

**Pollutant-induced changes in  
terrestrial  
nematode communities**

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**POLLUTANT-INDUCED CHANGES IN  
TERRESTRIAL  
NEMATODE COMMUNITIES**

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**Proefschrift**

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## Stellingen

1. Een nematodengemeenschap is indicatief en onderscheidend ten aanzien van vele abiotische en biotische stressfactoren.  
Dit proefschrift.
2. Ecotoxicologisch-onderzoek in afwezigheid van planten is niet representatief voor de effecten van bodemverontreiniging *in situ*.  
Dit proefschrift.
3. Bij de inschaling van nematoden in *cp*-groepen is geen sprake van cirkelredeneringen.  
Van Straalen, N.M. 1997. In: Biological indicators of soil health and sustainable productivity (Ed. Pankhorst, C.E., Doube, B.M. and Gupta, V.V.S.R.), pp. 235-264. CAB international, Wallingford.
4. In onderzoek met betrekking tot de bodemkwaliteit wordt onvoldoende aandacht besteed aan de bodembioïologie.
5. Proefdieren in kunstgrond hebben een beperkte relevantie voor de ecotoxicologie.
6. De hypothese dat nematoden een bijdrage leveren aan vegetatieveranderingen in natuurlijke systemen, is in de landbouw reeds lang bekend.  
Putten et al. 1993. Nature 362:53-55.
7. Voor het schrijven van een artikel met meerdere auteurs geldt de economische wet van de afnemende meeropbrengst.
8. Naast een algemene index voor de economie, zoals de Dow-Jones-index, dient er een algemene milieu-index te komen.
9. Natuurontwikkeling op uit produktie genomen landbouwsystemen is wetenschappelijk gezien een braak liggend terrein.
10. 'Jonge' onderzoekers zijn de nieuwe nomaden. Zij kunnen beter een camper kopen en geen partner hebben.

Stellingen behorend bij het proefschrift, getiteld 'Pollutant-induced changes in terrestrial nematode communities' door Gerard Korthals.

Wageningen, 26 mei 1997.

## ABSTRACT

This thesis concerns metal-induced changes in terrestrial nematode communities exposed in microcosm-experiments and in a manipulative field experiment. Indigenous nematode communities, present in freshly collected agricultural soil, were exposed to heavy metals applied singly (Cd, Cu, Ni and Zn) or in combination under different test conditions. Depending on abiotic characteristics such as soil pH and biotic characteristics such as the presence of vegetation, the nematode community structure responded very sensitively to increasing metal concentrations.

In general, the effects of the investigated metals were enhanced with increasing exposure time and decreasing soil pH (investigated for Cu only). Furthermore, the presence or absence of vegetation (*Lolium perenne* L.) seems a very important factor in determining the final ecotoxicological effects of metals to nematodes. In soil covered with *L. perenne* the effects of Cu and Zn became apparent only at higher metal concentrations, were less severe and were more often caused in an indirect manner. In an acid sandy soil containing Cu and Zn, it was demonstrated that the dissolved Cu or Zn concentrations measured after equilibrating soil samples with a 0.01 M solution of  $\text{CaCl}_2$  were not significantly different from single metal additions and that the final effects to the nematode community were all additive or less than additive. Metal-induced reductions in the population size of nematode taxa showed a low intra-taxon variation for the different metals tested. However, there were major differences between the sensitivities of the taxa. For example, some omnivorous and predatory nematodes, known to be "K-strategists", were very sensitive and disappeared at Cu and Zn concentrations exceeding  $50 \text{ mg kg}^{-1}$ . Classifications based on the different life-history and feeding groups both facilitated the interpretation of pollution-induced changes in the nematode community, despite the fact that on a lower level these classifications could not adequately predict the sensitivity of all nematode taxa.

It is concluded that the nematode community structure and some community parameters, such as the Maturity Index, offer excellent perspectives to assess effects of pollutants at the community level. The nematode community can provide an early and sensitive signal of increased Cu, Ni or Zn pollution in the soil. Moreover, it was demonstrated that the nematode structure may also provide opportunities to identify specific types of disturbance, *in casu* pollutants.

**Keywords:** Nematode community structure, heavy metals, soil pollution, Maturity Index, bioavailability, risk assessment, life-history strategy

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# Chapter 1

## INTRODUCTION

The ultimate goal in ecotoxicology is to detect, monitor and predict the effects of pollutants on ecosystems (Moriarty, 1983). Since ecotoxicology originates from several disciplines, i.e. toxicology, environmental chemistry, biochemistry and ecology, there is a long tradition in discussing the contributions of each separate discipline (Koeman, 1983; Cairns, 1990; Forbes and Forbes, 1994). Although the number of multidisciplinary studies performed in recent years has greatly increased, several approaches remain for the investigation of the potential risks of pollutants on soil fauna. They can be divided into two extremes and are here referred to as the bottom up and top down approach.

In the much used bottom up approach, pollutant effects are investigated in systems with a low complexity, such as standardized single species tests. These tests are performed on species which meet certain criteria, i.e. ease of culturing, simple food requirements and a high rate of reproduction (Edwards, 1989). Besides the many advantages of these systems, such as the strong relationship between exposure and ecophysiological response of the tested species, there are several disadvantages. One disadvantage is the over-simplification of the actual conditions in the field, or as stated by Van Straalen (1994): 'the action of toxicants cannot be studied without considering all other environmental factors affecting the animal'. This seems to be true, not only for abiotic factors, of which most may be mimicked in test systems, but especially for biotic factors which are often more difficult to manipulate in test systems. To circumvent this over-simplification, ecotoxicologists have started to increase the ecological relevance of the test systems for example by studying effects of pollutants on sublethal parameters, in multi-species systems and under variable abiotic conditions. Nevertheless, it remains difficult to generalize results due to the limited number of species which can be investigated. This can result in a gap in our knowledge of the risks for some species, i.e. with a long iteroparous life history strategy (Laskowski *et al.*, 1996), those depending on sexual reproduction (Siepel, 1994) or those species which are difficult to culture.

At present, the top down approach is gaining more attention. In it, the effects of pollutants are investigated in the field where the highest complexity can be found. These studies realistically reflect the impact of pollutants, since among other things, indirect effects of pollutants and the responses of many taxa which cannot be cultured or kept under laboratory conditions are implicitly accounted for. Despite the realism of studying



polluted sites in the field (observational field studies), these studies often have severe drawbacks, such as inadequate reference sites, mixtures of different pollutants and a high natural variability (Edwards *et al.*, 1996), which may complicate the establishment of cause-and-effect relationships and the extrapolation of results to other polluted sites.

A meaningful approach between single-species laboratory tests (bottom up approach) and ecosystem studies (top down approach) seems to either study naturally occurring fauna communities which are intentionally exposed to a pollutant in field experiments or in microcosms or mesocosm tests (Cairns, 1985; Cairns, 1986; Sheppard, 1994). Especially microcosms using freshly collected field soil containing (components of) the indigenous soil organisms seems promising (Kappers and Van Esbroek, 1987; Kappers and Manger, 1990; Parmelee *et al.*, 1993). It is essential for each choice of approach that a critical selection of the organisms is used for micro- or mesocosm experiments (Edwards *et al.*, 1996). In recent years the suitability of nematodes or roundworms as test species in ecotoxicology has been strongly advocated (Vranken and Heip, 1986; Saimoilof, 1987; Freckman, 1988; Bongers, 1990; Bongers *et al.*, 1991; Vranken *et al.*, 1991; De Goede *et al.*, 1993; Yeates, 1994).

For the studies described in this thesis, nematodes were chosen as test species. The terrestrial nematode fauna is characterized by high densities and a species diversity, of which Andrassy (1991) estimated that already more than 11000 free-living species have been described. Nematodes are present in almost every habitat where they play a prominent role, for example in terrestrial food webs (Yeates, 1987; De Ruiter *et al.*, 1995). Nematodes are easy to sample and identify on the genus level and are representative of soil samples in which they are found as a consequence of their low mobility. Although the main interest in nematodes originates from the harmful effects some plant parasitic nematodes can exert on agricultural crops, there are many other nematode species contributing to soil fertility by influencing decomposition and mineralization (Tietjen, 1980; Ingham *et al.*, 1985; Yeates, 1987; Alkemade *et al.*, 1992). In order to evaluate the impact of pollution on the structure of nematode communities, there is often a need for community parameters which can facilitate the interpretation. Besides classical diversity indices such as the Shannon index, specific nematode community parameters have been developed. Some examples are the percentage of dorylaimids, the percentage of omnivorous nematodes and the proportion of "Dauer larvae" within the Rhabditida (Wasilewska, 1974; Sohlenius and Bostrom, 1984; Zullini and Peretti, 1986).

Many of the above mentioned parameters are related to the more classical approaches of classifying organisms on the basis of their life-history strategy, such as the *r-K* concept

(MacArthur and Wilson, 1967) or R-C-S concept (Grime, 1977; Greenslade, 1983). In this line Bongers (1990) proposed the Maturity Index (MI) and the Plant Parasite Index (PPI) for which nematode families were classified on the basis of their ability to colonize new habitats. These indices are based on a tentative colonizer-persister (*c-p*) scale ranging from 1 to 5. Nematode families comprising species that rapidly increase in number during the early stages of succession, were considered as colonizers and received a low *c-p* value. They have similar characteristics to *r*-strategists. The persisters among the nematodes have more characteristics in common with *K*-strategists. For further information of life-history theory and the MI concept see Whittaker (1975), Stearns (1976), Southwood (1977), Grime (1985), Bongers *et al.* (1991), De Goede *et al.* (1993), Yeates (1994), Bongers *et al.* (1995), Korthals *et al.* (1996).

It is often stated that differences between the relative abundance between different life-history strategy groups might reflect not only the successional stage of a habitat, but also the effects of disturbances (Grime, 1977; Odum, 1985). Therefore, the weighed mean of the distribution between the different *c-p* groupings, i.e. the Maturity Index, might also indicate disturbances. Promising results have been obtained with the MI concept in ecological studies. For example, it has been used to differentiate between tillage regimes (Freckman & Ettema, 1993; Yeates & Bird, 1994; Neher & Campbell, 1994), or to study the influence of manuring (Ettema and Bongers, 1993), as well as in ecotoxicological studies to measure xenobiotic-induced stress (Bongers *et al.*, 1991; Weiss and Larink, 1991; Popovici and Korthals, 1995), the effects of acidification and liming (De Goede and Dekker, 1993), organic pollution in estuarine systems (Essink and Romeyn, 1994) and ammonia deposition (Tamis, 1986 cited in Bongers, 1990).

However, most of the studies to date were observational field studies and only few dealt with the influence of persistent pollutants such as heavy metals. The present thesis will report on microcosm-experiments and one field experiment in which heavy metal-induced changes in terrestrial nematode communities were experimentally imposed and the effects studied.

### Scope of the thesis

The main objectives of the present thesis are to increase our knowledge on pollutant-induced changes in terrestrial nematode communities and to evaluate the Maturity Index and some other community parameters with respect to biomonitoring and risk assessment

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of soil pollution. Therefore, the following experiments, in increasing complexity and, hence, ecological relevance will be presented:

In Chapter 2 the short-term effects of cadmium, copper, nickel and zinc on an indigenous nematode community from an agroecosystem are investigated. This 'natural soil method' can provide a valuable tool for the comparison of the effects of different pollutants on nematodes from several feeding and life-history strategy groups. In Chapter 3, the long-term effects of copper and zinc in relation to the presence of vegetation were investigated, in order to examine the influence of vegetation on ecotoxicological effects. Another aspect of heavy metal pollution in the field is the simultaneous presence of several pollutants. Since today's knowledge on these risks is still poorly developed, the interaction between Cu and Zn upon their bioavailability and their final joint toxicity on soil nematodes are described in Chapter 4. Chapter 5 focuses on the long-term effects of copper and pH on the nematode community under the most realistic conditions, i.e. in a field experiment with normal agricultural practices. In Chapter 6 the nematode community structure and parameters such as the Maturity Index are discussed in the light of their potential for future risk assessment of soil pollution. Finally, the conclusion of these chapters and possible implications for future risk assessment of soil pollution are discussed in Chapter 7.

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## Chapter 2

### SHORT-TERM EFFECTS OF CADMIUM, COPPER, NICKEL AND ZINC ON SOIL NEMATODES FROM DIFFERENT FEEDING AND LIFE-HISTORY STRATEGY GROUPS

#### Abstract

The effects of cadmium, copper, nickel and zinc on a nematode community were examined with a 'natural soil method'. Changes in the indigenous nematode community structure were studied 1-2 weeks after the addition of these metals (as sulphates) to soil collected from an agroecosystem. The soil was acid and only contained a moderate quantity of organic matter as the main metal-binding constituent. As a result, its metal-binding capacity was rather low. The nematode community was found to be affected by increasing concentrations of Cu, Ni and Zn up to 1600 mg kg<sup>-1</sup>, but not by Cd up to 160 mg kg<sup>-1</sup>. EC<sub>50</sub> values for the reduction in population size of individual taxa showed a low intra-taxon variation for Cu, Ni and Zn. For these heavy metals, uptake and elimination processes as well as their final effect appear similar within the same taxon. Omnivorous and predatory nematodes, known to be *K*-strategist, were among the most sensitive taxa, and were already significantly affected by 100 mg kg<sup>-1</sup> Cu, Ni or Zn added to the soil. The relative abundance of the different life-history groups and, to a lesser extent, the different feeding groups indicated pollution-induced changes in the soil community. However, neither classification predicts the acute effects of Cu, Ni and Zn on different nematode genera in an adequate way.

**Keywords:** Nematode community structure; Heavy metals; Life-history strategy; Maturity Index; Bioavailability, Soil pollution

#### Introduction

It has been stated that ecologically relevant bioindicator systems should be based on information relating to different species and ecological processes (Van Straalen and Krivolutsky, 1996). Studying the effects of pollutants on naturally occurring communities of terrestrial organisms can partially fulfil this requirement.

One of the advantages of community level studies is that indirect effects of pollutants are implicitly accounted for. Food availability, competitive interactions between species or the abiotic environment, may also become influenced by pollutants, and finally affect the community in an indirect way (Clements, 1994). Another advantage is that community studies comprise the responses of many taxa which cannot be cultured or kept under laboratory conditions.

Several studies have shown that nematode communities reflect not only soil characteristics, vegetation and management practices (Yeates, 1981; Bongers, 1989; Wasilewska, 1989; Freckman and Ettema, 1993; De Goede and Bongers, 1994; Neher and

Campbell, 1994), but also the effects of pollutants such as heavy metals (Sturhan, 1986; Weiss and Larink, 1991; Parmelee *et al.*, 1993; Yeates *et al.*, 1994). However, despite the high ecological relevance of such studies, relationships between cause and effects are often complex and difficult to assess.

Several nematode classifications have been developed to facilitate the interpretation of changes in the nematode community structure. One of these classifications is based on the different feeding modes among nematodes (e.g. Freckman and Ettema, 1993; Yeates *et al.*, 1993). Another classification is based on the different life-history strategies among nematode families (Bongers, 1990; Bongers *et al.*, 1991; Bongers *et al.*, 1995). In order to calculate the Maturity Index of a nematode community, all non-plant feeding nematode taxa were placed on a colonizer-persister continuum and subsequently assigned *c-p* values ranging from 1 (colonizer, *r*-strategist, tolerant to disturbances) to 5 (persister, *K*-strategist, sensitive to disturbances).

Both the classification of nematodes in feeding groups and in life-history groups could improve the interpretation of pollution-induced changes in nematode communities as well. Therefore the present study will investigate the short-term effects of Cd, Cu, Ni and Zn on an indigenous nematode community from field collected soil of an agroecosystem. The metal addition rate at which the population size of individual taxa is reduced by 50% ( $EC_{50}$ ) will be one of the parameters to compare the sensitivities of nematode taxa belonging to different feeding and life-history groups. As nematode exposure to metals strongly depends on soil-metal interactions, we also obtained information on the metal-binding characteristics of the soil.

## Materials and methods

### *Soil characteristics and analysis*

In October 1992 soil was collected from the top 10 cm of an arable field on sandy soil located 3 km North North East of Wageningen, the Netherlands. From 1980 onwards the field had been cropped with silage maize, starch potatoes and oats in a 3-year rotation (silage maize in 1992). For more details on site and soil see Korthals *et al.* (1996).

Fresh soil was passed through a 10 mm sieve to remove stones, stubble and coarse roots. After mixing, samples were taken for the determination of soil characteristics and initial metal contents. These samples were dried at 30°C and sieved to pass 2 mm. Soil texture was determined by sieve and pipette, organic carbon by acid dichromate oxidation, and the actual cation exchange capacity by the unbuffered barium chloride method. pH-KCl was measured 2 h after suspending 10 ml of soil in 50 ml 1 M KCl (Houba *et al.*, 1989).

Copper and zinc contents were determined by flame atomic absorption spectroscopy (F-AAS) with Smith-Hieftje background correction after digesting samples with a mixture of concentrated nitric and sulphuric acid (1:1 by volume). Nickel was determined in the same

digest by electrothermal atomization AAS (ETA-AAS) with Zeeman background correction. The Cd content was determined by ETA-AAS after extracting samples with 3 M hydrochloric acid for 2 h on a boiling water bath.

#### *Experimental design*

The fresh soil was allowed to dry for 2 days by which time its water content had decreased to 10.9% by weight. Portions of soil equivalent to 3 kg dry weight were then treated with solutions of either Cd, Cu, Ni, or Zn sulphate and demineralized water to a total volume of 100 ml. Sulphates were used so as to limit osmotic effects of the added salts. Precipitation, as hydrated calcium sulphate (gypsum), of added sulphate and calcium ions displaced from exchange sites by adsorbing heavy-metal ions, keeps the dissolved salt concentration at a low level. Cd was applied at rates of 0, 10, 20, 40, 80, and 160 mg kg<sup>-1</sup> dry weight. Cu, Ni and Zn were applied at rates of 0, 100, 200, 400, 800, and 1600 mg kg<sup>-1</sup>. Differences in sulphate additions were balanced by adding calcium sulphate. There were two controls treated with CaSO<sub>4</sub> only: a low CaSO<sub>4</sub> treatment (1.4 mmol kg<sup>-1</sup>) corresponding to the Cd series and a high CaSO<sub>4</sub> treatment (25.6 mmol kg<sup>-1</sup>) corresponding to the other metals series. Each treatment was replicated six times. The treated soil was thoroughly mixed by hand, placed in polythene bags, covered against light and kept at 15°C.

#### *Nematode sampling*

Three of the six replicates were sampled after 1 week. The soil was mixed in its own bag and approximately 100 g was taken out to extract nematodes (Oostenbrink, 1960). Sampling of the remaining three replicates was carried out similarly after 2 weeks. The total abundance of nematodes was estimated by counting approximately 10% of the total sample under a dissecting microscope. The abundance of nematodes was expressed per 100 g dry soil after a correction for material left on the top sieve (2 mm) of the Oostenbrink elutriation apparatus (mainly fine gravel). Nematodes were heat-killed and fixed with formalin (4% at 90°C) and placed on a permanent mass-slide. At least 150 nematodes were identified at 400x-1000x according to Bongers (1988) and allocated to feeding groups according to Yeates *et al.* (1993) and *c-p* groups according to Bongers (1990) and Bongers *et al.* (1995).

#### *Metal adsorption experiments*

Metal binding characteristics of the soil were studied in a batch experiment. Twenty grams of air-dried soil that passed through a 2 mm sieve were shaken end-over-end in 250 ml centrifuge tubes for up to 2 weeks with 200 ml 0.01 M CaCl<sub>2</sub> initially containing 1, 4 or 16 mg l<sup>-1</sup> Cd or 10, 40 or 160 mg l<sup>-1</sup> Cu, Ni or Zn. Metals were added as sulphates and calcium sulphate was added to balance the sulphate additions as in the main experiment. After 1, 3, 6, 24, 48, 96, 192 and 384 h pH was measured in the suspension and a sample was taken from the supernatant obtained by centrifugation at 7,000 rev min<sup>-1</sup> for 5 minutes. Metal concentrations in the supernatant were determined by F-AAS. The amount of metal adsorbed to the soil was calculated from the metal concentration and the total amount of metal added, corrected for the amounts removed by successive samplings.

#### *Data analysis*

Data on nematodes were analyzed by analysis of variance (Sokal and Rohlf, 1981). If necessary, logarithmic transformations were applied to meet assumptions of normality and homogeneity of variances. Tukey's multiple range test was employed to test for differences among treatments.

Replicates of week 1 and week 2 were combined in one group with an exposure time of 1-2 weeks. It was not our intention to focus on differences between 1 or 2 weeks of exposure, but because nematode extraction from all 132 samples could not be done at



once, we decided to sample three replicates after 1 week and the remaining three in the following week. Furthermore, after analyzing the data with ANOVA we detected no major differences between week 1 and week 2.

Metal addition rates at which the population size of individual taxa was reduced by 50% ( $EC_{50}$ ) compared with the control soil, were estimated by using a nonlinear regression estimation procedure (Bruce and Versteeg, 1992). In a number of cases the procedure did not converge. For successful cases, only  $EC_{50}$  values and 95% confidence intervals that fell within the range of metal addition rates will be discussed. Data on rare taxa (with a relative abundance approximately less than 0.5% of the total number of identified nematodes) will be not presented in detail.

## Results

### *Soil characteristics*

The soil characteristics are presented in Table 1. Soil texture and organic matter content are normal for arable land on cover sand, but pH-KCl and actual CEC are rather low. Their values reflect that the soil had not been limed for over a decade. Initial heavy metal contents are within the normal range for Dutch sandy soils and well below the official Dutch reference values that aim to distinguish between uncontaminated and contaminated soils. For this soil the reference values, which depend on clay and organic matter content (De Haan *et al.*, 1990), are 0.50 (Cd), 20 (Cu), 14 (Ni) and 68 (Zn)  $mg\ kg^{-1}$ .

Table 1. Soil characteristics at the beginning of the experiment

|  |      |
|--|------|
| Texture (% m/m on the mineral matter)        |      |
| clay (<2 $\mu m$ )                           | 4    |
| silt (2-50 $\mu m$ )                         | 11   |
| sand (>50 $\mu m$ )                          | 85   |
| Organic C (% m/m on the dry soil)            | 1.9  |
| CEC (cmol <sub>c</sub> $kg^{-1}$ dry weight) | 3.6  |
| pH-KCl                                       | 4.1  |
| Metal content ( $mg\ kg^{-1}$ dry weight)    |      |
| Cd   | 0.33 |
| Cu   | 11   |
| Ni   | 4.1  |
| Zn   | 38   |

% m/m; percentage by mass

### *Metal adsorption*

During the 2 weeks of the metal adsorption experiment, metal concentrations in solution continued to decrease. The pH increased by 0.2 units, irrespective of the metal treatment.

In treatments without heavy metals (only  $\text{CaSO}_4$  added) pH measured 4.40 after 1 h and 4.59 after 384 h. Metal treatments lowered pH, the size of the effect depending on both the metal and its addition rate (Fig. 1) but not on sampling time. There was a linear relationship between pH and the logarithm of time ( $t$ , h). Averaged over all treatments, this relationship was described by:

$$\text{pH} = 4.29 + 0.079(\pm 0.006) \log t$$

which accounted for 96.6% of the variance due to the time factor. Higher-order terms did not significantly improve the relationship.

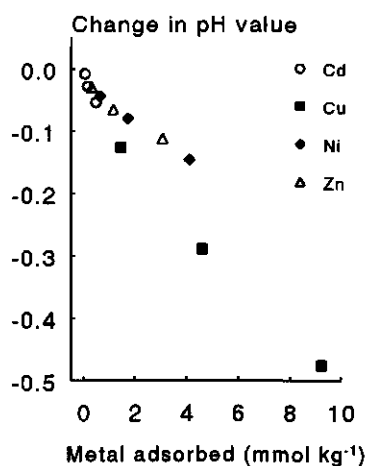


Figure 1. Effects of metal addition on pH at  $t = 384$  h (LSD = 0.02;  $P < 0.05$ )

Table 2. Parameters of the regression model  $\log q = \alpha + \beta \cdot \log c + \Gamma \cdot \log t$  ( $q$  is the amount of metal adsorbed in  $\text{mg kg}^{-1}$  and  $c$  is the metal concentration in solution in  $\text{mg l}^{-1}$ )

| Metal | $\alpha$ | $\beta$ | $\Gamma$ | $R^2$ <sup>a</sup> | SE <sup>b</sup> |
|-------|----------|---------|----------|--------------------|-----------------|
| Cd    | 0.4995   | 0.7941  | 0.1468   | 0.9934             | 0.0373          |
| Cu    | 1.7033   | 0.4082  | 0.1228   | 0.9872             | 0.0392          |
| Ni    | 0.6364   | 0.5899  | 0.1661   | 0.9698             | 0.0622          |
| Zn    | 0.2261   | 0.8350  | 0.1435   | 0.9660             | 0.0862          |

<sup>a</sup> Coefficient of determination. <sup>b</sup> Standard error of regression.

In general, data on metal adsorption cannot be described by single-reaction kinetic models (Amacher *et al.*, 1986). In this case, the relationship between pH and time suggested a Freundlich equation, which, extended with a time factor, could serve to summarize the data. The basic equation reads

$$\log q = \alpha + \beta \cdot \log c + \Gamma \cdot \log t$$

where  $q$  is the amount of metal adsorbed ( $\text{mg kg}^{-1}$ ) and  $c$  is the metal concentration in solution ( $\text{mg l}^{-1}$ ). The results are shown in Table 2.

