

Occurrence and diversity of endophytic colonization of *Taraxacum officinale* by *Botrytis* species: A preliminary study

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Abstract

The genus *Botrytis* (sexual form *Botryotinia* Whetzel) contains renowned plant pathogenic species with a necrotrophic lifestyle in which kill and feed on its hosts. Recent reports of symptomless endophytic colonization by *Botrytis cinerea* in cultivated plants of lettuce (*Lactuca sativa*) and *Botrytis* spp. in the wild plants dandelion (*Taraxacum agg*) and spotted knapweed (*Centaurea stoebe*) reveal a different “side” of *Botrytis* spp. These findings, expose additional strategies of *Botrytis* species to disseminate and grow in wild plants and also highlight the possibility that there exist (novel?) *Botrytis* species lacking virulence factors or with novel adaptations which could include some benefits to its hosts. In this work, the occurrence of endophytic colonization by *Botrytis* spp. in dandelion plants was investigated. The results, confirmed that symptomless dandelion plants were infected with *Botrytis* spp.. From these plants, 23 *Botrytis* isolates were obtained. By sequencing based on the genes *HSP60* (Heat-shock Protein 60), *G3PDH* (Glyceraldehyde-3-Phosphate Dehydrogenase) and *RPB2* (DNA-dependent RNA polymerase subunit II), 6 different genotypes were identified 3 of them showed the strongest phylogenetic association with *B. cinerea* whereas the other 3 were associated with *B. pseudocinerea*. Importantly, a pathogenicity test showed the capability of these isolates to cause disease lesions in tomato and *Nicotiana benthamiana*. In this report, the implications of the ecology of *Botrytis* species are discussed.

1. Introduction

The genus *Botrytis* (sexual form *Botryotinia* Whetzel) contains renowned plant pathogenic species with worldwide distribution which includes both host generalists and specialists. *Botrytis cinerea*, a generalist species, affects more than 200 eudicots whereas the others are specialized species which affect a single or closely related hosts (Staats et al., 2005). *Botrytis* species are important pathogens in fruits, vegetables, bulbous monocotyledons (members of the families *Liliaceae*, *Amaryllidaceae*, and *Iridaceae*) and green house crops (Jarvis, 1977).

Commonly, *Botrytis* species are known as pathogens with a necrotrophic lifestyle which kill and then feed on the dead cells of plants using in this process a variety of toxic molecules and lytic enzymes (van Kan et al., 2014). For example, in *Botrytis cinerea* host penetration is attributed more to the secretion of H_2O_2 and the degrading enzymes such as lipases, cutinases and oxidases rather than the physical pressure by the pathogen (van Kan et al., 2014). Then, for successful infection, cell death is induced using toxins that are effective against a large spectrum of plants in the case of *Botrytis cinerea* (Choquer et al., 2007). In this species, the toxins botrydial and botcinic acid have been identified as virulence factors (Choquer et al., 2007). In addition, secretion of H_2O_2 by both, the pathogen and the host (hypersensitive response), in response to the pathogen attack, contribute to the pathogen growth (van Kan, 2006; Choquer et al., 2007).

Recent reports of symptomless endophytic colonization by *Botrytis cinerea* in cultivated plants of lettuce (*Lactuca sativa*) (Sowley et al., 2010) and *Botrytis* spp. in wild plants such as dandelion (*Taraxacum agg*) (Shafia, 2009) and spotted knapweed (*Centaurea stoebe*) (Shipunov et al., 2008) provide evidence of the flexibility of *Botrytis* species to establish unnoticeable associations with plants. Although, this finding exposes the additional strategies of pathogenic *Botrytis* species to disseminate and grow in wild plants also highlights the possibility that there exist (novel?) *Botrytis* species lacking virulence factors or with novel adaptations which could include some benefits to its hosts.

In this work, the occurrence of endophytic colonization by *Botrytis* spp. in dandelion plants was investigated and we tried to answer the following research questions:

- How many different *Botrytis* genotypes are detected?
- To which species do these fungi belong?
- Are they novel species or isolates of a known species?

- The plants from which the isolates are sampled are not diseased, but are these isolates capable of causing disease?

2. Materials and methods

2.1 Plant sampling

Dandelion plants were sampled from 4 sites in Wageningen (The Netherlands) (Figure 1, Appendix 1) in May 2014. Sampling sites differed in soil composition and the surrounding vegetation. From each site 22-25 symptomless plants were collected and in total 97 plants were sampled.

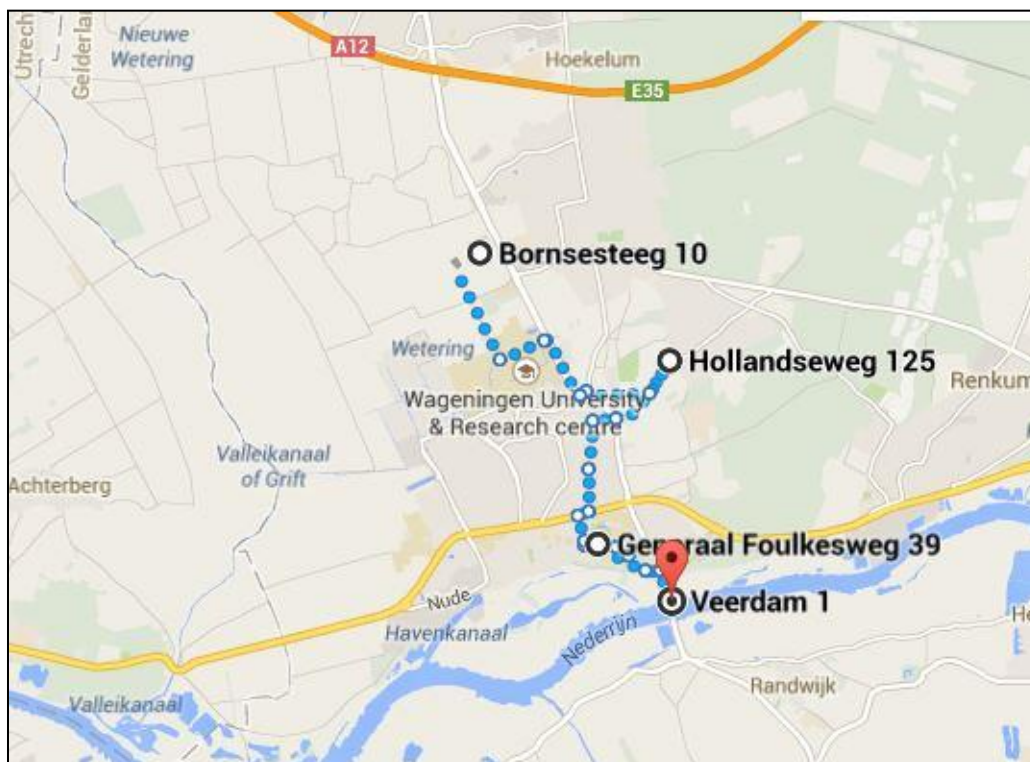


Figure 1. Sampling sites of *Taraxacum officinale* for *Botrytis* isolation in Wageningen

2.2 Botrytis isolation

Botrytis spp. were isolated from stems, leaves and flowers tissues after surface sterilization. Plant tissues were treated 30 s in a bleach solution (2%), 30 s in an ethanol solution (48%) and finally rinsed with water; 4 to 5 pieces from leaves, stems and flowers were plated separately in selective media for *Botrytis* (SBM, supplemented basal medium) (Kerssies, 1990). The media consisted of the following components (g/L distilled water): NaNO_3 , 1.0; K_2HPO_4 , 1.2; $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$, 0.2; KCl, 0.15; glucose, 20.0 and agar, 25.0. The medium was sterilized, cooled

to 65 °C and supplemented with the following ingredients (g/1 distilled water): terrachlor (PCNB, pentachlorobenzene 75%WP), 15×10^{-3} ; tetracycline, 2×10^{-2} ; chloramphenicol (antibiotic), 5×10^{-2} ; CuSO₄, 1.0; tannic acid, 5.0. The pH of the supplemented basal medium (SBM) was adjusted to 4.5 with 5.0 N NaOH.

A total of 291 SBM plates were employed for isolation of *Botrytis* sp. from the different plant tissues. Fungal cultures appeared after 7-10 days of plating and they were screened for the production of dark pigments in the medium due to the degradation of tannic acid, a characteristic property of *Botrytis* spp. Cultures were purified by transferring hyphae from the primary culture with minimal agar onto a fresh sterile plate with the selective media (SBM). Pure cultures were grown on MEA (Malt Extract Agar, DIFCO) plates over a cellophane membrane (1 cm²) which facilitated the sampling of mycelium without agar.

An immunoassay test, using a kit QuickStix strips for *Botrytis* (Envirologix, catalog Number AS 049 GP 25) was employed to identify *Botrytis* colonies among the cultures. For the test, a small sample of mycelium (around 0.5 mm²) was immersed in a eppendorf tube containing 300 µl of a buffer provided with the kit. The mycelium was mixed with the buffer by vortex and then the QuickStix strip for *Botrytis* was placed into the prepared suspension. Positive detection for *Botrytis* sp. was indicated by strips displaying two blue lines (control line and test line).

2.3 Species identification

2.3.1 DNA extraction and sequencing

Mycelial tissue (10-20 mg) from the pure cultures grown over the cellophane membrane was harvested, lyophilized and finely ground to extract DNA, using a modified protocol of Puregene DNA Purification Kit (Gentra Systems Inc./Biozym systems. Landgraaf, The Netherlands). The ground mycelium was placed in a 1.5 ml eppendorf tube (kept in ice) containing 600 µl of cell lysis solution and 3 µl of proteinase K solution (20mg/ml) and mixed by vortexing. Then, the cell lysate was incubated at 55°C for 1.5 h and vortexed periodically every 20 min. Tubes containing the cell lysate were cooled to room temperature and then 200µl of a protein precipitation solution were added and mixed thoroughly. Tubes were incubated on ice for 10-15 min and then centrifuged at 16000 x g for 3 min. 500µl of the supernatant were poured into a new 1.5 ml eppendorf tube containing 500µl of 100% isopropanol. The solution was mixed gently by inverting the tube. The mixture was centrifuged at 16000 x g for 1 min to obtain a DNA pellet, which was then, washed twice

using 600 µl of 70% ethanol. The ethanol was discarded after centrifugation at 16000 x g for 1 min. The DNA was dried in an oven at 37°C around 1 min and then rehydrated and dissolved in 60µl of sterile ultrapure water.

Amplification of regions of the three nuclear genes *HSP60* (Heat-shock Protein 60), *G3PDH* (Glyceraldehyde-3-Phosphate Dehydrogenase) and *RPB2* (DNA-dependent RNA polymerase subunit II) described by Staats et al. (2005) was done using the primers described in the Table 1.

Table 1. Primers employed for PCR amplification of regions of the genes *HSP60*, *G3PDH* and *RPB2* in *Botrytis* spp.

Primer name	Target region	Primer Sequence (5'-3')
G3PDHfor+	G3PDH	GTTTCCAGTCACGACATTGACATCGTCGCTGT
G3PDHrev+		CAGGAAACAGTCATGACACCCACATCGTTGTCGT
HSP60for+	HSP60	GTTTCCAGTCACGACCAACAATTGAGATTTCCCCACAAG
HSP60rev+		CAGGAAACAGCTATGACGATGGATCCAGTGGTACCGAGCAT
RPB2for+	RPB2	GTTTCCAGTCAGGACGATGATCGTGATCATTT
RPB2rev+		CAGGAAACAGTCATGACCCCATAGCTTGCTTACC

PCR amplification was carried out in a 25 µl reaction mixture containing 30-50 ng of DNA, 5X Colorless GoTaq Reaction Buffer (Promega, The Netherlands), 200µM each deoxynucleoside triphosphate (Promega, The Netherlands), 0.2 pmol of each primer (Amersham Pharmacia Biotech), and 1.0 U of GoTaq G2 DNA polymerase (Promega, The Netherlands). PCR conditions for *HSP60* and *RPB2* gene fragment amplification were as follows: 94°C for 5 min (1 cycle); 94°C for 30 s, 55°C for 30 s, and 72°C for 90 s (35 cycles), and then 72°C for 10 min (1 cycle). The same program with an annealing temperature of 64°C was used for *G3PDH* gene. The purified amplified fragments were sequenced by MacroGen Inc. (The Netherlands).

2.3.2 Phylogenetic analysis

From the collection of isolates obtained, initially 12 were sequenced based on *HSP60* gene. The sequence data obtained were employed to perform multiple alignments and sequence comparisons across isolates to select a set representing probably different genotypes or species. Then, the selected isolates were sequenced based on *G3PDH* and *RPB2* genes.

Sequence data of fragments of the genes *HSP60*, *G3PDH* and *RPB2* were analysed, assembled and consensus sequences were obtained using BioEdit software (Biological sequence alignment editor). Multiple sequences alignments and comparisons across isolates were performed by using Clustalw2 (<http://www.ebi.ac.uk/Tools/msa/clustalw2/>) .

