

QBOL: Development of a new diagnostic tool using DNA barcoding to identify quarantine organisms in support of plant health

Peter Bonants, EMBL-EBI, 20 oct 2008



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Introduction

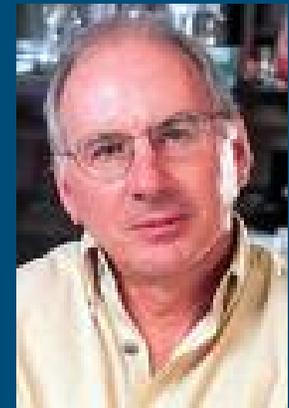
- What is DNA barcoding?
- Examples
- EU Call FP7
- Consortium / Origin
- Which genes?
- Plant Health: QBOL
- Future



The **Consortium for the Barcode of Life** (CBOL) is an international initiative devoted to developing DNA barcoding as a global standard for the identification of biological species. [DNA barcoding](#) is a new technique that uses a short DNA sequence from a standardized and agreed-upon position in the genome as a molecular diagnostic for species-level identification

CBOL has more than 150 [Member Organizations](#) from more than 45 countries including:

- Natural history museums, zoos, herbaria, and botanical gardens;
- University departments of biology and molecular biology;
- Biodiversity and conservation organizations, NGOs;
- Governmental and intergovernmental organizations; and
- Private biotech companies.

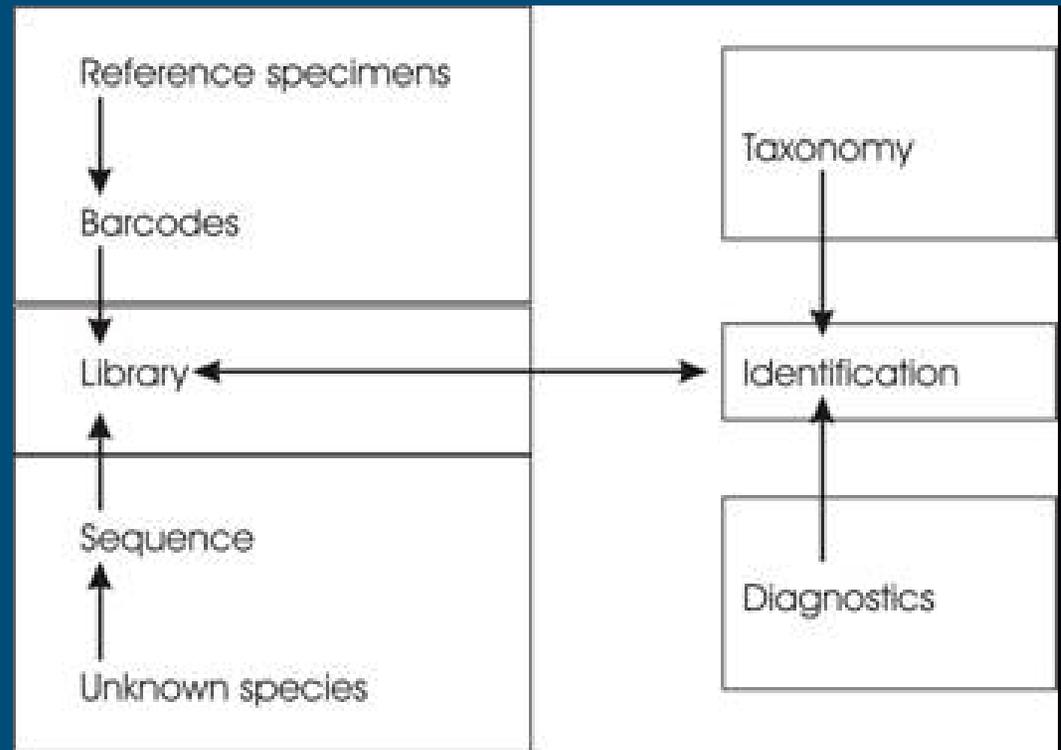


Paul Hebert

DNA Barcoding

DNA Barcoding projects have four components:

- *The Specimens*
- *The Laboratory Analysis*
- *The Database*
- *The Data Analysis*



Major Barcoding Campaigns



Polar Bears



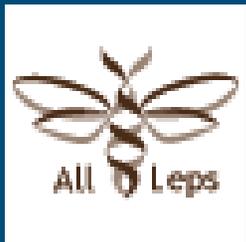
All Birds



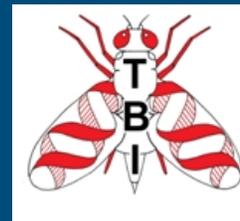
Bats



Fish



Lepidoptera



Tephritid





marine barcode of life

[MarBOL Blog](#)[Partners](#)[MarBOL Team](#)[MarBOL Project](#)[Species Checklist](#) | [Progress Report](#) | [Protocols](#) | [Upcoming Meetings](#)

Meetings

Upcoming Meetings

9th International Meeting - Oct 30, 2008

MEEGID IX, University of California at Irvine, 30 October - 1 November 2008

Communications on genetics, genomics, proteomics, phylogenetics, population biology, mathematical modeling, and bioinformatics are welcome. They can report on the host, the pathogen, or the vector for vector-borne diseases.

Papers considering host + pathogen or pathogen + vector (co-evolution) are particularly encouraged. All pathogens are within the scope of MEEGID: viruses, parasitic protozoa, helminths, fungal organisms, and prions. All infectious models can be explored, including those of veterinary or agronomical relevance.

progress

specimens barcoded

26361

species barcoded

4993

[View detailed progress reports](#)



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FP7 in brief / Specific Programmes

***Cooperation* – Collaborative research**

***Ideas* – Frontier Research**

***People* – Human Potential**

***Capacities* – Research Capacity**



FP7 in brief / Cooperation

- 10 themes:
 - Health
 - **Food, Agriculture and Biotechnology**
 - ICT
 - Nanosciences
 - Energy
 - Environment (incl. climate change)
 - Transport
 - Socio-economic sciences and Humanities
 - Security
 - Space



Text Call FP7-KBBE-2008-2B

KBBE-2008-1-4-01: Development of new diagnostic methods in support of Plant Health policy Call: FP7-KBBE-2008-2B

This Project will build a **sustainable diagnostic resource** to enable ‘**DNA-barcode identification**’ ultimately for **all quarantine plant pests or pathogens** of statutory importance. Key work will include: obtaining or producing relevant vouchered sequence data for individual pests or pest groups and position them in a correct taxonomic context, developing generic diagnostic tools based on these barcode sequences; linking vouchered sequence information to published biological information; developing strategic approaches and methodologies to enable the establishment of DNA banks and access to digital voucher specimens.

Funding scheme: Small collaborative project.



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FP7: Call KBBE-2008-2B

Proposal full title:

Development of a new diagnostic tool using DNA barcoding of Quarantine organisms in support of plant health

Proposal acronym:

QBOL



Type of funding scheme: Small Collaborative Project



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Targets Quarantine

■ Which?

