The IL-6–gp130–STAT3 pathway in hepatocytes triggers liver protection in T cell–mediated liver injury

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Introduction
Various etiologies result in acute and chronic liver diseases. Particularly in autoimmune and viral hepatitis, immune-mediated mechanisms play a central role and thus determine disease progression. In recent years, major progress has been made in treating these patients. However, there are still a high percentage of individuals for whom all treatment options fail and liver transplantation is required. Therefore, alternative treatment options are needed; therapeutic approaches that either inhibit immune-mediated mechanisms or directly inhibit liver cell damage are promising and might enter into the clinic.

IL-6 is a pleiotropic cytokine with beneficial effects for the liver. This cytokine promotes liver regeneration and protects against a multitude of liver-damaging influences such as alcohol and CCl4 intoxication (1–4). IL-6 prevents apoptosis and exhibits positive effects in both ischemia/reperfusion models and fatty liver transplantation (5–7). Additionally, IL-6 is known to efficiently reduce effects in both ischemia/reperfusion models and fatty liver transplantation (5–7). Additionally, IL-6 is known to efficiently reduce fatty liver transplantation (5–7). Additionally, IL-6 is known to efficiently reduce fatty liver transplantation. This cytokine promotes liver regeneration and protects against a multitude of liver-damaging influences such as alcohol and CCl4 intoxication (1–4). IL-6 prevents apoptosis and exhibits positive effects in both ischemia/reperfusion models and fatty liver transplantation (5–7). Additionally, IL-6 is known to efficiently reduce fatty liver transplantation (5–7). Additionally, IL-6 is known to efficiently reduce fatty liver transplantation. However, there are still a high percentage of individuals for whom all treatment options fail and liver transplantation is required. Therefore, alternative treatment options are needed; therapeutic approaches that either inhibit immune-mediated mechanisms or directly inhibit liver cell damage are promising and might enter into the clinic.

Increasing evidence demonstrates that IL-6 has a protective role during liver injury. IL-6 activates intracellular pathways via the gp130 receptor. In order to identify IL-6–gp130 pathways involved in mediating liver protection, we analyzed hepatocyte-specific gp130 knockout mice in a concanavalin A–induced (Con A–induced) model of immune-mediated hepatitis. We demonstrated that IL-6–gp130–dependent pathways in hepatocytes alone are sufficient for triggering protection in Con A–induced hepatitis. gp130-STAT3 signaling in hepatocytes mediates the IL-6–triggered protective effect. This was demonstrated by analysis of IL-6–induced protection in mice selectively deficient for gp130-dependent STAT1/3 or gp130-SHP2-RAS signaling in hepatocytes. To identify IL-6–gp130–STAT1/3–dependent expressed liver-protective factors, we performed gene array analysis of hepatic gene expression in hepatocyte-specific gp130−/− mice as well as in gp130-STAT1/3– and gp130-SHP2-RAS–deficient mice. The mouse IL-8 ortholog KC (also known as Gro-α) and serum amyloid A2 (SAA2) was identified as differentially IL-6–gp130–STAT3–regulated genes. Hepatic expression of KC and SAA2 mediate the liver-protective potential of IL-6, since treatment with recombinant KC or serum SAA2 effectively reduced liver injury during Con A–induced hepatitis. In summary, this study defines IL-6–gp130–STAT3–dependent gene expression in hepatocytes that mediates IL-6–triggered protection in immune-mediated Con A–induced hepatitis. Additionally, we identified the IL-6–gp130–STAT3–dependent proteins KC and SAA2 as new candidates for therapeutic targets in liver diseases.

Con A–induced liver damage is a well-defined experimental animal model of acute hepatitis. It mimics human viral and autoimmune hepatitis in many aspects (8–10). Con A–induced hepatitis is accompanied by an increase in the serum concentration of several cytokines, including TNF-α, IFN-γ, IL-6, and IL-1. During the early stages, TNF-α and IFN-γ directly mediate liver cell damage (11). The cell types involved in the initiation and propagation of Con A–induced hepatitis (neutrophils: CD4+ T cells, CD8+ T cells, NK T cells [NKT cells], and Kupffer cells) have been directly implicated in a number of human liver diseases, including autoimmune, viral, alcoholic, and ischemia/reperfusion injury (12–18).

The IL-6–triggered molecular mechanisms leading to liver protection are not completely understood. For signal transduction, IL-6 binds the membrane-bound IL-6 receptor α (gp80). The IL-6–gp80 dimer interacts with gp130, the signal transducing unit in this complex. Gp130 is the common signal transducer used by the IL-6 family of cytokines, which includes IL-6, leukemia inhibitory factor (LIF), IL-11, cardiotoxin-1 (CT-1), oncostatin-M (OSM), and the ciliary neurotrophic factor (19). The initiated intracellular signaling is strictly dependent on the function of gp130 (19). Formation of gp130-containing complexes results in activation of intracellular JAKs, which phosphorylate tyrosine residues in the cytoplasmic domain of gp130. The phosphorylation of gp130 leads to dimerization of gp130 and activation of STAT1 and STAT3 as well as the SHP2-RAS-MAPK pathway (where SHP2 is Src homology 2 [SH2] domain–containing protein tyrosine phosphatase) (20–22).

