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Environmental risk limits for various chlorobenzenes

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Abstract

Environmental Risk Limits for several chlorobenzenes

This report documents the derivation of environmental risk limits for several chlorobenzenes in water, groundwater, soil and air. The following substances were selected: monochlorobenzene, dichlorobenzenes and tetrachlorobenzenes. Chlorobenzenes are used as intermediates in the production of other substances. High concentrations of chlorobenzenes are hazardous to the environment.

For deriving the ERLs, RIVM used up-to-date ecotoxicological data in combination with the methodology as required by the European Water Framework Directive. The newly derived ERLs are lower than earlier derived ERLs. However, monitoring data from the river Rhine in the period 2001 – 2006 show only few cases of exceedance of the new ERLs.

ERLs were not derived for the sediment compartment, because sorption to sediment is below the trigger value to derive such risk limits.

Based on the comparable ecotoxicity of the individual isomers of dichlorobenzenes and tetrachlorobenzenes, ERLs for these substances were derived based on combined datasets and the use of sum limits is proposed. The respective chlorobenzenes may occur simultaneously in the environment and an additive effect cannot be excluded.

Environmental risk limits form the scientific basis on which the Interdepartmental Steering Group for substances sets environmental quality standards. The government uses these quality standards for carrying out the national policy concerning substances and the European Water Framework Directive.

Key words:

environmental risk limits, maximum permissible concentration, maximum acceptable concentration, monochlorobenzene, dichlorobenzene, tetrachlorobenzene

Rapport in het kort

Milieurisicogrenzen voor verscheidene chloorbenzenen

Het RIVM heeft milieurisicogrenzen afgeleid voor een serie chloorbenzenen in water, grondwater, bodem en lucht. De groep stoffen omvat monochloorbenzeen, dichloorbenzenen en tetrachloorbenzenen. Ze worden gebruikt als tussenproduct om andere stoffen te maken. Te hoge concentraties van deze stoffen zijn schadelijk voor het milieu.

Voor dit onderzoek zijn actuele ecotoxicologische gegevens gebruikt, gecombineerd met de methodiek die is voorgeschreven door de Europese Kaderrichtlijn Water (KRW). De nieuwe milieurisicogrenzen zijn lager dan de nu geldende afgeleide normen. Dit komt omdat nu niet alleen de directe schadelijke effecten zijn onderzocht, maar ook de indirekte effecten op mensen en op vogels en zoogdieren door het eten van vis. Tussen 2001 en 2006 zijn de stoffen een enkele keer aangetroffen in de Rijn, maar het is niet waarschijnlijk dat de nieuw afgeleide risicogrenzen langdurig zijn overschreden.

Voor de waterbodem zijn geen milieurisicogrenzen afgeleid, omdat de stoffen naar verwachting nauwelijks aan de waterbodem binden.

Normaal gesproken worden de milieurisicogrenzen afgeleid op basis van de eigenschappen van individuele stoffen. Van de di- en tetrachloorbenzenen bestaan echter verschillende vormen die gelijktijdig voorkomen en een vergelijkbare toxiciteit hebben. Daarom is voor deze stoffen een zogeheten somnorm afgeleid, die voorkomt dat de effecten van individuele stoffen worden gestapeld. Deze somnorm is gebaseerd op de *gezamenlijke* gegevens en effecten van vergelijkbare stoffen.

Milieurisicogrenzen zijn niet bindend, maar zijn de wetenschappelijke basis waarop de Nederlandse Interdepartementale Stuurgroep Stoffen de milieukwaliteitsnormen vaststelt. De overheid hanteert deze normen bij de uitvoering van het nationale stoffenbeleid en de KRW.

Trefwoorden:

milieurisicogrenzen, MTR, MAC, dichloorbenzeen, tetrachloorbenzeen

Preface

The goal of this report is to derive risk limits that protect both man and the environment. This is done in accordance with the methodology of the Water Framework Directive (WFD) that is incorporated in the present INS methodology, following the Guidance for the derivation of environmental risk limits within the INS framework (Van Vlaardingen and Verbruggen, 2007).

The results presented in this report have been discussed by the members of the scientific advisory group for the project ‘International and National Environmental Quality Standards for Substances in the Netherlands’ (WK-INS). This advisory group provides non binding scientific advice on the final draft of a report in order to advise the Dutch Steering Group for Substances on the scientific merits of the report.

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Summary

Environmental risk limits are derived using ecotoxicological, physico-chemical and human toxicological data. They represent environmental concentrations of a substance offering different levels of protection to man and ecosystems. It should be noted that the ERLs are scientifically derived values. They serve as advisory values for the Dutch Steering Group for Substances, which was appointed to set the Environmental Quality Standards (EQSs) from these ERLs. ERLs should thus be considered as preliminary values that do not have official status.

In this report, the risk limits negligible concentration (NC), maximum permissible concentration (MPC), maximum acceptable concentration for ecosystems (MAC_{eco}) and serious risk concentration for ecosystems (SRC_{eco}) are derived for monochlorobenzene, dichlorobenzenes (1,2-dichlorobenzene, 1,3-dichlorobenzene, and 1,4-dichlorobenzene) and tetrachlorobenzenes (1,2,3,4-tetrachlorobenzene, 1,2,3,5-tetrachlorobenzene and 1,2,4,5-tetrachlorobenzene) in water, groundwater, soil and air. No risk limits were derived for the sediment compartment because the triggers to derive such limits were not met.

For the derivation of the MPC and MAC_{eco} for water, the methodology used is in accordance with the Water Framework Directive. This methodology is based on the Technical Guidance Document on risk assessment for new and existing substances and biocides (European Commission (Joint Research Centre), 2003). For the NC and the SRC_{eco} , the guidance developed for the project ‘International and National Environmental Quality Standards for Substances in the Netherlands’ was used (Van Vlaardingen and Verbruggen, 2007). An overview of the derived environmental risk limits is given in Table 1.

Because of the small difference in the toxicity of the individual isomers, the risk limits for di- and tetrachlorobenzenes refer to a sum limit. In this approach, the sum of the concentrations of the individual di- or tetrachlorobenzenes should not exceed the ERLs for the group. It should be noted that the sum limit only applies when all isomers are monitored. Furthermore, chlorobenzenes may occur simultaneously in the environment and they are considered to have additive effects. Therefore, a toxic unit approach is recommended when monitoring data are compared with risk limits.

Table 1. Derived NC, MPC, MAC_{eco}, and SRC_{eco} values.

ERL	Unit	Substance	NC	MPC	MAC_{eco}	SRC_{eco}
Water	µg/L	monochlorobenzene	0.32	32	40	1.9×10^3
Marine	µg/L	monochlorobenzene	3.2×10^{-2}	3.2	4.0	1.9×10^3
Soil	µg/kg	monochlorobenzene	4.3	4.3×10^2	n.a.	2.6×10^4
Groundwater	µg/L	monochlorobenzene	0.32	32	n.a.	1.9×10^3
Air	µg/m ³	monochlorobenzene	n.a.	5.0×10^2	n.a.	n.a.
Drinking water	µg/L	monochlorobenzene	n.a.	2.1×10^2	n.a.	n.a.
Water	µg/L	dichlorobenzenes	6.9×10^{-2}	6.9	20	6.8×10^2
Marine	µg/L	dichlorobenzenes	2.0×10^{-2}	2.0	2.0	6.8×10^2
Soil	µg/kg	dichlorobenzenes	3.1	3.1×10^2	n.a.	2.0×10^4
Groundwater	µg/L	dichlorobenzenes	0.20	20	n.a.	6.8×10^2
Air	µg/m ³	dichlorobenzenes	n.a.	n.d.	n.a.	n.a.
Drinking water	µg/L	dichlorobenzenes	n.a.	3.8×10^2	n.a.	n.a.
Water	µg/L	tetrachlorobenzenes	1.6×10^{-5}	1.6×10^{-3}	0.13	58
Marine	µg/L	tetrachlorobenzenes	1.6×10^{-5}	1.6×10^{-3}	1.3×10^{-2}	58
Soil	µg/kg	tetrachlorobenzenes	2.6×10^{-2}	2.6	n.a.	2.5×10^4
Groundwater	µg/L	tetrachlorobenzenes	4.6×10^{-3}	0.46	n.a.	58
Air	µg/m ³	tetrachlorobenzenes	n.a.	n.d.	n.a.	n.a.
Drinking water	µg/L	tetrachlorobenzenes	n.a.	0.74	n.a.	n.a.

n.d. = not derived.

n.a. = not applicable.

1 Introduction

1.1 Project framework

In this report, environmental risk limits (ERLs) for surface water (freshwater and marine), soil, groundwater and air are derived for monochlorobenzene, dichlorobenzenes (1,2-dichlorobenzene, 1,3-dichlorobenzene and 1,4-dichlorobenzene) and tetrachlorobenzenes (1,2,3,4-tetrachlorobenzene, 1,2,3,5-tetrachlorobenzene and 1,2,4,5-tetrachlorobenzene). The following ERLs are considered (VROM, 2004):

- Negligible Concentration (NC) – concentration at which effects to ecosystems are expected to be negligible and functional properties of ecosystems must be safeguarded fully. It defines a safety margin which should exclude combination toxicity. The NC is derived by dividing the MPC (see next bullet) by a factor of 100.
- Maximum Permissible Concentration (MPC) – concentration in an environmental compartment at which:
 1. no effect to be rated as negative is to be expected for ecosystems;
 - 2a no effect to be rated as negative is to be expected for humans (for non-carcinogenic substances);
 - 2b for humans, no more than a probability of 10^{-6} per year of death can be calculated (for carcinogenic substances). Within the scope of the Water Framework Directive, a probability of 10^{-6} on a life-time basis is used.

Within the scope of the Water Framework Directive, the AA-EQS (equivalent to the MPC) specifically refers to long-term exposure.

- Maximum Acceptable Concentration (MAC_{eco}) – concentration protecting aquatic ecosystems for effects due to short-term exposure or concentration peaks.
- Serious Risk Concentration (SRC_{eco}) – concentration at which possibly serious ecotoxicological effects are to be expected.

These ERLs serve as advisory values that are used by the Steering Group for Substances to set environmental quality standards (EQS) for various policy purposes. EQSs are all legally and non-legally binding standards that are used in Dutch environmental policy.

1.2 Selection of substances

ERLs are derived for several chlorobenzenes (Table 2), which are selected by the Netherlands in the scope of ‘(Inter)national and National Environmental Quality Standards for Substances in the Netherlands’ (INS).

Table 2. Selected compounds.

Compound	CAS number
monochlorobenzene	108-90-7
1,2-dichlorobenzene	95-50-1
1,3-dichlorobenzene	541-73-1
1,4-dichlorobenzene	106-46-7
1,2,3,4-tetrachlorobenzene	634-66-2
1,2,3,5-tetrachlorobenzene	634-90-2
1,2,4,5-tetrachlorobenzene	95-94-3

2 Methods

2.1 Guidance followed for this project

In this report ERLs are derived following the methodology of the project ‘International and National Environmental Quality Standards for Substances in the Netherlands’ (INS). This INS-guidance by Van Vlaardingen and Verbruggen (2007) is in accordance with the guidance by Lepper (2005), which forms part of the Priority Substances Daughter Directive (2006/0129 (COD)) amending the Water Framework Directive (2000/60/EC).

The WFD guidance applies to the derivation of ERLs for water and sediment, for both the freshwater and marine compartment. The WFD guidance introduces a new ERL, which is the Maximum Acceptable Concentration (MAC_{eco}), a concentration that protects aquatic ecosystems from adverse effects caused by short-term exposure or concentration peaks.

Further, two MPC values are considered for the water compartment that are based on a human toxicological risk limit (TL_{hh}), which might be an ADI or TDI, etc. Discerned are (1) the $MPC_{hh, food, water}$, which is the concentration in water that should protect humans against adverse effects from the substance via fish and shellfish consumption; (2) the $MPC_{dw, water}$, which is the concentration in water that should protect humans against adverse effects of the substance in drinking water. Note that each of these two MPCs is allowed to contribute only 10% to the TL_{hh} .

Two other MPCs are derived for the water compartment, based on ecotoxicological data. These are (1) the $MPC_{eco, water}$, which is based on direct aquatic ecotoxicological data and (2) the $MPC_{sp, water}$, which is derived in case secondary poisoning in the environment is thought to be of concern. It is important to note that MPC and NC derivation integrates both ecotoxicological data and a human toxicological threshold value. The height of the final ‘environmental risk limit’ can be determined by either one of these protection objectives.

2.2 Data collection and evaluation

An on-line literature search was performed using TOXLINE (literature from 1985 to 2001) and Current contents (literature from 1997 to 2007). In addition to this, all relevant references in the RIVM e-tox base and EPA’s ECOTOX database were evaluated. All relevant toxicity data are reported in the Appendices.

Ecotoxicity studies were screened for relevant endpoints (i.e., those endpoints that have consequences at the population level of the test species). All ecotoxicity and bioaccumulation tests were then thoroughly evaluated with respect to the validity (scientific reliability) of the study according to the criteria of Klimisch (1997). A detailed description of the evaluation procedure is given in the INS-Guidance sections 2.2.2 and 2.3.2. In short, the following reliability indices were assigned:

- Ri 1: Reliable without restriction
‘Studies or data ... generated according to generally valid and/or internationally accepted testing guidelines (preferably performed according to GLP) or in which the test parameters documented are based on a specific (national) testing guideline ... or in which all parameters described are closely related/comparable to a guideline method.’

- Ri 2: Reliable with restrictions
 'Studies or data ... (mostly not performed according to GLP), in which the test parameters documented do not totally comply with the specific testing guideline but are sufficient to accept the data or in which investigations are described which cannot be subsumed under a testing guideline but are nevertheless well documented and scientifically acceptable.'
- Ri 3: Not reliable
 'Studies or data ... in which there are interferences between the measuring system and the test substance or in which organisms/test systems were used that are not relevant in relation to the exposure (e.g., non physiological pathways of application) or which were carried out or generated according to a method that is not acceptable, the documentation of which is not sufficient for an assessment and is not convincing for an expert judgment.' Since monochlorobenzenes and dichlorobenzenes are volatile substances, studies using an open static or renewal system in which actual concentrations are not monitored are awarded Ri 3. An exception is made for the studies on *Vibrio fischeri*, because the duration of these studies (5-15 minutes) is too short for a significant loss of the test substance due to volatilisation.
- Ri 4: Not assignable
 'Studies or data ... which do not give sufficient experimental details and are only listed in short abstracts or secondary literature (books, reviews, etc.).'
- Ri 4*: Data from other sources
 'Studies or data ... which are most likely copied from other sources'

All available studies were summarised in data-tables, which are included as Appendices to this report. These tables contain information on species characteristics, test conditions and endpoints. Explanatory notes are included with respect to the assignment of the reliability indices.

Endpoints with Ri 1 or 2 are accepted as valid, but this does not automatically mean that the endpoint is selected for the derivation of ERLs. The validity scores are assigned on the basis of scientific reliability, but valid endpoints may not be relevant for the purpose of ERL-derivation (e.g., due to inappropriate exposure times or test conditions that are not relevant for the Dutch situation).

After data collection and validation, toxicity data were combined into an aggregated data table with one effect value per species according to section 2.2.6 of the INS-Guidance. Results from studies in which test concentrations are measured continuously or at the beginning and end of the test period are always preferred above studies without analyses of concentrations. When several effect data were available for a species, the geometric mean of multiple values for the same endpoint was calculated where possible. Subsequently, when several endpoints were available for one species, the lowest of these endpoints (per species) is reported in the aggregated data table.

2.3 Derivation of ERLs for drinking water and MAC_{marine}

2.3.1 Drinking water

The INS Guidance includes the MPC for surface waters intended for the abstraction of drinking water ($MPC_{dw, water}$) as one of the MPCs from which the lowest value should be selected as the general MPC_{water} (see INS-Guidance, section 3.1.6 and 3.1.7). According to the daughter directive Priority Substances, however, the derivation of the AA-EQS (= MPC) should be based on direct exposure, secondary poisoning and human exposure, due to the consumption of fishery products. Drinking water was not included in the proposal and is thus not determinative for the general MPC value. The $MPC_{dw, water}$ is therefore presented as a separate value in this report. The MPC_{water} is thus derived considering the individual MPCs based on direct exposure ($MPC_{eco, water}$), secondary poisoning ($MPC_{sp, water}$) or human consumption of fishery products ($MPC_{hh food, water}$); the need for derivation of the latter two depends on the characteristics of the compound.

Related to this is the inclusion of water treatment for the derivation of the $MPC_{dw, water}$. According to the INS-Guidance (section 3.1.7), a substance specific removal efficiency related to simple water treatment should be derived in case the $MPC_{dw, water}$ is lower than the other MPCs. However, since no agreed method is currently available, in this report removal efficiency is set to 0% (= no removal).

2.3.2 MAC_{eco, marine}

The assessment factor for the derivation of the MAC_{eco, marine} is based on:

- the assessment factor for the MAC_{eco, water} when acute toxicity data for at least two specific marine taxa are available, or
- the assessment factor for the MAC_{eco, water} value with an additional assessment factor of 5 when acute toxicity data for only one specific marine taxon are available (analogous to the derivation of the MPC according to Van Vlaardingen and Verbruggen, 2007), or
- the assessment factor for the MAC_{eco, water} value with an additional assessment factor of 10 when no acute toxicity data are available for specific marine taxa.

If freshwater and marine data sets are not combined, the MAC_{eco, marine} is derived using the marine toxicity data and the additional assessment factors mentioned above. It has to be noted that this procedure is currently under discussion. Therefore, the MAC_{eco, marine} value might need to be re-evaluated once an agreed procedure is available.

3 Monochlorobenzene

3.1 Substance identification, physico-chemical properties, fate and human toxicology

3.1.1 Identity

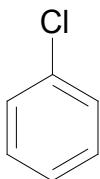


Figure 1. Structural formula of monochlorobenzene.

Table 3. Identification of monochlorobenzene

Parameter	Value	Source
Common/trivial/other name	Chlorobenzene	Mackay et al., 2006
Chemical name	Chlorobenzene	Mackay et al., 2006
CAS number	108-90-7	Mackay et al., 2006
EC number	203-628-5	ECB, ESIS
SMILES code	Clc1ccccc1	Chemsketch
Mode of action	Narcosis	ECB, ESIS

3.1.2 Physico-chemical properties

Table 4. Physico-chemical properties of monochlorobenzene

Parameter	Unit	Value	Remark	Reference ^a
Molecular weight	[g/mol]	112.6		Mackay et al., 2006
Water solubility	[mg/L]	495	10–70 °C	Mackay et al., 2006 (Hovarth and Getzen, 1985)
$\log K_{OW}$	[$-$]	2.89		Mackay et al., 2006 (Hansch et al., 1995)
$\log K_{OC}$	[$-$]	2.34		Otte et al., 2001
Vapour pressure	[Pa]	1333	22.2 °C	Mackay et al., 2006 (Stull, 1947)
Melting point	[°C]	-45.31		Mackay et al., 2006 (Lide, 2003)
Boiling point	[°C]	131.72		Mackay et al., 2006 (Lide, 2003)
Henry's law constant	[Pa.m ³ /mol]	297	20 °C	Mackay et al., 2006 (Staudinger and Roberts, 1996)

^a: reference between brackets refers to citation by Mackay et al., 2006

3.1.3 Behaviour in the environment

Table 5. Selected environmental properties of monochlorobenzene

Parameter	Unit	Value	Remark	Reference ^a
Hydrolysis half-life	DT50 [d]		No hydrolysis at environmentally relevant conditions	Mackay et al., 2006; (Mabey et al., 1982)
Photolysis half-life	DT50 [d]		No photolysis at environmentally relevant conditions	Mackay et al., 2006; (Mabey et al., 1982)
Readily biodegradable	No	OECD 301C		IUCLID, 2000a

^a: reference between brackets refers to citation by Mackay et al., 2006

In water, volatilisation is the main removal process.

3.1.4 Bioconcentration and biomagnification

An overview of the bioaccumulation data for monochlorobenzene is given in Table 6. Detailed bioaccumulation data for monochlorobenzene are tabulated in Appendix 1.

Table 6. Overview of bioaccumulation data for monochlorobenzene

Parameter	Unit	Value	Remark	Reference
BCF (fish)	[L/kg]	11	<i>Cyprinus carpio</i>	METI (http://www.safe.nite.go.jp)
BCF (fish)	[L/kg]	25	<i>Cyprinus carpio</i>	METI (http://www.safe.nite.go.jp)
BCF (fish)	[L/kg]	84	<i>Pimephales promelas</i>	De Wolf and Lieder, 1998
BMF	[kg/kg]	1	default value for BCF < 2000 L/kg	Van Vlaardingen and Verbruggen, 2007

3.1.5 Human toxicological threshold limits and carcinogenicity

The following R-phrases are assigned to monochlorobenzene: R10, R20 and R51/53 (ECB,ESIS). Based on a NOAEL of 60 mg/kg body weight per day in a 2-year rat study (dosing by gavage), and applying an assessment factor of 500 (100 for inter- and intraspecies variation and 5 for limited evidence of carcinogenicity) a TDI of 85.7 µg/kg body weight for 5 days/week exposure was calculated (WHO, 2004). When corrected for daily intake (7 instead of 5 days/week), the TDI is 61.2 µg/kg bw/day.

3.2 Trigger values

This section reports on the trigger values for ERL water derivation (as required in the WFD framework).

Table 7. Monochlorobenzene: collected properties for comparison to MPC triggers

Parameter	Value	Unit	Method/Source	Derived at section
Log $K_{p,\text{susp-water}}$	1.34	[-]	$K_{OC} \times f_{OC,\text{susp}}^{\text{a}}$	K_{OC} : 3.1.2
BCF	84	[L/kg]		3.1.4
BMF	1	[kg/kg]		3.1.4
Log K_{ow}	2.89	[-]		3.1.2
R-phrases	R10, R20, R51/53	[-]		3.1.5

^a: $f_{OC,\text{susp}} = 0.1 \text{ kg}_{OC}/\text{kg}_{\text{solid}}$ (EC, 2003).

- monochlorobenzene has a $K_{p,\text{susp-water}} < 3$; derivation of MPC_{sediment} is not triggered
- monochlorobenzene has a $K_{p,\text{susp-water}} < 3$; expression of the MPC_{water} as MPC_{susp, water} is not required
- monochlorobenzene has a BCF < 100 L/kg; assessment of secondary poisoning is not triggered
- monochlorobenzene has no classification (R-phrase) upon which an MPC_{water} for human health via food (fish) consumption (MPC_{hh food, water}) should be derived. However, since limited evidence for carcinogenicity of monochlorobenzene is available, the MPC_{hh food, water} is derived
- for monochlorobenzene, no specific A1 value or Drinking Water value is available from Council Directives 75/440, EEC and 98/83/EC, respectively.

3.3 Derivation of ERLs for water

3.3.1 Aquatic toxicity data

An overview of the selected freshwater toxicity data for monochlorobenzene is given in Table 8 and selected marine toxicity data are given in Table 9. Detailed toxicity data for monochlorobenzene are tabulated in Appendix 2.

Table 8. Monochlorobenzene: selected freshwater toxicity data for ERL derivation

Chronic ^a Taxonomic group	NOEC/EC10 (mg/L)	Acute ^a Taxonomic group	L(E)C50 (mg/L)
<u>Algae</u> <i>Pseudokirchneriella subcapitata</i>	6.8 ^b	<u>Bacteria</u> <i>Pseudomonas fluorescens</i>	118.5
<u>Crustacea</u> <i>Daphnia magna</i>	0.32^c	<u>Algae</u> <i>Pseudokirchneriella subcapitata</i>	12.5
<u>Amphibia</u> <i>Ambystoma gracile</i>	0.872	<u>Crustacea</u> <i>Ceriodaphnia dubia</i>	5.29 ^d
<u>Pisces</u> <i>Danio rerio</i>	4.8^c	<i>Daphnia carinata</i>	3.99
<i>Oncorhynchus mykiss</i>	2.9	<i>Daphnia magna</i>	10.6 ^e
		<u>Pisces</u> <i>Danio rerio</i>	10.5
		<i>Lepomis macrochirus</i>	5.77 ^f
		<i>Oncorhynchus mykiss</i>	4.7 ^g
		<i>Pimephales promelas</i>	16.9 ^g

^a For detailed information see Appendix 2. Bold values are used for MPC derivation.

- ^b Test duration 96 hours.
^c Most sensitive endpoint; growth.
^d Most sensitive endpoint; immobilisation.
^e Geometric mean of 26 and 4.3 mg/L, measured concentrations.
^f Geometric mean of 4.5 and 7.4 mg/L, test duration 96 hours.
^g Based on measured concentration, test duration 96 hours.

Table 9. Monochlorobenzene: selected marine toxicity data for ERL derivation

Chronic ^a Taxonomic group	NOEC/EC10 (mg/L)	Acute ^a Taxonomic group	L(E)C50 (mg/L)
		<u>Bacteria</u>	
		<i>Vibrio fischeri</i>	15 ^b
		<u>Pisces</u>	
		<i>Platichthys flesus</i>	6.6
		<i>Solea solea</i>	5.8

^a For detailed information see Appendix 2.

^b Microtox test, test duration 15 minutes. Based on measured concentration.

3.3.2 Treatment of fresh- and saltwater toxicity data

In line with the Fraunhofer Guidance (Lepper, 2005), the datasets for freshwater and marine water are combined.

3.3.3 Mesocosm studies

No mesocosm studies are available for monochlorobenzene.

3.3.4 Derivation of MPC_{water} and MPC_{marine}

3.3.4.1 MPC_{eco, water} and MPC_{eco, marine}

Freshwater

The base set is complete. Additionally, chronic data are available for four taxonomic groups (algae, crustaceans, amphibians and fish). Based on this dataset, the MPC_{eco, water} is derived using an assessment factor of 10 on the lowest NOEC value (*Daphnia magna*, 0.32 mg/L), resulting in an MPC_{eco, water} of $0.32 \text{ mg/L} / 10 = 32 \mu\text{g/L}$

Marine water

Based on the data in Tables 8 and 9 the MPC_{eco, marine} is derived using an assessment factor of 100 on the lowest NOEC value. Thus, the MPC_{eco, marine} is $0.32 \text{ mg/L} / 100 = 3.2 \mu\text{g/L}$.

3.3.4.2 MPC_{sp, water} and MPC_{sp, marine}

Since the BCF of monochlorobenzene is < 100 L/kg, ERLs for secondary poisoning are not derived.

3.3.4.3 MPC_{hh food, water}

Based on the limited evidence for carcinogenicity, an MPC_{hh food, water} should be derived for monochlorobenzene. Using the TDI of 61.2 µg/kg bw day and the formulas in section 3.1.5 of the INS Guidance, the MPC_{hh food, water} is 44 µg/L. This value is valid for freshwater as well as for the marine compartment.

3.3.5 Selection of MPC_{water} and MPC_{marine}

Freshwater

The lowest value of the routes included is the MPC_{eco, water} (32 µg/L). Therefore, the MPC_{water} for monochlorobenzene is 32 µg/L.

Marine water

The lowest value of the routes included is the MPC_{eco, marine} (3.2 µg/L). Therefore, the MPC_{marine} for monochlorobenzene is 3.2 µg/L.

3.3.6 MPC_{dw, water}

Because no A1 value or DWS is available for monochlorobenzene, the MPC_{dw, water, provisional} is derived using the formula in section 3.1.6 of the INS guidance. Using the TDI of 61.2 µg/kg bw day, an average bodyweight of 70 kg and an average uptake of drinking water of 2 L, the MPC_{dw, water, provisional} is 2.1×10^2 µg/L. The MPC_{dw, water, provisional} is not taken into account for the derivation of the final MPC_{water}.

3.3.7 Derivation of MAC_{eco, water} and MAC_{eco, marine}

Freshwater

The base set is complete and acute data are available for two additional taxonomic groups (bacteria and protozoa). Monochlorobenzene has a BCF < 100 L/kg. Therefore, an assessment factor of 100 is used on the lowest L(E)C₅₀ value (3.99 mg/L for *Daphnia carinata*), resulting in a MAC_{eco, water} of $3.99 \text{ mg/L} / 100 = 40 \mu\text{g/L}$.

