Colletotrichum – names in current use

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Filamentous fungi in the genus *Colletotrichum* are destructive pathogens that cause disease and crop losses in plants worldwide. Taxonomy and nomenclature in the group is confusing, even to scientists working in the field, and inaccurate diagnosis of species is not uncommon. In this review, we provide a overview of the 66 *Colletotrichum* names that are in common use, and the 19 recently used names which are regarded as doubtful. This paper represents the first comprehensive overview of the genus in 17 years, and is the first summary treatment of *Colletotrichum* to incorporate data generated through DNA analysis and phylogenetic systematics. Species are listed alphabetically and annotated with their taxonomic entry, teleomorph, hosts and disease, brief summaries of taxonomic and phylogenetic research, and outstanding issues for the genus that are neccesary to stabilize species names. Sequence data and type culture collection resources are also summarized. The paper serves to provide a new starting point for usage of current names in *Colletotrichum* and indicates future work needed.

Key words: anthracnose, molecular phylogeny, nomenclature, plant disease, plant pathogens, plant pathology, quarantine, taxonomy

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Introduction

Colletotrichum is a taxonomically confused genus in urgent need of revision, especially as its members are important plant pathogens (Cannon et al., 2000; Johnston, 2000). Species estimates range from 11 in von Arx (1957), 22 in Sutton (1980), about 40 in Sutton (1992) and 60 in the Dictionary of the Fungi (Kirk et al., 2008), while there are 688 names in Index Fungorum (www.index fungorum.org; accessed 24 November 2009).

This paper uses Sutton (1980) as a starting point for names in the genus (updated in Sutton, 1992), with additional names from literature records since 1980. We have also accessed the USDA database (Farr and Rossman, 2009). There were 129 taxa of Colletotrichum recorded from North America alone, but on further checking many of these were excluded as they were pre-1980 records. Many of the recent compilations (e.g. Index of Plant Diseases in Hawaii, Alfieri et al., 1984; Database of Plant Disease Names in Japan, Sato et al., 2009, http://www.gene.affrc.go. jp/databases-micro pl diseases en.php) names from older checklists that are rarely used these days and are treated as uncertain. Notes for each accepted species are provided, including (putative) hosts, location and requirements for further research (e.g. need for epitypification, host-specificity pathogenicity testing). Forms and varieties are generally not listed as an understanding of species is prerequisite. The entries are made by authoritative mycologists in the field of Colletotrichum research in order to provide a comprehensive understanding of the needs for the genus.

Colletotrichum species are anamorphic Glomerella species (Sutton, 1992; Armstrong-Cho and Banniza, 2006; Pfenning et al., 2007) and names such as Colletotrichum gloeosporioides and Glomerella cingulata refer in the broad sense to the same biological organism, i.e. they are synonyms. However, the species concepts applied to anamorphs and teleomorphs do not always coincide, making simple one-to-one links difficult to achieve (Cannon et al., 2000). There is an imperative to classify fungi under one name, either the teleomorphic or anamorphic name, giving preference to the

older generic name (Shenoy et al., 2007; Crous, 2009) as has been adopted in *Index Fungorum* (see entry under *Colletotrichum falcatum*). In order to follow this approach we need first to clarify the taxonomy of these important pathogens at the species level. Strains producing teleomorphs seem less often to cause serious disease of plants, but many species are heterothallic and have been recognized as holomorphic following laboratory mating experiments (e.g. Guerber and Correll, 2001; Armstrong-Cho and Banniza, 2006). Future revisions may need to adopt the generic name *Colletotrichum* for biological species.

Results and discussion

The *Colletotrichum* names in current use are listed alphabetically below with notes as follows.

- The species name is given with the authorities and publication details.
- Synonyms are not given as these can be searched in *Index Fungorum*. The main focus of *Index Fungorum* is, however, on name, and should not be relied upon in isolation to provide a complete synonymy. Furthermore, synonymies are generally based on subjective understanding based on morphology and are not the topic of this paper.
- The putative diseases caused and known hosts are listed. This is not a rigorous list as it is impossible to verify at this stage whether collections of each taxon on a host are correctly identified.
- Notes are provided on each species and include taxonomic and phylogenetic research needs.
- Recently used doubtful species are discussed at the end of this section.

Table 1 lists the species annotated below and the location of type and ex-type cultures, and genes presently sequenced.

Colletotrichum acutatum J.H. Simmonds ex J.H. Simmonds, Queensland Journal of Agricultural and Animal Science 25: 178A (1968).

Teleomorph: Glomerella acutata Guerber & J.C. Correll

Table 1. Species of *Colletotrichum* treated as currently used, and location of type specimens and their sequenced genes, if available.

Species	Type strain	ITS	Calmodulin	Actin	GAPDH	Tub2	GS	Mat1	Tub1	CHS-1	HIS3
C. acutatum	IMI 117617	AF411700	X	X	Х	X	X	X	Х	X	X
C. agaves	X	X	X	X	X	X	X	X	X	X	X
C. anthrisci	CBS 125334	GU227845	X	GU227943	GU228237	GU228139	X	X	X	GU228335	GU228041
C. asianum	MFU 090233	FJ 972612	FJ 917506	FJ 907424	FJ972576	FJ 907439	FJ 972595	X	X	X	X
C. axonopodi	IMI 279189	X	X	X	X	X	X	FJ377907	X	X	X
C. boninense	MAFF 305972	AB051400	X	X	GQ221769	X	X	X	X	X	X
C. capsici	CBS 120709	EF683603	X	X	X	EF683602	X	X	X	X	X
C. caudatum	MAFF 305700 ¹	EU554110 ¹	X	X	X	X	X	X	X	X	X
C. cereale	KS 20BIG	DQ126177	x	X	X	X	X	DQ131946	X	X	X
C. chlorophyti	IMI 103806	GU227894	x	GU227992	GU228286	GU228188	X	X	X	GU228384	GU228090
C. circinans	CBS 221.81	GU227855	x	GU227953	GU228247	GU228149	X	X	X	GU228345	GU228051
C. cliviae	CBS 125375	GQ485607	GQ849464	GQ856777	GQ856756	GQ849440	X	X	X	X	X
C. coccodes	X	X	x	X	X	X	X	X	X	X	X
C. crassipes	X	X	x	X	X	X	X	X	X	X	X
C. curcumae	IMI 288937	GU227893	x	GU227991	GU228285	GU228187	X	X	X	GU228383	GU228089
C. dematium	CBS 125.25	GU227819	X	GU227917	GU228211	GU228113	X	X	X	GU228309	GU228015
C. destructivum	X	X	X	X	X	X	X	X	X	X	X
C. dracaenophilum	X	X	X	X	X	X	X	X	X	X	X
C. echinochloae	MAFF 511473	AB439811	X	X	X	X	X	X	X	X	X
C. eleusines	MAFF 511155	EU554131	X	X	X	X	X	X	X	X	X
C. falcatum	X	X	X	X	X	X	X	X	X	X	X
C. fiorinae	EHS 58	EF464594	X	X	X	EF593325	X	X	X	X	X
C. fragariae	CBS 142.31	GU174546	X	X	GU174564	X	X	X	X	X	X
C. fructi	CBS 346.37 =		X				X	X	X		
	CCT 4806	GU227844		GU227942	GU228236	GU228138				GU228334	GU228040
C. fructicola	MFU 090228	FJ972603	FJ917508	FJ907426	FJ972578	FJ907441	FJ972593	X	X	X	X

Table 1 (continued). Species treated as currently used in this paper and location of type specimens of *Colletotrichum* and their sequenced genes if available.

Species	Type strain	ITS	Calmodulin	Actin	GAPDH	Tub2	GS	Mat1	Tub1	CHS-1	HIS3
C. fuscum	X	X	X	X	X	X	X	X	X	X	X
C. gloeosporioides	IMI 356878 = CBS 953.97	EU371022, AY376532, FJ976209	FJ917512	FJ907430	FJ972582	FJ907445	FJ972589	X	X	X	X
C. gossypii	X	X	X	X	X	X	X	X	X	X	X
C. graminicola	M 1.001	DO003110	X	X	X	X	X	FJ377994	X	X	X
C. hanaui	MAFF 305404	EU554101	X	X	X	X	X	FJ377922	X	X	X
C. higginsianum	X	X	X	X	X	X	X	X	X	X	X
C. hippeastri	CBS 125376	GQ485599	GQ849469	GQ856788	GQ856764	GQ849446	X	X	X	X	X
C. horii	ICMP 10492	GQ329690	X	X	GQ329681	X	X	X	X	X	X
C. hymenocallidis	CBS 125378	GQ485600	GQ849463	GQ856775	GQ856757	GQ849438	X	X	X	X	X
C. jacksonii	MAFF 305460	EU554108	X	X	X	X	X	X	X	X	X
C. kahawae	IMI 319418	GU174550	X	X	GU174562	X	X	X	X	X	X
C. lilii	X	X	X	X	X	X	X	X	X	X	X
C. lindemuthianum	X	X	X	X	X	X	X	X	X	X	X
C. lineola	CBS 125337	GU227829	X	GU227927	GU228221	GU228123	X	X	X	GU228319	GU228025
C. linicola	X	X	X	X	X	X	X	X	X	X	X
C. liriopes	CBS 119444	GU227804	X	GU227902	GU228196	GU228098	X	X	X	GU228294	GU228000
C. lupini	BBA 70884	X	X	X	X	X	X	DQ174704	AJ301948	X	X
C. lupini var. setosum	BBA 70352	X	X	X	X	X	X	DQ174702	AJ301923	X	X
C. malvarum	X	X	X	X	X	X	X	X	X	X	X
C. miscanthi	MAFF 510857	EU554121	X	X	X	X	X	EU365028	X	X	X
C. musae	X	X	X	X	X	X	X	X	X	X	X
C. navitas	CBS 125086	GQ919067	X	X	X	X	X	GQ919071	X	X	X
C. nicholsonii	MAFF 511115	EU554126	x	X	X	X	X	FJ377946	X	X	X

Table 1 (continued). Species treated as currently used in this paper and location of type specimens of *Colletotrichum* and their sequenced genes if available.

Species	Type strain	ITS	Calmodulin	Actin	GAPDH	Tub2	GS	Mat1	Tub1	CHS-1	HIS3
C. nymphaeae	X	Х	X	X	X	X	X	X	X	X	X
C. orbiculare	X	X	X	X	X	X	X	X	X	X	X
C. paspali	MAFF 305403	EU554100	X	X	X	X	X	FJ377921	X	X	X
C. phaseolorum	X	X	X	X	X	X	X	X	X	X	X
C. phormii	X	X	X	X	X	X	X	X	X	X	X
C. phyllachoroides	X	X	X	X	X	X	X	X	X	X	X
C. rusci	CBS 119206	GU227818	x	GU227916	GU228210	GU228112	X	X	X	GU228308	GU228014
C. sansevieriae	MAFF 239721	AB212991	x	X	X	X	X	X	X	x	X
C. siamense	MFU 090230	FJ972631	FJ917505	FJ907423	FJ972575	FJ907438	FJ972596	X	X	X	X
C. simmondsii	BRIP 28519	FJ972601	FJ917510	FJ907428	FJ972580	FJ907443	FJ972591	X	X	X	X
C. spaethianum	CBS 167.49 =		X				X	X	X		
-	BBA 4804	GU227807		GU227905	GU228199	GU228101				GU228297	GU228003
C. spinaciae	X	X	X	X	X	X	X	X	X	X	X
C. sublineola	S 3.001	DQ003114	X	X	X	X	X	FJ378029	X	X	X
C. tofieldiae	X	X	X	X	X	X	X	X	X	X	X
C. trichellum	X	X	X	X	X	X	X	X	X	X	X
C. trifolii	X	X	X	X	X	X	X	X	X	X	X
C. truncatum	CBS 151.35	GU227862	X	GU227960	GU228254	GU228156	X	X	X	GU228352	GU228058
C. verruculosum	IMI 45525	GU227806	X	GU227904	GU228198	GU228100	X	X	X	GU228296	GU228002
C. xanthorrhoeae	BRIP 45094	GU048667	X	X	GU174563	X	X	X	X	x	X
C. yunnanense	AS 3.9167	EF369490	x	X	X	X	X	X	X	X	X

The *Colletotrichum caudatum* strain listed is provided as a representative of the species aggregate and is provided only for reference; it is not a type strain. The distinctive caudate condia of this isolate clearly identifies it as an authentic member of the species as currently described through morphology. See listing below for additional information.

Hosts and disease: Widely known as a fruit rot pathogen (both in the field and postharvest), but also able to affect other parts of host plants causing symptoms such as blossom blight, crown rot, defoliation, fruit drop, leaf spot, leaf curl and root necrosis, generally described as anthracnose (Wharton Diéguez-Uribeondo, 2004). Commercial crops with significant diseases reported to be caused by C. acutatum include almond (Förster and Adaskaveg, 1999), apple (Biggs and Miller, 2001), chilli (Than *et al.*, 2008b), grape (Yamamoto et al., 1999; Whitelaw-Weckert et al., 2007), strawberry (Freeman et al., 2001a), olive (Talhinhas et al., 2005), and rubber (Fernando et al., 2000). Colletotrichum acutatum f. sp. pineum Dingley & J.W. Gilmour (1972) causes terminal crook disease of pine seedlings. Colletotrichum acutatum f. sp. hakeae Lubbe, Denman, P.F. Cannon, J.Z. Groenew., Lampr. & Crous was reported to be one of the most devastating fungal pathogens of Proteaceae in South Africa where it is used as a biological control agent of weedy Hakea (Lubbe et al., 2004). Colletotrichum fioriniae (as C. acutatum var. fioriniae Marcelino & Gouli was reported to be parasitic on a scale insect (Marcelino et al., 2008). Putative hosts include (from Walker et al., 1991; Johnston and Jones, 1997; Sreenivasaprasad and Talhinhas, 2005; Sato, 1997; The Phytopathological Society of Japan, 2000, 2009 [http://www.gene. affrc.go.jp/databases-micro_pl_diseases_en. php]) Actinidiaceae (Actinidia), Adoxaceae (Sambucus), Amaryllidaceae (Crinum, Amacrinum), Anacardiaceae (Mangifera, Cotinus), Annonaceae (Annona), Apiaceae (Apium), Asteraceae (Calendula, Carthamus, Chrysanthemum, Cosmos, Xanthium, Zinnia), Balsaminaceae (Impatiens) Berberidaceae (Nandina), Bignoniaceae (Bignonia, Parmentiera), Caricaceae (Carica), Cucurbitaceae (Cucurbita), Cruciferae (Matthiola), Dryopteridaceae (Rumohra), Ebenaceae (Diospyros), Ericaceae (Rhododendron, Vaccinium), Euphorbiaceae (Hevea), Fabaceae (Lathyrus, Lupinus, Vigna), Fagaceae (Castanopsis), Gentianaceae (Eustoma, Gentiana), Juglandaceae (Carya, Juglans), Lardizabalaceae (Akebia) Lauraceae (Cinnamonium, Persea), Leguminosae (Lathyrus), Liliaceae (Tricyrtis, Tulipa), Magnoliaceae (Liriodendron, Magnolia), Moraceae (Ficus,

Morus), Myrsinaceae (Cyclamen), Myrtaceae (Acca), Nelumbonaceae (Nelumbo), Oleaceae (Olea), Pinaceae (Pinus, Tsuga), Plumbaginaceae (Limonium), Polemoniaceae (Phlox), Primulaceae (Primula), Proteaceae (Hakea, Leucadendron, Leucospermum, Protea), Ranunculaceae (Anemone, Delphinium, Ranunculus), Rhamnaceae (Ceanothus), Rhodora-(Vaccinium), Rosaceae (Cydonia, Eriobotrya, Fragaria, Malus, Photonia, Prunus, Pyrus, Rubus, Sanguisorba), Rubiaceae (Coffea), Rutaceae (Casimiroa, Citrus, Flindersia), Salicaceae (Salix), Salvinaceae (Salvinia), Saxifragaceae (Bergenia), Simmondsiaceae (Simmondsia), Solanaceae (Capsicum, Cyphomandra, Lycopersicon, Physalis, Solanum), Theaceae (Camelia, Stewartia), Tiliaceae (Grewia, Corchorus), Umbelliferae (Apium), Urticaceae (Urtica), Verbenaceae (Verbena, Vitex), Vitaceae (Vitis). There is also a record of C. acutatum causing lung and kidney infection in a sea turtle (Manire et al., 2002).

Notes: Colletotrichum acutatum is a heterogeneous species (Lardner et al., 1991; Sreenivasaprasad and Talhinhas, 2005), which is difficult to distinguish morphologically from C. gloeosporioides as both exhibit extensive cultural variability and have overlapping host ranges (Wharton and Diéguez-Uribeondo, 2004). Within C. acutatum various groups have been recognised based on physiology (viz. C. acutatum f.sp. chromogenum A.P. Baxter, Van der Westh. & Eicker (1983) was distinguished on the basis of red pigments produced in agar culture media), morphology (Lardner et al., 1999), vegetative compatibility analysis (Lardner et al., 1999), arbitrarily primed PCR (Freeman and Rodriguez, 1995; Lardner et al., 1999; Freeman et al., 2001b) and DNA sequence analysis (Johnston and Jones, 1997; Freeman et al., 2001b; Sreenivasaprasad and Talhinhas, 2005; Whitelaw-Weckert et al., 2007; Than et al., 2008a).

Simmonds (1968) designated a holotype (IMI 117617) and six paratypes from three with morphological hosts variable molecular characteristics. The ex-paratype culture that Simmonds (coll. no. 16633D) sent (1783),New Zealand **ICMP** and subsequently deposited in ATCC as ATCC 56816 (page 225, Guerber and Correll, 2001)

has an ITS sequence that differs from that of the holotype by one base pair (Vinnere *et al.*, 2002; Farr *et al.*, 2005).

Than et al. (2008a) designated an isolate of C. acutatum from Carica papaya in southeast Queensland, Australia as an epitype to help clarify Simmonds' species concept despite the fact that ITS sequences from the holotype (IMI 117617) and one of the paratypes (IMI 117619) (Vinnere et al., 2002) have been determined, and the ex-holotype culture is available in CBS. It is likely that some of the various morphological and molecular groups that have been recognised within C. acutatum represent discrete species, as have been found in some Australian isolates (Shivas and Tan, 2009).

Colletotrichum agaves Cavara, Fungi Longobardiae Exsiccati No 100 (1892).

Teleomorph: Unknown.

Hosts and disease: Leaf disease of Agave, Furcraea.

Notes: This taxon has been detected on dying Agave plants in greenhouses in Missouri Botanical Garden in St Louis (Farr et al., 2006). It has been considered to be a synonym of C. gloeosporioides (von Arx, 1957) and C. coccodes (Hughes, 1958), the latter presently being followed in Index Fungorum. Molecular data (combined ITS and LSU) show this taxon to be a distinct species which can easily be distinguished from other species on Agavaceae by the numerous black setae that develop throughout the conidiogenous region in acervuli (Farr et al., 2006).

Colletotrichum anthrisci Damm, P.F. Cannon & Crous, Fungal Diversity 39: 56 (2009).

Teleomorph: Unknown.

Host: Anthriscus sylvestris.

Notes: This species was described from dead stems of *Anthriscus* in the Netherlands, where it was found to be associated with stem lesions (Damm *et al.*, 2009). The biology and life cycle of this fungus is still unknown.

Colletotrichum asianum Prihastuti, L. Cai & K.D. Hyde, Fungal Diversity 39: 96 (2009).

Teleomorph: Unknown.

Hosts and disease: Endophyte, epiphyte and pathogen of coffee berries (Coffea spp.),

and anthracnose of chilli (Capsicum) and mango (Mangifera).