The use of distinct downstream signaling pathways from different...
phosphorylation sites of the gp130 intracellular domain enables ligand- and tissue-specific activation of specific target genes. For these reasons, IL-6 signaling allows a complex plasticity (23, 24).

The aim of the present study is to identify the IL-6–gp130-dependent pathways and the cell type that confer protection during Con A–induced liver injury. The identification of the responsible target cell and the essential intracellular signal cascade will help to clarify liver protective mechanisms and to identify IL-6–dependent molecular mediators that contribute to beneficial effects in immune-mediated liver diseases.

Results

IL-6–gp130–dependent signaling in hepatocytes confers protection in the Con A model. The cytokine IL-6 triggers liver protection in the Con A model of T cell–mediated hepatitis. IL-6 controls gene transcription in various cell types that might be involved in protective mechanisms, e.g., hepatocytes and immune-activated cells. Hepatocytes are the predominant hepatic cells; therefore, we evaluated whether hepatocytes mediate IL-6–dependent protection in the Con A model. We generated mice with a hepatocyte-specific deletion of the IL-6 signal–transducing gp130 receptor (25) and analyzed the effect of IL-6 treatment in this experimental hepatitis model. Hepatocyte-specific gp130-deficient mice were generated by crossbreeding alfpCre and gp130loxP mice, in which loxP sites flank exon 16, encoding the transmembrane domain of gp130 (26). Expression of Cre recombinase is controlled by a hepatocyte-specific albumin promoter, and results in a more than 90% deletion of exon 16 in adult liver (25). Since nonparenchymal cells show no gp130 deletion, the overall extent of hepatic gp130 inactivation is likely to reflect complete gp130 deletion in hepatocytes.

Gp130 wild-type (gp130loxP/loxP) animals were treated with IL-6 or vehicle before Con A injection. In order to monitor liver cell damage, AST levels were determined before and at various time points after treatment. IL-6 pretreatment completely blocked liver cell damage. No increase in transaminases was evident (Figure 1A). In alfpCre gp130loxP/loxP mice, IL-6 failed to have a protective effect, which indicates that IL-6–dependent liver protection is dependent on gp130-mediated signaling in hepatocytes (Figure 1B). Additionally, all alfpCre gp130loxP/loxP mice died within the first 20 hours after Con A injection. In contrast, all NaCl- and IL-6–treated wild-type animals survived during this phase, indicating a higher sensitivity of alfpCre gp130loxP/loxP mice toward Con A.

Lack of gp130 expression in hepatocytes has no impact on liver-damaging cytokine secretion after Con A injection. During Con A–induced hepatitis, immune cells are activated, and as a consequence, cytokines that promote liver cell damage are released. TNF-α and IFN-γ are essential in triggering Con A–induced liver injury. Additionally, IFN-γ and TNF-α are markers of immune-cell activation in this model. They are expressed by activated macrophages or CD4+ and CD8+ cells (1, 3, 27, 28). Therefore, we were interested in studying whether lack of gp130 expression in hepatocytes and IL-6 pretreatment have an impact on cytokine secretion.

IL-6 treatment of gp130loxP/loxP mice resulted in a slight reduction of TNF-α levels compared with those in animals treated with the carrier solution (Figure 2A). The same observation was evident in alfpCre gp130loxP/loxP animals. Slightly reduced TNF-α levels were observed in the IL-6–treated group (Figure 2B). IL-6 pretreatment had no impact on IFN-γ secretion in wild-type and alfpCre gp130loxP/loxP animals. The endogenous IL-6 levels during Con A–induced hepatitis are significantly elevated in alfpCre gp130loxP/loxP mice. In wild-type mice, a significant increase in IL-6 serum concentration was observed during the first hours of Con A–induced hepatitis (Figure 2E). The IL-6 serum levels returned to basal levels 24 hours after Con A injection. IL-6 pretreatment reduced IL-6 serum levels at later time points (Figure 2E). A different IL-6 kinetic was evident in alfpCre gp130loxP/loxP animals. In both groups, a marked increase in IL-6 serum levels was obvious during the first hours after Con A injection. Additionally, IL-6 remained elevated during the first 24 hours in both groups (Figure 2F).

The IL-6–gp130–dependent activation of STAT3 in wild-type mice is suppressed in alfpCre gp130loxP/loxP animals. To distinguish between the intracellular events that occur in wild-type and alfpCre gp130loxP/loxP animals during Con A–induced liver cell damage, we analyzed hepatic STAT1 and STAT3 activation during disease progression. Earlier studies indicated that STAT1 plays a harmful role during activation of CD4+ and NKT cells (29). Therefore, we studied nuclear translocation of phosphorylated STAT1 and STAT3 in the liver.