Marine water

Based on the data in Tables 8 and 9 and the non-bioaccumulative properties of monochlorobenzene, an assessment factor of 1000 is used on the lowest L(E)C₅₀ value (3.99 mg/L for *Daphnia carinata*), resulting in a MAC_{eco, marine} of $3.99 \text{ mg/L} / 1000 = 4.0 \mu\text{g/L}$.

3.3.8 Derivation of NC_{water}

Freshwater

The NC is set a factor of 100 below the MPC. The NC_{water} is $32 \mu\text{g/L} / 100 = 0.32 \mu\text{g/L}$.

Marine water

The NC is set a factor of 100 below the MPC. The NC_{marine} is $3.2 \mu\text{g/L} / 100 = 3.2 \times 10^{-2} \mu\text{g/L}$.

3.3.9 Derivation of SRC_{eco, water}

The SRC_{eco, water} for monochlorobenzene is based on the geometric mean of the NOEC values, since data are available for algae, *Daphnia* and fish. The SRC_{eco, water} is $1925 = 1.9 \times 10^3 \mu\text{g/L}$. This value is valid for freshwater as well as for the marine compartment.

3.4 Derivation of ERLs for sediment

The log K_{p, susp-water} of monochlorobenzene is below the trigger value of 3, therefore, ERLs are not derived for sediment.

3.5 Derivation of ERLs for soil

3.5.1 Toxicity data for soil

Table 10. Monochlorobenzene: selected soil toxicity data for ERL derivation

Chronic ^a Taxonomic group	NOEC/EC10 (mg/kg)	Acute ^a Taxonomic group	L(E)C50 (mg/kg)
<u>Annelida</u>			
		<i>Eisenia andrei</i>	551
		<i>Eisenia andrei</i>	649
		<i>Lumbricus rubellus</i>	1367
		<i>Lumbricus rubellus</i>	1478

^a For detailed information see Appendix 3.

^b Results normalised for Dutch standard soil.

3.5.2 Derivation of MPC_{soil}

3.5.2.1 MPC_{eco, soil}

The dataset for monochlorobenzene contains two L(E)C₅₀ values for two species: 551 and 649 mg/kg_{dwt} standard soil for *Eisenia andrei* and 1367 and 1478 mg/kg_{dwt} for *Lumbricus rubellus*. Since these values are comparable, the geometric means are used: 580 mg/kg_{dwt} for *Eisenia andrei* and 1421 mg/kg_{dwt} for *Lumbricus rubellus*. According to the INS guidance, an assessment factor of 1000 should be applied to the lowest L(E)C₅₀ for the derivation of the MPC_{eco, soil}. Therefore, the MPC_{eco, soil} is 580 mg/kg_{dwt} / 1000 = 5.8 x 10² µg/kg_{dwt} standard soil. Since data for only one taxonomic group are available, this value should be compared with the MPC_{eco, soil} derived using the equilibrium partitioning method. The MPC_{eco, soil, equilibrium partitioning} is 4.3 x 10² µg/kg_{dwt}, which is lower than the value based on experimental soil data. The MPC_{eco, soil} for monochlorobenzene is set to 4.3 x 10² µg/kg_{dwt}.

3.5.2.2 MPC_{sp, soil}

The log K_{ow} of monochlorobenzene is below the trigger value of 3, therefore, no MPC_{sp, soil} is derived.

3.5.2.3 MPC_{human, soil}

The MPC_{human, soil} is derived using the formulas in section 3.3.6 of the INS Guidance (Van Vlaardingen and Verbruggen, 2007). These calculations result in an MPC_{human, soil} of 1236 = 1.2 x 10² µg/kg_{dwt} for consumption of root crops.

3.5.3 Selection of MPC_{soil}

The lowest value of the routes included is the MPC_{eco, soil}. The MPC_{soil} for monochlorobenzene is set to 4.3 x 10² µg/kg_{dwt}.

3.5.4 Derivation of NC_{soil}

The NC is set a factor 100 of below the MPC. Therefore, the NC_{soil} is 426 µg/kg_{dwt} / 100 = 4.3 µg/kg_{dwt}.

3.5.5 Derivation of SRC_{eco, soil}

Since only acute values for the toxicity of monochlorobenzene in soil are available, the SRC_{eco, soil} is derived by comparing the geometric mean of the L(E)C₅₀ values with an assessment factor of 10 with the result of the equilibrium partitioning method using the SRC_{eco, water} (Van Vlaardingen and Verbruggen, 2007). For monochlorobenzene, the geometric mean of the four L(E)C₅₀ values is 922 mg/kg_{dwt}. Using an assessment factor of 10, the SRC_{eco, soil} is 92.2 mg/kg_{dwt} = 92200 µg/kg_{dwt}. The SRC_{eco, soil} derived using the equilibrium partitioning method is 25623 µg/kg_{dwt}. Therefore, the SRC_{eco, soil} is 2.6×10^4 µg/kg_{dwt} standard soil.

3.6 Derivation of ERLs for groundwater

3.6.1 Derivation of MPC_{gw}

3.6.1.1 MPC_{eco, gw}

Since no ecotoxicity data are available for the groundwater compartment, the MPC_{eco, water} for surface water is taken as substitute. The MPC_{eco, gw} for monochlorobenzene is 32 µg/L.

3.6.1.2 MPC_{human, gw}

The MPC_{dw, water, provisional} is used as substitute for the MPC_{human, gw}. Therefore, the MPC_{human, gw} is 2.1×10^2 µg/L.

3.6.2 Selection of MPC_{gw}

The lowest value of the routes included is the MPC_{eco, gw}. Therefore, the MPC_{gw} is 32 µg/L.

3.6.3 Derivation of NC_{gw}

The NC is set a factor of 100 below the MPC. For groundwater, the NC_{gw} is $32 \mu\text{g}/\text{L} / 100 = 0.32 \mu\text{g}/\text{L}$.

3.6.4 Derivation of SRC_{eco, gw}

Since no toxicity data are available for the groundwater compartment, the SRC_{eco, water} for surface water is taken as a substitute. Therefore, the SRC_{eco, gw} is 1.9×10^3 µg/L.

3.7 Derivation of ERLs for air

3.7.1.1 MPC_{eco, air}

Since no data on the ecotoxicity of monochlorobenzene are available, no MPC_{eco, air} can be derived.

3.7.1.2 MPC_{human, air}

The MPC_{human, air} is set at the same value as the Tolerable Concentration in Air (TCA). The TCA and therefore, the MPC_{human, air} is $5.0 \times 10^2 \mu\text{g}/\text{m}^3$ (Lijzen et al., 2002).

3.7.2 Selection of MPC_{air}

The only value of the routes included is the MPC_{human, air}. Therefore, the MPC_{air} is $5.0 \times 10^2 \mu\text{g}/\text{m}^3$.

3.8 Overview of ERLs for monochlorobenzene

In Table 11, an overview of the ERLs derived for monochlorobenzene is given.

Table 11. Monochlorobenzene: derived NC, MPC, MAC_{eco} and SRC_{eco}

ERL	Unit	NC	MPC	MAC _{eco}	SRC _{eco}
Water	µg/L	0.32	32	40	1.9 x 10 ³
Marine	µg/L	3.2 x 10 ⁻²	3.2	4.0	1.9 x 10 ³
Soil	µg/kg	4.3	4.3 x 10 ²	n.a.	2.6 x 10 ⁴
Groundwater	µg/L	0.32	32	n.a.	1.9 x 10 ³
Air	µg/m ³	n.a.	5.0 x 10 ²	n.a.	n.a.
Drinking water	µg/L	n.a.	2.1 x 10 ²	n.a.	n.a.

n.a. = not applicable

3.9 Comparison of derived ERLs with monitoring data

Monitoring data for the Rhine from the years 2001–2006, obtained from RIWA (Association of River Waterworks), show that at all sampling occasions and locations, the concentration of monochlorobenzene in water was usually below detection limits (0.01 µg/L). On two occasions in 2005, a concentration of 0.15 µg/L was measured, a value well below the new MPC_{water}. Therefore, no exceedance is expected.

4 Dichlorobzenes

Since the differences in toxicity for the individual dichlorobzenes are small, ERLs are derived for the dichlorobzenes as a group, based on the combined datasets of 1,2-dichlorobenzene, 1,3-dichlorobenzene and 1,4-dichlorobenzene. ERLs for the group are only derived when relevant for the individual substances, i.e., when an individual substance triggers derivation.

4.1 Substance identification, physico-chemical properties, fate and human toxicology

4.1.1 Identity

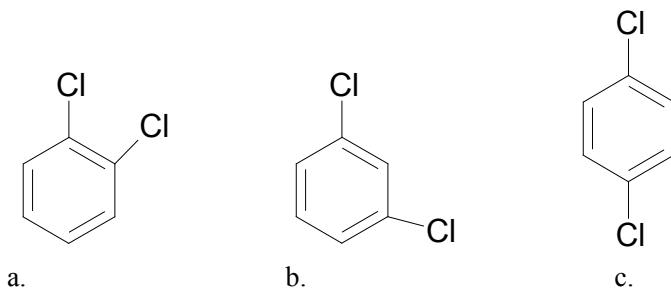


Figure 2. Structural formulas of dichlorobzenes.
a = 1,2-dichlorobenzene, b = 1,3-dichlorobenzene, c = 1,4-dichlorobenzene.

Table 12. Identification of dichlorobzenes.

Parameter	Value	Substance	Source
Common/trivial/other name	dichlorobzenes		Mackay et al., 2006
Chemical name	1,2-dichlorobenzene 1,3-dichlorobenzene 1,4-dichlorobenzene		Mackay et al., 2006
CAS number	95-50-1 541-73-1 106-46-7	1,2-dichlorobenzene 1,3-dichlorobenzene 1,4-dichlorobenzene	Mackay et al., 2006
EC number	202-425-9 208-792-1 203-400-5	1,2-dichlorobenzene 1,3-dichlorobenzene 1,4-dichlorobenzene	ECB, ESIS
SMILES code	Clc1ccccc1Cl Clc1cc(Cl)ccc1 Clc1ccc(Cl)cc1	1,2-dichlorobenzene 1,3-dichlorobenzene 1,4-dichlorobenzene	Chemsketch
Mode of action	Narcosis		ECB, ESIS

4.1.2 Physico-chemical properties

Table 13. Physico-chemical properties of dichlorobenzenes

Parameter	Unit	Value	Remark	Reference ^a
Molecular weight	[g/mol]	147		Mackay et al., 2006
Water solubility	[mg/L]	92.3 124 90.0	25 °C, 1,2-dichlorobenzene 25 °C, 1,3-dichlorobenzene 25 °C, 1,4-dichlorobenzene	Mackay et al., 2006; (Miller, 1984; Hovarth, 1982)
$\log K_{\text{ow}}$	[-]	3.45	worst-case, $\log K_{\text{ow}}$ of 1,4-dichlorobenzene	Mackay et al., 2006; (Sangster, 1993)
$\log K_{\text{oc}}$	[-]	2.69	worst-case, $\log K_{\text{oc}}$ of 1,3-dichlorobenzene	Otte et al., 1991
Vapour pressure	[Pa]	133.8 185.0 170	20 °C, 1,2-dichlorobenzene 20 °C, 1,3-dichlorobenzene 20 °C, 1,4-dichlorobenzene	Mackay et al., 2006; (Roháč et al., 1999)
Melting point	[°C]	-17.0 -24.8 53.09	1,2-dichlorobenzene 1,3-dichlorobenzene 1,4-dichlorobenzene	Mackay et al., 2006; (Lide, 2003)
Boiling point	[°C]	180 183 174	1,2-dichlorobenzene 1,3-dichlorobenzene 1,4-dichlorobenzene	Mackay et al., 2006; (Lide, 2003)
Henry's law constant	[Pa.m ³ /mol]	133 288 275	20 °C, 1,2-dichlorobenzene 20 °C, 1,3-dichlorobenzene 20 °C, 1,4-dichlorobenzene	Mackay et al., 2006; (Staudinger and Roberts, 2001)

^a: references between brackets refers to citation by Mackay et al., 2006

4.1.3 Behaviour in the environment

Table 14. Selected environmental properties of dichlorobenzenes

Parameter	Unit	Value	Remark	Reference
Hydrolysis half-life	DT50 [d]		No hydrolysis at environmentally relevant concentrations	Mackay et al., 2006
Photolysis half-life	DT50 [d]		No photolysis at environmentally relevant concentrations	Mackay et al., 2006
Readily biodegradable		No Yes	1,2-dichlorobenzene 1,4-dichlorobenzene	IUCLID, 2000b EC, 2004

4.1.4 Bioconcentration and biomagnification

An overview of the bioaccumulation data for dichlorobenzenes is given in Table 15. Detailed bioaccumulation data are tabulated in Appendix 1.

Table 15. Overview of bioaccumulation data for dichlorobenzenes

Parameter	Unit	Value	Remark	Reference
BCF (fish)	[L/kg]	178	1,2-dichlorobenzene, <i>Cyprinus carpio</i>	METI (http://www.safe.nite.go.jp)
		174	1,2-dichlorobenzene, <i>Cyprinus carpio</i>	METI (http://www.safe.nite.go.jp)
		454	1,2-dichlorobenzene, <i>Pimephales promelas</i>	Sijm et al., 1993
		138	1,3-dichlorobenzene, <i>Cyprinus carpio</i>	METI (http://www.safe.nite.go.jp)
		195	1,3-dichlorobenzene, <i>Cyprinus carpio</i>	METI (http://www.safe.nite.go.jp)
		728	1,3-dichlorobenzene, <i>Pimephales promelas</i>	De Wolf and Lieder, 1998
		69	1,4-dichlorobenzene, <i>Cyprinus carpio</i>	METI (http://www.safe.nite.go.jp)
		80	1,4-dichlorobenzene, <i>Cyprinus carpio</i>	METI (http://www.safe.nite.go.jp)
		296	1,4-dichlorobenzene, <i>Jordanella floridae</i>	EC, 2004; Smith et al., 1990
		1	default value for BCF < 2000 L/kg	Van Vlaardingen and Verbruggen, 2007
BMF	[kg/kg]			

4.1.5 Human toxicological threshold limits and carcinogenicity

1,2-dichlorobenzene

The following R-phrases are assigned to 1,2-dichlorobenzene: R22, R36/37/38 and R50/53 (ECB, ESIS). Based on a NOAEL of 60 mg/kg bw per day in a 2-year mouse study (dosing by gavage), and applying an uncertainty factor of 100, a TDI of 429 µg/kg bw/day was derived (WHO, 2004).

1,3-dichlorobenzene

The following R-phrases are assigned to 1,3-dichlorobenzene: R22 and R51/53 (ECB, ESIS). Insufficient reliable toxicological data on 1,3-dichlorobenzene are available to derive an ADI or TDI value.

1,4-dichlorobenzene

The following R-phrases are assigned to 1,4-dichlorobenzene: R36, R40 and R50/53 (ECB, ESIS). 1,4-dichlorobenzene is not considered to be genotoxic and the relevance for humans of the tumours observed in animals is doubtful. It is therefore valid to calculate a guideline value using the TDI approach. A TDI of 107 µg/kg of body weight has been calculated by applying an uncertainty factor of 1000 (100 for inter- and intraspecies variation and 10 for the use of a LOAEL instead of a NOAEL and because the toxic end-point is carcinogenicity) to a LOAEL of 150 mg/kg of body weight per day for kidney effects observed in a 2-year rat gavage study (administration 5 days per week) (WHO, 2004).

The derivation of the $MPC_{sp, water}$ and the $MPC_{hh food, water}$ is based on the worst-case scenario that all dichlorobenzenes are as toxic as 1,4-dichlorobenzene, e.g., on a TDI of 107 µg/kg bw and a NOAEL of 15 mg/kg bw.

4.2 Trigger values

This section reports on the trigger values for ERL water derivation (as demanded in the WFD framework).

Table 16. Dichlorobenzenes: collected properties for comparison to MPC triggers

Parameter	Value	Unit	Method/Source	Derived at section
Log $K_{p, \text{susp-water}}$	1.69	[-]	$K_{OC} \times f_{OC, \text{susp}}^a$	K_{OC} : 4.1.2
BCF	728	[L/kg]	worst-case, BCF of 1,3-dichlorobenzene	4.1.4
BMF	1	[kg/kg]		4.1.4
Log K_{ow}	3.45	[-]	worst-case, log K_{ow} of 1,4-dichlorobenzene	4.1.2
R-phrases	R22, R36/37/38, R50/53 R22, R51/53 R36, R40, R50/53	[-]	1,2-dichlorobenzene 1,3-dichlorobenzene 1,4-dichloroebenzene	4.1.5

^a: $f_{OC, \text{susp}} = 0.1 \text{ kg}_{OC}/\text{kg}_{solid}$ (EC, 2003).

- dichlorobenzenes have a worst-case of $\log K_{p, \text{susp-water}} < 3$; derivation of $MPC_{sediment}$ is not triggered
- dichlorobenzenes have a worst-case of $\log K_{p, \text{susp-water}} < 3$; expression of the MPC_{water} as $MPC_{susp, water}$ is not required
- dichlorobenzenes have a BCF > 100 L/kg; assessment of secondary poisoning is triggered
- dichlorobenzenes have the potential to bioaccumulate and an R22 classification (1,2- and 1,3-dichlorobenzene). 1,4-dichlorobenze has an R40 classification. Therefore, an MPC_{water} for human health via food (fish) consumption ($MPC_{hh food, water}$) should be derived
- for dichlorobenzenes, no specific A1 values or Drinking Water values are available from Council Directives 75/440, EEC and 98/83/EC, respectively.

4.3 Derivation of ERLs for water

4.3.1 Aquatic toxicity data

An overview of the selected freshwater toxicity data for dichlorobenzenes is given in Table 17 and marine toxicity data are given in Table 18. When selected data were available for multiple dichlorobenzenes, the lowest (worst-case) value was chosen. Detailed toxicity data for dichlorobenzene are tabulated in Appendix 2.

Table 17. Dichlorobenzenes: selected freshwater toxicity data for ERL derivation

Chronic ^a Taxonomic group	NOEC/EC10 (mg/L)	Acute ^a Taxonomic group	L(E)C50 (mg/L)
<u>Algae</u>		<u>Bacteria</u>	
<i>Scenedesmus subspicatus</i>	16 ^b	<i>Pseudomonas fluorescens</i>	21.6 ^e
<u>Crustacea</u>		<i>Nitrosomonas spec.</i>	86 ^b
<i>Daphnia magna</i>	0.22 ^{b,c}	<u>Algae</u>	
<u>Pisces</u>		<i>Pseudokirchneriella subcapitata</i>	1.6 ^b
<i>Danio rerio</i>	0.44 ^b	<i>Scenedesmus pannonicus</i>	17 ^e
<i>Jordanella floridae</i>	0.2^b	<u>Crustacea</u>	
<i>Oncorhynchus mykiss</i>	0.56 ^d	<i>Daphnia magna</i>	0.74^{e,f}
<i>Pimephales promelas</i>	0.57 ^b	<u>Insecta</u>	
		<i>Tanytarsus dissimilis</i>	12 ^e
		<u>Pisces</u>	
		<i>Danio rerio</i>	2.1 ^b
		<i>Jordanella floridae</i>	2.1 ^b
		<i>Oncorhynchus mykiss</i>	1.59 ^{e,h,i}
		<i>Oryzias latipes</i>	9.9 ^e
		<i>Pimephales promelas</i>	3.6 ^{b,j}
		<i>Poecilia reticulata</i>	4.8 ^e

^a For detailed information see Appendix 2. Bold values are used for ERL derivation.

^b Data for 1,4-dichlorobenzene

^c Most sensitive endpoint; fertility

^d Data for 1,3-dichlorobenzene

^e Data for 1,2-dichlorobenzene

^h Most relevant test duration 96 hours.

ⁱ Geometric mean of 1.61 and 1.58 mg/L; based on measured concentration.

^j ELS test

Table 18. Dichlorobenzenes: selected marine toxicity data for ERL derivation

Chronic ^a Taxonomic group	NOEC/EC10 (mg/L)	Acute ^a Taxonomic group	L(E)C50 (mg/L)
		<u>Bacteria</u>	
		<i>Vibrio fischeri</i>	4.4 ^{b,c}
		<u>Pisces</u>	
		<i>Cyprinodon variegatus</i>	7.4 ^d
		<i>Platichthys flesus</i>	4.6 ^b
		<i>Solea solea</i>	4.2 ^b

^a For detailed information see Appendix 2.

^b Data for 1,2-dichlorobenzene.

^c Geometric mean of 6.1 and 3.14 mg/L; most relevant test duration 15 minutes.

^d Data for 1,4-dichlorobenzene.

4.3.2 Treatment of fresh- and saltwater toxicity data

In line with the Fraunhofer guidance (Lepper, 2005), the datasets for freshwater and marine water are combined.

4.3.3 Mesocosm studies

No mesocosm studies are available for the dichlorobenzenes.

4.3.4 Derivation of MPC_{water} and MPC_{marine}

4.3.4.1 MPC_{eco, water} and MPC_{eco, marine}

Freshwater

Chronic toxicity data are available for three taxonomic groups (algae, crustaceans and fish). Therefore, the MPC_{eco, water} is derived using an assessment factor of 10 on the lowest NOEC value (0.2 mg/L for *Jordanella floridae*). Thus, the MPC_{eco, water} is $0.2 \text{ mg/L} / 10 = 0.02 \text{ mg/L} = 20 \mu\text{g/L}$

Marine water

Based on the data in Tables 17 and 18, an additional assessment factor of 10 is used on the lowest NOEC for the derivation of the MPC_{eco, marine}. Thus, the MPC_{eco, marine} is $20 \mu\text{g/L} / 10 = 2.0 \mu\text{g/L}$.

4.3.4.2 MPC_{sp, water} and MPC_{sp, marine}

Freshwater

All dichlorobenzenes have a BCF > 100 L/kg, thus assessment of secondary poisoning is triggered. The MPC_{sp, water} is based on the worst-case scenario that all dichlorobenzenes are as toxic as 1,4-dichlorobenzene. The LOAEL value of 150 mg/kg bw (WHO, 2003) with an additional assessment factor of 10 is used as a NOAEL value for the calculation of the MPC_{oral}. Using an assessment factor of 30, the MPC_{oral} is 5 mg/kg food. Applying the formulas in the INS guidance using the BCF of 728 L/kg, the MPC_{sp, water} is 6.9 µg/L.

Marine water

The MPC_{sp, marine} is based on the same data as the MPC_{sp, water}. Therefore, the MPC_{sp, marine} is 6.9 µg/L.

4.3.4.3 MPC_{hh food,water}

Derivation of MPC_{hh food, water} for dichlorobenzenes is required (Table 16). Based on a TDI of 107 µg/kg bw per day and using the formulas in the INS guidance, the MPC_{hh food, water} is 8.9 µg/L. This value is valid for freshwater as well as for the marine compartment.

4.3.5 Selection of MPC_{water} and MPC_{marine}

Freshwater

The lowest value of the routes included is the MPC_{sp, water}. Therefore, the MPC_{water} is 6.9 µg/L.

Marine water

The lowest value of the routes included is the MPC_{eco, water}. Therefore, the MPC_{marine} is 2.0 µg/L.

4.3.6 MPC_{dw, water}

Because no A1 value or DWS is available for dichlorobenzenes, the MPC_{dw, water, provisional} is derived using the formula in section 3.1.6 of the INS guidance. Using the TDI of 107 µg/kg bw per day, an average bodyweight of 70 kg and an average uptake of drinking water of 2 L, the MPC_{dw, water, provisional} is 375 µg/L = 3.8×10^2 µg/L. The MPC_{dw, water, provisional} is not taken into account for the derivation of the final MPC_{water}.

4.3.7 Derivation of MAC_{eco, water} and MAC_{eco, marine}

Freshwater

The acute dataset contains data for bacteria, algae, crustaceans, insects and fish. All dichlorobenzenes have a BCF > 100 L/kg. Therefore, an assessment factor of 1000 is used on the lowest L(E)C₅₀ value (0.74 mg/L, *Daphnia magna*) for the derivation of the MAC_{eco, water}, resulting in a MAC_{eco, water} of 0.74 mg/L / 1000 = 0.74 µg/L. This MAC_{eco, water} value is lower than the MPC_{eco, water}. Since the MPC_{eco, water} is an ERL which should protect the environment from the adverse effects of long-term exposure to dichlorobenzenes, it is also considered protective for short-term exposure. Therefore, the MAC_{eco, water} is set equal to the MPC_{eco, water} with a value of 20 µg/L.

Marine water

Based on the data in Tables 17 and 18 and the bioaccumulative properties of dichlorobenzenes, the MAC_{eco, marine} is derived by applying an assessment factor of 10000 on the lowest L(E)C₅₀ value. Thus, the MAC_{eco, marine} is 0.74 mg/L / 10000 = 0.074 µg/L. This MAC_{eco, marine} value is lower than the MPC_{eco, marine}. Since the MPC_{eco, marine} is an ERL which should protect the environment from the adverse effects of long-term exposure to dichlorobenzenes, it is also considered protective for short-term exposure. Therefore, the MAC_{eco, marine} is set equal to the MPC_{eco, marine} with a value of 2.0 µg/L.

4.3.8 Derivation of NC_{water}

Freshwater

The NC is set a factor of 100 below the MPC. Therefore, the NC_{water} is 6.9 µg/L / 100 = 6.9×10^{-2} µg/L.

Marine water

The NC is set a factor of 100 below the MPC. Therefore, the NC_{marine} is 2.0 µg/L / 100 = 2.0×10^{-2} µg/L.

4.3.9 Derivation of SRC_{eco, water}

The SRC_{eco, water} for dichlorobenzenes is based on the geometric mean of the NOEC values using an assessment factor of 1, because NOEC values are available for three taxonomic groups. Thus, the SRC_{eco, water} is 0.68 mg/L = 6.8×10^{-2} µg/L. This value is valid for freshwater as well as for the marine compartment.

4.4 Derivation of ERLs for sediment

The log K_{p, susp-water} of all dichlorobenzenes is below the trigger value of 3, therefore, ERLs are not derived for sediment.

4.5 Derivation of ERLs for soil

4.5.1 Toxicity data for soil

Table 19. Dichlorobenzenes: selected soil toxicity data for ERL derivation

Chronic Taxonomic group	NOEC/EC10 (mg/kg)	Acute Taxonomic group	L(E)C50 (mg/kg)
<u>Annelida</u>			
		<i>Eisenia andrei</i>	347 ^a
		<i>Eisenia andrei</i>	274 ^a
		<i>Lumbricus rubellus</i>	497 ^a
		<i>Lumbricus rubellus</i>	759 ^a

^a Results are normalised to an organic matter content of 10%.

4.5.2 Derivation of MPC_{soil}

4.5.2.1 MPC_{eco, soil}

The dataset of dichlorobenzenes contains four L(E)C₅₀ values for two species (all data are for 1,4-dichlorobenzene): 347 and 274 mg/kg_{dwt} for *Eisenia andrei* (geometric mean 308 mg/kg_{dwt}) and 497 and 759 mg/kg_{dwt} for *Lumbricus rubellus* (geometric mean 614 mg/kg_{dwt}). According to the INS guidance, an assessment factor of 1000 should be applied to the lowest L(E)C₅₀ for the derivation of the MPC_{eco, soil}. Therefore, the MPC_{eco, soil} is 3080 mg/kg_{dwt} / 1000 = 3.1 x 10² µg/kg_{dwt}

4.5.2.2 MPC_{sp, soil}

The log K_{ow} of the dichlorobenzenes exceeds the trigger value of 3 (worst-case 3.45) and the MPC_{sp, soil} should be derived. Since no bioconcentration data are available, a QSAR was used to estimate the BCF for earthworms (Van Vlaardingen and Verbruggen, section 3.3.5). Using an MPC_{oral} of 5 mg/kg_{bw}, and a BCF of 34.66 L/kg for earthworms, an MPC_{sp, soil} of 1.6 x 10³ µg/kg_{dwt} standard soil was calculated.

