

Herpesviruses of Fish, Amphibians and Invertebrates

Steven van Beurden and Marc Engelsma
Central Veterinary Institute, part of Wageningen UR
Faculty of Veterinary Medicine, Utrecht University
The Netherlands

1. Introduction

Herpesviruses are large and complex DNA viruses which infect a wide range of vertebrates and invertebrates, including humans and domestic animals (Davison et al., 2005a). Severe infections are usually only observed in foetuses and very young or immunocompromised individuals, but economic consequences for livestock can be significant. Certain herpesviruses may also cause serious disease in non-reservoir host species - for example, Bovine malignant catharral fever. Prevention and treatment are complicated due to the ability of herpesviruses to persist as a latent infection.

In general, the occurrence of herpesviruses is host-specific (Davison, 2002). Most herpesviruses have evolved with their hosts over long periods of time and are well adapted to them. This view is supported by molecular phylogenetic data, and consistent with the generally modest pathogenicity of these viruses. In order to understand the evolutionary origin and consequences of herpesviruses infecting higher vertebrates, a profound understanding of the taxonomy of lower vertebrate and invertebrate herpesviruses is needed.

In many of the major aquaculture species, such as carp, salmon, catfish, eel, sturgeon and oyster, herpesviruses have been identified (Table 1). These herpesviruses merely represent the focus of current fish disease research, and it is expected that many other herpesviruses of lower vertebrates exist. The herpesviruses infecting fish, amphibians, and invertebrates make up two families, which are only distantly related to the well-known family of herpesviruses of mammals, birds, and reptiles (Davison et al., 2009).

In the last decades several mass mortality outbreaks in fish and oysters were found to be associated with herpesvirus infections. Management strategies are taken at fish farms to reduce the production losses due to these viruses. Several less-pathogenic fish herpesviruses are known to induce skin tumours and tumour-like proliferations (Anders & Yoshimizu, 1994), and one amphibian herpesvirus causes renal adenocarcinoma (McKinnell & Carlson, 1997). Experimental studies on these spontaneously-occurring viral tumours may serve as a model for viral carcinogenesis in humans. Interestingly, the course of infection of both the high pathogenic and tumour-inducing alloherpesviruses seems to be dependent on host age and ambient water temperature.

Genus	Virus name	Common name	Natural host(s)	Key reference for taxonomy	
<i>Batrachovirus</i>	<i>Ranid herpesvirus 1</i>	Lucké tumour herpesvirus	Leopard frog (<i>R. pipiens</i>)	(Davison et al., 2006)	
	<i>Ranid herpesvirus 2</i>	Frog virus 4	Leopard frog (<i>R. pipiens</i>)	(Davison et al., 2006)	
<i>Cyprinivirus</i>	<i>Cyprinid herpesvirus 1</i>	Carp pox	Common carp (<i>C. carpio</i>)	(Waltzek et al., 2005)	
	<i>Cyprinid herpesvirus 2</i>	Goldfish hematopoietic necrosis virus	Goldfish (<i>C. auratus</i>)	(Waltzek et al., 2005)	
	<i>Cyprinid herpesvirus 3</i>	Koi herpesvirus	Common carp (<i>C. carpio carpio</i>) & Koi (<i>C. carpio koi</i>)	(Waltzek et al., 2005)	
<i>Ictalurivirus</i>	<i>Acipenserid herpesvirus 2</i>	White sturgeon herpesvirus 2	White sturgeon (<i>A. ransmontatus</i>)	(Kurobe et al., 2008)	
	<i>Ictalurid herpesvirus 1</i>	Channel catfish virus	Channel catfish (<i>I. punctatus</i>)	(Davison, 1992)	
	<i>Ictalurid herpesvirus 2</i>	Ictalurus melas herpesvirus	Black bullhead (<i>A. melas</i>) & Channel catfish (<i>I. punctatus</i>)	(Doszpöly et al., 2008)	
<i>Salmonivirus</i>	<i>Salmonid herpesvirus 1</i>	Herpesvirus salmonis	Rainbow trout (<i>O. mykiss</i>)	(Waltzek et al., 2009)	
	<i>Salmonid herpesvirus 2</i>	Coho salmon herpesvirus	Salmon spp. (<i>Oncorhynchus</i> spp.)	(Waltzek et al., 2009)	
	<i>Salmonid herpesvirus 3</i>	Epizootic epitheliotrophic disease virus	Lake trout (<i>S. namaycush</i>)	(Waltzek et al., 2009)	
Unclassified alloherpesviruses	<i>Acipenserid herpesvirus 1</i>	White sturgeon herpesvirus 1	Sturgeon spp. (<i>Acipenser</i> spp.)	(Kurobe et al., 2008)	
	<i>Anguillid herpesvirus 1</i>	Eel herpesvirus	European eel (<i>A. anguilla</i>) & Japanese eel (<i>A. japonica</i>)	(van Beurden et al., 2010)	
	-	Pilchard herpesvirus	Pacific sardine (<i>S. sagax</i>)	(Doszpöly et al., 2011a)	
Uncharacterized alloherpesviruses	<i>Esocid herpesvirus 1</i>	Blue spot disease virus	Northern pike (<i>E. lucius</i>) & muskellunge (<i>E. masquinongy</i>)	-	
	<i>Percid herpesvirus 1</i>	Herpesvirus vitreum	Walleye (<i>S. vitreum</i>)	-	
		Pleuronectid herpesvirus 1	Herpesvirus scophthalmi	Turbot (<i>S. maximua</i>)	-
			Atlantic salmon papillomatosis virus	Atlantic salmon (<i>S. salar</i>)	-
			Herpesvirus of osmerus eperlanus 1	European smelt (<i>O. eperlanus</i>) & rainbow smelt (<i>O. mordax</i>)	-
			Tilapia larvae encephalitis virus	Tilapia (<i>Oreochromis</i> spp.)	-
			Viral epidermal hyperplasia/necrosis	Japanese flounder (<i>P. esus</i>)	-
			-	Angelfish (<i>P. altum</i>)	-
			-	Golden ide (<i>L. ide</i>)	-
			-	Red striped rockfish (<i>S. proriger</i>)	-
			-	Smooth dogfish (<i>M. canis</i>)	-
	<i>Ostreavirus</i>	<i>Ostreid herpesvirus 1</i>	Oyster herpesvirus	Japanese oyster (<i>C. gigas</i>) & other bivalves	(Davison et al., 2005b)
	Unclassified malacoherpesvirus	<i>Abalone herpesvirus 1</i>	Abalone herpesvirus	Abalone (<i>H. diversicolor supertexta</i>)	(Savin et al., 2010)

Table 1. Classified, characterized and uncharacterized fish and amphibian herpesviruses of the family *Alloherpesviridae*, and invertebrate herpesviruses of the family *Malacoherpesviridae*

Herpesviruses of fish, amphibians and invertebrates form an important and potentially large group of many yet-undiscovered pathogens. Although the fundamental characteristics of herpesviruses of humans and several other mammals have been thoroughly studied, there is still little knowledge of the herpesviruses of lower vertebrates and invertebrates. This lack of knowledge hampers the development of therapeutic and preventive strategies. This book chapter describes the fundamentals of herpesviruses infecting fish, amphibians and invertebrates, including their biology, classification and taxonomy, capsid structure and structural proteins, and genome organization and gene conservation. Where possible, a comparison with herpesviruses infecting higher vertebrates is made. An overview of the current knowledge on gene expression, latency and virulence factors of alloherpesviruses, as well as the characterization of specific genes and the development of vaccines, is given at the end of this chapter.

2. Biology of fish, amphibian and invertebrate herpesviruses

2.1 Frog herpesviruses

The North American leopard frog (*Rana pipiens*) is occasionally affected with a renal adenocarcinoma known as the Lucké tumour. A viral aetiology was proposed based on the presence of acidophilic inclusions in tumour cell nuclei, and transmission experiments (Lucké, 1934; 1938). Viral particles were observed by electron microscopy (EM) about 20 years later (Fawcett, 1956). Yet another decade later, the virus was characterized as a herpes-type virus (Lunger, 1964), later designated *Ranid herpesvirus 1* (RaHV-1). Tumour formation could be induced by injection with purified RaHV-1 (Mizell et al., 1969), and Koch-Henle's postulates were fulfilled (Naegele et al., 1974). Virus replication is promoted by low temperature (Granoff, 1999), whereas induction of metastasis is promoted by high temperature (Lucké & Schlumberger, 1949; McKinnell & Tarin, 1984). Although RaHV-1 cannot be cultured in cell lines, it was the first amphibian herpesvirus subjected to extensive genomic studies (Davison et al., 1999).

During an attempt to isolate the causative agent of the Lucké tumour from pooled urine of tumour-bearing frogs, another virus - designated frog virus 4 - was isolated using a frog embryo cell line (Rafferty, 1965). The virus was shown to possess the morphological characteristics of a herpesvirus (Gravell et al., 1968), but appeared to be clearly distinctive from the Lucké tumour herpesvirus with regard to the possibility of *in vitro* propagation and genomic properties (Gravell, 1971). In addition, frog virus 4 has no oncogenic potential, although it infects leopard frog embryos and larvae effectively (Granoff, 1999). Frog virus 4 was later designated *Ranid herpesvirus 2* (RaHV-2).

2.2 Catfish herpesviruses

In the United States, during the rapid expansion of the pond-cultured channel catfish (*Ictalurus punctatus*) industry in the late 1960s, high mortalities were reported in fingerlings and fry shortly after transfer from the hatchery to the fry ponds (Wolf, 1988). Moribund fish showed behavioural changes (swimming in spirals and hanging vertically with the head at the water surface), exophthalmus, pale or haemorrhagic gills, external and internal haemorrhages, and distension of abdomen (ascites and oedema) and stomach (Fijan et al., 1971). In 1968 a virus was isolated from various fish farms, designated channel catfish virus. A year later the virus was shown to be a herpesvirus which replicated best at 25-33 °C (Wolf

& Darlington, 1971), later designated *Ictalurid herpesvirus 1* (IcHV-1). Although mortality among young channel catfish may be very high, the effects of the disease can be minimized through management practices (Wolf, 1988). IcHV-1 has been studied extensively ever since (Kucuktas & Brady, 1999), starting with the biological properties (including pathogenicity and diagnostic possibilities), followed by molecular and structural studies, as well as vaccine development. IcHV-1 was the first fish herpesvirus for which the genome sequence became available (Davison, 1992), which had a significant impact on herpesvirus taxonomy. In the summer of 1994 another catfish herpesvirus was isolated from adult black bullhead (*Ameiurus melas*) from two different farms in Italy (Alborali et al., 1996). Morbidity and mortality were very high (80-90%), with clinical signs being unexpected spiral movements, swimming in a vertical position and death. Internal findings included haemorrhages on skin, spleen and liver. This catfish herpesvirus was shown to be different from IcHV-1, but appeared to be highly virulent for channel catfish fry and juveniles (Hedrick et al., 2003). Hence, the virus was later designated *Ictalurid herpesvirus 2* (Waltzek et al., 2009).

2.3 Carp herpesviruses

Pox disease of carp was described as early as 1563 by Conrad Gessner. Four hundred years later, herpesvirus-like particles were found to be associated with the pox-like lesions in carp (Schubert, 1966). The agent was eventually isolated in Japan, and designated *Cyprinid herpesvirus 1* (CyHV-1) (Sano et al., 1985a; Sano et al., 1985b). Infection trials showed that the virus was highly pathogenic for carp-fry, but not for older carp (Sano et al., 1990b; Sano et al., 1991). The majority of surviving carp-fry and a number of older carp developed papillomas several months after infection. Mortality and regression of the papillomas appeared to be temperature dependent (Sano et al., 1993).