IL-6 pretreatment had no impact on hepatic STAT1 activation in wild-type or in alfpCre gp130loxP/loxP mice (Figure 3A; 0 hours, Con A) (25). Con A injection resulted in STAT1 activation in wild-type and alfpCre gp130loxP/loxP mice, but no remarkable differences were found between the 2 groups (Figure 3A). IL-6 pretreatment resulted in a slight increase of STAT1 phosphorylation in wild-type mice 2 hours and 4 hours after Con A injection (Figure 3A). In alfpCre gp130loxP/loxP animals, IL-6 injection resulted in a stronger and earlier STAT1 phosphorylation than in wild-type mice. Eight hours after Con A injection, STAT1 activation returned to basal levels (Figure 3A).

Activation of STAT3 during Con A–induced hepatitis can be divided into 3 phases (Figure 3B). The prephase, before Con A injection (0 hours), the immediate phase, 2 hours after injection, and 2 later phases, 4 and 8 hours after initiation of the disease.
To differentiate between STAT3 activation in hepatocytes and other liver cells we performed confocal fluorescence stainings of CD11b and P-STAT3. In the prephase, STAT3 phosphorylation was exclusively detectable in hepatocytes of IL-6–pretreated wild-type animals, but not in IL-6–injected alfpCre gp130LoxP/LoxP animals (Figure 3B; 0 hours, Con A). During this phase, STAT3 was predominately expressed in activated monocytes, NK cells, and macrophages (Figure 3C; CD11b and P-STAT3). In the prephase, STAT3 phosphorylation was detected in hepatocytes isolated from wild-type (gp130LoxP/LoxP) and alfpCre gp130LoxP/LoxP mice. STAT3 phosphorylation was exclusively detectable in hepatocytes of IL-6–pretreated wild-type animals compared to that in the NaCl-treated groups (Figure 3B). In hepatocyte-specific gp130Y757F/LoxP animals (Figure 3B; 0 hours, Con A), STAT3 phosphorylation was not detectable. This activation was independent of IL-6 pretreatment (Figure 3D).

Generation and molecular characterization of hepatocyte-specific gp130ΔSTAT/LoxP and gp130ΔY757F/LoxP mice. Since gp130 expression in hepatocytes is essential for IL-6–induced protection in Con A–induced hepatitis, we aimed to identify the intracellular signaling pathways that are responsible for mediating protection. For this purpose, we generated mice with hepatocyte-specific deletions of either gp130ΔSTAT1/3 or gp130ΔSHP2–RAS-MAPK signaling. The activation of the 2 distinct signaling cascades in response to ligand-mediated gp130 dimerization is mediated by phosphorylation of specific tyrosines (Y) in the cytoplasmic domain of gp130. Y757 mediates the engagement of the cytoplasmic Src-homology tyrosine phosphatase (SHP2) and the activation of the RAS-ERK pathway (30). The 4-membrane distal phosphotyrosines are binding sites for the SH2 domains of the latent transcription factors STAT1 and STAT3 (31).

The alfpCre gp130ΔSTAT/LoxP and alfpCre gp130ΔY757F/LoxP animals were obtained by crossing alfpCre gp130LoxP/LoxP with either gp130ΔSTAT/STAT or gp130ΔY757F/Y757F knockin mice (32). Gp130ΔSTAT/STAT mice express a truncated gp130 protein that lacks the part of the cytoplasmic domain required for STAT1 and STAT3 binding and activation. Gp130ΔY757F/Y757F animals express a gp130 protein with a phenylalanine substitution of the Y757 residue which, in its phosphorylated form, is essential for SHP2 recruitment and subsequent activation of the RAS-MAPK cascade. In the resulting alfpCre gp130ΔSTAT/LoxP and alfpCre gp130ΔY757F/LoxP animals, the LoxP-flanked gp130 is deleted selectively in hepatocytes as expression of Cre recombinase is controlled by a hepatocyte-specific promoter (25). This approach therefore enables the selective expression of mutant gp130 protein in hepatocytes. Each mouse was genotyped by PCR analysis for the genomic existence of Cre, gp130LoxP, and the respective mutant gp130 allele.

Functional characterization of the alfpCre gp130ΔSTAT/LoxP and alfpCre gp130ΔY757F/LoxP animals was performed by IL-6 stimulation in primary hepatocytes isolated from the 4 strains. IL-6–induced activation of gp130-dependent pathways was studied by the monitoring of P-STAT1/3 and P-ERK2 expression. Treatment of wild-type primary hepatocytes resulted in phosphorylation of STAT3 and ERK2 (Figure 4B). Neither protein was phosphorylated (and hence activated) in hepatocytes derived from alfpCre gp130ΔSTAT/LoxP mice (Figure 4C). However, in hepatocytes isolated from IL-6–stimulated alfpCre gp130ΔSTAT/LoxP or alfpCre gp130ΔSTAT/LoxP mice, we observed selective activation of P-STAT3 or P-ERK2, respectively (Figure 4, D and E). P-STAT1 activation was neither induced in IL-6–stimulated primary hepatocytes isolated from wild-type, alfpCre gp130ΔSTAT/LoxP, alfpCre gp130ΔY757F/LoxP, nor alfpCre gp130ΔSTAT/LoxP animals (data not shown).

