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Nematology

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<https://doi.org/10.1163/156854109X456862>

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# A phylogenetic tree of nematodes based on about 1200 full-length small subunit ribosomal DNA sequences

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Received: 8 December 2008; revised: 30 April 2009

Accepted for publication: 1 May 2009

**Summary** – As a result of the scarcity of informative morphological and anatomical characters, nematode systematics have always been volatile. Differences in the appreciation of these characters have resulted in numerous classifications and this greatly confuses scientific communication. An advantage of the use of molecular data is that it allows for an enormous expansion of the number of characters. Here we present a phylogenetic tree based on 1215 small subunit ribosomal DNA sequences (*ca* 1700 bp each) covering a wide range of nematode taxa. Of the 19 nematode orders mentioned by De Ley *et al.* (2006) 15 are represented here. Compared with Holterman *et al.* (2006) the number of taxa analysed has been tripled. This did not result in major changes in the clade subdivision of the phylum, although a decrease in the number of well supported nodes was observed. Especially at the family level and below we observed a considerable congruence between morphology and ribosomal DNA-based nematode systematics and, in case of discrepancies, morphological or anatomical support could be found for the alternative grouping in most instances. The extensiveness of convergent evolution is one of the most striking phenomena observed in the phylogenetic tree presented here – it is hard to find a morphological, ecological or biological characteristic that has not arisen at least twice during nematode evolution. Convergent evolution appears to be an important additional explanation for the seemingly persistent volatility of nematode systematics.

**Keywords** – convergent evolution, DNA barcoding, molecular, nematode evolution, phylogeny.

As in many other major invertebrate clades, the early members of the phylum Nematoda are thought to have arisen in marine habitats during the so-called Cambrian Explosion (600–550 million years ago). However, fossil records from nematodes are extremely rare and no hard evidence is available for this statement. So far, the oldest nematode fossil is *Palaeonema phyticum* that was found in association with *Aglaophyton major*, a free-sporing land plant of the early Devonian (416–396 million years ago; Poinar *et al.*, 2008). More, and older, fossil records are available from water bears or tardigrades, a group of animals relatively closely related to nematodes (*e.g.*, Dunn *et al.*, 2008). Fossils from marine tardigrades show that the earliest members of the Tardigrada arose in the

mid-Cambrian (for review, see Labandeira, 2005). This observation could be considered as indirect support for the marine origin of the phylum Nematoda during the Cambrian.

Compared with related phyla, such as the Priapulida, the Kinorhyncha, and its closest relative, the Nematomorpha (horsehair worms; *ca* 320 described species), the phylum Nematoda can be seen as a success story. Nematodes are speciose and are present in huge numbers in virtually all marine, freshwater and terrestrial environments. Analysis of large EST data sets recently reconfirmed the placement of the phylum Nematoda within the superphylum Ecdysozoa (Dunn *et al.*, 2008), a major animal clade proposed by Aguinaldo *et al.* (1997) that unites all moulting

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ing animals.

Being relatively small and colourless, most nematodes are inconspicuous organisms. They are best known to the general public as the causal agents of a number of human, animal and plant diseases. For instance, the pinworm, *Enterobius vermicularis*, is a ubiquitous parasite of man (particularly children), and the number of people infected is estimated at about 400 million (Lukeš *et al.*, 2005). *Ascaris lumbricoides* is a major intestinal parasite with more than a billion people estimated to be infected by this worm (with the highest incidence in areas with poor sanitation; Lukeš *et al.*, 2005). Other high impact nematode parasites of humans include *Necator americanus* and *Ancylostoma duodenale* (hookworms), *Wucheria bancrofti* and *Brugia malayi* (the causal agents of filariasis). Animal parasitism by nematodes is widespread and affects livestock such as sheep (*Haemonchus contortus*), pigs (*Ascaris suum*) and chickens (*Ascaridia galli*). Plant-parasitic nematodes are also ubiquitous with cyst (members of the genera *Heterodera* and *Globodera*), root-knot (representatives of the genus *Meloidogyne*) and lesion nematodes (*Pratylenchus* spp.) seriously affecting economically important crops such as soybean, potato and tomato. Plant-parasitic nematodes cause worldwide losses of about \$80 billion annually (Agrios, 2005). It should be noted that parasites mostly constitute a (small) minority within nematode assemblages, the majority being non-parasitic organisms that play essential roles in terrestrial and sediment food webs.

For decades people have tried to design a phylogenetic framework for the members of this old and highly diverse phylum. Micoletzky (1922) supposed that differences in the morphology of the stoma ('mouth') could be used to determine relationships between major groups. However, the subdivision of the phylum into five stoma-based families appeared to be unstable. The bipartite system proposed by Chitwood and Chitwood (1933) has been very influential. They proposed a subdivision of the phylum into the Aphasmidia and the Phasmidia. In 1958, Chitwood decided to change the names into Adenophorea ('gland bearers') and Secernentea ('secretors'), respectively, and for decades nearly all nematologists have adhered to this deep subdivision. Lorenzen (1981) was the first to propose a phylogenetic scheme for nematodes based on cladistic principles. He also indicated that the establishment of a more definitive cladogram was severely hindered by the scarcity of informative characters. Over the last two decades this impediment has been bypassed by the availability of a virtually unlimited number of DNA

characters for any taxon. Blaxter *et al.* (1998) (53 taxa) and Aleshin *et al.* (1998) (19 taxa) were among the first to exploit the potential of ribosomal DNA sequence data to resolve phylogenetic relationships among nematodes. Holterman *et al.* (2006) presented a subdivision of the phylum Nematoda into 12 clades based on a series of mostly well supported bifurcations in the backbone of the tree (339 taxa).

Over the last few years, an impressive number of molecular data-based papers have been published that focused on specific taxonomic groups. In the basal part of the nematode tree, the resolution among Dorylaimida, remarkably poor with SSU rDNA data, was substantially improved by using the 5' region of the LSU rDNA (Holterman *et al.*, 2008a). The under-representation of marine nematodes in the phylogenetic overview presented so far was, to some extent, lifted by SSU rDNA-based papers from Meldal *et al.* (2007) and Holterman *et al.* (2008a). Nadler *et al.* (2007) greatly increased our insight into the relationships among animal-parasitic nematodes by analysing 113 SSU rDNA sequences. Bert *et al.* (2008) investigated relationships within the suborder Tylenchina (covering four infraorders, namely Panagrolaimomorpha, Cephalobomorpha, Drilonematomorpha and Tylenchomorpha) by combining SSU rDNA data and morphological information on the female gonoduct. Holterman *et al.* (2009) concentrated on the Tylenchomorpha (mainly insect and plant parasites) and revealed phylogenetic relationships among some of the major plant parasites based on 116 SSU rDNA sequences.

In the current overview paper we made a selection of ca 1200 (nearly) full-length SSU rDNA sequences from representatives throughout the phylum Nematoda. Although, to the best of our knowledge, it is the most species rich and diverse nematode tree based on molecular data published so far, it is biased towards terrestrial nematodes living in moderate climate zones. It is, even at the ordinal level, still incomplete (no representatives from Muspiceida, Marimermithida, Benthimermithida and Rhaptothyreida), and it is based on only a single gene. Nevertheless, it provides numerous insights into the evolutionary relationships within the Nematoda in all its trophic and ecological diversity. Apart from this scientific merit, this framework can be used to (quantitatively) detect single targets in highly complex DNA backgrounds, such as specific plant parasites in a soil community. Currently, we are field testing SSU rDNA-based nematode community analysis in soil at family level and preliminary results will be published in the near future.

## Materials and methods

### SPECIMEN COLLECTION

Nematodes were collected from various habitats throughout The Netherlands, and extracted from the soil using standard techniques. Prior to DNA extraction, individual nematodes were identified using a light microscope (Zeiss Axioscope) equipped with DIC optics. A CCD camera (CoolSnap, RS Photometrics) was used to take a series of digital images from each nematode. For the nomenclature of taxonomic groups we essentially conformed to the systematics proposed by De Ley *et al.* (2006), except for the order Tylenchida for which we adhered to the systematics proposed by Siddiqi (2000).

### DNA EXTRACTION, AMPLIFICATION AND SEQUENCING

DNA extraction, amplification and sequencing were performed as detailed in Holterman *et al.* (2006). Newly generated SSU rDNA sequences ( $n = 109$ ) were deposited at GenBank under the accession numbers FJ040398-FJ040506.

### SEQUENCE ALIGNMENT

Newly generated nematode SSU rDNA sequences were supplemented with publicly available sequences (for a full list, see Supplementary Table S1 in the online edition of this journal, which can be accessed *via* <http://www.brill.nl/nemy>). The outgroup consisted of other Ecdysozoa ( $n = 10$ ). The SSU rDNA sequences were aligned using the ClustalW algorithm as implemented in BioEdit 5.0.9 (Hall, 1999) and manually improved using secondary structure information from *Loricea foveata* ([http://bioinformatics.psb.ugent.be/webtools/rRNA/secmodel/Lfov\\_SSU.html](http://bioinformatics.psb.ugent.be/webtools/rRNA/secmodel/Lfov_SSU.html)), in accordance with Ben Ali *et al.* (1999). Thirteen length variable regions (LVRs) were present within the alignment (length and positioning are given in Figure S1, parts 1, 2, that can be found in the online edition of this journal, which can be accessed *via* <http://www.brill.nl/nemy>). The software package mfold (Washington University, St. Louis, MO, USA; <http://mfold.bioinfo.rpi.edu/cgi-bin/dna-form1.cgi>) was used to predict the most likely secondary structure, and this information was used to align sequences with one or more LVRs. The final alignment consisted of 1225 SSU rDNA sequences and contained 2967 aligned positions (including gaps). In a second analysis, the 13 LVRs as defined in Supplementary

Figure S1 were removed to the study the effect on the phylogenetic analysis.

### PHYLOGENETIC ANALYSIS

The program Modeltest v.3.06 (Posada & Crandall, 1998) selected the GTR + I +  $\Gamma$  model as the best fitting model using both the likelihood ratio test and Akaike Information Criterion. The phylogenetic tree was constructed with a fast maximum likelihood method. The SSU rDNA alignment was analysed at a distant server (<http://phylobench.vital-it.ch/raxml-bb/index.php>) running the program RAXML-VI-HPC v.4.0.0 (Randomized Axelerated Maximum Likelihood for High Performance Computing (Stamatakis, 2006)). A GTR model with invariable sites and gamma distribution was used and the dataset was divided in a stem and loop partition. The tree presented is the result of two independent runs. The values of the shape parameter  $\alpha$  for the stem partition were estimated by RAXML at 0.626539 and 0.624500 for run 1 and 2, respectively. For the loop partition,  $\alpha$  was 0.638105 and 0.638054. The proportion of invariable sites was 0.037952 and 0.037696 (stem), and 0.041264 and 0.041248 (loop) for run 1 and 2, respectively. Two hundred bootstraps were performed.

In addition, the alignment without LVRs was analysed using RAXML and the tree with the best likelihood is presented in Figure S1 in the online edition of this journal, which can be accessed *via* <http://www.brill.nl/nemy>. The values of the shape parameter  $\alpha$  for the stem partition were estimated by RAXML at 0.635468 and 0.642951 for run 1 and 2, respectively. For the loop partition  $\alpha$  was 0.657278 and 0.658856. The proportion of invariable sites was 0.032238 and 0.032557 (stem), and 0.032853 and 0.032846 (loop) for run 1 and 2, respectively. Two hundred bootstraps were performed.

## Results and discussion

### BACKBONE

Previously, Holterman *et al.* (2006) proposed a subdivision of the phylum Nematoda into 12 clades on the basis of *ca* 360 SSU rDNA sequences that gave rise to a series of mostly well supported bifurcations in the backbone of the phylogenetic tree. The increase in the number of sequences analysed to 1215, the use of a fast maximum likelihood method (instead of a more time consum-



**Table 1.** Support values for deep phylogenetic relationships among nematode clades based on (nearly) full length SSU rDNA sequences.

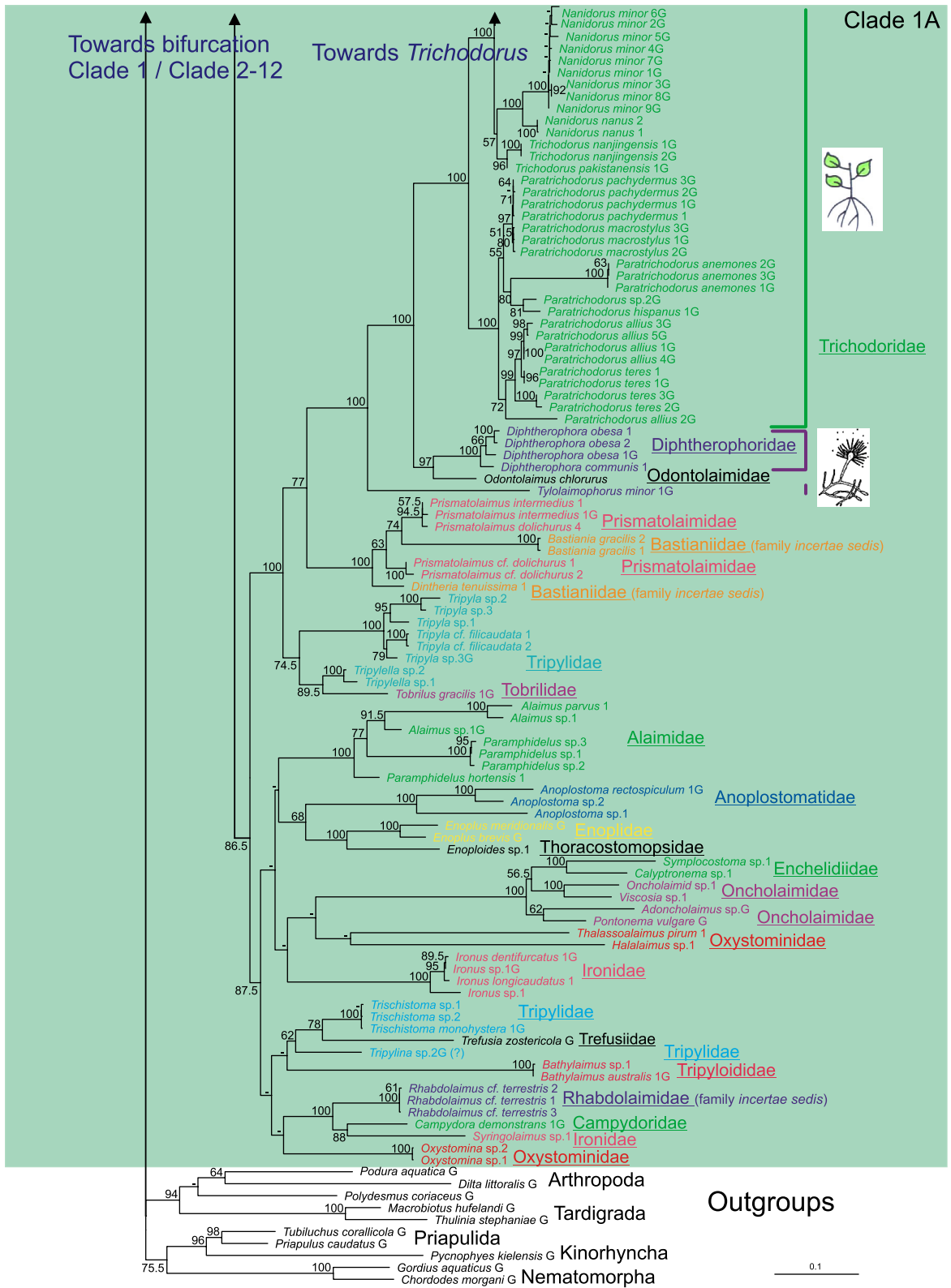
	Bifurcation between	Bootstrap value (%) (this paper) (>65% is robust)	Posterior probability (Holterman <i>et al.</i> , 2006) (>0.95 is robust)
Clade 1	Clade 2-12	100	1.00
Clade 2	Clade 3-12	63	0.81
Monoposthiidae	Haliplectidae and Desmodoridae, and Clade 3-12	100	– (no Monoposthiidae included)
Haliplectidae and Desmodoridae	Clade 3-12	–	– (no Haliplectidae included)
Clade 3	Clade 4-12	–	1.00
Clade 4	Desmoscolecidae and Clade 5ABC-12	64	1.00
Desmoscolecidae	Clade 5ABC-12	68.5	– (no Desmoscolecidae included)
Clade 5ABC	Clade 6-12	–	0.71
Clade 6	Clade 7-12	100	1.00
Clade 7	Clade 8-12	97	1.00
Clade 8	Clade 9-12	97	1.00
Clade 9	Clade 10-12	–	0.64
Clade 10	Clade 11-12	–	1.00
Clade 11	Clade 12 (and Clade 10A)	70	0.95 (Clade 10A not included)
Clade 12	Clade 10A	–	–

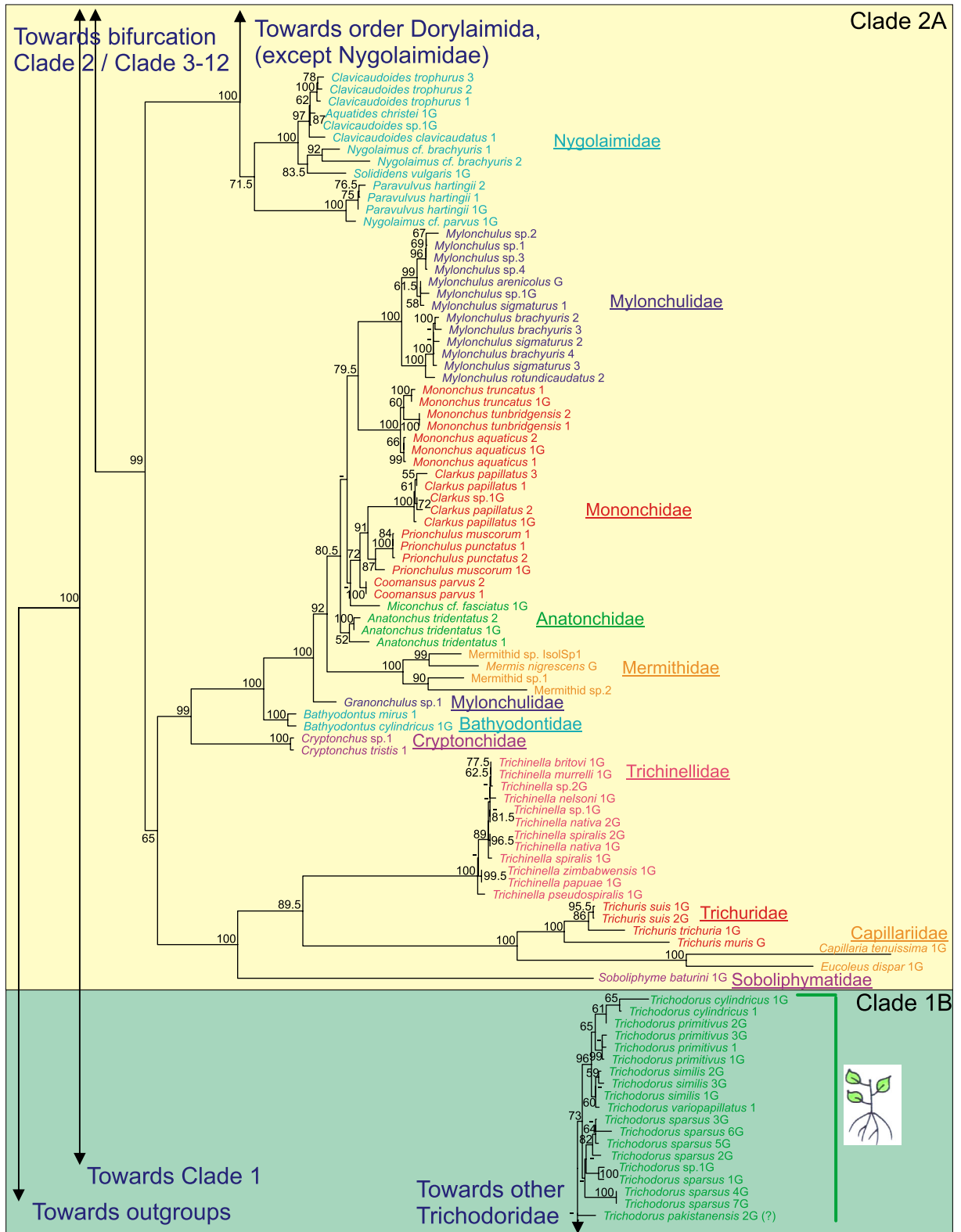
ing Bayesian analysis) and the inclusion of the gamma parameter in this analysis resulted in a phylogenetic tree with an overall topology that resembles the previous one. However, the support values for the backbone tend to be lower (Table 1). This can be explained, at least partially, by the inclusion of families that had not been considered previously or taxa that previously were not taken into consideration because of their strong destabilising effect on the tree ('rogue taxa', such as *Prodesmodora circulata* and *Desmoscolex* sp. in Holterman *et al.*, 2008b). Monoposthiidae (one representative), Desmoscololecidae (one representative) and a cluster with Haliplectidae (one representative) and Desmodoridae (subfamily Prodesmodorinae only) appeared as new, distinct, groups but their positioning in the overall tree remains unclear. The splitting of Clade 10 into two parts could be seen as a main difference between Holterman *et al.* (2006) and the current analysis but as there is no support for this alternative topology, it will not be discussed further. The use of subclades in this paper (e.g., 8A, 8B) is merely for the aid of discussion and no phylogenetic meaning is attached to them (indeed some are not even monophyletic).

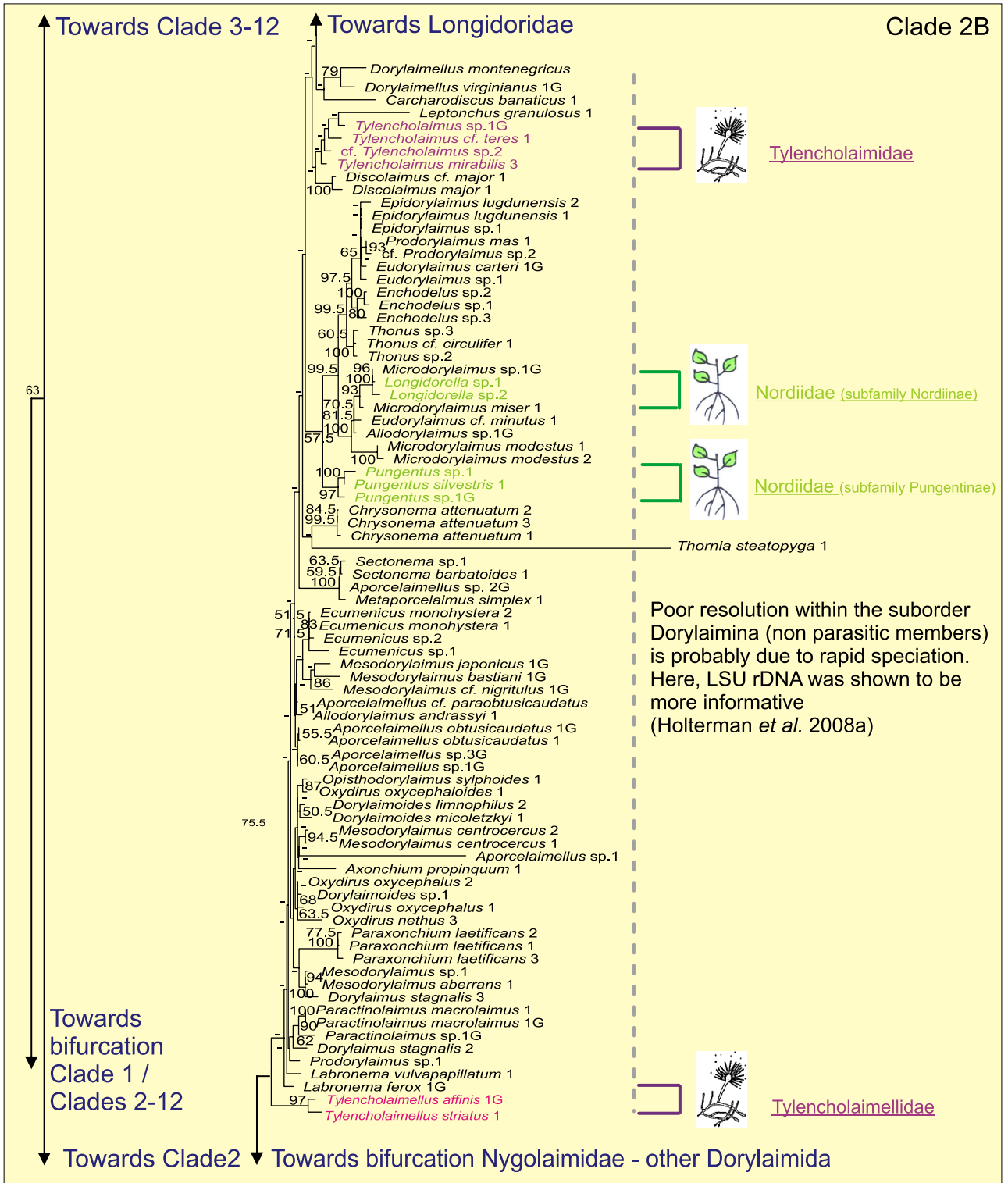
In the second analysis, we removed 13 LVRs as defined in Figure S1 (part 1) in the online edition of this journal, which can be accessed via <http://www.brill.nl/nemy>.