The causative agent of herpesviral haematopoietic necrosis of goldfish was first identified in the early 1990s in Japan (Jung & Miyazaki, 1995), and later designated *Cyprinid herpesvirus 2* (CyHV-2). This virus may cause severe mortality especially among juvenile goldfish (Chang et al., 1999; Groff et al., 1998; Jeffery et al., 2007; Jung & Miyazaki, 1995), at water temperatures between 15 and 25 °C (Jeffery et al., 2007). Clinical signs include lethargy, anorexia and inappetence. Gross pathology includes pale gills, and swollen spleen and kidney (Jeffery et al., 2007). The disease has been reported in the USA (Goodwin et al., 2006; Goodwin et al., 2009; Groff et al., 1998), Taiwan (Chang et al., 1999), Australia (Stephens et al., 2004), UK (Jeffery et al., 2007), and Hungary (Dospoly et al., 2011b). CyHV-2 seems to be widespread on commercial goldfish farms, with outbreaks occurring when fish are subjected to stress during permissive temperatures (Goodwin et al., 2009).

In the late 1990s, mass mortalities associated with gill and skin disease occurred in the koi and common carp (*Cyprinus carpio* spp.) industries worldwide (Haenen et al., 2004). Affected fish were lethargic, anorexic and showed increased respiratory movements (Bretzinger et al., 1999; Walster, 1999). The disease is characterized by epidermal lesions, extensive gill necrosis, and an enlarged anterior kidney showing moderate damage histologically. The course is acute or peracute in most cases, with high morbidity and mortality, depending on the water temperature (15-28 °C). EM analyses revealed the presence of herpesvirus-like particles in respiratory epithelial cells of gills of affected koi carp (Bretzinger et al., 1999), and River's postulates were fulfilled subsequently (Hedrick et al., 2000). The etiological agent koi herpesvirus was shown to differ from CyHV-1 (Gilad et al., 2002), and later designated *Cyprinid herpesvirus 3* (CyHV-3) (Waltzek et al., 2005). Common and koi carp are

among the most economically important aquaculture species worldwide, with CyHV-3 still being one of the most significant threats (Michel et al., 2010). CyHV-3 has therefore been the subject of advanced fundamental and applied research in the past decade.

2.4 Salmon herpesviruses

During the early 1970s a herpesvirus was isolated from a rainbow trout (*Oncorhynchus mykiss*) hatchery in the USA, which had increased post spawning mortalities (30-50%) for several years (Wolf, 1976). The virus could only be propagated in cell cultures of salmonid origin at low temperatures (10°C) (Wolf et al., 1978). General characteristics of a herpesvirus were demonstrated by EM. Rainbow trout fry could be infected with this virus, with mortality ranging from 50-100%, but related salmonid species could not (Wolf, 1976). Symptoms observed included inappetence, lethargy, dark pigmentation, pale gills, and sometimes haemorrhagic exophthalmia (Wolf & Smith, 1981). Visceral organs and the heart showed major pathological changes, with the liver and kidneys being prime targets for the virus. The disease failed to spread by cohabitation. The etiologic agent was later designated *Salmonid herpesvirus 1* (SalHV-1).

In the same period a number of herpesviruses was isolated in Japan from different salmonid species, such as kokanee salmon (*Oncorhynchus nerka*) (Sano, 1976), masu salmon (*Oncorhynchus masou*) (Hayashi et al., 1986; Kimura et al., 1981b), yamame (*Oncorhynchus masou*) (Sano et al., 1983), coho salmon (*Oncorhynchus kisutch*) (Kumagai et al., 1994), and rainbow trout (Yoshimizu et al., 1995). The viruses appeared to be highly pathogenic particularly for young fry of different salmonid species (Kimura et al., 1983; Yoshimizu et al., 1995). The liver and kidney were the primary target organs, characterized by necrosis (Tanaka et al., 1984). Interestingly, surviving fish developed epithelial tumours around the mouth (Kimura et al., 1981a; Sano et al., 1983; Yoshimizu et al., 1987). Subsequent serological and DNA restriction endonuclease cleavage analysis demonstrated that all isolates could be considered as a single virus species, designated *Salmonid herpesvirus 2* (SalHV-2) (Hayashi et al., 1986; Hayashi et al., 1989; Hedrick et al., 1987; Yoshimizu et al., 1995).

A third salmonid herpesvirus caused high mortalities among hatchery-reared juvenile lake trout (*Salvelinus namaycush*) in the Great Lakes Region of the USA for several subsequent springs and falls (6-15°C) during the mid-1980s (Kurobe et al., 2009). Epidemics were characterized by the rapid onset of mortality, followed by a number of nonspecific clinical signs, including corkscrew swimming, lethargy, and periods of hyperexcitability (Bradley et al., 1989). External symptoms included haemorrhages in the eyes, fin and skin degeneration, and secondary fungus infections (Bradley et al., 1989; McAllister & Herman, 1989). Mortality could be as much as 100%, with fry being more susceptible than fingerlings. In the late 1980s herpesvirus-like particles were associated with the disease. Transmission experiments demonstrated that *Salmonid herpesvirus 3* (SalHV-3) was the etiologic agent of the epizootic epitheliotropic disease restricted to lake trout. Propagation of the virus in cell culture is still impossible, which hampered detection until very recently (Kurobe et al., 2009).

A fourth salmonid herpesvirus has been described, but not yet characterized. In the late 1970s benign proliferative epidermal papillomatous lesions of cultured Atlantic salmon (*Salmo salar*) in Scandinavia and the UK were investigated (Bylund et al., 1980; Carlisle & Roberts, 1977). Papillomas developed in July and August, after which they sloughed, and in December nearly all were gone. The papillomas appeared as white plaques which raised several millimetres and were up to several centimetres in diameter. They were frequently

multiple and found anywhere on the skin and fins posterior to the head. Morbidity and mortality were generally low. Virus-like particles were observed using EM in samples from papillomatous tissue (Carlisle, 1977), later characterized as a herpesvirus in Atlantic salmon from Russia (Shchelkunov et al., 1992). Attempts to isolate the observed virus in cell culture failed (Carlisle, 1977; Shchelkunov et al., 1992).

2.5 Sturgeon herpesviruses

In 1991 a herpesvirus was isolated from juvenile white sturgeon (*Acipenser transmontanus*) from a commercial farm in California, USA (Hedrick et al., 1991). The white sturgeon herpesvirus 1, later designated *Acipenserid herpesvirus 1* (AciHV-1), was associated with infections of the tegument and oropharyngeal mucosa, and mortality among the juvenile white sturgeons. Experimentally induced infections also resulted in mortality. AciHV-1 was later isolated from other farmed white sturgeons in California and Italy (Kelley et al., 2005; Kurobe et al., 2008).

Few years later another herpesvirus called white sturgeon herpesvirus 2, or *Acipenserid herpesvirus 2* (AciHV-2), was isolated from dermal lesions of subadult white sturgeon and ovarian fluids of a mature white sturgeon (Watson et al., 1995). Mortality among experimentally infected juvenile white sturgeons reached a cumulative total of 80%. AciHV-2 has since been isolated from wild white sturgeon in Idaho and Oregon (USA), from farmed shortnose sturgeon (*Acipenser brevirostrum*) from Canada, and from Siberian sturgeon (*Acipenser baeri*) in Russia (Doszpoly & Shchelkunov, 2010; Kelley et al., 2005; Kurobe et al., 2008; Shchelkunov et al., 2009).

2.6 Eel herpesvirus

Herpesvirus-like particles in wild European eels (*Anguilla anguilla*) were described for the first time in 1986 (Békési et al., 1986). A herpesvirus was later isolated from cultured European eels and Japanese eels (*A. japonica*) in Japan (Sano et al., 1990a). Serological, molecular, and sequence data indicated that Asian and European eel herpesvirus isolates can be considered as a single virus species (Chang et al., 2002; Lee et al., 1999; Rijsewijk et al., 2005; Sano et al., 1990a; Ueno et al., 1992; Ueno et al., 1996; Waltzek et al., 2009), designated *Anguillid herpesvirus 1* (AnGHV-1). Clinical and pathological findings of the infection varied among and within outbreaks, and were predominantly apathy, haemorrhages and ulcerative lesions in skin and fins, haemorrhagic or pale and congested gills, a pale spleen, a pale and haemorrhagic liver, a distended gall bladder, and ascites (Chang et al., 2002; Davidse et al., 1999; Haenen et al., 2002; Lee et al., 1999; Sano et al., 1990a). AnGHV-1 infection in cultured eels resulted in decreased growth rates and an increased mortality (Haenen et al., 2002). The virus is also frequently observed in wild European eels (Haenen et al., 2010; Jørgensen et al., 1994; van Ginneken et al., 2004).

2.7 Pilchard herpesvirus

In 1995, a massive epizootic occurred in adult Australasian pilchards (*Sardinops sagax neopilchardus*) in south Australia (O'Neill, 1995). In several months it spread thousands of kilometres bidirectionally along the Australian coastline, and then to New Zealand (Whittington et al., 1997). A similar event occurred a few years later in 1998/1999 (Murray et al., 2003). Affected pilchards showed progressive gill inflammation followed by epithelial hypertrophy and hyperplasia (Whittington et al., 1997). Consequent clinical symptoms

included hypoxaemia and hypercapnea, resulting in an estimated mortality of at least 10%. Involvement of an infectious agent was suggested, and PCR analysis revealed the putative involvement of a herpesvirus (Tham & Moon, 1996), which was soon confirmed by EM (Hyatt et al., 1997). Diagnostic tools for detection of the pilchard herpesvirus have been developed (Crockford et al., 2005; Crockford et al., 2008), revealing that the virus is now endemic in Australian pilchard populations (Whittington et al., 2008). Although the pilchard herpesvirus has not yet been isolated in cell culture, hampering further studies, limited phylogenetic analysis showed its relation to other fish and frog herpesviruses (Doszpoly et al., 2011a).

2.8 Other fish herpesviruses

Several other herpesvirus-like particles have been observed and found to be associated with disease in other fish species. Many of these viruses have not been isolated yet, however, and limited sequence availability hampers official classification.

In the late 1970s a herpes-type viral infection of the epithelia of the skin and gills of turbot (*Scophthalmus maximus*) was found, presumably associated with heavy mortalities among farmed turbot (Buchanan & Madeley, 1978; Buchanan et al., 1978). Although the virus could not be isolated in cell culture, EM observations clearly demonstrated herpesvirus characteristics. The virus was tentatively named herpesvirus scophthalmi, later referred to as pleuronectid herpesvirus 1 (PIHV-1).

Few years later a herpesvirus was isolated from hyperplastic epidermal tissue from a walleye (*Stizostedion vitreum vitreum*) taken in Saskatchewan, Canada (Kelly et al., 1983). The virus was isolated in a walleye ovarian cell line and identified as a herpesvirus based on size, morphology, and apparent pattern of replication. The virus was initially called herpesvirus vitreum, but later designated percid herpesvirus 1 (PeHV-1).

A year later the same research group observed typical herpesvirus particles in epidermal hyperplasia or blue spot disease of northern pike (*Esox lucius*) in several waters of central Canada (Yamamoto et al., 1984). The virus, tentatively designated esocid herpesvirus 1 (EsHV-1), could not be isolated in cell culture, however. EsHV-1 was later also observed in northern pike and muskellunge (*Esox masquinongy*) in the USA, with clinical signs present only for a short period when water temperatures were between 2 and 13 °C (Margenau et al., 1995), and in northern pike in Ireland (Graham et al., 2004).

In the early 1980s, in a quarantined population of golden ide (*Leuciscus idus*) imported from Germany to the USA, 5% of the fish developed carp pox-like lesions (McAllister et al., 1985). EM analysis showed herpesvirus-like particles associated with the lesions, but the virus could not be isolated in cell culture.

In 1985 herpesvirus-like particles were observed in so-called spawning papillomas on the skin and fins of smelt (*Osmerus eperlanus*) in Germany (Anders & Möller, 1985). Herpesvirus-like particles were also observed in similar epidermal tumours of rainbow smelt (*O. mordax*) during spawning-time in Canada and the USA (Herman et al., 1997; Morrison et al., 1996). Isolation and genomic analysis of the herpesvirus of smelt have failed so far (Jakob et al., 2010).

In the same year a viral dermatitis was observed in a small percentage of wild and captive smooth dogfish (*Mustelus canis*) (Leibovitz & Leboutitz, 1985). Most skin eruptions were found on the tail and fins, often exceeding 1 cm in diameter, and histologically characterized by epidermal cell degeneration. EM revealed the progressive development of cellular

pathology associated with a specific epidermal viral infection, and viral particles showed the characteristics of a herpesvirus.