HSP60, *G3PDH* and *RPB2* sequences of 26 *Botrytis* species were downloaded from TreeBASE (<http://treebase.org/treebase->

web/home.html;jsessionid=B0D9A31750551B6430F2C506C36EF512) and phylogenetic trees based on *HSP60*, *RPB2*, *G3PDH* and all three genes were reconstructed by Maximum-likelihood with MEGA 6 (Tamura et al., 2013). *Sclerotinia sclerotiorum* and/or *Monilinia fructigena* were included in the phylogenetic analysis as out groups.

2.4 Pathogenicity assay

Pathogenicity tests with five of the *Botrytis* sp. isolates (V18F, V17S, B1LA, A7LA, B18F, A25SB) were carried out on detached leaves of tomato (*Solanum lycopersicum*, cv. Moneymaker) and *Nicotiana benthamiana* plants as described by Zhang and van Kan (2012).

In order to test each selected isolate, two composite tomato leaves and two *N. benthamiana* plants 5-week old were employed. Leaves were cut from tomato plants and stacked in horizontal position with their backsides down in strips of soak oasis placed in petri dishes containing a layer of water. The detached leaves were maintained in plastic trays with a lid to create a humid chamber. Similar moisture conditions were provided for *N. benthamiana* plants in pots.

For leaf inoculation, sporulating *Botrytis* sp. isolates grown for 8-10 days on malt extract agar plates at 18-20°C were flooded with 20 ml of sterile distilled water; the spores were gently removed using a Drigalski spatula and filtered through a glass fibre to eliminate fungi debris. The spore suspension obtained was adjusted to 10⁶ spores /ml using a haemocytometer and a diluted (1:2) potato dextrose solution. Three 2-µl drops of the spore suspension were inoculated on the front side of tomato leaflets and *N. benthamiana* leaves.

One of the tomato detached leaves and one of the *N. benthamiana* plants was inoculated with *Botrytis cinerea* strain B05.10 (control) and with one of the testing *Botrytis* isolates. Each of these was inoculated on one-half of a leaflet/leaf. The other detached tomato leaf and *N. benthamiana* plant was inoculated with the testing isolate alone.

Infective developing lesions were monitored after 24 h and expanding lesions were measured (mm) using a calliper after 48 h.

3. Results

3.1 *Botrytis* isolates

Colonization in dandelion plants (*Taraxacum officinale*) by *Botrytis* sp. isolates was detected in 21% of the plants sampled (N=97) based on the visual observation of dark pigments in the selective medium (an indication of tannic acid degradation, a property of *Botrytis* spp.) and

the immunoassay test. From the 291 SBM plates containing the plant tissues sampled in the 4 sites for isolation, 23 fungal cultures were identified as *Botrytis* by the immunoassay test (Table 2). The percentage of isolates obtained from flowers, stems and leaves were 48%, 39% and 13% respectively (Figure 2). 5 additional fungal cultures identified as *Botrytis* by the immunoassay were not considered (Table 2, Figure 3). In these isolates, formation of sclerotia and conidiphores was absent, they presented slow growth and their morphology was quite distinct to the majority of isolates obtained and identified as *Botrytis*.

Table 2. *Botrytis* cultures identified by immunoassay test (kit QuickStix strips for *Botrytis* spp.) isolated from symptomless dandelion plants (*Taraxacum officinale*).

#	Isolate name	#	Isolate name
1	B1LA	15	A14L
2	B1FA	16	A25SA
3	B13F	17	A25SB
4	B14FA	18	A12F
5	B18F	19	A24FA
6	B21F	20	H17SA
7	B24F	21	H23SA
8	B25F	22	H23SB
9	V8S	23	H24S
10	V9S	24	A1LA*
11	V17S	25	A11FA*
12	V5F	26	B2L*
13	V18F	27	H1L*
14	A7LA	28	H15S*

*Unlikely *Botrytis* isolates

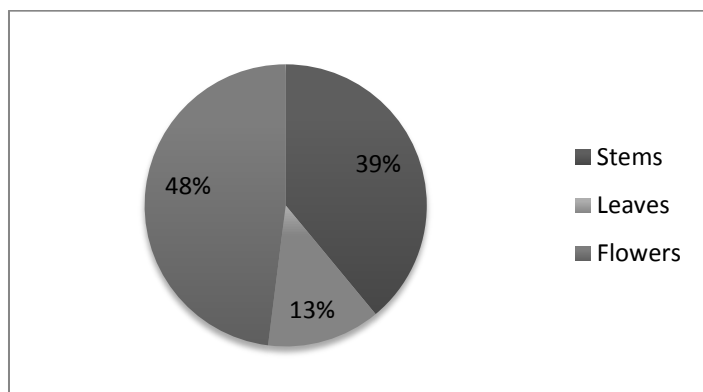


Figure 2. Percentage of *Botrytis* isolates obtained from different tissues of symptomless *Taraxacum officinale* plants (N=23).

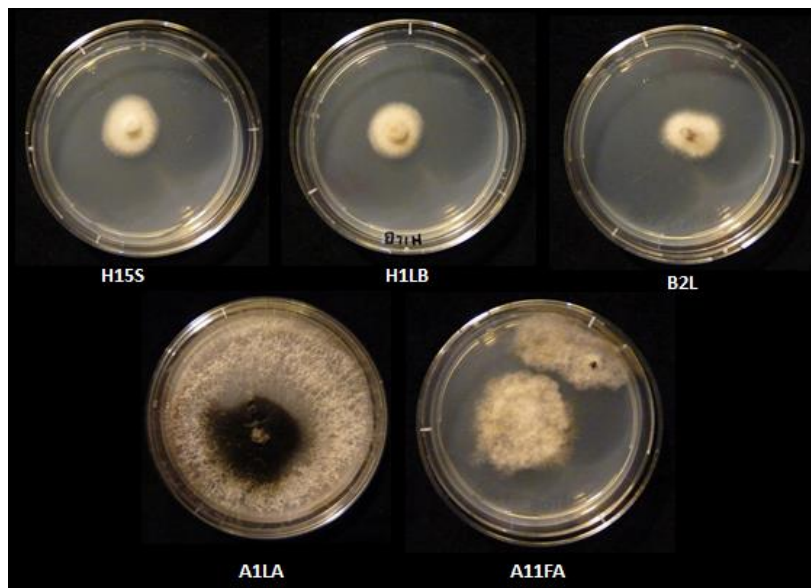


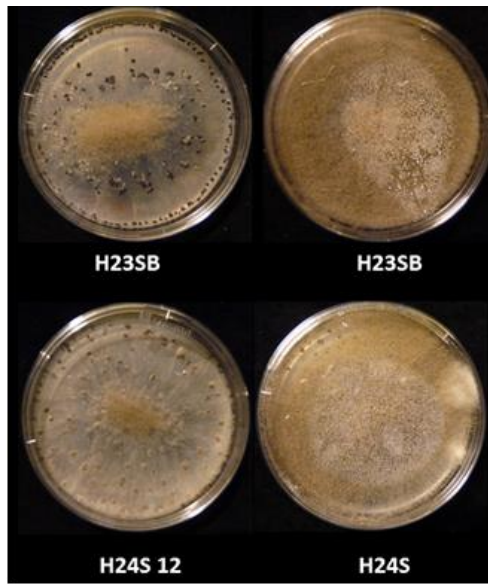
Figure 3. Morphology of unlikely *Botrytis* cultures (7 days old) on PDA with a positive identification by using QuickStix strips immunoassay test for *Botrytis* spp (Envirologix).

3.2 *Botrytis* species associated to *Taraxacum officinale*

The immunoassay test detected 23 isolates as *Botrytis* sp.. The phylogenetic analysis based on the *HSP60* gene of 12 of these isolates namely, B1LA, V17S, H24S, H23SA, H23SB, V5F, V9S, B24F, A24F, V18F, A14L and A25SA clustered them in the clade containing members; 3 different genotypes were found that showed the strongest phylogenetic association with *B. cinerea*; the genotype B1LA clustered closest to *B. pelargonii* strain 459.50; whereas V17S and a group of 5 strains (H24S, H23SA, H23SB, V5F, V9S) clustered as different genotypes together with *B. cinerea* strain MUCL87 (Figure 4, 5). Three other genotypes were distinguished that were associated with *B. pseudocinerea* they were B24F, A24F and a cluster of 3 isolates (V18F, A14L and A25SA) (Figure 4, 5). A set of 5 genotypes was chosen for further phylogenetic analysis.

The phylogenetic analysis based on *G3PDH* showed similar results; 4 of the genotypes clustered together with the species *B. cinerea* and 1 genotype was clustered with the species *B. pseudocinerea* (Figure 6). These results, were consistent with the phylogenetic analysis based *RPB2* and on all 3 genes (Figures 6, 7, 8). Unfortunately, for the phylogenetic analyses based on *RPB2* and all 3 genes the genotype V18F identified as *B. pseudocinerea* was excluded due to the low quality of the sequence data generated.

A

*B. cinerea*

B

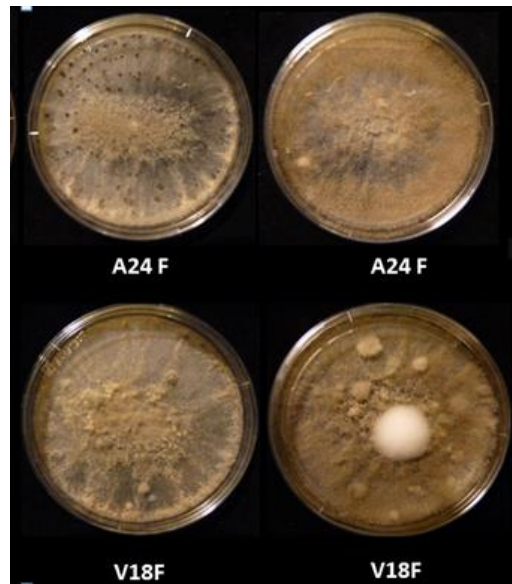
*B. pseudocinerea*

Figure 4. Cultures of *Botrytis* species *pseudocinerea* (A) and *cinerea* (B) on malt extract agar isolated from symptomless dandelion plants (*Taraxacum officinale*). Sclerotia in cultures of 12 days old grown in darkness (left side in the pictures) and sporulating cultures of 15 days old (right side of the pictures).

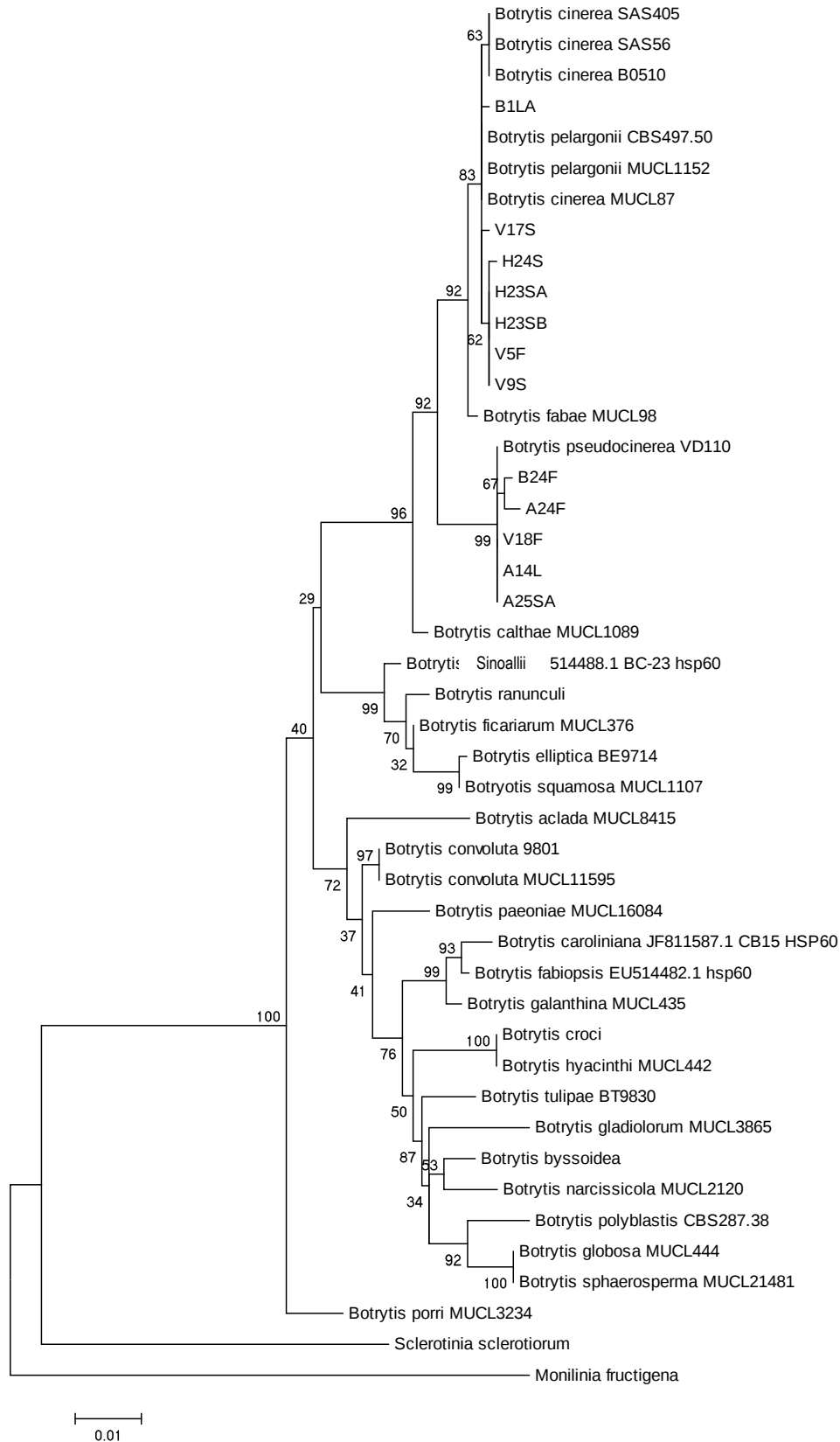


Figure 5. Phylogenetic tree based on *HSP60* gene for *Botrytis* species isolated from symptomless dandelion plants (*Taraxacum officinale*)

