage insects. *Biotrop Newsletter* 45(3): 3.
1159. Prasad, V.S., Devi, P.U., Rao, B.S. & Kamath, R., 1996. Radiosensitizing effect of plumbagin on mouse melanoma cells grown in vitro. *Indian Journal of Experimental Biology* 34(9): 857-858.
1160. Prendergast, H.D.V., Etkin, N.L., Harris, D.R. & Houghton, P.J. (Editors), 1998. *Plants for food and medicine. Proceedings of the joint conference of the Society for Economic Botany and the International Society for Ethnopharmacology, London, 1-6 July 1996.* Royal Botanic Gardens, Kew, United Kingdom. 438 pp.
1161. Prévost, M.-F., 1978. Modular construction and its distribution in tropical woody plants. In: Tomlinson, P.B. & Zimmermann, M.H. (Editors): *Tropical trees as living systems. The proceedings of the Fourth Cabot Symposium held at Harvard Forest, Petersham, Massachusetts on April 26-30, 1976.* Cambridge University Press, Cambridge, United Kingdom. pp. 223-231.
1162. Protacio, D.B., 1977. Evidence of cadang-cadang disease as coming from the weed *Elephantopus mollis*. *Philippine Phytopathology* 13: 1-2.
1163. Prucksumand, C., Indrasukesri, B., Leethochawalit, H., Nilvises, N., Prijavudhi, A. & Wimolwattanapun, S., 1986. Effect of the long turmeric (*Curcuma longa* Linn.) on healing of peptic ulcer: A preliminary report of 10 cases study. *Thai Journal of Pharmacology* 8(3): 139-151.
1164. Pu, Z., Lu, B.Y., Liu, W.Y. & Jin, S.W., 1996. Characterization of the enzymatic mechanism of gamma-momorcharin, a novel ribosome-inactivating protein with lower molecular weight of 11 500 purified from the seeds of bitter melon (*Momordica charantia*). *Biochemical and Biophysical Research Communications* 229(1): 287-294.
1165. Pui-Hay But, P., 1987. Tropical plants used in Chinese medicine: potential leads for pharmaceutical developments. In: Leeuwenberg, A.J.M. (Compiler): *Medicinal and poisonous plants of the tropics. Proceedings of Symposium 5-35 of the 14th International Botanical Congress, Berlin, 24 July - 1 August 1987.* Pudoc, Wageningen, the Netherlands. pp. 24-35.
1166. Puri, A., Saxena, R., Saxena, R.P., Saxena, K.C., Srivastava, V. & Tandon, J.S., 1994. Immunostimulant agents from *Andrographis paniculata*. *Journal of Natural Products* 56(7): 995-999.
1167. Purseglove, J.W., 1968-1972. *Tropical crops.* Longman, London. United Kingdom. Dicotyledons. 2 volumes (1968), 719 pp. Monocotyledons. 2 volumes (1972), 607 pp.
1168. Purushothaman, K.K., Chandrasekharan, S. & Balakrishna, K., 1975. Gangeticin and desmodin, two minor pterocarpanoids of *Desmodium gangeticum*. *Phytochemistry* 14: 1129-1130.
1169. Purwanto, Y. & Poerba, Y.S., 1990. Effects of drying, temperature and storage time on *Amaranthus spinosus*, *A. blitum* and *A. gracilis* seeds.

- Biotrop Special Publication No 38. SEAMEO Regional Center for Tropical Biology, Bogor, Indonesia. pp. 85-93.
1170. Purwantoro, R.S. & Roemantyo, 1993. Konservasi tempuyung dan saga manis – suatu studi Kebun Raya Bogor [Conservation of *Sonchus arvensis* and *Abrus precatorius* in the Bogor Botanical Gardens – a study]. *Warta Tumbuhan Obat Indonesia* 2(2): 18-20.
1171. Pushpalatha, E. & Muthukrishnan, J., 1995. Larvicidal activity of a few plant extracts against *Culex quinquefasciatus* and *Anopheles stephensi*. *Indian Journal of Malariology* 32(1): 14-23.
1172. Qing Yu, 1993. Comparison of the antinematode activity of compounds isolated from *Cylindrocarpon olidum* and *Cannabis sativa*. PhD thesis, Katholieke Universiteit Leuven, Belgium. 112 pp.
1173. Qiu, D.Y., Zhu, G. & Zhu, Z.Q., 1996. Study on production of trichosanthin from the hairy roots of *Trichosanthes kirilowii* Maxim. *Acta Botanica Sinica* 38(6): 439-443.
1174. Qu, C.F., Yang, Z.Q. & Xiang, J.M., 1993. *Alternanthera philoxeroides* (Mavt.) Griseb. protection against fetal epidemic hemorrhagic fever virus infection in suckling mice. *Chung Kuo Chung Yao Tsa Chih* 18: 304-305, 320.
1175. Quijano, L., Calderon, J.S., Gomez, F. & Rios, T., 1982. Two polymethoxyflavones from *Ageratum houstonianum*. *Phytochemistry* 21(12): 2965-2967.
1176. Quintana, E.G., Saludez, J.D., Batoon, M.P. & Generalao, M.L., 1982. Agricultural production of selected medicinal plants: propagation to postharvest handling. PCARRD [Philippine Council for Agriculture, Forestry, and Natural Resources Research and Development] Monitor 10(4): 8-10.
1177. Quirk, M.F., Bushell, J.J., Jones, R.J., Megarrity, R.G. & Butler, K.L., 1988. Live-weight gains on leucaena and native grass pastures after dosing cattle with rumen bacteria capable of degrading DHP, a ruminal metabolite from leucaena. *Journal of Agricultural Science (Cambridge)* 111: 165-170.
1178. Quisumbing, E., 1978. Medicinal plants of the Philippines. Katha Publishing Co., Quezon City, the Philippines. 1262 pp.
1179. Raadts, E., 1977. The genus *Kalanchoe* (Crassulaceae) in tropical East Africa. *Willdenowia* 8: 101-157.
1180. Racchi, M.L., 1988. Using in vitro culture to study the biosynthesis of secondary products in *Aloe ferox*. *Rivista di Agricoltura Subtropicale e Tropicale* 82(4): 707-714.
1181. Radcliffe-Smith, A., 1972. *Euphorbia* L. In: Airy Shaw, H. K. The Euphorbiaceae of Siam. *Kew Bulletin* 26: 261-268.
1182. Radcliffe-Smith, A., 1975. *Euphorbia* L. In: Airy Shaw, H. K. The Euphorbiaceae of Borneo. *Kew Bulletin Additional Series IV*. Her Majesty's Stationary Office, London, United Kingdom. pp. 10-112.
1183. Radcliffe-Smith, A., 1980. *Euphorbia* L. In: Airy Shaw, H.K. The Euphorbiaceae of New Guinea. *Kew Bulletin Additional Series VIII*. Her Majesty's Stationary Office, London, United Kingdom. pp. 81-88.
1184. Radcliffe-Smith, A., 1981. *Euphorbia* L. In: Airy Shaw, H.K. The Euphorbiaceae of Sumatra. *Kew Bulletin* 36: 294-296.

1185. Radcliffe-Smith, A., 1981. *Euphorbia* L. In: Airy Shaw, H.K. An alphabetical enumeration of the Euphorbiaceae of the Philippine Islands. Royal Botanic Gardens, Kew, United Kingdom. pp. 24–26.
1186. Radcliffe-Smith, A., 1982. *Euphorbia* L. In: Airy Shaw, H. K. The Euphorbiaceae of Central Malesia. *Kew Bulletin* 37: 18–20.
1187. Radcliffe-Smith, A., 1987. Euphorbiaceae (Part 1). In: Polhill, R. (Editor): *Flora of Tropical East Africa*. A.A. Balkema, Rotterdam, the Netherlands & Boston, United States. pp. 1–407.
1188. Raghava Reddy, J. & Purnachandra Reddi, A., 1980. Powdery mildews on plants of Cappariaceae and Euphorbiaceae. *Acta Botanica Indica* 8: 87–90.
1189. Raghunathan, K., Hariharan, V. & Rangaswami, S., 1974. Chrysophanol-1- $\beta$ -gentiobioside, a new anthraquinone glycoside from *Cassia tora* Linn. *Indian Journal of Chemistry* 12: 1251–1253.
1190. Rahmani, M., Yin, L.T. & Lajis, N.H., 1989. Toxicity studies of plant extracts on two species of fish. *Pertanika* 12(2): 189–192.
1191. Rai, S.N., Nagaveni, H.C. & Ananth Padmanabha, H.S., 1988. Germination and nursery technique of four species of *Ficus*. *Indian Forester* 114(2): 63–68.
1192. Raina, M.K., 1982. *Aloe*. In: Atal, C.K. & Kapur, B.M. (Editors): *Cultivation and utilization of medicinal plants*. Regional Research Laboratory, Council of Scientific and Industrial Research, Jammu-Tawi, India. pp. 368–374.
1193. Raja Reddy, K., 1988. Folk medicine from Chittoor District, Andhra Pradesh, India, used in the treatment of jaundice. *International Journal of Crude Drug Research* 26(3): 137–140.
1194. Rajarama Rao, M.R. & Siddiqui, H.H., 1964. Pharmacological studies on *Emblca officinalis* Gaertn. *Indian Journal of Experimental Biology* 2: 29–31.
1195. Rajeswari, E. & Mariappan, V., 1992. Effect of plant extracts on in vitro growth of rice blast (Bl) pathogen *Pyricularia oryzae*. *International Rice Research Newsletter* 17(6): 24.
1196. Ramos Ruiz, A.R., De la Torre, R.A., Alonso, N., Villaescusa, A., Betancourt, J. & Vizoso, A., 1996. Screening of medicinal plants for induction of somatic segregation activity in *Aspergillus nidulans*. *Journal of Ethnopharmacology* 52(3): 123–127.
1197. Randell, B.R., 1995. Taxonomy and evolution of *Senna obtusifolia* and *S. tora*. *Journal of the Adelaide Botanic Gardens* 16: 55–58.
1198. Randeria, A.J., 1960. The composite genus *Blumea*, a taxonomic revision. *Blumea* 10: 176–317.
1199. Rao, C.B., Rao, T.N. & Muralikrishna, B., 1977. Flavonoids from *Blumea lacera*. *Planta Medica* 31(3): 235–237.
1200. Rao, J.V., Sreenivasan, C. & Makkapati, A.K., 1996. Plumbagin effect on growth and metamorphosis of housefly *Musca domestica* L. (Diptera: Muscidae). *International Pest Control* 38: 24–27.
1201. Rao, M.V., Shah, K.D. & Rajani, M., 1997. Contraceptive effects of *Phyllanthus amarus* extract in the male mouse (*Mus musculus*) *Phytotherapy Research* 11(8): 594–596.
1202. Rao, S.G., Hrishikeshavan, H.J., Prasad, K.V.R.S.G. & Guruswami,

- M.N., 1985. Evaluation of an experimental model for studying urolithiasis effect of *Aerva lanata* on urinary stones. *Indian Drugs* 22(12): 640–643.
1203. Rao, V.G., Alaka Pande & Patwardhan, P.G., 1989. Three new records of fungal diseases of economic plants in Maharashtra State. *Biovigyanam* 15(1): 51–53.
1204. Rao, Y.S., Israel, P. & Biswas, H., 1970. Weed and rotation crop plants as hosts for the rice root-knot nematode, *Meloidogyne graminicola* (Golden and Birchfield). *Oryza* 7(2): 137–142.
1205. Rasco, S.M., 1993. In vitro culture of derris (*Derris elliptica* (Wall.) Benth.). University of the Philippines at Los Baños, College, Laguna, the Philippines. 97 pp.
1206. Ratnasooriya, W.D., Amarasekera, A.S., Perera, N.S.D. & Premakumara, G.A.S., 1991. Sperm antimotility properties of a seed extract of *Abrus precatorius*. *Journal of Ethnopharmacology* 33(1–2): 85–90.
1207. Raves, M.L., Harel, M., Pang, Y.P., Silman, I., Kozikowski, A.P. & Sussman, J.L., 1997. Structure of acetylcholinesterase complexed with the nootropic alkaloid, (-)-huperzine A. *Nature Structural Biology* 4(1): 57–63.
1208. Ravn, H., Nishibe, S., Sasahara, M. & Xuebo, L., 1990. Phenolic compounds from *Plantago asiatica*. *Phytochemistry* 29(11): 3627–3631.
1209. Ray, A.B., Chattopadhyay, S., Tripathi, R.M., Gambhir, S.S. & Das, P.K., 1979. Isolation and pharmacological action of epistephanine, an alkaloid of *Stephania hernandifolia*. *Planta Medica* 35(2): 167–173.
1210. Ray, P.G. & Majumdar, S.K., 1976. Antimicrobial activity of some Indian plants. *Economic Botany* 30: 317–320.
1211. Rehm, S. (Editor), 1989. *Handbuch der Landwirtschaft und Ernährung in den Entwicklungsländern. Band 4. Spezieller Pflanzenbau in den Tropen und Subtropen* [Handbook of agriculture and nutrition in developing countries. Vol. 4. Special crop cultivation in the tropics and subtropics]. 2nd Edition. E. Ulmer, Stuttgart, Germany. 653 pp.
1212. Rehm, S. & Espig, G., 1991. *The cultivated plants of the tropics and subtropics. Cultivation, economic value, utilization*. Technical Centre for Agriculture and Rural Co-operation (CTA), Ede, the Netherlands & Verlag Josef Margraf, Weikersheim, Germany. 552 pp.
1213. Rejesus, B.M. & Tantengco, G.B., 1986. Biological activity of flower extracts as insecticides. *National Science and Technology Authority Technology Journal (Philippines)* 11(1): 37–46.
1214. Reyes, A.I. & Ferreras, C.M., 1989. Dermatitis property of makahiya plant (*Mimosa pudica* Linn.). BSc thesis, Manila Central University, the Philippines.
1215. Reyes, L.J., 1938. *Philippine woods. Technical Bulletin No 7. Commonwealth of the Philippines, Department of Agriculture and Commerce. Bureau of Printing, Manila, the Philippines. 536 pp. + 88 plates.*
1216. Reyes, M.E.C., Gildemacher, B.H. & Jansen, G.J., 1993. *Momordica L.* In: Siemonsma, J.S. & Kasem Piluek (Editors): *Plant Resources of South-East Asia No 8. Vegetables*. Pudoc Scientific Publishers, Wageningen, the Netherlands. pp. 206–210.
1217. Reynaud, J., Couble, A. & Reynaud, J., 1989. *Les flavonoides de Verbena*

- officinalis L. (Verbénacées) [The flavonoids of *Verbena officinalis* L. (Verbenaceae)]. *Journal of Plant Physiology* 135: 380–381.
1218. Reynolds, G.W., 1966. The Aloes of Tropical Africa and Madagascar. The Trustees the Aloe Book Fund, Swaziland. 537 pp.
1219. Reynolds, G.W., 1982. The Aloes of South Africa, 4th Edition. A.A. Balkema, Rotterdam, the Netherlands, 538 pp.
1220. Reynolds, J.E.F. (Editor), 1989. Martindale. The extra Pharmacopoeia, 29<sup>th</sup> Edition. The Pharmaceutical Press, London, United Kingdom. 1896 pp.
1221. Reynolds, S.T. & West, J.G., 1985. Sapindaceae. In: George, A.S. (Editor): *Flora of Australia*. Vol. 25. Melianthaceae to Simaroubaceae. Australian Government Publishing Service, Canberra, Australia. pp. 4–164.
1222. Reynolds, T., 1985. The compounds in Aloë leaf exudates: a review. *Botanical Journal of the Linnean Society* 90: 157–177.
1223. Rez-Ul-Jilal, Jabbar, A. & Hasan, C.M., 1986. Hypoglycemic activities of the glycosides of *Momordica cochinchinensis*. *Journal of the Bangladesh Academy of Science* 10(1): 25–30.
1224. Rhee, J.K., Woo, K.J., Baek, B.K. & Ahn, B.J., 1982. Screening of the wormicidal Chinese raw drugs on *Clonorchis sinensis*. *American Journal of Chinese Medicine* 9(4): 277–284.
1225. Richards, A.J., 1985. Sectional nomenclature in *Taraxacum* (Asteraceae). *Taxon* 34: 633–644.
1226. Rickard, P.P. & Cox, P.A., 1986. Use of Derris as a fish poison in Guadalcanal, Solomon Islands. *Economic Botany* 40(4): 479–484.
1227. Ridley, H.N., 1922–1925. The flora of the Malay Peninsula. 5 volumes. Government of the Straits Settlements and Federated Malay States. L. Reeve & Co, London, United Kingdom.
1228. Ridley, H.N., 1930. The dispersal of plants throughout the world. L. Reeve & Co., Ashford, United Kingdom. 744 pp.
1229. Riedl, H., 1997. Boraginaceae. In: Kalkman, C., Kirkup, D.W., Nootboom, H.P., Stevens, P.F. & de Wilde, W.J.J.O. (Editors): *Flora Malesiana*. Series 1. Vol. 13. Rijksherbarium/Hortus Botanicus, Leiden, the Netherlands. pp. 43–144.
1230. Rifai, M.A., 1973. Catatan botani tentang tabat barito [A botanical note on 'tabat barito' (*Ficus deltoidea* var. *deltoidea*)]. *Acta Pharmaceutica* 4(3–4): 67–71.
1231. Riggs, D.R., Dehaven, J.I. & Lamm, D.L., 1997. *Allium sativum* (garlic) treatment for murine transitional cell carcinoma. *Cancer* 79(10) 1987–1994.
1232. Rimando, A.M., Inoshiri, S., Otsuka, H., Kohda, H., Yamasaki, K., Padolina, W.G., Torres, L., Quintana, E.G. & Cantoria, M.G., 1987. Screening for mast cell histamine release inhibitory activity of Philippine medicinal plants active constituent of *Ehretia microphylla*. *Shoyakugaku Zasshi* 41(3): 242–247. (in Japanese)
1233. Rimpler, H. & Schäfer, B., 1979. Hastatosid, ein neues Iridoid aus *Verbena hastata* L. und *Verbena officinalis* L. [Hastatoside, a new iridoid from *Verbena hastata* L. and *Verbena officinalis* L.]. *Zeitschrift für Naturforschung*. Section C, Biosciences 34: 311–318.

1234. Rinderle, S.J., Goldstein, I.J., Matta, K.L. & Ratchliffe, R.M., 1989. Isolation and characterization of Amaranthin, a lectin present in the seeds of *A. caudatus*. *Journal of Biological Chemistry* 264(27): 16123–16131.
1235. Rizk, A.M., 1982. Constituents of plants growing in Qatar I. A chemical survey of 60 plants. *Fitoterapia* 53: 35–44.
1236. Rizk, A.M., 1987. The chemical constituents and economic plants of the Euphorbiaceae. *Botanical Journal of the Linnean Society* 94: 293–326.
1237. Robillos, Y.U., 1976. Some medicinal forest trees in the Philippines. FORPRIDECOM Technical Note No 169. 3 pp.
1238. Robson, N.K.B., 1974. *Hypericum*. In: van Steenis, C.G.G.J. (General editor): *Flora Malesiana*. Series 1, Vol. 8. Sijthoff & Noordhoff International Publishers, Alphen aan den Rijn, the Netherlands. pp. 14–29.
1239. Robson, N.K.B., 1977. Studies in the genus *Hypericum* L. (Guttiferae) I. Infrageneric classification. *Bulletin of the British Museum (Natural History)*, Botany 5(6): 291–355.
1240. Robson, N.K.B., 1980. *Hypericum* L. In: Townsend, C.C. & Guest, E.R. (Editors): *Flora of Iraq*. Vol. 4. Ministry of Agriculture and Agrarian Reform, Bagdad, Iraq. pp. 363–381.
1241. Robson, N.K.B., 1981. Studies in the genus *Hypericum* L. (Guttiferae). I. Characters of the genus. *Bulletin of the British Museum (Natural History)*, Botany 8(2): 55–226.
1242. Robson, N.K.B., 1990. Studies in the genus *Hypericum* L. (Guttiferae). 8. Sections 29. *Brathys* (part 2) and 30. *Trigynobathys*. *Bulletin of the British Museum (Natural History)*, Botany 20(1): 1–151.
1243. Robson, N.K.B., 1996. Guttiferae. In: Huang, T.-C. (Editor): *Flora of Taiwan*. 2nd Edition. Vol. 2. Editorial Committee of the Flora of Taiwan, Taipei, Taiwan, Republic of China. pp. 694–714.
1244. Roddick, J.G., 1986. Steroidal alkaloids of the Solanaceae. In: d'Arcy, W.G. (Editor): *Solanaceae: biology and systematics*. Columbia University Press, New York, United States. pp. 201–222.
1245. Roddick, J.G., 1991. The importance of the Solanaceae in medicine and drug therapy. In: Hawkes, J.G., Lester, R.N., Nee, M. & Estrada, N. (Editors): *Solanaceae III: Taxonomy, chemistry, evolution*. Royal Botanic Gardens, Kew & the Linnean Society of London, London, United Kingdom. pp. 7–23.
1246. Roder, E., Wiedenfeld, H., Kersten, R. & Kroger, R., 1990. Determination of open chain pyrrolizidine alkaloids by capillary gas chromatography. *Planta Medica* 56(6): 522.
1247. Rodriguez, A., 1992. An introduction to *Jatropha*. *The Plantsman* 14: 48–53.
1248. Rodriguez-Bigas, M., Cruz, N.I. & Suarez, A., 1988. Comparative evaluation of *Aloe vera* in the management of burn wounds in guinea pigs. *Plastic and Reconstructive Surgery* 81(3): 386–389.
1249. Rodriguez-Moran, M., Guerrero-Romero, F. & Laczano-Burciaga, C., 1998. Lipid- and glucose-lowering efficiency of *Plantago psyllium* in type II diabetes. *Journal of Diabetes and its Complications* 12(5): 273–278.
1250. Roe, K.E., 1972. A revision of *Solanum* section *Brevantherum* (Solanaceae). *Brittonia* 24: 239–278.
1251. Roe, K.E., 1979. Dispersal and speciation in *Solanum*, section *Brevan-*

- therum. In: Hawkes, J.G., Lester, R.N. & Skelding, A.D. (Editors): The biology and taxonomy of the Solanaceae. Linnean Society Symposium Series 7. Academic Press, London, United Kingdom. p. 563–567.
1252. Rohwer, J.G., 1993. Moraceae. In: Kubitzki, K., Rohwer, J.G. & Bittrich, V. (Editors): The families and genera of vascular plants. Volume 2. Springer Verlag, Berlin, Germany. p. 438–453.
1253. Roja, G. & Heble, M.R., 1996. Indole alkaloids in clonal propagules of *Rauwolfia serpentina* Benth. ex Kurz. *Plant Cell, Tissue and Organ Culture* 44(2): 111–115.
1254. Rojanapo, W., Tepsuwan, A. & Siripong, P., 1990. Mutagenicity and antimutagenicity of Thai medicinal plants. *Basic Life Sciences* 52: 447–452.
1255. Rossignol, L., Rossignol, M. & Haicour, R., 1987. A systematic revision of *Phyllanthus* subsection *Urinaria*, Euphorbiaceae. *American Journal of Botany* 74(12): 1853–1862.
1256. Roy, A.K., 1979. Some new leaf spot diseases in India. *Geobios* 6(1): 36–37.
1257. Roy, A.K., Dhir, H., Sharma, A. & Talukder, G., 1991. *Phyllanthus emblica* fruit extract and ascorbic acid modify hepatotoxic and renotoxic effects of metals in mice. *International Journal of Pharmacognosy* 29(2): 117–126.
1258. Roy Shamal, K. & Datta, P.C., 1985. Establishment of high-yielding genetic line of *Andrographis paniculata* through cell culture. In: Proceedings of the 12th Annual Bangladesh Science Conference (Section 2). Bangladesh Association for the Advancement of Science, Dhaka, Bangladesh. p. 15.
1259. Roy Shamal, K. & Datta, P.C., 1988. Chromosomal biotypes of *Andrographis paniculata* in India and Bangladesh. *Cytologia* 53(2): 369–378.
1260. Roychoudhury, R., 1980. Effects of extracts of certain solanaceous plants on plant virus infection. *Acta Botanica Indica* 8(1): 91–94.
1261. Ruangrunsi, N., Tappayuthpijarn, P., Tantivatana, P., Borris, B.P. & Cordell, G.A., 1981. Traditional medicinal plants of Thailand. I. Isolation and structure elucidation of two new flavonoids (2r,3r)-dihydroquercetin-4-methyl ether and (2r,3r)-dihydroquercetin-4',7-dimethyl ether from *Blumea balsamifera* (Compositae). *Journal of Natural Products* 44: 541–545.
1262. Rucker, G., Walter, R.D., Manns, D. & Mayer, R., 1991. Antimalarial activity of some natural peroxides. *Planta Medica* 57(3): 295–296.
1263. Rugayah & de Wilde, W.J.J.O., 1997. *Trichosanthes* L. (Cucurbitaceae) in Java. *Blumea* 42(2): 471–482.
1264. Rukmana, R., 1994. Kencur [*Kaempferia galanga*]. Penerbit Kanisius, Yogyakarta, Indonesia. 29 pp.
1265. Rumphius, G.E., 1741–1747. *Herbarium Amboinense; Het Amboinsch Kruidboek*. 7 volumes (Latin, Dutch). Joannus Burmannus, Amsterdam, The Netherlands. Vol. 1: XXXIII+200 pp., Vol. 2: 270 pp., Vol. 3: 218 pp., Vol. 4: 154 pp., vol. 5: IV+492 pp., vol. 6: IIII+256 pp., vol. 7: 74 pp.+ XXI (Index Universalis).
1266. Ruyter, C.M., Akram, M., Illahi, I. & Stöckigt, J., 1991. Investigation of the alkaloid content of *Rauwolfia serpentina* roots from regenerated plants. *Planta Medica* 57(4): 328–330.



1267. Rwangabo, P.C., Claeys, M., Pieters, L., Corthout, J., Berghe, D.A.V. & Vlietinck, A.J., 1988. Umuhengerin, a new antimicrobially active flavonoid from *Lantana trifolia*. *Journal of Natural Products* 51(5): 966-968.
1268. Sahu, T.R., 1982. Taxonomic studies of the genus *Ageratum* L. in India. *Feddes Repertorium* 93(1-2): 61-65.
1269. Saito, H., Ishiguro, T., Imanishi, K. & Suzuki, I., 1982. Pharmacological studies on a plant lectin, Aloctin A. II. Inhibitory effect of aloctin A on experimental models of inflammation in rats. *Japanese Journal of Pharmacology* 32(1): 139-142.
1270. Sakai, W., 1916. Pharmacological action of *Ligusticum acutilobum* Sieb. & Zucc. *Tokyo Igakukai Zasshi* 30(24): 19-52. (in Japanese)
1271. Sakaki, T., Yoshimura, S., Ishibashi, M., Tsuyuki, T., Takahashi, T., Honda, T. & Nakanishi, T., 1984. New quassinoid glycosides, yadanziosides A - H, from *Brucea javanica*. *Chemical and Pharmaceutical Bulletin (Tokyo)* 32(11): 4702-4705.
1272. Sakaki, T., Yoshimura, S., Tsuyuki, T., Takahashi, T. & Honda, T., 1986. Yadanzioside P, a new antileukemic quassinoid glycoside from *Brucea javanica* (L.) Merr. with the 3-O-( $\beta$ -D-glucopyranosyl)bruceantin structure. *Chemical and Pharmaceutical Bulletin (Tokyo)* 34(10): 4447-4450.
1273. Sakaki, T., Yoshimura, S., Tsuyuki, T., Takahashi, T., Honda, T. & Nakanishi, T., 1986. Two new quassinoid glycosides, yadanziosides N and O isolated from seeds of *Brucea javanica* (L.) Merr. *Tetrahedron Letters* 27(5): 593-596.
1274. Salvosa, F.M., 1963. *Lexicon of Philippine trees*. Bulletin No 1. Forest Products Research Institute, College, Laguna, the Philippines. 136 pp.
1275. Samsijah, 1980. Pengaruh bentuk dan frekuensi pemangkasan tanaman murbei terhadap produksi daunnya [Effect of shape and frequency of mulberry pruning on leaf production]. Laporan 333. Lembaga Penelitian Hutan, Bogor, Indonesia. 29 pp.
1276. Samsijah, 1986. Jenis daun murbei dan ras ulat yang cocok untuk pengembangan persuteraan alam di Payakumbuh, Sumatra Barat [Leaves of mulberry species and silkworm races suitable for the development of the silk industry in the Payakumbuh area, West Sumatra]. *Buletin Penelitian Hutan* 484: 17-40.
1277. Samuelsson, G. (Editor), 1992. *Drugs of natural origin, a textbook of pharmacognosy*. Swedish Pharmaceutical Press, Stockholm, Sweden. 320 pp.
1278. Sands, D.C., Darlington, L., McCarthy, M.K., Pilgeram, A.L. & Ford, E.F., 1995. An effective and host-specific pathogen of *Erythroxylum* spp. *Phytopathology* 85: 1118.
1279. Sankara Subramanian, S., Nagarajan, S. & Sulochana, N., 1971. Flavonoids of some euphorbiaceous plants. *Phytochemistry* 10: 2548-2549.
1280. Sankaram, A.V.B., Srinivasa Rao, A. & Shoolery, J.N., 1979. Zeylanone and isozelanone, two novel quinones from *Plumbago zeylanica*. *Tetrahedron Letters* 35: 1777-1782.
1281. Sankaram, A.V.B., Srinivasa Rao, A. & Sidhu, G.S., 1976. Chitranone - a new binaphtaquinone from *Plumbago zeylanica*. *Phytochemistry* 15: 237-238.

1282. Santa, I.G.P., 1996. Studi taksonomi sambiloto *Andrographis paniculata* (Burm.f.) Nees [Taxonomic study of sambiloto *Andrographis paniculata* (Burm.f.) Nees]. *Warta Tumbuhan Obat Indonesia* 3(1): 14–15.
1283. Santamour, F.S. & Riedel, L.G.H., 1997. A new name for *Sophora japonica*. *Journal of Arboriculture* 23(4): 166–167.
1284. Santhakumari, G., Saralamma, P.G. & Radhakrishnan, N., 1980. Effect of plumbagin on cell growth and mitosis. *Indian Journal of Experimental Biology* 18(3): 215–218.
1285. Santhakumari, G. & Suganthan, D., 1980. Antigonadotrophic activity of plumbagin. *Planta Medica* 39: 244.
1286. Santos, A.R., Filho, V.C., Yunes, R.A. & Calixto, J.B., 1995. Analysis of the mechanisms underlying the antinociceptive effect of the extracts of plants from the genus *Phyllanthus*. *General Pharmacology* 26(7): 1499–1506.
1287. Saralamp, P., Tamsiririrkkul, R., Chuakul, W., Riewpaiboon, A., Prathanturug, S., Suthisisang, C. & Pongcharoensuk, P. (Editors), 1996. *Medicinal plants in the Siri Ruckhachati Garden*. 2nd Edition. Siambooks and Publications Co., Bangkok, Thailand. 263 pp.
1288. Sashida, Y., Kubo, S., Mimaki, Y., Nikaido, T. & Ohmoto, T., 1992. Steroidal saponins from *Smilax riparia* and *S. china*. *Phytochemistry* 31(7): 2439–2443.
1289. Sastrapradja, S. & Afristiani, J.J., 1984. Kerabat beringin [The genus *Ficus*]. *Seri Sumber Daya Alam* 115. Lembaga Biologi Nasional – LIPI, Bogor, Indonesia. 118 pp.
1290. Sastrapradja, S. & Aminah, S.H., 1970. Factors affecting fruit production in *Curcuma* species. *Annales Bogorienses* 5(2): 99–107.
1291. Satayavivad, J., Bunyapraphatsara, N. & Saivises, R., 1980. Pharmacological and toxicological studies to the constituents of *Schefflera venulosa* (Araliaceae). In: Department of Chemistry, Faculty of Science, Mahidol University: 4th Asian Symposium on Medicinal Plants and Spices (Abstracts). Mahidol University, Bangkok, Thailand. p. 47.
1292. Satyan, K.S., Prakash, A., Singh, R.P. & Srivastava, R.S., 1995. Phthalic acid bis-ester and other phytoconstituents of *Phyllanthus urinaria*. *Planta Medica* 61(3): 293–294.
1293. Saxena, B.P., Thappa, R.K., Tikku, K., Sharma, A. & Suri, O.P., 1996. Effect of plumbagin on gonadotrophic cycle of the housefly, *Musca domestica* L. *Indian Journal of Experimental Biology* 34(8): 739–744.
1294. Saxena, R.C., Jayashree, S., Padma, S. & Dixit, O.P., 1994. Evaluation of growth disrupting activity of *Ageratum conyzoides* crude extract on *Culex quinquefasciatus* (Diptera: Culicidae). *Journal of Environmental Biology* 15(1): 67–74.
1295. Sayavathi, D.V.L. & Narayana, L.L., 1990. The distribution pattern of free amino acids in certain Boraginaceae. *Feddes Repertorium* 101(11-12): 593–599.
1296. Scheibelreiter, G.K., 1978. The poppy-cephid *Pachycephus smyrnensis* Stein (Hymenoptera: Cephidae). *Zeitschrift für Angewandte Entomologie* 86(1): 19–25.
1297. Schilling, E.E. & Andersen, R.N., 1990. The black nightshades (*Solanum* section *Solanum*) of the Indian subcontinent. *Botanical Journal of the*

- Linnean Society 102: 253–259.
1298. Schliemann, W., Joy, R.W., Komanine, A., Metzger, J.W., Nimtz, M., Wray, V. & Strack, D., 1996. Betacyanins from plants and cell cultures of *Phytolacca americana*. *Phytochemistry* 42(4): 1039–1046.
1299. Schoental, R., 1968. Toxicology and carcinogenic action of pyrrolizidine alkaloids. *Cancer Research* 28: 2237.
1300. Scholz, E., 1995. Amla – ein exotische Vitamin C-Quelle. [Amla – an exotic vitamin C source]. *Naturwissenschaftliche Rundschau* 48(8): 308–310.
1301. Schultes, R.E. & Raffauf, R.F., 1990. The healing forest. Medicinal and toxic plants of the northwest Amazonia. Dioscorides Press, Portland, Oregon, United States. 484 pp.
1302. Schütte, H.-R., 1991. Secondary plant substances: monoterpene indole alkaloids. *Progress in Botany* 52: 84–96.
1303. Science Education Center, University of the Philippines, 1971. Plants of the Philippines. M. & L. Licudine Enterprises, St. Michael Village, Las Piñas, Metro Manila, the Philippines. 512 pp.
1304. Seddigh, M., Jolliff, G.D., Calhoun, W. & Crane, J.M., 1982. *Papaver bracteatum*, potential commercial source of codeine. *Economic Botany* 36(4): 433–441.
1305. Seigler, D.S. & Kawahara, W., 1976. New reports of cyanolipids from sapindaceous plants. *Biochemical Systematics and Ecology* 4: 263–265.
1306. Sekulic, D., Jovanovic, Z., Kostic, M. & Sekulovic, D., 1995. Preliminary testing of plant extracts for acaricide activity. *Pharmazie* 50(12): 835.
1307. Selvaraj, C. & Narayanasamy, P., 1991. Effect of plant extracts in controlling rice tungro. *International Rice Research Newsletter* 16(2): 21–22.
1308. Sengupta, A., Gupta, J.K., Dutta, J. & Ghosh, A., 1973. The component fatty acids of chaulmoogra oil. *Journal of the Science of Food and Agriculture* 24(6): 669–674.
1309. Sengupta, P. & Mukhopadhyay, J., 1966. Terpenoids and related compounds –VII. Triterpenoids of *Phyllanthus acidus* Skeels. *Phytochemistry* 5: 531–534.
1310. Serrame, E. & Lim-Sylianco, C.Y., 1995. Anti-tumor promoting activity of decoctions and expressed juices from Philippine medicinal plants. *Philippine Journal of Science* 124(3): 275–281.
1311. Seshagirirao, K., Narasimha, M. & Prasad, V., 1995. Purification and partial characterization of a lectin from *Euphorbia neriifolia* latex. *Biochemistry and Molecular Biology International* 35(6): 1199–1204.
1312. Shah, S.C. & Gupta, L.K., 1977. Note on pharmacognostic study of the rhizome of *Acorus calamus* Linn. *Indian Journal of Agricultural Research* 11(2): 113–115.
1313. Shanavas, K.R., Elyas, K.K. & Vasudevan, D.M., 1995. Purification and some properties of a lectin from the seeds of *Trichosanthes anguina*. *Biologia Plantarum* 37(3): 417–422.
1314. Shang, C.B., 1984. Le genre *Schefflera* (Araliacées) en Chine et en Indochine. *Candollea* 39: 453–486.
1315. Shanmugasundaram, E.R.B., Akbar, G.K.M. & Shanmugasundaram, K.R., 1991. Brahmighritham, an Ayurvedic herbal formula for the control of epilepsy. *Journal of Ethnopharmacology* 33(3): 269–276.
1316. Sharaf, A., 1969. Food plants as a possible factor in fertility control.

- Qualitas Plantarum et Materiae Vegetabilis 17: 153.
1317. Sharaf, A. & Mansour, M.Y., 1964. Pharmacological studies on the leaves of *Morus alba*, with special reference to its hypoglycemic activity. *Planta Medica* 12(1): 71–76.
1318. Sharma, D.K. & Hall, I.H., 1991. Hypolipidemic, anti-inflammatory, and antineoplastic activity and cytotoxicity of flavonolignans isolated from *Hydnocarpus wightiana* seeds. *Journal of Natural Products* 54(5): 1298–1302.
1319. Sharma, M.P., 1991. Lesser-known medicinal uses of plants from Mewat (District Gungaon), Haryana, India. *Economic Botany* 45: 435–436.
1320. Sharma, N. & Chandel, K.P.S., 1992. Low-temperature storage of *Rauvolfia serpentina* Benth. ex Kurz: An endangered, endemic medicinal plant. *Plant Cell Reports* 11(4): 200–203.
1321. Sharma, O.P., Dawra, R.K. & Pattabhi, V., 1991. Molecular structure, polymorphism, and toxicity of lantadene A, the pentacyclic triterpenoid from the hepatotoxic plant *Lantana camara*. *Journal of Biochemistry and Toxicology* 6(1): 57–63.
1322. Sharma, O.P., Makkar, H.P. & Dawra, R.K., 1988. A review of the noxious plant *Lantana camara*. *Toxicon* 26(11): 975–987.
1323. Sharma, O.P., Vaid, J., Pattabhi, V. & Bhutani, K.K., 1992. Biological action of lantadene C, a new hepatotoxicant from *Lantana camara* var. *aculeata*. *Journal of Biochemistry and Toxicology* 7(2): 73–79.
1324. Sharma, V.S., 1987. Comments on the identity of *Ageratum conyzoides* L., and *A. houstonianum* Mill. – two naturalized weeds in India. *Feddes Repertorium* 98(11–12): 557–560.
1325. Shaw, P.C., Chan, W.L., Yeung, H.W. & Ng, T.B., 1994. Trichosanthin – A protein with multiple pharmacological properties. *Life Sciences* 55(4): 253–262.
1326. Shen, Y.C. & Chen, C.H., 1989. Novel secoiridoid lactones from *Jasminum multiflorum*. *Journal of Natural Products* 52(5): 1060–1070.
1327. Shen, Y.C., Lin, C.Y. & Chen, C.H., 1990. Secoiridoid glycosides from *Jasminum multiflorum*. *Phytochemistry* 29(9): 2905–2912.
1328. Sheng, C.K., Sheng, C.C., Lin, H.C., Jin, B.W., Jih, P.W. & Che, M.T., 1995. Potent antiplatelet, anti-inflammatory and antiallergic isoflavan-quinones from the roots of *Abrus precatorius*. *Planta Medica* 61: 307–312.
1329. Sherff, E.E., 1937. The genus *Bidens*. 2 parts. *Field Museum of Natural History* 16: 1–709.
1330. Sheu, S.J., Ho, Y.S., Chen, Y.P. & Hsu, H.Y., 1987. Analysis and processing of Chinese herbal drugs. VI. The study of *Angelicae Radix*. *Planta Medica* 53(4): 377–378.
1331. Shih, N.J.R., McDonald, K.A., Dandekar, A.M., Girbes, T., Iglesias, R. & Jackman, A.P., 1998. A novel type-1 ribosome-inactivating protein isolated from the supernatant of transformed suspension cultures of *Trichosanthes kirilowii*. *Plant Cell Reports* 17(6–7): 531–537.
1332. Shih, N.J.R., McDonald, K.A., Jackman, A.P., Girbes, T. & Iglesias, R., 1997. Bifunctional plant defence enzymes with chitinase and ribosome inactivating activities from *Trichosanthes kirilowii* cell cultures. *Plant Science* 130(2): 145–150.

1333. Shinazaki, H., Hirate, K. & Ishida, M., 1987. Modification of drug-induced tremor by systematic administration of kainic acid and quisqualic acid in mice. *Neuropharmacology* 26(1): 9-18.
1334. Shingla, A.K. & Pathak, K., 1989. Anti-inflammatory studies on *Euphorbia prostrata*. *Journal of Ethnopharmacology* 27(1-2): 55-62.
1335. Shingla, A.K. & Pathak, K., 1990. Topical anti-inflammatory effects of *Euphorbia prostrata* on carrageenan-induced footpad oedema in mice. *Journal of Ethnopharmacology* 29(3): 291-294.
1336. Shirahata, A. & Takahashi K., 1982. Detection and production of anti microbial substances in leaves of mulberry and other Moraceae tree plants. *Bulletin Sericulture Experimental Station* 28(5): 707-718. (in Japanese)
1337. Shivanna, S., Lingappa, S. & Patil, B.V., 1994. Effectiveness of selected plant materials as protectants against pulse beetle, *Callosobruchus chinensis* (Linn.) during storage of red gram. *Karnataka Journal of Agricultural Sciences* 7(3): 285-290.
1338. Shook, J.E. & Burks, T.F., 1989. Psychoactive cannabinoids reduce gastrointestinal propulsion and motility in rodents. *Journal of Pharmacology and Experimental Therapeutics* 249: 444-449.
1339. Shoyama, Y., Yagi, M. & Nishioka, I., 1975. Biosynthesis of cannabinoid acids. *Phytochemistry* 14: 2189-2192.
1340. Shoyama, Y., Yamauchi, T. & Nishioka, I., 1970. Cannabigerolic acid monomethylether and cannabinolic acid. *Chemical and Pharmaceutical Bulletin* 18: 1327-1332.
1341. Shubhashish Sarkar, Maddali Pranava & Marita, A.R., 1996. Demonstration of the hypoglycemic action of *Momordica charantia* in a validated animal model of diabetes. *Pharmacological Research* 33(1): 1-4.
1342. Shukla, O.P. & Crishna-Murti, C.R., 1971. The biochemistry of plant latex. *Journal of Scientific and Industrial Research* 30: 640-662.
1343. Shukla, S.C. & Das, S.R., 1988. Cure of amoebiasis by seed powder of *Cassia fistula*. *International Journal of Crude Drug Research* 26(3):141-144.
1344. Si-man, Z., Yong-sheng, H., Tabbal, H.D. & Smith, K.M., 1988. Inhibitor against the human immunodeficiency virus in aqueous extracts of *Alternanthera philoxeroides*. *Chinese Medical Journal* 101: 861-886.
1345. Siddaramaiah, A.L. & Hegde, R.K., 1989. Development and severity of *Cercospora* leaf spot of mulberry in relation to environmental factors. *Mysore Journal of Agricultural Sciences* 23(2): 189-192.
1346. Siddharta, P. & Chaudhuri, A.K.N., 1990. Anti-inflammatory action of *Bryophyllum pinnatum* leaf extract. *Fitoterapia* 41(6): 527-533.
1347. Siddharta, P. & Chaudhuri, A.K.N., 1991. Studies on the anti-ulcer activity of a *Bryophyllum pinnatum* leaf extract in experimental animals. *Journal of Ethnopharmacology* 33: 97-102.
1348. Siddiqui, B.S., Usmani, S.B., Begum, S. & Siddiqui, S., 1993. Steroidal alkaloids and an androstane derivative from the bark of *Holarrhena pubescens*. *Phytochemistry* 33(4): 925-928.
1349. Siddiqui, B.S., Usmani, S.B., Begum, S. & Siddiqui, S., 1994. Steroidal constituents of *Holarrhena pubescens*. *Journal of Natural Products (Lloydia)* 57(1): 27-31.
1350. Siddiqui, M.B., Alam, M.M. & Husnia, W., 1989. Traditional treatment of

- skin diseases in Uttar Pradesh, India. *Economic Botany* 43: 480–486.
1351. Siddiqui, S., Faizi, S., Siddiqui, B.S. & Sultana, S., 1989. Triterpenoids and phenanthrenes from leaves of *Bryophyllum pinnatum*. *Phytochemistry* 28(9): 2433–2438.
1352. Siddiqui, S. & Shamsuddin, B.A., 1989. Isolation and structure of holar-rifine, a new alkaloid from the bark of *Holarrhena antidysenterica*. *Pakistan Journal of Scientific and Industrial Research* 32(1): 1–3.
1353. Sidhu, G.S. & Sankaram, A.V.B., 1971. A new biplumbagin and 3-chloro-plumbagin from *Plumbago zeylanica*. *Tetrahedron Letters* 26: 2385–2388.
1354. Siebertz, R., Proksch, P. & Witte, L., 1990. Accumulation and biosynthesis of the chromenes precocene I and II in *Ageratum houstonianum*. *Phytochemistry* 29(7): 2135–2138.
1355. Siegers, C.P., von Hertzberg-Lottin, E., Otte, M. & Schneider, B., 1993. Anthranoid laxative abuse – a risk for colorectal cancer? *Gut* 34(8): 1099–1101.
1356. Siemonsma, J.S. & Kasem Piluek (Editors), 1993. *Plant Resources of South-East Asia No 8. Vegetables*. Pudoc Scientific Publishers, Wageningen, the Netherlands. 412 pp.
1357. Sigounas, G., Hooker, J., Anagnostou, A. & Steiner, M., 1997. S-allylmercaptocysteine inhibits cell proliferation and reduces the viability of erythroleukemia, breast and prostate cancer cell lines. *Nutrition and Cancer* 27(2): 186–191.
1358. Sills, M.A., 1996. Strategic considerations for screening natural products. *Network Science* 2(5): <http://www.netsci.org/Science/Screening/feature10.html>.
1359. Silverstein, F.S., Chen, R. & Johnston, M.V., 1986. The glutamate analogue, quisqualic acid in neurotoxic in striatum and hippocampus of immature rat brain. *Neuroscience Letters* 71(1): 13–18.
1360. Sinchaisri, P., 1989. Phon khong suan sakat khong phut totan mateng thimi, to alkylating activity khong ethyl methane sulphonate [Effects of solvent fractions of some anti-carcinogenic plants on the alkylating activity of ethyl methane sulphonate]. *Warasan Wicha kan kaset* 7(1–3): 27–33.
1361. Singh, A. & Singh, D.K., 1994. Pestoban, a potent herbal molluscicide. *Biological Agriculture and Horticulture* 10(3): 175–178.
1362. Singh, A., Singh, S.P. & Bamezai, R., 1998. *Momordica charantia* (bitter gourd) peel, pulp, seed and whole fruit extract inhibits mouse skin papillomagenesis. *Toxicology Letters* 94(1): 37–46.
1363. Singh, I.S., Ali, W. & Pathak, R.K., 1994. New varieties of Indian gooseberry. *Indian Horticulture* 39(1): 3–5.
1364. Singh, K., Singh, A. & Singh, D.K., 1995. Molluscicidal activity of different combinations of the plant products used in the molluscicide Pestoban. *Biological Agriculture & Horticulture* 12(3): 253–261.
1365. Singh, M., Alam, S.P. & Siddiqui, M.O., 1996. Study on germination behaviour of *Blumea* spp. *Acta Botanica Indica* 24(1): 11–15.
1366. Singh, P. & Sinha, K.K., 1986. Inhibition of aflatoxin production on some agricultural commodities through aqueous plant extracts. *Journal of the Indian Botanical Society* 65(1): 30–32.

1367. Singh, S.S., 1992. Rooting and regeneration potential of stem cuttings of *Aerva sanguinolenta* L. as influenced by ethyl hydrogen-1-propyl phosphonate (Niagara). *Acta Botanica Indica* 20(2): 294–296.
1368. Singh, S.S. & Paliwal, G.S., 1985. Effect of stik on rooting and regeneration of stem cuttings of *Aerva sanguinolenta*. *Current Science* (Bangalore) 54(18): 920–923.
1369. Singh, V., Mathur, K., Sethia, M. & Nag, T.N., 1988. Desert medicinal plants a source of steroids. *Oikoassay* 5(2): 41–42.
1370. Sinha, P., Arora, V.K., Ansari, M.S. & Wahi, S.P., 1986. Pharmacognostic studies on the stem of *Alternanthera sessilis* R.Br. *International Journal of Crude Drug Research* 24(1): 7–15.
1371. Sirirugsa, P., 1989. The genus *Kaempferia* (Zingiberaceae) in Thailand. *Nordic Journal of Botany* 9: 257–260.
1372. Sirirugsa, P., 1992. Taxonomy of the genus *Kaempferia* (Zingiberaceae) in Thailand. *Thai Forest Bulletin (Botany)* 19: 1–15.
1373. Sivarajan, V.V. & Biju, S.D., 1990. Taxonomic and nomenclatural notes on the *Hedyotis corymbosa-diffusa* complex (Rubiaceae) in India. *Taxon* 39(4): 665–674.
1374. Siwon, J., 1982. A pharmacognostical study of some Indonesian plants of the family Menispermaceae. Thesis. Faculty of Pharmacy, Airlangga University, Surabaya, Indonesia. 108 pp.
1375. Skerman, P.J. & Riveros, F., 1990. Tropical grasses. FAO, Rome, Italy. 832 pp.
1376. Sleumer, H., 1938. Monographie der Gattung *Hydnocarpus* Gaertner [Monograph of the genus *Hydnocarpus* Gaertner]. *Botanische Jahrbücher* 69: 1–94.
1377. Sleumer, H., 1954. Flacourtiaceae. In: van Steenis, C.G.G.J. (General editor): *Flora Malesiana*. Series 1, Vol. 5. Noordhoff-Kolff N.V., Djakarta, Indonesia. pp. 1–106.
1378. Sleumer, H., 1985. The Flacourtiaceae of Thailand. *Blumea* 30: 217–250.
1379. Smith, R.M., 1985. A review of Bornean Zingiberaceae: 1 (Alpineae p.p.). *Notes from the Royal Botanic Garden Edinburgh* 42(2): 261–314.
1380. Smitinand, T., 1980. Thai plant names. Royal Forest Department, Bangkok, Thailand. 379 pp.
1381. Snoeijer, W., 1996. *Catharanthus roseus*, the Madagascar periwinkle, a review of its cultivars. *Wageningen Agricultural Papers* 96-3: 47–120.
1382. Soepadmo, E., 1991. Conservation status of medicinal plants in Peninsular Malaysia. In: Shaari, K., Kadir, A.A. & Ali, A.R.H. (Editors): *Medicinal products from tropical rain forests*. Proceedings of the conference, Forest Research Institute Malaysia, May 13–15, 1991. Forest Research Institute Malaysia, Kepong, Selangor, Malaysia. pp. 13–23.
1383. Soepadmo, E., Wong, K.M. & Saw, L.G. (Editors), 1995–. *Tree flora of Sabah and Sarawak*. Sabah Forestry Department, Forest Research Institute Malaysia and Sarawak Forestry Department, Kepong, Malaysia.
1384. Soepardi Poerwokoesomo, R., 1981. Tumbuh-tumbuhan dalam hutan jati yang berkhasiat obat [Medicinal plants from teak forests]. Perum Perhutani, Jakarta, Indonesia. 61 pp.
1385. Soerjani, M., 1970. Alang-alang, *Imperata cylindrica* (L.) Beauv., pattern of growth as related to its problem of control. *Biotrop Bulletin* No 1.

- SEAMEO Regional Centre for Tropical Biology, Bogor, Indonesia. 88 pp.
1386. Soerjani, M., Kostermans, A.J.G.H. & Tjitrosoepomo, G., (Editors) 1987. Weeds of rice in Indonesia. Balai Pustaka, Jakarta, Indonesia. 716 pp.
1387. Soetopo, D., Sufiani, S. & Hamid A., 1990. Tanaman mentha (*Mentha piperita* L. dan *Mentha arvensis* L.). [Mentha (*Mentha piperita* L. and *Mentha arvensis* L.)]. Penelitian Tanaman Rempah dan Obat 6(1): 38–44.
1388. Solsoloy, A.D., Cacayorin, N.D. & Cano, L.C., 1991. Insecticidal and fungicidal action of some indigenous plant extracts against cotton pests. Cotton Research Journal 4(1–2): 1–11.
1389. Solsoloy, A.D., Dumlao, N.A. & Solsoloy, T.S., 1987. Insecticidal activity of indigenous plant extracts against bollworms (*Helicoverpa armigera* Hubn.). Technical Report calendar year 1985–86, Cotton Research and Development Institute, Batac, Ilocos Norte, the Philippines. pp. 175–184.
1390. Somanabandhu, A., Nitayangkura, S., Mahidol, C., Ruchirawat, S., Likhitwitayawuid, K., Shieh, H.L., Chai, H., Pezzuto, J.M. & Cordell, G.A., 1993. <sup>1</sup>H- and <sup>13</sup>C-NMR assignments of phyllanthin and hypophyllanthin: lignans that enhance cytotoxic responses with cultured multidrug-resistant cells. Journal of Natural Products 56(2): 233–239.
1391. Sonck, C.E., 1987. A new *Taraxacum* species, *T. indonesicum*, from Java. Annales Botanici Fennici 24: 307–309.
1392. Sosef, M.S.M., Hong, L.T. & Prawirohatmodjo, S. (Editors), 1998. Plant Resources of South-East Asia No 5(3). Timber trees: Lesser-known timbers. Backhuys Publishers, Leiden, the Netherlands. 859 pp.
1393. Sprent, J.I. & McKey, D. (Editors), 1994. Advances in legume systematics. Part 5. The nitrogen factor. The Royal Botanic Gardens, Kew, United Kingdom. pp. 241.
1394. Sreedevi & Chaturvedi, A., 1993. Effect of vegetable fibre on post prandial glycemia. Plant Foods for Human Nutrition 44(1): 71–78.
1395. Srivastava, A.K., Pathak, R.K. & Singh, I.S., 1997. Genetic diversity in aonla (*Emblica officinalis* Gaertn). Indian Horticulture 42(1): 7, 10.
1396. Srivastava, A.K. & Purnima, 1990. Numerical and structural inconstancy in the chromosome complements of *Belamcanda chinensis* DC. (Iridaceae). Proceedings of the Indian Academy of Sciences (Plant Sciences) 100(3): 205–210.
1397. Srivastava, R.C., 1987. Rare and less known plants of Madhya Pradesh – II. Journal of Economic and Taxonomic Botany 11: 91–94.
1398. Srivastava, R.C. & Shukla, B.K., 1984. The genus *Blumea* DC. (Asteraceae) in Madhya Pradesh. Journal of Economic and Taxonomic Botany 5: 285–291.
1399. Srividya, N. & Periwal, S., 1995. Diuretic, hypotensive and hypoglycaemic effect of *Phyllanthus amarus*. Indian Journal of Experimental Biology 33(11): 861–864.
1400. Staubmann, R. et al., 1997. Biogas production from *Jatropha curcas* press-cake. Applied Biochemistry and Biotechnology, Part A, Enzyme Engineering and Biotechnology 63–65: 457–467.
1401. Steger, R.W., Murphy, L.L., Bartke, A. & Smith, M.S., 1990. Effects of psychoactive and non-psychoactive cannabinoids on the hypothalamic-pituitary axis of the adult male rat. Pharmacology, Biochemistry and



- Behavior 37: 299–302.
1402. Stienswat, W., Buachan, W. & Sukhiwong, R., 1983. Phaya rai bai phut petroleum [Euphorbia tirucalli – petroleum plant]. Faculty of Agriculture, Kasetsart University, Bangkok, Thailand. 21 pp.
  1403. Strzelecka, H., Glinkowska, G., Skopinska-Rozewska, E., Malkowska-Zwierz, W., Sikorska, E. & Sokolnicka, I., 1995. Immunotropic activity of plant extracts. I. Influence of water extracts of chosen crude drugs on humoral and cellular immune response. *Herba Polonica* 41(1): 23–32. (in Polish)
  1404. Subramanian, P.M. & Misra, G.S., 1978. Chemical constituents of *Ficus bengalensis*. *Polish Journal of Pharmacology and Pharmacy* 30: 559–562.
  1405. Subrata Biswas, Das, N.K., Qadri, S.M.H. & Saratchandra, B., 1995. Evaluating different plant extracts against three major diseases of mulberry. *Indian Phytopathology* 48(3): 342–346.
  1406. Sudiarto, 1989. *Kaempferia galanga* L. in Central Java and West Sumatra. In: Siemonsma, J.S. & Wulijarni-Soetjipto, N. (Editors): *Plant Resources of South-East Asia. Proceedings of the First PROSEA International Symposium, May 22–25, 1989, Jakarta, Indonesia*. Pudoc, Wageningen, the Netherlands. pp. 306–308.
  1407. Sudiarto, Rivaie, A.A. & Riyanto, 1991. Pengaruh kedalaman tanam dan macam bibit terhadap hasil rimpang kencur [The influence of planting depth and planting material on the yield of *Kaempferia galanga* rhizomes]. *Buletin Penelitian Tanaman Industri* 2: 14–19.
  1408. Sujatha, C.H., Vasuki, V., Mariappan, T., Kalyanasundaram, M. & Das, P.K., 1988. Evaluation of plant extracts for biological activity against mosquitoes. *International Pest Control* 30(5): 122, 124.
  1409. Sujatha, M. & Mukta, N., 1996. Morphogenesis and plant regeneration from tissue cultures of *Jatropha curcas*. *Plant Cell, Tissue and Organ Culture* 44(2): 135–141.
  1410. Sukanto, Manohara, D. & Wahyuno, D., 1996. Penyakit layu Sclerotium pada tanaman sambiloto [Sclerotium wilt disease on *Andrographis paniculata* Nees]. *Warta Tumbuhan Obat Indonesia* 3(1): 41–42.
  1411. Sukandar, E.Y., Suganda, A.G. & Lydia, B., 1992. Uji aktivitas antifungi beberapa tanaman suku Compositae terhadap Dermatophyta [Antifungal activity screening of several Compositae plants against Dermatophyta]. *Acta Pharmaceutica Indonesia* 17(1): 26–35.
  1412. Suksamrarn, A., Weerawattanametin, K. & Brophy, J.J., 1991. Variation of essential oil constituents in *Vitex trifolia* species. *Flavour and Fragrance Journal* 6(1): 97–99.
  1413. Sunarno, B., Lemmens, R.H.M.J. & Ani binti Sulaiman, 1995. *Vitex* L. In: Lemmens, R.H.M.J., Soerianegara, I. & Wong, W.C. (Editors): *Plant Resources of South-East Asia No 5(2). Timber trees: Minor commercial timbers*. Backhuys Publishers, Leiden, the Netherlands. pp. 502–509.
  1414. Sundar Rao, K. & Lakshminarayana, G., 1987. Characteristics and composition of six newer seeds and the oils. *Fett: Wissenschaft, Technologie* 89(8):324–326.
  1415. Sundarrao, K., Burrows, I., Kuduk, M., Yi, Y.D., Chung, M.H., Suh, N.J. & Chang, I.M., 1993. Preliminary screening of antibacterial and antitumor activities of Papua New Guinean native medicinal plants. *Inter-*

- national Journal of Pharmacognosy 31(1): 3-6.
1416. Sung, T.V. & Adam, G., 1991. A sulphated triterpenoid saponin from *Schefflera octophylla*. *Phytochemistry* 30(8): 2717-2720.
1417. Sung, T.V., Lavaud, C., Porzel, A., Steglich, W. & Adam, G., 1992. Triterpenoids and their glycosides from the bark of *Schefflera octophylla*. *Phytochemistry* 31(1): 227-231.
1418. Sung, T.V., Peter-Katalinic, J. & Adam, G., 1991. A bidesmosidic triterpenoid saponin from *Schefflera octophylla*. *Phytochemistry* 30(11): 3717-3720.
1419. Sung, T.V., Steglich, W. & Adam, G., 1991. Triterpene glycosides from *Schefflera octophylla*. *Phytochemistry* 30(7): 2349-2356.
1420. Supriadi, 1987. Penyakit layu bakteri pada bangle putih dan temu mangga [Bacterial wilt disease on bangle putih (*Zingiber cassumunar*) and temu mangga (*Curcuma mangga*)]. *Pemberitaan Lembaga Penelitian Tanaman Industri* 13(1-2): 28-32.
1421. Suresh Babu, P. & Srinivasan, K., 1997. Hypolipidemic action of curcumin, the active principle of turmeric (*Curcuma longa*) in streptozotocin-induced diabetic rats. *Molecular and Cellular Biochemistry* 166(1-2): 169-175.
1422. Suresh, K. & Vasudevan, D.M., 1994. Augmentation of murine natural killer cell and antibody dependent cellular cytotoxicity activities by *Phyllanthus emblica*, a new immunomodulator. *Journal of Ethnopharmacology* 44(1): 55-60.
1423. Sutarjadi, Santosa, M.H., Bendryman & Dyatmiko, W., 1991. Immunomodulatory activity of *Piper betle*, *Zingiber aromatica*, *Andrographis paniculata*, *Allium sativum*, and *Oldenlandia corymbosa* grown in Indonesia. *Planta Medica* 57, Supplement Issue 2: A136.
1424. Suteri, B.D., Joshi, C.C. & Bala, S., 1979. Some ornamentals and weeds as reservoirs of potato virus Y and cucumber mosaic virus in Kumaon. *Indian Phytopathology* 32(4): 640.
1425. Suwunnamek, U., 1994. *Euphorbia heterophylla* L. Weed Info Sheet 20. The Southeast Asian Information Centre, Bogor, Indonesia. 2 pp.
1426. Suzuki, I., 1981. Anti-inflammatory agent. *Eur. Pat. Appl.* 2 873 (CI A61k 35/78), 01 Apr. 1981.
1427. Syamasundar, K.V., Singh, B., Thakur, R.S., Husain, A., Kiso, Y. & Hikino, H., 1985. Antihepatotoxic principles of *Phyllanthus niruri* herbs. *Journal of Ethnopharmacology* 14(1): 41-44.
1428. Syamsuhidayat, S.S. & Hutapea, J.R., 1991. Inventaris tanaman obat Indonesia I [An inventory of Indonesian medicinal plants I]. *Badan Penelitian dan Pengembangan Kesehatan, Jakarta, Indonesia*. 616 pp.
1429. Sydskis, R.J., Owen, D.G., Lohr, J.L., Rosler, K.H. & Blomster, R.N., 1991. Inactivation of enveloped viruses by anthraquinones extracted from plants. *Antimicrobial Agents and Chemotherapy* 35(12): 2463-2466.
1430. Syed, T.A., Ahmad, S.A., Holt, A.H., Ahmad, S.A., Ahmad, S.H. & Afzal, M., 1996. Management of psoriasis with *Aloe vera* extract in a hydrophylic cream: a placebo-controlled, double-blind study. *Tropical Medicine and International Health* 1(4): 505-509.
1431. Symon, D.E., 1979. Fruit diversity and dispersal in *Solanum* in

- Australia. *Journal of the Adelaide Botanic Gardens* 1: 321–331.
1432. Symon, D.E., 1981. A revision of the genus *Solanum* in Australia. *Journal of the Adelaide Botanic Gardens* 4: 1–367.
1433. Symon, D.E., 1985. The Solanaceae of New Guinea. *Journal of Adelaide Botanic Gardens* 8: 1–171.
1434. t Mannetje, L. & Jones, R.M. (Editors), 1992. *Plant Resources of South-East Asia. No 4. Forages*. Pudoc Scientific Publishers, Wageningen, the Netherlands. 300 pp.
1435. Taesotikul, T., Panthong, A. & Kanjanapothi, D., 1980. Bronchodilator activity of *Schefflera venulosa* (family Araliaceae): preliminary investigation. In: Department of Chemistry, Faculty of Science, Mahidol University: 4th Asian Symposium on Medicinal Plants and Spices: Abstracts. Mahidol University, Bangkok, Thailand. p. 46.
1436. Taga, T., Akimoto, N. & Ibuka, T., 1984. Stephadiamine, a new skeletal alkaloid from *Stephania japonica*: the first example of a C-norhasubanan alkaloid. *Chemical and Pharmaceutical Bulletin* 32(10): 4223–4225.
1437. Tahir, S.K. & Zimmerman, A.M., 1991. Influence of marihuana on cellular structures and biochemical activities. *Pharmacology, Biochemistry and Behavior* 40: 617–623.
1438. Taibi, M. & Zamarlik, H., 1991. Synthèse des precocenes en une seule étape [Single step synthesis of precocenes]. *Tetrahedron Letters* 32(49): 7251–7252.
1439. Taikun, Z., Tanaka, T., Sakai, E., Yoshida, M. & Matsuo, K., 1990. Pharmacognostical studies of *Plantaginis Herba* 6. Anatomy of *Plantago major* in Hokkaido, Japan. *Shoyakugaku Zasshi* 44(3): 145–150. (in Japanese)
1440. Takada, N., Yano, Y., Wanibuchi, H., Otani, S. & Fukushima, S., 1997. S-methylcysteine and cysteine are inhibitors of induction of glutathione S-transferase placental form-positive foci during initiation of promotion phases of rat hepatocarcinogenesis. *Japanese Journal of Cancer Research* 88(5): 435–442.
1441. Takano, F., Yoshiaki, F., Suzuki, K., Suya, N., Ando, T. & Hisamichi, S., 1990. Anti-ulcer effects of *Trichosanthes* fruits. *Chemical and Pharmaceutical Bulletin* 38(5): 1313–1316.
1442. Taylor, R.S.L., Manandhar, N.P., Hudson, J.B. & Towers, G.H.N., 1996. Antiviral activities of Nepalese medicinal plants. *Journal of Ethnopharmacology* 52(3): 157–163.
1443. Telford, I.R., 1982. Cucurbitaceae. In: George, A.S. (Editor): *Flora of Australia. Vol. 8. Lecythidales to Batales*. Australian Government Publishing Service, Canberra, Australia. pp. 158–198.
1444. Terada, M., Sano, M., Ishii, A.I., Kino, H., Fukushima, S. & Noro, T., 1982. Studies on chemotherapy of parasitic helminths (IV). Effects of alkaloids from *Sophora flavescens* on the motility of parasitic helminths and isolated host tissues. *Nippon Yakurigaku Zasshi* 79(2): 105–111. (in Japanese)
1445. Terawaki, K., Nose, M., Kondo, T., Kojima, K., Mizukami, H. & Ogihara, Y., 1997. Effect of karasurin-A on nitric oxide production by murine macrophages and mitogenic response of murine splenocytes in vitro. *Biological and Pharmaceutical Bulletin* 20(4): 435–437.

