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# GENEROSE: Genetic Evaluation of European Rose Resources for Conservation and Horticultural Use

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## Abstract

Under the adagio “Conservation by utilisation is the best way forward for a long-term sustainable protection of the remaining resources” the GENEROSE-project focuses on 3 major objectives: 1) sustainable conservation of wild resources by attributing them an extra value in landscaping or for disease resistance breeding; 2) development of efficient screening techniques for fungal disease resistance and 3) strategies to overcome crossing barriers between wild species and cultivated roses. The project integrates biotechnology (DNA markers for biodiversity evaluation and resistance mapping, flow cytometry for pollen sorting) with original breeding work (use of wild species) and direct potential end-use evaluation by rose breeders and growers. Apart from the ornamental value of possible new wild features, disease resistant cultivars will promote rose production with a lower environmental impact.

## INTRODUCTION

Inventories of the occurrence and distribution of indigenous rose species have been performed in many European countries. These inventories are frequently limited to morphological and taxonomic descriptions although studies on inter- and intra-specific variability have been conducted in dogroses (*Rosa* sect. *Caninae*) using RAPDs (Olsson et al., 2000; Werlemark et al., 1999). In many other plant species, similar studies have been carried out on a large scale using also AFLP, DNA microsatellites and other related techniques. In roses, results are available on the use of these techniques for cultivar identification, for taxonomic studies in the genus *Rosa* and in the first genetic linkage maps (Debener and Mattiesch, 1999; De Riek et al., 2001; Zhang et al., 2000). Except for the studies on dogroses, most of the work in roses has up till now been conducted on cultivated material.

Cultivated roses have a very ancient history, and the first selections were reported already in the early 16<sup>th</sup> century. Later on, artificial crossings led to what is today perceived as the ‘modern rose cultivars’. However, the genetic basis on which these modern rose cultivars are established is very small. Several authors conclude that only between 8 and 20 species out of about 200 have contributed to the origin of our present-day rose cultivars (De Vries and Dubois, 1996; Gudin, 2001; Reynders-Aloisi and Bollereau, 1996). Many of the as yet unexploited rose species may contain valuable genes and traits of interest, which could be used for extending the present genetic diversity in breeding stock material.

Unfortunately, interspecific hybridization has frequently been limited by crossing barriers, which are mainly caused by differences in ploidy level. A modern approach, incorporating techniques for pollen sorting and embryo rescue, should be able to overcome these problems. Pollen sorting has been used successfully in other plant species like *Brassica napus* (Deslauriers et al., 1991; Schulze and Pauls, 2001). Chromosome doubling in roses has mainly been attempted with colchicine, which has not been very successful. The effects of other, more recently applied chemicals like oryzalin still needs to be researched in roses. Recently, dihaploids (parthenogenetically derived by using irradiated pollen and in vitro embryo rescue from tetraploid modern cultivars) have been successfully crossed with diploid species. Interestingly, these dihaploids and their progenies (issued from crosses with wild diploid species) have been shown to produce often unreduced (2n) gametes in large quantities (Crespel et al., 2002).

Other techniques to increase genetic diversity in roses such as radiation, chemical mutagens, somaclonal variation and protoplast fusion have shown limited success (Gudin, 2001). Genetic transformation is an alternative approach to improve specific traits in roses. Some initial results have already been reported, but so far they do not concern agronomic traits (except one publication showing enhanced blackspot resistance) (Marchant et al., 1998). However, the strategy of our research project is to explore the existing diversity in the rose genus itself by developing new breeding strategies based on increased knowledge about the sexual physiology of this polyploid crop and on conventional genetic advances.

Current rose varieties are attacked by a large number of pathogens, under both field and greenhouse conditions. Among the most severe are the fungal diseases blackspot, powdery mildew, downy mildew and rust. Reports on resistance versus susceptibility of different rose varieties and species for all these diseases have been published over the last decades. However, only a few studies have employed systematic screening of the diversity of the pathogens and the respective host species (Debener et al., 1998; Yokoya et al., 2002). Defined genetic material in the form of single conidial isolates and characterized fungal races have only been reported for blackspot. Furthermore, following the genetic characterization of a resistance gene from *R. multiflora*, closely linked markers have been found and the molecular isolation of this gene by positional cloning is currently underway (von Malek et al., 2000). The first evidences for the occurrence of physiological races have also been reported for powdery mildew although detailed studies are still lacking (Bender and Coyier, 1984). Currently, analyses are being undertaken concerning the race structure for both powdery mildew and rust.

