Aetiological study of the presumed ocular histoplasmosis syndrome in the Netherlands


Abstract

Aim—To investigate whether presumed ocular histoplasmosis syndrome in the Netherlands is caused by Histoplasma capsulatum and whether other risk factors might play a role in the pathogenesis of this syndrome.

Methods—23 patients were clinically diagnosed as having presumed ocular histoplasmosis syndrome based on the following criteria: peripapillary atrophy, punched out lesions, a macular disciform lesion or scar in one eye without vitritis. As controls, 66 sex and age matched healthy volunteers were used. Serum samples from both patients and controls were tested for the presence of antibodies against H capsulatum, Toxoplasma gondii, Toxocara canis et cati, Ascaris sp, and for the presence of antigens of Cryptococcus neoformans. Serum samples were also tested for the presence of autoantibodies against retinal or choroidal proteins. To investigate other risk factors, patients and controls were asked to fill in a health and travel related questionnaire. Ten patients with ocular toxoplasmosis were used as a disease control group.

Results—None of the patients with presumed ocular histoplasmosis syndrome or controls had circulating antibodies directed against H capsulatum. No risk factors could be identified and no indications for autoimmunity and no evidence for the role of the other infectious agents could be demonstrated.

Conclusions—In a Dutch group of patients fulfilling the criteria of a disease currently named presumed ocular histoplasmosis syndrome, no risk factors or relation with the fungus H capsulatum could be detected.

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Presumed ocular histoplasmosis is a distinct syndrome characterised by a macular disciform lesion or scar caused by choroidal neovascularisation, peripapillary and choroidal scarring, and multiple, often sharply circumscribed white atrophic scars (punched out lesions) in combination with a clear vitreous (Fig 1A and B). This syndrome is mainly seen in the midwest of the United States and is thought to be caused by Histoplasma capsulatum, endemic in this area. In Europe a clinically similar syndrome has been observed in areas where the infectious agent H capsulatum is absent.
of Utrecht and Amsterdam as suffering from presumed ocular histoplasmosis. Only patients with both fundus photography and data concerning the ophthalmic examination and therapy were included. All patients had to exhibit a macular disciform lesion or scar caused by a choroidal neovascularisation, peri-papillary and chorioretinal scarring, and multiple, often sharply circumscribed white atrophic scars (punched out lesions) in combination with a clear vitreous.

Fundus photographs were judged by an experienced ophthalmologist (MdS) in a masked fashion; fundus photographs from patients were mixed in a file with fundus photographs of patients with other macular disciform lesions. Patients were included in the study when the fundus pictures were judged as typical as described above.

From the patients who were diagnosed as having presumed ocular histoplasmosis syndrome, a serum sample was obtained and stored at the serum bank at the Netherlands Ophthalmic Research Institute.

Patients who were immunocompromised, or who suffered from any ocular disease that may mimic this syndrome, such as high myopia, age related macular degeneration, and other white dot syndromes were excluded from this study.

CONTROLS

Two control groups were used: group I consisted of 66 sex and age matched healthy volunteers; group II contained 10 patients diagnosed as having a toxoplasma chorioretinitis by means of clinical diagnosis which had been confirmed by intraocular antibody production against Toxoplasma gondii (and in five of these 10 cases also DNA of T gondii was detected by polymerase chain reaction (PCR)). All subjects included in the study gave their informed consent to participate in this study.

METHODS

All patients and all healthy controls answered a detailed questionnaire. Questions were asked about travel to areas endemic for H capsulatum; possible contacts with infectious agents, use of untreated or poorly treated drinking water and swimming in endemic areas; contacts with pets or professional animal contacts; contact with soil and sand; eating habits particularly the consumption of raw meat, or the frequency of barbecuing. In addition, a medical history regarding systemic infectious diseases and previous blood transfusions was recorded.

LABORATORY TESTS

To exclude H capsulatum as a possible causative agent in our patients suffering from presumed ocular histoplasmosis, antibodies against this fungus were measured. Antibodies against Toxoplasma gondii, Ascaris sp, Toxocara canis et cati were tested since these parasites can also give symptoms of inflammation of the posterior segment. In order to exclude the presence of any other fungal infection, antigens of Cryptococcus neoformans were tested, since this fungus is also present in Europe. Antibodies against Candida sp were not tested, as Candida sp is endemic in this region and all sera were expected to be positive. Sera were not tested for antibodies against Blastomyces and Coccidioides because both these fungi are not present in Europe and no positive serology has been described in European patients.

