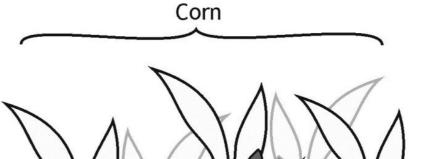


Chloramphenicol can occur in crops naturally

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Introduction

Chloramphenicol (CAP) is a broad-spectrum antibiotic that is banned for use in foodproducing animals. During the last decade, findings of CAP residues in animal products have had a major impact on international trade. Recently CAP was detected in straw and animal feed. CAP is produced for commercial use by chemical synthesis but it is also biosynthesized by soil-dwelling Actinomycetes.



We studied the hypothesis that CAP can be transferred to arable crops after biosynthesis by *Streptomyces venezuelae* in soil. Crops are processed to animal feed and used as stall bedding and thus, if the hypothesis is accepted, this can explain unexpected noncompliant CAP findings in feed and products of animal origin.

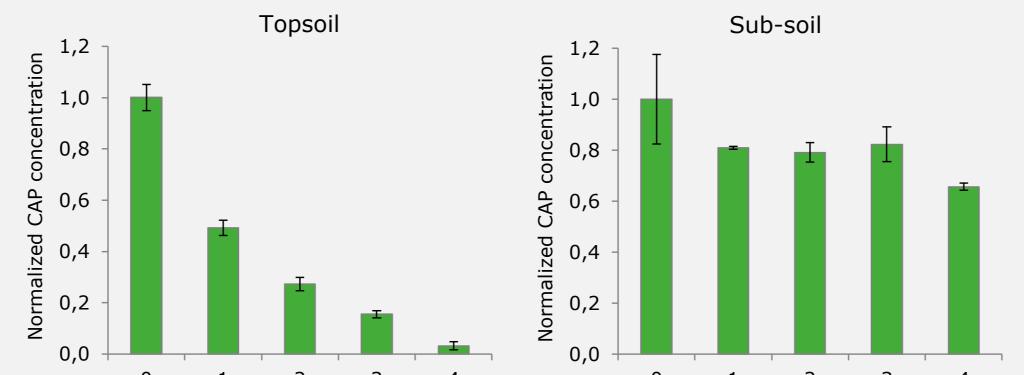
Three conditions have to be fulfilled to confirm the posed hypothesis: (A) CAP should be sufficiently stable in soil to enable transfer, (B) soil bacteria must be able to produce CAP in soil and (C) CAP should be transferred from the soil to the above ground biomass.

Wheat Stalk -30 cm -80 cm -120 cm CAP biosynthesis CAP Topsoil Sub-soil Actinomycetes

Tested hypothesis showing CAP biosyntesis in soil and the subsequent uptake by wheat and corn.

Results & discussion

A. Stability in soil CAP was added to 2 g aliquots of topsoil and subsoil and placed at room temperature. At t = 1, 2 and 3 days two aliquots of each soil were stored at -70 °C. At day 4 the CAP concentration in all aliquots was determined using a



- CAP degradation depends on soil composition; bacterial activity and organic content are the main factors.
- CAP is rapidly degraded in topsoil with a half-life of approximately one day.

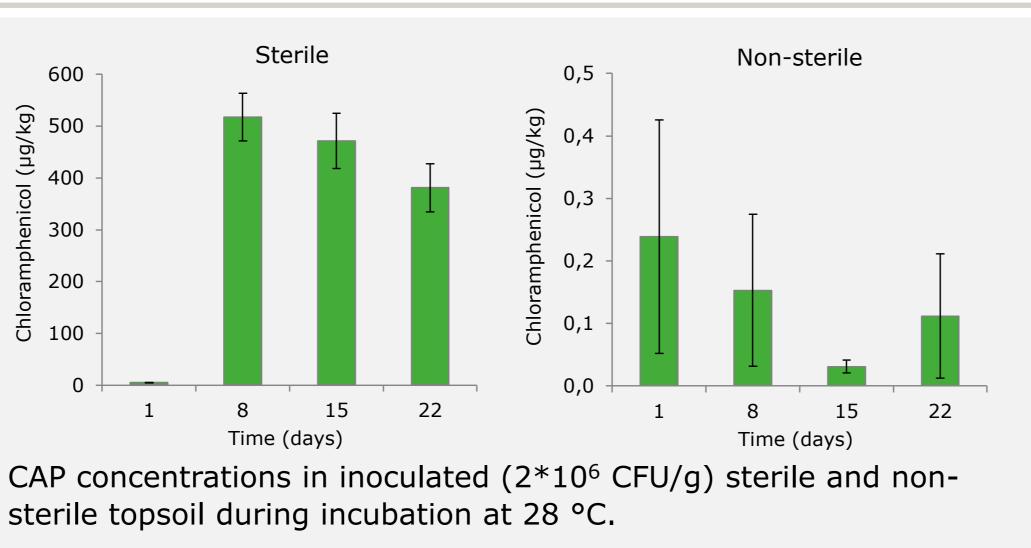
validated LC-MS/MS method.

Time (days) Time (days)

Normalized CAP concentrations in topsoil and sub-soil during incubation at room temperature.

B. Production in soil

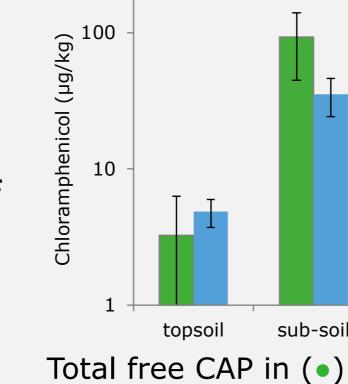
Sterilized and non-sterilized topsoil were inoculated with *Streptomyces* venezuelae at 2*10⁶ CFU g⁻¹ soil in duplicate and incubated at 28 °C. At day 1, 8, 15 and 22 the CAP concentration was determined using a validated LC-MS/MS method.



- Substantial CAP production in the sterile soil samples.
- Low CAP production in non-sterile soil: (competing) microbial flora triggers a physiological response.
- The collapse of the CAP concentration is explained by the degradation of CAP in topsoil.
- CAP is produced continuously during the experiment.

C. Transfer to crops

Wheat and corn were planted in topsoil and sub-soil. After germination and a 2 week growth phase, CAP was weekly added via 100 mL solution gifts (total of 7.5 mg CAP) via a plastic cylinder installed in the pot. After harvest the stems, spikes, stalks and cobs were analyzed separately using a validated LC-MS/MS method.





- The CAP concentration in the wheat stems and the corn stalks was significantly higher than that in the wheat spikes and maize cobs.
- <0.5% of the total administered amount of CAP was recovered in the crops as free CAP.
- CAP transfer is related to the bioavailability of the drug and thus highly depends on the experimental setup.

corn and (•) wheat harvested on topsoil and sub-soil.

sub-soil

Transfer experiment in the final growing stage.

Calculations

- For wheat, the production of in total 8 µg CAP/kg soil during the growing period is needed to yield 0.1 µg/kg CAP in wheat assuming a plant density of 200 plants/m² and an average wheat plant mass of 16 g.
- For corn this is 2 µg CAP/kg soil assuming a plant density of 10 plants/m² and an average corn plant mass of 185 g.

Conclusions

- CAP is degraded in soil but the degradation kinetics allow uptake.
- CAP can be produced by bacteria in soil in reasonable amounts.
- CAP transfer from soil to crops depends on the bioavailability of CAP.
- Low-ppb concentrations of CAP in crops can be explained by the natural production of CAP by soil bacteria and the subsequent uptake of the drug.



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