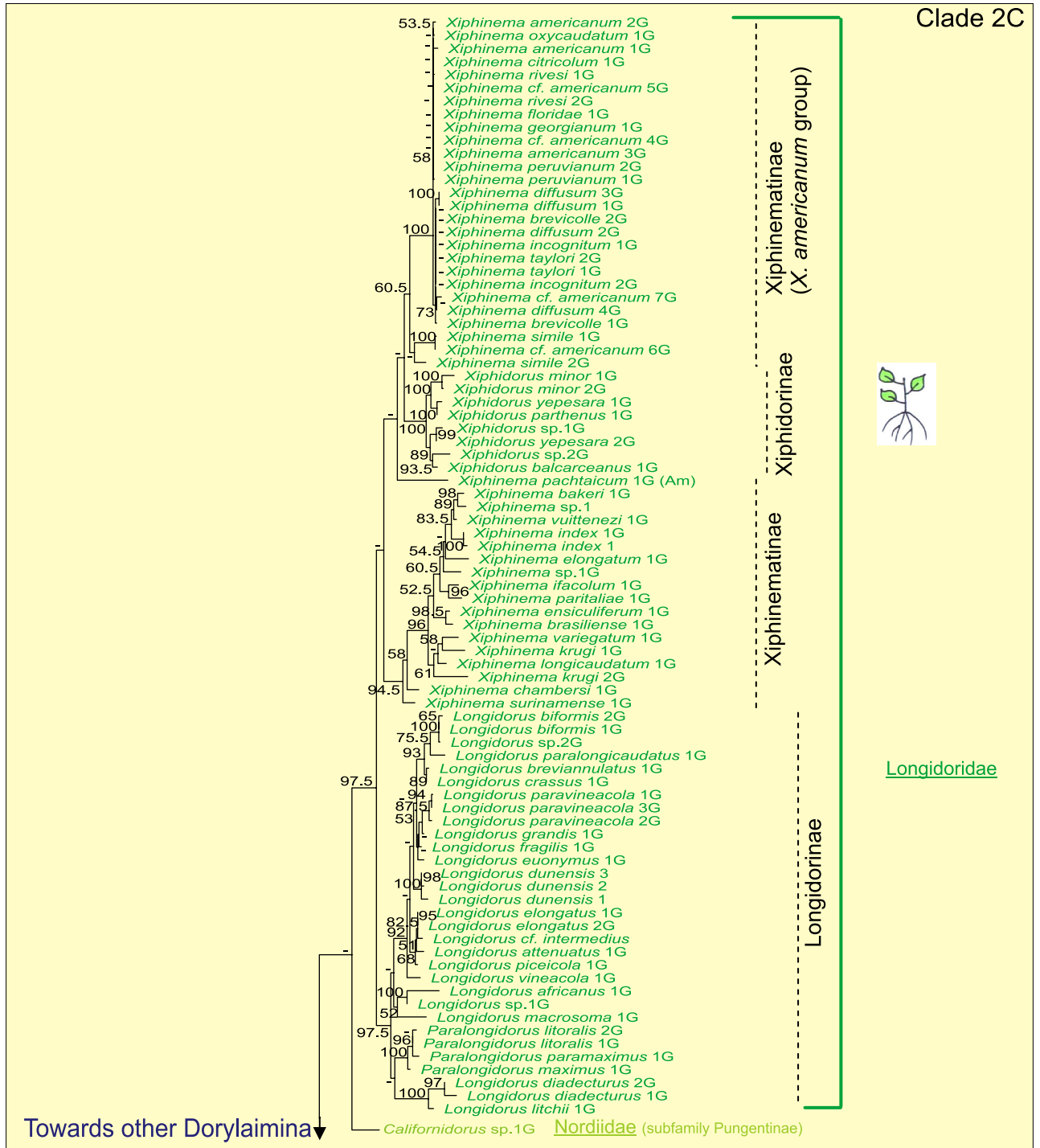
The positioning of the LVRs in a SSU rRNA secondary structure model is given in Figure S1 (part 2) in the online edition of this journal, which can be accessed via <http://www.brill.nl/nemy>. Maximum likelihood analysis resulted in a phylogenetic tree with a topology similar to that presented in Figure S1 (part 3) in the online edition of this journal, which can be accessed via <http://www.brill.nl/nemy>, although the support values for the major bifurcations tended to be lower. It is noted that some families with a poorly supported position in the original tree, such as Panagrolaimidae and Aphelenchidae, were repositioned in the analysis without LVRs. However, the alternative positioning received no significant bootstrap support.

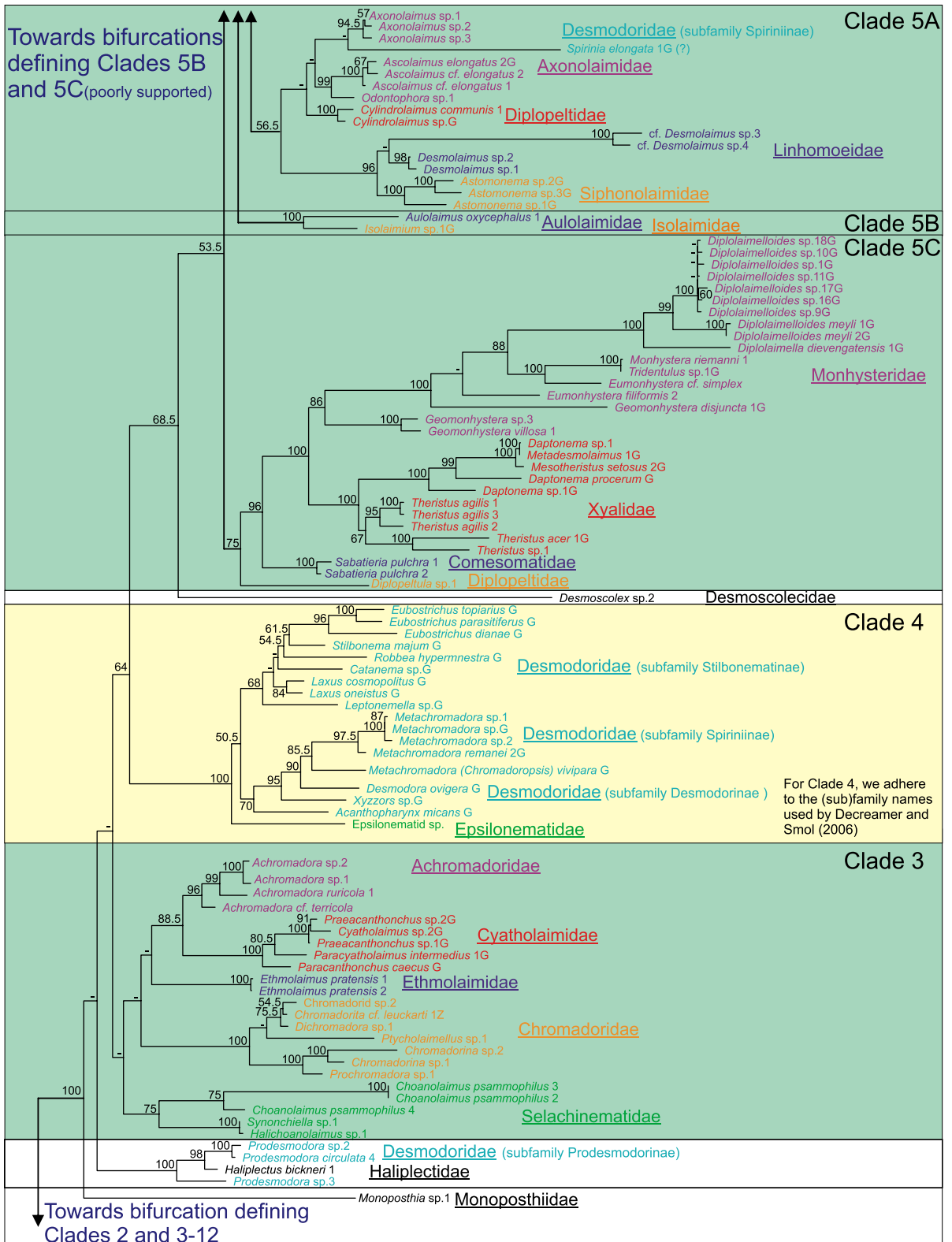
**Fig. 1.** Best likelihood phylogenetic tree recovered by RAxML on the basis of SSU rDNA sequences. Alternating pale yellow and green backgrounds define clades. White backgrounds indicate an uncertain clade position. For the nomenclature of taxonomic groups we essentially conformed to De Ley *et al.* (2006); deviations from this system are indicated in the tree. Family names are underlined, and family members have identical colours. Only bootstrap values >50% are given next to the nodes. A 'G' behind a nematode name indicates that the sequence was acquired from GenBank.

