



Sensitive MS method for the detection of palytoxins in shellfish using Li^+ cationization

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Background

Palytoxins (PITXs) are produced by marine dinoflagellates (*Ostreopsis spp.*), cyanobacteria and certain corals. Palytoxin is a neurotoxic compound that can end-up in shellfish. PITXs are within the classes of marine biotoxins the largest non-protein molecules with a molecular weight of $\sim 2700\text{Da}$. PITX is a polyhydroxylated compound with both lipophilic and hydrophilic parts, partially an unsaturated aliphatic backbone containing cyclic ethers and hydroxyl and amide functional groups (figure 1). To protect shellfish consumers EFSA established an acute reference dose of $0.2\text{ }\mu\text{g/kg}$ b.w. which can be translated to $30\text{ }\mu\text{g}$ PITX-eq/kg shellfish (400g shellfish by 60kg person). Sensitive methods for analysis of intact palytoxins in shellfish are lacking.

Objective

- Development of a sensitive MS detection method for intact palytoxins
- Validation of the developed method for shellfish monitoring

Method

- 1 g of shellfish, triplicate extracted with 3mL $50\text{v/v}\%$ MeOH.
- SPE Strata-X, after conditioning and sample extract applied, cartridge washed with water and subsequently eluted with 100% MeOH containing $0.1\text{v/v}\%$ acetic acid.
- LC column: Kinetex C_{18} $100 \times 2.1\text{mm}$, $1.7\text{ }\mu\text{m}$
- Mobile phase: A: H_2O and B: ACN: H_2O (9:1), both containing 0.25mM LiI and $0.00125\text{v/v}\%$ formic acid
- Flow: 0.4 mL/min
- Gradient: Linear, total 10 min run time
- Injection volume: $10\text{ }\mu\text{L}$
- MS: Thermo Q-Exacte Orbitrap and Waters Xevo TQ-S