Notes: This taxon was introduced for Colletotrichum species from coffee in northern Thailand. Epiphytic and endophytic strains were isolated from apparently healthy berries, and it was also isolated as a pathogen. This species is similar in growth rate and colony colour to Colletotrichum kahawae but differs in conidial shape, straight in C. kahawae versus cylindrical with narrowed centre in C. asianum. They are also genetically distinct (Prihastuti et al., 2009). The isolates named as C. gloeosporioides which were shown to be the causal agents of chilli (Capsicum annuum) anthracnose in Thailand (Than et al., 2008b) also belong to this species (L. Cai, pers. comm.). The species appears to be widespread in Asia and occurs on several hosts and further research is required to establish its distribution, host range, biology and hosts it infects.

Colletotrichum axonopodi J.A. Crouch, B.B. Clarke, J.F. White & B.I. Hillman, Mycologia 101: 727 (2009).

Teleomorph: Unknown.

Hosts and disease: Anthracnose of Axonopus affinis, A. compressus.

Notes: Colletotrichum axonopodi is morphologically indistinguishable from the majority of falcate-spored species described from graminicolous hosts, and is diagnosed through molecular phylogenetics (Crouch et al., 2009a). Historically, Axonopus is not listed as a host to any pathogenic Colletotrichum species in the standard publication sources (i.e., Sprague, 1950; Farr et al., 1989), but the taxon has been documented from the USA, Honduras and Australia as herbarium specimens (labelled as C. graminicola or C. sublineola) since the 1930s. Because the host plant is a cosmopolitan weed, the taxon is of no economic importance as a pathogen of agronomic crops.

Colletotrichum boninense Moriwaki, Toy. Sato & Tsukib., Mycoscience 44: 48 (2003).

Teleomorph: Unknown.

Hosts and disease: Leaf spots and endophytes of Carapa, Catostemma, Cattleya, Clivia, Crinum, Cucumis, Cymbidium, Dendrobium, Dracaena, Eperua, Eucalyptus, Euony-

mus, Goupia, Leucospermum, Manilkara, Mora, Passiflora, Protea and Prunus.

Notes: This taxon can be distinguished based on morphological characters (e.g. wide conidia with hilum-like base, 1/b ratio: (1.8-)2-3(-3.3) and molecular data, as well as three isolates being slightly more susceptible to benomyl than a putative C. gloeosporioides isolate (Sato, 1997). This name was first used for isolates from eight plants on the Pacific Coast of Japan (Moriwaki et al., 2003). Later, this species was reported as an endophyte of several plants in Guyana (Lu et al., 2004) and in Australia and southern Africa (Lubbe et al., 2004), and causing anthracnose on Dracaena sanderiana and Euonymus japonica (Lee et al., 2005a,b; Farr et al., 2006). Colletotrichum boninense appears to be a distinct phytopathogen and endophyte on a number of hosts worldwide. The lifestyle should be carefully investigated. Cultures are available at MAFF and thus further sequences can be obtained if required. Johnston and Jones (1997, as Colletotrichum Group 1) and Johnston et al. (2005) reported several genetically and morphologically distinct groups within C. boninense, some of which are consistently associated with unnamed teleomorphs. Sequence data obtained from various isolates at CBS indicate that several undescribed species are presently included in boninense (Damm et al., unpublished).

Colletotrichum caudatum (Peck ex Sacc.) Peck, Bulletin of the New York State Museum 131: 81 (1909).

Teleomorph: Unknown.

Hosts and disease: Anthracnose and leaf spot of Agropyron, Andropogon, Aristida, Bothriochloa, Cymbopogon, Eragrostis, Eremochloa, Eulaliopsis, Imperata, Roetboellia, Schizachyrium, Setaria, Sorghastrum, Sporobolus and Zoysia.

Notes: This is one of the most distinctive of Colletotrichum species, as the falcate conidia have a cauda (appendage) at the apex. Colletotrichum caudatum was originally described as Ellisiella caudata (based on "Colletotrichum caudatum Peck", nom. nud.), by Saccardo (1880) from the foliage of Sorghastrum nutans. Later this name was recombined in Colletotrichum by Peck (1909).

Von Arx (1957) included the taxon as a synonym of C. graminicola, but subsequent morphological examinations and multilocus sequence analysis upheld the distinctiveness of C. caudatum (Sutton, 1980, 1992; Crouch et al., 2009a,b). Although generally considered a single species with a broad range of warmseason grasses, the taxon appears to be an assemblage of host-specific phylogenetic species (Crouch et al., 2009b; Zeiders, 1987). Further sampling is needed to resolve this issue, epitypification should stabilize nomenclature. Potential epitype strains of C. caudatum are currently being evaluated alongside the 1880 type specimen (Crouch, unpubl. data).

Colletotrichum cereale Manns, Ohio Agricultural Experimental Station Bulletin 203: 207 (1909).

Teleomorph: Unknown.

Hosts and disease: Occurring as endophyte and causing leaf spots and stem rots on Agrostis, Avena, Bromus, Calamagrostis, Dactylis, Elymus, Festuca, Hierochloe, Holcus, Hordeum, Lolium, Poa, Polypogon and Triticum.

Notes: This taxon is widely distributed across temperate regions worldwide and inhabits numerous species of cool-season grasses of the subfamily Pooideae. Colletotrichum cereale can be found as either a plant pathogen or an endophyte, and host specialization is supported through phylogenetic studies (Crouch et al., 2009b). Colletotrichum cereale is a falcate-spored species first described by Selby and Manns (1909), but was synonymised with C. graminicola by Wilson just five years later. Wilson's treatment was upheld and expanded by von Arx (1957), but Sutton (1980, 1992), based on appressorial morphology, excluded from C. graminicola all but the fungus pathogenic to Zea. Wheat anthracnose outbreaks caused by C. cereale were problematic during the 1940s in the USA, but presently do not limit production of this crop. despite the fact that the fungus still inhabits wheat plants (see Crouch and Beirn, 2009 for review). Destructive disease outbreaks caused by this fungus on Poa annua and Agrostis stolonifera in golf course greens since the late 1990s highlighted the need to determine the

identity of this organism, which continued to be identified as *C. graminicola* even after Sutton's narrow circumscription. Multi-locus molecular phylogenetic study demonstrated the unique identity of this taxon and corresponded with the host range and morphology documented in the original *C. cereale* species description (Crouch *et al.*, 2006). Epitype strains have been established for the major populations (Crouch *et al.*, 2006).

Colletotrichum chlorophyti S. Chandra & Tandon [as '*chlorophytumi*'], Current Science 34: 565 (1965).

Teleomorph: Unknown.

Hosts and disease: Leaf spot of Chlorophytum. Also reported from Stylosanthes hamata (Damm et al., 2009).

Notes: Recent molecular analysis of the type strain confirmed *C. chlorophyti* to be a distinct species (Damm *et al.*, 2009).

Colletotrichum circinans (Berk.) Voglino, Annali della Reale Accademia d'Agricoltura di Torino 49:175 (1907).

Teleomorph: Unknown. The report of a teleomorph for this species as Cleistothecopsis circinans F. Stevens & E.Y. True (Stevens and True, 1919) must be highly doubtful, because the fungus forms muriform septate ascospores and may be a synonym of *Pleospora* (Dothideomycetes).

Hosts and disease: Causes onion smudge of Allium species. The taxon has also been reported as pathogen of beet (Beta vulgaris), leaf spot on hairy violet (Viola hirta), on dead stem of wild chervil (Anthriscus sylvestris) (Damm et al., 2009) and from several other putative hosts (Farr et al., 2009) which require confirmation.

Notes: This taxon was considered as a form of C. dematium by von Arx (1957). Sutton (1980) maintained C. circinans as a distinct species specific to Allium spp., characterised by falcate, fusiform conidia that are gradually tapered at each end and $19-21 \times 3.5 \, \mu m$, which was confirmed by Damm et al. (2009) in a multi-gene analysis. In the same study, an epitype of this species was designated. Fagbola and Abang (2004), who suspected C. circinans and C. coccodes to be the same species, distinguished them based on DGGE

analysis of PCR-amplified 18S rDNA fragments.

Colletotrichum cliviae Y.L. Yang, Zuo Y. Liu, K.D. Hyde & L. Cai, Fungi Diversity 39: 133 (2009)

Teleomorph: Unknown.

Hosts and disease: Leaf spots of Clivia miniata.

Notes: This species is similar to Colleto-trichum boninense and C. crassipes in terms of conidial width. Shape is however, different, conidia of C. crassipes are truncate at the base, while those in C. boninense have a low hilumlike protuberance at the base (Sutton, 1980; Moriwaki et al., 2003). Although the conidial width in C. cliviae, C. crassipes, C. orbiculare and C. boninense overlap, C. cliviae grows faster than C. orbiculare and C. boninense. In addition, it is different from any similar taxa in nu-rDNA ITS, CAL, GDPH, actin, chitin synthase A and β-tubulin sequences (Yang et al., 2009).

Colletotrichum coccodes (Wallr.) S. Hughes, Canadian Journal of Botany 36: 754 (1958).

Teleomorph: Unknown.

Hosts and disease: The most important diseases are potato black dot (Lees and Hilton, 2003) and anthracnose of tomato (Ben-Daniel et al., 2009), but it has also been reported from chilli and mint (Nitzan et al., 2006b; Than et al., 2008c) as well as several rotation crops and Curcurbitaceae (Nitzan et al., 2006a; Tsror and Johnson, 2000).

Notes: This species was described from potato in Germany in 1833 as Chaetomium coccodes and is now regarded as a species of Colletotrichum. It has since been shown to cause disease in tomatoes and numerous other hosts and even human infection (Cano et al., 2004). It has several synonyms from a wide range of hosts. The conidia are similar to those of C gloeosporioides but are slightly constricted in the centre and taper abruptly at each end (Sutton, 1992). The taxon has been isolated as a common endophyte of root segments of transgenic potatoes (Götz et al., 2006). Vegetative compatibility groups have been reported for isolates from Israel and northern Europe (Shcolnick et al., 2007) and USA (Heilmann et al., 2006; Nitzan et al., 2002, 2006a,b). Several

recent papers deal with various aspects of the fungus and the diseases it causes (Sanogo *et al.*, 2003; Costa *et al.*, 2006; Aqeel *et al.*, 2008; Minuto *et al.*, 2008; Nitzan *et al.*, 2008; Ben-Daniel *et al.*, 2009). It has not been epitypified, which is necessary in order to establish whether this is a distinct species and stabilize the characters and name for future studies.

Colletotrichum crassipes (Speg.) Arx, Verhandelingen Koninklijke Nederlandse Akademie van Wetenschappen, tweede sect. 51(3): 77 (1957).

Teleomorph: Unknown.

Hosts: Grape anthracnose (Vitis).

Notes: This species was originally described from fruit of Vitis vinifera (grape) from Conegliano, Italy. The species was accepted by Sutton (1980, 1992) and is similar to C. gloeosporioides, but differs in its wider. generally longer conidia and deeply lobed appressoria. The name has rarely been used recently and the records from rattans (Mohanan. 2005) and jacaranda - Dalbergia nigra (Dhingra et al., 2003) need verification. It has also been putatively identified, mainly based on the morphology of the appressoria with crenate or deeply lobed margins and size of conidia, as causing human phaeohyphomycotic cysts (Castro et al., 2001). Colletotrichum acutatum is also known from grape (Melksham et al., 2002). Sutton (1992) suggested this taxon migh be composite and comprise several taxa. Fresh collections of this taxon are needed from grapes in Italy for molecular analysis.

Colletotrichum curcumae (Syd.) E.J. Butler & Bisby, The Fungi of India: 153 (1931).

Teleomorph: Unknown.

Hosts and disease: Leaf spot of turmeric (Curcuma longa) (Palarpawar and Ghurde, 1994).

Notes: In a multi-gene analysis, this taxon was shown to be a distinct species and epitypified (Damm *et al.*, 2009).

Colletotrichum dematium (Pers.) Grove, Journal of Botany, British and Foreign, London 56: 341 (1918).

Teleomorph: Unknown.

Hosts and disease: Originally described on stem necroses on Eryngium campestre, and

also reported as pathogenic on potato stems (Solanum tuberosum), from leaf spots of Genista tinctoria, as an endophyte in grapevine (Vitis vinifera), on dead stems of Apiaceae and on Xanthium. Other hosts need to be confirmed. Notes: Many Colletotrichum species with curved conidia had been synonymised with C. dematium by von Arx (1957), who considered this taxon to be a common saprotroph on many herbaceous plants with several hostspecific forms. Descriptions of C. dematium in the literature are very variable and confusing (von Arx, 1957; Sutton, 1980; Baxter, 1983). Therefore, this taxon has been epitypified and compared with other curved spored species from herbaceous hosts based on morphological and molecular data (Damm et al., 2009). Colletotrichum dematium was confirmed to be plurivorous, probably having pathogenic, saprotrophic and endophytic lifestyles. However, since many strains originally identified as C. dematium proved to belong to different species, for example C. lineola, C. spaethiana C. truncatum, host and biological information of this taxon in previous studies might be wrong. This includes the causal organisms of several economically important diseases, such as leaf blight of Japanese radish seedlings, anthracnose of mulberry, narukoyuri (Polygonatum falcatum) and statice (Yoshida and Shirata, 1999; Sato et al., 2005; Babu et al., 2008; Tomioka et al., 2008; Bobev et al., 2009), while the anthracnose pathogen of Rhododendron in Sweden (Vinnere et al., 2002) does belong to C. dematium or the closely related C. lineola (Damm et al., 2009).

Colletotrichum destructivum O'Gara, Mycologia 7: 38 (1915).

Teleomorph: Glomerella glycines Lehman & F.A. Wolf

Hosts and disease: Anthracnose in Brassicaceae, Cuscutaceae, Leguminosae, Solanum, Perilla.

Notes: This taxon causes anthracnose of soybean usually affecting aging plants at harvest or stubble (Manandhar et al., 1986) and of lucerne as a sole or secondary pathogen (Boland and Brochu, 1989). It was also found to cause anthracnose of perilla (Kawaradani et al., 2008). It is a hemibiotrophic species in Medicago sativa, M. truncatula, Vigna ungui-

species culata, several Nicotiana and Arabidopsis thaliana (Latunde-Dada et al., 1996, 1997; Shen et al., 2001; O'Connell et al., 2004) and produces large, prominently multilobed infection structures that are localized within the initially infected epidermal cells of the infected host. This feature is the same in C. linicola and C. higginsianum, and represents a key taxonomic character (Latunde-Dada and Lucas, 2007; O'Connell et al., 2004). A multilocus comparison approach is required to determine their relationships.

Colletotrichum dracaenophilum D.F. Farr & M.E. Palm, Mycological Research 110: 1401 (2006).

Teleomorph: Unknown.

Hosts and disease: Leaf anthracnose of Dracaena.

Notes: This species was recently described and ex-type cultures and sequences are available. The taxon is known only from China and can be distinguished from other taxa on Agavaceae by its long conidia (averaging more than 28 μm long). Molecular data support the uniqueness of this species (Farr et al., 2006; L. Cai, pers. comm.). Other taxa from Dracaena have been discussed by Farr et al. (2006).

Colletotrichum echinochloae Moriwaki & Tsukib., Mycoscience 50: 275 (2009).

Teleomorph: Unknown.

Hosts and disease: Leaf blight and greyish-white lesions with brown margins on leaves of Echinochloa esculenta.

Notes: This taxon was previously identified as C. graminicola but differs from this species in its falcate and short conidia, 18-22.2 µm in length, cultural characteristics, and specific pathogenicity to Echinochloa esculenta. Molecular phylogenetic analyses using rDNA-ITS, HMG, and Sod2 sequences also show this species to be unique (Moriwaki and Tsukiboshi, 2009). C. echinochloae originates from the same host plant as C. jacksonii, and while the two species are clearly sister taxa, the type strains of C. echinochloa and C. jacksonii fall into two distinct phylogenetic lineages (Crouch et al., 2009a; Moriwaki and Tsukiboshi, 2009).

Colletotrichum eleusines Pavgi & U.P. Singh

[as 'eleusinis'], Mycopathologia et Mycologia Applicata 27: 85 (1965).

Teleomorph: Unknown.

Hosts: Anthracnose of Eleusine indica.

Notes: Wilson (1914) included isolates of Colletotrichum from Eleusine in his description of C. graminicola. Colletotrichum eleusines was first introduced in 1965, but the taxon was not accepted by Sutton (1980, 1992) in any of his subsequent treatments. Crouch et al. (2009a) reintroduced C. eleusines based on molecular phylogenetic data. The taxon is morphologically indistinguishable from many closely related falcate-spored Colletotrichum species associated with grasses and is diagnosed using molecular analysis.

Colletotrichum falcatum Went, Archives Java Suikerindustrie 1: 265 (1893).

Teleomorph: Glomerella tucumanensis (Speg.) Arx & E. Müll.

Hosts and disease: Red rot and red streak of *Saccharum* (sugarcane).

Notes: Red rot can infect mature stalks of sugarcane, leaf mid ribs and cause rot of planting material which results in substantial losses in crop yield and sugar quality (Rao et al., 2004). This fungus was isolated in 1892 from a sugarcane field at Tjomal, Java, Indonesia but the type material could not be located in any herbaria. Remarkably the same sugarcane field still exists and it is possible to designate an epitype in order to stabilize the taxonomic position. The taxon was recently isolated from the same original site and host species allowing it to be epitypified (Prihastuti et al., unpublished).

Colletotrichum fioriniae (Marcelino & Gouli) R.G. Shivas & Y.P. Tan, Fungal Diversity 39: 117 (2009).

Teleomorph: Glomerella acutata var. fioriniae J.A.P. Marcelino & S. Gouli Hosts and disease: Leaf and stem blight of Acacia acuminata, fruit rot of Persea americana, endophyte in Mangifera indica and 28 other species of plants, apparently entomopathogenic on elongate hemlock scale (Fiorinia externa).

Notes: Originally established to accommodate entomopathogenic forms of C. acuta-

tum associated with an epizoote of scale insects in eastern USA, where it also occurred as an endophyte in 28 species of plants (Marcelino *et al.*, 2008). Differs culturally from *Colletotrichum acutatum* Simmonds by having colonies on PDA that are grey cottony with aerial mycelium in compact tufts, and in reverse pale brownish-pink with dark flecking, as well as by its ITS and β-tubulin sequences. *Colletotrichum fioriniae* accommodates *C. acutatum* group C according to Lardner *et al.* (1999) or *C. acutatum* group A3 according to Sreenivasa-prasad and Talhinhas (2005).

Colletotrichum fragariae A.N. Brooks, Phytopathology 21: 113 (1931).

Teleomorph: Unknown.

Hosts and disease: Crown rot, stolon lesions and fruit rot of $Fragaria \times ananassa$ (strawberry).

Notes: This species is one of the causal agents of strawberry anthracnose, which is usually caused by three species (including C. acutatum and C. gloeosporioides). Each of these species produces similar symptoms on strawberry which include crown rot, fruit rot and stolon lesions. It has been regarded as a synonym of C. gloeosporioides by von Arx (1957) but then as specific to Fragaria (von Arx, 1981; Sutton, 1992). It is recognized as a distinct species differentiated from C. gloeosporioides and C. acutatum by both conidia and setae on a strawberry leaf agar medium (Gunnell et al., 1992), but its status as a distinct species is under review (Martínez-Culebras et al., 2003). Molecular data based on the rDNA ITS sequences has shown that the divergences between this taxon and a putative C. gloeosporioides strain were too low to distinguish them as separate species (Sreenivasaprasad et al., 1996; Buddie et al., 1999; Martínez-Culebras et al., 2003), but it can be discriminated from other members of the C. gloeosporioides aggregate isolated from strawberry by an MvnI restriction site within the ITS1 region (Martínez-Culebras et al., 2000, 2003). Recent phylogenetic research using a wider range of genes (Johnston, in litt.) supports the work of MacKenzie et al. (2007) indicating that C. fragariae is distinguishable from the rest of the C. gloeosporioides aggregate, but that it is not restricted to strawberry.