1446. Teuscher, E., 1990. Pharmaceutische Biologie [Pharmaceutical Biology]. 4th Edition. Friedr. Vieweg & Sohn., Braunschweig, Wiesbaden, Germany. 664 pp.
1447. Tewari, P.V., Sharma, P.V. & Prasad, D.N., 1972. Experimental studies on the ecobolic properties of *Gloriosa superba* (Kalihari). *Journal of Research on Indian Medicine* 7: 27.
1448. Tezuka, T., Kusuda, S., Higashida, T., Matsumura, H., Horikawa, T. & Tamaki, A., 1993. The clinical effects of mugwort extract on pruritic skin lesions. *Skin Research* 35(2): 303-311. (in Japanese)
1449. Thabrew, M.I., de Silva, K.T., Labadie, R.P., de Bie, P.A. & van der Berg, B., 1991. Immunomodulatory activity of three Sri-Lankan medicinal plants used in hepatic disorders. *Journal of Ethnopharmacology* 33(1-2): 63-66.
1450. Thakur, D.K., Mishra, S.K. & Choudhuri, P.C., 1983. The use of *Leucas aspera* Spreng. and *Curcuma longa* L. extracts in the treatment of *Trichophyton verrucosum* ringworm in cattle. *Sri Lanka Veterinary Journal* 31(1-2): 37-39.
1451. Thamlikitkul, V., Bunyapraphatsara, N., Riewpaiboon, W., Therapong, S., Chautrakul, C., Thanaveerasuman, T., Nimitnon, S., Wongkonkaptape, S., Riewpaiboon, A. & Tenambergen, E., 1991. Clinical trial of *Aloe vera* Linn for treatment of minor burns. *Siriraj Hospital Gazette* 43: 413-416.
1452. Thamlikitkul, V. et al., 1989. Randomized double blind study of *Curcuma domestica* Val. for dyspepsia. *Journal of the Medical Association of Thailand* 72(11): 613-620.
1453. Thamlikitkul, V. et al., 1991. Efficacy of *Andrographis paniculata* Nees for pharyngotonsillitis in adults. *Journal of the Medical Association of Thailand* 74(10): 437-442.
1454. Thebtaranonth, C., Thebtaranonth, Y., Wanauppathamkul, S. & Yuthavong, Y., 1995. Antimalarial sesquiterpenes from tubers of *Cyperus rotundus*. *Phytochemistry* 40: 125-128.
1455. Theresa, Y.M., Rajadurai, S., Sastry, K.N.S. & Nayudamma, Y., 1967. Biosynthesis of tannins in indigenous plants. XIII. Occurrence of a new gallotannin, amlaic acid, in amla leaves (*Phyllanthus emblica*). *Chemical Abstracts* 67: 8304-8305.
1456. Theresa, Y.M., Sastry, K.N.S. & Nayudamma, Y., 1969. Amla (*Phyllanthus emblica*) tannins. I. Isolation of trigalloylglucose, terchebin, corilagin and ellagic acid from amla fruits. *Chemical Abstracts* 71: 70-71.
1457. Theuns, H.G., Theuns, H.L. & Lousberg, R.J.J.C., 1986. Search for new natural sources of morphinans. *Economic Botany* 40(4): 485-497.
1458. Thresiamma, K.C., George, J. & Kuttan, R., 1996. Protective effect of curcumin, ellagic acid and bixin on radiation induced toxicity. *Indian Journal of Experimental Biology* 34(9): 845-847.
1459. Thyagarajan, S.P., Subramanian, S., Thirunalasundari, T., Venkateswaran, P.S. & Blumberg, B.S., 1988. Effect of *Phyllanthus amarus* on chronic carriers of hepatitis B virus. *The Lancet* 332(8614): 764-766.
1460. Tirel-Roudet, C., 1972. Loganiaceae. In: Vidal, J.E. & Galibert, Y. (Editors): *Flore du Cambodge, du Laos et du Vietnam* [Flora of Cambodia, Laos and Vietnam]. Vol. 13. Muséum National d'Histoire

- Naturelle, Paris, France. pp. 3–89.
1461. Tomimori, T., Miyaichi, Y., Imoto, Y., Kizu, H. & Namba T., 1985. Studies on the Nepalese crude drugs V. On the flavonoid constituents of the root of *Scutellaria discolor* 1. *Chemical and Pharmaceutical Bulletin (Tokyo)* 33(10): 4457–4463.
1462. Tomimori, T., Miyaichi, Y., Imoto, Y., Kizu, H. & Namba T., 1986. Studies on the Nepalese crude drugs VI. On the flavonoid constituents of the root of *Scutellaria discolor* 2. *Chemical and Pharmaceutical Bulletin (Tokyo)* 34(1): 406–408.
1463. Tomimori, T., Miyaichi, Y., Imoto, Y., Kizu, H. & Namba T., 1988. Studies on the Nepalese crude drugs XI. On the flavonoid constituents of the aerial parts of *Scutellaria discolor* Colebr. *Chemical and Pharmaceutical Bulletin (Tokyo)* 36(9): 3654–3658.
1464. Tomoda, M., Takada, K., Shimizu, N., Gonda, R. & Ohara, N., 1991. Reticuloendothelial system-potentiating and alkaline phosphatase-inducing activities of *Plantago-mucilage* A, the main mucilage from the seed of *Plantago asiatica*, and its five modification products. *Chemical and Pharmaceutical Bulletin* 39(8): 2068–2071.
1465. Toong, Y.C., Schooley, D.A. & Baker, F.C., 1988. Isolation of insect juvenile hormone III from a plant. *Nature (United Kingdom)* 333(6169): 170–171.
1466. Toriizuka, K., Nishiyama, P., Adachi, I., Kawashiri, N., Ueno, M., Terasawa, K., & Horikoshi, I., 1986. Isolation of a platelet aggregation inhibitor from *Angelicae Radix*. *Chemical and Pharmaceutical Bulletin* 34(12): 5011–5015.
1467. Torres, R.C. et al., 1995. Antibacterial essential oils from some Philippine plants. *Philippine Journal of Biotechnology* 6(1): 58–59.
1468. Townsend, C.C., 1974. Leguminales. In: Townsend, C.C. & Guest, E. (Editors): *Flora of Iraq* 3. The Ministry of Agriculture and Agrarian Reform, Baghdad, Iraq. 662 pp.
1469. Townsend, C.C., 1974. Notes on *Amaranthaceae*–2. *Kew Bulletin* 29(3): 461–475.
1470. Townsend, C.C., 1980. *Amaranthaceae*. In: Dassanayake, M.D. & Fosberg, F.R. (Editors): *A revised handbook to the flora of Ceylon*. Vol. 1. Amerind Publishing Co., New Delhi, India. pp. 1–57.
1471. Toxopeus, H.J., 1952. Studies in the breeding of *Derris elliptica* and *Derris malaccensis*. *Euphytica* 1: 34–42, 175–183.
1472. Toyokawa, S., Takeda, T., Kato, Y., Wakabayashi, K. & Ogihara, Y., 1991. Presence of protein polymorphism in karasurin, an abortifacient and anti-tumor protein, identified with physicochemical properties. *Chemical and Pharmaceutical Bulletin* 39(8): 2132–2134.
1473. Toyota, T., Fukamiya, N., Okano, M., Tagahara, K., Chang, J.J. & Lee, K.H., 1990. Antitumor agents, 118. The isolation and characterization of bruceanic acid A, its methyl ester, and the new bruceanic acids B, C, and D, from *Brucea antidysenterica*. *Journal of Natural Products* 53(6): 1526–1532.
1474. Tozyo, T., Yoshimura, Y., Sakurai, K., Uchida, N., Takeda, Y., Nakai, H. & Ishii, H., 1994. Novel antitumor sesquiterpenoids in *A. millefolium*. *Chemical and Pharmaceutical Bulletin (Tokyo)* 42: 1096–1100.

1475. Tran Cong Khanh, 1987. Beitrag zur Kenntnis der Sippenstruktur des Genus *Strychnos* L. (Loganiaceae) in der Flora Vietnams. Teil 2: Zur Gliederung der Gattung *Strychnos* in Vietnam [Contribution to the knowledge on affinity structure of the genus *Strychnos* L. (Loganiaceae) in the flora of Vietnam. Part 2: on the division of the genus *Strychnos* in Vietnam]. Feddes Repertorium 98(1-2): 75-104.
1476. Tran Dinh Ly, 1993. 1900 Loai cay co ich o Viet nam [1900 useful plant species in Vietnam]. Hanoi, Vietnam. 544 pp.
1477. Tripathi, G.S. & Tripathi, Y.B., 1991. Choleric action of andrographolide obtained from *Andrographis paniculata* in rats. Phytotherapy Research 5(4): 176-178.
1478. Troup, R.S., 1921. Silviculture of Indian trees. 3 volumes. Clarendon Press, Oxford, United Kingdom.
1479. Tsao, S.W., Ng, T.B. & Yeung, H.W., 1990. Toxicities of trichosanthin and  $\alpha$ -momorcharin, abortifacient proteins from Chinese medicinal plants, on cultured tumor cell lines. Toxicol 28(10): 1183-1192.
1480. Tsoong, P.-C. & Ma, C.-Y., 1981. A study on the genus *Sophora* Linn. Acta Phytotaxonomica Sinica 19: 1-22, 143-167.
1481. Tyler, V.E., Brady, L.R. & Robbers, J.E., 1988. Pharmacognosy. 9th Edition. Lea & Febiger, Philadelphia, United States. 519 pp.
1482. Uchiyama, T., Numata, M., Terada, S. & Hosino, T., 1993. Production and composition of extracellular polysaccharide from cell suspension cultures of *Mentha*. Plant Cell, Tissue and Organ Culture 32: 153-159.
1483. Udupihille, M. & Jiffry, M.T.M., 1986. Diuretic effect of *Aerva lanata* with water normal saline and coriander as controls. Indian Journal of Physiology and Pharmacology 30(1): 91-97.
1484. Ueda, J., Yokota, T., Takahashi, N., Yoshida, M. & Kato, J., 1986. A root growth-promoting factor, capillarol, from *Artemisia capillaris* Thunb. Agricultural and Biological Chemistry 50(12): 3083-3086.
1485. Ueno, H.M., Doyama, J.T., Padovani, C.R. & Salata, E., 1996. Efeito de *Momordica charantia* L. em camundongos infectados por *Plasmodium berghei* [Effect of *Momordica charantia* L. in mice infected with *Plasmodium berghei*]. Revista da Sociedade Brasileira de Medicina Tropical 29(5): 455-460.
1486. Umi Kalsom, Y. & Noor, H., 1995. Flavone-O-glycosides from *Tinospora crispa*. Fitoterapia 66(3): 280.
1487. Unander, D.W., 1991. Callus induction in *Phyllanthus* species and inhibition of viral DNA polymerase and reverse transcriptase by callus extracts. Plant Cell Reports 10: 461-466.
1488. Unander, D.W. & Blumberg, B.S., 1991. In vitro activity of *Phyllanthus* (Euphorbiaceae) species against the DNA polymerase of hepatitis viruses: effects of growing environment and inter- and intra-specific differences. Economic Botany 45(2): 225-242.
1489. Unander, D.W., Bryan, H.H., Lance, C.J. & McMillan Jr., R.T., 1993. Cultivation of *Phyllanthus amarus* and evaluation of variables potentially affecting yield and the inhibition of viral DNA polymerase. Economic Botany 47(1): 79-88.
1490. Unander, D.W., Bryan, H.H., Lance, C.J. & McMillan Jr., R.T., 1995. Factors affecting germination and stand establishment of *Phyllanthus*

- amarus (Euphorbiaceae). *Economic Botany* 49(1): 49–55.
1491. Ungson, L.B. & Sastrapradja, S., 1976. Variation in *Smilax* species of Java. *Biotrop Bulletin* No 12. SEAMEO Regional Centre for Tropical Biology, Bogor, Indonesia. 24 pp.
1492. Valdés III, L.J., Mislankar, S.G. & Paul, A.G., 1987. *Coleus barbatus* (C. forskohlii) (Lamiaceae) and the potential new drug forskolin (coleonol). *Economic Botany* 41(4): 474–483.
1493. Valencia, E., Madinaveitia, A., Bermejo, J., Gonzalez, A.G. & Gupta, M.P., 1995. Alkaloids from *Cassia grandis*. *Fitoterapia* 66(5): 476–477.
1494. Valeton, T., 1904. Ueber neue und unvollständig bekannte Zingiberaceae aus West-Java und Buitenzorg [About new and incompletely known Zingiberaceae from West Java]. *Bulletin de l'Institute Botanique de Buitenzorg* No 20. 99 pp.
1495. Valeton, T., 1914. Tabula CCULXXXVI. *Kaempferia undulata* T. et B. *Icones Bogorienses* 4: 240–241.
1496. Valeton, T., 1918. New notes on the Zingiberaceae of Java and the Malayan Archipelago. *Bulletin du Jardin Botanique de Buitenzorg, Série II*, 27: 1–167.
1497. Valeton, T., 1920. Some new and newnamed species of *Amomum* sectio *Euamomum* K. Sch. *Bulletin du Jardin Botanique de Buitenzorg, Série III*, 2: 354–356.
1498. van Bergen, M., 1996. Revision of *Catharanthus* G. Don. Series of revisions of Apocynaceae XLI. Wageningen Agricultural Papers 96-3: 9–46.
1499. Van Damme, P., 1989. Studie van *Euphorbia tirucalli* L., morfologie, fysiologie, teeltvoorwaarden [Study of *Euphorbia tirucalli* L., morphology, physiology, agronomy]. Thesis, Gent University, Belgium. 375 pp. + appendices.
1500. van den Berg, A.J.J., Horsten, S.F.A.J., Kettenes-van den Bosch, J.J., Kroes, B.H., Beukelman, C.J., Leeftang, B.R. & Labadie, R.P., 1995. Curcacycline A, a novel cyclic octapeptide isolated from the latex of *Jatropha curcas*. *FEBS Letters* 358: 215–218.
1501. van den Berg, A.J.J., Horsten, S.F.A.J., Kettenes-van den Bosch, J.J., Kroes, B.H. & Labadie, R.P., 1995b. Multifidin, a cyanoglucoside in the latex of *Jatropha multifida*. *Phytochemistry* 40: 597–598.
1502. van der Veen, W., Malingré, Th.M. & Zwaving, J.H., 1979. *Orthosiphon stamineus*, een geneeskruid met een diuretische werking [Orthosiphon stamineus, a medicinal plant with diuretic action]. *Pharmaceutisch Weekblad* (114)35: 965–970.
1503. van der Woerd, L.A., 1941. The native medicines of the East Indian archipelago. VII. The diuretic action of some of the most common herbs used in the Netherlands and East Indies in native medicines against diseases of the urinary system. *Geneeskundig Tijdschrift voor Nederlandsch-Indie* 81: 1963–1980.
1504. van Hall, C.J.J. & van de Koppel, C. (Editors). 1946–1950. *De landbouw in den Indischen Archipel* [Agriculture in the Indonesian Archipelago]. 4 volumes. K. van Hoeve, 's-Gravenhage, the Netherlands. Vol. 1: 423, Vol. 2: 905, Vol. 3: 784, Vol. 4: 756 pp.
1505. van Puyvelde, L., Geiser, I., Rwangabo, P.C. & Sebikali, B., 1983. Rwandese herbal remedies used against gonorrhoea. *Journal of Ethno-*

- pharmacology 8(3): 279–286.
1506. van Schaik, A.H., 1994. Growing Aloe vera for gel production: report on four years of agricultural research on Aruba. Department of Agronomy, Wageningen Agricultural University, the Netherlands & Department of Agriculture, Husbandry and Fisheries, Aruba, Dutch Caribbean. 58 pp.
1507. van Steenis, C.G.G.J., 1949. Plumbaginaceae. In: van Steenis, C.G.G.J. (General editor): *Flora Malesiana*. Series 1, Vol. 4. Noordhoff-Kolff N.V., Djakarta, Indonesia. pp. 107–112.
1508. van Steenis-Kruseman, M.J., 1953. Select Indonesian medicinal plants. Bulletin No 18. Organization for Scientific Research in Indonesia, Djakarta, Indonesia. 90 pp.
1509. van Valkenburg, J.L.C.H., 1997. Non-timber forest products of East Kalimantan – potentials for sustainable forest use. (Tropenbos Series 16). The Tropenbos Foundation, Wageningen, the Netherlands. 202 pp.
1510. van Wyk, B.E., van Rheede van Oudtshoorn, M.C.B. & Smith, G.F., 1995. Geographical variation in the major compounds of Aloe ferox leaf exudate. *Planta Medica* 61: 250–253.
1511. van Zijp, C., 1915. Beiträge zur Kenntnis der Zingiberaceen [Contributions to the knowledge of Zingiberaceae]. *Receuil des Travaux Botaniques Néerlandais* 12: 340–347.
1512. Varshney, M.D., Sharma, B.B. & Gupta, D.N., 1986. Antifertility screening of plants. Part II. Effect of ten indigenous plants on early and late pregnancy in albino rats. *Comparative Physiology and Ecology* 11(4): 183–189.
1513. Vasanth, S., Gopal, R.H. & Rao, R.B., 1990. Plant anti-malarial agents. *Journal of Scientific and Industrial Research* 49(2): 68–77.
1514. Vedavathy, S. & Rao, K.N., 1991. Antipyretic activity of six indigenous medicinal plants of Tirumala Hills, Andhra Pradesh, India. *Journal of Ethnopharmacology* 33(1–2): 193–196.
1515. Veldkamp, J.F., 1970. Oxalidaceae. In: Smitinand, T. & Larsen, K. (Editors): *Flora of Thailand*. Vol. 2. The Forest Herbarium, Royal Forest Department, Bangkok, Thailand. pp. 16–23.
1516. Venkataraman, A., 1982. Problems and prospects in ginger and turmeric. In: Nair, M.K., Premkumar, T., Ravindran, P.N. & Sarma, Y.R. (Editors): *Proceedings of the National Seminar on Ginger and Turmeric*, Calicut, April 8–9, 1980. Central Plantation Crops Research Institute, Kerala, India. pp. 5–9.
1517. Vera, R., 1993. Chemical composition of the essential oil of *Ageratum conyzoides* L. (Asteraceae) from Réunion. *Flavour and Fragrance Journal* 8(5): 257–260.
1518. Vera, R., Mondon, J.M. & Pieribattesti, J.C., 1993. Chemical composition of the essential oil and aqueous extract of *Plectranthus amboinicus*. *Planta Medica* 59(2): 182–183.
1519. Verdcourt, B., 1970. Studies in the Leguminosae-Papilionoideae for the 'Flora of Tropical East Africa': II. *Kew Bulletin* 24(2): 235–307.
1520. Verdcourt, B., 1979. A manual of New Guinea legumes. Botany Bulletin No 11. Office of Forests, Division of Botany, Lae, Papua New Guinea. 645 pp.
1521. Verheij, E.W.M. & Coronel, R.E. (Editors), 1991. *Plant Resources of*



- South-East Asia No 2. Edible fruits and nuts. Pudoc, Wageningen, the Netherlands. 446 pp.
1522. Verma, H.N. & Srivastava, A., 1985. A potent systematic inhibitor of plant virus infection from *Aerva sanguinolenta* Blume. *Current Science* (Bangalore) 54(11): 526–528.
1523. Verma, R.K., Garg, B.D., Kharole, M.U. & Ahmad, A., 1981. Chronic toxicity studies on *Kalanchoe integra* in sheep. *Indian Journal of Animal Sciences* 51(5): 522–526.
1524. Verma, S.C. & Misra, P.N., 1989. Biomass and energy production in coppice stands of *Vitex negundo* L. in high density plantations on marginal lands. *Biomass* 19(3): 189–194.
1525. Vidal, J., 1962. Noms vernaculaires de plantes en usage au Laos [Vernacular names of plants used in Laos]. *Ecole française d'Extrême-Orient*, Paris, France. 197 pp.
1526. Viguier, R., 1923. Araliacées [Araliaceae]. In: Gagnepain, F. (Editor): *Flore générale de l'Indo-Chine* [General flora of Indo-China]. Vol. 2. Masson & Cie, Paris, France. pp. 1158–1182.
1527. Vijaya, K., Ananthan, S. & Nalini, R., 1995. Antibacterial effect of theaflavin, polyphenon 60 (*Camellia sinensis*) and *Euphorbia hirta* on *Shigella* spp. – a cell culture study. *Journal of Ethnopharmacology* 49(2): 115–118.
1528. Vinayagamooty, T., 1982. Antibacterial activity of some medicinal plants of Sri Lanka. *Ceylon Journal of Science, Biological Sciences* 15(1–2): 50–59.
1529. Viswanathan, S., Kulenthavel, P., Nazimudeen, S.K. & Vinayakam, T., 1981. Effect of apigenin 7,4'-di-o-methyl-ether from *Andrographis paniculata* on experimentally induced ulcers. *Indian Journal of Pharmaceutical Sciences* 43: 159–161.
1530. Vongverachai, C., 1985. Phon khong 2,4-D lae kinetin to kan phalit callus khong phut samunphrai bang chanit [Effects of 2,4-D and kinetin on callus production of some medicinal herbs]. Chiang Mai University, Thailand. 129 pp.
1531. Vu Ngoc Lo et al., 1984. Nhung ket qua buoc nghien cuu Xuyen tam lien (*Andrographis paniculata*) [Preliminary results of the study of Xuyen tam lien' (*Andrographis paniculata*)]. *Tap chi Duoc hoc* 2: 5–7. (in Vietnamese)
1532. Vu Van Chuyen, 1985. Scientific name of 'cai troi'. *Tap chi Duoc hoc* 2: 15. (in Vietnamese)
1533. Vu Van Dien & Pham Xuan Sinh, 1993. On the difference between *C. rotundus* L. and *C. stoloniferus* Retz. *Pharmaceutical Journal* (Hanoi) 4: 20–21. (in Vietnamese)
1534. Vu Xuan Phuong, 1995. Family Lamiaceae L. in *Flora of Vietnam*. *Journal of Biology* 17(4): 33–46. (in Vietnamese)
1535. Wagner, H., 1989. Search for new plant constituents with potential antiphlogistic and antiallergic activity. *Planta Medica* 55(3): 235–241.
1536. Wagner, K., Fischer, M. & Lotter, H., 1985. Isolation and structure determination of daigremontianin, a novel bufadienolide from *Kalanchoe daigremontiana*. *Planta Medica* 51: 169–170.
1537. Wahi, P., 1978. Ecology of *Andrographis paniculata* – phenology and

- growth habit. *Indian Journal of Ecology* 5: 43-47.
1538. Wahi, S.P., Wahi, A.K., Khosa, R.L. & Sharma, M., 1977. A simple method of distinguishing *Holarrhena antidysenterica* from its adulterant, *Wrightia tomentosa*. *Indian Journal of Pharmacy* 39(1): 25.
1539. Walker, E.H., 1976. *Flora of Okinawa and the southern Ryukyu Islands*. Smithsonian Institution Press, Washington D.C., United States. 1159 pp.
1540. Walters, T.W. & Decker-Walters, D.S., 1988. Notes on Economic plants. Balsampear (*Momordica charantia*, Cucurbitaceae). *Economic Botany* 42: 286-287.
1541. Wambebe, C. & Amosun, L., 1984. Some neuromuscular effects of the crude extracts of the leaves of *Abrus precatorius*. *Journal of Ethnopharmacology* 11: 49-58.
1542. Wang, B.H., Ternai, B & Polya, G., 1997. Specific inhibition of cyclic AMP dependent protein kinase by warangalone and robustic acid. *Phytochemistry* 44: 787-796.
1543. Wang, C.M., Ohta, S. & Shinoda, M., 1990. Studies on chemical protectors against radiation. XXXII. Protective effects of methanol extracts of various Taiwan crude drugs on radiation. *Yakugaku Zasshi* 110(11): 885-889. (in Japanese)
1544. Wang, D.W. & Zhao, H.Y., 1993. Experimental studies on prevention of atherosclerotic arterial stenosis and restenosis after angioplasty with *Andrographis paniculata* Nees and fish oil. *Journal of the Tongji Medical University* 13(4): 193-198.
1545. Wang, H.B., Zheng, Q.Y., Qian, D.H., Fang, J. & Ju, D.W., 1993. Effects of *Phytolacca acinosa* polysaccharides I on immune function in mice. *Acta Pharmacologica Sinica* 14(3): 243-246.
1546. Wang, Q.C., Ying, W.B., Xie, H., Hang, Z.C., Yang, Z.H. & Ling, L.Q., 1991. Trichosanthin-mono-clonal antibody conjugate specifically cytotoxic to human hepatoma cells in vitro. *Cancer Research* 51(13): 3353-3355.
1547. Wang, S.-C. & Huffman, J.B., 1981. Botanochemicals: supplements to petrochemicals. *Economic Botany* 35(4): 369-382.
1548. Wang, Z.Q., 1992. Combined therapy of brain metastasis in lung cancer. *Chung Kuo Chung Hsi I Chieh Ho Tsa Chih* 12(10): 581, 609-610. (in Chinese)
1549. Wang, Z.W., Shi, D.W., Chen, Z.D., Zheng, L.X. & Chen, C.Y., 1993. Quantitative determination of curculigoside in rhizomes of *Curculigo orchoides* by TLC-densitometry. *Acta Academiae Medicinae Shanghai* 20(1): 55-58. (in Chinese)
1550. Wariso & Rifai, M.A., 1983. Identitas jamur parasit daun *Mimosa pudica* [The identity of a parasitic fungus on *Mimosa pudica*]. *Berita Biologi* 2(7): 149.
1551. Wat, C.K., Biswas, R., Graham, E., Bohm, L. & Towers, G.H.N., 1978. UV-mediated antibiotic activity of phenylheptatryne in *Bidens pilosa*. *Planta Medica* 33(3): 309-310.
1552. Watanabe, K. et al., 1986. Antiulcer activity of extracts and isolated compounds from Zedoary *Gajutsu* cultivated in Yakushima, Japan. *Yakugaku Zasshi* 106(12): 1137-1142. (in Japanese)
1553. Watanabe, K., Takada, Y., Matsuo, N. & Nishimura, H., 1995. *Rotundial*,

- a new natural mosquito repellent from the leaves of *Vitex rotundifolia*. *Bioscience, Biotechnology and Biochemistry* 59(10): 1979–1980.
1554. Watt, J.M. & Breyer-Brandwijk, M.G., 1962. The medicinal and poisonous plants of southern and eastern Africa – being an account of their medicinal and other uses, chemical composition, pharmacological effects and toxicology in man and animal. Livingstone, Edinburgh, United Kingdom. 1457 pp.
1555. Webster, G.L., 1956–1958. A monographic study of the West Indian species of *Phyllanthus*. *Journal of the Arnold Arboretum* 37: 91–122, 217–268, 340–359; 38: 51–80, 170–198, 295–373; 39: 49–100, 111–212.
1556. Webster, G.L., 1986. A revision of *Phyllanthus* (Euphorbiaceae) in Eastern Melanesia. *Pacific Science* 40: 88–105.
1557. Webster, G.L., 1994. Synopsis of the genera and suprageneric taxa of Euphorbiaceae. *Annals of the Missouri Botanical Garden* 81: 33–144.
1558. Weenen, H., Nkunya, M.H.H., Bray, D.H., Mwasumbi, L.B., Kinabo, L.S. & Kilimali, V.A.E.B., 1990. Antimalarial activity of Tanzanian medicinal plants. *Planta Medica* 56: 368–370.
1559. Weenen, H., Nkunya, M.H.H., Bray, D.H., Mwasumbi, L.B., Kinabo, L.S., Kilimali, V.A.E.B. & Wijnberg, J.B.P.A., 1990. Antimalarial compounds containing an  $\alpha,\beta$ -unsaturated carbonyl moiety from Tanzanian medicinal plants. *Planta Medica* 56: 371–373.
1560. Wehtje, G.R., Gilliam, C.H. & Reeder, J.A., 1992. Germination and growth of leafhopper (*Phyllanthus urinaria*) as affected by cultural conditions and herbicides. *Weed Technology* 6: 139–143.
1561. Weidelt, H.J. (Editor), 1976. Manual of reforestation and erosion control for the Philippines. Schriftenreihe No 22. Deutsche Gesellschaft für Technische Zusammenarbeit (GTZ) GmbH, Eschborn, Germany. 569 pp.
1562. Werawattanametin, K., Podimuang, V. & Suksamrarn, A., 1986. Ecdysteroids from *Vitex glabrata*. *Journal of Natural Products* 49(2): 365–366.
1563. Westphal, E. & Jansen, P.C.M. (Editors), 1989. Plant Resources of South-East Asia, A selection. Pudoc, Wageningen, the Netherlands. 322 pp.
1564. Whitmore, T.C. & Ng, F.S.P. (Editors), 1972–1989. Tree flora of Malaya. A manual for foresters. 2nd Edition. 4 volumes. Malayan Forest Records No 26. Longman Malaysia Sdn. Berhad, Kuala Lumpur & Petaling Jaya, Malaysia.
1565. Whittington, D.P. & Zehr, E.I., 1992. Populations of *Criconebella xenoplax* on peach interplanted with certain herbaceous plants. *Journal of Nematology* 24(4, Supplement): 688–692.
1566. Wichtl, M. (Editor), 1994. Herbal drugs and phytopharmaceuticals. CRC Press, Boca Raton, Florida, United States. 800 pp.
1567. Wickens, G.E., 1973. Combretaceae. In: Polhill, R.M. (Editor): *Flora of Tropical East Africa*. A.A. Balkema, Rotterdam, the Netherlands. 99 pp.
1568. Wickens, G.E., 1982. Miscellaneous notes on *Crassula*, *Bryophyllum* and *Kalanchoe*. *Studies in the Crassulaceae for the Flora of Tropical East Africa*: 3. *Kew Bulletin* 36(4): 665–674.
1569. Wickens, G.E., 1987. Crassulaceae. In: Polhill, R.M. (Editor): *Flora of Tropical East Africa*. A.A. Balkema, Rotterdam, the Netherlands &

- Boston, United States. 67 pp.
1570. Wijayakusuma, H.M.H., Dalimartha, S. & Wirian, S.W., 1994. Tanaman berkhasiat obat di Indonesia [Plants yielding medicine in Indonesia]. Vol. 3. Pustaka Kartini, Jakarta, Indonesia. 143 pp.
1571. Wijayakusuma, H.M.H., Dalimartha, S., Wirian, S.W., Yaputra, T. & Wibowo, B., 1993. Tanaman berkhasiat obat di Indonesia [Plants yielding medicine in Indonesia]. Vol. 2. Pustaka Kartini, Jakarta, Indonesia. 138 pp.
1572. Wijayakusuma, H.M.H., Wirian, S.W., Yaputra, T., Dalimartha, S. & Wibowo, B., 1992. Tanaman berkhasiat obat di Indonesia [Plants yielding medicine in Indonesia]. Vol. 1. Pustaka Kartini, Jakarta, Indonesia. 122 pp.
1573. Wijesekera, R.O.B., 1991. The medicinal plant industry. CRC Press, Boca Raton, Florida, United States. 269 pp.
1574. Wijnands, D.O., 1983. The Botany of the Commelins. A.A. Balkema, Rotterdam, the Netherlands. 232 pp.
1575. Wijnands, D.O., 1989. De correcte citering van Aloe vera en Aloe succotrina [The correct citation of Aloe vera and Aloe succotrina]. Succulenta 65(3): 69–71.
1576. Wild, H., 1969. The Compositae of the Flora Zambesiaca area, 2. *Kirkia* 7: 121–135.
1577. Williams, C.A., Goldstone, F. & Greenham, J., 1996. Flavonoids, cinnamic acids and coumarins from the different tissues and medicinal preparations of *Taraxacum officinale*. *Phytochemistry* 42(1): 121–127.
1578. Williams, L.A.D., 1997. Angiotensin converting enzyme inhibiting and anti-dipsogenic activities of *Euphorbia hirta* extracts. *Phytotherapy Research* 11(5): 401–402.
1579. Williams, L.A.D. & Mansingh, A., 1993. Pesticidal potentials of tropical plants – I. Insecticidal activity in leaf extracts of sixty plants. *Insect Science and Its Application* 14(5–6): 696–700.
1580. Wills, S., 1995. The use of cannabis in multiple sclerosis. *Pharmaceutical Journal* 255: 237–238.
1581. Wilmot-Dear, C.M., 1976. Plumbaginaceae. In: Polhill, R.M. (Editor): *Flora of Tropical East Africa. Crown Agents from Oversea Governments and Administrations*, London, United Kingdom. 12 pp.
1582. Wilson, A.K., 1981. *Euphorbia heterophylla*: a review of distribution, importance and control. *Tropical Pest Management* 27: 32–38.
1583. Winde, E., Echaust, I. & Hänsel, R., 1961. *Verbena officinalis*: Vorkommen von Adenosin und  $\beta$ -Carotin [Verbena officinalis: occurrence of adenosine and  $\beta$ -carotene]. *Archiv der Pharmazie* 294: 220–229.
1584. Winter, K. & Smith, J.A.C. (Editors), 1996. Crassulacean acid metabolism: biochemistry, ecophysiology and evolution. *Ecological Studies*, Vol. 114. Springer Verlag, Berlin, Germany. 449 pp.
1585. Woerdenbag, H.J., Moskal, T.A., Pras, N., Malingré, T.M., El-Feraly, F.S., Kampinga, H.H. & Konings, A.W.T., 1993. Cytotoxicity of artemisinin-related endoperoxides to Ehrlich ascites tumour cells. *Journal of Natural Products* 56: 849–856.
1586. Womersley, J.S. (Editor), 1978. *Handbooks of the flora of Papua New Guinea*. Vol. 1. Melbourne University Press, Carlton, Australia. 278 pp.

1587. Wong, B.Y.Y., Lau, B.H.S., Tadi, P.P. & Teel, R.W., 1992. Chinese medicinal herbs modulate mutagenesis, DNA binding and metabolism of aflatoxin B1. *Mutation Research* 279(3): 209–216.
1588. Wong, B.Y.Y., Lau, B.H.S., Yamasaki, T. & Teel, R.W., 1993. Modulation of cytochrome P-450IA1-mediated mutagenicity DNA binding and metabolism of benzo-a-pyrene by Chinese medicinal herbs. *Cancer Letters* 68(1): 75–82.
1589. Wong, C.M., Ng, T.B. & Yeung, H.W., 1985. Screening of *Trichosanthes kirilowii*, *Momordica charantia* and *Cucurbita maxima* (Family Cucurbitaceae) for compounds with antilipolytic activity. *Journal of Ethnopharmacology* 13(3): 313–321.
1590. Wong, K.C. & Tan, G.L., 1994. Essential oil of *Centella asiatica* (L.) Urb. *Journal of Essential Oil Research* 6(3): 307–309.
1591. Wong, K.M., 1989. *Ixora* Linn. In: Ng, F.S.P. (Editor): *Tree flora of Malaya*. Vol. 4. Longman Malaysia, Petaling Jaya, Selangor, Malaysia. pp. 356–364.
1592. Wong, N.S., Dong, X.T., Ng, T.B., Choi, W.T. & Yeung, H.W., 1996.  $\alpha$ -Kirilowin, a novel ribosome-inactivating protein from seeds of *Trichosanthes kirilowii* (family Cucurbitaceae): a comparison with  $\beta$ -kirilowin and other related proteins. *International Journal of Peptide and Protein Research* 47(1–2): 103–109.
1593. Wong, S.M., Swanson, M.M. & Harrison, B.D., 1993. A geminivirus causing vein yellowing of *Ageratum conyzoides* in Singapore. *Plant Pathology* 42(1): 137–139.
1594. Wong, S.M., Wong, M.M., Seligmann, O. & Wagner, H., 1989. New anti-hepatotoxic naphtho-pyrone glycosides from the seeds of *Cassia tora*. *Planta Medica* 55: 276–280.
1595. Wongsatit, C., 1992. Genetic and taxonomic studies for the improvement of high alkaloid producing progenies in Madagascar Periwinkle (*Catharanthus roseus* (L.) G.Don). PhD thesis, Department of Agronomy, Graduate School, Kasetsart University, Bangkok, Thailand.
1596. Woodward, F.I., 1997. Life at the edge: a 14-year study of a *Verbena officinalis* population's interactions with climate. *Journal of Ecology* 85: 899–906.
1597. Wu, J. & Xie, F.S., 1987. Dimethyl-1-curine dimethochloride, an alternative for D-tubocurarine chloride. *Chinese Medical Journal* 100(3): 173–176.
1598. Wu, K.M., 1962. Chemical analysis and animal experimentation on *Quisqualis indica*. *National Medical Journal of China* 12: 161–170.
1599. Wu, M.A., He, C.H., Guo, Z.H. & Sun, B.S., 1982. Clinical efficacy of extracts of radix *Trichosanthes* and *Wikstroemia chamaedaphne* on midterm labour induction. Analysis of 248 cases. *Chung Kuo I Hsueh Ko Hsueh Yuan Hsueh Pao* 4(4): 241–242.
1600. Wu, T.S., Hsu, H.C., Wu, P.L., Teng, C.M. & Wu, Y.C., 1998. Rhinacanthin-Q, a naphthoquinone from *Rhinacanthus nasutus* and its biological activity. *Phytochemistry* 49(7): 2001–2003.
1601. Xu, S.B., Chen, W.F., Liang, H.Q., Lin, Y.C., Deng, Y.J. & Long, K.H., 1993. Protective action of blumeatin against experimental liver injuries. *Acta Pharmacologica Sinica* 14(4): 376–378.

1602. Xu, S.B., Chung, J.Y., Hsien, S.Y. & Sun, Y.S., 1980. Cardio-cerebrovascular effect of total flavonoids from *Desmodium styracifolium* (Osbeck) Merr. *Chung Tsao Yao* 11: 265-267.
1603. Yadav, S.K., 1987. Protection against radiation induced chromosome damage by *Embllica officinalis* fruit extract. *Caryologia* 40(3): 261-266.
1604. Yamagishi, T., Haruna, M., Yan, X.Z., Chang, J.J. & Lee, K.H., 1989. Antitumor agents, 110. Bryophyllin B, a novel potent cytotoxic bufadienolide from *Bryophyllum pinnatum*. *Journal of Natural Products* 52(5): 1071-1079.
1605. Yamagishi, T., Yan, X.Z., Wu, R.Y., McPhail, D.R., McPhail, A.T. & Lee, K.H., 1988. Structure and stereochemistry of bryophyllin-A, a novel potent cytotoxic bufadienolide orthoacetate from *Bryophyllum pinnatum*. *Chemical and Pharmaceutical Bulletin* 36(4): 1615-1617.
1606. Yamahara, J., Kobayashi, G., Matsuda, H., Katayama, T. & Fujimura, H., 1989a. Vascular dilatatory action of *Artemisia capillaris* bud extracts and their active constituent. *Journal of Ethnopharmacology* 26: 129-136.
1607. Yamahara, J., Kobayashi, G., Matsuda, H., Katayama, T. & Fujimura, H., 1989b. The effect of scoparone, a coumarin derivative isolated from the Chinese crude drug *Artemisiae Capillaris Flos*, on the heart. *Chemical and Pharmaceutical Bulletin* 37(5): 1297-1299.
1608. Yamahara, J., Mochizuki, M., Chisaka, T., Fujimura, H. & Tamai, Y., 1990. The antiulcer action of *Sophora* and the active constituent in *Sophora*. II. The antiulcer action of vexibinol. *Chemical and Pharmaceutical Bulletin (Tokyo)* 38(4): 1039-1044.
1609. Yamahara, J., Mochizuki, M., Fujimura, H., Takaishi, Y., Yoshida, M., Tomimatsu, T. & Tamai, Y., 1990. Antiulcer action of *Sophora flavescens* root and an active constituent. I. *Journal of Ethnopharmacology* 29(2): 173-177.
1610. Yamamoto et al., 1981. Chemical constituents of cultured cells of *Euphorbia tirucalli* and *millii*. *Plant Cell Reports* 1(1): 29-30.
1611. Yamamura, Y. & Matsui, M., 1985. Alkaloids from the fruits of *Stephania japonica* 2. Structures of oxostephabenine and N,O-dimethyl-oxostephine. *Journal of Natural Products* 48(5): 746-750.
1612. Yamasaki, N., Absar, N. & Funatsu, G., 1988. States and roles of tryptophan residues in *Abrus precatorius* agglutinin. *Agricultural and Biological Chemistry* 52(2): 569-575.
1613. Yamatake, Y., Shibata, M. & Nagai, M., 1976. Pharmacological studies on root bark of mulberry tree (*Morus alba* L.). *Japanese Journal of Pharmacology* 26(4): 461-469.
1614. Yang, L.Q., Singh, M., Yap, E.H., Ng, G.C., Xu, H.X. & Sim, K.Y., 1996. In vitro response of *Blastocystis hominis* against traditional Chinese medicine. *Journal of Ethnopharmacology* 55(1): 35-42.
1615. Yang, Q.Z. & Lin, L.R., 1981. Mode of action of dimethyl-levo curine dimethochloride on neuro-muscular transmission. *Acta Pharmacologica Sinica* 2(1): 19-23. (in Chinese)
1616. Yang, Z.Q., Zhang, M.Y., Liu, J.J., Hu, Z.J., Zhu, B.L., Liu, Y.W., Wang, G.Z., Wan, M. & Wu, X.L., 1989. Extraction of effective parts of *Alternanthera philoxeroides* (Mart.) Griseb. and its antiviral effect. *Chung Kuo Chung Yao Tsa Chih* 14: 488-490, 511-512. (in Chinese)

1617. Yano, K., 1975. Variation in acetylene content of different ecotypes of *Artemisia capillaris*. *Phytochemistry* 14(8): 1783-1784.
1618. Yano, K., 1983. Insect antifeeding phenylacetylenes from growing buds of *Artemisia capillaris*. *Journal of Agricultural and Food Chemistry* 31(3): 667-668.
1619. Yano, K., 1987. Minor components from growing buds of *Artemisia capillaris* that act as insect antifeedants. *Journal of Agricultural and Food Chemistry* 35(6): 889-891.
1620. Yano, K. & Ishizu, T., 1994. Capillen, a seed germination inhibitor from *Artemisia capillaris* roots. *Phytochemistry* 37(3): 689-690.
1621. Yao, Q.Q. & Zuo, C.X., 1993. Chemical studies on the constituents of *Phyllanthus urinaria* L. *Acta Pharmaceutica Sinica* 28(11): 829-835. (in Chinese)
1622. Yasni, S. et al., 1993. Effects of *Curcuma xanthorrhiza* Roxb. and curcuminoids on the level of serum and liver lipids serum apolipoprotein A-I and lipogenic enzymes in rats. *Food and Chemical Toxicology* 31(3): 213-218.
1623. Yasni, S. et al., 1994. Identification of an active principle in essential oils and hexane-soluble fractions of *Curcuma xanthorrhiza* Roxb. showing triglyceride-lowering action in rats. *Food and Chemical Toxicology* 32(3): 273-278.
1624. Yasni, S., Imaizumi, K. & Sugano, M., 1991. Effects of an Indonesian medicinal plant *Curcuma xanthorrhiza* Roxb. on the levels of serum glucose and triglyceride fatty acid desaturation and bile acid excretion in streptozotocin-induced diabetic rats. *Agricultural and Biological Chemistry* 55(12): 3005-3010.
1625. Yasukawa, K., Akihisa, T., Tamura, T. & Takido, M., 1994. Inhibitory effect of karounidiol on 12-*O*-tetradecanoylphorbol-13-acetate-induced tumor promotion. *Biological and Pharmaceutical Bulletin* 17(3): 460-462.
1626. Yeh, S.F., Hong, C.Y., Huang, Y.L., Liu, T.Y., Choo, K.B. & Chou, C.K., 1993. Effect of an extract from *Phyllanthus amarus* on hepatitis B surface antigen gene expression in human hepatoma cells. *Antiviral Research* 20(3): 185-192.
1627. Yeung, H.W., Feng, Z., Li, W.W., Cheung W.K. & Ng, T.B., 1987. Abortifacient activity in leaves, roots and seeds of *Phytolacca acinosa*. *Journal of Ethnopharmacology* 21(1): 31-36.
1628. Yeung, H.W. & Li, W.W., 1987.  $\beta$ -Trichosanthin, a new abortifacient protein from the Chinese drug wangua, *Trichosanthes cucumeroides*. *International Journal of Peptide and Protein Research* 29(3): 289-292.
1629. Yeung, H.W., Ng, T.B., Wong, N.S. & Li, W.W., 1987. Isolation and characterization of an abortifacient protein momorcochin from root tubers of *Momordica cochinchinensis*, family Cucurbitaceae. *International Journal of Peptide and Protein Research* 30(1): 135-140.
1630. Yeung, H.W., Wong, D.M. & Li, W.W., 1980. A galactose-binding lectin from the Chinese herb, Tien-Hua-Fen, *Trichosanthes kirilowii*. In: 4th Asian Symposium on Medicinal Plants and Spices: Abstracts. Department of Chemistry, Faculty of Science, Mahidol University, Bangkok, Thailand. p. 94.
1631. Yi, Y.H. & Dai, F.B., 1991. A new triterpenoid and its glycoside from

- Phytolacca esculenta*. *Planta Medica* 57(2): 162–164.
1632. Yi, Y.H. & Wang, C.L., 1989. A new active saponin from *Phytolacca esculenta*. *Planta Medica* 55(6): 551–552.
1633. Yi, Y.J., Cao, Z.Z., Yang, W.H., Hong, W.Q., Cao, Y. & Leng, Z.K., 1995. Chemical studies of *Smilax glabra* (III). Isolation and identification of smiglanin from *Smilax glabra*. *Acta Pharmaceutica Sinica* 30(9): 718–720.
1634. Yin, X.J., Liu, D.X., Wang, H.C. & Zhou, Y., 1991. A study on the mutagenicity of 102 raw pharmaceuticals used in Chinese traditional medicine. *Mutation Research* 260(1): 73–82.
1635. Yongchaiyudha, S., Rungpitarangsi, V., Bunyaphratharsara, N. & Chokechaijaroenporn, A., 1996. Antidiabetic activity of *Aloe vera* L. juice. I. Clinical trial in new cases of diabetes mellitus. *Phytomedicine* 3(3): 241–243.
1636. Yonizawa, M., 1993. Radioprotective activity in some medicinal herbs. *Shoyakugaku Zasshi* 47(3): 338–341. (in Japanese)
1637. Yoshida, Y., Wang, M.Q., Liu, J.N., Shan, B.E. & Yamashita, U., 1997. Immunomodulating activity of Chinese medicinal herbs and *Oldenlandia diffusa* in particular. *International Journal of Immunopharmacology* 19(7): 359–370.
1638. Yupraphat, T., Pachaly, P. & Zymalkowski, F., 1974. Alkaloide aus der thailändischen Menispermaceen-Droge Krung Kha Mao (*Cyclea barbata*), 3. Mitt. Isolierung und Identifizierung weiterer Bisbenzylisochinolin-Alkaloide vom Berbamin- bzw. Oxyacanthin-Typ. [Alkaloids from the Thai Menispermaceae drug krung kha mao (*Cyclea barbata*), part 3. Isolation and identification of further bisbenzylisochinoline alkaloids of the berbamine, resp. oxyacanthine type]. *Planta Medica* 25(4): 315–324.
1639. Zaczek, R. & Coyle, J.T., 1982. Excitatory amino acid analogs: neurotoxicity and seizures. *Neuropharmacology* 21(1): 15–26.
1640. Zandvoort, E.A. & Staritsky, G., 1983. In vitro propagation of *Cinchona ledgeriana*. *Tropical Crops Communications* 6. Agricultural University Wageningen, the Netherlands. 12 pp.
1641. Zapesochnaya, G.G., Kurkin, V.A., Okhanov, V.V. & Miroshnikov, A.I., 1992. Canthin-6-one and  $\beta$ -carboline alkaloids from *Aerva lanata*. *Planta Medica* 58(2): 192–196.
1642. Zapesochnaya, G.G., Kurkin, V.A., Okhanov, V.V., Pervykh, L.N. & Miroshnikov, A.I., 1991. Structure of the alkaloids of *Aerva lanata*. *Chemistry of Natural Compounds* 27(6): 725–728.
1643. Zapesochnaya, G.G., Kurkin, V.A. & Pervykh, L.N., 1990. A study of the herb *Aerva lanata* II. Feruloylamines. *Chemistry of Natural Compounds* 26(5): 590–591.
1644. Zapesochnaya, G.G., Pervykh, L.N. & Kurkin, V.A., 1991. A study of the herb *Aerva lanata*. *Chemistry of Natural Compounds* 27(3): 336–340.
1645. Zava, D.T., Dollbaum, C.M. & Blen, M., 1998. Estrogen and progestin bioactivity of foods, herbs and spices. *Proceedings of the Society for Experimental Biology and Medicine* 217(3): 369–378.
1646. Zelnik, R., Lavie, D., Levy, E.C., Wang, A.H.J. & Paul, I.C., 1977. Barbatusin and cyclobutatusin, two novel diterpenoids from *Coleus barbatus* Benth. *Tetrahedron* 33(12): 1457–1467.



1647. Zepernick, B., Langhammer, L. & Lüdcke, J.B.P., 1984. *Lexicon der ofizinellen Arzneipflanzen* [Lexicon of medicinal plants]. Walter de Gruyter & Co. Berlin, Germany. 546 pp.
1648. Zeven, A.C. & de Wet, J.M.J., 1982. *Dictionary of cultivated plants and their regions of diversity: excluding most ornamentals, forest trees and lower plants*. Pudoc, Wageningen, the Netherlands. 263 pp.
1649. Zhan, Y.Z., Tang, J.Z. & Zhang, Y.J., 1994. Study of *Andrographis paniculata* extract on platelet aggregation and release reaction and its mechanism. *Chung Kuo Chung Hsi I Chieh Ho Tsa Chih* 14(1): 28–30. (in Chinese)
1650. Zhang, C.Y. & Tan, B.K., 1996. Hypotensive activity of aqueous extract of *Andrographis paniculata* in rats. *Clinical and Experimental Pharmacology and Physiology* 23(8): 675–678.
1651. Zhang, L. & Tizard, I.R., 1996. Activation of a mouse macrophage cell line by acemannan: the major carbohydrate fraction from *Aloe vera* gel. *Immunopharmacology* 35(2): 119–128.
1652. Zhang, Z.-Y., & Lu, A.-M. & D'Arcy, W., 1994. Solanaceae. In: Wu, Z.-Y. & Raven, P.H. (Editors): *Flora of China* 17. Science Press, Beijing, China & Missouri Botanical Garden, St. Louis, United States. pp. 300–332.
1653. Zhao, H.Y. & Fang, W.Y., 1991. Antithrombotic effects of *Andrographis paniculata* Nees in preventing myocardial infarction. *Chinese Medicinal Journal of England* 104(9): 770–775.
1654. Zheng, G.Q., 1994. Cytotoxic terpenoids and flavonoids from *Artemisia annua*. *Planta Medica* 60: 54–57.
1655. Zheng, M.S., 1989. An experimental study of the anti-HSV-II action of 500 herbal drugs. *Journal of Traditional Chinese Medicine* 9(2): 113–116.
1656. Zheng, Y.T., Zhang, W.F., Ben, K.L. & Wang, J.B., 1995. In vitro immunotoxicity and cytotoxicity of trichosanthin against human normal immunocytes and leukemia-lymphoma cells. *Immunopharmacology and Immunotoxicology* 17(1): 69–79.
1657. Zhou, B.N., 1972. Preparation and application of the injection from *Trichosanthes kirilowii* Maxim. used as an abortifacient agent. *Yiyao Gongye* 7: 26–33. (in Chinese)
1658. Zhou, S., 1993. Cultivation of *Amomum villosum* in tropical forests. *Forest Ecology and Management* 60(1–2): 157–162.
1659. Zhu, L.-F., Li, Y.-H., Li, B.-L., Lu, B.-Y., Xia, N.-H. & Zhang, W.-L., 1993. *Aromatic plants and essential constituents*. Hai Feng Publishing, Hong Kong, China. 300 pp.
1660. Zhu, X. & Hu, Z., 1989. Preparation of the antiviral protein from poke-weed seeds and assay of its toxicity. *Acta Botanica Yunnanica* 11(4): 440–448.

## Acknowledgments

Our thanks are due to

- the Department of International Development Cooperation (DIDC), Finland, for financial support;
- the Commission of the European Union, DG-I Programme ‘Tropical Forests’, Brussels, Belgium, for financial support;
- the Netherlands Ministry of Agriculture, Nature Management and Fisheries for financial support;
- the Netherlands Ministry of Foreign Affairs, Directorate-General for International Cooperation (DGIS), for financial support;
- the Netherlands Ministry of Education, Culture and Science for financial support;
- the ‘Yayasan Sarana Wanajaya’, Indonesia, for financial support;
- the Indonesian Ministry of Forestry for financial support;
- Glaxo Wellcome for financial support;
- the Netherlands Association of Producers and Importers of Homeopathic Drugs (NEHOMA), for financial support;
- the Chairman of the Indonesian Institute of Sciences (LIPI), Jakarta, Indonesia, for supporting the Prosea programme, and the Research and Development Centre for Biology (RDCB), Bogor, Indonesia, for providing facilities for the Prosea Network Office in the Herbarium Bogoriense;
- the Executive Board of Wageningen Agricultural University, the Netherlands, for supporting the Prosea programme, and the Departments of Agronomy and Plant Taxonomy, for providing facilities for the Prosea Publication Office;
- the coordinating institutions of the Prosea programme in Indonesia, Malaysia, Papua New Guinea, the Philippines, Thailand and Vietnam, for providing facilities for the Prosea Country Offices;
- the Centre for Agricultural Publishing and Documentation (PUDOC-DLO), Wageningen, the Netherlands, for support and documentation facilities;
- the Prosea Country Offices in South-East Asia, for their search work on less-accessible literature, and for their editorial support concerning statistics;
- Professor Dr N.R. Farnsworth and Professor Dr H.S. Fong, College of Pharmacy, University of Illinois, Chicago, United States, for the permission to use the NAPRALERT database, and for critically reading the text of the introduction;
- Professor Dr R.P. Labadie, Nieuwegein, the Netherlands, and Professor Dr A.M. Latiff, University Kebangsaan, Bangi, Malaysia, for their contribution to the Prosea Task Force on medicinal and poisonous plants;
- Dr F.A.C.B. Adema, Rijksherbarium/Hortus Botanicus Leiden, the Netherlands, for his comments on nomenclature of *Abrus* and *Derris*;

- Mr S. Massalt, Department of Plant Cytology and Morphology, Wageningen Agricultural University, the Netherlands, for scanning the illustrations and chemical structures;
- all persons, institutions, publishers and authors mentioned in the list 'Sources of illustrations', for authorization to use these illustrations.

## Acronyms of organizations

- CITES: Convention on International Trade in Endangered Species of Wild Fauna and Flora (Lausanne, Switzerland).
- DGIS: Directorate-General for International Cooperation of the Netherlands Ministry of Foreign Affairs (Den Haag, the Netherlands).
- FAO: Food and Agriculture Organization of the United Nations (Rome, Italy).
- FRIM: Forest Research Institute Malaysia (Kepong, Malaysia).
- IEBR: Institute of Ecology and Biological Resources (Hanoi, Vietnam).
- LIPI: Indonesian Institute of Sciences (Jakarta, Indonesia).
- PCARRD: Philippine Council for Agriculture, Forestry and Natural Resources Research and Development (Los Baños, the Philippines).
- PROSEA: Plant Resources of South-East Asia (Bogor, Indonesia).
- RDCB: Research and Development Centre for Biology (Bogor, Indonesia).
- TISTR: Thailand Institute of Scientific and Technological Research (Bangkok, Thailand).
- UNITECH: Papua New Guinea University of Technology (Lae, Papua New Guinea).
- UPLB: University of the Philippines at Los Baños (Los Baños, the Philippines).
- WAU: Wageningen Agricultural University (Wageningen, the Netherlands).

## Glossary

- abaxial*: on the side facing away from the axis or stem (dorsal)
- abomasal*: pertaining to the abomasum, the fourth stomach of a ruminant animal
- abortifacient*: causing abortion; an agent that causes abortion
- abortive*: imperfectly developed; effecting an abortion (abortifacient)
- abortivum*: agent inducing abortion
- abscess*: a swollen, inflamed area in body tissues, in which pus gathers
- abstinence syndrome*: withdrawal syndrome
- acaricidal*: destroying or controlling mites
- accession*: in germplasm collections: plant material of a particular collection, usually indicated with a number
- acrescent*: increasing in size with age
- acetate pathway*: the biosynthetic route which leads to acetate, see also acetates
- acetates*: a large group of secondary metabolites, in which acetylcoenzyme A ('acetate') is the building block; acetate itself is derived from primary metabolism carbohydrates via pyruvic acid
- acetogenins*: a group of long-chain aliphatic compounds, ending with a  $\gamma$ -lactone, most often unsaturated and cyclized into one or two tetrahydrofuran rings that may or may not be adjacent
- achene*: a small dry indehiscent one-seeded fruit
- acne*: inflammatory disease affecting hair follicles and glands of the skin; frequently used to designate *acne vulgaris*, with lesions on the face, chest and back
- actinomorphic*: radially symmetrical; applied to flowers which can be bisected in more than one vertical plane
- aculeate*: furnished with prickles; prickly
- acumen*: the point of an acuminate leaf; the drip tip
- acuminate*: ending in a narrowed, tapering point with concave sides
- acute*: in botany: sharp; ending in a point with straight or slightly convex sides; in medicine: with a short and relatively severe course
- acute toxicity*: toxicity characterized by a sudden onset, sharp rise and short course
- adaptogenic*: strengthening the resistance of the body to stress
- adaxial*: on the side facing the axis (ventral)
- adenocarcinoma*: carcinoma derived from glandular tissue or in which the tumour cells form recognizable glandular structures
- adenovirus (AV)*: a virus belonging to the *Adenoviridae*, a family of DNA viruses
- adnate*: united with another part; with unlike parts fused, e.g. ovary and calyx tube
- adpressed (appressed)*: lying flat for the whole length of the organ
- adrenergic*: activated by, characteristic of, or secreting epinephrine or related substances, particularly referring to the sympathetic nerve fibres that release norepinephrine when a nerve impulse passes
- adrenoceptor, adrenoreceptor*: adrenergic receptor, i.e. postulated site on effector organs innervated by adrenergic fibres of the sympathetic nervous system
- adrenocortical hormone*: = corticosteroid
- adrenolytic*: inhibiting the action of adrenergic nerves
- adventitious*: not in the usual place, e.g. roots on stems, or buds produced in other than terminal or axillary positions on stems
- aerial root*: any root that grows above the ground
- aetiology*: the causes or origin of a disease or disorder
- aflatoxin*: a toxic factor produced by *Aspergillus flavus* and *A. parasiticus* and implicated as a cause of human hepatic carcinoma
- agglutinin*: antibody which aggregates a particular antigen; any other substance that is capable of agglutinating particles
- aglycones*: the non-sugar part of glycosides is called the aglycone part or simply the aglycone; aglycones and their glycosides may be present in the same plant; furthermore, the combination of aglycone and sugar will alter the properties of the molecule

- agonist*: a drug that has affinity for and stimulates physiological activity at cell receptors normally stimulated by naturally occurring substances (see also antagonist)
- agroforestry*: land-use systems in which trees or shrubs are grown in association with crops (agricultural crops or pastures) in a spatial arrangement or a rotation and in which there are both ecological and economic interactions between the trees and the other components of the system
- ague*: a fever of malarial character marked by paroxysms of chills, fever, and sweating that recur at regular intervals
- AIDS*: acquired immune deficiency syndrome (acquired immunodeficiency syndrome), an epidemic, transmissible retroviral disease due to infection with HIV (human immunodeficiency virus), in severe cases manifested as a profound depression of cell-mediated immunity
- ailment*: any bodily or mental disorder; illness, especially one that is not severe
- air layering*: a form of layering in which soil (rooting medium) is brought to the branch to be layered; the ball of soil in a polyethene cover is wrapped around the girdled branch; after adventitious roots grow out above the girdle, the layer can be separated
- albino*: a person, animal or plant lacking normal pigmentation
- albumen*: the nutritive material stored within the seed, and in many cases surrounding the embryo (endosperm)
- alexipharmac*: an antidote or remedy for poisoning
- alkaloids, pseudo-alkaloids*: compounds, derived from plant sources, with basic properties, containing one or more nitrogen atoms (usually in a heterocyclic ring), they usually have a marked physiological action on man or animals
- allelochemical*: a plant compound inhibiting the growth of another plant species
- allelopathy*: the reputed baneful influence of one living plant upon another due to secretion of toxic substances
- allergenic*: acting as an allergen; inducing allergy
- allergic*: pertaining to, caused by, affected with, or of the nature of allergy
- allergy*: a state of hypersensitivity induced by exposure to a particular antigen (allergen) resulting in harmful immunological reactions on subsequent exposures
- allopathic*: pertaining to or characteristic of allopathy (a system of therapeutics in which diseases are treated by producing a condition incompatible with or antagonistic to the condition to be cured or alleviated; also called heteropathy)
- alopecia*: baldness; absence of the hair from skin areas where it is normally present
- alterative*: tending to change gradually the condition of the body to a normal state; a drug having this effect
- alternate*: leaves, etc., inserted at different levels along the stem, as distinct from opposite or whorled
- alveolate*: marked as though honeycombed
- amastigote*: any of the bodies representing the morphological (leishmanial) stage in the life cycle of trypanosomatid protozoa resembling the adult form of members of the genus *Leishmania*, with the oval or round cell having a nucleus, kinetoplast and basal body, and the flagellum being very small or absent (see also: promastigote)
- amenorrhoea*: abnormal absence or suppression of the menses
- amoebiasis*: the state of being affected by amoebae, especially with *Entamoeba histolytica*
- amoebicidal*: destroying amoebae
- amplexicaul*: stem-clasping, when the base of a sessile leaf or a stipule is dilated at the base, and embraces the stem
- ampliate*: enlarged
- anabolic steroid*: a (synthetic) derivative of testosterone, having pronounced anabolic and weak androgenic properties; clinically mainly used to promote growth and repair of body tissues in senility, debilitating illness and convalescence
- anaemia*: a condition in which the blood is deficient in red blood cells, in haemoglobin, or in total volume
- anaesthesia*: loss of the ability to feel pain, caused by administration of a drug or by other medical interventions
- anaesthetic*: producing loss of sensation; producing loss of the ability to feel pain; an agent used to abolish the sensation of pain
- analeptic*: restorative, especially a stimulant to the central nervous system
- analgesia*: absence of sensibility to pain; the relief of pain without loss of consciousness
- analgesic*: relieving pain; not sensitive to pain; an agent alleviating pain without causing loss of consciousness
- anamorph*: the asexual (imperfect) stage or state of a given fungus
- anaphylaxis*: a general term, originally applied to the situation in which exposure to a toxin re-

- sulted not in immunity (prophylaxis), but in hypersensitivity; extended to include all cases of systemic anaphylaxis in response to foreign antigens and to include a range of experimental models
- anastomosis*: cross connection of branches or roots; union of one vein or parenchyma band with another, the connection forming a reticulation
- anatropous*: of an ovule, reversed, with micropyle close to the side of the hilum and the chalaza at the opposite end
- androdioecious*: having male and bisexual flowers on different plants
- androecium*: the male element; the stamens as a unit of the flower
- androgen*: any substance that promotes masculinization
- andromonoecious*: having bisexual and male flowers, but no female flowers, on the same plant
- aneuploid*: with other than the exact multiple of the haploid chromosome complement
- aneusomy*: the condition in which an organism is made up of cells containing different numbers of chromosomes
- angina pectoris*: a paroxysmal thoracic pain, often radiating to the arms. It is most often due to deficiency of blood in the myocardium and precipitated by effort or excitement
- angiogenic*: inducing the growth of new blood vessels; arising in the vascular system
- angioplasty*: elimination of areas of narrowing in blood vessels
- angiotensin*: any of a family of polypeptide hormones formed by the catalytic action of renin on renin substrate and stimulating contraction of the muscular tissue of the capillaries and arteries
- annual*: a plant which completes its life cycle in one year
- annular*: used of any organs disposed in a circle
- anodyne*: relieving pain; a medicine that relieves pain
- anorexia*: lack or loss of the appetite for food
- antagonist*: a substance that tends to nullify the action of another, as a drug that binds to a cell receptor without eliciting a biological response (see also agonist)
- antalgic*: analgesic
- anterior*: of time, previous; of place, position in front, or turned away from the axis
- anthelmintic*: destructive to worms; a drug or agent that destroys worms
- anther*: the part of the stamen containing the pollen
- anthesis*: the time the flower is expanded, or, more strictly, the time when pollination may take place
- anthocyanidins*: the aglycone part of anthocyanins, compounds closely related to the flavonoids but derived from the 2-phenyl benzopyrylium cation
- anthocyanins*: glycosides of the anthocyanidins
- anthracnose*: a disease characterized by distinctive limited lesions on stem, leaf or fruit, often accompanied by dieback and usually caused by a *Gloeosporium* or a *Colletotrichum*, imperfect fungi. The perfect state of the fungus, when known, is *Gnomonia* or *Glomerella*
- anthraquinones*: a subgroup of the quinones, in which the dione is conjugated to the condensed polycyclic aromatic system of anthracene
- anti-inflammatory*: suppressing or counteracting inflammation; an agent that suppresses or counteracts the inflammatory process
- antiarrhythmic*: preventing or alleviating arrhythmia; an agent that prevents or alleviates arrhythmia
- antibiotic*: any of a large class of substances produced by various micro-organisms and fungi and having the power of arresting the growth of other micro-organisms or destroying them; a chemical, produced by plants, animals or synthetically, having similar properties
- antibody*: an immunoglobulin molecule formed in the body in response to a foreign substance (antigen) and serving to neutralize that substance
- anticholinergic*: blocking the passage of impulses through the parasympathetic nerves; an agent that blocks the parasympathetic nerves
- anticomplementary*: reducing or destroying the power of a complement (a complex system of heat-sensitive proteins present in serum and reacting with antibodies to destroy antigens)
- antidote*: anything counteracting the effects of a poison
- antifeedant*: preventing something from being eaten
- antihepatotoxic*: counteracting injuries to the liver
- antimicrobial*: killing micro-organisms, or suppressing their growth or multiplication; an agent acting so
- antioxidant*: a substance that opposes oxidation or inhibits reactions promoted by oxygen or peroxides; many of these substances are used as preservatives in various products
- antiperiodic*: remedial of periodic diseases, as qui-

- nine for malaria; a remedy for such diseases
- antiphlogistic*: counteracting inflammation and fever; an agent counteracting inflammation and fever
- antiplasmodial*: destroying plasmodia
- antipyretic*: relieving or reducing fever; an agent that relieves or reduces fever
- antiscorbutic*: relieving or preventing scurvy; a remedy for scurvy
- antiseptic*: pertaining to asepsis (prevention of contact with micro-organisms); preventing decay or putrefaction; a substance inhibiting the growth and development of micro-organisms without necessarily killing them
- antispasmodic*: relieving spasm; an agent that relieves spasm
- antitoxic*: counteracting poison
- antitussive*: preventing or relieving cough; an agent that prevents or relieves cough
- antrorse*: directed upwards (opposed to retrorse)
- anuria*: complete suppression of urinary secretion by the kidneys
- anxiolytic*: reducing anxiety
- aperient*: a mild or gentle purgative; also called laxative
- aperture*: gap or mouth
- apex (plural apices)*: the tip or summit of an organ
- aphrodisiac*: stimulating sexual desire; a drug arousing the sexual instinct
- aphtha*: small ulcer
- apthae*: plural of aphtha; recurrent inflammation of the oral mucous membranes, characterized by the presence of small ulcers
- apical*: at the apex of any structure
- apiculate*: ending abruptly in a short point
- apocarp*: a fruit which is apocarpous
- apocarpous*: with the carpels free from each other
- apomict*: an organism reproducing by apomixis
- apomixis*: reproduction by seed formed without sexual fusion (apomictic)
- apoplexy*: sudden neurologic impairment due to a cerebrovascular disorder, either an arterial occlusion or an intracranial haemorrhage; copious extravasation of blood within any organ
- apoptosis*: fragmentation of a cell into membrane-bound particles, which are eliminated by phagocytosis
- aporphine alkaloids*: a subgroup of the alkaloids
- appendage*: a part added to another; attached secondary or subsidiary part, sometimes projecting or hanging
- appressed (adpressed)*: lying flat for the whole length of the organ
- aquaretic diuresis*: diuresis without disturbing the electrolyte balance
- arachnoid*: like a cobweb
- arboreal*: of, relating to, or resembling a tree; inhabiting or frequenting trees
- arborescent*: attaining the size or character of a tree
- areolate*: with irregular squares or angular spaces marked out on a surface, e.g. of a fruit; with small cells or cavities
- areole*: an irregular square or angular space marked out on a surface, e.g. of a fruit; a small cell or cavity
- aril*: an expansion of the funicle enveloping the seed, arising from the placenta; sometimes occurring as a pulpy cover (arillus)
- arillate*: possessing an aril
- armed*: bearing some form of spines
- arrhythmia*: any variation from the normal rhythm of the heartbeat
- arteriosclerosis*: a group of diseases characterized by thickening and loss of elasticity of arterial walls
- arthritis*: inflammation of a joint or joints
- article*: a segment of a constricted pod or fruit, as in *Desmodium*
- articulate*: jointed, or with places where separation takes place naturally
- articulation*: a joint, popularly applied to nodes of grasses
- ascariasis*: infection by the roundworm *Ascaris lumbricoides*, which is found in the small intestine and causes colicky pains and diarrhoea, especially in children
- ascendent, ascending*: curving or sloping upwards
- ascites*: effusion and accumulation of serous fluid in the abdominal cavity
- ascitic*: pertaining to or characterized by ascites
- asexual*: sexless; not involving union of gametes
- asphyxia*: pathological changes caused by lack of oxygen in respired air, resulting in hypoxia and hypercapnia
- asthma*: a chronic disorder characterized by paroxysms of the bronchi, shortness of breath, wheezing, a suffocating feeling, and laboured coughing to remove tenacious mucus from the air passages
- astrigent*: causing contraction; an agent or substance causing contraction of the skin, mucous membranes or raw or exposed tissues. As such, ethanol is used in skin-toning lotions and aluminium chlorohydrate in anti-perspirants
- ataxia*: failure of muscular coordination; irregularity of muscular action
- atonic*: lacking normal tone or strength



- atony*: lack of normal tone or strength
- attenuate*: gradually tapering
- auct.*: auctorum (Latin); of authors
- auct. non*: auctorum non (Latin); of authors not ... (author name); used after a scientific name when this name is erroneously applied by several authors to material actually belonging to a different species than the species described by the author mentioned
- auricle*: a small lobe or ear
- auriculate*: eared, having auricles
- autotriploid*: an autopolyploid with three similar sets of chromosomes
- auxin*: an organic substance characterized by its ability in low concentrations to promote growth of plant shoots and to produce other effects such as root formation and bud inhibition
- awn*: a bristle-like appendage, especially occurring on the glumes of grasses
- axil*: the upper angle between the leaf and the stem
- axile*: (placenta) belonging to or situated in an axis
- axillary*: arising from the axil
- axis*: the main or central line of development of a plant or organ
- azoospermia*: absence of spermatozoa in the semen or failure of formation of spermatozoa
- bacillary dysentery*: infectious disease caused by bacteria of the genus *Shigella*, and marked by intestinal pain, tenesmus, diarrhoea with mucus and blood in the stools, and variable toxæmia
- bactericidal*: destroying bacteria
- bactericide*: an agent that destroys bacteria
- bacteriostatic*: inhibiting the growth or multiplication of bacteria
- barbiturate*: any of a group of sedative-hypnotic agents derived from barbituric acid or thiobarbituric acid
- bark*: the tissue external to the vascular cambium collectively, being the secondary phloem, cortex and periderm
- basifixed*: attached or fixed by the base
- basophil*: granular leucocyte, from which vasoactive amines like histamine and serotonin are released on stimulation
- batik*: an Indonesian method of hand-printing textiles by coating parts of the fabric with wax to resist dye, dipping in a cold dye solution, boiling off the wax, and repeating the process for each colour used
- beak*: a long, prominent and substantial point, applied particularly to prolongations of fruits
- beaked*: used of fruits which end in a long point
- bearded*: awned; having tufts of hairs
- bechic*: a remedy or treatment of cough
- benzoquinones*: see quinones
- beri-beri*: a disease caused by a deficiency of thiamine (vitamin B1), marked by inflammatory or degenerative changes of the nerves and heart, and edema
- berry*: a juicy indehiscent fruit with the seeds immersed in pulp; usually several-seeded without a stony layer surrounding the seeds
- biconvex*: convex on both sides
- bidentate*: having two teeth; doubly dentate, as when the marginal teeth are also toothed
- biennial*: a plant which flowers, fruits and dies in its second year or season
- bifid*: forked, divided in two but not to the base
- bilabiate*: two-lipped
- bilateral (botany)*: having 2, often opposite, sides
- bilharzia*: schistosomiasis
- biliousness*: a symptom complex with nausea, abdominal discomfort, headache and constipation, formerly attributed to excessive secretion of bile
- bilocular*: with two compartments or cells
- biogenic*: having origins in biological processes
- biological half-life*: the time required for a living tissue, organ or organism to eliminate one-half of a radioactive substance which has been introduced to it
- biomimetic*: imitative of biological processes
- bipinnate*: when the primary divisions (pinnae) of a pinnate leaf are themselves pinnate
- bisbenzylisoquinoline alkaloids*: a subgroup of the isoquinoline alkaloids
- biseriate*: arranged in two rows
- bisexual*: having both sexes present and functional in the same flower
- blade*: the expanded part, e.g. of a leaf or petal
- blastogenic*: pertaining to or characterized by blastogenesis, i.e. the transformation of small lymphocytes into larger cells resembling blast cells
- blennorrhoea*: an excessive discharge of mucus; former name for gonorrhoea
- blight*: a general term applied to any of a wide range of unrelated plant diseases
- blotched*: see variegated
- bole*: the main trunk of a tree, generally from the base up to the first main branch
- bollworm*: any of several genera of moths belonging to the *Noctuidae*
- bract*: a reduced leaf subtending a flower, flower stalk or the whole or part of an inflorescence
- bracteole*: a secondary bract on the pedicel or close under the flower
- bradycardia*: slowness of the heartbeat, as evi-