## **PROJECT**

The GENEROSE-project started in 2003 within the European Research programme "Quality of Life and Management of Living Resources". Seven research groups and companies from Belgium, France, Germany, The Netherlands and Sweden participate in the consortium. The project consists of 4 major work programs which all focus on the conservation and use of natural resources in the genus *Rosa*.

## **Genetic and Phylogenetic Relationships in Roses**

Attempts to preserve rose gene pools without sufficient knowledge about the genetic background may counter-act efforts to increase biodiversity, compromise the maintenance of (rare) indigenous species in landscaping and make it difficult to (re)introduce autochthonous plant material in the wild. Given that genetic authenticity is an important feature in nature conservancy, the use of molecular tools to perform an in-depth investigation of genetic variability and relatedness is of utmost importance. Within the framework of the GENEROSE-project, a total 316 wild populations belonging to 28 different species have been sampled and described (Table 1). By the use of AFLP and microsatellite DNA markers, the genetic relationships between the different European species in the selected study areas and the biodiversity within and between populations of

wild rose species is studied. Furthermore, a phylogenetic study is conducted, using material of the wild European species, cultivated roses and their supposed ancestors.

### **Characterization of Fungal Diseases and Sources of Resistance**

In this project, the four main fungal diseases on roses (blackspot, powdery mildew, downy mildew and rust) are studied. The project aims to provide representative isolates of the different fungi to be used in standardized protocols for disease resistance screening. The collected wild rose species will be tested in order to point out which material can be useful for resistance breeding. Finally, genetic markers linked with disease resistance will be identified.

### **Crossing Strategies for Interspecific Hybridizations**

A bottleneck in interspecific crosses with rose species at different ploidy levels is caused by the fact that the progeny has an uneven chromosome number (mostly triploids). Within the GENEROSE-project, different strategies to overcome these barriers are investigated, such as pollen sorting, polyploidization and dihaploidisation. Also the occurrence of unreduced gametes in roses is studied. Finally, gene transfer by diploid pollen derived from triploid plants is studied: DNA profiles are developed from haploid and diploid pollen, and compared to the profiles of the parent plants.

### **Evaluation for Horticultural Use of Wild Rose Species**

Collected wild material is multiplied and evaluated for its potential use for landscaping and for horticulturally valuable traits like disease resistance, hip production, perfume, winter hardiness etc. Multiplication methods and cultivation techniques are tested in order to produce valuable plant material of autochthonous species.

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### **Literature Cited**

- Bender, C.L. and Coyier, D.L. 1984. Isolation and identification of races of *Sphaerotheca pannosa* var. *rosae*. *Phytopath.* 73:100-103.
- Crespel, L., Gudin, S., Meynet, J. and Zhang, D. 2002. AFLP-based estimation of 2n gametophytic heterozygosity in two parthenogenetically derived dihaploids of *Rosa hybrida* L. *Theor. Appl. Genet.* 104:451-456.
- Debener, T., Drewes-Alvarez, R. and Rockstroh, K. 1998. Identification of five physiological races of black spot, *Diplocarpon rosae* Wolf. on roses. *Plant Breed.* 117:267-270.
- Debener, T. and Mattiesch, L. 1999. Construction of a genetic linkage map for roses using RAPD and AFLP markers. *Theor. Appl. Genet.* 99:891-899.
- De Riek, J., Dendauw, J., Leus, L., De Loose, M. and Van Bockstaele, E. 2001. Variety protection by use of molecular markers: some case studies on ornamentals. *Plant Biosystems* 135:107-113.
- Deslauriers, C., Powell, A.D., Fuchs, K. and Pauls, K.P. 1991. Flow cytometric characterization and sorting of cultured *Brassica napus* microspores. *Biochim. Biophys. Acta* 1091:165-172.
- De Vries, D.P. and Dubois, L. 1996. Rose breeding: past, present, prospects. *Acta Hort.* 424:241-248.
- Gudin, S. 2001. Rose breeding technologies. *Acta Hort.* 547:23-26.
- Marchant, R., Dazey, M.R., Lucas, J.A., Lamb, C.J., Dixon, R.A. and Power, J.B. 1998. Expression of kitinase transgen in rose (*Rosa hybrida* L.) reduces development of