Colletotrichum fructi (F. Stevens & J.G. Hall) Sacc. [as 'fructus'], Sylloge Fungorum (Abellini) 22: 1201 (1913).

Teleomorph: Unknown.

Hosts and disease: Fruit rot of apple (Malus domestica). Also reported on pear (Pyrus communis) and walnut (Juglans regia) (Farr et al., 2009), which needs to be confirmed.

Notes: This taxon has been revealed to be a distinct species based on multi-gene analyses and an epitype has been designated (Damm *et al.*, 2009).

Colletotrichum fructicola Prihastuti, L. Cai & K.D. Hyde, Fungal Diversity 39: 96.

Teleomorph: Glomerella sp.

Hosts and disease: Endophyte, epiphyte and pathogen isolated from coffee berries (Coffea sp.) and leaf spots of peanut (Arachis). Notes: This species was first reported from coffee berries in Thailand, isolated as epiphytes, endophytes and pathogens. Conidia of this species are similar to C. kahawae which has been reported associated with coffee berry disease, but they can be distinguished by biochemical tests (Prihastuti et al., 2009). The taxon was introduced as a new species based on molecular data. Subsequent collections in Thailand have shown this species to have a wide host range (L. Cai, pers. comm.).

Colletotrichum fuscum Laubert, Gartenwelt 31: 675 (1927).

Teleomorph: Unknown.

Hosts and disease: Leaf spots on Scrophulariaceae sensu lato (Antirrhinum majus, Digitalis spp., Linaria maroccana, Nemesia strumosa).

Notes: Laubert described *C. fuscum* from diseased leaves of *Digitalis purpurea* in 1927 from Germany. A similar disease on *Digitalis* and related plants is common in New Zealand (Sutton 1980; Anon., 2001–2009) and Japan (e.g. Tomioka *et al.*, 2001). This fungus is genetically distinct from *C. gloeosporioides*, based on sequences from the ITS region (Moriwaki *et al.*, 2003, Cannon *et al.*, 2008) and GPDH intron 2 (Weir, unpubl. data). Both Moriwaki *et al.* (2002) and Cannon *et al.* (2008) recognized that *C. fuscum* was closely related to isolates referred variously to *C. destructivum*,

C. higginsianum, and C. linicola on the basis of ITS sequences. Each of these names is used for fungi causing leaf-spotting diseases of Scrophulariaceae, legumes, crucifers, and Linum respectively. A comparison of published descriptions shows that there are morphological as well as genetic similarities between isolates given these names. For example, all have distinctively-shaped conidia with a slight curve and tapering to narrowly rounded ends (e.g. Spilsbury, 1953; von Arx, 1970; Sutton, 1980; Tomioka et al., 2001). There has been no modern study of the type specimens of these taxa and whether or not they are genetically and biologically distinct remains unresolved.

Colletotrichum gloeosporioides (Penz.) Penz. & Sacc., Atti del Reale Istituto Veneto di Scienze, Lettere ed Arti, Serie 6, 2: 670 (1884). Teleomorph: Glomerella cingulata (Stoneman) Spauld. & H. Schrenk

Hosts and disease: Colletotrichum gloeosporioides (Penz.) Penz. & Sacc. is one of the most common and widely distributed plant pathogens in the world (Sutton, 1992; Cannon et al., 2000). Since its original description (as Vermicularia gloeosporioides Penz.) it has been associated with at least 470 different host genera (Sutton 1980), either as a primary disease-causing organism, or isolated from deteriorated plant parts. It is especially prevalent in the tropics, but has been recorded also from a wide range of temperate and subtropical habitats. The species is well known as a latent pathogen causing post-harvest problems (Prusky and Plumbley, 1992), endophytic strains are commonly isolated from symptomless plant parts (Cannon and Simmons, 2002; Lu et al., 2004; Photita et al., 2004, 2005), and reduced-pathogenicity mutants have been explored as control mechanisms for virulent strains (Yakoby et al., 2002). Some strains are highly host-specific, and are used or planned for use as bioherbicides in several parts of the world (e.g. Watson et al., 2000; Goodwin, 2001; Kaewchai et al., 2009).

Colletotrichum gloeosporioides has been implicated as an economically important pathogen of a wide range of plants, including Agavaceae (Farr et al., 2006), apple (Carvalho et al., 2000), avocado (Mills et al., 1992; Freeman et al., 2000), citrus (Adaskaveg and

Förster, 2000; Timmer and Brown, 2000; Ramos et al., 2006), coffee (Waller and Bridge, 2000), mango (Mills et al., 1992; Afanador Kafuri et al., 2003), olive (Martin and Garcia Figueres, 1999), papaya (Mills et al., 1992), passion fruit (Afanador Kafuri et al., 2003), proteas (Lubbe et al., 2004), stylosanthes (Manners and He, 1997; Munaut et al., 2002), strawberry (Buddie et al., 1999; Smith and Black, 1990; Xiao et al., 2004) and yam (Abang et al., 2002; Mignouna et al., 2002).

Notes: Colletotrichum gloeosporioides as currently circumscribed is a polymorphic species aggregate containing a number of subgroups which show varying degrees of pathogenicity, host-specificity and genetic homogeneity. It was first recognized as an aggregate taxon by von Arx (1957) who placed at least 600 taxa into synonymy (the precise number is difficult to determine due to the high level of homonymy and complications due to teleomorph/anamorph links). Some of these (e.g. C. higginsianum, C. phormii) are now recognized as belonging to separate species, but the vast majority have never been reevaluated. In the absence of living cultures of type specimens with extractable DNA, there are strong practical arguments against their resurrection.

The Colletotrichum gloeosporioides aggregate has been defined using morphological methods by e.g. Sutton (1980) and Baxter et al. (1983), primarily using characteristics of the conidia - they were considered to be cylindrical with rounded ends and less than 4.5 µm in diameter. Such features are not considered to be reliable; ,specially as *Colletotrichum* species in culture frequently produce secondary conidia that are highly variable in size and shape. The aggregate is now more objectively defined using molecular methods, and sequences including rDNA-ITS, β-tubulin, MAT1-2 and GDPH (Morikawi et al., 2002; Du et al., 2005; Gonzalez et al., 2006; Cannon et al., 2008; Than et al., 2008a) may be used to assign strains to C. gloeosporioides in its currently defined sense.

Some recently accepted taxa are now known to belong within the *C. gloeosporioides* aggregate. The best studied of these is *C. kahawae* (Waller *et al.*, 1993), described for the important African pathogen of *Coffea*

known to pathologists as coffee berry disease. Although its recognition as a distinct taxonomic unit is very important in the applied sciences, it is now believed to be an almost clonal entity that can only be reliably distinguished from other taxa within the C. gloeosporioides aggregate using biochemical tests. Colletotrichum fragariae (Brooks, 1931) constitutes a further subgroup within the C. gloeosporioides aggregate, but this exhibits substantially more genetic polymorphism compared with C. kahawae. Recent studies (Weir and Johnston, in litt.) suggest that it can be recognized as a distinct taxonomic entity within the aggregate, but is not specific to strawberry.

ITS analysis has not proved informative in distinguishing subgroups within the *C. gloeosporioides* aggregate, although there are suggestions that a sequence dichotomy exists that is reflected in the gene's secondary structure (Bridge *et al.*, 2008b). The aggregate is currently the subject of intensive phylogenetic research using multiple genes, and it is likely that a series of well-supported monophyletic (though not host-specific) clades will be identified.

Colletotrichum gossypii Southw., Journal of Mycology 6: 100 (1890).

Teleomorph: Glomerella gossypii Edgerton

Hosts and disease: Anthracnose, pink boll rot or seedling blight of cotton (Gossypium).

Notes: The name C. gossypii has been frequently used in the literature (e.g. Silva-Mann et al., 2007; Alves and Pozza, 2009; Chitarra et al., 2009). It was suggested that the two varieties can be differentiated by AFLP, but this observation was based on only ten strains (Silva-Mann et al., 2005). Colletotrichum gossypii var. gossypii supposedly causes anthracnose, mostly resulting in seedling dieback, while C. gossypii var. cephalosporioides is proposed to be the causal agent of ramulose of cotton (Mathieson and Mangano, 1985; Chitarra et al., 2009; Monteiro et al., 2009). There is no consistent report about morphological or molecular markers to differentiate the two varieties. Many authors have considered C. gossypii to be a synonym of C.

gloeosporioides and the taxon was not accepted in Sutton (1992). Colletotrichum gossypii has quarantine significance (EPPO, 2009), particularly as it is seed dispersed. Fresh collections are needed to establish if this is a distinct species and for epitypification.

Colletotrichum graminicola (Ces.) G.W. Wilson [as 'graminicolum'], Phytopathology 4: 110 (1914)

Teleomorph: Glomerella graminicola D.J. Politis

Hosts and disease: Anthracnose of Zea mays (corn).

Notes: Colletotrichum graminicola was introduced in 1914, combining several morphologically similar taxa from Colletotrichum, Dicladium, Colletotrichopsis, and Steirochaete, including C. cereale and what would later be described as C. eleusines, C. jacksonii, C. miscanthi, C. navitas, C. nicholsonii and C. paspali (Crouch et al., 2009a,c). The taxon has been the subject of much controversy over the past 50 years, beginning with the work of von Arx (1957). In order to connect the conidial state of the graminicolous species with a newly introduced sexual morph, Glomerella tucumanensis (von Arx and Müller, 1954), von Arx synonymized all Colletotrichum associated with grasses under the name C. graminicola, including the previously distinct taxa C. sublineola, C. falcatum and C. caudatum. Colletotrichum graminicola sensu lato Arx was considered by most researchers as a group species and in subsequent years the taxon was increasingly subdivided, eventually into 12 species. The identification of a unique teleomorph associated with C. graminicola from Zea mays (Politis, 1975) and subsequent morphological studies by Sutton (1966, 1968) resulted in C. graminicola being established as a taxon limited to this host plant, and C. sublineola, C. falcatum and C. caudatum were re-established as discrete species. Molecular phylogenetic studies confirmed the distinction of C. graminicola sensu stricto (Du et al., 2005: Crouch et al., 2006, 2009a,b, 2010). Although this fungus emerged as an important pathogen of corn crops during the 1970s and 1980s, anthracnose caused by C. graminicola no longer impacts crop production in most developed areas due to the deployment of

resistant cultivars. *Colletotrichum graminicola* is the first *Colletotrichum* species to have its entire genome sequenced (Lisa Vaillancourt, pers. comm.) and is a well established model for fungus-plant interactions, pathogen biology and hemibiotrophic infection (reviewed in Crouch and Beirn, 2009). An epitype strain (M1.001) has been established (Crouch *et al.*, 2006); this is the same strain sequenced through the Broad Institute's Fungal Genome Initiative (http://www.broad.mit.edu/science/projects/fun gal-genome-initiative/current-fgi-sequence-projects).

Colletotrichum hanaui J.A. Crouch, B.B. Clarke, J.F. White & B.I. Hillman, Mycologia 101: 728 (2009).

Teleomorph: Unknown.

Hosts: Anthracnose of *Digitaria* (crabgrass).

Notes: Colletotrichum associated with crabgrass has traditionally been referred to as C. graminicola, although Wilson (1914) did not include isolates of Colletotrichum from crabgrass in his treatment of the species, and Sutton (1980, 1992) limited C. graminicola to the fungus pathogenic to Zea mays. Systematic studies of this morphologically cryptic phylogenetic species were first performed in 2009, when it was described as a distinct taxon based upon multilocus molecular data (Crouch et al., 2009a).

Colletotrichum higginsianum Sacc., Journal of Agricultural Research, Washington 10: 161 (1917).

Teleomorph: Unknown.

Hosts and disease: Anthracnose of Brassicaceae.

Notes: This taxon causes anthracnose leaf spot disease on a wide range of cruciferous plants (Brassicaceae) in South Atlantic and Gulf states of the United States, the West Indies, Japan, and Southeast Asia (Higgins, 1917; Scheffer, 1950; Sutton, 1980; Moriwaki et al., 1997). It was regarded as a synonym of C. gloeosporioides by von Arx (1957), but subsequently has been recognized as a distinct species on the basis of its conidial morphology and consistent association with cruciferous hosts (Sutton, 1980, 1992). It has however, been considered as a synonym of C. destruc-

tivum based on morphology of conidia and appressoria. The infection process closely resembled that of *C. destructivum* on other hosts (e.g. alfalfa, cowpea, and tobacco) and rDNA sequences are similar to *C. destructivum* (O'Connell *et al.*, 2004). More recently, its host range was also considered an unambiguous criterion of delimitation of two species (Sun and Zhang, 2009). Molecular data analysis based on a multilocus comparison is needed before the status of the two taxa can be resolved.

Colletotrichum hippeastri Y.L. Yang, Zuo Y. Liu, K.D. Hyde & L. Cai, Fungal Diversity 39: 133 (2009)

Teleomorph: Unknown.

Hosts and disease: Anthracnose of Hippeastrum vittatum.

Notes: The conidia of this taxon are distinct in being usually narrower near the ends or at centre and germinating conidia form 2-4 cells. This character is distinct from other species of *Colletotrichum*. In addition, *C. hippeastri* clustered separately from any morphologically similar species (e.g. *C. dracaenophilum*, *C. sansevieriae* and *C. nupharicola* (Farr *et al.*, 2006; Johnson *et al.*, 1997; Nakamura *et al.*, 2006) in phylograms inferred from nu-rDNA ITS, calmodulin, β-tubulin, GDPH, actin, chitin synthase A and/or, six combined genes datasets (Yang *et al.*, 2009).

Colletotrichum horii B. Weir & P.R. Johnst., Mycotaxon (in press).

Teleomorph: Unknown.

Hosts and disease: Associated with lesions on unripe fruit, young stems and twigs of persimmon (Diospyros kaki).

Notes: An anthracnose disease of persimmon fruits was described in a Japanese language article by Shotaro Hori in 1910 as Gloeosporium kaki. The next year Seiya Ito (1911), probably unaware of the earlier work, described a pathogen from persimmon fruits and twigs which he also named Gloeosporium kaki, although based on different specimens. This has led to some confusion as the authority has variably been cited as either Ito or Hori. Maffei (1921) described a leaf spot pathogen of persimmon from a specimen collected in Italy as Colletotrichum kaki. Von Arx (1957, 1970)

considered the fungi described by Hori and Maffei to be the same, and synonymous with Colletotrichum gloeosporioides. However, on the basis of biological and morphological differences, Weir and Johnston (in press) concluded that the two species were distinct. They examined specimens from China, Japan and New Zealand, neotypified Gloeosporium kaki Hori and, based on molecular and morphological evidence, renamed the persimmon pathogen Colletotrichum horii, a member of the C. gloeosporioides sensu lato species complex.

Colletotrichum hymenocallidis Y.L. Yang, Zuo Y. Liu, K.D. Hyde & L. Cai, Fungal Diversity 39: 138 (2009).

Teleomorph: Unknown.

Hosts and disease: leaf spot of Hymeno-callis americana.

Notes: The conidial dimensions and shapes of Colletotrichum hymenocallidis are close to C. siamense, whereas their colony characters differ. Colonies of C. siamense are yellowish-white in reverse versus greenish-black in C. hymenocallidis; the mean size of conidia, conidial and mycelial appressorium in C. hymenocallidis are also larger than that of C. siamense (Prihastuti et al., 2009; Yang et al., 2009). Phylogenetic analysis using chitin synthase A gene region and multiple gene loci showed that C. hymenocallidis is a distinct species (Yang et al., 2009).

Colletotrichum jacksonii J.A. Crouch, B.B. Clarke, J.F. White & B.I. Hillman, Mycologia 101: 729 (2009).

Teleomorph: Unknown.

Hosts and disease: Anthracnose of Echinochloa (barnyard grass).

Notes: As with many Colletotrichum species associated with grass hosts, this organism is referred to in the literature as C. graminicola. Lectotype material for C. graminicola (as Dicladium graminicola; Cesati, 1852) was present on both Echinochloa and Zea host substrates, but Sutton (1965, 1980, 1992) constrained C. graminicola to Colletotrichum pathogenic on Zea mays. As described by Crouch et al. (2009a), C. jacksonii is a falcatespored phylogenetic species morphologically

indistinguishable from several closely related graminicolous inhabiting species; it is diagnosable through molecular characters. Although closely related to *C. echinochloae*, a taxon also described from *Echinochloa*, the type of *C. jacksonii* is part of a phylogenetic lineage distinct from *C. echinichloae* (Crouch *et al.*, 2009a; Moriwaki and Tsukiboshi, 2009).

Colletotrichum kahawae J.M. Waller & Bridge, Mycological Research 97: 993 (1993).

Teleomorph: Unknown.

Hosts and disease: Anthracnose of coffee berry (*Coffea arabica*).

Notes: The name C. coffeanum was applied to Colletotrichum species found on coffee in Brazil by F. Noack at the end of the 19th century, where coffee berry disease (CBD) is not present, and is now considered as a poorly defined host-linked group within the C. gloeosporioides aggregate. Therefore, the use of *C. coffeanum* for the CBD pathogen was a misapplication (Sutton, 1992). Waller et al. (1993) renamed the highly pathogenic strain causing coffee berry disease in Africa as C. kahawae based on differences in growth rate and ability to utilize carbon sources. Correll et al. (2000) used SSREP and ALFP techniques to show that C. kahawae is a clonal population and similar results were found by Varzea et al. (2002) using VCG. Its status as a distinct species or infraspecific taxon is discussed in detail by Cannon et al. (2000). The name has continued to be used (see Bridge et al., 2008a; Gichuru et al., 2008; Durand et al., 2009; Van Der Vossen and Walyaro, 2009; Zeru et al., 2009). Data presented by Cai et al. (2009) and Prihastiti et al. (2009) in this issue may indicate that this is a distinct species. In Brazil, different populations of C. gloeosporioides are quite common as endophytes of the coffee tree, an observation already made by Rayner (1948). Others are considered the etiologic agent of a foliar disease known as "mancha mantecosa" or blister spot, described first in Costa Rica (Vargas and Gonzales 1972; Ferreira et al., 2009; Pereira et al., 2009). Nevertheless, conclusive pathogenicity tests or phylogenetic analyses have still to be conducted. C. coffeanum could be a suitable name for this fungus.

Colletotrichum lilii Plakidas ex Boerema & Hamers, Netherlands Journal of Plant Pathology 94(suppl. 1): 12 (1988).

Teleomorph: Unknown.

Hosts and disease: Black scale disease of lily bulbs (*Lilium longiflorum*). Also reported on other *Lilium* spp. and on *Hemerocallis* sp. (Farr *et al.*, 2009), which however needs to be confirmed.

Notes: In a molecular study of curved spored species from herbaceous hosts, this taxon has been shown to be a distinct species (Damm *et al.*, 2009).

Colletotrichum lindemuthianum (Sacc. & Magnus) Briosi & Cavara, I Funghi Parassiti delle Piante Coltivate ed Utili Essicati, Delineati e Descriti, Fasc. 2: no. 50 (1889).

Teleomorph: Glomerella lindemuthiana Shear

Hosts and Disease: Primarily observed on Phaseolus and Vigna species (Fabaceae), causing anthracnose of leaves, stems and fruits (pods). Also reported from a wide range of other leguminous plants, including species of Cassia, Dolichos, Glycine, Lablab, Medicago and Mucuna (Sutton, 1980). Biological control of the fungus using Pseudomonas species has been investigated (Bardas et al., 2009a,b).

Notes: Colletotrichum lindemuthianum as widely circumscribed has notably short, slightly ovoid conidia (9.5-11.5 \times 3.5-4.5 μ m; Sutton, 1992) that do not become septate before germination (Sherriff et al., 1994). It has been accepted as part of the C. orbiculare aggregate based on both morphological and molecular evidence (Sherriff et al., 1994; Sreenivasaprasad et al., 1996), and Liu et al. (2007) considered that there was a good argument for combining the complex into a single species. Mating types were investigated by Rodríguez-Guerra et al. (2005) and García-Serrano et al. (2008), who found that all strains investigated were heterothallic. Several studies on its population biology have been published (e.g. Balardin et al., 1997; Bardas et al., 2009b; Fabre et al., 1995; González et al., 1998; Sicard et al., 1997).