Table 3. Relative abundances (as a percentage of the total nematode abundance), feeding groups,  $c$ - $p$  score and total abundances in the different control treatments (mean  $\pm$  SE;  $n = 6$ ).

| Taxon                          | $c$ - $p$<br>score | Feeding<br>group <sup>a</sup> | Untreated soil<br>$t=0$      | Low $\text{CaSO}_4$<br>$t=1-2$ weeks | High $\text{CaSO}_4$<br>$t=1-2$ weeks | $P^b$ |
|--------------------------------|--------------------|-------------------------------|------------------------------|--------------------------------------|---------------------------------------|-------|
| <i>Filenchus</i>               |                    | P                             | 4.1 $\pm$ 0.7                | 6.0 $\pm$ 1.2                        | 5.5 $\pm$ 1.0                         |       |
| <i>Tylenchus</i>               |                    | P                             | 0.9 $\pm$ 0.2                | 0.8 $\pm$ 0.5                        | 0.8 $\pm$ 0.5                         |       |
| <i>Pratylenchus</i>            |                    | P                             | 12.7 $\pm$ 1.0 <sup>a</sup>  | 20.4 $\pm$ 3.2 <sup>b</sup>          | 19.7 $\pm$ 1.1 <sup>b</sup>           | *     |
| <i>Rotylenchus</i>             |                    | P                             | 0.7 $\pm$ 0.2                | 1.5 $\pm$ 0.7                        | 1.0 $\pm$ 0.4                         |       |
| <i>Tylenchorhynchus</i>        |                    | P                             | 9.6 $\pm$ 1.1                | 13.7 $\pm$ 1.8                       | 12.1 $\pm$ 1.1                        |       |
| Dauer-larvae only              | 1                  | B                             | 15.5 $\pm$ 1.6 <sup>b</sup>  | 9.1 $\pm$ 4.3 <sup>ab</sup>          | 3.4 $\pm$ 1.6 <sup>a</sup>            | *     |
| Rhabditidae                    | 1                  | B                             | 42.3 $\pm$ 1.8 <sup>b</sup>  | 30.6 $\pm$ 3.8 <sup>a</sup>          | 33.9 $\pm$ 2.2 <sup>ab</sup>          | *     |
| <i>Acrobeles</i>               | 2                  | B                             | 1.3 $\pm$ 0.3                | 1.8 $\pm$ 0.5                        | 1.2 $\pm$ 0.2                         |       |
| <i>Acrobeloides</i>            | 2                  | B                             | 7.5 $\pm$ 0.6                | 4.8 $\pm$ 1.0                        | 6.2 $\pm$ 1.1                         |       |
| <i>Cephalobus</i>              | 2                  | B                             | 0.2 $\pm$ 0.1                | 1.6 $\pm$ 0.7                        | 0.8 $\pm$ 0.5                         |       |
| <i>Cervidellus</i>             | 2                  | B                             | 0.1 $\pm$ 0.1                | 0.1 $\pm$ 0.1                        | 0.3 $\pm$ 0.1                         |       |
| <i>Eucephalobus</i>            | 2                  | B                             | 3.1 $\pm$ 0.4 <sup>b</sup>   | 0.8 $\pm$ 0.5 <sup>a</sup>           | 1.7 $\pm$ 0.5 <sup>ab</sup>           | **    |
| <i>Plectus</i>                 | 2                  | B                             | 1.5 $\pm$ 0.3                | 1.7 $\pm$ 0.2                        | 1.6 $\pm$ 0.1                         |       |
| <i>Alaimus</i>                 | 4                  | B                             | 0.3 $\pm$ 0.1                | 0.7 $\pm$ 0.5                        | 0.5 $\pm$ 0.2                         |       |
| <i>Aphelenchoides</i>          | 2                  | H                             | 4.2 $\pm$ 0.6                | 4.4 $\pm$ 1.1                        | 5.5 $\pm$ 1.3                         |       |
| <i>Ditylenchus</i>             | 2                  | H                             | 1.3 $\pm$ 0.3                | 0.4 $\pm$ 0.2                        | 0.3 $\pm$ 0.3                         |       |
| <i>Pseudhalenchus</i>          | 2                  | H                             | 2.8 $\pm$ 0.6                | 2.3 $\pm$ 0.9                        | 2.6 $\pm$ 0.2                         |       |
| <i>Clarkus</i>                 | 4                  | C                             | 2.3 $\pm$ 0.5                | 1.9 $\pm$ 0.8                        | 1.3 $\pm$ 0.4                         |       |
| <i>Aporcelaimellus</i>         | 5                  | O                             | 1.4 $\pm$ 0.3                | 3.0 $\pm$ 0.6                        | 1.9 $\pm$ 0.4                         |       |
| Plant feeding                  |                    |                               | 28.6 $\pm$ 1.3 <sup>a</sup>  | 42.7 $\pm$ 5.0 <sup>b</sup>          | 39.9 $\pm$ 2.6 <sup>b</sup>           | *     |
| Bacterial feeding              |                    |                               | 58.0 $\pm$ 1.7 <sup>b</sup>  | 44.5 $\pm$ 3.7 <sup>a</sup>          | 47.2 $\pm$ 2.5 <sup>a</sup>           | **    |
| Hyphal feeding                 |                    |                               | 9.5 $\pm$ 1.4                | 7.9 $\pm$ 1.4                        | 9.5 $\pm$ 1.3                         |       |
| Omnivorous/predators           |                    |                               | 3.9 $\pm$ 0.7                | 4.9 $\pm$ 0.7                        | 3.4 $\pm$ 0.6                         |       |
| $c$ - $p$ 1                    |                    |                               | 60.0 $\pm$ 2.0               | 52.7 $\pm$ 2.4                       | 56.6 $\pm$ 2.5                        |       |
| $c$ - $p$ 2                    |                    |                               | 32.8 $\pm$ 2.2               | 35.2 $\pm$ 1.4                       | 35.7 $\pm$ 2.5                        |       |
| $c$ - $p$ 3                    |                    |                               | 1.3 $\pm$ 0.4                | 1.8 $\pm$ 0.6                        | 1.1 $\pm$ 0.3                         |       |
| $c$ - $p$ 4                    |                    |                               | 4.0 $\pm$ 0.9                | 4.7 $\pm$ 1.3                        | 3.3 $\pm$ 0.8                         |       |
| $c$ - $p$ 5                    |                    |                               | 2.0 $\pm$ 0.4 <sup>a</sup>   | 5.6 $\pm$ 1.5 <sup>b</sup>           | 3.3 $\pm$ 0.7 <sup>ab</sup>           | *     |
| Maturity index                 |                    |                               | 1.55 $\pm$ 0.03 <sup>a</sup> | 1.75 $\pm$ 0.07 <sup>b</sup>         | 1.61 $\pm$ 0.05 <sup>ab</sup>         | *     |
| Total abundance<br>(per 100 g) |                    |                               | 2927 $\pm$ 162 <sup>b</sup>  | 1879 $\pm$ 143 <sup>a</sup>          | 2140 $\pm$ 77 <sup>a</sup>            | ***   |

<sup>a</sup> P, plant feeding; B, bacterial feeding; H, hyphal feeding; O, omnivorous; C, predators.

<sup>b</sup> \*  $P < 0.05$ ; \*\*  $P < 0.01$ ; \*\*\*  $P < 0.001$ . Significant differences between the three means within rows ( $P < 0.05$ , HSD) are indicated by different letters.

*Nematodes: Control soils*

The total nematode fauna consisted of 36 genera among 25 families, of which on average 17 different taxa were found in the control samples. Most taxa belonged to the bacterial feeders or plant feeders, of which Rhabditidae, Pratylenchidae, Dolichodoridae, Cephalobidae and Tylenchidae formed approximately 80% of the total nematode fauna (Table 3). Some parameters of the nematode community found at the start of the experiment were significantly different from data obtained for both control soils after 1-2 weeks experimentation. However, no significant differences could be detected as a consequence of the different amounts of  $\text{CaSO}_4$  added.

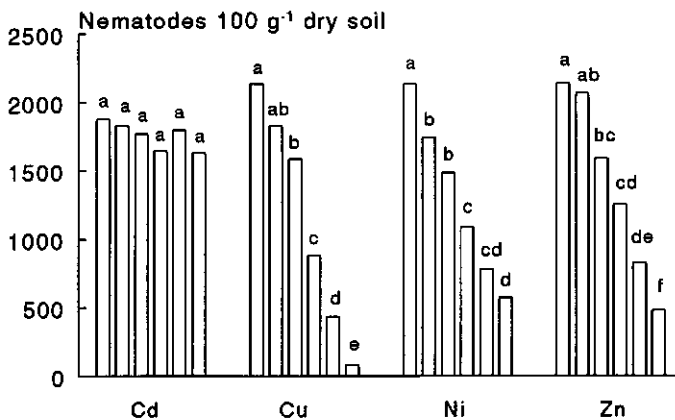


Figure 2. Effects of Cd (0, 10, 20, 40, 80 and 160  $\text{mg kg}^{-1}$ ) and Cu, Ni and Zn (0, 100, 200, 400, 800 and 1600  $\text{mg kg}^{-1}$ ) on total nematode abundance after 1-2 weeks exposure. Different letters indicate significant differences between treatments with one metal (HSD test,  $P < 0.05$ ).

*Nematodes: Metal addition experiments**1. Total abundance of nematodes and trophic structure*

The total abundance of nematodes significantly decreased with higher Cu, Ni and Zn dosages, but were not affected by the investigated range of Cd concentrations (Fig. 2). Omnivorous and predatory nematodes appeared very sensitive and their population size was already significantly smaller in soils with Cu, Ni or Zn additions of 100  $\text{mg kg}^{-1}$  (Fig. 3).

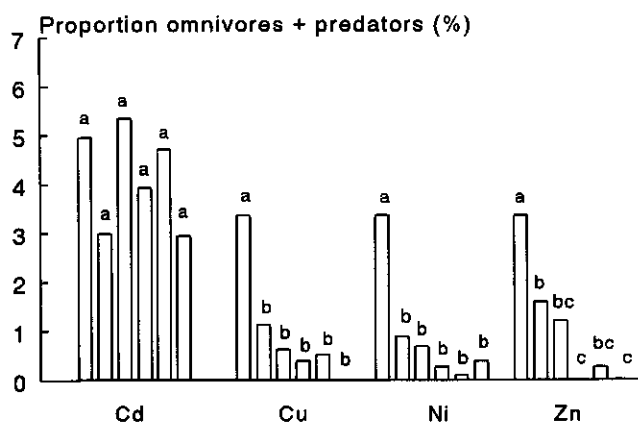


Figure 3. Effects on omnivores and predators (as a percentage of the total nematode community) of exposure to Cd (0, 10, 20, 40, 80 and 160 mg kg<sup>-1</sup>) and Cu, Ni and Zn (0, 100, 200, 400, 800 and 1600 mg kg<sup>-1</sup>) for 1-2 weeks. Different letters indicate significant differences between treatments with one metal (HSD test,  $P < 0.5$ ).

The relative abundances of bacterial, plant and hyphal feeding nematodes, were only significantly affected at the highest Cu addition of 1600 mg kg<sup>-1</sup> (Table 4). The relative abundance of bacterial feeding nematodes was significantly higher, and the relative abundances of the plant and hyphal feeding nematodes were significantly lower than in the control soils (Table 4).

## 2. Composition of taxa

The addition of Cu, Ni and Zn changed the composition of nematode taxa (Table 4). The relative abundances of most taxa were reduced at the highest metal additions, but for some metals *Pratylenchus* (Zn), Dauer-larvae of the Rhabditidae (Cu), Rhabditidae (Cu) and *Ditylenchus* (Ni and Zn) were found in significantly higher relative abundances than in the control soils. The EC<sub>50</sub> values for several nematode taxa are presented in Table 5. The differences between the lowest and highest EC<sub>50</sub> values found for taxa exposed to the same metal differed by a factor 11, 38 or 30 for Cu, Ni and Zn, respectively. Intra-taxon differences in the EC<sub>50</sub> values were always less than a factor 4.5. In general, *Pseudhalenchus*, Dauer-larvae of the Rhabditidae, *Aphelenchoides*, *Pratylenchus* and

*Tylenchorhynchus* were the most tolerant to Cu, Ni and Zn. *Plectus*, *Clarkus*, *Aporcelaimellus*, *Prismatolaimus*, *Alaimus* and *Acrobeles* appeared to be the most sensitive taxa.

Table 4. Relative abundances (as a percentage of the total nematode abundance) , feeding groups, c-p score and total abundances after 1-2 weeks exposure to the highest metal concentrations (mean  $\pm$  SE; n = 6)

| Taxon                       | c-p score | Feeding group <sup>a</sup> | Cadmium (160 mg kg <sup>-1</sup> ) | Copper (1600 mg kg <sup>-1</sup> ) | Nickel (1600 mg kg <sup>-1</sup> ) | Zinc (1600 mg kg <sup>-1</sup> )   |
|-----------------------------|-----------|----------------------------|------------------------------------|------------------------------------|------------------------------------|------------------------------------|
| <i>Filenchus</i>            |           | P                          | 3.1 $\pm$ 0.5                      | 0.4 $\pm$ 0.4*                     | 1.3 $\pm$ 0.5*                     | 2.0 $\pm$ 0.4*                     |
| <i>Tylenchus</i>            |           | P                          | 0.7 $\pm$ 0.3                      | 0.4 $\pm$ 0.4                      | 0.0 $\pm$ 0.0                      | 0.5 $\pm$ 0.2                      |
| <i>Pratylenchus</i>         |           | P                          | 24.7 $\pm$ 3.1                     | 17.6 $\pm$ 3.0                     | 26.0 $\pm$ 3.3                     | <b>29.5 <math>\pm</math> 1.2*</b>  |
| <i>Rotylenchus</i>          |           | P                          | 1.0 $\pm$ 0.5                      | 1.3 $\pm$ 1.3                      | 2.3 $\pm$ 0.7                      | 1.8 $\pm$ 0.6                      |
| <i>Tylenchorhynchus</i>     |           | P                          | 13.1 $\pm$ 1.2                     | 3.2 $\pm$ 1.4*                     | 14.8 $\pm$ 1.4                     | 13.3 $\pm$ 1.8                     |
| Dauer-larvae only           | 1         | B                          | 0.0 $\pm$ 0.0                      | <b>59.9 <math>\pm</math> 5.5*</b>  | 5.0 $\pm$ 2.3                      | 14.1 $\pm$ 7.0                     |
| Rhabditidae                 | 1         | B                          | 30.0 $\pm$ 2.4                     | <b>73.7 <math>\pm</math> 4.5*</b>  | 43.2 $\pm$ 3.0                     | 39.3 $\pm$ 2.9                     |
| <i>Acrobeles</i>            | 2         | B                          | 0.5 $\pm$ 0.4                      | 0.0 $\pm$ 0.0*                     | 0.2 $\pm$ 0.1*                     | 0.0 $\pm$ 0.0*                     |
| <i>Acrobeloides</i>         | 2         | B                          | 9.1 $\pm$ 1.4                      | 0.0 $\pm$ 0.0*                     | 1.7 $\pm$ 0.3*                     | 2.0 $\pm$ 0.5*                     |
| <i>Cephalobus</i>           | 2         | B                          | 0.8 $\pm$ 0.4                      | 0.0 $\pm$ 0.0                      | 0.6 $\pm$ 0.2                      | 0.3 $\pm$ 0.2                      |
| <i>Cervidellus</i>          | 2         | B                          | 0.3 $\pm$ 0.1                      | 0.0 $\pm$ 0.0                      | 0.1 $\pm$ 0.1                      | 0.0 $\pm$ 0.0*                     |
| <i>Eucephalobus</i>         | 2         | B                          | 3.4 $\pm$ 0.4                      | 0.0 $\pm$ 0.0                      | 0.1 $\pm$ 0.1*                     | 0.0 $\pm$ 0.0                      |
| <i>Heterocephalobus</i>     | 2         | B                          | 0.3 $\pm$ 0.3                      | 0.0 $\pm$ 0.0                      | 0.0 $\pm$ 0.0                      | 0.0 $\pm$ 0.0                      |
| <i>Plectus</i>              | 2         | B                          | 0.7 $\pm$ 0.4                      | 0.4 $\pm$ 0.4*                     | 0.2 $\pm$ 0.2*                     | 0.0 $\pm$ 0.0*                     |
| <i>Prismatolaimus</i>       | 3         | B                          | 1.0 $\pm$ 0.1                      | 0.0 $\pm$ 0.0                      | 0.0 $\pm$ 0.0*                     | 0.1 $\pm$ 0.1                      |
| <i>Alaimus</i>              | 4         | B                          | 0.6 $\pm$ 0.2                      | 0.0 $\pm$ 0.0                      | 0.0 $\pm$ 0.0                      | 0.0 $\pm$ 0.0                      |
| <i>Aphelenchoides</i>       | 2         | H                          | 2.9 $\pm$ 0.6                      | 1.8 $\pm$ 0.8*                     | 2.2 $\pm$ 0.5                      | 3.2 $\pm$ 0.4                      |
| <i>Ditylenchus</i>          | 2         | H                          | 1.0 $\pm$ 0.5                      | 0.0 $\pm$ 0.0                      | <b>2.3 <math>\pm</math> 0.5*</b>   | <b>2.6 <math>\pm</math> 0.7*</b>   |
| <i>Pseudhalenchus</i>       | 2         | H                          | 3.3 $\pm$ 0.8                      | 0.7 $\pm$ 0.7*                     | 3.0 $\pm$ 0.5                      | 4.7 $\pm$ 0.9                      |
| <i>Clarkus</i>              | 4         | C                          | 1.1 $\pm$ 0.3                      | 0.0 $\pm$ 0.0*                     | 0.1 $\pm$ 0.1*                     | 0.0 $\pm$ 0.0*                     |
| <i>Aporcelaimellus</i>      | 5         | O                          | 1.7 $\pm$ 0.4                      | 0.0 $\pm$ 0.0*                     | 0.1 $\pm$ 0.1*                     | 0.0 $\pm$ 0.0*                     |
| Plant feeding               |           |                            | 42.7 $\pm$ 2.5                     | 23.4 $\pm$ 3.2 *                   | 45.2 $\pm$ 3.9                     | 47.6 $\pm$ 2.1                     |
| Bacterial feeding           |           |                            | 46.8 $\pm$ 2.0                     | <b>73.8 <math>\pm</math> 3.1 *</b> | 46.1 $\pm$ 3.2                     | 41.8 $\pm$ 2.4                     |
| Hyphal feeding              |           |                            | 7.5 $\pm$ 0.7                      | 2.9 $\pm$ 0.9 *                    | 8.2 $\pm$ 1.0                      | 10.5 $\pm$ 0.9                     |
| c-p 1                       |           |                            | 52.2 $\pm$ 2.3                     | <b>95.1 <math>\pm</math> 1.6 *</b> | <b>79.2 <math>\pm</math> 2.1 *</b> | <b>74.8 <math>\pm</math> 2.6 *</b> |
| c-p 2                       |           |                            | 39.9 $\pm$ 2.5                     | 4.3 $\pm$ 1.3 *                    | 20.1 $\pm$ 1.8 *                   | 25.0 $\pm$ 2.5                     |
| c-p 3                       |           |                            | 1.8 $\pm$ 0.3                      | 0.6 $\pm$ 0.4                      | 0.0 $\pm$ 0.0                      | 0.2 $\pm$ 0.2                      |
| c-p 4                       |           |                            | 3.0 $\pm$ 0.6                      | 0.0 $\pm$ 0.0*                     | 0.2 $\pm$ 0.2*                     | 0.0 $\pm$ 0.0*                     |
| c-p 5                       |           |                            | 3.0 $\pm$ 0.7                      | 0.0 $\pm$ 0.0*                     | 0.2 $\pm$ 0.2*                     | 0.0 $\pm$ 0.0*                     |
| Total abundance (per 100 g) |           |                            | 1635 $\pm$ 65                      | 84 $\pm$ 15*                       | 573 $\pm$ 77*                      | 479 $\pm$ 89 *                     |

<sup>a</sup> P, plant feeding; B, bacterial feeding; H, hyphal feeding; O, omnivorous; C, predators.

Values significantly different from controls at  $P < 0.05$  are indicated by asterisks (HSD within one metal series). Values in bold type are significantly higher than the control.

Table 5. Estimates of  $EC_{50}$  ( $mg\ kg^{-1}$ ) after an exposure period of 1-2 weeks (95% confidence interval in parentheses,  $n = 36$ ).

| Taxon                   | c-p score | Feeding group <sup>a</sup> | Copper |           | Nickel |            | Zinc  |             |
|-------------------------|-----------|----------------------------|--------|-----------|--------|------------|-------|-------------|
| <i>Filenchus</i>        |           | P                          | 455    | (253-820) | 364    | (250-478)  | 141   | (45-438)    |
| <i>Pratylenchus</i>     |           | P                          | 508    | (322-802) | 883    | (335-2328) | 902   | (418-1945)  |
| <i>Tylenchorhynchus</i> |           | P                          | 175    | (91-335)  | 682    | (353-1315) | 710   | (385-1303)  |
| Dauer-larvae only       | 1         | B                          | >1600  |           | 658    | (211-2046) | 1538  | (1294-1824) |
| Rhabditidae             | 1         | B                          | 497    | (352-697) | 321    | (89-1496)  | 444   | (155-1268)  |
| <i>Acrobeles</i>        | 2         | B                          | 147    | (72-299)  | 64     | (12-355)   |       |             |
| <i>Acrobeloides</i>     | 2         | B                          | 287    | (183-451) | 386    | (222-671)  | 493   | (308-789)   |
| <i>Eucephalobus</i>     | 2         | B                          | 206    | (107-394) |        |            | 300   | (135-670)   |
| <i>Plectus</i>          | 2         | B                          | 97     | (36-262)  | 23     | (3-167)    | 52    | (20-138)    |
| <i>Prismatolaimus</i>   | 3         | B                          |        |           | 62     | (10-400)   |       |             |
| <i>Alaimus</i>          | 4         | B                          | 110    | (43-281)  | <100   |            |       |             |
| <i>Aphelenchoides</i>   | 2         | H                          | 416    | (244-710) | 286    | (87-942)   | 527   | (318-873)   |
| <i>Pseudhalenchus</i>   | 2         | H                          | 538    | (361-800) | 836    | (245-2838) | >1600 |             |
| <i>Clarkus</i>          | 4         | C                          | 65     | (20-206)  | 62     | (15-253)   | <100  |             |
| <i>Aporcelaimellus</i>  | 5         | O                          | 48     | (13-172)  | <100   |            | 145   | (69-303)    |

<sup>a</sup> P, plant feeding; B, bacterial feeding; H, hyphal feeding; O, omnivorous; C, predators.

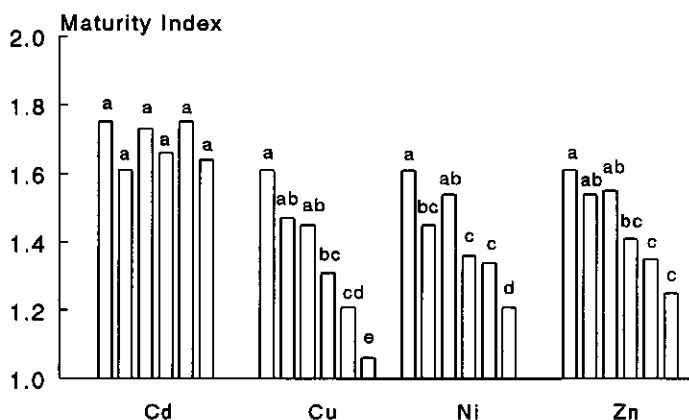


Figure 4. Effects of Cd (0, 10, 20, 40, 80 and 160  $mg\ kg^{-1}$ ) and Cu, Ni and Zn (0, 100, 200, 400, 800 and 1600  $mg\ kg^{-1}$ ) on the maturity index after 1-2 weeks exposure. Different letters indicate significant differences between treatments with one metal (HSD test,  $P < 0.05$ ).

### 3. Maturity Index

The Maturity Index was significantly lower in soils with higher concentrations of Cu, Ni and Zn (Fig. 4). These differences were driven by relative changes in the *c-p* group distribution among the non-plant feeding nematodes (Table 4), mainly because some taxa with *c-p* values of 1 or 2, e.g. Rhabditidae (including the Dauer-larvae), *Aphelenchoides* and *Pseudhalenchus*, were relatively less sensitive to higher Cu, Ni and Zn dosages than most other taxa.

### Discussion

Since the present study was originally started as a range finding experiment focusing on short-term effects, the metal addition rates used were high. For this soil the official Dutch intervention values that separate seriously and not seriously contaminated soils are approximately 8 (Cd), 100 (Cu), 80 (Ni) and 340 (Zn) mg kg<sup>-1</sup>. Except for Zn, the lowest addition rate already increased the metal content in soil to a level exceeding the intervention value.

Cd additions up to 160 mg kg<sup>-1</sup> had no acute effects on the nematode community. Several other studies have mentioned the relatively high tolerance of nematodes to Cd (Haight *et al.*, 1982; Williams and Dusenbery, 1990; Kammenga *et al.*, 1994). The results of the adsorption experiment indicate that the absence of Cd effects in this study is not a consequence of a much stronger adsorption of Cd compared to the other metals, but that it is due to the lower addition rates. Cd was added at rates 10 times lower than the other metals because its intervention value is also lower by an order of magnitude. The rationale behind the Dutch intervention values for all 4 metals tested is the existence of 'serious danger' for a soil ecosystem when the survival of 50% of its species is threatened because the NOEC for effects on vital life-history traits like reproduction and growth is exceeded (Denneman and Van Gestel, 1990). The method applied in the derivation of intervention values takes account of toxicological data on all species (Van Straalen and Denneman, 1989). The ten times lower intervention value for Cd compared to Cu can be traced back to Cd being much more toxic to plants, as well as to oribatid mites and springtails (when exposed via food). Effects of Cd and Cu on earthworm fecundity, however, occur at quite similar concentrations in soil and would, when considered separately, not justify such a large difference in intervention values.

The Cu, Ni and Zn additions up to 1600 mg kg<sup>-1</sup> significantly affected many parameters of the nematode community structure, such as the populations of certain omnivorous and



predatory nematodes with a *K*-strategist type of life-history. The populations of several nematode taxa were already significantly affected by the lowest Cu, Ni and Zn addition of 100 mg kg<sup>-1</sup>. This agrees with other studies on the effects of pollution in the short-term (Parmelee *et al.*, 1993; Kammenga *et al.*, 1994) as well as in the long-term (Zullini and Peretti, 1986; Weiss and Larink, 1991; Popovici and Korthals, 1995), although for some predatory nematode taxa contradicting results have been mentioned (Kappers and Wondergem-van Eijk, 1988; Yeates *et al.*, 1994).

The differences between the lowest and highest EC<sub>50</sub> for different taxa exposed to the same metal were much larger than between the EC<sub>50</sub> values for Cu, Ni or Zn within the same taxon. This may indicate that, at least for Cu, Ni and Zn, uptake and elimination processes as well as their final effect do not differ very much within a nematode taxon. Although this may not be true for organic pollutants, such as pentachlorophenol (Kammenga *et al.*, 1994), it emphasizes the importance of investigating which characteristics of nematodes define their sensitivity to these heavy metals.

Our data showed that closely related genera with similar feeding modes and *c-p* values, such as *Acrobelloides* and *Acrobeles* within the family Cephalobidae, can have very different toxicological responses. Furthermore, it was found that some genera with a *c-p* value of 2 (e.g. *Plectus* and *Acrobeles*) were as sensitive as genera with higher *c-p* values. These results indicate that our current knowledge is not sufficient to correctly place all nematode taxa in *c-p* groups or that the relationship between this classification and the sensitivity to short-term effects of these heavy metals may not be as straightforward as presumed.

One of the problems seems to be that present knowledge on feeding behaviour and life-history strategies among nematodes is poorly developed, and that most information is only available at a broad level of taxonomic resolution. Phylogenetic relationships, mainly based on morphological aspects, are used to make assumptions on characteristics for other, less well studied taxa. One of the difficulties with this approach is that the present classification of nematodes does not always reflect monophyletic groups (Bongers *et al.*, 1991) and that differences in traits defining life-history strategies, such as developmental rate and reproductive mode, between nematode families as well as between closely related genera do exist.

The results on the Rhabditidae demonstrate that increasing knowledge on ecological characteristics of nematode taxa can help to understand their ecotoxicological response. In contrast with the Cu-induced decline among the Rhabditidae, the abundance of Dauer-larvae was hardly affected by the treatments. It seems that Dauer-larvae cannot only

survive periods of low food availability, but also periods of exposure to pollutants. This capacity is found for the enrichment opportunists (De Goede *et al.*, 1993) among nematodes and was in fact an important additional criterium to classify nematodes in *c-p* group 1 (Bongers *et al.*, 1995).

Although the present paper only studied the short-term effects of fairly high Cd, Cu, Ni and Zn additions in one soil type, we believe that experiments exposing indigenous nematode populations in their natural soil probably lead to less under- or overestimation of the real risks, then experiments carried out in water or artificial soil. The use of natural soil, however, does give rise to some complications.