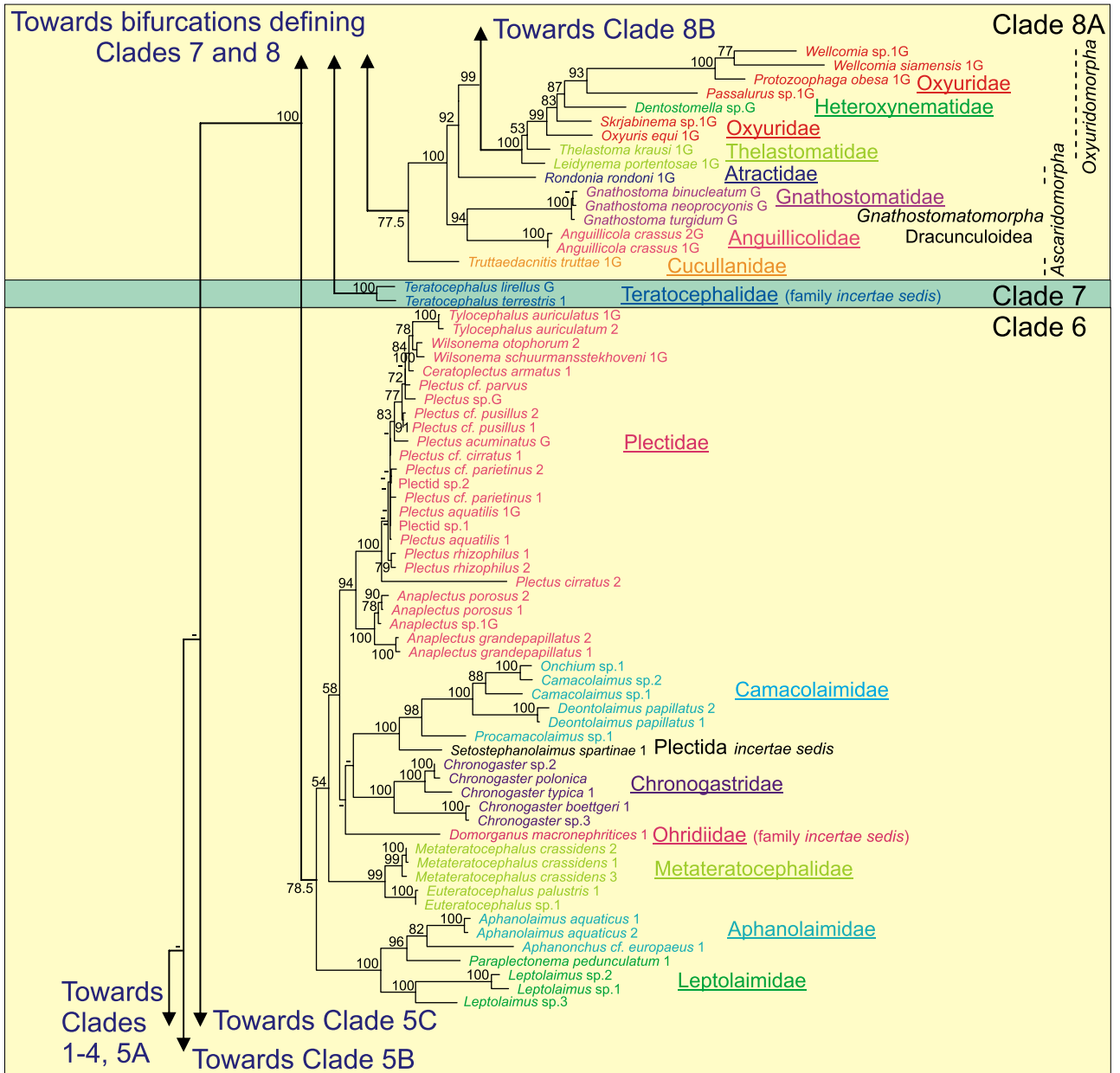




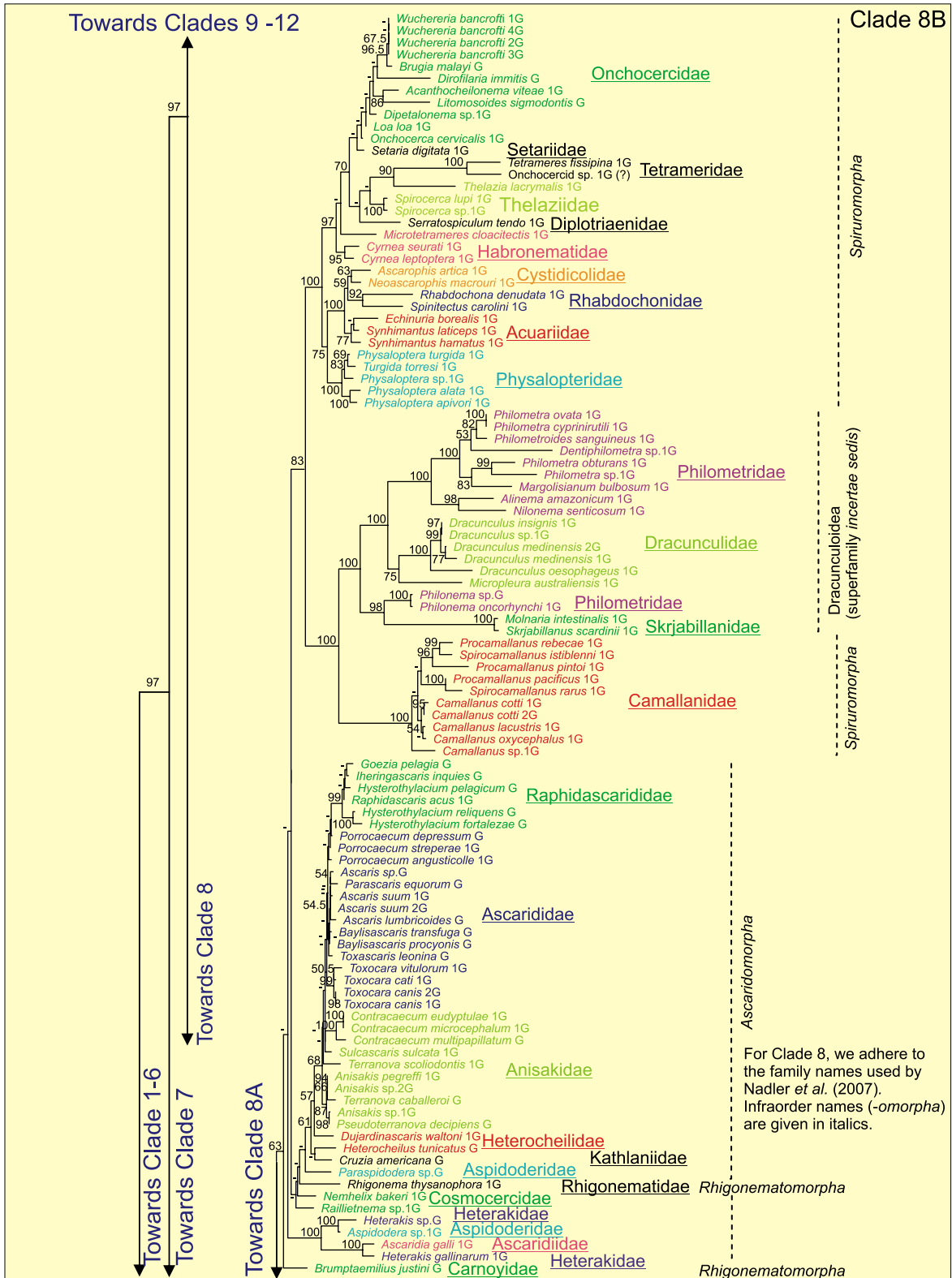


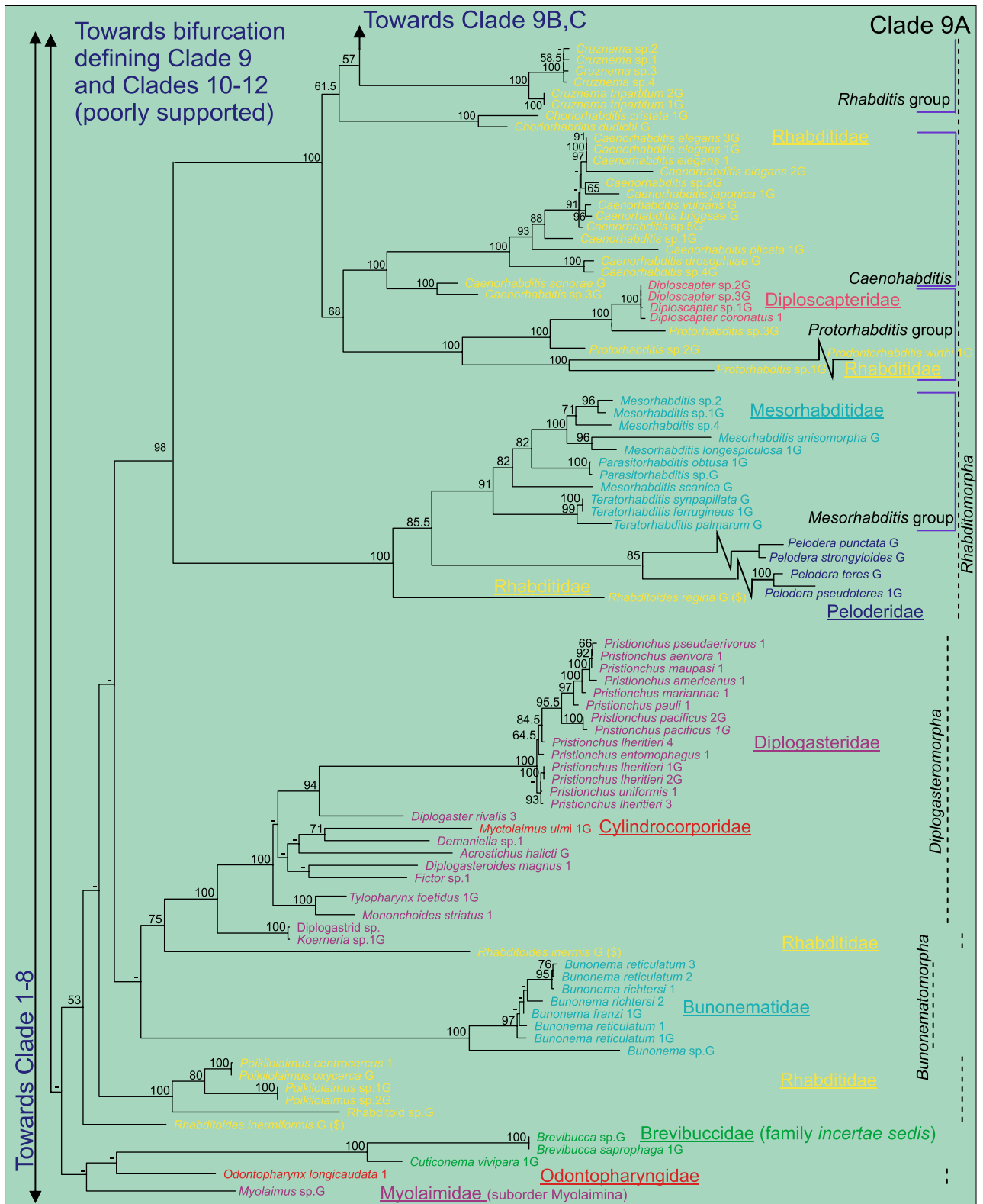


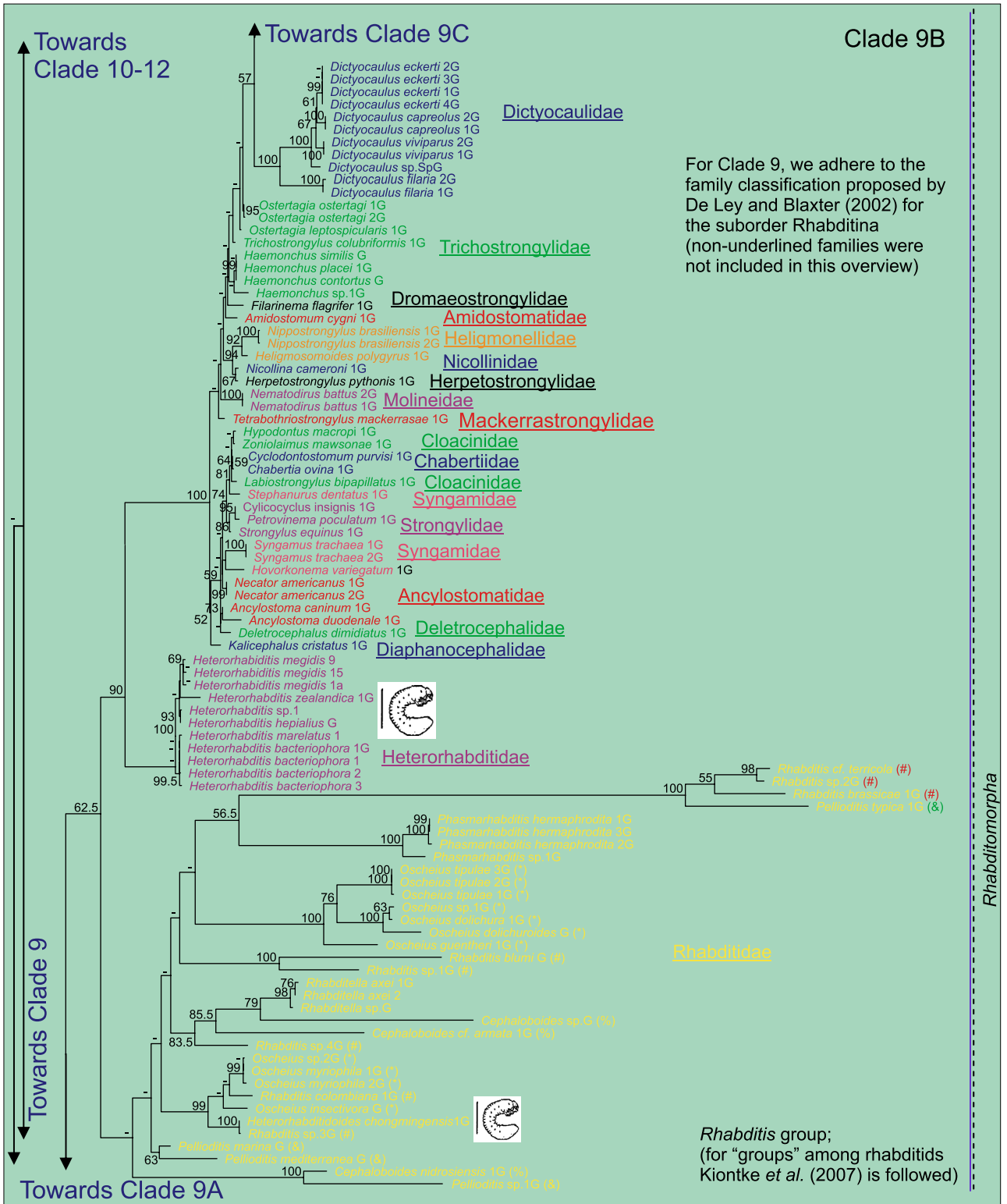


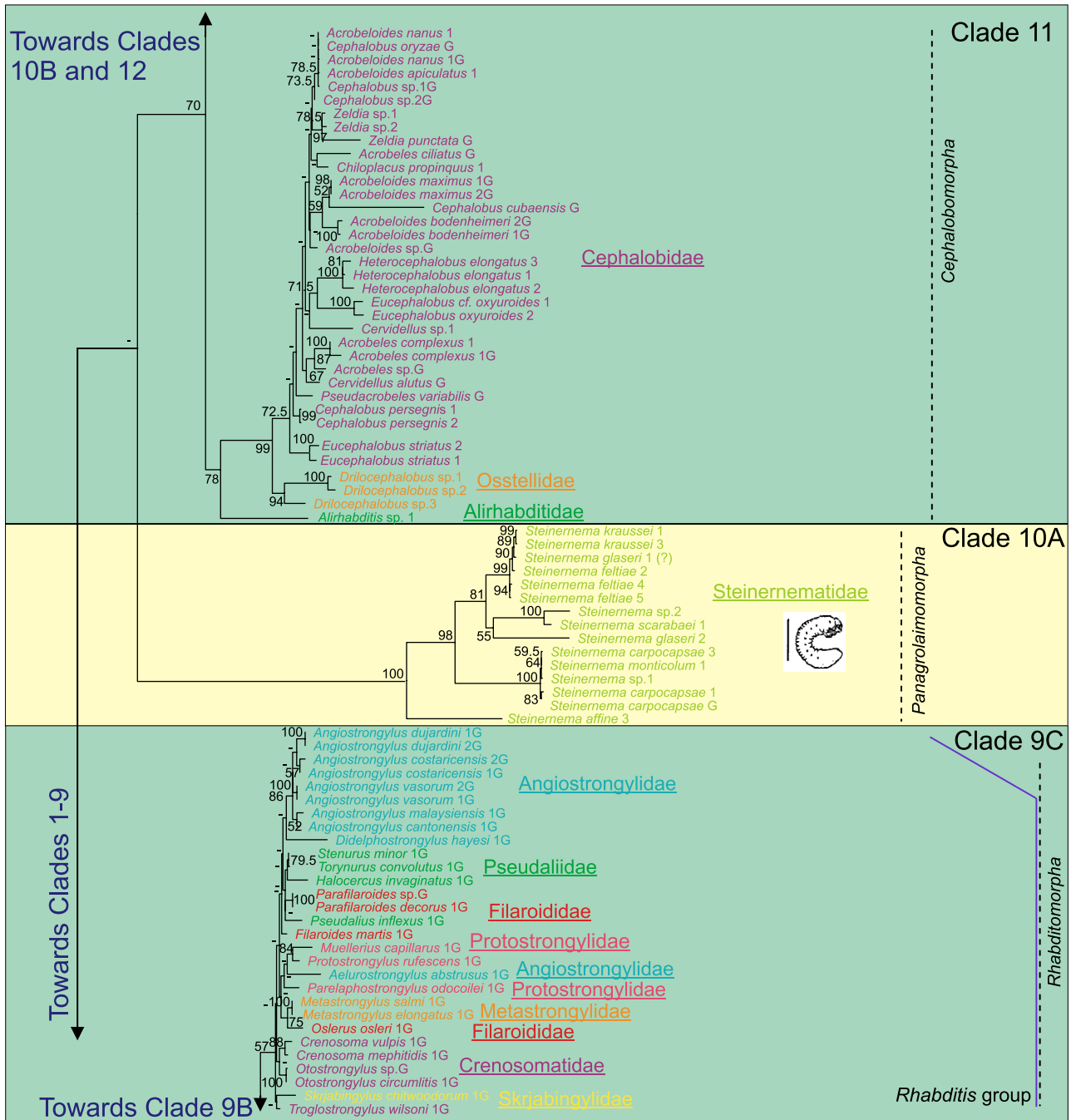


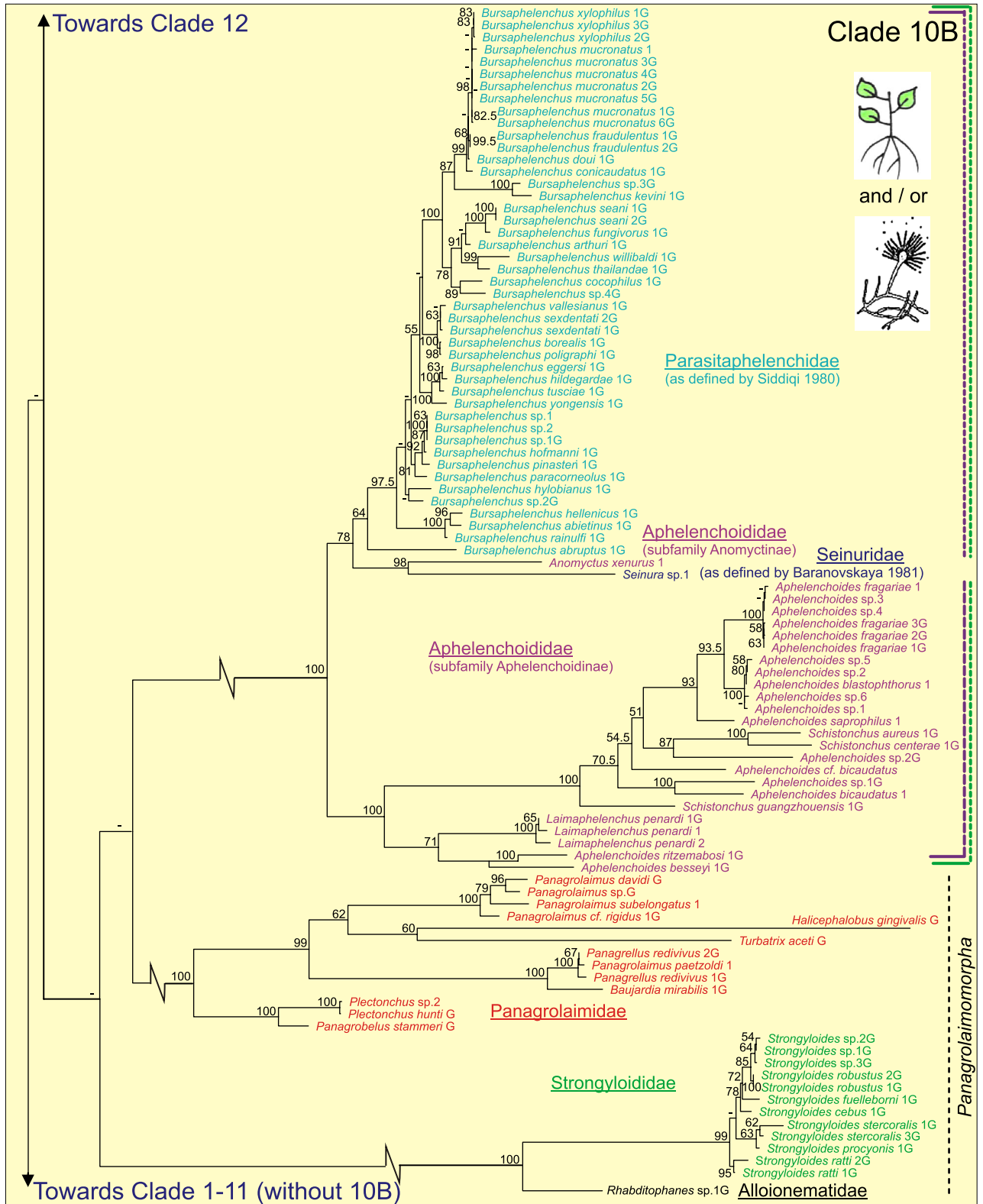


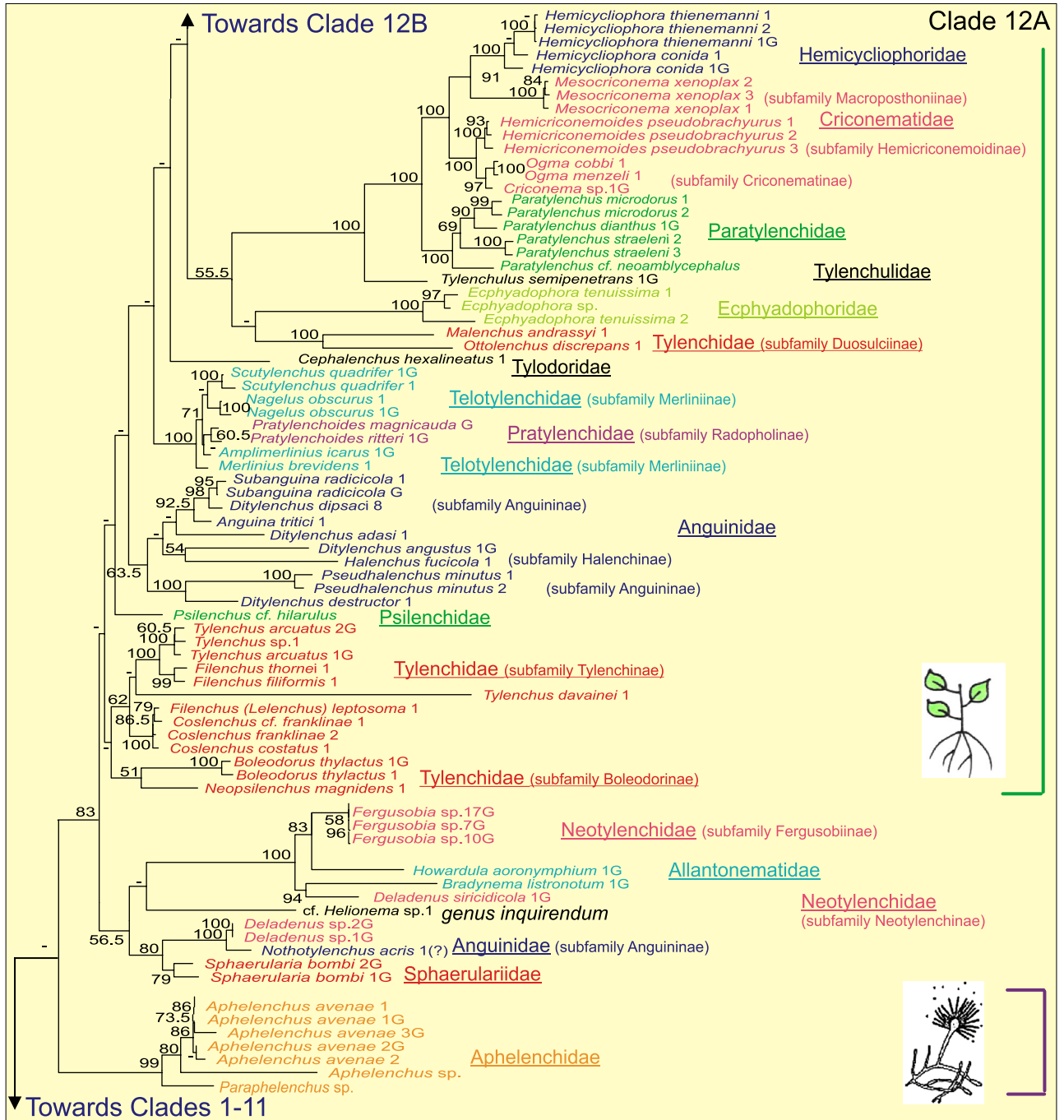








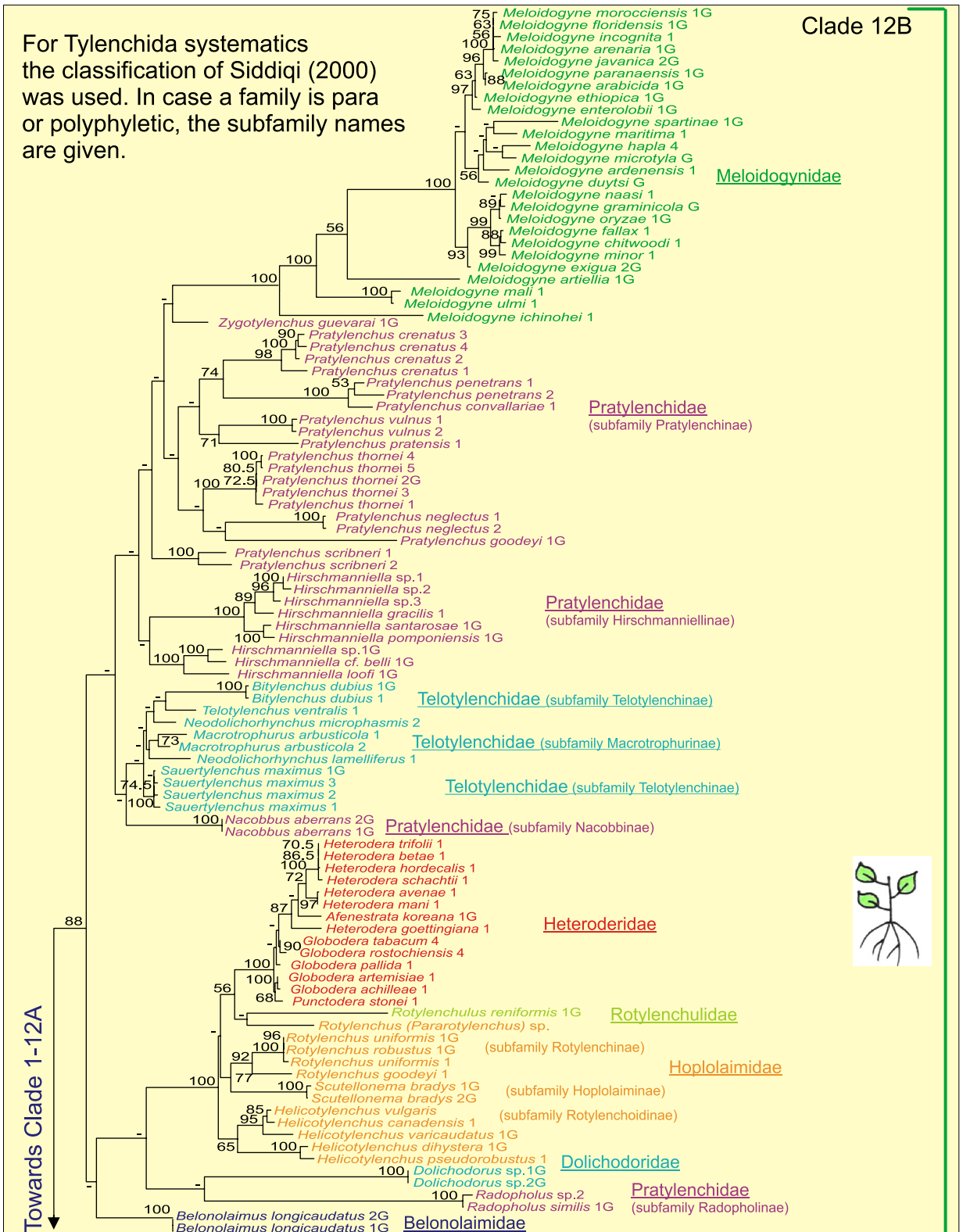






Clade 12B

For Tylenchida systematics the classification of Siddiqi (2000) was used. In case a family is para or polyphyletic, the subfamily names are given.





## CLADE 1 – ENOPLIDA AND TRIPLONCHIDA

Since the bootstrap support for the second major node between Clade 2 (subclass Dorylaimia) and Clades 3-12 (63%) is just below the threshold value (65%), we cannot further confirm the basal positioning of Clade 1 as proposed by Holterman *et al.* (2006) on the basis of SSU rDNA sequences and embryological and morphological arguments. Within this clade there is a well-supported split between the Enoplida and the Triplonchida. In a single case, representatives of the family Tripylidae (usually placed within the order Triplonchida) are found in both orders. *Trischistoma* and *Tripylina* reside among the Enoplida, whereas *Tripyla* and *Tripylella* members are positioned among the Triplonchida. Similarities in the morphology of the labial region, the amphids, the digestive system and the male copulatory apparatus support the sister relationship between *Trischistoma* and *Trefusia*. For similar reasons Brzeski and Winiszewska-Ślipińska (1993) proposed the removal of the genus *Trischistoma* (but not *Tripylella*) from the Tripylidae.

Within the suborder Diphtherophorina, members of the Diphtherophoridae are positioned at the base of the Trichodoridae. The subdivision of the Trichodoridae (most sequences were generated by Dr Konstantina Boutsika in the framework of her Ph.D. thesis: see Boutsika, 2002) is remarkably similar to the grouping of (*Para*)*Trichodorus* on the basis of the cuticle fine structure as presented by Karanastasi *et al.* (2001); Type 1, represented by *T. cylindricus*, *T. primitivus* and *T. similis* (here supplemented with *T. variopapillatus*), constitutes a single cluster (see Clade 1B). This also holds for Type 2, represented here by *Nanidorus* (*Paratrichodorus*) *minor*, *N. nanus*, *T. nanjingensis* and *T. pakistanensis*. The SSU rDNA based analyses suggests a sister relationship between Type 1 and Type 2 (*Para*)*Trichodorus* groups. A third group (Type 3), defined on the basis of the cuticle structure, which included *Paratrichodorus pachydermus*, *P. anemones*, *P. hispanus*, and *P. teres*, was confirmed by molecular data. The current analysis suggests that *P. macrostylus* and *P. allius* also belong to this group and points at a sister relationship between Types 1 and 2 on the one hand, and Type 3 on the other.

The close relationship between the Bastianiidae (family *incertae sedis*) and the Prismatolaimidae (order Triplonchida, suborder Tobrilina) confirms earlier results from Coomans and Raski (1988) who pointed out a remarkable number of similarities between *Prismatolaimus* and *Bastiania* in the structure of sensilla, amphid, cardia, supplements, spicules and gubernaculum. The genera

*Prismatolaimus*, *Bastiania* and *Dintheria*, which constitute a well supported subclade sister to the Diphtherophorina, share several characteristics of the male copulatory apparatus, including spicule and gubernaculum shape, arrangement of spicule protractors and supplement shape (Holovachov, 2006).

Within the order Enoplida, a well-supported cluster was formed by members of the genera *Rhabdolaimus* (Rhabdolaimidae, family *incertae sedis*), *Campydora* (Campydoridae) and *Syringolaimus* (Ironidae). Previously, Chitwood (1951) united the genera *Rhabdolaimus* and *Syringolaimus* in the subfamily Rhabdolaiminae, having three minute, outwardly directed, teeth as a common characteristic. *Campydora* species have only one, elongated, dorsal tooth (Andrássy, 2007). The three genera are united by a strongly sclerotised lumen in the swollen, bulb-like, posterior end of the pharynx (Chitwood, 1951; Andrássy, 2007).

## CLADE 2 – SUBCLASS DORYLAIMIA

Most of the orders residing within the subclass Dorylaimia (*i.e.*, Dorylaimida, Mononchida, Isolaimida, Diocotophymatida, Muspiceida, Marimermithida, Mermithida and Trichinellida) are represented in Clade 2. Muspiceida and Marimermithida (marine nematodes) are missing as no ribosomal DNA data are available. Isolaimida, here represented by a single SSU rDNA sequence from *Isolaimium* sp., was placed in Clade 5B. This is an elaboration on the results presented by Mullin *et al.* (2005) proposing an exclusion of the Isolaimida from the Dorylaimia. The positioning of *Isolaimium* and *Aulolaimus* in Clade 5B is discussed in detail in Holterman *et al.* (2008b).

As compared with Holterman *et al.* (2008a), one family is added, namely the Capillariidae. A sister relationship was observed between the two members of the Capillariidae, *Capillaria tenuissima* and *Eucoleus dispar* (both isolated from the digestive system of the common buzzard (Honisch & Krone, 2008)) and representatives of the Trichuridae. This is a confirmation of morphology-based taxonomy which places both families within the superfamily Trichinelloidea.

Within the Longidoridae, three subfamilies are indicated in Figure 1 (Clade 2C), *viz.*, Xiphinematinae, Xiphidorinae and Longidorinae (see Hunt, 1993). For the *Xiphinema americanum*-group we used the species list from Lamberti *et al.* (2000). Most of the Longidoridae data shown here were generated by and discussed in Neilson *et al.* (2004), and the data set was supple-

mented by unpublished GenBank sequences from Dr S.S. Lazarova and co-workers. For the status and the positioning of the genus *Xiphidorus* we refer to Oliveira *et al.* (2004). Originally, the atypical genus *Californidorus* was placed in a separate subfamily (Californidorinae) under the Longidoridae. Mainly based on the number of pharyngeal glands (five instead of three in the longidorids), Jairajpuri (1982) decided to place this genus under the Nordiidae. For further discussion on the Dorylaimida we refer to Holterman *et al.* (2008a).

#### CLADES 3 TO 7 – BASAL CHROMADORIA

As compared with Holterman *et al.* (2008b), the inclusion of SSU rDNA sequences from additional taxa resulted in a re-positioning of the Selachinematidae. This family is usually placed within the suborder Chromadorina and, in the current analysis, it is indeed placed within Clade 3, although without bootstrap support.

Most remarkable is the highly polyphyletic nature of the Desmodoridae. Within this family, six subfamilies are distinguished, *viz.*, Desmodorinae (predominantly marine), Spiriniinae (marine), Prodesmodorinae (limno-terrestrial), Stilbonematinae (marine), Pseudonchinae (marine) and Molgolaiminae (marine). Only the first four are represented in the current SSU rDNA-based tree. Within Clade 4, the Stilbonematinae appear as a single monophyletic group, whereas the Desmodorinae and most of the Spiriniinae (exception: *Spirinia elongata* among Axonolaimidae) also form a single cluster. The positioning of the genus *Prodesmodora* together with *Haliplectus bickneri* sister to Clades 3-12 (no support) questions whether this single-genus subfamily should indeed reside in the family Desmodoridae.

The clustering of representatives of the Linhomoeidae and Siphonolaimidae fits well in current nematode systematics as both reside within the superfamily Siphonolaimoidea. For further discussion of subclades 5A, B and C and the polyphyletic nature of the Monhysterida and Araeolaimida see Holterman *et al.* (2008b).

*Domorganus macronephritices* (Ohridiidae – family *incertae sedis*) was placed amidst the Plectida families in Clade 6, but the relationship with other Plectida families remains unclear.

#### CLADE 8 – BASAL RHABDITIDA

Clade 8 comprises the Rhabditida ('Secernentea') superfamily Dracunculoidea, and the infraorders Spiruromorpha, Ascaridomorpha, Oxyuridomorpha, Gnathosto-

matomorpha and Rhigonematomorpha. Because the use of infraorders for zooparasitic nematodes is relatively new, their representation is indicated in Figure 1 (Clade 8A, B). The superfamily Dracunculoidea (except *Anguillicola crassus*) is robustly placed within the Spiruromorpha. Within Spiruromorpha *sensu lato* a subdivision is observed between the Dracunculoidea and the Camallanidae on the one hand, and the other Spiruromorpha on the other. The Rhigonematomorpha are represented by two species only, belonging to different families. Both rhigonematid rDNA sequences were associated with Ascaridomorpha representatives (but without bootstrap support). Two families, Atractidae and Cucullanidae, were placed outside the Ascaridomorpha but, as they are each represented by one species only, there is no basis for firm conclusions with regard to their systematic position. Most SSU rDNA sequences in Clade 8 were presented in Nadler *et al.* (2007). There is no essential difference between the topology presented here and the trees included in this paper and for an excellent further discussion we refer to Nadler *et al.* (2007).

#### CLADE 9 – RHABDITIDAE AND RELATED FAMILIES

The species-rich Clade 9 includes the infraorders Bunonematomorpha, Diplogasteromorpha and Rhabditomorpha. Most deep relationships within Clade 9 are unresolved, except for a major bifurcation in the Rhabditomorpha between the superfamily Mesorhabditoidea (= Mesorhabditidae and Peloderidae) on the one hand and the Rhabditoidea (= Rhabditidae and Diploscapteridae) and the Strongyloidea (Heterorhabditidae and animal parasite families) on the other. Apart from animal-parasitic and insect-parasitic nematodes, more loose associations with arthropods and gastropods are found within the superfamilies Rhabditoidea and Mesorhabditoidea. These associations involve either the facilitation of transport (phoresy) or necromeny. Necromeny implies that dauer juveniles only start feeding and develop into adults after the death of its host (apparently without promoting this) upon the colonisation of the insect cadaver by saprophytic bacteria.

Within the *Rhabditis* group, as defined by Kiontke *et al.* (2007), and apart from the family Heterorhabditidae, another member of the group of entomopathogenic nematodes (EPN) was found in Eastern China, namely *Heterorhabditidoides chongmingensis* (Zhang *et al.*, 2008). It is noteworthy that *Rhabditis* sp. 3, with a SSU rDNA sequence almost identical to that of *H. chongmingensis*, was also collected from China. Two close relatives of

this novel EPN, *Rhabditis (Oscheius) colombiana* and *Oscheius tipulae*, are associated with the subterranean burrower bug (*Cyrtomenus bergi*; Stock *et al.*, 2005) and with leatherjackets (*Tipula paludosa*; Sudhaus, 1993). Members of the genus *Caenorhabditis* are not normal soil inhabitants, being found only in nutrient rich environments such as compost heaps or in association with arthropods or gastropods (for overview see Kiontke & Sudhaus, 2006). A similar phenomenon was described for a member of the Mesorhabditidae: originally *Teratorhabditis synpallata* was found in cow dung from Bali (Indonesia), but was recently described in association with the red palm weevil (*Rhynchophorus ferrugineus*; see Kanzaki *et al.*, 2008). A related species, *Parasitorhabditis platidontus*, was described as a parasite of *Cyclocephala signaticollis* (Reboredo & Camino, 2000). Occasionally, true parasitic interactions occur among Rhabditoidea and Mesorhabditoidea (e.g., *Phasmarhabditis hermaphrodita* on slugs and *H. chongmingensis* on insects). Hence, loose associations between members of the Rhabditomorpha and arthropods and gastropods are widespread and not confined to one or two groups.

Rhabditidae genera within the *Rhabditis*-group are often poly- and/or paraphyletic (e.g., *Rhabditis*, *Pellioiditis*, *Cephaloboides* and *Oscheius* – each indicated with a distinct symbol in Figure 1). In part this can be explained by the unstable state of Rhabditidae systematics. For instance, *Pellioiditis marina* is occasionally placed within the genus *Rhabditis* (Derycke *et al.*, 2008), *Oscheius tipulae* is sometimes placed within the genus *Rhabditis* (Sudhaus, 1993), whereas *Rhabditis colombiana* is also known as *Oscheius colombiana* (Stock *et al.*, 2005). Apart from this, molecular and morphological data indicate that the actual diversity among Rhabditidae could be substantially larger than that suggested by classical taxonomists. This can be illustrated by *P. marina* that was recently shown to be an ‘umbrella taxon’ for a huge species complex (Derycke *et al.*, 2008).

It is noted that the Brevibuccidae, a family *incertae sedis* according to De Ley *et al.* (2006), was formerly placed in Clade 10 (Holterman *et al.*, 2006). This new positioning will not be discussed here because in both instances it received insufficient support.

#### CLADE 10 – APHELENCHOIDEA PART I (PARASITAPHELENCHIDAE, APHELENCHOIDIDAE AND SEINURIDAE) AND THE PANAGROLAIMOMORPHA

In a previous analysis by Holterman *et al.* (2006), a sister relationship between the Steinernematidae and

Strongyloididae/Alloionematidae was observed. The inclusion of more taxa and the use of a new, faster, maximum likelihood program resulted in Clade 10 splitting into part A and B. One of the reasons underlying the instability of Clade 10 could be the elevated AT-contents of *Bursaphelenchus* spp. (De Ley & Blaxter, 2002), and the Panagrolaimomorpha (AT-content *ca* 57%; Holterman *et al.*, 2006). Hence, the current composition of Clade 10 is probably biased by long-branch attraction (LBA) artefacts although, as there is as yet no properly supported alternative, there is little point proposing an alternative.

At the base of the Aphelenchoididae, there is a bifurcation between *Laimaphelenchus*, and two *Aphelenchoides* species, viz., *A. besseyi* (the causal agent of ‘white tip’ in rice) and *A. ritzemabosi* (chrysanthemum leaf nematode) on the one hand, and the remaining *Aphelenchoides* species and the representatives of the genus *Schistonchus* on the other. Although it is realised that only a subset of the described species is included here, it is remarkable to see that this split coincides with the tail tip morphology, both *A. besseyi* and *A. ritzemabosi* having a mucro with two to four pointed processes, whereas *Laimaphelenchus* species are usually characterised by fringed tubercles on the tail tip (Hunt, 1993). *Aphelenchoides fragariae*, *A. blastophthorus* and *A. saprophilus* possess a single, poorly to well developed mucro devoid of any processes. *Schistonchus* species have a tail with a mucronate tip (Hunt, 1993) and in *A. bicaudatus* the tail is terminally bifurcate. Hence, the presence or absence of processes on the tail tip seems to support the split within the Aphelenchoididae as revealed by ribosomal DNA data. Based on these results it may be worthwhile investigating in more detail whether *Schistonchus* should be maintained as a separate genus.

Another remarkable feature of Clade 10 is the positioning of *Anomyctus* and *Seinura* at the base of the Parasitaphelenchidae. A characteristic that coincides with this split is the morphology of the cephalic region. Contrary to the Aphelenchoidinae, Anomyctinae have a high cephalic region which is strongly offset by a constriction. This high and offset cephalic region is also typical for *Seinura* and *Bursaphelenchus* species.

#### CLADE 11 – CEPHALOBOMORPHA

The infraorder Cephalobomorpha includes five families: Cephalobidae, Osstellidae, Alirhabditidae, Elaphonematidae and Bicirronematidae. The first three families are represented here. It is noted that the non-represented families comprise only a small number of genera (Elaphone-

matidae: *Acromoldavicus*, *Kirjanovia* and *Elaphonema* (Baldwin *et al.*, 2001) and Bicirronematidae: *Trualaimus*, *Tricirronema* and *Bicirronema* (Holovachov *et al.*, 2003). Except for *Acromoldavicus*, for which some partial LSU rDNA sequences are available, no molecular data have been generated from these families. In accordance with a previous study based on partial LSU rDNA data (Nadler *et al.*, 2006), SSU rDNA sequences confirm the monophyly of the taxa presenting the superfamily Cephaloboidea, the only superfamily within the Cephalobomorpha. Compared with most other nematode families, SSU rDNA variation between representatives of the Cephalobidae is very limited. Moreover, most genera do not appear as monophyletic groups (see for instance *Cephalobus* or *Acrobeloides*). Attempts to resolve Cephalobidae relationships on the basis of LSU rDNA D1-D3 domains failed; D1 and D3 were too conserved, whereas the high variability of the D2 domains among outgroup species hampered proper alignment (O. Holovachov, unpubl.). The para- or polyphyly of genera can further be explained by the fact that, in the past, Cephalobidae systematics was mainly based on labial morphology (probolae). Phenotypic plasticity of probolae morphology and homoplasy – parallel and convergent evolution of lip morphology among Cephalobidae members – make lip morphology a poor choice as a basis for cephalobid systematics. The current data set does not allow us to say whether complex probolae are an ancestral (Nadler *et al.*, 2006) or a derived (De Ley *et al.*, 1993) character.

#### CLADE 12 – APHELENCHOIDEA PART II (APHELENCHIDAE) AND THE TYLENCHIDA

Out of the two families of the superfamily Aphelenchoidea, the Aphelenchidae and Aphelenchoididae, only the first family is found at the base of Clade 12. The Aphelenchidae genera *Aphelenchus* and *Paraphelenchus* are predominantly mycetophagous. Additional molecular data are needed to confirm their position as an immediate sistergroup of the Tylenchida. It is noted that the other family of the Aphelenchoidea, the Aphelenchoididae, was placed in Clade 10 in a sister relationship with (mainly) the Parasitaphelenchidae. Interestingly, the cellulases from Clade 12 members investigated so far, including *Aphelenchus avenae* (Dr Taisei Kikuchi, Tsukuba, Japan, pers. comm.), all belong to glycoside hydrolase family (GHF) 5, whereas cellulases from the Parasitaphelenchidae (Clade 11, close to the Aphelenchoididae) belong to GHF 45 (Kikuchi *et al.*, 2004).