- denced by a slowing of the pulse rate to less than 60 in an adult
- bradykinin*: a nonapeptide which is produced by activation of the kinin system in a range of inflammatory conditions; it is an extremely powerful vasodilator, which also increases vascular permeability, stimulates pain receptors and causes contraction of extravascular smooth muscles
- bradypnea*: abnormal slowness of breathing
- bran*: the husks or outer coats of ground corn, separated from the flour by bolting
- breeding*: the propagation of plants or animals to improve certain characteristics
- bristle*: a stiff hair or a hair-like stiff slender body
- broadcast*: to sow seed scattered, not in lines or pockets
- bronchitis*: inflammation of one or more bronchi
- bronchoconstriction*: the act or process of decreasing the diameter of a bronchus
- bronchodilation*: the act or process of increasing the diameter of a bronchus
- bronchus* (*plural bronchi*): any of the larger air passages of the lungs
- bud*: the nascent state of a flower or branch; often applied to those primordial vegetative or reproductive branches that are enclosed in a prophyllum and have a resting stage
- budding*: the process of inserting a scion, which consists of the bud in a leaf axil on a shield of rind, with or without a small piece of wood attached, into a plant (rootstock) with the intention that it will unite and grow there, usually in order to propagate a desired cultivar
- bufadienolides*: cardiac glycosides in which the side chain of the steroid aglycone is a 5-membered lactone ring
- bulb*: an underground storage organ with a much-shortened stem bearing fleshy leaf bases or scale leaves enclosing the next year's bud
- bulbil*: an aerial bulb or bud produced in a leaf axil or replacing the flower, which, on separation, is capable of propagating the plant
- bullate*: surface much blistered or puckered
- bush*: a low thick shrub without a distinct trunk
- buttress*: the enlargement of the base of trunks of tropical trees that ranges from a small spur or swelling to massive structures, partly root, partly stem, reaching as high as 10 m up the stem, thin and flat to thick, twisted or anastomose
- cachexia*: general ill health and malnutrition
- caducous*: falling off
- calcareous*: consisting of or containing chalk (calcium carbonate)
- calculus*: an abnormal concretion within the body and usually consisting of mineral salts; also called stone
- callus*: in plants, small hard outgrowth at the base of spikelets in some grasses, or tissue that forms over cut or damaged plant surface; in humans, localized hyperplasia of the horny layer of the epidermis due to pressure or friction, or an unorganized meshwork of woven bone which is formed after a fracture of a bone
- calyx*: the outer envelope of the flower, consisting of sepals, free or united
- campanulate*: bell-shaped
- canaliculate*: channelled, with a longitudinal groove
- cancer*: a malignant neoplasm or tumour, characterized by a morbid proliferation of epithelial cells in different parts of the body, resulting in progressive degeneration and often ending fatally
- cancerous*: of the nature of or pertaining to cancer
- canker*: a sunken, necrotic lesion of main root, stem or branch, due to disintegration of tissue outside the xylem cylinder, sometimes limited in extent because of host reactions resulting in overgrowth of surrounding tissues
- canopy*: the uppermost leafy layer of a tree, forest or crop
- capillaroscopy*: examination of the capillaries with the microscope
- capitate*: headed, like the head of a pin in some stigmas, or collected into compact headlike clusters as in some inflorescences
- capitulate*: with a capitulum
- capituliform*: shaped somewhat like a head
- capitulum*: a dense inflorescence of an aggregation of usually sessile flowers, as in *Compositae*
- capsule*: in botany: a dry dehiscent fruit composed of two or more carpels and either splitting when ripe into valves, or opening by slits or pores; in medicine: a structure in which something is enclosed, e.g. a hard or soft, soluble container enclosing a dose of medicine
- carbohydrates*: compounds formed from water and carbon dioxide; they can be grouped into sugars and polysaccharides
- carbuncle*: a necrotizing infection of skin and subcutaneous tissue consisting of a cluster of boils, and with multiple formed or incipient drainage sinuses; it is usually caused by *Staphylococcus aureus*
- carcinogenesis*: the production of carcinoma
- carcinogenic*: producing carcinoma
- carcinoma*: a malignant new growth consisting of

- epithelial cells, which tends to infiltrate surrounding tissues and give rise to metastases
- cardenolides*: cardiac glycosides in which the side chain of the steroid aglycone is a 5-membered lactone ring
- cardiac*: pertaining to, situated near, or affecting the heart; pertaining to the opening between the oesophagus and the stomach
- cardiac glycosides*: natural products characterized by a specific effect on myocardial contraction and atrioventricular conduction
- cardioactive*: having an effect on the heart
- cardiotonic*: having a tonic effect on the heart; an agent that has a tonic effect on the heart
- cardiotropic*: affecting, acting upon or attracted to the heart
- cardiovascular*: pertaining to the heart and blood vessels
- carina*: keel, the two inner united petals of a papilionaceous flower
- carinate*: keeled
- cariogenic*: leading to the production of caries
- carminative*: relieving flatulence; an agent relieving flatulence and assuaging pain
- carotenoids*: a subgroup of the terpenoids, containing 8 isoprene units (C<sub>40</sub>) named after β-carotene
- carpel*: one of the foliar units of a compound pistil or ovary; a simple pistil has only one carpel
- carpophore*: the part of the receptacle which is prolonged between the carpels as a central axis
- cartilaginous*: hard and tough
- caruncle*: an outgrowth of a seed near the hilum
- caryopsis*: the fruit of a grass, in which the outer layer (testa) of the seed proper is fused to the ovary wall
- castaneous*: chestnut-coloured, dark brown
- cataphyll*: vestigial or scale-like leaf present in certain seedlings on the lower stem nodes and sometimes elsewhere on the seedling stem
- cataplasma*: poultice or soft external application
- catarrh*: inflammation of the lining tissue of various organs, particularly of the nose, throat, and air passages, and characterized by an outpouring of mucus
- catch-cropping*: a form of intercropping in which a perennial crop (e.g. cocoa) is interplanted in its juvenile phase with a secondary annual or short-perennial crop (e.g. banana) to obtain income during the interim period, to check weeds, to provide shade and to control the spread of insect pests; a form of sequential cropping in which, for instance, a green manure crop precedes the main crop
- cathartic*: causing evacuation of the bowels; an agent that causes evacuation of the bowels by increasing bulk, stimulating peristaltic action etc.; also called purgative
- catkin*: a close bracteate, often pendulous spike, usually with unisexual flowers
- caudate*: with a tail-like appendage
- cauliflorous*: with the flowers borne on the trunk
- cauline*: belonging to the stem or arising from it
- cellulitis*: acute, suppurative inflammation of the deep subcutaneous tissues and sometimes muscle, usually resulting from wound infection by bacteria such as group A streptococci and *Staphylococcus aureus*
- cephalalgia*: headache
- cerebriform*: having an irregular, brain-like appearance, like the kernel of a walnut
- cerebrovascular*: pertaining to the blood vessels of the cerebrum or brain
- Chagas' disease*: a form of trypanosomiasis, occurring widely in Central and South America and caused by *Trypanosoma cruzi*
- chalcones*: a subgroup of the flavonoids
- channelled*: grooved, hollowed out like a gutter
- chartaceous*: papery
- chemiluminescence*: luminescence resulting from the direct transformation of chemical energy into light energy
- chemotype*: taxon which is morphologically similar to another one but with different chemical content
- chlorosis (in medicine)*: a disorder characterized by greenish-yellow discoloration of the skin and generally affecting adolescent females in the nineteenth century; believed to be associated with iron deficiency anaemia
- cholagogue*: an agent that promotes an increased flow of bile
- cholangitis*: inflammation of a bile duct
- cholecystitis*: inflammation of the gall bladder
- cholekinesis*: activation of the bile flow
- cholera*: acute, infectious inflammation of the intestine, caused by an enterotoxin elaborated by *Vibrio cholerae*, and characterized by severe, watery diarrhoea
- choleraogenic*: producing cholera
- choleresis*: the secretion of bile by the liver
- choleric*: stimulating the production of bile by the liver
- cholestasis*: suppression or stoppage of the bile flow
- cholinergic*: stimulated, activated or transmitted by acetylcholine; applied to the sympathetic and parasympathetic nerve fibres that liberate

- acetylcholine at a synapse when a nerve impulse passes (see also: anticholinergic)
- cholinesterase*: enzyme that catalyses the cleavage of the acyl group from various esters of choline, including acetylcholine, and several related compounds, and which occurs primarily in the serum, liver and pancreas
- choriocarcinoma*: an epithelial malignancy, formed by abnormal proliferation of certain placental epithelium cells
- chromophore*: any chemical group whose presence gives a decided colour to a compound and which unites with certain other groups to form dyes
- chromosome*: a structural unit in the nucleus which carries the genes in a linear constant order; the number is typically constant in any species
- chronic*: persisting over a long period of time
- ciliate*: with a fringe of hairs along the edge
- cincinnus*: a monochasial cymose inflorescence with branches alternating from one side of the vertical axis to the other and normally curved to one side
- circumscissile*: dehiscing or falling off along a circular line
- cirrhosis*: liver disease characterized by diffuse, interlacing bands of fibrous tissue that divide the hepatic parenchyma into nodular areas
- clavate*: club-shaped or thickened towards the end
- claw*: the basal, narrow part of a petal or sepal
- clawed*: furnished with a basal, narrow part (the claw)
- cleft*: cut halfway down
- cleistogamous*: pollination and fertilization taking place within the unopened flower
- clone*: a group of plants originating by vegetative propagation from a single plant and therefore of the same genotype
- clustered*: compactly gathered together; with several stems
- coccidial*: pertaining to coccidia, i.e. organisms of a subclass (*Coccidia*) of parasitic protozoa, found in vertebrates and higher invertebrates
- coherent*: the incorporation of one part with another, as the petals to form a tubular corolla
- cohobation*: re-use of distillation waters for the purpose of recovering dissolved essential oil
- colic*: acute, spasmodic pain in the bowels; pertaining to the bowels
- collagen*: an insoluble fibrous protein that occurs in vertebrates as the chief constituent of the connective tissues, as in skin, cartilage, bone and hair
- collar*: the boundary between the above- and underground portions of the axis of a plant
- colleter*: a multicellular glandular hair
- colliculate*: covered with small round elevations
- column (botany)*: a cylindrical body, e.g. a tube of connate stamen filaments or the central axis of a fruit
- coma*: in medicine: a state of unconsciousness from which the patient cannot be aroused; in botany: the hairs at the end of some seeds; a tuft of leafy bracts or leaves at the top of an inflorescence (e.g. pineapple)
- compatibility*: in floral biology: capable of cross- or self-fertilization; in plant propagation: stock-scion combinations resulting in a lasting union of the two parts
- compound*: of two or more similar parts in one organ, as in a compound leaf or compound fruit
- concave*: hollow
- concolourous*: similarly coloured on both sides or throughout; of the same colour as a specified structure
- condensed tannins*: see tannins
- conduplicate*: folded lengthwise
- confluent*: blended into one, passing by degrees from one into the other
- congeneric*: belonging to the same genus or forming a single genus
- conical*: having the shape of a cone (cone-shaped)
- conjunctiva*: the membrane that lines the eyelid and covers the anterior part of the eyeball
- conjunctivitis*: inflammation of the conjunctiva
- connate*: united or joined
- connective (botany)*: tissue between the pollen sacs of an anther
- connivent*: having a gradually inward direction, as in many petals (convergent)
- conspicuous*: belonging to the same species
- constipation*: a condition of the bowels in which the expulsion of waste matter is infrequent and difficult
- contorted*: twisted or bent
- contraceptive*: reducing the likelihood of or preventing conception; an agent that reduces the likelihood of or prevents conception
- covalent bond*: a chemical bond between two atoms or radicals formed by the sharing of one, two or three pairs of electrons
- convex*: having a more or less rounded surface
- convulsion*: a violent and involuntary contraction of the voluntary muscles
- coppice*: a small wood which is regularly cut at stated intervals, the new growth arising from the stools
- coralline*: pertaining to, composed of, or having

- the structure of corals, as coralline limestone
- cordate*: heart-shaped, as seen at the base of a leaf, etc., which is deeply notched
- coriaceous*: of leathery texture
- corm*: in botany: a solid, short, swollen underground stem, usually erect and tunicated, of one year's duration, with that of the next year at the top or close to the old one
- corolla*: the inner envelope of the flower consisting of free or united petals
- corrugate (corrugated)*: wrinkled
- cortex*: the bark or rind
- corticosteroid*: any of the 21-carbon steroids elaborated by the adrenal cortex (excluding sex hormones of adrenal origin) in response to the release of ACTH or angiotensin II; used clinically for hormone replacement therapy, for suppression of ACTH secretion, for suppression of immune responses and as antineoplastic, anti-allergic and anti-inflammatory agents
- corymb*: a flat-topped indeterminate inflorescence in which the branches or pedicels sprout from different points, but attain approximately the same level, with the outer flowers opening first
- corymbose*: flowers arranged to resemble a corymb
- cotyledon*: seed-leaf, the primary leaf; dicotylous embryos have two cotyledons and monocotylous embryos have one
- coumarins*: benzo- $\alpha$ -pyrone ( $\alpha$ -chromone) derivatives
- counterirritant*: producing a counterirritation, i.e. a superficial irritation in one part of the body, intended to relieve an irritation in another part; an agent which causes counterirritation
- cover crop*: a close-growing crop primarily grown for the purpose of protecting and improving soil between periods of regular crop production or between trees or vines in orchards and plantations
- crenate*: the margin notched with blunt or rounded teeth
- crenulate*: slightly crenate, with small teeth
- cross-incompatibility*: not capable of cross fertilization
- cross-pollination*: the transfer of pollen from one flower to the stigma of a flower of another plant which is not of the same clone
- crossing-over*: the process of exchange of genetic material between homologous chromosomes during meiosis, consisting of the breakage of homologous chromatids at corresponding sites and their reunion with each other
- crown*: the aerial expanse of a tree, not including the trunk; corona; a short rootstock with leaves; the base of a tufted, herbaceous, perennial grass
- crustaceous*: of hard but brittle texture
- cryopreservation*: maintaining the viability of tissue or organs by storage at very low temperatures
- crystalline*: of the nature of or relating to a crystal or crystals
- culm*: the stem of grasses and sedges
- cultigen*: a plant species or race that has arisen or is known only in cultivation
- cultivar (cv., plural cvs)*: an agricultural or horticultural variety that has originated and persisted under cultivation, as distinct from a botanical variety; a cultivar name should always be written with an initial capital letter and given single quotation marks (e.g. banana 'Gros Michel')
- cuneate*: wedge-shaped; triangular, with the narrow end at the point of attachment, as the bases of leaves or petals
- cupular*: furnished with or subtended by a cupule
- cupule*: a small cup-like structure; the cup of such fruits as the acorn, consisting of an involucre composed of adherent bracts
- cupuliform*: cupule-shaped
- cuspid*: a sharp, rigid but small point
- cuspidate*: abruptly tipped with a sharp rigid point
- cutting*: a portion of a plant, used for vegetative propagation
- cyanogenic glycosides*: a group of secondary metabolites that form hydrocyanic acid on hydrolysis
- cymbiform*: boat-shaped
- cyme*: a determinate inflorescence, often flat-topped, in which each growing point ends in a flower and the central flowers open first
- cymose*: bearing cymes or inflorescences related to cymes
- cyst*: a stage in the life cycle of certain parasites, during which they have a protective wall
- cysticidal*: destroying cysts, e.g. those of *Entamoeba histolytica*
- cystitis*: inflammation of the urinary bladder
- cystolith (botany)*: mineral concretions, usually of calcium carbonate on a cellulose stalk
- cytokine*: generic term for nonantibody proteins released by a cell population on contact with a specific antigen and acting as intercellular mediators
- cytokinin*: any of a class of phytohormones having as main functions the induction of cell division and the regulation of tissue differentiation
- cytoplasm*: the protoplasm of a cell, excluding the nucleus

- cytosol*: the liquid medium of the cytoplasm  
*cytosolic*: pertaining to or contained in the cytosol  
*cytotoxic*: pertaining to or exhibiting a destructive effect on certain cells  
*damping-off*: a disease of seeds or seedlings caused by fungi which cause various effects, from failure to germinate to the dying off of the seedling  
*deciduous*: shedding, applied to leaves, petals, etc.  
*declinate*: bent or curved downward or forward  
*decoction*: a medicinal preparation or other substance made by boiling, especially in water  
*decompound*: several times divided or compounded  
*decongestant*: an agent that reduces congestion or swelling  
*decumbent*: reclining or lying on the ground, but with the summit ascending  
*decurrent*: extending down and adnate to the petiole or stem, as occurs in some leaves  
*decussate*: of leaves, arranged in opposite pairs on the stem, with each pair perpendicular to the preceding pair  
*deflexed (reflexed)*: abruptly recurved; bent downwards or backwards  
*dehiscent*: opening spontaneously when ripe, e.g. of capsules, anthers  
*deltoid*: shaped like an equilateral triangle  
*demulcent*: allaying the irritation of abraded or inflamed body surfaces, soothing; a soothing, mucilaginous or oily medicine or application  
*density*: weight (kg) per volume (cubic m) at a certain moisture content  
*dentate*: margin prominently toothed with the pointed teeth directed outwards  
*denticulate*: minutely toothed  
*deobstruent*: having the power to remove obstructions; an agent that removes obstructions  
*depilatory*: having the power to remove the hair  
*depressant*: diminishing functional activity; a medicine or drug which lowers functional activity and vital energy in general  
*depurative*: tending to purify or cleanse  
*dermatitis*: inflammation of the skin  
*dermatomycosis*: superficial fungal infection of the skin or its appendages, with the term including dermatophytosis  
*dermatophyte*: a fungus parasitic on the skin  
*dermatophytosis*: any superficial fungal infection caused by a dermatophyte and involving the horny layer of skin, hair and nails  
*dermatosis*: any skin disease, especially one without inflammation  
*determinate*: of inflorescences, when the terminal or central flower of an inflorescence opens first and the prolongation of the axis is arrested; of shoot growth, when extension growth takes the form of a flush, i.e. only the previously formed leaf primordia unfold; for pulses also used to indicate bush-shaped plants with short duration flowering in one plane  
*diabetes*: a general term referring to disorders characterized by the excretion of excessive amounts of urine. When used alone, usually referring to diabetes mellitus, i.e. a chronic syndrome of impaired carbohydrate, protein and fat metabolism owing to insufficient secretion of insulin or tissue insulin resistance  
*diadelphous*: in two bundles  
*dialysable*: capable of dialysis or of passing through a membrane  
*dialysis*: the process of separating solutions of mixed substances of unequal diffusibility by means of membranes or septa  
*diapause*: a state of inactivity and arrested development, accompanied by greatly reduced metabolism  
*diaphoretic*: pertaining to, characterized by, or promoting (profuse) perspiration; an agent inducing sweating, having the power to increase perspiration  
*diarrhoea*: a profuse, frequent, and loose discharge from the bowels  
*dichasium (plural dichasia)*: a cymose inflorescence with 2 equal or nearly equal lateral branches arising below the terminal flower, this pattern being repeated or not (compound and simple dichasium respectively)  
*dichotomous*: forked, parted by pairs  
*dicotyledon*: angiosperm with two cotyledons or seed-leaves  
*didynamous*: with the stamens in two pairs, two long and two short ones  
*dieback*: the dying off of parts of the aboveground structure of the plant, generally from the top downward  
*digestibility*: the percentage of a foodstuff taken into the digestive tract that is absorbed into the body  
*digitate*: a compound leaf whose leaflets diverge from the same point like the fingers of a hand  
*dilated (botany)*: expanded into a flat structure  
*dimer*: a compound formed by combination of two identical simpler molecules  
*dimeric*: showing the characteristics of a dimer  
*dimerous*: with two members  
*dimorphic*: of two forms, as may occur with branches, etc.

- dimorphous*: dimorphic
- dioecious*: with unisexual flowers and with the staminate and pistillate flowers on different plants (dioecy)
- diosgenin*: a complex steroid obtained from certain species of yam and which can be converted into 16-dehydropregnenolone, one of the main active ingredients in oral contraceptives
- diploid*: with two sets (genomes) of chromosomes, as occurs in somatic or body cells; usually written  $2n$ , having twice the basic chromosome number of the haploid germ cells
- dipterocarp forest*: woodland dominated by trees belonging to the family *Dipterocarpaceae*
- disc*: = disk
- disciform*: shaped like a disk
- discoid*: resembling a disk or discus, being flat and circular, e.g. of a leaf with a round thickened lamina and rounded margins
- discutient*: causing a disappearance; an agent which causes a disappearance
- disk*: a fleshy or elevated development of the receptacle within the calyx, corolla or stamens, often lobed and nectariferous
- dispersal*: the various ways by which seeds are scattered, e.g. by wind, water or animals
- dissected*: divided into many slender segments
- distal*: situated farthest from the place of attachment
- distichous*: regularly arranged in two opposite rows on either side of an axis
- distillation*: the process of transforming (fractions of) a liquid or solid into the vapour state, and condensing the vapour back to liquid or solid, named the distillate
- distylous*: referring to flowers of a species which possess one of two style types
- diterpenes*: a subgroup of the isoprenoids, formed by coupling of 4  $C_5$  units
- diuresis*: increased discharge of urine
- diuretic*: tending to increase the flow of urine; an agent that promotes the excretion of urine
- divaricate*: extremely divergent
- domatium (plural domatia)*: a modified projection that provides shelter for other organisms
- dormancy*: a term used to denote the inability of a resting plant or plant part (e.g. the seed, bulb, tuber, or in tree crops usually the buds) to grow or to leaf out, even under favourable environmental conditions
- dorsal*: back; referring to the back or outer surface of a part or organ (abaxial)
- dorsifixed*: attached by the back, as in the case of the attachment of a filament to an anther
- double blind*: pertaining to a clinical trial or other experiment in which neither the subject nor the person administering treatment knows which treatment any particular subject is receiving
- double-flowered*: petals monstrously increased at the expense of other organs, especially the stamens
- downy*: covered with very short and weak soft hairs
- dropsy*: oedema
- drupaceous*: resembling a drupe, whether actually a drupe or not
- drupe*: a fleshy one-seeded indehiscent fruit with the seed enclosed in a strong endocarp
- drupelet*: a small drupe
- dysentery*: any of various diseases characterized by inflammation of the intestines, abdominal pain and frequent bloody, mucous faeces
- dysmenorrhoea*: painful menstruation
- dyspepsia*: a condition of disturbed digestion
- dyspeptic*: relating to or having dyspepsia
- dysphoria*: malaise, disquiet, restlessness; opposite of euphoria
- dyspnoea (dyspnea)*: laboured or difficult breathing
- dystonia*: distorted or impaired movements resulting from disordered muscle tonicity
- dystonic*: pertaining to or characterized by dystonia
- dysuria*: difficult or painful urination
- ebracteate*: without bracts
- EC<sub>50</sub>*: median effective concentration, i.e. the concentration that produces the desired effect in fifty percent of a test population
- ecbolic*: increasing uterine contractions and aiding in or hastening expulsion or delivery of child during birth; an agent acting so
- ecotype*: a biotype resulting from selection in a particular habitat
- ecto-*: in compositions, referring to the outside or the outer surface or part
- ectoparasite*: a parasite that lives on the outside of the hosts's body
- eczema*: a disease of the skin characterized by inflammation, redness, itching, and the formation of vesicles which exude a watery substance that evaporates and leaves the skin covered with crusts
- ED<sub>50</sub>*: median effective dose, i.e. the dose that produces the desired effect in fifty percent of a population
- edaphic*: pertaining to or influenced by conditions of the soil
- eglandular*: without glands

- elaiosome*: a seed or fruit outgrowth in which oil is stored. Serves as food for ants
- ellipsoid*: a solid which is elliptical in outline
- elliptical*: oval in outline but widest about the middle
- emarginate*: notched at the extremity
- embryo*: in plants, the rudimentary plant within a seed, developed from a zygote (sexual) or from other nuclei in the embryo sac or cells of the nucellus or integuments (apomictic); in animals, those derivatives of the fertilized ovum that will become the offspring, during their period of most rapid development; in humans, the developing organism from the end of the 2nd week after fertilization to the end of the 8th week
- emergent*: of a tree, one of which the crown reaches distinctly above the forest canopy; of cotyledons, becoming free from the seed coat and other external tissues
- emesis*: vomiting
- emetic*: tending to induce or cause vomiting; an agent that induces or causes vomiting
- emeto-cathartic*: an agent that is both emetic and cathartic
- emmenagogue*: a substance or measure that induces menstruation
- emollient*: soothing and softening; an agent that soothes or softens the skin or soothes an irritated internal surface
- emphysema*: pathological accumulation of air in organs or tissues, especially applied to the lungs
- encephalitis*: inflammation of the brain
- endemic*: exclusively native to a specified or comparatively small region; also used as a noun for a taxon thus distributed
- endo-*: prefix, referring to the inside or the inner surface or part
- endocarp*: the innermost layer of the pericarp or fruit wall
- endodermis*: a sheath of one or more layers of modified parenchymatous cells between the cortex and the central zone of the stem or root
- endogenous*: originating from within the organism
- endosperm*: the starchy or oily nutritive material stored within some seeds, sometimes referred to as albumen; it is triploid, having arisen from the triple fusion of a sperm nucleus and the two polar nuclei of the embryo sac
- enema*: a liquid injected into the rectum
- enfleurage*: the process of transferring the volatile compounds responsible for the scent of picked flowers to a fixed oil or fat spread out on a glass plate. Enfleurage is successful only with flowers that continue to produce aroma compounds for several hours after picking, such as jasmine and tuberose
- ensiform*: sword-shaped
- enterobiasis*: infection with nematode worms of the genus *Enterobius*, especially *E. vermicularis* (seatworm, threadworm or pinworm)
- entire (botany)*: with an even margin without teeth, lobes, etc.
- epicotyl*: the young stem above the cotyledons
- epidermis*: in plants, the true cellular skin or covering of a plant below the cuticle; in humans, the outermost and nonvascular layer of the skin
- epidermoid*: belonging to or resembling the epidermis
- epididymal*: pertaining to the epididymis
- epididymis*: the cordlike structure at the posterior part of the testis, whose coiled duct provides for storage, transit and maturation of spermatozoa
- epigeal*: above the ground; in epigeal germination the cotyledons are raised above the ground
- epigynous*: on the pistil, apparently above the ovary
- epilepsy*: any of a group of syndromes characterized by recurrent, transient disturbances of the brain function, with manifestations including unconsciousness and uncontrolled motion
- epileptic*: pertaining to epilepsy
- epimer*: either of two diastereomers that differ in the configuration around one asymmetrical carbon atom
- epipetalous*: borne upon or placed before the petals
- epiphyte*: a plant that grows on another plant but without deriving nourishment from it
- epistaxis*: nosebleed
- epitepalous*: borne upon or placed before the tepals
- epithelium*: in plants, the layer of secretory parenchymatous cells that surrounds an intercellular canal or cavity; in humans, the covering of internal and external body surfaces, including the lining of vessels and other small cavities
- epithet*: the second part of the scientific name of a species, the first part denoting the genus to which the species belongs
- erect*: directed towards summit, not decumbent
- erecto-patent*: between spreading and erect
- erysipelas*: an acute, superficial form of dermatitis, usually caused by group A streptococci and characterized by a spreading, red, hot plaque
- erythema*: name applied to skin redness produced by congestion of the capillaries
- essential oil*: a volatile product, obtained from a natural source, which agrees with that source in odour and name. In a narrow sense, only volatile products obtained by steam or water



- distillation are called essential oils
- eutrophic*: providing adequate or with a large supply of nutrition
- evergreen*: bearing foliage all year long; a plant that changes its leaves gradually
- ex situ*: in an artificial environment or unnatural habitat
- exalbuminous*: lacking albumen
- exanthem, exanthema*: a skin eruption or rash; a disease in which skin eruptions or rashes are a prominent manifestation
- exocarp*: the outer layer of the pericarp or fruit wall
- exocytosis*: the discharge from a cell of particles which are too large to diffuse through the wall
- expectorant*: promoting the ejection of mucus or other fluids from the respiratory tract; an agent tending to promote discharge of mucus or other fluids from the respiratory tract
- exsert, exserted*: protrude beyond, as stamens beyond the tube of the corolla
- exstipulate*: without stipules
- extra-axillary*: beyond or outside the axil
- extract*: a concentrated preparation of a vegetal or animal drug obtained by removing the active constituents with a suitable solvent
- extrafloral*: of nectaries, beyond the flower
- extrorse*: directed outward, as the dehiscence of an anther
- F<sub>1</sub>, F<sub>2</sub>, etc.*: symbols used to designate the first generation, second generation, etc., after a cross
- falcate*: sickle-shaped
- fallow*: land resting from cropping, often covered by natural vegetation or planted with fast growing herbs, shrubs or trees (fallow crop)
- fascicle*: a cluster of flowers, leaves, etc., arising from the same point
- fasciculate*: connected or drawn into a fascicle
- favus*: type of ringworm, usually caused by *Trichophyton schoenleinii*
- febrifuge*: an agent serving to reduce fever
- fermentation*: a chemical change accompanied by effervescence and suggestive of changes produced in organic materials by yeasts
- fertile*: in plants: capable of completing fertilization and producing seed; producing seed capable of germination; having functional sexual organs; in humans: having the capacity to reproduce; capable of developing into a new individual (said of ova)
- fertilization (biology)*: union of the gametes (egg and sperm) to form a zygote
- fibre*: in plants: any long, narrow cell of wood or bark other than vessel or parenchyma elements; in humans: an elongated, threadlike structure
- fibrinolytic*: pertaining to, characterized by or causing fibrinolysis, i.e. the dissolution of fibrin by enzymatic action
- fibroblast*: connective tissue cell
- fibroplasia*: the formation of fibrous tissue, as occurs normally in the healing of wounds and abnormally in some tissues
- fibrosarcoma*: a malignant tumour consisting of cells and fibres derived from fibroblasts
- fibrosis*: the formation of fibrous tissue
- fibrous*: composed of or containing fibres
- fig*: the fleshy multiple fruit, derived from the inflorescence of *Ficus* spp. (syconium)
- filament*: thread; the stalk supporting the anther
- filariasis*: a diseased state due to the presence of nematode worms of the superfamily *Filarioideae* in the body
- filiform*: slender; threadlike
- fimbriate*: fringed
- fissured*: provided with fissures (cracks of considerable length and depth), e.g. in the bark of some trees
- flaky*: lamelliform, in the shape of a plate or scale
- flatulence*: the presence of excessive amounts of air or gases in the intestine
- flavonoles*: a subgroup of the flavonoids
- flavanones*: a subgroup of the flavonoids
- flavones*: a subgroup of the flavonoids
- flavonoids*: a group of natural products in which the basic structure is the 2-phenyl-chromane skeleton
- flavonoles*: a subgroup of the flavonoids
- fleshy*: succulent
- floret*: a small flower, one of a cluster as in grasses or *Compositae*; a grass floret typically consists of a lemma, palea, 2 lodicules, 3 stamens and a pistil with 2 plumose stigmas
- flush*: a brief period of rapid shoot growth, with unfolding of the leaf primordia which had accumulated during the previous quiescent period
- fluted*: of a bole, with rounded grooves and folds
- fodder*: something fed to domesticated animals, especially coarse, dried food from plants (hay, straw, leaves)
- foliaceous*: leaf-like
- foliolate*: 2-, 3-, 4- etc., with 2, 3, 4 leaflets
- follicle*: in plants: a dry, unilocular fruit, dehiscing by the ventral suture to which the seeds are attached; in humans: a sac or pouchlike depression or cavity, e.g. hair follicle
- follicular atresia*: the degeneration and resorption of an ovarian follicle before it reaches maturity and ruptures

- fomentation*: treatment by the application of warm, moist substances; the substance thus applied
- forage*: grassland and fodder plants suitable as feed for herbivores, usually with lower nutrient concentration and digestibility than concentrates such as grain
- foveolate*: with small pits
- framboesia*: see yaws
- free*: neither adhering nor united
- free radical*: a radical (a group of atoms which enters into and goes out of chemical combination without change and forms one of the fundamental constituents of a molecule) which is extremely reactive, has a very short half-life, and carries an unpaired electron
- fringed*: fimbriate; with hair-like appendages along the margin
- frondose*: frond-like
- frugivorous*: feeding on fruit
- fruit*: the ripened ovary with adnate parts
- fugaceous*: withering or falling off rapidly or early
- fulminant hepatitis*: massive hepatic necrosis, usually resulting from hepatitis B or non-A, non-B hepatitis
- fulvous*: yellow, tawny
- fungicidal*: destroying fungi
- fungicide*: an agent that destroys fungi or inhibits their growth
- fungistatic*: inhibiting the growth of fungi
- fungitoxicity*: the quality of having a toxic effect on fungi
- funicle (funiculus)*: the little cord which attaches the ovule or seed to the placenta
- furuncle*: a painful nodule in the skin caused by inflammation of the dermis and subcutaneous tissue, enclosing a central core; it is caused by staphylococci which enter through hair follicles
- fusiform*: spindle-shaped; tapering towards each end from a swollen centre
- galactagogue, galactogogue*: promoting the flow of milk; an agent that promotes the flow of milk
- galactophoritis*: inflammation of the milk ducts
- galenicals*: medicines prepared according to the formulas of Galen; the term is now used to denote preparations containing one or more organic ingredients, as contrasted with pure chemical substances
- gallery forest*: fringing forest, forest growing along a watercourse in an otherwise non-forested area
- gamete*: either of two mature reproductive cells, an ovum or sperm, which in uniting produce a zygote; the malarial parasite in its sexual form
- gametocidal*: capable of destroying gametes or gametocytes
- gametocyte*: a cell capable of dividing to form gametes
- gamopetalous*: with united petals either throughout their length or at the base
- gamophyllous*: with leaves which are united by their edges
- gargle*: to rinse or medicate the throat and mouth with a liquid kept in motion by the slow expulsion of air from the lungs; a solution used for rinsing or medicating the throat and mouth
- gastralgia*: gastric colic
- gastric*: pertaining to, originating in, or affecting the stomach
- gastritis*: inflammation of the stomach
- gastro-enteritis*: acute inflammation of the lining of stomach and intestines, which may be caused by food poisoning, ingestion of irritating food or drinks, or psychological factors
- gastropathy*: any disease of the stomach
- gene*: the unit of inheritance located on the chromosome
- genetic erosion*: the decline or loss of genetic variability
- geniculate*: abruptly bent so as to resemble the knee-joint
- genin*: aglycone
- genome*: a set of chromosomes as contained within the gamete and corresponding to the haploid chromosome number of the species
- genotype*: the genetic makeup of an organism comprising the sum total of its genes, both dominant and recessive; a group of organisms with the same genetic makeup
- genus (plural genera)*: the smallest natural group containing distinct species
- geocarpic*: having the fruits mature underground
- germplasm*: the genetic material that provides the physical basis of heredity
- giardiasis*: common infection of the small intestine with the flagellate protozoan *Giardia lamblia*
- glabrescent*: becoming glabrous or nearly so
- glabrous*: devoid of hairs
- glandular*: in botany: having or bearing secreting organs or glands; in medicine: pertaining to or of the nature of a gland
- glaucoma*: a group of eye diseases characterized by an increased intraocular pressure which causes pathological changes in the eye and impaired vision, and which may lead to blindness
- glaucous*: pale bluish-green, or with a whitish bloom which rubs off
- glioma*: a tumour composed of tissue representing

- neuroglia (the supporting tissue of the nervous system); sometimes extended to include all primary, intrinsic neoplasms of the central nervous system
- globose*: spherical or nearly so
- glomerule*: a condensed head of almost sessile flowers; a cluster of heads in a common involucre
- glossitis*: inflammation of the tongue
- glume* (*plural glumes*): the chaffy or membranous two-ranked members of the inflorescence of grasses and similar plants; lower glume and upper glume, two sterile bracts at the base of a grass spikelet
- glutinous*: sticky
- glycaemia*: the presence of glucose in the blood
- glycosides*: see aglycones
- goitre*: an enlargement of the thyroid gland, resulting in a swelling in the front part of the neck
- goitrogen*: a goitre-producing compound
- goitrogenic*: producing goitre
- gonadotrophic, gonadotropic*: stimulating the gamete-producing glands
- gonadotrophin, gonadotropin*: any hormone that stimulates the gamete-producing glands (gonads), like ovary and testis
- gonorrhoea*: a venereal disease characterized by inflammation of the mucous membrane of the genitourinary tract and a discharge of mucus and pus
- gout*: a group of disorders of (purine) metabolism, characterized by inflammation of a joint, paroxysmal recurrent pain and an excess of uric acid in the blood
- gouty*: characterized by inflammation of a joint, paroxysmal recurrent pain and an excess of uric acid in the blood
- graft*: a union of different individuals by apposition, the rooted plant being termed the stock, the portion inserted the scion
- grafting*: the process of inserting a scion, which consists of a piece of stem and two or more buds of the plant to be propagated, into another plant (rootstock) with the intention that it will unite and grow
- grain* (*botany*): a general term for cereals, those grasses cultivated for food; the caryopsis or the fruit of cereals
- gram-negative*: losing the stain or decolorized by alcohol in Gram's staining method, which is a primary characteristic for bacteria with a cell wall consisting of a thin layer of peptidoglycan with an outer membrane of lipoprotein and lipopolysaccharide
- gram-positive*: retaining the stain or resisting decolorization by alcohol in Gram's staining method, which is a primary characteristic for bacteria with a cell wall consisting of a thick layer of peptidoglycan with attached teichoic acids
- granulocyte*: any cell containing granules, especially leucocytes containing cytoplasmic granules
- granulocytopenia*: a complex of symptoms characterized by a marked decrease in the number of granulocytes and by lesions of the throat and other mucous membranes, the gastro-intestinal tract and the skin; also known as agranulocytosis
- granuloma*: imprecise term, applied to aggregations of either mononuclear inflammatory cells or modified macrophages; granuloma formation represents a chronic inflammatory response
- granulose* (*granular*): composed of or covered with grain-like minute particles
- green manure*: green leafy material applied to and mostly worked into the soil to enrich the soil with nutrients and organic matter
- greenwood cutting*: a cutting of immature and still soft and pliable tissue
- gregarious*: growing in associated groups or clusters but not matted; at the same time; in bamboos gregarious flowering is used to indicate that a whole population flowers over a period of 2-3 years and then dies, although sometimes the rhizomes remain alive
- gum*: a colloidal polysaccharide substance that is gelatinous when moist but hardens on drying; gum is exuded by plants or extracted from them
- gynobase*: an elongated or enlarged receptacle on which the pistil is borne
- gynobasic*: of a style, attached to a prolongation upwards of the receptacle between the carpels
- gynodioecious*: having female and bisexual flowers on different plants
- gynoecium*: the female part or pistil of a flower, consisting, when complete, of one or more ovaries with their styles and stigmas
- gynostemium*: the androecium and gynoecium combined
- habit* (*botany*): external appearance or way of growth of a plant
- habitat*: the kind of locality in which a plant grows
- haemagglutination*: agglutination of erythrocytes
- haematometra*: an accumulation of blood in the uterus
- haematopoietic*: pertaining to or effecting the formation of blood cells; an agent that promotes

- the formation of blood cells
- haematotoxic*: poisonous to the blood and the haematopoietic system
- haematuria*: the presence of blood in the urine
- haemin*: a porphyrin chelate of iron
- haemolysis*: disruption of the integrity of the red blood cell membrane, causing release of haemoglobin
- haemolytic*: pertaining to, characterized by, or producing haemolysis
- haemoptysis*: expectoration of blood or blood-stained sputum from some part of the respiratory tract
- haemorrhage*: bleeding; the escape of blood from blood vessels
- haemorrhoid*: a mass of dilated veins in swollen tissue situated near the anal sphincter
- haemostasis*: the arrest of bleeding
- haemostatic*: arresting the flow of blood; an agent that checks the flow of blood
- haemostyptic*: haemostatic
- hallucinogenic*: inducing hallucinations
- hardwood cutting*: a cutting consisting of mature woody tissue
- head*: a dense inflorescence of small crowded often stalkless flowers (a capitulum)
- heartwood*: wood from the inner portion of a tree in which the cells are dead and no longer engaged in sap conduction and food storage
- hemi-*: prefix, meaning half
- hemiplegia*: paralysis of one side of the body
- hepatitis*: inflammation of the liver
- hepatocyte*: liver cell
- hepatomegaly*: enlargement of the liver
- hepatotoxic*: having a toxic effect on liver cells
- herb*: any vascular plant which is not woody
- herbaceous*: with the texture, colour and properties of a herb; not woody
- herbivore*: a plant-eating animal
- hermaphrodite*: bisexual; in flowers, with stamens and pistil in the same flower
- herpes*: any of several inflammatory diseases of the skin caused by a herpesvirus and characterized by clusters of vesicles
- herpes simplex*: group of acute infections caused by herpes simplex virus type 1 or type 2, characterized by the development of one or more small fluid-filled vesicles on the skin or mucous membrane, and occurring as a primary infection or recurring because of reactivation of a latent infection
- heterodistylous*: with two kinds of plants, having either short or long styles
- heteromorphic*: varying in number or form
- heterosis*: exceptional vigour of organisms through crossbreeding between two different types
- heterostylous*: having styles of two or more distinct forms or of different lengths
- heterotristylous*: with three kinds of plants, having long, short or intermediate styles
- hexaploid*: having six sets of chromosomes (6n)
- hilum*: the scar left on a seed indicating its point of attachment
- hirsute*: with rather coarse stiff hairs
- hispid*: covered with long rigid hairs or bristles
- hispidulous*: minutely hispid
- histiocytosis*: a condition characterized by the abnormal appearance of macrophages in the blood
- histochemical*: pertaining to the chemical components or activities of cells and tissues
- histopathological*: pertaining to the histology of diseased tissues
- HIV (human immunodeficiency virus)*: a virus that is the aetiological agent of acquired immunodeficiency syndrome (AIDS). Two serotypes are distinguished: HIV-1, with a worldwide distribution, and HIV-2, which is largely confined to West Africa
- Hodgkin's disease*: a form of malignant lymphoma characterized by painless, progressive enlargement of the lymph nodes, spleen and general lymphoid tissue
- homeostasis*: a tendency to stability in the normal body states of the organism
- homogamous*: bearing one kind of flowers
- homostylous*: having styles of the same length and/or shape
- husk*: the outer covering of certain fruits or seeds
- hyaline*: almost transparent
- hybrid*: the first generation offspring of a cross between two individuals of different species or taxa
- hybridization*: the crossing of individuals of different species or taxa
- hydrolysable tannins*: see tannins
- hydrolysis*: a chemical reaction of water in which a bond in the reactant other than water is split and hydrogen and hydroxyl are added
- hydrophilic*: having a strong affinity for water
- hydrophobia*: any morbid dread of water; rabies
- hydrophobic*: resistant to or avoiding wetting
- hydrophyte*: plant living in water or wet ground
- hypanthium*: a cup-like receptacle usually derived from the fusion of the floral envelopes and androecium on which are seemingly borne the calyx, corolla and stamens
- hyperaemia*: an excess of blood in a part of the body

- hyperaesthesia*: increased sensitivity, particularly a painful sensation from a normally painless touch stimulus
- hyperbilirubinaemia*: the presence of excessive concentrations of bilirubin in the blood, which may lead to jaundice
- hypercalcaemia*: an excess of calcium in the blood
- hypercholesterolaemia*: an excess of cholesterol in the blood
- hypercholesterolaemic*: pertaining to, characterized by, or tending to produce hypercholesterolaemia
- hyperglycaemia*: an abnormally increased glucose concentration in the blood
- hyperglycaemic*: pertaining to, characterized by, or causing hyperglycaemia
- hyperlipemia*: hyperlipidemia, a general term for elevated concentrations of any lipid or all lipids in the plasma
- hyperphylloerythrinaemia*: the presence of excessive concentrations of phylloerythrin in the blood
- hypertension*: high arterial blood pressure
- hypertrophic*: pertaining to or marked by hypertrophy, i.e. the enlargement of (part of) an organ due to an increased size of its cells
- hypo-thyroidism*: deficiency of thyroid activity; in adults characterized by a decreased basal metabolic rate, fatigue and lethargy, sensitivity to cold and menstrual disturbances; in children it may lead to arrested physical and mental development
- hypocholesterolaemia*: an abnormally diminished amount of cholesterol in the blood
- hypocholesterolaemic*: pertaining to, characterized by, or producing hypocholesterolaemia
- hypocotyl*: the young stem below the cotyledons
- hypocrateriform*: saucer-shaped, with a long and narrow tube and limbs at right angles to the tube
- hypogeal*: below ground; in hypogeal germination the cotyledons remain below ground within the testa
- hypoglycaemic*: pertaining to, characterized by, or producing an abnormally decreased glucose concentration in the blood (hypoglycaemia)
- hypolipidaemic*: promoting the reduction of lipid concentration in the serum
- hypopodium*: the basal portion of a leaf, including the stalk
- hyposensitization*: the act or process of reducing the ability to react to a specific allergen by applying repeated and gradually increasing doses of the offending substance
- hypotension*: an abnormally low blood pressure
- ichthyotoxic*: poisonous to fishes
- imbricate*: overlapping like tiles; in a flower bud when one sepal or petal is wholly external and one wholly internal and the others overlapping at the edges only
- imidazole alkaloids*: a subgroup of the alkaloids
- imparipinnate*: of leaves, pinnate with an unpaired terminal leaflet
- impetigo*: a contagious, purulent skin disease, caused by group A streptococci or *Staphylococcus aureus*, and mostly seen in children, usually on the face
- implantation*: the embedding of the fertilized egg in the uterus; the insertion or grafting of material into the body
- impotence*: lack of power, specifically lack of copulative power in the male due to failure to initiate or maintain an erection
- in situ*: in the natural environment; in medicine: in the natural or normal place
- in vitro*: outside the living body and in an artificial environment
- inarching*: grafting by approach, the scion remaining attached to its parent until union has taken place
- incised*: cut deeply
- incompatibility*: in floral biology: not capable of cross- or self-fertilization; in plant propagation: not capable of making stock-scion combinations resulting in a lasting union
- indehiscent*: not opening when ripe
- indented*: forced inward to form a depression
- indeterminate*: of inflorescences, a sequence in which the terminal flowers are the last to open, so that the floral axis may be prolonged indefinitely by the terminal meristem; of shoot growth: when the shoot apex forms and unfolds leaves during extension growth, so that shoot growth can continue indefinitely
- indigenous*: native to a particular area or region
- indigestion*: lack or failure of digestion
- indole alkaloids*: a subgroup of the alkaloids
- indumentum*: a covering, as of hairs, scales, etc.
- induplicate*: with the margins bent inwards and the external face of these edges applied to each other without twisting; V-shaped in cross section, trough-shaped
- inequilateral*: unequal-sided
- inferior*: beneath, lower, below; an inferior ovary is one which is situated below the sepals, petals and stamens
- inflammation*: a protective response of the body in response to injury, infection, irritation, etc.,

- aimed at destroying or isolating the injurious agent and injured tissue, and characterized by redness, pain, heat, and swelling
- inflexed*: bent or curved inward toward the centre
- inflorescence*: the arrangement and mode of development of the flowers on the floral axis; the branch that bears the flowers, including all its bracts and branches
- influenza*: an acute highly contagious virus disease characterized by sudden onset, fever, prostration, severe aches and pains, and progressive inflammation of the respiratory mucous membrane
- infrageneric*: referring to any taxon below the genus level
- infraspecific*: referring to any taxon below the species level
- infructescence*: a ripened inflorescence in the fruiting stage
- infundibular*: funnel-shaped
- infusion*: a liquid extract obtained by steeping or soaking something in a liquid for the purpose of extracting its medicinal principles without boiling; the therapeutic introduction of a fluid, other than blood, into a vein
- inner bark*: the secondary phloem; the living part of the tissue outside the cambium
- inoculation*: grafting, more properly budding, a single bud only being inserted; introduction of microorganisms, infective material, serum and other substances into tissues of living plants and animals, or culture media promote growth
- inoculum*: material used for inoculation, e.g. rhizobia in soil to promote the growth of certain *Leguminosae*
- inotropic*: affecting the force or energy of muscular contractions (positive: increasing the force; negative: weakening the force)
- insecticide*: an agent that destroys insects
- insomnia*: sleeplessness
- insulin*: a protein hormone produced by  $\beta$ -cells of the islets of Langerhans in the pancreas, which is secreted in response to elevated glucose and amino acid levels in the blood and promotes their storage and utilization. Insulin deficiency is often the cause of diabetes, and exogenous insulin is used to control that disease
- interacuminal*: between acumens
- intergeneric hybrid*: hybrid between species of two or more genera
- internode*: the portion of the stem (culm) between two nodes
- interpetiolar*: of stipules placed between the petioles of opposite leaves
- intracerebro ventricular*: within one of the several cavities of the brain which are filled with cerebrospinal fluid
- intragastic*: within the stomach
- intraluminal*: within the lumen of a tube, e.g. of a blood vessel or intestine
- intramarginal*: placed within the margin near the edge; of a vein, running near and parallel with the margin
- intrapetiolar*: of stipules, positioned within the petiole axil
- intraspecific*: occurring within a species or involving members of one species
- intrastaminal*: within the stamens
- introrse*: turned inward, towards the axis, as the dehiscence of an anther
- involucral*: belonging to an involucre
- involucre*: a ring of bracts (involucral bracts) surrounding several flowers or their supports, as in the heads of *Compositae* or the umbels in *Umbelliferae*
- involute*: having the edges of the leaves rolled inwards
- iridoids*: monoterpenes ( $C_{10}$ ) characterized by a cyclopentanotetrahydropyran ring system, also known as the iridane skeleton; seco-iridoids can be regarded as being formed from iridoids by opening of the cyclopentane ring between  $C_7$  and  $C_8$
- ischias*: pain in the inferior dorsal portion of the hip bone (ischium)
- islets of Langerhans*: irregular microscopic structures scattered throughout the pancreas and comprising its endocrine part
- isoflavonoids*: a subgroup of the flavonoids, in which the basic structure is the 3-phenyl chromane skeleton
- isomer*: a compound, radical or ion containing the same numbers of atoms of the same elements in the molecule as one or more others, and hence having the same molecular formula, but differing in the structural arrangement of the atoms and consequently in one or more properties
- isoprenoids or terpenoids*: a large group of secondary metabolites, in which isopentenyl pyrophosphate ('active isoprene' or ' $C_5$ -unit') is the building block; isopentenyl pyrophosphate is derived via the mevalonic acid pathway; mevalonic acid itself is formed from 3 molecules of acetate, but the mevalonic acid pathway channels acetate into a different series of compounds than does the acetate pathway
- isoquinoline alkaloids*: a subgroup of the alkaloids
- jaundice*: a syndrome marked by hyperbilirubi-

- naemia and deposition of bile pigments in the skin, mucous membranes and eyeball, resulting in yellowish pigmentation of these body parts
- joint*; *jointed*: an articulation, like a node in plants and a place of union of two bones in the human body; articulated
- jugate*: connected or yoked together; e.g. in leaves 1-n-jugate: with 1-n pairs of leaflets
- karyology*: the science of the nucleus and its development and vital history
- karyotype*: the full chromosome set of the nucleus of a cell
- keel (carina)*: a ridge like the keel of a boat; the two anterior and united petals of a papilionaceous corolla; the principal vein of a sepal or glume
- keeled (carinate)*: having a keel or carina
- keloid*: an overgrowth of scar tissue
- keratinophilic*: having an affinity for keratin, i.e. any of a family of scleroproteins which form the primary constituents of epidermis, hair, nails and horny tissues
- kernel*: the nucellus of an ovule or of a seed, that is, the whole body within the coats
- labellum*: lip; the lowest petal of an orchid; petaloid anterior staminode in *Zingiberaceae*
- lac insect*: a scale insect (*Laccifer lacca*, synonym *Kerria lacca*) that produces lac, a resinous gold-coloured substance used for lacquerware
- lacinate*: slashed, cut into narrow lobes
- lactifuge*: an agent that checks the secretion of milk
- laevodopa*: the laevorotatory isomer of dopa, used as an agent against parkinsonism
- lamina*: see blade
- lanate*: with woolly hairs
- lanceolate*: lance-shaped; much longer than broad, being widest at the base and tapering to the apex
- lanuginose*: woolly or cottony
- larvicidal*: destroying insect larvae
- larvicide*: an agent destructive to insect larvae
- laryngitis*: inflammation of the larynx
- lateral*: on or at the side
- laterite*: a red soil that shows intensive weathering and chemical change and leaching away of bases and silica, leaving aluminium and iron oxides
- latex*: a juice, usually white and sometimes sticky, exuding from broken surfaces of some plants
- laticiferous*: latex-bearing
- lax*: loose, distant
- laxative*: aperient, mildly purgative; an agent that promotes evacuation of the bowel
- layer*: a branch caused to root while still connected to the parent and used for propagation (layering)
- LC<sub>50</sub>*: median lethal concentration, i.e. the concentration of a chemical that kills fifty percent of the organisms in a test population
- LD<sub>50</sub>*: median lethal dose, i.e. the amount of an agent that kills fifty percent of the organisms in a test population
- leaflet*: one part of a compound leaf
- lecitins*: glycerophosphoric acid derivatives, in which 2 free hydroxyl groups of the glycerol are esterified with fatty acids, while 1 of the 2 remaining groups of the phosphoric acid residue is esterified to an alcohol
- lectins*: proteins of glycoproteins, which are not antibodies or enzymes, but which have the ability to attach themselves to specific sugars; the binding is not covalent, and the sugar can either be free or constituent part of a larger molecule, which may be present, e.g. in a membrane
- leishmanial*: pertaining to or caused by leishmaniasis, i.e. protozoa of the genus *Leishmania*
- Leishmaniasis ('kala azar')*: infection caused by *Leishmania*, and classified into cutaneous, mucocutaneous and visceral leishmaniasis (kala-azar)
- lemma*: the lower of the two glumes which surround each floret in the spikelet of grasses
- lenticel*: lenticular masses of loose cells protruding through fissures in the periderm on stems, fruits and roots, usually arising beneath individual stomata; their main function is gaseous exchange
- lenticellate*: having lenticels
- lenticular*: shaped like a double-convex lens
- leprosy*: a chronic, infectious, slowly progressive disease, caused by *Mycobacterium leprae*, characterized by lesions in the skin, mucous membranes, nerves, bones and viscera, and manifested by a broad range of clinical symptoms
- Letterer-Siwe disease*: a specific form of histiocytosis, characterized by a haemorrhagic tendency, eczematoid skin eruption, enlargement of the liver, spleen and lymph nodes, and progressive anaemia
- leucopaenia*: reduction in the number of leucocytes (white blood cells) in the blood
- leucopaenic*: pertaining to, characterized by, or causing leucopaenia
- leucorrhoea*: a whitish, viscid discharge from the female genitals
- leucotriens*: see prostaglandins
- leukaemia*: a malignant, progressive disease of the

- blood-forming organs, with distorted proliferation and development of the white corpuscles (leucocytes) and their precursors
- liana*: a woody climbing vine
- ligand*: a molecule that binds to another molecule, used especially to refer to a small molecule that binds specifically to a larger molecule, e.g. an antigen binding to an antibody, or a hormone or neurotransmitter binding to a receptor
- lignans*: a group of natural products (dimers) derived from condensation of 2 phenylpropane units
- lignins*: see lignans
- ligulate*: possessing an elongated flattened strap-shaped structure or ligule
- ligule*: an elongated flattened strap-shaped structure; a membranous outgrowth on the upper surface of a grass leaf at the junction of the sheath and the blade which may be presented by a ridge or by a line of hairs; in palms it is a distal projection of the leaf sheath, often coriaceous
- limb (botany)*: the expanded part of a tubular corolla, as distinct from the tube or throat; the lamina of a leaf or of a petal; the branch of a tree
- line*: used in plant breeding for a group of individuals from a common ancestry
- linear*: long and narrow with parallel sides
- liniment*: an oily liquid preparation to be used on the skin
- lipolytic*: pertaining to, characterized by, or causing the decomposition or splitting up of fat
- lipomatosis*: a condition characterized by abnormal localized, or tumour-like, accumulations of fat in the tissues
- lipophilic*: having an affinity to fat; dissolving in lipids
- lithiasis*: the formation of stones or calculi in the body
- lithotriptic*: pertaining to or producing lithotripsy, i.e. the destruction of a stone in the gallbladder or urinary system stone in the bladder or kidneys
- lobe*: any division of an organ or specially rounded division
- lobed*: divided, but not to the base
- lochia*: the vaginal discharge taking place during the first week or two after giving birth
- locular*: divided by internal partitions into compartments as in anthers and ovaries
- locule*: the cavity of an ovary or anther
- loculicidal*: the cavity of a pericarp dehiscent by the back, the dorsal suture
- long-day plant*: a plant which only reaches a certain development stage (usually flowering), or reaches it more rapidly, if there are more than a certain number of hours of light in each 24 h period
- longitudinal*: lengthwise
- lotion*: a liquid suspension or dispersion for external application to the body
- lumbago*: pain in the lumbar region of the back (loins); lumbar rheumatism
- lupus*: name originally given to localized destruction or degeneration of the skin caused by various cutaneous diseases; formerly the term was used to designate lupus vulgaris and lupus erythematosus, nowadays it is only used with modifier
- lymphadenopathy*: disease of the lymph nodes
- lymphoma*: any neoplastic disorder of the lymphoid tissue; the term is often used alone to denote malignant lymphoma
- lyophilization*: the creation of a stable preparation of a biological substance (e.g. blood plasma, serum) by rapid freezing and dehydration of the frozen product under high vacuum
- lysosome*: an intracellular body containing various hydrolytic enzymes and normally involved in the process of intracellular digestion. Injury to a lysosome is followed by release into the cell, which may damage the cell and give rise to wasting and other pathological aspects of certain diseases
- lysozyme*: an enzyme of the hydrolase class, occurring in saliva, tears, egg white and many animal fluids, and catalysing the breakdown of some bacterial cell walls
- macerate*: to reduce to a soft mass by soaking
- maceration*: a method of extract preparation in which the matter to be extracted is mixed with the prescribed extraction solvent, and allowed to stand in a closed container for an appropriate time; the residue is separated from the extraction solvent, and if necessary, pressed out; in the latter case, the two liquids obtained are combined; see also percolation
- macrophage*: any of the many forms of mononuclear phagocytes (cells capable of ingesting particulate matter) found in tissues
- Malesia*: the biogeographical region including Malaysia, Indonesia, the Philippines, Singapore, Brunei and Papua New Guinea
- mangrove*: a brackish-water coastal swamp of tropical and subtropical areas that is partly inundated by tidal flow
- margin*: the edge or boundary line of a body
- mast cell*: a connective tissue cell whose specific



- physiological function remains unknown
- masticatory*: used for chewing (mastication)
- mastitis*: inflammation of the mammary gland or breast
- median*: belonging to the middle
- medifixed*: attached or fixed by the middle
- melanoma*: a tumour arising from the melanocytic system of the skin and other organs; when used alone, the term refers to malignant melanoma
- membranous*: thin and semi-transparent, like a fine membrane
- meniscoid*: thin and concavo-convex
- menorrhagia*: excessive uterine bleeding, occurring at regular intervals, with the period of flow being of usual duration; also called hypermenorrhoea
- mericarp*: one of the separate halves or parts of a fruit, as in *Umbelliferae*
- meristem*: undifferentiated tissue of the growing point whose cells are capable of dividing and developing into various organs and tissues
- merous*: 4-, 5- etc., with 4, 5 etc. parts or numbers of sepals, petals etc.
- merozoite*: a stage in the life cycle of certain sporozoan protozoa, resulting from asexual reproduction
- mesocarp*: the middle layer of the pericarp or fruit wall which is often fleshy or succulent
- mesophyte*: plant requiring medium moisture conditions; intermediate between hydrophyte and xerophyte
- metabolic acidosis*: a disturbance in which the acid-base status of the body shifts towards acidity as a result of the loss of bases or the retention of acids
- metabolism*: the sum of all the physical and chemical processes by which living organized substance is produced and maintained, and also the transformation by which energy is made available for the uses of the organism; biotransformation
- metabolite*: any substance produced by metabolism or by a particular metabolic process
- metastasis (plural metastases)*: growth of abnormal cells or pathogenic microorganisms distant from the site which was primarily involved by the morbid process
- metritis*: inflammation of the uterus
- metrorrhagia*: uterine bleeding, usually of variable amount, occurring at completely irregular but frequent intervals, the period of flow sometimes being prolonged
- mevalonic acid pathway*: the biosynthetic pathway which leads to isoprenoids; see also isoprenoids
- midrib*: the main vein of a leaf which is a continuation of the petiole
- mildew*: a superficial, usually whitish growth on living plants produced by fungi
- miliaria*: a syndrome of cutaneous changes associated with the retention of sweat and the extravasation of sweat at different levels in the skin
- mistletoe*: any of numerous hemiparasitic plants of the family *Loranthaceae*
- mitogen*: a substance that induces blast transformation, the synthesis of DNA, RNA and proteins, and the proliferation of lymphocytes
- mitogenic*: causing or inducing mitosis or cell proliferation
- mitosis*: a method of indirect division of a cell, consisting of a complex of various processes, through which the two daughter nuclei normally receive identical complements of the chromosomes
- module*: shoot unit with determinate growth either by apical abortion or conversion of the apex to an inflorescence
- molluscicide*: an agent that destroys snails and other molluscs; also called molluscicide
- monadelphous*: of stamens, united into one group by their filaments
- monochasium*: a cymose inflorescence where a pattern of a single lateral branch arising below the terminal flower is repeated
- monocotyledon*: angiosperm having a single cotyledon or seed-leaf
- monoecious*: with unisexual flowers, but male and female flowers borne on the same plant
- monomer*: the simple unpolymerized form of a chemical compound having relatively low molecular weight
- monomeric*: formed of a single member, e.g. a fruit of one carpel
- monopodial*: of a primary axis which continues its original line of growth from the same apical meristem to produce successive lateral branches; in bamboos used to designate a type of rhizome (see leptomorph)
- monoterpenes*: a subgroup of the isoprenoids, formed by coupling of 2 C<sub>5</sub> units
- monotherapy*: treatment by means of a single drug
- monotypic*: consisting of a single element, e.g. of a genus consisting of only one species
- monsoon forest*: a deciduous tropical woodland experiencing periodic drought
- morphogenetic*: relating to the development of normal organic form

- mucilage (mucilaginous)*: a gelatinous substance that is similar to gum but that swells in water without dissolving and forms a slimy mass
- mucolytic*: an agent that destroys or dissolves mucin
- mucous*: pertaining to, resembling, producing, containing or covered with mucus
- mucronate*: ending abruptly in a short stiff point
- mucronulate*: diminutive of mucronate
- mulch*: plant or non-living materials used to cover the soil surface with the object of protecting it from the impact of rainfall, controlling weeds, temperature and evaporation
- multiple sclerosis (MS)*: disease caused by sclerosis occurring in patches in the brain and/or spinal cord, leading to tremors, failure of coordination and various nervous and mental symptoms
- musculotropic*: affecting, acting upon, or attracted to muscular tissue
- mutagen*: an agent inducing or increasing genetic mutations by causing changes in DNA
- mutagenesis*: the induction of genetic mutation
- mutagenic*: capable of inducing genetic mutation
- myalgia*: pain in a muscle
- mycoherbicide*: a fungus deliberately used to cause disease in weeds or undesired crops
- mycotoxigenic*: producing or elaborating fungal toxins (mycotoxins)
- mydriasis*: physiological, morbid or drug-induced dilation of the pupil
- myelogenic*: produced in the bone marrow
- myelosuppression*: suppression of bone marrow activity, resulting in reduction in the number of platelets, red cells and white cells
- myocardial ischemia*: deficiency of blood supply to the heart muscle, due to constriction or obstruction of the coronary arteries
- myocardium*: the middle and thickest layer of the heart wall, composed of cardiac muscle
- myorelaxant*: an agent that aids in reducing muscle tension
- myosis*: contraction of the pupil
- naphthaquinones*: a subgroup of the quinones, in which the dione is conjugated to the condensed polycyclic aromatic system of naphthalene
- narcotic*: pertaining to or producing narcosis or stupor; an agent that in moderate doses dulls the senses, relieves pain and induces sleep, but in excessive doses may cause stupor, coma, convulsions and death
- nasopharynx*: the upper part of the alimentary canal continuous with the nasal passages
- naturalized*: introduced into a new area and established there, giving the impression of wild growth
- nausea (nauseous)*: an uncomfortable feeling in and about the stomach associated with aversion to food and a need to vomit
- necrobiosis*: swelling, basophilia (abnormal accumulation of basophilic blood cells in the blood) and distortion of collagen bundles in the dermis, sometimes with obliteration of the normal structure, but short of actual necrosis
- necrosis*: in plants, death of a portion of tissue often characterized by a brown or black discoloration; in humans, the sum of morphological changes indicative of cell death and affecting groups of cells, parts of structures, or organs
- nectar*: a sweet fluid exuded from various parts of the plant (e.g. by the flower to attract pollinators)
- nectary*: a group of modified subepidermal cells in flowers or leaves (extrafloral) secreting nectar
- nematicide*: an agent that destroys nematodes
- nematode*: small elongated cylindrical worm-like micro-organism, free-living in soil or water, or parasitic in animals or plants
- neolignans*: see lignans
- nephritis*: inflammation of the kidney
- nephroblastoma*: a rapidly developing malignant mixed tumour of the kidneys, usually affecting children less than five years old; also called Wilms' tumour
- nephrotic syndrome*: general name for a group of diseases involving defective kidney glomeruli
- nephrotoxicity*: the quality of being toxic or destructive to kidney cells
- nerve*: in botany: a strand of strengthening and/or conducting tissue running through a leaf, which starts from the midrib and diverges or branches throughout the blade; in medicine: a cordlike structure consisting of nerve fibres, which convey impulses between the central nervous system and other body parts
- neuralgia*: pain radiating along the course of one or more nerves
- neurasthenia*: a syndrome of chronic mental and physical weakness and fatigue, which was thought to be caused by exhaustion of the nervous system
- neuroblastoma*: sarcoma consisting of malignant neuroblasts (immature nerve cells)
- neurohypophysial*: pertaining to the neurohypophysis, i.e. the posterior lobe of the pituitary gland
- neurolethyrism*: a morbid condition resulting from excessive ingestion of *Lathyrus* seeds
- neuroleptic*: term referring to effects of antipsy-

- chotic drugs, such as producing a state of apathy, lack of initiative, limited range of emotion, and, in psychotic patients, normalization of psychomotor activity and reduced confusion and agitation
- neuropathy*: a functional disturbance or pathological change in the peripheral nervous system
- neurotrophic*: pertaining to neurotrophs, i.e. the nutrition and maintenance of nervous tissue
- neutrophil*: a granular leucocyte, having the properties of chemotaxis, adherence to immune complexes and phagocytosis
- nidation*: implantation of the fertilized egg in the uterus
- nociception*: sensation of pain
- node*: the point on the stem or branch at which a leaf or lateral shoot is borne
- nodulation*: formation of root-nodules
- nodule*: a small knot or rounded body, often in roots of leguminous plants, where bacteria of the genus *Rhizobium* are active in the fixation of nitrogen from the air
- norlignans*: see lignans
- norlupinane alkaloids*: a subgroup of the alkaloids
- nut*: a one- to many-seeded indehiscent fruit with a hard dry pericarp or shell
- nutlet*: a little nut
- ob-*: prefix, indication inverse or opposite condition (obtriangular, obcordate, etc.)
- oblanceolate*: reverse of lanceolate
- oblate*: more or less spherical but flattened at the poles
- oblique*: slanting; of unequal sides
- oblong*: longer than broad, with the sides parallel or almost so
- obovate*: reverse of ovate
- obstructive jaundice*: jaundice which is due to an impediment to the bile flow from the liver cells to the duodenum
- obtuse*: blunt or rounded at the end
- octoploid*: having eight times the basic number of chromosomes ( $8n$ )
- odontalgic*: pertaining to or characterized by toothache (odontalgia)
- oedema*: the presence of abnormally large amounts of fluid in the intercellular tissue spaces of the body
- oestrogen*: a sex hormone produced especially in the ovaries
- oliguria*: reduced urine excretion
- ombrophilous*: rain-loving
- oncogenic*: giving rise to tumours (either benign or malignant) or causing tumour formation
- oncolytic*: pertaining to, characterized by, or causing the lysis or destruction of tumour cells
- ophthalmia*: severe inflammation of the eye, or of the conjunctiva or deeper structures of the eye
- opposite*: of leaves and branches when two are borne at the same node on opposite sides of the stem
- optical activity*: in organic chemistry, the property of a compound, containing an asymmetrical carbon atom, of rotating the plane of polarized light, clockwise in the case of dextrorotatory (abbreviated d-) compounds, and counterclockwise in the case of laevorotatory (abbreviated l-) compounds. In perfumery d- and l-compounds may have different odours, e.g. d- and l-limonene
- optical isomerism*: isomerism in which the molecular structures of the molecules are mirror-images of one another. Optical isomers have the same structural formula, but their molecules cannot be superimposed
- orbicular*: flat with a more or less circular outline
- orchitis*: inflammation of a testis
- organoleptic*: of or pertaining to sensations and their evaluation
- orthotropic*: having a more or less vertical direction of growth
- osmolality*: the concentration of osmotically active particles in solution
- ostalgia*: pain in a bone or in the bones
- osteolathyrism*: a skeletal disorder produced by diets containing *Lathyrus* spp. or their active principles
- osteosarcoma*: a malignant primary tumour of bone
- outcross*: cross-pollination, usually by natural means, with plants differing in genetic constitution
- outer bark*: the periderm or rhytidome; the non-living layer of fibrous or corky tissue outside the cambium in woody plants which may be shed or retained
- oval*: see ovate
- ovary*: in plants, that part of the pistil, usually the enlarged base, which contains the ovules and eventually becomes the fruit; in humans, one of the two sexual glands in which the female reproductive cells (ova) are formed
- ovate*: egg-shaped in outline or in section; a flat surface which is scarcely twice as long as broad with the widest portion below the middle
- ovicidal*: destructive to the eggs of certain organisms
- ovoid*: a solid object which is egg-shaped (ovate in section)