For example, the soil used in this experiment was acid and only contained a moderate quantity of organic matter as the main metal-binding constituent. As a result, its metal-binding capacity was rather low. After an equilibration period of 14 days, the percentage of Cu retained by the soil ranged from 93.2% at the lowest addition rate to 36.9% at the highest rate. For Zn, the least strongly bound metal, the corresponding figures were only 20.6% and 13.0%, respectively. From the results of the adsorption experiment (Fig. 1, Table 2) it appeared that the binding of Cu and Cd was mainly effected by chemisorption, but the binding of Zn and Ni, at least at the higher addition rates, mainly by cation exchange (McBride, 1994). Had this experiment been done with a soil with a higher binding capacity, the metal concentration in solution would have been lower, metal exposure less intense and  $EC_{50}$  values higher than found now. As the relationships between soil composition and pH on the one hand and metal ion adsorption on the other differ quantitatively between heavy metals, it is also conceivable that  $EC_{50}$  values for different metals change order when another soil is used.

This also seems true for the local nematode community. Soil characteristics not only affect the bioavailability of pollutants, but they can have a pronounced influence on the nematode community structure itself. The nematode community at the start of the experiment was comparable with data obtained from other arable agroecosystems (Wasilewska, 1989; Weiss and Larink, 1991; Freckman and Ettema, 1993) and in general quite different from nematode communities found in other ecosystems. Although not investigated, it can be assumed that these differences among nematode communities will affect ecotoxicological data such as  $EC_{50}$  values for the Maturity Index or percentage bacterivorous nematodes. In the present study there were no major differences in most parameters obtained from the control soil at the start and at the end of the two weeks, which indicated that the influence of mixing the soil and additions of water and  $CaSO_4$  probably did not influence the present data to a great extent. However, had this experiment

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been done with a nematode community from a more mature ecosystem like a forest, then the effect of mixing was probably more pronounced and could have interfered with the final effect of metal additions.

## Conclusion

Changes in the relative abundances of nematode life-history groups, reflected by the *c-p* scaling, as well as in the nematode feeding groups, formed an indication of short-term effects of Cu, Ni and Zn on the soil nematode community. However, both characteristics cannot adequately predict differences in sensitivity found among nematode genera. Nevertheless, both classification methods do not only facilitate the interpretation of pollution-induced changes in nematode communities, but can also help to investigate which characteristics within the different groups of nematode taxa are of importance in defining their sensitivity to pollutants. The 'natural soil method' presented in this study seems very promising for this kind of investigations. However, we conclude that unless much effort is put in measuring the bioavailability of pollutants, it seems that the advantages of using this method mainly lie in fundamental studies. Additionally, the 'natural soil method' may be valuable in site-specific risk assessment studies.

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## Chapter 3

### INFLUENCE OF PERENNIAL RYEGRASS ON A COPPER AND ZINC AFFECTED TERRESTRIAL NEMATODE COMMUNITY

#### Abstract

Effects of copper or zinc (0, 25, 50, 100, 200 and 400 mg kg<sup>-1</sup>) to soil containing an indigenous nematode community were examined in the presence or absence of ryegrass (*Lolium perenne* L.). Increasing Cu and Zn additions had negative effects on the total abundance, average number of taxa, proportion of plant-feeding nematodes, proportion of omnivorous and carnivorous nematodes, nematode taxa from *c-p* groups 4 and 5 and the Maturity Index, both in the presence and absence of *L. perenne*. At intermediate Cu or Zn concentrations of 25-200 mg kg<sup>-1</sup>, certain taxa had higher absolute abundances than in the control, which indicated indirect effects. Final effects of Cu or Zn were less extreme and were caused more often in an indirect way in the presence than in the absence of *L. perenne*. These results imply that the presence of vegetation is an important factor in determining the final ecotoxicological effect of pollutants. The risk assessment of pollutants should not only include the response of communities of soil organisms, but this response should be investigated in a realistic way, e.g. in the presence of plants.

**Keywords:** Nematodes; Zinc; Copper; Community; Soil pollution; Bioavailability

#### Introduction

Potential risks of pollutants have often been assessed on the basis of single species toxicity tests in the laboratory in combination with pollutant concentrations measured in the field. Apart from the difficulty of measuring relevant concentrations of pollutants in the field, discrepancies between results of laboratory and field studies may also be caused by interactions among organisms and between organisms and their environment (Cairns, 1983). In contrast to direct effects that follow on the action of a toxicant on receptor sites within the organism itself, these indirect effects occur when the pollutant interferes with for example the food availability or predator-prey interactions. It has been suggested that under realistic conditions, indirect effects may be more important for ecosystem functioning than direct effects, although they are more difficult to predict (Yodzis, 1988).

In order to improve our understanding of indirect effects, multispecies micro- and mesocosms tests are currently in use. They enable us to judge whether 'rebuilding nature' is necessary for the risk assessment of pollutants. A step in the direction of increasing ecological complexity is to expose indigenous nematode communities in micro- or mesocosms (Parmelee *et al.*, 1993; Korthals *et al.*, 1996b). Among soil ecotoxicologists these tests are typically done without plants. Plants, however, have a pronounced impact

on biotic and abiotic processes in soil. With respect to nematodes, this is most straightforward for plant feeding nematodes, of which some taxa are dependent on a single host species. Some other examples of interactions between vegetation and nematodes are the observations that grass species diversity is correlated with the nematode diversity (Wasilewska, 1995), and that certain plant-feeding nematode species play an essential role in the succession of vegetation (Van der Putten *et al.*, 1993).

With respect to the impact of pollutants, plants may also affect the bioavailability of pollutants, as observed earlier for aquatic ecosystems (Brock *et al.*, 1992), and the toxic effects on plants may in turn influence the nematode community. Hence, it seems obvious that the relationship between vegetation and nematodes can strongly influence the impact of pollutants on the nematode community as well. Therefore, we examined the effects of copper and zinc on nematode communities in bare soil and soil covered with *L. perenne*. This species is the most widely utilized in West European grasslands and is not very sensitive to heavy metals (Dijkshoorn *et al.*, 1979). It is hypothesized that changes in the nematode community structure will depend not only on the metal concentrations in soil, but also on the presence of vegetation.

## Materials and methods

### *Soil and treatments*

In October 1992, soil was collected from the top 10 cm of an arable field on sandy soil located 3 km NNE of Wageningen, the Netherlands. From 1980 onwards the field had been cropped with silage maize, starch potatoes and oats in a 3-year rotation (silage maize in 1992). Some soil characteristics are listed in Table 1. For more details on site and soil see Korthals *et al.* (1996a).

After removing stones and organic material of > 1 cm, the fresh soil was mixed and dried to a water content of 10.9% by weight, to allow the addition of metal solutions without exceeding the field capacity. Copper and zinc were added to the soil in two steps in order to prevent the immediate extinction of nematodes, due to the higher bioavailable metal concentrations before an equilibrium between free and adsorbed metals has been reached.

Firstly, both metals (as sulphates) were applied at 0, 100, 200, 400, 800 and 1600 mg kg<sup>-1</sup> by mixing 0-100 ml stock solutions with 3.3 kg fresh soil (equivalent to 3 kg dry soil). Differences in water and sulphate additions were balanced by adding demineralized water and calcium sulphate. Each treatment was replicated 6 times. Treated soils were placed in plastic bags, covered against light and kept at 15 °C for a period of 2 weeks, by which an equilibrium was assumed to have come about. During this period the short term effects on the nematode community were assessed (Korthals *et al.*, 1996b).

After 2 weeks, portions of treated soil equivalent to 2.5 kg dry weight were thoroughly mixed with portions of fresh and untreated soil equivalent to 7.5 kg dry soil and watered to field capacity (16.5% by weight), resulting in nominal Cu and Zn concentrations of 0, 25, 50, 100, 200 and 400 mg kg<sup>-1</sup>.

Treated soils were placed in plastic 10 l pots (surface area of 452 cm<sup>2</sup>) and *Lolium perenne* L. (10 g seed per pot) was sown on half of the pots; the other half was covered

with 400 g fine gravel (heated to 120 °C to kill any nematodes present) to prevent growth of mosses and algae and to diminish evaporation. Pots were placed at random in a greenhouse ( $\pm 15$  °C) and the soil water content was maintained near field capacity. No artificial light was applied and screens were used to keep the temperature as close to 15 °C as possible during sunny spells. The grass was cut 4 cm above the soil surface each month, dried at 70 °C and weighed. Three or four clippings from each pot were combined for heavy metal analysis.

Table 1. Soil characteristics at the beginning of the experiment.

|  |      |
|--|------|
| <i>Texture</i> (% m/m on the mineral matter)               |      |
| clay (<2 $\mu\text{m}$ )                                   | 4    |
| silt (2-50 $\mu\text{m}$ )                                 | 11   |
| sand (>50 $\mu\text{m}$ )                                  | 85   |
| <i>Organic C</i> (% m/m on the dry soil)                   | 1.9  |
| <i>CEC</i> (cmol <sub>c</sub> kg <sup>-1</sup> dry weight) | 3.6  |
| <i>pH-KCl</i>  | 4.1  |
| <i>Metal content</i> (mg kg <sup>-1</sup> dry weight)      |      |
| Cd   | 0.33 |
| Cu   | 11   |
| Ni   | 4.1  |
| Zn   | 38   |

% m/m; percentage by mass

The grass was fertilized 4 times (on 15 January, 24 March, 15 June and 23 August) with 7.6 mmol KNO<sub>3</sub> and 5.8 mmol Ca(NO<sub>3</sub>)<sub>2</sub> per pot, equivalent to 60 kg N and 80 kg K<sub>2</sub>O per ha. The phosphate status of the soil precluded the need for the addition of P. At the beginning of April, another 3 g of seed was applied to 9 pots to improve the density of the stand, and 200 g gravel was added to all pots without grass to keep the soil well covered. Any emerging weeds were removed during the experiment.

#### Sampling

After 1 year each pot was sampled by taking 10 cores (diameter 17 mm) from the top 10 cm of the soil. After mixing the subsamples, 100 g was used for chemical analyses and 100 g served to extract nematodes.

#### Chemical analyses

The grass samples were digested with HF, HNO<sub>3</sub> and H<sub>2</sub>O<sub>2</sub> (Novozamsky *et al.*, 1993a). Copper and Zn concentrations in the digest were determined by flame atomic absorption spectroscopy (F-AAS) using D<sub>2</sub> background correction for Zn only.

The water content in the soil samples was determined by drying at 105 °C. Soil samples were also dried at 30 °C, sieved (2 mm) and analyzed for pH and metal concentrations by the CaCl<sub>2</sub> method (Novozamsky *et al.*, 1993b). Copper and Zn were determined by F-AAS, but solutions with low Cu concentrations (< 200  $\mu\text{g l}^{-1}$ ) were reanalysed using electrothermal AAS.

#### Nematode analyses

Nematodes were extracted from 100 g soil, using a modified Oostenbrink elutriator (Oostenbrink, 1960). The total number of nematodes was estimated by counting 2 subsamples ( $\pm 10$  % of the total sample) under a dissecting microscope. Nematode numbers were expressed per 100 g dry soil after a correction for material left on the



topsoil (mainly gravel and plant roots). Nematodes were heat-killed and fixed in 4% formalin of 80 °C and included in a permanent mass-slide. At least 150 nematodes were identified at 400x-1000x according to Bongers (1988) and allocated to feeding groups according to Yeates *et al.* (1993). To calculate the Maturity Index, the nematode taxa were assigned a *c-p* value from 1 (colonizer, *r*-strategist, tolerant to disturbances) to 5 (persister, *K*-strategist, sensitive to disturbances), according to Bongers (1990) and Bongers *et al.* (1995).

#### Data processing

Effects on grass growth were evaluated from the cumulative yield over time. This relationship appeared to be linear, with an average coefficient of determination of 0.983. The metal content in grass was log transformed before being subjected to ANOVA.

Data on nematodes were analyzed by analysis of variance. If necessary, logarithmic transformation was applied to meet assumptions of normality and homogeneity of variances. Tukey's multiple range test was employed to test for differences among treatments.

### Results

#### Soil analysis

Table 2 shows the results of soil analysis at the termination of the experiment. Averaged over all Cu and Zn treatments *L. perenne* increased pH by 0.12 ( $\pm 0.02$ ) from 4.24 to 4.36, as a consequence of nitrogen being supplied and taken up as nitrate (Dijkshoorn *et al.*, 1983). *L. perenne* decreased the Cu concentration, but had no consistent effect on the Zn concentration.

Table 2. Average pH and metal concentrations in 0.01 M CaCl<sub>2</sub> at the end of the experiment. Cu and Zn (pots without *Lolium perenne*); Cu L and Zn L (pots with *Lolium perenne*).

| Metal addition<br>(mg kg <sup>-1</sup> dry soil) | pH   |      |      |      |
|--|------|------|------|------|
|  | Cu   | Cu L | Zn   | Zn L |
| 0  | 4.28 | 4.39 | 4.28 | 4.39 |
| 25   | 4.20 | 4.38 | 4.28 | 4.39 |
| 50   | 4.26 | 4.38 | 4.29 | 4.40 |
| 100  | 4.26 | 4.29 | 4.27 | 4.41 |
| 200  | 4.18 | 4.32 | 4.29 | 4.32 |
| 400  | 4.15 | 4.25 | 4.19 | 4.41 |

|     | Cu concentration<br>(mg l <sup>-1</sup> ) |      | Zn concentration<br>(mg l <sup>-1</sup> ) |       |
|-----|---|------|---|-------|
|     | Cu  | Cu L | Zn  | Zn L  |
| 0   | 0.01                                      | 0.01 | 0.73                                      | 0.54  |
| 25  | 0.09                                      | 0.06 | 1.99                                      | 1.88  |
| 50  | 0.20                                      | 0.16 | 3.39                                      | 3.30  |
| 100 | 0.61                                      | 0.57 | 6.07                                      | 6.31  |
| 200 | 2.35                                      | 1.62 | 10.60                                     | 11.00 |
| 400 | 7.27                                      | 6.51 | 18.20                                     | 21.80 |

*Effects on grass growth and metal content*

The effects of metal treatments on *L. perenne* are summarized in figures 1 and 2. The rate of grass growth was rather low because conditions in the greenhouse (temperature, light intensity) were aimed at avoiding large fluctuations in soil temperature and water content, rather than maximizing grass growth. Copper reduced grass growth by 50 % at a metal addition of  $243 \text{ mg kg}^{-1}$  dry soil, whereas Zn reduced grass growth by 50 % at a metal addition of  $385 \text{ mg kg}^{-1}$  dry soil.

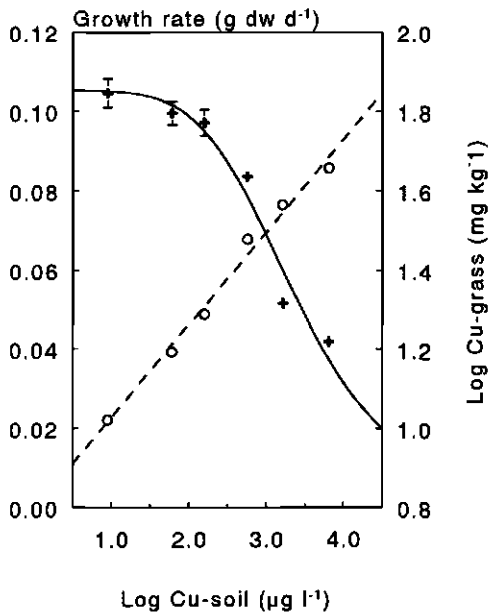


Fig. 1. Effect of Cu in the soil on the growth rate ( $\text{g dw d}^{-1}$ ) and Cu uptake ( $\text{mg kg}^{-1}$ ) of *Lolium perenne*.

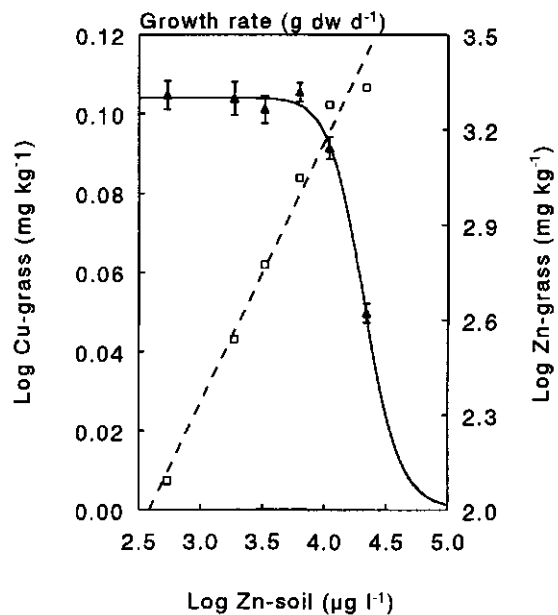


Fig. 2. Effect of Zn in the soil on the growth rate ( $\text{g dw d}^{-1}$ ) and Zn uptake ( $\text{mg kg}^{-1}$ ) of *Lolium perenne*.

Metal contents in *L. perenne* also strongly responded to the treatments. Grass from the control contained  $10.5 \text{ mg Cu}$  and  $124 \text{ mg Zn}$  per  $\text{kg DW}$ . At 50 % reduction in growth rate the Cu content had reached  $39 \text{ mg kg}^{-1} \text{ DW}$  and the Zn content  $2200 \text{ mg kg}^{-1} \text{ DW}$ . Cumulative Cu uptake reached a maximum of  $0.72 \text{ mg}$  per pot at a Cu addition rate of  $100 \text{ mg kg}^{-1}$  soil. Zinc uptake reached a maximum of  $51 \text{ mg}$  per pot when  $200 \text{ mg kg}^{-1}$  had been added to the soil. Relative to the amount of metal added, the Cu uptake reached a maximum of  $0.055 \%$  (at  $25 \text{ mg kg}^{-1}$  added). The Zn uptake reached a maximum of  $3.1 \%$  when  $100 \text{ mg kg}^{-1}$  had been added.

## ECOLOGICAL EFFECTS

*Total nematode abundance and number of taxa*

Some data on the nematode community at the start and of the control soils with or without *L. perenne* after one year are listed in Table 3. The total nematode abundance in the control soils increased significantly in the presence of *L. perenne*, but decreased

Table 3. Absolute numbers of nematodes, number of taxa, number of feeding groups, Maturity Indices and absolute numbers of individual taxa in the control treatments at the start of the experiment and after 1 year.

|                            | Control<br>t=0 weeks |        | Control, Lolium<br>t=52 weeks |        | Control, bare soil<br>t=52 weeks |        | P-value |
|----------------------------|----------------------|--------|-------------------------------|--------|----------------------------------|--------|---------|
|                            | Mean                 | SE     | Mean                          | SE     | Mean                             | SE     |         |
| N (100 g <sup>-1</sup> DW) | 2927                 | 162 b  | 7278                          | 426 c  | 1718                             | 257 a  | ***     |
| Nr of taxa                 | 18.5                 | 0.8    | 18.0                          | 1.5    | 14.7                             | 1.5    |         |
| % P                        | 28.6                 | 1.3 a  | 58.3                          | 5.7 b  | 49.1                             | 5.2 b  | ***     |
| % B                        | 58.0                 | 1.7 b  | 32.4                          | 5.3 a  | 48.8                             | 5.5 b  | ***     |
| % H                        | 9.5                  | 1.4 b  | 2.7                           | 1.7 a  | 1.2                              | 0.6 a  | **      |
| % C                        | 2.3                  | 0.5    | 3.6                           | 2.7    | 0.8                              | 0.3    |         |
| % O                        | 1.6                  | 0.4 ab | 3.0                           | 1.0 b  | 0.0                              | 0.0 a  | *       |
| % c-p 1                    | 60.0                 | 2.0 b  | 15.3                          | 4.1 a  | 9.8                              | 4.2 a  | ***     |
| % c-p 2                    | 32.8                 | 2.2 a  | 64.5                          | 10.2 b | 29.0                             | 2.0 a  | ***     |
| % c-p 3                    | 1.3                  | 0.4 a  | 1.5                           | 1.0 a  | 60.0                             | 6.1 b  | ***     |
| % c-p 4                    | 4.0                  | 0.9    | 12.0                          | 7.6    | 1.3                              | 0.4    |         |
| % c-p 5                    | 2.0                  | 0.4 a  | 6.7                           | 1.8 b  | 0.0                              | 0.0 a  | **      |
| MI                         | 1.55                 | 0.03 a | 2.30                          | 0.20 b | 2.53                             | 0.10 b | ***     |
| MI2-5                      | 2.39                 | 0.07   | 2.54                          | 0.22   | 2.69                             | 0.03   |         |
| <i>Acrobeles</i>           | 36                   | 8 a    | 509                           | 280 b  | 8                                | 6 a    | **      |
| <i>Acroboloides</i>        | 217                  | 17 b   | 618                           | 333 b  | 52                               | 12 a   | ***     |
| <i>Alaimus</i>             | 11                   | 5 a    | 52                            | 9 b    | 0                                | 0 a    | ***     |
| <i>Aphelenchoides</i>      | 125                  | 24     | 154                           | 131    | 13                               | 8      |         |
| <i>Aporcelaimellus</i>     | 41                   | 10 b   | 211                           | 78 b   | 0                                | 0 a    | ***     |
| <i>Cephalobus</i>          | 7                    | 3      | 24                            | 14     | 0                                | 0      |         |
| <i>Clarkus</i>             | 70                   | 15     | 275                           | 210    | 12                               | 4      |         |
| <i>Ditylenchus</i>         | 38                   | 10     | 8                             | 8      | 0                                | 0      |         |
| <i>Eucephalobus</i>        | 89                   | 11 b   | 481                           | 72 c   | 9                                | 2 a    | ***     |
| <i>Filenchus</i>           | 115                  | 17 a   | 371                           | 44 b   | 249                              | 51 b   | ***     |
| <i>Monhystera</i>          | 0                    | 0      | 33                            | 33     | 71                               | 40     |         |
| <i>Plectus</i>             | 43                   | 8      | 96                            | 35     | 35                               | 14     |         |
| <i>Pratylenchus</i>        | 369                  | 30 b   | 150                           | 91 a   | 473                              | 59 b   | **      |
| <i>Prismatolaimus</i>      | 27                   | 9 a    | 41                            | 30 a   | 564                              | 185 b  | *       |
| <i>Pseudhalenchus</i>      | 82                   | 22     | 36                            | 20     | 4                                | 4      |         |
| Rhabditidae                | 1233                 | 52 c   | 424                           | 51 b   | 67                               | 21 a   | ***     |
| <i>Rotylenchus</i>         | 22                   | 7 a    | 669                           | 168 b  | 11                               | 11 a   | **      |
| <i>Tylenchorhynchus</i>    | 281                  | 35 b   | 2908                          | 626 c  | 57                               | 14 a   | ***     |

Significant differences between means ( $P < 0.05$ , HSD) are indicated by different letters in the same row. MI=Maturity Index; MI2-5=Maturity Index excluding the c-p 1 value. Feeding groups are given as follows: P=plant feeding; B=bacterial feeding; H=hyphal feeding; C=carnivorous; O=omnivorous. P-values: \*=0.05>P>0.01; \*\*=0.01>P>0.001; \*\*\*=P<0.001.

significantly in soils without *L. perenne*. The average number of taxa had not changed in the presence of *L. perenne* (18), but had declined, although not significantly, to 15 in the absence of *L. perenne*. This was mainly due to the disappearance of taxa such as *Alaimus*, *Aporcelaimellus*, *Cephalobus* and *Ditylenchus* in the control with bare soil. However, certain taxa, such as *Monhystera*, were only detected in both controls at the termination of the experiment.

#### *Composition of taxa*

In comparison with the initial abundances, the abundances of most taxa were higher in the presence of *L. perenne* and lower in the absence of *L. perenne*. The increase in absolute numbers in the presence of *L. perenne* was most obvious for plant-feeding genera such as *Rotylenchus* and *Tylenchorhynchus*, but also for taxa from other feeding groups such as *Acrobeles*, *Eucephalobus*, *Alaimus*, *Aporcelaimellus* and *Clarkus*. The opposite was detected for the genera *Pratylenchus* and *Prismatolaimus*, which reached the highest abundances in bare soil.

#### *Trophic structure*

Compared to the initial proportions it was observed that, irrespective of the presence of *L. perenne*, the relative abundance of plant-feeding nematodes increased, while that of bacterial and hyphal-feeding nematodes decreased. Proportions of the less dominant carnivorous and omnivorous nematodes were, in comparison with the start, higher in the presence of *L. perenne*, but lower in bare soils.

#### *Maturity Indices*

The MI and MI2-5 had higher values at the end of the experiment, both in the presence and absence of *L. perenne*. Although the MI values in the controls with or without *L. perenne* were not significantly different, the *c-p* group distribution indicated that the increases in MI values had different causes. The MI increase for soils with *L. perenne* was mainly due to the decrease in *c-p* 1 and increase in *c-p* 2, 4 and 5. The *c-p* group distribution in soil without *L. perenne* indicated that an increase in *c-p* 3 was the main contributor to the final MI increase.