The presence of antibodies against H capsulatum was tested by immunodiffusion (Fungal immunodiffusion system, Meridian diagnostics, Inc, OH, USA) following the manufacturer’s protocol. Immunodiffusion is highly specific for the detection of antibodies directed against H capsulatum. As a positive control, serum from a patient with an active systemic histoplasmosis was used (courtesy of Dr Wheat, Indianapolis, IN, USA).

The presence of Cryptococcus neoformans antigens was tested by a latex agglutination test (Crypto-LA test, Whampoule Laboratories, International Biological Labs, Inc, Cranbury, USA) according to the manufacturer’s instructions.

The presence of antibodies to T gondii was tested in a 1:20 dilution and the presence of antibodies directed to Toxocara canis et cati and Ascaris sp was tested in a 1:40 dilution. These antibodies were measured by an in house enzyme linked immunosorbent assay at the Diagnostic Laboratory for Infectious Diseases and Perinatal Screening, National Institute of Public Health and the Environment, Bilthoven, Netherlands. Tests were considered positive when the respective dilutions showed a signal.

To exclude a possible autoimmune component we also tested for the presence of autoantibodies against retinal or choroidal proteins. Retina and choroid were retrieved from human donor eyes. Total protein extracts were obtained by dissolving retina or choroid tissue in SDS-PAGE loading buffer containing 2% SDS and 5% β-mercapto-ethanol followed by boiling for 5 minutes. After centrifugation, the proteins were size fractioned by SDS-PAGE gel electrophoresis and followed by western blotting. Blots were incubated with a 1:50 dilution of sera from patients and controls. After 1 hour incubation, blots were washed and incubated with peroxidase labelled rabbit anti-human immunoglobulins (Dako) for 1 hour. Blots were stained with diaminobenzidine tetrahydrochloride (Sigma-Aldrich Chemie, Zwijndrecht, Netherlands) and photographed directly after the reaction was stopped. Sera that gave bands other than the control sera were considered positive.

To further investigate the presence of autoantibodies, cryosections of rat eyes were incubated with sera of 13 patients with presumed ocular histoplasmosis in a 1:100 dilution for 2 hours. After washing one cryosection was incubated with peroxidase labelled rabbit anti-human immunoglobulins and one with peroxidase labelled rabbit anti-human IgG. The cryosections were stained with diaminobenzidine tetrahydrochloride. Cryosections were independently judged by two observers. As a negative control a sample of pooled normal human sera was used, and as a positive control a serum sample was used
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Table 1  Participant characteristics

<table>
<thead>
<tr>
<th></th>
<th>Patients (n=23)</th>
<th>Controls (n=66)</th>
<th>Disease controls (n=10)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Medicinal age (years)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Female</td>
<td>40 (range 29–60)</td>
<td>42 (range 26–60)</td>
<td>46 (range 29–55)</td>
</tr>
<tr>
<td>Male</td>
<td>33 (range 28–48)</td>
<td>36 (range 26–44)</td>
<td>53 (range 36–65)</td>
</tr>
<tr>
<td>Mean age at start of symptoms</td>
<td>30 (SD 6.5) (range 18–44)</td>
<td>30 (SD 6.5) (range 18–44)</td>
<td>30 (SD 6.5) (range 18–44)</td>
</tr>
<tr>
<td>Sex</td>
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<tr>
<td>Male</td>
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<tr>
<td>Female</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Time interval between start of symptoms and sample collection (years)</td>
<td>8 (SD 6.7) (range 0.5–23)</td>
<td>16 F/7 M</td>
<td>49 F/17 M</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>7 F/3 M</td>
</tr>
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</table>

Table 2  Results of tests to detect antibodies

<table>
<thead>
<tr>
<th></th>
<th>H capsulatum</th>
<th>C neoformans*</th>
<th>T gondii</th>
<th>Toxocara sp</th>
<th>Ascaris sp</th>
</tr>
</thead>
<tbody>
<tr>
<td>Patients with presumed ocular histoplasmosis syndrome (n=23)</td>
<td>0 (0%)†</td>
<td>0 (0%)‡</td>
<td>13 (57%)</td>
<td>2 (8%)</td>
<td>6 (26%)</td>
</tr>
<tr>
<td>Controls (n=66)</td>
<td>0 (0%)</td>
<td>0 (0%)</td>
<td>42 (64%)</td>
<td>4 (6%)</td>
<td>22 (33%)</td>
</tr>
<tr>
<td>Controls with toxoplasma chorioretinitis (n=10)</td>
<td>0 (0%)</td>
<td>0 (0%)</td>
<td>10 (100%)</td>
<td>2 (20%)</td>
<td>8 (80%)</td>
</tr>
</tbody>
</table>

*Sera were tested for the presence of antigens of C neoformans.
†The positive control serum used showed only the presence of antibodies against H capsulatum and was negative in all other tests.
‡The positive control was included in the kit.