- Fungi
- Arthropods
- Bacteria
- Nematodes
- Viruses
- Phytoplasmas

Council Directive 2000/29/EC
EPPO list A1 and A2



State of the Art

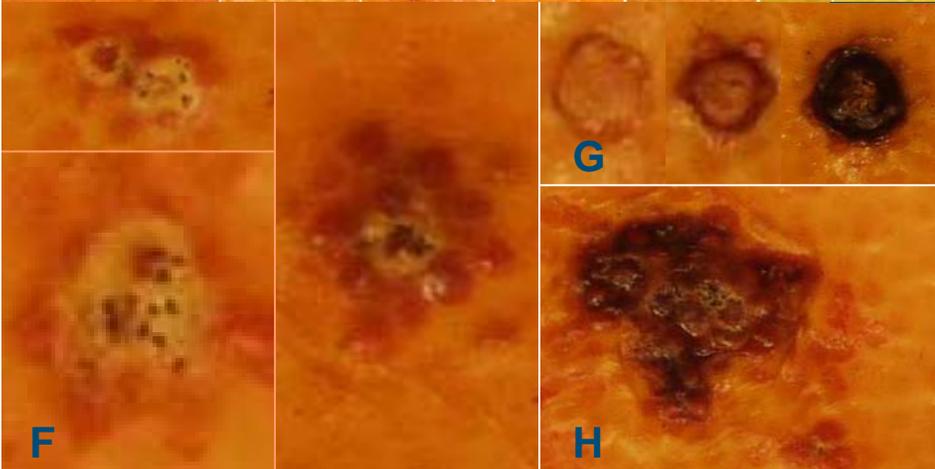
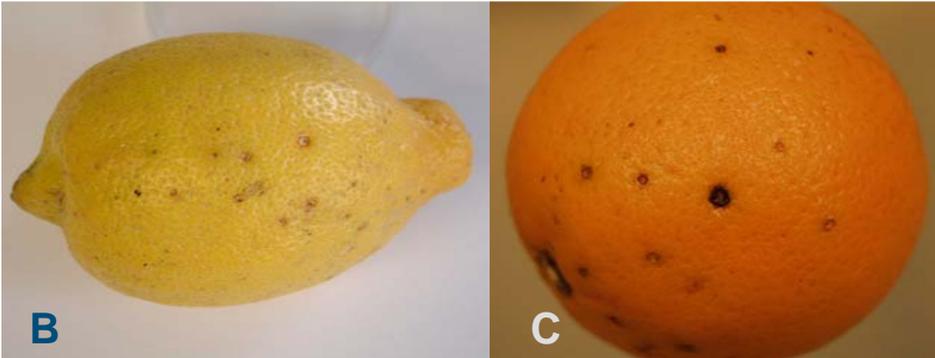
- *Collections and Taxonomic Experience*
- *Quarantine organisms on EU Directive and EPPO list*
- *Sequencing of Q-organisms*
- *Identification methods*
- *DNA Barcoding projects*
- *Databases*



Fungi

Guignardia citricarpa

Phytophthora ramorum



Barcoding Q Arthropods

OBJECTIVES

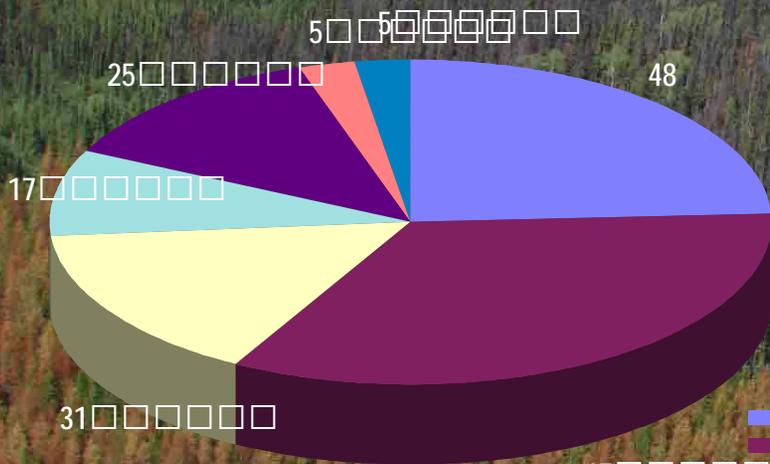
1. generate 5-10 barcode sequences (Cox1 and ITS) for about 100 species of Q arthropods and 50 closely related species
2. establish a prioritized list of all EU Q arthropods
3. collect all Q arthropods from different localities and crops
4. develop DNA extraction procedures for larvae and adults
5. develop and test primers for PCR amplification
6. collect DNA barcode sequence data
7. test the usefulness of this dataset





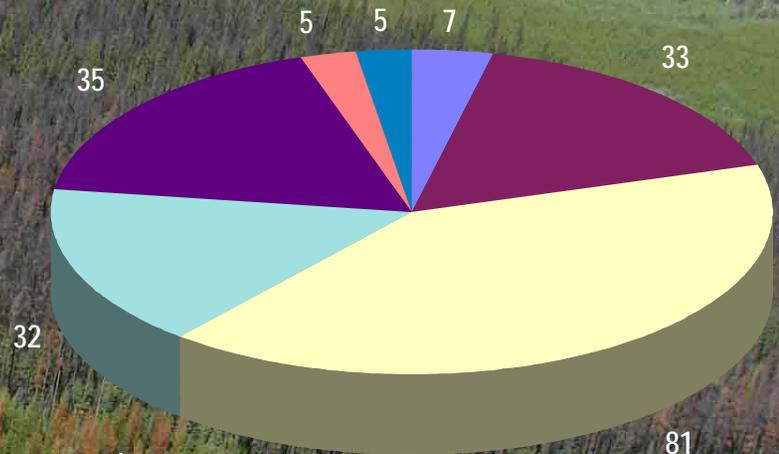
Barcoding Q Arthropods

Q pest origin



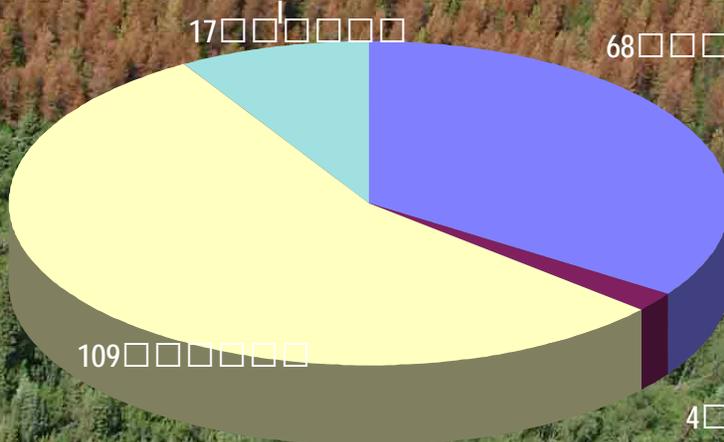
- Palearctic
- Nearctic
- Neotropical
- Afrotropical
- Oriental
- Australasia
- Undetermined

Q arthropod orders

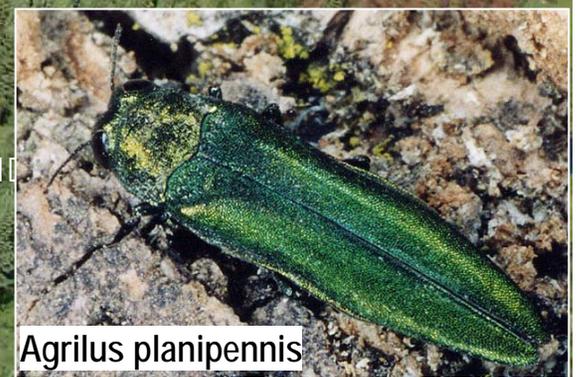


- Acari
- Lepidoptera
- Coleoptera
- Hemiptera
- Diptera
- Hymenoptera
- Thysanoptera