Colletotrichum gloeosporioides occurs commonly on legumes, and some descriptions (e.g. Mordue, 1971) appear to be composites of the two species. The name has been used

frequently in the literature (see Bardas *et al.*, 2009a,b; Campa *et al.*, 2009; Davide and de Souza, 2009; Munda *et al.*, 2009) and thus typification of *C. lindemuthianum* needs to be addressed, with conservation if necessary to preserve the wider application of the name.

Colletotrichum lineola Corda, in Sturm, Deutschlands Flora (Nürnberg) 3: 41 (1831).

Teleomorph: Unknown.

Hosts: Primarily observed on a dead stem of an unknown species in Apiaceae. Also reported on dead stems of Anthriscus sp., Heracleum sp. and Allium giganteum, on petioles of Fragaria sp., rotten fruit of Prunus domestica, as pathogenic on Clarkia elegans, from leaf spot of Trillium sp., on Astrantia major, Tussilago farfara, Euphorbia egula, Lupinis polyphyllus, and Symplocarpus foetidus (Damm et al., 2009).

Notes: This is the type species of the genus but has rarely been mentioned in recent literature. The species was not recognised by von Arx (1957) and Sutton (1980, 1992) regarded it as a synonym of C. dematium. The epitype however, was isolated from a plant belonging to the Apiaceae collected near the location of the original specimen described from Prague, Czech Republic, and the species has been confirmed to be distinct from C. dematium and other species with curved conidia from herbaceous hosts (Damm et al., Colletotrichum lineola had been considered as occuring on Dactylis glomerata and other grass species (Grove, 1937; Farr et al., 2009), which is doubtful, because of the separate host spectrum of Colletotrichum species with curved conidia on graminicolous and herbaceous hosts (Crouch et al., 2009a, Damm et al., 2009). This was recently confirmed for the anthracnose pathogen of switchgrass that was recognised as a new species, C. navitas (Crouch et al. 2010). While many disease reports might have been confused C. lineola with other species with curved conidia, the species was confirmed to be plurivorous having pathogenic and saprotrophic livestyles (Damm et al., 2009).

Colletotrichum linicola Pethybr. & Laff. Scientific Proceedings of the Royal Dublin Society, N.S. 15(no. 30): 368 (1918).

Teleomorph: Unknown.

Hosts and disease: Anthracnose of flax (Linum). It has also been reported from field bindweed (Convolvulus arvensis) in Turkey as C. cf. linicola (Tunali et al., 2008) and may be a potential biocontrol agent.

Notes: Sutton (1980) listed this species under the name C. lini (Westerd.) Tochinai, but later (Sutton, 1992) referred to Dickson's (1956) opinion that the basionym, Gloeosporium lini Westerd. was probably synonymous with Polyspora lini Laff. And not with Colletotrichum linicola. The names Gloeosporium lini Westerd. and Colletotrichum lini (Westerd.) Tochinai should be treated as of uncertain application, and deleted from the synonymy of C. linicola. The species was described from flax (Linum) and appears to occur mainly on this host (http://www.ism pminet.org/Resources/common/names/flax.asp). This name has been used in the literature as a distinct species (e.g. Sreenivasaprasad et al., 1996: Latunde-Dada and Lucas, 2007; Liu et al., 2007; Diederichsen et al., 2008), although it is not clear whether this is a unique species from flax. Fresh collections are needed to establish if this is a distinct species by molecular data analysis and epitypification is necessary.

Colletotrichum liriopes Damm, P.F. Cannon & Crous, Fungal Diversity 39: 71 (2009).

Teleomorph: Unknown.

Hosts: Lilyturf (*Liriope muscari*).

Notes: This species was described on Liriope muscari from Mexico (Damm et al., 2009). This taxon might be identical with the cause of anthracnose of Liriope platyphylla in Korea (Lee 1997), but this needs to be verified.

Colletotrichum lupini (Bondar) Nirenberg, Feiler & Hagedorn, Mycologia 94: 309 (2002).

Teleomorph: Unknown.

Hosts and disease: Anthracnose lesions of lupin (*Lupinus*). The most distinctive symptom is the bending and twisting of stems.

Notes: This taxon was first introduced as Gloeosporium lupini by Bondar (1912) from Brazil. Nirenberg et al. (2002) accepted this species and formally transferred it to Colletotrichum. They designated a neotype with living cultures and also introduced a variety setosum.

It is difficult to distinguish this species from other Colletotrichum species with cylindrical conidia, such as C. gloeosporioides and C. fragariae. This species has been recognised by RAPD banding patterns, nu-rDNA, ITS1 and ITS2 sequence data (Nirenberg et al., 2002; Lotter and Berger, 2005). Colletotrichum lupini strains form a homogeneous group supported by a high bootstrap value of 96% based on DNA sequence data. This species causes a devastating disease on lupins worldwide (Lotter and Berger, 2005) and interaction of infection between the species and host have been studied (Bonivento et al., 2007; Oelofse et al., 2009). The name has been extensively used in the recent literature (e.g. Thomas et al., 2008; Adhikari et al., 2009; Muth et al., 2009).

Colletotrichum malvarum (A. Braun & Casp.) Southw., Journal of Mycology 6: 116 (1891).

Teleomorph: Unknown.

Hosts and disease: Leaf spot on Althaea, Lavatera, Malva, Sida.

Notes: This species was originally described from Malva in Europe as Steirochaete malvarum A. Braun & Casp. and was reported to have very small conidia (8-9 \times 3-4 um). It was later reported from hollyhock and Sida, but with larger conidia (11-28 \times 5 μ m) and Althaea (Tosi et al., 2004). It has also been reported from Chinese mallow (Malva) in Korea (Kim et al., 2008). Colletotrichum malvarum was proposed to be a synonym of C. orbiculare based on conidial morphology, affinity for the Bauhinia purpurea agglutinin (BPA) lectin and a monoclonal antibody (UB 20), infection hyphae and analysis of rDNA sequence data (Bailey et al., 1996). Liu et al. (2007) showed the isolates from Sida spinosa differed from C. orbiculare and several other closely related species based on distinct RFLPs. The species has also been proposed for use as a biological control agent to control prickly sida (Kirkpatrick et al., 1982). There is obvious confusion surrounding this species which may be specific to Malvaceae or even to genera within this family. New collections from hosts in this family are needed to establish whether this is a known species such as C. orbiculare, a distinct species in its own right or even a complex of species.

Colletotrichum miscanthi J.A. Crouch, B.B. Clarke, J.F. White & B.I. Hillman, Mycologia 101: 729 (2009).

Teleomorph: Unknown.

Host and disease: Anthracnose of Miscanthus.

Notes: Wilson (1914) included Colleto-trichum strains associated with Miscanthus in his broad interpretation of C. graminicola, but Sutton limited this species to Zea pathogens (1980; 1992). Colletotrichum miscanthi was introduced in 2009 based on molecular phylogeny.

Colletotrichum musae (Berk. & M.A. Curtis) Arx, Verhandelingen Koninklijke Nederlandse Akademie van Wetenschappen, tweede sect. 51(3): 107 (1957).

Telemorph: Glomerella musarum Petch Hosts and disease: Endophyte, anthracnose, blossom end rot, crown rot and tip rot diseases of banana (Musa spp.).

Notes: Von Arx (1981) included this under C. gloeosporioides as specific to Musa, while Sutton (1980, 1982) accepted this as a distinct species, which is supported by recent molecular work (L. Cai, pers. comm.). This taxon has been reported as the major causal organism of anthracnose and is also responsible for causing crown rot, blossom end rot and tip rot of banana (Nazriya et al., 2007). This taxon has been found on Musa balbisiana, M. cavendishii, M. paradisiaca, and M. sapientum throughout the world (Sutton, 1980), and also has been found on M. acuminata from Jordan (Israeli and Temkin-Gorodeiski, 1977), West Indies (Chillet et al., 2006) and Thailand (Nuangmek et al., 2008). In Sri Lanka, C. musae causes post-harvest disease of many varieties of banana (Antony et al., 2004; Nazriya et al., 2007). This taxon can be isolated from healthy leaves and roots of M. acuminata as an endophyte (Pereira et al., 1999; Photita et al., 2001, 2005; Alvindia and Natsuaki, 2008). Recently, Mahadtanapuk et al. (2007) found C. musae was a pathogen causing anthracnose on curcuma flowers (Curcuma alismatifolia Gagnep.). However, identification of C. musae was not based on molecular data. Colletotrichum musae has been used frequently in the literature (e.g. Daundasekera et al., 2008;

Da Silva *et al.*, 2008; Demerutis *et al.*, 2008; Niroshini Gunasinghe and Karunaratne, 2009) and appears to be a distinct species that needs to be epitypified.

Colletotrichum navitas J.A. Crouch, Mycological Research 113: 1417 (2009).

Teleomorph: Unknown.

Host and disease: Anthracnose of Panicum virgatum.

Notes: Colletotrichum pathogenic to P. virgatum (switchgrass) has been observed from the native range of the grass across the USA since the late 1880s. Prior to the description of C. navitas by Crouch et al. (2009c), the fungus was referred to as either C. lineola (before 1914; the type species for the genus) or C. graminicola (post-1914). Wilson included a specimen of Colletotrichum from switchgrass in his description of C. graminicola, but Sutton (1980) excluded all but isolates from Zea in his treatment of the species. Unlike most Colletotrichum species described from grass hosts, C. navitas is distinguished by relatively large conidia (34.2-40.5 µm long) with a distinct hockey-stick shape. Colletotrichum navitas is the sister taxa to C. graminicola, and may be uniquely diagnosed through its distinct multilocus phylogenetic signature (Crouch et al., 2009c).

Colletotrichum nicholsonii J.A. Crouch, B.B. Clarke, J.F. White & B.I. Hillman, Mycologia 101: 730 (2009).

Teleomorph: Unknown.

Host and disease: Anthracnose of Paspalum dilatatum.

Notes: Prior to its introduction in 2009 as a morphologically cryptic phylogenetic species, Colletotrichum associated with Paspalum hosts were referred to as C. graminicola. Neither Wilson (1914), von Arx (1957) nor Sutton (1980, 1992) included Paspalum strains in their treatments of C. graminicola, but the name was applied because the host was a graminicolous plant (sensu von Arx) rather than as a result of systematic study. It is one of two cryptic species that have been described from Paspalum (see also C. paspali), and is identified through molecular phylogenetics.

Colletotrichum nymphaeae (Pass.) Aa, Netherlands Journal of Plant Pathology 84: 110 (1978).

Teleomorph: Unknown.

Host and disease: Leaf spots arranged in two lines, almost parallel to the midrib of Nymphaea and Nuphar.

Notes: This taxon was considered restricted to Nymphaeaceae by Aa et al. (1978; 1990). It causes anthracnose on water-lilies leaves in the Netherlands and other European countries (Aa et al., 1978). The taxonomic novelty of this species is inconclusive. Although this species is thought to be specific to Nymphaeaceae and morphologically different from C. gloeosporioides. Conidia of C. nymphaeae are ellipsoidal or cylindrical, straight or slightly curved, rounded at the apex, somewhat attenuated, occasionally truncate at the base, whereas conidia of C. gloeosporioides are straight, cylindrical, apex obtuse, tapered towards the truncate base. Aa et al. (1990) considered these criteria were inadequate to conclude this is a distinct species. Johnson et al. (1997) observed this species on Nymphaeceae and compared it with C. nupharicola using morphological and molecular (RAPD and RFLP-ITS) sequence data. Both appressoria and conidia of C. nymphaeae were wider than C. nupharicola. Digestion of the PCR-ITS products with four enzymes produced distinct restriction digest phenotypes, combining other characters, C. nupharicola was introduced as a new species. C. nymphaeae requires further work using ITS and other genes, since only 28s rDNA sequences from only one strain are available.

Colletotrichum orbiculare (Berk. & Mont.) Arx, Verhandelingen Koninklijke Nederlandse Akademie van Wetenschappen, tweede sect. 51(3): 112 (1957).

Teleomorph: Glomerella lagenaria F. Stevens

Hosts and Disease: Anthracnose of Cucurbitaceae, especially melons (Cucumis), watermelon (Citrullus) and cucumber (Cucurbita); see e.g. Correll et al (1993). Walker et al. (1991) showed that strains from cucurbits could infect a wide range of plants, and morphologically similar strains from Xanthium (cocklebur, Asteraceae) could infect cucurbits.

Notes: Colletotrichum orbiculare (also widely referred to in the older literature as C. lagenarium) is traditionally separated from the C. gloeosporioides aggregate in morphological terms by its conidia, which tend to have a smaller length/breadth ratio and which do not become septate at germination. However, the difference is not absolute and von Arx (1957) considered the species to be a divergent ('abweichend') form of C. gloeosporioides. Several molecular studies using ribosomal DNA (Sherriff et al., 1994; Bailey et al., 1996; Sreenivasaprasad et al., 1996, Johnston and Jones, 1997) have shown that *C. orbiculare* is distinct from the C. gloeosporioides aggregate, and belongs to its own aggregate taxon that also include C. lindemuthianum, C. malvarum and C. trifolii. Phylogenetic analysis using multiple genes has not yet been completed for this complex, but a preliminary study (Liu et al., 2007) suggested that the complex should be regarded as a single operational species with a number of host-specific subunits, and strains of C. orbiculare from Xanthium represented a separate infraspecific taxon. Studies required using strains from a wider range of hosts and geographical origin before the systematic arrangement proposed by Liu and colleagues can be confirmed. As this name is commonly used in the literature (e.g. Asakura et al., 2009; Shimizu et al., 2009; Zhang et al., 2009), research on typification and designation of epitype cultures is urgently needed.

The teleomorph of *Colletotrichum orbiculare* was described from UV-irradiated cultures of the anamorph by Stevens (1931) under the name *Glomerella lagenaria*, and also superfluously by Watanabe and Tamura (1952).

Colletotrichum paspali J.A. Crouch, B.B. Clarke, J.F. White & B.I. Hillman, Mycologia 101: 730 (2009).

Teleomorph: Unknown.

Host and disease: Anthracnose of Paspalum notatum.

Notes: This is one of two morphologically cryptic taxa described from *Paspalum*; see *Colletotrichum nicholsonii*.

Colletotrichum phaseolorum S. Takim., Annals of the Phytopathological Society of Japan 5: 21 (1934).

Teleomorph: Unknown.

Hosts and disease: Anthracnose of Azuki bean (Vigna angularis syn. Phaseolus radiatus var. aurea) and cowpea (Vigna unguiculata subsp. cylindrica, syn. Vigna catiang var. sinensis).

Notes: According to a molecular study, the taxon is distinct from other species with curved conidia (Damm et al., 2009). However, while strains from Azuki bean and cowpea were regarded as belonging to the same species in the original description (Takimoto 1934), they differ in DNA sequence data from each other (Damm et al., 2009).

Colletotrichum phormii (Henn.) D.F. Farr & Rossman, Mycological Research 110: 1403 (2006).

Teleomorph: Glomerella phormii (J. Schröt.) D.F. Farr & Rossman.

Hosts and disease: Causing leaf lesions in Phormium.

Notes: This species was revisited by Farr et al. (2006) who provided a detailed account of the nomenclature of this species and its synonyms following examination of the holotype. Their illustrations and the cultures used in barcoding are derived from recent collections or cultures ex Kinghorn (1936) and there is no ex-type isolate. Based on ITS and LSU data C. phormii is a unique species. The taxon is only known from *Phormium* species (New Zealand flax), is the slowest growing species on Agavaceae and is differentiated from other similar species by longer, wider conidia (Farr et al., 2006). All 14 C. phormii sequences in NCBI, are named as Fusarium phormii, but are clearly C. phormii (accession numbers DQ286134 to DQ286147, accessed 21 April 2009). An epitype should be designated.

Colletotrichum phyllachoroides (Ellis & Everh.) Arx, Verh Verhandelingen Koninklijke Nederlandse Akademie van Wetenschappen, tweede sect. 51(3): 119 (1957).

Teleomorph: Unknown.

Hosts and Disease: Necrotic leaf spot of Artemisia species (Asteraceae; von Arx, 1957; Sutton, 1980), the conidiomata occupying most of the lesions. Records on tomato (Lycopersicon, Solanaceae) in BPI and on Suaeda

fruticosa (Chenopodiaceae; see below) need confirmation.

Notes: This is a poorly known species that is not definitely known from culture, recognized by its distinctive black conidiomata and its rather large, wide, falcate conidia. The only definitively identified collections are from California, USA.

Fisher and Petrini (1987) reported Colletotrichum phyllachoroides as a semidominant endophyte in leaves of Suaeda fruticosa from Dorset, UK. The cultures were presumably identified using morphological features, but apparently none was preserved in either living or dried condition, so the identification must be regarded as tentative. This taxon needs recollecting and DNA sequencing to establish if it is a unique species.

Colletotrichum rusci Damm, P.F. Cannon & Crous, Fungal Diversity 39: 72 (2009).

Teleomorph: Unknown.

Hosts: Ruscus.

Notes: This species was described on Ruscus from Italy (Damm et al., 2009). The biology and life cycle of this fungus is still unknown.

Colletotrichum sansevieriae M. Nakamura & M. Ohzono, Journal of General Plant Pathology 72: 253 (2006).

Host and disease: Causing water-soaked lesions on leaves of *Sansevieria*.

Notes: This species cannot be distinguished from C. boninense, C. gloeosporioides, and other Colletotrichum species with broad and cylindrical conidia, as their conidia sizes and shape overlap. This species shows high specificity to Sansevieria (Agavaceae) causing leaf anthracnose; pathogenicity testing showed that it does not cause disease on selected plant species belonging to 11 families (Nakamura et al., 2006). It can also be distinguished from other similar species based on ITS sequences.

Colletotrichum siamense Prihastuti, L. Cai & K.D. Hyde, Fungal Diversity 39: 98 (2009).

Teleomorph: Unknown.

Hosts and disease: Endophyte, epiphyte and pathogen isolated from coffee berries (Coffea sp.).

Notes: This taxon comprises epiphytes, endophytes and pathogens from coffee and was described based on morphology and multigene sequence data. It resembles *C. acutatum* in cultural characters and conidial shape, however *C. siamense* conidia are fusiform with obtuse slightly rounded ends versus fusiform in *C. acutatum* (Prihastuti *et al.*, 2009). Subsequent collections in Thailand have shown this species to have a narrow host range (L. Cai, pers. comm.).

Colletotrichum simmondsii R.G. Shivas & Y.P. Tan, Fungal Diversity 39:119 (2009)

Teleomorph: Unknown.

Hosts and disease: Endophyte in Actinidia chinensis, fruit rot of Capsicum frutescens, Carica papaya, Cyphomandra, Fragaria x ananassa, Litchi chinensis, Lycopersicon esculentum, Mangifera indica, Nephelium lappaceum, Persea americana, Vaccinium corym-bosum.

Notes: Morphologically similar to Colletotrichum acutatum Simmonds but differs culturally by having colonies on PDA that are grey cottony and in reverse pale grey to pale orange sometimes with dark flecking, as well as by its ITS and β-tubulin sequences. Colletotrichum simmondsii accommodates C. acutatum group D according to Lardner et al. (1999) or C. acutatum group A2 according to Sreenivasaprasad and Talhinhas (2005).

Colletotrichum spaethianum (Allesch.) Damm, P.F. Cannon & Crous, Fungal Diversity 39: 74 (2009).

Teleomorph: Unknown.

Hosts: Hosta sieboldiana, Lilium sp., Hemerocallis sp.