4.5.2.3 MPC_{human, soil}

The MPC_{human, soil} was derived using the formulas in section 3.3.6 of the INS Guidance (Van Vlaardingen and Verbruggen, 2007). These calculations resulted in an MPC_{human, soil} of 1.4 x 10³ µg/kg_{dwt} for consumption of root crops.

4.5.3 Selection of MPC_{soil}

The lowest value of the routes included is the MPC_{eco, soil}. Therefore, the MPC_{soil} for dichlorobenzenes is 3.1 x 10² µg/kg_{dwt}.

4.5.4 Derivation of NC_{soil}

The NC is set a factor of 100 below the MPC. Therefore, the NC_{soil} is
 $308 \text{ } \mu\text{g}/\text{kg}_{\text{dwt}} / 100 = 3.1 \text{ } \mu\text{g}/\text{kg}_{\text{dwt}}$.

4.5.5 Derivation of SRC_{eco, soil}

Since only acute values for the toxicity of 1,4-dichlorobenzene in soil are available, the SRC_{eco, soil} is derived by comparing the geometric mean of the L(E)C₅₀ values with an assessment factor of 10 with the result of the equilibrium partitioning method (Van Vlaardingen and Verbruggen, 2007). For 1,4-dichlorobenzene, the geometric mean of the L(E)C₅₀ values is 435 mg/kg_{dwt}. Using an assessment factor of 10, the SRC_{eco, soil} is $435 \text{ mg}/\text{kg}_{\text{dwt}} / 10 = 43500 \text{ } \mu\text{g}/\text{kg}_{\text{dwt}}$. The SRC_{eco, soil} derived using the equilibrium partitioning method is 19890 µg/kg_{dwt}, which is lower than the value based on experimental data. Therefore, the SRC_{eco, soil} is $2.0 \times 10^4 \text{ } \mu\text{g}/\text{kg}_{\text{dwt}}$.

4.6 Derivation of ERLs for groundwater

4.6.1 Derivation of MPC_{gw}

4.6.1.1 MPC_{eco, gw}

Since no toxicity data are available for the groundwater compartment, the MPC_{eco, water} for surface water is taken as a substitute. Therefore, the MPC_{eco, gw} is 20 µg/L.

4.6.1.2 MPC_{human, gw}

The MPC_{dw, water, provisional} is used as a substitute for the MPC_{human, gw}. Therefore, the MPC_{human, gw} is $375 \text{ } \mu\text{g}/\text{L} = 3.8 \times 10^2 \text{ } \mu\text{g}/\text{L}$.

4.6.2 Selection of MPC_{gw}

The lowest value of the routes included is the MPC_{eco, water}. Therefore, the MPC_{gw} is 20 µg/L.

4.6.3 Derivation of NC_{gw}

The NC is set a factor 100 below the MPC. For groundwater therefore, the NC_{gw} is
 $20 \text{ } \mu\text{g}/\text{L} / 100 = 0.20 \text{ } \mu\text{g}/\text{L}$.

4.6.4 Derivation of SRC_{eco, gw}

Since no toxicity data are available for the groundwater compartment, the SRC_{eco, water} for surface water is taken as a substitute. Therefore, the SRC_{eco, gw} is $6.8 \times 10^2 \text{ } \mu\text{g}/\text{L}$.

4.7 Derivation of ERLs for air

4.7.1.1 MPC_{eco, air}

Since no data on the ecotoxicity of dichlorobenzenes in air are available, no MPC_{eco, air} can be derived.

4.7.1.2 MPC_{human, air}

The MPC_{human, air} is set at the same value as the TCA. Since no TCA for dichlorobenzenes is available, no MPC_{human, air} can be derived.

4.7.2 Selection of MPC_{air}

The MPC_{air} for dichlorobenzenes cannot be derived.

4.8 Overview of MPCs for dichlorobenzenes

In Table 20, an overview of the ERLs derived for dichlorobenzenes is given.

Table 20. Dichlorobenzenes: derived NC, MPC, MAC_{eco} and SRC_{eco}

ERL	Unit	NC	MPC	MAC _{eco}	SRC _{eco}
Water	µg/L	6.9 x 10 ⁻²	6.9	20	6.8 x 10 ²
Marine	µg/L	2.0 x 10 ⁻²	2.0	2.0	6.8 x 10 ²
Soil	µg/kg	3.1	3.1 x 10 ²	n.a.	2.0 x 10 ⁴
Groundwater	µg/L	0.20	20	n.a.	6.8 x 10 ²
Air	µg/m ³	n.a.	n.d.	n.a.	n.a.
Drinking water	µg/L	n.a.	3.8 x 10 ²	n.a.	n.a.

4.9 Comparison of derived ERLs with monitoring data

Monitoring data for the Rhine from the years 2003–2006, obtained from RIWA (Association of River Waterworks), show that at all sampling occasions and locations, the concentration of dichlorobenzenes in water was usually below detection limits (0.01 µg/L).

4.10 Sum limits

The small difference in toxicity of the individual dichlorobenzenes can result in unacceptable risks when a mixture of dichlorobenzenes is present at one location. In order to prevent this accumulation of toxicity, the use of a sum limit for dichlorobenzenes is proposed. In this approach, the sum of the concentrations of the combination of the three individual dichlorobenzenes should not exceed the ERLs for the grouped dichlorobenzenes (Table 20). It should be noted that the sum limit only applies when all three dichlorobenzenes are monitored.

5 Tetrachlorobzenes

Since the differences in toxicity for the individual tetrachlorobzenes are small and data availability is limited, ERLs are derived for the tetrachlorobzenes as a group based on the combined datasets of 1,2,3,4-tetrachlorobenzene, 1,2,3,5-tetrachlorobenzene, 1,2,4,5-tetrachlorobenzene. ERLs for the group are only derived when relevant for the individual substances, i.e., when an individual substance triggers derivation.

5.1 Substance identification, physico-chemical properties, fate and human toxicology

5.1.1 Identity

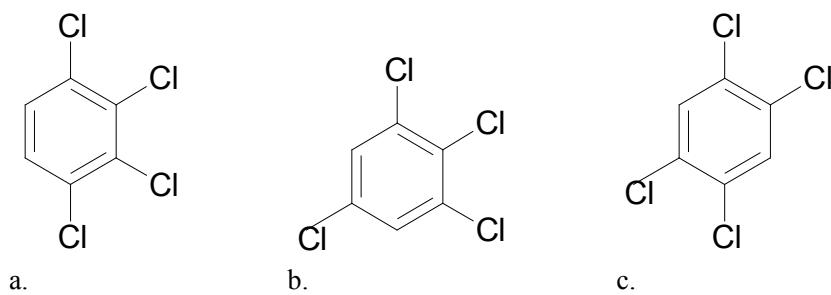


Figure 3. Structural formulas of tetrachlorobzenes.

a = 1,2,3,4-tetrachlorobenzene, b = 1,2,3,5-tetrachlorobenzene, c = 1,2,4,5-tetrachlorobenzene.

Table 21. Identification of tetrachlorobzenes

Parameter	Value	Substance	Source
Common/trivial/other name	tetrachlorobzenes		Mackay et al., 2006
Chemical name	1,2,3,4-tetrachlorobenzene 1,2,3,5-tetrachlorobenzene 1,2,4,5-tetrachlorobenzene		Mackay et al., 2006
CAS number	634-66-2 634-90-2 95-94-3	1,2,3,4-tetrachlorobenzene 1,2,3,5-tetrachlorobenzene 1,2,4,5-tetrachlorobenzene	Mackay et al., 2006
EC number	211-214-0 211-217-7 202-466-2	1,2,3,4-tetrachlorobenzene 1,2,3,5-tetrachlorobenzene 1,2,4,5-tetrachlorobenzene	ECB, ESIS
SMILES code	Clc1ccc(Cl)c(Cl)c1Cl Clc1cc(Cl)c(Cl)c(Cl)c1 ClC1=CC(Cl)C(Cl)C=C1Cl	1,2,3,4-tetrachlorobenzene 1,2,3,5-tetrachlorobenzene 1,2,4,5-tetrachlorobenzene	Chemsketch
Mode of action	Narcosis		ECB, ESIS

5.1.2 Physico-chemical properties

Table 22. Physico-chemical properties of tetrachlorobenzenes

Parameter	Unit	Value	Remark	Reference ^a
Molecular weight	[g/mol]	215.9		Mackay <i>et al.</i> , 2006
Water solubility	[mg/L]	12.2	25 °C, 1,2,3,4-tetrachlorobenzene	Mackay <i>et al.</i> , 2006; (Miller, 1984)
		2.89	25 °C, 1,2,3,5-tetrachlorobenzene	
		2.35	25 °C, 1,2,4,5-tetrachlorobenzene	
log K_{OW}	[-]	4.70	worst-case, log Kow of 1,2,4,5-tetrachlorobenzene	Mackay <i>et al.</i> , 2006; (Hansch <i>et al.</i> , 1995)
log K_{OC}	[-]	3.91	worst-case, log Kow of 1,2,3,4-tetrachlorobenzene	Otte <i>et al.</i> , 1991
Vapour pressure	[Pa]	4.11	25°C, geometric mean of 6.29, 5.085 and 2.163 Pa	Mackay <i>et al.</i> , 2006; (Rordorf, 1985)
Melting point	[°C]	47.5	1,2,3,4-tetrachlorobenzene	Mackay <i>et al.</i> , 2006; (Lide, 2003)
		54.5	1,2,3,5-tetrachlorobenzene	
		139.5	1,2,4,5-tetrachlorobenzene	
Boiling point	[°C]	254	1,2,3,4-tetrachlorobenzene	Mackay <i>et al.</i> , 2006; (Lide, 2003)
		246	1,2,3,5-tetrachlorobenzene	
		244.5	1,2,4,5-tetrachlorobenzene	
Henry's law constant	[Pa.m ³ /mol]	58.5	20 °C, 1,2,3,4-tetrachlorobenzene	Mackay <i>et al.</i> , 2006; (Staudinger and Roberts, 2001)
		99	1,2,3,5-tetrachlorobenzene	
		101	1,2,4,5-tetrachlorobenzene	

^a: reference between brackets refers to citation by Mackay *et al.*, 2006

5.1.3 Behaviour in the environment

Table 23. Selected environmental properties of tetrachlorobenzenes

Parameter	Unit	Value	Remark	Reference
Hydrolysis half-life	DT50 [d]		No hydrolysis at environmentally relevant concentrations	Mackay <i>et al.</i> , 2006
Photolysis half-life	DT50 [d]		No photolysis at environmentally relevant concentrations	Mackay <i>et al.</i> , 2006
Readily biodegradable				

5.1.4 Bioconcentration and biomagnification

An overview of the bioaccumulation data for tetrachlorobenzenes is given in Table 24. Detailed bioaccumulation data are tabulated in Appendix 1. Based on the studies by Muir *et al.* (2003) and Kelly *et al.* (2007), BMF values were calculated for the aquatic (BMF₁) and the aquatic-dependent (BMF₂) organisms for the tetrachlorobenzenes.

Table 24. Overview of bioaccumulation data for tetrachlorobenzenes

Parameter	Unit	Value	Remark	Reference
BCF (fish)	[L/kg]	1122	1,2,3,4-tetrachlorobenzene, <i>Cyprinus carpio</i>	METI (http://www.safe.nite.go.jp)
		1159	1,2,3,4-tetrachlorobenzene, <i>Cyprinus carpio</i>	METI (http://www.safe.nite.go.jp)
		2310	1,2,3,4-tetrachlorobenzene, <i>Poecilia reticulata</i>	Sijm <i>et al.</i> , 1993
		3888	1,2,3,5-tetrachlorobenzene, <i>Poecilia reticulata</i>	Könemann and Van Leeuwen, 1980
		3543	1,2,4,5-tetrachlorobenzene, <i>Cyprinus carpio</i>	METI (http://www.safe.nite.go.jp)
		3128	1,2,4,5-tetrachlorobenzene, <i>Cyprinus carpio</i>	METI (http://www.safe.nite.go.jp)
		4050	1,2,4,5-tetrachlorobenzene, <i>Jordamella floridæ</i>	Smith <i>et al.</i> , 1990
BMF ₁	[kg/kg]	2		
BMF ₂	[kg/kg]	3		

5.1.5 Human toxicological threshold limits and carcinogenicity

No R-phrases are assigned to the tetrachlorobenzenes.

1,2,3,4-tetrachlorobenzene

Based on a NOAEL of 34 mg/kg bw/day for male rats exposed by gavage and using an assessment factor of 10000 (10 for intraspecies variation; x 10 for interspecies variation; x 10 for less than chronic study; x 10 for limited data base including lack of adequate data on carcinogenicity and chronic and reproductive toxicity), a TDI of 3.4 µg/kg bw/day was derived (Health Canada, 1993).

1,2,3,5-tetrachlorobenzene

No R-phrases are assigned to 1,2,3,5-tetrachlorobenzene. Based on a NOAEL of 4.1 mg/kg bw/day for male rats exposed via the diet and using an assessment factor of 10000 (10 for intraspecies variation; x 10 for interspecies variation; x 10 for less than chronic study; x 10 for limited database including lack of adequate data on carcinogenicity and chronic and reproductive toxicity), a TDI of 0.41 µg/kg bw/day was derived (Health Canada, 1993).

1,2,4,5-tetrachlorobenzene

No R-phrases are assigned to 1,2,4,5-tetrachlorobenzene. Based on a NOAEL of 2.1 mg/kg bw/day for rats exposed via the diet and using an assessment factor of 10000 (10 for intraspecies variation; x 10 for interspecies variation; x 10 for less than chronic study; x 10 for limited database including lack of adequate data on carcinogenicity and chronic and reproductive toxicity), a TDI of 0.21 µg/kg bw/day was derived (Health Canada, 1993).

The derivation of the $MPC_{sp, water}$ and the $MPC_{hh food, water}$ is based on the worst-case scenario that all tetrachlorobenzenes are as toxic as 1,2,4,5-tetrachlorobenzene, e.g., on a TDI of 0.21 µg/kg bw/day and a NOAEL of 2.1 mg/kg bw/day.

5.2 Trigger values

This section reports on the trigger values for ERL water derivation (as required in the WFD framework).

Table 25. Tetrachlorobenzenes: collected properties for comparison to MPC triggers

Parameter	Value	Unit	Method/Source	Derived at section
Log $K_{p,susp-water}$	2.91	[-]	$K_{OC} \times f_{OC,susp}$ ^a	K_{OC} : 5.1.2
BCF	4050	[L/kg]	worst-case, BCF of 1,2,3,4-tetrachlorobenzene	5.1.4
BMF ₁	2	[kg/kg]		5.1.4
BMF ₂	3	[kg/kg]		5.1.4
Log K_{ow}	4.70	[-]	worst-case, log K_{ow} of 1,2,4,5-tetrachlorobenzene	5.1.2
R-phrases	-	[-]		5.1.5

^a. $f_{OC,susp} = 0.1 \text{ kg}_{OC}/\text{kg}_{solid}$ (EC, 2003).

- tetrachlorobenzenes have a worst-case of $\log K_{p, susp-water} < 3$; derivation of $MPC_{sediment}$ is not triggered
- tetrachlorobenzenes have a worst-case of $\log K_{p, susp-water} < 3$; expression of the MPC_{water} as $MPC_{susp, water}$ is not required
- tetrachlorobenzenes have a BCF > 100 L/kg; assessment of secondary poisoning is triggered
- tetrachlorobenzenes have no R classifications. However, since tetrachlorobenzenes have a high BCF value and a large uncertainty factor (10000) in the calculation of the TDI, the $MPC_{hh food water}$ is derived.

5.3 Derivation of ERLs for water

5.3.1 Aquatic toxicity data

An overview of the selected freshwater toxicity data for tetrachlorobenzenes is given in Table 26 and marine toxicity data are given in Table 27. Detailed toxicity data for tetrachlorobenzene are tabulated in Appendix 2.

Table 26. Tetrachlorobenzenes: selected freshwater toxicity data for ERL derivation

Chronic ^a Taxonomic group	NOEC/EC10 (mg/L)	Acute ^a Taxonomic group	L(E)C50 (mg/L)
<u>Crustacea</u>		<u>Bacteria</u>	
<i>Daphnia magna</i>	0.023^b	<i>Pseudomonas fluorescens</i>	2.16 ^d
<u>Insecta</u>		<u>Crustacea</u>	
<i>Chironomus riparius</i>	0.052	<i>Ceriodaphnia cf. dubia</i>	0.13
<u>Pisces</u>		<i>Daphnia carinata</i>	0.51
<i>Danio rerio</i>	0.1 ^c	<i>Daphnia magna</i>	0.54
<i>Jordanella floridae</i>	0.067	<u>Pisces</u>	
<i>Oncorhynchus mykiss</i>	0.245 ^c	<i>Jordanella floridae</i>	2.01
<i>Pimephales promelas</i>	0.25	<i>Pimephales promelas</i>	0.3 ^d
<i>Poecilia reticulata</i>	0.4	<i>Poecilia reticulata</i>	0.4 ^e

^a For detailed information see Appendix 2. Bold values are used for ERL derivation.

^b Geometric mean of 0.01 and 0.055 mg/L; most sensitive endpoint; reproduction. Data from 1,2,3,4-tetrachlorobenzene.

^c Most sensitive endpoint; growth.

^d Data from 1,2,4,5-tetrachlorobenzene.

^e Test duration 96 hours.

Table 27. Tetrachlorobenzenes: selected marine toxicity data for ERL derivation

Chronic ^a Taxonomic group	NOEC/EC10 (mg/L)	Acute ^a Taxonomic group	L(E)C50 (mg/L)
<u>Pisces</u>		<u>Bacteria</u>	
<i>Cyprinodon variegatus</i>	0.18	<i>Vibrio fischeri</i>	1.13 ^b
		<u>Pisces</u>	
		<i>Cyprinodon variegatus</i>	0.33

^a For detailed information see Appendix 2.

^b Geometric mean of 1.9 and 0.67 mg/L; exposure duration 15 minutes.

5.3.2 Treatment of fresh- and saltwater toxicity data

In line with the Fraunhofer guidance (Lepper, 2005), the datasets for freshwater and marine water are combined.

5.3.3 Mesocosm studies

No mesocosm studies are available for tetrachlorobenzenes.

5.3.4 Derivation of MPC_{water} and MPC_{marine}

5.3.4.1 MPC_{eco, water} and MPC_{eco, marine}

Freshwater

Reliable data for algae are lacking in both the acute and the chronic dataset. However, the less reliable (effects above the water solubility) or unassignable data for algae show that algae are a less sensitive taxonomic group. Therefore, ERLs can be derived for the tetrachlorobenzenes.

Chronic values are available for three taxonomic groups (crustaceans, insects and fish). Therefore, the $MPC_{eco, water}$ is derived using an assessment factor of 50 on the lowest NOEC value (0.023 mg/L for *Daphnia*). Thus, the $MPC_{eco, water}$ is $0.023 \text{ mg/L} / 50 = 0.00046 \text{ mg/L} = 0.46 \mu\text{g/L}$

Marine water

Based on the data in Tables 26 and 27, an additional assessment factor of 10 is used on the lowest NOEC for the derivation of the $MPC_{eco, marine}$. Thus, the $MPC_{eco, marine}$ is $0.46 \mu\text{g/L} / 10 = 4.6 \times 10^{-2} \mu\text{g/L}$.

5.3.4.2 $MPC_{sp, water}$ and $MPC_{sp, marine}$

Freshwater

All tetrachlorobenzenes have a BCF > 100 L/kg, thus assessment of secondary poisoning is triggered. Based on a worst-case $MPC_{oral, mammal}$ of 0.47 mg/kg fd (based on a 90-day rat study with a NOAEL of 2.1 mg/kg bw per day by NTP, 1991), a BCF of 4050 L/kg and a BMF₁ of 2 kg/kg, the $MPC_{sp, water}$ is $5.8 \times 10^{-2} \mu\text{g/L}$ (Van Vlaardingen and Verbruggen, 2007, section 3.1.4).

Marine water

The $MPC_{sp, marine}$ is based on the same data as the $MPC_{sp, water}$. In order to account for the longer food chains in the marine environment, an additional biomagnification factor is used. Because of the BCF value of 2310 L/kg, the BMF₂ has a value of 3. Therefore, the $MPC_{sp, marine}$ is $1.9 \times 10^{-2} \mu\text{g/L}$.

5.3.4.3 $MPC_{hh food, water}$

Derivation of $MPC_{hh food, water}$ for tetrachlorobenzenes is not triggered (Table 25). However, since tetrachlorobenzenes have a high BCF value and a large uncertainty factor (10000) in the calculation of the TDI (0.21 µg/kg bw day), the $MPC_{hh food, water}$ is calculated. Using the formulas in the INS guidance, the $MPC_{hh food, water}$ is $0.0016 \mu\text{g/L} = 1.6 \times 10^{-3} \mu\text{g/L}$. This value is valid for freshwater as well as for the marine compartment.

5.3.5 Selection of MPC_{water} and MPC_{marine}

Freshwater

The lowest value of the routes included is the $MPC_{hh food, water}$. Therefore, the MPC_{water} is $1.6 \times 10^{-3} \mu\text{g/L}$.

Marine water

The lowest value of the routes included is the $MPC_{hh food, water}$. Therefore, the MPC_{marine} is $1.6 \times 10^{-3} \mu\text{g/L}$.

5.3.6 $MPC_{dw, water}$

Because no A1 value or DWS is available for tetrachlorobenzenes, the $MPC_{dw, water, provisional}$ is derived using the formula in section 3.1.6 of the INS guidance. Using the TDI of 0.21 µg/kg bw day, an average bodyweight of 70 kg and an average uptake of drinking water of 2 L, the $MPC_{dw, water, provisional}$ is 0.74 µg/L. The $MPC_{dw, water, provisional}$ is not taken into account for the derivation of the final MPC_{water} .

5.3.7 Derivation of $MAC_{eco, water}$ and $MAC_{eco, marine}$

Freshwater

The acute dataset contains data for bacteria, crustaceans and fish. Reliable data for algae are lacking in both the acute and the chronic dataset. However, the less reliable (effects above the water solubility) or

unassignable data for algae indicate that algae a less sensitive taxonomic group. Therefore, a MAC value is derived for tetrachlorobenzenes. All tetrachlorobenzenes have a BCF > 100 L/kg. Therefore, an assessment factor of 1000 is used on the lowest L(E)C₅₀ value (0.13 mg/L, *Ceriodaphnia dubia*) for the derivation of the MAC_{eco, water}, resulting in a MAC_{eco, water} of 0.13 mg/L / 1000 = 0.13 µg/L.

Marine water

Based on the data in Tables 32 and 33 and the bioaccumulative properties of 1,2,3,4-tetrachlorobenzene, the MAC_{eco, marine} is derived by applying an assessment factor of 10000 to the lowest L(E)C₅₀ value. Thus, the MAC_{eco, marine} is 0.13 mg/L / 10000 = 1.3 x 10⁻² µg/L.

5.3.8 Derivation of NC

Freshwater

The NC is set a factor of 100 below the MPC. Therefore, the NC_{water} is 1.6 x 10⁻³ µg/L / 100 = 1.6 x 10⁻⁵ µg/L.

Marine water

The NC is set a factor of 100 below the MPC. Therefore, the NC_{water} is 1.6 x 10⁻³ µg/L / 100 = 1.6 x 10⁻⁵ µg/L.

5.3.9 Derivation of SRC_{eco, water}

The SRC_{eco, water} for tetrachlorobenzenes is based on the geometric mean of the L(E)C₅₀ values using an assessment factor of 10, because NOEC values are available for both *Daphnia* and fish and the geometric mean of the L(E)C₅₀ value is less than 10 times the geometric mean of the NOEC values. Thus, the SRC_{eco, water} is 0.58 mg/L / 10 = 0.058 mg/L = 58 µg/L. This value is valid for freshwater as well as for the marine compartment.

5.4 Derivation of ERLs for sediment

The log K_{p, susp-water} of all tetrachlorobenzenes is below the trigger value of 3, therefore, ERLs are not derived for sediment.

5.5 Derivation of ERLs for soil

5.5.1 Toxicity data for soil

Table 28. tetrachlorobenzenes: selected soil toxicity data for ERL derivation

Chronic ^a Taxonomic group	NOEC/EC10 (mg/kg)	Acute ^a Taxonomic group	L(E)C50 (mg/kg)
<u>Annelida</u>			
		<i>Eisenia andrei</i>	203 ^b
		<i>Eisenia andrei</i>	275 ^b
		<i>Lumbricus rubellus</i>	248 ^b
		<i>Lumbricus rubellus</i>	303 ^b

^a For detailed information see Appendix 3.

^b Normalised to standard soil.

5.5.2 Derivation of MPC_{soil}

5.5.2.1 MPC_{eco, soil}

The dataset of tetrachlorobenzenes contains four L(E)C₅₀ values for two species: 203 and 275 mg/kg_{dwt} for *Eisenia andrei* (geometric mean 236 mg/kg_{dwt}) and 248 and 303 mg/kg_{dwt} for *Lumbricus rubellus* (geometric mean 274 mg/kg_{dwt}). According to the INS guidance, an assessment factor of 1000 should be applied to the lowest L(E)C₅₀ for the derivation of the MPC_{eco, soil}. Therefore, the MPC_{eco, soil} is 236 mg/kg_{dwt} / 1000 = 2.4 x 10² µg/kg_{dwt}.

5.5.2.2 MPC_{sp, soil}

The log K_{ow} of the tetrachlorobenzenes exceeds the trigger value of 3 (4.70 for 1,2,4,5-tetrachlorobenzene), so the MPC_{sp, soil} should be derived. One BSAF value of 104 g/g lipid was available for earthworms (Belfroid *et al.*, 1994). Using an earthworm lipid content of 1.2% and a correction to standard Dutch soil in the formulas in section 3.3.5 of the INS guidance, the MPC_{sp, soil} is 0.488 mg/kg_{dwt} = 4.9 x 10² µg/kg_{dwt}.

5.5.2.3 MPC_{human, soil}

The MPC_{human, soil} was derived using the formulas in section 3.3.6 of the INS Guidance (Van Vlaardingen en Verbruggen, 2007). These calculations resulted in an MPC_{human, soil} of 2.6 µg/kg_{dwt} for the consumption of root crops.

5.5.3 Selection of MPC_{soil}

The lowest value of the routes included is the MPC_{human, soil}. Therefore, the MPC_{soil} for tetrachlorobenzenes is 2.6 µg/kg_{dwt}.

5.5.4 Derivation of NC_{soil}

The NC is set a factor of 100 below the MPC. Therefore, the NC_{soil} is 2.6 µg/kg_{dwt} / 100 = 2.6 x 10⁻² µg/kg_{dwt}.

5.5.5 Derivation of SRC_{eco, soil}

Since only acute values for the toxicity of 1,2,3,4-tetrachlorobenzene in soil are available, the SRC_{eco, soil} is derived by comparing the geometric mean of the L(E)C₅₀ values with an assessment factor of 10 with the result of the equilibrium partitioning method (Van Vlaardingen and Verbruggen, 2007). For 1,2,3,4-tetrachlorobenzene, the geometric mean of the L(E)C₅₀ values is 254 mg/kg_{dwt}. Using an assessment factor of 10, the SRC_{eco, soil} is 254 mg/kg_{dwt} / 10 = 2.5 x 10⁴ µg/kg_{dwt}. The SRC_{eco, soil} derived using the equilibrium partitioning method is 2.9 x 10⁴ µg/kg_{dwt}. Therefore, the SRC_{eco, soil} is set to 2.5 x 10⁴ µg/kg_{dwt}.

5.6 Derivation of ERLs for groundwater

5.6.1 Derivation of MPC_{gw}

5.6.1.1 MPC_{eco, gw}

Since no toxicity data are available for the groundwater compartment, the MPC_{eco, water} for surface water is taken as a substitute. Therefore, the MPC_{eco, gw} is 0.46 µg/L.