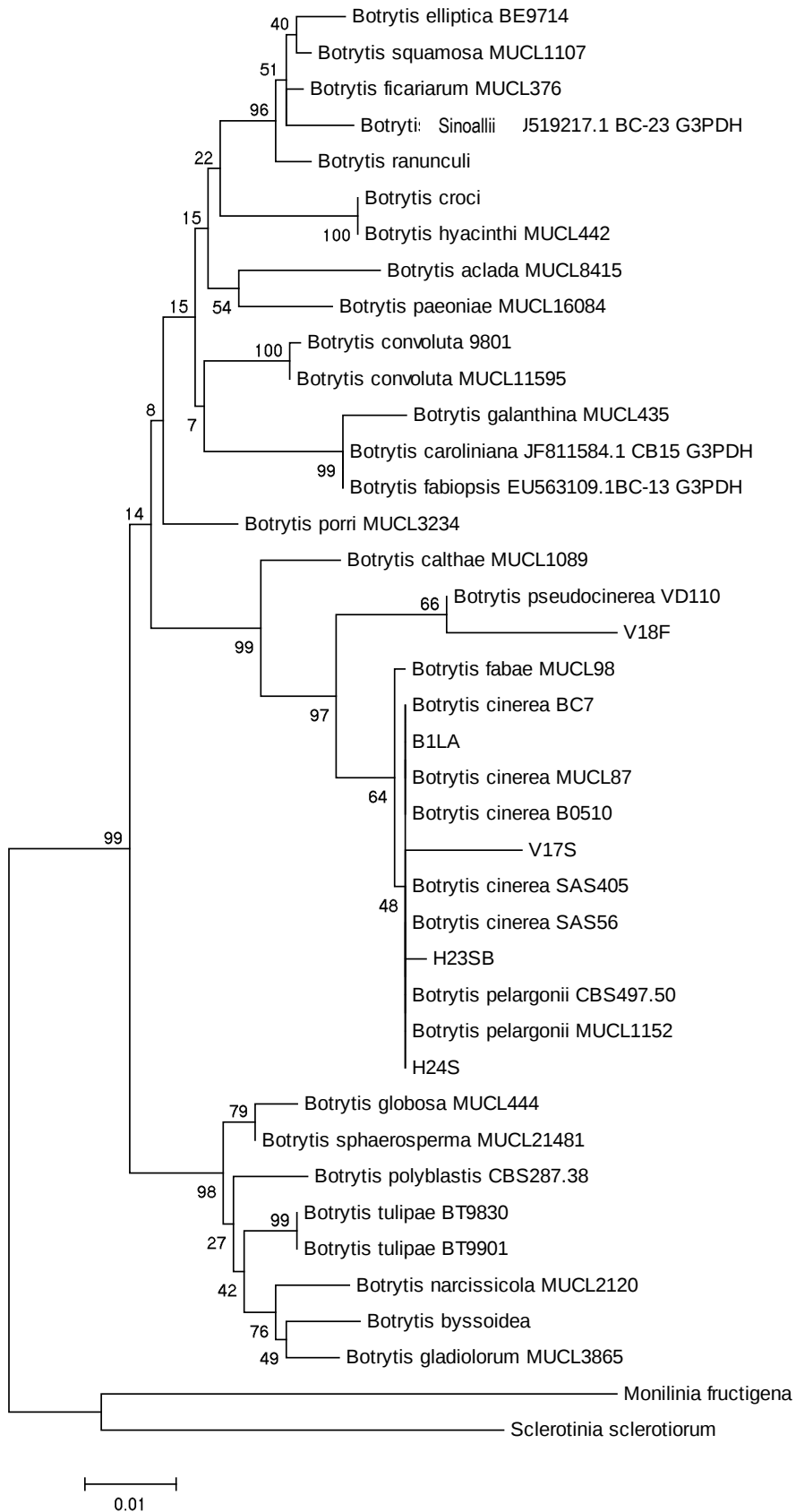


Figure 6. Phylogenetic tree based on *G3PDH* gene for *Botrytis* species isolated from symptomless dandelion plants (*Taraxacum officinale*)

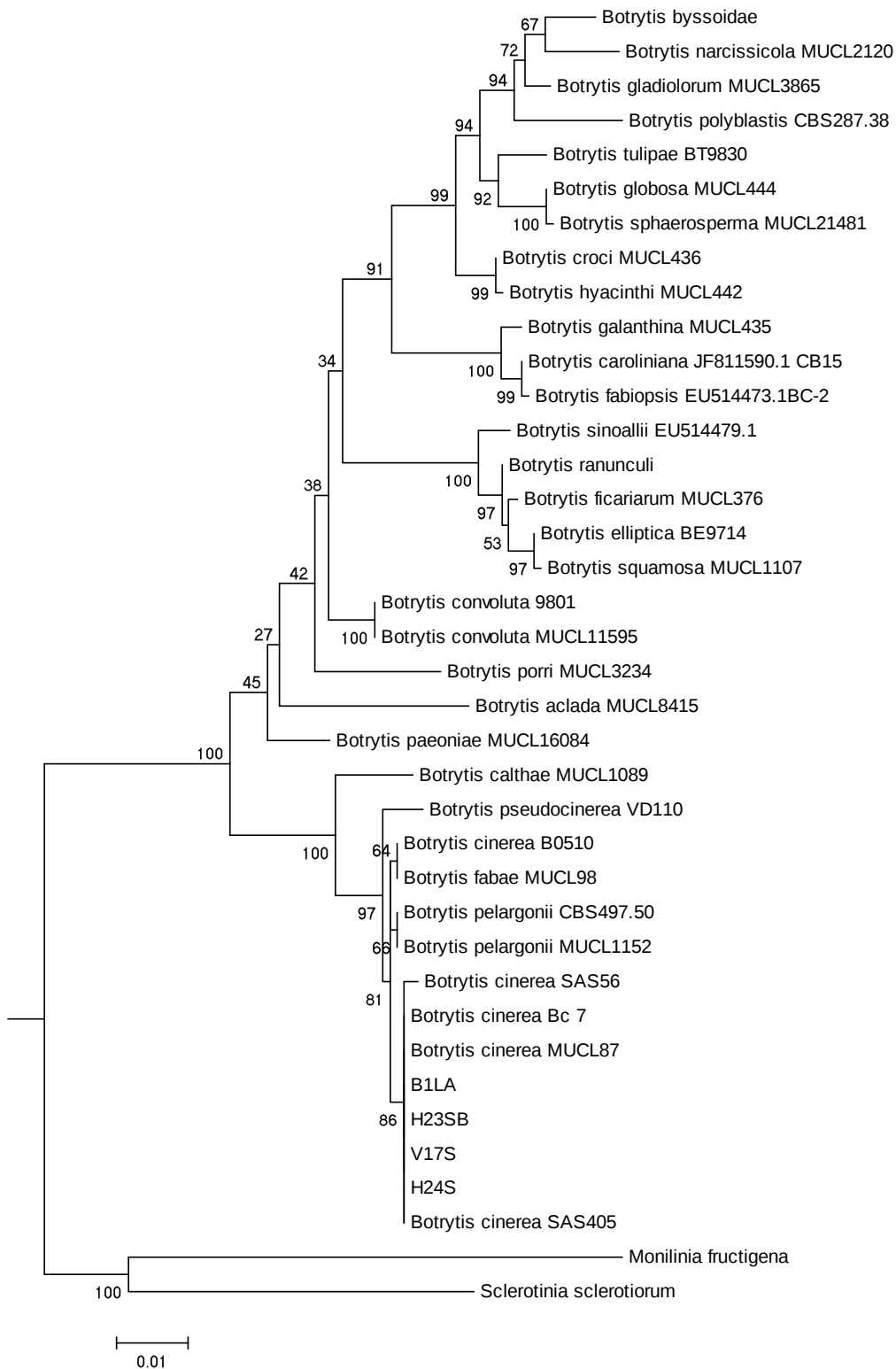


Figure 7. Phylogenetic tree based on *RPB2* gene for *Botrytis* species isolated from symptomless dandelion plants (*Taraxacum officinale*)

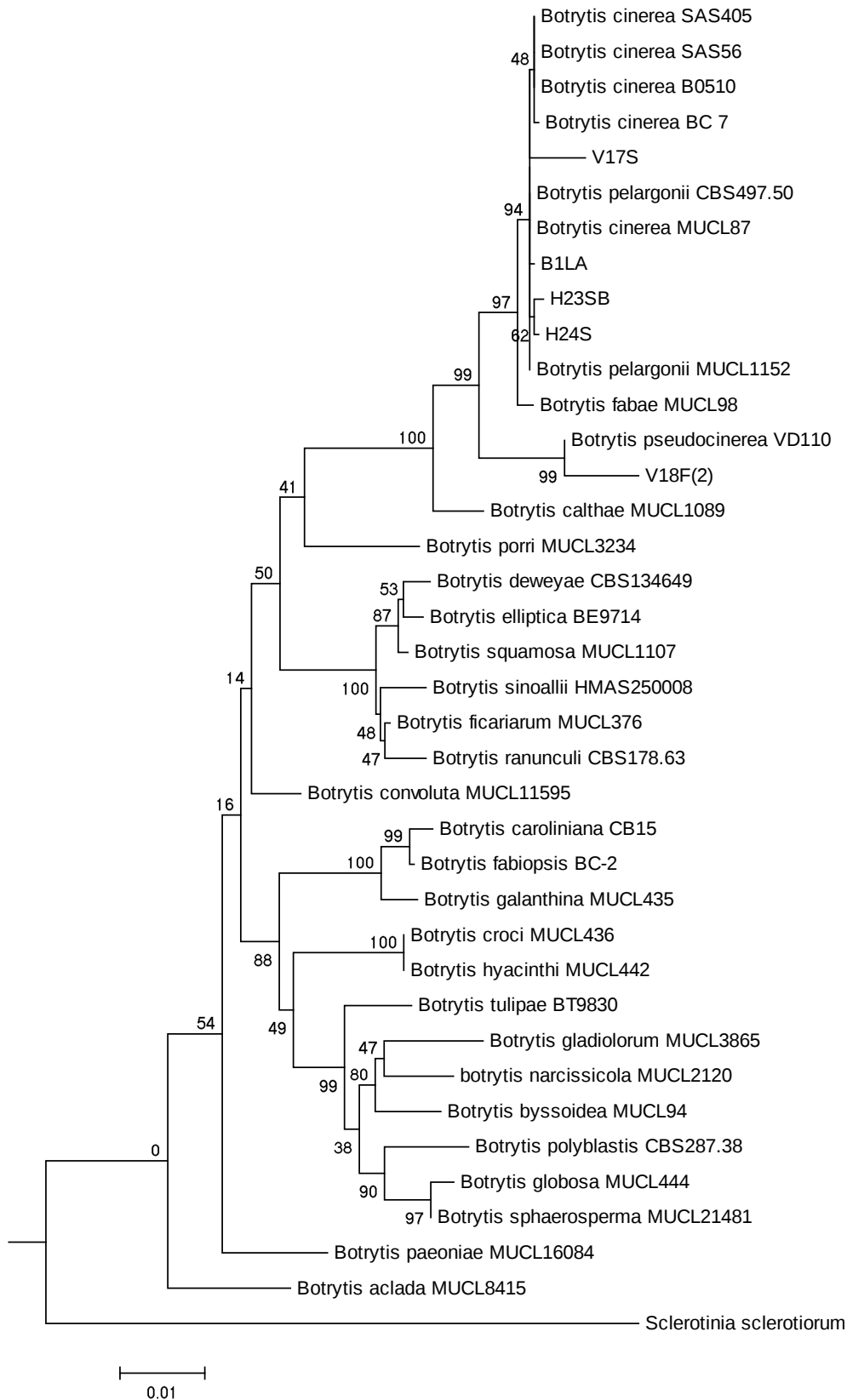


Figure 8. Phylogenetic tree based on *HSP60* and *G3PDH* genes of *Botrytis* species isolated from symptomless dandelion plants (*Taraxacum officinale*).