Results

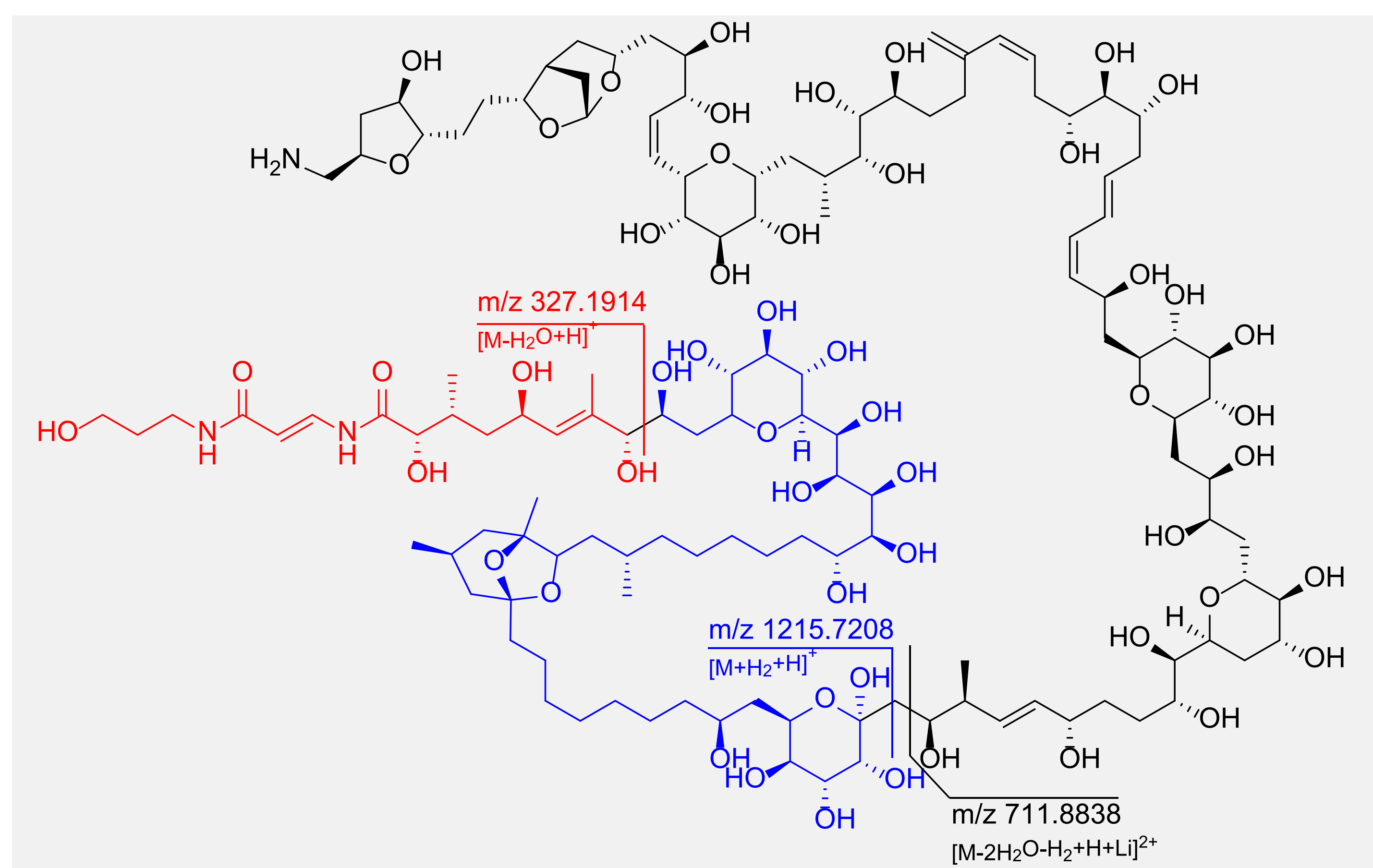


Figure 1. Proposed fragmentation of PITX under acidic mobile phase conditions with LiI as additive

Results

- LiI (0.25mM) combined with formic acid ($0.00125\text{ v/v}\%$) in the mobile phase showed best response in the MS for PITX.
- Only one abundant ion was formed (m/z 897.8413 , $[\text{M}+\text{H}+2\text{Li}]^{3+}$).
- Q-exactive MS^2 spectra revealed both non specific fragments (losses of H_2O) and structural related fragments (figure 1 and 2).
- MRM method developed at Waters Xevo TQS, MRM transitions $898.2>880.2$, $898.2>874.2$ and $898.2>711.9$.
- Sensitivity of the applied methodology on the TQ-S was good with LODs of 8 and $22\text{ }\mu\text{g/kg}$ for respectively mussels and oysters.

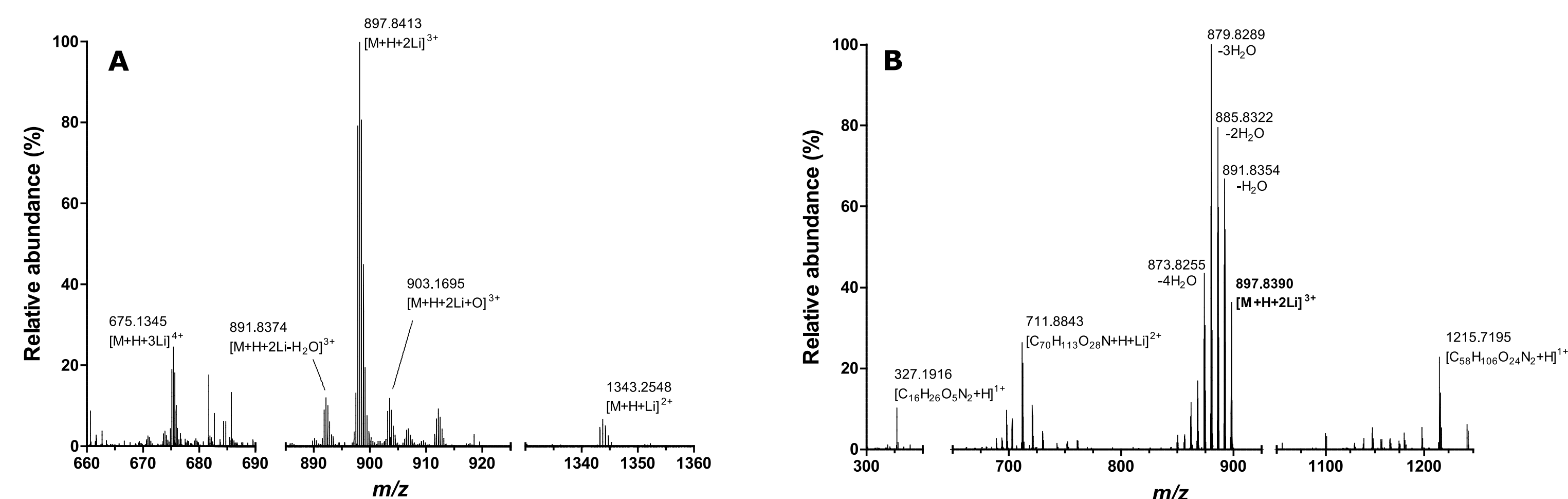


Figure 2. A) MS^1 spectra of palytoxin and B) MS^2 spectra of palytoxins with m/z 987.8 as precursor mass. Both recorded under acidic mobile phase conditions with LiI as additive.