Activation of the IL-6–gp130–STAT pathway in hepatocytes confers protection during Con A–induced liver damage. We used alfpCre gp130ΔY757F/LoxP and alfpCre gp130ΔSTAT/LoxP animals to identify the
gp130-dependent pathways in hepatocytes essential for mediating IL-6–dependent protection during Con A–induced hepatitis. IL-6 pretreatment of alfpCre gp130<sup>Y757F/LoxP</sup> mice resulted in complete prevention of liver cell damage (Figure 5A). In contrast, alfpCre gp130<sup>ΔSTAT/LoxP</sup> animals showed enhanced sensitivity toward Con A when compared to wild-type mice (Figure 5B). Independently of IL-6 pretreatment, all alfpCre gp130<sup>ΔSTAT/LoxP</sup> mice died within 8 hours after Con A injection in contrast to a 100% survival rate for all analyzed alfpCre gp130<sup>Y757F/LoxP</sup> animals.

To analyze possible modifications of immune-cell activation in these animals, serum TNF-α, IFN-γ, and IL-6 levels were determined before and after Con A injection (Figure 5C and D). IL-6 pretreatment in alfpCre gp130<sup>Y757F/LoxP</sup> or alfpCre gp130<sup>ΔSTAT/LoxP</sup> animals had no significant impact on TNF-α serum levels (Figure 5D). In both mouse strains, IL-6 treatment resulted in slightly elevated IFN-γ serum levels (Figure 5, B and C). Additionally, IL-6 serum levels did not differ between the vehicle-treated alfpCre gp130<sup>Y757F/LoxP</sup> and alfpCre gp130<sup>ΔSTAT/LoxP</sup> animals. In both mouse strains, we detected maximal IL-6 concentrations 8 hours after Con A injection (Figure 5D). In contrast, pretreatment with recombinant IL-6 resulted in a reduction of endogenous IL-6 serum concentrations during Con A–induced hepatitis in alfpCre gp130<sup>Y757F/LoxP</sup> animals (Figure 5C). No reduction of IL-6 serum levels was observed in alfpCre gp130<sup>ΔSTAT/LoxP</sup> mice (Figure 5D).

Gene expression profiles of IL-6–gp130 pathways in hepatocytes. The analysis of hepatocyte-specific gp130 mutations during Con A–induced hepatitis demonstrated that IL-6–gp130–STAT3–dependent signals appear to confer protection. To identify potential target genes involved in mediating protection, hepatic gene expression profiles were determined in IL-6–treated wild-type, alfpCre gp130<sup>LoxP/LoxP</sup>, alfpCre gp130<sup>Y757F/LoxP</sup>, and alfpCre gp130<sup>ΔSTAT/LoxP</sup> mice. Figure 6 depicts a selection of genes that are significantly regulated by gp130-dependent pathways in hepatocytes in vivo. These can be divided into 2 groups: genes involved in controlling gp130-dependent signaling in the cell, e.g., <i>stat-1</i>, <i>stat-3</i>, <i>socs-1</i> (suppressor of cytokine signaling-1) and <i>socs-3</i>; and genes regulating immune-mediated mechanisms, e.g., serum amyloid A2 (<i>saa2</i>) and keratinocyte growth factor (<i>kc</i>). As the gp130-STAT3–dependent pathway confers protection in the Con A model, we hypothesized that indirect immune-mediated mechanisms were essential for protection against liver damage in the Con A model.

Treatment with SAA2 and KC protects against Con A–induced liver damage. We analyzed in more detail the regulation of <i>saa</i> and <i>kc</i> and their potential roles during Con A–induced liver failure. Both
genes are selectively induced via IL-6–gp130–STAT3. The analysis of SAA serum expression showed that pronounced transcriptional induction of the saa gene strictly depended on IL-6–gp130–STAT3 signaling in hepatocytes. In order to confirm the array analysis, we also investigated hepatic mRNA expression (data not shown) and serum levels of SAA and found that both were elevated only in wild-type and alfpCre gp130ΔSTAT/LoxP mice (Figure 7A).

SAA is known to interact with metalloproteinases and to influence the activity of immune cells (33). Therefore, we studied the impact of SAA on Con A–induced hepatitis by pretreating wild-type animals with recombinant SAA 1 hour before Con A injection. SAA injection resulted in a significant reduction in the increase of AST levels compared with those in vehicle-treated control animals, indicating that SAA may have a protective effect for Con A–induced liver damage (Figure 7B).