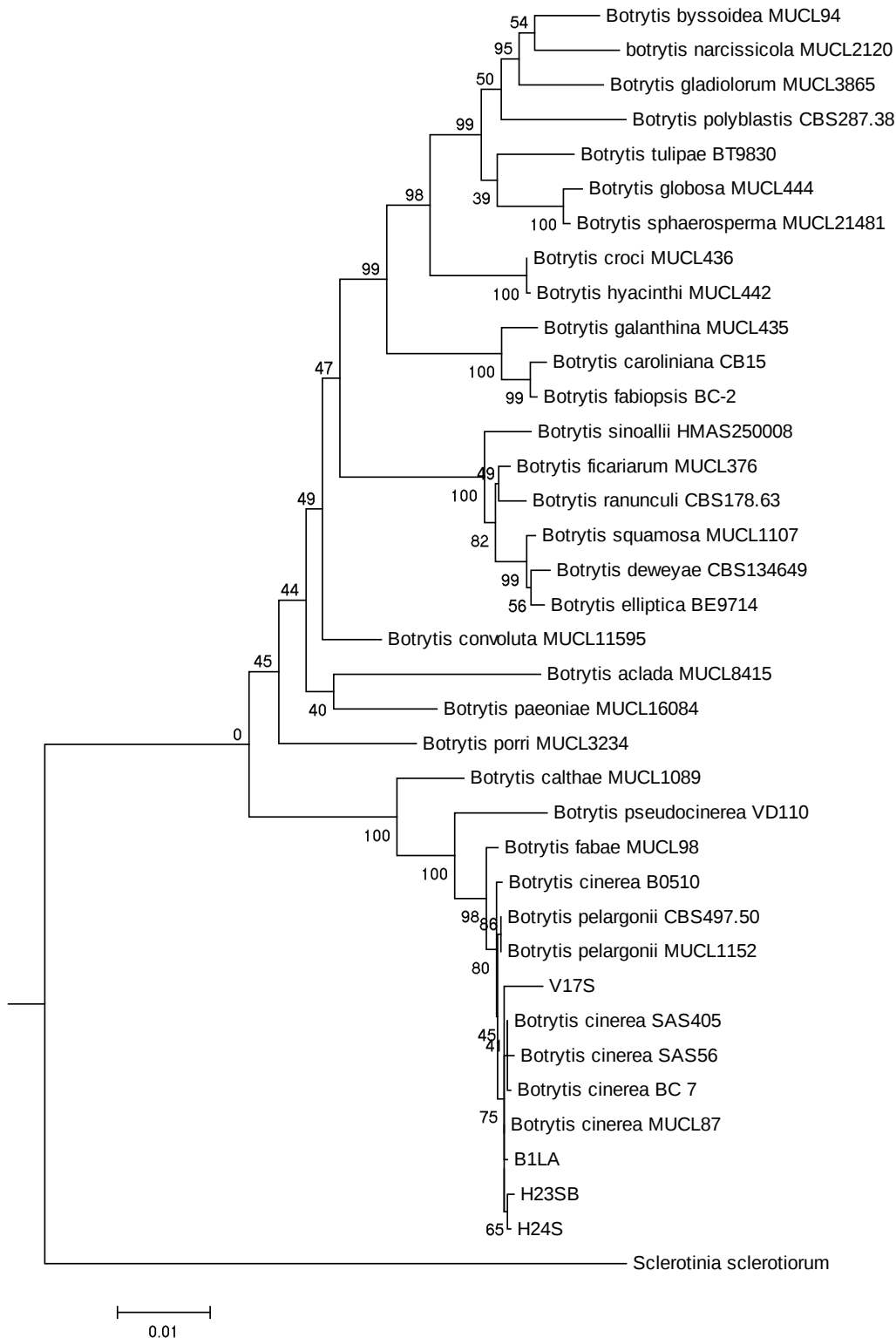
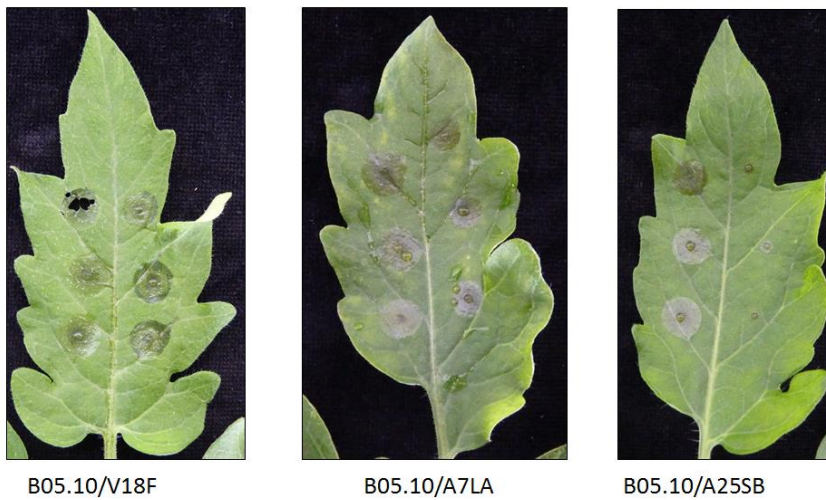


Figure 9. Phylogenetic tree based on *HSP60*, *G3PDH* and *RPB2* genes of *Botrytis* species isolated from symptomless dandelion plants (*Taraxacum officinale*)

Pathogenicity assay

Inoculation of tomato leaves and *N. benthamiana* plants demonstrated that 5 of the 6 testing *Botrytis* sp. isolates had the capacity to cause infective expanding lesions. In some of the isolates, lesions were comparable to those caused by *Botrytis cinerea* strain B05.10 employed as control (Figure 10, Table 3). Among the *Botrytis* isolates tested, inoculations with A25SB resulted in smaller lesions which developed slower compared with the other isolates inoculated.

A



B

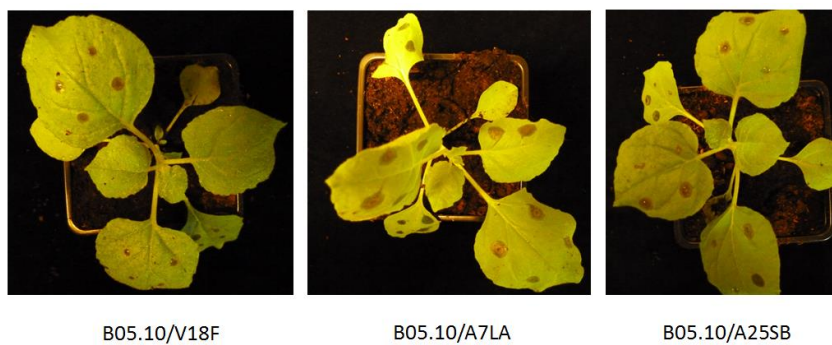


Figure 10. Lesion development (24hpi) on (A) tomato leaflets (cv. Moneymaker) and (B) *N. benthamiana* leaves inoculated (10^6 spores/ml) with *Botrytis cinerea* B05.10 (control) (leaf left side) and *Botrytis* sp. (leaf right side) isolated from symptomless *Taraxacum officinale* plants.

Table 3. Infective lesions (mm) on tomato (*S. lycopersicum*) and *N. benthamiana* leaves, produced by *Botrytis* spp. isolated from symptomless *Taraxacum officinale* plants.

<i>Botrytis</i> isolates	Tomato	<i>N. benthamiana</i>
	Lesion diameter (mm)	Lesion diameter (mm)
<i>B. cinerea</i> strain B05.10 (control)	18	14
<i>Botrytis pseudocinerea</i> (V18F)	16	10
<i>Botrytis cinerea</i> (V17S)	15	14
<i>Botrytis cinerea</i> (B1LA)	13	12
<i>Botrytis</i> sp A7LA	13	12
<i>Botrytis</i> sp B18F	12	13
<i>Botrytis</i> sp A25SB	10	7

Discussion

Botrytis cinerea is known as a polyphagous necrotrophic pathogen of plants which kills the host's cells by secreting toxic compounds and degrading enzymes, and inducing a programmed cell death by the host itself (van Kan, 2006; Choquer et al., 2007; Shlezinger et al., 2011). However, recent studies show a different “side” of this pathogen, in which it behaves as an endophyte and causes symptomless systemic infections in some plants (Sowley et al., 2010; Shipunov et al., 2008). Our results confirm the flexibility of this pathogen to behave as an endophyte. Dandelion plants investigated for endophytic colonization by *Botrytis* species resulted in isolation of different strains, which were identified by sequencing as *B. cinerea* and *B. pseudocinera*. In this study, the limited set of isolates sequenced made it difficult to elucidate if *Botrytis* species less related to *B. cinerea* were also present.

The flexible behaviour by *Botrytis* species agrees with a phylogenetic study of some fungal lineages in which it was determined that an interchangeable switch between a necrotrophic and endophytic behaviour occurred multiple times (Delaye et al., 2013). In contrast, a biotrophic condition in pathogen fungi was considered a more stable trait (Delaye et al., 2013).

Endophytes have commonly been defined as organisms which live inside plants without causing symptoms of disease (Carroll, 1988). However, this definition should be considered as a temporary status which includes a variety of microorganisms with different life history strategies (Schulz and Boyle, 2006). Endophytes can display variable associations which range from mutualism, commensalism, latent pathogenicity, and exploitation (Saikkonen et al.,

1998; Schulz and Boyle, 2006). The flexibility of these associations depends on the genetic dispositions of the two partners, their developmental stage and nutritional status as well as environmental factors (Schulz and Boyle 2006). For example, van Kan et al. (2014) suggest that a reduced genetic diversity in cultivated *Hemerocallis* hybrids could be a significant factor in the abrupt occurrence of a foliar disease called “spring sickness” that emerged from a switch of *B. deweyae* from endophytic to pathogenic infections in these plants. In grape, kiwi, strawberry and other hosts, *B. cinerea* may produce dormant infections which are then activated by the onset of senescence or stress of the host tissue (Sowley et al., 2010).

In the present study, the pathogenicity tests carried out on tomato and *N. benthamiana* with 5 of the 6 strains isolated from symptomless dandelion plants revealed that these strains were able to degrade plant tissues and cause the typical expanding disease lesions observed in aggressive infections by *Botrytis* pathogens. These results were similar to those reported by Sowley et al. 2010 in which *Botrytis cinerea* isolated from symptomless lettuce plants and later drop-inoculated on detached lettuce leaf pieces developed typical disease lesions. Endophytes and pathogens can have many of the same virulence factors which are required to infect and colonize the host (Schulz and Boyle, 2006). However, if the fungal virulence and the defensive response (which could include defensive metabolites and a general defence) by the plant are balanced, this interaction is asymptomatic (Schulz and Boyle, 2006). The conditions under which endophytic symptomless colonization by *Botrytis* species occurs have not yet been elucidated (Sowley et al. 2010; van Kan et al., 2014). Sowley et. al. (2010) suggest that during endophytic colonization by *B. cinerea* in lettuce, toxic compounds are not produced or are maintained at low levels to minimize host plant defence responses, and also mentions that active and aggressive infections by *B. cinerea* are only found in ripened fruits and wounded plant tissues. Van Kan et al. (2014) suggest that the switch from one lifestyle to another as described in *Botrytis* sp. is controlled by a mechanism which prevents or induce the expression of toxic compounds and degrading enzymes.

Overall this work we consider some information from some of the isolates is still lacking. Among the cultures identified as *Botrytis* by the immunoassay 5 isolates were considered unlikely members of the *Botrytis* genus due to their distinct morphology. However, the immunoassay test employed in this work is thought to be highly specific, therefore, obtaining information of these isolates by sequencing and a morphological description could confirm our assumption. Similarly, information by sequencing of the isolate A25SB could contribute

to explain if its reduced aggression to produce disease lesions is due to its adaptation as an endophyte or due to species particularities.

This study supports previous research in which the flexibility of *Botrytis* species to behave as necrotrophic pathogens or endophytes was established (Shipunov et al., 2008; Sowley et al. 2010), and highlights the necessity to generate new knowledge about the complex interactions which *Botrytis* species display with plants as well as the need to elucidate what could be the key factors capable of modulating such contrasting behaviours in these fungal species.

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Appendices

Appendix 1.

Table 1. Sampling sites of *Taraxacum officinale* for *Botrytis* isolation in Wageningen

	Sites	Latitude	Longitude
1	Bornsesteeg 6721 Bennekom	51.995452	5.653710
2	Hollandseweg 6705 BC Wageningen	51.985182	5.689045
3	Arboretum de Dreijen 6703 BL Wageningen	51.967068	5.677716
4	Veerdam 6703 PA, Wageningen	51.963223	5.688082

Appendix 2.