In the mid-1980s a new disease characterized by skin and fin opacity, and associated with high mortality occurred in larvae and juveniles of Japanese flounder (*Paralichthys olivaceus*) hatcheries in Japan (Iida et al., 1989; Miyazaki et al., 1989). Microscopically, epidermal cells at the surface were rounded, and histopathologically the epidermis appeared to be hyperplastic and necrotic (Iida et al., 1991; Miyazaki, 2005). Although the causative agent could not be isolated, herpesvirus-like particles were observed by EM. Infection trials showed that especially larvae <10 mm were susceptible, at water temperatures around 20 °C (Masumura et al., 1989).

After a stress-trigger, all three angelfish (*Pterophyllum altum*) in a fish tank showed loss of equilibrium and hung apathetic at the water surface (Møllergaard & Bloch, 1988). The fish had skin haemorrhages, pale gills and distended liver and spleen. Histopathological findings of the spleen were all suggestive of severe haemolytic anaemia. Herpesvirus-like particles were observed by EM in the nuclei of spleen macrophages and monocytes.

A wild redstriped rockfish (*Sebastes proriger*) demonstrated hepatic lesions, which were histologically suggestive of a herpesvirus-type infection (Kent & Myers, 2000). EM analysis of suboptimally preserved liver tissue revealed the presence of intranuclear particles, consistent in size and shape with herpesvirus capsids.

Recently, an outbreak of a novel disease characterized by a whirling syndrome and high mortality rates occurred in laboratory-reared tilapia larvae (*Oreochromis* spp.) (Shlapobersky et al., 2010). The disease was designated viral encephalitis of tilapia larvae. The morphological characteristics of virus particles found in the brains of diseased larvae as observed by EM were comparable to those of a herpesvirus.

2.9 Oyster herpesvirus

The first description of a herpesvirus-like virus in oysters originates from the United States (Farley et al., 1972). As for the fish and amphibian herpesviruses, it was only the virus morphology that identified this oyster virus as a herpesvirus. The principle susceptible species is the Japanese oyster (*Crassostrea gigas*), and the virus was designated *Ostreid herpesvirus 1* (OsHV-1). In Europe, where the virus was found for the first time in the early 1990s, OsHV-1 has been associated with high mortalities in oyster seedlings (Garcia et al., 2011). Herpesvirus-like particles – later confirmed to be OsHV-1 by molecular analyses – have also been observed in various other bivalve species, associated with mortalities (Arzul et al., 2001; Burge et al., 2011). Recently, an OsHV-1 variant, designated OsHV-1 μ var, was identified as the causative agent of high mortalities in juvenile oysters (Segarra et al., 2010).

2.10 Abalone herpesvirus

A second herpesvirus with an invertebrate host has been identified by EM in cultured abalone (*Haliotis diversicolor supertexta*) in Taiwan in early 2003 and in Australia in late 2005 (Chang et al., 2005; Tan et al., 2008). Abalones of all ages suffered from the disease, which was characterized by mantle recession and muscle stiffness, followed by high mortality within a few days. Histologically the nerve system appeared to be the primary target tissue, characterized by tissue necrosis with infiltration of haemocytes. The virus has been designated *Abalone herpesvirus 1* (AbHV-1).

3. Classification and taxonomy of the order *Herpesvirales*

Herpesviruses are large and complex DNA viruses with a distinctive virion morphology, which consists of four distinct structures: the core, capsid, tegument, and envelope (Davison et al., 2005a). The order *Herpesvirales* is subdivided into three families (Davison et al., 2009), which separated about 500 million years ago (McGeoch et al., 2006). The family *Herpesviridae* comprises the herpesviruses with mammalian, avian and reptilian hosts, and is further subdivided into the subfamilies *Alpha-*, *Beta-* and *Gammaherpesvirinae*, which fall into genera.

The family *Alloherpesviridae* comprises the piscine and amphibian herpesviruses (Waltzek et al., 2009). The criteria for the establishment of genera within the family *Alloherpesviridae* have not been defined yet, nor have rules been formulated for deciding whether species should belong to a particular genus. Currently, genera and their respective species are assigned largely based on available phylogenetic analyses, with phylogenetically closely related species assigned to the same genus. To date, four genera in the family *Alloherpesviridae* have been established: *Batrachovirus*, *Cyprinivirus*, *Ictalurivirus* and *Salmonivirus*, comprising a total of 11 different species (Table 1, Fig. 1). In the majority of the cases the classification follows the grouping of alloherpesviruses infecting the same host. At least another 14 herpesviruses infecting fish have been described, but not yet characterized sufficiently to allow classification.

The oyster herpesvirus OsHV-1 has been assigned to the genus *Ostreavirus* of the third herpesvirus family *Malacoherpesviridae*, which comprises the invertebrate herpesviruses (Davison et al., 2009; Davison et al., 2005b). The other invertebrate herpesvirus infecting abalone AbHV-1 has been shown to be related to OsHV-1 (Savin et al., 2010). Despite the similarities in capsid structure between the herpesviruses of mammals, birds and reptiles, fish and amphibians, and invertebrates, the three families are highly divergent. Only a single gene, encoding a DNA packaging enzyme complex distantly related to the ATPase subunit of bacteriophage T4 terminase, is convincingly conserved among all herpesviruses (Davison, 1992; 2002).

4. Conservation of herpesvirus capsid architecture & structural proteins

4.1 Capsid architecture of herpesviruses

Despite their diversity by genes, host range, and genome size, herpesvirus structure is conserved throughout the entire order *Herpesvirales*. Herpesvirus virions invariably consist of a large (diameter >100 nm) thick-walled icosahedral nucleocapsid ($T=16$), surrounded by a host-derived envelope with a diameter of about 200 nm, with an intervening proteinaceous layer called the tegument (Booy et al., 1996). This appearance is distinctively different from that of any other animal virus. Key to this morphological conservatism is the icosahedral nucleocapsid made up of hollow capsomers. The functionality of the genes involved in capsid assembly is largely conserved.

The capsid structures of alloherpesvirus IcHV-1 and malacoherpesvirus OsHV-1 have been studied by cryoelectron microscopy and three-dimensional image reconstruction (Booy et al., 1996; Davison et al., 2005b). Both viruses have capsids with diameters of approximately 116 nm, which is slightly smaller than the capsids of the human model herpesvirus, Herpes simplex virus type 1 (HSV-1, diameter 125 nm). The capsids of IcHV-1 and OsHV-1 are roughly hexagonal in outline and reconstruction revealed an icosahedral structure with a

triangulation number of $T=16$. The icosahedral facets of ICHV-1 appeared to be flatter, and the shell is about 20% thinner (12.4 nm vs. 15 nm) than that of HSV-1, consistent with the smaller size of its major capsid protein. The hexons and pentons showed protrusions with an axial channel through each capsomer. The outer surface is composed of heterotrimeric complexes or triplexes at the sites of local threefold symmetry, and the inner surface has a relatively flat and featureless appearance. ICHV-1 and OsHV-1 capsids are therefore similar in overall appearance to those of all other herpesviruses studied to date.

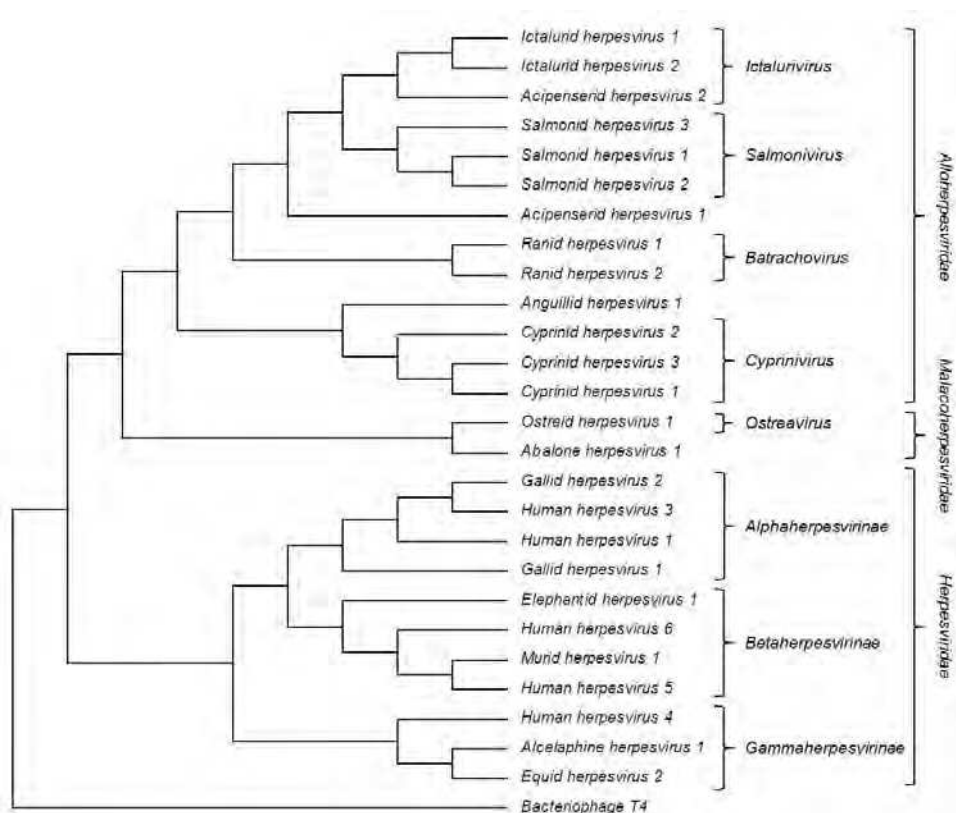


Fig. 1. Phylogenetic tree depicting the relationships (topology only) among viruses in the order *Herpesvirales*, based on partially deduced amino acid sequence of the terminase gene (107 residues, including gaps), analysed with the maximum likelihood method, using the JTT matrix (1,000 replicates), and rooted with bacteriophage T4

4.2 Structural proteins of *Alloherpesviridae*

Complete herpesvirus virions are made up of about 40 viral proteins. Recent mass spectrometry (MS) analyses showed that HSV-1 virions comprise at least 8 viral capsid proteins, 13 viral glycoproteins, 23 potential viral teguments, and 49 host proteins (Loret et al., 2008). When in 1995 the structural proteins of ICHV-1 were identified by MS (Davison & Davison, 1995), a total of 11 genes encoding 15 virion proteins was identified. Based on

analyses of capsid, capsid-tegument, and envelope fractions, the proteins could be assigned to the different compartments of the virion.

More recent studies on the structural proteins of CyHV-3 and AngHV-1 used liquid chromatography tandem MS-based proteomic approaches. For CyHV-3 a total of 40 structural proteins was identified, which were classified based on homology and bioinformatic analyses as capsid (3), envelope (13), tegument (2) and unclassified (22) structural proteins (Michel et al., 2010). For AngHV-1, by analysing separately prepared fractions of capsid, tegument, and envelope, the identified proteins could also be functionally characterized. A total of 40 structural proteins was identified, of which 7 could be assigned to the capsid, 11 to the envelope, and 22 to the tegument (van Beurden et al., 2011).

The protein composition of fish herpesvirus capsids generally mirrors that of mammalian herpesvirus capsids. MS analyses of purified alloherpesvirus capsids demonstrated the presence of the major capsid protein, the capsid triplex protein 2, and, presumably, the capsid triplex protein 1 (Davison & Davison, 1995; van Beurden et al., 2011). The protease-and-scaffolding protein is only abundantly present in premature capsids, and later replaced by viral DNA during maturation. The small protein which forms the hexon tips in mammalian herpesviruses has not been found in alloherpesvirus capsids.

Conservation of tegument and envelope proteins among fish herpesviruses is rather limited (van Beurden et al., 2011). A large tegument protein was found to be partially conserved, and two envelope proteins might be conserved. The higher conservation of structural proteins between members of the family *Herpesviridae* resembles the greater evolutionary distance of the family *Alloherpesviridae*.