KC controls chemotaxis and activation of polymorph nuclear cells (PMNs) in areas of inflammation. Serum KC expression was also strictly dependent on an intact IL-6–gp130–STAT3–signaling axis in hepatocytes (Figures 6 and 7C). No hepatic KC regulation was found in alfpCre gp130ΔSTAT/LoxP and alfpCre gp130ΔSTAT/ΔSTAT mice. Earlier results showed that the preexistence of high KC concentration in the serum leads to decreased PMN infiltration during inflammation (34). This effect is caused by reduced sensitivity of downstream pathways of the corresponding chemokine receptor (34). We thus analyzed the influence of IL-6 pretreatment on PMN infiltration during Con A–induced hepatitis. IL-6 treatment resulted in a significant reduction of PMN infiltration 4 hours after Con A injection (Figure 7D). This time coincided with the maximum of PMN infiltration. In order to further investigate the relevance of this observation, recombinant KC was injected before Con A administration. KC had a protective effect on Con A–induced T cell–mediated liver failure. The increase in transaminases was significantly blocked in the animals treated with KC (Figure 7E). Additionally, KC treatment leads to 100% survival during Con A–induced hepatitis compared with 70% in NaCl-treated animals.

Discussion
Con A injection into mice results in T cell activation in the liver as well as hepatocyte damage. The mechanisms during Con A–induced disease progression resemble human liver diseases, e.g., autoimmune disease and viral hepatitis (12–18). Therefore, the concentrations of inflammatory cytokines and chemokines are increased in the serum and liver during Con A–induced hepatitis.

Figure 4
Generation and functional characterization of hepatocyte-specific gp130ΔSTAT/LoxP and gp130Y757F/LoxP mice. (A) alfpCre gp130LoxP/LoxP mice were generated by breeding alfpCre mice with animals expressing LoxP-flanked gp130 alleles (25). Hepatocyte-specific gp130ΔSTAT/LoxP animals (alfpCre gp130ΔSTAT/LoxP) were generated by crossing alfpCre gp130LoxP/LoxP with gp130ΔSTAT/ΔSTAT mice, which express a truncated gp130 protein that lacks the domains mediating STAT1 and STAT3 activation (26). Hepatocyte-specific gp130Y757F/LoxP animals (alfpCre gp130Y757F/LoxP) were bred by crossing alfpCre gp130LoxP/LoxP with gp130Y757F/ΔSTAT mice, which express a gp130 allele encoding a phenylalanine substitution of the Y757 residue (32), thereby rendering gp130 incapable of recruiting SHP2 and activating the RAS-MAPK pathway. Animals were genotyped by PCR analysis for alfpCre, gp130LoxP, and gp130Y757F alleles. (B and E) The functional characterization of hepatocyte-specific gp130 mutant mice. Gp130 downstream-signaling pathways were analyzed by monitoring of phosphorylated STAT3 and phosphorylated ERK2 (p42) expression in whole cell extracts of primary hepatocytes isolated from wild-type (B), alfpCre gp130LoxP/LoxP (C), alfpCre gp130ΔSTAT/LoxP (E), or alfp Cre gp130ΔSTAT/ΔSTAT mice (E). Activation of phosphorylated STAT1 was not detected in any of the 4 mouse strains.
The model is attractive for exploring mechanisms that might ultimately result in new therapeutic options.

We analyzed mice with mutations in the gp130 receptor in hepatocytes that selectively impair the signaling capacity of this receptor. Functional inactivation of this molecule in hepatocytes resulted in lack of signal activation after stimulation with ligands of the IL-6 cytokine family, e.g., IL-6 or OSM. Further experiments in hepatocyte-specific knockout animals showed that lack of gp130 expression in hepatocytes renders these animals more susceptible to injury during various pathophysiological conditions of the liver (25). Therefore, the aim of the present study was to identify molecular mechanisms that are involved in mediating IL-6–gp130–dependent protection in hepatocytes.

Earlier studies demonstrated that IL-6 attenuated liver injury in Con A–induced hepatitis (1). Additionally, recent reports demonstrated that other IL-6 family members, e.g., IL-11 and CT-1, also confer protection in this model (35, 36). As all IL-6 family members use the gp130 receptor for signal transduction, our aim was to determine the cell type that confers protection in this model. Interestingly, when gp130 was selectively deleted in hepatocytes, IL-6 no longer conferred a protective effect (Figure 1). Moreover, in hepatocyte-specific gp130 knockout mice, Con A–induced liver injury was more pronounced and resulted in death of the animals within 24 hours, indicating that gp130-dependent pathways in liver cells are also protective in the later phase of Con A–induced liver injury.

In a recent study, Sun et al. also suggested that IL-6 might attenuate liver injury in Con A–induced hepatitis (37) by suppressing the activity of NKT cells. NKT cell activation is an important event in the initial phase of Con A–induced hepatitis (38). However, the IL-6 doses Sun et al. used were 10-fold higher than in the IL-6–treatment schedule used in this study. Thus, the effect of IL-6 on NKT cell activation might be added to the more prominent effect triggered by IL-6 in hepatocytes.