Multiple sequence alignment (Clustal 2.1) of *Botrytis* spp. based on consensus sequences of *HSP60*, *G3PDH*, *RPB2* genes

HSP60

CLUSTAL 2.1 multiple sequence alignment

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V17S      -----TTTAAGATACGACATGGATATCTGGTGATT 30
B1LA      -----TGGTGATT 8
H23SA     -----CCACGTCGCTCTAAGATACGACATGGATATCTGGTGATT 39
V9S       -----GCTCTAAGATACGACATGGATATCTGGTGATT 32
H24S      -----TCCACGTCGCTCTAAGATACGACATGGATATCTGGTGATT 40
V5F       -----ATT 3
H23SB     -----CAGTCCACGTCGCTTTAAGATACGACATGGATATCTGGTGATT 43
V18F      CTATGTGTGGGTGACTTCAGTCCCATCGCTCTAAGATACGACATAGATATTTGGTGATT 60
A24F      -----
B24F      -----
A14L      -----
A25SA     -----
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V17S      GTAAACTGATCATGTTTTTAATTTAGGAGCTCAAATTCGGTGTTGAGGGCAGAGCAGCTC 90
B1LA      GTAAACTGATCATGTTTTTAATTTAGGAGCTCAAATTCGGTGTTGAGGGCAGAGCAGCTC 68
H23SA     GTAAACTGATCATGTTTTTAATTTAGGAGCTCAAATTCGGTGTTGAGGGCAGAGCAGCTC 99
V9S       GTAAACTGATCATGTTTTTAATTTAGGAGCTCAAATTCGGTGTTGAGGGCAGAGCAGCTC 92
H24S      GTAAACTGATCATGTTTTTAATTTAGGAGCTCAAATTCGGTGTTGAGGGCAGAGCAGCTC 100
V5F       GTAAACTGATCATGTTTTTAATTTAGGAGCTCAAATTCGGTGTTGAGGGCAGAGCAGCTC 63
H23SB     GTAAACTGATCATGTTTTTAATTTAGGAGCTCAAATTCGGTGTTGAGGGCAGAGCAGCTC 103
V18F      GTAAACTGATCATGTTTTTAATTTAGGAGCTCAAATTCGGTGTTGAGGGCAGAGCAGCTC 120
A24F      -----TGAACCTGTGTTTAAATTTATGAGCTCAAATTCGGGTGTTGAGGGCAGAGCAGCTC 54
B24F      ----ACTGATCATGTTTTTAATTTAGGAGCTCGAATTCGGGGTTGAGGGCAGAGCAGCTC 56
A14L      GTAAACTGATCATGTTTTTAATTTAGGAGCTCAAATTCGGTGTTGAGGGCAGAGCAGCTC 60
A25SA     --AAACTGATCATGTTTTTAATTTAGGAGCTCAAATTCGGTGTTGAGGGCAGAGCAGCTC 58
          ***.*.*** ***** *****.****** * *****
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V17S      TTCTTGCTGGTGTTGAGACTTTGGCAAAAGCTGTTGCTACAACCTTGGGTCCCCAAAGGCC 150
B1LA      TTCTTGCTGGTGTTGAGACTTTGGCAAAAGCTGTTGCTACAACCTTGGGTCCCCAAAGGCC 128
H23SA     TTCTTGCTGGTGTTGAGACTTTGGCAAAAGCTGTTGCTACAACCTTGGGTCCCCAAAGGCC 159
V9S       TTCTTGCTGGTGTTGAGACTTTGGCAAAAGCTGTTGCTACAACCTTGGGTCCCCAAAGGCC 152
H24S      TTCTTGCTGGTGTTGAGACTTTGGCAAAAGCTGTTGCTACAACCTTGGGTCCCCAAAGGCC 160
V5F       TTCTTGCTGGTGTTGAGACTTTGGCAAAAGCTGTTGCTACAACCTTGGGTCCCCAAAGGCC 123
H23SB     TTCTTGCTGGTGTTGAGACTTTGGCAAAAGCTGTTGCTACAACCTTGGGTCCCCAAAGGCC 163
V18F      TTCTTGCTGGTGTTGAGACTTTGGCAAAAGCTGTTGCTACAACCTTGGGTCCCCAAAGGCC 180
A24F      TTCTTGCTGGTGTTGAGACTTTGGCAAAAGCTGTTGCTACAACCTTGGGTCCCCAAAGGCC 114
B24F      TTCTTGCTGGTGTTGAGACTTTGGCAAAAGCTGTTGCTACAACCTTGGGTCCCCAAAGGCC 116
A14L      TTCTTGCTGGTGTTGAGACTTTGGCAAAAGCTGTTGCTACAACCTTGGGTCCCCAAAGGCC 120
A25SA     TTCTTGCTGGTGTTGAGACTTTGGCAAAAGCTGTTGCTACAACCTTGGGTCCCCAAAGGCC 118
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V17S      GAAATGTTCTTATTGAGTCAGCATATGGCTCCCCAAAGATCACTAAAGGTTTGCAAAACT 210
B1LA      GAAATGTTCTTATTGAGTCAGCATATGGCTCCCCAAAGATCACTAAAGGTTTGCAAAACT 188
H23SA     GAAATGTTCTTATTGAGTCAGCATATGGCTCCCCAAAGATCACTAAAGGTTTGCAAAACT 219
V9S       GAAATGTTCTTATTGAGTCAGCATATGGCTCCCCAAAGATCACTAAAGGTTTGCAAAACT 212
H24S      GAAATGTTCTTATTGAGTCAGCATATGGCTCCCCAAAGATCACTAAAGGTTTGCAAAACT 220
V5F       GAAATGTTCTTATTGAGTCAGCATATGGCTCCCCAAAGATCACTAAAGGTTTGCAAAACT 183
H23SB     GAAATGTTCTTATTGAGTCAGCATATGGCTCCCCAAAGATCACTAAAGGTTTGCAAAACT 223
V18F      GAAATGTTCTTATTGAGTCAGCATATGGCTCTCCAAAGATCACTAAAGGTTTGTGAAACT 240
A24F      GAAATGTTCTTATTGAGTCAGCATATGGCTCTCCAAAGATCACTAAAGGTTTGTGAAACT 174
B24F      GAAATGTTCTTATTGAGTCAGCATATGGCTCTCCAAAGATCACTAAAGGTTTGTGAAACT 176
A14L      GAAATGTTCTTATTGAGTCAGCATATGGCTCTCCAAAGATCACTAAAGGTTTGTGAAACT 180
A25SA     GAAATGTTCTTATTGAGTCAGCATATGGCTCTCCAAAGATCACTAAAGGTTTGTGAAACT 178
          *****
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V17S	CCCGGCTACCTAGTTTCAAGATTCTAATTATTGGTGAATAGATGGTGTAAACCGTTGCCAG	270
B1LA	CCCGGCTACCTAGTTTCAAGATTCTAATTATTGGTGAATAGATGGTGTAAACCGTTGCCAG	248
H23SA	CCCGGCTACCTAGTTTCAAGATTCTAATTATTGGTGAATAGATGGTGTAAACCGTTGCCAG	279
V9S	CCCGGCTACCTAGTTTCAAGATTCTAATTATTGGTGAATAGATGGTGTAAACCGTTGCCAG	272
H24S	CCCGGCTACCTAGTTTCAAGATTCTAATTATTGGTGAATAGATGGTGTAAACCGTTGCCAG	280
V5F	CCCGGCTACCTAGTTTCAAGATTCTAATTATTGGTGAATAGATGGTGTAAACCGTTGCCAG	243
H23SB	CCCGGCTACCTAGTTTCAAGATTCTAATTATTGGTGAATAGATGGTGTAAACCGTTGCCAG	283
V18F	CCCGGCTACCTAGTTTCAAGATTCTAATTATTGATGAATAGATGGTGTAAACCGTTGCCAG	300
A24F	CCCGGCTACCTAGTTTCAAGATTCTAATTATTGATGAATAGATGGTGTAAACCGTTGCCAG	234
B24F	CCCGGCTACCTAGTTTCAAGATTCTAATTATTGATGAATAGATGGTGTAAACCGTTGCCAG	236
A14L	CCCGGCTACCTAGTTTCAAGATTCTAATTATTGATGAATAGATGGTGTAAACCGTTGCCAG	240
A25SA	CCCGGCTACCTAGTTTCAAGATTCTAATTATTGATGAATAGATGGTGTAAACCGTTGCCAG	238

V17S	AGCTATTTCCCTCAAGGACAAATTCGAGAACCTCGGTGCTAGACTTATCCAAGATGTTGC	330
B1LA	AGCTATTTCCCTCAAGGACAAATTCGAGAACCTCGGTGCTAGACTTATCCAAGATGTTGC	308
H23SA	AGCTATTTCCCTCAAGGACAAATTCGAGAACCTCGGTGCTAGACTTATCCAAGATGTTGC	339
V9S	AGCTATTTCCCTCAAGGACAAATTCGAGAACCTCGGTGCTAGACTTATCCAAGATGTTGC	332
H24S	AGCTATTTCCCTCAAGGACAAATTCGAGAACCTCGGTGCTAGACTTATCCAAGATGTTGC	340
V5F	AGCTATTTCCCTCAAGGACAAATTCGAGAACCTCGGTGCTAGACTTATCCAAGATGTTGC	303
H23SB	AGCTATTTCCCTCAAGGACAAATTCGAGAACCTCGGTGCTAGACTTATCCAAGATGTTGC	343
V18F	AGCTATTTCCCTCAAGGACAAATTCGAGAACCTCGGTGCTAGACTTATCCAAGATGTTGC	360
A24F	AGCTATTTCCCTCAAGGACAAATTCGAGAACCTCGGTGCTAGACTTATCCAAGATGTTGC	294
B24F	AGCTATTTCCCTCAAGGACAAATTCGAGAACCTCGGTGCTAGACTTATCCAAGATGTTGC	296
A14L	AGCTATTTCCCTCAAGGACAAATTCGAGAACCTCGGTGCTAGACTTATCCAAGATGTTGC	300
A25SA	AGCTATTTCCCTCAAGGACAAATTCGAGAACCTCGGTGCTAGACTTATCCAAGATGTTGC	298

V17S	CTCGAAAACCAACGAGACCGCTGGTGATGGAACCACAACCGCTACTGTCCTTGCTAAATC	390
B1LA	CTCGAAAACCAACGAGACCGCTGGTGATGGAACCACAACCGCTACTGTCCTTGCTAAATC	368
H23SA	CTCGAAAACCAACGAGACCGCTGGTGATGGAACCACAACCGCTACTGTCCTTGCTAAATC	399
V9S	CTCGAAAACCAACGAGACCGCTGGTGATGGAACCACAACCGCTACTGTCCTTGCTAAATC	392
H24S	CTCGAAAACCAACGAGACCGCTGGTGATGGAACCACAACCGCTACTGTCCTTGCTAAATC	400
V5F	CTCGAAAACCAACGAGACCGCTGGTGATGGAACCACAACCGCTACTGTCCTTGCTAAATC	363
H23SB	CTCGAAAACCAACGAGACCGCTGGTGATGGAACCACAACCGCTACTGTCCTTGCTAAATC	403
V18F	CTCGAAAACCAACGAGACCGCTGGTGATGGAACCACAACCGCTACTGTCCTTGCTAAATC	420
A24F	CTCGAAAACCAACGAGACCGCTGGTGATGGAACCACAACCGCTACTGTCCTTGCTAAATC	354
B24F	CTCGAAAACCAACGAGACCGCTGGTGATGGAACCACAACCGCTACTGTCCTTGCTAAATC	356
A14L	CTCGAAAACCAACGAGACCGCTGGTGATGGAACCACAACCGCTACTGTCCTTGCTAAATC	360
A25SA	CTCGAAAACCAACGAGACCGCTGGTGATGGAACCACAACCGCTACTGTCCTTGCTAAATC	358

V17S	TATTTTCTCCGAGACCGTAAAGAACGTCGCCGAGGATGCAACCCAATGGACTTGCGCAG	450
B1LA	TATTTTCTCCGAGACCGTAAAGAACGTCGCCGAGGATGCAACCCAATGGACTTGCGCAG	428
H23SA	TATTTTCTCCGAGACCGTAAAGAACGTCGCCGAGGATGCAACCCAATGGACTTGCGCAG	459
V9S	TATTTTCTCCGAGACCGTAAAGAACGTCGCCGAGGATGCAACCCAATGGACTTGCGCAG	452
H24S	TATTTTCTCCGAGACCGTAAAGAACGTCGCCGAGGATGCAACCCAATGGACTTGCGCAG	460
V5F	TATTTTCTCCGAGACCGTAAAGAACGTCGCCGAGGATGCAACCCAATGGACTTGCGCAG	423
H23SB	TATTTTCTCCGAGACCGTAAAGAACGTCGCCGAGGATGCAACCCAATGGACTTGCGCAG	463
V18F	TATTTTCTCCGAGACCGTAAAGAACGTCGCCGAGGATGCAACCCAATGGACTTGCGCAG	480
A24F	TATTTTCTCCGAGACCGTAAAGAACGTCGCCGAGGATGCAACCCAATGGACTTGCGCAG	414
B24F	TATTTTCTCCGAGACCGTAAAGAACGTCGCCGAGGATGCAACCCAATGGACTTGCGCAG	416
A14L	TATTTTCTCCGAGACCGTAAAGAACGTCGCCGAGGATGCAACCCAATGGACTTGCGCAG	420
A25SA	TATTTTCTCCGAGACCGTAAAGAACGTCGCCGAGGATGCAACCCAATGGACTTGCGCAG	418