Labouratory test results

Testing for H capsulatum antibodies was negative in all people tested, both in patients and in controls (Table 2). The serum of the positive control, from a patient with systemic histoplasmosis, showed H and M bands in the immunodiffusion test (Fig 2).

All sera of patients and controls were negative for C neoformans antigens.

Thirteen of 23 patients (57%) with presumed ocular histoplasmosis syndrome were positive for IgG antibodies against T gondii compared with 42 of 66 healthy controls (64%). This was not significantly different (p >0.01).

IgG antibodies to Toxocara canis et cati were positive in two out of 23 patients with presumed ocular histoplasmosis syndrome compared with four out of 66 healthy controls and three out of 10 patients with toxoplasma chorioretinitis (p >0.01).

Six of 23 patients with presumed ocular histoplasmosis syndrome had IgG antibodies to Ascaris sp in their serum. Twenty two out of 66 controls (33%) had IgG antibodies, and eight of 10 patients (80%) with toxoplasma chorioretinitis. The patients with toxoplasma chorioretinitis had antibodies against Ascaris sp significantly more often than the healthy controls and patients with presumed ocular histoplasmosis (p <0.01).

Sera from 23 patients and 20 healthy controls were tested for the presence of retinal or choroidal autoantibodies by western blotting. Eight patients had chorioretinal autoantibodies compared with five of the healthy controls. No distinctive banding pattern was seen between the two groups.

Analysis of chorioretinal autoantibodies using immunohistochemistry on cryosections did not reveal a positive staining in the patients tested.

Figure 2  Immunodiffusion test for antibodies against H capsulatum. Left: positive control serum, showing H and M bands. Wells 1 and 4 positive serum provided with the kit, wells 2 and 5 serum from a patient positive for antibodies against H capsulatum, wells 3 and 6 serum from a patient negative for antibodies against H capsulatum. Well 7 Histoplasmin.

Right: two negative sera. Wells 1 and 4 positive serum provided with the kit, wells 2, 3, 5, and 6 serum from two patients negative for antibodies against H capsulatum. Well 7 Histoplasmin.
Ongkosuwito, Kortbeek, Van der Lelij, et al

Syndrome in the USA has been described, that called presumed ocular histoplasmosis. In Europe, an ocular syndrome identical to presumed ocular histoplasmosis syndrome has been described only twice. In these two patients, the clinical picture did not resemble the typical clinical characteristics of presumed ocular histoplasmosis syndrome. Serologically there has been no confirmation of a direct link between infection with *H capsulatum* and presumed ocular histoplasmosis syndrome, even in endemic areas. In histopathological studies of eyes from patients with presumed ocular histoplasmosis syndrome, no direct correlation could be demonstrated by immunohistochemical staining. In surgically removed membranes of patients suffering from presumed ocular histoplasmosis no fungi could be detected by electron microscopy and only non-specific inflammatory reaction tissue could be shown. Therefore, the ocular changes might be late immunological reactions to a previous infection with *H capsulatum*, or *H capsulatum* does not play a role at all in this syndrome.

In our study, no risk factors could be identified through our questionnaire that might have played a role in the pathogenesis of presumed ocular histoplasmosis syndrome. Careful assessment of travelling, eating habits, and animal contacts did not reveal any shared possible infectious risks.

None of our patients with presumed ocular histoplasmosis syndrome showed serological evidence of previous infection with *H capsulatum*, as was also the case in the group of healthy controls and disease controls. It is generally assumed that the manifestations of presumed ocular histoplasmosis syndrome is a late sequela of a previous infection with *H capsulatum* and that the interval between the primary infection and the first ocular symptoms might take many years. IgG antibodies against *H capsulatum* persist for years after an acute infection. Therefore, at least some of our patients with this ocular syndrome should have shown serological evidence of previous contact. Skin testing was not performed because of described reactivation of the ocular lesions in patients with macular disease.

It was possible to exclude as a causative mechanism other infectious agents that are present in Europe and that might induce choriororetinopathy, such as *C neoformans*, *T gondii*, *Toxocara* sp, and *Ascaris* sp. Of these, the first was negative in all people, while antibodies to the others were evenly distributed in all groups. When the group of patients with ocular toxoplasmosis (eight out of 10 had antibodies against *Ascaris* sp) was compared with the controls (22 out of 66 had antibodies against *Ascaris* sp), this difference seemed statistically significant (p = 0.05). Patients indicated the start of a new job, moving to another town or pregnancy among stress factors.