Crop types



Crop types



Insects



Disciplin Entomology PD-NRL



DNA barcodes for biosecurity: invasive species identification

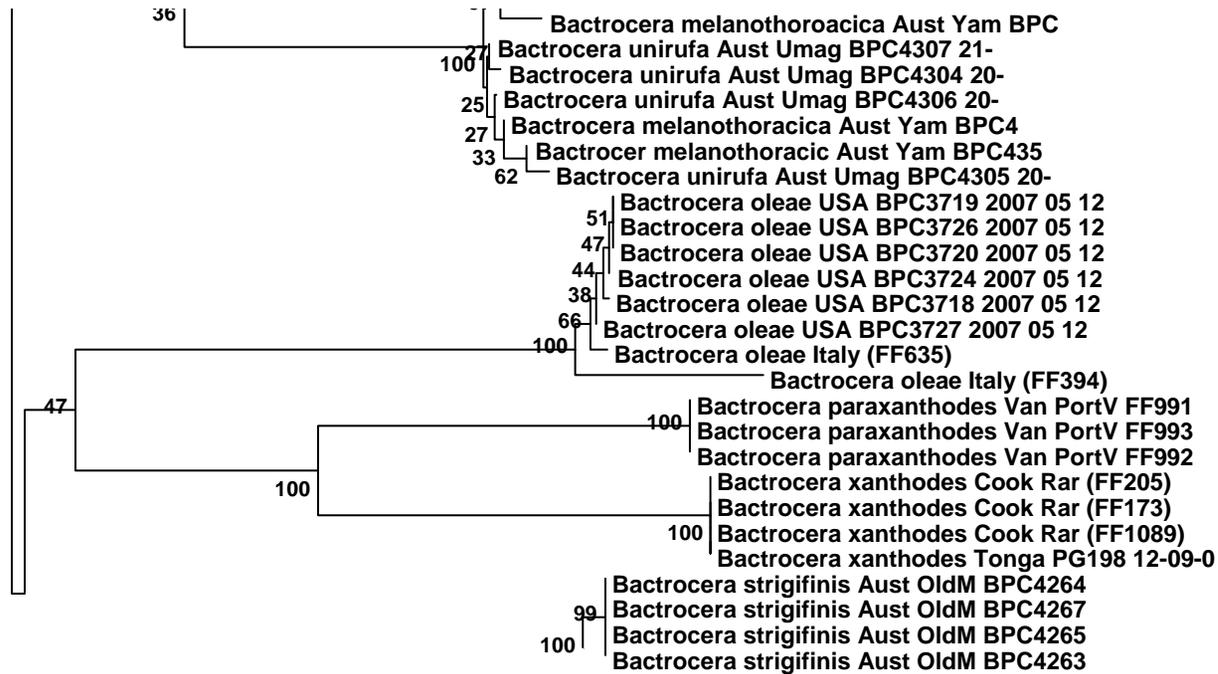
K. F. Armstrong* and S. L. Ball

*National Centre for Advanced Bio-Protection Technologies, PO Box 84,
Lincoln University, Canterbury, New Zealand*

Biosecurity encompasses protecting against any risk through ‘biological harm’, not least being the economic impact from the spread of pest insects. Molecular diagnostic tools provide valuable support for the rapid and accurate identification of morphologically indistinct alien species. However, these tools currently lack standardization. They are not conducive to adaptation by multiple sectors or countries, or to coping with changing pest priorities. The data presented here identifies DNA barcodes as a very promising opportunity to address this. DNA of tussock moth and fruit fly specimens intercepted at the New Zealand border over the last decade were reanalysed using the *cox1* sequence barcode approach. Species identifications were compared with the historical dataset obtained by PCR–RFLP of nuclear rDNA. There was 90 and 96% agreement between the methods for these species, respectively. Improvements included previous tussock moth ‘unknowns’ being placed to family, genera or species and further resolution within fruit fly species complexes. The analyses highlight several advantages of DNA barcodes, especially their adaptability and predictive value. This approach is a realistic platform on which to build a much more flexible system, with the potential to be adopted globally for the rapid and accurate identification of invasive alien species.

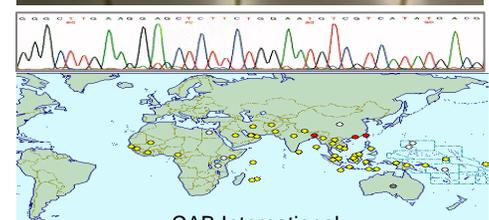
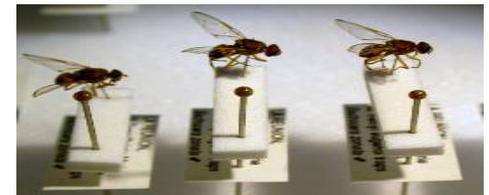
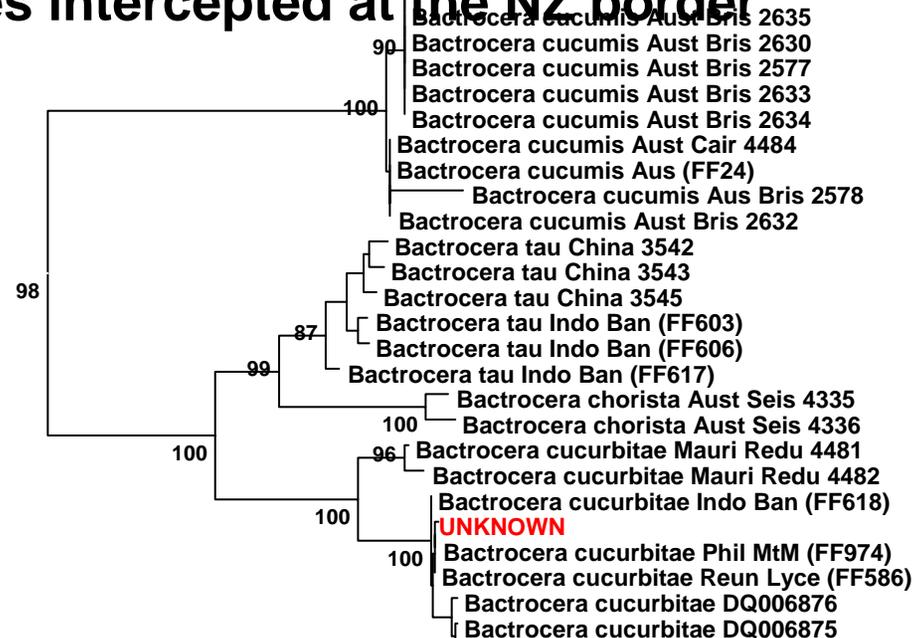
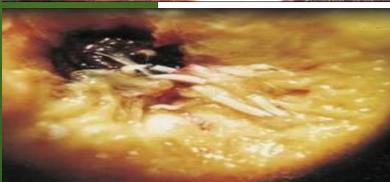
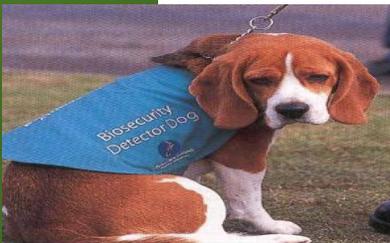
Keywords: mitochondrial DNA; cytochrome oxidase subunit I; COI; molecular diagnostics; quarantine; insects





DNA barcodes to identify fruit fly immature life stages intercepted at the NZ border