- ovule (botany)*: the immature seed (egg) in the ovary before fertilization
- oxidation*: the processes of combining a compound with oxygen, dehydrogenating, or increasing the proportion of the electro-negative part
- palmate*: of leaflets, leaf-lobes or veins, with the different elements arising from the same point
- palsy*: paralysis
- palyology*: a branch of science studying pollen
- panacea*: a universal remedy; a herb credited with remarkable healing properties
- pancreas*: a compound glandular organ associated with the gut in most vertebrates, and secreting the hormones insulin and glucagon from the endocrine part (the islets of Langerhans) and pancreatic juice with digestive enzymes from the exocrine part
- panelling*: to furnish or decorate with panels (rectangular boards)
- panicle*: an indeterminate branched racemose inflorescence
- paniculate*: resembling a panicle
- pantropical*: distributed throughout the tropics
- papilionaceous flower*: a butterfly-like, pea-like flower, with standard, wings and keel
- papillate*: having minute nipple-like protuberances
- papilloma*: a benign epithelial tumour producing finger-like or warty projections
- papillomatosis*: the development of multiple papillomas
- papillose*: covered with minute nipple-like protuberances
- pappus*: the various tufts of hairs on achenes or fruits; the limb of the calyx of *Compositae* florets
- parasympatholytic*: producing effects which resemble those of interruption of the parasympathetic nerve supply to a body part; an agent that opposes the effects of parasympathetic nerve impulses; also called anticholinergic
- parasympathomimetic*: producing effects resembling those of stimulation of the parasympathetic nerve supply to a part
- parenchyma*: in plants: ground tissue composed of thin-walled, relatively undifferentiated cells, e.g. the pith and mesophyll; in humans: the soft cellular substance of glandular and other organs, or the essential elements of an organ
- paresthesia*: an abnormal touch sensation, e.g. burning, itching or prickling, often without an external stimulus
- parietal*: placentation type, when the ovules are attached to the wall of a one-celled ovary
- paripinnate*: a pinnate leaf with all leaflets in pairs
- parkinsonism*: a group of neurological disorders marked by abnormally decreased motor function (hypokinesia), tremor and muscular rigidity
- partite (parted)*: cleft, but not quite to the base
- patent (botany)*: spreading out widely
- pathophysiology*: the physiology of disordered function
- pectin*: a substance yielding viscous solutions with water and, in combination with acid and sugar, forming a gel constituting the base of fruit jellies
- pectinate*: pinnately cleft with narrow segments set close like the teeth of a comb
- pectoral*: of, or pertaining to, the chest or thorax; relieving disorders of the respiratory tract; any medicine against ailments of the chest
- pedicel*: the stalk of an individual flower
- pedicellate*: furnished with a pedicel
- peduncle*: the stalk of an inflorescence or partial inflorescence
- pedunculate*: furnished with a peduncle
- pellucid*: translucent
- peltate*: of a leaf, with the stalk attached to the lower surface, not at the edge
- pendent, pendulous*: drooping; hanging down from its support
- pentagonal*: with five angles
- pentamerous*: having five parts in a flower-whorl
- percolate*: a liquid that has been submitted to percolation
- percolation*: a method of extract preparation in which the matter to be extracted is mixed with a portion of the prescribed extraction solvent, and allowed to stand for an appropriate time; the mass is then transferred to a percolator and the remaining extraction solvent is allowed to flow slowly, making sure that the matter to be extracted is always covered with liquid; the residue may be pressed out and the expressed fluid combined with the percolate; see also maceration
- perennial*: a plant living for many years and usually flowering each year
- perianth*: the floral leaves as a whole, including both sepals and petals if both are present
- pericarp*: the wall of the ripened ovary or fruit whose layers may be fused into one, or may be more or less divisible into exocarp, mesocarp and endocarp
- peristalsis*: the movement by which the digestive tract and other tubular organs with both longi-

- tudinal and circular muscle fibres propel their contents
- peritonitis*: inflammation of the peritoneum, i.e. the serous membrane that lines the abdominal cavity
- persistent*: remaining attached; not falling off, not deciduous; applies to organs that remain in place after they have fulfilled their natural functions
- petal*: a member of the inner series of perianth segments (corolla) which are often brightly coloured
- petiolar*: borne on, or pertaining to a petiole
- petiolate*: having a petiole
- petiole*: the stalk of a leaf
- petiolulate*: having a petiolule
- petiolule*: the stalk of a leaflet
- phagocytosis*: endocytosis (uptake by a cell of material by invagination of its plasma membrane) of particulate material, such as microorganisms and cell fragments
- pharmacokinetics*: the activity or fate of drugs in the body over a period of time, including the processes of absorption, distribution, localization in tissues, biotransformation and excretion
- pharmacopoeia*: an authoritative treatise on drugs and their preparations; a book containing a list of products used in medicine, with descriptions, chemical tests for determining identity and purity, formulas for certain mixtures of these substances, and generally also statements of average dosage
- pharyngitis*: inflammation of the pharynx
- pharyngolaryngeal*: pertaining to the pharynx (the passage between the mouth and the larynx and oesophagus) and the larynx (the structure connected to the upper part of the trachea and to the pharynx, guarding the entrance into the trachea and functioning as the organ of voice)
- phenolics*: phenols are compounds which have an aromatic ring with an alcoholic group attached to it
- phenotype*: the physical or external appearance of an organism as distinguished from its genetic constitution (genotype); a group of organisms with similar physical or external make-up
- pheromone*: a substance which is secreted to the outside of the body by an individual and which is perceived (e.g. by smell) by another individual of the same species, leading to a specific reaction of behaviour in the percipient
- phlebitis*: inflammation of a vein
- phlebology*: the study of the veins and their diseases
- phlegm*: a viscid, stringy mucous secretion, like that produced by the mucous membranes of the respiratory tract, as during a cold
- phloroglucinols*: derivatives of 1,3,5-trihydroxybenzene
- photodermatitis*: an abnormal state of the skin in which light is an important causative factor
- photoperiod*: the relative duration of illumination in a cycle of light and darkness, whether occurring naturally (day and night) or imposed in an artificial way
- photosensitization*: the development of abnormally heightened reactivity of the skin to sunlight
- phototoxicity*: a nonimmunological, chemically induced type of photosensitivity
- phthisis*: wasting away of (a part of) the body; tuberculosis, especially of the lungs
- phyllody*: transformation of flower parts into leaves
- phylogenetic*: based on natural evolutionary and genealogical relationships
- phytophotodermatitis*: dermatitis induced by the sequential exposure to certain plants containing psoralen-type photosensitizers and then to sunlight
- phytosterins*: see phytosterols
- phytosterols*: a group name for the widespread plant sterols sitosterol, campesterol and stigmasterol
- phytotherapy*: treatment by use of plants
- pickle*: steep or soak in a solution for preservation, conditioning etc.; a preserving, flavouring liquid; an object preserved in a pickle
- pilose*: hairy with rather long soft hairs
- pinna (plural pinnae)*: a primary division or leaflet of a pinnate leaf
- pinnate*: arranged in pairs along each side of a common axis
- pinnatifid*: pinnately divided about halfway to the midrib
- pinnatilobed*: pinnately divided to about half-way to the midrib
- pinnatipartite*: pinnately parted
- pinnatisect*: pinnately divided down to the midrib
- pioneer species*: a species able to establish itself on bare ground, starting primary succession, often showing rapid growth and producing large amounts of diaspores
- piperidine alkaloids*: a subgroup of the alkaloids
- piscicidal*: poisonous to or controlling fish
- pistil*: the female part of a flower (gynoecium) of one or more carpels, consisting, when complete, of one or more ovaries, styles and stigmas
- pistillate*: a unisexual flower with pistil, but no

- stamens
- pistillode*: a sterile, often reduced pistil
- pith*: the soft core occurring in the structural centre of a log; the tissue, sometimes soft, in the centre of the stem of a non-woody dicotyledon
- placenta*: in plants, the part of the ovary to which the ovules are attached; in higher mammals, the vascular, spongy organ of interlocking maternal and foetal tissue by which the foetus is nourished in the uterus
- placentation (botany)*: the way in which the placentae are arranged in the ovary
- plagiotropic*: having an oblique or horizontal direction of growth
- plano-convex*: flat on one side and convex on the other
- platelet activating factor*: a substance released by basophils and mast cells in immediate hypersensitivity reactions and macrophages and neutrophils in other inflammatory reactions. It is an extremely potent mediator of bronchoconstriction and of the platelet aggregation and release reactions
- pleurisy, pleuritis*: inflammation of the pleura (the membrane between thorax and lung), which may be acute or chronic
- pleuritis*: pleurisy
- plicate*: folded to and fro, like a fan
- ploidy*: degree or repetition of the basic number of chromosomes
- plumule*: the primary bud of an embryo or germinating seed
- plywood*: a panel material consisting of wood veneers glued together with the grains of adjacent layers arranged at right angles or at a wide angle
- pneumonia*: inflammation of the lungs, with the lungs becoming firm following the filling of air spaces with exudate
- pod*: a dry fruit composed of a single carpel and dehiscent by sutures, as in legumes; a general term for a dry dehiscent fruit
- pollarding*: cutting a tree back to the trunk to promote the growth of a dense head of foliage
- pollen*: spores or grains borne by the anthers containing the male element (gametophyte)
- pollination*: the transfer of pollen from the dehiscing anther to the receptive stigma
- polygamous*: with unisexual and bisexual flowers in the same plant
- polymorphic, polymorphous*: with several or various forms; variable as to habit
- polyploidy*: the state of having more than two full sets of homologous chromosomes
- polyuria*: the passage of a large volume of urine in a given period, a characteristic of diabetes
- posology*: the science of dosage
- postphlebotic syndrome*: the different complications associated with deep vein thrombosis
- postsynaptic membrane*: the area of the plasma membrane of postsynaptic cells (cells situated beyond or distal to a synapse), either a muscle fibre or a neuron, which is within the synapse and has areas specifically adapted for receiving neurotransmitters
- poultice*: a soft, moist, usually heated and sometimes medicated mass spread on cloth and applied to sores or other lesions to create moist local heat or counterirritation
- prickle*: a sharp, relatively stout outgrowth from the outer layers
- primary immunization*: first contact with an antigen
- primary vegetation*: the original, undisturbed plant cover
- proanthocyanidins*: see tannins
- procumbent*: lying along the ground; in wood anatomy also of ray parenchyma cells with their longest dimension in radial direction
- progeny*: offspring
- progestational*: promoting gestation
- progestogen*: a term applied to any substance which possesses progestational activity
- promastigote*: any of the bodies representing the morphological (leptomonad) stage in the life cycle of certain trypanosomatid protozoa resembling the adult form of members of the genus *Leptomonas*, with the elongate or pear-shaped cell having a central nucleus, and at the anterior end a kinetoplast and a basal body from which arises a long, slender flagellum (see also: amastigote)
- prop roots*: aerial roots
- propagule*: a part of a plant that becomes detached and grows into a new plant
- prophyll*: the first bract borne on the inflorescence; the bracteole at the base of an individual flower
- prostaglandins*: the prostaglandins, leucotrienes and thromboxanes are a large group of modified C<sub>20</sub> fatty acids; they are known to occur widely in animal tissues, but only in tiny amounts, and they have been found to exert a wide variety of pharmacological effects (e.g. mediators of inflammation, platelet aggregation) on humans and animals
- prostrate*: lying flat on the ground
- protandrous*: of flowers, shedding pollen before

- the stigma is receptive
- proteolytic*: pertaining to, characterized by, or promoting proteolysis
- proto-alkaloids*: see alkaloids (Introduction)
- protogynous, proterogynous*: of flowers, the stigma is receptive before the pollen is shed; of inflorescences, the female flowers mature before the male ones
- provenance*: origin; a collection of pollen, seed or propagules from a certain restricted locality
- proximal*: in botany: the part nearest the axis (as opposed to distal); in human anatomy: relatively nearer to the central part of the body or point of origin
- pruning*: cutting off the superfluous branches or shoots of a plant for better shape or more fruitful growth; M-shaped pruning in bamboos: pruning a clump by removing culms so that the culm-less part looks like a letter M
- pruritic*: pertaining to or characterized by pruritus
- pruritus*: itching; any of various conditions marked by itching
- pseudo-alkaloids*: see alkaloids (Introduction)
- pseudopetiole*: a structure resembling a petiole, but not being one
- pseudoraceme*: raceme-like inflorescence but not a true raceme
- pseudostem*: an axis with the appearance of a stem but made up of other organs, e.g. leaf sheaths in *Musa* and *Curcuma*
- psoriasis*: a common chronic, scaly dermatosis with polygenic inheritance and a fluctuating course
- psychomotor*: pertaining to motor effects of cerebral or psychic activity
- psychosis*: a mental disorder marked by gross impairment in reality testing, reflected in delusions, hallucinations, incoherent speech or disorganized and agitated behaviour; also used in a more general sense for mental disorders in which impairment of mental functioning interferes with the capacity to meet the ordinary demands of life
- psychotomimetic*: pertaining to, characterized by, or producing manifestations resembling those of a psychosis, e.g. hallucinations, distortion of perception and schizophrenia-like behaviour
- psychotropic*: affecting the mind
- psyllid*: belonging to the homopterous insect family *Psyllidae*, which includes the jumping plant lice
- puberulent*: covered with down or fine hairs
- puberulous*: minutely pubescent
- pubescent*: covered with soft short hairs
- pulp*: the soft fleshy part of the fruit; mechanically ground or chemically digested wood used in manufacturing paper and allied products
- pulses*: dry edible seeds of legumes
- pulvinate*: cushion-shaped
- punctate*: marked with dots or translucent glands
- pungent*: bearing a sharp point; causing a sharp or irritating sensation
- purgative*: causing evacuation of the bowels; an agent causing evacuation of the bowels, especially through stimulating peristaltic action; also called cathartic
- purine alkaloids*: a subgroup of the alkaloids
- pustular, pustulate*: with blister-like prominences
- pyloric*: pertaining to the pylorus (the opening between the stomach and the small intestine) or to the pyloric part of the stomach
- pyrene*: a nutlet or kernel; the stone of a drupe or similar fruit
- pyrethrins*: pyrethrins are esters of the monoterpenes pyrethic acid and chrysanthemic acid, with a cyclopentenolone bearing an unsaturated side chain; often these compounds are toxic to cold-blooded animals (e.g. fish, insects) but are harmless to man and other warm-blooded animals
- pyrexia*: fever; an abnormal elevation of the body temperature
- pyridine alkaloids*: a subgroup of the alkaloids
- pyriform*: resembling a pear in shape
- pyrrole alkaloids*: a subgroup of the alkaloids
- pyrrolidine alkaloids*: a subgroup of the alkaloids
- pyrrolizidine alkaloids*: a subgroup of the alkaloids
- Q-fever*: an acute rickettsial infection, caused by *Coxiella burnietti* and characterized by fever, chills, headache, myalgia, malaise and, rarely, rash
- quadrangular*: four-cornered or four-edged
- quadrate*: approximately square or cubical
- quassinoids*: a subgroup of the saponins; the aglycone is a modified triterpene which has lost 10 carbons, and thus could be misinterpreted as a diterpene; most quassinoid structures also include a lactone function in the molecule
- quincuncial*: in aestivation partially imbricated of five parts, two being exterior, two interior and the fifth having one margin exterior, the other interior
- quinoline alkaloids*: a subgroup of the alkaloids
- quinones*: a group of oxygen-containing homologues of aromatic derivatives, characterized by a diketo pattern (dione-structure)

- raceme*: an unbranched elongated indeterminate inflorescence with stalked flowers opening from the base upwards
- racemic*: made up of two enantiomorphous isomers (stereoisomers which have molecules which are mirror images of each other) and therefore optically inactive
- racemose*: raceme-like
- rachilla*: a diminutive or secondary axis, e.g. the branch that bears a flower or the stalk of the spikelet in grasses
- rachis* (*plural rachides*): the principal axis of an inflorescence or a compound leaf beyond the peduncle or petiole
- radial*: lengthwise, in a plane that passes through the pith; radiating, as from a centre (cf. tangential)
- radical*: arising from the root, or its crown
- radicle*: the first root of an embryo or germinating seed
- ramenta* (*plural ramenta*): thin, chaffy scale of the epidermis
- ramification*: branching
- ramified*: branched
- ramiflorous*: bearing flowers on the branches
- raphe*: a ridge on a seed, formed by a portion of the funicle that is adnate to the ovule, as in an anatropous ovule
- rash*: a temporary eruption on the skin, as in urticaria
- ratoon*: shoots in perennial crops such as the pineapple, left on the plants after harvest to produce the subsequent crop (ratoon crop)
- ray*: the radiating branch of an umbel; the outer floret of an inflorescence of the *Compositae* with straplike perianth which differs from those in the centre or disk
- receptacle* (*botany*): the flat, concave or convex part of the axis from which the parts of the flower arise
- recombination*: new gene combination as a result of cross-fertilization between individuals differing in genotype
- recurved*: bent or curved downward or backward
- reduced*: subnormal in size; connotes also either a failure to fulfil a normal function, or a diminution the expected number of parts in a set (of stamens, for example)
- reflexed*: abruptly bent or turned downward or backward
- reforestation*: the planting of a formerly forested area with forest trees
- refrigerant*: in medicine: an agent that relieves fever and thirst
- regular*: of a radially symmetrical flower; actinomorphic
- renal calculi*: kidney-stones
- reniform*: kidney-shaped
- repand*: with an undulating margin
- resin*: solid to soft semisolid amorphous fusible flammable substance obtained as exudate or as an extract of plants
- resolvent*: promoting resolution or the dissipation of a pathological growth
- restenosis*: recurrent stenosis, especially of a valve of the heart, after surgical correction of the primary condition
- restorative*: capable of restoring health, strength, consciousness; an agent having this capability
- reticulate*: netted, as when the smallest veins of a leaf are connected together like the meshes of a net
- reticuloendothelium*: the tissue of the reticuloendothelial system (a group of cells, e.g. in liver, spleen and bone marrow, which are able to take up and sequester inert particles and dyes)
- retinaculum* (*plural retinacula*): the funicle in most *Acanthaceae*, which is curved like a hook and retains the seed until maturity
- retorse*: turned or directed backward or downward (opposed to antrorse)
- retrovirus*: any virus belonging to the family of the *Retroviridae*, a family of RNA viruses, most of which are oncogenic
- retuse*: with a shallow notch at a rounded apex
- revolute*: of leaves with the margins, rolled downwards towards the midrib
- rhabdomyosarcoma*: a highly malignant tumour of striated muscle
- rheumatism*: any of various disorders, characterized by inflammation, degeneration, or metabolic derangement of the connective tissue structures of the body, especially the joints and related structures, and accompanied by pain, stiffness or limited mobility of these parts
- rhinitis*: inflammation of the mucous membrane of the nose
- rhizome*: an underground stem which is distinguished from a root by the presence of nodes, buds, and leaves or scales
- rhizosphere*: the area of soil which immediately surrounds plant roots and is influenced by them
- rhombic*: shaped like a rhomb, an equilateral oblique-angled figure
- rhomboid* (*botany*): quadrangular, diamond-shaped with the lateral angles obtuse
- ringworm*: popular name for tinea, which is a term used to describe various fungal skin infections;



- the name refers to the ring-shaped lesions
- riparian*: growing on the banks of streams or rivers
- riverine forest*: = riparian forest
- root sucker*: a shoot originating from adventitious buds on the roots
- rootstock*: see rhizome; a stock for grafting consisting of a root and part of the main axis
- rosette*: a cluster of leaves or other organs in a circular form
- rosulate*: collected in a rosette
- rot*: disintegration of tissue due to the action of invading organisms, usually bacteria or fungi; a disease so characterized
- rotate*: wheel-shaped; circular and flat
- rotund*: rounded in outline, somewhat orbicular, but a little inclined towards oblong
- rubefacient*: reddening the skin by causing hyperaemia (an excess of blood); an agent that reddens the skin
- rudimentary*: of organs, imperfectly developed and non-functional
- rufous*: reddish
- rugose*: wrinkled
- ruminant*: an animal that chews again what has been swallowed (e.g. sheep, cows, camels, goats)
- ruminant*: of endosperm, mottled in appearance, due to the infolding of a dark inner layer of the seed-coat into the paler coloured endosperm
- runner*: a specialized stem that develops from a leaf axil at the crown of a plant, grows horizontally along the ground, and forms a new plant at one of the nodes, usually at or near the tip (as in strawberry)
- rust*: a disease caused by, and a species in, the class *Urediniomycetes*, order *Uredinales*; so called because of the yellowish to orange brown colour of the spores
- sagittate*: shaped like an arrowhead; of a leaf base with two acute straight lobes directed downwards
- saluretic*: pertaining to, characterized by or promoting the excretion of sodium and chloride ions in the urine (saluresis)
- sapling*: a young tree of more than 1.5 m tall and with a bole of less than 10 cm in diameter
- saponin*: a glycoside with soap properties
- saponins*: the term is applied to a group of glycosides which have the ability to lower the surface tension of aqueous solutions
- sapwood*: the outer layers of wood adjacent to the bark which in the living tree contain living cells and reserve materials
- sarcoma*: any of a group of tumours usually arising from connective tissue, most of which are malignant
- sarcotesta*: the fleshy outer seed-coat
- scaberulous*: somewhat rough
- scabies*: a contagious dermatitis caused by the itch mite (*Sarcoptes scabiei*) that burrows under the skin and deposit eggs, causing intense itching
- scabrid*, *scabrous*: rough to the touch
- scabridulous*: slightly rough
- scalariform*: ladder-like, having markings or perforations suggestive of a ladder
- scale*: a thin scarious body, often a degenerate leaf or a trichome of epidermal origin
- scandent*: climbing
- scape*: a leafless floral axis or peduncle arising from the ground
- scarification*: scratching or making incisions, e.g. to harvest latex from *Papaver somniferum*; of seed, the cutting or softening of the wall of a hard seed to hasten germination
- scarify*: to treat a hard-coated seed by mechanical abrasion or with acid to facilitate germination
- schistosomiasis*: infection with flukes of the genus *Schistosoma*; sometimes called bilharzia
- schizocarpous*: in the form of a schizocarp
- schizonticide*: an agent that destroys schizonts, i.e. specific development stages or forms of certain protozoa
- sciatica*: pain in the lower back, buttocks, hips or adjacent parts of the body
- scion*: the plant being propagated vegetatively in grafting; the part of the plant above the graft union
- scleroderma*: chronic hardening and thickening of the skin, which may be a finding in several different diseases
- scorpioid*: circinate; coiled as to resemble a scorpion
- scrofula*: tuberculosis of the lymph nodes of the neck
- scrub*: vegetation whose growth is stunted because of lack of water coupled with strong transpiration
- scurf*: abnormal skin condition in which small flakes or scales become detached
- scurvy*: a disease resulting from a deficiency of vitamin C in the body, characterized by weakness, anaemia, spongy gums, bleeding from mucous membrane, etc.
- scutelliform*: platter-shaped
- seborrhoea*: excessive secretion of an oily substance (sebum) from certain glands of the skin (sebaceous glands), which are abundant on the scalp, face, chest, back, armpit and groin

- seco-iridoids*: see iridoids
- secondary immunization*: second contact with the same antigen
- secondary vegetation*: a plant cover that has been disturbed by natural causes or by man
- section (botany)*: a taxonomic rank between the genus and the species accommodating a single or several related species
- sedative*: allaying activity and excitement; an agent that allays excitement
- seed*: the reproductive unit formed from a fertilized ovule, consisting of embryo and seed-coat, and, in some cases, also endosperm
- seedling*: a plant produced from seed; a juvenile plant, grown from a seed
- segment*: one of the divisions into which a plant organ, as a leaf or a calyx, may be cleft; the division of a palmate or costapalmate leaf
- self-compatible*: capable of fertilization and setting seed after self-pollination
- self-fertile*: capable of fertilization and setting seed after self-pollination
- self-incompatible*: self-sterile, i.e. not capable of producing seed without cross-pollination
- self-pollination*: pollination with pollen from the same flower or from other flowers of plants of the same clone
- selfing*: fertilization of female gametes with male gametes from the same individual
- semi-*: prefix, meaning half or incompletely, e.g. semi-inferior
- seminal vesicle*: either of the paired, sacculated pouches attached to the posterior part of the urinary bladder
- senescence*: advancing in age
- sensu lato (s.l.)*: in the broad sense
- sensu stricto (s.s.)*: in the strict sense
- sepal*: a member of the outer series of perianth segments
- sepsis*: the presence of pathogens or their toxins in the blood or other tissues
- septate*: divided by one or more partitions
- septicidal*: dehiscing along the septa of the ovary
- septum (plural septa)*: a partition or cross-wall
- seriate*: serial, disposed in series of rows
- sericeous*: silky
- serotonergic, serotoninergic*: containing or activated by serotonin
- serrate*: toothed like a saw, with regular pointed teeth pointing forwards
- sesquiterpenes*: a subgroup of the isoprenoids, formed by coupling of 3 C<sub>5</sub> units
- sessile*: without a stalk
- setose*: set with bristles or bristle-like elements
- setulose*: set with small bristles or bristle-like elements
- sheath*: a tubular structure surrounding an organ or part, as the lower part of the leaf clasping the stem in grasses
- sheathing organ*: any sheathing structure inserted at a node of any vegetative or reproductive axis in a gramineous plant; among the bamboos, distinguishable types of sheathing organs are rhizome sheaths, neck sheaths, culm sheaths, branch sheaths, leaf sheaths, prophylla, bracts, empty glumes, lemmas, and paleas
- Shikimic acid pathway*: shikimic acid is formed from the primary metabolism intermediates phosphoenolpyruvate and erythrose-4-phosphate; the shikimic acid pathway leads to a variety of secondary metabolites e.g. phenols, cinnamic acids and other phenylpropane derivatives, tannins and coumarins
- shingles*: an acute, infectious skin disease, characterized by neuralgia and eruptions sometimes extending half round the body like a girdle; also called herpes zoster
- shoot*: the ascending axis, when segmented into dissimilar members it becomes a stem; a young growing branch or twig
- shrub*: a woody plant which branches from the base, all branches being equivalent (see also tree)
- sialagogue*: an agent promoting the flow of saliva
- side graft*: a graft made by any of various methods in which the scion is inserted into the side of the stock, which is not beheaded until the union is complete
- sigmoid, sigmoidal*: doubly curved in opposite directions, like the letter s
- silicosis*: a pulmonary disease due to the inhalation of the dust of stone, sand or flint containing silicon dioxide
- simple (botany)*: not compound, as in leaves with a single blade
- sinuate*: with a deep wavy margin
- sinuous*: wavy
- sinusitis*: inflammation of a sinus (cavity)
- solitary*: single stemmed, not clustering
- somatic embryogenesis*: the production of embryo-like structures (embryoids) from sporophytic or somatic cells of the plant, as opposed to gametophytic or germ cells (zygotic embryogenesis)
- sooty mould*: saprophytic fungus of the family *Capnodiaceae* or other families of the order *Dothideales*, which forms superficial, brown to black colonies on living plants, is often associated with insect secretion, and can be detrimental

- to the plant
- sporific*: a drug or other agent that induces sleep
- sore*: popular term for almost any lesion of the skin or mucous membranes
- spadix*: a flower spike with a fleshy or thickened axis, as in aroids and some palms
- spasmodic*: of the nature of a spasm, i.e. a sudden, violent, involuntary contraction of a muscle or of a group of muscles
- spasmolytic*: checking spasms; antispasmodic
- spastic*: of the nature of or characterized by spasms
- spat(h)ulate*: spoon-shaped
- spathaceous*: resembling a spathe
- spathe*: a large bract enclosing a spadix, or two or more bracts enclosing a flower cluster
- spectroscopy*: examination by means of a spectroscope, i.e. an optical instrument for forming and analysing spectra emitted by substances or bodies
- spermatogenesis*: the process of formation of spermatozoa
- spermatorrhoea*: involuntary, abnormally frequent, and excessive emission of semen without copulation
- spherical*: globular
- spicate*: spike-like
- spiciform*: with the form of a spike
- spike*: a simple indeterminate inflorescence with sessile flowers along a single axis
- spikelet*: a secondary spike, one of the units of which the inflorescence is made in grasses, consisting of one or more florets on a thin axis, subtended by a common pair of glumes
- spine (botany)*: a short, stiff, straight, sharp-pointed, hard structure usually arising from the wood of a stem
- spinose, spinous*: having spines
- spiral*: as though wound round an axis
- splenocyte*: the monocyte (mononuclear phagocytic leucocyte) characteristic of the spleen
- spore*: in cryptogams a cell which becomes free and capable of direct development into a new bion; the analogue of seed in phanerogams
- sporozoite*: the elongate, nucleated, motile infective stage of certain protozoa, like *Plasmodium* spp.
- sprue*: a chronic deficiency syndrome due to subnormal absorption of dietary constituents
- stain*: discoloration or variation from natural colour due to fungi, chemical action or other causes
- stamen*: one of the male reproductive organs of a flower; a unit of the androecium
- staminode*: an abortive or rudimentary stamen without or with an imperfect anther
- standard (botany)*: the fifth, posterior or upper petal of a papilionaceous corolla
- starch*: polysaccharide made up of a long chain of glucose units joined by  $\alpha$ -1,4 linkages, either unbranched (amylose) or branched (amylopectin) at a  $\alpha$ -1,6 linkage, and which is the storage carbohydrate in plants, occurring as starch granules in amyloplasts, and which is hydrolysed by animals during digestion by amylases, maltase and dextrinases to glucose via dextrins and maltose
- stellate*: star-shaped, as of hairs with radiating branches, or of petals arranged in the form of a star
- stem*: the main ascending axis of a plant; in bamboos usually named culm, in other plant groups occasionally
- stenocardia*: angina pectoris
- stenosis*: narrowing or stricture of a duct or canal
- stereoisomer*: one of two or more isomers that have the same structure (linkages between atoms) but different configurations (spatial arrangements)
- steric*: of or pertaining to the 3-dimensional arrangement of the atoms in a molecule
- sterile*: unable to produce offspring; in plants: failing to complete fertilization and produce seed as a result of defective pollen or ovules; not producing seed capable of germination; lacking functional sexual organs (sterility)
- steroid saponins*: a subgroup of the saponins
- steroidal alkaloids*: a subgroup of the alkaloids
- steroids*: a group of modified triterpenes which contain a ring system of three 6-membered and one 5-membered rings
- stigma*: the portion of the pistil which receives the pollen
- stilt root*: an oblique adventitious root as in mangrove trees and similar forms
- stimulant*: producing a temporary increase of the functional activity or efficiency of an organism or any of its parts; an agent acting so
- stipe*: the stalk supporting a carpel or gynoeceium
- stipule*: small secondary stipule at the base of a leaflet
- stipitate*: borne on a stipe or short stalk
- stipulate*: with or bearing stipules
- stipule*: a scale-like or leaf-like appendage at the base of a petiole
- stolon*: a trailing stem usually above the ground which is capable of producing roots and shoots at its nodes

- stoloniferous*: bearing a stolon or stolons
- stoma (plural stomata)*: a breathing pore or aperture in the epidermis
- stomachic*: pertaining to the stomach; a medicine stimulating the action of the stomach
- straggling*: extremely divergent, spreading very far apart; irregular, bushy
- strain*: a group of individuals of a common origin, usually a more narrowly defined group than a cultivar
- strangury*: slow and painful urination
- stratification*: a moist, cold treatment of seed to overcome physiological dormancy
- striate*: marked with fine longitudinal parallel lines, as grooves or ridges
- strigose*: with short stiff hairs lying close along the surface
- strongyloidiasis*: infection with *Strongyloides stercoralis*, a roundworm which occurs widely in the tropics and subtropics and causes diarrhoea and ulceration of the small intestine
- stump*: seedling with trimmed roots and shoot and used as planting stock; the part of anything that remains after the main part has been removed, e.g. the part of a tree remaining attached to the root after the trunk is cut
- style*: the part of the pistil connecting the ovary with the stigma
- styptic*: astringent, tending to check bleeding through astringent properties; a remedy which is astringent and arrests bleeding
- sub-*: prefix, meaning somewhat or slightly (e.g. subacute), or below (e.g. subterranean) or less than, imperfectly
- subfamily*: a taxonomic rank between the family and the tribe denoting a part of a family
- subglobose*: nearly globular
- subshrub*: a small shrub which may have partially herbaceous stems
- subspecies*: a subdivision of a species, in rank between a variety and a species
- subulate*: awl-shaped, sharply pointed
- succulent*: juicy, fleshy
- sucher*: a shoot, usually originating from adventitious buds on the roots or basal stem parts, which does not fit in the architectural model, but is capable of repeating the model
- sudorific*: causing or promoting the flow of sweat; an agent causing sweating
- suffrutescent*: obscurely shrubby
- superior*: of an ovary, with the perianth inserted below or around its base, the ovary being attached at its base only
- supraventricular*: above the ventricles (lower heart chambers), especially applied to rhythms originating from centres proximal to the ventricles
- suture*: the line of junction of two carpels; the line or mark of splitting open
- syconium*: a multiple, hollow fruit, like a fig
- symbiosis*: the intimate living together of two dissimilar organisms in a mutually beneficial relationship
- sympatholytic*: opposing the effects of impulses conveyed by adrenergic postganglionic fibres of the sympathetic nervous system
- sympathomimetic*: mimicking the effects of impulses conveyed by adrenergic postganglionic fibres of the sympathetic nervous system
- sympatrically*: occupying an area together with another species
- sympodial*: of a stem in which the growing point either terminates in an inflorescence or dies, growth being continued by a new lateral growing point; in bamboos also used to designate the branching habit of a rhizome (see pachymorph)
- synandrium*: the cohesion of the anthers of each male flower, e.g. in certain *Araceae*
- synapse*: the site of functional apposition between neurons or between neurons and effector organs, at which an impulse is transmitted, usually by a chemical neurotransmitter
- syncarp*: a multiple or fleshy aggregate fruit, including fruit produced from a more or less entire inflorescence (as in *Artocarpus*, *Ananas*, *Morus*)
- synergistic effect*: the phenomenon of a mutually cooperating activity of substances, which in a mixture produce a greater effect than when taken alone
- syphilis*: a disease usually communicated by sexual contact, or via the blood or bite of an infected person, caused by a spirochete (*Treponema pallidum*) and characterized by a clinical course in 3 stages continued over many years
- tachycardia*: excessive rapidity of the heartbeat, usually applied to a pulse rate of more than 100 in an adult
- tachypnea*: excessive rapidity of respiration
- taeniacide*: an agent that destroys tapeworms
- taeniafuge*: an agent expelling tapeworms
- tannins*: a large group of plant-derived phenolic compounds
- taproot*: the primary descending root, forming a direct continuation of the radicle
- taxon (plural taxa)*: a term applied to any taxonomic unit irrespective of its classification level, e.g. variety, species, genus, etc.
- taxonomy*: the study of principles and practice of

- classifying living organisms (systematics)
- tendril*: a thread-like climbing organ formed from the whole or part of a stem, leaf or petiole
- tepal*: a segment of a perianth, applied when no distinction between sepals and petals can be made
- teratogenic*: tending to produce anomalies of formation or development
- teratological*: pertaining to teratology, i.e. that division of embryology and pathology which deals with abnormal development and congenital anomalies
- terete*: cylindrical; circular in transverse section
- terminal*: placed at the end or apex; a termination, end or extremity
- termite*: ant-like organism of the order *Isoptera* damaging wood by characteristic irregular honeycombing or wide channels with dry bore-dust or dust cemented together
- ternate*: in threes
- terpenes*: see isoprenoids
- terpenoids*: see isoprenoids
- terrestrial*: on or in the ground
- tertiary venation*: generally the collection of the smallest veins of a leaf blade
- testa*: the outer coat of the seed
- tetanic spasm*: physiological tetanus; a state of sustained muscular contraction without periods of relaxation
- tetanus*: an acute, often fatal, infectious disease characterized by muscular contractions and abnormal reflexes, and caused by a toxin produced by *Clostridium tetani*, a bacillus which is usually introduced through a wound
- tetraploid*: having four times ( $4n$ ) the basic number of chromosomes or twice the diploid number ( $2n$ )
- theca (plural thecae)*: a spore- or pollen-case
- theileriasis*: a group of diseases due to protozoa of the genus *Theileria*, which cause an infection with fever
- thinning*: removing trees, stems or plants from immature or mature stands in order to stimulate the growth of the remaining trees, stems or plants
- thorn*: a woody sharp-pointed structure formed from a modified branch
- throat (botany)*: of a corolla, the orifice of a gamopetalous corolla
- thrombocyte*: a blood platelet
- thrombocytopenia*: a decrease in the number of blood platelets
- thrombosis*: the formation, development or presence of an aggregation of blood factors (thrombus), often causing vascular obstruction
- thrombotic*: pertaining to or affected with thrombosis
- thromboxanes*: see prostaglandins
- thrombus*: an aggregation of blood factors, frequently causing vascular obstruction at the point of its formation
- thrush*: infection of the mucous membrane of the mouth with a fungus of the genus *Candida*, especially *C. albicans*, and characterized by the formation of creamy, white, somewhat elevated lesions
- thyrses (thyrsus)*: a compound inflorescence composed of a panicle (indeterminate axis) with the secondary and ultimate axes cymose (determinate)
- thyrsoid*: like a thyrses
- tiller*: a shoot from the axils of the lower leaves, e.g. in some grasses and palms (making such shoots: tillering)
- timber*: any wood other than fuelwood
- tincture*: an alcoholic or hydroalcoholic solution of some principle used in medicine
- tinnitus*: a noise in the ears, like ringing, buzzing, roaring or clicking
- tissue culture*: a body of tissue growing in a culture medium outside the organism
- tomentose*: densely covered with short soft hairs
- tomentulose*: slightly tomentose
- tongue graft*: a graft in which a tongue cut on the scion is fitted into a slit cut slopingly in the stock
- tonic*: restoring or producing the normal tone (degree of vigour and tension) of tissue or organs; characterized by continuous tension (e.g. tonic spasm); medicinal preparation believed to have the power of restoring normal tone to tissue or organs
- tonsillitis*: inflammation of the tonsils
- topical*: pertaining to a particular surface area, as a topical anti-infective applied to a certain area of the skin and affecting only the area to which it is applied
- tortuous*: bent or twisted in different directions
- toxigenic, toxicogenic*: producing or elaborating toxins
- trailing*: prostrate, but not rooting
- tranquillizer*: a drug with a calming, soothing effect
- transgenic*: pertaining to the experimental splicing of a segment of DNA from one genome to DNA of a different genome
- transverse*: straight across; of tertiary veins, connecting the secondary veins, not necessarily in a

- perpendicular way
- trapezoid*: like a trapezium, a figure of four unequal sides
- trauma*: a wound or injury, whether physical or psychic
- tree*: a perennial woody plant with a single evident trunk (see also shrub)
- trematodiasis*: infection with trematodes, i.e. parasitic animal organisms belonging to the *Trematoda*, a class of *Platyhelminthes* including the flukes
- tribe (plural tribae)*: a taxonomic rank between the family and the genus
- trifid*: cleft in three parts
- trigonal*: three-angled, with plane faces
- triploid*: having three times the basic number of chromosomes, usually written  $3n$
- triquetrous*: three-edged, with three salient angles
- tristichous*: in 3 vertical ranks
- triterpene saponins*: a subgroup of the saponins
- triterpenes*: a subgroup of the isoprenoids, formed by coupling of 6  $C_5$  units
- tropane alkaloids*: a subgroup of the alkaloids
- trophozoite*: the active, motile, feeding stage of a protozoan organism, as contrasted with the non-motile encysted stage
- truncate*: cut off more or less squarely at the end
- trunk*: the main stem of a tree apart from its limbs and roots
- trypanosomiasis*: the state of being infected with protozoa of the genus *Trypanosoma* (trypanosomes), which destroy the red corpuscles and cause serious and even fatal diseases, as the sleeping sickness
- tuber*: the swollen portion of an underground stem or root which acts as a storage organ and propagule; it is usually of one year's duration, those of successive years not arising directly from the old ones nor bearing any constant relation to them
- tubercle*: a small tuber-like excrescence
- tuberculate*: covered with warty protuberances
- tuberculosis*: any of the diseases in man and animals caused by *Mycobacterium* spp, characterized by the formation of lesions (tubercles) and necrosis in the tissue of the lung or other organs and having a tendency to great chronicity
- tuberculostatic*: inhibiting the growth of *Mycobacterium tuberculosis*, the causative agent of tuberculosis
- tuberous*: producing tubers or resembling a tuber
- tufted*: growing in tufts (caespitose)
- tumorigenic*: giving rise to either benign or malignant tumours
- tumour necrosis factor*: a substance (lymphokine) produced by macrophages, capable of causing in vivo haemorrhagic necrosis of certain tumour cells, but not affecting normal cells
- tunic*: the coat of a bulb
- turbinate*: top-shaped
- turgid*: swollen, but not with air
- twining*: winding spirally
- tympanites*: swelling of the abdomen, due to the accumulation of gas or air in the intestine or in the peritoneal cavity
- tympanitis*: inflammation of the lining membrane of the tympanum (middle ear)
- ulcer*: an open sore on an external or internal body surface, usually accompanied by disintegration of tissue and formation of pus
- umbel*: an indeterminate, often flat-topped inflorescence whose divergent peduncles (rays) and pedicels arise from a common point; in a compound umbel each ray itself bears an umbellule (small umbel)
- umbelliform*: umbrella-shaped
- umbellule*: diminutive of umbel
- unarmed*: devoid of thorns, spines or prickles
- undershrub*: any low shrub; partially herbaceous shrub, the ends of the branches perishing during the winter
- undulate*: wavy, said for instance of a leaf margin if the waves run in a plane at right angles to the plane of the leaf blade
- unifoliolate*: with one leaflet only, but in origin a compound leaf
- unilateral*: one-sided
- unilocular*: one-celled
- uniparous*: bearing one, as a cyme giving forth one axis at each branching
- unisexual*: of one sex, having stamens or pistils only
- urceolate*: urn-shaped
- urticaria*: a vascular reaction, acute or chronic, which can have various causes and is characterized by the development of weals on the skin
- uterotonic*: giving muscular tone to the uterus
- utricle*: a small bladder pericarp
- uveitis*: inflammation of the uvea, the vascular middle coat of the eye
- vagotomy*: interruption of the impulses carried by the vagus nerve or nerves
- valvate*: of perianth segments, with their edges in contact, but not overlapping in the bud
- valve*: one of the parts produced by a dehiscing capsule
- variegated*: irregularly coloured in patches, blotched

- variety*: a botanical variety which is a subdivision of a species; an agricultural or horticultural variety is referred to as a cultivar
- vas deferens*: the excretory duct of the testis, which unites with the excretory duct of the seminal vesicle to form the ejaculatory duct; also called ductus deferens (deferent duct)
- vasoconstriction*: diminution of the calibre of vessels, especially of arterioles
- vasodilatation*: a state of increased calibre of the blood vessels
- vasodilation*: dilation of a vessel, especially dilation of arterioles leading to increased blood flow to a part
- vasomotor centres*: centres in the central nervous system that regulate the calibre of the blood vessels and increase or decrease the heart rate and contractility
- vein (botany)*: a strand of vascular tissue in a flat organ, such as a leaf
- velutinous*: see velvety
- velvety*: with a coating of fine soft hairs; the same as tomentose but denser so that the surface resembles (and feels like) velvet
- venation (botany)*: the arrangement of the veins in a leaf
- venereal*: pertaining or related to or transmitted by sexual contact
- venereal disease*: any of a diverse group of contagious diseases (as gonorrhoea or syphilis) that are typically transmitted by sexual contact
- venous*: of or pertaining to the veins
- ventral*: in botany: facing the central axis (adaxial), opposed to dorsal (abaxial); in human anatomy: pertaining to the abdomen, or denoting a position more toward the belly surface than some reference object
- ventricose*: with a swelling or inflation on one side
- ventricular*: pertaining to a ventricle, i.e. a small cavity, such as one of the several cavities of the brain, or one of the lower chambers of the heart
- vermiculite*: lightweight highly water-absorbent material, usually resulting from expansion of the granules of mica at high temperature
- vermifuge*: an agent expelling worms or intestinal animal parasites; an anthelmintic
- verrucose*: warty
- verruculose*: very warty, much covered with warts
- verticillaster*: a false whorl, composed of a pair of opposed cymes, as in *Labiatae*
- verticillate*: in a whorl with several elements arising at the same node
- vertigo*: an illusory sense that the surroundings or one's own body are revolving
- vesicant*: causing blisters; an agent that induces blistering
- vesicle (botany)*: a small bladder or cavity
- vesicular arbuscular mycorrhiza*: a common endomycorrhizal association characterized by 2 types of fungal structures: small structures within root cells known as arbuscules, and storage organs between root cells known as vesicles
- vesicular stomatitis*: a vesicular eruption caused by a virus and affecting pigs, cattle and horses
- vestigial*: small and imperfectly developed
- vexillum*: see standard
- viability*: ability to live, grow and develop
- villose (villous)*: with long weak hairs
- vine*: a plant having a stem that is too slender to hold itself erect and therefore supports itself by climbing over an object; the stem itself
- viricidal*: capable of neutralizing or destroying a virus
- virion*: complete, individual virus particle, found extracellularly and capable of surviving in crystalline form and infecting a living cell
- vitiligo*: a chronic, usually progressive, pigmentary anomaly of the skin, manifested by depigmented white patches that may be surrounded by a hyperpigmented border
- volatile*: a volatile substance is one that evaporates at room temperature. It is an essential property of odorous materials
- volatile oils*: see essential oils
- vulnerary*: pertaining to wounds or the healing of wounds; an agent promoting the healing of wounds
- wart (in medicine)*: a small, usually hard and non-malignant, excrescence on the skin
- warty*: covered with firm roundish excrescences
- waterlogged*: flooded with water, generally for a period of at least a few weeks
- whipworm: Trichuris trichiura*, an intestinal nematode parasite
- whorl*: arrangement with more than two organs of the same kind arising at the same level
- wilt*: loss of turgidity, usually in leaves, typically caused by pathogens which colonize the vascular system
- wing*: any membraneous expansion attached to an organ; a lateral petal of a papilionaceous corolla
- withdrawal syndrome*: a substance-specific organic mental syndrome that follows the cessation of use or reduction of intake of a psychoactive substance that had been regularly used to induce a state of intoxication
- wood*: the hard, compact, fibrous substance between pith and bark

*woolly*: clothed with long and tortuous or matted hairs

*wormicidal*: destroying worms

*xenobiotic*: a chemical foreign to the biological system

*xerophthalmia*: dryness of the conjunctiva and cornea (the transparent anterior part of the eyeball) due to vitamin A deficiency

*xerophytic*: relating to a plant structurally adapted for life and growth with a limited water supply

*xerosis*: abnormal dryness, as of the eye, mouth or skin

*xerostomia*: dryness of the mouth due to malfunction of salivary glands

*yaws*: an infectious, tropical disease caused by a spirochete (*Treponema pertenue*), usually affecting children under 15, and marked by skin elevations (papules) and papilloma, with later manifestations including deformation of skin, bone and joints (also called framboesia)

*zooflagellate*: an animal-like flagellate protozoan of the class *Zoomastigophorea*

*zygomorphic*: irregular and divisible into equal halves in one plane only



## Sources of illustrations

- Abrus precatorius*: Westphal, E. & Jansen, P.C.M. (Editors): Plant Resources of South-East Asia. A selection. Pudoc, Wageningen, The Netherlands. p. 27 (young shoot, inflorescence, infructescence, seeds).
- Achillea millefolium*: Ross-Craig, S., 1961. Drawings of British plants, part XVI. G. Bell and Sons, London, United Kingdom. pl. 1 (plant habit, middle part of stem with leaf, upper part of flowering stem, ray flower and involucre bract, disk flower and receptacle scale, achene from disk flower, achene from ray flower). Redrawn and adapted by Iskak Syamsudin.
- Acorus calamus*: Cook, C.D.K., Gut, B.J., Rix, E.M., Schneller, J. & Seitz, M., 1974. Water plants of the world. A manual for the identification of the genera of freshwater macrophytes. Dr. W. Junk Publishers, The Hague, The Netherlands. p. 139, fig. 49 (plant habit); Dahlgren, R.M.T., Clifford, H.T. & Yeo, P.F., 1985. The families of the Monocotyledons. Structure, evolution, and taxonomy. Springer Verlag, Berlin, Heidelberg, New York, Tokyo. p. 276, fig. 127A-B (inflorescence, flower). Redrawn and adapted by Achmad Satiri Nurhaman.
- Aerva lanata*: Backer, C.A., 1935. Onkruidflora der Javasche suikerrietgronden. Handboek ten dienste van de suikerriet cultuur en de rietsuiker fabricage op Java. Deel 7(8) (atlas) [Weed flora of Javanese sugar-cane fields. Handbook for the cultivation of sugar-cane and manufacturing of cane-sugar in Java. Vol. 7(8) (atlas)]. Vereniging het Proefstation voor de Java-Suikerindustrie, Pasuruan, Indonesia. pl. 235 (plant habit, flower); Matthew, K.M., 1988. Further illustrations on the Flora of Tamilnadu Carnatic. The Rapinat Herbarium, St. Joseph's College, Tiruchirapalli, India. p. 506, pl. 506 (branchlet with young inflorescences, opened staminal tube with stamens and staminodes). Redrawn and adapted by Achmad Satiri Nurhaman.
- Ageratum conyzoides*: Holm, L.G., Plucknett, D.L., Pancho, J.V. & Herberger, J.P., 1977. The world's worst weeds. Distribution and biology. East-West Center. The University Press of Hawaii, Honolulu, United States. p. 147, fig. 55 (plant habit); Soerjani, M., Kostermans, A.J. G.H. & Tjitrosoepomo, G. (Editors), 1987. Weeds of rice in Indonesia. Balai Pustaka, Jakarta, Indonesia. p. 61, fig. 4.16 (flowering head, flower, fruit with pappus). Redrawn and adapted by Achmad Satiri Nurhaman.
- Allium cepa* cv. group *Aggregatum*: Siemonsma, J.S. & Kasem Piluek (Editors), 1993. Plant resources of South-East Asia No 8. Vegetables. Pudoc Scientific Publishers, Wageningen, The Netherlands. p. 65 (flowering plants, inflorescence, mature bulbs).
- Allium sativum*: Siemonsma, J.S. & Kasem Piluek (Editors), 1993. Plant resources of South-East Asia No 8. Vegetables. Pudoc Scientific Publishers, Wageningen, The Netherlands. p. 78 (plant habit, bulb, plantlets, inflorescence).
- Aloe vera*: Wijnands, D.O., 1983. The botany of the Commelins. A.A. Balkema, Rotterdam, The Netherlands. p. 128 (plant habit); Baillon, H., 1892. Histoire des plantes [History of plants]. Vol. 12. Hachette & Cie, London, United Kingdom. p. 439, fig. 471 (part of inflorescence); Krause, K., 1930. Liliaceae. In: Engler, A. & Prantl, K. (Editors): Die natürlichen Pflanzenfamilien [The natural plant families]. Second edition, Band 15A. Wilhelm Engelmann, Leipzig, Germany. p. 306, fig. 115 (flower in longitudinal section).
- Alternanthera sessilis*: Backer, C.A., 1935. Onkruidflora der Javasche suikerrietgronden. Handboek ten dienste van de suikerriet cultuur en de rietsuiker fabricage op Java. Deel 7(8) (atlas) [Weed flora of Javanese sugar-cane fields. Handbook for the cultivation of sugar-cane and manufacturing of cane-sugar in Java. Vol. 7(8) (atlas)]. Vereniging het Proefstation voor de Java-Suikerindustrie, Pasuruan, Indonesia. pl. 241 (plant habit); Larsen, K., 1989. Caryophyllales. In: Lescot, M. (Editor): Flore du Cambodge, du Laos et du Vietnam [Flora of Cambodia, Laos and Vietnam]. Vol. 24. Muséum Na-

- tionale d'Histoire Naturelle, Paris, France. p. 51, pl. 7 (flower with bract and bracteoles, flower with tepals removed); Matthew, K.M., 1982. Illustrations on the Flora of Tamilnadu Carnatic. The Rapinat Herbarium, St. Joseph's College, Tiruchirapalli, India. pl. 584 (fruit). Redrawn and adapted by Achmad Satiri Nurhaman.
- Amaranthus spinosus*: Backer, C.A., 1935. Onkruidflora der Javasche suikerrietgronden. Handboek ten dienste van de suikerriet cultuur en de rietsuiker fabricage op Java. Deel 7(8) (atlas) [Weed flora of Javanese sugar-cane fields. Handbook for the cultivation of sugar-cane and manufacturing of cane-sugar in Java. Vol. 7(8) (atlas)]. Vereniging het Proefstation voor de Java-Suikerindustrie, Pasuruan, Indonesia. pl. 229 (part of flowering plant); Zohary, M., 1966. Flora Palestina. Part. 1, plates. The Israel Academy of Sciences and Humanities. pl. 266 (male flower with bracteoles); Townsend, C.C., 1974. Amaranthaceae. In: Nasir, E. & Ali, S.I. (Editors): Flora of West Pakistan. No 71. Steward Herbarium, Gordon College, Rawalpindi, and Department of Botany, University of Karachi, Karachi, Pakistan. fig. 2D (fruit). Redrawn and adapted by Achmad Satiri Nurhaman.
- Amomum hochreutineri*: Valetton, T., 1906. Icones Bogorienses. Vol. 2(4). E.J. Brill, Leiden, The Netherlands. pl. 195 (stem base with inflorescence, flower, stamen, infructescence). Redrawn and adapted by Achmad Satiri Nurhaman.
- Amomum villosum*: Beijing Botanical Research Institute, 1976. Iconographia Cormophytorum Sinicorum [Line drawings of Chinese vascular plants]. Vol. 5. Beijing, China. p. 593, pl. 8015 (plant base with infructescence, part of leafy stem, flower). Redrawn and adapted by Achmad Satiri Nurhaman.
- Andrographis paniculata*: Backer, C.A. & van Steenis, C.G.G.J., 1973. Atlas of 220 weeds of sugar-cane fields in Java. Greshoff's Rumphius Fund, Amsterdam, The Netherlands. pl. 628 (flowering stem); Barker, R.M., 1986. A taxonomic revision of Australian Acanthaceae. Journal of the Adelaide Botanic Garden 9. p. 126, fig. 19 (flower with detail of style and stamen arrangement, seed, dehisced fruit); Hansen, B., 1985. Notes on *Andrographis* and *Gymnostachyum* (Acanthaceae). Nordic Journal of Botany 5. p. 353, fig. 1 (flower). Redrawn and adapted by Iskak Syamsudin.
- Angelica acutiloba*: Kariyone, T. & Koiso, R., 1971. Atlas of medicinal plants. Takeda Chemical Industries, Osaka, Japan. pl. 68 (root, stem with flowers and fruits). Redrawn and adapted by Iskak Syamsudin.
- Antiaris toxicaria*: Greshoff, M., 1894. Schetsen nuttige Indische planten [Sketches of useful Indonesian plants]. Series 1. Extra Bulletin van het Koloniaal Museum. J.H. De Bussy, Amsterdam, The Netherlands. pl. 15 (fertile twig); Berg, C.C., 1977. Revision of African Moraceae (excluding *Dorstenia*, *Ficus*, *Musanga* and *Myrianthus*). Bulletin du Jardin Botanique National de Belgique 47. p. 315, fig. 9 (female inflorescence); Chew, W.L., 1989. Moraceae. Flora of Australia. Vol. 3. Hammelidales to Casuarinales. Australian Government Publishing Service, Canberra, Australia. p. 25, fig. 29 E (male inflorescence). Redrawn and adapted by Iskak Syamsudin.
- Arcangelisia flava*: Westphal, E. & Jansen, P.C.M. (Editors): Plant Resources of South-East Asia. A selection. Pudoc, Wageningen, The Netherlands. p. 49 (intertwining flowering and fruiting stems, closed male flower, opened male flower, part of woody stem, detail of stem in cross and longitudinal section).
- Aristolochia tagala*: Beijing Botanical Research Institute, 1972. Iconographia Cormophytorum Sinicorum [Line drawings of Chinese vascular plants]. Vol. 1. Beijing, China. p. 548, pl. 1096 (flowering stem, fruiting stem); Phuphathana-phong, L., 1987. Aristolochiaceae. In: Smitinand, T. & Larsen, K. (Editors): Flora of Thailand. Vol. 5, Part 1. The Forest Herbarium, Royal Forest Department, Bangkok, Thailand. p. 16, fig. 10 (dehisced fruit). Redrawn and adapted by Achmad Satiri Nurhaman.
- Artemisia annua*: Hess, H.E., Landolt, E. & Hirzel, R., 1972. Flora der Schweiz un angrenzender Gebiete [Flora of Switzerland and neighbouring regions]. Band 3. Birkhäuser Verlag, Basel & Stuttgart, Switzerland & Germany. p. 580 (flowering stem, stem base and roots); Hegi, G., 1929. Illustrierte Flora von Mittel Europa. VI. Band, 2. Hälfte [Illustrated flora of Central Europe, 6st Volume, 2nd half]. J.F. Lehmanns Verlag, München, Germany. p. 630, fig. 341 (flower head). Redrawn and adapted by Achmad Satiri Nurhaman.
- Belamcanda chinensis*: Beijing Botanical Research Institute, 1976. Iconographia Cormophytorum Sinicorum [Line drawings of Chinese vascular plants]. Vol. 5. Beijing, China. p. 570, fig. 7970 (flowering plant habit, flower); Britton, N. & Brown, A., 1947. An illustrated flora of the

- northern United States, Canada and the British possessions. Vol. 1. The New York Botanical Garden, New York, United States. p. 542, fig. 1342 (dehisced fruit showing the seeds). Redrawn and adapted by Achmad Satiri Nurhaman.
- Bidens pilosa*: Soerjani, M., Kostermans, A.J.G.H. & Tjitrosoepomo, G. (Editors), 1987. Weeds of rice in Indonesia. Balai Pustaka, Jakarta, Indonesia. p. 65, fig. 4.18 (flowering and fruiting plant, flowering head, ray floret, disk floret, achenes).
- Blumea lacera*: Soerjani, M., Kostermans, A.J.G.H. & Tjitrosoepomo, G. (Editors), 1987. Weeds of rice in Indonesia. Balai Pustaka, Jakarta, Indonesia. p. 67, fig. 4.19 (plant habit, capitulum, marginal flower, disk flower, achene with pappus removed). Redrawn and adapted by Iskak Syamsudin.
- Brucea javanica*: Nooteboom, H.P., 1962. Simaroubaceae. In: van Steenis, C.G.G.J. (General editor): Flora Malesiana. Ser. I. Vol. 6. Wolters-Noordhoff Publishing, Groningen, The Netherlands. p. 210, fig. 12 (twig with flowers and fruits, female flower, male flower, fruit). Redrawn and adapted by Iskak Syamsudin.
- Bryophyllum pinnatum*: Liu, T.-S. & Chung, N.-J., 1993. *Bryophyllum* Salisb. In: Huang, T.-C. (Editor in Chief): Flora of Taiwan. 2nd edition. Vol. 3. Editorial Committee of the Flora of Taiwan, Taipei, Taiwan, Republic of China. p. 12, pl. 5 (upper part of stem); Wickens, G.E., 1987. Crasulaceae. In: Polhill, R.M. (Editor): Flora of Tropical East Africa. A.A. Balkema, Rotterdam, Boston. p. 29, fig. 5 (inflorescence, flower in longitudinal section). Redrawn and adapted by Achmad Satiri Nurhaman.
- Cannabis sativa*: Bruneton, J., 1995. Pharmacognosy, phytochemistry, medicinal plants. Technique & Documentation. Lavoisier, Paris, France. p. 372 (branch of female plant); Mansfeld, R., 1986. Verzeichnis landwirtschaftlicher und gärtnerischer Kulturpflanzen (ohne Zierpflanzen) [Register of agricultural and horticultural plants (excluding ornamentals)]. Vol. 1. 2nd edition by Schultze-Motel. J. Springer Verlag, Berlin, Germany. p. 88, 89, fig. 12A, 12B (part of branch with female inflorescence, female flower, part of branch of male plant, male flower); Backer, C.A., 1951. Cannabinaceae. In: van Steenis, C.G.G.J. (General editor): Flora Malesiana. Ser. I. Vol. 4. Noordhoff-Kolff, Jakarta, Indonesia. p. 222, fig. 1 (fruit). Redrawn and adapted by Iskak Syamsudin.
- Cardiospermum halicacabum*: Matthew, K.M., 1988. Further illustrations on the Flora of Tamilnadu Carnatic. The Rapinat Herbarium, St. Joseph's College, Tiruchirappalli, India. p. 107, pl. 107 (male flower with sepals and petals removed, pistillode of male flower, female flower with sepals and petals removed, pistil of female flower); Leenhouts, P.W., 1994. *Cardiospermum*. In: Kalkman, C. et al. (Editors): Flora Malesiana. Series I. Vol. 11(3). Rijksherbarium/Hortus Botanicus, Leiden, the Netherlands. p. 485, fig. 12 (plant habit, fruit, seed). Redrawn and adapted by Achmad Satiri Nurhaman.
- Carmona retusa*: Matthew, K.M., 1988. Further illustrations on the Flora of Tamilnadu Carnatic. The Rapinat Herbarium, St. Joseph's College, Tiruchirappalli, India. p. 383, pl. 383 (flower, 4-merous corolla, 5-merous corolla); Riedl, H., 1997. Boraginaceae. In: Kalkman, C. et al. (Editors): Flora Malesiana. Series I. Vol. 13. Rijksherbarium/Hortus Botanicus, Leiden, The Netherlands. p. 66, fig. 2 (fruiting twig, fruit). Redrawn and adapted by Iskak Syamsudin.
- Cassia fistula*: Brown, W.H., 1954. Useful plants of the Philippines. Reprint of the 1941-1943 edition. Vol. 2. Technical Bulletin No 10. Department of Agriculture and Natural Resources, Bureau of Printing, Manila, The Philippines. p. 103, fig. 42 (section of pod showing seeds); Larsen, K., Larsen, S.S. & Vidal, J.E., 1980. Flore du Cambodge, du Laos et du Vietnam. Vol. 18. Légumineuses-Caesalpinioïdées [Flora of Cambodia, Laos and Vietnam. Vol. 18. Leguminosae-Caesalpinioideae]. Muséum National d'Histoire Naturelle, Paris, France. p. 81, fig. 14 (flower); Verdcourt, B., 1979. A manual of New Guinea legumes. Botany Bulletin No 11. Office of Forests, Division of Botany, Lae, Papua New Guinea. p. 44, fig. 9 (branch with inflorescence, mature pod). Redrawn and adapted by Achmad Satiri Nurhaman.
- Catharanthus roseus*: Lawrence, G.H.M., 1959. *Vinca* and *Catharanthus*. *Baileya* 7(4). p. 117, fig. 38 (flowering twig); Stearn, W.T., 1966. *Catharanthus roseus*, the correct name for the Madagascar periwinkle. *Lloydia* 29(3). p. 197, fig. 1 (flower); Plaizier, A.C., 1981. A revision of *Catharanthus roseus* (L.) G. Don (Apocynaceae). *Mededelingen Landbouwhogeschool Wageningen* 81-9. p. 4, fig. 1 (base and top of corolla tube in longitudinal section, fruit, seed). Redrawn and adapted by Achmad Satiri Nurhaman.
- Centella asiatica*: Backer, C.A. & van Slooten, D.F., 1924. Geïllustreerd handboek der Javaan-

- sche theeonkruiden en hunne beteekenis voor de cultuur [Illustrated handbook of the weeds of Javanese tea plantations and their significance for tea growing]. Ruygrok, Batavia, Indonesia. pl. 185 (plant habit); Townsend, C.C., 1989. Umbelliferae. In: Polhill, R.M. (Editor): Flora of Tropical East Africa. A.A. Balkema, Rotterdam, Brookfield, p. 16, fig. 3 (leaf, stem base with young leaf, flowers and fruits, fruit); Kao, M.-T., 1993. Umbelliferae. In: Huang, T.-C. (Editor in Chief): Flora of Taiwan. 2nd edition. Vol. 3. Editorial Committee of the Flora of Taiwan, Taipei, Taiwan, Republic of China. p. 1018, pl. 507 (inflorescence, flower). Redrawn and adapted by Iskak Syamsudin.
- Chenopodium ambrosioides*: Hitchcock, C.L., Cronquist, A., Ownbey, M. & Thompson, J.W., 1964. Vascular plants of the Pacific Northwest. Part 2. University of Washington Press, Seattle, United States. p. 197 (flowering plant, fruit enclosed by perianth); Ulbrich, E., 1934. Chenopodiaceae. In: Engler, A. & Prantl, K. (Editors): Die natürlichen Pflanzenfamilien [The natural plant families]. Second edition, Band 16C. Verlag von Wilhelm Engelmann, Leipzig, Germany. p. 481, fig. 183 (bisexual flower, female flower, ovary with glands, fruit after removal of perianth). Redrawn and adapted by Achmad Satiri Nurhaman.
- Cinchona officinalis*: Moens, J.C.B., 1882. De kina-cultuur in Azië [The kina culture in Asia]. Ernst & Co., Batavia, Indonesia. tab. 1 (tree habit); Van den Abeele, M. & Vandenput, R., 1951. De voornaamste cultures van Belgisch-Congo [The most important cultures of Belgian Congo]. 2nd edition. Ministerie van Koloniën, Directie voor Landbouw, Veeteelt en Kolonisatie, Brussel, Belgium. p. 445, fig. 235 (flowering twig); Purseglove, J.W., 1968. Tropical crops. Dicotyledons 2. Longmans, London & Harlow, United Kingdom. p. 455, fig. 72 (flowers in longitudinal section showing heterodistyly); Schumann, K., 1891. Rubiaceae. In: Engler, A. & Prantl, K. (Editors): Die natürlichen Pflanzenfamilien. Band 4(4) [The natural plant families. Volume 4(4)]. Verlag von Wilhelm Engelmann, Leipzig, Germany. p. 41, fig. 13C (fruiting twig). Redrawn and adapted by Achmad Satiri Nurhaman.
- Cissampelos pareira*: Troupin, G., 1956. Menispermaceae. In: Turrill, W.B. & Milne-Redhead, E. (Editors): Flora of Tropical East Africa. Crown Agents for Oversea Governments and Administrations, London, United Kingdom. p. 24, fig. 6 (flowering stems of male plant, flowering stem of female plant, male flower, sectioned male flower, part of female inflorescence, endocarp). Redrawn and adapted by Iskak Syamsudin.
- Curculigo orchoides*: Nasir, E., 1980. Amaryllidaceae. In: Nasir, E. & Ali, S.I. (Editors): Flora of Pakistan. No 134. National Herbarium, Pakistan Agricultural Research Council, Islamabad, Pakistan. p. 4, fig. 1 (flowering plant, flower); Bhaskaran, K. & Padmanabhan, D., 1983. Leaf development in *Curculigo orchoides*. Phytomorphology 31. p. 3, fig. 1 (rhizome); Matthew, K.M., 1982. Illustrations on the Flora of Tamilnadu Carnatic. The Rapinat Herbarium, St. Joseph's College, Tiruchirapalli, India. pl. 733 (fruit, seed). Redrawn and adapted by Achmad Satiri Nurhaman.
- Curcuma longa*: Brown, W.H., 1951. Useful plants of the Philippines. Reprint of the 1941-1943 edition. Vol. 1. Technical Bulletin No 10. Department of Agriculture and Natural Resources, Bureau of Printing, Manila, The Philippines. p. 426, fig. 172 (rhizome); Ochse, J.J. & Bakhuizen van den Brink, R.C., 1980. Vegetables of the Dutch East Indies. 3rd English edition (translation of 'Indische groenten', 1931). Asher & Co., Amsterdam, The Netherlands. p. 737, fig. 445 (flowering clump). Redrawn and adapted by P. Verheij-Hayes.
- Curcuma zedoaria*: Flach, M. & Rumawas, F., 1996. Plant Resources of South-East Asia No 9. Plants yielding non-seed carbohydrates. Backhuys Publishers, Leiden, The Netherlands. p. 77 (leafy shoot and inflorescence, rhizome, flower with bract and bracteole, flower in lateral view, flower in front view, stamen, lateral staminodes and pistil).
- Cyclea barbata*: Sastrapradja, S. (Editor), 1978. Tumbuhan obat [Medicinal plants]. Lembaga Biologi Nasional-LIPI, Bogor, Indonesia. p. 16 (leafy stem); Forman, L.L., 1991. Menispermaceae. In: Smitinand, T. & Larsen, K. (Editors): Flora of Thailand. Vol. 5, Part 3. The Forest Herbarium, Royal Forest Department, Bangkok, Thailand. p. 329, fig. 73K-O (stem with leaf and male inflorescence, male flower, male flower with calyx removed, fruit, endocarp). Redrawn and adapted by Iskak Syamsudin.
- Cyperus brevifolius*: Backer, C.A. & van Slooten, D.F., 1924. Geïllustreerd handboek der Javaansche theeonkruiden en hunne beteekenis voor de cultuur [Illustrated handbook of the weeds of Javanese tea plantations and their significance for tea growing]. Ruygrok, Batavia, Indonesia.