5.6.1.2 MPC_{human, gw}

The MPC_{dw, water, provisional} is used as a substitute for the MPC_{human, gw}. Therefore, the MPC_{human, gw} is 0.74 µg/L.

5.6.2 Selection of MPC_{gw}

The lowest value of the routes included is the MPC_{eco, water}. Therefore, the MPC_{gw} is 0.46 µg/L.

5.6.3 Derivation of NC_{gw}

The NC is set a factor of 100 below the MPC. For groundwater therefore, the NC_{gw} is $0.46 \mu\text{g/L} / 100 = 4.6 \times 10^{-3} \mu\text{g/L}$.

5.6.4 Derivation of SRC_{eco, gw}

Since no toxicity data are available for the groundwater compartment, the SRC_{eco, water} for surface water is taken as a substitute. Therefore, the SRC_{eco, gw} is 58 µg/L.

5.7 Derivation of ERLs for air

5.7.1.1 MPC_{eco, air}

Since no data on the ecotoxicity of tetrachlorobenzenes in air are available, no MPC_{eco, air} can be derived.

5.7.1.2 MPC_{human, air}

The MPC_{human, air} is set at the same value as the TCA. Since no TCA for tetrachlorobenzenes is available, no MPC_{human, air} can be derived.

5.7.2 Selection of MPC_{air}

The MPC_{air} for tetrachlorobenzenes cannot be derived.

5.8 Overview of MPCs for tetrachlorobenzenes

In Table 29, an overview of the ERLs derived for tetrachlorobenzenes is given.

Table 29. Tetrachlorobenzenes: derived NC, MPC, MAC_{eco} and SRC_{eco}

ERL	Unit	NC	MPC	MAC _{eco}	SRC _{eco}
Water	µg/L	1.6×10^{-5}	1.6×10^{-3}	0.13	58
Marine	µg/L	1.6×10^{-5}	1.6×10^{-3}	1.3×10^{-2}	58
Soil	µg/kg	2.6×10^{-2}	2.6	n.a.	2.5×10^3
Groundwater	µg/L	4.6×10^{-3}	0.46	n.a.	58
Air	µg/m ³	n.a.	n.d.	n.a.	n.a.
Drinking water	µg/L	n.a.	0.74	n.a.	n.a.

5.9 Comparison of derived ERLs with monitoring data

Monitoring data for the Rhine from the years 2003–2006, obtained from RIWA (Association of River Waterworks), show that at all sampling occasions and locations, the concentration of tetrachlorobenzenes in water was usually below detection limits (0.01 µg/L). However, it should be noted that monitoring data are not reported for all tetrachlorobenzenes. In 2003–2005, measured concentrations are reported for 1,2,3,4- and 1,2,4,5-tetrachlorobenzene. In 2006, only for 1,2,4,5-tetrachlorobenzene concentrations were reported.

On two occasions in 2004, a concentration of 0.05 µg 1,2,3,4-tetrachlorobenzene/L was measured, which would result in an exceedance of the MPC_{water} and potentially the MAC for tetrachlorobenzenes. In more recent monitoring reports, no tetrachlorobenzenes were measured above detection limits.

5.10 Sum limits

The small differences in toxicity of the individual tetrachlorobenzenes can result in unacceptable risks when a mixture of tetrachlorobenzenes is present at one location. In order to prevent this accumulation of toxicity, the use of a sum limit for tetrachlorobenzenes is proposed. In this approach, the sum of the concentrations of the combination of the three individual tetrachlorobenzenes should not exceed the ERLs for the grouped tetrachlorobenzenes (Table 29). It should be noted that the sum limit only applies when all three tetrachlorobenzenes are included in the analysis.

6 Conclusions

In this report, the risk limits negligible concentration (NC), maximum permissible concentration (MPC), maximum acceptable concentration for ecosystems (MAC_{eco}), and serious risk concentration for ecosystems (SRC_{eco}) are derived for monochlorobenzene, the group of three dichlorobenzenes (1,2-dichlorobenzene, 1,3-dichlorobenzene, and 1,4-dichlorobenzene) and the group of three tetrachlorobenzenes (1,2,3,4-tetrachlorobenzene, 1,2,3,5-tetrachlorobenzene and 1,2,4,5-tetrachlorobenzene) in water, groundwater, soil and air. No risk limits were derived for water organisms via the sediment compartment because the triggers to derive such limits were not exceeded.

The ERLs that were obtained are summarised in the table below. Because of the small difference in toxicity of the individual isomers, the risk limits for di- and tetrachlorobenzenes refer to a sum limit. In this approach, the sum of the concentrations of the individual di- or tetrachlorobenzenes should not exceed the ERLs for the group. The sum limit only applies when all isomers are included in the analysis. It should further be noted that the respective chlorobenzenes may occur simultaneously, and an additive effect cannot be excluded.

Table 30. Derived NC, MPC, MAC_{eco}, and SRC_{eco} values

ERL	Unit	Substance	NC	MPC	MAC _{eco}	SRC _{eco}
Water	µg/L	monochlorobenzene	0.32	32	40	1.9×10^3
Marine	µg/L	monochlorobenzene	3.2×10^{-2}	3.2	4.0	1.9×10^3
Soil	µg/kg	monochlorobenzene	4.3	4.3×10^2	n.a.	2.6×10^4
Groundwater	µg/L	monochlorobenzene	0.32	32	n.a.	1.9×10^3
Air	µg/m ³	monochlorobenzene	n.a.	5.0×10^2	n.a.	n.a.
Drinking water	µg/L	monochlorobenzene	n.a.	2.1×10^2	n.a.	n.a.
Water	µg/L	dichlorobenzenes	6.9×10^{-2}	6.9	20	6.8×10^2
Marine	µg/L	dichlorobenzenes	2.0×10^{-2}	2.0	2.0	6.8×10^2
Soil	µg/kg	dichlorobenzenes	3.1	3.1×10^2	n.a.	2.0×10^4
Groundwater	µg/L	dichlorobenzenes	0.20	20	n.a.	6.8×10^2
Air	µg/m ³	dichlorobenzenes	n.a.	n.d.	n.a.	n.a.
Drinking water	µg/L	dichlorobenzenes	n.a.	3.8×10^2	n.a.	n.a.
Water	µg/L	tetrachlorobenzenes	1.6×10^{-5}	1.6×10^{-3}	0.13	58
Marine	µg/L	tetrachlorobenzenes	1.6×10^{-5}	1.6×10^{-3}	1.3×10^{-2}	58
Soil	µg/kg	tetrachlorobenzenes	2.6×10^{-2}	2.6	n.a.	2.5×10^4
Groundwater	µg/L	tetrachlorobenzenes	4.6×10^{-3}	0.46	n.a.	58
Air	µg/m ³	tetrachlorobenzenes	n.a.	n.d.	n.a.	n.a.
Drinking water	µg/L	tetrachlorobenzenes	n.a.	0.74	n.a.	n.a.

n.d. = not derived.

n.a. = not applicable.

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Appendix 1. Information on bioconcentration

Detailed information on bioconcentration is presented in the following tables. For explanation of symbols and abbreviations, see Van Vlaardingen and Verbruggen, 2007. For information on the reliability index, see section 2.2 of the main text. In case of citations, the original reference is given behind the secondary source. If cited references could be retrieved, they were checked and included in the reference list. Otherwise, evaluation was based on the secondary source.

Legend to column headings													
A	test water analysed Y(es)/N(o)												
Test type	S = static; Sc = static closed; R = renewal; F = flow through; CF = continuous flow; IF = intermittent flow system												
Purity	refers to purity of active substance or content of active in formulation												
Test water	am = artificial medium; dtw = dechlorinated tap water; dw = deionised/dechlorinated/distilled water; nw= natural water; rw = reconstituted water; rtw = reconstituted tap water; tw = tap water												
T	temperature												
Ri	Reliability index according to Klimisch <i>et al.</i> (1997); asterisk indicates citation												

Table A1.1. Bioconcentration data for monochlorobenzene

Species	Species properties	Subst. purity [%]	A	Test type	Test water	pH	Hardness/ Salinity	Temp. [°C]	Exp. time	Exp. conc. [mg/L]	BCF	BCF type	Method	Ri	Notes	Reference
Algae																
<i>Chlorella fusca</i>									24h	0.05	50			4		Freitag <i>et al.</i> , 1984
<i>Oedogonium cardiacum</i>									24h	0.01-0.1	4185	cosm		4	2	Lu and Metcalf, 1975
<i>Pseudokirchneriella subcapitata</i>									24h	10	2172			4		Casserly <i>et al.</i> , 1983
Crustacea																
<i>Daphnia magna</i>									24h	0.01-0.1	2789	cosm		4	2	Lu and Metcalf, 1975
Insecta																
<i>Culex quinquefasciatus</i>									24h	0.01-0.1	1292	cosm		4	2	Lu and Metcalf, 1975
Mollusca																
<i>Physa</i>									24h	0.01-0.1	1313	cosm		4	2	Lu and Metcalf, 1975
Pisces																
<i>Gambusia affinis</i>																Lu and Metcalf, 1975
<i>Lepomis macrochirus</i>																IUCLID, 2000: Monsanto and Bayer AG
<i>Leuciscus idus</i>																Freitag <i>et al.</i> , 1984
<i>Leuciscus idus</i>																Freitag, 1987
<i>Leuciscus idus</i>																Freitag <i>et al.</i> , 1984
<i>Pimephales promelas</i>	mature female			Sc	nw	>6.6	75-85	21-22	until steady state	3.6	84	kinetic		2		De Wolf and Lieder, 1998
<i>Pimephales promelas</i>									28d		447			4		BUA.054: ref. 31

Species	Species properties	Subst. purity [%]	A	Test type	Test water	pH	Hardness/ Salinity	Temp. [°C]	Exp. time	Exp. conc. [mg/L]	BCF	BCF type	Method	Ri	Notes	Reference
Cyprinus carpio	32 g, 12 cm	GC-FID	CF					25±2	56d	46	25	wwt	kinetic	2	5	METI (http://www.safe.nite.go.jp)
Cyprinus carpio fish	32 g, 12 cm	GC-FID	CF					25±2	56d	8.6	11	wwt	kinetic	2	5	METI (http://www.safe.nite.go.jp)
			F								12-450			4		Hesse et al., 1991; Kenaga, 1980 and EPA, 1984
											10-447			4		IUCLID, 2000: Petrasol BV

Notes

- 1 Unit unclear
- 2 Based on dry weight
- 3 According to ASTM guidelines
- 4 Exposure duration too short
- 5 Recalculated from original data

Table A1.2. Bioconcentration data for 1,2-dichlorobenzene

Species	Species properties	Subst. purity [%]	A	Test type	Test water	pH	Hardness/ Salinity	Temp. [°C]	Exp. time	Exp. conc. [mg/L]	BCF	BCF type	Method	Ri	Notes	Reference	
Algae																	
<i>cyanobacteria and green algae</i>								40h		6212			4			BUA.053: Davis et al., 1983	
<i>Pseudokirchneriella subcapitata</i>								24h	10	19700			4			Casserly et al., 1983	
<i>Pseudokirchneriella subcapitata</i>								24h	2	10080			3	1		Casserly et al., 1983	
Crustacea																	
<i>Callinectes sapidus</i>	0.5% lipid		GC-MS						9 ng/L	144			equi	4	2	Pereira et al., 1988	
Pisces																	
<i>Cynoscion nebulosus</i>	2.3% lipid		GC-MS							9 ng/L	142			4	2	Pereira et al., 1988	
<i>Cyprinus carpio</i>								21d	70.3 µg/L	40			equi	4		BUA.053: Tadokoro and Tomita, 1987	
<i>Cyprinus carpio</i>								56d	0.01	90-260			4	3		IUCLID, 2000: Bayer	
<i>Cyprinus carpio</i>								56d	0.1	150-230			4	3		IUCLID, 2000: Bayer	
<i>Cyprinus carpio</i>	31 g, 11 cm		GC-ECD	CF				25±2	56d	53	178	wwt	k1/k2	2	6	METI (http://www.safe.nite.go.jp)	
<i>Cyprinus carpio</i>	31 g, 11 cm		GC-ECD	CF				25±2	56d	6.3	174	wwt	k1/k2	2	6	METI (http://www.safe.nite.go.jp)	
<i>Ictalurus furcatus</i>	3.3% lipid		GC-MS							9 ng/L	218			equi	4	2	Pereira et al., 1988
<i>Lepomis macrochirus</i>	0.37-0.95 g		IF	nw	7.1	35		16	14d	0.00789	89			4*	4	Veith et al., 1980	
<i>Lepomis macrochirus</i>	0.37-0.95 g		F	nw	6.3-7.9			16	14d	0.00789	89				2	4	Barrows et al., 1978
<i>Micropogonias undulatus</i>										9 ng/L	192			equi	4	2	BUA.053: Pereira et al., 1988
<i>Oncorhynchus mykiss</i>									119d	0.047 µg/L	270			equi	3	5	Oliver and Niimi, 1983
<i>Oncorhynchus mykiss</i>									105d	0.940 µg/L	560			equi	3	5	Oliver and Niimi, 1983
<i>Pimephales promelas</i>	0.39 g	95	GC	R	tw				8d	10.3	454	steady state	k1/k2	2	3	Sijm et al., 1993	
<i>Poecilia reticulata</i>	3 m/o	95	GC	R	tw				8d	1.5	251	steady state	k1/k2	2	3	Sijm et al., 1993	

Notes

- 1 Mixture with 7 other aromatic compounds
- 2 Field study in contaminated estuary
- 3 According to OECD guidelines
- 4 Closed system; equilibrium reached after 14 days
- 5 Equilibrium after 7-8 days; loading too high (18 g fish/L)
- 6 Recalculated from original data

Table A1.3. Bioconcentration data for 1,3-dichlorobenzene

Species	Species properties	Subst. A purity [%]	Test type	Test water	pH	Hardness/ Salinity	Temp. [°C]	Exp. time	Exp. conc. [mg/L]	BCF	BCF type	Method	Ri	Notes	Reference
Pisces															
<i>Cyprinus carpio</i>			F					8w	0.1	57-229			4		BUA.133: MITI, 1992
<i>Cyprinus carpio</i>			F					8w	0.01	58-370			4		BUA.133: MITI, 1992
<i>Cyprinus carpio</i>			F				25	64d	0.1	57-229			4*		IUCLID, 2000: Hoechst AG and Clariant GmbH
<i>Cyprinus carpio</i>			F				25	64d	0.01	58-370			4*		IUCLID, 2000: Hoechst AG and Clariant GmbH
<i>Cyprinus carpio</i>	4.8% lipids, 32.2 g, 10.7 cm	98	GC-MS	CF				25±2	56d	83	138	wwt	k1/k2	2 5	METI (http://www.safe.nite.go.jp)
<i>Cyprinus carpio</i>	4.8% lipids, 32.2 g, 10.7 cm	98	GC-MS	CF				25±2	56d	8.9	195	wwt	k1/k2	2 5	METI (http://www.safe.nite.go.jp)
<i>Lepomis macrochirus</i>	0.37-0.95 g			F nw	6.3-7.9			16	14d	0.107	66			2	Barrows et al., 1978
<i>Lepomis macrochirus</i>	0.37-0.94 g		F	nw	7.1	35	16	14d	0.107	66		k1/k2	4*	Veith, 1981	
<i>Lepomis macrochirus</i>			F	nw	7.1	35	16	14d	0.107	66		k1/k2	4*	IUCLID, 2000: Hoechst AG and Clariant GmbH	
<i>Oncorhynchus mykiss</i>			F					119d	0.000028	420			3 2		Oliver and Niimi, 1983
<i>Oncorhynchus mykiss</i>			F					105d	0.00069	740			3 2		Oliver and Niimi, 1983
<i>Oncorhynchus mykiss</i>			F				15	105d	0.00069	740		k1/k2	4* 1		IUCLID, 2000: Hoechst AG and Clariant GmbH
<i>Oncorhynchus mykiss</i>			F				15	119d	0.000028	420		k1/k2	4* 1		IUCLID, 2000: Hoechst AG and Clariant GmbH
<i>Pimephales promelas</i>										98			4		Veith, 1981
<i>Pimephales promelas</i>		98					25	32d		97			4		IUCLID, 2000: Hoechst AG and Clariant GmbH
<i>Pimephales promelas</i>								30d		98			4*		IUCLID, 2000: Hoechst AG and Clariant GmbH
<i>Pimephales promelas</i>	mature female, 1.45-4.71 g	98		Sc nw	>6.6	75-85	21-22	until steady state	2.5	728	kinetic		2		De Wolf and Lieder, 1998
<i>Poecilia reticulata</i>	2y old; 15-20 mm; 206-283 mg; 5% lipids	>97	GC-ECD	S tw/dw			13	48h		294	steady state	Cf/Cw	3 3,4		Opperhuizen et al., 1988
<i>Poecilia reticulata</i>	2y old; 15-20 mm; 206-283 mg; 5% lipids	>97	GC-ECD	S tw/dw			19	48h		301	steady state	Cf/Cw	3 3,4		Opperhuizen et al., 1988
<i>Poecilia reticulata</i>	2y old; 15-20 mm; 206-283 mg; 5% lipids	>97	GC-ECD	S tw/dw			28	48h		330	steady state	Cf/Cw	3 3,4		Opperhuizen et al., 1988
<i>Poecilia reticulata</i>	2y old; 15-20 mm; 206-283 mg; 5% lipids	>97	GC-ECD	S tw/dw			33	48h		346	steady state	Cf/Cw	3 3,4		Opperhuizen et al., 1988
<i>Poecilia reticulata</i>		>97						33	48h	sol. limit	6918			4	IUCLID, 2000: Hoechst AG and Clariant GmbH

Notes

- 1 Closed system; equilibrium reached after 14 days
 2 Loading too high (18 g fish/L)
 3 Exposure in a mixture
 4 Equilibrium was reached in a preliminary study at 48h but this does not seem to be reliable; original BCF in ref reported based on lipid contents, recalculated using 5% lipids
 5 Recalculated from original data

Table A1.4. Bioconcentration data for 1,4-dichlorobenzene

Species	Species properties	Subst. purity [%]	A	Test type	Test water	pH	Hardness/ Salinity	Temp. [°C]	Exp. time	Exp. conc. [mg/L]	BCF	BCF type	Method	Ri	Notes	Reference
Pisces																
<i>Cyprinus carpio</i>		98.9	HPLC-UV	CF		7.3-7.6		25±2	35d	1.9	69	wwt	k1/k2	2	1	METI (http://www.safe.nite.go.jp)
<i>Cyprinus carpio</i>		98.9	HPLC-UV	CF		7.3-7.6		25±2	35d	0.18	80	wwt	k1/k2	2	1	METI (http://www.safe.nite.go.jp)
<i>Poecilia reticulata</i>	female, 0.62 g; 5.4% lipid weight	>97	GC	F	tw		89	21	2 d	0.116	97	steady state	Cf/Cw	2	2,3	Könemann and Van Leeuwen, 1980

Notes

- 1 Recalculated from original data
 2 Exposure in a mixture
 3 Total exposure was 19 d but BCF was calculated after 2 d of exposure. BCF recalculated from original value (1800), which was based on lipid content.

Table A1.5. Bioconcentration data for 1,2,3,4-tetrachlorobenzene.

Species	Species properties	Subst. purity [%]	A	Test type	Test water	pH	Hardness/ Salinity	Temperature [°C]	Exp. time	Exp. conc. [mg/L]	BCF	BCF type	Method	Ri	Notes	Reference
Pisces																
<i>Cyprinus carpio</i>	4.5% lipids, 26.7 g, 10.1 cm	97	GC-ECD	CF				25±1	56 d	9.0	1122	wwt	k1/k2	2	1	METI (http://www.safe.nite.go.jp)
<i>Cyprinus carpio</i>	4.5% lipids, 26.7 g, 10.1 cm	97	GC-ECD	CF				25±1	56 d	0.84	1159	wwt	k1/k2	2	1	METI (http://www.safe.nite.go.jp)
<i>Poecilia reticulata</i>	3 m/o	95	GC	R	tw			8d	0.36	2310	steady state	k1/k2	2	2	Sijm et al., 1993	
<i>Poecilia reticulata</i>	2y old; 15-20 mm; 206-283 mg; 5% lipids	>97	GC-ECD	S	tw/dw		13	48h		2506	steady state	Cf/Cw	3	3,4	Opperhuizen et al., 1988	
<i>Poecilia reticulata</i>	2y old; 15-20 mm; 206-283 mg; 5% lipids	>97	GC-ECD	S	tw/dw		19	48h		2748	steady state	Cf/Cw	3	3,4	Opperhuizen et al., 1988	
<i>Poecilia reticulata</i>	2y old; 15-20 mm; 206-283 mg; 5% lipids	>97	GC-ECD	S	tw/dw		28	48h		2812	steady state	Cf/Cw	3	3,4	Opperhuizen et al., 1988	
<i>Poecilia reticulata</i>	2y old; 15-20 mm; 206-283 mg; 5% lipids	>97	GC-ECD	S	tw/dw		33	48h		3459	steady state	Cf/Cw	3	3,4	Opperhuizen et al., 1988	
<i>Poecilia reticulata</i>	150 mg, 18 mm	>95		Sc	tw/dw			290h	0.06	6600	whole fish	equi	3	5	Schrap and Opperhuizen, 1990	

Notes

- 1 Recalculated from original data
 2 According to OECD guidelines
 3 Exposure in a mixture
 4 Equilibrium was reached in a preliminary study at 48 h but this does not seem to be reliable; original BCF in ref reported based on lipid contents, recalculated using 5% lipids mortality during test

Table A1.5. Bioconcentration data for 1,2,3,5-tetrachlorobenzene

Species	Species properties	Subst. purity [%]	A	Test type	Test water	pH	Hardness/ Salinity	Temp. [°C]	Exp. time	Exp. conc. [mg/L]	BCF	BCF type	Method	Ri	Notes	Reference
Pisces																
<i>Lepomis macrochirus</i>	0.37-0.95 g			F	nw	6.3-7.9		16	28d	0.00772	1800			3	1	Barrows et al., 1978
<i>Poecilia reticulata</i>	female, 0.62 g, lipid content 5.4%		GC	F	tw			21	7d	0.0012	3888	steady state	Cf/Cw	2	2	Könemann and Van Leeuwen, 1980

Notes

1 Equilibrium not reached after 28 days

2 Total exposure was 19 d, but BCF was calculated after 7 d of exposure. BCF recalculated from original value (72000), which was based on lipid content. Original value lipid based (72000 L/kg) was recalculated; exposure to a mixture of chlorobenzenes; actual concentration measured

Table A1.6. Bioconcentration data for 1,2,4,5-tetrachlorobenzene

Species	Species properties	Subst. purity [%]	A	Test type	Test water	pH	Hardness/ Salinity	Temp. [°C]	Exp. time	Exp. conc.. [mg/L]	BCF	BCF type	Method	Ri	Notes	Reference
Algae																
<i>Seleniastrum capricornutum</i>	exponential growth phase										7700		equi	4		BUA.086: Mailhot, 1987
Pisces																
<i>Cyprinus carpio</i>	5.7% lipids, 24.7 g, 9.7 cm	99	GC-ECD	CF				25±1	56 d	7.2	3543	wwt	k1/k2	2	1	METI (http://www.safe.nite.go.jp)
<i>Cyprinus carpio</i>	5.7% lipids, 24.7 g, 9.7 cm	99	GC-ECD	CF				25±1	56 d	0.73	3128	wwt	k1/k2	2	1	METI (http://www.safe.nite.go.jp)
<i>Cyprinus carpio</i>	20-30 g			F				25	28d	7.24 µg/L	1122		equi	4		BUA.086: Tadokoro and Tomita, 1987
<i>Cyprinus carpio</i>	20-30 g			F				25	28d	7.24 µg/L	3981		equi	4		BUA.086: Tadokoro and Tomita, 1987
<i>Jordanella floridae</i>	4-6mo; lipid content 8.5%			F				25	28d	3.02 µg/L	4050		k1/k2	2		Smith et al., 1990
<i>Oncorhynchus mykiss</i>	not yet fully grown			F	nw				119d	1 ± 0.5	6200		equi	3	2	Oliver and Niimi, 1983
<i>Oncorhynchus mykiss</i>	not yet fully grown			F	nw				105d	21 ± 13 ng/L	15000		equi	3	2	Oliver and Niimi, 1983

Notes

1 Recalculated from original data

2 Loading too high (18 g fish/L)

Appendix 2. Detailed aquatic toxicity data

Detailed information on bioconcentration is presented in the following tables. For an explanation of the symbols and abbreviations, see Van Vlaardingen and Verbruggen, 2007. For information on the reliability index, see section 2.2 of the main text. In case of citations, the original reference is given behind the secondary source. If cited references could be retrieved, they were checked and included in the reference list. Otherwise, evaluation was based on the secondary source.