Close to the base, there is a major split between the insect-associated and the plant-parasitic Tylenchida. Among the more basal Tylenchida families the resolution is poor; in fact we observe a large polytomy closely resembling the one presented by Holterman *et al.* (2009). Worthwhile mentioning is the positioning of *Halenchus fucicola*, a parasite of marine plants. Together with *Hirschmanniella* spp., such as *H. zostericola*, *H. mexicana*, and *H. marina* (species from which no rDNA data are available yet) (Sher, 1968), *H. fucicola* constitutes one of the rare examples in the Tylenchida that made the transition from terrestrial to marine habitats.

Recently, only two tylenchid groups were reported to be monophyletic, namely the Criconematoidea and the Hoplolaimidae (including the cyst and cystoid nematodes) (Bert *et al.*, 2008). Concerning the superfamily Criconematoidea, it should be noted that Bert *et al.* (2008) followed the Criconematoidea definition given by De Ley *et al.* (2006), including the families Criconematidae, Hemicycliophoridae and Tylenchulidae. We adhered to the Tylenchida systematics of Siddiqi (2000) in which the same superfamily name comprises the Criconematidae only. The suborder Criconematina *sensu* Siddiqi (2000) corresponds to the superfamily Criconematoidea *sensu* De Ley *et al.* (2006) and this cluster within Clade 12 indeed appears to be monophyletic.

In a more distal part of the tree (Clade 12B), a major split is observed between (mainly) the Heteroderidae and Hoplolaimidae, and most Pratylenchidae and Meloidogynidae. In Bert *et al.* (2008) the Hoplolaimidae include cyst and cystoid nematodes and the reniform nematode *Rotylenchulus reniformis*. Here, the inclusion of more taxa (from 11 to 27) resulted in a confirmation of the monophyletic nature of the Hoplolaimidae *sensu lato* (Hoplolaimidae, Heteroderidae and Rotylenchulidae).

According to Siddiqi (2000), “Meloidogynidae and Heteroderidae most probably originated from ancestors similar to the contemporary, migratory Pratylenchidae and Hoplolaimidae, respectively” (p. 372). SSU rDNA data presented here and in a recent paper on Tylenchida evolution (Holterman *et al.*, 2009) tentatively support the first hypothesis. We hypothesise that members of the genera *Belonolaimus*, *Radopholus* or *Dolichodorus* could be relatively closely related to the common ancestor of the Hoplolaimidae and the Heteroderidae.

#### Concluding remarks

From the 19 nematode orders included in the classification framework presented by De Ley *et al.* (2006), 15

are represented in this paper. As compared with Holterman *et al.* (2006), the number of SSU rDNA sequences was more than tripled and, instead of Bayesian inference, a fast maximum likelihood method was used (including the gamma parameter). The resulting phylogenetic tree is presumably the largest published so far and the subdivision of the phylum Nematoda into 12 clades as proposed by Holterman *et al.* (2006) still seems to hold. However, the deep subdivision of the phylum Nematoda should be regarded as ‘work in progress’, and a multi loci approach will be required for a more definitive framework.

The SSU rDNA-based molecular framework of the phylum Nematoda shows the extensiveness of parallel evolution among nematodes. Irrespective of whether ecological (*e.g.*, trophic ecology), biological (*e.g.*, mode of reproduction), or morphological (*e.g.*, tail shape) characteristics are taken into consideration, in the vast majority of cases similar characteristics are found on independent branches within Figure 1. Hence, similar ecological challenges independently gave rise to common adaptive phenotypes. Of course, this phenomenon greatly complicates the deduction of phylogenetic relationships and this co-explains the volatility of nematode systematics. Extensive convergent evolution has occurred in many organismal groups and comparison of DNA sequences from truly orthologous genes can be very helpful to elucidate evolutionary relationships. This point can be illustrated with an example from the plant kingdom, the family Brassicaceae. This is a large plant family (338 genera and 3700 species) and convergent evolution in nearly every morphological character lead to major problems in taxa description. The use of neutral molecular data – in this case internal transcribed spacer (ITS) sequences – substantially contributed to a more stable phylogeny (Bailey *et al.*, 2006). In another example, two major ascomycete classes Dothideomycetes and Sordariomycetes, the morphological characteristics used for the classification underwent major parallel evolution and, hence such characteristics appeared to be of little use for phylogenetics. In this case, the combined use LSU rDNA and a RNA polymerase II subunit (RPB2) resulted in a more stable systematic framework (Shenoy *et al.*, 2006).

We are currently using the framework described in this paper for DNA barcode-based nematode detection and community analysis. With regard to detection we have focused so far on plant-parasitic nematodes, often a small minority with a nematode community. On the basis of the framework presented here, it is (in most cases) possible to define species-specific sequence signatures and to

design simple and cheap PCR primers that allow real-time PCR-based detection and quantification of pathogenic nematodes in complex DNA backgrounds. At the same time, the SSU rDNA alignment was used to design dozens of family-specific PCR primers (see for example Holterman *et al.* (2008a) and we are currently testing quantitative, DNA barcode-based nematode community analyses under field conditions.

## Acknowledgements

H.v.M., S.v.d.E. and M.H. contributed equally to this work. The authors thank Dr Matthias Herrmann from the Max Planck Institute for Developmental Biology, Department of Evolutionary Biology, Tübingen, Germany, for providing us with cultures of a number of Diplogastriidae species. H.v.M., S.v.d.E. and J.H. acknowledge financial support from SenterNovem grant IS043076 (SenterNovem is an agency of the Dutch Ministry of Economic Affairs). M.H. was supported by the Dutch Technology Foundation (STW) grant WBI 4725.

## References

- AGRIOS, G.N. (2005). *Plant pathology*. London, UK, Elsevier Academic Press, 922 pp.
- AGUINALDO, A.M.A., TURBEVILLE, J.M., LINFORD, L.S., RIVERA, M.C., GAREY, J.R., RAFF, R.A. & LAKE, J.A. (1997). Evidence for a clade of nematodes, arthropods and other moulting animals. *Nature* 387, 489-493.
- ALESHIN, V.V., KEDROVA, O.S., MILYUTINA, I.A., VLADYCHENSKAYA, N.S. & PETROV, N.B. (1998). Relationships among nematodes based on the analysis of 18S rRNA gene sequences: molecular evidence for monophyly of chromadorian and secernentian nematodes. *Russian Journal of Nematology* 6, 175-184.
- ANDRÁSSY, I. (2007). *Free-living nematodes of Hungary (Nematoda errantia)*, II. *Pedozoologica Hungarica* No. 4. Budapest, Hungary, Hungarian Natural History Museum, 496 pp.
- BAILEY, C.D., KOCH, M.A., MAYER, M., MUMMENHOFF, K., O’KANE JR, S.L., WARWICK, S.I., WINDHAM, M.D. & AL-SHEHBAZ, I.A. (2006). Toward a global phylogeny of the Brassicaceae. *Molecular Biology and Evolution* 23, 2142-2160.
- BALDWIN, J.G., DE LEY, I.T., MUNDO-OCAMPO, M., DE LEY, P., NADLER, S.A. & GEBRE, M. (2001). *Acromoldavicus mojavicus* n. sp. (Nematoda: Cephaloidea) from the Mojave Desert, California. *Nematology* 3, 343-353.
- BEN ALI, A., WUYTS, J., DE WACHTER, R., MEYER, A. & VAN DE PEER, Y. (1999). Construction of a variability map

- for eukaryotic large subunit ribosomal RNA. *Nucleic Acids Research* 27, 2825-2831.
- BERT, W., LELIAERT, F., VIERSTRAETE, A.R., VANFLETEREN, J.R. & BORGONIE, G. (2008). Molecular phylogeny of the Tylenchida and evolution of the female gonoduct (Nematoda: Rhabditida). *Molecular Phylogenetics and Evolution* 48, 728-744.
- BLAXTER, M.L., DE LEY, P., GAREY, J.R., LIU, L.X., SCHELDAMAN, P., VIERSTRAETE, A., VANFLETEREN, J.R., MACKAY, L.Y., DORRIS, M., FRISSE, L.M., ET AL. (1998). A molecular evolutionary framework for the phylum Nematoda. *Nature* 392, 71-75.
- BOUTSIKA, K. (2002). *Molecular identification and phylogenies of virus and non-virus vector trichodorid nematodes*. Ph.D. Thesis, University of Dundee, Dundee, UK.
- BRZESKI, M.W. & WINISZEWSKA-ŚLIPIŃSKA, G. (1993). Taxonomy of Tripyliidae (Nematoda, Enoplia). *Nematologica* 39, 12-52.
- CHITWOOD, B.G. (1951). North American marine nematodes. *The Texas Journal of Science* 4, 617-672.
- CHITWOOD, B.G. (1958). The designation of official names for higher taxa of invertebrates. *Bulletin of Zoological Nomenclature* 15, 860-895.
- CHITWOOD, B.G. & CHITWOOD, M.B. (1933). The characters of a protonematode. *Journal of Parasitology* 20, 130.
- COOMANS, A.V. & RASKI, D.J. (1988). 2. New species of *Prismatolaimus* de Man, 1880 (Nemata, Prismatolaimidae) in southern Chile. *Journal of Nematology* 20, 288-303.
- DE LEY, P. & BLAXTER, M.L. (2002). Systematic position and phylogeny. In: Lee, D.L. (Ed.). *The biology of nematodes*. London, UK, Taylor & Francis, pp. 1-30.
- DE LEY, P., SIDDIQI, M.R. & BOSTRÖM, S. (1993). A revision of the genus *Pseudacrobeles* Steiner, 1938 (Nematoda: Cephalobidae). Part 2. *Bunobus* subgen. n., problematical species, discussion and key. *Fundamental and Applied Nematology* 16, 289-308.
- DE LEY, P., DECRAEMER, W. & ABEBE, E. (2006). Introduction: Summary of present knowledge and research addressing the ecology and taxonomy of freshwater nematodes. In: Abebe, E., Andrásy, I. & Traunspurger, W. (Eds). *Freshwater nematodes, ecology and taxonomy*. Wallingford, CABI Publishing, pp. 3-30.
- DERYCKE, S., FONSECA, G., VIERSTRAETE, A., VANFLETEREN, J., VINCX, M. & MOENS, T. (2008). Disentangling taxonomy within the *Rhabditis (Pellioiditis) marina* (Nematoda, Rhabditidae) species complex using molecular and morphological tools. *Zoological Journal of the Linnean Society* 152, 1-15.
- DUNN, C.W., HEJNOL, A., MATUS, D.Q., PANG, K., BROWNE, W.E., SMITH, S.A., SEAVER, E., ROUSE, G.W., OBST, M., EDGEcombe, G.D., ET AL. (2008). Broad phylogenomic sampling improves resolution of the animal tree of life. *Nature* 452, 745-749.
- HALL, T.A. (1999). BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. *Nucleic Acids Symposium Series* 41, 95-98.
- HOLOVACHOV, O. (2006). *Morphology and systematics of the order Plectida Malakhov, 1982 (Nematoda)*. Ph.D. Thesis. Wageningen University and Research Centre, Wageningen, The Netherlands. 246 pp.
- HOLOVACHOV, O., ESQUIVEL, A. & BONGERS, T. (2003). Free-living nematodes from nature reserves in Costa Rica. 4. Cephalobina. *Nematology* 5, 1-15.
- HOLTERMAN, M., VAN DER WURFF, A., VAN DEN ELSEN, S., VAN MEGEN, H., BONGERS, T., HOLOVACHOV, O., BAKKER, J. & HELDER, J. (2006). Phylum-wide analysis of SSU rDNA reveals deep phylogenetic relationships among nematodes and accelerated evolution toward crown clades. *Molecular Biology and Evolution* 23, 1792-1800.
- HOLTERMAN, M., RYBARCZYK, K., VAN DEN ELSEN, S., VAN MEGEN, H., MOOYMAN, P., PEÑA-SANTIAGO, R., BONGERS, T., BAKKER, J. & HELDER, J. (2008a). A ribosomal DNA-based framework for the detection and quantification of stress-sensitive nematode families in terrestrial habitats. *Molecular Ecology Resources* 8, 23-34.
- HOLTERMAN, M., HOLOVACHOV, O., VAN DEN ELSEN, S., VAN MEGEN, H., BONGERS, T., BAKKER, J. & HELDER, J. (2008b). Small subunit ribosomal DNA-based phylogeny of basal Chromadoria (Nematoda) suggests that transitions from marine to terrestrial habitats (and vice versa) require relatively simple adaptations. *Molecular Phylogenetics and Evolution* 48, 785-763.
- HOLTERMAN, M., KARSSSEN, G., VAN DEN ELSEN, S., VAN MEGEN, H., BAKKER, J. & HELDER, J. (2009). Small subunit rDNA-based phylogeny of the Tylenchida sheds light on relationships among some high-impact plant-parasitic nematodes and the evolution of plant feeding. *Phytopathology* 99, 227-235.
- HONISCH, M. & KRONE, O. (2008). Phylogenetic relationships of Spiruromorpha from birds of prey based on 18S rDNA. *Journal of Helminthology* 82, 129-133.
- HUNT, D.J. (1993). *Aphelenchida, Longidoridae and Trichodoridae: their systematics and bionomics*. Wallingford, UK, CABI Publishing, 352 pp.
- JAIRAJPURI, M.S. (1982). The systematic position of *Californidorus* Robbins & Weiner, 1978 (Nematoda: Dorylaimida). *Systematic Parasitology* 4, 135-137.
- KANZAKI, N., ABE, F., GIBLIN-DAVIS, R.M., KIONTKE, K., FITCH, D.H.A., HATA, K. & SONE, K. (2008). *Teratorhabditis synpapillata* Sudhaus, 1985 (Rhabditida: Rhabditidae) is an associate of the red palm weevil, *Rhynchophorus ferrugineus* (Coleoptera: Curculionidae). *Nematology* 10, 207-218.
- KARANASTASI, E., DECRAEMER, W., ZHENG, J.W., DE ALMEIDA, M.T.M. & BROWN, D.J.F. (2001). Interspecific differences in the fine structure of the body cuticle of Trichodoridae Thorne, 1935 (Nematoda: Diphtherophorina) and



- review of anchoring structures of the epidermis. *Nematology* 3, 525-533.
- KIKUCHI, T., JONES, J.T., AIKAWA, T., KOSAKA, H. & OGIURA, N. (2004). A family of glycosyl hydrolase family 45 cellulases from the pine wood nematode *Bursaphelenchus xylophilus*. *FEBS Letters* 572, 201-205.
- KIONTKE, K. & SUDHAUS, W. (2006). Ecology of *Caenorhabditis* species. The *C. elegans* Research Community, WormBook, doi/10.1895/wormbook.1.37.1, <http://www.wormbook.org>.
- KIONTKE, K., BARRIÈRE, A., KOLOTUEV, I., PODBILEWICZ, B., SOMMER, R., FITCH, D.H.A. & FÉLIX, M.A. (2007). Trends, stasis, and drift in the evolution of nematode vulva development. *Current Biology* 17, 1925-1937.
- LABANDEIRA, C.C. (2005). Invasion of the continents: cyanobacterial crusts to tree-inhabiting arthropods. *Trends in Ecology and Evolution* 20, 253-262.
- LAMBERTI, F., MOLINARI, S., MOENS, M. & BROWN, D.J.F. (2000). The *Xiphinema americanum* group. I. Putative species, their geographical occurrence and distribution, and regional polytomous identification keys for the group. *Russian Journal of Nematology* 8, 65-84.
- LORENZEN, S. (1981). Entwurf eines phylogenetischen Systems der freilebenden Nematoden. *Veröffentlichungen des Institut für Meeresforschungen Bremerhaven, Supplement* 7, 1-472.
- LUKEŠ, J., HORÁK, A. & SCHOLZ, T. (2005). Helminth genome projects: all or nothing. *Trends in Parasitology* 21, 265-266.
- MELDAL, B.H.M., DEBENHAM, N.J., DE LEY, P., TANDINGAN DE LEY, I., VANFLETEREN, J., VIERSTRAETE, A., BERT, W., BORGONIE, G., MOENS, T., TYLER, P.A., ET AL. (2007). An improved molecular phylogeny of the Nematoda with special emphasis on marine taxa. *Molecular Phylogenetics and Evolution* 42, 622-636.
- MICOLETZKY, H. (1922). Die freilebenden Erd-Nematoden mit besonderer Berücksichtigung der Steiermark und der Bukowina, zugleich mit einer Revision sämtlicher nicht mariner, freilebender Nematoden in Form von Genus – Beschreibung un Bestimmungsschlüsseln. *Archiv für Naturgeschichte, Abteilung A* 87, 1-650.
- MULLIN, P.G., HARRIS, T.S. & POWERS, T.O. (2005). Phylogenetic relationships of Nysolaimina and Dorylaimina (Nematoda: Dorylaimida) inferred from small subunit ribosomal DNA sequences. *Nematology* 7, 59-79.
- NADLER, S.A., DE LEY, P., MUNDO-OCAMPO, M., SMYTHE, A.B., STOCK, S.P., BUMBARGER, D., ADAMS, B.J., DE LEY, I.T., HOLOVACHOV, O. & BALDWIN, J.G. (2006). Phylogeny of Cephalobina (Nematoda): molecular evidence for recurrent evolution of probolae and incongruence with traditional classifications. *Molecular Phylogenetics and Evolution* 40, 696-711.
- NADLER, S.A., CARRENO, R.A., MEJIA-MADRID, H., ULLBERG, J., PAGAN, C., HOUSTON, R. & HUGOT, J.P. (2007). Molecular phylogeny of clade III nematodes reveals multiple origins of tissue parasitism. *Parasitology* 134, 1421-1442.
- NEILSON, R., YE, W., OLIVEIRA, C.M.G., HÜBSCHEN, J., ROBBINS, R.T., BROWN, D.J.F. & SZALANSKI, A.L. (2004). Phylogenetic relationships of Longidoridae species (Nematoda: Dorylaimida) from North America inferred from 18S rDNA sequence data. *Helminthologia* 41, 209-215.
- OLIVEIRA, C.M.G., FERRAZ, L.C.C.B., MONTEIRO, A.R., FENTON, B., MALLOCH, G. & NEILSON, R. (2004). Molecular and morphometric analyses of *Xiphidurus* species (Nematoda: Longidoridae). *Nematology* 6, 715-727.
- POINAR JR, G., KERP, H. & HASS, H. (2008). *Palaeonema phyticum* gen. n., sp. n. (Nematoda: Palaeonematidae fam. n.), a Devonian nematode associated with early land plants. *Nematology* 10, 9-14.
- POSADA, D. & CRANDALL, K.A. (1998). MODELTEST: testing the model of DNA substitution. *Bioinformatics* 14, 817-818.
- REBOREDO, G.R. & CAMINO, N.B. (2000). Two new Rhabditida species (Nematoda: Rhabditidae) parasites of *Cyclocephala signaticollis* (Coleoptera: Scarabaeidae) in Argentina. *Journal of Parasitology* 86, 819-821.
- SHENOY, B.D., JEEWON, R., WU, W.P., BHAT, D.J. & HYDE, K.D. (2006). Ribosomal and RPB2 DNA sequence analyses suggest that *Sporidesmium* and morphologically similar genera are polyphyletic. *Mycological Research* 110, 916-928.
- SHER, S.A. (1968). Revision of the genus *Hirschmanniella* Luc & Goodey, 1963 (Nematoda: Tylenchoidea). *Nematologica* 14, 243-275.
- SIDDIQI, M.R. (2000). *Tylenchida parasites of plants and insects*, 2nd edition. Wallingford, UK, CABI Publishing, 833 pp.
- STAMATAKIS, A. (2006). RAxML-VI-HPC: Maximum likelihood-based phylogenetic analyses with thousands of taxa and mixed models. *Bioinformatics* 22, 2688-2690.
- STOCK, S.P., CAICEDO, A.M. & CALATAYUD, P.A. (2005). *Rhabditis (Oscheius) colombiana* n. sp. (Nematoda: Rhabditidae), a necromenic associate of the subterranean burrower bug *Cyrtomenus bergi* (Hemiptera: Cydnidae) from the Cauca Valley, Colombia. *Nematology* 7, 363-373.
- SUDHAUS, W. (1993). Redescription of *Rhabditis (Oscheius) tipulae* (Nematoda: Rhabditidae) associated with leatherjackets, larvae of *Tipula paludosa* (Diptera: Tipulidae). *Nematologica* 39, 234-239.
- ZHANG, C., LIU, J., XU, M., SUN, J., YANG, S., AN, X., GAO, G., LIN, M., LAI, R., HE, Z., WU, Y. & ZHANG, K. (2008). *Heterorhabditoides chongmingensis* gen. nov., sp. nov. (Rhabditida: Rhabditidae), a novel member of the entomopathogenic nematodes. *Journal of Invertebrate Pathology* 98, 153-168.



## Supplementary material

**Positioning and lengths of Length Variable Regions (LVRs)**

LVR 1:	14-51	(length 38 nt)
LVR 2:	104-132	(length 29 nt)
LVR 3:	186-234	(length 349 nt)
LVR 4:	269-447	(length 179 nt)
LVR 5:	698-711	(length 14 nt)
LVR 6:	991-1257	(length 267 nt)
LVR 7:	1304-1321	(length 18 nt)
LVR 8:	1750-1770	(length 21 nt)
LVR 9:	2151-2284	(length 134 nt)
LVR 10:	2293-2306	(length 14 nt)
LVR 11:	2337-2352	(length 16 nt)
LVR 12:	2456-2473	(length 17 nt)
LVR 13:	2756-2904	(length 149 nt)

In total 945 positions have been removed

Including LVRs: 2967 aligned positions

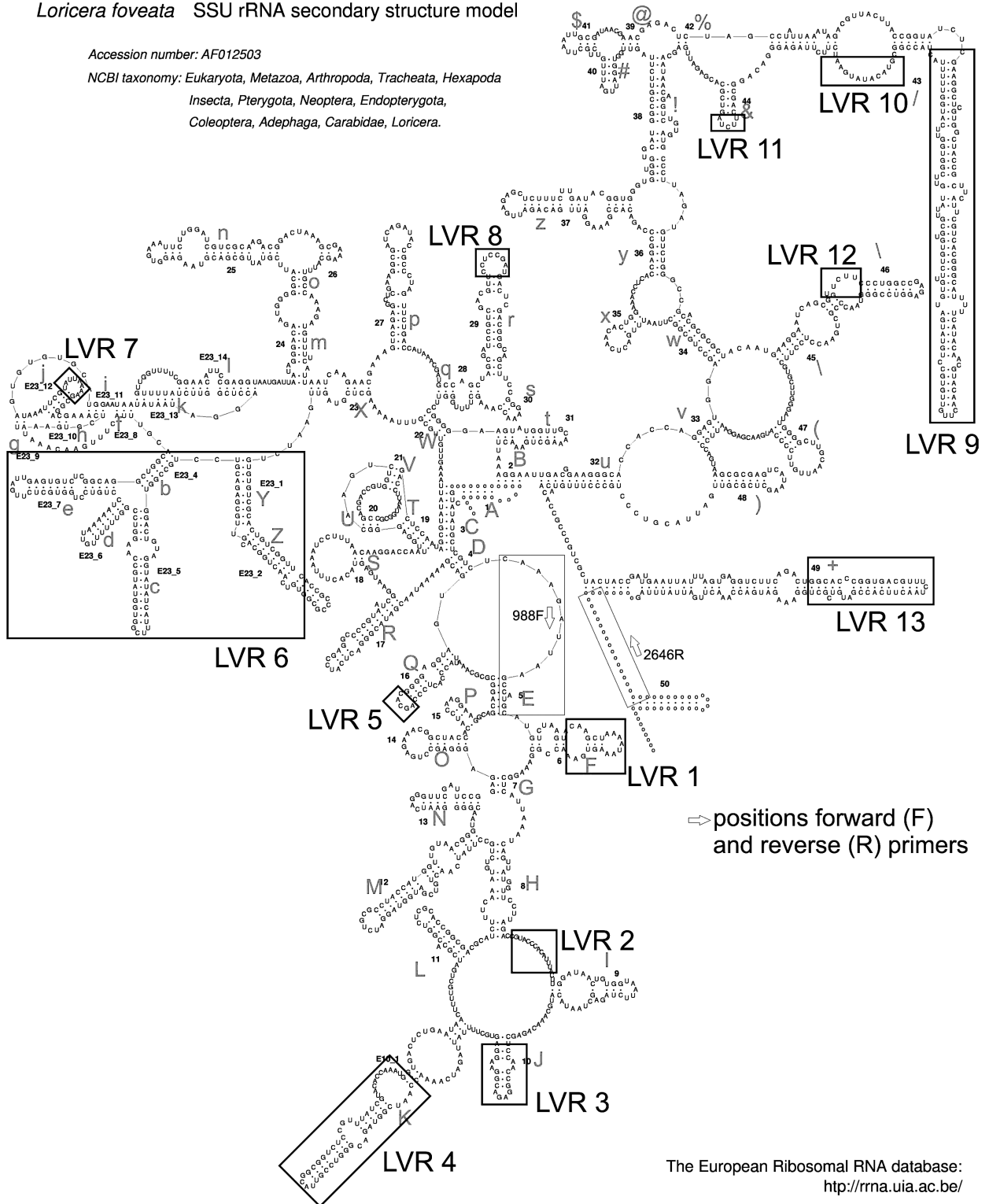
Excluding LVRs: 2022 aligned positions

**Fig. S1. Part 1.** *Effect of the removal of SSU rDNA length variable regions (LVRs) on the overall topology of a SSU rDNA-based ML tree. Positioning and length of the LVRs are given in part 1; the positioning of the LVR within the SSU rRNA is given in part 2. The best likelihood phylogenetic tree recovered by RAxML on the basis of SSU rDNA sequences after removal of LVRs is presented in part 3. Only bootstrap values >50% are given next to the nodes. Branch lengths are given in parentheses.*

*Loricera foveata* SSU rRNA secondary structure model

Accession number: AF012503

NCBI taxonomy: Eukaryota, Metazoa, Arthropoda, Tracheata, Hexapoda  
 Insecta, Pterygota, Neoptera, Endopterygota,  
 Coleoptera, Adephaga, Carabidae, Loricera.



The European Ribosomal RNA database:  
<http://rrna.uia.ac.be/>

Fig. S1. Part 2.