V17S	AGGTACTCAAGCTGCCGTGGAGGCCGTTGTTGAGTTTTTGCAAAAGAACAAGCGTGATAT	510
B1LA	AGGTACCCAAGCTGCCGTGGAGGCCGTTGTTGAGTTTTTGCAAAAGAACAAGCGTGATAT	488
H23SA	AGGTACCCAAGCTGCCGTGGAGGCCGTTGTTGAGTTTTTGCAAAAGAACAAGCGTGATAT	519
V9S	AGGTACCCAAGCTGCCGTGGAGGCCGTTGTTGAGTTTTTGCAAAAGAACAAGCGTGATAT	512
H24S	AGGTACCCAAGCTGCCGTGGAGGCCGTTGTTGAGTTTTTGCAAAAGAACAAGCGTGATAT	520
V5F	AGGTACCCAAGCTGCCGTGGAGGCCGTTGTTGAGTTTTTGCAAAAGAACAAGCGTGATAT	483
H23SB	AGGTACCCAAGCTGCCGTGGAGGCCGTTGTTGAGTTTTTGCAAAAGAACAAGCGTGATAT	523
V18F	AGGTACCCAAGCTGCCGTGGAGGCCGTTGTTGAGTTTTTGCAAAAGAACAAGCGTGATAT	540
A24F	AGGTACCCAAGCTGCCGTGGAGGCCGTTGTTGAGTTTTTGCAAAAGAACAAGCGTGATAT	474
B24F	AGGTACCCAAGCTGCCGTGGAGGCCGTTGTTGAGTTTTTGCAAAAGAACAAGCGTGATAT	476
A14L	AGGTACCCAAGCTGCCGTGGAGGCCGTTGTTGAGTTTTTGCAAAAGAACAAGCGTGATAT	480
A25SA	AGGTACCCAAGCTGCCGTGGAGGCCGTTGTTGAGTTTTTGCAAAAGAACAAGCGTGATAT	478
***** .*****		
V17S	CACAACAAGCGAGGAAATCGCACAAAGTTGCGACTATCAGTGCAAACGGTGATACCCACAT	570
B1LA	CACAACAAGCGAGGAAATCGCACAAAGTTGCGACTATCAGTGCAAACGGTGATACCCACAT	578
H23SA	CACAACAAGCGAGGAAATCGCACAAAGTTGCGACTATCAGTGCAAACGGTGATACCCACAT	579
V9S	CACAACAAGCGAGGAAATCGCACAAAGTTGCGACTATCAGTGCAAACGGTGATACCCACAT	572
H24S	CACAACAAGCGAGGAAATCGCACAAAGTTGCGACTATCAGTGCAAACGGTGATACCCACAT	580
V5F	CACAACAAGCGAGGAAATCGCACAAAGTTGCGACTATCAGTGCAAACGGTGATACCCACAT	543
H23SB	CACAACAAGCGAGGAAATCGCACAAAGTTGCGACTATCAGTGCAAACGGTGATACCCACAT	583
V18F	CACAACCAGCGAGGAAATCGCACAAAGTTGCGACTATTAGTGCAAACGGTGATACCCACAT	600
A24F	CACAACCAGCGAGGAAATCGCACAAAGTTGCGACTATTAGTGCAAACGGTGATACCCACAT	534
B24F	CACAACCAGCGAGGAAATCGCACAAAGTTGCGACTATTAGTGCAAACGGTGATACCCACAT	536
A14L	CACAACCAGCGAGGAAATCGCACAAAGTTGCGACTATTAGTGCAAACGGTGATACCCACAT	540
A25SA	CACAACCAGCGAGGAAATCGCACAAAGTTGCGACTATTAGTGCAAACGGTGATACCCACAT	538
***** .*****		
V17S	CGGAAAATTGATTGCCAACGCTATGGAAAAGGTTGGAAAGGAAGGTGTTATCACAGTTAA	630
B1LA	CGGAAAATTGATTGCCAACGCTATGGAAAAGGTTGGAAAGGAAGGTGTTATCACAGTTAA	608
H23SA	CGGAAAATTGATTGCCAACGCTATGGAAAAGGTTGGAAAGGAAGGTGTTATCACAGTTAA	639
V9S	CGGAAAATTGATTGCCAACGCTATGGAAAAGGTTGGAAAGGAAGGTGTTATCACAGTTAA	632
H24S	CGGAAAATTGATTGCCAACGCTATGGAAAAGGTTGGAAAGGAAGGTGTTATCACAGTTAA	640
V5F	CGGAAAATTGATTGCCAACGCTATGGAAAAGGTTGGAAAGGAAGGTGTTATCACAGTTAA	603
H23SB	CGGAAAATTGATTGCCAACGCTATGGAAAAGGTTGGAAAGGAAGGTGTTATCACAGTTAA	643
V18F	CGGAAAATTGATTGCCAACGCTATGGAAAAGGTTGGAAAGGAAGGTGTTATCACAGTTAA	660
A24F	CGGAAAATTGATTGCCAACGCTATGGAAAAGGTTGGAAAGGAAGGTGTTATCACAGTTAA	594
B24F	CGGAAAATTGATTGCCAACGCTATGGAAAAGGTTGGAAAGGAAGGTGTTATCACAGTTAA	596
A14L	CGGAAAATTGATTGCCAACGCTATGGAAAAGGTTGGAAAGGAAGGTGTTATCACAGTTAA	600
A25SA	CGGAAAATTGATTGCCAACGCTATGGAAAAGGTTGGAAAGGAAGGTGTTATCACAGTTAA	598

V17S	GGAAGGAAAGACCATGGAGGACGAACTCGATATTACCGAGGGAATGAGATTTGACCGCGG	690
B1LA	GGAAGGAAAGACCATGGAGGACGAACTCGATATTACCGAGGGAATGAGATTTGACCGCGG	668
H23SA	GGAAGGAAAGACCATGGAGGACGAACTCGATATTACCGAGGGAATGAGATTTGACCGCGG	699
V9S	GGAAGGAAAGACCATGGAGGACGAACTCGATATTACCGAGGGAATGAGATTTGACCGCGG	692
H24S	GGAAGGAAAGACCATGGAGGACGAACTCGATATTACCGAGGGAATGAGATTTGACCGCGG	700
V5F	GGAAGGAAAGACCATGGAGGACGAACTCGATATTACCGAGGGAATGAGATTTGACCGCGG	663
H23SB	GGAAGGAAAGACCATGGAGGACGAACTCGATATTACCGAGGGAATGAGATTTGACCGCGG	703
V18F	GGAAGGTAAGACCATGGAGGATGAACTCGATATTACCGAGGGAATGAGATTTGACCGCGG	720
A24F	GGAAGGTAAGACCATGGAGGATGAACTCGATATTACCGAGGGAATGAGATTTGACCGCGG	654
B24F	GGAAGGTAAGACCATGGAGGATGAACTCGATATTACCGAGGGAATGAGATTTGACCGCGG	656
A14L	GGAAGGTAAGACCATGGAGGATGAACTCGATATTACCGAGGGAATGAGATTTGACCGCGG	660
A25SA	GGAAGGTAAGACCATGGAGGATGAACTCGATATTACCGAGGGAATGAGATTTGACCGCGG	658
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V17S	TTATGTTTCCCCATACTTCATCACCGATACCAAGTCGCAAAAGGTGGAATTCGAGAAGCC	750
B1LA	TTATGTTTCCCCATACTTCATCACCGATACCAAGTCGCAAAAGGTGGAATTCGAGAAGCC	728
H23SA	TTATGTTTCTCCATACTTCATCACCGATACCAAGTCGCAAAAGGTGGAATTCGAGAAGCC	759
V9S	TTATGTTTCTCCATACTTCATCACCGATACCAAGTCGCAAAAGGTGGAATTCGAGAAGCC	752
H24S	TTATGTTTCTCCATACTTCATCACCGATACCAAGTCGCAAAAGGTGGAATTCGAGAAGCC	760
V5F	TTATGTTTCTCCATACTTCATCACCGATACCAAGTCGCAAAAGGTGGAATTCGAGAAGCC	723
H23SB	TTATGTTTCTCCATACTTCATCACCGATACCAAGTCGCAAAAGGTGGAATTCGAGAAGCC	763
V18F	TTATGTTTCCCCATACTTCATCACCGATACCAAGTCGCAAAAGGTGGAATTCGAGAAGCC	780
A24F	TTATGTTTCCCCATACTTCATCACCGATACCAAGTCGCAAAAGGTGGAATTCGAGAAGCC	714
B24F	TTATGTTTCCCCATACTTCATCACCGATACCAAGTCGCAAAAGGTGGAATTCGAGAAGCC	716
A14L	TTATGTTTCCCCATACTTCATCACCGATACCAAGTCGCAAAAGGTGGAATTCGAGAAGCC	720
A25SA	TTATGTTTCCCCATACTTCATCACCGATACCAAGTCGCAAAAGGTGGAATTCGAGAAGCC	718

V17S	ATTGATTCTCCTTTCTGAGAAGAAGATTTCAAACGTCCAAGATATTATCCCAGCACTTGA	810
B1LA	ATTGATTCTCCTTTCTGAGAAGAAGATTTCAAACGTCCAAGATATTATCCCAGCACTTGA	788
H23SA	ATTGATTCTCCTTTCTGAGAAGAAGATTTCAAACGTCCAAGATATTATCCCAGCACTTGA	819
V9S	ATTGATTCTCCTTTCTGAGAAGAAGATTTCAAACGTCCAAGATATTATCCCAGCACTTGA	812
H24S	ATTGATTCTCCTTTCTGAGAAGAAGATTTCAAACGTCCAAGATATTATCCCAGCACTTGA	820
V5F	ATTGATTCTCCTTTCTGAGAAGAAGATTTCAAACGTCCAAGATATTATCCCAGCACTTGA	783
H23SB	ATTGATTCTCCTTTCTGAGAAGAAGATTTCAAACGTCCAAGATATTATCCCAGCACTTGA	823
V18F	ATTGATTCTCCTTTCTGAGAAGAAGATTTCAAACGTCCAAGATATTATCCCAGCACTTGA	840
A24F	ATTGATTCTCCTTTCTGAGAAGAAGATTTCAAACGTCCAAGATATTATCCCAGCACTTGA	774
B24F	ATTGATTCTCCTTTCTGAGAAGAAGATTTCAAACGTCCAAGATATTATCCCAGCACTTGA	776
A14L	ATTGATTCTCCTTTCTGAGAAGAAGATTTCAAACGTCCAAGATATTATCCCAGCACTTGA	780
A25SA	ATTGATTCTCCTTTCTGAGAAGAAGATTTCAAACGTCCAAGATATTATCCCAGCACTTGA	778

V17S	GGCGTCTACTCAACTTCGTCGTCCTTTGGTCATCATTGCTGAAGATATCGATGGAGAAGC	870
B1LA	GGCGTCTACTCAACTTCGTCGTCCTTTGGTCATCATTGCTGAAGATATCGATGGAGAAGC	848
H23SA	GGCGTCTACTCAACTTCGTCGTCCTTTGGTCATCATTGCTGAAGATATCGATGGAGAAGC	879
V9S	GGCGTCTACTCAACTTCGTCGTCCTTTGGTCATCATTGCTGAAGATATCGATGGAGAAGC	872
H24S	GGCGTCTACTCAACTTCGTCGTCCTTTGGTCATCATTGCTGAAGATATCGATGGAGAAGC	880
V5F	GGCGTCTACTCAACTTCGTCGTCCTTTGGTCATCATTGCTGAAGATATCGATGGAGAAGC	843
H23SB	GGCGTCTACTCAACTTCGTCGTCCTTTGGTCATCATTGCTGAAGATATCGATGGAGAAGC	883
V18F	GGCGTCTACTCAACTTCGCCGTCCTTTGGTCATCATTGCTGAAGATATCGATGGAGAAGC	900
A24F	GGCGTCTACTCAACTTCGCCGTCCTTTGGTCATCATTGCTGAAGATATCGATGGAGAAGC	834
B24F	GGCGTCTACTCAACTTCGCCGTCCTTTGGTCATCATTGCTGAAGATATCGATGGAGAAGC	836
A14L	GGCGTCTACTCAACTTCGCCGTCCTTTGGTCATCATTGCTGAAGATATCGATGGAGAAGC	840
A25SA	GGCGTCTACTCAACTTCGCCGTCCTTTGGTCATCATTGCTGAAGATATCGATGGAGAAGC	838