5. Genome organization and gene conservation

5.1 Genome properties

The single linear double stranded DNA genomes of herpesviruses vary greatly in size, ranging from 124 kbp for Simian varicella virus, to more than 250 kbp. This enormous variation is also present within the family *Alloherpesviridae*, in which ICHV-1 has the smallest genome of 134 kbp, and AngHV-1 and CyHV-3 represent the largest herpesvirus genomes known of 249 and 295 kbp, respectively (Table 2). G+C content varies from 32 to 75% within the family *Herpesviridae* (McGeoch et al., 2006), but seems to be more restricted among members of the family *Alloherpesviridae* (52.8 to 59.2%), and is rather low for OsHV-1 (38.7%).

5.2 Genome organization

Herpesvirus genomes characteristically contain one or two regions of unique sequence flanked by direct or inverted repeats (McGeoch et al., 2006). To date, six different classes of genome organization have been identified (Davison, 2007). The genome organization of six alloherpesviruses has been determined. The genomes of AngHV-1, CyHV-3, ICHV-1, RaHV-1 and RaHV-2 all consist of one long unique region (U) flanked by two short direct repeat regions (TR) at the termini (Fig. 2). This genome structure (A) is also represented among the *Betaherpesvirinae* (Davison, 2007). Interestingly, the terminal repeats of the ranid herpesviruses are considerably shorter (<1-kbp) than those of the fish herpesviruses (>10-kbp). Genome organization A does not seem to be a general feature of the *Alloherpesviridae*,

since SalHV-1 is known to have a long unique region (U_L) linked to a short unique region (U_S) flanked by an inverted repeat (IR_S and TR_S) (Davison, 1998). This genome structure (D) is characteristic of *Alphaherpesvirinae* in the *Varicellovirus* genus (Davison, 2007).

Name	Genome length	Terminal repeat length	G+C content	Number of ORFs	ORF density per kpb	Refseq accession	Key reference
<i>Anguillid herpesvirus 1</i>	248,531 bp	10,634 bp	53.0%	136	0.57	NC_013668	(van Beurden et al., 2010)
<i>Cyprinid herpesvirus 3</i>	295,146 bp	22,469 bp	59.2%	156	0.57	NC_009127	(Aoki et al., 2007)
<i>Ictalurid herpesvirus 1</i>	134,226 bp	18,556 bp	56.2%	77	0.67	NC_001493	(Davison, 1992)
<i>Ranid herpesvirus 1</i>	220,859 bp	636 bp	54.6%	132	0.60	NC_008211	(Davison et al., 2006)
<i>Ranid herpesvirus 2</i>	231,801 bp	912 bp	52.8%	147	0.64	NC_008210	(Davison et al., 2006)
<i>Ostreid herpesvirus 1</i>	207,439 bp	7584 bp 9774 bp	38.7%	124	0.65	NC_005881	(Davison et al., 2005b)

Table 2. Genome characteristics of completely sequenced members of the families *Alloherpesviridae* and *Malacoherpesviridae*

Genome structure E is the most complex genome structure and characteristic of *Alphaherpesvirinae* in the *Simplexvirus* genus, and certain members of the genus *Cytomegalovirus* of the *Betaherpesvirinae* (Davison, 2007). The gross genome organization of OsHV-1 is a combination of class D and E genomes, consisting of two invertible unique regions (U_L & U_S), each flanked by inverted repeats (TR_L & IR_L and IR_S & TR_S), with an additional short unique sequence (X) between the inverted repeats (Davison et al., 2005b).

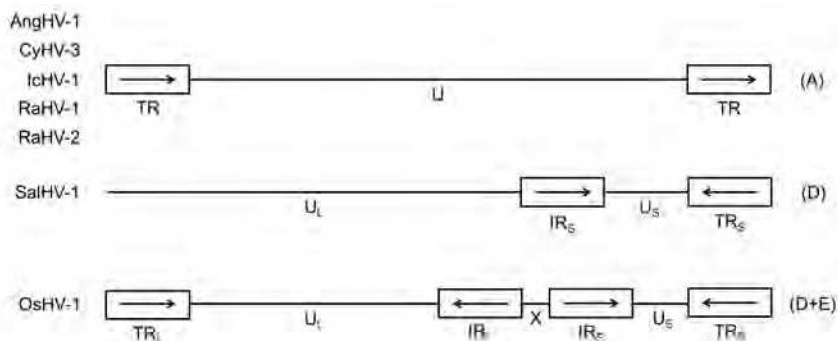


Fig. 2. Schematic representation of the genome organization of selected members of the families *Alloherpesviridae* and *Malacoherpesviridae*

5.3 Gene conservation

Herpesvirus genomes show a wide range in number of genes, ranging from about 70 (*Varicella zoster virus*) to more than 200 (*Human cytomegalovirus*) genes (Davison, 2007; McGeoch et al., 2006). Herpesvirus genes are divided into core genes and non-core genes.

The core genes are inherited from a common ancestor and fundamental to replication, being involved in capsid assembly and structure, DNA replication machinery, processing and packaging of DNA, egress of capsids from the nucleus, and control and modulation. The non-core genes represent accessory systems that developed more recently and fit a virus to a particular biological niche. These genes are involved in cellular tropisms, control of cellular processes, manipulation or evasion from the host immune system, and latency. Among the *Herpesviridae*, there is a subset of 43 core genes (McGeoch & Davison, 1999).

Only 13 genes are convincingly conserved among all members of the family *Alloherpesviridae* (Aoki et al., 2007). These genes encode proteins putatively involved in capsid morphogenesis, DNA replication and DNA packaging (Table 3). Five of the identified conserved genes encode proteins with unknown functions. With only 13 genes conserved among all family members, the family *Alloherpesviridae* appears to be considerably more divergent than the family *Herpesviridae*, reflecting the greater divergence of its host species.

Function	AngHV-1	CyHV-3	IcHV-1	RaHV-1	RaHV-2
ATPase subunit of terminase	ORF10	ORF33	ORF62	ORF42	ORF68
Primase	ORF21	ORF46	ORF63	ORF87	ORF121
<i>Unknown</i>	ORF22	ORF47	ORF64	ORF88	ORF122
Capsid triplex protein 2	ORF36	ORF72	ORF27	ORF95	ORF131
DNA helicase	ORF37	ORF71	ORF25	ORF93	ORF129
<i>Unknown</i>	ORF52	ORF80	ORF60	ORF84	ORF118
DNA polymerase	ORF55	ORF79	ORF57	ORF72	ORF110
Capsid protease and scaffolding protein	ORF57	ORF78	ORF28	ORF63	ORF88
Large envelope glycoprotein	ORF67	ORF99	ORF46	ORF46	ORF72
<i>Unknown</i>	ORF82	ORF61	ORF54	ORF75	ORF113
<i>Unknown</i>	ORF98	ORF107	ORF56	ORF73	ORF111
<i>Unknown</i>	ORF100	ORF90	ORF37	ORF52	ORF78
Major capsid protein	ORF104	ORF92	ORF39	ORF54	ORF80

Table 3. Conserved genes of the family *Alloherpesviridae*

The arrangement of homologous genes in AngHV-1 and CyHV-3 appears to be conserved in clusters (van Beurden et al., 2010). Several conserved gene blocks can be identified, either in the same or in inverse orientation. At the *Alloherpesviridae* family level, the 13 conserved genes seem to be conserved within the same clusters, which was previously shown also for IcHV-1 and SalHV-1 (Davison, 1998), and appears to apply for all completely sequenced *Alloherpesviridae* (Fig. 3). This resembles the seven blocks of core genes that are typically arranged throughout the family *Herpesviridae* (McGeoch et al., 2006).

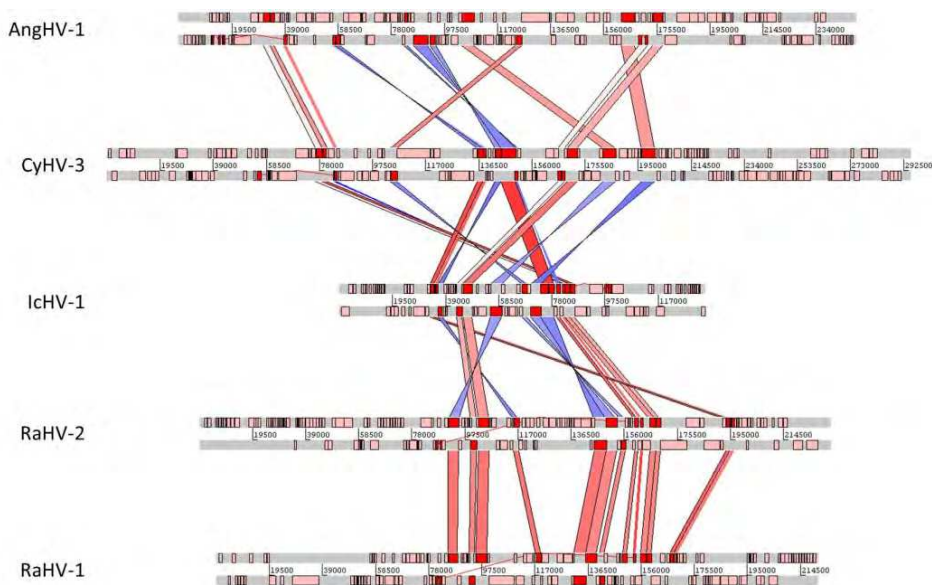


Fig. 3. Geneblock conservation throughout the family *Alloherpesviridae*: conserved genes are coloured red and connected through the genomes by coloured bars; red bars indicate similar and reading frame direction, blue bars indicate reversed reading frame direction; percentage sequence identity at amino acid level is indicated by bar colour intensity

6. Extended characterization of *Alloherpesviridae*

6.1 Gene expression

Gene expression of herpesviruses is regulated in a temporal fashion, with genes being classified on the basis of their expression kinetics as immediate early, early, and late genes (McGeoch et al., 2006). Temporal gene expression has been suggested for IcHV-1 based on polypeptide analyses of *in vitro* infection experiments (Dixon & Farber, 1980). Transcription studies of selected ORFs, especially of the terminal repeat region of IcHV-1, confirmed the temporal expression of these genes (Huang & Hanson, 1998; Silverstein et al., 1995; Silverstein et al., 1998; Stingley & Gray, 2000). Expression of certain ORFs of CyHV-3 at lower and higher temperatures *in vitro* has been studied using RT-PCR (Dishon et al., 2007). Initial experiments to determine the complete transcription profile of AngHV-1 by quantitative RT-PCR showed similar temporal expression kinetics as have been determined for members of the family *Herpesviridae* (van Beurden et al., unpublished results).

6.2 Latency

The ability to establish a latent infection is one of the hallmarks of herpesviruses. Latent infections are characterized by the absence of infectious virus particles and regular viral transcription and replication, but the presence of intact viral genomic DNA and transcription of latency associated transcripts. Accordingly, the presence of viral DNA (and not infectious virus) has been demonstrated in fish surviving infections with IcHV-1 and CyHV-3 (Eide et al., 2011a; Gray et al., 1999), whereas viral replication was absent in such

fish (Eide et al., 2011b; Stingley et al., 2003). It has been shown *in vitro* that CyHV-3 is able to persist in cultured cells at a nonpermissive temperature, with viral propagation and viral gene transcription being turned off, and reactivated upon return to the permissive temperature (Dishon et al., 2007). Reactivation of fish herpesviruses *in vivo* has been demonstrated for AngHV-1 following dexamethasone treatment (van Nieuwstadt et al., 2001), and for CyHV-3 following temperature stress (Eide et al., 2011b; St-Hilaire et al., 2005).

6.3 Gene characterization

Identification and characterization of most alloherpesvirus ORFs is based on sequence homology and bioinformatics. Recent probabilistic mapping of IchV-1 (Kunec et al., 2009), and complete transcriptome analysis of AngHV-1 (van Beurden et al., unpublished results), show, however, that there might still be a large discrepancy between the predicted and actual numbers of ORFs.