Notes: Originally described on Funkia univittata (= Hosta sieboldiana) in Germany, this taxon was combined in the genus Colletotrichum, epitypified and confirmed as a discrete species. The taxon is closely related or identical with the causal agents of anthracnose of Japanese radish (Raphanus sativus var. narukovuri hortensis) and (Polygonatum falcatum) (Damm et al., 2009) that were recently reported from Japan (Sato et al., 2005; Tomioka et al., 2008). However, further molecular studies are necessary to confirm the identity of these pathogens and to study the possible synonymy of the taxon with *C. liliacearum*.

Colletotrichum spinaciae Ellis & Halst., Journal of Mycology 6: 34 (1890).

Teleomorph: Unknown.

Hosts and disease: Leaf spots and anthracnose of spinach (Spinacea oleracea). Also reported on white goosefoot (Chenopodium album), purslane (Portulaca oleracea) and alfalfa (Medicago sativa) (Damm et al., 2009).

Notes: Regarded as C. dematium forma spinaciae by von Arx (1957), but confirmed as a discrete species in a molecular study (Damm et al., 2009). According to von Arx (1957), the species develops only weak symptoms on hosts other than spinach.

Colletotrichum sublineola Henn. ex Sacc. & Trotter, ['as sublineolum'] Sylloge Fungorum 22: 1206 (1913).

Teleomorph: Currently undescribed; but see notes below.

Hosts and disease: Anthracnose of Sorghum bicolor, S. halepense, Eremochloa ophiuroides.

Notes: Von Arx (1957) considered this taxon, along with C. falcatum and C. caudatum, as synonyms of C. graminicola, but Sutton (1980) re-established these species as distinct taxa based on appressorial characters. Molecular fingerprinting and phylogenetic analysis supports C. sublineola as a distinct taxon (for review, see Crouch et al., 2006), and an epitype strain has been established (S3001). Due to the taxonomic changes associated with this and other graminicolous Colletotrichum, the host range of C. sublineola is poorly defined, with only Sorghum bicolor, S. halepense, and Eremochloa ophiuroides (centipedegrass) confirmed as hosts through molecular analysis (Crouch, unpubl. data). Historical observations of C. sublineola from other host plants based upon morphology may be erroneous, as the fungus is morphologically indistinguishable from several other graminicolous Colletotrichum species (Crouch et al., 2009a). The teleomorph of this fungus has been identified (Vaillancourt and Hanau, 1992), but not yet formally described or named.

Colletotrichum tofieldiae (Pat.) Damm, P.F. Cannon & Crous, Fungal Diversity 39: 77 (2009).

Teleomorph: Unknown.

Hosts: Tofieldia calyculata, lupin (Lupinus polyphyllus), Dianthus sp.

Notes: This species, originally described on *Tofieldia* sp. from Sichuan, was combined in the genus *Colletotrichum* and confirmed as a discrete species (Damm *et al.*, 2009). The taxon is closely related or identical with the causal agent of anthracnose of statice (*Goniolimon tataricum*) (Damm *et al.*, 2009) that was recently reported from Bulgaria (Bobev *et al.*, 2009). However, further molecular studies are necessary to confirm the identity of this pathogen.

Colletotrichum trichellum (Fr.) Duke, Transactions of the British Mycological Society 13: 173 (1928).

Teleomorph: Unknown.

Host and disease: leaf spots of Ivy (Hedera spp.).

Notes: Von Arx (1957) included this species as a facultative synonym of C. dematium. Sutton (1962) noted significant morphological differences between the two species with C. trichellum showing thinly developed acervuli, and C. dematium showing extensive and dark-walled acervuli. conidia of C. trichellum were found to be less falcate and tapered abruptly at each end as opposed to those of C. dematium which are strongly falcate with long acute apices. The most marked morphological difference between the two species is the appearance of the appressoria. Those of C. trichellum are relatively darker with irregular, crenate edges while those of C. dematium are cinnamon buff and rarely develop complex edges (Sutton, 1962). Based on these observations, Sutton (1962) regarded them as separate species and von Arx (1981) also accepted C. trichellum as a distinct taxon, which was confirmed in a molecular study (Damm et al., 2009).

The taxon causes leaf spots on species of *Hedera* (ivy) worldwide, especially *H. helix*. Additional species of *Colletotrichu*m have also been observed from ivy, including *C. dematium* (Sutton, 1962) and a new collection of an unnamed *Colletotrichum* species from ivy

plants in Thailand (Hyde, unpubl. data). The *Colletotrichum* species occurring on ivy as well as the identity of *C. trichellum* require further research

The type specimen for *C. trichellum* is old (pre-1817) and in poor condition; thus there is a need of epitypification. Since it is unlikely that DNA can be extracted from the type, an epitype from a living culture originating from the UK has been identified (Crouch and Hyde, unpubl. data).

Colletotrichum trifolii Bain, Journal of Mycology 12: 193 (1906).

Teleomorph: Unknown.

Hosts and disease: Anthracnose of flower heads and petioles and rarely leaves of Medicago (alfalfa), Trifolium (red clover).

Notes: This species was described from Tennessee and West Virginia in 1906 causing devastating disease of red clover (Bain and Essary, 1906). The conidia were reported as 11-13 × 3-4 μm and straight with rounded ends as in the present day "acutatum" type. The protein and gene levels of this species and signalling pathways during infection as well as genes for host resistance have been recently studied (Warwar and Dickman, 1996; Yang and Dickman, 1999; Chen and Dickman, 2004; Armour et al., 2008; Yang et al., 2009). C. trifolii was erected more than a hundred years ago and its status cannot clearly be determined; thus there is a need for epitypification.

Colletotrichum truncatum (Schwein.) Andrus & W.D. Moore, Phytopathology 25: 122 (1935).

Teleomorph: Unknown. The teleomorph of the anthracnose pathogen of lentil named as Glomerella truncata C.L. Armstr. & Banniza is not related to *C. truncatum*, but to the *C. destructivum* aggregate (Latunde-Dada and Lucas 2007, Damm *et al.*, 2009).

Hosts and disease: Anthracnose on legume crops such as bean (Phaseolus lunatus, P. vulgaris), soybean (Glycine max), peanut (Arachis hypogea), and lucerne (Medicago sativa), anthracnose of chilli pepper (Capsicum annuum, C. frutescens), peppers (Capsicum annuum), and several other hosts in a wide range of plant families (Pring et al., 1995; Shenoy et al., 2007; Damm et al., 2009; Farr et al., 2009),

of which many need to be confirmed. Associated with symptoms of leaf tip die-back, foliar blight, leaf spot, leaf lesions and boll rot of various plants (Shenoy *et al.*, 2007). The taxon was also identified from a corneal ulcer of human eye (Damm *et al.*, 2009).

Notes: The taxon, originally described on Phaseolus lunatus, was epitypified recognised as distinct (Damm et al., 2009). The major pathogen of the anthracnose disease complex of chilli pepper, known as C. capsici The causal organism of lentil anthracnose referred to C. truncatum by Ford et al. (2004) does not belong to this species. Latunde-Dada and Lucas (2007) showed that C. truncatum isolates from lentil, peas and cowpeas had similar phylogeny based on ITS sequences to C. destructivum isolated from faba beans and lucerne, and C. linicola isolates from flax. Further research is required to clarify the identity of these anthracnose pathogens. The species has a great deal of intraspecific variability and a wide host range (Damm et al., 2009).

Colletotrichum verruculosum Damm, P.F. Cannon & Crous, Fungal Diversity 39: 81 (2009).

Teleomorph: Unknown.

Hosts: Crotalaria juncea.

Notes: This species was described on Crotalaria juncea from Zimbabwe (Damm et al., 2009). The biology and life cycle of this fungus is still unknown.

Colletotrichum xanthorrhoeae R.G. Shivas, Bathgate & Podger, Mycological Research 102: 280 (1998).

Teleomorph: Unknown.

Hosts and disease: Leaf spots of Xanthorrhoea.

Notes: The pathogenicity of this species was demonstrated on young leaves of seedling plants (Shivas et al., 1998). Cannon et al. (2008) considered this species a segregate within the *C. gloeosporioides* aggregate that remained to be further characterized using molecular techniques (Table 1).

Colletotrichum yunnanense Xiao Ying Liu & W.P. Wu, Mycotaxon 100: 139 (2007).

Teleomorph: Unknown.

Host: Buxus sp.

Notes: This recently reported species is only known as an endophyte from Buxus sp. Since other known endophytic species, such as C. acutatum, C. asianum, C. boninense, C. fructicola, C. gloeosporioides and C. siamense are also well known plant pathogens (Lu et al., 2004; Prihastuti et al., 2009), it is highly likely that C. yunnanense has other life styles. The biology and life cycle of this fungus needs further investigation. Phylogenetically and morphologically this fungus is most closely related to C. dracaenophilum, but it produces smaller conidia and has a faster growth rate in culture.

Recently used doubtful species

Colletotrichum aquatile R. Sprague [as 'aquatilis'], Mycologia 49: 838 (1958) [1957].

Hosts: Catabrosa, Puccinellia.

Notes: Based upon the host range (coolseason grasses) and morphology, this taxon is likely to be *C. cereale*. This species was not accepted by Sutton (1980, 1992). Fresh collections are required before this taxon can be established as distinct from *C. cereale*.

Colletotrichum araliae Kamal & R.P. Singh, Indian Phytopathology 33: 594 (1981) [1980].

Hosts: Aralia.

Notes: This taxon is the second species described from *Aralia*, and is poorly justified. The conidia are falcate and fresh collections are needed to establish whether it is a distinct species.

Colletotrichum artocarpi Delacr. ex Sacc. & D. Sacc., Sylloge Fungorum 18: 466 (1906).

Teleomorph: Glomerella artocarpi Delacr.

Host and disease: Leaf spots of Artocarpus spp. (jackfruit).

Notes: This name was recently used for a fungus causing leaf spots of Artocarpus heterophyllus Lam. in Thailand (Sanyong and Amarakul, 2001). Dieback of the same host was also reported to be caused by C. gloeosporioides. It is also reported as a pathogen in Hawaii (http://www.extento. hawaii.edu/Kbase/Crop/Type/col_prim.htm), Guam (http://www.prevalentfungi.org/subject. cfm?id=56164) and

French Polynesia (http://www.agriculture.gov.pf/UserFiles/organismes%20nuisibles%20PF.pdf). Otherwise the name has been little used. The taxon needs re-evaluation following fresh collection, morphological examination and sequence data analysis.

Colletotrichum atramentarium (Berk. & Broome) Taubenh., Memoirs of the New York Botanical Garden 6: 549 (1916).

Hosts and disease: reported to cause anthracnose of several solanaceous hosts (Farr et al., 2009).

Notes: This species is listed as a synonym of *C. coccodes* in *Index Fungorum*. The name has been used occasionally in some recent nontaxonomic publications (e.g. Miyazawa *et al.*, 2000; Carpinella *et al.*, 2003), but does not appear to represent a distinct species.

"Colletotrichum bromeliacearum" Birge, USDA Bureau of Plant Industry Bulletin 226: 14 (1912), nom. nud.

Hosts: Billbergia, Tillandsia recurvata. Disease: Anthracnose on stems.

Notes: This name is used in Farr et al. (1989), but is not listed in Index Fungorum. It needs to be established whether the Colletotrichum species on Bromeliaceae are distinct species.

Colletotrichum capsici (Syd.) E.J. Butler & Bisby, The Fungi of India, Imperial Council of Agricultural Research Scientific Monograph 1: 152 (1931).

Teleomorph: Unknown.

Hosts and disease: Anthracnose of chilli pepper (Capsicum annuum, C. frutescens) and has been reported to have a wide putative host range (121 genera in 45 plant families, Shenoy et al., 2007). The taxon has been reported to cause anthracnose on cotton, peppers, tomatoes and a wide range of legume species (Pring et al., 1995). It has also been reported to infect and survive in weed and flower hosts such as hibiscus and chrysanthemum (Roberts et al., 2001) and associated with symptoms of leaf tip die-back, foliar blight, leaf spot, leaf lesions and boll rot of various plants (Shenoy et al., 2007).

Notes: Because of its importance as a plant pathogen *C. capsici* has been well-studied

and epitypification by Shenoy *et al.* (2007) established a connection between the original morphological species concept and DNA-sequence based assessments. Subsequent research on multi-locus phylogenies and morphology of the curved spore *Colletotrichum* species, which includes *C. capsici*, synonymised this taxon with *C. truncatum* (Damm *et al.*, 2009), along with *C. curvatum*. Not all researchers are, however, in agreement with the synonomy proposed by Damm *et al.* (2009).

Colletotrichum caricae F. Stevens & G.J. Hall, Zeitschrift für Pflanzenkrankheiten 19: 68 (1909).

Host: Ficus carica.

This species was listed in Sutton (1992) although there is no modern description. Von Arx (1957) had reduced it to synonymy with *C. gloeosporioides* but later (von Arx, 1981) accepted it as a species specific to *Ficus* (as *Carica*). This species therefore needs to be recollected from *Ficus* and sequenced to establish whether it is distinct.

Colletotrichum corchori Pavgi & U.P. Singh, Mycopathologia et Mycologia Applicata 27: 84 (1965).

Teleomorph: Unknown.

Hosts and disease: Anthracnose spots on leaves of *Corchorus* (jute).

Notes: This taxon has been introduced twice from jute plants, firstly for a species of Colletotrichum occurring on Corchorus capsularis in Japan (Ikata and Yoshida, 1940) and then on C. aestruans (= C. acutangulus) (jute) in India (Pavgi and Singh, 1965). There have been a few subsequent studies on this species (e.g. Khan and Strange, 1975, 1976; Rahman and Junaid, 2008), one showing it comprises at least six races (Choudhury and Ahmed, 1969). Sutton (1980) accepted this as a distinct species and its current name is listed as C. corchori Pavgi & U.P. Singh in Index Fungorum. In a multigene analysis, a strain from Corchorus capsularis from Bangladesh, originally identified as C. corchori, proved to be C. truncatum (Damm et al., 2009). The collection from Zea in Brazil (Mendes et al., 1998) is doubtful. Fresh collections of this species are needed to establish whether this is a distinct species.

Colletotrichum curvatum Briant & E.B. Martyn, Tropical Agriculture 6: 258 (1929).

Host: Crotalaria.

Notes: There is no modern description for this taxon and Sutton (1992) and von Arx (1957) considered it a synonym of *C. dematium*. However, Damm *et al.* (2009) showed the taxon to be a synonym of *C. truncatum*.

Colletotrichum fusarioides (Ellis & Kellerm.) O'Gara, Mycologia 16: 169 (1924).

Teleomorph: Glomerella fusarioides Edgerton

Host: Asclepias.

Notes: This species was mentioned in Sutton (1992) but has been little used otherwise since its introduction.

Colletotrichum gnaphalii Syd., Annales Mycologici 37: 419 (1939).

Host: Gnaphalium.

Notes: This species has rarely been mentioned other than in Sutton (1992). It has small conidia (8-15 \times 4-6 μ m) and very little is known about it. Fresh collections are needed.

Colletotrichum helichrysi (G. Winter) Arx, Verhandelingen Koninklijke Nederlandse Akademie van Wetenschappen, tweede sect. 51(3): 92 (1957).

Host: Helichrysum.

Notes: This species was accepted by von Arx (1957), excluded in Sutton (1980), but discussed in Sutton (1992). Very little is known about this species and it must be considered doubtful until fresh collections are made.

Colletotrichum liliacearum Ferraris, Malpighia 16: 35 (1902).

Hosts: Hemerocallis (daylily).

Notes: Sutton (1992) considered C. lilii Plakidas ex Boerema & Hamers (from Lilium longiflorum) a synonym of C. liliacearum. It was suggested by Sobers and Plakidas (1962) that C. liliacearum should be used for non-pathogenic forms and C. lilii for pathogenic forms. These taxa require fresh collections for further work. Damm et al. (2009) found two distinct species occuring on Lilium, C. lilii and C. spaethianum. More collections from Hemerocallis and Lilium are needed to establish

whether this taxon is a distinct species or a synonym of *C. spaethianum*.

Colletotrichum neriicola Hüseyın & Selçuk [as 'neriicolum'], Lidia 5: 149 (2001)

Host and disease: On leaves of cultivated *Nerium oleander*.

Notes: This species has not been mentioned since it was introduced. Fresh collections and isolates are needed to establish if it is a distinct species.

Colletotrichum nigrum Ellis & Halst., Bulletin of the Torrey Botanical Club 18:15 (1891).

Host: Capsicum annuum.

Notes: This species has been placed in synonymy with *C. gloeosporioides* (von Arx, 1957) and later maintained as a species restricted to *Capsicum* (von Arx, 1981). The name has rarely been used and should be considered doubtful.

Colletotrichum nupharicola D.A. Johnson, Carris & J.D. Rogers, Mycological Research 101: 647 (1997).

Hosts: Nuphar, Nymphaea.

Notes: This species falls into C. gloeo-sporioides sensu lato based on ITS and GPDH sequences (Weir, unpubl. data) and may not represent a unique species. Cultures are available so further analysis is needed using multigenes to establish its status.

Colletotrichum paludosum (Ellis & Galloway) Arx, Verhandelingen Koninklijke Nederlandse Akademie van Wetenschappen, tweede sect. 51(3): 115 (1957).

Host: Peltandra.

Notes: Von Arx (1957) considered this species to be close to *C. crassipes*. The name has not been used in recent literature.

Colletotrichum psoraleae (Peck) Arx, Verhandelingen Koninklijke Nederlandse Akademie van Wetenschappen, tweede sect. 51: 125 (1957).

Host: Psoralea.

Notes: This species is mentioned in Sutton (1992) but has not been referred to in the recent literature.

Colletotrichum taiwanense Sivan. & W.H. Hsieh, Mycological Research 97: 1525 (1993). Teleomorph: Glomerella septospora Sivan. & W.H. Hsieh

Host: Styrax formosanus.

Notes: This is a very unusual species and is unlikely to be Glomerella / Colletotrichum. The Glomerella-like teleomorph has ascomata with a well-develop wall or even stroma, asci lack an apical ring and ascospores are 3-septate and relatively massive (60-95(-100) µm long). Asci with Glomerella-like ascospores were found mixed with asci containing 3-septate ascospores in the same perithecium (Sivanesan and Hsieh, 1993). The anamorphic state also has unusual conidiogeneous cells for the genus and conidia are also relatively large, 22-35(45) \times 5-8 µm in 0-3-septate conidia and (50-58 \times 5-8 um) in 4-5-septate conidia. Confirmation that this is a Glomerella species with a Colletotrichum anamorph is required.

Colletotrichum typhae H.C. Greene, Transactions of the Wisconsin Academy of Science and Arts, Letters 44: 41 (1956) [1955].

Host: Typha latifolia

Notes: The species was mentioned in Sutton (1992) but has not otherwise been mentioned recently in the literature.

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References

- Aa, H.A. van der (1978). A leaf spot of *Nymphaea alba* in the Netherlands. Netherlands Journal of Plant Pathology 84: 109-115.
- Aa, H.A. van der, Noordeloos, M.E. and de Gruyter, J. (1990). Species concepts in some larger genera of the coelomycetes. Studies in Mycology 32: 3-19.
- Abang, M.M., Winter, S., Green, K.R., Hoffmann, P., Mignouna, H.D. and Wolf, G.A. (2002). Molecular identification of *Colletotrichum gloeosporioides* causing yam anthracnose in Nigeria. Plant Pathology 51: 63-71.
- Adaskaveg, J.E. and Förster, H. (2000). Occurrence and management of anthracnose epidemics caused by *Colletotrichum* species on tree fruit crops in California. In: *Colletotrichum. Host Specificity*,

Pathology and Host-Pathogen Interaction (eds. D. Prusky, S. Freeman and M.B. Dickman). APS Press, St Paul, Minnesota: 317-336.