Intracellular signaling mediated by gp130 is dependent on phosphorylation of tyrosine residues in the intracellular part of the receptor. To dissect the contribution of the 2 major pathways engaged by gp130 in this model, we focused on the gp130-STAT3 signaling pathway in hepatocytes.
In further experiments, we analyzed whether IL-6 pretreatment induces changes in gp130-dependent signaling in hepatocytes. IL-6 stimulation may have effects on cytokine secretion during Con A–induced hepatitis. In particular, TNF-α and IFN-γ have been shown to play an essential role (41, 42). TNF-α is produced primarily by Kupffer cells in Con A–induced hepatitis (16). In wild-type mice, TNF-α levels were reduced after IL-6 stimulation. But in alfpCre gp130Y757F/LoxP mice, which retained sensitivity to IL-6–dependent liver protection, no changes in TNF-α levels were detected. These results indicate that a change in TNF-α serum levels is not essential for the protective effect mediated by IL-6.

IFN-γ is a marker of activated CD4+ cells and is strongly elevated after Con A injection (27, 42). IFN-γ levels in alfpCre gp130Y757F/LoxP and wild-type mice were not affected by pretreatment with IL-6, despite our observation that both strains showed protection from Con A–induced liver damage (Figure 2, C and D; Figure 5, C and D). Thus, serum levels of TNF-α and IFN-γ showed no correlation with the capacity of IL-6 to provide protection from liver damage, indicating that alterations in concentrations of these cytokines are unlikely to explain protection.

We next concentrated on mechanisms related to IL-6–dependent signaling in hepatocytes. IL-6 serum concentrations correlated with survival rates. High IL-6 serum concentrations after Con A injection were detectable in animals with a disruption of the IL-6–gp130–STAT3 pathway and correlated with increased morbidity. In contrast, IL-6–induced activation of STAT3 in hepatocytes prior to or 4 hours after Con A challenge was associated with survival of the animals. Lack of STAT3 activation in hepatocytes at these time points, as observed in alfpCre gp130+/-P/LoxP (Figure 1B) and alfpCre gp130Y757F/LoxP (Figure 5B) animals, correlated with poor prognosis.

STAT3 activation 2 hours after Con A injection occurred predominantly in activated immune cells, such as NK cells and infiltrating monocytes (CD11b+ cells) (Figure 3C, NaCl-treated groups). The pathways leading to STAT3 activation in these cells are currently unclear. gp130-dependent or gp130-independent factors, e.g., IFN-γ, could be relevant in explaining this observation (29).

IL-6 pretreatment increased the amount of phosphorylated STAT3 in nonparenchymal cells 2 hours after Con A injection by increasing numbers of P-STAT3–positive cells (Figure 3C). The increased number of STAT3-positive cells likely reflects IL-6–induced sensitization of NK cells, monocytes, or Kupffer cells to STAT3-activating pathways.

Four hours after Con A injection, STAT3 activation occurred predominately in hepatocytes in a gp130-dependent manner (Figure 3, B and D). Strong STAT3 activation in this phase correlated with maximal IL-6 serum levels (Figure 2E). IL-6 pretreatment inhibited the increase of STAT3 activation in hepatocytes of wild-type mice, probably by inducing negative feedback mechanisms, e.g., induction of SOCS-3 in hepatocytes (Figure 6).

It has been shown that SOCS-3 negatively regulates IL-6 signaling in vivo (43). In agreement with these findings, we demonstrated that activation of STAT3 is transitional. Hepatocytes of IL-6–pretreated wild-type mice showed STAT3 activation 3 hours after IL-6 injection. Up to 4 hours after initiation of Con A–induced hepatitis, these cells were not able to activate STAT3, although high IL-6 serum levels were detectable (Figures 3D and 2E).

In order to define potential IL-6–gp130–STAT3 target genes in hepatocytes that might be involved in conferring protection during Con A–induced liver failure, we performed gene expression profiling. Interestingly some acute phase genes were strictly