## ECOTOXICOLOGICAL EFFECTS

*Total nematode abundance and number of taxa*

The total nematode abundance declined significantly with increasing Cu or Zn concentrations (Table 4). Increasing metal concentrations lowered the average number of

Table 4. Absolute numbers of nematodes and number of taxa, relative numbers of feeding groups and *c-p* value groups (%) and Maturity Indices after 1 year exposure to Cu and Zn in the soil.

| Metal addition (mg kg <sup>-1</sup> ) | Copper Lolium (CuL) |         |         |         |         |        | P   |
|---------------------------------------|---------------------|---------|---------|---------|---------|--------|-----|
|                                       | 0                   | 25      | 50      | 100     | 200     | 400    |     |
| N (100 g <sup>-1</sup> DW)            | 7278 b              | 7336 b  | 7005 b  | 7409 b  | 3677 ab | 1519 a | **  |
| Nr of taxa                            | 18.0                | 21.3    | 19.3    | 16.3    | 14.3    | 8.3    |     |
| % P                                   | 58.3 bc             | 57.3 bc | 65.3 c  | 44.1 bc | 34.6 b  | 5.7 a  | *** |
| % B                                   | 32.4 a              | 36.3 ab | 31.5 a  | 51.5 ab | 57.1 b  | 83.0 c | *** |
| % H                                   | 2.7 ab              | 2.6 ab  | 1.8 a   | 1.4 a   | 6.1 ab  | 10.5 b | *   |
| % C                                   | 3.6                 | 2.7     | 0.9     | 3.1     | 2.2     | 0.8    |     |
| % O                                   | 3.0 b               | 1.1 ab  | 0.5 a   | 0.0 a   | 0.0 a   | 0.0 a  | **  |
| % <i>c-p</i> 1                        | 15.3                | 24.7    | 19.2    | 9.6     | 7.4     | 26.8   |     |
| % <i>c-p</i> 2                        | 64.5                | 62.2    | 72.8    | 80.5    | 92.3    | 73.0   |     |
| % <i>c-p</i> 3                        | 1.5                 | 2.9     | 4.3     | 6.7     | 0.3     | 0.2    |     |
| % <i>c-p</i> 4                        | 12.0                | 8.2     | 2.9     | 3.3     | 0.0     | 0.0    |     |
| % <i>c-p</i> 5                        | 6.7 c               | 2.0 b   | 0.8 ab  | 0.0 a   | 0.0 a   | 0.0 a  | *** |
| MI                                    | 2.30 b              | 2.01 ab | 1.93 ab | 2.04 ab | 1.93 ab | 1.73 a | *   |
| MI2-5                                 | 2.54 b              | 2.32 ab | 2.16 ab | 2.15 ab | 2.00 a  | 2.00 a | *   |
| Metal addition (mg kg <sup>-1</sup> ) | Zinc Lolium (ZnL)   |         |         |         |         |        | P   |
|                                       | 0                   | 25      | 50      | 100     | 200     | 400    |     |
| N (100 g <sup>-1</sup> DW)            | 7278 ab             | 8507 b  | 7520 ab | 8909 b  | 6534 ab | 4722 a | *   |
| Nr of taxa                            | 18.0                | 19.7    | 14.7    | 15.3    | 13.3    | 9.3    |     |
| % P                                   | 58.3 b              | 68.7 b  | 75.7 b  | 64.4 b  | 56.4 b  | 26.4 a | *** |
| % B                                   | 32.4 a              | 28.0 a  | 21.9 a  | 28.4 a  | 36.8 ab | 56.0 b | *** |
| % H                                   | 2.7 a               | 1.9 a   | 2.3 a   | 7.0 a   | 6.7 a   | 17.6 b | *** |
| % C                                   | 3.6 b               | 0.5 ab  | 0.0 a   | 0.1 ab  | 0.2 ab  | 0.0 a  | *   |
| % O                                   | 3.0 b               | 0.9 ab  | 0.1 a   | 0.0 a   | 0.0 a   | 0.0 a  | **  |
| % <i>c-p</i> 1                        | 15.3                | 23.6    | 37.1    | 18.8    | 27.5    | 24.6   |     |
| % <i>c-p</i> 2                        | 64.5                | 72.1    | 62.1    | 79.5    | 71.9    | 75.4   |     |
| % <i>c-p</i> 3                        | 1.5                 | 0.3     | 0.0     | 0.5     | 0.5     | 0.0    |     |
| % <i>c-p</i> 4                        | 12.0 b              | 2.5 ab  | 0.6 a   | 1.2 ab  | 0.0 a   | 0.0 a  | **  |
| % <i>c-p</i> 5                        | 6.7 b               | 1.6 a   | 0.2 a   | 0.0 a   | 0.0 a   | 0.0 a  | *** |
| MI                                    | 2.30 b              | 1.87 ab | 1.65 a  | 1.84 a  | 1.73 a  | 1.75 a | **  |
| MI2-5                                 | 2.54 b              | 2.14 ab | 2.03 a  | 2.04 a  | 2.01 a  | 2.00 a | **  |

Significant differences between means ( $P < 0.05$ , HSD) are indicated by different letters in the same row. MI=Maturity Index; MI2-5=Maturity Index excluding the *c-p* 1 value. Feeding groups are given as follows: P=plant feeding; B=bacterial feeding; H=hyphal feeding; C=carnivorous; O=omnivorous. *P*-values: \*=0.05> $P$ >0.01; \*\*=0.01> $P$ >0.001; \*\*\*= $P$ <0.001.

taxa to a maximum reduction of 54% and 59% for the highest Cu concentration or 48% and 37% for the highest Zn concentration, in soil with or without *L. perenne*, respectively.

Table 4 (extended)

| Metal addition (mg kg <sup>-1</sup> ) | Copper bare soil (Cu) |         |          |         |         |          | P   |
|---------------------------------------|-----------------------|---------|----------|---------|---------|----------|-----|
|                                       | 0                     | 25      | 50       | 100     | 200     | 400      |     |
| N (100 g <sup>-1</sup> DW)            | 1718 b                | 1836 b  | 1690 b   | 967 ab  | 412 a   | 144 a    | *** |
| Nr of taxa                            | 14.7                  | 13.3    | 13.0     | 13.0    | 9.0     | 6.0      |     |
| % P                                   | 49.1 b                | 47.3 b  | 41.7 b   | 49.6 b  | 42.0 b  | 8.6 a    | *** |
| % B                                   | 48.8 a                | 50.0 ab | 55.0 ab  | 45.8 a  | 51.2 ab | 68.6 b   | *   |
| % H                                   | 1.2 a                 | 2.1 ab  | 2.9 abc  | 4.4 bc  | 6.8 c   | 22.8 d   | *** |
| % C                                   | 0.8                   | 0.5     | 0.3      | 0.2     | 0.0     | 0.0      |     |
| % O                                   | 0.0                   | 0.1     | 0.0      | 0.0     | 0.0     | 0.0      |     |
| % c-p 1                               | 9.8                   | 13.4    | 12.9     | 18.8    | 29.0    | 18.4     |     |
| % c-p 2                               | 29.0 a                | 26.5 a  | 20.1 a   | 35.8 ab | 64.3 bc | 81.6 c   | *** |
| % c-p 3                               | 60.0 c                | 59.1 c  | 66.4 c   | 45.4 bc | 6.7 ab  | 0.0 a    | *** |
| % c-p 4                               | 1.3                   | 1.0     | 0.6      | 0.0     | 0.0     | 0.0      |     |
| % c-p 5                               | 0.0                   | 0.0     | 0.0      | 0.0     | 0.0     | 0.0      |     |
| MI                                    | 2.53 b                | 2.48 b  | 2.55 b   | 2.27 ab | 1.78 a  | 1.82 a   | **  |
| MI2-5                                 | 2.69 c                | 2.70 c  | 2.78 c   | 2.52 bc | 2.10 ab | 2.00 a   | *** |
|                                       | Zinc bare soil (Zn)   |         |          |         |         |          | P   |
|                                       | 0                     | 25      | 50       | 100     | 200     | 400      |     |
| N (100 g <sup>-1</sup> DW)            | 1718 d                | 1476 cd | 1000 bc  | 658 ab  | 524 ab  | 336 a    | *** |
| Nr of taxa                            | 14.7                  | 15.7    | 17.3     | 13.3    | 12.7    | 9.3      |     |
| % P                                   | 49.1 ab               | 64.3 bc | 73.4 c   | 66.9 bc | 69.4 bc | 31.6 a   | *** |
| % B                                   | 48.8 b                | 30.5 ab | 20.4 a   | 25.7 a  | 25.7 a  | 46.0 b   | *** |
| % H                                   | 1.2 a                 | 3.8 ab  | 3.3 ab   | 7.2 bc  | 4.7 abc | 22.4 c   | *** |
| % C                                   | 0.8 b                 | 0.4 ab  | 1.8 c    | 0.0 a   | 0.0 a   | 0.0 a    | *** |
| % O                                   | 0.0                   | 0.1     | 0.1      | 0.0     | 0.0     | 0.0      |     |
| % c-p 1                               | 9.8 a                 | 20.8 ab | 23.7 ab  | 28.2 ab | 34.2 b  | 23.3 ab  | *   |
| % c-p 2                               | 29.0 a                | 39.3 ab | 43.4 ab  | 69.8 c  | 64.3 bc | 76.7 c   | *** |
| % c-p 3                               | 60.0 c                | 36.5 bc | 23.4 ab  | 1.7 a   | 0.9 a   | 0.0 a    | *** |
| % c-p 4                               | 1.3 a                 | 3.4 a   | 9.5 b    | 0.3 a   | 0.6 a   | 0.0 a    | *** |
| % c-p 5                               | 0.0                   | 0.0     | 0.0      | 0.0     | 0.0     | 0.0      |     |
| MI                                    | 2.53 d                | 2.22 cd | 2.19 bcd | 1.74 ab | 1.68 a  | 1.77 abc | *** |
| MI2-5                                 | 2.69 b                | 2.52 b  | 2.56 b   | 2.03 a  | 2.03 a  | 2.00 a   | *** |

Table 5. Mean numbers of nematode taxa after 1 year (numbers 100 g<sup>-1</sup> dry soil).

|                                       |     |   | Copper Lolium (CuL) |        |        |        |        |        | P   |
|---------------------------------------|-----|---|---------------------|--------|--------|--------|--------|--------|-----|
| Metal addition (mg kg <sup>-1</sup> ) |     |   | 0                   | 25     | 50     | 100    | 200    | 400    |     |
| Taxon                                 | c-p | T |                     |        |        |        |        |        |     |
| <i>Acrobeles</i>                      | 2   | B | 509 b               | 106 ab | 49 a   | 0 a    | 0 a    | 0 a    | *** |
| <i>Acrobeloides</i>                   | 2   | B | 618                 | 589    | 897    | 825    | 1251   | 879    |     |
| <i>Alaimus</i>                        | 4   | B | 52 b                | 28 ab  | 0 a    | 0 a    | 0 a    | 0 a    | **  |
| <i>Anaplectus</i>                     | 2   | B | 41                  | 17     | 0      | 0      | 0      | 0      |     |
| <i>Aphelenchoides</i>                 | 2   | H | 154                 | 70     | 72     | 51     | 134    | 133    |     |
| <i>Aporcelaimellus</i>                | 5   | O | 211 c               | 29 bc  | 19 ab  | 0 a    | 0 a    | 0 a    | *** |
| <i>Cephalobus</i>                     | 2   | B | 24 a                | 7 a    | 39 a   | 336 c  | 219 b  | 0 a    | *** |
| <i>Clarkus</i>                        | 4   | C | 275                 | 194    | 68     | 126    | 0      | 0      |     |
| <i>Ditylenchus</i>                    | 2   | H | 8                   | 61     | 38     | 0      | 25     | 13     |     |
| <i>Driiocephalobus</i>                | 2   | B | 23                  | 19     | 26     | 0      | 0      | 0      |     |
| <i>Eucephalobus</i>                   | 2   | B | 481 b               | 732 b  | 525 b  | 1732 b | 245 b  | 6 a    | *** |
| <i>Filenchus</i>                      |     | P | 371                 | 314    | 405    | 202    | 23     | 18     |     |
| <i>Monhystera</i>                     | 2   | B | 33                  | 76     | 0      | 0      | 0      | 0      |     |
| <i>Plectus</i>                        | 2   | B | 96 ab               | 49 ab  | 26 ab  | 173 b  | 91 ab  | 0 a    | *   |
| <i>Pratylenchus</i>                   |     | P | 150                 | 200    | 183    | 315    | 131    | 46     |     |
| <i>Prismatolaimus</i>                 | 3   | B | 41                  | 87     | 125    | 250    | 6      | 2      |     |
| <i>Pseudhalenchus</i>                 | 2   | F | 36                  | 40     | 21     | 62     | 29     | 29     |     |
| Rhabditidae                           | 1   | B | 424 b               | 596 b  | 173 ab | 223 ab | 57 a   | 344 ab | *   |
| <i>Rotylenchus</i>                    |     | P | 669 ab              | 696 ab | 1002 b | 1098 b | 524 ab | 11 a   | *   |
| <i>Seinura</i>                        | 2   | H | 0                   | 7      | 0      | 97     | 68     | 10     |     |
| <i>Trichodorus</i>                    |     | P | 76                  | 78     | 0      | 0      | 0      | 0      |     |
| <i>Tylenchorhynchus</i>               |     | P | 2908 b              | 2892 b | 2868 b | 1580 b | 745 b  | 3 a    | *** |
| <i>Tylenchus</i>                      |     | P | 44                  | 40     | 53     | 38     | 0      | 0      |     |

|                         |   |   | Zinc Lolium (ZnL) |        |        |         |        |        | P   |
|-------------------------|---|---|-------------------|--------|--------|---------|--------|--------|-----|
|                         |   |   | 0                 | 25     | 50     | 100     | 200    | 400    |     |
| <i>Acrobeles</i>        | 2 | B | 509 b             | 38 a   | 0 a    | 0 a     | 0 a    | 0 a    | *** |
| <i>Acrobeloides</i>     | 2 | B | 618               | 570    | 396    | 1290    | 1359   | 1775   |     |
| <i>Alaimus</i>          | 4 | B | 52 b              | 5 ab   | 10 ab  | 34 ab   | 0 a    | 0 a    | *   |
| <i>Anaplectus</i>       | 2 | B | 41                | 28     | 0      | 0       | 0      | 0      |     |
| <i>Aphelenchoides</i>   | 2 | H | 154               | 73     | 29     | 356     | 197    | 756    |     |
| <i>Aporcelaimellus</i>  | 5 | O | 211 b             | 56 ab  | 5 a    | 0 a     | 0 a    | 0 a    | **  |
| <i>Cephalobus</i>       | 2 | B | 24                | 28     | 39     | 17      | 4      | 0      |     |
| <i>Clarkus</i>          | 4 | C | 275               | 45     | 0      | 0       | 0      | 0      |     |
| <i>Ditylenchus</i>      | 2 | H | 8                 | 67     | 46     | 125     | 49     | 0      |     |
| <i>Driiocephalobus</i>  | 2 | B | 23                | 60     | 20     | 11      | 0      | 0      |     |
| <i>Eucephalobus</i>     | 2 | B | 481               | 815    | 479    | 599     | 225    | 0      |     |
| <i>Filenchus</i>        |   | P | 371 b             | 221 ab | 147 ab | 143 ab  | 154 ab | 43 a   | *   |
| <i>Monhystera</i>       | 2 | B | 33                | 22     | 4      | 0       | 0      | 0      |     |
| <i>Plectus</i>          | 2 | B | 96 b              | 11 a   | 0 a    | 0 a     | 0 a    | 0 a    | **  |
| <i>Pratylenchus</i>     |   | P | 150 ab            | 217 ab | 72 a   | 437 bc  | 697 c  | 225 ab | *** |
| <i>Prismatolaimus</i>   | 3 | B | 41                | 11     | 0      | 0       | 8      | 0      |     |
| <i>Pseudhalenchus</i>   | 2 | F | 36                | 33     | 95     | 132     | 187    | 76     |     |
| Rhabditidae             | 1 | B | 424               | 623    | 591    | 414     | 855    | 566    |     |
| <i>Rotylenchus</i>      |   | P | 669               | 448    | 1277   | 2072    | 1800   | 929    |     |
| <i>Seinura</i>          | 2 | H | 0                 | 0      | 0      | 4       | 8      | 0      |     |
| <i>Trichodorus</i>      |   | P | 76                | 0      | 0      | 0       | 0      | 0      |     |
| <i>Tylenchorhynchus</i> |   | P | 2908 ab           | 4806 b | 4158 b | 2998 ab | 825 a  | 0 a    | **  |
| <i>Tylenchus</i>        |   | P | 44                | 0      | 0      | 0       | 0      | 0      |     |

Significant differences between mean abundances of one taxon within the same experiment are indicated by different letters ( $P < 0.05$ , HSD) in the same row. c-p value and T (feeding group) are given. P=plant feeding; B=bacterial feeding; H=hyphal feeding; C=carnivorous; O=omnivorous. Differences between treatment means are given by: \*= $0.05 > P > 0.01$ ; \*\*= $0.01 > P > 0.001$ ; \*\*\*= $P < 0.001$ .

## Influence of ryegrass on a nematode community

Table 5 (extended)

| Metal addition (mg kg <sup>-1</sup> ) |     |   | Copper bare soil (Cu) |        |        |        |        |      | P   |
|---------------------------------------|-----|---|-----------------------|--------|--------|--------|--------|------|-----|
|                                       |     |   | 0                     | 25     | 50     | 100    | 200    | 400  |     |
| Taxon                                 | c-p | T |                       |        |        |        |        |      |     |
| <i>Acrobeles</i>                      | 2   | B | 8                     | 0      | 0      | 0      | 0      | 0    |     |
| <i>Acrobeloides</i>                   | 2   | B | 52                    | 101    | 86     | 88     | 119    | 69   |     |
| <i>Alaimus</i>                        | 4   | B | 0                     | 0      | 0      | 0      | 0      | 0    |     |
| <i>Anaplectus</i>                     | 2   | B | 0                     | 0      | 0      | 0      | 0      | 0    |     |
| <i>Aphelenchoides</i>                 | 2   | H | 13                    | 0      | 8      | 10     | 16     | 28   |     |
| <i>Aporcelaimellus</i>                | 5   | O | 0                     | 0      | 0      | 0      | 0      | 0    |     |
| <i>Cephalobus</i>                     | 2   | B | 0                     | 0      | 0      | 1      | 1      | 0    |     |
| <i>Clarkus</i>                        | 4   | C | 12                    | 8      | 6      | 0      | 0      | 0    |     |
| <i>Ditylenchus</i>                    | 2   | H | 0 a                   | 21 b   | 15 b   | 19 b   | 2 a    | 2 a  | *** |
| <i>Driiocephalobus</i>                | 2   | B | 56                    | 58     | 23     | 0      | 0      | 0    |     |
| <i>Eucephalobus</i>                   | 2   | B | 9                     | 4      | 14     | 1      | 0      | 0    |     |
| <i>Filenchus</i>                      |     | P | 249 abc               | 364 c  | 284 bc | 97 ab  | 5 a    | 0 a  | **  |
| <i>Monhystera</i>                     | 2   | B | 71 c                  | 32 bc  | 11 b   | 0 a    | 0 a    | 0 a  | *** |
| <i>Plectus</i>                        | 2   | B | 35                    | 16     | 9      | 16     | 0      | 0    |     |
| <i>Pratylenchus</i>                   |     | P | 473 c                 | 457 c  | 390 bc | 352 bc | 165 ab | 12 a | *** |
| <i>Prismatolaimus</i>                 | 3   | B | 564 b                 | 590 b  | 647 b  | 264 ab | 19 a   | 0 a  | **  |
| <i>Pseudhalenchus</i>                 | 2   | F | 4                     | 18     | 21     | 12     | 11     | 3    |     |
| <i>Rhabditidae</i>                    | 1   | B | 67 ab                 | 103 ab | 105 b  | 71 ab  | 57 ab  | 28 a | *   |
| <i>Rotylenchus</i>                    |     | P | 11                    | 5      | 6      | 8      | 2      | 0    |     |
| <i>Seinura</i>                        | 2   | H | 4                     | 3      | 0      | 2      | 0      | 0    |     |
| <i>Trichodorus</i>                    |     | P | 0                     | 0      | 0      | 0      | 0      | 0    |     |
| <i>Tylenchorhynchus</i>               |     | P | 57 b                  | 31 ab  | 36 ab  | 11 ab  | 2 a    | 0 a  | *   |
| <i>Tylenchus</i>                      |     | P | 15                    | 0      | 0      | 4      | 0      | 0    |     |

|                         |   |   | Zinc bare soil (Zn) |        |        |        |         |       | P   |
|-------------------------|---|---|---------------------|--------|--------|--------|---------|-------|-----|
|                         |   |   | 0                   | 25     | 50     | 100    | 200     | 400   |     |
| <i>Acrobeles</i>        | 2 | B | 8                   | 2      | 0      | 0      | 0       | 0     |     |
| <i>Acrobeloides</i>     | 2 | B | 52                  | 69     | 32     | 72     | 72      | 95    |     |
| <i>Alaimus</i>          | 4 | B | 0                   | 10     | 6      | 0      | 1       | 0     |     |
| <i>Anaplectus</i>       | 2 | B | 0                   | 0      | 0      | 0      | 0       | 0     |     |
| <i>Aphelenchoides</i>   | 2 | H | 13                  | 7      | 4      | 11     | 5       | 71    |     |
| <i>Aporcelaimellus</i>  | 5 | O | 0                   | 0      | 0      | 0      | 0       | 0     |     |
| <i>Cephalobus</i>       | 2 | B | 0                   | 0      | 0      | 0      | 0       | 0     |     |
| <i>Clarkus</i>          | 4 | C | 12 ab               | 6 ab   | 17 c   | 0 a    | 0 a     | 0 a   | **  |
| <i>Ditylenchus</i>      | 2 | H | 0 a                 | 20 ab  | 24 ab  | 29 b   | 17 ab   | 10 ab | *   |
| <i>Driiocephalobus</i>  | 2 | B | 56 ab               | 43 b   | 39 b   | 26 b   | 0 a     | 0 a   | **  |
| <i>Eucephalobus</i>     | 2 | B | 9                   | 6      | 2      | 0      | 0       | 0     |     |
| <i>Filenchus</i>        |   | P | 249 bc              | 309 c  | 248 bc | 94 ab  | 150 abc | 23 a  | **  |
| <i>Monhystera</i>       | 2 | B | 71                  | 0      | 0      | 0      | 0       | 0     |     |
| <i>Plectus</i>          | 2 | B | 35 b                | 23 ab  | 3 ab   | 0 a    | 0 a     | 0 a   | *   |
| <i>Pratylenchus</i>     |   | P | 473 cd              | 568 d  | 457 cd | 315 bc | 197 ab  | 60 a  | *** |
| <i>Prismatolaimus</i>   | 3 | B | 564 c               | 188 bc | 47 b   | 1 a    | 0 a     | 0 a   | *** |
| <i>Pseudhalenchus</i>   | 2 | F | 4                   | 16     | 4      | 6      | 1       | 5     |     |
| <i>Rhabditidae</i>      | 1 | B | 67                  | 104    | 59     | 57     | 53      | 51    |     |
| <i>Rotylenchus</i>      |   | P | 11                  | 13     | 5      | 17     | 7       | 8     |     |
| <i>Seinura</i>          | 2 | H | 4                   | 0      | 1      | 0      | 0       | 0     |     |
| <i>Trichodorus</i>      |   | P | 0                   | 0      | 0      | 0      | 0       | 0     |     |
| <i>Tylenchorhynchus</i> |   | P | 57 c                | 62 bc  | 26 abc | 8 abc  | 3 ab    | 1 a   | **  |
| <i>Tylenchus</i>        |   | P | 15                  | 0      | 0      | 0      | 0       | 0     |     |



*Trophic structure*

Increasing Cu and Zn concentrations caused a gradual decrease in the proportion of plant-feeding nematodes and an increase in that of the hyphal-feeding and to a lesser extent bacterial-feeding nematodes (Table 4). The decrease in plant-feeding nematodes was most extreme in **CuL**, whereas the relative increase was greatest for the hyphal-feeding nematodes in **Cu** and **Zn**. Intermediate Zn concentrations (50-200 mg kg<sup>-1</sup>) tended to increase the proportion of plant feeders and to decrease the proportion of bacterial feeders. In all Cu and Zn treatments, the proportion of omnivorous and carnivorous nematodes declined, that of the omnivorous nematodes the most.

*Maturity index*

Irrespective of the presence of *L. perenne*, the Maturity Index and MI2-5 declined with increasing Cu and Zn additions (Table 4). Maturity Index values at concentrations of 400 (**CuL**), 200 (**Cu**), 50 (**ZnL**) or 100 (**Zn**) mg kg<sup>-1</sup> were significantly lower than in the control. The decline in MI values was mainly affected by the increase in the proportion of taxa with *c-p* value 1 or 2 (i.e. *Acrobeloides*, *Aphelenchoides* and *Rhabditidae*), while that of most other taxa declined. From these taxa, the decline in *Prismatolaimus* in **Cu** and **Zn** was most obvious and had the largest impact on the final MI value.

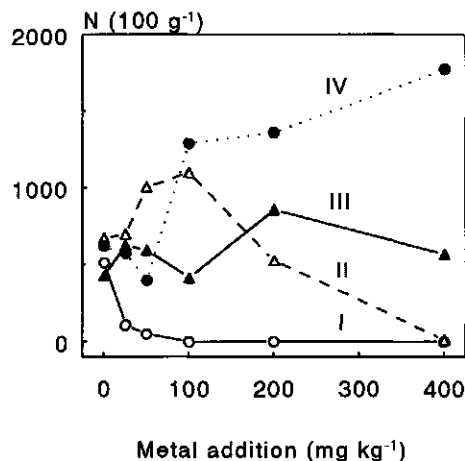
*Composition of taxa*

Fig. 3. Change in population numbers of 4 taxa to increasing concentrations of Cu (open symbols) and Zn (closed symbols) in the presence of *Lolium perenne*. (○-) *Acrobeloides*; (△-) *Rotylenchus*; (▲-) *Rhabditidae*; (●-) *Acrobeloides*.

Irrespective of the presence of *L. perenne*, most taxa had declining population numbers with increasing metal concentrations (Table 5). This response (response type I Fig. 3) is most common from a toxicological point of view. However, certain taxa responded differently to the metal additions in one or more of the treatments. The absolute numbers of some taxa were negatively affected by increasing Cu and Zn additions in the absence of *L. perenne*, but were increased at one or more intermediate concentrations in pots with *L. perenne* (response type II Fig. 3). A type II response was found for *Rotylenchus* and *Seinura* in both the **CuL** and **ZnL** experiments, for *Tylenchorhynchus* and *Pratylenchus* in the **ZnL** and for *Cephalobus*, *Prismatolaimus* and *Eucephalobus* in the **CuL**. Some taxa were hardly affected (response type III Fig. 3) or tended to increase at intermediate or high Cu and Zn concentrations (response type IV Fig. 3). Response types III and IV were found for *Pseudhalenchus*, *Ditylenchus*, Rhabditidae, *Acrobeloides* and *Aphelenchoides*.

## Discussion

### ECOLOGICAL EFFECTS

The nematode community at the start of the study was comparable with that in other arable agroecosystems (Wasilewska, 1989; Weiss and Larink, 1991; Freckman and Ettema, 1993). Based on the high number of bacterivorous nematodes with a *c-p* value 1, observed at the start and the significant decline in both control soils at the termination of the experiment we assume that initially the soil was disturbed, probably because the soil was collected shortly after harvest, and that during the experiment the nematode communities followed trends previously observed after disturbance and subsequent recovery (Ettema and Bongers, 1993; De Goede *et al.*, 1993). However, the presence or absence of *L. perenne*, did influence this process of secondary succession.

Total nematode numbers in control soils with *L. perenne* increased, which was most pronounced for some obligate plant-feeding taxa (*Rotylenchus* and *Tylenchorhynchus*), as well as for a few omnivorous and carnivorous taxa. This follows the observation that the quantity and quality of plants, in combination with the addition of plant nutrients can increase the population size of not only plant-feeding taxa, but also taxa from other feeding groups (Yeates, 1987).

In contrast, the total number of nematodes, the average number of omnivorous and carnivorous nematodes and taxa from *c-p* groups 4 and 5 had declined in soils without *L.*

## Chapter 3

*perenne*. These changes indicate unnatural conditions in the bare soils, causing a steady decline in food-availability (indicated by a significant decrease in the number of Rhabditidae). Unnatural conditions may also have been caused by covering bare soil with gravel. Although not quantified, the absence of plant roots in combination with the gravel resulted in a more compact soil with a higher, less variable soil moisture content, all important "driving" factors for nematode communities (c.f. Yeates, 1981).

There were very few taxa not affected (*Filenchus*) or which benefitted (*Pratylenchus* and *Prismatolaimus*) from the conditions in the bare control soils. For *Prismatolaimus*, a bacterial feeder preferring wet conditions, this is less surprising than for the other two taxa, which are both plant feeding taxa. No living roots were present in these soils, but it is possible that in comparison to the initial populations, *Pratylenchus* could persist, or in the case of *Filenchus* even increase, since this taxon can probably feed on fungi as well.

### ECOTOXICOLOGICAL EFFECTS

The purpose of this experiment was to compare the effects of Cu and Zn on a nematode community in bare soil and soil with *L. perenne*. At the community level, Cu and Zn reduced the total nematode abundance, average number of taxa, proportion of plant feeding nematodes, proportion of omnivorous and carnivorous nematodes and the Maturity Index. In general these findings are in agreement with results obtained earlier in field studies (Zullini and Peretti, 1986; Weiss and Larink, 1991; Popovici and Korthals, 1995; Korthals et al., 1996a).