Karen Armstrong
Lincoln University



Combined EU / EPPO list

Q and Q-alert bacteria

Annex I	Part A	Section I
	1 <i>Xyllela fastidiosa</i>	
Annex I	Part A	Section II
	1 <i>Clavibacter michiganensis</i> ssp. <i>sepedonicus</i>	
	2 <i>Pseudomonas solanacearum</i> = <i>Ralstonia solanacearum</i> (race 1, race 3)	
Annex I	Part B	
Annex II	Part A	Section I
	1 <i>Erwinia stewartii</i>	
	2 <i>Xanthomonas campestris</i> strains pathogenic to Citrus (as <i>Xanthomonas axonopodis</i> pv. <i>citri</i>)	
	3 <i>Xanthomonas campestris</i> pv. <i>oryzae</i> & <i>oryzicola</i> = <i>Xanthomonas oryzae</i> pv. <i>oryzae</i> & <i>oryzicola</i>	
Annex II	Part A	Section II
	1 <i>Clavibacter michiganensis</i> ssp. <i>insidiosus</i>	
	2 <i>Clavibacter michiganensis</i> ssp. <i>michiganensis</i>	
	3 <i>Erwinia amylovora</i>	
	4 <i>Erwinia chrysanthemi</i> pv. <i>dianthicola</i> = <i>Dickeya dianthicola</i>	
	5 <i>Pseudomonas caryophylli</i> = <i>Burkholderia caryophylli</i>	
	6 <i>Pseudomonas syringae</i> pv. <i>persicae</i>	
	7 <i>Xanthomonas campestris</i> pv. <i>phaseoli</i> <i>Xanthomonas arboricola</i> pv. <i>corylina</i> (EPPO list)	
	8 <i>Xanthomonas campestris</i> pv. <i>pruni</i> = <i>Xanthomonas arboricola</i> pv. <i>pruni</i>	
	9 <i>Xanthomonas campestris</i> pv. <i>vesicatoria</i> = <i>Xanthomonas vesicatoria</i> & <i>Xanthomonas axonopodis</i> pv. <i>vesicatoria</i>	
	10 <i>Xanthomonas fragariae</i>	
	11 <i>Xylophilus ampelinus</i>	
Annex II	Part B	
	1 <i>Curtobacterium flaccumfaciens</i> pv. <i>flaccumfaciens</i>	
Q-alert EPPO		
	<i>Xanthomonas axonopodis</i> pv. <i>dieffenbachiae</i>	
	<i>Xanthomonas arboricola</i> pv. <i>fragariae</i>	
	<i>Xanthomonas axonopodis</i> pv. <i>allii</i>	

Bacteria



Xanthomonas fragariae



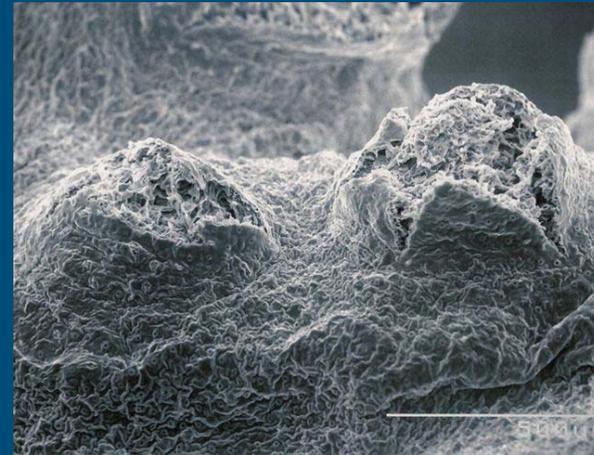
Clavibacter michiganensis subsp. *sepedonicus*