Legend to column headings													
A	test water analysed Y(es)/N(o)												
Test type	S = static; Sc = static closed; R = renewal; F = flow through; CF = continuous flow; IF = intermittent flow system												
Purity	refers to purity of active substance or content of active substance in formulation												
Test water	am = artificial medium; dtw = dechlorinated tap water; dw = deionised/dechlorinated/distilled water; nw= natural water; rw = reconstituted water; rtw = reconstituted tap water; tw = tap water												
T	temperature												
Ri	Reliability index according to Klimisch et al. (1997); asterisk indicates citation												

Table A2.1. Acute toxicity of monochlorobenzene to freshwater organisms

Species	Species properties	A	Test type	Subst. purity [%]	Test water	pH	Temp. [°C]	Hardness CaCO ₃ [mg/L]	Exp. time	Criterion	Test endpoint	Value	Ri	Notes	Reference		
Bacteria																	
<i>Activated sludge</i>									30min	EC50	resp. inhibition	140	4	1	IUCLID, 2000		
<i>Activated sludge</i>									30min	EC50	resp. inhibition	2950	3	2, 40	IUCLID, 2000: Bayer AG		
<i>Activated sludge</i>									24d	EC50		20	4	3	IUCLID, 2000: Bayer AG		
<i>Aerobic heterotrophs</i>									15h	EC50	resp. inhibition	310	4	4	IUCLID, 2000: Bayer AG		
<i>Pseudomonas fluorescens</i>										EC50	luminescence	118.3	4*	8	Warne et al., 1999		
<i>Pseudomonas fluorescens</i>		Y	S	>98	am	7.1	25			EC50	luminescence	118.5	2	6	Boyd et al., 1998		
Protozoa																	
<i>Tetrahymena pyriformis</i>										Sc	growth inhibition	83.5	3	9	Schultz, 1999		
Algae																	
<i>Ankistrodesmus falcatus</i>										N	photosynth.	50.3	3	10	Wong et al., 1984		
<i>Chlamydomonas angulosa</i>	5*10 ⁴ cells/mL									N	photosynth.	57	3	11	Hutchinson et al., 1980		
<i>Clamydomonas sp.</i>											EC50	56.6	4*		IUCLID, 2000: Petrasol BV		
<i>Chlorella vulgaris</i>	20*10 ⁴ cells/mL									N	photosynth.	99	3	12	Hutchinson et al., 1980		
<i>Cyclotella sp.</i>											EC50	235.7	4	39	IUCLID, 2000: Petrasol BV		
<i>Pseudokirchneriella subcapitata</i>											EC50	280	4		IUCLID, 2000: Bayer AG		
<i>Pseudokirchneriella subcapitata</i>		Y	Sc		am						96h	EC50	growth	12.5	2	15	Calamari et al., 1983

Species	Species properties	A	Test type	Subst. purity [%]	Test water	pH	Temp. [°C]	Hardness CaCO ₃ [mg/L]	Exp. time	Criterion	Test endpoint	Value [mg/L]			Ri	Notes	Reference
												Value [mg/L]	Ri	Notes			
<i>Pseudokirchneriella subcapitata</i>	5mln cells/l	Y	Sc		am		20		96h	EC50	growth inhibition	12.5	2	16			Galassi and Vighi, 1981
<i>Pseudokirchneriella subcapitata</i>			Sc						96h	EC50	cell number	12.5	4*				IUCLID, 2000: Bayer AG
<i>Pseudokirchneriella subcapitata</i>									96h	EC50		224	4				IUCLID, 2000: Petrasol BV
<i>Pseudokirchneriella subcapitata</i>									96h	EC50	cell growth	224	4*				US-EPA, 1980
<i>Pseudokirchneriella subcapitata</i>									96h	EC50	cell number	232	4				US-EPA, 1980
<i>Scenedesmus subspicatus</i>									48h	EC50	biomass	110	4	17			IUCLID, 2000: Bayer AG
<i>Scenedesmus subspicatus</i>									48h	EC50	growth rate	220	4	17			IUCLID, 2000: Bayer AG
Crustacea																	
<i>Ceriodaphnia dubia</i>	< 12 h	N	S		nw	8.2± 0.2	25±2	90-110	48h	LC50	mortality	47	3	13			Cowgill and Milazzo, 1991
<i>Ceriodaphnia cf. dubia</i>	< 24 h	Y	S	>97		7.7	23±1	65.2	48h	EC50	immobility	5.29	2				Rose et al., 1998
<i>Ceriodaphnia dubia</i>	< 24 h	Y	Sc	>99.9	nw	7.6	25	45.5	24h	LC50	mortality	7.6	2	18			Marchini et al., 1993
<i>Ceriodaphnia dubia/affinis</i>	neonates	N	Sc		nw	8.2-9.0	20	90	48h	LC50	mortality	8.9	3	19			Hesse et al., 1991: Cowgill et al., 1985
<i>Ceriodaphnia dubia/affinis</i>	neonates	N	Sc		nw	8.2-8.8	24	90	48h	LC50	mortality	11.1	3	19			Hesse et al., 1991: Cowgill et al., 1985
<i>Daphnia carinata</i>	< 24 h	Y	S			20			48h	EC50	immobility	3.99	2				Warne et al., 1999: Khalil, 1998
<i>Daphnia magna</i>			Sc		nw	7			24h	EC50	immobility	16	3	20			Bazin et al., 1987
<i>Daphnia magna</i>	<72 h		Sc		am	7.8-8.2	20	200	24h	EC50	immobility	12	3	21			Devillers et al., 1987
<i>Daphnia magna</i>	<24 h	N	S		am	8.0± 0.2	20	250.2	24h	EC50	immobility	195	3	13			Bringmann and Kühn, 1982
<i>Daphnia magna</i>	<24 h	N	S		tw	7.6-7.7	20-22	286	24h	EC50	immobility	310	3	13			Bringmann and Kühn, 1977b
<i>Daphnia magna</i>		Y	Sc		am				24h	EC50	immobility	4.3	2	20, 22			Calamari et al., 1983
<i>Daphnia magna</i>	< 24 h	N	S		am	7.5± 0.3	21±2	115±8	24h	LC50	immobility	6.2	3	23			Wei et al., 1999
<i>Daphnia magna</i>	1.5 mm	N	Sc		aw	6-7			48h	EC50	immobility	0.58	3	23			Bobra et al., 1985
<i>Daphnia magna</i>			Sc			7			48h	LC50	immobility	0.59	4				IUCLID, 2000: Hoechst AG and Clariant GmbH
<i>Daphnia magna</i>	< 48 h	Y	S		am (DSW)		22±1	100	48h	LC50	immobility	26	2	24			Hermens et al., 1984
<i>Daphnia magna</i>			Sc						24h	LC50	mortality	16	4				IUCLID, 2000: Bayer AG
<i>Daphnia magna</i>			S						24h	LC50	mortality	34	3	13			IUCLID, 2000: Bayer AG
<i>Daphnia magna</i>			Sc						24h	LC50	mortality	140	4				IUCLID, 2000: Bayer AG
<i>Daphnia magna</i>			S						24h	LC50	mortality	310	4				IUCLID, 2000: Bayer AG
<i>Daphnia magna</i>		N	S						48h	EC50		86	3	13			US-EPA, 1980
<i>Daphnia magna</i>	neonates	N	Sc		aw	8	20	160	48h	LC50	mortality	12.9	3				Hesse et al., 1991: Cowgill et al., 1985
<i>Daphnia magna</i>			S		nw	8	20	170	48h	LC50	mortality	12.9	4*				Gersich et al., 1986
<i>Daphnia magna</i>	neonates	N	Sc		aw	8	24	160	48h	LC50	mortality	17.3	3				Hutchinson et al., 1980
<i>Daphnia magna</i>	< 12 h	N	S		nw	8.2± 0.2	25±2	160-180	48h	LC50	mortality	31	3	13			Cowgill and Milazzo, 1991
<i>Daphnia magna</i>			S			8.2	25		48h	LC50	mortality	31	4*				IUCLID, 2000: Bayer AG
<i>Daphnia magna</i>		N	S		nw	7.7-8.3	19.8-	157±4	48h	LC50	mortality	13.9	3	13			Gersich et al., 1986
<i>Daphnia magna</i>			S			20.9			48h	LC50	mortality	47	4				IUCLID, 2000: Bayer AG
<i>Daphnia magna</i>						8.2	25		48h	LC50	mortality	19.9	4	25			IUCLID, 2000: Monsanto and Bayer AG
<i>Daphnia magna</i>									48h	LC50	mortality	0.59	4	26			IUCLID, 2000: Petrasol BV
<i>Daphnia magna</i>									48h	LC50	mortality	5.8	4	27			IUCLID, 2000: Petrasol BV
<i>Daphnia magna</i>	< 24 h	N	Sc	-	rw	6.7-8.1	22±1	72±6	48h	LC50	mortality	86	3	28			LeBlanc, 1980
<i>Daphnia magna</i>									48h	LC50	mortality	5.8	4	29			Zhao et al., 1993
<i>Daphnia magna</i>	4-6 days	N	Sc	>97%	dw		23±2		48h	LC50	mortality	0.58	3				Abernethy et al., 1986

Species	Species properties	A	Test type	Subst. purity [%]	Test water	pH	Temp. [°C]	Hardness CaCO ₃ [mg/L]	Exp. time	Criterion	Test endpoint	Value [mg/L]			Ri	Notes	Reference
												Value [mg/L]	Ri	Notes			
<i>Daphnia pulex</i>	<24 h	N	S	ag	am		20	293.6	96h	LC50	mortality	32.2	3	13			Ikemoto et al., 1992
Pisces																	
<i>Carassius auratus</i>	3.8-6.4 cm, 1-2 g	N	S		nw diluted	7.5	25	soft water	96h	LC50	mortality	51.62	3	13, 29			Pickering and Henderson, 1966
<i>Carassius auratus</i>			S			7	12.8	300	24h	LC50	mortality	5	4	30			BUA.054: Wood, 1953
<i>Cyprinus carpio</i>	5 g, 5 cm	N	R			20±1			48h	LC50	mortality	19	3				Zhao et al., 1993
<i>Danio rerio</i>		Y	Sc		am	7.4	23	310	48h	LC50	mortality	10.5	2	31			Calamari et al., 1983.
<i>Danio rerio</i>	3 +/- 0.5 cm		S				22		96h	LC50	mortality	91	4	32			BUA.054: Rompps Chemie-Lexikon, 1979
<i>Danio rerio</i>																	IUCLID, 2000 Bayer AG
<i>Lepomis macrochirus</i>	juv. 3.65±0.72 cm, 0.90±0.22 g	Y	S	reagent grade	dtw		22±1	30.2	96h	LC50	mortality	91	4*				Bailey et al., 1985
<i>Lepomis macrochirus</i>			S							LC50	mortality	4.5	2	28, 33			IUCLID, 2000: Bayer AG
<i>Lepomis macrochirus</i>	juv. 3.65±0.72 cm, 0.90±0.22 g	Y	F	reagent grade	dtw		22±1	30.2	96h	LC50	mortality	4.5	4*				Bailey et al., 1985
<i>Lepomis macrochirus</i>										LC50	mortality	7.4	2	28, 33			IUCLID, 2000: Bayer AG
<i>Lepomis macrochirus</i>	0.31-1.2 g	Y	F						96h	LC50	mortality	7.4	4*	28			Buccafusco et al., 1981
<i>Lepomis macrochirus</i>		N	S	>80%	rw	6.5-7.9	21-23	31-48	96h	LC50	mortality	16	3	23			US-EPA, 1980
<i>Lepomis macrochirus</i>	3.8-6.4 cm, 1-2 g	N	S		nw diluted	7.5	25	~20	96h	LC50	mortality	15.9	4*	13			Pickering and Henderson, 1966
<i>Lepomis macrochirus</i>												24	3	13, 29			
<i>Lepomis macrochirus</i>	juveniles		S		nw	6.7-7.8	22	28-33	96h	LC50	mortality	19.95	4	28			BUA.054: Janardan et al., 1984
<i>Lepomis macrochirus</i>			S			7	12.8	300	24h	LC50	mortality	6	3	28			Buccafusco et al., 1981
<i>Lepomis macrochirus</i>										LC50	mortality	5	3	30			BUA.054: Wood, 1953
<i>Leuciscus idus</i>			S						96h	LC50	mortality	42.2	4				IUCLID, 2000: Bayer AG
<i>Leuciscus idus melanotus</i>		N	S	-	tw	7-8	20±1	255	48h	LC50	mortality	24	3	13, 34			Juhnke and Lüdemann, 1978
<i>Oncorhynchus mykiss</i>		Y	Sc		am	7.4	15	310	48h	LC50	mortality	4.1	2	31			Calamari et al., 1983
<i>Oncorhynchus mykiss</i>	200-400 g	Y	F		nw		15		96h	LC50	mortality	4.7	2				IUCLID, 2000: Bayer AG
<i>Oncorhynchus mykiss</i>		Y	F						96h	LC50	mortality	4.7	4*				Dalich et al., 1982
<i>Oncorhynchus mykiss</i>	4.6-6.4 cm		F		am	7.6-8.2	15		96h	LC50	mortality	4.7	4*				McCart et al., 1985
<i>Oncorhynchus mykiss</i>			F						96h	LC50	mortality	7.47	2				Hodson et al., 1984
<i>Oryzias latipes</i>									96h	LC50	mortality	7.46	4*				IUCLID, 2000: Petrasol BV
<i>Oryzias latipes</i>	0.2 g	N	S	ag	am		20	293.6	24h	LC50	mortality	17	4	35			IUCLID, 2000: Bayer AG
<i>Perca fluviatilis</i>			S			7	12.8	300	24h	LC50	mortality	55.1	3	13			Ikemoto et al., 1992
<i>Pimephales promelas</i>	30-34 d	N	F		rw		25	43.3-48.5	96h	LC50	mortality	5	3	30			BUA.054: Wood, 1953
<i>Pimephales promelas</i>										LC50	mortality	19	2				Hall et al., 1989
<i>Pimephales promelas</i>	3.8-6.4 cm, 1-2 g	N	S		nw diluted	7.5	25	soft water	96h	LC50	mortality	27	4	8, 36			Nendza and Russom, 1991
<i>Pimephales promelas</i>												29.12	3	13, 29			Pickering and Henderson, 1966
<i>Pimephales promelas</i>	3.8-6.4 cm, 1-2 g	N	S		nw diluted	7.5	25	hard water	96h	LC50	mortality	33.93	3	13, 29			Pickering and Henderson, 1966
<i>Pimephales promelas</i>									96h	LC50	mortality	19	4	8			Zhao et al., 1993
<i>Pimephales promelas</i>	28-36 d	Y	F		nw		25±1		96h	LC50	mortality	16.9	2				Fathead minnow database, US- EPA, 2008
<i>Pimephales promelas</i>	fry	N	Sc	rg	rw	7.6-8.3		96-125	96h	LC50	mortality	22.3	4*				Hesse et al., 1991: Mayes et al., 1983
<i>Pimephales promelas</i>			F						96h	LC50	mortality	22.2	4				IUCLID, 2000a: Petrasol BV
<i>Pimephales promelas</i>	30-34 d		Sc				22		96h	LC50	mortality	35.4	2				Pickering and Henderson, 1966
<i>Pimephales promelas</i>	60-100 d		Sc				22		96h	LC50	mortality	22.2	3				Pickering and Henderson, 1966
<i>Pimephales promelas</i>			F						96h	LC50	mortality	19.12	4				IUCLID, 2000: Bayer AG
<i>Pimephales promelas</i>									48h	LC50	mortality	26	4				IUCLID, 2000: Bayer AG

Species	Species properties	A	Test type	Subst. purity [%]	Test water	pH	Temp. [°C]	Hardness CaCO ₃ [mg/L]	Exp. time	Criterion	Test endpoint	Value	Ri	Notes	Reference
												[mg/L]			
<i>Pimephales promelas</i>	larvae	Y	F	>99.9	nw	7.6	25	45.5	24h	LC50	mortality	7.7	2	37	Marchini et al., 1993
<i>Pimephales promelas</i>	30-34 days old	N	F		nw		25	43.3-48.5	96h	LC50	mortality	19.1	2		Hall et al., 1984
<i>Poecilia reticulata</i>	6 m old, 1.9-2.5 cm, 0.1-0.2 g	N	S		nw diluted	7.5	25	~20	96h	LC50	mortality	44	3	13, 29	Pickering and Henderson, 1966
<i>Poecilia reticulata</i>									7-14d	LC50	mortality	19	4	8	Zhao et al., 1993
<i>Poecilia reticulata</i>	2-3 m/o		S						24h	LC50	mortality	5.63	4		Benoit-Guyod et al., 1984
<i>Poecilia reticulata</i>	2-3 m/o		Rc					25	14d	LC50	mortality	19.1	4		IUCLID, 2000: Bayer AG
<i>Poecilia reticulata</i>		Y	F						96h	LC50		23	4	8	McCarty et al., 1985
<i>Salmo sp.</i>		Y	S						96h	LC50	mortality	10.4	4		IUCLID, 2000: Monsanto and Bayer AG
<i>Salmo trutta</i>			S			7	12.8	300	24h	LC50	mortality	5	3	13, 38	BUA.054: Wood, 1953

Notes

- 1 According to OECD guideline 209
 2 According to ISO 8192
 3 ETAD fermentation tube method
 4 Prolonged incubation compared to ISO 8192
 5 Determined by graphic interpolation
 6 Not stated whether open or closed
 7 Similar to ISO/DIS 9509
 8 Cited in reference
 9 Methods in Schultz 1997
 10 Light intensity 5000 lux, stocks in acetone (>0.1% total conc), recalculated from log EC50 in mM
 11 Light intensity 400 foot candles; 12h light:dark
 12 Light intensity 400 foot candles; 12h light:dark
 13 Open system, no analysis of concentrations
 14 Light intensity 4500 lux; loss of compound was less than 10% of initial value
 15 EPA algal assay
 16 EPA AAPBT test
 17 According to modified DIN 37412 T.9 method
 18 Analysis at beginning and end of test, value estimated by interpolation
 19 Test according to ASTM
 20 AFNOR test
 21 AFNOR test with modifications
 22 AFNOR test; loss of compound was less than 15% of initial value
 23 Test solution constituted with saturated compound
 24 Test according to NEN 6501; results based on measured concentrations.
 25 According to OECD guideline 202
 26 Recalculated from 5.2 µmol/L
 27 Recalculated from 51.6 µmol/L
 28 EPA method
 29 Open system, no analysis of concentrations; TIm is used as LC50
 30 No information exposure concentrations; a concentration of 5 mg/L was classified as 'non-toxic'. Aerated, open systems
 31 IRSA test; loss of compound was less than 10% of initial value
 32 According to UBA, 1984
 33 Based on average measured concentration; EPA method
 34 Open system, no analysis of concentrations; test according to Mann, 1976
 35 According to Japanese Industrial Standard method.
 36 Cited in reference, source EPA, according to ASTM guideline
 37 Concentration measured twice during test, EPA method, value estimated by interpolation

38 Open system, no analysis of concentrations; according to OECD guideline 203; based on nominal concentrations
 39 Species unclear
 40 Exceeds water solubility

Table A2.2. Chronic toxicity of monochlorobenzene to freshwater organisms

Species	Species properties	A	Test type	Subst. purity [%]	Test water	pH	Temp. [°C]	Hardness CaCO ₃ [mg/L]	Exp. time	Criterion	Test endpoint	Value [mg/L]	Ri	Notes	Reference
Bacteria															
<i>Pseudomonas putida</i>		N	Sc		am	7.0	25	81.2	16h	NOEC	growth	17	3	1	Bringmann and Kühn, 1976, 1977a, 1979
Cyanobacteria															
<i>Microcystis aeruginosa</i>		N	Sc		am	7.0	27	55	8d	NOEC	growth	120	3	1, 2	Bringmann and Kühn, 1976, 1978a,b
Protozoa															
<i>Chilomonas paramaecium</i>		N	Sc		am	6.9	20	74.6	48h	NOEC	growth	>195	3	1	Bringmann et al., 1980
<i>Entosiphon sulcatum</i>	Stein	N	Sc		am	6.9	25	75.1	72h	NOEC	growth	>390	3	1	Bringmann and Kühn, 1979, 1980b
<i>Uronema parduczi</i>	Chatton-Lwoff	N	Sc		am	6.9	25	75.1	20h	NOEC	growth	>392	3	1	Bringmann and Kühn, 1980a
Algae															
<i>Scenedesmus quadricauda</i>		N	Sc		am	7.0	27	55	8d	NOEC	growth	>390	3	1, 2	Bringmann and Kühn, 1977a, 1978ab, 1979
<i>Scenedesmus subspicatus</i>									48h	EC10	biomass	38	4	3	IUCLID: Bayer AG
<i>Scenedesmus subspicatus</i>									48h	EC10	growth rate	50	4	3	IUCLID: Bayer AG
<i>Pseudokirchneriella subcapitata</i>		Y	Sc		am				96h	NOEC	growth	6.8	2	4	Calamari et al., 1983
Crustacea															
<i>Ceriodaphnia dubia</i>	< 12 h	N	R		nw	8.2±0.2	25±2	90-110	10d	NOEC	mortality	3.89	3	5	Cowgill and Milazzo, 1991
<i>Ceriodaphnia dubia</i>	< 12 h	N	R		nw	8.2±0.2	25±2	90-110	10d	NOEC	reproduction	12	3	5	Cowgill and Milazzo, 1991
<i>Daphnia magna</i>	< 24 h	Y	S		am		19±1		16d	NOEC	reproduction	0.32	2		Hermens et al., 1984
<i>Daphnia magna</i>	< 24 h	Y	S		am		19±1		16d	NOEC	growth	0.32	4*		De Wolf et al., 1988
<i>Daphnia magna</i>	< 24 h	Y	S		am		19±1		16d	NOEC	reproduction	1	4*		De Wolf et al., 1988
<i>Daphnia magna</i>	< 24 h	Y	S		am		19±1		16d	NOEC	growth	0.32	4*		Hermens et al., 1985a
<i>Daphnia magna</i>			R						16d	NOEC	growth	0.32	4*		IUCLID, 2000: Bayer AG
<i>Daphnia magna</i>	< 12 h	N	R		nw	8.2±0.2	25±2	160-180	10d	NOEC	mortality	<1.4	3	5	Cowgill and Milazzo, 1991
<i>Daphnia magna</i>	< 24 h	Y	S		am		19±1		16d	NOEC	mortality	1	2		Hermens et al., 1984
<i>Daphnia magna</i>	< 12 h	N	R		nw	8.2±0.2	25±2	160-180	10d	NOEC	reproduction	6.5	3	5	Cowgill and Milazzo, 1991
<i>Daphnia magna</i>			Sc						21d	NOEC	reproduction	2.5	4		IUCLID, 2000: Bayer AG
<i>Daphnia magna</i>									9d	NOEC	reproduction	11	4		IUCLID, 2000: Petrasol
<i>Daphnia magna</i>									14d	NOEC	reproduction	2.5	4*		IUCLID, 2000: Petrasol
Amphibia															
<i>Ambystoma gracile</i>	embryo-larva		F					100	9.5d	LC10	mortality	0.872	2	6	Black et al., 1982
Pisces															
<i>Danio rerio</i>	<1d old --> fry	Y	R	>99	rw	7.4-8.4		210	28d	NOEC	growth	4.8	2		Hesse et al., 1991: Van Leeuwen et al., 1990
<i>Danio rerio</i>	<1d old --> fry	Y	R	>99	rw	7.4-8.4		210	28d	NOEC	mortality	8.5	2		Hesse et al., 1991: Van Leeuwen et al., 1990
<i>Danio rerio</i>									28d	NOEC	reproduction	8.5	4*		IUCLID, 2000: Petrasol
<i>Danio rerio</i>									28d	NOEC		10.3	4		IUCLID, 2000: Petrasol
<i>Oncorhynchus mykiss</i>	(sub)adult	Y	CF		nw				30d	NOEC	mortality	2.9	2	7	Hesse et al., 1991: Dalich et al., 1982

Notes

- 1 Toxicity threshold is used as a NOEC
 2 Light intensity 2800 lm
 3 According to modified DIN 38412 T.9 method
 4 EPA algal assay
- 5 Renewal every other day
 6 LC50 = 1.15 mg/L; LC50 after 5.5d exposure = 1.65 mg/L
 7 Reported value is the average of the nominal NOEC and the lowest analysed concentration during the test

Table A2.3. Acute toxicity of monochlorobenzene to marine organisms

Species	Species properties	A	Test type	Subst. purity [%]	Test water	pH	Temp. [°C]	Salinity [%]	Exp. time	Criterion	Test endpoint	Value [mg/L]	Ri	Notes	Reference
Bacteria															
<i>Vibrio fischeri</i>									10min	EC50	bioluminescence	20	4	1	Bazin et al., 1987
<i>Vibrio fischeri</i>									10min	EC50	bioluminescence	18	2	1	BUA.054: Ferard et al., 1983
<i>Vibrio fischeri</i>		S			am	5-9			15min	EC50	bioluminescence	20	4		Kaiser and Palabrica, 1991: Bazin et al., 1987
<i>Vibrio fischeri</i>		S			am	5-9			15min	EC50	bioluminescence	14.8	4		Kaiser and Palabrica, 1991: Hermens et al., 1985
<i>Vibrio fischeri</i>									15min	EC50	bioluminescence	14.8	4*	1	Hermens et al., 1985b
<i>Vibrio fischeri</i>									15min	EC50	bioluminescence	11.5	2	1	Ribo and Kaiser, 1983
<i>Vibrio fischeri</i>									15min	EC50	bioluminescence	11.5	4*	2	Kaiser and Ribo, 1988: Ribo and Kaiser, 1983
<i>Vibrio fischeri</i>		S			am	5-8	15		15min	EC50	bioluminescence	15	2		Hermens et al., 1985
<i>Vibrio fischeri</i>		Y			am		15		15min	EC50	bioluminescence	16	2		Zhao et al., 1993
<i>Vibrio fischeri</i>		N	S				20		30m	EC50	bioluminescence	11.3	4		Kaiser and Palabrica, 1991: Ribo and Kaiser, 1983
<i>Vibrio fischeri</i>			S		am	5-9			30min	EC50	bioluminescence	11.3	4*	1	Ribo and Kaiser, 1983
<i>Vibrio fischeri</i>									30min	EC50	bioluminescence	11.26	4*	1	IUCOLID: Bayer AG
<i>Vibrio fischeri</i>			S		am	5-8	15		30min	EC50	bioluminescence	11.3	4*	2	Kaiser and Ribo, 1988: Ribo and Kaiser, 1983
<i>Vibrio fischeri</i>									30min	EC50	bioluminescence	11	4	2	Sixt et al., 1995
<i>Vibrio fischeri</i>									30min	EC50	bioluminescence	11.25	4	2	Warne et al., 1999
<i>Vibrio fischeri</i>		N	Sc			6.5-7.5	15		5min	EC50	bioluminescence	9.4	2		Blum and Speece, 1991
<i>Vibrio fischeri</i>									5min	EC50	bioluminescence	14.6	2	1	BUA.054: Ferard et al., 1983
<i>Vibrio fischeri</i>									5min	EC50	bioluminescence	9.4	2	1	Ribo et al., 1983
<i>Vibrio fischeri</i>									5min	EC50	bioluminescence	9.4	4*	1	IUCOLID: Bayer AG
<i>Vibrio fischeri</i>			S		am	5-8	15		5min	EC50	bioluminescence	9.36	4*	2	Kaiser and Ribo, 1988: Ribo and Kaiser, 1983
<i>Vibrio fischeri</i>									5min	EC50	bioluminescence	20	4	1	IUCOLID: Bayer AG
Algae															
<i>Chlorella marine</i>		N	Sc				20		72h	EC50	growth rate	20.2	3	4	Ma et al., 1997
<i>Nannochloropsis oculata</i>		N	Sc				20		72h	EC50	growth rate	26.2	3	4	Ma et al., 1997
<i>Phaeodactylum tricornutum</i>		N	Sc				20		72h	EC50	growth rate	19.6	3	4	Ma et al., 1997
<i>Platymonas subcordiformis</i>		N	Sc				20		72h	EC50	growth rate	30.2	3	4	Ma et al., 1997
<i>Pyramidomonas sp.</i>		N	Sc				20		72h	EC50	growth rate	37.2	3	4	Ma et al., 1997
<i>Skeletonema costatum</i>									96h	EC50	growth rate	341	4		US-EPA, 1980
<i>Skeletonema costatum</i>			Sc						120h	EC50	cell number	203	3		Cowgill et al., 1989
Crustacea															
<i>Artemia spp</i>															Zhao et al., 1993
<i>Artemia salina</i>			Sc		nw	7.7-8.1	10.5-22	29.2-30.7	24h	LC50	mortality	41	4*	2	Wells et al., 1982
<i>Americamysis bahia</i>			S						48h	LC50	mortality	40.6	3		US-EPA, 1980
Pisces															
<i>Cyprinodon variegatus</i>	8-15 mm, 14-28 d	N	S	>80%	nw	-	25-31	10-31	96h	LC50	mortality	10	3	3	Heitmuller et al., 1981
<i>Cyprinodon variegatus</i>			S					31	96h	LC50	mortality	10	4*	5	IUCOLID, 2000: Bayer AG
<i>Cyprinodon variegatus</i>		N	S						96h	LC50	mortality	10.5	3	3	US-EPA, 1980
<i>Platichthys flesus</i>	56.2 g	Y	CF	97-99	rw	6	5		96h	LC50	mortality	6.6	2	6	Furay and Smith, 1995
<i>Solea solea</i>	45 g	Y	CF	97-99	rw	6	22		96h	LC50	mortality	5.8	2	6	Furay and Smith, 1995

Notes

- 1 Microtox test
- 2 Cited in reference
- 3 Open system, no analysis of concentrations
- 4 Culturing described by Otsuki et al., 1987, 12h light, nominal concentrations
- 5 EPA-method
- 6 OECD, fish collected in Thames, aeration, 12h light, recovery >80%, recalculated from µM

Table A2.4. Chronic toxicity of monochlorobenzene to marine organisms

Species	Species properties	A	Test type	Subst. purity [%]	Test water	pH	Temp. [°C]	Salinity [%]	Exp. time	Criterion	Test endpoint	Value	Ri	Notes	Reference
												[mg/L]			
Algae															
<i>Skeletonema costatum</i>			Sc						5d	NOEC	cell volume, cell number	100	4		US-EPA, 1980
Crustacea															
<i>Portunus pelagicus</i>	First instar	N	S			26			NR	EC10	growth	0.25	3	1	Mortimer and Connell, 1995
<i>Portunus pelagicus</i>	Sixth instar	N	S			26			NR	EC10	growth	0.25	3	1	Mortimer and Connell, 1995