V17S	TCTCGCAGTATGCATTCTTAACAAGCTCCGTGGTCAACTCCAAGTTGCCGCTGTCAAGGC	930
B1LA	TCTCGCAGTATGCATTCTTAACAAGCTCCGTGGTCAACTCCAAGTTGCCGCTGTCAAGGC	908
H23SA	TCTCGCAGTATGCATTCTTAACAAGCTCCGTGGTCAACTCCAAGTTGCCGCTGTCAAGGC	939
V9S	TCTCGCAGTATGCATTCTTAACAAGCTCCGTGGTCAACTCCAAGTTGCCGCTGTCAAGGC	932
H24S	TCTCGCAGTATGCATTCTTAACAAGCTCCGTGGTCAACTCCAAGTTGCCGCTGTCAAGGC	940
V5F	TCTCGCAGTATGCATTCTTAACAAGCTCCGTGGTCAACTCCAAGTTGCCGCTGTCAAGGC	903
H23SB	TCTCGCAGTATGCATTCTTAACAAGCTCCGTGGTCAACTCCAAGTTGCCGCTGTCAAGGC	943
V18F	TCTCGCTGTATGCATTCTTAACAAGCTCCGTGGTCAACTCCAAGTTGCCGCTGTCAAGGC	960
A24F	TCTCGCTGTATGCATTCTTAACAAGCTCCGTGGTCAACTCCAAGTTGCCGCTGTCAAGGC	894
B24F	TCTCGCTGTATGCATTCTTAACAAGCTCCGTGGTCAACTCCAAGTTGCCGCTGTCAAGGC	896
A14L	TCTCGCTGTATGCATTCTTAACAAGCTCCGTGGTCAACTCCAAGTTGCCGCTGTCAAGGC	900
A25SA	TCTCGCTGTATGCATTCTTAACAAGCTCCGTGGTCAACTCCAAGTTGCCGCTGTCAAGGC	898

V17S	CCCCGGTTTTCGGTGATAACCGAAAAGTCCATCCTCGGCGATCTCGGTATCTTGACCAATGC	990
B1LA	CCCCGGTTTTCGGTGATAACCGAAAAGTCCATCCTCGGCGATCTCGGTATCTTGACCAATGC	968
H23SA	CCCCGGTTTTCGGTGATAACCGAAAAGTCCATCCTCGGCGATCTCGGTATCTTGACCAATGC	999
V9S	CCCCGGTTTTCGGTGATAACCGAAAAGTCCATCCTCGGCGATCTCGGTATCTTGACCAATGC	992
H24S	CCCCGGTTTTCGGTGATAACCGAAAAGTCCATCCTCGGCGATCTCGGTATCTTGACCAATGC	1000
V5F	CCCCGGTTTTCGGTGATAACCGAAAAGTCCATCCTCGGCGATCTCGGTATCTTGACCAATGC	963
H23SB	CCCCGGTTTTCGGTGATAACCGAAAAGTCCATCCTCGGCGATCTCGGTATCTTGACCAATGC	1003
V18F	CCCCGGTTTTCGGTGATAACCGAAAAGTCCATCCTCGGCGATCTCGGTATCTTGACCAATGC	1020
A24F	CCCCGG-----	900
B24F	CCCCGG-----	902
A14L	CCCCGGTTTTCGGTGATAACCGAAAAGTTCATCCTCGGCGATCTCGGTATCTTGACCAATG-	959
A25SA	CCCCGGTTTTCGGTGATAACCGAAAAGTCCATCCTCGGCGATCTCGGTATCTT-----	949

V17S	TACCGTCTTCACTGACGAGCTTGATCTCAA-----	1020
B1LA	TACCGTCTTCACTGACGA-----	986
H23SA	TACCGTCTTCACTGACGAGCTTGATCTCA-----	1028
V9S	TACCGTCTTCACTGACGAGCTTGATCTCAA-----	1022
H24S	TACCGTCTTCACTGACGAGCTTGATCTCAAGCTCGA	1036
V5F	TACCGTCTTCACTGACGAGCTTGATCTCAAGCTCGA	999
H23SB	TACCGTCTTCACTGACGAGCTTGATCTCAAGCTCG-	1038
V18F	TACCGTCTTCACTGACGAGCTTGATCTCAAG-----	1051
A24F	-----	
B24F	-----	
A14L	-----	
A25SA	-----	

G3PDH

CLUSTAL 2.1 multiple sequence alignment

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H24S      -----TTTCCGCTATCGGAGCTCCCGCAGATATCAAGGACCCGAGCTAATTTA 48
B1LA      --ATGCCGTAAGTTTCCGCTATCGGAGCTCCCGCAGATATCAAGGACCCGAGCTAATTTA 58
V17S      --ATGCCGTAAGTTTCCGCTATCGGAGCTCCCGCAGATATCAAGGACCCGAGCTAATTTA 58
H23SB     --ATGCCGTAAGTTTCCGCTATCGGAGCTCCCGCAGATATCAAGGACCCGAGCTAATTTA 58
V18F      ATATGCCGTAAGTTTCCGCTATCGGACTTCCCGCAGATATTACGGACCCGAGCTAATTTA 60
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          *****
          *

H24S      TGTTTTGCACAGGCATACATGTTGAAGTATGATTCCACCCACGGTCAATTCAAGGGTGAC 108
B1LA      TGTTTTGCACAGGCATACATGTTGAAGTATGATTCCACCCACGGTCAATTCAAGGGTGAC 118
V17S      TGTTTTGCACAGGCATACATGTTGAAGTATGATTCCACCCACGGTCAATTCAAGGGTGAC 118
H23SB     TGTTTTGCACAGGCATACATGTTGAAGTATGATTCCACCCACGGTCAATTCAAGGGTGAC 118
V18F      TATTTTGCACAGGCATACATGTTGAAGTATGATTCCACCCACGGTCAATTCAAGGGTGAC 120
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          *****

H24S      ATCAAGGTCCTTGCCGATGGATTGGAGGTCAATGGCAAGAAGGTCAAGTTCTACACCGAG 168
B1LA      ATCAAGGTCCTTGCCGATGGATTGGAGGTCAATGGCAAGAAGGTCAAGTTCTACACCGAG 178
V17S      ATCAAGGTCCTTGCCGATGGATTGGAGGTCAATGGCAAGAAGGTCAAGTTCTACACCGAG 178
H23SB     ATCAAGGTCCTTGCCGATGGATTGGAGGTCAATGGCAAGAAGGTCAAGTTCTACACCGAG 178
V18F      ATCAAGGTCCTTGCCGATGGATTGGAGGTCAATGGCAAGAAGGTCAAGTTCTACACCGAG 180
          *****

H24S      AGAGACCCAGCCAACATCCCATGGGCTGAGTCTGAGGCATACTACGTCGTCGAGTCCACC 228
B1LA      AGAGACCCAGCCAACATCCCATGGGCTGAGTCTGAGGCATACTACGTCGTCGAGTCCACC 238
V17S      AGAGACCCAGCCAACATCCCATGGGCTGAGTCTGAGGCATACTACGTCGTCGAGTCCACC 238
H23SB     AGAGACCCAGCCAACATCCCATGGGCTGAGTCTGAGGCATACTACGTCGTCGAGTCCACC 238
V18F      AGAGACCCAGCCAACATCCCATGGGCTGAGTCTGAGGCATACTACGTCGTCGAGTCCACC 240
          *****

H24S      GGTGTTTTTACCACCACCGAGAAGGCCAAGGCACATTTGAAGGGTGGTGCCAAGAAGGTT 288
B1LA      GGTGTTTTTACCACCACCGAGAAGGCCAAGGCACATTTGAAGGGTGGTGCCAAGAAGGTT 298
V17S      GGTGTTTTTACCACCACCGAGAAGGCCAAGGCACATTTGAAGGGTGGTGCCAAGAAGGTT 298
H23SB     GGTGTTTTTACCACCACCGAGAAGGCCAAGGCACATTTGAAGGGTGGTGCCAAGAAGGTT 298
V18F      GGTGTTTTTACCACCACCGAGAAGGCTAAGGCACATTTGAAGGGTGGTGCCAAGAAGGTT 300
          *****

H24S      GTTATCTCTGCTCCTTCTGCCGATGCCCCAATGTACGTTATGGGTGTCAACAACGAGACC 348
B1LA      GTTATCTCTGCTCCTTCTGCCGATGCCCCAATGTACGTTATGGGTGTCAACAACGAGACC 358
V17S      GTTATCTCTGCTCCTTCTGCCGATGCCCCAATGTACGTTATGGGTGTCAACAACGAGACC 358
H23SB     GTTATCTCTGCTCCTTCTGCCGATGCCCCAATGTACGTTATGGGTGTCAACAACGAGACC 358
V18F      GTTATCTCTGCTCCTTCTGCCGACGCCCAATGTACGTTATGGGTGTCAACAACGAGACC 360
          *****

H24S      TACACTGGTGATGTTGATGTTATCTCCAACGCCTCTTGACACAACCAACTGCTTGGCTCCT 408
B1LA      TACACTGGTGATGTTGATGTTATCTCCAACGCCTCTTGACACAACCAACTGCTTGGCTCCT 418
V17S      TACACTGGTGATGTTGATGTTATCTCCAACGCCTCTTGACACAACCAACTGCTTGGCTCCT 418
H23SB     TACACTGGTGATGTTGATGTTATCTCCAACGCCTCTTGACACAACCAACTGCTTGGCTCCT 418
V18F      TACACTGGTGATGTTGATGTTATCTCCAACGCCTCTTGACACAACCAACTGCTTGGCTCCT 420
          *****

H24S      CTCGCCAAGGTCATCAACGATGAGTTCACCATCATTTGAAGGTTTGATGACCACCATCCAC 468
B1LA      CTCGCCAAGGTCATCAACGATGAGTTCACCATCATTTGAAGGTTTGATGACCACCATCCAC 478
V17S      CTCGCCAAGGTCATCAACGATGAGTTCACCATCATTTGAAGGTTTGATGACCACCATCCAC 478
H23SB     CTCGCCAAGGTCATCAACGATGAGTTCACCATCATTTGAAGGTTTGATGACCACCATCCAC 478
V18F      CTCGCCAAGGTCATCAACGATGAGTTCACCATCATTTGAAGGTTTGATGACCACCATCCAC 480
          *****

H24S      TCCTACACCGCTACCCAAAAGACCGTTGATGGTCCATCCGCTAAGGATTGGCGTGGAGGA 528
B1LA      TCCTACACCGCTACCCAAAAGACCGTTGATGGTCCATCCGCTAAGGATTGGCGTGGAGGA 538
V17S      TCCTACACCGCTACCCAAAAGACCGTTGATGGTCCATCCGCTAAGGATTGGCGTGGAGGA 538
H23SB     TCCTACACCGCTACCCAAAAGACCGTTGATGGTCCATCCGCTAAGGATTGGCGTGGAGGA 538
V18F      TCCTACACCGCTACCCAAAAGACCGTTGACGGTCCATCCGCTAAGGATTGGCGTGGAGGA 540
          *****

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H24S	CGTACCGCTGCTCAAAACATCATCCCATCGAGCACCGGTGCTGCCAAGGCTGTCGGAAAG	588
B1LA	CGTACCGCTGCTCAAAACATCATCCCATCGAGCACCGGTGCTGCCAAGGCTGTCGGAAAG	598
V17S	CGTACCGCTGCTCAAAACATCATCCCATCGAGCACCGGTGCTGCCAAGGCTGTCGGAAAG	598
H23SB	CGTACCGCTGCTCAAAACATCATCCCATCGAGCACCGGTGCTGCCAAGGCTGTCGGAAAG	598
V18F	CGTACCGCTGCTCAAAACATCATCCCATCGAGCACCGGTGCTGCCAAGGCTGTCGGAAAG	600

H24S	GTCATCCCAGTCCTTAACGGCAAACCTACCGGAATGTCCATGCGTGTTCCAAGTCCAAC	648
B1LA	GTCATCCCAGTCCTTAACGGCAAACCTACCGGAATGTCCATGCGTGTTCCAAGTCCAAC	658
V17S	GTCATCCCAGTCCTTAACGGCAAACCTACCGGAATGTCCATGCGTGTTCCAAGTCCAAC	658
H23SB	GTCATCCCAGTCCTTAACGGCAAACCTACCGGAATGTCCATGCGTGTTCCAAGTCCAAC	658
V18F	GTCATCCCAGTCCTTAACGGCAAACCTACCGGAATGTCCATGCGTGTTCCAAGTCCAAC	660

H24S	GTCTCAGTTGTTGACTTGACTGTCCGCATTGAGAAGGGTGCTTCTTACGATGAGATCAAG	708
B1LA	GTCTCAGTTGTTGACTTGACTGTCCGCATTGAGAAGGGTGCTTCTTACGATGAGATCAAG	718
V17S	GTCTCAGTTGTTGACTTGACTGTCCGCATTGAGAAGGGTGCTTCTTACGATGAGATCAAG	718
H23SB	GTCTCAGTTGTTGACTTGACTGTCCGCATTGAGAAGGGTGCTTCTTACGATGAGATCAAG	718
V18F	GTCTCAGTTGTTGACTTGACTGTCCGCATTGAGAAGGGTGCTTCTTATGATGAGATTAAG	720