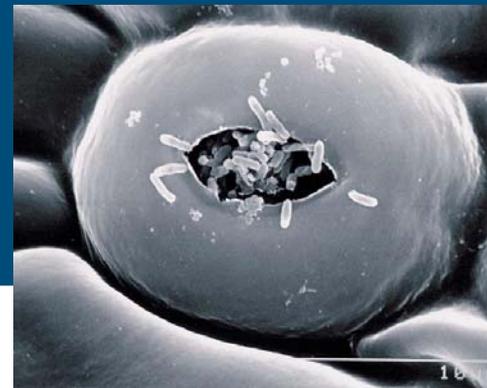
Ralstonia solanacearum



Bacteria



Citrus Canker, *Xanthomonas citri* subsp. *citri*



Nematodes



Pine wood nematode



Root knot nematode



Viruses

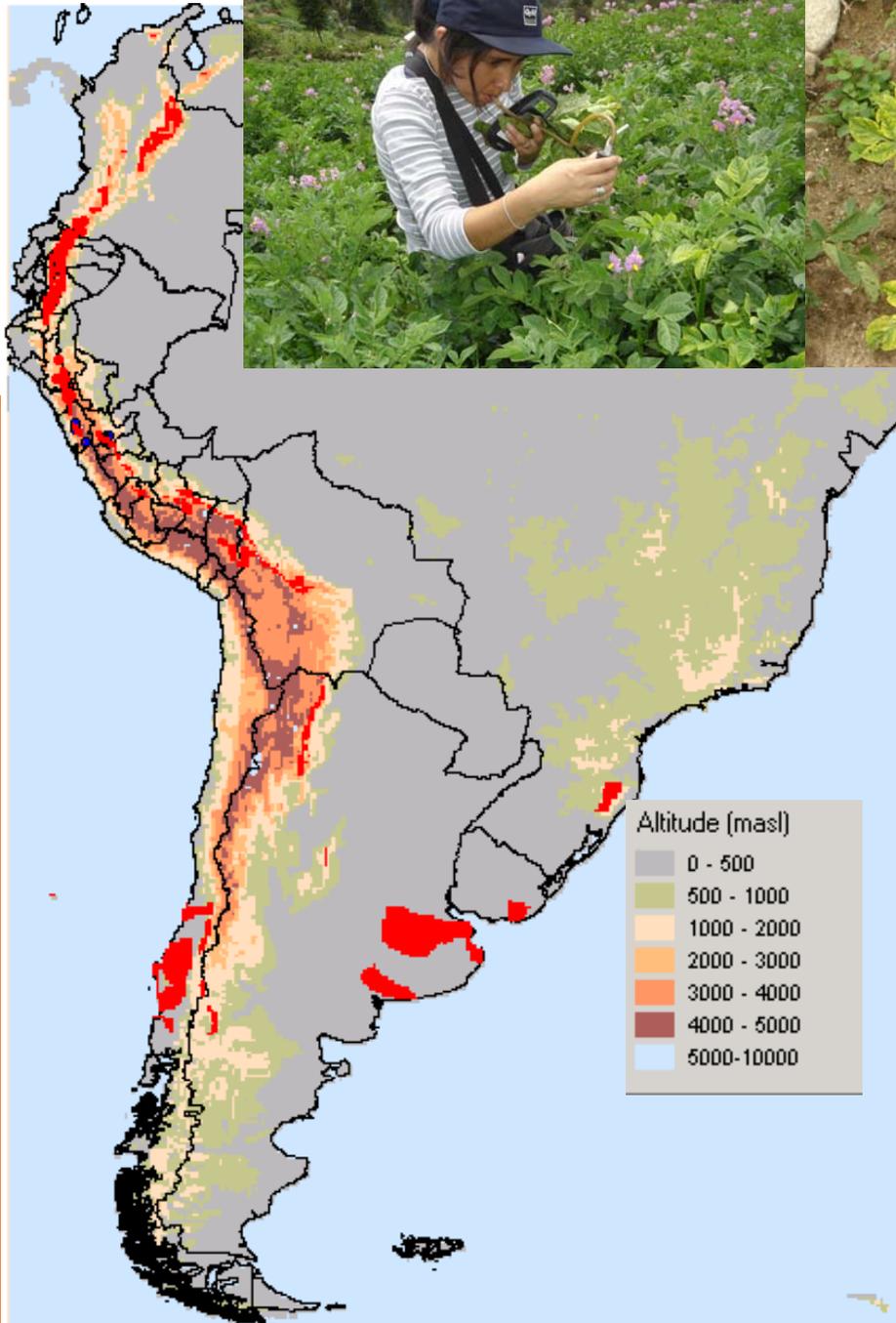


Alstroemeria Yellow Spot Topovirus



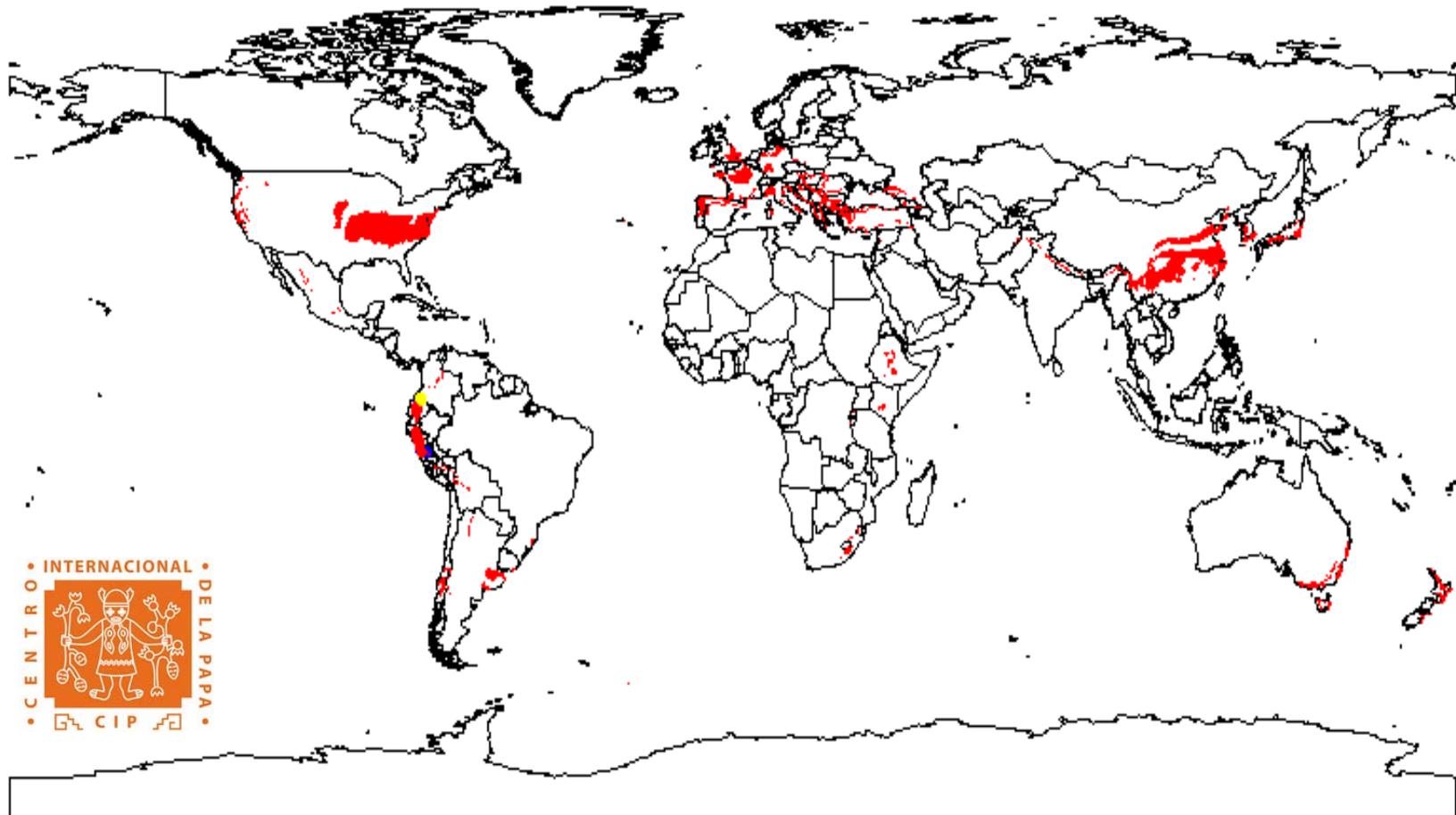
Prunus Necrotic Ringspot Virus





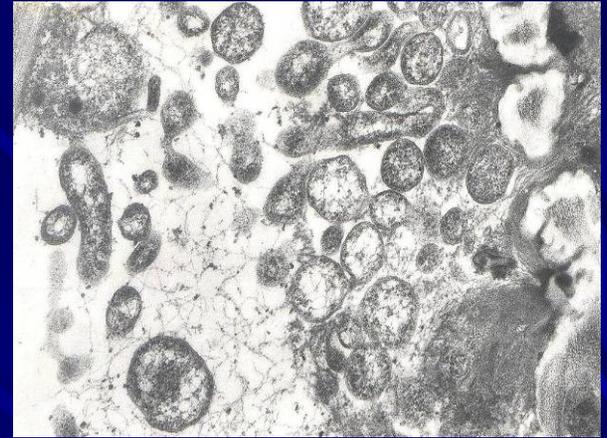
- GPS survey work, laboratory diagnosis and GIS modeling predicts *Potato yellow vein virus* (a model system for emergent disease) threat to Southern Peru and Bolivia.
- Relevant National bodies alerted.
- Climate change scenarios indicate increase in range

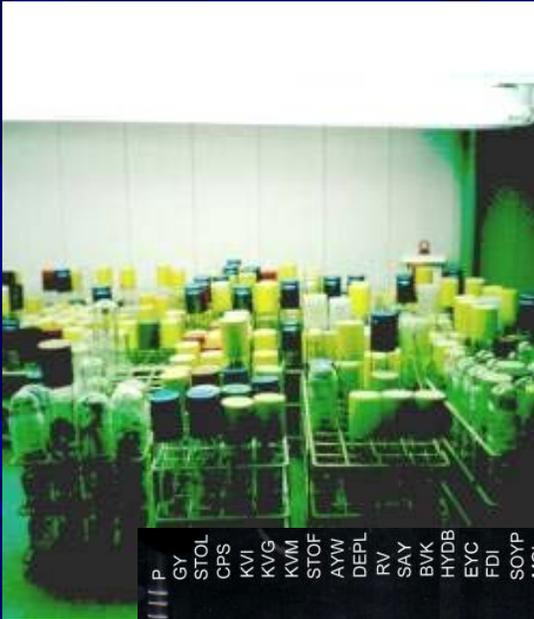
Current global potential threat of establishment of PVV.



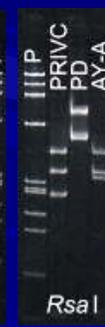
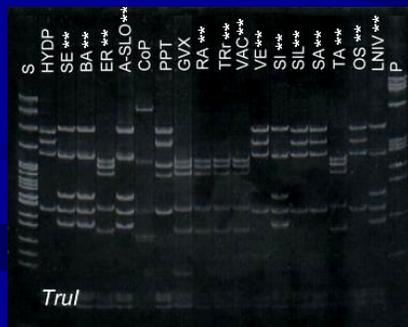
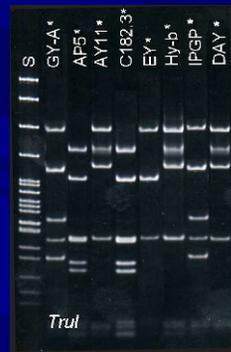
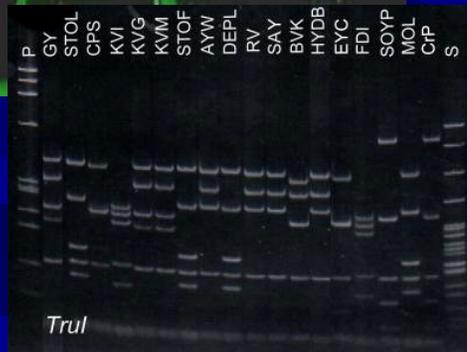
1. What are Phytoplasmas?