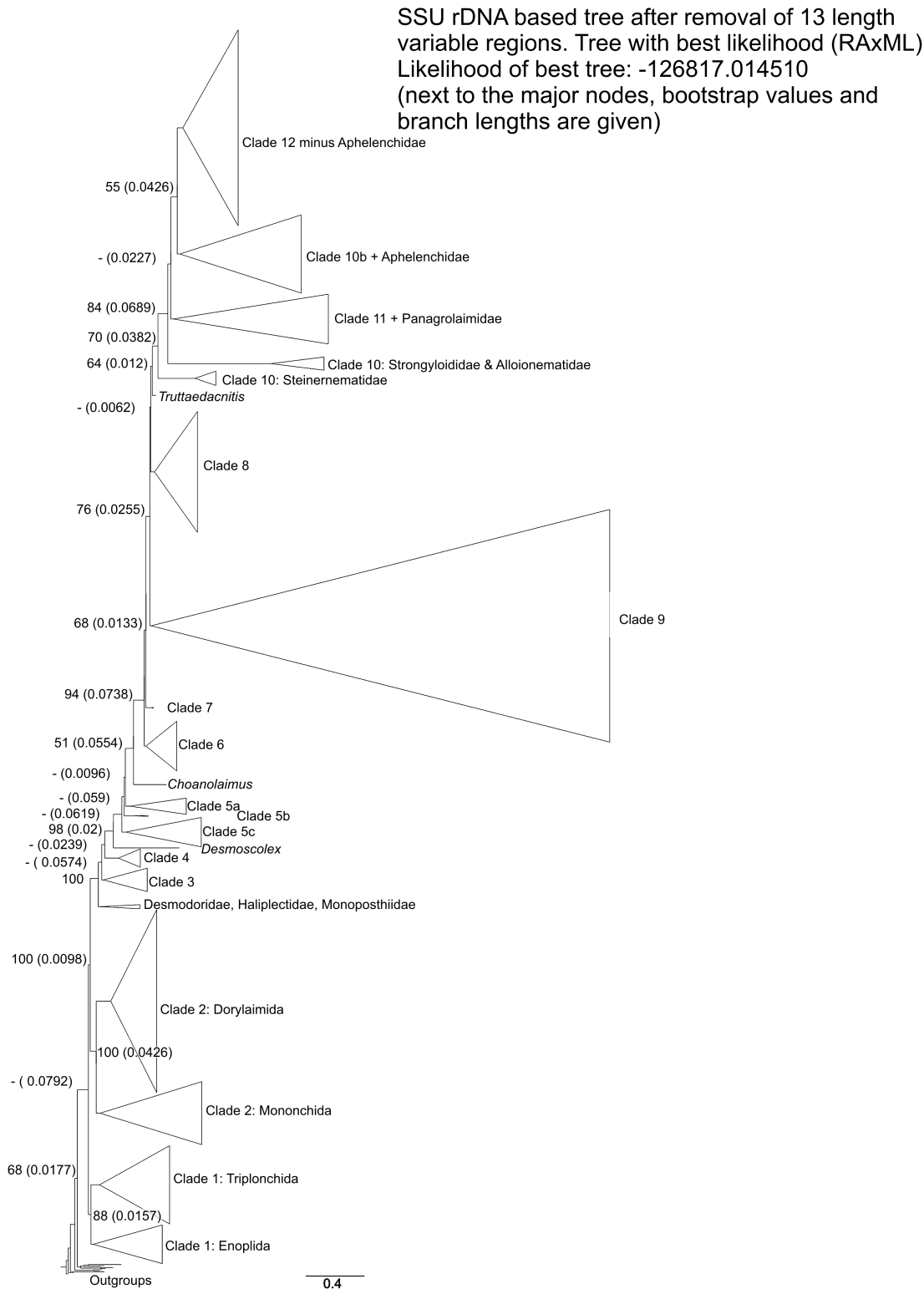


Fig. S1. Part 3.

**Table S1.** List of all SSU rDNA sequences used in this paper (scientific names, specimen identifiers, and corresponding GenBank accession numbers). A 'G' behind a nematode name indicates that the sequence was acquired from GenBank. The outgroup members are in the shaded lines.

Genus	Species	ID	GenBank No.
<i>Acanthocheilonema</i>	<i>viteae</i>	ACheVit1G	DQ094171
<i>Acanthopharynx</i>	<i>micans</i>	AcanMicG	Y16911
<i>Achromadora</i>	cf. <i>terricola</i>	AchrTerZ	AY593940
<i>Achromadora</i>	<i>ruvicola</i> 1	AchrRur1	AY593941
<i>Achromadora</i>	sp.	AchrSp2	AY284718
<i>Achromadora</i>	sp.	AchrSp1	AY284717
<i>Acrobeles</i>	<i>ciliatus</i>	AcroCilG	AF202148
<i>Acrobeles</i>	<i>complexus</i>	AcroCom1G	U81577
<i>Acrobeles</i>	<i>complexus</i> 1	AcroCom1	AY284671
<i>Acrobeles</i>	sp.	AcroSpG	U81576
<i>Acrobeloides</i>	<i>apiculatus</i> 1	AcLoApi1	AY284673
<i>Acrobeloides</i>	<i>bodenheimeri</i>	AcLoBod1G	AF202159
<i>Acrobeloides</i>	<i>bodenheimeri</i>	AcLoBod2G	AF202162
<i>Acrobeloides</i>	<i>maximus</i>	AcLoMax1G	EU196016
<i>Acrobeloides</i>	<i>maximus</i>	AcLoMax2G	EU306344
<i>Acrobeloides</i>	<i>nanus</i>	AcLoNan1G	DQ102707
<i>Acrobeloides</i>	<i>nanus</i> 1	AcLoNan1	AY284672
<i>Acrobeloides</i>	sp.	AcLoSpG	AF034391
<i>Acrostichus</i>	<i>halicti</i>	AcStHalG	U61759
<i>Adoncholaimus</i>	sp.	AdonSpG	AF036642
<i>Aelurostrongylus</i>	<i>abstrusus</i>	AeluAbs1G	AJ920366
<i>Afenestrata</i>	<i>koreana</i>	AfenKor1G	EU306357
<i>Alaimus</i>	<i>parvus</i> 1	AlaiPar1	AY284738
<i>Alaimus</i>	sp.	AlaiSp1G	AJ966514
<i>Alaimus</i>	sp. 1	AlaiSp1	FJ040489
<i>Alinema</i>	<i>amazonicum</i>	AlinAma1G	DQ442672
<i>Allodorylaimus</i>	<i>andrassyi</i>	AlloAnd1	AY284801
<i>Allodorylaimus</i>	sp.	AlloSp1G	AJ966472
<i>Amidostomum</i>	<i>cygni</i>	AmidCyg1G	AJ920353
<i>Amplimerlinius</i>	<i>icarus</i>	AmplIca1G	EU306351
<i>Anaplectus</i>	<i>grandepapillatus</i>	AnapGra1	AY284697
<i>Anaplectus</i>	<i>grandepapillatus</i>	AnapGra2	AY284698
<i>Anaplectus</i>	<i>porosus</i>	AnapPor1	AY284696
<i>Anaplectus</i>	<i>porosus</i>	AnapPor2	FJ040453
<i>Anaplectus</i>	sp.	AnapSp1G	AJ966473
<i>Anatonchus</i>	<i>tridentatus</i>	AnatTri1	AY284768
<i>Anatonchus</i>	<i>tridentatus</i>	AnatTri2	AY284769
<i>Anatonchus</i>	<i>tridentatus</i>	AnatTri1G	AJ966474
<i>Ancylostoma</i>	<i>caninum</i>	AncyCan1G	AJ920347
<i>Ancylostoma</i>	<i>duodenale</i>	AncyDuo1G	EU344798
<i>Angiostrongylus</i>	<i>cantonensis</i>	AngiCan1G	AY295804
<i>Angiostrongylus</i>	<i>costaricensis</i>	AngiCos1G	DQ116748
<i>Angiostrongylus</i>	<i>costaricensis</i>	AngiCos2G	EF514913
<i>Angiostrongylus</i>	<i>dujardini</i>	AngiDuj1G	AY542282
<i>Angiostrongylus</i>	<i>dujardini</i>	AngiDuj2G	EF514915
<i>Angiostrongylus</i>	<i>malaysiensis</i>	AngiMal1G	EF514914
<i>Angiostrongylus</i>	<i>vasorum</i>	AngiVas1G	AJ920365
<i>Angiostrongylus</i>	<i>vasorum</i>	AngiVas2G	EF514916

Table S1. (Continued).

Genus	Species	ID	GenBank No.
<i>Anguillicola</i>	<i>crassus</i>	AngCCra1G	DQ118535
<i>Anguillicola</i>	<i>crassus</i>	AngCCra2G	DQ490223
<i>Anguina</i>	<i>tritici</i> 1	AnguTri1	AY593913
<i>Anisakis</i>	<i>pegreffii</i>	AnisPeg1G	EF180082
<i>Anisakis</i>	sp.	AnisSp1G	U81575
<i>Anisakis</i>	sp.	AnisSp2G	U94365
<i>Anomyctus</i>	<i>xenurus</i>	AnomXen1	FJ040413
<i>Anoplostoma</i>	<i>rectospiculum</i>	AnopRec1G	AY590149
<i>Anoplostoma</i>	sp. 1	AnopSp1	FJ040491
<i>Anoplostoma</i>	sp. 2	AnopSp2	FJ040492
<i>Aphanolaimus</i>	<i>aquaticus</i>	AphaAqu2	AY593933
<i>Aphanolaimus</i>	<i>aquaticus</i> 1	AphaAqu1	AY593932
<i>Aphanonchus</i>	cf. <i>europaeus</i>	AphNEurZ1	EF591319
<i>Aphelenchoides</i>	1	AChoSp1	AY284646
<i>Aphelenchoides</i>	2	AChoSp2	AY284647
<i>Aphelenchoides</i>	<i>besseyi</i>	AChoBes1G	AY508035
<i>Aphelenchoides</i>	<i>bicaudatus</i> 1	AChoBic1	AY284643
<i>Aphelenchoides</i>	<i>blastophorus</i> 1	AChoBla1	AY284644
<i>Aphelenchoides</i>	cf. <i>bicaudatus</i>	AcHoBicZ	FJ040407
<i>Aphelenchoides</i>	<i>fragariae</i>	AChoFra1G	AB067755
<i>Aphelenchoides</i>	<i>fragariae</i>	AChoFra2G	AJ966475
<i>Aphelenchoides</i>	<i>fragariae</i>	AChoFra3G	DQ901551
<i>Aphelenchoides</i>	<i>fragariae</i> 1	AChoFra1	AY284645
<i>Aphelenchoides</i>	<i>ritzemabosi</i>	AChoRit1G	DQ901554
<i>Aphelenchoides</i>	<i>saprophilus</i>	AChoSap1	FJ040408
<i>Aphelenchoides</i>	sp.	AChoSp1G	DQ901550
<i>Aphelenchoides</i>	sp.	AChoSp2G	DQ901552
<i>Aphelenchoides</i>	sp. 3	AChoSp3	FJ040409
<i>Aphelenchoides</i>	sp. 4	AChoSp4	FJ040410
<i>Aphelenchoides</i>	sp. 5	AChoSp5	FJ040411
<i>Aphelenchoides</i>	sp. 6	AChoSp6	FJ040412
<i>Aphelenchus</i>	<i>avenae</i>	ApheAve1	AY284639
<i>Aphelenchus</i>	<i>avenae</i>	ApheAve2	AY284640
<i>Aphelenchus</i>	<i>avenae</i>	ApheAve1G	AF036586
<i>Aphelenchus</i>	<i>avenae</i>	ApheAve2G	AB368918
<i>Aphelenchus</i>	<i>avenae</i>	ApheAve3G	EU306347
<i>Aphelenchus</i>	sp.	ApheSp	AY284641
<i>Aporcelaimellus</i>	cf. <i>paraobtusicaudatus</i>	ApoEParZ	AY284812
<i>Aporcelaimellus</i>	<i>obtusicaudatus</i>	ApoEObt1	AY284811
<i>Aporcelaimellus</i>	<i>obtusicaudatus</i>	ApoEObt1G	DQ141212
<i>Aporcelaimellus</i>	sp.	ApoESp1G	AJ875154
<i>Aporcelaimellus</i>	sp.	ApoESp2G	AJ875153
<i>Aporcelaimellus</i>	sp.	ApoESp3G	AJ875155
<i>Aporcelaimellus</i>	sp. 1	ApoESp1	AY284813
<i>Aquatides</i>	<i>christei</i>	AquaCri1G	AY552963
<i>Ascaridia</i>	<i>galli</i>	AsDiGal1G	EF180058
<i>Ascaris</i>	<i>lumbricoides</i>	AscaLumG	U94366
<i>Ascaris</i>	sp.	AscaSpG	M58348
<i>Ascaris</i>	<i>suum</i>	AscaSuu1G	U94367
<i>Ascaris</i>	<i>suum</i>	AscaSuu2G	AF036587

**Table S1.** (Continued).

Genus	Species	ID	GenBank No.
<i>Ascarophis</i>	<i>arctica</i>	AscPArc1G	DQ094172
<i>Ascolaimus</i>	cf. <i>elongatus</i>	AscoElo1Z	FJ040460
<i>Ascolaimus</i>	cf. <i>elongatus</i>	AscoElo2Z	EF591330
<i>Ascolaimus</i>	<i>elongatus</i>	AscoElo2G	AM234617
<i>Aspidodera</i>	sp.	AspiSp1G	EF180070
<i>Astomonema</i>	sp.	AstoSp1G	DQ408759
<i>Astomonema</i>	sp.	AstoSp2G	DQ408760
<i>Astomonema</i>	sp.	AstoSp3G	DQ408761
<i>Aulolaimus</i>	<i>oxycephalus</i>	AuloOxy1	AY284724
<i>Axonchium</i>	<i>propinquum</i>	AxonPro1	AY284820
<i>Axonolaimus</i>	sp. 1	AxLaSp1	FJ040461
<i>Axonolaimus</i>	sp. 2	AxLaSp2	EF591331
<i>Axonolaimus</i>	sp. 3	AxLaSp3	FJ040462
<i>Bastania</i>	<i>gracilis</i>	BastGra1	AY284725
<i>Bastania</i>	<i>gracilis</i>	BastGra2	AY284726
<i>Bathylaimus</i>	<i>australis</i>	BaLaAus1G	AJ966476
<i>Bathylaimus</i>	sp. 1	BaLaSp1	FJ040504
<i>Bathyodontus</i>	<i>cylindricus</i>	BathCyl1G	AY552964
<i>Bathyodontus</i>	<i>mirus</i>	BathMir1	AY284744
<i>Baujardia</i>	<i>mirabilis</i>	BaujMir1G	AF547385
<i>Baylisascaris</i>	<i>procyonis</i>	BaylproG	U94368
<i>Baylisascaris</i>	<i>transfuga</i>	BayltraG	U94369
<i>Belonolaimus</i>	<i>longicaudatus</i>	BeloLon1G	AY633449
<i>Belonolaimus</i>	<i>longicaudatus</i>	BeloLon2G	DQ912919
<i>Bitylenchus</i>	<i>dubius</i>	BityDub1G	EU306352
<i>Bitylenchus</i>	<i>dubius</i> 1	BityDub1	AY284601
<i>Boleodorus</i>	<i>thylactus</i>	BoleThy1	AY593915
<i>Boleodorus</i>	<i>thylactus</i>	BoleThy1G	AY993976
<i>Bradynema</i>	<i>listronotum</i>	BradLis1G	DQ915805
<i>Brevibucca</i>	<i>saprophaga</i>	BrevSap1G	EU196018
<i>Brevibucca</i>	sp.	BrevSpG	AF202163
<i>Brugia</i>	<i>malayi</i>	BrugMalG	AF036588
<i>Brumptaemilius</i>	<i>justini</i>	BrumJusG	AF036589
<i>Bunonema</i>	<i>franzi</i>	BunoFra1G	AJ966477
<i>Bunonema</i>	<i>reticulatum</i>	BunoRet2	AY593925
<i>Bunonema</i>	<i>reticulatum</i>	BunoRet3	FJ040450
<i>Bunonema</i>	<i>reticulatum</i>	BunoRet1G	EU196017
<i>Bunonema</i>	<i>reticulatum</i> 1	BunoRet1	AY284661
<i>Bunonema</i>	<i>richtersi</i>	BunoRic1	FJ040451
<i>Bunonema</i>	<i>richtersi</i>	BunoRic2	FJ040452
<i>Bunonema</i>	sp.	BunoSpG	U81582
<i>Bursaphelenchus</i>	1	BursSp1	AY284649
<i>Bursaphelenchus</i>	2	BursSp2	AY284650
<i>Bursaphelenchus</i>	<i>abietinus</i>	BursAbi1G	AY508011
<i>Bursaphelenchus</i>	<i>abruptus</i>	BursAbr1G	AY508010
<i>Bursaphelenchus</i>	<i>arthuri</i>	BursArt1G	AM397010
<i>Bursaphelenchus</i>	<i>borealis</i>	BursBor1G	AY508012
<i>Bursaphelenchus</i>	<i>cocophilus</i>	BursCoc1G	AY509153
<i>Bursaphelenchus</i>	<i>conicaudatus</i>	BursCon1G	AM397011
<i>Bursaphelenchus</i>	<i>doui</i>	BursDou1G	AM397012

Table S1. (Continued).

Genus	Species	ID	GenBank No.
<i>Bursaphelenchus</i>	<i>eggersi</i>	BursEgg1G	AY508013
<i>Bursaphelenchus</i>	<i>fraudulentus</i>	BursFra1G	AY508014
<i>Bursaphelenchus</i>	<i>fraudulentus</i>	BursFra2G	AY508015
<i>Bursaphelenchus</i>	<i>fungivorus</i>	BursFun1G	AY508016
<i>Bursaphelenchus</i>	<i>hellenicus</i>	BursHel1G	AY508017
<i>Bursaphelenchus</i>	<i>hildegardae</i>	BursHil1G	AM397013
<i>Bursaphelenchus</i>	<i>hofmanni</i>	BursHof1G	AY508018
<i>Bursaphelenchus</i>	<i>hylobianus</i>	BursHyl1G	AY508019
<i>Bursaphelenchus</i>	<i>kevinci</i>	BursKev1G	AY753531
<i>Bursaphelenchus</i>	<i>mucronatus</i>	BursMuc1G	AB067759
<i>Bursaphelenchus</i>	<i>mucronatus</i>	BursMuc2G	AY508020
<i>Bursaphelenchus</i>	<i>mucronatus</i>	BursMuc3G	AY508021
<i>Bursaphelenchus</i>	<i>mucronatus</i>	BursMuc4G	AY508022
<i>Bursaphelenchus</i>	<i>mucronatus</i>	BursMuc5G	AY508023
<i>Bursaphelenchus</i>	<i>mucronatus</i>	BursMuc6G	AM397015
<i>Bursaphelenchus</i>	<i>mucronatus</i> 1	BursMuc1	AY284648
<i>Bursaphelenchus</i>	<i>paracorneolus</i>	BursPar1G	AY508027
<i>Bursaphelenchus</i>	<i>pinasteri</i>	BursPin1G	AM397016
<i>Bursaphelenchus</i>	<i>poligraphi</i>	BursPol1G	AY508028
<i>Bursaphelenchus</i>	<i>rainulfi</i>	BursRai1G	AM397017
<i>Bursaphelenchus</i>	<i>seani</i>	BursSea1G	AY508029
<i>Bursaphelenchus</i>	<i>seani</i>	BursSea2G	AY508030
<i>Bursaphelenchus</i>	<i>sexdentati</i>	BursSex1G	AY508031
<i>Bursaphelenchus</i>	<i>sexdentati</i>	BursSex2G	AY508032
<i>Bursaphelenchus</i>	sp.	BursSp1G	AF037369
<i>Bursaphelenchus</i>	sp.	BursSp2G	AY508024
<i>Bursaphelenchus</i>	sp.	BursSp3G	AY508025
<i>Bursaphelenchus</i>	sp.	BursSp4G	AY508026
<i>Bursaphelenchus</i>	<i>thailandae</i>	BursTha1G	AM397019
<i>Bursaphelenchus</i>	<i>tusciae</i>	BursTus1G	AY508033
<i>Bursaphelenchus</i>	<i>vallesianus</i>	BursVal1G	AM397020
<i>Bursaphelenchus</i>	<i>willibaldi</i>	BursWil1G	AM397021
<i>Bursaphelenchus</i>	<i>xylophilus</i>	BursXyl1G	AB067760
<i>Bursaphelenchus</i>	<i>xylophilus</i>	BursXyl2G	AY508034
<i>Bursaphelenchus</i>	<i>xylophilus</i>	BursXyl3G	AM397022
<i>Bursaphelenchus</i>	<i>yongensis</i>	BursYon1G	AM397023
<i>Caenorhabditis</i>	<i>briggsae</i>	CaenBriG	U13929
<i>Caenorhabditis</i>	<i>drosophilae</i>	CaenDroG	AF083025
<i>Caenorhabditis</i>	<i>elegans</i>	CaenEle1G	X03680
<i>Caenorhabditis</i>	<i>elegans</i>	CaenEle2G	AY268117
<i>Caenorhabditis</i>	<i>elegans</i>	CaenEle3G	EU196001
<i>Caenorhabditis</i>	<i>elegans</i> 1	CaenEle1	AY284652
<i>Caenorhabditis</i>	<i>japonica</i>	CaenJap1G	AY602182
<i>Caenorhabditis</i>	<i>plicata</i>	CaenPli1G	AY602178
<i>Caenorhabditis</i>	<i>sonorae</i>	CaenSonG	AF083026
<i>Caenorhabditis</i>	sp.	CaenSp2G	U13930
<i>Caenorhabditis</i>	sp.	CaenSp1G	AF083006
<i>Caenorhabditis</i>	sp.	CaenSp3G	AY602180
<i>Caenorhabditis</i>	sp.	CaenSp4G	AY602181
<i>Caenorhabditis</i>	sp.	CaenSp5G	EU196000



**Table S1.** (Continued).

Genus	Species	ID	GenBank No.
<i>Caenorhabditis</i>	<i>vulgaris</i>	CaenVulG	U13931
<i>Californidorus</i>	sp.	CaliSp1G	AY283155
<i>Calyptonema</i>	sp. 1	CalySp1	FJ040503
<i>Camacolaimus</i>	sp. 1	CamaSp1	EF591325
<i>Camacolaimus</i>	sp. 2	CamaSp2	EF591327
<i>Camallanus</i>	<i>cotti</i>	CaLaCot1G	DQ442662
<i>Camallanus</i>	<i>cotti</i>	CaLaCot2G	EF180071
<i>Camallanus</i>	<i>lacustris</i>	CaLaLac1G	DQ442663
<i>Camallanus</i>	<i>oxycephalus</i>	CaLaOxy1G	DQ503463
<i>Camallanus</i>	sp.	CaLaSp1G	DQ442664
<i>Campydora</i>	<i>demonstrans</i>	CampDem1G	AY552965
<i>Capillaria</i>	<i>tenuissima</i>	CapiTen1G	EU004822
<i>Carcharodiscus</i>	<i>banaticus</i>	CarcBan1	AY284827
<i>Catanema</i>	sp.	CataSpG	Y16912
<i>Cephalenchus</i>	<i>hexalineatus</i> 1	CeLeHex1	AY284594
<i>Cephalobidae</i>		Cephfamil	FJ040406
<i>Cephaloboides</i>	cf. <i>armata</i>	CeBoArm1ZG	EU196005
<i>Cephaloboides</i>	<i>nidrosiensis</i>	CeBoNid1G	EU196020
<i>Cephaloboides</i>	sp.	CeBoSpG	AF083027
<i>Cephalobus</i>	<i>cubaensis</i>	CephCubG	AF202161
<i>Cephalobus</i>	<i>oryzae</i>	CephOryG	AF034390
<i>Cephalobus</i>	<i>persegnis</i>	CephPer1	AY284662
<i>Cephalobus</i>	<i>persegnis</i>	CephPer2	AY284663
<i>Cephalobus</i>	sp.	CephSp1G	AF202158
<i>Cephalobus</i>	sp.	CephSp2G	AF202160
<i>Ceratoplectus</i>	<i>armatus</i>	CeraArm1	AY284706
<i>Cervidellus</i>	<i>alutus</i>	CervAluG	AF202152
<i>Cervidellus</i>	sp. 1	CervSp1	AY284674
<i>Chabertia</i>	<i>ovina</i>	ChabOvi1G	AJ920341
<i>Chiloplacus</i>	<i>propinquus</i> 1	ChilPro1	AY284677
<i>Choanolaimus</i>	<i>psammophilus</i>	ChoaPsa4	FJ040467
<i>Choanolaimus</i>	<i>psammophilus</i> 2	ChoaPsa2	AY284715
<i>Choanolaimus</i>	<i>psammophilus</i> 3	ChoaPsa3	AY284716
<i>Chordodes</i>	<i>morgani</i>	ChDoMorG	AF036639
<i>Choriorhabditis</i>	<i>cristata</i>	ChorCri1G	EU196013
<i>Choriorhabditis</i>	<i>dudichi</i>	ChorDudG	AF083012
<i>Chromadoridae</i>	1	ChMafamil1	AY284713
<i>Chromadoridae</i>	sp. 2	ChMafamil2	FJ040474
<i>Chromadorina</i>	sp. 1	ChInSp1	FJ040470
<i>Chromadorina</i>	sp. 2	ChInSp2	FJ040471
<i>Chromadorita</i>	<i>leuckarti</i>	ChItleu1Z	FJ040473
<i>Chromadoropsis</i> (= <i>Atrochromadora</i> )	<i>vivipara</i>	ChOpVivG	AF047891
<i>Chronogaster</i>	3	ChGaSp3	FJ040455
<i>Chronogaster</i>	<i>boettgeri</i> 1	ChGaBoe1	AY593931
<i>Chronogaster</i>	<i>polonica</i>	ChGaSp1	AY284708
<i>Chronogaster</i>	sp. 2	ChGaSp2	AY284709
<i>Chronogaster</i>	<i>typica</i> 1	ChGaTyp1	FJ040456
<i>Chrysonema</i>	<i>attenuatum</i>	ChryAtt1	AY593945
<i>Chrysonema</i>	<i>attenuatum</i>	ChryAtt2	AY284779

Table S1. (Continued).