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Figure 6
IL-6–induced gene expression profile of wild-type, alfpCre gp130+/+Stat3/LoxP, alfpCre gp130+/+Y757F/LoxP, and alfpCre gp130+/+Stat3/LoxP mice. Affymetrix GeneChip analysis was used to detect gene expression of IL-6–stimulated gp130+/+Stat3/LoxP, alfpCre gp130+/+Stat3/LoxP, alfpCre gp130+/+Stat3/LoxP, and alfpCre gp130+/+Y757F/LoxP mice. Three allele groups were per group were treated in parallel by i.p. injection of recombinant IL-6–IL-6 rα (200 μg/kg) or NaCl. Liver RNA was isolated, and gene expression was compared. Most of the examples shown are differentially regulated by gp130-STAT signaling. Glycine receptor (Glycine R) and cytochrome p450 (p450) represent examples that are regulated by gp130-ras-MAPK signaling. The change of gene expression level is marked by increasing color intensity. LBP, LPS binding protein.
regulated via IL-6–gp130–STAT3. SAA2 showed the most prominent regulation. SAA2 is a known target of MMP and MMP activity induced during T cell–mediated hepatitis. Earlier experiments have shown that inhibition of MMPs attenuates liver failure (44). Indeed, treatment with recombinant SAA (0.8 mg/kg) leads to a significant reduction of AST levels in the time course of Con A–induced hepatitis in wild-type mice. Either SAA (dotted line) or NaCl (solid line) was injected intravenously 1 hour before induction of Con A–induced hepatitis. (C) Serum KC levels after IL-6 injection. Indeed, squares represent wild-type mice, open squares show alfpCre gp130LoxP/LoxP mice, filled triangles mark KpcCre gp130Y757F/LoxP mice, and open triangles represent alfpCre gp130ΔSTAT/LoxP mice. (D) IL-6 treatment results in a significant reduction of PMN infiltration during Con A–induced hepatitis. Four hours after Con A injection, the influx of PMNs was counted on H&E-stained paraffin sections by an experienced liver pathologist. The arrows mark liver infiltrating PMNs. hPF, high powered field. (E) KC (40 μg/kg) treatment leads to a significant reduction of AST levels during Con A–induced hepatitis. Eight mice per group were treated with either KC (dotted line) or NaCl (solid line) 1 hour before Con A injection. *P < 0.001 and *P < 0.05 vs. corresponding control groups at the same time point.

Figure 7
Acute-phase protein SAA2 and the chemokine KC are IL-6–gp130–STAT–induced, liver-protective proteins. (A) SAA2 serum levels after IL-6 (200 μg/kg) injection. Each time point represents the serum of 3–5 animals. Filled squares represent wild-type mice, open squares show alfpCre gp130LoxP/LoxP mice, filled triangles mark alfpCre gp130Y757F/LoxP mice, and open triangles represent alfpCre gp130ΔSTAT/LoxP. (B) Pretreatment with recombinant SAA (0.8 mg/kg) leads to a significant reduction of AST levels in the time course of Con A–induced hepatitis in wild-type mice. Either SAA (dotted line) or NaCl (solid line) was injected intravenously 1 hour before induction of Con A–induced hepatitis. (C) Serum KC levels after IL-6 injection. (D) IL-6 treatment results in a significant reduction of PMN infiltration during Con A–induced hepatitis. Four hours after Con A injection, the influx of PMNs was counted on H&E-stained paraffin sections by an experienced liver pathologist. The arrows mark liver infiltrating PMNs. hPF, high powered field. (E) KC (40 μg/kg) treatment leads to a significant reduction of AST levels during Con A–induced hepatitis. Eight mice per group were treated with either KC (dotted line) or NaCl (solid line) 1 hour before Con A injection. *P < 0.001 and *P < 0.05 vs. corresponding control groups at the same time point.

In conclusion, our present analysis suggests that IL-6–gp130–dependent gene activation in hepatocytes confers protection against T cell hepatitis induced by Con A. Functional dissection of gp130-dependent pathways in hepatocytes demonstrates that
IL-6–mediated STAT3 activation correlates with reduced liver damage and induces hepatic expression of liver-protective proteins SAA2 and KC. Both proteins are potent inhibitors of Con A–induced hepatitis and might also be of interest in the treatment of immune-mediated liver diseases in humans.

Methods

Animals
Specified pathogen–free 8- to 10-week-old male BALB/c mice were obtained from the Animal Research Institute of the Medizinische Hochschule Hannover. H. Hedrich, state head of the animal facility at the Hannover Medical School, approved the care of animals and experiments. For each time point and group, 3–5 animals were treated in parallel.

Generation and genotyping of gp130-mutated animals

AlfpCre gp130L^{oxP/LoxP}. Hepatocyte-specific gp130^{+/-} mice were generated by breeding alfpCre mice with mice expressing LoxP-flanked gp130 alleles. Genomic DNA was isolated and analyzed by PCR as described previously (25). Gp130L^{oxP/LoxP} mice without Cre expression were used as control (wild-type) animals.

AlfpCre gp130^{A57T/LoxP}. Hepatocyte-specific gp130^{+/-} animals were generated by breeding alfpCre mice with mice expressing LoxP-flanked gp130^{A57T/LoxP} knockin mice. Gp130^{A57T/LoxP} expresses a truncated gp130 knockin allele that lacks the essential region for the activation of STAT1 and -3 signaling (48). The genotype was detected by PCR analysis for alfpCre, gp130^{LoxP/LoxP}, and gp130^{A57T/LoxP} allele as described previously (32, 48).

AlfpCre gp130^{Y757F/LoxP}. Hepatocyte-specific gp130^{Y757F/LoxP} animals were generated by crossing alfpCre gp130^{LoxP/LoxP} with gp130^{Y757F/LoxP} knockin mice. The genotype was analyzed by PCR analysis for alfpCre, gp130^{LoxP/LoxP}, and gp130^{Y757F/LoxP} allele as described previously (25, 32).