- pl. 83 (plant habit); Kern, J.H., 1952. Notes on Malaysian Cyperaceae. *Reinwardtia* 2(1). p. 127, fig. 14 (inflorescence enclosed by involucre bracts, spikelet, nuts). Redrawn and adapted by Achmad Satiri Nurhaman.
- Cyperus rotundus*: Mercado, B.L., 1979. A monograph on *Cyperus rotundus* L. Biotrop Bulletin No 15. Biotrop, SEAMEO Regional Center for Tropical Biology, Bogor, Indonesia. p. 15 & 16, fig. 5 & 6 (young plant with rhizomes and tubers, tubers); Kern, J.H., 1974. Cyperaceae. In: van Steenis, C.G.G.J. (General editor): *Flora Malesiana*. Ser. I. Vol. 7. Noordhoff International Publishing, Leiden, The Netherlands. p. 594, fig. 49 (flowering plant). Redrawn and adapted by Achmad Satiri Nurhaman.
- Datura metel*: Backer, C.A. & van Steenis, C.G.G.J., 1973. Atlas of 220 weeds of sugar-cane fields in Java. Greshoff's Rumphius Fund, Amsterdam, The Netherlands. pl. 567 (flowering stem and young fruit); Mansfeld, R., 1986. *Verzeichnis landwirtschaftlicher und gärtnerischer Kulturpflanzen (ohne Zierpflanzen)* [Register of agricultural and horticultural plants (excluding ornamentals)]. Vol. 3. 2nd edition by Schultze-Motel. J. Springer Verlag, Berlin, Germany. p. 1217, fig. 177 (fruit, seeds). Redrawn and adapted by Iskak Syamsudin.
- Derris elliptica*: Spoon, W. & Toxopeus, H.J., 1950. Derriswortel [Derris root]. In: van Hall, C.J.J. & van de Koppel, C. (Editors): *De landbouw in de Indische archipel* [Agriculture in the East Indian archipelago]. Vol. 3. W. v. Hoeve, The Hague, The Netherlands. p. 596, fig. 10 (root system); Greshoff, M., 1894. *Schetsen van nuttige Indische planten* [Sketches of useful Indonesian plants]. Serie 1. Extra Bulletin van het Koloniaal Museum. J.H. de Bussy, Amsterdam, The Netherlands. p. 99, fig. 25 (leaf, inflorescences and young leaves); Geesink, R., 1984. *Scala Millettiearum*. A survey of the genera of the tribe Millettieae (Legum.-Pap.) with methodological considerations. Leiden Botanical Series 8. E.J. Brill / Leiden University Press, Leiden, The Netherlands. p. 67, pl. V, fig. 39 (pod). Redrawn and adapted by Iskak Syamsudin.
- Desmodium gangeticum*: Backer, C.A., 1938. *Onkruidflora der Javasche suikerrietgronden*. Handboek ten dienste van de suikerriet cultuur en de rietsuiker fabricage op Java. Deel 7(11-12) (atlas) [Weed flora of Javanese sugarcane fields. Handbook for the cultivation of sugarcane and manufacturing of cane-sugar in Java. Vol. 7(11-12) (atlas)]. Vereniging het Proefstation voor de Java-Suikerindustrie. Pasuruan. pl. 328 (plant habit); Matthew, K.M., 1988. Further illustrations on the Flora of Tamilnadu Carnatic. The Rapinat Herbarium, St. Joseph's College, Tiruchirapalli, India. p. 155, pl. 155 (branch with infructescence, flower); Verdcourt, B., 1979. A manual of New Guinea legumes. Botany Bulletin No 11. Office of Forests, Division of Botany, Lae, Papua New Guinea. p. 395, fig. 93 (mature pod). Redrawn and adapted by Achmad Satiri Nurhaman.
- Desmodium styracifolium*: Dy Phon, P., Ohashi, H. & Vidal, J.E., 1994. *Flore du Cambodge du Laos et du Vietnam*. Vol. 27. Légumineuses (Fabaceae) Papilionoidea - Desmodiées. [Flora of Cambodia, Laos and Vietnam. Vol. 27. Leguminosae (Fabaceae) Papilionoideae - Desmodiidae]. Muséum National d'Histoire Naturelle, Paris, France. p. 91, pl. 19 (flowering stem, young inflorescence, flowerbud, young pod, mature pod). Redrawn and adapted by Achmad Satiri Nurhaman.
- Elephantopus scaber*: Backer, C.A. & van Slooten, D.F., 1924. *Geïllustreerd handboek der Javaansche theeonkruiden en hunne beteekenis voor de cultuur* [Illustrated handbook of the weeds of Javanese tea plantations and their significance for tea growing]. Ruygrok, Batavia, Indonesia. pl. 215 (plant habit); Matthew, K.M., 1982. Illustrations on the Flora of Tamilnadu Carnatic. The Rapinat Herbarium, St. Joseph's College, Tiruchirapalli, India. pl. 366 (flower). Redrawn and adapted by Iskak Syamsudin.
- Embelia ribes*: Matthew, K.M., 1988. Further illustrations on the Flora of Tamilnadu Carnatic. The Rapinat Herbarium, St. Joseph's College, Tiruchirapalli, India. p. 340, pl. 340 (flowering twig, infructescence, 4-merous flower, 5-merous flower, petal with stamen). Redrawn and adapted by Iskak Syamsudin.
- Erythroxylum novogranatense*: Schulz, O.E., 1907. Erythroxylaceae. In: Engler, A. (Editor): *Das Pflanzenreich* [The plant kingdom]. IV, 134. Wilhelm Engelmann, Leipzig, Germany. p. 86, fig. 18 (flowering twig, short-styled flower with petals removed, long-styled flower with petals removed, petal showing 3-lobed ligule-like appendage, fresh fruit, dried fruit). Redrawn and adapted by Achmad Satiri Nurhaman.
- Euphorbia heterophylla*: Soerjani, M., Kostermans, A.J.G.H. & Tjitrosopomo, G. (Editors), 1987. *Weeds of rice in Indonesia*. Balai Pustaka, Jakarta, Indonesia. p. 289, fig. 4.129 (plant habit, seed); Backer, C.A., 1939. *Onkruidflora*

- der Javasche suikerrietgronden. Handboek ten dienste van de suikerriet cultuur en de rietsuiker fabricage op Java. Deel 7(13) (atlas) [Weed flora of Javanese sugar-cane fields. Handbook for the cultivation of sugar-cane and manufacturing of cane-sugar in Java. Vol. 7(13) (atlas)]. Vereniging het Proefstation voor de Java-Suikerindustrie, Pasuruan. pl. 396 (leaf); Dressler, R.L., 1962. A synopsis of *Poinsetta* (Euphorbiaceae). *Annals of the Missouri Botanical Garden* 48. p. 331, fig. 1B (cyathium). Redrawn and adapted by Achmad Satiri Nurhaman.
- Euphorbia hirta*: Soerjani, M., Kostermans, A.J.G.H. & Tjitrosoepomo, G. (Editors), 1987. Weeds of rice in Indonesia. Balai Pustaka, Jakarta, Indonesia. p. 285, fig. 4.127 (plant habit, young cyathium, mature cyathium, seed). Redrawn and adapted by Achmad Satiri Nurhaman.
- Eurycoma longifolia*: Greshoff, M., 1894. Schetsen nuttige Indische planten [Sketches of useful Indonesian plants]. Serie 1. Extra Bulletin van het Koloniaal Museum. J.H. De Bussy, Amsterdam, The Netherlands. pl. 23 (tree habit, detached leaf and inflorescence); Engler, A., 1931. Simaroubaceae. In: Engler, A. & Prantl, K. (Editors): Die natürlichen Pflanzenfamilien [The natural plant families]. Second edition, Band 19a. Wilhelm Engelmann, Leipzig. p. 381, fig. 175 (male flower, female flower, fruit). Redrawn and adapted by Achmad Satiri Nurhaman.
- Fatoua villosa*: Koorders, S.H., 1913. Exkursionsflora von Java [Excursion flora of Java]. Vol. 4. Gustav Fischer, Jena, Germany. p. 464, fig. 743 (female flower); Liao, J.-C., 1996. Moraceae. In: Huang, T.-C. (Editor in Chief): Flora of Taiwan. 2nd edition. Vol. 2. Editorial Committee of the Flora of Taiwan, Taipei, Taiwan, Republic of China. p. 144, pl. 70 (flowering stem, male flower, fruits with perianth removed). Redrawn and adapted by Achmad Satiri Nurhaman.
- Ficus benghalensis*: Ghafoor, A., 1985. Moraceae. In: Nasir, E. & Ali, S.I. (Editors): Flora of Pakistan. No 171. National Herbarium (Stewart Collection), Pakistan Agricultural Research Council, Islamabad, Pakistan. p. 25, fig. 4 (fruiting twig, halfed fig, female flower, male flower). Redrawn and adapted by Achmad Satiri Nurhaman.
- Ficus religiosa*: Kunkel, G., 1969. Arboles exóticos. Los arboles cultivados en Gran Canaria I [Exotic trees. The cultivated trees of Gran Canaria]. Ediciones del Excmo. Cabildo Insular de Gran Canaria, Canary Islands. p. 141 (fruiting twig); Matthew, K.M., 1988. Further illustrations on the Flora of Tamilnadu Carnatic. The Rapinat Herbarium, St. Joseph's College, Tiruchirapalli, India. p. 606, pl. 606 (fig in cross section, part of receptacle with flowers, female flower, male flower, infructescence). Redrawn and adapted by Achmad Satiri Nurhaman.
- Gloriosa superba*: Matthew, K.M., 1982. Illustrations on the Flora of Tamilnadu Carnatic. The Rapinat Herbarium, St. Joseph's College, Tiruchirapalli, India. p. 743, pl. 743 (flowering stem); Saralamp, P. et al. (Editors), 1996. Medicinal plants in the Sri Ruckhachati Garden. 2nd edition. Siambooks and Publications Co., Bangkok, Thailand. p. 115 (tubers); Watt, J.M. & Breyer-Brandwijk, M.G., 1962. The medicinal and poisonous plants of southern and eastern Africa. Second edition. E. & S. Livingstone Ltd., Edinburgh & London, United Kingdom. p. 701, fig. 193 (fruit). Redrawn and adapted by Iskak Syamsudin.
- Heliotropium indicum*: Gutierrez, H.G., 1982. An illustrated manual of Philippine materia medica. Vol. 2. Natural Research Council of the Philippines, Tagig, Metro Manila, The Philippines. p. 442 (plant habit); Soerjani, M., Kostermans, A.J.G.H. & Tjitrosoepomo, G., (Editors) 1987. Weeds of rice in Indonesia. Balai Pustaka, Jakarta, Indonesia. p. 115, fig. 4.43 (flower, bilobed fruit, cross-section of fruit). Redrawn and adapted by Iskak Syamsudin.
- Holarrhena pubescens*: de Kruif, A.P.M., 1981. A revision of *Holarrhena* R. Br. (Apocynaceae). Mededelingen Landbouwhogeschool Wageningen 81-2. p. 19, fig. 5 (flowering twig, opened flower, fruit, seed); Saralamp, P. et al. (Editors), 1996. Medicinal plants in the Sri Ruckhachati Garden. 2nd edition. Siambooks and Publications Co., Bangkok, Thailand. p. 120 (tree habit). Redrawn and adapted by Iskak Syamsudin.
- Hydnocarpus alcalae*: Brown, W.H., 1954. Useful plants of the Philippines. Reprint of the 1941-1943 edition. Vol. 2. Technical Bulletin 10. Department of Agriculture and Natural Resources. Bureau of Printing, Manila, The Philippines. p. 505, fig. 237 (leafy twig, male flower, female flower, fruits, fruit in longitudinal section). Redrawn and adapted by Achmad Satiri Nurhaman.
- Hypericum japonicum*: Robson, N.K.B., 1996. *Hypericum*. In: Huang, T.-C. (Editor in Chief): Flora of Taiwan. 2nd edition. Vol. 2. Editorial Committee of the Flora of Taiwan, Taipei, Taiwan,

- Republic of China. p. 706, pl. 332 (plant habit, detail of stem, leaf, dehisced capsule, ovary in cross-section); Matthew, K.M., 1982. Illustrations on the Flora of Tamilnadu Carnatic. The Rapinat Herbarium, St. Joseph's College, Tiruchirapalli, India. p. 54, pl. 54 (flower). Redrawn and adapted by Achmad Satiri Nurhaman.
- Imperata cylindrica*: 't Mannetje, L. & Jones, R.M., 1992. Plant Resources of South-East Asia No 4. Forages. Pudoc Scientific Publishers, Wageningen, The Netherlands. p. 141 (plant habit, ligule, inflorescence, spikelet, caryopsis in front and side view).
- Ixora coccinea*: Gutierrez, H.G., 1982. An illustrated manual of Philippine materia medica. Vol. 2. Natural Research Council of the Philippines, Tagig, Metro Manila, The Philippines. p. 417 (flowering twig); Matthew, K.M., 1988. Further illustrations on the Flora of Tamilnadu Carnatic. The Rapinat Herbarium, St. Joseph's College, Tiruchirapalli, India. p. 302, pl. 302 (flower, flower as seen from above). Redrawn and adapted by Iskak Syamsudin.
- Jasminum sambac*: original drawing by P.H. Yap.
- Jatropha curcas*: Heller, J., 1996. Physic nut. *Jatropha curcas* L. Promoting the conservation and use of underutilized and neglected crops. 1. Institute of Plant Genetics and Crop Plant Research, Gatersleben, Germany and International Plant Genetic Resources Institute, Rome, Italy. p. 11, fig. 2 (flowering stem, fruits, fruit in longitudinal section, seed); Jones, N. & Miller, J.H., 1992. *Jatropha curcas*. A multipurpose species for problematic sites. Land Resources Series No 1. Agriculture Division, Asia Technical Department, The World Bank. Fig. 1 (female flower, opened female flower, male flower, opened male flower). Redrawn and adapted by Achmad Satiri Nurhaman.
- Justicia gendarussa*: Backer, C.A. & van Steenis, C.G.G.J., 1973. Atlas of 220 weeds of sugar-cane fields in Java. Greshoff's Rumphius Fund, Amsterdam, The Netherlands. pl. 640 (flowering twig); Hsieh, C.-F. & Huang, T.-C., 1978. Acanthaceae. In: Li, H.-L. et al. (Editors): Flora of Taiwan. Vol. 4. Epoch Publishing Co., Taipei, Taiwan, Republic of China. p. 633, pl. 1135 (flower); Matthew, K.M., 1988. Further illustrations on the Flora of Tamilnadu Carnatic. The Rapinat Herbarium, St. Joseph's College, Tiruchirapalli, India. pl. 456 (stamens and pistil). Redrawn and adapted by Iskak Syamsudin.
- Kaempferia galanga*: Departemen Kesehatan Republik Indonesia, 1983. Pemanfaatan tanaman obat, edisi III [Usefulness of medicinal plants, 3rd edition]. p. 199 (plant habit); Wight, R., 1844. Icones plantarum Indiae orientalis [Drawings of East Indian plants]. Vol. 3. J.B. Pharoah, Madras, India. t. 899 (flower, apex of fertile stamen, style apex). Redrawn and adapted by Achmad Satiri Nurhaman.
- Kalanchoe crenata*: Wickens, G.E., 1987. Crassulaceae. In: Polhill, R.M. (Editor): Flora of tropical East Africa. A.A. Balkema, Rotterdam, Boston. p. 44, fig. 6. (plant habit, flower, flower in longitudinal section). Redrawn and adapted by Achmad Satiri Nurhaman.
- Lantana camara*: Holm, L.G., Plucknett, D.L., Pancho, J.V. & Herberger, J.P., 1977. The world's worst weeds. Distribution and biology. East-West Center. The University Press of Hawaii, Honolulu, United States. p. 300, fig. 124 (root system, flower with bract, flower in longitudinal section, infructescence, seed); Backer, C.A. & van Slooten, D.F., 1924. Geïllustreerd handboek der Javaansche theekonkruiden en hunne beteekenis voor de cultuur [Illustrated handbook of the weeds of Javanese tea plantations and their significance for tea growing]. Ruygrok, Batavia, Indonesia. pl. 186 (flowering and fruiting twig). Redrawn and adapted by Achmad Satiri Nurhaman.
- Melochia corchorifolia*: Soerjani, M., Kostermans, A.J.G.H. & Tjitrosoepomo, G. (Editors), 1987. Weeds of rice in Indonesia. Balai Pustaka, Jakarta, Indonesia. p. 555, fig. 4.261 (flowering plant, flowers viewed from different angles, fruit, seed). Redrawn and adapted by Iskak Syamsudin.
- Mentha arvensis*: Backer, C.A. & van Steenis, C.G.G.J., 1973. Atlas of 220 weeds of sugar-cane fields in Java. Greshoff's Rumphius Fund, Amsterdam, The Netherlands. pl. 534 (flowering plant); Keng, H., 1978. Labiatae. In: van Steenis, C.G.G.J. (General editor): Flora Malesiana. Ser. I. Vol. 8. Sijthoff & Noordhoff International Publishers, Alphen aan den Rijn, The Netherlands. p. 344, fig. 16 (upper part of flowering stem, flower, nutlet). Redrawn and adapted by Achmad Satiri Nurhaman.
- Mimosa pudica*: 't Mannetje, L. & Jones, R.M., 1992. Plant Resources of South-East Asia No 4. Forages. Pudoc Scientific Publishers, Wageningen, The Netherlands. p. 168 (flowering and fruiting stem, leaflet, flower, pod, one-seeded pod segment, seed).
- Momordica charantia*: Siemonsma, J.S. & Kasem

- Piluek (Editors), 1993. Plant Resources of South-East Asia No 8. Vegetables. Pudoc Scientific Publishers, Wageningen, The Netherlands. p. 208 (leafy shoot, male flower in longitudinal section, female flower in longitudinal section, fruit); Wight, R., 1840-1843. *Icones plantarum Indiae orientalis* [Drawings of East Indian plants]. Vol. 2(2). J.B. Pharoah, Madras, India. t. 504 (male flower in cross section with petals removed). Redrawn and adapted by P. Verheij-Hayes.
- Morus alba*: Ghafoor, A., 1985. Moraceae. In: Nasir, E. & Ali, S.I. (Editors): Flora of Pakistan. No. 171. Shamim Printing Press, Karachi, Pakistan. p. 51, fig. 10 (fruiting twig, infructescence, female flower); Townsend, C.C., 1980. 27. Moraceae. In: Townsend, C.C. & Guest, E. (Editors): Flora of Iraq 4(1). The Ministry of Agriculture, Baghdad, Iraq. p. 83, pl. 16 (male inflorescence, male flower). Redrawn and adapted by Achmad Satiri Nurhaman.
- Oldenlandia corymbosa*: Sivarajan, V.V. & Biju, S.D., 1990. Taxonomic and nomenclatural notes on the Hedyotis corymbosa-diffusa complex (Rubiaceae) in India. *Taxon* 39(4). p. 668, pl. 1 (plant habit, flower, infructescence); Halford, D.A., 1992. Review of the genus *Oldenlandia* L. (Rubiaceae) and related genera in Australia. *Austrobaileya* 3(4). p. 691, fig. 2 (flowering and fruiting stem). Redrawn and adapted by Iskak Syamsudin.
- Orthosiphon aristatus*: Keng, H., 1978. Labiatae. In: van Steenis, C.G.G.J. (General editor): Flora Malesiana. Ser. I. Vol. 8. Sijthoff & Noordhoff International Publishers, Alphen aan den Rijn, The Netherlands. p. 381, fig. 31 (flowering stem); Keng, H., 1969. Flora Malesianae Precursores XLVIII. A revision of Malesian Labiatae. The Gardens' Bulletin, Singapore 24. p. 133, fig. 25 (flower, fruiting calyx and nutlet). Redrawn and adapted by Achmad Satiri Nurhaman.
- Oxalis corniculata*: Backer, C.A., 1938. Onkruidflora der Javasche suikerrietgronden. Handboek ten dienste van de suikerriet cultuur en de rietsuiker fabricage op Java. Deel 7(11-12) (atlas) [Weed flora of Javanese sugar-cane fields. Handbook for the cultivation of sugar-cane and manufacturing of cane-sugar in Java. Vol. 7(11-12) (atlas)]. Vereniging het Proefstation voor de Java-Suikerindustrie, Pasuruan, Indonesia. pl. 371 (creeping stem with flowers and fruits); Huang, T.-C. & Liu, T.-S., 1977. Labiatae. In: Li, H.-L. et al. (Editors): Flora of Taiwan. Vol. 4. Epoch Publishing Co., Taipei, Taiwan, Republic of China. p. 425, pl. 670 (part of stem with flowers and fruits, flower in longitudinal section, fruit with seeds); Holm, L.G., Plucknett, D.L., Pancho, J.V. & Herberger, J.P., 1977. The world's worst weeds. Distribution and biology. East-West Center. The University Press of Hawaii, Honolulu, United States. p. 143, fig. 344 (seed). Redrawn and adapted by Iskak Syamsudin.
- Papaver somniferum*: Hess, H.E., Landolt, E. & Hirzel, R., 1970. Flora der Schweiz un angrenzender Gebiete [Flora of Switzerland and neighbouring regions]. Band 2. Birkhäuser Verlag, Basel & Stuttgart, Switzerland & Germany. p. 111 (flowering stem, stem base, fruit); Jávorka, S. & Csapody, V., 1975. *Iconographia florum partis austro-orientalis europae centralis* [Drawings of flowers from the south-eastern part of central Europe]. Akadémiai Kiadó, Budapest, Hungary. p. 179, drawing 1391 (flower, stamen, seed). Redrawn and adapted by Achmad Satiri Nurhaman.
- Peperomia pellucida*: Gutierrez, H.G., 1982. An illustrated manual of Philippine materia medica. Vol. 2. Natural Research Council of the Philippines, Manila. p. 452 (flowering plant, part of infructescence). Redrawn and adapted by Achmad Satiri Nurhaman.
- Phyllanthus amarus*: Berhaut, J., 1975. Flore illustrée du Sénégal [Illustrated flora of Senegal]. Ministère du Développement Rural et de l'Hydraulique, Direction des Eaux et Forêts, Dakar, Senegal. p. 552 (plant habit); Brunel, J.F. & Roux, J., 1984. South-East Asian Phyllanthaceae II. Some Phyllanthus of subsect. Swartziani. *Nordic Journal of Botany* 4(4). p. 70, fig. 1-6 (cataphyll and stipules, leaves and inflorescences, fruit, seed); Webster, G.L., 1957. A monographic study of the West Indian species of Phyllanthus. *Journal of the Arnold Arboretum* 38(4). p. 305, pl. 19 (male flower, female flower). Redrawn and adapted by Achmad Satiri Nurhaman.
- Phyllanthus emblica*: Lemmens, R.H.M.J. & Wuljarni-Soetjipto, N. (Editors), 1991. Plant Resources of South-East Asia No 3. Dye and tannin-producing plants. Pudoc, Wageningen, The Netherlands. p. 107 (flowering twig, male flower, female flower, fruiting twig).
- Phytolacca acinosa*: Liu, T.-S., 1976. Phytolaccaceae. In: Li, H.-L. et al. (Editors): Flora of Taiwan. Vol. 3. Epoch Publishing Co., Taipei, Taiwan, Republic of China. p. 294, pl. 300 (flowering and fruiting stem, mature fruit); Mansfeld,



- R., 1986. Verzeichnis landwirtschaftlicher und gärtnerischer Kulturpflanzen (ohne Zierpflanzen) [Register of agricultural and horticultural plants (excluding ornamentals)]. Vol. 1. 2nd edition by Schultze-Motel. J. Springer Verlag, Berlin, Germany. p. 125, fig. 23 (flower, young fruit). Redrawn and adapted by Iskak Syamsudin.
- Plantago major*: Holm, L.G., Plucknett, D.L., Panch, J.V. & Herberger, J.P., 1977. The world's worst weeds. Distribution and biology. East-West Center. The University Press of Hawaii, Honolulu, United States. p. 387, fig. 162 (fruiting plant, flower, dehisced fruit); Ross-Craig, S., 1968. Drawings of British plants, part XXV. G. Bell and Sons, London, United Kingdom. pl. 5 (inflorescence, seeds). Redrawn and adapted by Achmad Satiri Nurhaman.
- Plectranthus scutellarioides*: Huang, T.-C. & Liu, T.-S., 1977. Labiatae. In: Li, H.-L. et al. (Editors): Flora of Taiwan. Vol. 4. Epoch Publishing Co., Taipei, Taiwan, Republic of China. p. 459, pl. 1069 (flowering stem, leaf, flower, opened calyx, nutlets). Redrawn and adapted by Achmad Satiri Nurhaman.
- Plumbago indica*: Backer, C.A., 1941. Onkruidflora der Javasche suikerrietgronden. Handboek ten dienste van de suikerriet cultuur en de riet-suiker fabricage op Java. Deel 7(15) (atlas) [Weed flora of Javanese sugar-cane fields. Handbook for the cultivation of sugar-cane and manufacturing of cane-sugar in Java. Vol. 7(15) (atlas)]. Vereniging het Proefstation voor de Java-Suikerindustrie, Pasuruan, Indonesia. pl. 453 (plant habit); Saralamp, P. et al. (Editors), 1996. Medicinal plants in the Sri Ruckhachati Garden. 2nd edition. Siambooks and Publications Co., Bangkok, Thailand. p. 169 (roots); Baillon, H., 1892. Histoire des plantes [History of plants]. Vol. 11. Hachette & Cie, London, United Kingdom. p. 355, fig. 402 (flower). Redrawn and adapted by Iskak Syamsudin.
- Premna herbacea*: Munir, A.A., 1984. A taxonomic revision of the genus *Premna* L. (Verbenaceae) in Australia. Journal of the Adelaide Botanic Gardens 7(1). p. 9, fig. 1 (plant habit, flower, opened flower, fruit). Redrawn and adapted by Iskak Syamsudin.
- Premna serratifolia*: Koorders, S.H. & Valetton, Th., 1914. Atlas der Baumarten von Java [Atlas of tree species from Java]. Vol. 4. P.W.M. Trap, Leiden, The Netherlands. fig. 291 (tree habit); Munir, A.A., 1984. A taxonomic revision of the genus *Premna* L. (Verbenaceae) in Australia. Journal of the Adelaide Botanic Gardens 7(1). p. 16, fig. 2 (flowering twig, flower, fruit). Redrawn and adapted by Iskak Syamsudin.
- Pueraria montana*: Huang, T.-C. & Ohashi, H., 1977. Leguminosae. In: Li, H.-L. et al. (Editors): Flora of Taiwan. Vol. 3. Epoch Publishing Co., Taipei, Taiwan, Republic of China. p. 368, pl. 636 (part of flowering stem, flower, part of infructescence, seed). Redrawn and adapted by Achmad Satiri Nurhaman.
- Quisqualis indica*: Coode, M.J.E., 1978. Combretaceae. In: Womersley, J.S. (Editor): Handbooks of the flora of Papua New Guinea. Vol. 1. Melbourne University Press, Carlton, Australia. p. 52, fig. 16 (flowering twig); Townsend, C.C., 1980. 53. Combretaceae. In: Townsend, C.C. & Guest, E. (Editors): Flora of Iraq. Vol. 4(1). Ministry of Agriculture & Agrarian Reform, Baghdad, Iraq. p. 408, pl. 71 (opened flower); Lecompte, O., 1969. Combretaceae. In: Tardieu-Blot, M.L. (Editor): Flore du Cambodge du Laos et du Viêt-nam [Flora of Cambodia, Laos and Vietnam]. Fasc. 10. Muséum National d'Histoire Naturelle, Paris, France. p. 23, pl. IV (fruit). Redrawn and adapted by Iskak Syamsudin.
- Rauwolfia serpentina*: Perrot, E. & Paris, R., 1971. Les plantes médicinales [The medicinal plants]. Vol. 2. Presses Universitaires de France. plate opposite p. 198 (plant habit, piece of root, flower, opened corolla, fruit, stone). Redrawn and adapted by Iskak Syamsudin.
- Rhinacanthus nasutus*: Greshoff, M., 1894. Schetsen nuttige indische planten [Sketches of useful Indonesian plants]. Series 1 (1-50). Extra Bulletin van het Koloniaal Museum. J.H. de Bussy, Amsterdam, the Netherlands. pl. 40 (flowering twig); Saralamp, P. et al. (Editors), 1996. Medicinal plants in the Sri Ruckhachati Garden. 2nd edition. Siambooks and Publications Co., Bangkok, Thailand. p. 178. (upper part of flower); Wight, R., 1841. Icones plantarum Indiae Orientalis [Drawings of East Indian plants]. Vol. 2. J.B. Pharoah, Madras, India. pl. 464 (fruit). Redrawn and adapted by Achmad Satiri Nurhaman.
- Schefflera elliptica*: Koorders, S.H. & Valetton, Th., 1916. Atlas der Baumarten von Java [Atlas of tree species from Java]. Vol. 4. P.W.M. Trap, Leiden, The Netherlands. fig. 688 (flowering twig, more narrow leaflet, flower, fruit). Redrawn and adapted by Iskak Syamsudin.
- Schefflera heptaphylla*: Doan Thi Nhu, Nguyen Thuong Thuc, Do Huy Bich & Vu Thuy Huyen

- (Editors), 1990. Les plantes médicinales au Vietnam (livre 1) [The medicinal plants of Vietnam (volume 1)]. Médecine traditionnelle et pharmacopée. Agence de coopération Culturelle et Technique, Paris, France. p. 26 (flowering twig); Ohashi, H., 1993. 106. Araliaceae. In: Huang, T.-C. (Editor): Flora of Taiwan. 2nd edition. Vol. 3. Editorial Committee of the Flora of Taiwan, Taipei, Taiwan, Republic of China. p. 1004, pl. 500 (part of infructescence, flower, flower in longitudinal section, fruit). Redrawn and adapted by Iskak Syamsudin.
- Scutellaria indica*: Wu, C.-Y. & Li, H.-W., 1977. Flora Reipublicae Popularis Sinicae [Flora of the Peoples Republic of China] 65(2). Institutum Botanicum Provinciae Yunnanicae & Institutum Botanicum Pekinense Academiae Sinicae, China. p. 173, pl. 38 (flowering stem, opened corolla); Huang, T.-C. & Cheng, W.-T., 1978. Labiatae. In: Li, H. et al. (Editors): Flora of Taiwan, Vol. 4. Epoch Publishing Co., Taipei, Taiwan. p. 522, pl. 1091 (flower, calyx with scutellum, nutlets). Redrawn and adapted by Achmad Satiri Nurhaman.
- Senna tora*: Busson, F., 1965. Plantes alimentaires de l'Ouest Africain. [Food plants of West Africa]. Le Ministère de la Coopération et al., Paris, France. p. 261, fig. 101 (twig with flowers and fruits, detail of gland on leaf rachis, detail of stipules, detail of stamens and pistil); Holm, J., Doll, J. & Holm, E., 1997. World weeds: natural histories and distribution. Wiley, New York, United States. p. 160, fig. 19-2 (root system). Redrawn and adapted by Achmad Satiri Nurhaman.
- Smilax china*: Beijing Botanical Research Institute, 1976. Iconographia Cormophytorum Sini-corum [Line drawings of Chinese vascular plants]. Vol. 5. Beijing, China. p. 535, fig. 7900 (rhizome, part of flowering stem, male flower). Redrawn and adapted by Iskak Syamsudin.
- Smilax glabra*: Beijing Botanical Research Institute, 1976. Iconographia Cormophytorum Sini-corum [Line drawings of Chinese vascular plants]. Vol. 5. Beijing, China. p. 539, fig. 7908 (rhizome); Koyama, T., 1975. Smilacaceae. In: Smitinand, T. & Larsen, K. (Editors): Flora of Thailand. Vol. 2. The Forest Herbarium, Royal Forest Department, Bangkok, Thailand. p. 241, fig. 34 (stem with male inflorescences, stem with infructescences, female flower, male flower). Redrawn and adapted by Iskak Syamsudin.
- Solanum erianthum*: Symon, D.E., 1985. The Solanaceae of New Guinea. Journal of the Adelaide Botanic Gardens 8. p. 82, fig. 32 (flowering stem, flower, infructescence). Redrawn and adapted by Achmad Satiri Nurhaman.
- Solanum nigrum*: Ross-Craig, S., 1965. Drawings of British plants, part XXI, Boraginaceae, Convolvulaceae, Solanaceae. G. Bell and Sons, London, United Kingdom. pl. 29 (flowering stem, part of fruiting stem, opened calyx and gynoe-cium, petal and stamens, stamen, seed from below, seed in side view). Redrawn and adapted by Iskak Syamsudin.
- Sophora tomentosa*: Verdcourt, B., 1979. A manual of New Guinea legumes. Botany Bulletin No 11. Office of Forests, Division of Botany, Lae, Papua New Guinea. p. 290, fig. 64 (flowering twig, flower, flower with petals removed, seed); Nguyen Van Thuan, Dy Phom, P. & Niyomdham, C., 1987. Flore du Cambodge, du Laos et du Vietnam [Flora of Cambodia, Laos and Vietnam]. Vol. 23. Leguminosae (Fabaceae) Papilionoideae. Muséum National d'Histoire Naturelle, Paris, France. p. 17, pl. 1 (infructescence). Redrawn and adapted by Iskak Syamsudin.
- Stephania japonica*: Backer, C.A. & van Slooten, D.F., 1924. Geïllustreerd handboek der Javaansche theonkruiden en hunne beteekenis voor de cultuur [Illustrated handbook of the weeds of Javanese tea plantations and their significance for tea growing]. Ruygrok, Batavia, Indonesia. pl. 113 (flowering twig); Forman, L.L., 1986. Menispermaceae. In: van Steenis, C.G.G.J. & de Wilde, W.J.J.O. (General editors): Flora Malesiana. Ser. I. Vol. 10. Kluwer Academic Publishers, Dordrecht, the Netherlands. p. 246, fig. 19 (part of stem with leaf and inflorescence, male flower, female flower, fruit, endocarp). Redrawn and adapted by Achmad Satiri Nurhaman.
- Strychnos ignatii*: Leenhouts, P.W., 1962. Loganiaceae. In: van Steenis, C.G.G.J. (General editor): Flora Malesiana. Ser. I. Vol. 6. Wolters-Noordhoff Publishing, Groningen, The Netherlands. p. 348, fig. 28 (flowering twig, flower, twig with fruits, lenticular-shaped seed, irregularly shaped seeds). Redrawn and adapted by Iskak Syamsudin.
- Styphnolobium japonicum*: Hegi, G., 1924. Illustrierte Flora von Mittel-Europa. Band IV, Teil 3 [Illustrated flora of Central Europe. Vol. 4, Part 3]. J.F. Lehmanns, München, Germany. p. 1145, fig. 1302 (flowering twig, flower, stamens and ovary); Nguyen Van Thuan, Dy Phom, P. & Niyomdham, C., 1987. Flore du Cambodge, du Laos

- et du Vietnam [Flora of Cambodia, Laos and Vietnam]. Vol. 23. Leguminosae (Fabaceae) Papilionoideae. Muséum National d'Histoire Naturelle, Paris, France. p. 17, pl. 1 (pod). Redrawn and adapted by Achmad Satiri Nurhaman.
- Taraxacum officinale*: Ross-Craig, S., 1963. Drawings of British plants, part XVIII. G. Bell and Sons, London, United Kingdom. pl. 27 (plant habit, inner involucre bract, flower, fruit, upper part of achene). Redrawn and adapted by Achmad Satiri Nurhaman.
- Tinospora crispa*: Santos, J.K., 1928. Stem and leaf structure of *Tinospora rumphii* Boerlage and *Tinospora reticulata* Miers. The Philippine Journal of Science 35(2): 187-208. pl. 1 (part of stem as found in the market, part of stem with leafy shoot); Forman, L.L., 1991. Menispermaceae. In: Smitinand, T. & Larsen, K. (Editors): Flora of Thailand. Vol. 5, Part 3. The Forest Herbarium, Royal Forest Department, Bangkok, Thailand. pl. 24(30) (part of stem with male inflorescence); Brown, W.H., 1954. Useful plants of the Philippines. Reprint of the 1941-1943 edition. Vol. 1. Technical Bulletin 10. Department of Agriculture and Natural Resources. Bureau of Printing, Manila, The Philippines. p. 534, fig. 22 (male flower, fruit); Forman, L.L., 1986. Menispermaceae. In: van Steenis, C.G.G.J. & de Wilde, W.J.J.O. (General editors): Flora Malesiana. Ser. I. Vol. 10. Kluwer Academic Publishers, Dordrecht, the Netherlands. p. 196, fig. 7 (endocarp in dorsal and ventral view). Redrawn and adapted by Iskak Syamsudin.
- Trichosanthes cucumerina*: Siemonsma, J.S. & Kasem Piluek (Editors), 1993. Plant Resources of South-East Asia No 8. Vegetables. Pudoc Scientific Publishers, Wageningen, The Netherlands. p. 272 (flowering shoot, top of female flower in longitudinal section, fruit of cultivated plant, seed).
- Trichosanthes kirilowii*: Beijing Botanical Research Institute, 1975. Iconographia Cormophyta Sinicorum [Line drawings of Chinese Vascular plants]. Vol. 4. Beijing, China. p. 367 (main root, twig with male inflorescence, female flower, ovary and styles, fruit, seed). Redrawn and adapted by Iskak Syamsudin.
- Verbena officinalis*: Ross-Craig, S., 1966. Drawings of British plants, part XXIII. G. Bell and Sons, London, United Kingdom. pl. 38 (plant habit, part of stem, leaf, inflorescence, back of flower with bracteole, corolla opened out, fruit with part of calyx removed). Redrawn and adapted by Iskak Syamsudin.
- Vernonia cinerea*: Soerjani, M., Kostermans, A.J.G.H. & Tjitrosoepomo, G. (Editors), 1987. Weeds of rice in Indonesia. Balai Pustaka, Jakarta, Indonesia. p. 111, fig. 4.41 (flowering plant, flower head, flower, achene without inner pappus row). Redrawn and adapted by Achmad Satiri Nurhaman.
- Vitex negundo*: Hsiao, J.-Y., 1978. Verbenaceae. In: Li, H. et al. (Editors): Flora of Taiwan, Vol. 4. Epoch Publishing Co., Taipei, Taiwan. p. 433, pl. 1060 (flowering twig, flower, opened corolla, calyx and style); Koorders, S.H. & Valetton, Th., 1914. Atlas der Baumarten von Java [Atlas of tree species from Java]. Vol. 2. P.W.M. Trap, Leiden, The Netherlands. fig. 293 (fruit). Redrawn and adapted by Achmad Satiri Nurhaman.

## Index of compounds

- abrin (N-methyl-L-tryptophan) 73, 74  
abruquinones A, B, D and F 74  
abrus agglutinin 73  
abrusogenin 74  
abrusosides A, B, C and D 74  
acetylenes, antimalarial/antimicrobial 151  
3-O-acetylhellebrigenin 336  
achimillic acid A, B and C 79  
acteoside (verbascoside) 398  
ageratochromene (precocene II) 89  
ajmalicine (raubasine) 186, 187  
ajmaline 425  
ajoenes 95  
allicin (diallyldisulphide-mono-S-oxide) 94  
alliin (S-allyl-L-(+)-cysteine sulphoxide) 94  
aloctin A and B 101  
aloe-emodin 443  
aloin 101  
americanol 393  
(S)- $\alpha$ -amino-3,5-dioxo-1,2,4-oxodiazolidine-2-propanoic acid (L-quisqualic acid) 421  
andrographiside 121  
andrographolide, and related diterpenes 120  
9,10-anthraquinones 442  
 $\alpha$ ,  $\beta$  and  $\gamma$ -antiarin 127  
antiviral protein 393  
antofine 278  
apigenin 78  
apigenin-7,4'-di-o-methyl-ether 121  
apiol 379  
ar-turmerone 212  
arabinoxylans 397  
arcapillin 142  
ardisianone A 148  
aristolic acid 135  
aristolochic acids 133  
artemisinin, and related compounds 140  
 $\alpha$ -asarone (trans-isoasarone) 82  
 $\beta$ -asarone (cis-isoasarone) 82  
ascarirole 78, 79, 195  
ascorbic acid 383  
Asiatic acid 191, 434  
Asiaticoside 191, 434  
(-)-asimilobine 464  
 $\beta$ -atlantone 212  
aucubin 398  
aurantio-obtusin 443  
baicalein (5,6,7-trihydroxyflavone) 439  
baicalin (5,6-dihydroxyflavone-7-O- $\beta$ D-glucuronide) 439  
barakol 443  
barbaloin 101  
barbatusin 405  
barbatusol 404  
belamcandaquinones A and B 148  
belamcandol A and B 148  
benzyl benzoate 332  
berberine 129, 480  
bergapten 278  
bersaldegenins 164  
biobollein 322  
bis-andrographolides (A, B, and C) 121  
bisdesmethoxy-curcumin 211  
blumeatin (5,3',5'-trihydroxy-7-methoxy-dihydroflavone) 156  
(-)-borneol 156  
boujotinolone A 272  
brahminoside 191  
brahmoside 191  
bruceajavanins A and B 161  
bruceanic acids 161  
bruceanols 161  
bruceantin 161  
bruceantanol 161  
bruceantinosides 161  
bruceines A, B and C 161  
bruceolides 161  
bruceosides A, B and C 161  
brucine 468  
brusatol 161  
bryonolic acid 485  
bryophyllins A and B 164  
bryotoxins A, B and C 164  
calystegin B2 and C1 360  
calystegins 360  
(-)-camphor 156  
cannabinoids 169  
canthin-6-one alkaloids 161

- cardiospermin 176  
carpesterol 455  
carvacrol 403  
cassiaside 443  
casticin 498  
(+)-catechin 322  
cepharanthine 464  
chacoine 454  
chamazulene 78  
chaulmoogric acid 300  
choline 124, 264  
R,S-chondocurine 220  
chryso-obtusin 443  
chrysophanol 443  
chrysoplenol D 498  
cinchonidine 200  
cinchonine 200  
cinchophyllamine 200  
1,8-cineol (eucalyptol) 113  
cinerin I and II 494  
cocaine (methylbenzoyl ecgonine) 259  
codeine 374  
colchicine 289  
columestrol 419  
commersonine 454  
conessine 296  
p-coumaric acid 135  
coumarin 328  
crotepoxide 332  
curcacycline A 321  
curcain 321  
curcin 321  
curculigosaponins A-J 208  
curculigoside 208  
 $\alpha$ -curcumene 211, 212  
curcumin 210  
curcumin I 210  
curcumin II and III 211  
curcuminoids 210  
(-)-curine 220  
(-)-cycleanine 464  
L-cycleanine 206  
(-)-cycleapeltine 219  
cylindol A 308  
cylindrene 308  
 $\alpha$ -cyperone 223  
(-)-cytisine 461  
daidzein 418  
daidzein-4',7-diglucoside 418  
daidzin 418  
deguelin 235  
5,6-dehydroeurycomalactone 272  
11-dehydroklaineanonone 272  
dehydrosoyasaponin I 243  
demissidine 454  
demissine 454  
deoxyelephantopin 250  
12-deoxy-16-hydroxyphorbol 321  
deoxynojirimycins 360  
deserpidine 425  
desmethoxy-curcumin 211  
1,4-dideoxy-1,4-imino-D-arabinitol 360  
dihydrobruceajavanin A 161  
dihydroelephantopin 250  
13 $\beta$ ,18-dihydroeurycomanol 272  
dihydroniloticin 272  
14,15 $\beta$ -dihydroxyklaineanonone 272  
3'- $\gamma,\gamma$ -dimethylallylwighteone 236  
8- $\gamma,\gamma$ -dimethylallylwighteone 236  
dimethyl-thiosulphinate 95  
trans-1,7-diphenyl-1,3-hepten-5-ol 211  
trans,trans-1,7-diphenyl-1,3-heptadien-5-ol 211  
trans,trans-1,7-diphenyl-1,3-heptadien-4-one  
(alnustone) 211  
1E,3E,1,7-diphenylheptadien-5-one 211  
diphenylthiosulphinate 95  
dioscin 448  
diphyllin 327  
diphyllin apioside 327  
diphyllin apioside-5-acetate 327  
ehretianone 179  
elephantopin 250  
elliptone 235  
embelin 254  
emblicanin A and B 383  
emodin 443  
3-epibryonolol 486  
(-)-epicatechin 322  
3-epikarounidiol 485  
3-episapelin A 272  
esculentoside 393  
ethyl cinnamate 332  
ethyl-p-methoxy-trans-cinnamate 332  
ethyl-p-methoxycinnamate 332  
eupatolitin 142  
eurycomalactone 272  
eurycomanol 272  
eurycomanol-2-O- $\beta$ -glucopyranoside 272  
eurycomanone 272, 273  
fagomine 360  
falcarindiol 124  
falcarinol 124  
falcarinolone 124  
fangchinoline 220  
ferulic acid (4-hydroxy-3-methoxycinnamic acid)  
311  
ficin 278  
ficuseptine 278

- formononetin 418  
forskolin 404  
furanodienone 212  
furanogermerone 212  
gangetin 243  
genistein 418, 419  
genistin 418, 419  
(4S,5S)-(+)-germacrone-4,5-epoxide 212  
gracillin 448  
graminone B 308  
grandirubrine 206  
hayatinine, and related alkaloids 206  
hellicoside 398  
hispidone 272  
(+)-homoaromoline 131, 219, 220  
hordenine 243  
hydnocarpic acid 300  
hydnocarpin 300  
hydnowightin 300  
p-hydroxybenzoic acid 498  
9-hydroxy-canthin-6-one 272  
9-hydroxy-canthin-6-one-N-oxide 272  
6-hydroxy-5,6-dehydroeurycomalactone 272  
6 $\alpha$ -hydroxyeurycomalactone 272  
3-hydroxy-4-methoxycinnamic acid 311  
10-hydroxyoleuropein 316  
hydroxyphorbols (esters) 322  
3'-hydroxy-5,6,7,4'-tetramethoxyflavone 368  
(-)-hyoscyamine 231  
hypaphorine 74  
hypericin 303  
hypophyllanthin 382  
imperanene 308  
indicine 293  
indicine-N-oxide 293  
insularine 464  
inulin 476  
isoamericanol 393  
isomerubrine 206  
isoorientin 498  
isorhamnetin-(2-(3-methoxy-4-hydroxyphenyl)-  
3,5,7-trihydroxy-4H-1-benzopyran-4-one  
473  
isotetrandrine 220  
japonicine A 304  
jasmolactones 316  
jasmolin I and II 494  
jatrophone, and related compounds 321  
justicidin A and B 327  
kaempferol-3-sophoroside 443  
kalambrsides A-C 336  
karasurin-A 485  
karasurins 484  
karounidiol 486  
kirilowins 485  
kwakhurin 419  
L-quisqualic acid ( $\beta$ -(3,5-dioxo-1,2,4-oxodiazol-  
lidin-2-yl)-L-alanine) 421  
labaditin 322  
lanceotoxin A and B 336  
lantadenes A and C 340  
leptidine 454  
lettucenin A 477  
leucocyanidin-3-O- $\beta$ -galactosyl-cellobioside 278  
leucopelargonidin-3-O- $\alpha$ -L-rhamnoside 278  
leurosidine 187  
leurosine 187  
(-)-limacine 219  
longilactone 272  
luteolin 498  
luteolin-7-O-glucoside 78  
M. charantia trypsin inhibitor-II 354  
madecassic acid 191  
magnoflorine 135  
MAP30 354  
(+)-matrine 461  
(+)-matrine-N-oxide 461  
melianone 272  
(-)-(R)-melochinine 343  
(-)-menthol ((1R,3R,4S)-menthol) 345  
9-methoxy-canthin-6-one 272  
9-methoxy-canthin-6-one-N-oxide 272  
7-methoxy- $\beta$ -carboline-1-propionic acid 272  
p-methoxycinnamic acid 332  
7-methoxy-2,2-dimethylchromene (precocene I)  
89  
5-methoxy-psoralen (5-MOP or bergapten) 275  
7-methoxyvasicine 328  
methyl-allyl-trisulphide 95  
(-)-N-methylcytisine 461  
microphyllone 179  
mimosine (N-(3-allyl)-3-hydroxy-4-pyridone)  
350  
mirificin 419  
mirificoumestan 419  
mirificoumestan hydrate 419  
miroestrol 419  
molephantin 250  
molephantinin 250  
momorcharins 353  
momorcochin-S 354  
momordicines I and II 355  
momordin 354  
morin 360  
morphine 374  
morusinol 360  
multifidol 322  
multifidol glucoside 322

- multifloroside 316  
N-methyl-L-tryptophan (abrine) 74  
N-p-coumaroyltyramine 95  
N-trans-feruloyltyramine 95  
neoandrographolide 121  
neohydnocarpin 300  
niloticin 272  
noscapine 374  
oleanolic-acid derivatives (saponins) 393  
oxalic acid (salts of) 371  
7-oxo-10 $\alpha$ -cucurbitadienol 486  
7-oxodihydrokarounidiol 486  
7-oxoisomultiflorenol 486  
oxymatrine 461  
P. americana antiviral protein 393  
palmatine 131, 480  
papaverine 374  
pareirubrine A and B 206  
pareitropone 206  
pasakbumin A and B 273  
pedunculagin 383  
3,3',4',5,7-pentahydroxyflavone-3-rutinoside 473  
10,12-peroxycalamenene 223  
phantomolin 250  
phorbols (diterpene esters) 264  
phyllanthin 382  
phyllembin 383  
physcion 443  
phytolaccoside 393  
piscidinol 272  
plantamajoside 398  
plumbagin (2-methyl-5-hydroxy-1,4-naphthoquinone) 409  
pokeweed mitogen 393  
polysaccharides, immunoactive 124  
polysaccharides, anti-tumour 124  
precocene I and II 90  
premnazole 413  
pseudohypericin 303  
psoralen 278  
puerarin 418  
punigluconin 383  
putranjivain A 383  
pyrethrin I and II 494  
quercetagenin-6,7,3',4'-tetramethylether 141  
quercetin 383  
quercetin-3-rutinoside 473  
quercitrin 264  
quinidine 200  
quinine 200  
raubasine (ajmalicine) 425  
rescinamine 425  
reserpine 425  
retronecine 293  
rhein 181, 443  
rhinacanthins 431, 432  
rhinacanthone 432  
robustic acid 236  
rosmarinic acid 179, 405  
rotenone 235  
rotundinal 498  
rubrofusarin-gentobioside 443  
rutin 473  
rutoside 473  
S. japonicum agglutinin (SJA) 473  
saroaspidin A, B and C 304  
sarrothralen A, B, C and D 304  
sarrothralin 304  
schaftoside 78  
scoparone (6,7-dimethoxycoumarin) 142  
(-)-scopolamine 231  
(-)-scopolamine (= (-)-hyoscyne) 230  
scutellarin 439  
sennidins 181  
sennosides 181  
shikimic acid 264  
simplexoside 327  
sinensetin 368, 369  
 $\beta$ -sitosterol-D-glucoside 278  
sobatum 455  
solanidine 454  
solanine 454  
solasodine 454  
sophorine 461  
soyasaponin 243  
spegatrine 426  
(-)-stepholidine 464  
strychnine 468  
swertisin 78  
TAP-29 484  
 $\delta^9$ -tetrahydrocannabinol 169  
(-)-tetrahydropalmatine 464  
tetramethylscutellarein 369  
(+)-tetrandrine 219  
S,S-tetrandrine 220, 464  
5,7,2',3'-tetrahydroxyflavone 439  
2(S)-5,7,2',6'-tetrahydroxyflavanone 439  
(+)-thalarugosine 219  
(Z)-thiopropional-S-oxide 94  
TK-35 485  
tomatidenol 454  
tomatidine 454  
tomatine 454  
tomenphantopin-A and -B 250  
toxicarol 235  
trachelanthamidine 293  
triacontanol 382  
trichokirin 485

trichosans A, B, C, D and E 486  
trichosanthins 484, 485  
 $\beta$ -trichosanthin 485  
5,7,2'-trihydroxy-flavone 439  
5,7,4'-trihydroxy-8-methoxyflavone 439  
5,7,3'-trihydroxy,6,8,4'-trimethoxy flavones 498  
tubaic acid 236  
ulexine 461  
umbelliferone 328  
umuhengerin (5-hydroxy-6,7,3',4',5'-pen-  
tamethoxyflavone) 339  
ursolic acid 311  
vasakin 327  
vasicine 327  
vasicinone 327  
verbascoside (= acetoside) 339, 492  
verbenalin (= verbanalosite, cornin) 492  
verbenin (= aucubin) 492  
vernolepin 494  
vernonin 494  
verticillatine 426  
vinblastine 187  
vincristine 187  
vindoline 187  
warangalone 236  
xanthorrhizol 212  
yadanziosides 161  
yadanziosides A-H, O and P 161  
yamolosite 393



# Index of pharmaceutical terms

- abortifacient 45, 75, 78, 79, 105, 109, 135, 227, 229, 235, 254, 256, 267, 292, 320, 326, 329, 335, 353, 380, 392, 393, 395, 409, 412, 413, 419, 458, 484, 485, 487
- abortion 73, 110, 208, 316, 354, 387, 485, 492
- abortive 74, 99, 138, 327
- abortivum 129, 408, 494
- abscesses 111, 205, 235, 281, 294, 296, 306, 320, 353, 379, 391, 475, 491
- absinthism 142
- abstinence syndrome 255
- acaricidal 83, 432
- acetylcholine-induced contractions 75
- acne 142, 218, 475
- ACTH 404
- acute toxicity 121, 169, 418, 481
- adaptogenic 208, 480
- addiction 375
- adenocarcinoma 141, 142, 300, 308
- adenovirus 106
- adjunct 191, 252, 398, 479
- adjuvant 164, 368, 439, 494
- adjuvant-induced arthritis 164
- adrenoceptor 131, 461
- adrenocortical hormones 296
- adrenolytic 236
- agonist 255, 464, 492
- ague 267, 314, 318, 364, 367
- AIDS 54, 168, 171, 175, 266, 354, 383, 385, 443, 487
- alexipharmic 494
- allergy 90, 148, 330, 495
- alopecia 104, 186, 187, 350, 468, 472
- alterative 167, 179, 246, 254, 286, 289, 306, 331, 391
- amenorrhoea 98, 119, 187, 227, 313, 330
- amoebiasis 130, 182, 183
- amoebicidal 181, 200, 204, 296, 298, 332
- anaemia 121, 124, 156, 158, 250, 253, 254, 257, 284, 286, 355, 389, 467, 480, 491
- anaesthesia 219, 296
- anaesthetic 38, 150, 229, 230, 258, 259, 261, 316, 319
- analeptic 208
- analgesia 169, 177, 255, 375
- analgesic 36, 38, 41, 44, 47, 82, 83, 90, 120, 124, 139, 167, 168, 191, 193, 196, 200, 226, 243, 251, 255–258, 263, 266, 289, 295, 313, 328, 345, 359, 360, 375, 383, 403, 408, 443, 461, 463, 469, 477, 486, 492
- anaphylaxis 404, 439
- androgen 498
- angina pectoris 94, 420
- angiotensin 120, 142, 176, 264, 360
- anodyne 167, 374, 378, 467, 497, 501
- anorexia 146, 147, 217, 218, 254, 257
- antalgic 458
- anthelmintic 45, 79, 82, 98, 99, 104, 121, 133, 135, 139, 141, 147, 153, 156, 159, 167, 182, 184, 195, 198, 215, 223, 243, 246, 250, 253, 254, 258, 271, 278, 286, 287, 296, 320, 325, 371, 396, 408, 421–424, 448, 451, 491, 494, 497, 498, 500, 502
- anti-acne 142
- anti-amoebic 83, 89, 161, 182, 183, 192, 296, 319, 330, 366, 469
- anti-androgenic 121, 255
- anti-anginal 142
- anti-arrhythmic 199, 321, 426
- anti-arthritis 242, 289
- anti-asthma 147, 171
- anti-emesis 169
- anti-emetic 117, 119, 168, 218, 223, 489
- anti-epileptic 191
- anti-exudative 360
- anti-implantation 254, 350, 409, 419
- anti-inflammatory 36, 38, 41, 44, 47, 74–76, 79, 83, 86, 90, 100, 101, 112, 119, 120, 123, 139, 145, 147, 151, 152, 164, 176, 177, 208, 211, 223, 225, 243, 250, 263, 266, 272, 280, 293, 294, 300, 301, 308, 309, 311, 313, 328, 337, 346, 350, 359, 362, 369, 383, 386, 393, 398, 402, 405, 413, 414, 431, 439, 447, 448, 455–457, 461, 462, 464, 475, 476, 479, 480, 485–487, 494
- anti-influenza 407
- anti-oedema 42
- anti-oestrogenic 74, 255
- anti-ovipositional 414
- anti-ulcer 85, 121, 151, 164, 166, 194, 273, 274,

- 461, 469, 473, 477, 479  
anti-ulcerative 106, 107  
anti-ulcerogenic 41, 83, 457  
antibacterial 34, 38, 41, 45, 53, 79, 82, 83, 90,  
92–94, 102, 110, 119, 130, 164, 179, 180, 212,  
236, 243, 251, 252, 263, 278, 296, 301, 304, 329,  
346, 360, 369, 370, 380–383, 398, 405, 410, 414,  
439, 443, 455, 477, 479, 480, 486, 500  
antibiotic 74, 153, 250, 266, 271, 340, 341, 369,  
381, 383, 389, 403, 427  
antibronchitis 252  
anticholeric 467, 470  
anticholinergic 229, 230  
anticholinesterase 131  
anticoccidial 141, 421, 423  
anticomplementary 124, 126, 398  
anticonvulsant 83, 167, 168, 171, 359  
anticough 330  
antidepressant 303–305  
antidiabetic 101, 277, 278, 280, 281, 286, 476,  
480, 483  
antidiarrhoeal 41, 121, 179, 218, 263, 313, 407  
antidiuretic 375  
antidote 73, 82, 110, 133, 179, 208, 212, 227, 229,  
265, 272, 283, 288, 334, 339, 340, 342, 371, 452,  
460, 491  
antidysenteric 219, 296, 313, 315, 463  
antidyspeptic 404, 412  
antifebrile 418  
antifeedant 83, 90, 121, 196, 296, 298, 300, 410,  
414, 481  
antifertility 40, 121, 133, 254, 256, 257, 407, 419  
antifibrillatory 83  
antifungal 35, 41, 45, 82, 83, 90, 94, 113, 135,  
141, 156, 182, 183, 195, 196, 198, 212, 223, 243,  
271, 278, 332, 340, 360, 381–383, 395, 421,  
431–433, 443, 445, 454, 455, 469, 477, 480, 498  
antigalactagogue 316  
antigastralgie 501  
antigiardiasis 328  
antiglaucoma 168  
antigonorrhoeal 286  
antihaemorrhagic 303, 491  
antihaemorrhoid 289  
antihepatic 252  
antihepatotoxic 41, 211, 214, 223, 308  
antihistamine 83, 156, 179, 308, 498  
antihyperglycaemic 94, 95, 151, 360, 426, 486  
antihypertensive 35, 45, 94, 404, 406, 424, 431,  
442, 463, 492  
antijuvenile hormone 90, 92  
antileishmanial 243, 481, 495  
antileprotic 289, 480  
antileukaemia 76  
antileukaemic 207  
antimalarial 36, 45, 86, 113, 115, 116, 121, 131,  
139–141, 144–146, 151–153, 161, 163, 196, 199,  
204, 205, 219, 221, 223, 272, 274, 304, 339, 355,  
430, 464, 470, 479, 480  
antimicrobial 79, 97, 142, 151, 200, 211, 236, 243,  
264, 275, 280, 304, 305, 333, 339, 351, 352, 356,  
360, 369, 369, 385, 397, 403, 406, 411, 432, 469,  
492  
antimitotic 187, 410  
antimutagenic 141, 156, 158, 164, 165, 179, 214,  
311–313, 339, 346, 354, 380, 443, 448, 449, 496  
antineoplastic 300, 301, 321, 464  
antinociceptive 124, 126, 346, 383, 385  
antioxidant 41, 113, 196, 211, 255, 383, 405, 418  
antiparasitic 151, 153, 321, 325, 392, 479  
antiperiodic 289  
antiphlogistic 78, 90, 133, 329, 364, 365, 392,  
447, 451, 458  
antiplasmodial 200, 464, 469  
antiproliferative 304, 439, 440  
antiprotozoal 79, 278, 480  
antipyretic 79, 83, 89, 110, 113, 120, 125, 133,  
135, 139, 147, 176, 191, 218, 225, 226, 233, 249,  
263, 266, 272, 294, 320, 330, 359, 360, 362, 366,  
383, 402, 418, 431, 439, 442, 446, 451, 463, 472,  
481  
antiretroviral 303  
antirheumatic 43, 82, 133, 176, 195, 277, 367,  
392, 402, 479  
antiscorbutic 388, 433, 436, 491  
antisecretory 83, 121, 404, 407  
antiseptis 235  
antiseptic 93, 99, 100, 119, 129, 150, 152, 163,  
165, 185, 248, 249, 269, 284, 311, 313, 335, 337,  
338, 387, 391, 403, 427, 479  
antispasmodic 36, 79, 81, 83, 100, 101, 125, 153,  
155, 158, 191, 195, 205, 212, 216, 232, 235, 242,  
243, 258, 271, 321, 345, 346, 455, 464, 491, 498  
antispastic 464  
antispermatogenic 121, 256, 419  
antistress 191, 194, 480, 481  
antithrombotic 74, 121  
antitoxic 74, 117  
antitubercular 296  
antitumour 33, 34, 41, 43, 74, 75, 83, 95, 97, 124,  
126, 134, 142, 156, 188, 192, 194, 212, 236, 266,  
278, 308, 311–314, 329, 339, 341, 354, 356, 360,  
393, 406, 410, 411, 443, 445, 477, 485, 487, 495,  
498, 500  
antitussive 47, 74, 75, 133, 148, 230, 359, 363,  
374, 375, 378  
antivenin 330  
anuria 253, 371

- anxiety 43, 169, 170, 259, 425, 468  
anxiolytic 426  
aphrodisiac 73, 81, 94, 208, 253, 258, 272-274,  
281, 283, 286, 350, 381, 419, 431, 447, 450, 453,  
468, 472, 480  
apoplexy 304, 392  
appetizer 219, 479  
arteriosclerosis 93, 95, 368  
arthritis 93, 164, 208, 212, 220, 242, 289, 330,  
350, 359, 360, 417, 439, 447, 458, 472, 480  
ascariasis 463  
ascites 73, 124, 130, 141, 192, 250, 286, 311, 369,  
383, 392, 404, 405, 455  
ascitic 124, 134  
asphyxia 346, 469  
asthma 77, 81, 82, 89, 93, 104, 105, 109, 110,  
119, 147, 148, 153, 155, 160, 171, 176, 208, 210,  
214, 229, 230, 233, 243, 245, 253, 263, 266, 268,  
269, 271, 278, 284, 287, 307, 316, 319, 327,  
329-332, 334, 350, 353, 359, 363, 364, 386, 389,  
391, 397, 402, 403, 407, 415, 433, 450, 454, 460,  
463, 467, 470, 496, 501  
astringent 48, 150, 151, 155, 159, 163, 188, 215,  
223, 227, 246, 250, 254, 271, 277, 281, 286, 296,  
302, 303, 306, 311-313, 319, 331, 334, 335, 345,  
373, 379, 383, 387, 389, 402, 433, 442, 445, 459,  
460, 472, 489, 491, 500, 501  
ataxia 251  
atonic 81, 191  
azoospermia 187  
bacillary dysentery 227  
bactericidal 130, 298, 481  
bactericide 100  
bacteriostatic 130, 192, 355, 369, 434  
bechic 242, 374, 423, 436, 489  
beri-beri 330, 416  
bilharzia 392, 394, 396  
biliousness 133, 246, 353, 366, 389  
blastogenic 74  
blennorrhoea 205, 253  
boils 89, 93, 105, 109-111, 146, 150, 154, 155,  
158, 160, 161, 163, 164, 166, 217, 229, 263, 266,  
267, 277, 283, 284, 288, 296, 321, 325, 326, 331,  
379, 390, 392, 401, 408, 421, 423, 439, 441, 447,  
452, 454, 475, 489, 490  
bradycardia 130, 176, 223, 230, 343, 375, 404,  
426  
bradykinin 124, 359  
bronchitis 75, 86, 93, 105, 109, 110, 119, 155,  
159, 160, 229, 230, 296, 313, 316, 319, 327, 329,  
339, 342, 347, 359, 363, 364, 389, 392, 397, 401,  
403, 407, 445, 454, 491, 496  
bronchoconstriction 339  
bronchodilating 404  
bronchodilation 43, 171, 339  
bronchodilator 83, 327, 329, 434, 435  
bruises 164, 195, 216, 299, 303, 335, 353, 364,  
367, 408, 454  
burns 100, 104, 110, 150, 154, 163, 164, 166, 191,  
270, 353, 371, 398, 402, 403, 407, 458, 471, 491  
cachexia 179  
carcinogen 83, 106, 212, 311, 383  
carcinogenic 83, 85, 135, 137, 151, 192, 211, 214,  
293, 321, 322, 324, 354, 428, 432, 433  
carcinoma 95, 124, 141, 142, 161, 170, 236, 250,  
293, 296, 311, 321, 327, 332, 354, 405  
cardiac 46, 47, 125-128, 164, 196, 200, 220, 321,  
327, 336, 416, 472, 492  
cardioactive 46, 404, 406  
cardiotonic 95, 226, 318, 330, 454, 464  
cardiotropic 316  
carminative 78, 81, 82, 89, 99, 113, 124, 133, 138,  
148, 155, 195, 210, 216, 218, 242, 254, 258, 331,  
345, 346, 353, 389, 402, 407, 412, 413, 416, 441,  
472  
catarrh 86, 88, 89, 176, 210, 216, 265, 268, 303,  
316, 318, 319, 339, 342, 364, 368, 386, 397, 492  
cathartic 101, 104, 181, 264, 267, 320, 324-326,  
359, 373, 442, 443  
cellulitis 191  
central neurotoxic effects 187  
cephalalgia 330  
childbirth 82, 116, 138, 148, 155, 159, 179, 214,  
217-219, 222, 235, 272, 274, 275, 313, 314, 315,  
318, 320, 330, 331, 367, 390, 407, 413, 414, 417,  
433, 435, 437, 438, 447, 450, 453, 458, 502  
cholagogue 105, 109, 139, 147, 289, 313, 329, 475  
cholangitis 369  
cholecystitis 139, 147, 369  
cholera 119, 163, 229, 286, 336, 351, 415, 424,  
430, 460, 467, 470, 479, 483, 501  
cholesterol 94, 95, 97, 99, 110, 120, 182, 211, 236,  
255, 297, 300, 383, 398, 428, 486, 498  
choriocarcinoma(s) 186, 485  
cirrhosis 250  
colds 99, 113, 120, 123, 195, 216, 218, 250, 277,  
288, 339, 345, 363, 414, 418, 440, 491, 493, 497,  
501  
colic 81, 113, 155, 160, 205, 214, 217, 229, 230,  
248, 250, 269, 284, 311, 314, 326, 345, 379, 387,  
388, 403, 408, 417, 460, 491  
coma 192, 213, 215-219, 231  
condiment 94, 114, 117, 119, 155, 159, 195, 254,  
257, 345, 358, 403, 407, 408, 490  
congestion 101, 408, 425, 491  
conjunctivitis 73, 153, 250, 268, 278, 296, 381,  
382, 494  
constipation 83, 137, 150, 154, 187, 218, 339, 342,

- 367, 375, 397, 398, 489  
contact dermatitis 141, 143  
contact sensitivity 192  
contact toxicity 83  
contraceptive 74, 208, 447, 455, 456  
contusions 104, 294, 303, 335, 387, 402, 408, 491, 502  
convulsant 375  
convulsion 229  
coronary-dilating 316, 318  
cosmetic 25, 78, 100, 104, 216, 334, 335, 345  
cough(s) 36, 73, 75, 88, 89, 93, 104, 113, 118, 119, 133, 133, 148, 150, 154, 161, 163, 164, 179, 205, 210, 214, 216, 217, 229, 246, 250, 253, 253, 257, 277, 283, 286, 288, 307, 325, 327, 330, 330, 331, 334, 335, 345, 359, 359, 363, 363, 371, 373, 375, 379, 381, 386, 387, 389, 397, 399, 401, 402, 403, 407, 413, 415-417, 421, 433, 446, 447, 453, 454, 480, 488, 494, 494, 496, 498, 501, 502  
counter-irritant 165, 235, 242, 337, 359, 412  
cysticidal 181  
cystitis 205  
cytotoxic 41, 47, 79, 127, 130, 135, 140, 161, 163, 164, 166, 193, 194, 206, 219, 221, 236, 239, 250, 252, 264, 272-274, 293, 296, 321, 329, 333, 336, 354, 383, 410, 414, 433, 448, 455, 464, 485-487, 495  
decongestant 319  
dehydration 74, 290  
delayed-type hypersensitivity 220, 322  
delirium 231, 290  
demulcent 88, 167, 402  
deobstruent 148, 159, 387  
depressant 83, 125, 191, 223, 230, 231, 268, 316, 327, 360, 383, 414, 443  
depression 103, 176, 177, 187, 290, 332, 375, 410, 424-426  
depurant 475  
depurative 104, 185, 214, 218, 219, 275, 353, 359, 391, 449, 475, 489, 491  
dermatitis 79, 83, 141, 143, 239, 256, 350, 353, 497  
dermatophyte 94, 447  
dermatosis 147, 227, 276  
detoxicant 431  
diabetes 86, 119, 142, 145, 150, 154, 181, 185, 187, 353-355, 357, 359, 362, 368, 381, 387, 389, 398, 402, 459, 467, 470, 471, 479, 480, 483  
diaphoretic 99, 176, 185, 248, 250, 253, 272, 281, 330, 339, 342, 390, 418, 491, 502  
diarrhoea 40, 74, 78, 81-83, 89, 92, 93, 98, 101, 105, 106, 109, 110, 113, 117, 119, 130, 139, 147, 150, 154, 161, 163, 164, 176, 185, 205, 208, 216, 218, 223, 225, 227, 242, 243, 246, 249, 250, 253, 257, 264, 271, 277, 281, 283, 285, 286, 288, 289, 306, 307, 309, 310, 313, 314, 321, 325, 330, 334, 336, 345, 350, 353, 364, 366, 373, 379, 381, 382, 387-389, 393, 397, 398, 402, 403, 408, 413, 417, 418, 421, 423, 424, 430, 454, 458, 459, 463, 467, 479, 480, 490, 494, 496, 497, 501  
digestive 73, 76, 81, 113, 139, 256, 310, 313, 346, 360, 375, 382, 400, 418, 454, 458, 491  
diuresis 86, 251, 330, 368, 410, 479, 481  
diuretic 73, 86-89, 93, 97, 98, 105, 109-112, 119, 121, 133, 139, 147, 148, 150, 151, 155, 156, 158, 159, 163, 164, 167, 176, 177, 185, 195, 205, 208, 218, 219, 223, 224-228, 241-243, 246-248, 250-253, 269, 271, 275, 281, 286, 288, 303, 307, 308, 310, 313, 330, 331, 339, 350, 351, 359, 363, 368-371, 380-382, 387, 388-392, 394, 396, 397, 399, 401, 402, 413, 415-417, 424, 426, 433, 437, 439, 446, 447, 451, 459, 463, 475, 476, 479-481, 484, 489, 491, 494, 497, 501, 502  
dressing 202, 291, 299, 428, 456  
dropsy 253, 266, 286, 307, 359, 387, 392, 413, 415, 417, 436, 437, 459, 467, 470, 491, 494  
drowsiness 196, 425  
dysentery 73, 81, 82, 89, 104, 105, 109, 117, 119, 127, 129, 155, 161, 163, 176, 179, 185, 191, 195, 205, 218, 223, 227, 242, 243, 246, 249, 250, 253, 254, 257, 263, 264, 266, 269, 271, 272, 281, 286, 288, 289, 295, 296, 298, 307, 310, 311, 313, 326, 334, 336, 342, 350, 353, 367, 371, 373, 387, 389, 391, 392, 397, 402, 416, 418, 424, 427, 430, 442, 454, 458, 460, 463, 466, 467, 480  
dysmenorrhoea 124, 235, 242, 243, 303, 313, 320, 350, 463  
dyspepsia 81, 104, 119, 146, 167, 185, 195, 212, 214, 250, 253, 331, 334, 387, 389, 403, 407, 408, 463  
dyspeptic 124  
dysphoria 169, 171  
dyspnoea 74  
dystonia 171  
dystonic 171  
dysuria 133, 330, 463  
eczema 110, 133, 155, 195, 218, 250, 253, 254, 257, 294, 320, 321, 324-326, 330, 397, 431, 442, 445, 467, 471, 475, 497  
embryotoxicity 455  
emesis 74, 169, 185, 264  
emetic 81, 89, 119, 168, 176, 188, 205, 218, 223, 252, 267, 271, 283, 284, 287, 288, 319, 326, 330, 339, 350, 386, 489, 490  
emetocathartic 442, 460  
emmenagogue 78, 89, 100, 104, 110, 113, 124, 129, 133, 137-139, 147, 150, 151, 153, 155, 167, 195, 205, 208, 223, 224, 240, 241, 250, 253, 267,