Genus	Species	ID	GenBank No.
<i>Chrysonema</i>	<i>attenuatum</i>	ChryAtt3	EF207245
<i>Clarkus</i>	<i>papillatus</i>	ClarPap1	AY284748
<i>Clarkus</i>	<i>papillatus</i>	ClarPap3	AY284749
<i>Clarkus</i>	<i>papillatus</i>	ClarPap2	AY284750
<i>Clarkus</i>	<i>papillatus</i>	ClarPap1G	AY552966
<i>Clarkus</i>	sp.	ClarSp1G	AJ966479
<i>Clavicaudoides</i>	<i>clavicaudatus</i>	ClavCla1	AY593944
<i>Clavicaudoides</i>	sp.	ClavSp1G	AY552967
<i>Clavicaudoides</i>	<i>trophurus</i>	ClavTro1	AY284772
<i>Clavicaudoides</i>	<i>trophurus</i>	ClavTro2	AY284773
<i>Clavicaudoides</i>	<i>trophurus</i>	ClavTro3	AY593943
<i>Contraecum</i>	<i>eudypulatae</i>	ContEud1G	EF180072
<i>Contraecum</i>	<i>microcephalum</i>	ContMic1G	AY702702
<i>Contraecum</i>	<i>multipapillatum</i>	ContMulG	U94370
<i>Coomansus</i>	<i>parvus</i>	CoomPar1	AY284766
<i>Coomansus</i>	<i>parvus</i>	CoomPar2	AY284767
<i>Coslenchus</i>	cf. <i>franklinae</i>	CoslFra1Z	AY284582
<i>Coslenchus</i>	<i>costatus</i> 1	CoslCos1	AY284581
<i>Coslenchus</i>	<i>franklinae</i>	CoslFra2	AY284583
<i>Crenosoma</i>	<i>mephitidis</i>	CrenMep1G	AY295805
<i>Crenosoma</i>	<i>vulpis</i>	CrenVul1G	AJ920367
<i>Criconema</i>	sp. 1	CricSp1G	AJ966480
<i>Cruzia</i>	<i>americana</i>	CrZiAmeG	U94371
<i>Cruznema</i>	1	CruzSp1	AY284655
<i>Cruznema</i>	2	CruzSp2	AY284656
<i>Cruznema</i>	3	CruzSp3	AY284657
<i>Cruznema</i>	4	CruzSp4	AY284658
<i>Cruznema</i>	<i>tripartita</i>	CruzTri1G	U73449
<i>Cruznema</i>	<i>tripartitum</i>	CruzTri2G	EU196012
<i>Cryptonchus</i>	sp. 1	CrypSp1	FJ040479
<i>Cryptonchus</i>	<i>tristis</i> 1	CrypTri1	EF207244
<i>Cuticonema</i>	<i>vivipara</i>	CuCoViv1G	EU196019
<i>Cyatholaimus</i>	sp.	CyatSp2G	AM234618
<i>Cyclodontostomum</i>	<i>purvisi</i>	CyclPur1G	AJ920340
<i>Cylicocyclus</i>	<i>insignis</i>	CyCyIns1G	AJ920342
<i>Cylindrolaimus</i>	<i>communis</i> 1	CyliCom1	AY593939
<i>Cylindrolaimus</i>	sp.	CyliSpG	AF202149
<i>Cyrnea</i>	<i>leptoptera</i>	CyrnLep1G	EU004815
<i>Cyrnea</i>	<i>seurati</i>	CyrnSeu1G	EU004816
<i>Daptonema</i>	<i>procerum</i>	DaptProG	AF047889
<i>Daptonema</i>	sp.	DaptSp1G	EF436228
<i>Daptonema</i>	sp. 1	DaptSp1	FJ040463
<i>Deladenus</i>	<i>siricidicola</i>	DeDeSir1G	AY633447
<i>Deladenus</i>	sp.	DeDeSp1G	AJ966481
<i>Deladenus</i>	sp.	DeDeSp2G	EU306345
<i>Deletrocephalus</i>	<i>dimidiatus</i>	DeleDim1G	AJ920346
<i>Demaniella</i>	sp. 1	DemaSp1	FJ040438
<i>Dentiphilometra</i>	sp.	DPhiSp1G	DQ442673
<i>Dentostomella</i>	sp.	DentSpG	AF036590
<i>Deontolaimus</i>	<i>papillatus</i>	DeonPap1	EF591322

Table S1. (Continued).

Genus	Species	ID	GenBank No.
<i>Deontolaimus</i>	<i>papillatus</i>	DeonPap2	FJ040457
<i>Desmodora</i>	<i>ovigera</i>	DeRaOviG	Y16913
<i>Desmolaimus</i>	sp. 1	DeLaSp1	EF591332
<i>Desmolaimus</i>	sp. 2	DeLaSp2	EF591333
cf. <i>Desmolaimus</i>		DeLaSp3Z	EF591336
cf. <i>Desmolaimus</i>		DeLaSp4Z	EF591337
<i>Desmoscolex</i>	sp. 2	DeCoSp2	EF591342
<i>Dichromadora</i>	sp. 1	DichSp1	FJ040506
<i>Dictyocaulus</i>	<i>capreolus</i>	DictCap1G	AY168862
<i>Dictyocaulus</i>	<i>capreolus</i>	DictCap2G	AY168859
<i>Dictyocaulus</i>	<i>eckerti</i>	DictEck1G	AY168857
<i>Dictyocaulus</i>	<i>eckerti</i>	DictEck2G	AY168858
<i>Dictyocaulus</i>	<i>eckerti</i>	DictEck3G	AY168863
<i>Dictyocaulus</i>	<i>eckerti</i>	DictEck4G	AY168864
<i>Dictyocaulus</i>	<i>filaria</i>	DictFil1G	AY168861
<i>Dictyocaulus</i>	<i>filaria</i>	DictFil2G	AJ920362
<i>Dictyocaulus</i>	sp.	DictSpG	AY168860
<i>Dictyocaulus</i>	<i>viviparus</i>	DictViv1G	AY168856
<i>Dictyocaulus</i>	<i>viviparus</i>	DictViv2G	AJ920361
<i>Didelphostrongylus</i>	<i>hayesi</i>	DideHay1G	AY295806
<i>Dilta</i>	<i>littoralis</i>	DiltLitG	AF005457
<i>Dintheria</i>	<i>tenuissima</i>	DintTen1	FJ040487
<i>Dipetalonema</i>	sp.	DiTaSp1G	DQ531723
<i>Diphtherophora</i>	<i>communis</i>	DiphCom1	AY593955
<i>Diphtherophora</i>	<i>obesa</i>	DiphObe1	AY284838
<i>Diphtherophora</i>	<i>obesa</i>	DiphObe2	AY284839
<i>Diphtherophora</i>	<i>obesus</i>	DiphObe1G	AY552968
<i>Diplogaster</i>	<i>rivalis</i> 1	DiGaRiv3	AY284688
<i>Diplogastridae</i>		DiGafamil	AY284689
<i>Diplogasteroides</i>	<i>magnus</i>	DiGOMag1	FJ040448
<i>Diplolaimella</i>	<i>dievengatensis</i>	DiplDie1G	AJ966482
<i>Diplolaimelloides</i>	<i>meyli</i>	DiLaMey1G	AF036644
<i>Diplolaimelloides</i>	<i>meyli</i>	DiLaMey2G	AF036611
<i>Diplolaimelloides</i>	sp.	DiLaSp1G	EF659926
<i>Diplolaimelloides</i>	sp.	DiLaSp9G	EF659927
<i>Diplolaimelloides</i>	sp.	DiLaSp10G	EF659925
<i>Diplolaimelloides</i>	sp.	DiLaSp11G	EF659924
<i>Diplolaimelloides</i>	sp.	DiLaSp16G	EF659919
<i>Diplolaimelloides</i>	sp.	DiLaSp17G	EF659918
<i>Diplolaimelloides</i>	sp.	DiLaSp18G	EF659917
<i>Diplopeltula</i>	sp. 1	DiPeSp1	EF591329
<i>Diploscapter</i>	<i>coronatus</i> 1	DiScCor1	AY593921
<i>Diploscapter</i>	sp.	DiScSp1G	U81586
<i>Diploscapter</i>	sp.	DiScSp2G	AF083009
<i>Diploscapter</i>	sp.	DiScSp3G	EU196003
<i>Dirofilaria</i>	<i>immitis</i>	DiroImmG	AF036638
<i>Discolaimus</i>	cf. <i>major</i>	DisLMaj1Z	EF207252
<i>Discolaimus</i>	<i>major</i>	DisLMaj1	AY284828
<i>Ditylenchus</i>	<i>adasi</i>	DityAda1	EU669909
<i>Ditylenchus</i>	<i>angustus</i>	DityAng1G	AJ966483

Table S1. (Continued).

Genus	Species	ID	GenBank No.
<i>Ditylenchus</i>	<i>destructor</i>	DityDes1	AY593912
<i>Ditylenchus</i>	<i>dipsaci</i>	DityDip8	EU669931
<i>Dolichodoros</i>	sp.	DoRuSp1G	DQ912918
<i>Dolichodoros</i>	sp.	DoRuSp2G	EF025336
<i>Domorganus</i>	<i>macronephritices</i>	DoGaMac1	FJ040454
<i>Dorylaimellus</i>	<i>montenegricus</i>	DoMeMon	AY284821
<i>Dorylaimellus</i>	<i>virginianus</i>	DoMeVir1G	AY552969
<i>Dorylaimoides</i>	<i>limnophilus</i>	DoMoLim2	AY593950
<i>Dorylaimoides</i>	<i>micoletskyi</i>	DoMoMic1	AY284830
<i>Dorylaimoides</i>	sp. 1	DoMoSp1	AY593951
<i>Dorylaimus</i>	<i>stagnalis</i>	DomuSta2	AY284776
<i>Dorylaimus</i>	<i>stagnalis</i>	DoMuSta3	AY284777
<i>Dracunculus</i>	<i>insignis</i>	DracIns1G	AY947719
<i>Dracunculus</i>	<i>medinensis</i>	DracMed1G	AY852268
<i>Dracunculus</i>	<i>medinensis</i>	DracMed2G	AY947720
<i>Dracunculus</i>	<i>oesophageus</i>	DracOes1G	AY852269
<i>Dracunculus</i>	sp.	DracSp1G	DQ503457
<i>Drilocephalobus</i>	1	DrilSp1	AY284678
<i>Drilocephalobus</i>	2	DrilSp2	AY284679
<i>Drilocephalobus</i>	3	DrilSp3	AY284680
<i>Dujardinascaris</i>	<i>waltoni</i>	DujaWal1G	EF180081
<i>Echinuria</i>	<i>borealis</i>	EchiBor1G	EF180064
<i>Ecphyadophora</i>	sp.	EcphSp	AY593917
<i>Ecphyadophora</i>	<i>tenuissima</i>	EcphTen1	EU669910
<i>Ecphyadophora</i>	<i>tenuissima</i>	EcphTen2	EU669911
<i>Ecumenicus</i>	1	EcumSp1	AY284781
<i>Ecumenicus</i>	2	EcumSp2	AY284782
<i>Ecumenicus</i>	<i>monohystera</i>	EcumMon1	AY284783
<i>Ecumenicus</i>	<i>monohystera</i>	EcumMon2	AY284784
<i>Enchodelus</i>	sp.	EnchSp3	EF207247
<i>Enchodelus</i>	sp. 1	EnchSp1	AY284792
<i>Enchodelus</i>	sp. 2	EnchSp2	AY284793
<i>Enoploides</i>	sp. 1	EPloSp1	FJ040490
<i>Enoplus</i>	<i>brevis</i>	EnopBreG	U88336
<i>Enoplus</i>	<i>meridionalis</i>	EnopMerG	Y16914
<i>Epidorylaimus</i>	<i>lugdunensis</i>	EpidLug1	AY284802
<i>Epidorylaimus</i>	<i>lugdunensis</i>	EpidLug2	AY284803
<i>Epidorylaimus</i>	sp. 1	EpidSp1	FJ040478
<i>Epsilonematidae</i>	sp. 1	Epsifamil1	EF591340
<i>Ethmolaimus</i>	<i>pratensis</i>	EthmPra2	FJ040475
<i>Ethmolaimus</i>	<i>pratensis</i> 1	EthmPra1	AY593942
<i>Eubostrichus</i>	<i>dianae</i>	EuboDiaG	Y16915
<i>Eubostrichus</i>	<i>parasitiferus</i>	EuboParG	Y16916
<i>Eubostrichus</i>	<i>toparius</i>	EuboTopG	Y16917
<i>Eucephalobus</i>	cf. <i>oxyuroides</i>	EuceOxy1Z	AY284664
<i>Eucephalobus</i>	<i>oxyuroides</i>	EuceOxy2	AY284665
<i>Eucephalobus</i>	<i>striatus</i>	EuceStr1	AY284666
<i>Eucephalobus</i>	<i>striatus</i>	EuceStr2	AY284667
<i>Eucoleus</i>	<i>dispar</i>	EucoDis1G	EU004821
<i>Eudorylaimus</i>	1	EudoSp1	AY284800

Table S1. (Continued).

Genus	Species	ID	GenBank No.
<i>Eudorylaimus</i>	<i>carteri</i>	EudoCar1G	AJ966484
<i>Eudorylaimus</i>	cf. <i>minutus</i>	EudoMin1Z	AY284794
<i>Eumonhystera</i>	cf. <i>simplex</i>	EumoSimZ	AY284692
<i>Eumonhystera</i>	<i>filiformis</i>	EumoFil2	AY593937
<i>Euteratocephalus</i>	<i>palustris</i>	EutePal1	AY284684
<i>Euteratocephalus</i>	sp.	EuteSp1	AY284685
<i>Fergusobia</i>	sp.	FergSp7G	AY589299
<i>Fergusobia</i>	sp.	FergSp10G	AY589302
<i>Fergusobia</i>	sp.	FergSp17G	EF011668
<i>Fictor</i>	sp. 1	FictSp1	FJ040437
<i>Filarinema</i>	<i>flagrifer</i>	FiNeFla1G	AJ920354
<i>Filaroides</i>	<i>martis</i>	FilaMar1G	AY295807
<i>Filenchus</i>	<i>filiformis</i> 1	FileFil1	AY284592
<i>Filenchus</i>	<i>thornei</i> 1	FileTh1	AY284591
<i>Geomonhystera</i>	<i>disjuncta</i>	GeomDis1G	AJ966485
<i>Geomonhystera</i>	sp. 3	GeomSp3	FJ040465
<i>Geomonhystera</i>	<i>villosa</i> 1	GeomVil1	EF591334
<i>Globodera</i>	<i>achilleae</i>	GlobAch1	FJ040399
<i>Globodera</i>	<i>artemisiae</i>	GlobArt1	FJ040400
<i>Globodera</i>	<i>pallida</i>	GlobPal1	AY284618
<i>Globodera</i>	<i>rostochiensis</i>	GlobRos4	AY593880
<i>Globodera</i>	<i>tabacum</i>	GlobTab4	FJ040401
<i>Gnathostoma</i>	<i>binucleatum</i>	GnatBinG	Z96946
<i>Gnathostoma</i>	<i>neoprocyonis</i>	GnatNeoG	Z96947
<i>Gnathostoma</i>	<i>turgidum</i>	GnatTurG	Z96948
<i>Goezia</i>	<i>pelagia</i>	GoezPelG	U94372
<b><i>Gordius</i></b>	<b><i>aquaticus</i></b>	<b>GordAquG</b>	<b>X80233</b>
<i>Granonchulus</i>	sp. 1	GranSp1	AY593953
<i>Haemonchus</i>	<i>contortus</i>	HaemConG	L04153
<i>Haemonchus</i>	<i>placei</i>	HaemPla1G	L04154
<i>Haemonchus</i>	<i>similis</i>	HaemSimG	L04152
<i>Haemonchus</i>	sp.	HaemSp1G	DQ503465
<i>Halalaimus</i>	sp. 1	HalaSp1	FJ040501
<i>Halenchus</i>	<i>fucicola</i>	HaleFuc1	EU669912
<i>Halicephalobus</i>	<i>gingivalis</i>	HaliGinG	AF202156
<i>Halichoanolaimus</i>	sp. 1	HaChSp1	EF591338
<i>Haliplectus</i>	<i>bickneri</i>	HaPlSp1	AY593935
<i>Halocercus</i>	<i>invaginatus</i>	HaloInv1G	AY295808
<i>Helicotylenchus</i>	<i>canadensis</i>	HeliCan1	AY284605
<i>Helicotylenchus</i>	<i>dihystera</i>	HeliDih1G	AJ966486
<i>Helicotylenchus</i>	<i>pseudorobustus</i>	HeliPse1	AY284606
<i>Helicotylenchus</i>	<i>varicaudatus</i>	HeliVar1G	EU306354
<i>Helicotylenchus</i>	<i>vulgaris</i>	HeliVul	AY284607
<i>Heligmosomoides</i>	<i>polygyrus</i>	HeSoPol1G	AJ920355
cf. <i>Helionema</i>	sp. 1	HelOSp1	EU669913
<i>Hemicriconemoides</i>	<i>pseudobrachyurus</i>	HCriPse1	AY284622
<i>Hemicriconemoides</i>	<i>pseudobrachyurus</i>	HCriPse2	AY284623
<i>Hemicriconemoides</i>	<i>pseudobrachyurus</i>	HCriPse3	AY284624
<i>Hemicycliophora</i>	<i>conida</i>	HemiCon1	EU669914
<i>Hemicycliophora</i>	<i>conida</i>	HemiCon1G	AJ966471

Table S1. (Continued).

Genus	Species	ID	GenBank No.
<i>Hemicycliophora</i>	<i>thienemanni</i>	HemiThi1	AY284628
<i>Hemicycliophora</i>	<i>thienemanni</i>	HemiThi2	AY284629
<i>Hemicycliophora</i>	<i>thienemanni</i>	HemiThi1G	EU306341
<i>Herpetostrongylus</i>	<i>pythonis</i>	HerpPyt1G	AJ920358
<i>Heterakis</i>	<i>gallarum</i>	HeRaGal1G	DQ503462
<i>Heterakis</i>	sp.	HeRaSpG	AF083003
<i>Heterocephalobus</i>	<i>elongatus</i>	HCepElo1	AY284668
<i>Heterocephalobus</i>	<i>elongatus</i>	HCepElo2	AY284669
<i>Heterocephalobus</i>	<i>elongatus</i>	HCepElo3	AY284670
<i>Heterocheilus</i>	<i>tunicatus</i>	HeChTunG	U94373
<i>Heterodera</i>	<i>avenae</i> 1	HeDeAve1	FJ040403
<i>Heterodera</i>	<i>betae</i>	HeDeBet1	FJ040404
<i>Heterodera</i>	<i>goettingiana</i>	HeDeGoe1	EU669915
<i>Heterodera</i>	<i>hordecalis</i>	HeDeHor1	FJ040405
<i>Heterodera</i>	<i>mani</i>	HeDeMan1	EU669916
<i>Heterodera</i>	<i>schachtii</i>	HeDeSch1	AY284617
<i>Heterodera</i>	<i>trifolii</i> 1	HeDeTri1	FJ040402
<i>Heterorhabditis</i>	<i>bacteriophora</i>	HeRhBac1G	AF036593
<i>Heterorhabditis</i>	<i>bacteriophora</i>	HeRhBac1	FJ040428
<i>Heterorhabditis</i>	<i>bacteriophora</i>	HeRhBac2	FJ040429
<i>Heterorhabditis</i>	<i>bacteriophora</i>	HeRhBac3	FJ040430
<i>Heterorhabditis</i>	<i>hepialius</i>	HeRhHepG	AF083004
<i>Heterorhabditis</i>	<i>marelatus</i>	HeRhMar1	FJ040431
<i>Heterorhabditis</i>	<i>megidis</i>	HeRhMeg1	FJ040432
<i>Heterorhabditis</i>	<i>megidis</i>	HeRhMeg9	FJ040433
<i>Heterorhabditis</i>	<i>megidis</i>	HeRhMeg15	FJ040434
<i>Heterorhabditis</i>	sp.	HeRhSp1	FJ040435
<i>Heterorhabditis</i>	<i>zealandica</i>	HeRhZea1G	AJ920368
<i>Heterorhabditoides</i>	<i>chongmingensis</i>	HRTtoCho1G	EF503692
<i>Hirschmanniella</i>	cf. <i>belli</i>	HirsBel1GZ	EF029856
<i>Hirschmanniella</i>	<i>gracilis</i>	HirsGra1	EU669959
<i>Hirschmanniella</i>	<i>loofi</i>	HirsLoo1G	EU306353
<i>Hirschmanniella</i>	<i>pomponiensis</i>	HirsPom1G	EF029854
<i>Hirschmanniella</i>	<i>santarosae</i>	HirsSan1G	EF029855
<i>Hirschmanniella</i>	sp.	HirsSp1G	EF029857
<i>Hirschmanniella</i>	sp. 1	HirsSp1	AY284614
<i>Hirschmanniella</i>	sp. 2	HirsSp2	AY284615
<i>Hirschmanniella</i>	sp. 3	HirsSp3	AY284616
<i>Hovorkonema</i>	<i>variegatum</i>	HovoVar1G	AY702705
<i>Howardula</i>	<i>aoronymphium</i>	HowaAor1G	AY589304
<i>Hypodontus</i>	<i>macropi</i>	HypoMac1G	AJ920339
<i>Hysterothylacium</i>	<i>fortalezae</i>	HystForG	U94374
<i>Hysterothylacium</i>	<i>pelagicum</i>	HystPelG	U94375
<i>Hysterothylacium</i>	<i>reliquens</i>	HystRelG	U94376
<i>Iheringascaris</i>	<i>inquires</i>	IhAsInqG	U94377
<i>Ironus</i>	<i>dentifurcatus</i>	IronDen1G	AJ966487
<i>Ironus</i>	<i>longicaudatus</i>	IronLon1	FJ040495
<i>Ironus</i>	sp.	IronSp1G	AY552970
<i>Ironus</i>	sp. 1	IronSp1	FJ040496
<i>Isolaimium</i>	sp.	IsolSp1G	AY552971
<i>Isolaimium</i>	sp. 1	IsolSp1	AY552971

**Table S1.** (Continued).

Genus	Species	ID	GenBank No.
<i>Kalicephalus</i>	<i>cristatus</i>	KaliCri1G	AJ920349
<i>Koerneria</i>	sp.	KoerSp1G	EU196025
<i>Labiostromyulus</i>	<i>bipapillosus</i>	LabiBip1G	AJ920337
<i>Labronema</i>	<i>ferox</i>	LabrFer1G	AY552972
<i>Labronema</i>	<i>vulvapapillatum</i>	LabrVul1	AY284807
<i>Laimaphelenchus</i>	<i>penardi</i>	LaimPen2	AY593919
<i>Laimaphelenchus</i>	<i>penardi</i>	LaimPen1G	EU306346
<i>Laimaphelenchus</i>	<i>penardi</i> 1	LaimPen1	AY593918
<i>Laxus</i>	<i>cosmopolitus</i>	LaxuCosG	Y16918
<i>Laxus</i>	<i>oneistus</i>	LaxuOneG	Y16919
<i>Leidynema</i>	<i>portentosae</i>	LeidPor1G	EF180073
<i>Lelenchus</i>	<i>leptosoma</i>	LeleLep1	AY284584
<i>Leptolaimus</i>	sp. 1	LeLaSp1	EF591323
<i>Leptolaimus</i>	sp. 2	LeLaSp2	EF591324
<i>Leptolaimus</i>	sp. 3	LeLaSp3	FJ040458
<i>Leptonchus</i>	<i>granulosus</i>	LeOnGra1	AY284831
<i>Leptonemella</i>	sp.	LeptSpG	Y16920
<i>Litomosoides</i>	<i>sigmodontis</i>	LitoSigG	AF227233
<i>Loa</i>	<i>loa</i>	LoaLoa1G	DQ094173
<i>Longidorella</i>	sp. 1	LoReSp1	AY284789
<i>Longidorella</i>	sp. 2	LoReSp2	AY284790
<i>Longidorus</i>	<i>africanus</i>	LoRuAfr1G	AY283164
<i>Longidorus</i>	<i>attenuatus</i>	LoRuAtt1G	AY687994
<i>Longidorus</i>	<i>biformis</i>	LoRuBif1G	AY283162
<i>Longidorus</i>	<i>biformis</i>	LoRuBif2G	AY283171
<i>Longidorus</i>	<i>breviannulatus</i>	LoRuBre1G	AY283161
<i>Longidorus</i>	cf. <i>intermedius</i>	LoRuIntZ	AY284816
<i>Longidorus</i>	<i>crassus</i>	LoRuCra1G	AY283158
<i>Longidorus</i>	<i>diadecturus</i>	LoRuDia1G	AY283166
<i>Longidorus</i>	<i>diadecturus</i>	LoRuDia2G	AY283167
<i>Longidorus</i>	<i>dunensis</i>	LoRuDun1	AY284817
<i>Longidorus</i>	<i>dunensis</i>	LoRuDun2	AY284818
<i>Longidorus</i>	<i>dunensis</i>	LoRuDun3	AY284819
<i>Longidorus</i>	<i>elongatus</i>	LoRuElo1G	AF036594
<i>Longidorus</i>	<i>elongatus</i>	LoRuElo2G	AY687992
<i>Longidorus</i>	<i>euonymus</i>	LoRuEuo1G	AY687995
<i>Longidorus</i>	<i>fragilis</i>	LoRuFra1G	AY283172
<i>Longidorus</i>	<i>grandis</i>	LoRuGra1G	AY283165
<i>Longidorus</i>	<i>litchii</i>	LoRuLit1G	AY687996
<i>Longidorus</i>	<i>macrosoma</i>	LoRuMac1G	AY580055
<i>Longidorus</i>	<i>paralongicaudatus</i>	LoRuPar1G	AY283160
<i>Longidorus</i>	<i>paravineacola</i>	LoRuPaV1G	AY283156
<i>Longidorus</i>	<i>paravineacola</i>	LoRuPaV2G	AY283157
<i>Longidorus</i>	<i>paravineacola</i>	LoRuPaV3G	AY283159
<i>Longidorus</i>	<i>piceicola</i>	LoRuPic1G	AY687993
<i>Longidorus</i>	sp.	LoRuSp1G	AY283163
<i>Longidorus</i>	sp.	LoRuSp2G	AY283168
<i>Longidorus</i>	<i>vineacola</i>	LoRuVin1G	AY283169
<i>Macrobiotus</i>	<i>hufelandi</i>	MaBiHufG	X81442
<i>Macrotriphurus</i>	<i>arbusticola</i>	MaTrArb2	AY284596

Table S1. (Continued).

Genus	Species	ID	GenBank No.
<i>Macrotrophurus</i>	<i>arbusticola</i> 1	MaTrArb1	AY284595
<i>Malenchus</i>	<i>andrassyi</i> 1	MaleAnd1	AY284587
<i>Margolisianum</i>	<i>bulbosum</i>	MargBul1G	AB185161
<i>Meloidogyne</i>	<i>arabica</i>	MeloAra1G	AY942625
<i>Meloidogyne</i>	<i>ardenensis</i>	MeloArd1	AY593894
<i>Meloidogyne</i>	<i>arenaria</i>	MeloAre1G	U42342
<i>Meloidogyne</i>	<i>artiellia</i>	MeloArt1G	AF248477
<i>Meloidogyne</i>	<i>chitwoodi</i>	MeloChi1	AY593883
<i>Meloidogyne</i>	<i>duytsi</i>	MeloDuyG	AF442197
<i>Meloidogyne</i>	<i>ethiopica</i>	MeloEth1G	AY942630
<i>Meloidogyne</i>	<i>exigua</i>	MeloExi2G	AY942627
<i>Meloidogyne</i>	<i>fallax</i>	MeloFal1	AY593895
<i>Meloidogyne</i>	<i>floridensis</i>	MeloFlo1G	AY942621
<i>Meloidogyne</i>	<i>graminicola</i>	MeloGraG	AF442196
<i>Meloidogyne</i>	<i>hapla</i>	MeloHap4	AY593892
<i>Meloidogyne</i>	<i>ichinohei</i>	MeloIch1	EU669953
<i>Meloidogyne</i>	<i>incognita</i>	MeloInc1	AY284621
<i>Meloidogyne</i>	<i>javanica</i>	MeloJav2G	AY268121
<i>Meloidogyne</i>	<i>mali</i>	MeloMal1	EU669948
<i>Meloidogyne</i>	<i>maritima</i>	MeloMar1	EU669944
<i>Meloidogyne</i>	<i>mayaguensis</i>	MeloMay1G	AY942629
<i>Meloidogyne</i>	<i>microtyla</i>	MeloMicG	AF442198
<i>Meloidogyne</i>	<i>minor</i>	MeloMin1	AY593899
<i>Meloidogyne</i>	<i>morocciensis</i>	MeloMor1G	AY942632
<i>Meloidogyne</i>	<i>naasi</i>	MeloNaa1	AY593900
<i>Meloidogyne</i>	<i>oryzae</i>	MeloOry1G	AY942631
<i>Meloidogyne</i>	<i>paranaensis</i>	MeloPar1G	AY942622
<i>Meloidogyne</i>	<i>spartinae</i>	MeloSpa1G	EF189177
<i>Meloidogyne</i>	<i>ulmi</i>	MeloUlm1	EU669947
<i>Merlinius</i>	<i>brevicens</i>	MerlBre1	AY284597
<i>Mermis</i>	<i>nigrescens</i>	MermNigG	AF036641
<i>Mermiidae</i>	sp. 2	Mermfamil2	FJ040480
<i>Mermiidae</i>		Mermfamil	AY284743
<i>Mesocriconema</i>	<i>xenoplax</i>	MCriXen1	AY284625
<i>Mesocriconema</i>	<i>xenoplax</i>	MCriXen2	AY284626
<i>Mesocriconema</i>	<i>xenoplax</i>	MCriXen3	AY284627
<i>Mesodorylaimus</i>	<i>aberrans</i>	MesDAbe1	AY593947
<i>Mesodorylaimus</i>	<i>bastiani</i>	MesDBas1G	AJ966488
<i>Mesodorylaimus</i>	<i>centrocercus</i>	MesDCen1	AY284799
<i>Mesodorylaimus</i>	<i>centrocercus</i>	MesDCen2	EF207248
<i>Mesodorylaimus</i>	cf. <i>nigritulus</i>	MesDNig1ZG	AJ966490
<i>Mesodorylaimus</i>	<i>japonicus</i>	MesDJap1G	AJ966489
<i>Mesodorylaimus</i>	sp. 1	MesDSp1	AY284780
<i>Mesorhabditis</i>	2	MRhaSp2	AY284660
<i>Mesorhabditis</i>	<i>anisomorpha</i>	MRhaAniG	AF083013
<i>Mesorhabditis</i>	<i>longespiculosa</i>	MRhaLon1G	EU196014
<i>Mesorhabditis</i>	<i>n</i>	MRhaSp4	AY593922
<i>Mesorhabditis</i>	<i>scanica</i>	MRhaScaG	AF083014
<i>Mesorhabditis</i>	sp.	MRhaSp1G	U73452
<i>Mesotheristus</i>	<i>setosus</i>	MTheSet2G	AM234045



Table S1. (Continued).