Notes

1 Open system, no analysis of concentrations

Table A2.5. Acute toxicity of 1,2-dichlorobenzene to freshwater organisms

Species	Species properties	A	Test type	Subst. purity [%]	Test water	pH	Temp. [°C]	Hardness CaCO ₃ [mg/L]	Exp. time	Criterion	Test endpoint	Value	Ri	Notes	Reference
												[mg/L]			
Bacteria															
<i>Pseudomonas fluorescens</i>		Y	S	>98	am	7.1	25		20 min	EC50	luminescence	21.6	2	2	Boyd et al., 1998
<i>Pseudomonas fluorescens</i>										EC50	luminescence	21.6	4*	1	Warne et al., 1999
Protozoa															
<i>Tetrahymena pyriformis</i>		Sc	>95		am	7.35	27		40h	IC50	growth inhibition	1471	3	3, 25	Schultz, 1999
<i>Tetrahymena pyriformis</i>		S					30		24h	EC50	cell proliferation	51	4	2	Yoshioka et al., 1985
Algae															
<i>Ankistrodesmus falcatus</i>		N	Sc		am	8	20		4h	EC50	photosynth.	20.8	3	4	Wong et al., 1984
<i>Chlorella vulgaris</i>									6h	EC50	growth	38	4	26	BUA.053: Kramer and Trumper, 1986
<i>Pseudokirchneriella subcapitata</i>		Y	Sc		am		20±1		3h	EC50	photosynth.	10	2	5	Calamari et al., 1983
<i>Pseudokirchneriella subcapitata</i>		Y	Sc	ag	am				96h	EC50	growth	2.2	4*	6	Calamari et al., 1983
<i>Pseudokirchneriella subcapitata</i>									EC50			91.6	4	7	LeBlanc, 1984
<i>Pseudokirchneriella subcapitata</i>	5 mln cells/L	Y	Sc		am				96h	EC50	cell growth	98	4		US-EPA, 1980
<i>Scenedesmus pannonicus</i>		Y	Sc	99.9	rw		20		96h	EC50	growth inhibition	2.2	2	8	Galassi and Vighi, 1981
<i>Selenastrum subspicatus</i>	8681 SAG	N	Sc		am	8.1-9.6	24±1	55	24h	EC50	growth rate	17	2		Canton et al., 1985
<i>Scenedesmus subspicatus</i>									48h	EC50	growth rate	13.5	3	10	Kühn and Pattard, 1990
<i>Scenedesmus subspicatus</i>									48h	EC50	biomass	13.5	4*	9	IUCLID, 2000: Bayer AG
Crustacea															
<i>Daphnia magna</i>	<24 h	N	S		tw	7.6-7.7	20-22	286	24h	EC50	immobility	68	3	11	Bringmann and Kühn, 1977b
<i>Daphnia magna</i>	<24 h, IRCHA	N	S		am	8.0±0.2	20	250.2	24h	EC50	immobility	45	3	11	Bringmann and Kühn, 1982
<i>Daphnia magna</i>		Y	Sc		am				24h	EC50	immobility	0.78	2	12	Calamari et al., 1983
<i>Daphnia magna</i>	< 24 h	N	S		am	7.5±0.3	21±2	115±8	24h	LC50	immobility	3.4	3	11	Wei et al., 1999
<i>Daphnia magna</i>	1.5 mm	N	Sc		aw	6-7			48h	EC50	immobility	2.35	3	13	Bobra et al., 1985
<i>Daphnia magna</i>	< 48 h	Y	Sc		am		22±1	100	48h	LC50	immobility	3.8	2	14	Hermens et al., 1984
<i>Daphnia magna</i>		Y	Sc	99.9	rw				48h	EC50	immobility	0.74	2		Canton et al., 1985
<i>Daphnia magna</i>	<24 h, IRCHA	N	S	-	am	8.0±0.2	25±1	250	24h	EC50	mortality	1.7	3	11	Kühn et al., 1989
<i>Daphnia magna</i>									48h	EC50	mortality	1.7	4*	16	IUCLID, 2000: Bayer AG
<i>Daphnia magna</i>		Y	Sc	99.9	rw				48h	LC50	mortality	2.2	2		Canton et al., 1985
<i>Daphnia magna</i>	< 24 h	N	Sc	-	rw	6.7-8.1	22±1	72±6	48h	LC50	mortality	2.4	3		LeBlanc, 1980
<i>Daphnia magna</i>		Y	R	> 98	rw			20	48h	LC50	mortality	0.13	2		Pawlisz and Peters, 1995
<i>Daphnia magna</i>									48h	LC50	mortality	2.3	4	7	Zhao et al., 1993
<i>Daphnia magna</i>					S				48h	EC50	mortality	2.44	4	18	US-EPA, 1980

Species	Species properties	A	Test type	Subst. purity [%]	Test water	pH	Temp. [°C]	Hardness CaCO ₃ [mg/L]	Exp time	Criterion	Test endpoint	Value	Ri	Notes	Reference		
																[mg/L]	
<i>Daphnia pulex</i>	<24 h	N	S	ag	am		20	293.6	96h	LC50	mortality	8.27	3	11	Ikemoto et al., 1992		
Insecta																	
<i>Aedes sticticus</i>	adults						25		48h	LC50	mortality	>40	4		BUA.053: Nishiuchi, 1981		
<i>Chironomidae</i>	larvae								2h	LC52	mortality	3.9	4		BUA.053: Fellton, 1940		
<i>Chironomidae</i>	larvae								68h	LC100	mortality	7.8	4		BUA.053: Fellton, 1940		
<i>Cloeon</i>	nymphs								48h	LC50	mortality	48	4		BUA.053: Nishiuchi and Asano, 1979		
<i>Cloeon dipeterum</i>	larvae						25		48h	LC50	mortality	26	4		BUA.053: Nishiuchi, 1981		
<i>Orthetrum albistylum speciosum</i>	larvae						25		48h	LC50	mortality	>40	4		BUA.053: Nishiuchi, 1981		
<i>Sympetrum frequens</i>	larvae						25		48h	LC50	mortality	>40	4		BUA.053: Nishiuchi, 1981		
<i>Micronecta sedula</i>	larvae						25		48h	LC50	mortality	>40	4		BUA.053: Nishiuchi, 1981		
<i>Sigara substrata</i>	larvae								48h	LC50	mortality	>40	4		BUA.053: Nishiuchi, 1981		
<i>Tanytarsus dissimilis</i>	3-4 instart, 2-3.5 mm	Y	S			7.6	20	47	24h	LC50	mortality	20.9	2		Call et al., 1983		
<i>Tanytarsus dissimilis</i>	3-4 instar	Y	S			7.6	20	47	48h	LC50	mortality	12	2		Call et al., 1983		
<i>Tanytarsus dissimilis</i>	larvae								96h	LC50	mortality	11.8	4		US-EPA, 1980		
Pisces																	
<i>Cyprinus carpio</i>	5 g, 5 cm	N	R				20±1		48h	LC50	mortality	9.5	3	11	Zhao et al., 1993		
<i>Danio rerio</i>		Y	Sc		am	7.4	23	320	48h	LC50	mortality	6.8	2	19	Calamari et al., 1983		
<i>Danio rerio</i>		F							48h	LC50	mortality	10	2		Slooff, 1979		
<i>Danio rerio</i>		Y	F						96h	LC50	mortality	5.2	2	20	IUCLID, 2000: Bayer AG		
<i>Lepomis macrochirus</i>	0.32-1.2 g	N	S	>80%	rw	6.5-7.9	21-23	32-48	96h	LC50	mortality	5.6	3	11	Buccafusco et al., 1981		
<i>Lepomis macrochirus</i>									96h	LC50	mortality	5.6	4*		US-EPA, 1980		
<i>Lepomis macrochirus</i>	33-75 mm	N	S	-	nw	7.6-7.9	23	55	96h	LC50	mortality	27	3	11	Dawson et al., 1975/77		
<i>Lepomis macrochirus</i>	33-75 mm	N	S	-	nw	7.6-7.9	23	55	96h	LC50	mortality	29	3	11, 21	Dawson et al., 1975/77		
<i>Lepomis macrochirus</i>		S						22	96h	LC50	mortality	5.6	4*		IUCLID, 2000: Bayer AG		
<i>Leuciscus idus</i>									48h	LC50	mortality	29	4*		IUCLID, 2000: Bayer AG		
<i>Leuciscus idus melanotus</i>		N	S	-	tw	7-8	20±1	255	48h	LC50	mortality	29	3	11, 22	Juhnke and Lüdemann, 1978		
<i>Leuciscus idus melanotus</i>		N	S	-	tw	7-8	20±1	255	48h	LC50	mortality	20	3	11, 22	Juhnke and Lüdemann, 1978		
<i>Oncorhynchus mykiss</i>	4.3 g	Y	F			6.8-7.5	11.6-12.7	51-56.8	96h	LC50	mortality	1.61	2		Ahmad et al., 1984		
<i>Oncorhynchus mykiss</i>	5.6 cm, 2.69 g	Y	F			7.5	12	57	96h	LC50	mortality	1.58	2		Call et al., 1983		
<i>Oncorhynchus mykiss</i>	5.6 cm, 2.69 g	Y	F			7.5	12	57	6 d	LC50	mortality	1.54	2		Call et al., 1983		
<i>Oncorhynchus mykiss</i>		Y	Sc		am	7.4	15	320	48h	LC50	mortality	2.3	2	19	Calamari et al., 1983		
<i>Oncorhynchus mykiss</i>									96h	LC50	mortality	1.1	4		BUA.053: Ribo and Kaiser, 1983		
<i>Oncorhynchus mykiss</i>			F						96h	LC50	mortality	1.67	4		US-EPA, 1980		
<i>Oncorhynchus mykiss</i>			F						96h	LC50	mortality	1.58	4*		US-EPA, 1980		
<i>Oncorhynchus mykiss</i>			F						96h	LC50	mortality	1.58	4*		IUCLID, 2000: Bayer AG		
<i>Oncorhynchus mykiss</i>			Sc				15		48h	LC50	mortality	2.3	4		IUCLID, 2000: Bayer AG		
<i>Oncorhynchus mykiss</i>		Y	F						96h	LC50	mortality	1.6	4	7	McCartry et al., 1985		
<i>Oryzias latipes</i>	3 cm; 0.3g	Y	R						48h	LC50	mortality	9.9	2		Yoshioka et al., 1986		
<i>Oryzias latipes</i>									48h	LC50	mortality	10	4*	23	IUCLID, 2000: Bayer AG		
<i>Oryzias latipes</i>	0.2 g	N	S	ag	am		20	293.6	24h	LC50	mortality	34.5	3	11	Ikemoto et al., 1992		
<i>Pimephales promelas</i>			F						96h	LC50	mortality	17	4	7	Blum and Speece, 1991		
<i>Pimephales promelas</i>	28-36 d	Y	F		nw		25±1		96h	LC50	mortality	9.47	2		Fathead minnow database, US-EPA, 2008		
<i>Pimephales promelas</i>	30-35 d	N	F		rw		25	43.3-48.5	96h	LC50	mortality	5.9	4	7	Zhao et al., 1993		
<i>Pimephales promelas</i>		N	S				22		96h	LC50	mortality	5.8	4		Hall et al., 1989		
<i>Pimephales promelas</i>									96h	LC50	mortality	9.5	4	7	Nendza and Russom, 1991		
<i>Pimephales promelas</i>									96h	LC50	mortality	57	3	18	Curtis et al., 1979		
<i>Pimephales promelas</i>			S				22		96h	LC50	mortality	5.8	4		BUA.053: Könenmann, 1979		
<i>Pimephales promelas</i>									96h	LC50	mortality	9.5	4	7	Mekyan and Veith, 1993		
<i>Pimephales promelas</i>									96h	LC50	mortality	57	4*		IUCLID, 2000: Bayer AG		

Species	Species properties	A	Test type	Subst. purity [%]	Test water	pH	Temp. [°C]	Hardness CaCO ₃ [mg/L]	Exp time	Criterion	Test endpoint	Value [mg/L]	Ri	Notes	Reference
<i>Pimephales promelas</i>									96h	LC50	mortality	5.8	4*		IUCLID, 2000: Bayer AG
<i>Pimephales promelas</i>	3 months, 3.6 cm	Y	R	95					96h	LC50	mortality	6	2	20	Sijm et al., 1993
<i>Pimephales promelas</i>	30-35 days old	N	F		nw		25	43.3-48.5	96h	LC50	mortality	5.85	2		Hall et al., 1984
<i>Poecilia reticulata</i>	3 months, 2.6 cm	Y	R	95					96h	LC50	mortality	4.8	2	20	Sijm et al., 1993
<i>Poecilia reticulata</i>	2-3 months old	N	R			am	22	25	14 d	LC50	mortality	5.85	2	24	Könemann, 1981
<i>Poecilia reticulata</i>									7-14d	LC50	mortality	5.9	4*	7	Zhao et al., 1993
Trout (not specified)									48h	LC50	mortality	6	4		BUA.053: Durig, 1976

Notes

- 1 Cited in reference, unit unclear
 2 Not stated whether system is open or closed
 3 Methods in Schultz, 1997
 4 Light intensity 5000 lux, stocks in acetone (>0.1% total conc), recalculated from log EC50 in mM
 5 Light intensity 4500 lx; loss of compound was less than 10% of initial value
 6 EPA algal assay
 7 Cited in reference
 8 EPA AAPBT test
 9 Probably *Selenastrum subspicatus*; DIN 38412
 10 fluorescent light with 17.0 W/m²; initial cell density: 1e4 cells/L
 11 Open system, no analysis of concentrations
 12 AFNOR test; loss of compound was less than 15% of initial value; closed system
 13 Test solution constituted with saturated compound
 14 Test according to NEN 6501.
 15 Photoperiod 9:15 light:dark with fluorescent light
 16 According to DIN 38412
 17 n = 23 with 7 different populations
 18 Open system, results based on nominal concentrations
 19 IRSA test; loss of compound was less than 10% of initial value
 20 According to OECD guideline 203
 21 LC50 recalculated using nonlinear regression
 22 Test according to Mann, 2076; aerated systems
 23 According to Japanese Industrial Standard method
 24 Covered with glass, acute tests with 96 hours exposure time are preferred
 25 Exceeds water solubility
 26 Short exposure duration

Table A2.6. Chronic toxicity of 1,2-dichlorobenzene to freshwater organisms

Species	Species properties	A	Test type	Subst. purity [%]	Test water	pH	Temp. [°C]	Hardness CaCO ₃ [mg/L]	Exp. time	Criterion	Test endpoint	Value	Ri	Notes	Reference
												[mg/L]			
Bacteria															
<i>Aeromonos hydrophila</i>										NOEC	growth	150	3	1, 12	BUA.053: Schubert, 1979
<i>Bacillus subtilis</i>										NOEC	growth	150	3	12	BUA.053: Schubert, 1979
<i>Pseudomonas cepacia</i>										NOEC	growth	150	3	1, 12	BUA.053: Schubert, 1979
<i>Pseudomonas fluorescens</i>										NOEC	growth	250	3	2, 12	IUCLID, 2000: Bayer AG
<i>Pseudomonas putida</i>		N	Sc		am	7.0	25	81.2	24h 16h	NOEC	growth	15	3	3	Bringmann and Kühn, 1976
Cyanobacteria															
<i>Microcystis aeruginosa</i>		N	Sc		am	7.0	27	55	8d	NOEC	growth	53	3	2, 3	Bringmann, 1975
Protozoa															
<i>Chilomonas paramaeicum</i>		N	Sc		am	6.9	20	74.6	48h	NOEC	growth	>60	3	3, 5	Bringmann et al., 1980
<i>Entosiphon sulcatum</i>	Stein	N	Sc		am	6.9	25	75.1	72h	NOEC	growth	>64	3	3, 5	Bringmann, 1978, Bringmann and Kühn, 1979, 1980b
<i>Uronema parduzci</i>	Chatton-Lwoff	N	Sc		am	6.9	25	75.1	20h	NOEC	growth	80	3	3, 5	Bringmann and Kühn, 1980a
Algae															
<i>Pseudokirchneriella subcapitata</i>	PRINTZ								96h	NOEC	growth	0.88	4		BUA.053: Calamari et al., 1983
<i>Pseudokirchneriella subcapitata</i>	PRINTZ								96h	NOEC	growth	< 0.9	4		BUA.053: Galassi and Vighi, 1981
<i>Pseudokirchneriella subcapitata</i>		Y	Sc		am		20±1		3h	NOEC	photosynth.	< 0.4	2		Calamari et al., 1983
<i>Pseudokirchneriella subcapitata</i>									96h	NOEC	cell growth	<12.9	4		US-EPA, 1980
<i>Pseudokirchneriella subcapitata</i>	10E4 cells/ml	N	S		am		22		120h	LOEC	growth	80	3	6	Millington et al., 1988
<i>Pseudokirchneriella subcapitata</i>	10E4 cells/ml	N	S		am		22		120h	LOEC	growth	80	3	6	Millington et al., 1988
<i>Pseudokirchneriella subcapitata</i>	10E4 cells/ml	N	S		am		22		120h	LOEC	growth	80	3	6	Millington et al., 1988
<i>Scenedesmus quadricauda</i>		N	Sc		am	7.0	27	55	8d	NOEC	growth	>100	3	3, 4, 12	Bringmann and Kühn, 1977a
<i>Scenedesmus subspicatus</i>			Sc						48h	EC10	biomass	3	4	7	IUCLID, 2000: Bayer AG
<i>Scenedesmus subspicatus</i>	10E4 cells/ml	N	S		am		22		48h	EC10	growth rate	7.8	3	7	Kuhn and Pattard, 1990
<i>Scenedesmus subspicatus</i>	10E4 cells/ml	N	S		am		22		120h	LOEC	growth	50	3	6	Millington et al., 1988
<i>Scenedesmus subspicatus</i>	10E4 cells/ml	N	S		am		22		120h	LOEC	growth	100	3	6	Millington et al., 1988
<i>Scenedesmus subspicatus</i>	10E4 cells/ml	N	S		am		22		120h	LOEC	growth	100	3	6	Millington et al., 1988
Crustacea															
<i>Daphnia magna</i>	<24 h, IRCHA	N	R	-	am	8.0±0.2	25±1	250	21d	NOEC	reproduction, mortality	0.63	3	8	Kühn et al., 1989
<i>Daphnia magna</i>			R						21d	NOEC	reproduction, mortality	0.63	4*	7	IUCLID, 2000: Bayer AG
<i>Daphnia magna</i>		Y	R		tw	6.6-7.6		16	21d	NOEC	reproduction, mortality	0.505	2	9	Hesse et al., 1991: Kühn et al., 1989
Pisces															
<i>Oncorhynchus mykiss</i>			F						119d	NOEC	mortality	>0.047	3	10	Oliver and Niimi, 1983
<i>Oncorhynchus mykiss</i>			F						105d	NOEC	mortality	>0.94	3	10	Oliver and Niimi, 1983
<i>Pimephales promelas</i>	larvae									NOEC	mortality	2	4		US-EPA, 1980

Notes

1 Growth stimulation

2 German standard methods

3 Toxicity threshold is used as a NOEC

4 Light intensity 2800 lm

5 1.5*10⁴ cells/ml

6 Open system, no analysis of concentrations; every 24h cell count, acetone as solvent max. 100 µl/l, BBM medium

7 According to DIN 38412

8 photoperiod 9:15 light:dark with fluorescent light

9 Reported value is the average of the nominal NOEC and the lowest analysed concentration during the test

10 Experiment to determine BCF; one concentration; no deaths occurred; loading 18 g fish/L

11 Data originates from US EPA, 1980

12 Exceeds water solubility

Table A2.7. Acute toxicity of 1,2-dichlorobenzene to marine organisms

Species	Species properties	A	Test type	Subst. purity [%]	Test water	pH	Temp. [°C]	Salinity [%]	Exp. time	Criterion	Test endpoint	Value [mg/L]	Ri	Notes	Reference	
Bacteria																
<i>Vibrio fischeri</i>		N	Sc			6.5-7.5	15		30min	EC50	bioluminescence	6	2	1, 3	Sixt et al., 1995	
<i>Vibrio fischeri</i>		N	S				20		5min	EC50	bioluminescence	2.7	2	3	Blum and Speece, 1991	
<i>Vibrio fischeri</i>									15min	EC50	bioluminescence	6.1	2	3	Zhao et al., 1993	
<i>Vibrio fischeri</i>		S		am	5-8	15			5min	EC50	bioluminescence	10.2	2	1, 3	Kaiser and Ribo, 1988: McFeters et al., 1985	
<i>Vibrio fischeri</i>		S		am	5-8	15			5min	EC50	bioluminescence	2.74	2	1, 3	Kaiser and Ribo, 1988: Ribo and Kaiser, 1983	
<i>Vibrio fischeri</i>		S		am	5-8	15			15min	EC50	bioluminescence	3.14	2	1, 3	Kaiser and Ribo, 1988: Ribo and Kaiser, 1983	
<i>Vibrio fischeri</i>		S		am	5-8	15			30min	EC50	bioluminescence	4.04	4*	1, 3	Kaiser and Ribo, 1988: Ribo and Kaiser, 1983	
<i>Vibrio fischeri</i>									5min	EC50	bioluminescence	2.7	4*	3	IUCLID: Bayer AG	
<i>Vibrio fischeri</i>									30min	EC50	bioluminescence	5.99	2	1, 3	Warne et al., 1999	
<i>Vibrio fischeri</i>		S		am	5<pH<9				30m	EC50	bioluminescence	5.99	4*	3	Kaiser and Palabrica, 1991	
<i>Vibrio fischeri</i>		S		am	5<pH<9				30m	EC50	bioluminescence	4.05	4*	3	Kaiser and Palabrica, 1991: Ribo and Kaiser, 1983	
<i>Vibrio fischeri</i>		S		am	5<pH<9				15m	EC50	bioluminescence	4.24	4	3	Kaiser and Palabrica, 1991: Bazin et al., 1987	
<i>Vibrio fischeri</i>		S		am	5<pH<9				5m	EC50	bioluminescence	10.2	4*	3	Kaiser and Palabrica, 1991: McFeters et al., 1983	
Algae																
<i>Skeletonema costatum</i>										EC50		44.2	4	1	LeBlanc, 1984	
<i>Skeletonema costatum</i>									96h	EC50	growth rate	44.1	4		BUA.053: US-EPA, 1980	
<i>Skeletonema costatum</i>									96h	EC50	growth rate	44.1	4*		WHO, 1991: orig ref US-EPA, 1980	
<i>Pyramidomonas sp.</i>		N	Sc			20			72h	EC50	growth rate	17.9	3	4	Ma et al., 1997	
<i>Platymonas subcordiformis</i>		N	Sc			20			72h	EC50	growth rate	16.9	3	4	Ma et al., 1997	
<i>Nonnochloropsis oculata</i>		N	Sc			20			72h	EC50	growth rate	13.1	3	4	Ma et al., 1997	
<i>Chlorella marine</i>		N	Sc			20			72h	EC50	growth rate	16.2	3	4	Ma et al., 1997	
<i>Phaeodactylum tricornutum</i>		N	Sc			20			72h	EC50	growth rate	12.8	3	4	Ma et al., 1997	
Crustacea																
<i>Artemia spp</i>									24h	LC50	mortality	15	4*	1	Zhao et al., 1993	
<i>Artemia spp</i>	larae	N	Sc	>97%			20±1		24h	LC50	mortality	15	3		Abernethy et al., 1986	
<i>Americamysis bahia</i>										LC50		1.97	4*	1	LeBlanc, 1984	
<i>Americamysis bahia</i>										LC50	mortality	1.97	4		US-EPA, 1980	
<i>Palaeomonetes pugio</i>									96h	LC50	mortality	9.4	4		BUA.053: Curtis et al., 1979	
<i>Palaeomonetes pugio</i>		Y	S	am	8.3-8.7	22		25	96h	LC50	mortality	10	3	8,9	BUA.053: Curtis and Ward, 1981	
<i>Palaeomonetes pugio</i>		S				22			96h	LC50	mortality	10	3*		IUCLID, 2000: Bayer AG	
Echinodermata																
<i>Paracentrotus lividus</i>	embryo	N	S	>99	nw	8.2-8.4			37.6-37.8	48h	NOEC	development	0.147	3	2	Pagano et al., 1988
<i>Paracentrotus lividus</i>	embryo	N	S	>99	nw	8.2-8.4			37.6-37.8	48h	LOEC	genotoxicity	0.147	3	2	Pagano et al., 1988
<i>Paracentrotus lividus</i>	sperm	N	S	>99	nw	8.2-8.4			37.6-37.8		LOEC	fertilisation success	1.47	3	2	Pagano et al., 1988
<i>Paracentrotus lividus</i>	sperm	N	S	>99	nw	8.2-8.4			37.6-37.8		LOEC	development	1.47	3	2	Pagano et al., 1988
Pisces																
<i>Cyprinodon variegatus</i>	8-15 mm, 14-28 d	N	S	>80%	nw	-	25-31	10-31	96h	LC50	mortality	9.7	3	2	Heitmuller et al., 1981	
<i>Cyprinodon variegatus</i>									96h	LC50	mortality	9.7	4*		IUCLID, 2000: Bayer AG	
<i>Menidia beryllina</i>	40-100 mm	N	S	-	rw	-	20	-	96h	LC50	mortality	7.3	3	2	Dawson et al., 1975/77	
<i>Menidia beryllina</i>	40-100 mm	N	S	-	rw	-	20	-	96h	LC50	mortality	9.2	3	2, 5	Dawson et al., 1975/77	
<i>Menidia beryllina</i>	40-100 mm	N	S	-	rw	-	20	-	96h	LC10	mortality	8.5	3	2, 6	Dawson et al., 1975/77	
<i>Platichthys flesus</i>	56.2	Y	CF	97-99	rw		6	5	96h	LC50	mortality	4.6	2	7	Furay and Smith, 1995	
<i>Solea solea</i>	45	Y	CF	97-99	rw		6	22	96h	LC50	mortality	4.2	2	7	Furay and Smith, 1995	

Notes

1 Cited in reference

2 Open system, no analysis of concentrations

3 Microtox test
 4 Culturing described by Otsuki et al., 1987, 12h light, nominal concentrations
 5 LC50 recalculated with nonlinear regression
 6 LC10 calculated from original data
 7 OECD, fish collected in Thames, aeration, 12h light/dark, recovery >80%, recalculated from uM
 8 Not stated whether open or closed
 9 Analysed but results not reported and endpoint based on nominal value

Table A2.8. Chronic toxicity of 1,2-dichlorobenzene to marine organisms

Species	Species properties	A	Test type	Substance purity [%]	Test water	pH	Temp. [°C]	Salinity [%]	Exp. time	Criterion	Test endpoint	Value [mg/L]	Ri	Notes	Reference
Algae															
<i>Monochrysis lutheri</i>							20.5		10d	NOEC	growth	7.6	3	1	Ukeles, 1962
<i>Dunaliella euchlora</i>							20.5		10d	NOEC	growth	7.6	3	1	Ukeles, 1962
<i>Chlorella sp.</i>							20.5		10d	NOEC	growth	7.6	3	1	Ukeles, 1962
<i>Protococcus sp.</i>							20.5		10d	NOEC	growth	7.6	3	1	Ukeles, 1962
<i>Phacodactylum tricornutum</i>									10d	NOEC	growth	7.6	3	1	Ukeles, 1962
<i>Skeletonema costatum</i>									96h	NOEC	cell growth and Chl-a content	<12.8	4		US-EPA, 1980
Mollusca															
<i>Mercenaria mercenaria</i>	eggs	N	R		nw		24		48h	NOEC	eggs developing	5	3		Davis and Hidu, 1969
<i>Mercenaria mercenaria</i>	2 d old larvae	N	R		nw		24		10d	NOEC	survival	>1000	3	2	Davis and Hidu, 1969

Notes

- 1 Formulation containing 130 mg 1,2-dichlorobenzene/L
 2 Exceeds water solubility