- Adhikari, K.N., Buirchell, B.J., Thomas, G.J., Sweetingham, M.W. and Yang H. (2009). Identification of anthracnose resistance in *Lupinus albus* L. and its transfer from landraces to modern cultivars. Crop and Pasture Science 60: 472-479.
- Afanador-Kafuri, L., Minz, D., Maymon, M. and Freeman, S. (2003). Characterization of *Colletotrichum* isolates from tamarillo, *Passiflora* and mango in Colombia and identification of a unique species from the genus. Phytopathology 93: 579-587.
- Alfieri Jr., S.A., Langdon, K.R., Wehlburg, C., and Kimbrough, J.W. (1984). Index of Plant Diseases in Florida (Revised). Florida Department of Agriculture and Consumer Services, Division of Plant Industries, Bulletin 11: 1-389.
- Alves, M.D.C. and Pozza, E.A. (2009). Scanning electron microscopy applied to seed-borne fungi examination. Microscopy Research and Technique 72: 482-488.
- Alvindia, D.G. and Natsuaki, K.T. (2008). Evaluation of fungal epiphytes isolated from banana fruit surfaces for biocontrol of banana crown rot disease. Crop Protection 27: 1200-1207.
- Anon. 2001-2009. NZFungi database of New Zealand fungi. Landcare Research, New Zealand. http://nzfungi.landcareresearch.co.nz
- Anthony, S., Abeywickrama, K., Dayananda, R., Wijeratnam, S. and Arambewela, L. (2004). Fungal pathogens associated with banana fruit in Sri Lanka, and their treatment with essential oils. Mycopathologia 157: 91-97.
- Aqeel, A.M., Pasche, J.S. and Gudmestad, N.C. (2008). Variability in morphology and aggressiveness among North American vegetative compatibility groups of *Colletotrichum coccodes*. Phytopathology 98: 901-909.
- Armour, D.J., Mackie, J.M., Musial, J.M. and Irwin, J.A.G. (2008). Transfer of anthracnose resistance and pod coiling traits from *Medicago arborea* to *M. sativa* by sexual reproduction. Theoretical and Applied Genetics 117: 149-156.
- Armstrong-Cho, C.L. and Banniza, S. (2006). Glomerella truncata sp. nov., the teleomorph of Colletotrichum truncatum. Mycological Research 110: 951-956.
- Arx, J.A. von (1957). Die Arten der Gattung *Colleto-trichum* Cda. Phytopathologische Zeitschrift 29: 414-468.
- Arx, J.A. von (1970). A revision of the fungi classified as *Gloeosporium*. Bibliotheca Mycologica 24: 1-203.
- Arx, J.A. von (1981). The Genera of Fungi Sporulating in Pure Culture. J. Cramer, Vaduz.
- Arx, J.A. von and Müller, E. (1954). Die amerosporen Gattungen der Pyrenomyceten. Beitrage zur Kryptogamenflora der Schweiz 11: 1-434.

- Asakura, M., Ninomiya, S., Sugimoto, M., Oku, M., Yamashita, S.I., Okuno, T., Sakai, Y. and Takano, Y. (2009). Atg26-mediated pexophagy is required for host invasion by the plant pathogenic fungus *Colletotrichum orbiculare*. Plant Cell 21: 1291-1304.
- Babu, A.M., Chowdary, N.B., Kumar, V., Rajan, M.V. and Dandin, S.B. (2008). Infection process of *Colletotrichum dematium* on mulberry leaves: An unusual method of sporulation. Archives of Phytopathology and Plant Protection 41: 290-295.
- Bailey, J.A., O'Connell, R.J., Pring, R.J. and Nash, C. (1992). Infection strategies of *Colletotrichum* species. In: *Colletotrichum: Biology, Pathology and Control* (eds. J.A. Bailey and M.J. Jeger). CAB International: Wallingford: 88-120.
- Bailey, J.A., Nash, C., Morgan, L.W., O'Connell, R.J. and TeBeest, D.O. (1996). Molecular taxonomy of *Colletotrichum* species causing anthracnose on the Malvaceae. Phytopathology 86: 1076-1083.
- Bain, S.M. and Essary, S.H. (1906). A new anthracnose of alfalfa and red clover. Journal of Mycology 12: 192-193.
- Balardin, R.S., Jarosz, A.M. and Kelly, J.D. (1997). Virulence and molecular diversity in *Colleto-trichum lindemuthianum* from South, Central and North America. Phytopathology 87: 1184-1191.
- Bardas, G.A., Lagopodi, A.L., Kadoglidou, K. and Tzavella-Klonari, K. (2009a). Biological control of three *Colletotrichum lindemuthianum* races using *Pseudomonas chlororaphis* PCL1391 and *Pseudomonas fluorescens* WCS365. Biological Control 49: 139-145.
- Bardas, G.A.; Koutita, O. and Tzavella-Klonari, K. (2009b). Molecular diversity and assessment of biological characteristics of Greek *Colletotrichum lindemuthianum* populations. Journal of Phytopathology 157: 311-318.
- Baxter A.P., Westhuizen, G.C.A. van der and Eicker, A. (1983). Morphology and taxonomy of South African isolates of *Colletotrichum*. South African Journal of Botany 2: 259-289.
- Begum, M.M., Sariah, M., Puteh, A.B. and Abidin, M.A.Z. (2008a). Pathogenicity of *Colletotrichum truncatum* and its influence on soybean seed quality. International Journal of Agriculture and Biology 10: 393-398.
- Begum, M.M., Sariah, M., Abidin, M.A.Z., Puteh, A.B. and Rahman, M.A. (2008b). Antagonistic potential of selected fungal and bacterial biocontrol agents against *Colletotrichum truncatum* of soybean seeds. Pertanika Journal of Tropical Agricultural Science 31: 45-53.
- Begum, M.M., Sariah, M., Puteh, A.B. and Zainal Abidin, M.A. (2008c). Detection of seed-borne fungi and site of infection by *Colletotrichum truncatum* in naturally-infected soybean seeds. International Journal of Agricultural Research 2: 812-819.
- Ben-Daniel, B., Bar-Zvi, D. and Tsror, L. (2009). An improved large-scale screening method for assessment of *Colletotrichum coccodes*

- aggressiveness using mature green tomatoes. Plant Pathology 58: 497-503.
- Biggs, A.R. and Miller, S.S. (2001). Relative susceptibility of selected apple cultivars to *Colletotrichum acutatum*. Plant Disease 85: 657-660.
- Bobev, S.G., Jelev, Z.J., Zveibil, A., Maymon, M. and Freeman, S. (2009). First report of anthracnose caused by *Colletotrichum dematium* on statice (goniolimon tataricum, synonym limonium tataricum) in Bulgaria. Plant Disease 93: 552.
- Boland, G.J. and Brochu, L.D. (1989). *Colletotrichum dsetructivum* on alfalfa in Ontario and cultivar response to anthracnose. Canadian Journal of Plant Pathology 11: 303-307.
- Bondar, G. (1912). Tremoco branco e suas molestias. Boletim de Agricultura São Paulo 13: 427-432.
- Bonivento, D. Pontiggia, D. Matteo, A.D. Fernandez-Recio, J., Salvi, G. Tsernoglou, D. Cervone, F. Lorenzo, G.D. and Federici, L.(2007). Crystal structure of the endopolygalacturonase from the phytopathogenic fungus *Colletotrichum lupini* and its interaction with polygalacturonase-inhibiting proteins. Proteins 70: 294-299.
- Bridge, P.D., Waller, J.M., Davies, D. and Buddie, A.G. (2008a). Variability of *Colletotrichum kahawae* in relation to other *Colletotrichum* species from tropical perennial crops and the development of diagnostic techniques. Journal of Phytopathology 156: 274-280.
- Bridge, P.D., Cannon, P.F., Buddie, A.G., Baker, M., and Borman, A.M. (2008b). Domain II hairpin structure in ITS1 sequences as an aid in differentiating recently evolved animal and plant pathogenic fungi. Mycopathologia 166: 1-16.
- Brooks, A.N. (1931). Anthracnose of strawberry caused by *Colletotrichum fragariae*, n. sp. Phytopathology 21: 739-744.
- Buddie, A.G., Martinez-Culebras, P., Bridge, P.D., Garcia, M.D., Querol, A., Cannon, P.F. and Monte, E. (1999). Molecular characterization of *Colletotrichum* strains derived from strawberry. Mycological Research 103: 385-394.
- Cai, L., Hyde, K.D., Taylor, P.W.J., Weir, B., Waller, J., Abang, M.M., Zhang, J.Z., Yang, Y.L., Phoulivong, S., Liu, Z.Y., Prihastuti, H., Shivas, R.G., McKenzie, E.H.C. and Johnston, P.R. (2009). A polyphasic approach for studying *Colletotrichum*. Fungal Diversity 39: 183-204.
- Campa, A., Giraldez, R. and Ferreira, J.J. (2009). Genetic dissection of the resistance to nine anthracnose races in the common bean differential cultivars MDRK and TU. Theoretical and Applied Genetics 119: 1-11.
- Cannon, P.F. and Simmons, C.M. (2002). Diversity and host preference of leaf endophytic fungi in the Iwokrama Forest Reserve, Guyana. Mycologia 94: 210-220.
- Cannon, P.F., Bridge, P.D. and Monte, E. (2000). Linking the past, present and future of *Colletotrichum* systematics. In: *Colletotrichum*. *Host Specificity, Pathology and Host-Pathogen*

Interaction (eds. D. Prusky, S. Freeman and M.B. Dickman). APS Press, St Paul, Minnesota: 1-20.

- Cannon, P.F., Buddie, A.G. and Bridge, P.D. (2008). The typification of *Colletotrichum gloeo-sporioides*. Mycotaxon 104: 189-204.
- Cano, J., Guarro, J. and Gene, J. (2004). Molecular and morphological identification of *Colletotrichum* species of clinical interest. Journal of Clinical Microbiology 42: 2450-2454.
- Carpinella, M.C., Ferrayoli, C.G. and Palacios, M.S. (2003). Antimycotic activity of the members of Meliaceae. In: *Plant Derived Antimycotics* (eds. M. Rai and D. Mares). Food Products Press, USA: 81-115.
- Carvalho, F.M.S., Leite Junior, R.P. and Ueno, B. (2000). Pathogenic characterization of *Colletotrichum* spp. associated with apple diseases in southern Brazil. Fitopatologia Brasileira 25: 72-78.
- Castro, L.G.M., Silva Lacaz, da C., Guarro, J., Gené, J., Heins-Vaccari, E.M., Freitas Leite, de R.S., Arriagada, G.L.H., Reguera, M.M.O., Ito, E.M., Valente, N.Y.Y. and Nunes, R.S. (2001). Phaeohyphomycotic cyst caused by *Colletotrichum crassipes*. Journal of Clinical Microbiology 39: 2321-2324.
- Cesati, V. de, (1852). Klotzsch, herbarium vivum mycologium, sistens fungorum per totam Germaniam crescentium collectionem perfectam. Century XVII. Berlin. Flora 35: 398.
- Chakrabarti, A., Shivaprakash, M.R., Singh, R., Tarai, B., George, V.K., Fomda, B.A. and Gupta, A. (2008). Fungal endophthalmitis: Fourteen years' experience from a center in India. Retina 28: 1400-1407.
- Chen, C.B. and Dickman, M.B. (2004). Dominant active Rac and dominant negative Rac revert the dominant active Ras phenotype in *Colletotrichum trifolii* by distinct signalling pathways. Molecular Microbiology 51: 1493-1507.
- Chillet, M., Hubert, O., Rives, M.J. and de Lapeyre de Bellaire, L. (2006). Effects of the physiological age of bananas on their susceptibility to wound anthracnose due to *Colletotrichum musae*. Plant Disease 90:1181-1185.
- Chitarra, L.G., Goulart, A.C.P. and Zorato, M.F. (2009). Cottonseeds treatment with fungicides for the control of seedling damping-off pathogens. Revista Brasileira de Sementes 31: 168-176.
- Choudhury, M. and Ahmed, Q.A. (1969). Physiologic specialization of *Colletotrichum corchori*, Ikata and Yoshida, the causal organism of anthracnose of jute (*Corchorus capsularis* L.). Mycopathologia et Mycologia Applicata 38: 161-168.
- Correll, J.C., Rhoads, D.D. and Guerber, J.C. (1993). Examination of mitochondrial DNA restriction fragment length polymorphisms, DNA fingerprints and randomly amplified polymorphic DNA of *Colletotrichum orbiculare*. Phytopathology 83: 1199-1204.
- Costa, M.H.D., Pfenning, L.H. and Pozza, E.A. (2006) Colletotrichum coccodes patógeno de solanáceas no Brasil. Fitopatologia Brasileira 31: 315 (in portugese).

Crouch, J.A. and Beirn, L.A. (2009). Anthracnoses of cereals and grasses. Fungal Diversity 39: 19-44.

- Crouch, J.A., Clarke, B.B. and Hillman, B.I. (2006). Unraveling evolutionary relationships among the divergent lineages of *Colletotrichum* causing anthracnose disease in turfgrass and maize. Phytopathology 96: 46-60.
- Crouch, J.A., Clarke, B.B., White, J.F. and Hillman, B.I. (2009a). Systematic analysis of falcate-spored graminicolous *Colletotrichum* and a description of six new species from warm-season grasses. Mycologia: 101: 717-732.
- Crouch, J.A., Tredway, L.P., Clarke, B.B. and Hillman, B.I. (2009b). Phylogenetic and population genetic divergence correspond with habitat for the pathogen *Colletotrichum cereale* and allied taxa across diverse grass communities. Molecular Ecology 18: 123-135.
- Crouch, J.A., Beirn, L.A., Cortese, L.M., Bonos, S.B. and Clarke, B.B. (2009c). Anthracnose disease of switchgrass caused by the novel fungal species *Colletotrichum navitas*. Mycological Research 113: 1411-1421.
- Crous, P.W. (2009). Taxonomy and phylogeny of the genus *Mycosphaerella* and its anamorphs. Fungal Diversity 38: 1-24.
- Da Silva, M.B., Da Costa, A.S.V., Rufini, J.C.M., Galvao, E.R. and Zambolim, L. (2008). Thermotherapy and prochloraz to control of anthracnose on 'Prata Anã' bananas in post harvest conditions. Summa Phytopathologica 34: 364-365.
- Damm, U., Woudenberg, J.H.C., Cannon, P.F. and Crous, P.W. (2009). *Colletotrichum* species with curved conidia from herbaceous hosts. Fungal Diversity 39: 45-87.
- Daundasekera, W.A.M., Joyce, D.C., Adikaram, N.K.B. and Terry, L.A. (2008). Pathogen-produced ethylene and the *Colletotrichum musae*-banana fruit pathosystem. Australasian Plant Pathology 37: 448-453.
- Davide, L.M.C. and De Souza, E.A. (2009). Pathogenic variability within race 65 of *Colletotrichum lindemuthianum* and its implications for common bean breeding. Crop Breeding and Applied Biotechnology 9: 23-29.
- Demerutis, C., Quiros, L., Martinuz, A., Alvarado, E., Williams, R.N. and Ellis M.A. (2008). Evaluation of an organic treatment for post-harvest control of crown rot of banana. Ecological Engineering 34: 324-327.
- Dhingra, O.D., Lustosa, D.C., Maia, C.B. and Mesquita, J.B. (2003). Seedborne fungal pathogens of Jacaranda (*Dalbergia nigra*) tree. Seed Science and Technology 31: 341-349.
- Dickson, J.G. (1956). *Diseases of Field Crops*. 2nd edn. McGraw-Hill, New York.
- Diederichsen, A., Rozhmina, T.A. and Kudrjavceva, L.P. (2008). Variation patterns within 153 flax (*Linum usitatissimum* L.) genebank accessions based on evaluation for resistance to fusarium wilt,

- anthracnose and pasmo. Plant Genetic Resources: Characterisation and Utilisation 6: 22-32.
- Dingley, J.M. and Gilmour, J.W. (1972). *Colletotrichum acutatum*: Simmds. f. sp. *pinea* associated with "terminal crook" disease of *Pinus* spp. New Zealand Journal of Forestry Science 2: 192-201.
- Du, M., Schardl, C. L., Nuckles, E. M. and Vaillancourt, L. (2005). Using mating-type gene sequences for improved phylogenetic resolution of *Colletotrichum* species complexes. Mycologia 97: 641-658.
- Durand, N., Bertrand, B., Guyot, B., Guiraud, J.P. and Fontana Tachon, A. (2009). Study on the *Coffea arabica/Colletotrichum kahawae* pathosystem: Impact of a biological plant protection product Journal of Plant Diseases and Protection 116: 78-85.
- EPPO (2009). Datasheets on quarantine pests. Glomerella gossypii. EPPO quarantine pest. http://www.eppo.org/QUARANTINE/fungi/Glom erella_gossypii/GLOMGO_ds.pdf, accessed 26 August 2009.
- Fabre, J.V., Julien, J., Parisot, D. and Dron, M. (1995). Analysis of diverse isolates of *Colletotrichum lindemuthianum* infecting common bean using molecular markers. Mycological Research 99: 429-435.
- Fagbola, O. and Abang, M.M. (2004). *Colletotrichum circinans* and *Colletotrichum coccodes* can be distinguished by DGGE analysis of PCR-amplified 18S rDNA fragments. African Journal of Biotechnology 3: 195-198.
- Farr, D.F., Bills, G.F., Chamuris, G.P. and Rossman, A.Y. (1989). Fungi on plants and plant products in the United States. APS Press, Minnesota, USA.
- Farr, D.F., Aime, M.C., Rossman, A.Y. and Palm, M.E. (2006). Species of *Colletotrichum* on *Agavaceae*. Mycological Research 110: 1395-1408.
- Farr, D.F. and Rossman, A.Y. (2009). Fungal Databases, Systematic Mycology and Microbiology Laboratory, ARS, USDA. Retrieved April 15, 2009, from http://nt.ars-grin.gov/fungaldatabases/
- Fernando, T.H.P.S., Jayasinghe, C.K. and Wijesundera, R.L.C. (2000). Factors affecting spore production, germination and viability of *Colletotrichum acutatum* isolates from *Hevea brasiliensis*. Mycological Research 104: 681-685.
- Ferreira, J.B., Abreu, M.S. and Pereira, I.S. (2009). Análise da dinâmica, estrutura de focos e arranjo espacial da mancha manteigosa em campo. Ciência e Agrotecnologia 33: 24-30 (in portugese).
- Fiala, J.V., Tullu, A., Banniza, S., Ginette, S.S. and Vandenberg, A. (2009). Interspecies transfer of resistance to anthracnose in lentil (*Lens culinaris* Medic.). Crop Science 49: 825-830.
- Fisher, P.J. and Petrini, O. (1987). Location of fungal endophytes in tissues of *Suaeda fruticosa*: a preliminary study. Transactions of the British Mycological Society 89: 246-249.
- Ford, R., Banniza, S., Photita, W. and Taylor, P.W.J. (2004). Morphological and molecular discrimination of *Colletotrichum truncatum*

- causing anthracnose on lentil in Canada. Australasian Plant Pathology 33: 559-569.
- Förster, H. and Adaskaveg, J.E. (1999). Identification of subpopulations of *Colletotrichum acutatum* and epidemiology of almond anthracnose in California. Phytopathology 89: 1056-1065.
- Freeman, S. and Rodriguez, R.J. (1995). Differentiation of *Colletotrichum* species responsible for anthracnose of strawberry by arbitrarily primed PCR. Mycological Research 99: 501-504.
- Freeman, S., Minz, D., Jurkevitch, E., Maymon, M. and Shabi, E. (2000). Molecular analyses of *Colleto-trichum* species from almond and other fruits. Phytopathology 90: 608-614.
- Freeman, S., Horowitz, S. and Sharon, A. (2001a). Pathogenic and nonpathogenic lifestyles in *Colletotrichum acutatum* from strawberry and other plants. Phytopathology 91: 986-992.
- Freeman, S., Minz, D., Maymon, M. and Zveibil, A. (2001b). Genetic diversity within *Colletotrichum acutatum sensu* Simmonds. Phytopathology 91: 586-592.
- García-Serrano, M., Laguna, E.A., Rodríguez-Guerra, R. and Simpson, J. (2008). Analysis of the *MAT1-2-1* gene of *Colletotrichum lindemuthianum*. Mycoscience 49: 312-317.
- Gichuru, E.K., Agwanda, C.O., Combes, M.C., Mutitu,
 E.W., Ngugi, E.C.K., Bertrand, B. and Lashermes,
 P. (2008). Identification of molecular markers linked to a gene conferring resistance to coffee berry disease (*Colletotrichum kahawae*) in *Coffea Arabica*. Plant Pathology 57: 1117-1124.
- González, E., Sutton, T.B. and Correll, J.C. (2006). Clarification of the etiology of *Glomerella* leaf spot and bitter rot of apple caused by *Colletotrichum* spp. based on morphology and genetic, molecular and pathogenicity tests. Phytopathology 96: 982-992.
- González, M.M., Rodríguez, R., Zavala, M.E., Jacobo, J.L., Hernández, F., Acosta, J., Martánez, O. and Simpson, J. (1998). Characterization of Mexican isolates of *Colletotrichum lindemuthianum* by using differential cultivars and molecular markers. Phytopathology 88: 292-299.
- Goodwin, P.H. (2001). A molecular weed-mycoherbicide interaction: *Colletotrichum gloeo-sporioides* f. sp. *malvae* and round-leaved mallow, *Malva pusilla*. Canadian Journal of Plant Pathology 23: 28-35.
- Göre, M.E. and Bucak, C. (2007). Geographical and seasonal influences on the distribution of fungal endophytes in *Laurus nobilis*. Forest Pathology 37: 281-288.
- Gossen, B.D., Anderson, K.L. and Buchwaldt, L. (2009). Host specificity of *Colletotrichum truncatum* from lentil. Canadian Journal of Plant Pathology 31: 65-73.
- Götz, M., Nirenberg, H., Krause, S., Wolters, H., Draeger, S., Buchner, A., Lottmann, J., Berg, G. and Smalla, K. (2006). Fungal endophytes in potato roots studied by traditional isolation and cultivation-independent DNA-based methods. FEMS Microbiology Ecology 58: 404-413.