At the population level, most taxa showed the expected response to both metals, i.e. numbers declined with increasing metal concentrations. However, at intermediate Cu or Zn concentrations (25-200 mg kg<sup>-1</sup>), certain taxa appeared to be stimulated (response type II, Fig. 3) in absolute abundances. Based on short-term exposure periods, Cu and Zn ions may have stimulated the hatching of nematode eggs (Clarke and Shepherd, 1966). However, since our data were obtained after a period of one year, the results are more likely due to less food competition and/or predation experienced by the tolerant taxa (Hendrix and Parmelee, 1985). This kind of results form an example of indirect effects, which to our opinion may be expected more often when exposing whole communities to pollutants under realistic conditions.

By comparing the experiments done in bare soil with those in the presence of *L. perenne*, it became clear that many of the observed effects occurred at higher metal addition rates, were less extreme and may have been caused in an indirect way. One possible explanation is that *L. perenne* and fertilizers enriched the soil, which caused not

only 'ecological effects' but interfered with the ecotoxicological effects of Cu or Zn as well. The observation that the presence of vegetation reduced the 'stress' to the nematode community, agrees with the view that quantity and quality of food can influence the response of individual species as well as whole communities to pollutants (Vanni and Lampert, 1992; Norberg-King and Schmidt, 1993; Barreiro Lozano and Pratt, 1994).

One other possible explanation is that *L. perenne* may have influenced metal bioavailability. Based on available metal concentrations measured with the  $\text{CaCl}_2$  method, the nematodes were exposed to fairly similar concentrations in the presence or absence of the grass. These concentrations, however, characterized the bulk soil and not the rhizosphere soil, where the grass may have effected a larger increase in soil pH due to excess anion uptake when supplied with nitrate as nitrogen source. The influence of nitrate or ammonium uptake on soil pH and metal bioavailability has been observed earlier (Dijkshoorn *et al.*, 1979). Compared with the experiment by Dijkshoorn *et al.* (1979), in which a very similar soil was used, metal toxicity to *L. perenne* appeared much lower in our experiment. For example, the metal addition rates associated with a 50 % reduction in yield were approximately 2.4 times higher in our case. The slower rate of growth due to less favourable growing conditions is likely to be responsible for this apparent increase in metal tolerance. The demands placed on the nutrient and water uptake function of the roots and, consequently, the effects of root malfunctioning will be stronger in vigorously growing plants than in plants with a limited light supply.

Metal uptake by *L. perenne* could also have contributed to lower metal availability. The total amount of metal removed with the grass clippings was small, but the effect of metal binding to and in the roots was not accounted for. As both the increase in pH and metal uptake may be expected to have their largest effect on bioavailability in the rhizosphere, where many nematode taxa aggregate, they could have affected the present results.

The effects of Cu and Zn on *L. perenne* itself may have influenced the quantity and quality of the food source for several nematode taxa, especially those which are completely dependent on plants. This is not only negative, since, especially with intermediate pollutant concentrations, it may also be beneficial to herbivores or plant-associated organisms, for example due to an increase in N-availability, a higher leakage of root exudates or a breakdown of the plant's defence strategy (White, 1984). This may play a role in the increased abundances of *Tylenchorhynchus* at intermediate Zn concentrations in **ZnL**.

The present study demonstrated that a field collected nematode community changes due to biotic (presence of *L. perenne*) as well as abiotic (heavy metals) factors. It is clear that the presence of vegetation is a very important factor in determining the final

ecotoxicological effects of Cu and Zn. In soils covered with *L. perenne* it was found that the effects of Cu and Zn became apparent at higher metal concentrations, were less severe and were more often caused in an indirect way. The present data were based on a monoculture of *L. perenne* during one year. The effects of pollution on nematode communities in soil with a more diverse vegetation can be expected to be even more complex. Therefore, it is recommended that future risk assessment studies are based on experimental methods using increased ecological complexity. To expose natural nematode communities in micro- or mesocosms in the presence of vegetation seems necessary.

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## Chapter 4

### JOINT TOXICITY OF COPPER AND ZINC TO A TERRESTRIAL NEMATODE COMMUNITY IN AN ACID SANDY SOIL

#### Abstract

Heavy metal toxicity to an indigenous nematode community was examined following the addition of Cu and Zn, alone or in combination, to agricultural soil. The dissolved Cu or Zn concentrations measured after equilibrating soil samples with a 0.01 M solution of  $\text{CaCl}_2$  showed that the metal concentrations found in soils with combined metal additions were not significantly different from those with single metal additions. After an exposure period of six months, many nematode community parameters and individual nematode taxa were significantly affected by increasing concentrations of Cu and Zn up to  $200 \text{ mg kg}^{-1}$ . Some nematode taxa, such as *Thonus*, *Alaimus* and *Aporcelaimellus* were very sensitive and disappeared at Cu and Zn concentrations exceeding  $50 \text{ mg kg}^{-1}$ . For several nematode community parameters and nematode taxa,  $\text{EC}_{50}$  values for single metal exposures were used to calculate  $\text{TU}_{50}$  values for the joint toxicity of Cu and Zn. Based on these calculations, it is concluded that the effects of a combined exposure to Cu and Zn were additive or less than additive. Before this conclusion can be generalized, however, more data are needed on other types of soil, other pH values and other combinations of pollutants.

**Keywords:** Nematodes; Copper; Zinc; Joint toxicity; Bioavailability

#### Introduction

An important aspect in the protection of our environment is the definition of soil quality criteria. There is a strong tradition in the Netherlands to relate these criteria with ecotoxicological risk assessment. For example, the method applied in the derivation of limit values accounts for all available No Observed Effect Concentrations (NOEC) on life-history traits (growth, reproduction, mortality) of different species (Van Straalen and Denneman, 1989; Denneman and Van Gestel, 1990). To date, these limit values have been divided by a safety factor of 100 in order to define target values (Anon., 1989; Anon., 1991).

One of the arguments for the use of this safety factor was the uncertainty of the obtained limit values. Can the often very limited number of NOEC's, obtained for different organisms exposed to a single pollutant, indeed reflect the potential risk of that particular pollutant to an entire ecosystem? An even more important argument for the use of this safety factor is the simultaneous presence of several pollutants, which is the rule rather than the exception in the field. Knowledge on joint toxicity of pollutants, especially with respect to terrestrial animals, is still poor (Eifac, 1987; Hensbergen and Van Gestel, 1995).

There is general agreement that the simultaneous presence of several pollutants, although each may be below their specific limit concentrations, may still imply a risk. In the most simple case, the joint toxicity of two pollutants is additive. Even then, methods to estimate the potential risks of mixtures are only in an early stage of development (Ikeda, 1994). Besides this simple case, there are several studies indicating that the joint toxicity of pollutants can be larger (Sprague and Ramsay, 1965; Babich *et al.*, 1986; Spehar and Fiandt, 1986; Asztalos *et al.*, 1988; Kraak *et al.*, 1994) or smaller (Spehar *et al.*, 1978; Babich *et al.*, 1986; Vranken *et al.*, 1988; Kraak *et al.*, 1993) than predicted on the basis of addition of effects of single exposures. This makes standards based on data obtained from single pollutant dose-response relationships unreliable.

This holds more strongly in soils where pollutants interact with the soil matrix. Pollutants such as heavy metals may compete for binding sites in the soil. Where two or more metals are present in the soil, competition may lead to different concentrations in solution than expected from the binding behaviour of the individual metals (Van Riemsdijk and Hiemstra, 1993).

This article describes the joint toxicity of Cu and Zn to an indigenous nematode community present in agricultural soil. The phylum Nematoda is comprised of many species showing a high diversity of life cycles, feeding types and sensitivities to pollutants. Since terrestrial nematodes live in the soil pore water, they are assumed to be exposed to the pollutant concentration in the soil solution, which offers good perspectives to assess effects of pollutants in relation to their bioavailability in soil. Furthermore, by studying effects on the nematode community, one has the opportunity to examine the joint toxic effects on various parameters simultaneously, from a low (population) to a high (community) level of biological organization.

## Materials and methods

### *Experimental design*

In October 1993 soil was collected from the top 10 cm of an arable field on sandy soil located 3 km NNE of Wageningen, the Netherlands. Fresh soil was passed through a 9 mm sieve to remove stones, stubble and coarse roots. After mixing, samples were taken, dried (30°C) and sieved (2 mm) to determine some soil characteristics. The soil used in this experiment was a loamy sand (4% clay, 11% silt, 85% sand) with an organic carbon content of 1.9% by mass. The initial pH, measured in 1 M KCl, was 4.1 and the actual CEC amounted to 3.6 cmol<sub>c</sub> kg<sup>-1</sup> (unbuffered BaCl<sub>2</sub>). Initial Cu and Zn contents, as determined following digestion with a mixture of concentrated nitric and sulphuric acid, were 11 and 38 mg kg<sup>-1</sup>, respectively. For more details on site and soil see Korthals *et al.* (1996a).



The fresh soil was allowed to dry to a water content of 10.6% by weight, so as to allow for metal solutions to be added without exceeding the field capacity of the soil. Copper and zinc (sulphates) were added to the soil in two steps, in order to increase the survival chances of nematode species.

First, eight concentrations of Cu (0, 100, 140, 200, 280, 400, 560 and 800 mg kg<sup>-1</sup> dry weight) were combined with five concentrations of Zn (0, 100, 200, 400 and 800 mg kg<sup>-1</sup>) by mixing 0-200 ml stock solutions with 3.3 kg moist soil (equivalent to 3 kg dry soil). Each combination was replicated three times. Differences in water and sulphate additions were balanced between the treatments by adding demineralized water and calcium sulphate. The treated soil was thoroughly mixed by hand, placed in polythene bags and kept at 15°C in the dark for a period of 3 weeks, after which an equilibrium between added metals and the soil was assumed.

Following this treatment, portions of treated soil (2 kg dry weight) were thoroughly mixed with portions of untreated fresh soil equivalent to 6 kg dry soil and brought to field capacity (17.7% by weight). Thus, the final metal concentration ranges were lowered by a factor 4. Plastic 7.5 l pots were filled with treated soil and covered with 800 g fine gravel (heated to 120°C to kill any nematodes present). The pots were placed at random in a greenhouse at 15°C, watered to field capacity and kept free of weeds.

### Sampling

After 6 months 10 soil cores (diameter 17 mm) were taken from the top 10 cm of the soil from each pot. After mixing the subsamples, 100 g was used for chemical analyses and 100 g served to sample nematodes.

### Chemical analyses

The soil samples were dried at 40 °C for 24 h and sieved through a 2 mm mesh size. Soil pH was determined in suspension according to Novozamsky *et al.*, (1993) after shaking soil samples end-over-end for 24 h with 0.01 M CaCl<sub>2</sub> in a soil to solution ratio of 1:10. A sample was taken from the supernatant obtained by centrifugation at 5000 rev min<sup>-1</sup> for 10 min. and acidified by adding 1 % (by volume) of concentrated HNO<sub>3</sub>, to prevent the adsorption of heavy metals during storage (Houba *et al.*, 1993). Copper and Zn concentrations were then determined by flame atomic absorption spectroscopy (F-AAS) with Smith-Hieftje background correction. Low Cu concentrations (< 200 mg l<sup>-1</sup>) were re-analyzed using electrothermal AAS.

### Nematode analyses

Nematodes were extracted from 100 g fresh soil, using a modified Oostenbrink elutriator (Oostenbrink, 1960). The total number of nematodes was estimated by counting 2 subsamples ( $\pm 10\%$  of the total sample) under a dissecting microscope. Nematode numbers were expressed per 100 g dry soil after a correction for material left on the top sieve (mainly gravel and plant roots). Nematodes were heat-killed and fixed with formalin (90°C, 4%) and placed on a permanent mass-slide. At least 150 nematodes were identified at 400x-1000x according to Bongers (1988) and allocated to feeding groups according to Yeates *et al.* (1993) and allocated to c-p value groups according to Bongers (1990) and Bongers *et al.* (1995) in order to calculate Maturity Index values.

### Data processing

Data on nematodes were analyzed by analysis of variance (Sokal and Rohlf, 1981). If necessary, logarithmic transformations were applied to meet assumptions of normality and homogeneity of variances.  $EC_{50}$  values for Cu and Zn were estimated by using non-linear regression estimation (Bruce and Versteeg, 1992) of metal concentrations against several nematode community parameters from the single metal exposure data. The concentrations of all Cu and Zn combinations were expressed in Toxic Units (TU) by dividing the sum of the individual Cu or Zn concentrations by their specific  $EC_{50}$  obtained for the single metal exposure series (Sprague, 1970; Könemann, 1981). For several nematode parameters, the exposure-response relationship (in Toxic Units) was used to estimate where a parameter was lowered by 50%, hereafter referred as  $TU_{50}$  value.  $TU_{50}$  values were used to estimate whether the effects of a combined exposure to Cu and Zn were additive ( $TU_{50} = 1$  TU), less than additive ( $TU_{50} > 1$  TU) or more than additive ( $TU_{50} < 1$  TU).

## Results

### Chemical analysis

After six months, at the end of the experiment, the pH of the soil suspended in 0.01 M  $CaCl_2$  was 4.32, with a standard deviation of 0.13. Differences between treatments were not significant. The Cu and Zn concentrations in 0.01 M  $CaCl_2$  are shown in Figs. 1 and 2, respectively. Metal concentrations increased with the rate at which they had been added,

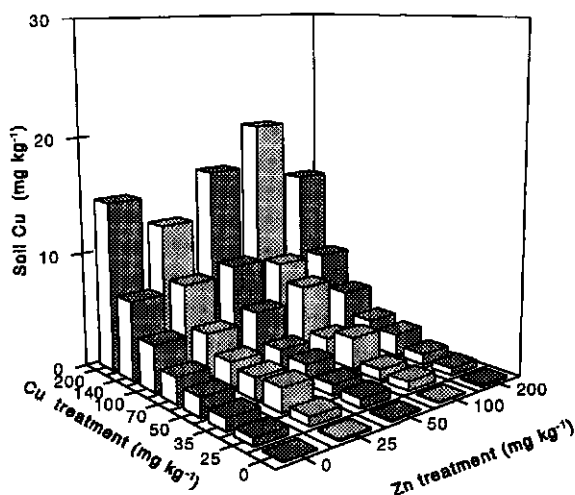


Fig. 1. Measured Cu concentrations ( $mg\ kg^{-1}$ ) in 0.01 M  $CaCl_2$ .

but they were not affected by addition of the other metal. Much more Zn than Cu was extracted, 55.5% and 4.3%, respectively, indicating that Zn was less strongly bound to the soil matrix. The amount of Cu extracted as a percentage of applied Cu, increased with the higher amounts applied, whereas in the case of Zn it was always ca. 55%.

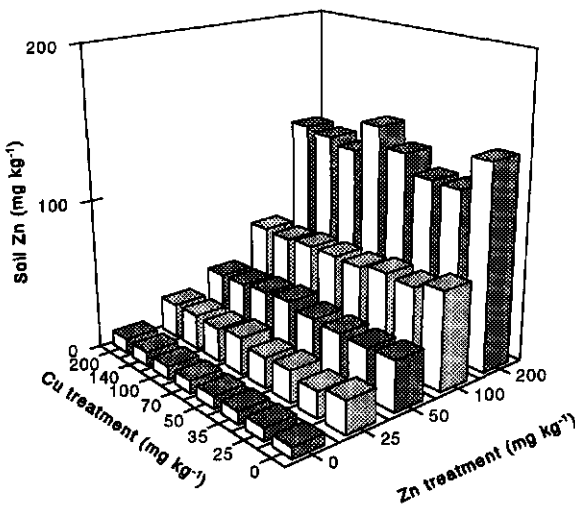


Fig. 2. Measured Zn concentrations ( $\text{mg kg}^{-1}$ ) in 0.01 M  $\text{CaCl}_2$ .

#### Effects on nematodes

The mean values of several nematode community parameters and nematode taxa (for some selected treatments only), and results of ANOVA (carried out on all treatments) are presented in Tables 1 and 2. Increasing Cu and Zn caused a significant decline in most community structure parameters, with the exception of the total number of nematodes with a  $c$ - $p$  values of 1 and 3 which were only significantly affected by Zn. Nematode taxa with  $c$ - $p$  values 3, 4 and 5 (such as *Thonus*, *Alaimus* and *Aporcelaimellus*) were present in low numbers in the control soil and disappeared at Cu and Zn addition rates exceeding 50  $\text{mg kg}^{-1}$ . Few nematode taxa were significantly reduced in numbers by Cu only (*Ditylenchus* and *Aphelenchus*) or Zn only (*Dauer-larvae* and *Prismatolaimus*).

Table 1. Absolute abundances (per 100 g dry soil) for some selected treatments only (means  $\pm$  SE; n=3). Results of ANOVA are based on all treatments.

| Taxon                     | T | c-p | Applied Cu (mg kg <sup>-1</sup> ) |               |                |              | Applied Zn (mg kg <sup>-1</sup> ) |     |     |     | Main effect |     |     |     | Interaction |     |
|---------------------------|---|-----|-----------------------------------|---------------|----------------|--------------|-----------------------------------|-----|-----|-----|-------------|-----|-----|-----|-------------|-----|
|                           |   |     | 0                                 | 200           | 0              | 200          | 0                                 | 200 | 0   | 200 | Cu          | Zn  | P   | P   | Cu*Zn       | P   |
| <i>Pratylenchus</i>       | P | -   | 370 $\pm$ 68                      | 135 $\pm$ 31  | 205 $\pm$ 27   | 184 $\pm$ 32 | 62 $\pm$ 31                       | *** | *** | *** | ***         | *** | *** | *** | ns          | ns  |
| <i>Tylenchorhynchus</i>   | P | -   | 168 $\pm$ 51                      | 29 $\pm$ 2    | 76 $\pm$ 7     | 140 $\pm$ 29 | 28 $\pm$ 5                        | *** | *** | *** | ***         | *** | *** | *** | ***         | *** |
| <i>Filenchus</i>          | P | -   | 78 $\pm$ 51                       | 5 $\pm$ 5     | 0 $\pm$ 0      | 0 $\pm$ 0    | 0 $\pm$ 0                         | *** | *** | *** | ***         | *** | *** | *** | ***         | ns  |
| <i>Basiria</i>            | P | -   | 29 $\pm$ 29                       | 3 $\pm$ 3     | 10 $\pm$ 5     | 23 $\pm$ 12  | 12 $\pm$ 3                        | ns  | ns  | ns  | ns          | ns  | ns  | ns  | ns          | ns  |
| <i>Rotylenchus</i>        | P | -   | 19 $\pm$ 10                       | 17 $\pm$ 10   | 8 $\pm$ 8      | 20 $\pm$ 6   | 3 $\pm$ 3                         | ns  | ns  | ns  | ns          | ns  | ns  | ns  | ns          | ns  |
| <i>Diplogasteridae</i>    | B | 1   | 22 $\pm$ 13                       | 26 $\pm$ 11   | 66 $\pm$ 3     | 51 $\pm$ 7   | 22 $\pm$ 14                       | ns  | ns  | ns  | ns          | ns  | ns  | ns  | ***         | *** |
| <i>Rhabditidae</i>        | B | 1   | 1359 $\pm$ 56                     | 996 $\pm$ 173 | 1078 $\pm$ 144 | 984 $\pm$ 27 | 873 $\pm$ 6                       | *   | ns  | ns  | *           | *** | *** | *** | ns          | ns  |
| Dauer-larvae <sup>1</sup> | B | 1   | 55 $\pm$ 45                       | 65 $\pm$ 45   | 13 $\pm$ 13    | 13 $\pm$ 7   | 9 $\pm$ 0                         | ns  | ns  | ns  | ns          | *   | *** | *** | ns          | ns  |
| <i>Driolophobus</i>       | B | 2   | 61 $\pm$ 20                       | 0 $\pm$ 0     | 0 $\pm$ 0      | 0 $\pm$ 0    | 0 $\pm$ 0                         | *** | *** | *** | ***         | *** | *** | *** | ***         | *** |
| <i>Eucephalobus</i>       | B | 2   | 76 $\pm$ 10                       | 0 $\pm$ 0     | 0 $\pm$ 0      | 0 $\pm$ 0    | 0 $\pm$ 0                         | *** | *** | *** | ***         | *** | *** | *** | ***         | *** |
| <i>Cervidellus</i>        | B | 2   | 0 $\pm$ 0                         | 0 $\pm$ 0     | 4 $\pm$ 4      | 0 $\pm$ 0    | 0 $\pm$ 0                         | ns  | ns  | ns  | ns          | ns  | ns  | ns  | ns          | ns  |
| <i>Acrobeloides</i>       | B | 2   | 329 $\pm$ 78                      | 141 $\pm$ 25  | 140 $\pm$ 35   | 145 $\pm$ 24 | 16 $\pm$ 6                        | *** | *** | *** | ***         | *** | *** | *** | ***         | *** |
| <i>Acrobeles</i>          | B | 2   | 53 $\pm$ 30                       | 0 $\pm$ 0     | 0 $\pm$ 0      | 0 $\pm$ 0    | 0 $\pm$ 0                         | *** | *** | *** | ***         | *** | *** | *** | *           | *   |
| <i>Prismatolaimus</i>     | B | 3   | 53 $\pm$ 27                       | 0 $\pm$ 0     | 0 $\pm$ 0      | 0 $\pm$ 0    | 0 $\pm$ 0                         | ns  | ns  | ns  | ns          | ns  | ns  | ns  | ns          | ns  |
| <i>Pseudhalenchus</i>     | H | 2   | 0 $\pm$ 0                         | 17 $\pm$ 5    | 21 $\pm$ 11    | 13 $\pm$ 8   | 0 $\pm$ 0                         | ns  | ns  | ns  | ns          | ns  | ns  | ns  | ns          | ns  |
| <i>Aphelenchus</i>        | H | 2   | 39 $\pm$ 29                       | 0 $\pm$ 0     | 10 $\pm$ 5     | 9 $\pm$ 4    | 0 $\pm$ 0                         | *   | *   | *   | *           | *   | *   | *   | ns          | ns  |
| <i>Ditylenchus</i>        | H | 2   | 22 $\pm$ 13                       | 11 $\pm$ 7    | 10 $\pm$ 5     | 4 $\pm$ 4    | 6 $\pm$ 6                         | *   | *   | *   | *           | *   | *   | *   | ns          | ns  |
| <i>Aphelenchoides</i>     | H | 2   | 238 $\pm$ 80                      | 86 $\pm$ 8    | 130 $\pm$ 24   | 146 $\pm$ 36 | 47 $\pm$ 6                        | **  | **  | **  | **          | **  | **  | **  | ***         | ns  |

c-p=c-p value; T: feeding group (P plant feeder, B bacterial feeder, H hyphal feeder); ANOVA results based on data from all treatments: ns:  $P > 0.05$ ; \*  $0.05 > P > 0.01$ ; \*\*  $0.01 > P > 0.001$ ; \*\*\*  $P < 0.001$ ; <sup>1</sup> Dauer-larvae of Rhabditidae

Table 2. Absolute abundances (per 100 g dry soil) and Maturity Index values for some selected treatments only (means  $\pm$  SE; n=3). Results of ANOVA are based on all treatments.

| Applied Cu (mg kg <sup>-1</sup> ) | 0               | 200             | 0               | 100             | 200             | Treatment effects |     |       |
|-----------------------------------|-----------------|-----------------|-----------------|-----------------|-----------------|-------------------|-----|-------|
| Applied Zn (mg kg <sup>-1</sup> ) | 0               | 0               | 200             | 100             | 200             | Cu                | Zn  | Cu*Zn |
| Total abundance                   | 3100 $\pm$ 104  | 1538 $\pm$ 164  | 1789 $\pm$ 149  | 1731 $\pm$ 53   | 1085 $\pm$ 48   | ***               | *** | ns    |
| Plant feeders                     | 684 $\pm$ 82    | 197 $\pm$ 19    | 310 $\pm$ 27    | 367 $\pm$ 39    | 112 $\pm$ 33    | ***               | *** | ns    |
| Bacterial feeders                 | 2101 $\pm$ 71   | 1228 $\pm$ 187  | 1310 $\pm$ 129  | 1192 $\pm$ 36   | 920 $\pm$ 13    | ***               | *** | ns    |
| Hyphal feeders                    | 300 $\pm$ 59    | 114 $\pm$ 9     | 169 $\pm$ 31    | 172 $\pm$ 51    | 53 $\pm$ 4      | ***               | *** | ns    |
| c-p 1                             | 1443 $\pm$ 83   | 1086 $\pm$ 166  | 1162 $\pm$ 129  | 1047 $\pm$ 13   | 904 $\pm$ 15    | ns                | **  | ns    |
| c-p 2                             | 850 $\pm$ 103   | 255 $\pm$ 16    | 318 $\pm$ 61    | 317 $\pm$ 41    | 68 $\pm$ 9      | ***               | *** | ns    |
| c-p 3                             | 61 $\pm$ 31     | 0 $\pm$ 0       | 0 $\pm$ 0       | 0 $\pm$ 0       | 0 $\pm$ 0       | ns                | *** | ns    |
| c-p 4                             | 46 $\pm$ 14     | 0 $\pm$ 0       | 0 $\pm$ 0       | 0 $\pm$ 0       | 0 $\pm$ 0       | ***               | *** | ***   |
| c-p 5                             | 15 $\pm$ 15     | 0 $\pm$ 0       | 0 $\pm$ 0       | 0 $\pm$ 0       | 0 $\pm$ 0       | ns                | ns  | ns    |
| Maturity Index                    | 1.47 $\pm$ 0.04 | 1.19 $\pm$ 0.02 | 1.22 $\pm$ 0.04 | 1.23 $\pm$ 0.02 | 1.07 $\pm$ 0.01 | ***               | *** | ns    |

ANOVA based on data from all treatments: ns:  $p > 0.05$ ; \*  $0.05 > p > 0.01$ ; \*\*  $0.01 > p > 0.001$ ; \*\*\*  $p < 0.001$ 

Data obtained for the single metal treatments allowed estimation of  $EC_{50}$  values for added metal concentrations and concentrations measured in  $CaCl_2$  (Table 3). It was found that several nematode taxa were too tolerant (e.g. Rhabditidae and Diplogasteridae) or too sensitive (e.g. *Basiria* and *Prismatolaimus*) to estimate an  $EC_{50}$ .  $EC_{50}$  values based on nominal added Cu concentrations ranged from the 234 mg kg<sup>-1</sup> for the number of nematode taxa to 32 mg kg<sup>-1</sup> for *Acrobeles*, corresponding with Cu concentrations in  $CaCl_2$  of 32 and 1 mg l<sup>-1</sup>, respectively. Within the same parameter,  $EC_{50}$  values based on nominal added Zn

Table 3. Estimates of  $EC_{50}$  (mg kg<sup>-1</sup>) for Cu and Zn applied singly (95% confidence interval in parentheses).  $EC_{50}$  values are based on nominal concentrations and concentrations obtained after extraction with 0.01 M  $CaCl_2$ .

| Parameter                  | Cu nominal |           | Cu $CaCl_2$ |            | Zn nominal |           | Zn $CaCl_2$ |            |
|----------------------------|------------|-----------|-------------|------------|------------|-----------|-------------|------------|
| N (100 g <sup>-1</sup> DW) | 234        | (126-447) | 23          | (5.4-100)  | 295        | (224-398) | 204         | (123-331)  |
| Nr. of Taxa                | 234        | (155-363) | 32          | (5.6-182)  | 457        | (251-813) | 457         | (129-1585) |
| Plant feeding              | 148        | (112-196) | 6.6         | (3.6-12)   | 224        | (170-295) | 126         | (85-191)   |
| c-p 2                      | 170        | (120-240) | 8.5         | (4.3-17)   | 204        | (155-269) | 112         | (76-162)   |
| p-p 2                      | 63         | (47-85)   | 1.3         | (0.8-2)    | 110        | (49-240)  | 44          | (16-117)   |
| p-p 3                      | 170        | (126-229) | 8.7         | (4.4-17)   | 245        | (186-324) | 141         | (93-214)   |
| <i>Pratylenchus</i>        | 200        | (146-269) | 13          | (5.9-28)   | 269        | (102-708) | 166         | (47-589)   |
| <i>Tylenchorhynchus</i>    | 120        | (91-162)  | 4.4         | (2.6-7.4)  | -          | -         | 129         | (65-257)   |
| <i>Acrobeloides</i>        | 196        | (124-268) | 14          | (5.3-37)   | 219        | (158-309) | 123         | (78-195)   |
| <i>Eucephalobus</i>        | 56         | (35-89)   | 2.2         | (1.4-3.5)  | -          | -         | 30          | (25-35)    |
| <i>Aphelenchus</i>         | 43         | (16-117)  | 1.6         | (0.58-4.3) | -          | -         | -           | -          |
| <i>Acrobeles</i>           | 32         | (19-54)   | 1           | (0.89-1.1) | 60         | (25-141)  | 21          | (12-35)    |

c-p=c-p value group; p-p=p-p value group

concentrations were always higher by a factor 1.1 to 2, but ranking the different parameters from low to high  $EC_{50}$  values results in a more or less similar order of parameters for both metals. Comparing  $EC_{50}$  for metal concentrations in  $CaCl_2$  shows larger differences between Cu and Zn.