- 292, 294, 319, 326, 339, 345, 359, 371, 387, 392, 408, 412, 413, 449, 472, 491, 501
- emollient 110, 111, 164, 250, 253, 350, 397, 399, 402, 458, 491
- emphysema 74, 268
- encephalitis 78, 250, 332
- epilepsy 127, 167, 171, 191, 225, 229, 245, 403, 424, 430, 446, 467
- epistaxis 147, 253, 310, 472
- erysipelas 195, 267
- erythema 171
- euphoria 43, 48, 169, 258
- expectorant 47, 81, 99, 110, 129, 133, 147, 148, 150, 153, 155, 241, 242, 246, 249, 254, 268, 331, 345, 353, 359, 363, 364, 387, 397, 445, 446, 459, 460, 489, 497, 501
- fatigue 100, 215, 229, 234, 258, 379
- febrifuge 89, 92, 98, 99, 105, 109–111, 117, 119, 126, 127, 129, 133, 138, 139, 146, 147, 156, 159, 161, 163, 167, 179, 205, 219, 223, 224, 227, 242, 246, 250, 252, 253, 254, 266, 272, 274, 310, 318, 326, 330, 339, 342, 353, 359, 367, 382, 387, 389, 392, 413, 414, 416–419, 429, 441, 467, 468, 470, 472, 489, 491, 494, 497, 501
- febrile stiffness 350
- fecundity 74
- fertility 76, 135, 189, 192, 243, 254, 284, 286, 308, 325, 352, 382, 385, 402, 410, 419
- fibrinolytic 35, 95
- fibrosarcoma(s) 73, 124, 236, 255, 273, 311, 477
- filariasis 92, 250
- fish poison 126, 129, 132, 177, 206, 235, 237, 239–242, 254, 256, 257, 259, 263, 266, 269, 270, 271, 283, 299, 321, 326, 414, 419, 435, 460, 471
- flatulence 78, 93, 119, 146, 331, 345, 398, 417, 501
- fractures 242, 316, 319
- framboesia 219, 429, 450
- fungicidal 53, 110, 137, 187, 216, 236, 263, 328, 329, 380, 381, 394, 395, 405, 443, 454
- fungicide 41, 237
- fungistatic 393, 396
- fungitoxic 196, 198, 346
- fungitoxicity 114, 196, 198, 349, 426
- furuncles 147, 401
- galactagogue 105, 109–111, 124, 188, 191, 218, 223, 242, 250, 283, 286, 359, 417, 419, 430, 484, 489, 491, 492, 501
- gallstones 150, 155, 210, 218, 368, 476
- gargle 275, 295, 318, 331, 339, 353, 371, 374, 423, 491
- gastralgia 73, 75, 119, 138
- gastric 113, 121, 133, 164, 170, 191, 212, 274, 303, 313, 335, 367, 382, 404, 425, 443, 455, 457, 476
- gastric ulcer 191, 455
- gastritis 82, 139
- gastro-enteritis 74, 83, 345, 393
- gastropathy 83
- genotoxicity 426
- giardiasis 328
- giddiness 118, 284
- glaucoma 48, 171, 175
- glossitis 367
- glucose tolerance 182, 278, 355, 480
- gonorrhoea 88, 89, 105, 109, 110, 119, 148, 208, 242, 269, 271, 281, 303, 309, 310, 313, 330, 339, 342, 364, 366, 387, 390, 402, 403, 442, 447, 450, 451, 459, 460, 480
- gout 36, 289, 292, 320, 325, 353, 368, 379, 447, 491
- granulocytopaenia 187
- growth regulator(s) / hormones 59, 87, 193, 232, 432
- haemagglutinating 393, 486
- haemagglutination 35, 263, 486
- haemagglutinator 73
- haematuria 86, 88, 290, 350, 491
- haemolysis 454
- haemoptysis 113, 281, 286, 310, 313, 460, 472
- haemorrhage(s) 74, 89, 124, 139, 156, 159, 167, 185, 205, 216, 307, 310, 313, 329, 389, 454, 472
- haemorrhoids 78, 101, 104, 110, 147, 161, 163, 199, 218, 219, 229, 269, 271, 286, 295, 296, 353, 371, 398, 403, 408, 412, 458, 472, 502
- haemostasis 89
- haemostatic 79, 142, 147, 150, 153, 185, 195, 269, 303, 310, 320, 321, 324, 398, 401, 473
- hallucination(s) 43, 48, 169, 231, 375, 393
- hallucinogenic 83, 230
- hay fever 110, 268
- hemiplegia 330
- hepatitis 105, 109, 191, 206, 220, 250, 306, 381, 382, 384–386, 464, 466
- hepatomegaly 340
- hepatoprotective 121, 250, 252, 383, 399, 480, 498, 500
- hepatotoxic 40, 211, 293, 294, 340, 410, 418, 485
- herpes 106, 107, 135, 192, 271, 288, 294, 304, 354, 357, 431, 450
- histiocytosis 186
- HIV 40, 41, 54, 106, 121, 148, 150, 212, 251, 278, 303, 354, 356, 357, 383, 439, 455, 464, 484, 485, 487
- HIV-1 54, 106, 148, 150, 212, 303, 354, 439, 455, 464
- HIV-1 protease 150

- Hodgkin's disease 186  
homeopathic 73, 135  
hookworms 195  
hormonal / hormone(s) 46, 90, 92, 166, 170, 170, 212, 223, 225, 238, 255, 296, 313, 323, 350, 355, 373, 404, 404, 407, 411, 448, 448, 449, 455, 456, 499  
hyperaemia 171  
hyperaesthesia 231  
hyperbilirubinaemia 340  
hypercalcaemic 419  
hypercholesterolaemia 182, 211, 406  
hypercholesterolaemic 182, 183  
hyperglycaemia 121, 354, 355, 454  
hyperglycaemic 94, 95, 355, 357, 360, 426, 486  
hyperlipemia 93  
hyperphylloerythrinaemia 340  
hypersecretion 425  
hypertension 119, 161, 208, 220, 223, 310, 313, 339, 418, 420, 424, 426, 431, 446, 472  
hypertensive 120, 406, 424, 431  
hypnotic 350, 373  
hypo-sensitization 110  
hypo-thyroidism 350  
hypocholesterolaemic 182, 183, 455  
hypoglycaemic 33, 121, 185, 187, 188, 206, 264, 277, 278, 296, 327, 354, 355, 357, 360, 362, 382, 383, 398, 414, 426, 448, 476, 480, 481, 484, 486  
hypolipidemic 300, 301, 486  
hypopodia 290  
hypopodial 290, 291  
hypotension 131, 135, 220, 290, 296, 343, 375, 426, 454, 494  
hypotensive 83, 99, 113, 120, 125, 130, 135, 139, 151, 171, 177, 188, 189, 191, 206, 223, 243, 316, 327, 350, 359, 382, 393, 418, 455  
hypotensive relief 83  
hypothalamic 170  
hypothermia 135  
hypothermic 83, 106, 196, 243, 312  
immune response 170, 364, 365  
immune system 170, 440  
immunization 170  
immunoglobulins 124  
immunomodulatory 33, 101, 134, 364, 365, 382, 399, 440, 487  
immunostimulant 41, 124, 134, 192, 208, 404, 464  
immunosuppression 119, 220, 464, 473, 485  
immunotherapeutic 481  
immunotoxin 73, 354, 357, 485  
impetigo 294, 320  
implantation 134, 135, 177, 243, 254, 350, 409, 419  
impotence 259, 286  
indigestion 116, 117, 119, 129, 185, 195, 218, 254, 257, 258, 272, 309, 321, 326, 345, 367, 463  
infarction 93, 121, 123, 230, 420  
inflammation 53, 73, 78, 82, 90, 101, 105, 109, 139, 147, 178, 211, 216, 218, 220, 294, 330, 335, 342, 343, 350, 353, 360, 368, 389, 397, 402, 447, 448, 451, 459, 461, 466, 476, 485, 489  
inflammatory exudation 359  
inflammatory mediators 220  
influenza 116, 119, 250, 303, 307, 345, 407, 418, 432, 433, 439  
inotropic action 404, 434  
insecticidal 34, 74, 76, 83, 90-92, 114, 131, 133, 135, 137, 141, 151, 156, 196, 212, 216, 223, 236, 241, 263, 266, 298, 322, 327, 329, 333, 343, 351, 352, 372, 393, 395, 410, 454, 455, 461, 465, 494, 498  
insecticide 76, 82, 85, 89, 90, 92, 93, 132, 161, 182, 235, 237, 239-241, 271, 341, 456, 491, 494  
insomnia 231, 259, 349, 350, 363, 373, 379, 403, 446, 467  
insulinotropic 480, 483  
interferon 124  
interleukin-1 220  
interstitial 463, 466  
intoxication 74, 272, 290, 293, 350, 418, 468  
intoxications 380  
ischias 93  
itch / itches 78, 89, 210, 214, 216, 235, 247, 272, 281, 287, 304, 321, 325, 326, 345, 371, 390, 397, 399, 413, 424, 430, 463, 466, 479, 490  
jamu 25, 82, 218, 331, 334  
jaundice 119, 129, 139, 146, 147, 205, 208, 210, 216, 218, 219, 242, 253, 284, 303, 307, 327, 329, 330, 340, 353, 364, 367, 368, 374, 381, 385, 387, 389, 475, 479, 480, 481, 483, 489, 491  
juvenile hormone 223, 225  
keloid 191  
kidney 49, 104, 150, 151, 155, 158, 196, 243, 246, 248, 307, 310, 318, 367-369, 379, 380, 387, 390, 399, 475, 488, 491, 492  
kidney-stone(s) 150, 155, 158, 243, 248, 318, 369, 399, 492  
kinetic experiments 142  
lactifuge 316, 319  
larvicidal 308, 332, 333  
larvicide 79  
laryngitis 148  
laxative 41, 100, 101, 103, 104, 167, 176, 181-185, 196, 241, 242, 245, 268, 269, 271, 286, 331, 353, 360, 386, 388, 389, 392, 396-399, 401-403, 429, 442, 443, 445, 446, 459, 475, 476  
Leishmaniasis 195, 481, 496

- leprosy 191, 194, 229, 235, 284, 299–302, 326, 412, 463, 468, 471, 494  
leucopaenia 464  
leucorrhoea 208, 250, 253, 281, 283, 294, 313, 320, 339, 342, 442, 458  
leukaemia 76, 79, 141, 161, 186, 212, 236, 250, 278, 280, 290, 293, 300, 311, 321, 327, 405, 439, 485, 494, 495  
Lewis lung carcinoma 170, 250, 405  
LH-releasing hormone 170  
lithiasis 86, 164, 336  
lithotriptic 371  
liver 74, 90, 113, 117, 119, 121, 131, 134, 139–142, 145, 147, 148, 156–158, 182, 199, 210, 211, 212, 214, 216, 218, 236, 277, 284, 293, 340, 353, 355, 364, 380, 382, 386, 387, 404, 405, 408, 413, 418, 424, 442, 443, 459, 475, 476, 480, 486, 491, 498, 500–502  
longevity 94, 258, 336  
lumbago 82, 93, 116, 148, 176, 181, 184, 208, 272, 281, 286, 330, 353, 364, 366, 367  
lungs 171, 364, 404, 439, 489  
luteinizing hormone 170  
lymphadenopathy 320  
malaria 28, 73, 79, 86, 92, 113, 114, 139–141, 143, 145–147, 151, 161–163, 185, 199, 200, 204, 219, 225, 247, 253, 254, 258, 272, 274, 280, 306, 329, 330, 353, 387, 392, 402, 442, 454, 470, 479, 480, 483, 498  
malignancy 135  
malignant 290, 354  
masticatory 82, 181, 258, 262, 331, 389  
mastitis 320, 475, 502  
measles 227  
menorrhagia 110, 139, 398  
metabolic acidosis 290  
metritis 320  
metrorrhagia 472  
microcirculation 95, 192  
microtubule 187  
migraine 167, 387  
miliaria 371  
mitosis 187  
molluscicide 256, 392, 395  
motion sickness 230  
mouthwash 100, 150, 154, 185, 223, 242, 249, 268, 316, 402, 445  
mucilage 100, 208, 213, 397–399, 401, 403, 492  
mucolytic(s) 327  
multiple sclerosis 168, 171, 175  
mumps 229, 353, 416  
muscle-relaxing system 360  
musculotropic 418  
mutagenesis 364, 365, 487  
mutagenic 31, 74, 83, 135, 293, 346, 364, 365, 410, 473  
myalgia 187  
mycoherbicide 261  
mycotoxigenic 142  
mydriasis 231  
mydriatic 230  
myelogenic 290  
myelosuppression 186  
myorelaxant 193  
myosis 426  
narcois 296  
narcotic 36, 104, 167, 168, 173, 175, 191, 192, 256, 258, 269, 288, 373, 375, 378  
nausea 116, 168, 170, 196, 289, 302, 307, 313, 373, 421, 423, 502  
nauseant 81  
necrosis 74, 187, 206, 220, 251, 296, 421, 444, 464  
necrotic 426  
nematicidal 83, 90, 91, 142, 171, 187, 216, 223, 290, 300, 351, 352  
nematode(s) 22, 91, 96, 112, 142, 144, 171, 174, 174, 195, 196, 198, 225, 230, 244, 244, 256, 292, 329, 351, 356, 357, 362, 370, 373, 385, 399, 406, 420, 422, 456, 465, 478, 482, 486, 487, 495  
nephritis 164, 208, 310, 368, 381, 421, 423, 463  
nephrotic syndrome 86, 87  
nephrotoxicity 135  
nervine 195, 266, 472  
neuralgia 164, 167, 187, 345, 417, 491  
neurasthenia 147, 463  
neuroleptic 83, 425  
neuropathy 290  
neurotoxic 142, 164, 187, 336  
neurotoxicity 139, 186  
neurotrophic 393, 395  
nociception 375  
non-Hodgkin's lymphoma 186  
obese 223  
obstinate vomiting 167, 281, 467, 470  
obstructive jaundice 340, 481  
oedema 42, 74, 79, 120, 133, 135, 163, 164, 176, 192, 211, 223, 246, 248, 251, 253, 263, 266, 303, 311, 330, 346, 353, 359, 363, 381, 398, 455, 463, 476, 479, 486, 491, 498  
oestrogen(s) / oestrogenic 47, 255, 264, 296, 419, 472  
oliguria 290, 363, 463  
ophthalmia 104, 164, 199, 331, 339, 408, 446, 491  
opiodergic receptor(s) 191  
opiod peptide(s) 375  
opiod receptor(s) 375  
ostalgia 320

- osteosarcoma 300  
ovicidal 90  
oviposition 121, 136, 421, 422, 481  
palsy 171  
panacea 81, 93, 502  
panic 43, 169  
papaverine-like 418  
papilloma(s) 212, 311, 354  
papillomatosis 135  
paralysis 164, 235, 242, 290, 325, 330, 343, 410, 412, 472, 491, 492  
parasympatholytic(s) 229, 231, 278  
parkinsonism 171, 230, 231  
parturition 124, 167, 250, 253, 460, 491, 494  
pectoral 195, 205, 267, 359, 402, 407, 423  
peptic ulcers 208, 212, 214, 367, 425  
peripheral neurotoxic effects 187  
peripheral vascular resistance 404  
peristalsis 101, 127, 181, 398, 443  
phagocytic 110, 134, 135, 481  
phagocytosis 364, 365, 399  
pharmacokinetic 139  
pharyngitis 148, 250, 294, 367, 392  
pheromone 79  
phlebitis 191  
phlebology 473  
photodermatitis 304  
photosensitization 48, 276, 340  
phototoxic 53, 275  
phototoxicity 212, 278  
phototoxins 275  
phthisis 124, 163, 336, 359  
phytohormones 132  
phytophotodermatitis 79  
phytosterol(s) 151, 250, 328, 368, 427, 432, 439, 478  
phytotherapeutic 97, 405  
phytotherapy 82, 92, 123, 139, 150, 194, 214, 344, 373, 397, 399, 470, 479  
phytotoxic 151, 196, 380  
piles 73, 119, 208, 222, 403, 454, 458, 459, 491  
pimples 104, 379, 402, 467  
pinworms 215, 353, 391, 484  
piscicidal 237, 241, 327  
platelet activating factor 220  
platelet aggregation 34, 74, 95, 97, 98, 121, 142, 200, 264, 308, 309, 327, 360, 431, 464, 494  
pleurisy 359  
pleuritis 392  
pneumonia 89  
post-phlebotic syndrome 192  
poultice 98, 104, 111, 116, 118, 119, 127, 133, 148, 156, 160, 161, 163, 164, 166, 176, 214, 217, 223, 225, 227, 242, 248, 249, 257, 267, 271, 272, 274, 281, 284, 288, 294, 315, 316, 318–320, 325, 327, 330, 331, 335, 350, 371, 379, 387, 392, 397, 401, 412, 413, 439, 441, 467, 475, 490, 491, 494, 496, 497, 501, 502  
pre-anaesthetic 229, 230  
primary humoral immune response 170  
progestogens 296  
prolactin 135, 316  
proliferation 170, 192, 208, 212, 321, 336, 464, 485  
prophylactic 21, 99, 141  
prostration 251  
pruritic 139, 141, 147  
psoriasis 101, 276, 386, 443, 447–449, 451, 475  
psychiatric 424  
psychoactive 54, 82, 171  
psychosis 170, 259  
psychotropic 170  
purgative 34, 100, 101, 104, 133, 148, 176, 181–183, 185, 188, 205, 254, 263, 266, 267, 269, 270, 289, 295, 321, 322, 326, 359, 381, 386, 401, 421, 438, 442, 443, 445, 446, 447, 488, 490, 491  
quotidian fever 320  
rabies 468  
radioprotective 125  
rash(es) 288, 458, 471  
refrigerant 104, 155, 164, 345, 359, 388, 389, 391  
repellants 199  
repellency 481  
repellent 90, 141, 223, 340, 405, 421, 494, 498  
restenosis 120, 123, 439  
restorative 359  
retroviruses 303, 382  
reverse mutation 426  
reverse transcriptase 54, 251, 383, 385, 386, 484  
rheumatism 82, 99, 113, 117, 119, 139, 147, 156, 160, 164, 167, 176, 192, 205, 215, 216, 227, 229, 235, 242, 250, 266, 281, 286, 288, 289, 294, 299, 303, 320, 321, 325, 327, 330, 331, 334, 339, 341, 353, 359, 363, 364, 367, 368, 379, 409, 412, 413, 415, 421, 423, 447, 449–452, 454, 467, 470, 480, 491, 502  
rhinitis 89, 110, 278, 442  
ringworm 9, 161, 181, 196, 212, 229, 271, 321, 325, 346, 371, 412, 431, 442, 443, 445, 446  
roundworm(s) 133, 195  
rubefacient 82, 93, 176, 266, 321, 325, 345, 413, 472, 491  
saluretic 86, 476  
sarcoma(s) 73, 124, 130, 134, 142, 156, 186, 250, 308, 311, 494, 498  
scabies 205, 217, 253, 280, 286, 288, 296, 321, 325, 326, 407, 408, 413, 439, 441, 442, 445, 446, 471, 479, 483



- scalds 163, 164, 166  
schistosomiasis 73, 74, 141, 322, 393  
schizonticide 140  
sciatica 299  
scrofula 303, 306, 458, 491  
scurf 161, 210, 431  
scurvy 21, 214, 318, 371  
seborrhoea 82  
secondary humoral immune response 170  
secondary immunization 170  
sedative 81, 82, 139, 147, 164, 167, 168, 196, 208,  
231, 246, 263, 269, 310, 311, 313, 316, 319, 327,  
345, 350, 352, 359, 374, 378, 408, 425, 431, 438,  
454, 459, 464  
senility 186, 191, 425  
sepsis 481  
serotonergic 425  
sex pheromone 79  
shingles 227  
silicosis 464  
sinusitis 89, 120, 155, 159, 345  
skin grafting 191  
smallpox 216, 218, 229, 342, 381, 391, 392, 408,  
479  
soporific 167  
spasmodic colitis 399  
spasmolytic 40, 78, 83, 168, 187, 346, 383, 418,  
420, 426, 480  
spasm(s) 137, 167, 168, 171, 229, 230, 346, 350,  
375  
spastic 171  
spasticity 171  
spermatogenesis 255  
spermatorrhoea 281, 467  
spice 83, 114, 117, 119, 210, 214, 216, 403, 407  
sprain(s) 229, 234, 294, 299, 316, 319, 325, 334,  
497, 502  
sprue 129, 132, 253, 353, 494  
stenocardia 94  
stimulant 43, 81, 82, 94, 124, 126, 133, 135, 139,  
142, 151, 153, 155, 156, 159, 216, 218, 223, 227,  
229, 235, 242, 243, 254, 259, 271, 290, 313, 327,  
331, 339, 342, 345, 360, 414, 426, 469, 472  
stomachic 78, 81, 82, 98, 100, 104, 113, 118, 129,  
133, 137, 139, 147, 155, 158, 179, 195, 206, 210,  
216, 218, 223, 224, 227, 229, 242, 296, 306, 311,  
313, 326, 331, 339, 345, 346, 353, 367, 412, 413,  
417, 429, 460, 467, 470-472, 502  
styptic 151, 153, 210, 216, 284, 325, 335, 336,  
373, 472  
sudorific 110, 155, 156, 158, 160, 195, 223, 269,  
288, 331, 339, 345, 374, 378  
sympatholytic 425  
sympathomimetic 231, 243  
syphilis 104, 179, 229, 295, 391, 447, 450, 452,  
453  
T lymphocytes 354  
T-cell dependent immune responses 220  
T-independent antigens 124  
tachycardia 258, 454  
tachypnea 454  
taeniafuge 254, 257, 258  
teratogenic 31, 74, 187, 256, 409  
testicles 350, 502  
tetanic spasms 375  
tetanus 167  
thrombocytopaenia 186, 293  
thrombosis 95, 97, 98  
thrush 294, 330, 407, 448  
tight chest 105, 109  
tinnitus 186  
tonic 82, 105, 109, 113, 119, 123, 124, 129, 133,  
138, 139, 147, 151, 167, 190-192, 199, 208, 219,  
223, 227, 240, 242, 246, 253, 272, 274, 281, 283,  
286, 289, 296, 310, 315, 326, 331, 332, 339, 353,  
359, 367, 386, 387, 402, 413, 417, 419, 424, 433,  
435, 437, 439, 441, 442, 447, 450, 453, 463, 467,  
468, 470, 472, 475, 479, 480, 489, 491, 494, 497,  
500-502  
tonsillitis 148, 250  
tranquilizer 313, 424  
traumas 186, 425  
tremor(s) 142, 171, 423, 426, 498  
tuberculosis 113, 146, 147, 161, 192, 216, 247,  
311, 313, 327, 329, 431, 434, 463, 502  
tuberculostatic 463  
tumorigenic 101, 343, 344  
tympanites 138, 438  
ulcer(s) 73, 85, 89, 104, 121, 121, 129, 133, 139,  
147, 150, 151, 154, 155, 161, 164, 164, 166, 167,  
181, 184, 191, 191, 195, 206, 208, 210, 212, 214,  
218, 223, 224, 227, 242, 246, 249, 272-274, 273,  
274, 285, 286, 292, 295, 296, 313, 314, 316,  
319-322, 325, 326, 334, 335, 337, 341, 350, 353,  
364, 367, 387, 398, 402, 404, 407, 421, 423, 425,  
447, 455, 459, 461, 469, 473, 477, 479, 489, 491,  
501  
urinary 73, 74, 86, 131, 139, 176, 205, 216, 229,  
243, 245, 269, 278, 288, 342, 350, 368, 370, 387,  
407, 408, 447, 450, 451, 454, 463, 466, 475, 479,  
480, 491  
urologic 223  
urticaria 292, 386, 442  
uterine 113, 135, 138, 167, 205, 208, 218, 290,  
300, 360, 414, 424, 426, 430, 448, 469, 491  
uterotonic 329  
uveitis 230  
vasoconstriction 131

vasodilation 492  
vasodilative 309  
vasodilator 41, 142, 223, 404  
vasodilatory 131  
venereal diseases 245, 253, 319, 326, 381, 387,  
442, 447  
vermifuge 81, 86, 88, 93, 100, 104, 119, 139, 147,  
155, 158, 185, 195, 196, 219, 222, 226, 254, 257,  
269, 270, 272, 287, 302, 353, 359, 367, 412, 421,  
423, 430, 442, 446, 460, 467, 468, 470, 471, 479,  
483, 488, 494  
verruculose 391, 450  
vertigo 192, 318, 367, 458  
veterinary 22, 67, 100, 104, 133, 230, 299, 395,  
412, 417, 423, 430, 479, 480, 498  
viricidal 137  
virility 459  
vitiligo 119  
vomitive 185, 188  
vulnerable 89, 100, 147, 252, 253, 296, 303, 306,  
310, 316, 319, 325, 407, 433, 436  
wart(s) 75, 161, 244, 263, 266, 268, 269, 294, 371  
whipworm 195  
wormicidal 418, 449  
wound(s) 78, 86, 89, 90, 92, 92, 93, 98, 99,  
100-103, 101, 104, 105, 109, 110, 129, 150, 151,  
153, 154, 159, 163, 164, 166, 167, 180, 181, 185,  
191, 191, 206, 210, 216, 218, 242, 246, 248-250,  
252, 263, 271, 272, 274, 277, 284, 286, 288, 292,  
293, 294, 299, 303, 306, 310, 319-322, 321,  
324-326, 324, 326, 328, 330, 334-336, 339, 342,  
350, 353, 359, 360, 364, 366, 371, 382, 391, 399,  
401-403, 407, 407, 417, 424, 429-431, 434, 435,  
446, 447, 451, 454, 459, 463, 467, 471, 475, 479,  
480, 483, 489, 491, 494, 496, 501  
xerosis 141  
xerostomia 360  
yaws 318, 459

# Index of scientific plant names

Page numbers printed in bold refer to main treatment.

- Abies 448  
Abreae 75  
Abrus Adanson **73**  
Abrus fruticosus Wight & Arn. **73, 76**  
Abrus melanospermus Hassk. 76  
Abrus precatorius L. 35, **73, 77**  
– subsp. africanus Verdc. 77  
– subsp. precatorius 77  
Abrus pulchellus Wallich ex Thwaites 76  
Acacia Miller 235  
Acanthoideae 122, 432  
Achillea L. 156  
Achillea collina J. Becker ex Reichenb. 80  
Achillea lanulosa Nutt 80  
Achillea millefolium L. 56, **77, 90**  
– subsp. collina (J. Becker ex Reichenb.) Weiss 80  
Achyranthes lanata L. 87  
Achyranthes sanguinolenta L. 88  
Achyranthes villosa Forssk. 87  
Acoraceae 81, 84  
Acoreae 84  
Acorus L. 82, 84  
Acorus asiaticus Nakai 81  
Acorus calamus L. 65, **81, 369**  
– var. americanus (Raf.) Wulff 84  
– var. angustatus Bess. 82, 84  
– var. calamus 82, 84  
– var. verus L. 84  
Acorus gramineus Soland. ex Aiton 83, 85  
Acorus terrestris Spreng. 81  
Adenoropium multifidum (L.) Pohl 326  
Adhatoda Miller 328  
Adhatoda vasica Nees 329  
Adhatoda zeylanica Medic. 329  
Aegle marmelos (L.) Correa 24  
Aerva Forssk. **86, 111**  
Aerva cochinchinensis Gagn. 87  
Aerva curtisii Oliv. 87  
Aerva lanata (L.) A.L. Juss. ex Schultes **86, 87**  
Aerva sanguinolenta (L.) Blume **86, 88**  
Aerva scandens (Roxb.) Wallich ex Moq. 88  
Aerva timorensis Moq. 88  
Agalma Miq. 434  
Aganope Miq. 237  
Agave L. 47, 448  
Ageratina riparia (Regel) R.M. King & H. Robinson 369  
Ageratum L. **88**  
Ageratum conyzoides L. **88, 92, 93**  
– var. houstonianum (Miller) Sahu 91  
Ageratum houstonianum Miller **88, 92**  
Ageratum mexicanum Sims 92  
Ailanthus excelsa Roxb. 328  
Albizia chinensis (Osbeck) Merr. 115  
Alliaceae 93  
Allium L. **93**  
Allium altaicum Pallas 96  
Allium bakeri Hoop. non Regel 98  
Allium bakeri Regel 98  
Allium bouddhae O. Debeaux 98  
Allium cepa L. **93, 97**  
– cv. group Aggregatum 97, 98  
– cv. group Common Onion 97, 98  
Allium chinense G. Don 93–95, **98**  
Allium fistulosum L. 93, 94, **98**  
Allium longicuspis Regel 96  
Allium odorum auct. non L. 99  
Allium sativum L. 83, **93, 99, 383**  
– cv. group Common Garlic 99  
– cv. group Ophioscorodon 99  
Allium schoenoprasum auct. non L. 98  
Allium senescens Miq. 99  
Allium tuberosum Rottler ex Sprengel 93–95, **99**  
Allium uliginosum G. Don 99  
Aloe L. 41, 65, **100**  
Aloe arborescens Miller 100  
Aloe barbadensis Miller 102, 104  
Aloe camperii Schweinf. 100  
Aloe ferox Miller 100–103, **104**  
Aloe perfoliata L.  
– var. vera L. 104  
Aloe perryi Baker. 100  
Aloe saponaria (Aiton) Haw. 100  
Aloe vera (L.) Burm.f. **100, 104**  
Aloeaceae 102  
Alpinia Roxb. 114  
Alpinia galanga (L.) Willd. 83

- Alpinia officinarum* Hance 83  
 Alpinieae 114  
*Alternanthera* Forssk. **105**  
*Alternanthera amoena* Backer & v. Slooten 108  
*Alternanthera bettzickiana* (Regel) Nicholson 108  
*Alternanthera brasiliana* (Torner) O. Kuntze 105, 106, 107, **108**  
*Alternanthera denticulata* R.Br. 109  
*Alternanthera ficoidea* (L.) P. Beauv. 105, 106, 107, **108**  
 – var. *bettzickiana* (Nicholson) Backer 108  
 – var. *versicolor* (Lem.) Backer 108  
*Alternanthera manillensis* (Walp.) Kanis 108  
*Alternanthera nodiflora* R.Br. 109  
*Alternanthera philoxeroides* (Mart.) Griseb. 105, 106, 107, **108**  
*Alternanthera pungens* Kunth 105, **108**  
*Alternanthera repens* (L.) Link (1821) non Gmelin 108  
*Alternanthera sessilis* (L.) DC. **105, 109**  
*Alternanthera strigosa* Hassk. 108  
*Alternanthera tenella* Colla 106, 108  
 – var. *versicolor* (Lem.) Veldk. 108  
*Alternanthera triandra* Lamk 109  
*Althaea officinalis* L. 399  
*Alyxia stellata* (Forst.) Roem. & Schultes 217  
 Amaranthaceae 86, 105, 110, 111  
*Amaranthus* L. 111  
*Amaranthus caudatus* L. 110  
*Amaranthus cruentus* L. 112  
*Amaranthus leucocarpus* S. Watson 110  
*Amaranthus spinosus* L. **110**  
*Amaranthus tricolor* L. 111  
*Amaranthus viridis* L. 111  
 Amaryllidaceae 36, 209  
*Amomum* Roxb. **113**  
*Amomum aculeatum* Roxb. 113, **116**  
*Amomum cardamomum* L. 114  
*Amomum ciliatum* Blume 116  
*Amomum compactum* Soland. ex Maton 114, 115  
*Amomum echinospaera* K. Schumann ex Gagnep. 118  
*Amomum fenzlii* Kurz 113, 114  
*Amomum flavum* Ridley 116  
*Amomum gracile* Blume **116**  
*Amomum hochreutineri* Valetton **116**  
*Amomum krervanh* Pierre ex Gagnep. **113, 117**  
*Amomum ligulatum* R.M. Smith **117**  
*Amomum longiligulare* T.L. Wu 113, 117  
*Amomum squarrosum* Ridley 113, **117**  
*Amomum stenocarpum* Valetton **118**  
*Amomum subulatum* Roxb. 114  
*Amomum testaceum* Ridley 117  
*Amomum tsao-ko* Crevost & Lem. 113, 114  
*Amomum uliginosum* J.G. König ex Retz. 113, **118**  
*Amomum villosum* Lour. **113, 118**  
 – var. *xanthioides* (Wallich ex Baker) T.L. 118  
*Amomum xanthioides* Wallich ex Baker **113, 118**  
*Anamirta cocculus* (L.) Wight & Arnott 132  
 Andrographideae 122  
*Andrographis* Wallich ex Nees 122  
*Andrographis paniculata* (Burm.f.) Wallich ex Nees **119, 368**  
*Andrographis subspathulata* C.B. Clarke 119  
*Angelica* L. 123, 124, 125, 126  
*Angelica acutiloba* (Siebold & Zucc.) Kitagawa **123**  
 – var. *acutiloba* 125  
 – var. *iwatenis* (Kitagawa) Hikino 125  
*Angelica archangelica* L. 124  
*Angelica dahurica* (Fisch. ex Hoffm.) Benth. & Hook.f. ex Franchet & Savat. 124, 126  
*Angelica polymorpha* Maxim. 124, 126  
*Angelica sinensis* (Oliv.) Diels 124  
*Anisomeria* D. Don 394  
*Annona* L. 34  
*Annona squamosa* L. 237, 481  
 Annonaceae 34  
*Anthurium* Schott 84  
*Antiaris* Lesch. 126, 127, 128  
*Antiaris africana* Engl. 126  
*Antiaris macrophylla* R.Br. 126  
*Antiaris toxicaria* Lesch. **126, 342, 468**  
 – subsp. *macrophylla* (R.Br.) C.C. Berg 128  
 – subsp. *toxicaria* 128  
*Antiaris welwitschii* Engl. 126  
*Antidesma buniis* (L.) Sprengel 24  
 Apocynaceae 36, 56, 187, 426, 469  
 Araceae 35, 84  
*Araucaria* Juss. 472  
*Arcangelisia* Becc. 129, 132  
*Arcangelisia flava* (L.) Merr. **129**  
*Arcangelisia lemniscata* (Miers) Becc. 129  
*Arcangelisia loureiri* (Pierre) Diels 129  
*Arcangelisia tympanoda* (Lauterb. & K. Schumann) Diels 132  
*Aristolochia* L. **133, 463**  
*Aristolochia bracteata* Retz. 133  
*Aristolochia bracteolata* Lamk 133–136  
*Aristolochia contorta* Bunge 133, 134  
*Aristolochia debilis* Sieb. & Zucc. 133, 135, 136  
*Aristolochia elegans* Masters 133, 134  
*Aristolochia fangchi* Y.C. Wu ex L.D. Chou & S.M. Hwang 133, 134  
*Aristolochia heterophylla* Hemsley 133, 134  
*Aristolochia imbricata* Masters 138

- Aristolochia indica* L. 133-137  
*Aristolochia mandshuriensis* Kom. 133, 134, 135  
*Aristolochia megalophylla* K. Schumann 138  
*Aristolochia membranacea* Merr. 138  
*Aristolochia mindanaensis* Warb. 138  
*Aristolochia philippinensis* Warb. 137  
*Aristolochia roxburghiana* Klotzsch 138  
*Aristolochia rumphii* Kostel. 137  
*Aristolochia sericea* Blanco 138  
*Aristolochia serpentaria* L. 133  
*Aristolochia tagala* Cham. 133, 138  
*Aristolochia westlandii* Hemsley 133  
*Armeria* Willd. 411  
*Artemisia* L. 139, 156, 196, 224, 369  
 – section *Abrotanum* 143  
 – section *Dracunculus* 143  
*Artemisia absinthium* L. 139, 141, 142  
*Artemisia annua* L. 56, 79, 113, 139, 146, 162, 200, 223  
*Artemisia apiacea* Hance 139, 143, 145, 147  
*Artemisia campestris* L. 143, 147  
*Artemisia capillaris* Thunb. 139, 147  
*Artemisia cina* Berg ex Poljakov 139, 142, 145  
*Artemisia dracunculus* L. 139  
*Artemisia indica* Willd. 143, 147  
*Artemisia maritima* L. 139, 421  
*Artemisia nilagirica* (C.B. Clarke) Pampan. 139, 143  
*Artemisia princeps* Pampan. 143  
*Artemisia scoparia* Waldst. & Kit. 139, 143, 147  
*Artemisia vulgaris* L. 139, 147  
*Asclepias curassavica* L. 127  
 Asphodelaceae 100, 102  
*Aster* L. 95  
 Asterales 45  
*Astroloba* Uitew. 102  
*Atropa* L. 233  
*Atropa belladonna* L. 231  
*Aucuba* Thunb. 399  
*Azadirachta indica* A.H.L. Juss. 23, 83, 90, 143, 200, 237, 255, 481  
*Ballota* L. 405  
*Barleria* L. 368  
*Belamcanda* Adans. 149  
*Belamcanda chinensis* (L.) DC. 148  
*Belamcanda punctata* Moench 148  
 Berberidaceae 36, 131  
*Berberis* L. 131, 465  
*Bidens* L. 150  
*Bidens abyssinica* Sch. Bip. 153  
*Bidens bipinnata* L. 150, 151, 152, 153  
*Bidens biternata* (Lour.) Merr. & Sherff 150, 151, 152, 153  
*Bidens chinensis* Willd. 153  
*Bidens leucorrhiza* (Lour.) DC. 154  
*Bidens pilosa* L. 150, 154  
 – var. *bipinnata* (L.) Hook.f. 153  
 – var. *minor* (Blume) Sherff 154  
*Bidens sundaica* Blume 154  
*Bidens tripartita* L. 150, 152, 155  
 Bignoniaceae 41  
*Blumea* DC. 155  
*Blumea appendiculata* (Blume) DC. 158  
*Blumea arfakiana* Martelli 155, 156, 157, 158  
*Blumea arnakidophora* Mattf. 157, 158  
*Blumea balfourii* Hemsl. 158  
*Blumea balsamifera* (L.) DC. 155, 158, 368  
*Blumea bodinieri* Vaniot 159  
*Blumea chinensis* auct. non (L.) DC. 157, 160  
*Blumea conspicua* Hayata 160  
*Blumea grandis* (Wallich) DC. 158  
*Blumea lacera* (Burm.f.) DC. 155, 159  
*Blumea lanceolaria* (Roxb.) Druce 155, 160  
*Blumea laxiflora* Elmer 160  
*Blumea myriocephala* DC. 160  
*Blumea pubigera* auct. non (L.) Merr. 157, 160  
*Blumea riparia* (Blume) DC. 155, 157, 160  
*Blumea runcinata* DC. 159  
*Blumea thyrsoides* Sch. Bip. 159  
*Blumea zollingeriana* C.B. Clarke 158  
*Blumeopsis* Gagnep. 157  
*Boesenbergia* O. Kuntze 332  
 Boraginaceae 34, 41, 179, 180, 292, 405  
*Borassus flabellifer* L. 24, 269  
*Brachiaria decumbens* Stapf 352  
*Brachypterum robustum* (Roxb. ex DC.) Dalz. & Gibs. 241  
*Brachypterum scandens* (Roxb.) Benth. 241  
*Brassaia* Endl. 434  
*Breynia* J.R. Forster & J.G. Forster 384  
*Bucea* J.F. Miller 161  
*Bucea acuminata* Li 162  
*Bucea amarissima* (Lour.) Desv. ex Gomes 160  
*Bucea antidysenterica* J.F. Miller 161, 162  
*Bucea javanica* (L.) Merr. 143, 160, 200  
*Bucea luzoniensis* S.Vidal 162  
*Bucea macrobotrys* Merr. 162  
*Bucea mollis* Wallich ex Kurz 162  
*Bucea sumatrana* Roxb. 160  
*Brugmansia* Pers. 232  
*Bryophyllum* Salisb. 163, 336  
*Bryophyllum calycinum* Salisb. 166  
*Bryophyllum daigremontianum* (Hamet & Perr.) Berger 164  
*Bryophyllum pinnatum* (Lamk) Oken 59, 163, 166, 336  
*Bryophyllum proliferum* Bowie 163, 164, 165, 167

- Bryophyllum serrata* Blanco 337  
*Bryophyllum tubiflorum* Harv. 164  
*Bulbostylis* Kunth 224  
 Cactaceae 42  
*Cajanus cajan* (L.) Millsp. 112, 212  
*Calophyllum inophyllum* L. 24  
*Calotropis gigantea* (L.) Dryander 127  
*Calystegia sepium* (L.) R. Br. 360  
*Camellia sinensis* (L.) Kuntze 260  
*Cannabis* L. 167, 172  
*Cannabis sativa* L. 43, 59, **167**, 375  
*Canthium horridum* Blume 220  
*Capsicum* L. 38, 74  
*Cardiospermum corindum* L. 176  
*Cardiospermum grandiflorum* Swartz 178  
*Cardiospermum halicacabum* L. **176**  
*Cardiospermum luridum* Blume 176  
*Cardiospermum microcarpum* Humb. 176  
*Carmona* Cav. 179, 180  
*Carmona microphylla* (Lamk) G. Don 178  
*Carmona retusa* (Vahl) Masam. **178**  
 Caryophyllaceae 42  
 Caryophyllales 42  
*Cassia* L. 101, 103, **181**, 369, 443, 444  
*Cassia agnes* (de Wit) Brenan 185  
*Cassia alata* L. 445  
*Cassia bakeriana* Craib 181  
*Cassia bartonii* F.M. Bailey 185  
*Cassia borneensis* Miq. 446  
*Cassia fistula* L. **181**, **183**  
*Cassia grandis* L.f. **181**, **184**  
*Cassia javanica* L. 181, 182, 183, **185**  
 – subsp. *agnes* (de Wit) K. Larsen 185  
 – subsp. *nodosa* (Roxb.) K. & S.S. Larsen 185  
*Cassia nodosa* Roxb. 185  
*Cassia pachycarpa* de Wit 184  
*Cassia siamea* Lamk 115  
*Cassia sophera* L. 446  
*Cassia tora* L. 446  
*Casuarina* L. 336, 338  
*Casuarina equisetifolia* L. 484  
*Catharanthus* G. Don 188, 190, 469  
*Catharanthus lanceus* (Bojer ex A.DC.) Pichon 188  
*Catharanthus pusillus* (Murr.) G. Don 185  
*Catharanthus roseus* (L.) G. Don 21, 59, 66, **185**, 426  
 – cv. *Albus* 188  
*Catharanthus trichophyllus* (Baker) Pichon 190  
*Cedrus deodara* Loud. 255  
*Centella* L. 190, 193  
*Centella asiatica* (L.) Urb. 101, **190**, 434  
*Cephaloscheffera blancoi* (Merr.) Merr. 435  
*Cerantonia siliqua* L. 473  
*Ceratostigma* Bunge 411  
*Cerbera odollam* Gaertner 127  
*Chamaecrista* Moench 182, 444  
*Chamaecrista mimosoides* (L.) Greene 443  
*Chamaesyce* Gray 265  
*Chamaesyce atoto* (J.G. Forster) Croizat 267  
*Chamaesyce hirta* (L.) Millsp. 268  
*Chamaesyce pilulifera* (L.) Small 268  
*Chamaesyce prostrata* (Aiton) Small 270  
*Chamaesyce thymifolia* (L.) Millsp. 271  
*Chamomilla recutita* (L.) Rauschert 79  
 Chenopodiaceae 42  
*Chenopodium* L. 197  
*Chenopodium album* L. 195  
*Chenopodium ambrosioides* L. 79, **194**  
 – var. *ambrosioides* 197  
 – var. *anthelminticum* (L.) A. Gray 197, 198  
*Chenopodium anthelminticum* L. 197  
*Chenopodium chilense* Schrader 197  
*Chenopodium graveolens* Willd. 197  
*Chenopodium multifidum* L. 197  
*Chenopodium scoparia* L. 195  
*Chlaenandra* Miq. 481  
*Chondrodendron* Ruiz & Pav. 468  
*Chondrodendron tomentosum* Ruiz & Pavón 206, 220, 468  
*Choripetalum benthamii* Hance 257  
*Chromolaena odorata* (L.) R. King & H. Robinson. 91  
*Chrysanthemum* L. 95  
*Cicca* L. 384  
*Cicca acida* (L.) Merr. 386  
*Cichorium intybus* L. 477  
*Cinchona* L. 21, 56, 57, 59, 143, 161, **198**, 469  
*Cinchona calisaya* Wedd. 204  
*Cinchona cordifolia* Mutis 204  
*Cinchona ledgeriana* Moens ex Trimen 201, 204  
*Cinchona officinalis* L. **198**, **204**  
*Cinchona pubescens* Vahl **198**, **204**  
*Cinchona succirubra* Pav. ex Klotzsch 201, 204  
*Cinnamomum camphora* (L.) J.S. Presl 156  
*Cinnamomum porrectum* (Roxb.) Kosterm. 115  
*Cissampelos* L. 205, 465  
*Cissampelos owariensis* P. Beauv. ex DC. 207  
*Cissampelos pareira* L. **205**, 219, 220  
 – var. *hirsuta* (Buch.-Ham. ex DC.) Forman 205, 207  
 – var. *orbiculata* (DC.) Miq. 207  
 – var. *pareira* 207  
 – var. *peltata* Scheff. 207  
 – var. *typica* Diels 207  
 ×*Citrofortunella microcarpa* (Bunge) Wijnands 416  
*Clerodendrum* L. 369, 402

- Clusiaceae 304  
 Codariocalyx Hassk. 244  
 Coffea arabica L. 112  
 Cola nitida (Vent.) Schott & Endl. 258  
 Colchicaceae 290  
 Colchicum autumnale L. 193, 290  
 Colchinaceae 291  
 Coldenia L. 180  
 Coleus Lour. 405  
 Coleus amboinicus Lour. 407  
 Coleus aromaticus Benth. 407  
 Coleus atropurpureus Benth. 408  
 Coleus barbatus (Andrews) Benth. 408  
 Coleus blumei Benth. 408  
 Coleus carnosa Hassk. 407  
 Coleus forskohlii (Willd.) Briq. 404, 408  
 Coleus scutellarioides (L.) Benth. 408  
 Colocasia esculenta (L.) Schott 403  
 Combretum Loefl. 422  
 Compositae 45, 56, 77, 88, 95, 156, 251, 292, 369, 476, 477, 494  
 Conyza cinerea L. 496  
 Coptis Salisb. 465  
 Coptis teeta Wallich 131  
 Corchorus capsularis L. 127  
 Corchorus olitorius L. 127  
 Cordia L. 180  
 Cordia alliodora (Ruiz & Pavon) Oken 403  
 Cordiaceae 180  
 Cordioideae 180  
 Coreopsis L. 151, 152  
 Coreopsis biternata Lour. 153  
 Cornaceae 399  
 Cornus florida L. 492  
 Coronilleae 244  
 Corydalis DC. 465  
 Coscinium fenestratum (Gaertner) Colebr. 131  
 Cotyledon crenata (Andrews) Vent. 337  
 Cotyledon laciniata L. 338  
 Cotyledon pinnata Lamk 166  
 Crassulaceae 164  
 Cratoxyleae 304  
 Cratoxylum Blume 304  
 Crepis L. 478  
 Crotalaria L. 292  
 Crotalaria trichotoma Bojer 201  
 Croton tiglium L. 237, 481  
 Crotonoideae 323  
 Cucurbitaceae 75  
 Cucurbitoidae 356, 486  
 Curcas indica A. Rich. 324  
 Curcas purgans Medik. 324  
 Curculigo Gaertner 209  
 Curculigo ensifolia R. Br. 209  
 Curculigo latifolia Dryander 208, 209  
 Curculigo orchoides Gaertner 207  
 Curcuma L. 138, **210**  
 – subgenus Curcuma 213  
 – subgenus Paracurcuma 213  
 Curcuma aeruginosa Roxb. 211, 213, 215, **214**  
 Curcuma aurantiaca v. Zijp 210, 213, **215**  
 Curcuma domestica Valetton 215  
 Curcuma euchroma Valetton **215**  
 Curcuma heyneana Valetton & v. Zijp **215**  
 Curcuma longa L. 23, 83, **210, 215**  
 Curcuma mangga Valetton & v. Zijp 214, **216**  
 Curcuma pallida Lour. 218  
 Curcuma petiolata Roxb. 210, **217**  
 Curcuma purpurascens Blume 213, **217**  
 Curcuma soloensis Valetton **217**  
 Curcuma xanthorrhiza Roxb. **210, 217**, 368  
 Curcuma zedoaria (Christm.) Roscoe **210, 218**  
 Curcuma zerumbet Roxb. 218  
 Cuscuta L. 189  
 Cuscuta europaea L. 174  
 Cyclea Arn. ex Wight **219**, 464, 465, 481  
 Cyclea barbata Miers 143, 200, 206, **219, 221**, 466  
 Cyclea kinabaluensis Forman 221  
 Cyclea laxiflora Miers **219, 222**  
 Cyclea peltata auct. non (Lamk) Hook.f. & Thomson 221  
 Cymbopogon Spreng. 224  
 Cymbopogon nardus Rendle 23  
 Cynanchum caudatum Maxim. 394  
 Cypereae 224  
 Cyperoideae 224  
 Cyperus L. **222**  
 – subgenus Kyllinga 224  
 – subgenus Pycneus 224  
 Cyperus brevifolius (Rottb.) Hassk. 222, 224, **225**  
 Cyperus cyperoides (L.) O. Kuntze **226**  
 Cyperus diffusus Vahl **226**  
 Cyperus halpan L. **226**  
 – subsp. halpan 227  
 – subsp. juncooides (Lamk) Kük. 227  
 Cyperus iria L. 222, 223, 224, **227**  
 Cyperus kyllingia Endl. **222, 227**  
 Cyperus malaccensis Lamk **228**  
 Cyperus retzii Nees 228  
 Cyperus rotundus L. **222, 228**  
 – subsp. retzii (Nees) Kük. 228  
 – subsp. rotundus 228, 229  
 Cyperus stoloniferus Retz. **229**  
 Cyrtosiphona sumatrana (Jack) Miq. 430  
 Cyrtosiphonia madurensis Teijsm. & Binnend. 430  
 Dalbergia sissoo Roxb. ex DC. 230

- Dammaropsis kingiana* Warb. 283  
*Datura* L. 66, **229**  
 – sect. *Brugmansia* (Persoon) Bernh. 232  
*Datura fastuosa* L. 233, 234  
*Datura innoxia* Miller 232  
*Datura metel* L. 24, 57, **229**, **233**, 269  
*Datura stramonium* L. 59, 229–233, **234**  
*Datura tatula* L. 234  
*Deguelia* Aubl. 237  
*Deguelia malaccensis* (Benth.) Blake 241  
*Dendrolobium* (Wight & Arn.) Benth. 244  
*Derris* Lour. 128, **234**, 481  
 – section *Brachypterum* (Wight & Arn.) Benth. 237  
 – section *Derris* 237  
 – section *Dipteroderris* Benth. 237  
 – section *Paraderris* Miq. 237  
*Derris acuminata* Benth. 234, 237  
*Derris amoena* Benth. 237  
*Derris cauliflora* Pulle 240  
*Derris elegans* Graham ex Benth. 234, 235, 237, **240**  
 – var. *elegans* 240  
 – var. *gracillima* (Hemsley) Verdc. 240  
 – var. *vestita* (Baker) Prain 240  
*Derris elliptica* (Wallich) Benth. 23, 90, **234**, **240**  
*Derris ferruginea* (Roxb.) Benth. 237  
*Derris heterophylla* (Willd.) Backer ex K. Heyne 241  
*Derris malaccensis* (Benth.) Prain **234**, **241**  
*Derris momiensis* Kanehira & Hatusima 240  
*Derris montana* Benth. 237  
*Derris multiflora* Benth. 237  
*Derris papuana* Pulle 240  
*Derris philippinensis* Merr. 237  
*Derris polyantha* Perk. 237  
*Derris polyphylla* (Miq.) Benth. 241  
*Derris pubipetala* Miq. 237  
*Derris robusta* (Roxb. ex DC.) Benth. 234–237, **241**  
*Derris rufula* Lauterb. & K. Schumann 240  
*Derris scandens* (Roxb.) Benth. 234–237, **241**  
*Derris trifoliata* Lour. **234**, **241**  
*Derris uliginosa* (Willd.) Benth. 241  
*Derris vestita* Baker 240  
*Desmodiinae* 244  
*Desmodium* Desv. 201, 224, **242**, 369  
*Desmodium adscendens* (Sw.) DC. **242**, **245**  
*Desmodium ancistrotrichum* K. Schumann & Lauterb. 247  
*Desmodium auricomum* Grah. ex Benth. 242  
*Desmodium canum* Schinz & Thell. 242  
*Desmodium capitatum* Burm.f. 248  
*Desmodium caudatum* (Thunb. ex Murray) DC. 242, 243  
*Desmodium diffusum* DC. 242, 244, **245**  
*Desmodium gangeticum* (L.) DC. **242**, **245**  
*Desmodium heterocarpon* (L.) DC. 242  
 – subsp. *angustifolium* H. Ohashi 242  
*Desmodium heterophyllum* (Willd.) DC. 242  
*Desmodium incanum* DC. 242, 243  
*Desmodium lasiocarpum* (P. Beauv.) DC. 249  
*Desmodium latifolium* (Roxb. ex Ker.) DC. 249  
*Desmodium laxiflorum* auct. non DC. 245  
*Desmodium laxiflorum* DC.  
 – subsp. *parvifolium* H. Ohashi & Chen 245  
*Desmodium microphyllum* (Thunb. ex Murray) DC. 242, 244, **246**  
*Desmodium ormocarpoides* DC. 242, **247**  
*Desmodium oxalidifolium* G. Don 245  
*Desmodium parvifolium* Blanco 249  
*Desmodium parvifolium* DC. 246  
*Desmodium recurvatum* (Roxb.) Wight & Arn. 245  
*Desmodium renifolium* Schindler 242  
*Desmodium repandum* (Vahl) DC. 242, 244, **247**  
*Desmodium retroflexum* (L.) DC. 248  
*Desmodium scalpe* DC. 247  
*Desmodium sequax* Wallich 242, 244, **247**  
*Desmodium sinuatum* (Miq.) Blume ex Baker 247  
*Desmodium strigillosum* Schindler **248**  
*Desmodium styracifolium* (Osbeck) Merr. **242**, **248**  
*Desmodium triflorum* (L.) DC. **242**, **249**  
*Desmodium trifoliastrum* Miq. 245  
*Desmodium velutinum* (Willd.) DC. 242, 244, **249**  
 – subsp. *longibracteatum* (Schindler) H. Ohashi 250  
 – subsp. *velutinum* 250  
*Diasperus pulcher* (Wallich ex Muell. Arg.) Kuntze 390  
*Dicerma* DC. 244  
*Dichroa* Lour. 162  
*Dichroa febrifuga* Lour. 143, 200  
*Digitalis* L. 47, 127  
*Dimocarpus longan* Lour. 482  
*Dioscorea* L. 21, 47, 448, 455, 456  
*Dissolena verticillata* Lour. 431  
*Droseraceae* 41  
*Duboisia* R. Br. 231, 233  
*Ebenaceae* 41  
*Echinochloa colona* (L.) Link. 344  
*Echinochloa crus-galli* (L.) P. Beauv. 309  
*Eclipta prostrata* (L.) L. 105  
*Ehretia* P. Browne 180  
*Ehretia buxifolia* Roxb. 178  
*Ehretia microphylla* Lamk 178



- Ehretioideae 180  
 Elephantopus L. **250**, 369, 495  
 – subgenus *Pseudelephantopus* (Rohr) C. Jeffrey 251  
 Elephantopus mollis Kunth **250**, **252**  
 Elephantopus scaber L. 224, **250**, **253**  
 Elephantopus spicatus Juss. ex Aublet 250–252, **253**  
 Elephantopus tomentosus auct. non L. 252  
 Elephantopus tomentosus L. 250  
 Elettaria Maton 114  
 Elettaria cardamomum (L.) Maton 114  
 Embelia Burm.f. **254**  
 Embelia coriacea Wallich ex A.DC. 254  
 Embelia garciniifolia Wallich ex Miq. 257  
 Embelia laeta (L.) Mez. 254, **257**  
 Embelia obovata Hemsl. 257  
 Embelia philippinensis A.DC. 254  
 Embelia ribes Burm.f. **254**, **257**  
 Embelia robusta C.B. Clarke non Roxb. 258  
 Embelia robusta Roxb. 254, 256, **258**  
 Embelia schimperi Vatke 254  
 Embelia tsjeriam-cottam (Roemer & Schultes) A. DC. 254, 256, **258**  
 Emblica Gaertner 384  
 Emblica arborea Raf. 388  
 Emblica grandis Gaertner 388  
 Emblica officinalis Gaertner 388  
 Ercilla A. Juss. 394  
 Ericaceae 41  
 Ervatamia (A. DC.) Stapf 426  
 Erythroxyloaceae 260  
 Erythroxyllum P. Browne **258**  
 Erythroxyllum bolivianum Burck 262  
 Erythroxyllum coca Lamk 59, **258**, **262**  
 – var. coca 259–262  
 – var. ipadu Plowman 259–262  
 – var. novogranatense Morris 262  
 – var. spruceanum Burck 262  
 Erythroxyllum cuneatum (Miq.) Kurz 259  
 Erythroxyllum ecarinatum Burck 259  
 Erythroxyllum novogranatense (Morris) Hieron. 59, **258**, **262**  
 – var. novogranatense 259, 260, 262  
 – var. truxillense (Rusby) Plowman 259, 260, 262  
 Erythroxyllum peruvianum Prescott 262  
 Erythroxyllum truxillense Rusby 262  
 Eucalyptus L'Hér. 235, 473  
 Eucalyptus macrorhyncha F. v. Mueller ex Benth. 473  
 Euphorbia L. **263**  
 – subgenus *Chamaesyce* 265  
 – subgenus *Euphorbia* 265  
 – subgenus *Poinsettia* 265  
 Euphorbia antiquorum L. **263**, **266**, 271  
 Euphorbia atoto J.G. Forster 265, **267**  
 Euphorbia barnhartii Croizat 263, 265, **267**  
 Euphorbia buxoides Radcl.-Sm. 265  
 Euphorbia cyathophora Murray 265, **267**  
 Euphorbia halophila Miq. 267  
 Euphorbia heterophylla L. 263, 265, **268**  
 – var. cyathophora (Murray) Griseb. 267  
 Euphorbia hirta L. **263**, **268**  
 Euphorbia laevis Poir. 267  
 Euphorbia ligularia Roxb. 269  
 Euphorbia media N.E.Br. 271  
 Euphorbia neriifolia L. 263, **269**, 271  
 Euphorbia pilulifera L. 268  
 Euphorbia plumerioides Teijsm. ex Hassk. 265, **270**  
 Euphorbia prostrata Aiton 263–265, **270**  
 Euphorbia prunifolia Jacq. 268  
 Euphorbia rhipsaloides Lem. 271  
 Euphorbia ridleyi Croizat 270  
 Euphorbia scoparia N.E. Br. 271  
 Euphorbia serrulata Reinw. ex Blume non Thuill. 265  
 Euphorbia synadenium Ridley **270**  
 Euphorbia taiwaniana Ying 268  
 Euphorbia thymifolia L. **263**, **270**, **271**  
 Euphorbia tirucalli L. 263–266, **271**  
 Euphorbia trigona Roxb. non Miller 267  
 Euphorbia vachellii Hook. & Arn. 265  
 Euphorbiaceae 35, 41, 382  
 Eurycoma Jack 162, **272**  
 Eurycoma apiculata A.W. Bennett **272**, **274**  
 Eurycoma longifolia Jack **272**, **274**  
 – subsp. eglanulosa (Merr.) Nootboom 274  
 Fagopyrum esculentum Moench 473  
 Fatoua Gaudich. 275  
 Fatoua japonica (Thunb. ex Murray) Blume 275  
 Fatoua pilosa Gaudich. 275, 276  
 Fatoua villosa (Thunb. ex Murray) Nakai **275**  
 Fibraurea Lour. 481  
 Fibraurea tinctoria Lour. 131  
 Ficus L. **277**  
 Ficus adenosperma Miq. **280**  
 Ficus aechmophylla Summerh. 285  
 Ficus ampelas Burm.f. **280**  
 Ficus aurantiacea Griffith **281**  
 Ficus bauerleni King **281**  
 Ficus banyana Oken 281  
 Ficus barnesii Merr. 282  
 Ficus benghalensis L. **277**, **281**  
 Ficus benjamina L. 277, 278, 280  
 Ficus blepharosepala Warb. 280  
 Ficus botryocarpa Miq. **282**

- Ficus brassii* Summerh. 285  
*Ficus cairnsii* Warb. 284  
*Ficus callicarpa* Miq. 281  
*Ficus calopilina* Diels 282  
*Ficus casearia* F. v. Mueller ex Benth. 288  
*Ficus caudata* Stokes 286  
*Ficus cerasiformis* Desf. 285  
*Ficus conciliorum* Oken 287  
*Ficus copiosa* Steud. 282  
*Ficus cordifolia* Roxb. 287  
*Ficus crininervia* Miq. 287  
*Ficus damit* Gagnep. 287  
*Ficus dammaropsis* Diels 283  
*Ficus deltoidea* Jack 278, 279, 283  
*Ficus diversifolia* Blume 283  
*Ficus du Lauterb. & K. Schumann* 284  
*Ficus elastica* Roxb. ex Hornem. 277  
*Ficus eulampra* K. Schumann 289  
*Ficus formosa* Summerh. 288  
*Ficus grandifolia* Wallich ex Miq. 285  
*Ficus grandis* King 285  
*Ficus hauili* Blanco 288  
*Ficus hispida* L.f. 277, 283  
*Ficus hollrungii* Lauterb. & K. Schumann 281  
*Ficus hypoglauca* Lauterb. & K. Schumann 285  
*Ficus indica* L. 281  
*Ficus kalingaensis* Merr. 286  
*Ficus kaukauensis* Hayata 288  
*Ficus kingiana* Hemsley 280  
*Ficus krausseana* Rechinger 282  
*Ficus lasiophylla* Link 281  
*Ficus laurentina* Diels 281  
*Ficus letaqui* Lév. & Van. 283  
*Ficus linearifolia* Elmer 282  
*Ficus longipedunculata* Rechinger 282  
*Ficus lutescens* Desf. 283  
*Ficus magnifolia* F. v. Mueller 282  
*Ficus mangiferifolia* Lauterb. & K. Schumann 285  
*Ficus megacarpa* Merr. 281  
*Ficus mespiloides* King 281  
*Ficus microcarpa* L.f. 279, 284  
*Ficus minahassae* (Teysm. & de Vriese) Miq. 277  
*Ficus mindorensis* Merr. 282  
*Ficus motleyana* Miq. 283  
*Ficus myriocarpa* Miq. 286  
*Ficus nasuta* Summerh. 284  
*Ficus nodosa* Teijsm. & Binnend. 284  
*Ficus nubigena* Diels 289  
*Ficus ovalifolia* Ridley 286  
*Ficus pachyrrachis* Lauterb. & K. Schumann 278, 285  
*Ficus pachystemon* Warb. 285  
*Ficus pachythyrssa* Diels 285  
*Ficus parietalis* Blume 285  
*Ficus pauper* King 280  
*Ficus peepul* Griffith 286  
*Ficus poilanei* Gagnep. 283  
*Ficus pomifera* Kurz 281  
*Ficus pumila* L. 280, 285  
*Ficus pungens* Reinw. ex Blume 286  
*Ficus racemosa* L. 277-279  
*Ficus ramentacea* Roxb. 287  
*Ficus ramosii* Merr. ex Sata 287  
*Ficus religiosa* L. 277, 286  
*Ficus repens* Hort.  
 – var. *lutchuensis* Koidz. 285  
*Ficus retusa* auct. non L.f. 284  
*Ficus retusa* L.f. 277  
*Ficus retusifolmis* Lév. 284  
*Ficus rhodocarpa* Summerh. 289  
*Ficus rumphii* Blume 277, 287  
*Ficus sagittata* J. König ex Vahl 287  
*Ficus scandens* Lamk 285  
*Ficus septica* Burm.f. 277, 288  
*Ficus setistyla* Warb. 282  
*Ficus soronensis* King 280  
*Ficus stipulata* Thunb. 285  
*Ficus stoechotricha* Diels 288  
*Ficus subcuneata* Miq. 288  
*Ficus sublimbata* Corner 288  
*Ficus superstitiosa* Link 286  
*Ficus turbinata* Ridley 280  
*Ficus wassa* Roxb. 289  
*Fimbristylis* Vahl 224  
 Flacourtiaceae 300  
*Garrya Douglas* ex Lindl. 399  
 Garryaceae 399  
*Gasteria Duval* 102  
*Gemmingia chinensis* (L.) O. Kuntze 148  
*Gendarussa* Nees 328  
*Gendarussa vulgaris* Nees 330  
*Gliricidia sepium* (Jacq.) Kunth ex Walp. 75  
*Globba* L. 114  
*Glochidion* J.R. Forster & J.G. Forster 384  
*Gloriosa* L. 291  
*Gloriosa rothschildiana* O'Brien 289, 291  
*Gloriosa simplex* auct. 289  
*Gloriosa superba* L. 289  
*Glycyrrhiza glabra* L. 75  
*Gmelina* L. 414  
*Gomphrena* L. 106  
*Gomphrena ficoidea* L. 108  
*Goniothalamus* (Blume) Hook.f. & Thomson 34  
 Gramineae 35  
*Grenacheria* Mez 256  
 Guttiferae 304  
*Gynocardia odorata* R.Br. 301

- Haplochorema* K. Schumann 332  
*Haworthia* Duval 102  
*Hedychieae* 213, 332  
*Hedyotis* L. 365  
*Hedyotis affinis* Roemer & Schultes 366  
*Hedyotis biflora* (L.) Lamk 366  
*Hedyotis brachypoda* (DC.) Sivar. & Biju 366  
*Hedyotis capitellata* Wallich ex G. Don 366  
*Hedyotis corymbosa* (L.) Lamk 367  
*Hedyotis dichotoma* Heyne ex Roth 366  
*Hedyotis diffusa* auct. non Willd. 366  
*Hedyotis herbacea* L. 367  
*Hedysarum adhaerens* Poir. 247  
*Hedysarum adscendens* Sw. 245  
*Hedysarum gangeticum* L. 245  
*Hedysarum microphyllum* Thunb. ex Murray 246  
*Hedysarum ormocarpum* Desv. ex Poir. 247  
*Hedysarum repandum* Vahl 247  
*Hedysarum styracifolium* Osbeck 248  
*Hedysarum triflorum* L. 249  
*Hegnere* Schindl. 244  
*Heliantheae* 152  
*Heliotropioideae* 293  
*Heliotropium* L. **292**  
*Heliotropium arborescens* L. 292  
*Heliotropium curassavicum* L. 292, 293, **294**  
*Heliotropium cyrtostachyum* Miq. 295  
*Heliotropium indicum* L. 224, **292, 294**  
*Heliotropium marifolium* Retz. 295  
*Heliotropium ovalifolium* Forssk. 292, **295**  
*Heliotropium peruvianum* L. 292  
*Heliotropium scabrum* Retz. 293, **295**  
*Heptapleurum* Gaertner 434  
*Heptapleurum caudatum* S. Vidal 435  
*Heptapleurum cumingii* Seem. 436  
*Heptapleurum insularum* Seem. 438  
*Heptapleurum junghuhniana* (Miq.) Seem. 437  
*Heterosmilax* Kunth 448  
*Hevea brasiliensis* (Willd. ex A.L. Juss.) Muell.-Arg. 115  
*Hibiscus* L. 224  
*Hibiscus sabdariffa* L. 394  
*Holarrhena* R. Br. **296**  
*Holarrhena antidysenterica* (L.) Wallich ex A.DC. 297, 298  
*Holarrhena curtisii* King & Gamble 296, 297, **298**  
*Holarrhena densiflora* Ridley 298  
*Holarrhena latifolia* Ridley 298  
*Holarrhena macrocarpa* (Hassk.) Villar 298  
*Holarrhena malaccensis* Wight 298  
*Holarrhena mitis* (Vahl) R.Br. 296, 297  
*Holarrhena pubescens* Wallich ex G. Don **296, 298**  
*Holarrhena pulcherrima* Ridley 298  
*Huperzia serrata* (Thunb. ex Murray) Trevisan 51  
*Hydnocarpus* Gaertner **299**  
*Hydnocarpus alcalae* C.DC. **299, 301**  
*Hydnocarpus alpina* Wight  
– var. *elongata* Boerl. 302  
– var. *macrocarpa* Boerl. 302  
*Hydnocarpus anthelmintica* Pierre ex Lanessan **299, 302**  
*Hydnocarpus cauliflora* Merr. 300  
*Hydnocarpus heterophylla* Blume 301  
*Hydnocarpus kunstleri* 301  
*Hydnocarpus kurzii* (King) Warb. **299, 302**  
– subsp. *australis* Sleum. 303  
– subsp. *kurzii* 303  
*Hydnocarpus laurifolia* (Dennst.) Sleumer 299, 300  
*Hydnocarpus pentandrus* (Ham.) Oken 299  
*Hydnocarpus subfalcata* Merr. 300  
*Hydnocarpus venenata* Gaertner 299  
*Hydnocarpus wightiana* Blume 299  
*Hydnocarpus woodii* Merr. 300, 301  
*Hydrocotyle* L. 190, 193  
*Hydrocotyle asiatica* L. 190  
*Hyoscyamus niger* L. 231  
*Hypericaceae* 304  
*Hypericeae* 304  
*Hypericoideae* 304  
*Hypericum* L. **303**  
*Hypericum ascyron* L. 105, 303  
*Hypericum chinense* L. 306  
*Hypericum chinense* L. non Osbeck 306  
*Hypericum gramineum* G. Forster 303, **305**  
*Hypericum habbemense* A.C. Smith 307  
*Hypericum helwigii* Laut. 307  
*Hypericum involutum* (Labill.) Choisy 305  
*Hypericum japonicum* Thunb. ex Murray **303, 306**  
*Hypericum monogynum* L. **303, 306**  
*Hypericum mutilum* Maxim. 306  
*Hypericum papuanum* Ridley 303, 304, **307**  
*Hypericum patulum* Thunb. ex Murray 303, 304  
– var. *uralum* (Buch.-Ham. ex D. Don) Koehne 307  
*Hypericum perforatum* L. 303, 304  
*Hypericum pusillum* Choisy 306  
*Hypericum sampsonii* Hance 303  
*Hypericum uralum* Buch.-Ham. ex D. Don 303, 304, **307**  
*Hypoxidaceae* 209  
*Illecebrum lanatum* (L.) L. 87  
*Illicium verum* Hook.f. 83, 237  
*Imperata* Cirillo 159, 209, **307**  
– section *Eriopogon* Endl. 308