Validation

- For recovery and repeatability 20 samples ($n=5$ per matrix), mussels, oysters, cockles and ensis were fortified at 10 , 30 and $60\text{ }\mu\text{g}$ PITX/kg shellfish.
- For selectivity 20 blank samples ($n=5$ per matrix) were used
- For quantification and linearity matrix matched calibration curves were constructed at concentrations 0 , 5 , 10 , 30 , 60 and $100\text{ }\mu\text{g}$ PITX/kg blank mussel homogenate.

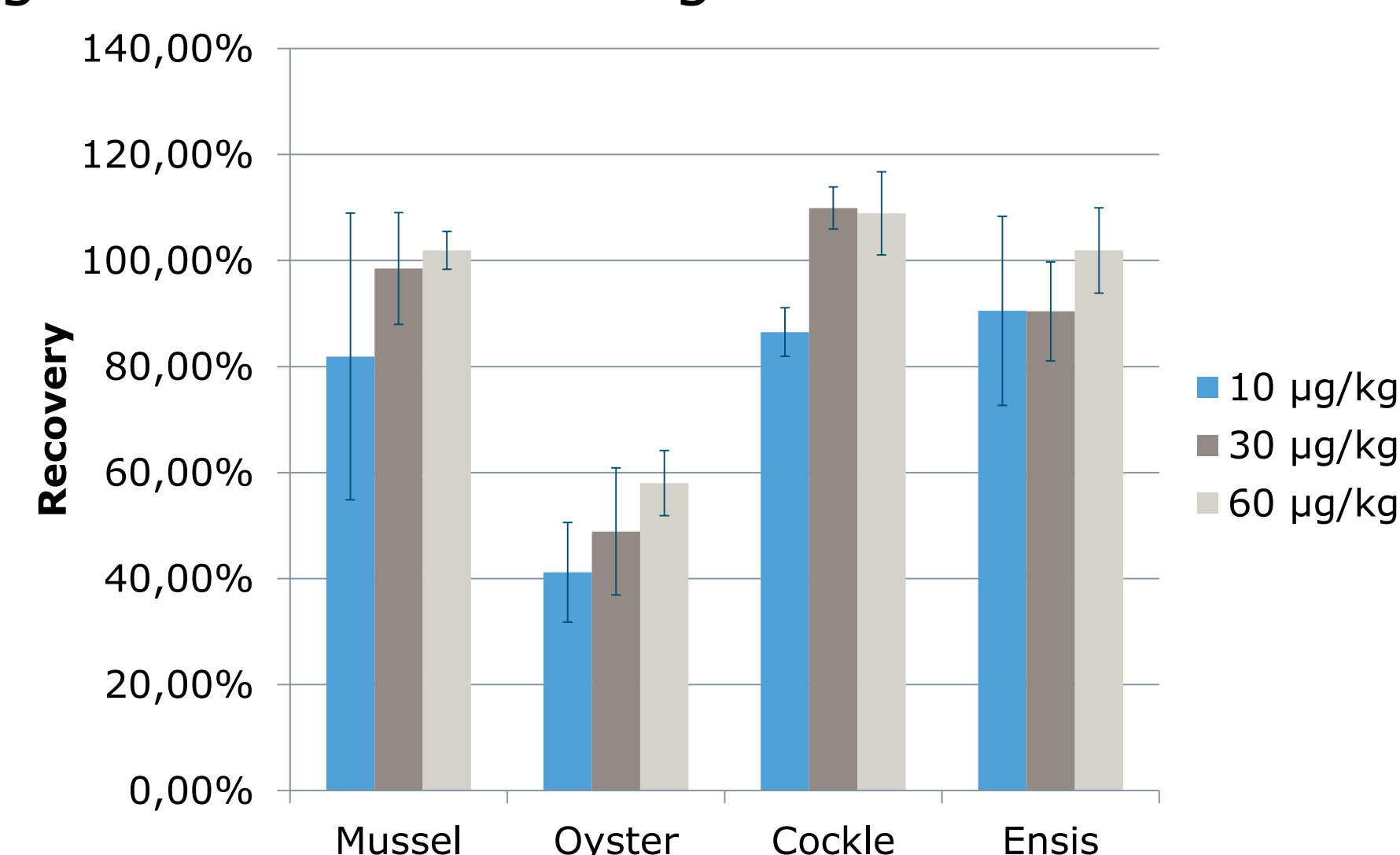


Figure 3. Recovery and repeatability error (RSD $n=5$) for various shellfish matrices.

Conclusions

- Sensitive LC-MS/MS method for the detection of PITXs and analogues in shellfish is developed.
- Validation was successful with exception of repeatability at $10\text{ }\mu\text{g/kg}$ which had $\text{RSDs}>20\%$, further for oysters recoveries in general were low ($40\text{--}60\%$) (figure 3).

Acknowledgements

- This project was financed by the Ministry of Agriculture, Nature and Food Quality (KB-23-002-021)