A significant interaction between Cu and Zn was found for 25% of the investigated parameters (Tables 1 and 2). However, there were no obvious trends, indicating that the combined exposure to Cu and Zn differed from that expected on the basis of single

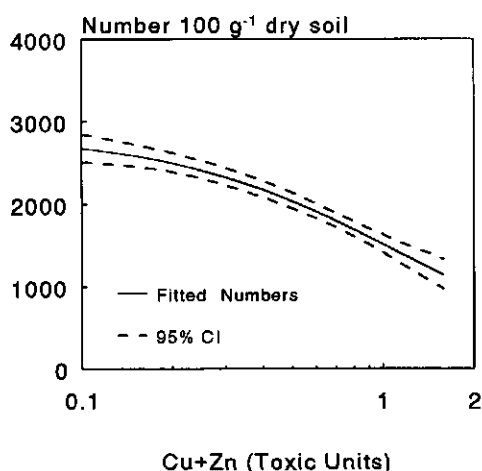


Fig. 3. Influence of the joint toxicity of Cu and Zn (in TU) on the number of nematodes in the soil. The mean relationship is given as well as the 95% confidence intervals.

Table 4. Estimates of  $TU_{50}$  for Cu and Zn (95% confidence interval in parentheses).  $TU_{50}$  values are based on  $EC_{50}$  values for the singel metal exposures expressed in nominal concentrations and concentrations obtained after extraction with 0.01 M  $CaCl_2$ .

| Parameter                  | $TU_{50}$ nominal |             | $TU_{50}$ $CaCl_2$ |             |
|----------------------------|-------------------|-------------|--------------------|-------------|
| N (100 g <sup>-1</sup> DW) | 1.11              | (0.75-1.46) | 0.83               | (0.58-1.08) |
| Plant feeding              | 1.12              | (0.84-1.41) | 0.89               | (0.48-1.29) |
| c-p 2                      | 1.26              | (1.09-1.43) | 1.12               | (0.88-1.36) |
| p-p 2                      | 1.3               | (0.91-1.69) | 1.27               | (0.67-1.88) |
| p-p 3                      | 1.26              | (1.04-1.47) | 1.06               | (0.72-1.41) |
| <i>Pratylenchus</i>        | 1.31              | (1.04-1.57) | 0.99               | (0.57-1.41) |
| <i>Tylenchorhynchus</i>    | 1.25              | (1.02-1.48) | 1.16               | (0.78-1.54) |
| <i>Acrobeloides</i>        | 1.3               | (1.18-1.42) | 1.01               | (0.88-1.15) |
| <i>Eucephalobus</i>        | 1.32              | (1.01-1.63) | 1.3                | (1.03-1.57) |
| <i>Acrobeles</i>           | 0.96              | (-0.19-2.1) | 0.88               | (0.46-1.31) |

c-p=c-p value group; p-p=p-p value group

exposures. For some nematode parameters it was possible to examine the joint toxicity by expressing all metal treatments in Toxic Units. One example of such a dose-response relationship for the joint toxicity of Cu and Zn to the total nematode abundance is shown in Figure 3. For these dose-response relationships  $TU_{50}$  values for the joint toxicity of Cu and Zn was estimated (Table 4). Most of the  $TU_{50}$  values based on nominal concentrations were above, or close to 1 TU, indicating that the effects of combinations of Cu and Zn to the nematode community of this soil were additive or less than additive.  $TU_{50}$  values based on metal concentrations measured in  $CaCl_2$  were always lower than those based on nominal concentrations, and with the exception of *Eucephalobus*, all indicate additive effects.

## Discussion

Although there is an increasing amount of data on joint toxicity, the present study is still one of the few based on terrestrial invertebrates exposed to pollutants in the soil i.e. in the presence of abiotic binding sites. Heavy metals may compete for binding sites in the soil and where two or more metals are present, this may change the bioavailable concentrations (Calamari and Alabaster, 1980; Christensen, 1987; Van Gestel and Hensbergen, 1997), which in turn might affect biota in a different manner than expected on the basis of exposure to a single metal. However, the present study demonstrated that the Cu or Zn concentrations measured in 0.01 M  $CaCl_2$  were not influenced by the applied concentration of the other metal.

A likely explanation for the lack of competition between copper and zinc for binding sites in the soil is that different binding mechanisms were involved. The results of the  $CaCl_2$  extraction indicate that Cu was mainly bound by chemisorption, whereas at the low pH of the soil used, Zn was bound by electrostatic adsorption (ion exchange). One would expect chemisorption of Zn to gain importance with increasing pH; therefore competition could also become more important at higher pH.

Although the soil pH between treatments in the present study was not significantly different, a previous study (Korthals *et al.*, 1996b) demonstrated that soil pH may change, depending on the metal and the amount added. Since soil pH influences the availability of heavy metals (Korthals *et al.*, 1996a), this might lead to differences in metal toxicity between soils in which metals are applied singly or in combination, which in turn may cause deviations from additivity, i.e. antagonistic or synergistic effects.

Several authors have argued that the effects at the highest metal concentrations may be enhanced by the high anion levels introduced (Weltje *et al.*, 1995; Van Gestel and Hensbergen, 1997). In the present study this was circumvented by adding the metals as sulphates and balancing differences in sulphates between the treatments by adding calcium sulphate. Precipitation of added sulphate and calcium ions displaced from exchange sites by adsorbing heavy-metal ions (gypsum), probably kept the dissolved salt concentration low and comparable between the treatments. Another advantage of using  $\text{CuSO}_4$  and  $\text{ZnSO}_4$  is that the total amount of  $\text{SO}_4^{2-}$  anions is only half of that when using metal chlorides or nitrates. Another possibility for removing excessive soluble salts is by percolating the treated soil with water prior to the experiments (Van Gestel and Hensbergen, 1997). This seems less advisable, however, when using a 'natural soil' in which the test organisms are already present in the soil before the metals are added.

The present study demonstrated that the effects on a terrestrial nematode community jointly exposed to Cu and Zn were all additive or less than additive. This is in agreement with the conclusions of two reviews (Ikeda, 1994; Hensbergen and Van Gestel, 1995), that in more than 90% of 210 studies, the joint toxicity of pollutant combinations were additive or less than additive. Despite the fact that in the present study Cu and Zn did not compete for binding sites, there are still many factors which could have caused differential effects on the biota. For example the chemical analyses only characterized the bulk soil at the end of the experiment. The nematode community integrates the effects of (changes in) soil conditions during a period of six months, which in fact is one of the major advantages of using organisms for monitoring the quality of the environment in comparison to chemical analyses. Furthermore, interactions between Cu and Zn could have occurred in uptake processes and in binding processes in the target organ(s) of organisms.

For the soil used in the present study, the official Dutch reference values that aim to distinguish between uncontaminated and contaminated soils are 20 (Cu) and 68 (Zn)  $\text{mg kg}^{-1}$ . These reference values were derived from current metal concentrations in the top soil of rural areas. They depend on the clay and organic matter content of the soil (De Haan *et al.*, 1990). Several nematode taxa were negatively affected by metal concentrations below the reference values, such as *Acrobeles* with an  $\text{EC}_{50}$  value of 60  $\text{mg kg}^{-1}$  for Zn. This result is in agreement with Lexmond and Edelman (1987) who have stated that concentrations above the reference values do not necessarily cause negative effects, whereas concentrations below the reference values may still cause negative effects.

The official Dutch intervention values that separate seriously and not seriously contaminated soils calculated for the soil used in this study are approximately 100 (Cu) and



340 (Zn) mg kg<sup>-1</sup>. Taking into consideration that at these values approximately 50% of the species are supposed to be endangered, the intervention value for Zn seems to high for this type of soil. More than half of the investigated nematode taxa were already seriously affected by the highest Zn additions of 200 mg kg<sup>-1</sup>. Other species, especially those species which are not living in the soil pore water such as litter dwelling organisms, could still be protected at these metal concentrations. Nevertheless, it is realistic to assume that the negative effects on this nematode community are illustrative for many species living in the soil pore water. Furthermore, since nematodes play an important role in the functioning of the whole food web, other organisms may also be affected with yet unknown impacts on ecosystem functioning.

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## Chapter 5

### LONG-TERM EFFECTS OF COPPER AND pH ON THE NEMATODE COMMUNITY IN AN AGROECOSYSTEM

#### Abstract

Four copper (0, 250, 500 and 750 kg Cu ha<sup>-1</sup>) and pH (4.0, 4.7, 5.4 and 6.1 in 1M KCl) treatments were applied to an arable agroecosystem. Effects on the nematode community were assessed after 10 years of exposure under field conditions. Both copper and pH had major influences on nematodes. The effect of copper was generally enhanced with decreasing soil pH. The lowest copper application rate which had a significant negative effect on the total number of nematodes was 250 kg ha<sup>-1</sup> at pH 4.0, which is equivalent to a copper concentration of 0.32 mg l<sup>-1</sup> in 0.01 M calcium chloride (Cu-CaCl<sub>2</sub>) in 1992. Species composition and the abundance of trophic groups were more sensitive than the total number of nematodes. Combinations of high copper and low pH significantly reduced the number of bacterial feeding nematodes, whereas the number of hyphal feeding nematodes increased. Omnivorous and predacious nematodes showed the most sensitive response, becoming extinct when Cu-CaCl<sub>2</sub> was 0.8-1.4 mg l<sup>-1</sup>. Plant feeding nematodes showed the largest differences in abundance and appeared to reflect the effects of Cu and pH on primary production. The results suggest that the nematode community was also affected indirectly by copper and pH via other components of the soil food web. It is concluded that nematodes offer excellent perspectives to assess effects of pollutants at the community level.

*Keywords:* Nematodes; Copper; Community; pH; Soil

#### Introduction

The ultimate goal of ecotoxicology is to predict the effects of pollutants on ecosystems. Experiments on the long-term effects of pollutants on whole fauna communities are important to reach this objective. The advantage of these studies is that they include mechanisms in which the pollutant indirectly affects the community, by changing the food availability and the interactions between species. Several authors (Underwood and Peterson, 1988; Clements, 1994) suggested that the importance of indirect effects increases in complex soil systems and that they are of major concern for the interpretation of ecotoxicological effects.

Studying fauna communities of polluted sites in the field (observational field studies) is a suitable approach, but mainly because of inadequate reference sites, mixtures of different pollutants and natural variability, the interpretation of results and establishment of cause-and-effect relationships are very complicated. To circumvent these problems, studies in which naturally occurring fauna communities

are intentionally exposed to a pollutant (field experiments) probably provide better possibilities for documenting a causal relationship between contaminants and their effects.

Nematodes offer perspectives for the study of pollutant effects at the community level. The phylum Nematoda is comprised of many species showing a high diversity of life cycles, feeding types and sensitivities to pollutants. Based on the food source, nematodes have been assigned to trophic groups which reflect the trophic structure of the soil food web. Furthermore, they are abundant and important for key processes such as nutrient cycling (Freckman, 1988). Finally, terrestrial nematodes live in the soil pore water and are therefore assumed to be in close contact with the bioavailable concentration of pollutants (Houx and Aben, 1993).

It has been demonstrated that terrestrial nematode communities can indicate disturbances caused by manipulating soil pH (Ruess and Funke, 1992; de Goede and Dekker, 1993), tillage (Freckman and Ettema, 1993; Yeates, 1990) or manuring (Ettema and Bongers, 1993). Information on effects of heavy metals on terrestrial nematodes relates mainly to short-term single-species laboratory studies, although there are some observational field studies in which the structure of a nematode community was investigated (Zullini and Peretti, 1986; Yeates *et al.*, 1994; Popovici and Korthals, 1995). However, field experiments in which the long-term effects of an intentional heavy metal pollution on nematodes have been studied are rare.

The objective of the present study was to examine the long-term effects of copper and pH on the nematode community of an agroecosystem. It was hypothesized that the species composition and abundance of trophic groups will be sensitive to changes in the agroecosystem. In a previously unpolluted arable field copper concentrations and soil pH were manipulated. Copper was chosen because of the ongoing accumulation of this heavy metal in the top soil layer of many agroecosystems, mainly as a result of adding pig manure and sewage sludge (van Driel and Smilde, 1990). The pH was selected as a main variable because both the binding of copper to the soil and its phytotoxicity are pH dependent (Lexmond, 1980). After 10 years of normal arable land use, the effects of copper and pH were assessed by studying the composition of the nematode fauna.

## Materials and methods

### *Study site*

The experimental site is located on the eastern side of the Gelderse Vallei, ca. 3 km NNE of Wageningen, The Netherlands. It belongs to an area known as the Bovenbuurt pastures (Buringh, 1951). The soil parent material is cover sand, consisting of slightly loamy, moderately fine sand (de Bakker, 1979). The dominant soil type is a fomic anthrosol (FAO, 1988) with an A-horizon between 50 and 60 cm thick. The subdominant soil type can be classified as a dystric gleysol. The presence of a fomic A-horizon indicates that the field had been used as arable land in the past, which is confirmed by historical records.

### *Experimental design*

In 1978 the experimental field and two adjacent areas, which had been used as permanent pasture for at least 30 years, were ploughed and the surface smoothed. As a result, the A-horizon was reduced to 30–40 cm. Silage maize was grown for three successive years, followed by starch potatoes in 1981. During these four years the crops were fertilized with both liquid cattle manure and mineral fertilizers. After 1981, only mineral fertilizers were applied and no organic materials other than crop residues entered the soil. In autumn 1981, the ploughed layer was sampled on 42 10\*10 m plots, randomly distributed over the field, to measure the variability of some important soil characteristics. The organic carbon content was  $2.1 \pm 0.3\%$  by mass, pH-KCl  $4.7 \pm 0.4$  and copper extractable with dilute nitric acid (Novozamsky *et al.*, 1993)  $3.9 \pm 0.4 \text{ mg kg}^{-1}$ . The soil had a texture of 3% clay, 10% silt and 87% sand and a CEC of 5.6 cmol<sub>c</sub> per kg (NH<sub>4</sub>acetate, pH 7). In 1982 oats were grown on ninety 3\*10 m plots with a mean grain yield of 5730 kg dry matter ha<sup>-1</sup> (coefficient of variation 4.3%). The field thus appeared to be sufficiently uniform for experimentation.

In the autumn of 1982 the experimental field (48\*176 m) was divided into 128 plots of 6\*11 m each. The plots were arranged in 8 blocks of 22\*48 m each. Four copper (Cu) levels were introduced by applying CuSO<sub>4</sub>.5H<sub>2</sub>O at rates of 0, 250, 500 and 750 kg Cu ha<sup>-1</sup>. The pH was adjusted to pH-KCl 4.0, 4.7, 5.4 or 6.1 by adding flower of sulphur or ground calcitic limestone at rates of -1310, 510, 2330 and 4150 kg CaO-equivalents ha<sup>-1</sup> respectively. Half of the Cu, lime or sulphur required were applied in September 1982 and were worked in with a rotary tiller. In October 1982, the field was ploughed and the remaining half of the chemicals was applied. Each Cu and pH treatment was represented and distributed randomly once in each of the 8 blocks.

From 1983 onwards, the crop rotation of silage maize, starch potatoes and oat started in 1980 was continued. During this period yields of maize and potatoes were determined. Mineral fertilizers applied to maize, potatoes and oat averaged 210, 230 and 60 kg N, 120, 110 and 30 kg P<sub>2</sub>O<sub>5</sub> and 170, 210 and 110 kg K<sub>2</sub>O, ha<sup>-1</sup> yr<sup>-1</sup>, respectively. In 1988 pH levels were readjusted to their nominal values by application of ground dolomitic limestone at rates of 0, 1440, 2280 and 2930 kg CaO-equivalents ha<sup>-1</sup> for pH-KCl 4.0, 4.7, 5.4 or 6.1 respectively.

### *Sampling*

Soil samples were taken in March 1992. In the centre of each plot (4 m x 9 m) 30 cores (diameter 17 mm) from the top 10 cm were taken in a regular pattern, mixed and divided into two portions.

One portion was dried at room temperature and sieved (mesh-size 2 mm). pH-KCl was measured after 10 ml soil was suspended in 50 ml 1 M KCl. To

characterize the copper status of the soil, soil samples were extracted with 0.43 M  $\text{HNO}_3$  or 0.01 M  $\text{CaCl}_2$ , so as to estimate the quantity ( $\text{Cu-HNO}_3$ ) or the intensity ( $\text{Cu-CaCl}_2$ ) of available copper respectively (Westerhoff, 1955). Copper ( $\text{Cu-HNO}_3$ , in  $\text{mg kg}^{-1}$  dry soil) was extracted from 10 g soil by shaking for 2 h with 100 ml 0.43 M  $\text{HNO}_3$  (Novozamsky *et al.*, 1993). Another 10 g dry soil was suspended in 100 ml  $\text{CaCl}_2$  (0.01 M) for 20 h. The suspension was centrifuged to determine the copper concentration in solution ( $\text{Cu-CaCl}_2$  in  $\text{mg l}^{-1}$ ). Metal analyses were performed by atomic absorption spectrometry (Houba *et al.*, 1989; Houba *et al.*, 1993). In addition, the vertical distribution of Cu was investigated by sampling the soil profile in 10 cm layers to a depth of 60 cm. Corresponding layers from each of the eight treatment plots were combined and analyzed for  $\text{Cu-HNO}_3$ .

Nematodes were extracted from the other portion of the soil sample using a modified Oostenbrink elutriator (Oostenbrink, 1960). The total number of nematodes was estimated by counting 2 subsamples (approx. 10 % of the total sample) under a dissecting microscope. Nematode numbers were expressed per 100 g dry soil after a correction for material left on the topsieve (mainly stones). Nematodes were heat-killed, fixed in 4% formalin and brought on a permanent mass-slide of which at least 150 nematodes were identified at 400x-1000x according to Bongers (1988). Nematodes were allocated to feeding groups according to Yeates *et al.* (1993).

#### *Statistical analysis*

A two factorial design with 8 completely randomized blocks was used. Data were analyzed by analysis of variance (ANOVA) to test the main effects of copper and pH, and their interaction. If necessary logarithmic transformations were applied to meet assumptions of normality and homogeneity of variances (Sokal and Rohlf, 1981). Tukey's multiple range test was employed to test for differences among treatments. All statistical analyses were performed with the software program Statgraphics 2.6 (Manugistics, 1986).

## **Results**

#### *Soil analysis*

The copper and pH levels found after 10 years of experimentation are presented in Table 1.  $\text{Cu-HNO}_3$  increased linearly with the level of applied copper. Within the copper treatments,  $\text{Cu-HNO}_3$  tended to decrease with decreasing nominal pH, which is probably due to leaching into deeper soil layers, as is shown for the two most obvious treatments (Fig. 1).  $\text{Cu-CaCl}_2$  increased with increasing copper addition and decreasing pH level (Fig. 2). Differences between the actual pH-values were smaller than between the nominal pH-values.

#### *Crop yield*

Maize and potatoes showed strong treatment effects (Table 1). The effects of Cu, pH and their interaction were all highly significant ( $p < 0.01$ ) in every single year.

Table 1. Actual pH and Cu values in 1992 ( $n = 8$ ; Means  $\pm$  SE) and crop yield during 1983-1993.

| Treatment |                        | Soil data in 1992 |   |   | Crop Yield         |                       |
|-----------|------------------------|-------------------|---|---|--------------------|-----------------------|
| pH-KCl    | kg Cu ha <sup>-1</sup> | pH-KCl            | Cu-HNO <sub>3</sub><br>(mg kg <sup>-1</sup> ) | Cu-CaCl <sub>2</sub><br>(mg l <sup>-1</sup> ) | Maize <sup>a</sup> | Potatoes <sup>b</sup> |
| 4.0       | 0                      | 3.88 $\pm$ 0.03   | 25 $\pm$ 3                                    | 0.10 $\pm$ 0.02                               | 9.5                | 36.2                  |
| 4.0       | 250                    | 3.96 $\pm$ 0.03   | 65 $\pm$ 3                                    | 0.32 $\pm$ 0.04                               | 8.7                | 41.0                  |
| 4.0       | 500                    | 3.90 $\pm$ 0.03   | 100 $\pm$ 4                                   | 0.77 $\pm$ 0.05                               | 2.7                | 28.0                  |
| 4.0       | 750                    | 3.82 $\pm$ 0.04   | 134 $\pm$ 6                                   | 1.40 $\pm$ 0.07                               | 0.9                | 8.8                   |
| 4.7       | 0                      | 4.30 $\pm$ 0.04   | 27 $\pm$ 3                                    | 0.04 $\pm$ 0.01                               | 14.6               | 50.3                  |
| 4.7       | 250                    | 4.29 $\pm$ 0.05   | 78 $\pm$ 4                                    | 0.18 $\pm$ 0.01                               | 12.5               | 48.4                  |
| 4.7       | 500                    | 4.32 $\pm$ 0.07   | 104 $\pm$ 5                                   | 0.36 $\pm$ 0.07                               | 6.6                | 41.4                  |
| 4.7       | 750                    | 4.27 $\pm$ 0.07   | 151 $\pm$ 6                                   | 0.71 $\pm$ 1.05                               | 2.1                | 21.4                  |
| 5.4       | 0                      | 5.05 $\pm$ 0.12   | 27 $\pm$ 3                                    | 0.03 $\pm$ 0.00                               | 14.1               | 48.6                  |
| 5.4       | 250                    | 4.97 $\pm$ 0.08   | 74 $\pm$ 3                                    | 0.08 $\pm$ 0.01                               | 14.6               | 48.7                  |
| 5.4       | 500                    | 4.74 $\pm$ 0.07   | 108 $\pm$ 7                                   | 0.18 $\pm$ 0.01                               | 11.2               | 47.0                  |
| 5.4       | 750                    | 4.75 $\pm$ 0.08   | 160 $\pm$ 9                                   | 0.25 $\pm$ 0.02                               | 5.2                | 34.6                  |
| 6.1       | 0                      | 5.65 $\pm$ 0.07   | 29 $\pm$ 5                                    | 0.02 $\pm$ 0.00                               | 15.0               | 45.9                  |
| 6.1       | 250                    | 5.38 $\pm$ 0.08   | 65 $\pm$ 4                                    | 0.06 $\pm$ 0.00                               | 15.2               | 47.3                  |
| 6.1       | 500                    | 5.45 $\pm$ 0.05   | 119 $\pm$ 4                                   | 0.10 $\pm$ 0.00                               | 13.5               | 46.6                  |
| 6.1       | 750                    | 5.37 $\pm$ 0.07   | 168 $\pm$ 6                                   | 0.14 $\pm$ 0.00                               | 10.5               | 42.0                  |

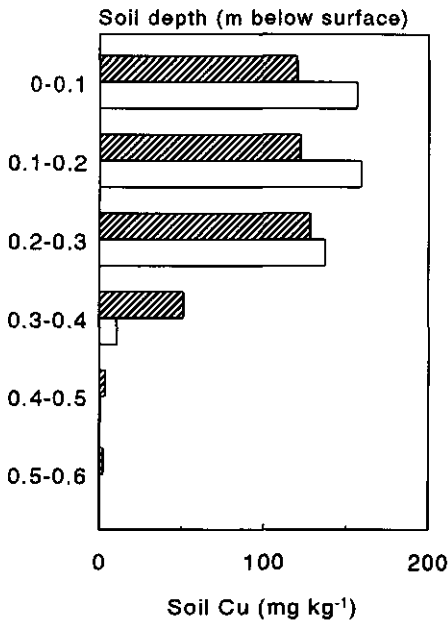
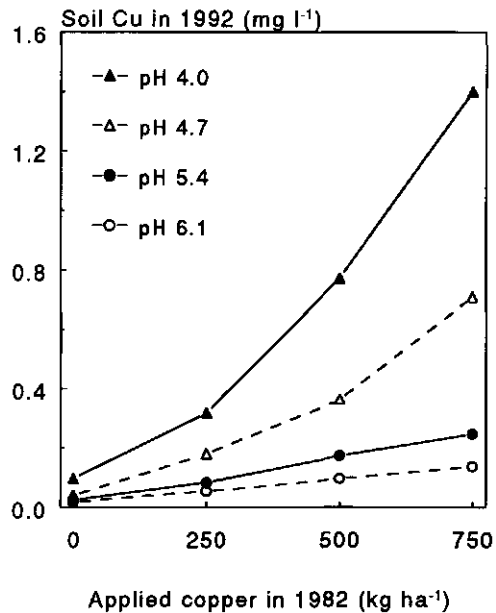
<sup>a</sup>: mean dry matter yield in ton ha<sup>-1</sup> in 1983, 1986, 1989 and 1992<sup>b</sup>: mean tuber yield in ton ha<sup>-1</sup> in 1984, 1987, 1990 and 1993Fig. 1. Mean Cu concentrations (mg kg<sup>-1</sup>) at different depths in 1992 for the applications of 750 kg ha<sup>-1</sup> at pH-KCl 4.0 (hatched bars) and 6.1 (open bars) ( $n = 8$ ).Fig. 2. Mean Cu-CaCl<sub>2</sub> concentrations (mg l<sup>-1</sup>) 10 years after copper and pH treatments in 1982, ( $n = 8$ ).