IL-6, SAA, KC, and Con A treatment

Con A–induced hepatitis was induced by intravenous injection of 32.5 mg/kg Con A. IL-6 (200 μg/kg) was injected i.p. 3 hours before Con A injection. Recombinant SAA (0.8 mg/kg) and recombinant KC (40 μg/kg) were applied by i.v. injection 1 hour before Con A administration. Control animals were injected with an equal volume of carrier solution (NaCl). Livers of IL-6–treated animals used for array analysis (at 0 hours) were perfused. To ensure that infiltrated cells in livers of Con A–treated mice were not eliminated, we washed the organs intensively by shaking the organs for 15 minutes in cold PBS.

Preparation of liver nuclear extracts

Liver nuclear extracts were prepared as described previously (49).

Quantification of hepatic PMNs

Sections of the livers of 4 mice per group were analyzed. From each liver slide, PMNs in 6 representative high-power fields were counted on an H&E stain by an experienced liver pathologist. Student’s t test was performed to evaluate the significance of detected differences.

Detection of nuclear p-STAT3, p-STAT1, and p-ERK

Activated STAT and ERK were analyzed by detection of phosphorylated nuclear proteins. Five μg of liver nuclear proteins were separated on a 10% SDS-polyacrylamide gel and blotted onto nitrocellulose membrane. Western blot, using an antibody directed against tyrosine phosphorylated STAT3 or STAT1 (Cell Signaling Technology). Phospho-ERK1/2 expression was analyzed by use of an antibody directed against phosphorylated ERK1/2 (Cell Signaling Technology). The antigen-antibody complexes were visualized using the ECL detection system as recommended by the manufacturer (Amersham Biosciences). To verify equal loading of protein samples, a Coomassie-blue stained SDS Gel was quantified.

Detection of p-STAT3 and CD11b⁺ cells

Cryosections of frozen livers were performed and slides were fixed in a combination of methanol and acetone. We performed CD11b–P-STAT3 double immunofluorescence staining. CD11b reacts to activated macrophages and Con A–activated monocytes. It thus serves as a marker for differentiation between STAT3-positive hepatocytes and STAT3-positive Con A–activated immune cells. CD11b–P-STAT3 double staining was performed by using rat anti-mouse CD11b antibody (BD Biosciences — Pharmingen) in combination with rabbit anti-mouse P-STAT3 (Cell Signaling Technology). As a second antibody, a mix of Alexa Fluor 488 donkey anti-rat and Alexa Fluor 594 donkey anti-rabbit was used (Molecular Probes).

Determination of IL-6, TNF-α, IFN, KC, and SAA concentrations and AST activity

Blood was withdrawn by retro-orbital puncture at the indicated time points. TNF-α, IL-6, IFN-γ, KC, and SAA serum concentrations were determined by ELISA (mouse TNF; BD Biosciences — Pharmingen), mouse IL-6 (BD Biosciences — Pharmingen), mouse SAA (BioSource International), mouse KC (R&D Systems) as described previously (3).

Determination of ASTs

AST activity was determined by an automated enzyme assay as described previously (3).

Recombinant proteins

Recombinant IL-6 was produced and provided by S. Rose-John. Recombinant SAA was purchased from Cell Sciences. Recombinant KC was purchased from Strathmann Biotec AG.

Gene expression profile analysis

Affymetrix mouse genome GeneChip arrays were used to analyze gene expression of IL-6–stimulated gp130^{LoxP/LoxP} (wild-type), alfpCre gp130^{LoxP/LoxP}, alfpCre gp130^{A57T/LoxP}, and alfpCre gp130^{Y757F/LoxP} mice. Three animals per group were treated in parallel by i.p.-injected recombinant IL-6 (200 μg/kg) or NaCl. After 3 hours, animals were sacrificed. Liver RNA was isolated by guanidine isothiocyanate isolation. The RNA was used for the Affymetrix GeneChips, as described in the manufacturer’s protocol. RNA of 3 mice per group was pooled. For each mouse strain, we compared mice treated with NaCl for 3 hours with those treated with IL-6 for 3 hours. Additionally, nuclear proteins were isolated from each liver, and the activation of STAT3 and ERK1/2 was analyzed by Western blot to verify the integrity of IL-6 signaling in each mouse.

Isolation and stimulation of primary hepatocytes

Primary mouse hepatocytes were isolated by collagenase perfusion (50). Mouse liver was perfused with HEPES buffer, then with collagenase solution in a retrograde fashion. Isolated hepatocytes were washed with PBS. 1 × 10⁶ cells were plated on 6 cm dishes in DMEM containing 10% fetal calf serum and a combination of penicillin and streptomycin. The medium was changed after 4 hours. The cells were cultured under serum-free conditions overnight. The next day, cells were stimulated with 200 ng/ml recombinant

IL-6. Cells were harvested at the indicated time points, and whole cell proteins, RNA, and nuclear proteins were isolated.

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