Table A2.9. Acute toxicity of 1,3-dichlorobenzene to freshwater organisms

Species	Species properties	A	Test type	Subst. purity [%]	Test water	pH	Temp. [°C]	Hardness CaCO ₃ [mg/L]	Exp. time	Criterion	Test endpoint	Value [mg/L]	Ri	Notes	Reference
Bacteria															
<i>aerobic bacteria, adapted</i>										EC100		≥ 20	4		IUCLID, 2000: Hoechst AG and Clariant GmbH
<i>Pseudomonas fluorescens</i>		Y	S	>98	am	7.1	25		20m	EC50	luminescence	34.6	2	4	Boyd et al., 1998
<i>Pseudomonas fluorescens</i>										EC50	luminescence	34.6	4*	3	Warne et al., 1999
Protozoa															
<i>Tetrahymenis pyriformis</i>									24h	EC50		130	4	4	Yoshioka et al., 1985
<i>Tetrahymenis pyriformis</i>							30		24h	EC50		130	4*		IUCLID, 2000: Hoechst AG and Clariant GmbH
Algae															
<i>Algae (unspecified)</i>				99.4					96h	EC50	growth rate	31	4		IUCLID, 2000: Hoechst AG and Clariant GmbH
<i>Ankistrodesmus falcatulus</i>		N	Sc		am	8	20		4h	EC50	photosynth.	22.8	3	5	Wong et al., 1984
<i>Chlorella vulgaris</i>		N							6h	EC50	growth	35-38	4	6	IUCLID, 2000: Hoechst AG
<i>Pseudokirchneriella subcapitata</i>									96h	EC50		179	3	23	BUA.008: US-EPA, 1978
<i>Pseudokirchneriella subcapitata</i>									96h	EC50		149	3	23	BUA.008: US-EPA, 1978
<i>Pseudokirchneriella subcapitata</i>									96h	EC50	growth rate	149	4*		IUCLID, 2000: Hoechst AG and Clariant GmbH
<i>Scenedesmus pannonicus</i>		Y	Sc	99.4	rw	8.1-9.6	24±1	55	24h	EC50		31	2		Canton et al., 1985
<i>Scenedesmus subspicatus</i>	8681 SAG	N	Sc	-	am	8.0±0.2	25±1	250	48h	EC50	growth rate	30	3	8	Kühn and Pattard, 1990
Crustacea															
<i>Daphnia magna</i>	<24h, IRCHA	N	S	-	am	8.0±0.2	25±1	250	24h	EC50	mortality	7	3	7, 9	Kühn et al, 1989

Species	Species properties	A	Test type	Subst. purity [%]	Test water	pH	Temp. [°C]	Hardness CaCO ₃ [mg/L]	Exp time	Criterion	Test endpoint	Value [mg/L]	Ri	Notes	Reference	
<i>Daphnia magna</i>															IUCLID, 2000: Hoechst AG and Clariant GmbH	
<i>Daphnia magna</i>	< 24 h	Y	Sc	95-99%	nw	7.0-7.5	20±1	44.7 (43.5-47.5)	48h	EC50	immobility	7	4*	18	Richter et al., 1983	
<i>Daphnia magna</i>		N	Sc		nw	7			24h	EC50	immobility	6	3	11	Bazin et al., 1987	
<i>Daphnia magna</i>		N	S		am	7.5±0.3	21±2	115±8	24h	EC50	immobility	6	3	10	IUCLID, 2000: Hoechst AG and Clariant GmbH	
<i>Daphnia magna</i>	< 24 h	N	Sc	99.9	rw				48h	EC50	immobility	5.7	3	7	Wei et al 1999	
<i>Daphnia magna</i>		Y	Sc	99.4					24h	EC50	immobility	1.2	2		Canton et al., 1985	
<i>Daphnia magna</i>	< 24 h	Y	Sc	95-99%	nw	7.1-7.7	20±1	44.7 (43.5-47.5)	48h	EC50	immobility	4.2	2	12	Richter et al., 1983	
<i>Daphnia magna</i>		Y	Sc	99.9	rw				48h	LC50	mortality	4.2	4*	14	IUCLID, 2000: Hoechst AG and Clariant GmbH	
<i>Daphnia magna</i>		Y	Sc	99.4					24h	LC50	mortality	6.8	2		Hesse et al., 1991; Canton et al., 1985	
<i>Daphnia magna</i>	< 24 h	Y	Sc	95-99%	nw	7.1-7.7	20±1	44.7 (43.5-47.5)	48h	LC50	mortality	6.8	4*	13	IUCLID, 2000: Hoechst AG and Clariant GmbH	
<i>Daphnia magna</i>		Y	Sc	-	rw	6.7-8.1	22±1	72±6	48h	LC50	mortality	7.4	2	16	Richter et al., 1983	
<i>Daphnia magna</i>	< 24 h	N	Sc		rw	6.7-8.1	22±1	72±6	48h	LC50	mortality	7.4	4*	14	IUCLID, 2000: Hoechst AG and Clariant GmbH	
<i>Daphnia magna</i>		N	Sc						48h	LC50	mortality	28	3		LeBlanc, 1980	
<i>Daphnia magna</i>	< 24 h	Y	Sc	95-99%	nw	7.0-7.5	20±1	44.7	48h	LC50	mortality	28	4*	15	IUCLID, 2000: Hoechst AG and Clariant GmbH	
<i>Daphnia magna</i>		Y	Sc	95-99%	nw	7.0-7.5	20±1	44.7	48h	LC50	mortality	7.2	3	11	Richter et al., 1983	
<i>Daphnia magna</i>		Y	Sc	95-99%	nw	7.0-7.5	20±1	44.7	48h	LC50	mortality	4.9	4	17	Zhao et al., 1993	
<i>Daphnia magna</i>									24h	EC50	mortality	6.3-12.6	4		BUA.008: Hoechst, 1979	
<i>Daphnia pulex</i>									96h	LC50	mortality	8.08	3	7	Ikemoto et al., 1992	
Insecta																
<i>Chironomus riparius</i>	4th-instar	Y	R	>95	nw			19 ± 0.5				0.0368	2		van der Zandt et al., 1994	
<i>Chironomus riparius</i>	4th instar	R	R						96h	NOEC	behaviour	0.0367	4*		IUCLID, 2000: Hoechst AG and Clariant GmbH	
Pisces																
<i>Cyprinus carpio</i>	5 g, 5 cm	N	R					20±1							Zhao et al., 1993	
<i>Danio rerio</i>		N	S						48h	LC50	mortality	3.15-6.3	4	19	IUCLID, 2000: Hoechst AG and Clariant GmbH	
<i>Lepomis macrochirus</i>	0.32-1.2 g	N	Sc	>80%	rw	6.5-7.9	21-23	32-48	96h	LC50	mortality	5	3	20	Buccafusco et al., 1981; also stated in BUA.008	
<i>Lepomis macrochirus</i>		S							96h	LC50	mortality	5.02	4*		BUA.008: US-EPA, 1978	
<i>Lepomis macrochirus</i>		S							96h	LC50	mortality	5	4*		IUCLID, 2000: Hoechst AG and Clariant GmbH	
<i>Lepomis macrochirus</i>		S							96h	LC50	mortality	5.02	4*		IUCLID, 2000: Hoechst AG and Clariant GmbH	
<i>Leuciscus idus</i>									96h	LC50	mortality	13.9	4		BUA.008: Hoechst, 1980	
<i>Leuciscus idus melanotus</i>		S							96h	LC50	mortality	13.9	4*	13	IUCLID, 2000: Hoechst AG and Clariant GmbH	
<i>Leuciscus idus</i>									48h	LC50	mortality	3.1-6.3	4		BUA.008: Hoechst, 1979	
<i>Leuciscus idus melanotus</i>		S							48h	LC50	mortality	3.1-6.3	4*	18	IUCLID, 2000: Hoechst AG and Clariant GmbH	
<i>Oryzias latipes</i>									48h	LC50	mortality	9	4		BUA.133: MITI, 1992	
<i>Oryzias latipes</i>	0.2 g	Y	R	ag	am			20	293.6	48h	LC50	mortality	9	4*		IUCLID, 2000: Hoechst AG and Clariant GmbH
<i>Paramisgurnus dabryanus</i>		N	S	ag	am				24h	LC50	mortality	31.4	3	7	Ikemoto et al., 1992	
<i>Phoxinus sp.</i>		S							48h	LC50	mortality	5.97	4		IUCLID, 2000: Hoechst AG and Clariant GmbH	
<i>Pimephales promelas</i>		F							96h	LC50	mortality	7.37	4		IUCLID, 2000: Hoechst AG and Clariant GmbH	
<i>Pimephales promelas</i>									96h	LC50	mortality	9.5	4	17	Blum and Speece, 1991	
<i>Pimephales promelas</i>	30-35 d	N	F		rw			25	43.3-48.5	96h	LC50	mortality	7.4	4	17	Zhao et al., 1993
<i>Pimephales promelas</i>									96h	LC50	mortality	14	2		Hall et al., 1989	
<i>Pimephales promelas</i>	28-36 d	Y	F		nw			25±1		96h	LC50	mortality	8	4	17	Nendza and Russom, 1991
<i>Pimephales promelas</i>		Y	F	98					96h	LC50	mortality	8.03	2		Fathead minnow database, US-EPA, 2008	
<i>Pimephales promelas</i>	30 d, 0.12 g	Y	F	98	nw	7.5	25±1	45.5		96h	LC50	mortality	8.03	4*	21	IUCLID, 2000: Hoechst AG
<i>Pimephales promelas</i>	30 d, 0.1-6-0.160 g	Y	F	98	nw	7.3-7.6	25	44-46		96h	LC50	mortality	7.8	2		Veith et al., 1983
<i>Pimephales promelas</i>									96h	LC50	mortality	7.8	4*		Carlson and Kosian, 1987	
<i>Pimephales promelas</i>									96h	LC50	mortality	7.8	4*		IUCLID, 2000: Hoechst AG and Clariant GmbH	
<i>Pimephales promelas</i>		Y	F						96h	LC50	mortality	7.79	4*		WHO 1991: US-EPA, 1980	

Species	Species properties	A	Test type	Subst. purity [%]	Test water	pH	Temp. [°C]	Hardness CaCO ₃ [mg/L]	Exp. time	Criterion	Test endpoint	Value [mg/L]	Ri	Notes	Reference
<i>Pimephales promelas</i>										LC50	mortality	8.1	4	17	Mekenyen and Veith, 1993
<i>Pimephales promelas</i>		Y	S		rw	7.2-7.9	22	40-48	96h	LC50	mortality	12.7	3	4,	BUA.008: Curtis and Ward, 1981
<i>Pimephales promelas</i>			F						96h	LC50	mortality	7.36	4	15	IUCLID, 2000: Hoechst AG and Clariant GmbH
<i>Pimephales promelas</i>			S	98					96h	LC50	mortality	7.7	4	21	IUCLID, 2000: Hoechst AG and Clariant GmbH
<i>Pimephales promelas</i>	juvenile 28-34 d	Y	CF	high	nw	7.6	25	44	96h	LC50	mortality	9.12	2		Broderius and Kahl, 1985
<i>Pimephales promelas</i>	28-34 d		F	99					96h	LC50	mortality	9.12	4*	14	IUCLID, 2000: Hoechst AG and Clariant GmbH
<i>Pimephales promelas</i>	30-35 days old	N	F		nw		25	43.3-48.5	96h	LC50	mortality	7.37	2		Hall et al., 1984
<i>Pimephales promelas</i>		Y	F						96h	LC50	mortality	7.79	4*	17	McCarty et al., 1985
<i>Poecilia reticulata</i>									7-14 d	LC50	mortality	7.4	4	17	Zhao et al., 1993
<i>Poecilia reticulata</i>	2-3 months old	N	R		am		22	25	14 d	LC50	mortality	7.4	2	22	Könemann, 1981
<i>Poecilia reticulata</i>			R					22	14 d	LC50	mortality	7.37	4*		IUCLID, 2000: Hoechst AG and Clariant GmbH

Notes

- 1 Concentration exceeds water solubility
 2 Microtox test
 3 Cited in reference, unit unclear
 4 Not stated whether system is open or closed
 5 Light intensity 5000 lux, stocks in acetone (>0.1% total conc), recalculated from log EC50 in mM
 6 Measured is extinction at 680 nm; method according to Wachstumstest of Boehm et al., 1972
 7 Open system, no analysis of concentrations
 8 Fluorescent light with 17.0 W/m²; initial cell density: 1e4 cells/L
 9 Photoperiod 9:15 light:dark with fluorescent light
 10 According to AFNOR methods
 11 Animals were fed 20 mg/L dw trout chow and yeast during test; based on mean concentrations; 16:8h light::dark at 344 lm; binomial
 12 Based on mean concentrations; 16:8h light:dark at 344 lm; binomial
 13 According to OECD guidelines
 14 According to ASTM standard test
 15 According to US EPA guidelines
 16 Based on mean concentrations; 16:8h light:dark at 344 lm; probit analysis
 17 Cited in reference
 18 According to DIN 38412
 19 According to AFNOR T90 303
 20 Undissolved chemical present at highest concentrations
 21 According to APHA standard methods
 22 Covered with glass
 23 Exceeds water solubility
 24 Concentrations measured, results not reported and endpoint based on nominal value

Table A2.10. Chronic toxicity of 1,3-dichlorobenzene to freshwater organisms

Species	Species properties	A	Test type	Subst. purity [%]	Test water	pH	Temp. [°C]	Hardness CaCO ₃ [mg/L]	Exp time	Criterion	Test endpoint	Value [mg/L]	Ri	Notes	Reference
Bacteria															
Anaerobic bacteria from a domestic STP		N							40h	NOEC		40	4	1	IUCLID, 2000: Hoechst AG and Clariant GmbH
Anaerobic microorganisms							37		193h	NOEC	gas production	50	4	2	IUCLID, 2000: Hoechst AG and Clariant GmbH
Algae															
<i>Pseudokirchneriella subcapitata</i>									96h	NOEC		41.8	4		BUA.008: US-EPA, 1978
<i>Pseudokirchneriella subcapitata</i>		S							96h	NOEC		41.8	4*		IUCLID, 2000: Hoechst AG and Clariant GmbH
<i>Scenedesmus subspicatus</i>		Sc							48h	EC10	growth rate	12.5	3		Kühn and Pattard, 1990
Crustacea															
<i>Daphnia magna</i>	<24 h, IRCHA	N	R	-	am	8.0±0.2	25±1	250	21d	NOEC	reproduction	0.8	4	3	Kühn et al., 1989
<i>Daphnia magna</i>	<24 h, IRCHA	Y	R	-	am	8.0±0.2	25±1	250	21d	NOEC	reproduction	0.5	4	3	Kühn et al., 1989
<i>Daphnia magna</i>	< 24 h	Y	R	95-99%	nw	6.6-7.9	20±1	44.7 (43.5-47.5)	28d	NOEC	reproduction	0.69	2	4	Richter et al., 1983
<i>Daphnia magna</i>		Y	CF	98	nw	6.9-7.4		42	28d	NOEC	reproduction	0.69	4*		Hesse et al., 1991: Call et al., 1989
<i>Daphnia magna</i>		Y							21d	NOEC	reproduction rate	0.69	4	7	IUCLID, 2000: Hoechst AG and Clariant GmbH
<i>Daphnia magna</i>	< 24 h	Y	R	95-99%	nw	6.6-7.9	20±1	44.7 (43.5-47.5)	28d	EC10	growth	1.3	2	4	Richter et al., 1983
<i>Daphnia magna</i>		Y	Rc		tw	6.6-7.6		16	21d	NOEC	reproduction	0.65	2	5	Hesse et al., 1991; Kühn et al., 1989
<i>Daphnia magna</i>	< 24 h		R		tw			19	16d	NOEC	growth	0.3	4	6	Deneer et al., 1988
<i>Daphnia magna</i>			R						21d	NOEC	reproduction rate	0.54	4		IUCLID, 2000: Hoechst AG and Clariant GmbH
Pisces															
<i>Oncorhynchus mykiss</i>		Y	F			7.4	25.0	45	32d	NOEC	mortality	0.555	2		Ahmad et al., 1984
<i>Oncorhynchus mykiss</i>		Y	F			7.4	25.0	45	32d	NOEC	growth	0.555	2		Ahmad et al., 1984
<i>Pimephales promelas</i>	4-12 h embryo	Y	F		nw	7.3-7.6	25	44-46	32d	NOEC	mortality	1	2		Carlson and Kosian, 1987
<i>Pimephales promelas</i>										NOEL		1.51	4		US-EPA, 1980
<i>Pimephales promelas</i>	embryo to early juvenile	Y	F	98					31-33d	NOEC	mortality, development	1	4*		IUCLID, 2000: Hoechst AG and Clariant GmbH
<i>Pimephales promelas</i>	ELS	Y	F									1.51	4*	8	McCarty et al., 1985

Notes

1 ETAD fermentation tube method

2 EC15 = ca. 100 mg/L

3 Photoperiod 9:15 light:dark with fluorescent light

4 Based on mean concentrations; 16:8 h light:dark at 344 lm; Dunnett; control mortality <30%

5 Reported value is the average of the nominal NOEC and the lowest analysed concentration during the test

6 NEN 6502, 6501; EC10=0.95 mg/L

7 ASTM guidelines; LOEC = 1.5 mg/L; same as Richter et al.

8 Cited in reference

Table A2.11. Acute toxicity of 1,3-dichlorobenzene to marine organisms

Species	Species properties	A	Test type	Subst. purity [%]	Test water	pH	Temp. [°C]	Salinity [%]	Exp. time	Criterion	Test endpoint	Value [mg/L]	Ri	Notes	Reference
Bacteria															
<i>Vibrio fischeri</i>									5m	EC50	luminescence	2.9	4		IUCLID, 2000: Hoechst AG and Clariant GmbH
<i>Vibrio fischeri</i>									30min	EC50	bioluminescence	5.1	4	1	Sixt et al., 1995
<i>Vibrio fischeri</i>	N	S			6.5-7.5	15			5min	EC50	bioluminescence	3.1	2		Blum and Speece, 1991
<i>Vibrio fischeri</i>	N	S				20			15min	EC50	bioluminescence	8.5	2		Zhao et al., 1993
<i>Vibrio fischeri</i>	N	Sc			am	15			15min	EC50	bioluminescence	3.29	2		Hermens et al., 1985b
<i>Vibrio fischeri</i>		S			am	5-8	15		5min	EC50	bioluminescence	3.07	4	1	Kaiser and Ribo, 1988: McFeters et al., 1985
<i>Vibrio fischeri</i>		S			am	5-8	15		15min	EC50	bioluminescence	4.14	4	1	Kaiser and Ribo, 1988: McFeters et al., 1985
<i>Vibrio fischeri</i>		S			am	5-8	15		30min	EC50	bioluminescence	5.1	4	1	Kaiser and Ribo, 1988: McFeters et al., 1985
<i>Vibrio fischeri</i>									10min	EC50	bioluminescence	4.2	4	3	Bazin et al., 1987
<i>Vibrio fischeri</i>									30min	EC50	bioluminescence	5.1	4	1	Warne et al., 1999
<i>Vibrio fischeri</i>		S			am	5< pH <9			30min	EC50	bioluminescence	5.1	4*		Kaiser and Palabrica, 1991
<i>Vibrio fischeri</i>		S			am	5< pH <9			30min	EC50	bioluminescence	5.1	4*		Kaiser and Palabrica, 1991: Ribo and Kaiser, 1983
<i>Vibrio fischeri</i>		S			am	5< pH <9			15min	EC50	bioluminescence	3.29	4*		Kaiser and Palabrica, 1991: Hermens et al., 1985
<i>Vibrio fischeri</i>		S			am	5< pH <9			30min	EC50	bioluminescence	71.6	4		Kaiser and Palabrica, 1991: R. Speece, 1987
<i>Vibrio fischeri</i>									30min	EC50	bioluminescence	5.1	4		BUA.008: Ribo and Kaiser, 1983
<i>Vibrio fischeri</i>									5min	EC50	bioluminescence	3.3	4		BUA.008: Kamlet et al., 1986
Algae															
<i>Skeletonema costatum</i>									96h	EC50		52.8	4		BUA.008: US-EPA, 1978
<i>Skeletonema costatum</i>									96h	EC50		49.6	4		BUA.008: US-EPA, 1978
Crustacea															
<i>Artemia spp</i>									24h	LC50	mortality	11	4	1	Zhao et al., 1993
<i>Americamysis bahia</i>									96h	LC50	mortality	2.85	4	4	BUA.008: US-EPA, 1978
<i>Americamysis bahia</i>		S							96h	LC50	mortality	2.85	4*	4	IUCLID, 2000: Hoechst AG and Clariant GmbH
Pisces															
<i>Cyprinodon variegatus</i>	8-15 mm	N	S	>80%	nw	-	25-31	10-31	96h	LC50	mortality	7.8	3	2	Heitmuller et al., 1981
<i>Cyprinodon variegatus</i>									96h	LC50	mortality	7.8	4*		IUCLID, 2000: Hoechst AG and Clariant GmbH

Notes

- 1 Cited in reference
- 2 Open system, no analysis of concentrations
- 3 Microtox test
- 4 NOEC < 1.3mg/L

Table A2.12. Chronic toxicity of 1,3-dichlorobenzene to marine organisms

Species	Species properties	A	Test type	Subst. purity [%]	Test water	pH	Temp. [°C]	Salinity [%]	Exp. time	Criterion	Test endpoint	Value [mg/L]	Ri	Notes	Reference
Algae															
<i>Skeletonema costatum</i>									96h	NOEC		7.3	4	BUA.008: US-EPA, 1978	
<i>Skeletonema costatum</i>		S							96h	NOEC		7.3	4*	IUCLID, 2000: Hoechst AG and Clariant GmbH	

Table A2.13. Acute toxicity of 1,4-dichlorobenzene to freshwater organisms

Species	Species properties	A	Test type	Subst. purity [%]	Test water	pH	Temp. [°C]	Hardness CaCO ₃ [mg/L]	Exp. time	Criterion	Test endpoint	Value [mg/L]	Ri	Notes	Reference	
Bacteria																
Methanogenic sewage sludge	900 mg/L		Sc		7	35			48h	EC50	gas production	86	2	EU-RAR: Blum and Speece, 1991		
<i>Nitrosomonas</i> spec.	450 mg/L		Sc		6.5-8	25			12h	EC50	NH4-uptake	86	2	EU-RAR: Blum and Speece, 1991		
Sewage sludge	200 mg/L		Sc		7	35			12h	EC50	O2-uptake	330	2	EU-RAR: Blum and Speece, 1991		
Algae																
<i>Cyclotella meneghiniana</i>		Y							48h	EC50	growth	34.3	2	1	EU-RAR: Figueroa and Simmons, 1991	
<i>Pseudokirchneriella subcapitata</i>		Y	Sc						96h	EC50	growth	1.6	2	1	EU-RAR: Calamari et al., 1983	
<i>Scenedesmus subspicatus</i>		N	S						48h	EC50	proliferation inhibition	38	2		EU-RAR: Kühn and Pattard, 1990	
<i>Scenedesmus pannonicus</i>		Y	S						72h	EC50	growth	31	2		EU-RAR: Canton et al., 1985	
Crustacea																
<i>Daphnia magna</i>		Y	S						24h	EC50	immobilisation	1.6	2		EU-RAR; Calamari et al., 1982	
<i>Daphnia magna</i>		N	S						24h	EC50	immobilisation	3.2	2		EU-RAR: Kühn et al., 1989	
<i>Daphnia magna</i>		Y	S						48h	EC50	immobilisation	0.7	2		EU-RAR: Canton et al., 1985	
<i>Daphnia magna</i>		Y	S						48h	LC50	mortality	2.2	2		EU-RAR: Canton et al., 1985	
Pisces																
<i>Danio rerio</i>		Y	FT						96h	LC50	mortality	2.1	2		EU-RAR: Röderer, 1990	
<i>Jordanella floridae</i>		Y	S						96h	LC50	mortality	4.5	2		EU-RAR: Smith et al., 1991	
<i>Jordanella floridae</i>		Y	FT						96h	LC50	mortality	2.1	2		EU-RAR: Smith et al., 1991	
<i>Oncorhynchus mykiss</i>		Y	FT						96h	LC50	mortality	1.12	2		EU-RAR: Call et al., 1983	
<i>Pimephales promelas</i>	larvae	N	Sc						96h	LC50	mortality	3.6	2		EU-RAR: Mayes et al., 1983	
<i>Pimephales promelas</i>	juvenile	N	Sc						96h	LC50	mortality	14.2	2		EU-RAR: Mayes et al., 1983	
<i>Pimephales promelas</i>	adult	N	Sc						96h	LC50	mortality	11.7	2		EU-RAR: Mayes et al., 1983	
<i>Pimephales promelas</i>	30 d old	Y	FT						96h	LC50	mortality	4.2	2		EU-RAR: Carlson and Kosian, 1987	

Notes

1 Analysis at test start.

Table A2.14. Chronic toxicity of 1,4-dichlorobenzene to freshwater organisms

Species	Species properties	A	Test type	Subst. purity [%]	Test water	pH	Temp. [°C]	Hardness CaCO ₃ [mg/L]	Exp. time	Criterion	Test endpoint	Value [mg/L]	Ri	Notes	Reference
Algae															
<i>Scenedesmus subspicatus</i>		N	S						48h	EC10	proliferation inhibition	16	2		EU-RAR: Kühn and Pattard, 1990
Crustacea															
<i>Daphnia magna</i>		N	R						21d	NOEC	hatching	0.4		1	EU-RAR: Kühn et al., 1989
<i>Daphnia magna</i>		Y	R						28d	NOEC	fertility	0.22	2		EU-RAR: Calamari et al., 1982
Pisces															
<i>Danio rerio</i>		Y	FT						14d	NOEC	mortality, weight, behaviour	0.44	2		EU-RAR: Röderer, 1990
<i>Jordanella floridae</i>	embryo	Y	FT						16d	NOEC	hatching, survival	0.2	2		EU-RAR: Smith et al., 1991
<i>Oncorhynchus mykiss</i>	embryo	Y	FT						60d	NOEC		> 0.1	2	2	EU-RAR: Calamari et al., 1983
<i>Pimephales promelas</i>	embryo	Y	FT						28d	NOEC	hatching, survival	0.57	2		EU-RAR: Carlson and Kosian, 1987

Notes

1 Test concentration decreased from 0.5 to 0.3 mg/L between renewals. 0.4 mg/L is the mean concentration.

2 No effects at any tested concentration

Table A2.15. Acute toxicity of 1,4-dichlorobenzene to marine organisms

Species	Species properties	A	Test type	Subst. purity [%]	Test water	pH	Temp. [°C]	Salinity [‰]	Exp. time	Criterion	Test endpoint	Value [mg/L]	Ri	Notes	Reference
Pisces															
<i>Cyprinodon variegatus</i>		N	S						96h	LC50	mortality	7.4	2		EU-RAR; Heitmüller et al., 1981

Table A2.16. Chronic toxicity of 1,4-dichlorobenzene to marine organisms

No data on the chronic toxicity of 1,4-dichlorobenzene to marine organisms were available in the EU-RAR.