Table 2. Abundance of nematode taxa (N 100 g<sup>-1</sup>) in 1992

| Treatment              |  | 4.0      |         |         |          | 4.7      |          |         |          |
|------------------------|--|----------|---------|---------|----------|----------|----------|---------|----------|
| pH-KCl                 |  |          |         |         |          |          |          |         |          |
| kg Cu ha <sup>-1</sup> |  | 0        | 250     | 500     | 750      | 0        | 250      | 500     | 750      |
| Total number           |  | 3379 bc  | 3379 bc | 2366 ab | 1027 a   | 4094 cd  | 4114 cd  | 3297 bc | 2125 ab  |
| Plant-feeding          |  |          |         |         |          |          |          |         |          |
| <i>Trichodorus</i>     |  | 0 a      | 4 ab    | 0 a     | 0 a      | 66 cd    | 15 abc   | 0 a     | 0 a      |
| <i>Filenchus</i>       |  | 116      | 89      | 34      | 12       | 77       | 101      | 41      | 32       |
| <i>Basiria</i>         |  | 5 abc    | 15 abcd | 0 a     | 0 a      | 172 e    | 125 de   | 83 cde  | 3 ab     |
| <i>Merlinius</i>       |  | 14 a     | 58 abc  | 10 a    | 0 a      | 62 abc   | 88 abc   | 59 ab   | 9 a      |
| <i>Bitylenchus</i>     |  | 91       | 51      | 11      | 2        | 409      | 204      | 47      | 6        |
| <i>Pratylenchus</i>    |  | 934      | 1023    | 354     | 111      | 1153     | 1630     | 1270    | 319      |
| Bacterial-feeding      |  |          |         |         |          |          |          |         |          |
| <i>Acrobeloides</i>    |  | 678 bcd  | 389 bcd | 327 bc  | 118 a    | 561 bcd  | 380 bcd  | 300 bc  | 253 ab   |
| <i>Chiloplacus</i>     |  | 142 abcd | 282 bcd | 368 d   | 150 abcd | 154 abcd | 335 cd   | 270 cd  | 228 abcd |
| <i>Plectus</i>         |  | 163      | 223     | 82      | 12       | 187      | 205      | 165     | 114      |
| <i>Rhabditis</i>       |  | 95       | 118     | 115     | 58       | 145      | 100      | 97      | 150      |
| <i>Eucephalobus</i>    |  | 85       | 85      | 74      | 59       | 70       | 63       | 80      | 68       |
| <i>Protorhabditis</i>  |  | 86       | 79      | 75      | 82       | 82       | 45       | 82      | 149      |
| <i>Panagrolaimus</i>   |  | 50       | 79      | 35      | 11       | 112      | 102      | 77      | 70       |
| <i>Cephalobus</i>      |  | 12       | 28      | 20      | 5        | 29       | 57       | 46      | 54       |
| <i>Acrobeles</i>       |  | 6 a      | 0 a     | 0 a     | 0 a      | 70 bc    | 8 a      | 0 a     | 2 a      |
| <i>Mesorhabditis</i>   |  | 3 ab     | 8 abcd  | 0 a     | 0 a      | 48 cde   | 25 abcde | 10 abcd | 5 abc    |
| <i>Cervidellus</i>     |  | 65 bc    | 34 abc  | 0 a     | 1 a      | 45 abc   | 30 abc   | 12 ab   | 0 a      |
| <i>Pristionchus</i>    |  | 53 abc   | 12 abc  | 0 a     | 3 ab     | 17 abc   | 18 abc   | 16 abc  | 9 ab     |
| <i>Drilocephalobus</i> |  | 48       | 30      | 13      | 3        | 21       | 77       | 48      | 13       |
| Hyphal-feeding         |  |          |         |         |          |          |          |         |          |
| <i>Aphelenchoides</i>  |  | 496 b    | 510 b   | 626 b   | 344 ab   | 272 ab   | 169 ab   | 370 ab  | 426 b    |
| <i>Ditylenchus</i>     |  | 65 abc   | 31 ab   | 75 abc  | 11 a     | 71 abc   | 99 bc    | 86 bc   | 85 bc    |
| <i>Pseudhalenchus</i>  |  | 108      | 185     | 124     | 33       | 49       | 91       | 75      | 59       |
| <i>Diptherophora</i>   |  | 0 a      | 0 a     | 0 a     | 1 a      | 44 bcd   | 14 ab    | 10 ab   | 0 a      |
| <i>Nothotylenchus</i>  |  | 9        | 4       | 9       | 2        | 25       | 14       | 12      | 5        |
| <i>Aphelenchus</i>     |  | 0 a      | 3 a     | 0 a     | 1 a      | 6 a      | 5 a      | 11 ab   | 7 a      |

<sup>a</sup> Average abundances ( $n = 8$ ) within one row followed by different letters differed significantly ( $P < 0.05$ )

<sup>b</sup> Asterisks indicate significant treatment effects: \*  $0.05 > P \geq 0.01$ , \*\*  $0.01 > P \geq 0.001$ , \*\*\*  $P < 0.001$

Table 3. Abundance of trophic groups (N 100 g<sup>-1</sup>) in 1992

| Treatment              |  | 4.0        |           |         |         | 4.7        |            |           |          |
|------------------------|--|------------|-----------|---------|---------|------------|------------|-----------|----------|
| pH-KCl                 |  |            |           |         |         |            |            |           |          |
| kg Cu ha <sup>-1</sup> |  | 0          | 250       | 500     | 750     | 0          | 250        | 500       | 750      |
| Bacterial-feeding      |  | 1503 bcdef | 1381 bcde | 1110 ab | 506 a   | 1587 bcdef | 1489 bcdef | 1212 abcd | 1126 abc |
| Hyphal-feeding         |  | 684 abc    | 739 abc   | 836 abc | 394 abc | 477 abc    | 394 abc    | 564 abc   | 582 abc  |
| Plant-feeding          |  | 1180       | 1252      | 420     | 127     | 1978       | 2216       | 1500      | 407      |
| Omnivores              |  | 10 ab      | 6 ab      | 0 a     | 0 a     | 33 bc      | 2 ab       | 13 ab     | 3 ab     |
| Carnivores             |  | 3          | 2         | 0       | 0       | 19         | 13         | 9         | 8        |

<sup>a</sup> Average abundances ( $n = 8$ ) within one row followed by different letters differed significantly ( $P < 0.05$ )

<sup>b</sup> Asterisks indicate significant treatment effects: \*  $0.05 > P \geq 0.01$ , \*\*  $0.01 > P \geq 0.001$ , \*\*\*  $P < 0.001$

Table 2. Extended

| 5.4      |          |          |          | 6.1     |         |         |          | Treatment effect |     |         |
|----------|----------|----------|----------|---------|---------|---------|----------|------------------|-----|---------|
| 0        | 250      | 500      | 750      | 0       | 250     | 500     | 750      | Cu               | pH  | Cu * pH |
| 5022 d   | 4555 cd  | 4468 cd  | 4080 cd  | 4401 cd | 3923 cd | 4898 d  | 4585 cd  | ***              | *** | ***     |
| 82 bcd   | 68 d     | 5 ab     | 0 a      | 79 bcd  | 50 abcd | 55 abcd | 12 abc   | ***              | *** | ***     |
| 91       | 115      | 144      | 107      | 180     | 60      | 195     | 144      |                  |     |         |
| 265 e    | 295 e    | 67 bcde  | 97 abcde | 168 e   | 190 e   | 158 de  | 141 de   | ***              | *** |         |
| 447 c    | 309 bc   | 125 bc   | 116 abc  | 368 bc  | 270 bc  | 317 c   | 237 bcd  | ***              | *** |         |
| 131      | 138      | 122      | 96       | 245     | 381     | 109     | 120      |                  | *   |         |
| 1255     | 1054     | 1219     | 866      | 1100    | 961     | 1209    | 1076     |                  |     |         |
| 849 d    | 669 cd   | 393 bcd  | 359 b    | 732 cd  | 552 bcd | 725 cd  | 909 d    | ***              | *** |         |
| 165 abcd | 151 abcd | 250 abcd | 319 bcd  | 20 a    | 87 ab   | 115 abc | 242 abcd | **               |     |         |
| 156      | 138      | 212      | 210      | 94      | 94      | 100     | 66       |                  |     |         |
| 48       | 43       | 161      | 151      | 55      | 80      | 91      | 85       |                  |     |         |
| 106      | 119      | 175      | 129      | 100     | 95      | 147     | 186      |                  | *** |         |
| 40       | 58       | 142      | 140      | 55      | 78      | 55      | 97       |                  |     |         |
| 110      | 135      | 186      | 115      | 92      | 87      | 184     | 107      |                  |     |         |
| 129      | 118      | 111      | 213      | 46      | 63      | 93      | 172      |                  | *** |         |
| 247 c    | 94 c     | 15 ab    | 6 a      | 206 c   | 159 c   | 87 bc   | 9 a      | ***              | *** | **      |
| 104 e    | 116 e    | 24 abcde | 58 bcde  | 82 cde  | 57 bcde | 69 cde  | 65 de    |                  | *** |         |
| 95 c     | 47 abc   | 11 ab    | 12 ab    | 124 c   | 66 bc   | 61 bc   | 11 ab    | ***              | *** |         |
| 36 abc   | 25 abc   | 47 bc    | 105 bc   | 18 abc  | 40 bc   | 84 abc  | 159 c    |                  | *** | **      |
| 64       | 36       | 41       | 24       | 38      | 29      | 72      | 29       |                  |     | **      |
| 168 ab   | 265 ab   | 373 ab   | 529 b    | 74 a    | 105 a   | 161 ab  | 264 ab   |                  | *** | *       |
| 159 bc   | 203 c    | 263 c    | 214 c    | 100 bc  | 149 bc  | 222 c   | 148 bc   |                  | **  |         |
| 28       | 28       | 119      | 67       | 16      | 13      | 34      | 29       |                  | *** |         |
| 39 bcd   | 74 cd    | 35 bcd   | 17 abc   | 67 cd   | 72 d    | 69 cd   | 24 abcd  | **               | *** | *       |
| 6        | 36       | 103      | 29       | 11      | 10      | 90      | 84       |                  | **  |         |
| 34 abc   | 20 abc   | 13 ab    | 22 ab    | 64 c    | 10 ab   | 35 abc  | 54 bc    |                  | *** |         |

Table 3. Extended

| 5.4     |           |            |          | 6.1        |            |         |         | Treatment effect |     |         |
|---------|-----------|------------|----------|------------|------------|---------|---------|------------------|-----|---------|
| 0       | 250       | 500        | 750      | 0          | 250        | 500     | 750     | Cu               | pH  | Cu * pH |
| 2197 f  | 1860 cdef | 1833 bcdef | 1889 def | 1739 bcdef | 1522 bcdef | 2004 ef | 2211 f  | *                | *** | ***     |
| 477 abc | 641 abc   | 908 c      | 877 bc   | 351 a      | 366 ab     | 631 abc | 602 abc | **               | **  | *       |
| 2348    | 2031      | 1708       | 1304     | 2202       | 1963       | 2242    | 1754    |                  |     |         |
| 16 abc  | 11 ab     | 11 abc     | 0 a      | 100 c      | 39 bc      | 14 abc  | 10 ab   | ***              | *** |         |
| 14      | 12        | 8          | 10       | 9          | 34         | 7       | 8       |                  | **  |         |

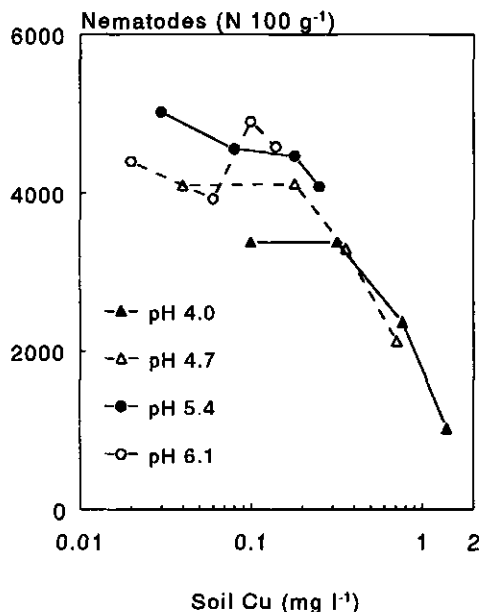


Fig. 3. Mean total number of nematodes found in 1992 for each treatment combination, ( $n = 8$ ).

The coefficient of variation between years amounted to 15% for maize and 24% for potatoes. Since the yield of oats was not determined, no quantitative information can be provided on crop yields in the growing season preceding sampling.

#### *Density and number of nematode taxa*

The total number of nematodes was significantly lower in soils with pH adjusted to 4.7 or 4.0 and loaded with more than 250 kg Cu ha<sup>-1</sup> (Fig. 3, Table 2). These copper and pH combinations were equivalent to CaCl<sub>2</sub> extracted Cu concentrations of above 0.32 mg l<sup>-1</sup>.

In total 74 different taxa were identified. In plots where no copper was added an average of 22 taxa was counted and this number declined to 18 taxa in plots with the highest copper concentrations. Soil pH caused a shift in the mean number of taxa from 15 to 25 for pH 4.0 and pH 6.1 respectively. *Pratylenchidae*, *Cephalobidae*, *Plectidae*, *Rhabditidae* and *Aphelenchoidae* were the most dominant genera.

### *Composition of the nematode fauna*

Taxa with an overall abundance of more than 1 % are summarized in Table 2. Copper significantly reduced numbers of *Trichodorus*, *Basiria*, *Merlinius*, *Acrobeloides*, *Acrobeles*, *Cervidellus* and *Diptherophora*. However, the number of *Chiloplacus* showed an opposite response. An increase in soil pH had a significant positive effect upon the numbers of most of the genera included in Table 2, except for *Aphelenchoides* and *Pseudhalenchus*, which were significantly reduced in numbers at pH 6.1. For *Trichodorus*, *Acrobeloides*, *Acrobeles*, *Pristionchus*, *Driiocephalobus* and *Diptherophora* a significant interaction of copper and pH on nematode numbers occurred with negative effects being most apparent at high copper concentrations and low pH. However numbers of *Aphelenchoides* were found to increase in the presence of copper in combination with a lower pH, except for plots where 750 kg Cu ha<sup>-1</sup> was added and the nominal pH was adjusted to 4.0.

### *Trophic groups*

Increasing copper significantly reduced the total number of bacterial feeding nematodes and had a significant positive effect on the number of hyphal feeding nematodes with highest densities in combinations where 500 kg Cu ha<sup>-1</sup> was added (Table 3). Although omnivorous nematodes were not very abundant, they showed the most sensitive response to copper, decreasing significantly by more than 90 % in the treatment combinations with the highest copper additions.

Increasing pH had a significantly positive effect on the total number of bacterial feeding nematodes. Conversely, an increase in soil pH led to significant reductions of the total number of hyphal feeders, although the highest densities were found in plots where the pH was adjusted to 5.4. With respect to pH as well as to copper, omnivorous and predacious nematodes were the most sensitive trophic groups, totally disappearing when Cu-CaCl<sub>2</sub> concentrations were 0.8-1.4 mg l<sup>-1</sup>. A significant interaction between copper and pH was found for bacterial and hyphal feeding nematodes.

## Discussion

The present study provides an assessment of the effect of copper and pH treatments on the nematode community of an agroecosystem after an exposure period exceeding 10 years. The composition of the nematode fauna and abundance of trophic groups were sensitive measures for the direct and indirect effects of Cu and pH.

A significant interaction between copper and pH was found for many nematode genera, with negative effects being most apparent in plots treated with a high copper dose and a low pH. This reflects that pH affects heavy metal adsorption to the soil, as best described by Cu-CaCl<sub>2</sub>. Although it has been found that increasing pH also affects the relationship between exposure and final effect (toxicity) (Nederlof *et al.*, 1993), the present study indicates that a lower pH enhances the toxicity of copper, which has also been observed for other organisms and heavy metals (Ma, 1982; van Gestel and van Dis, 1988).

The present study showed that the total number of nematodes did respond to an increase in Cu-CaCl<sub>2</sub>, although earlier reports suggested that this is not a sensitive indicator of heavy metal pollution (Bisessar, 1982; Sturhan, 1989). The most likely reason for this discrepancy is the difference in bioavailability, which emphasizes the advantage of the CaCl<sub>2</sub> extraction method. The fact that changes at the genus or family level were better detectable than changes in the total number of nematodes is in agreement with many other studies (Bisessar, 1982; Zullini and Peretti, 1986; Sturhan, 1989; Yeates *et al.*, 1994; Popovici and Korthals, 1995).

There are several possible ways in which copper might have had a direct effect on individual nematodes and thus on the nematode community in this study. One way is via ingestion of contaminated food (Donkin and Dusenbery, 1993; Doelman *et al.*, 1984). Since it can be expected that a change in soil pH has a more or less similar effect on the binding of Cu to the soil as to biotic surfaces (Nederlof *et al.*, 1993), this ingestion can not explain the Cu and pH interaction on nematodes found in the present study.

Another way of copper uptake is via the cuticle. It is possible that in the present study Cu<sup>2+</sup> passed through the cuticle and accumulated to toxic concentrations, which has also been observed for cadmium (Popham and Webster, 1979). It has been suggested that differences in cuticle characteristics contribute to the large

variations in sensitivity to the acute effects of pollutants among nematode species (Kammenga *et al.*, 1994).

Changes in soil pH itself could have led to direct effects on the nematode community. In order to regulate their osmotic pressure, nematodes exchange several ions through their cuticle (Castro and Thomason, 1971). It has been suggested that soil acidification can lead to increasing ion concentrations in the soil pore water to such an extent that nematodes might experience problems in regulating their water status (Bååth *et al.*, 1980). The fact that a lower pH leads to enhanced copper bioavailability and to more ion exchange through the nematode cuticle, is in agreement with the observed interaction between the influence of copper and pH on nematodes.

Copper and pH could have also indirectly affected the nematode community, i.e. by influencing food availability, by interfering with the competitive interactions between species or by affecting the abiotic environment. Although it was not the objective of this study to investigate differences between direct and indirect effects of copper and pH, there are indications that indirect effects did influence the nematode community.

One indication that indirect effects did occur is found among the trophic group comprising of plant feeding nematodes. This trophic group showed the largest differences at the population level and seemed to reflect the effects of Cu and pH on primary production. Although, due to their more aggregated distribution, the present data appeared to be not significant, other studies have indicated that primary production influences densities of plant feeding nematodes (Ingham *et al.*, 1985; Yeates, 1987). Furthermore, it can be expected that in the present study copper reduced the root biomass (Marschner, 1986), which is the food source of plant feeding nematodes. Another indication for indirect effects is the shift from bacterial feeding nematodes to fungal feeding nematodes in plots with high Cu-CaCl<sub>2</sub>. Several other studies have shown that a decrease in soil pH reduces the biomass of bacteria in favour of that of fungi and that such changes are reflected by the trophic composition of the nematode community (Ruess and Funke, 1992; de Goede and Dekker, 1993; Bassus, 1960; Heungens, 1981; Moore and De Ruiter, 1993).

*Aphelenchoides* species have been found to be tolerant to direct effects of pH (Schouten and Van der Brugge, 1989) and heavy metals (Pitcher and McNamara, 1972). Hence, the absolute increase in abundance with low pH and high copper

concentrations found in this study has to be an indirect effect, probably caused by increased fungal biomass, reduced food competition with other organisms, reduced predation pressure or combinations of these factors.

The observation that omnivorous and predacious nematodes were the most sensitive trophic groups agrees with earlier observations on direct effects of pollutants (Kammenga *et al.*, 1994), as well as with long-term investigations in which populations could have been affected indirectly (Ferris and Ferris, 1974; Wasilewska, 1979). Feeding behaviour and life histories of most of the species belonging to these trophic groups are not well understood, but the fact that they generally have a rather long generation time and a more permeable cuticle seems important. Research on the relationship between these characteristics and the effects of pollutants seems to be of major importance to understand why these species are more affected by changes in their environment.

Whereas some earlier studies found an increased abundance or proportion of predacious nematodes in soil contaminated with heavy metals (Yeates *et al.*, 1994; Weiss and Larink, 1991), the present study showed a decrease. This discrepancy may be explained by differences in food availability or bioavailability of the metals, since in the present study metals were applied as salts whereas the earlier studies added metals in combination with organic materials.

While the relative importance of direct and indirect effects of metals on communities remain to be assessed, it is clear that soil nematodes are sensitive indicators of environmental stress induced by copper and pH. Studying trophic groups and species composition of soil nematode communities can provide a sensitive measure of soil quality in the field.

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## Chapter 6

### THE MATURITY INDEX AS AN INSTRUMENT FOR RISK ASSESSMENT OF SOIL POLLUTION

#### Abstract

It is widely acknowledged that the impact of contaminants on ecosystems is difficult to predict from single-species laboratory tests. Moreover there is growing evidence that the ultimate effect of a toxicant can be mediated through indirect effects. Studies carried out at the community level provide a meaningful step between single-species laboratory test and ecosystem studies. In order to interpret complex changes in community structure, there is an urgent need to develop new approaches to monitor soil pollution.

The present paper reports on methods to detect various kinds of disturbances by studying changes in the composition of natural occurring nematode communities. We examined whether the Maturity Index (MI), originally developed to monitor colonization and subsequent development of the nematode fauna, can also provide an instrument to measure the effects of soil pollution. It was found that within the Maturity Index concept, most taxa originally scaled in  $c-p$  1 indicate food-rich conditions. Withholding these taxa from the calculation of the MI results in an index (MI2-5) to indicate disturbances resulting from chronic soil pollution, such as heavy metals.

**Keywords:** Soil pollution; Nematode community; Maturity Index; Biomonitoring

#### Introduction

The ultimate goal of ecotoxicology is to detect, predict and monitor the effects of pollutants on ecosystems (Moriarty, 1983). To overcome the existing gap between our knowledge of the effects that pollutants have on single species or on ecosystems, there is an urgent need to develop new bioindicator systems with more environmental realism (Cairns, 1992). Studying pollution-induced changes in the structure of natural occurring faunal communities can be an important step to bridge this gap. The advantage of such studies is that they also include mechanisms in which the pollutant indirectly affects the community, like for example via changes in the food availability. This can lead to a better understanding of the role ecological interactions play in toxicological responses of more complex systems (Moore and De Ruiter, 1993; Clements, 1994).

The objective of the present paper is to demonstrate new approaches in the development of bioindicator systems based on changes in the structure of nematode communities. Therefore, first a general overview on the successional stages in the development of faunal communities in general and nematode communities in particular will

be given. Next we will test the hypothesis that effects of disturbances can be described as a retrogression in the succession of the nematode community. To test this hypothesis the effects of several kinds of disturbances on the nematode community are studied in terms of changes in the proportion of colonizers and persisters, i.e. *r*- and *K*-strategists, respectively. Finally, the prospects of the nematode Maturity Index (Bongers, 1990) an index based on the relative abundance of colonizers and persisters, as an instrument to discriminate between several types of disturbances of the soil ecosystem, will be discussed.

### **Changes in the structure of communities during succession**

Faunal communities within an ecosystem are in a continuous process of change. It has been suggested that certain trends can be distinguished during successional changes of ecosystems (Odum, 1969). Newly formed habitats are invaded by colonizers (I), which results in an initial dominance of rapidly reproducing species (II). Subsequently, more organisms invade the system and the species diversity and structural and functional complexity increases (III). Finally, the system matures and becomes less controlled by external disturbances but more by biotic interactions (IV). In this stage there is a strong selection for species with a high competitive ability (V).

Although it can be debated whether succession is an unidirectional process and comprise predictable changes, there is a long tradition among ecologists to unravel the biotic and abiotic properties which determine the ecological range of the species within a successional sequence. The theory of *r*- and *K*-strategies showed that the degree of crowding can be one of the explaining factors. Although the meaning of the *r*-*K* concept has broadened, originally it was suggested that *r*-selected species are characterized by a high population growth in uncrowded populations, whereas selected species exhibit a high competitive ability in crowded populations (MacArthur and Wilson, 1967). The *r*-strategists are the successful colonizers. They have a high rate of population increase and a high dispersal ability. In uncrowded environments, like early stages of succession, these species are in favour. *K*-strategists show opposite characteristics and are found in more stable habitats, in which they maximize their carrying capacity (Pianka, 1978; Brown and Southwood, 1987).

Theoretically all species can be placed on a  $r$ - $K$  continuum of strategies, of which some important characteristics are listed below:

| Type of strategy            | $r$ -strategy | $K$ -strategy |
|-----------------------------|---------------|---------------|
| <u>Community attributes</u> |               |               |
| Diversity                   | Low           | High          |
| Interspecific competition   | Occasional    | Frequent      |
| Degree of specialization    | Low           | High          |
| <u>Species attributes</u>   |               |               |
| Colonization ability        | High          | Low           |
| Distribution                | Wide          | Restricted    |
| Length of life              | Short         | Long          |
| Maturity                    | Early         | Late          |
| Rate of development         | Rapid         | Slow          |
| Fecundity                   | High          | Low           |
| Population density          | Variable      | Constant      |
| Capacity for dormancy       | Variable      | Low           |

The  $r$ - $K$  theory has been debated thoroughly. Most of the criticism was based on the fact that other factors than crowding can explain the diversity in life-history strategies as well (Wilbur *et al.*, 1974; Parry, 1981; Southwood, 1977), or that certain species exhibit a dynamic strategy involving shifts between relative  $r$ - $K$  positions along the  $r$ - $K$  continuum (Nichols *et al.*, 1976), or the fact that some pioneer species do have certain characteristics which are not in line with the  $r$ - $K$  concept (Bengtsson and Baur, 1993). Despite this criticism there is much empirical support for the  $r$ - $K$  concept, which warrants an analyses of the application of this ecological concept in studying changes in the structure of faunal communities within ecotoxicological studies.

#### Changes in the structure of nematode communities during succession

A helpful tool in studying changes in the structure of nematode communities during natural succession is the nematode Maturity Index (Bongers, 1990). Based on their ability to colonize new habitats, nematode families were classified on a tentative colonizer-persister ( $c$ - $p$ ) scale ranging from 1 to 5. Nematode families comprising species that rapidly increase in number in early stages of succession, were considered as colonizers and received a low  $c$ - $p$  value. They have similar characteristics as  $r$ -strategists. In general colonizers live in unstable habitats. Species of the families Rhabditidae, Panagrolaimidae and

Diplogasteridae represent typical colonizers. The persisters among the nematodes are comparable with *K*-strategists and they generally live in habitats with a long durational stability. The most extreme persisters are found among the families Nygolaimidae, Thomematidae, Belondiridae, Actinolaimidae and Discolaimidae. During the last couple of years, further information on this MI-concept has become available (Bongers *et al.*, 1991; Bongers *et al.*, 1995; De Goede and Bongers, 1997).

The MI is calculated as the weighted mean of the *c-p* values assigned to the constituent nematode families (and genera and species they contain), such that:

$$MI = \frac{\sum_{i=1}^n (v(i) \cdot a(i))}{\sum_{i=1}^n a(i)}$$

where *v(i)* is the *c-p* value assigned to taxon *i* and *a(i)* is the abundance of taxon *i* in a sample. If, for example, a community exists of 10 Rhabditidae (*c-p* value=1), 10 Diplogasteridae (*c-p* value=1), 30 Cephalobidae (*c-p* value=2) and 50 Dorylaimidae (*c-p* value=5) the MI is  $\{(10+10)*1 + (30)*2 + (50)*5\}/(10+10+30+50) = (330/100) = 3.3$ .