H24S	GCCGTCATCAAGAAGGCTGCTGATGGTCCTCTCAAGGGTAAGTTACTCCATTACTCTTTC	768
B1LA	GCCGTCATCAAGAAGGCTGCTGATGGTCCTCTCAAGGGTAAGTTACTCCATTACTCTTTC	778
V17S	GCCGTCATCAAGAAGGCTGCTGATGGTCCTCTCAAGGGTAAGTTACTCCATTACTCTTTC	778
H23SB	GCCGTCATCAAGAAGGCTGCTGATGGTCCTCTCAAGGGTAAGTTACTCCATTACTCTTTC	776
V18F	GCCGTCATCAAGAAGGCTGCTGATGGTCCTCTCAAGGGCAGGTTACTCCATCACTCTTTC	780

H24S	TTCGGCTCTAATTTGCTAATCGTAACACAGGCATATTGGCTTACACTGAGGACGATGTTG	828
B1LA	TTCGGCTCTAATTTGCTAATCGTAACACAGGCATATTGGCTTACACTGAGGACGATGTTG	838
V17S	TTCGGCTCTAATTTGCTAATCGTAACACAGGCATATTGGCTTACACTGAGGACGATGTTG	838
H23SB	-TTCGGCTCTAATTTGCTAATCGTAACACAGGCATATTGGCTTACACTGAGGACGATGTCG	835
V18F	TTTGGCTCTAATTTACTAATCATAATACAGGCATATTGGCTTACACTGAGGACGATGTTG	840
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H24S	TCTCCACTGACATGAACGG-TGACAACCA-CTCCTCCATCTT-CGATGCCA-AGGCTGGT	884
B1LA	TCTCCACTGACATGAACGG-TGACAACCA-CTCCTCCATCTT-CGATGCCA-AGGCTGGT	894
V17S	TCTCCACTGACATGAACGGT-GACAACCA-CTCCTCCATCTTCTGATGCCATAGGCTGGT	896
H23SB	TCTCCACTGACATGAACGGT-GACAACCA-CTCCTCCATCTT-CGATGCCA-AGGCTGGT	891
V18F	TCTCCACTGACATGAACGG-TGACAACCA-CTCCTCCATCTTTCGATGCCAAGGCCGGT	898

H24S	ATCTCCCTCAACGCAAACCTTCGT-----	907
B1LA	ATCTCCCTCAACGCAAACCTTCGTCAAGTTG	924
V17S	ATCTCC-----	902
H23SB	ATCTCCCTCA-----	901
V18F	ATCTCC-----	905

RPB2

CLUSTAL 2.1 multiple sequence alignment

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V17S      -----CCGCAGATTGACAACAGACGTGTACAGATACTTGCAACGTTGCGTGGAAAACAAC 55
H24S      -----ACAGACGTGTACAGATACTTGCAACGTTGCGTGGAAAACAAC 42
B1LA      CCGGTCCGCAGATTGACAACAGACGTGTACAGATACTTGCAACGTTGCGTGGAAAACAAC 60
H23SB     CCTGTCCGCAGATTGACAACAGACGTGTACAGATACTTGCAACGTTGCGTGGAAAACAAC 60
          *****

V17S      CGAGAGTTTAATTTGACTTTGGGTGTGAAATCAACAACAATCACCAACGGTCTGAAATAT 115
H24S      CGAGAGTTTAATTTGACTTTGGGTGTGAAATCAACAACAATCACCAACGGTCTGAAATAT 102
B1LA      CGAGAGTTTAATTTGACTTTGGGTGTGAAATCAACAACAATCACCAACGGTCTGAAATAT 120
H23SB     CGAGAGTTTAATTTGACTTTGGGTGTGAAATCAACAACAATCACCAACGGTCTGAAATAT 120
          *****

V17S      TCTTTGGCCACAGGTAAGTGGGGTGACCAGAAGAAGGCAGCAAGTTCTACCGCCGGAGTG 175
H24S      TCTTTGGCCACAGGTAAGTGGGGTGACCAGAAGAAGGCAGCAAGTTCTACCGCCGGAGTG 162
B1LA      TCTTTGGCCACAGGTAAGTGGGGTGACCAGAAGAAGGCAGCAAGTTCTACCGCCGGAGTG 180
H23SB     TCTTTGGCCACAGGTAAGTGGGGTGACCAGAAGAAGGCAGCAAGTTCTACCGCCGGAGTG 180
          *****

V17S      TCTCAAGTGTTGAACAGATATACCTTTGCATCCACACTTTCTCATTTGCGCCGAACCAAT 235
H24S      TCTCAAGTGTTGAACAGATATACCTTTGCATCCACACTTTCTCATTTGCGCCGAACCAAT 222
B1LA      TCTCAAGTGTTGAACAGATATACCTTTGCATCCACACTTTCTCATTTGCGCCGAACCAAT 240
H23SB     TCTCAAGTGTTGAACAGATATACCTTTGCATCCACACTTTCTCATTTGCGCCGAACCAAT 240
          *****

V17S      ACACCCATTGGACGTGATGGAAAGATCGCCAAACCTAGACAGCTGCATAATACCCATTGG 295
H24S      ACACCCATTGGACGTGATGGAAAGATCGCCAAACCTAGACAGCTGCATAATACCCATTGG 282
B1LA      ACACCCATTGGACGTGATGGAAAGATCGCCAAACCTAGACAGCTGCATAATACCCATTGG 300
H23SB     ACACCCATTGGACGTGATGGAAAGATCGCCAAACCTAGACAGCTGCATAATACCCATTGG 300
          *****

V17S      GGCTTGGTCTGTCCGGCAGAGACGCCGAAGGACAAGCTTGTGGTTTGGTTAAGAATTTG 355
H24S      GGCTTGGTCTGTCCGGCAGAGACGCCGAAGGACAAGCTTGTGGTTTGGTTAAGAATTTG 342
B1LA      GGCTTGGTCTGTCCGGCAGAGACGCCGAAGGACAAGCTTGTGGTTTGGTTAAGAATTTG 360
H23SB     GGCTTGGTCTGTCCGGCAGAGACGCCGAAGGACAAGCTTGTGGTTTGGTTAAGAATTTG 360
          *****

V17S      GCTCTGATGTGTTACGTTACAGTCGGTACGCCAAGTGATCCAATCGTTGAGTTCATGATT 415
H24S      GCTCTGATGTGTTACGTTACAGTCGGTACGCCAAGTGATCCAATCGTTGAGTTCATGATT 402
B1LA      GCTCTGATGTGTTACGTTACAGTCGGTACGCCAAGTGATCCAATCGTTGAGTTCATGATT 420
H23SB     GCTCTGATGTGTTACGTTACAGTCGGTACGCCAAGTGATCCAATCGTTGAGTTCATGATT 420
          *****

V17S      CAACGAAATATGGAAGTATTGGAGGAGTATGAACCACTCCGAGCCCCCAATGCAACAAAG 475
H24S      CAACGAAATATGGAAGTATTGGAGGAGTATGAACCACTCCGAGCCCCCAATGCAACAAAG 462
B1LA      CAACGAAATATGGAAGTATTGGAGGAGTATGAACCACTCCGAGCCCCCAATGCAACAAAG 480
H23SB     CAACGAAATATGGAAGTATTGGAGGAGTATGAACCACTCCGAGCCCCCAATGCAACAAAG 480
          *****

V17S      GTTTTCGTCAATGGTGTGTTGGGTGGTATTTCATCGAGATCCTGCTCATTTGGTCAAATGT 535
H24S      GTTTTCGTCAATGGTGTGTTGGGTGGTATTTCATCGAGATCCTGCTCATTTGGTCAAATGT 522
B1LA      GTTTTCGTCAATGGTGTGTTGGGTGGTATTTCATCGAGATCCTGCTCATTTGGTCAAATGT 540
H23SB     GTTTTCGTCAATGGTGTGTTGGGTGGTATTTCATCGAGATCCTGCTCATTTGGTCAAATGT 540
          *****

V17S      GTCCAAGATCTTCGTAGATCACACTTGATCTCTCATGAAGTTTCACTTATTCGAGAAATT 595
H24S      GTCCAAGATCTTCGTAGATCACACTTGATCTCTCATGAAGTTTCACTTATTCGAGAAATT 582
B1LA      GTCCAAGATCTTCGTAGATCACACTTGATCTCTCATGAAGTTTCACTTATTCGAGAAATT 600
H23SB     GTCCAAGATCTTCGTAGATCACACTTGATCTCTCATGAAGTTTCACTTATTCGAGAAATT 600
          *****

V17S      CGTGATAGAGAATTCAAGATTTTCACAGATGCAGGACGAGTGTGCAGACCTCTATTGGTT 655
H24S      CGTGATAGAGAATTCAAGATTTTCACAGATGCAGGACGAGTGTGCAGACCTCTATTGGTT 642
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B1LA	CGTGATAGAGAATTCAAGATTTTCACAGATGCAGGACGAGTGTGCAGACCTCTATTGGTT	660
H23SB	CGTGATAGAGAATTCAAGATTTTCACAGATGCAGGACGAGTGTGCAGACCTCTATTGGTT	660

V17S	ATTGACAATGATCCTGACAGCGCAAACAAAGGTAACCTGGTGTTGAACAAGGATCACATT	715
H24S	ATTGACAATGATCCTGACAGCGCAAACAAAGGTAACCTGGTGTTGAACAAGGATCACATT	702
B1LA	ATTGACAATGATCCTGACAGCGCAAACAAAGGTAACCTGGTGTTGAACAAGGATCACATT	720
H23SB	ATTGACAATGATCCTGACAGCGCAAACAAAGGTAACCTGGTGTTGAACAAGGATCACATT	720

V17S	CGCCGTCTGGAGGATGATCAGTTGCTACCAGCAAACATGGATAAGGATGAGAAAGTAAGA	775
H24S	CGCCGTCTGGAGGATGATCAGTTGCTACCAGCAAACATGGATAAGGATGAGAAAGTAAGA	762
B1LA	CGCCGTCTGGAGGATGATCAGTTGCTACCAGCAAACATGGATAAGGATGAGAAAGTAAGA	780
H23SB	CGCCGTCTGGAGGATGATCAGTTGCTACCAGCAAACATGGATAAGGATGAGAAAGTAAGA	780

V17S	AACGGATACTATGGATTCCAAGGTTTGATTAATGACGGTGTGGTTGAGTACCTGGATGCC	835
H24S	AACGGATACTATGGATTCCAAGGTTTGATTAATGACGGTGTGGTTGAGTACCTGGATGCC	822
B1LA	AACGGATACTATGGATTCCAAGGTTTGATTAATGACGGTGTGGTTGAGTACCTGGATGCC	840
H23SB	AACGGATACTATGGATTCCAAGGTTTGATTAATGACGGTGTGGTTGAGTACCTGGATGCC	840

V17S	GAGGAAGAAGAGACTGTCATGATTACAATGACACCTGAAGATCTGGACATCTCCCGACAG	895
H24S	GAGGAAGAAGAGACTGTCATGATTACAATGACACCTGAAGATCTGGACATCTCCCGACAG	882
B1LA	GAGGAAGAAGAGACTGTCATGATTACAATGACACCTGAAGATCTGGACATCTCCCGACAG	900
H23SB	GAGGAAGAAGAGACTGTCATGATTACAATGACACCTGAAGATCTGGACATCTCCCGACAG	900

V17S	CTTCAGGCTGGTTACCAAATTCGTCCTGACGAAAGTGGTGATTGTAACAAGCGTGTTAAG	955
H24S	CTTCAGGCTGGTTACCAAATTCGTCCTGACGAAAGTGGTGATTGTAACAAGCGTGTTAAG	942
B1LA	CTTCAGGCTGGTTACCAAATTCGTCCTGACGAAAGTGGTGATTGTAACAAGCGTGTTAAG	960
H23SB	CTTCAGGCTGGTTACCAAATTCGTCCTGACGAAAGTGGTGATTGTAACAAGCGTGTTAAG	960

V17S	GCACCTATCAATCCAACCTGCTCATGTCTGGACTCATTGTGAAATTCATCCAAGTATGATC	1015
H24S	GCACCTATCAATCCAACCTGCTCATGTCTGGACTCATTGTGAAATTCATCCAAGTATGATC	1002
B1LA	GCACCTATCAATCCAACCTGCTCATGTCTGGACTCATTGTGAAATTCATCCAAGTATGATC	1020
H23SB	GCACCTATCAATCCAACCTGCTCATGTCTGGACTCATTGTGAAATTCATCCAAGTATGATC	1020

V17S	TTGGGTATCTGCGCAAGCATTATTCCCTTCCCGGATCACAATCAGGTAAGCTTTAAGTTC	1075
H24S	TTGGGTATCTGCGCAAGCATTATTCCCTTCCCGGATCACAATCAGGTAAGCTTTA-----	1057
B1LA	TTGGGTATCTGCGCAAGCATTATTCCCTTCCCGGATCACAATCAGGTAAGCTTTAAGTTC	1080
H23SB	TTGGGTATCTGCGCAAGCATTATTCCCTTCCCGGATCACAATCAGGTAAGCTTTAAGTTC	1080

V17S	AATGAATTTATTTG	1089
H24S	-----	
B1LA	AATGAATTTAT---	1091
H23SB	AATGAA-----	1086