Table A2.17. Acute toxicity of 1,2,3,4-tetrachlorobenzene to freshwater organisms

Species	Species properties	A	Test type	Subst. purity [%]	Test water	pH	Temp. [°C]	Hardness CaCO ₃ [mg/L]	Exp. time	Criterion	Test endpoint	Value	Ri	Notes	Reference
															[mg/L]
Bacteria															
<i>Pseudomonas fluorescens</i>		Y	S	>98	am	7.1	25		20 min	EC50	luminescence	11.8	4	1	Warne et al., 1999
<i>Pseudomonas fluorescens</i>										EC50	luminescence	12.1	2	2	Boyd et al., 1998
Algae															
<i>Ankistrodesmus falcatus</i>		N	Sc		am	8	20		4h	EC50	growth	4.1	3	3	Wong et al., 1984
Crustacea															
<i>Ceriodaphnia cf. dubia</i>	< 24 h	Y	S	>97%		7.7	23±1	65.2	48h	EC50	immobility	0.13	2		Rose et al., 1998
<i>Daphnia carinata</i>	< 24 h	Y	S				20±1		48h	EC50	immobility	0.51	2		Warne et al., 1999; Khalil, 1998
<i>Daphnia magna</i>	< 48 h	Y	S		am		22±1	100		LC50	immobility	0.54	2	4	Hermens et al., 1984
<i>Daphnia magna</i>	<48 h	Y	S		am		22	100	48h	EC50	reproduction	0.09	2	5	De Wolf et al., 1988
<i>Daphnia pulex</i>	<24 h	N	S	ag	am		20	293.6	96h	LC50	mortality	0.18	3	6	Ikemoto et al., 1992
Pisces															
<i>Jordanella floridae</i>		Y	F						96h	LC50		2.01	2		McCarty et al., 1985
<i>Oryzias latipes</i>	0.2 g	N	S	ag	am		20	293.6	24h	LC50	mortality	1.93	3	6	Ikemoto et al., 1992
<i>Pimephales promelas</i>	30-35 d	N	F		rw		25	43.3-48.5	96h	LC50	mortality	0.8	2		Hall et al., 1989
<i>Pimephales promelas</i>	30 d, 0.12 g	Y	F		nw	7.5	25±1	45.5	96h	LC50	mortality	1.1	2		Veith et al., 1983
<i>Pimephales promelas</i>	30 d	Y	F			7.3-7.6	25	44-46	96h	LC50	mortality	1.1	4*		Carlson and Kosian, 1987
<i>Pimephales promelas</i>		Y	F						96h	LC50	mortality	1.1	4*		Mekenyany and Veith, 1993
<i>Pimephales promelas</i>	30-35 days old	N	F		nw		25	43.3-48.5	96h	LC50	mortality	0.8	4*		US-EPA, 1980
<i>Pimephales promelas</i>		Y	F						96h	LC50	mortality	1.08	4		Hall et al., 1984
<i>Poecilia reticulata</i>	1yr, male, 10-14mm	Y	CF	98	tw, dw		20		96h	LC50	mortality	0.4	2		McCarty et al., 1985
															van Hoogen and Opperhuizen, 1988

Notes

- 1 Cited in reference; unit unclear
- 2 Not stated whether system is open or closed
- 3 Light intensity 5000 lux, stocks in acetone (>0.1% total conc.), recalculated from log EC50 in mM
- 4 Test according to NEN 6501, 6502
- 5 Test conditions according to Hermens et al., 1984, recalculated from log(µl/l)
- 6 Open system, no analysis of concentrations

Table A2.18. Chronic toxicity of 1,2,3,4-tetrachlorobenzene to freshwater organisms

Species	Species properties	A	Test type	Subst. purity [%]	Test water	pH	Temp. [°C]	Hardness CaCO ₃ [mg/L]	Exp time	Criterion	Test endpoint	Value [mg/L]	Ri	Notes	Reference
Crustacea															
<i>Daphnia magna</i>	<24 h	Y	R		am		19±1		16d	NOEC	reproduction	0.01	2		Hermens et al., 1984
<i>Daphnia magna</i>	<24 h	Y	R		am		19±1		16d	NOEC	mortality	0.1	2		Hermens et al., 1984
<i>Daphnia magna</i>	<24 h	Y	R		am		19±1		16d	NOEC	growth	0.055	2		Hermens et al., 1985a
<i>Daphnia magna</i>	<24 h	Y	R		am		19±1		16d	NOEC	reproduction	0.055	2		De Wolf et al., 1988
<i>Daphnia magna</i>	<24 h	Y	R		am		19±1		16d	NOEC	growth	0.055	2		De Wolf et al., 1988
Insecta															
<i>Chironomus riparius</i>	3rd instar	N	S	>99%	aw	8.2±0.2	21±2	210	48h	LC10	mortality	0.62	3	1	Roghair et al., 1994
<i>Chironomus riparius</i>	eggs to 2nd instar	Y	R	99%	am	8.2	18 ± 1	209	28d	NOEC	growth	0.0518	2		Leslie et al., 2004
Pisces															
<i>Danio rerio</i>	<1d old --> fry	Y	R	99	rw	7.4-8.4		210	28d	NOEC	growth	0.1	2		Hesse et al., 1991: Van Leeuwen et al., 1990
<i>Danio rerio</i>	<1d old --> fry	Y	R	99	rw	7.4-8.4		210	28d	NOEC	mortality	0.31	2		Hesse et al., 1991: Van Leeuwen et al., 1990
<i>Oncorhynchus mykiss</i>		Y	F			7.4	25.0	45	32d	NOEC	mortality	0.245	2		Ahmad et al., 1984
<i>Pimephales promelas</i>	4-12 h embryos	Y	F			7.3-7.6	25	44-46	32d	NOEC	mortality	0.25	2		Carlson and Kosian, 1987
<i>Pimephales promelas</i>	early life stages	Y	F									0.318	4	3	US-EPA, 1980
<i>Pimephales promelas</i>	early life stages	Y	F									0.32	4	4	McCarty et al., 1985
<i>Pimephales promelas</i>	early life stages	Y	F									0.71	4	4	McCarty et al., 1985

Notes

- 1 Open system, no analysis of concentrations; recalculated
- 2 Recalculated few data in lower concentration range
- 3 Endpoint undetermined
- 4 Cited in reference

Table A2.19. Acute toxicity of 1,2,3,4-tetrachlorobenzene to marine organisms

Species	Species properties	A	Test type	Subst. purity	Test water	pH	Temp. [°C]	Salinity [%]	Exp. time	Criterion	Test endpoint	Value	Ri	Notes	Reference	[mg/L]
Bacteria																
<i>Vibrio fischeri</i>									30min	EC50	bioluminescence	4	4	1	Sixt et al., 1995	
<i>Vibrio fischeri</i>	N	Sc				6.5-7.5	15		5min	EC50	bioluminescence	2.3	2		Blum and Speece, 1991	
<i>Vibrio fischeri</i>	N	Sc		am			15		15min	EC50	bioluminescence	1.9	2		Hermens et al., 1985b	
<i>Vibrio fischeri</i>									30min	EC50	bioluminescence	4.02	4*	1	Warne et al., 1999	
<i>Vibrio fischeri</i>	S		am	5<pH<9					30min	EC50	bioluminescence	4.02	4		Kaiser and Palabrica, 1991; Ribo and Kaiser, 1983	
<i>Vibrio fischeri</i>	S		am	5<pH<9					15min	EC50	bioluminescence	1.88	4		Hermens et al., 1985b	
Algae																
<i>Pyramidomonas sp.</i>	N	Sc				20			72h	EC50	growth rate	3.55	3	2	Ma et al., 1997	
<i>Platymonas subcordiformis</i>	N	Sc				20			72h	EC50	growth rate	3.4	3	2	Ma et al., 1997	
<i>Nannochloropsis oculata</i>	N	Sc				20			72h	EC50	growth rate	2.92	3	2	Ma et al., 1997	
<i>Chlorella marine</i>	N	Sc				20			72h	EC50	growth rate	2.24	3	2	Ma et al., 1997	
<i>Phaeodactylum tricornutum</i>	N	Sc				20			72h	EC50	growth rate	1.88	3	2	Ma et al., 1997	

Notes

1 Cited in reference

2 Culturing described by Otsuki et al., 1987, 12h light, nominal concentrations

Table A2.20. Chronic toxicity of 1,2,3,4-tetrachlorobenzene to marine organisms

Species	Species properties	A	Test type	Substance purity [%]	Test water	pH	Temp. [°C]	Salinity [%]	Exp. time	Criterion	Test endpoint	Value	Ri	Notes	Reference	[mg/L]
Echinodermata																
<i>Portunus pelagicus</i>	1st instar	N	S				26		NR	EC10	growth	0.035	3	1	Mortimer and Connell, 1995	

Notes

1 Open system, no analysis of concentrations

Table A2.21. Acute toxicity of 1,2,3,5-tetrachlorobenzene to freshwater organisms

Species	Species properties	A	Test type	Subst. purity [%]	Test water	pH	Temp. [°C]	Hardness CaCO ₃ [mg/L]	Exp time	Criterion	Test endpoint	Value	Ri	Notes	Reference
Algae															
<i>Ankistrodesmus falcatus</i>		N	Sc		am	8	20		4h	EC50	photosynth.	3	3	1	Wong et al., 1984
<i>Chlamydomonas angulosa</i>		N	Sc		am	6.5	19		3h	EC50	photosynth.	1.6	3	2	Hutchinson et al., 1980
<i>Chlorella vulgaris</i>		N	Sc		am	6.5	19		3h	EC50	photosynth.	2.5	3	3	Hutchinson et al., 1980
<i>Pseudokirchneriella subcapitata</i>									96h	EC50	cell numbers	17.7	3	7	US-EPA, 1980
Crustacea															
<i>Daphnia magna</i>	4-6 days	N	Sc	>97%	dw		23±2		48h	LC50	mortality	0.86	4*		Abernethy et al., 1986
<i>Daphnia magna</i>	< 24 h	N	Sc	-	rw	7.4-9.4	22±1	173±13	48h	LC50	mortality	9.7	3		LeBlanc, 1980
<i>Daphnia magna</i>	1.5 mm	N	Sc		aw	6-7			48h	EC50	immobility	0.86	3	4	Bobra et al., 1985
<i>Daphnia magna</i>		N	S						48h	EC50		50.2	3	5, 7	US-EPA, 1980
Pisces															
<i>Lepomis macrochirus</i>	0.32-1.2 g	N	S	>80%	rw	6.5-7.9	21-23	32-48	96h	LC50	mortality	6.4	3	4, 5, 7	Buccafusco et al., 1981
<i>Lepomis macrochirus</i>		N	S						96h	LC50	mortality	6.42	4*	5, 7	US-EPA, 1980
<i>Poecilia reticulata</i>	2-3 m/o	N	R		am		22	25	14d	LC50	mortality	3.7	2	6	Könemann, 1981

Notes

- 1 Light intensity 5000 lux, recalculated from log EC50 in mM
- 2 Light intensity 400 foot candles; 12h light:dark; 5*10⁴ cells/mL
- 3 Light intensity 400 foot candles; 12h light:dark; 20*10⁴ cells/mL
- 4 Test solution constituted with saturated compound
- 5 Open system, no analysis of concentrations
- 6 Covered with glass
- 7 Exceeds water solubility

Table A2.22. Chronic toxicity of 1,2,3,5-tetrachlorobenzene to freshwater organisms

No data were available on the chronic toxicity of 1,2,3,5-tetrachlorobenzene to freshwater organisms.

Table A2.23. Acute toxicity of 1,2,3,5-tetrachlorobenzene to marine organisms

Species	Species properties	A	Test type	Subst. purity [%]	Test water	pH	Temp. [°C]	Salinity [%]	Exp. time	Criterion	Test endpoint	Value [mg/L]	Ri	Notes	Reference
Bacteria															
<i>Vibrio fischeri</i>		N	Sc			6.5-7.5	15		30 min	EC50	bioluminescence	2.5	4	1	Sixt et al., 1995
<i>Vibrio fischeri</i>			S		am	5<pH<9			5 min	EC50	bioluminescence	3.3	2		Blum and Speece, 1991
<i>Vibrio fischeri</i>									30 min	EC50	bioluminescence	2.48	4		Kaiser and Palabrica, 1991
<i>Vibrio fischeri</i>									30 min	EC50	bioluminescence	2.48	4*	1	Warne et al., 1999
Algae															
<i>Skeletonema costatum</i>									96 h	EC50	cell growth	0.7	4		US-EPA, 1980
<i>Skeletonema costatum</i>									96 h	EC50	cell growth	0.7	4*		US-EPA, 1980
Crustacea															
<i>Mysidopsis bahia</i>			S						96 h	LC50	mortality	0.34	4		US-EPA, 1980
<i>Mysidopsis bahia</i>		N	S						96 h	LC50	mortality	0.34	3	2	US-EPA, 1980
Pisces															
<i>Cyprinodon variegatus</i>	8-15 mm, 14-28 d	N	S	>80%	nw	-	25-31	10-31	96 h	LC50	mortality	3.7	3	2	Heitmuller et al., 1981
<i>Cyprinodon variegatus</i>		N	S						96 h	LC50	mortality	3.7	4*	2	US-EPA, 1980

Notes

1 Cited in reference

2 Open system, no analysis of concentrations

Table A2.24. Chronic toxicity of 1,2,3,5-tetrachlorobenzene to marine organisms

No data were available on the chronic toxicity of 1,2,3,5-tetrachlorobenzene to marine organisms.

Table A2.25. Acute toxicity of 1,2,4,5-tetrachlorobenzene to freshwater organisms

Species	Species properties	A	Test type	Subst. purity [%]	Test water	pH	Temp. [°C]	Hardness CaCO ₃ [mg/L]	Exp time	Criterion	Test endpoint	Value [mg/L]	Ri	Notes	Reference
Bacteria															
<i>Pseudomonas fluorescens</i>		Y	S	>98	am	7.1	25		20 min	EC50	luminescence	1.27	4	1	Warne et al., 1999
<i>Pseudomonas fluorescens</i>										EC50	luminescence	2.16	2	2	Boyd et al., 1998
Protozoa															
<i>Tetrahymena pyriformis</i>		S		>95	am	7.35	30		24h	LC50	mortality	20	3	3, 14	Yoshioka, 1985
<i>Tetrahymena pyriformis</i>							27		40h	IC50	growth inhibition	100	3	4, 14	Schultz, 1999
Algae															
<i>Ankistrodesmus falcatus</i>		N	Sc		am	8	20		4h	EC50	growth rate	4.9	3	5, 14	Wong et al., 1984
<i>Pseudokirchneriella subcapitata</i>									96h	EC50	growth rate	52.9	3	14	US-EPA, 1980: US-EPA, 1978
<i>Pseudokirchneriella subcapitata</i>									96h	EC50	growth rate	46.8	3	14	US-EPA, 1980
Crustacea															
<i>Daphnia magna</i>	<24 h	N	Sc	-	rw	7.4-9.4	22±1	173±13	48h	LC50	mortality	>530	3	6, 14	LeBlanc, 1980
<i>Daphnia magna</i>	<24 h	N	Sc	-	rw	7.4-9.4	22±1	173±13	48h	NOEC	mortality	320	3	6, 14	LeBlanc, 1980
Pisces															
<i>Cyprinus carpio</i>	5 g, 5 cm	N	R				20±1		48h	LC50	mortality	2.8	3	7, 10	Zhao et al., 1993
<i>Jordanella floridae</i>	2-4 month	N	R		nw	6.95±0.35	25±1	48±2.13	96h	LC50	mortality	2.08	3	8, 9	Smith et al., 1991
<i>Jordanella floridae</i>	2-4 mo	Y	F		nw	6.95±0.35	25±1	48±2.13	96h	LC50	mortality	2.15	3	9	Smith et al., 1991
<i>Lepomis macrochirus</i>	0.32-1.2 g	N	S	>80%	rw	6.5-7.9	21-23	32-48	96h	LC50	mortality	1.6	3	9, 10	Buccafusco et al., 1981
<i>Lepomis macrochirus</i>		N	S						96h	LC50	mortality	1.55	3	10	US-EPA, 1980
<i>Leuciscus idus melanotus</i>		N	S	-	tw	7-8	20±1	255	48h	LC50	mortality	30	3	10, 11, 14	Juhnke and Lüdemann, 1978
<i>Leuciscus idus melanotus</i>		N	S	-	tw	7-8	20±1	255	48h	LC0	mortality	10	3	10, 11, 14	Juhnke and Lüdemann, 1978
<i>Pimephales promelas</i>	30-35 d	N	F		rw		25	43.3-48.5	96h	LC50	mortality	0.3	2		Hall et al., 1989
<i>Pimephales promelas</i>									96h	LC50	mortality	0.3	4	12	Zhao et al., 1993
<i>Poecilia reticulata</i>	2-3 m/o	N	R		am		22	25	14 d	LC50	mortality	1.4	2	13	Könemann, 1981

Notes

- 1 Cited in reference, unit unclear
- 2 Not stated whether system is open or closed
- 3 Concentration exceeds water solubility; estimated value
- 4 Methods in Schultz, 1997
- 5 Light intensity 5000 lux, stocks in acetone (>0.1% total conc), recalculated from log EC50 in mM
- 6 According to US EPA, 1975
- 7 Extrapolated value
- 8 Renewal every 24h
- 9 According to US EPA; precipitate present
- 10 Open system, no analysis of concentrations
- 11 Test according to Mann, 1976
- 12 Cited in reference
- 13 Covered with glass
- 14 Exceeds water solubility

Table A2.26. Chronic toxicity of 1,2,4,5-tetrachlorobenzene to freshwater organisms

Species	Species properties	A	Test type	Subst. purity [%]	Test water	pH	Temp. [°C]	Hardness CaCO ₃ [mg/L]	Exp. time	Criterion	Test endpoint	Value [mg/L]	Ri	Notes	Reference
Pisces															
<i>Jordanella floridae</i>	eggs, < 24 h old	Y	F		nw	6.95±0.35	25±1	48±2.13	4-6d	NOEC	hatchability	>0.238	2		Smith et al., 1991
<i>Jordanella floridae</i>	2 weeks	Y	F		nw	6.95±0.35	25±1	48±2.13	10d	NOEC	mortality	>0.238	2		Smith et al., 1991
<i>Jordanella floridae</i>	1 week	Y	F		nw	6.95±0.35	25±1	48±2.13	28d	NOEC	growth	0.067	2		Smith et al., 1991
<i>Jordanella floridae</i>	early life stages	Y	F						28d		fry growth, survival	0.084	2		McCarty et al., 1985
<i>Oncorhynchus mykiss</i>	egg		R			7.2	10	50	96h	NOEC	mortality	10	3	1	Van Leeuwen et al., 1985
<i>Oncorhynchus mykiss</i>	egg		R			7.2	10	50	96h	NOEC	mortality	10	3	1	Van Leeuwen et al., 1985
<i>Oncorhynchus mykiss</i>	14 d		R			7.2	10	50	96h	NOEC	mortality	10	3	1	Van Leeuwen et al., 1985
<i>Oncorhynchus mykiss</i>	28 d		R			7.2	10	50	96h	NOEC	mortality	10	3	1	Van Leeuwen et al., 1985
<i>Oncorhynchus mykiss</i>	42 d		R			7.2	10	50	96h	NOEC	mortality	10	3	1	Van Leeuwen et al., 1985
<i>Oncorhynchus mykiss</i>	young larva		R			7.2	10	50	96h	NOEC	mortality	1.2	2		Van Leeuwen et al., 1985
<i>Salvelinus fontinalis</i>	early life stages	Y	F						28d		fry growth, survival	< 0.24	2		McCarty et al., 1985

Notes

1 Exceeds water solubility

Table A2.27. Acute toxicity of 1,2,4,5-tetrachlorobenzene to marine organisms

Species	Species properties	A	Test type	Subst. purity [%]	Test water	pH	Temp. [°C]	Salinity [%]	Exp. time	Criterion	Test endpoint	Value [mg/L]	Ri	Notes	Reference
Bacteria															
<i>Vibrio fischeri</i>		N	Sc			6.5-7.5	15		5min	EC50	bioluminescence	10	3	4	Blum and Speece, 1991
<i>Vibrio fischeri</i>									30min	EC50	bioluminescence	4.5	4	1, 4	Sixt et al., 1995
<i>Vibrio fischeri</i>		N	S				20		15min	EC50	bioluminescence	0.67	2		Zhao et al., 1993
<i>Vibrio fischeri</i>							15		30min	EC50	bioluminescence	4.51	3	4	Ribo and Kaiser, 1983
<i>Vibrio fischeri</i>									30min	EC50	bioluminescence	4.51	4*	1, 4	Warne et al., 1999
<i>Vibrio fischeri</i>					S	am	5<pH<9		30min	EC50	bioluminescence	4.51	4*	4	Kaiser and Palabrica, 1991
Algae															
<i>Skeletonema costatum</i>									96h	EC50	growth	52.9	3	4	US-EPA, 1978
<i>Skeletonema costatum</i>									96	EC50	cell growth	7.3	3	4	US-EPA, 1980
Crustacea															
<i>Americamysis bahia</i>		N	S						96h	LC50	mortality	1.48	4		US-EPA, 1980
Pisces															
<i>Cyprinodon variegatus</i>	8-15 mm, 14-28 d	N	S	>80%	nw	-	25-31	10-31	96h	LC50	mortality	0.8	3	2	Heitmuller et al., 1981
<i>Cyprinodon variegatus</i>	<20 d old	Y	CF		nw	7.3-8.3	30	12-28	96h	LC50	mortality	0.33	2		Ward et al., 1981
<i>Cyprinodon variegatus</i>		N	S						96h	LC50	mortality	0.84	3	2	US-EPA, 1980

Notes

1 Cited in reference

2 Open system, no analysis of concentrations; extrapolated value

3 Up to 5% methanol as dissolvent

4 Exceeds water solubility

Table A2.28. Chronic toxicity of 1,2,4,5-tetrachlorobenzene to marine organisms

Species	Species properties	A	Test type	Subst. purity [%]	Test water	pH	Temp. [°C]	Salinity [%]	Exp. time	Criterion	Test endpoint	Value [mg/L]	Ri	Notes	Reference
Pisces															
<i>Cyprinodon variegatus</i>	<1d old --> 28 d post-hatching	Y	CF		nw (filtered)	7.3-8.3	30	12-28	>28d	NOEC	mortality	0.18	2		Ward et al., 1981
<i>Cyprinodon variegatus</i>	<1d old --> 28 d post-hatching	Y	CF		nw (filtered)	7.3-8.3	30	12-28	>28d	NOEC	hatchability	>0.52	3	1	Ward et al., 1981
<i>Cyprinodon variegatus</i>												0.129	4		US-EPA, 1980

Notes

1 Value exceeds LC₅₀

Appendix 3. Detailed soil toxicity data

Table A3.1. Toxicity of monochlorobenzene to soil organisms

Species	Species properties	Soil type	A	Subst. purity [%]	pH	o.m. [%]	Clay [%]	T [°C]	Exp. time	Criterion	Test endpoint	Result test soil [mg/kgd.w.]	Result stand. soil [mg/kgd.w.]	Ri	Notes	Reference
Bacteria																
unspecified		sandy loam	Y		0.7			25	8-10h	IC50	O2 consumption	330	1714	4	2	Arulgnanendran and Nirmakhandan, 1998
polytox, mixed microbial culture of 12 strains		sandy loam	Y		0.7			25	8-10h	IC50	O2 consumption	35576	508229	4	2	Arulgnanendran and Nirmakhandan, 1998
Annelida																
<i>Eisenia andrei</i>	adult	sand	Y >98	4.8	3.7	1.4	23	14d		LC50		240	649	2	1	Van Gestel et al., 1991
<i>Eisenia andrei</i>	adult	OECD artificial	Y >98	5.9	8.1	8.1	23	14d		LC50		446	551	2	1	Van Gestel et al., 1991
<i>Lumbricus rubellus</i>	adult	sandy	Y >98	4.8	3.7	1.4	15	14d		LC50		547	1478	2	1	Van Gestel et al., 1991
<i>Lumbricus rubellus</i>	adult	OECD artificial	Y >98	5.9	8.1	8.1	15	14d		LC50		1107	1367	2	1	Van Gestel et al., 1991

Notes

- 1 Data corrected for recovery
2 Exposure duration too short

A3.2. Toxicity of 1,2-dichlorobenzene to soil organisms

Species	Species properties	Soil type	A	Subst. purity [%]	pH	o.m. [%]	Clay [%]	T [°C]	Exp. time	Criterion	Test endpoint	Result test soil [mg/kgd.w.]	Result stand. soil [mg/kgd.w.]	Ri	Notes	Reference
Bacteria																
unspecified		sandy loam	Y		0.7			25	8-10 h	IC50	O2 consumption	120	1714	4	3	Arulgnanendran and Nirmakhandan, 1998
Pseudomonas sp.		sandy loam	Y		0.7			25	8-10 h	IC50	O2 consumption	282587	4036957	4	3	Arulgnanendran and Nirmakhandan, 1998
mixed soil culture		clay-loam	N		7.4				14 d	LOEC	viable bacterial numbers	10		4	1	Meharg et al., 1998
Pseudomonas sp.		clay-loam	Y >99	7.4	7.5	30			49 d	NOEC	colony development	≥ 3200	≥ 4256	4	2,3	Thompson et al., 1999a
mixed soil culture		clay-loam	Y >99	7.4	7.5	30			7 d	NOEC	viable bacterial numbers	1300	1733	4	2,3	Thompson et al., 1999a
mixed soil culture		clay-loam	Y >99	7.4	7.5	30			14 d	NOEC	viable bacterial numbers	1300	1733	4	2,3	Thompson et al., 1999a
mixed soil culture		clay-loam	Y >99	7.4	7.5	30			49 d	NOEC	viable bacterial numbers	1300	1733	4	2,3	Thompson et al., 1999a
mixed soil culture		clay-loam	Y >99	7.4	7.5	30			22 w	LOEC	viable bacterial numbers	100	133	4	2,3	Thompson et al., 1999b
mixed soil culture		clay-loam	Y >99	7.4	7.5	30			22 w	LOEC	bacterial taxa composition	100	133	4	2,3	Thompson et al., 1999b
Fungi																
mixed soil culture		clay-loam	N		7.4				14 d	NOEC	viable fungal hyphal lengths	10		4	1	Meharg et al. 1998
mixed soil culture		clay-loam	Y >99	7.4	7.5	30			7 d	LOEC	viable fungal hyphal lengths	65	87	4	2,3	Thompson et al., 1999a
mixed soil culture		clay-loam	Y >99	7.4	7.5	30			14 d	LOEC	viable fungal hyphal lengths	65	87	4	2,3	Thompson et al., 1999a
mixed soil culture		clay-loam	Y >99	7.4	7.5	30			49 d	LOEC	viable fungal hyphal lengths	65	87	4	2,3	Thompson et al., 1999a
mixed soil culture		clay-loam	Y >99	7.4	7.5	30			22 w	LOEC	viable fungal hyphal lengths	100	133	4	2,3	Thompson et al., 1999b

Notes

- 1 O.m. content of soil not given, recalculation to standard soil not possible
2 Endpoint expressed as nominal; total recovery of extractable radioactivity in concurrent system 42-78% of nominal at the end of the experiment; actual exposure not known
3 Exposure duration too short

A3.3. Toxicity of 1,3-dichlorobenzene to soil organisms

Species	Species properties	Soil type	A	Subst. purity [%]	pH	o.m.	Clay	T	Exp. time	Criterion	Test endpoint	Result test soil [mg/kgd.w.]	Result stand. soil [mg/kgd.w.]	Ri	Notes	Reference
Bacteria																
unspecified	sandy loam	Y		0.7		25	8-10h	IC50		O2 consumption	140	2000	4	1	Arulgnanendran and Nirmakhandan, 1998	
polytox, mixed microbial culture of 12 strains	sandy loam	Y		0.7		25	8-10h	IC50		O2 consumption	417977	5971100	4	1	Arulgnanendran and Nirmakhandan, 1998	

Notes

1 Exposure duration too short

A3.4. Toxicity of 1,2,3,4-tetrachlorobenzene to soil organisms

Species	Species properties	Soil type	A	Subst. purity [%]	pH	o.m.	Clay	T	Exp. time	Criterion	Test endpoint	Result test soil [mg/kgd.w.]	Result stand. soil [mg/kgd.w.]	Ri	Notes	Reference
<i>Eisenia andrei</i>	adult	sandy	Y	>98	4.8	3.7	1.4	23	14d	LC50		75	203	2	1	van Gestel et al., 1991
<i>Eisenia andrei</i>	adult	OECD artificial	Y	>98	5.9	8.1	8.1	23	14d	LC50		223	275	2	1	van Gestel et al., 1991
<i>Lumbricus rubellus</i>	adult	sandy	Y	>98	4.8	3.7	1.4	15	14d	LC50		112	303	2	1	van Gestel et al., 1991
<i>Lumbricus rubellus</i>	adult	OECD artificial	Y	>98	5.9	8.1	8.1	15	14d	LC50		201	248	2	1	van Gestel et al., 1991

Notes

1 Data corrected for recovery

A3.5. Toxicity of 1,2,3,5-tetrachlorobenzene to soil organisms

No data were available on the toxicity of 1,2,3,5-tetrachlorobenzene to soil organisms.

A3.6. Toxicity of 1,2,4,5-tetrachlorobenzene to soil organisms

No data were available on the toxicity of 1,2,4,5-tetrachlorobenzene to soil organisms.

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