MS analysis of purified capsids resulted in the identification of the structural proteins of IchV-1, CyHV-3 and AngHV-1 (Davison & Davison, 1995; Michel et al., 2010; van Beurden et al., 2011). Based on the localization of the structural proteins in the different virion compartments, several of these proteins could be functionally characterized. Two alloherpesvirus proteins have been characterized in more detail: IchV-1 ORF50 has been shown to encode a secreted mucin-like glycoprotein (Vanderheijden et al., 1999), and CyHV-3 ORF81 has been identified as an envelope protein (Rosenkranz et al., 2008).

6.4 Virulence factors and vaccine development

In many herpesviruses the homologues of cellular enzymes are nonessential for virus replication *in vitro*, but relevant for virulence *in vivo*. Attenuated IchV-1 and CyHV-3 strains with specific gene deletions have been developed and tested for virulence. Thymidine kinase-negative mutants of IchV-1 resembled wild type IchV-1 *in vitro*, but appeared to be much less pathogenic *in vivo* (Zhang & Hanson, 1995). Similarly, CyHV-3 recombinants possessing deletions within the viral ribonucleotide reductase, thymidine kinase and dUTPase genes were developed and tested for attenuation (Fuchs et al., 2011). The use of the attenuated strains as modern live vaccines in challenges with wild-type virus showed promising results.

Several other attempts of vaccine development have been undertaken for IchV-1 and CyHV-3. For CyHV-3, immunization with wild type and attenuated virus has been tested in relation to different water temperatures (Ronen et al., 2003). For IchV-1, experimental subunit and attenuated vaccines were developed and tested (Dixon, 1997). Also, DNA vaccination with several different ORFs has been tested for IchV-1 (Nusbaum et al., 2002). Later, an overlapping and a full BAC have been developed for IchV-1 (Kunec et al., 2008) and CyHV-3 (Costes et al., 2008), respectively. The latter was used to test attenuation after disruption of the thymidine kinase locus and deletion of ORF16. To date no satisfying vaccine has been developed nor registered for any of the fish herpesviruses.

7. Conclusions

In conclusion, herpesviruses of fish, amphibians and invertebrates form a potentially large group of important pathogens. Understanding the origins of the distantly related families

Alloherpesviridae and *Malacoherpesviridae* provides insights into the evolution of the family *Herpesviridae*. The association of many fish herpesviruses with severe diseases in important aquaculture species has stimulated fundamental and applied research. For a selected number of alloherpesviruses, complete genome sequences and structural analyses of the virus particles are now available and have revealed similarity to the basic composition of higher vertebrate herpesviruses. Extended characterization of gene expression, latency, the function of specific genes and virulence factors as well as vaccine development is currently ongoing. These studies will add to the control and prevention of fish herpesvirus associated diseases, and might serve as models for research on human and domestic animal herpesviruses.

8. Acknowledgements

We thank Alex Bossers (Central Veterinary Institute (CVI), part of Wageningen UR) for generating Fig. 3. We are grateful to Olga Haenen (CVI), Ben Peeters (CVI), Fred van Zijderveld (CVI), and Peter Rottier (Faculty of Veterinary Medicine, Utrecht University) for a critical reading of this book chapter.

9. References

- Alborali, L., Bovo, G., Cappellaro, H. & Guadagini, P.F. (1996). Isolation of an herpesvirus in breeding catfish (*Ictalurus mela*). *Bull Eur Ass Fish Pathol*, Vol. 16, No. 4, pp. 134-137
- Anders, K. & Möller, H. (1985). Spawning papillomatosis of smelt, *Osmerus eperlanus* L., from the Elbe estuary. *J Fish Dis*, Vol. 8, pp. 233-235
- Anders, K. & Yoshimizu, M. (1994). Role of viruses in the induction of skin tumours and tumour-like proliferations of fish. *Dis Aquat Org*, Vol. 19, pp. 215-232
- Aoki, T., Hirono, I., Kurokawa, K., Fukuda, H., Nahary, R., Eldar, A., Davison, A.J., Waltzek, T.B., Bercovier, H. & Hedrick, R.P. (2007). Genome sequences of three koi herpesvirus isolates representing the expanding distribution of an emerging disease threatening koi and common carp worldwide. *J Virol*, Vol. 81, No. 10, pp. 5058-5065
- Arzul, I., Renault, T., Lipart, C. & Davison, A.J. (2001). Evidence for interspecies transmission of oyster herpesvirus in marine bivalves. *J Gen Virol*, Vol. 82, No. 4, pp. 865-870
- Békési, L., Horváth, I., Kovács-Gayer, E. & Csaba, G. (1986). Demonstration of herpesvirus like particles in skin lesions of European eel (*Anguilla anguilla*). *J Appl Ichthyol*, Vol. 4, pp. 190-192
- Booy, F.P., Trus, B.L., Davison, A.J. & Steven, A.C. (1996). The capsid architecture of channel catfish virus, an evolutionarily distant herpesvirus, is largely conserved in the absence of discernible sequence homology with herpes simplex virus. *Virology*, Vol. 215, No. 2, pp. 134-141
- Bradley, T.M., Medina, D.J., Chang, P.W. & McClain, J. (1989). Epizootic epitheliotropic disease of lake trout (*Salvelinus namaycush*): history and viral etiology. *Dis Aquat Org*, Vol. 7, pp. 195-201
- Bretzinger, A., Fischer-Scherl, T., Oumouna, M., Hoffmann, R. & Truyen, U. (1999). Mass mortalities in Koi carp, *Cyprinus carpio*, associated with gill and skin disease. *Bull Eur Ass Fish Pathol*, Vol. 19, No. 5, pp. 182-185

- Buchanan, J.S. & Madeley, C.R. (1978). Studies on *Herpesvirus scophthalmi* infection of turbot *Scophthalmus maximus* (L.) ultrastructural observations. *J Fish Dis*, Vol. 1, pp. 283-295
- Buchanan, J.S., Richards, R.H., Sommerville, C. & Madeley, C.R. (1978). A herpes-type virus from turbot (*Scophthalmus maximus* L.). *Vet Rec*, Vol. 102, No. 24, pp. 527-528
- Burge, C.A., Strenge, R.E. & Friedman, C.S. (2011). Detection of the oyster herpesvirus in commercial bivalve in northern California, USA: conventional and quantitative PCR. *Dis Aquat Org*, Vol. 94, No. 2, pp. 106-116
- Bylund, G., Valtonen, E.T. & Niemelä, E. (1980). Observations on epidermal papillomata in wild and cultured Atlantic salmon *Salmo salar* L. in Finland. *J Fish Dis*, Vol. 3, pp. 525-528
- Carlisle, J.C. (1977). An epidermal papilloma of the Atlantic salmon II: Ultrastructure and etiology. *J Wildl Dis*, Vol. 13, No. 3, pp. 235-239
- Carlisle, J.C. & Roberts, R.J. (1977). An epidermal papilloma of the Atlantic salmon I: Epizootiology, pathology and immunology. *J Wildl Dis*, Vol. 13, No. 3, pp. 230-234
- Chang, P.H., Kuo, S.T., Lai, S.H., Yang, H.S., Ting, Y.Y., Hsu, C.L. & Chen, H.C. (2005). Herpes-like virus infection causing mortality of cultured abalone *Haliotis diversicolor supertexta* in Taiwan. *Dis Aquat Org*, Vol. 65, No. 1, pp. 23-27
- Chang, P.H., Lee, S.H., Chiang, H.C. & Jong, M.H. (1999). Epizootic of herpes-like virus infection in Goldfish, *Carassius auratus* in Taiwan. *Fish Pathol*, Vol. 34, No. 4, pp. 209-210
- Chang, P.H., Pan, Y.H., Wu, C.M., Kuo, S.T. & Chung, H.Y. (2002). Isolation and molecular characterization of herpesvirus from cultured European eels *Anguilla anguilla* in Taiwan. *Dis Aquat Org*, Vol. 50, No. 2, pp. 111-118
- Costes, B., Fournier, G., Michel, B., Delforge, C., Raj, V.S., Dewals, B., Gillet, L., Drion, P., Body, A., Schynts, F., Liefbrig, F. & Vanderplasschen, A. (2008). Cloning of the koi herpesvirus genome as an infectious bacterial artificial chromosome demonstrates that disruption of the thymidine kinase locus induces partial attenuation in *Cyprinus carpio koi*. *J Virol*, Vol. 82, No. 10, pp. 4955-4964
- Crockford, M., Jones, J.B., Crane, M.S. & Wilcox, G.E. (2005). Molecular detection of a virus, Pilchard herpesvirus, associated with epizootics in Australasian pilchards *Sardinops sagax neopilchardus*. *Dis Aquat Org*, Vol. 68, No. 1, pp. 1-5
- Crockford, M., Jones, J.B., McColl, K. & Whittington, R.J. (2008). Comparison of three molecular methods for the detection of pilchard herpesvirus in archived paraffin-embedded tissue and frozen tissue. *Dis Aquat Org*, Vol. 82, pp. 37-44
- Davidse, A., Haenen, O.L.M., Dijkstra, S.G., van Nieuwstadt, A.P., van der Vorst, T.J.K., Wagenaar, F. & Wellenberg, G.J. (1999). First isolation of herpesvirus of eel (*Herpesvirus Anguillae*) in diseased European eel (*Anguilla anguilla* L.) in Europe. *Bull Eur Ass Fish Pathol*, Vol. 19, No. 4, pp. 137-141
- Davison, A.J. (1992). Channel catfish virus: a new type of herpesvirus. *Virology*, Vol. 186, No. 1, pp. 9-14
- Davison, A.J. (1998). The genome of salmonid herpesvirus 1. *J Virol*, Vol. 72, No. 3, pp. 1974-1982
- Davison, A.J. (2002). Evolution of the herpesviruses. *Vet Microbiol*, Vol. 86, No. 1-2, pp. 69-88
- Davison, A.J. (2007). Comparative analysis of the genomes, In: *Human Herpesviruses: Biology, Therapy, and Immunoprophylaxis* A. Arvin, G. Campadelli-Fiume, E. Mocarski, P.S.