- Phloem-limited insect-transmitted plant pathogenic bacteria that lack a cell wall
- Related to Gram positive bacteria (*Bacillus*) do not encode typical genes related to pathogenicity present in other plant pathogenic bacteria
- Can not be grown in axenic culture
- Genomes of some have been sequenced (sizes range from 530 – 1200 kb) they are the smallest known self-replicating life form
- Symptoms include yellowing, phyllody, proliferation, stunting, general decline, witches' broom



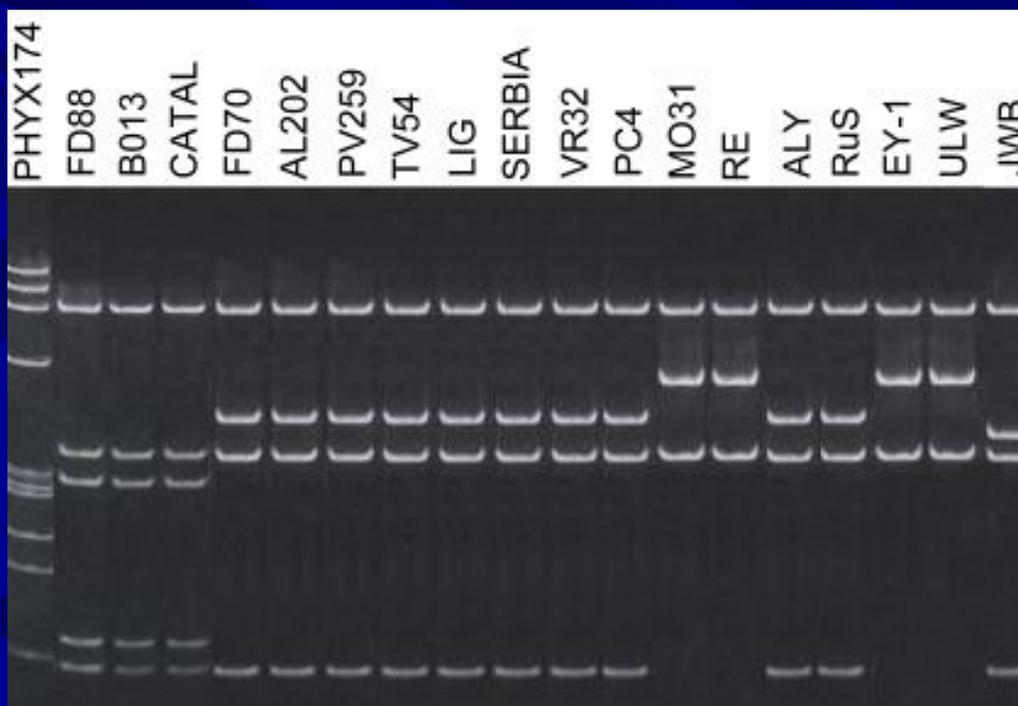


Collection of in periwinkle micropropagated phytoplasma strains

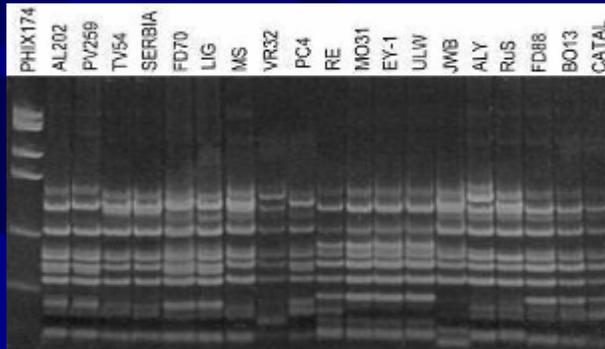


http://www.dista.unibo.it/person/collectionseptember_2003.pdf

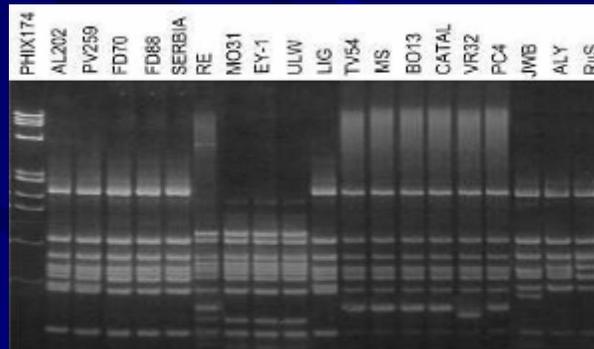
Polymorphisms as shown by RFLP analyses with selected restriction enzymes on three genes of different strains of phytoplasmas related to *Flavescence dorée*