Genus	Species	ID	GenBank No.
<i>Metachromadora</i>	<i>remanei</i>	MAchRem2G	AM234620
<i>Metachromadora</i>	sp.	MAchSpG	AF036595
<i>Metachromadora</i>	sp. 1	MAchSp1	EF591339
<i>Metachromadora</i>	sp. 2	MAchSp2	FJ040469
<i>Metadesmolaimus</i>	sp.	MDesSp1G	AJ966491
<i>Metaporcelaimus</i>	<i>simplex</i>	MApoSim1	AY593948
<i>Metastrongylus</i>	<i>elongatus</i>	MStrElo1G	AJ920363
<i>Metastrongylus</i>	<i>salmi</i>	MStrSal1G	AY295809
<i>Metateratocephalus</i>	<i>crassidens</i>	MTCeCra1	AY284686
<i>Metateratocephalus</i>	<i>crassidens</i>	MTCeCra2	AY284687
<i>Metateratocephalus</i>	<i>crassidens</i>	MTCeCra3	AY593934
<i>Miconchus</i>	cf. <i>fasciatus</i>	MicoFas1ZG	AY552973
<i>Microdorylaimus</i>	<i>miser</i>	MicDMis1	AY284804
<i>Microdorylaimus</i>	<i>modestus</i>	MicDMod1	AY284805
<i>Microdorylaimus</i>	<i>modestus</i>	MicDMod2	AY284806
<i>Microdorylaimus</i>	sp.	MicDSp1G	AJ966492
<i>Micropleura</i>	<i>australiensis</i>	MicPAus1G	DQ442678
<i>Microtetrameres</i>	<i>cloacitectus</i>	MicTClo1G	EU004814
<i>Molnaria</i>	<i>intestinalis</i>	MolnInt1G	DQ442668
<i>Monhystera</i>	<i>riemanni</i>	MonhRie1	AY593938
<i>Mononchoides</i>	<i>striatus</i> 1	MonEStr1	AY593924
<i>Mononchus</i>	<i>aquaticus</i>	MonCAqu1	AY284764
<i>Mononchus</i>	<i>aquaticus</i>	MonCAqu2	AY284765
<i>Mononchus</i>	<i>aquaticus</i>	MonCAqu1G	AY297821
<i>Mononchus</i>	<i>truncatus</i>	MonCTru1	AY284762
<i>Mononchus</i>	<i>truncatus</i>	MonCTru1G	AJ966493
<i>Mononchus</i>	<i>tunbridgensis</i>	MonCTun1	AY284763
<i>Mononchus</i>	<i>tunbridgensis</i>	MonCTun2	AY593954
<i>Monoposthia</i>	sp. 1	MonPSP1	FJ040505
<i>Muellerius</i>	<i>capillaris</i>	MuelCap1G	AY295810
<i>Myctolaimus</i>	<i>ulmi</i>	MyctUlm1G	EU196024
<i>Mylonchulus</i>	1	MyloSp1	AY284758
<i>Mylonchulus</i>	2	MyloSp2	AY284759
<i>Mylonchulus</i>	3	MyloSp3	AY284760
<i>Mylonchulus</i>	4	MyloSp4	AY284761
<i>Mylonchulus</i>	<i>arenicolus</i>	MyloAreG	AF036596
<i>Mylonchulus</i>	<i>brachyuris</i>	MyloBra2	AY284752
<i>Mylonchulus</i>	<i>brachyuris</i>	MyloBra3	AY284753
<i>Mylonchulus</i>	<i>brachyuris</i>	MyloBra4	AY284754
<i>Mylonchulus</i>	<i>rotundicaudatus</i>	MyloRot2	AY284751
<i>Mylonchulus</i>	<i>sigmaturus</i>	MyloSig1	AY284755
<i>Mylonchulus</i>	<i>sigmaturus</i>	MyloSig2	AY284756
<i>Mylonchulus</i>	<i>sigmaturus</i>	MyloSig3	AY284757
<i>Mylonchulus</i>	sp.	MyloSp1G	AJ875156
<i>Myolaimus</i>	sp.	MyolSpG	U81585
<i>Nacobbus</i>	<i>aberrans</i>	NacoAbe1G	AF442190
<i>Nacobbus</i>	<i>aberrans</i>	NacoAbe2G	AJ966494
<i>Nagelus</i>	<i>obscurus</i>	NageObs1G	EU306350
<i>Nagelus</i>	<i>obscurus</i> 1	NageObs1	AY593904
<i>Nanidorus</i>	<i>minor</i>	NaniMin1G	AJ438052

Table S1. (Continued).

Genus	Species	ID	GenBank No.
<i>Nanidorus</i>	<i>minor</i>	NaniMin2G	AJ438053
<i>Nanidorus</i>	<i>minor</i>	NaniMin3G	AJ438054
<i>Nanidorus</i>	<i>minor</i>	NaniMin4G	AJ438055
<i>Nanidorus</i>	<i>minor</i>	NaniMin5G	AJ438056
<i>Nanidorus</i>	<i>minor</i>	NaniMin6G	AJ438057
<i>Nanidorus</i>	<i>minor</i>	NaniMin7G	AJ438058
<i>Nanidorus</i>	<i>minor</i>	NaniMin8G	AJ439571
<i>Nanidorus</i>	<i>minor</i>	NaniMin9G	AM269897
<i>Nanidorus</i>	<i>nanus</i>	NaniNan1	FJ040485
<i>Nanidorus</i>	<i>nanus</i>	NaniNan2	FJ040486
<i>Necator</i>	<i>americanus</i>	NecaAme1G	AY295811
<i>Necator</i>	<i>americanus</i>	NecaAme2G	AJ920348
<i>Nematodirus</i>	<i>battus</i>	NemaBat1G	U01230
<i>Nematodirus</i>	<i>battus</i>	NemaBat2G	AJ920360
<i>Nemhelix</i>	<i>bakeri</i>	NemhBak1G	DQ118537
<i>Neoscarophis</i>	<i>macrouri</i>	NeoaMac1G	DQ442660
<i>Neodolichorhynchus</i>	<i>lamelliferus</i> 1	NeodLam1	AY284598
<i>Neodolichorhynchus</i>	<i>microphasmis</i>	NeodMic2	EU669917
<i>Neopsilenchus</i>	<i>magnidens</i> 1	Neopmag1	AY284585
<i>Nicollina</i>	<i>cameroni</i>	NicoCam1G	AJ920357
<i>Nilonema</i>	<i>senticosum</i>	NiloSen1G	DQ442671
<i>Nippostrongylus</i>	<i>brasiliensis</i>	NippBra1G	AF036597
<i>Nippostrongylus</i>	<i>brasiliensis</i>	NippBra2G	AJ920356
<i>Nothotylenchus</i>	<i>acris</i> 1	NothAcr1	AY593914
<i>Nygolaimus</i>	cf. <i>brachyuris</i>	NygoBra1Z	AY284770
<i>Nygolaimus</i>	cf. <i>brachyuris</i>	NygoBra2Z	AY284771
<i>Nygolaimus</i>	cf. <i>parvus</i>	NygoPar1ZG	AY552974
<i>Odontolaimus</i>	<i>chlorurus</i>	OdLaChl	AY284723
<i>Odontopharynx</i>	<i>longicaudata</i>	OdPhLon1	FJ040449
<i>Odontophora</i>	sp. 1	OdonSp1	FJ040459
<i>Ogma</i>	<i>cobbi</i>	OgmaCob1	EU669918
<i>Ogma</i>	<i>menzeli</i>	OgmaMen1	EU669919
<i>Onchium</i>	sp. 1	OChiSp1	EF59138
<i>Onchocerca</i>	<i>cervicalis</i>	OnCeCer1G	DQ094174
<i>Onchocercidae</i>	sp.	OnCefamil1G	DQ103704
<i>Oncholaimidae</i>	sp. 1	Onchfamil1	FJ040493
<i>Opisthodorylaimus</i>	<i>sylphoides</i>	OpiDSyl1	AY284785
<i>Oscheius</i>	<i>dolichurus</i>	OschDoA1G	EU196010
<i>Oscheius</i>	<i>dolichuroides</i>	OschDoI1G	AF082998
<i>Oscheius</i>	<i>guentheri</i>	OschGue1G	EU196022
<i>Oscheius</i>	<i>insectivorus</i>	OschInsG	AF083019
<i>Oscheius</i>	<i>myriophila</i>	OschMyr1G	U81588
<i>Oscheius</i>	<i>myriophila</i>	OschMyr2G	U13936
<i>Oscheius</i>	sp.	OschSp1G	AF082995
<i>Oscheius</i>	sp.	OschSp2G	AF082994
<i>Oscheius</i>	<i>tipulae</i>	OschTip1G	U81587
<i>Oscheius</i>	<i>tipulae</i>	OschTip2G	AF036591
<i>Oscheius</i>	<i>tipulae</i>	OschTip3G	EU196009
<i>Oslerus</i>	<i>osleri</i>	OsleOsl1G	AY295812
<i>Ostertagia</i>	<i>leptospicularis</i>	OsteLep1G	AJ920351

**Table S1.** (Continued).

Genus	Species	ID	GenBank No.
<i>Ostertagia</i>	<i>ostertagi</i>	OsteOst1G	AF036598
<i>Ostertagia</i>	<i>ostertagi</i>	OsteOst2G	AJ920352
<i>Otostrongylus</i>	<i>circumlitus</i>	OtosCir1G	AY295813
<i>Otostrongylus</i>	sp.	OtosSpG	U81589
<i>Ottolenchus</i>	<i>discrepans</i> 1	OttoDis1	AY284590
<i>Oxydirus</i>	<i>nethus</i>	OxydNet3	EF207251
<i>Oxydirus</i>	<i>oxycephaloides</i>	OxydOCO1	AY284823
<i>Oxydirus</i>	<i>oxycephalus</i>	OxydOxy1	AY284824
<i>Oxydirus</i>	<i>oxycephalus</i>	OxydOxy2	AY284825
<i>Oxystomina</i>	sp. 1	OxysSp1	FJ040498
<i>Oxystomina</i>	sp. 2	OxysSp2	FJ040499
<i>Oxyuris</i>	<i>equi</i>	OxyuEqu1G	EF180062
<i>Panagrellus</i>	<i>redivivus</i>	PGreRed1G	AF036599
<i>Panagrellus</i>	<i>redivivus</i>	PGreRed2G	AF083007
<i>Panagrobelus</i>	<i>stammeri</i>	PGBeStaG	AF202153
<i>Panagrolaimoid</i>		RhPhSp2GZ	U81580
<i>Panagrolaimus</i>	<i>cf. rigidus</i>	PGLaRig1GZ	DQ285636
<i>Panagrolaimus</i>	<i>dauidi</i>	PGLaDavG	AJ567385
<i>Panagrolaimus</i>	<i>paetzoldi</i>	PGLaPae1	FJ040414
<i>Panagrolaimus</i>	sp.	PGLaSpG	U81579
<i>Panagrolaimus</i>	<i>subelongatus</i> 1	PGLaSub1	AY284681
<i>Paracanthochus</i>	<i>caecus</i>	PCanCaeG	AF047888
<i>Paractinolaimus</i>	<i>macrolaimus</i>	PActMac1	AY284826
<i>Paractinolaimus</i>	<i>macrolaimus</i>	PActMac1G	AY993978
<i>Paractinolaimus</i>	sp.	PActSp1G	AY552975
<i>Paracyatholaimus</i>	<i>intermedius</i>	PCyaInt1G	AJ966495
<i>Parafilaroides</i>	<i>decorus</i>	PFilDec1G	AY295814
<i>Parafilaroides</i>	sp.	PFilSpG	U81590
<i>Paralongidorus</i>	<i>litoralis</i>	PLonLit1G	EU026158
<i>Paralongidorus</i>	<i>litoralis</i>	PLonLit2G	EU026159
<i>Paralongidorus</i>	<i>maximus</i>	PLonMax1G	AJ875152
<i>Paralongidorus</i>	<i>paramaximus</i>	PLonPar1G	EU026157
<i>Paramphidelus</i>	1	PAmpSp1	AY284740
<i>Paramphidelus</i>	2	PAmpSp2	AY284741
<i>Paramphidelus</i>	3	PAmpSp3	AY284742
<i>Paramphidelus</i>	<i>hortensis</i>	PAmpHor1	AY284739
<i>Paraphelenchus</i>	sp.	PAphSp	AY284642
<i>Paraplectonema</i>	<i>pedunculatum</i>	PaPIPed1	EF591320
<i>Parascaris</i>	<i>equorum</i>	PAscEquG	U94378
<i>Parasitorhabditis</i>	<i>obtusa</i>	PaRhObt1G	EU003189
<i>Parasitorhabditis</i>	sp.	PaRhSpG	AF083028
<i>Paraspidodera</i>	sp.	PSpiSpG	AF083005
<i>Paratrichodorus</i>	<i>allius</i>	PtrCAI1G	AJ439572
<i>Paratrichodorus</i>	<i>allius</i>	PtrCAI2G	AJ439623
<i>Paratrichodorus</i>	<i>allius</i>	PtrCAI3G	AM067124
<i>Paratrichodorus</i>	<i>allius</i>	PtrCAI4G	AJ439569
<i>Paratrichodorus</i>	<i>allius</i>	PtrCAI5G	AM269895
<i>Paratrichodorus</i>	<i>anemones</i>	PtrCAne1G	AF036600
<i>Paratrichodorus</i>	<i>anemones</i>	PtrCAne2G	AJ439573
<i>Paratrichodorus</i>	<i>anemones</i>	PtrCAne3G	AJ439570

Table S1. (Continued).

Genus	Species	ID	GenBank No.
<i>Paratrichodorus</i>	<i>hispanus</i>	PTrCHis1G	AJ439577
<i>Paratrichodorus</i>	<i>macrostylus</i>	PTrCMac1G	AJ439507
<i>Paratrichodorus</i>	<i>macrostylus</i>	PTrCMac2G	AJ439621
<i>Paratrichodorus</i>	<i>macrostylus</i>	PTrCMac3G	AJ439622
<i>Paratrichodorus</i>	<i>pachydermus</i>	PTrCPac1	FJ040483
<i>Paratrichodorus</i>	<i>pachydermus</i>	PTrCPac1G	AF036601
<i>Paratrichodorus</i>	<i>pachydermus</i>	PTrCPac2G	AJ439512
<i>Paratrichodorus</i>	<i>pachydermus</i>	PTrCPac3G	AJ439574
<i>Paratrichodorus</i>	sp.	PTrCSp2G	AJ439576
<i>Paratrichodorus</i>	<i>teres</i>	PTrCTer1	FJ040484
<i>Paratrichodorus</i>	<i>teres</i>	PTrCTer1G	AJ439575
<i>Paratrichodorus</i>	<i>teres</i>	PTrCTer2G	AM087125
<i>Paratrichodorus</i>	<i>teres</i>	PTrCTer3G	AM269896
<i>Paratylenchus</i>	cf. <i>neoamblycephalus</i>	PTylNeoZ	AY284634
<i>Paratylenchus</i>	<i>dianthus</i>	PTylDia1G	AJ966496
<i>Paratylenchus</i>	<i>microdorus</i>	PTylMic1	AY284632
<i>Paratylenchus</i>	<i>microdorus</i>	PTylMic2	AY284633
<i>Paratylenchus</i>	<i>straeleni</i>	PTylStr2	AY284630
<i>Paratylenchus</i>	<i>straeleni</i>	PTylStr3	AY284631
<i>Paravulvulus</i>	<i>hartingii</i>	PVulHar1	AY284774
<i>Paravulvulus</i>	<i>hartingii</i>	PVulHar2	AY284775
<i>Paravulvulus</i>	<i>hartingii</i>	PVulHar1G	AY552976
<i>Paraxonchium</i>	<i>laetificans</i>	PAxoLae1	AY284808
<i>Paraxonchium</i>	<i>laetificans</i>	PAxoLae2	AY284809
<i>Paraxonchium</i>	<i>laetificans</i>	PAxoLae3	AY284810
<i>Parelaphostrongylus</i>	<i>odocoilei</i>	PElaOdo1G	AY295815
<i>Passalurus</i>	sp.	PassSp1G	EF180061
<i>Pellioiditis</i>	<i>marina</i>	PellMarG	AF083021
<i>Pellioiditis</i>	<i>mediterranea</i>	PellMedG	AF083020
<i>Pellioiditis</i>	sp.	PellSp1G	EU196011
<i>Pellioiditis</i>	<i>typica</i>	PellTyp1G	U13933
<i>Pelodera</i>	<i>cylindrica</i>	PeloCyl1G	EU196021
<i>Pelodera</i>	<i>pseudoteres</i>	PeloPse1G	EU196023
<i>Pelodera</i>	<i>punctata</i>	PeloPunG	AF083018
<i>Pelodera</i>	<i>strongyloides</i>	PeloStrG	U13932
<i>Pelodera</i>	<i>teres</i>	PeloTerG	AF083002
<i>Petrovinema</i>	<i>poculatum</i>	PetrPoc1G	AJ920343
<i>Phasmarhabditis</i>	<i>hermaphrodita</i>	PhasHer1G	DQ639980
<i>Phasmarhabditis</i>	<i>hermaphrodita</i>	PhasHer2G	DQ639981
<i>Phasmarhabditis</i>	<i>hermaphrodita</i>	PhasHer3G	DQ639980
<i>Phasmarhabditis</i>	sp.	PhasSp1G	EU196008
<i>Philometra</i>	<i>cyprinirutili</i>	PhiMCyp1G	DQ442675
<i>Philometra</i>	<i>obturans</i>	PhiMObt1G	AY852267
<i>Philometra</i>	<i>ovata</i>	PhiMOva1G	DQ442677
<i>Philometra</i>	sp.	PhiMSp1G	DQ442674
<i>Philometroides</i>	<i>sanguineus</i>	PhMOSan1G	DQ442676
<i>Philonema</i>	<i>oncorhynchi</i>	PhilOnc1G	DQ442670
<i>Philonema</i>	sp.	PhilSpG	U81574
<i>Physaloptera</i>	<i>alata</i>	PhysAla1G	AY702703
<i>Physaloptera</i>	<i>apivori</i>	PhysApi1G	EU004817

Table S1. (Continued).

Genus	Species	ID	GenBank No.
<i>Physaloptera</i>	sp.	PhysSp1G	EF180065
<i>Physaloptera</i>	<i>turgida</i>	PhysTur1G	DQ503459
<i>Plectid</i>		Plectfamil2G	AJ966508
<i>Plectidae</i>	sp.	Plectfamil1G	AJ966478
<i>Plectonchus</i>	<i>hunti</i>	PIChHunG	AF202154
<i>Plectonchus</i>	sp. 1	PIChSp2	AY593920
<i>Plectus</i>	<i>acuminatus</i>	PlecAcuG	AF037628
<i>Plectus</i>	<i>aquatilis</i>	PlecAqu1G	AF036602
<i>Plectus</i>	<i>aquatilis</i> 1	PlecAqu1	AY284700
<i>Plectus</i>	cf. <i>cirratus</i>	PlecCir1Z	AY284701
<i>Plectus</i>	cf. <i>parietinus</i>	PlecPRi1Z	AY284702
<i>Plectus</i>	cf. <i>parietinus</i>	PlecPRi2Z	AY284703
<i>Plectus</i>	cf. <i>parvus</i>	PlecParZ	AY284699
<i>Plectus</i>	cf. <i>pusillus</i>	PlecPus1Z	AY284704
<i>Plectus</i>	cf. <i>pusillus</i>	PlecPus2Z	AY284705
<i>Plectus</i>	<i>cirratus</i> 1	PlecCir2	AY593930
<i>Plectus</i>	<i>rhizophilus</i>	PlecRhi1	AY593928
<i>Plectus</i>	<i>rhizophilus</i>	PlecRhi2	AY593929
<i>Plectus</i>	sp.	PlecSpG	U61761
<i>Podura</i>	<i>aquatica</i>	PoduAquG	AF005452
<i>Poikilolaimus</i>	<i>centrocercus</i> 1	PoikCen1	FJ040436
<i>Poikilolaimus</i>	<i>oxycerca</i>	PoikOxyG	AF083023
<i>Poikilolaimus</i>	sp.	PoikSp1G	U81583
<i>Poikilolaimus</i>	sp.	PoikSp2G	DQ385848
<i>Polydesmus</i>	<i>coriaceus</i>	PolyCorG	AF005449
<i>Pontonema</i>	<i>vulgare</i>	PontVulG	AF047890
<i>Porrocaecum</i>	<i>angusticolle</i>	PorrAng1G	EU004820
<i>Porrocaecum</i>	<i>depressum</i>	PorrDepG	U94379
<i>Porrocaecum</i>	<i>streperae</i>	PorrStr1G	EF180074
<i>Praeacanthochus</i>	sp.	PraeSp1G	AF036612
<i>Praeacanthochus</i>	sp.	PraeSp2G	AM234046
<i>Pratylenchoides</i>	<i>magnicauda</i>	PrChMagG	AF202157
<i>Pratylenchoides</i>	<i>ritteri</i>	PrChRit1G	AJ966497
<i>Pratylenchus</i>	<i>convallariae</i>	PratCon1	EU669957
<i>Pratylenchus</i>	<i>crenatus</i>	PratCre1	AY284610
<i>Pratylenchus</i>	<i>crenatus</i>	PratCre2	EU669920
<i>Pratylenchus</i>	<i>crenatus</i>	PratCre3	EU669921
<i>Pratylenchus</i>	<i>crenatus</i>	PratCre4	EU669922
<i>Pratylenchus</i>	<i>goodeyi</i>	PratGoo1G	AJ966498
<i>Pratylenchus</i>	<i>neglectus</i>	PratNeg1	EU669923
<i>Pratylenchus</i>	<i>neglectus</i>	PratNeg2	EU669924
<i>Pratylenchus</i>	<i>penetrans</i>	PratPen1	EU669925
<i>Pratylenchus</i>	<i>penetrans</i>	PratPen2	EU669926
<i>Pratylenchus</i>	<i>pratensis</i>	PratPra1	AY284611
<i>Pratylenchus</i>	<i>scribneri</i>	PratScr1	EU669927
<i>Pratylenchus</i>	<i>scribneri</i>	PratScr2	EU669958
<i>Pratylenchus</i>	<i>thornei</i>	PratTho2G	AJ966499
<i>Pratylenchus</i>	<i>thornei</i>	PratTho1	AY284612
<i>Pratylenchus</i>	<i>thornei</i>	PratTho3	EU669928
<i>Pratylenchus</i>	<i>thornei</i>	PratTho4	AJ966499

Table S1. (Continued).

Genus	Species	ID	GenBank No.
<i>Pratylenchus</i>	<i>thornei</i>	PratTho5	EU669930
<i>Pratylenchus</i>	<i>vulnus</i>	PratVul1	EU669955
<i>Pratylenchus</i>	<i>vulnus</i>	PratVul2	EU669956
<i>Priapulus</i>	<i>caudatus</i>	PriaCauG	Z38009
<i>Prionchulus</i>	<i>muscorum</i>	PrioMus1	AY284745
<i>Prionchulus</i>	<i>muscorum</i>	PrioMus1G	AJ966500
<i>Prionchulus</i>	<i>punctatus</i>	PrioPun1	AY284746
<i>Prionchulus</i>	<i>punctatus</i>	PrioPun2	AY284747
<i>Prismatolaimus</i>	cf. <i>dolichurus</i>	PriMDol1Z	AY284727
<i>Prismatolaimus</i>	cf. <i>dolichurus</i>	PriMDol2Z	AY284728
<i>Prismatolaimus</i>	<i>dolichurus</i>	PriMDol4	AY593957
<i>Prismatolaimus</i>	<i>intermedius</i>	PriMInt1	AY284729
<i>Prismatolaimus</i>	<i>intermedius</i>	PriMInt1G	AF036603
<i>Pristionchus</i>	<i>aerivora</i>	PriTAer1	FJ040440
<i>Pristionchus</i>	<i>americanus</i>	PriTAME1	FJ040445
<i>Pristionchus</i>	<i>entomophagus</i>	PriTEnt1	FJ040441
<i>Pristionchus</i>	<i>lherithieri</i>	PriTLhe2G	AF036643
<i>Pristionchus</i>	<i>lheritieri</i>	PriTLhe1G	AF036640
<i>Pristionchus</i>	<i>lheritieri</i>	PriTLhe4	FJ040439
<i>Pristionchus</i>	<i>lheritieri</i> 1	PriTLhe1	AY284690
<i>Pristionchus</i>	<i>lheritieri</i> 2	PriTLhe3	AY593923
<i>Pristionchus</i>	<i>marianneae</i>	PriTMar1	FJ040442
<i>Pristionchus</i>	<i>maupasi</i>	PriTMau1	FJ040443
<i>Pristionchus</i>	<i>pacificus</i>	PriTPac1G	U81584
<i>Pristionchus</i>	<i>pacificus</i>	PriTPac2G	AF083010
<i>Pristionchus</i>	<i>pauli</i>	PriTPau1	FJ040446
<i>Pristionchus</i>	<i>pseudaerivorus</i>	PriTPse1	FJ040447
<i>Pristionchus</i>	<i>uniformis</i>	PriTUni1	FJ040444
<i>Procamacolaimus</i>	sp. 1	PrCoSp1	EF591326
<i>Procamallanus</i>	<i>pacificus</i>	PrCaPac1G	DQ442665
<i>Procamallanus</i>	<i>pintoi</i>	PrCaPin1G	DQ442666
<i>Procamallanus</i>	<i>rebecae</i>	PrCaReb1G	DQ442667
<i>Prodesmodora</i>	<i>circulata</i> 4	PrDeCir4	AY284722
<i>Prodesmodora</i>	sp. 2	PrDeSp2	FJ040476
<i>Prodesmodora</i>	sp. 3	PrDeSp3	FJ040477
<i>Prodontorhabditis</i>	<i>wirthi</i>	PDRhWir1G	AY602179
<i>Prodorylaimus</i>	<i>mas</i>	PrDoMas1	AY593946
<i>Prodorylaimus</i>	sp.	PrDoSp1	EF207246
cf. <i>Prodorylaimus</i>		PrDoSp2Z	AY284778
<i>Protorhabditis</i>	sp.	PRhaSp2G	AF083024
<i>Protorhabditis</i>	sp.	PRhaSp1G	AF083001
<i>Protorhabditis</i>	sp.	PRhaSp3G	EU196002
<i>Protostrongylus</i>	<i>rufescens</i>	PrStRuf1G	AJ920364
<i>Protozoophaga</i>	<i>obesa</i>	PrZoObe1G	EF180075
<i>Pseudacrobeles</i>	<i>variabilis</i>	PseAVarG	AF202150
<i>Pseudalius</i>	<i>inflexus</i>	PsAllnf1G	AY295816
<i>Pseudhalenchus</i>	<i>minutus</i>	PseHMin2	AY593916
<i>Pseudhalenchus</i>	<i>minutus</i> 1	PseHMin1	AY284638
<i>Pseudoterranova</i>	<i>decepiens</i>	PseTDecG	U94380
<i>Psilenchus</i>	cf. <i>hilarulus</i>	PsilHilZ	AY284593

Table S1. (Continued).