- Moore, B. Roizman, R. Whitley & K. Yamanishi, Cambridge University Press, Cambridge
- Davison, A.J., Cunningham, C., Sauerbier, W. & McKinnell, R.G. (2006). Genome sequences of two frog herpesviruses. *J Gen Virol*, Vol. 87, No. 12, pp. 3509-3514
- Davison, A.J. & Davison, M.D. (1995). Identification of structural proteins of channel catfish virus by mass spectrometry. *Virology*, Vol. 206, No. 2, pp. 1035-1043
- Davison, A.J., Eberle, R., Ehlers, B., Hayward, G.S., McGeoch, D.J., Minson, A.C., Pellett, P.E., Roizman, B., Studdert, M.J. & Thiry, E. (2009). The order Herpesvirales. *Arch Virol*, Vol. 154, No. 1, pp. 171-177
- Davison, A.J., Eberle, R., Hayward, G.S., McGeoch, D.J., Minson, A.C., Pellet, P.E., Roizman, B., Studdert, M.J. & Thiry, E. (2005a). *Herpesviridae*, In: *Virus Taxonomy: VIIIth Report of the International Committee on Taxonomy of Viruses*, C.M. Fauquet, M.A. Mayo, J. Maniloff, U. Desselberger & L.A. Ball, pp. 193-212, Elsevier Academic Press, Amsterdam
- Davison, A.J., Sauerbier, W., Dolan, A., Addison, C. & McKinnell, R.G. (1999). Genomic studies of the Lucke tumor herpesvirus (RaHV-1). *J Cancer Res Clin Oncol*, Vol. 125, No. 3-4, pp. 232-238
- Davison, A.J., Trus, B.L., Cheng, N., Steven, A.C., Watson, M.S., Cunningham, C., Le Deuff, R.M. & Renault, T. (2005b). A novel class of herpesvirus with bivalve hosts. *J Gen Virol*, Vol. 86, No. 1, pp. 41-53
- Dishon, A., Davidovich, M., Ilouze, M. & Kotler, M. (2007). Persistence of cyprinid herpesvirus 3 in infected cultured carp cells. *J Virol*, Vol. 81, No. 9, pp. 4828-4836
- Dixon, P. (1997). Immunization with viral antigens: viral diseases of carp and catfish. *Dev Biol Stand*, Vol. 90, pp. 221-232
- Dixon, R.A. & Farber, F.E. (1980). Channel catfish virus: physicochemical properties of the viral genome and identification of viral polypeptides. *Virology*, Vol. 103, No. 2, pp. 267-278
- Doszpoly, A., Benkő, M., Bovo, G., LaPatra, S.E. & Harrach, B. (2011a). Comparative analysis of a conserved gene block from the genome of the members of the genus Ictalurivirus. *Intervirology*, pp. 1-8
- Doszpoly, A., Benkő, M., Csaba, G., Dán, Á., Lang, M. & Harrach, B. (2011b). Introduction of the family Alloherpesviridae: The first molecular detection of herpesviruses of cyprinid fish in Hungary. *Magyar Allatorvosok Lapja*, Vol. 133, No. 3, pp. 174-181
- Doszpoly, A., Kovacs, E.R., Bovo, G., LaPatra, S.E., Harrach, B. & Benko, M. (2008). Molecular confirmation of a new herpesvirus from catfish (*Ameiurus melas*) by testing the performance of a novel PCR method, designed to target the DNA polymerase gene of alloherpesviruses. *Arch Virol*, Vol. 153, No. 11, pp. 2123-2127
- Doszpoly, A. & Shchelkunov, I.S. (2010). Partial genome analysis of Siberian sturgeon alloherpesvirus suggests its close relation to AcHV-2. *Acta Vet Hung*, Vol. 58, No. 2, pp. 269-274
- Eide, K., Miller-Morgan, T., Heidel, J., Bildfell, R. & Jin, L. (2011a). Results of total DNA measurement in koi tissue by Koi Herpes Virus real-time PCR. *J Virol Methods*, Vol. 172, No. 1-2, pp. 81-84
- Eide, K.E., Miller-Morgan, T., Heidel, J.R., Kent, M.L., Bildfell, R.J., Lapatra, S., Watson, G. & Jin, L. (2011b). Investigation of koi herpesvirus latency in koi. *J Virol*, Vol. 85, No. 10, pp. 4954-4962

- Farley, C.A., Banfield, W.G., Kasnic, G.J. & Foster, W.S. (1972). Oyster herpes-type virus. *Science*, Vol. 178, pp. 759-760
- Fawcett, D.W. (1956). Electron microscope observations on intracellular virus-like particles associated with the cells of the Lucke renal adenocarcinoma. *J Biophys Biochem Cytol*, Vol. 2, No. 6, pp. 725-741
- Fijan, N., Petrinc, Z., Sulimanovic, D. & Zwillenberg, L.O. (1971). Isolation of the viral causative agent from the acute form of infectious dropsy of carp. *Veternarski arhiv*, Vol. 41, pp. 125-138
- Fuchs, W., Fichtner, D., Bergmann, S.M. & Mettenleiter, T.C. (2011). Generation and characterization of koi herpesvirus recombinants lacking viral enzymes of nucleotide metabolism. *Arch Virol*, Vol. 156, No. 6, pp. 1059-1063
- Garcia, C., Thebault, A., Degremont, L., Arzul, I., Miossec, L., Robert, M., Chollet, B., Francois, C., Joly, J.P., Ferrand, S., Kerdudou, N. & Renault, T. (2011). Ostreid herpesvirus 1 detection and relationship with *Crassostrea gigas* spat mortality in France between 1998 and 2006. *Vet Res*, Vol. 42: 73
- Gilad, O., Yun, S., Andree, K.B., Adkison, M.A., Zlotkin, A., Bercovier, H., Eldar, A. & Hedrick, R.P. (2002). Initial characteristics of koi herpesvirus and development of a polymerase chain reaction assay to detect the virus in koi, *Cyprinus carpio koi*. *Dis Aquat Org*, Vol. 48, No. 2, pp. 101-108
- Goodwin, A.E., Khoo, L., LaPatra, S.E., Bonar, A., Key, D.W., Garner, M., Lee, M.V. & Hanson, L. (2006). Goldfish Hematopoietic Necrosis Herpesvirus (Cyprinid Herpesvirus 2) in the USA: Molecular confirmation of isolates from diseased fish. *J Aquat Anim Health*, Vol. 18, pp. 11-18
- Goodwin, A.E., Sadler, J., Merry, G.E. & Marecaux, E.N. (2009). Herpesviral haematopoietic necrosis virus (CyHV-2) infection: case studies from commercial goldfish farms. *J Fish Dis*, Vol. 32, No. 3, pp. 271-278
- Graham, D.A., Curran, W.L., Geoghegan, F., McKiernan, F. & Foyle, K.L. (2004). First observation of herpes-like virus particles in northern pike, *Esox lucius* L., associated with bluespot-like disease in Ireland. *J Fish Dis*, Vol. 27, No. 9, pp. 543-549
- Granoff, A. (1999). Amphibian herpesviruses (*Herpesviridae*). In: *Encyclopedia of Virology*, A. Granoff & R. Webster, pp. 51-53, Academic Press, London
- Gravell, M. (1971). Viruses and renal carcinoma of *Rana pipiens*. X. Comparison of herpes-type viruses associated with Lucke tumor-bearing frogs. *Virology*, Vol. 43, No. 3, pp. 730-733
- Gravell, M., Granoff, A. & Darlington, R.W. (1968). Viruses and renal carcinoma of *Rana pipiens*. VII. Propagation of a herpes-type frog virus. *Virology*, Vol. 36, No. 3, pp. 467-475
- Gray, W.L., Williams, R.J., Jordan, R.L. & Griffin, B.R. (1999). Detection of channel catfish virus DNA in latently infected catfish. *J Gen Virol*, Vol. 80, No. 7, pp. 1817-1822
- Groff, J.M., LaPatra, S.E., Munn, R.J. & Zinkl, J.G. (1998). A viral epizootic in cultured populations of juvenile goldfish due to a putative herpesvirus etiology. *J Vet Diagn Invest*, Vol. 10, No. 4, pp. 375-378
- Haenen, O.L.M., Dijkstra, S.G., Tulden, P.W., Davidse, A., van Nieuwstadt, A.P., Wagenaar, F. & Wellenberg, G.J. (2002). Herpesvirus anguillae (HVA) isolations from disease outbreaks in cultured European eel, *Anguilla anguilla* in the Netherlands since 1996. *Bull Eur Ass Fish Pathol*, Vol. 22, No. 4, pp. 247-257

- Haenen, O.L.M., Lehmann, J., Engelsma, M.Y., Stürenberg, F.-J., Roozenburg, I., Kerkhoff, S. & Klein Breteler, J. (2010). The health status of European silver eels, *Anguilla anguilla*, in the Dutch River Rhine Watershed and Lake IJsselmeer. *Aquaculture*, Vol. 309, No. 1-4, pp. 15-24
- Haenen, O.L.M., Way, K., Bergmann, S.M. & Ariel, E. (2004). The emergence of Koi herpesvirus and its significance to European aquaculture. *Bull Eur Ass Fish Pathol*, Vol. 24, No. 6, pp. 293-307
- Hayashi, Y., Kodoma, H., Ishigaki, K., Mikami, T., Izawa, H. & Sakai, D.K. (1986). Characteristics of a new herpesviral isolate from salmonid fish. *Jpn J Vet Sci*, Vol. 48, No. 5, pp. 915-924
- Hayashi, Y., Kodoma, H., Mikami, T. & Izawa, H. (1989). Serological and genetic relationships of three herpesvirus strains from salmonid fish. *Arch Virol*, Vol. 104, pp. 163-168
- Hedrick, R.P., Gilad, O., Yun, S., Spangenberg, J.V., Marty, G.D., Nordhausen, R.W., Kebus, M.J., Bercovier, H. & Eldar, A. (2000). A herpesvirus associated with mass mortality of juvenile and adult koi, a strain of common carp. *J Aquat Anim Health*, Vol. 12, No. 1, pp. 44-57
- Hedrick, R.P., McDowell, T., Eaton, W.D., Kimura, T. & Sano, T. (1987). Serological relationships of five herpesviruses isolated from salmonid fishes. *J Appl Ichthyol*, Vol. 3, pp. 87-92
- Hedrick, R.P., McDowell, T.S., Groff, J.M., Yun, S. & Wingfield, W.H. (1991). Isolation of an epitheliotropic herpesvirus from white sturgeon *Acipenser transmontanus*. *Dis Aquat Org*, Vol. 11, pp. 49-56
- Hedrick, R.P., McDowell, T.S., Gilad, O., Adkison, M. & Bovo, G. (2003). Systemic herpes-like virus in catfish *Ictalurus melas* (Italy) differs from Ictalurid herpesvirus 1 (North America). *Dis Aquat Org*, Vol. 55, No. 2, pp. 85-92
- Herman, R.L., Burke, C.N. & Perry, S. (1997). Epidermal tumors of rainbow smelt with associated virus. *J Wildl Dis*, Vol. 33, No. 4, pp. 925-929
- Huang, S. & Hanson, L.A. (1998). Temporal gene regulation of the channel catfish virus (Ictalurid herpesvirus 1). *J Virol*, Vol. 72, No. 3, pp. 1910-1917
- Hyatt, A.D., Hine, P.M., Whittington, R.J., Kearns, C., Wise, T.G., Crane, M.S. & Williams, L.M. (1997). Epizootic mortality in the pilchard *Sardinops sagax neopilchardus* in Australia and New Zealand in 1995. II. Identification of a herpesvirus within the gill epithelium. *Dis Aquat Org*, Vol. 28, pp. 17-29
- Iida, Y., Masumura, K., Nakai, T., Sorimachi, M. & Matsuda, H. (1989). A viral disease in larvae and juveniles of the Japanese flounder *Paralichthys olivaceus*. *J Aquat Anim Health*, Vol. 1, pp. 7-12
- Iida, Y., Nakai, T., Sorimachi, M. & Masumura, K. (1991). Histopathology of a herpesvirus infection in larvae of Japanese flounder *Paralichthys olivaceus*. *Dis Aquat Org*, Vol. 10, pp. 59-63
- Jakob, N.J., Kehm, R. & Gelderblom, H.R. (2010). A novel fish herpesvirus of *Osmerus eperlanus*. *Virus genes*, Vol. 41, No. 1, pp. 81-85
- Jeffery, K.R., Bateman, K., Bayley, A., Feist, S.W., Hulland, J., Longshaw, C., Stone, D., Woolford, G. & Way, K. (2007). Isolation of a cyprinid herpesvirus 2 from goldfish, *Carassius auratus* (L.), in the UK. *J Fish Dis*, Vol. 30, No. 11, pp. 649-656