TaqI on 16Sr gene

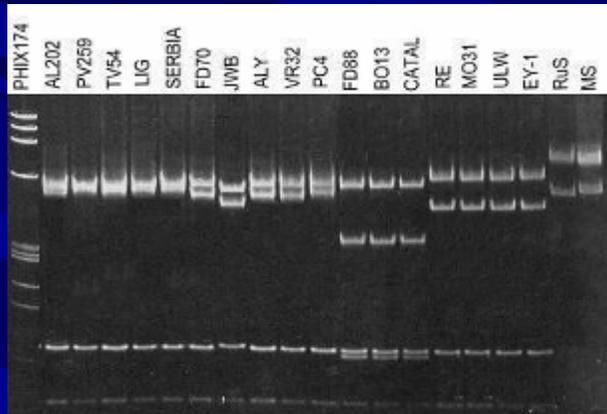


TruI

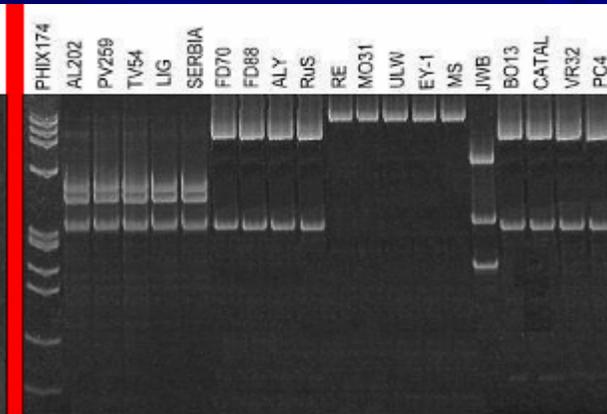


Tsp509I

rpS3 gene

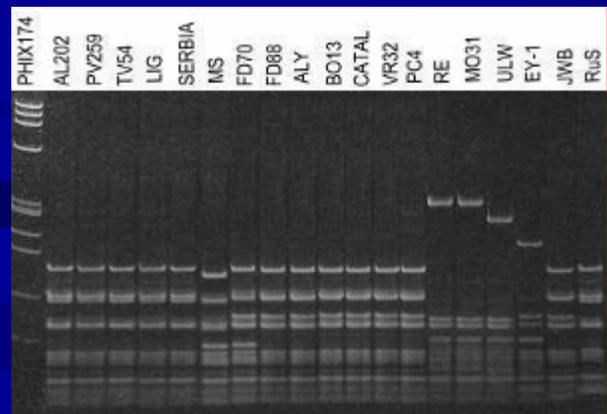


AluI

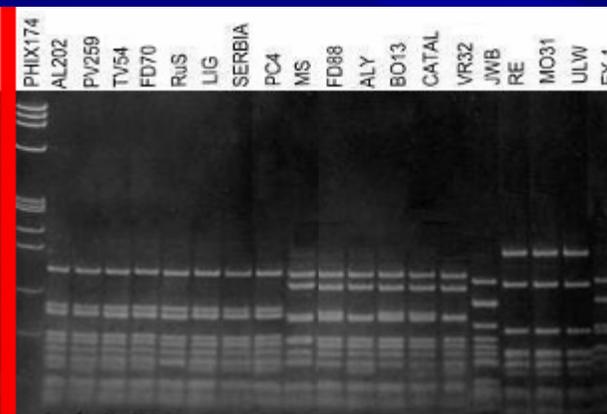


TaqI

SecY gene



TruI



Tsp509I

Three principle QBOL Objectives

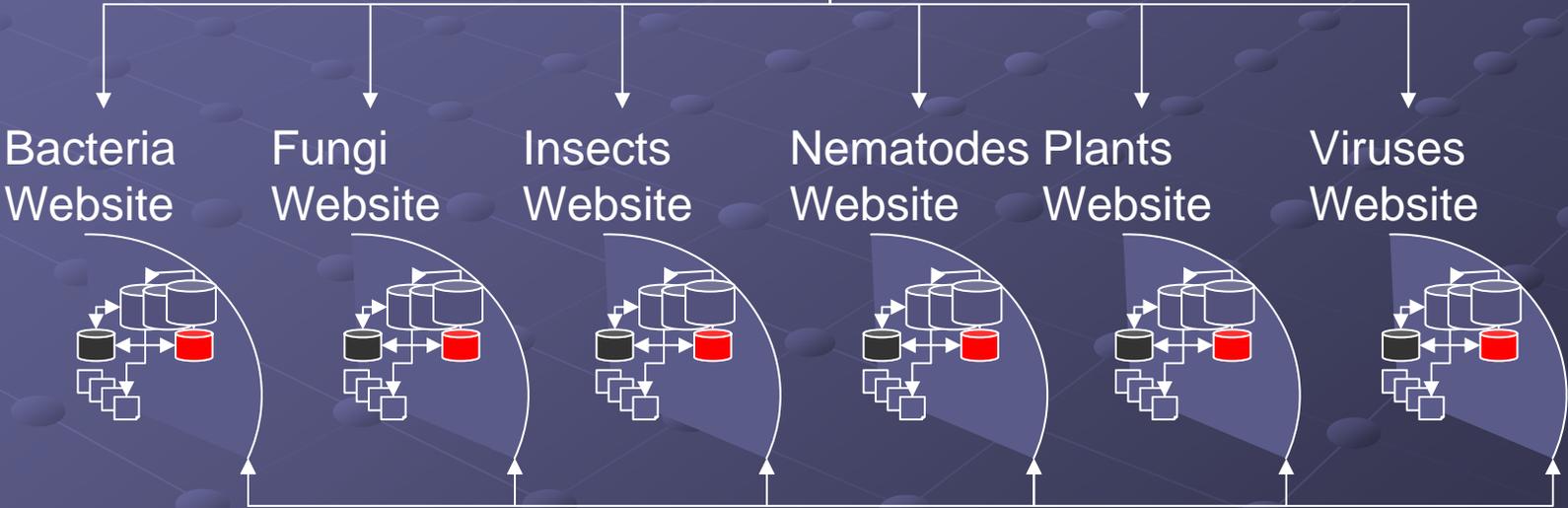
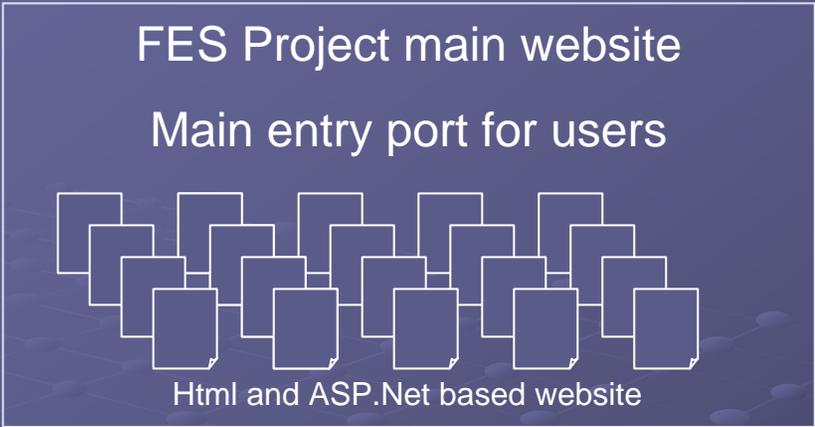
- *to DNA barcode relevant Q-organisms + morphologically and/or taxonomically related organisms*
- *to develop a database of DNA barcode sequences plus relevant taxonomic/geographic/host data*
- *to develop a DNA bank for the selected set of Q-organisms + morphologically and/or taxonomically related organisms*



Databases

- BOLD
- GenBank, EMBL
- FES Programme (NL)
 - *BioMICS*
 - *Mycobank* (www.mycobank.org)





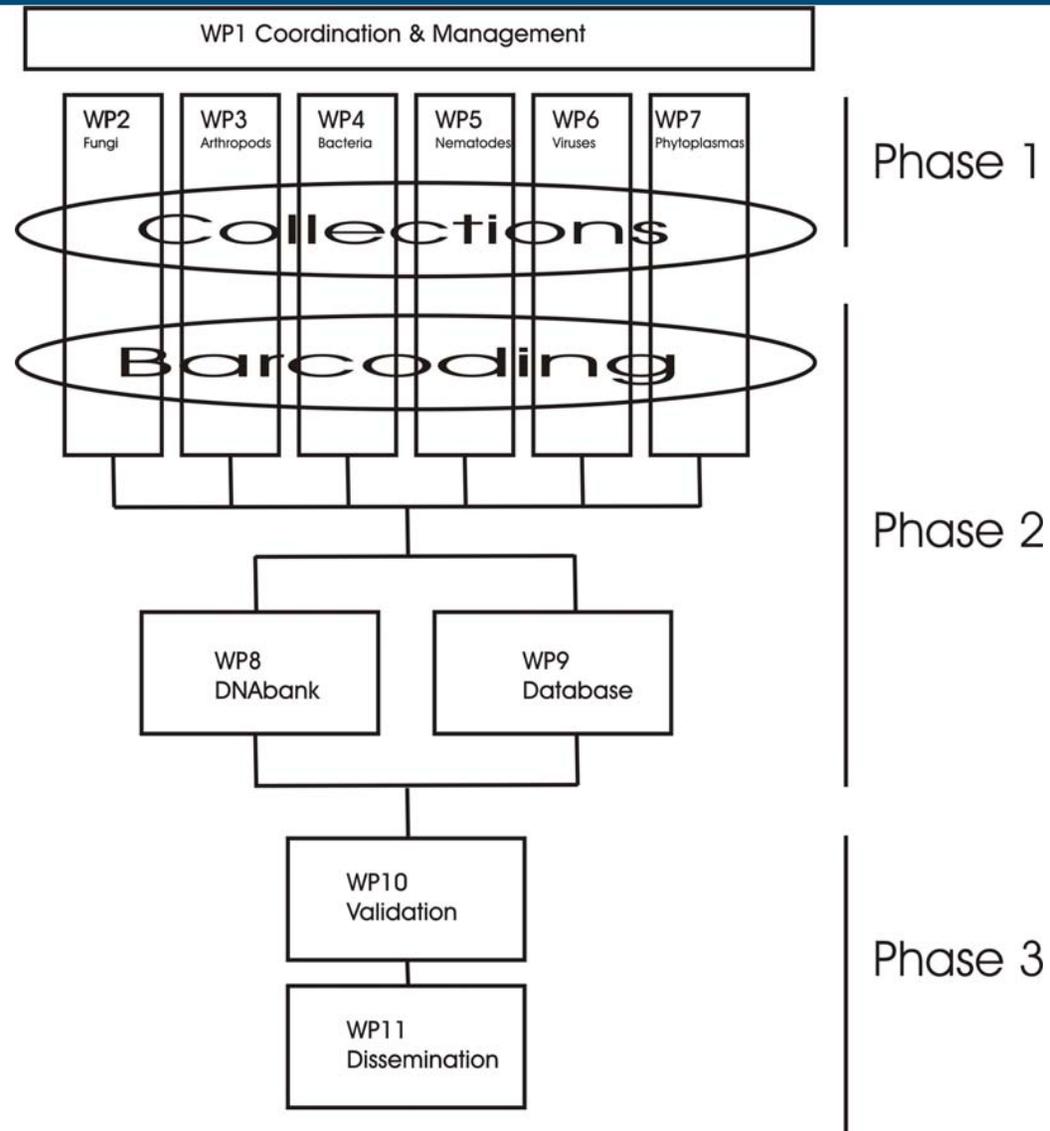
Real linking of databases not only via hyperlinks. Direct access possible