Genus	Species	ID	GenBank No.
<i>Ptycholaimellus</i>	sp. 1	PtycSp1	FJ040472
<i>Punctodera</i>	<i>stonei</i>	PuncSto1	EU682391
<i>Pungentus</i>	<i>silvestris</i>	PungSil1	AY284788
<i>Pungentus</i>	sp.	PungSp1	AY284791
<i>Pungentus</i>	sp.	PungSp1G	AJ966501
<i>Pycnophyes</i>	<i>kielensis</i>	PycnKieG	U67997
<i>Radopholus</i>	<i>similis</i>	RadoSim1G	AJ966502
<i>Radopholus</i>	sp. 2	RadoSp2	FJ040398
<i>Raillietnema</i>	sp.	RailSp1G	DQ503461
<i>Raphidascaris</i>	<i>acus</i>	RaphAcu1G	DQ503460
<i>Rhabditella</i>	<i>axei</i>	RhTeAxe2	AY284654
<i>Rhabditella</i>	<i>axei</i>	RhTeAxe1G	U13934
<i>Rhabditella</i>	sp.	RhTeSpG	AF083000
<i>Rhabditis</i>	<i>blumi</i>	RhabBluG	U13935
<i>Rhabditis</i>	<i>brassicae</i>	RhabBra1G	EU196006
<i>Rhabditis</i>	cf. <i>terricola</i>	RhabTerZ	AY284653
<i>Rhabditis</i>	<i>colombiana</i>	RhabCol1G	AY751546
<i>Rhabditis</i>	sp.	RhabSp1G	AF083008
<i>Rhabditis</i>	sp.	RhabSp2G	EU196007
<i>Rhabditis</i>	sp.	RhabSp3G	EU273597
<i>Rhabditis</i>	sp.	RhabSp4G	EU196004
<i>Rhabditoid</i>	sp.	Rhabditoid1G	EU196015
<i>Rhabditoides</i>	<i>inermiformis</i>	RhToInFG	AF083017
<i>Rhabditoides</i>	<i>inermis</i>	RhToIneG	AF082996
<i>Rhabditoides</i>	<i>regina</i>	RhToRegG	AF082997
<i>Rhabditophanes</i>	sp.	RhPhSp1G	AF202151
<i>Rhabdochona</i>	<i>denudata</i>	RhChDen1G	DQ442659
<i>Rhabdolaimus</i>	cf. <i>terrestris</i>	RhDoTer1Z	AY284710
<i>Rhabdolaimus</i>	cf. <i>terrestris</i>	RhDoTer2Z	AY284711
<i>Rhabdolaimus</i>	cf. <i>terrestris</i>	RhDoTer3Z	AY284712
<i>Rhigonema</i>	<i>thysanophora</i>	RhigThy1G	EF180067
<i>Robbea</i>	<i>hypermnestra</i>	RobbHypG	Y16921
<i>Rondonia</i>	<i>rondoni</i>	RondRon1G	DQ442679
<i>Rotylenchulus</i>	<i>reniformis</i>	RoChRen1G	EU306342
<i>Rotylenchus</i>	<i>goodeyi</i>	RotyGoo1	AY284609
<i>Rotylenchus</i>	<i>robustus</i>	RotyRob1G	AJ966503
<i>Rotylenchus</i>	sp. 1	RotySp	AY284608
<i>Rotylenchus</i>	<i>uniformis</i>	RotyUni1	AY593882
<i>Rotylenchus</i>	<i>uniformis</i>	RotyUni1G	EU306356
<i>Sabatieria</i>	<i>pulchra</i>	SabaPul1	EF591335
<i>Sabatieria</i>	<i>pulchra</i>	SabaPul2	FJ040466
<i>Sauertylenchus</i>	<i>maximus</i>	SaueMax2	AY284603
<i>Sauertylenchus</i>	<i>maximus</i>	SaueMax3	AY284604
<i>Sauertylenchus</i>	<i>maximus</i>	SaueMax1G	AY993979
<i>Sauertylenchus</i>	<i>maximus</i> 1	SaueMax1	AY284602
<i>Schistonchus</i>	<i>aureus</i>	SchiAur1G	DQ912922
<i>Schistonchus</i>	<i>centerae</i>	SchiCen1G	DQ912923
<i>Schistonchus</i>	<i>guangzhouensis</i>	SchiGua1G	DQ912924
<i>Scutellonema</i>	<i>bradys</i>	ScNeBra1G	AY271723
<i>Scutellonema</i>	<i>bradys</i>	ScNeBra2G	AJ966504

Table S1. (Continued).

Genus	Species	ID	GenBank No.
<i>Scutylenchus</i>	<i>quadrifer</i>	ScutQua1	AY284599
<i>Scutylenchus</i>	<i>quadrifer</i>	ScutQua1G	AY993977
<i>Sectonema</i>	<i>barbatoides</i>	SectBar1	AY284814
<i>Sectonema</i>	sp. 1	SectSp1	AY284815
<i>Seinura</i>	sp. 1	SeinSp1	AY284651
<i>Serratospiculum</i>	<i>tendo</i>	SerrTen1G	AY702704
<i>Setaria</i>	<i>digitata</i>	SetaDig1G	DQ094175
<i>SetoStephanolaimus</i>	<i>spartinae</i>	SeStSpa1	EF591321
<i>Skrijabillanus</i>	<i>scardinii</i>	SkLaSca1G	DQ442669
<i>Skrijabinema</i>	sp.	SkNeSp1G	EF180060
<i>Skrijabingylus</i>	<i>chitwoodorum</i>	SkriChi1G	AY295819
<i>Soboliphyme</i>	<i>baturini</i>	SoboBat1G	AY277895
<i>Solididens</i>	<i>vulgaris</i>	SoliVul1G	AY552977
<i>Sphaerularia</i>	<i>bombi</i>	SpRuBom1G	AB250212
<i>Sphaerularia</i>	<i>bombi</i>	SpRuBom2G	AB250213
<i>Spinitectus</i>	<i>carolini</i>	SpinCar1G	DQ503464
<i>Spirinia</i>	<i>elongata</i>	SpirElo1G	EF527426
<i>Spirocamallanus</i>	<i>istiblenni</i>	SpCaIst1G	EF180076
<i>Spirocamallanus</i>	<i>rarus</i>	SpCaRar1G	DQ494195
<i>Spirocerca</i>	<i>lupi</i>	SpCeLup1G	AY751497
<i>Spirocerca</i>	sp.	SpCeSp1G	AY751498
<i>Steinernema</i>	<i>affine</i>	SteiAff3-27	FJ040425
<i>Steinernema</i>	<i>carpocapsae</i>	SteiCarG	AF036604
<i>Steinernema</i>	<i>carpocapsae</i>	SteiCar1	FJ040415
<i>Steinernema</i>	<i>carpocapsae</i>	SteiCar3	FJ040416
<i>Steinernema</i>	<i>feltiae</i>	SteiFel2	FJ040417
<i>Steinernema</i>	<i>feltiae</i>	SteiFel4	FJ040418
<i>Steinernema</i>	<i>feltiae</i>	SteiFel5	FJ040419
<i>Steinernema</i>	<i>glaseri</i>	SteiGla2	FJ040422
<i>Steinernema</i>	<i>glaseri</i> 1	SteiGla1	AY284682
<i>Steinernema</i>	<i>kraussei</i>	SteiKra1	FJ040420
<i>Steinernema</i>	<i>kraussei</i>	SteiKra3	FJ040421
<i>Steinernema</i>	<i>monticolum</i>	SteiMon1	FJ040423
<i>Steinernema</i>	<i>scarabaei</i>	SteiSca1	FJ040424
<i>Steinernema</i>	sp. 1	SteiSp1	FJ040426
<i>Steinernema</i>	sp. 2	SteiSp2	FJ040427
<i>Stenurus</i>	<i>minor</i>	StenMin1G	AY295817
<i>Stephanurus</i>	<i>dentatus</i>	StepDen1G	AJ920345
<i>Stilbonema</i>	<i>majum</i>	StilMajG	Y16922
<i>Strongyloides</i>	<i>cebus</i>	StroCeb1G	AB272236
<i>Strongyloides</i>	<i>fuelleborni</i>	StroFue1G	AB272235
<i>Strongyloides</i>	<i>procyonis</i>	StroPro1G	AB272234
<i>Strongyloides</i>	<i>ratti</i>	StroRat1G	U81581
<i>Strongyloides</i>	<i>ratti</i>	StroRat2G	AF036605
<i>Strongyloides</i>	<i>robustus</i>	StroRob1G	AB272232
<i>Strongyloides</i>	<i>robustus</i>	StroRob2G	AB272233
<i>Strongyloides</i>	sp.	StroSp1G	AB272229
<i>Strongyloides</i>	sp.	StroSp2G	AB272230
<i>Strongyloides</i>	sp.	StroSp3G	AB272231
<i>Strongyloides</i>	<i>stercoralis</i>	StroSte1G	M84229



**Table S1.** (Continued).

Genus	Species	ID	GenBank No.
<i>Strongyloides</i>	<i>stercoralis</i>	StroSte3G	AF279916
<i>Strongylus</i>	<i>equinus</i>	StGyEqu1G	DQ094176
<i>Subanguina</i>	<i>radicicola</i>	SubaRadG	AF202164
<i>Subanguina</i>	<i>radicicola</i>	SubaRad1	EU682392
<i>Sulcascaris</i>	<i>sulcata</i>	SulcSul1G	EF180080
<i>Symplocostoma</i>	sp. 1	SympSp1	FJ040502
<i>Syngamus</i>	<i>trachea</i>	SyngTra1G	AF036606
<i>Syngamus</i>	<i>trachea</i>	SyngTra2G	AJ920344
<i>Synhimantus</i>	<i>hamatus</i>	SynhHam1G	EU004819
<i>Synhimantus</i>	<i>laticeps</i>	SynhLat1G	EU004818
<i>Synonchiella</i>	sp. 1	SynoSp1	FJ040468
<i>Syngolaimus</i>	sp. 1	SyriSp1	FJ040497
<i>Telotylenchus</i>	<i>ventralis</i> 1	TeloVen1	AY593905
<i>Teratocephalus</i>	<i>lirellus</i>	TCepLirG	AF036607
<i>Teratocephalus</i>	<i>terrestris</i>	TCepTer1	AY284683
<i>Teratorhabditis</i>	<i>ferrugineus</i>	TRhaFer1G	AB269816
<i>Teratorhabditis</i>	<i>palmarum</i>	TRhaPalG	U13937
<i>Teratorhabditis</i>	<i>synpapillata</i>	TRhaSynG	AF083015
<i>Terranova</i>	<i>caballeroi</i>	TerrCabG	U94381
<i>Terranova</i>	<i>scoliodontis</i>	TerrSco1G	DQ442661
<i>Tetrabothriostrogylus</i>	<i>mackerrasae</i>	TetrMac1G	AJ920359
<i>Tetrameres</i>	<i>fissipina</i>	TetMFis1G	EF180077
<i>Thalassoalaimus</i>	<i>pirum</i>	ThalPir1	FJ040500
<i>Thelastoma</i>	<i>krausi</i>	ThSoKra1G	EF180068
<i>Thelazia</i>	<i>lacrymalis</i>	ThelLac1G	DQ503458
<i>Theristus</i>	<i>acer</i>	TherAce1G	AJ966505
<i>Theristus</i>	<i>agilis</i>	TherAgi1	AY284693
<i>Theristus</i>	<i>agilis</i>	TherAgi3	AY284695
<i>Theristus</i>	<i>agilis</i>	TherAgi2	AY284694
<i>Theristus</i>	sp. 1	TherSp1	FJ040464
<i>Thonus</i>	2	ThonSp2	AY284797
<i>Thonus</i>	cf. <i>circulifer</i>	ThonCir1Z	AY284795
<i>Thonus</i>	sp.	ThonSp3	AY284798
<i>Thornia</i>	<i>steatopyga</i> 1	ThorSte1	AY284787
<i>Thulinia</i>	<i>stephaniae</i>	ThulSteG	AF056023
<i>Tobrilus</i>	<i>gracilis</i>	TobrGra1G	AJ966506
<i>Torynurus</i>	<i>convolutus</i>	ToryCon1G	AY295818
<i>Toxascaris</i>	<i>leonina</i>	TAscLeoG	U94383
<i>Toxocara</i>	<i>canis</i>	ToxoCan1G	U94382
<i>Toxocara</i>	<i>canis</i>	ToxoCan2G	AF036608
<i>Toxocara</i>	<i>cati</i>	ToxoCat1G	EF180059
<i>Toxocara</i>	<i>vitulorum</i>	ToxoVit1G	EF180078
<i>Trefusia</i>	<i>zostericola</i>	TrefZosG	AF329937
<i>Trichidorus</i>	<i>primitivus</i>	TricPri1	FJ040481
<i>Trichinella</i>	<i>britovi</i>	TriNBri1G	AY851257
<i>Trichinella</i>	<i>murrelli</i>	TriNMur1G	AY851259
<i>Trichinella</i>	<i>nativa</i>	TriNNat1G	AY487254
<i>Trichinella</i>	<i>nativa</i>	TriNNat2G	AY851256
<i>Trichinella</i>	<i>nelsoni</i>	TriNNel1G	AY851261
<i>Trichinella</i>	<i>papuae</i>	TriNPap1G	AY851263

Table S1. (Continued).

Genus	Species	ID	GenBank No.
<i>Trichinella</i>	<i>pseudospiralis</i>	TriNPse1G	AY851258
<i>Trichinella</i>	sp.	TriNSp1G	AY851260
<i>Trichinella</i>	sp.	TriNSp2G	AY851262
<i>Trichinella</i>	<i>spiralis</i>	TriNSpi1G	U60231
<i>Trichinella</i>	<i>spiralis</i>	TriNSpi2G	AY497012
<i>Trichinella</i>	<i>zimbabwensis</i>	TriNZim1G	AY851264
<i>Trichodorus</i>	<i>cylindricus</i>	TricCyl1	FJ040482
<i>Trichodorus</i>	<i>cylindricus</i>	TricCyl1G	AJ439578
<i>Trichodorus</i>	<i>nanjingensis</i>	TricNan1G	AJ439579
<i>Trichodorus</i>	<i>nanjingensis</i>	TricNan2G	AJ439580
<i>Trichodorus</i>	<i>pakistanensis</i>	TricPak1G	AJ439581
<i>Trichodorus</i>	<i>pakistanensis</i>	TricPak2G	AJ439582
<i>Trichodorus</i>	<i>primitivus</i>	TricPri1G	AF036609
<i>Trichodorus</i>	<i>primitivus</i>	TricPri2G	AJ439517
<i>Trichodorus</i>	<i>primitivus</i>	TricPri3G	AJ439583
<i>Trichodorus</i>	<i>similis</i>	TricSim1G	AJ439522
<i>Trichodorus</i>	<i>similis</i>	TricSim2G	AJ439584
<i>Trichodorus</i>	<i>similis</i>	TricSim3G	AJ439585
<i>Trichodorus</i>	sp.	TricSp1G	AJ439588
<i>Trichodorus</i>	<i>sparsus</i>	TricSpa1G	AJ439586
<i>Trichodorus</i>	<i>sparsus</i>	TricSpa2G	AJ439587
<i>Trichodorus</i>	<i>sparsus</i>	TricSpa3G	AJ439589
<i>Trichodorus</i>	<i>sparsus</i>	TricSpa4G	AJ439590
<i>Trichodorus</i>	<i>sparsus</i>	TricSpa5G	AJ439624
<i>Trichodorus</i>	<i>sparsus</i>	TricSpa6G	AJ439625
<i>Trichodorus</i>	<i>sparsus</i>	TricSpa7G	AJ439591
<i>Trichodorus</i>	<i>variopapillatus</i>	TricVar1	AY284841
<i>Trichostrongylus</i>	<i>colubriformis</i>	TrStCol1G	AJ920350
<i>Trichuris</i>	<i>muris</i>	TriUMurG	AF036637
<i>Trichuris</i>	<i>suis</i>	TriUSui1G	AY851265
<i>Trichuris</i>	<i>suis</i>	TriUSui2G	AY856093
<i>Trichuris</i>	<i>trichiura</i>	TriUTri1G	DQ118536
<i>Tridentulus</i>	sp.	TridSp1G	AJ966507
<i>Tripyla</i>	cf. <i>filicaudata</i>	TripFil1Z	AY284730
<i>Tripyla</i>	cf. <i>filicaudata</i>	TripFil2Z	AY284731
<i>Tripyla</i>	sp.	TripSp3G	EF197734
<i>Tripyla</i>	sp. 1	TripSp1	AY284732
<i>Tripylella</i>	sp.	TrLeSp1	AY284737
<i>Tripylella</i>	sp.	TrLeSp2	FJ040488
<i>Tripylina</i>	sp.	TrLiSp2G	EF197728
<i>Trischistoma</i>	1	TrisSp1	AY284735
<i>Trischistoma</i>	2	TrisSp2	AY284736
<i>Trischistoma</i>	<i>monohystera</i>	TrisMon1G	AJ966509
<i>Troglostrongylus</i>	<i>wilsoni</i>	TrogWil1G	AY295820
<i>Truttaedacnitis</i>	<i>truttae</i>	TrutTru1G	EF180063
<i>Trypila</i>	sp. 2	TripSp2	AY284733
<i>Trypila</i>	sp. 4	TripSp3	AY284734
<i>Tubiluchus</i>	<i>corallicola</i>	TubiCorG	AF119086
<i>Turbatrix</i>	<i>aceti</i>	TurbAceG	AF202165
<i>Turgida</i>	<i>torresi</i>	TurgTor1G	EF180069

**Table S1.** (Continued).

Genus	Species	ID	GenBank No.
<i>Tylencholaimellus</i>	<i>affinis</i>	TyMeAff1G	AY552978
<i>Tylencholaimellus</i>	<i>striatus</i>	TyMeStr1	AY284837
<i>Tylencholaimus</i>	cf. <i>teres</i>	TyLaTer1Z	EF207254
<i>Tylencholaimus</i>	sp. 2	TyLaSp2Z	AY284833
<i>Tylencholaimus</i>	<i>mirabilis</i>	TyLaMir2	AY284836
<i>Tylencholaimus</i>	<i>mirabilis</i>	TyLaMir3	EF207253
<i>Tylencholaimus</i>	sp.	TyLaSp1G	AJ966510
<i>Tylenchulus</i>	<i>semipenetrans</i>	TyChSem1G	AJ966511
<i>Tylenchus</i>	<i>arcuatus</i>	TyleArc1G	EU306348
<i>Tylenchus</i>	<i>arcuatus</i>	TyleArc2G	EU306349
<i>Tylenchus</i>	<i>davainei</i> 1	TyleDav1	AY284588
<i>Tylenchus</i>	sp.	TyleSp1	AY284589
<i>Tylocephalus</i>	<i>auriculatus</i>	TyCeAur2	AY284707
<i>Tylocephalus</i>	<i>auriculatus</i>	TyCeAur1G	AF202155
<i>Tylolaimophorus</i>	<i>minor</i>	TyloMin1G	AJ966512
<i>Tylopharynx</i>	<i>foetida</i>	TyPhFoe1G	EU306343
<i>Viscosia</i>	sp. 1	ViscSp1	FJ040494
<i>Wellcomia</i>	<i>siamensis</i>	WellSia1G	EF180079
<i>Wellcomia</i>	sp.	WellSp1G	EF180066
<i>Wilsonema</i>	<i>otophorum</i>	WilsOto2	AY593927
<i>Wilsonema</i>	<i>schuurmansstekhoveni</i>	WilsSch1G	AJ966513
<i>Wuchereria</i>	<i>bancrofti</i>	WuchBan1G	AF227234
<i>Wuchereria</i>	<i>bancrofti</i>	WuchBan2G	AY843436
<i>Wuchereria</i>	<i>bancrofti</i>	WuchBan3G	AY843437
<i>Wuchereria</i>	<i>bancrofti</i>	WuchBan4G	AY843438
<i>Xiphidorus</i>	<i>balcarceanus</i>	XiDoBal1G	AY297839
<i>Xiphidorus</i>	<i>minor</i>	XiDoMin1G	AY297830
<i>Xiphidorus</i>	<i>minor</i>	XiDoMin2G	AY604181
<i>Xiphidorus</i>	<i>parthenus</i>	XiDoPar1G	AY604182
<i>Xiphidorus</i>	sp.	XiDoSp1G	AY604183
<i>Xiphidorus</i>	sp.	XiDoSp2G	AY297841
<i>Xiphidorus</i>	<i>yepesara</i>	XiDoYep1G	AY297837
<i>Xiphidorus</i>	<i>yepesara</i>	XiDoYep2G	AY297838
<i>Xiphinema</i>	<i>americanum</i>	XiphAme1G	AY283170
<i>Xiphinema</i>	<i>americanum</i>	XiphAme2G	AY580056
<i>Xiphinema</i>	<i>americanum</i>	XiphAme3G	AM086684
<i>Xiphinema</i>	<i>bakeri</i>	XiphBak1G	AY283173
<i>Xiphinema</i>	<i>brasiliense</i>	XiphBra1G	AY297836
<i>Xiphinema</i>	<i>brevicolle</i>	XiphBre1G	AY297822
<i>Xiphinema</i>	<i>brevicolle</i>	XiphBre2G	AY580057
<i>Xiphinema</i>	cf. <i>americanum</i>	XiphAme4GZ	AM086671
<i>Xiphinema</i>	cf. <i>americanum</i>	XiphAme5GZ	AM086672
<i>Xiphinema</i>	cf. <i>americanum</i>	XiphAme6GZ	AM086679
<i>Xiphinema</i>	cf. <i>americanum</i>	XiphAme7GZ	AM086683
<i>Xiphinema</i>	<i>chambersi</i>	XiphCha1G	AY283174
<i>Xiphinema</i>	<i>citricolum</i>	XiphCit1G	AM086686
<i>Xiphinema</i>	<i>diffusum</i>	XiphDif1G	AY297823
<i>Xiphinema</i>	<i>diffusum</i>	XiphDif2G	AM086669
<i>Xiphinema</i>	<i>diffusum</i>	XiphDif3G	AM086677
<i>Xiphinema</i>	<i>diffusum</i>	XiphDif4G	AM086685

Table S1. (Continued).

Genus	Species	ID	GenBank No.
<i>Xiphinema</i>	<i>elongatum</i>	XiphElo1G	AY297824
<i>Xiphinema</i>	<i>ensiculiferum</i>	XiphEns1G	AY297825
<i>Xiphinema</i>	<i>floridae</i>	XiphFlo1G	AM086687
<i>Xiphinema</i>	<i>georgianum</i>	XiphGeo1G	AM086688
<i>Xiphinema</i>	<i>ifacolum</i>	XiphIfa1G	AY297826
<i>Xiphinema</i>	<i>incognitum</i>	XiphInc1G	AM086670
<i>Xiphinema</i>	<i>incognitum</i>	XiphInc2G	AM086678
<i>Xiphinema</i>	<i>index</i>	XiphInd1	EF207249
<i>Xiphinema</i>	<i>index</i>	XiphInd1G	AY687997
<i>Xiphinema</i>	<i>krugi</i>	XiphKru1G	AY297827
<i>Xiphinema</i>	<i>krugi</i>	XiphKru2G	AY297828
<i>Xiphinema</i>	<i>longicaudatum</i>	XiphLon1G	AY297829
<i>Xiphinema</i>	<i>oxycaudatum</i>	XiphOxy1G	AY297835
<i>Xiphinema</i>	<i>pachtaicum</i>	XiphPac1G	AM086682
<i>Xiphinema</i>	<i>paritaliae</i>	XiphPar1G	AY297831
<i>Xiphinema</i>	<i>peruvianum</i>	XiphPer1G	AY297832
<i>Xiphinema</i>	<i>peruvianum</i>	XiphPer2G	AM086674
<i>Xiphinema</i>	<i>rivesi</i>	XiphRiv1G	AF036610
<i>Xiphinema</i>	<i>rivesi</i>	XiphRiv2G	AM086673
<i>Xiphinema</i>	<i>simile</i>	XiphSim1G	AM086680
<i>Xiphinema</i>	<i>simile</i>	XiphSim2G	AM086681
<i>Xiphinema</i>	sp.	XiphSp1G	AY297840
<i>Xiphinema</i>	sp. 1	XiphSp1	EF207250
<i>Xiphinema</i>	<i>surinamense</i>	XiphSur1G	AY297833
<i>Xiphinema</i>	<i>taylori</i>	XiphTay1G	AM086675
<i>Xiphinema</i>	<i>taylori</i>	XiphTay2G	AM086676
<i>Xiphinema</i>	<i>variegatum</i>	XiphVar1G	AY297834
<i>Xiphinema</i>	<i>vuittenezi</i>	XiphVui1G	AY552979
<i>Xyzzors</i>	sp.	XyzzSpG	Y16923
<i>Zeldia</i>	<i>punctata</i>	ZeldPunG	U61760
<i>Zeldia</i>	sp. 1	ZeldSp1	AY284675
<i>Zeldia</i>	sp. 2	ZeldSp2	AY284676
<i>Zoniolaimus</i>	<i>mawsonae</i>	ZoniMaw1G	AJ920338
<i>Zygotylenchus</i>	<i>guevarai</i>	ZygoGue1G	AF442189