- section Imperata 308
- Imperata arundinacea Cirillo 310
- Imperata conferta (J.S. Presl) Ohwi **307, 309**
- Imperata cylindrica (L.) Raeuschel 221, **307, 310**
- Imperata exaltata (Roxb.) Brongn. 309
- Inuleae 157
- Iphigenia Kunth 290
- Ipomoea batatas (L.) Lamk 484
- Irideae 149
- Iris L. 148
- Iris dichotoma Pall. 149
- Iris fulva Ker Gawler 149
- Iris japonica Thunb. 148
- Ixia chinensis L. 148
- Ixora L. **311**
- Ixora amboinica (Blume) DC. 315
- Ixora amoena Wallich ex G. Don 314
- Ixora chinensis Lamk **311, 313**
- Ixora coccinea L. **311, 313**
- Ixora crassifolia Ridley 314
- Ixora fulgens auct. non Roxb. 315
- Ixora grandiflora Loddiges 313
- Ixora grandifolia Zoll. & Moritz **312, 314**
- Ixora javanica (Blume) DC. **311, 314**
- Ixora lobbii Loudon **312, 315**
- Ixora longifolia J.E. Smith **311, 315**
- Ixora montana Lour. 313
- Ixora nigricans R.Br. ex Wight & Arn. **311, 315**
- Ixora philippinensis Merr. 311
- Ixora ridleyi Bremek. 314
- Ixora stricta Roxb. 313
- Jasmineae 317
- Jasminoideae 317
- Jasminum L. 59, **315**
- Jasminum aemulum R.Br. 318
- Jasminum bifarium Wallich ex G. Don 318
- Jasminum elongatum (Bergius) Willd. **315, 318**
- Jasminum grandiflorum L. 316, 317
- Jasminum multiflorum (Burm.f.) Andr. **315, 319**
- Jasminum multiflorum sensu Bakhuizen f. non (Burm. f.) Andr. 318
- Jasminum officinale L. 317
- Jasminum pubescens (Retz.) Willd. 319
- Jasminum pubescens sensu Backer non (Retz.) Willd. 318
- Jasminum sambac (L.) Aiton **315, 319**
- Jasminum subtriplinerve Blume 316, **320**
- Jatropha L. **320**
  - subgenus Curcas 323
  - subgenus Jatropha 323
- Jatropha afrocurcas Pax 324
- Jatropha curcas L. **320, 324**
- Jatropha elegans (Pohl) Klotzsch 325
- Jatropha gossypifolia L. **320, 325**
  - var. elegans (Pohl) Muell. Arg. 326
  - var. gossypifolia 326
- Jatropha janipha Blanco 326
- Jatropha multifida L. **320, 326**
- Jatropheae 323
- Joliffieae 356
- Juglandaceae 41
- Justicia L. **327**
- Justicia adhatoda L. **327, 329**
- Justicia cydoniifolia (Nees) Lindau 328
- Justicia gendarussa Burm.f. **327, 330**
- Justicia japonica Thunb. 331
- Justicia pectoralis Jacq. 328
- Justicia procumbens L. **327, 331**
- Justicia simplex D. Don 331
- Justicia spicigera Schltld. 328
- Justicieae 432
- Kaempferia L. **331**
- Kaempferia angustifolia Roscoe 331, 332, **334**
- Kaempferia galanga L. **331, 334**
- Kaempferia gilbertii W. Bull 334
- Kaempferia longa Jacq. 335
- Kaempferia rotunda L. **331, 335**
- Kaempferia roxburghiana Schult. 334
- Kaempferia undulata Teijsm. & Binnend. non Link 334
- Kalanchoe Adans. 165, **335**
- Kalanchoe blossfeldiana Poelln. 336
- Kalanchoe brasiliensis Camb. 336
- Kalanchoe ceratophylla Haw. 165, **335, 337**
- Kalanchoe craibii Raymond-Hamet 338
- Kalanchoe crenata (Andrews) Haw. 165, **335, 337**
- Kalanchoe integra auct. non (Medic.) O. Kuntze 336, 337
- Kalanchoe laciniata (L.) DC. 335, 336, **338**
- Kalanchoe laciniata auct. non (L.) DC. 337
- Kalanchoe lanceolata (Forssk.) Pers. 336
- Kalanchoe pinnata (Lamk) Pers. 166
- Kalanchoe prolifera (Bowie) Hamet 167
- Kalanchoe spathulata DC. 337
- Kitchingia Baker 165
- Kochia scoparia (L.) Schrader 195
- Kopsia Blume 426
- Kyllinga brevifolia Rottb. 225
- Kyllinga monocephala Rottb. 227
- Labiatae 57, 346, 405, 439, 440
- Laburnum anagyroides Medik. 462
- Lactuceae 478
- Laggera Sch. Bip. ex Benth. 157
- Lagurus cylindricus L. 310
- Lamiales 45
- Lantana L. 237, **338, 481**
- Lantana aculeata L. 341
- Lantana camara L. **338, 341**

- Lantana trifolia* L. **338, 342**  
*Lathyrus sativus* L. 35  
*Lavandula* L. 346  
*Lawsonia inermis* L. 41  
 Leguminosae 35, 36, 40, 41, 73  
*Leonurus* L. 224  
*Leucaena* Benth. 35  
*Leucaena leucocephala* (Lamk) de Wit 75, 201, 351  
*Ligusticum acutilobum* Siebold & Zucc. 123  
 Liliaceae 36, 41, 102, 209, 291, 448  
*Limacia* Lour. 131  
*Limonium* Miller 411  
 Linaceae 260  
*Linum usitatissimum* L. 399  
*Lippia* L. 340  
*Lippia rehmanii* Pears. 340  
*Liquidambar* L. 448  
*Lochnera rosea* (L.) Rcheichenb. ex Endl. 185  
 Loganiaceae 469  
*Lonchocarpus* Kunth 236, 237, 481  
*Ludwigia* L. 369  
*Luffa* Miller 75  
*Lycopodium serratum* Thunb. ex Murray 51  
 Lythraceae 41  
*Macaranga Thouars* 384  
*Macaranga triloba* (Blume) Muell.-Arg. 452  
 Malpighiaceae 260  
*Malva sylvestris* L. 399  
 Malvaceae 399  
*Mangifera indica* L. 115  
*Mariscus sieberianus* Nees ex Clarke 226  
*Marrubium* L. 405  
*Matricaria chamomilla* auct. non L. 79  
*Matricaria recutita* L. 79  
*Medicago sativa* L. 418  
*Meibomia gangetica* (L.) O. Kuntze 245  
*Meibomia repanda* (Vahl) O. Kuntze 247  
*Melia azedarach* L. 83, 237, 481  
*Melochia* L. 342, 344  
*Melochia concatenata* L. 342  
*Melochia corchorifolia* L. **342**  
*Melochia pyramidata* L. 343  
*Melochia tomentosa* L. 343  
*Melochia umbellata* (Houtt.) Stapf 343  
 Menispermaceae 56, 129, 131, 135, 206, 219, 220, 465, 466, 481  
 Menispermeae 465  
*Menodora* Bonpl. 317  
*Mentha* L. 59, 344, 345, 347-349  
*Mentha aquatica* L. 347  
*Mentha arvensis* L. 59, **344**, 369  
 - cv. Jombang 348  
 - subsp. *haplocalyx* (Briq.) Briq. 347  
 - var. *arvensis* 344, 347  
 - var. *javanica* (Blume) Hook.f. 344, 345, 347  
 - var. *piperascens* Malinv. ex Holmes 345, 347  
*Mentha canadensis* L. 347  
*Mentha xcordifolia* Opiz ex Fresen 346, 347, 348  
*Mentha haplocalyx* Briq. 347  
*Mentha javanica* Blume 344  
*Mentha longifolia* L. 347  
*Mentha xpiperita* L. 57, 347, 348  
*Mentha pulegium* L. 347  
*Mentha xrotundifolia* (L.) Huds. 347  
*Mentha spicata* L. 347  
*Mentha suaveolens* Ehrh. 347  
*Mentha xvillosa* Huds. 347  
*Millettia* Wight & Arn. 236, 237  
 Millettieae 237  
*Mimosa* L. 351  
*Mimosa pudica* L. **349**  
 - var. *hispida* Brenan 351  
 - var. *pudica* 351  
 - var. *tetrandra* (Humb. & Bonpl. ex Willd.) DC. 351  
 - var. *unijuga* (Duchass. & Walp.) Griseb. 352  
*Mimosa tetrandra* Humb. & Bonpl. ex Willd. 352  
*Mimosa unijuga* Duchass. & Walp. 352  
 Mimoseae 351  
 Mimosoideae 351  
 Molluginaceae 42  
*Momordica* L. 75, **353**, 486, 491  
*Momordica charantia* L. **353, 357**  
*Momordica chinensis* Spreng. 357  
*Momordica cochinchinensis* (Lour.) Spreng. **353, 358**  
*Momordica elegans* Salisb. 357  
*Momordica indica* L. 357  
*Momordica meloniflora* Hand.-Mazz. 358  
*Momordica mixta* Roxb. 358  
*Monochoria vaginalis* (N.L. Burmann) Kunth 309  
*Morus* L. 328, **359**, 372  
*Morus acidosa* Griffith 363  
*Morus alba* L. **359, 362**  
*Morus atropurpurea* Roxb. 362  
*Morus australis* Poir. 359, 360, **363**  
*Morus bombycis* Koidz. 360  
*Morus cathayana* Hemsl. 359  
*Morus cavaleriei* H. Lév. 363  
*Morus indica* L. 362  
*Morus inusitata* H. Lév. 363  
*Morus laciniata* Miller 363  
*Morus latifolia* Poir. 359  
*Morus macroura* Miq. 359  
*Morus morettiana* Jacq. ex Burr. 362  
*Morus nigra* L. **359, 363**  
*Morus scabra* Moretti 363

- Muricia cochinchinensis* Lour. 358  
*Musa textilis* Née 252  
*Myrica esculenta* Buch.-Ham 237  
*Myristica fragrans* Houtt. 84  
*Nerium oleander* L. 127  
*Nicotiana tabacum* L. 237, 481  
*Norysca urala* (Buch.-Ham. ex D. Don) K. Koch 307  
*Nyctanthes sambac* L. 319  
*Nypa Steck* 466  
*Ocimum* L. 83, 346  
*Oldenlandia* L. **364**  
*Oldenlandia affinis* (Roemer & Schultes) DC. 364, **366**  
*Oldenlandia biflora* L. **366**  
*Oldenlandia brachypoda* DC. **364, 366**  
*Oldenlandia capitellata* (Wallich ex G. Don) O. Kuntze **364, 366**  
*Oldenlandia corymbosa* L. **364, 367**  
*Oldenlandia dichotoma* (Heyne ex Roth) Hook.f. 366  
*Oldenlandia diffusa* (Willd.) Roxb. 365  
*Oldenlandia diffusa* auct. non (Willd.) Roxb. 366  
*Oldenlandia herbacea* (L.) Roxb. 364, **367**  
*Oldenlandia recurva* (Korth.) Miq. 366  
*Oldenlandia umbellata* L. 364, 365  
Oleaceae 317  
Onagraceae 34  
*Ophioxylon serpentinum* L. 430  
*Orobanche racemosa* L. 174  
*Orthopappus* Gleason 251  
*Orthosiphon* Benth. 368, 369  
*Orthosiphon aristatus* (Blume) Miq. 86, 119, **368, 405**  
*Orthosiphon grandiflorum* auct. non Terrac. 368  
*Orthosiphon spicatus* auct. non Benth. 368  
*Orthosiphon stamineus* Benth. 368  
*Orthosiphon thymiflorus* (Roth) v.d. Sleseen 370  
*Ostryocarpus* Hook.f. 237  
*Oxalis* L. 371, 372  
*Oxalis corniculata* L. **371**  
*Oxalis corymbosa* DC. 372  
*Oxalis javanica* Blume 371  
*Oxalis magellanica* J.G. Forster 372  
*Oxalis repens* Thunb. 371  
*Pangium edule* Reinw. 299  
*Papaver* L. 66, **373**  
*Papaver bracteatum* Lindley 375, 376, 377  
*Papaver hortense* Hussenot 378  
*Papaver nudicaule* L. 374  
*Papaver officinale* C.C. Gmelin 378  
*Papaver orientale* L. 377  
*Papaver rhoeas* L. 373–376, **378**  
*Papaver setigerum* DC. 376, 378  
*Papaver somniferum* L. 59, **373, 378**  
– subsp. *setigerum* (DC.) Corb. 376  
– subsp. *somniferum* 376  
– subsp. *songaricum* Basil. 376  
Papaveraceae 36  
Papilionoideae 75, 237, 244, 462  
*Parabaena* Miers 481  
*Paraserianthes falcataria* (L.) Nielsen 115  
*Paratropia* (Blume) DC. 434  
*Pardanthus chinensis* (L.) Ker Gawler 148  
Passifloraceae 35  
*Pavetta* L. 312  
*Peperomia* Ruiz & Pav. 379, 380  
*Peperomia pellucida* (L.) Kunth **379**  
*Peperomia tetraphylla* (J.G. Forster) Hook. & Arn. 380  
Peperomiaceae 380  
*Pericampylus glaucus* (Lamk) Merr. 207  
*Petroselinum crispum* (Miller) Nyman ex A.W. Hill 380  
*Phaseolus vulgaris* L. 196  
*Phyllanthodendron* Hemsl. 384  
Phyllanthoideae 384  
*Phyllanthus* L. 369, **381**  
– subgenus *Isocladus* 384  
– subgenus *Phyllanthus* 384  
*Phyllanthus acidissimus* (Blanco) Muell. Arg. 386  
*Phyllanthus acidus* (L.) Skeels **381, 386**  
*Phyllanthus amarus* Schum. **381, 387**  
*Phyllanthus dalbergioides* Wallich ex J.J. Smith 391  
*Phyllanthus debilis* Klein ex Willd. 382, 384, 385, **388**  
*Phyllanthus discoides* Muell. Arg. 382  
*Phyllanthus distichus* (L.) Muell. Arg. 386  
*Phyllanthus elegans* Wallich 382  
*Phyllanthus emblica* L. **381, 388**  
*Phyllanthus erythrocarpus* Ridley 391  
*Phyllanthus fraternus* Webster 368, 382, 384  
*Phyllanthus frondosus* Wallich ex Muell. Arg. 390  
*Phyllanthus gomphocarpus* Hook.f. 382  
*Phyllanthus gueinzii* Muell. Arg. 389  
*Phyllanthus hasskarlianus* Muell. Arg. 390  
*Phyllanthus kunstleri* Hook.f. 390  
*Phyllanthus lepidocarpus* Siebold & Zucc. 392  
*Phyllanthus leprocarpus* Wight 392  
*Phyllanthus maderaspatensis* L. 381, 382, 384, **389**  
*Phyllanthus microcarpus* (Benth.) Muell. Arg. 391  
*Phyllanthus nanus* Hook.f. 387  
*Phyllanthus niruri* auct. non L. 384, 387, 388  
*Phyllanthus niruri* L. 384

- var. *debilis* (Klein ex Willd.) Muell. Arg. 388  
 - var. *javanicus* Muell. Arg. 388  
*Phyllanthus oxyphyllus* Miq. **390**  
*Phyllanthus pulcher* Wallich ex Muell. Arg. 381, **390**  
*Phyllanthus reticulatus* Poir. **381, 391**  
*Phyllanthus simplex* Retz. 381, 382, 384, 385, **391**  
*Phyllanthus swartzii* Kostel. 387  
*Phyllanthus urinaria* L. 381-385, **392**  
 - subsp. *nudicarpus* Rossignol & Haicour 392  
*Phyllanthus vaccinioides* Klotzsch 389  
*Phyllanthus venosus* A. Rich. 389  
*Phyllanthus verrucosus* Elmer 392  
*Phyllanthus virgatus* P. Forst. 384  
*Phyllodium* Desv. 244  
*Phytolacca* L. **392**  
*Phytolacca abyssinica* Hoffm. 396  
*Phytolacca acinosa* Roxb. **392, 395**  
*Phytolacca americana* L. **392, 396**  
*Phytolacca decandra* L. 396  
*Phytolacca dodecandra* L'Hér. 392-395, **396**  
*Phytolacca esculenta* van Houtte 395  
*Phytolacca octandra* L. 392, 393, **396**  
*Phytolacchoideae* 394  
*Picrasma* Blume 162  
*Piper betle* L. 148  
*Piper exiguum* Blume 379  
*Piper nigrum* L. 32, 256  
*Piper pellucidum* L. 379  
*Piperaceae* 380  
*Piscidia* L. 236  
*Plantago* L. 59, 101, 103, 369, **397**  
*Plantago afra* L. 397-400, **401**  
*Plantago arenaria* Waldst. & Kit. 399  
*Plantago asiatica* L. 397-400, **401**  
*Plantago indica* L. 399  
*Plantago lanceolata* L. 397-400, **401**  
*Plantago major* L. **397, 402**  
 - var. *asiatica* (L.) Decne. 401  
*Plantago ovata* Forssk. 397-400, **402**  
*Plantago psyllium* auct. non L. 401  
*Plectranthus* L'Hér. **403**  
*Plectranthus amboinicus* (Lour.) Spreng. **403, 407**  
*Plectranthus apoensis* (Elmer) H. Keng 406  
*Plectranthus barbatus* Andrews **403, 408**  
*Plectranthus congestus* R.Br. 406, **408**  
*Plectranthus kunstleri* Prain 406  
*Plectranthus merrillii* H. Keng 406  
*Plectranthus petraeus* Back. ex Adelb. 406  
*Plectranthus scutellarioides* (L.) R.Br. **403, 408**  
*Plectranthus sparsiflorus* (Elmer) H. Keng 406  
*Plectranthus steenisii* H. Keng 406  
*Plucheeae* 157  
*Plumbaginaceae* 41, 409  
*Plumbaginoideae* 411  
*Plumbago* L. **409**  
*Plumbago aphylla* Bojer ex. Boiss. 409  
*Plumbago auriculata* Lamk 409  
*Plumbago caerulea* Humb., Bonpl. & Kunth 409  
*Plumbago capensis* Thunb. 409  
*Plumbago europaea* L. 409, 411  
*Plumbago indica* L. **409, 411**  
*Plumbago pearsonii* L. Bolus 409  
*Plumbago pulchella* Boiss. 409  
*Plumbago rosea* L. 411  
*Plumbago scandens* L. 409  
*Plumbago zeylanica* L. **409, 412**  
*Podocarpus* L'Hér. ex Pers. 441  
*Poinsettia* Graham 265  
*Poinsettia cyathophora* (Murray) Klotzsch & Garcke 267  
*Poinsettia geniculata* (Ort.) Klotzsch & Garcke ex Klotzsch 268  
*Poinsettia graminifolia* (Michx.) Millsp. 267  
*Poinsettia heterophylla* (L.) Klotzsch & Garcke ex Klotzsch 268  
*Polygonaceae* 41  
*Populus* L. 85, 144, 329, 400  
*Pothoideae* 84  
*Pothos* L. 84  
*Premna* L. **413, 499**  
*Premna benthamiana* Domin 416  
*Premna cardiophylla* Schauer 415  
*Premna cordifolia* Roxb. 414  
*Premna corymbosa* Rottl. & Willd. 416  
*Premna cumingiana* Schauer 414, **415**  
*Premna curranii* H.J. Lam 416  
*Premna divaricata* Wallich ex Schauer 414  
*Premna flavida* Miq. 417  
*Premna foetida* Reinw. ex Blume 416  
*Premna herbacea* Roxb. **413, 415**  
*Premna inaequilateralis* E. Beer & H.J. Lam 416  
*Premna integrifolia* L. 416  
*Premna nauseosa* Blanco 414  
*Premna obtusifolia* R.Br. 416  
*Premna odorata* Blanco **413, 416**  
*Premna oligotricha* Baker 414  
*Premna parasitica* Blume 414  
*Premna peekelii* H.J. Lam 414, **416**  
*Premna pubescens* Blume 416  
*Premna pyramidata* Wallich 417  
*Premna schimperii* Engl. 414  
*Premna serratifolia* L. **413, 416**  
*Premna tomentosa* Willd. **413, 417**  
*Proteaceae* 41  
*Pseudelephantopus* Rohr 251

- Pseudelephantopus spicatus* (Juss. ex Aublet)  
 Rohr 253  
*Psilotrichopsis* C.C. Towns. 87  
*Pueraria* DC. 417  
*Pueraria candollei* Grah. ex. Benth.  
 – var. *mirifica* (Airy Shaw & Suvat.) Niyomdham  
 419  
*Pueraria lobata* (Willd.) Ohwi 417, 419  
 – var. *thomsonii* (Benth.) v.d. Maesen 419  
*Pueraria mirifica* Airy Shaw & Suvat. 419  
*Pueraria montana* (Lour.) Merr. 417  
 – var. *chinensis* (Ohwi) v.d. Maesen & Almeida  
 419  
 – var. *lobata* (Willd.) v.d. Maesen & Almeida  
 417–420  
 – var. *montana* 419  
*Pueraria phaseoloides* (Roxb.) Benth. 420  
*Pueraria thunbergiana* (Sieb. & Zucc.) Benth.  
 417  
*Pueraria tuberosa* (Roxb. ex Willd.) DC. 419  
*Pygmaeopremna* Merr. 414  
*Pygmaeopremna herbacea* (Roxb.) Moldenke 415  
*Pygmaeopremna humilis* Merr. 415  
*Pyrus pyrifolia* (N.L. Burman) Nakai 235  
*Quassia* L. 162, 273  
*Quisqualis* L. 421  
*Quisqualis conferta* (Jack) Exell 421, 422, 423  
*Quisqualis densiflora* Wallich ex Miq. 423  
*Quisqualis glabra* Burm.f. 423  
*Quisqualis indica* L. 421, 423  
 – var. *indica* 424  
 – var. *pierrei* (Gagnep.) O. Lecompte 424  
 – var. *villosa* (Roxb. ex DC.) Kurz 424  
*Quisqualis pierrei* Gagnep. 424  
*Quisqualis prostrata* Craib 423  
*Quisqualis pubescens* Burm.f. 423  
*Quisqualis spinosa* Blanco 423  
*Quisqualis thorelli* Exell 423  
*Rabdosia* Hassk. 405  
 Ranunculaceae 36, 131  
*Rauvolfia* L. 59, 66, 187, 424, 469  
*Rauvolfia amsoniifolia* DC. 429  
*Rauvolfia cambodiana* Pierre ex Pitard 427  
*Rauvolfia chaudocensis* Pierre ex Pitard 427, 428  
*Rauvolfia chinensis* (Spreng.) Hemsl. 431  
*Rauvolfia indochinensis* Pichon 427, 428  
*Rauvolfia javanica* Koord. & Valetton 426, 429  
*Rauvolfia madurensis* (Teijsm. & Binnend.) Boerl.  
 430  
*Rauvolfia perakensis* King & Gamble 431  
*Rauvolfia reflexa* Teijsm. & Binnend. 426, 429  
*Rauvolfia serpentina* (L.) Benth. ex Kurz 59, 65,  
 410, 424, 430  
*Rauvolfia sumatrana* Jack 424, 426, 429, 430  
*Rauvolfia tetraphylla* L. 427  
*Rauvolfia verticillata* (Lour.) Baillon 424, 431  
*Rauvolfia vietnamensis* Ly 427, 428  
*Rauvolfia vomitoria* Afzel. 427  
 Rhamnaceae 41  
*Rhazya stricta* Decne. 428  
*Rheum tanguticum* Maxim. ex Balf. 208  
*Rhinacanthus* Nees 432  
*Rhinacanthus communis* Nees 431  
*Rhinacanthus nasutus* (L.) Kurz 431  
*Rhipogonum* J.R. Forster & J.G. Forster 448  
*Ricinus communis* L. 35, 74, 75  
*Riedelia* Oliv. 114  
*Rollinia* A.St.-Hil. 34  
 Rosaceae 35  
*Rosmarinus* L. 405  
*Rostellaria* Nees 328  
*Rostellularia* Reichenb. 328  
*Rostellularia procumbens* (L.) Nees 331  
*Rotula* Lour. 180  
 Rubiaceae 36, 41, 469  
*Rungia* Nees 328  
*Saccharum confertum* J.S. Presl 309  
*Salvia* L. 346, 405  
*Salvia aegyptiaca* L. 399  
*Samara robusta* (Roxb.) Kurz 258  
 Sapindaceae 176  
*Sapindus saponaria* L. 237  
*Sarothra japonica* (Thunb. ex Murray) Y. Kimura  
 306  
*Sauropus* Blume 384  
 Saxifragaceae 34, 41, 162  
*Scaphochlamys* Baker 332  
*Schefflera* J.R. Forster & J.G. Forster 433  
*Schefflera acuminatissima* Merr. 435  
*Schefflera affinis* (King) R. Vig. 438  
*Schefflera bengalensis* Gamble 436  
*Schefflera blancoi* Merr. 435  
*Schefflera caudata* (S. Vidal) Merr. & Rolfe 435  
 – var. *piperioides* (Elmer) Frodin 435  
*Schefflera cumingii* (Seem.) Harms 436  
*Schefflera curtisii* (King) Ridley 437  
*Schefflera elliptica* (Blume) Harms 433, 436  
*Schefflera elliptifoliola* Merr. 436  
*Schefflera heptaphylla* (L.) Frodin 192, 433, 437  
*Schefflera heterophylla* (Wallich ex G. Don)  
 Harms 437  
*Schefflera insularum* (Seem.) Harms 438  
*Schefflera junghuhniana* (Miq.) Harms 437  
*Schefflera klossii* Ridley 438  
*Schefflera leucantha* R. Vig. 433, 434  
*Schefflera mindanaensis* Merr. 438  
*Schefflera octophylla* (Lour.) Harms 192, 434,  
 437



- Schefflera odorata* (Blanco) Merr. & Rolfe 436  
*Schefflera oxyphylla* (Miq.) R. Vig. **438**  
*Schefflera piperoides* Elmer 435  
*Schefflera simulans* Craib **438**  
*Schefflera subracemosa* R. Vig. 438  
*Schefflera subulata* (Miq.) R. Vig. 438  
*Schefflera trifoliata* Merr. & Rolfe **438**  
*Schefflera venulosa* (Wight & Arn.) Harms 436  
*Scheffleropsis* Ridl. 434  
*Sciadophyllum* P. Browne 434  
 Scrophulariaceae 41, 399  
*Scutellaria* L. **438**  
   – section *Scutellaria* 440  
   – subgenus *Apeltanthus* 440  
   – subgenus *Scutellaria* 440  
*Scutellaria baicalensis* Georgi 439, 440  
*Scutellaria copelandii* Merr. 441  
*Scutellaria cyrtopoda* Miq. 440  
*Scutellaria discolor* Wallich ex Benth. 438, 439, **440**  
   – var. *cyrtopoda* (Miq.) Adelb. 441  
   – var. *discolor* 441  
   – var. *hirta* Handel-Mazzetti 441  
*Scutellaria heteropoda* Miq. 440  
*Scutellaria horsfieldiana* Miq. 441  
*Scutellaria indica* L. 440, **441**  
   – var. *indica* 441  
*Scutellaria javanica* Jungh. **438, 441**  
   – var. *luzonica* (Rolfe) H. Keng 442  
*Scutellaria luzonica* Rolfe 441  
*Scutellaria russeliaefolia* Vatke 441  
*Scutellaria sumatrana* Miq. 441  
*Scutellaria zollingeriana* Briq. 440  
 Scutellarioideae 440  
*Senecio* L. 293  
*Senna* Miller 41, 60, 101, 103, 182, **442**  
*Senna alata* (L.) Roxb. 101, **442, 445**  
*Senna auriculata* (L.) Roxb. 442  
*Senna garrettiana* (Craib) Irwin & Barneby 442, 443  
*Senna hirsuta* (L.) Irwin & Barneby 442, 443  
*Senna obtusifolia* (L.) Irwin & Barneby 442, 444  
*Senna occidentalis* (L.) Link 442, 443, 444  
*Senna siamea* (Lamk) Irwin & Barneby 442, 443  
*Senna sophora* (L.) Roxb. **442, 446**  
*Senna surattensis* (Burm. f.) Irwin & Barneby 442  
*Senna timoriensis* (DC.) Irwin & Barneby 442  
*Senna tora* (L.) Roxb. **442, 446**  
*Shuteria vestita* Wight & Arnott 201  
*Sida* L. 369  
*Sideritis* L. 405  
 Simaroubaceae 162, 273  
*Siphonochilus* J.M. Wood & Franks 332  
 Smilacaceae 448  
*Smilax* L. 21, 47, **447, 455**  
*Smilax aristolochiifolia* Miller 447  
*Smilax australis* R.Br. 447, 448  
*Smilax balansaeana* H. Bon ex Gagnepain 451  
*Smilax blumei* A.DC. **449, 452**  
*Smilax bracteata* K. Presl 447, **449**  
   – subsp. *bracteata* 450  
   – subsp. *verruculosa* (Merr.) T. Koyama 450  
*Smilax calophylla* Wallich ex A.DC. 447, **450**  
*Smilax celebica* Blume 453  
*Smilax chapaensis* Gagnepain 452  
*Smilax china* L. **447, 450, 453**  
*Smilax corbularia* Kunth **451**  
   – subsp. *corbularia* 451  
   – subsp. *synandra* (Gagnepain) T. Koyama 451  
*Smilax glabra* Wallich ex Roxb. **447, 451**  
*Smilax glycopylla* Smith 447  
*Smilax helferi* A.DC. 452  
*Smilax hemsleyana* Craib 453  
*Smilax hypoglauca* Benth. 451  
*Smilax laevis* Wallich ex A.DC. 452  
*Smilax lanceifolia* Roxb. **452**  
   – subsp. *lanceifolia* 452  
   – subsp. *reflexa* (Norton) T. Koyama 452  
*Smilax leucophylla* Blume **447, 449, 452**  
*Smilax lundellii* Killip & C. Morton 448  
*Smilax luzonensis* K. Presl **452**  
*Smilax macrocarpa* Blume 447  
*Smilax megacarpa* A.DC. & C.DC. 447, **452**  
*Smilax micropoda* A.DC. 452  
*Smilax myosotiflora* A.DC. 447, **453**  
*Smilax odoratissima* Blume 450  
*Smilax officinalis* Kunth 447  
*Smilax opaca* (A.DC.) Norton 452  
*Smilax papyracea* Duhamel 447  
*Smilax peguana* A.DC. 451  
*Smilax regelii* Killip & C. Morton 447  
*Smilax sarsaparilla* L. 447, 448  
*Smilax simulans* T. Koyama 453  
*Smilax stenopetala* A. Gray 450  
*Smilax synandra* Gagnepain 451  
*Smilax verruculosa* Merr. 450  
*Smilax verticalis* Gagnepain **453**  
*Smilax wightii* A.DC. 447  
*Smilax zeylanica* L. 447, 448, **453**  
   – subsp. *hemsleyana* (Craib) T. Koyama 453  
   – subsp. *zeylanica* 453  
 Solanaceae 36, 231, 261, 454  
*Solanum* L. **453**  
   – subgenus *Brevantherum* 456  
   – subgenus *Leptostemon* 456  
   – subgenus *Solanum* 456  
*Solanum acetosaefolium* Lamk 460

- Solanum aculeatissimum* auct. non Jacq. 457  
*Solanum aculeatissimum* Jacq. 456  
*Solanum americanum* Miller 456  
*Solanum aviculare* J.G. Forster 456  
*Solanum canaranum* Miq. ex C.B. Clarke 460  
*Solanum capsicoides* All. 453, 454, 455, **457**  
*Solanum ciliare* Willd. 457  
*Solanum ciliatum* Lamk 457  
*Solanum erianthum* D. Don **453, 457**  
*Solanum khasianum* C.B. Clarke 456  
*Solanum laciniatum* Aiton 456  
*Solanum maingayi* O.Kuntze 460  
*Solanum mammosum* L. 453, 455, 456  
*Solanum mauritianum* Blanco 457  
*Solanum melongena* L. 456  
*Solanum nigrum* L. **453, 458**  
*Solanum procumbens* Lour. 456  
*Solanum sanitwongsei* Craib **454, 459**  
*Solanum sarmentosum* Nees **460**  
*Solanum schultesii* Opiz 458  
*Solanum spirale* Roxb. 456  
*Solanum surattense* Burm.f. 456  
*Solanum torvum* Swartz 455  
*Solanum trilobatum* L. 453–456, **460**  
*Solanum verbascifolium* auct. non L. 457  
*Solanum verbascifolium* L. 456  
*Solanum villosum* Miller 456  
*Solanum xanthocarpum* Schrad. & J.C. Wendl. 456  
*Solenostemon* Thonn. 405  
*Solenostemon scutellarioides* (L.) Codd 408  
*Sonchus* L. 368  
*Sophora* L. 460, 462, 474  
*Sophora crassifolia* J. St.-Hil. 460  
*Sophora flavescens* Aiton 461  
*Sophora havanensis* Jacq. 460  
*Sophora heptaphylla* L. 460  
*Sophora japonica* L. 462, 472  
*Sophora subprostrata* Chun & Chen 461  
*Sophora tomentosa* L. 128, **460**  
– subsp. *tomentosa* 462  
Sophoreae 462  
*Sphagnum* L. 149  
Statioideae 411  
*Stephania* Lour. 135, 136, 221, **463, 481**  
*Stephania capitata* (Blume) Spreng. 220, **466**  
*Stephania cepharantha* Hayata 463, 464, 465  
*Stephania erecta* Craib 464  
*Stephania forsteri* (DC.) A. Gray 466  
*Stephania glabra* (Roxb.) Miers 463, 464, 465  
*Stephania hernandiifolia* (Willd.) Walp. 466  
*Stephania japonica* (Thunb.) Miers **463, 466**  
– var. *discolor* (Miq.) Forman 463, 466  
– var. *japonica* 466  
– var. *timoriensis* (DC.) Forman 466  
*Stephania kwangsiensis* H.S. Lo 465  
*Stephania longa* Lour. 465  
*Stephania pierreii* Diels 463, 464, 465  
*Stephania rotunda* Lour. 464, 465  
*Stephania sinica* Diels **463, 467**  
*Stephania tetrandra* S. Moore 463–465  
*Stephania venosa* (Blume) Spreng. 463–465, **467**  
*Strophanthus* DC. 127  
*Strychnos* L. 126–128, 273, **467**  
*Strychnos axillaris* Colebr. 468  
*Strychnos beccarii* Gilg 470  
*Strychnos castelnaei* Wedd. 468  
*Strychnos colubrina* auct. non L. 471  
*Strychnos gautheriana* Pierre ex Dop 468  
*Strychnos guianensis* (Aubl.) Mart. 468  
*Strychnos ignatii* Bergius 127, **467, 470**  
*Strychnos laurina* Wallich ex DC. 471  
*Strychnos ligustrina* Blume 471  
*Strychnos lucida* R.Br. 468, 469, **471**  
*Strychnos minor* Dennst. **471**  
*Strychnos multiflora* Benth. 471  
*Strychnos nux-vomica* L. **467, 472**  
*Strychnos ovalifolia* Wallich ex G. Don 470  
*Strychnos potatorum* L. 468, 469  
*Strychnos rufa* C.B. Clarke 468  
*Strychnos tieute* Lesch. 470  
*Strychnos toxifera* Schomb. ex Benth. 468  
*Strychnos vanprukii* Craib 468  
*Strychnos wallichiana* Steudel ex DC. 468  
*Styphnolobium* Schott ex Endl. 462, 474  
*Styphnolobium japonicum* (L.) Schott 462, **472**  
*Swietenia* Jacq. 235  
*Symphytum* L. 293  
*Syzygium cumini* (L.) Skeels 24  
*Tacca leontopetaloides* (L.) O. Kuntze 394  
*Tadehagi* H. Ohashi 244  
*Tagetes* L. 60, 90, 237, 481  
*Tamarindus indica* L. 394  
*Tanacetum cinerariifolium* (Trev.) Schultz-Bip. 90, 237, 481  
*Taraktogenos kurzii* auct. non King 302  
*Taraxacum* Weber ex F.H. Wigg. 478  
– section *Ruderalia* J. Kirschn., H. Ollg. & Stepanek 478  
– section *Vulgaria* Dahlst. 478  
*Taraxacum indonesicum* Sonck 478  
*Taraxacum javanicum* v. Soest 478  
*Taraxacum officinale* Weber ex F.H. Wigg. **475**  
*Taxus* L. 45  
*Tectona grandis* L.f. 348, 500  
*Teijsmanniodendron* Koord. 499  
*Telanthera philoxeroides* (Mart.) Moq. 108  
*Telanthera pungens* (Kunth) Moq. 109

- Tephrosia* Pers. 236  
*Tephrosia noctiflora* Bojer ex Baker 238  
*Terminalia brassii* Exell 192, 434  
*Terminalia complanata* K. Schumann 192, 434  
*Teucrium* L. 405  
*Thevetia peruviana* (Pers.) K. Schumann 127  
*Thladiantha* Bunge 356  
*Tinomisium* Miers ex Hook.f. & Thomson 481  
*Tinospora* Miers 131, 224, 237, **479**  
*Tinospora cordifolia* (Willd.) Miers 479–482  
*Tinospora coriacea* (Blume) Beumée ex K. Heyne 484  
*Tinospora crispa* (L.) Hook.f. & Thomson **479**, **483**  
*Tinospora glabra* (Burm.f.) Merr. **479**, **484**  
*Tinospora malabarica* (Lamk.) Hook.f. & Thomson 480  
*Tinospora reticulata* Miers 484  
*Tinospora rumphii* Boerl. 481, 483  
*Tinospora sinensis* (Lour.) Merr. 480  
*Tinospora tomentosa* (Colebr.) Hook.f. & Thomson 480  
*Tinospora tuberculata* (Lamk) Beumée ex K. Heyne 483  
*Tinosporeae* 481  
*Tournefortia* L. 293  
*Trichosantheae* 486  
*Trichosanthes* L. 75, 356, **484**  
*Trichosanthes anguina* L. 488  
*Trichosanthes borneensis* Cogn. **488**  
*Trichosanthes bracteata* (Lamk) Voigt 486, 490  
*Trichosanthes cucumerina* L. 484, 486, 487, **488**  
– cv. group Snakegourd 488  
– subsp. *anguina* (L.) Greb. 488  
– var. *anguina* (L.) Haines 488  
*Trichosanthes cucumeroides* (Ser.) Maxim. 489  
*Trichosanthes grandibracteata* Kurz 488  
*Trichosanthes himalensis* C.B. Clarke 489  
*Trichosanthes horsfieldii* Miq. 489  
*Trichosanthes japonica* (Miq.) Regel 489  
*Trichosanthes kerrii* Craib 490  
*Trichosanthes kirilowii* Maxim. **484**, **489**  
– var. *japonica* (Miq.) Kitam 484, 485, 489  
*Trichosanthes ovigera* Blume 484, 485, **489**  
*Trichosanthes pedatifolia* Miq. 488  
*Trichosanthes pubera* Blume 486, 490  
*Trichosanthes quadricirra* Miq. 489  
*Trichosanthes quinquantulata* A. Gray 486, **490**  
*Trichosanthes reniformis* Miq. 488  
*Trichosanthes rosthornii* Harms 489  
*Trichosanthes tricuspidata* Lour. 484, 486, **490**  
*Trichosanthes tricuspis* Miq. 490  
*Trichosanthes trifolia* (L.) Blume 491  
*Trichosanthes trifolia* auct. non (L.) Blume 491  
*Trichosanthes villosa* Blume **490**  
*Trichosanthes wallichiana* Ridley non (Ser.) Wight 488  
*Trichosanthes wawrae* Cogn. **491**  
*Tupidanthus* Hook.f. & Thomson 434  
*Ulex europaeus* L. 462  
*Umbelliferae* 56  
*Uncaria* Schreb. 469  
*Uvaria* L. 34  
*Verbascum* L. 399  
*Verbena* L. 340, 491, 493  
*Verbena bonariensis* L. 491, 492  
*Verbena hallei* Small 493  
*Verbena xhybrida* Voss 491  
*Verbena laciniata* (L.) Briq. 491  
*Verbena officinalis* L. **491**  
– subsp. *africana* R. Fernandes & Verdcourt 493  
– subsp. *hallei* (Small) Barber 493  
– subsp. *officinalis* 493  
*Verbena rigida* Spreng. 491, 492  
*Verbenaceae* 41  
*Verbenoideae* 340  
*Vernonia* Schreber 251, **493**  
– subgenus *Orbisvestus* 495  
– subgenus *Vernonia* 495  
*Vernonia amygdalina* Delile 494, 495  
*Vernonia anthelmintica* (L.) Willd. 493–495  
*Vernonia arborea* Buch.-Ham. 494  
*Vernonia chinensis* Less. 497  
*Vernonia cinerea* (L.) Less. **493**, **496**  
*Vernonia elaeagnifolia* DC. 493, 494  
*Vernonia hymenolepis* A. Rich. 251  
*Vernonia leptophylla* DC. 496  
*Vernonia patula* (Dryander) Merr. **493**, **497**  
*Vernonieae* 251, 495  
*Vigna hosei* (Craib) Backer ex K. Heyne 197  
*Vigna mungo* (L.) Hepper 112  
*Vigna radiata* (L.) Wilczek 110, 196, 212  
*Vinca* L. 188  
*Vinca minor* L. 428  
*Vinca rosea* L. 185  
*Vismieae* 304  
*Vitex* L. 414, **497**  
*Vitex altissima* L.f. 498  
*Vitex celebica* Koord. 501  
*Vitex glabrata* R.Br. 498, 499, **500**  
*Vitex helogiton* K. Schumann 500  
*Vitex heterophylla* Roxb. 501  
*Vitex incisa* Lamk 501  
*Vitex lagundi* Ridley 502  
*Vitex leucoxydon* Blanco 501  
*Vitex leucoxydon* L.f. 498  
*Vitex minahassae* Koord. 500  
*Vitex negundo* L. 64, 237, 481, **497**, **501**

- Vitex paniculata* Lamk 501  
*Vitex parviflora* A.L. Juss. 498  
*Vitex pentaphylla* Merr. 500  
*Vitex pinnata* L. 498  
*Vitex quinata* (Lour.) F.N. Williams 499, **501**  
*Vitex repens* Blanco 502  
*Vitex rotundifolia* L.f. 502  
*Vitex sumatrana* Miq. 501  
*Vitex trifolia* L. **497, 502**  
Viticoideae 499  
*Vitis heptaphylla* L. 437  
*Voacanga* Thouars 469  
*Withania somnifera* (L.) Dunal 455  
*Wollastonia chinensis* (Osbeck) Merr. 105  
*Wrightia tinctoria* R.Br. 297  
*Yucca* L. 448  
*Zingiber officinale* Roscoe 32  
Zingiberaceae 57, 114

# Index of vernacular plant names

Page numbers printed in bold refer to main treatment. For transcriptions of Vietnamese names, see at the end of the index.

- abanico 148  
abaniko 148  
abisrana 166  
abkal 435  
abutra 129  
achillée millefeuille 77  
acore odorant 81  
acore vrai 81  
acoro 81  
adang-adang 408  
adlabon 194  
agagil 247  
agas-moro 496  
agdau 416  
ahos-ahos 228  
ahus 99  
ai tuban 502  
ail 99  
ajeran 154, 407  
ajos 99  
akar ali 453  
akar banar 452  
akar beringin 287  
akar binasa 411  
akar chabang lima 437  
akar dani 421, 423  
akar dedingin 453  
akar gadong 447  
akar gadung tikus 452  
akar gasing bukit 222  
akar ipoh 467, 470  
akar jalar-jalar 281  
akar kancil 450  
akar katah 246  
akar kemenyan hantu 366  
akar ketola hutan 138  
akar kuning 275  
akar lampu bukit 452  
akar pala-pala 281  
akar patah bubul 366  
akar patah gogoh 366  
akar pesat bedak 438  
akar petola hutan 138  
akar pulurun 137  
akar putarwali 483  
akar rebanar 452  
akar rempenang 222  
akar restong 450  
akar sepakan 438  
akar sesudu 438  
akar sulur kerang 257  
akar tanding 453  
akar tiga chabang 491  
akar timun gagak 488  
akar tuba 240  
akkhe thawaan thale 416  
akler 371  
alagao 416  
alang-alang 307, 310  
alangit 178  
alfalfa 418  
alimpuyas 218  
alinang 227  
alligator weed **108**  
aloe **100**  
alpasotis 194  
aluy 119  
am aai 391  
am ai<sup>2</sup> 391  
Amazonian coca 260, 262  
ambin jantan 268  
amin buah 392  
amingit 362  
amniyas 185  
amoras 359, 362  
ampadu 119  
ampalaya 358  
ampalayu 500  
ampelas 282  
amperu lemah 440  
amput di imayyaw 99  
an di[eefn c]or] 367  
an di[eefn hai hoa 366  
ancak 287  
ancar 126  
andadasi 445, 446  
andawali 483  
andudukut 246  
angkanh 442  
anian 97  
anitia 431  
anobran 416  
antanan gede 190  
anti 458  
antiaris 126  
antong sar 272  
antoung sar 272  
antsoan 185  
anuang 227  
aonla **388**  
apiyanga 116  
apple-mint 347  
apugapugan 87  
apulid-gapang 229  
ara 277  
ara bebari 436  
ara bumbing 283  
ara burong 283  
ara jelateh 283  
ara kelumpung 277  
ara kesinai 285  
ara landang puteh 285  
ara seniah 283  
ara sinigai 283  
ara tanah 277, 283  
ara tandok 281  
Arabian jasmine **319**  
arasagat 436  
arbre corail 326  
areng-arengan 311  
areuy baduyut 490  
areuy camcau minyak 466  
areuy geureung 466  
areuy ki koneng 129  
areuy ki tonggeret 241  
areuy tiwuk 489  
aristoloche 133  
asaihe tuni 453  
aseik 126  
asiasimanan 241  
Asiatic pennywort **190**  
asin-asin 390  
asthma herb **268**  
ati-ati besar 408  
ati-ati merah 408  
avavaia 289  
awar-awar 288  
ba d[aa]ju nam 324  
ba g[aj]c hoa d[or] 430  
ba g[aj]c thu[oo]c 430  
ba g[aj]c v[of]ng 431  
ba g[aj]c [aas]n d[ooj] 430  
b[aa]e d[aa]fju 496  
baai 417  
babadotan 92  
babain 349  
babawangan 227  
badiara 408  
b[af]i ng[af]i 246  
baga-as 228  
baguai 286  
baho-baho 341  
bahuerueru 288  
bahug-bahug 92  
bai mat 158  
bai ngai 245  
bai taang dok 268  
bait 269  
b[aj]c d[aa]fju 497  
b[aj]c d[aa]fju l[as] ng[aws]n 225  
b[aj]ch hoa x[af] 412  
b[aj]ch h[aj]c 431  
b[aj]ch ph[uj] t[uwr] 326  
b[aj]ch d[aa]ju kh[aas]u 117  
bala-balangutan 227  
balacai 320, 324

- balacai batai 326  
 balangot 228  
 balaniog 160  
 balatong-aso 446  
 balbas-pusa 368  
 balete 277  
 baleteng-liitan 284  
 bali susuk 282  
 bali-bali 271  
 balitadham 423  
 balloon vine **176**  
 balsam pear 357  
 bama 412  
 ban 306  
 banaba 415  
 banag 447, 449, 452  
 banagan 449  
 banar babi 452  
 bandaul pech 483  
 bandira 287  
 bandotan 294  
 bangbang 412  
 bangisi 138  
 bangkiling 386  
 bangun bangun batu  
   441  
 bangun bangun na  
   gerger 441  
 bangun-bangun 407  
 bankalanan 342  
 bânkráp 349  
 banod 441  
 banogan 429  
 banyan 281  
 banyan tree **281**  
 bao 302  
 barak 218  
 Barbados aloe **104**  
 barsanga 227  
 barsanga-bakir 226  
 baru cina 147  
 barubo 137  
 basikong 282  
 basil 83  
 bastard cardamom **118**  
 bataka 334  
 batu lincar 159  
 bauang pula 97  
 baume des champs 344  
 bawang 93, 99  
 bawang bakung 98  
 bawang besar 97  
 bawang beureum 97  
 bawang bodas 99  
 bawang bombay 97  
 bawang daun 98  
 bawang ganda 98  
 bawang kecil 97  
 bawang merah 97  
 bawang onchang 98  
 bawang putih 99  
 b[aw]ng phi[ees]n 158  
 bayam berduri 110  
 bayam duri 110  
 bayam hutan 110  
 bayam merah 108  
 bayam rusa 342  
 bayem eri 110  
 bayem kremah 109  
 bean 35  
 bebesaran 359  
 bebuas 417  
 bebulang handak 417  
 bebusok 185, 442  
 bedara merah 272  
 bedara putih 272  
 beggar-tick **150**  
 bekak rengat 438  
 bekel 241  
 belai hitam 470  
 benchachat 166  
 bendan 241  
 bendingin 166  
 bereksa 183  
 beringin india 281  
 besaran 362  
 beseng 272  
 besi-besi 330  
 beunghar kucicing 147  
 bi l[eej] 285  
 bidani 423  
 bidara laut 272, 273,  
   471  
 bident 150  
 bident bipenné 153  
 bident triparti 155  
 b[if]nh linh 497  
 b[if]nh linh nh[awx]n  
   500  
 b[if]nh v[oo]i 463, 467  
 bihbul 500  
 bijanggut 344  
 bilanamanut 109  
 billy goat weed 88  
 bina 470  
 bintang berahi 176  
 birthwort **133**  
 bisoro 283  
 bitok 417  
 bitsula 184  
 bitter cucumber 357  
 bitter gourd **357**  
 bitter melon 357  
 black jack **154**  
 black mulberry **363**  
 black nightshade **458**  
 black pepper 256  
 black psyllium 401  
 blackberry lily **148**  
 blond psyllium 402  
 bô prúk' 185  
 bo tree **286**  
 bobondelan 183, 185  
 bobose 458  
 bodhi 286  
 bodhi tree 286  
 bohdi 281  
 bois à enivrer 381  
 bois de gaulettes 381  
 boking-boking 185  
 Bolivian coca 260, 262  
 bongoog 500  
 bonne femme 401  
 b[oof] b[oof] n[ees]p 81  
 b[oof] c[aj]p d[or] 184  
 b[oof] c[oo]ng anh 475  
 b[ooj]ng trang d[or] 313  
 b[ool]ng ng[os]t 388  
 b[ool]ng trang 311  
 b[ool]ng trang d[or] 313  
 b[ool]ng trang tr[aws]ng  
   315  
 boraphet 483  
 boraphet yang daeng  
   467  
 borobotones 227  
 borsa-nga-dadakkal  
   227  
 boto-botones 228  
 boto-botonisan 225  
 botobotonis 268  
 brai xiem 184  
 brojo lintang 148  
 brotowali 483  
 buah bok 190  
 buanal 450  
 buap ngu 488  
 buas-buas 416  
 bubula 195  
 buckhorn plantain **401**  
 b[uf] c[aj]p 185  
 b[uf]m r[uj]n 179  
 bugahin 471  
 bukuan 471  
 bulak-manok 92, 496  
 bulang 417  
 bulum 310  
 bunching onion 98  
 bunga ayam hutan  
   bateh 147  
 bunga chakar ayam  
   366  
 bunga pagar 341  
 bunga pagar puteh 342  
 bunga raja raja 104  
 bunga selang 315  
 bunga serdadu 185  
 bunga tahi ayam 341  
 bunga-bunga 109  
 bungot-bungot 227  
 bunlao 330  
 buntiris 163, 166, 167,  
   337  
 buntot-leon 294  
 buntut tikus 294  
 bunut 277  
 bur-marigold **150**  
 burburtak 154  
 Burma creeper **423**  
 busok-busok 185  
 buyah 98  
 buyok-buyok 358  
 buyung-buyung 496  
 bwax falangx 97  
 ca bia 240  
 c[aa]f]n xa 167  
 c[aa]m k[ee]ch 449  
 c[aa]m dia la 335  
 c[aa]y anh t[us]c 378  
 c[aa]y ba g[aj]c 424  
 c[aa]y b[as] b[ee]nh  
   272  
 c[aa]y b[of] c[aj]p  
   n[uw][ows]c 184  
 c[aa]y b[oo]f d[ee]f 286  
 c[aa]y b[oo]ng c[uws]t  
   heo 92  
 c[aa]y b[oo]ng d[uw]fa  
   185  
 c[aa]y b[oo]ng th[us]i  
   92  
 c[aa]y b[oo]ng [oor]i  
   341  
 c[aa]y ch[os] d[er] 387  
 (c[aa]y) chu[oo]xi h[oo]jt  
   460

- c[aa]y clas]ch 416  
 c[aa]y da 286  
 c[aa]y da b[oo]f] d[ee]f] 286  
 c[aa]y d[aa]f]u giun 195  
 c[aa]y d[aa]f]u h[oo]i] 195  
 c[aa]y d[aj] c[aa]r]m 366  
 c[aa]y hoa c[uws]t l[ow]n 92  
 c[aa]y h[of]e 472  
 c[aa]y la 458  
 c[aa]y lo[es]t m[oo]f]m 366  
 c[aa]y m[aj]n kinh 502  
 c[aa]y m[aws]c c[ow]r] 349  
 c[aa]y m[ux]n 257  
 c[aa]y ng[os]t ngh[ex]o 289  
 c[aa]y nh[us] nho[as]i 289  
 c[aa]y n[oox] 391  
 c[aa]y sui 126  
 c[aa]y su[oo]s]t 160  
 c[aa]y th[aw]f]n l[aw]f]n 285  
 c[aa]y thu[oo]c d[of]i 267  
 c[aa]y thu[oo]s]c b[or]ng 166  
 c[aa]y thu[oo]s]c phi[ee]j]n 378  
 c[aa]y th[os]c l[es]p 246  
 c[aa]y trinh n[uwx] 349  
 c[aa]y tr[aa]m [oor]i 341  
 c[aa]y tr[uw]l[ow]f]ng sinh 166  
 c[aa]y t[uws] qu[is] 341  
 c[aa]y v[of]i voi 294  
 c[aa]y x[aa]s]u h[oor] 349  
 c[aa]y x[oo]j]p x[oo]j]p 285  
 c[aa]y x[uw]l[ow]ng kh[oo] 271  
 cacabutan 116  
 c[af] d[oo]j]c d[uw]l[ow]j]c 233  
 c[af] h[oo]i 458  
 c[af] kheo 337  
 c[af] n[us]t [as]o 458  
 c[af] t[of]m 467  
 c[af]ba th[uf]y 460  
 c[af]d[oo]j]c d[uw]l[ow]j]c 229  
 calamansi 416  
 calamus 81  
 calingcing 371  
 cam [is]ch 452  
 camcao 222  
 camcauh 222  
 camphrier 158  
 canar bokor 452  
 caneficier 183  
 canh ki na 198  
 capa 155  
 Cape aloe **104**  
 capo 155  
 caramele 386  
 c[ar]i ma 159  
 carob 473  
 c[as]ch c[or] 415  
 casse fétide 446  
 c[as]t c[aw]n 417  
 castor 34, 35  
 cekeng 226  
 cekluk 423  
 cekur 334  
 cekur jawa 334  
 cenet 176  
 cengkur 334  
 ceraka merah 411  
 ceremai 386  
 cerisier de Tahiti 386  
 cerme 386  
 cetek 470  
 Ceylon leadwort **412**  
 cha lueat 416  
 cha mang 423  
 chaa 178  
 chaa kruut 379  
 chaa yeepun 178  
 ch[aa]n chim 437  
 ch[aa]n chim leo 436  
 ch[aa]n voi gi[es] 253  
 chaek 431  
 chaiaphruk 185  
 chakar bebek 337  
 chan bat day 266  
 chann tanea 311  
 chantoe phnom kok 371  
 chanvre 167  
 chapa 155, 158, 160  
 chapor 158  
 chaulmoogra 299, 302  
 cheepuk 431  
 ch[ef] v[aw]f]ng 320  
 chenamah gajah 436  
 chë:ng bângko:ng 109  
 cheraka 412  
 cheraka merah 411  
 cherek hantu 390  
 chermala 386  
 chermala 386  
 chettamun phloeng daeng 411  
 chettamun phloeng khaao 412  
 chi 277  
 Chiang phraa mon 330  
 chichirica 185  
 China root **450**  
 Chinese banyan **284**  
 Chinese chives **99**  
 Chinese honeysuckle 423  
 Chinese ixora **313**  
 Chinese sarsaparilla 450  
 Chinese scholar tree 472  
 cho' tau quân 490  
 chocolate-weed **342**  
 ch[os] d[er] r[aw]ng c[uw]a 392  
 chou pruc 257  
 chu me 388  
 chua me ba ch[if]a 371  
 chua m[es]o 257  
 chua ng[us]t 257  
 chüa tau kung 417  
 ch[uf]m bao l[ows]n 302  
 ch[uf]m phong 176  
 ch[uf]m ru[oo]j]t 386  
 chum het lek 446  
 chum het tai 446  
 chumhet thet 445  
 chumhet yai 445  
 chuvondacovallie 411  
 ciboule 98  
 cincau 222  
 cinchona **198**  
 citronella 23  
 climbing lily 289  
 club moss 51  
 coca 59, **258, 262**  
 coeur des Indes 176  
 cogon grass **307, 310**  
 Colombian coca 260, 262  
 common bean 35  
 common blue vitex **502**  
 common jasmine 317  
 common milk hedge **269**  
 common mulberry 363  
 common nightshade 458  
 common onion 97  
 comphor seed 117  
 concombres diable 233  
 congcong belut 391  
 c[oo] chi 472  
 c[oo]ng c[oo]j]ng 119  
 coquelicot 378  
 c[or] s[uwx]a l[as] nh[or] 271  
 c[or] b[aj]c d[aa]u 227  
 c[or] ch[as]y 246  
 c[or] g[aa]s]u n[uw]l[ows]c 226  
 c[or] lau 309  
 c[or] l[uw]l[ow]x]i m[ef]lo 253  
 c[or] m[ur] 268  
 c[or] n[us]t [as]o 227  
 c[or] roi ng[uw]j]a 491  
 c[or] s[uwx]a d[aa]s]t 271  
 c[or] s[uwx]a l[ows]n l[as] 268  
 c[or] s[uwx]a n[aw]f]m 270  
 c[or] s[uwx]ra 268  
 c[or] tranh 310  
 coral plant **326**  
 corn poppy **378**  
 c[os]c k[ef]n 234, 241  
 c[os]c k[ef]n leo 241  
 c[os]c k[ef]n m[aj]nh 241  
 c[os]c k[es]n n[uw]l[ows]c 241  
 c[os]c m[aw]r]n 367  
 c[os]i 222, 228  
 c[os]i g[aj]o 227  
 cotton-leaved physic nut **325**  
 country borage **407**  
 country gooseberry 386  
 creat **119**  
 creeping fig **285**

- creeping wood-sorrel 371  
 crown cinchona **204**  
 c[uf]m r[uj]n 179  
 cup 'khoa<sup>2</sup> 'nhai<sup>1</sup> 358  
 c[ur] chi 472  
 c[ur] g[aa]s[u] 228  
 c[ur] g[aa]s[u] bi[eer]n 229  
 c[ur] m[ooj]t 467  
 Curaçao aloe 104  
 curcuma **210, 215**  
 c[us] c[ow]m 227  
 c[us]c chi thi[ee]n 253  
 c[us]c ch[ir] thi[ee]n 250  
 c[uws]t chu[ooj]t 160  
 cyperus 222  
 d[aa]f[u] lai ti[as] 326  
 d[aa]f[u] m[ef] 324  
 d[aa]f[u] m[ef] d[or] 326  
 d[aa]j[u] t[aa]f[m] 362  
 d[aa]j[u] t[af]u 363  
 d[aa]j[u] t[aw]f[m] 363  
 d[aa]j[y] c[aar]m v[aa]n 320  
 d[aa]j[y] v[aw]f[ng] 320  
 d[aa]ng'hët khmaoch ni 446  
 d[aa]u b[is]ch 275  
 d[aa]u g[ur]a den 363  
 d[aa]u ta 363  
 d[aa]u t[af]u 359  
 d[aa]u t[aw]f[m] 359  
 d[aa]y ch[aws]t 451  
 d[aa]y c[os]c 241, 483  
 d[aa]y d[or] m[or] 490  
 d[aa]y g[aa]s[c] 358  
 d[aa]y g[aj]o 451  
 d[aa]y giun 423  
 d[aa]y giun nh[or] 423  
 d[aa]y khum 451  
 d[aa]y kh[oo]s r[as]ch 138  
 d[aa]y kim cang 447, 451, 452  
 d[aa]y k[ys] ninh 483  
 d[aa]y l[ox]i ti[ee]f[n] 466  
 d[aa]y m[aa]j[t] 240  
 d[aa]y man lees[t] 452  
 d[aa]y m[oo]s[i] 205  
 d[aa]y na t[aa]y 488  
 d[aa]y ng[os] 257  
 d[aa]y ng[us]t 257  
 d[aa]y s[aa]m 222  
 d[aa]y th[aa]f[n] th[oo]ng 483  
 d[aa]y thu[oo]s[c] c[as] 240  
 dadayem 154  
 daing-daing 282  
 d[aj] h[uw]l[ow]ng ng[uw]u 496  
 d[aj] bi 158  
 d[aj]i bi r[as]ch 159  
 d[aj]i phong t[uw]r 302  
 dalit 126  
 dalumpang 366  
 damli thnang 160  
 damong-mabaho 159  
 damong-maria 147  
 dandelion **475**  
 dang het 445  
 dang het khmoch 445  
 dangla 502  
 daniri 366  
 dao 452  
 dap yaang 458  
 dara laut 471  
 daraisig 371  
 darandan 287  
 daraw 81  
 darengdeng 228  
 daringo 81  
 dartrier 445  
 daun akar wali 483  
 daun asem kecil 371  
 daun ati-ati 408  
 daun bawang 98  
 daun biji kacang 268  
 daun bulan 129  
 daun bulu ayam 246  
 daun burung 431  
 daun encok 412  
 daun jari buaya 488  
 daun jarong 153  
 daun jinten 407  
 daun kaki kuda 190  
 daun kambing 416  
 daun kucing 407  
 daun kukur 440  
 daun kukuran 441  
 daun kupang 445  
 daun kurap 445  
 daun mules 249  
 daun picah 246  
 daun poko 344  
 daun salawar 457  
 daun sejuk 166  
 daun sendok 402  
 daun telinga kerbau 457  
 daun tolod 109  
 daun urat 402  
 dawai-dawai 450  
 da l[as] tr[of]n 281  
 da m[is]t 287  
 d[as]ng nhi[ee]f[u] g[aa]n 436  
 d[as]ng thu[oo]n 436  
 dia d[ar]m th[ar]o 253  
 d[ow]n 311  
 dee khon 160  
 d[ej]n ba l[as] 502  
 delah 282  
 dent de lion 475  
 deringu 81  
 derris **23, 234, 240**  
 devil's apple 234  
 dia li[ee]f[n] 334  
 di[ee]f[n] c[ow] holaf[ng] 306  
 di[ee]p h[aj] ch[aa]u 387, 392  
 di[ee]p h[aj] ch[aa]u y[ees]u 388  
 digambi 247  
 dikit-dikit 246  
 dila-dila 253  
 dilang-aso 253  
 dilang-buwaya 104  
 dilang-halo 104  
 dilaw 216  
 disok 334  
 do mai ruu lom 253  
 doeum tuk das 298  
 do:k khaix ped 88  
 dok ung 423  
 dom pur 286  
 dong dueng 289  
 dong preah phnom 423  
 d[oo]f[ng] ti[ee]f[n] (l[oo]ng) 248  
 d[ow]n bu[oo]s[t] 154  
 d[ow]n d[or] 313  
 d[ow]n tr[aws]ng 315  
 downy thorn apple **233**  
 do:yz tük hma: 246  
 dringo 81  
 dua<sup>1</sup> pong<sup>1</sup> 283  
 dudoa 301  
 duea plong 283  
 duea pong 283  
 dukong anak 387, 392  
 dukong-dukong anak 387  
 du[oo]i c[oo]ng 409, 412  
 du[oo]i c[oo]ng hoa d[or] 412  
 du[oo]i c[oo]ng hoa tr[aws]ng 412  
 du[oo]i c[oo]ng tr[aws]ng 412  
 du[oo]s[i] c[or] 275  
 dus-bedusan 92  
 dusol 334  
 dutchman's pipe 133  
 d[uw]a n[us]i 488  
 d[uw]f[a] c[aj]n 185  
 d[uw]l[ow]ng kim hoa 233  
 d[uw]l[ow]ng k[yf] th[ar]o 77  
 d[uw]l[ow]ng xu[aa]n sa 118  
 dyeberry 396  
 East-Indian galangal **334**  
 echalote 97  
 ecorce de conessie 298  
 ee pae 502  
 ekor anjing 402  
 ela-ela 116  
 elephant's foot **250**  
 embalau padang 160  
 emblic myrobalan 388  
 emblique officinale 388  
 enaime 307  
 enceng-enceng 446  
 endod **396**  
 epiap 457  
 epinard malabre 110  
 erbaka 147  
 eseue 81  
 eupatoire bleue 92  
 euphorbe 263  
 euphorbe à feuilles de thym 271  
 euphorbe à fleurs en tête 268  
 euphorbe des anciens 266  
 euphorbe effilé 271  
 euphorbe pilulifère 268



- fa thalaai 119  
 faa laep 452  
 faa paeng 458  
 fai tai din 412  
 fak-khao 358  
 false elephant's foot  
     **253**  
 fangkis 280  
 f'a:z langab 349  
 fèves de Saint-Ignace  
     470  
 ficus **277**  
 field mint 344  
 fig **277**  
 fig ivy 285  
 figue 277  
 fin ton 326  
 finger tree **271**  
 fir 448  
 fistula 183  
 five-leaved chaste tree  
     **501**  
 flame lily **289**  
 flax 399  
 flea seed **402**  
 floppers 166  
 foetid cassia **446**  
 French physic nut 326  
 gabajekni 289  
 gadel 241  
 gadong china 450  
 gadong saberang 450  
 gadung cina 447, 450  
 gai m[ef]o 167  
 gajahan 294  
 galamai 436  
 galamai-amo 436, 438  
 galangal **331**  
 galik 99  
 galumi 502  
 gambir hutan 318  
 gamo 241  
 ganda 99  
 gandarusa 330  
 ganja 167  
 garamut 497  
 garden nightshade 458  
 garlic 35, 83, **99**  
 garogira 417  
 gatas-gatas 268  
 gauai-gauai 438  
 gelang pasir 270, 271  
 gelang susu 271  
 gelenggang 445  
 gelenggang kecil 446  
 gelenggang kecil 446  
 gelenggang padang 446  
 gendiran 342  
 gentian 43  
 gentileng 500  
 German chamomile 79  
 ghadhung tambha 450  
 gia c[aa]fju 458  
 giant spine gourd 358  
 gi[ee]f'n gai 110  
 gilbas 147  
 ginato bobudo 466  
 ginje jawa 167  
 ginseng 46, 47  
 gisol 334  
 gisol na bilog 335  
 giwit 277  
 goan-goan 138  
 goatweed **88, 92, 93**  
 gobul 241  
 gofasa 502  
 gogoat 109  
 gogon 309, 310  
 gohi 282, 289  
 golden shower **183**  
 gondang kasih 88  
 gorong bodas 467  
 gosau ma dungi 387  
 gosau ma dungi roriha  
     392  
 gososo 286  
 gotu-cola 190  
 grain amaranth 111  
 grand plantain 402  
 grass pea 315  
 great plantain **402**  
 green chireta 119  
 groseillier de Ceylan  
     388  
 groundnut 34, 35  
 gumadep 249  
 g[uw]fa 284  
 haang khaao 241  
 haang khaao phaa 81  
 haang takhe 104  
 hae phan chan 272  
 h[af]n the 242, 249  
 h[af]nh c[ur] 97  
 h[af]nh hoa 98  
 h[af]nh h[uw][ow]ng 98  
 h[af]nh t[aa]y 97  
 h[af]nh t[aw]m 97  
 hahang-halo 190  
 hai 277  
 'hai' 358  
 hairy spurge 268  
 hampelas 280  
 hampelas telpe 287  
 hand of Mary 502  
 hang khao nam 81  
 hang lai daeng 240  
 hareuga 150, 153, 154  
 harum 110  
 hatsakhun thet 298  
 hauili 288  
 heart pea 176  
 hedge euphorbia 269  
 h[ej] 99  
 heliotrope **292**  
 hemp 59, **167, 375**  
 henna 41  
 herbe à calalou 458  
 herbe au chagrin 387  
 herbe sacrée 491  
 herbe-à-cinq-côtes 401  
 hèwz hmu: 228  
 himainat 435, 438  
 himbispuyo 249  
 hiya 147  
 hmô-gyin 371  
 hnha:z kh'a: 310  
 hnha:z phak kè:b 246  
 hnwàd mèew 368  
 hoa b[as]t 489  
 hoa h[ar]j d[aw]fng 185  
 h[of]e l[oo]ng 460  
 hogcreeper **241**  
 holarrhena **296, 298**  
 holy wort 491  
 hom chaeo naa 366  
 hom duan hu suea 407  
 hom duan luang 407  
 hom farang 97  
 hom haem 337  
 hòm hna:m 110  
 hom hua yai 97  
 hom proh 334  
 hom-cheen 98  
 hom-paen 99  
 hom-paenyuak 98  
 hom-prang 98  
 hom-tiam 99  
 hom-ton 98  
 Honduras-sarsaparilla  
     447  
 h[oo]f li[ee]n l[as] nh[or]  
     298  
 h[oo]f li[ee]n l[as] to  
     298  
 h[oo]fng anh 378  
 hoom bwàx 97  
 horse cassia **184**  
 horseshoe vitex 501  
 hua khaao-yen nuea  
     451  
 hua khaao-yen wok  
     451  
 Huánoco coca 260, 262  
 h[us]ng chanh 407  
 h[uw][ow]ng ph[uj] 228  
 h[uw][ow]ng ph[uj]  
     bi[eer]n 229  
 hydrocotyle asiatique  
     190  
 hymaseik 126  
 i-nio 246  
 iba 386  
 ibaiba-an 392  
 ibon-ibonan 431  
 ich kone 160  
 igasud 470  
 ilalang 310  
 imora 240  
 Indian banyan 281  
 Indian borage 407  
 Indian gooseberry 388  
 Indian heliotrope **294**  
 Indian hemp 167  
 Indian laburnum 183  
 Indian liquorice **77**  
 Indian pennywort 190  
 Indian poke **395**  
 Indian sorrel **371**  
 inkweed **396**  
 ipo 126  
 ipoh 126  
 ipoh akar 128  
 ipoh akar besar 470  
 ipu tanah 471  
 ivu na mag 154  
 jamaka 148  
 jangata 408  
 janggot 344  
 Japanese arrowroot  
     417  
 Japanese mint 345  
 Japanese pagoda tree  
     **472**  
 Japanese poinsettia  
     **268**  
 jarak 320, 412