- blackspot disease (*Diplocarpon rosae* Wolf). Mol. Breeding 4:187-194.
- Olsson, A., Nybom, H. and Prentice, H.C. 2000. Relationships between Nordic dogroses (*Rosa* L. sect. *Caninae*, Rosaceae) assessed by RAPDs and elliptic Fourier analysis of leaflet shape. Syst. Bot. 25:511-521.
- Reynders-Aloisi, S. and Bollereau, P. 1996. Characterisation of genetic diversity in genus *Rosa* by RAPD. Acta Hort. 424:253-259.
- Schulze, D. and Pauls, P. 1998. Flow cytometric characterisation of embryogenic and gametophytic development in *Brassica napus* microspore cultures. Plant Cell Physiol. 39:226-234.
- Von Malek, B., Weber, W.E. and Debener, T. 2000. Identification of molecular markers linked to *Rdr1*, a gene conferring resistance to blackspot in roses. Theor. Appl. Genet. 101:977-983.
- Werlemark, G., Uggla, M. and Nybom, H. 1999. Morphological and RAPD markers show a highly skewed distribution in a pair of reciprocal crosses between hemisexual dogrose species, *Rosa* sect. *Caninae*. Theor. Appl. Genet. 98:557-563.
- Yokoya, K., Kandasamy, K.I., Walker, S., Mandegaran, Z. and Roberts, A.V. 2000. Resistance of roses to pathotypes of *Diplocarpon rosae* Wolf. Ann. Appl. Biol. 136:15-20.
- Zhang, D., Germain, E., Reynders-Aloisi, S. and Gandelin, M.H. 2002. Development of AFLP markers for variety identification in rose. Acta Hort. 502:113-120.

## Tables

Table 1. Overview of the number of collected wild rose populations per country.

Rose species	B*	D	F	NL	Sca
<b>NON-DOGROSES</b>					
<i>R. arvensis</i> (incl. <i>R. sempervirens</i> )	3	6	5	3	
<i>R. majalis</i>		5			2
<i>R. pendulina</i> ( <i>R. alpina</i> )		2	2		
<i>R. spinosissima</i> ( <i>R. pimpinellifolia</i> )	2	4	6	4	
<b>INTERMEDIATE GROUP</b>					
<i>R. glauca</i> Pourr. ( <i>R. rubrifolia</i> ) (Sect. <i>Cinnamomeae</i> or <i>Caninae</i> )	1	2	4		
<i>R. jundzilli</i> (dogrose x <i>R. gallica</i> hybrid)		2			
<i>R. gallica</i>		2	8		
<b>DOGROSES</b>					
<b>Subsection <i>Caninae</i></b>					
<i>R. canina</i>	4	12	3	23	8
<i>R. subcanina</i>	2	1		5	
<i>R. dumalis</i>	1	1	1	5	14
<i>R. corymbifera</i> ( <i>R. dumetorum</i> )	2	10	4	16	
<i>R. subcollina</i>				5	
<i>R. caesia</i> ( <i>R. coriifolia</i> )	1	1	1	1	2
<i>R. montana</i>			3		
<i>R. stylosa</i>	3				
<i>R. canina</i> var. <i>andegavensis</i>	1	1			
<b>Subsection <i>Rubigineae</i></b>					
<i>R. micrantha</i>	2	2			
<i>R. columnifera</i>				8	2
<i>R. rubiginosa</i> ( <i>R. eglanteria</i> )	5	4	4	11	11
<i>R. agrestis</i>	2		4	3	
<i>R. inodora</i>					2
<i>R. elliptica</i>		1	2	1	
<b>Subsection <i>Vestitae</i></b>					
<i>R. tomentosa</i>	3		3	10	
<i>R. pseudoscabriuscula</i>	6	1			
<i>R. sherardii</i>		3	1		2
<i>R. mollis</i>		4			4
<i>R. villosa</i>	2		2		
<b>Subsection <i>Tomentellae</i></b>					
<i>R. tomentella</i>	4			13	

\*B = Belgium, D = Germany, F = France, NL = The Netherlands, Sca = Scandinavian region