Guerber, J.C. and Correll, J.C. (2001). Characterization of *Glomerella acutata*, the teleomorph of *Colletotrichum acutatum*. Mycologia 93: 216-229.

- Gunnell, P.S. and Gubler, W.D. (1992). Taxonomy and morphology of *Colletotrichum* species pathogenic to strawberry. Mycologia 84: 157-165.
- Heilmann, L.J., Nitzan, N., Johnson, D.A., Pasche, J.S., Doetkott, C. and Gudmestad, N.C. (2006). Genetic variability in the potato pathogen *Colletotrichum coccodes* as determined by amplified fragment length polymorphism and vegetative compatibility group analyses. Phytopathology 96: 1097-1107.
- Higgins, B.B. (1917). A *Colletotrichum* leafspot of turnips. Journal of Agricultural Research 10: 157-161
- Hori, S. (1910). Kaki no Shinbyogai Tansobyo. Engei no Tomo 6: 21-24.
- Hughes, S.J. (1958). Revisiones hyphomycetum aliquot cum appendice de nominibus rejiciendis. Canadian Journal of Botany 36: 727-836.
- Ikata, S. and Yoshida, M. (1940) Studies on the disease of matrush. 1. Blight Special Bulletin, Okayama Agricultural Experiment Station 42: 1-47
- Israeli, Y. and Temkin-Gorodeiski, N. (1997). The mode of infection of *Colletotrichum musae* (Cke. and Massee) causing tip rot of banana fruits in the plantation. Phytoparasitica 5: 167-172.
- Ito, S. (1911). Gloeosporiose of the Japanese persimmon. The Botanical Magazine (Tokyo) 25: 197-202.
- Johnson, D.E. Carris, L.M. and Rogers, J.D. (1997). Morphological and molecular characterization of *Colletotrichum nymphaeae* and *C. nupharicola* sp. nov. on water-lilies (*Nymphaea* and *Nuphar*). Mycological Research 101: 641-649.
- Johnston, P.R. (2000). The importance of phylogeny in understanding host relationships with *Colletotrichum*. In: *Colletotrichum*. Host Specificity, *Pathology and Host-Pathogen Interaction* (eds. D. Prusky, S. Freeman and M.B. Dickman). APS Press, St Paul, Minnesota: 21-28.
- Johnston, P.R. and Jones, D. (1997). Relationships among *Colletotrichum* isolates from fruit-rots assessed using rDNA sequences. Mycologia 89: 420-430.
- Johnston, P.R.; Pennycook, S.R. and M.A. Manning. (2005). Taxonomy of fruit rotting fungal pathogens: What's really out there? New Zealand Plant Protection 58: 42-46.
- Kaewchai, S., Soytong, K. and Hyde, K.D. (2009). Mycofungicides and fungal biofertilizers. Fungal Diversity 38: 25-50.
- Kawaradani, M., Nishimura, A., Moriwaki, J., Tomioka, K., Sato, T., Okada, K. and Nakasone W. (2008). Anthracnose of Perilla caused by *Colletotrichum destructivum*. Japanese Journal of Phytopathology 74: 335-339 (in Japanese).
- Khan, S.R. and Strange, R.N. (1975). Evidence for the role of a fungal stimulant as a determinant of differential susceptibility of jute cultivars to *Colletotrichum corchori*. Physiological Plant Pathology 5: 157-162.

Khan, S.R. and Strange, R.N. (1976). Partial purification and properties of a fungal stimulant from jute affecting the growth in vitro and in vivo of *Colletotrichum corchori*. Physiological Plant Pathology 9: 273-278.

- Kim, W.G.H., Sung, K.K. and Jin, H. (2008). Occurrence of anthracnose on Chinese mallow caused by *Colletotrichum malvarum*. Mycobiology 36: 139-140.
- Kirk, P.M., Cannon, P.F., Minter, D.W. and Stalpers, J.A. (2008). *The Dictionary of the Fungi*, 10th edition. CABI Bioscience. UK.
- Kirkpatrick, T.L., Templeton, G.E., and TeBeest, D.O. (1982). Potential of *Colletotrichum malvarum* for biological control of prickly sida. Plant Disease 66: 323-325.
- Lardner, R., Johnston, P.R., Plummer, K.M. and Pearson, M.N. (1999). Morphological and molecular analysis of *Colletotrichum acutatum sensu lato*. Mycological Research 103: 275-285.
- Latunde-Dada, A.O., O'Connell, R.J., Nash, C., Pring, R.J., Lucas, J.A. and Bailey J.A. (1996). Infection process and identity of the hemibiotrophic anthracnose fungus (*Colletotrichum destructivum*) from cowpea (*Vigna unguiculata*). Mycological Research 100: 1133-1141.
- Latunde-Dada, A.O., Bailey, J.A., Lucas, J.A. (1997). Infection process of *Colletotrichum destructivum* O'Gara from lucerne (*Medicago sativa* L.). European Journal of Plant Pathology 103: 35-41.
- Latunde-Dada, A.O. and Lucas, J.A. (2007). Localized hemibiotrophy in *Colletotrichum*: cytological and molecular taxonomic similarities among *C. destructivum*, *C. linicola* and *C. truncatum*. Plant Pathology 56: 437-447.
- Lee, H.B., Park, J.Y. and Jung, H.S. (2005a). First report of leaf anthracnose caused by *Colletotrichum boninense* on spindle trees.Plant Pathology 54: 254
- Lee, H.B., Park, J.Y. and Jung, H.S. (2005b). Identification, growth and pathogenicity of *Colletotrichum boninense* causing leaf anthracnose on Spindle tree. Plant Pathology Journal 21: 27-32.
- Lees, A.K. and Hilton, A.J. (2003). Black dot (*Colletotrichum coccodes*): An increasingly important disease of potato. Plant Pathology 52: 3-12.
- Liu, B., Wasilwa, L.A., Morelock, T.E., O'Neill, N.R. and Correll, J.C. (2007). Comparison of *Colletotrichum orbiculare* and several allied *Colletotrichum* spp. for mtDNA RFLPs, intron RFLP and sequence variation, vegetative compatibility and host specificity. Phytopathology 97: 1305-1314.
- Liu, G., Kennedy, R., Greenshields, D.L., Peng, G., Forseille, L., Selvaraj, G. and Wei, Y. (2007). Detached and attached *Arabidopsis* leaf assays reveal distinctive defense responses against hemibiotrophic *Colletotrichum* spp. Molecular Plant-Microbe Interactions 20: 1308-1319.
- Lotter, H.C. and Berger, D.K. (2005). Anthracnose of

- lupins in South Africa is caused by *Colletotrichum lupine* var. *setosum*. Australasian Plant Pathology 34: 385-392.
- Lu, G.Z., Cannon, P.F., Reid A. and Simmons, C.M. (2004). Diversity and molecular relationships of endophytic *Colletotrichum* isolates from the Iwokrama Forest Reserve, Guyana. Mycological Research 108: 53-63.
- Lubbe, C.M., Denman, S., Cannon, P.F., Groenewald, J.Z., Lamprecht, S.C. and Crous, P.W. (2004). Characterization of *Colletotrichum gloeosporioides* and similar species associated with anthracnose and dieback of *Proteaceae*. Mycologia 96: 1268-1279.
- MacKenzie, S.J.; Seijo, T.E.; Legard, D.E.; Timmer, P.W. and N.A. Peres (2007). Selection for pathogenicity to strawberry in populations of *Colletotrichum gloeosporioides* from native plants. Phytopathology 97: 1130-1140.
- Maffei, L. (1921). Una malattia delle foglie del "Kaki" dovuta al *Colletotrichum kaki* n. sp. Rivista di Patologia Vegetale 11: 116-118.
- Mahadtanapuk, S., Sanguansermsri, M., Cutler, R.W., Sardsud, V. and Anuntalabhochai, S. (2007). Control of anthracnose caused by *Colletotrichum musae* on *Curcuma alismatifolia* Gagnep. using antagonistic *Bacillus* spp. American Journal of Agricultural and Biological Sciences 2: 54-61.
- Manandhar, J.B., Hargman, G.L. and Sinclair, J.B. (1986). *Colletotrichum destructivum*, the anamorph of *Glomerella glycines*. Phytopathology 76: 282-285.
- Manire, C.A., Rhinehart, H.L., Sutton, D.A., Thompson, E.H., Rinaldi, M.G., Buck, J.D. and Jacobson, E. (2002). Disseminated mycotic infection caused by *Colletotrichum acutatum* in a Kemp's ridley sea turtle (*Lepidochelys kempi*). Journal of Clinical Microbiology 40: 4273-4280.
- Manners, J.M. and He, C. (1997). Recent advances in studies of anthracnose of *Stylosanthes*. IV. Molecular approaches to studies of *Colletotrichum gloeosporioides* causing anthracnose of *Stylosanthes* in Australia. Tropical Grasslands 31: 435-444.
- Marcelino, J., Giordano, R., Gouli, S., Gouli, V., Parker, B.L., Skinner, M., TeBeest, D. and Cesnik, R. (2008). *Colletotrichum acutatum* var. *fioriniae* (teleomorph: *Glomerella acutata* var. *fioriniae* var. nov.) infection of a scale insect. Mycologia 100: 353-374.
- Martin, M.P. and Garcia Figueres, F. (1999). *Colletotrichum acutatum* and *C. gloeosporioides* cause anthracnose on olives. European Journal of Plant Pathology 105: 733-741.
- Martinez-Culebras, P.V., Barrio, E., Garciá, M.D. and Querol. A. (2000). Identification of *Colletotrichum* species responsible for anthracnose of strawberry based on the internal transcribed spacers of the ribosomal region. FEMS Microbiology Letters 189: 97-101.
- Martinez-Culebras, P.V., Querol, A., Suarez-Fernadez, M.B., Garcia-Lopez, M.D. and Barrio, E. (2003). Phylogenetic relationships among *Colletotrichum*

- pathogens of strawberry and design of PCR primers for their identification. Journal of Phytopathology 151: 135-143.
- Mathieson, J.T. and Mangano, V. (1985). Ramulose, a new cotton disease in Paraguay caused by *Colletotrichum gossypii* var. *cephalosporioides*. Summa Phytopathologica 11 115-118.
- Melksham, K.J., Weckert, M.A. and Steel C.C. (2002). An unusual bunch rot of grapes in sub-tropical regions of Australia caused by *Colletotrichum acutatum*. Australasian Plant Pathology 31: 193-194
- Mendes, M.A.S., da Silva, V.L. and Dianese, J.C. (1998). Fungos em plantas no Brasil. Embrapa-SPI/Embrapa-Cenargen, Brasilia: 1-555.
- Mignouna, H.D., Abang, M.M., Onasanya, A. and Asiedu, R. (2002). Identification and application of RAPD markers for anthracnose resistance in water yam (*Dioscorea alata*). Annals of Applied Biology 141: 61-66.
- Mills, P.R., Hodson, A. and Brown, A.E. (1992). Molecular differentiation of *Colletotrichum* isolates infecting tropical fruits. In: *Colletotrichum: Biology, Pathology and Control* (eds. J.A. Bailey and M.J. Jeger). CAB International: Wallingford: 269-288.
- Minuto, A., Gilardi, G., Gullino, M.L. and Garibaldi, A.. (2008). Increasing severity of attacks of *Colletotrichum coccodes* on grafted tomatoes. Acta Horticulturae 789: 101-106.
- Miyazawa, M., Okamura, S., Yamaguchi, M. and Kameoka, H. (2000). Biological stereoselective reduction of 4-methylcyclohexanone and 4-ethylcyclohexanone by anthracnose fungi. Journal of Chemical Technology & Biotechnology 75: 143-146
- Mohanan, C. (2005). Diseases of rattan in nurseries, plantations and natural stands in Kerala, India. Journal of Bamboo and Rattan 4: 151-162.
- Monteiro, J.E.B.A., Sentelhas, P.C., Gleason, M.L., Esker, P.D. and Chiavegato, E.J. (2009). Development of ramulosis disease of cotton under controlled environment and field conditions. Phytopathology 99: 659-665.
- Mordue, J.M. (1971). *Colletotrichum lindemuthianum*. IMI Descriptions of Pathogenic Fungi and Bacteria no. 316. Kew, Surrey: Commonwealth Mycological Institute.
- Moriwaki, J. and Tsukiboshi, T. (2009). *Colletotrichum echinochloae*, a new species on Japanese barnyard millet (*Echinochloa utilis*). Mycoscience 50: 273-280.
- Moriwaki, J., Ohkubo, H., Horie, H., Kasuyama, S. and Sato, T. (1997). Genotypic differences between *Colletotrichum higginsianum* and *C. gloeosporioides* based on arbitrarily primed PCR analysis. Annals of the Phytopathological Society of Japan 63: 395-398.
- Moriwaki, J., Tsukiboshi, T. and Sato, T. (2002). Grouping of *Colletotrichum* species in Japan based on rDNA sequences. Journal of General Plant Pathology 68: 307-320.

Moriwaki, J., Sato, T. and Tsukiboshi, T. (2003). Morphological and molecular characterization of *Colletotrichum boninense* sp. nov. from Japan. Mycoscience 44: 47-53.

- Munaut, F., Hamaide, N. and Maraite, H. (2002). Genomic and pathogenic diversity in *Colletotrichum gloeosporioides* from wild native Mexican *Stylosanthes* spp., and taxonomic implications. Mycological Research 106: 579-593.
- Munda, A., Radisek, S., Sustar-Vozlic, J. and Javornik, B. (2009). Genetic variability of *Colletotrichum lindemuthianum* isolates from Slovenia and resistance of local *Phaseolus vulgaris* germplasm. Journal of Plant Diseases and Protection 116: 23-29.
- Muth, D., Kachlicki, P., Krajewski, P., Przystalski, M. and Stobiecki, M. (2009). Differential metabolic response of narrow leafed lupine (*Lupinus angustifolius*) leaves to infection with *Colletotrichum lupine*. Metabolomics (in press).
- Nakamura, M., Ohzono, M. and Kei Arai, H.S. (2006). Anthracnose of *Sansevieria trifasciata* caused by *Colletotrichum sansevieriae* sp. nov. Journal of General Plant Pathology 72: 253-256.
- Nazriya, N.F., De Costa, D.M. and Azhaar, A.S. (2007). Genomic variations of *Colletotrichum musae* morphotypes infecting banana varieties of Sri Lanka. Proceedings of the Peradeniya University Research Sessions, Sri Lanka 12: 1-2.
- Nirenberg, H.I., Feiler, U. and Hagedorn, G. (2002). Description of *Colletotrichum lupini* comb. nov. in modern terms. Mycologia 94: 307-320.
- Niroshini Gunasinghe, W.K.R. and Karunaratne, A.M. (2009). Interactions of *Colletotrichum musae* and *Lasiodiplodia theobromae* and their biocontrol by *Pantoea agglomerans* and *Flavobacterium* sp. in expression of crown rot of "Embul" banana. BioControl 54: 587-596.
- Nitzan, N., Hazanovsky, M., Tal, M. and Tsror, L. (2002). Vegetative compatibility groups in *Colletotrichum coccodes*, the causal agent of black dot on potato. Phytopathology 92: 827-832.
- Nitzan, N., Lucas, B.S. and Christ, B.J. (2006a). Colonization of rotation crops and weeds by the potato black dot pathogen *Colletotrichum coccodes*. American Journal of Potato Research 83: 503-507.
- Nitzan, N., Tsror, L. and Johnson, D.A. (2006b). Vegetative compatibility groups and aggressiveness of North American isolates of *Colletotrichum coccodes*, the causal agent of potato black dot. Plant Disease 90: 1287-1292.
- Nitzan, N., Cummings, T.F. and Johnson, D.A. (2008). Disease potential of soil- and tuberborne inocula of *Colletotrichum coccodes* and black dot severity on potato. Plant Disease 92: 1497-1502.
- Nuangmek, W., McKenzie, E.H.C. and Lumyong, S. (2008). Endophytic fungi from wild banana (*Musa acuminata* Colla) works against anthracnose disease caused by *Colletotrichum musae*. Research Journal of Microbiology 3: 368-374.