Tailor-made
website for FES

BioIMICS Net

BioIMICS
Main software

Work Packages QBOL



Partners QBOL 1

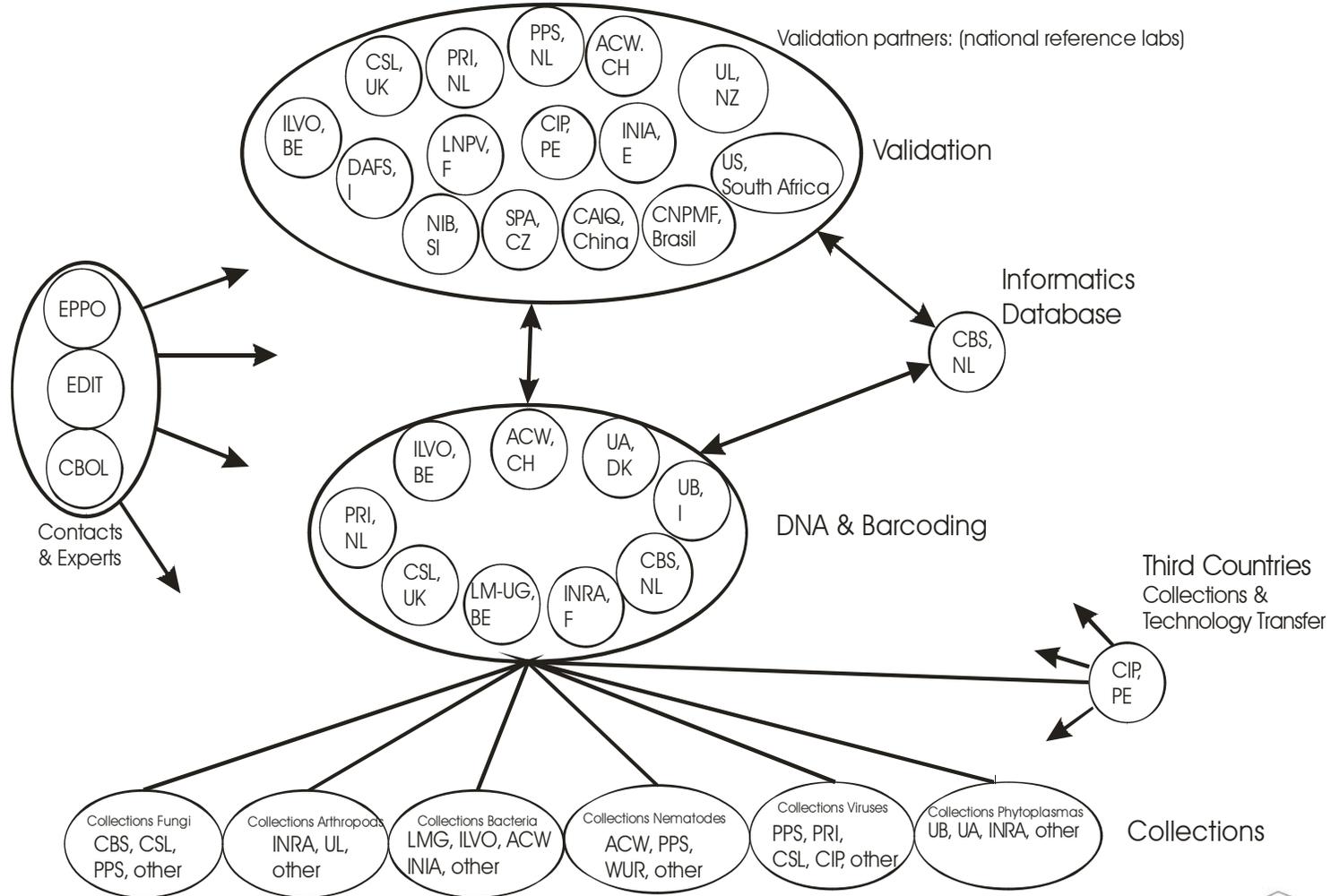
Plant Research International	(PRI, The Netherlands)
Central Science Laboratory	(CSL, United Kingdom)
Institute for Agricultural and Fisheries Research	(ILVO, Belgium))
Laboratory of Microbiology, Univ Gent	(LM-UG, Belgium)
Agroscope Changins-Wädenswil Research Station	(ACW, Switzerland)
INRA - Centre de Biologie et de Gestion des Populations	(INRA, France)
University of Aarhus	(UA, Denmark)
University of Bologna	(UB, Italy)
Centraalbureau voor Schimmelcultures	(CBS, The Netherlands)
International Potato Center	(CIP, Peru)
Plant Protection Service	(PPS, The Netherlands)

Partners QBOL 2

National Institute of Biology	(NIB, Slovenia)
State Phytosanitary Administration	(SPA, Czech Republic)
French national laboratory for plant health	(LNPV, France)
Chinese Academy of Inspection and Quarantine	(CAIQ, China)
Lincoln University, Bio-Protection Research Centre	(LU, New Zealand)
Embrapa Mandioca e Fruticultura Tropical	(CNPMPF, Brasil)
Dept. Agricultural and Food Sciences	(DAFS, Italy)
	(INIA, Spain)
University of Stellenbosch	(US, South Africa)

Consortium OBOI

CONSORTIUM QBOL



Advisory Board QBOL

EPPO: European Plant Protection Organisation

Dr. Francoise Petter

BOLD: Barcoding of Life Database

Dr. Paul Hebert

Deliverables QBOL

Culture Collections and Taxonomic Experience

A worldwide network of relevant culture collections containing selected Q-organisms and the taxonomic expertise related to those collections.

Quarantine organisms EU Directive, EPPO list (the EPPO pest lists include 298 pests)

Many of the quarantine or regulated organisms of the EU Directive / EPPO list will be addressed. Priorities will be made based upon availability of target specimens and closest relatives in different regions in the world.

Deliverables QBOL

Sequencing of Q-organisms

Multi gene analysis will be executed on selected quarantine/ regulated organisms

Barcoding projects BOLD

Close contact with BOLD is guaranteed and data will be shared with BOLD

DNAbank

A DNA bank will be build containing DNA samples of barcoded quarantine/regulated organisms and their taxonomically related species

Deliverables QBOL

Database

A web based Database system will be build containing all relevant DNA barcodes and freely accessible by the end-users

Validation

Developed protocols and the database containing relevant DNA barcodes + taxonomic data will be validated by end-users

Implementation- Dissemination

End-users will be trained, developed protocols and the database will be distributed, a workshop/symposium will be organized. Effort will be put on implementation of Database into an (inter)national platform within and outside EU laboratories.

QBOL: Barcoding for Plant Health

- Genes with sequence difference between Q-organisms and closely related organisms
- Easy to amplify with generic primers
- Culture collections
- Taxonomic experience
- Accessible Database
- Development of ID and DET methods
- Validation – EPPO/IPPC
- Implementation – PPS

The Future



Acknowledgements

- Karen Armstrong, Lincoln University, New Zealand
- Jean-Yves Rasplus, INRA, France
- Martine Maes, ILVO and Paul de Vos, LM-UG, Belgium
- Neil Boonham, CSL, UK
- Hans de Gruyter/Linda Kox, PPS, The Netherlands
- Jaime Cubero, INIA, Spain
- Juerg Frey/Sebastian Kiewnick , ACW, Switzerland
- Assunta Bertaccini, UB, Italy
- Ian Barker, CIP, Peru
- QBOL partners



Thank you very much for your attention



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