- Jørgensen, P., Castric, J., Hill, B., Ljungberg, O. & De Kinkelin, P. (1994). The occurrence of virus infections in elvers and eels (*Anguilla anguilla*) in Europe with particular reference to VHSV and IHNV. *Aquaculture*, Vol. 123, No. 1-2, pp. 11-19
- Jung, S.J. & Miyazaki, T. (1995). Herpesviral haematopoietic necrosis of goldfish, *Carassius auratus* (L.). *J Fish Dis*, Vol. 18, pp. 211-220
- Kelley, G.O., Waltzek, T.B., McDowell, T.S., Yun, S.C., LaPatra, S.E. & Hedrick, R.P. (2005). Genetic Relationships among Herpes-Like Viruses Isolated from Sturgeon. *J Aquat Anim Health*, Vol. 17, No. 4, pp. 297-303
- Kelly, R.K., Nielsen, O., Mitchell, S.C. & Yamamoto, T. (1983). Characterization of Herpesvirus vitreum isolated from hyperplastic epidermal tissue of walleye, *Stizostedion vitreum vitreum* (Mitchill). *J Fish Dis*, Vol. 6, pp. 249-260
- Kent, M.L. & Myers, M.S. (2000). Hepatic lesions in a redstriper rockfish (*Sebastes proriger*) suggestive of a herpesvirus infection. *Dis Aquat Org*, Vol. 41, pp. 237-239
- Kimura, T., Yoshimizu, M. & Tanaka, M. (1981a). Studies on a new virus (OMV) from *Oncorhynchus masou* - II. Oncogenic nature. *Fish Pathol*, Vol. 15, No. 3/4, pp. 149-153
- Kimura, T., Yoshimizu, M. & Tanaka, M. (1983). Susceptibility of different fry stages of representative salmonid species to oncorhynchus masou virus (OMV). *Fish Pathol*, Vol. 17, No. 4, pp. 251-258
- Kimura, T., Yoshimizu, M., Tanaka, M. & Sannohe, H. (1981b). Studies on a new virus (OMV) from *Oncorhynchus masou* - I. Characteristics and pathogenicity. *Fish Pathol*, Vol. 15, No. 3/4, pp. 143-147
- Kucuktas, H. & Brady, Y.J. (1999). Molecular biology of channel catfish virus. *Aquaculture*, Vol. 172, No. 1-2, pp. 147-161
- Kumagai, A., Takahashi, K. & Fukuda, H. (1994). Epizootics caused by salmonid herpesvirus type 2 infection in maricultured coho salmon. *Fish Pathol*, Vol. 29, No. 2, pp. 127-134
- Kunec, D., Hanson, L.A., van Haren, S., Nieuwenhuizen, I.F. & Burgess, S.C. (2008). An overlapping bacterial artificial chromosome system that generates vectorless progeny for channel catfish herpesvirus. *J Virol*, Vol. 82, No. 8, pp. 3872-3881
- Kunec, D., Nanduri, B. & Burgess, S.C. (2009). Experimental annotation of channel catfish virus by probabilistic proteogenomic mapping. *Proteomics*, Vol. 9, No. 10, pp. 2634-2647
- Kurobe, T., Kelley, G.O., Waltzek, T.B. & Hedrick, R.P. (2008). Revised Phylogenetic Relationships among Herpesviruses Isolated from Sturgeons. *J Aquat Anim Health*, Vol. 20, No. 2, pp. 96-102
- Kurobe, T., Marcquenski, S. & Hedrick, R.P. (2009). PCR assay for improved diagnostics of epitheliotropic disease virus (EEDV) in lake trout *Salvelinus namaycush*. *Dis Aquat Org*, Vol. 84, No. 1, pp. 17-24
- Lee, N.-S., Kobayashi, J. & Miyazaki, T. (1999). Gill filament necrosis in farmed Japanese eels, *Anguilla japonica* (Temminck & Schlegel), infected with *Herpesvirus anguillae*. *J Fish Dis*, Vol. 22, No. 6, pp. 457-463
- Leibovitz, L. & Leboutz, S.S. (1985). A viral dermatitis of the smooth dogfish, *Mustelus canis* (Mitchill). *J Fish Dis*, Vol. 8, pp. 273-279
- Loret, S., Guay, G. & Lippe, R. (2008). Comprehensive characterization of extracellular herpes simplex virus type 1 virions. *J Virol*, Vol. 82, No. 17, pp. 8605-8618
- Lucké, B. (1934). A neoplastic disease of the kidney of the frog, *Rana pipiens*. *Am J Cancer*, Vol. 20, pp. 352-379

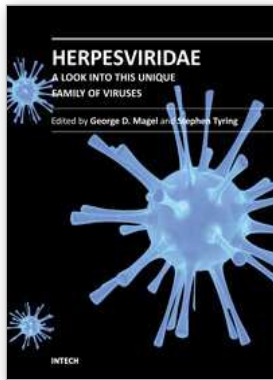
- Lucké, B. (1938). Carcinoma in the Leopard Frog: Its Probable Causation by a Virus. *J Exp Med*, Vol. 68, No. 4, pp. 457-468
- Lucké, B. & Schlumberger, H. (1949). Induction of metastasis of frog carcinoma by increase of environmental temperature. *J Exp Med*, Vol. 89, No. 3, pp. 269-278
- Lunger, P.D. (1964). The Isolation and Morphology of the Luck'e Frog Kidney Tumor Virus. *Virology*, Vol. 24, pp. 138-145
- Margenau, T.L., Marcquenski, S.V., Rasmussen, P.W. & MacConnell, E. (1995). Prevalence of Blue spot disease (Esocid herpesvirus-1) on northern pike and muskellunge in Wisconsin. *J Aquat Anim Health*, Vol. 7, pp. 29-33
- Masumura, K., Iida, Y., Nakai, T. & Mekuchi, T. (1989). The effects of water temperature and fish age on a herpesvirus infection of Japanese flounder larvae, *Paralichthys olivaceus*. *Fish Pathol*, Vol. 24, No. 2, pp. 111-114
- McAllister, P.E. & Herman, R.L. (1989). Epizootic mortality in hatchery-reared lake trout *Salvelinus namaycush* caused by a putative virus possibly of the herpesvirus group. *Dis Aquat Org*, Vol. 6, pp. 113-119
- McAllister, P.E., Lidgerding, B.C., Herman, R.L., Hoyer, L.C. & Hankins, J. (1985). Viral diseases of fish: first report of carp pox in golden ide (*Leuciscus idus*) in North America. *J Wildl Dis*, Vol. 21, No. 3, pp. 199-204
- McGeoch, D.J. & Davison, A. (1999). The molecular evolutionary history of the herpesviruses, In: *Origin and Evolution of Viruses*, E. Domingo, R. Webster & J. Holland, pp. 441-465, Academic Press, London
- McGeoch, D.J., Rixon, F.J. & Davison, A.J. (2006). Topics in herpesvirus genomics and evolution. *Virus Res*, Vol. 117, No. 1, pp. 90-104
- McKinnell, R.G. & Carlson, D.L. (1997). Lucke renal adenocarcinoma, an anuran neoplasm: studies at the interface of pathology, virology, and differentiation competence. *J Cell Physiol*, Vol. 173, No. 2, pp. 115-118
- McKinnell, R.G. & Tarin, D. (1984). Temperature-dependent metastasis of the Lucke renal carcinoma and its significance for studies on mechanisms of metastasis. *Cancer Metastasis Rev*, Vol. 3, No. 4, pp. 373-386
- Mellergaard, S. & Bloch, B. (1988). Herpesvirus-like particles in angelfish *Pterophyllum altum*. *Dis Aquat Org*, Vol. 5, pp. 151-155
- Michel, B., Fournier, G., Loeffrig, F., Costes, B. & Vanderplasschen, A. (2010). Cyprinid herpesvirus 3. *Emerg Infect Dis*, Vol. 16, No. 12, pp. 1835-1843
- Michel, B., Leroy, B., Stalin Raj, V., Loeffrig, F., Mast, J., Wattiez, R., Vanderplasschen, A. & Costes, B. (2010). The genome of cyprinid herpesvirus 3 encodes 40 proteins incorporated in mature virions. *J Gen Virol*, Vol. 91, No. 2, pp. 452-462
- Miyazaki, K., Fujiwara, K., Kobara, J., Matsumoto, N., Abe, M. & Nagano, T. (1989). Histopathology associated with two viral diseases of larval and juvenile fishes: epidermal necrosis of the Japanese flounder *Paralichthys olivaceus* and epithelial necrosis of black sea bream *Acanthopagrus schlegeli*. *J Aquat Anim Health*, Vol. 1, No. 2, pp. 85-93
- Miyazaki, T. (2005). Ultrastructural features of herpesvirus-infected cells in epidermal lesions in larvae of the Japanese flounder *Paralichthys olivaceus*. *Dis Aquat Org*, Vol. 66, No. 2, pp. 159-162

- Mizell, M., Toplin, I. & Isaacs, J.J. (1969). Tumor induction in developing frog kidneys by a zonal centrifuge purified fraction of the frog herpes-type virus. *Science*, Vol. 165, No. 898, pp. 1134-1137
- Morrison, C.M., Leggiardro, C.T. & Martell, D.J. (1996). Visualization of viruses in tumors of rainbow smelt *Osmerus mordax*. *Dis Aquat Org*, Vol. 26, pp. 19-23
- Murray, A.G., O'Callaghan, M. & Jones, B. (2003). A model of spatially evolving herpesvirus epidemics causing mass mortality in Australian pilchard *Sardinops sagax*. *Dis Aquat Org*, Vol. 54, pp. 1-14
- Naegele, R.F., Granoff, A. & Darlington, R.W. (1974). The presence of the Lucke herpesvirus genome in induced tadpole tumors and its oncogenicity: Koch-Henle postulates fulfilled. *Proc Natl Acad Sci U S A*, Vol. 71, No. 3, pp. 830-834
- Nusbaum, K.E., Smith, B.F., DeInnocentes, P. & Bird, R.C. (2002). Protective immunity induced by DNA vaccination of channel catfish with early and late transcripts of the channel catfish herpesvirus (IHV-1). *Vet Immunol Immunopathol*, Vol. 84, No. 3-4, pp. 151-168
- O'Neill, G. (1995). Ocean anomaly triggers record fish kill. *Science*, Vol. 268, No. 5216, p. 1431
- Rafferty, K.A., Jr. (1965). The cultivation of inclusion-associated viruses from Lucke tumor frogs. *Ann N Y Acad Sci*, Vol. 126, No. 1, pp. 3-21
- Rijsewijk, F., Pritz-Verschuren, S., Kerkhoff, S., Botter, A., Willemsen, M., van Nieuwstadt, T. & Haenen, O. (2005). Development of a polymerase chain reaction for the detection of Anguillid herpesvirus DNA in eels based on the herpesvirus DNA polymerase gene. *J Virol Methods*, Vol. 124, No. 1-2, pp. 87-94
- Ronen, A., Perelberg, A., Abramowitz, J., Hutoran, M., Tinman, S., Bejerano, I., Steinitz, M. & Kotler, M. (2003). Efficient vaccine against the virus causing a lethal disease in cultured *Cyprinus carpio*. *Vaccine*, Vol. 21, No. 32, pp. 4677-4684
- Rosenkranz, D., Klupp, B.G., Teifke, J.P., Granzow, H., Fichtner, D., Mettenleiter, T.C. & Fuchs, W. (2008). Identification of envelope protein pORF81 of koi herpesvirus. *J Gen Virol*, Vol. 89, No. 4, pp. 896-900
- Sano, M., Fukuda, H. & Sano, T. (1990a). Isolation and characterization of a new herpesvirus from eel. In: *Pathology in Marine Sciences*, F.O. Perkins & T.C. Cheng, pp. 15-31, Academic Press, San Diego
- Sano, N., Moriwake, M. & Sano, T. (1993). *Herpesvirus cyprini*: thermal effects on pathogenicity and oncogenicity. *Fish Pathol*, Vol. 28, No. 4, pp. 171-175
- Sano, T. (1976). Viral diseases of cultured fishes in Japan. *Fish Pathol*, Vol. 10, pp. 221-226
- Sano, T., Fukuda, H. & Furukawa, M. (1985a). Herpesvirus cyprini: biological and oncogenic properties. *Fish Pathol*, Vol. 20, No. 4, pp. 381-388
- Sano, T., Fukuda, H., Furukawa, M., Hosoya, H. & Moriya, Y. (1985b). A herpesvirus isolated from carp papilloma in Japan. In: *Fish and Shellfish Pathology*, A.E. Ellis, pp. 307-311, Academic Press, London
- Sano, T., Fukuda, H., Okamoto, N. & Kaneko, F. (1983). Yamame tumor virus: lethality and oncogenicity. *Bull Jpn Soc Sci Fish*, Vol. 49, No. 8, pp. 1159-1163
- Sano, T., Morita, N., Shima, N. & Akimoto, M. (1990b). A preliminary report on pathogenicity and oncogenicity of cyprinid herpesvirus. *Bull Eur Ass Fish Pathol*, Vol. 10, No. 1, pp. 11-13
- Sano, T., Morita, N., Shima, N. & Akimoto, M. (1991). *Herpesvirus cyprini*: lethality and oncogenicity. *J Fish Dis*, Vol. 14, pp. 533-543