- O'Connell, R., Herbert, C., Sreenivasaprasad, S., Khatib, M., Esquerré-Tugayé, M.T. and Dumas, B. (2004). A novel *Arabidopsis-Colletotrichum* pathosystem for the molecular dissection of plantfungal interactions. Molecular Plant-Microbe Interactions 17: 272-282.
- Oelofse, D., Dubery, I. and Berger, D.K. (2009). Exo-β-1,3-Glucanase from Yeast Inhibits *Colletotrichum lupini* and *Botrytis cinerea* Spore Germination. Journal of Phytopathology 157: 1-6.
- Pavgi, M.S. and Singh, U.P. (1965). Parasitic fungi from India IV. Mycopathologia and Mycologia Applicata 27: 81-88.
- Peck, C.H. (1909) Report of the state botanist, 1908. Bulletin of the New York State Museum 450: 5-220
- Pereira, J.O., Carneiro Vieira, M.L. and Azevedo, J.L. (1999). Endophytic fungi from *Musa acuminata* and their reintroduction into axenic plants. World Journal of Microbiology and Biotechnology 15: 43-46.
- Pereira, I.S., Abreu, M.S., Alves, E. and Ferreira, J.B. (2009). Estudos histopatológicos da interação *Colletotrichum gloeosporioides* cafeeiro. Bragantia 68: 117-123 (In Portugese).
- Pfenning, L.H., Salgado, M. and Lima, C.S. (2007). New concepts in the systematics of *Colletotrichum*. Fitopatologia Brasileira 32(Supl): 20-21.
- Photita W., Lumyong S., Lumyong P. and Hyde K.D. (2001). Endophytic fungi of wild banana (*Musa acuminata*) at Doi Suthep Pui National Park, Thailand. Mycological Research 105: 1508-1513.
- Photita W., Lumyong S., Lumyong P., McKenzie E.H.C. and Hyde K.D. (2004). Are some endophytes of Musa acuminata latent pathogens? Fungal Diversity 16: 131-140.
- Photita, W., Taylor, P.W.J., Ford, R., Hyde, K.D. and Lumyong, S. (2005). Morphological and molecular characterization of *Colletotrichum* species from herbaceous plants in Thailand. Fungal Diversity 18: 117-133.
- Politis, D.J. (1975). The identity and perfect state of Colletotrichum graminicola. Mycologia 67: 56-62
- Prihastuti, H., Cai, L., Chen, H. and Hyde, K.D. (2009). Characterization of *Colletotrichum* species associated with coffee berries in Chiang Mai, Thailand. Fungal Diversity 39: 89-109.
- Pring, R.J., Nash, C., Zakaria, M. and Bailey, J.A. (1995). Infection process and host range of *Colletotrichum capsici*. Physiological and Molecular Plant Pathology 46: 137-152.
- Prusky, D. and Plumbley, R.A. (1992). Quiescent infections of *Colletotrichum* in tropical and subtropical fruits. In: *Colletotrichum: Biology, Pathology and Control* (eds. J.A. Bailey and M.J. Jeger). CAB International: Wallingford: 289-307.
- Rahman, S. and Junaid, M. (2008). Antimicrobial activity of leaf extracts of *Eupatorium triplinerve* Vehl. against some human pathogenic bacteria and phytopathogenic fungi. Bangladesh Journal of Botany 37: 89-92.

- Ramos, A.P., Merali, Z., Talhinhas, P., Sreenivasaprasad, S. and Oliveira, H. (2006). Molecular and morphological characterisation of *Colletotrichum* species involved in citrus anthracnose in Portugal. Bulletin OILB/SROP 29: 317-326.
- Rao, G.P. (2004). Sugarcane Pathology Volume II. Oxford & IBH Publishing Co, UK.
- Rao, G.P., Bergamin Filho, A., Magarey, R.C. and Autrey. L.J.C. (eds) (1999). Sugarcane Pathology: Volume I. Fungal Diseases. Oxford & IBH Publishing Co, UK.
- Rayner, R.W. (1948). Latent infection in *Coffea arabica* L. Nature 4085: 245-246.
- Roberts, P.D., Pernezny, K., Kucharek and T.A. (2001). Anthracnose caused by *Colletotrichum* sp. on pepper. *Journal of University of Florida/Institute of Food and Agricultural Sciences.* 2001 (Available from: http://edis.ifas.ufl.edu/PP104). (Accessed 25/8/2009).
- Rodriguez-Guerra, R., Ramírez-Rueda, M.-T., Cabral-Enciso, M., García-Serrano, M., Lira-Maldonado, Z., Guevara-González, R., González-Chavira, M. and Simpson, J. (2005). Heterothallic mating observed between Mexican isolates of *Glomerella lindemuthiana*. Mycologia 97: 793-803.
- Saccardo, P.A. (1980). Fungorum extra-european pugillus. Michellia 2: 136-149.
- Sanogo, S., Stevenson, R.E. and Pennypacker, S.P. (2003). Appressorium formation and tomato fruit infection by *Colletotrichum coccodes*. Plant Disease 87: 336-340.
- Sanyong, S. and Amarakul, V. (2001). GIS applications for plant diseases distribution on Jack fruit, Bael fruit and Ma Kiang trees in lower northern part of Thailand. In: 22nd Asian Conference on Remote Sensing, 5-9 Septemebr 2001, Singapore.
- Sato. T. (1997). Characters and identication of a plurivorous anthracnose fungus, *Colletotrichum acutatum*. Plant Protection Shikoku 32: 1-19 (in Japanese).
- Sato, T., Muta, T., Imamura, Y., Nojima, H., Moriwaki, J. and Yaguchi, Y. (2005). Anthracnose of Japanese radish caused by *Colletotrichum dematium*. Journal of General Plant Pathology 71: 380-383.
- Sato, T., Yamasaki, F., and Takeya, M. (2009). Common name database of plant diseases in Japan. Plant Protection 63: 587-591 (in Japanese).
- Scheffer, R.P. (1950). Anthracnose leaf spot of crucifers. North Carolina Agricultural Experiment Station Technical Bulletin, 92.
- Selby, A.D. and Manns, T.F. (1909). Studies in diseases of cereals and grasses. Ohio Agricultural Experimental Station Bulletin 203: 187-236.
- Shcolnick, S., Dinoor, A. and Tsror, L. (2007). Additional vegetative compatibility groups in *Colletotrichum coccodes* subpopulations from Europe and Israel. Plant Disease 91: 805-808.
- Shen S., Goodwin P. and Hsiang T. (2001). Hemibiotrophic infection and identity of the fungus, *Colletotrichum destructivum*, causing anthracnose of tobacco. Mycological Research 105: 1340-1347.

- Shenoy, B.D., Jeewon, R., Lam, W.H., Bhat, D.J., Than, P.P., Taylor, P.W.J. and Hyde, K.D. (2007). Morphomolecular characterisation and epitypification of *Colletotrichum capsici*, causal agent of anthracnose in chilli. Fungal Diversity 27: 197-211.
- Sherriff, C., Whelan, M.J., Arnold, G.M., Lafay, J.F., Brygoo, Y. and Bailey, J.A. (1994). Ribosomal DNA sequence analysis reveals new species groupings in the genus *Colletotrichum*. Experimental Mycology 18: 121-138.
- Shimizu, M., Yazawa, S. and Ushijima, Y. (2009). A promising strain of endophytic *Streptomyces* sp. for biological control of cucumber anthracnose. Journal of General Plant Pathology 75: 27-36.
- Shivas, R.G. and Tan, Y.P. (2009). A taxonomic reassessment of *Colletotrichum acutatum*, introducing *C. fioriniae* comb. et stat. nov. and *C. simmondsii* sp. nov. Fungal Diversity 39: 111-122.
- Shivas, R.G., Bathgate, J. and Podger, F.D. (1998). *Colletotrichum xanthorrhoeae* sp. nov. on *Xanthorrhoea* in Western Australia. Mycological Research 102: 280-282.
- Sicard, D., Michalakis, Y., Dron, M. and Neema, C. (1997). Genetic diversity and pathogenic variation of *Colletotrichum lindemuthianum* in three centres of diversity of its host, *Phaseolus vulgaris*. Phytopathology 87: 807-813.
- Silva-Mann, R., Vieira, M.G.G.C., Machado, J.C., Filho, J.R.B., Salgado, K.C.C. and Stevens, M.R. (2005). AFLP markers differentiate *Colletotrichum gossypii* from *C. gossypii* var. *cephalosporioides*. Fitopatologia Brasileira 30: 169-172.
- Silva-Mann, R., Filho, J.R.B., Mann, E.N., Vieira, M.G.G.C., Machado, J.C., Santos, M.F. and Boari, A.J. (2007). Instability of ramulosis reaction of cotton cultivars with respect to protection and registration procedures. Crop Breeding and Applied Biotechnology 7: 367-372.
- Simmonds, J.H. (1968). Type specimens of *Colletotrichum gloeosporioides* var. *minor* and *Colletotrichum acutatum*, Queensland Journal of Agricultural and Animal Science 25: 178A.
- Sivanesan, A. and Hsieh, W.H. (1993). A new ascomycete, *Glomerella septospora* sp. nov. and its coelomycete anamorph, *Colletotrichum taiwanense* sp. nov. Mycological Research 97: 1523-1529.
- Smith, B.J. and Black, L.L. (1990). Morphological, cultural and pathogenic variation among *Colleto-trichum* species isolated from strawberry. Plant Disease 74: 69-76.
- Smith, J.E., Korsten, L. and Aveling, T.A.S. (1999). Infection process of *Colletotrichum dematium* on cowpea stems. Mycological Research 103: 230-234.
- Sobers, E.K. and Plakidas, A.G. (1962). Colletotrichums associated with lily bulbs. Phytopathology 52: 884-887.
- Spilsbury, J.F. (1953). Some fungus diseases of *Digitalis lanata*. Transactions of the British Mycological Society 36: 335-346.

Sprague, R. (1950) *Diseases of cereals and grasses in North America*. The Ronald Press Company, NY.

- Sreenivasaprasad, S. and Talhinhas, P. (2005). Genotypic and phenotypic diversity in *Colletotrichum acutatum*, a cosmopolitan pathogen causing anthracnose on a wide range of hosts. Molecular Plant Pathology 6: 361-378.
- Sreenivasaprasad, S., Mills, P.R., Meehan, B.M. and Brown, A.E. (1996). Phylogeny and systematics of 18 *Colletotrichum* species based on ribosomal DNA spacer sequences. Genome 39: 499-512.
- Stevens, F.L. (1931). The ascigerous stage of *Colletotrichum lagenarium* induced by ultraviolet radiation. Mycologia 23: 134-139.
- Stevens, F.L. and True, E.Y. (1919). Black spot of onion sets. Agricultural Experiment Station Bulletin. 220: 507-532.
- Sun, H. and Zhang, J.Z. (2009). *Colletotrichum destructivum* from cowpea infecting *Arabidopsis thaliana* and its identity to *C. higginsianum*. European Journal of Plant Pathology, 125: 459-469
- Sutton, B.C. (1962). *Colletotrichum dematium* (Pers. ex Fr.) Grove and *C. trichellum* (Fr. ex Fr.) Duke. Transactions of the British Mycological Society 45: 222-232.
- Sutton, B.C. (1965) Studies on the taxonomy of Colletotrichum Cda with especial reference to C. graminicola (Ces.) Wilson. University of London, London.
- Sutton, B.C. (1966) Development of fruitifications in *Colletotrichum graminicola* (Ces.) Wils. and related species. Canadian Journal of Botany 44: 887-897.
- Sutton, B.C. (1968). The appressoria of *Colletotrichum graminicola* and *C. falcatum*. Canadian Journal of Botany 46: 873-876.
- Sutton, B.C. (1980). *The Coelomycetes*. Commonwealth Mycological Institute, Kew, London.
- Sutton, B.C. (1992). The genus *Glomerella* and its anamorph *Colletotrichum*. In: *Colletotrichum: Biology, Pathology and Control* (eds. J.A. Bailey and M.J. Jeger). CAB International: Wallingford: 1-26.
- Talhinhas, P., Sreenivasaprasad, S., Neves-Martin, J. and Oliveira, H. (2005). Molecular and phenotypic analyses reveal association of diverse *Colletotrichum acutatum* groups and a low level of *C. gloeosporioides* with olive anthracnose. Applied and Environmental Microbiology 71: 2987-2998.
- Tank, D.C., Beardsley, P.M., Kelchner, S.A. and Olmstead, R.G. (2006). Review of the systematics of *Scrophulariaceae* s.l. and their current disposition. Australian Systematic Botany 19: 289-307.
- Than, P.P., Shivas, R.G., Jeewon, R., Pongsupasamit, S., Marney, T.S., Taylor, P.W.J. and Hyde, K.D. (2008a). Epitypification and phylogeny of *Colletotrichum acutatum* J.H. Simmonds. Fungal Diversity 28: 97-108.

- Than, P.P., Jeewon, R., Hyde, K.D., Pongsupasamit, S., Mongkolporn, O. and Taylor, P.W.J. (2008b). Characterization and pathogenicity of *Colletotrichum* species associated with anthracnose on chilli (*Capsicum* spp) in Thailand. Plant Pathology 57: 562-572.
- Than P.P., Prihastuti H., Phoulivong S., Taylor P.W.J. and Hyde K.D. (2008c). Chilli anthracnose disease caused by *Colletotrichum* species. Journal of Zhejiang University: Science B 9: 764-778.
- The Phytopathological Society of Japan (Ed.) (2000). Common names of plant diseases in Japan. Japan Plant Protection Association, Tokyo
- The Phytopathological Society of Japan (ed.) (2009).

 The supplement of the Common names of plant diseases in Japan. http://www.ppsj.org/pdf/misctsuiroku090602.pdf
- Thomas, G.J., Sweetingham, M.W., Yang, H.A. and Speijers, J. (2008). Effect of temperature on growth of *Colletotrichum lupini* and on anthracnose infection and resistance in lupins. Australasian Plant Pathology 37: 35-39.
- Timmer, L.W. and Brown, G.E. (2000). Biology and control of anthracnose diseases of citrus. In: *Colletotrichum. Host Specificity, Pathology and Host-Pathogen Interaction* (eds. D. Prusky, S. Freeman and M.B. Dickman). APS Press, St Paul, Minnesota: 300-316.
- Tomioka, K., Sato, T. and Koganezawa, H. (2001). Anthracnose of *Nemesia strumosa* caused by *Colletotrichum fuscum*. Journal of General Plant Pathology 67: 111–115.
- Tomioka, K., Morawaki, J. and Sato, T. (2008). Anthracnose of *Polygonatum falcatum* caused by *Colletotrichum dematium*, Journal of General Plant Pathology 74: 402-404.
- Tosi, L., Buonaurio, R. and Cappelli, C. (2004). Occurrence of anthracnose caused by *Colletotrichum malvarum* on *Althaea officinalis* in Italy. Plant Disease 88: 425.
- Tsror, L. and Johnson, D.A. (2000). Colletotrichum coccodes on potato. In: Colletotrichum. Host Specificity, Pathology and Host-Pathogen Interaction (eds. D. Prusky, S. Freeman and M.B. Dickman). APS Press, St Paul, Minnesota: 362-373.
- Tunali, B., Berner, D.K. and Dubin, H. (2008). First report of leaf spot caused by *Colletotrichum* cf. *linicola* on field bindweed in Turkey. Plant Disease 92: 316.
- Van Der Vossen, H.A.M., and Walyaro, D.J. (2009).

 Additional evidence for oligogenic inheritance of durable host resistance to coffee berry disease (*Colletotrichum kahawae*) in arabica coffee (*Coffea arabica* L.) Euphytica 165: 105-111.

- Vargas, E. and Gonzalez, L.C. (1972). La mancha mantecosa del café causada por *Colletotrichum* spp. Turrialba 22: 129-135.
- Varzea, V.M.P., Rodrigues Junior, C.J. and Lewis, B.G. (2002). Distinguishing characteristics and vegetative compatibility of *Colletotrichum kahawae* in comparison with other related species from coffee. Plant Pathology 51: 202-207.
- Vaillancourt, L.J. and Hanau, R.M. (1992) Genetic and morphological comparisons of *Glomerella* (*Colletotrichum*) isolates from maize and from sorghum. Experimental Mycology 16:219-229.
- Vinnere, O., Fatehi, J., Wright, S.A.I. and Gerhardson, B. (2002). The causal agent of anthracnose of *Rhododendron* in Sweden and Latvia. Mycological Research 106: 60-69.
- Waller, J.M. and Bridge, P.D. (2000). Recent advantages in understanding *Colletotrichum* diseases of some tropical perennial crops. In: *Colletotrichum. Host Specificity, Pathology and Host-Pathogen Interaction* (eds. D. Prusky, S. Freeman and M.B. Dickman). APS Press, St Paul, Minnesota: 337-345.
- Walker, J., Nikandrow, A. and Millar, G.D. (1991). Species of *Colletotrichum* on *Xanthium* (Asteraceae) with comments on some taxonomic and nomenclatural problems in *Colletotrichum*. Mycological Research 95: 1175-1193.
- Waller, J.M., Bridge, P.D., Black, R. and Hakiza, G. (1993). Characterization of the coffee berry disease pathogen, *Colletotrichum kahawae* sp. nov. Mycological Research 97: 989-994.
- Warwar, V. and Dickman, M.B. (1996). Effects of calcium and calmodulin on spore germination and appressorium development in *Colletotrichum trifolii*. Applied and Environmental Microbiology 62: 74-79.
- Watanabe, T. and Tamura, M. (1952). Studies on the perfect stage of the causal fungus of the anthracnose of cucumber. Annals of the Phytopathological Society of Japan 16: 137-140.
- Watson, A.K., Gressel, J., Sharon, A. and Dinoor, A. (2000). Colletotrichum strains for weed control. In: Colletotrichum. Host Specificity, Pathology and Host-Pathogen Interaction (eds. D. Prusky, S. Freeman and M.B. Dickman). APS Press, St Paul, Minnesota: 245-265.
- Weir, B.S. and Johnston P.R. (in press). Characterisation and neotypification of *Gloeosporium kaki* Hori as *Colletotrichum horii* nom. nov. Mycotaxon (in press).
- Wharton, P.S. and Diéguez-Uribeondo, J. (2004). The biology of *Colletotrichum acutatum*. Anales del Jardín Botánico de Madrid 61: 3-22.
- Whitelaw-Weckert, M.A., Curtin, S.J., Huang, R., Steel, C.C., Blanchard C.L. and Roffey, P.E. (2007)

- Phylogenetic relationships and pathogenicity of *Colletotrichum acutatum* isolates from grape in subtropical Australia. Plant Pathology 56: 448–463.
- Wilson, G.W. (1914). The identity of the anthracnose of grasses. Phytopathology 4: 106-112.
- Xiao, C.L., MacKenzie, S.J. and Legard, D.E. (2004). Genetic and pathogenic analyses of *Colletotrichum gloeosporioides* isolates from strawberry and noncultivated hosts. Phytopathology 94: 446-453.
- Yakoby, N., Beno-Moualem, D., Kobiler, I. and Prusky, D. (2002). The analysis of fruit protection mechanisms provided by reduced-pathogenicity mutants of *Colletotrichum gloeosporioides* obtained by restriction enzyme mediated integration. Phytopathology 92: 1196-1201.
- Yamamoto, J., Sato, T. and Tomioka, K. (1999).

 Occurrence of ripe rot of grape (*Vitis vinifera* L.) caused by *Colletotrichum acutatum* Simmonds ex Simmonds. Annals of the Phytopathological Society Japan 65: 83-86.
- Yang, S., Gao, M., Xu, C., Gao, J., Deshpande, S., Lin, S., Roe, B.A. and Zhu, H. (2008). Alfalfa benefits from *Medicago truncatula*: The RCT1 gene from *M. truncatula* confers broad-spectrum resistance to anthracnose in alfalfa. Proceedings of the National Academy of Sciences of the United States of America 105: 12164-12169.
- Yang, Y.L., Liu, Z.Y., Cai, L., Hyde, K.D., Yu, Z.N. and McKenzie, E.H.C. (2009). Colletotrichum anthracnose of Amaryllidaceae. Fungal Diversity 39: 123-149.
- Yang, Z.H. and Dickman, M.B. (1999). *Colletotrichum trifolii* mutants disrupted in the catalytic subunit of camp-dependent protein kinase are nonpathogenic. Molecular Plant-Microbe Interactions 12: 430-439.
- Yoshida S. and Shirata A. (1999). Survival of *Colletotrichum dematium* in soil and infected mulberry leaves. Plant Disease 83: 465-468.
- Zeiders, K.E. (1987). Leaf spot of Indiangrass caused by Colletotrichum caudatum. Plant Disease 71: 348-
- Zeru, A., Assefa, F., Adugna, G. and Hindorf, H. (2009).
 Occurrence of fungal diseases of *Coffea arabica*L. in montane rainforests of Ethiopia. Journal of Applied Botany and Food Quality 82: 148-151.
- Zhang, P.Y., Wang, J.C., Liu, S.H. and Chen, K.S. (2009). A novel burdock fructooligosaccharide induces changes in the croduction of calicylates, activates defence enzymes and induces systemic acquired resistance to colletotrichum orbiculare in cucumber seedlings. Journal of Phytopathology 157: 201-207.