Following this classification of nematode taxa into *c-p* groups it is expected that early stages in the succession of a habitat will be characterized by relatively low MI values. The first colonizers comprise species with the characteristics of *r*-strategists and thus with low *c-p* values. Subsequently, during further development of the habitat the MI will increase, because of increasing numbers of *K*-strategists belonging to the higher *c-p* values. Such pattern of nematode fauna development was observed in several studies comprising natural successional series (De Goede *et al.*, 1993).

### Changes in the structure of nematode communities and MI after disturbances

Natural succession of habitats coincides with patterns of nematode community development, generally resulting in an increasing value of the Maturity Index. However, the process of succession can be influenced by external disturbances, resulting in a retrogression or an arrestment of the succession of habitats (Regier and Cowell, 1972; Whittaker, 1975; Odum, 1985). If environmental disturbances also lead to a retrogression or arrestment of the development of nematode communities, then this should result in decreased MI values.

In terms of the *c-p* group classification within the MI-concept, two kinds of changes in the composition of the nematode fauna after a disturbance were suggested (Bongers, 1990; De Goede *et al.*, 1993). A type I response is found when disturbances result in increased numbers of taxa with low *c-p* values (1-2), whereas taxa from the higher *c-p* groups hardly respond (see e.g. Ettema and Bongers, 1993). A second type of response (response type II), is found when disturbances result in an absolute decrease among most taxa, but in particular those with high *c-p* values (Ruess *et al.*, 1993; Korthals *et al.*, 1996).

Despite essential differences between these types of response of the nematode community after a disturbance, both response types can be seen as a retrogression in the succession of the nematode community; that is, from the perspective of the *r*- and *K*-concept and in terms of the *c-p* classification. Therefore, the MI is not only an index to monitor natural succession, but can also be applied to detect disturbances and to monitor any subsequent recovery of the ecosystem. A review of applications of the MI in environmental studies comprising among others effects of water pollution, heavy metals, eutrophication, oil spill, liming, acidification, physical stresses and tillage regimes is given by De Goede and Bongers, (1997). These studies showed that in general, disturbances are followed by a decrease in MI, whereas recovery or natural succession coincided with an increase in MI.

Bioindicators should give opportunities to distinguish between different stress factors (Bongers and Schouten, 1991). De Goede *et al.* (1993) indicated that the MI-concept offers possibilities to discriminate between effects of eutrophication and pollution, which correspond to changes in the composition of the nematode fauna following the earlier mentioned response types I and II, respectively. Examples of studies demonstrating response type I involve effects of eutrophication, fertilization, oil spill, liming and several physical stresses (De Goede and Bongers, 1997). These disturbances cause such an increase in nutrient availability that, as a result of the (temporal) increased microbial activity, *c-p* 1 taxa ('enrichment opportunists' *sensu* Ettema & Bongers, 1993) can increase in number. When the nematode food supply decreases the densities of the *c-p* 1 taxa will decrease, while often at the same time the densities of *c-p* 2 taxa can increase. The higher scaled taxa (*c-p* 3-5) hardly react numerically to these changes in food supply (Ettema and Bongers, 1993).

At present, examples of the type II response are less often documented. Type II responses were found for soil acidification (Ruess *et al.*, 1993) and for the chronic effects of heavy metals (Korthals *et al.*, 1996; Popovici and Korthals, 1995). Although further confirmation is required, it seems that a type II response can be expected for long-term

effects of disturbances where toxic stress is accompanied with conditions of low food availability. Under such conditions most taxa, but especially those belonging to higher *c-p* groups, decrease in number and the food availability is too low to have high populations of *c-p* 1 taxa. Certain taxa from the *c-p* 2 group (the 'general opportunists' *sensu* Ettema & Bongers, 1993) are the most tolerant to survive these severe conditions, or even benefit of it and increase in number (Korthals *et al.*, 1996).

Table 1. Correlation coefficients (*r*) for heavy metal content versus MI and MI2-5 for 20 agricultural soils in The Netherlands with mixed pollutants (Bongers, 1992).

|         | MI     | MI2-5  |
|---------|--------|--------|
| Copper  | -0.300 | -0.496 |
| Zinc    | -0.318 | -0.502 |
| Cadmium | -0.178 | -0.466 |
| Lead    | -0.154 | -0.461 |

The response types I and II both result in a decrease of the MI. As a rule of thumb a decrease of the MI below 2 as a consequence of an absolute increase found among *c-p* 1 taxa, usually indicates a type I response (eutrophication), whereas samples characterized by MI values close to 2, and a lowered species diversity consisting mainly of taxa with *c-p* value 2, indicates a type II response (pollution). However, it is possible that in habitats with pollution, organic inputs can lead to a period in which *c-p* 1 taxa predominate the nematode

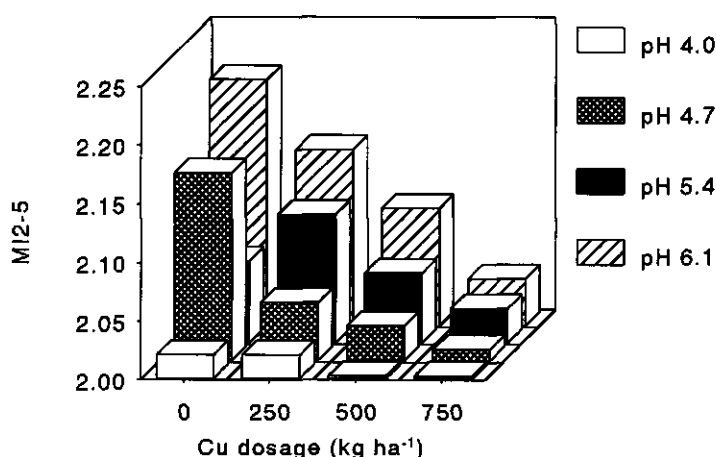


Fig. 1. Applications of MI2-5.



fauna such that the MI is below 2. Under these conditions the MI is less effective to identify pollution (type II response). To reduce this effect of enrichment in measuring pollution induced stress, it might have advantages to omit taxa scaled in *c-p* 1 from the calculation of the MI. We propose to express this new index as the MI2-5. Compared to the MI, the MI2-5 has proved to be more sensitive to indicate pollution as demonstrated in Table 1. Moreover, comparison of the scores of both indices can help to discriminate between pollution induced stress or eutrophication. First applications of this MI2-5 can be found in Figure 1., Korthals *et al.* (1993) and in Popovici (1994).

### Discussion

The classification of nematode taxa into *c-p* groups reflects a first attempt to categorize nematodes in ecological groups with similar life-history characteristics. Although promising results were obtained when using this concept, further calibration of the *c-p* scaling might be necessary. Moreover, substantial more knowledge is needed on the relationship

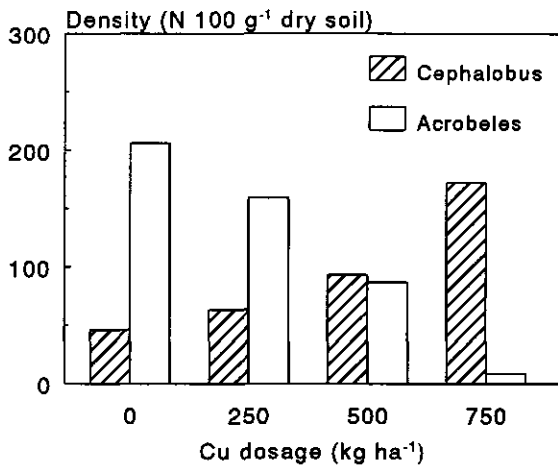


Fig. 2. Relationship between life-history and soil pollution.

between the life-history of certain species and their sensitivity towards soil pollution. Figure 2., for example, shows the effects of Cu on two closely related genera within the Cephalobidae. Although both genera have the same *c-p* value (*c-p* 2), their sensitivity towards Cu

differs greatly. When it becomes possible to unravel these relationships, any improvements of *c-p* scaling can result in an increased sensitivity of the MI and MI2-5.

In addition to the MI and MI2-5, examination of the changes within each of the *c-p* groups on which the indices are based, can provide additional information on the nature of the underlying changes in the habitat. De Goede *et al.* (1993) presented a *c-p* triangle in which they schematically indicated the main directions along which changes in the nematode fauna composition may occur. Since this method makes it possible to visualize the changes in *c-p* 1, 2 and 3-5 distribution, it can also be used to identify type I and type II responses within the nematode community. Moreover, examination of *c-p* groups can also be helpful to detect simultaneous increase or decrease of opportunists and persisters which may remain unnoticed when using only the MI.

As was mentioned in the recommendations of the NATO Advanced Research Workshop (Van Straalen and Krivolutsky, 1996) ecological relevant bioindicator systems should, among others, be based on information obtained from different species and processes. Studies carried out at the community level provide opportunities in this direction. However, in order to interpret complex changes in the structure of whole communities, there is an urgent need to develop new methods which can be used by decision makers as well. The MI-concept can become one of these tools, especially because the changes in MI not only indicate the disturbance acting on a nematode community, but also provide opportunities to identify specific types of disturbance. Therefore, it seems worthwhile to investigate the usefulness of this MI-concept also for other groups of organisms, especially for those organisms that, in comparison to nematodes, have a different route of exposure.

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## Chapter 7

### GENERAL DISCUSSION

The largest problem ecotoxicologists face is how to deal with the high degree of variation between field sites. One possibility is to 'remove' much of the heterogeneity by using a well-defined artificial soil (Edwards, 1983; Edwards, 1984). Since it seems impossible to 'rebuild' whole faunal communities in an artificial soil, the applicability of this kind of standardized toxicity testing will be limited to an initial screening of new pollutants, which should always be succeeded by investigations with a higher field relevance.

Another possibility is to focus on the chemical aspects. Increasing knowledge of the pollutant and the most important factors which control its bioavailability can lead to the incorporation of these factors in standard settings, as has been done earlier for clay and organic matter content in the official Dutch reference values (Lexmond and Edelman, 1987; De Haan *et al.*, 1993). That soil pH is another important factor which should be incorporated in soil standard settings was clearly demonstrated in Chapter 5, where a lower pH increased the bioavailability and enhanced the toxicity of copper to nematodes. This has also been observed for other organisms and heavy metals (Ma, 1982; Van Gestel and Van Dis, 1988). Therefore, the influence of other soil properties and abiotic environmental factors in relation to the bioavailability of pollutants should be investigated in the future.

In this line, increasing efforts in developing methods to measure bioavailable pollutant concentrations should be mentioned. To characterize the metal status in the different experiments of the present thesis, soil samples were extracted with 0.43 M  $\text{HNO}_3$  or 0.01 M  $\text{CaCl}_2$ , in order to estimate the total quantity or the presumed bioavailability of the metals, respectively (Novozamky *et al.* 1993). Both estimates are important with respect to risk assessment, but based on the present thesis, it seems that for comparing the actual risks for nematodes at different sites, the last method seems most suitable.

Standard settings improved with our increasing knowledge of pollutant bioavailability are a useful tool for evaluating the degree of contamination. However, they cannot be used for soil quality assessment in relation to soil functioning (De Haan *et al.*, 1993), unless the (cor)relations with the effects on biota are incorporated. A first attempt to incorporate more ecotoxicology in risk analysis methods is shown by Van Straalen and Denneman (1989), in which No Observed Effect Concentrations (NOEC) for several organisms are incorporated. But in this approach as well, it is necessary that the ecotoxicological tests have a higher relevance for real soil.

Studying changes in nematode communities by exposing field-collected soil to pollutants seems to be a method with a higher degree of realism. Most nematode taxa showed declining numbers with increasing metal concentrations, some of them being very sensitive (*Plectus*, *Clarkus*, *Aporcelaimellus*, *Prismatolaimus*, *Alaimus* and *Acrobeles*) and others being very tolerant (*Pseudhalenchus*, Dauer-larvae of the Rhabditidae, *Aphelenchoides*, *Acrobeloides*, *Pratylenchus* and *Tylenchorhynchus*). Several experiments indicated that indirect effects did occur, such as the absolute increase in abundance of *Aphelenchoides* in soils with low pH and high copper concentrations (Chapter 5). This type of indirect effect was probably caused by increased fungal biomass, reduced food competition by less tolerant organisms, a reduced predation pressure by sensitive predators, or combinations of these factors. Evidence was obtained that these factors are more likely to occur in community-level micro- or mesocosm experiments with 'natural soil', which may cause less, under-, or overestimation of the actual risks, than do results based on soil organisms exposed to pollution in water or artificial soil.

In Chapter 3 it was demonstrated that a field-collected nematode community changed not only due to an abiotic factor (heavy metals), but also due to a biotic factor (presence of *Lolium perenne*). Compared to bare soils, the effects of Cu and Zn in soil covered with *L. perenne* only became apparent at higher metal concentrations, were less severe and were more often caused indirectly. This result was based only on experiments with a monoculture of *L. perenne*, so the final effect of pollutants in a soil covered by a more diverse vegetation may differ to some extent. Therefore, it is recommended that future ecotoxicological experiments with nematode communities should be carried out in the presence of a more natural (diverse) vegetation.

One other important aspect in exposing whole nematode communities to pollutants is that certain soil and vegetation characteristics not only influence the bioavailability of a pollutant, but also influence the structure of the original (unexposed) nematode community. The conclusions of the present thesis were based on a nematode community collected from one agroecosystem which was dominated by some bacterial feeders and plant feeders, of which Rhabditidae, Pratylenchidae, Dolichodoridae, Cephalobidae and Tylenchidae constituted approximately 80% of the total nematode community. This type of community may be quite different from nematode communities found in other ecosystems (De Goede and Bongers, 1994) and, although not investigated, it can be assumed that these differences will affect ecotoxicological results based on community parameters.

Ecotoxicological effects on the community level are the final outcome of the effects on the constituting species. To increase our knowledge on pollutant-induced changes in

terrestrial nematode communities we have attempted to discern general patterns in the sensitivities among different nematode groupings, based on feeding strategies or life-history strategies. An example of classifying life-history strategies among nematodes has formed the basis for the Maturity Index (Bongers, 1990) and has been used to facilitate comparisons of the metal-induced changes in nematode communities in the present thesis. It was hypothesized that external disturbances might retrogress or arrest the succession of faunal communities (Regier and Cowell, 1972; Whittaker, 1975; Odum, 1985). Since the Maturity Index was originally developed as an ecological measure of the state of succession, disturbances should also be reflected in changed MI values.

It was demonstrated that, depending on the disturbance, two kinds of change in the composition of the nematode fauna may occur. Some disturbances, such as eutrophication, fertilization, oil spills, liming and several physical stresses (De Goede and Bongers, 1997), result in increased numbers of taxa with low *c-p* values (1-2), whereas taxa from the higher *c-p* groups hardly respond (e.g. Ettema and Bongers, 1993). A second type of disturbance results in an absolute decrease in most taxa, in particular those with high *c-p* values. Although further confirmation is required for other pollutants than the heavy metals investigated in this thesis, it seems that the second type of response can be expected for long-term effects of pollutants, especially those which are accompanied with conditions of low food availability. To improve the discrimination between "pollution stress" and "eutrophication stress", both of which result in a decreased MI, we proposed to omit taxa scaled in *c-p* 1 from the calculation of the MI and to express this new index as the MI2-5 (Chapter 6). In addition to the MI2-5, the so-called *c-p* triangles, in which it is possible to visualize the changes in *c-p* 1, 2 and 3-5 distribution, can also be used to detect the simultaneous increase or decrease of opportunists and persisters (De Goede *et al.*, 1993). These changes may otherwise remain unnoticed when using only the MI.

The classification of nematode taxa in *c-p* groups, reflects a first attempt to categorize nematodes in ecological groups with similar life-history characteristics. Although promising results were obtained when using this concept, further calibration of the *c-p* scaling seems necessary. The present thesis showed that closely related genera with similar feeding modes and *c-p* values, such as *Acrobeloides*, *Acrobeles*, *Cephalobus* and *Chiloplacus*, within the family Cephalobidae, can have very different toxicological responses. Furthermore, it was found that some genera with a *c-p* value of 2 (e.g. *Plectus* and *Acrobeles*) were as sensitive as genera with higher *c-p* values. These results indicate that substantially more knowledge on the ecology and life-history characteristics of nematodes

on the genus- or species-level in relation to soil pollution is needed. Improvements of c-p scaling are likely to result in an increased indicator value of the MI or related indices.

Indices such as the MI are relative measures; their usage in field monitoring is limited to a signal function. To increase their usability, there is a strong need for a reference system, i.e. an estimate of what kind of nematode community might be expected on a certain site, under natural vegetation and unpolluted conditions. For The Netherlands (Bongers *et al.*, 1989; De Goede and Bongers, 1994; Alkemade and Van Esbroek, 1994), this type of knowledge is gradually increasing. Studying the taxa lists also remains useful in biomonitoring. The number of species, the trophic structure, absolute densities and the dominance of certain indicator species all help to differentiate between sites and to identify the most dominant stress factor.

Since nematodes feeding on higher plants were excluded from the calculation of the MI, a separate Plant Parasite Index (PPI) was proposed (Bongers, 1990). More recently, Yeates (1994) and Wasilewska (1994) proposed the inclusion of plant feeders in the MI. However, the present thesis demonstrated that under certain circumstances, the inclusion of plant parasites can lead to an index which is less sensitive to disturbances than the Maturity Index. Therefore, it is proposed to maintain them separately or to use the ratio between Maturity Index and Plant Parasite Index (Bongers *et al.*, 1997). In the future, the advantages of a set of indices for every trophic level above the MI calculated for all nematodes should be investigated. These separate MI's may provide more detailed information regarding shifts within trophic groups, which is more closely related to changes in the functioning of ecosystems.

To further increase our knowledge on potential risks of pollutants at the community or ecosystem level, an integration of pollutant effects on different taxonomic groups in relation to the functioning of the soil seems important. The advantages of such an approach have been illustrated by Edwards *et al.* (1996). We hope that the work of many nematologists and the present thesis have demonstrated that nematodes should be included in such investigations.

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## SUMMARY

One way to assess the quality of our environment is by comparing chemical data against existing soil quality standards. Preferably these standards are based on effect concentrations obtained for different organisms. Unfortunately there is only a limited number of test species, the test conditions are often very unrealistic and it is very difficult to measure bioavailable pollutant concentrations.

One other approach is to compare biological data collected from the site of which the quality must be assessed with that of a reference site, i.e. those having a good quality. Main difficulties of this approach are the limited knowledge of reference sites and the complexity of interpreting the data.

Within both approaches there is a strong need for more knowledge on the effects of pollutants on whole communities. Therefore the research presented in this thesis mainly investigated heavy metal induced changes in terrestrial nematode communities exposed in microcosms tests or in a field experiment. Nematodes were chosen as test species since they are present in almost every habitat and their communities are normally characterized by high densities and a high species diversity. Nematodes play a prominent role in terrestrial food webs, are easy to sample and are representative of the soil being sampled. Furthermore, some existing functional groupings, based on for example feeding strategy or on life-history strategies, might facilitate the interpretation of pollutant induced changes in nematode communities.

In several experiments whole nematode communities, present in freshly collected agricultural soil, were exposed to cadmium, copper, nickel or zinc. Our main objective was to increase the complexity of the successive experiments, in order to get a better understanding of the effects of some heavy metals upon terrestrial nematode communities in the real world.

Therefore, in Chapter 2 the nematode community structure was studied one to two weeks after the addition of cadmium (Cd), copper (Cu), nickel (Ni) or zinc (Zn) to soil collected from an agroecosystem. The nematode community was found to be affected by increasing concentrations of Cu, Ni and Zn up to  $1600 \text{ mg kg}^{-1}$ , but not by Cd up to  $160 \text{ mg kg}^{-1}$ .  $\text{EC}_{50}$  values for the reduction in population size of individual taxa showed a low intra-taxon variation for Cu, Ni and Zn, which seemed to indicate that for these metals, uptake and elimination processes as well as their final effect appeared similar within the same taxon. However, major differences between the sensitivities of different nematode taxa were detected, with for example some omnivorous and predatory nematodes, known to be

## Summary

'K-strategist', already significantly affected by 100 mg kg<sup>-1</sup> Cu, Ni or Zn added to the soil. The relative abundance of the different life-history groups and, to a lesser extent, the different feeding groups indicated pollution-induced changes in the soil community. However, these classifications couldn't predict the sensitivity of different nematode genera in an adequate way.

In Chapter 3 the same 'natural soil method' was used to study the long-term effects of copper and zinc (0, 25, 50, 100, 200 and 400 mg kg<sup>-1</sup>) in the presence or absence of ryegrass (*Lolium perenne* L.). It was demonstrated that the presence of vegetation is a very important factor in determining the final ecotoxicological effects of Cu and Zn. In soils covered with *L. perenne* it was found that the effects of Cu and Zn became only apparent at higher metal concentrations, were less severe and were more often caused in an indirect way. Therefore it was recommended that future risk assessment based on results obtained from micro- or mesocosms should include vegetation.

In Chapter 4 the possible consequences of another realistic aspect of pollution in the real world, namely the simultaneously presence of several pollutants, were studied. After an exposure period of a half year, it was found that many nematode community parameters were affected by increasing concentrations of Cu and Zn up to 200 mg kg<sup>-1</sup>. However the bioavailable Cu or Zn concentrations measured in soils with combined additions of Cu and Zn were not significantly different from single metal additions, indicating that in the present study Cu and Zn did not affect each other's bioavailability. After evaluating changes in the nematode community with the Toxic Unit model, it was concluded that the potential risks of a combined exposure to Cu and Zn can be judged by assuming additiveness or less than additiveness.

Chapter 5 focused on the long-term effects of copper and pH on a nematode community under the most realistic conditions, that is in an agroecosystem. Both copper and pH had major influences on nematodes. The effect of copper was generally enhanced with decreasing soil pH. The lowest copper application rate which had a significant negative effect on the total number of nematodes was a copper concentration of 0.32 mg l<sup>-1</sup>. Species composition and the abundance of trophic groups were even more sensitive than the total number of nematodes. Combinations of high copper and low pH significantly reduced the number of bacterial-feeding nematodes, whereas the number of hyphal-feeding nematodes increased. Omnivorous and predacious nematodes showed the most sensitive response, becoming extinct when the Cu concentrations were between 0.8-1.4 mg l<sup>-1</sup>.

In Chapters 6 and 7 the nematode community structure and some community parameters, such as the Maturity Index, are discussed in the light of their potential for future risk assessment of soil pollution.

Based on the present results, it is concluded that nematodes offer excellent perspectives to assess effects of pollutants at the community level. Nematode community parameters, such as the Maturity Index and the distribution between different trophic groups, gave an early and sensitive signal of increased Cu, Ni or Zn pollution in the soil. Moreover, it was demonstrated that the nematode structure may also provide opportunities to identify specific types of disturbance, i.e. pollutants.

## Samenvatting

nematodengemeenschap veranderde bij toenemende concentraties Cu, Ni en Zn, maar niet bij Cd waarvan de hoogste onderzochte concentratie 160 mg kg<sup>-1</sup> was. EC<sub>50</sub> waarden voor een 50% reductie in de populatie van individuele nematodentaxa gaven kleine intra-taxon verschillen voor Cu, Ni and Zn te zien, wat aantoont dat voor deze metalen het uiteindelijke effect vergelijkbaar is binnen eenzelfde geslacht. Er werden echter wel grote verschillen gevonden tussen de gevoeligheid van verschillende taxa. In een aantal experimenten bleek bijvoorbeeld dat de populatie omnivoren en carnivoren, die kenmerken bezitten van K-strategen, al significant verlaagd waren bij concentraties Cu, Ni en Zn van 100 mg kg<sup>-1</sup>. Relatieve verschuivingen binnen de levensstrategiegroepen, en in mindere mate ook binnen de voedselgroepen, waren indicatief voor metaal-geïnduceerde veranderingen in de bodem. Deze classificaties waren echter niet geschikt genoeg om de gevoeligheid van alle nematodensoorten te voorspellen.

In hoofdstuk 3 werd ook gebruik gemaakt van de 'natuurlijke grond methode' om de lange-termijn effecten van koper en zink (0, 25, 50, 100, 200 and 400 mg kg<sup>-1</sup>) in relatie tot de aan- of afwezigheid van engels raaigras (*Lolium perenne* L.) te bestuderen. Na 1 jaar bleek de aanwezigheid van vegetatie een belangrijke factor in het uiteindelijke ecotoxicologische effect van Cu en Zn. In grond begroeid met *L. perenne* werden de effecten van Cu en Zn pas duidelijk bij hogere metaalconcentraties, waren minder sterk en kwamen vaker op een indirecte manier tot stand. Daarom is het raadzaam om in de toekomst de ecotoxicologische effecten van verontreinigende stoffen op nematoden in de aanwezigheid van vegetatie te onderzoeken.

In hoofdstuk 4 werd een ander belangrijk aspect van verontreinigingen onderzocht, namelijk het gelijktijdig voorkomen van meerdere toxische stoffen. De biologisch beschikbare concentraties Cu en Zn, gemeten nadat de grond was geëxtraheerd met 0.01 M CaCl<sub>2</sub>, toonden aan dat de metaalconcentraties in grond met Cu en Zn niet significant verschillend waren ten opzichte van grond waar slechts één metaal aanwezig was. Na een half jaar waren veel gemeenschapsparameters en nematodenpopulaties significant beïnvloed door toenemende concentraties Cu en Zn tot maximaal 200 mg kg<sup>-1</sup>. De effecten op nematoden die gelijktijdig aan Cu en Zn waren blootgesteld waren echter altijd additief of antagonistisch.

In hoofdstuk 5 werden de lange-termijn effecten van Cu en pH op nematoden onderzocht. Dit vond plaats onder de meest realistische omstandigheden, namelijk in een akker. Zowel Cu als pH hadden een grote invloed op nematoden. In het algemeen was het effect van Cu groter bij afnemende bodem-pH. De laagste Cu-concentratie, waarbij een significant negatief effect op het totaal aantal nematoden werd gevonden, was 0.32 mg l<sup>-1</sup>.

De soortsaamenstelling en verdeling tussen de verschillende voedselgroepen waren zelfs gevoeliger dan het totaal aantal nematoden. Combinaties van een hoge Cu concentratie en lage pH gaven een significante verlaging van het aantal bacterie-eters, terwijl het aantal schimmel-eters toenam. Omnivoren en carnivoren waren het meest gevoelig en verdwenen bij Cu-concentraties tussen de 0.8-1.4 mg l<sup>-1</sup>.

In hoofdstuk 6 en 7 worden de structuur van een nematodengemeenschap en sommige gemeenschapsparameters, zoals de Maturity Index, besproken met betrekking tot hun geschiktheid om verontreinigde bodems te beoordelen. Gebaseerd op de huidige resultaten luidt de conclusie dat nematoden zeer geschikt zijn om de effecten van verontreinigingen op het niveau van de levensgemeenschap te beoordelen. Parameters zoals de Maturity Index en de verdeling tussen de verschillende voedselgroepen, gaven een snel en gevoelig signaal bij verhoogde concentraties Cu, Ni en Zn in de bodem. Daarnaast werd er aangetoond dat de structuur van een nematodengemeenschap ook mogelijkheden biedt om verschillen tussen stress-factoren, waaronder giftige stoffen, op te sporen.

## Samenvatting

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