- jarak belanda 324  
 jarak beremah 326  
 jarak cina 326  
 jarak gurita 326  
 jarak hitam 326  
 jarak keling 324  
 jarak kosta jarak pagar 324  
 jarak kosta merah 326  
 jarak merah 326  
 jarak pagar 324  
 jarak pendek 234  
 jarak ulung 326  
 jaring 342  
 jaringan 154  
 jarong-jarong 315  
 jarum hutan 314  
 jarum-jarum 311  
 jarum-jarum merah 313  
 jasmin 316  
 jasmin d'arabie 319  
 jasmine 59, **316**  
 jatropha **320**  
 Java coca **262**  
 Java tea 86, **368**  
 Javanese ixora **314**  
 jawer kotok 408, 440  
 jawi jawi 284  
 jebat harimau 341  
 jejawi 284  
 jelutung laut 267  
 jequirity bean 35  
 jerangau 81  
 jerango 81  
 jerangoh 81  
 jeune 408  
 jimsonweed 234  
 jombang 475  
 jonge areuy 160  
 jukut bebalean 226  
 jukut jarem 249  
 jukut jatinangor 108  
 jukut pendul 225  
 jukut pendul bodas 227  
 jukut riyud 349  
 ka bao 302  
 ka chaplak 160  
 ka yom 430  
 ka-ngap 349  
 kaam kung 341  
 kaan thuup 496  
 kaang plaa 390  
 kaang plaa khruca 391  
 kabaiura 445  
 kabkabron 253  
 kabling-gubat 368  
 kaburon 252  
 kacembang 257  
 kachaay 99  
 kachiëw 210  
 kacubung 233  
 kacubung wulung 234  
 kadel 324  
 kadkadot 225  
 kaempferia **331**  
 kagua 282  
 kaguno 452  
 kaiga 226  
 kalaad 205  
 kalabaga 216  
 kalalphruk 185  
 kalam-phak 266  
 kalamphoh 240  
 kalangkang 438  
 kalanum-uak 490  
 kalaw 299, 302  
 kalaw-wa 302  
 kalayar 138, 490  
 kaleke bacu 326  
 Kalinga fig 286  
 kalingan 342  
 kalipapa 502  
 kaliskis-dalag 249  
 kalkalapikap 253  
 kalkugamat 436  
 kalokalo 416  
 kalunai 110  
 kam lam ko 388  
 kam phung 158  
 kam ron tea 313  
 kam rontea 313  
 kam thuat 388  
 kamagsa-obat 449  
 kambra-kambra 294  
 kamkamaulau 233  
 kamlang hualamaan 366  
 kamphaeng jedchunum 129  
 kampupot 319  
 kamu maeng 160  
 kan-tot 388  
 kana 176  
 kana-pistula 184  
 kanabaw 108  
 kancha 167  
 kancha cheen 167  
 kanching bayu jantan 225  
 kanchopni 411  
 kancing baju 154  
 kandolamo 490  
 kang<sup>2</sup> pa 391  
 kanpaphruek 184  
 kantotai 185  
 kântouot srók 386  
 kântûët 386  
 kântûët préi 388  
 kantui damrey 294  
 kantutay 341  
 kaol haol 118  
 kapanitulot 330  
 kapen prey 368  
 kapiat 417  
 kapunten 441  
 karelawai 228  
 karet 277  
 karet rambat 285  
 karitana 166  
 karlatan 87  
 karmay 386  
 karn lam 388  
 karpus 299  
 karvanh 117  
 katakataka 166  
 katanda 445, 446  
 katarai 160  
 katbalonga 470  
 katchibong 233  
 katapan 248  
 katepengleutik 446  
 kath'iem 99  
 katimbau 490  
 katongkat 289  
 katumpangan uler 87  
 kauili 288  
 kawo 330  
 kaya-an 391  
 kayu cina utan 453  
 kayu darah belut 391  
 kayu patah tulang 271  
 kayu penawar 460  
 kayu ular 471  
 kayu urip 271, 337  
 kayut-bulang 391  
 k'biehs 240  
 kechubong 229, 233  
 kechubong hitam 233  
 kechubong puteh 233  
 kecicak abang 108  
 kecubung 229, 233  
 kecupong 233  
 kedusan 241  
 kee fai nok khuun 253  
 kee nok sai 154  
 kelat tandok 314  
 kelurut tanjong 390  
 kelusan 268  
 kem kem 284  
 keman jolok 392  
 kemangur 386  
 kemani bali 246  
 kembang santen merah 313  
 kembang soka 313  
 kembang sungsang 289  
 kembang telek 341  
 kembang tembaga 185  
 kembili-kembili 227  
 kemloko 388  
 kemunting china 185  
 kencur 334, 335  
 ké:ng hmu: 452  
 kéng no:yz ngwà liaz 248  
 kenkhyokeni 411  
 kentangan 408  
 kepijit 160  
 kepleng 466  
 keremak 109  
 keremak susu 268  
 kermak bukit 109  
 keroten 154  
 ket 'hoy 391  
 ketepeng 445  
 ketepeng kebo 445  
 ketepeng kecil 446  
 ketileng 500, 502  
 ketipes 176  
 ketola ular 488  
 ketul 150, 153, 154  
 ketumpangan air **379**  
 kha Chiang chee 81  
 kha om 458  
 khaa taai 458  
 khaam poomz 388  
 khaang amphai 391  
 khaang hua lek 415  
 khaang paak put 342  
 khainao 500  
 khaix ped 109  
 khamin 210, 216  
 khamin chan 216  
 khamin kaeng 216  
 khamin khao 216

- khamin khrua 129  
 khamin khun 218  
 khamin oi 218  
 khan haam suea 437  
 kh'a:ng lua:yx 453  
 khang sai chang 241  
 khao ham 392  
 khao ham 'sano khok 392  
 khao taek 319  
 khauz pièd 417  
 kee hen 500  
 kee kae 341  
 kee-pang-hee 119  
 kheekaa din 491  
 kheekaa khom 490  
 kheekhaak 445  
 kheemoot 241  
 khem baan 313  
 khem daeng 315, 430  
 khem farang 313  
 khem nam 315  
 khem nuu 313  
 khem phuut maa 315  
 khem thong 314  
 khem tuut maa 315  
 khem yai 314  
 kheme 311  
 'khi<sup>2</sup> doy<sup>2</sup> 391  
 'khi<sup>2</sup> hai bai 271  
 'khi<sup>2</sup> hen 500  
 khi lek 442  
 khi let ban 445  
 khi min 216  
 'khi<sup>2</sup> nok 500  
 khia cheen 271  
 khia phaa 266  
 khia thian 271  
 khika-khrua 358  
 khleng kong 298  
 'khmin<sup>2</sup> 'khun<sup>2</sup> 216  
 'khmin<sup>2</sup> khai 218  
 khminz 210  
 khoi cheen 178  
 khok krasun 109  
 khok krasun lek 109  
 khom khwaan 500  
 khon dinso 502  
 khon thee khamao 502  
 khon thiso 502  
 khong khamao 205  
 khong saamyaan 337, 338  
 kh[oor] di[eej]p 119  
 kh[oor] qua 358  
 khoun 184  
 khoun loy<sup>2</sup> 185  
 khrua daao 453  
 khrua khaao tok 88  
 khrua khao nang 241  
 khrua ma noi 205  
 khtüm barang 97  
 khtüm krâhââm 97  
 khtüm sââ 99  
 khtüm sânlök 98  
 kh'u:a hmu: 'wa:k 452  
 khua hung 423  
 khua kao ho 483  
 khua mak 'khao<sup>2</sup> 358  
 kh'üa s'a:thwa' 247  
 khua:ng 447  
 khua:ng khau 451  
 khua:ng khua 449  
 khua:ng la:y 451  
 khua:ng no:yz 453  
 khua:ng th'ô:n 449, 452, 453  
 khuea hin 457  
 khueang 447, 452  
 khueang thon 449  
 khuun 184  
 khwaeng khia 460  
 khwaep thale 241  
 khwum taai ngaai pen 166  
 ki bangbara 502  
 ki benteli 429  
 ki ciyat 288  
 ki encok 412  
 ki malaka 388  
 ki mules 270, 271  
 ki oray 119  
 ki pahang 416  
 ki sambang 88  
 ki soka 311, 314  
 ki ucing 460  
 ki urat 402  
 ki[eej]lu 98  
 ki[ees]n c[of] 431  
 kihitir 116  
 kiki kana kuku 268  
 kikilé 392  
 kim cang 452  
 kim chang d[uws]ng 453  
 kim chang trung 450  
 kim li[ee]n 179  
 kim lu[oo]ng 471  
 kim ti[ee]f[n th[ar]o 248  
 kina 198  
 kinangan 178  
 kini:n 198  
 kling klang dong 467  
 klobop 183  
 ko tan 437  
 koe hee 310  
 kogon 307, 309, 310  
 kogon-lake 309  
 kok huadaeng 227  
 kok kra om 176  
 kok naa 227  
 kok pa pay 502  
 kokavu 371  
 kolokagama 436  
 kolong-kugon 496  
 kom roi 160  
 kon pit 466  
 koncham 153  
 koneng 216  
 koneng gede 217  
 koneng hideung 214  
 koneng kalamasu 215  
 koneng lalab 216  
 koneng pinggang 216  
 koneng tegal 218  
 koneng tinggang 216  
 kong saamyaan 337  
 konti 458  
 korat nasi 248  
 Korean mulberry 363  
 koronitas 341  
 kot chulaalamphuaua 147  
 kotek 184  
 kotek mamak 184  
 koudzou 417  
 kra niat 329  
 krabao 299, 302  
 krabao yai 302  
 krabaou phlae thom 302  
 krabaou thom 302  
 krachao mot 138  
 krachao pheemot 138  
 krachee 472  
 kradueng chang phueak 490  
 kraduuk kaidam 330  
 kramat hujan 315  
 krang 281  
 krathiam 99  
 krathiam-chin 98  
 kráva:nh chru:k 228  
 krawaan 113  
 krawaan khaao 117  
 krawaan paa 118  
 kreete 184  
 kreko krervanh 117  
 kremahan 159  
 kremek 109  
 krewanh 117  
 krokot 108  
 krung badan 222  
 krung khamao 205, 222  
 ku-no kaa-mo 294  
 ku-no-kaa-mo 502  
 kua 416  
 kuang 324  
 kub nyuj 396  
 kucai 99  
 kuchai 99  
 kucubung 233  
 kucubung leutik 234  
 kudzu 417  
 kuichai 99  
 kuinin 198  
 kulaa khaao 329  
 kulau 302  
 kulim papa 497  
 kumboomba 457  
 kumbu 228  
 kumis kucing 368  
 kumis ucing 368  
 kunai 310  
 kunchur 218  
 kunci kunot 334  
 kunci menir 334  
 kunci pepet 334, 335  
 kunik 216  
 kunir 216  
 kunir putih 335  
 kunit 138  
 kunyit 215, 216  
 kunyit putih 335  
 kupiupi 226  
 kuren 466  
 kurukalunggai 387  
 kurukuru 310  
 kusum 160  
 kuti 458  
 kutsay 99  
 kuuning 502  
 kuwalot 160  
 kyet-mei 310

- kyetyo 497  
 kyetyo-po 502  
 la cho 148  
 la r[uw]fng 458  
 laa laeng 310  
 l[aa]m v[oo]f 287  
 l[aa]u s[as]c 490  
 labu ayer hutan 488  
 lada pahit 160  
 l[af]i, hoa nh[af]i 319  
 l[af]ng nhi[ee]f]u hoa 423  
 laggundi 497  
 lagoon spurge **388**  
 lagundi 64, 497, 502  
 lagundi laut laki-laki 502  
 lagunding-dagat 502  
 lai nam 240  
 laiolaioan 392  
 l[aj]c d[ij]a sinh c[aw]n 166  
 lak khoei lak kluea 185  
 laka 388  
 lakadbulan 158  
 lalang 310  
 lalang jawa 309  
 lalangkaplan 160  
 lam 324  
 lame lalaki 430  
 lameh 424, 429  
 lameh utan 429  
 lamingo 267  
 lamlampaka 159  
 lamphong 229, 233  
 langsia siam 272  
 lanh m[as]n 167  
 lankat 160  
 lanphong khao 234  
 lansi-lansinaan 326  
 lantin 402  
 lanting 402  
 lanting haba 402  
 lanting-haba 401  
 lapak 241  
 lapmuen noi 446  
 l[as]c 222, 228  
 lasona 97  
 latai 407  
 laura 284  
 laurel 411  
 laurel fig 284  
 lavakaliu 416  
 lavar 216  
 lawean 287  
 leadwort **409**  
 leaf flower **392**  
 leban 497  
 leban bunga 502  
 leban capo 417  
 leban tandok 502  
 Ledger cinchona 204  
 legundi 497, 502  
 lekha 119  
 lemak kepiting 342  
 lemak ketam 342  
 lemuning 502  
 leng-elengan 247  
 lenggundi 502  
 lengkoyan 471  
 leopard flower 148  
 leopard lily 148  
 lep mu nang 436  
 lep mue naang 421, 423  
 lep nguak 266  
 lepe 81  
 leunca 458  
 levoanna 445  
 liak 233  
 lidah buaya 100, 104  
 life plant **166**  
 lima-lima 435, 436  
 lin suea 285  
 lip anian 97  
 liquorice 46, 47, 75, 77  
 little ironweed 496  
 liuangkag 457  
 llantin 402  
 lo[af]ng n[af]n 470  
 lohong 324  
 lohong khvang kraham 326  
 l[oj] n[oo]f]i 302  
 lokyo 98  
 lom laeng 184  
 long k[es]n 241  
 long zedoary 218  
 l[oo] h[oo]j]i 100, 104  
 l[ox]i ti[ee]f]n d[or] 467  
 lu lu d[uw]j]c 458  
 lubigan 81  
 ludanggan 445  
 lukrabo 299  
 lumai hutan 159  
 lupingan 441  
 luuk khoei taai mae yai  
 tham sop 268  
 luuk tai bai 387, 391  
 luupleep khrueta 176  
 l[uw] h[oo]j]i 104  
 luwing 283  
 l[uw]l[owx]i r[aws]n 367  
 ma 500  
 ma duuk 302  
 ma hai 285  
 ma hung daeng 326  
 ma khaa kong 289  
 ma khaam pom din 387, 392  
 ma khuea ba 229, 233  
 ma khuea kham 457  
 ma khuea khuen 457  
 ma noi 488  
 ma phraao nok khao 267  
 ma waeng khrueta 459, 460  
 ma waeng nok 458  
 ma yao 324  
 ma yom 386  
 ma-khaam pom 388  
 maak ee 118  
 maak khai lang 392  
 maak lek maak noi 502  
 Madagascar periwinkle **185**  
 maduea khon 285  
 maduea plong 283  
 madueo thao 285  
 magi 284  
 magilik 415  
 magkapayos 160  
 magwu 408  
 maha 358  
 mai h[oo]f]n 366  
 maiyaraap 349  
 m[aj]n d[af] la 233  
 m[aj]n kinh 502  
 mak dip nam khang 366  
 mak kham pom 388  
 mak kheua ba 233  
 mak mouk kuay 298  
 mak nhom 386  
 makabuhay 479, 483, 484  
 makahiya 349  
 makhang 502  
 makikitot 271  
 makkao 358  
 makulu 299  
 Malabar nut tree **329**  
 malabuta 466  
 maladita 429  
 malaina 408  
 malaka 388  
 malako farang 326  
 malasaga 241  
 malatabako 252  
 malatalong 457  
 malati leuweung 318  
 malatinta 391  
 malaulasiman 367  
 Malay gooseberry 386  
 Malayan banyan 284  
 Malayan spurge tree **266**  
 malbas-damo 275  
 mali luei 318  
 mali som 318  
 mali son 319  
 malu-malu 349  
 malulee 318  
 malur 160  
 man kai 416  
 mana 326  
 manaba 415  
 mandakaki 342  
 mang-lang-du 110  
 mangaloke 205  
 mangguiau 460  
 mangilang 226  
 manibwohebwahe 288  
 manjinimbi 387  
 manol 318  
 manongao-bobi 160  
 manul 319  
 mao v[ix] d[or] 88  
 mao v[ix] l[oo]ng 87  
 mara 358  
 maragatas 268  
 marasiksik 371  
 maratabako 253  
 maratugi 466  
 margose 357  
 marihuana 167  
 marotti 299  
 marphin choo 491  
 maryuna 496  
 mat kham 391  
 matang-buiud 391  
 matmat 349  
 matricaria 79  
 matuum kaa 490

- mawseed 378  
 m[ax] d[eeɸ] 401  
 m[ax] ti[eeɸ] l[as]ng 471  
 m[ax] ti[eeɸ]n 472  
 m[ax] ti[ee]n th[ar]o  
 491  
 may khoum 184  
 mayana 408  
 me d[aa]st 371  
 me l[as] l[ee]jch 390  
 me r[uw]fng 381, 388  
*médecinier d'Espagne*  
 326  
 m[ef] tr[as] l[af] 117  
 m[ef] tr[es] b[af] 118  
 mehulatu 411  
 melaka 388  
 melaran 227  
 melati 319  
 meliburigan 483  
 melikan 248  
 melon-daga 488  
 melor 319  
 melor hutan 318  
 memalu 349  
 membangun 407  
 memeniran 387, 388,  
 392  
 menderong darat 228  
 menderong ekur tupai  
 226  
 mengkunyit 129  
 meniran 387, 388, 392  
 menur 319  
 merboh 502  
 meroyan puteri 390  
 metel thorn apple 233  
 metimun tikus 222  
 Mexican fireweed 268  
 Mexican tea 194  
 milfoil 77  
 millepertuis 303  
 min-kuabin 190  
 mint 59, **344**  
 mistletoe fig 283  
 mo noi 401  
 mock bodh tree **287**  
 mok noi 298  
 mok thung 298  
 mok yai 298  
 mole plant 268  
 molih 319  
 momordica **353**  
 mon 359, 362
- mongmong 283  
 monhnyin-bin 228  
 m[oo]jlc hoa tr[aws]ng  
 298  
 m[oo]jlc mi[ees]t t[uwr]  
 358  
 moon 359  
 m[oo]ng g[af] 88  
 m[oo]si tr[of]n 205  
 mora 362  
 moral 359  
 moral blanco 362  
 moral negro 363  
 moratti 299  
 morelle 453  
 morelle à grappe 396  
 morelle noire 458  
 morera 359  
 morera blanca 362  
 morera negra 363  
 mota 228  
 mouk may 268  
 mouk nhai<sup>2</sup> 298  
 mouk noy<sup>2</sup> 298  
 mreah 358  
 mu masang 160  
 mue phranaaraai 436  
 mugwort **139, 147**  
 mulberry **359**  
 mulmul 160  
 mungla 160  
 muning 502  
 mu[oof] ng[os]t 446  
 mu[oof]ng 442  
 mu[oof]ng b[of] 185  
 mu[oof]ng ho[af]ng  
 y[ees]n 184  
 mu[oof]ng h[of]e 446  
 mu[oof]ng h[oo]i 446  
 mu[oof]ng l[as]c 445  
 mu[oof]ng ng[ur] 446  
 mu[oof]ng tr[aa]u 445  
 murbei 359, 362  
 muricie 358  
 mûrier 359, 363  
 mûrier blanc 362  
 mûrier noire 363  
 mutha 228  
 m[uw] [ows]p d[aws]ng  
 358  
 m[uw] [ows]p m[ur]  
 358  
 m[uw] [ows]p t[aa]y  
 488
- m[uws]c hoa tr[aws]ng  
 298  
 myrobalan emblic 388  
 naam dao 452  
 naat me khlaen 253  
 naat wua 159  
 naat yai 158  
 naga buana 390  
 nagerus 138  
 nam nom raatchasee  
 268  
 nam nom raatchasee  
 thale 267  
 nam s[aa]m 437  
 namnom raatchasee lek  
 271  
 nanangkaan 268  
 nang dong laang 491  
 'nat 155, 158  
 nê: ti:d kho:x 109  
 neak naeng 118  
 neem 23, 83  
 nelli 388  
 n[es]n t[af]u 99  
 nga tru[aa]jt 218  
 ngaa yoi 302  
 ngai camphor plant **158**  
 ng[air] si 146  
 ng[ar]i cau 207  
 ng[ar]i c[uws]u 147  
 ng[ar]i l[as] kim 147  
 ng[ar]i m[as]u 335  
 ng[ar]i t[is]m 218  
 ng[as]i 283  
 ngh[eej] 210, 216  
 ngh[eej] den 218  
 ngh[eej] r[ee]x v[af]ng  
 218  
 ngh[eej] ten d[oof]ng  
 214  
 ngh[eej] v[af]ng 216,  
 218  
 ngoo ngèewz 488  
 ngot 446  
 ngotokong 306  
 ng[ux] tr[ar]o 502  
 nha d[ar]m 104  
 nha khau mau ri 245  
 nha lap mun 446  
 nha leung meum 446  
 nha nguong xang 294  
 nha tük hma 245  
 nh[aa]n tr[aa]f]n b[aws]c  
 147
- nh[af]i nhi[eeɸ]u hoa  
 319  
 nhao 320, 324  
 nhao luat 326  
 nh[ar] m[uw]jlc n[oj]i  
 271  
 nhayang ung baynoy  
 271  
 nhôm baanz 386  
 nhom ban<sup>2</sup> 386  
 ni khrot 281  
 niam hu suea 407  
 nieo maa 245  
 nieo yai 249  
 nightshade **453**  
 nilam bukit 440  
 ninggrp 416  
 nipple grass 402  
 niyog-niyogan 421, 423  
 noisetier purgatif 326  
 n[oj]c s[owr]i c[or] ban  
 306  
 noksai 150  
 nom in 185  
 nom phichit 488  
 nom ra sa 'si 268  
 nong 126  
 'nong<sup>2</sup> 126  
 noti 270  
 nuat plaa muek khao  
 436  
 nunok 277  
 nusu 280  
 nut grass 222  
 nut sedge **222**  
 nutmeg 84  
 nuut phra phuu 246  
 nux-vomica tree **472**  
 nyaung 277  
 officinal leadwort 411  
 ohohone 286  
 oignon 97  
 okavu 408  
 okokiang 226, 227  
 olasiman-ihalas 379  
 oleander spurge 269  
 omia 288  
 onion **93, 97**  
 [oo] m[oo]i 184  
 opium poppy 59, **378**  
 orang-arang 342  
 orange-barked vitex  
**501**  
 oregano 407

- orere 445  
 otaheite gooseberry **386**  
 oyod santenan 281  
 oyod tungkul 240  
 pa-na-e khaa-doh 190  
 pa-tue 110  
 paa-ul 445  
 paang 167  
 pacing tawa 289  
 pādông fai 248  
 pagang-pagang 160  
 paillotte 310  
 painted leaf **267**  
 painted nettle **408**  
 pakupis 488  
 palarapdap 366  
 palia 358  
 paliaban 483  
 palipit 450  
 palma 148  
 palochina 445  
 pamaynap 87  
 pamedang 429  
 pampasapit 411  
 pamulaklakim 471  
 pan 311  
 panakomo 436  
 pancasona 484  
 pancasuda 318  
 pang pon 466  
 pangalangan 460  
 pangisi 138  
 panicked milkwort 446  
 panyawan vine 483  
 papaitan 484  
 paparau 502  
 papari 358  
 pâprâ:hs 451  
 para-para 226  
 parah-parah 226  
 parahulu 116  
 parajito 431  
 pararan 438  
 pare 357  
 pare welut 488  
 parempasa 429  
 paria 357, 358  
 paria belut 488  
 paria gunung 176  
 paria-aso 176  
 paroka 357  
 parol-parolan 176  
 parsley 380  
 paruk-paruk 358  
 pasak bumi 272  
 patah tulang 271  
 patikan cina 270, 271  
 patikan kebo 268  
 patolang-gubat 490  
 patole 488  
 pau-pau pasir 154  
 pavot 378  
 pavot officinale 378  
 pavot rouge 378  
 pechah priok 311, 313,  
 315  
 pega pega 245  
 pega-pega 246  
 pegaga 190  
 pegagan 190  
 pekan hutan 318  
 pekan jantan 318  
 pekapar 268  
 pelas kebo 285  
 pelir kambing 430  
 pelochok 460  
 pelotok 460  
 pencil tree 271  
 pennyroyal 347  
 pepalut 253  
 pepita de San Ignacio  
 470  
 pepita-sa-katbalogan  
 470  
 peppermint 347  
 perasi putih 497  
 peria 358  
 peria bulan 176  
 peria laut 358  
 periok 358  
 periwinkle 59  
 Peru coca 262  
 Peruvian coca 262  
 petagar mangas 315  
 petawali 483  
 petit tamarind rouge  
 392  
 petola ular 488  
 peundang 450  
 pha nha hay 436  
 phaa laa 118  
 phaen din yen 415  
 phaeng kham hoi 391  
 phaengphuai bok 185  
 'phak 'ha 358  
 phak haak kluai 379  
 phak hom nam 110  
 phak kaat nam 401  
 phak khlet 446  
 phak khuang 366  
 phak krasang 379  
 phak nok 190  
 phak ph'ê:w 109  
 phak thaep 241  
 phak waan baan 446  
 phak waen 190, 371  
 phak-khao 358  
 phakaa krong 342  
 phakha 358  
 phakpeetpe 417  
 phakpet 108  
 phakpet daeng 108  
 phakpet farang 108  
 phakpet khaao 109  
 phakpet thai 109  
 phakpet-nam 108  
 phakpot bok 185  
 phan ma ha 289  
 phan nguu yai 88  
 phan sanai 241  
 phang nhot pang 160  
 phayaa mue lek 470,  
 471  
 phayaa muun lek 471  
 phayaa rai bai 271  
 phee suea nam 248  
 phee suea noi 502  
 ph[ef]n den 381, 391  
 ph'è:ng kh'am h'o:yz  
 246  
 phi ma 'sen 158  
 'phi 'sua<sup>2</sup> noy<sup>2</sup> 502  
 phia<sup>2</sup> fan 160  
 phiak 272  
 pho 286  
 pho kee nok 287  
 pho om 176  
 pho prasaat 287  
 pho see ma haa pho  
 286  
 pho tua phuu 287  
 ph[of]ng k[yr] 138  
 phong t[uw]r 299  
 phrommi daeng 108  
 phti: bânla: 110  
 phut nam 298  
 phyllanthe 381  
 phyllanthus **381**  
 physic nut **320, 324**  
 pied d'éléphant 250  
 piggyback tree **390**  
 pignon d'Inde 324  
 pila 280  
 pill-bearing spurge 268  
 pinaan 178  
 pink river ixora **314**  
 pink shower 184, 185  
 pintado 267  
 pipal tree 286  
 pis kucing 349  
 pisau-pisau 154  
 pisek 366  
 pissenlit 475  
 pit pi' khaao 412  
 pit piu daeng 411  
 pit piu khaao 412  
 pitipitikoto 502  
 plaalai phuengk 272  
 plantain **397**  
 plantain lancéolé 401  
 plantain majeur 402  
 plao lueat khruua 467  
 pohok 344  
 pois de coeur 176  
 pokeweed **392, 396**  
 pokok bajang beranak  
 438  
 pokok batu pelir kamb-  
 ing 431  
 pokok buru hantu 416  
 pokok susu hutan 270  
 pokok telur belangkas  
 367  
 pokru 470  
 polay lakek 430  
 pomme épineuse 229,  
 234  
 pong dam 330  
 ponna 311  
 popoul ach 500  
 popoul tuk 500  
 poppy **373**  
 posa 362  
 potato tree **457**  
 potong kujang 247  
 poughère 324  
 poun po 166  
 poun tay 166  
 pounng-ma-theing 158  
 prab samut 334  
 prahulu 116  
 prak phlè 392  
 pramat monus 160  
 prâpééh chhmóól 391  
 prâtiël prèah 'àngkaól  
 218

- préák phlè 387  
 preh 284  
 prickly amaranth 110  
 prickly-leaved elephant's foot **253**  
 procumbent yellow sorrel 371  
 promoi damrey 294  
 psyllium 59, 101, **401**  
 puar gajah 118  
 puar hijau 118  
 puar tadah embun 117  
 pudoh 109  
 puen noksai 154  
 pugo-pugo 225  
 pukul sedapan 226  
 pulau pipit 430  
 pule pandak 430  
 pupia 358  
 purging nut 324  
 purple nut grass **228**  
 purple nut sedge 228  
 puso-pusoan 137  
 puteri malu 349  
 putiana ma gitipi 436  
 putputai 178  
 putri malu 349  
 putrowali 483  
 puyan 138  
 puyengan 342  
 pyi-nyaung 281  
 qua l[aa]u 489  
 qua l[aa]u nh[aa]n 489  
 quan [aa]m 502  
 qu[ar] giun 421, 423  
 quinin 198  
 quinine **198**  
 quinquina 198  
 qulengapaie 116  
 q[ur]y tr[aa]m th[ar]o 154  
 ra yom 430  
 ra yom teenpet 430  
 r[aa]u m[ef]o 368  
 racine de Chine 450  
 ragi 408  
 raisin d'Amérique 396  
 rajah kayu 183  
 rak luukmaa 270  
 rakkyo **98**  
 rakot 154  
 rami buah 387  
 ranggitan 275  
 Rangoon creeper 423  
 ranosandang 471  
 ranti 458  
 ratchadat 160  
 ratchaphruek 184  
 rau [ax]n g[ox]i 160  
 rau bao 147  
 rau c[af]ng cua 379  
 rau chua 88  
 rau d[eef]n gai 110  
 rau d[ee]ju 109  
 rau d[ee]ju d[or] 108  
 rau m[as] 190  
 rau t[aa]f[n] 407  
 rau t[aa]f[n] d[aa]f[y] l[as] 407  
 raun suluk 445  
 reach 184  
 reach chhpoeus 184  
 reach speu 184  
 red cinchona **204**  
 red ixora **313**  
 red milkweed 267  
 red poppy 378  
 redweed 342  
 r[ef] d[oos]m 258  
 rembang 226  
 rempelas 280  
 remuk jung 368  
 reo dong 118  
 r[er] qu[aj]t 148  
 reut jeum bang 257  
 ribwort 401  
 ringworm bush **445**  
 ro miet 216  
 roabe 245  
 roi des amers 119  
 rosy-flowered leadwort **411**  
 rougette 271  
 rough-leaved stem fig **283**  
 round Siam cardamom **117**  
 round zedoary 218  
 round-rooted galangal **335**  
 rubber euphorbia 271  
 ruese phasom laeo 408  
 ruk ku ning taa no 248  
 ruku gajah 497  
 rumput angka 366  
 rumput barah 271  
 rumput barek sisek putih 249  
 rumput bilis jantan 226  
 rumput bumbat 226  
 rumput butang 227  
 rumput chukor karbau 226  
 rumput ekur kuching 294  
 rumput haliya hitan 228  
 rumput jalang 185  
 rumput janggut baung 226  
 rumput jangot 271  
 rumput jekeng kungit 227  
 rumput kala jenkeng 294  
 rumput kapas 225  
 rumput kudung 226  
 rumput kuluwing 228  
 rumput malu 349  
 rumput mesiyang 226  
 rumput oleh 294  
 rumput pereh jarang 92  
 rumput roman 147  
 rumput sadanan 225  
 rumput sekedok 92  
 rumput sekepet burit 225  
 rumput silupak 227  
 rumput sumbu 226  
 rumput tahi babi 496  
 rumput teki 225, 227  
 rumput upas-upasan 87  
 rusty-leaved bush fig **283**  
 sa aeng bai mon 342  
 'sa 'khang<sup>1</sup> 497  
 'sa mang<sup>2</sup> 423  
 sa nh[aa]n 113, 117, 118  
 sa nh[aa]n nam vang 117  
 s[aa]fju d[aa]u 160  
 s[aa]m cau 207  
 s[aa]m l[oo]ng 222  
 saam paang 417  
 saapraeng saapkaa 62  
 saaraphat phit 460  
 sabila 104  
 sabu lueat 326  
 sabuu daeng 326  
 sabuu dam 324  
 sac phle 184  
 sacking tree 126  
 sadao din 366  
 safran des Indes 215  
 saga 73  
 sagaba 435  
 sage **338, 341**  
 sahakepo 391  
 sai 277  
 sai<sup>1</sup> 358  
 sai khao 284  
 sai rayong 284  
 sai yoi bai thu 284  
 saihe maruani 453  
 sainat 435  
 Saint Ignatius bean **470**  
 sak dam ray 407  
 sak khe kai 417  
 salaeng thom 472  
 salatdai paa 266  
 salee 286  
 saliarra 341  
 salot daeng 326  
 salot paa 324  
 salung-salung 431  
 sam t[oo] 431  
 sambang colok 88  
 sambilata 119  
 sambong 155, 158  
 sampagitang-gubat 318  
 sampagitang-sunsong 319  
 sampaguita 319  
 sampare 205  
 sampasampalukan 387  
 san h[oo] xanh 271  
 san pedro 387  
 sândaèk préi srâmot 460  
 sang mong peng 457  
 sang mou 457  
 sangawnaw 484  
 sangdikit 412  
 saniat moraa 329  
 sansau **205**  
 santan 311, 313  
 santan-pula 313  
 santan-tsina 313  
 sarang burong 417  
 sarap 497  
 sari cina 472  
 sari kuning 472

- sarsaparilla **447**  
 sarsaparillang-china  
     450  
 sarsaparillang-puti **452**  
 sarungkar-a-babassit  
     275  
 asaladaan 379  
 sasawi langit 496  
 satintail 310  
 sausage tree **302**  
 s[aws]n d[aa]y 417  
 sbô:w 310  
 schefflera **433**  
 scutellaire 438  
 seacoast laburnum **460**  
 seaside laurel 381  
 sedawai 450  
 sedingin 163, 166, 335,  
     337  
 segading jantan 314  
 seketan 366  
 selimpas 421, 423  
 semanggen 249  
 semangnen 371  
 sembong 155, 158  
 sembung 155, 158  
 sembung gantung 158  
 sembung lalaki 159  
 sembung utan 158  
 semelit patong 390  
 semiyo akar 471  
 semprit 148  
 'sêng bua<sup>1</sup> 472  
 seng lek 342  
 senggang cucuk 110  
 sensitive 349  
 sensitive plant **349**  
 sepedeh 285, 287  
 seprah 285  
 seregang 437  
 serengan 246  
 serigen 166  
 seringin 163, 166, 335  
 serpent végétal 488  
 serut lanang 178  
 sesudu 266, 267, 269  
 sesudu bukit 270  
 sesudu hutan 270  
 setaceous poppy 376  
 setaka 411  
 setawar kampong 337  
 setawar padang 166  
 setumpol 299  
 seven golden candle-  
     sticks 445  
 shallot onion 97  
 siantan 313  
 siantan hutan 313  
 sibakong 424, 429  
 sibuyas tagalog 97  
 sidit 441  
 siempreviva 337  
 sieo duuk 471  
 sikap dada 371  
 sikatan 314  
 sikir **275**  
 siku dengan 367  
 siku-siku 367  
 silasila 241  
 silver bush 460  
 simbahu 282  
 simbu 270  
 simpa 280  
 sinat 438  
 singao 226  
 singilan 92  
 singkil alas 416  
 sinta 119  
 sio 288  
 sip 268  
 sirawan 129  
 sisek tenggiling 249  
 skullcap **438**  
 slaêng 472  
 slaêng thom 472  
 slak 233  
 slap chravea 401  
 smao kak kdam 227  
 smau bânla 349  
 smau kaè lolook 249  
 smau srâlâb pôpôok  
     248  
 snake gourd **488**  
 snakeroot 133  
 snakewood **424**  
 so-thue 298  
 soap berry 396  
 sobi 241  
 soka 311  
 soka beureum 313  
 soka merah 313  
 solanum **453**  
 som chao 269  
 som din 371  
 som kang 415  
 som kung 257  
 som sangka 371  
 som ten kalm 371  
 song nha k[es]p 153  
 sopan malu 349  
 sornet 154  
 soro-soro 269  
 sorog-sorog 267, 269  
 sosa kecil 282  
 sosor bebek 166  
 souchet rond 228  
 s[ow]n nai 334  
 soya 35  
 Spanish needles **153**  
 Spanish physic nut 326  
 spearmint 347  
 spiny amaranth **110**  
 spiny bitter cucumber  
     358  
 spiny pigweed 110  
 spring onion 98  
 spurge **263**  
 squine 450  
 srâka: niék 248  
 St John's wort **303**  
 star anise 83, 237  
 star jasmine **319**  
 stramoine 229, 234  
 stramoine metel 233  
 strychnine plant 472  
 subsub 158  
 sud-sud 227  
 sudu-sudu 266, 267,  
     269  
 suelda-con-suelda 271  
 suerda 271  
 suganda 407  
 suket ganjahan 147  
 suket lumbungan 226  
 sulsulitik 207  
 suma 129  
 sumang 423  
 sumpat kendi 466  
 sung 277  
 sung cam 281  
 sung d[af]u t[ee]n  
     287  
 sungot-olang 391  
 superb lily 289  
 suruh-suruhan 379  
 s[us]c sa m[aa]t 118  
 susuan i lawanan 267  
 susudu 266, 267, 269  
 susuru 266, 267, 269  
 susuukan 248  
 s[uw]fng h[uw]owu  
     337  
 s[uwr] qu[aa]n 423  
 sweet flag 81  
 sweet gourd **358**  
 sweet potato 484  
 sweet root 81  
 sweet wormwood **146**  
 sycamine 363  
 ta-sha-pen 388  
 t[aa]fm phong 176  
 t[aa]fm ru[oo]t 386  
 t[aa]fn c[uwr]u 330  
 t[aa]fn giao 330  
 t[aa]fn mu[oo]si 451  
 tabang-ahas 87  
 tabat barito 283  
 tabatabakohan 253  
 taboguak 358  
 tabtabako 252  
 tabubok 488  
 tagak-tagak 431  
 tagalolo 288  
 tagpayan 330  
 tagumbau 320  
 tagumbau-a-nalabaga  
     326  
 tagumbau-na-purau  
     324  
 tahaunon 417  
 tahi anjing 92  
 tahi ayam 185, 342  
 taingan-daga 371  
 tairas 267  
 tajungan 484  
 takip-kohol 190  
 takumtakum 392  
 takuta 458  
 talang-pulo 500  
 talankan 412  
 taloangi **207**  
 talong-punay 229, 233  
 talong-siam 459  
 talpak tana 253  
 tam n[aj]i 334  
 tamahilan 218  
 tamaravirua 216  
 tambak-tambak 496  
 tambal-balanding 137  
 tambalisa 446, 460  
 tambara marica 160  
 tampa badak 430  
 tampal besi 391  
 tampu taura 337  
 tamyakhrua 417  
 tang 362



- tang kui 123  
 tang<sup>2</sup> to<sup>1</sup> 436  
 tanganan 436  
 tangon-tangon 379  
 tangpupo 313  
 tap tao 466  
 tapak dara 185  
 tapak leman 253  
 tapak liman 250, 253  
 tappingan-daga 190  
 tarebak 431  
 tarera intalun 282  
 tartaraok 423  
 tarump 445  
 tasem 126  
 tatai 126  
 tataluangi 207  
 taum pauv hmab 248  
 tavoy cardamom 118  
 t[awf]m tang 359  
 tè prey 311  
 tebi 417  
 teenpet lek 430  
 teki 222, 228  
 tembako utan 457  
 tembaroh 417  
 tembelean 341  
 temenggong melela 330  
 temo labak 217  
 temo pao 216  
 temp 270  
 temu 210  
 temu badur 216  
 temu bayi 217  
 temu blenyeh 217  
 temu blobo 215  
 temu erang 214  
 temu giring 215  
 temu glenyeh 217  
 temu hitam 214  
 temu ireng 214  
 temu kuning 216, 218  
 temu kunyit 216  
 temu lawak 217, 218  
 temu lawas 218  
 temu mangga 216  
 temu pauh 216  
 temu purot 215  
 temu putih 218, 335  
 temu putri 216  
 temu rapet 335  
 temu raya 218  
 temu tihing 216  
 temu tis 216
- tengguli 183  
 tengkok biawak hitam 281  
 tentulang 271  
 tepus merah 118  
 tereba jepang 431  
 terong asam hutan 457  
 terong belah 457  
 terong kori 457  
 terong meranti 458  
 terong parachichit 458  
 terong perat 457  
 terong pipit 460  
 terong puyoh 457, 460  
 terong raya 457  
 terong tenang 457  
 terong tikus 460  
 teruah 358  
 tetemung 226  
 teter 457  
 thai-mong 423  
 thanh cao 147  
 thanh hao 146  
 thanh hao hoa v[af]ng 146  
 thanuwen 218  
 thao ca 489  
 thao nhang hang 334  
 thao yang dong 452  
 thaowan priang 241  
 th[ar]o cao 147  
 th[ar]o quy[ees]t minh 446  
 thé de Java 368  
 thiam mae haang 92  
 thi[ee]n li[ee]n l[as] h[ej]p 334  
 thi[ee]n kim d[aw]fng 466  
 thinbozihpyoo 386  
 tho nan 272  
 thong kan<sup>2</sup> sang 431  
 thong khan chang 431  
 thong phan chang 431  
 thong saamyaan 337, 338  
 th[oor] kinh gi[ows]i 195  
 th[oor] ph[uj]c linh 451  
 thopthaep thale 241  
 thorn apple 229, 234  
 thousand weed 77  
 three-leaved chaste tree 502
- three-lobed butterbur 155  
 thu[ax]n 438  
 thu[ax]n [aas]n d[ooj] 441  
 thu[ax]n java 441  
 thu[ax]n nhi[ee]flu m[af]u 440  
 th'üb nhub 349  
 th[uf]n m[ux]n 257  
 thu[oor]c m[oj]li 471  
 thu[oo]c b[aws]n 126  
 thu[oo]c c[uws]u 147  
 thu[oo]c tr[aw]c 330  
 th[ur]y song nha 155  
 th[ur]y x[uw]lowng b[oo]f 81  
 th[uw]lowng l[uj]c 392, 396  
 th[uw]lowng l[uj]c nh[or] 395  
 tiamuun 319  
 ti[as] t[oo] t[aa]y 408  
 tick clovers 242  
 ti[ee]n mao 207  
 ti[ees]t d[ee] 205  
 tigau 502  
 tikel balung 271  
 tikog 228  
 timbangan 138  
 timun bengkok 488  
 timun dendang 484  
 timun gagak 484  
 tin nok 497, 500  
 tinnok 497  
 tintinalino 280  
 tirucalli 271  
 t[is]ch ti[ee]n 431  
 t[is]ch tuy[ees]t th[ar]o 190  
 tius 216  
 toan t[uw]lowng th[ar]o 371  
 tobacco tree 457  
 tobi 417  
 todong periok 311  
 tolod 108  
 toma 440  
 tombak-tombak 160  
 ton tai bai pen 166  
 tong haeng 366  
 tongkat ali 272  
 topu 285  
 tor[or]i 99
- torobuk 358  
 torog-torog 349  
 toropu 358  
 touba 234, 240  
 tournesol indien 294  
 trachiek kranh 190  
 tr[af]ng qu[ar] ba hoa 246  
 trailing red spurge 270  
 tr[aj]ng nguy[ee]n ghi ta 267  
 tr[as]i g[aa]c 358  
 trasiet 502  
 trembilu 391  
 trengguli 183, 185  
 trifold bur-marigold 155  
 trôm sva: 248  
 trompa ng elephante 294  
 tropillo 457  
 trueng baa daan 390  
 Trujillo coca 260, 262  
 tr[uws]ng cua 342  
 tr[uw]owfng sanh r[as]ch l[as] 337  
 tr[uw]owfng sinh l[as] to 337  
 Truxillo coca 262  
 tsaang gubat 178  
 tuba 234, 240, 320, 324  
 tuba bekut 241  
 tuba merah 241  
 tuba root 234, 240  
 tuba-tuba 326  
 tubang amerikano 326  
 tubang-bakod 324  
 tubang-kabayo 159  
 tubli 234, 240  
 tugling-pula 240  
 tuhe tutunu 137  
 tuhog-dalag 226  
 tuk das khla 298  
 tük hma: 246  
 tulang-tulang 271  
 tum kaa daeng 471  
 tum kaa khao 471  
 tungat ali 272  
 tupa-aui 207  
 turk's cap 289  
 turmeric 23, 83, 210, 215  
 tutup bumi 250, 253  
 tuumka daeng 472  
 tuwa areuy 241

- tuwa leteng 240  
 t[uwf] bi 158  
 t[uw][ows]c s[af]ng 331  
 t[uwr] t[oo] hoang 154  
 ulaas]t kim 216  
 uban kayu 176  
 ubut bele sa'ai 117  
 udani 421, 423  
 ueang din 335  
 ufi pata 247  
 ulasiman-aso 367  
 ulasiman-bato 379  
 ulasiman-kalat 366  
 umbrella tree 472  
 ung<sup>1</sup> yang 268  
 ungali 457  
 upar upar 441  
 upas 126  
 upas biji 128, 460  
 upas kamarungi 128  
 upas tree **126**  
 upei 240  
 upling-gubat 280  
 urai 110  
 urat sugi 330  
 urinaire de Malabar  
     392  
 utu guraci 275  
 v[aar]y [oos]c 391  
 v[ai] gi [aas]y 342  
 v[aj]n th[oj] t[aa]y 153  
 'van<sup>2</sup> toup 'moup 334  
 'van<sup>2</sup> 'hom 334  
 var sleng dong dang  
     289  
 v[ar]y r[oof]ng 248  
 v[ar]y d[aws]ng 129  
 Veracruz-sarsaparilla  
     447  
 verbena 491  
 vernonia **493, 496**  
 vervain **491**  
 verveine officinale 491  
 vitex **497**  
 voë bâng-he:t khmô:t  
     452  
 voë chon 451  
 voë mē:m thnam chôn  
     453  
 voë pâpâhs 453  
 voë pâprâ:hs 447  
 voë pâprohs dâmbrèi  
     449  
 voë srâ:m 451  
 voë vè:t 248  
 v[oj]ng c[as]ch 416  
 vor romiet nhi 423  
 waan faimai 104  
 waan haangchaang 148  
 waan haaon non 335  
 waan hang chorakhe  
     104  
 waan hom 334  
 waan kaam puu 289  
 waan lueat haeng 408  
 waan mahaamek 214  
 waan meetyap 148  
 waan nonlap 335  
 waan oi chaang 436  
 waan phraao 207  
 waan teen din 334  
 waan thoraanee saan  
     390  
 wale ammelaum 471  
 waliketupa 246  
 waliketupa sapi 247  
 waluh leuweung 490  
 wamala 81  
 wan chakmotluk 218  
 wan nam 81  
 wanabekira 452  
 waringin jawa 287  
 warosbot 137  
 wase wages 484  
 wassa 289  
 wassa laki-laki 289  
 water mint 347  
 wawaina 285  
 wawulitan 391  
 waybread 402  
 wedusan 92, 241  
 welsh onion **98**  
 white mulberry **362**  
 white-flowered lead-  
     wort 412  
 wild poinsettia 267  
 wild sage 338, 341  
 wola waliyan 116  
 wopope 286  
 wormseed 79, **194**  
 wormwood **139**  
 xa ti[ee]f]n 401  
 x[aj] can 148  
 x[is]ch hoa x[af] 412  
 xu[aa]n ti[ees]t 329  
 x[uw][ow]ng b[oof] 81  
 x[uw][ow]ng c[as] 271  
 x[uw][ow]ng r[oof]ng  
     266, 269  
 x[uw][ow]ng r[oof]ng  
     c[aj]nh 266  
 x[uw][ow]ng r[oof]ng  
     r[af]o 269  
 x[uw][ow]ng r[oof]ng ta  
     269  
 x[uw][ow]ng s[oo]ng  
     160  
 xuy[ee]n t[aa]m li[ee]n  
     119  
 ya fin 378  
 ya haeo 222  
 ya: hu:a 451  
 ya kok 222  
 ya pi 378  
 ya saap raeng 92  
 ya thopthaep 366  
 ya tom tok 458  
 ya-kha 310  
 ya-klethoi 249  
 ya-tanhoi 249  
 ya-tansai 249  
 yaa dok khaao 88, 496  
 yaa enyuet 401  
 yaa haeo muu 228  
 yaa hua 451  
 yaa hua mong 225  
 yaa kae haak khom  
     431  
 yaa kannguu 119  
 yaa khon muu 228  
 yaa kok chaai 227  
 yaa kok dok khaao 225  
 yaa kok lek 227  
 yaa kok saai 227  
 yaa koncham khaao  
     154  
 yaa linnuu 367  
 yaa man kai 431  
 yaa nam muek 268  
 yaa nguang chaang  
     294  
 yaa nguang chaang noi  
     294  
 yaa nuai faai 227  
 yaa nuat maeo 368  
 yaa pan yot 349  
 yaa rang-kaa 226  
 yaa saam liam 228  
 yaa saam wan 496  
 yaa song plong 249  
 yaa tai bai 387, 392  
 yaa teen kaa 226  
 yaa tuet maeo 246  
 yaa tumhu 227  
 yaa yaang 268  
 yaa-lang-ueng 268  
 yaai chuung laan 390  
 yaai theep laan 390  
 yaan thaat 452  
 yan bueang thuai 366  
 yang nong 126  
 yarrow 77  
 yawun 497  
 yeusun 341  
 yellow cinchona 204  
 yellow-fruited moon-  
     seed **129**  
 yerba buena 346  
 yoekiyapinba 166  
 yom hin 390  
 yong 286  
 yuan 126  
 zédoaire 218  
 zedoary **218**

**Transcriptions of Vietnamese characters**

[aa]	= â	[ar]	= ă	[ax]	= ã	[ej]	= ẹ	[oo]	= ô	[ow]	= ơ	[uj]	= ư	[uwx]	= ữ
[aaf]	= à	[as]	= á	[ee]	= ê	[er]	= é	[oof]	= ò	[owf]	= ò	[ur]	= ù	[ux]	= ù
[aaj]	= â	[aw]	= ă	[eef]	= è	[es]	= é	[ooj]	= ô	[owj]	= ơ	[us]	= ú		
[aar]	= ả	[awf]	= ả	[eej]	= ê	[ex]	= ẽ	[oor]	= ỏ	[owr]	= ờ	[uw]	= ư		
[aas]	= ấ	[awj]	= ấ	[eer]	= ê	[if]	= ì	[oos]	= ố	[ows]	= ớ	[uwf]	= ừ		
[aax]	= ã	[awr]	= ả	[ees]	= ế	[is]	= í	[oox]	= ồ	[owx]	= ỡ	[uwj]	= ư		
[af]	= à	[aws]	= ấ	[eex]	= ề	[of]	= ò	[or]	= ỏ	[ox]	= ố	[uwr]	= ử		
[aj]	= ạ	[awx]	= ả	[ef]	= ẹ	[oj]	= ọ	[os]	= ó	[uf]	= ù	[uws]	= ứ		

# The Prosea Foundation (Plant Resources of South-East Asia)

## **Name, location, legal status and structure**

- Prosea is a Foundation under Indonesian law, with an international charter, domiciled in Bogor. It is an autonomous, non-profit, international agency, governed by a Board of Trustees. It seeks linkage with existing regional and international organizations;
- Prosea is an international programme focusing on the documentation of information on plant resources of South-East Asia;
- Prosea consists of a Network Office in Bogor (Indonesia) coordinating 6 Country Offices in South-East Asia, and a Publication Office in Wageningen (the Netherlands).

## **Participating institutions**

- Forest Research Institute of Malaysia (FRIM), Karung Berkunci 201, Jalan FRIM, Kepong, 52109 Kuala Lumpur, Malaysia;
- Indonesian Institute of Sciences (LIPI), Sasana Widya Sarwono, Jalan Gatot Subroto 10, Jakarta 12710, Indonesia;
- Institute of Ecology and Biological Resources (IEBR), Nghia Do, Cau Giay, Hanoi, Vietnam;
- Papua New Guinea University of Technology (UNITECH), Private Mail Bag, Lae 411, Papua New Guinea;
- Philippine Council for Agriculture, Forestry and Natural Resources Research and Development (PCARRD), Los Baños, Laguna, the Philippines;
- Thailand Institute of Scientific and Technological Research (TISTR), 196 Phahonyothin Road, Chatuchak, Bangkok 10900, Thailand;
- Wageningen Agricultural University (WAU), Costerweg 50, 6701 BH Wageningen, the Netherlands.

## **Objectives**

- to document and make available the existing wealth of information on the plant resources of South-East Asia for education, extension work, research and industry;
- to make operational a computerized data bank on the plant resources of South-East Asia;
- to publish the results in the form of an illustrated, multi-volume handbook in English;
- to promote the dissemination of the information gathered.

### **Target groups**

- those professionally concerned with plant resources in South-East Asia and working in education, extension work, research and commercial production (direct users);
- those in South-East Asia depending directly on plant resources, obtaining relevant information through extension (indirect users).

### **Activities**

- the establishment and operation of data bases;
- the publication of books;
- the sponsorship, support and organization of training courses;
- research into topics relevant to Prosea's purpose;
- the publication and dissemination of reports and the research results.

### **Implementation**

The programme period has been tentatively divided into 3 phases:

- preliminary phase (1985–1986): publication of 'Plant Resources of South-East Asia, Proposal for a Handbook' (1986);
- preparatory phase (1987–1990): establishing cooperation with South-East Asia through internationalization, documentation, consultation and publication; reaching agreement on the scientific, organizational and financial structure of Prosea;
- implementation phase (1991–2000): compiling, editing and publishing of the handbook; making operational the computerized data bank with the texts and additional information; promoting the dissemination of the information obtained.

### **Documentation**

A documentation system has been developed for information storage and retrieval called Prosea Data Bank. It consists of 7 data bases:

- BASELIST: primarily a checklist of more than 6200 plant species;
- CATALOG: references to secondary literature;
- PREPHASE: references to literature from South-East Asia;
- ORGANYM: references to institutions and their research activities;
- PERSONYM: references to specialists;
- TEXTFILE: all Prosea publications and additional information;
- PHOTFILE: photographs of useful plants of South-East Asia.

### **Publication**

The handbook in blue cover (hardbound) is distributed by Backhuys Publishers, Leiden, the Netherlands (formerly by Pudoc, Wageningen, the Netherlands). The handbook in green cover (paperback) is distributed in two price-classes: a low-price paperback, distributed by Prosea South-East Asia for all developing countries; a medium-price paperback, distributed by Backhuys

Publishers, Leiden, the Netherlands, and by Prosea South-East Asia for developed countries (becoming available two years after publication of the hard-bound edition). The bibliographies are distributed by Prosea South-East Asia.

*The handbook*

- No 1. Pulses. L.J.G. van der Maesen and Sadikin Somaatmadja (Editors). Pudoc, Wageningen. 1989/ESCAP CGPRT Centre, Bogor. 1990 (out of print)/Prosea, Bogor. 1992.
- No 2. Edible fruits and nuts. E.W.M. Verheij and R.E. Coronel (Editors). Pudoc, Wageningen. 1991/Prosea, Bogor. 1992.
- No 3. Dye and tannin-producing plants. R.H.M.J. Lemmens and N. Wulijarni-Soetjipto (Editors). Pudoc, Wageningen. 1991/Prosea, Bogor. 1992.
- No 4. Forages. L. 't Mannetje and R.M. Jones (Editors). Pudoc, Wageningen. 1992/Prosea, Bogor. 1992.
- No 5(1). Timber trees. Major commercial timbers. I. Soerianegara and R.H.M.J. Lemmens (Editors). Pudoc, Wageningen. 1993/Prosea, Bogor. 1994.
- No 5(2). Timber trees. Minor commercial timbers. R.H.M.J. Lemmens, I. Soerianegara and Wong Wing Chong (Editors). Backhuys Publishers, Leiden. 1995/Prosea, Bogor. 1995.
- No 5(3). Timber trees. Lesser-known timbers. M.S.M. Sosef, L.T. Hong and S. Prawirohatmodjo (Editors). Backhuys Publishers, Leiden. 1998/Prosea, Bogor. 1998.
- No 6. Rattans. J. Dransfield and N. Manokaran (Editors). Pudoc, Wageningen. 1993/Prosea, Bogor. 1994.
- No 7. Bamboos. S. Dransfield and E.A. Widjaja (Editors). Backhuys Publishers, Leiden. 1995/Prosea, Bogor. 1995.
- No 8. Vegetables. J.S. Siemonsma and Kasem Piluek (Editors). Pudoc, Wageningen. 1993/Prosea, Bogor. 1994.
- No 9. Plants yielding non-seed carbohydrates. M. Flach and F. Rumawas (Editors). Backhuys Publishers, Leiden. 1996/Prosea, Bogor. 1996.
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- No 11. Auxiliary plants. I. Faridah Hanum and L.J.G. van der Maesen (Editors). Backhuys Publishers, Leiden. 1997/Prosea, Bogor. 1997.
- No 12(1). Medicinal and poisonous plants 1. L.S. de Padua, N. Bunyapraphatsara and R.H.M.J. Lemmens (Editors). Backhuys Publishers, Leiden. 1999/Prosea, Bogor. 1999.
- No 12(2). Medicinal and poisonous plants 2. N. Bunyapraphatsara, L.S. de Padua and J.L.C.H. van Valkenburg (Editors). (expected publication date 2001).
- No 12(3). Medicinal and poisonous plants 3. R.H.M.J. Lemmens, N. Bunyapraphatsara and L.S. de Padua (Editors). (expected publication date 2002).
- No 13. Spices. C.C. de Guzman and J.S. Siemonsma (Editors). (expected publication date 1999).
- No 14. Vegetable oils and fats. H.A.M. van der Vossen and B.E. Umali (Editors). (expected publication date 2001).
- No 15(1). Cryptogams. Algae. W.F. Prud'homme van Reine and G.C. Trono Jr (Editors). (expected publication date 2000).

- No 15(2). Cryptogams. Ferns.
- No 15(3). Cryptogams. Fungi.
- No 16. Stimulants. H.A.M. van der Vossen and M. Wessel (Editors). (expected publication date 1999).
- No 17. Fibre plants.
- No 18. Plants producing exudates. E. Boer and A.B. Ella (Editors). (expected publication date 2000).
- No 19. Essential-oil plants. L.P.A. Oyen and Nguyen Xuan Dung (Editors). Backhuys Publishers, Leiden. 1999/Prosea, Bogor. 1999.
- No 20. Ornamental plants.

### *Bibliographies*

- Bibliography 1: Pulses. Edition 1. N. Wulijarni-Soetjipto and J.S. Siemonsma (Editors). Prosea, Bogor. 1990.
- Bibliography 2: Edible fruits and nuts. Edition 1. Part 1 and part 2. N. Wulijarni-Soetjipto and J.S. Siemonsma (Editors). Prosea, Bogor/Pudoc, Wageningen. 1993.
- Bibliography 3: Dye and tannin-producing plants. Edition 1. N. Wulijarni-Soetjipto and J.S. Siemonsma (Editors). Prosea, Bogor/Pudoc, Wageningen. 1991.
- Bibliography 4: Forages. Edition 1. N. Wulijarni-Soetjipto (Editor). Prosea, Bogor/Pudoc, Wageningen. 1994.
- Bibliography 5(1): Timber trees: Major commercial timbers. Edition 1. Part 1 and part 2. Sarkat Danimihardja and Soedarsono Riswan (Editors). Prosea, Bogor/Pudoc, Wageningen. 1994.
- Bibliography 5(2): Timber trees: Minor commercial timbers. Edition 1. Sarkat Danimihardja and Djunaedi Gandawidjaja (Editors). Prosea, Bogor. 1996.
- Bibliography 5(3): Timber trees: Lesser-known timbers. Edition 1. Sarkat Danimihardja and Djunaedi Gandawidjaja (Editors). Prosea, Bogor. 1998.
- Bibliography 6: Rattans. Edition 1. N. Wulijarni-Soetjipto and Sarkat Danimihardja (Editors). Prosea, Bogor. 1995.
- Bibliography 7: Bamboos. Edition 1. N. Wulijarni-Soetjipto and Sarkat Danimihardja (Editors). Prosea, Bogor. 1996.
- Bibliography 8: Vegetables. Edition 1. Part 1 and part 2. Sarkat Danimihardja and M.H. van den Bergh (Editors). Prosea, Bogor. 1995.
- Bibliography 9 (CD-ROM & Floppies): Plants yielding non-seed carbohydrates. Edition 1. Sarkat Danimihardja and Djunaedi Gandawidjaja (Editors). Irfan Afandi (Electronic design). Prosea, Bogor. 1999.
- Bibliography 11: Auxiliary plants. Edition 1. Sarkat Danimihardja and Djunaedi Gandawidjaja (Editors). Prosea, Bogor. 1997.

### *Miscellaneous*

- A Selection. E. Westphal and P.C.M. Jansen (Editors). Pudoc, Wageningen. 1989/Prosea, Bogor. 1993.
- Basic list of species and commodity grouping. Version 1. R.H.M.J. Lemmens, P.C.M. Jansen, J.S. Siemonsma, F.M. Stavast (Editors). Prosea Project, Wageningen. 1989. (out of print).
- Basic list of species and commodity grouping. Final version. P.C.M. Jansen,

- R.H.M.J. Lemmens, L.P.A. Oyen, J.S. Siemonsma, F.M. Stavast and J.L.C.H. van Valkenburg (Editors). Pudoc, Wageningen. 1991/Prosea, Bogor. 1993.
- Proceedings of the First Prosea International Symposium, May 22–25, 1989, Jakarta, Indonesia. J.S. Siemonsma and N. Wulijarni-Soetjipto (Editors). Pudoc, Wageningen. 1989. (out of print).
  - Proceedings of the Second Prosea International Workshop, November 7–9, 1994, Jakarta and Cisarua, Indonesia. Rusdy E. Nasution and N. Wulijarni-Soetjipto (Editors). Prosea, Bogor. 1995. (out of print).

### **In brief, Prosea is**

- an international programme, focused on plant resources of South-East Asia;
- interdisciplinary, covering the fields of agriculture, forestry, horticulture and botany;
- a research programme, making knowledge available for education and extension;
- ecologically focused on promoting plant resources for sustainable tropical land-use systems;
- committed to conservation of biodiversity;
- committed to rural development through diversification of resources and application of farmers' knowledge.

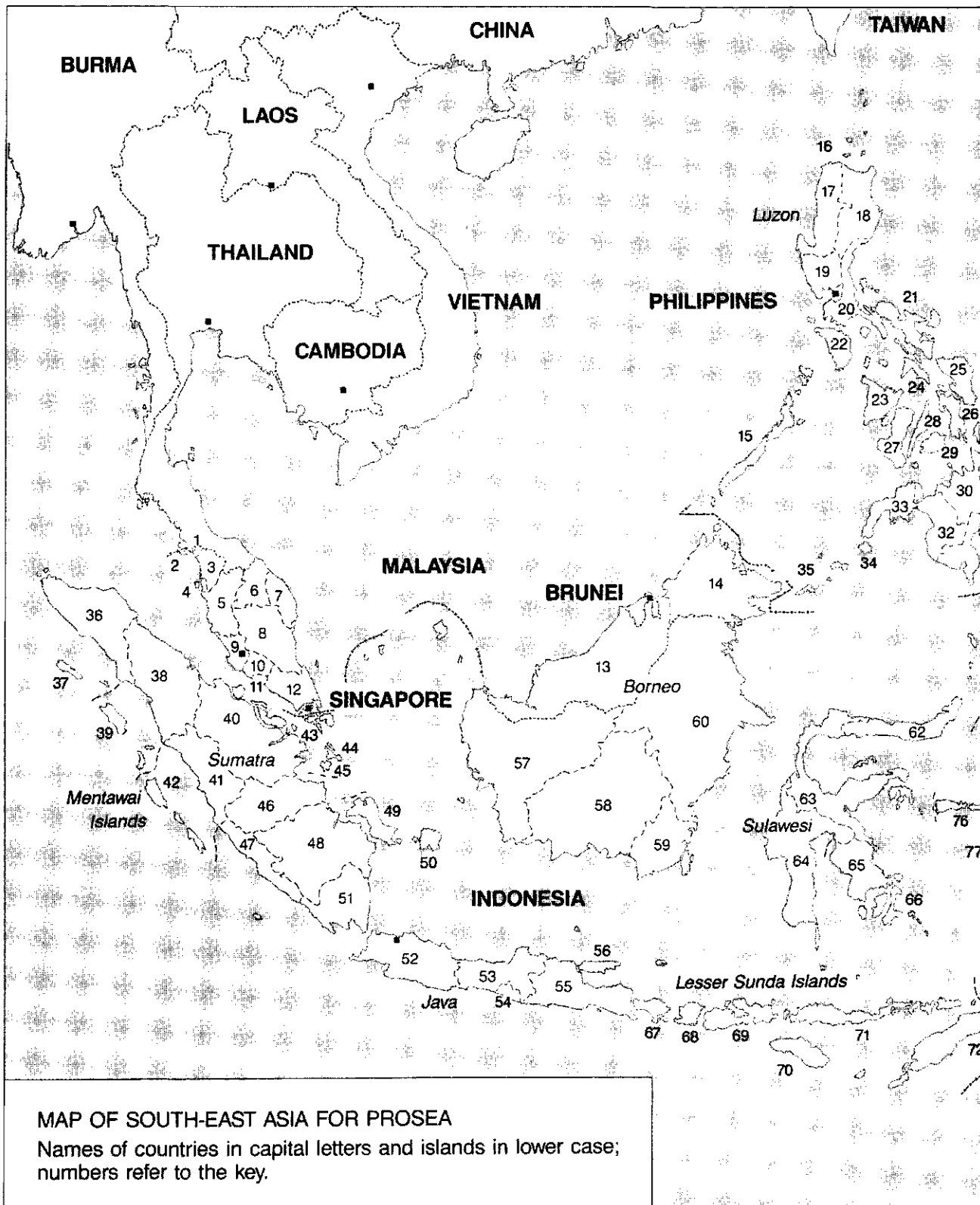
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Key of islands (i), states (s), regions (r) and provinces (p).

**MALAYSIA**

East Malaysia *r* 13-14  
 Johor *s* 12  
 Kedah *s* 3  
 Kelantan *s* 6  
 Langkawi *i* 2  
 Melaka *s* 11  
 Negeri Sembilan *s* 10  
 Pahang *s* 8  
 Peninsular Malaysia  
 (West Malaysia) *r* 1-12  
 Perak *s* 5  
 Perlis *s* 1  
 Pinang *s* 4  
 Sabah *s* 14  
 Sarawak *s* 13  
 Selangor *s* 9  
 Terengganu *s* 7

**PHILIPPINES**

Babuyan Islands *i* 16  
 Basilan *i* 34  
 Bicol *r* 21  
 Bohol *i* 29  
 Cagayan Valley *r* 18  
 Cebu *i* 28  
 Central Mindanao *r* 32  
 Central Luzon *r* 19  
 Ilocos *r* 17  
 Leyte *i* 26  
 Masbate *i* 24  
 Mindoro *i* 22  
 Negros *i* 27

Northern Mindanao *r* 30  
 Palawan *i* 15  
 Panay *i* 23  
 Samar *i* 25  
 Southern Tagalog *r* 20  
 Southern Mindanao *r* 31  
 Sulu Archipelago *i* 35  
 Western Mindanao *r* 33

**INDONESIA**

Aceh *p* 36  
 Ambon *i* 79  
 Aru Islands *i* 82  
 Bali *i* 67  
 Bangka *i* 49  
 Belitung *i* 50  
 Bengkulu *p* 47  
 Buru *i* 77  
 Butung *i* 66  
 Central Java *p* 53  
 Central Kalimantan *p* 58  
 Central Sulawesi *p* 63  
 East Java *p* 55  
 East Kalimantan *p* 60  
 Flores *i* 71  
 Halmahera *i* 74  
 Irian Jaya *p* 84  
 Jambi *p* 46  
 Kai Islands *i* 83  
 Lampung *p* 51  
 Lingga *i* 44  
 Lombok *i* 68  
 Madura *i* 56

Morotai *i* 73  
 Nias *i* 39  
 North Sulawesi *p* 62  
 North Sumatra *p* 38  
 Obi *i* 75  
 Riau *p* 40  
 Riau Archipelago *i* 43  
 Seram *i* 78  
 Siberut *i* 42  
 Simeulue *i* 37  
 Singkep *i* 45  
 South-East Sulawesi *p* 65  
 South Kalimantan *p* 59  
 South Sulawesi *p* 64  
 South Sumatra *p* 48  
 Sula Islands *i* 76  
 Sumba *i* 70  
 Sumbawa *i* 69  
 Talaud Islands *i* 61  
 Timor *i* 72  
 West Daya Islands *i* 80  
 West Java *p* 52  
 West Kalimantan *p* 57  
 West Sumatra *p* 41  
 Yogyakarta *p* 54

**PAPUA NEW GUINEA**  
 Bougainville Island *i* 87  
 D'Entrecasteaux Islands *i* 88  
 Louisiade Archipelago *i* 89  
 New Britain *i* 86  
 Papua *r* 85

Mindanao

51