- Savin, K.W., Cocks, B.G., Wong, F., Sawbridge, T., Cogan, N., Savage, D. & Warner, S. (2010). A neurotropic herpesvirus infecting the gastropod, abalone, shares ancestry with oyster herpesvirus and a herpesvirus associated with the amphioxus genome. *Virology*, Vol. 7: 308
- Schubert, G.H. (1966). The infective agent in carp pox. *Bull Off Int Epiz*, Vol. 65, No. 7-8, pp. 1011-1022
- Segarra, A., Pepin, J.F., Arzul, I., Morga, B., Faury, N. & Renault, T. (2010). Detection and description of a particular Ostreid herpesvirus 1 genotype associated with massive mortality outbreaks of Pacific oysters, *Crassostrea gigas*, in France in 2008. *Virus Res*, Vol. 153, No. 1, pp. 92-99
- Shchelkunov, I.S., Karaseva, T.A. & Kadoshnikov, Y.U.P. (1992). Atlantic salmon papillomatosis: visualization of herpesvirus-like particles in skin growths of affected fish. *Bull Eur Ass Fish Pathol*, Vol. 12, No. 1, pp. 28-31
- Shchelkunov, I.S., Shchelkunova, T.I., Shchelkunov, A.I., Kolbassova, Y.P., Didenko, L.V. & Bykovsky, A.P. (2009). First detection of a viral agent causing disease in farmed sturgeon in Russia. *Dis Aquat Org*, Vol. 86, No. 3, pp. 193-203
- Shlapobersky, M., Sinyakov, M.S., Katzenellenbogen, M., Sarid, R., Don, J. & Avtalion, R.R. (2010). Viral encephalitis of tilapia larvae: Primary characterization of a novel herpes-like virus. *Virology*, Vol. 399, No. 2, pp. 239-247
- Silverstein, P.S., Bird, R.C., van Santen, V.L. & Nusbaum, K.E. (1995). Immediate-early transcription from the channel catfish virus genome: characterization of two immediate-early transcripts. *J Virol*, Vol. 69, No. 5, pp. 3161-3166
- Silverstein, P.S., van Santen, V.L., Nusbaum, K.E. & Bird, R.C. (1998). Expression kinetics and mapping of the thymidine kinase transcript and an immediate-early transcript from channel catfish virus. *J Virol*, Vol. 72, No. 5, pp. 3900-3906
- St-Hilaire, S., Beevers, N., Way, K., Le Deuff, R.M., Martin, P. & Joiner, C. (2005). Reactivation of koi herpesvirus infections in common carp *Cyprinus carpio*. *Dis Aquat Org*, Vol. 67, No. 1-2, pp. 15-23
- Stephens, F.J., Roldal, S.R. & Jones, B. (2004). Haematopoietic necrosis in a goldfish (*Carassius auratus*) associated with an agent morphologically similar to herpesvirus. *Aust Vet J*, Vol. 82, No. 3, pp. 167-169
- Stingley, R.L. & Gray, W.L. (2000). Transcriptional regulation of the channel catfish virus genome direct repeat region. *J Gen Virol*, Vol. 81, No. 8, pp. 2005-2010
- Stingley, R.L., Griffin, B.R. & Gray, W.L. (2003). Channel catfish virus gene expression in experimentally infected channel catfish, *Ictalurus punctatus* (Rafinesque). *J Fish Dis*, Vol. 26, No. 8, pp. 487-493
- Tan, J., Lancaster, M., Hyatt, A., van Driel, R., Wong, F. & Warner, S. (2008). Purification of a herpes-like virus from abalone (*Haliotis* spp.) with ganglioneuritis and detection by transmission electron microscopy. *J Virol Methods*, Vol. 149, No. 2, pp. 338-341
- Tanaka, M., Yoshimizu, M. & Kimura, T. (1984). *Oncorhynchus masou* virus: Pathological changes in masu salmon (*Oncorhynchus masou*), chum salmon (*O. keta*), coho salmon (*O. kisutch*) fry infected with OMV by immersion method. *Bull Jpn Soc Sci Fish*, Vol. 50, No. 3, pp. 431-437
- Tham, K.M. & Moon, C.D. (1996). Polymerase chain reaction amplification of the thymidine kinase and protein kinase-related genes of channel catfish virus and a putative pilchard herpesvirus. *J Virol Methods*, Vol. 61, No. 1-2, pp. 65-72

- Ueno, Y., Kitao, T., Chen, S.-N., Aoki, T. & Kou, G.-H. (1992). Characterization of A Herpes-like Virus Isolated from Cultured Japanese Eels in Taiwan. *Fish Pathol*, Vol. 27, No. 1, pp. 7-17
- Ueno, Y., Shi, J.-W., Yoshida, T., Kitao, T., Sakai, M., Chen, S.-N. & Kou, G.H. (1996). Biological and serological comparisons of eel herpesvirus in Formosa (EHVF) and Herpesvirus anguillae (HVA). *J Appl Ichthyol*, Vol. 12, No. 1, pp. 49-51
- van Beurden, S.J., Bossers, A., Voorbergen-Laarman, M.H., Haenen, O.L.M., Peters, S., Abma-Henkens, M.H., Peeters, B.P., Rottier, P.J. & Engelsma, M.Y. (2010). Complete genome sequence and taxonomic position of anguillid herpesvirus 1. *J Gen Virol*, Vol. 91, No. 4, pp. 880-887
- van Beurden, S.J., Leroy, B., Wattiez, R., Haenen, O.L.M., Boeren, S., Vervoort, J.J.M., Peeters, B.P.H., Rottier, P.J.M., Engelsma, M.Y. & Vanderplasschen, A.F. (2011). Identification and localization of the structural proteins of anguillid herpesvirus 1. *Vet Res*, Vol. 42: 105
- van Ginneken, V., Haenen, O., Coldenhoff, K., Willemze, R., Antonissen, E., van Tulden, P., Dijkstra, S., Wagenaar, F. & van den Thillart, G. (2004). Presence of eel viruses in eel species from various geographic regions. *Bull Eur Ass Fish Pathol*, Vol. 24, No. 5, pp. 268-272
- van Nieuwstadt, A.P., Dijkstra, S.G. & Haenen, O.L. (2001). Persistence of herpesvirus of eel Herpesvirus anguillae in farmed European eel *Anguilla anguilla*. *Dis Aquat Org*, Vol. 45, No. 2, pp. 103-107
- Vanderheijden, N., Hanson, L.A., Thiry, E. & Martial, J.A. (1999). Channel catfish virus gene 50 encodes a secreted, mucin-like glycoprotein. *Virology*, Vol. 257, No. 1, pp. 220-227
- Walster, C.I. (1999). Clinical observations of severe mortalities in Koi carp, *Cyprinus carpio*, with gill disease. *Fish Vet Journal*, Vol. 3, pp. 54-58
- Waltzek, T.B., Kelley, G.O., Alfaro, M.E., Kurobe, T., Davison, A.J. & Hedrick, R.P. (2009). Phylogenetic relationships in the family *Alloherpesviridae*. *Dis Aquat Org*, Vol. 84, pp. 179-194
- Waltzek, T.B., Kelley, G.O., Stone, D.M., Way, K., Hanson, L., Fukuda, H., Hirono, I., Aoki, T., Davison, A.J. & Hedrick, R.P. (2005). Koi herpesvirus represents a third cyprinid herpesvirus (CyHV-3) in the family Herpesviridae. *J Gen Virol*, Vol. 86, No. 6, pp. 1659-1667
- Watson, L.R., Yun, S.C., Groff, J.M. & Hedrick, R.P. (1995). Characteristics and pathogenicity of a novel herpesvirus isolated from adult and subadult white sturgeon *Acipenser transmontanus*. *Dis Aquat Org*, Vol. 22, pp. 199-210
- Whittington, R.J., Crockford, M., Jordan, D. & Jones, B. (2008). Herpesvirus that caused epizootic mortality in 1995 and 1998 in pilchard, *Sardinops sagax neopilchardus* (Steindachner), in Australia is now endemic. *J Fish Dis*, Vol. 31, No. 2, pp. 97-105
- Whittington, R.J., Jones, J.B., Hine, P.M. & Hyatt, A.D. (1997). Epizootic mortality in the pilchard *Sardinops sagax neopilchardus* in Australia and New Zealand in 1995. I. Pathology and epizootiology. *Dis Aquat Org*, Vol. 28, pp. 1-16
- Wolf, K. (1976). Fish viral diseases in North America, 1971-75, and recent research of the Eastern fish disease laboratory, U.S.A. *Fish Pathol*, Vol. 10, No. 2, pp. 135-154
- Wolf, K. (1988). Channel catfish virus disease, In: *Fish viruses and fish viral diseases*, K. Wolf, pp. 21-42, Cornell University Press, Ithaca, New York

- Wolf, K. & Darlington, R.W. (1971). Channel catfish virus: a new herpesvirus of ictalurid fish. *J Virol*, Vol. 8, No. 4, pp. 525-533
- Wolf, K., Darlington, R.W., Taylor, W.G., Quimby, M.C. & Nagabayashi, T. (1978). Herpesvirus salmonis: Characterization of a New Pathogen of Rainbow Trout. *J Virol*, Vol. 27, No. 3, pp. 659-666
- Wolf, K. & Smith, C.E. (1981). *Herpesvirus salmonis*: pathological changes in parenterally-infected rainbow trout, *Salmo gairdneri* Richardson, fry. *J Fish Dis*, Vol. 4, pp. 445-457
- Yamamoto, T., Kelly, R.K. & Nielsen, O. (1984). Epidermal hyperplasias of northern pike (*Esox lucius*) associated with herpesvirus and C-type particles. *Arch Virol*, Vol. 79, No. 3-4, pp. 255-272
- Yoshimizu, M., Fukuda, H., Sano, T. & Kimura, T. (1995). Salmonid herpesvirus 2. Epizootiology and serological relationship. *Vet Res*, Vol. 26, No. 5-6, pp. 486-492
- Yoshimizu, M., Tanaka, M. & Kimura, H. (1987). *Oncorhynchus masou* virus (OMV): Incidence of tumor development among experimentally infected representative salmonid species. *Fish Pathol*, Vol. 22, No. 1, pp. 7-10
- Zhang, H.G. & Hanson, L.A. (1995). Deletion of thymidine kinase gene attenuates channel catfish herpesvirus while maintaining infectivity. *Virology*, Vol. 209, No. 2, pp. 658-663



Herpesviridae - A Look Into This Unique Family of Viruses

Edited by Dr. George Dimitri Magel

ISBN 978-953-51-0186-4

Hard cover, 320 pages

Publisher InTech

Published online 07, March, 2012

Published in print edition March, 2012

In order to fully understand the nature of viruses, it is important to look at them from both, their basic science and clinical, standpoints. Our goal with this book was to dissect Herpesviridae into its biological properties and clinical significance in order to provide a logical, as well as practical, approach to understanding and treating the various conditions caused by this unique family of viruses. In addition to their up-to-date and extensive text, each chapter is laced with a variety of diagrams, tables, charts, and images, aimed at helping us achieve our goal. We hope that this book will serve as a reference tool for clinicians of various specialties worldwide.

How to reference

In order to correctly reference this scholarly work, feel free to copy and paste the following:

Steven van Beurden and Marc Engelsma (2012). Herpesviruses of Fish, Amphibians and Invertebrates, Herpesviridae - A Look Into This Unique Family of Viruses, Dr. George Dimitri Magel (Ed.), ISBN: 978-953-51-0186-4, InTech, Available from: <http://www.intechopen.com/books/herpesviridae-a-look-into-this-unique-family-of-viruses/herpesviruses-of-fish-amphibians-and-invertebrates>

INTECH

open science | open minds

InTech Europe

University Campus STeP Ri
Slavka Krautzeka 83/A
51000 Rijeka, Croatia
Phone: +385 (51) 770 447
Fax: +385 (51) 686 166
www.intechopen.com

InTech China

Unit 405, Office Block, Hotel Equatorial Shanghai
No.65, Yan An Road (West), Shanghai, 200040, China
中国上海市延安西路65号上海国际贵都大饭店办公楼405单元
Phone: +86-21-62489820
Fax: +86-21-62489821