Discussion

In Europe, an ocular syndrome identical to that called presumed ocular histoplasmosis syndrome in the USA has been described, while *H capsulatum* is not endemic in this area. Several studies support an association between *H capsulatum* infection and the classic ocular triad of presumed ocular histoplasmosis in humans. This association is mainly based on epidemiological data from endemic areas and animal models, in which *H capsulatum* was injected into the carotid artery of primates which developed atypical choroidal lesions 4 weeks after inoculation. There are no reports of positive *H capsulatum* cultures in patients with presumed ocular histoplasmosis syndrome, but *H capsulatum* has been cultured from a patient with postoperative endophthalmitis. In another report, two immunocompetent patients with systemic histoplasmosis (confirmed by antibodies in serum as well as in urine) showed choroiditis, but no intraocular fluid analysis was performed. In these two patients the clinical picture did not show the complete typical clinical characteristics of presumed ocular histoplasmosis syndrome. Furthermore, one would expect that opportunistic ocular infections should be more frequently seen in immunocompromised patients, as up to 16% of these patients have a disseminated infection by *H capsulatum* in endemic areas. In the literature, however, positive cultures from ocular fluid samples obtained from patients with AIDS are described only twice. In these two patients the clinical picture did not resemble the typical

(57%) controls before obtaining the blood sample. Animal contact was not significantly different between patients and controls. No difference could be demonstrated when comparing patients and controls who gardened with or without gloves.

In both groups, one participant was vegetarian, and no difference could be found in the frequency of barbecuing or eating “raw” meat.

None of the patients, nor any controls, had had therapy for infection with herpes viruses, *T gondii*, *H capsulatum*, or *Cryptococcus neoformans*.

One of the patients and eight of the controls remembered having had contact with *Ascaris* sp or being treated with mebendazole for an infection with *Ascaris* sp; this was not statistically significant.

Three of 22 patients had a blood transfusion some time before the onset of ocular symptoms, as did six of 66 controls. One of the patients and also one of the controls had suffered from gonorrhoea in the past.

Eight out of 22 patients did mention a period of stress around the onset of ocular symptoms, compared with 11 of 66 controls during a similar period before blood sampling. This seemed statistically significant (p = 0.05). Patients indicated the start of a new job, moving to another town or pregnancy among stress factors.

Serologically there has been no confirmation of a direct link between infection with *H capsulatum* and presumed ocular histoplasmosis syndrome, even in endemic areas. In histopathological studies of eyes from patients with presumed ocular histoplasmosis syndrome, no direct correlation could be demonstrated by immunohistochemical staining. In surgically removed membranes of patients suffering from presumed ocular histoplasmosis no fungi could be detected by electron microscopy and only non-specific inflammatory reaction tissue could be shown. Therefore, the ocular changes might be late immunological reactions to a previous infection with *H capsulatum*, or *H capsulatum* does not play a role at all in this syndrome.

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Autoimmunity might also play an aetiological role in this syndrome. There have been reports of a relation with HLA-DRw2 or HLA-B7, showing a higher frequency in these patients with disciform lesions compared with
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healthy people. Racial differences might also play a role as this ocular disease occurs predominantly in white females. This is in accordance with our results, which included only white Europeans. Nevertheless, no clear evidence was found by western blotting and immunohistochemistry to suggest that the ocular syndrome might be the result of autoantibodies against choroid or retina.

In conclusion, no risk factors and no relation with the fungus *H capsulatum* could be detected. We therefore suggest that this ocular syndrome in patients from non-endemic areas should no longer be called presumed ocular histoplasmosis. The cause in our patients is probably multifactorial and might depend, as in other white dot syndromes, on interactions between triggering factors involved in presentation of antigens and the host immune system. Subsequent inflammation around choroidal dendritic cells can lead to scars seen in several white dot syndromes. Therefore, in European patients, it might be better to use as basis for the nomenclature not the aetiology but the clinical and histopathological presentation. Even in the United States presumed ocular histoplasmosis syndrome might be a misnomer; “multifocal choroidopathy” is, in our opinion, a more appropriate description.

We thank Dr L J Wheat from the Wishard Memorial Hospital, Indianapolis, Indiana, USA, for providing us with the serum of a patient with systemic histoplasmosis.

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